

Supplementary Information for

Programmable Regiodivergent Light-Driven Cyclisation of Acyclic 1,5-Dienes

Unlocks Rigid Bicyclic Architectures

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1. Materials and General Methods

1.1. Glassware, Solvents and Reagents

All reactions were conducted under an inert atmosphere of nitrogen using standard Schlenk manifold techniques unless mentioned otherwise. All glassware was oven-dried at $>100\text{ }^{\circ}\text{C}$ and allowed to cool to room temperature under a positive pressure of nitrogen. Reactions were monitored by TLC until deemed complete using aluminium-backed silica plates. Plates were visualised under ultraviolet light (254 nm) and/or by staining with KMnO_4 solution. Cooling of reaction mixtures to $0\text{ }^{\circ}\text{C}$ was achieved using an ice-water bath. Cooling of reaction mixtures to $-78\text{ }^{\circ}\text{C}$ was achieved using a dry ice-acetone bath. "Room temperature" refers to an ambient temperature of $23 \pm 2\text{ }^{\circ}\text{C}$.

All anhydrous solvents (MeOH, CH_3CN , acetone, DMF, DMSO, 1,4-dioxane, Et_2O , THF, CH_2Cl_2 , 1,2-Dichloroethane, etc.) were commercially supplied (ACROS). All reagents were purchased from commercial sources [Sigma Aldrich (Merck), Across, Fischer, Fluorochem Ltd, TCI, etc.] and used as received unless otherwise stated. Irradiation of reaction mixtures was achieved using a 40 W KSPR160L-427 nm (or 370 and 390 nm) Kessil light (100% intensity). Brine refers to a saturated aqueous solution of NaCl.

1.2. Chromatography and Instrumentation

Flash column chromatography (FCC) was carried out using the Biotage IsoleraTM flash purification system equipped with a high-efficiency silica column (Biotage[®] Sfär Silica HC D High Capacity Duo, $20\text{ }\mu\text{m}$).

NMR spectra were recorded at various field strengths, as indicated, using Bruker 400 MHz and Bruker 500 MHz for ^1H , ^{13}C , ^{19}F acquisitions. All NMR spectra were recorded at $\sim 25\text{ }^{\circ}\text{C}$ in CDCl_3 unless otherwise stated. All reported ^1H and ^{13}C chemical shifts (δ_{H} , δ_{C}) are referenced to the residual signal of deuterated solvents (CDCl_3 : $\delta_{\text{H}} = 7.26\text{ ppm}$, $\delta_{\text{C}} = 77.16\text{ ppm}$; $(\text{CD}_3)_2\text{SO}$: $\delta_{\text{H}} = 2.50\text{ ppm}$, $\delta_{\text{C}} = 39.52\text{ ppm}$; $(\text{CD}_3)_2\text{CO}$: $\delta_{\text{H}} = 2.05\text{ ppm}$, $\delta_{\text{C}} = 206.26\text{ ppm}$; CD_3CN : $\delta_{\text{H}} = 1.94\text{ ppm}$, $\delta_{\text{C}} = 118.26\text{ ppm}$; CD_2Cl_2 : $\delta_{\text{H}} = 5.32\text{ ppm}$, $\delta_{\text{C}} = 53.84\text{ ppm}$). Chemical shifts (δ) are reported in parts per million (ppm) to the nearest 0.01 ppm for ^1H , ^{19}F NMR and 0.1 ppm for ^{13}C NMR. Coupling constants (J) are reported in Hertz (Hz). Multiplicities are reported as follows: s (singlet), d (doublet), t (triplet), q (quartet), pent. (quintet), hept. (septet), m (multiplet), br. (broad signal), app. (apparent). The ^1H NMR spectra are reported as follows: chemical shift (multiplicity, coupling constants, number of protons).

High resolution mass spectra (HRMS) were recorded on a Bruker Daltonics MicroTOF II by Electrospray Ionisation (ESI); a Thermo Scientific QExactive by Electron Ionisation (EI); a Thermo Scientific Orbitrap Elite by ESI or Atmospheric Pressure Chemical Ionisation (APCI); or a Bruker UltrafleXtreme by Matrix-assisted Laser Desorption/Ionisation (MALDI).

IR spectra were recorded neat as a thin film on a Perkin Elmer Spectrum One FT-IR. Selected absorption maxima (ν_{max}) are reported in wavenumbers (cm^{-1})

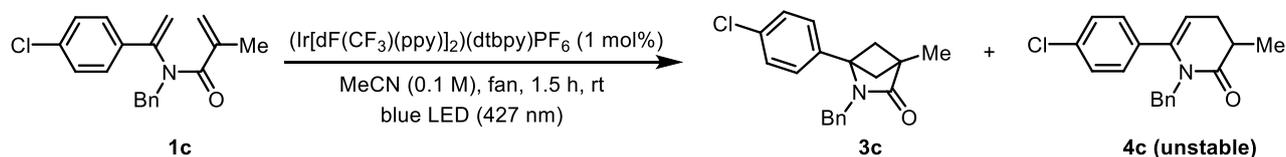
1.3. Naming of Compounds

Compound names are those generated by ChemDraw Professional 20.0 software (PerkinElmer), following the IUPAC nomenclature.

2. Experimental Data

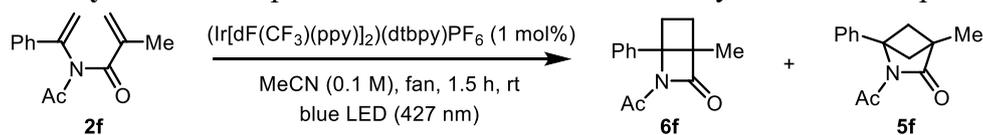
2.1. Reaction Optimisation Studies

Supplementary Table S1. Optimisation of the intramolecular cycloaddition of precursor **1c**



Entry	variation	E _T (kcal/mol)	E _{1/2} [M ^{•+} /M [*]] (V)	E _{1/2} [M ^{•+} /M [*]] (V)	3c (% yield)	4c (% yield)
1	[Ru(bpy) ₃](PF ₆) ₂ as PC	46.5	+0.77	-0.81	0	0
2	[Ir(ppy) ₂ (dtbbpy)]PF ₆ as PC	49.2	+0.66	-0.96	52	35
3	<i>fac</i> -[Ir(ppy) ₃] as PC	58.1	+0.31	-1.73	52	35
4	(Ir[dF(CF ₃)(ppy)] ₂)(dtbbpy)PF ₆ as PC	61.8	+1.21	-0.89	54 (52)	16 (14)
5	<i>fac</i> -[Ir(dF(ppy)) ₃] as PC	63.5	+0.34	-1.44	51	34
6	thioxanthone as PC ^a	65.6	+ 1.18	-1.11	45	0
7	THF as solvent				52	33
8	CH ₂ Cl ₂ as solvent ^b				48	45
9	EtOAc as solvent				52	31
10	Acetone as solvent				54	30
11	Et ₂ O as solvent				54	33
12	HFIP as solvent				0	72 (72)
13	15 min (reaction time)				47	35
14	30 min (reaction time)				51	20
15	0.2 M (concentration)				52	26
16	0.5 M (concentration)				51	27
17	without PC				0	0
18	without light				0	0

Yields determined by quantitative NMR spectroscopy of the crude reaction mixture using CH₂Br₂ as the internal standard. Isolated yields in the parentheses. a. 390 nm blue LED was used. b. gram scal, Ir[dF(CF₃)(ppy)]₂(dtbbpy)PF₆ (0.5 mol%), 1.5 h.

Supplementary Table S2. Optimisation of the intramolecular cycloaddition of precursor **2f**

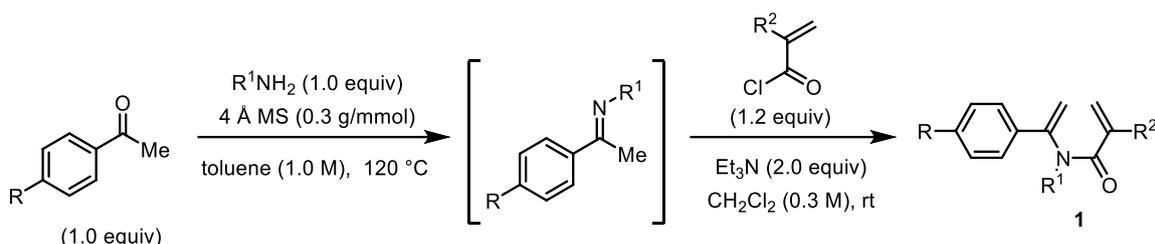
Entry	variation	E _T (kcal/mol)	E _{1/2} [M [*] /M [*]] (V)	E _{1/2} [M ⁺⁺ /M [*]] (V)	6f (% yield)	5f (% yield)
1	[Ru(bpy) ₃](PF ₆) ₂ as PC	46.5	+0.77	-0.81	0	0
2	[Ir(ppy) ₂ (dtbbpy)]PF ₆ as PC	49.2	+0.66	-0.96	36	3
3	<i>fac</i> -[Ir(ppy) ₃] as PC	58.1	+0.31	-1.73	48	5
4	(Ir[dF(CF ₃)(ppy) ₂](dtbpy)PF ₆ as PC	61.8	+1.21	-0.89	80 (76)	8 (5)
5	<i>fac</i> -[Ir(dF(ppy)) ₃] as PC	63.5	+0.34	-1.44	80	8
6	thioxanthone as PC ^a	65.6	+ 1.18	-1.11	79	8
7	THF as solvent				79	10
8	CH ₂ Cl ₂ as solvent				80	9
9	EtOAc as solvent				80	9
10	Acetone as solvent				80	9
11	Et ₂ O as solvent				79	10
12	HFIP as solvent				85 (79)	3
13	15 min (reaction time)				80	8
14	30 min (reaction time)				80	8
15	0.2 M (concentration)				80	8
16	0.5 M (concentration)				79	9
17	without PC				0	0
18	without light				0	0

Yields determined by quantitative NMR spectroscopy of the crude reaction mixture using CH₂Br₂ as the internal standard. Isolated yields in the parentheses. a. 390 nm blue LED was used.

For substrates bearing *N*-substituents with varying electronic properties (**Figure 2A in the main text**), the product distributions resulting from the competing 5-*exo*-trig and 6-*endo*-trig cyclisations under the optimal conditions, together with the corresponding product characterisation data, are provided in **Section 2.4 (Synthesis of Cyclisation Products)** of the Supporting Information.

2.2 General Procedure

2.2.1. General Procedure A: Synthesis of Cyclisation Precursors (1)

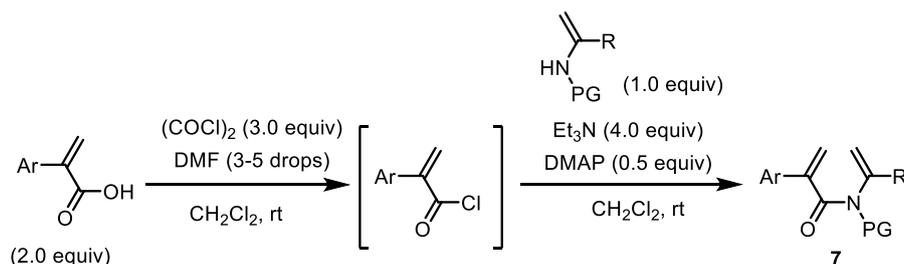


An oven-dried round-bottom flask equipped with a magnetic stir bar and a 15 cm condenser was charged with 4 Å molecular sieves (powder, 0.3 g/mmol) under an argon atmosphere in a glovebox. The flask was sealed with a rubber septum, removed from the glovebox, and kept under a nitrogen atmosphere using a balloon. Anhydrous toluene (1.0 M), the ketone (1.0 equiv), and amine (1.0 equiv) were added sequentially via syringe. The reaction mixture was heated to 120 °C under nitrogen for 5 h, then cooled to room temperature. The resulting mixture was rapidly filtered through anhydrous $MgSO_4$ and washed with anhydrous CH_2Cl_2 (approximately 0.3 M with respect to ketone).^A The filtrate was collected in an oven-dried round-bottom flask and the crude imine was used directly in the next step without further purification.

The flask containing the crude imine (1.0 equiv) was sealed and kept under a nitrogen atmosphere using a balloon. Triethylamine (2.0 equiv) was added, followed by the acyl chloride (1.2 equiv). The reaction mixture was stirred at room temperature until completion of the reaction (TLC). The reaction was quenched with saturated aqueous $NaHCO_3$ and extracted with CH_2Cl_2 three times. The combined organic extracts were dried over anhydrous $MgSO_4$, filtered, and concentrated *in vacuo*. The crude product was purified using a Biotage automated flash chromatography system to afford the desired products.

Notes: (A) Due to the inherent instability of the imine, which is prone to hydrolysis, the crude imine was immediately and rapidly filtered (over $MgSO_4$) upon cooling to room temperature, and the residue was washed with anhydrous CH_2Cl_2 . The receiving round-bottom flask was also oven-dried to minimise exposure to moisture and achieve the highest possible yield.

2.2.2. General Procedure A: Synthesis of Cyclisation Precursors (7)



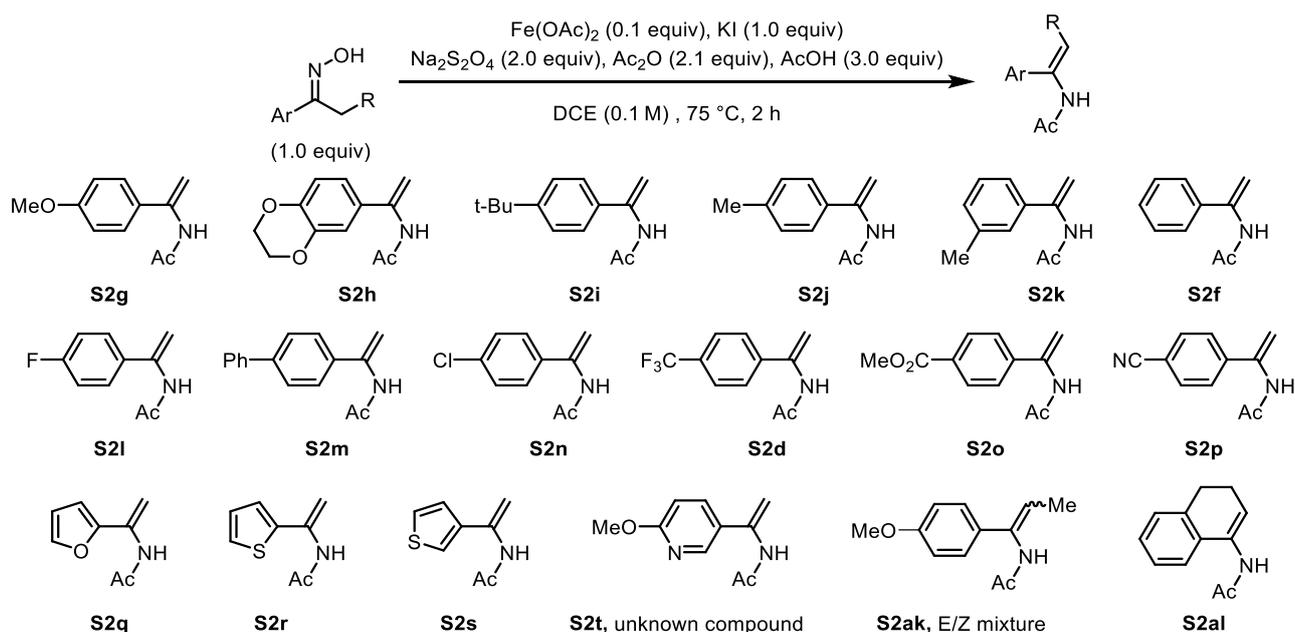
An oven-dried round-bottom flask containing carboxylic acid (2.0 equiv) was sealed and subjected to three N₂ evacuation/refill cycles before pre-sparged anhydrous CH₂Cl₂ (0.3 M with respect to carboxylic acid) was added. The solution was cooled to 0 °C before oxalyl chloride (3.0 equiv) was added, followed by DMF (3-5 drops). The reaction mixture was allowed to warm to room temperature and stirred for 1 h. The reaction was then concentrated under reduced pressure to afford the crude acyl chloride which was used directly for the next step without further purification.

The crude acyl chloride (2.0 equiv) was re-dissolved in anhydrous CH₂Cl₂ (0.3 M with respect to enamide) at room temperature. Et₃N (4.0 equiv) was then added, followed by the addition of the enamide (1.0 equiv) in anhydrous CH₂Cl₂ (1.0 M with respect to enamide) and DMAP (0.5 equiv). The reaction mixture was stirred overnight until completion of the reaction (TLC). The reaction was quenched with brine and extracted with CH₂Cl₂ three times. The combined extracts were dried over anhydrous MgSO₄ and concentrated *in vacuo*. The crude product was purified using a Biotage automated flash chromatography system to afford the desired product.

2.2.3. General Procedure C: Synthesis of Cyclisation Precursors (2)

Almost all *N*-acetyl enamides used in this study (shown in **Supplementary Figure S1**) are known compounds and were synthesised according to the modified literature-reported procedure shown below.¹ The ¹H and ¹³C NMR spectra of our samples were consistent with the reported data.¹⁻⁵ For enamides that have not been previously reported, full characterisation data are provided.

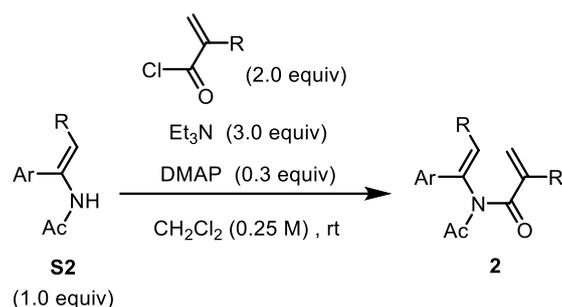
Supplementary Figure S1. List of *N*-acetyl enamides used as starting materials in the synthesis of cyclisation precursors



To an oven-dried round-bottom flask equipped with a magnetic stir bar and a 15 cm condenser, acetophenone oxime (1.0 equiv), KI (1.0 equiv), Na₂S₂O₄ (2.0 equiv), and Fe(OAc)₂ (0.1 equiv) were added under an argon atmosphere in a glovebox. The flask was sealed with a rubber septum, removed from the glovebox, and kept under a nitrogen atmosphere using a balloon. 1,2-Dichloroethane (DCE, 0.1 M), acetic acid (3.0 equiv), and acetic anhydride (2.1 equiv) were added to the mixture via syringe. The reaction mixture was heated to 75 °C for 2 h, then cooled to room temperature. The resulting mixture was quenched with saturated NaHCO₃ solution and extracted with CH₂Cl₂ three times. The combined organic layers were dried over anhydrous MgSO₄, filtered, concentrated under reduced pressure, and purified using a Biotage automated flash chromatography system to afford the desired product.^A

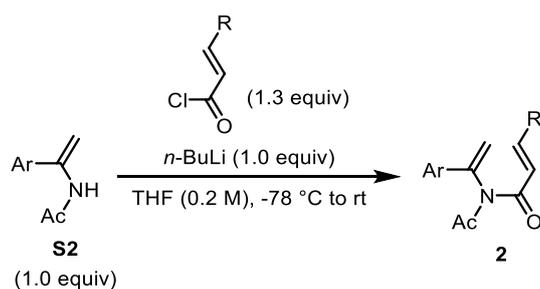
Notes: (A) If the crude product after column chromatography is not sufficiently pure, further purification can be performed either by a second round of chromatography or by recrystallisation. Since the product is typically a solid, it often precipitates during rotary evaporation. In such cases, recrystallisation from a petroleum ether/Et₂O solvent system can be used for purification.

2.2.3.1 General Procedure C1: Synthesis of Cyclisation Precursors (2)



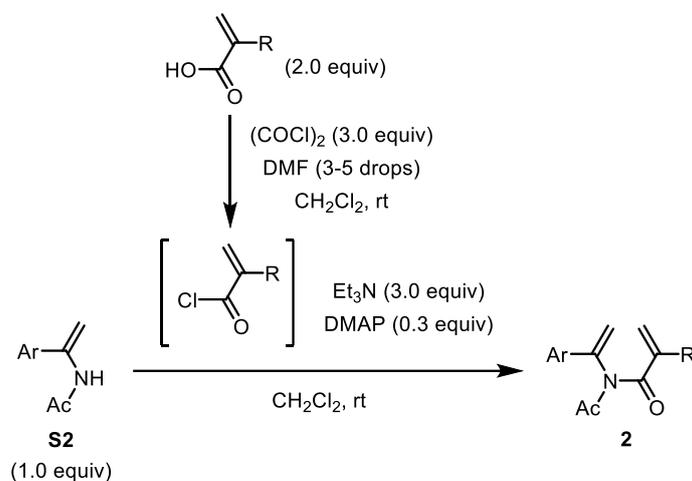
In an oven-dried round-bottom flask, the *N*-acetyl enamide **S2** (1.0 equiv) was dissolved in anhydrous CH₂Cl₂ (0.25 M) at room temperature. Et₃N (3.0 equiv) was then added, followed by the addition of acyl chloride (2.0 equiv) and DMAP (0.3 equiv). The reaction mixture was stirred overnight. The reaction was quenched with brine and extracted with CH₂Cl₂ three times. The combined extracts were dried over anhydrous MgSO₄ and concentrated *in vacuo*. The crude product was purified by Biotage automated flash chromatography system to afford the needed product **2**.

2.2.3.2 General Procedure C2: Synthesis of Cyclisation Precursors (2)



An oven-dried round-bottom flask charged with *N*-acetyl enamide **S2** (1.0 equiv) was sealed and subjected to three cycles of evacuation and nitrogen backfilling. Anhydrous THF (0.2 M) was then added, and the reaction mixture was cooled to $-78\text{ }^{\circ}\text{C}$. *n*-BuLi (1.0 equiv) was added dropwise at $-78\text{ }^{\circ}\text{C}$, and the mixture was stirred at this temperature for 10 minutes. Acyl chloride (1.3 equiv) was subsequently added at $-78\text{ }^{\circ}\text{C}$, and the reaction was allowed to warm to room temperature over 1 h. The reaction was then quenched with brine and extracted with ethyl acetate three times. The combined organic layers were dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude product was purified by a Biotage automated flash chromatography system to afford the desired compound **2**.

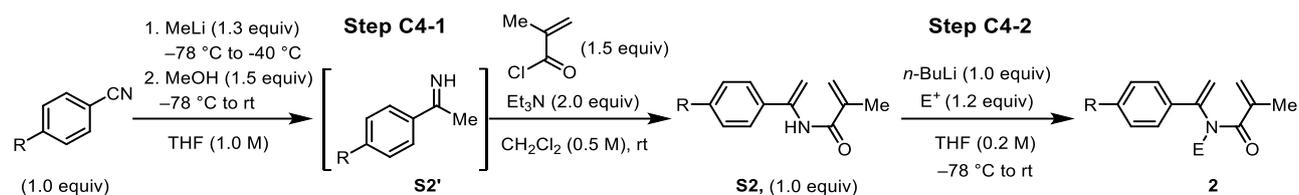
2.2.3.3 General Procedure C3: Synthesis of Cyclisation Precursors (2)



An oven-dried round-bottom flask containing carboxylic acid (2.0 equiv) was sealed and subjected to three N_2 evacuation/refill cycles before pre-sparged anhydrous CH_2Cl_2 (0.3 M with respect to carboxylic acid) was added. Oxalyl chloride (3.0 equiv) was then added, followed by the addition of DMF (3-5 drops). The reaction mixture was allowed to stir at room temperature for 1 h. The reaction was then concentrated under reduced pressure to afford the crude acyl chloride which was used directly for the next step without further purification.

The crude acyl chloride (2.0 equiv) was redissolved in anhydrous CH_2Cl_2 (0.25 M with respect to *N*-acetyl enamide) at room temperature. Et_3N (3.0 equiv) was then added, followed by the addition of *N*-acetyl enamide **S2** (1.0 equiv) and DMAP (0.3 equiv). The reaction mixture was stirred overnight until completion of the reaction (TLC). The reaction was quenched with brine and extracted with CH_2Cl_2 three times. The combined extracts were dried over anhydrous MgSO_4 and concentrated *in vacuo*. The crude product was purified by a Biotage automated flash chromatography system to afford the desired product **2**.

2.2.3.4 General Procedure C4: Synthesis of Cyclisation Precursors (2)



Step C4-1: To an oven-dried round-bottom flask charged with benzonitrile (1.0 equiv) and a magnetic stir bar was added anhydrous THF (1.0 M) under a nitrogen atmosphere. The solution was cooled to $-78\text{ }^{\circ}\text{C}$, and MeLi solution (1.3 equiv) was added dropwise. After stirring at $-78\text{ }^{\circ}\text{C}$ for 5 min, the reaction was warmed to $-40\text{ }^{\circ}\text{C}$ and stirred for 2 h. Then the mixture was cooled again to $-78\text{ }^{\circ}\text{C}$, followed by the dropwise addition of anhydrous MeOH (1.5 equiv). The reaction was allowed to warm to room temperature over the course of 1 h. The reaction mixture was then immediately filtered through anhydrous MgSO_4 under reduced pressure and washed with anhydrous CH_2Cl_2 (approximately 0.5 M with respect to benzonitrile) under nitrogen protection to prevent decomposition of the imine intermediate.^A The filtrate was collected in an oven-dried round-bottom flask, sealed with a rubber septum, and protected under a nitrogen balloon. Et_3N (2.0 equiv) and methacryloyl chloride (1.5 equiv) were then added to the filtrate under a nitrogen atmosphere. The reaction mixture was stirred at room temperature for 12 h. Upon completion, the reaction mixture was then quenched with brine and extracted with ethyl acetate three times. The combined organic layers were dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude product was purified by a Biotage automated flash chromatography system to afford *N*-H 1,5-diene **S2**.

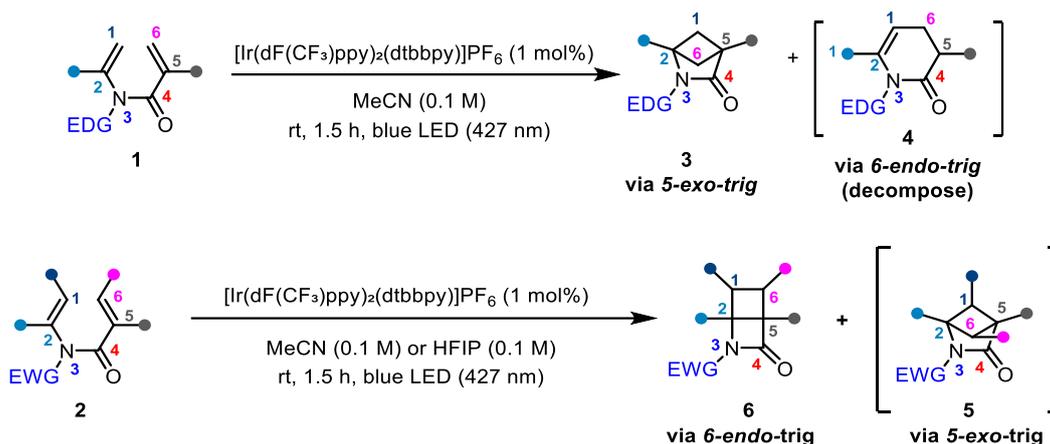
Notes: **(A)** The *N*-H imine intermediate **S2'** is unstable at room temperature; hence, the MgSO_4 filtration of the crude *N*-H imine intermediate **S2'** should be carried out rapidly under reduced pressure and under a nitrogen atmosphere. The filtrate containing the crude *N*-H imine **S2'** was immediately subjected to acylation without further purification.

(B) Although acylation steps typically do not require long reaction times, we found that upon addition of methacryloyl chloride, the reaction mixture rapidly formed a mixture of enamine **S2** and its imine isomer, as evidenced by crude ^1H NMR analysis. Importantly, stirring the reaction at room temperature for 12 h led to substantial conversion of the imine into the enamine, thereby affording **S2** in improved yield.

Step C4-2: An oven-dried round-bottom flask charged with *N*-H 1,5-diene **S2** (1.0 equiv) was sealed and subjected to three cycles of evacuation and nitrogen backfilling. Anhydrous THF (0.2 M) was then added, and the reaction mixture was cooled to $-78\text{ }^{\circ}\text{C}$. *n*-BuLi solution (1.0 equiv) was added dropwise at $-78\text{ }^{\circ}\text{C}$, and the mixture was stirred at this temperature for 10 minutes. Electrophile (1.2

equiv) was subsequently added at $-78\text{ }^{\circ}\text{C}$, and the reaction was allowed to warm to room temperature over 1 hour. The reaction was then quenched with brine and extracted with ethyl acetate three times. The combined organic layers were dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude product was purified by a Biotage automated flash chromatography system to afford the desired compound **2**.

2.2.4. General Procedure D: Synthesis of Cyclisation Products



An oven-dried round-bottom flask containing the substrate (1.0 equiv) and $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbbpy})]\text{PF}_6$ (1.0 mol%) was sealed and subjected to three evacuation–backfill cycles with N_2 . Anhydrous solvent^A (0.1 M) pre-sparged with N_2 was added, and the flask was positioned ~ 1 cm from a 40 W KSPR160L-427 nm Kessil light source (100% intensity).^B The mixture was stirred under continuous irradiation at ambient temperature^C under N_2 for 1.5 h (judged by TLC).^{D,E} Concentration *in vacuo* and purification by Biotage automated flash chromatography system afforded the desired product.^{F,G}

Notes: (A) All substrates were evaluated in MeCN as the solvent. For substrates exhibiting poor selectivity toward the 6-*endo*-trig pathway, experiments using HFIP as the solvent were additionally performed and are specifically indicated in the corresponding characterisation data.

