

1 Supplementary for the Paper: Multiphasic
2 Transport in the Human Cranial Dura: How
3 Meningeal Lymphatic Vessels Drive Waste
4 Clearance from Brain Cerebrospinal Fluid

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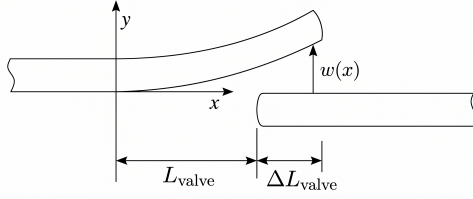


Fig. A: Schematic representation of the lymphatic valve [1]

24 **A mLV Valve Model**

25 We use the model of [1] to obtain an explicit solution of the valve opening of initial
 26 lymphatics and consequently estimate the flow passing through this valve. The single
 27 primary lymphatic valve is formed by an overlapping endothelial flap of length L_{valve}
 28 (free portion) (see Figure A). The flap behaves as an Euler–Bernoulli beam clamped
 29 at $x = 0$ and free at $x = L_{\text{valve}} + \Delta L_{\text{valve}}$. We assume a constant trans-wall pressure
 30 difference Δp to obtain the explicit solution.

31 The beam equation is

$$EI w''''(x) = \Delta p, \quad (1)$$

32 with boundary conditions

$$w(0) = 0, \quad w'(0) = 0, \quad w''(L_{\text{valve}} + \Delta L_{\text{valve}}) = 0, \quad w'''(L_{\text{valve}} + \Delta L_{\text{valve}}) = 0. \quad (2)$$

33 Solving

$$w(x) = \frac{\Delta p}{24 EI} x^2 (x^2 - 4(L_{\text{valve}} + \Delta L_{\text{valve}})x + 6(L_{\text{valve}} + \Delta L_{\text{valve}})^2), \quad 0 \leq x \leq L_{\text{valve}} + \Delta L_{\text{valve}}. \quad (3)$$

34 We consider steady, incompressible flow between two plates separated by the height
 35 $w(x)$. Let μ be the dynamic viscosity, T the valve depth, and Q the volumetric flow
 36 rate.

37 The lubrication equation is

$$\frac{dp}{dx} = -\frac{12\mu q}{w(x)^3} = -\frac{12\mu Q}{T w(x)^3}. \quad (4)$$

38 If we assume a constant pressure gradient, it follows that

$$Q = \frac{\Delta P w^3(x) T}{12 L_{\text{valve}} \mu}, \quad (5)$$

39 taking the average over x , we have:

$$\bar{Q} = \frac{\Delta P T}{12 L_{\text{valve}}^2 \mu} \int_0^{L+\Delta L} w^3(x) dx \equiv Q_{\text{valve}}. \quad (6)$$

40 From [2], the permeability relation can be written as

$$L_p = \frac{Q_{\text{prod}}}{S_{\text{mLV}} \Delta p}. \quad (7)$$

41 At the *microscale*, we apply (7) to a single primary lymphatic valve:

$$L_{p,\text{micro}} = \frac{Q_{\text{valve}}}{S_{\text{valve}} \Delta p}, \quad (8)$$

42 where S_{valve} is the local valve surface area.

43 Then we have that the estimated number of valves can be computed by

$$N_{\text{valve}} = \frac{Q_{\text{prod}}}{Q_{\text{valve}}}. \quad (9)$$

44 It follows that we can write the following surface-area fraction

$$\phi_{\text{valve}} = \frac{N_{\text{valve}} S_{\text{valve}}}{S_{\text{mLV}}}, \quad (10)$$

45 and we have

$$L_p = \phi_{\text{valve}} L_{p,\text{micro}}. \quad (11)$$

46 Similarly, the structural model of the valve opening can be extended to estimate
 47 the molecular permeability P_{lymph} . We consider the transport of a solute across the
 48 valve governed by both diffusion and advection. At the microscale, the local solute
 49 flux $J(x)$ (molar or mass rate) through the cross-section $A(x) = w(x)T$ is described
 50 by Fick's first law and the convective contribution:

$$J(x) = -DTw(x)\frac{dC}{dx} + (1 - \sigma)Q_{\text{valve}}C(x), \quad (12)$$

51 where D is the diffusion coefficient of the solute in the lymph and σ is the reflection
 52 coefficient. For the open valve configuration where $w(x)$ is significantly larger than the
 53 solute's molecular radius, we assume $\sigma \approx 0$.

54 To obtain an explicit solution for the effective molecular permeability, we consider
 55 a steady-state concentration difference $\Delta C = C_i - C_L$, where C_i is the interstitial
 56 concentration and C_L is the concentration in the lymphatic lumen. Under the phys-
 57 iological assumption of a *sink condition* in the lymphatic vessels, where the solute is
 58 rapidly cleared by the flow, we set $C_L = 0$ and $\Delta C = C_i$.

59 The total solute flux passing through a single primary valve, J_{valve} , is the sum of
 60 the diffusive and advective components:

$$J_{\text{valve}} = J_{\text{diff}} + J_{\text{adv}} = \left(\frac{DT}{\int_0^{L_{\text{valve}} + \Delta L_{\text{valve}}} \frac{1}{w(x)} dx} \right) \Delta C + Q_{\text{valve}} \bar{C}, \quad (13)$$

61 where \bar{C} is the average concentration of the solute in the trans-valve flow. Under the
 62 sink condition $C_L = 0$ and assuming the fluid enters the valve from the interstitium,
 63 we have $\bar{C} = C_i = \Delta C$.

64 We define the microscale effective molecular permeability as:

$$P_{\text{micro,eff}} = \frac{J_{\text{valve}}}{S_{\text{valve}}\Delta C} = P_{\text{micro,diff}} + \frac{Q_{\text{valve}}}{S_{\text{valve}}}, \quad (14)$$

65 where $P_{\text{micro,diff}}$ is the purely diffusive permeability obtained from the integral of the
66 inverse gap height $1/w(x)$.

67 Finally, applying the same upscaling framework used for the hydraulic permeabil-
68 ity, the macroscale effective molecular permeability P_{eff} of the lymphatic vessel wall
69 is given by:

$$P_{\text{eff}} = \phi_{\text{valve}}P_{\text{micro,eff}}. \quad (15)$$

70 Substituting the surface-area fraction ϕ_{valve} , we obtain the final relation:

$$P_{\text{eff}} = \phi_{\text{valve}}P_{\text{micro,diff}} + \frac{Q_{\text{flux}}}{S_{\text{mLV}}}, \quad (16)$$

71 where Q_{flux} is the net volumetric drainage of the network. This formulation allows us
72 to estimate the total solute clearance $F_{\text{solute}} = P_{\text{eff}}S_{\text{mLV}}C_i$ directly from the structural
73 parameters of the Mendoza model.

74 A.1 QMC Analysis

75 The QMC simulation is based on 60 million samples. The validity of each parameter
76 set was evaluated against a multi-objective cost function. The algorithm minimizes
77 the cumulative relative error between the model outputs and two specific physiological
78 targets:

- 79 1. **Net Lymphatic Flux (Q_{flux}):** The drainage capacity of the network. We set as
80 target the production rate of $0.5 \text{ L/day} \approx 5.8 \times 10^{-9} \text{ m}^3/\text{s}$ [3] against our computed
81 total $Q_{\text{flux}} = -L_p S_{\text{mLV}} \Delta P_{\text{at}}$.

