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1. General Considerations

Air and moisture sensitive reactions were carried out in glovebox or in oven-dried glassware sealed with rubber septa using standard schlenk techniques. Solvents used in moisture and oxygen sensitive reactions were firstly degassed by three freeze-pump-thaw cycles and then distilled under Ar protection after dehydration (ether solvents were dried over sodium metal). Regular chemicals were purchased from commercial sources with purity over 95% and used without further purification. NMR spectra were collected using a Bruker 400 MHz spectrometer and JEOL JNM-ECZL500G. Chemical shifts are reported in ppm relative to the deuterated solvents. The peak information was described as: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet; coupling constants in Hz. GC analysis was carried out on an SHIMADAZU GC 2010 PLUS system. GC/MS analysis was carried out on a Thermo Trace ISQ system equipped with a Thermo SCIENTIFIC TG-5MS (30 m x 250 μ m x 25 μ m). High resolution exact mass measurements (HRMS) were performed on Thermo SCIENTIFIC Q EXACTIVE. Cyclic voltammetry was recorded using a CHI760E potentiostation.

2. Synthesis and Characterization Data of Substrates

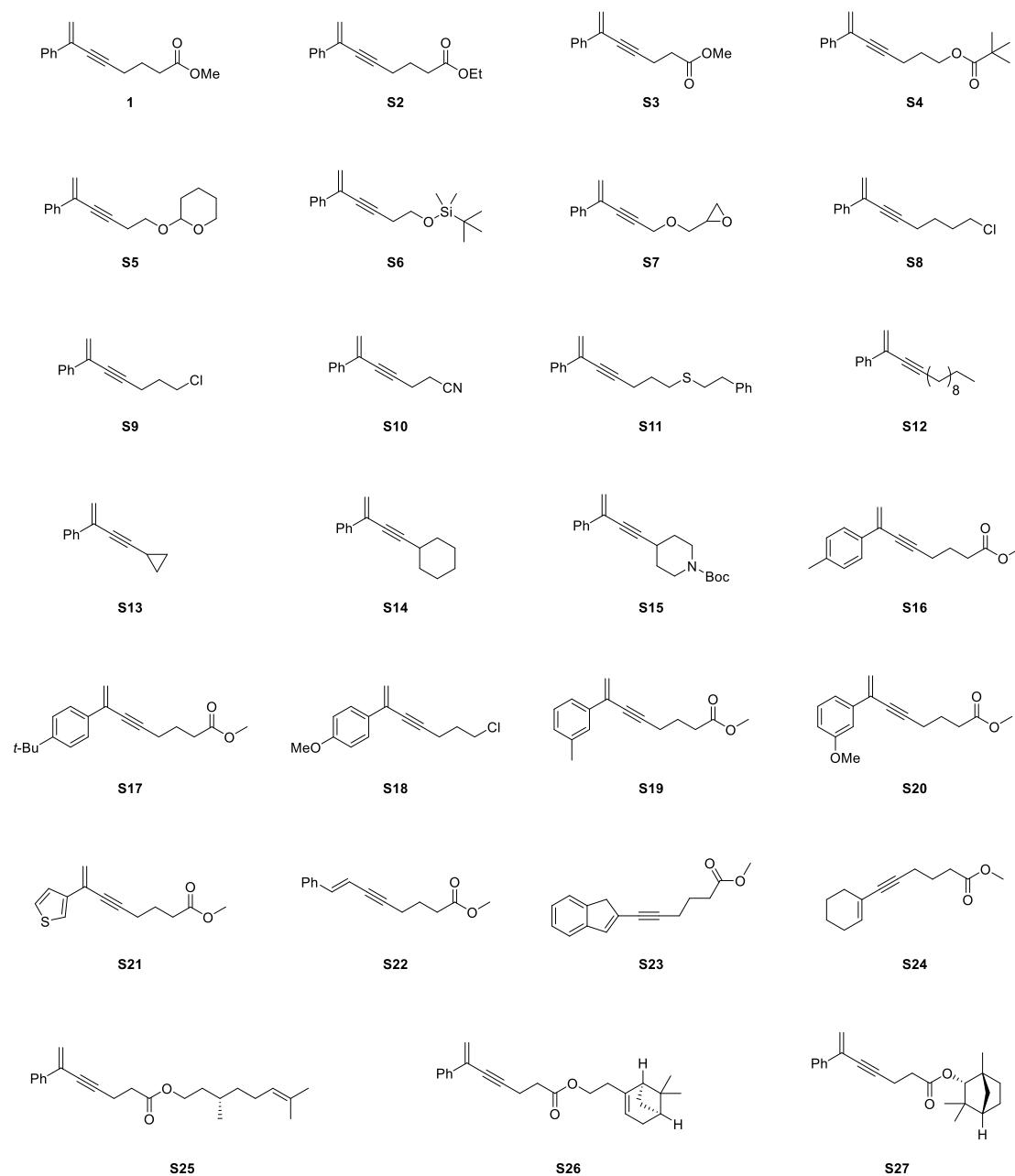
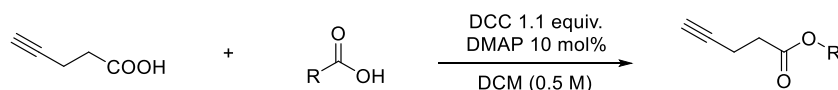


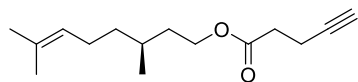
Fig. S1 Investigated enynes.

General procedure for synthesis of alkynes



To a solution of pent-4-ynoic acid (1.96 g, 20 mmol) and ROH (1.0 equiv., 20 mmol) in DCM (40 mL) were added N, N'-Dicyclohexylcarbodiimide (4.54 g, 22 mmol), 4-dimethylaminopyridine (244 mg, 2.0 mmol). After stirring at room temperature for 12 h, the solution was extracted with DCM (2 × 50 mL). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated in vacuo¹.

(S)-3,7-dimethyloct-6-en-1-yl pent-4-ynoate



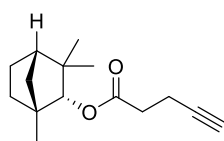
Compound **S28** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (4.06 g, 86% yield).

¹H NMR (400 MHz, CDCl₃) δ 5.17-4.98 (m, 1H), 4.19-4.08 (m, 2H), 2.58-2.43 (m, 4H), 2.05-1.87 (m, 3H), 1.70-1.63 (m, 4H), 1.59 (s, 3H), 1.57-1.50 (m, 1H), 1.48-1.40 (m, 1H), 1.36-1.29 (m, 1H), 1.22-1.12 (m, 1H), 0.90 (d, *J* = 6.5 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.86, 131.41, 124.62, 82.59, 69.02, 63.35, 37.03, 35.50, 33.51, 29.52, 25.76, 25.45, 19.46, 17.71, 14.47.

HRMS (APCI) calcd. for C₁₅H₂₅O₂⁺ [M+H]⁺: 237.1850; found: 237.1848.

(1R,2R,4S)-1,3,3-trimethylbicyclo[2.2.1]heptan-2-yl pent-4-ynoate



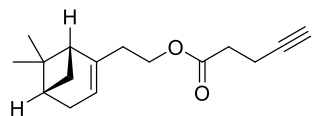
Compound **S29** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (3.5 g, 75% yield).

¹H NMR (400 MHz, CDCl₃) δ 4.38 (d, *J* = 2.0 Hz, 1H), 2.64-2.56 (m, 2H), 2.55-2.46 (m, 2H), 1.96 (t, *J* = 2.5 Hz, 1H), 1.77-1.65 (m, 3H), 1.60-1.54 (m, 1H), 1.49-1.39 (m, 1H), 1.18 (dd, *J* = 10.3, 1.7 Hz, 1H), 1.09 (s, 3H), 1.04 (s, 3H), 0.77 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 172.07, 86.59, 82.65, 69.01, 48.33, 48.27, 41.36, 39.41, 33.58, 29.67, 26.62, 25.79, 20.13, 19.34, 14.57.

HRMS (APCI) calcd. for C₁₅H₂₃O₂⁺ [M+H]⁺: 235.1693; found: 235.1687.

2-((1R,5S)-6,6-dimethylbicyclo[3.1.1]hept-2-en-2-yl)ethyl pent-4-ynoate

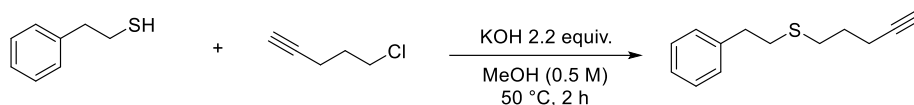


Compound **S30** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (4.51 g, 92% yield).

¹H NMR (400 MHz, CDCl₃) δ 5.33-5.24 (m, 1H), 4.15-4.05 (m, 2H), 2.55-2.45 (m, 4H), 2.35 (dt, *J* = 8.5, 5.6 Hz, 1H), 2.30-2.25 (m, 2H), 2.24-2.09 (m, 2H), 2.09-1.99 (m, 2H), 1.96 (t, *J* = 2.4 Hz, 1H), 1.26 (s, 3H), 1.13 (d, *J* = 8.6 Hz, 1H), 0.81 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 171.81, 131.35, 124.55, 82.52, 68.96, 63.28, 36.95, 35.42, 33.43, 29.44, 25.70, 25.37, 19.38, 17.64, 14.40.

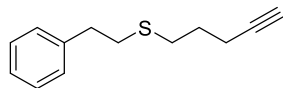
HRMS (APCI) calcd. for C₁₆H₂₃O₂⁺ [M+H]⁺: 247.1693; found: 247.1689.



KOH (2.47 g, 44 mmol) was dissolved in MeOH (40 mL) in a 100 mL round bottom flask with a magnetic stir bar. 2-phenylethane-1-thiol (2.7 mL, 20 mmol) was added to the solution, and the resulting mixture

was stirring at room temperature for 10 min. The 5-chloropent-1-yne (2.3 mL, 22 mmol) was then added to the solution. After stirring at 50 °C for 2 h, the reaction was concentrated in vacuo. Water (20 mL) was added to the reaction mixture, followed by neutralization with HCl aq. (3M). The solution was extracted with DCM (2 × 30 mL). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by silica gel flash chromatography to give the products².

pent-4-yn-1-yl(phenethyl)sulfane

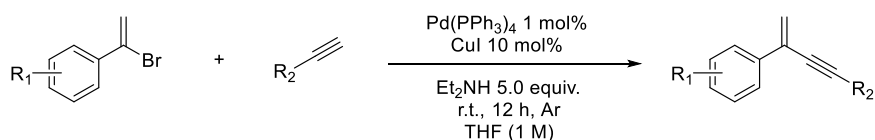


Compound **S31** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (3.15 g, 77% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.33-7.27 (m, 2H), 7.25-7.17 (m, 3H), 2.90 (dd, *J* = 9.7, 6.4 Hz, 2H), 2.78 (dd, *J* = 9.0, 5.9 Hz, 2H), 2.65 (t, *J* = 7.2 Hz, 2H), 2.32 (td, *J* = 6.9, 2.6 Hz, 2H), 1.97 (t, *J* = 2.6 Hz, 1H), 1.86-1.76 (m, 2H).

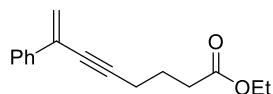
¹³C NMR (101 MHz, CDCl₃) δ 140.57, 128.49, 126.37, 83.53, 68.94, 36.36, 33.61, 30.99, 28.32, 17.51.

HRMS (APCI) calcd. for C₁₃H₁₇S⁺ [M+H]⁺: 205.1046; found: 205.1041.



An oven-dried 100 mL Schlenk tube, which was equipped with a magnetic stir bar and charged with Pd (PPh₃)₄ (116 mg, 1 mol%) and CuI (190 mg, 10 mol%). Then aryl bromide (1.1 equiv., 11 mmol), substituted alkyne (1.0 equiv., 10 mmol), and Et₂NH (5.2 mL, 50 mmol) were added under Ar. After stirring at room temperature for 12 h, the reaction mixture was quenched by sat. aq. NH₄Cl (40 mL). Then the solution was extracted with ethyl acetate (2 × 50 mL). The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by silica gel flash chromatography to give the products^{3,4}.

ethyl 7-phenyloct-7-en-5-ynoate



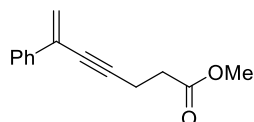
Compound **S2** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (1.98 g, 82% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.77-7.56 (m, 2H), 7.41-7.33 (m, 2H), 7.33-7.26 (m, 1H), 5.87 (s, 1H), 5.61 (s, 1H), 4.15 (q, *J* = 7.2 Hz, 2H), 2.50 (dd, *J* = 8.4, 6.8 Hz, 4H), 1.98-1.90 (m, 2H), 1.27 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 173.14, 137.63, 130.80, 128.32, 128.20, 126.04, 119.79, 90.52, 80.61, 60.40, 33.21, 23.98, 18.89, 14.26.

HRMS (APCI) calcd. for C₁₆H₁₉O₂⁺ [M+H]⁺: 243.1380; found: 243.1378.

methyl 6-phenylhept-6-en-4-ynoate



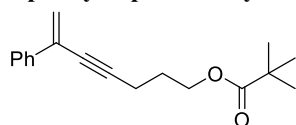
Compound **S3** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (1.59 g, 74% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.63 (d, *J* = 7.0 Hz, 2H), 7.35 (t, *J* = 7.1 Hz, 2H), 7.30 (d, *J* = 6.7 Hz, 1H), 5.87 (s, 1H), 5.60 (s, 1H), 3.72 (s, 3H), 2.75 (t, *J* = 7.6 Hz, 2H), 2.65 (t, *J* = 7.0 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 172.38, 137.46, 130.58, 128.31, 128.22, 126.03, 120.00, 89.54, 80.46, 51.85, 33.44, 15.42.

HRMS (APCI) calcd. for C₁₄H₁₅O₂⁺ [M+H]⁺: 215.1067; found: 215.1065.

6-phenylhept-6-en-4-yn-1-yl pivalate



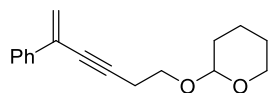
Compound **S4** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (1.45 g, 66% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.73-7.56 (m, 2H), 7.40-7.28 (m, 3H), 5.86 (s, 1H), 5.60 (s, 1H), 4.22 (t, *J* = 6.3 Hz, 2H), 2.53 (t, *J* = 7.1 Hz, 2H), 2.00-1.91 (m, 2H), 1.22 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 178.47, 137.63, 130.78, 128.32, 128.18, 126.04, 119.84, 90.20, 80.46, 63.07, 38.81, 27.92, 27.22, 16.21.

HRMS (APCI) calcd. for C₁₈H₂₃O₂⁺ [M+H]⁺: 271.1693; found: 271.1690.

2-((5-phenylhex-5-en-3-yn-1-yl)oxy)tetrahydro-2H-pyran



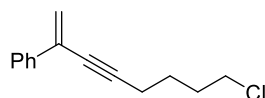
Compound **S5** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (1.23 g, 48% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.75-7.54 (m, 2H), 7.37-7.28 (m, 3H), 5.86 (s, 1H), 5.60 (s, 1H), 4.70 (t, *J* = 3.6 Hz, 1H), 3.95-3.88 (m, 2H), 3.65 (dt, *J* = 9.6, 7.0 Hz, 1H), 3.55-3.49 (m, 1H), 2.73 (t, *J* = 7.0 Hz, 2H), 1.91-1.82 (m, 1H), 1.77-1.70 (m, 1H), 1.68-1.62 (m, 1H), 1.59-1.48 (m, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 137.56, 130.76, 128.26, 128.16, 126.06, 119.71, 98.76, 88.66, 80.67, 65.72, 62.13, 30.60, 25.46, 20.96, 19.35.

HRMS (APCI) calcd. for C₁₇H₂₁O₂⁺ [M+H]⁺: 257.1536; found: 219.1506.

(8-chlorooct-1-en-3-yn-2-yl)benzene



Compound **S8** was isolated via column chromatography (PE/EA=80:1~40:1) as pale yellow oil (700 mg, 32% yield).

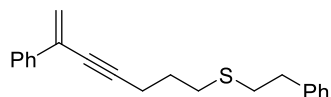
¹H NMR (400 MHz, CDCl₃) δ 7.66 (d, *J* = 7.3 Hz, 2H), 7.37 (t, *J* = 7.2 Hz, 2H), 7.32 (d, *J* = 7.0 Hz,

1H), 5.87 (s, 1H), 5.61 (s, 1H), 3.61 (t, $J = 6.5$ Hz, 2H), 2.49 (t, $J = 6.9$ Hz, 2H), 2.00-1.94 (m, 2H), 1.82-1.76 (m, 2H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 137.68, 130.83, 128.33, 128.20, 126.05, 119.73, 90.87, 80.44, 44.56, 31.69, 25.92, 18.76.

HRMS (APCI) calcd. for $\text{C}_{14}\text{H}_{16}\text{Cl}^+$ $[\text{M}+\text{H}]^+$: 219.0936; found: 219.0933.

phenethyl(6-phenylhept-6-en-4-yn-1-yl)sulfane



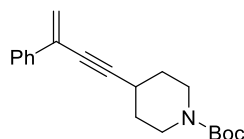
Compound **S11** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (2.39 g, 78% yield).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.78-7.58 (m, 2H), 7.39-7.30 (m, 5H), 7.26-7.22 (m, 3H), 5.89 (s, 1H), 5.63 (s, 1H), 2.94 (t, $J = 7.7$ Hz, 2H), 2.86-2.81 (m, 2H), 2.76-2.72 (m, 2H), 2.59 (td, $J = 7.0, 2.0$ Hz, 2H), 1.97-1.89 (m, 2H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 140.60, 137.71, 130.86, 128.53, 128.37, 128.24, 126.41, 126.09, 119.81, 90.77, 80.58, 36.41, 33.73, 31.30, 28.68, 18.60.

HRMS (APCI) calcd. for $\text{C}_{21}\text{H}_{23}\text{S}^+$ $[\text{M}+\text{H}]^+$: 307.1515; found: 307.1509.

tert-butyl 4-(3-phenylbut-3-en-1-yn-1-yl)piperidine-1-carboxylate



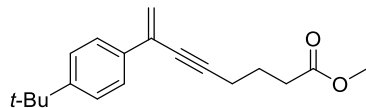
Compound **S15** was isolated via column chromatography (PE/EA=20:1~10:1) as pale yellow oil (2.03 g, 65% yield).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.63 (d, $J = 7.2$ Hz, 2H), 7.36-7.32 (m, 2H), 7.29 (d, $J = 6.9$ Hz, 1H), 5.86 (s, 1H), 5.59 (s, 1H), 3.73 (dt, $J = 10.6, 3.0$ Hz, 2H), 3.29-3.21 (m, 2H), 2.80 (tt, $J = 7.9, 4.0$ Hz, 1H), 1.89-1.81 (m, 2H), 1.73-1.64 (m, 2H), 1.46 (s, 9H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 154.77, 137.58, 130.65, 128.33, 128.21, 125.99, 119.88, 93.38, 81.16, 79.48, 42.27, 31.44, 28.47, 27.64.

HRMS (APCI) calcd. for $\text{C}_{20}\text{H}_{26}\text{ON}_2^+$ $[\text{M}+\text{H}]^+$: 312.1958; found: 312.1931.

methyl 7-(4-(tert-butyl)phenyl)oct-7-en-5-ynoate



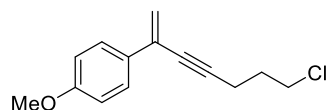
Compound **S17** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (2.35 g, 50% yield).

$^1\text{H NMR}$ (500 MHz, $\text{CHLOROFORM-}D$) δ 7.63-7.54 (m, 2H), 7.40-7.35 (m, 2H), 5.83 (s, 1H), 5.56 (s, 1H), 3.69 (s, 3H), 2.51 (dt, $J = 9.8, 7.2$ Hz, 4H), 1.98-1.91 (m, 2H), 1.34 (s, 9H).

$^{13}\text{C NMR}$ (126 MHz, $\text{CHLOROFORM-}D$) δ 173.69, 151.36, 134.93, 130.58, 125.83, 125.32, 119.15, 90.21, 80.87, 51.68, 34.66, 33.03, 31.37, 24.03, 18.97.

HRMS (APCI) calcd. for $\text{C}_{19}\text{H}_{25}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 285.1850; found: 285.1842.

1-(7-chlorohept-1-en-3-yn-2-yl)-4-methoxybenzene



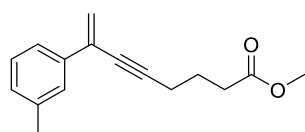
Compound **S18** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (1.51 g, 64% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.60 (d, *J* = 8.9 Hz, 2H), 6.90 (d, *J* = 8.9 Hz, 2H), 5.78 (s, 1H), 5.52 (s, 1H), 3.85 (s, 3H), 3.73 (t, *J* = 6.4 Hz, 2H), 2.64 (t, *J* = 6.8 Hz, 2H), 2.13-2.04 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 159.72, 130.22, 129.99, 127.25, 118.05, 113.67, 89.33, 80.97, 55.33, 43.77, 31.45, 16.87.

HRMS (APCI) calcd. for C₁₄H₁₆ClO⁺ [M+H]⁺: 235.0885; found: 235.0882.

methyl 7-(*m*-tolyl)oct-7-en-5-ynoate



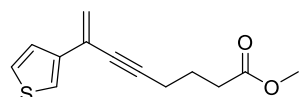
Compound **S19** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (1.84 g, 76% yield).

¹H NMR (500 MHz, CHLOROFORM-*D*) δ 7.44 (d, *J* = 8.2 Hz, 2H), 7.24 (t, *J* = 7.5 Hz, 1H), 7.12 (d, *J* = 7.5 Hz, 1H), 5.84 (s, 1H), 5.58 (s, 1H), 3.69 (s, 3H), 2.51 (dt, *J* = 8.8, 7.2 Hz, 4H), 2.38 (s, 3H), 1.97-1.91 (m, 2H).

¹³C NMR (126 MHz, CHLOROFORM-*D*) δ 173.70, 137.96, 137.66, 130.89, 129.03, 128.29, 126.83, 123.24, 119.81, 90.33, 80.84, 51.69, 33.00, 24.00, 21.55, 18.97.

HRMS (APCI) calcd. for C₁₆H₁₉O₂⁺ [M+H]⁺: 243.1380; found: 243.1376.

methyl 7-(thiophen-3-yl)oct-7-en-5-ynoate



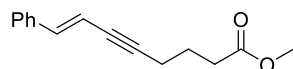
Compound **S21** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (1.06 g, 43% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.44 (t, *J* = 2.1 Hz, 1H), 7.26-7.23 (m, 2H), 5.72 (s, 1H), 5.48 (s, 1H), 3.66 (s, 3H), 2.50-2.44 (m, 4H), 1.94-1.88 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 173.50, 140.19, 126.03, 125.63, 124.80, 123.14, 118.53, 89.29, 80.57, 51.58, 32.90, 23.92, 18.79.

HRMS (APCI) calcd. for C₁₃H₁₅O₂S⁺ [M+H]⁺: 235.0788; found: 235.0784.

methyl (*E*)-8-phenyloct-7-en-5-ynoate



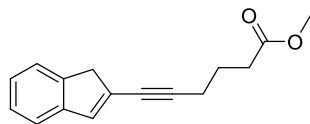
Compound **S22** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (1.89 g, 83% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.38 (d, *J* = 7.8 Hz, 2H), 7.34 (t, *J* = 7.2 Hz, 2H), 7.31-7.26 (m, 1H), 6.90

(d, $J = 16.2$ Hz, 1H), 6.17 (dt, $J = 16.3, 2.3$ Hz, 1H), 3.71 (s, 3H), 2.54-2.45 (m, 4H), 1.96-1.88 (m, 2H).
 $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 173.61, 140.41, 136.47, 128.68, 128.34, 126.10, 108.59, 91.35, 80.66, 51.60, 32.91, 23.96, 19.13.

HRMS (APCI) calcd. for $\text{C}_{15}\text{H}_{17}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 229.1224; found: 229.1218.

methyl 6-(1H-inden-2-yl)hex-5-ynoate



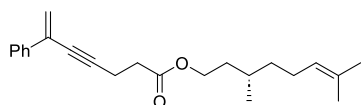
Compound **S23** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (1.78 g, 74% yield).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.38 (d, $J = 7.3$ Hz, 1H), 7.33 (d, $J = 7.4$ Hz, 1H), 7.25 (t, $J = 7.4$ Hz, 1H), 7.19 (td, $J = 7.3, 1.2$ Hz, 1H), 6.96 (s, 1H), 3.68 (s, 3H), 3.47 (s, 2H), 2.50 (t, $J = 6.9$ Hz, 4H), 1.91 (q, $J = 7.2$ Hz, 2H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 173.60, 144.26, 142.66, 136.13, 127.91, 126.67, 125.40, 123.47, 121.12, 93.85, 78.62, 51.61, 42.87, 32.94, 23.93, 19.29.

HRMS (APCI) calcd. for $\text{C}_{16}\text{H}_{17}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 241.1224; found: 241.1219.

(S)-3,7-dimethyloct-6-en-1-yl 6-phenylhept-6-en-4-ynoate



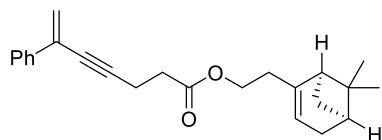
Compound **S25** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (2.49 g, 72% yield).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.63 (d, $J = 7.0$ Hz, 2H), 7.38-7.32 (m, 2H), 7.31-7.27 (m, 1H), 5.86 (s, 1H), 5.59 (s, 1H), 5.11-5.04 (m, 1H), 4.20-4.12 (m, 2H), 2.74 (t, $J = 7.2$ Hz, 2H), 2.63 (t, $J = 7.2$ Hz, 2H), 2.03-1.90 (m, 2H), 1.72-1.66 (m, 4H), 1.60 (s, 3H), 1.58-1.52 (m, 1H), 1.49-1.42 (m, 1H), 1.37-1.29 (m, 1H), 1.22-1.14 (m, 1H), 0.90 (d, $J = 6.5$ Hz, 3H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 171.97, 137.49, 131.36, 130.63, 128.28, 128.18, 126.03, 124.57, 119.93, 89.61, 80.43, 63.32, 36.97, 35.44, 33.74, 29.49, 25.72, 25.38, 19.40, 17.66, 15.45.

HRMS (APCI) calcd. for $\text{C}_{23}\text{H}_{31}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 339.2319; found: 339.2315.

2-((1R,5S)-6,6-dimethylbicyclo[3.1.1]hept-2-en-2-yl)ethyl 6-phenylhept-6-en-4-ynoate



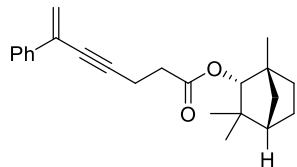
Compound **S26** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (2.03 g, 65% yield).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.63 (d, $J = 6.8$ Hz, 2H), 7.44-7.32 (m, 2H), 7.32-7.27 (m, 1H), 5.86 (s, 1H), 5.59 (s, 1H), 5.32-5.28 (m, 1H), 4.13 (td, $J = 7.0, 4.4$ Hz, 2H), 2.73 (t, $J = 7.3$ Hz, 2H), 2.64-2.59 (m, 2H), 2.39-2.33 (m, 1H), 2.29 (t, $J = 7.0$ Hz, 2H), 2.25-2.16 (m, 2H), 2.10-2.02 (m, 2H), 1.27 (s, 3H), 1.14 (d, $J = 8.5$ Hz, 1H), 0.82 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 171.86, 144.02, 137.50, 130.63, 128.29, 128.19, 126.04, 119.95, 118.87, 89.63, 80.42, 62.99, 45.67, 40.73, 38.00, 35.93, 33.70, 31.66, 31.36, 26.28, 21.12, 15.41.

HRMS (APCI) calcd. for $\text{C}_{24}\text{H}_{29}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 349.2163; found: 349.2133.

(1*R*,2*R*,4*S*)-1,3,3-trimethylbicyclo[2.2.1]heptan-2-yl 6-phenylhept-6-en-4-ynoate



Compound **S27** was isolated via column chromatography (PE/EA=50:1~20:1) as pale yellow oil (2.68 g, 80% yield).

^1H NMR (400 MHz, CDCl_3) δ 7.63 (d, $J = 7.1$ Hz, 2H), 7.37-7.32 (m, 2H), 7.30 (d, $J = 6.9$ Hz, 1H), 5.85 (s, 1H), 5.58 (s, 1H), 4.42 (d, $J = 2.0$ Hz, 1H), 2.82-2.73 (m, 2H), 2.71-2.64 (m, 2H), 1.78-1.67 (m, 3H), 1.61-1.56 (m, 1H), 1.49-1.40 (m, 1H), 1.19 (dd, $J = 10.3, 1.6$ Hz, 1H), 1.11 (s, 4H), 1.04 (s, 3H), 0.79 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 172.22, 137.52, 130.68, 128.28, 128.16, 126.06, 119.96, 89.75, 86.60, 80.43, 48.34, 48.30, 41.37, 39.44, 33.84, 29.70, 26.64, 25.83, 20.17, 19.37, 15.58.

HRMS (APCI) calcd. for $\text{C}_{23}\text{H}_{29}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 337.2163; found: 337.2159.

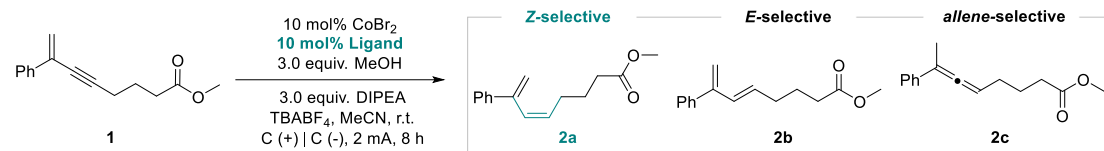
3. Optimization of Reaction Conditions for Hydrogenation

General procedure

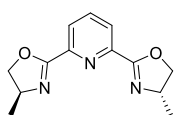
In the glovebox, a 15 mL tube equipped with a magnetic stirrer was charged with electrolyte, ligand, cobalt salt, then solvent was added. The resulting solution was stirred for 10 min at room temperature. substrate **1**, proton source and other reagent were added sequentially to the tube. The electrochemical cell was then sealed, brought out from the glove box. The resulting solution was electrolyzed under a constant current. Yields, *Z/E* and r.r. value were determined by GC using diphenyl as the internal standard.

Optimization of reaction condition for *Z*-selective hydrogenation

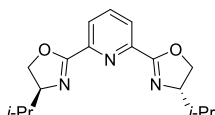
Table S1 Screening of ligands



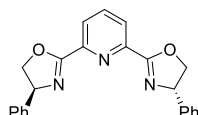
Entry	Ligand	Yield(%)	<i>Z/E</i> (2a:2b)	r.r.(2a+2b:2c)
1	L1	37	12:1	9:1
2	L2	35	15:1	18:1
3	L3	39	17:1	>20:1
4	L4	36	>20:1	18:1
5	L8	24	10:1	1:1
6	L9	22	8:1	15:1
7	L10	36	4:1	18:1
8	L11	n.p.	-	-
9	L12	n.p.	-	-
10	L13	trace	-	-
11	L14	trace	-	-
12	L15	trace	-	-



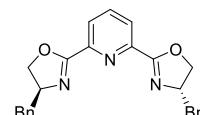
L1



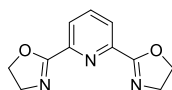
L2



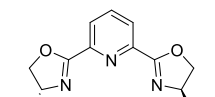
L3



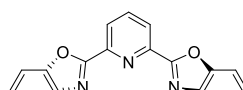
L4



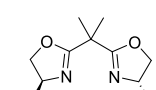
L8



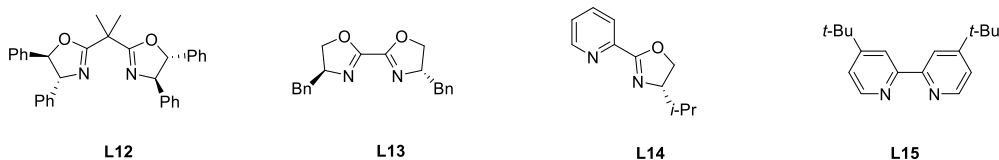
L9



L10

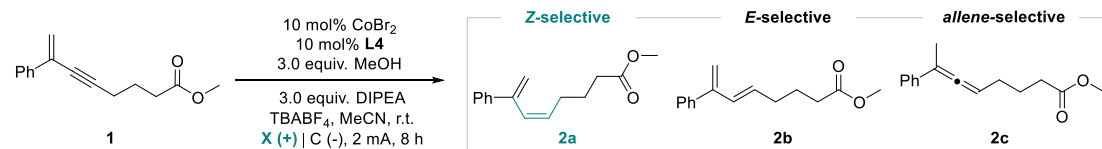


L11



Reaction Conditions: substrate **1** (0.2 mmol), CoBr₂ (10 mol%), Ligand (10 mol%), MeOH (3.0 equiv.), DIPEA (3.0 equiv.), TBABF₄ (0.1 M), Ar, 2 mA, undivided cell, C felt (+) | C felt (-), in MeCN (5 mL) at r.t. for 8 h. Yields were determined by GC using diphenyl as the internal standard, *Z/E* and r.r. value were determined by GC analysis.

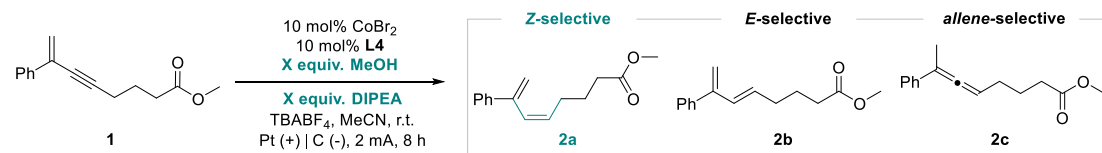
Table S2 Screening of electrode



Entry	Anode	Yield(%)	<i>Z/E</i> (2a:2b)	r.r.(2a+2b:2c)
1	C felt	36	>20:1	18:1
2	Pt	65	>20:1	>20:1
3	RVC	19	>20:1	2:1
4	SS	60	-	1:17
5	Ni foam	trace	-	-
6	Zn, no DIPEA	messy	-	-

Reaction Conditions: substrate **1** (0.2 mmol), CoBr₂ (10 mol%), L4 (10 mol%), MeOH (3.0 equiv.), DIPEA (3.0 equiv.), TBABF₄ (0.1 M), Ar, 2 mA, undivided cell, Anode (+) | C felt (-), in MeCN (5 mL) at r.t. for 8 h. Yields were determined by GC using diphenyl as the internal standard, *Z/E* and r.r. value were determined by GC analysis.

Table S3 Screening of the amount of DIPEA and MeOH

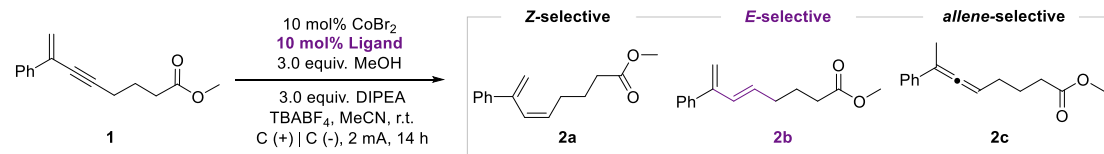


Entry	X equiv.	Yield(%)	<i>Z/E</i> (2a:2b)	r.r.(2a+2b:2c)
1	3.0	65	>20:1	>20:1
2	4.5	77	>20:1	>20:1
3	6.0	73	>20:1	>20:1
4	10.0	69	>20:1	>20:1

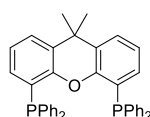
Reaction Conditions: substrate **1** (0.2 mmol), CoBr₂ (10 mol%), L4 (10 mol%), MeOH (X equiv.), DIPEA (X equiv.), TBABF₄ (0.1 M), Ar, 2 mA, undivided cell, Pt (+) | C felt (-), in MeCN (5 mL) at r.t. for 8 h. Yields were determined by GC using diphenyl as the internal standard, *Z/E* and r.r. value were determined by GC analysis.

Optimization of reaction condition for *E*-selective hydrogenation

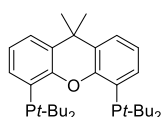
Table S4 Screening of ligands



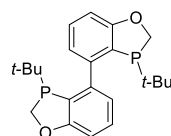
Entry	Ligand	Yield(%)	<i>E/Z</i> (2b:2a)	r.r.(2a+2b:2c)
1	L5	13	5:1	12:1
2	L6	6	2:1	5:1
3	L7	25	1.5:1	12:1
4	L16	trace	-	-
5	L17	11	2:1	4:1
6	L18	trace	-	-
7	L19	trace	-	-
8 ^a	L5	27	-	<1:20
9^a	L7	40	14:1	16:1



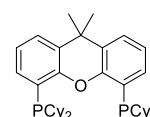
L5



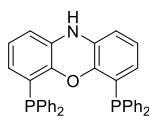
L6



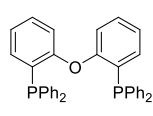
L7



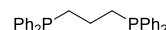
L16



L17



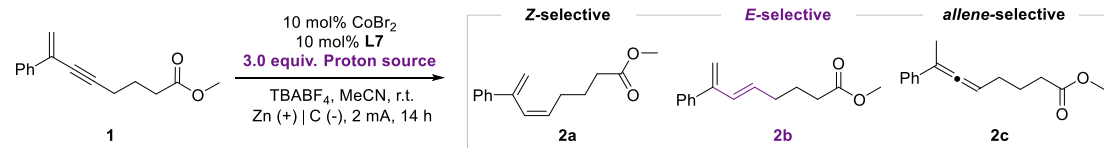
L18



L19

Reaction Conditions: substrate **1** (0.2 mmol), CoBr₂ (10 mol%), Ligand (10 mol%), MeOH (3.0 equiv.), DIPEA (3.0 equiv.), TBABF₄ (0.1 M), Ar, 2 mA, undivided cell, C felt (+) | C felt (-), in MeCN (5 mL) at r.t. for 8 h. Yields were determined by GC using diphenyl as the internal standard, *E/Z* and r.r. value were determined by GC analysis. ^aZn (+) instead of C felt (+), no DIPEA.

Table S5 Screening of proton sources

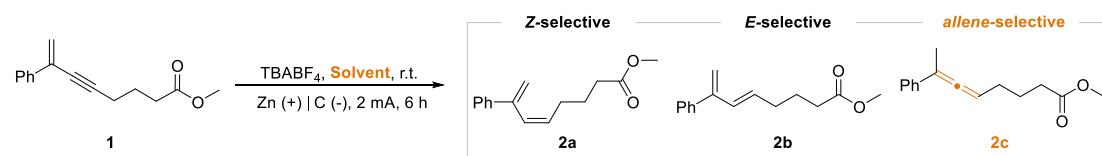


Entry	Proton source	Yield(%)	<i>E/Z</i> (2b:2a)	r.r.(2a+2b:2c)
1	MeOH	40	14:1	16:1
2	4-tert-Butylphenol	54	18:1	8:1
3	4-Trifluoromethylphenol	27	7:1	4:1
4	HOAc	61	2:1	5:1
5	HFIP	28	>20:1	5:1
6	CF ₃ CH ₂ OH	71	20:1	13:1

Reaction Conditions: substrate **1** (0.2 mmol), CoBr₂ (10 mol%), L7 (10 mol%), Proton source (3.0 equiv.), TBABF₄ (0.1 M), Ar, 2 mA, undivided cell, Zn (+) | C felt (-), in MeCN (5 mL) at r.t. for 14 h. Yields were determined by GC using diphenyl as the internal standard, *E/Z* and r.r. value were determined by GC analysis.

Optimization of reaction condition for *allene*-selective hydrogenation

Table S6 Screening of solvents



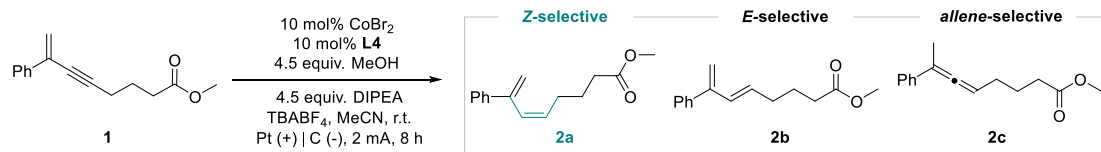
Entry	Solvent	Yield(%)	r.r.(2c:2a+2b)
1	Acetone	50	>20:1
2	MeCN	85	17:1
3	DMA	43	19:1
4	NMP	44	>20:1
5	THF	52	17:1
6 ^a	MeCN	95	>20:1

Reaction Conditions: substrate **1** (0.2 mmol), MeOH (2.0 equiv.), TBABF₄ (0.1 M), Ar, 2 mA, undivided cell, Zn (+) | C felt (-), in MeCN (5 mL) at r.t. for 6 h. Yields were determined by GC using diphenyl as the internal standard, r.r. value was determined by GC analysis.

^astainless steel felt (-) instead of C (-)

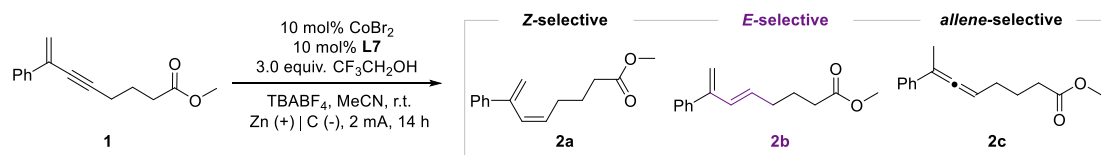
Reaction procedures of hydrogenation

Z condition:



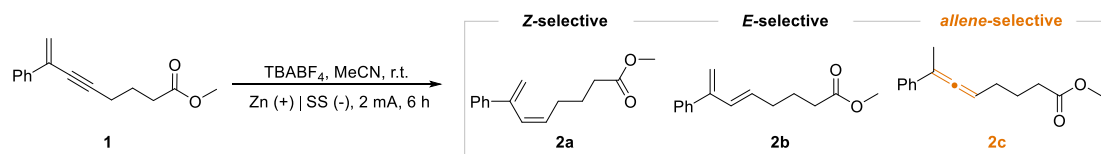
Carbon felt electrodes were polished with 600 grit silicon carbide sandpaper before use. In the glovebox, a 15 mL dried tube equipped with a magnetic stirrer was charged with TBABF₄ (165mg, 0.5 mmol), L4 (7.9 mg, 0.02 mmol), CoBr₂ (4.3 mg, 0.02 mmol), then 5 mL MeCN was added, the resulting solution was stirred for 10 min at room temperature. Substrate 1 (41 μ L, 0.2 mmol), MeOH (37 μ L, 0.9 mmol) and DIPEA (157 μ L, 0.9 mmol) were added sequentially to the tube. The tube was sealed with the pre-assembled electrochemical cap, equipped with anode (Pt felt, 25 \times 10 \times 1 mm) and cathode (carbon felt, 25 \times 10 \times 1 mm), and then the assembly was brought out of the glove box. The resulting solution was electrolyzed under a constant current of 2 mA for 8 h at room temperature. Once reaction was completed, the reaction solvent was concentrated in vacuo. The resulting residue was purified by silica gel flash chromatography to give the products.

E condition:



Carbon felt electrodes were polished with 600 grit silicon carbide sandpaper before use. In the glovebox, a 15 mL dried tube equipped with a magnetic stirrer was charged with TBABF₄ (165mg, 0.5 mmol), L7 (7.7 mg, 0.02 mmol), CoBr₂ (4.3 mg, 0.02 mmol), then 5 mL MeCN was added, the resulting solution was stirred for 10 min at room temperature. Substrate 1 (41 μ L, 0.2 mmol) and CF₃CH₂OH (43 μ L, 0.6 mmol) were added sequentially to the tube. The tube was sealed with the pre-assembled electrochemical cap, equipped with anode (Zn felt, 25 \times 10 \times 1 mm) and cathode (carbon felt, 25 \times 10 \times 1 mm), and then the assembly was brought out of the glove box. The resulting solution was electrolyzed under a constant current of 2 mA for 14 h at room temperature. Once reaction was completed, the reaction solvent was concentrated in vacuo. The resulting residue was purified by silica gel flash chromatography to give the products.

Allene condition:

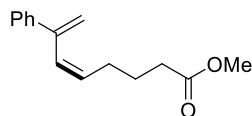


Carbon felt electrodes were polished with 600 grit silicon carbide sandpaper before use. In the glovebox, a 15 mL dried tube equipped with a magnetic stirrer was charged with TBABF₄ (165mg, 0.5 mmol), then 5 mL MeCN was added, the resulting solution was stirred for 10 min at room temperature. Substrate 1 (41 μ L, 0.2 mmol) and MeOH (16 μ L, 0.4 mmol) were added sequentially to the tube. The tube was sealed with the pre-assembled electrochemical cap, equipped with anode (Zn felt, 25 \times 10 \times 1 mm) and

cathode (stainless steel felt, 25×10×1 mm), and then the assembly was brought out of the glove box. The resulting solution was electrolyzed under a constant current of 2 mA for 6 h at room temperature. Once reaction was completed, the reaction solvent was concentrated in vacuo. The resulting residue was purified by silica gel flash chromatography to give the products.

4. Characterization Data of Products

methyl (Z)-7-phenylocta-5,7-dienoate



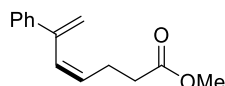
Compound **2a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (34.5 mg, 75% yield, *Z:E* >20:1, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.40 (d, *J* = 6.8 Hz, 2H), 7.32 (t, *J* = 7.7 Hz, 2H), 7.29 (d, *J* = 8.6 Hz, 1H), 6.19 (d, *J* = 11.5 Hz, 1H), 5.68 (dt, *J* = 11.5, 7.3 Hz, 1H), 5.54 (s, 1H), 5.13 (s, 1H), 3.63 (s, 3H), 2.28 (t, *J* = 7.6 Hz, 2H), 2.17 (t, *J* = 7.5 Hz, 2H), 1.75-1.68 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 173.97, 144.33, 140.67, 133.03, 129.90, 128.26, 127.58, 126.53, 114.90, 51.46, 33.58, 27.98, 25.05.

HRMS (APCI) calcd. for C₁₅H₁₉O₂⁺ [M+H]⁺: 231.1380; found: 231.1372.

methyl (Z)-6-phenylhepta-4,6-dienoate



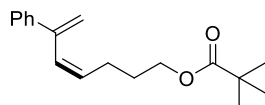
Compound **3a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (32.4 mg, 72% yield, *Z:E* = 17:1, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, *J* = 8.5 Hz, 2H), 7.33 (t, *J* = 7.8 Hz, 2H), 7.29 (d, *J* = 6.8 Hz, 1H), 6.20 (d, *J* = 11.4 Hz, 1H), 5.69 (dt, *J* = 11.4, 7.2 Hz, 1H), 5.58 (s, 1H), 5.16 (s, 1H), 3.65 (s, 3H), 2.46 (t, *J* = 7.5 Hz, 2H), 2.38 (t, *J* = 7.2 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 173.46, 144.19, 140.53, 131.81, 130.40, 128.36, 127.71, 126.57, 115.12, 51.60, 34.24, 24.17.

HRMS (APCI) calcd. for C₁₄H₁₇O₂⁺ [M+H]⁺: 217.1224; found: 217.1217.

(Z)-6-phenylhepta-4,6-dien-1-yl pivalate



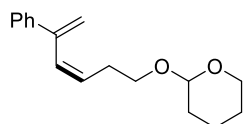
Compound **4a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (41.4 mg, 76% yield, *Z:E* >20:1, r.r. = 9:1).

¹H NMR (400 MHz, CDCl₃) δ 7.43-7.38 (m, 2H), 7.35-7.28 (m, 3H), 6.19 (dd, *J* = 11.5, 1.6 Hz, 1H), 5.71 (dt, *J* = 11.4, 7.3 Hz, 1H), 5.54 (d, *J* = 1.7 Hz, 1H), 5.14 (s, 1H), 4.03 (t, *J* = 6.5 Hz, 2H), 2.21 (qd, *J* = 7.5, 1.8 Hz, 2H), 1.74-1.68 (m, 2H), 1.15 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 178.55, 144.33, 140.64, 132.92, 129.86, 128.28, 127.60, 126.52, 114.83, 63.80, 38.70, 28.83, 27.22, 27.16, 25.12.

HRMS (APCI) calcd. for C₁₈H₂₅O₂⁺ [M+H]⁺: 273.1850; found: 273.1838.

(Z)-2-((5-phenylhexa-3,5-dien-1-yl)oxy)tetrahydro-2H-pyran



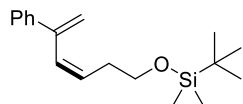
Compound **5a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (39.3 mg, 76% yield, *Z:E* >20:1, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.42 (dd, *J* = 8.2, 1.6 Hz, 2H), 7.36-7.28 (m, 3H), 6.23 (dd, *J* = 11.5, 1.7 Hz, 1H), 5.79 (dt, *J* = 11.6, 7.3 Hz, 1H), 5.55 (d, *J* = 1.7 Hz, 1H), 5.20 (s, 1H), 4.58 (t, *J* = 3.5 Hz, 1H), 3.85 (td, *J* = 8.1, 4.0 Hz, 1H), 3.79-3.74 (m, 1H), 3.51-3.46 (m, 1H), 3.46-3.41 (m, 1H), 2.46 (qd, *J* = 7.0, 1.8 Hz, 2H), 1.86-1.79 (m, 1H), 1.72-1.67 (m, 1H), 1.61-1.58 (m, 1H), 1.55-1.51 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 144.28, 140.65, 130.58, 130.46, 128.24, 127.56, 126.57, 114.96, 98.62, 66.98, 62.18, 30.67, 29.27, 25.49, 19.50.

HRMS (APCI) calcd. for C₁₇H₂₃O₂⁺ [M+H]⁺: 259.1693; found: 259.1685.

(Z)-tert-butyltrimethyl((5-phenylhexa-3,5-dien-1-yl)oxy)silane



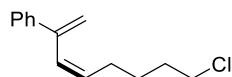
Compound **6a** was isolated via column chromatography (PE/EA=100:1~70:1) as pale yellow oil (35.2 mg, 61% yield, *Z:E* = 14:1, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, *J* = 6.7 Hz, 2H), 7.35-7.29 (m, 3H), 6.23 (d, *J* = 11.5 Hz, 1H), 5.77 (dt, *J* = 11.6, 7.3 Hz, 1H), 5.55 (s, 1H), 5.19 (s, 1H), 3.65 (t, *J* = 6.6 Hz, 2H), 2.37 (qd, *J* = 6.8, 1.8 Hz, 2H), 0.90 (s, 9H), 0.05 (s, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 144.41, 140.73, 130.59, 130.46, 128.24, 127.54, 126.58, 114.83, 62.99, 32.37, 25.97, 18.37, -5.26.

HRMS (APCI) calcd. for C₁₈H₂₉OSi⁺ [M+H]⁺: 289.1983; found: 289.1984.

(Z)-(8-chloroocta-1,3-dien-2-yl)benzene



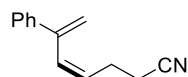
Compound **7a** was isolated via column chromatography (PE/EA=100:1~70:1) as pale yellow oil (21.2 mg, 48% yield, *Z:E* = 17:1, r.r. = 8:1).

¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, *J* = 8.5 Hz, 2H), 7.33 (t, *J* = 7.2 Hz, 2H), 7.29 (d, *J* = 7.0 Hz, 1H), 6.18 (dd, *J* = 11.5, 1.6 Hz, 1H), 5.69 (dt, *J* = 11.6, 7.4 Hz, 1H), 5.54 (d, *J* = 1.7 Hz, 1H), 5.14 (s, 1H), 3.48 (t, *J* = 6.7 Hz, 2H), 2.14 (qd, *J* = 7.5, 1.8 Hz, 2H), 1.77-1.71 (m, 2H), 1.54-1.50 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 144.47, 140.77, 133.53, 129.56, 128.28, 127.58, 126.55, 114.91, 44.84, 32.13, 27.76, 26.96.

HRMS (APCI) calcd. for C₁₄H₁₈Cl⁺ [M+H]⁺: 221.1092; found: 221.1088.

(Z)-6-phenylhepta-4,6-dienenitrile



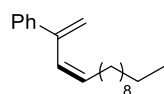
Compound **8a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (22.7 mg, 62% yield, *Z:E* >20:1, r.r. = 5:1).

¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, *J* = 8.6 Hz, 2H), 7.34 (d, *J* = 7.7 Hz, 2H), 7.31 (d, *J* = 6.8 Hz, 1H), 6.35 (d, *J* = 11.4 Hz, 1H), 5.71 (dt, *J* = 11.5, 7.1 Hz, 1H), 5.59 (s, 1H), 5.17 (s, 1H), 2.47-2.43 (m, 2H), 2.38-2.34 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 143.92, 140.00, 132.41, 129.05, 128.45, 127.93, 126.53, 119.21, 115.69, 24.42, 17.51.

HRMS (APCI) calcd. for C₁₃H₁₄N⁺ [M+H]⁺: 184.1121; found: 184.1116.

(Z)-tetradeca-1,3-dien-2-ylbenzene



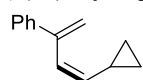
Compound **9a** was isolated via column chromatography (PE) as pale yellow oil (36.2 mg, 67% yield, *Z:E* = 14:1, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, *J* = 7.6 Hz, 2H), 7.35-7.29 (m, 3H), 6.12 (d, *J* = 11.5 Hz, 1H), 5.71 (dt, *J* = 11.5, 7.4 Hz, 1H), 5.53 (s, 1H), 5.14 (s, 1H), 2.11 (q, *J* = 7.3 Hz, 2H), 1.28-1.22 (m, 16H), 0.88 (t, *J* = 6.7 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 144.58, 140.97, 134.71, 128.71, 128.20, 127.47, 126.58, 114.64, 31.92, 29.85, 29.62, 29.47, 29.35, 29.31, 28.64, 22.70, 14.12.

HRMS (APCI) calcd. for C₂₀H₃₁⁺ [M+H]⁺: 271.2421; found: 271.2412.

(Z)-(4-cyclopropylbuta-1,3-dien-2-yl)benzene



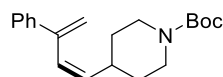
Compound **10a** was isolated via column chromatography (PE) as pale yellow oil (14.6 mg, 43% yield, *Z:E* >20:1, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.49 (d, *J* = 7.1 Hz, 2H), 7.39-7.31 (m, 3H), 6.09 (d, *J* = 11.4 Hz, 1H), 5.56 (s, 1H), 5.37 (s, 1H), 5.07 (t, *J* = 10.7 Hz, 1H), 1.67-1.57 (m, 1H), 0.73-0.69 (m, 2H), 0.43-0.40 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 145.05, 141.09, 138.53, 128.20, 127.49, 126.95, 126.84, 114.96, 11.51, 7.76.

HRMS (APCI) calcd. for C₁₃H₁₅⁺ [M+H]⁺: 171.1169; found: 171.1161.

tert-butyl (Z)-4-(3-phenylbuta-1,3-dien-1-yl)piperidine-1-carboxylate



Compound **11a** was isolated via column chromatography (PE/EA=20:1~10:1) as pale yellow oil (45.0 mg, 72% yield, *Z:E* >20:1, r.r. >20:1).

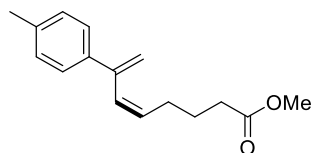
¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, *J* = 8.4 Hz, 2H), 7.35-7.29 (m, 3H), 6.11 (d, *J* = 11.4 Hz, 1H), 5.53-5.46 (m, 2H), 5.14 (s, 1H), 4.03 (s, 2H), 2.58 (t, *J* = 12.8 Hz, 2H), 2.48 (dt, *J* = 10.2, 3.8 Hz, 1H),

1.54 (d, $J = 10.6$ Hz, 2H), 1.45 (s, 9H), 1.36-1.28 (m, 2H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 154.86, 144.74, 140.51, 137.85, 128.57, 128.28, 127.70, 126.55, 114.45, 79.30, 43.50, 35.26, 32.09, 28.47.

HRMS (ESI) calcd. for $\text{C}_{20}\text{H}_{28}\text{NO}_2^+$ $[\text{M}+\text{H}]^+$: 314.2115; found: 314.2112.

methyl (Z)-7-(p-tolyl)octa-5,7-dienoate



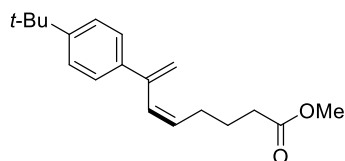
Compound **12a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (32.5 mg, 67% yield, $Z:E >20:1$, r.r. = 8:1).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.30 (d, $J = 7.9$ Hz, 2H), 7.13 (d, $J = 7.9$ Hz, 2H), 6.17 (d, $J = 11.4$ Hz, 1H), 5.67 (dt, $J = 11.3, 7.3$ Hz, 1H), 5.51 (s, 1H), 5.08 (s, 1H), 3.64 (s, 3H), 2.35 (s, 3H), 2.28 (t, $J = 7.6$ Hz, 2H), 2.17 (q, $J = 7.5$ Hz, 2H), 1.74-1.69 (m, 2H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 174.02, 144.05, 137.74, 137.38, 132.87, 130.06, 128.97, 126.39, 114.06, 51.47, 33.60, 27.97, 25.08, 21.14.

HRMS (APCI) calcd. for $\text{C}_{16}\text{H}_{21}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 245.1537; found: 245.1528.

methyl (Z)-7-(4-(tert-butyl)phenyl)octa-5,7-dienoate



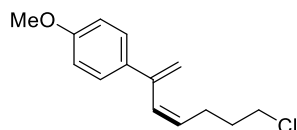
Compound **13a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (36.7 mg, 64% yield, $Z:E >20:1$, r.r. $>20:1$).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.36 (s, 4H), 6.18 (d, $J = 11.5$ Hz, 1H), 5.69 (dt, $J = 11.5, 7.3$ Hz, 1H), 5.54 (s, 1H), 5.09 (s, 1H), 3.64 (s, 3H), 2.29 (t, $J = 7.6$ Hz, 2H), 2.19 (q, $J = 7.8$ Hz, 2H), 1.75-1.69 (m, 2H), 1.33 (s, 9H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 174.03, 150.67, 143.83, 137.60, 132.87, 130.03, 126.14, 125.19, 114.07, 51.48, 34.54, 33.62, 31.33, 27.95, 25.10.

HRMS (APCI) calcd. for $\text{C}_{19}\text{H}_{27}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 287.2006; found: 287.1996.

(Z)-1-(7-chlorohepta-1,3-dien-2-yl)-4-methoxybenzene



Compound **14a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (27.9 mg, 59% yield, $Z:E = 14:1$, r.r. $>20:1$).

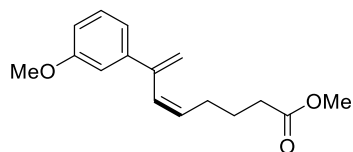
$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.35 (d, $J = 8.8$ Hz, 2H), 6.87 (d, $J = 8.7$ Hz, 2H), 6.20 (dd, $J = 11.4, 1.7$ Hz, 1H), 5.67 (dt, $J = 11.5, 7.3$ Hz, 1H), 5.48 (d, $J = 1.6$ Hz, 1H), 5.05 (s, 1H), 3.82 (s, 3H), 3.50 (t, $J = 6.8$ Hz, 2H), 2.29 (qd, $J = 7.5, 1.7$ Hz, 2H), 1.88-1.81 (m, 2H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 159.32, 143.54, 133.09, 132.01, 130.54, 127.65, 113.67, 113.32, 55.31,

44.48, 32.79, 26.01.

HRMS (APCI) calcd. for $C_{14}H_{18}ClO^+$ $[M+H]^+$: 237.1041; found: 237.1032.

methyl (Z)-7-(3-methoxyphenyl)octa-5,7-dienoate



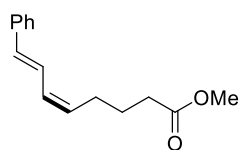
Compound **15a** was isolated via column chromatography (PE/EA=30:1~15:1) as pale yellow oil (32.3 mg, 62% yield, *Z:E* = 10:1, r.r. >20:1).

¹H NMR (400 MHz, $CDCl_3$) δ 7.23 (d, *J* = 8.0 Hz, 1H), 7.00 (d, *J* = 7.7 Hz, 1H), 6.93 (d, *J* = 2.5 Hz, 1H), 6.83 (dd, *J* = 8.3, 2.5 Hz, 1H), 6.17 (d, *J* = 11.5 Hz, 1H), 5.67 (dt, *J* = 11.4, 7.3 Hz, 1H), 5.54 (s, 1H), 5.13 (s, 1H), 3.82 (s, 3H), 3.63 (s, 3H), 2.28 (t, *J* = 7.6 Hz, 2H), 2.16 (q, *J* = 7.4 Hz, 2H), 1.74-1.68 (m, 2H).

¹³C NMR (101 MHz, $CDCl_3$) δ 173.98, 159.56, 144.19, 142.19, 133.10, 129.80, 129.24, 119.09, 115.11, 113.01, 112.30, 55.24, 51.48, 33.58, 27.97, 25.04.

HRMS (APCI) calcd. for $C_{16}H_{21}O_3^+$ $[M+H]^+$: 261.1486; found: 261.1481.

methyl (5Z,7E)-8-phenylocta-5,7-dienoate



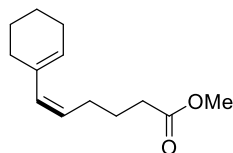
Compound **16a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (30.0 mg, 66% yield, *Z:E* >20:1, r.r. >20:1).

¹H NMR (400 MHz, $CDCl_3$) δ 7.42 (d, *J* = 7.3 Hz, 2H), 7.32 (t, *J* = 7.5 Hz, 2H), 7.22 (t, *J* = 7.3 Hz, 1H), 7.03 (ddd, *J* = 15.5, 11.0, 1.1 Hz, 1H), 6.54 (d, *J* = 15.6 Hz, 1H), 6.21 (t, *J* = 10.9 Hz, 1H), 5.49 (dt, *J* = 10.8, 7.7 Hz, 1H), 3.67 (s, 3H), 2.40-2.32 (m, 4H), 1.83-1.75 (m, 2H).

¹³C NMR (101 MHz, $CDCl_3$) δ 173.97, 137.54, 132.61, 131.44, 129.84, 128.60, 127.47, 126.38, 124.13, 51.50, 33.32, 27.22, 24.83.

HRMS (APCI) calcd. for $C_{15}H_{19}O_2^+$ $[M+H]^+$: 231.1380; found: 230.1374.

methyl (Z)-6-(cyclohex-1-en-1-yl)hex-5-enoate



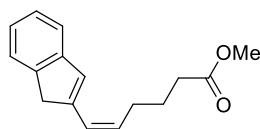
Compound **17a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (24.6 mg, 59% yield, *Z:E* >20:1, r.r. >20:1).

¹H NMR (400 MHz, $CDCl_3$) δ 5.77 (d, *J* = 11.8 Hz, 1H), 5.60 (s, 1H), 5.22 (dt, *J* = 11.7, 7.4 Hz, 1H), 3.66 (s, 3H), 2.32 (t, *J* = 7.6 Hz, 2H), 2.31-2.25 (m, 2H), 2.13-2.07 (m, 4H), 1.74-1.68 (m, 2H), 1.65-1.61 (m, 2H), 1.60-1.54 (m, 2H).

¹³C NMR (101 MHz, $CDCl_3$) δ 174.14, 135.26, 132.69, 128.03, 127.47, 51.49, 33.60, 28.99, 28.21, 25.60, 25.49, 22.89, 22.11.

HRMS (APCI) calcd. for $C_{13}H_{21}O_2^+$ $[M+H]^+$: 209.1537; found: 209.1535.

methyl (Z)-6-(1H-inden-2-yl)hex-5-enoate



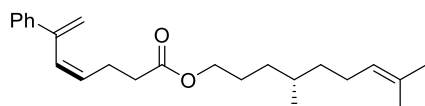
Compound **18a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (35.4 mg, 73% yield, *Z:E* = 12:1, r.r. >20:1).

¹H NMR (400 MHz, $CDCl_3$) δ 7.42 (d, *J* = 8.2 Hz, 1H), 7.35 (d, *J* = 7.5 Hz, 1H), 7.25 (t, *J* = 7.5 Hz, 1H), 7.16 (td, *J* = 7.4, 1.2 Hz, 1H), 6.76 (s, 1H), 6.36 (d, *J* = 11.5 Hz, 1H), 5.51 (dt, *J* = 11.6, 7.4 Hz, 1H), 3.68 (s, 3H), 3.60 (s, 2H), 2.49-2.43 (m, 2H), 2.41 (t, *J* = 7.5 Hz, 2H), 1.87-1.79 (m, 2H).

¹³C NMR (101 MHz, $CDCl_3$) δ 173.97, 144.91, 144.62, 143.42, 131.71, 131.14, 126.54, 125.71, 124.67, 123.52, 120.83, 51.58, 41.13, 33.56, 28.64, 25.07.

HRMS (APCI) calcd. for $C_{16}H_{19}O_2^+$ $[M+H]^+$: 243.1380; found: 243.1374.

(S)-4,8-dimethylnon-7-en-1-yl (Z)-6-phenylhepta-4,6-dienoate



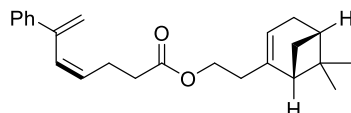
Compound **19a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (51.0 mg, 75% yield, *Z:E* >20:1, r.r. >20:1).

¹H NMR (400 MHz, $CDCl_3$) δ 7.40 (d, *J* = 8.1 Hz, 2H), 7.33 (t, *J* = 7.3 Hz, 2H), 7.29 (d, *J* = 7.0 Hz, 1H), 6.20 (dd, *J* = 11.4, 1.6 Hz, 1H), 5.70 (dt, *J* = 11.5, 7.2 Hz, 1H), 5.57 (d, *J* = 1.6 Hz, 1H), 5.16 (s, 1H), 5.11-5.07 (m, 1H), 4.12-4.06 (m, 2H), 2.48-2.43 (m, 2H), 2.38-2.33 (m, 2H), 2.02-1.93 (m, 2H), 1.69 (s, 3H), 1.61 (s, 3H), 1.56-1.50 (m, 1H), 1.47-1.28 (m, 3H), 1.21-1.14 (m, 1H), 0.90 (d, *J* = 6.6 Hz, 3H).

¹³C NMR (101 MHz, $CDCl_3$) δ 173.07, 144.13, 140.50, 131.87, 131.33, 130.24, 128.29, 127.63, 126.51, 124.59, 115.08, 62.99, 36.99, 35.45, 34.47, 29.51, 25.72, 25.40, 24.17, 19.42, 17.66.

HRMS (APCI) calcd. for $C_{23}H_{33}O_2^+$ $[M+H]^+$: 341.2476; found: 341.2467.

2-((1R,5S)-6,6-dimethylbicyclo[3.1.1]hept-2-en-2-yl)ethyl (Z)-6-phenylhepta-4,6-dienoate



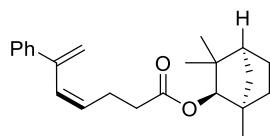
Compound **20a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (40.0 mg, 57% yield, *Z:E* = 12:1, r.r. >20:1).

¹H NMR (400 MHz, $CDCl_3$) δ 7.40 (d, *J* = 7.6 Hz, 2H), 7.33 (t, *J* = 7.2 Hz, 2H), 7.28 (d, *J* = 7.2 Hz, 1H), 6.19 (d, *J* = 11.4 Hz, 1H), 5.69 (dt, *J* = 11.4, 7.1 Hz, 1H), 5.56 (s, 1H), 5.27 (s, 1H), 5.15 (s, 1H), 4.06 (td, *J* = 6.8, 4.6 Hz, 2H), 2.44 (t, *J* = 7.3 Hz, 2H), 2.37-2.32 (m, 3H), 2.28-2.19 (m, 4H), 2.09-2.06 (m, 1H), 2.04-2.01 (m, 1H), 1.26 (s, 3H), 1.13 (d, *J* = 8.5 Hz, 1H), 0.81 (s, 3H).

¹³C NMR (101 MHz, $CDCl_3$) δ 172.99, 144.11, 144.09, 140.48, 131.89, 130.20, 128.29, 127.65, 126.50, 118.77, 115.06, 62.69, 45.66, 40.72, 38.00, 35.93, 34.46, 31.65, 31.36, 26.28, 24.14, 21.12.

HRMS (APCI) calcd. for $C_{24}H_{31}O_2^+$ $[M+H]^+$: 351.2319; found: 351.2311.

(1R,2R,4S)-1,3,3-trimethylbicyclo[2.2.1]heptan-2-yl (Z)-6-phenylhepta-4,6-dienoate



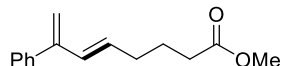
Compound **21a** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (39.0 mg, 58% yield, *Z:E* >20:1, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, *J* = 8.4 Hz, 2H), 7.33 (t, *J* = 7.1 Hz, 2H), 7.29 (d, *J* = 7.6 Hz, 1H), 6.21 (d, *J* = 11.5 Hz, 1H), 5.70 (dt, *J* = 11.8, 7.0 Hz, 1H), 5.55 (s, 1H), 5.17 (s, 1H), 4.35 (s, 1H), 2.48-2.42 (m, 2H), 2.41-2.36 (m, 2H), 1.73-1.66 (m, 3H), 1.62-1.52 (m, 2H), 1.49-1.41 (m, 1H), 1.19-1.15 (m, 1H), 1.09 (s, 3H), 1.01 (s, 3H), 0.73 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 173.38, 144.29, 140.65, 132.02, 130.15, 128.29, 127.63, 126.59, 115.31, 86.15, 48.34, 48.24, 41.36, 39.39, 34.57, 29.69, 26.64, 25.83, 24.44, 20.19, 19.39.

HRMS (APCI) calcd. for C₂₃H₃₁O₂⁺ [M+H]⁺: 339.2319; found: 339.2312.

methyl (E)-7-phenylocta-5,7-dienoate



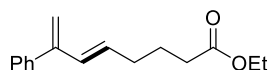
Compound **2b** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (32.7 mg, 71% yield, *E:Z* >20:1, r.r. = 13:1).

¹H NMR (400 MHz, CDCl₃) δ 7.35-7.29 (m, 5H), 6.32 (d, *J* = 15.6 Hz, 1H), 5.61 (dt, *J* = 15.8, 7.1 Hz, 1H), 5.20 (s, 1H), 5.08 (s, 1H), 3.66 (s, 3H), 2.32 (t, *J* = 7.5 Hz, 2H), 2.17 (q, *J* = 7.1 Hz, 2H), 1.78-1.70 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 173.99, 147.91, 140.53, 132.86, 132.40, 128.22, 128.08, 127.35, 115.06, 51.50, 33.45, 32.16, 24.36.

HRMS (APCI) calcd. for C₁₅H₁₉O₂⁺ [M+H]⁺: 231.1380; found: 231.1371.

ethyl (E)-7-phenylocta-5,7-dienoate



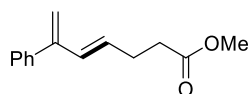
Compound **3b** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (28.0 mg, 57% yield, *E:Z* >20:1, r.r. = 8:1).

¹H NMR (400 MHz, CDCl₃) δ 7.37-7.27 (m, 5H), 6.32 (d, *J* = 15.6 Hz, 1H), 5.61 (dt, *J* = 14.9, 7.0 Hz, 1H), 5.19 (s, 1H), 5.08 (s, 1H), 4.12 (q, *J* = 7.1 Hz, 2H), 2.30 (t, *J* = 7.5 Hz, 2H), 2.17 (q, *J* = 7.3 Hz, 2H), 1.77-1.69 (m, 2H), 1.23 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 173.60, 147.92, 140.53, 132.96, 132.36, 128.26, 128.09, 127.37, 115.10, 60.29, 33.74, 32.20, 24.40, 14.27.

HRMS (APCI) calcd. for C₁₆H₂₀O₂⁺ [M+H]⁺: 245.1537; found: 245.1528.

methyl (E)-6-phenylhepta-4,6-dienoate



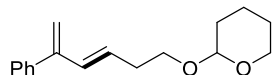
Compound **4b** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (28.5 mg, 66% yield, *E:Z* >20:1, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.34-7.28 (m, 5H), 6.34 (d, *J* = 15.7 Hz, 1H), 5.63 (dt, *J* = 15.6, 6.3 Hz, 1H), 5.21 (s, 1H), 5.10 (s, 1H), 3.67 (s, 3H), 2.47-2.41 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 173.42, 147.70, 140.37, 132.50, 131.72, 128.22, 128.11, 127.42, 115.49, 51.62, 33.68, 28.10.

HRMS (APCI) calcd. for C₁₄H₁₇O₂⁺ [M+H]⁺: 217.1224; found: 217.1220.

(*E*)-2-((5-phenylhexa-3,5-dien-1-yl)oxy)tetrahydro-2H-pyran



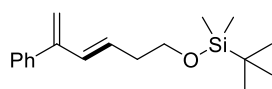
Compound **5b** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (29.8 mg, 58% yield, *E:Z* = 11:1, r.r. = 10:1).

¹H NMR (400 MHz, CDCl₃) δ 7.34-7.29 (m, 5H), 6.38 (d, *J* = 15.7 Hz, 1H), 5.66 (dt, *J* = 14.9, 7.0 Hz, 1H), 5.20 (s, 1H), 5.09 (s, 1H), 4.60 (t, *J* = 3.5 Hz, 1H), 3.86-3.81 (m, 1H), 3.79-3.73 (m, 1H), 3.51-3.48 (m, 1H), 3.47-3.43 (m, 1H), 1.84-1.78 (m, 1H), 1.73-1.68 (m, 1H), 1.60-1.49 (m, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 147.98, 140.50, 133.17, 130.48, 128.25, 128.04, 127.34, 115.11, 98.67, 66.79, 62.17, 33.28, 30.66, 25.50, 19.48.

HRMS (APCI) calcd. for C₁₇H₂₃O₂⁺ [M+H]⁺: 259.1693; found: 259.1687.

(*E*)-tert-butyltrimethyl((5-phenylhexa-3,5-dien-1-yl)oxy)silane



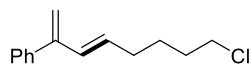
Compound **6b** was isolated via column chromatography (PE/EA=100:1~70:1) as pale yellow oil (35.3 mg, 62% yield, *E:Z* >20:1, r.r. = 8:1).

¹H NMR (400 MHz, CDCl₃) δ 7.33-7.29 (m, 5H), 6.36 (d, *J* = 15.7 Hz, 1H), 5.64 (dt, *J* = 15.5, 7.4 Hz, 1H), 5.20 (s, 1H), 5.08 (s, 1H), 3.65 (t, *J* = 6.8 Hz, 2H), 2.35 (q, *J* = 6.9 Hz, 2H), 0.88 (s, 9H), 0.03 (s, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 148.09, 140.55, 133.23, 130.65, 128.27, 128.23, 128.02, 127.29, 114.96, 62.80, 36.43, 25.92, 18.33, -5.29.

HRMS (APCI) calcd. for C₁₈H₂₉OSi⁺ [M+H]⁺: 289.1983; found: 289.1976.

(*E*)-(8-chloroocta-1,3-dien-2-yl)benzene



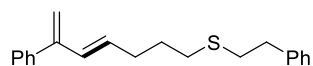
Compound **7b** was isolated via column chromatography (PE/EA=100:1~70:1) as pale yellow oil (18.1 mg, 41% yield, *E:Z* >20:1, r.r. = 5:1).

¹H NMR (400 MHz, CDCl₃) δ 7.35-7.29 (m, 5H), 6.32 (d, *J* = 15.5 Hz, 1H), 5.63 (dt, *J* = 15.8, 7.0 Hz, 1H), 5.19 (s, 1H), 5.08 (d, *J* = 1.9 Hz, 1H), 3.54 (t, *J* = 6.7 Hz, 2H), 2.16 (q, *J* = 7.9 Hz, 2H), 1.82-1.75 (m, 2H), 1.63-1.55 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 147.98, 140.59, 133.43, 132.05, 128.24, 128.08, 127.34, 114.94, 44.90, 32.10, 32.03, 26.41.

HRMS (APCI) calcd. for C₁₄H₁₈Cl⁺ [M+H]⁺: 221.1092; found: 221.1085.

(E)-phenethyl(6-phenylhepta-4,6-dien-1-yl)sulfane



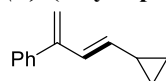
Compound **8b** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (33.9 mg, 55% yield, *E:Z* = 12:1, r.r. = 12:1).

¹H NMR (400 MHz, CDCl₃) δ 7.35-7.29 (m, 5H), 7.22 (t, *J* = 7.6 Hz, 5H), 6.34 (d, *J* = 15.6 Hz, 1H), 5.62 (dt, *J* = 14.8, 6.8 Hz, 1H), 5.20 (s, 1H), 5.09 (s, 1H), 2.89 (s, 2H), 2.78 (d, *J* = 8.1 Hz, 2H), 2.54 (t, *J* = 7.9 Hz, 2H), 2.22 (t, *J* = 7.2 Hz, 2H), 1.70 (p, *J* = 7.4 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 147.96, 140.65, 140.57, 133.08, 132.28, 128.49, 128.27, 128.10, 127.37, 126.35, 115.05, 36.39, 33.63, 31.90, 31.71, 29.08.

HRMS (APCI) calcd. for C₂₁H₂₅S⁺ [M+H]⁺: 309.1672; found:309.1662.

(E)-(4-cyclopropylbuta-1,3-dien-2-yl)benzene



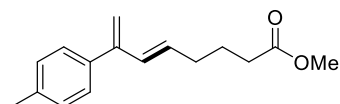
Compound **9b** was isolated via column chromatography (PE) as pale yellow oil (22.5 mg, 66% yield, *E:Z* = 7:1, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.35-7.28 (m, 5H), 6.41 (d, *J* = 15.6 Hz, 1H), 5.19-5.11 (m, 2H), 5.00 (s, 1H), 1.52-1.44 (m, 1H), 0.79-0.72 (m, 2H), 0.40-0.35 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 147.92, 140.73, 138.17, 129.10, 128.30, 128.04, 127.26, 113.91, 14.37, 7.25.

HRMS (APCI) calcd. for C₁₃H₁₅⁺ [M+H]⁺: 171.1169; found: 171.1163.

methyl (E)-7-(p-tolyl)octa-5,7-dienoate



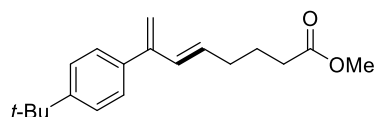
Compound **10b** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (30.3 mg, 62% yield, *E:Z* >20:1, r.r. = 7:1).

¹H NMR (400 MHz, CDCl₃) δ 7.20 (d, *J* = 7.9 Hz, 2H), 7.15 (d, *J* = 8.0 Hz, 2H), 6.30 (d, *J* = 15.6 Hz, 1H), 5.62 (dt, *J* = 15.0, 7.0 Hz, 1H), 5.16 (s, 1H), 5.06 (s, 1H), 3.66 (s, 3H), 2.36 (s, 3H), 2.32-2.28 (m, 2H), 2.16 (q, *J* = 7.2 Hz, 2H), 1.76-1.69 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 174.04, 147.73, 137.57, 137.08, 132.72, 132.49, 128.79, 128.11, 114.69, 51.53, 33.45, 32.16, 24.37, 21.17.

HRMS (APCI) calcd. for C₁₆H₂₁O₂⁺ [M+H]⁺: 245.1537; found: 245.1529.

methyl (E)-7-(4-(tert-butyl)phenyl)octa-5,7-dienoate



Compound **11b** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (26.9 mg, 47% yield, *E:Z* >20:1, r.r. = 5:1).

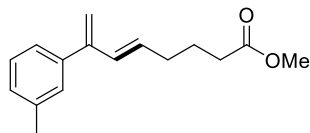
¹H NMR (500 MHz, CDCl₃) δ 7.38 (d, *J* = 8.4 Hz, 2H), 7.27 (d, *J* = 1.9 Hz, 2H), 6.32 (dd, *J* = 15.6, 1.9 Hz, 1H), 5.68 (dt, *J* = 15.6, 7.0 Hz, 1H), 5.18 (d, *J* = 1.8 Hz, 1H), 5.10 (d, *J* = 1.9 Hz, 1H), 3.68 (s, 3H),

2.34 (t, $J = 7.5$ Hz, 2H), 2.20-2.16 (m, 2H), 1.78-1.73 (m, 2H), 1.36 (s, 9H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 174.11, 150.37, 147.69, 137.55, 132.73, 132.48, 127.89, 125.06, 114.74, 51.59, 34.61, 33.54, 32.27, 31.44, 31.40, 24.47, 24.45.

HRMS (APCI) calcd. for $\text{C}_{19}\text{H}_{27}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 287.2006; found: 287.1996.

methyl (*E*)-7-(*m*-tolyl)octa-5,7-dienoate



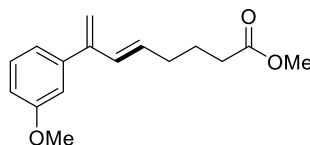
Compound **12b** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (24.9 mg, 51% yield, $E:Z > 20:1$, r.r. = 6:1).

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.22-7.18 (m, 1H), 7.14-7.00 (m, 3H), 6.30 (d, $J = 15.6$ Hz, 1H), 5.61 (dt, $J = 15.0, 7.0$ Hz, 1H), 5.17 (s, 1H), 5.06 (s, 1H), 3.66 (s, 3H), 2.36 (s, 3H), 2.31 (t, $J = 7.6$ Hz, 2H), 2.16 (q, $J = 6.9$ Hz, 2H), 1.77-1.68 (m, 2H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 174.09, 148.06, 140.54, 137.75, 132.86, 132.51, 129.00, 128.17, 128.01, 125.39, 114.97, 51.60, 33.52, 32.23, 24.44, 21.53.

HRMS (APCI) calcd. for $\text{C}_{16}\text{H}_{21}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 245.1537; found: 245.1531.

methyl (*E*)-7-(3-methoxyphenyl)octa-5,7-dienoate



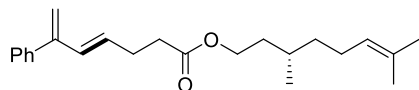
Compound **13b** was isolated via column chromatography (PE/EA=30:1~15:1) as pale yellow oil (27.1 mg, 52% yield, $E:Z = 12:1$, r.r. $> 20:1$).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.26-7.22 (m, 1H), 6.89 (dd, $J = 7.7, 1.3$ Hz, 1H), 6.87-6.83 (m, 2H), 6.29 (d, $J = 15.6$ Hz, 1H), 5.63 (dt, $J = 15.0, 7.0$ Hz, 1H), 5.18 (s, 1H), 5.09 (s, 1H), 3.82 (s, 3H), 3.66 (s, 3H), 2.32 (t, $J = 7.5$ Hz, 2H), 2.16 (q, $J = 7.2$ Hz, 2H), 1.76-1.69 (m, 2H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 174.01, 159.35, 147.76, 141.98, 132.95, 132.22, 129.06, 120.76, 115.09, 113.94, 112.79, 55.23, 51.53, 33.45, 32.16, 24.34.

HRMS (APCI) calcd. for $\text{C}_{16}\text{H}_{21}\text{O}_3^+$ $[\text{M}+\text{H}]^+$: 261.1486; found: 261.1484.

(*S*)-3,7-dimethyloct-6-en-1-yl (*E*)-6-phenylhepta-4,6-dienoate



Compound **14b** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (37.4 mg, 55% yield, $E:Z = 14:1$, r.r. $> 20:1$).

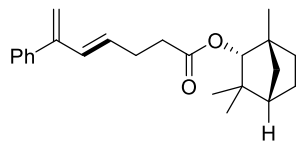
$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.37-7.28 (m, 5H), 6.34 (d, $J = 15.6$ Hz, 1H), 5.64 (dt, $J = 14.4, 6.4$ Hz, 1H), 5.20 (s, 1H), 5.09 (s, 2H), 4.14-4.08 (m, 2H), 2.49-2.39 (m, 4H), 2.01-1.93 (m, 2H), 1.68 (s, 3H), 1.60 (s, 3H), 1.53 (t, $J = 6.8$ Hz, 1H), 1.46-1.41 (m, 1H), 1.39-1.28 (m, 3H), 1.23-1.17 (m, 1H), 0.90 (d, $J = 6.5$ Hz, 3H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 173.05, 147.74, 132.47, 131.82, 131.36, 128.22, 128.09, 127.40, 124.59,

115.42, 63.00, 36.99, 35.46, 33.98, 29.50, 28.17, 25.73, 25.39, 19.41, 17.67.

HRMS (APCI) calcd. for $C_{23}H_{33}O_2^+$ $[M+H]^+$: 341.2476; found: 341.2467.

(1*R*,2*R*,4*S*)-1,3,3-trimethylbicyclo[2.2.1]heptan-2-yl (*E*)-6-phenylhepta-4,6-dienoate



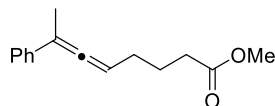
Compound **15b** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (46.1 mg, 68% yield, *E*:*Z*>20:1, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.35-7.27 (m, 5H), 6.36 (d, *J* = 15.6 Hz, 1H), 5.64 (dt, *J* = 16.5, 6.2 Hz, 1H), 5.19 (s, 1H), 5.08 (s, 1H), 4.36 (d, *J* = 1.9 Hz, 1H), 2.49-2.42 (m, 4H), 1.73-1.66 (m, 3H), 1.60-1.54 (m, 2H), 1.47-1.41 (m, 1H), 1.20-1.15 (m, 1H), 1.09 (s, 3H), 1.01 (s, 3H), 0.74 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 173.31, 147.82, 140.42, 132.51, 131.96, 128.25, 128.08, 127.35, 115.44, 86.22, 48.34, 48.26, 41.36, 39.39, 34.03, 29.70, 28.22, 26.64, 25.82, 20.19, 19.37.