(B) When thioxanthone (0.1 equiv) is used as a photosensitizer, a 40W KSPR160L-390 nm Kessil light (100% intensity) was used. When benzophenone (0.25 equiv) is used as a photosensitizer, a 40W KSPR160L-370 nm Kessil light (100% intensity) was used.

(C) The actual temperature of the reaction mixture was higher than room temperature. Fan-assisted cooling was used to maintain the temperature at around $35\text{--}40\text{ }^{\circ}\text{C}$. The solution temperature was monitored with a thermometer probe placed 1 cm below the solvent surface.

(D) Most substrates react completely in 1.5 h; substrates requiring extended reaction times are specifically indicated in the subsequent detailed data descriptions.

(E) The reaction was typically monitored by TLC. The product is usually slightly more polar than the starting material; however, in many cases, the products and starting precursor exhibit very similar R_f values. A relatively straightforward way to distinguish them is by their UV response under 254 nm light: product spots appear significantly weaker due to the loss of the styrenyl chromophore. Additionally, the product generally shows a slower colour development when treated with the KMnO_4 stain, as the reactive alkene moiety has been consumed. Of course, the most reliable method of identification is by sampling the crude reaction mixture for ^1H NMR analysis.

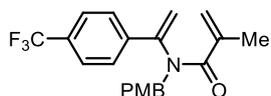
(F) When 1,5-diene **1** was subjected to the [2+2] cyclisation, the corresponding [2.1.1] bicyclic compound **3** was the only product that could be isolated. The six-membered-ring product (**4**) arising from the 6-*endo*-trig pathway could occasionally be detected only in trace amounts in the crude ^1H NMR spectrum. Time-course ^1H NMR monitoring at different reaction intervals (15, 30, 60, and 90 min) revealed that this species gradually decomposes over time. Furthermore, when an isolated sample of compound **4** was resubjected to the standard photochemical conditions, substantial decomposition was observed within 1 h. Owing to the intrinsic instability of this compound, except for a few selected substrates that could be isolated and fully characterised, only the yield and characterisation data of the [2.1.1] scaffolds (**3**) are reported.

(G) When using 1,5-diene **2** for the [2+2] cyclisation, both the [2.2.0] product (**6**) and the [2.1.1] product (**5**) could be observed in the crude ^1H NMR spectrum (shown below) and isolated. All products in this study were ultimately purified using a Biotage automated flash chromatography system equipped with a high-efficiency silica column (Biotage® Sfar Silica HC D High Capacity Duo, 20 μm). When the molar ratio of [2.2.0] product **6** to [2.1.1] product **5** in the crude mixture was less than 5:1, the [2.1.1] isomer **5** was also isolated and fully characterised.

2.3 Synthesis of Cyclisation Precursors

2.3.1 Synthesis and Characterisation of Precursors (1)

N-(4-methoxybenzyl)-*N*-(1-(4-(trifluoromethyl)phenyl)vinyl)methacrylamide (**1a**)



Prepared according to **General Procedure A** using 4-(trifluoromethyl)acetophenone (1.50 g, 8.00 mmol, 1.00 equiv), toluene (8.0 mL), 4-methoxybenzylamine (1.04 mL, 8.00 mmol, 1.00 equiv) and 4 Å molecular sieves (2.40 g). Then using CH₂Cl₂ (30.0 mL), Et₃N (2.23 mL, 16.0 mmol, 2.00 equiv.) and methacryloyl chloride (0.94 mL, 9.60 mmol, 1.20 equiv). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **1a** as a colourless oil (2.33 g, 6.21 mmol, 78%).

¹H NMR (500 MHz, CDCl₃) δ 7.60 (d, *J* = 8.2 Hz, 2H), 7.41 (d, *J* = 8.1 Hz, 2H), 7.20 (d, *J* = 8.6 Hz, 2H), 6.82 (d, *J* = 8.6 Hz, 2H), 5.46 (d, *J* = 0.9 Hz, 1H), 5.26 (s, 1H), 5.05 (s, 1H), 4.98 (d, *J* = 0.9 Hz, 1H), 4.73 (s, 2H), 3.78 (s, 3H), 1.73 (s, 3H).

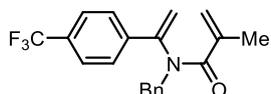
¹³C NMR (126 MHz, CDCl₃) δ 172.4, 159.2, 146.5, 141.2, 140.6, 130.8 (q, *J* = 32.6 Hz), 130.5, 129.4, 126.8, 125.9 (q, *J* = 3.8 Hz), 124.0 (q, *J* = 272.1 Hz), 118.3, 114.0, 113.9, 55.3, 50.5, 20.1.

¹⁹F NMR (471 MHz, CDCl₃) δ -62.68.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1654, 1612, 1511, 1384, 1246, 1165, 1113, 1063.

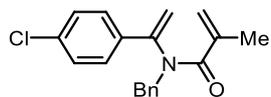
HRMS (ESI⁺) calcd. for C₂₁H₂₁F₃NO₂⁺ [M+H]⁺: 376.1519, found: 376.1519.

N-benzyl-*N*-(1-(4-(trifluoromethyl)phenyl)vinyl)methacrylamide (**1b**)



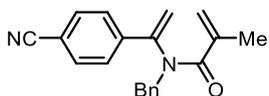
Prepared according to **General Procedure A**, known compound.⁶

N-benzyl-*N*-(1-(4-chlorophenyl)vinyl)methacrylamide (**1c**)



Prepared according to **General Procedure A**, known compound.⁶

N-benzyl-*N*-(1-(4-cyanophenyl)vinyl)methacrylamide (**1d**)



Prepared according to **General Procedure A** using 4-acetylbenzointrile (1.16 g, 8.00 mmol, 1.00 equiv), toluene (8.0 mL), benzylamine (0.87 mL, 8.0 mmol, 1.0 equiv) and 4 Å molecular sieves (2.40 g). Then using CH₂Cl₂ (30.0 mL), Et₃N (2.23 mL, 16.0 mmol, 2.00 equiv) and methacryloyl chloride (0.94 mL, 9.6 mmol, 1.2 equiv). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/CH₂Cl₂ (0% to 10%), affording **1d** as a colourless oil (1.30 g, 4.30 mmol, 54%).

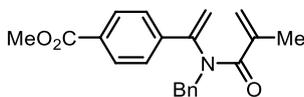
¹H NMR (500 MHz, CDCl₃) δ 7.62 (d, *J* = 8.5 Hz, 2H), 7.37 (d, *J* = 8.5 Hz, 2H), 7.31 – 7.25 (m, 5H), 5.47 (d, *J* = 1.1 Hz, 1H), 5.26 (s, 1H), 5.07 (s, 2H), 4.81 (s, 2H), 1.71 (t, *J* = 1.3 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 172.3, 146.5, 141.7, 141.0, 137.1, 132.6, 128.9, 128.7, 127.8, 127.0, 118.7, 118.5, 114.5, 112.4, 51.4, 20.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2228, 1655, 1614, 1495, 1453, 1352, 1261, 1197, 1030.

HRMS (ESI⁺) calcd. for C₂₀H₁₉N₂O⁺ [M+H]⁺: 303.1492, found: 303.1480.

Methyl 4-(1-(*N*-benzylmethacrylamido)vinyl)benzoate (**1e**)



Prepared according to **General Procedure A** using methyl 4-acetylbenzoate (1.42 g, 8.00 mmol, 1.00 equiv), toluene (8.0 mL), benzylamine (0.87 mL, 8.0 mmol, 1.0 equiv) and 4 Å molecular sieves (2.40 g). Then using CH₂Cl₂ (30.0 mL), Et₃N (2.23 mL, 16.0 mmol, 2.00 equiv) and methacryloyl chloride (0.94 mL, 9.6 mmol, 1.2 equiv). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (0% to 50%), affording **1e** as a colourless oil (588 mg, 1.76 mmol, 22%).

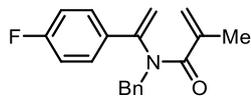
¹H NMR (500 MHz, CDCl₃) δ 8.01 (d, *J* = 8.5 Hz, 2H), 7.37 (d, *J* = 8.5 Hz, 2H), 7.31 – 7.25 (m, 5H), 5.46 (d, *J* = 0.9 Hz, 1H), 5.28 (s, 1H), 5.05 (app. p, *J* = 1.6 Hz, 1H), 5.00 (d, *J* = 0.9 Hz, 1H), 4.81 (s, 2H), 3.92 (s, 3H), 1.73 (t, *J* = 1.3 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 172.5, 166.6, 147.0, 141.5, 141.1, 137.3, 130.4, 130.1, 129.0, 128.6, 127.6, 126.4, 118.4, 113.7, 52.4, 51.2, 20.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1721, 1675, 1656, 1609, 1521, 1435, 1277, 1196, 1108.

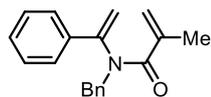
HRMS (ESI⁺) calcd. for C₂₁H₂₂NO₃⁺ [M+H]⁺: 336.1594, found: 336.1580.

***N*-benzyl-*N*-(1-(4-fluorophenyl)vinyl)methacrylamide (1f)**



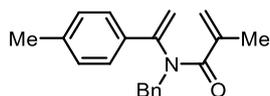
Prepared according to **General Procedure A**, known compound.⁶

***N*-benzyl-*N*-(1-phenylvinyl)methacrylamide (1g)**



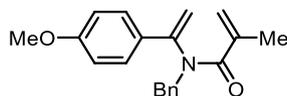
Prepared according to **General Procedure A**, known compound.⁶

***N*-benzyl-*N*-(1-(*p*-tolyl)vinyl)methacrylamide (1h)**



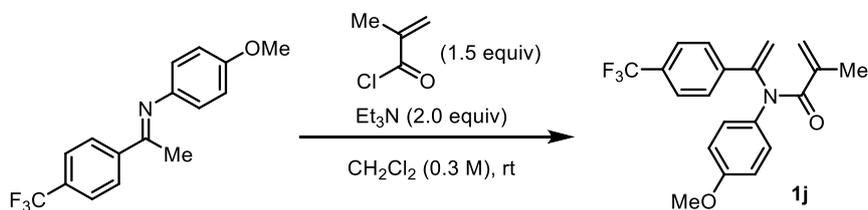
Prepared according to **General Procedure A**, known compound.⁷

***N*-benzyl-*N*-(1-(4-methoxyphenyl)vinyl)methacrylamide (1i)**



Prepared according to **General Procedure A**, known compound.⁶

***N*-(4-methoxyphenyl)-*N*-(1-(4-(trifluoromethyl)phenyl)vinyl)methacrylamide (1j)**



In an oven-dried round-bottom flask, *N*-(4-methoxyphenyl)-1-(4-(trifluoromethyl)phenyl)ethan-1-imine⁸ (1.04 g, 3.60 mmol, 1.00 equiv) was dissolved in anhydrous CH₂Cl₂ (12.0 mL) at room temperature. Et₃N (0.99 mL, 7.1 mmol, 2.0 equiv) was then added, followed by the addition of methacryloyl chloride (0.52 mL, 5.3 mmol, 1.5 equiv). The reaction mixture was stirred overnight. The reaction was then quenched with brine (100 mL) and extracted with CH₂Cl₂ (3 × 50 mL). The

combined extracts were dried over anhydrous MgSO₄ and concentrated *in vacuo*. The crude product was purified by Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 50%), affording **1j** as a pale-yellow oil (1.25 g, 3.46 mmol, 98%).

¹H NMR (500 MHz, CDCl₃) δ 7.62 (d, *J* = 9.1 Hz, 2H), 7.59 (d, *J* = 8.6 Hz, 2H), 7.15 (d, *J* = 9.0 Hz, 2H), 6.85 (d, *J* = 8.9 Hz, 2H), 5.59 (d, *J* = 1.0 Hz, 1H), 5.41 (s, 1H), 5.21 (app. p, *J* = 1.4 Hz, 1H), 5.13 (s, 1H), 3.79 (s, 3H), 1.82 (t, *J* = 1.3 Hz, 3H).

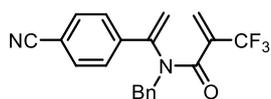
¹³C NMR (126 MHz, CDCl₃) δ 171.8, 158.1, 148.2, 141.0, 140.9, 135.2, 130.1 (q, *J* = 32.6 Hz), 127.4, 126.2, 125.5 (q, *J* = 3.8 Hz), 124.0 (q, *J* = 272.1 Hz), 120.5, 114.4, 113.6, 55.2, 19.6.

¹⁹F NMR (471 MHz, CDCl₃) δ -62.65.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1660, 1617, 1508, 1321, 1299, 1243, 1165, 1112, 1065.

HRMS (ESI⁺) calcd. for C₂₀H₁₉F₃NO₂⁺ [M+H]⁺: 362.1362, found: 362.1349.

***N*-benzyl-*N*-(1-(4-cyanophenyl)vinyl)-2-(trifluoromethyl)acrylamide (**1k**)**



Prepared according to **General Procedure A** using 4-acetylbenzonnitrile (870 mg, 6.00 mmol, 1.00 equiv), toluene (6.0 mL), benzylamine (0.65 mL, 6.0 mmol, 1.0 equiv) and 4 Å molecular sieves (1.80 g). Then using CH₂Cl₂ (20.0 mL), Et₃N (1.67 mL, 12.0 mmol, 2.00 equiv) and 2-(trifluoromethyl)-2-propenoyl chloride⁹ (1.14 g, 7.20 mmol, 1.20 equiv).^A The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of acetone/pentane (0% to 35%) and recrystallisation, affording **1k** as a white solid (1.60 g, 4.49 mmol, 75%).

Notes: (A) 2-(Trifluoromethyl)-2-propenoyl chloride⁹ was purified by distillation before use. It was stored in a glovebox freezer at -30 °C, where it remained stable for at least three months.

M.P.: 115-117 °C

¹H NMR (500 MHz, CDCl₃) δ 7.67 (d, *J* = 8.5 Hz, 2H), 7.43 (d, *J* = 8.6 Hz, 2H), 7.32 – 7.27 (m, 3H), 7.25 – 7.22 (m, 2H), 5.88 (s, 1H), 5.73 (s, 1H), 5.66 (d, *J* = 1.4 Hz, 1H), 5.08 (s, 1H), 4.80 (s, 2H).

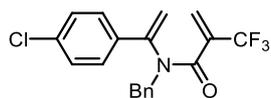
¹³C NMR (126 MHz, CDCl₃) δ 164.3, 144.8, 140.1, 136.0, 134.9 (q, *J* = 31.9 Hz), 133.0, 129.1, 128.8, 128.2, 126.7, 123.8 (q, *J* = 5.3 Hz), 121.5 (q, *J* = 274.1 Hz), 118.3, 117.6, 113.0, 51.1.

¹⁹F NMR (471 MHz, CDCl₃) δ -64.09.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2231, 1664, 1625, 1396, 1345, 1275, 1263, 1172, 1122.

HRMS (ESI⁺) calcd. for C₂₀H₁₆F₃N₂O⁺ [M+H]⁺: 357.1209, found: 357.1219.

N-benzyl-*N*-(1-(4-chlorophenyl)vinyl)-2-(trifluoromethyl)acrylamide (**1l**)



Prepared according to **General Procedure A** using 4-chloroacetophenone (0.78 mL, 6.0 mmol, 1.0 equiv), toluene (6.0 mL), benzylamine (0.65 mL, 6.0 mmol, 1.0 equiv) and 4 Å molecular sieves (1.80 g). Then using CH₂Cl₂ (20.0 mL), Et₃N (1.67 mL, 12.0 mmol, 2.00 equiv) and 2-(trifluoromethyl)-2-propenoyl chloride⁹ (1.14 g, 7.20 mmol, 1.20 equiv). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of acetone/pentane (0% to 20%), affording **1l** as a white solid (1.71 g, 4.68 mmol, 78%).

M.P.: 44-46 °C

¹H NMR (500 MHz, CDCl₃) δ 7.37 (d, *J* = 8.5 Hz, 2H), 7.33 – 7.27 (m, 5H), 7.27 – 7.23 (m, 2H), 5.86 (s, 1H), 5.74 (s, 1H), 5.53 (d, *J* = 1.2 Hz, 1H), 4.89 (s, 1H), 4.78 (s, 2H).

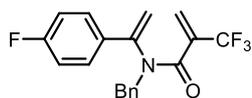
¹³C NMR (126 MHz, CDCl₃) δ 164.5, 145.0, 136.3, 135.5, 134.8 (q, *J* = 31.8 Hz), 134.0, 129.4, 129.1, 128.7, 128.0, 127.5, 123.1 (q, *J* = 5.3 Hz), 121.6 (q, *J* = 274.1 Hz), 115.2, 50.6.

¹⁹F NMR (471 MHz, CDCl₃) δ –64.12.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1663, 1626, 1396, 1342, 1276, 1261, 1171, 1136.

HRMS (ESI⁺) calcd. for C₁₉H₁₆ClF₃NO⁺ [M+H]⁺: 366.0867, found: 366.0860.

N-benzyl-*N*-(1-(4-fluorophenyl)vinyl)-2-(trifluoromethyl)acrylamide (**1m**)



Prepared according to **General Procedure A** using 4-fluoroacetophenone (0.73 mL, 6.0 mmol, 1.0 equiv), toluene (6.0 mL), benzylamine (0.65 mL, 6.0 mmol, 1.0 equiv) and 4 Å molecular sieves (1.80 g). Then using CH₂Cl₂ (20.0 mL), Et₃N (1.67 mL, 12.0 mmol, 2.00 equiv) and 2-(trifluoromethyl)-2-propenoyl chloride⁹ (1.14 g, 7.20 mmol, 1.20 equiv). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of CH₂Cl₂/pentane (0% to 100%), affording **1m** as a white solid (1.47 g, 4.21 mmol, 70%).

M.P.: 88-90 °C

¹H NMR (500 MHz, CDCl₃) δ 7.37 – 7.33 (m, 2H), 7.31 – 7.24 (m, 5H), 7.11 – 7.06 (m, 2H), 5.86 (s, 1H), 5.76 (s, 1H), 5.47 (d, *J* = 1.2 Hz, 1H), 4.87 (s, 1H), 4.79 (s, 2H).

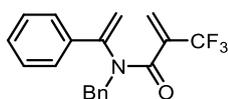
^{13}C NMR (126 MHz, CDCl_3) δ 164.5, 163.4 (d, $J = 249.9$ Hz), 145.1, 136.4, 134.9 (q, $J = 31.8$ Hz), 131.7 (d, $J = 3.5$ Hz), 129.1, 128.6, 128.1 (d, $J = 8.3$ Hz), 127.9, 123.0 (q, $J = 5.3$ Hz), 121.6 (q, $J = 273.9$ Hz), 116.2 (d, $J = 21.9$ Hz), 114.4, 50.7.

^{19}F NMR (471 MHz, CDCl_3) δ -64.13, -111.55.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1661, 1628, 1598, 1507, 1404, 1275, 1228, 1125.

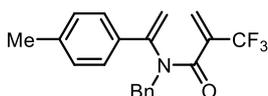
HRMS (ESI $^+$) calcd. for $\text{C}_{19}\text{H}_{16}\text{F}_4\text{NO}^+$ $[\text{M}+\text{H}]^+$: 350.1163, found: 350.1155.

N-benzyl-*N*-(1-phenylvinyl)-2-(trifluoromethyl)acrylamide (**1n**)



Prepared according to **General Procedure A**, known compound.¹⁰

N-benzyl-*N*-(1-(*p*-tolyl)vinyl)-2-(trifluoromethyl)acrylamide (**1o**)



Prepared according to **General Procedure A** using 4-methylacetophenone (0.80 mL, 6.0 mmol, 1.0 equiv), toluene (6.0 mL), benzylamine (0.65 mL, 6.0 mmol, 1.0 equiv) and 4 Å molecular sieves (1.80 g). Then using CH_2Cl_2 (20.0 mL), Et_3N (1.67 mL, 12.0 mmol, 2.00 equiv) and 2-(trifluoromethyl)-2-propenoyl chloride⁹ (1.14 g, 7.20 mmol, 1.20 equiv). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of CH_2Cl_2 /pentane (0% to 100%), affording **1o** as a white solid (610 mg, 1.77 mmol, 29%).

M.P.: 80-82 °C

^1H NMR (500 MHz, CDCl_3) δ 7.33 – 7.19 (m, 9H), 5.82 (s, 1H), 5.76 (s, 1H), 5.49 (s, 1H), 4.78 (s, 2H), 4.77 (s, 1H), 2.39 (s, 3H).

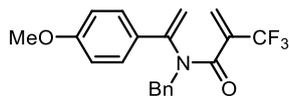
^{13}C NMR (126 MHz, CDCl_3) δ 164.6, 145.7, 139.6, 136.6, 134.8 (q, $J = 31.8$ Hz), 132.4, 129.9, 129.2, 128.6, 127.8, 126.1, 122.6 (q, $J = 5.3$ Hz), 121.7 (q, $J = 274.1$ Hz), 114.0, 50.4, 21.3.

^{19}F NMR (471 MHz, CDCl_3) δ -64.20.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1663, 1627, 1395, 1342, 1275, 1170, 1136, 1119.

HRMS (ESI $^+$) calcd. for $\text{C}_{20}\text{H}_{19}\text{F}_3\text{NO}^+$ $[\text{M}+\text{H}]^+$: 346.1413, found: 346.1404.

N-benzyl-*N*-(1-(4-methoxyphenyl)vinyl)-2-(trifluoromethyl)acrylamide (**1p**)



Prepared according to **General Procedure A** using 4-methoxyacetophenone (1.05 g, 7.00 mmol, 1.00 equiv), toluene (7.0 mL), benzylamine (0.76 mL, 7.0 mmol, 1.0 equiv) and 4 Å molecular sieves (2.1 g). Then using CH₂Cl₂ (25.0 mL), Et₃N (1.95 mL, 14.0 mmol, 2.00 equiv) and 2-(trifluoromethyl)-2-propenoyl chloride⁹ (1.33 g, 8.40 mmol, 1.20 equiv). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/CH₂Cl₂ (0% to 10%), affording **1p** as a colourless oil (1.38 g, 3.82 mmol, 55%).

¹H NMR (500 MHz, CDCl₃) δ 7.33 (d, *J* = 8.8 Hz, 2H), 7.31 – 7.24 (m, 5H), 6.93 (d, *J* = 8.9 Hz, 2H), 5.83 (s, 1H), 5.77 (s, 1H), 5.41 (s, 1H), 4.78 (s, 2H), 4.74 (s, 1H), 3.85 (s, 3H).

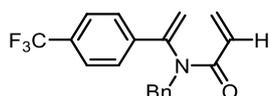
¹³C NMR (126 MHz, CDCl₃) δ 164.6, 160.6, 145.4, 136.6, 134.8 (q, *J* = 31.7 Hz), 129.2, 128.6, 127.79, 127.77, 127.6, 122.5 (q, *J* = 5.3 Hz), 121.7 (q, *J* = 273.9 Hz), 114.5, 112.9, 55.5, 50.4.

¹⁹F NMR (471 MHz, CDCl₃) δ –64.16.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1663, 1627, 1606, 1395, 1277, 1173, 1136, 1120.

HRMS (ESI⁺) calcd. for C₂₀H₁₉F₃NO₂⁺ [M+H]⁺: 362.1362, found: 362.1344.

N-benzyl-*N*-(1-(4-(trifluoromethyl)phenyl)vinyl)acrylamide (**1q**)



Prepared according to **General Procedure A** using 4-(trifluoromethyl)acetophenone (1.13 g, 6.00 mmol, 1.00 equiv), toluene (6.0 mL), benzylamine (0.65 mL, 6.0 mmol, 1.0 equiv) and 4 Å molecular sieves (1.80 g). Then using CH₂Cl₂ (20.0 mL), Et₃N (1.67 mL, 12.0 mmol, 2.00 equiv) and acryloyl chloride (0.59 mL, 7.2 mmol, 1.2 equiv). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **1q** as a colourless oil (1.55 g, 4.68 mmol, 78%).

¹H NMR (500 MHz, CDCl₃) δ 7.64 (d, *J* = 8.2 Hz, 2H), 7.50 (d, *J* = 8.2 Hz, 2H), 7.29 – 7.21 (m, 5H), 6.53 – 6.44 (m, 2H), 5.70 (s, 1H), 5.64 (dd, *J* = 7.6, 4.8 Hz, 1H), 5.00 (s, 1H), 4.70 (s, 2H).

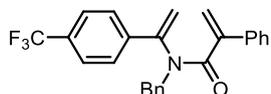
¹³C NMR (126 MHz, CDCl₃) δ 166.1, 144.5, 139.4, 137.0, 131.2 (q, *J* = 32.7 Hz), 129.1, 128.8, 128.4, 128.1, 127.6, 126.5, 126.0 (q, *J* = 3.8 Hz), 123.9 (q, *J* = 272.1 Hz), 117.5, 49.9.

¹⁹F NMR (471 MHz, CDCl₃) δ –62.69.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1657, 1616, 1407, 1321, 1275, 1267, 1223, 1166, 1115.

HRMS (ESI⁺) calcd. for C₁₉H₁₇F₃NO⁺ [M+H]⁺: 332.1257, found: 332.1249.

***N*-benzyl-2-phenyl-*N*-(1-(4-(trifluoromethyl)phenyl)vinyl)acrylamide (**1r**)**



Prepared according to **General Procedure A** using 4-(trifluoromethyl)acetophenone (1.50 g, 8.00 mmol, 1.00 equiv), toluene (8.0 mL), benzylamine (0.87 mL, 8.0 mmol, 1.0 equiv) and 4 Å molecular sieves (2.40 g). Then using CH₂Cl₂ (30.0 mL), Et₃N (2.23 mL, 16.0 mmol, 2.00 equiv) and crude 2-phenylacryloyl chloride^A (12.0 mmol, 1.50 equiv) in CH₂Cl₂ (9.0 mL). The crude reaction mixture was purified by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient elution of acetone/pentane (0% to 20%) and Et₂O/pentane (0% to 50%), affording **1r** as a colourless oil (1.81 g, 4.45 mmol, 56%).

Notes: (A) Crude 2-phenylacryloyl chloride was prepared according to **General Procedure B** (acyl chloride synthesis step) using 2-phenylacrylic acid (1.78 g, 12.0 mmol, 1.50 equiv), CH₂Cl₂ (40.0 mL), oxalyl chloride (1.22 mL, 14.4 mmol, 1.80 equiv) and DMF (5 drops).

¹H NMR (500 MHz, CDCl₃) δ 7.49 (d, J = 8.1 Hz, 2H), 7.33 – 7.21 (m, 12H), 5.46 (s, 1H), 5.39 (s, 1H), 5.37 (s, 1H), 4.84 (s, 2H), 4.79 (s, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 170.7, 145.5, 145.1, 139.7, 137.1, 136.8, 130.7 (q, J = 32.4 Hz), 129.1, 128.6, 128.5, 128.3, 127.8, 126.7, 126.2, 125.6 (q, J = 3.8 Hz), 124.0 (q, J = 272.3 Hz), 116.1, 116.0, 50.4.

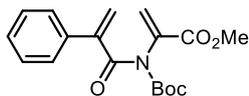
¹⁹F NMR (471 MHz, CDCl₃) δ -62.68.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1651, 1617, 1388, 1324, 1276, 1261, 1167, 1123, 1064.

HRMS (ESI⁺) calcd. for C₂₅H₂₁F₃NO⁺ [M+H]⁺: 408.1570, found: 408.1563.

2.3.2 Synthesis and Characterisation of Precursors (7)

Methyl 2-(*N*-(*tert*-butoxycarbonyl)-2-phenylacrylamido)acrylate (**7a**)



Prepared according to **General Procedure B** using 2-phenylacrylic acid (5.92 g, 40.0 mmol, 2.00 equiv), CH₂Cl₂ (133.0 mL), oxalyl chloride (5.15 mL, 60.0 mmol, 3.00 equiv) and DMF (5 drops). Then using CH₂Cl₂ (67.0 mL), Et₃N (11.2 mL, 80.0 mmol, 4.00 equiv), methyl 2-((*tert*-butoxycarbonyl)amino)acrylate¹¹ (4.02 g, 20.0 mmol, 1.00 equiv) in CH₂Cl₂ (20.0 mL), and DMAP (1.22 g, 10.0 mmol, 0.50 equiv). The crude reaction mixture was purified by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%) and ethyl acetate/CH₂Cl₂ (0% to 10%), affording **7a** (5.10 g, 15.41 mmol, 77%) as a colourless oil.

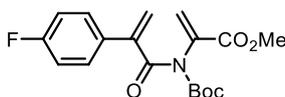
¹H NMR (500 MHz, CDCl₃) δ 7.52 – 7.48 (m, 2H), 7.37 – 7.32 (m, 2H), 7.31 – 7.28 (m, 1H), 6.49 (d, *J* = 0.9 Hz, 1H), 5.79 (s, 1H), 5.78 (d, *J* = 0.9 Hz, 1H), 5.74 (s, 1H), 3.80 (s, 3H), 1.24 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 171.5, 163.6, 151.2, 146.2, 135.9, 135.5, 128.5, 128.4, 126.7, 126.1, 117.8, 84.3, 52.6, 27.5.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1732, 1689, 1439, 1368, 1275, 1212, 1113, 1091.

HRMS (ESI⁺) calcd. for C₁₈H₂₁NO₅Na⁺ [M+Na]⁺: 354.1312, found: 354.1296.

