

Supplementary Information

for

Chemical diversity of bilin species defines the chiroptical landscape of phytochromes

Oliver Maximilian Eder^{1,*}, Chongyao Wei², Marie Reißbüchel¹, Igor Schapiro^{2,3,4,*}, Andreas Winkler^{1,5,*}

Affiliations

¹ Institute of Biochemistry, Graz University of Technology, 8010 Graz, Styria, Austria

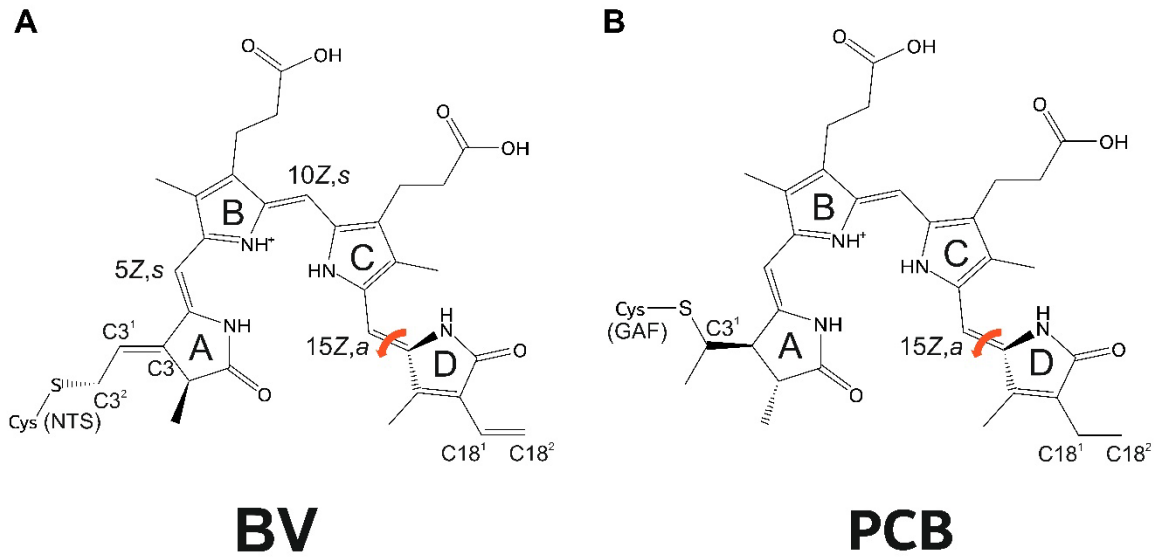
² Department of Physics, TU Dortmund University, Otto-Hahn-Str. 4, 44227 Dortmund, Germany.

³ Institute of Chemistry, The Hebrew University of Jerusalem, Jerusalem 9190401, Israel

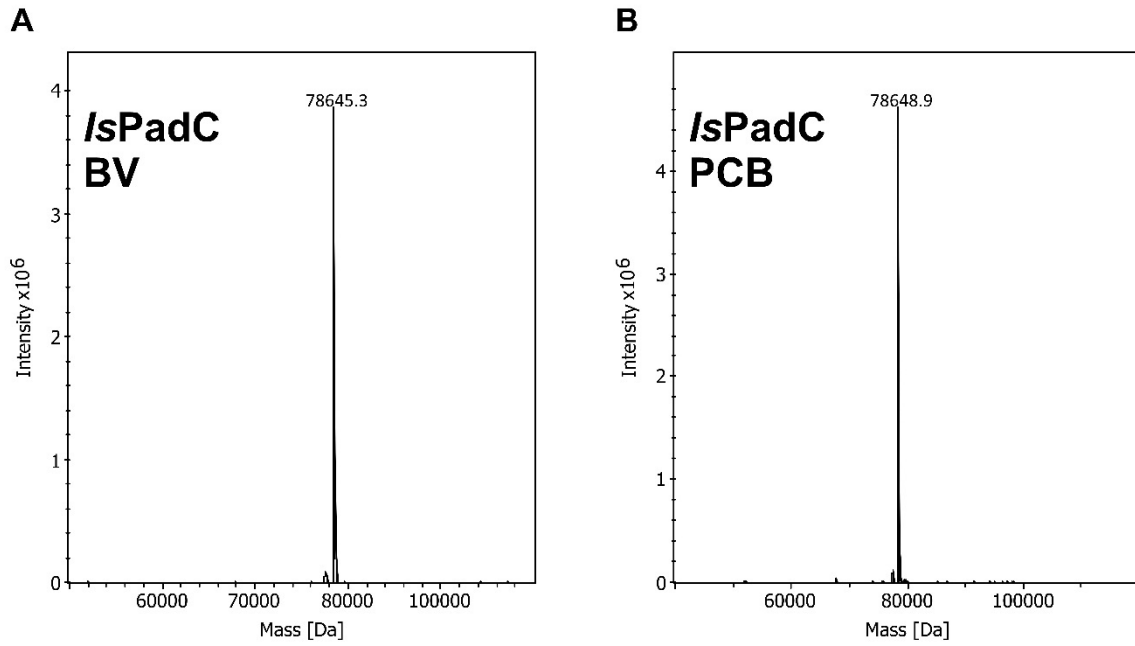
⁴ Research Center Chemical Sciences and Sustainability, University Alliance Ruhr, 44801 Bochum, Germany

