

Supplementary Information

Mechanistic insights into target searching, de-dimerization, and activation of *Mucilaginibacter paludis* Argonaute

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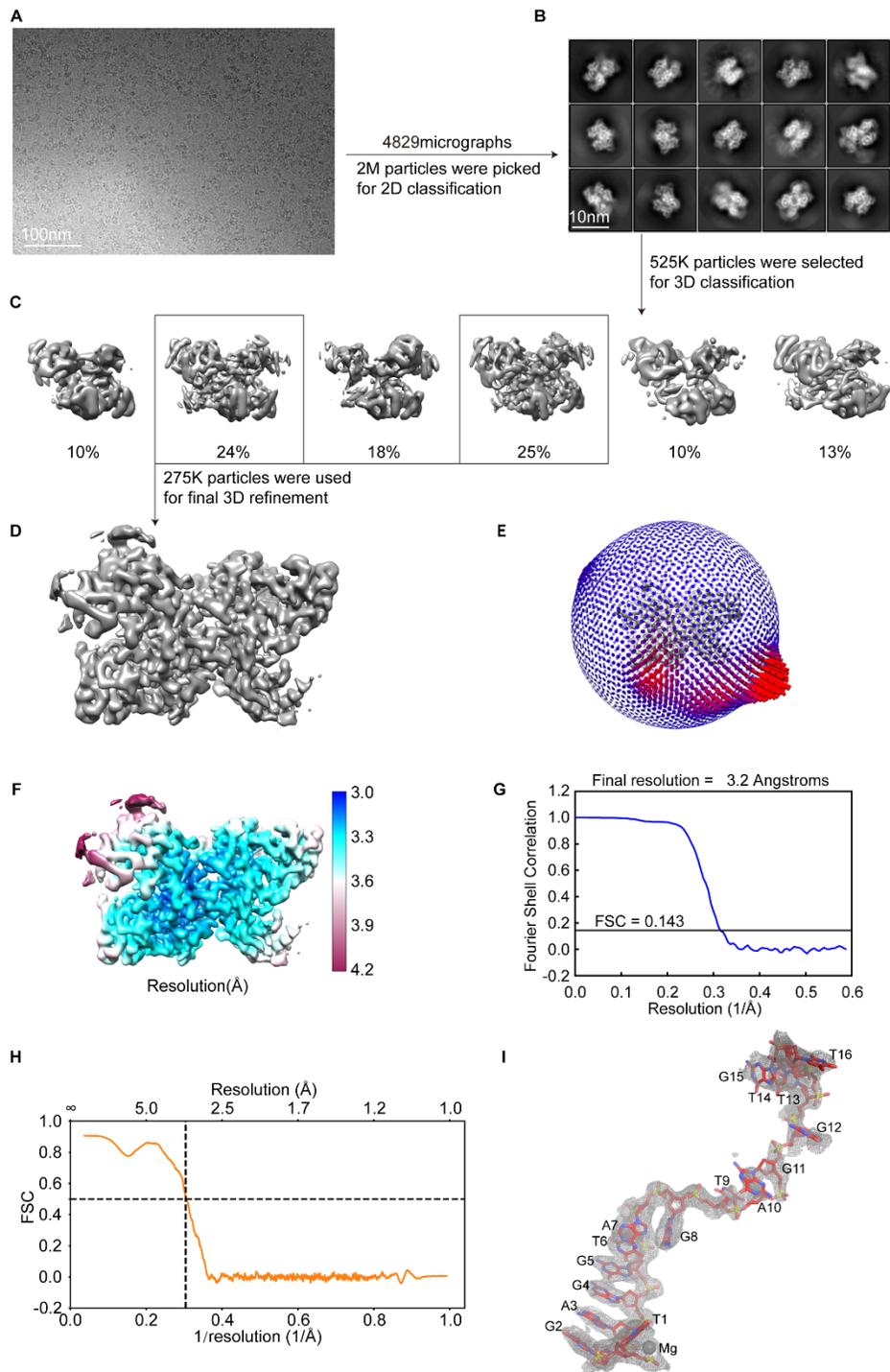
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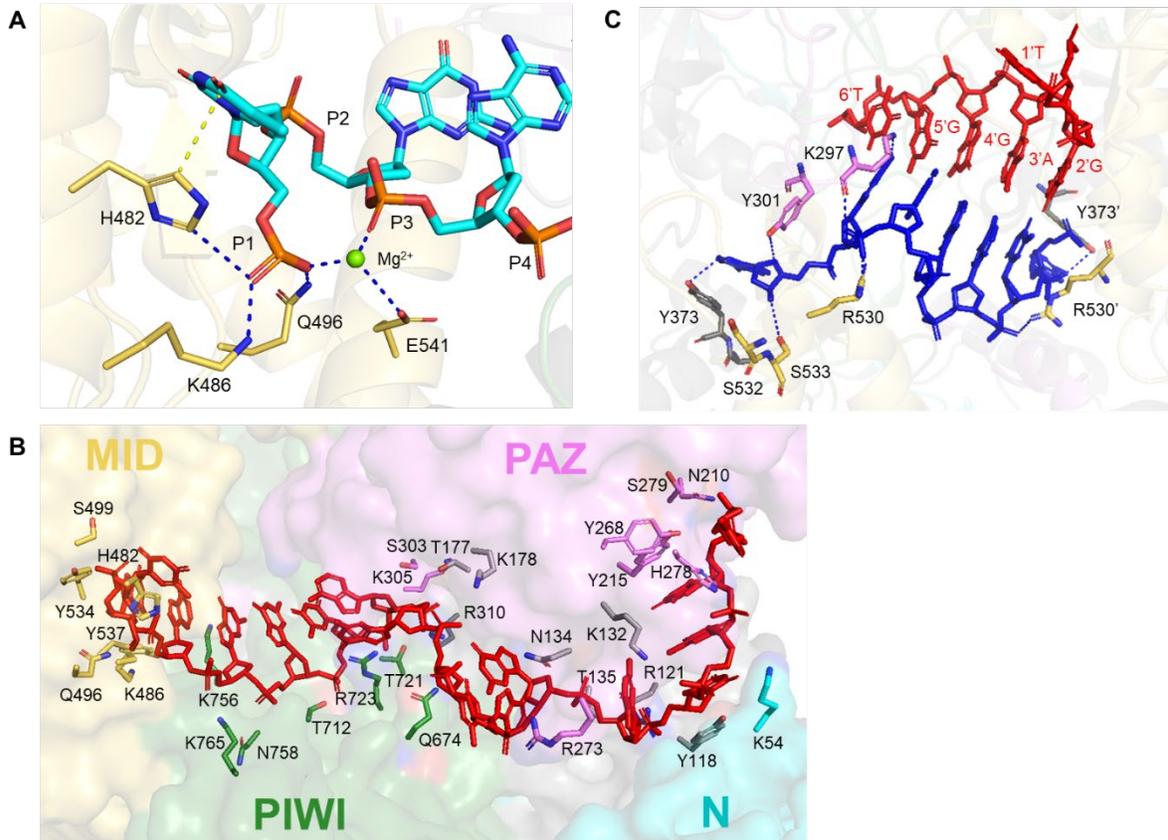
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18 **Supplementary figures**

