

Supplementary materials:

Not so dangerous? PET environmental-like microplastics and nanoplastics (PET-eIMNPs) effect on *Escherichia coli* and environmental bacteria

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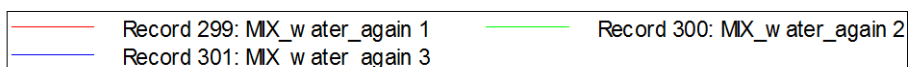
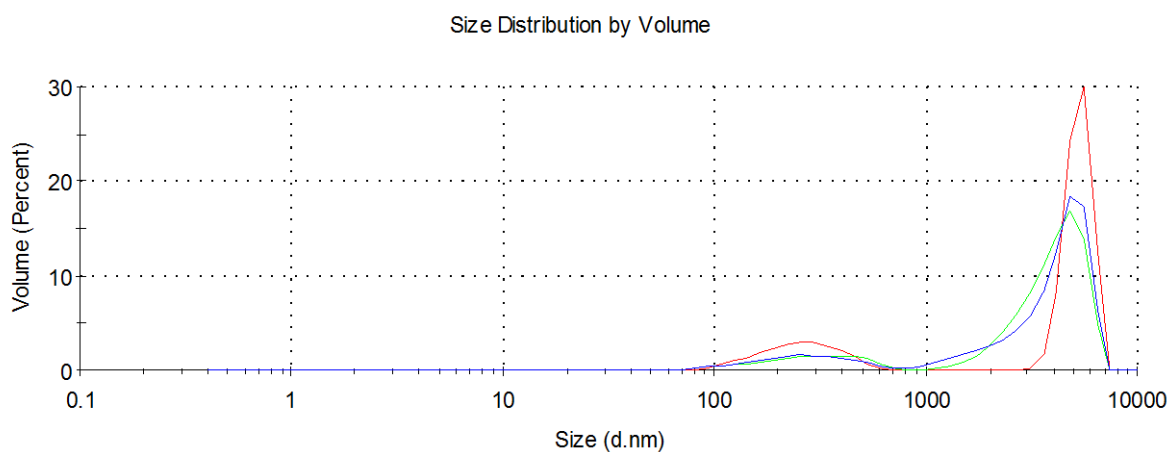
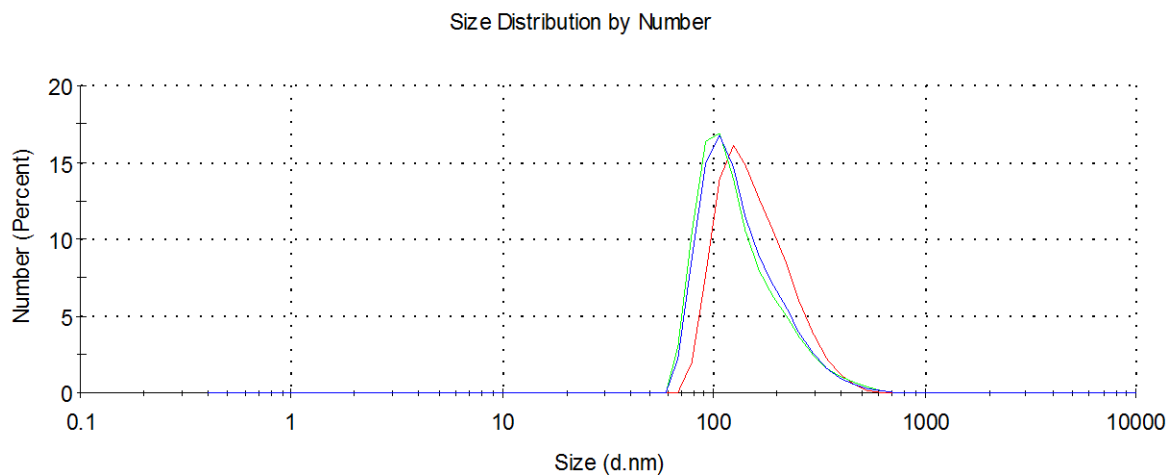
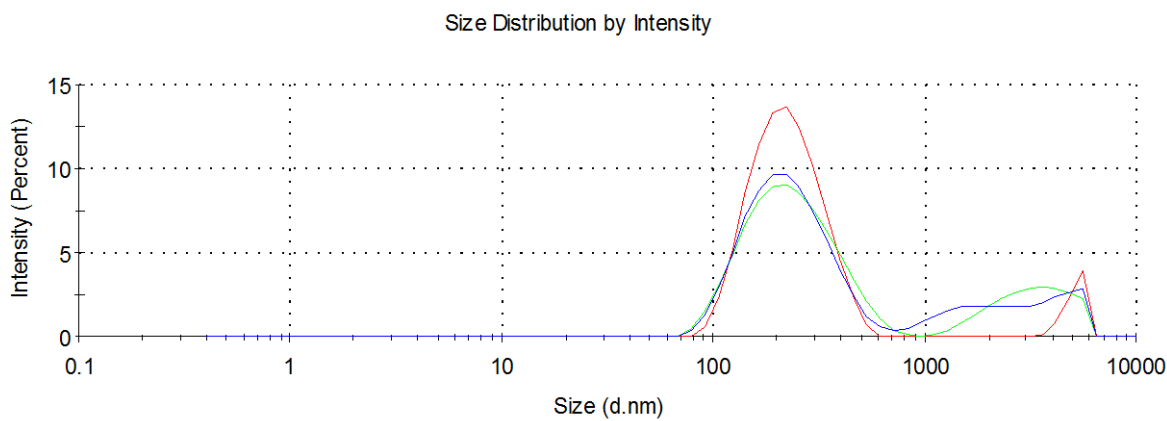
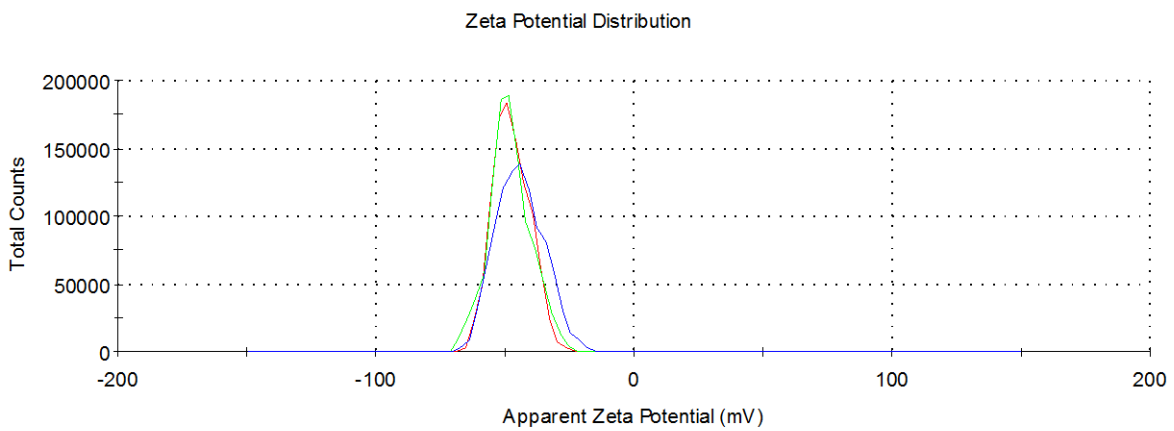
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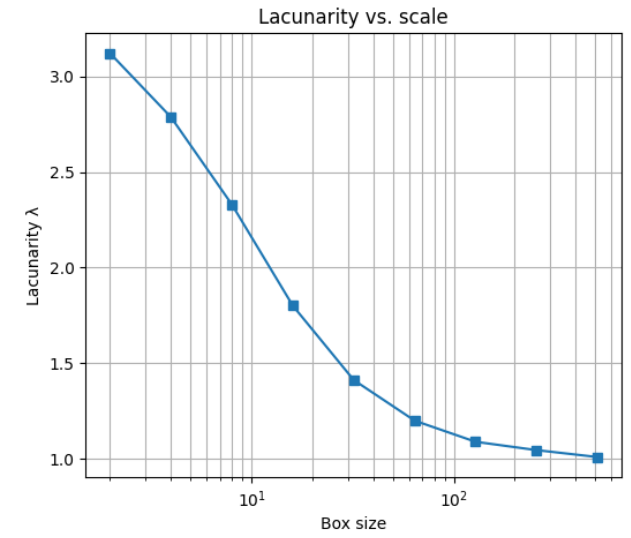
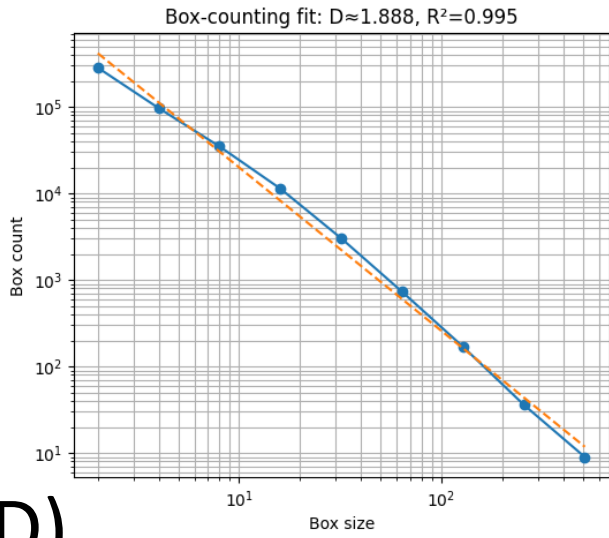
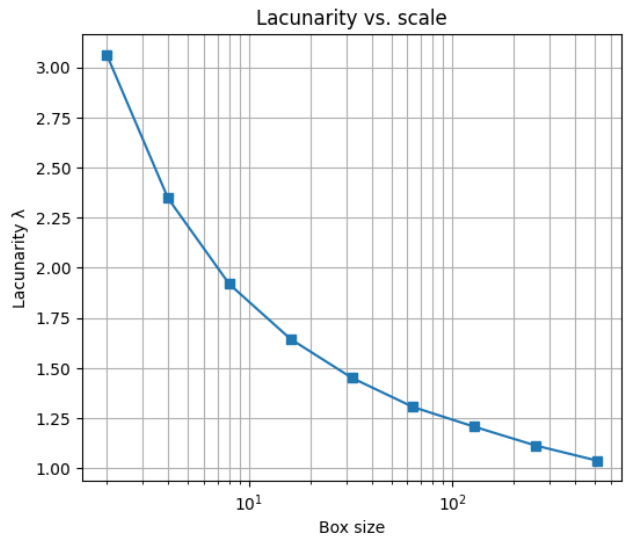
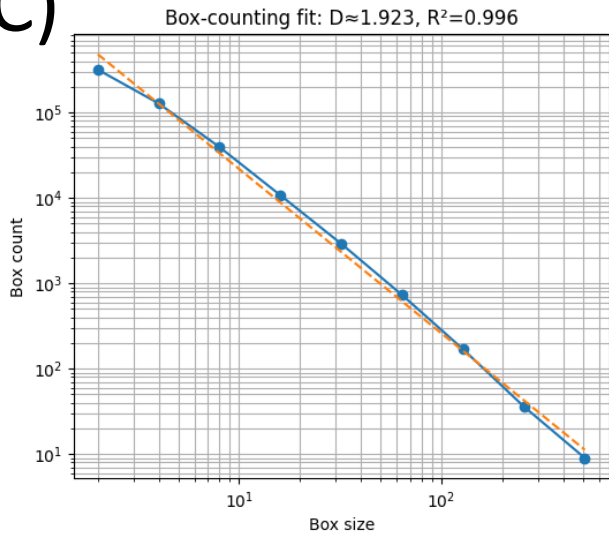
⁵Department of Agrochemistry, Soil Science, Microbiology and Plant Nutrition, Mendel University in Brno, Zemědělská 1665/1, 613 00, Brno, Czech Republic.