HRMS (APCI) calcd. for $C_{23}H_{31}O_2^+$ $[M+H]^+$: 339.2319; found: 339.2309.

methyl 7-phenylocta-5,6-dienoate



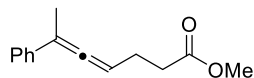
Compound **2c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (42.8 mg, 93% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.40 (d, *J* = 8.0 Hz, 2H), 7.32 (t, *J* = 7.7 Hz, 2H), 7.20 (t, *J* = 7.3 Hz, 1H), 5.44 (tq, *J* = 6.1, 2.9 Hz, 1H), 3.67 (s, 3H), 2.39 (t, *J* = 7.5 Hz, 2H), 2.16 (q, *J* = 6.8 Hz, 2H), 2.10 (d, *J* = 2.9 Hz, 3H), 1.87-1.79 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 204.31, 173.98, 137.47, 128.28, 126.45, 125.63, 100.91, 92.07, 51.49, 33.45, 28.40, 24.38, 17.17.

HRMS (APCI) calcd. for $C_{15}H_{19}O_2^+$ $[M+H]^+$: 231.1380; found: 231.1376.

methyl 6-phenylhepta-4,5-dienoate



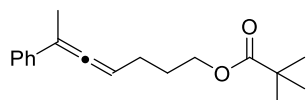
Compound **3c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (35.9 mg, 88% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.39 (d, *J* = 7.3 Hz, 2H), 7.32 (t, *J* = 7.6 Hz, 2H), 7.20 (t, *J* = 7.2 Hz, 1H), 5.56-5.50 (m, 1H), 3.59 (s, 3H), 2.50-2.42 (m, 4H), 2.08 (d, *J* = 2.9 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 203.90, 173.48, 137.18, 128.27, 126.59, 125.65, 102.18, 91.79, 51.54, 33.02, 24.03, 17.12.

HRMS (APCI) calcd. for $C_{14}H_{17}O_2^+$ $[M+H]^+$: 217.1224; found: 217.1221.

6-phenylhepta-4,5-dien-1-yl pivalate



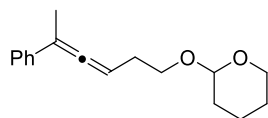
Compound **4c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (47.9 mg, 92% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, *J* = 8.4 Hz, 2H), 7.32 (t, *J* = 7.7 Hz, 2H), 7.20 (t, *J* = 7.3 Hz, 1H), 5.47 (tq, *J* = 6.0, 2.9 Hz, 1H), 4.13 (t, *J* = 6.4 Hz, 2H), 2.20 (q, *J* = 7.1 Hz, 2H), 2.10 (d, *J* = 2.9 Hz, 3H), 1.86-1.78 (m, 2H), 1.20 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 204.25, 178.58, 137.46, 128.30, 126.49, 125.62, 101.03, 92.00, 63.66, 38.77, 28.11, 27.22, 25.43, 17.17.

HRMS (APCI) calcd. for C₁₈H₂₅O₂⁺ [M+H]⁺: 273.1850; found: 273.1842.

2-((5-phenylhexa-3,4-dien-1-yl)oxy)tetrahydro-2H-pyran



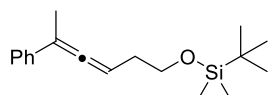
Compound **5c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (37.9 mg, 77% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.43 (d, *J* = 7.7 Hz, 2H), 7.32 (t, *J* = 7.6 Hz, 2H), 7.19 (t, *J* = 7.3 Hz, 1H), 5.53-5.46 (m, 1H), 4.62 (dt, *J* = 13.6, 3.7 Hz, 1H), 3.92-3.85 (m, 2H), 3.57-3.48 (m, 2H), 2.42 (q, *J* = 6.7 Hz, 2H), 2.10 (d, *J* = 2.9 Hz, 3H), 1.88-1.80 (m, 1H), 1.74-1.69 (m, 1H), 1.61-1.48 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 204.74, 204.71, 137.48, 128.23, 126.42, 125.71, 100.68, 100.62, 98.85, 98.81, 89.91, 66.87, 66.82, 62.20, 62.11, 30.71, 30.67, 29.67, 29.61, 25.51, 19.51, 19.46, 17.09.

HRMS (APCI) calcd. for C₁₇H₂₃O₂⁺ [M+H]⁺: 259.1693; found: 259.1685.

tert-butyldimethyl((5-phenylhexa-3,4-dien-1-yl)oxy)silane



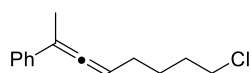
Compound **6c** was isolated via column chromatography (PE/EA=100:1~70:1) as pale yellow oil (43.1 mg, 78% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, *J* = 7.3 Hz, 2H), 7.32 (t, *J* = 7.6 Hz, 2H), 7.20 (t, *J* = 7.3 Hz, 1H), 5.47 (tq, *J* = 6.2, 2.9 Hz, 1H), 3.76 (t, *J* = 6.6 Hz, 2H), 2.35 (q, *J* = 6.7 Hz, 2H), 2.10 (d, *J* = 2.9 Hz, 3H), 0.92 (s, 9H), 0.08 (s, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 204.74, 137.55, 128.24, 126.37, 125.69, 100.25, 89.79, 63.02, 32.77, 25.98, 18.39, 17.13, -5.23.

HRMS (APCI) calcd. for C₁₈H₂₉OSi⁺ [M+H]⁺: 289.1983; found: 289.1980.

(8-chloroocta-2,3-dien-2-yl)benzene



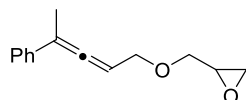
Compound **7c** was isolated via column chromatography (PE/EA=100:1~70:1) as pale yellow oil (25.1 mg, 60% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, *J* = 7.2 Hz, 2H), 7.33 (t, *J* = 7.7 Hz, 2H), 7.20 (t, *J* = 7.3 Hz, 1H), 5.45 (tq, *J* = 6.1, 2.9 Hz, 1H), 3.54 (t, *J* = 6.7 Hz, 2H), 2.16 (q, *J* = 7.1 Hz, 2H), 2.10 (d, *J* = 2.9 Hz, 3H), 1.90-1.82 (m, 2H), 1.64 (qd, *J* = 7.3, 2.2 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 204.23, 137.53, 128.29, 126.45, 125.61, 100.78, 92.40, 44.86, 32.08, 28.21, 26.35, 17.20.

HRMS (APCI) calcd. for C₁₄H₁₈Cl⁺ [M+H]⁺: 221.1092; found: 221.1086.

2-(((4-phenylpenta-2,3-dien-1-yl)oxy)methyl)oxirane



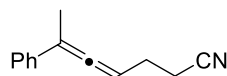
Compound **8c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (30.2 mg, 73% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.40 (d, *J* = 7.3 Hz, 2H), 7.33 (t, *J* = 7.6 Hz, 2H), 7.22 (t, *J* = 7.3 Hz, 1H), 5.59-5.52 (m, 1H), 4.21-4.13 (m, 2H), 3.79-3.74 (m, 1H), 3.47 (dd, *J* = 11.4, 5.7 Hz, 1H), 3.20-3.15 (m, 1H), 2.80 (t, *J* = 4.1 Hz, 1H), 2.65-2.60 (m, 1H), 2.13 (d, *J* = 2.9 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 205.19, 136.60, 128.37, 126.87, 125.80, 101.66, 90.33, 90.30, 70.50, 69.45, 50.73, 44.44, 17.00.

HRMS (APCI) calcd. for C₁₄H₁₇O₂⁺ [M+H]⁺: 217.1224; found: 184.1217.

6-phenylhepta-4,5-dienitrile



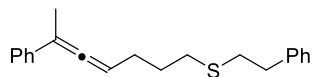
Compound **9c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (27.4 mg, 80% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, *J* = 7.7 Hz, 2H), 7.34 (t, *J* = 7.7 Hz, 2H), 7.23 (t, *J* = 7.3 Hz, 1H), 5.58-5.51 (m, *J* = 2.9 Hz, 1H), 2.49-2.44 (m, 4H), 2.15 (d, *J* = 2.9 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 204.20, 136.54, 128.41, 127.00, 125.76, 119.35, 103.48, 90.08, 24.99, 17.14, 16.66.

HRMS (APCI) calcd. for C₁₃H₁₄N⁺ [M+H]⁺: 184.1121; found: 184.1116.

phenethyl(6-phenylhepta-4,5-dien-1-yl)sulfane



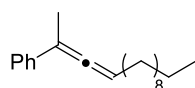
Compound **10c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (30.8 mg, 52% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.43 (d, *J* = 8.0 Hz, 2H), 7.37-7.30 (m, 4H), 7.22 (t, *J* = 8.5 Hz, 4H), 5.52-5.45 (m, 1H), 2.92-2.87 (m, 2H), 2.81-2.74 (m, 2H), 2.63 (t, *J* = 7.4 Hz, 2H), 2.25 (q, *J* = 7.0 Hz, 2H), 2.13 (d, *J* = 2.9 Hz, 3H), 1.85-1.76 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 204.30, 140.68, 137.50, 128.50, 128.33, 126.50, 126.35, 125.65, 100.96, 92.21, 36.43, 33.73, 31.79, 29.01, 28.07, 17.26.

HRMS (APCI) calcd. for C₂₁H₂₄SNa⁺ [M+Na]⁺: 331.1491; found: 331.1490.

tetradeca-2,3-dien-2-ylbenzene



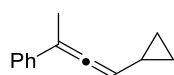
Compound **11c** was isolated via column chromatography (PE) as pale yellow oil (46.1 mg, 89% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, *J* = 7.7 Hz, 2H), 7.31 (t, *J* = 7.6 Hz, 2H), 7.18 (t, *J* = 7.3 Hz, 1H), 5.44 (tq, *J* = 6.2, 2.9 Hz, 1H), 2.09 (d, *J* = 3.0 Hz, 3H), 1.36-1.22 (m, 18H), 0.89 (t, *J* = 6.7 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 204.14, 137.84, 128.21, 126.25, 125.59, 100.22, 93.11, 31.92, 29.68, 29.62, 29.48, 29.35, 29.25, 29.22, 28.99, 22.69, 17.20, 14.12.

HRMS (APCI) calcd. for C₂₀H₃₁⁺ [M+H]⁺: 271.2421; found: 271.2415.

(4-cyclopropylbuta-2,3-dien-2-yl)benzene



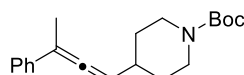
Compound **12c** was isolated via column chromatography (PE) as pale yellow oil (30.0 mg, 95% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, *J* = 7.0 Hz, 2H), 7.33 (t, *J* = 7.6 Hz, 2H), 7.20 (t, *J* = 7.4 Hz, 1H), 5.37-5.29 (m, 1H), 2.11 (d, *J* = 2.8 Hz, 3H), 1.37-1.31 (m, 1H), 0.77-0.71 (m, 2H), 0.46-0.41 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 203.64, 137.61, 128.26, 126.47, 125.66, 102.04, 97.45, 17.34, 9.72, 6.97, 6.87.

HRMS (APCI) calcd. for C₁₃H₁₅⁺ [M+H]⁺: 171.1169; found: 171.1165.

tert-butyl 4-(3-phenylbuta-1,2-dien-1-yl)piperidine-1-carboxylate



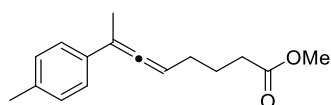
Compound **13c** was isolated via column chromatography (PE/EA=20:1~10:1) as pale yellow oil (57.0 mg, 91% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.40 (d, *J* = 7.4 Hz, 2H), 7.32 (t, *J* = 7.8 Hz, 2H), 7.20 (t, *J* = 7.3 Hz, 1H), 5.49-5.43 (m, 1H), 4.03 (s, 2H), 2.82 (d, *J* = 11.1 Hz, 2H), 2.31-2.22 (m, 1H), 2.10 (d, *J* = 2.9 Hz, 3H), 1.78 (d, *J* = 13.6 Hz, 2H), 1.46 (s, 9H), 1.41-1.32 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 203.27, 154.87, 137.27, 128.32, 126.54, 125.50, 102.01, 97.41, 79.33, 43.60, 35.91, 31.92, 28.47, 17.24.

HRMS (ESI) calcd. for C₂₀H₂₈NO₂⁺ [M+H]⁺: 314.2115; found: 314.2018.

methyl 7-(p-tolyl)octa-5,6-dienoate



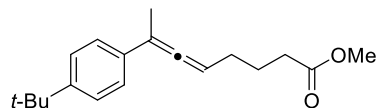
Compound **14c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (38.6 mg, 83% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.29 (d, *J* = 7.9 Hz, 2H), 7.13 (d, *J* = 7.9 Hz, 2H), 5.45-5.38 (m, 1H), 3.67 (s, 3H), 2.39 (t, *J* = 7.5 Hz, 2H), 2.34 (s, 3H), 2.15 (q, *J* = 7.1 Hz, 2H), 2.08 (d, *J* = 3.0 Hz, 3H), 1.85-1.77 (m, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 204.02, 174.04, 136.15, 134.51, 129.01, 125.53, 100.75, 91.93, 51.51, 33.47, 28.47, 24.38, 21.06, 17.25.

HRMS (APCI) calcd. for $\text{C}_{16}\text{H}_{21}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 245.1537; found: 245.1536.

methyl 7-(4-(tert-butyl)phenyl)octa-5,6-dienoate



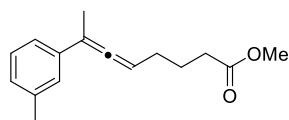
Compound **15c** was isolated via column chromatography (PE/EA=30:1~15:1) as pale yellow oil (44.4 mg, 81% yield, r.r. >20:1).

^1H NMR (500 MHz, CDCl_3) δ 7.35 (s, 4H), 5.46-5.39 (m, 1H), 3.67 (s, 3H), 2.39 (t, $J = 7.5$ Hz, 2H), 2.15 (q, $J = 6.9$ Hz, 2H), 2.09 (s, 3H), 1.85-1.78 (m, 2H), 1.32 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 204.23, 174.11, 149.53, 134.60, 125.40, 125.30, 100.70, 92.00, 51.59, 34.53, 33.57, 31.41, 28.57, 24.48, 17.26.

HRMS (APCI) calcd. for $\text{C}_{19}\text{H}_{27}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 287.2006; found: 287.1998.

methyl 7-(m-tolyl)octa-5,6-dienoate



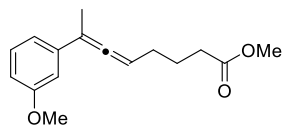
Compound **16c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (40.5 mg, 87% yield, r.r. >20:1).

^1H NMR (500 MHz, CDCl_3) δ 7.24-7.20 (m, 3H), 7.04-7.00 (m, 1H), 5.42 (tq, $J = 6.0, 2.9$ Hz, 1H), 3.67 (s, 3H), 2.42-2.38 (m, 2H), 2.36 (s, 3H), 2.19-2.13 (m, 2H), 2.09 (d, $J = 2.9$ Hz, 3H), 1.86-1.76 (m, 2H).

^{13}C NMR (126 MHz, CDCl_3) δ 204.36, 174.11, 137.86, 137.46, 128.26, 127.35, 126.43, 122.82, 100.95, 91.99, 51.60, 33.53, 28.50, 24.43, 21.61, 17.35.

HRMS (APCI) calcd. for $\text{C}_{16}\text{H}_{21}\text{O}_2^+$ $[\text{M}+\text{H}]^+$: 245.1537; found: 245.1531.

methyl 7-(3-methoxyphenyl)octa-5,6-dienoate



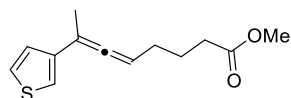
Compound **17c** was isolated via column chromatography (PE/EA=30:1~15:1) as pale yellow oil (45.7 mg, 92% yield, r.r. >20:1).

^1H NMR (400 MHz, CDCl_3) δ 7.22 (d, $J = 7.9$ Hz, 1H), 7.00 (d, $J = 8.5$ Hz, 1H), 6.95 (d, $J = 1.6$ Hz, 1H), 6.75 (dd, $J = 8.2, 2.6$ Hz, 1H), 5.43 (tq, $J = 6.1, 2.9$ Hz, 1H), 3.81 (s, 3H), 3.66 (s, 3H), 2.39 (t, $J = 7.5$ Hz, 2H), 2.15 (q, $J = 7.1$ Hz, 2H), 2.08 (d, $J = 2.9$ Hz, 3H), 1.85-1.77 (m, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 204.38, 173.98, 159.66, 139.09, 129.18, 118.22, 111.80, 111.53, 100.84, 92.12, 55.20, 51.52, 33.45, 28.37, 24.37, 17.23.

HRMS (APCI) calcd. for $\text{C}_{16}\text{H}_{21}\text{O}_3^+$ $[\text{M}+\text{H}]^+$: 261.1486; found: 261.1479.

methyl 7-(thiophen-3-yl)octa-5,6-dienoate



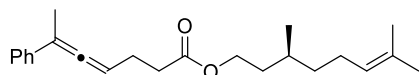
Compound **18c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (33.6 mg, 75% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.24 (dd, *J* = 5.1, 2.9 Hz, 1H), 7.09 (dd, *J* = 5.1, 1.3 Hz, 1H), 7.04-7.01 (m, 1H), 5.38 (tq, *J* = 6.2, 2.9 Hz, 1H), 3.66 (s, 3H), 2.38 (t, *J* = 7.5 Hz, 2H), 2.13 (q, *J* = 7.3 Hz, 2H), 2.06 (d, *J* = 2.8 Hz, 3H), 1.84-1.76 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 204.32, 173.98, 139.80, 126.65, 125.33, 118.69, 97.42, 91.62, 51.52, 33.44, 28.49, 24.35, 17.72.

HRMS (APCI) calcd. for C₁₃H₁₇O₂S⁺ [M+H]⁺: 237.0944; found: 237.0936.

(*S*)-3,7-dimethyloct-6-en-1-yl 6-phenylhepta-4,5-dienoate



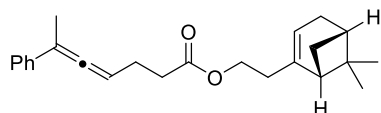
Compound **19c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (53.9 mg, 82% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.38 (d, *J* = 7.8 Hz, 2H), 7.31 (t, *J* = 7.6 Hz, 2H), 7.19 (t, *J* = 7.3 Hz, 1H), 5.55-5.49 (m, 1H), 5.08 (t, *J* = 7.3 Hz, 1H), 4.09-3.97 (m, 2H), 2.49-2.40 (m, 4H), 2.08 (d, *J* = 2.9 Hz, 3H), 2.02-1.90 (m, 2H), 1.68 (s, 3H), 1.60 (s, 3H), 1.54-1.46 (m, 1H), 1.44-1.31 (m, 2H), 1.30-1.25 (m, 1H), 1.20-1.11 (m, 1H), 0.88 (d, *J* = 6.5 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 203.92, 173.10, 137.20, 131.33, 128.26, 126.57, 125.64, 124.59, 102.04, 91.84, 63.01, 36.97, 35.42, 33.39, 29.47, 25.72, 25.38, 24.08, 19.40, 17.66, 17.13.

HRMS (APCI) calcd. for C₂₃H₃₃O₂⁺ [M+H]⁺: 341.2476; found: 341.2464.

2-((1*R*,5*S*)-6,6-dimethylbicyclo[3.1.1]hept-2-en-2-yl)ethyl 6-phenylhepta-4,5-dienoate



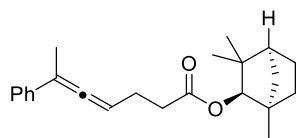
Compound **20c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (24.4 mg, 36% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.38 (d, *J* = 7.5 Hz, 2H), 7.31 (t, *J* = 7.6 Hz, 2H), 7.19 (t, *J* = 7.3 Hz, 1H), 5.55-5.48 (m, 1H), 5.25 (s, 1H), 4.05-3.95 (m, 2H), 2.48-2.40 (m, 4H), 2.37-2.32 (m, 1H), 2.29-2.09 (m, 5H), 2.08 (d, *J* = 3.0 Hz, 3H), 2.00 (t, *J* = 5.7 Hz, 1H), 1.26 (s, 3H), 1.12 (d, *J* = 8.5 Hz, 1H), 0.80 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 203.92, 172.99, 144.11, 137.21, 128.26, 126.57, 125.64, 118.71, 102.02, 91.84, 62.74, 45.68, 40.71, 37.99, 35.90, 33.40, 31.64, 31.35, 26.27, 24.09, 21.11, 17.14.

HRMS (APCI) calcd. for C₂₄H₃₁O₂⁺ [M+H]⁺: 351.2319; found: 351.2309.

(1*R*,2*R*,4*S*)-1,3,3-trimethylbicyclo[2.2.1]heptan-2-yl 6-phenylhepta-4,5-dienoate



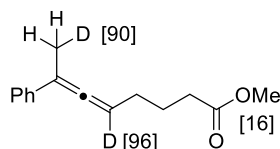
Compound **21c** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (54.2 mg, 83% yield, r.r. >20:1).

¹H NMR (400 MHz, CDCl₃) δ 7.41 (d, *J* = 7.6 Hz, 2H), 7.33 (t, *J* = 7.6 Hz, 2H), 7.20 (t, *J* = 7.3 Hz, 1H), 5.57-5.51 (m, 1H), 4.37 (dd, *J* = 11.5, 1.9 Hz, 1H), 2.54-2.49 (m, 2H), 2.48-2.43 (m, 2H), 2.10 (d, *J* = 2.9 Hz, 3H), 1.81-1.60 (m, 4H), 1.60-1.54 (m, 1H), 1.48-1.41 (m, 1H), 1.20-1.16 (m, 1H), 1.10 (d, *J* = 10.2 Hz, 3H), 1.02 (d, *J* = 16.7 Hz, 3H), 0.77 (d, *J* = 6.4 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 203.98, 173.36, 137.28, 137.25, 128.29, 126.57, 125.68, 101.83, 91.87, 86.19, 86.17, 48.36, 48.26, 48.24, 41.38, 39.43, 33.70, 33.67, 29.70, 29.67, 26.66, 25.85, 24.37, 24.33, 20.19, 20.16, 19.44, 19.37, 17.19, 17.16.

HRMS (APCI) calcd. for C₂₃H₃₁O₂⁺ [M+H]⁺: 339.2319; found: 339.2316.

methyl 7-phenylocta-5,6-dienoate-5,8-d₂



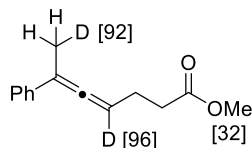
Compound **2d** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (34.8 mg, 75% yield, r.r. >20:1).

¹H NMR (500 MHz, CDCl₃) δ 7.40 (d, *J* = 7.2 Hz, 2H), 7.32 (t, *J* = 7.6 Hz, 2H), 7.24-7.16 (m, 1H), 3.67 (s, 3H), 2.39 (t, *J* = 7.5 Hz, 2H), 2.15 (t, *J* = 7.3 Hz, 2H), 2.09 (d, *J* = 7.3 Hz, 2H), 1.87-1.75 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 204.37, 174.09, 137.52, 128.35, 126.53, 125.68, 100.97, 91.91, 51.60, 33.54, 28.38, 24.41, 17.15, 17.00, 16.84.

HRMS (APCI) calcd. for C₁₅H₁₇D₂O₂⁺ [M+H]⁺: 233.1506; found: 233.1497.

methyl 6-phenylhepta-4,5-dienoate-4,7-d₂



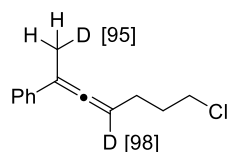
Compound **3d** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (35.8 mg, 82% yield, r.r. >20:1).

¹H NMR (500 MHz, CDCl₃) δ 7.39 (d, *J* = 7.8 Hz, 2H), 7.32 (t, *J* = 7.6 Hz, 2H), 7.20 (t, *J* = 7.2 Hz, 1H), 3.59 (s, 2H), 2.53-2.36 (m, 4H), 2.08 (d, *J* = 7.7 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 203.99, 173.54, 137.24, 128.34, 126.66, 125.72, 102.27, 91.86, 91.64, 91.44, 51.61, 33.07, 24.01, 17.20, 17.09, 16.94, 16.78.

HRMS (APCI) calcd. for C₁₄H₁₅D₂O₂⁺ [M+H]⁺: 219.13490; found: 219.1347.

(7-chlorohepta-2,3-dien-2-yl-1,4-d₂)benzene



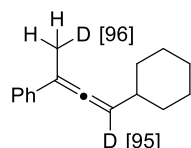
Compound **4d** was isolated via column chromatography (PE/EA=100:1~70:1) as pale yellow oil (37.0 mg, 89% yield, r.r. >20:1).

¹H NMR (500 MHz, CDCl₃) δ 7.41 (d, *J* = 7.7 Hz, 2H), 7.33 (t, *J* = 7.6 Hz, 2H), 7.21 (t, *J* = 7.3 Hz, 1H), 3.61 (t, *J* = 6.6 Hz, 2H), 2.27 (t, *J* = 7.3 Hz, 2H), 2.12-2.07 (m, 2H), 2.00-1.91 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 204.37, 137.39, 128.41, 126.64, 125.69, 101.37, 91.60, 91.40, 91.20, 44.53, 31.85, 26.07, 17.16, 17.01, 16.85.

HRMS (APCI) calcd. for C₁₃H₁₄D₂Cl⁺ [M+H]⁺: 209.1061; found: 209.1055.

(4-cyclohexylbuta-2,3-dien-2-yl-1,4-d₂)benzene



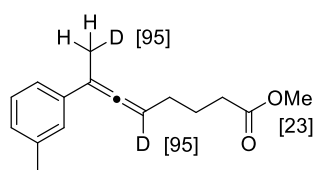
Compound **5d** was isolated via column chromatography (PE) as pale yellow oil (36.4 mg, 85% yield, r.r. >20:1).

¹H NMR (500 MHz, CDCl₃) δ 7.43 (d, *J* = 7.8 Hz, 2H), 7.32 (t, *J* = 7.5 Hz, 2H), 7.19 (t, *J* = 7.3 Hz, 1H), 2.18-2.04 (m, 3H), 1.84 (dd, *J* = 12.8, 3.9 Hz, 2H), 1.77-1.71 (m, 2H), 1.67-1.62 (m, 1H), 1.33-1.27 (m, 2H), 1.23-1.14 (m, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 203.14, 137.87, 128.32, 126.32, 125.53, 101.17, 99.17, 98.98, 98.78, 37.89, 33.33, 33.27, 26.26, 26.16, 17.39, 17.28, 17.13, 16.97.

HRMS (APCI) calcd. for C₁₆H₁₉D₂⁺ [M+H]⁺: 215.1764; found: 215.1757.

methyl 7-(m-tolyl)octa-5,6-dienoate-5,8-d₂



Compound **6d** was isolated via column chromatography (PE/EA=40:1~20:1) as pale yellow oil (37.4 mg, 76% yield, r.r. = 16:1).

¹H NMR (500 MHz, CDCl₃) δ 7.23-7.17 (m, 3H), 7.06-6.97 (m, 1H), 3.67 (s, 3H), 2.39 (t, *J* = 7.5 Hz, 2H), 2.35 (s, 3H), 2.15 (t, *J* = 7.2 Hz, 2H), 2.11-2.04 (m, 2H), 1.86-1.77 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 204.37, 174.10, 137.86, 137.45, 128.25, 127.34, 126.42, 122.81, 100.97, 91.98, 91.74, 91.55, 51.59, 33.53, 28.40, 24.40, 21.60, 17.23, 17.08, 16.92.

HRMS (APCI) calcd. for C₁₆H₁₉D₂O₂⁺ [M+H]⁺: 247.1662; found: 247.1653.

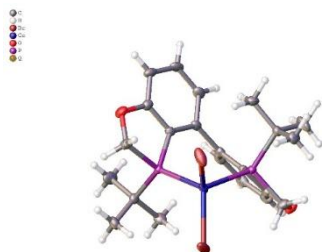


Fig. S2 Crystal structure of **CoBr₂·L7**

Crystal data and structure refinement for **CoBr₂·L7**

Compound	CoBr₂·L7
CCDC	2536099
Identification code	12.17_auto
Empirical formula	C ₂₂ H ₂₈ Br ₂ CoO ₂ P ₂
Formula weight	605.13
Temperature/K	100.00(10)
Crystal system	trigonal
Space group	P3 ₂ 21
a/Å	10.13310(10)
b/Å	10.13310(10)
c/Å	20.9219(3)
α/°	90
β/°	90
γ/°	120
Volume/Å ³	1860.44(5)
Z	3
ρ _{calc} /cm ³	1.620
μ/mm ⁻¹	10.540
F(000)	909.0
Crystal size/mm ³	0.2 × 0.2 × 0.1
Radiation	Cu Kα (λ = 1.54184)
2θ range for data collection/°	10.08 to 152.654
Index ranges	-12 ≤ h ≤ 12, -12 ≤ k ≤ 12, -25 ≤ l ≤ 26
Reflections collected	11981
Independent reflections	2522 [R _{int} = 0.0350, R _{sigma} = 0.0245]
Data/restraints/parameters	2522/0/136
Goodness-of-fit on F ²	1.042
Final R indexes [I ≥ 2σ (I)]	R ₁ = 0.0222, wR ₂ = 0.0519
Final R indexes [all data]	R ₁ = 0.0231, wR ₂ = 0.0522
Largest diff. peak/hole / e Å ⁻³	0.47/-0.51

5. Mechanistic Studies

Reaction Progress Kinetic Analysis (RPKA)

Z-selective hydrogenation kinetic analysis

General method for kinetic studies: according to the general procedure of Z-hydrogenation reactions, using substrate **1**, CoBr₂/L4, MeOH and DIPEA in several different equivalents, and diphenyl was used as an internal standard. The stopwatch was immediately started when the current was turned on. A small amount of the reaction mixture was taken out by syringe. The obtained mixture was immediately diluted by EA (0.5 mL), which was further sampled for GC analysis.

The rate on the concentration of CoBr₂/L4

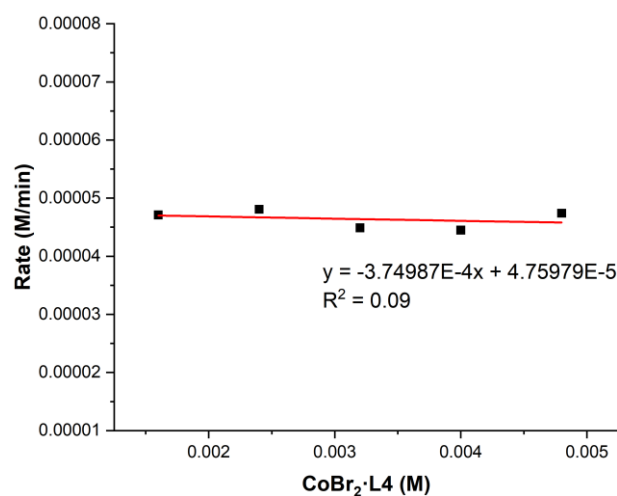
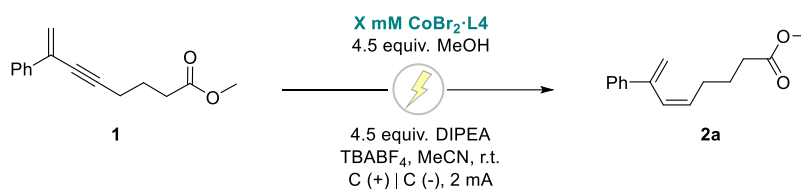


Fig. S3 Rate on the concentration of CoBr₂·L4 from the reaction of substrate **1** (0.04 M), MeOH (0.18 M), DIPEA (0.18 M), TBABF₄ (0.1 M), Current = 2 mA with 0.0016 M, 0.0024 M, 0.0032 M, 0.004 M, 0.0048 M of CoBr₂·L4.

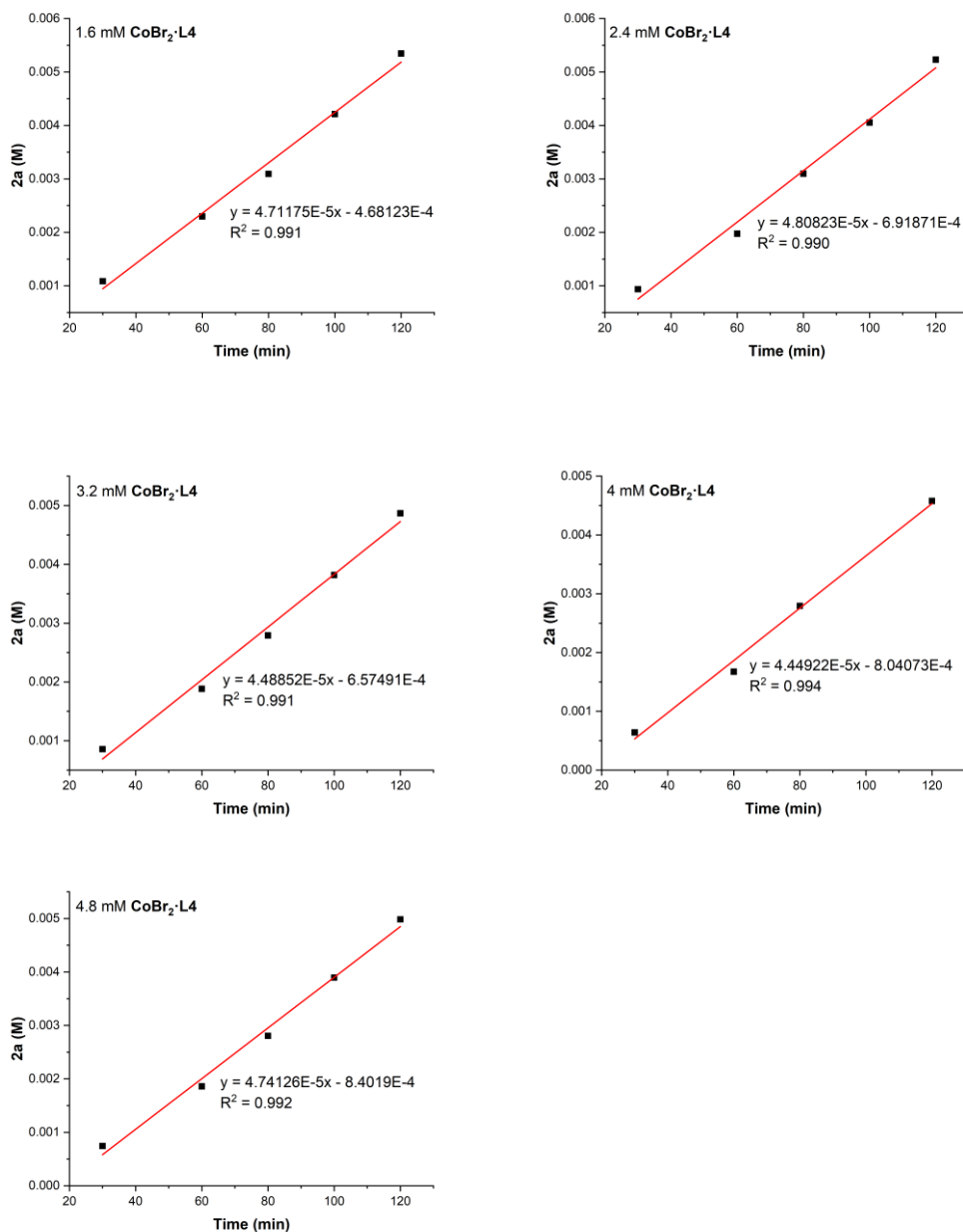


Fig. S4 Plot of the rise of product from the reaction of substrate **1** (0.04 M), MeOH (0.18 M), DIPEA (0.18 M), TBABF₄ (0.1 M), Current = 2 mA with 0.0016 M, 0.0024 M, 0.0032 M, 0.004 M, 0.0048 M of $\text{CoBr}_2 \cdot \text{L4}$.

The rate on the concentration of substrate 1

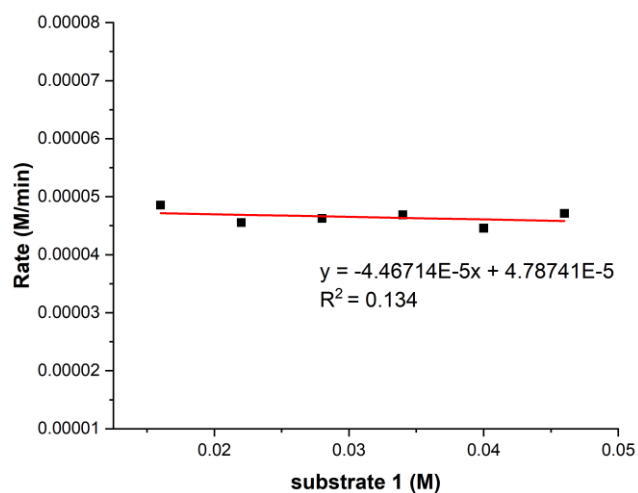
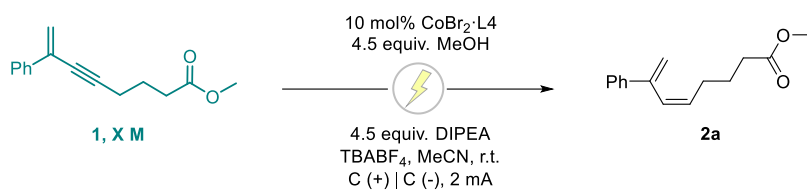
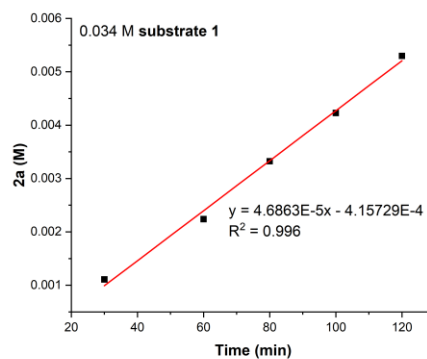
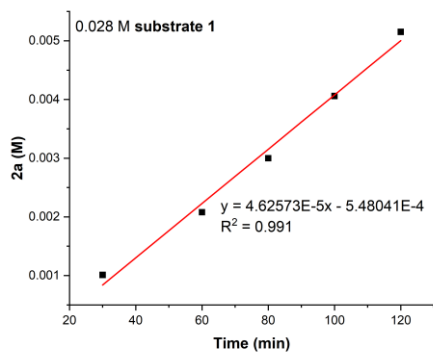
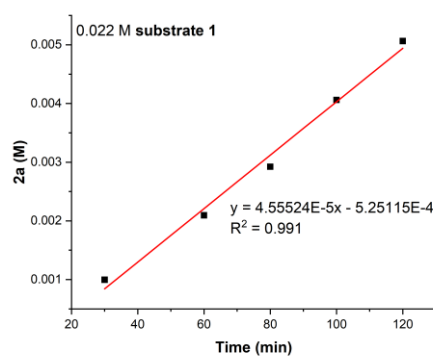
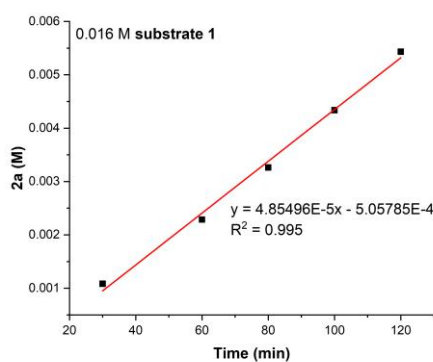


Fig. S5 Rate on the concentration of **substrate 1** from the reaction of CoBr₂·L4 (0.004 M), MeOH (0.18 M), DIPEA (0.18 M), TBABF₄ (0.1 M), Current = 2 mA with 0.016 M, 0.022 M, 0.028 M, 0.034 M, 0.04 M, 0.046 M of **substrate 1**.



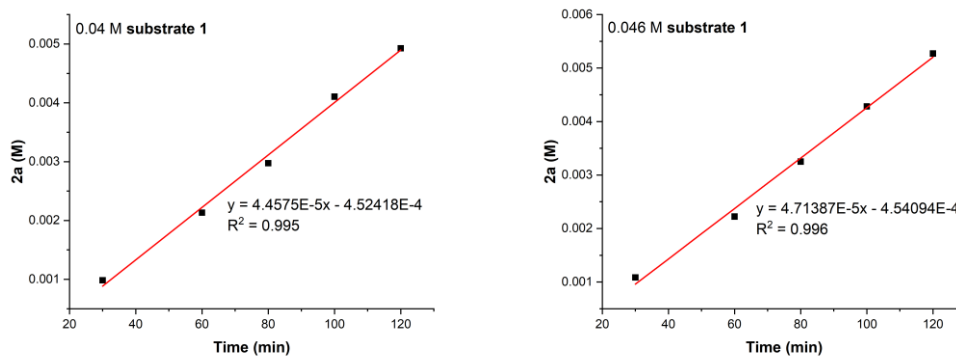


Fig. S6 Plot of the rise of product from the reaction of $\text{CoBr}_2 \cdot \text{L4}$ (0.004 M), MeOH (0.18 M), DIPEA (0.18 M), TBABF_4 (0.1 M), Current = 2 mA with 0.016 M, 0.022 M, 0.028 M, 0.034 M, 0.04 M, 0.046 M of substrate 1.

The rate on the concentration of MeOH

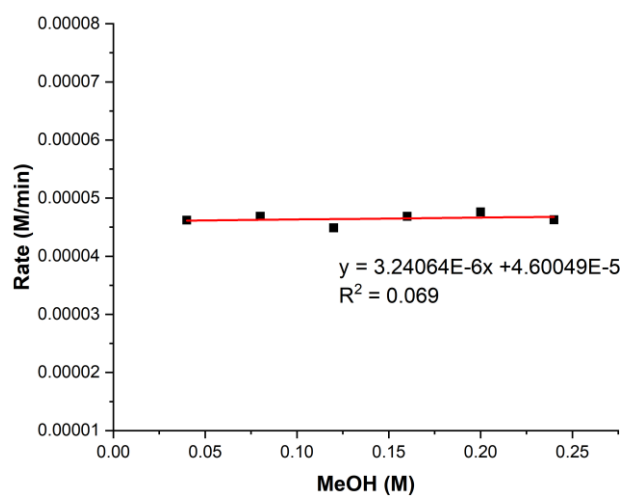
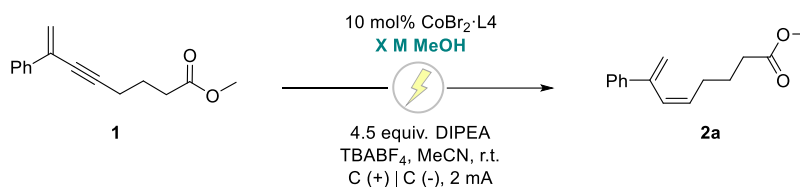


Fig. S7 Rate on the concentration of **MeOH** from the reaction of $\text{CoBr}_2 \cdot \text{L4}$ (0.004 M), substrate 1 (0.04 M), DIPEA (0.18 M), TBABF_4 (0.1 M), Current = 2 mA with 0.04 M, 0.08 M, 0.12 M, 0.16 M, 0.2 M, 0.24 M of **MeOH**.

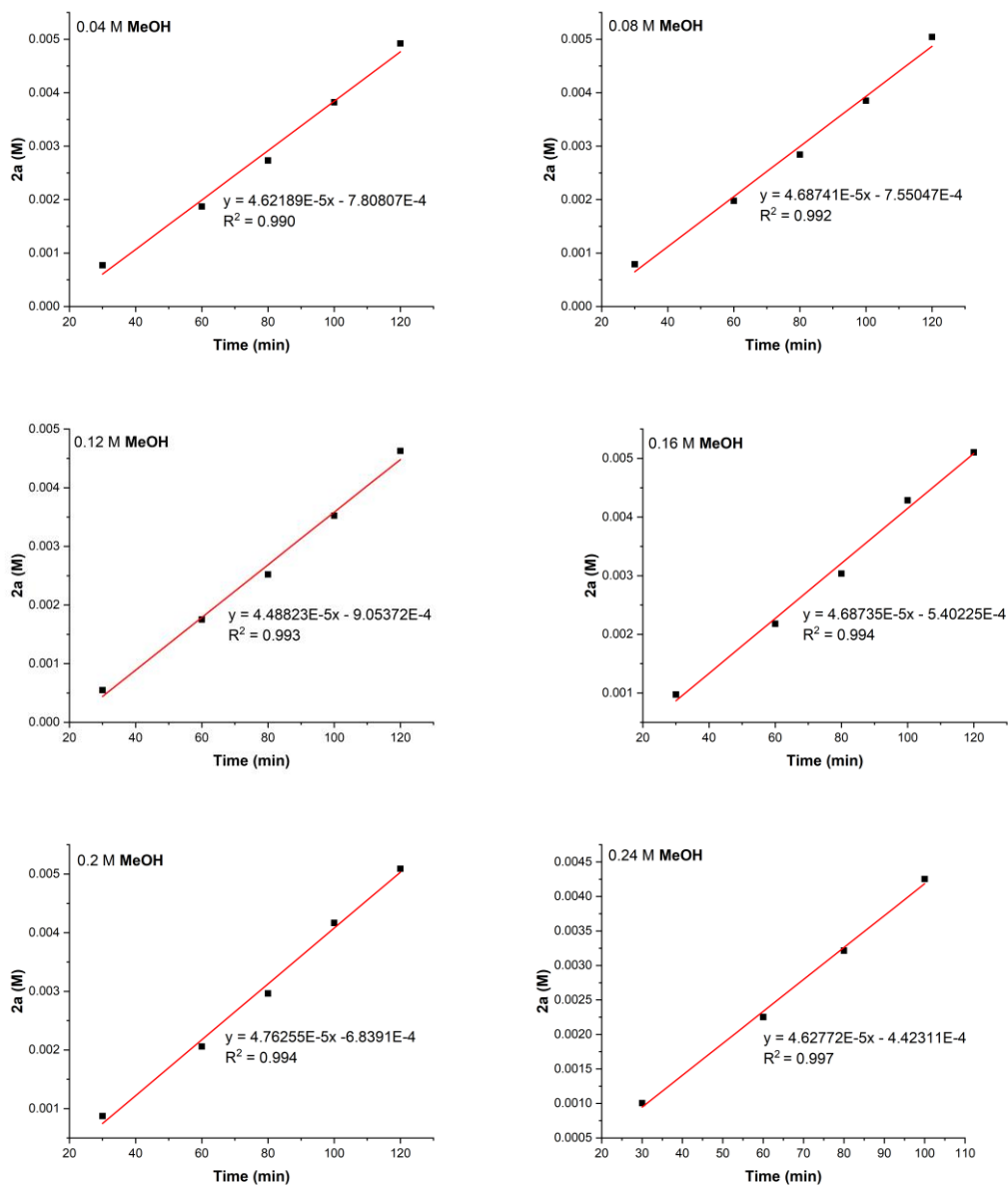


Fig. S8 Plot of the rise of product from the reaction of $\text{CoBr}_2 \cdot \text{L4}$ (0.004 M), substrate **1** (0.04 M), DIPEA (0.18 M), TBABF_4 (0.1 M), Current = 2 mA with 0.04 M, 0.08 M, 0.12 M, 0.16 M, 0.2 M, 0.24 M of MeOH.

The rate on the concentration of DIPEA

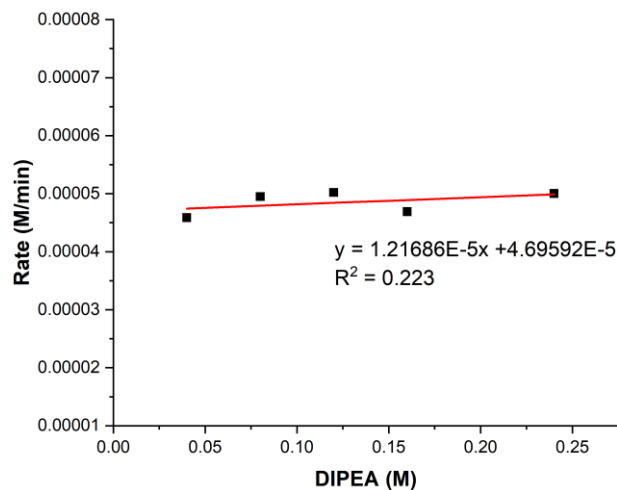
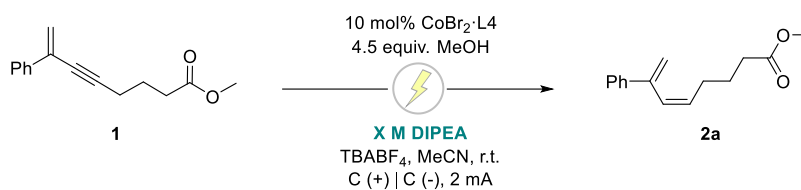
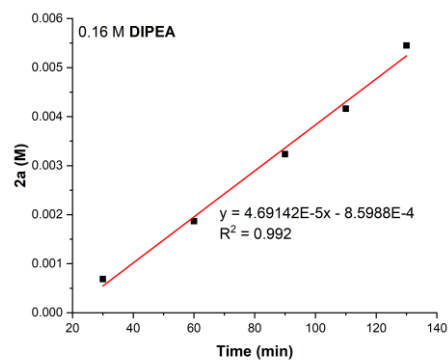
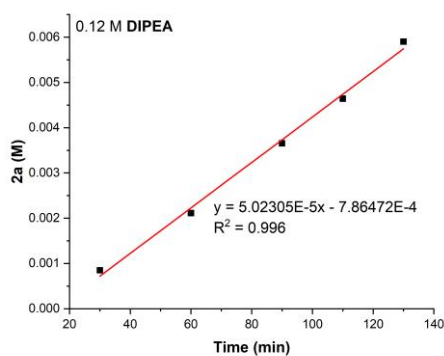
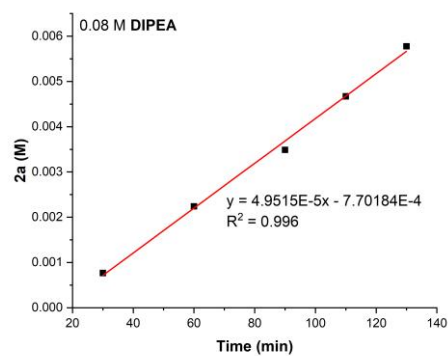
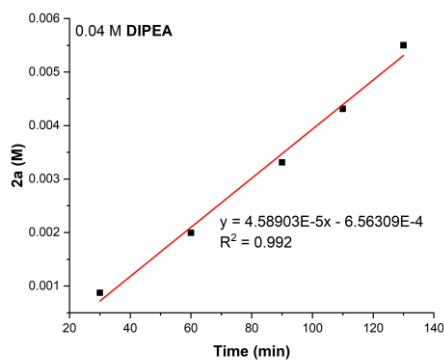


Fig. S9 Rate on the concentration of **DIPEA** from the reaction of CoBr₂·L4 (0.004 M), substrate **1** (0.04 M), MeOH (0.18 M), TBABF₄ (0.1 M), Current = 2 mA with 0.04 M, 0.08 M, 0.12 M, 0.16 M, 0.24 M of **DIPEA**.



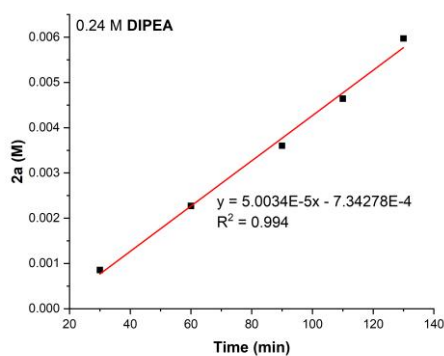


Fig. S10 Plot of the rise of product from the reaction of $\text{CoBr}_2 \cdot \text{L4}$ (0.004 M), substrate **1** (0.04 M), MeOH (0.18 M), TBABF_4 (0.1 M), Current = 2 mA with 0.04 M, 0.08 M, 0.12 M, 0.16 M, 0.24 M of **DIPEA**.

The rate on the value of current

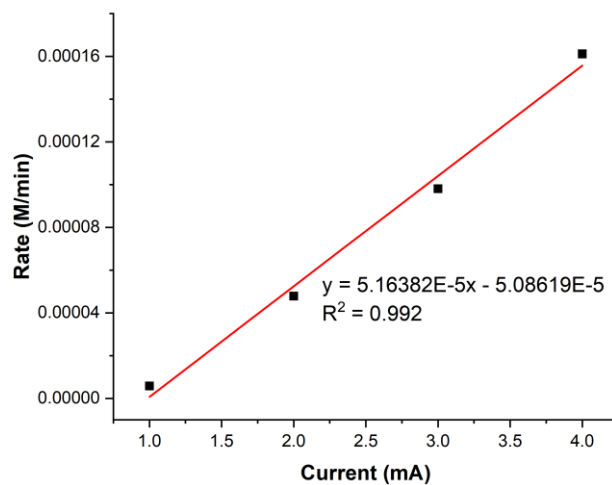
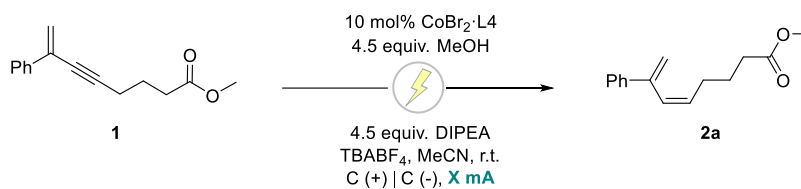


Fig. S11 Rate on the concentration of **Current** from the reaction of $\text{CoBr}_2 \cdot \text{L4}$ (0.004 M), substrate **1** (0.04 M), MeOH (0.18 M), DIPEA (0.18 M), TBABF_4 (0.1 M) with 1 mA, 2 mA, 3 mA, 4 mA of **Current**.

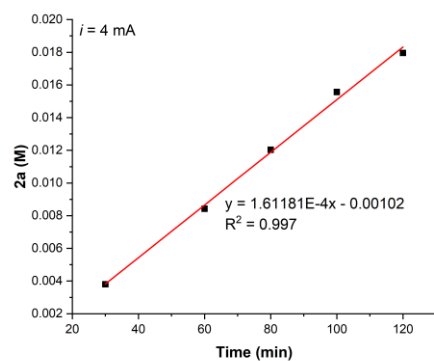
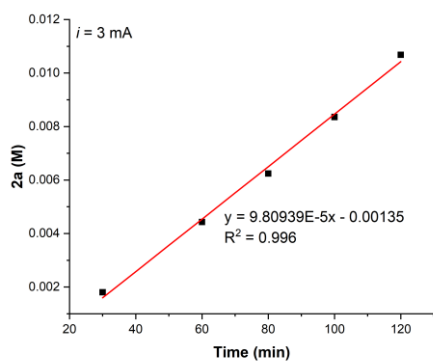
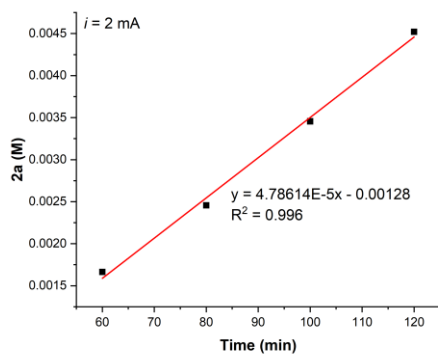
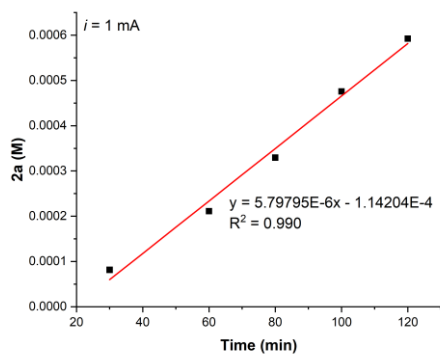


Fig. S12 Plot of the rise of product from the reaction of $\text{CoBr}_2 \cdot \text{L4}$ (0.004 M), substrate **1** (0.04 M), MeOH (0.18 M), DIPEA (0.18 M), TBABF₄ (0.1 M) with 1 mA, 2 mA, 3 mA, 4 mA of Current.

E-selective hydrogenation kinetic analysis

General method for kinetic studies: according to the general procedure of *E*-hydrogenation reactions, using substrate **1**, CoBr₂/L7 and CF₃CH₂OH in several different equivalents, and diphenyl was used as an internal standard. The stopwatch was immediately started when the current was turned on. A small amount of the reaction mixture was taken out by syringe. The obtained mixture was immediately diluted by EA (0.5 mL), which was further sampled for GC analysis.

The rate on the concentration of CoBr₂/L7

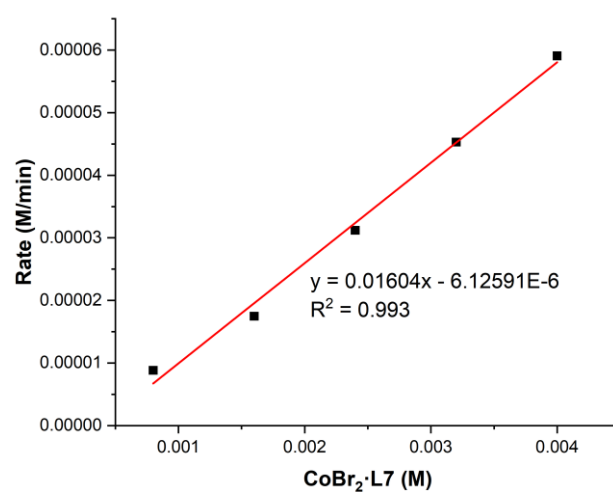
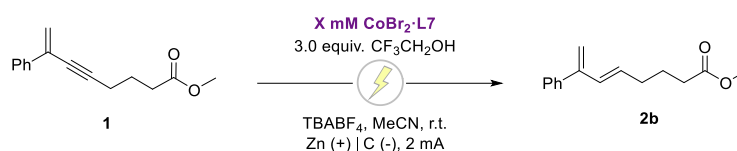
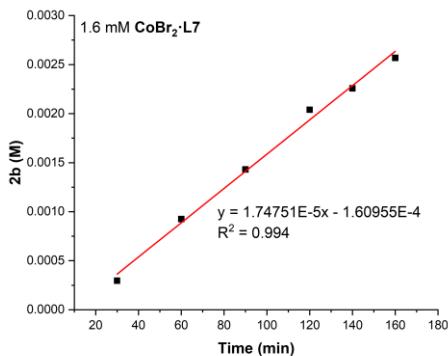
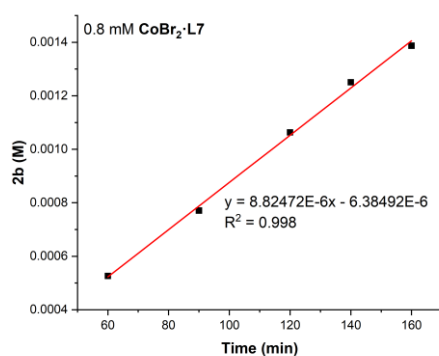


Fig. S13 Rate on the concentration of CoBr₂-L7 from the reaction of substrate **1** (0.04 M), CF₃CH₂OH (0.12 M), TBABF₄ (0.1 M), Current = 2 mA with 0.8 mM, 1.6 mM, 2.4 mM, 3.2 mM, 4 mM of CoBr₂-L7.



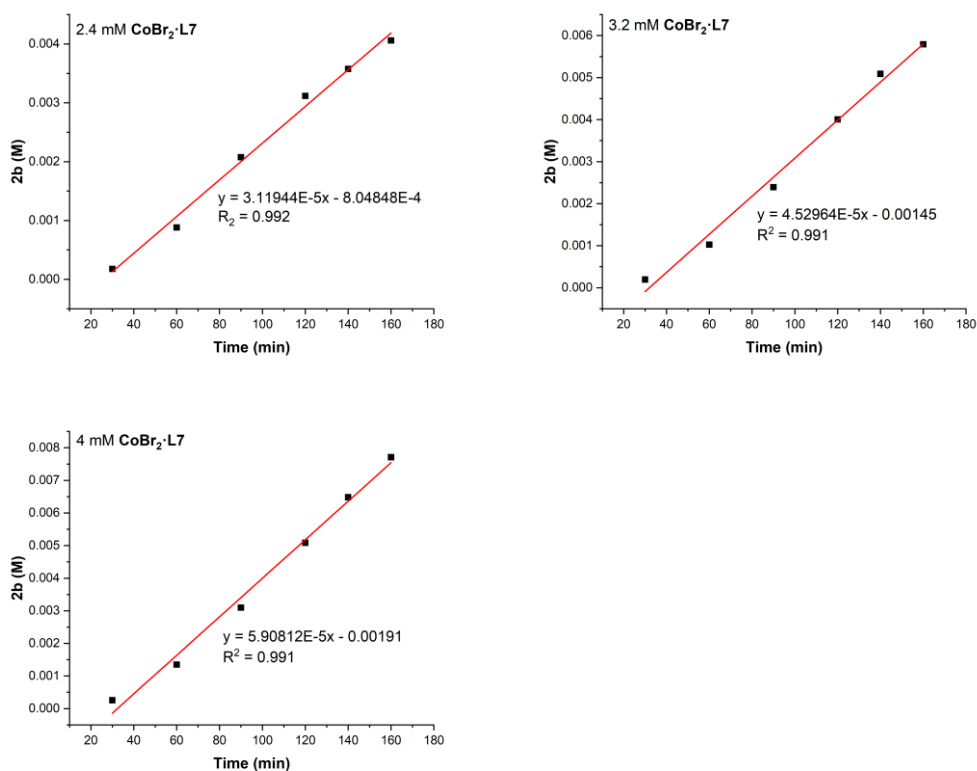


Fig. S14 Plot of the rise of product from the reaction of substrate **1** (0.04 M), CF₃CH₂OH (0.12 M), TBABF₄ (0.1 M), Current = 2 mA with 0.8 mM, 1.6 mM, 2.4 mM, 3.2 mM, 4 mM of CoBr₂·L7.

The rate on the concentration of substrate **1**

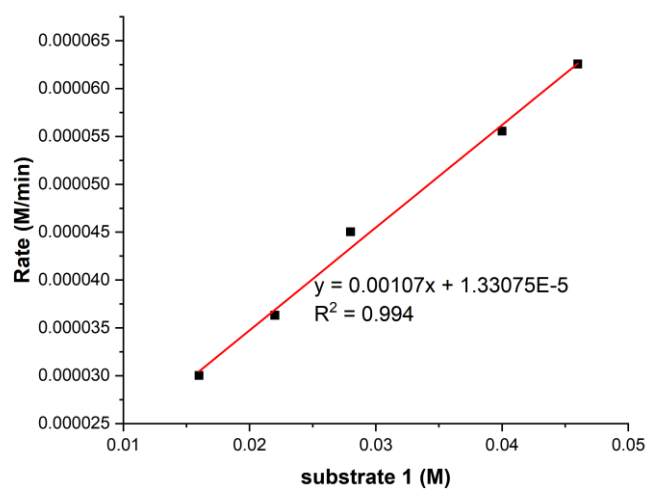
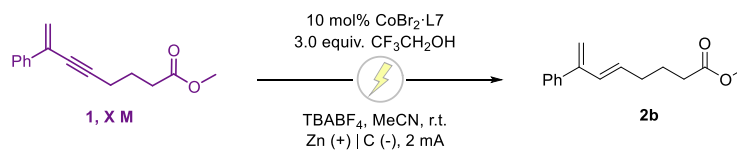


Fig. S15 Rate on the concentration of substrate **1** from the reaction of CoBr₂·L7 (0.004 M), CF₃CH₂OH (0.12 M), TBABF₄ (0.1 M), Current = 2 mA with 0.016 M, 0.022 M, 0.028 M, 0.04 M, 0.046 M of

substrate 1.

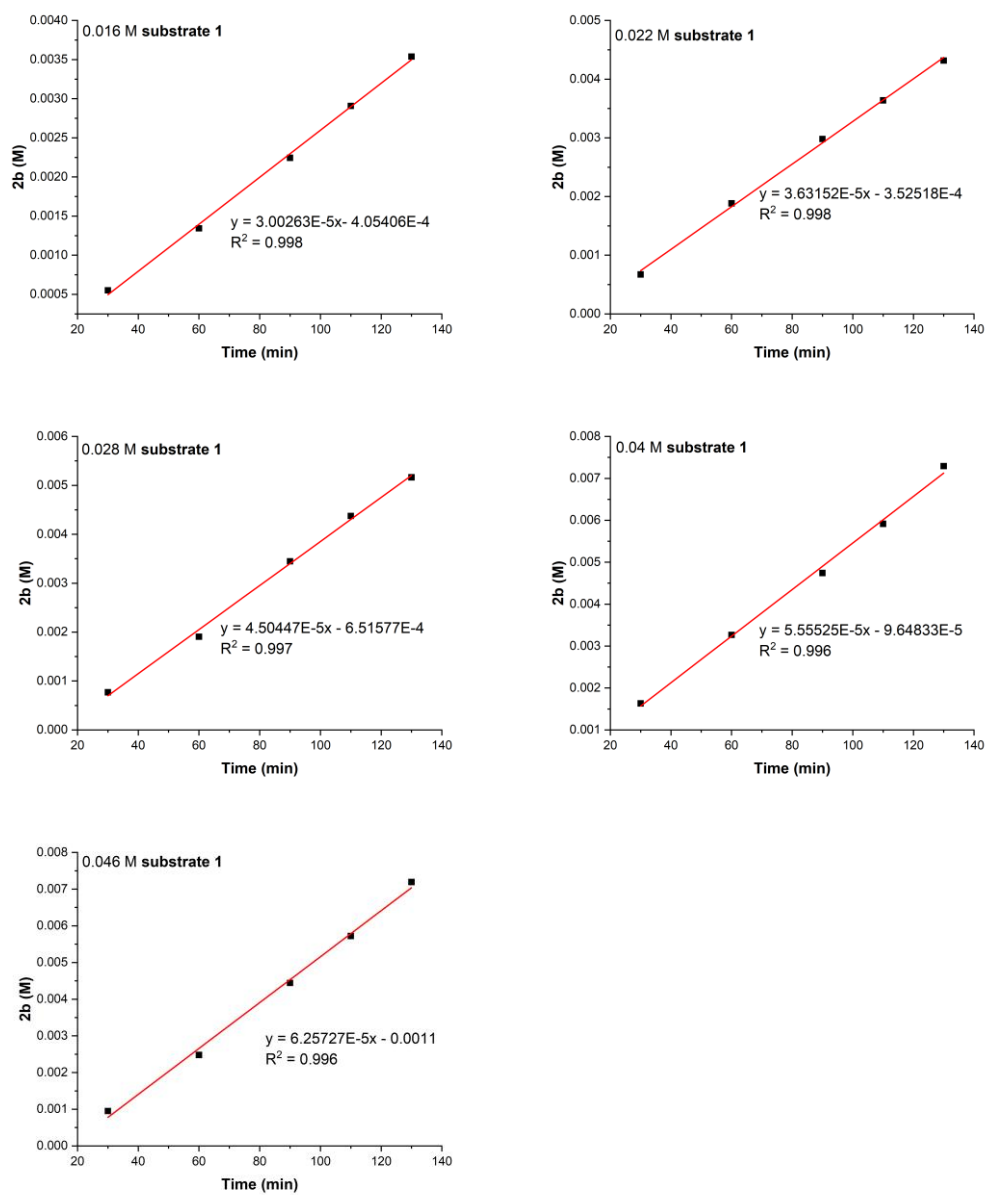


Fig. S16 Plot of the rise of product from the reaction of $\text{CoBr}_2 \cdot \text{L7}$ (0.004 M), $\text{CF}_3\text{CH}_2\text{OH}$ (0.12 M), TBABF_4 (0.1 M), Current = 2 mA with 0.016 M, 0.022 M, 0.028 M, 0.04 M, 0.046 M of **substrate 1**.

The rate on the concentration of CF₃CH₂OH

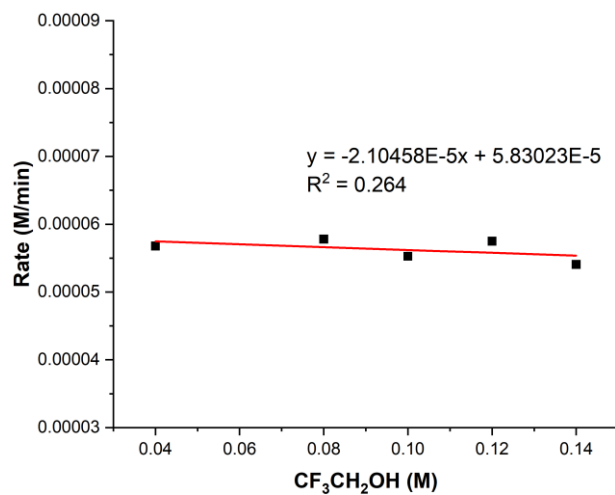
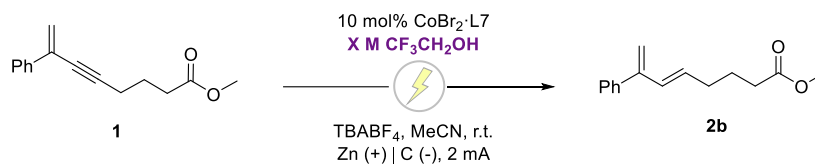
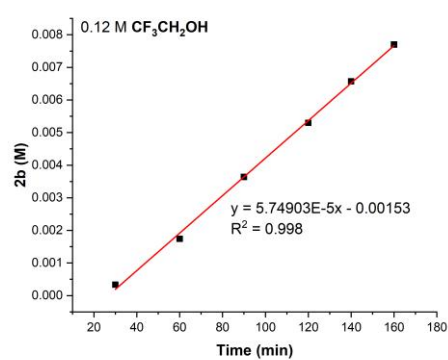
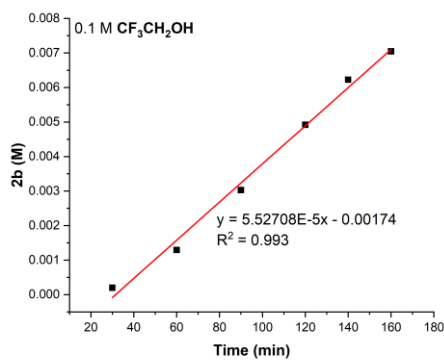
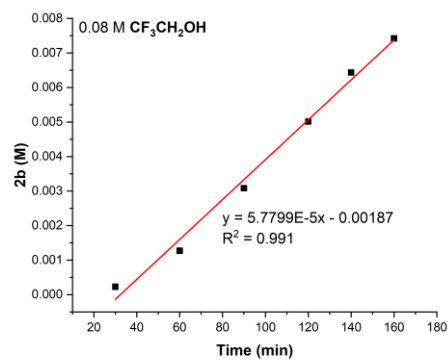
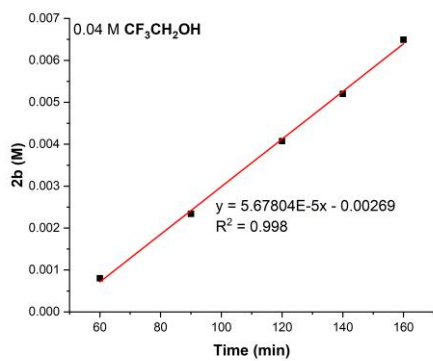


Fig. S17 Rate on the concentration of CF₃CH₂OH from the reaction of CoBr₂·L7 (0.004 M), substrate 1 (0.04 M), TBABF₄ (0.1 M), Current = 2 mA with 0.04 M, 0.08 M, 0.1 M, 0.12 M, 0.14 M of CF₃CH₂OH.



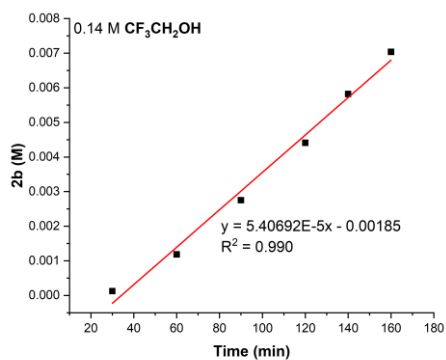


Fig. S18 Plot of the rise of product from the reaction of $\text{CoBr}_2 \cdot \text{L7}$ (0.004 M), substrate **1** (0.04 M), TBABF_4 (0.1 M), Current = 2 mA with 0.04 M, 0.08 M, 0.1 M, 0.12 M, 0.14 M of $\text{CF}_3\text{CH}_2\text{OH}$.

The rate on the value of current

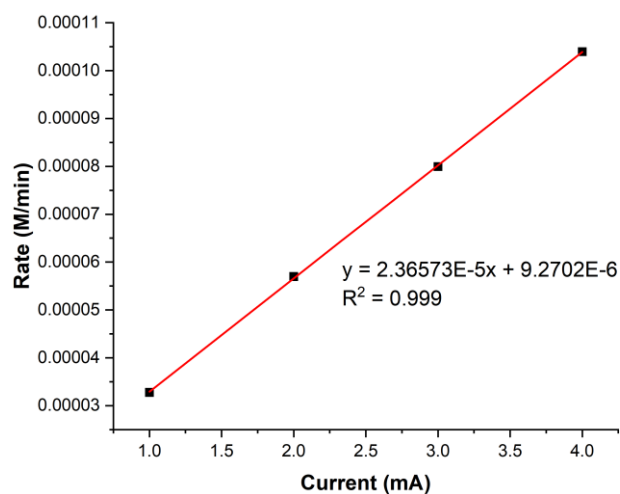
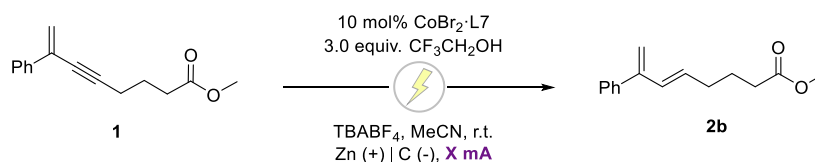


Fig. S19 Rate on the concentration of **Current** from the reaction of $\text{CoBr}_2 \cdot \text{L7}$ (0.004 M), substrate **1** (0.04 M), $\text{CF}_3\text{CH}_2\text{OH}$ (0.12 M), TBABF_4 (0.1 M) with 1 mA, 2 mA M, 3 mA, 4 mA of **Current**.

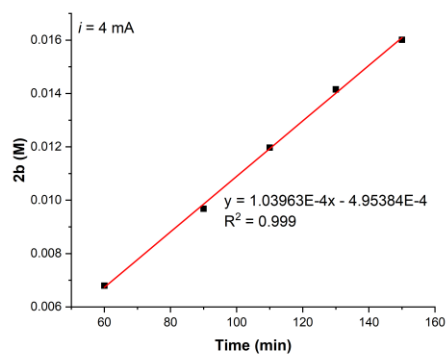
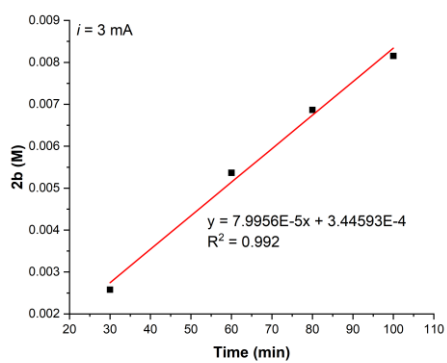
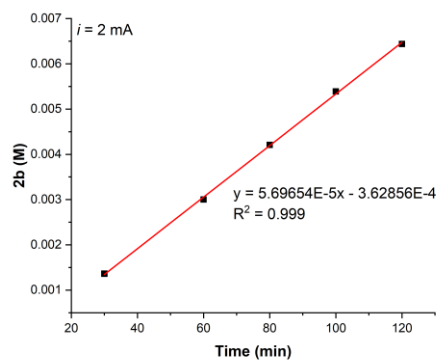
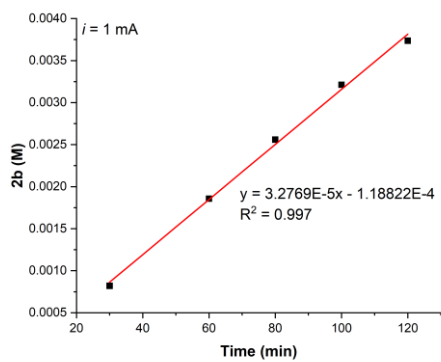


Fig. S20 Plot of the rise of product from the reaction of $\text{CoBr}_2 \cdot \text{L7}$ (0.004 M), substrate **1** (0.04 M), $\text{CF}_3\text{CH}_2\text{OH}$ (0.12 M), TBABF_4 (0.1 M) with 1 mA, 2 mA M, 3 mA, 4 mA of Current.

Cyclic Voltammetry (CV) Experiments

Cyclic voltammetry was recorded using a CHI760E potentiostat at room temperature. A glassy carbon electrode (3 mm diameter), Pt wire (0.5 mm diameter), and Ag/AgNO₃ (0.1 M in 0.1 M TBAPF₆ in CH₃CN) were used as the working, counter, and reference electrodes, respectively. All potentials were referenced to an internal Fc/Fc⁺ standard added in the last step of each experiment. Prior to the experiment, glassy carbon electrode was polished using 0.5 μm α-Al₂O₃ polishing powder. Before beginning electrochemical testing, the solution was purged with Ar gas for 10 minutes to remove oxygen from the solution. In all electrochemical studies, MeCN was used as the solvent in the presence of 0.1 M TBAPF₆ as supporting electrolyte. Unless otherwise noted, scan rate = 100 mV/s.

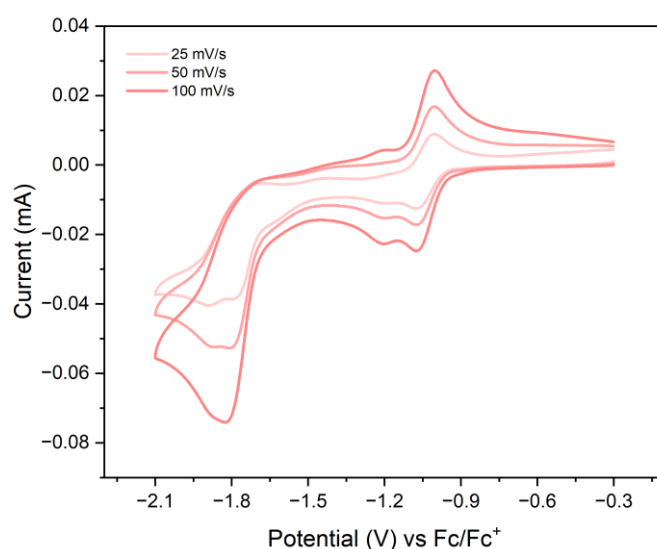


Fig. S21 Cyclic voltammetric profiles of 3.0 mM CoBr₂·L4 in 0.1 M TBAPF₆ in MeCN (8 mL) at varied sweep rates. Two reduction peaks at -1.07 V and -1.82 V vs. Fc⁺/Fc corresponds to the Co(II/I) and Co(I/0) reduction processes.

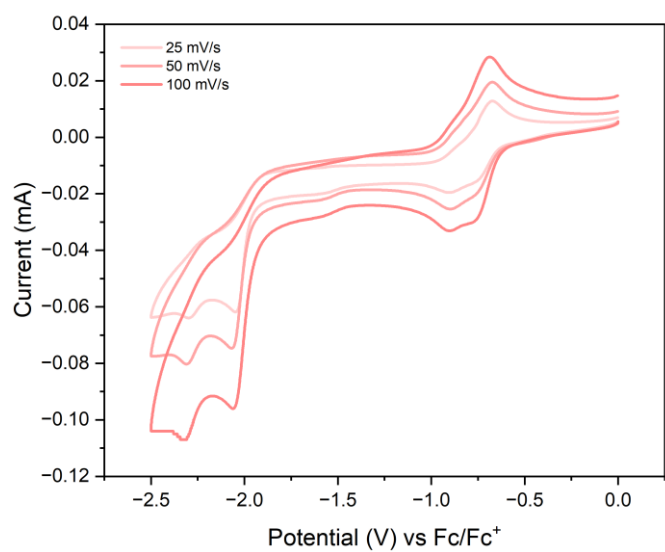


Fig. S22 Cyclic voltammetric profiles of 3.0 mM $\text{CoBr}_2 \cdot \text{L7}$ in 0.1 M TBAPF_6 in MeCN (8 mL) at varied sweep rates. Two reduction peaks at -0.83 V and -2.05 V vs. Fc^+/Fc corresponds to the Co(II/I) and Co(I/0) reduction processes.

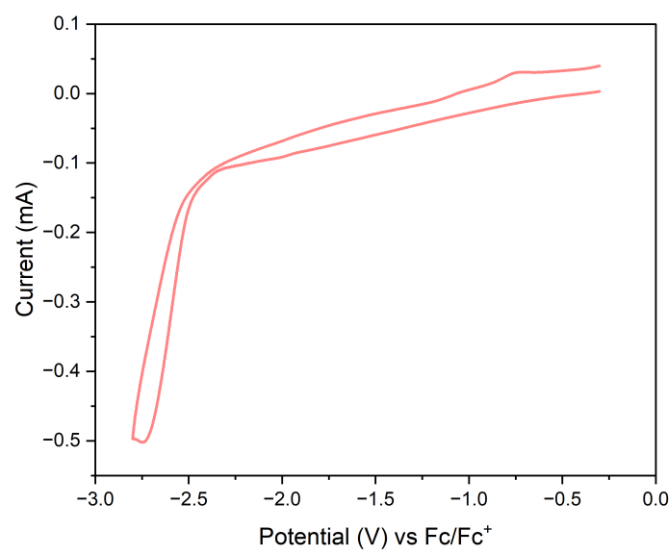


Fig. S23 Cyclic voltammetric profiles of 15.0 mM substrate **1** in 0.1 M TBAPF_6 in MeCN (8 mL). A reduction peak at -2.74 V vs. Fc^+/Fc corresponds to the substrate **1** reduction processes.

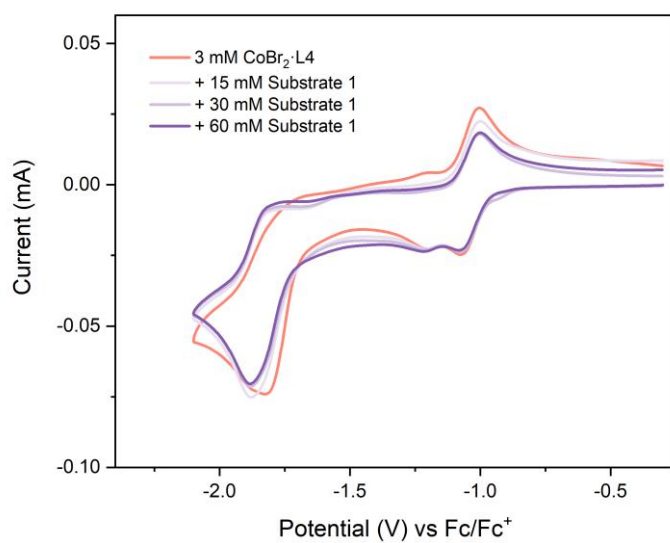


Fig. S24 Cyclic voltammetry of 3.0 mM CoBr₂·L4 in 0.1 M TBAPF₆ in MeCN (8 mL) in the presence of varying concentrations of substrate **1**. The cathodic shift may correspond to the coordination of substrate **1** with Co(0).

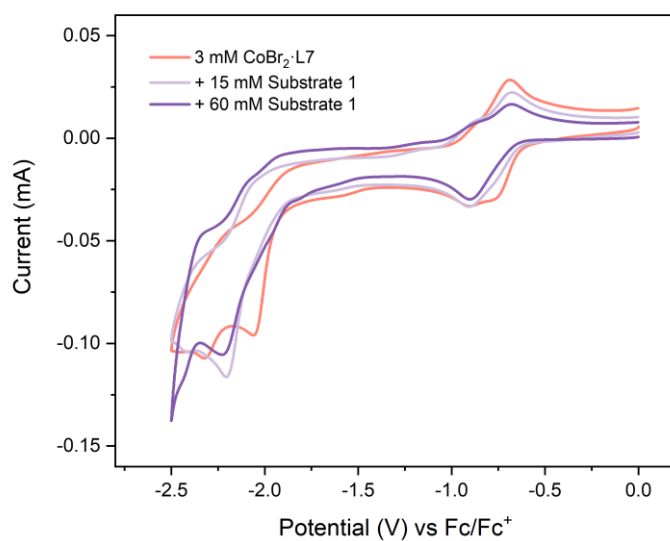


Fig. S25 Cyclic voltammetry of 3.0 mM CoBr₂·L7 in 0.1 M TBAPF₆ in MeCN (8 mL) in the presence of varying concentrations of substrate **1**. The cathodic shift may correspond to the coordination of substrate **1** with Co(0).

