

Fig. S1: A phylogenetic tree generated from the multiple sequence alignment (MSA) of the *NGR5* promoter and gene body regions, trimmed at a 5% gap threshold, showing the clustering of phylogenetically homogeneous core-set accessions. The x-axis depicts phylogenetic distance between clusters, and the triangle's size represents the number of accessions per cluster.

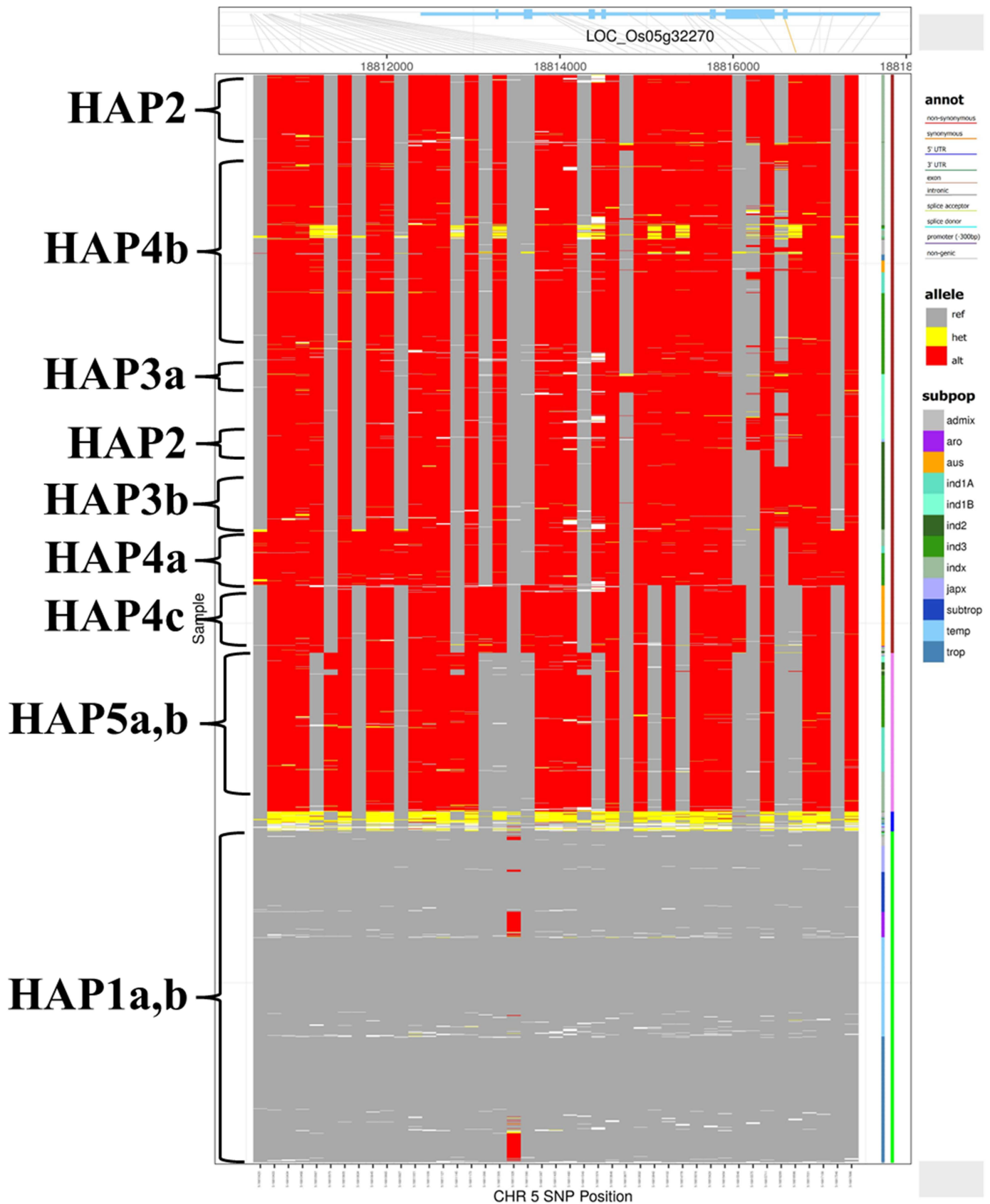


Fig. S2. Graphical representation of the *NGR5* haplotypes identified from the 3KRG SNP-Seek dataset. SNP positions across the *NGR5* gene body and promoter region are shown along the X-axis, while individual 3KRG accessions are arranged along the Y-axis. The top panel shows the *NGR5* gene structure and corresponding sequence variation, with blue boxes indicating exons, blue lines indicating introns, and the upstream, unfilled region representing the 2 kb promoter region. Grey, red, and yellow colours indicate reference alleles, alternate alleles, and heterozygous loci, respectively.