Methyl 2-(*N*-(*tert*-butoxycarbonyl)-2-(4-fluorophenyl)acrylamido)acrylate (**7b**)



Prepared according to **General Procedure B** using 2-(4-fluorophenyl)acrylic acid¹² (664 mg, 4.00 mmol, 2.00 equiv), CH₂Cl₂ (13.3 mL), oxalyl chloride (0.52 mL, 6.0 mmol, 3.0 equiv) and DMF (3 drops). Then using CH₂Cl₂ (6.70 mL), Et₃N (1.12 mL, 8.00 mmol, 4.00 equiv), methyl 2-((*tert*-butoxycarbonyl)amino)acrylate¹¹ (402 mg, 2.00 mmol, 1.00 equiv) in CH₂Cl₂ (2.0 mL), and DMAP (122 mg, 1.00 mmol, 0.50 equiv). The crude reaction mixture was purified by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 20%) and Et₂O/CH₂Cl₂ (5% to 10%), affording **7b** (402 mg, 1.15 mmol, 58%) as a colourless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.50 – 7.45 (m, 2H), 7.03 – 6.98 (m, 2H), 6.46 (d, *J* = 0.9 Hz, 1H), 5.74 (d, *J* = 0.9 Hz, 1H), 5.70 (s, 1H), 5.69 (s, 1H), 3.77 (s, 3H), 1.24 (s, 9H).

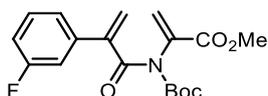
^{13}C NMR (126 MHz, CDCl_3) δ 171.2, 163.4, 162.7 (d, $J = 248.0$ Hz), 151.1, 145.1, 135.3, 132.1 (d, $J = 3.5$ Hz), 128.6 (d, $J = 8.2$ Hz), 126.1, 117.4, 115.3 (d, $J = 21.7$ Hz), 84.3, 52.6, 27.4.

^{19}F NMR (471 MHz, CDCl_3) δ -113.41.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1736, 1692, 1511, 1370, 1275, 1225, 1155, 1114.

HRMS (ESI⁺) calcd. for $\text{C}_{18}\text{H}_{20}\text{FNO}_5\text{Na}^+$ [$\text{M}+\text{Na}$]⁺: 372.1218, found: 372.1201.

Methyl 2-(*N*-(*tert*-butoxycarbonyl)-2-(3-fluorophenyl)acrylamido)acrylate (7c)



Prepared according to **General Procedure B** using 2-(3-fluorophenyl)acrylic acid¹² (664 mg, 4.00 mmol, 2.00 equiv), CH_2Cl_2 (13.3 mL), oxalyl chloride (0.52 mL, 6.0 mmol, 3.0 equiv) and DMF (3 drops). Then using CH_2Cl_2 (6.70 mL), Et_3N (1.12 mL, 8.00 mmol, 4.00 equiv), methyl 2-((*tert*-butoxycarbonyl)amino)acrylate¹¹ (402 mg, 2.00 mmol, 1.00 equiv) in CH_2Cl_2 (2.0 mL), and DMAP (122 mg, 1.00 mmol, 0.50 equiv). The crude reaction mixture was purified by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 20%) and $\text{Et}_2\text{O}/\text{CH}_2\text{Cl}_2$ (5% to 10%), affording **7c** (351 mg, 1.01 mmol, 50%) as a colourless oil.

^1H NMR (500 MHz, CDCl_3) δ 7.33 – 7.27 (m, 2H), 7.25 – 7.21 (m, 1H), 7.01 – 6.96 (m, 1H), 6.50 (d, $J = 0.9$ Hz, 1H), 5.79 (s, 1H), 5.771 (s, 1H), 5.768 (s, 1H), 3.81 (s, 3H), 1.25 (s, 9H).

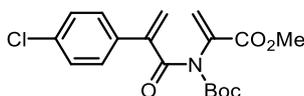
^{13}C NMR (126 MHz, CDCl_3) δ 170.9, 163.5, 162.9 (d, $J = 245.5$ Hz), 151.2, 145.2 (d, $J = 2.5$ Hz), 138.1 (d, $J = 7.8$ Hz), 135.3, 130.1 (d, $J = 8.3$ Hz), 126.3, 122.5 (d, $J = 3.0$ Hz), 118.6, 115.3 (d, $J = 21.0$ Hz), 113.7 (d, $J = 22.6$ Hz), 84.5, 52.7, 27.5.

^{19}F NMR (471 MHz, CDCl_3) δ -113.05.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1737, 1693, 1581, 1440, 1370, 1284, 1259, 1154, 1109.

HRMS (ESI⁺) calcd. for $\text{C}_{18}\text{H}_{20}\text{FNO}_5\text{Na}^+$ [$\text{M}+\text{Na}$]⁺: 372.1218, found: 372.1203.

Methyl 2-(*N*-(*tert*-butoxycarbonyl)-2-(4-chlorophenyl)acrylamido)acrylate (7d)



Prepared according to **General Procedure B** using 2-(4-chlorophenyl)acrylic acid¹³ (728 mg, 4.00 mmol, 2.00 equiv), CH_2Cl_2 (13.3 mL), oxalyl chloride (0.52 mL, 6.0 mmol, 3.0 equiv) and DMF (3

drops). Then using CH₂Cl₂ (6.70 mL), Et₃N (1.12 mL, 8.00 mmol, 4.00 equiv), methyl 2-((*tert*-butoxycarbonyl)amino)acrylate¹¹ (402 mg, 2.00 mmol, 1.00 equiv) in CH₂Cl₂ (2.0 mL) and DMAP (122 mg, 1.00 mmol, 0.50 equiv). The crude reaction mixture was purified by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 20%) and Et₂O/CH₂Cl₂ (5% to 10%), affording **7d** (410 mg, 1.12 mmol, 56%) as a colourless oil.

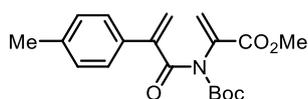
¹H NMR (500 MHz, CDCl₃) δ 7.44 (d, *J* = 8.7 Hz, 2H), 7.31 (d, *J* = 8.6 Hz, 2H), 6.48 (d, *J* = 0.9 Hz, 1H), 5.76 (d, *J* = 1.0 Hz, 1H), 5.75 (s, 1H), 5.73 (s, 1H), 3.79 (s, 3H), 1.25 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 171.1, 163.5, 151.2, 145.2, 135.3, 134.4, 134.3, 128.7, 128.1, 126.2, 118.0, 84.4, 52.7, 27.5.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1735, 1691, 1581, 1493, 1369, 1285, 1257, 1154, 1111.

HRMS (ESI⁺) calcd. for C₁₈H₂₀ClNO₅Na⁺ [M+Na]⁺: 388.0922, found: 388.0905.

Methyl 2-(*N*-(*tert*-butoxycarbonyl)-2-(*p*-tolyl)acrylamido)acrylate (**7e**)



Prepared according to **General Procedure B** using 2-(*p*-tolyl)acrylic acid (648 mg, 4.00 mmol, 2.00 equiv)¹², CH₂Cl₂ (13.3 mL), oxalyl chloride (0.52 mL, 6.0 mmol, 3.0 equiv) and DMF (3 drops). Then using CH₂Cl₂ (6.70 mL), Et₃N (1.12 mL, 8.00 mmol, 4.00 equiv), methyl 2-((*tert*-butoxycarbonyl)amino)acrylate¹¹ (402 mg, 2.00 mmol, 1.00 equiv) in CH₂Cl₂ (2.0 mL) and DMAP (122 mg, 1.00 mmol, 0.50 equiv). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 20%), affording **7e** (450 mg, 1.30 mmol, 65%) as a colourless oil.

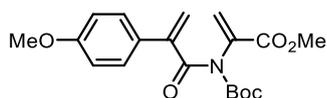
¹H NMR (500 MHz, CDCl₃) δ 7.39 (d, *J* = 8.2 Hz, 2H), 7.15 (d, *J* = 8.0 Hz, 2H), 6.48 (s, 1H), 5.77 (s, 1H), 5.75 (d, *J* = 1.0 Hz, 1H), 5.68 (d, *J* = 1.1 Hz, 1H), 3.80 (s, 3H), 2.33 (s, 3H), 1.26 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 171.7, 163.6, 151.3, 146.1, 138.3, 135.5, 133.0, 129.2, 126.6, 126.1, 116.6, 84.3, 52.6, 27.5, 21.3.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1734, 1691, 1643, 1515, 1439, 1369, 1286, 1211, 1155, 1084, 975.

HRMS (ESI⁺) calcd. for C₁₉H₂₃NO₅Na⁺ [M+Na]⁺: 368.1468, found: 368.1451.

Methyl 2-(*N*-(*tert*-butoxycarbonyl)-2-(4-methoxyphenyl)acrylamido)acrylate (**7f**)



Prepared according to **General Procedure B** using 2-(4-methoxyphenyl)acrylic acid¹² (712 mg, 4.00 mmol, 2.00 equiv), CH₂Cl₂ (13.3 mL), oxalyl chloride (0.52 mL, 6.0 mmol, 3.0 equiv) and DMF (3 drops). Then using CH₂Cl₂ (6.70 mL), Et₃N (1.12 mL, 8.00 mmol, 4.00 equiv), methyl 2-((*tert*-butoxycarbonyl)amino)acrylate¹¹ (402 mg, 2.00 mmol, 1.00 equiv) in CH₂Cl₂ (2.0 mL) and DMAP (122 mg, 1.00 mmol, 0.50 equiv). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **7f** (430 mg, 1.19 mmol, 60%) as a colourless oil.

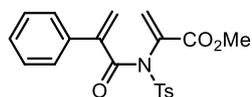
¹H NMR (500 MHz, CDCl₃) δ 7.44 (d, *J* = 8.9 Hz, 2H), 6.87 (d, *J* = 8.8 Hz, 2H), 6.48 (s, 1H), 5.77 (s, 1H), 5.69 (s, 1H), 5.62 (s, 1H), 3.81 (s, 3H), 3.79 (s, 3H), 1.27 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 171.8, 163.6, 159.9, 151.4, 145.7, 135.5, 128.5, 128.1, 126.1, 115.6, 113.9, 84.3, 55.4, 52.7, 27.6.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1733, 1690, 1608, 1514, 1440, 1369, 1279, 1216, 1154, 1113.

HRMS (ESI⁺) calcd. for C₁₉H₂₃NO₆Na⁺ [M+Na]⁺: 384.1418, found: 384.1406.

Methyl 2-(2-phenyl-*N*-tosylacrylamido)acrylate (**7g**)



Prepared according to **General Procedure B** using 2-phenylacrylic acid (4.74 g, 32.0 mmol, 2.00 equiv), CH₂Cl₂ (100.0 mL), oxalyl chloride (4.06 mL, 48.0 mmol, 3.00 equiv) and DMF (6 drops). Then using CH₂Cl₂ (54.0 mL), Et₃N (8.92 mL, 64.0 mmol, 4.00 equiv), methyl 2-((4-methylphenyl)sulfonamido)acrylate¹⁴ (4.10 g, 16.0 mmol, 1.00 equiv) in CH₂Cl₂ (16.0 mL) and DMAP (976 mg, 8.00 mmol, 0.50 equiv). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **7g** (1.55 g, 4.03 mmol, 25%) as a colourless oil.

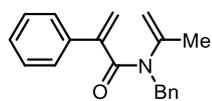
¹H NMR (500 MHz, CDCl₃) δ 8.02 (d, *J* = 8.4 Hz, 2H), 7.35 – 7.31 (m, 2H), 7.30 – 7.25 (m, 3H), 7.23 – 7.19 (m, 2H), 6.40 (d, *J* = 0.9 Hz, 1H), 5.67 (d, *J* = 0.9 Hz, 1H), 5.61 (s, 1H), 5.45 (s, 1H), 3.63 (s, 3H), 2.43 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 168.4, 163.3, 145.3, 144.1, 135.2, 134.7, 133.6, 132.0, 129.7, 129.2, 128.9, 128.7, 126.3, 119.0, 52.7, 21.7.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1733, 1699, 1631, 1438, 1362, 1321, 1206, 1166, 1084, 1048.

HRMS (ESI⁺) calcd. for C₂₀H₁₉NO₅SNa⁺ [M+Na]⁺: 408.0876, found: 408.0857.

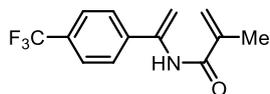
***N*-benzyl-2-phenyl-*N*-(prop-1-en-2-yl)acrylamide (7h)**



Prepared according to **General Procedure A**, known compound.¹⁵

2.3.3 Synthesis and Characterisation of Precursors (2)

N-(1-(4-(trifluoromethyl)phenyl)vinyl)methacrylamide (**S2a**)



Prepared according to **General Procedure C4 (step C4-1)** using 4-(trifluoromethyl)benzotrile (3.42 g, 20.0 mmol, 1.00 equiv), THF (20.0 mL), MeLi (8.40 mL, 3.1 M in diethoxymethane, 26.0 mmol, 1.30 equiv), MeOH (1.22 mL, 30.0 mmol, 1.50 equiv), CH₂Cl₂ (ca. 30–40 mL), Et₃N (5.58 mL, 40.0 mmol, 2.00 equiv) and methacryloyl chloride (2.93 mL, 30.0 mmol, 1.50 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 25%) afforded **S2a** (2.70 g, 10.6 mmol, 53%) as a white solid.

M.P.: 66–68 °C

¹H NMR (500 MHz, CDCl₃) δ 7.60 (d, *J* = 8.2 Hz, 2H), 7.51 (d, *J* = 8.2 Hz, 2H), 7.41 (s, 1H), 5.81 (s, 1H), 5.74 (s, 1H), 5.42 (q, *J* = 1.7 Hz, 1H), 5.18 (s, 1H), 1.99 (s, 3H).

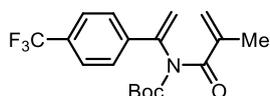
¹³C NMR (126 MHz, CDCl₃) δ 167.3, 141.8, 140.4, 139.8, 130.6 (q, *J* = 32.5 Hz), 126.4, 125.7 (q, *J* = 3.8 Hz), 124.0 (q, *J* = 272.1 Hz), 120.3, 105.5, 18.7.

¹⁹F NMR (471 MHz, CDCl₃) δ –62.68.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1664, 1619, 1500, 1320, 1163, 1114, 1064, 1015.

HRMS (ESI⁺) calcd. for C₁₃H₁₃F₃NO⁺ [M+H]⁺: 256.0944, found: 256.0941.

Tert-butyl methacryloyl(1-(4-(trifluoromethyl)phenyl)vinyl)carbamate (**2a**)



Prepared according to **General Procedure C4 (step C4-2)** using **S2a** (510 mg, 2.00 mmol, 1.00 equiv) and THF (10.0 mL), then *n*-BuLi (0.80 mL, 2.5 M in hexane, 2.0 mmol, 1.0 equiv), and di-*tert*-butyl dicarbonate (523 mg, 2.40 mmol, 1.20 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 30%) afforded **2a** (590 mg, 1.66 mmol, 83 %) as a white solid.

M.P.: 75–77 °C

¹H NMR (500 MHz, CDCl₃) δ 7.60 (d, *J* = 8.4 Hz, 2H), 7.55 (d, *J* = 8.5 Hz, 2H), 5.74 (d, *J* = 1.2 Hz, 1H), 5.58 (s, 1H), 5.41 (q, *J* = 1.8 Hz, 1H), 5.29 (d, *J* = 1.3 Hz, 1H), 2.07 (s, 3H), 1.31 (s, 9H).

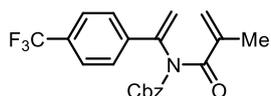
^{13}C NMR (126 MHz, CDCl_3) δ 173.8, 152.6, 143.5, 142.5, 140.9, 130.7 (q, $J = 32.5$ Hz), 126.2, 125.7 (q, $J = 3.8$ Hz), 124.1 (q, $J = 272.1$ Hz), 119.4, 115.5, 83.9, 27.7, 19.0.

^{19}F NMR (471 MHz, CDCl_3) δ -62.68.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1737, 1690, 1636, 1370, 1323, 1276, 1155, 1115, 1064.

HRMS (ESI⁺) calcd. for $\text{C}_{18}\text{H}_{20}\text{F}_3\text{NO}_3\text{Na}^+$ [$\text{M}+\text{Na}$]⁺: 378.1287, found: 378.1285.

Benzyl methacryloyl(1-(4-(trifluoromethyl)phenyl)vinyl)carbamate (**2b**)



Prepared according to **General Procedure C4 (step C4-2)** using **S2a** (510 mg, 2.00 mmol, 1.00 equiv) and THF (10.0 mL), then *n*-BuLi (0.80 mL, 2.5 M in hexane, 2.0 mmol, 1.0 equiv) and benzyl chloroformate (0.34 mL, 2.4 mmol, 1.2 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 30%) afforded **2b** (660 mg, 1.70 mmol, 85 %) as a white solid.

M.P.: 58-60 °C

^1H NMR (500 MHz, CDCl_3) δ 7.55 (d, $J = 8.2$ Hz, 2H), 7.49 (d, $J = 8.1$ Hz, 2H), 7.31 – 7.23 (m, 3H), 7.09 – 7.05 (m, 2H), 5.79 (d, $J = 1.4$ Hz, 1H), 5.55 (s, 1H), 5.39 (q, $J = 1.6$ Hz, 1H), 5.31 (d, $J = 1.4$ Hz, 1H), 5.10 (s, 2H), 2.04 (dd, $J = 1.6, 1.0$ Hz, 3H).

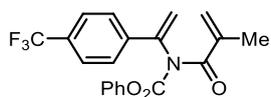
^{13}C NMR (126 MHz, CDCl_3) δ 173.5, 153.9, 143.0, 142.0, 140.3 (q, $J = 1.6$ Hz), 134.5, 130.7 (q, $J = 32.5$ Hz), 128.7, 128.6, 128.5, 126.0, 125.8 (q, $J = 3.8$ Hz), 124.1 (q, $J = 272.1$ Hz), 120.0, 116.1, 69.1, 18.9.

^{19}F NMR (471 MHz, CDCl_3) δ -62.64.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1739, 1694, 1636, 1456, 1408, 1380, 1322, 1261, 1166, 1115.

HRMS (ESI⁺) calcd. for $\text{C}_{21}\text{H}_{19}\text{F}_3\text{NO}_3^+$ [$\text{M}+\text{H}$]⁺: 390.1312, found: 390.1315.

Phenyl methacryloyl(1-(4-(trifluoromethyl)phenyl)vinyl)carbamate (**2c**)



Prepared according to **General Procedure C4 (step C4-2)** using **S2a** (510 mg, 2.00 mmol, 1.00 equiv) and THF (10.0 mL), then *n*-BuLi (0.80 mL, 2.5 M in hexane, 2.0 mmol, 1.0 equiv) and phenyl chloroformate (0.30 mL, 2.4 mmol, 1.2 equiv). Purification by Biotage automated flash

chromatography system (ethyl acetate/pentane, 0% to 30%) afforded **2c** (610 mg, 1.63 mmol, 81 %) as a white solid.

M.P.: 88-90 °C

¹H NMR (500 MHz, CDCl₃) δ 7.67 (s, 4H), 7.35 – 7.30 (m, 2H), 7.24 – 7.19 (m, 1H), 6.93 – 6.89 (m, 2H), 5.90 (d, *J* = 1.5 Hz, 1H), 5.74 (d, *J* = 1.2 Hz, 1H), 5.54 (q, *J* = 1.6 Hz, 1H), 5.49 (d, *J* = 1.5 Hz, 1H), 2.13 (t, *J* = 1.3 Hz, 3H).

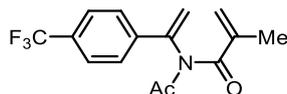
¹³C NMR (126 MHz, CDCl₃) δ 173.3, 152.6, 150.3, 142.9, 142.0, 140.2, 131.0 (q, *J* = 32.7 Hz), 129.6, 126.5, 126.2, 125.9 (q, *J* = 3.8 Hz), 124.0 (q, *J* = 272.1 Hz), 120.9, 120.4, 116.5, 18.9.

¹⁹F NMR (471 MHz, CDCl₃) δ –62.66.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1750, 1701, 1637, 1618, 1490, 1409, 1325, 1264, 1158, 1115, 1062.

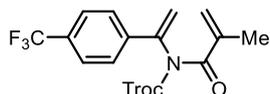
HRMS (ESI⁺) calcd. for C₂₀H₁₇F₃NO₃⁺ [M+H]⁺: 376.1155, found: 376.1152.

N-acetyl-*N*-(1-(4-(trifluoromethyl)phenyl)vinyl)methacrylamide (**2d**)



Prepared according to **General Procedure C1**, known compound.¹⁶

2,2,2-Trichloroethyl methacryloyl(1-(4-(trifluoromethyl)phenyl)vinyl)carbamate (**2e**)



Prepared according to **General Procedure C4 (step C4-2)** using **S2a** (510 mg, 2.00 mmol, 1.00 equiv) and THF (10.0 mL), then *n*-BuLi (0.80 mL, 2.5 M in hexane, 2.0 mmol, 1.0 equiv) and 2,2,2-trichloroethyl chloroformate (0.33 mL, 2.4 mmol, 1.2 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 25%) afforded **2e** (740 mg, 1.72 mmol, 86 %) as a colourless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.62 (d, *J* = 9.0 Hz, 2H), 7.58 (d, *J* = 8.4 Hz, 2H), 5.87 (d, *J* = 1.5 Hz, 1H), 5.64 (s, 1H), 5.51 (q, *J* = 1.6 Hz, 1H), 5.41 (d, *J* = 1.5 Hz, 1H), 4.72 (s, 2H), 2.09 (s, 3H).

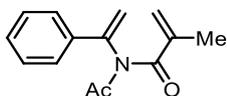
¹³C NMR (126 MHz, CDCl₃) δ 172.9, 152.3, 142.6, 141.5, 139.9, 131.0 (q, *J* = 32.7 Hz), 126.3, 125.8 (q, *J* = 3.7 Hz), 124.0 (q, *J* = 272.1 Hz), 121.0, 116.9, 94.1, 75.8, 18.9.

¹⁹F NMR (471 MHz, CDCl₃) δ –62.72.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1749, 1700, 1637, 1619, 1452, 1376, 1260, 1166, 1116, 1064.

HRMS (ESI⁺) calcd. for C₁₆H₁₄Cl₃F₃NO₃⁺ [M+H]⁺: 429.9986, found: 429.9986.

***N*-acetyl-*N*-(1-phenylvinyl) methacrylamide (2f)**



Prepared according to **General Procedure C1** using *N*-(1-phenylvinyl)acetamide **S2f** (483 mg, 3.00 mmol, 1.00 equiv) and CH₂Cl₂ (12.0 mL), then Et₃N (1.25 mL, 9.00 mmol, 3.00 equiv), methacryloyl chloride (0.59 mL, 6.0 mmol, 2.0 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 40%) afforded **2f** (460 mg, 2.01 mmol, 67 %) as a white solid.

M.P.: 65-67 °C

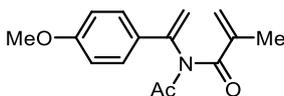
¹H NMR (500 MHz, CDCl₃) δ 7.45 – 7.30 (m, 5H), 5.83 (s, 1H), 5.51 (s, 1H), 5.33 (s, 1H), 5.21 (s, 1H), 2.32 (s, 3H), 1.92 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 174.2, 173.5, 144.7, 141.4, 136.0, 129.1, 128.9, 125.6, 120.2, 114.8, 25.3, 19.2.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1689, 1632, 1495, 1446, 1365, 1316, 1246, 1170, 1027.

HRMS (ESI⁺) calcd. for C₁₄H₁₅NO₂Na⁺ [M+Na]⁺: 252.0995, found: 252.0987.

***N*-acetyl-*N*-(1-(4-methoxyphenyl)vinyl)methacrylamide (2g)**



Prepared according to **General Procedure C1** using *N*-(1-(4-methoxyphenyl)vinyl)acetamide **S2g** (383 mg, 2.00 mmol, 1.00 equiv) and CH₂Cl₂ (8.0 mL), then Et₃N (0.84 mL, 6.0 mmol, 3.0 equiv), methacryloyl chloride (0.39 mL, 4.0 mmol, 2.0 equiv), and DMAP (73 mg, 0.60 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 50%) afforded **2g** (410 mg, 1.58 mmol, 79 %) as a colourless oil.

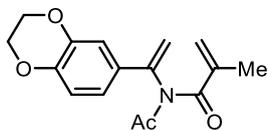
¹H NMR (500 MHz, CDCl₃) δ 7.33 (d, *J* = 8.8 Hz, 2H), 6.83 (d, *J* = 8.8 Hz, 2H), 5.67 (s, 1H), 5.47 (s, 1H), 5.28 (s, 1H), 5.06 (s, 1H), 3.74 (s, 3H), 2.27 (s, 3H), 1.90 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 174.1, 173.4, 160.1, 144.1, 141.2, 128.4, 126.9, 119.8, 114.1, 112.6, 55.2, 25.0, 19.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1691, 1606, 1511, 1366, 1302, 1248, 1176, 1028, 892.

HRMS (ESI⁺) calcd. for C₁₅H₁₇NO₃Na⁺ [M+Na]⁺: 282.1101, found: 282.1094.

***N*-acetyl-*N*-(1-(2,3-dihydrobenzo[*b*][1,4]dioxin-6-yl)vinyl)methacrylamide (2h)**



Prepared according to **General Procedure C1** using *N*-(1-(2,3-dihydrobenzo[*b*][1,4]dioxin-6-yl)vinyl)acetamide **S2h** (548 mg, 2.50 mmol, 1.00 equiv) and CH₂Cl₂ (10.0 mL), then Et₃N (1.05 mL, 7.50 mmol, 3.00 equiv), methacryloyl chloride (0.49 mL, 5.0 mmol, 2.0 equiv), and DMAP (92 mg, 0.75 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 50%) afforded **2h** (550 mg, 1.92 mmol, 77 %) as a colourless oil.

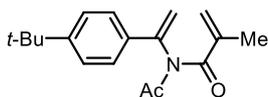
¹H NMR (500 MHz, CDCl₃) δ 6.94 – 6.90 (m, 2H), 6.81 (d, *J* = 8.3 Hz, 1H), 5.69 (s, 1H), 5.49 (s, 1H), 5.31 (d, *J* = 1.7 Hz, 1H), 5.08 (d, *J* = 1.1 Hz, 1H), 4.27 – 4.19 (m, 4H), 2.30 (s, 3H), 1.94 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 174.1, 173.5, 144.5, 144.0, 143.7, 141.4, 129.5, 120.0, 118.9, 117.7, 114.6, 113.3, 64.5, 64.3, 25.2, 19.2.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1691, 1629, 1580, 1506, 1457, 1366, 1276, 1199, 1065.

HRMS (ESI⁺) calcd. for C₁₆H₁₇NO₄Na⁺ [M+Na]⁺: 310.1050, found: 310.1039.

***N*-acetyl-*N*-(1-(4-(*tert*-butyl)phenyl)vinyl)methacrylamide (2i)**



Prepared according to **General Procedure C1** using *N*-(1-(4-(*tert*-butyl)phenyl)vinyl)acetamide **S2i** (651 mg, 3.00 mmol, 1.00 equiv) and CH₂Cl₂ (12.0 mL), then Et₃N (1.25 mL, 9.00 mmol, 3.00 equiv), methacryloyl chloride (0.59 mL, 6.0 mmol, 2.0 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 40%) afforded **2i** (630 mg, 2.21 mmol, 74 %) as a colourless oil.

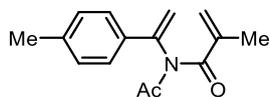
¹H NMR (500 MHz, CDCl₃) δ 7.37 (s, 4H), 5.81 (d, *J* = 1.0 Hz, 1H), 5.52 (s, 1H), 5.33 (d, *J* = 0.9 Hz, 1H), 5.16 (d, *J* = 1.0 Hz, 1H), 2.33 (s, 3H), 1.95 (s, 3H), 1.31 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 174.3, 173.6, 152.4, 144.6, 141.5, 133.0, 125.9, 125.4, 120.0, 114.0, 34.8, 31.3, 25.3, 19.3.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1713, 1695, 1632, 1366, 1299, 1274, 1258, 1176, 840.

HRMS (ESI⁺) calcd. for C₁₈H₂₃NO₂Na⁺ [M+Na]⁺: 308.1621, found: 308.1606.

***N*-acetyl-*N*-(1-(*p*-tolyl)vinyl)methacrylamide (2j)**



Prepared according to **General Procedure C1** using *N*-(1-(*p*-tolyl)vinyl)acetamide **S2j** (525 mg, 3.00 mmol, 1.00 equiv) and CH₂Cl₂ (12.0 mL), then Et₃N (1.25 mL, 9.00 mmol, 3.00 equiv), methacryloyl chloride (0.59 mL, 6.0 mmol, 2.0 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 40%) afforded **2j** (500 mg, 2.06 mmol, 69 %) as a white solid.

M.P.: 37-39 °C

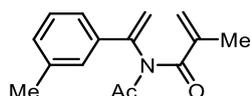
¹H NMR (500 MHz, CDCl₃) δ 7.32 (d, *J* = 8.2 Hz, 2H), 7.15 (d, *J* = 8.0 Hz, 2H), 5.78 (s, 1H), 5.51 (s, 1H), 5.31 (s, 1H), 5.15 (s, 1H), 2.33 (s, 3H), 2.31 (s, 3H), 1.93 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 174.2, 173.5, 144.6, 141.4, 139.1, 133.1, 129.6, 125.5, 120.0, 113.8, 25.2, 21.2, 19.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1691, 1629, 1512, 1452, 1366, 1315, 1295, 1250, 1187, 1109, 1017, 893.

HRMS (ESI⁺) calcd. for C₁₅H₁₇NO₂Na⁺ [M+Na]⁺: 266.1152, found: 266.1143.

