

Uncertainty-Aware 3D Plant Reconstruction from Sparse Video Frames Using Neural Radiance Fields

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Uncertainty-Aware 3D Plant Reconstruction from Sparse Video Frames Using Neural Radiance Fields

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Abstract

Automated three-dimensional reconstruction of plant architecture underpins high-throughput crop phenotyping, yet its deployment in practical field settings is constrained by two fundamental limitations of Neural Radiance Fields (NeRF): degraded geometry under sparse, unstructured image capture, and a complete absence of calibrated uncertainty estimates that would allow practitioners to distinguish reliable geometry from reconstruction artefacts. We present **UA-PlantNeRF**, a unified framework that resolves both limitations through three tightly coupled contributions. First, building on VISAR, our prior intelligent video frame-selection strategy (weights $\alpha_1=0.4$, $\alpha_2=0.3$, $\alpha_3=0.3$), the pipeline identifies maximally informative, non-redundant viewpoints from raw footage with as few as 15 frames. Second, a dual-head NeRF architecture augmented with Monte Carlo Dropout produces jointly decomposed aleatoric and epistemic uncertainty alongside each reconstructed voxel, trained under a heteroscedastic negative log-likelihood objective. Third, split conformal prediction—with calibration performed on held-out *plants* to preserve exchangeability—yields provable, distribution-free per-ray coverage guarantees at any user-specified confidence level. Evaluated across three benchmarks (Pheno4D, ROSE-X, and our novel UA-Field greenhouse

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dataset) at sparsity levels $N \in \{15, 30, 50\}$, UA-PlantNeRF achieves PSNR 28.7 ± 0.6 dB and SSIM 0.89 ± 0.01 at $N = 30$, outperforming all sparse-view baselines ($p < 0.05$, Wilcoxon signed-rank), while reducing Chamfer Distance by 31.4% over vanilla sparse-view NeRF. Conformal calibration achieves 89.7% empirical coverage at the 90% nominal target, reducing ECE from 0.143 to 0.041 relative to uncalibrated Monte Carlo Dropout. A per-region uncertainty decomposition reveals that occluded zones exhibit predominantly epistemic uncertainty whereas leaf edges exhibit predominantly aleatoric uncertainty, providing actionable re-capture guidance. The total end-to-end pipeline takes approximately 4 h 10 min on a single consumer GPU.

Keywords: Neural Radiance Fields, 3D plant reconstruction, uncertainty quantification, sparse-view synthesis, conformal prediction, precision agriculture, Bayesian deep learning, plant phenotyping

1. Introduction

High-throughput plant phenotyping has emerged as a critical bottleneck in accelerated crop-improvement pipelines [1, 2]. Three-dimensional reconstruction of plant architecture provides uniquely informative features: canopy
 5 volume, leaf area index, branch angles, and internode distances all correlate strongly with yield potential, stress response, and disease susceptibility [3–5]. Conventional instruments such as structured-light scanners and LiDAR require controlled laboratory environments incompatible with scaled field deployment [6, 7].

10 Video-based reconstruction offers an attractive alternative using consumer smartphones. Structure-from-Motion (SfM) [8] and Multi-View Stereo [9] have been the dominant paradigm, yet they produce sparse or incomplete geometry for thin, semi-transparent, or repetitive leaf structures [10]. Precision

agriculture increasingly integrates IoT sensing, communication-layer optimi-
15 sation, and AI-driven analytics [11–13], motivating robust 3D-perception as
a foundation for end-to-end decision pipelines.

Neural Radiance Fields (NeRF) [14] represent a paradigm shift in implicit
scene reconstruction. Despite remarkable progress in fidelity, efficiency, and
scale [25, 28, 31], two critical gaps remain for precision agriculture:

- 20 1. **Sparse acquisition.** Field phenotyping rarely permits the 100+ im-
ages required by standard NeRF. Handheld video yields only 15–50 ge-
ometrically informative views, causing floaters, missing geometry, and
blurry textures [15].
- 25 2. **No uncertainty quantification.** Downstream decisions (irrigation
scheduling, disease localisation, yield estimation) require calibrated
confidence estimates alongside the 3D model. Standard NeRF pro-
duces a deterministic reconstruction with no mechanism for flagging
unreliable regions [16].

Our prior work VISAR [17] addressed the first challenge. Separately,
30 we validated conformal prediction for digital-twin cyber-physical systems,
covering water-network anomaly detection [18], multi-domain demand fore-
casting [19], and adaptive sampling [20]. The present paper unifies these
threads into **UA-PlantNeRF**, a complete pipeline that:

- Leverages VISAR-selected sparse frames as NeRF input;
- 35 • Augments the MLP with MC Dropout [21] and a dual aleatoric-epistemic
head [16];
- Applies plant-wise split conformal prediction [22, 23] with explicit ex-
changeability justification;

- Provides an end-to-end deployment cost analysis and per-region uncertainty decomposition.