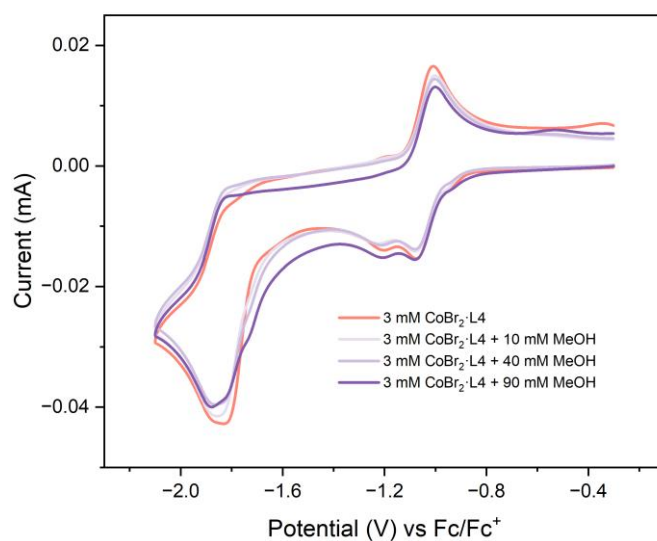


Fig. S26 Cyclic voltammetry of 3.0 mM CoBr₂·L4 in 0.1 M TBAPF₆ in MeCN (8 mL) in the presence of varying concentrations of MeOH.

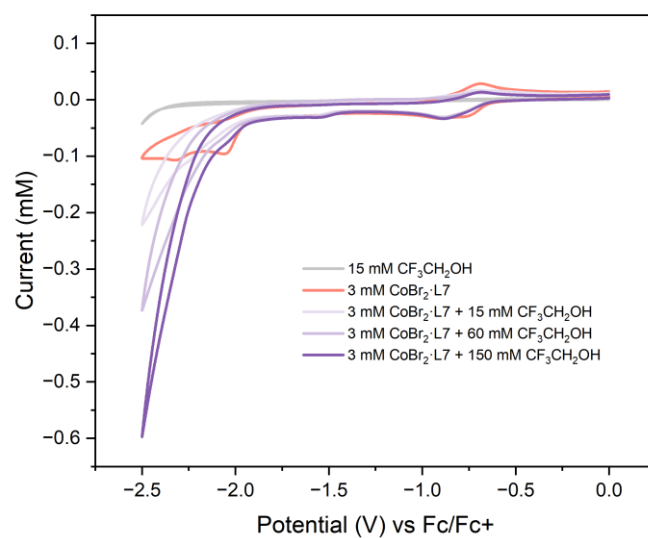


Fig. S27 Cyclic voltammetry of 3.0 mM CoBr₂·L7 in 0.1 M TBAPF₆ in MeCN (8 mL) in the presence of varying concentrations of CF₃CH₂OH. By increasing the concentration of CF₃CH₂OH, the catalytic current increases, the process corresponding to the production of Co(II)-H and the release of H₂.

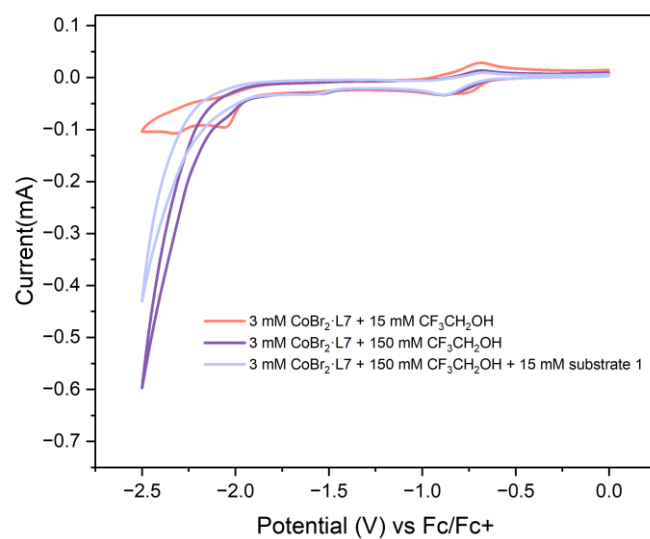


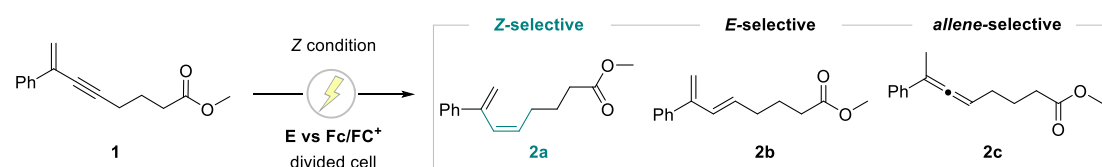
Fig. S28 Cyclic voltammetry of 3.0 mM CoBr₂·L7 and 150 mM CF₃CH₂OH in 0.1 M TBAPF₆ in MeCN (8 mL) in the presence of substrate **1**. By adding the substrate **1**, the catalytic current decreases, the process corresponding to the reaction of substrate **1** with Co(II)-H.

Controlled Potential Electrolysis

Controlled potential electrolysis was proceeded using a CHI760E potentiostat. Electrolyses were conducted in a two-chamber system using a glass frit to separate the two compartments. The two-chamber system was equipped with magnetic stir bars, and in each of the chambers TBAPF₆ (0.1 M) was dissolved in MeCN in the glove box. Either according to the general procedure of *Z*-selective, *E*-selective or *allene*-selective hydrogenation reactions, using substrate **1**, cobalt complex, proton source and additives, then equipped with the electrodes. The second chamber was equipped with Ag/AgNO₃ (0.1 M in 0.1 M TBAPF₆ in CH₃CN) reference electrode. Then the mixture was respectively electrolyzed under a constant voltage (vs Fc/Fc⁺). After reaction was completed, a small amount of the reaction mixture was taken out and was sampled for GC analysis.

Z-selective hydrogenation

Table S7 Controlled potential electrolysis of *Z*-selective hydrogenation

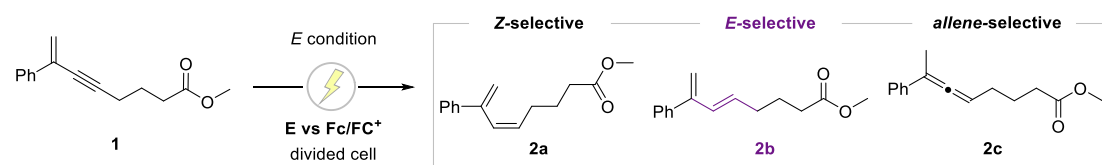


Entry	E vs. Fc/FC ⁺	Yield(%)	Z/E(2a:2b)	r.r.(2a+2b:2c)
1	-1.1 V	n.p.	-	-
2	-1.3 V	trace	-	-
3	-1.7 V	54	>20:1	15:1
4	-1.9 V	59	>20:1	7:1

Reaction Conditions: substrate **1** (0.32 mmol), CoBr₂·L4 (10 mol%), MeOH (4.5 equiv.), DIPEA (4.5 equiv.), TBAPF₆ (0.1 M), Ar, Pt (+) | C felt (-), MeCN (8 mL) at r.t. for 13 h. Yields and selectivity were determined by GC using diphenyl as the internal standard.

E-selective hydrogenation

Table S8 Controlled potential electrolysis of *E*-selective hydrogenation



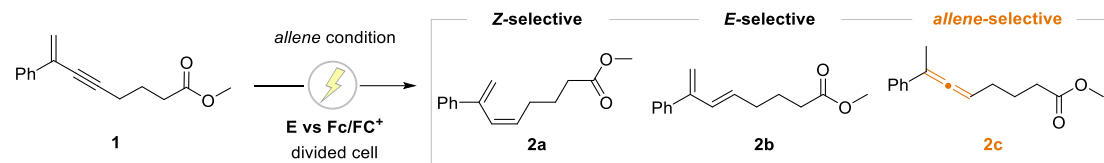
Entry	E vs. Fc/FC ⁺	Yield(%)	E/Z(2b:2a)	r.r.(2a+2b:2c)
1	-0.8 V	n.p.	-	-
2	-1.5 V	51	20:1	12:1
3	-1.6 V	71	>20:1	>20:1
4	-1.7 V	68	19:1	18:1
5	-1.8 V	60	6:1	15:1

Reaction Conditions: substrate **1** (0.32 mmol), CoBr₂·L7 (10 mol%), CF₃CH₂OH (3.0 equiv.), TBAPF₆ (0.1 M), Ar,

Zn (+) | C felt (-), MeCN (8 mL) at r.t. for 22 h. Yields and selectivity were determined by GC using diphenyl as the internal standard.

***allene*-selective hydrogenation**

Table S9 Controlled potential electrolysis of *allene*-selective hydrogenation



Entry	E vs. Fc/FC ⁺	Yield(%)	r.r.(2c:2a+2b)
1	-1.5 V	-	-
2	-2.0 V	-	-
3	-2.5 V	48	>20:1

Reaction Conditions: substrate **1** (0.32 mmol), MeOH (2.0 equiv.), TBAPF₆ (0.1 M), Ar, Zn (+) | SS felt (-), MeCN (8 mL) at r.t. for 10 h. Yields and selectivity were determined by GC using diphenyl as the internal standard.

Controlled Current Electrolysis

Controlled current electrolysis was proceeded using a CHI760E potentiostat. Electrolyses were conducted in a two-chamber system using a glass frit to separate the two compartments. The two-chamber system was equipped with magnetic stir bars, and in each of the chambers TBAPF₆ (0.1 M) was dissolved in MeCN (8 mL) in the glove box. Either according to the general procedure of *Z*-selective, *E*-selective or *allene*-selective hydrogenation reactions, using substrate **1**, cobalt complex, proton source and additives, then equipped with the electrodes. The second chamber was equipped with Ag/AgNO₃ (0.1 M in 0.1 M TBAPF₆ in CH₃CN) reference electrode. Then the mixture was electrolyzed under 2 mA.

Z-selective hydrogenation

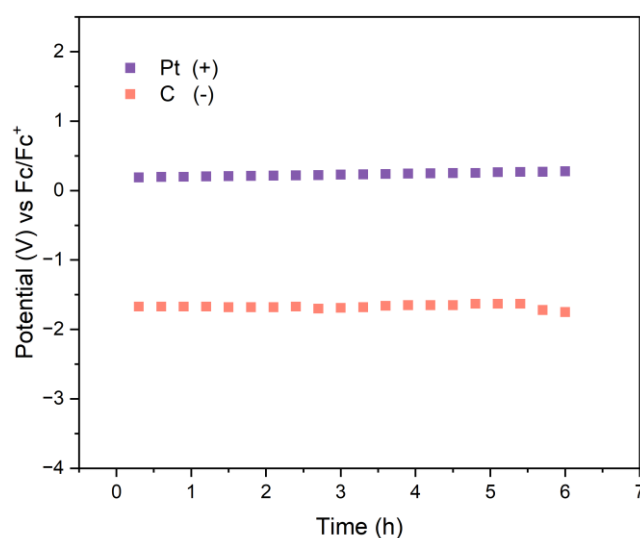


Fig. S29 Potential profiles of Pt felt cathode and carbon felt anode during the electrolysis under the *Z*-selective hydrogenation condition. The average cathode potential is about -1.7 V vs Fc/Fc⁺.

E-selective hydrogenation

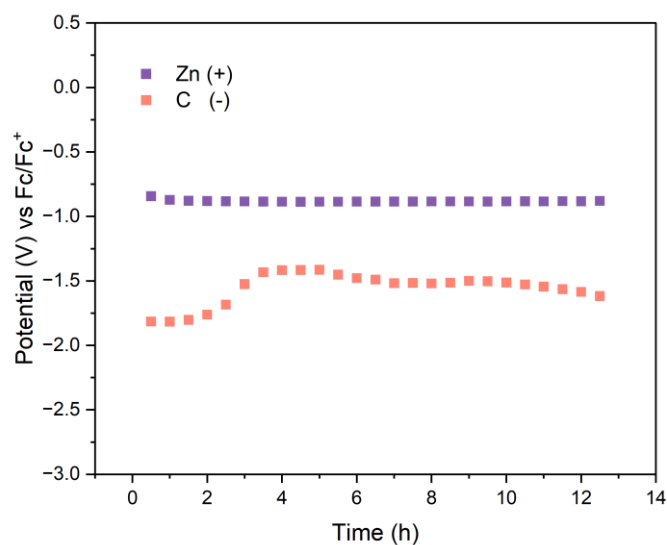


Fig. S30 Potential profiles of carbon felt cathode and Zn felt anode during the electrolysis under the *E*-selective hydrogenation condition. The average cathode potential is about -1.6 V vs Fc/Fc⁺.

allene-selective hydrogenation

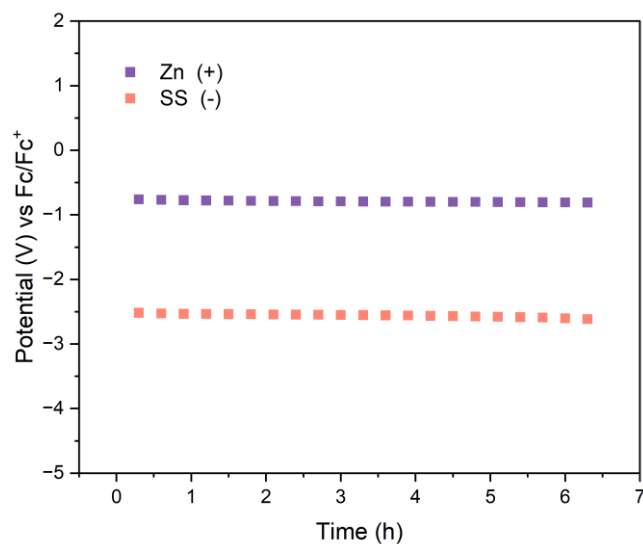


Fig. S31 Potential profiles of stainless steel felt cathode and Zn felt anode during the electrolysis under the *allene*-selective hydrogenation condition. The average cathode potential is about -2.5 V vs Fc/Fc⁺.

Hydrogenation Product Interconversion Experiments

General method for interconversion experiments: according to the general procedure of hydrogenation reactions, diphenyl was used as an internal standard. A small amount of the reaction mixture was taken out by syringe. The obtained mixture was immediately diluted by EA (0.5 mL), which was further sampled for GC analysis.

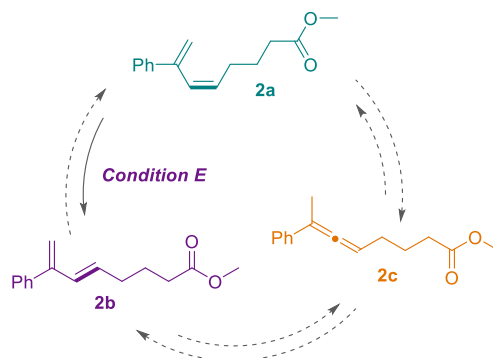
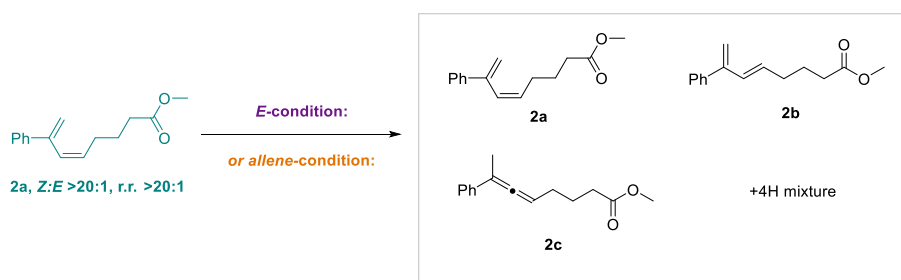


Fig. S32 Inter-isomerization of hydrogenated products

Z- product isomerization

Table S10 Z- product isomerization experiment



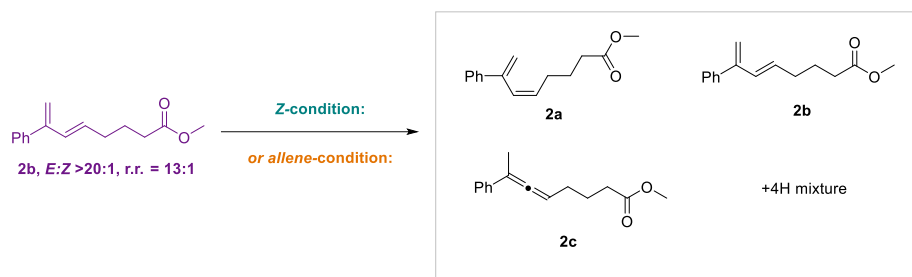
Entry	Condition	2a:2b:2c:+4H mixture
1	<i>E</i> -condition	6:66:4:14
2	allene-condition	50:3:3:44

Reaction Conditions: **2a** ($Z:E >20:1$, r.r. $>20:1$, 0.2 mmol). Yields and selectivity were determined by GC using diphenyl as the internal standard.

According to the results of entry 1, the *Z*- product can be converted into the *E*- product under standard *E*- conditions, but it cannot be concluded that the *E*- product is formed by isomerization of the *Z*- product under standard *E*- selective conditions. This requires further experimental verification or ruling out.

E- product isomerization

Table S11 *E*- product isomerization experiment



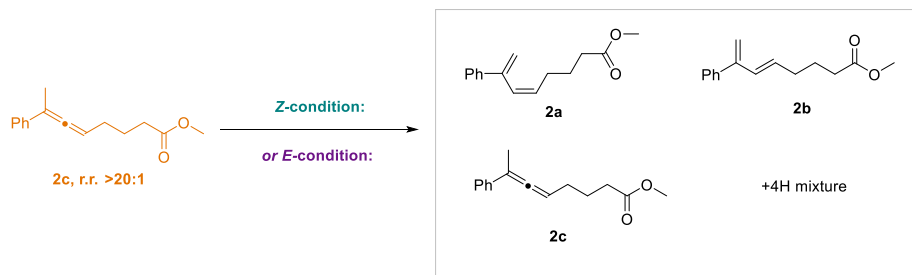
Entry	Condition	2a:2b:2c:+4H mixture
1	<i>Z</i> -condition	1:40:0:59
2	<i>allene</i> -condition	1:37:4:58

Reaction Conditions: **2b** ($E:Z >20:1$, $r.r. = 13:1$, 0.2 mmol). Yields and selectivity were determined by GC using diphenyl as the internal standard.

According to the results of entry 1, the *E*- product can't be converted to the *Z*- product under standard *Z*-selective conditions.

allene- product isomerization

Table S12 *allene*- product isomerization experiment



Entry	Condition	2a:2b:2c:+4H mixture
1	<i>Z</i> -condition	0:0:55:45
2	<i>E</i> -condition	4:6:77:13

Reaction Conditions: **2c** ($r.r. >20:1$, 0.2 mmol). Yields and selectivity were determined by GC using diphenyl as the internal standard.

Based on the results of entry 1, the *allene*- product cannot be converted to the *Z*- product under standard *Z*-selective conditions.

Based on the results of entry 2, the *allene*- product can be converted into the *E*- product in small amounts under standard *E*-selective conditions, but the *Z/E* selectivity is poor. It can be inferred that under standard *E*-selective conditions, the *E*- product is not formed through the isomerization of the allene.

Monitor the Reaction Process

General method: according to the general procedure of *E*-hydrogenation reactions, using substrate **1**, CoBr₂/L7 and CF₃CH₂OH, and diphenyl was used as an internal standard. The stopwatch was immediately started when the current was turned on. A small amount of the reaction mixture was taken out by syringe. The obtained mixture was immediately diluted by EA (0.5 mL), which was further sampled for GC analysis.

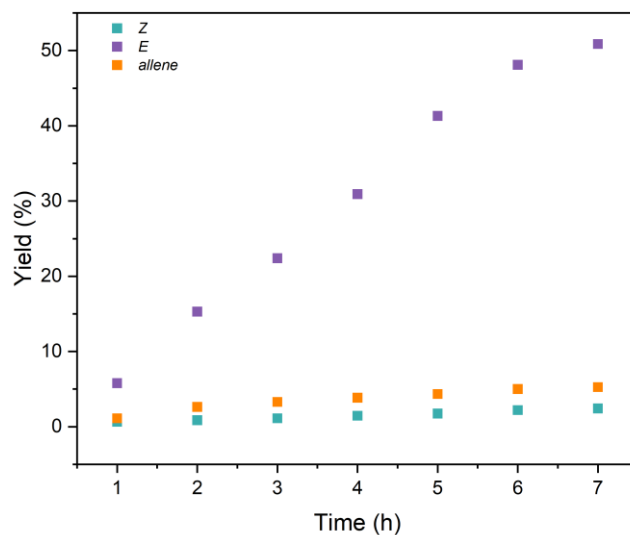
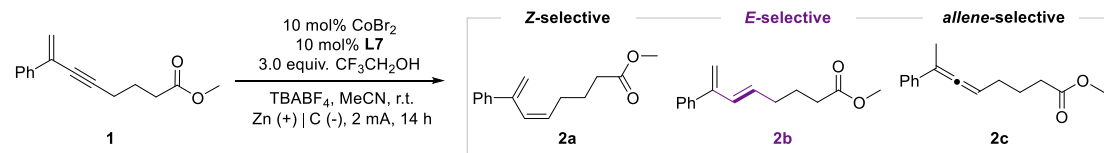


Fig. S33 Reaction process profiles under standard E-selective condition

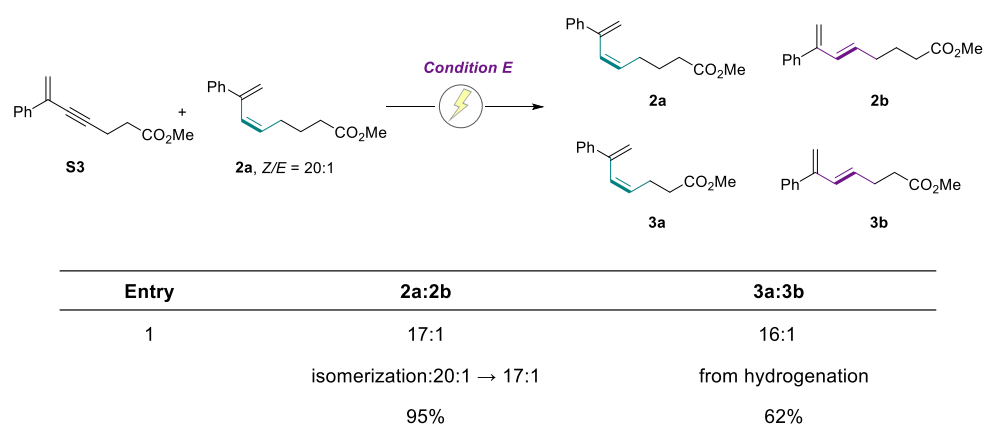
Reaction Conditions: substrate **1** (0.2 mmol), CoBr₂ (10 mol%), L7 (10 mol%), CF₃CH₂OH (3.0 equiv.), TBABF₄ (0.1 M), Ar, 2 mA, undivided cell, Zn (+) | C felt (-), in MeCN (5 mL) at r.t. Yields and selectivity were determined by GC using diphenyl as the internal standard.

Based on the results of monitoring the reaction, under standard *E*-selective conditions, we did not detect the formation or consumption of *Z*- products. This may indicate that under *E*- conditions, the formation of *E*- products may occur directly through hydrogenation, rather than through isomerization of *Z*- products.

Crossover Experiments

General method: according to the general procedure of *E*-hydrogenation reactions, using substrate **S3**, **2a**, CoBr₂/L7 and CF₃CH₂OH, and diphenyl was used as an internal standard. The resulting solution was electrolyzed under a constant current of 2 mA for 14 h at room temperature. A small amount of the reaction mixture was taken out by syringe. The obtained mixture was immediately diluted by EA (0.5 mL), which was further sampled for GC analysis. Additionally, crude NMR spectra were used to assist in analyzing the selectivity.

Table S13 Crossover experiments



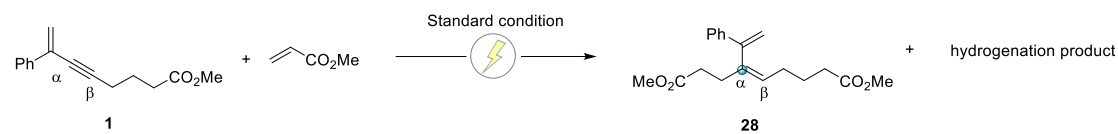
Reaction Conditions: substrate **S3** (0.2 mmol), **2a** (*Z*:*E* = 20:1, 0.2 mmol), CoBr₂ (10 mol%), L7 (10 mol%), CF₃CH₂OH (3.0 equiv.), TBABF₄ (0.1 M), Ar, 2 mA, undivided cell, Zn (+) | C felt (-), in MeCN (5 mL) at r.t. for 14 h. Selectivity were determined by GC and ¹H NMR.

Based on the aforementioned crossover experiments, it is evident that hydrogenation proceeds much faster than isomerization. Combined with the experimental observation that no accumulation or consumption of the *Z*-isomer product was detected during the monitoring of the reaction progress, it can be concluded that under standard *E*-selective conditions, the *E*-isomer is obtained directly through hydrogenation rather than through isomerization from the *Z*-isomer.

C-Co Bond Trapping Experiments

General method: according to the general procedure of *E*- or *Z*-hydrogenation reactions.

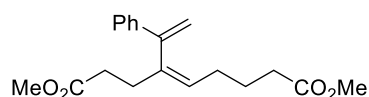
Table S14 C-Co bond trapping experiments



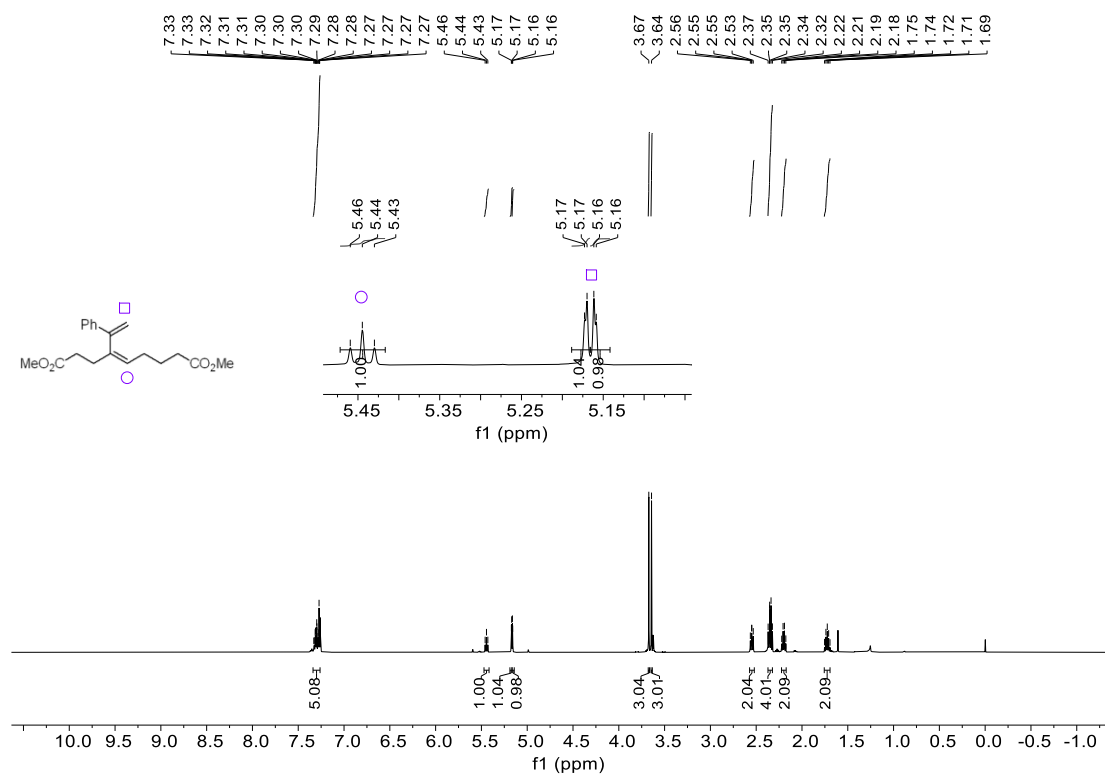
Entry	Condition	Yield of 28
1	<i>Z</i> -condition	4%
2	<i>E</i> -condition	10%

Reaction Conditions: substrate **1** (0.2 mmol), methyl acrylate (0.2 mmol). Yields were determined by GC using diphenyl as the internal standard.

dimethyl (*Z*)-4-(1-phenylvinyl)non-4-enedioate



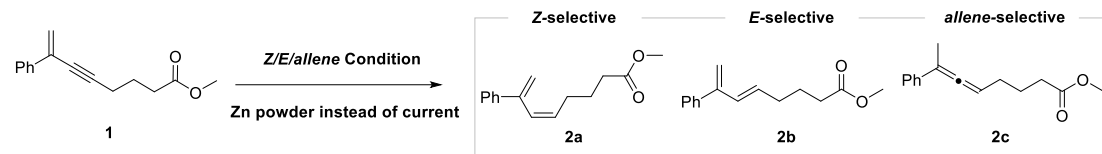
¹H NMR (500 MHz, CDCl₃) δ 7.34-7.26 (m, 5H), 5.44 (t, *J* = 7.4 Hz, 1H), 5.17 (d, *J* = 1.5 Hz, 1H), 5.16 (d, *J* = 1.5 Hz, 1H), 3.67 (s, 3H), 3.64 (s, 3H), 2.57-2.52 (m, 2H), 2.37-2.32 (m, 4H), 2.20 (q, *J* = 7.4 Hz, 2H), 1.76-1.69 (m, 2H).



Control Experiments

General method: according to the general procedure of *E*- or *Z*- hydrogenation reactions.

Table S15 Control experiments

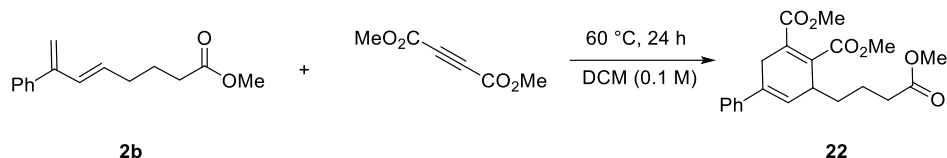


Entry	Condition	Yield
1	Condition <i>allene</i> , r.t.	-
2	Condition <i>allene</i> , 60 °C	-
3	Condition <i>allene</i> , 90 °C	-
4	Condition <i>Z</i> , r.t.	-
5	Condition <i>Z</i> , 60 °C	-
6	Condition <i>Z</i> , 90 °C	-
7	Condition <i>E</i> , r.t.	13% yield, <i>E/Z</i> = 12:1, r.r. = 8:1
8	Condition <i>E</i> , 60 °C	13% yield, <i>E/Z</i> = 7:1, r.r. > 20:1
9	Condition <i>E</i> , 90 °C	12% yield, <i>E/Z</i> = 3:1, r.r. > 20:1

Reaction Conditions: substrate **1** (0.2 mmol), Zn powder (2.0 equiv.) instead of current. Yields and selectivity were determined by GC using diphenyl as the internal standard.

6. Synthetic Applications

dimethyl 5-(4-methoxy-4-oxobutyl)-2,5-dihydro-[1,1'-biphenyl]-3,4-dicarboxylate



To a solution of **2b** (46 mg, 0.2 mmol) in DCM (2 mL) was added dimethyl acetyl dicarboxylate (30 μL , 0.24 mmol). After stirring at 60 $^\circ\text{C}$ for 24 hours, the solution was cooled to room temperature and concentrated in vacuo. The concentrated residue was purified by column chromatography on silica gel with a gradient eluent of PE/EA to give the desired products⁵.

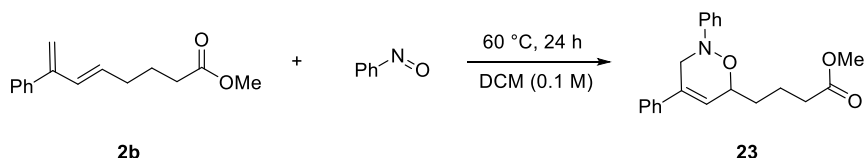
Compound **22** was isolated via column chromatography (PE/EA=10:1~3:1) as colorless oil (32.0 mg, 43% yield).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.43 (d, $J = 7.6$ Hz, 2H), 7.35 (t, $J = 7.4$ Hz, 2H), 7.29 (d, $J = 7.3$ Hz, 1H), 6.06 (s, 1H), 3.83 (s, 3H), 3.80 (s, 3H), 3.64 (s, 3H), 3.55-3.38 (m, 2H), 3.37-3.22 (m, 1H), 2.30 (t, $J = 5.5$ Hz, 2H), 1.73-1.58 (m, 4H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 173.73, 169.02, 167.41, 139.70, 138.51, 133.37, 130.86, 128.50, 127.68, 125.21, 123.52, 52.32, 51.54, 38.84, 34.07, 33.42, 29.43, 20.83.

HRMS (APCI) calcd. for $\text{C}_{21}\text{H}_{25}\text{O}_6$ $[\text{M}+\text{H}]^+$: 373.1646; found: 373.1642.

methyl 4-(2,4-diphenyl-3,6-dihydro-2H-1,2-oxazin-6-yl)butanoate



To a solution of **2b** (46 mg, 0.2 mmol) in DCM (2 mL) was added nitrosobenzene (32.2 mg, 0.3 mmol). After stirring at 60 $^\circ\text{C}$ for 24 hours, the solution was cooled to room temperature and concentrated in vacuo. The concentrated residue was purified by column chromatography on silica gel with a gradient eluent of PE/EA to give the desired products⁵.

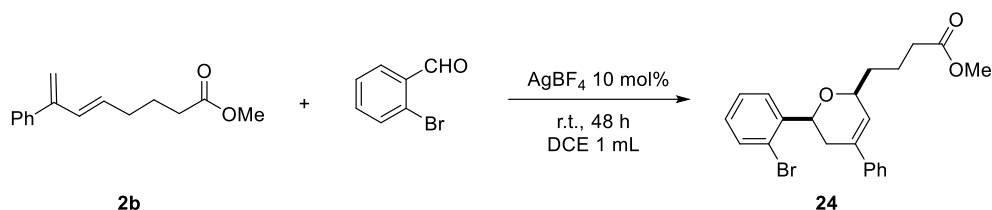
Compound **23** was isolated via column chromatography (PE/EA=20:1~10:1) as colorless oil (44.5 mg, 66% yield).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.44-7.30 (m, 7H), 7.20 (d, $J = 8.0$ Hz, 2H), 7.01 (t, $J = 7.3$ Hz, 1H), 6.19 (s, 1H), 4.71 (s, 1H), 4.23 (d, $J = 15.3$ Hz, 1H), 4.09 (d, $J = 15.3$ Hz, 1H), 3.67 (s, 3H), 2.41 (t, $J = 7.1$ Hz, 2H), 1.99-1.74 (m, 4H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 173.92, 150.50, 138.14, 134.08, 128.93, 128.68, 127.89, 125.72, 125.04, 122.35, 115.91, 77.10, 53.59, 51.58, 33.97, 33.07, 21.20.

HRMS (APCI) calcd. for $\text{C}_{21}\text{H}_{24}\text{NO}_3$ $[\text{M}+\text{H}]^+$: 338.1751; found: 338.1745.

methyl 4-((2*S*,6*S*)-6-(2-bromophenyl)-4-phenyl-5,6-dihydro-2*H*-pyran-2-yl)butanoate



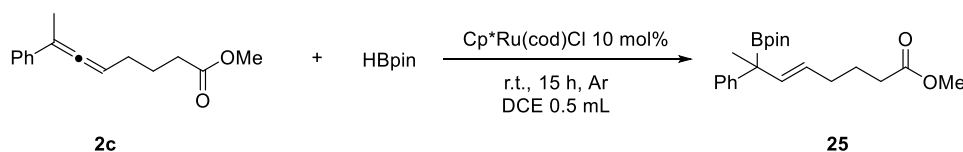
In a glove box, an oven dried 10 mL vial was charged with **2b** (92 mg, 0.4 mmol), AgBF₄ (2 mg, 10 mol%), 2-bromobenzaldehyde (12 μ L, 0.2 mmol) and DCE (1 mL). The vial was capped and removed from the glove box. The reaction mixture was stirred at room temperature for 48 h, and then concentrated in vacuo. The residue was purified by silica gel flash column chromatography to give the desired product⁶. Compound **24** was isolated via column chromatography (PE/EA=20:1~10:1) as colorless oil (48.0 mg, 55% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.66 (d, J = 7.7 Hz, 1H), 7.54 (d, J = 8.0 Hz, 1H), 7.40 (t, J = 7.8 Hz, 3H), 7.33 (t, J = 7.6 Hz, 2H), 7.25 (d, J = 4.5 Hz, 1H), 7.15 (t, J = 7.7 Hz, 1H), 6.10 (d, J = 2.5 Hz, 1H), 5.01 (dd, J = 10.7, 3.0 Hz, 1H), 4.58-4.48 (m, 1H), 3.67 (d, J = 1.3 Hz, 3H), 2.86 (dd, J = 16.5, 3.2 Hz, 1H), 2.46-2.37 (m, 3H), 1.93-1.72 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 174.11, 142.05, 139.91, 135.11, 132.48, 128.82, 128.48, 127.92, 127.55, 127.49, 125.67, 124.92, 121.63, 75.26, 75.07, 51.54, 35.00, 34.09, 33.51, 20.65.

HRMS (ESI) calcd. for C₂₂H₂₃BrO₃Na⁺ [M+H]⁺: 437.0723; found: 437.0724.

methyl (*E*)-7-phenyl-7-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)oct-5-enoate



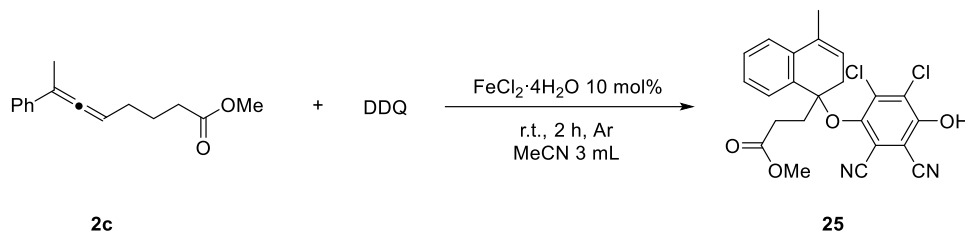
In a glove box, an oven-dried 10 mL vial was charged with **2b** (46 mg, 0.2 mmol), Cp*Ru(cod)Cl (7.6 mg, 10 mol%), pinacolborane (43 μ L, 0.3 mmol) and DCE (0.5 mL). The vial was capped and removed from the glove box. The reaction mixture was stirred at room temperature for 12 h, and then concentrated in vacuo. The residue was purified by silica gel flash column chromatography to give the desired product⁷. Compound **25** was isolated via column chromatography (PE/EA=20:1~10:1) as colorless oil (53.7 mg, 75% yield).

¹H NMR (500 MHz, CDCl₃) δ 7.27-7.23 (m, 3H), 7.16-7.12 (m, 1H), 5.84 (dt, J = 15.6, 1.4 Hz, 1H), 5.40 (dt, J = 15.7, 6.8 Hz, 1H), 3.66 (s, 3H), 2.34 (t, J = 7.7 Hz, 2H), 2.17-2.10 (m, 2H), 1.78-1.70 (m, 2H), 1.39 (s, 3H), 1.22 (d, J = 7.9 Hz, 12H).

¹³C NMR (126 MHz, CDCl₃) δ 174.36, 147.50, 136.98, 128.26, 127.42, 127.34, 125.46, 83.59, 51.56, 33.45, 32.40, 25.00, 24.60, 24.59, 23.46.

HRMS (APCI) calcd. for C₂₁H₃₂BO₄⁺ [M+H]⁺: 359.2389; found: 359.2382.

methyl 3-(1-(2,3-dichloro-5,6-dicyano-4-hydroxyphenoxy)-4-methyl-1,2-dihydronaphthalen-1-yl)propanoate



In a glove box, an oven dried 10 mL vial was charged with **2b** (46 mg, 0.2 mmol), FeCl₂·4H₂O (2.9 mg, 10 mol%), DDQ (45.4 mg, 0.2 mmol) and MeCN (3 mL). The vial was capped and removed from the glove box. The reaction mixture was stirred at room temperature for 48 h, and then concentrated in vacuo. The residue was purified by silica gel flash column chromatography to give the desired product⁸.

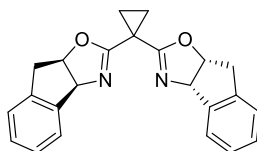
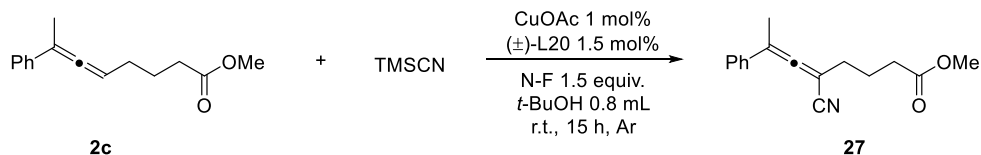
Compound **26** was isolated via column chromatography (PE/EA=20:1~10:1) as pale yellow oil (53.7 mg, 63% yield).

¹H NMR (500 MHz, CDCl₃) δ 7.55-7.50 (m, 2H), 7.45-7.42 (m, 2H), 6.18 (t, *J* = 7.8 Hz, 1H), 3.60 (s, 3H), 2.26 (q, *J* = 7.3 Hz, 2H), 2.07 (q, *J* = 7.7 Hz, 2H), 1.77-1.68 (m, 2H), 1.65 (s, 3H).

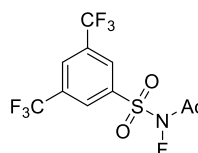
¹³C NMR (126 MHz, CDCl₃) δ 176.92, 175.20, 173.27, 146.18, 145.52, 138.07, 136.61, 131.98, 129.76, 129.70, 126.42, 113.00, 112.72, 63.35, 51.91, 33.15, 28.49, 24.89, 23.58.

HRMS (APCI) calcd. for C₂₃H₁₉Cl₂N₂O₄⁺ [M+H]⁺: 457.0717; found: 457.0710.

methyl 5-cyano-7-phenylocta-5,6-dienoate



(±)-L20



N-F

In a dried sealed 10 mL Schlenk tube, (±)-L20 (10.8 mg, 0.03 mmol), CuOAc (2.5 mg, 0.02 mmol) were dissolved in 2.0 mL of DCE. The resulting mixture was stirred for 1 hour. Then a catalyst solution (0.2 mL, 0.002 mol %) was added to a 10 mL sealed tube under an argon atmosphere, and the solvent was removed under vacuum. The residue was redissolved in 0.8 mL of ultra-dry *t*-BuOH. After that, N-F (134 mg, 0.3 mmol), **2c** (46mg, 0.2 mmol), and TMSCN (50 μL, 0.4 mmol) were added sequentially. After stirring at room temperature for 15 h, the reaction mixture was concentrated in vacuo. The concentrated residue was purified by column chromatography on silica gel with a gradient eluent of PE/EA to give the desired products⁹.

Compound **27** was isolated via column chromatography (PE/EA=20:1~10:1) as pale yellow oil (38.0 mg, 75% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.40-7.29 (m, 5H), 3.66 (s, 3H), 2.38 (dt, *J* = 14.0, 7.5 Hz, 4H), 2.19 (s, 3H), 1.94-1.86 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 212.53, 173.16, 133.59, 128.81, 128.54, 126.40, 115.27, 107.54, 83.00,

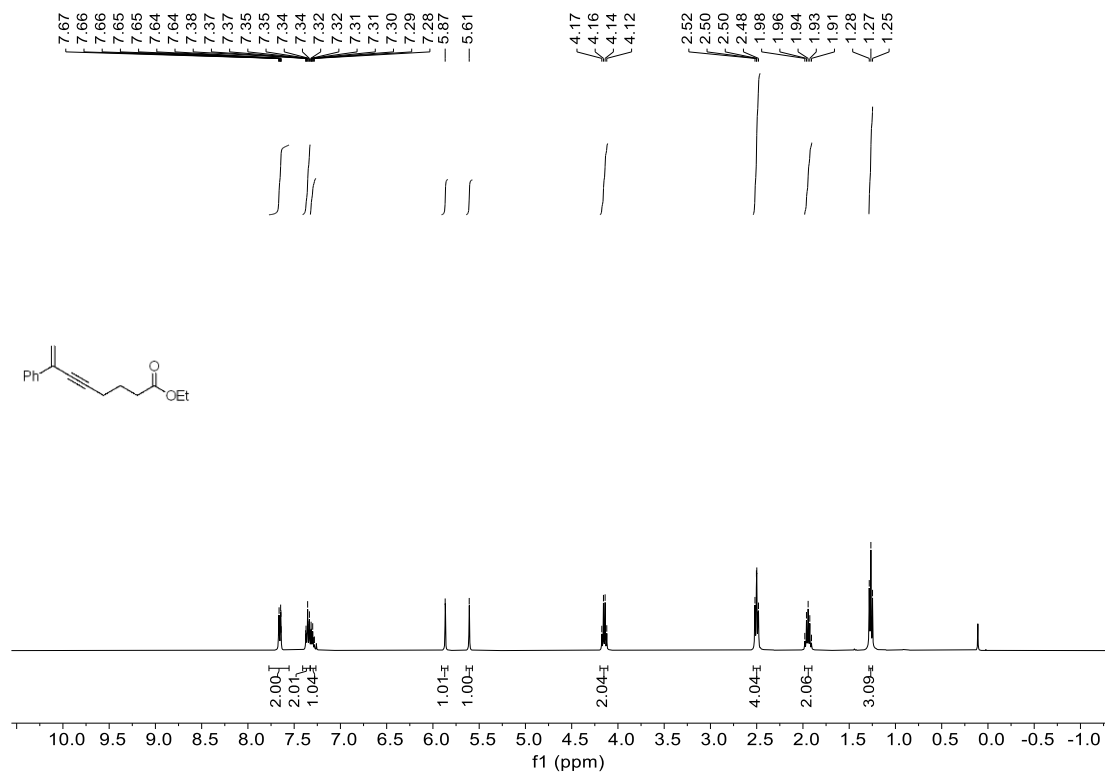
51.70, 32.77, 31.07, 22.84, 16.59.

HRMS (APCI) calcd. for $C_{16}H_{18}NO_2^+$ $[M+H]^+$: 256.1333; found: 256.1325.

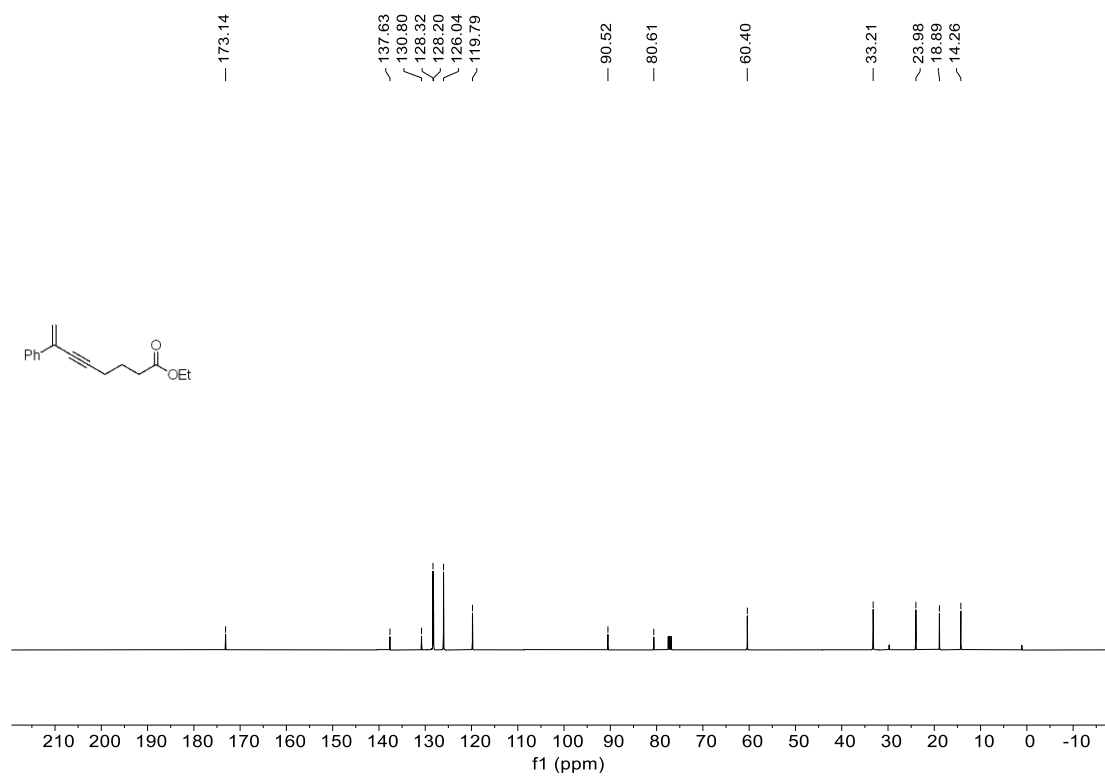
7. References

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- 7 Tan, Y.-X. *et al.* Ruthenium-Catalyzed α -Regioselective Hydroboration of Allenes. *Angew. Chem. Int. Ed.* **64**, e202420370 (2025).
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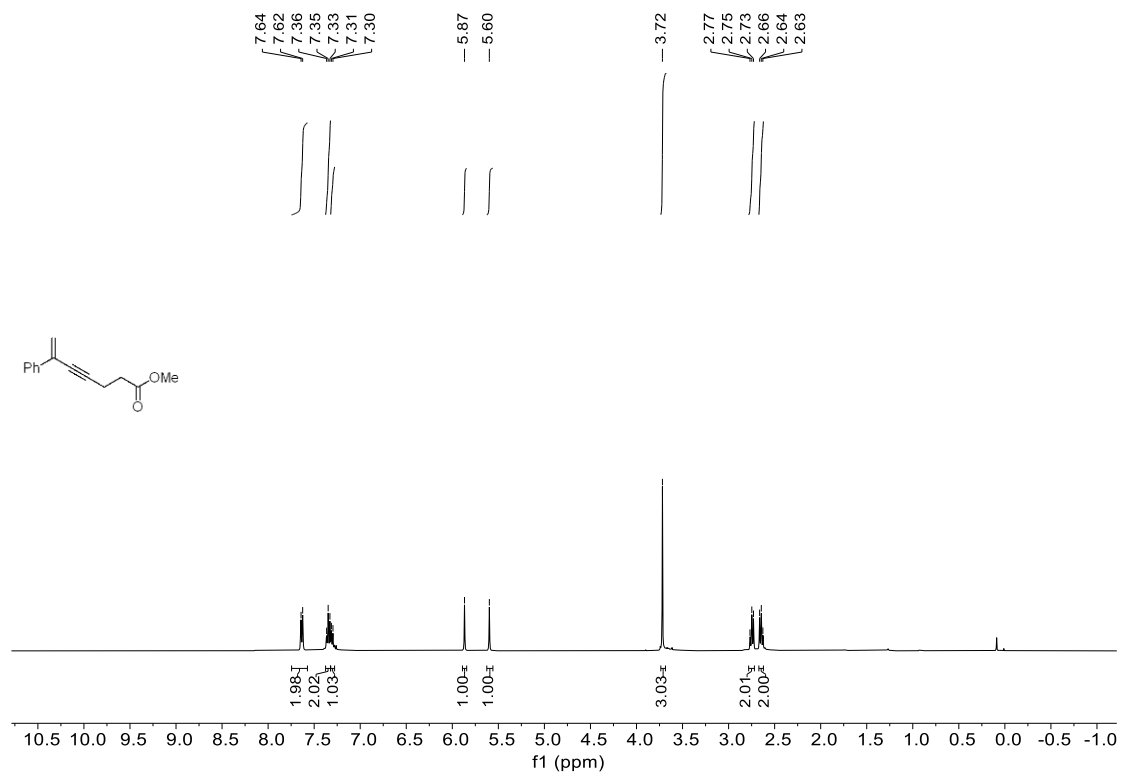
8. NMR Spectra



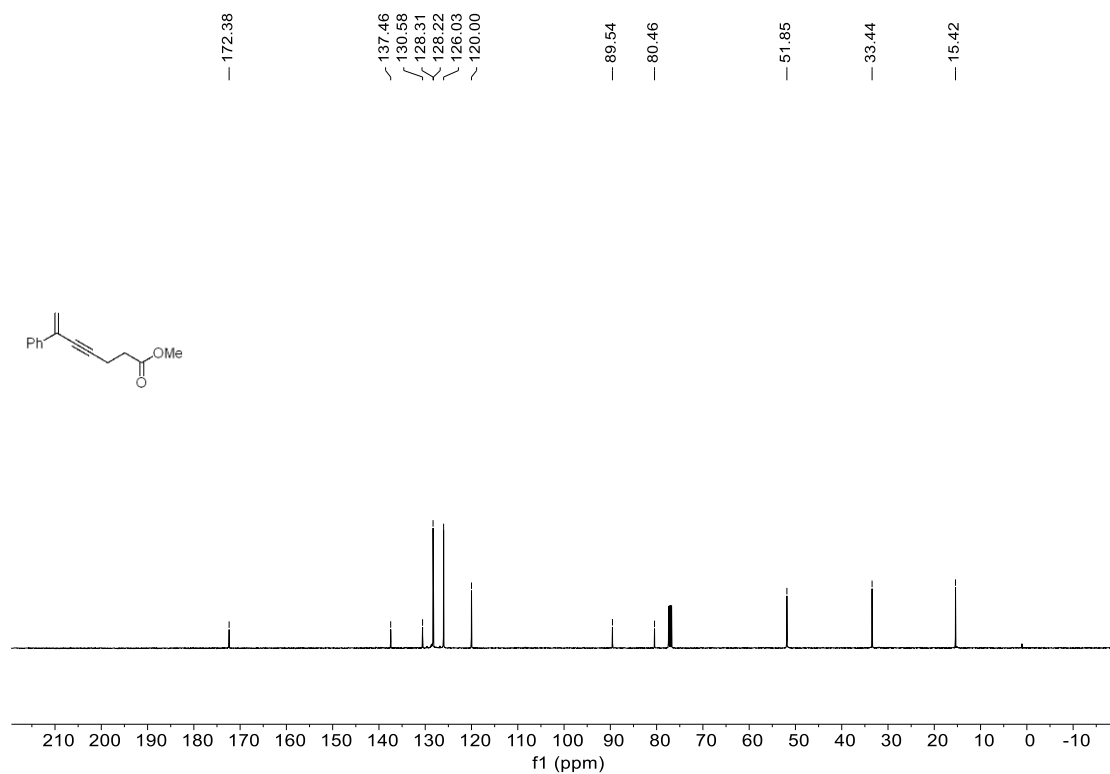
¹H NMR (400 MHz, CDCl₃) spectra of S2



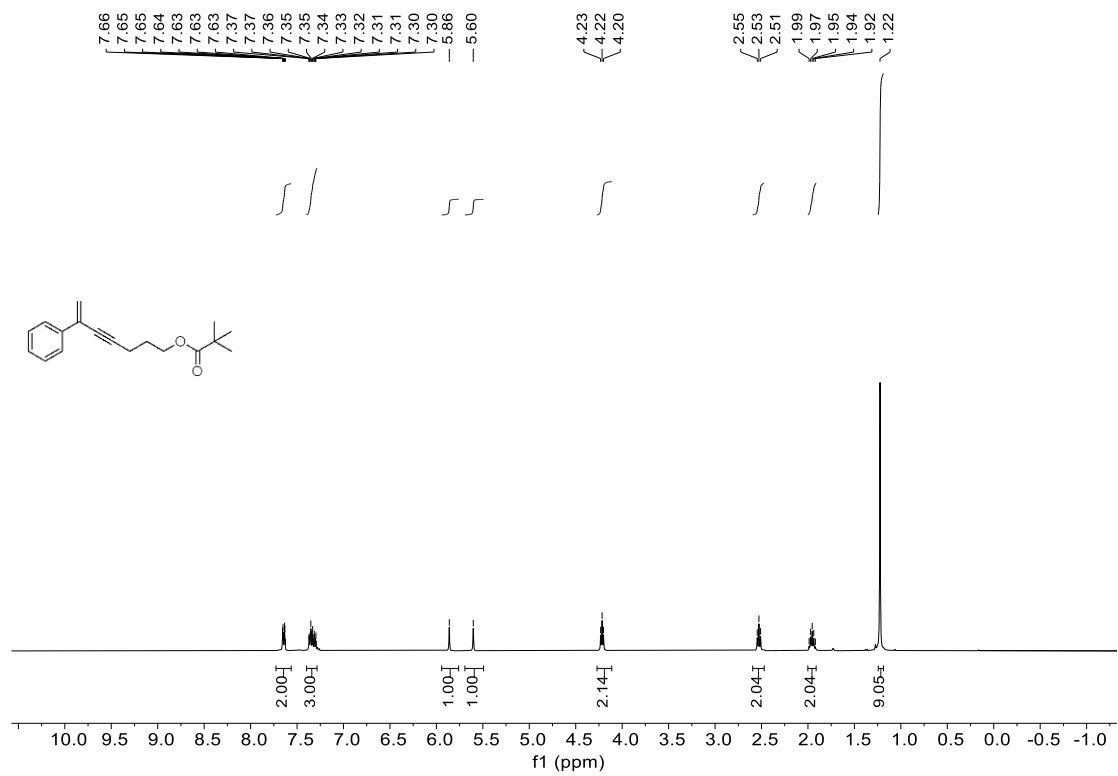
¹³C NMR (101 MHz, CDCl₃) spectra of S2



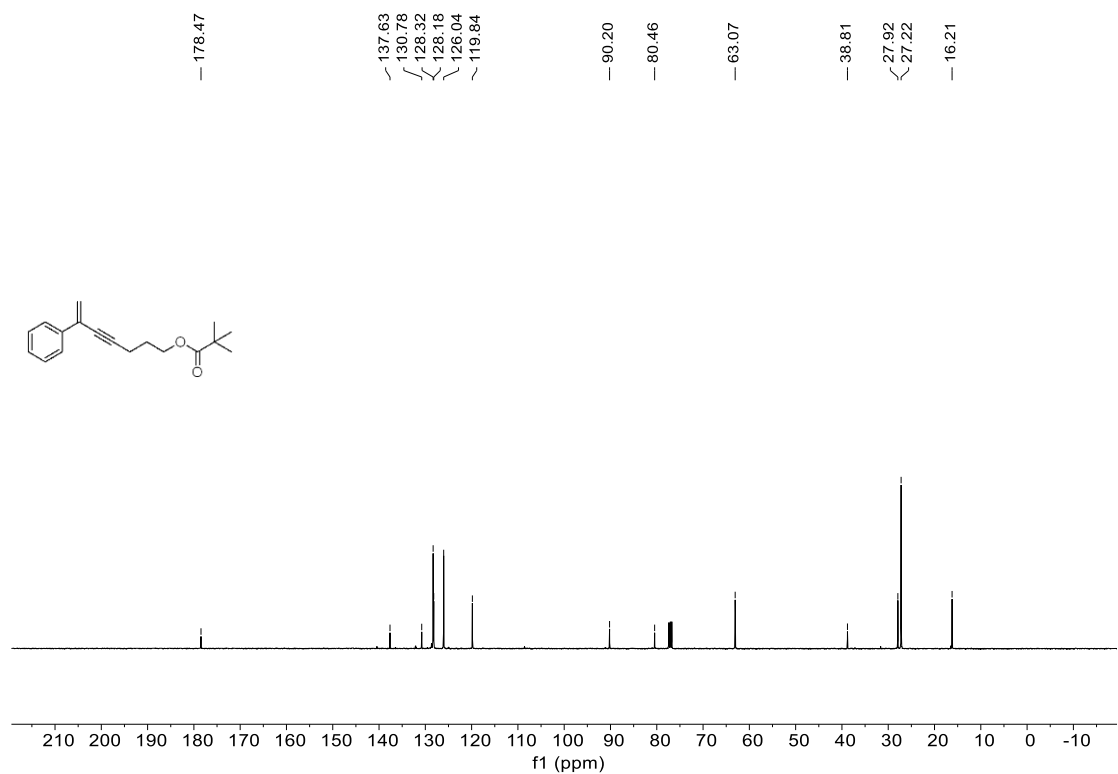
$^1\text{H NMR}$ (400 MHz, CDCl_3) spectra of S3



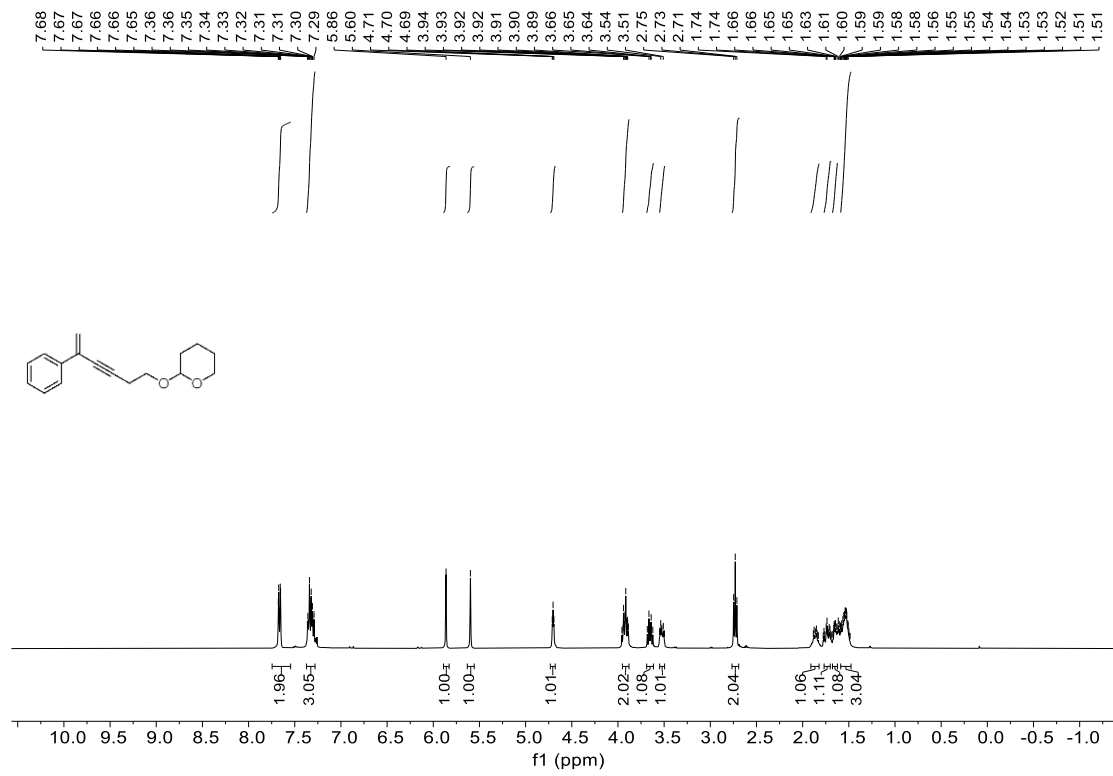
$^{13}\text{C NMR}$ (101 MHz, CDCl_3) spectra of S3