82 **2. Total Vascular Volume:** The volume occupied by the mLVs network; we test
83 two targets: $V_{\text{mLV}} \approx 6 \times 10^{-6} \text{ m}^3$ [4] and $V_{\text{mLV}} = 4.57 \times 10^{-7} \text{ m}^3$ (see Section F)
84 against our computed volume of $S_{\text{tot}}r/2$.

The sampled parameters and ranges are

$$\Delta P_{\text{at}} \in [-1750, -400] \text{ Pa}, \quad L_p \in [10^{-12}, 10^{-8}] \text{ m/Pa} \cdot \text{s},$$

$$r \in [5 \times 10^{-6}, 2 \times 10^{-5}] \text{ m}, \quad L_{\text{FIX}} = L_{\text{valve}} + \Delta L_{\text{valve}} \in [5 \times 10^{-7}, 6 \times 10^{-6}] \text{ m}.$$

85 For a porosity of 0.5%, the near-optimal subset (9,207 accepted samples) yields the
86 following mean values, 95% confidence intervals, and coefficients of variation (CV):

$$\Delta P_{\text{at}} = -6.71 \times 10^2 [-6.76 \times 10^2, -6.66 \times 10^2] \text{ Pa}, \quad (\text{CV} = 37.0\%)$$

$$L_p = 4.42 \times 10^{-10} [4.37 \times 10^{-10}, 4.46 \times 10^{-10}] \text{ m/Pa} \cdot \text{s}, \quad (\text{CV} = 48.5\%)$$

$$r = 4.02 \times 10^{-5} [4.00 \times 10^{-5}, 4.05 \times 10^{-5}] \text{ m}, \quad (\text{CV} = 34.4\%)$$

$$L_{\text{FIX}} = 6.11 \times 10^{-7} [6.09 \times 10^{-7}, 6.13 \times 10^{-7}] \text{ m}, \quad (\text{CV} = 13.2\%)$$

87 Using the confidence interval means, the derived total vessel surface area for the
88 active network is $S_{\text{mLV}} \approx 1.95 \times 10^{-2} \text{ m}^2$.

89 The global sensitivity analysis (Table A) evaluates both the linear (Pearson) and
90 monotonic nonlinear (Spearman rank) correlations to the estimated flux across the
91 entire valid parameter space. It indicates that the net flux shows near-zero marginal
92 correlation to individual parameters under both metrics.

93 Within this accepted near-optimal subset, strong structural correlations emerge
94 (Figure B). Hydraulic conductivity L_p shows strong positive correlation with both
95 radius r (0.71) and transmural pressure ΔP_{at} (0.58), while transmural pressure also
96 correlates positively with L_{FIX} (0.51).

Table A: Sensitivity Analysis (Global Correlations) - 0.5% Porosity

Parameter	Pearson w/ Q_{flux}	Spearman w/ Q_{flux}
ΔP_{at}	-0.0015	-0.0021
L_p	-0.0015	-0.0022
r	0.0007	0.0015
L_{FIX}	-0.0004	-0.0011

97 Conversely, for the case of 6.5% porosity, the near-optimal subset (727 accepted
98 samples) yields the following adjusted mean values, confidence intervals, and CVs:

$$\Delta P_{\text{at}} = -6.78 \times 10^2 [-6.97 \times 10^2, -6.60 \times 10^2] \text{ Pa, } (\text{CV} = 37.1\%)$$

$$L_p = 3.28 \times 10^{-11} [3.16 \times 10^{-11}, 3.40 \times 10^{-11}] \text{ m/Pa} \cdot \text{s, } (\text{CV} = 50.0\%)$$

$$r = 3.97 \times 10^{-5} [3.87 \times 10^{-5}, 4.07 \times 10^{-5}] \text{ m, } (\text{CV} = 35.7\%)$$

$$L_{\text{FIX}} = 6.08 \times 10^{-7} [6.03 \times 10^{-7}, 6.14 \times 10^{-7}] \text{ m, } (\text{CV} = 13.2\%)$$

99 Using these corresponding CI means, the derived total vessel surface area for the
100 active network scales up to $S_{\text{mLV}} \approx 2.60 \times 10^{-1} \text{ m}^2$.

101 The global sensitivity analysis for this higher porosity level (Figure B) evaluates
102 the same metrics. Similar to the lower porosity case, it indicates that the net flux
103 maintains a near-zero marginal correlation to individual parameters.

Table B: Sensitivity Analysis (Global Correlations) - 6.5% Porosity

Parameter	Pearson w/ Q_{flux}	Spearman w/ Q_{flux}
ΔP_{at}	-0.0018	-0.0035
L_p	-0.0004	-0.0004
r	0.0015	0.0029
L_{FIX}	-0.0017	-0.0023

104 Within this accepted near-optimal subset, strong structural correlations again
 105 emerge (Figure B). Consistent with the 0.5% porosity case, hydraulic conductivity L_p
 106 shows a strong positive correlation with both radius r (0.73) and transmural pressure
 107 ΔP_{at} (0.56), while transmural pressure continues to correlate with L_{FIX} (0.51).

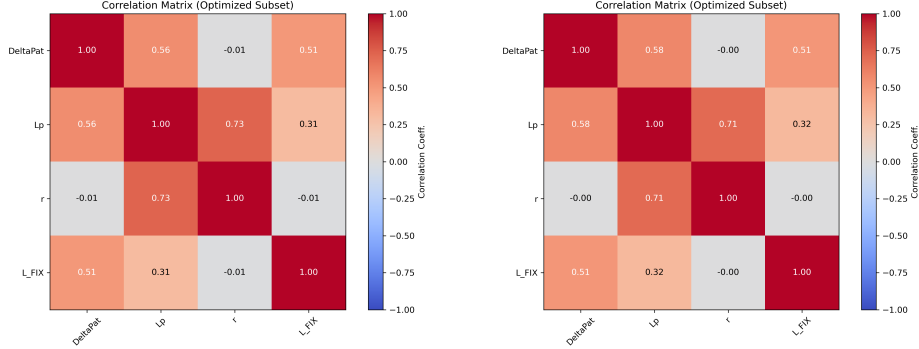


Fig. B: The correlation matrix of the near-optimal subset obtained from the Quasi-Monte Carlo simulation, illustrating the morphometric and mechanical trade-offs of the dural lymphatic network. **Left:** Case with porosity $\phi_L = 6.5\%$. **Right:** Case with porosity $\phi_L = 0.5\%$.

108 The global sensitivity analysis (Tables A and B) reveals that the Net Lymphatic
 109 Flux (Q_{flux}) exhibits near-zero marginal correlation to any individual parameter under
 110 both linear (Pearson) and rank-based (Spearman) metrics. This is due to the fact
 111 that ($Q_{flux} \propto -L_p S_{mLV} \Delta P_{at}$), which means that the drainage capacity cannot be dic-
 112 tated by a single "bottleneck" variable but requires the simultaneous co-optimization
 113 of permeability (L_p), network surface area (S_{mLV}), and the transmural pressure gra-
 114 dient (ΔP_{at}). Notably, while L_p and S_{mLV} differ significantly depending on the tissue
 115 porosity, their product ($L_p S_{mLV}$) remains consistent, emphasizing that the system
 116 fundamentally relies on the balance between permeability and available surface area
 117 to maintain adequate flux. The model successfully found an effective vessel average
 118 radius of $r \approx 39.7$ to $40.2 \mu m$ across the two porosity cases, which falls perfectly within
 119 the observed anatomical bounds for initial and collecting lymphatic vessels [5, 6].