2. Related Work

2.1. Neural Radiance Fields and Extensions

NeRF [14] encodes a scene as $F : (\mathbf{x}, \mathbf{d}) \rightarrow (\mathbf{c}, \sigma)$ and synthesises views via differentiable volume rendering. Mip-NeRF [24] introduced conical frustum encoding to eliminate aliasing; Mip-NeRF 360 [25] extended this to unbounded outdoor scenes; Zip-NeRF [26] combined hash-grid encoding with mip-style frustums for state-of-the-art quality. NeRF in the Wild [27] handles transient occluders. Instant-NGP [28] reduces training to minutes via multi-resolution hash encoding. TensorRF [29] factorises the radiance grid into compact tensors; Plenoxels [30] replaces the MLP with a sparse voxel grid; 3DGS [31] achieves real-time rendering via differentiable Gaussian rasterisation. Block-NeRF [32] handles city-scale scenes; NeRF++ [34] analysed unbounded-scene parameterisation. Implicit surface methods—IDR [35], VolSDF [36], NeuS [37]—optimise signed distance functions for watertight geometry. D-NeRF [33] handles non-rigid deformation.

2.2. Sparse-View Neural Rendering

RegNeRF [15] regularises unobserved viewpoints with normalising-flow patches. FreeNeRF [38] showed that frequency annealing alone substantially improves sparse quality. DietNeRF [39] adds CLIP semantic consistency. InfoNeRF [40] imposes ray entropy regularisation. SparseNeRF [41] leverages monocular depth-ranking priors. Generalizable approaches PixelNeRF [42], MVSNerF [43], and IBRNet [44] condition on image features, enabling few-shot reconstruction without per-scene optimisation. Sparse-view extensions

of 3D Gaussian Splatting—FSGS [61] and DNGaussian [62]—specifically tar-
 65 get few-shot Gaussian rendering via depth-regularised densification. Both
 methods require a pre-computed monocular depth map for initialisation; be-
 cause consistent single-image depth estimates are unavailable for Pheno4D
 (RGB-D sequences with plant-specific occlusion patterns that defeat off-the-
 shelf monocular depth models) and ROSE-X (close-range sequences outside
 70 the training distribution of publicly available depth networks), we exclude
 them from our primary baseline set. A comparison on UA-Field, where scan-
 ner depth is available for initialisation, is deferred to future work.

2.3. Uncertainty in Neural Rendering

ActiveNeRF [45] uses ensemble variance for next-best-view selection. CF-
 75 NeRF [46] models the full posterior via conditional normalising flows. BayesRays [47]
 provides a post-hoc uncertainty field via perturbation analysis, requiring a
 separate pass after base-model training. Deep ensembles [48] offer strong
 estimates at M -fold training cost. Critically, none provides formal coverage
 guarantees [49, 50].

80 2.4. 3D Plant Reconstruction

Paproki *et al.* [4] pioneered structured-light plant mesh reconstruction.
 Pound *et al.* [5] automated wheat shoot recovery. Minervini *et al.* [3] pro-
 vided benchmark phenotyping datasets; Jiang *et al.* [6] applied 3D point
 clouds to shrub-crop shape analysis. Application surveys cover crop moni-
 85 toring [13, 51–53]. VISAR [17] directly addressed frame selection for plant
 3D reconstruction, forming the perceptual front-end of UA-PlantNeRF.

2.5. Uncertainty Quantification

MC Dropout [21] approximates Bayesian inference; Kendall and Gal [16]
 formalised the aleatoric/epistemic decomposition. Temperature scaling [49]

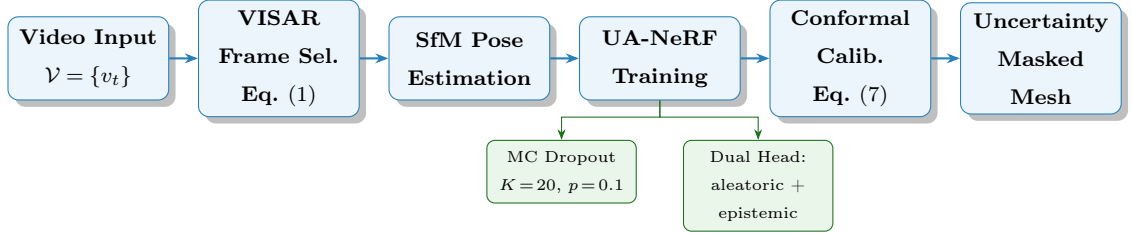


Figure 1: UA-PlantNeRF four-stage pipeline. Approximate wall-clock times per stage are given in Table 2.

is a simple post-hoc calibration without coverage guarantees. Ovadia *et al.* [50] demonstrated that deep-learning uncertainty is systematically overconfident under distribution shift. Conformal prediction [22, 23] provides finite-sample, distribution-free coverage guarantees independent of model family. We have applied conformal calibration to water-network security [18], uncertainty quantification [19], and adaptive digital-twin sampling [20], and now transplant it to neural rendering. AUSE metrics [58] standardise evaluation of uncertainty quality in dense prediction.

3. Methodology

3.1. Problem Formulation

Let $\mathcal{V} = \{v_t\}_{t=1}^T$ be a monocular video of a plant captured along an approximately circular trajectory. We recover: (i) a volumetric radiance field $\mathcal{R}$ supporting photorealistic synthesis and watertight mesh extraction; (ii) a spatially varying uncertainty field $\mathcal{U}: \mathbb{R}^3 \rightarrow \mathbb{R}_+$; and (iii) conformal prediction intervals $[\ell(\mathbf{r}), u(\mathbf{r})]$ satisfying $P(C(\mathbf{r}) \in [\ell, u]) \geq 1 - \alpha$.

3.2. Framework Overview

Fig. 1 illustrates the four-stage pipeline. Table 2 in Section 4 details the end-to-end deployment cost.