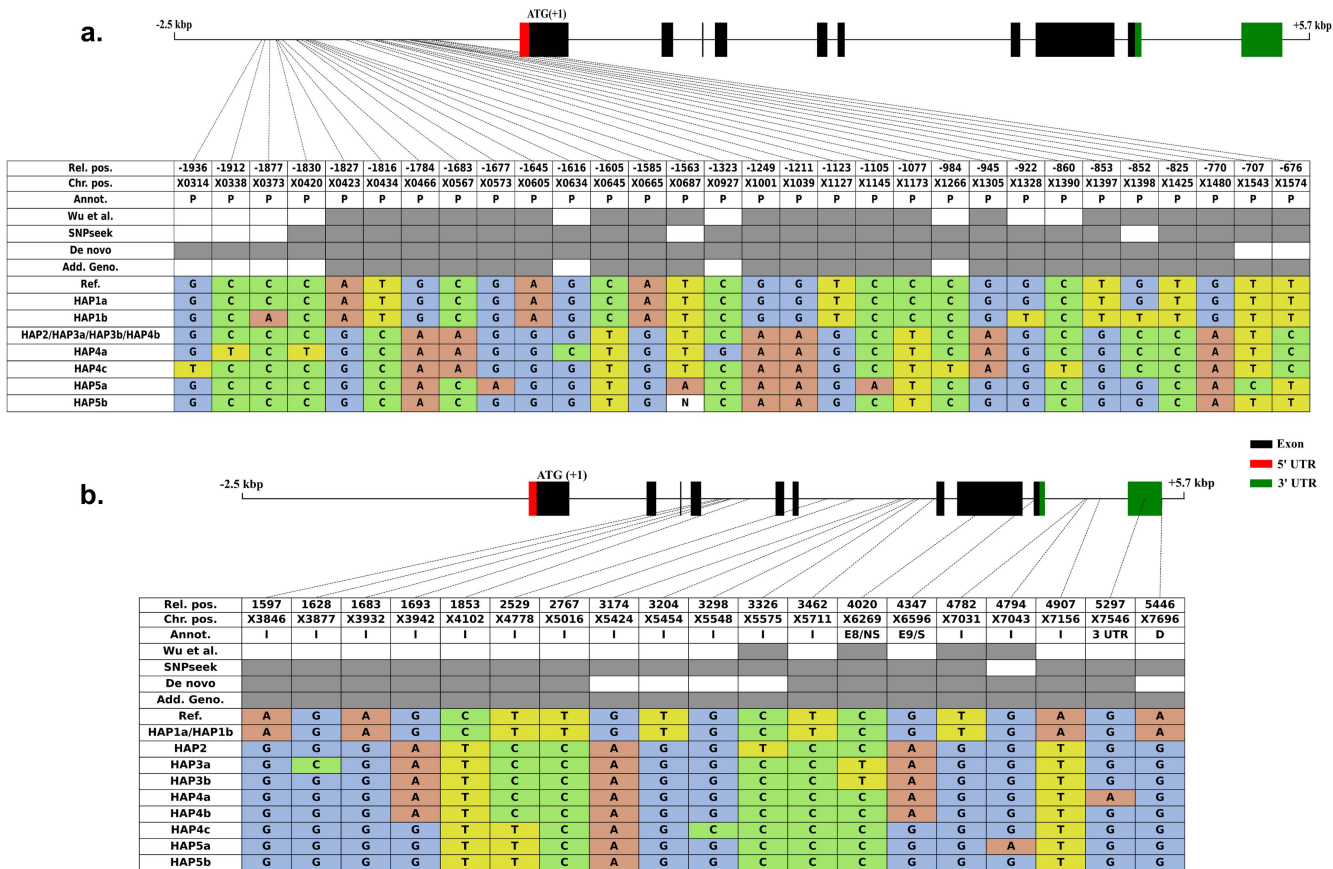


Fig. S3: Diagram showing haplotype-specific allelic variation in the *NGR5* promoter (a.) and gene body (b.) individually. The allelic variation presented here is a compilation of SNPs identified using three different approaches (as described in Figure 3). The 1st row (Rel. pos.) indicates relative chromosomal position with respect to the translation start site (TSS), while the 2nd row (Chr. pos.) indicates the actual chromosomal coordinates of the *NGR5* locus based on the IRGSP-1.0 reference genome. The first four digits of all positions (1881) are identical and therefore abbreviated as ‘X’. The 3rd row (Annot.) indicates the genomic annotation of each SNP site, where P: promoter, I: intron, E: exon, UTR: untranslated region, and D: downstream. The result of an SNP in an exon is classified as a NS (non-synonymous) or S (synonymous) mutation in the amino acid sequence. Rows 4–7 indicate SNP sites reported by Wu et al. (2020) and those identified using the different approaches used in this study. Row 8 (Ref.) indicates the nucleotide present at each position in the Nipponbare IRGSP-1.0 reference genome. Rows 9–15 (a.) and row 9-17 (b.) indicate the nucleotides present at the corresponding positions in the haplotypes.

a.

Primer Name	Sequence (5'→3')	Amplicon size (bp)
OsActin1_F	CTTCATAGGAATGGAAGCTGCGGGTA	197
OsActin1_R	CGACCACCTTGATCTTCATGCTGCTA	
NGR5_CDE6.1_F	GCAGTGCCTTTTCCAGGGGTTTAC	137
NGR5_CDE8.1_R	CCCATCCAGCGTGATGTCCTTC	

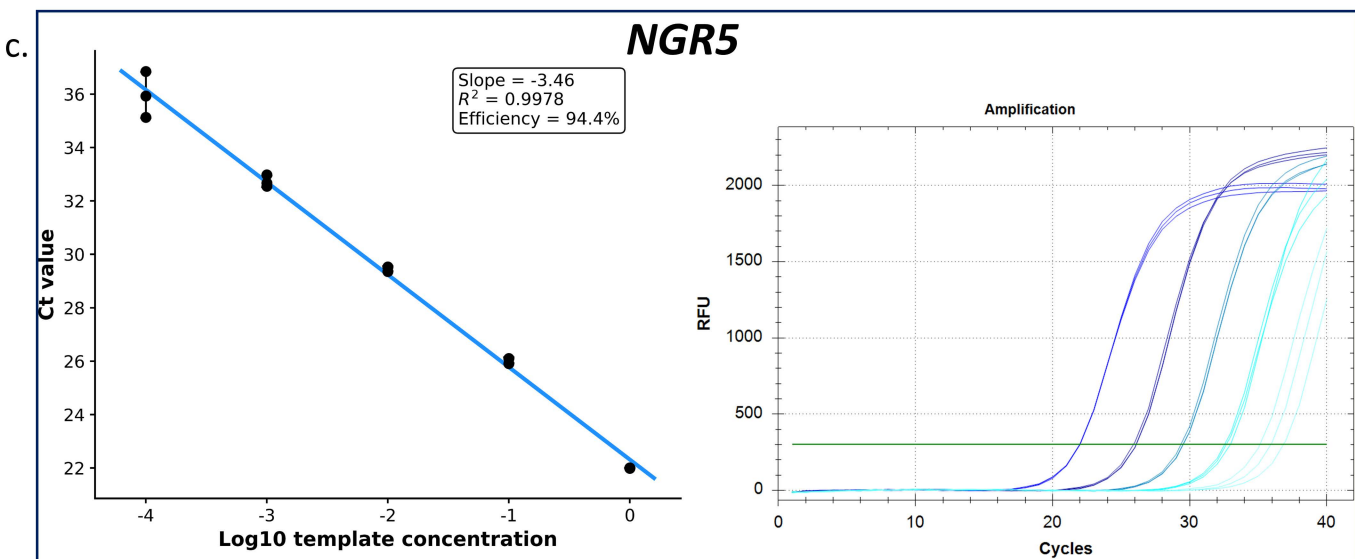
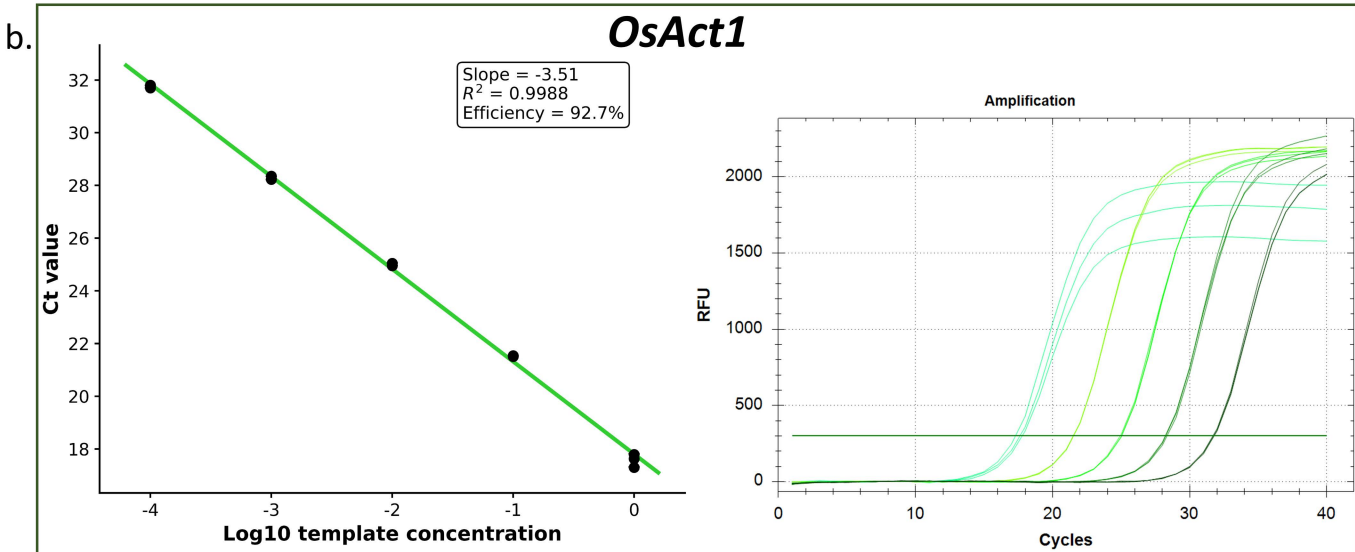