120 Using the estimation above, we can compute the effective permeability of the lym-
121 phatic vessels as well using equation (16). We test the simulation with different ϕ_L
122 and we notice that, as for the parameter L_p , what remains constant through different
123 lymphatic permeabilities is the product $P_{\text{eff}}S_{\text{mLV}} = 5.79 \times 10^{-9} \text{ m}^3/\text{s}$. For model-
124 ing purpose, we define the quantity $\beta = \frac{P_{\text{eff}}S_{\text{mLV}}}{V_{\text{dura}}} \approx 0.228 \text{ 1/h}$. By employing our
125 estimated $P_{\text{eff}}S_{\text{mLV}}$ and assuming a solute production rate of 3 g/day (equivalent to
126 a flux of 0.0018 mmol/h), we obtain an interstitial concentration of approximately
127 0.0864 mmol/L. This estimate is exceptionally consistent with the concentration values
128 derived from the gMRI segmentation reported in the main text.

129 B Parameters exploration in a Two Compartment 130 ODE Model

131 We describe the tracer transport between the CSF compartment concentration
132 $C_{\text{CSF}}(t)$ and the Dura compartment concentration in a simplified ODE model in the
133 following way:

$$\frac{d\phi_{\text{CSF}}C_{\text{CSF}}}{dt} = -\alpha(C_{\text{CSF}} - C_D) - \bar{k}_{10}C_{\text{CSF}} + I(t), \quad (17)$$

$$\frac{d\phi_D C_D}{dt} = \alpha(C_{\text{CSF}} - C_D) - \bar{k}_D C_D, \quad (18)$$

135 where $C_{\text{CSF}}(t)$ and $C_D(t)$ are the concentrations in the whole CSF and dural com-
136 partments, respectively, α [h^{-1}] is the influx rate constant (mass transfer coefficient)
137 from CSF to the dura, \bar{k}_{10} [h^{-1}] is the total efflux or clearance rate constant from the
138 CSF, \bar{k}_D [h^{-1}] is the total efflux or clearance rate constant from the dura, $I(t)$ is the
139 injection of gadobutrol in the CSF compartment (that needs to be modelize).

140 Initial conditions were set to $C_{\text{CSF}}(0) = C_D(0) = 0$.

141 Since we are not considering the space variable in this model, it is better to
142 reformulate it in terms of amounts instead of concentrations

143

$$\frac{dA_1}{dt} = -(k_{12} + k_{10}) A_1 + k_{21} \frac{V_{\text{CSF}}}{V_D} A_2 + I(t), \quad (19)$$

$$\frac{dA_2}{dt} = k_{12} \frac{V_D}{V_{\text{CSF}}} A_1 - (k_{21} + k_D) A_2, \quad (20)$$

where A_1 and A_2 are the amounts (in mmol) of gadobutrol in the CSF and dura compartment, respectively;

$$k_{12} = \frac{\alpha}{\phi_{\text{CSF}}}, \quad k_{21} = \frac{\alpha}{\phi_D},$$

$$k_{10} = \frac{\bar{k}_{10}}{\phi_{\text{CSF}}}, \quad k_D = \frac{\bar{k}_D}{\phi_D},$$

and the injection is modeled as the best fit between the following models

$$I(t) = S_{\text{in}} t^{B_{\text{in}}} \exp(-t/C_{\text{in}}), \quad I(t) = S_{\text{in}} (\exp(-t/B_{\text{in}}) - \exp(-t/C_{\text{in}})).$$

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B.1 QMC Analysis

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The search space consisted of seven dimensions representing the unknown parameters of the model: k_{10} (elimination from the CSF), k_{12} (transfer from CSF to Dura), k_{21} (return from Dura to CSF), k_D (clearance), S_{in} (scaling factor), B_{in} (first decay constant), and C_{in} (second decay constant). To reduce the number of dimensions for this analysis, the parameters S_{in} , B_{in} , and C_{in} are fitted against the CSF concentration data and then we vary them of 10% to decrease the uncertainty given by these parameters. Moreover, we apply an optimization method (L-BFGS-B) to the 90% confidence interval found by the Montecarlo simulations to obtain the parameters that best fit the results in that range. To evaluate the goodness-of-fit for each simulated trajectory against the experimental data, a Normalized Sum of Squared Residuals (SSR) was computed.

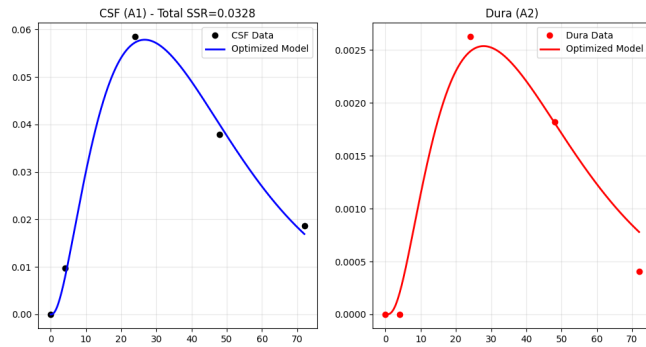


Fig. C: The best fit models of the two compartment (without the 4h data point)

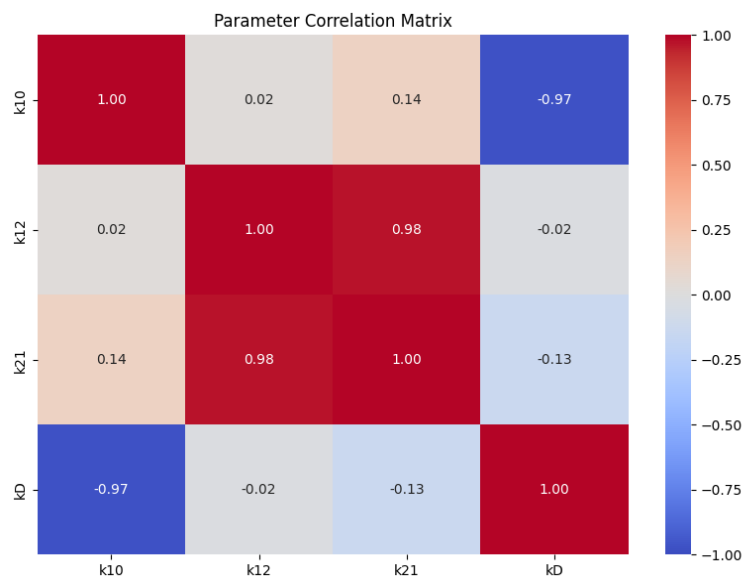


Fig. D: The correlation matrix of the 0D model.

156 The refined two-compartment model, successfully captured the macroscopic tissue
 157 kinetics with a final optimized Sum of Squared Residuals (SSR) of 0.0328 (see Figure
 158 C). Moreover, Figure D shows the correlation matrix between the parameters.

