

## **Supplementary Material**

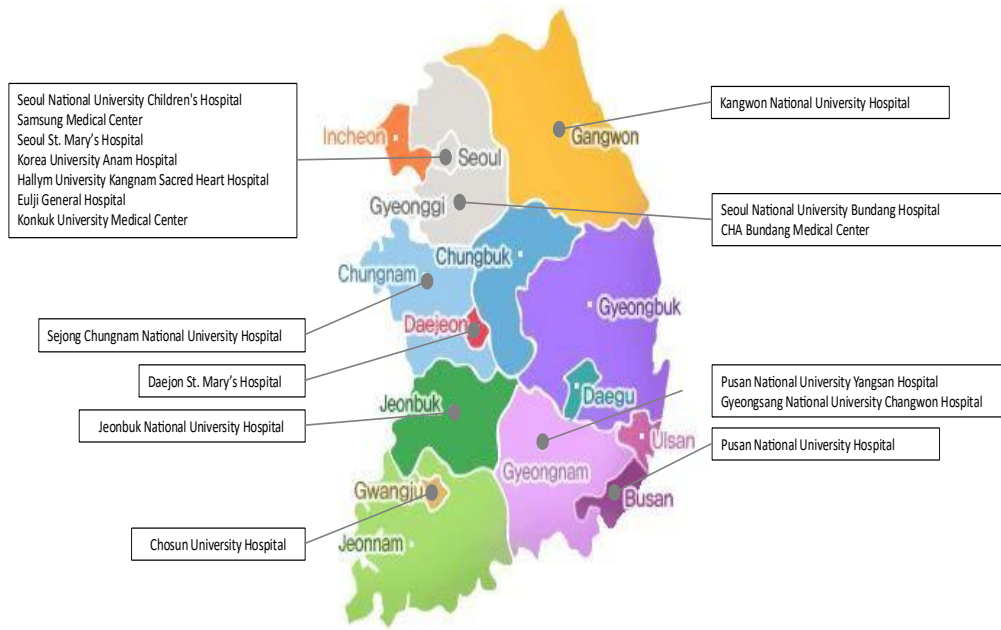
Supplementary Fig.S1. Distribution of 17 participating hospitals throughout South Korea

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**Supplementary Fig. S1. Distribution of 17 participating hospitals throughout South Korea**

**Supplementary Table S1. Categories and Variables collected using the electronic case report form (eCRF)**

Category	Variables collected
Demographics	Age, sex
Hospitalization	Hospital admission status, admission and discharge dates
Clinical presentation	Symptom onset date, fever, cough, sputum, rhinorrhea, sore throat, chest pain, tachypnea, dyspnea, gastrointestinal symptoms, rash, seizure
Vital signs and physical examination	Highest body temperature, highest respiratory rate, lowest oxygen saturation, auscultatory findings, chest retraction
Radiologic findings	Chest radiograph patterns including peribronchial infiltration, consolidation, pleural effusion, cavity or pneumatocele
Underlying medical conditions	Prematurity, congenital heart disease, chronic neurologic, renal, gastrointestinal, endocrine, or pulmonary disease, asthma or allergic disease, genetic disease or congenital anomaly, primary immunodeficiency, hematologic or oncologic disease, and history of transplantation
Microbiologic testing	Respiratory viral molecular testing, bacterial and atypical pathogen PCR assays, Mycoplasma pneumoniae serologic testing, bacterial cultures
Laboratory findings	White blood cell count, C-reactive protein, procalcitonin
Treatment	Antibiotic therapy, antiviral therapy, systemic corticosteroid use for $\geq 3$ days
Respiratory support	the use of respiratory support [either noninvasive (nasal prong, oxygen mask, high-flow nasal cannula, or noninvasive positive pressure ventilation) or invasive respiratory support (invasive mechanical ventilation or extracorporeal membrane oxygenation)].
Clinical outcomes	Intensive care unit admission, complications, mortality, length of hospital stay, outcome at last follow-up