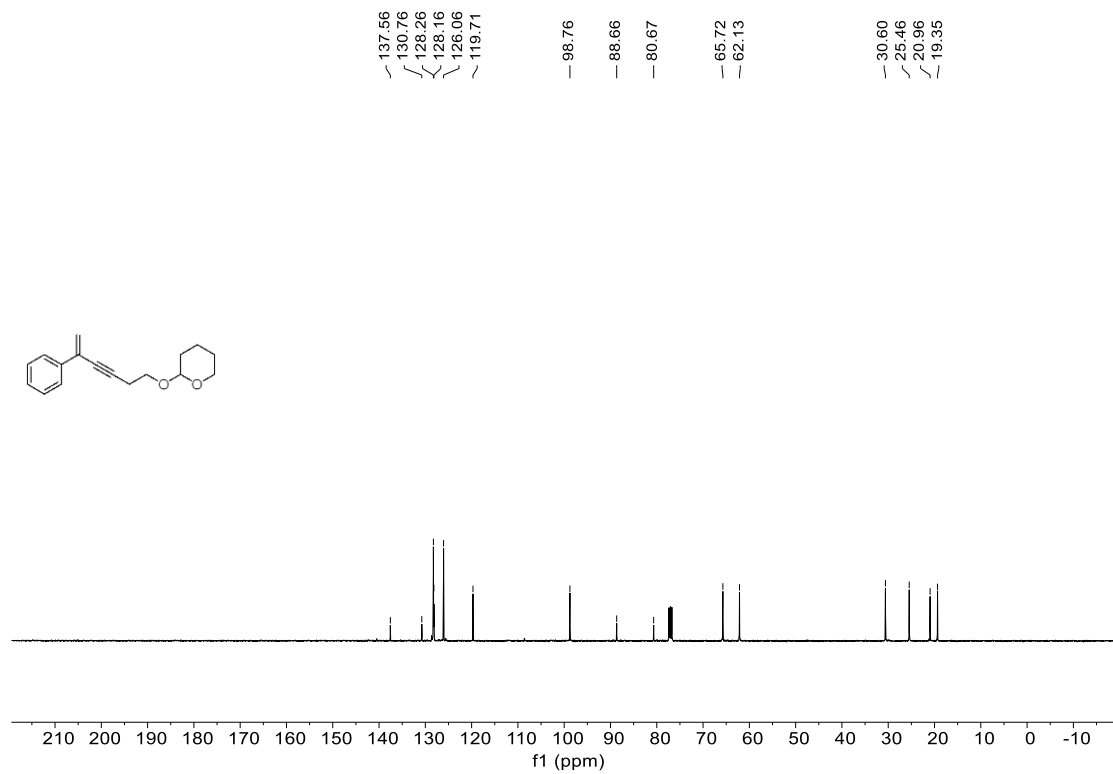
¹H NMR (400 MHz, CDCl₃) spectra of S4



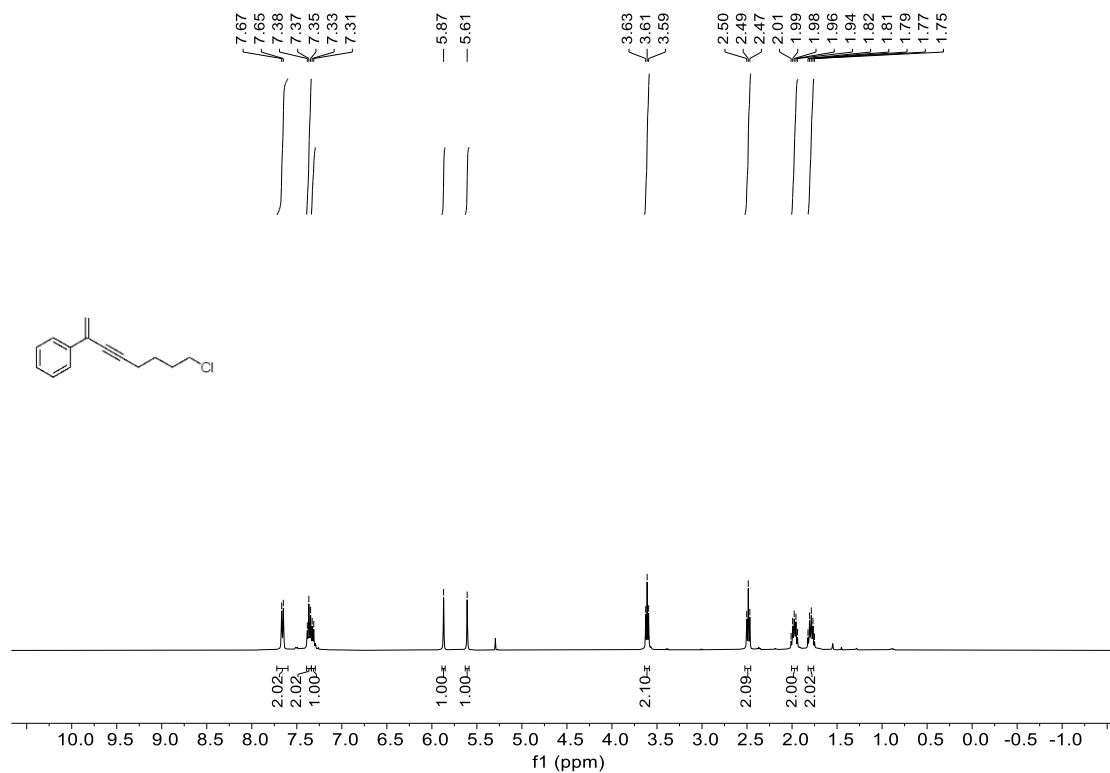
¹³C NMR (101 MHz, CDCl₃) spectra of S4



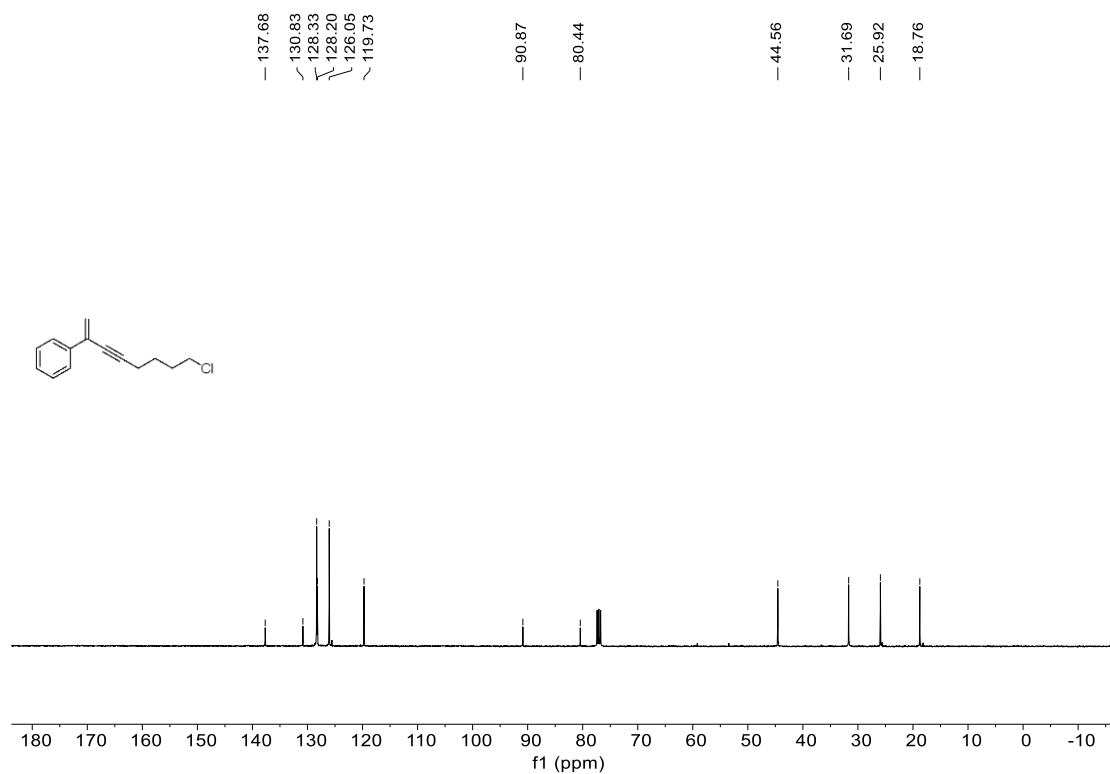
¹H NMR (400 MHz, CDCl₃) spectra of S5



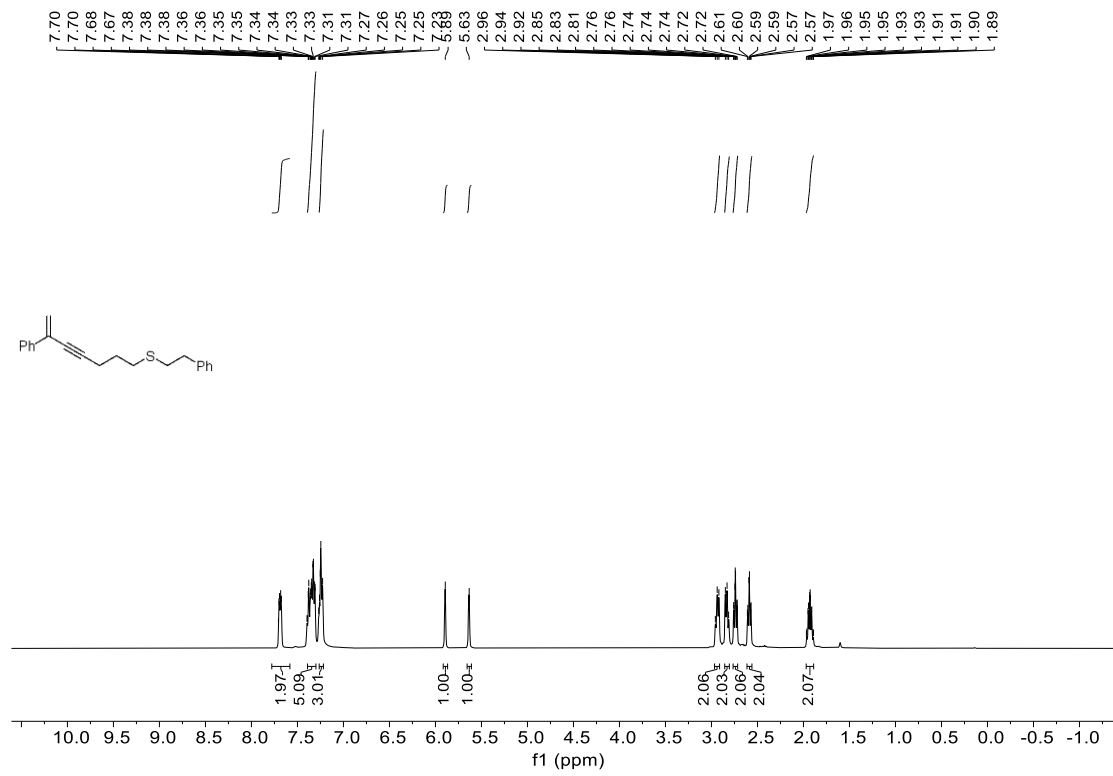
¹³C NMR (101 MHz, CDCl₃) spectra of S5



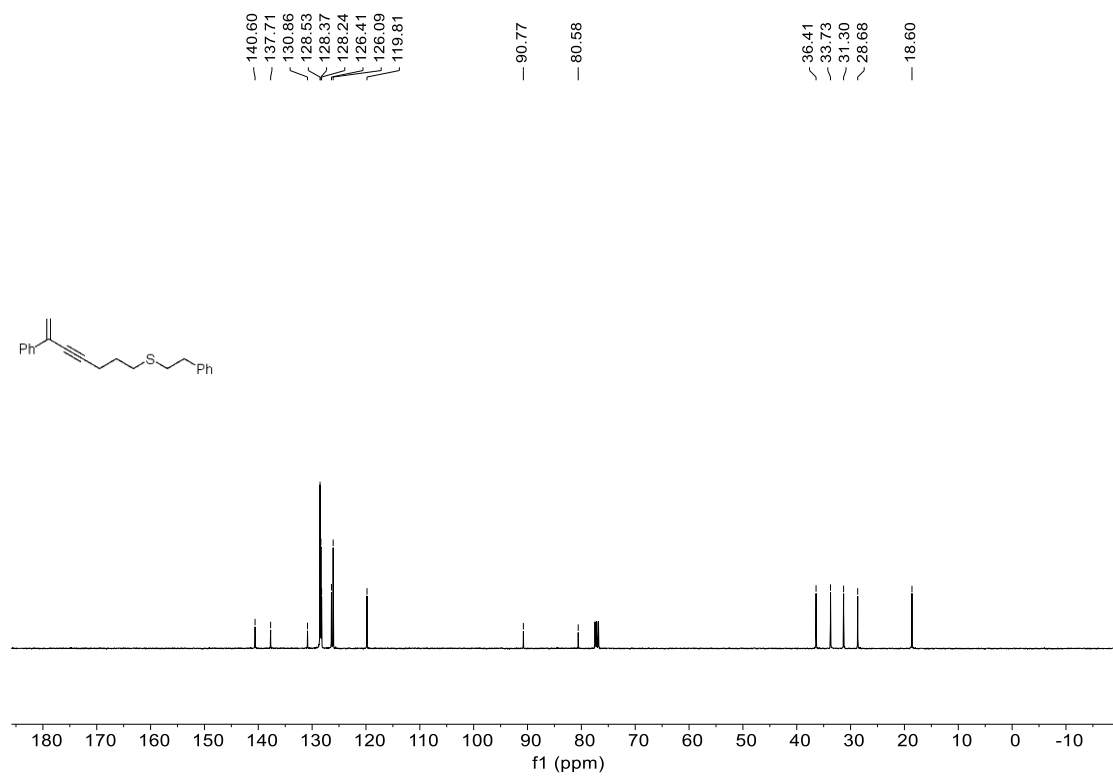
¹H NMR (400 MHz, CDCl₃) spectra of S8



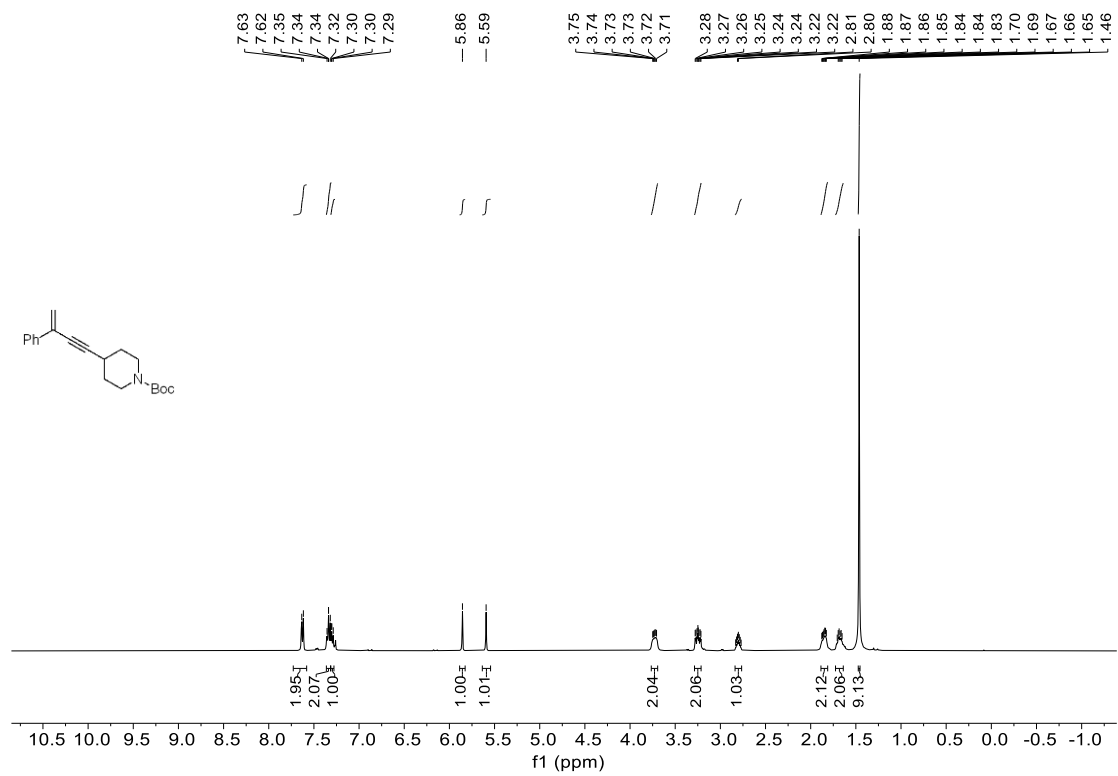
¹³C NMR (101 MHz, CDCl₃) spectra of S8



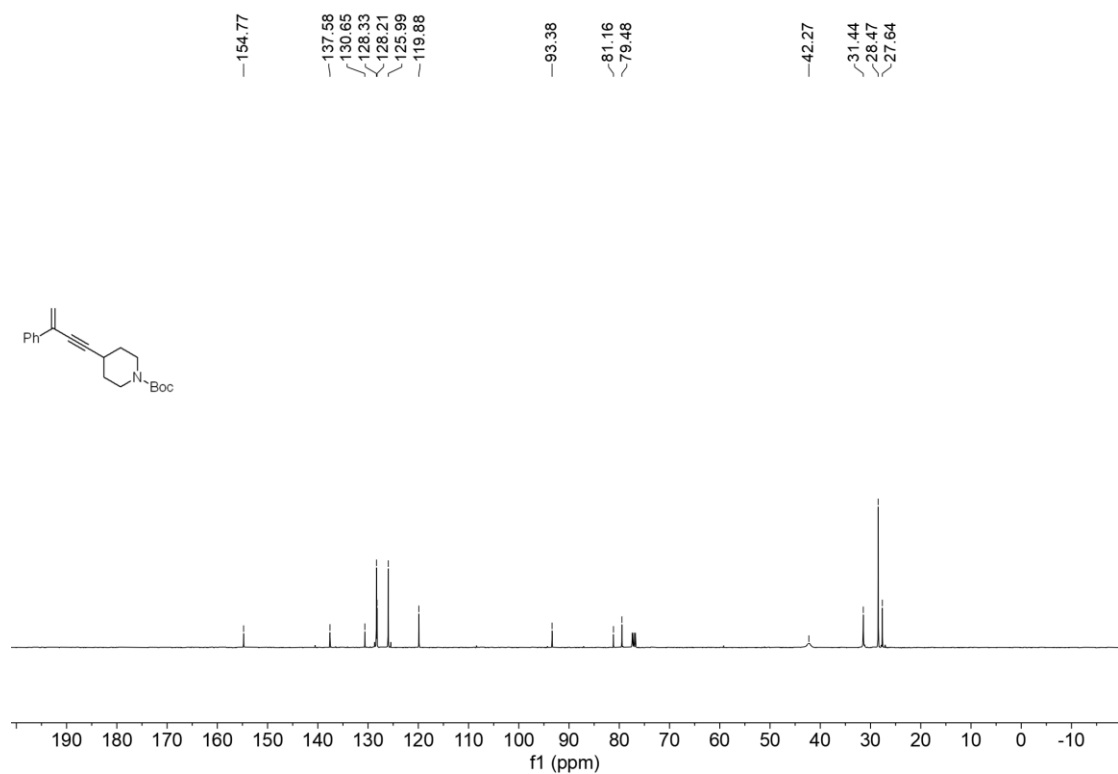
¹H NMR (400 MHz, CDCl₃) spectra of S11



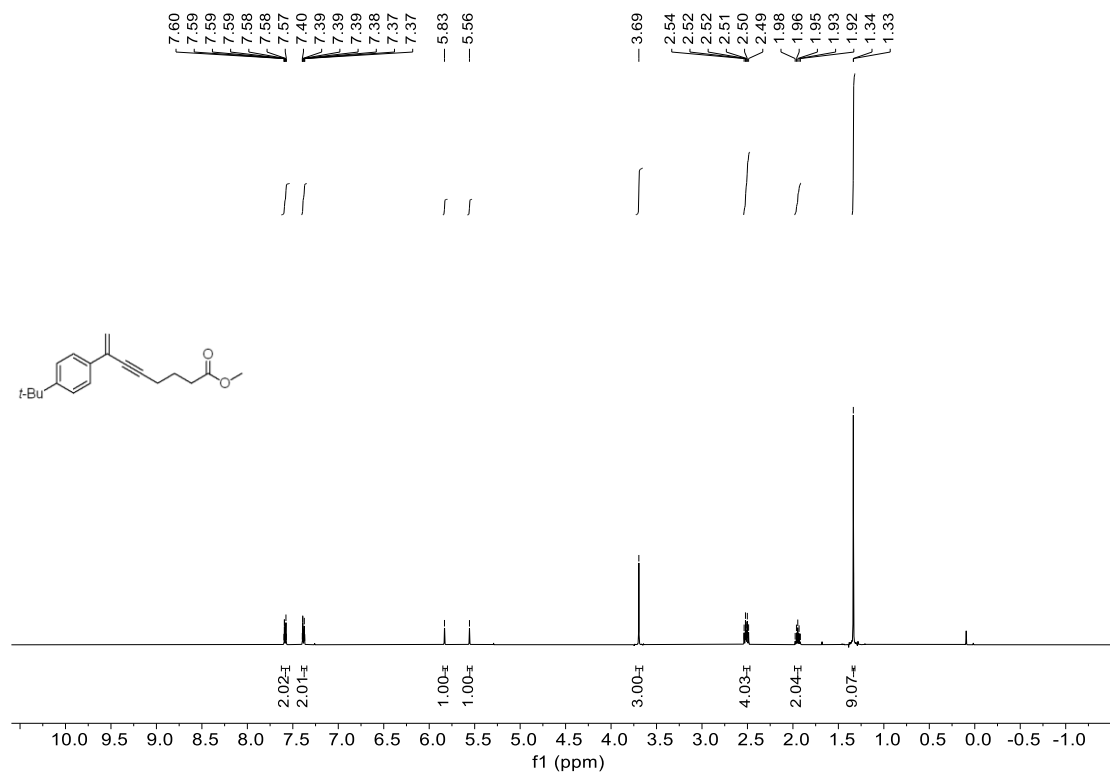
¹³C NMR (101 MHz, CDCl₃) spectra of S11



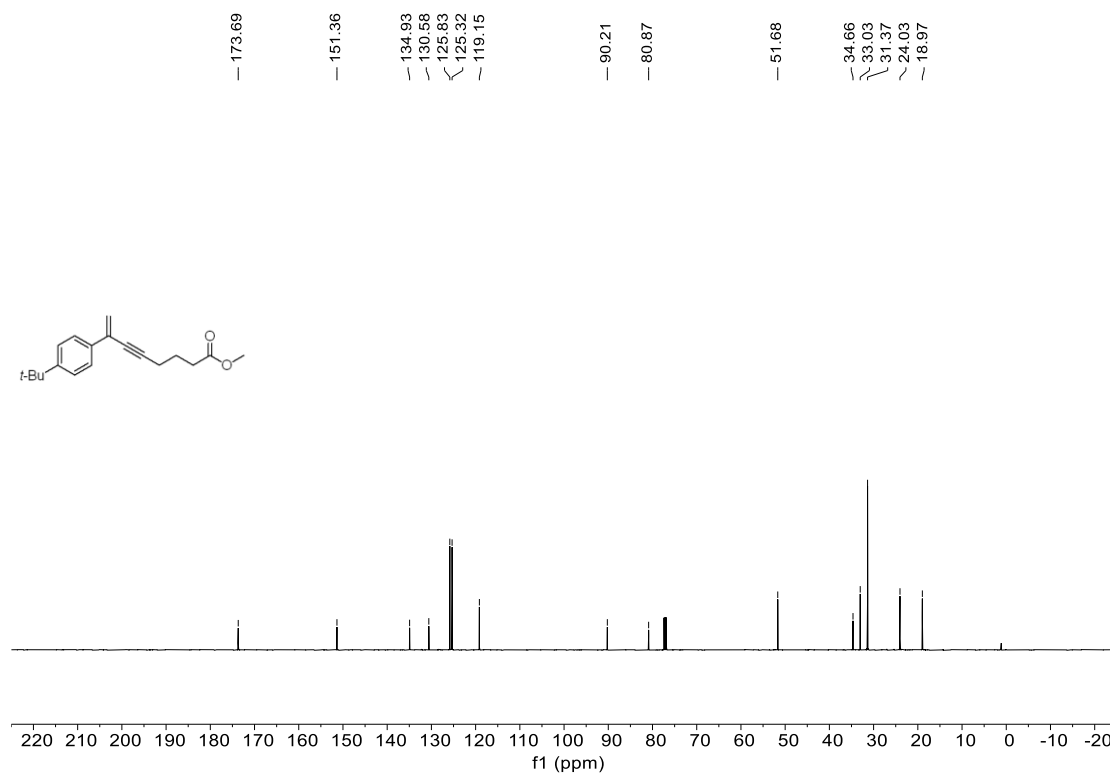
¹H NMR (400 MHz, CDCl₃) spectra of S15



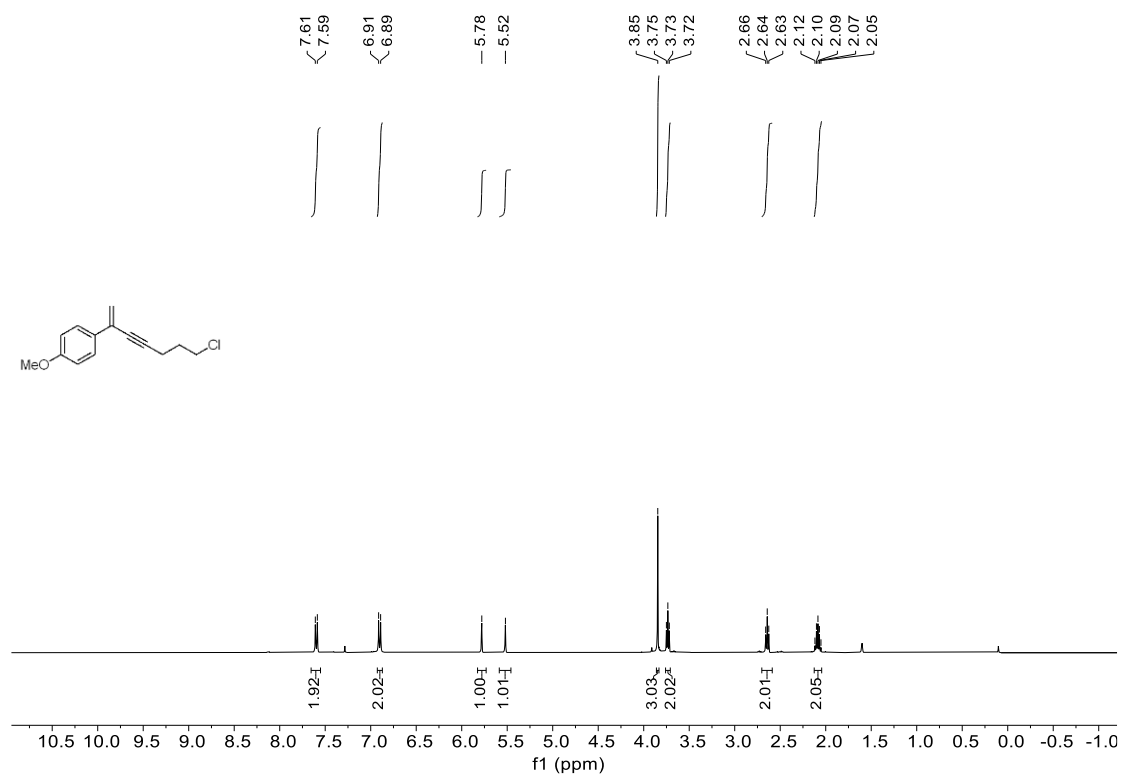
¹³C NMR (101 MHz, CDCl₃) spectra of S15



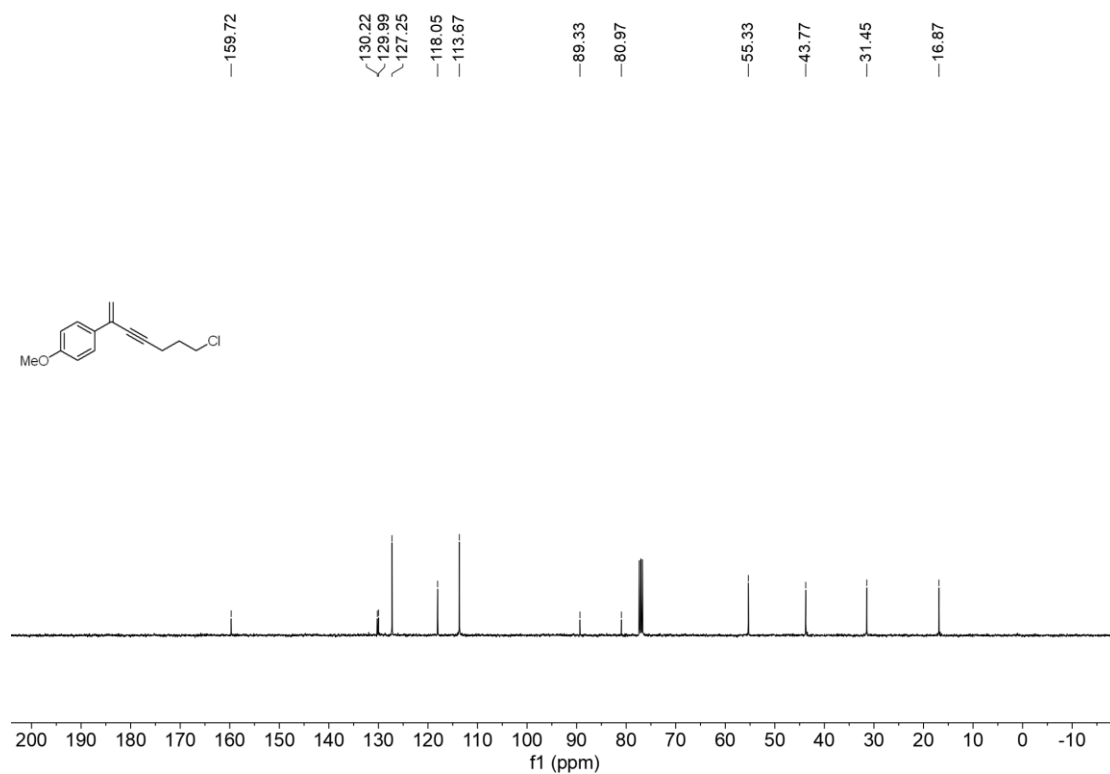
¹H NMR (500 MHz, CDCl₃) spectra of S17



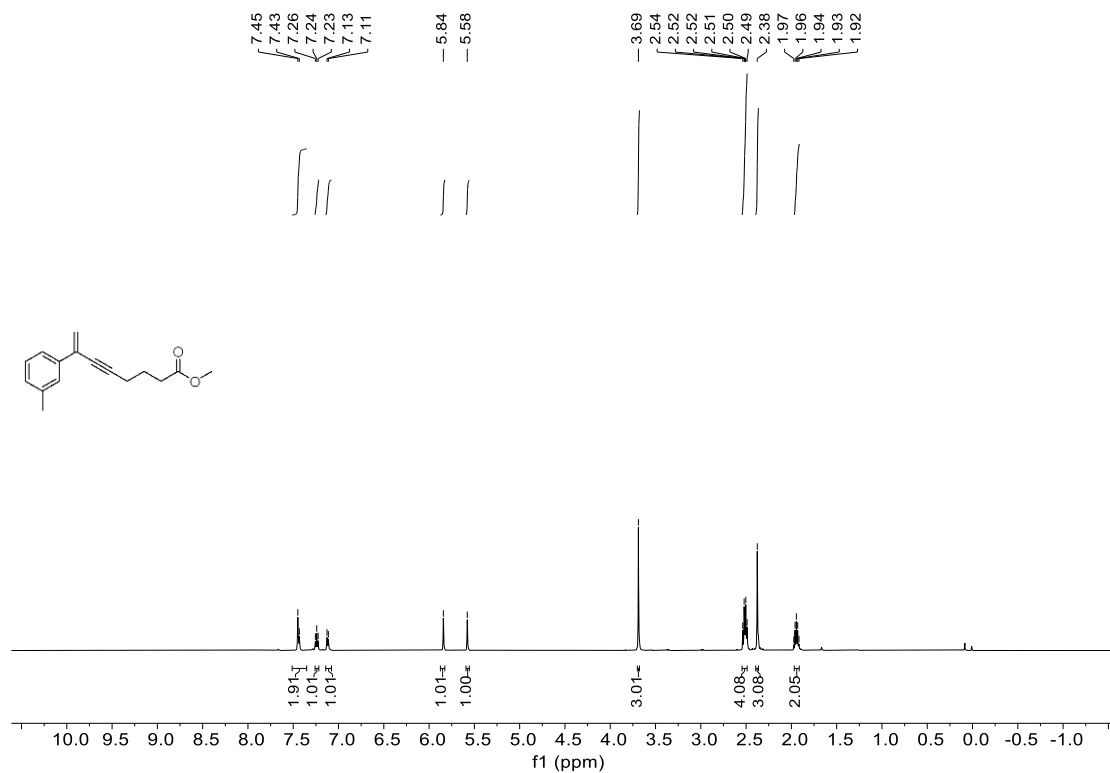
¹³C NMR (126 MHz, CDCl₃) spectra of S17



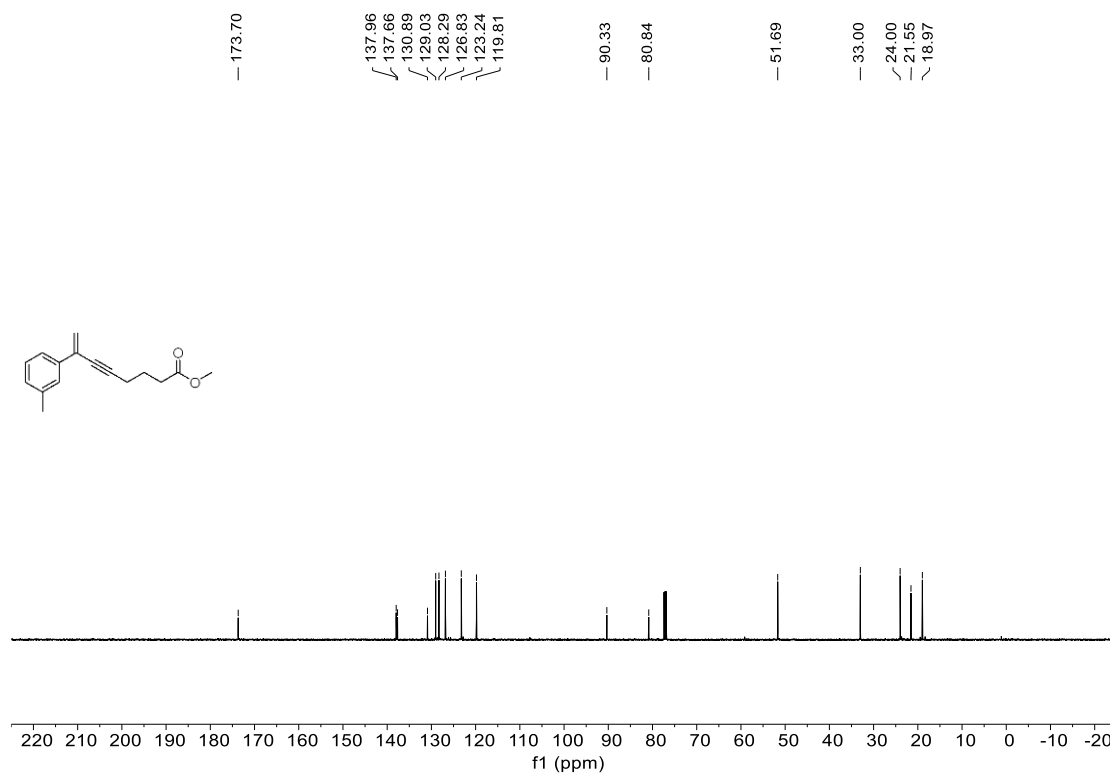
¹H NMR (400 MHz, CDCl₃) spectra of S18



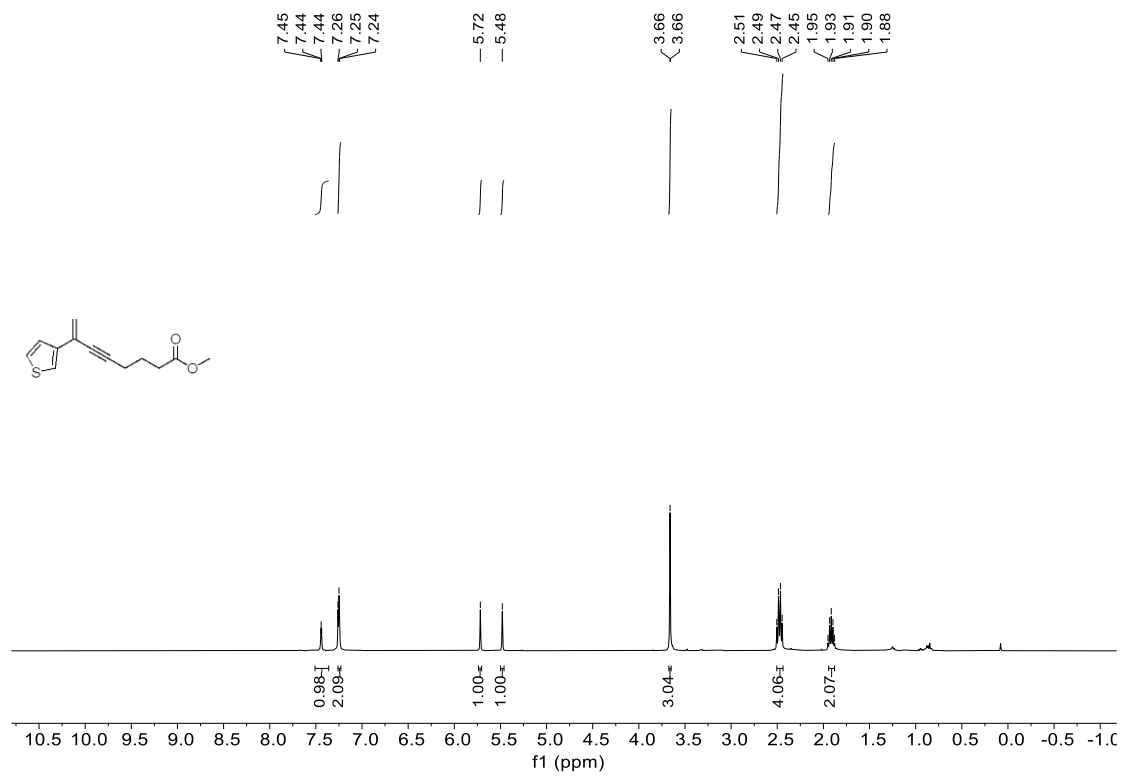
¹³C NMR (101 MHz, CDCl₃) spectra of S18



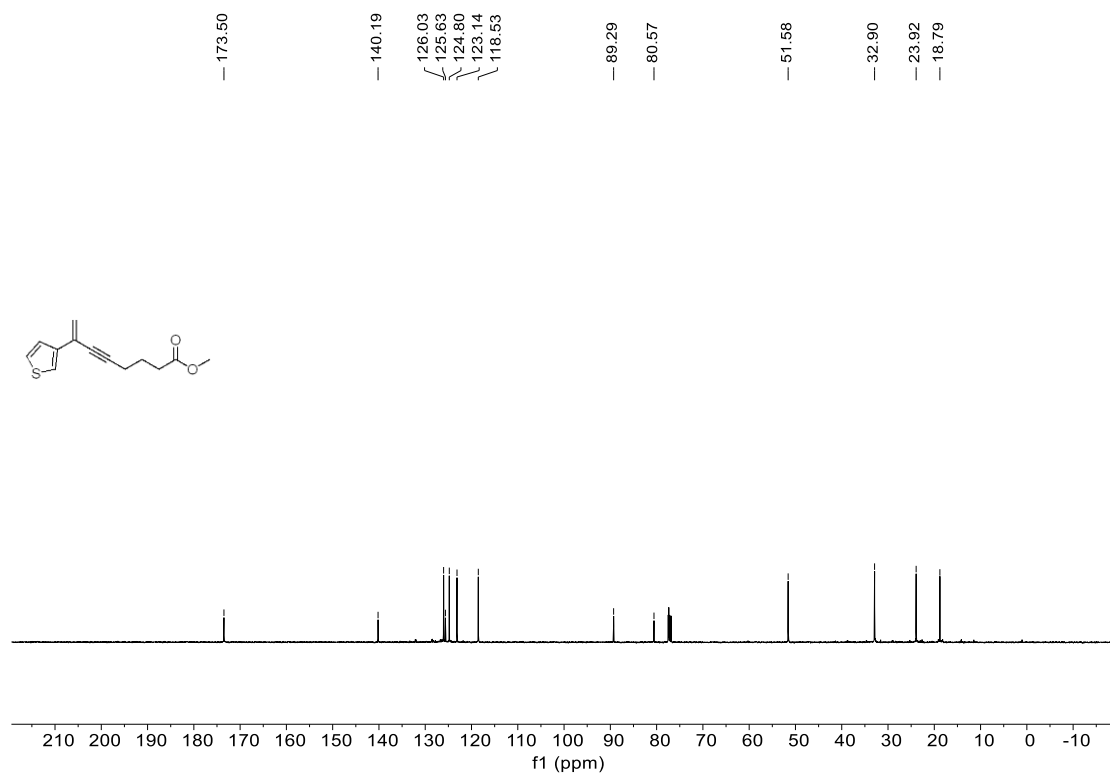
¹H NMR (400 MHz, CDCl₃) spectra of S19



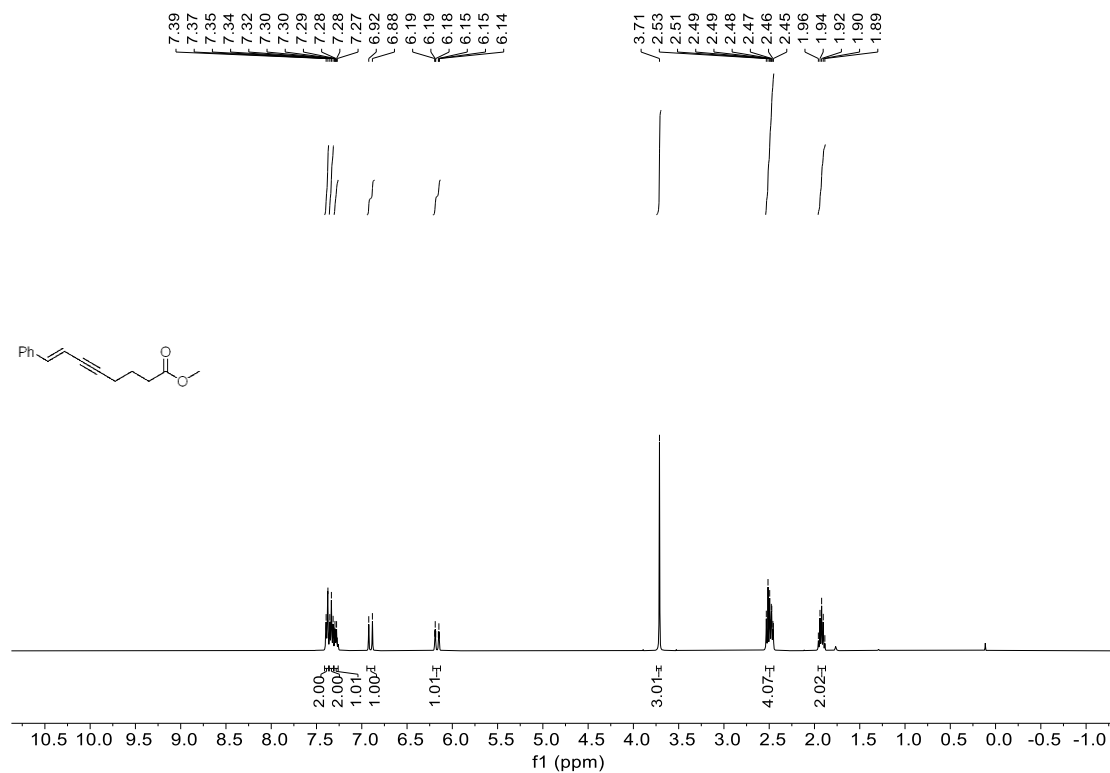
¹³C NMR (101 MHz, CDCl₃) spectra of S19



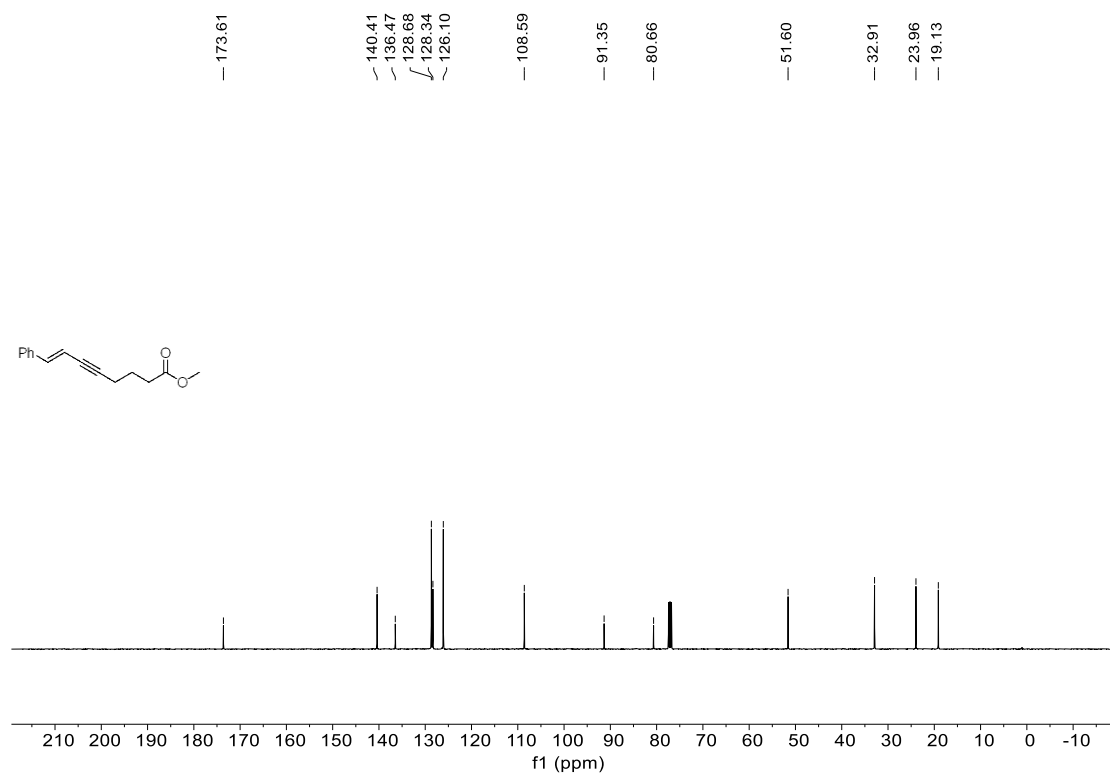
¹H NMR (400 MHz, CDCl₃) spectra of S21



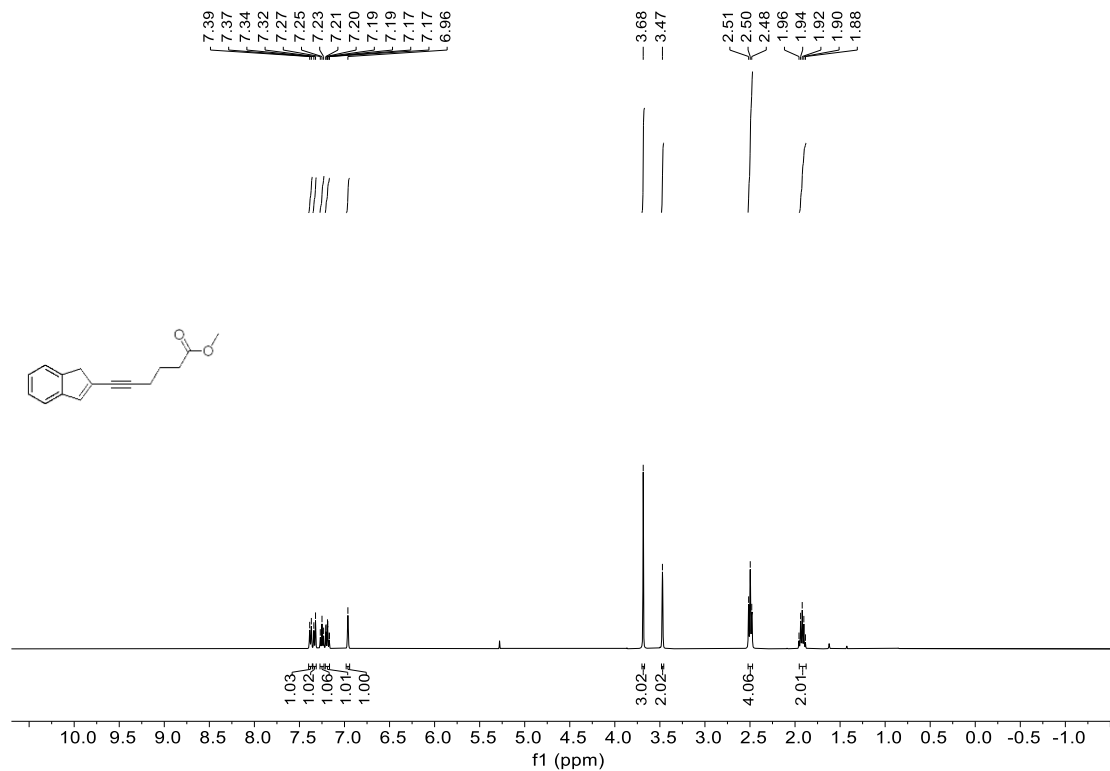
¹³C NMR (101 MHz, CDCl₃) spectra of S21



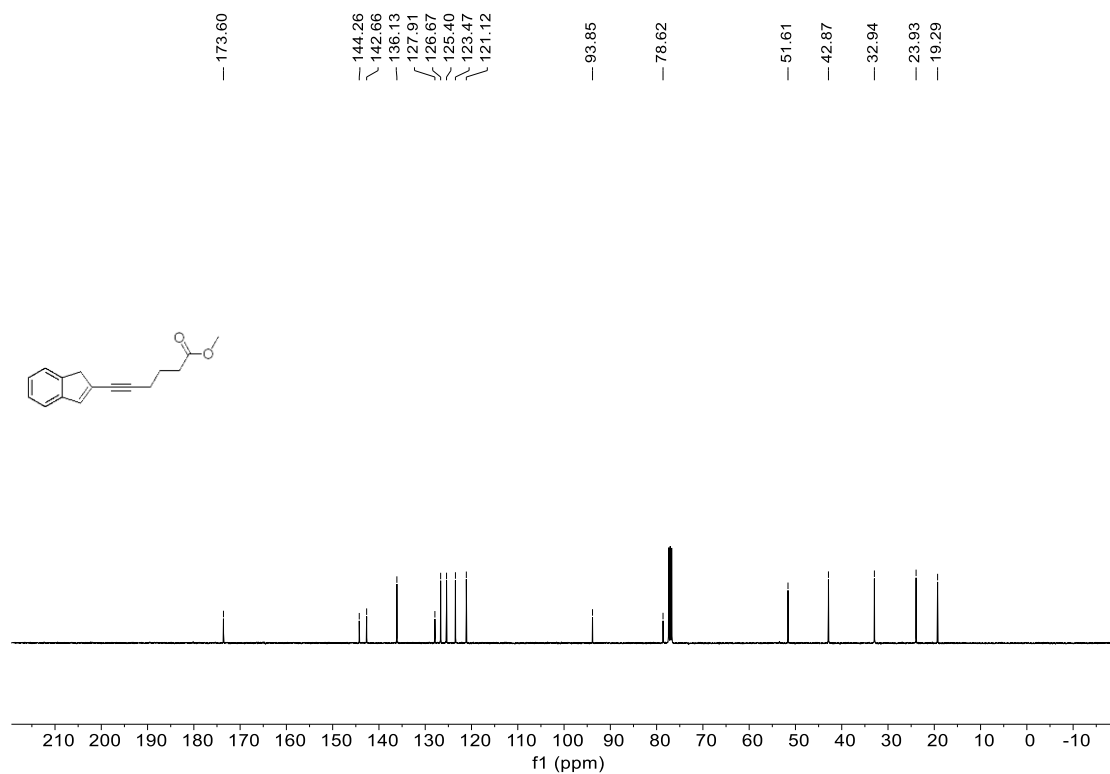
¹H NMR (400 MHz, CDCl₃) spectra of S22



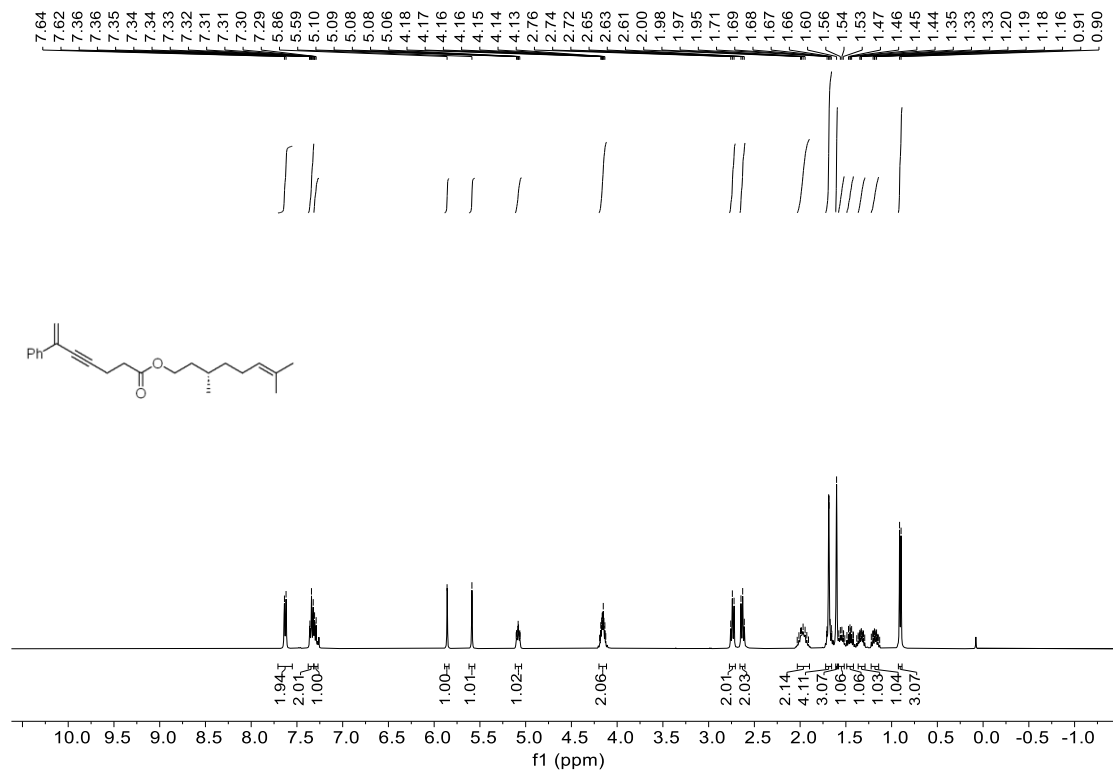
¹³C NMR (101 MHz, CDCl₃) spectra of S22



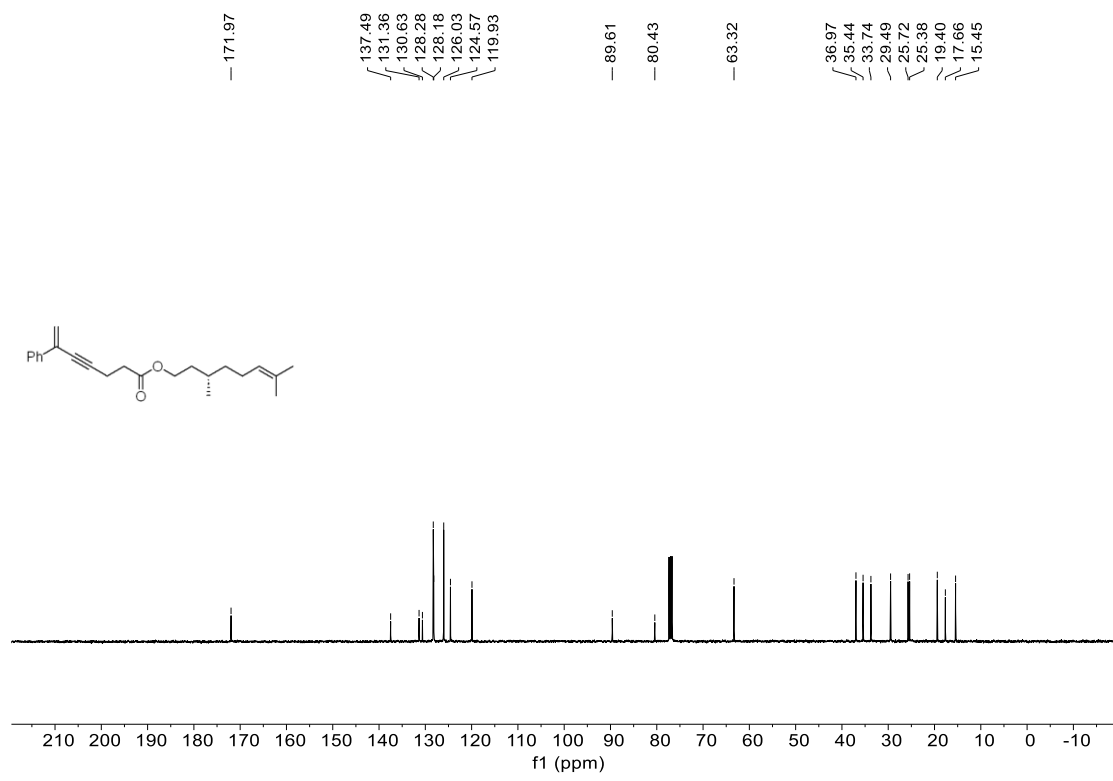
¹H NMR (400 MHz, CDCl₃) spectra of S23



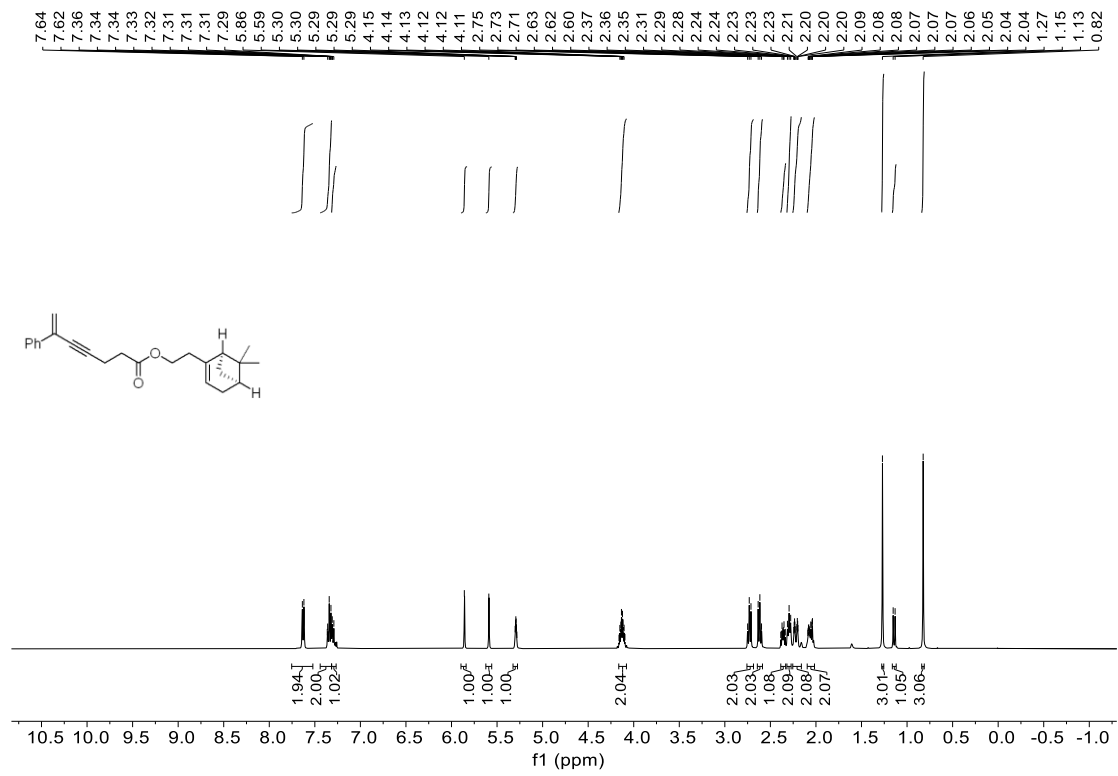
¹³C NMR (101 MHz, CDCl₃) spectra of S23



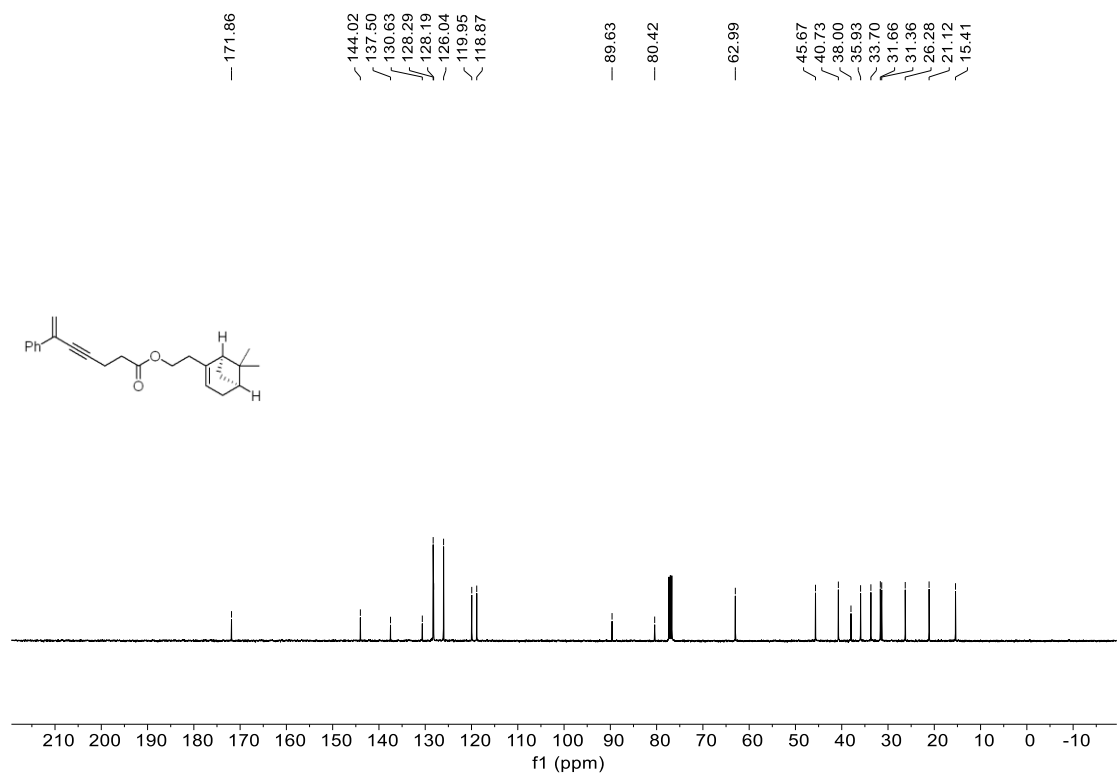
¹H NMR (400 MHz, CDCl₃) spectra of S25



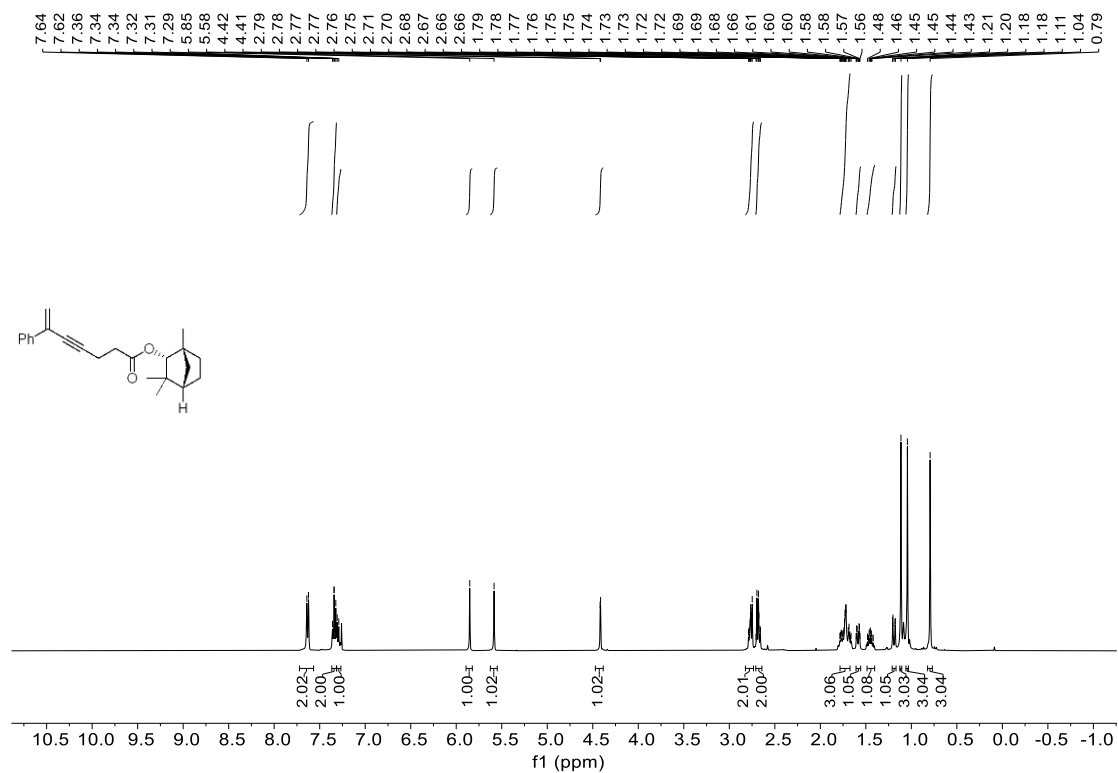
¹³C NMR (101 MHz, CDCl₃) spectra of S25



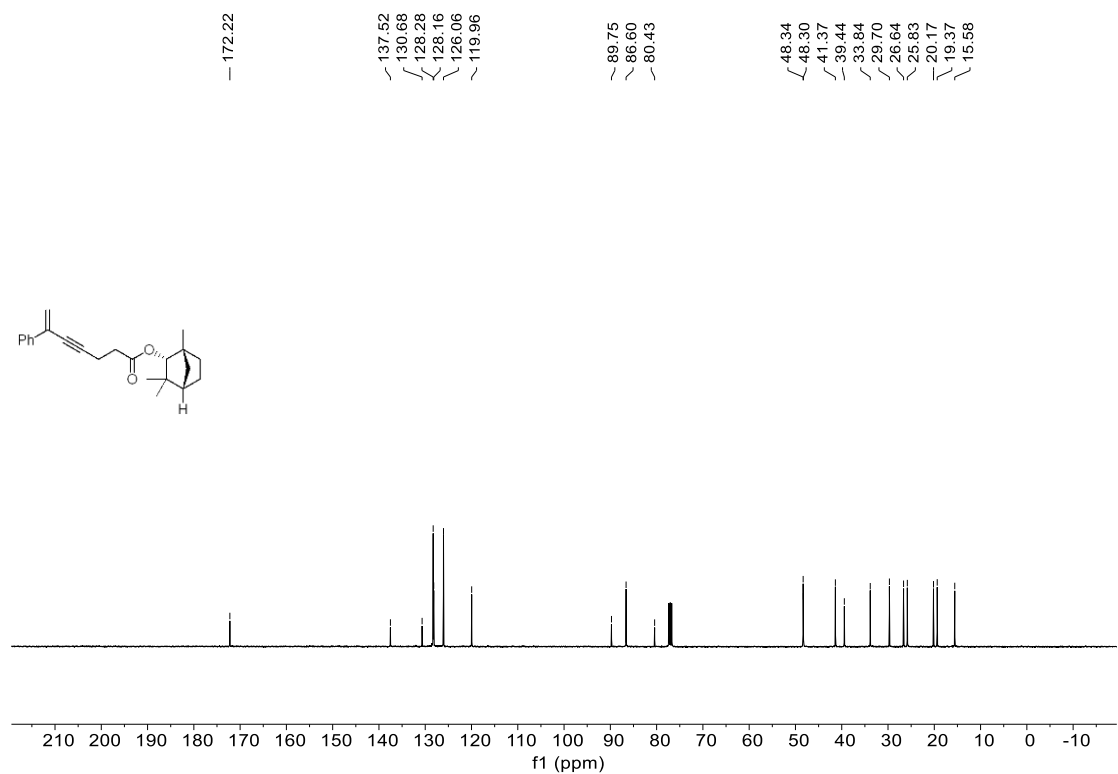
¹H NMR (400 MHz, CDCl₃) spectra of S26



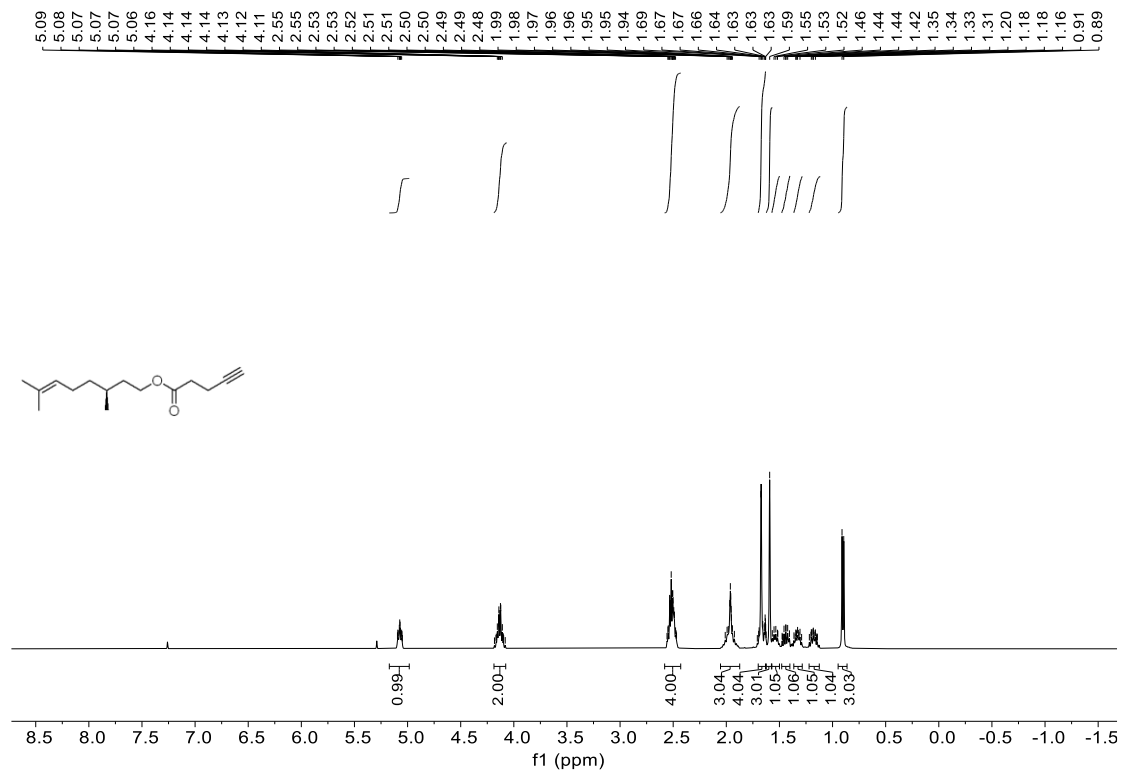
¹³C NMR (101 MHz, CDCl₃) spectra of S26



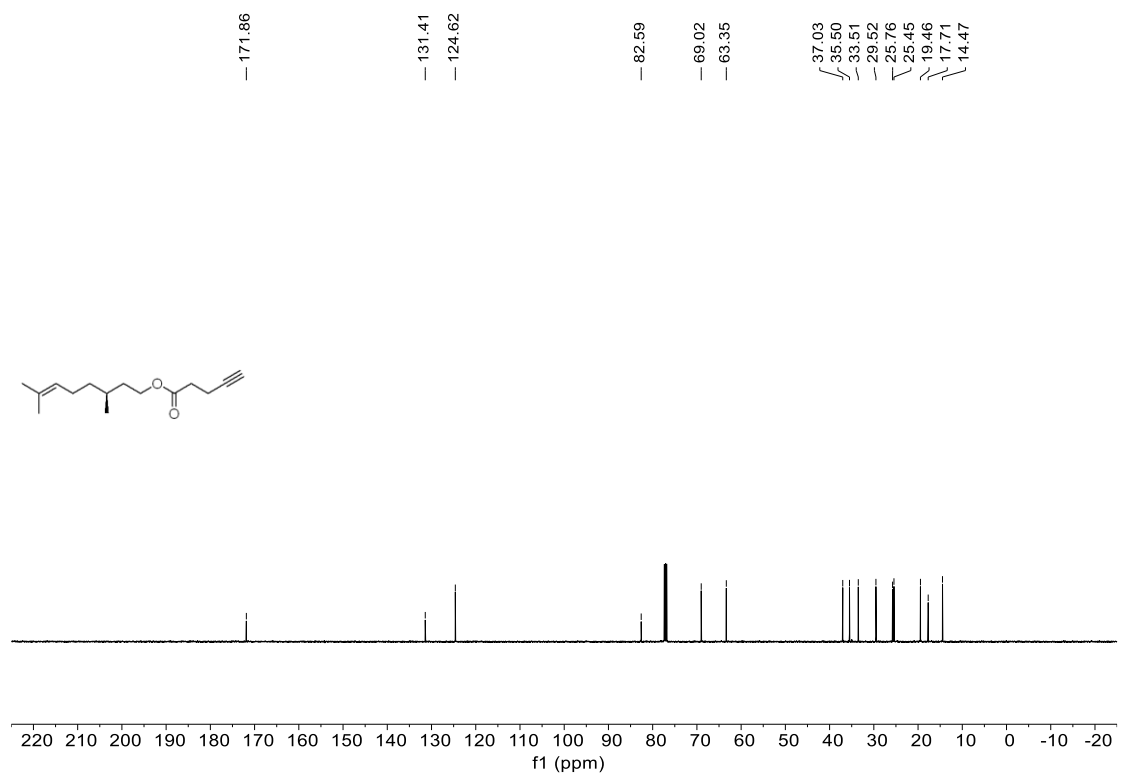
¹H NMR (400 MHz, CDCl₃) spectra of S27



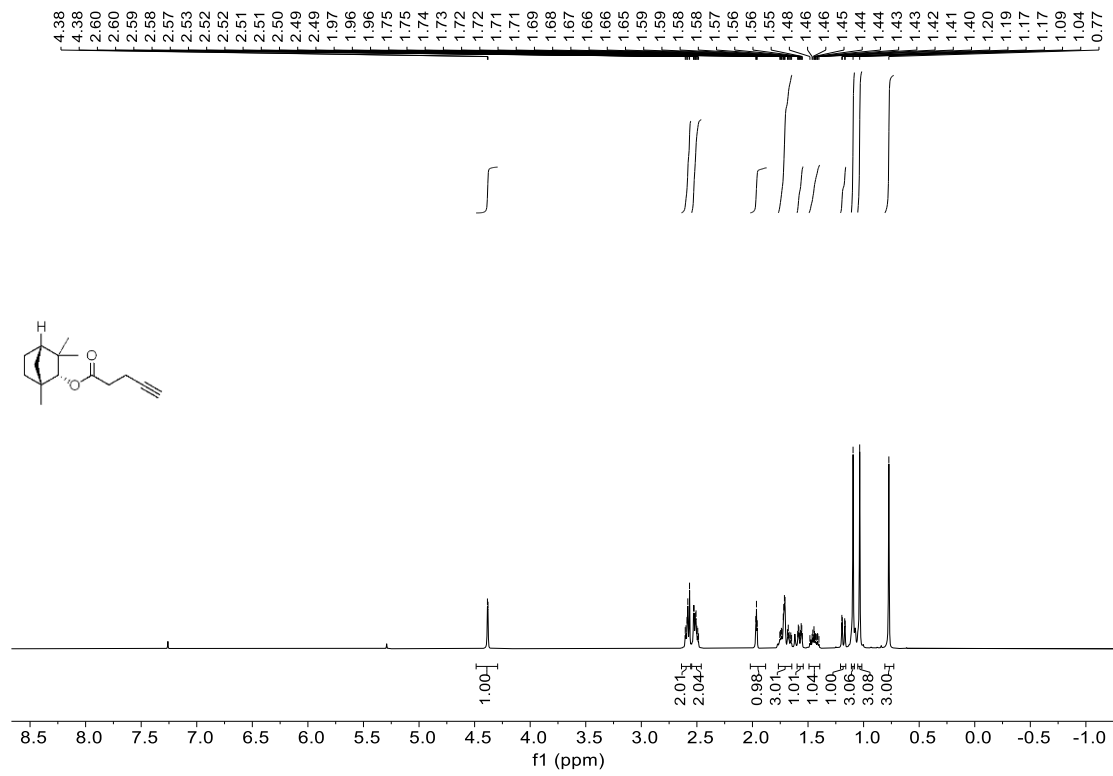
¹³C NMR (101 MHz, CDCl₃) spectra of S27



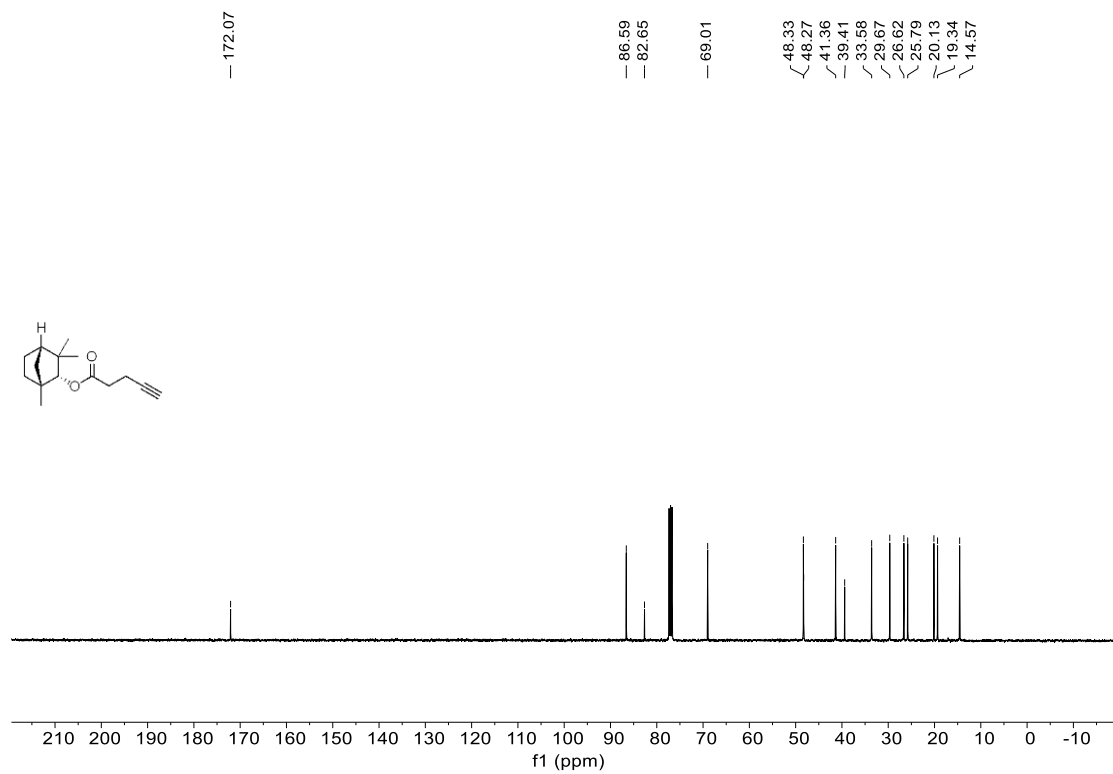
¹H NMR (400 MHz, CDCl₃) spectra of S28



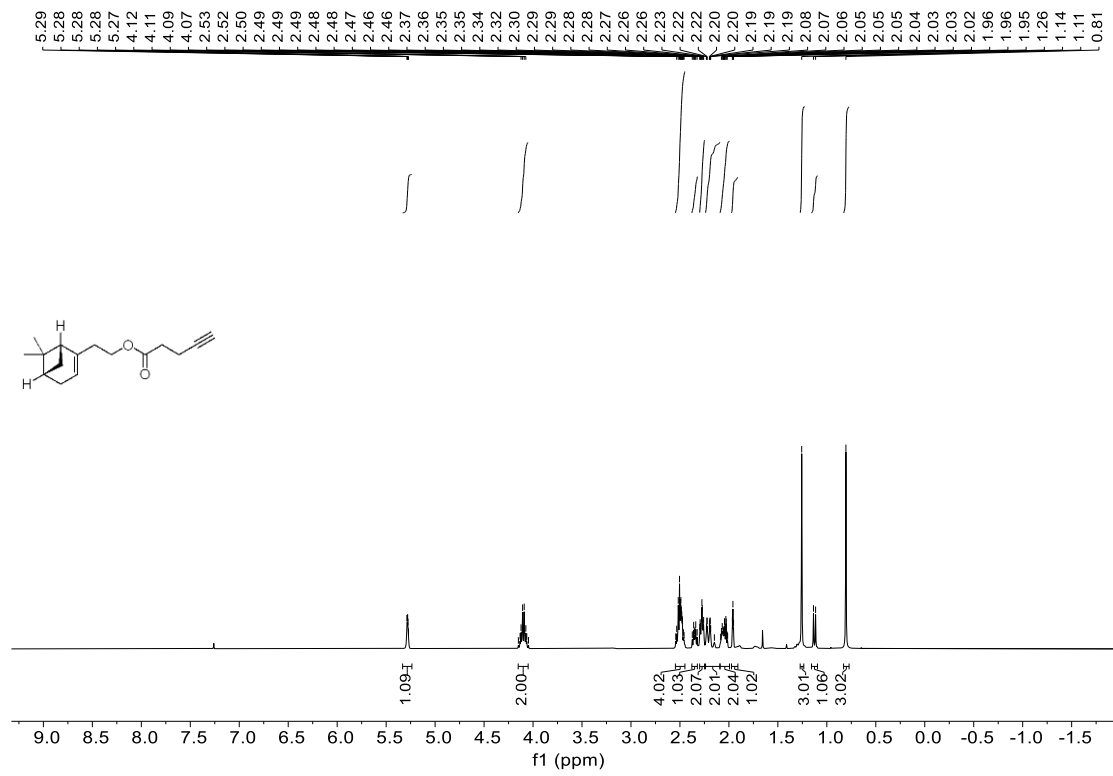
¹³C NMR (126 MHz, CDCl₃) spectra of S28



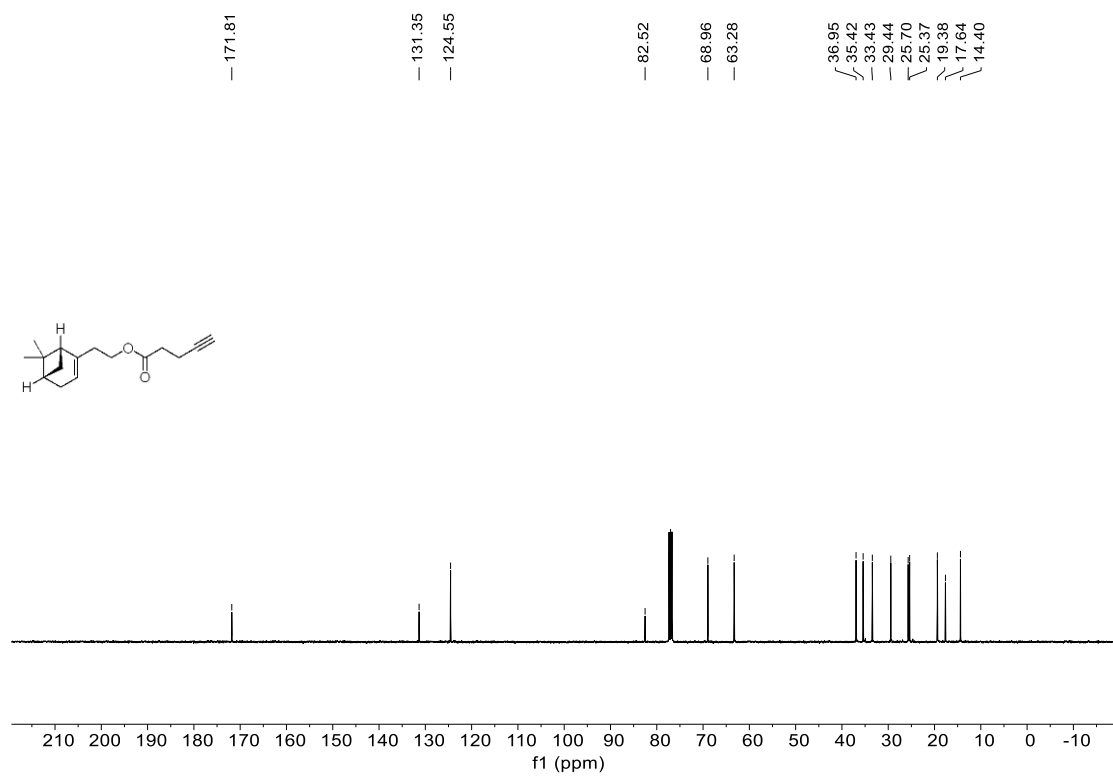
¹H NMR (400 MHz, CDCl₃) spectra of S29



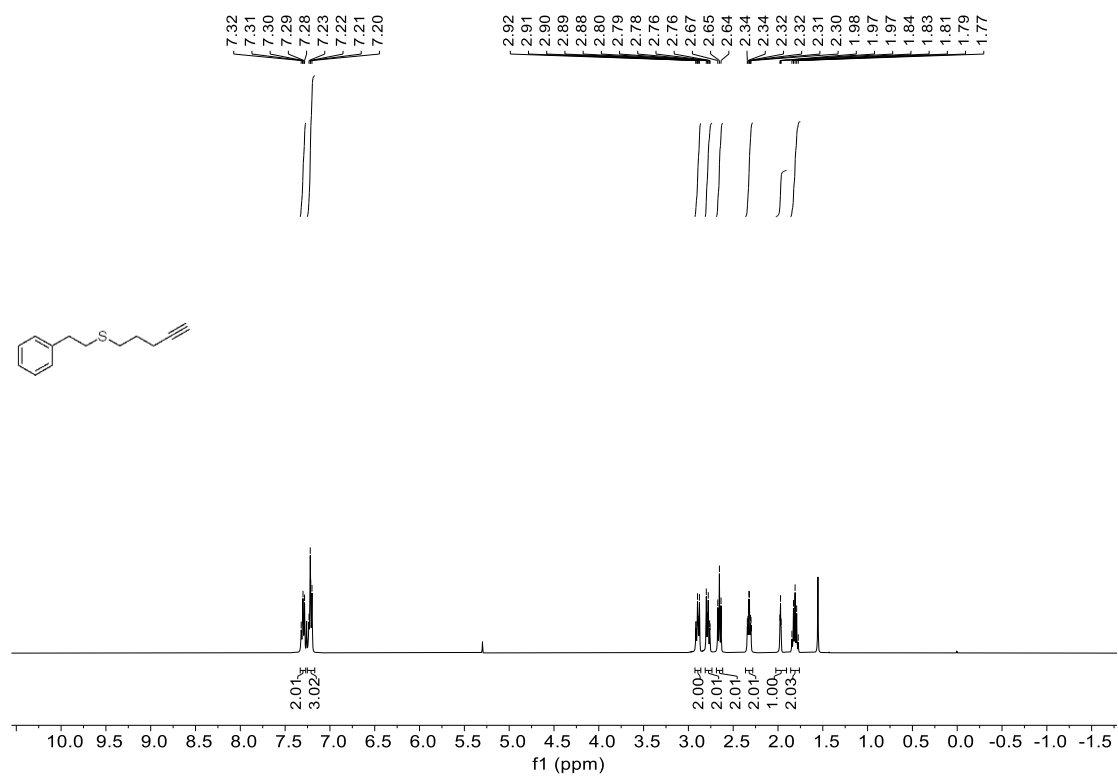
¹³C NMR (101 MHz, CDCl₃) spectra of S29



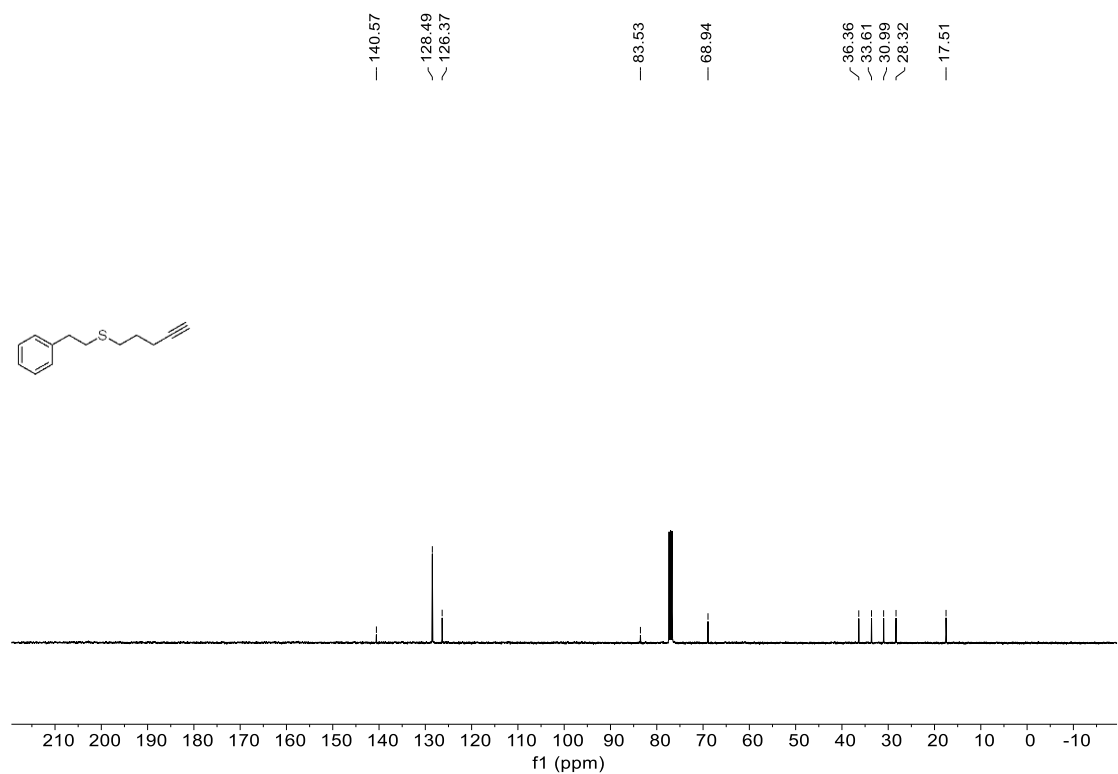
¹H NMR (400 MHz, CDCl₃) spectra of S30



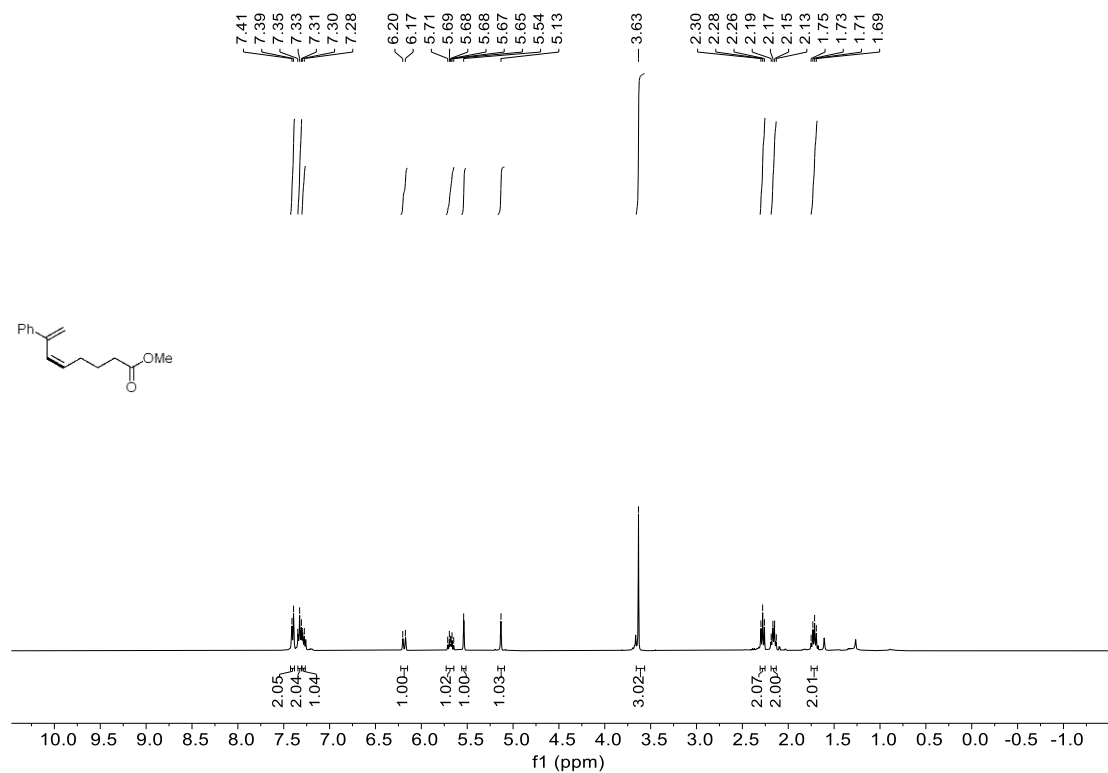
¹³C NMR (101 MHz, CDCl₃) spectra of S30



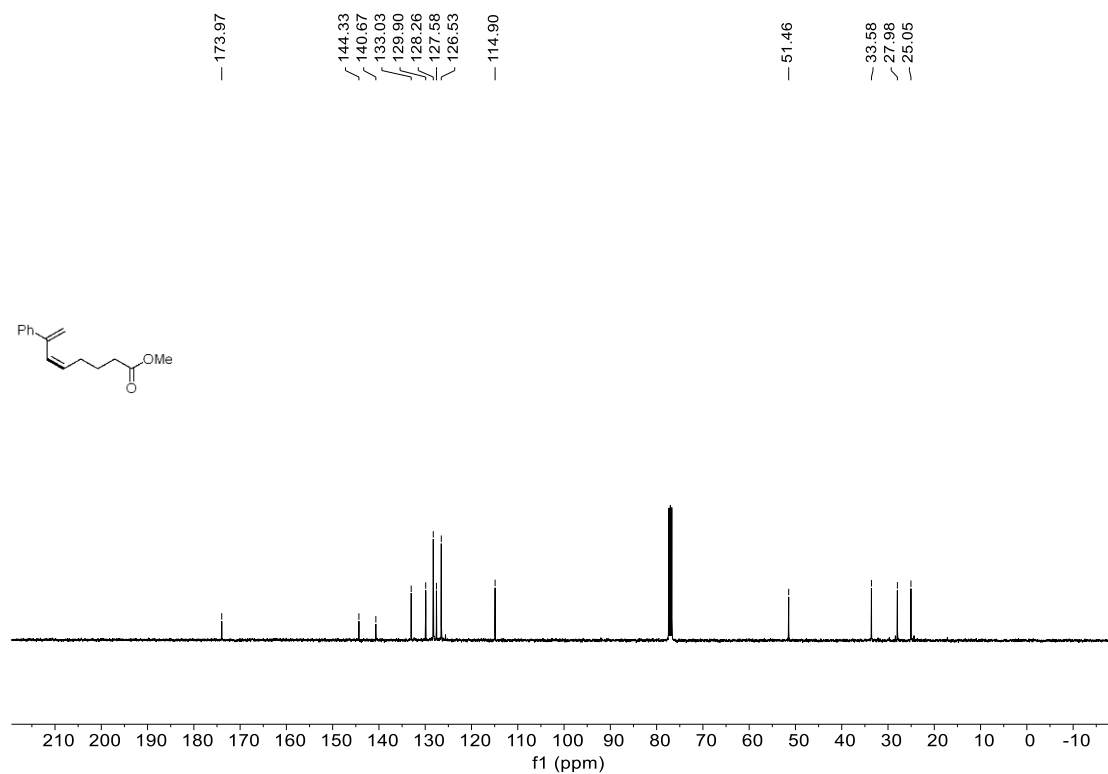
¹H NMR (400 MHz, CDCl₃) spectra of S31



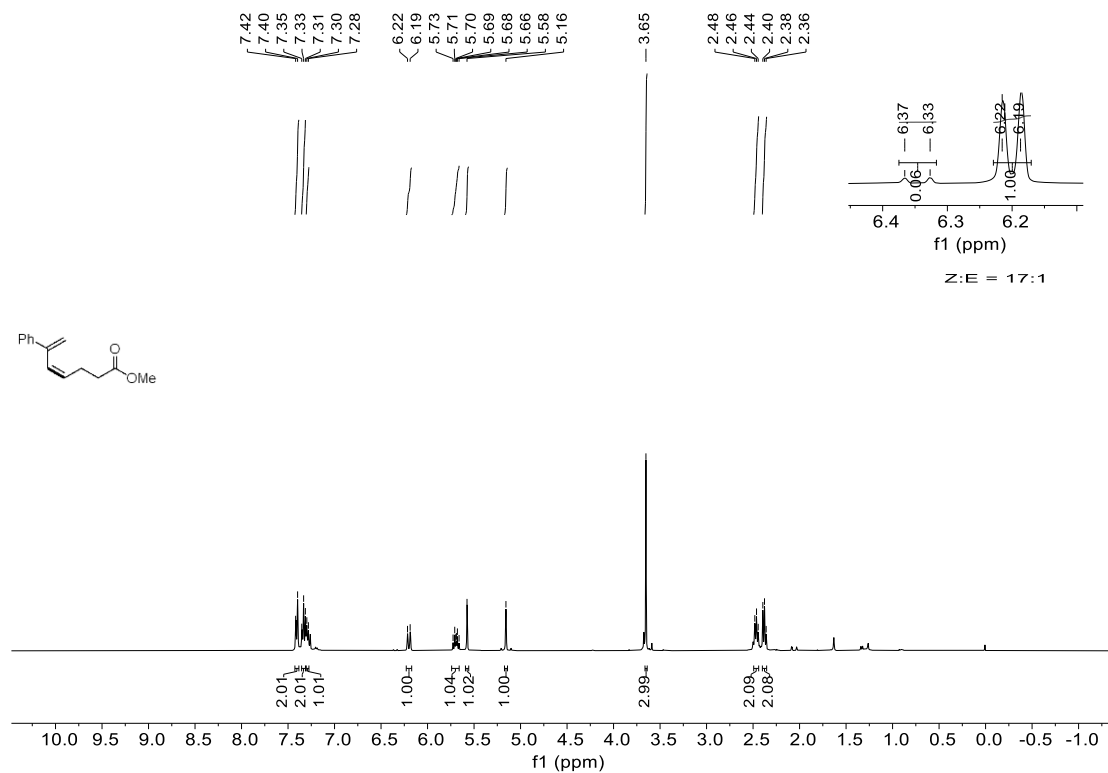
¹³C NMR (101 MHz, CDCl₃) spectra of S31



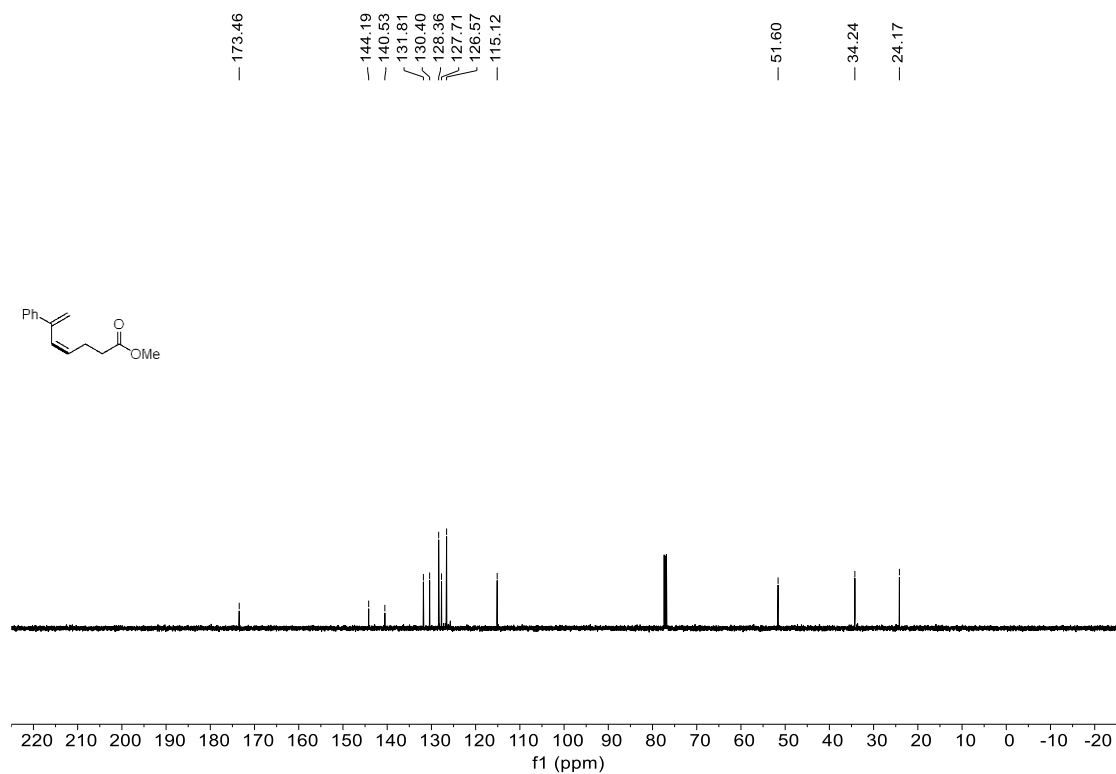
¹H NMR (400 MHz, CDCl₃) spectra of **2a**



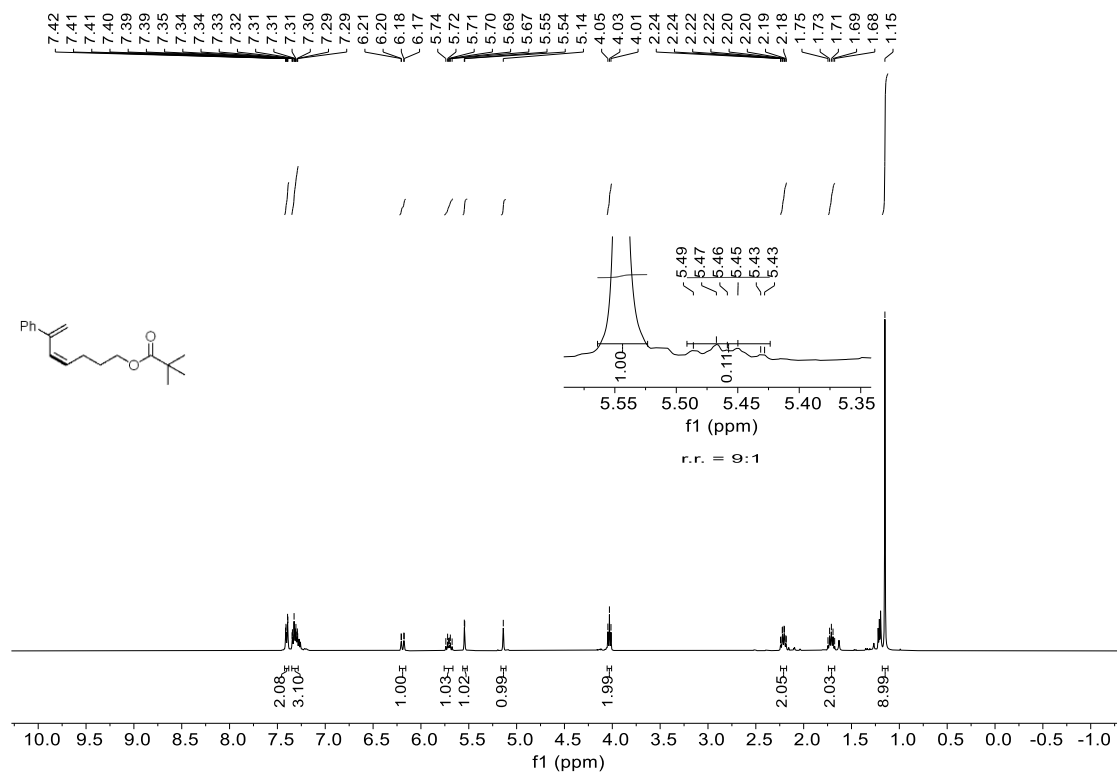
¹³C NMR (101 MHz, CDCl₃) spectra of **2a**



