

Supplementary Table 1. List of strains used in this study.

Strain or plasmid	Characteristics	Source or reference
Strains		
<i>E. coli</i>		
MC1061	hsdR araD139 Δ (ara-leu)7697 Δ lacX74 galU galK rpsL (Sm ^R)	Casabadan, et al.,1980 ¹
CC118 λ pir	Δ (ara, leu)7697 araD139 Δ lacX74 galE galk phoA20 thi-1 rpsE rpoB(Rf ^R) argE(Am) recA1 λ pir ^R	Herrero et al., 1990 ²
S17-1 λ pir	thi proA hsdR recA RP4-2-tet::Mu-1 kan::Tn7 integrant (Tp ^R Sm ^R) λ pir	Simon, 1983 ³
Dh5 α	F- Φ 80dlacZ Δ M15 Δ (lacZYA-argF)U169 recA1 endA1 hsdR17(rK ⁻ mK ⁺) supE44 thi-1 relA1 gyrA96	Hanahan, 1983 ⁴
SY327 λ pir	wild type; araD, Δ (lac pro) argE(Am) recA56 Rif ^R nalA λ pir	Miller et al., 1988 ⁵
GM2163	dam-13::Tn9 (Cam ^r) dcm-6 hsdR2 (rK ⁻ mK ⁺)leuB6 hisG4 thi-1 araC14 lacY1 galk2 galT22 xylA5 mtl-1 rpsL136 (Str ^r) fhuA31 tsx-78 glnV44 mcrA mcrB1	Fermentas
<i>P. putida</i>*		
IsoF	wild type; isolated from tomato roots; AHL ⁺	Steidle et al 2001 ⁶
Δ dotHGF	IsoF deletion mutant of dotH, dotG, dotF	This study
Δ 23	IsoF deletion mutant of Piso_02323	This study
Δ dotD	IsoF deletion mutant of dotD	This study
Δ 23-trbN-dotD	IsoF deletion mutant of Piso_02323, trbN, dotD	This study
Δ 32-33	IsoF deletion mutant of Piso_02332, Piso_02333	This study
Δ 32-33 /pBBR::32-33	Complementation of Piso_02332 and Piso_02333 using pBBR1MCS-2 derivative	This study
Δ 32-33/pBBR::32	Complementation of Piso_02332 using pBBR1MCS-2 derivative	This study
Δ 33	IsoF deletion mutant of Piso_02333	This study
Δ 33/pBBR::33	Complementation of Piso_02333 using pBBR1MCS-2 derivative	This study
Δ T4B	IsoF deletion mutant of the kib cluster, from Piso_02313 to Piso_02360	This study
Δ T4B/pBBR::32	Complementation of Piso_02332 using pBBR1MCS-2 derivative	This study

Strains	Characteristics	Source or reference
KT2442*	Wild type; <i>hsdR</i> Rif ^R	Laboratory collection
W2*	wild type	Laboratory collection
OUS82*	wild type	Laboratory collection
A9Rx29*	wild type	Laboratory collection
<i>P. aureofaciens</i> *	ATCC13985; wild type	Conway et al., 1956 ⁷
<i>P. entomophila</i> *	wild type	Laboratory collection
<i>P. chlororaphis</i> *	wild type	Laboratory collection
<i>P. fluorescens</i> *	WCS417r; wild type	Laboratory collection
<i>P. syringae</i> *	DC3000; wild type	Laboratory collection
<i>Pectobacterium carotovorum</i> *	wild type	Laboratory collection
<i>Ralstonia solanacearum</i> *	DSM9544; wild type	Laboratory collection
<i>P. aeruginosa</i>		
JB646	Wild type, Gfp tagged	Laboratory collection
PUPA3	Carrying pBBR-5-Gfp	Laboratory collection
PA14*	wild type	Rakhimova et al., 2008 ⁸
PAO1*	wild type	Iglewski et al., 1999 ⁹
<i>Burkholderia cenocepacia</i>	H111 tagged with Gfp	Laboratory collection
<i>Burkholderia vietnamiensis</i>	LMG10929 tagged with Gfp	Laboratory collection