⁶Delong Instruments a.s., Palackého třída 3019/153b, 612 00, Brno, Czech Republic

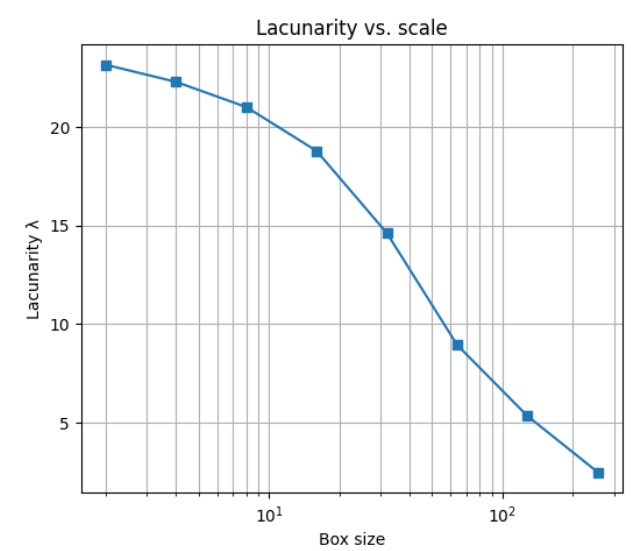
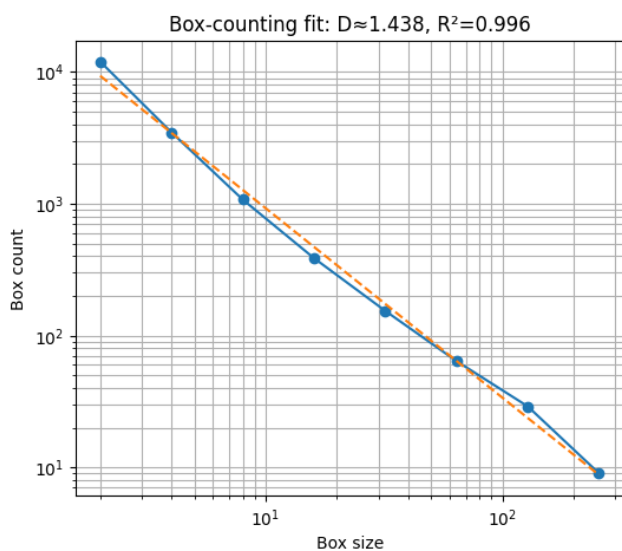
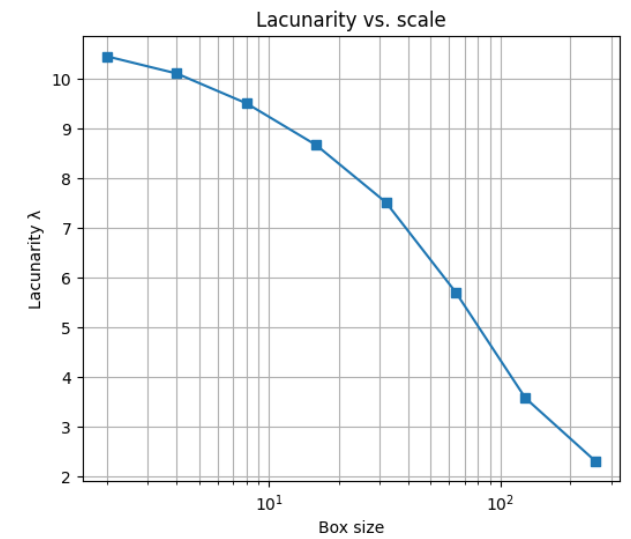
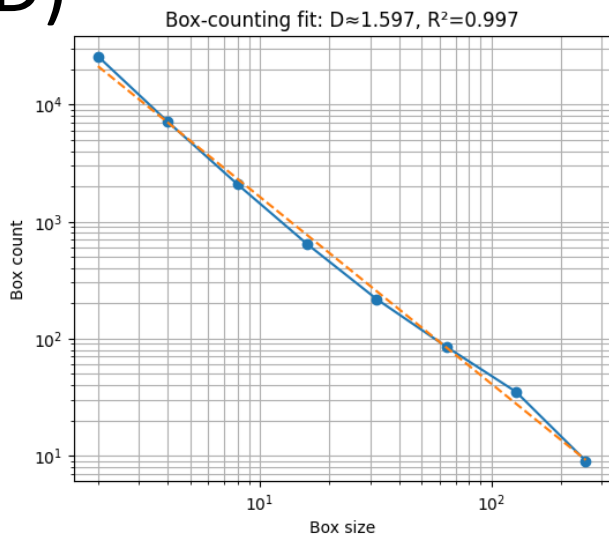