⁵ BioTechMed-Graz, Graz, Austria

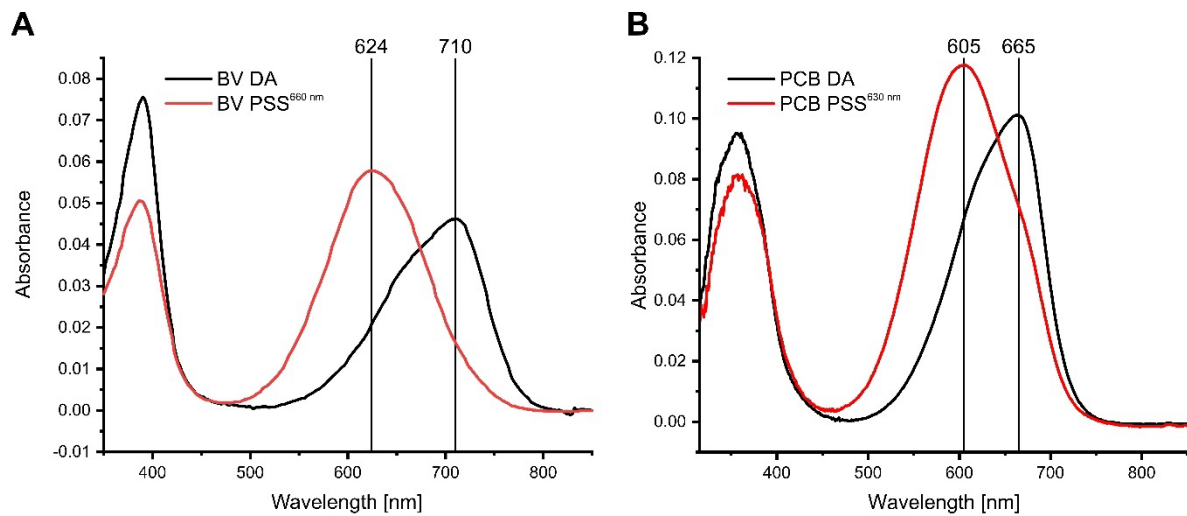
* Correspondence to: oliver.eder@tugraz.at, igor.schapiro@tu-dortmund.de,
andreas.winkler@tugraz.at