159 The QMC analysis of the model (19) – (20) provided a rigorous assessment of
 160 the practical identifiability of the physiological parameters. The statistical analy-
 161 sis revealed remarkably low Spearman sensitivities to the cost function across all
 162 parameters ($|\rho| < 0.05$). Consequently, the QMC 95% confidence intervals remained
 163 exceptionally broad for both the exchange rates (e.g., $k_{21} \in [0.0001, 77.03] \text{ h}^{-1}$) and
 164 the elimination pathways (e.g., $k_D \in [0.0002, 6.79] \text{ h}^{-1}$). However, this apparent lack
 165 of individual parameter identifiability is fundamentally explained by the presence of
 166 strong compensatory processes within the system, as evidenced by the Pearson corre-
 167 lation matrix. Specifically, we observed a near-perfect negative correlation between the
 168 two elimination pathways (k_{10} and k_D , $r \approx -0.97$), indicating that the model easily
 169 compensates for a lower direct CSF clearance by increasing dural clearance to maintain
 170 the overall systemic elimination rate. Similarly, a highly significant positive correla-
 171 tion was found between the compartment exchange rates (k_{12} and k_{21} , $r \approx 0.98$).
 172 While this strong coupling renders the exact individual magnitudes of k_{12} and k_{21}
 173 difficult to isolate independently, it carries critical physiological significance, but their
 174 ratio k_{12}/k_{21} is identifiable and it corresponds to the physiological parameter ratio
 175 $\eta = \phi_D/\phi_{\text{CSF}}$. Due to the broad parameter results range, $\eta \in [0.18, 244]$; removing the
 176 non-physiological values, we restrict the parameter interval at $\eta \in [0.18, 0.6]$.

177 **C Parameter Exploration in a One-Dimensional One** 178 **Compartment PDE Model**

179 **C.1 Model**

180 The transport of the solute is modeled within a spherically symmetric, single-porosity
 181 domain representing the tissue layer.

182 The geometric domain is defined as a spherical shell bounded by an inner radius
 183 R_1 and an outer radius R_2 . Assuming spherical symmetry, the concentration profile

184 $C(r, t)$ varies only with the radial distance $r \in [R_1, R_2]$ and time t . The values of R_1
 185 and R_2 were found such that $R_2 - R_1 \approx 1$ mm (approximately the thickness of the
 186 dura [7]) and a volume of $V_D \approx 0.0914$ L (found by the segmentation presented in
 187 Section 2.2). With these constrains, we found $R_1 = 0.848$ dm and $R_2 = 0.858$ dm.

188 The solute concentration $C(r, t)$ is governed by a one-dimensional radial reaction-
 189 diffusion equation. The model accounts for Fickian diffusion and a volumetric clearance
 190 (sink) term. The governing partial differential equation (PDE) is given by:

$$\frac{\partial C}{\partial t} = \nabla \cdot (D_D \nabla C) - \frac{K_D}{\phi} C; \quad (21)$$

191 expanded in spherical coordinates, this yields:

$$\frac{\partial C}{\partial t} = \frac{D_D}{r^2} \frac{\partial}{\partial r} \left(r^2 \frac{\partial C}{\partial r} \right) - \frac{K_D}{\phi} C, \quad (22)$$

192 where D_D is the effective diffusion coefficient (dm^2/h) that describes the diffusion
 193 and eventually the dispersion in the whole dura (including the venous and lymphatic
 194 vessels) and K_D is the clearance coefficient (h^{-1}).

195 **Inner Boundary** ($r = R_1$): Transport across the inner surface is dictated
 196 by a permeability coefficient P (dm/h). This is represented as a Robin (mixed)
 197 boundary condition where the diffusive flux matches the permeable flux driven by
 198 the concentration difference between the CSF driver $C_{\text{CSF}}(t)$ and the tissue surface
 199 $C(R_1, t)$:

$$-D_D \frac{\partial C}{\partial r} \Big|_{r=R_1} = \frac{P}{\phi} (\eta C_{\text{CSF}}(t) - C(R_1, t)), \quad (23)$$

where $\eta = \phi_D / \phi_{\text{CSF}}$. The CSF input function $C_{\text{CSF}}(t)$ is defined empirically using
 best fit between the following models

$$I(t) = S_{\text{in}} t^{B_{\text{in}}} \exp(-t/C_{\text{in}}), \quad C_{\text{CSF}}(t) = S_{\text{in}} (\exp(-t/B_{\text{in}}) - \exp(-t/C_{\text{in}})),$$

200 where S_{in} , B_{in} , and C_{in} are fixed parameters derived from prior curve fitting.

201 **Outer Boundary** ($r = R_2$): The outer boundary is assumed to be strictly
202 impermeable, yielding a homogeneous Neumann (no-flux) boundary condition:

$$\left. \frac{\partial C}{\partial r} \right|_{r=R_2} = 0. \quad (24)$$

203 **Initial Condition** ($t = 0$): The tissue is assumed to be initially free of the solute,
204 yielding:

$$C(r, 0) = 0 \quad \text{for} \quad R_1 \leq r \leq R_2 \quad (25)$$

205 To compare the continuous spatial model against bulk empirical data, the observ-
206 able concentration is defined as the volume-weighted average of $C(r, t)$ across the
207 entire spherical shell:

$$\bar{C}(t) = \frac{1}{V_{\text{total}}} \int_{R_1}^{R_2} C(r, t) \cdot 4\pi r^2 dr \quad (26)$$

208 where $V_{\text{total}} = \frac{4}{3}\pi(R_2^3 - R_1^3)$ is the total volume of the domain. This $\bar{C}(t)$ is subse-
209 quently used to compute the objective function (Normalized Root Mean Square Error)
210 during the parameter optimization process. We run QMC to estimate parameters 95
211 % confidence intervals. We can see the results in Table C.

212 C.2 QMC analysis and Results

213 We rely on the QMC method with Halton sequence to estimate the parameters D_D ,
214 K_D/ϕ , and P/ϕ . We choose as parameter range the following (motivated by biology
215 and range found in the 0D model of Section B): $D_D \in [1 \times 10^{-5}, 5 \times 10^{-3}] \text{ dm}^2/\text{h}$
216 $[8-10]$, $P/\phi \in [10^{-4}, 100] \text{ dm}/\text{h}$, $K_D/\phi \in [10^{-4}, 100] \text{ 1}/\text{h}$, and $\eta \in [0.18, 0.6]$.

217 To ensure a comprehensive and physiologically relevant exploration of the 1D
218 diffusion-reaction model, the parameter search space was constrained using insights
219 from the prior 0D compartmental analysis. Specifically, the search boundaries for the

220 effective clearance rate (K_D/ϕ) and the effective boundary permeability (P/ϕ) were
 221 informed directly by the confidence intervals of their macroscopic analogs derived from
 222 the 0D model. Conversely, the limit for the internal diffusion coefficient (D_D) was set
 223 broad enough to allow the numerical solver to freely explore both extreme diffusion-
 224 limited and permeability-limited (well-mixed) transport regimes, but the extrema were
 225 set inspired by biological limit [9–11].

226 The optimal parameter set for the 1D diffusion-reaction model yielded a minimum
 227 Sum of Squared Residuals (SSR) of approximately 0.068. To assess the structural
 228 identifiability and robustness of these estimates, a subset analysis was performed on
 229 the top 250 model fits. The statistical summary of the physical and derived parameters
 230 is presented in Table C.

Table C: Statistical Summary of Estimated Parameters (Top 250 Fits). CV denotes the Coefficient of Variation ($SD/Mean$), serving as a metric for parameter identifiability.

Parameter	Symbol	Mean	Std. Dev.	Median	Rel. IQR	CV (%)
Permeability (dm/h)	P/ϕ	0.0040	0.0015	0.0039	0.703	38.6%
Partition Coeff. (-)	η	0.3186	0.1290	0.2800	0.751	40.5%
Clearance Rate (h^{-1})	K_D/ϕ	0.2129	0.1363	0.2175	1.173	64.1%
Diffusion Coeff. (dm^2/h)	D_D	1.14×10^{-3}	1.26×10^{-3}	5.29×10^{-4}	3.025	110.8%

Table D: Statistical Summary of Estimated Parameters (Top 250 Fits). CV denotes the Coefficient of Variation ($SD/Mean$). Results refer to the single-compartment model with fixed $\eta = 0.3186$.