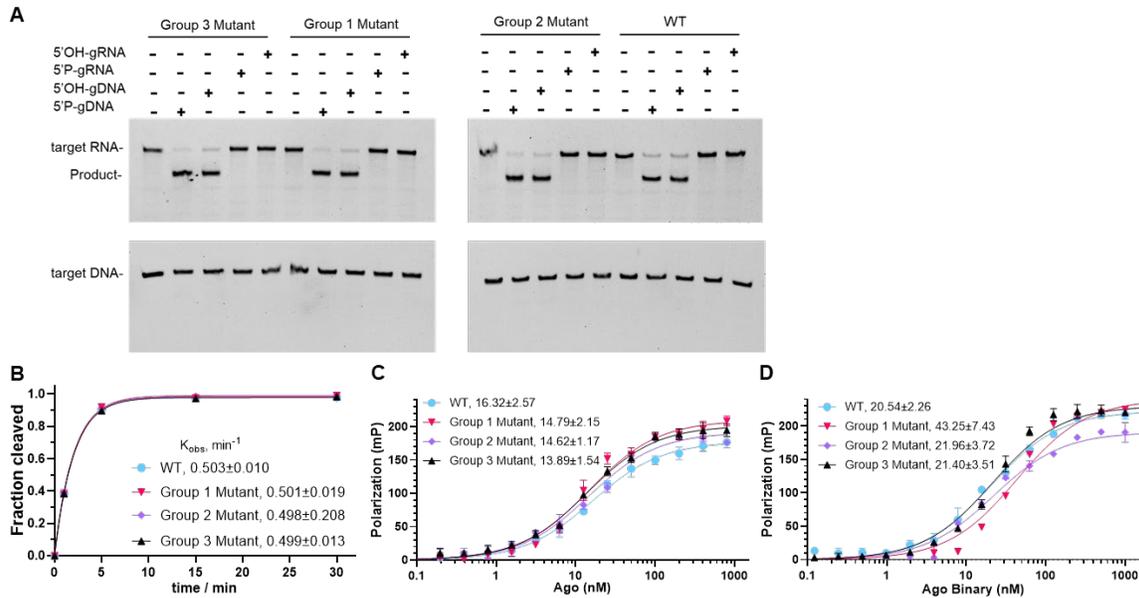


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 20 **Figure S1. Cryo-EM of MbpAgo-guide DNA-target DNA binary complex.** (A) Cryo-EM micrograph.
 21 (B) Representative 2D class averages. (C) Heterogeneous refinement with particle distribution. (D) Final
 22 reconstruction. (E) Angular distribution of the reconstruction shown in D. (F) Local resolution estimation
 23 on the reconstruction in D. (G) Global half-map FSC plot of the reconstruction shown in D. (H) Map-to-
 24 model FSC curve. (I) Local cryo-EM density of guide DNA.



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 26 **Figure S2. The interactions of MbpAgo with guide DNA.** (A) Interactions of the MID domain (yellow
 27 ribbon) with the 5'-phosphate of gDNA (stick representation) in MbpAgo-gDNA complex. Salt bridges
 28 between amino acid residues and phosphate groups, as well as coordination bonds of Mg^{2+} , are shown as
 29 blue dashed lines. The stacking histidine residue H482 with the 5'-end nucleotide of the gDNA is shown
 30 as yellow dashed lines. (B) A magnified view of the MbpAgo- gDNA interactions. MbpAgo is depicted as
 31 a partially transparent surface. DNA nucleotides and amino acid residues responsible for guide positioning
 32 are shown as stick models and labeled. (C) The binding and recognition candidate target DNA by MbpAgo.
 33 The view is the same as Figure 1A.