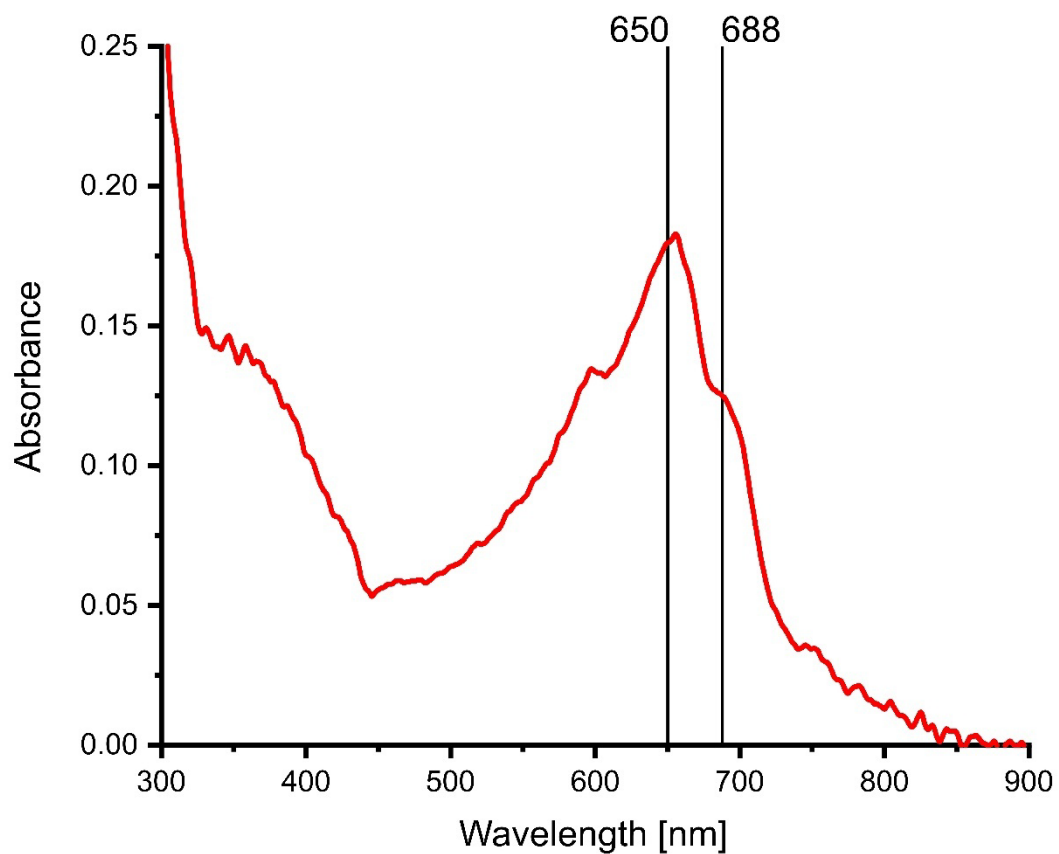
Supplementary Figure 1A+B: Chemical structures of biliverdin (BV) and phycocyanobilin (PCB), respectively. The location of the covalent attachment site to the protein is highlighted with either Cys (NTS) or Cys (GAF). In addition, the bilin atoms involved in the covalent attachment are shown which are either C3² (BV) or C3¹ (PCB). Note the nomenclature of the bilin A-D rings. Upon red light illumination, the C15=C16 double bond isomerizes from a 15Z to 15E configuration (red arrow).



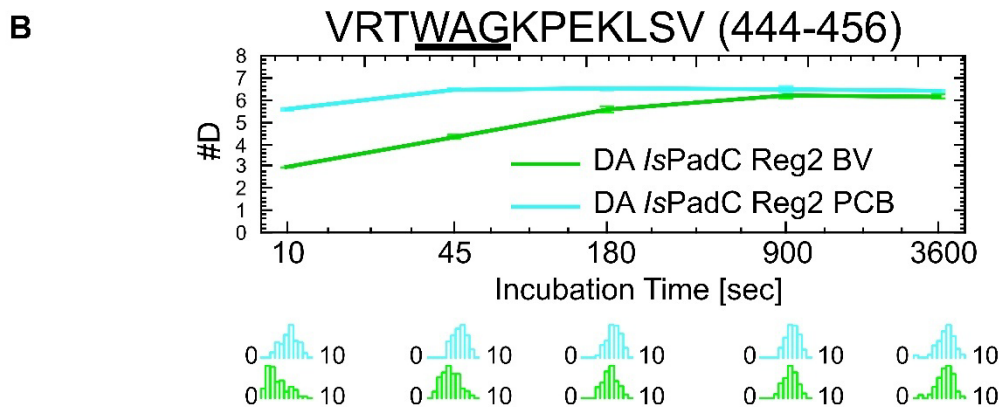
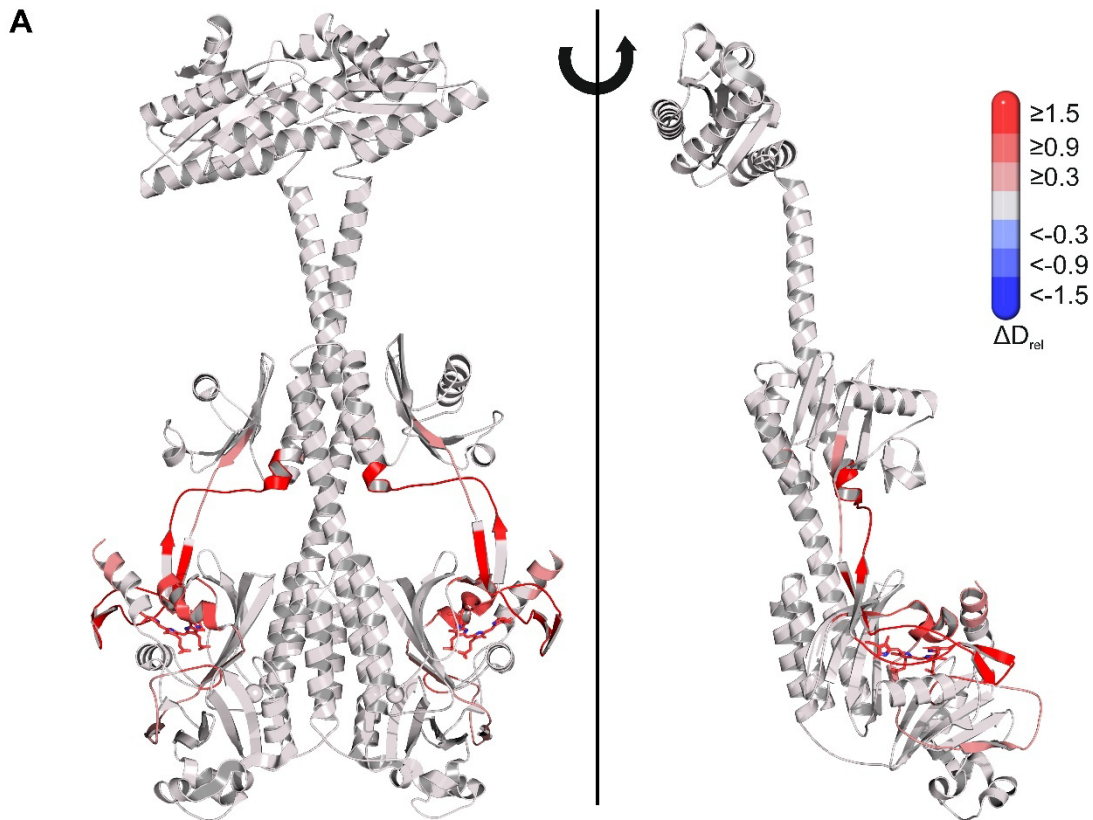
Supplementary Figure 2: Intact mass measurements prove complete loading of the proteins *IsPadC BV* (left) and *IsPadC PCB* (right) with their respective bilin cofactor. The theoretically calculated Holoprotein masses for *IsPadC BV* (78646,08 Da) and *IsPadC PCB* (78650,13 Da) agree well with the experimentally determined masses.