S1: Zeta-potential and DLS results (Intensity, Number, Volume size distribution). Settings: refractive index 1.575, adsorption coefficient 0.160, temperature 25 °C, equilibration time 120 s, measurement angle of 173° backscatter

C)

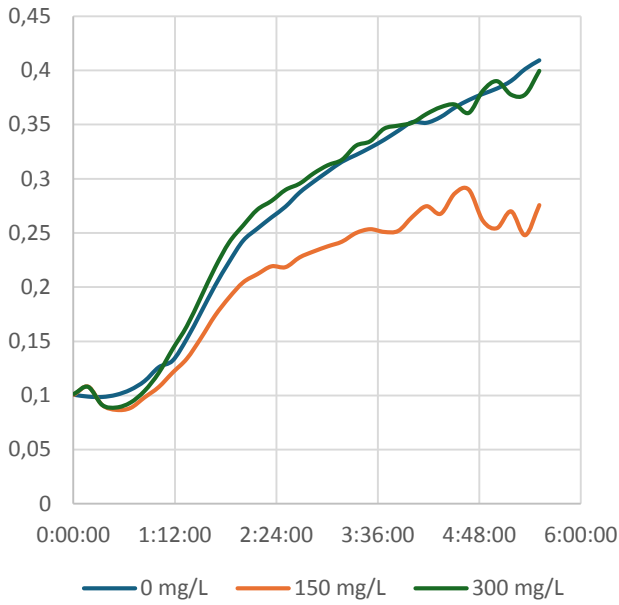


D)

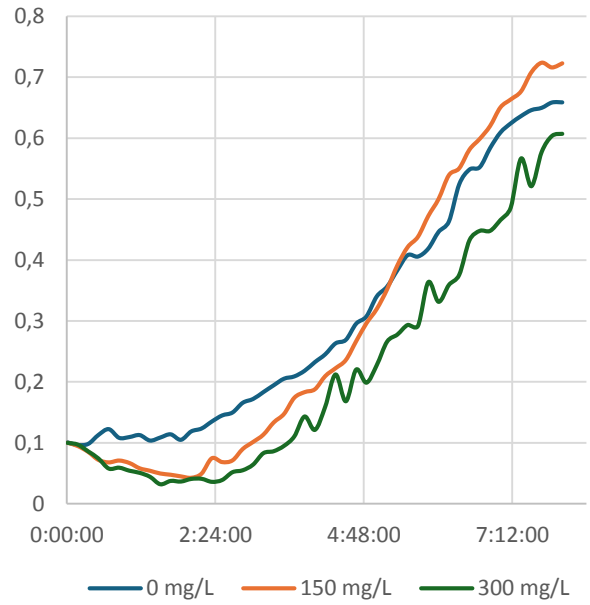


S2: C) SEM images fractal dimension and lacunarity, D) TEM images fractal dimension and lacunarity calculations