Fig. S4. Standard curve of qPCR primers. a. The table provides the primer sequence used in qPCR. Standard curve analysis of b. *OsAct1* and c. *NGR5* used in qPCR analysis

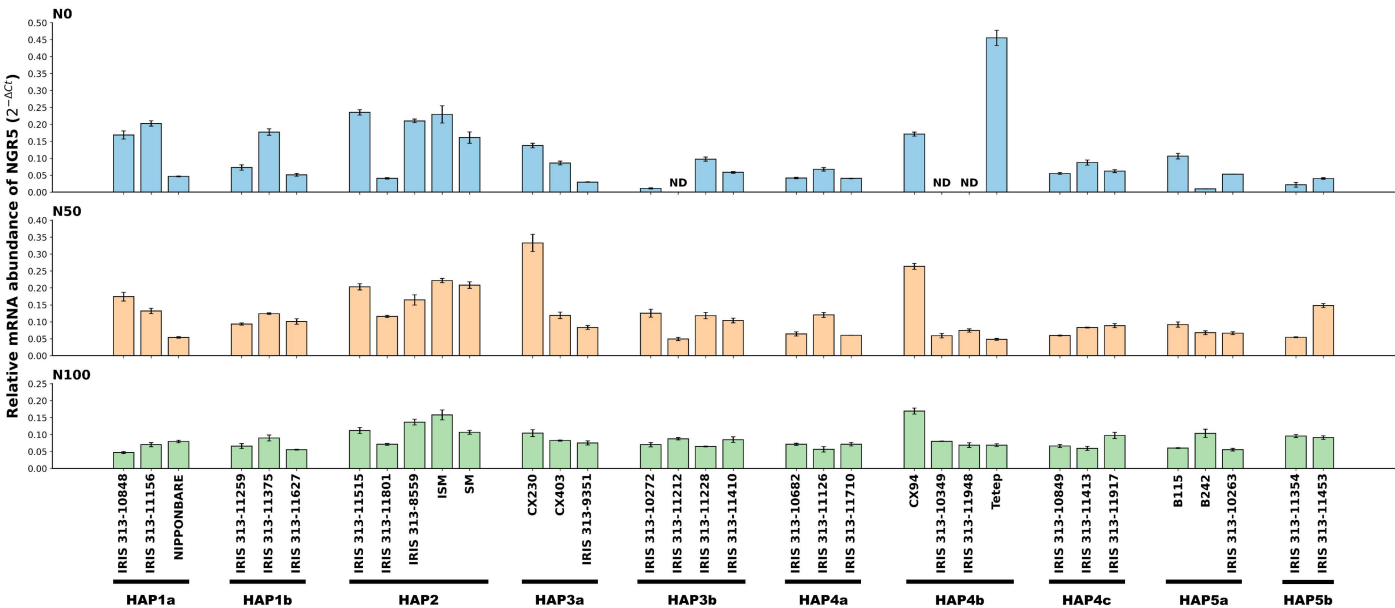
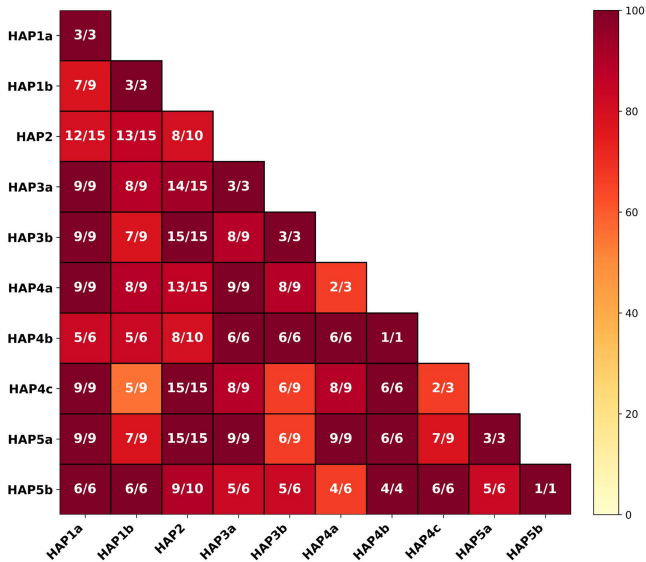
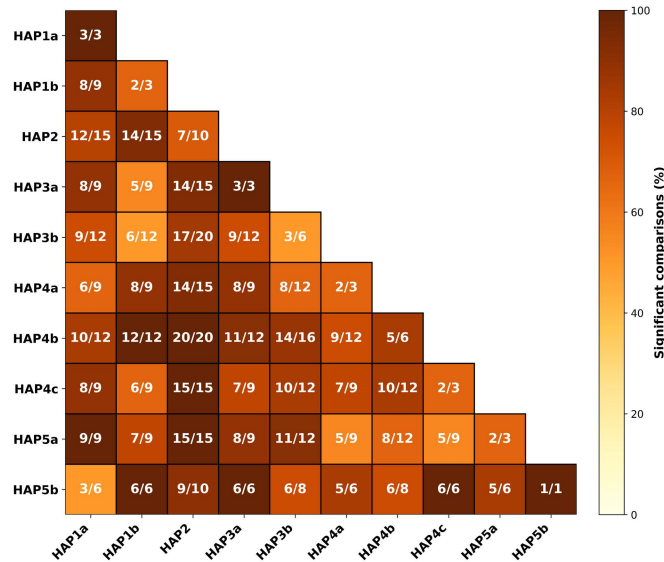


Fig. S5. Relative abundance of *NGR5* mRNA among rice genotypes of the different *NGR5* haplotypes under varying nitrogen conditions. Graphs show relative *NGR5* mRNA abundance in the shoot base tissue of rice genotypes grown under nitrogen-deficient (N0; 0 mM NH_4NO_3), moderate nitrogen (N50; 0.715 mM NH_4NO_3), and high nitrogen (N100; 1.43 mM NH_4NO_3) conditions. Data are shown as mean \pm SD of three technical replicates. Relative mRNA abundance was calculated as $2^{-\Delta Ct}$ using *OsActin1* as an internal control.