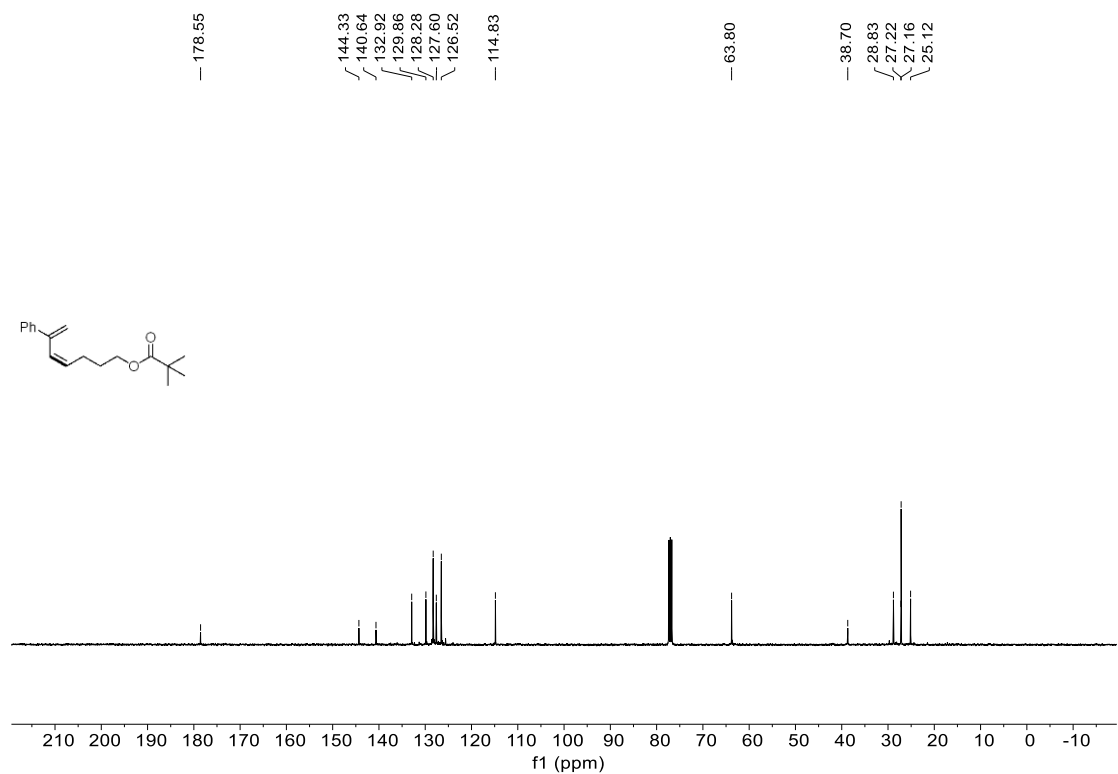
¹H NMR (400 MHz, CDCl₃) spectra of 3a



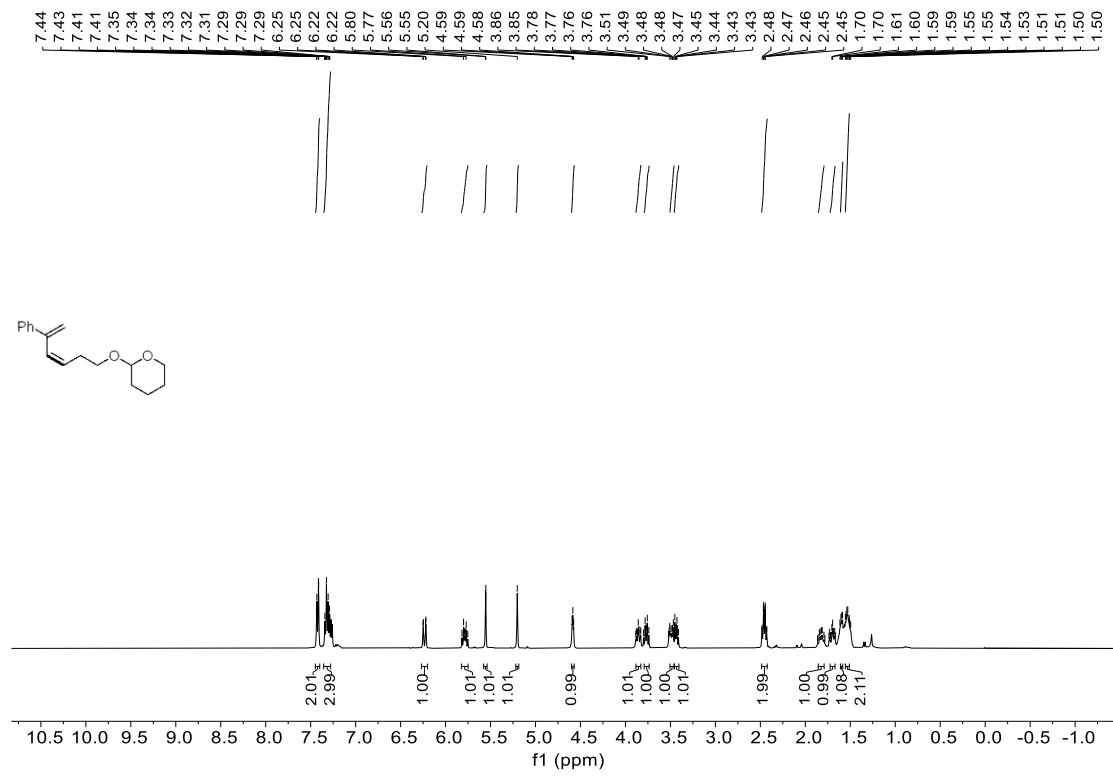
¹³C NMR (126 MHz, CDCl₃) spectra of 3a



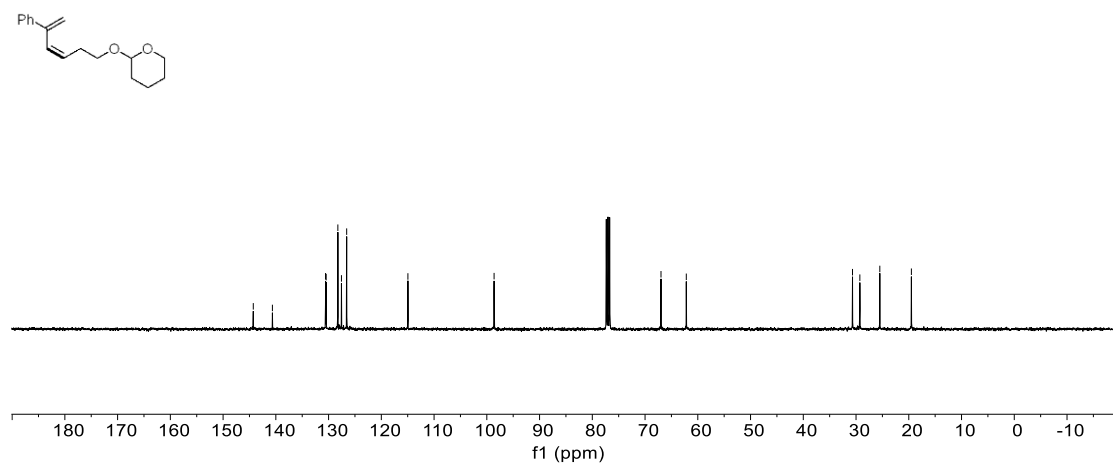
¹H NMR (400 MHz, CDCl₃) spectra of **4a**



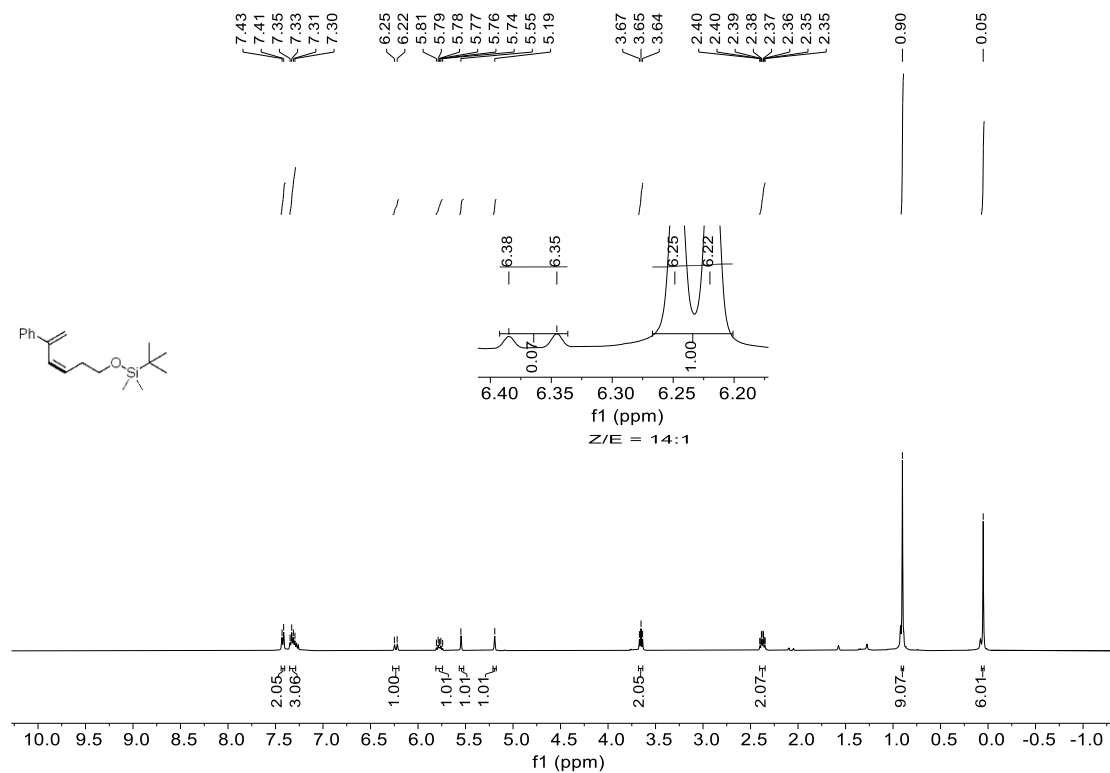
¹³C NMR (101 MHz, CDCl₃) spectra of **4a**



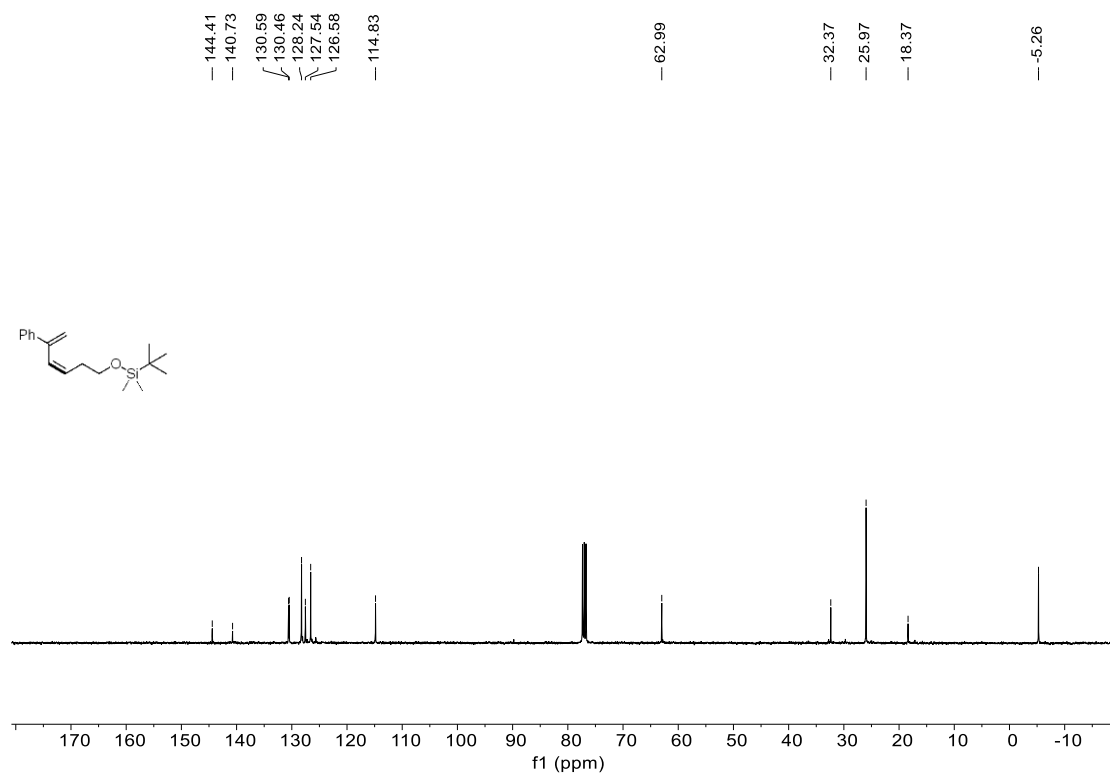
¹H NMR (400 MHz, CDCl₃) spectra of 5a



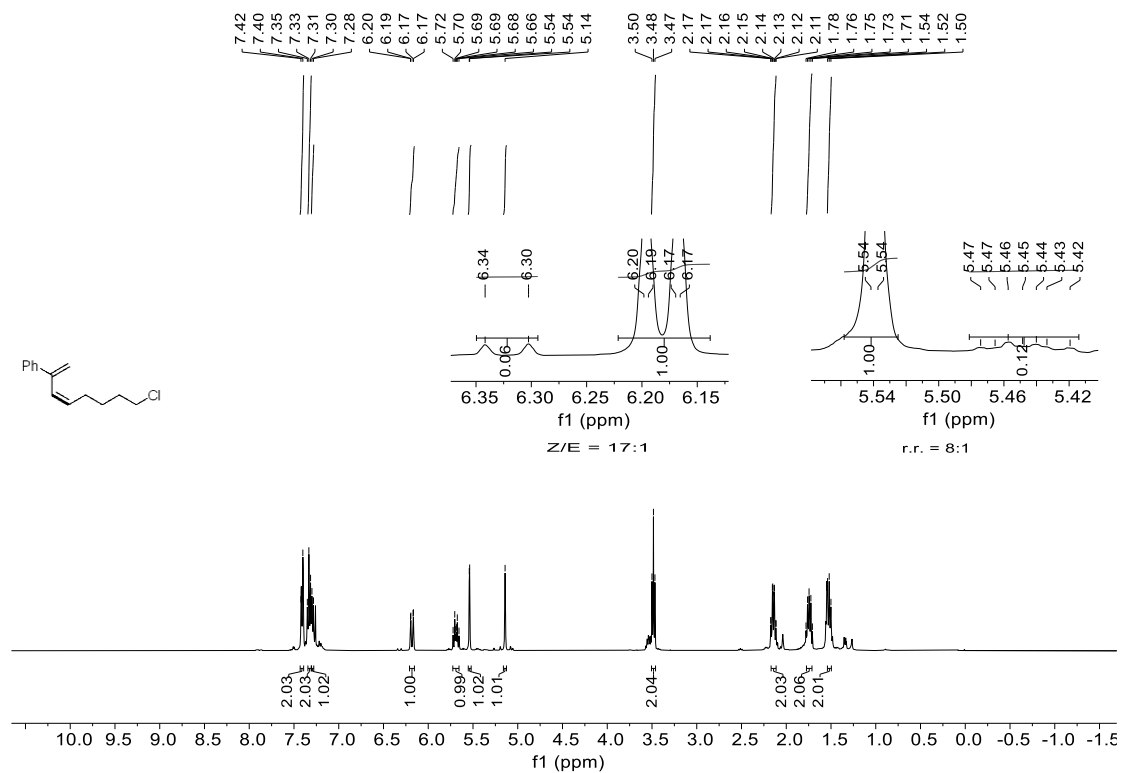
¹³C NMR (101 MHz, CDCl₃) spectra of 5a



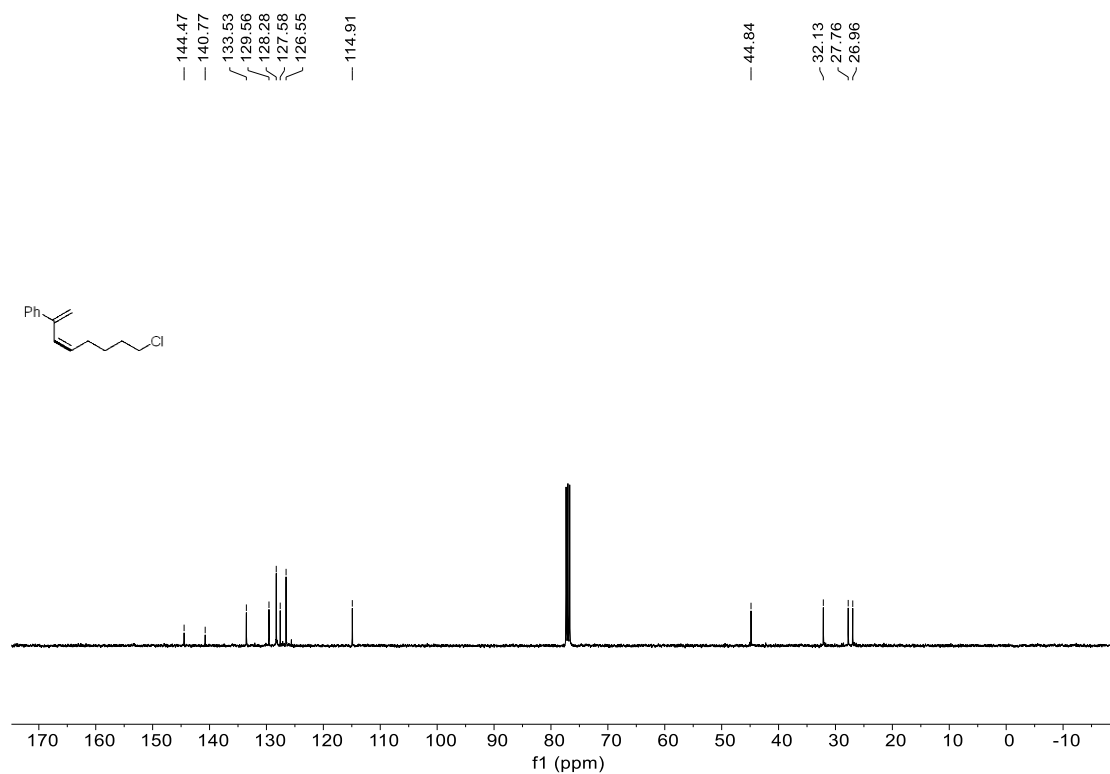
¹H NMR (400 MHz, CDCl₃) spectra of **6a**



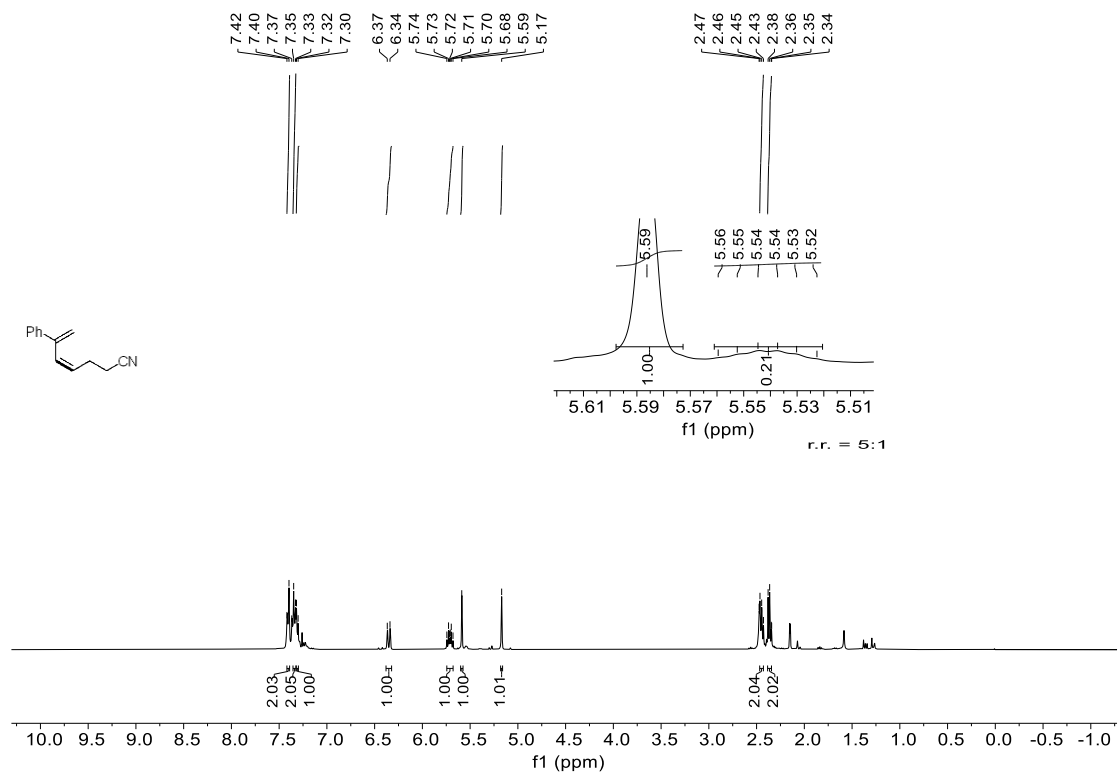
¹³C NMR (101 MHz, CDCl₃) spectra of **6a**



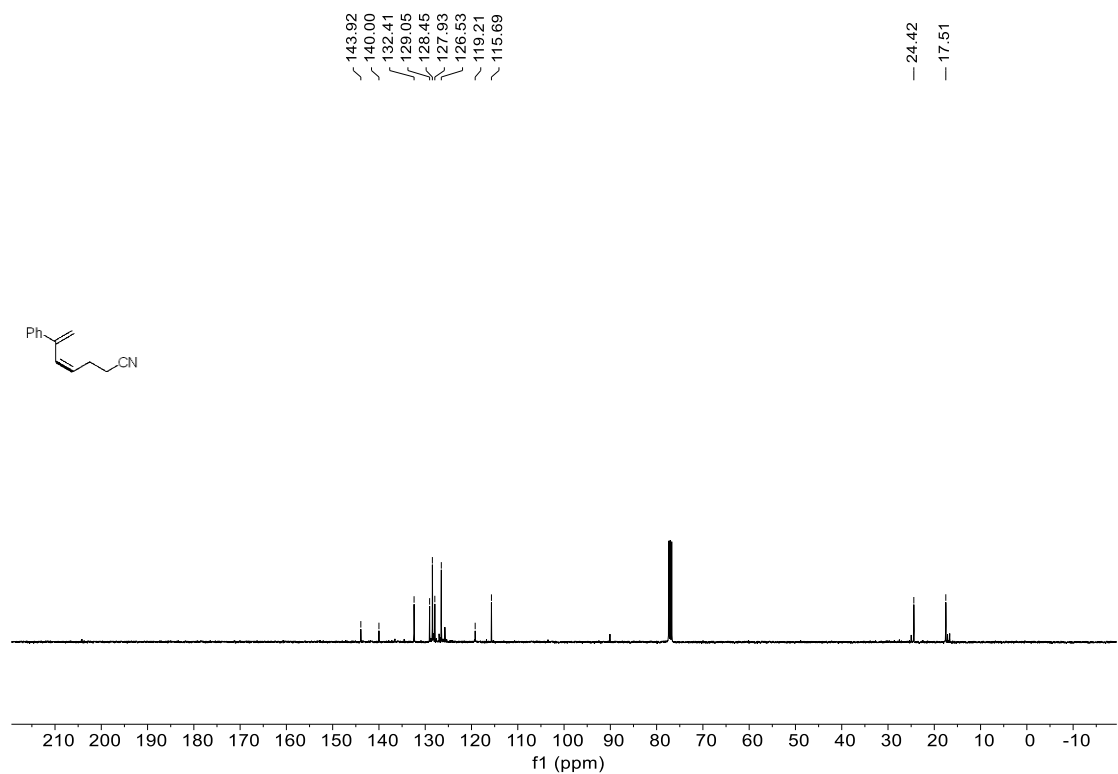
¹H NMR (400 MHz, CDCl₃) spectra of 7a



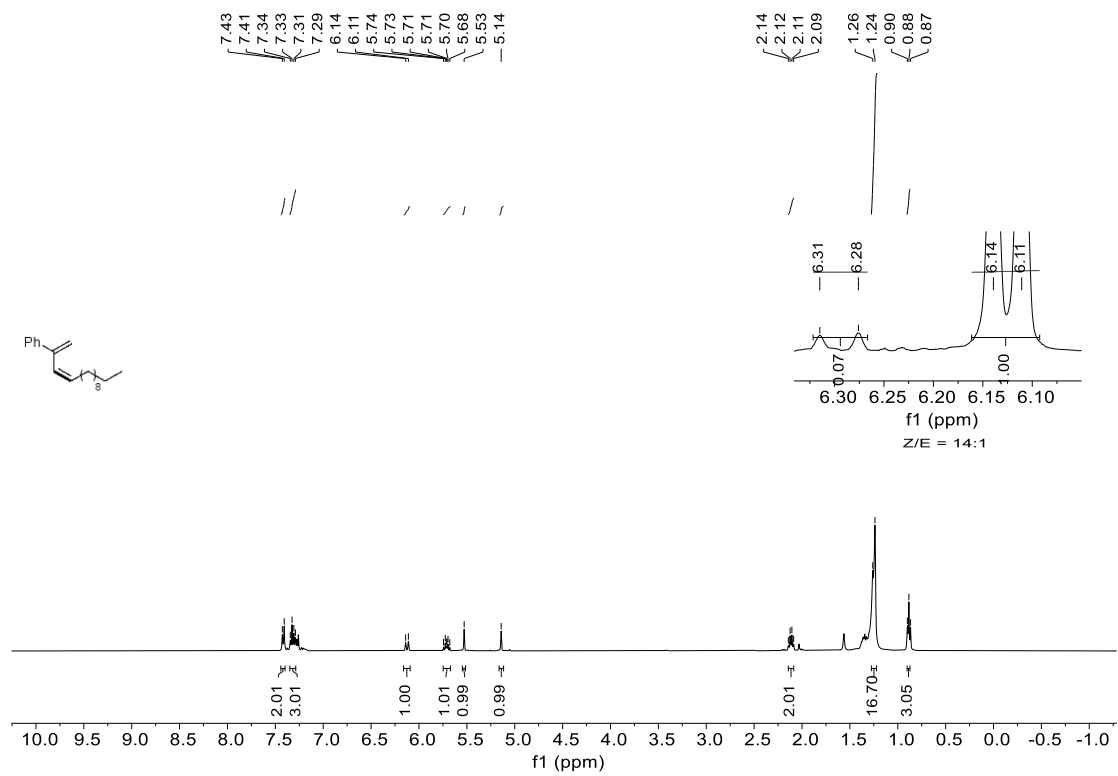
¹³C NMR (101 MHz, CDCl₃) spectra of 7a



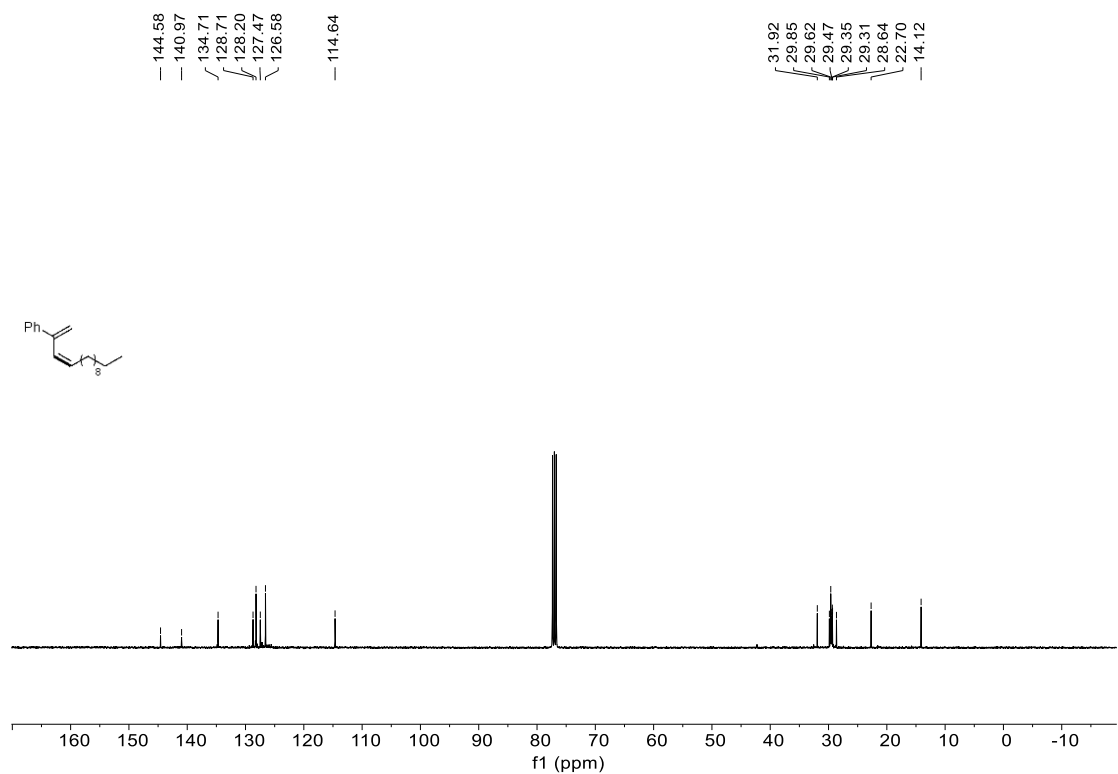
^1H NMR (400 MHz, CDCl_3) spectra of **8a**



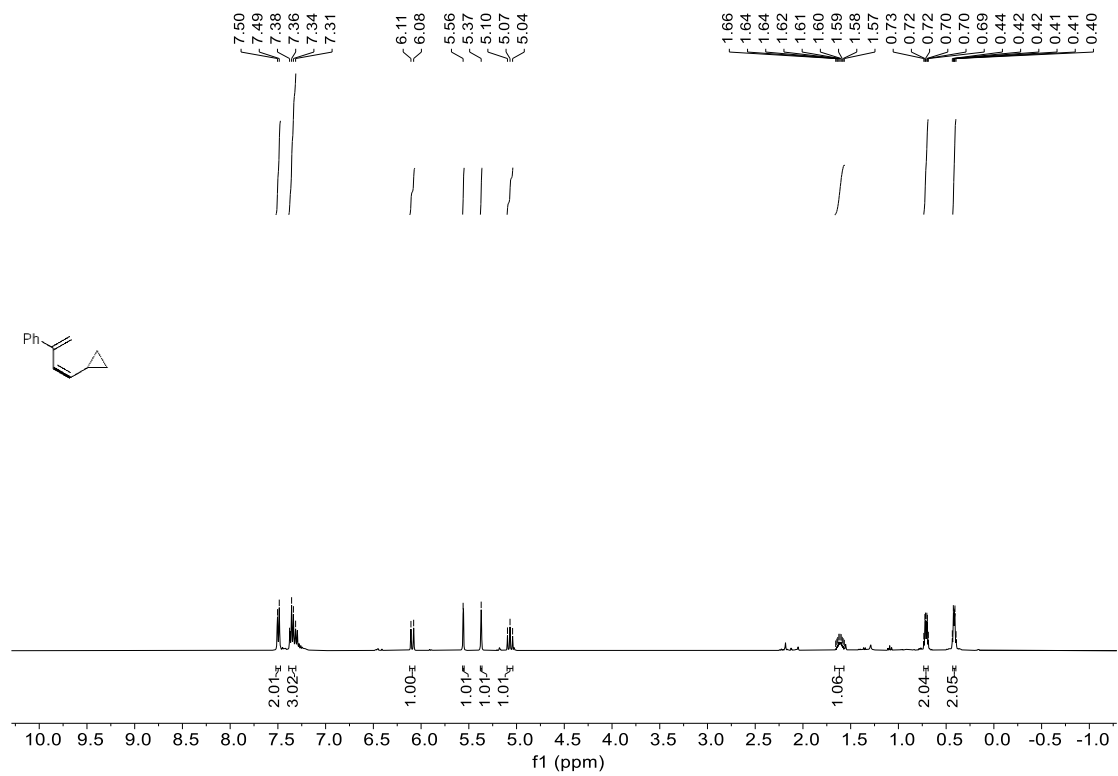
^{13}C NMR (101 MHz, CDCl_3) spectra of **8a**



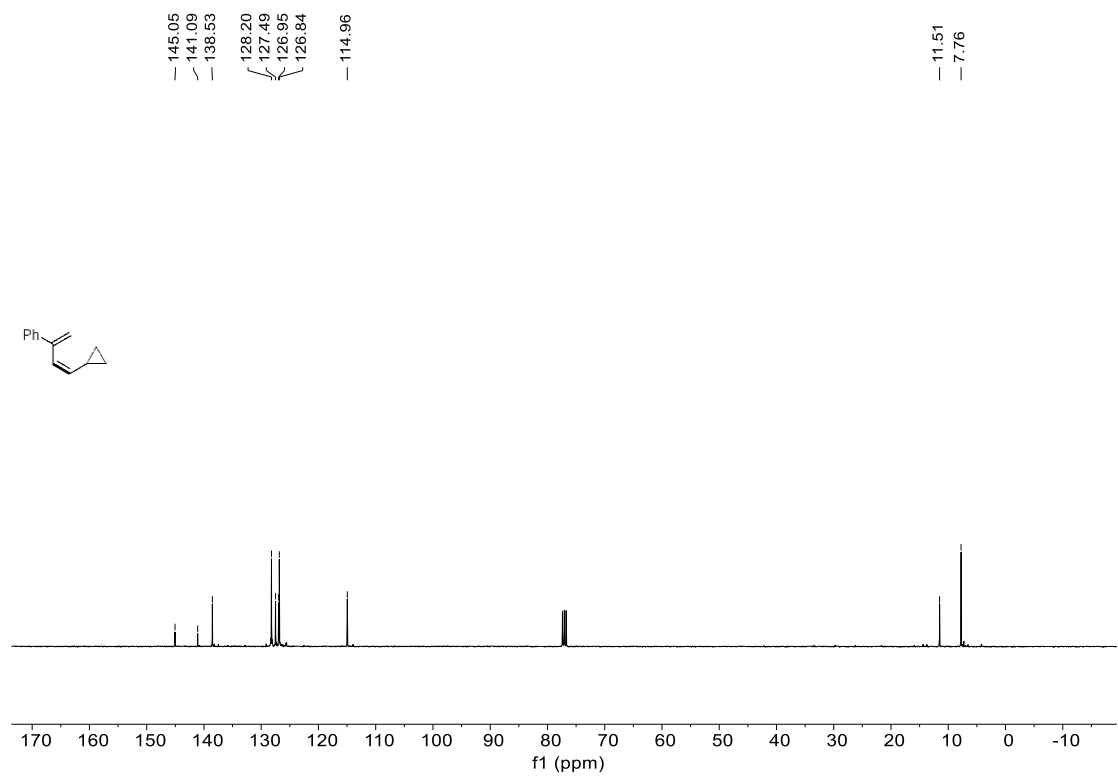
^1H NMR (400 MHz, CDCl_3) spectra of **9a**



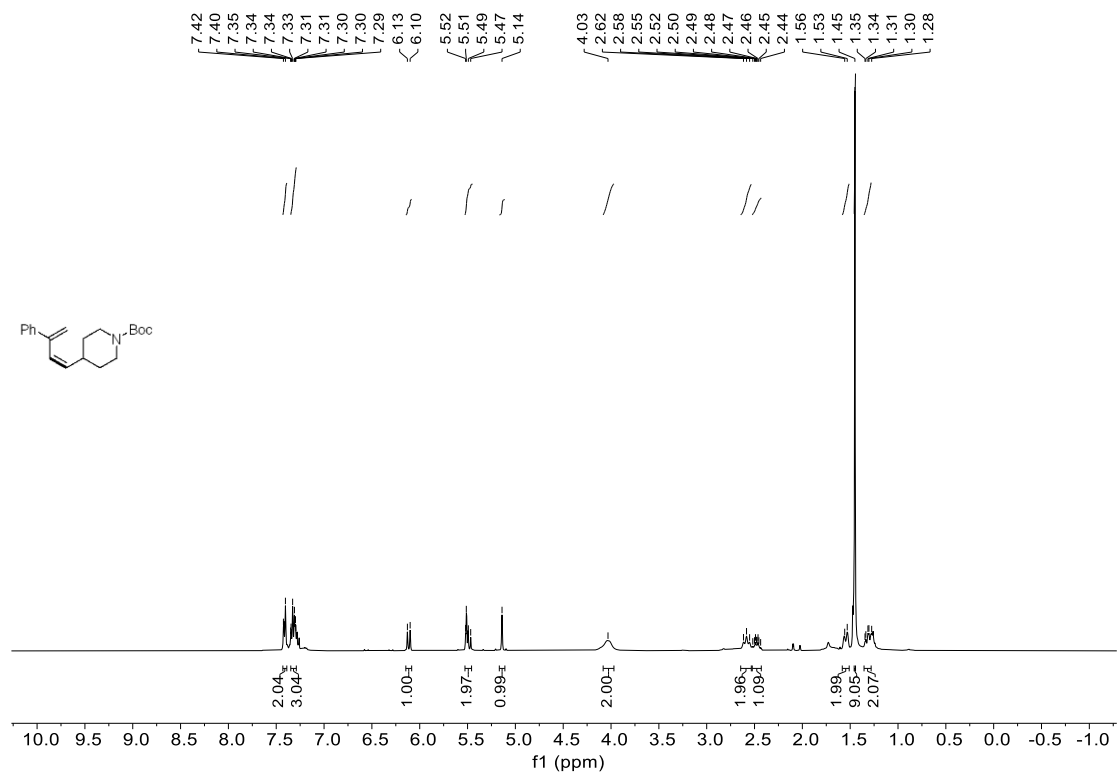
^{13}C NMR (101 MHz, CDCl_3) spectra of **9a**



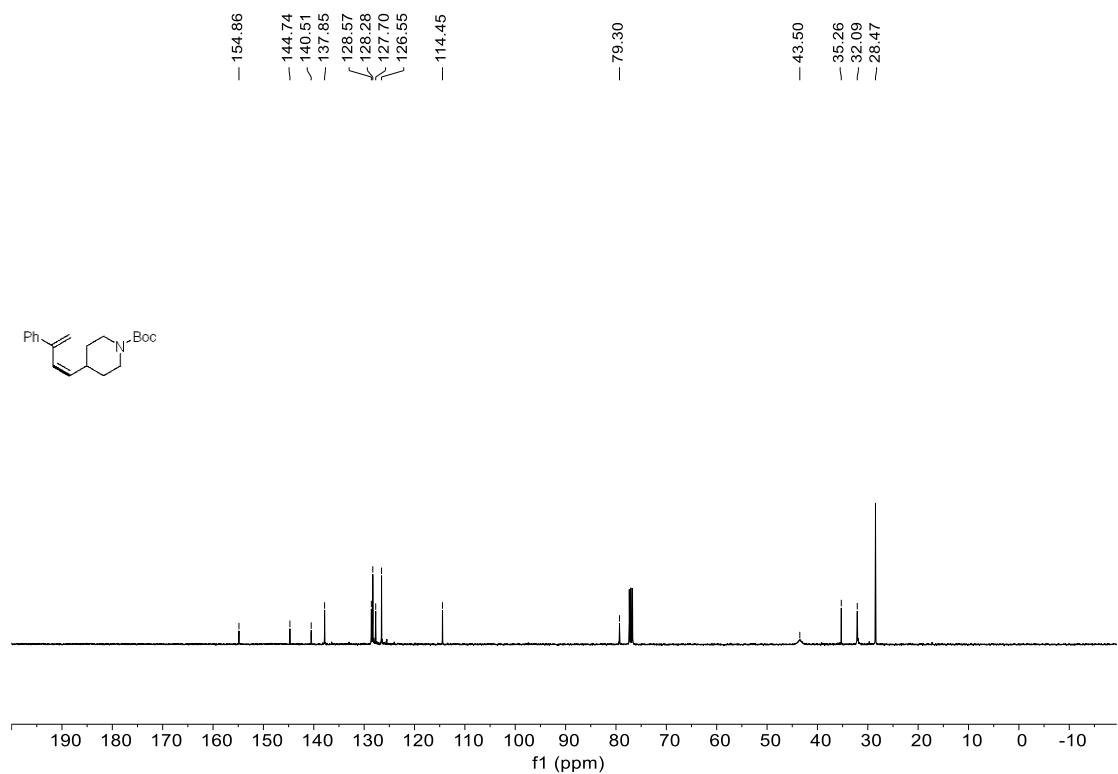
¹H NMR (400 MHz, CDCl₃) spectra of **10a**



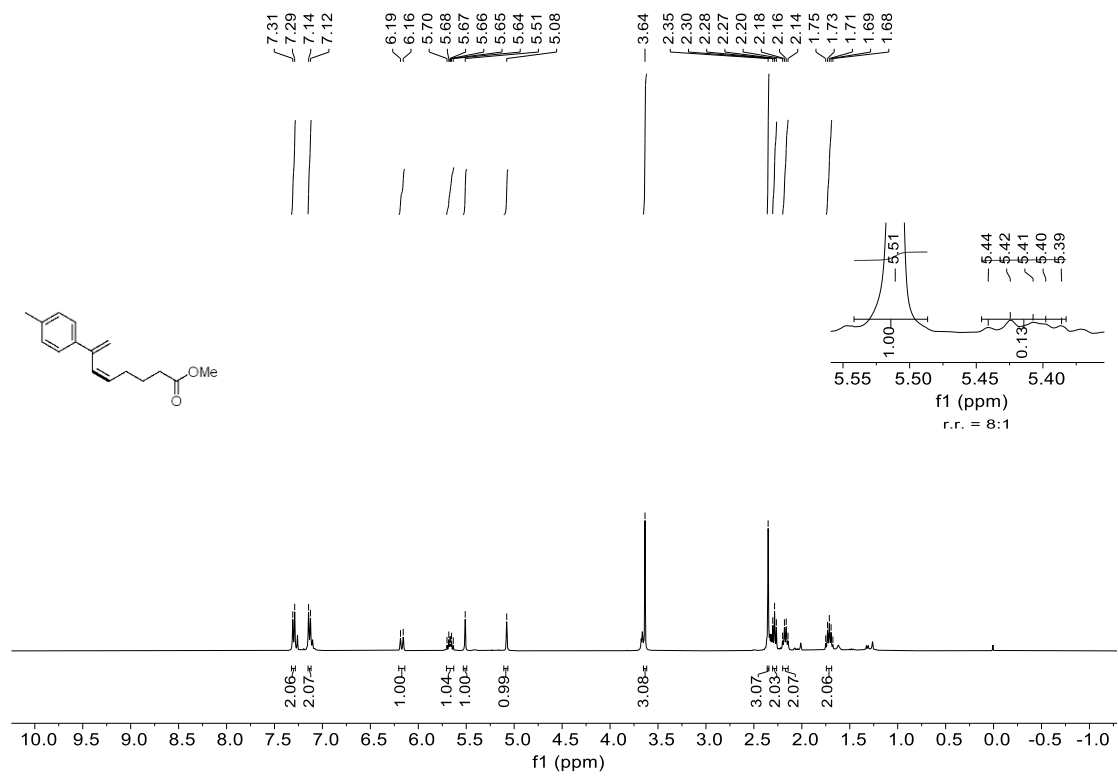
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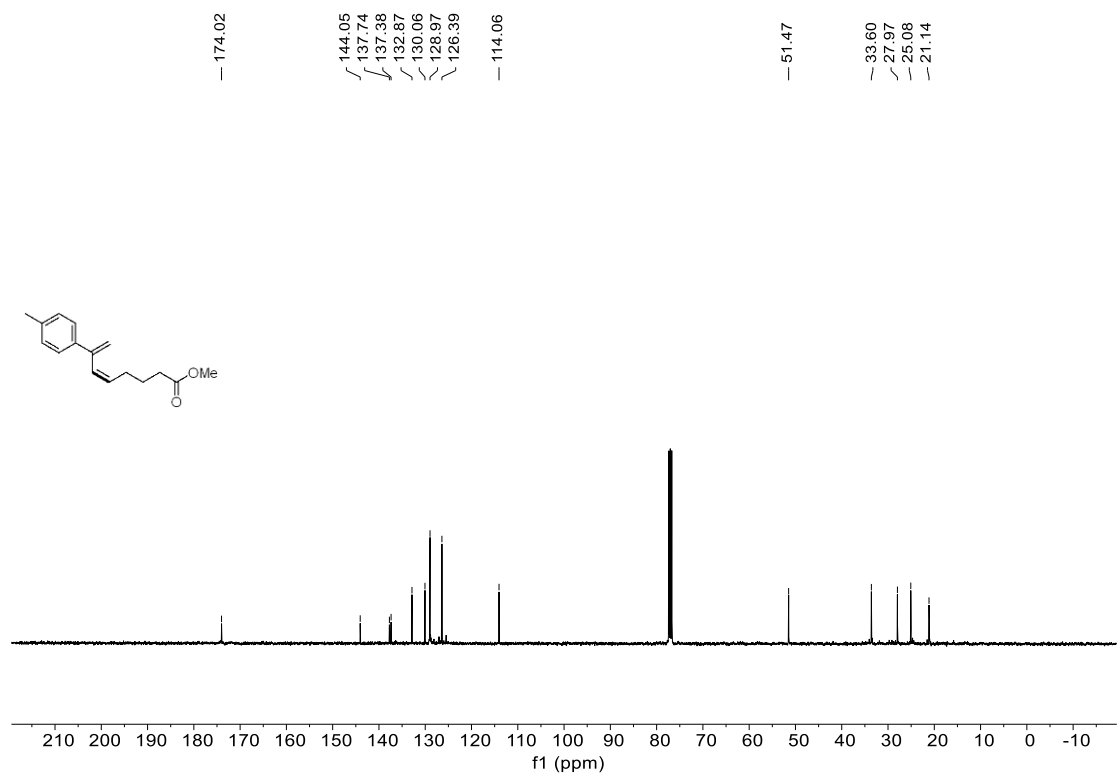
¹H NMR (400 MHz, CDCl₃) spectra of **11a**



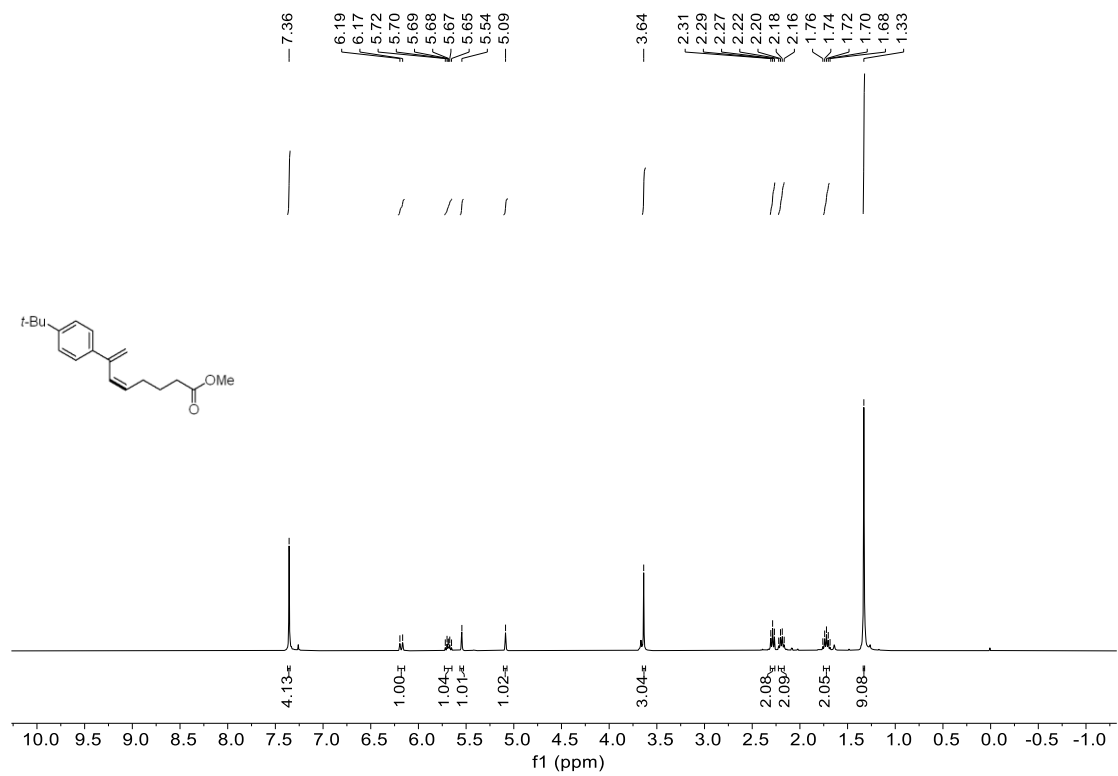
¹³C NMR (101 MHz, CDCl₃) spectra of **11a**



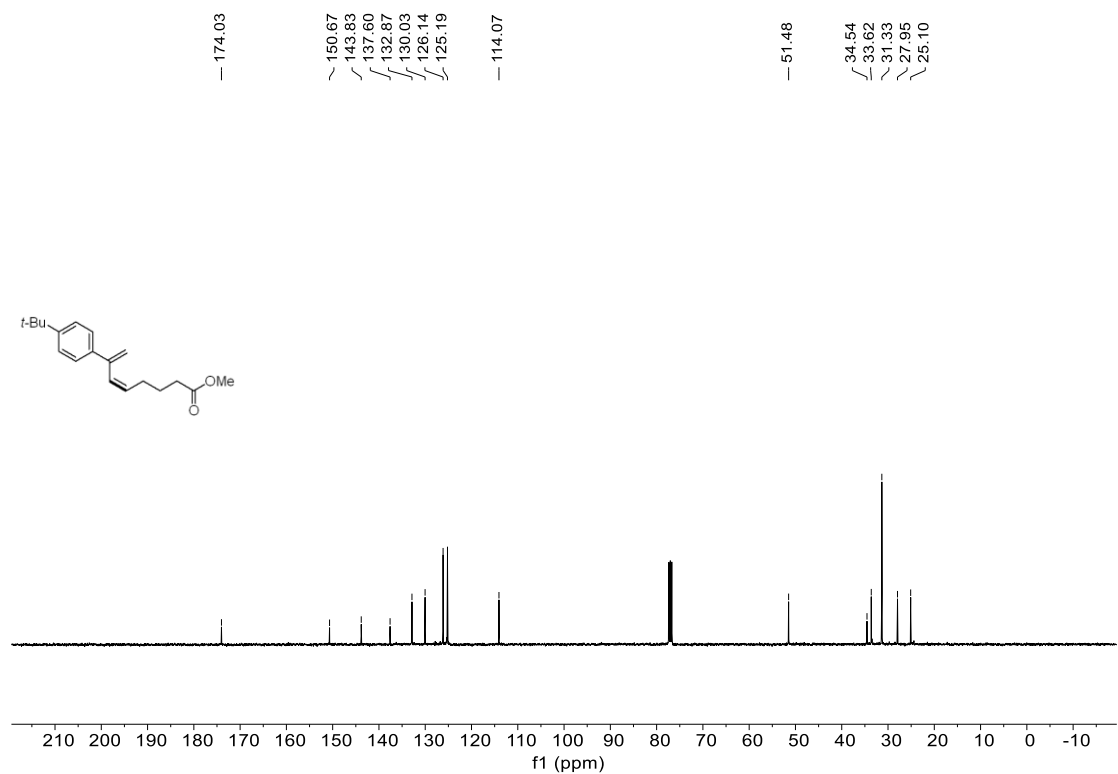
¹H NMR (400 MHz, CDCl₃) spectra of **12a**



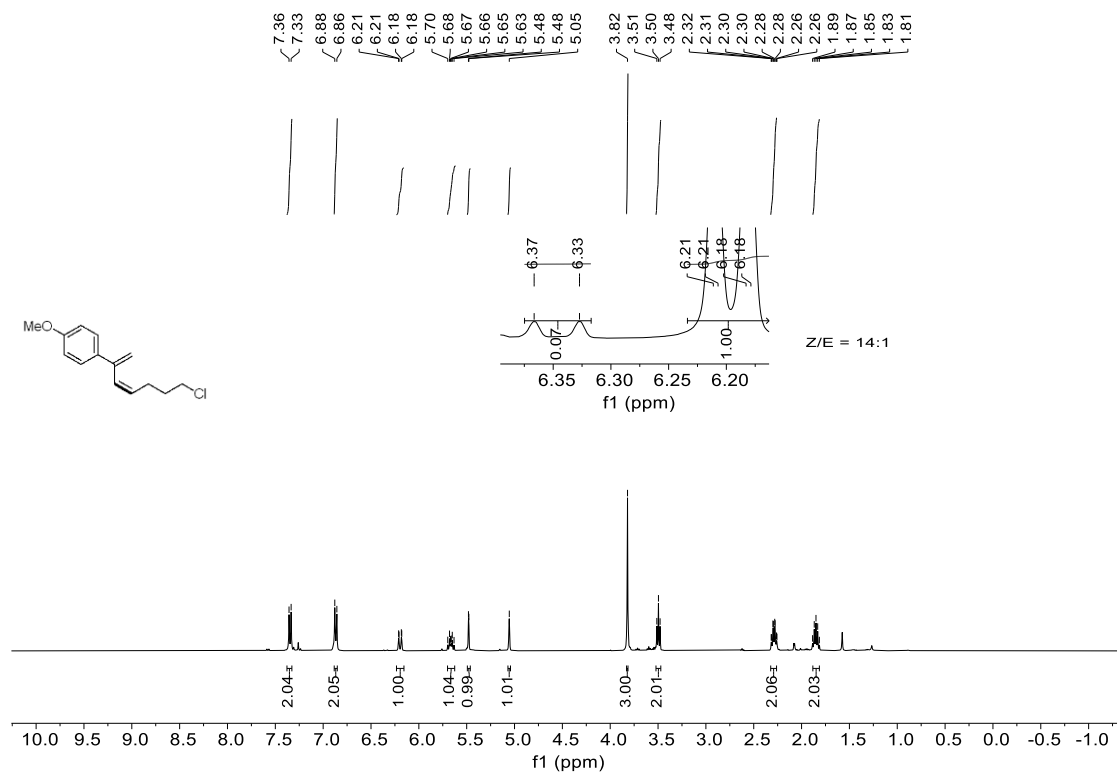
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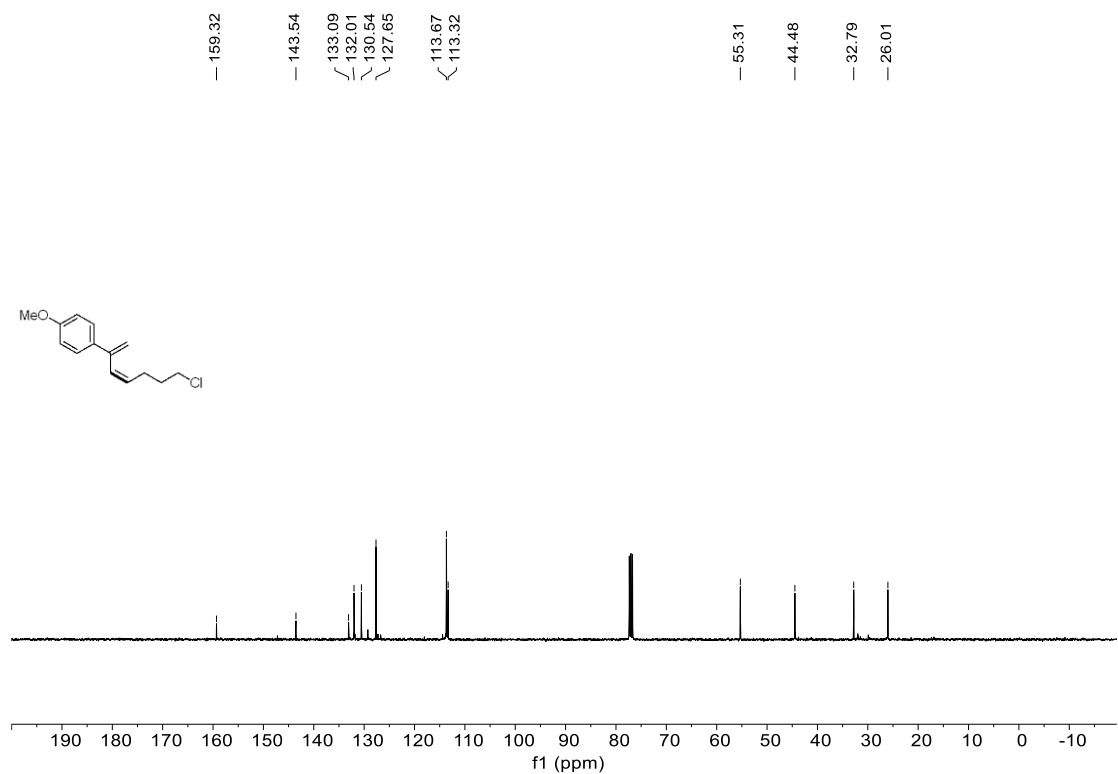
¹H NMR (400 MHz, CDCl₃) spectra of **13a**



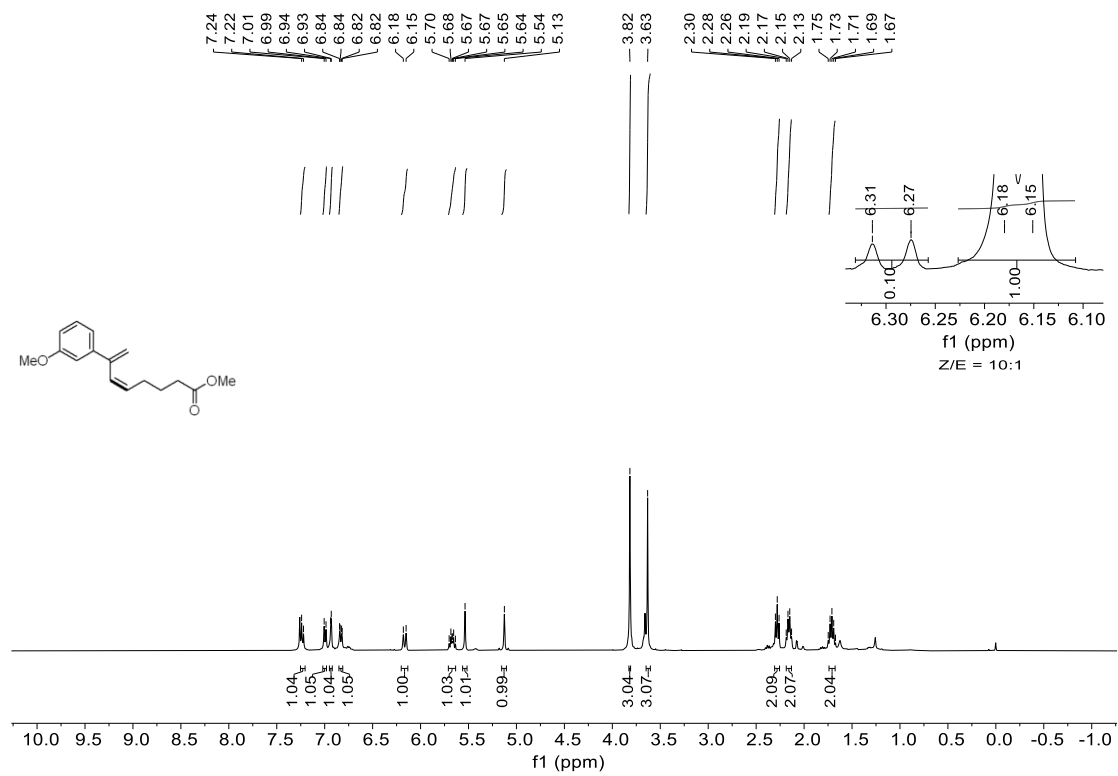
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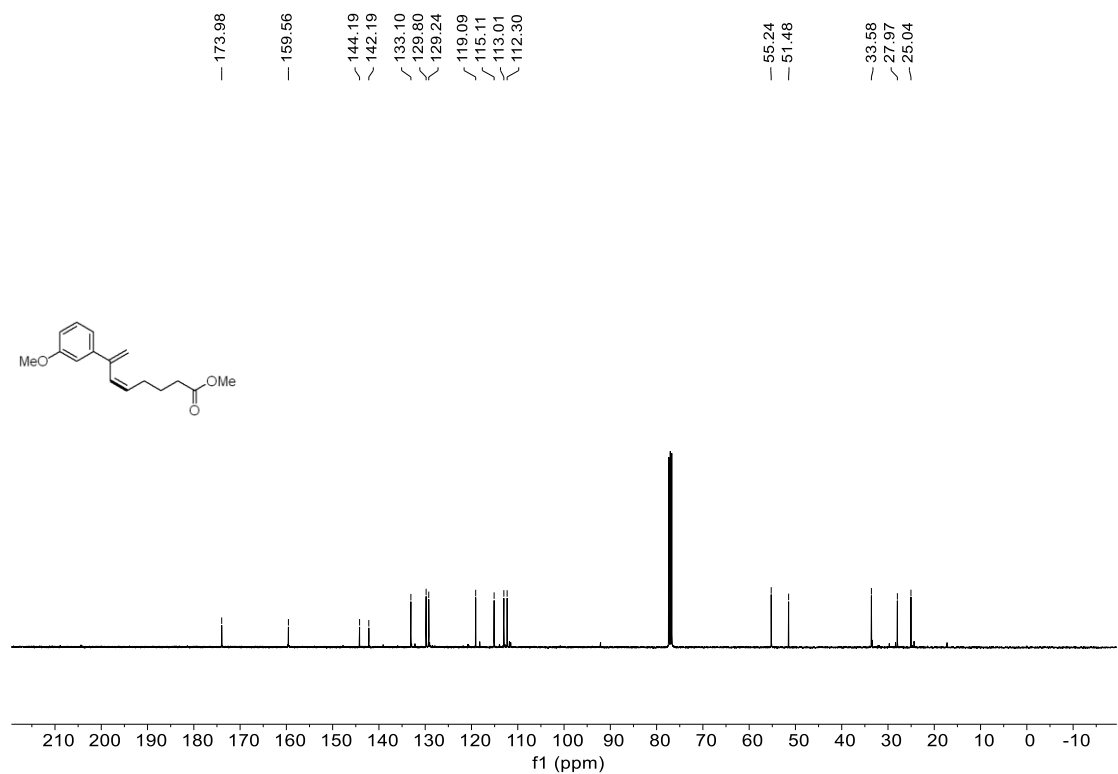
¹H NMR (400 MHz, CDCl₃) spectra of **14a**



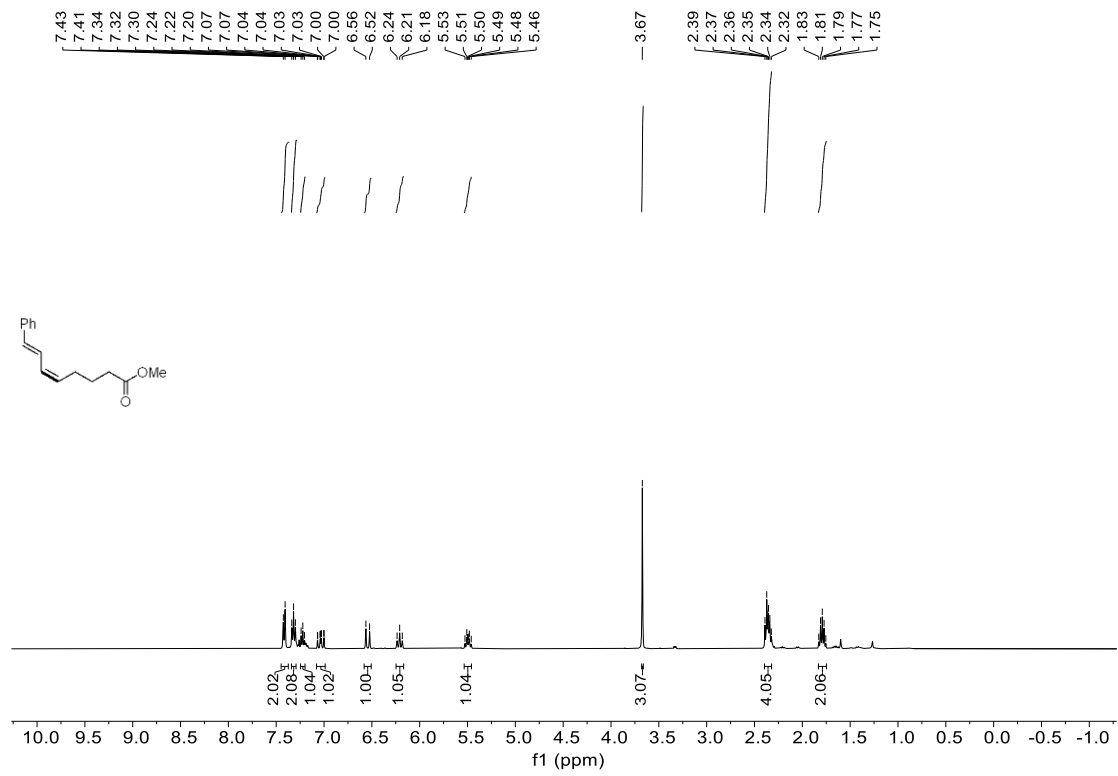
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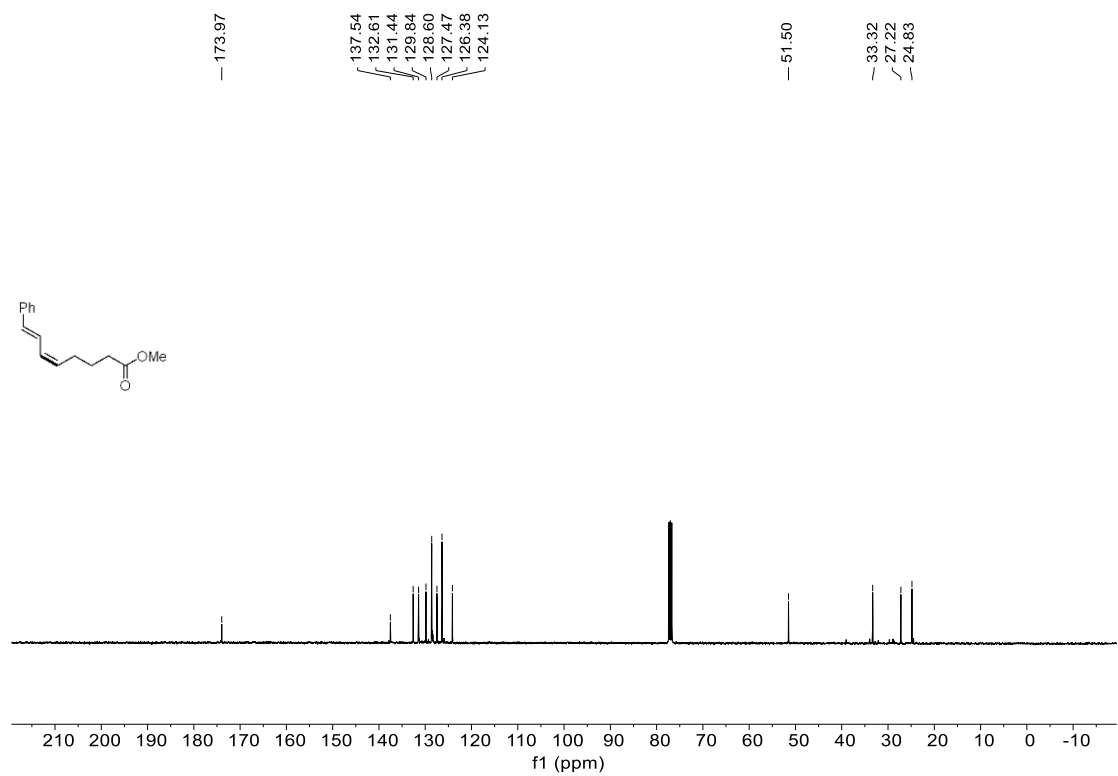
¹H NMR (400 MHz, CDCl₃) spectra of **15a**



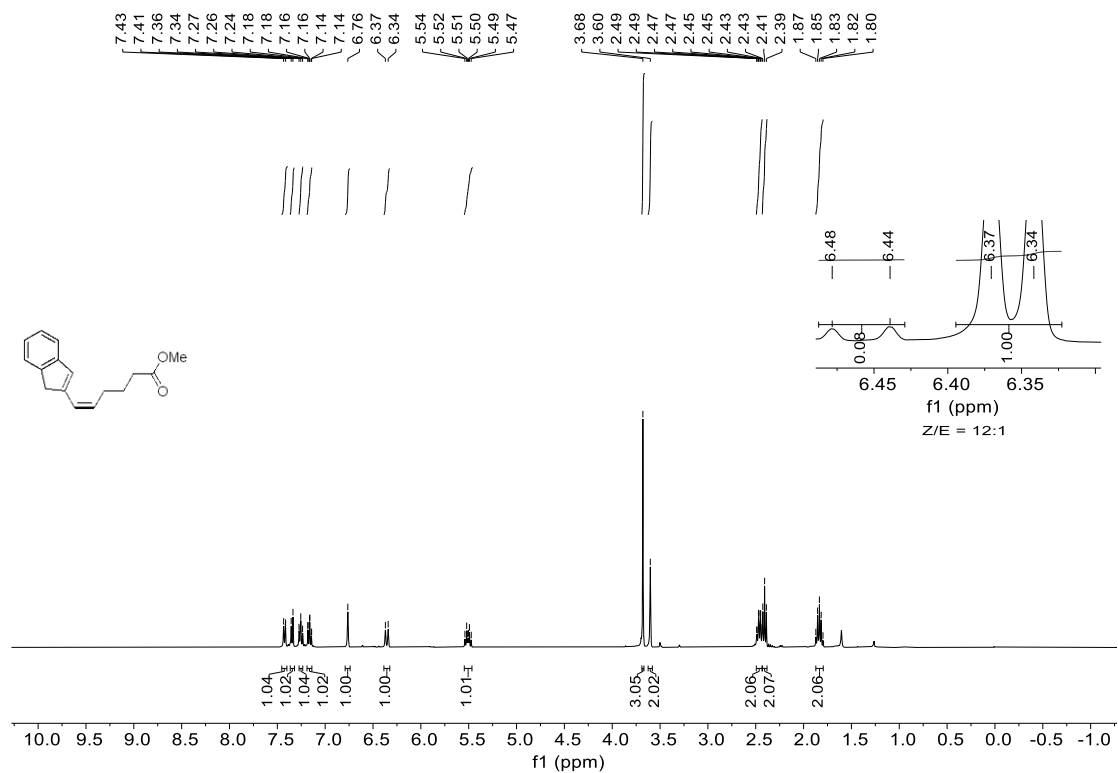
¹³C NMR (101 MHz, CDCl₃) spectra of **15a**



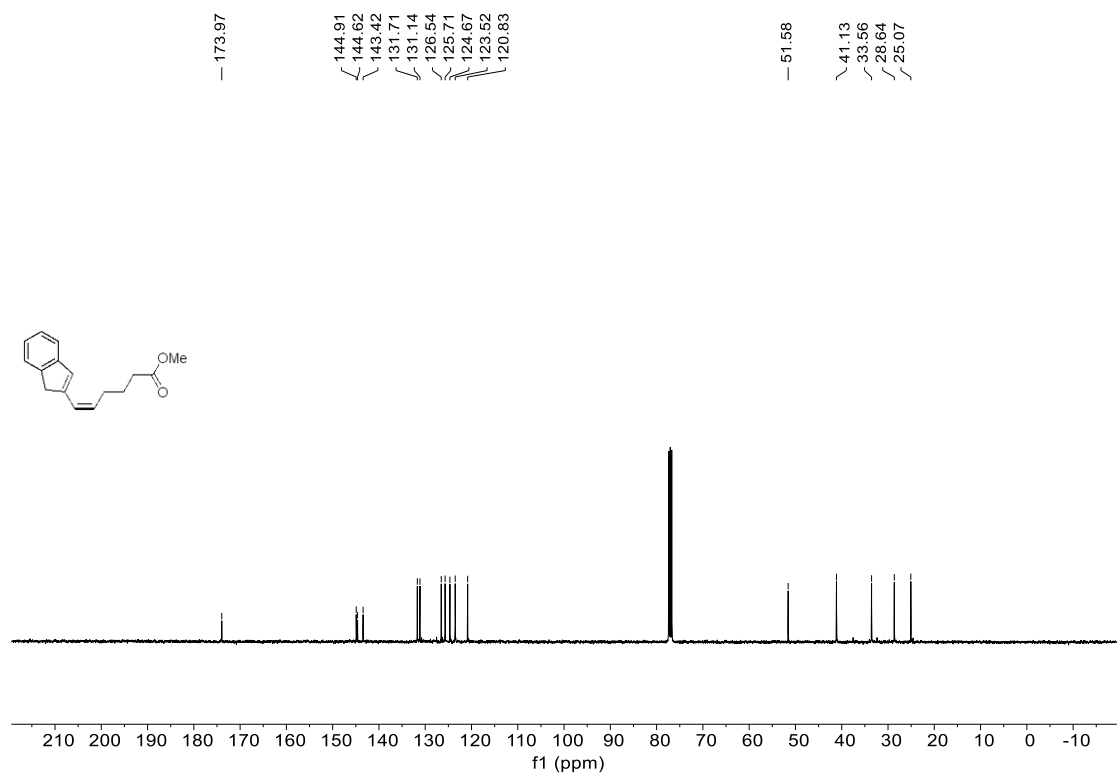
¹H NMR (400 MHz, CDCl₃) spectra of 16a



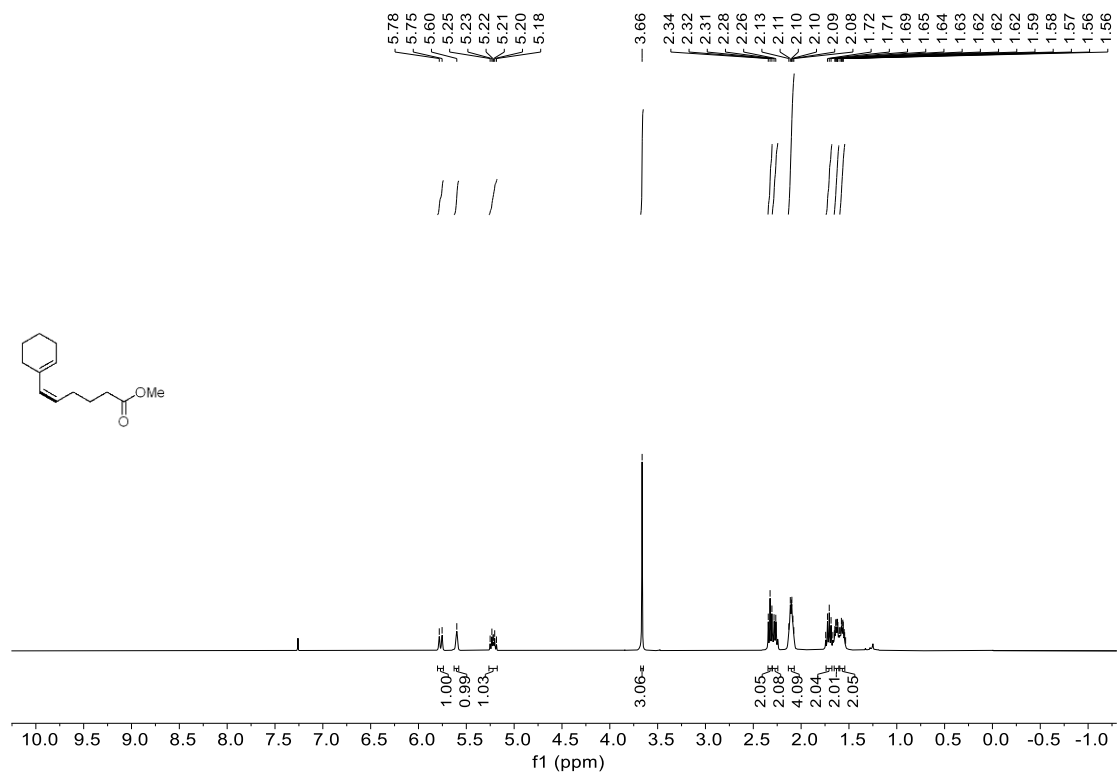
¹³C NMR (101 MHz, CDCl₃) spectra of 16a



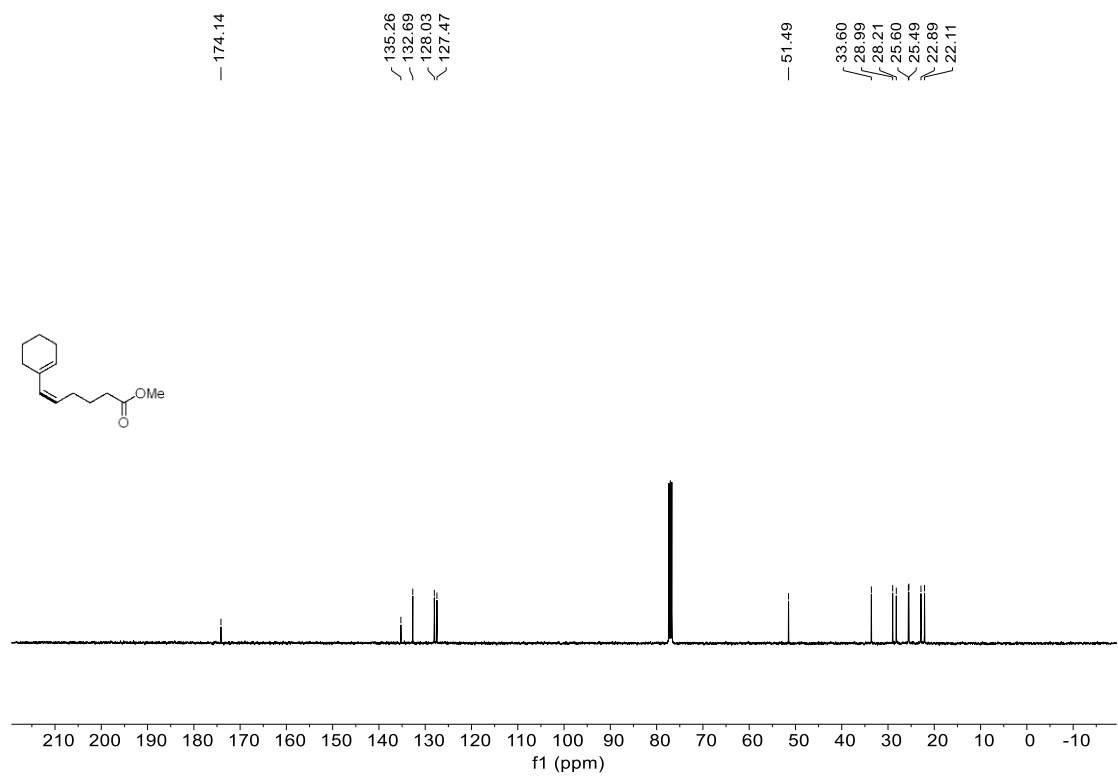
¹H NMR (400 MHz, CDCl₃) spectra of 17a



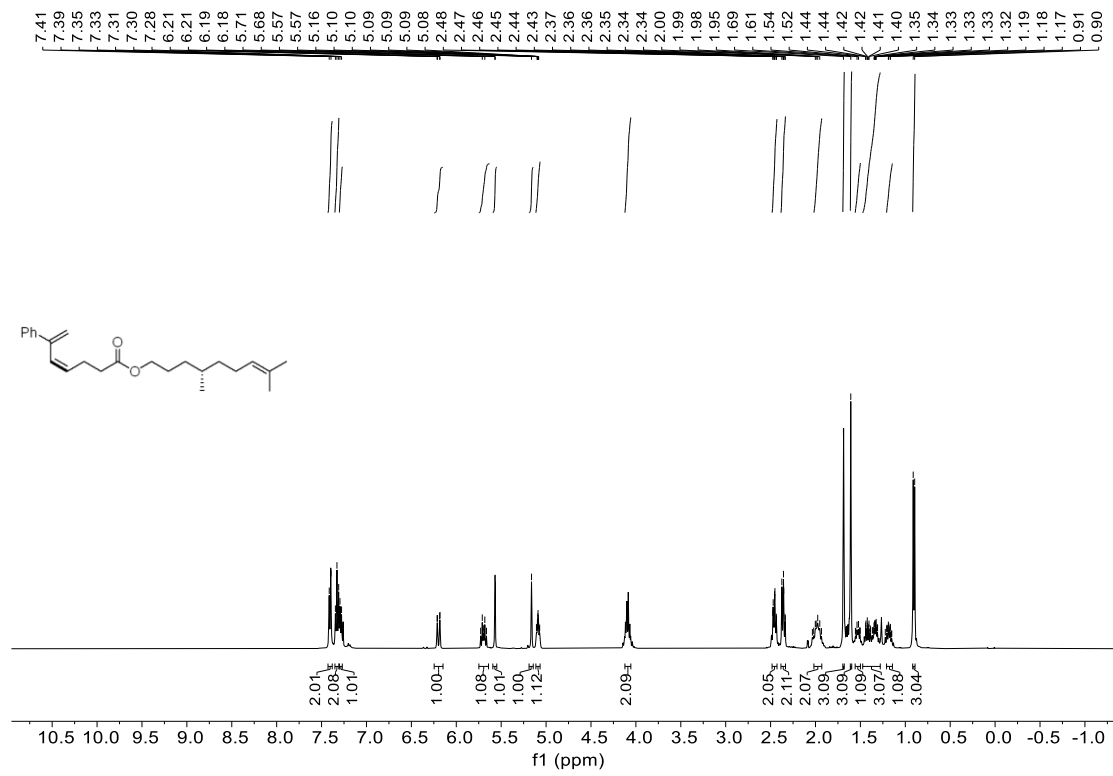
¹³C NMR (101 MHz, CDCl₃) spectra of 17a



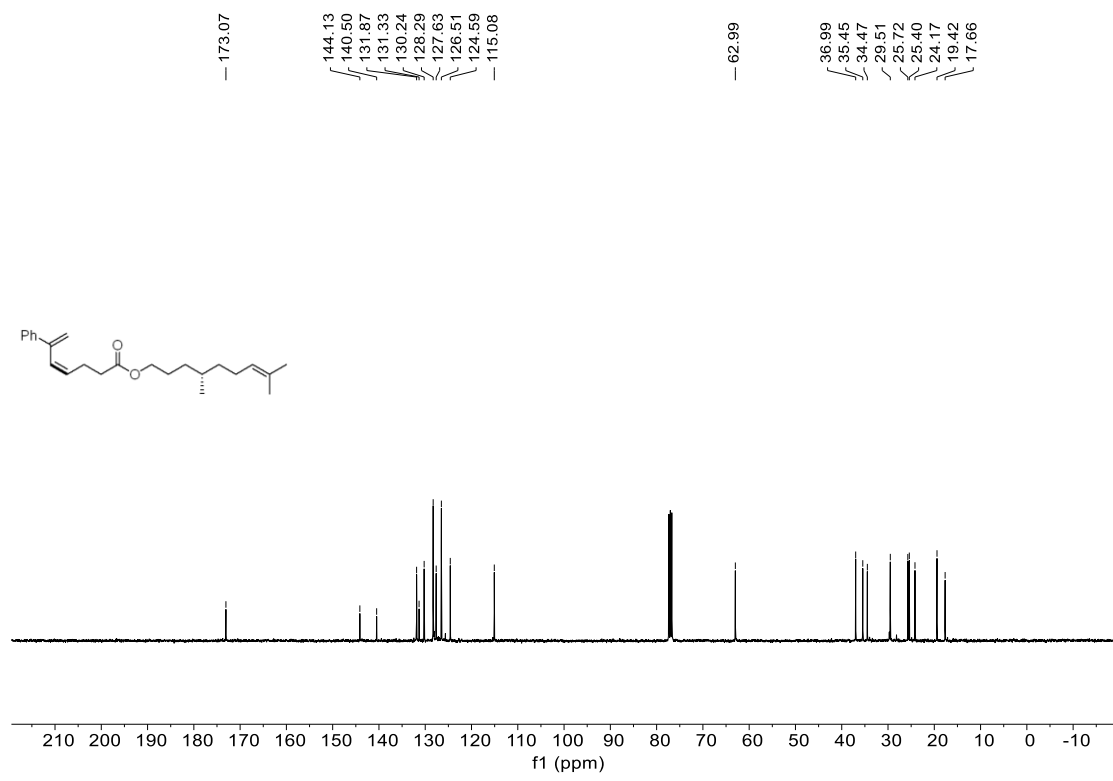
¹H NMR (400 MHz, CDCl₃) spectra of **18a**



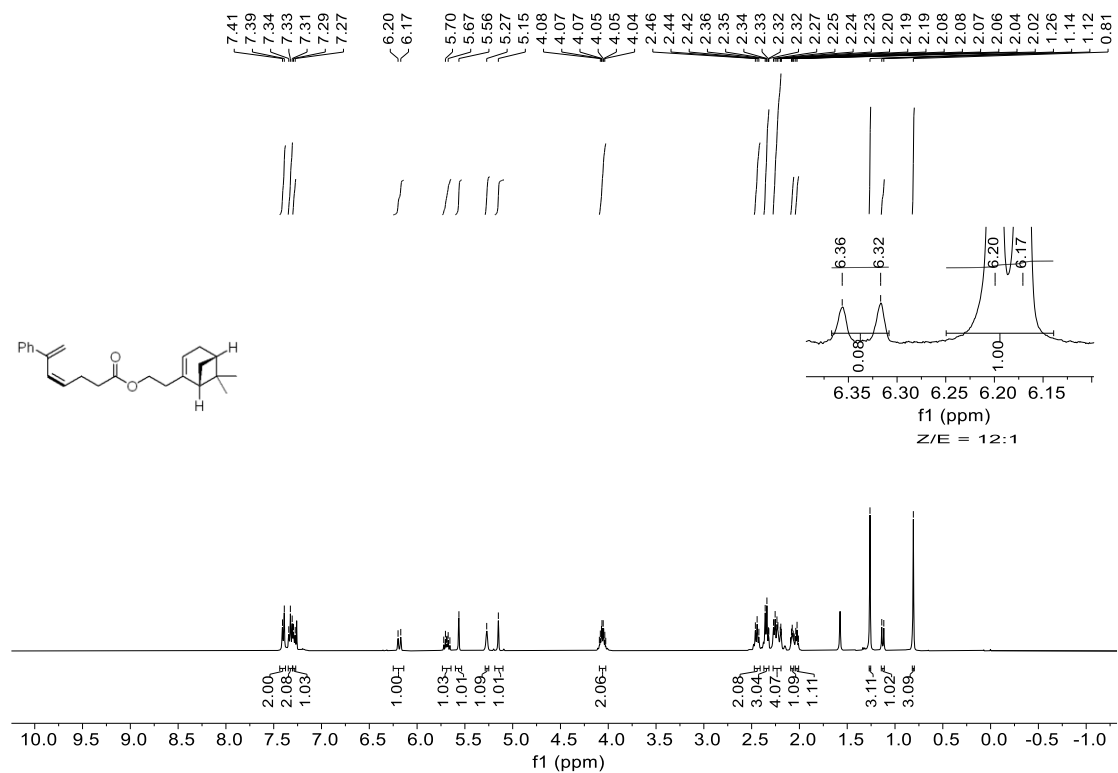
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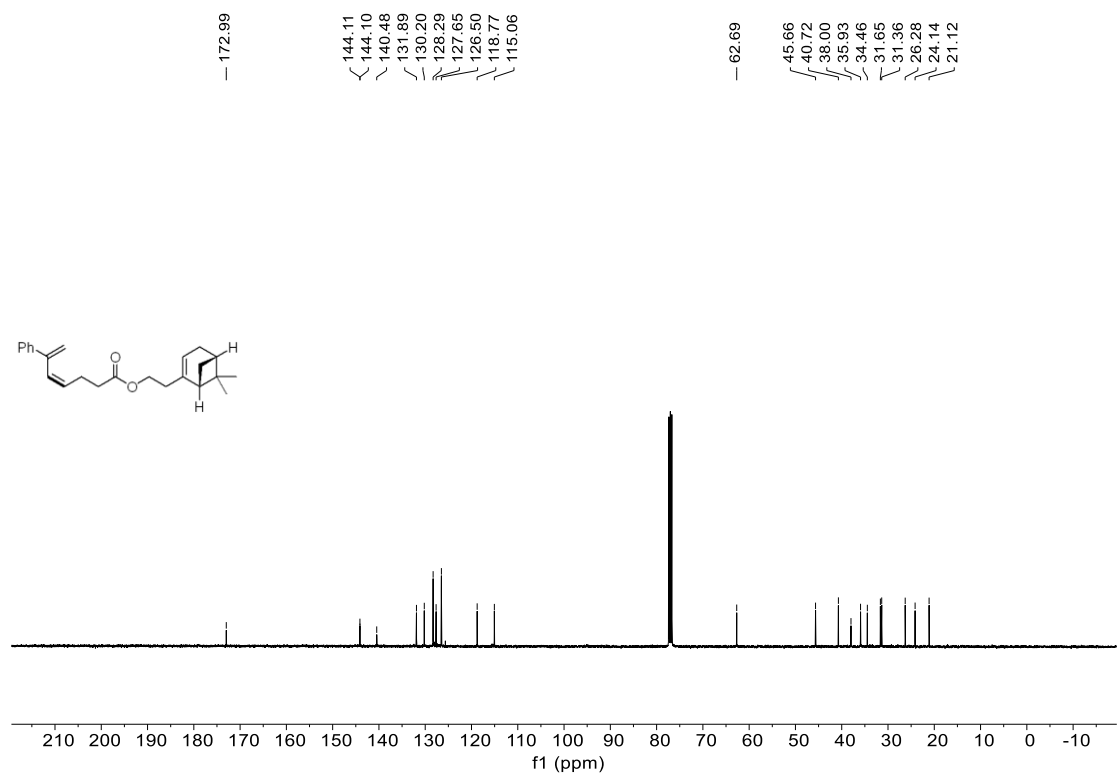
¹H NMR (400 MHz, CDCl₃) spectra of 19a