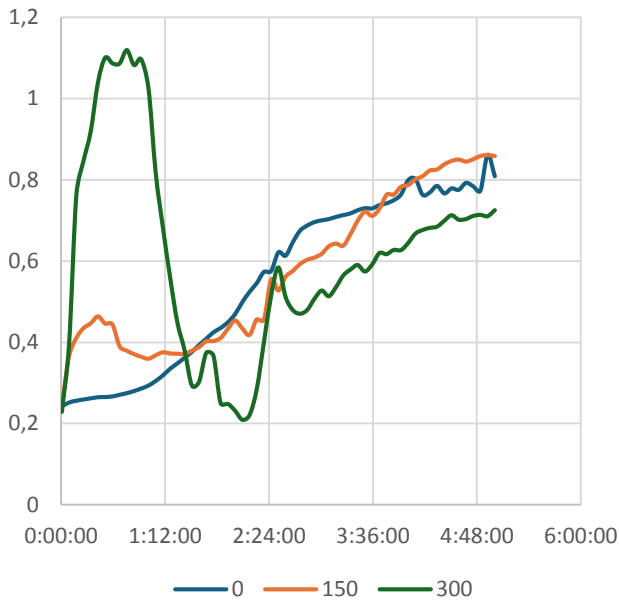
Escherichia coli



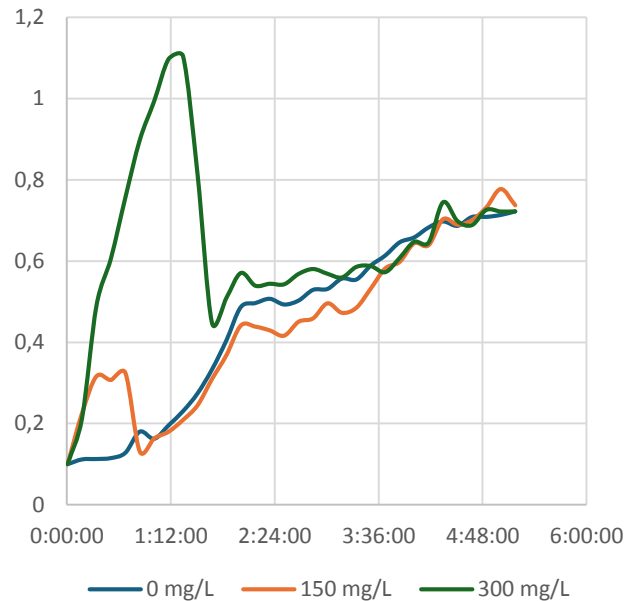
Pseudoaltermonas rubra



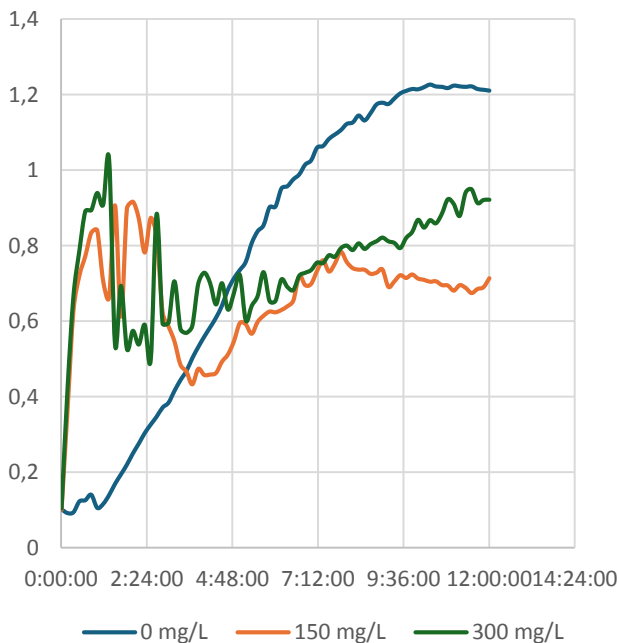
Vibrio natriegens



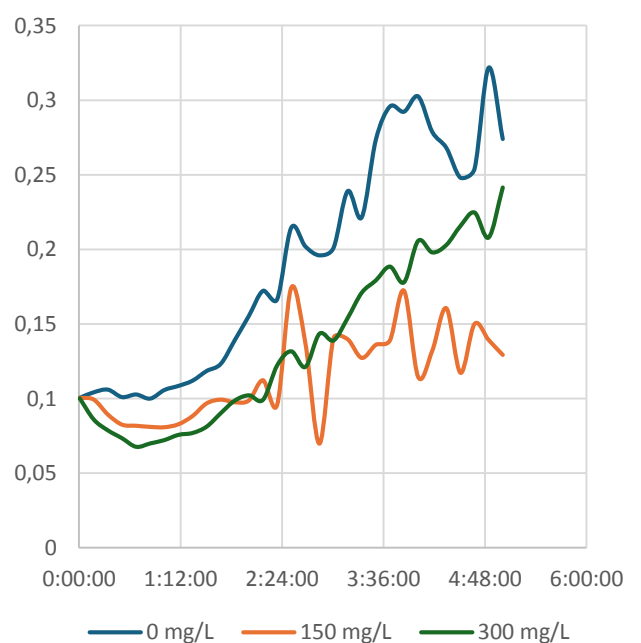
Vibrio natriegens - repeat



Curtobacterium oceanosedimentum



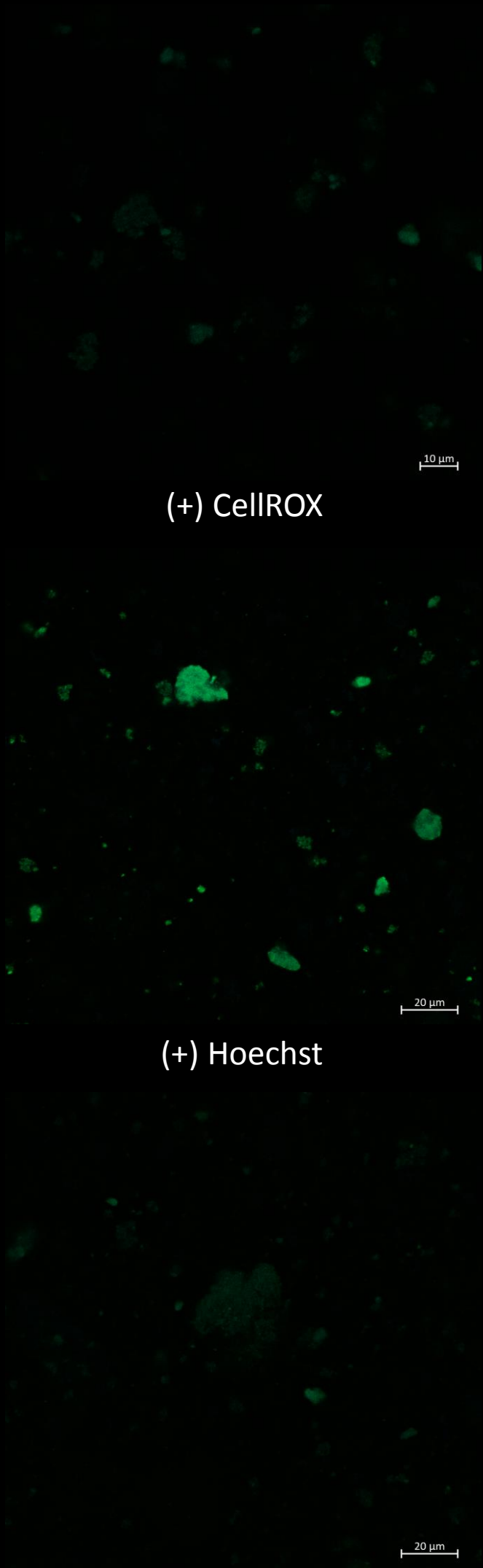
Bacillus mycoides



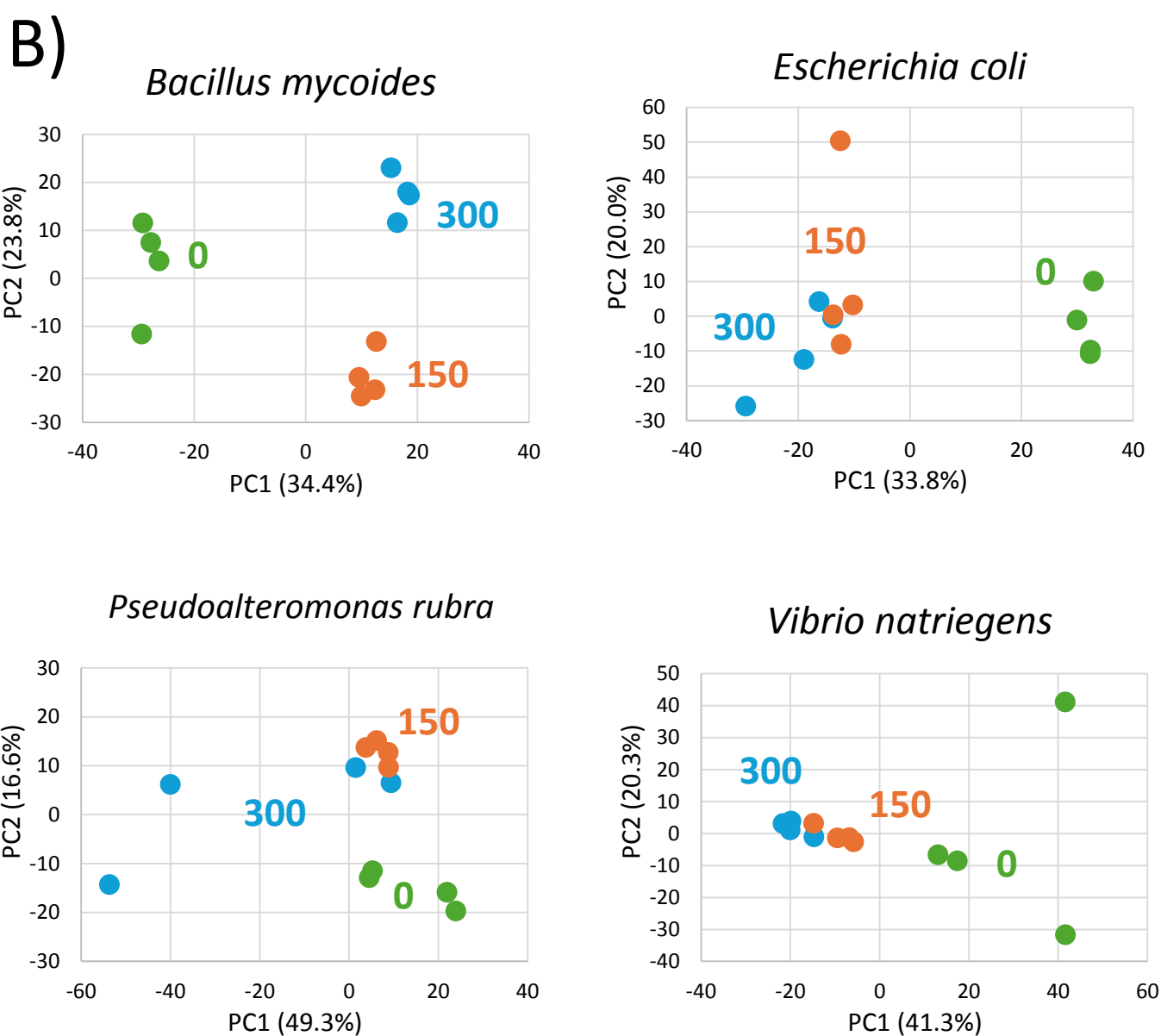
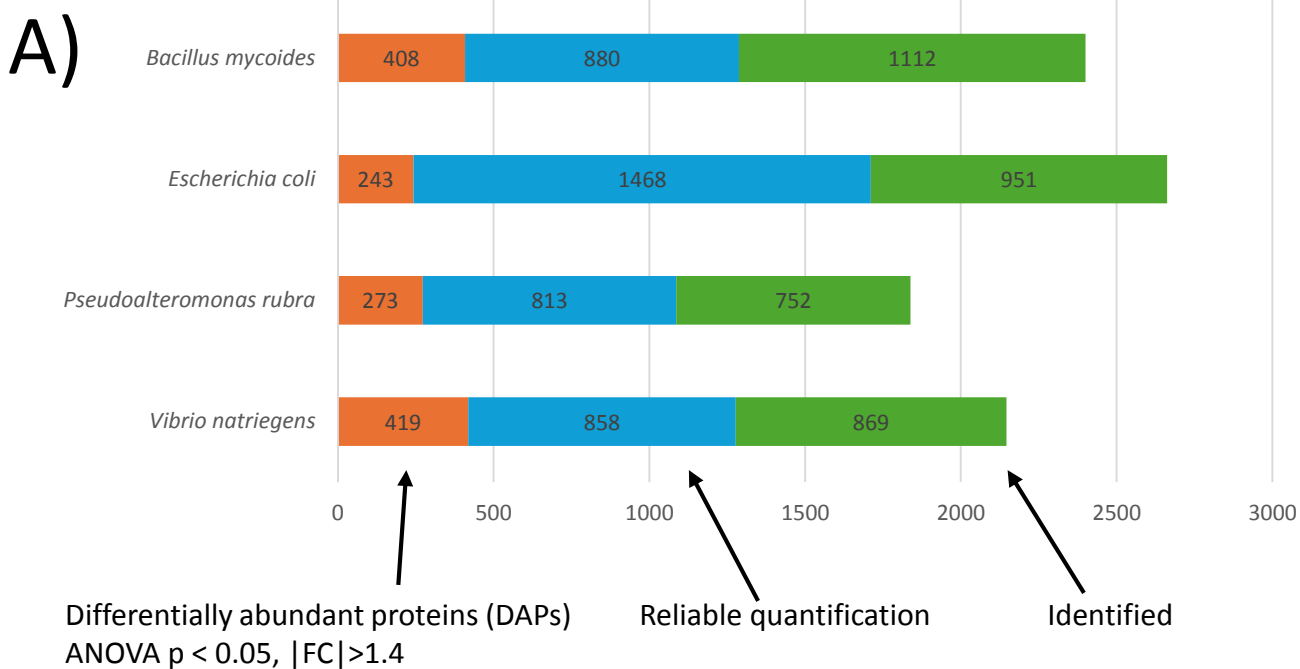