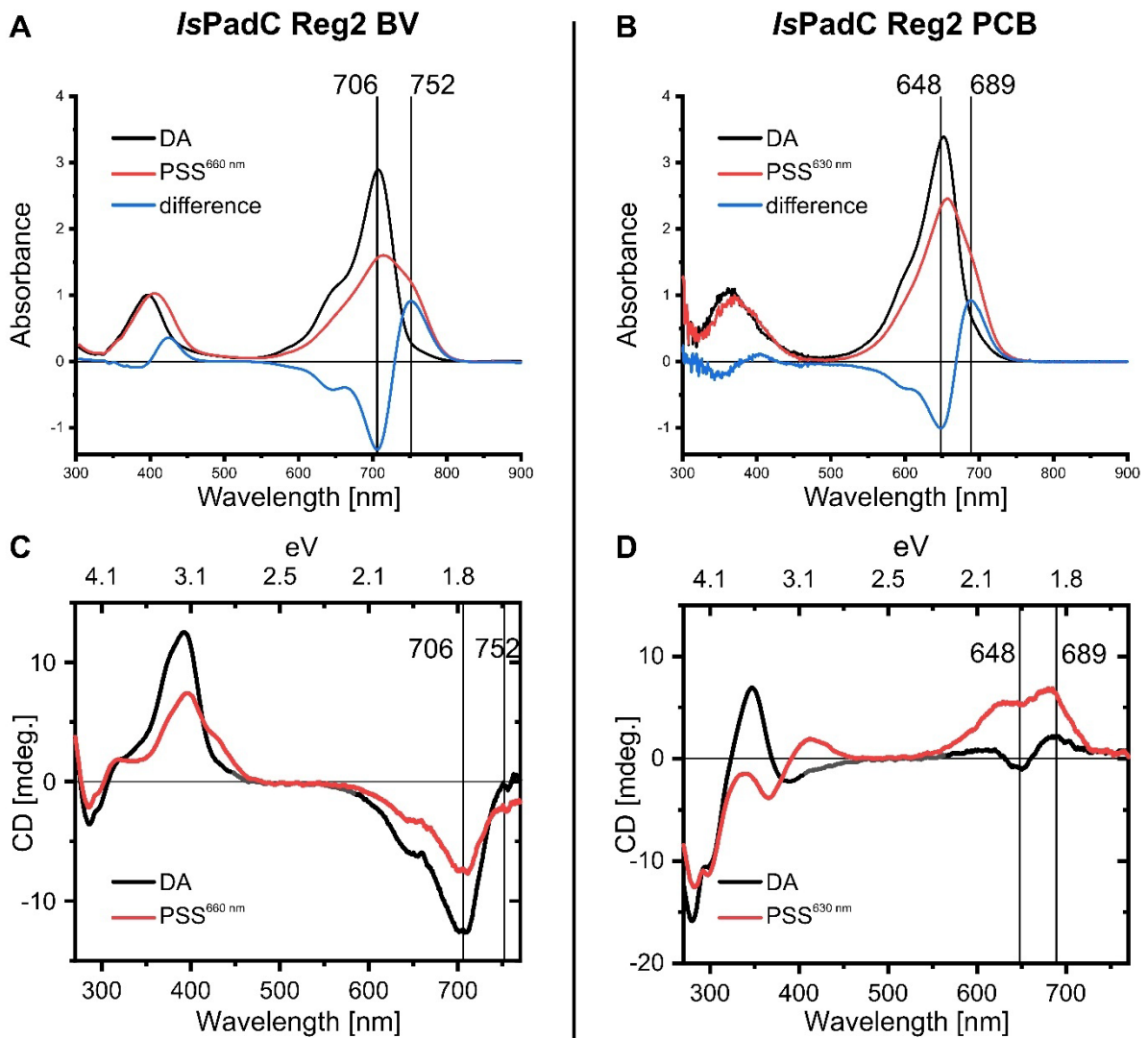
Supplementary Figure 3: A+B) Protein denaturation experiments of *IsPadC* BV and *IsPadC* PCB, respectively. Samples were either denatured using 0.1% TCA/Methanol (BV) or 6M Guanidinium HCl/1 % HCl (PCB). In both panels, the dark-adapted state (DA) is shown as a black line. For the illuminated samples (BV 660 nm, PCB: 630 nm), samples were treated with red light followed by immediate denaturation. Black vertical lines highlight the maxima of the respective UV/vis peak. BV data was taken from <https://doi.org/10.7554/eLife.34815>.