***N*-Acetyl-*N*-(1-(*m*-tolyl)vinyl)methacrylamide (2k)**



Prepared according to **General Procedure C1** using *N*-(1-(*m*-tolyl)vinyl)acetamide **S2k** (525 mg, 3.00 mmol, 1.00 equiv) and CH₂Cl₂ (12.0 mL), then Et₃N (1.25 mL, 9.00 mmol, 3.00 equiv), methacryloyl chloride (0.59 mL, 6.0 mmol, 2.0 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 35%) afforded **2k** (480 mg, 1.98 mmol, 66 %) as a white solid.

M.P.: 39-41 °C

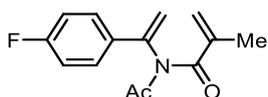
¹H NMR (500 MHz, CDCl₃) δ 7.26 – 7.20 (m, 3H), 7.15 – 7.12 (m, 1H), 5.81 (d, *J* = 1.0 Hz, 1H), 5.51 (s, 1H), 5.32 (d, *J* = 1.7 Hz, 1H), 5.19 (d, *J* = 1.1 Hz, 1H), 2.35 (s, 3H), 2.32 (s, 3H), 1.94 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 174.2, 173.5, 144.8, 141.4, 138.5, 135.9, 129.9, 128.8, 126.3, 122.8, 120.1, 114.5, 25.2, 21.5, 19.1.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1691, 1629, 1512, 1453, 1366, 1275, 1260, 1162, 907.

HRMS (ESI $^+$) calcd. for $\text{C}_{15}\text{H}_{17}\text{NO}_2\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 266.1152, found: 266.1144.

N-acetyl-*N*-(1-(4-fluorophenyl)vinyl)methacrylamide (**2l**)



Prepared according to **General Procedure C1** using *N*-(1-(4-fluorophenyl)vinyl)acetamide **S2l** (537 mg, 3.00 mmol, 1.00 equiv) and CH_2Cl_2 (12.0 mL), then Et_3N (1.25 mL, 9.00 mmol, 3.00 equiv), methacryloyl chloride (0.59 mL, 6.0 mmol, 2.0 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 40%) afforded **2l** (530 mg, 2.15 mmol, 72 %) as a white solid.

M.P.: 55-57 $^\circ\text{C}$

^1H NMR (500 MHz, CDCl_3) δ 7.41-7.36 (m, 2H), 7.06 – 6.99 (m, 2H), 5.74 (s, 1H), 5.48 (s, 1H), 5.33 (d, J = 2.1 Hz, 1H), 5.18 (s, 1H), 2.31 (s, 3H), 1.91 (s, 3H).

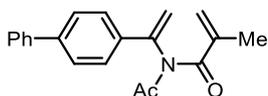
^{13}C NMR (126 MHz, CDCl_3) δ 174.1, 173.4, 163.1 (d, J = 249.4 Hz), 143.8, 141.3, 132.3 (d, J = 3.4 Hz), 127.6 (d, J = 8.3 Hz), 120.5, 115.9 (d, J = 21.9 Hz), 114.6, 25.2, 19.1.

^{19}F NMR (471 MHz, CDCl_3) δ -112.17.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1691, 1632, 1602, 1508, 1453, 1366, 1306, 1231, 1160, 1110, 1012, 895.

HRMS (ESI $^+$) calcd. for $\text{C}_{14}\text{H}_{14}\text{FNO}_2\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 270.0901, found: 270.0898.

N-(1-([1,1'-biphenyl]-4-yl)vinyl)-*N*-acetylmethacrylamide (**2m**)



Prepared according to **General Procedure C1** using *N*-(1-([1,1'-biphenyl]-4-yl)vinyl)acetamide **S2m** (474 mg, 2.00 mmol, 1.00 equiv) and CH_2Cl_2 (8.0 mL), then Et_3N (0.84 mL, 6.00 mmol, 3.00 equiv), methacryloyl chloride (0.39 mL, 4.0 mmol, 2.0 equiv), and DMAP (73 mg, 0.60 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 50%) afforded **2m** (440 mg, 1.44 mmol, 72 %) as a white solid.

M.P.: 140-142 °C

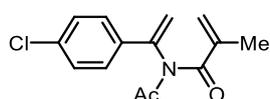
¹H NMR (500 MHz, CDCl₃) δ 7.63 – 7.58 (m, 4H), 7.54 – 7.50 (m, 2H), 7.48 – 7.43 (m, 2H), 7.39 – 7.34 (m, 1H), 5.90 (d, *J* = 1.2 Hz, 1H), 5.56 (s, 1H), 5.37 (q, *J* = 1.6 Hz, 1H), 5.25 (d, *J* = 1.2 Hz, 1H), 2.38 (s, 3H), 1.98 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 174.2, 173.5, 144.4, 141.9, 141.4, 140.2, 134.8, 128.9, 127.8, 127.6, 127.1, 126.1, 120.3, 114.7, 25.3, 19.2.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1712, 1685, 1637, 1607, 1487, 1366, 1311, 1265, 1166, 846.

HRMS (ESI⁺) calcd. for C₂₀H₁₉NO₂Na⁺ [M+Na]⁺: 328.1308, found: 328.1298.

***N*-acetyl-*N*-(1-(4-chlorophenyl)vinyl)methacrylamide (2n)**



Prepared according to **General Procedure C1** using *N*-(1-(4-chlorophenyl)vinyl)acetamide **S2n** (390 mg, 2.00 mmol, 1.00 equiv) and CH₂Cl₂ (8.0 mL), then Et₃N (0.84 mL, 6.0 mmol, 3.0 equiv), methacryloyl chloride (0.39 mL, 4.0 mmol, 2.0 equiv), and DMAP (73 mg, 0.60 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 50%) afforded **2n** (424 mg, 1.61 mmol, 81 %) as a colourless oil.

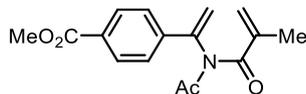
¹H NMR (500 MHz, CDCl₃) δ 7.33 (d, *J* = 8.7 Hz, 2H), 7.27 (d, *J* = 8.8 Hz, 2H), 5.76 (d, *J* = 1.4 Hz, 1H), 5.46 (s, 1H), 5.30 (q, *J* = 1.6 Hz, 1H), 5.18 (d, *J* = 1.4 Hz, 1H), 2.28 (s, 3H), 1.88 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.8, 173.1, 143.6, 141.0, 134.7, 134.5, 128.9, 126.8, 120.4, 115.1, 25.1, 18.9.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1693, 1632, 1491, 1367, 1259, 1172, 1090, 1011, 834.

HRMS (ESI⁺) calcd. for C₁₄H₁₅ClNO₂⁺ [M+H]⁺: 264.0786, found: 264.0783.

Methyl 4-(1-(*N*-acetylmethacrylamido)vinyl)benzoate (2o)



Prepared according to **General Procedure C1** using methyl 4-(1-acetamidovinyl)benzoate **S2o** (657 mg, 3.00 mmol, 1.00 equiv) and CH₂Cl₂ (12.0 mL), then Et₃N (1.25 mL, 9.00 mmol, 3.00 equiv), methacryloyl chloride (0.59 mL, 6.0 mmol, 2.0 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 40%) afforded **2o** (680 mg, 2.37 mmol, 79 %) as a colourless oil.

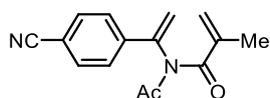
¹H NMR (500 MHz, CDCl₃) δ 7.99 (d, *J* = 8.6 Hz, 2H), 7.46 (d, *J* = 8.6 Hz, 2H), 5.90 (d, *J* = 1.4 Hz, 1H), 5.47 (s, 1H), 5.31 (q, *J* = 1.6 Hz, 1H), 5.29 (d, *J* = 1.4 Hz, 1H), 3.87 (s, 3H), 2.31 (s, 3H), 1.88 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.9, 173.3, 166.4, 143.9, 141.1, 140.3, 130.5, 130.2, 125.6, 120.7, 116.7, 52.2, 25.3, 19.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1713, 1703, 1631, 1609, 1436, 1368, 1276, 1261, 1187, 1107.

HRMS (ESI⁺) calcd. for C₁₆H₁₈NO₄⁺ [M+H]⁺: 288.1230, found: 288.1221.

N-acetyl-*N*-(1-(4-cyanophenyl)vinyl)methacrylamide (**2p**)



Prepared according to **General Procedure C1** using *N*-(1-(4-cyanophenyl)vinyl)acetamide **S2p** (558 mg, 3.00 mmol, 1.00 equiv) and CH₂Cl₂ (12.0 mL), then Et₃N (1.25 mL, 9.00 mmol, 3.00 equiv), methacryloyl chloride (0.59 mL, 6.0 mmol, 2.0 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 50%) afforded **2p** (630 mg, 2.48 mmol, 83 %) as a colourless oil.

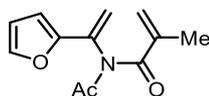
¹H NMR (500 MHz, CDCl₃) δ 7.61 (d, *J* = 8.5 Hz, 2H), 7.49 (d, *J* = 8.6 Hz, 2H), 5.90 (d, *J* = 1.6 Hz, 1H), 5.47 (d, *J* = 1.7 Hz, 1H), 5.34-5.32 (m, 2H), 2.30 (s, 3H), 1.88 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.7, 173.0, 143.2, 140.9, 140.3, 132.6, 126.1, 121.1, 118.3, 117.6, 112.4, 25.3, 19.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2228, 1692, 1631, 1506, 1367, 1315, 1299, 1239, 1188, 1108.

HRMS (ESI⁺) calcd. for C₁₅H₁₄N₂O₂Na⁺ [M+Na]⁺: 277.0947, found: 277.0953.

N-acetyl-*N*-(1-(furan-2-yl)vinyl)methacrylamide (**2q**)



Prepared according to **General Procedure C1** using *N*-(1-(furan-2-yl)vinyl)acetamide **S2q** (350 mg, 2.32 mmol, 1.00 equiv) and CH₂Cl₂ (9.3 mL) then Et₃N (0.97 mL, 7.0 mmol, 3.0 equiv), methacryloyl chloride (0.45 mL, 4.6 mmol, 2.0 equiv), and DMAP (85 mg, 0.70 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 40%) afforded **2q** (270 mg, 1.23 mmol, 53 %) as a pale-yellow oil.

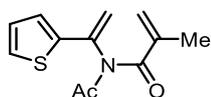
¹H NMR (500 MHz, CDCl₃) δ 7.32 (d, *J* = 1.8 Hz, 1H), 6.33 (dd, *J* = 3.5, 1.9 Hz, 1H), 6.29 (d, *J* = 3.5 Hz, 1H), 5.77 (s, 1H), 5.46 (q, *J* = 0.9 Hz, 1H), 5.27 (s, 1H), 5.06 (s, 1H), 2.29 (s, 3H), 1.90 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.4, 172.8, 150.3, 143.1, 140.8, 135.2, 120.0, 112.7, 111.6, 108.2, 24.7, 19.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1698, 1636, 1454, 1368, 1319, 1275, 1260, 1193, 1014.

HRMS (ESI⁺) calcd. for C₁₂H₁₃NO₃Na⁺ [M+Na]⁺: 242.0788, found: 242.0784.

N-acetyl-*N*-(1-(thiophen-2-yl)vinyl)methacrylamide (**2r**)



Prepared according to **General Procedure C1** using *N*-(1-(thiophen-2-yl)vinyl)acetamide **S2r** (334 mg, 2.00 mmol, 1.00 equiv) and CH₂Cl₂ (8.0 mL), then Et₃N (0.84 mL, 6.0 mmol, 3.0 equiv), methacryloyl chloride (0.39 mL, 4.0 mmol, 2.0 equiv), and DMAP (73 mg, 0.60 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 40%) afforded **2r** (343 mg, 1.46 mmol, 73 %) as a pale-yellow oil.

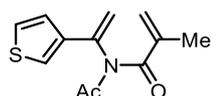
¹H NMR (500 MHz, CDCl₃) δ 7.23 (dd, *J* = 5.0, 1.2 Hz, 1H), 7.04 (dd, *J* = 3.7, 1.2 Hz, 1H), 6.96 (dd, *J* = 5.1, 3.6 Hz, 1H), 5.74 (d, *J* = 1.4 Hz, 1H), 5.52 (s, 1H), 5.32 (q, *J* = 1.7 Hz, 1H), 5.09 (d, *J* = 1.4 Hz, 1H), 2.35 (s, 3H), 1.96 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.6, 173.1, 141.1, 140.7, 139.0, 127.8, 126.3, 125.6, 120.1, 114.0, 25.0, 19.2.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1691, 1623, 1434, 1365, 1276, 1259, 1164, 1037.

HRMS (ESI⁺) calcd. for C₁₂H₁₃NO₂SNa⁺ [M+Na]⁺: 258.0559, found: 258.0553.

N-acetyl-*N*-(1-(thiophen-3-yl)vinyl)methacrylamide (**2s**)



Prepared according to **General Procedure C1** using *N*-(1-(thiophen-3-yl)vinyl)acetamide **S2s** (501 mg, 3.00 mmol, 1.00 equiv) and CH₂Cl₂ (12.0 mL), then Et₃N (1.25 mL, 9.00 mmol, 3.00 equiv), methacryloyl chloride (0.59 mL, 6.0 mmol, 2.0 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 35%) afforded **2s** (479 mg, 2.04 mmol, 68 %) as a pale-yellow solid.

M.P.: 47-49 °C

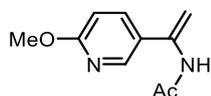
¹H NMR (500 MHz, CDCl₃) δ 7.28 (dd, *J* = 5.1, 3.0 Hz, 1H), 7.20 (dd, *J* = 3.0, 1.4 Hz, 1H), 7.16 (dd, *J* = 5.1, 1.4 Hz, 1H), 5.73 (d, *J* = 1.2 Hz, 1H), 5.47 (s, 1H), 5.30 (q, *J* = 1.6 Hz, 1H), 5.13 (d, *J* = 1.1 Hz, 1H), 2.31 (s, 3H), 1.93 (dd, *J* = 1.7, 1.1 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.9, 173.3, 141.1, 140.2, 138.6, 127.0, 125.3, 122.6, 120.0, 114.2, 25.0, 19.2.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1688, 1630, 1423, 1366, 1276, 1260, 1205, 1185, 1161.

HRMS (ESI⁺) calcd. for C₁₂H₁₃NO₂SNa⁺ [M+Na]⁺: 258.0559, found: 258.0554.

***N*-(1-(6-methoxypyridin-3-yl)vinyl)acetamide (S2t)**



Prepared according to **General Procedure C** (*N*-acetyl enamides synthesis) using (*E*)-1-(6-methoxypyridin-3-yl)ethan-1-one oxime¹⁷ (1.66 g, 10.0 mmol, 1.00 equiv), KI (1.66 g, 10.0 mmol, 1.00 equiv), Na₂S₂O₄ (3.48 g, 20.0 mmol, 2.00 equiv), Fe(OAc)₂ (174 mg, 1.00 mmol, 0.10 equiv), DCE (100 mL), acetic acid (1.72 mL, 30.0 mmol, 3.00 equiv), and acetic anhydride (1.99 mL, 21.0 mmol, 2.10 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 60%) and recrystallisation (pentane/Et₂O) afforded **S2t** (1.10 g, 5.73 mmol, 57 %) as a white solid.

M.P.: 74-76 °C

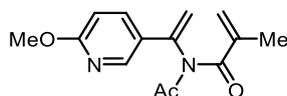
¹H NMR (500 MHz, CDCl₃) δ 8.19 (d, *J* = 2.5 Hz, 1H), 7.57 (dd, *J* = 8.7, 2.6 Hz, 1H), 7.09 (br. s, 1H), 6.70 (d, *J* = 8.5 Hz, 1H), 5.66 (s, 1H), 5.00 (s, 1H), 3.92 (s, 3H), 2.07 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 169.3, 164.4, 144.7, 138.0, 136.7, 127.4, 110.7, 103.3, 53.7, 24.4.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1667, 1633, 1603, 1527, 1489, 1368, 1281, 1254, 1131, 1022.

HRMS (ESI⁺) calcd. for C₁₀H₁₃N₂O₂⁺ [M+H]⁺: 193.0972, found: 193.0964.

***N*-acetyl-*N*-(1-(6-methoxypyridin-3-yl)vinyl)methacrylamide (2t)**



Prepared according to **General Procedure C1** using *N*-(1-(6-methoxypyridin-3-yl)vinyl)acetamide **S2t** (576 mg, 3.00 mmol, 1.00 equiv) and CH₂Cl₂ (12.0 mL), then Et₃N (1.25 mL, 9.00 mmol, 3.00

equiv), methacryloyl chloride (0.59 mL, 6.0 mmol, 2.0 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 40%) afforded **2t** (537 mg, 2.07 mmol, 69 %) as a colourless oil.

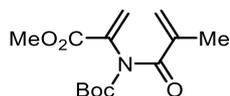
¹H NMR (500 MHz, CDCl₃) δ 8.17 (dd, *J* = 2.7, 0.8 Hz, 1H), 7.58 (dd, *J* = 8.7, 2.6 Hz, 1H), 6.66 (dd, *J* = 8.7, 0.8 Hz, 1H), 5.67 (d, *J* = 1.3 Hz, 1H), 5.43 (s, 1H), 5.27 (q, *J* = 1.7 Hz, 1H), 5.09 (d, *J* = 1.3 Hz, 1H), 3.86 (s, 3H), 2.27 (s, 3H), 1.88 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.8, 173.1, 164.3, 144.3, 142.0, 141.1, 135.9, 125.1, 120.3, 113.9, 111.1, 53.6, 25.1, 19.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1692, 1631, 1600, 1494, 1454, 1367, 1286, 1258, 1173, 1019.

HRMS (ESI⁺) calcd. for C₁₄H₁₇N₂O₃⁺ [M+H]⁺: 261.1234, found: 261.1223.

Methyl 2-(*N*-(*tert*-butoxycarbonyl)methacrylamido)acrylate (**2u**)



Prepared according to **General Procedure C2** using methyl 2-((*tert*-butoxycarbonyl)amino)acrylate¹¹ (402 mg, 2.00 mmol, 1.00 equiv) and THF (10.0 mL), then *n*-BuLi (0.80 mL, 2.5 M in hexane, 2.0 mmol, 1.0 equiv) and methacryloyl chloride (0.25 mL, 2.6 mmol, 1.3 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 20%) afforded **2u** (461 mg, 1.71 mmol, 86 %) as a colourless oil.

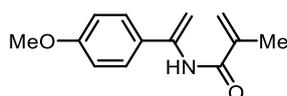
¹H NMR (500 MHz, CDCl₃) δ 6.42 (d, *J* = 0.8 Hz, 1H), 5.69 (d, *J* = 0.8 Hz, 1H), 5.54 (p, *J* = 1.1 Hz, 1H), 5.35 – 5.33 (m, 1H), 3.78 (s, 3H), 2.02 (s, 3H), 1.44 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 173.3, 163.7, 152.0, 142.1, 135.7, 125.6, 119.2, 84.1, 52.7, 27.8, 19.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1735, 1694, 1439, 1370, 1281, 1208, 1155, 1100.

HRMS (ESI⁺) calcd. for C₁₃H₁₉NO₅Na⁺ [M+Na]⁺: 292.1155, found: 292.1146.

N-(1-(4-methoxyphenyl)vinyl)methacrylamide (**S2v**)



Prepared according to **General Procedure C4 (Step C4-1)** using 4-methoxybenzotrile (2.66 g, 20.0 mmol, 1.00 equiv), THF (20.0 mL), MeLi (16.3 mL, 1.60 M in Et₂O, 26.0 mmol, 1.30 equiv), MeOH (1.22 mL, 30.0 mmol, 1.50 equiv), CH₂Cl₂ (ca. 30–40 mL), Et₃N (5.58 mL, 40.0 mmol, 2.00 equiv) and methacryloyl chloride (2.93 mL, 30.0 mmol, 1.50 equiv). Purification by Biotage automated flash

chromatography system (ethyl acetate/pentane, 0% to 30%) afforded **S2v** (1.09 g, 5.02 mmol, 25%) as a white solid.

M.P.: 41-43 °C

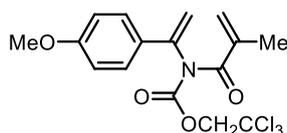
¹H NMR (500 MHz, CDCl₃) δ 7.36 (d, *J* = 8.8 Hz, 2H), 7.13 (br. s, 1H), 6.90 (d, *J* = 8.7 Hz, 2H), 5.86 (s, 1H), 5.74 (s, 1H), 5.44 – 5.41 (m, 1H), 5.05 (d, *J* = 1.0 Hz, 1H), 3.82 (s, 3H), 2.03 (dd, *J* = 1.5, 0.9 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 167.2, 160.1, 141.0, 140.1, 131.2, 127.3, 119.8, 114.3, 101.6, 55.5, 18.9.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1667, 1625, 1608, 1507, 1248, 1177, 1031.

HRMS (ESI⁺) calcd. for C₁₃H₁₆NO₂⁺ [M+H]⁺: 218.1176, found: 218.1187.

2,2,2-Trichloroethyl methacryloyl(1-(4-methoxyphenyl)vinyl)carbamate (**2v**)



Prepared according to **General Procedure C4 (Step C4-2)** using **S2v** (326 mg, 1.50 mmol, 1.00 equiv) and THF (7.5 mL), then *n*-BuLi (0.60 mL, 2.5 M in hexane, 1.5 mmol, 1.0 equiv) and 2,2,2-trichloroethyl chloroformate (0.25 mL, 1.8 mmol, 1.2 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 25%) afforded **2v** (475 mg, 1.21 mmol, 81 %) as a colourless oil.

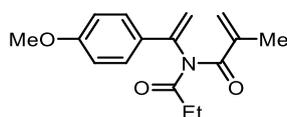
¹H NMR (500 MHz, CDCl₃) δ 7.40 (d, *J* = 8.8 Hz, 2H), 6.86 (d, *J* = 8.9 Hz, 2H), 5.67 (s, 1H), 5.62 (s, 1H), 5.46 (s, 1H), 5.18 (s, 1H), 4.72 (s, 2H), 3.79 (s, 3H), 2.06 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.0, 160.3, 152.7, 143.3, 141.6, 128.8, 127.3, 120.5, 114.1, 112.9, 94.2, 75.7, 55.4, 19.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1746, 1698, 1634, 1608, 1511, 1452, 1248, 1178, 1130, 1087.

HRMS (ESI⁺) calcd. for C₁₆H₁₆Cl₃NO₄Na⁺ [M+Na]⁺: 414.0037, found: 414.0026.

N-(1-(4-methoxyphenyl)vinyl)-*N*-propionylmethacrylamide (**2w**)



Prepared according to **General Procedure C4 (Step C4-2)** using **S2v** (326 mg, 1.50 mmol, 1.00 equiv) and THF (7.5 mL), then *n*-BuLi (0.60 mL, 2.5 M in hexane, 1.5 mmol, 1.0 equiv) and propionic anhydride (0.23 mL, 1.8 mmol, 1.2 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 30%) afforded **2w** (332 mg, 1.22 mmol, 81 %) as a colourless oil.

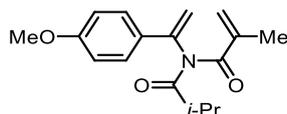
¹H NMR (500 MHz, CDCl₃) δ 7.36 (d, *J* = 8.8 Hz, 2H), 6.87 (d, *J* = 8.9 Hz, 2H), 5.71 (d, *J* = 1.2 Hz, 1H), 5.48 (s, 1H), 5.31 (dq, *J* = 2.2, 1.4 Hz, 1H), 5.10 (d, *J* = 1.1 Hz, 1H), 3.80 (s, 3H), 2.59 (q, *J* = 7.4 Hz, 2H), 1.97 (dd, *J* = 1.6, 1.0 Hz, 3H), 1.11 (t, *J* = 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 177.6, 174.5, 160.3, 144.2, 141.9, 128.8, 127.1, 119.4, 114.3, 112.9, 55.4, 30.6, 19.2, 9.4.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1690, 1627, 1607, 1511, 1454, 1252, 1172, 1029.

HRMS (ESI⁺) calcd. for C₁₆H₁₉NO₃Na⁺ [M+Na]⁺: 296.1257, found: 296.1266.

***N*-isobutyryl-*N*-(1-(4-methoxyphenyl)vinyl)methacrylamide (2x)**



Prepared according to **General Procedure C4 (Step C4-2)** using **S2v** (326 mg, 1.50 mmol, 1.00 equiv) and THF (7.5 mL), then *n*-BuLi (0.60 mL, 2.5 M in hexane, 1.5 mmol, 1.0 equiv) and isobutyric anhydride (0.30 mL, 1.8 mmol, 1.2 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 25%) afforded **2x** (370 mg, 1.29 mmol, 86 %) as a colourless oil.

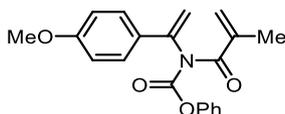
¹H NMR (500 MHz, CDCl₃) δ 7.38 (d, *J* = 8.9 Hz, 2H), 6.88 (d, *J* = 8.9 Hz, 2H), 5.69 (d, *J* = 1.0 Hz, 1H), 5.54 (s, 1H), 5.34 (qd, *J* = 1.6, 0.7 Hz, 1H), 5.09 (d, *J* = 1.0 Hz, 1H), 3.80 (s, 3H), 2.95 (hept, *J* = 6.7 Hz, 1H), 1.99 (dd, *J* = 1.6, 1.0 Hz, 3H), 1.12 (d, *J* = 6.8 Hz, 6H).

¹³C NMR (126 MHz, CDCl₃) δ 181.6, 175.0, 160.4, 144.4, 142.1, 129.1, 127.2, 119.4, 114.3, 112.5, 55.5, 35.2, 19.7, 19.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1691, 1627, 1607, 1511, 1458, 1383, 1301, 1251, 1168.

HRMS (ESI⁺) calcd. for C₁₇H₂₁NO₃Na⁺ [M+Na]⁺: 310.1414, found: 310.1412.

Phenyl methacryloyl(1-(4-methoxyphenyl)vinyl)carbamate (2y)



Prepared according to **General Procedure C4 (Step C4-2)** using **S2v** (326 mg, 1.50 mmol, 1.00 equiv) and THF (7.5 mL), then *n*-BuLi (0.60 mL, 2.5 M in hexane, 1.5 mmol, 1.0 equiv) and phenyl chloroformate (0.22 mL, 1.8 mmol, 1.2 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 25%) afforded **2y** (410 mg, 1.22 mmol, 81 %) as a white solid.

M.P.: 105-107 °C

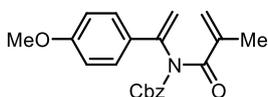
¹H NMR (500 MHz, CDCl₃) δ 7.48 (d, *J* = 8.9 Hz, 2H), 7.34 – 7.28 (m, 2H), 7.22 – 7.17 (m, 1H), 6.92 (d, *J* = 8.9 Hz, 2H), 6.91 – 6.88 (m, 2H), 5.72-5.69 (m, 2H), 5.48 (q, *J* = 1.6 Hz, 1H), 5.25 (d, *J* = 1.3 Hz, 1H), 3.83 (s, 3H), 2.10 (t, *J* = 1.3 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.5, 160.3, 153.0, 150.5, 143.7, 142.1, 129.5, 129.2, 127.2, 126.3, 121.1, 119.9, 114.3, 112.4, 55.5, 19.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1749, 1695, 1633, 1607, 1511, 1493, 1250, 1157, 1028.

HRMS (ESI⁺) calcd. for C₂₀H₁₉NO₄Na⁺ [M+Na]⁺: 360.1206, found: 360.1221.

Benzyl methacryloyl(1-(4-methoxyphenyl)vinyl) carbamate (**2z**)



Prepared according to **General Procedure C4 (Step C4-2)** using **S2v** (326 mg, 1.50 mmol, 1.00 equiv) and THF (7.5 mL), then *n*-BuLi (0.60 mL, 2.5 M in hexane, 1.5 mmol, 1.0 equiv) and benzyl chloroformate (0.26 mL, 1.8 mmol, 1.2 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 25%) afforded **2z** (430 mg, 1.23 mmol, 82 %) as a colourless oil.

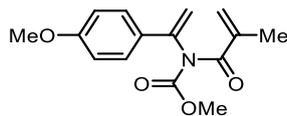
¹H NMR (500 MHz, CDCl₃) δ 7.35 (d, *J* = 8.8 Hz, 2H), 7.30 – 7.24 (m, 3H), 7.13 – 7.10 (m, 2H), 6.84 (d, *J* = 8.8 Hz, 2H), 5.61 (d, *J* = 1.1 Hz, 1H), 5.53 (s, 1H), 5.34 (q, *J* = 1.6 Hz, 1H), 5.11 (s, 2H), 5.10 (d, *J* = 1.1 Hz, 1H), 3.81 (s, 3H), 2.01 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.5, 160.2, 154.3, 143.6, 142.1, 134.8, 129.2, 128.5, 128.5, 128.3, 127.1, 119.4, 114.1, 112.1, 68.8, 55.4, 19.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1736, 1690, 1633, 1607, 1511, 1455, 1380, 1248, 1178, 1124, 1076.

HRMS (ESI⁺) calcd. for C₂₁H₂₁NO₄Na⁺ [M+Na]⁺: 374.1363, found: 374.1381.

Methyl methacryloyl(1-(4-methoxyphenyl)vinyl)carbamate (**2aa**)



Prepared according to **General Procedure C4 (Step C4-2)** using **S2v** (326 mg, 1.50 mmol, 1.00 equiv) and THF (7.5 mL), then *n*-BuLi (0.60 mL, 2.5 M in hexane, 1.5 mmol, 1.0 equiv) and methyl chloroformate (0.14 mL, 1.8 mmol, 1.2 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 30%) afforded **2aa** (350 mg, 1.27 mmol, 85 %) as a colourless oil.