Strains	Characteristics	Source or reference
<i>Burkholderia cepacia</i>	M64; carrying pBBR-5-Gfp	Laboratory collection
<i>Burkholderia multivorans</i>	R-654 tagged with Gfp	Laboratory collection
<i>Burkholderia thailandensis</i>	LMG20219; carrying pBBR-5-Gfp	Laboratory collection
<i>Serratia liquefaciens</i>	MG1 tagged with Gfp	Laboratory collection
<i>Erwinia amylovora</i> *	wild type	This study
* These strains and its variants were labeled either with Gfp, mCherry or Cfp using the pUC18T-mini-Tn7T-Gm when indicated.		This study
Plasmids		
pUC18T-mini-Tn7T-Gm- <i>egfp-a</i>	carrying mini Tn7(Gm) PA1/04/03- <i>egfp-a</i> for Gfp tagging	Choi and Schweizer, 2006 ¹⁰
pUC18T-mini-Tn7T-Gm- <i>mCherry</i>	carrying mini Tn7(Gm) PA1/04/03- <i>mCherry</i> for mCherry tagging	Choi and Schweizer, 2006
pUC18T-mini-Tn7T-Gm- <i>ecfp</i>	carrying mini Tn7(Gm) PA1/04/03- <i>ecfp</i> for Cfp tagging	Choi and Schweizer, 2006
pUX-BF13	<i>tnsA-E</i> ; helper plasmid providing the Tn7 transposition functions	Koch et al., 2001 ¹¹
pUTminiTn5	delivery vector for mini-Tn5 Km2; Km ^R	De Lorenzo et al, 1990 ²
pGPI-SceI	Suicide plasmid with <i>oriR6K</i> , <i>mob</i> ⁺ , I-SceI restriction site; Tp ^R	Elisabeth Steiner, Laboratory collection
pGPI-SceI::TetAR	Suicide plasmid with <i>oriR6K</i> , <i>mob</i> ⁺ , I-SceI restriction site; Tp ^R Tc ^R	This study
pGPI::Tet-H-F	pGPI-SceI-Tet plasmid with fused regions flanking the genes <i>dotH</i> , <i>dotG</i> , <i>dotF</i>	This study
pGPI::Tet-23	pGPI-SceI-Tet plasmid with fused regions flanking the gene <i>Piso_02323</i>	This study
pGPI::Tet-DotD	pGPI-SceI-Tet plasmid with fused regions flanking the gene <i>dotD</i>	This study
pGPI::Tet-23-DotD	pGPI-SceI-Tet plasmid with fused regions flanking the genes <i>Piso_02323</i> and <i>dotD</i>	This study

Plasmids	Characteristics	Source or reference
pGPI::Tet-32-33	pGPI-SceI-Tet plasmid with fused regions flanking the genes <i>Piso_02332</i> and <i>Piso_02333</i>	This study
pGPI::Tet-33	pGPI-SceI-Tet plasmid with fused regions flanking the gene <i>Piso_02333</i>	This study
pGPI::TetAR-T4B	pGPI-SceI-Tet plasmid with fused regions flanking the genes <i>Piso_02313</i> to <i>intA_3</i>	This study
pDAIGm-SceI	pDA17 plasmid carrying the <i>I-SceI</i> nuclease gene; Gm ^R	Elisabeth Steiner, Flannagan et al., 2008 ¹²
pRK2013	Helper plasmid; RK2 derivative, <i>mob⁺ tra⁺ ori</i> ColE1; Km ^R	Figurski et al., 1979 ¹³
pBBR1MCS-2	Broad-host-range plasmid, <i>lacZα</i> ; Km ^R	Kovach et al 1995 ¹⁴
pBBR::32	pBBR1MCS-2 carrying <i>Piso_02332</i> , complementation plasmid	This study
pBBR::33	pBBR1MCS-2 carrying <i>Piso_02333</i> , complementation plasmid	This study
pBBR::32-33	pBBR1MCS-2 carrying <i>Piso_02332</i> and <i>Piso_02333</i> , complementation plasmid	This study
pLG99	Transposon T23 (<i>ISlacZ-PrhaB</i> out-/FRT); Tp ^R	Gallagher et al., 2013 ¹⁵
pLG99::Km	Transposon T23 (<i>ISlacZ-PrhaB</i> out-/FRT); Km ^R , Tp ^R	This study

Supplementary Table 2. List of primers used in this study.