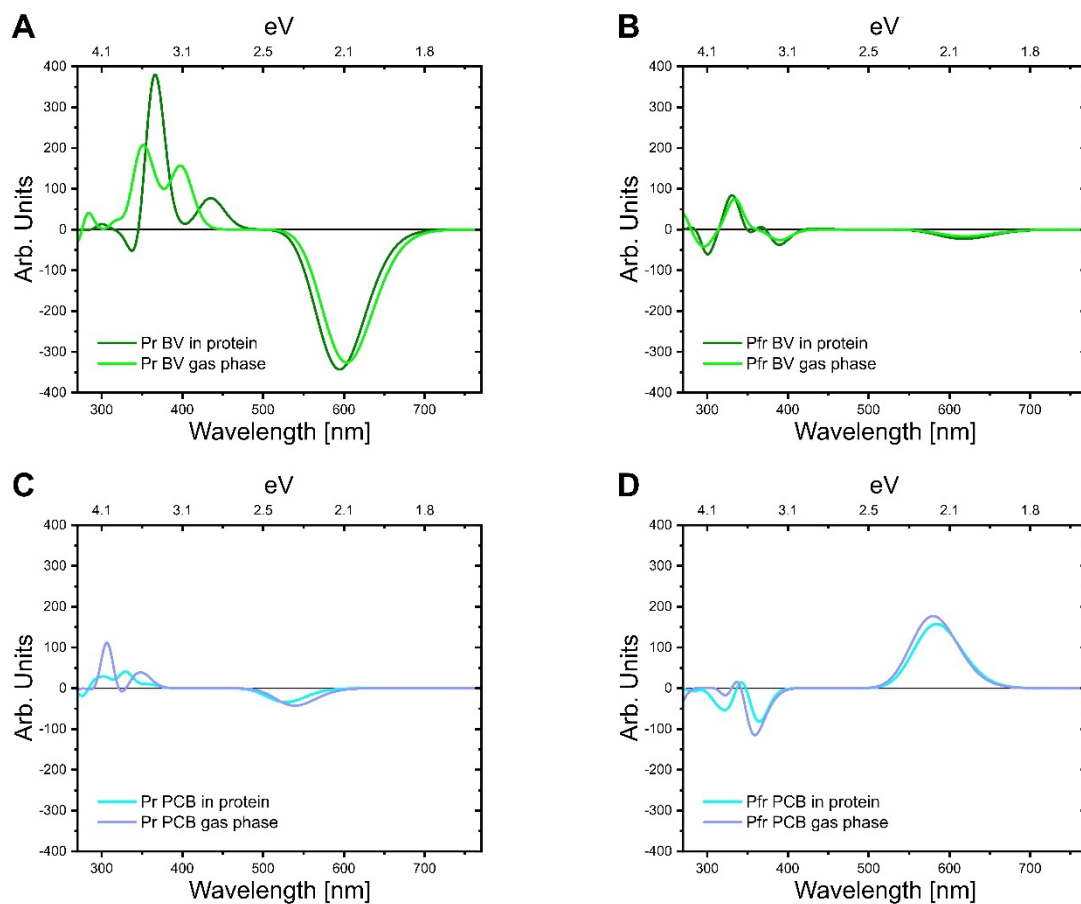
Supplementary Figure 4: *In crystallo* Optical Spectroscopy (icOS) data of *IsPadC* Reg2 PCB (C17L L251C S505V A526V) at 100 K. Before plotting, the spectrum was denoised. Note that the crystal was not illuminated but was kept in complete darkness. Nevertheless, the shape of the spectrum suggests the stabilization of a mixed Pr/Pfr state in the crystal.