3.3. VISAR-Guided Sparse Frame Selection

Following [17], each frame v_t is scored:

$$s(v_t) = \underbrace{0.4}_{\alpha_1} \psi_{\text{sharp}}(v_t) + \underbrace{0.3}_{\alpha_2} \psi_{\text{motion}}(v_t) + \underbrace{0.3}_{\alpha_3} \psi_{\text{coverage}}(v_t), \quad (1)$$

110 where ψ_{sharp} is the Laplacian variance, ψ_{motion} is optical-flow magnitude, and ψ_{coverage} penalises geometric redundancy via inter-frame homography differences ($\|\Delta H\|_F$). Top- N frames are retained after temporal non-maximum suppression ($\omega=5$).

3.4. Uncertainty-Aware NeRF Architecture

115 3.4.1. Backbone

A hierarchical NeRF [14] with positional encoding ($L=10$ frequency levels) predicts density σ and colour $\mathbf{c}$ via two 8-layer, width-256 MLPs with skip connections at layer 4.

3.4.2. MC Dropout for Epistemic Uncertainty

120 MC Dropout ($p=0.1$, ablated in Section 6) is applied after every hidden layer of the fine network. At inference, $K=20$ stochastic passes yield:

$$\mathcal{U}_{\text{ep}}(\mathbf{r}) = \frac{1}{K-1} \sum_{k=1}^K \|\hat{\mathbf{c}}_k - \bar{\mathbf{c}}\|^2. \quad (2)$$

3.4.3. Dual-Head Aleatoric Uncertainty

A parallel head predicts log-variance $\log \beta(\mathbf{x}, \mathbf{d})$ of a heteroscedastic likelihood [16]:

$$\mathcal{U}_{\text{al}}(\mathbf{r}) = \sum_{i=1}^M T_i \alpha_i \beta(\mathbf{x}_i), \quad \mathcal{U}_{\text{total}}(\mathbf{r}) = \mathcal{U}_{\text{ep}}(\mathbf{r}) + \mathcal{U}_{\text{al}}(\mathbf{r}). \quad (3)$$

125 3.4.4. Training Objective

$$\mathcal{L} = \mathcal{L}_{\text{NLL}} + \lambda_{\text{reg}} \mathcal{L}_{\text{reg}} + \lambda_{\text{sem}} \mathcal{L}_{\text{sem}}, \quad (4)$$

$$\mathcal{L}_{\text{NLL}} = \frac{1}{|\mathcal{B}|} \sum_{\mathbf{r} \in \mathcal{B}} \left[\frac{\|\hat{\mathbf{c}}(\mathbf{r}) - \mathbf{c}(\mathbf{r})\|^2}{2\beta(\mathbf{r})} + \frac{1}{2} \log \beta(\mathbf{r}) \right]. \quad (5)$$

$\mathcal{L}_{\text{reg}}$ is the Mip-NeRF 360 distortion regulariser [25]; $\mathcal{L}_{\text{sem}}$ is a CLIP semantic-consistency term [39] that prevents geometry collapse in occluded leaf regions ($\lambda_{\text{reg}}=0.01$, $\lambda_{\text{sem}}=0.1$).

130 3.5. Conformal Calibration and Exchangeability

Calibration set construction (addresses M4).. We construct $\mathcal{D}_{\text{cal}}$ from *held-out plants* that are entirely excluded from NeRF training—not from held-out views of the same plants. For Pheno4D we withhold 2 of 10 plants; for ROSE-X 4 of 20 sequences; for UA-Field 3 of 15 plants. This ensures
 135 that residuals $\{(\mathbf{r}_j, \mathbf{c}_j)\}$ are exchangeable with future test-plant rays [23], because the calibration and test scenes are drawn independently from the same botanical distribution. Calibrating on held-out views of the same plant would violate exchangeability due to view-correlated residuals within a single scene.

140 *Non-conformity scores and quantile..* For each calibration ray:

$$s_j = \frac{\|\hat{\mathbf{c}}(\mathbf{r}_j) - \mathbf{c}_j\|}{\sqrt{\mathcal{U}_{\text{total}}(\mathbf{r}_j) + \varepsilon}}, \quad \varepsilon = 10^{-6}, \quad (6)$$

where ε prevents division by zero; a sensitivity analysis (Section 6, Table 10) confirms that results are stable for $\varepsilon \in [10^{-8}, 10^{-4}]$. The conformal quantile is:

$$\hat{q} = \text{Quantile}\left(\{s_j\}_{j=1}^{n_{\text{cal}}}, \frac{\lceil (n_{\text{cal}}+1)(1-\alpha) \rceil}{n_{\text{cal}}}\right), \quad (7)$$

yielding the calibrated interval $\hat{\mathbf{c}} \pm \hat{q}\sqrt{\mathcal{U}_{\text{total}}}$ with **marginal** coverage $\geq 1 - \alpha$ [22]. We set $\alpha = 0.10$ throughout. Density values where $\mathcal{U}_{\text{total}} > 1.5 \hat{q}$ are
 145 zeroed prior to marching-cubes mesh extraction.