S3: Growth patterns from proteome/confocal samples. Settings are following: 30 °C (*E. coli* 37 °C), shaking duration 30 s, shaking amplitude 4 mm, wavelength 600 nm, interval 10 minutes, number of replicates: 4

A) (-) Hoechst + CellROX: control

B) (-) PI + FDA: control



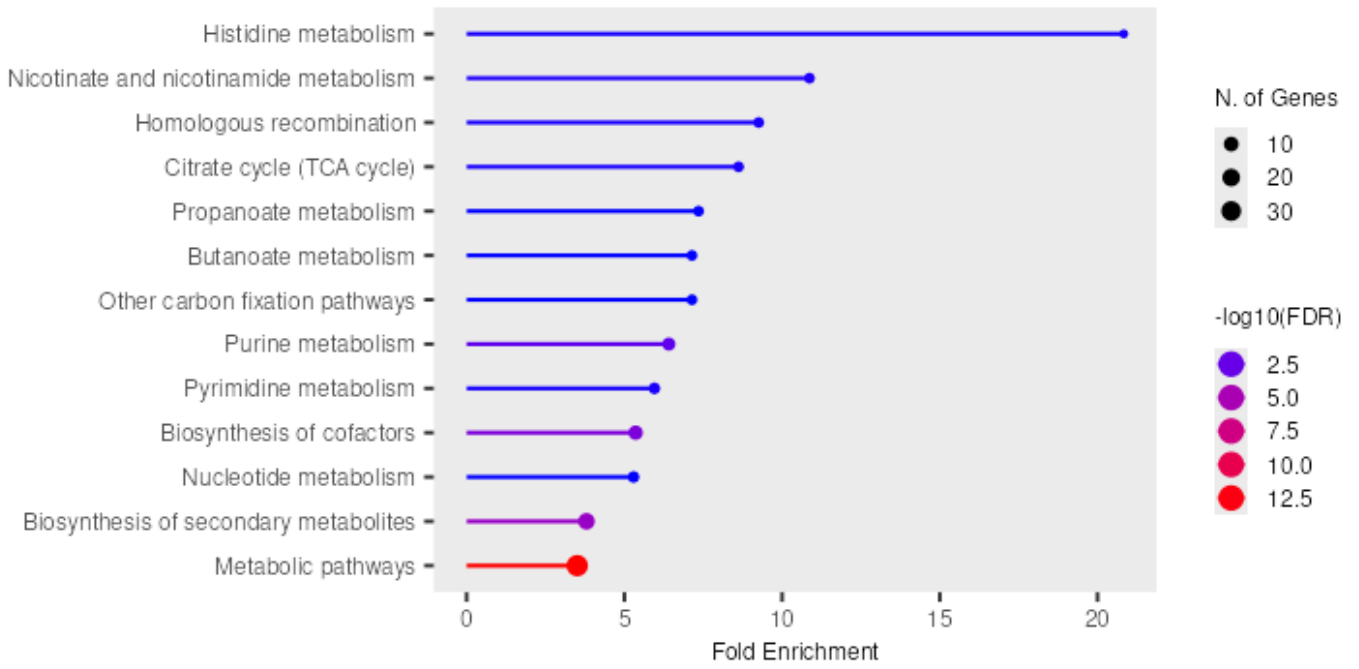
S4: Negative control for MNPs measurements: A) (up-down line) H + ROX: control (no staining), CellROX staining, Hoechst staining, B) (up-down line) FDA + PI: control (no staining), FDA staining, PI staining