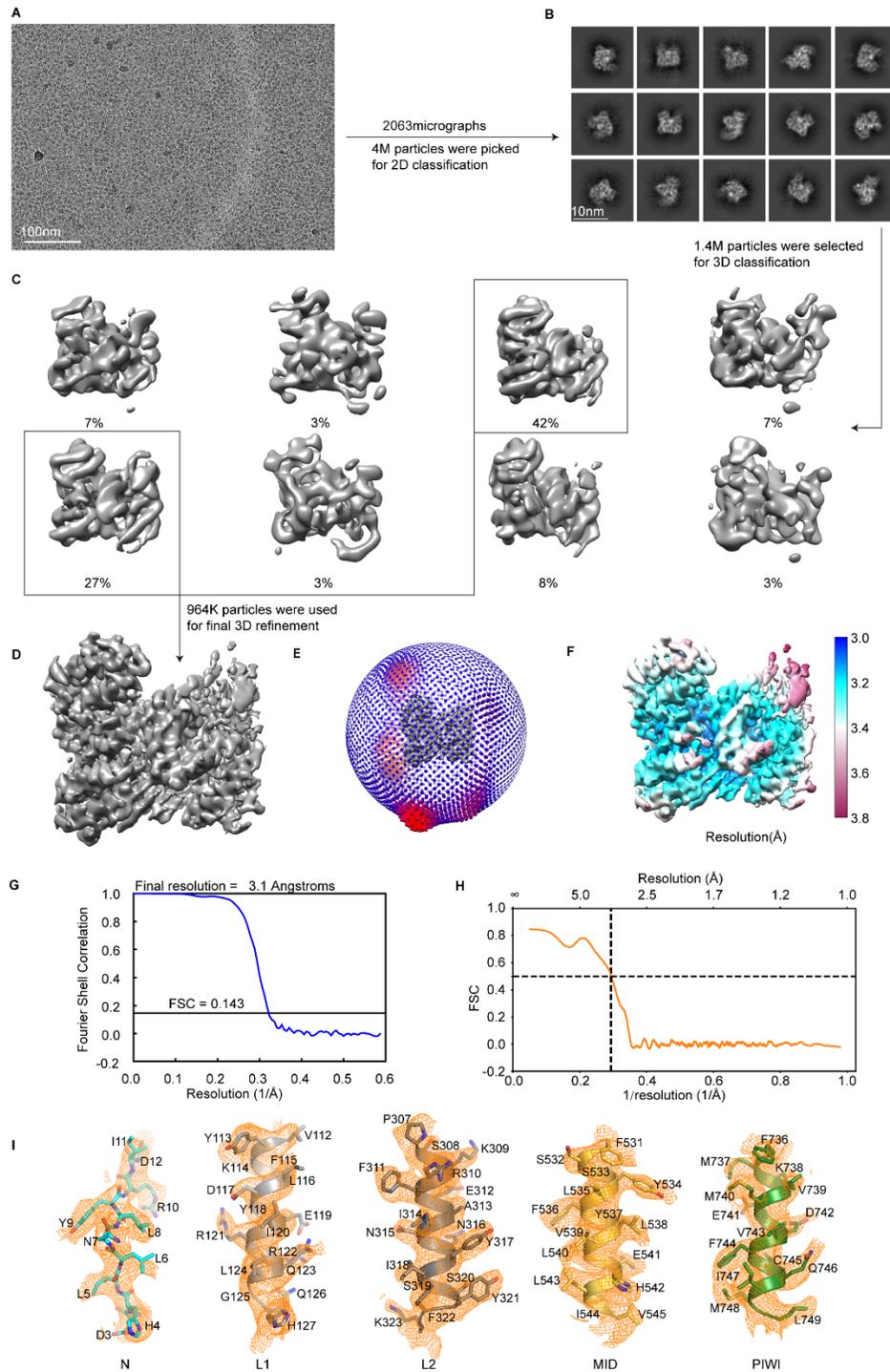
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36 **Figure S3. Target cleavage assay with wild-type MbpAgo and mutant MbpAgo (Group 1, Group 2**
 37 **and Group 3).** (A) Mutant MbpAgo (Group 1, Group 2 and Group 3) exhibits a preference for cleaving
 38 RNA consistent with the wild-type. (B) Target RNA cleavage assay with wild-type MbpAgo and mutant
 39 MbpAgo (Group 1, Group 2 and Group 3). (C) Binding of wild-type MbpAgo and mutant MbpAgo (Group
 40 1, Group 2 and Group 3) to 16 nt gDNA by fluorescence polarization assay. (D) Binding of wild-type
 41 MbpAgo and mutant MbpAgo (Group 1, Group 2 and Group 3) binary complex to 8 nt target DNA by
 42 fluorescence polarization assay. Data are represented as the mean ± standard deviation (SD) from three
 43 independent experiments.

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46 **Figure S4. Cryo-EM of MbpAgo-guide DNA-target DNA ternary complex.** (A) Cryo-EM micrograph.
 47 (B) Representative 2D class averages. (C) Heterogeneous refinement with particle distribution. (D) Final
 48 reconstruction. (E) Angular distribution of the reconstruction shown in D. (F) Local resolution estimation
 49 on the reconstruction in D. (G) Global half-map FSC plot of the reconstruction shown in D. (H) Map-to-
 50 model FSC curve. (I) Local cryo-EM density of representative helices from the individual domain of
 51 MbpAgo.

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53 **Supplementary tables**54 **Table S1. List of oligonucleotides used in this study.**