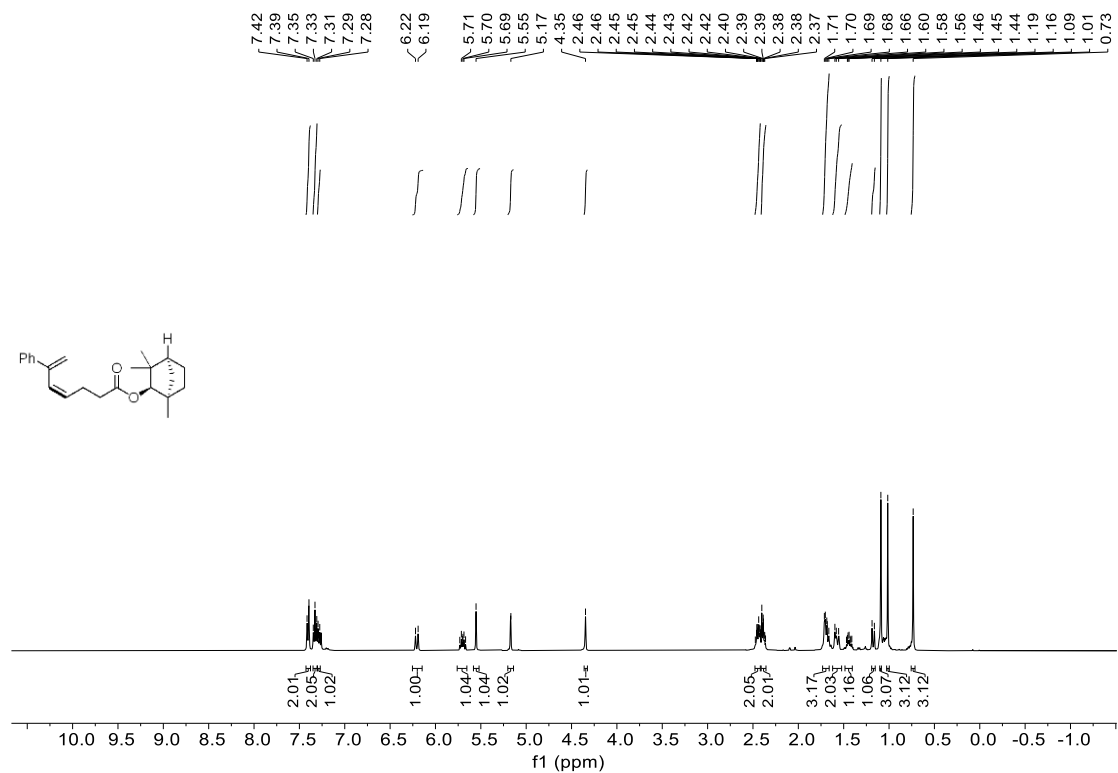
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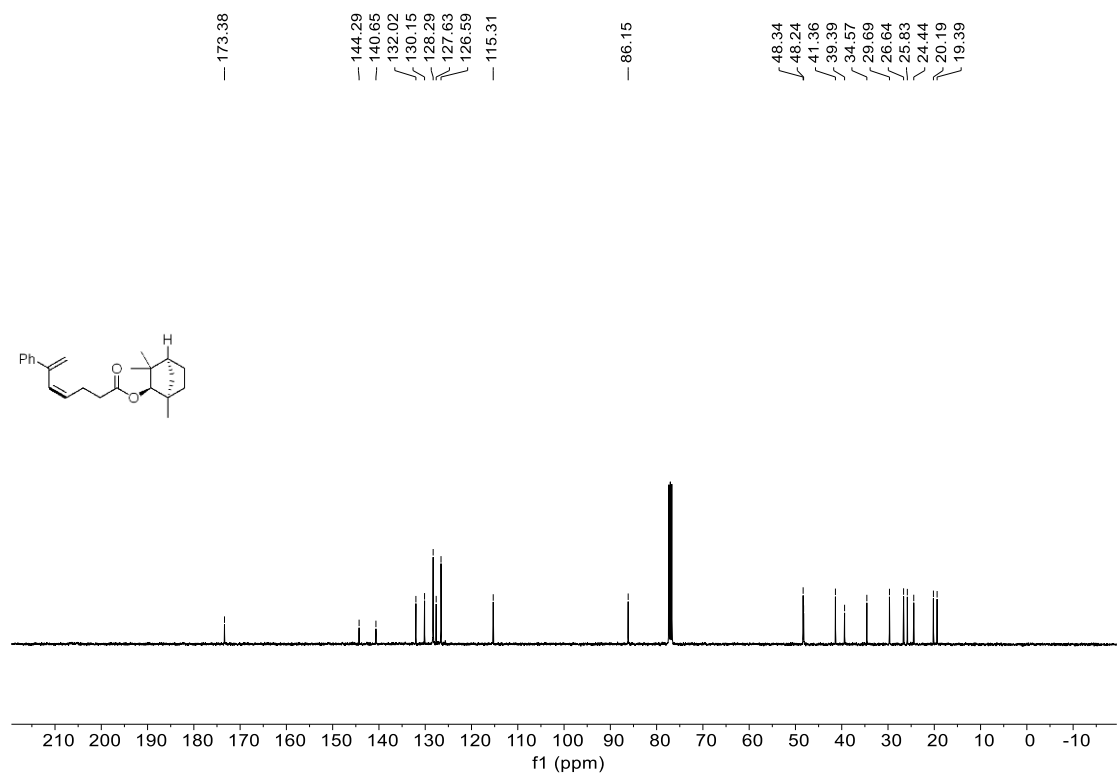
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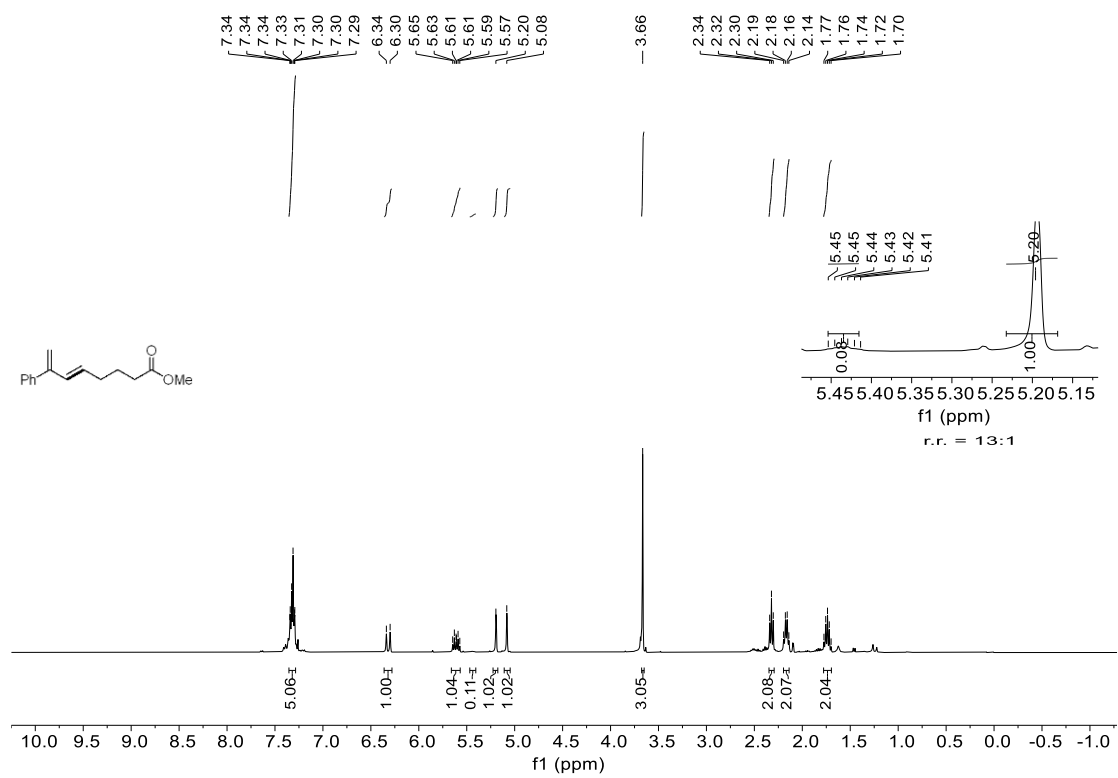
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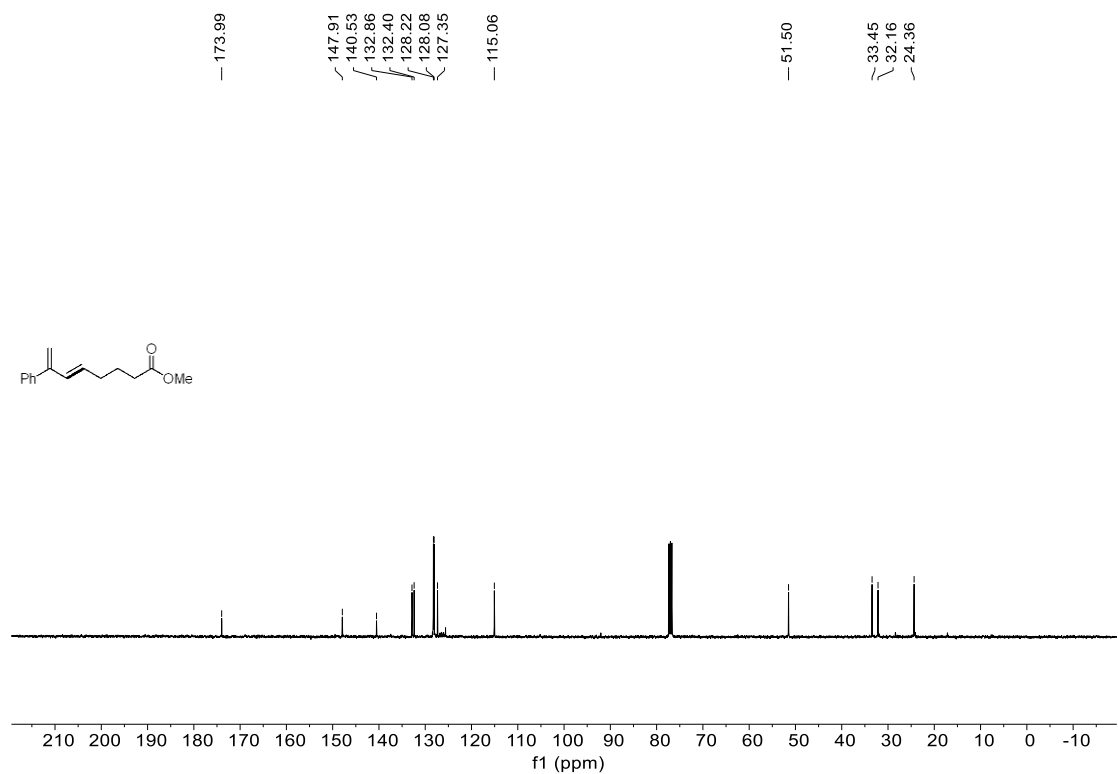
¹H NMR (400 MHz, CDCl₃) spectra of **21a**



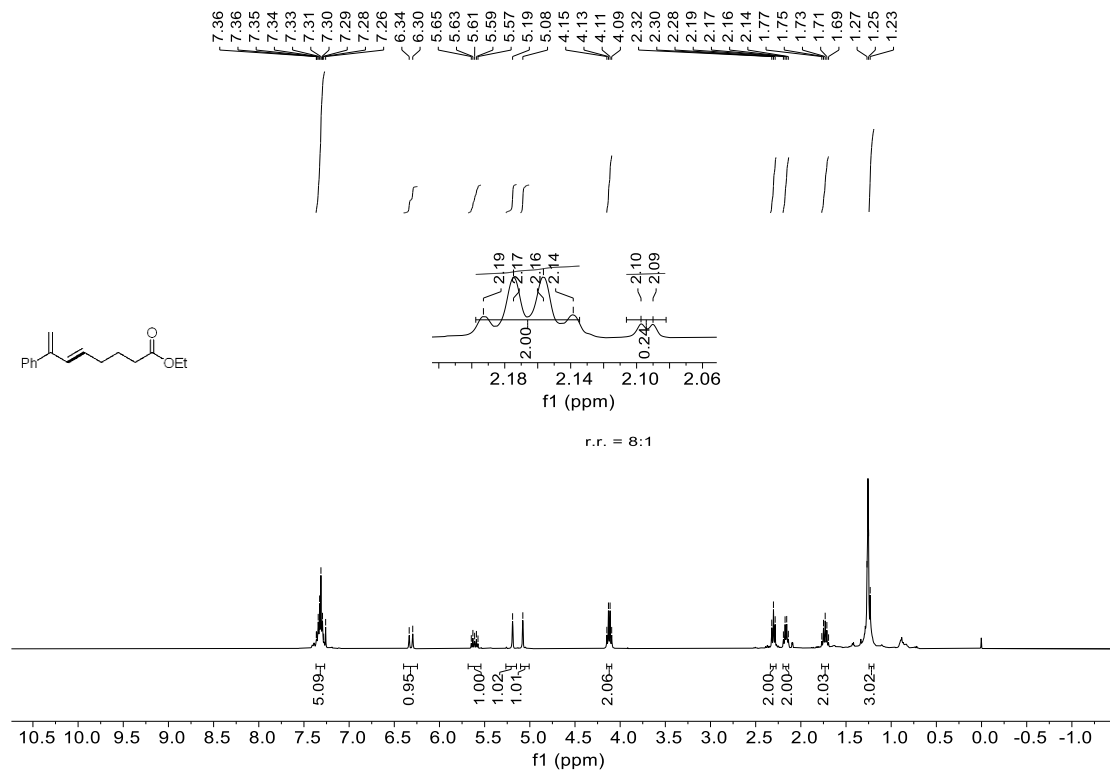
¹³C NMR (101 MHz, CDCl₃) spectra of **22a**



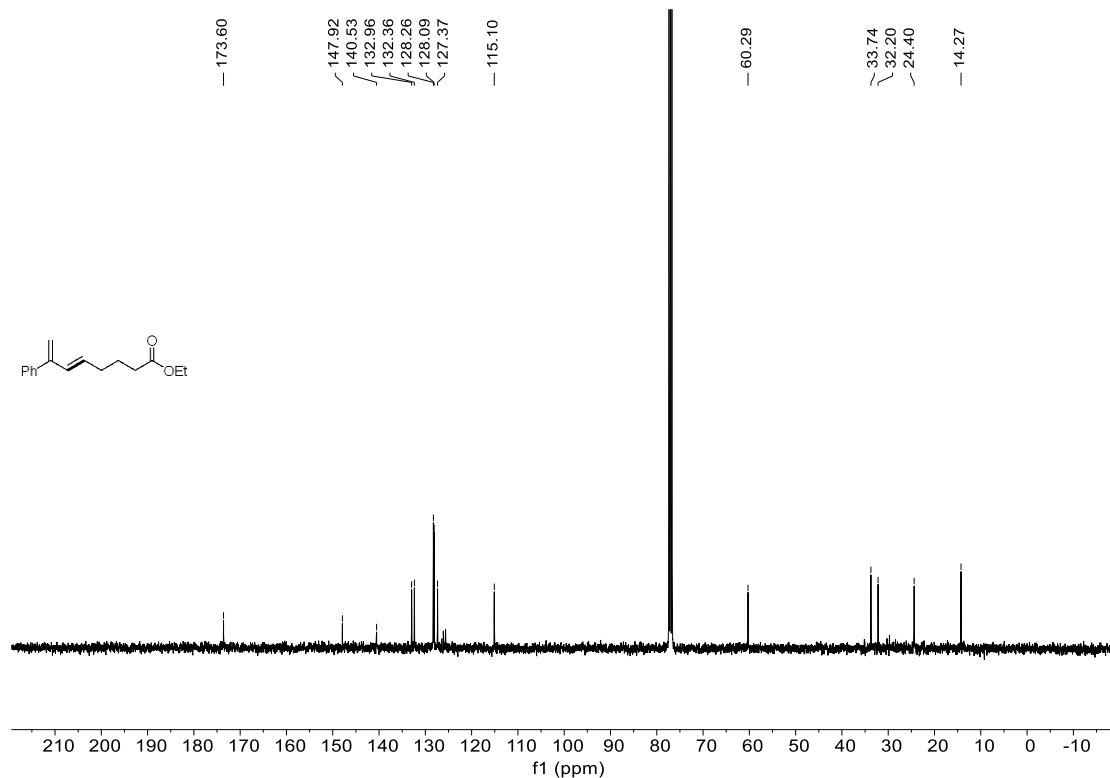
¹H NMR (400 MHz, CDCl₃) spectra of **2b**



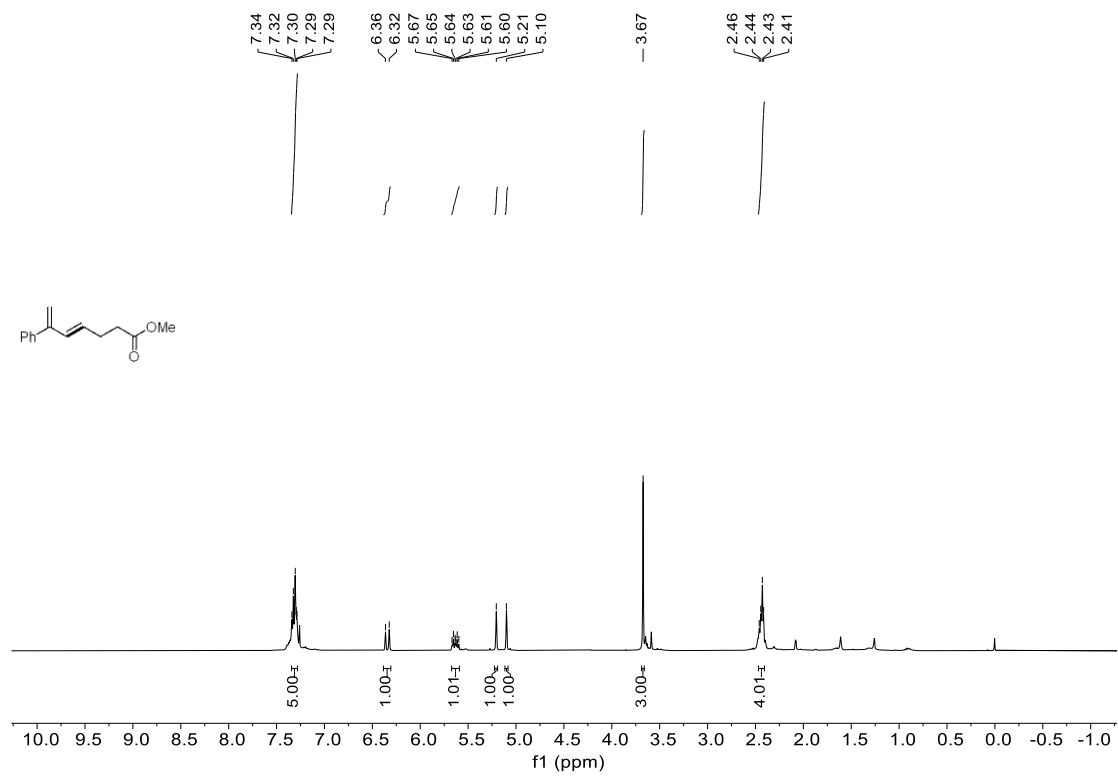
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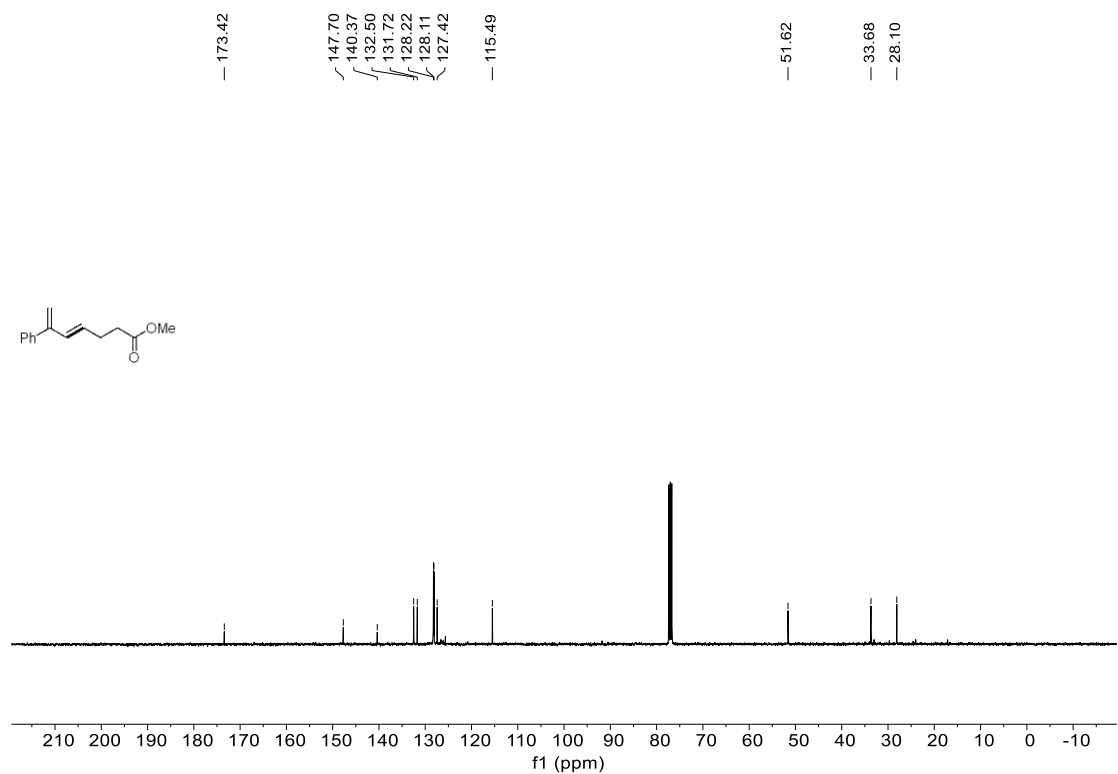
^1H NMR (400 MHz, CDCl_3) spectra of **3b**



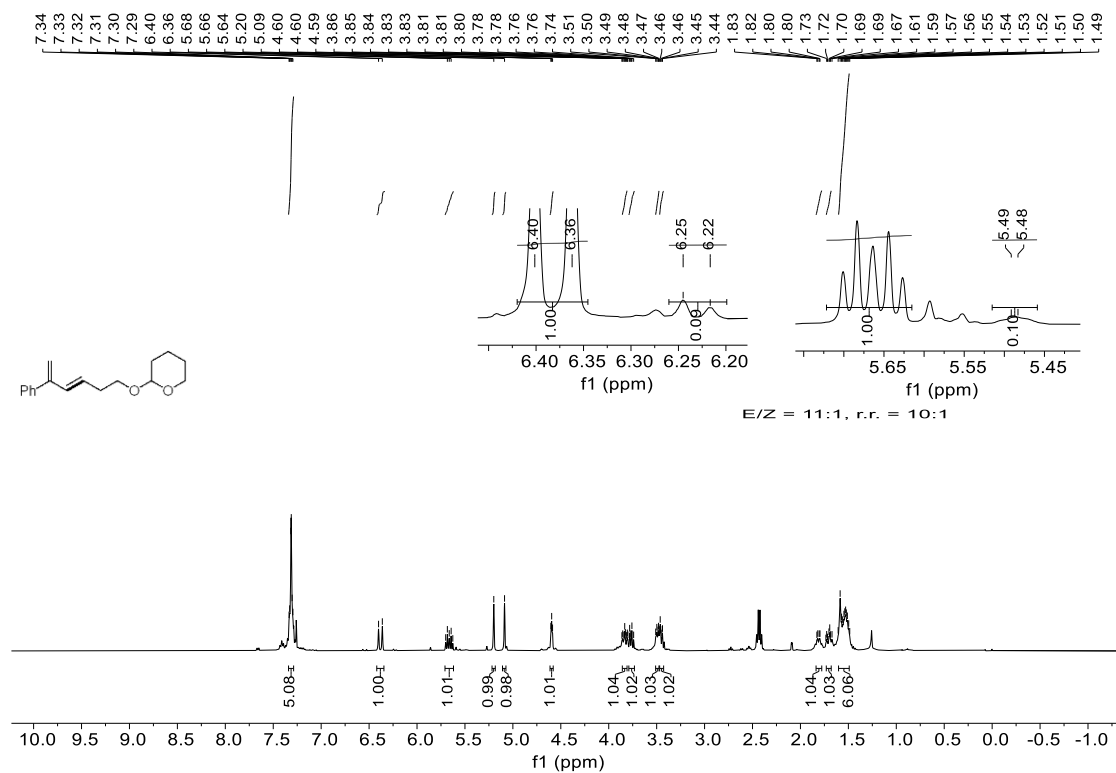
^{13}C NMR (101 MHz, CDCl_3) spectra of **3b**



¹H NMR (400 MHz, CDCl₃) spectra of **4b**

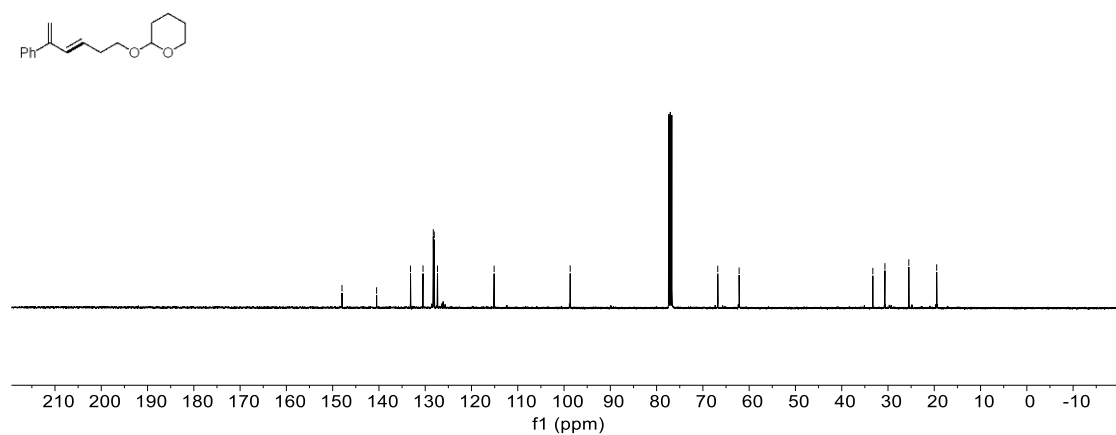


¹³C NMR (101 MHz, CDCl₃) spectra of **4b**

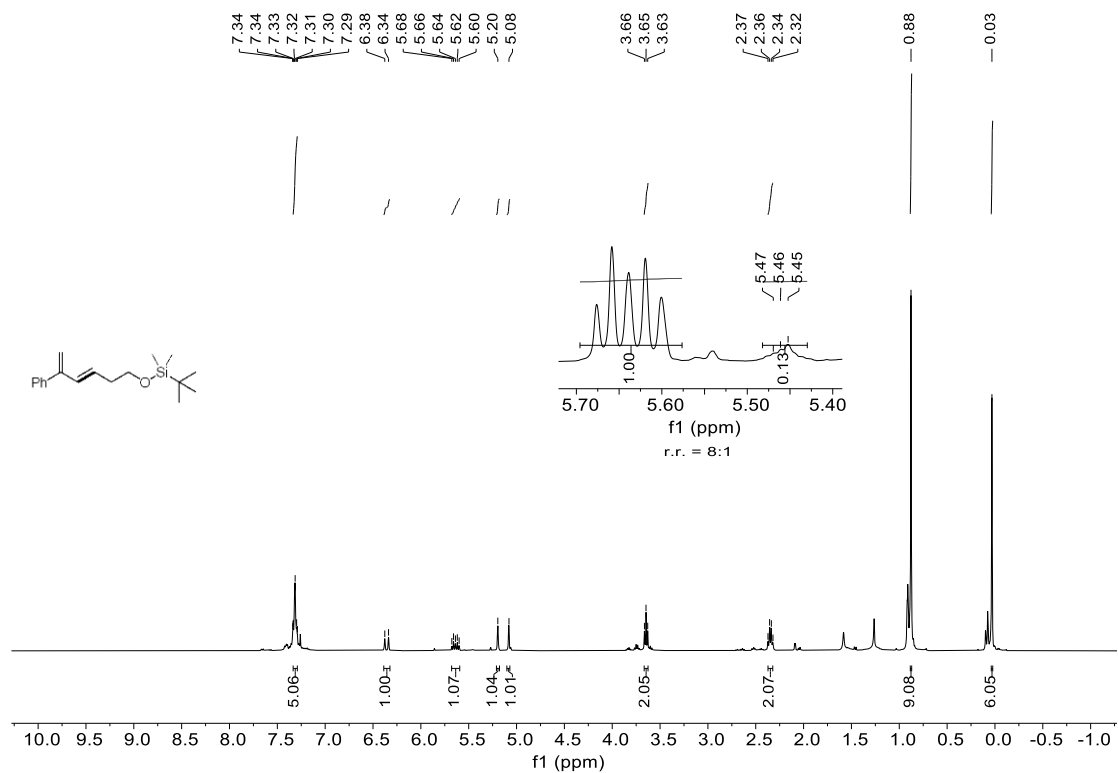


^1H NMR (400 MHz, CDCl_3) spectra of **5b**

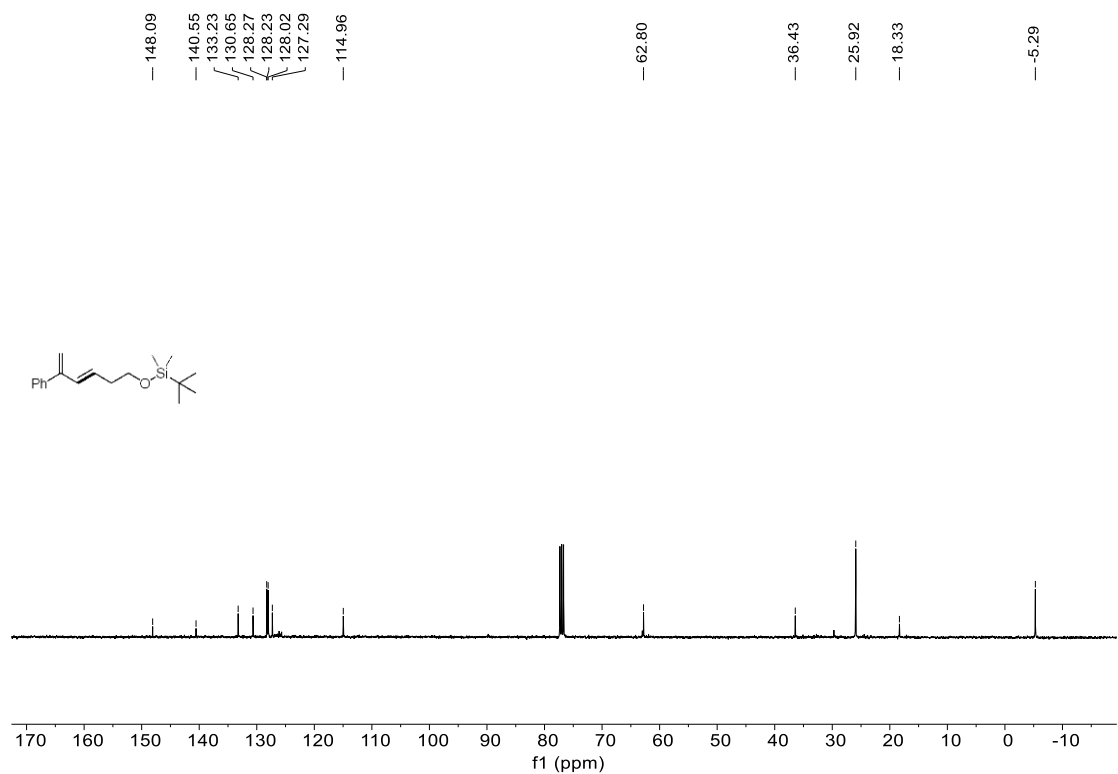
147.98, 140.50, 133.17, 130.48, 128.25, 128.04, 127.34, 115.11, 98.67, 66.79, 62.17, 33.28, 30.66, 25.50, 19.48



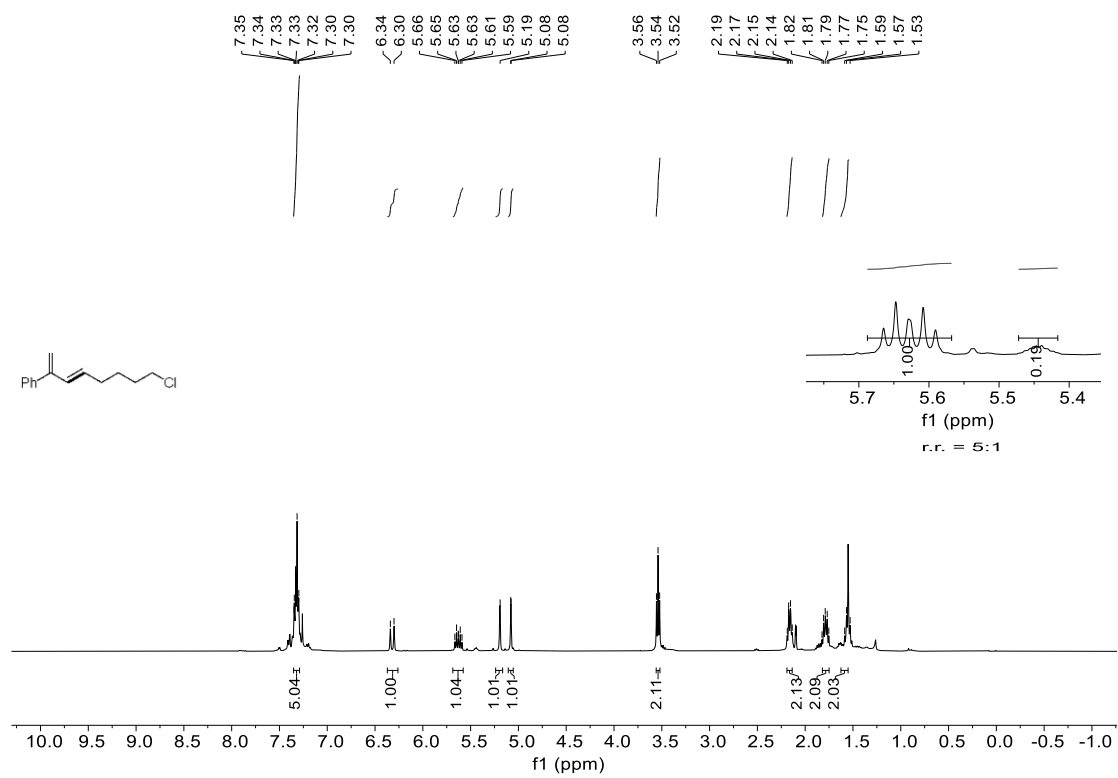
^{13}C NMR (101 MHz, CDCl_3) spectra of **5b**



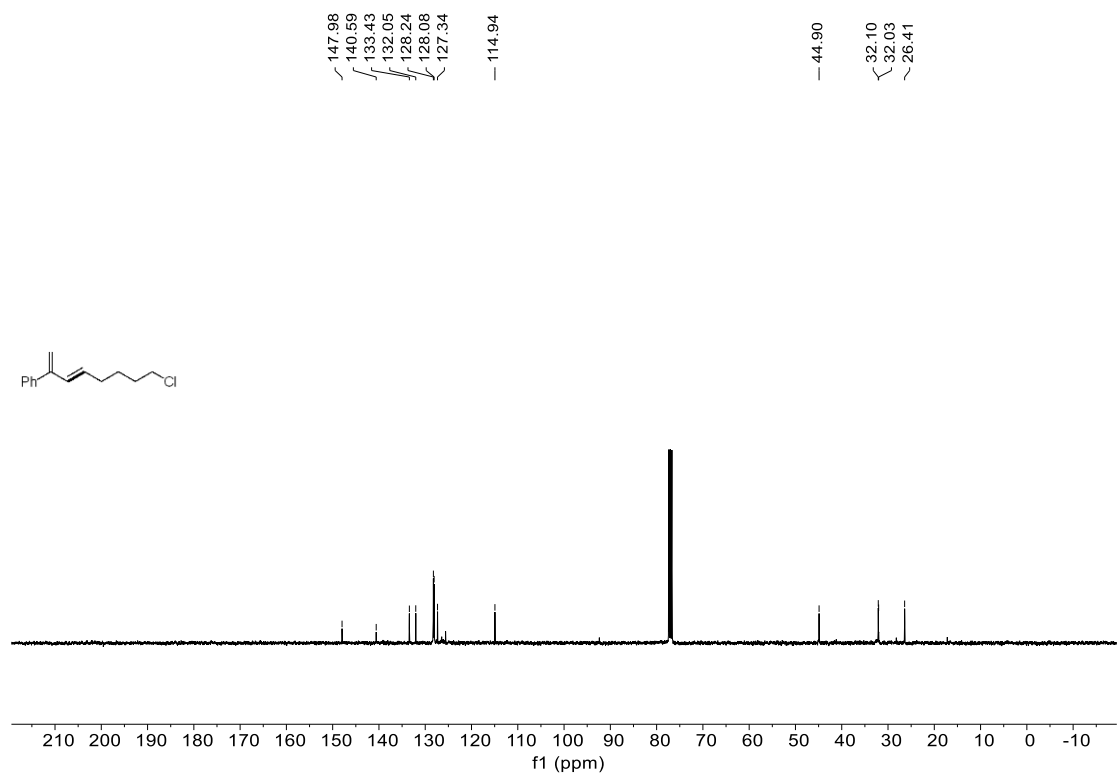
¹H NMR (400 MHz, CDCl₃) spectra of **6b**



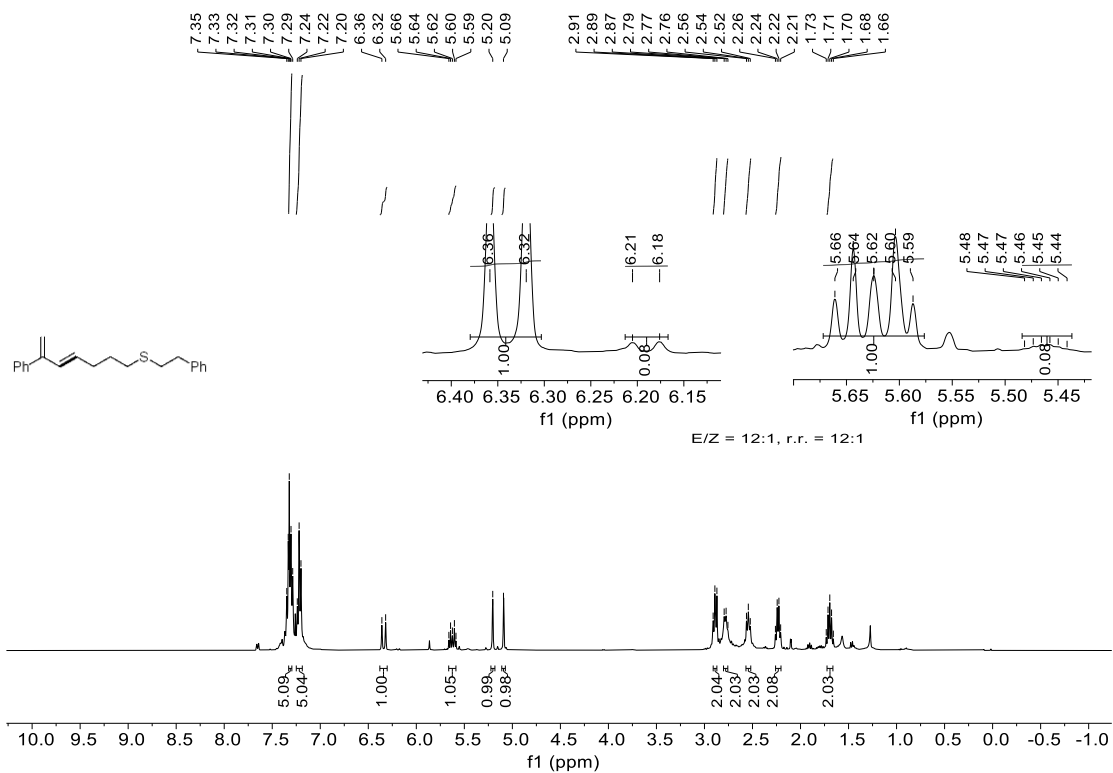
¹³C NMR (101 MHz, CDCl₃) spectra of **6b**



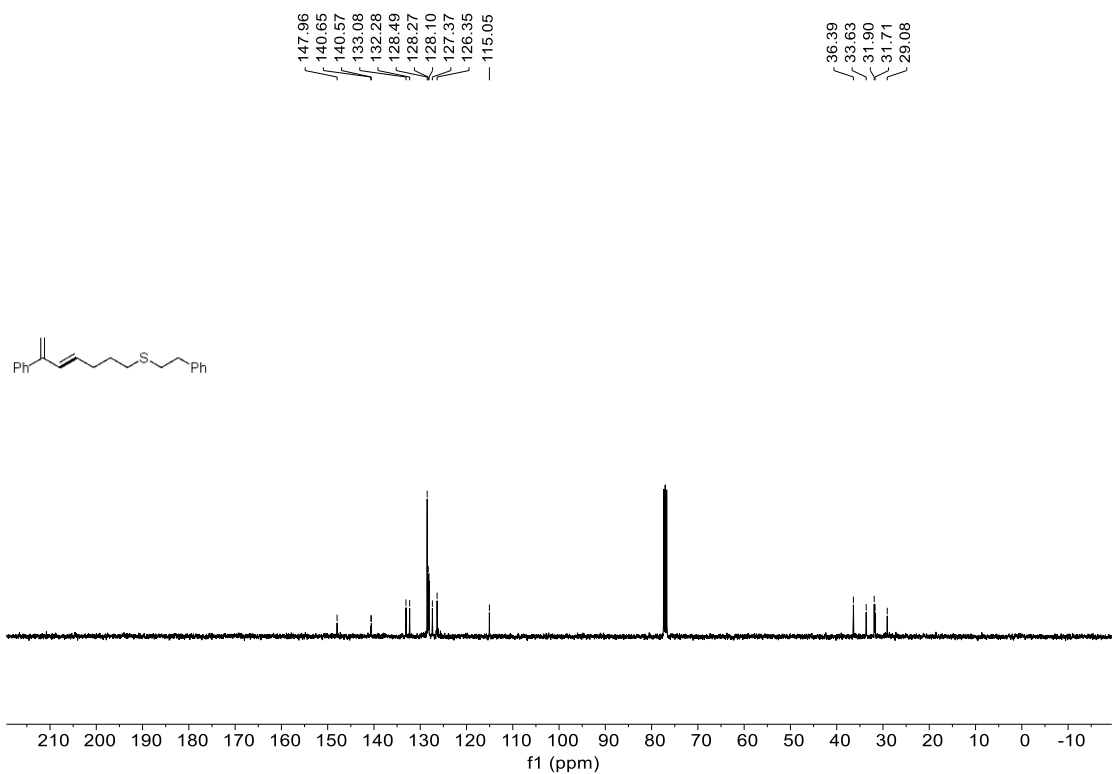
^1H NMR (400 MHz, CDCl_3) spectra of **7b**



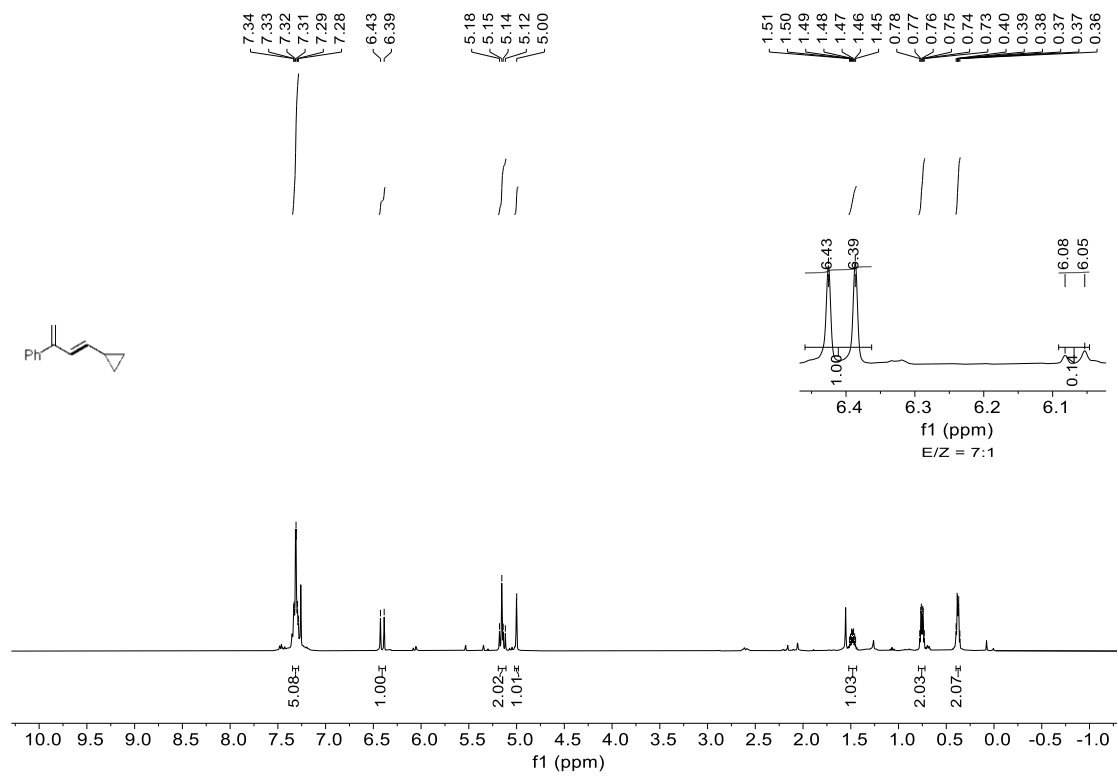
^{13}C NMR (101 MHz, CDCl_3) spectra of **7b**



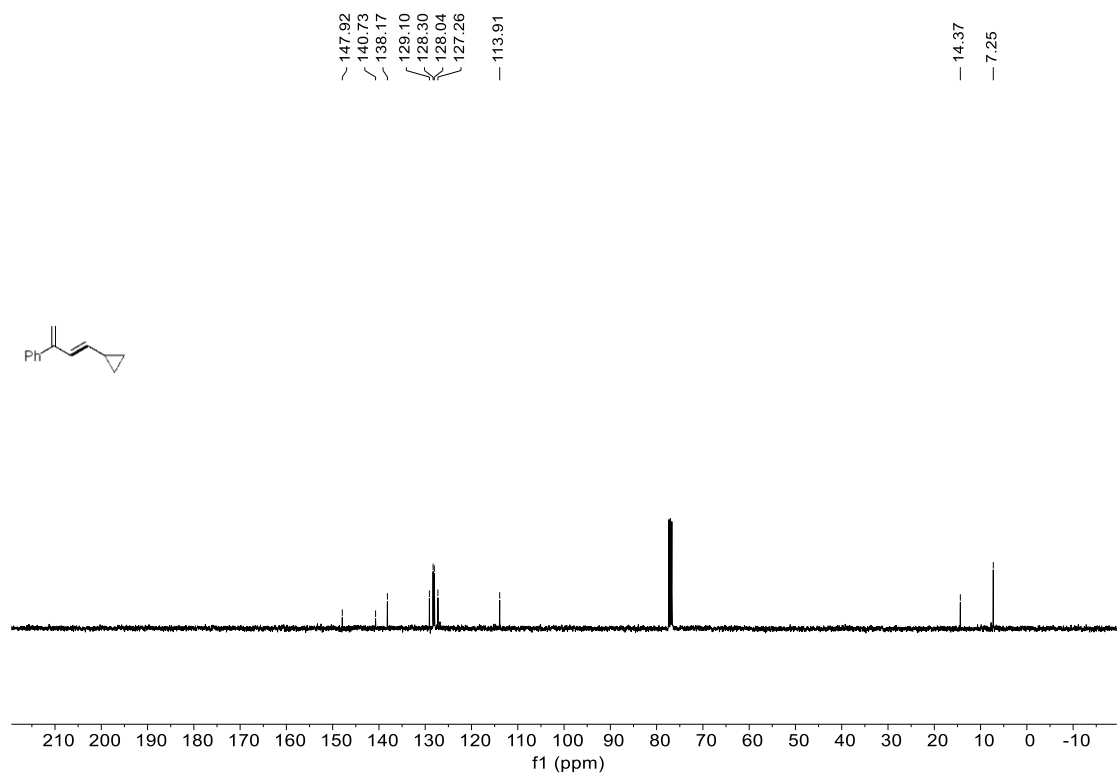
¹H NMR (400 MHz, CDCl₃) spectra of **8b**



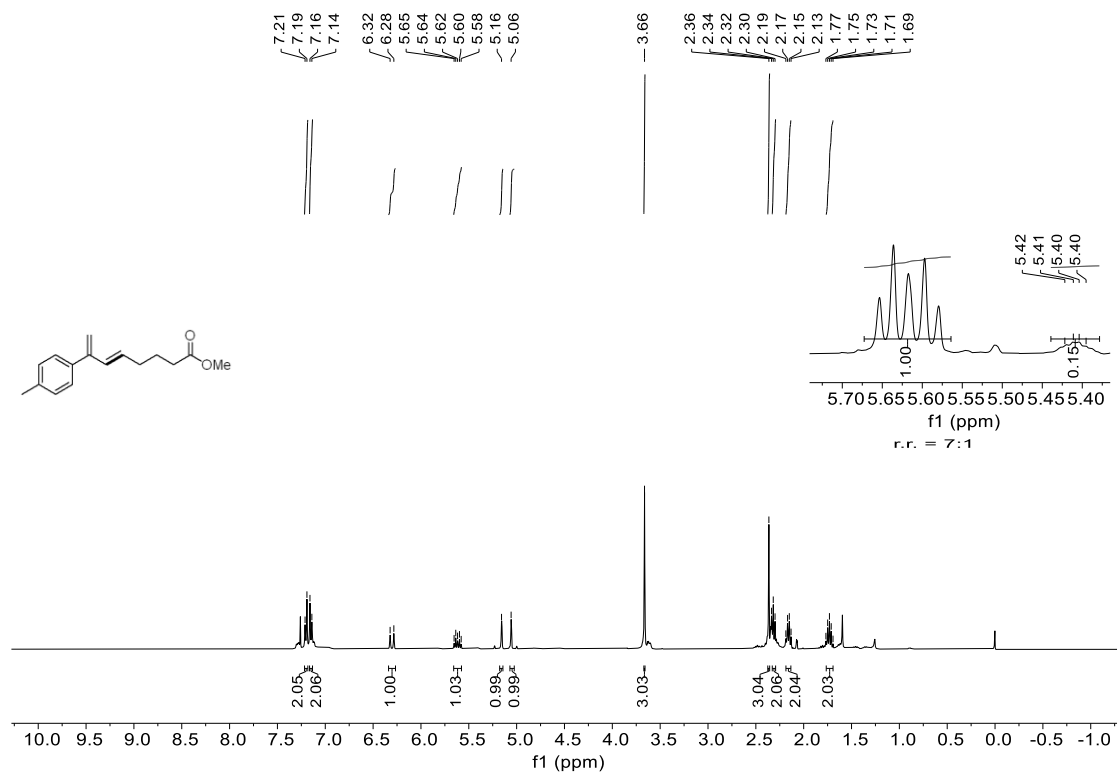
¹³C NMR (101 MHz, CDCl₃) spectra of **8b**



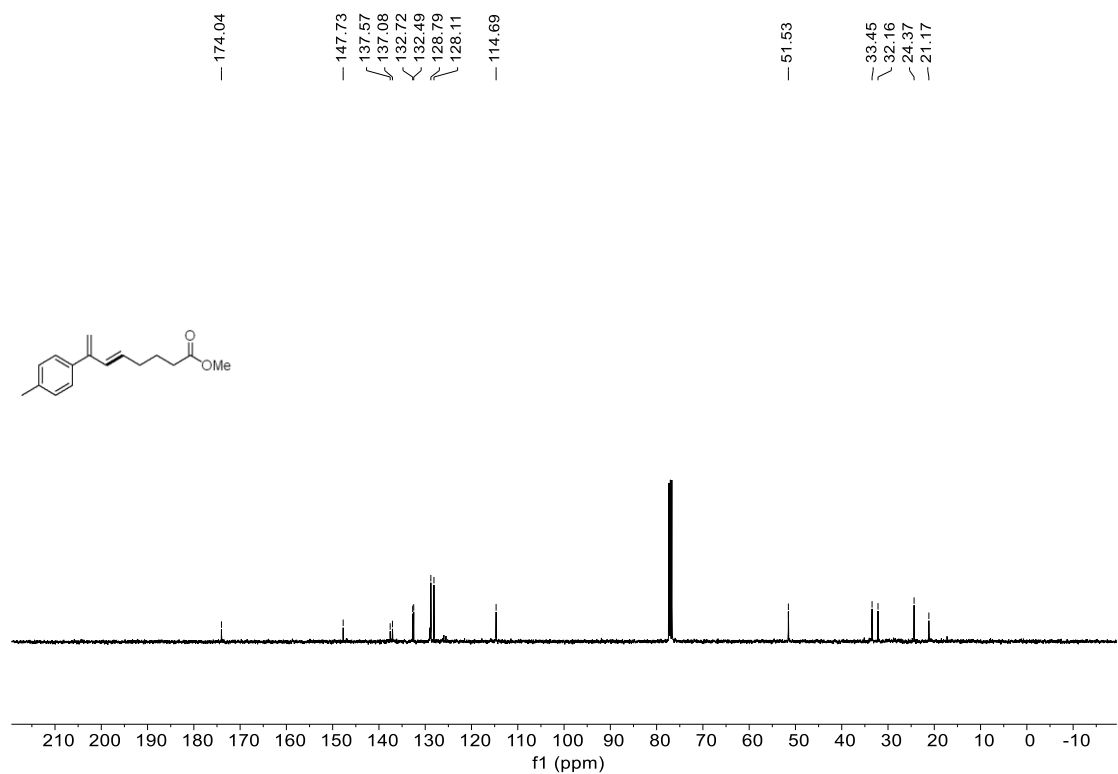
^1H NMR (400 MHz, CDCl_3) spectra of **9b**



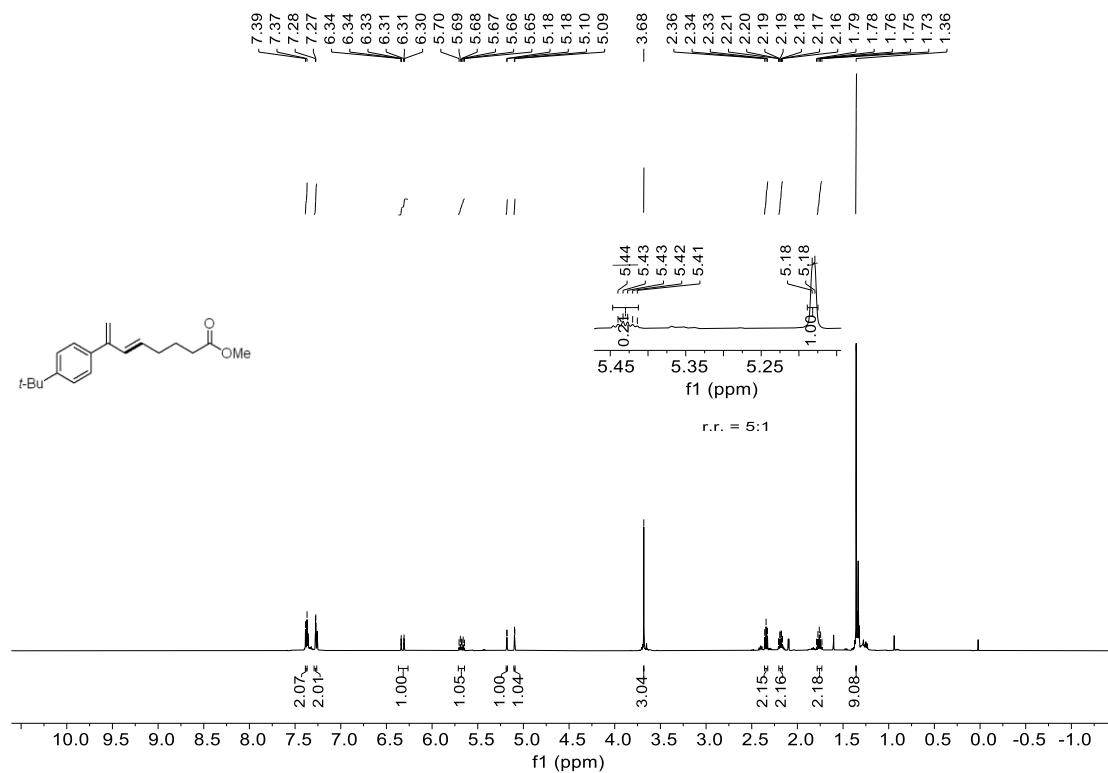
^{13}C NMR (101 MHz, CDCl_3) spectra of **9b**



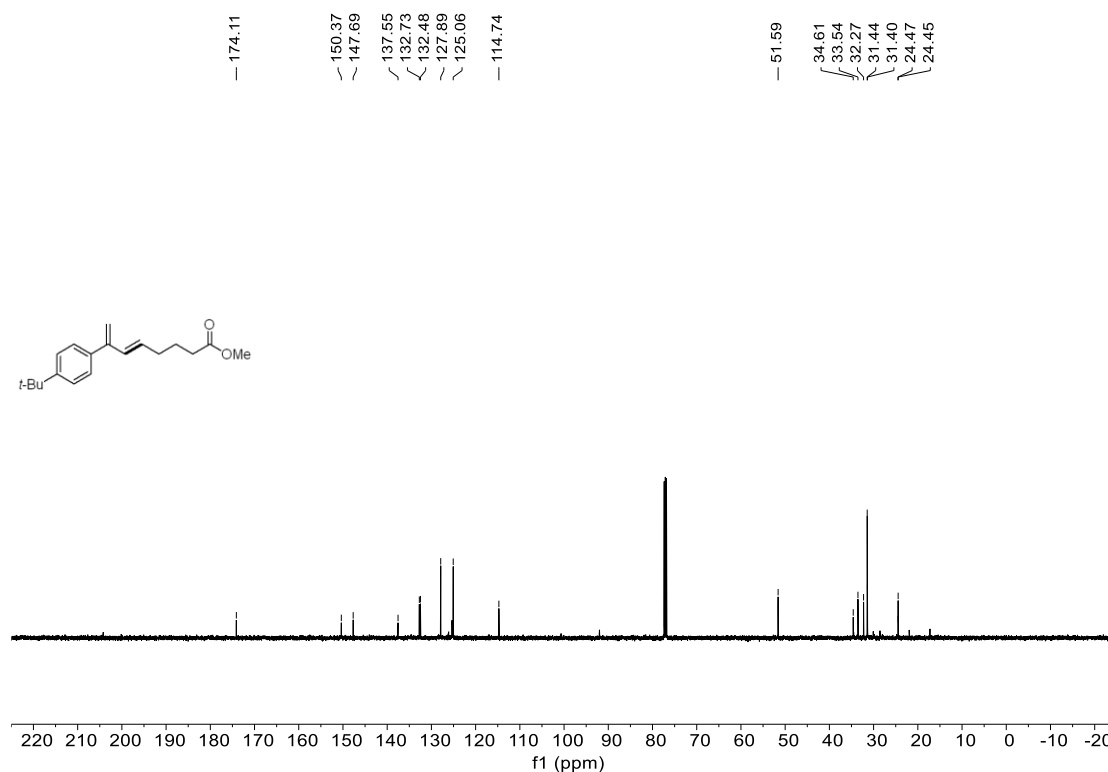
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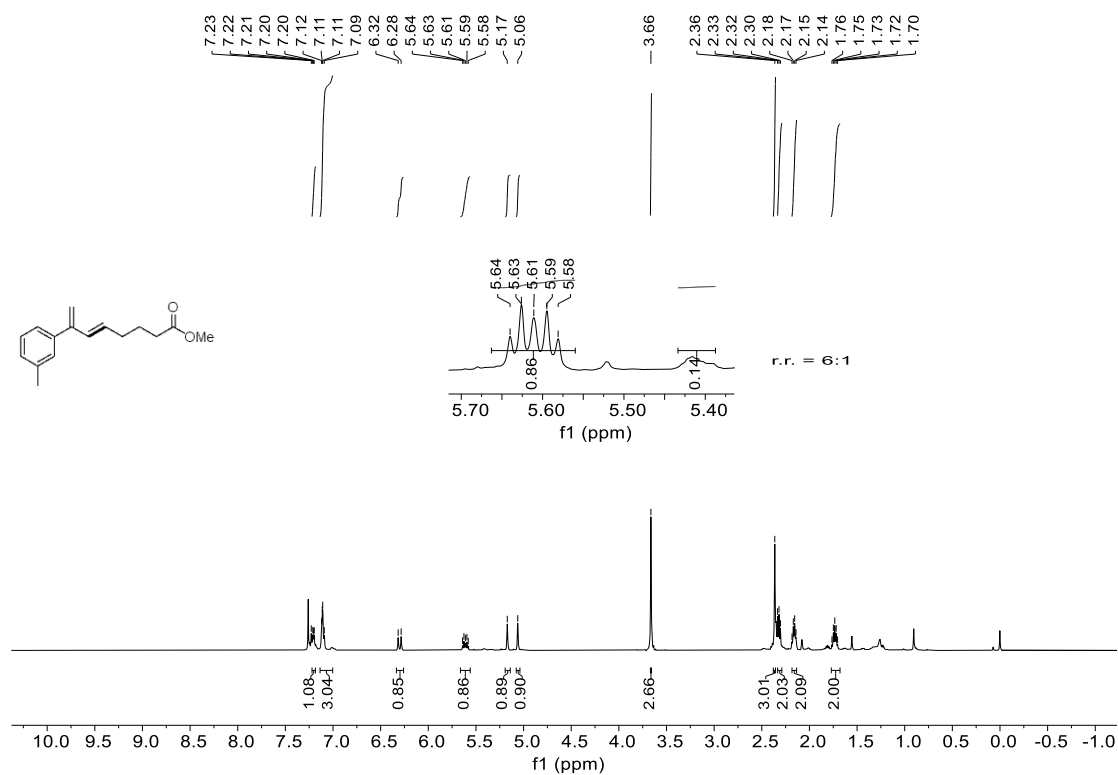
¹³C NMR (101 MHz, CDCl₃) spectra of **10b**



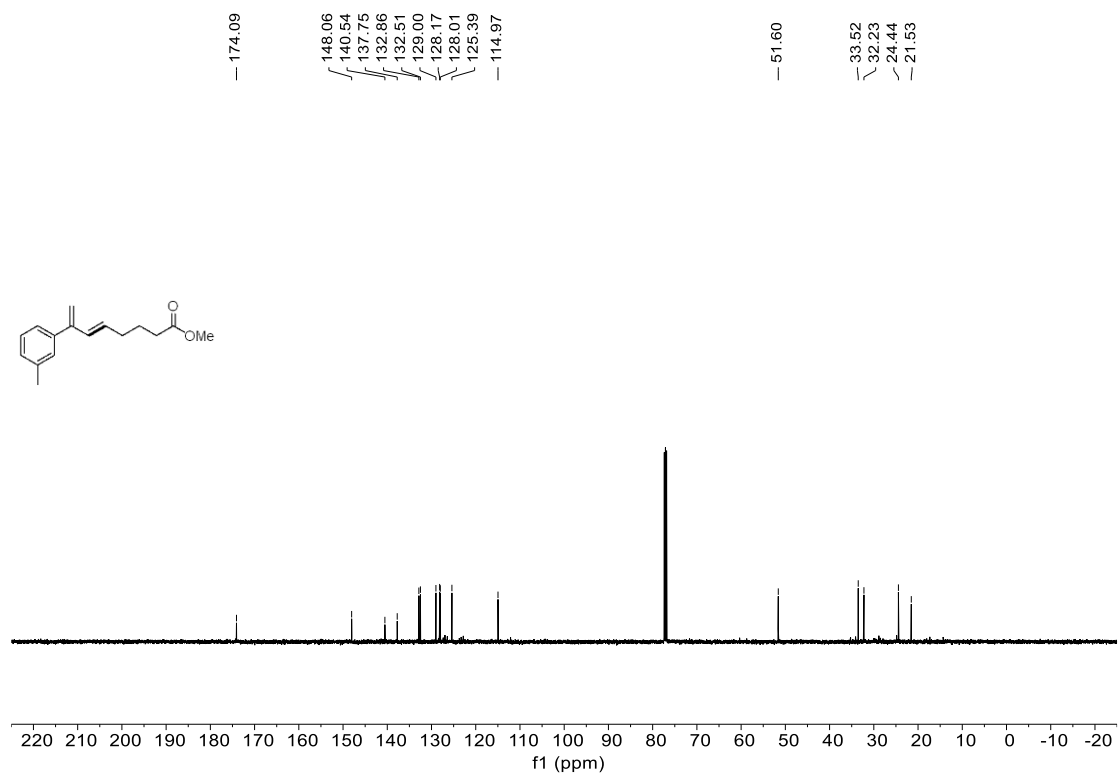
¹H NMR (500 MHz, CDCl₃) spectra of **11b**



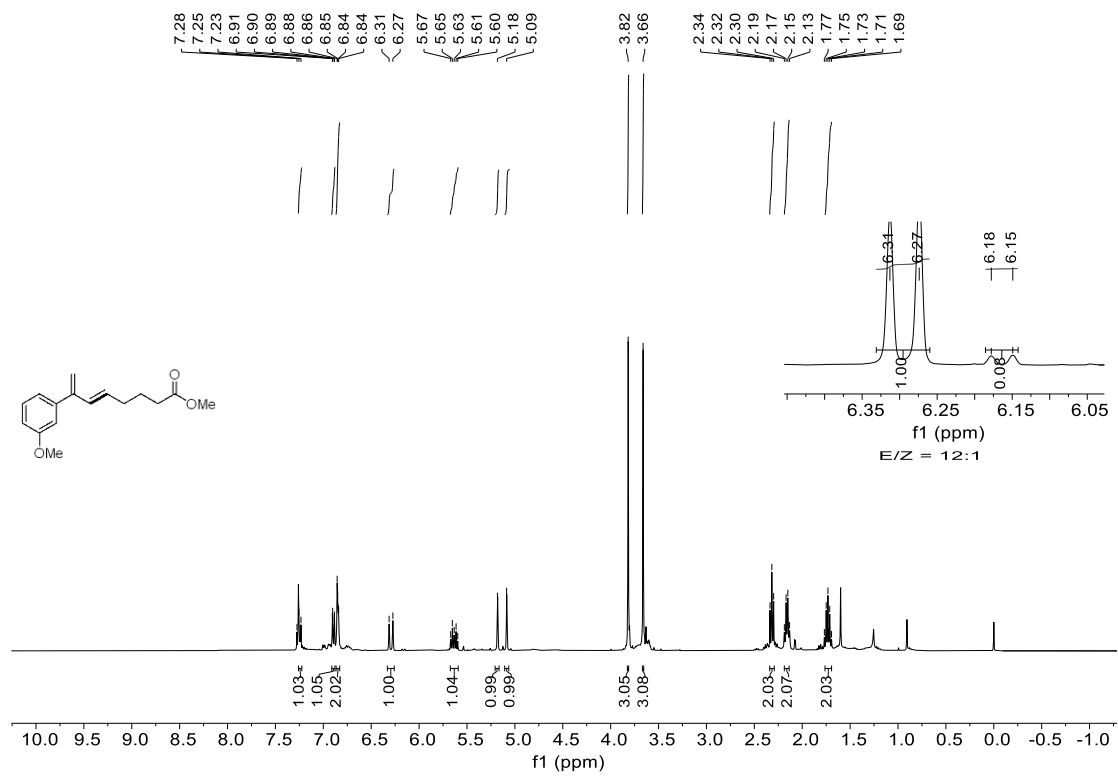
¹³C NMR (126 MHz, CDCl₃) spectra of **11b**



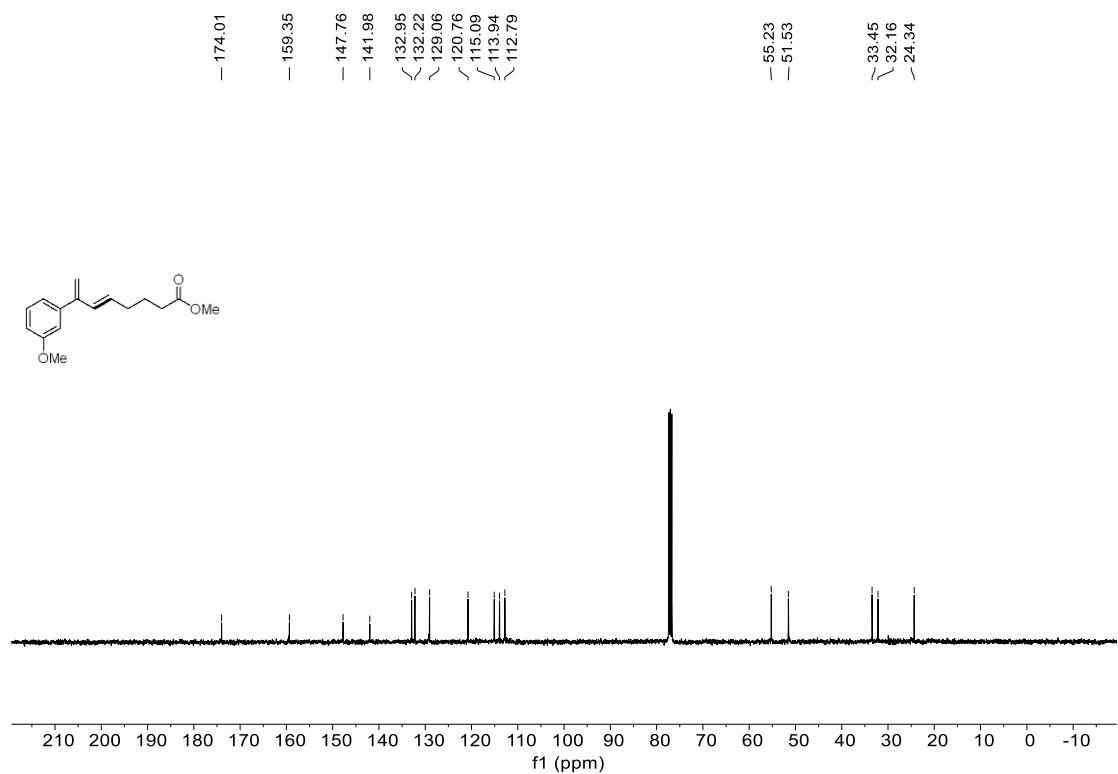
¹H NMR (500 MHz, CDCl₃) spectra of **12b**



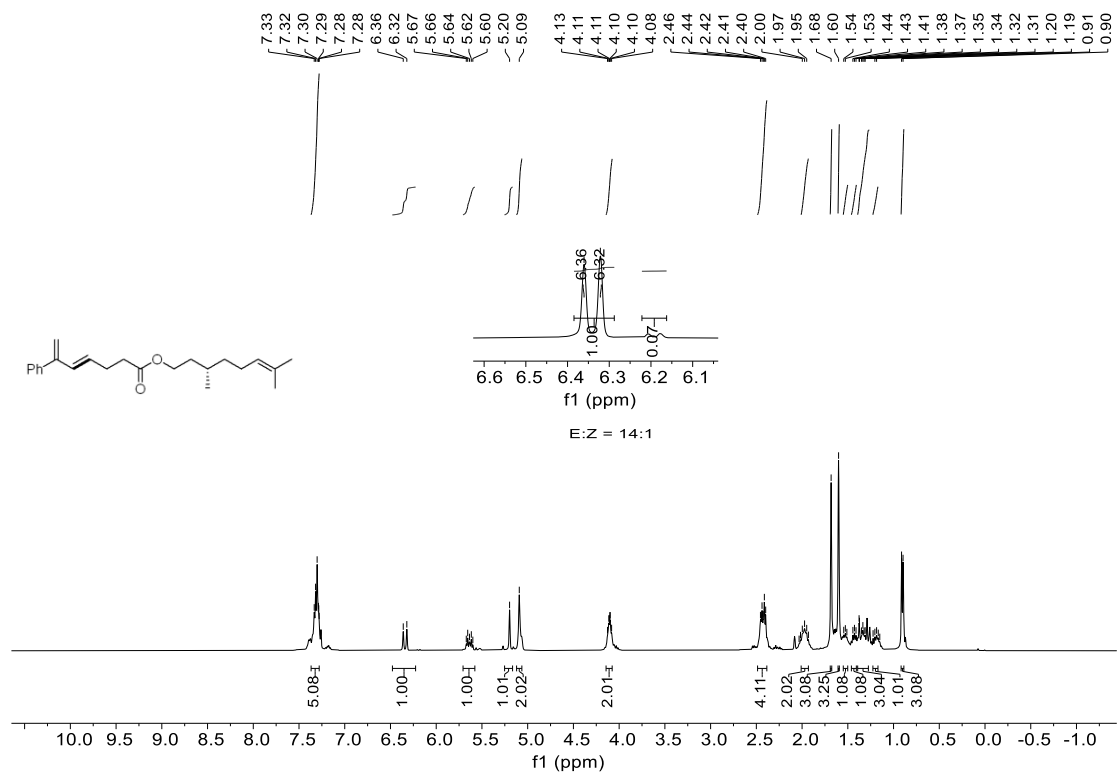
¹³C NMR (126 MHz, CDCl₃) spectra of **12b**



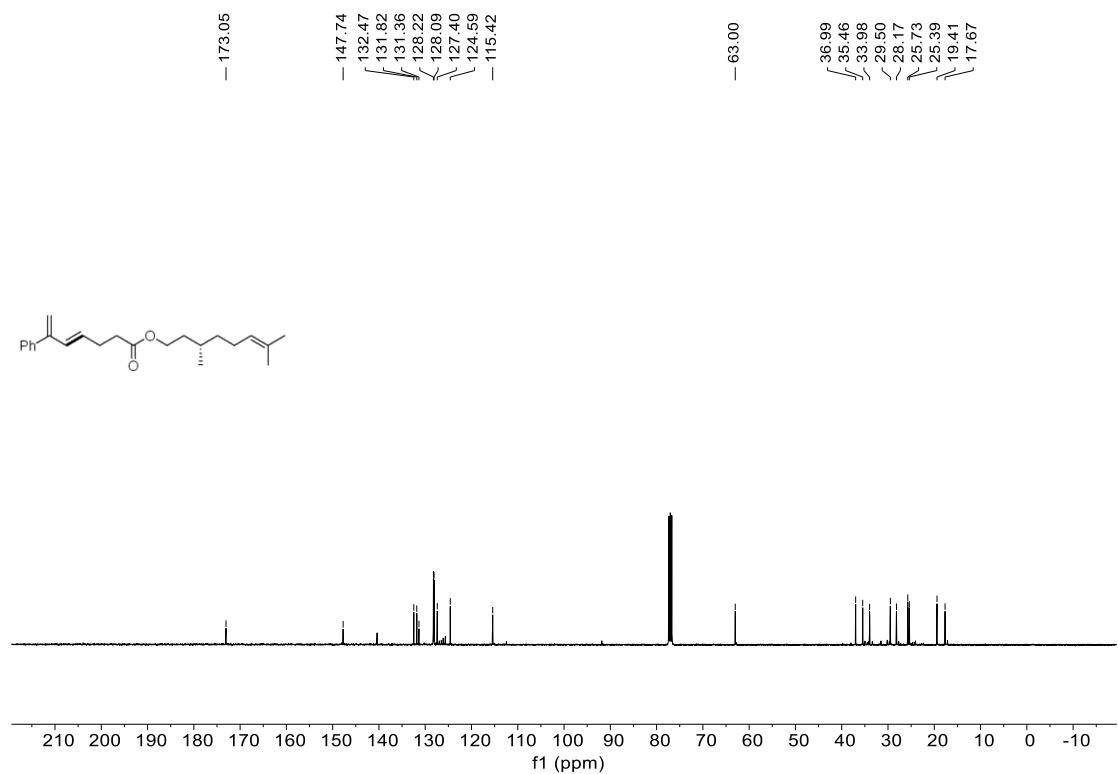
¹H NMR (400 MHz, CDCl₃) spectra of **13b**



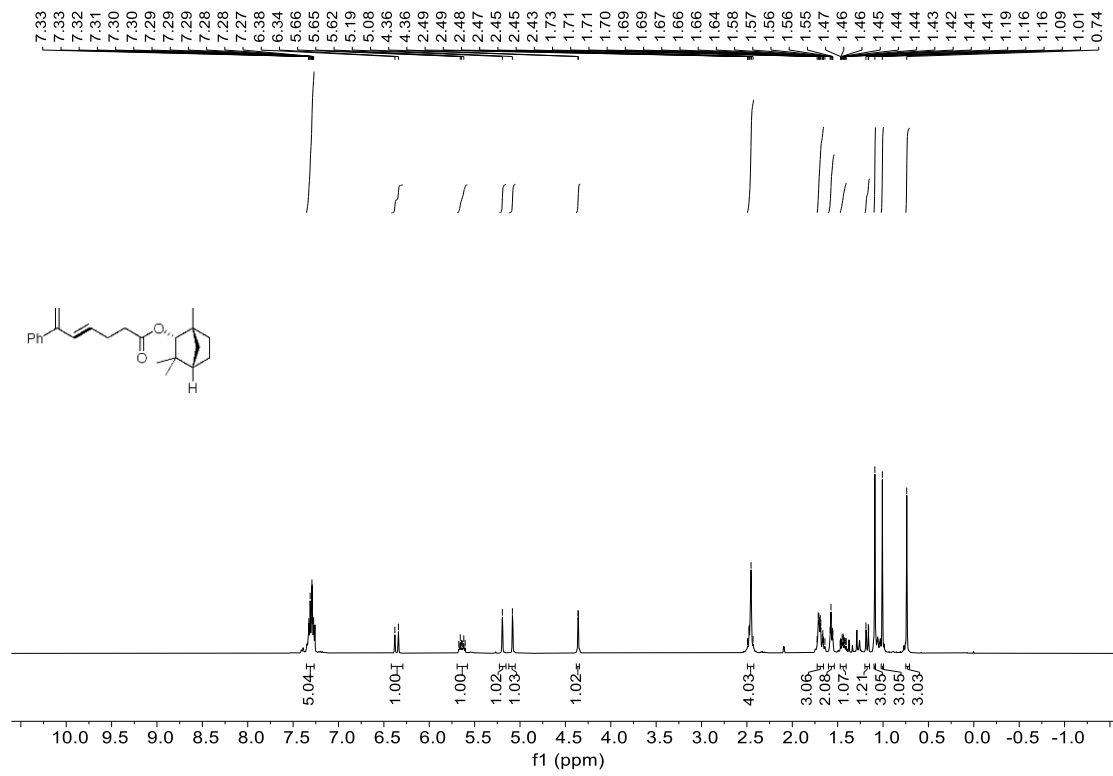
¹³C NMR (101 MHz, CDCl₃) spectra of **13b**



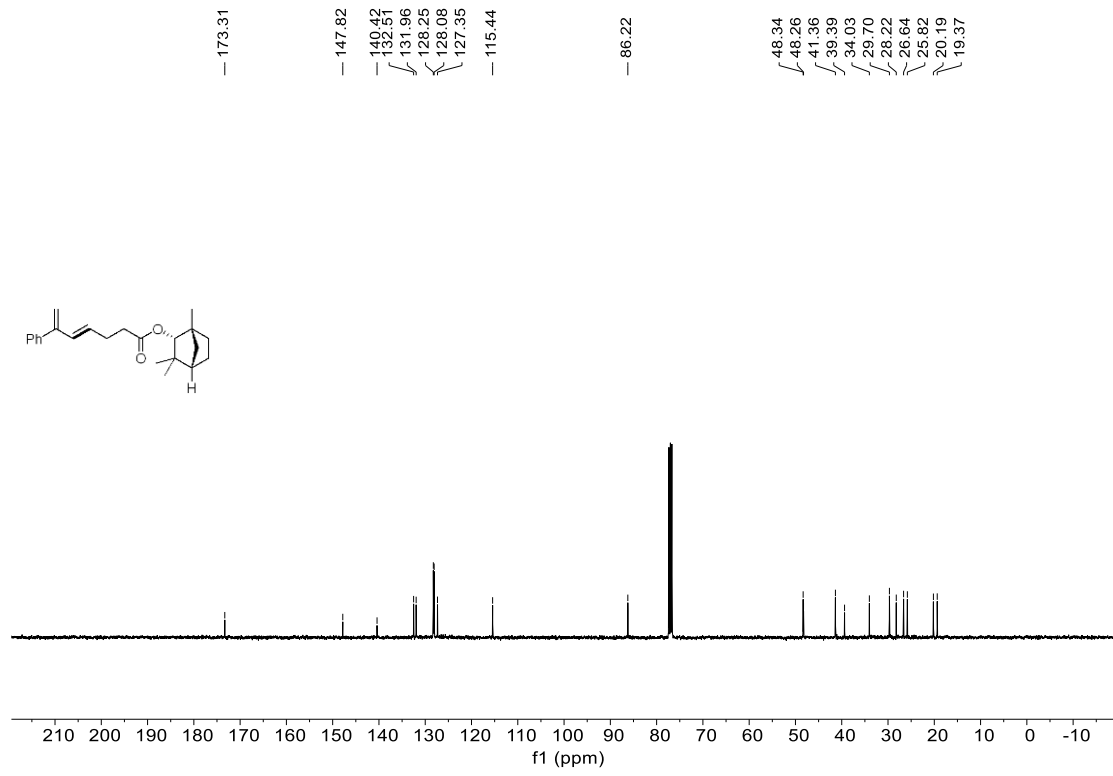
¹H NMR (400 MHz, CDCl₃) spectra of **14b**