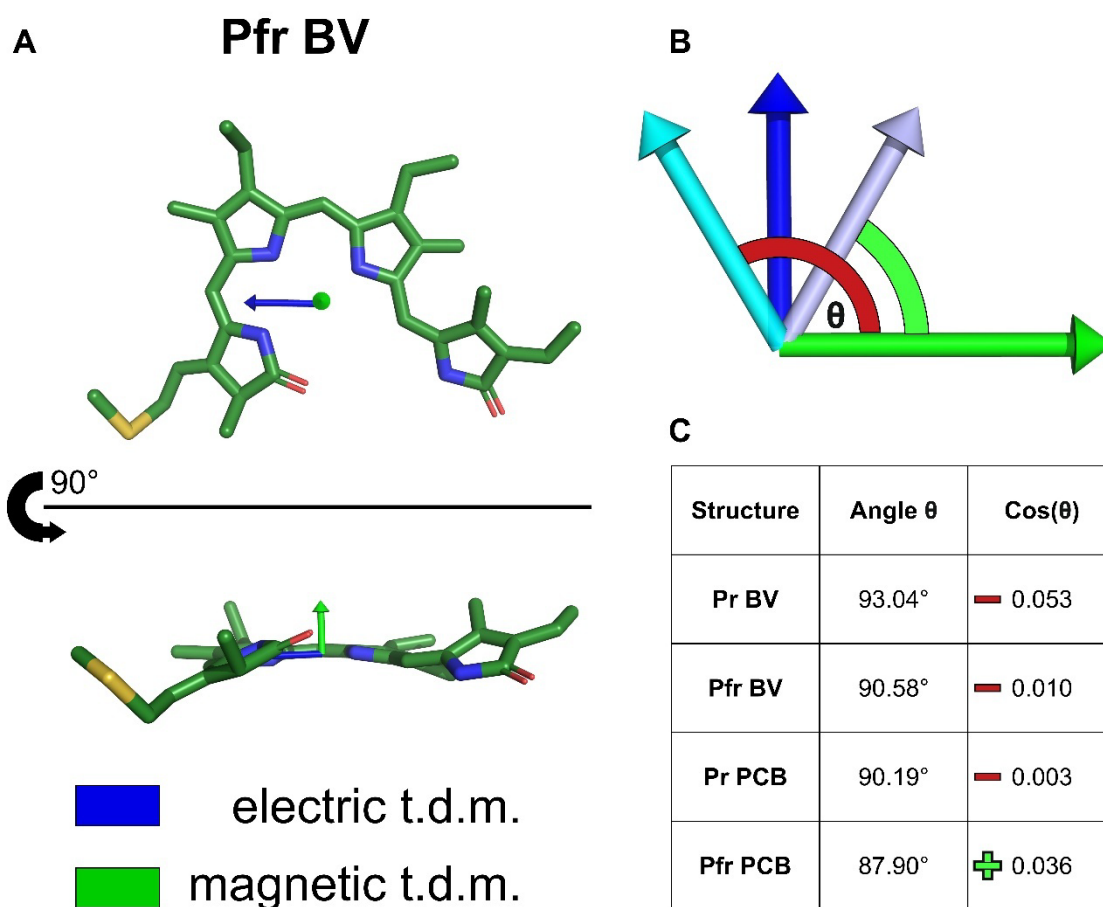
Supplementary Figure 5: Dark adapted (DA) HDX-MS data of a BV- and a PCB-bound *IsPadC* variant. A) Differences in deuterium exchange (ΔD_{rel}) of *IsPadC* Reg2 PCB minus *IsPadC* Reg2 BV ($t=10$ s) mapped onto the crystal structure of *IsPadC* (PDB: 5llw). In the regions highlighted in red, the PCB variant is significantly more dynamic than the BV variant. B) Example PHY tongue peptide with diverging deuterium incorporation in the dark-adapted (DA) states. This peptide contains the WAG motif (underlined) and harbors strongly increased conformational dynamics for the PCB variant.