N0



N50



N100

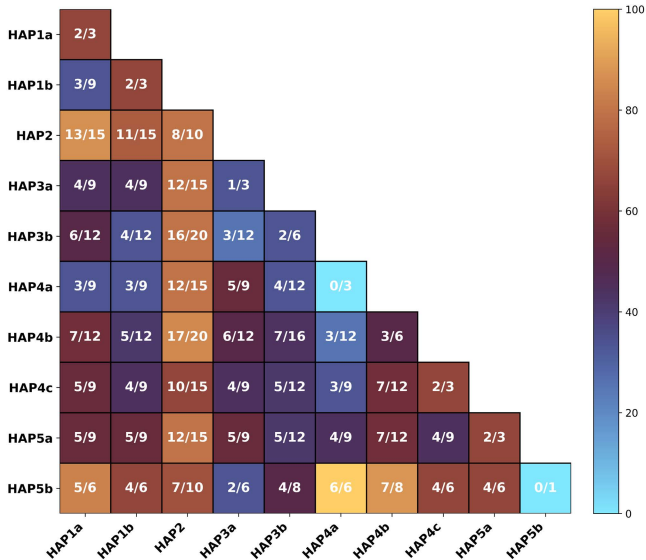
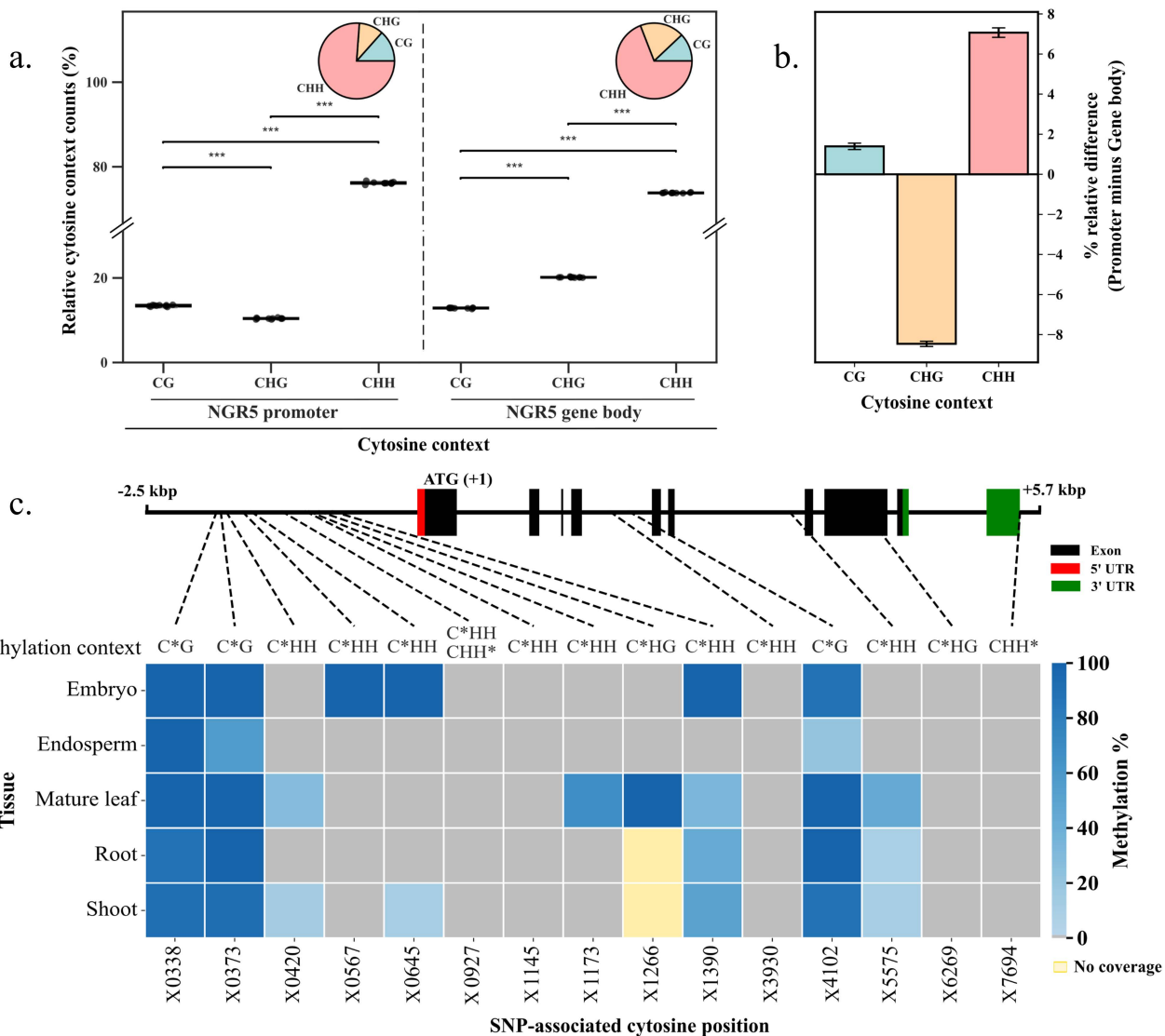


Fig. S6. Pairwise comparisons of *NGR5* expression levels reveal statistically significant differences both within and among haplotypes. The diagrams show pairwise comparisons of *NGR5* expression levels between genotypes across all haplotypes under a. N0, b. N50 and c. N100 conditions.