Parameter	Symbol	Mean	Std. Dev.	Median	Rel. IQR	CV (%)
Clearance Rate (h^{-1})	K_D/ϕ	0.2466	0.0169	0.2463	0.108	6.8%
Permeability (dm/h)	P/ϕ	0.0037	0.0003	0.0037	0.133	9.4%
Diffusion Coeff. (dm^2/h)	D_D	1.16×10^{-3}	1.29×10^{-3}	6.03×10^{-4}	2.617	110.7%

231 The correlation matrix of Figure E reveals high correlation between the parameters
 232 P , η , and K_D . The correlation between P and η is given by the product $P\eta$ at the

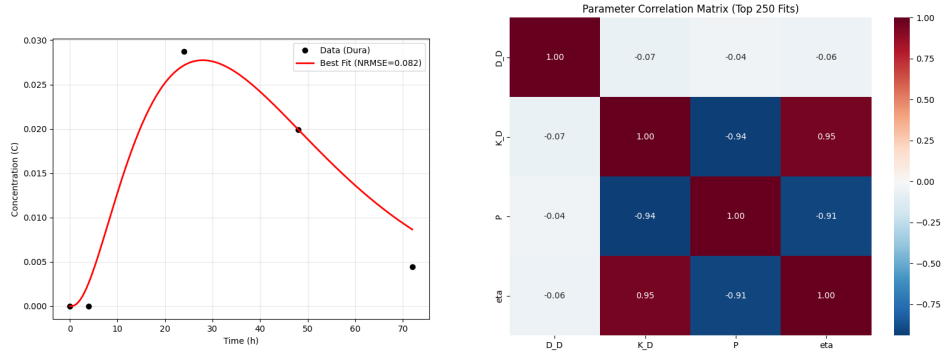


Fig. E: **Left:** The best fit models of the 1 compartment model on a sphere. **Right:** The correlation matrix of the spherical 1 compartment model.

233 boundary condition. By fixing the partition coefficient at its mean value ($\eta = 0.3186$),
 234 we successfully decoupled the boundary dynamics, yielding high identifiability for the
 235 clearance rate (K_D) and permeability (P), which exhibit low Coefficients of Variation
 236 (6.8% and 9.4%, respectively, see Table D). This stability confirms that the previously
 237 observed uncertainties were primarily numerical artifacts of the P - η correlation.

238 The parameter correlation matrix highlights a strong positive coupling between K_D
 239 and P ($r = 0.738$). This relationship elucidates a fundamental physiological trade-off:
 240 higher dural permeability, which increases tracer influx, must be balanced by a pro-
 241 portionally higher clearance rate to accurately replicate the washout decay observed
 242 in clinical data. This compensatory mechanism ensures that the model preserves the
 243 mass balance and the observed peak timing, regardless of the individual parameter
 244 magnitudes.

245 The internal diffusion estimation $D_D \approx 1.16 \times 10^{-3} \text{ dm}^2/\text{h}$ presents high uncer-
 246 tainty ($CV > 100\%$), but it offers a value higher than the gadobutrol diffusion in free
 247 water of $\approx 1.37 \times 10^{-4} \text{ dm}^2/\text{h}$, suggesting that the only-diffusion regime is not enough.

248 **D Parameter Exploration in a One-Dimensional Two**
 249 **Compartment PDE Model**

250 **D.1 Model**

251 The transport of the solute is modeled within a spherically symmetric, double-porosity
 252 domain representing the tissue layer. The geometric domain is defined as a spherical
 253 shell bounded by an inner radius R_1 and an outer radius R_2 .

254 To account for the complex microstructure of the tissue, the continuum is divided
 255 into two distinct, overlapping interacting compartments: the Dura (D) and the Lymph
 256 (L). The concentration profiles for both compartments, $C_D(r, t)$ and $C_L(r, t)$, vary
 257 with the radial distance $r \in [R_1, R_2]$ and time t .

258 In this model, we suppose that the entire clearance from the dura is given by
 259 the mLVs direct towards the cervical lymphatic vessels. We suppose this to test if
 260 it is possible to have a full clearance given by the lymphatics and to estimate the
 261 parameters which make this possible.

262 The spatio-temporal evolution of the solute is governed by a coupled system of
 263 one-dimensional radial reaction-diffusion equations.

264 For the primary tissue compartment (Dura, C_D), transport is driven by Fickian
 265 diffusion and mass exchange with the secondary compartment:

$$\frac{\partial C_D}{\partial t} = \frac{D_D}{r^2} \frac{\partial}{\partial r} \left(r^2 \frac{\partial C_D}{\partial r} \right) + \beta(\phi_R C_L - C_D), \quad (27)$$

266 For the secondary compartment (Lymph, C_L), transport involves diffusion, the
 267 reverse mass exchange, and a volumetric clearance (elimination) term:

$$\frac{\partial C_L}{\partial t} = \frac{D_L}{r^2} \frac{\partial}{\partial r} \left(r^2 \frac{\partial C_L}{\partial r} \right) - \beta(\phi_R C_L - C_D) - k_{\text{elim}} C_L \quad (28)$$

268 where D_D and D_L are the effective diffusion coefficients for the Dura and Lymph
 269 compartments, respectively (dm^2/h), $\beta = \bar{\beta}/\phi_D$ is the exchange rate between the two

270 compartments (h^{-1}), $\phi_R = \phi_D/\phi_L$, $\phi = \phi_D + \phi_L$ and ϕ_D takes into consideration
 271 the porosity of the dura matrix and the venous vessels inside the dura, and $k_{\text{elim}} =$
 272 $\bar{k}_{\text{elim}}/\phi_L$ is the lymphatic clearance coefficient given by cervical lymphatic vessels
 273 (h^{-1}).

274 **Inner Boundary ($r = R_1$):** Transport from the CSF into the Dura compartment
 275 is dictated by a permeability coefficient $P = \bar{P}/\phi_D$ (dm/h). A scaling factor $\eta_D =$
 276 ϕ_D/ϕ_{CSF} is applied to the CSF driving function. The Lymph compartment is assumed
 277 to have no direct exchange with the CSF at this boundary.

$$-D_D \frac{\partial C_D}{\partial r} \Big|_{r=R_1} = P(\eta C_{\text{CSF}}(t) - C_D(R_1, t)), \quad (29)$$

$$-D_L \frac{\partial C_L}{\partial r} \Big|_{r=R_1} = 0. \quad (30)$$

278 The CSF input function $C_{\text{CSF}}(t)$ is defined empirically using a bi-exponential
 279 model:

$$C_{\text{CSF}}(t) = St^B t e^{-t/C_{\text{rate}}} \quad (31)$$

280 where S , B , and C_{rate} are fixed parameters derived from prior data fitting.