**Supplementary Table S2. Diagnostic platforms and *Mycoplasma pneumoniae* serologic assays used across participating hospitals**

Location (Province)	Hospital	Respiratory virus PCR	Respiratory virus antigen test	Atypical PCR <sup>a</sup>	<i>Mycoplasma pneumoniae</i> serology <sup>b</sup>	Bacterial PCR <sup>c</sup>
Seoul	Seoul National University Children's Hospital	Allplex™ Respiratory Panel 1/2/3 (Seegene, Seoul, Korea)	BD Veritor System Flu A+B (Becton, Dickinson and Company, Franklin Lakes, NJ, USA), Ecoli Dx (Amplisens, Prague, Czech Republic) (Wonmed, Gyeonggi, Korea), BinaxNOW RSV Card (Abbott, Scarborough, ME, USA)		IgM LIAISON (DiaSorin), PA Serodia-Myc II (Fujirebio)	
	Samsung Medical Center	BIOFIRE® Respiratory 2.1 plus Panel (bioMérieux, Marcy-l'Étoile, France)	Not performed		<i>M. pneumoniae</i> PCR (SCL, Yongin, Korea); BIOFIRE® Respiratory 2.1 Panel (bioMérieux, Marcy-l'Étoile, France)	PA Serodia-Myc II Not performed
	Seoul St. Mary's Hospital	AdvanSure™ RV-plus Real-Time RT-PCR (LG Chem, Seoul, Korea), BIOFIRE® Respiratory 2.1 Panel (bioMérieux, Marcy-l'Étoile, France)	Wonmed Covid-19 Ag (Wonmed, Gyeonggi, Korea)	BioCore <i>M. pneumoniae</i> Real-Time PCR Kit (Invites BioCore, Seoul, Korea)	<i>M. pneumoniae</i> IgM CHORUS (Diesse), IgG LIAISON (DiaSorin)	
	Korea University Anam Hospital	Allplex™ Respiratory Panel 1/2/3 (Seegene, Seoul, Korea)	BD Veritor System Flu A+B (Becton, Dickinson and Company, Franklin Lakes, NJ, USA)	<i>M. pneumoniae</i> PCR (GCLabs, Yongin, Korea)	IgM LIAISON (DiaSorin)	

Location (Province)	Hospital	Respiratory virus PCR	Respiratory antigen test	virus Atypical PCR <sup>a</sup>	<i>Mycoplasma</i> serology <sup>b</sup>	Bacterial PCR <sup>c</sup>
		Korea)	Dickinson Company, Lakes, NJ, USA)	and Korea) Franklin		
	Hallym University Kangnam Sacred Heart Hospital	AllplexRespiratory 1/2/3 (Seegene, Korea), BIOFIRE® Respiratory 2.1 Plus Panel (bioMérieux, Marcy-l'Étoile, France)	BD Veritor System Flu A+B Dickinson Company, Lakes, NJ, USA)	(Becton, and Franklin Korea)	Allplex PneumoBacter Assay IgM LIAISON (DiaSorin) Seoul, Korea)	Allplex™ PneumoBacter Assay (Seegene, Seoul, Korea)
	Eulji General Hospital	AllplexRespiratory 1/2/3 (Seegene, Korea)	BD Veritor System Flu A+B Dickinson Company, Lakes, NJ, USA)	(Becton, and Franklin Yongin, Korea)	Bacterial Multiplex PCR (GCLabs, Diesse)	Bacterial Multiplex PCR (GCLabs, Yongin, Korea)
	Konkuk University Medical Center	BIOFIRE® Respiratory 2.1 Plus Panel (bioMérieux, Marcy-l'Étoile, France)	Not performed	BIOFIRE Respiratory 2.1 Plus Panel (bioMérieux, Marcy-l'Étoile, France)	IgM CHORUS (Diesse)	Not performed
Gyeonggi-do	CHA Bundang Medical Center	BIOFIRE® Respiratory 2.1 Panel (bioMérieux, Marcy-l'Étoile, France)	BD Veritor System Flu A+B Dickinson Company, Lakes, NJ, USA), STANDARD Q COVID-19 Ag Test	(Becton, and Franklin l'Étoile, France)	IgM LIAISON Respiratory 2.1 Panel (DiaSorin), CHORUS (Diesse)	Not performed