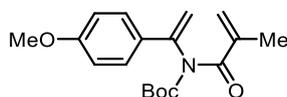
¹H NMR (500 MHz, CDCl₃) δ 7.37 (d, *J* = 8.8 Hz, 2H), 6.86 (d, *J* = 8.9 Hz, 2H), 5.63 (d, *J* = 1.2 Hz, 1H), 5.51 (s, 1H), 5.36 (qd, *J* = 1.6, 0.6 Hz, 1H), 5.11 (d, *J* = 1.1 Hz, 1H), 3.79 (s, 3H), 3.69 (s, 3H), 2.03 (dd, *J* = 1.7, 1.0 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.3, 160.2, 155.0, 143.5, 142.1, 129.0, 127.0, 119.0, 114.1, 112.2, 55.4, 53.9, 19.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1742, 1690, 1633, 1607, 1511, 1438, 1248, 1178, 1129, 1081.

HRMS (ESI⁺) calcd. for C₁₅H₁₇NO₄Na⁺ [M+Na]⁺: 298.1050, found: 298.1063.

Tert-butyl methacryloyl(1-(4-methoxyphenyl)vinyl)carbamate (**2ab**)



Prepared according to **General Procedure C4 (Step C4-2)** using **S2v** (326 mg, 1.50 mmol, 1.00 equiv) and THF (7.5 mL), then *n*-BuLi (0.60 mL, 2.5 M in hexane, 1.5 mmol, 1.0 equiv) and di-*tert*-butyl dicarbonate (392 mg, 1.80 mmol, 1.20 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 30%) afforded **2ab** (396 mg, 1.25 mmol, 83 %) as a colourless oil.

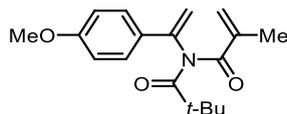
¹H NMR (500 MHz, CDCl₃) δ 7.37 (d, *J* = 8.9 Hz, 2H), 6.86 (d, *J* = 8.9 Hz, 2H), 5.56 (t, *J* = 1.0 Hz, 1H), 5.54 (d, *J* = 0.9 Hz, 1H), 5.39 – 5.34 (m, 1H), 5.07 (d, *J* = 0.9 Hz, 1H), 3.80 (s, 3H), 2.06 (t, *J* = 1.3 Hz, 3H), 1.31 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 173.9, 160.1, 153.0, 144.3, 142.7, 129.9, 127.2, 118.9, 114.0, 111.6, 83.4, 55.4, 27.7, 19.2.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1735, 1691, 1634, 1608, 1512, 1457, 1369, 1276, 1260, 1157, 1031.

HRMS (ESI⁺) calcd. for C₁₈H₂₃NO₄Na⁺ [M+Na]⁺: 340.1519, found: 340.1507.

N-(1-(4-methoxyphenyl)vinyl)-*N*-pivaloylmethacrylamide (**2ac**)



Prepared according to **General Procedure C4 (Step C4-2)** using **S2v** (326 mg, 1.50 mmol, 1.00 equiv) and THF (7.5 mL), then *n*-BuLi (0.60 mL, 2.5 M in hexane, 1.5 mmol, 1.0 equiv) and trimethylacetic anhydride (0.36 mL, 1.8 mmol, 1.2 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 25%) afforded **2ac** (381 mg, 1.27 mmol, 84 %) as a colourless oil.

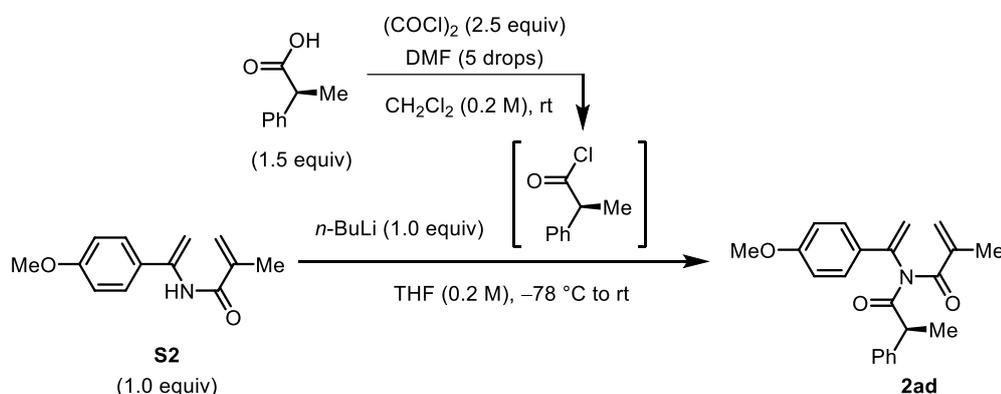
¹H NMR (500 MHz, CDCl₃) δ 7.40 (d, *J* = 8.8 Hz, 2H), 6.87 (d, *J* = 8.9 Hz, 2H), 5.71 (s, 1H), 5.52 (d, *J* = 1.1 Hz, 1H), 5.37 (s, 1H), 4.90 (d, *J* = 1.1 Hz, 1H), 3.80 (s, 3H), 1.88 (dd, *J* = 1.6, 0.9 Hz, 3H), 1.31 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 187.4, 175.2, 160.2, 145.0, 141.4, 129.0, 127.5, 120.8, 114.2, 110.7, 55.4, 43.5, 29.0, 19.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1689, 1608, 1511, 1459, 1276, 1258, 1177, 1144, 1031.

HRMS (ESI⁺) calcd. for C₁₈H₂₄NO₃⁺ [M+H]⁺: 302.1751, found: 302.1750.

(*S*)-*N*-(1-(4-methoxyphenyl)vinyl)-*N*-(2-phenylpropanoyl)methacrylamide (**2ad**)



An oven-dried 100 mL round-bottom flask containing (*S*)-(+)-2-phenylpropionic acid (266 mg, 1.77 mmol, 1.50 equiv) was sealed and subjected to three N₂ evacuation/refill cycles before pre-sparged anhydrous CH₂Cl₂ (9.0 mL) was added. Oxalyl chloride (0.25 mL, 3.0 mmol, 2.5 equiv) was then added at 0 °C, followed by the addition of DMF (5 drops). The reaction mixture was allowed to warm up to room temperature and stirred for 1 h. The reaction was then concentrated under reduced pressure to afford the crude acyl chloride and used directly for the next step without further purification.

An oven-dried 50 mL round-bottom flask charged with **S2v** (255 mg, 1.18 mmol, 1.00 equiv) was sealed and subjected to three cycles of evacuation and nitrogen backfilling. Anhydrous THF (6.0 mL)

was then added, and the reaction mixture was cooled to $-78\text{ }^{\circ}\text{C}$. *n*-BuLi (2.5 M in hexane, 0.47 mL, 1.2 mmol, 1.0 equiv) was added dropwise at $-78\text{ }^{\circ}\text{C}$, and the mixture was stirred at this temperature for 10 minutes. Crude acyl chloride (dissolved in 2.0 mL anhydrous THF) was subsequently added at $-78\text{ }^{\circ}\text{C}$, and the reaction was allowed to warm to room temperature over 1 hour. The reaction was then quenched with brine (50 mL) and extracted with ethyl acetate ($3 \times 40\text{ mL}$). The combined organic layers were dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude product was purified by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 25%) to afford **2ad** (165 mg, 0.47 mmol, 40 %) as a colourless oil.

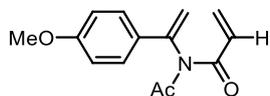
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.33 – 7.23 (m, 7H), 6.81 (d, $J = 8.8\text{ Hz}$, 2H), 5.63 (d, $J = 1.1\text{ Hz}$, 1H), 5.18 (d, $J = 1.7\text{ Hz}$, 1H), 5.15 (s, 1H), 4.89 (d, $J = 1.1\text{ Hz}$, 1H), 4.24 (q, $J = 6.9\text{ Hz}$, 1H), 3.79 (s, 3H), 1.88 (s, 3H), 1.43 (d, $J = 7.0\text{ Hz}$, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 178.1, 174.9, 160.3, 144.2, 141.4, 140.3, 128.8, 128.7, 128.1, 127.5, 127.2, 119.7, 114.2, 112.6, 55.4, 46.4, 19.1, 18.9.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1696, 1626, 1607, 1512, 1454, 1304, 1275, 1172, 1030.

HRMS (ESI^+) calcd. for $\text{C}_{22}\text{H}_{23}\text{NO}_3\text{Na}^+$ [$\text{M}+\text{Na}$] $^+$: 372.1570, found: 372.1568.

N-acetyl-*N*-(1-(4-methoxyphenyl)vinyl)acrylamide (**2ae**)



Prepared according to **General Procedure C1** using *N*-(1-(4-methoxyphenyl)vinyl)acetamide **S2g** (573 mg, 3.00 mmol, 1.00 equiv) and CH_2Cl_2 (12.0 mL), then Et_3N (1.25 mL, 9.00 mmol, 3.00 equiv) and acryloyl chloride (0.49 mL, 6.0 mmol, 2.0 equiv).^A Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 40%) afforded **2ae** (285 mg, 1.16 mmol, 39 %) as a pale-yellow oil.

Notes: (A) When acryloyl chloride is used as the electrophile, the addition of DMAP is not required to promote the reaction. Acryloyl chloride is a highly reactive reagent and is usually stored in a glovebox freezer at $-30\text{ }^{\circ}\text{C}$. The reaction was complete within 1 h.

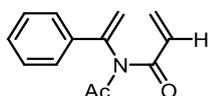
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.33 (d, $J = 8.9\text{ Hz}$, 2H), 6.85 (d, $J = 8.9\text{ Hz}$, 2H), 6.62 (dd, $J = 16.8, 10.2\text{ Hz}$, 1H), 6.44 (dd, $J = 16.8, 1.7\text{ Hz}$, 1H), 5.83 (d, $J = 1.1\text{ Hz}$, 1H), 5.70 (dd, $J = 10.2, 1.7\text{ Hz}$, 1H), 5.14 (d, $J = 1.2\text{ Hz}$, 1H), 3.75 (s, 3H), 2.46 (s, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 173.0, 167.5, 160.3, 143.3, 130.5, 130.1, 127.9, 126.4, 114.3, 114.1, 55.2, 26.2.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1693, 1606, 1511, 1461, 1400, 1367, 1303, 1256, 1218, 1121, 1028.

HRMS (ESI⁺) calcd. for C₁₄H₁₅NO₃Na⁺ [M+Na]⁺: 268.0944, found: 268.0934.

***N*-acetyl-*N*-(1-phenylvinyl)acrylamide (**2af**)**



Prepared according to **General Procedure C1** using *N*-(1-phenylvinyl)acetamide **S2f** (483 mg, 3.00 mmol, 1.00 equiv) and CH₂Cl₂ (12.0 mL), then Et₃N (1.25 mL, 9.00 mmol, 3.00 equiv) and acryloyl chloride (0.49 mL, 6.0 mmol, 2.0 equiv).^A Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 30%) afforded **2af** (230 mg, 1.07 mmol, 36 %) as a colourless oil.

Notes: (A) When acryloyl chloride is used as the electrophile, the addition of DMAP is not required to promote the reaction. Acryloyl chloride is a highly reactive reagent and is usually stored in a glovebox freezer at -30 °C. The reaction was complete within 1 h.

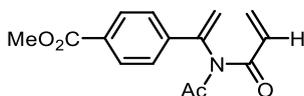
¹H NMR (500 MHz, CDCl₃) δ 7.44 – 7.41 (m, 2H), 7.39 – 7.32 (m, 3H), 6.65 (dd, *J* = 16.7, 10.2 Hz, 1H), 6.49 (dd, *J* = 16.8, 1.7 Hz, 1H), 5.99 (d, *J* = 1.1 Hz, 1H), 5.74 (dd, *J* = 10.2, 1.7 Hz, 1H), 5.29 (d, *J* = 1.1 Hz, 1H), 2.50 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.2, 167.7, 143.9, 135.5, 130.9, 130.2, 129.3, 129.1, 125.1, 116.3, 26.4.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1697, 1634, 1518, 1401, 1367, 1306, 1221, 1191, 1050.

HRMS (ESI⁺) calcd. for C₁₃H₁₃NO₂Na⁺ [M+Na]⁺: 238.0838, found: 238.0837.

Methyl 4-(1-(*N*-acetylacrylamido)vinyl)benzoate (2ag**)**



Prepared according to **General Procedure C1** using methyl 4-(1-acetamidovinyl)benzoate **S2o** (420 mg, 1.92 mmol, 1.00 equiv), CH₂Cl₂ (7.7 mL), Et₃N (0.80 mL, 5.8 mmol, 3.0 equiv) and acryloyl chloride (0.31 mL, 3.8 mmol, 2.0 equiv).^A The reaction mixture was stirred at room temperature for 1 h^B, after which a second portion of Et₃N (0.80 mL, 5.8 mmol, 3.0 equiv) and acryloyl chloride (0.31 mL, 3.8 mmol, 2.0 equiv) were added. The mixture was stirred at room temperature for an additional 1 h. Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 30%) afforded **2ag** (210 mg, 0.77 mmol, 40 %) as a colourless oil.

Notes: (A) When acryloyl chloride is used as the electrophile, the addition of DMAP is not required to promote the reaction. Acryloyl chloride is a highly reactive reagent and is usually stored in a glovebox freezer at $-30\text{ }^{\circ}\text{C}$.

(B) After 1 h, TLC analysis indicated the presence of unreacted starting material.

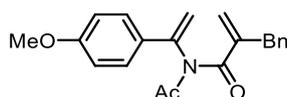
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.03 (d, $J = 8.5$ Hz, 2H), 7.48 (d, $J = 8.5$ Hz, 2H), 6.61 (dd, $J = 16.7$, 10.1 Hz, 1H), 6.50 (dd, $J = 16.8$, 1.8 Hz, 1H), 6.08 (d, $J = 1.3$ Hz, 1H), 5.77 (dd, $J = 10.1$, 1.7 Hz, 1H), 5.40 (d, $J = 1.2$ Hz, 1H), 3.91 (s, 3H), 2.50 (s, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 173.1, 167.5, 166.5, 143.3, 139.9, 131.4, 130.8, 130.4, 129.9, 125.2, 118.4, 52.4, 26.4.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1700, 1632, 1609, 1435, 1401, 1368, 1275, 1217, 1185, 1107.

HRMS (ESI⁺) calcd. for $\text{C}_{15}\text{H}_{15}\text{NO}_4\text{Na}^+$ [$\text{M}+\text{Na}$]⁺: 296.0893, found: 296.0895.

N-acetyl-2-benzyl-*N*-(1-(4-methoxyphenyl)vinyl)acrylamide (**2ah**)



Prepared according to **General Procedure C3** using 2-benzylacrylic acid (972 mg, 6.00 mmol, 2.00 equiv), CH_2Cl_2 (20.0 mL), oxalyl chloride (0.76 mL, 9.0 mmol, 3.0 equiv) and DMF (5 drops). Then using CH_2Cl_2 (12.0 mL), Et_3N (1.25 mL, 9.00 mmol, 3.00 equiv), *N*-(1-(4-methoxyphenyl)vinyl)acetamide **S2g** (573 mg, 3.00 mmol, 1.00 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by two successive flash chromatography separations using a Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 35% and ethyl acetate/ CH_2Cl_2 , 0% to 10%) afforded **2ah** (719 mg, 2.15 mmol, 72 %) as a pale-yellow oil.

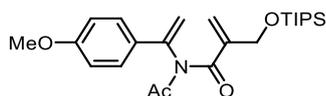
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.32 – 7.27 (m, 4H), 7.25 – 7.21 (m, 1H), 7.18 – 7.15 (m, 2H), 6.85 (d, $J = 8.9$ Hz, 2H), 5.63 – 5.60 (m, 2H), 5.17 (t, $J = 1.7$ Hz, 1H), 4.77 (d, $J = 1.1$ Hz, 1H), 3.80 (s, 3H), 3.62 (s, 2H), 2.29 (s, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 173.7, 173.6, 160.3, 145.3, 144.1, 137.6, 129.5, 128.6, 127.1, 126.7, 120.8, 114.3, 113.1, 55.4, 39.0, 25.3.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1688, 1627, 1605, 1511, 1366, 1252, 1178, 1029.

HRMS (ESI⁺) calcd. for $\text{C}_{21}\text{H}_{21}\text{NO}_3\text{Na}^+$ [$\text{M}+\text{Na}$]⁺: 358.1414, found: 358.1409.

N-acetyl-*N*-(1-(4-methoxyphenyl)vinyl)-2-(((triisopropylsilyloxy)methyl)acrylamide (**2ai**)



Prepared according to **General Procedure C3** using 2-(((triisopropylsilyloxy)methyl)acrylic acid¹⁸ (3.10 g, 12.0 mmol, 2.00 equiv), CH₂Cl₂ (40.0 mL), oxalyl chloride (1.52 mL, 18.0 mmol, 3.00 equiv) and DMF (8 drops). Then using CH₂Cl₂ (24.0 mL), Et₃N (2.51 mL, 18.0 mmol, 3.00 equiv), *N*-(1-(4-methoxyphenyl)vinyl)acetamide **S2g** (1.15 g, 6.00 mmol, 1.00 equiv), and DMAP (220 mg, 1.80 mmol, 0.30 equiv). Purification by two successive flash chromatography separations using a Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 35% and ethyl acetate/CH₂Cl₂, 0% to 10%) afforded **2ai** (1.73 g, 4.01 mmol, 67 %) as a pale-yellow oil.

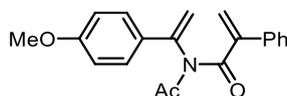
¹H NMR (500 MHz, CDCl₃) δ 7.36 (d, *J* = 8.9 Hz, 2H), 6.86 (d, *J* = 8.9 Hz, 2H), 5.70 (d, *J* = 1.1 Hz, 1H), 5.67 (td, *J* = 2.1, 0.8 Hz, 1H), 5.65 (td, *J* = 1.8 Hz, 0.9, 1H), 5.11 (d, *J* = 1.0 Hz, 1H), 4.40 (t, *J* = 1.9 Hz, 2H), 3.80 (s, 3H), 2.36 (s, 3H), 1.13 – 1.02 (m, 21H).

¹³C NMR (126 MHz, CDCl₃) δ 173.7, 172.7, 160.3, 145.0, 144.3, 128.7, 127.1, 118.6, 114.3, 113.0, 62.8, 55.5, 25.6, 18.1, 12.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2942, 2865, 1691, 1629, 1607, 1512, 1461, 1366, 1303, 1250, 1119.

HRMS (ESI⁺) calcd. for C₂₄H₃₈NO₄Si⁺ [M+H]⁺: 432.2565, found: 432.2581.

N-acetyl-*N*-(1-(4-methoxyphenyl)vinyl)-2-phenylacrylamide (**2aj**)



Prepared according to **General Procedure C3** using 2-phenylacrylic acid (888 mg, 6.00 mmol, 2.00 equiv), CH₂Cl₂ (20.0 mL), oxalyl chloride (0.76 mL, 9.0 mmol, 3.0 equiv), and DMF (5 drops). Then using CH₂Cl₂ (12.0 mL), Et₃N (1.25 mL, 9.00 mmol, 3.00 equiv), *N*-(1-(4-methoxyphenyl)vinyl)acetamide **S2g** (573 mg, 3.00 mmol, 1.00 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by two successive flash chromatography separations using a Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 30% and ethyl acetate/CH₂Cl₂, 0% to 8%) afforded **2aj** (800 mg, 2.49 mmol, 83 %) as a colourless oil.

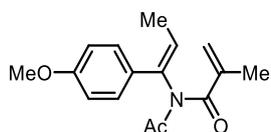
¹H NMR (500 MHz, CDCl₃) δ 7.38 – 7.30 (m, 5H), 7.29 (d, *J* = 8.9 Hz, 2H), 6.85 (d, *J* = 8.9 Hz, 2H), 5.70 (d, *J* = 1.2 Hz, 1H), 5.57 (s, 1H), 5.49 (s, 1H), 5.01 (d, *J* = 1.2 Hz, 1H), 3.80 (s, 3H), 2.36 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.0, 172.5, 160.3, 146.2, 143.5, 136.4, 128.6, 128.5, 128.5, 126.9, 126.7, 118.2, 114.2, 55.4, 25.7.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1692, 1630, 1606, 1511, 1366, 1256, 1179, 1029.

HRMS (ESI⁺) calcd. for C₂₀H₁₉NO₃Na⁺ [M+Na]⁺: 344.1257, found: 344.1254.

(E)-N-acetyl-N-(1-(4-methoxyphenyl)prop-1-en-1-yl)methacrylamide (2ak)



Prepared according to **General Procedure C1** using *N*-(1-phenylprop-1-en-1-yl)acetamide **S2ak** (*Z/E* mixture, 615 mg, 3.00 mmol, 1.00 equiv) and CH₂Cl₂ (12.0 mL), then Et₃N (1.25 mL, 9.00 mmol, 3.00 equiv), methacryloyl chloride (0.59 mL, 6.0 mmol, 2.0 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 30%) afforded **2ak** (680 mg, 2.49 mmol, 83 %) as a colourless oil.

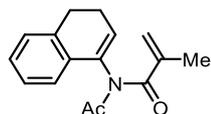
¹H NMR (500 MHz, CDCl₃) δ 7.22 (d, *J* = 8.9 Hz, 2H), 6.82 (d, *J* = 8.9 Hz, 2H), 6.12 (q, *J* = 7.0 Hz, 1H), 5.35 (s, 1H), 5.24 (q, *J* = 1.6 Hz, 1H), 3.75 (s, 3H), 2.28 (s, 3H), 1.84 (dd, *J* = 1.7, 1.0 Hz, 3H), 1.72 (d, *J* = 7.0 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.9, 173.5, 159.5, 141.5, 137.4, 130.1, 126.6, 123.1, 118.9, 114.2, 55.3, 25.1, 19.1, 13.7.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1709, 1691, 1608, 1511, 1443, 1365, 1275, 1259, 1176, 1031.

HRMS (ESI⁺) calcd. for C₁₆H₁₉NO₃Na⁺ [M+Na]⁺: 296.1257, found: 296.1246.

***N*-acetyl-*N*-(3,4-dihydronaphthalen-1-yl)methacrylamide (2al)**



Prepared according to **General Procedure C1** using *N*-(3,4-dihydronaphthalen-1-yl)acetamide **S2al** (561 mg, 3.00 mmol, 1.00 equiv) and CH₂Cl₂ (12.0 mL), then Et₃N (1.25 mL, 9.00 mmol, 3.00 equiv), methacryloyl chloride (0.59 mL, 6.0 mmol, 2.0 equiv), and DMAP (110 mg, 0.90 mmol, 0.30 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 35%) afforded **2al** (630 mg, 2.47 mmol, 82 %) as a colourless oil.

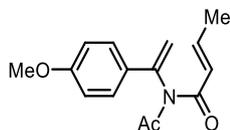
¹H NMR (500 MHz, CDCl₃) δ 7.21 – 7.14 (m, 3H), 7.04 – 6.99 (m, 1H), 5.92 (t, *J* = 4.7 Hz, 1H), 5.47 – 5.43 (m, 1H), 5.26 – 5.24 (m, 1H), 2.92 – 2.80 (m, 2H), 2.50 – 2.40 (m, 2H), 2.34 (s, 3H), 1.96 (dd, *J* = 1.7, 1.0 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 174.0, 173.4, 141.4, 136.4, 136.3, 131.6, 128.7, 128.1, 128.0, 126.8, 121.9, 119.2, 27.1, 25.0, 23.0, 19.4.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1688, 1635, 1451, 1427, 1365, 1316, 1275, 1261, 1195.

HRMS (ESI⁺) calcd. for C₁₆H₁₇NO₂Na⁺ [M+Na]⁺: 278.1152, found: 278.1153.

(*E*)-*N*-acetyl-*N*-(1-(4-methoxyphenyl)vinyl)but-2-enamide (**2am**)



Prepared according to **General Procedure C2** using *N*-(1-(4-methoxyphenyl)vinyl)acetamide **S2g** (252 mg, 1.32 mmol, 1.00 equiv) and THF (6.6 mL), then *n*-BuLi (0.53 mL, 2.5 M in hexane, 1.3 mmol, 1.0 equiv) and crotonoyl chloride (0.17 mL, 1.7 mmol, 1.3 equiv). Purification by Biotage automated flash chromatography system (ethyl acetate/pentane, 0% to 30%) afforded **2am** (140 mg, 0.54 mmol, 41 %) as a colourless oil.

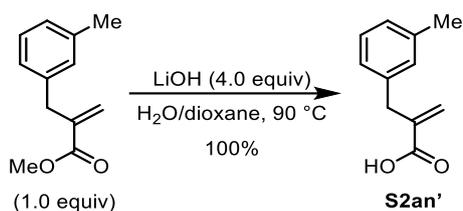
¹H NMR (500 MHz, CDCl₃) δ 7.35 (d, *J* = 8.9 Hz, 2H), 7.07 (dq, *J* = 15.1, 7.0 Hz, 1H), 6.87 (d, *J* = 8.9 Hz, 2H), 6.34 (dq, *J* = 15.1, 1.7 Hz, 1H), 5.83 (d, *J* = 1.0 Hz, 1H), 5.14 (d, *J* = 1.0 Hz, 1H), 3.78 (s, 3H), 2.46 (s, 3H), 1.84 (dd, *J* = 7.0, 1.7 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.2, 167.8, 160.3, 145.6, 143.6, 128.3, 126.5, 124.5, 114.3, 114.0, 55.4, 26.3, 18.3.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1693, 1636, 1606, 1513, 1462, 1367, 1275, 1260, 1214, 1181.

HRMS (ESI⁺) calcd. for C₁₅H₁₇NO₃Na⁺ [M+Na]⁺: 282.1101, found: 282.1089.

2-(3-Methylbenzyl)acrylic acid (**S2an'**)



In a 250 mL round-bottom flask containing a solution of methyl 2-(3-methylbenzyl)acrylate¹⁹ (11.3 g, 59.2 mmol, 1.00 equiv) in 1,4-dioxane (42.0 mL) and H₂O (120 mL) was added LiOH·H₂O (9.95 g, 237 mmol, 4.00 equiv) at room temperature. The mixture was then stirred at 90 °C for 2 h. After that, the reaction was cooled to room temperature and extracted with Et₂O (50 mL). The aqueous phase was acidified with 2 N HCl until the solution reached pH = 1 and extracted with ethyl acetate (3 × 100 mL). The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure, affording **S2an'** (10.4 g, 59.1 mmol, 100%) as a colourless oil.

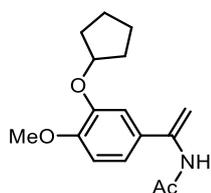
¹H NMR (500 MHz, CDCl₃) δ 11.96 (br. s, 1H), 7.25 (t, *J* = 7.4 Hz, 1H), 7.12-7.04 (m, 3H), 6.45 (d, *J* = 1.4 Hz, 1H), 5.64 (q, *J* = 1.5 Hz, 1H), 3.66 (s, 2H), 2.39 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 172.9, 139.7, 138.5, 138.2, 129.9, 128.8, 128.5, 127.3, 126.2, 37.5, 21.5.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1691, 1627, 1608, 1433, 1294, 1219, 1148.

HRMS (Nanospray) calcd. for C₁₁H₁₁O₂ [M-H]⁻: 175.0765, found: 175.0753.

N-(1-(3-(cyclopentyloxy)-4-methoxyphenyl)vinyl)acetamide (**S2an**)



Prepared according to **General Procedure C** using (*E*)-1-(3-(cyclopentyloxy)-4-methoxyphenyl)ethan-1-one oxime²⁰ (12.5 g, 50.0 mmol, 1.00 equiv), KI (8.30 g, 50.0 mmol, 1.00 equiv), Na₂S₂O₄ (17.4 g, 100 mmol, 2.00 equiv), Fe(OAc)₂ (870 mg, 5.00 mmol, 0.10 equiv), 1,2-dichloroethane (500 mL), acetic acid (8.65 mL, 150 mmol, 3.00 equiv), and acetic anhydride (9.92 mL, 105 mmol, 2.10 equiv). Purification using Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 60%) afforded **S2an** (6.45 g, 23.5 mmol, 47 %) as a pale-yellow oil.

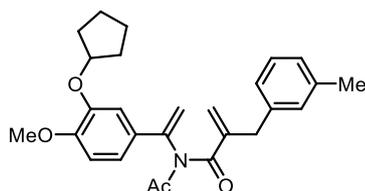
¹H NMR (500 MHz, CDCl₃) δ 7.26 (s, 1H), 6.91-6.86 (m, 2H), 6.78-6.73 (m, 1H), 5.70 (s, 1H), 4.94 (s, 1H), 4.74 (hept, *J* = 6.3, 3.2 Hz, 1H), 3.79 (s, 3H), 2.04 (s, 3H), 1.92 – 1.76 (m, 6H), 1.60 – 1.55 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 169.3, 150.5, 147.5, 140.5, 131.3, 118.4, 113.4, 111.6, 101.3, 80.6, 56.0, 32.8, 24.4, 24.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1670, 1509, 1258, 1172, 1142.

HRMS (ESI⁺) calcd. for C₁₆H₂₂NO₃⁺ [M+H]⁺: 276.1594, found: 276.1587.