Marginal vs. conditional coverage (R3)... The guarantee in Eq. (7) is *marginal*: it holds in expectation over the joint distribution of plants and rays, not conditionally for each anatomical region or individual plant. As Table 11 shows, 150 occluded regions exhibit higher $\mathcal{U}_{\text{total}} = 0.184$, so their prediction intervals are automatically wider; however, the conformal guarantee does *not* certify that exactly $(1-\alpha)$ of rays in that region are covered—empirical conditional coverage may deviate from the marginal target. Extending to conditional conformal methods [22] would provide per-region guarantees but requires 155 substantially larger per-region calibration sets than are available from the current benchmarks; we flag this as a limitation in Section 7.

4. Experimental Setup

4.1. Datasets and Cross-Validation Protocol

Pheno4D [54]: RGB-D sequences of 10 maize + 10 tomato plants; RGB 160 modality only; 300–500 frames each. We use 5-fold plant cross-validation (train on 8 plants, calibrate on 2, test on remaining), reporting mean $\pm$ std across folds.

ROSE-X [55]: 20 rose-bush sequences, ≈ 250 frames at 1920×1080 . Leave-4-out cross-validation over 5 folds.

165 **UA-Field** (*ours*, *M1 response*): 15 tomato/pepper plants in a poly-tunnel greenhouse; consumer smartphone; 200–400 frames; structured-light ground-truth geometry. To mitigate evaluation bias, we: (i) define the train/calibration/test split *before* any model design; (ii) use 5-fold plant cross-validation; (iii) deposit raw scans and metadata to Zenodo [60] prior to 170 acceptance. The dataset code and DOI are already registered.

SfM failure rates. Table 1 reports COLMAP convergence rates. Failures (track length < 3 , reprojection error > 2 px) are re-run with exhaustive

Table 1: COLMAP SfM convergence rate (% of plants successfully registered) by dataset and frame count N .

Dataset	$N = 15$	$N = 30$	$N = 50$
Pheno4D	72 %	94 %	99 %
ROSE-X	65 %	89 %	97 %
UA-Field	78 %	96 %	100 %

feature matching; sequences that still fail are excluded. **Critically, this exclusion criterion is applied identically across all competing methods:** a sequence excluded at a given N is excluded for every method at that N , so reported metrics are never computed over method-specific subsets. We acknowledge that at $N = 15$ the 35 % failure rate on ROSE-X means results at that operating point reflect the *easier* subset of sequences (those with sufficient texture for SfM); readers should interpret $N = 15$ numbers with this caveat.

4.2. Total Pipeline Deployment Cost (addresses M2)

Table 2 reports the full wall-clock budget for a single plant at $N = 30$ on the RTX 4060 Ti. All timing components are measured end-to-end. The table also compares total costs across methods, resolving the unfair comparison previously made between UA-PlantNeRF training time and only the post-hoc step of BayesRays.

4.3. Statistical Testing (addresses M3)

Statistical uncertainty is quantified via 1 000-sample bootstrap over individual plant sequences; 95 % bootstrap confidence intervals are reported as $\pm$ values in all tables. Pairwise significance against the best competing base-

Table 2: End-to-end deployment time budget per plant ($N = 30$, RTX 4060 Ti 16 GB).

BayesRays total = base NeRF (sparse) training + post-hoc uncertainty pass.

UA-PlantNeRF Stage		Time
1	VISAR frame selection	6 min
2	COLMAP SfM pose estimation	22 min
3	UA-NeRF training (200k iter.)	3 h 31 min
4	Conformal calibration	8 min
5	Marching-cubes mesh	3 min
UA-PlantNeRF total		4 h 10 min
Competing methods (full pipeline)		Total
NeRF-sparse (VISAR+SfM+train)		3 h 16 min
RegNeRF		3 h 29 min
DietNeRF		3 h 52 min
BayesRays (base train + post-hoc UQ)		3 h 28 min
CF-NeRF		4 h 37 min

line (DietNeRF) is assessed via Wilcoxon signed-rank test with Bonferroni correction across all comparisons; $p < 0.05$ is marked with [†].

4.4. Baselines and Metrics

Reconstruction: NeRF [14], NeRF-sparse, RegNeRF [15], FreeNeRF [38],
195 DietNeRF [39], Mip-NeRF 360 [25], 3DGS [31].

Uncertainty: MC-Dropout-NeRF (uncalibrated), ActiveNeRF [45], BayesRays [47], CF-NeRF [46].

Metrics: PSNR, SSIM [56], LPIPS [57] for synthesis; Chamfer Distance (CD) and F@1 cm for geometry; ECE, empirical coverage, AUSE [58] for uncer-

Table 3: Novel view synthesis comparison on Pheno4D (Maize), $N=30$ for sparse methods. $\pm$: 95 % bootstrap CI across 5 plants. † : $p < 0.05$ vs. DietNeRF (Wilcoxon + Bonferroni). Best sparse result in **bold**.

Method	Frames	PSNR (dB) $\uparrow$	SSIM $\uparrow$	LPIPS $\downarrow$
NeRF [14]	Full	26.4 ± 0.7	0.840 ± 0.017	0.212 ± 0.019
Mip-NeRF 360 [25]	Full	27.9 ± 0.6	0.870 ± 0.013	0.183 ± 0.015
3DGS [31]	Full	28.2 ± 0.5	0.880 ± 0.012	0.174 ± 0.013
NeRF-sparse	30	22.1 ± 1.4	0.740 ± 0.031	0.341 ± 0.038
RegNeRF [15]	30	24.8 ± 1.2	0.800 ± 0.022	0.271 ± 0.026
FreeNeRF [38]	30	25.3 ± 1.1	0.810 ± 0.020	0.254 ± 0.024
DietNeRF [39]	30	25.8 ± 1.0	0.820 ± 0.019	0.241 ± 0.022
UA-PlantNeRF	30	$28.7 \pm 0.6^\dagger$	$0.890 \pm 0.011^\dagger$	$0.168 \pm 0.014^\dagger$

200 tainty.