Location (Province)	Hospital	Respiratory virus PCR	Respiratory antigen test	virus	Atypical PCR <sup>a</sup>	<i>Mycoplasma</i> serology <sup>b</sup>	Bacterial PCR <sup>c</sup>
			(SD Biosensor, Suwon, Korea)				
	Seoul National University Bundang Hospital	Allplex Respiratory 1/2/3 (Seegene, Korea)	Panel (Seegene, Seoul, Korea)	Standard F Influenza A/B FIA (SD Biosensor, Seoul, Korea), Standard F Ag FIA (SD Biosensor, Seoul, Korea), Standard SARS-CoV-2 (SD Biosensor, Seoul, Korea)	NxTAG (Luminex Corporation, Austin, USA); Allplex™ PneumoBacter Assay (Seegene, Seoul, Korea)	RPP IgM R-FIND (SG Medical)	Not performed
<b>Daejeon</b>	Daejeon St. Mary's Hospital	AllplexRespiratory 1/2/3 (Seegene, Korea)	Panel (Seegene, Seoul, Korea)	Not performed	Allplex PneumoBacter Assay (Seegene, Seoul, Korea)	IgM CHORUS (Diesse)	Allplex PneumoBacter Assay (Seegene, Seoul, Korea)
<b>Sejong</b>	Sejong Chungnam National University Hospital	BIOFIRE Respiratory 2.1 Panel (bioMérieux, l'Étoile, France)	Panel (bioMérieux, Marcy-l'Étoile, France)	Not performed	BIOFIRE Respiratory 2.1 Panel (bioMérieux, Marcy-l'Étoile, France)	IgM LIAISON (DiaSorin)	Allplex PneumoBacter Assay (Seegene, Seoul, Korea)
<b>Jeonbuk</b>	Jeonbuk National University Hospital	BIOFIRE Respiratory 2.1 Panel (bioMérieux, l'Étoile, France)	Panel (bioMérieux, Marcy-l'Étoile, France)	Not performed	BIOFIRE Respiratory 2.1 Panel (bioMérieux, Marcy-l'Étoile, France)	IgM LIAISON (DiaSorin)	Not performed
<b>Gyeongnam</b>	Pusan National University	BIOFIRE Respiratory 2.1	QuantumPACK	Easy Bacterial	Multiplex IgM	LIAISON	Bacterial

Location (Province)	Hospital	Respiratory virus PCR	Respiratory antigen test	virus	Atypical PCR <sup>a</sup>	<i>Mycoplasma</i> serology <sup>b</sup>	Bacterial PCR <sup>c</sup>
	Yangsan Hospital	Panel (bioMérieux, Marcy-l'Étoile, France)	Influenza A+B( BioSquare, Korea), Covid-19 (Wonmed, Gyeonggi, Korea)		PCR (SCL, Yongin, Korea); <i>pneumoniae</i> PCR (Seegene, Seoul, Korea); BIOFIRE® Respiratory 2.1 Panel (bioMérieux, Marcy-l'Étoile, France)	<i>M. pneumoniae</i> PCR (DiaSorin)	Multiplex PCR (SCL, Yongin, Korea);
	Gyeongsang University Hospital	National BIOFIRE Respiratory 2.1 Panel (bioMérieux, Marcy-l'Étoile, France),	STANDARD Influenza A/B Biosensor, Korea)	FIA (SD Biosensor, Suwon, Korea)	BioFire Pneumonia Panel (bioMérieux, Marcy-l'Étoile, France)	<i>Mycoplasma pneumoniae</i> Antibody Diagnostic Reagent (Livzon Diagnostics, Zhuhai, China)	BioFire FilmArray Pneumonia Panel (bioMérieux, Marcy-l'Étoile, France)
<b>Busan</b>	Pusan National University Hospital	AdvanSure™ RV-plus Real-Time RT-PCR (Invitros, Seoul, Korea)	Not performed		Respiratory Pathogen Bacteria Panel [multiplex real-time PCR] (GC Labs, Yongin, Republic of Korea)	IgM CHORUS (Diesse)	Respiratory Pathogen Bacteria Panel [multiplex real-time PCR] (GC Labs, Yongin, Republic of Korea)
<b>Gangwon</b>	Kangwon National University Hospital	BIOFIRE Respiratory 2.1 Panel (bioMérieux, Marcy-l'Étoile, France)	STANDARD COVID-19	FIA	BIOFIRE Respiratory 2.1 Panel	IgM CHORUS (Diesse)	Allplex™ PneumoBacter