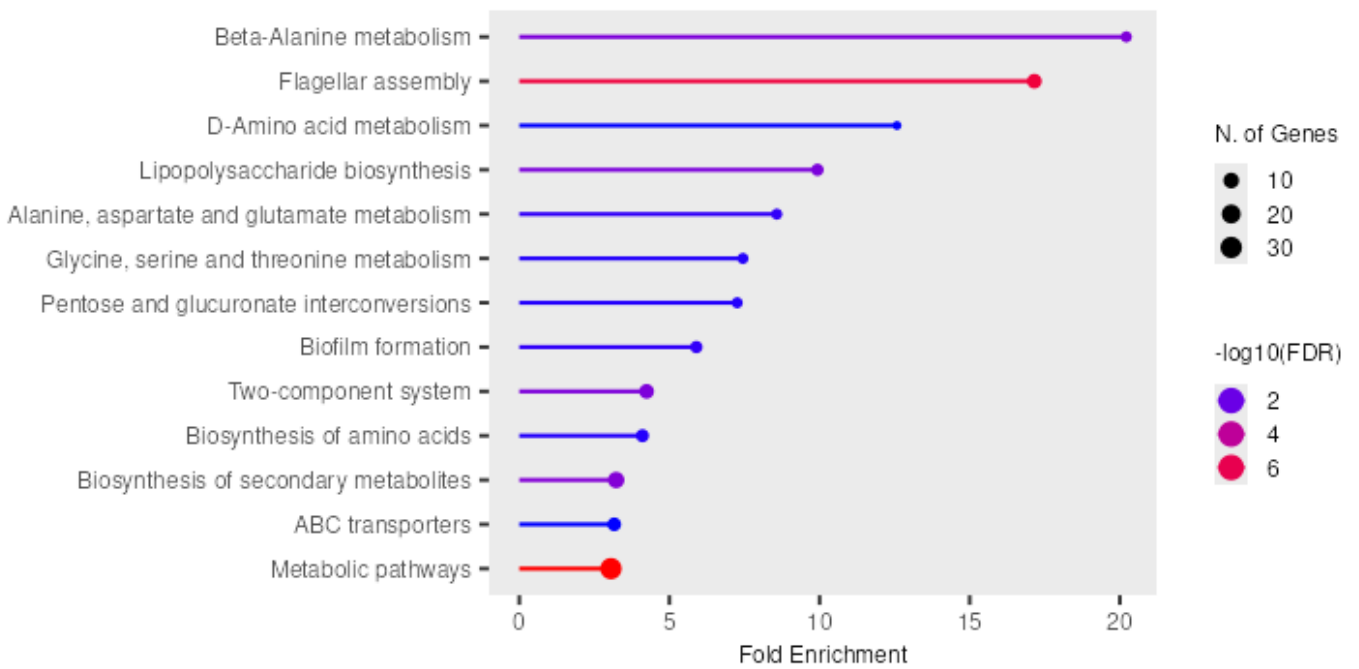
S5: Proteomic parameters: A) Protein count overview – DAPs, reliable quantified proteins, identified but non-quantified proteins. B) Principal component analysis (PCA) of bacteria.

Escherichia coli

A) DOWN



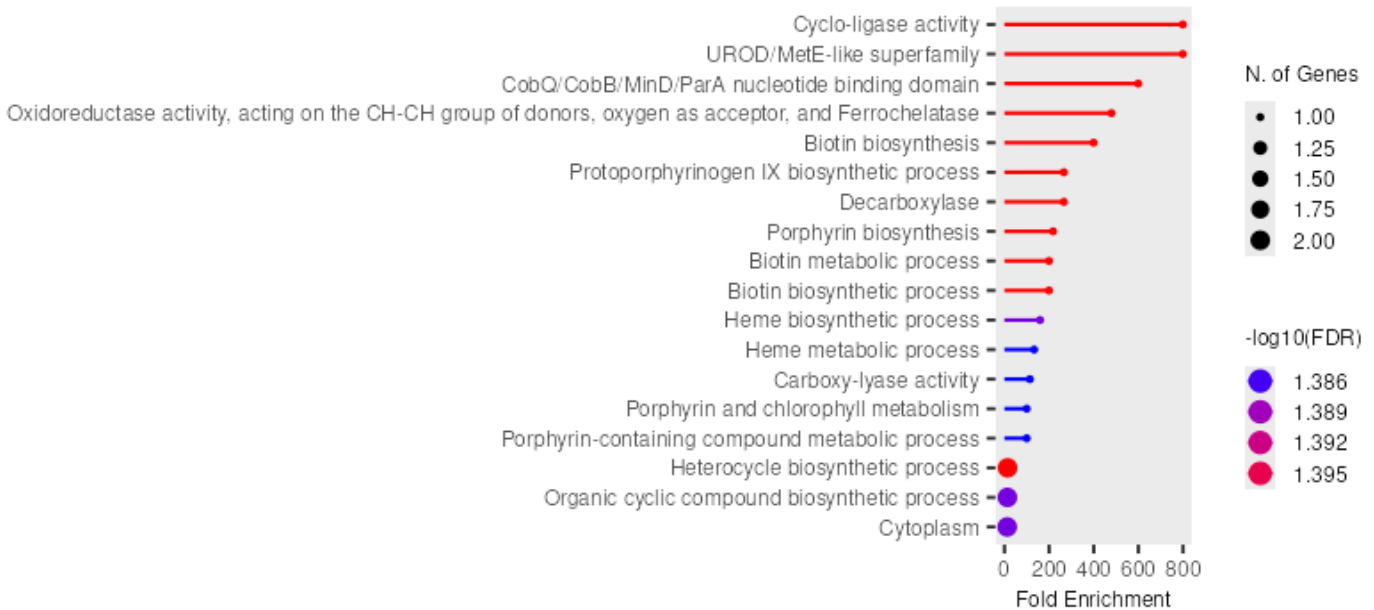
B) UP



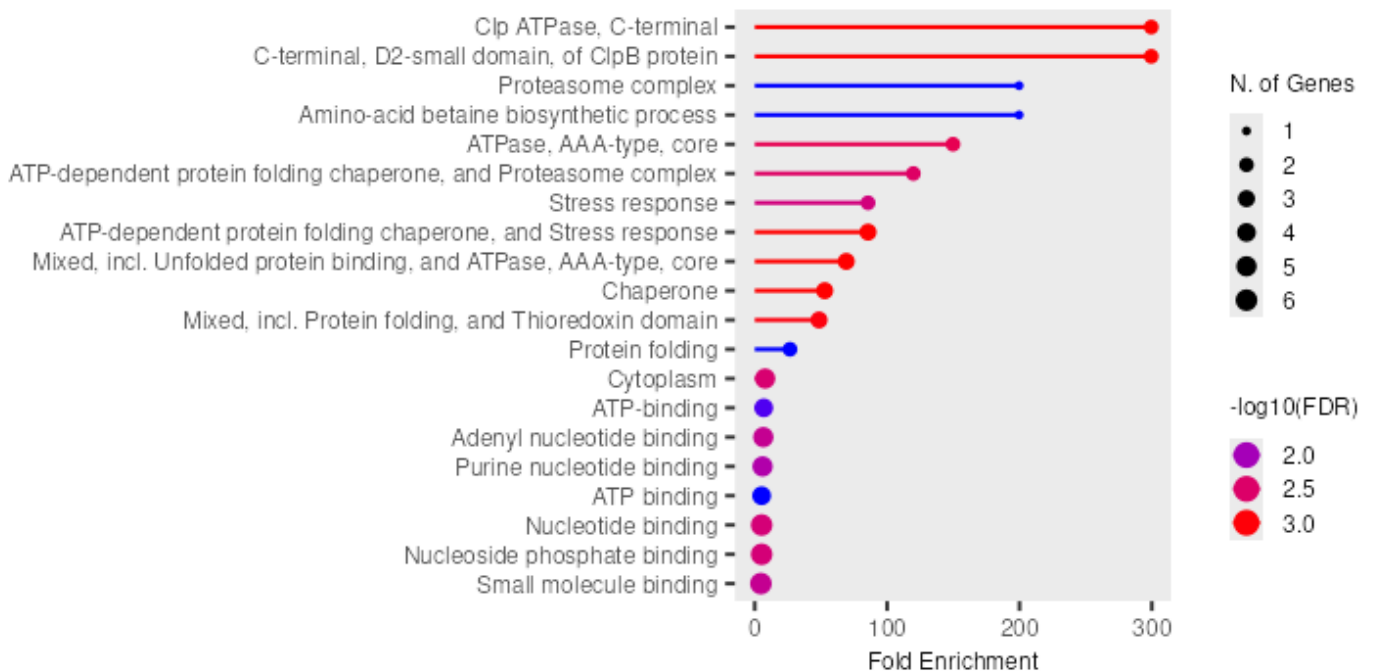
S6: ShinyGO gene ontology and enrichment for Escherichia coli K12: a) statistically significant different decreased protein abundance data set (DOWN) derived pathways, b) statistically significant different increased protein abundance data set (UP) derived pathways