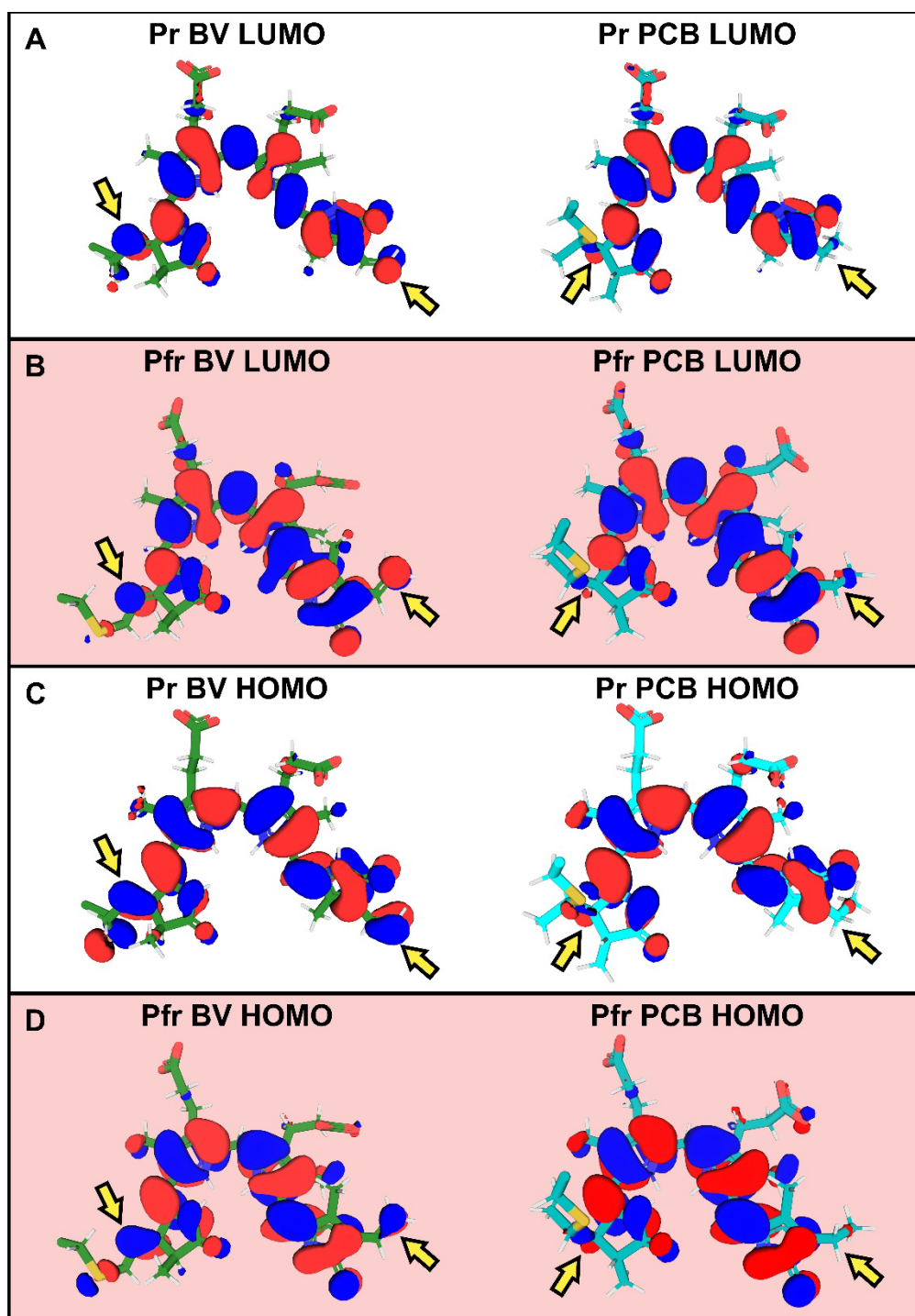
Supplementary Figure 6: Spectral properties of *IsPadC Reg2 BV* versus *PCB*. In all panels, spectra of dark adapted (DA) and red light illuminated (BV: PSS^{660 nm}, PCB: PSS^{630 nm}) samples are shown with black and red lines, respectively. A+B) UV/vis characterization of *IsPadC Reg2 BV* and *IsPadC Reg2 PCB*. Black vertical lines highlight the maxima and minima of the difference spectrum (PSS minus DA) in blue. Spectra are scaled to 1 according to their Soret band maximum (BV: 398 nm, PCB: 357 nm). C+D) Experimentally determined CD traces of *IsPadC Reg2 BV* and *IsPadC Reg2 PCB*. Black vertical lines indicate the maxima and minima of the UV/vis difference spectrum determined in A+B.