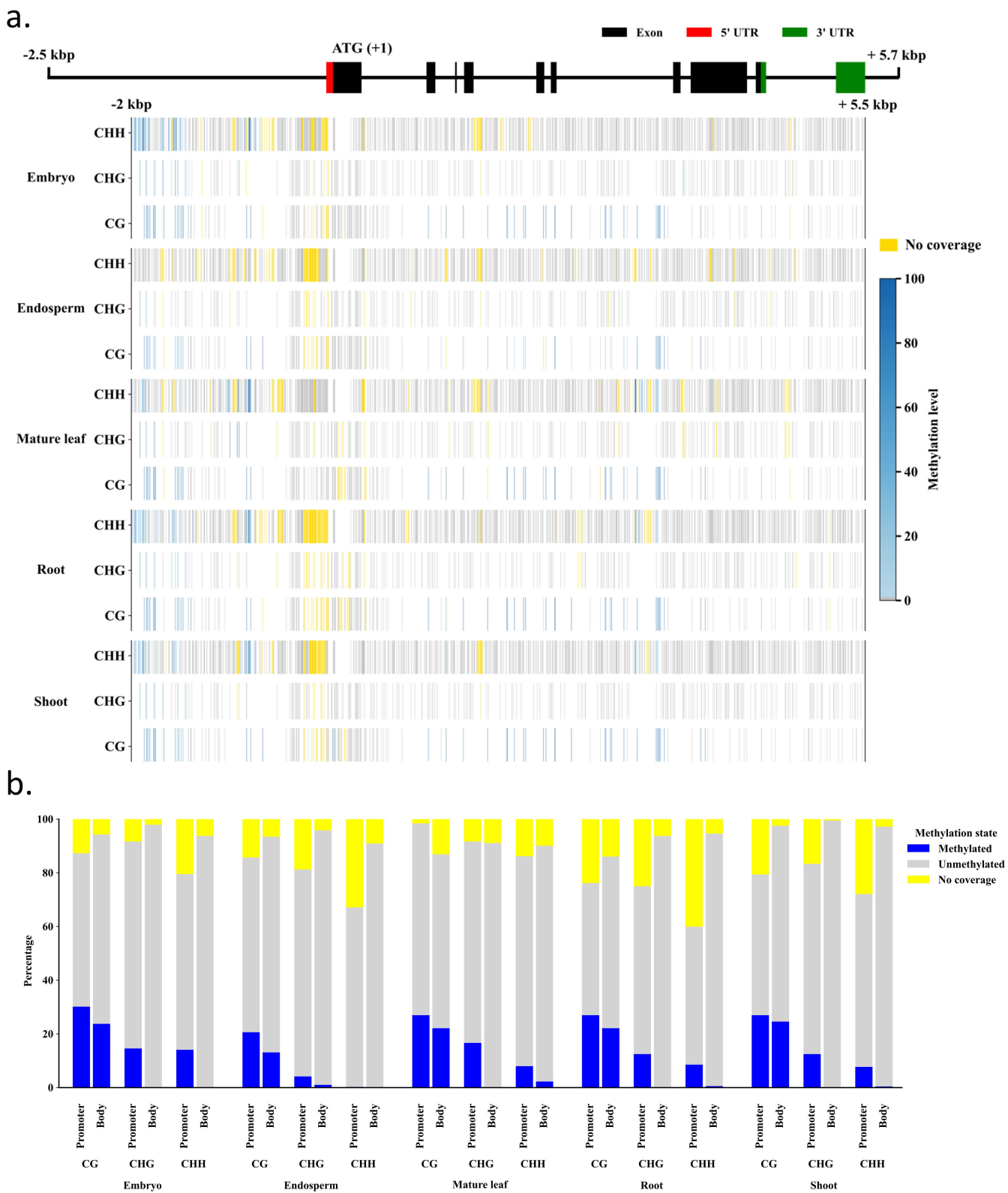


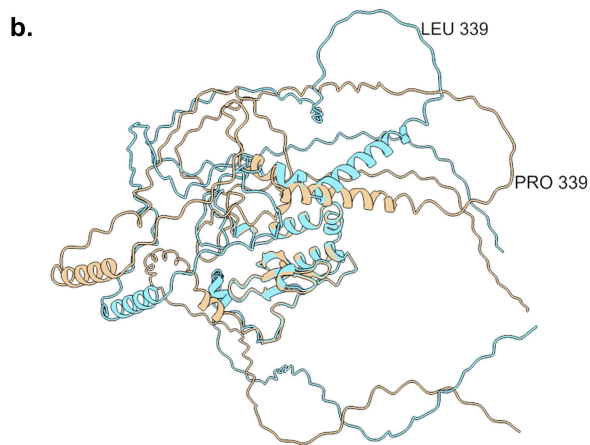
Fig. S8. Cytosine methylation landscape across the *NGR5* promoter and gene body.

a. Heatmap depicts the methylation landscape of all possible cytosine contexts (CG, CHG, and CHH) in the 2 kb upstream promoter region and the *NGR5* gene body (~5.5 kb downstream from the ATG (+1) site) across different tissues Nipponbare rice tissues. The upper panel represents the *NGR5* gene structure, where black boxes indicate exons, black lines indicate introns, and red and green regions represent the 5' and 3' untranslated regions (UTRs), respectively. Methylation levels are represented as a blue intensity gradient, where darker blue indicates higher methylation percentage and grey indicates absence of methylation, while yellow denotes positions lacking sequencing coverage. Methylation heatmap was generated using publicly available bisulfite sequencing data b. Bar graph representing the overall percentage distribution of methylated, unmethylated, and the non-covered cytosine sites across the promoter and gene body regions in different cytosine contexts across different tissues.