Vibrio natriegens

A) DOWN



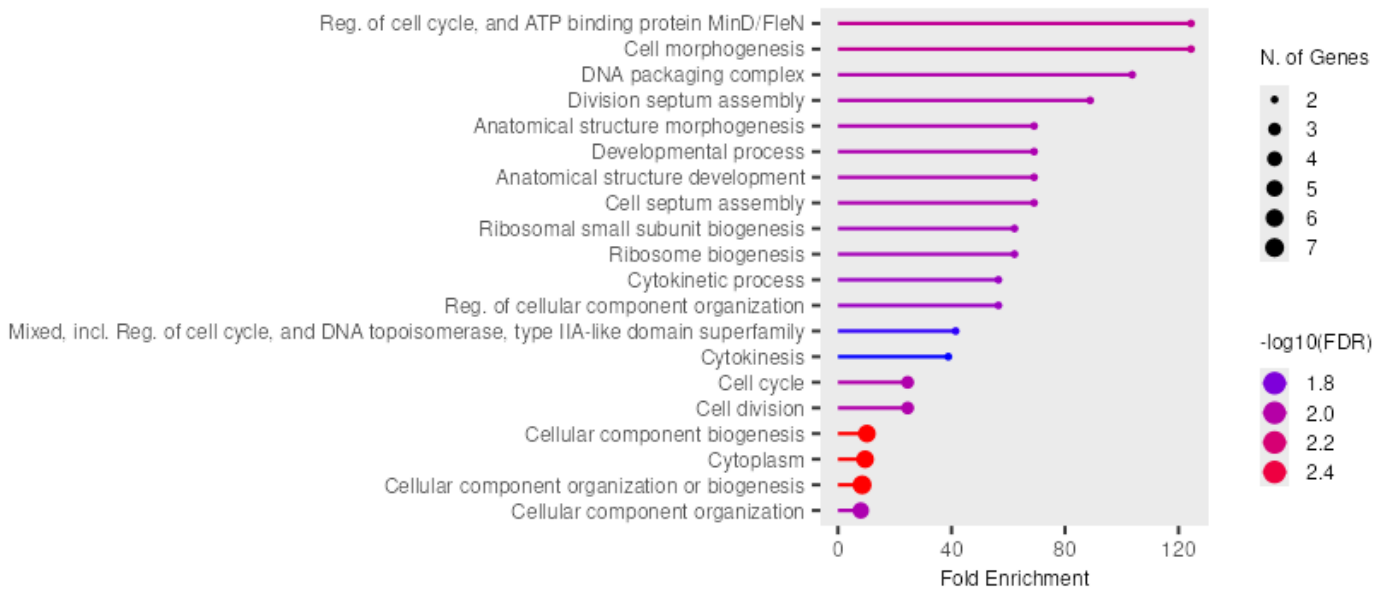
B) UP



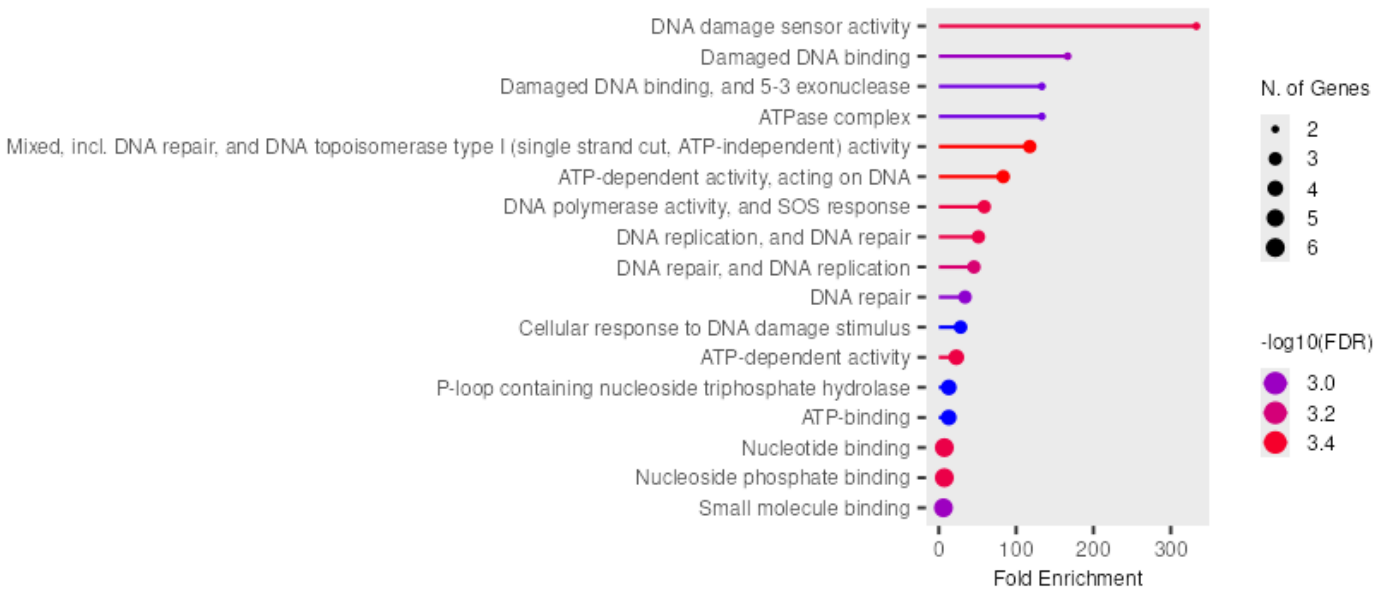
S7: ShinyGO gene ontology and enrichment for *Vibrio natriegens*: a) statistically significant different decreased protein abundance data set (DOWN) derived pathways, b) statistically significant different increased protein abundance data set (UP) derived pathways