Supplementary Figure 7 A-D): Comparison of CD spectra calculated from bilin structures in the protein context versus isolated bilin structures in the gas phase. In green colors, the biliverdin (BV) data and in blue colors the phycocyanobilin (PCB) data is shown.



Supplementary Figure 8: Specific calculations of transition dipole moments determining the sign of the rotatory strength and thus the CD behavior. A) Example structure of biliverdin (BV) in the Pfr configuration. Although the calculations were conducted in the protein context, the amino acids are not shown to aid visibility. Displayed at the center of mass of the molecule, the rescaled electric (blue) and magnetic (green) transition dipole moments (t.d.m.) are visible. B) Scheme to visualize the dependency of the sign of the rotatory strength with the angle between the electronic (blue shades) and magnetic (green) transition dipole moment. Due to the properties of the cosine function, angles greater than 90° lead to negative $\cos(\theta)$ values whereas angles smaller than 90° lead to positive $\cos(\theta)$ values. C) Overview of angles θ and $\cos(\theta)$ calculated for the biliverdin (BV) and phycocyanobilin (PCB) structures in the Pr or Pfr geometry, respectively.



Supplementary Figure 9: Calculated molecular orbitals of *IsPadC* BV and PCB in Pr and Pfr, respectively. A-D) HOMO and LUMO orbitals were calculated from the QM/MM optimized starting structures utilized for the *in silico* CD simulations. Positive phases are shown in red whereas negative phases are displayed in blue. Differences in orbitals between the BV and PCB cofactor are highlighted with yellow arrows. In BV, the C3=C3¹ interaction on the A-ring (left) and the C18¹=C18² interaction on the D-ring (right) has double bond character. The equivalent positions in PCB have single bond character.

Supplementary Table 1: Primers used to generate the *IsPadC C17L L251C* construct.

primer name	primer sequence
IsPSM_C17L _fwd	GCAGCATTGGATCAAGAACCGATTCATATTCCGAATGCAATTC
IsPSM_C17L _rev	TTGATCCAATGCTGCAATCAGTTTGCTGATATCATCACTACCC
IsPSM_L251C _fwd	GTTCTGCGTGCAGTTAGCCCTTGTCACATGCAGTATCTGCGTAATTTTGG
IsPSM_L251C _rev	AACTGCACGCAGAACACCGCTGCTCAGATTAAGTGC

Supplementary Table 2: HDX-MS dataset statistics for the shown datasets according to the conventions of the HDX-MS community

	<i>IsPadC Reg2 BV</i>	<i>IsPadC Reg2 PCB</i>	Peptides occurring in both datasets
Light conditions	Dark-adapted (DA)	Dark-adapted (DA)	-
HDX reaction details	10mM HEPES, 150mM NaCl, 2mM MgCl ₂ , pD=7.0, 20°C		-
HDX time course (s)	10, 45, 180, 900, 3600		-
HDX control samples	Unlabeled control (Reg2 BV DA)	Unlabeled control (Reg2 PCB DA)	-
Back-exchange	Not measured		-
# of Peptides	180	161	120
Sequence coverage	92%	93%	85%
Avg peptide length / Avg. redundancy	13/3	13.6/3	13.5/2
Replicates	3	3	-
Repeatability (average SD for each time point)	0.09, 0.06, 0.08, 0.07, 0.06	0.05, 0.05, 0.04, 0.06, 0.07	-
Significant differences in HDX	$\Delta\text{HDX} > 0.3 \text{ D}$		-