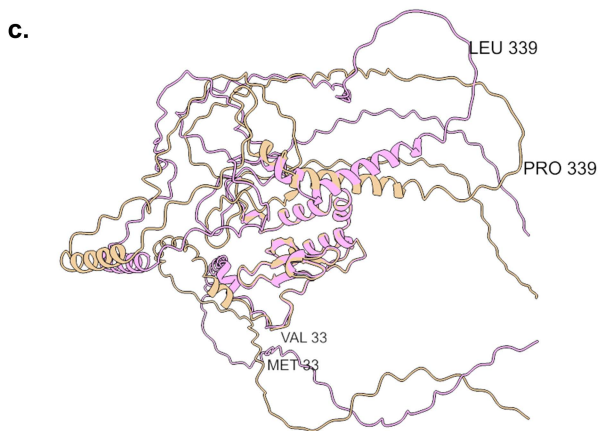
SA score = 2111.5

RMSD: 28 pruned atom pairs = 1.297Å;
across all 425 pairs = 15.774Å)



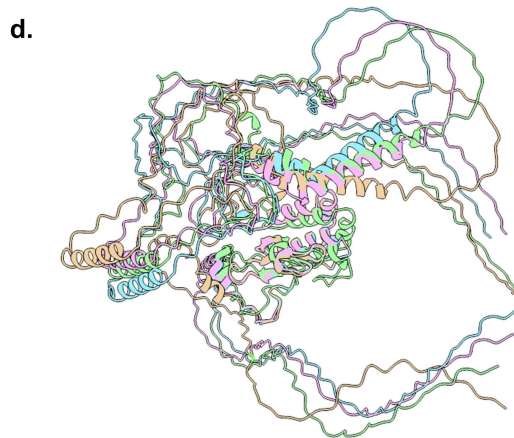
SA score = 2119.9

RMSD: 107 pruned atom pairs = 0.512Å;
across all 425 pairs = 18.445Å)



SA score = 2108.1

RMSD: 108 pruned atom pairs = 0.504Å;
across all 425 pairs = 15.215Å)



 **NGR5: WT**

 **NGR5: P339L**

 **NGR5: M33V**

 **NGR5: M33V & P339L**

Fig. S9. Predicted three-dimensional (3D) structural variation among NGR5 protein variants carrying haplotype-associated amino acid substitutions. Diagrams show AlphaFold predicted structures of NGR5 protein variants, superimposed on the predicted 3D structure of NGR5 of Nipponbare as the reference: a. M33V variant b. P339L variant c. M33V and P339L variants d. The superimposed structures of all the variants and the reference NGR5 protein. The structural similarity among the predicted models was evaluated using root-mean-square deviation (RMSD) and structure alignment (SA) scores.