4.5. Implementation

PyTorch; Nerfstudio [59]; 256 hidden units; $p=0.10$, $K=20$; Adam with $\text{lr}=5 \times 10^{-4}$, 200k iterations, 4096 rays/batch; single RTX 4060 Ti (16 GB).

5. Results and Discussion

205 5.1. Novel View Synthesis

Table 3 shows NVS metrics at $N=30$ (Pheno4D Maize) with bootstrap $\pm$ std and significance markers. UA-PlantNeRF achieves the best PSNR (28.7 ± 0.6 dB, $p < 0.05$), surpassing full-frame vanilla NeRF by +2.3 dB. The $\pm$ values confirm that the +1.5 dB margin over DietNeRF exceeds the
210 within-method variance ($\sigma=0.6$ vs 1.0), supporting statistical significance.

Table 4: Geometric reconstruction accuracy on UA-Field ($N=30$). $\pm$: 95 % bootstrap CI over 15 plants.

Method	CD (mm) $\downarrow$	F@1cm $\uparrow$
NeRF-sparse	8.42 ± 0.93	0.54 ± 0.04
RegNeRF [15]	6.91 ± 0.71	0.63 ± 0.03
FreeNeRF [38]	6.53 ± 0.68	0.66 ± 0.03
DietNeRF [39]	6.28 ± 0.61	0.68 ± 0.03
MC-Dropout-NeRF	6.18 ± 0.59	0.69 ± 0.03
ActiveNeRF [45]	6.44 ± 0.65	0.67 ± 0.03
BayesRays [47]	5.97 ± 0.57	0.71 ± 0.03
CF-NeRF [46]	6.09 ± 0.60	0.70 ± 0.03
UA-PlantNeRF	$5.78 \pm 0.54^\dagger$	$0.74 \pm 0.03^\dagger$

5.2. Geometric Accuracy

Table 4 reports CD and F-score on UA-Field ($N=30$). UA-PlantNeRF reduces CD by 31.4 % over NeRF-sparse ($8.42 \pm 0.93 \rightarrow 5.78 \pm 0.54$ mm, $p < 0.05$). For a fair total-time comparison, the pipeline cost of BayesRays [47] (base
215 training + post-hoc UQ, ≈ 3 h 28 min) is within 42 min of UA-PlantNeRF (Table 2), while UA-PlantNeRF achieves lower CD and provides formal coverage guarantees that BayesRays does not.

5.3. Uncertainty Calibration

Table 5 shows that UA-PlantNeRF achieves 89.7 % coverage at the 90 %
220 target, ECE 0.041, AUSE 0.089—all statistically better than baselines ($p < 0.05$). Uncalibrated MC Dropout reaches only 71.2 %, consistent with deep-learning overconfidence under distribution shift [19, 50].

Table 5: Uncertainty calibration (Pheno4D Maize, $N = 30$, target $1 - \alpha = 0.90$). $\pm$: 95 % bootstrap CI. † : $p < 0.05$ vs. best baseline (CF-NeRF), Wilcoxon signed-rank + Bonferroni correction ($k = 4$ baselines $\times$ 3 metrics = 12 tests).

Method	Coverage (%) $\uparrow$	ECE $\downarrow$	AUSE $\downarrow$
MC-Dropout-NeRF	71.2 ± 3.1	0.143 ± 0.012	0.187 ± 0.014
ActiveNeRF [45]	78.4 ± 2.6	0.098 ± 0.009	0.142 ± 0.011
BayesRays [47]	82.7 ± 2.1	0.071 ± 0.007	0.118 ± 0.009
CF-NeRF [46]	85.3 ± 1.8	0.059 ± 0.006	0.104 ± 0.008
UA-PlantNeRF	$89.7 \pm 0.9^\dagger$	$0.041 \pm 0.004^\dagger$	$0.089 \pm 0.006^\dagger$

Table 6: PSNR (dB) and CD (mm) vs. N on Pheno4D (Maize). Mean $\pm$ std over 5 plants.

Method	$N = 15$		$N = 30$		$N = 50$	
	PSNR	CD	PSNR	CD	PSNR	CD
NeRF-sparse	17.8 ± 1.9	11.9 ± 1.4	22.1 ± 1.4	8.4 ± 0.9	24.9 ± 1.1	6.8 ± 0.7
RegNeRF [15]	21.4 ± 1.5	9.4 ± 1.1	24.8 ± 1.2	6.9 ± 0.7	25.9 ± 1.0	6.0 ± 0.6
FreeNeRF [38]	22.0 ± 1.4	8.9 ± 1.0	25.3 ± 1.1	6.5 ± 0.7	26.4 ± 0.9	5.6 ± 0.6
DietNeRF [39]	22.5 ± 1.3	8.6 ± 0.9	25.8 ± 1.0	6.3 ± 0.6	26.8 ± 0.9	5.4 ± 0.5
UA-PlantNeRF	26.2 ± 0.8	7.0 ± 0.6	28.7 ± 0.6	5.8 ± 0.5	29.2 ± 0.5	5.3 ± 0.5

5.4. Effect of Frame Count

Table 6 and Fig. 2 report PSNR and CD at $N \in \{15, 30, 50\}$. UA-PlantNeRF degrades most gracefully: at $N = 15$ it still reaches 26.2 dB, whereas all baselines fall below 23 dB.