N-acetyl-*N*-(1-(3-(cyclopentyloxy)-4-methoxyphenyl)vinyl)-2-(3-methylbenzyl) acrylamide (**2an**)



Prepared according to **General Procedure C3** using 2-(3-methylbenzyl)acrylic acid **S2an'** (2.07 g, 11.8 mmol, 2.00 equiv), CH₂Cl₂ (440.0 mL), oxalyl chloride (1.50 mL, 17.7 mmol, 3.00 equiv), and DMF (8 drops). Then using CH₂Cl₂ (23.6 mL), Et₃N (2.47 mL, 17.7 mmol, 3.00 equiv), *N*-(1-(3-(cyclopentyloxy)-4-methoxyphenyl)vinyl)acetamide **S2an** (1.62 g, 5.90 mmol, 1.00 equiv) in CH₂Cl₂ (6.0 mL), and DMAP (216 mg, 1.77 mmol, 0.30 equiv). Purification by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%) and ethyl acetate/CH₂Cl₂ (0% to 10%) afforded **2an** (1.76 g, 4.06 mmol, 69 %) as a colourless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.17 (t, *J* = 7.5 Hz, 1H), 7.06-7.02 (m, 1H), 6.98-6.93 (m, 2H), 6.92 – 6.89 (m, 2H), 6.82-6.78 (m, 1H), 5.63 (s, 1H), 5.60 (d, *J* = 1.1 Hz, 1H), 5.18 (t, *J* = 1.7 Hz, 1H), 4.80 (d, *J* = 1.2 Hz, 1H), 4.73 (hept, *J* = 3.0 Hz, 1H), 3.83 (s, 3H), 3.57 (s, 2H), 2.32 (s, 3H), 2.30 (s, 3H), 1.92 – 1.80 (m, 6H), 1.64 – 1.57 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 173.70, 173.66, 150.9, 147.8, 145.2, 144.2, 138.1, 137.5, 130.2, 128.9, 128.4, 127.3, 126.5, 120.9, 118.5, 113.0, 112.4, 111.8, 80.6, 56.0, 38.7, 32.8, 25.2, 24.1, 21.4.

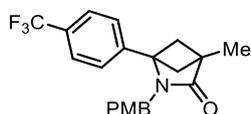
IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2958, 1689, 1628, 1602, 1512, 1421, 1365, 1256, 1222, 1140.

HRMS (ESI⁺) calcd. for C₂₇H₃₂NO₄⁺ [M+H]⁺: 434.2326, found: 434.2314.

2.4 Synthesis of Cyclisation Products

2.4.1 Synthesis and Characterisation of Products (3 and 4)

2-(4-Methoxybenzyl)-4-methyl-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.1.1]hexan-3-one (3a)



Prepared according to **General Procedure D** using **1a** (780 mg, 2.08 mmol, 1.00 equiv), CH₃CN (20.8 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (23.3 mg, 20.8 μmol, 1.0 mol%). The mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **3a** (513 mg, 1.37 mmol, 66%) as a white solid and **4a** (174 mg, 0.46 mmol, 22%) as a colourless oil.^A

Notes: (A) The reaction time was limited to 0.5 h. Compound **4a** is not stable under the optimised conditions and gradually decomposes as the reaction progresses. After 30 minutes, the ¹H NMR spectrum showed a 2.7:1 molar ratio of **3a/4a**, and the starting material **1a** was completely consumed (as confirmed by TLC).

M.P.: 109-111 °C

¹H NMR (500 MHz, CDCl₃) δ 7.48 (d, *J* = 7.9 Hz, 2H), 7.09 (d, *J* = 7.9 Hz, 2H), 6.60 (d, *J* = 8.6 Hz, 2H), 6.57 (d, *J* = 8.6 Hz, 2H), 4.11 (s, 2H), 3.66 (s, 3H), 2.65 (dd, *J* = 4.0, 1.9 Hz, 2H), 2.37-2.31 (m, 2H), 1.31 (s, 3H).

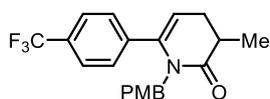
¹³C NMR (126 MHz, CDCl₃) δ 182.2, 158.6, 139.3, 130.4 (q, *J* = 32.4 Hz), 130.1, 129.3, 128.0, 125.1 (q, *J* = 3.8 Hz), 123.9 (q, *J* = 272.3 Hz), 113.4, 66.4, 57.9, 55.0, 48.2, 43.8, 13.3.

¹⁹F NMR (471 MHz, CDCl₃) δ -62.62.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1715, 1612, 1512, 1324, 1266, 1241, 1167, 1125, 1067.

HRMS (ESI⁺) calcd. for C₂₁H₂₁F₃NO₂⁺ [M+H]⁺: 376.1519, found: 376.1520.

1-(4-Methoxybenzyl)-3-methyl-6-(4-(trifluoromethyl)phenyl)-3,4-dihydropyridin-2(1H)-one (4a)



¹H NMR (500 MHz, CDCl₃) δ 7.55 (d, *J* = 8.0 Hz, 2H), 7.23 (d, *J* = 7.9 Hz, 2H), 6.77 (d, *J* = 8.7 Hz, 2H), 6.69 (d, *J* = 8.8 Hz, 2H), 5.37 (dd, *J* = 6.1, 3.9 Hz, 1H), 4.97 (d, *J* = 15.1 Hz, 1H), 4.34 (d, *J* =

15.3 Hz, 1H), 3.73 (s, 3H), 2.72 – 2.61 (m, 1H), 2.40 (dt, $J = 16.4, 6.0$ Hz, 1H), 2.13 (ddd, $J = 16.2, 11.7, 4.0$ Hz, 1H), 1.29 (d, $J = 7.0$ Hz, 3H).

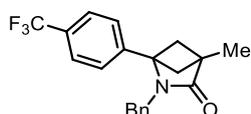
^{13}C NMR (126 MHz, CDCl_3) δ 174.5, 158.7, 141.3, 139.8, 130.2 (q, $J = 32.6$ Hz), 129.9, 128.7, 128.1, 125.3 (q, $J = 3.8$ Hz), 124.1 (q, $J = 272.1$ Hz), 113.7, 111.7, 55.2, 45.7, 35.7, 27.9, 15.5.

^{19}F NMR (471 MHz, CDCl_3) δ -62.58.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1671, 1614, 1512, 1457, 1384, 1323, 1246, 1164, 1066.

HRMS (ESI⁺) calcd. for $\text{C}_{21}\text{H}_{21}\text{F}_3\text{NO}_2^+$ $[\text{M}+\text{H}]^+$: 376.1519, found: 376.1521.

2-Benzyl-4-methyl-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.1.1]hexan-3-one (3b)



Prepared according to **General Procedure D** using **1b** (390 mg, 1.13 mmol, 1.00 equiv), CH_3CN (11.3 mL), and $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbbpy})](\text{PF}_6)$ (12.7 mg, 11.3 μmol , 1.0 mol%). The mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **3b** (239 mg, 0.69 mmol, 61%) as a white solid and **4b** (81 mg, 0.23 mmol, 21%) as a colourless oil.^A

Notes: (A) The reaction time was limited to 0.5 h. Compound **4b** is not stable under the optimised conditions and gradually decomposes as the reaction progresses. After 30 minutes, the ^1H NMR spectrum showed a 2.5:1 molar ratio of **3b/4b**, and the starting material **1b** was completely consumed (as confirmed by TLC).

M.P.: 60-62 °C

^1H NMR (500 MHz, CDCl_3) δ 7.48 (d, $J = 8.1$ Hz, 2H), 7.13 – 7.05 (m, 5H), 6.74 (d, $J = 6.4$ Hz, 2H), 4.20 (s, 2H), 2.73 (dd, $J = 4.0, 2.0$ Hz, 2H), 2.40 (dd, $J = 3.8, 2.1$ Hz, 2H), 1.37 (s, 3H).

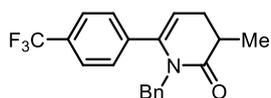
^{13}C NMR (126 MHz, CDCl_3) δ 182.3, 139.1, 138.2, 130.6 (q, $J = 32.4$ Hz), 128.14, 128.07, 128.05, 127.1, 125.2 (q, $J = 3.7$ Hz), 124.0 (q, $J = 272.1$ Hz), 66.6, 58.1, 48.4, 44.6, 13.5.

^{19}F NMR (471 MHz, CDCl_3) δ -62.65.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1722, 1623, 1496, 1411, 1378, 1323, 1276, 1164, 1121.

HRMS (ESI⁺) calcd. for $\text{C}_{20}\text{H}_{19}\text{F}_3\text{NO}^+$ $[\text{M}+\text{H}]^+$: 346.1413, found: 346.1401.

1-Benzyl-3-methyl-6-(4-(trifluoromethyl)phenyl)-3,4-dihydropyridin-2(1H)-one (4b)



¹H NMR (500 MHz, CDCl₃) δ 7.54 (d, *J* = 8.1 Hz, 2H), 7.22 (d, *J* = 8.0 Hz, 2H), 7.19 – 7.15 (m, 3H), 6.88 – 6.84 (m, 2H), 5.39 (dd, *J* = 6.0, 3.9 Hz, 1H), 5.03 (d, *J* = 15.3 Hz, 1H), 4.42 (d, *J* = 15.4 Hz, 1H), 2.74-2.66 (m, 1H), 2.44 (dt, *J* = 16.5, 6.1 Hz, 1H), 2.18 (ddd, *J* = 16.2, 11.8, 4.0 Hz, 1H), 1.31 (d, *J* = 7.0 Hz, 3H).

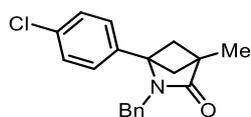
¹³C NMR (126 MHz, CDCl₃) δ 174.5, 141.4, 139.7 (q, *J* = 1.6 Hz), 137.8, 130.3 (q, *J* = 32.6 Hz), 128.4, 128.2, 127.3, 127.1, 125.3 (q, *J* = 3.7 Hz), 124.1 (d, *J* = 272.1 Hz), 111.6, 46.4, 35.8, 27.9, 15.6.

¹⁹F NMR (471 MHz, CDCl₃) δ -62.61.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1670, 1486, 1455, 1380, 1220, 1087, 1015.

HRMS (ESI⁺) calcd. for C₂₀H₁₉F₃NO⁺ [M+H]⁺: 346.1413, found: 346.1398.

2-Benzyl-1-(4-chlorophenyl)-4-methyl-2-azabicyclo[2.1.1]hexan-3-one (**3c**)



Prepared according to **General Procedure D** using **1c** (385 mg, 1.24 mmol, 1.00 equiv), CH₃CN (12.4 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (13.9 mg, 12.4 μmol, 1.0 mol%). The mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 40%), affording **3c** (201 mg, 0.64 mmol, 52%) as a white solid and **4c** (54 mg, 0.17 mmol, 14%) as a colourless oil.^{A,B}

Notes: (**A**) Due to the instability of **4c** under photoirradiation, a low yield (14%) was obtained. Notably, a higher yield of **4c** was observed when the reaction time was shortened to 0.5 h (see **Supplementary Table S1**).

(**B**) When HFIP was used as the solvent, **4c** was isolated in 72% yield while **3c** was detected in trace amounts in the crude NMR spectrum.

M.P.: 105-107 °C

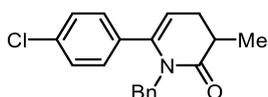
¹H NMR (500 MHz, CDCl₃) δ 7.23 (d, *J* = 8.4 Hz, 2H), 7.15 – 7.10 (m, 3H), 6.93 (d, *J* = 8.4 Hz, 2H), 6.83 – 6.79 (m, 2H), 4.20 (s, 2H), 2.69 (dd, *J* = 4.0, 1.8 Hz, 2H), 2.37 (dd, *J* = 4.0, 1.9 Hz, 2H), 1.36 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 182.5, 138.5, 134.5, 133.8, 129.1, 128.6, 128.2, 127.1, 66.7, 58.4, 48.4, 44.4, 13.6.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1722, 1601, 1495, 1400, 1377, 1276, 1260, 1090, 1016.

HRMS (ESI⁺) calcd. for $\text{C}_{19}\text{H}_{19}\text{ClNO}^+$ [$\text{M}+\text{H}$]⁺: 312.1150, found: 312.1138.

1-Benzyl-6-(4-chlorophenyl)-3-methyl-3,4-dihydropyridin-2(1H)-one (4c)



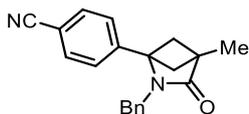
^1H NMR (500 MHz, CDCl_3) δ 7.25 (d, J = 8.5 Hz, 2H), 7.20 – 7.14 (m, 3H), 7.03 (d, J = 8.5 Hz, 2H), 6.91 – 6.87 (m, 2H), 5.31 (dd, J = 6.0, 3.9 Hz, 1H), 5.01 (d, J = 15.3 Hz, 1H), 4.42 (d, J = 15.4 Hz, 1H), 2.73 – 2.62 (m, 1H), 2.40 (dt, J = 16.4, 6.0 Hz, 1H), 2.14 (ddd, J = 16.2, 11.7, 3.9 Hz, 1H), 1.30 (d, J = 6.9 Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 174.4, 141.3, 137.9, 134.6, 134.1, 129.2, 128.5, 128.3, 127.2, 127.0, 110.5, 46.3, 35.7, 27.8, 15.5.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1672, 1488, 1454, 1381, 1222, 1088, 1015.

HRMS (ESI⁺) calcd. for $\text{C}_{19}\text{H}_{19}\text{ClNO}^+$ [$\text{M}+\text{H}$]⁺: 312.1150, found: 312.1149.

4-(2-Benzyl-4-methyl-3-oxo-2-azabicyclo[2.1.1]hexan-1-yl)benzonitrile (3d)



Prepared according to **General Procedure D** using **1d** (410 mg, 1.36 mmol, 1.00 equiv), CH_3CN (13.6 mL), and $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbbpy})](\text{PF}_6)$ (15.2 mg, 13.5 μmol , 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 50%), affording **3d** (275 mg, 0.91 mmol, 67%) as a white solid.

M.P.: 124-126 °C

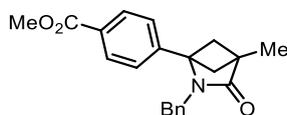
^1H NMR (500 MHz, CDCl_3) δ 7.52 (d, J = 7.9 Hz, 2H), 7.13 – 7.07 (m, 5H), 6.74 (d, J = 6.7 Hz, 2H), 4.20 (s, 2H), 2.72 (dd, J = 4.1, 1.8 Hz, 2H), 2.40 (dd, J = 4.3, 1.5 Hz, 2H), 1.37 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 182.3, 140.3, 138.0, 132.1, 128.5, 128.3, 128.1, 127.3, 118.5, 112.4, 66.6, 58.1, 48.4, 44.7, 13.5.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 2228, 1718, 1612, 1454, 1395, 1357, 1276, 1132, 1017.

HRMS (ESI⁺) calcd. for C₂₀H₁₉N₂O⁺ [M+H]⁺: 303.1492, found: 303.1486.

Methyl 4-(2-benzyl-4-methyl-3-oxo-2-azabicyclo[2.1.1]hexan-1-yl)benzoate (3e)



Prepared according to **General Procedure D** using **1e** (288 mg, 0.86 mmol, 1.00 equiv), CH₃CN (8.6 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (9.6 mg, 8.6 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 40%), affording **3e** (151 mg, 0.45 mmol, 52%) as a colourless oil.

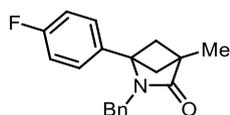
¹H NMR (500 MHz, CDCl₃) δ 7.90 (d, *J* = 8.4 Hz, 2H), 7.10 – 7.04 (m, 5H), 6.76 – 6.72 (m, 2H), 4.18 (s, 2H), 3.89 (s, 3H), 2.69 (dd, *J* = 4.0, 1.9 Hz, 2H), 2.38 (dd, *J* = 3.9, 2.0 Hz, 2H), 1.34 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 182.4, 166.6, 140.0, 138.2, 130.1, 129.5, 128.12, 128.09, 127.7, 127.0, 66.8, 58.2, 52.2, 48.2, 44.4, 13.5.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1717, 1615, 1495, 1435, 1377, 1275, 1178, 1110, 1020.

HRMS (ESI⁺) calcd. for C₂₁H₂₂NO₃⁺ [M+H]⁺: 336.1594, found: 336.1587.

2-Benzyl-1-(4-fluorophenyl)-4-methyl-2-azabicyclo[2.1.1]hexan-3-one (3f)



Prepared according to **General Procedure D** using **1f** (225 mg, 0.76 mmol, 1.00 equiv), CH₃CN (7.6 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (8.6 mg, 7.6 μmol, 1 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 40%), affording **3f** (98 mg, 0.33 mmol, 44%) as a white solid.

M.P.: 84-86 °C

¹H NMR (500 MHz, CDCl₃) δ 7.13 – 7.08 (m, 3H), 6.98 – 6.90 (m, 4H), 6.82 – 6.78 (m, 2H), 4.19 (s, 2H), 2.70 (dd, *J* = 4.0, 1.8 Hz, 2H), 2.37 (dd, *J* = 3.8, 2.0 Hz, 2H), 1.35 (s, 3H).

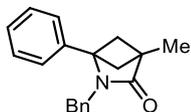
¹³C NMR (126 MHz, CDCl₃) δ 182.4, 162.7 (d, *J* = 248.0 Hz), 138.5, 131.1 (d, *J* = 3.2 Hz), 129.5 (d, *J* = 8.3 Hz), 128.103, 128.097, 127.0, 115.3 (d, *J* = 21.5 Hz), 66.6, 58.4, 48.2, 44.3, 13.6.

¹⁹F NMR (377 MHz, CDCl₃) δ -112.98.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1723, 1611, 1516, 1377, 1276, 1260, 1224, 1158, 1015.

HRMS (ESI⁺) calcd. for C₁₉H₁₉FNO⁺ [M+H]⁺: 296.1445, found: 296.1431.

2-Benzyl-4-methyl-1-phenyl-2-azabicyclo[2.1.1]hexan-3-one (**3g**)



Prepared according to **General Procedure D** using **1g** (290 mg, 1.05 mmol, 1.00 equiv), CH₃CN (10.5 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.7 mg, 10.4 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 40%), affording **3g** (136 mg, 0.49 mmol, 47%) as a colourless oil.

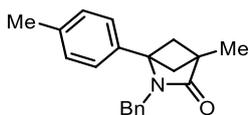
¹H NMR (500 MHz, CDCl₃) δ 7.33 – 7.24 (m, 3H), 7.13 – 7.08 (m, 3H), 7.03 – 7.00 (m, 2H), 6.82 – 6.78 (m, 2H), 4.20 (s, 2H), 2.71 (dd, *J* = 4.0, 1.9 Hz, 2H), 2.39 (dd, *J* = 3.9, 2.0 Hz, 2H), 1.36 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 182.6, 138.6, 135.1, 128.4, 128.3, 128.2, 128.0, 127.6, 126.8, 67.2, 58.3, 48.2, 44.2, 13.6.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1717, 1495, 1448, 1376, 1357, 1276, 1260, 1130.

HRMS (ESI⁺) calcd. for C₁₉H₂₀NO⁺ [M+H]⁺: 278.1539, found: 278.1534.

2-Benzyl-4-methyl-1-(*p*-tolyl)-2-azabicyclo[2.1.1]hexan-3-one (**3h**)



Prepared according to **General Procedure D** using **1h** (285 mg, 0.98 mmol, 1.00 equiv), CH₃CN (9.8 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.0 mg, 9.80 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **3h** (113 mg, 0.39 mmol, 40%) as a white solid.

M.P.: 71-73 °C

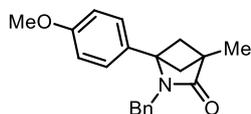
¹H NMR (500 MHz, CDCl₃) δ 7.14 – 7.10 (m, 3H), 7.08 (d, *J* = 7.9 Hz, 2H), 6.92 (d, *J* = 8.0 Hz, 2H), 6.85 – 6.82 (m, 2H), 4.20 (s, 2H), 2.70 (dd, *J* = 4.0, 1.9 Hz, 2H), 2.37 (dd, *J* = 4.0, 1.9 Hz, 2H), 2.35 (s, 3H), 1.36 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 182.7, 138.8, 138.3, 132.1, 129.0, 128.2, 128.0, 127.6, 126.8, 67.2, 58.4, 48.2, 44.2, 21.3, 13.6.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1722, 1475, 1454, 1394, 1376, 1276, 1261.

HRMS (ESI⁺) calcd. for C₂₀H₂₂NO⁺ [M+H]⁺: 292.1696, found: 292.1690.

2-Benzyl-1-(4-methoxyphenyl)-4-methyl-2-azabicyclo[2.1.1]hexan-3-one (**3i**)



Prepared according to **General Procedure D** using **1i** (307 mg, 1.00 mmol, 1.00 equiv), CH₃CN (10.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.2 mg, 10.0 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **3i** (74 mg, 0.24 mmol, 24%) as a white solid.

M.P.: 70-72 °C

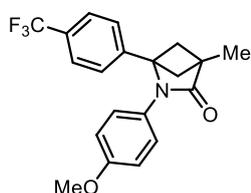
¹H NMR (500 MHz, CDCl₃) δ 7.14 – 7.10 (m, 3H), 6.92 (d, *J* = 8.7 Hz, 2H), 6.85 – 6.82 (m, 2H), 6.78 (d, *J* = 8.7 Hz, 2H), 4.19 (s, 2H), 3.80 (s, 3H), 2.69 (dd, *J* = 4.0, 1.9 Hz, 2H), 2.36 (dd, *J* = 3.7, 1.9 Hz, 2H), 1.35 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 182.6, 159.7, 138.8, 129.0, 128.2, 128.1, 127.3, 126.9, 113.7, 67.0, 58.5, 55.4, 48.2, 44.1, 13.6.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1716, 1615, 1519, 1376, 1247, 1175, 1032.

HRMS (ESI⁺) calcd. for C₂₀H₂₂NO₂⁺ [M+H]⁺: 308.1645, found: 308.1642.

2-(4-Methoxyphenyl)-4-methyl-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.1.1]hexan-3-one (**3j**)



Prepared according to **General Procedure D** using **1j** (600 mg, 1.66 mmol, 1.00 equiv), CH₃CN (16.6 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (18.6 mg, 16.6 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 50%), affording **3j** (363 mg, 1.01 mmol, 61%) as a pale-yellow solid.

M.P.: 80-82 °C

¹H NMR (500 MHz, CDCl₃) δ 7.49 (d, *J* = 8.2 Hz, 2H), 7.23 (d, *J* = 7.9 Hz, 2H), 6.73 (d, *J* = 9.0 Hz, 2H), 6.63 (d, *J* = 9.1 Hz, 2H), 3.63 (s, 3H), 2.95 (dd, *J* = 4.2, 2.0 Hz, 2H), 2.59 (dd, *J* = 4.2, 2.0 Hz, 2H), 1.40 (s, 3H).

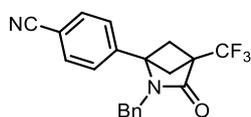
¹³C NMR (126 MHz, CDCl₃) δ 181.3, 157.3, 139.6, 130.2 (q, *J* = 32.5 Hz), 129.5, 127.8, 127.1, 125.3 (q, *J* = 3.7 Hz), 123.9 (q, *J* = 272.3 Hz), 113.9, 69.1, 57.4, 55.1, 48.6, 13.4.

¹⁹F NMR (471 MHz, CDCl₃) δ -62.57.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2248, 1748, 1725, 1509, 1324, 1290, 1165, 1123, 1067.

HRMS (ESI⁺) calcd. for C₂₀H₁₉F₃NO₂⁺ [M+H]⁺: 362.1362, found: 362.1351.

4-(2-Benzyl-3-oxo-4-(trifluoromethyl)-2-azabicyclo[2.1.1]hexan-1-yl)benzonitrile (**3k**)



Prepared according to **General Procedure D** using **1k** (356 mg, 1.00 mmol, 1.00 equiv), CH₃CN (10.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.2 mg, 10.0 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 50%), affording **3k** (295 mg, 0.83 mmol, 83%) as a white solid.

M.P.: 134-136 °C

¹H NMR (500 MHz, CDCl₃) δ 7.56 (d, *J* = 7.9 Hz, 2H), 7.23-7.01 (m, 5H), 6.72 (d, *J* = 7.5 Hz, 2H), 4.23 (s, 2H), 2.97 (s, 2H), 2.79 (s, 2H).

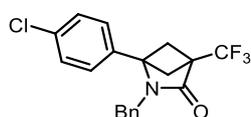
¹³C NMR (126 MHz, CDCl₃) δ 173.2, 137.9, 136.8, 132.3, 128.4, 128.3, 127.9, 127.6, 122.6 (q, *J* = 274.8 Hz), 118.0, 113.1, 65.8, 53.7, 51.5 (q, *J* = 34.7 Hz), 44.4.

¹⁹F NMR (471 MHz, CDCl₃) δ -71.21.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2231, 1737, 1482, 1405, 1265, 1218, 1182, 1161.

HRMS (ESI⁺) calcd. for C₂₀H₁₆F₃N₂O⁺ [M+H]⁺: 357.1209, found: 357.1210.

2-Benzyl-1-(4-chlorophenyl)-4-(trifluoromethyl)-2-azabicyclo[2.1.1]hexan-3-one (**3l**)



Prepared according to **General Procedure D** using **11** (425 mg, 1.16 mmol, 1.00 equiv), CH₃CN (11.6 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (13.1 mg, 11.7 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **31** (301 mg, 0.82 mmol, 71%) as a white solid.

M.P.: 67-69 °C

¹H NMR (500 MHz, CDCl₃) δ 7.28 (d, *J* = 8.5 Hz, 2H), 7.19 – 7.12 (m, 3H), 6.94 (d, *J* = 8.4 Hz, 2H), 6.81 – 6.77 (m, 2H), 4.24 (s, 2H), 2.93 (dd, *J* = 4.1, 1.9 Hz, 2H), 2.76 (dd, *J* = 4.1, 2.0 Hz, 2H).

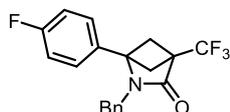
¹³C NMR (126 MHz, CDCl₃) δ 173.5 (q, *J* = 2.6 Hz), 137.3, 135.5, 131.5, 129.0, 128.9, 128.4, 128.2, 127.5, 122.8 (q, *J* = 274.9 Hz), 66.0, 53.9 (q, *J* = 2.6 Hz), 51.6 (q, *J* = 34.7 Hz), 44.3.

¹⁹F NMR (471 MHz, CDCl₃) δ -71.29.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1734, 1480, 1403, 1320, 1217, 1182, 1153, 1119, 1091.

HRMS (ESI⁺) calcd. for C₁₉H₁₆ClF₃NO⁺ [M+H]⁺: 366.0867, found: 366.0862.

2-Benzyl-1-(4-fluorophenyl)-4-(trifluoromethyl)-2-azabicyclo[2.1.1]hexan-3-one (**3m**)



Prepared according to **General Procedure D** using **1m** (349 mg, 1.00 mmol, 1.00 equiv), CH₃CN (10.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.2 mg, 10.0 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **3m** (192 mg, 0.55 mmol, 55%) as a colourless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.18 – 7.11 (m, 3H), 7.02 – 6.96 (m, 4H), 6.80 – 6.77 (m, 2H), 4.23 (s, 2H), 2.94 (dd, *J* = 4.1, 1.9 Hz, 2H), 2.76 (dd, *J* = 3.8, 2.1 Hz, 2H).

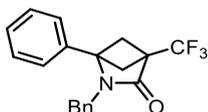
¹³C NMR (126 MHz, CDCl₃) δ 173.5 (q, *J* = 2.6 Hz), 163.2 (d, *J* = 249.2 Hz), 137.4, 129.5 (d, *J* = 8.4 Hz), 128.9 (d, *J* = 3.2 Hz), 128.4, 128.1, 127.4, 122.8 (q, *J* = 274.8 Hz), 115.7 (d, *J* = 21.8 Hz), 66.0, 54.0 (q, *J* = 2.3 Hz), 51.5 (q, *J* = 34.5 Hz), 44.2.

¹⁹F NMR (471 MHz, CDCl₃) δ -71.29, -111.37.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1733, 1604, 1518, 1481, 1403, 1276, 1217, 1181, 1118.

HRMS (ESI⁺) calcd. for C₁₉H₁₆F₄NO⁺ [M+H]⁺: 350.1163, found: 350.1159.

2-Benzyl-1-phenyl-4-(trifluoromethyl)-2-azabicyclo[2.1.1]hexan-3-one (**3n**)



Prepared according to **General Procedure D** using **1n** (185 mg, 0.56 mmol, 1.00 equiv), CH₃CN (5.6 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (6.3 mg, 5.6 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **3n** (120 mg, 0.36 mmol, 65%) as a white solid.

M.P.: 55-57 °C

¹H NMR (500 MHz, CDCl₃) δ 7.41 – 7.37 (m, 1H), 7.37 – 7.29 (m, 2H), 7.18 – 7.11 (m, 3H), 7.04 – 7.00 (m, 2H), 6.80 – 6.76 (m, 2H), 4.25 (s, 2H), 2.95 (dd, *J* = 4.1, 1.9 Hz, 2H), 2.78 (dd, *J* = 3.7, 2.2 Hz, 2H).

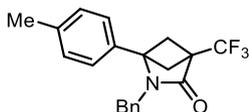
¹³C NMR (126 MHz, CDCl₃) δ 173.7 (q, *J* = 2.6 Hz), 137.5, 132.9, 129.3, 128.7, 128.3, 127.5, 127.3, 122.9 (q, *J* = 274.7 Hz), 66.6, 54.0 (q, *J* = 2.3 Hz), 51.6 (q, *J* = 34.5 Hz), 44.1.

¹⁹F NMR (471 MHz, CDCl₃) δ –71.26.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1735, 1481, 1450, 1404, 1321, 1276, 1261, 1217, 1181.

HRMS (ESI⁺) calcd. for C₁₉H₁₇F₃NO⁺ [M+H]⁺: 332.1257, found: 332.1249.