Oligonucleotide name	Sequence (5'-3')	Description	Used in figure
1. T-gDNA	TGAGGTAGTAGGTTGT	16 nt guide DNA pair with tDNA/ tRNA	Fig. 1-4; Fig. S3
2. 45-nt tDNA	FAM-AAACGACGGCCAGTGCCAAGC TTACTATACAACCTACTACCTCAT	5' FAM labeled 45-nt target DNA	Fig. S3A
3. 45-nt tRNA	FAM-AAACGACGGCCAGUGCCAAGC UUACUAUACAACCUACUACCUCAU	5' FAM labeled 45-nt target RNA	Fig. 3E-G; Fig. S3A, B
4. gRNA	UGAGGUAGUAGGUUGU	16 nt guide RNA pair with tDNA/ tRNA	Fig. S3A
5. gDNA iFAM	TGAGGTAGTAGGTI(FAM)GT	internally-labeled gDNA	Fig. S3C
6. 9 nt-TD	CTACCTCAT-FAM	3' FAM labeled 9-nt target DNA pair with gDNA form 8 bp duplex	Fig. S3D
7. 18-nt TD	TACAACCTACTACCTCAT	18-nt target DNA pair with gDNA form 16 bp duplex	Fig. 3A; Fig. 4A, D
8. 8-nt-TD	TACCTCAT	8-nt target DNA pair with gDNA form 7 bp duplex	Fig. 4A
9. 9 nt-TD	CTACCTCAT	9-nt target DNA pair with gDNA form 8 bp duplex	
10. 11 nt-TD	TACTACCTCAT	11-nt target DNA pair with gDNA form 10 bp duplex	
11. 13nt-TD	CCTACTACCTCAT	13-nt target DNA pair with gDNA form 12 bp duplex	
12. 15 nt-TD	AACCTACTACCTCAT	15-nt target DNA pair with gDNA form 14 bp duplex	
13. 8 nt-TD+10NT	ATGTTGGATGTACCTCAT	18-nt target DNA pair with gDNA form 7 bp matched duplex	Fig. 4B
14. 9 nt-TD+9NT	ATGTTGGATCTACCTCAT	18-nt target DNA pair with gDNA form 8 bp matched duplex	
15. 11 nt-TD+7NT	ATGTTGGTACTACCTCAT	18-nt target DNA pair with gDNA form 10 bp matched duplex	
16. 13 nt-TD+5NT	ATGTTCTACTACCTCAT	18-nt target DNA pair with gDNA form 12 bp matched duplex	
17. 15 nt-TD+3NT	ATGAACCTACTACCTCAT	18-nt target DNA pair with gDNA form 14 bp matched duplex	
18. 8-nt-TR	UACCUCAU	8-nt target RNA pair with gDNA form 7 bp duplex	Fig. 4C
19. 18-nt TR	UACAACCUACUACCUCAU	18-nt target RNA pair with gDNA form 17 bp duplex	
20. 3'-Cy3 gDNA	TGAGGTAGTAGGTTGT-Cy3	3' Cy3 labeled guide DNA	Fig. 4D
21. 3'-Cy5 gDNA	TGAGGTAGTAGGTTGT-Cy5	5' Cy3 labeled guide DNA	

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57 **Table S2 Primers used in constructing MbpAgo mutant.**

Primer	Sequence (5'-3')	template
Constructing Group 1 mutant		
22.	R530A-F	TGAAGGGCgcATTCAGCTCCTACCTGTTCTAC
23.	R530A-R	AGCTGAATgcGCCCTTCAGGGCTTCCACC
24.	Y301A-F	CCCTGCACgcCCTGTCTATCAAGGGCCCC
25.	Y301A-R	TAGACAGGgcGTGCAGGGACTTCACTCTC
26.	Y373A-F	GACTTCGCCgcCCAGAGAAAGCAGCTGATTAAGAAC
27.	Y373A-R	CTTTCTCTGGgcGGCGAAGTCCCGCATGCC
Constructing Group 2 mutant		
28.	Y364AR369A-F	CgcCAACAGCGGCATGgcGGACTTCGCCTACC
29.	Y364AR369A-R	CgcCATGCCGCTGTTGgcGCCGGTGATCTTC
30.	E312A-F	AGAGATTCgcGGCCATCAACAATTACATCTCC
31.	E312A-R	GTTGATGGCCgCGAATCTCTTGCTGGGGCC
32.	Y688AV695A-F	CGATCAGCTGGAAAACCCCGcGGCCGGCTCCTATAAG
33.	Y688AV695A-R	GGGGTTTTCCAGCTGATCGgcGCTGTTGGTCTTGCCGGC
34.	K2A-F	gcggcATGgcGGACCACATCCTGAACCTG
35.	K2A-R	TGTGGTCCgcCATgccgctgtgatg
36.	Y714A-F	GCACAGGCgcCCCCTTCGACATCAAGGG
37.	Y714A-R	CGAAGGGGgcGCCTGTGCTGAAGATAA
38.	Constructing Group 3 mutant	
39.	684-690del-F	CGCCGCCCTGGAAAACCCCGTGGCCGGCTC
40.	684-690del-R	GGGTTTTCCAGGGCGGCGATTCTGATTGG
41.	684-690GG-F	CGCCggcggctTGAAAACCCCGTGGCCGGCTC
42.	684-690GG-R	TTTTCCAGaccgccGGCGGCGATTCTGATTGG
43.	Constructing N686C mutant	
44.	N686C-F	GCCAAGACCTGCAGCTACGATCAGCTGG
45.	N686C-R	GTAGCTGCAGGTCTTGCCGGCGATTCC

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