Pseudoalteromonas rubra

A) DOWN



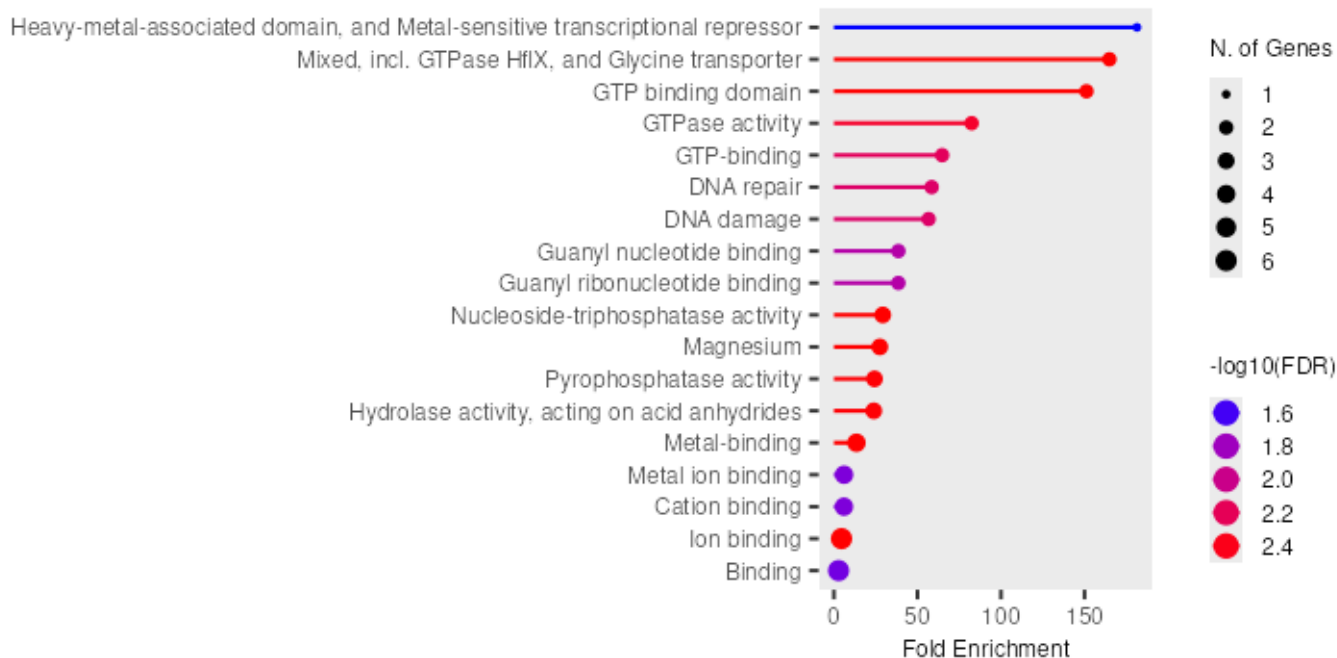
B) UP



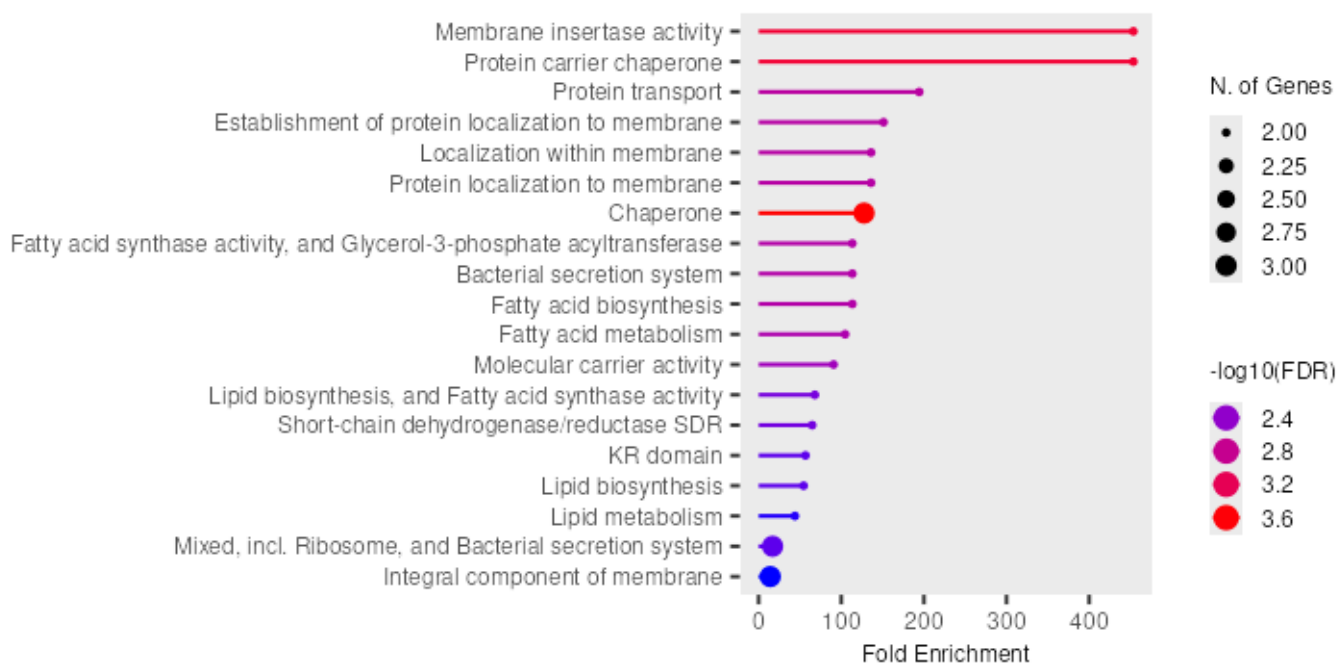
S8: ShinyGO gene ontology and enrichment for *Pseudoalteromonas rubra*: a) statistically significant different decreased protein abundance data set (DOWN) derived pathways, b) statistically significant different increased protein abundance data set (UP) derived pathways

Bacillus mycooides

A) DOWN

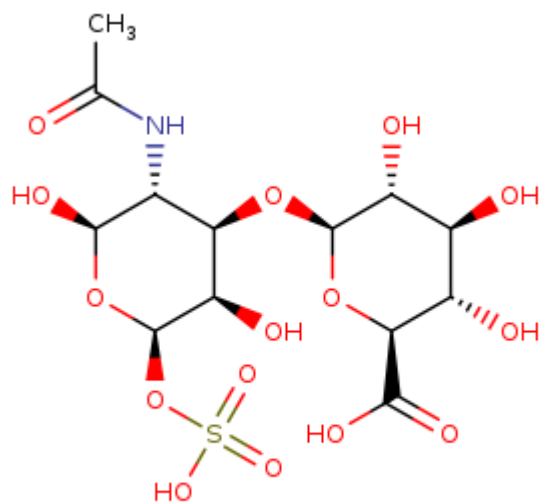


B) UP

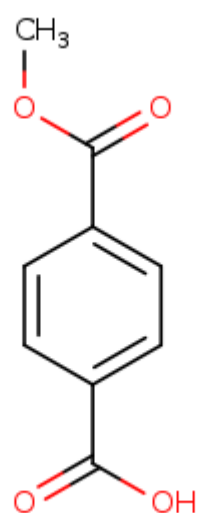


S9: ShinyGO gene ontology and enrichment for *Bacillus mycooides*: a) decreased statistically significant different protein abundance data set (DOWN) derived pathways, b) statistically significant different increased protein abundance data set (UP) derived pathways

A)



B)



Model	Calculated affinity (kcal/mol)
1	-5.020
2	-4.982
3	-4.908
4	-4.749
5	-4.632
6	-4.629
7	-4.505
8	-4.493
9	-4.392
10	-4.379
11	-4.304
12	-4.291
13	-4.266
14	-4.239
15	-4.235
16	-4.195
17	-4.115
18	-4.111
19	-4.073
20	-4.067

Model	Calculated affinity (kcal/mol)
1	-4.419
2	-4.400
3	-4.394
4	-4.291
5	-4.289
6	-4.271
7	-4.257
8	-4.257
9	-4.230
10	-4.216
11	-4.204
12	-4.203
13	-4.200
14	-4.183
15	-4.113
16	-4.097
17	-4.015
18	-3.960
19	-3.945
20	-3.932

S10: AutoDock Vina binding affinities for chondroitin AC-lyase: A) Native chondroitin-sulfate monomer substrate binding affinity energy. B) MHET (monohydroxy-ethylene-terephthalate) substrate binding affinity energy.