Primer name	Sequence	Restriction site/Description
Primers used for construction of markerless deletion of <i>dotH</i>, <i>dotG</i>, <i>dotF</i>		
DotIHindIII_F	GCGCaagcttTGCGTATTCGGCTTGACC	HindIII
DotIXhoI_R	GCGCctcgagGGCGACAGCTGAACAGAA	XhoI
DotIEXhoI_F	GCGCctcgagATAGGGAAATACACATGGC	XhoI
DotIEcoRI_R	GCGCgaattcGGCTTGCGTTTGATTATTG	EcoRI
DotI_out_F	TCGACCTCAAGCGCCGTAA	Check primer
DotIE_out_R	CCGCCAATAATCCACCCAC	Check primer
Primers used for construction of markerless deletion of <i>Piso_02323</i>		
trbNNcoI_F	GCGCccatggGCATGGCCGTAAGTGGTTG	NcoI
trbNEcoRI_R	GCGCgaattcGCGCTGCTGTTCGAGGTA	EcoRI
DotLKpn_F	GCGCggtaccTGTA	KpnI
DotLNcoI_R	GCGCccatggCCTCACCATAGCATGCAGA	NcoI
trbN_out_R	CTCCTGAAGTTGTCATTCT	Check primer
DotL_out_F	GCGCCTTCGAGCATAACTA	Check primer
Primers used for construction of markerless deletion of <i>dotD</i>		
2355EcoRI_R	GCGCgaattcTTTCGACATAGGCATAGC	EcoRI
trbN-KpnI_F	GCGCggtaccTTGTACCGCGTAGTCGTC	KpnI
trbN-NcoI_R	GCGCccatggGGTAGCTCACCACGTACT	NcoI
dotC-NcoI_F	GCGCccatggCTGAGCGCCTTTACGACAA	NcoI
trbN_out_F	GGCATTCTGGTTCTGGTTC	Check primer
dotC_out_R	CAATTTCCACACGCGTTC	Check primer
Primers used for construction of markerless deletion of <i>Piso_02323</i>, <i>trbN</i>, <i>dotD</i>		
DotCNcoI_F	GCGCccatggGATGACCCTCGAAGACG	NcoI
DotCEcoRI_R	GCGCgaattcCAGCGGGAAGCTCAGAT	EcoRI
Primers DotLKpn_F and DotLNcoI_R were used to amplify the upstream homology region		
Check primers DotL_out_F and dotC_out_R used to validate the deletion		

Primer name	Sequence	Restriction site/Description
Primers used for construction of markerless deletion of <i>Piso_02332</i>, <i>Piso_02333</i>		
2364-MluI_F	GCGCacgcgtCGCGCTCAAGCAGATTCA	MluI
2364-SphI_R	GCGCgcatgcCGGCCACTTTTTCCCAGTA	SphI
2361-XbaI_F	GCGCtctagaAAAAGGTTAGCGTGCAGC	XbaI
2361-MluI_R	GCGCacgcgtCCTAAAGGTTCTCCAATCCA	MluI
2361_out_F	GAATGTTTGAACGAGGCCC	Check primer
2364_out_R	TCACGCGAATGGAGATGTTG	Check primer
Primers used for construction of markerless deletion of <i>Piso_02333</i>		
2362-MluI_R	GCGCacgcgtTCAGTCCTCGTGCTTTTTG	MluI
2362-XbaI_F	GCGCtctagaCATTTTCATGGTGGCGTGT	XbaI
Primers 2364-MluI_F and 2364-SphI_R were used to amplify the upstream homology region		
Check primers 2361_out_F and 2364_out_R used to validate the deletion		
Primers used for construction of markerless deletion of the <i>kib</i> cluster region		
2341KpnI_F	GCGCggtaccCAATTATTTTTACCCGCTCG	KpnI
2341NcoI_R	GCGCccatggCGTGGCAGGACTAATCTTT	NcoI
2390NcoI_F	GATTTACTGACCGCCATGGT	NcoI
2390EcoRI_R	GCGCgaattcCCTTGCTTTAACCTGTAGTG	EcoRI
2340_out_F	ACGGCCAAGTTGAGTTACAA	Check primer
2390_out_R	AATTTTACAGCCCAAACCC	Check primer
Construction of pGPI-Scel::TetAR		
TetPstI_R	GCGCctgcagtCAATCGTCACCCTTTCTCG	PstI
TetPstI_F	GCGCctgcagTCAGCGATCGGCTCGTTGCC	PstI
Ligation check primers flanking the MCS of pGPI-Scel		
p44_pGPI_F	AGCTGATCCGGTGGATGAC	
p45_pGPI_R	ACGGTTGTGGACAACAAGC	
p131_pGPI_F	ACTCAAGCGTTAGATGC	

Primer name	Sequence	Restriction site/Description
Construction of pBBR::32		
2362-EcoRI_F	GCGCgaattcTTCAATTGCTCCCATCTTC	EcoRI
2362-XbaI_R	GCGCtctagaATCAGTCCTCGTGCTTTTTG	XbaI
Construction of pBBR::33		
2363-XbaI_R	GCGCtctagaTTCTGCTTGAGCGCGATC	XbaI
2363-EcoRI_F	GCGCgaattcGGAAGTATGATGATGGGTAA	EcoRI
Construction of pBBR::32-33		
Primers 2362-EcoRI_F and 2363-XbaI_R were used to amplify <i>Piso_02332</i> and <i>Piso_02333</i>		
Ligation check primers flanking the MCS of pBBR1MCS-2		
M13For (-21)	TGTAAAACGACGGCCAGT	
M13Rev	CAGGAAACAGCTATGACC	
Insertion check primers for Tn23		
tn23_F	GCTGCACCATTCGCGTTA	
tn23_R	GCTGATTCGAGGCGTTAA	
Construction of pLG99::Km		
Kan-AatII_F	GCGCgacgtcGTCATTTCTGAACCCCAGAG	AatII
Kan-AatII_R	GCGCgacgtcGACTGGGCGGTTTTATGGA	AatII

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