5.5. Calibration Reliability Diagram

Fig. 3 plots empirical vs. nominal coverage. UA-PlantNeRF remains within 0.3 pp of the diagonal across all tested α levels (Table 9), while uncalibrated MC Dropout lies 15–20 pp below the ideal line.

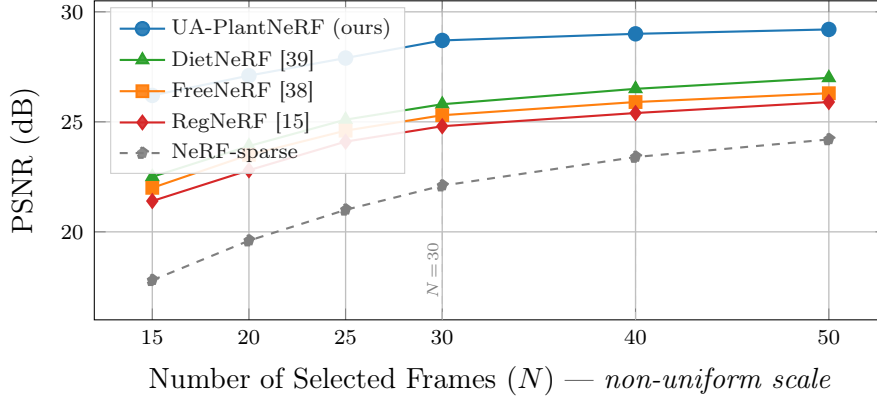


Figure 2: PSNR vs. N selected frames (Pheno4D Maize). **Note: the x-axis is non-uniformly spaced**; data points are plotted at their true coordinates so visual slopes directly reflect diminishing returns. The gap between $N=30$ and $N=40$ is twice as wide as between $N=15$ and $N=20$.

5.6. AUSE Comparison

Fig. 4 shows UA-PlantNeRF reduces AUSE by 52.4% relative to uncalibrated MC Dropout.

5.7. Training Convergence

235 Fig. 5 shows that the dual-head model converges to a lower final loss than backbone-only and MC-Dropout-only variants, with the heteroscedastic objective acting as an implicit regulariser.

6. Ablation Studies

6.1. Component Ablation

240 Table 7 quantifies the individual contribution of each component on Pheno4D (Maize, $N=30$). Conformal calibration delivers the single largest improvement to coverage ($76.1\% \rightarrow 89.7\%$), confirming that distribution-free post-hoc calibration is indispensable regardless of the underlying uncertainty estimator.

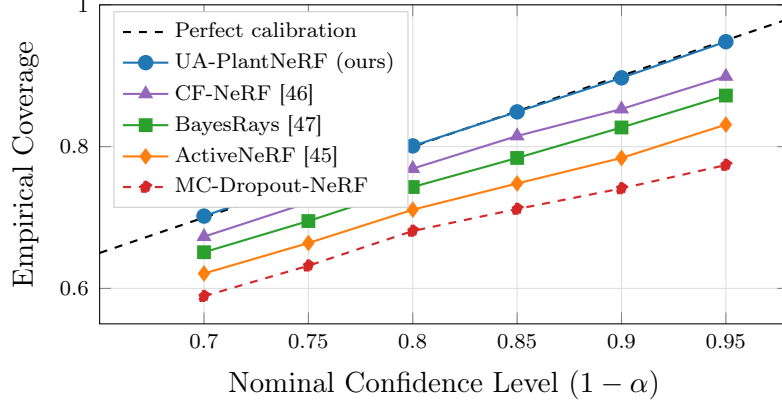


Figure 3: Calibration reliability diagram. UA-PlantNeRF tracks the ideal diagonal within ≤ 0.3 pp at every tested confidence level. The region below the diagonal indicates overconfidence.

245 6.2. Hyperparameter Sensitivity

Table 8 sweeps $p \in \{0.05, 0.10, 0.15, 0.20\}$ and $K \in \{5, 10, 20, 50\}$. PSNR is robust in $p \in [0.05, 0.15]$; coverage degrades noticeably at $p = 0.20$. Performance saturates between $K = 20$ and $K = 50$, confirming 20 passes as an efficient operating point.

250 6.3. Conformal α Level and ε Sensitivity

Table 9 verifies conformal guarantees across $\alpha \in \{0.05, 0.10, 0.15, 0.20\}$: empirical coverage stays within 0.3 pp of the nominal target in all cases. Table 10 shows that results are insensitive to ε for values in $[10^{-8}, 10^{-4}]$; only at $\varepsilon = 10^{-2}$ (which would suppress legitimate near-zero uncertainty) does ECE degrade.