281 **Outer Boundary ($r = R_2$):** The outer boundary is assumed to be strictly imper-
 282 meable for both compartments, yielding homogeneous Neumann (no-flux) boundary
 283 conditions:

$$\frac{\partial C_D}{\partial r} \Big|_{r=R_2} = 0, \quad \frac{\partial C_L}{\partial r} \Big|_{r=R_2} = 0 \quad (32)$$

284 **Initial Condition ($t = 0$):** Both compartments are assumed to be initially free
 285 of the solute:

$$C_D(r, 0) = 0 \quad \text{and} \quad C_L(r, 0) = 0 \quad \text{for} \quad R_1 \leq r \leq R_2 \quad (33)$$

286 To evaluate the mathematical model against bulk tissue empirical data, the observ-
 287 able concentration is defined as the sum of the volume-weighted averages of both
 288 compartments across the entire spherical shell:

$$\bar{C}_{\text{Total}}(t) = \frac{1}{V_{\text{total}}} \int_{R_1}^{R_2} (C_D(r, t) + C_L(r, t)) \cdot 4\pi r^2 dr \quad (34)$$

289 where $V_{\text{total}} = \frac{4}{3}\pi(R_2^3 - R_1^3)$ is the total volume of the domain. This aggregate
 290 $\bar{C}_{\text{Total}}(t)$ is utilized to compute the objective function (Normalized Root Mean Square
 291 Error) during Montecarlo simulations. We run QMC to estimate parameters 95 %
 292 confidence intervals. We can see the results in Table ?? and the details about the
 293 QMC sampling in Supplementary D.

294 D.2 QMC Analysis

295 The search space consisted of six dimensions representing the unknown parameters
 296 of the model: the diffusion coefficient of the dura phase D_D (dm^2/h), the dura-
 297 lymphatic exchange β/ϕ_D ($1/\text{h}$), the porosity ratio ϕ_R , the diffusion coefficient of the
 298 lymph phase D_L (dm^2/h), the volumetric sink k_{elim}/ϕ_L ($1/\text{h}$), and the permeability
 299 between the dura and the CSF P/ϕ_D (dm/h). To have a parameter range that takes
 300 into account the 1 dimensional 1 compartment model, the following relationship are
 301 considered. First, we have that $\frac{k_{\text{elim}}}{\phi_L} = \frac{K_D}{\phi}(\phi_R + 1)$.

302 For the diffusion coefficients D_D and D_L , from the fact that the 1-compartment
 303 diffusion coefficient $D_D^{1\text{comp}} = \frac{\phi_D + \phi_L \gamma_L}{\phi_D + \phi_L} D$ where γ_L represents the enhancement dif-
 304 fusion given by the convection in the lymphatics, we use the following relationship:
 305 $D_L = D_D^{1\text{comp}}(\phi_R + 1) - \phi_R D_D$, where $D_D \in [2 \times 10^{-5}, 5 \times 10^{-5}] \text{ dm}^2/\text{h}$ adapted
 306 from [12]; $\frac{k_{\text{elim}}}{\phi_L} \in [0.1(\phi_R + 1), 1(\phi_R + 1)]$, $\phi_R = \phi_D/\phi_L$ supposing a bound for ϕ_D
 307 similar to the gray matter ($\approx 0.1 - 0.3$) and for $\phi_L \in [0.005, 0.07]$, $\phi_R \in [2, 60]$, from

308 the (wider) 1-compartment model we have $P/\phi \in [0.001, 0.1]$, in our case we need
 309 $\frac{P}{\phi_D} = \frac{P}{\phi} \left(1 + \frac{1}{\phi_R}\right)$, the $\eta_D = \frac{\phi_D}{\phi_{CSF}} = \eta \left(\frac{\phi_R}{\phi_R+1}\right)$ where $\eta = \frac{\phi_D}{\phi_{CSF}}$ is given in the 1-
 310 compartment model as $\eta = 0.31$, and $\beta/\phi_D \in [0.5, 3]$ from Supplementary A and the
 311 range for ϕ_D .

312 The optimal parameter set for this double-porosity model yielded a minimum Sum
 313 of Squared Residuals (SSR) of approximately 0.082. To assess structural identifiability
 314 and the robustness of these constrained estimates, a subset analysis was performed on
 315 the top 250 model fits (the upper 5% convergence region). The statistical summary of
 316 the macroscopic and derived parameters is presented in Table E.

317 The estimated lymphatic diffusion coefficient ($D_L \approx 1.69 \times 10^{-2} \text{ dm}^2/\text{h}$) is approx-
 318 imately two orders of magnitude higher than the established free-water diffusion
 319 coefficient of Gadobutrol ($1.37 \times 10^{-4} \text{ dm}^2/\text{h}$). Such an unphysiologically high diffusiv-
 320 ity suggests that the QMC algorithm is mathematically forcing the diffusion term to
 321 compensate for macroscopic advective transport, which is inherently absent from the
 322 current governing equations. This phenomenon strongly indicates that a pure diffusion
 323 model is insufficient to fully capture the complex, rapid transport dynamics within the
 324 dural lymphatic and vascular networks, pointing to advection-driven flow as a critical
 325 mechanism that must be accounted for to accurately describe dural clearance.

326 To further validate whether this behavior was an artifact of boundary conditions,
 327 we tested the model using a different partition coefficient ($\eta = 0.18$). As anticipated
 328 from the parameter coupling observed in the single-compartment analysis presented in
 329 Supplementary C, modifying η induced expected proportional shifts in the boundary
 330 permeability (P) and elimination (k_{elim}) parameters to preserve the overall mass
 331 balance. Crucially, however, the artificially high magnitude of D_L remained consistent.
 332 This persistence confirms that the inflated D_L is not a numerical artifact driven by
 333 boundary correlations, but rather a fundamental structural limitation.

Table E: Statistical Summary of Estimated Parameters for the 2-Compartment Model (Top 250 Fits). *CV* denotes the Coefficient of Variation ($SD/Mean$), serving as a metric for parameter identifiability.

Parameter	Symbol	Mean	Std. Dev.	Median	Rel. IQR	CV (%)
Permeability (dm/h)	P	0.0045	0.0003	0.0045	0.057	6.1%
Primary Diffusion (dm ² /h)	D_D	4.50×10^{-5}	3.00×10^{-6}	4.60×10^{-5}	0.105	7.7%
Exchange Rate (h ⁻¹)	β	2.25	0.51	2.29	0.334	22.6%
Lymphatic Diffusion (dm ² /h)	D_L	1.69×10^{-2}	7.87×10^{-3}	1.76×10^{-2}	0.752	46.6%
Elimination Rate (h ⁻¹)	k_{elim}	9.51	4.48	10.01	0.733	47.1%
Porosity Ratio (-)	ϕ	33.79	16.24	35.37	0.764	48.1%

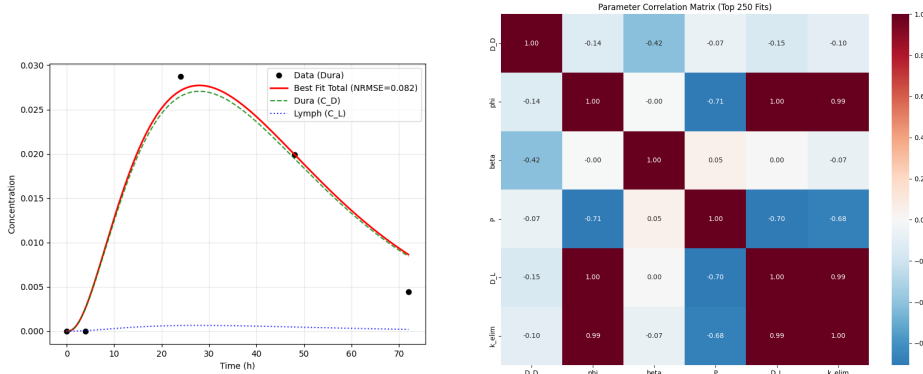


Fig. F: Left: The best fit models of the 2 compartment model on a sphere. Right: The correlation matrix of the spherical 2 compartment model.