Location (Province)	Hospital	Respiratory virus PCR	Respiratory virus antigen test	Atypical PCR <sup>a</sup>	<i>Mycoplasma</i> serology <sup>b</sup>	Bacterial PCR <sup>c</sup>
		l'Étoile, France), Allplex™ (SD Biosensor, Suwon, Korea)		(bioMérieux, Marcy-l'Étoile, France); Allplex™ PneumoBacter Assay (Seegene, Seoul, Korea)		Assay (Seegene, Seoul, Korea)
<b>Gwangju</b>	Chosun University Hospital	BIOFIRE Respiratory 2.1 Panel (bioMérieux, Marcy-l'Étoile, France);	Sofia Influenza A+B FIA (QuidelOrtho, San Diego, CA, USA)	BIOFIRE Respiratory 2.1 Panel (bioMérieux, Marcy-l'Étoile, France); Allplex™ PneumoBacter Assay (Seegene, Seoul, Korea)	IgM LIAISON (DiaSorin)	Allplex™ PneumoBacter Assay (Seegene, Seoul, Korea)

Blank cells indicate that the corresponding diagnostic test was not performed at that institution.

<sup>a</sup> Atypical PCR included assays for *Mycoplasma pneumoniae*, *Chlamydia pneumoniae*, *Legionella pneumophila*, *Bordetella pertussis*, and *Bordetella parapertussis*.

<sup>b</sup> Serologic assays included particle agglutination (PA), chemiluminescent immunoassay (CLIA), and enzyme-linked immunosorbent assay (ELISA).

<sup>c</sup> Bacterial PCR included assays for *Streptococcus pneumoniae* and *Haemophilus influenzae* and was performed only when clinically indicated.

**Supplementary Table S3.** Seasonal Distribution of Etiologic groups in Children with Community-Acquired Pneumonia (CAP) by age group.

Age group	Season <sup>a</sup>	Atypical CAP	Pyogenic CAP	Viral CAP	No pathogen	Total	
All ages	Fall	22 (30.1)	5 (6.8)	21 (28.8)	25 (34.2)	73	
	Winter	6 (23.1)	3 (11.5)	12 (46.2)	5 (19.2)	26	
	Spring	27 (25.0)	2 (1.9)	36 (33.3)	43 (39.8)	108	
	Summer	132 (64.4)	3 (1.5)	22 (10.7)	48 (23.4)	205	
	Fall <sup>b</sup>	109 (64.9)	2 (1.2)	17 (10.1)	40 (23.8)	168	
	Winter <sup>b</sup>	20 (19.2)	3 (2.9)	61 (58.7)	20 (19.2)	104	
	Spring	7 (12.5)	2 (3.6)	29 (51.8)	18 (32.1)	56	
	<5 years	Fall	8 (19.0)	2 (4.8)	17 (40.5)	15 (35.7)	42
		Winter	1 (6.3)	1 (6.3)	12 (75.0)	2 (12.5)	16
		Spring	4 (8.0)	0 (0.0)	23 (46.0)	23 (46.0)	50
Summer <sup>b</sup>		25 (43.9)	1 (1.8)	13 (22.8)	18 (31.6)	57	
Fall <sup>b</sup>		30 (51.7)	0 (0.0)	11 (19.0)	17 (29.3)	58	
Winter		7 (10.6)	1 (1.5)	47 (71.2)	11 (16.7)	66	
Spring		3 (7.0)	0 (0.0)	25 (58.1)	15 (34.9)	43	
≥5 years		Fall	14 (45.2)	3 (9.7)	4 (12.9)	10 (32.3)	31
		Winter	5 (50.0)	2 (20.0)	0 (0.0)	3 (30.0)	10
		Spring	23 (39.7)	2 (3.4)	13 (22.4)	20 (34.5)	58
	Summer <sup>b</sup>	107 (72.3)	2 (1.4)	9 (6.1)	30 (20.3)	148	
	Fall <sup>b</sup>	79 (71.8)	2 (1.8)	6 (5.5)	23 (20.9)	110	
	Winter	13 (34.2)	2 (5.3)	14 (36.8)	9 (23.7)	38	
	Spring	4 (30.8)	2 (15.4)	4 (30.8)	3 (23.1)	13	