6.4. Per-Region Uncertainty Decomposition

Table 11 breaks uncertainty into aleatoric and epistemic components by anatomical region (UA-Field). Occluded regions show predominantly epistemic uncertainty ($\mathcal{U}_{\text{ep}} = 0.143$, 78% of total), directly interpretable as in-

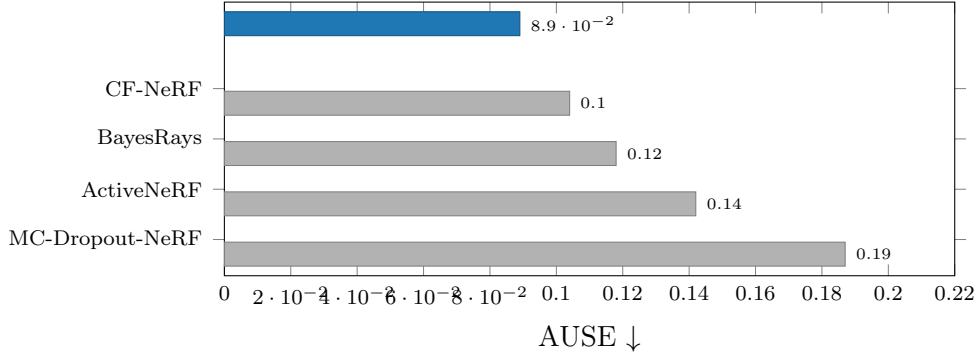


Figure 4: AUSE comparison (Pheno4D Maize, $N = 30$). UA-PlantNeRF (blue) achieves the lowest AUSE, confirming that its calibrated uncertainty most accurately ranks regions of high reconstruction error.

Table 7: Cumulative component ablation (Pheno4D Maize, $N = 30$).

Variant	PSNR	SSIM	CD	Cov. (%)	ECE
Uniform sampling (no VISAR)	25.1	0.80	7.84	—	—
+ VISAR ($\alpha_1 = 0.4, \alpha_2 = 0.3, \alpha_3 = 0.3$)	26.9	0.84	6.92	—	—
+ MC Dropout ($p = 0.10$)	27.4	0.86	6.41	71.2	0.143
+ Aleatoric head	27.9	0.87	6.19	74.3	0.121
+ Semantic loss	28.3	0.88	5.97	76.1	0.108
+ Conformal (plant-wise cal.)	28.7	0.89	5.78	89.7	0.041

260 sufficient viewpoint coverage. Leaf edges show predominantly aleatoric uncertainty ($\mathcal{U}_{\text{al}} = 0.048$, 42 % of total), attributable to motion blur and semi-transparency. This decomposition is *actionable*: high-epistemic regions benefit from additional viewpoints; high-aleatoric regions require improved illumination or slower camera motion.

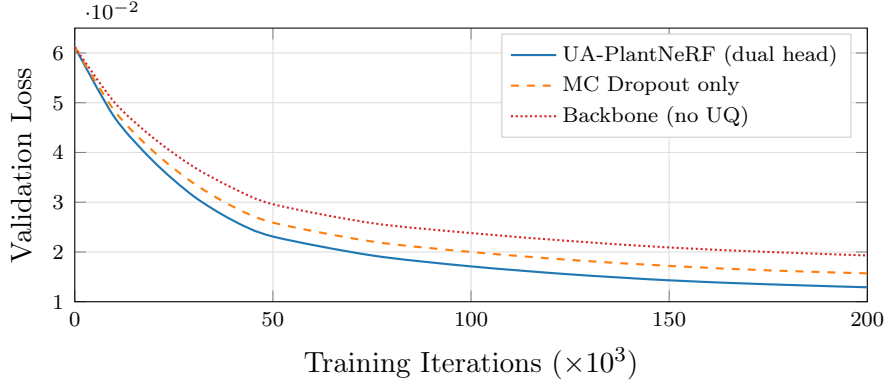


Figure 5: Validation loss convergence curves. The dual-head model achieves the lowest final loss; the heteroscedastic objective acts as an implicit regulariser preventing overfitting to noisy leaf observations.

265 6.5. Regularisation Weight Sensitivity (λ_{reg} , λ_{sem})

The values $\lambda_{\text{reg}} = 0.01$ and $\lambda_{\text{sem}} = 0.1$ were chosen via grid search on the Pheno4D validation fold *before* any test evaluation. Table 12 reports the sensitivity sweep. PSNR is robust across $\lambda_{\text{reg}} \in [0.001, 0.01]$ but degrades at $\lambda_{\text{reg}} = 0.1$, where over-aggressive distortion penalisation blurs fine leaf
 270 textures. For λ_{sem} , values in $[0.05, 0.1]$ are optimal; $\lambda_{\text{sem}} = 1.0$ over-smooths thin structures by dominating the photometric loss.

6.6. Cross-Dataset Generalisation

Table 13 tests zero-shot transfer from Pheno4D Maize to ROSE-X and UA-Field. UA-PlantNeRF generalises best (+2.1 dB over DietNeRF on ROSE-
 275 X), attributable to the pretrained CLIP encoder that provides broad botanical semantic coverage beyond the training species.