334 E Dura and Lymphatic Permeabilities

Using the explicit formula for the permeability $K = \frac{Q\Delta x}{A\Delta p}$, we have that that $A = V/\Delta x$, using the $V = 9.489 \times 10^{-5} \text{ m}^3$ and $\Delta x = 0.001 \text{ m}$, assuming a flux of $Q = 0.5 \text{ L/day} \approx 5.8 \times 10^{-9} \text{ m}^3/\text{s}$, we have the relationship

$$K_D = \mu K = \frac{4.28 \times 10^{-14}}{\Delta P}.$$

335 Using the data estimated in Supplementary A, we have an estimated permeability
 336 value of $K_D \approx 5.2 \times 10^{-17} \text{ m}^2$.

337 Using the same approach, we can estimate the permeability of the mLVs as well.
 338 For the mLVs we need to consider that the flow goes in the "polar" direction, hence
 339 we have that as characteristic length we use an average value of $L \approx 10$ cm; Moreover,
 340 we assume that the pressure at the upper part of the brain is $\approx 1.4 \times 10^3$ Pa [13] and
 341 is 0 Pa in the exit region of the brain (cervical lymphatic vessels) [14, 15]; these lead to
 342 a permeability of $K_L \approx 2.39 \times 10^{-13}$ m², smaller of one order of magnitude to the one
 343 found using an homogenization modeling approach in dermal lymphatic capillaries in
 344 [16].

345 F mLV Volume

We assume that the meningeal lymphatic vessels absorb all the fluid produced by the brain over one day ($Q_{\text{prod}} \approx 0.5$ L/day $\approx 5.79 \times 10^{-9}$ m³/s). Furthermore, the average velocity in the basal mLVs is $\bar{v} \approx 2.2 - 2.7$ mm/s [17]. Using these data and assuming a spherical brain, we can estimate the porosity of the mLVs to be

$$\phi_L \approx \frac{Q_{\text{prod}}}{\bar{v}A_{\perp}} \approx 0.005 - 0.015 = 0.5\% - 1.5\%.$$

346 This value is lower respect to the porosity found of the dermal lymphatic vessels
 347 [16] ($\approx 5 - 20\%$). It follows that the mLVs volume is $V_{\text{mLV}} \approx 4.57 \times 10^{-7} - 1.37 \times$
 348 10^{-6} m³. Assuming a spherical shell geometry for the dura, the cross-sectional area A_{\perp}
 349 perpendicular to the polar lymph flow is given by $A_{\perp} = V_{\text{dura}}/L$, where L represents
 350 the characteristic path length of the mLV (in polar direction, way through the cervical
 351 lymphatic vessels), and V_{dura} is the total volume.

352 In [4] they measured the volume of the meningeal lymphatic vessels and the venous
 353 vessels inside the dura. We have that the average volume of the mLV is $V_{\text{mLV}} \approx$
 354 6×10^{-6} m³, which means a porosity of the mLV of $\phi_L \approx 0.063$. In [4] they did not
 355 measure the volume of the smaller mLVs found in [5], that represents a very small

356 fraction of the mLVs. Hence we can estimate that the porosity of the mLVs to be
 357 between 0.065 – 0.07. Due to the uncertainty of these values, in this work we will
 358 consider a range for the porosity of the mLVs between 0.5 – 7%.

359 G Structural Identifiability and the Role of Local 360 Dural Clearance (k_D)

361 To evaluate the impact of alternative drainage pathways, an extended formulation of
 362 the model was tested, incorporating a non-specific dural clearance parameter (k_D)
 363 adding the term $-\frac{k_D}{\phi_D}C_{D,i}$ to equation (2) of the main text. This parameter simulates
 364 the local reabsorption of the tracer in the dura mater without entering the lym-
 365 phatic system, thereby acting in competition with the macroscopic lymphatic drainage
 366 towards the deep cervical lymph nodes (P_{DcLV}).

367 The Quasi-Monte Carlo (QMC) simulations of the extended model yielded an
 368 excellent experimental fit, with an optimal loss function (SSR = 0.055) identical to
 369 that of the base model presented in the main text. The ensemble statistics for this
 370 competitive scenario are detailed in Table F.

Table F: Optimal parameters and derived quantities (Top 200 QMC ensemble) for the extended model including local dural clearance (k_D).

Parameter	Unit	Median	IQR [25%, 75%]	95% CI
<i>Model Parameters</i>				
Partition Coeff. (ϕ)	[-]	7.69	[3.74, 14.57]	[2.26, 33.58]
CSF Permeability (P_{bot})	[h ⁻¹]	0.434	[0.351, 0.557]	[0.239, 0.829]
Dispersion Coeff. (D_L)	[-]	0.0006	[0.0001, 0.0020]	[0.00001, 0.0043]
Deep Clearance (P_{DcLV})	[h ⁻¹]	5.84	[0.085, 22.65]	[0.001, 231.54]
Dural Clearance (k_D)	[h ⁻¹]	1.04	[0.086, 1.55]	[0.0002, 2.24]
Lymphatic Porosity (ϕ_L)	[-]	0.026	[0.013, 0.042]	[0.006, 0.066]
<i>Derived Quantities</i>				
Péclet Number (Pe)	[-]	54.22	[16.52, 513.74]	[2.92, 15855.82]
Biot Number (Bi)	[-]	177.21	[143.23, 227.44]	[70.28, 458.45]
Advection Time (T_{adv})	[h]	59.09	[28.95, 94.69]	[11.75, 154.43]
Diffusion Time (T_{diff})	[h]	3419.89	[999.64, 23042.68]	[430.72, 201782.79]

371 **G.1 Discussion of Results and Equifinality**

372 The inclusion of k_D significantly alters the hydrodynamic balance of the system. By
373 allowing the dura mater to act as a local clearance pathway (with an estimated half-life
374 of approximately 6.3 hours, $k_D \approx 0.11 \text{ h}^{-1}$), the burden on the lymphatic system is
375 drastically reduced. Consequently, the model predicts a slower advection time ($T_{adv} \approx$
376 46.7 h) and a markedly reduced basal clearance P_{DcLV} . Nevertheless, the transport
377 regime within the lymphatic vessels remains undeniably dominated by convection, as
378 confirmed by a median Péclet number firmly greater than unity ($Pe \approx 54.22$).

379 Despite the physiological plausibility of this extended scenario, the simultaneous
380 inclusion of k_D and P_{DcLV} introduces a well-known mathematical issue of *structural*
381 *identifiability* (equifinality). Because contrast-enhanced MRI captures the global signal
382 decay within a voxel but lacks the resolution to differentiate the specific molecular exit
383 route or distinguish between interstitial or lymphatic enrichment, the QMC analysis
384 can seamlessly compensate for an increase in one clearance parameter by decreasing
385 the other, without altering the Sum of Squared Residuals (SSR).

386 **G.2 Justification for the Base Model**

387 In light of this, the model imposing $k_D = 0$ was adopted for the main text. Through
388 this choice, the meningeal lymphatic network is forced to handle 100% of the tracer
389 clearance.