2-Benzyl-1-(*p*-tolyl)-4-(trifluoromethyl)-2-azabicyclo[2.1.1]hexan-3-one (**3o**)



Prepared according to **General Procedure D** using **1o** (345 mg, 1.00 mmol, 1.00 equiv), CH₃CN (10.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.2 mg, 10.0 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **3o** (178 mg, 0.52 mmol, 52%) as a white solid.

M.P.: 66-68 °C

¹H NMR (500 MHz, CDCl₃) δ 7.19 – 7.12 (m, 5H), 6.92 (d, *J* = 8.2 Hz, 2H), 6.84 – 6.80 (m, 2H), 4.24 (s, 2H), 2.93 (dd, *J* = 4.1, 1.9 Hz, 2H), 2.75 (dd, *J* = 4.0, 2.1 Hz, 2H), 2.39 (s, 3H).

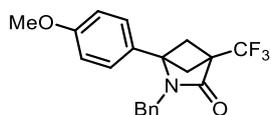
¹³C NMR (126 MHz, CDCl₃) δ 173.8 (q, *J* = 2.3 Hz), 139.3, 137.6, 129.9, 129.3, 128.3, 128.2, 127.4, 127.3, 122.9 (q, *J* = 274.8 Hz), 66.5, 54.0 (q, *J* = 2.3 Hz), 51.6 (q, *J* = 34.3 Hz), 44.1, 21.3.

¹⁹F NMR (471 MHz, CDCl₃) δ –71.26.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1733, 1403, 1275, 1261, 1218, 1180, 1150, 1109.

HRMS (ESI⁺) calcd. for C₂₀H₁₉F₃NO⁺ [M+H]⁺: 346.1413, found: 346.1403.

2-Benzyl-1-(4-methoxyphenyl)-4-(trifluoromethyl)-2-azabicyclo[2.1.1]hexan-3-one (**3p**)



Prepared according to **General Procedure D** using **1p** (375 mg, 1.04 mmol, 1.00 equiv), CH₃CN (10.4 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.7 mg, 10.4 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (0% to 50%), affording **3p** (135 mg, 0.37 mmol, 36%) as a colourless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.18 – 7.12 (m, 3H), 6.92 (d, *J* = 8.7 Hz, 2H), 6.86 – 6.80 (m, 4H), 4.23 (s, 2H), 3.83 (s, 3H), 2.92 (dd, *J* = 4.1, 1.9 Hz, 2H), 2.74 (dd, *J* = 3.9, 2.0 Hz, 2H).

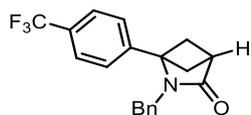
¹³C NMR (126 MHz, CDCl₃) δ 173.7 (q, *J* = 2.6 Hz), 160.3, 137.7, 128.9, 128.29, 128.28, 127.3, 125.0, 122.9 (q, *J* = 274.9 Hz), 114.1, 66.4, 55.5, 54.1 (q, *J* = 2.2 Hz), 51.6 (q, *J* = 34.3 Hz), 44.0.

¹⁹F NMR (471 MHz, CDCl₃) δ –71.29.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1732, 1616, 1521, 1481, 1403, 1275, 1217, 1179, 1152.

HRMS (ESI⁺) calcd. for C₂₀H₁₉F₃NO₂⁺ [M+H]⁺: 362.1362, found: 362.1353.

2-Benzyl-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.1.1]hexan-3-one (**3q**)



Prepared according to **General Procedure D** using **1q** (331 mg, 1.00 mmol, 1.00 equiv), CH₃CN (11.2 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.2 mg, 10.0 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 50%), affording **3q** (156 mg, 0.47 mmol, 47%) as a pale-yellow solid.

M.P.: 52-54 °C

¹H NMR (500 MHz, CDCl₃) δ 7.50 (d, *J* = 8.0 Hz, 2H), 7.14 – 7.07 (m, 5H), 6.77 – 6.74 (m, 2H), 4.21 (s, 2H), 2.93 (t, *J* = 2.7 Hz, 1H), 2.80 – 2.75 (m, 2H), 2.62 – 2.59 (m, 2H).

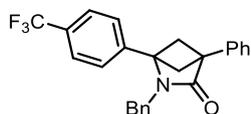
¹³C NMR (126 MHz, CDCl₃) δ 181.9, 139.1 (q, *J* = 1.6 Hz), 138.0, 130.8 (q, *J* = 32.6 Hz), 128.2, 128.1, 128.0, 127.2, 125.4 (q, *J* = 3.8 Hz), 124.0 (q, *J* = 272.1 Hz), 70.6, 54.3, 44.3, 43.2.

¹⁹F NMR (471 MHz, CDCl₃) δ –62.70.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1721, 1621, 1496, 1411, 1382, 1322, 1164, 1119, 1105.

HRMS (ESI⁺) calcd. for C₁₉H₁₇F₃NO⁺ [M+H]⁺: 332.1257, found: 332.1250.

2-Benzyl-4-phenyl-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.1.1]hexan-3-one (3r)



Prepared according to **General Procedure D** using **1r** (430 mg, 1.06 mmol, 1.00 equiv), CH₃CN (10.6 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.9 mg, 10.6 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **3r** (180 mg, 0.44 mmol, 42%) as a white solid.

M.P.: 89-91 °C

¹H NMR (500 MHz, CDCl₃) δ 7.58 (d, *J* = 8.0 Hz, 2H), 7.46 – 7.40 (m, 4H), 7.37 – 7.33 (m, 1H), 7.23 (d, *J* = 8.0 Hz, 2H), 7.20 – 7.14 (m, 3H), 6.87 – 6.83 (m, 2H), 4.34 (s, 2H), 3.18 (dd, *J* = 4.0, 1.9 Hz, 2H), 2.82 (dd, *J* = 3.9, 1.9 Hz, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 179.9, 138.8, 138.0, 135.8, 130.8 (q, *J* = 32.7 Hz), 128.5, 128.24, 128.22, 128.1, 127.6, 127.22, 127.17, 125.4 (q, *J* = 3.7 Hz), 124.0 (q, *J* = 272.1 Hz), 65.8, 57.6, 55.0, 44.8.

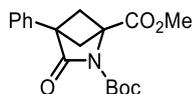
¹⁹F NMR (471 MHz, CDCl₃) δ -62.57.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1722, 1411, 1321, 1165, 1120, 1067.

HRMS (ESI⁺) calcd. for C₂₅H₂₁F₃NO⁺ [M+H]⁺: 408.1570, found: 408.1562.

2.4.2 Synthesis and Characterisation of Products (8)

2-(*tert*-Butyl) 1-methyl 3-oxo-4-phenyl-2-azabicyclo[2.1.1]hexane-1,2-dicarboxylate (**8a**)



Prepared according to **General Procedure D** using **7a** (460 mg, 1.39 mmol, 1.00 equiv), CH₃CN (14.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (15.6 mg, 13.9 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **8a** (382 mg, 1.15 mmol, 83%) as a white solid.

M.P.: 124-126 °C

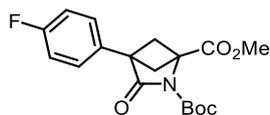
¹H NMR (500 MHz, CDCl₃) δ 7.39 – 7.35 (m, 2H), 7.34 – 7.30 (m, 1H), 7.25 – 7.22 (m, 2H), 3.83 (s, 3H), 2.91 (dd, *J* = 4.8, 2.0 Hz, 2H), 2.81 (dd, *J* = 4.8, 2.0 Hz, 2H), 1.51 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 174.2, 166.7, 149.6, 133.7, 128.7, 128.2, 127.0, 83.9, 62.9, 56.1, 53.0, 52.6, 28.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1801, 1751, 1718, 1439, 1370, 1309, 1259, 1218, 1117.

HRMS (ESI⁺) calcd. for C₁₈H₂₁NO₅Na⁺ [M+Na]⁺: 354.1312, found: 354.1296.

2-(*tert*-Butyl) 1-methyl 4-(4-fluorophenyl)-3-oxo-2-azabicyclo[2.1.1]hexane-1,2-dicarboxylate (**8b**)



Prepared according to **General Procedure D** using **7b** (386 mg, 1.10 mmol, 1.00 equiv), CH₃CN (11.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (12.4 mg, 11.1 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 25%), affording **8b** (330 mg, 0.95 mmol, 85%) as a white solid.

M.P.: 99-101 °C

¹H NMR (500 MHz, CDCl₃) δ 7.19 – 7.16 (m, 2H), 7.02 – 6.97 (m, 2H), 3.78 (s, 3H), 2.84 (dd, *J* = 4.8, 2.0 Hz, 2H), 2.76 (dd, *J* = 4.8, 2.0 Hz, 2H), 1.46 (s, 9H).

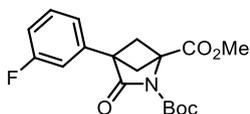
¹³C NMR (126 MHz, CDCl₃) δ 174.0, 166.4, 162.4 (d, *J* = 247.0 Hz), 149.3, 129.5 (d, *J* = 3.1 Hz), 128.8 (d, *J* = 8.3 Hz), 115.5 (d, *J* = 21.5 Hz), 83.8, 62.7, 55.3, 52.9, 52.4, 27.9.

¹⁹F NMR (471 MHz, CDCl₃) δ -113.63.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1798, 1749, 1716, 1610, 1517, 1439, 1369, 1218, 1116.

HRMS (ESI⁺) calcd. for C₁₈H₂₀FNO₅Na⁺ [M+Na]⁺: 372.1218, found: 372.1200.

2-(*tert*-Butyl) 1-methyl 4-(3-fluorophenyl)-3-oxo-2-azabicyclo[2.1.1]hexane-1,2-dicarboxylate (8c)



Prepared according to **General Procedure D** using **7c** (174 mg, 0.50 mmol, 1.00 equiv), CH₃CN (5.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (5.6 mg, 5.0 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 25%), affording **8c** (151 mg, 0.43 mmol, 87%) as a white solid.

M.P.: 128-130 °C

¹H NMR (500 MHz, CDCl₃) δ 7.32 (td, *J* = 8.0, 5.8 Hz, 1H), 7.02 – 6.97 (m, 2H), 6.95 (ddd, *J* = 9.6, 2.6, 1.7 Hz, 1H), 3.82 (s, 3H), 2.89 (dd, *J* = 4.8, 2.0 Hz, 2H), 2.79 (dd, *J* = 4.8, 2.0 Hz, 2H), 1.49 (s, 9H).

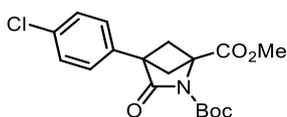
¹³C NMR (126 MHz, CDCl₃) δ 173.7, 166.5, 162.8 (d, *J* = 246.7 Hz), 149.4, 136.1 (d, *J* = 7.7 Hz), 130.3 (d, *J* = 8.3 Hz), 122.8 (d, *J* = 3.0 Hz), 115.2 (d, *J* = 21.0 Hz), 114.3 (d, *J* = 22.3 Hz), 84.0, 62.8, 55.6 (d, *J* = 2.1 Hz), 53.0, 52.6, 28.0.

¹⁹F NMR (471 MHz, CDCl₃) δ -112.36.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1801, 1787, 1749, 1716, 1589, 1442, 1369, 1300, 1156.

HRMS (ESI⁺) calcd. for C₁₈H₂₀FNO₅Na⁺ [M+Na]⁺: 372.1218, found: 372.1201.

2-(*tert*-Butyl) 1-methyl 4-(4-chlorophenyl)-3-oxo-2-azabicyclo[2.1.1]hexane-1,2-dicarboxylate (8d)



Prepared according to **General Procedure D** using **7d** (295 mg, 0.81 mmol, 1.00 equiv), CH₃CN (8.1 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (9.1 mg, 8.1 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 25%), affording **8d** (256 mg, 0.70 mmol, 88%) as a white solid.

M.P.: 109-111 °C

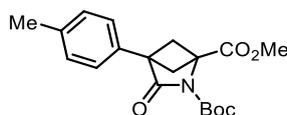
¹H NMR (500 MHz, CDCl₃) δ 7.31 (d, *J* = 8.5 Hz, 2H), 7.15 (d, *J* = 8.5 Hz, 2H), 3.80 (s, 3H), 2.86 (dd, *J* = 4.8, 2.0 Hz, 2H), 2.77 (dd, *J* = 4.8, 2.0 Hz, 2H), 1.48 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 173.8, 166.4, 149.3, 134.1, 132.2, 128.8, 128.5, 83.9, 62.8, 55.4, 52.9, 52.5, 27.9.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1798, 1748, 1716, 1498, 1369, 1301, 1218, 1149.

HRMS (ESI⁺) calcd. for C₁₈H₂₀ClNO₅Na⁺ [M+Na]⁺: 388.0922, found: 388.0903.

2-(*tert*-Butyl) 1-methyl 3-oxo-4-(*p*-tolyl)-2-azabicyclo[2.1.1]hexane-1,2-dicarboxylate (**8e**)



Prepared according to **General Procedure D** using **7e** (365 mg, 1.06 mmol, 1.00 equiv), CH₃CN (10.6 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.9 mg, 10.6 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 25%), affording **8e** (281 mg, 0.81 mmol, 77%) as a white solid.

M.P.: 140-142 °C

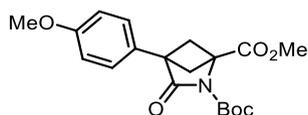
¹H NMR (500 MHz, CDCl₃) δ 7.17 (d, *J* = 7.9 Hz, 2H), 7.12 (d, *J* = 8.3 Hz, 2H), 3.82 (s, 3H), 2.87 (dd, *J* = 4.7, 2.0 Hz, 2H), 2.77 (dd, *J* = 4.8, 2.0 Hz, 2H), 2.33 (s, 3H), 1.51 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 174.2, 166.6, 149.4, 137.8, 130.6, 129.2, 126.8, 83.6, 62.7, 55.7, 52.8, 52.4, 27.9, 21.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1785, 1751, 1718, 1444, 1369, 1309, 1210, 1151, 1113.

HRMS (ESI⁺) calcd. for C₁₉H₂₃NO₅Na⁺ [M+Na]⁺: 368.1468, found: 368.1453.

2-(*tert*-Butyl) 1-methyl 4-(4-methoxyphenyl)-3-oxo-2-azabicyclo[2.1.1]hexane-1,2-dicarboxylate (**8f**)



Prepared according to **General Procedure D** using **7f** (330 mg, 0.91 mmol, 1.00 equiv), CH₃CN (9.1 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (10.2 mg, 9.1 μmol, 1 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **8f** (252 mg, 0.70 mmol, 76%) as a white solid.

M.P.: 125-127 °C

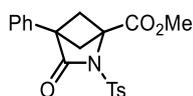
¹H NMR (500 MHz, CDCl₃) δ 7.14 (d, *J* = 8.7 Hz, 2H), 6.87 (d, *J* = 8.8 Hz, 2H), 3.80 (s, 3H), 3.75 (s, 3H), 2.84 (dd, *J* = 4.7, 2.0 Hz, 2H), 2.75 (dd, *J* = 4.8, 1.9 Hz, 2H), 1.48 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 174.4, 166.7, 159.3, 149.4, 128.1, 125.7, 114.0, 83.6, 62.7, 55.5, 55.2, 52.9, 52.4, 27.9.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1799, 1786, 1748, 1715, 1616, 1519, 1439, 1369, 1248, 1152.

HRMS (ESI⁺) calcd. for C₁₉H₂₃NO₆Na⁺ [M+Na]⁺: 384.1418, found: 384.1405.

Methyl 3-oxo-4-phenyl-2-tosyl-2-azabicyclo[2.1.1]hexane-1-carboxylate (**8g**)



Prepared according to **General Procedure D** using **7g** (1.19 g, 3.09 mmol, 1.00 equiv), CH₃CN (31.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (34.8 mg, 31.0 μmol, 1 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 40%), affording **8g** (1.05 g, 2.73 mmol, 88%) as a white solid.

M.P.: 138-140 °C

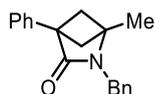
¹H NMR (500 MHz, CDCl₃) δ 7.92 (d, *J* = 8.3 Hz, 2H), 7.39 – 7.28 (m, 5H), 7.13 (d, *J* = 8.3 Hz, 2H), 3.92 (s, 3H), 2.81-2.73 (m, 4H), 2.44 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 174.4, 166.0, 145.3, 136.2, 132.9, 129.9, 128.7, 128.4, 128.0, 126.9, 66.6, 55.9, 53.1, 52.9, 21.8.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1782, 1747, 1597, 1439, 1363, 1225, 1201, 1140, 1089.

HRMS (ESI⁺) calcd. for C₂₀H₁₉NO₅SNa⁺ [M+Na]⁺: 408.0876, found: 408.0860.

2-Benzyl-1-methyl-4-phenyl-2-azabicyclo[2.1.1]hexan-3-one (**8h**)



Prepared according to **General Procedure D** using **7h** (1.26 g, 4.55 mmol, 1.00 equiv), CH₃CN (45.5 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (51.1 mg, 0.046 mmol, 1 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **8h** (845 mg, 3.05 mmol, 67%) as a colourless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.42 – 7.26 (m, 10H), 4.58 (s, 2H), 2.76 (dd, *J* = 4.0, 1.9 Hz, 2H), 2.51 (dd, *J* = 3.9, 2.0 Hz, 2H), 1.37 (s, 3H).

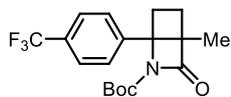
¹³C NMR (126 MHz, CDCl₃) δ 181.0, 139.4, 136.6, 128.6, 128.4, 127.7, 127.23, 127.18, 62.1, 59.3, 55.4, 43.8, 16.5.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1713, 1604, 1503, 1454, 1399, 1334, 1276, 1164, 1075.

HRMS (ESI⁺) calcd. for C₁₉H₂₀NO⁺ [M+H]⁺: 278.1539, found: 278.1533.

2.4.3 Synthesis and Characterisation of Products (5, 6 and 9)

tert-Butyl 4-methyl-3-oxo-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.2.0]hexane-2-carboxylate (**6a**)



Prepared according to **General Procedure D** using **2a** (490 mg, 1.38 mmol, 1.00 equiv), CH₃CN (13.8 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (15.5 mg, 13.8 mmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **6a** (132 mg, 0.37 mmol, 27%) as a white solid and **5a** (222 mg, 0.63 mmol, 45 %) as a white solid.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 1:1.6 molar ratio of **6a**/**5a**.

M.P.: 94-96 °C

¹H NMR (500 MHz, CDCl₃) δ 7.65 (d, *J* = 8.2 Hz, 2H), 7.41 (d, *J* = 8.2 Hz, 2H), 3.11 – 2.96 (m, 1H), 2.57 – 2.48 (m, 2H), 2.22 – 2.12 (m, 1H), 1.41 (s, 9H), 1.06 (s, 3H).

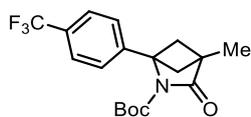
¹³C NMR (126 MHz, CDCl₃) δ 170.6, 147.7, 139.7, 130.1 (q, *J* = 32.7 Hz), 126.7, 125.6 (q, *J* = 3.7 Hz), 124.1 (q, *J* = 272.1 Hz), 83.7, 65.9, 61.9, 28.0, 26.3, 25.7, 13.1.

¹⁹F NMR (471 MHz, CDCl₃) δ –62.63.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1800, 1720, 1620, 1371, 1320, 1278, 1151, 1119, 1067.

HRMS (ESI⁺) calcd. for C₁₈H₂₀F₃NO₃Na⁺ [M+Na]⁺: 378.1287, found: 378.1288.

tert-Butyl 4-methyl-3-oxo-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.1.1]hexane-2-carboxylate (**5a**)



M.P.: 121-123 °C

¹H NMR (500 MHz, CDCl₃) δ 7.61 (d, *J* = 8.1 Hz, 2H), 7.37 (d, *J* = 8.1 Hz, 2H), 2.73 (dd, *J* = 4.7, 2.0 Hz, 2H), 2.41 (dd, *J* = 4.7, 2.0 Hz, 2H), 1.35 (s, 3H), 1.13 (s, 9H).

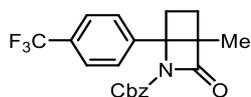
¹³C NMR (126 MHz, CDCl₃) δ 178.6, 149.5, 141.4, 130.2 (q, *J* = 32.4 Hz), 127.0, 125.4 (q, *J* = 3.8 Hz), 124.1 (q, *J* = 272.1 Hz), 82.5, 67.7, 54.8, 49.9, 27.6, 13.2.

¹⁹F NMR (471 MHz, CDCl₃) δ –62.62.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1790, 1720, 1623, 1368, 1324, 1266, 1159, 1125, 1068.

HRMS (ESI⁺) calcd. for C₁₈H₂₀F₃NO₃Na⁺ [M+Na]⁺: 378.1287, found: 378.1289.

Benzyl 4-methyl-3-oxo-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.2.0]hexane-2-carboxylate (6b)



Prepared according to **General Procedure D** using **2b** (530 mg, 1.36 mmol, 1.00 equiv), CH₃CN (13.6 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (15.3 mg, 13.6 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/CH₂Cl₂ (2% to 5%), affording **6b** (176 mg, 0.45 mmol, 33%) as a white solid and **5b** (178 mg, 0.45 mmol, 33 %) as a white solid.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 1:1 molar ratio of **6b/5b**.

M.P.: 142-144 °C

¹H NMR (500 MHz, CDCl₃) δ 7.64 (d, *J* = 8.2 Hz, 2H), 7.41 (d, *J* = 8.1 Hz, 2H), 7.32 – 7.29 (m, 3H), 7.25 – 7.21 (m, 2H), 5.26 (d, *J* = 12.4 Hz, 1H), 5.19 (d, *J* = 12.4 Hz, 1H), 3.10 – 3.00 (m, 1H), 2.61 – 2.50 (m, 2H), 2.25 – 2.15 (m, 1H), 1.10 (s, 3H).

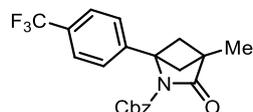
¹³C NMR (126 MHz, CDCl₃) δ 170.1, 148.8, 139.2, 134.9, 130.2 (q, *J* = 32.6 Hz), 128.6, 128.5, 128.1, 126.8, 125.7 (q, *J* = 3.8 Hz), 124.0 (q, *J* = 272.1 Hz), 68.1, 66.0, 62.4, 26.3, 25.6, 13.0.

¹⁹F NMR (471 MHz, CDCl₃) δ –62.58.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1812, 1715, 1620, 1393, 1320, 1205, 1169, 1116, 1067.

HRMS (ESI⁺) calcd. for C₂₁H₁₉F₃NO₃⁺ [M+H]⁺: 390.1312, found: 390.1317.

Benzyl 4-methyl-3-oxo-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.1.1]hexane-2-carboxylate (5b)



M.P.: 124-126 °C

¹H NMR (500 MHz, CDCl₃) δ 7.56 (d, *J* = 8.1 Hz, 2H), 7.36 (d, *J* = 7.6 Hz, 2H), 7.29 – 7.26 (m, 3H), 7.15 – 7.11 (m, 2H), 5.01 (s, 2H), 2.77 (dd, *J* = 4.8, 2.0 Hz, 2H), 2.45 (dd, *J* = 4.7, 2.0 Hz, 2H), 1.39 (s, 3H).

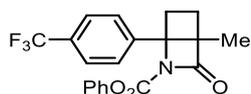
¹³C NMR (126 MHz, CDCl₃) δ 177.8, 151.0, 140.4, 134.9, 130.2 (q, *J* = 32.4 Hz), 128.4, 128.2, 127.9, 127.1, 125.2 (q, *J* = 3.8 Hz), 124.0 (q, *J* = 272.2 Hz), 67.9, 67.7, 54.4, 49.9, 13.1.

^{19}F NMR (471 MHz, CDCl_3) δ -62.44.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1797, 1720, 1623, 1412, 1382, 1324, 1276, 1265, 1169, 1123.

HRMS (ESI⁺) calcd. for $\text{C}_{21}\text{H}_{19}\text{F}_3\text{NO}_3^+$ $[\text{M}+\text{H}]^+$: 390.1312, found: 390.1313.

Phenyl 4-methyl-3-oxo-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.2.0]hexane-2-carboxylate (6c)



Prepared according to **General Procedure D** using **2c** (490 mg, 1.31 mmol, 1.00 equiv), CH_3CN (13.1 mL), and $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbbpy})](\text{PF}_6)$ (14.7 mg, 13.1 μmol , 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of $\text{Et}_2\text{O}/\text{CH}_2\text{Cl}_2$ (2% to 8%), affording **6c** (220 mg, 0.59 mmol, 45%) as a white solid and **5c** (90 mg, 0.24 mmol, 18 %) as a white solid.^A

Notes: (A) The crude ^1H NMR spectrum indicated a 2:1 molar ratio of **6c/5c**.

M.P.: 134-136 °C

^1H NMR (500 MHz, CDCl_3) δ 7.70 (d, J = 8.2 Hz, 2H), 7.52 (d, J = 8.1 Hz, 2H), 7.37 – 7.33 (m, 2H), 7.24 – 7.20 (m, 1H), 7.16 – 7.13 (m, 2H), 3.12 (ddd, J = 13.6, 11.5, 6.3 Hz, 1H), 2.72 (ddd, J = 13.8, 8.7, 5.6 Hz, 1H), 2.63 (ddd, J = 13.0, 8.7, 6.2 Hz, 1H), 2.27 (ddd, J = 13.1, 11.5, 5.6 Hz, 1H), 1.15 (s, 3H).

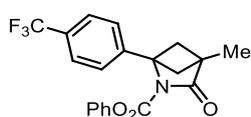
^{13}C NMR (126 MHz, CDCl_3) δ 170.0, 149.9, 147.1, 139.0, 130.3 (q, J = 32.6 Hz), 129.5, 126.8, 126.4, 125.8 (q, J = 3.8 Hz), 124.0 (q, J = 272.1 Hz), 121.2, 66.3, 62.6, 26.4, 25.8, 13.1.

^{19}F NMR (471 MHz, CDCl_3) δ -62.58.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1810, 1736, 1620, 1494, 1411, 1321, 1275, 1190, 1164, 1067.

HRMS (ESI⁺) calcd. for $\text{C}_{20}\text{H}_{16}\text{F}_3\text{NO}_3\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 398.0974, found: 398.0970.

Phenyl 4-methyl-3-oxo-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.1.1]hexane-2-carboxylate (5c)



M.P.: 154-156 °C

¹H NMR (500 MHz, CDCl₃) δ 7.63 (d, *J* = 8.1 Hz, 2H), 7.46 (d, *J* = 7.8 Hz, 2H), 7.29 – 7.25 (m, 2H), 7.17 – 7.13 (m, 1H), 6.99 – 6.95 (m, 2H), 2.88 (dd, *J* = 4.8, 2.1 Hz, 2H), 2.52 (dd, *J* = 4.8, 2.0 Hz, 2H), 1.44 (s, 3H).

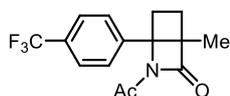
¹³C NMR (126 MHz, CDCl₃) δ 177.3, 150.2, 149.6, 140.0, 130.5 (q, *J* = 32.5 Hz), 129.4, 127.3, 126.0, 125.4 (q, *J* = 3.8 Hz), 124.0 (d, *J* = 272.1 Hz), 121.3, 68.2, 54.3, 50.1, 13.2.

¹⁹F NMR (471 MHz, CDCl₃) δ –62.61.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1793, 1733, 1623, 1494, 1413, 1324, 1275, 1199, 1166, 1067.

HRMS (ESI⁺) calcd. for C₂₀H₁₆F₃NO₃Na⁺ [M+Na]⁺: 398.0974, found: 398.0971.

2-acetyl-4-methyl-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.2.0]hexan-3-one (6d)



Prepared according to **General Procedure D** using **2d** (280 mg, 0.94 mmol, 1.00 equiv), CH₃CN (9.4 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (10.5 mg, 9.4 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (0% to 60%), affording **6d** (175 mg, 0.59 mmol, 63%) as a white solid and **5d** (49 mg, 0.16 mmol, 18%) as a white solid.^{A,B}

Notes: (A) The crude ¹H NMR spectrum indicated a 3.4:1 molar ratio of **6d**/**5d**.

(B) When HFIP was used as the solvent, **6d** was isolated in 78% yield. The crude ¹H NMR spectrum indicated a 12:1 molar ratio of **6d**:**5d**.

M.P.: 70–72 °C

¹H NMR (500 MHz, CDCl₃) δ 7.63 (d, *J* = 8.2 Hz, 2H), 7.37 (d, *J* = 8.1 Hz, 2H), 3.07 – 2.96 (m, 1H), 2.60 – 2.47 (m, 5H), 2.29 – 2.20 (m, 1H), 1.09 (s, 3H).

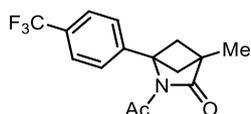
¹³C NMR (126 MHz, CDCl₃) δ 170.9, 168.1, 139.1, 130.1 (q, *J* = 32.6 Hz), 126.6, 125.7 (q, *J* = 3.8 Hz), 124.0 (q, *J* = 272.1 Hz), 65.6, 61.5, 26.4, 26.4, 24.4, 13.0.

¹⁹F NMR (471 MHz, CDCl₃) δ –62.65.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1777, 1703, 1619, 1411, 1374, 1320, 1275, 1267, 1163, 1066.

HRMS (ESI⁺) calcd. for C₁₅H₁₄F₃NO₂Na⁺ [M+Na]⁺: 320.0869, found: 320.0865.