Values are presented as n (%), with percentages calculated for each season and age group.

<sup>a</sup> Seasons are presented in chronological order from fall 2023 through spring 2025. The *M. pneumoniae* epidemic period corresponded to the summer and fall 2024.

<sup>b</sup> *Mycoplasma pneumoniae* epidemic period.

**Supplementary Table S4. Clinical Features of Children With Community-Acquired Pneumonia (CAP) According to Etiologic Group**

<b>Variable</b>	<b>Viral CAP (N=198)</b>	<b>Atypical (N=323)</b>	<b>Pyogenic (N=20)</b>	<b><i>P</i> value</b>
<b>Clinical Presentation</b>				
Fever	173 (87.4)	307 (95.0)	19 (95.0)	0.006
Cough	190 (96.0)	313 (96.9)	14 (70.0)	<0.001
Rhinorrhea	83 (41.9)	66 (20.4)	3 (15.0)	<0.001
Dyspnea	64 (32.3)	48 (14.9)	8 (40.0)	<0.001
Rales	111 (56.1)	159 (49.2)	8 (40.0)	0.185
Wheezing	69 (34.8)	41 (12.7)	2 (10.0)	<0.001
Decreased breath sounds	35 (17.7)	120 (37.2)	4 (20.0)	<0.001
Chest retraction	36 (18.2)	28 (8.7)	1 (5.0)	0.003
<b>Radiographic finding</b>				
Bronchial infiltration	132 (66.7)	107 (33.1)	3 (15.0)	<0.001
Consolidation	94 (47.5)	269 (83.3)	16 (80.0)	<0.001
Pleural effusion	8 (4.0)	51 (15.8)	7 (35.0)	<0.001
<b>Treatment and Hospitalization</b>				
IV antibiotics $\geq 3$ days	126 (63.6)	183 (56.7)	20 (100.0)	<0.001
Steroid $\geq 3$ days	56 (28.3)	153 (47.4)	3 (15.0)	<0.001
Length of stay, days, median (IQR) [n = 191/283/20]	5.0 (4.0–10.0)	6 (4.0–8.0)	12.0 (7.0–26.0)	<0.001
<b>Inflammatory markers</b>				
WBC, $\times 10^3/\mu\text{L}$ , median (IQR) [n = 191/316/20]	9.5 (6.2–14.0)	8.1 (6.2–11.3)	22.2 (14.4–31.8)	<0.001
CRP, mg/dL, median (IQR) [n = 198/321/20]	2.2 (0.5–5.6)	3.5 (1.2–8.7)	15.3 (7.8–23.5)	<0.001
Procalcitonin, ng/mL, median (IQR) [n = 100/161/17]	0.2 (0.1–1.6)	0.1 (0.1–0.3)	0.5 (0.2–6.2)	<0.001

Sample sizes for each variable are indicated where applicable.

CAP, community-acquired pneumonia; IQR, interquartile range.