390 This approach allows us to test the theoretical maximum capacity of the lymphatic
391 network under a "worst-case scenario" approach. The results presented in the main
392 text demonstrate that, even under this extreme load, the lymphatic system is physi-
393 cally capable of clearing the entire tracer volume using geometrically, physiologically,
394 and anatomically realistic parameters. Therefore, the base model provides robust,
395 independent computational validation of the clinical hypothesis that the meningeal

396 lymphatic network possesses sufficient hydraulic capacity to be the primary driver of
397 brain clearance.

398 Data Availability

399 All the code and setup used in this supplementary material is available upon request.

400 References

- 401 [1] Mendoza, E., Schmid-Schönbein, G.W.: A model for mechanics of primary lym-
402 phatic valves. *Journal of Biomechanical Engineering* **125**(3), 407–414 (2003)
403 <https://doi.org/10.1115/1.1568128>
- 404 [2] Koch, T., Vinje, V., Mardal, K.: Estimates of the permeability of extra-cellular
405 pathways through the astrocyte endfoot sheath. *Fluids and Barriers of the CNS*
406 **20**(20) (2023) <https://doi.org/10.1186/s12987-023-00421-8>
- 407 [3] Pardridge, W.M.: Csf, blood-brain barrier, and brain drug delivery. *Expert*
408 *Opin Drug Deliv* **13**(7), 963–975 (2016) [https://doi.org/10.1517/17425247.2016.](https://doi.org/10.1517/17425247.2016.1171315)
409 [1171315](https://doi.org/10.1517/17425247.2016.1171315)
- 410 [4] Jacob, L., Brito Neto, J., Lenck, S., Corcy, C., Benbelkacem, F., Geraldo, L.H.,
411 Xu, Y., Thomas, J.M., El Kamouh, M.R., Spajer, M., Potier, M.C., Haik, S.,
412 Kalamarides, M., Stankoff, B., Lehericy, S., Eichmann, A., Thomas, J.L.: Con-
413 served meningeal lymphatic drainage circuits in mice and humans. *The Journal*
414 *of experimental medicine* **219** (2022) <https://doi.org/10.1084/jem.20220891>
- 415 [5] Vera Quesada, C.L., Rao, S.B., Torp, R., Eide, P.K.: Widespread distribution of
416 lymphatic vessels in human dura mater remote from sinus veins. *Frontiers in Cell*
417 *and Developmental Biology* **Volume 11 - 2023** (2023) [https://doi.org/10.3389/](https://doi.org/10.3389/fcell.2023.1228344)
418 [fcell.2023.1228344](https://doi.org/10.3389/fcell.2023.1228344)

- 419 [6] Çavdar, S., Köse, B., Altınöz, D., Söyler, G., Cingöz, A., Gürses, .I.A., Özkan,
420 M., Aşilyüksek, H., Çakır, H.: Lymphatic vessels accompanying dorsal and basal
421 dural sinuses in the human brain. *Journal of Chemical Neuroanatomy* **134**, 102357
422 (2023) <https://doi.org/10.1016/j.jchemneu.2023.102357>
- 423 [7] Protasoni, M., Sangiorgi, S., Cividini, A., Culivaris, G.T., Tomei, G., Dell’Orbo,
424 C., Raspanti, M., Balbi, S., Reguzzoni, M.: The collagenic architecture of human
425 dura mater. *J Neurosurg* **114**, 1723–1730 (2011) [https://doi.org/10.3171/2010.](https://doi.org/10.3171/2010.12.JNS101732)
426 [12.JNS101732](https://doi.org/10.3171/2010.12.JNS101732)
- 427 [8] Riseth, J.N., Koch, T., Lian, S.L., Holck Storås, T., Zikatanov, L.T., Valnes,
428 L.M., Nordengen, K., Mardal, K.-A.: Human brain MRI data of intrathecally
429 injected tracer evolution over 72 hours for data-integrated simulations. *Scientific*
430 *Data* (2026)
- 431 [9] Riseth, J.N., Koch, T., Mardal, K.-A.: Two-compartment modeling of tracer
432 transport in the brain. In: Dokken, J.S., Mardal, K.-A., Rognes, M.E., Valnes,
433 L.M., Vinje, V. (eds.) *Mathematical Modelling of the Human Brain II: From*
434 *Glymphatics to Deep Learning*, pp. 105–122. Springer, ??? (2025)
- 435 [10] Hornkjøl, M., Valnes, L.M., Dokken, J.S.: Segmenting, meshing, and modeling
436 CSF spaces. In: Dokken, J.S., Mardal, K.-A., Rognes, M.E., Valnes, L.M., Vinje,
437 V. (eds.) *Mathematical Modelling of the Human Brain II: From Glymphatics to*
438 *Deep Learning. Simula SpringerBriefs on Computing*, vol. 18, pp. 35–46. Springer,
439 Cham (2026). [https://doi.org/10.1007/978-3-032-00679-0.3](https://doi.org/10.1007/978-3-032-00679-0_3)
- 440 [11] Riseth, J., Koch, T., Lian, S., Storås, T., Zikatanov, L.T., Valnes, L.M., Norden-
441 gen, K., Mardal, K.-A.: The Gonzo Dataset: Human Brain MRI Data of CSF
442 Tracer Evolution Over 72h For Data-Integrated Simulations. [https://doi.org/10.](https://doi.org/10.5281/zenodo.14266867)
443 [5281/zenodo.14266867](https://doi.org/10.5281/zenodo.14266867) . <https://doi.org/10.5281/zenodo.14266867>

- 444 [12] Bashkatov, A.N., Genina, E.A., Sinichkin, Y.P., Kochubey, V.I., Lakodina, N.A.,
445 Tuchin, V.V.: Glucose and mannitol diffusion in human dura mater. *Biophys-*
446 *ical Journal* **85**(5), 3310–3318 (2003) [https://doi.org/10.1016/S0006-3495\(03\)](https://doi.org/10.1016/S0006-3495(03)74750-X)
447 [74750-X](https://doi.org/10.1016/S0006-3495(03)74750-X)
- 448 [13] Vinje, V., Eklund, A., Mardal, K., Rognes, M., Støverud, K.: Intracranial pressure
449 elevation alters csf clearance pathways. *Fluids and Barriers of the CNS* **17** (2020)
450 <https://doi.org/10.1186/s12987-020-00189-1>
- 451 [14] Kim, D., Tithof, J.: Lumped parameter simulations of cervical lymphatic vessels:
452 dynamics of murine cerebrospinal fluid efflux from the skull. *Fluids and Barriers*
453 *of the CNS* **21** (2024) <https://doi.org/10.1186/s12987-024-00605-w>
- 454 [15] Kim, D., Tithof, J.: Fluid-structure interaction dynamics
455 of cervical lymphatic vessel pumping and valvular func-
456 tion. *bioRxiv* (2025) <https://doi.org/10.1101/2025.11.21.689846>
457 <https://www.biorxiv.org/content/early/2025/11/25/2025.11.21.689846.full.pdf>
- 458 [16] Roose, T., Swartz, M.A.: Multiscale modeling of lymphatic drainage from tis-
459 sues using homogenization theory. *Journal of Biomechanics* **45**(1), 107–115 (2012)
460 <https://doi.org/10.1016/j.jbiomech.2011.09.015>
- 461 [17] Kim, J., Yoo, R., Choi, S.H., Park, S.: Non-invasive flow mapping of parasagittal
462 meningeal lymphatics using 2d interslice flow saturation MRI. *Fluids and Barriers*
463 *of the CNS* **20** (2023) <https://doi.org/10.1186/s12987-023-00446-z>