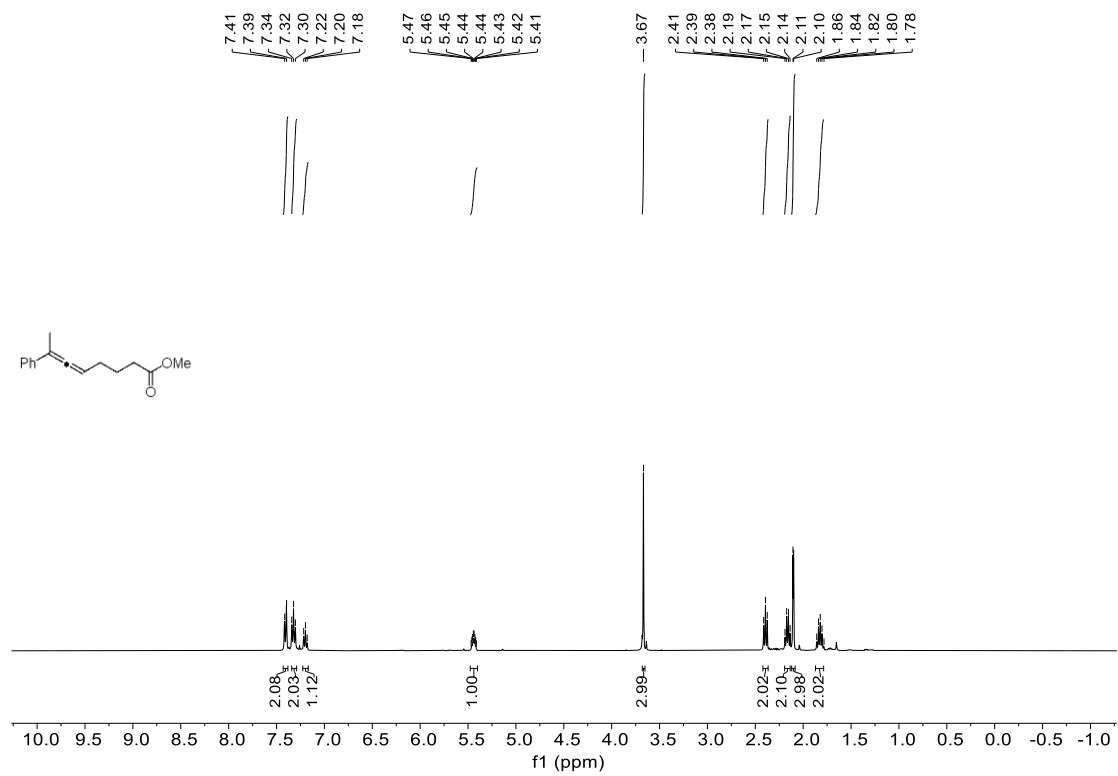
¹³C NMR (101 MHz, CDCl₃) spectra of **14b**



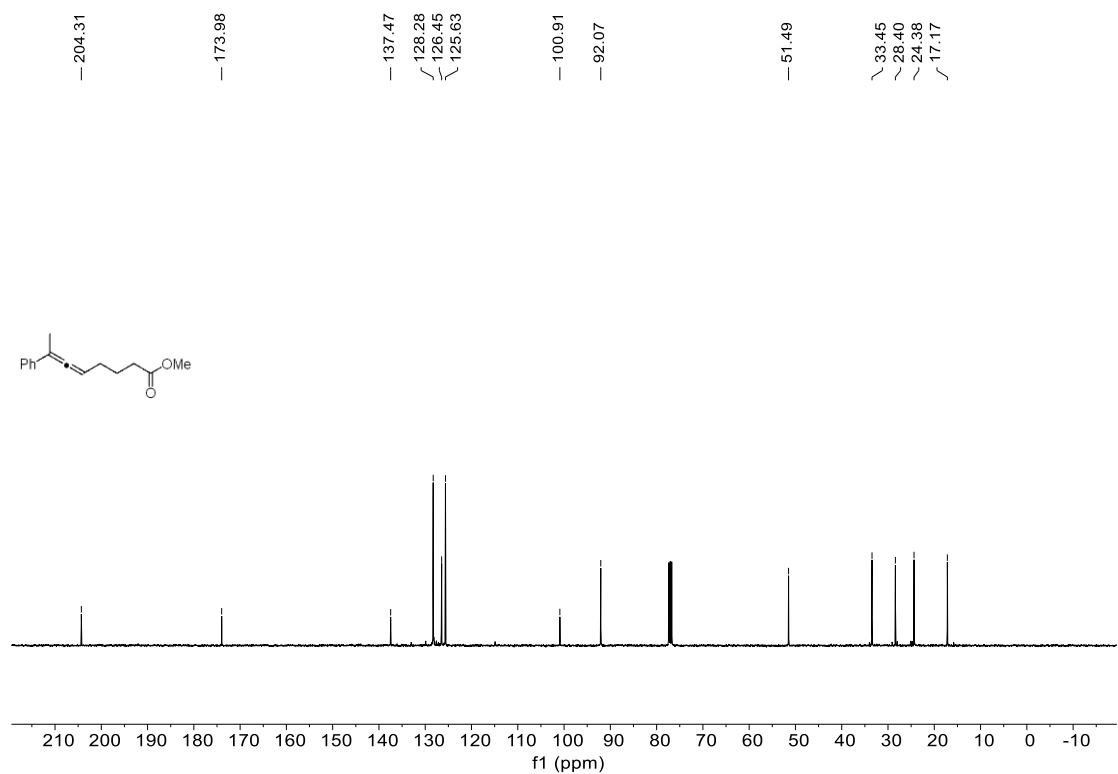
¹H NMR (400 MHz, CDCl₃) spectra of **15b**



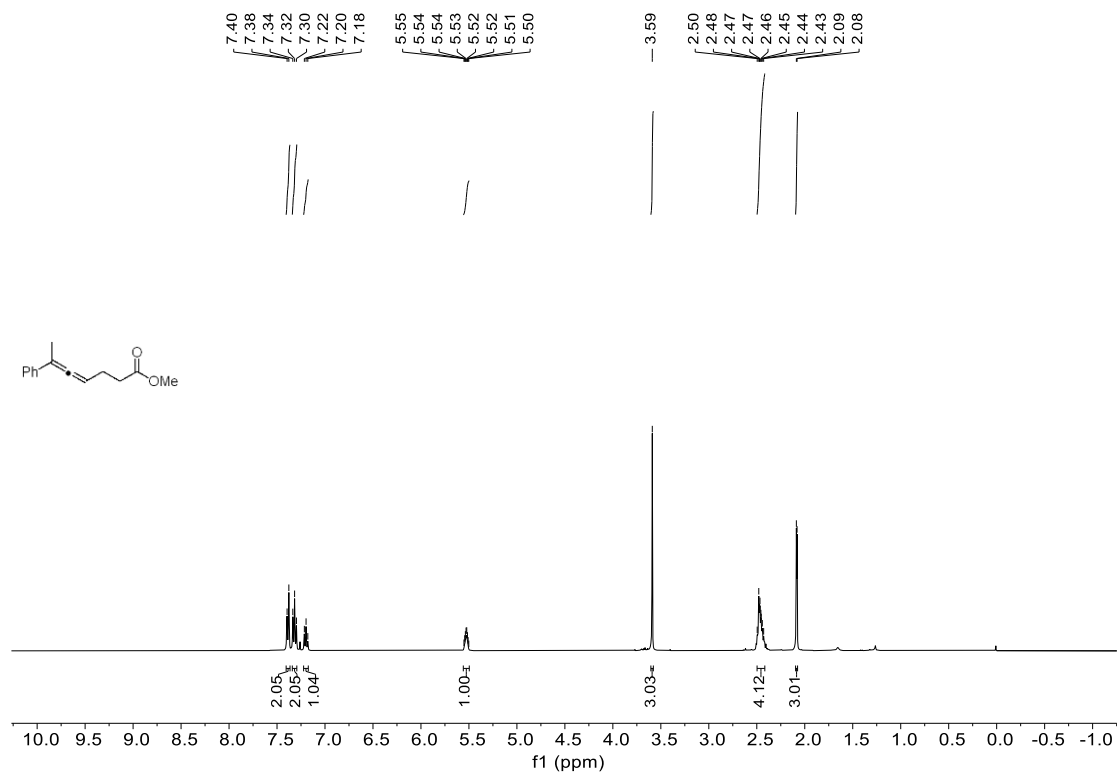
¹³C NMR (101 MHz, CDCl₃) spectra of **15b**



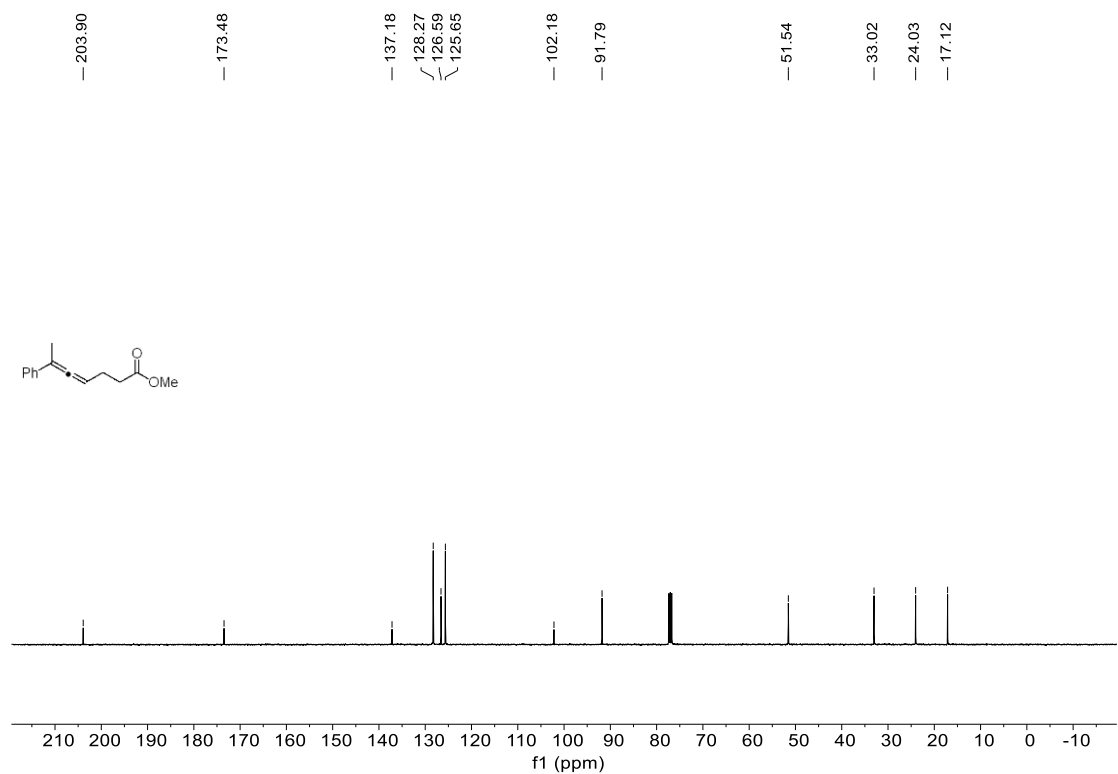
¹H NMR (400 MHz, CDCl₃) spectra of **2c**



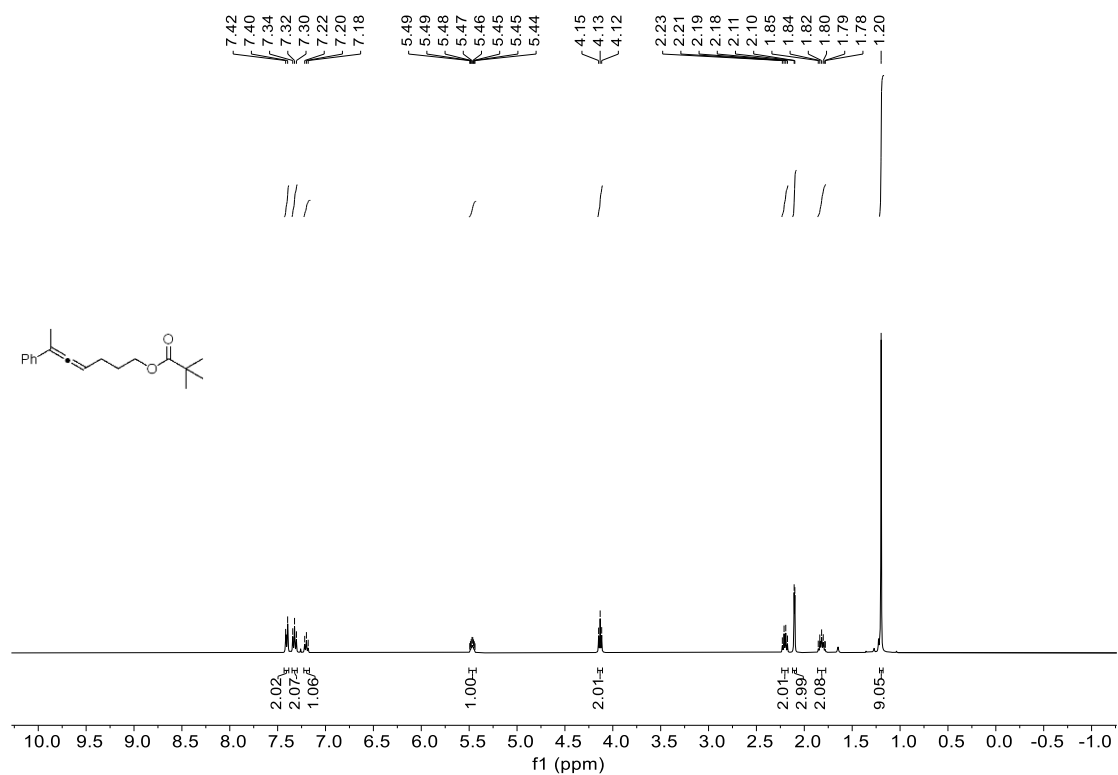
¹³C NMR (101 MHz, CDCl₃) spectra of **2c**



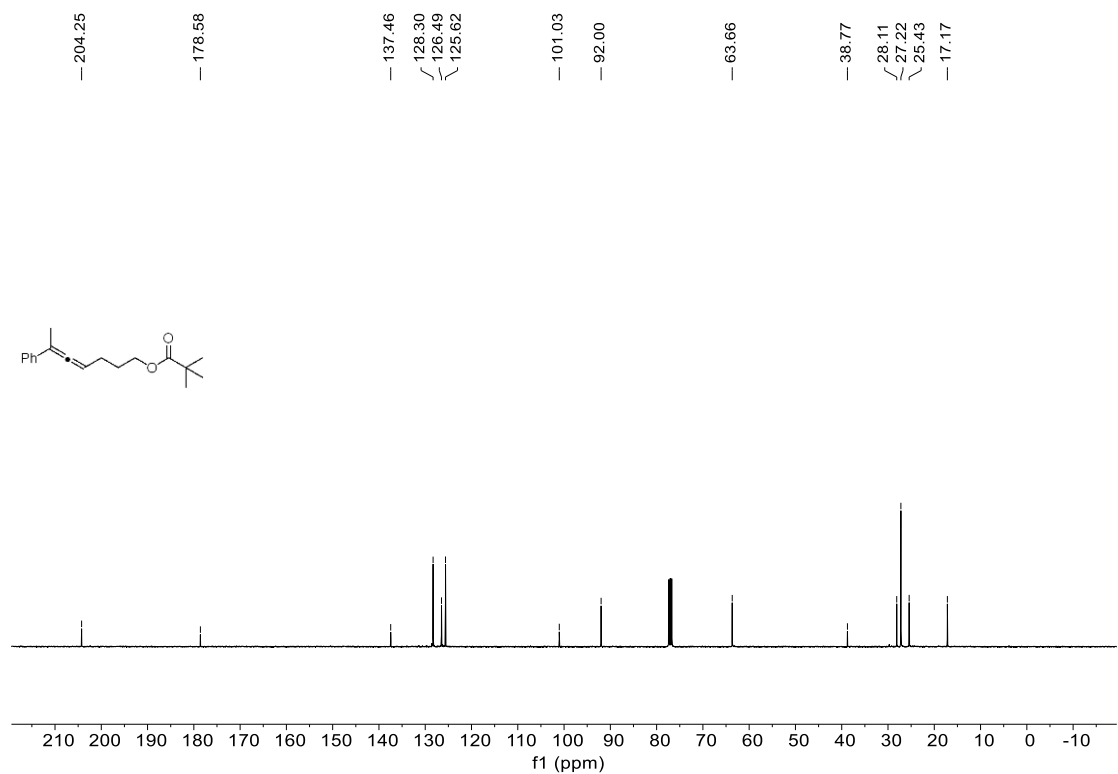
¹H NMR (400 MHz, CDCl₃) spectra of **3c**



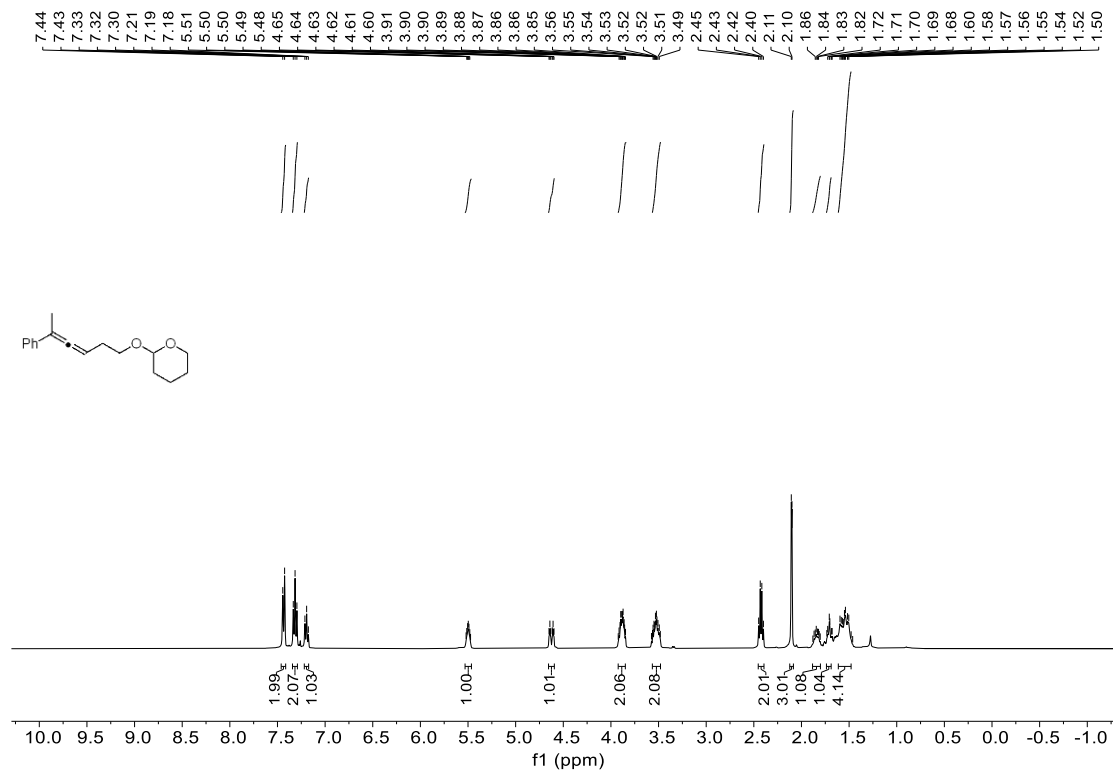
¹³C NMR (101 MHz, CDCl₃) spectra of **3c**



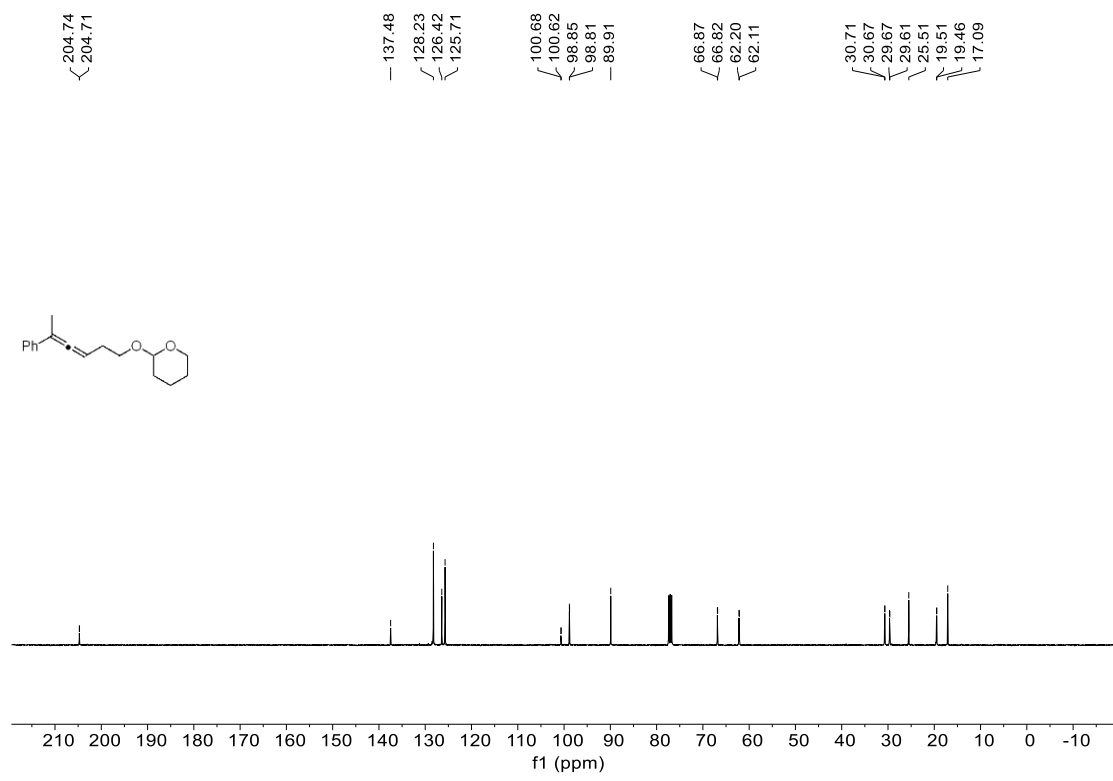
¹H NMR (400 MHz, CDCl₃) spectra of 4c



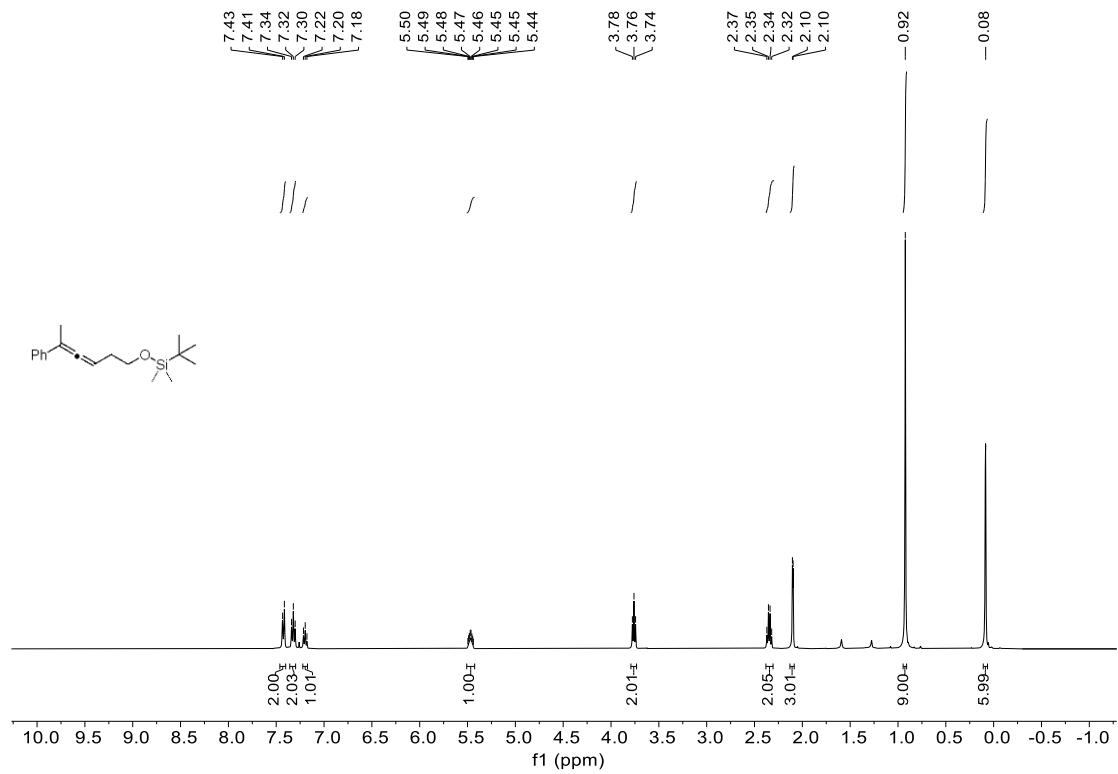
¹³C NMR (101 MHz, CDCl₃) spectra of 4c



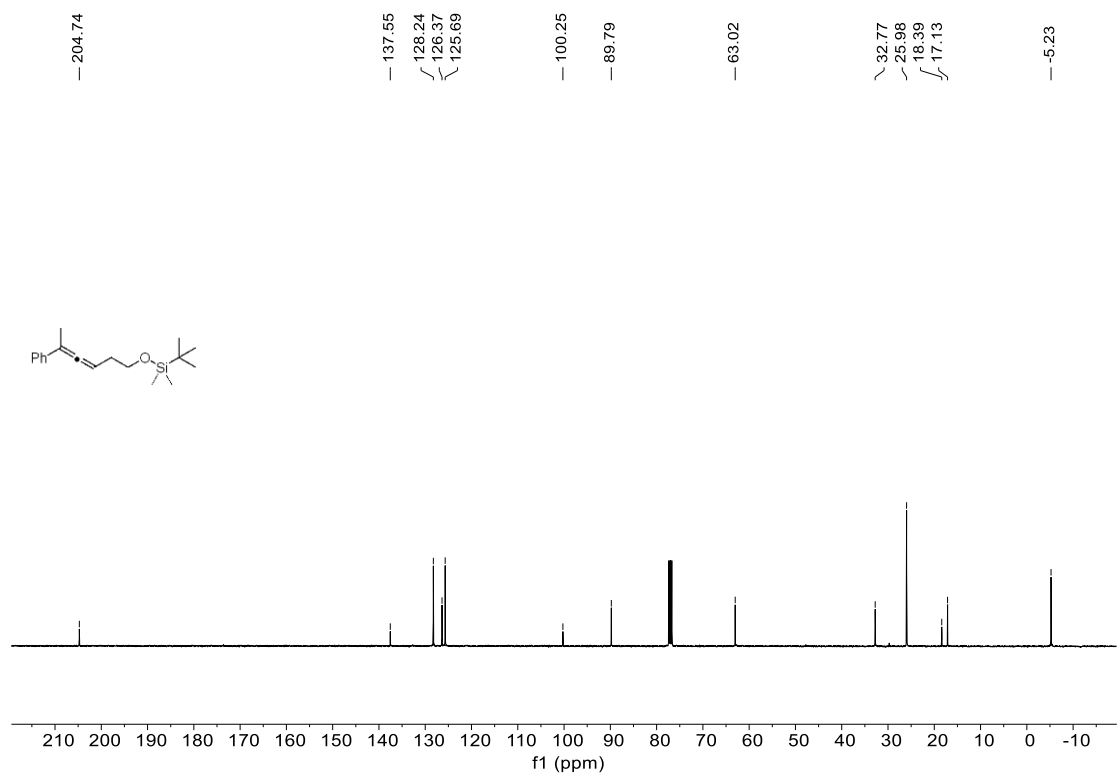
¹H NMR (400 MHz, CDCl₃) spectra of 5c



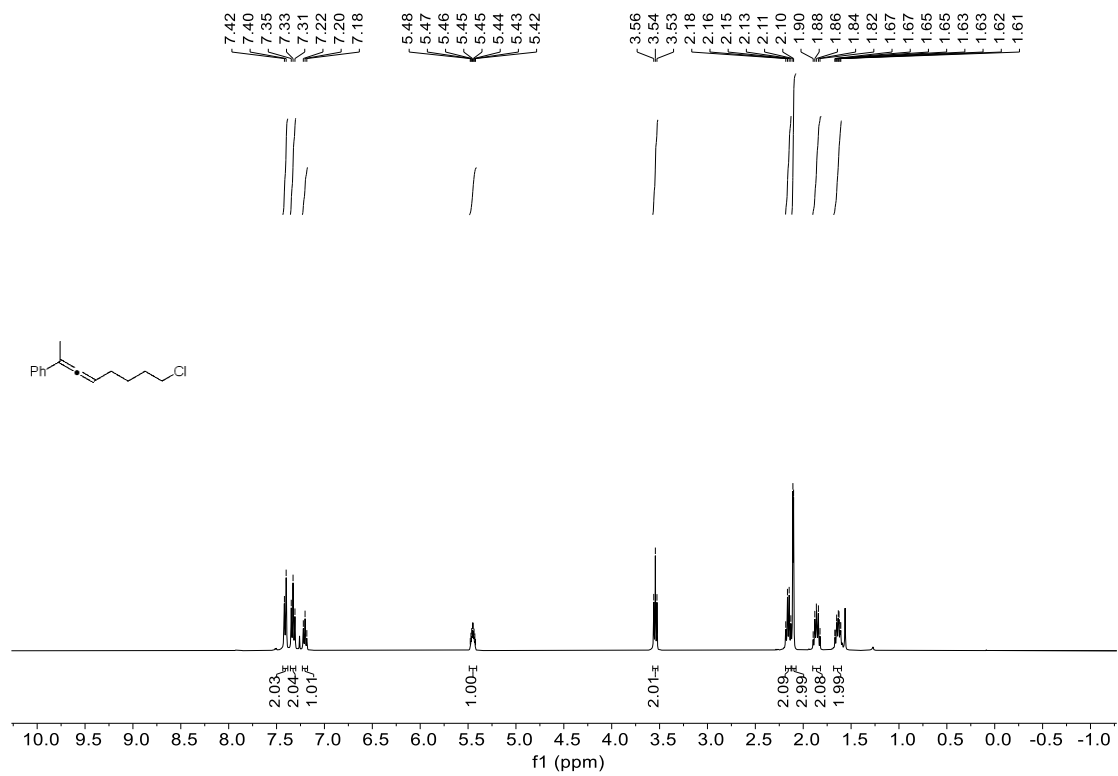
¹³C NMR (101 MHz, CDCl₃) spectra of 5c



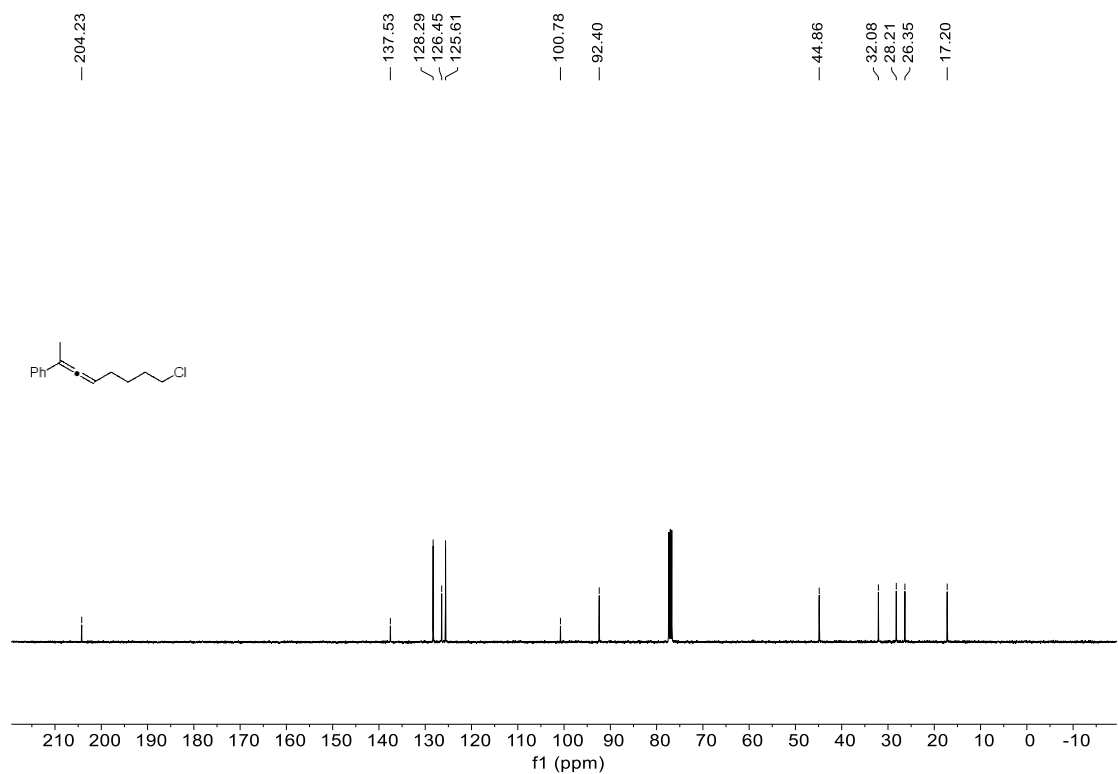
¹H NMR (400 MHz, CDCl₃) spectra of **6c**



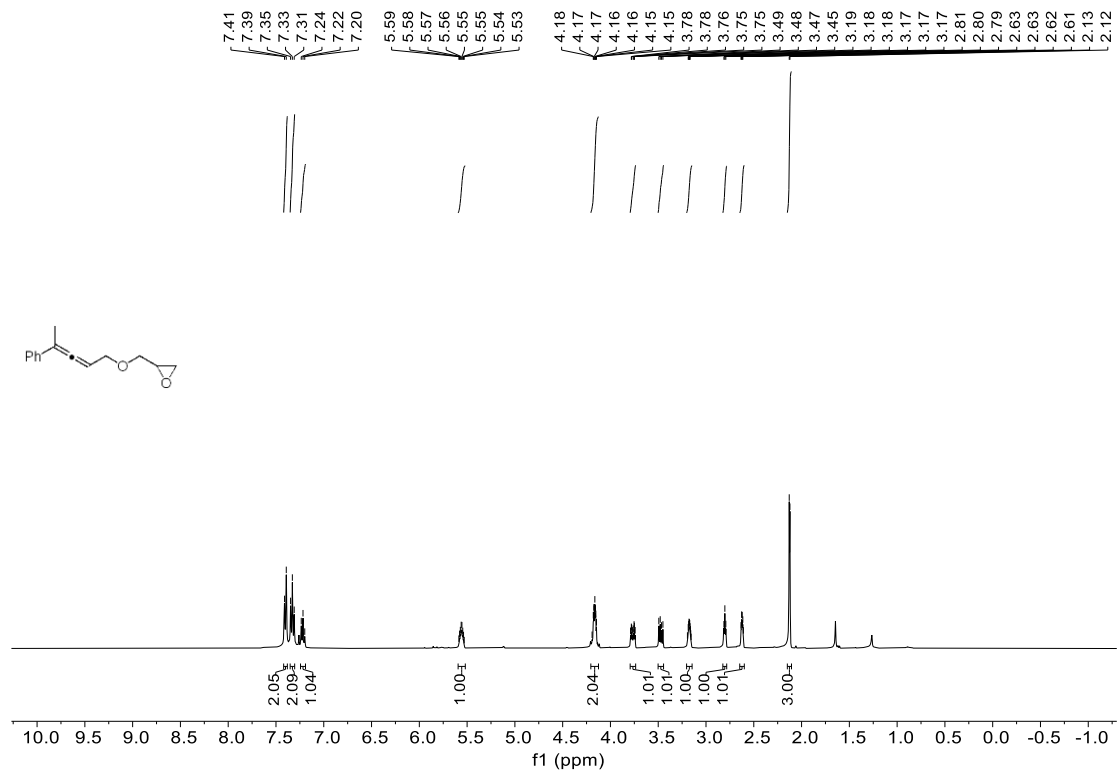
¹³C NMR (101 MHz, CDCl₃) spectra of **6c**



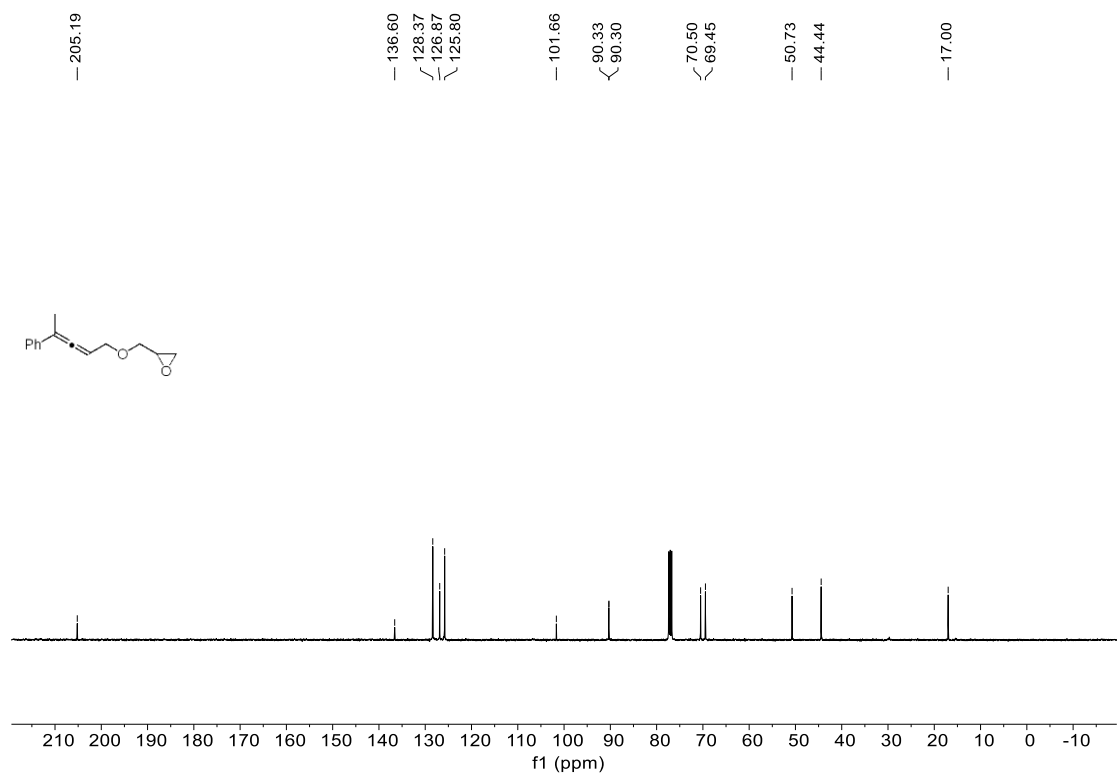
$^1\text{H NMR}$ (400 MHz, CDCl_3) spectra of 7c



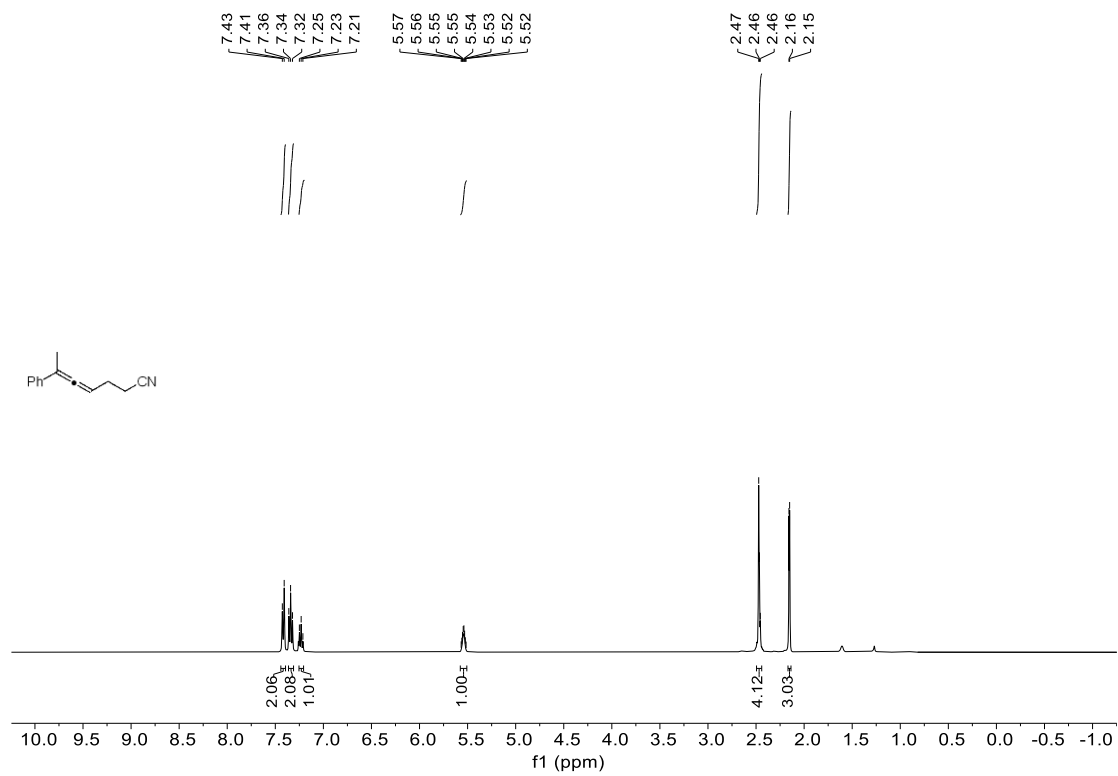
$^{13}\text{C NMR}$ (101 MHz, CDCl_3) spectra of 7c



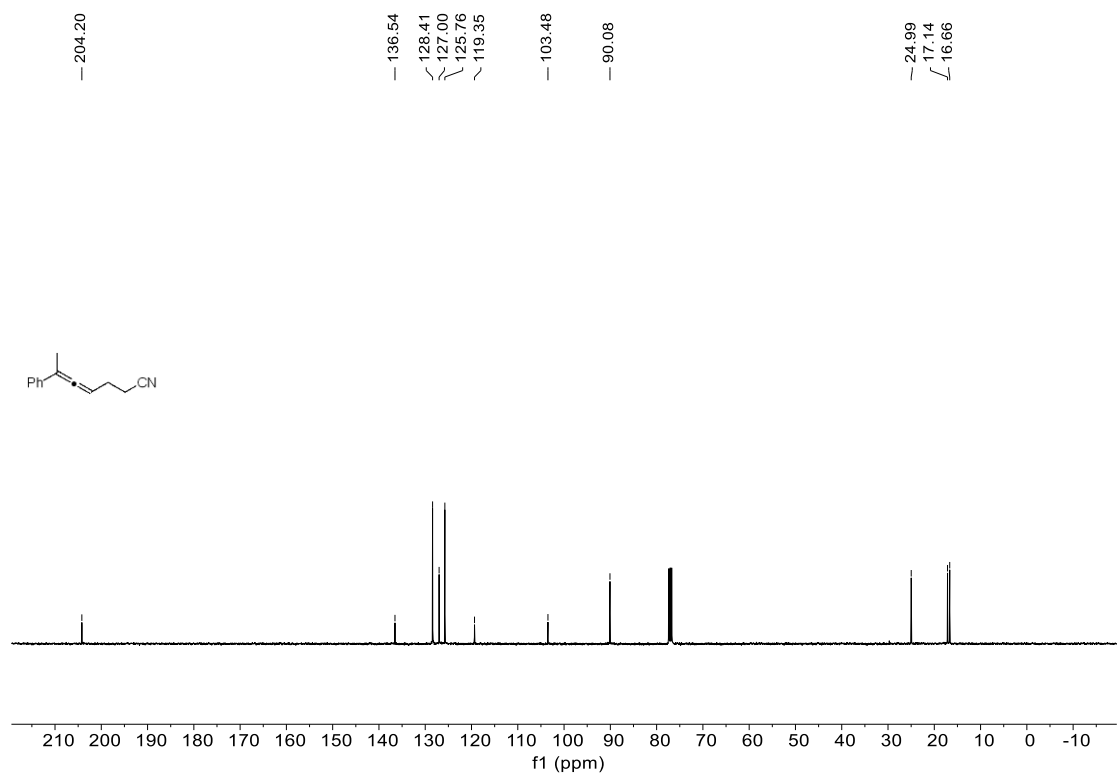
¹H NMR (400 MHz, CDCl₃) spectra of **8c**



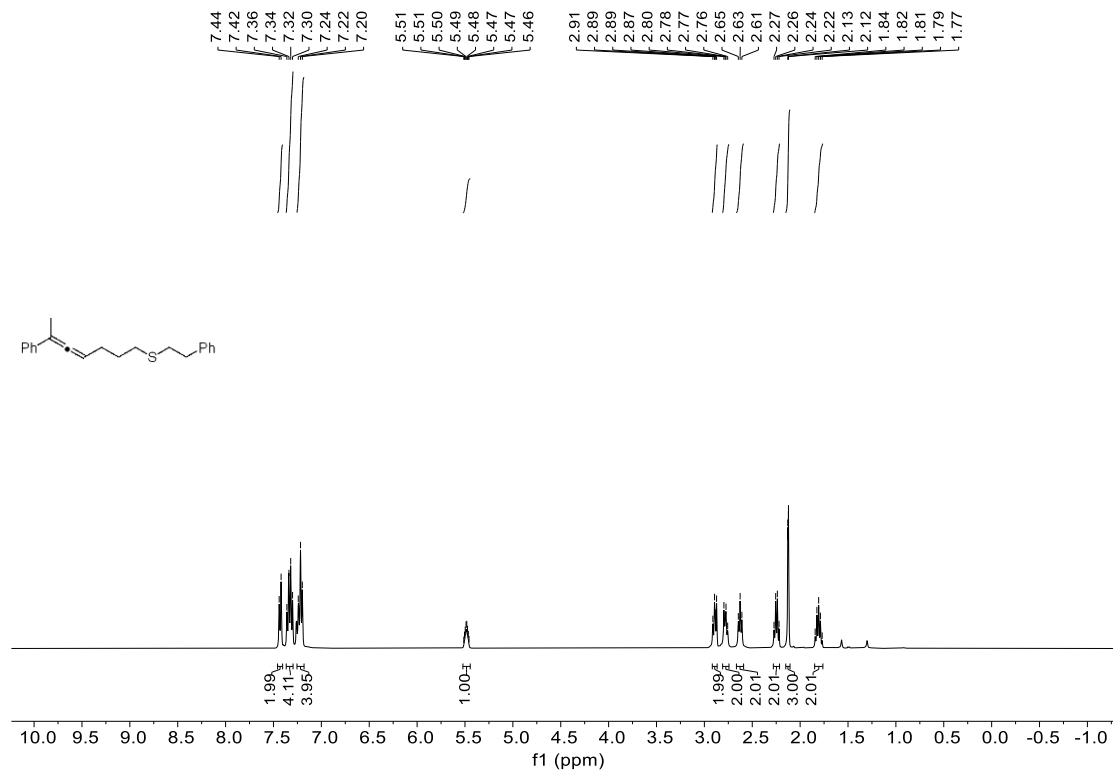
¹³C NMR (101 MHz, CDCl₃) spectra of **8c**



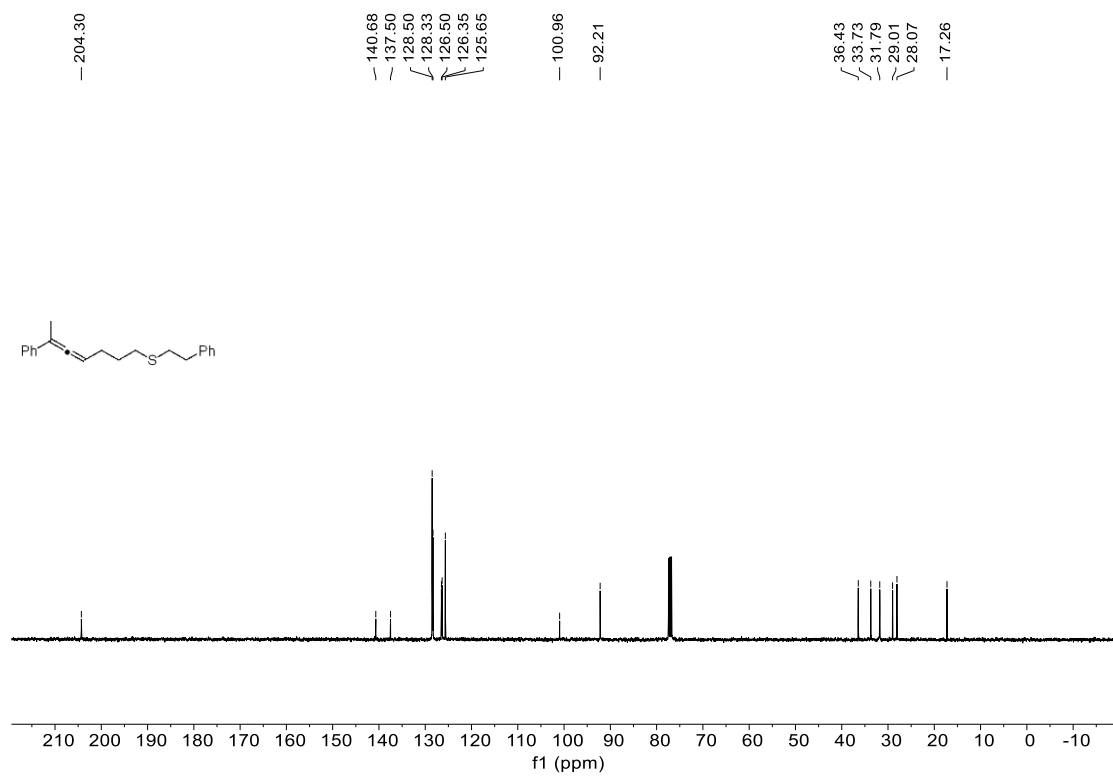
¹H NMR (400 MHz, CDCl₃) spectra of **9c**



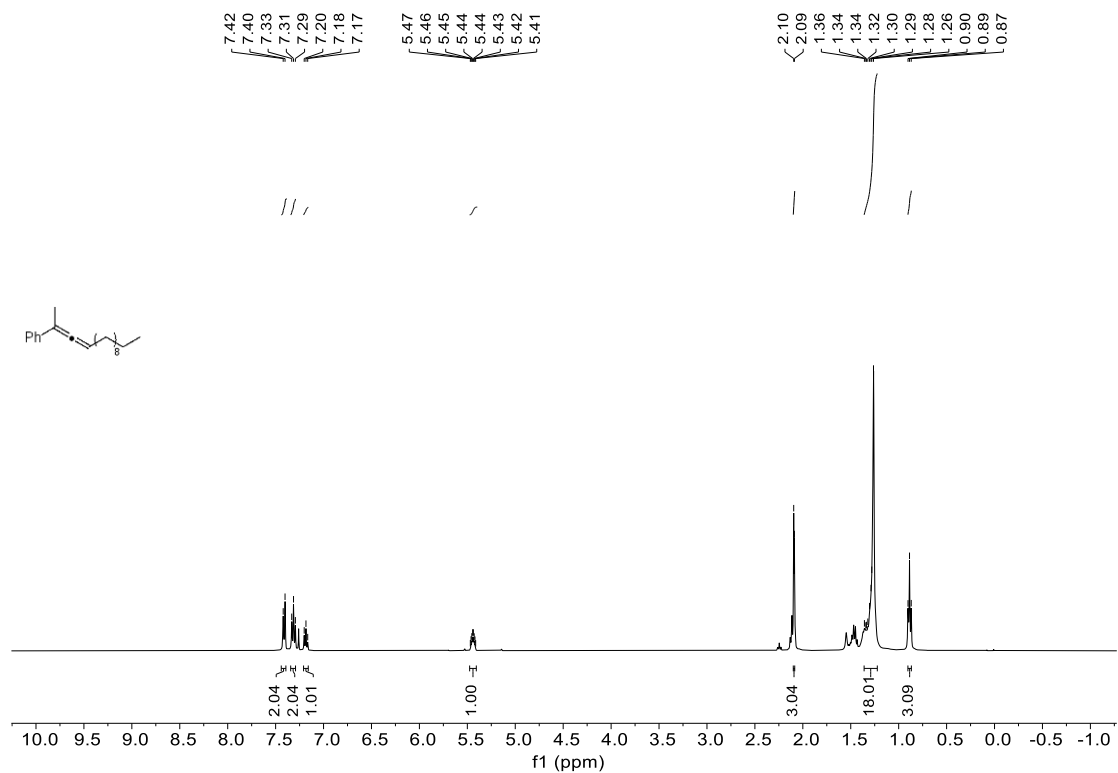
¹³C NMR (101 MHz, CDCl₃) spectra of **9c**



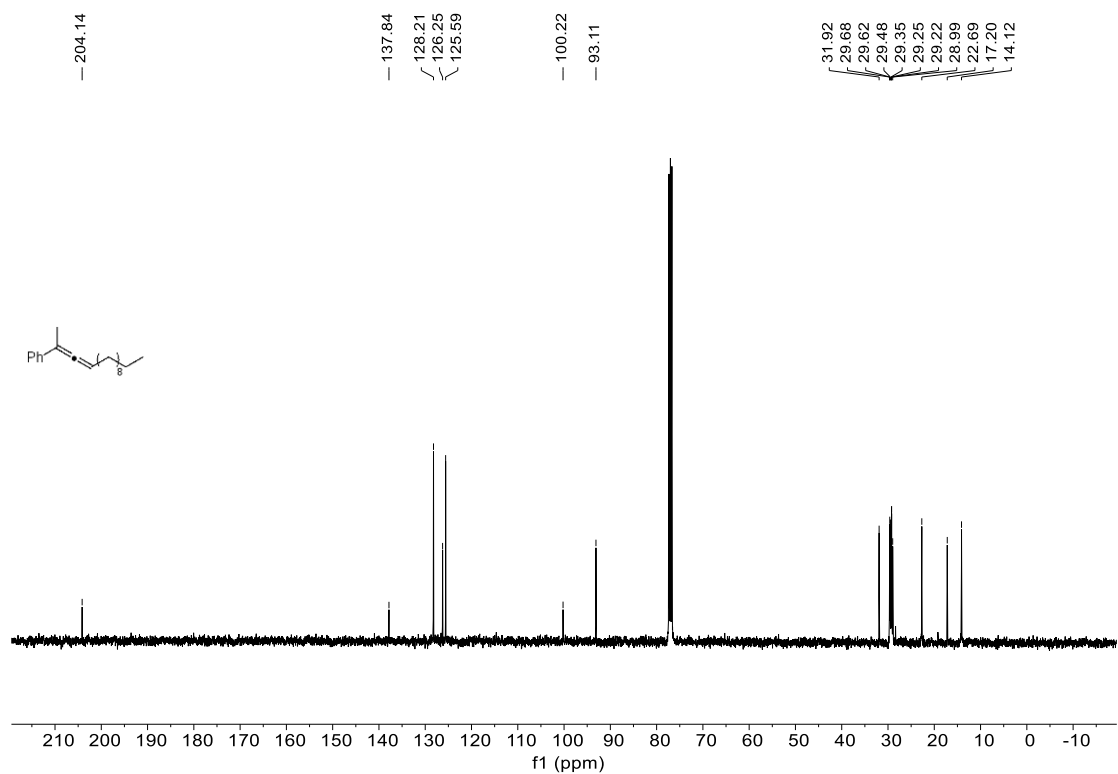
¹H NMR (400 MHz, CDCl₃) spectra of 10c



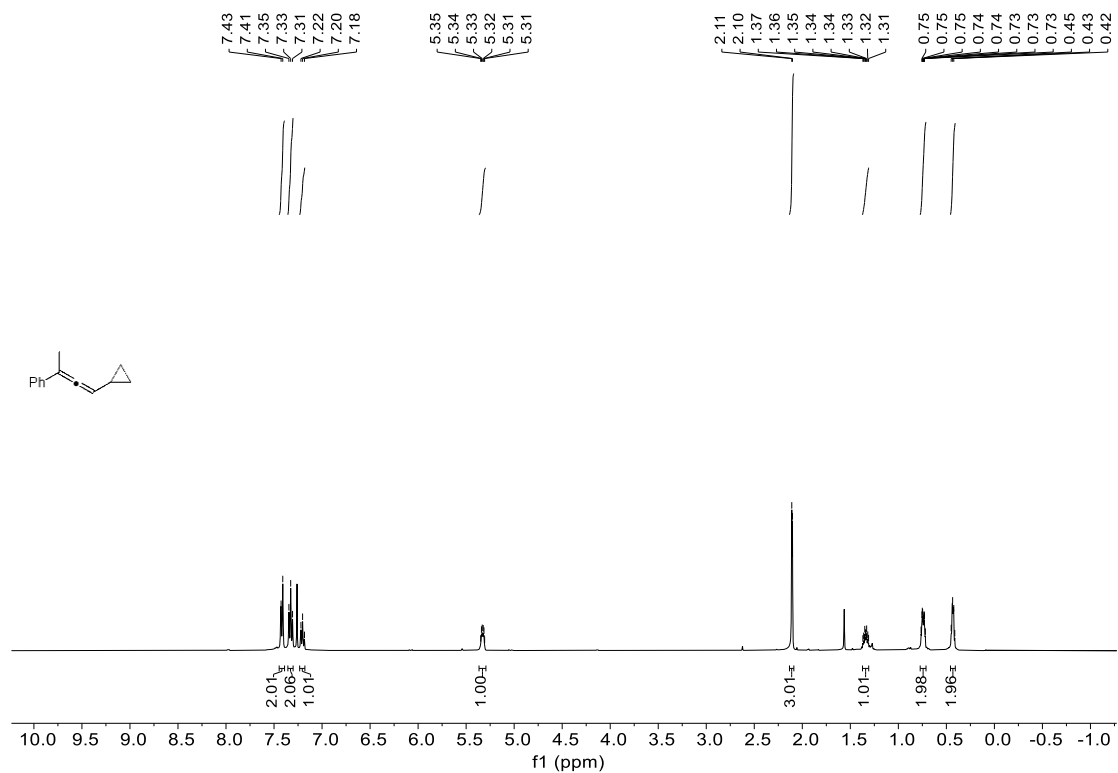
¹³C NMR (101 MHz, CDCl₃) spectra of 10c



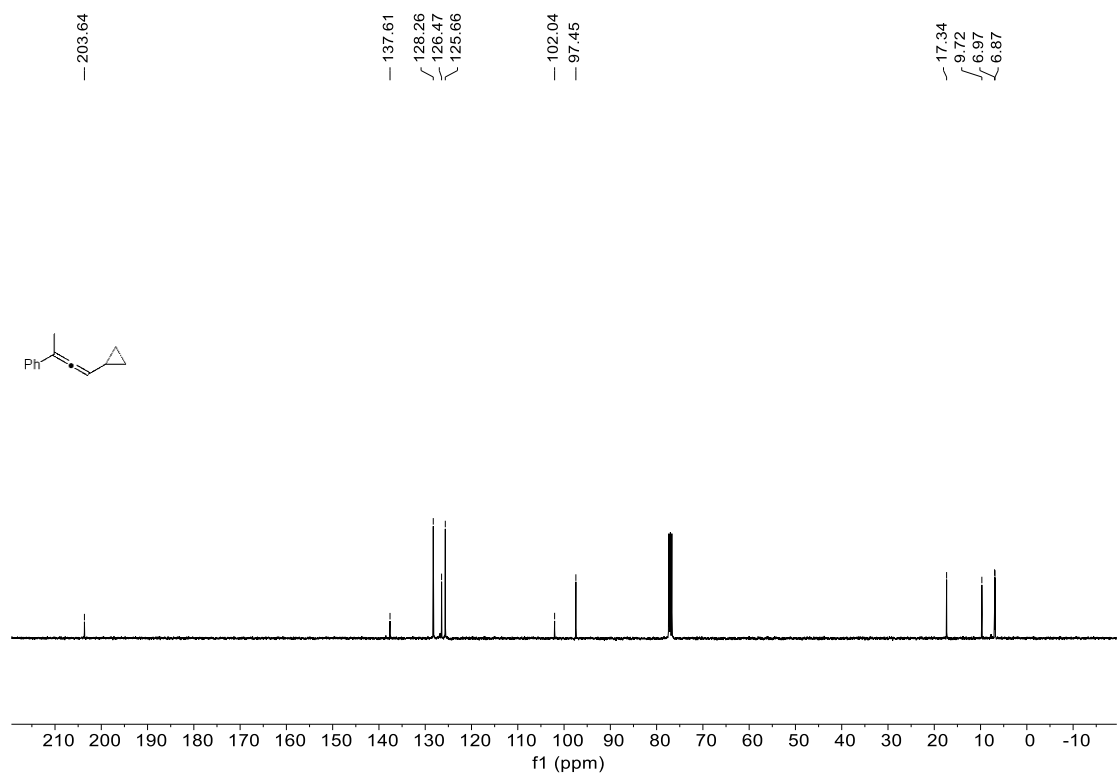
¹H NMR (400 MHz, CDCl₃) spectra of **11c**



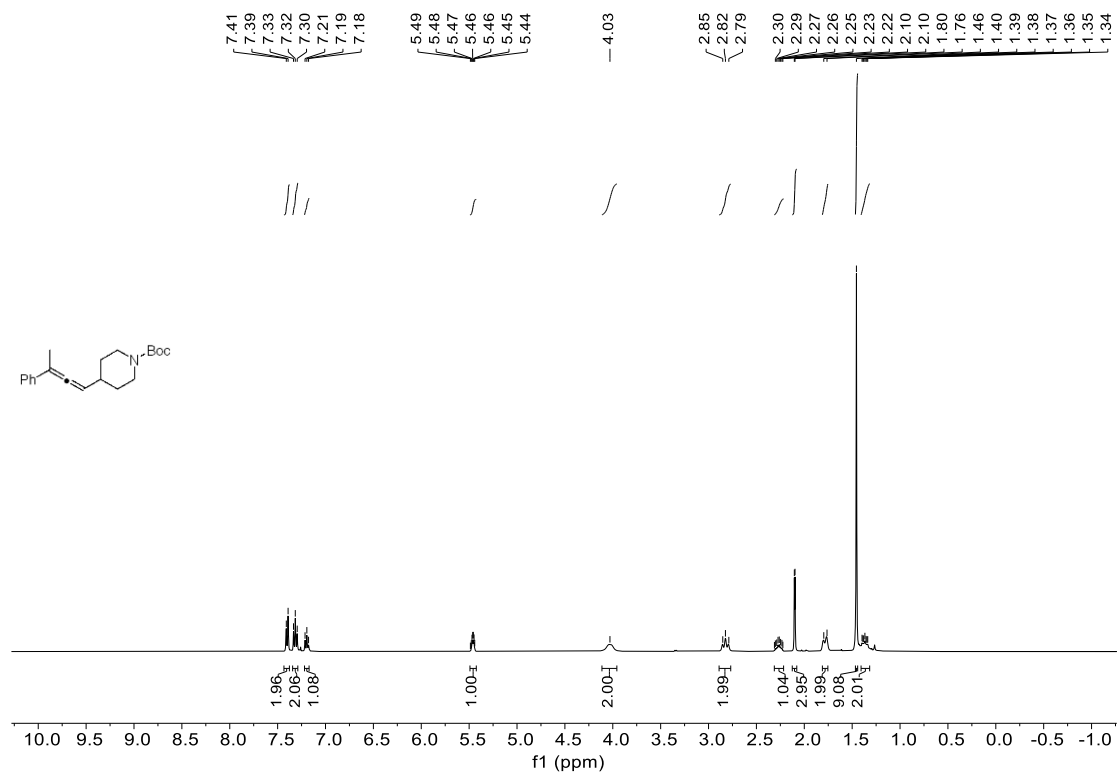
¹³C NMR (101 MHz, CDCl₃) spectra of **11c**



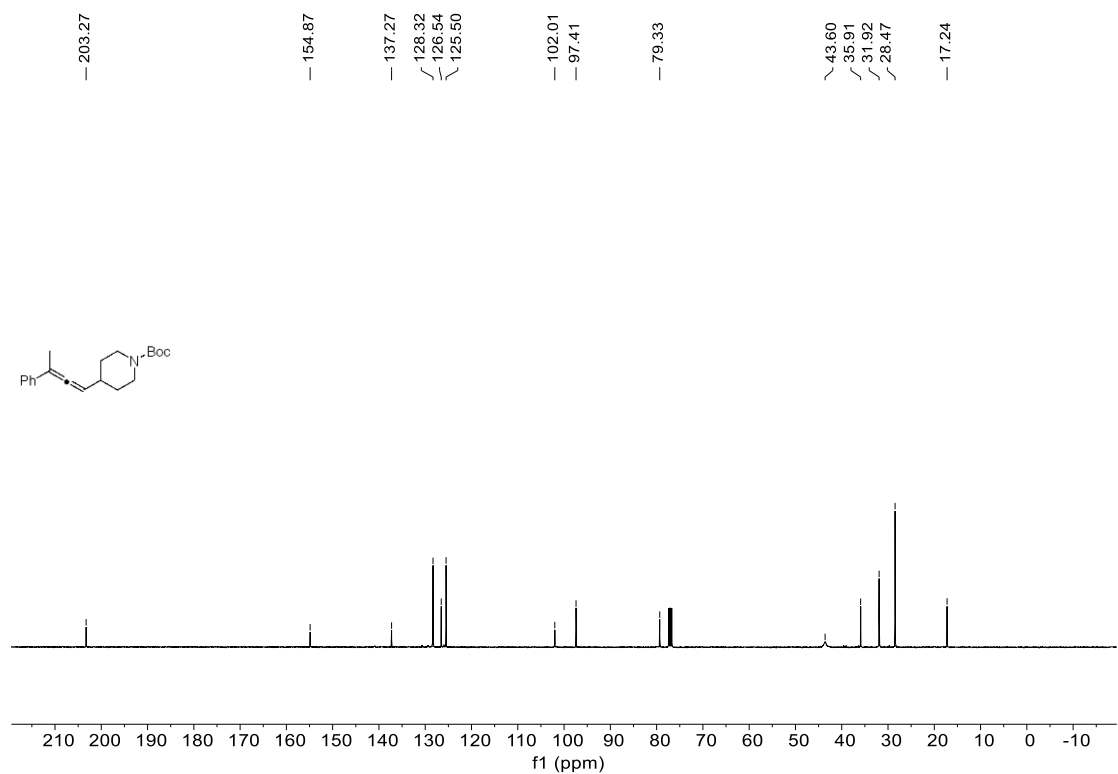
¹H NMR (400 MHz, CDCl₃) spectra of **12c**



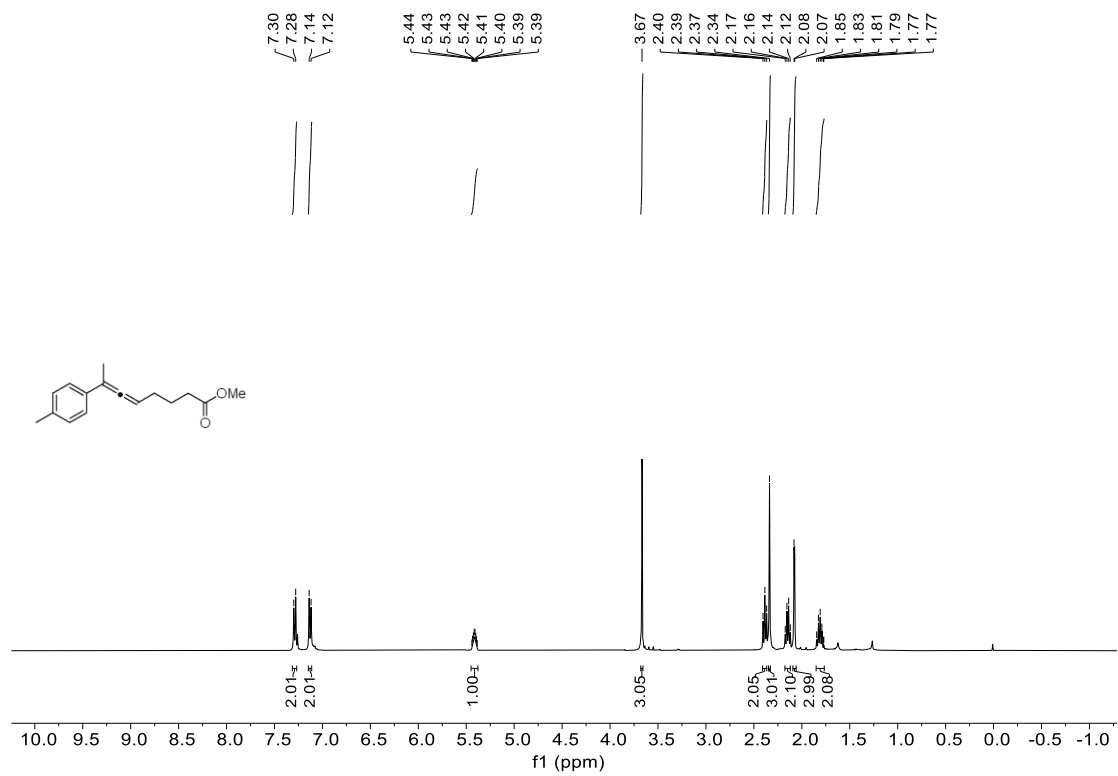
¹³C NMR (101 MHz, CDCl₃) spectra of **12c**



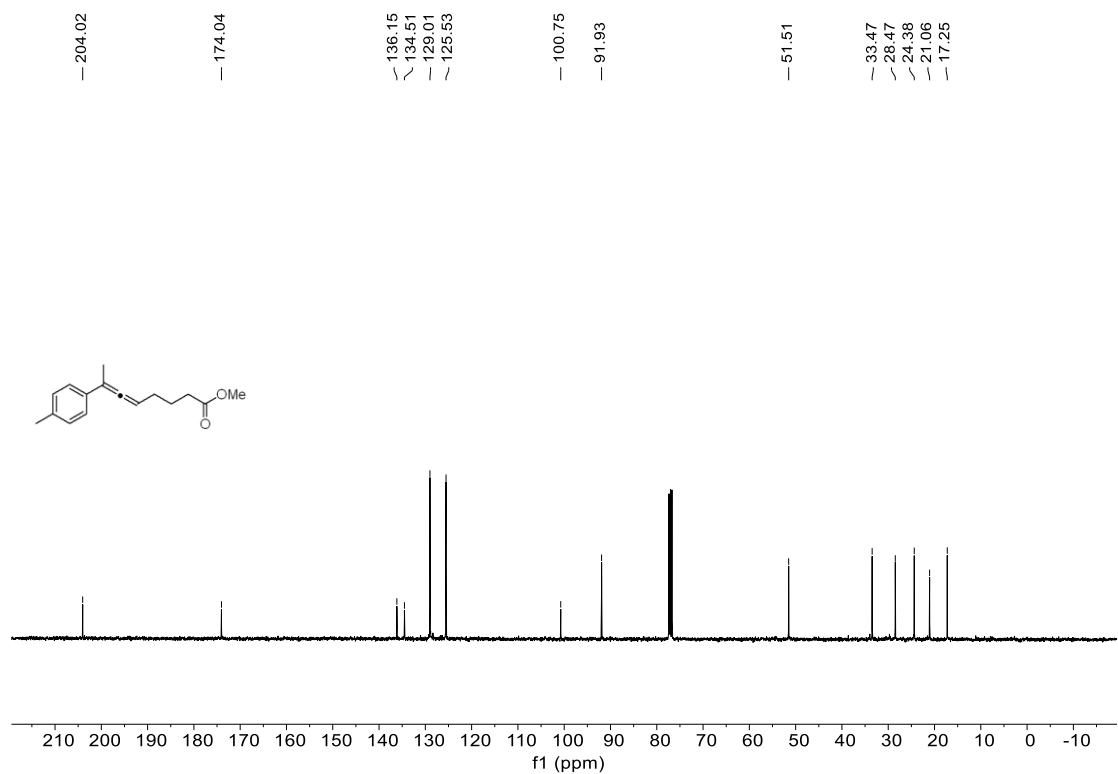
^1H NMR (400 MHz, CDCl_3) spectra of **13c**



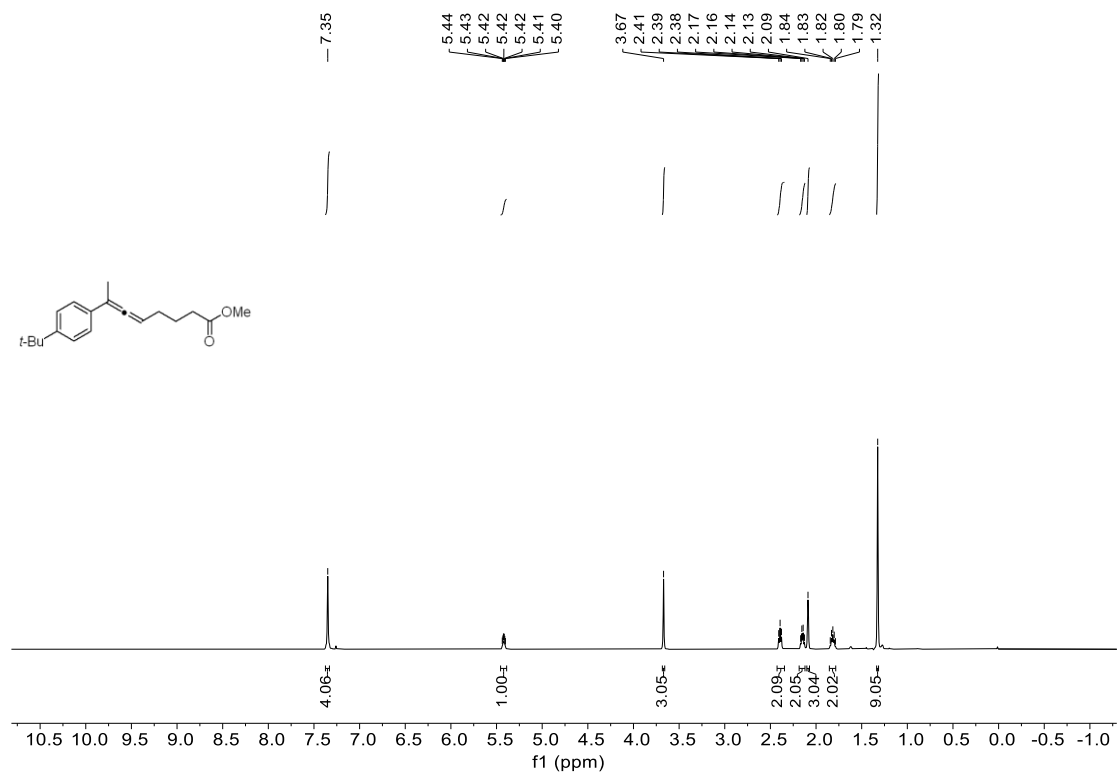
^{13}C NMR (101 MHz, CDCl_3) spectra of **13c**



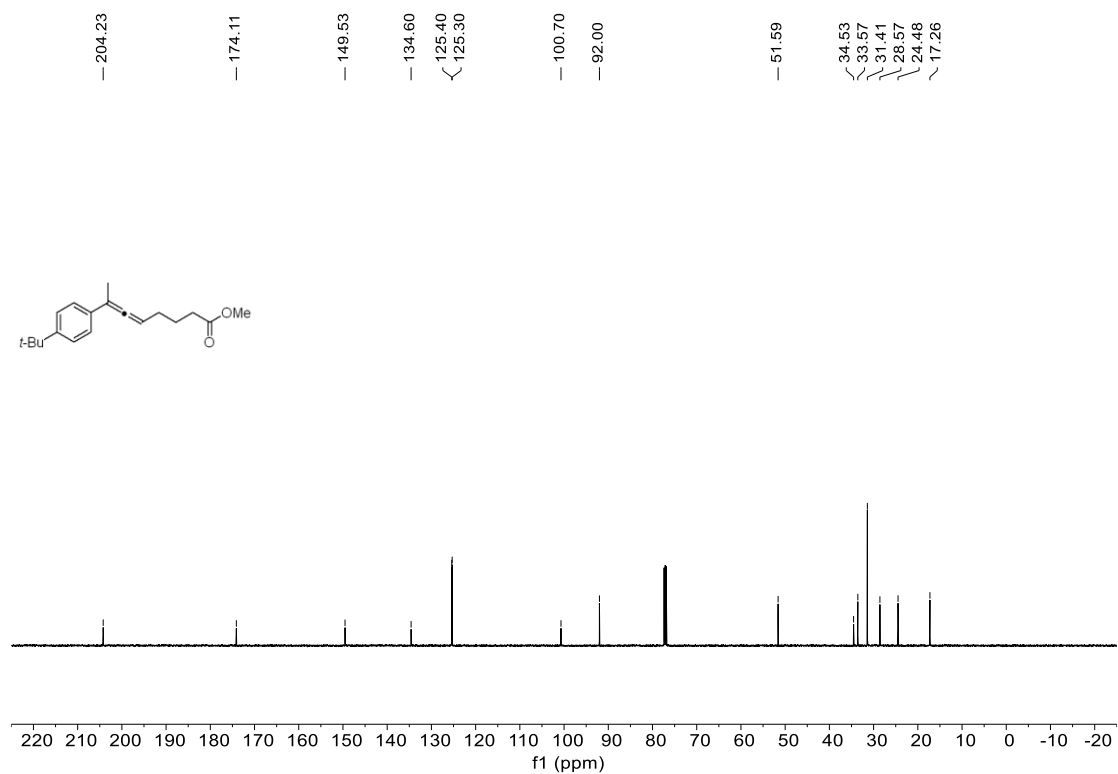
¹H NMR (400 MHz, CDCl₃) spectra of 14c



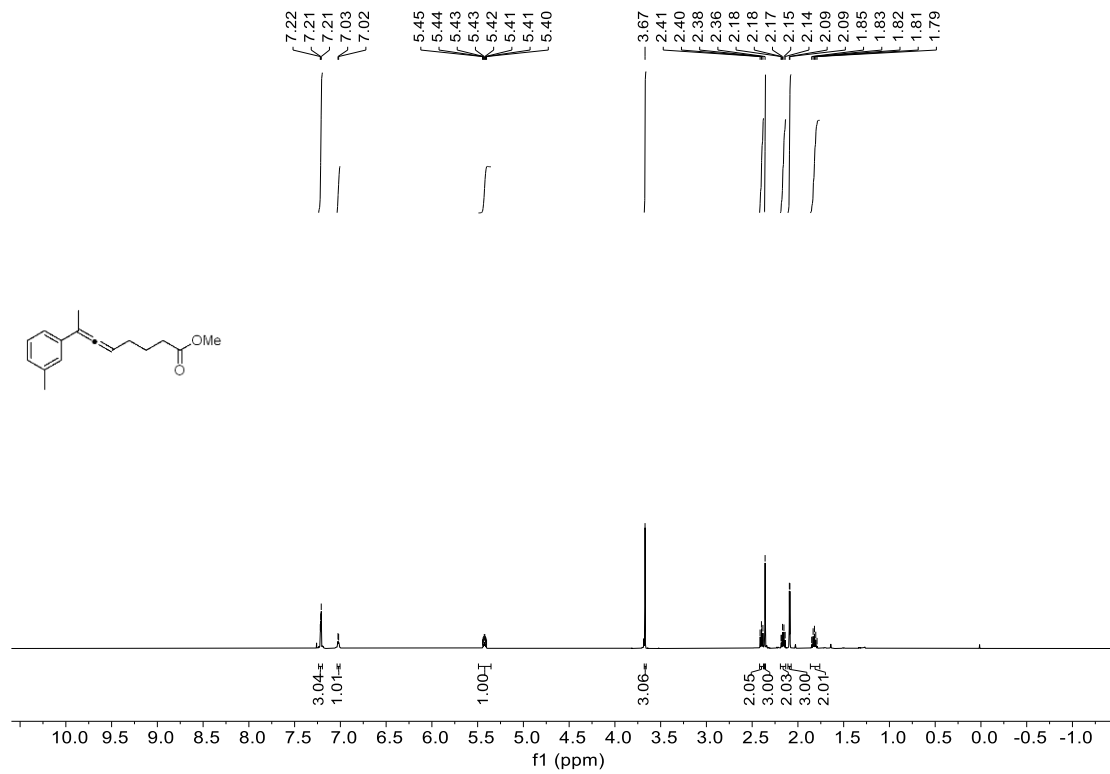
¹³C NMR (101 MHz, CDCl₃) spectra of 14c



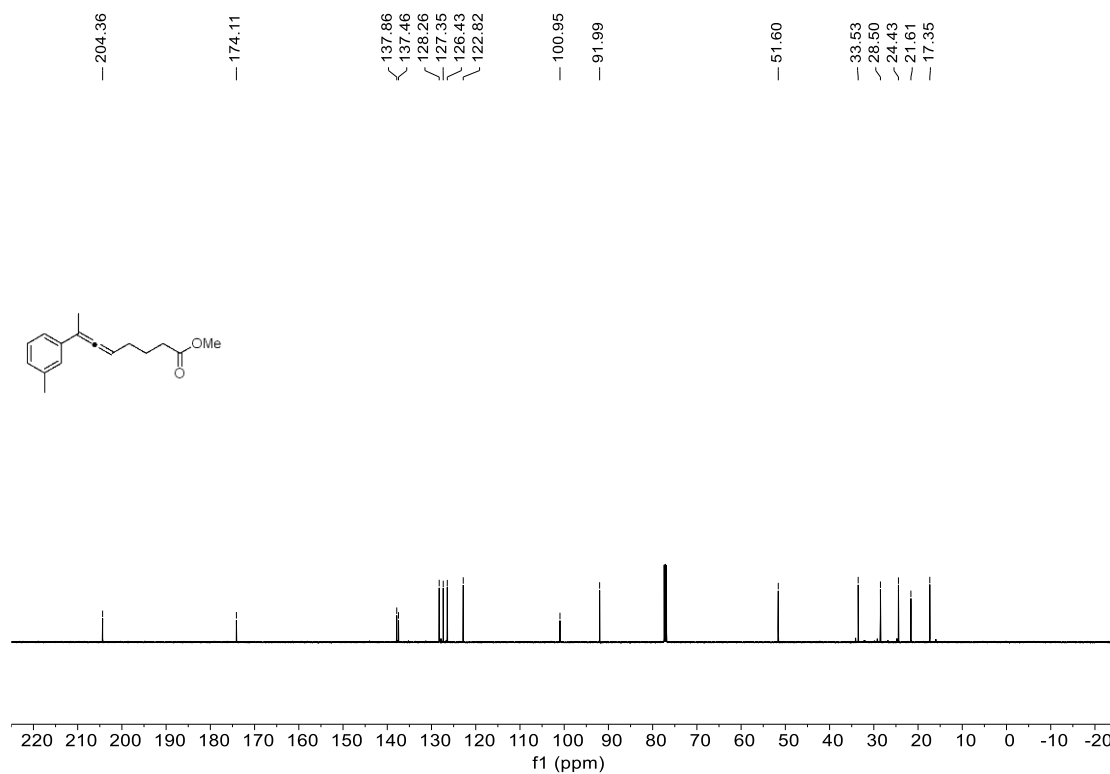
^1H NMR (500 MHz, CDCl_3) spectra of **15c**



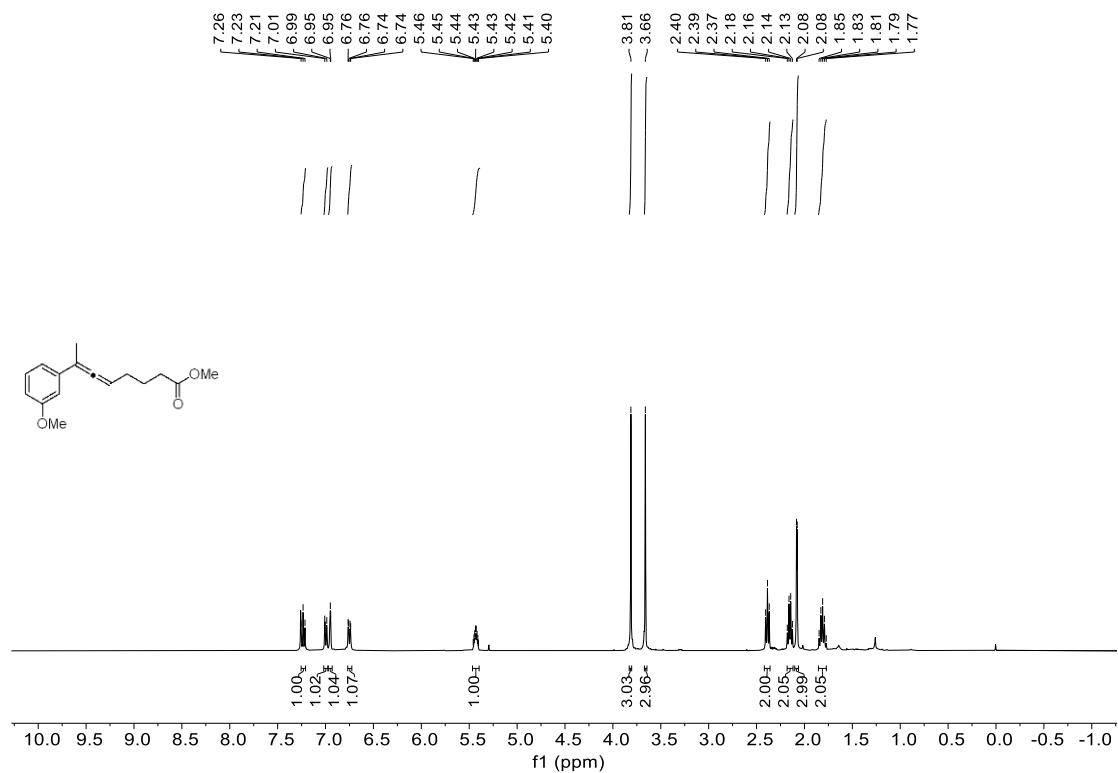
^{13}C NMR (126 MHz, CDCl_3) spectra of **15c**