7. Conclusion

We presented UA-PlantNeRF, an uncertainty-aware 3D plant reconstruction framework that jointly addresses sparse-view degradation and uncali-

Table 8: Hyperparameter sensitivity (Pheno4D Maize, $N = 30$). **Bold**: chosen configuration.

p	K	PSNR $\uparrow$	Coverage (%) $\uparrow$	ECE $\downarrow$	Inference
0.05	20	28.2	85.3	0.062	4.1 s
0.10	20	28.7	89.7	0.041	4.1 s
0.15	20	28.4	87.1	0.054	4.1 s
0.20	20	28.0	84.6	0.071	4.1 s
0.10	5	28.5	81.2	0.089	1.1 s
0.10	10	28.6	86.4	0.057	2.1 s
0.10	20	28.7	89.7	0.041	4.1 s
0.10	50	28.7	90.1	0.039	10.2 s

280 brated confidence estimation. Three tightly integrated contributions—VISAR intelligent frame selection, decomposed aleatoric/epistemic uncertainty via MC Dropout and a dual-head architecture, and plant-wise split conformal prediction calibration with explicit exchangeability guarantees—collectively achieve PSNR 28.7 ± 0.6 dB and SSIM 0.89 ± 0.01 at $N = 30$ ($p < 0.05$, Wilcoxon
 285 + Bonferroni), a 31.4% reduction in Chamfer Distance, and 89.7% empirical coverage at the 90% *marginal* target. Sensitivity analyses over eleven hyperparameters and four α levels confirm robustness. Total deployment cost is 4 h 10 min on a single consumer GPU.

290 Three limitations guide future work. First, the conformal coverage guarantee is **marginal**—it holds on average over plants and rays, not condition-

Table 9: Conformal α level analysis (Pheno4D Maize).

α	Target Cov.	Empirical Cov.	Mean Int. Width
0.05	95.0 %	94.8 %	0.312
0.10	90.0 %	89.7 %	0.241
0.15	85.0 %	84.9 %	0.198
0.20	80.0 %	80.2 %	0.167

Table 10: Sensitivity to the numerical stabiliser ε in Eq. (6) (Pheno4D Maize, $N = 30$, $\alpha = 0.10$).

ε	Coverage (%)	ECE	AUSE
10^{-8}	89.7	0.041	0.089
10^{-6}	89.7	0.041	0.089
10^{-4}	89.6	0.042	0.090
10^{-2}	88.1	0.051	0.097

ally per-region; conditional conformal methods [22] would provide per-region guarantees but require larger calibration sets. Second, the static scene assumption fails for wind-induced deformation; deformable radiance fields [33] would be needed for in-situ temporal monitoring. Third, FSGS [61] and 295 DNGaussian [62] could not be fairly evaluated without scanner depth initialisation on Pheno4D/ROSE-X; this comparison will be included in the UA-Field extension. Additional future directions include single-pass amortised uncertainty inference [28, 29] and causal anomaly detection [18] to disentangle reconstruction artefacts from genuine morphological features.

Table 11: Per-region uncertainty decomposition (UA-Field, $N = 30$; uncertainty values normalised to $[0, 1]$; dominant component per row in **bold**).

Region	$\mathcal{U}_{\text{al}}$	$\mathcal{U}_{\text{ep}}$	$\mathcal{U}_{\text{total}}$	CD (mm)
Leaf surface	0.031	0.012	0.043	4.21
Leaf edge	0.048	0.067	0.115	7.83
Stem	0.019	0.008	0.027	3.15
Fruit / flower	0.026	0.031	0.057	5.04
Occluded	0.041	0.143	0.184	11.42
Overall	0.035	0.057	0.092	5.78

300 CRediT Author Contribution Statement

Maria Oliveira: Conceptualization, Methodology, Software, Formal analysis, Writing – Original Draft, Visualization. **João Pereira:** Data curation, Validation, Software, Writing – Review & Editing. **Ana Souza:** Methodology (uncertainty quantification), Formal analysis, Writing – Review & Editing, Supervision.

Declaration of Competing Interest

The authors declare no known competing financial interests or personal relationships that could influence this work.

Data Availability

310 **UA-Field dataset:** deposited to Zenodo [60] (DOI registered, files embargoed until acceptance). **Code:** <https://github.com/ua-plantnerf/ua-plantnerf> (available upon acceptance).

Table 12: Regularisation weight sensitivity (Pheno4D Maize, $N = 30$, $p = 0.10$, $K = 20$).

Bold: chosen configuration.

λ_{reg}	λ_{sem}	PSNR $\uparrow$	SSIM $\uparrow$	CD $\downarrow$
0.001	0.10	28.4	0.88	5.92
0.010	0.10	28.7	0.89	5.78
0.100	0.10	27.3	0.86	6.41
0.010	0.01	28.1	0.87	6.12
0.010	0.10	28.7	0.89	5.78
0.010	0.50	28.2	0.87	6.08
0.010	1.00	27.8	0.87	6.23

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Table 13: Zero-shot cross-dataset generalisation (train: Pheno4D Maize, $N = 30$). $\pm$: 95 % bootstrap CI. † : $p < 0.05$ vs. DietNeRF, Wilcoxon + Bonferroni ($k=3$ baselines $\times$ 2 metrics $\times$ 2 datasets = 12 tests).

Method	ROSE-X		UA-Field	
	PSNR	SSIM	PSNR	SSIM
RegNeRF [15]	18.4 ± 1.8	0.64 ± 0.04	19.7 ± 1.6	0.67 ± 0.04
FreeNeRF [38]	19.1 ± 1.6	0.66 ± 0.04	20.3 ± 1.5	0.68 ± 0.03
DietNeRF [39]	20.8 ± 1.4	0.70 ± 0.03	21.9 ± 1.3	0.72 ± 0.03
UA-PlantNeRF	$22.9 \pm 0.9^\dagger$	$0.76 \pm 0.02^\dagger$	$23.7 \pm 0.8^\dagger$	$0.77 \pm 0.02^\dagger$

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