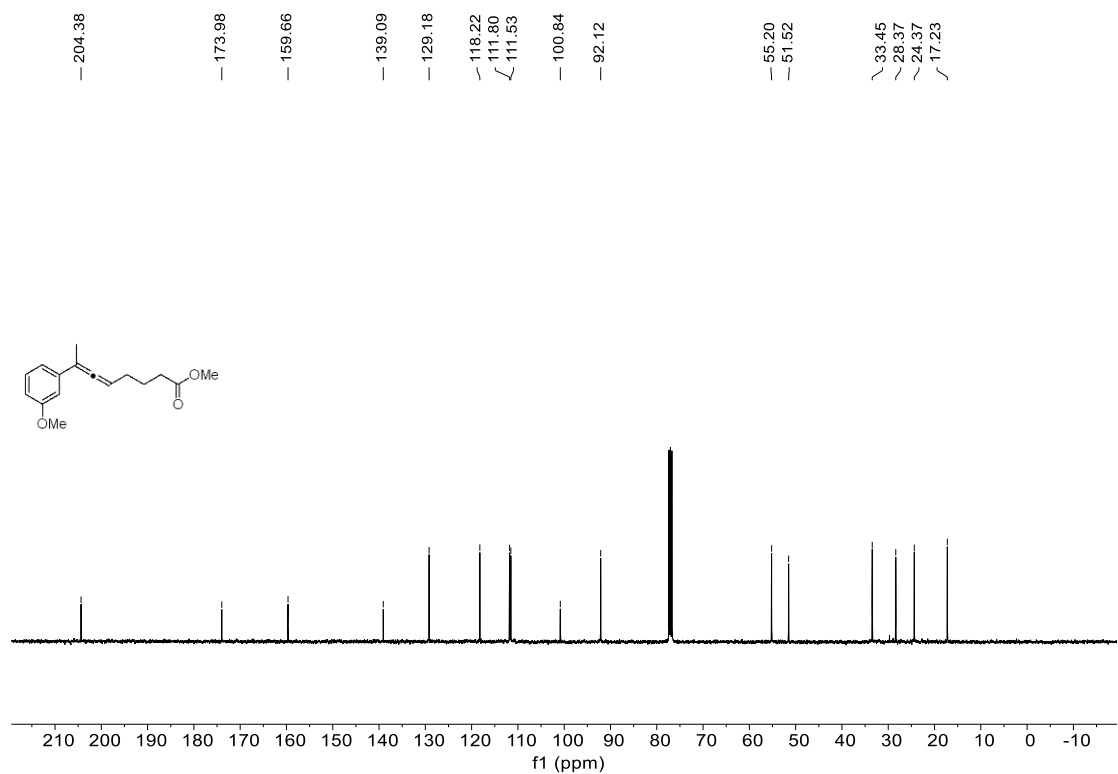
¹H NMR (500 MHz, CDCl₃) spectra of **16c**



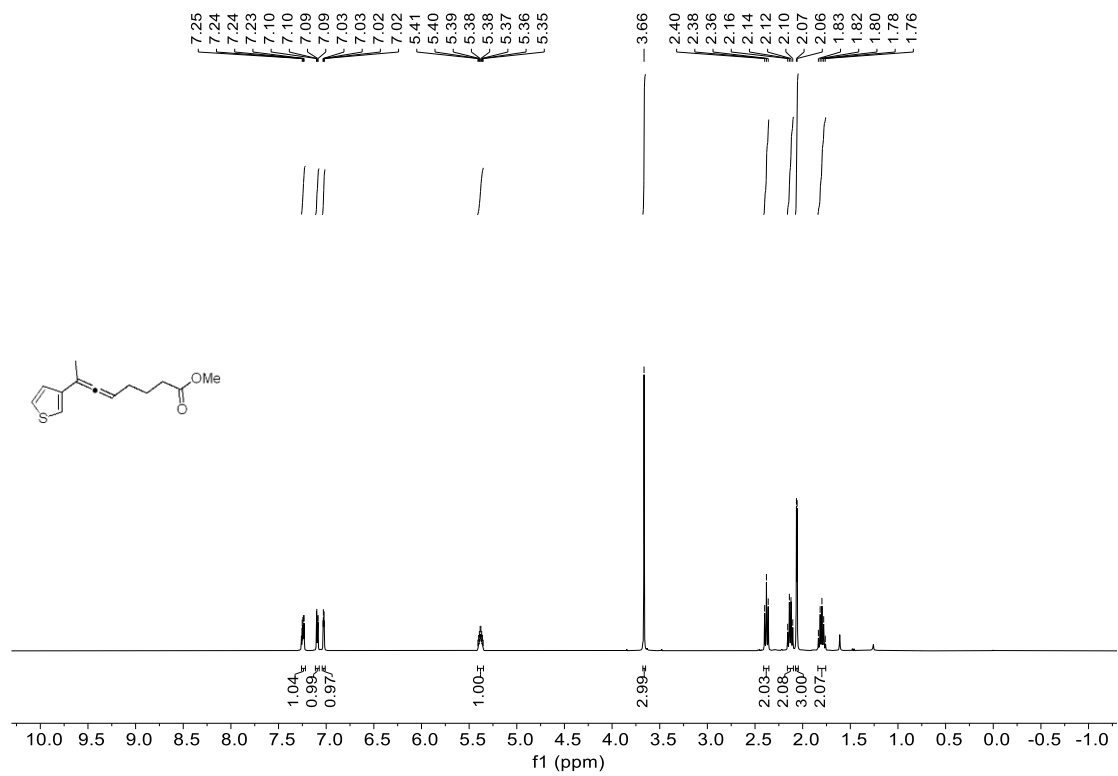
¹³C NMR (126 MHz, CDCl₃) spectra of **16c**



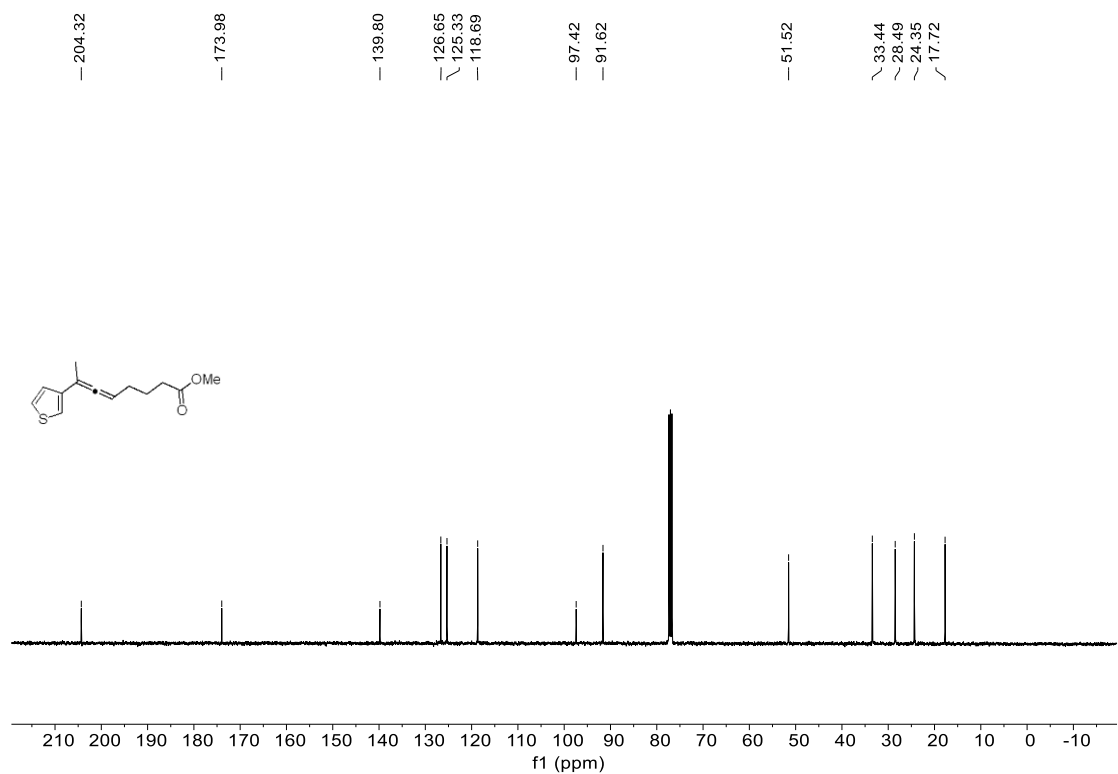
¹H NMR (400 MHz, CDCl₃) spectra of 17c



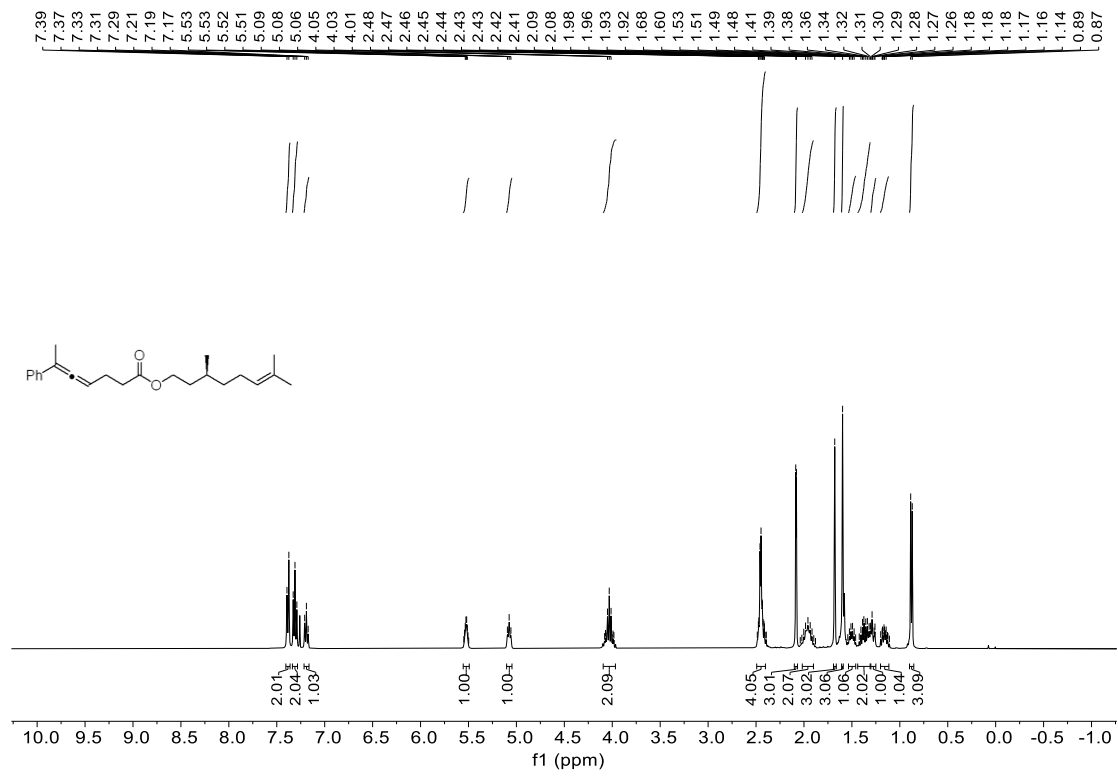
¹³C NMR (101 MHz, CDCl₃) spectra of 17c



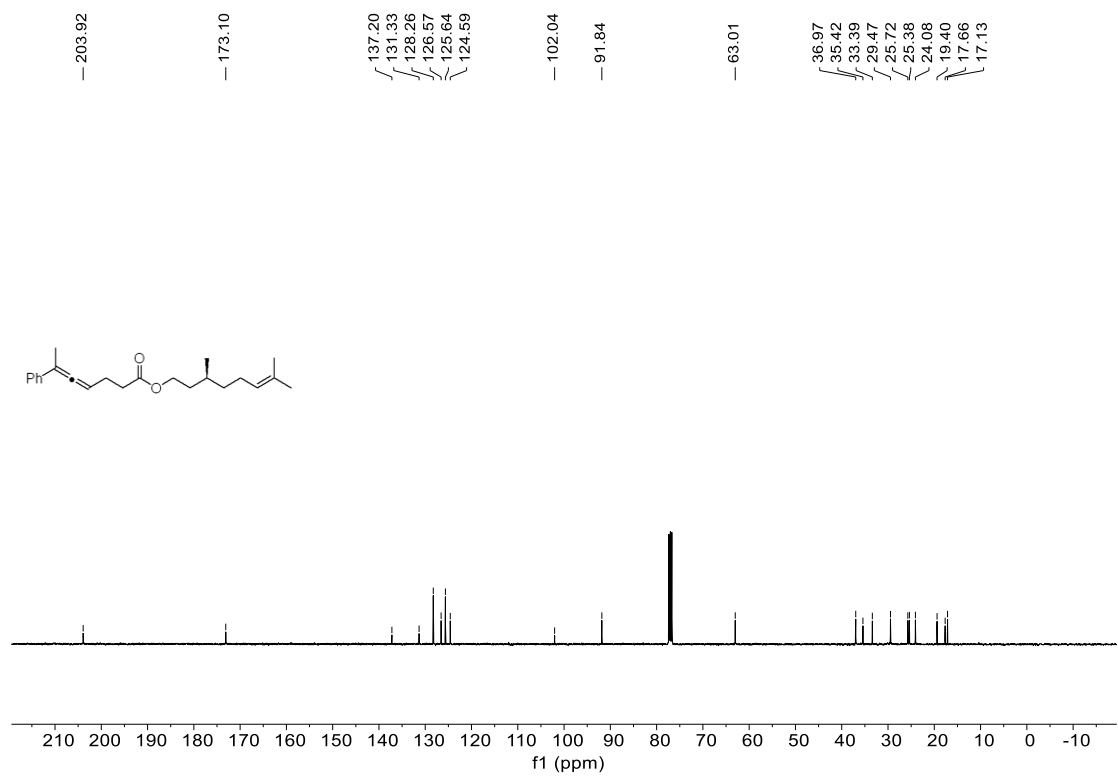
¹H NMR (400 MHz, CDCl₃) spectra of **18c**



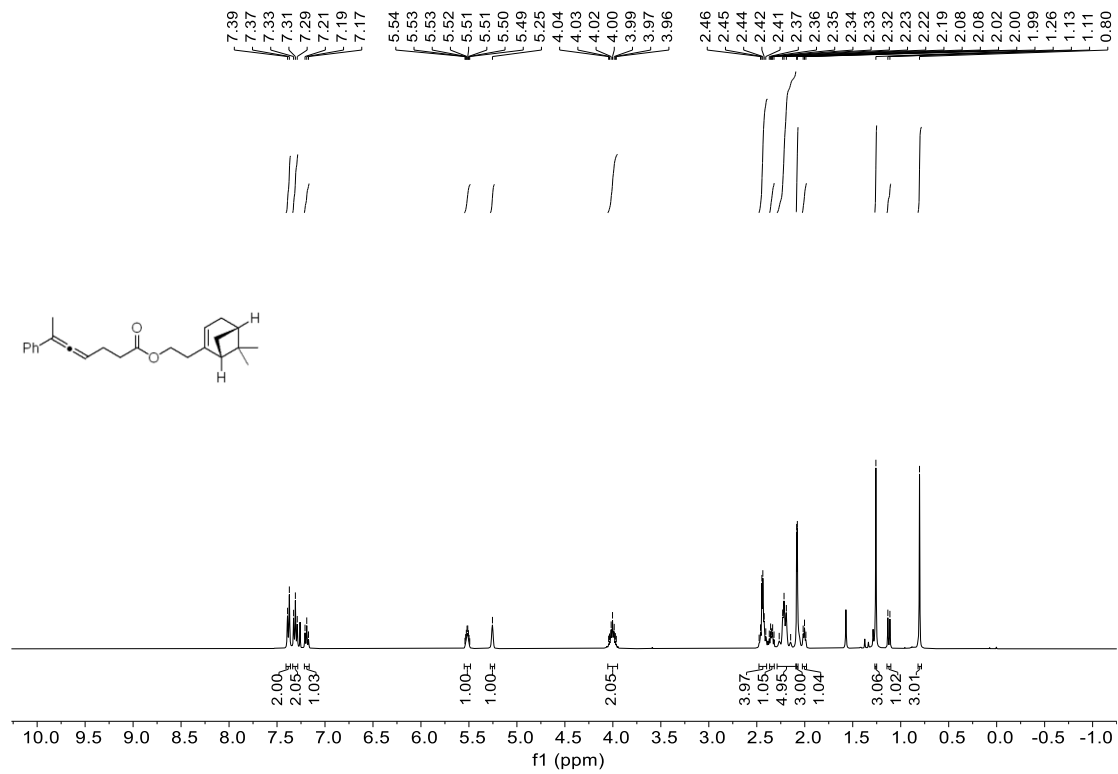
¹³C NMR (101 MHz, CDCl₃) spectra of **18c**



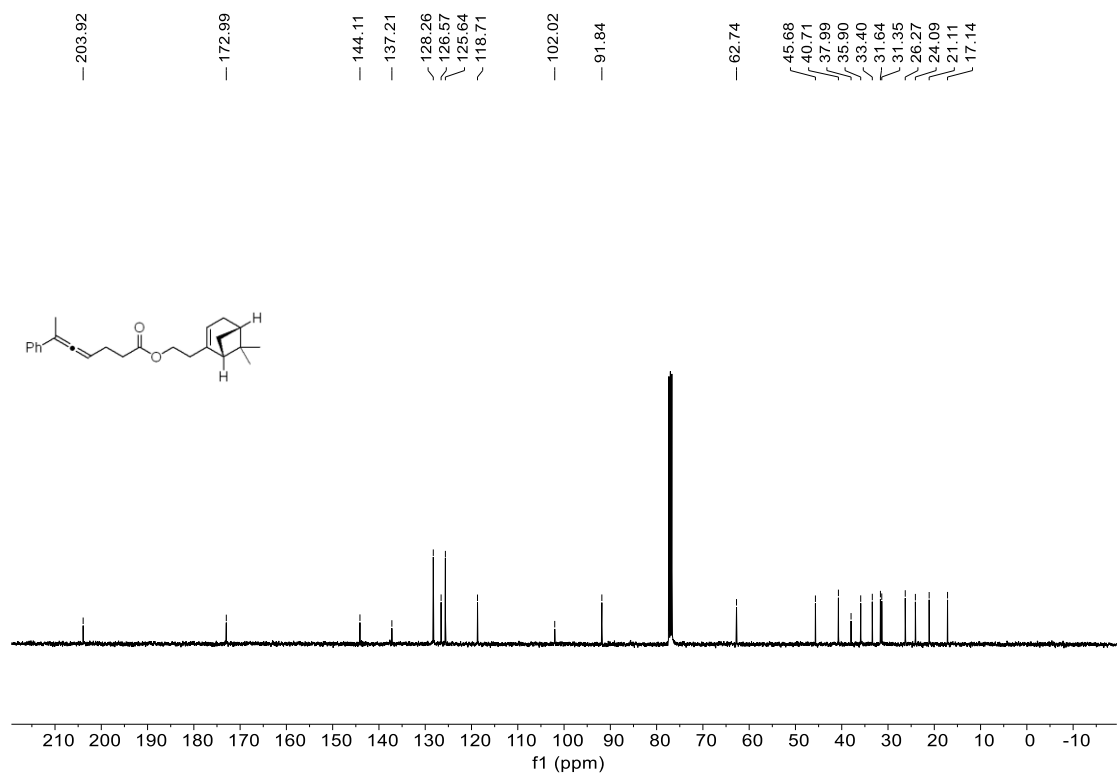
¹H NMR (400 MHz, CDCl₃) spectra of **19c**



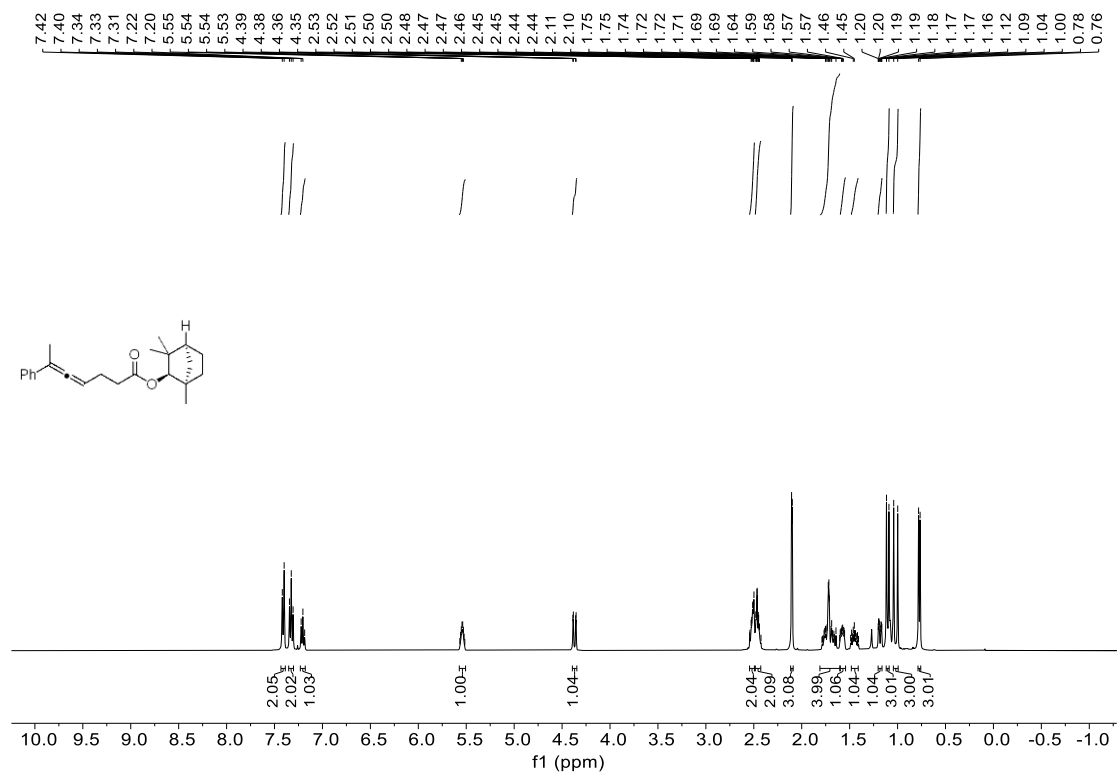
¹³C NMR (101 MHz, CDCl₃) spectra of **19c**



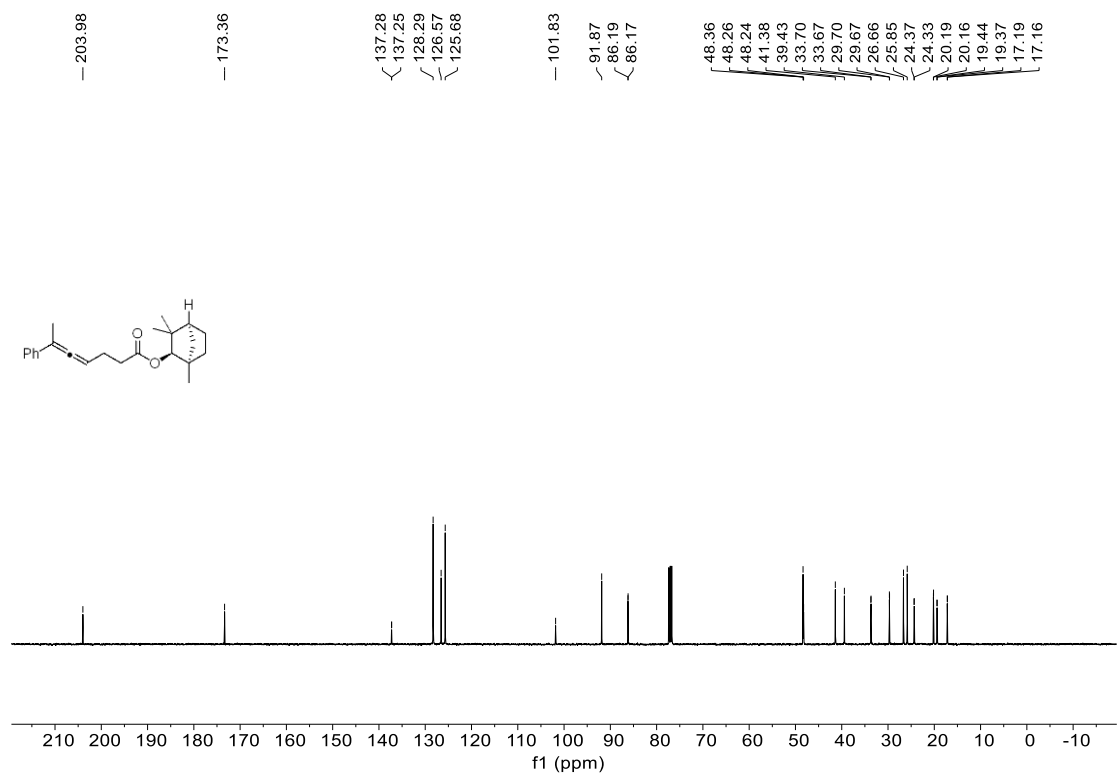
¹H NMR (400 MHz, CDCl₃) spectra of 20c



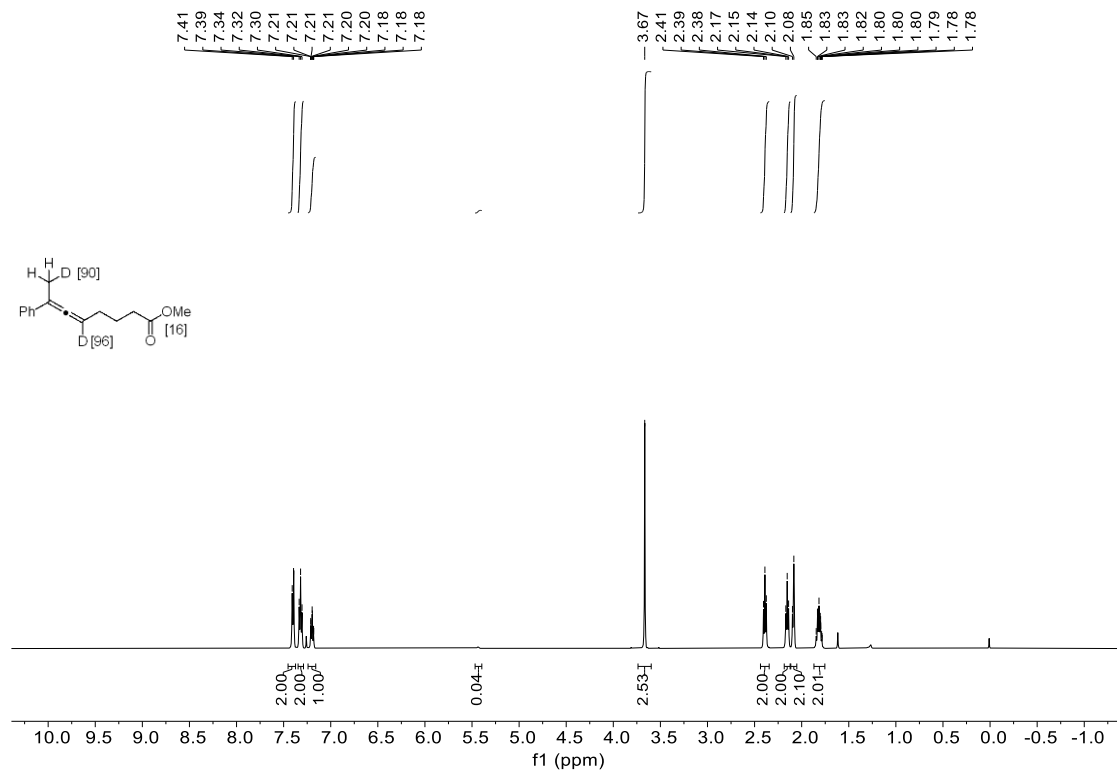
¹³C NMR (101 MHz, CDCl₃) spectra of 20c



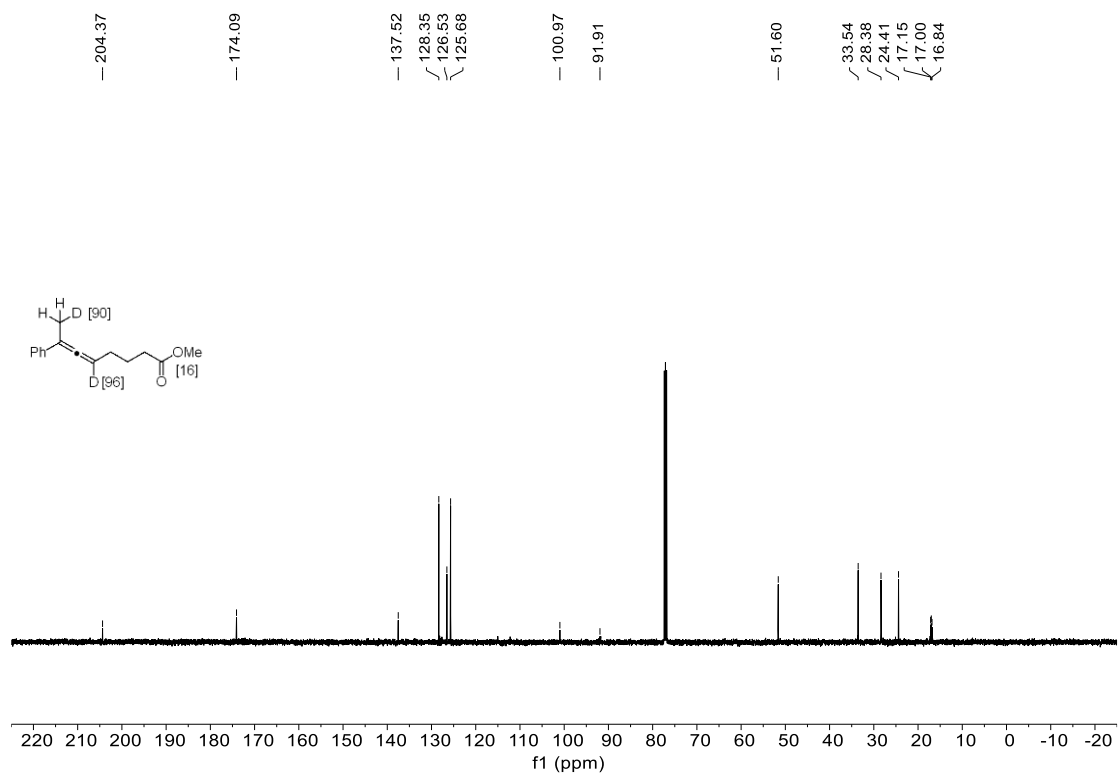
¹H NMR (400 MHz, CDCl₃) spectra of **21c**



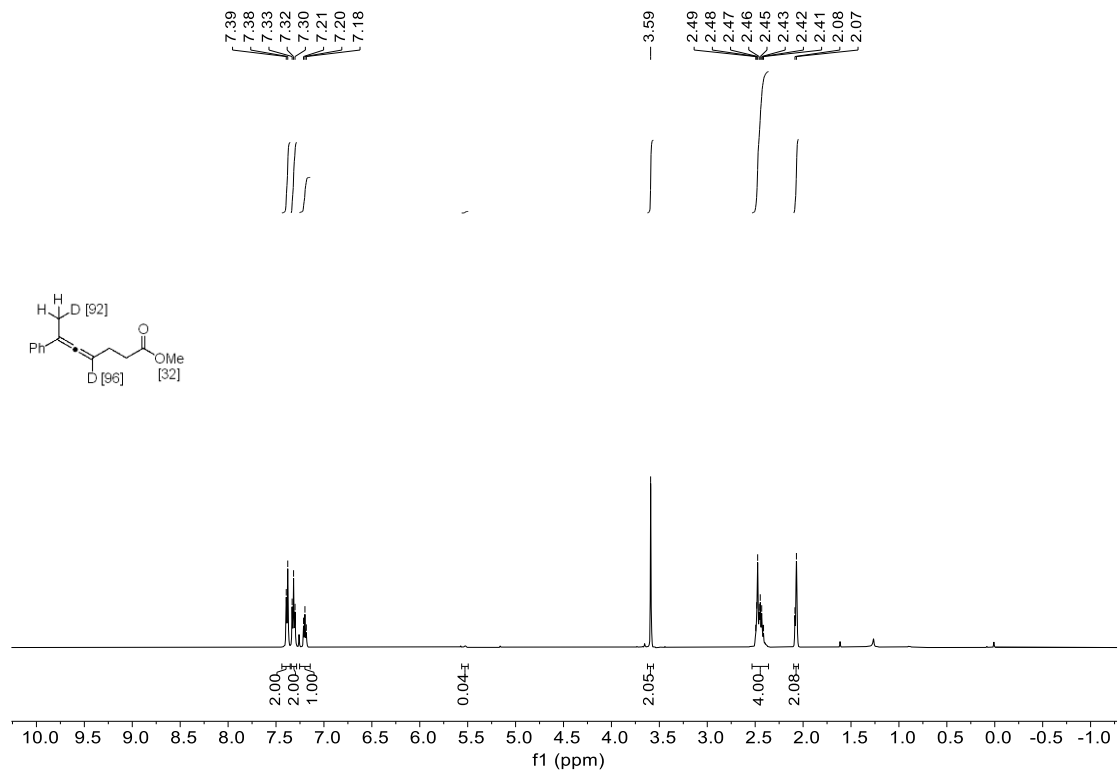
¹³C NMR (101 MHz, CDCl₃) spectra of **21c**



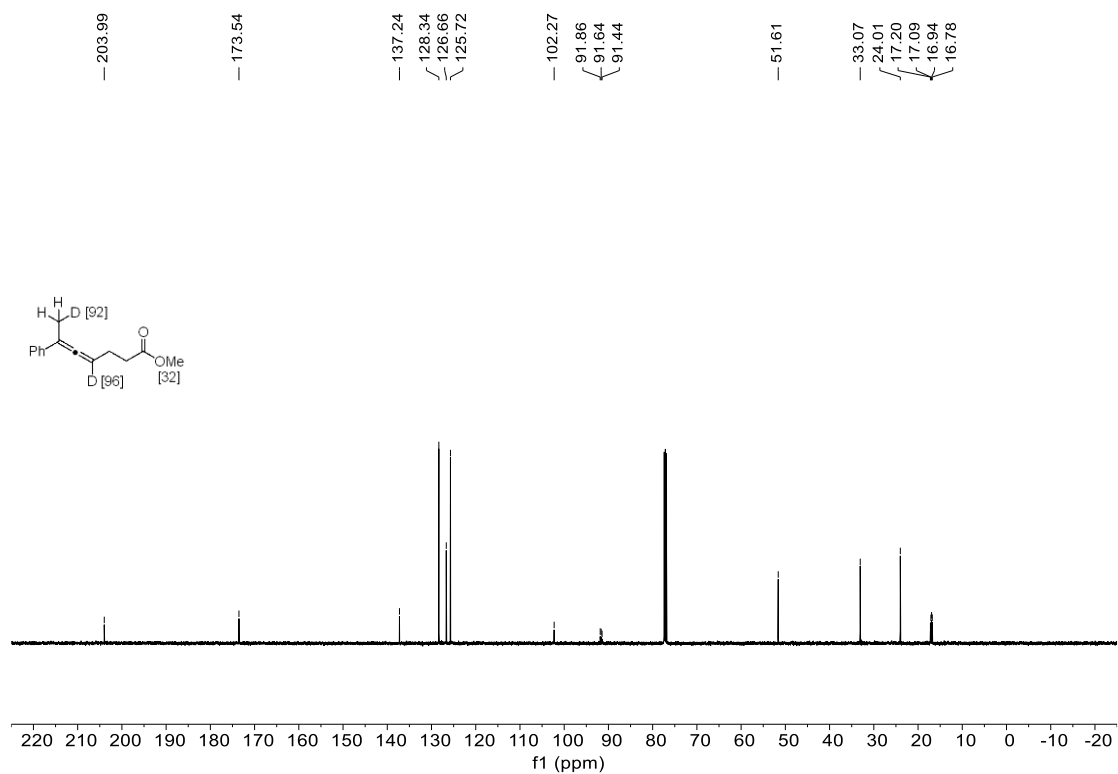
¹H NMR (500 MHz, CDCl₃) spectra of 2d



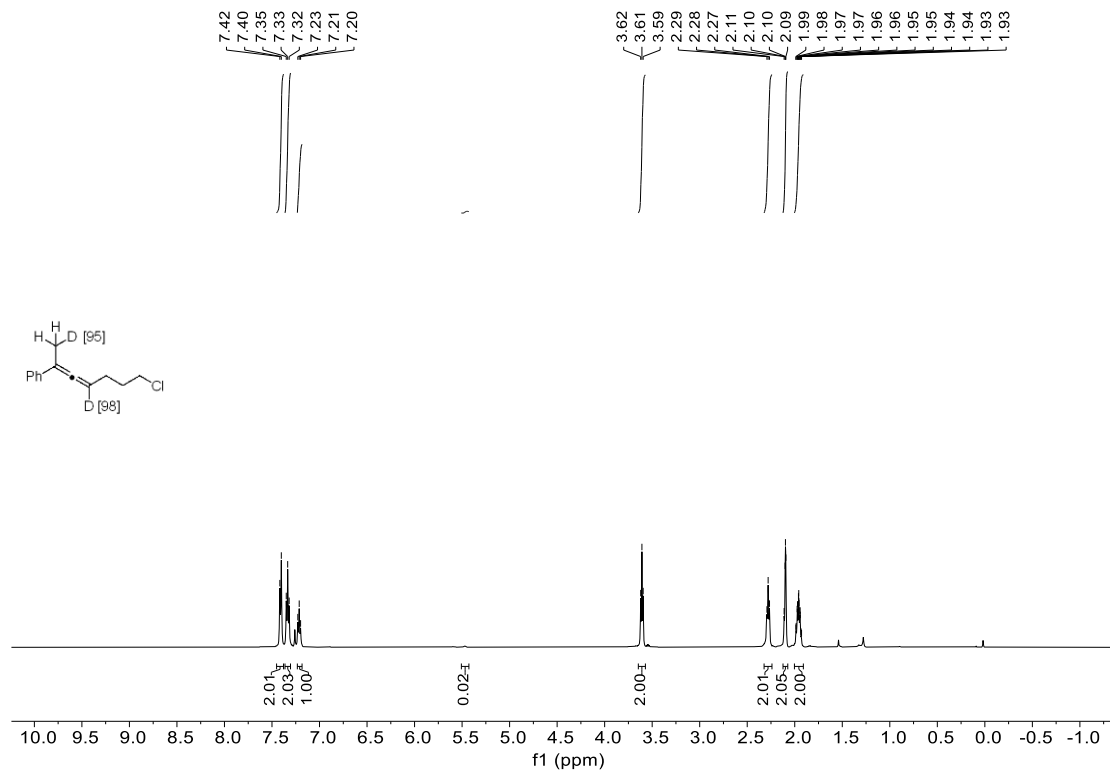
¹³C NMR (126 MHz, CDCl₃) spectra of 2d



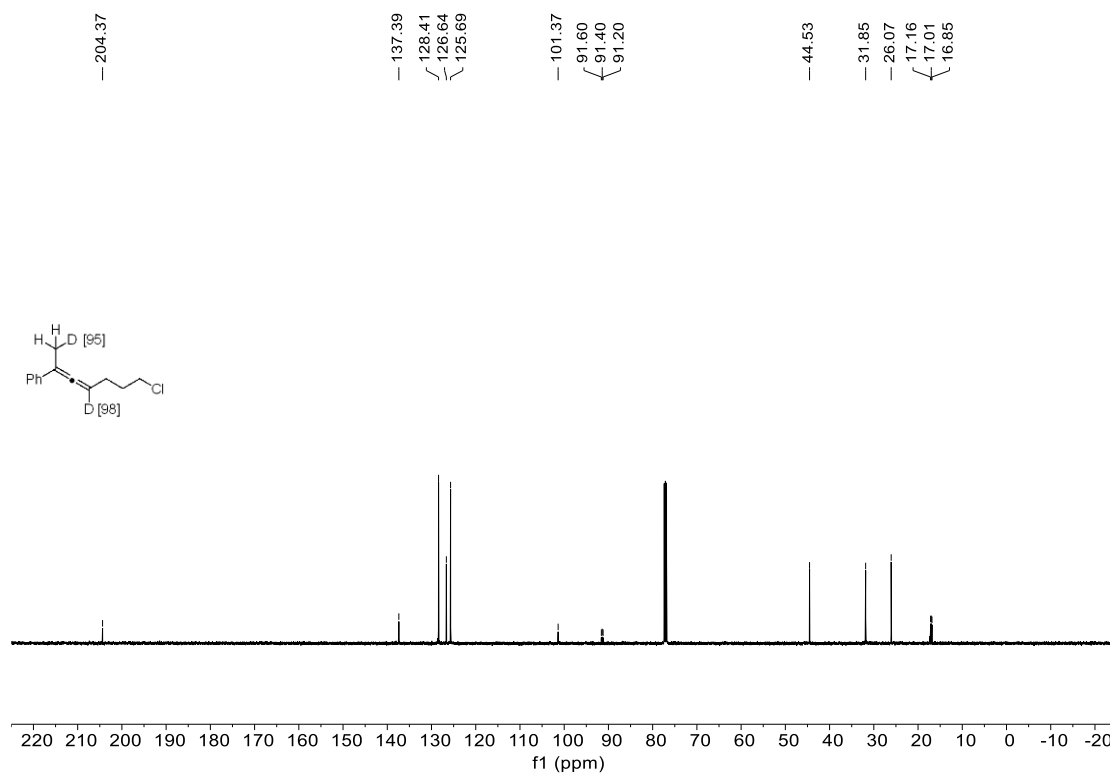
^1H NMR (500 MHz, CDCl_3) spectra of **3d**



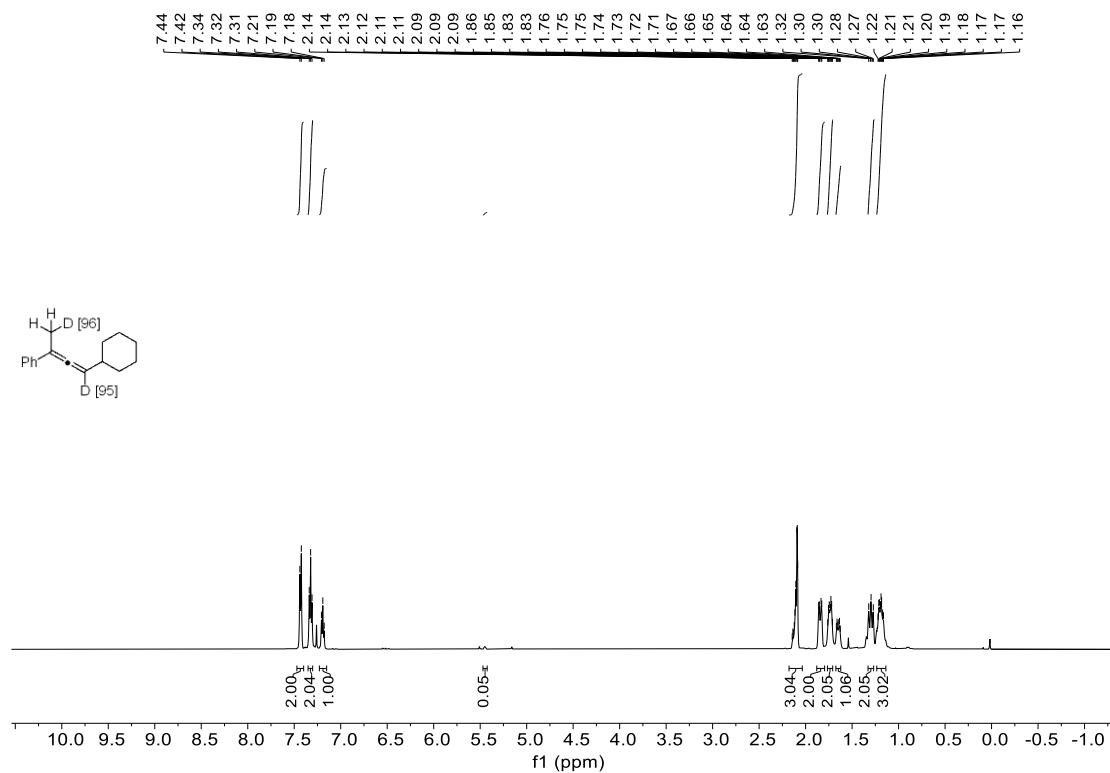
^{13}C NMR (500 MHz, CDCl_3) spectra of **3d**



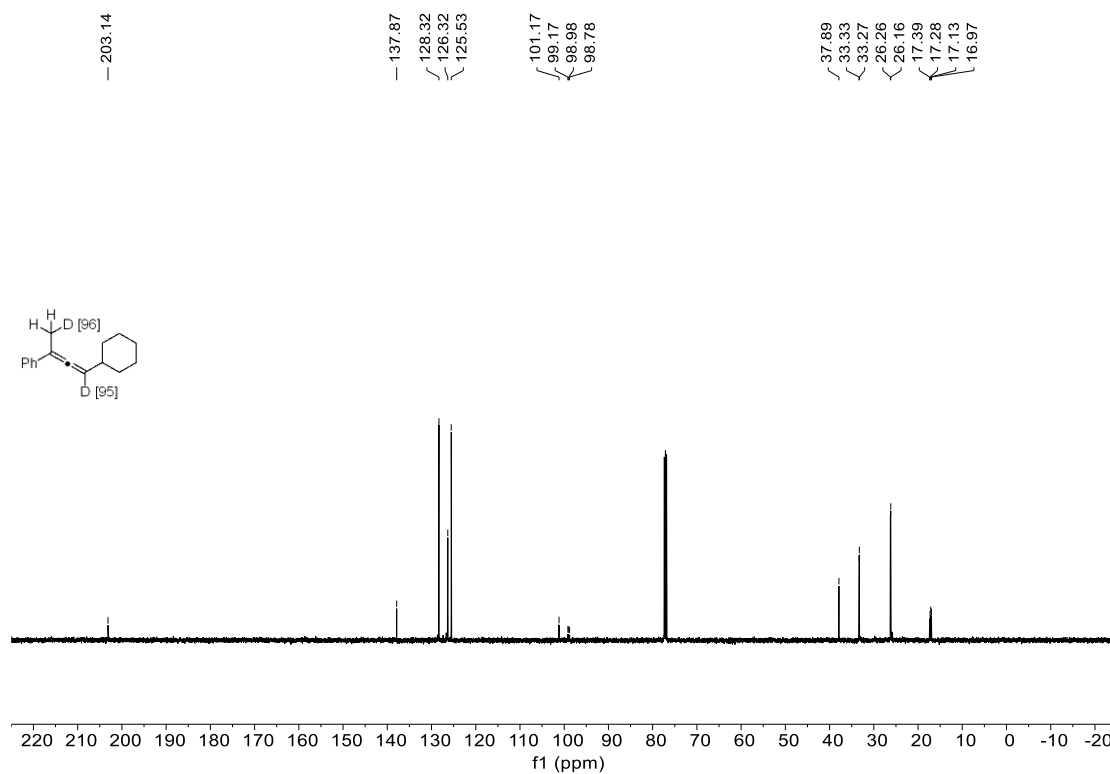
¹H NMR (500 MHz, CDCl₃) spectra of 4d



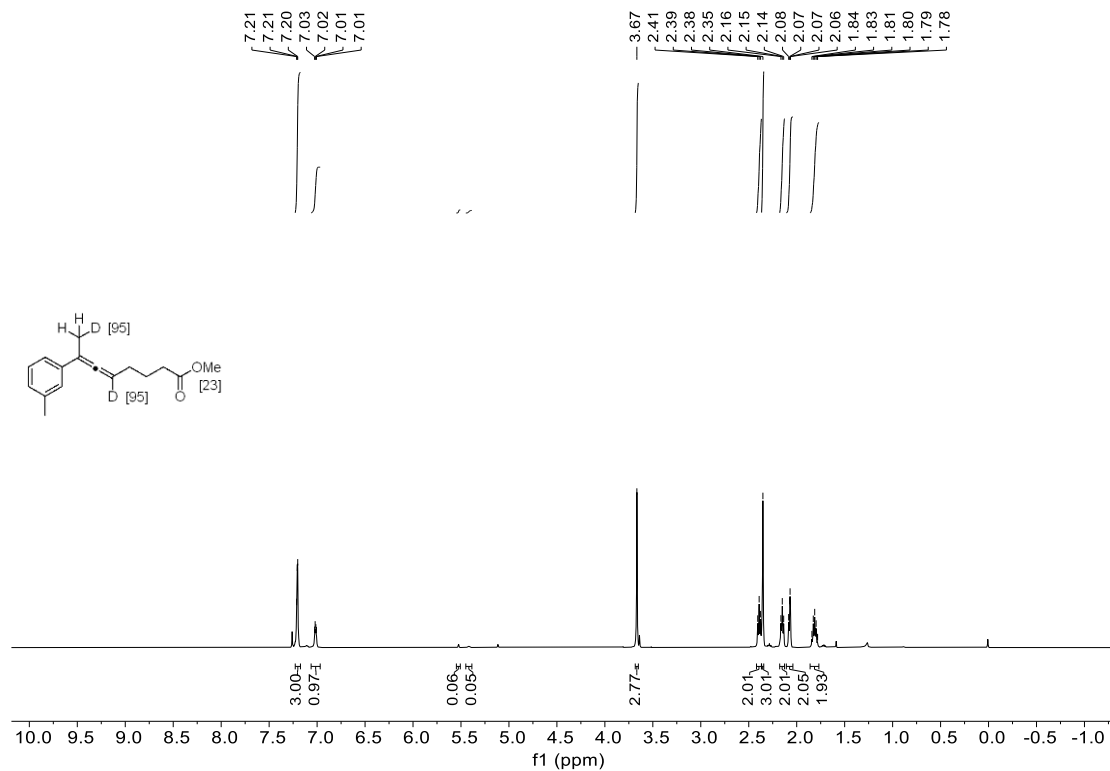
¹³C NMR (126 MHz, CDCl₃) spectra of 4d



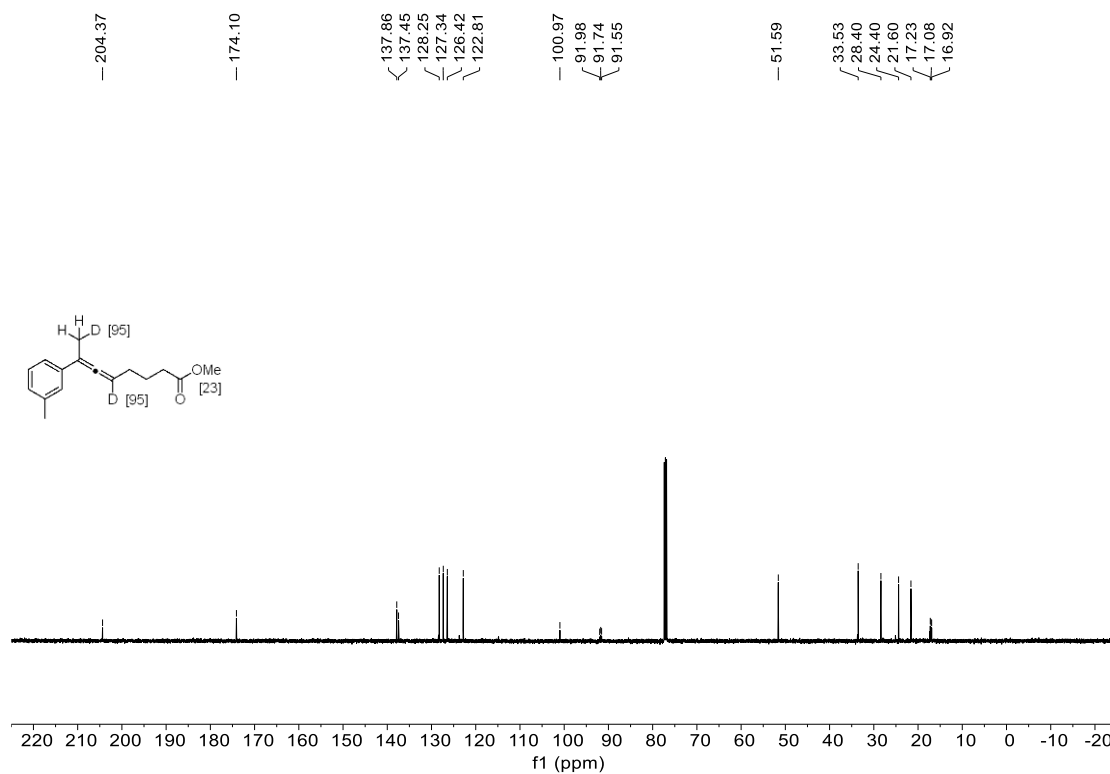
¹H NMR (500 MHz, CDCl₃) spectra of 5d



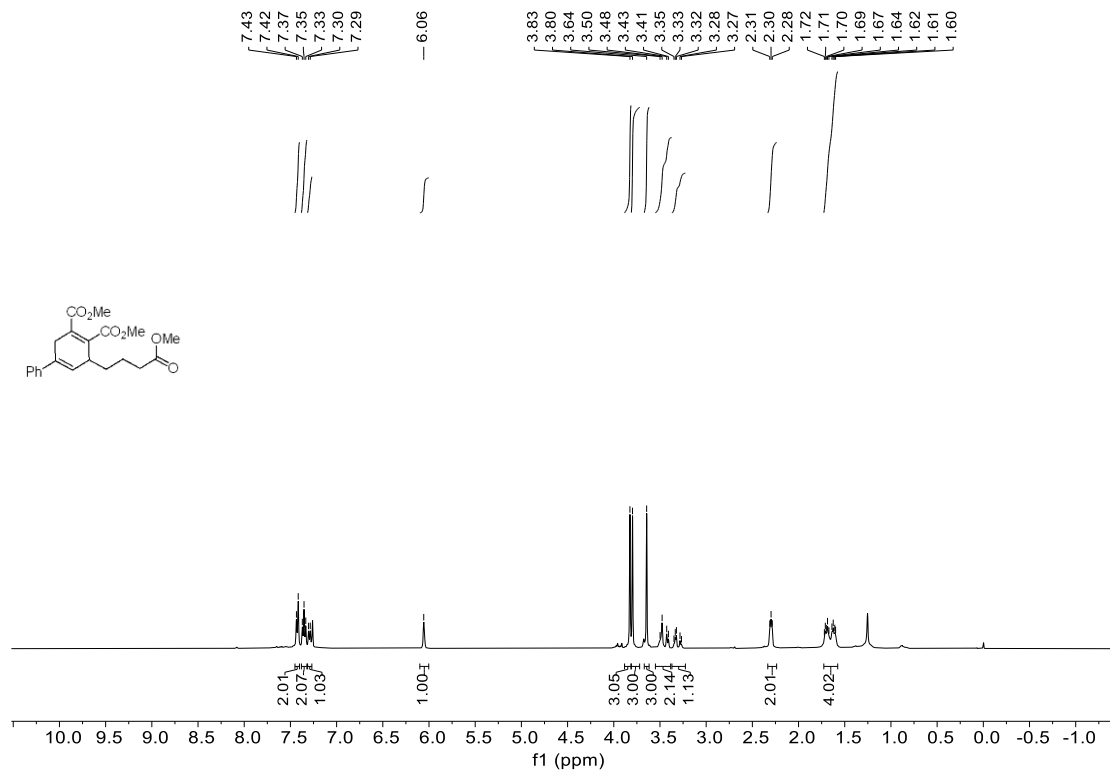
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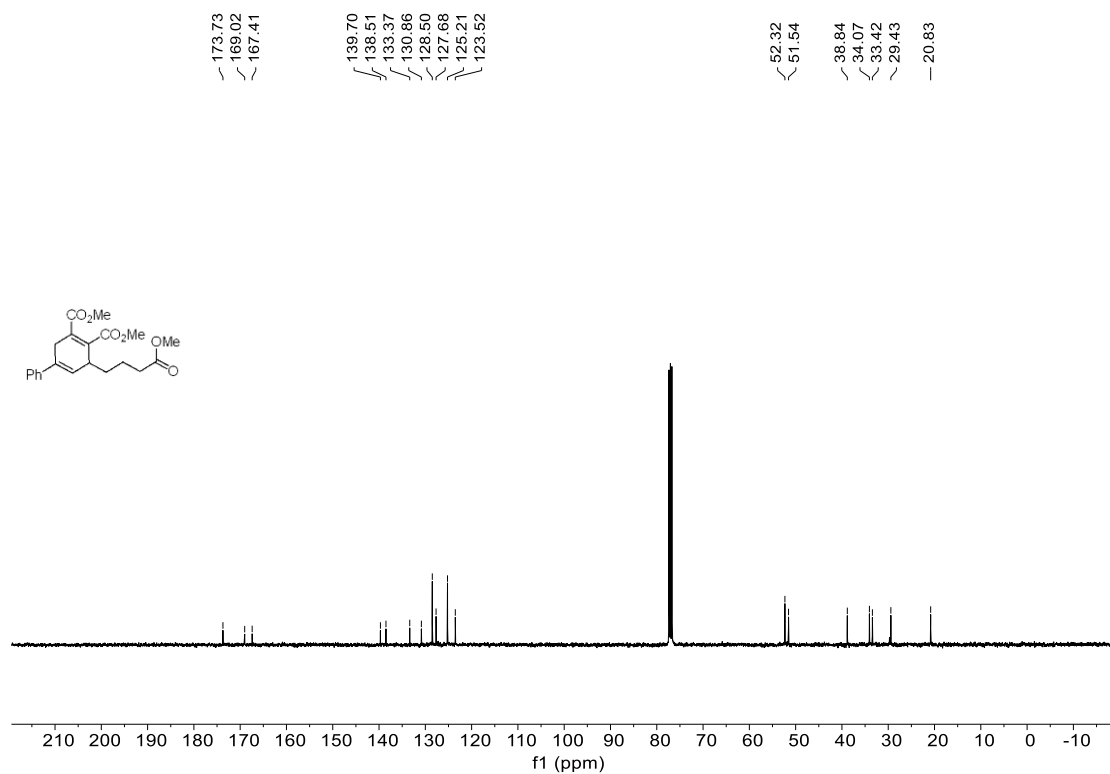
¹H NMR (500 MHz, CDCl₃) spectra of 6d



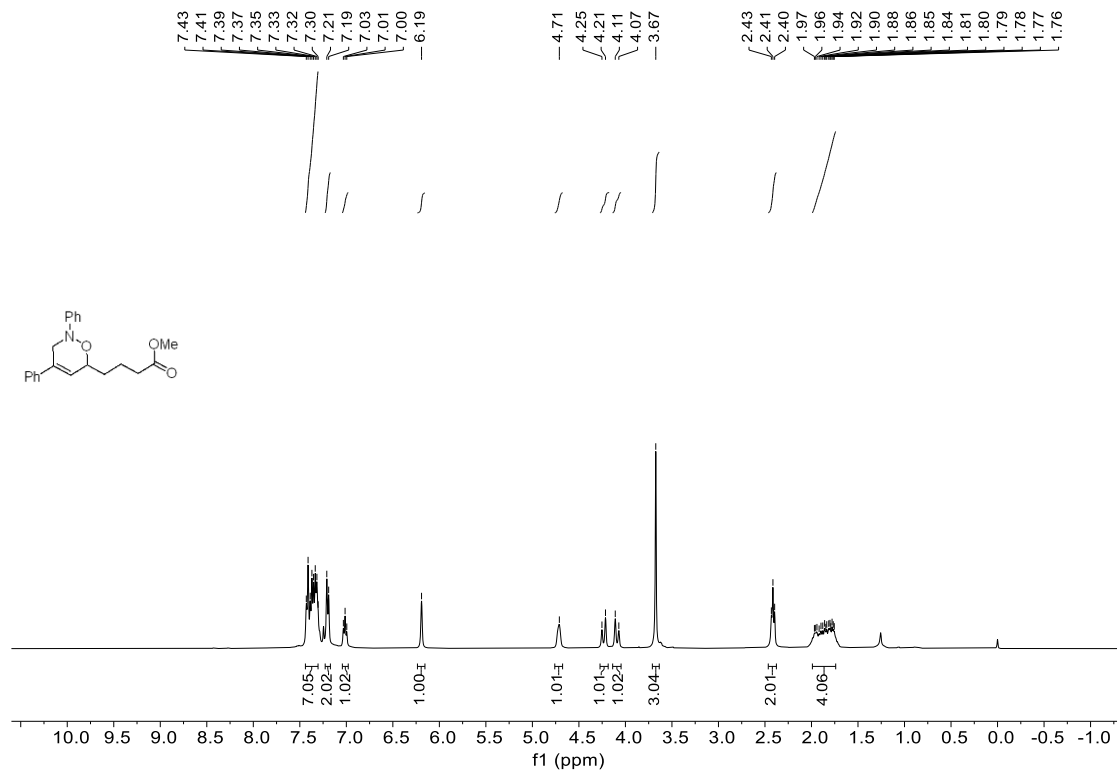
¹³C NMR (126 MHz, CDCl₃) spectra of 6d



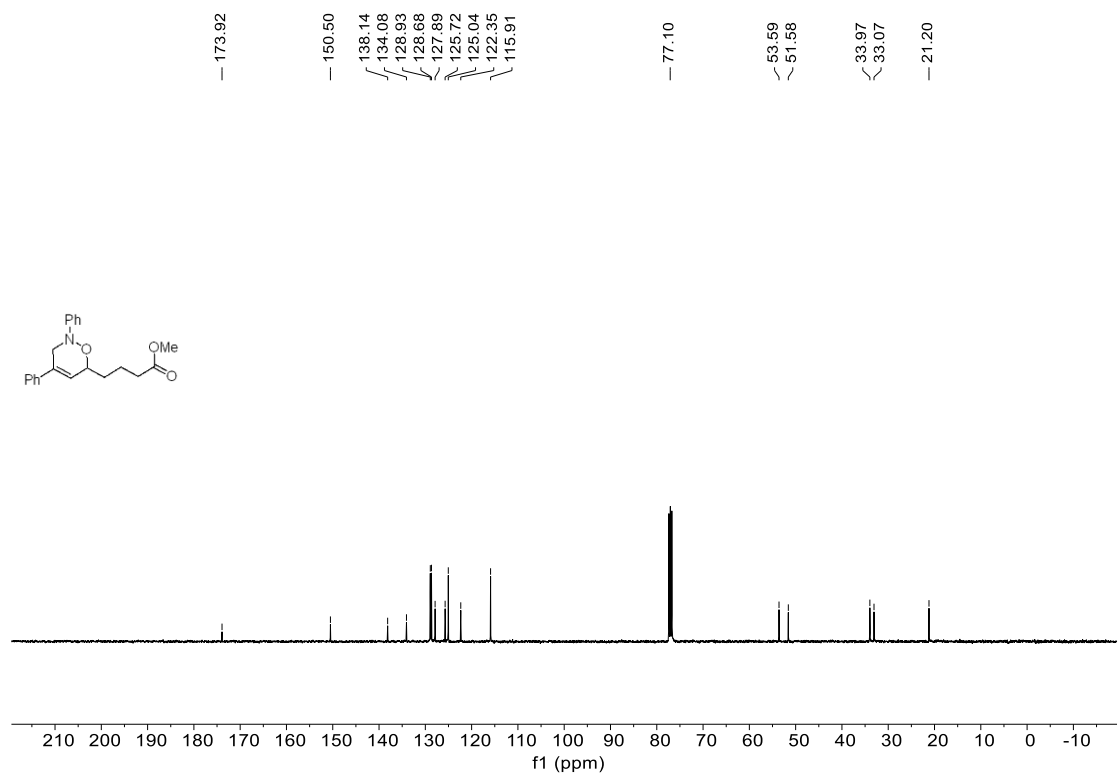
¹H NMR (400 MHz, CDCl₃) spectra of **22**



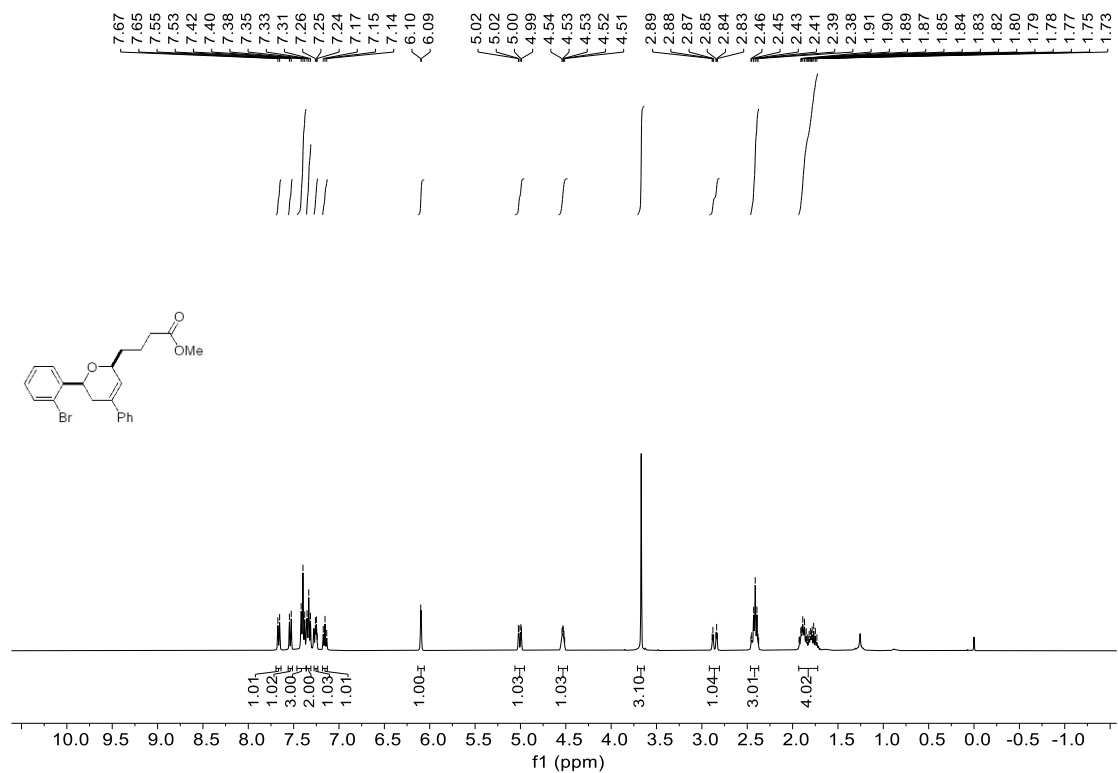
¹³C NMR (101 MHz, CDCl₃) spectra of **22**



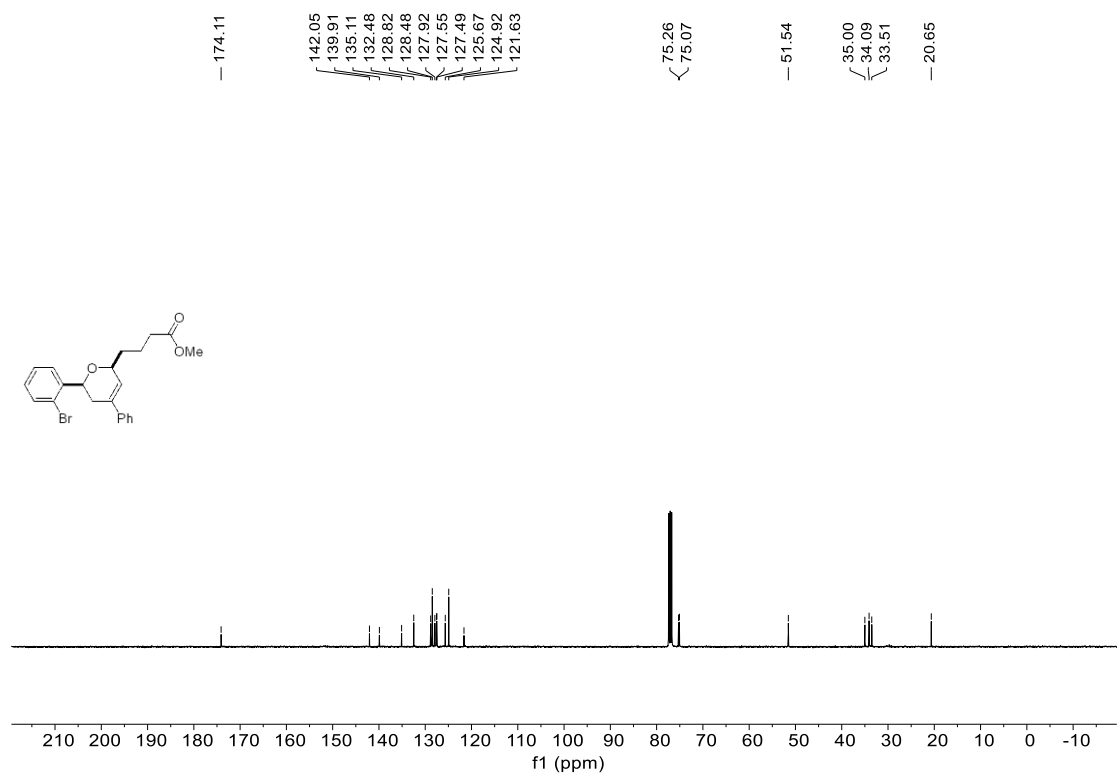
¹H NMR (400 MHz, CDCl₃) spectra of **23**



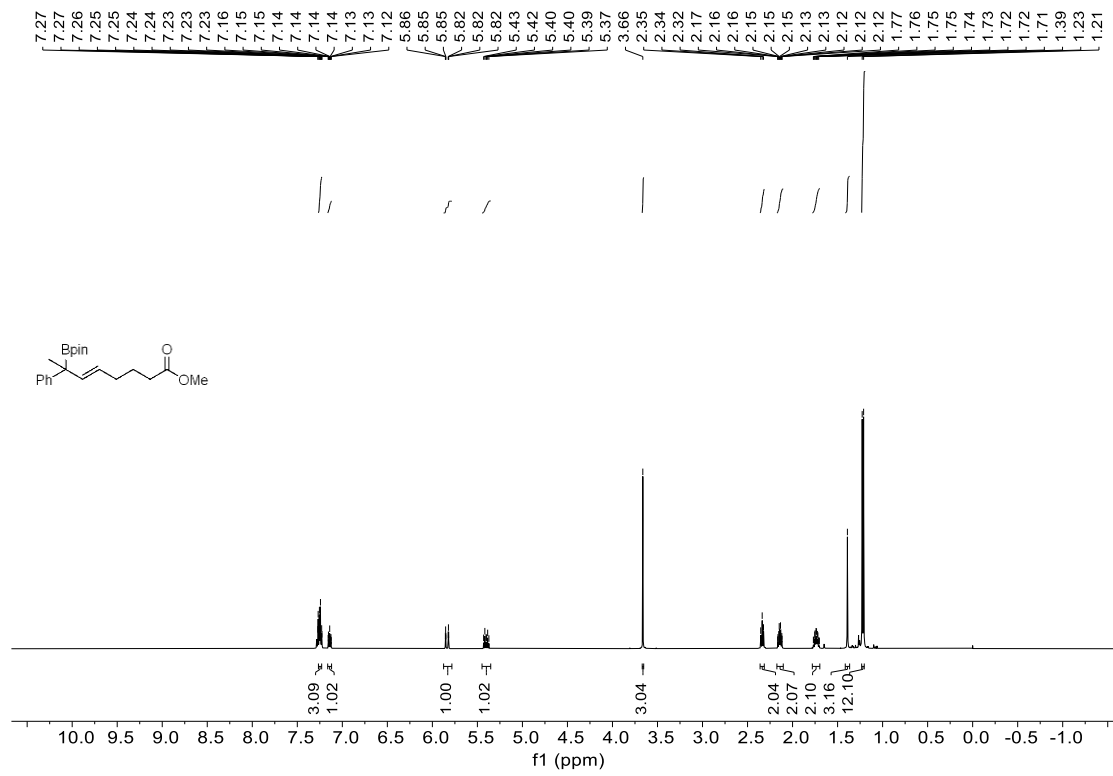
¹³C NMR (101 MHz, CDCl₃) spectra of **23**



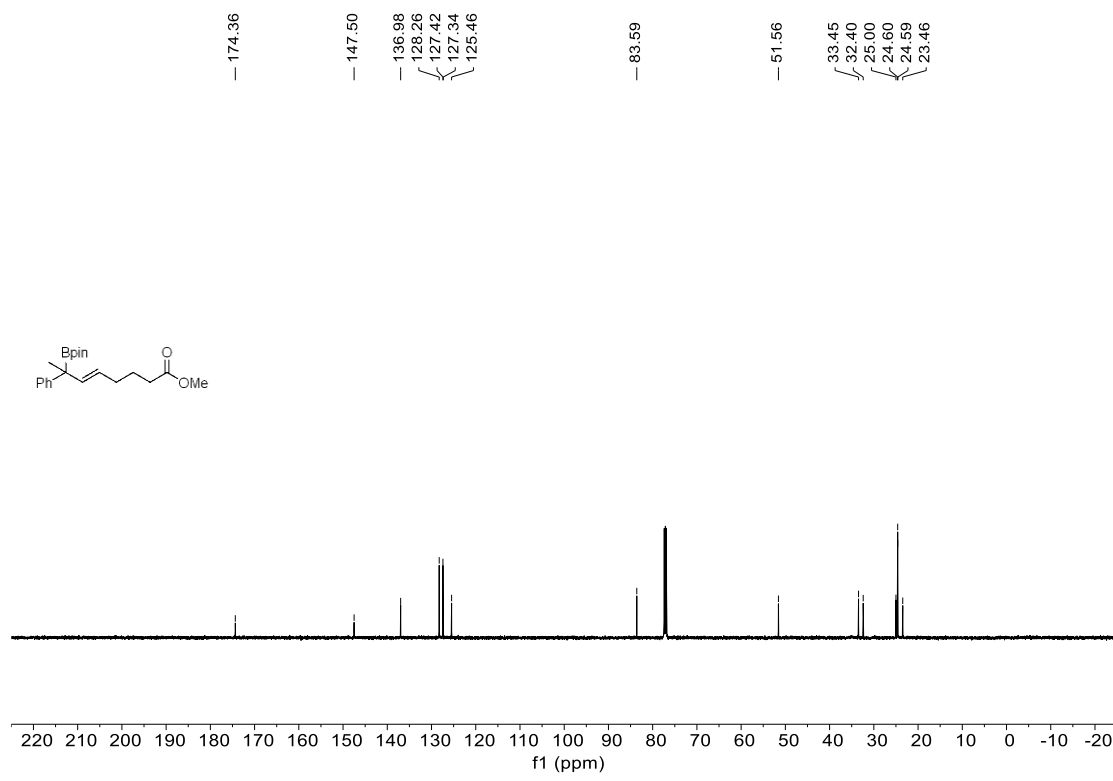
¹H NMR (400 MHz, CDCl₃) spectra of 24



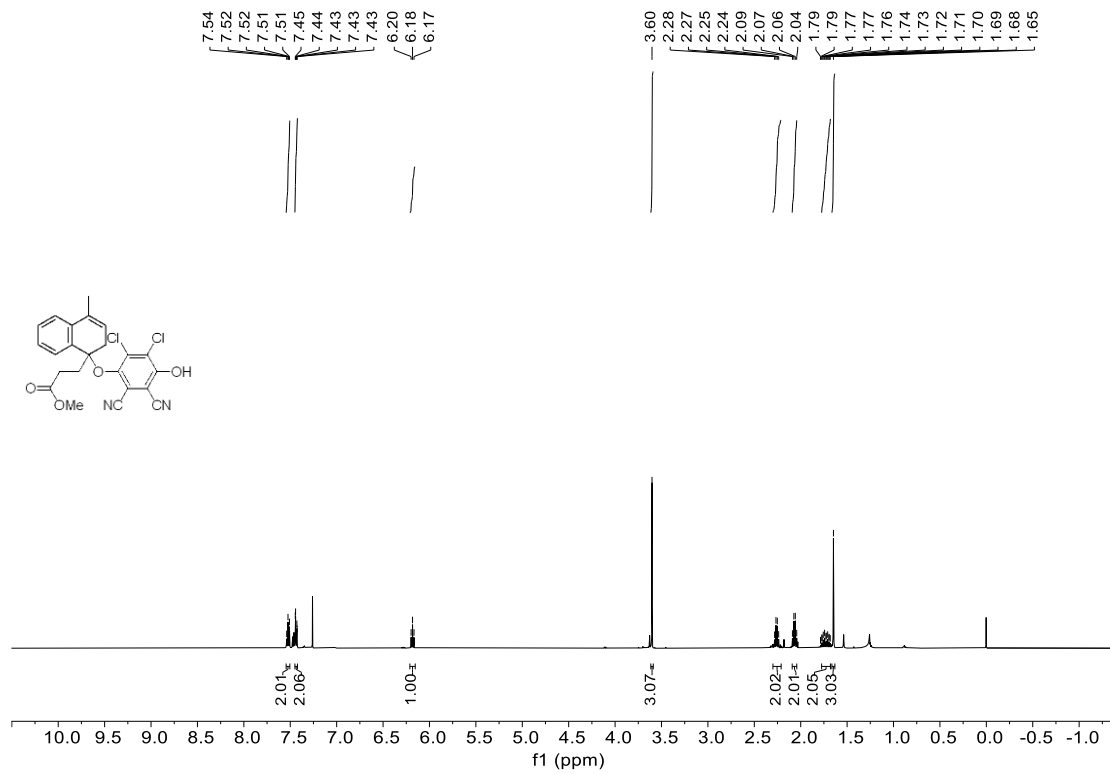
¹³C NMR (101 MHz, CDCl₃) spectra of 24



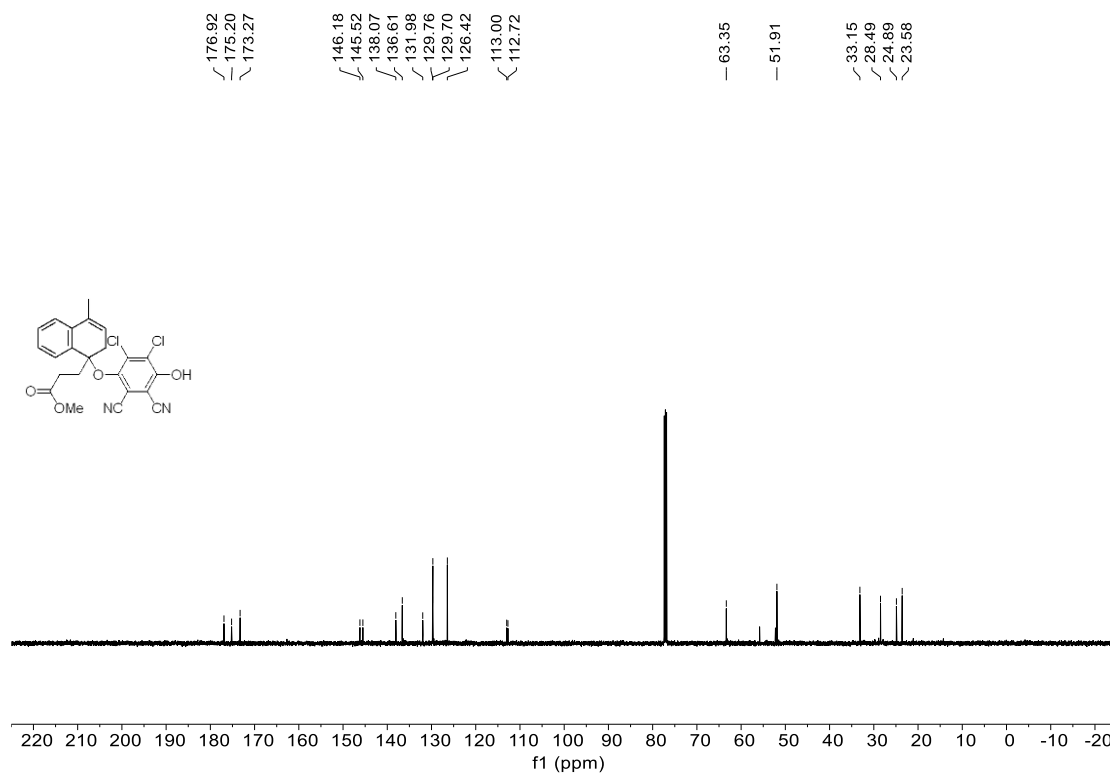
¹H NMR (400 MHz, CDCl₃) spectra of **25**



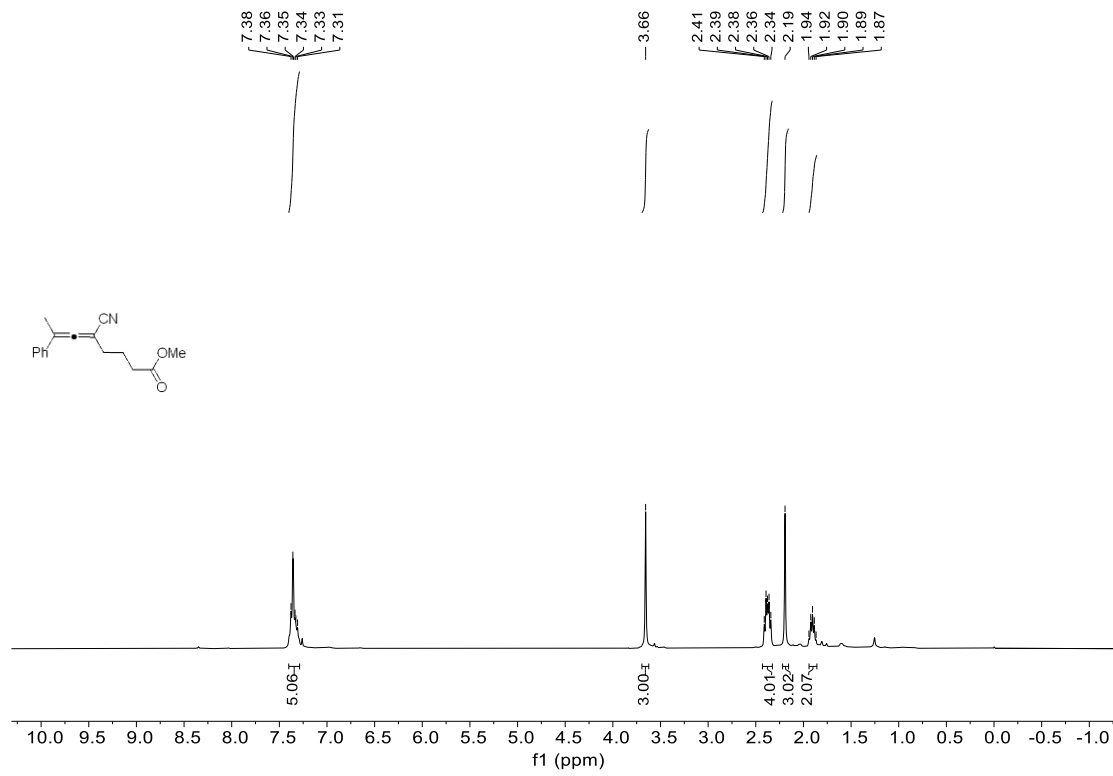
¹³C NMR (101 MHz, CDCl₃) spectra of **25**



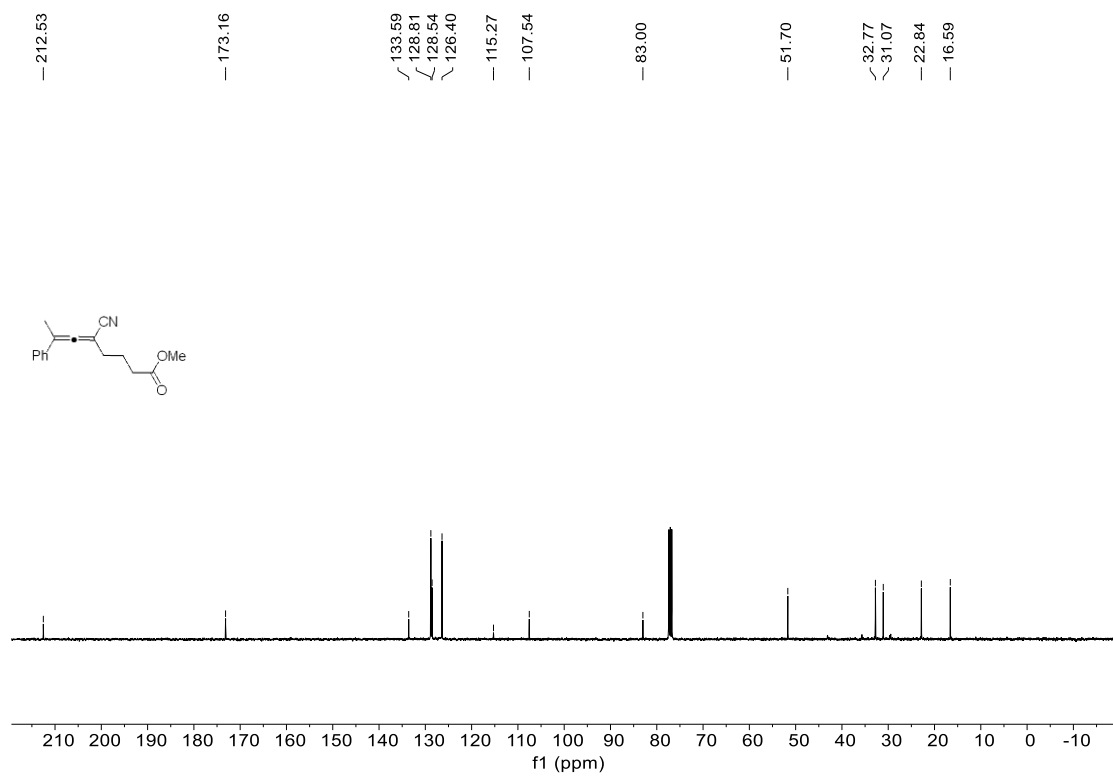
¹H NMR (400 MHz, CDCl₃) spectra of **26**



¹³C NMR (101 MHz, CDCl₃) spectra of **26**



¹H NMR (400 MHz, CDCl₃) spectra of **27**



¹³C NMR (101 MHz, CDCl₃) spectra of **27**