2-Acetyl-4-methyl-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.1.1]hexan-3-one (5d)



M.P.: 60-62 °C

¹H NMR (500 MHz, CDCl₃) δ 7.61 (d, *J* = 8.0 Hz, 2H), 7.37 (d, *J* = 7.8 Hz, 2H), 2.74 (dd, *J* = 4.6, 2.0 Hz, 2H), 2.44 (dd, *J* = 4.7, 2.1 Hz, 2H), 2.40 (s, 3H), 1.39 (s, 3H).

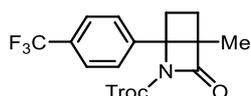
¹³C NMR (126 MHz, CDCl₃) δ 179.3, 169.6, 141.0, 130.2 (q, *J* = 32.5 Hz), 126.9, 125.3 (q, *J* = 3.8 Hz), 124.2 (q, *J* = 272.1 Hz), 67.9, 54.6, 49.9, 25.8, 13.3.

¹⁹F NMR (471 MHz, CDCl₃) δ -62.58.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1763, 1696, 1372, 1320, 1287, 1161, 1119, 1067.

HRMS (ESI⁺) calcd. for C₁₅H₁₄F₃NO₂Na⁺ [M+Na]⁺: 320.0869, found: 320.0863.

2,2,2-Trichloroethyl 4-methyl-3-oxo-1-(4-(trifluoromethyl)phenyl)-2-azabicyclo[2.2.0]hexane-2-carboxylate (6e)



Prepared according to **General Procedure D** using **2e** (580 mg, 1.35 mmol, 1.00 equiv), CH₃CN (13.5 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (15.2 mg, 13.5 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/CH₂Cl₂ (2% to 5%), affording **6e** (375 mg, 0.87 mmol, 65 %) as a white solid.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 4:1 molar ratio of **6e/5e**.

M.P.: 117-119 °C

¹H NMR (500 MHz, CDCl₃) δ 7.65 (d, *J* = 8.2 Hz, 2H), 7.45 (d, *J* = 8.1 Hz, 2H), 4.77 (d, *J* = 11.9 Hz, 1H), 4.72 (d, *J* = 12.0 Hz, 1H), 3.12 (ddd, *J* = 13.5, 11.4, 6.3 Hz, 1H), 2.64 (ddd, *J* = 13.7, 8.7, 5.5 Hz, 1H), 2.56 (ddd, *J* = 13.1, 8.7, 6.2 Hz, 1H), 2.22 (ddd, *J* = 13.2, 11.5, 5.5 Hz, 1H), 1.13 (s, 3H).

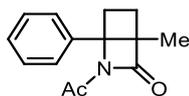
¹³C NMR (126 MHz, CDCl₃) δ 170.1, 147.0, 138.7, 130.4 (q, *J* = 32.6 Hz), 126.9, 125.8 (q, *J* = 3.8 Hz), 123.9 (q, *J* = 272.4 Hz), 94.2, 74.8, 66.6, 63.1, 26.1, 25.5, 13.0.

¹⁹F NMR (471 MHz, CDCl₃) δ -62.66.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1814, 1733, 1620, 1379, 1316, 1276, 1166, 1118, 1067, 1015.

HRMS (ESI⁺) calcd. for C₁₆H₁₄Cl₃F₃NO₃⁺ [M+H]⁺: 429.9986, found: 429.9986.

2-acetyl-4-methyl-1-phenyl-2-azabicyclo[2.2.0]hexan-3-one (6f)



Prepared according to **General Procedure D** using **2f** (540 mg, 2.36 mmol, 1.00 equiv), CH₃CN (23.6 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (26.5 mg, 23.6 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/CH₂Cl₂ (2% to 6%), affording **6f** (411 mg, 1.79 mmol, 76%) as a colourless oil and **5f** (26 mg, 0.11 mmol, 5 %) as a white solid.^{A,B}

Notes: (A) The crude ¹H NMR spectrum indicated a 10:1 molar ratio of **6f**/**5f**.

(B) When HFIP was used as the solvent, **6f** was isolated in 79% yield. The crude ¹H NMR spectrum indicated a 30:1 molar ratio of **6f**:**5f**.

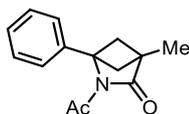
¹H NMR (500 MHz, CDCl₃) δ 7.41 – 7.34 (m, 2H), 7.31 – 7.27 (m, 1H), 7.27 – 7.24 (m, 2H), 3.08 – 2.99 (m, 1H), 2.58 – 2.46 (m, 5H), 2.24 – 2.17 (m, 1H), 1.09 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.6, 168.1, 134.9, 128.7, 127.9, 126.1, 66.2, 61.1, 26.5, 26.4, 24.5, 13.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1771, 1701, 1497, 1371, 1309, 1270, 1193, 1075, 970.

HRMS (ESI⁺) calcd. for C₁₄H₁₅NO₂Na⁺ [M+Na]⁺: 252.0995, found: 252.0986.

2-Acetyl-4-methyl-1-phenyl-2-azabicyclo[2.1.1]hexan-3-one (5f)



M.P.: 100-102 °C

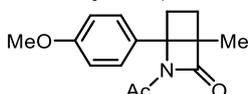
¹H NMR (500 MHz, CDCl₃) δ 7.39 – 7.31 (m, 3H), 7.29 – 7.26 (m, 2H), 2.73 (dd, *J* = 4.7, 2.1 Hz, 2H), 2.42 (dd, *J* = 4.6, 2.1 Hz, 2H), 2.38 (s, 3H), 1.38 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 179.9, 169.4, 137.1, 128.3, 128.1, 126.5, 68.7, 54.6, 49.7, 26.0, 13.3.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1766, 1681, 1449, 1374, 1359, 1297, 1163.

HRMS (ESI⁺) calcd. for C₁₄H₁₆NO₂⁺ [M+H]⁺: 230.1176, found: 230.1169.

2-Acetyl-1-(4-methoxyphenyl)-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (6g)



Gram scale: Prepared according to **General Procedure D** using **2g** (3.31 g, 12.8 mmol, 1.00 equiv), CH₃CN (128 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (71.7 mg, 63.9 μmol, 0.50 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (0% to 60%), affording **6g** (2.71 g, 10.5 mmol, 82%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 24:1 molar ratio of **6g/5g**.

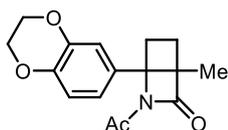
¹H NMR (500 MHz, CDCl₃) δ 7.17 (d, *J* = 8.8 Hz, 2H), 6.89 (d, *J* = 8.8 Hz, 2H), 3.76 (s, 3H), 2.97 (ddd, *J* = 12.9, 11.4, 6.0 Hz, 1H), 2.56 – 2.42 (m, 5H), 2.20 – 2.11 (m, 1H), 1.08 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.7, 168.0, 159.2, 127.3, 127.0, 114.0, 65.9, 60.8, 55.2, 26.4, 26.3, 24.4, 12.9.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1771, 1700, 1614, 1515, 1445, 1371, 1309, 1272, 1179, 1104, 1031.

HRMS (ESI⁺) calcd. for C₁₅H₁₇NO₃Na⁺ [M+Na]⁺: 282.1101, found: 282.1093.

2-Acetyl-1-(2,3-dihydrobenzo[*b*][1,4]dioxin-6-yl)-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (**6h**)



Prepared according to **General Procedure D** using **2h** (430 mg, 1.50 mmol, 1.00 equiv), CH₃CN (15.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (16.8 mg, 15.0 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of (Et₂O/ethyl acetate 3:1)/pentane (10% to 50%), affording **6h** (225 mg, 0.78 mmol, 52%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 15:1 molar ratio of **6h/5h**.

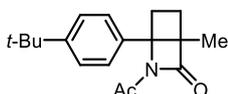
¹H NMR (500 MHz, CDCl₃) δ 6.83 (d, *J* = 8.4 Hz, 1H), 6.73 (d, *J* = 2.3 Hz, 1H), 6.70 (dd, *J* = 8.4, 2.2 Hz, 1H), 4.20 (s, 4H), 2.97 – 2.89 (m, 1H), 2.51 – 2.39 (m, 5H), 2.17 – 2.10 (m, 1H), 1.07 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.6, 168.0, 143.6, 143.3, 128.1, 119.1, 117.4, 115.2, 65.7, 64.31, 64.30, 60.9, 26.23, 26.21, 24.4, 12.9.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1770, 1701, 1588, 1510, 1425, 1372, 1310, 1276, 1261, 1194, 1124.

HRMS (ESI⁺) calcd. for C₁₆H₁₇NO₄Na⁺ [M+Na]⁺: 310.1050, found: 310.1039.

2-acetyl-1-(4-(*tert*-butyl)phenyl)-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (**6i**)



Prepared according to **General Procedure D** using **2i** (540 mg, 1.90 mmol, 1.00 equiv), CH₃CN (19.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (21.3 mg, 19.0 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (10% to 50%), affording **6i** (350 mg, 1.23 mmol, 65%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 13:1 molar ratio of **6i**:**5i**.

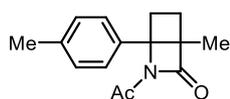
¹H NMR (500 MHz, CDCl₃) δ 7.38 (d, *J* = 8.5 Hz, 2H), 7.18 (d, *J* = 8.5 Hz, 2H), 3.01 (ddd, *J* = 12.6, 11.4, 5.8 Hz, 1H), 2.58 – 2.44 (m, 5H), 2.24 – 2.15 (m, 1H), 1.30 (s, 9H), 1.10 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.7, 168.1, 150.7, 131.8, 125.8, 125.6, 66.1, 61.0, 34.6, 31.3, 26.48, 26.46, 24.5, 13.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1774, 1703, 1371, 1311, 1277, 1191, 970.

HRMS (ESI⁺) calcd. for C₁₈H₂₃NO₂Na⁺ [M+Na]⁺: 308.1621, found: 308.1629.

2-Acetyl-4-methyl-1-(*p*-tolyl)-2-azabicyclo[2.2.0]hexan-3-one (**6j**)



Prepared according to **General Procedure D** using **2j** (302 mg, 1.24 mmol, 1.00 equiv), CH₃CN (12.4 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (13.9 mg, 12.4 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (0% to 60%), affording **6j** (206 mg, 0.85 mmol, 68%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 15:1 molar ratio of **6j**:**5j**.

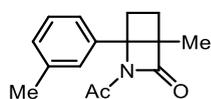
¹H NMR (500 MHz, CDCl₃) δ 7.19 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 8.2 Hz, 2H), 3.06 – 2.98 (m, 1H), 2.58 – 2.44 (m, 5H), 2.34 (s, 3H), 2.24 – 2.13 (m, 1H), 1.10 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.8, 168.1, 137.8, 132.0, 129.5, 126.1, 66.2, 61.0, 26.5, 26.4, 24.6, 21.2, 13.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1772, 1702, 1518, 1447, 1371, 1310, 1270, 1186, 1105, 1037.

HRMS (ESI⁺) calcd. for C₁₅H₁₇NO₂Na⁺ [M+Na]⁺: 266.1152, found: 266.1140.

2-Acetyl-4-methyl-1-(*m*-tolyl)-2-azabicyclo[2.2.0]hexan-3-one (**6k**)



Prepared according to **General Procedure D** using **2k** (365 mg, 1.50 mmol, 1.00 equiv), CH₃CN (15.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (16.9 mg, 15.1 mmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (0% to 60%), affording **6k** (272 mg, 1.12 mmol, 75%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 11:1 molar ratio of **6k/5k**.

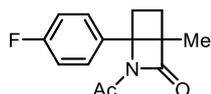
¹H NMR (500 MHz, CDCl₃) δ 7.29 – 7.25 (m, 1H), 7.11 (dp, *J* = 7.5, 0.9 Hz, 1H), 7.07 – 7.03 (m, 2H), 3.07 – 2.98 (m, 1H), 2.57 – 2.46 (m, 5H), 2.36 (s, 3H), 2.25 – 2.17 (m, 1H), 1.09 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.7, 168.1, 138.3, 134.9, 128.8, 128.6, 126.8, 123.2, 66.2, 61.0, 26.5, 26.3, 24.5, 21.7, 13.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1772, 1702, 1609, 1448, 1371, 1310, 1275, 1276, 1193, 1038.

HRMS (ESI⁺) calcd. for C₁₅H₁₇NO₂Na⁺ [M+Na]⁺: 266.1152, found: 266.1142.

2-Acetyl-1-(4-fluorophenyl)-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (**6l**)



Prepared according to **General Procedure D** using **2l** (305 mg, 1.23 mmol, 1.00 equiv), CH₃CN (12.3 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (13.9 mg, 12.4 mmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of (Et₂O/ethyl acetate 3:1)/pentane (10% to 40%), affording **6l** (208 mg, 0.84 mmol, 68%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 11:1 molar ratio of **6l/5l**.

¹H NMR (500 MHz, CDCl₃) δ 7.27 – 7.22 (m, 2H), 7.11 – 7.05 (m, 2H), 3.00 (ddd, *J* = 13.0, 11.5, 6.0 Hz, 1H), 2.60 – 2.45 (m, 5H), 2.26 – 2.18 (m, 1H), 1.11 (s, 3H).

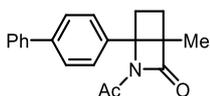
¹³C NMR (126 MHz, CDCl₃) δ 171.3, 168.1, 162.3 (d, *J* = 247.2 Hz), 130.9 (d, *J* = 3.2 Hz), 127.9 (d, *J* = 8.3 Hz), 115.6 (d, *J* = 21.8 Hz), 65.6, 61.0, 26.5, 26.4, 24.4, 12.9.

¹⁹F NMR (471 MHz, CDCl₃) δ -114.23.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1773, 1701, 1603, 1513, 1448, 1372, 1311, 1269, 1230, 1161, 1097.

HRMS (ESI⁺) calcd. for C₁₄H₁₄FNO₂Na⁺ [M+Na]⁺: 270.0901, found: 270.0893.

1-([1,1'-biphenyl]-4-yl)-2-acetyl-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (**6m**)



Prepared according to **General Procedure D** using **2m** (305 mg, 1.00 mmol, 1.00 equiv), CH₃CN (10.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.2 mg, 10.0 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of (Et₂O/ethyl acetate 3:1)/pentane (10% to 40%), affording **6m** (194 mg, 0.64 mmol, 64%) as a white solid.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 7:1 molar ratio of **6m/5m**.

M.P.: 77-79 °C

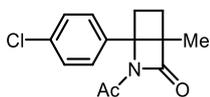
¹H NMR (500 MHz, CDCl₃) δ 7.62 (d, *J* = 8.4 Hz, 2H), 7.61 – 7.58 (m, 2H), 7.45 (t, *J* = 7.6 Hz, 2H), 7.38 – 7.33 (m, 3H), 3.08 (ddd, *J* = 12.8, 11.5, 5.9 Hz, 1H), 2.63 – 2.50 (m, 5H), 2.28 – 2.21 (m, 1H), 1.16 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.6, 168.2, 140.8, 140.5, 133.9, 128.9, 127.6, 127.4, 127.2, 126.6, 66.1, 61.2, 26.49, 26.47, 24.5, 13.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1773, 1702, 1487, 1371, 1311, 1276, 1191, 1104, 970.

HRMS (ESI⁺) calcd. for C₂₀H₁₉NO₂Na⁺ [M+Na]⁺: 328.1308, found: 328.1299.

2-acetyl-1-(4-chlorophenyl)-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (**6n**)



Prepared according to **General Procedure D** using **2n** (380 mg, 1.44 mmol, 1.00 equiv), CH₃CN (14.4 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (16.2 mg, 14.4 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (10% to 60%), affording **6n** (294 mg, 1.12 mmol, 77%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 7:1 molar ratio of **6n/5n**.

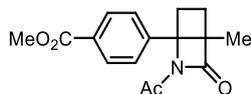
¹H NMR (500 MHz, CDCl₃) δ 7.32 (d, *J* = 8.6 Hz, 2H), 7.17 (d, *J* = 8.6 Hz, 2H), 2.99 – 2.91 (m, 1H), 2.54 – 2.42 (m, 5H), 2.21 – 2.14 (m, 1H), 1.06 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.1, 167.9, 133.8, 133.6, 128.8, 127.5, 65.5, 61.1, 26.29, 26.27, 24.3, 12.9.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1773, 1701, 1495, 1371, 1310, 1277, 1264, 1194, 1094.

HRMS (ESI⁺) calcd. for C₁₄H₁₄ClNO₂Na⁺ [M+Na]⁺: 286.0605, found: 286.0601.

Methyl 4-(2-acetyl-4-methyl-3-oxo-2-azabicyclo[2.2.0]hexan-1-yl)benzoate (**6o**)



Prepared according to **General Procedure D** using **2o** (550 mg, 1.92 mmol, 1.00 equiv), CH₃CN (19.2 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (21.5 mg, 19.2 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of CH₂Cl₂/ethyl acetate (0% to 10%), affording **6o** (345 mg, 1.20 mmol, 63%) as a colourless oil and **5o** (95 mg, 0.33 mmol, 17 %) as a white solid.^{A,B}

Notes: (A) The crude ¹H NMR spectrum indicated a 3:1 molar ratio of **6o/5o**.

(B) When HFIP was used as the solvent, **6o** was isolated in 71% yield. The crude ¹H NMR spectrum indicated a 8:1 molar ratio of **6o/5o**.

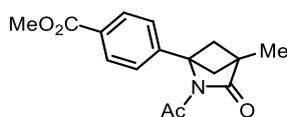
¹H NMR (500 MHz, CDCl₃) δ 7.99 (d, *J* = 8.5 Hz, 2H), 7.28 (d, *J* = 8.5 Hz, 2H), 3.84 (s, 3H), 3.04 – 2.93 (m, 1H), 2.54 – 2.42 (m, 5H), 2.24 – 2.15 (m, 1H), 1.02 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.8, 167.8, 166.4, 139.9, 129.8, 129.5, 126.1, 65.6, 61.4, 52.1, 26.3, 26.2, 24.3, 12.8.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1775, 1702, 1612, 1435, 1372, 1311, 1275, 1192, 1110.

HRMS (ESI⁺) calcd. for C₁₆H₁₇NO₄Na⁺ [M+Na]⁺: 310.1050, found: 310.1056.

Methyl 4-(2-acetyl-4-methyl-3-oxo-2-azabicyclo[2.1.1]hexan-1-yl)benzoate (**5o**)



M.P.: 141-143 °C

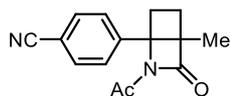
¹H NMR (500 MHz, CDCl₃) δ 8.01 (d, *J* = 8.5 Hz, 2H), 7.31 (d, *J* = 8.4 Hz, 2H), 3.89 (s, 3H), 2.71 (dd, *J* = 4.6, 2.1 Hz, 2H), 2.43 (dd, *J* = 4.6, 2.1 Hz, 2H), 2.37 (s, 3H), 1.36 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 179.3, 169.2, 166.7, 141.9, 129.6, 129.5, 126.4, 67.9, 54.4, 52.1, 49.7, 25.7, 13.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1758, 1721, 1692, 1368, 1275, 1262, 1180, 1099.

HRMS (ESI⁺) calcd. for C₁₆H₁₇NO₄Na⁺ [M+Na]⁺: 310.1050, found: 310.1040.

4-(2-Acetyl-4-methyl-3-oxo-2-azabicyclo[2.2.0]hexan-1-yl)benzonitrile (**6p**)



Prepared according to **General Procedure D** using **2p** (508 mg, 2.00 mmol, 1.00 equiv), CH₃CN (20.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (22.4 mg, 20.0 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/CH₂Cl₂ (0% to 10%), affording **6p** (272 mg, 1.07 mmol, 54%) as a white solid and **5p** (125 mg, 0.49 mmol, 25 %) as a white solid.^{A,B}

Notes: (A) The crude ¹H NMR spectrum indicated a 2:1 molar ratio of **6p**/**5p**.

(B) When HFIP was used as the solvent, **6p** was isolated in 64% yield. The crude ¹H NMR spectrum indicated a 6:1 molar ratio of **6p**:**5pp**.

M.P.: 79-81 °C

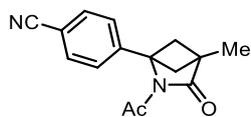
¹H NMR (500 MHz, CDCl₃) δ 7.64 (d, *J* = 8.5 Hz, 2H), 7.34 (d, *J* = 8.7 Hz, 2H), 3.02 – 2.92 (m, 1H), 2.55 – 2.47 (m, 2H), 2.45 (s, 3H), 2.27 – 2.19 (m, 1H), 1.05 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.4, 167.9, 140.3, 132.4, 126.9, 118.4, 111.6, 65.3, 61.6, 26.3, 26.2, 24.3, 12.9.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2228, 1775, 1701, 1609, 1372, 1310, 1275, 1266, 1190, 970.

HRMS (ESI⁺) calcd. for C₁₅H₁₄N₂O₂Na⁺ [M+Na]⁺: 277.0947, found: 277.0937.

4-(2-Acetyl-4-methyl-3-oxo-2-azabicyclo[2.1.1]hexan-1-yl)benzonitrile (**5p**)



M.P.: 141-143 °C

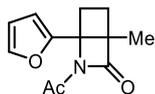
¹H NMR (500 MHz, CDCl₃) δ 7.62 (d, *J* = 8.4 Hz, 2H), 7.35 (d, *J* = 8.4 Hz, 2H), 2.71 (dd, *J* = 4.7, 2.1 Hz, 2H), 2.44 (dd, *J* = 4.6, 2.1 Hz, 2H), 2.37 (s, 3H), 1.37 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 178.9, 169.4, 142.2, 132.0, 127.2, 118.7, 111.7, 67.6, 54.3, 49.8, 25.6, 13.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2227, 1762, 1693, 1613, 1371, 1357, 1276, 1261, 1175.

HRMS (ESI⁺) calcd. for C₁₅H₁₅N₂O₂⁺ [M+H]⁺: 255.1128, found: 255.1120.

2-Acetyl-1-(furan-2-yl)-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (6q)



Prepared according to **General Procedure D** using **2q** (257 mg, 1.17 mmol, 1.00 equiv), CH₃CN (11.7 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (13.2 mg, 11.8 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (0% to 60%), affording **6q** (136 mg, 0.62 mmol, 53%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a >25:1 molar ratio of **6q**/**5q**.

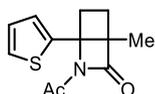
¹H NMR (500 MHz, CDCl₃) δ 7.35 (dd, *J* = 1.9, 0.9 Hz, 1H), 6.33 (dd, *J* = 3.3, 1.8 Hz, 1H), 6.28 (dd, *J* = 3.3, 0.9 Hz, 1H), 2.90 (ddd, *J* = 13.2, 11.4, 6.5 Hz, 1H), 2.48 (ddd, *J* = 13.5, 8.6, 5.4 Hz, 1H), 2.43 – 2.36 (m, 4H), 2.15 (ddd, *J* = 13.0, 11.4, 5.4 Hz, 1H), 1.17 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.7, 167.8, 148.9, 143.1, 110.7, 108.9, 61.6, 60.8, 25.80, 25.76, 24.2, 13.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1775, 1704, 1449, 1372, 1275, 1268, 1190, 970.

HRMS (ESI⁺) calcd. for C₁₂H₁₃NO₃Na⁺ [M+Na]⁺: 242.0788, found: 242.0785.

2-Acetyl-4-methyl-1-(thiophen-2-yl)-2-azabicyclo[2.2.0]hexan-3-one (6r)



Prepared according to **General Procedure D** using **2r** (310 mg, 1.32 mmol, 1.00 equiv), CH₃CN (13.2 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (14.8 mg, 13.2 μmol, 1 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (0% to 60%), affording **6r** (244 mg, 1.04 mmol, 79%) as a pale-yellow oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a >25:1 molar ratio of **6r**/**5r**.

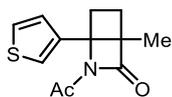
¹H NMR (500 MHz, CDCl₃) δ 7.25 (dd, *J* = 5.1, 1.3 Hz, 1H), 6.98 (dd, *J* = 5.0, 3.5 Hz, 1H), 6.94 (dd, *J* = 3.6, 1.3 Hz, 1H), 2.91 (ddd, *J* = 13.6, 11.4, 6.6 Hz, 1H), 2.63 (ddd, *J* = 13.9, 8.6, 5.6 Hz, 1H), 2.46 – 2.38 (m, 4H), 2.17 (ddd, *J* = 13.1, 11.4, 5.6 Hz, 1H), 1.13 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.8, 167.7, 139.0, 127.5, 125.7, 125.5, 63.4, 62.0, 28.3, 25.9, 24.3, 12.7.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1773, 1703, 1442, 1370, 1293, 1266, 1188, 962.

HRMS (ESI⁺) calcd. for C₁₂H₁₃NO₂SNa⁺ [M+Na]⁺: 258.0559, found: 258.0553.

2-Acetyl-4-methyl-1-(thiophen-3-yl)-2-azabicyclo[2.2.0]hexan-3-one (6s)



Prepared according to **General Procedure D** using **2s** (300 mg, 1.28 mmol, 1.00 equiv), CH₃CN (12.8 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (14.3 mg, 12.7 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/CH₂Cl₂ (0% to 10%), affording **6s** (210 mg, 0.89 mmol, 70%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 20:1 molar ratio of **6s**/**5s**.

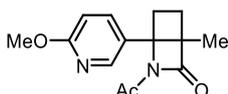
¹H NMR (500 MHz, CDCl₃) δ 7.32 (ddd, *J* = 5.2, 3.0, 1.3 Hz, 1H), 7.15 (dt, *J* = 3.1, 1.1 Hz, 1H), 6.94 (dt, *J* = 5.0, 1.1 Hz, 1H), 2.89 (ddd, *J* = 13.6, 11.4, 6.6 Hz, 1H), 2.59 (ddd, *J* = 13.7, 8.7, 5.7 Hz, 1H), 2.48 – 2.42 (m, 4H), 2.19 (ddd, *J* = 13.0, 10.7, 5.6 Hz, 1H), 1.12 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.4, 168.1, 137.0, 126.7, 125.5, 122.6, 63.7, 61.2, 27.8, 26.2, 24.4, 13.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1770, 1699, 1371, 1295, 1275, 1266, 1200, 1036.

HRMS (ESI⁺) calcd. for C₁₂H₁₃NO₂SNa⁺ [M+Na]⁺: 258.0559, found: 258.0551.

2-Acetyl-1-(6-methoxypyridin-3-yl)-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (6t)



Prepared according to **General Procedure D** using **2t** (370 mg, 1.42 mmol, 1.00 equiv), CH₃CN (14.2 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (16.0 mg, 14.3 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 40%), affording **6t** (275 mg, 1.06 mmol, 74%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 11:1 molar ratio of **6t**/**5t**.

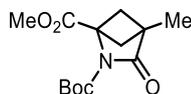
¹H NMR (500 MHz, CDCl₃) δ 8.01 (d, *J* = 2.6 Hz, 1H), 7.37 (dd, *J* = 8.7, 2.7 Hz, 1H), 6.65 (d, *J* = 8.6 Hz, 1H), 3.82 (s, 3H), 2.87 (ddd, *J* = 13.0, 11.4, 6.0 Hz, 1H), 2.50 – 2.34 (m, 5H), 2.11 (td, *J* = 11.8, 5.4 Hz, 1H), 1.03 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.9, 167.8, 163.6, 144.9, 136.4, 123.4, 110.7, 64.1, 61.0, 53.4, 26.19, 26.15, 24.2, 12.8.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1774, 1701, 1607, 1568, 1497, 1446, 1375, 1311, 1277, 1192, 1023.

HRMS (ESI⁺) calcd. for C₁₄H₁₇N₂O₃⁺ [M+H]⁺: 261.1234, found: 261.1224.

2-(*tert*-Butyl) 1-methyl 4-methyl-3-oxo-2-azabicyclo[2.1.1]hexane-1,2-dicarboxylate (5u)



Prepared according to **General Procedure D** using **2u** (1.05 g, 3.90 mmol, 1.00 equiv), CH₃CN (240 mL, 0.016 M) and benzophenone (177 mg, 0.97 mmol, 0.25 equiv).^A The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **5u** (830 mg, 3.09 mmol, 79%) as a white solid.

Notes: (A) 40 W Kessil KSPR160L lamp (370 nm, 100% intensity) was used.

M.P.: 77-79 °C

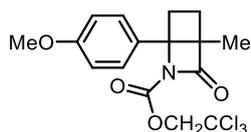
¹H NMR (500 MHz, CDCl₃) δ 3.76 (s, 3H), 2.46 (dd, *J* = 4.9, 1.8 Hz, 2H), 2.41 (dd, *J* = 4.9, 1.8 Hz, 2H), 1.45 (s, 9H), 1.26 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 176.7, 166.9, 149.5, 83.5, 63.8, 53.4, 52.4, 49.7, 28.0, 13.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1801, 1749, 1715, 1439, 1306, 1210, 1157, 1088, 1036.

HRMS (ESI⁺) calcd. for C₁₃H₁₉NO₅Na⁺ [M+Na]⁺: 292.1155, found: 292.1145.

2,2,2-Trichloroethyl 1-(4-methoxyphenyl)-4-methyl-3-oxo-2-azabicyclo[2.2.0]hexane-2-carboxylate (6v)



Prepared according to **General Procedure D** using **2v** (630 mg, 1.61 mmol, 1.00 equiv), CH₃CN (16.1 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (18.0 mg, 16.0 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 25%), affording **6v** (470 mg, 1.20 mmol, 75%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a >25:1 molar ratio of **6v**/**5v**.

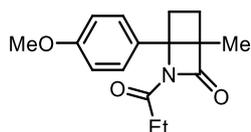
¹H NMR (500 MHz, CDCl₃) δ 7.21 (d, *J* = 8.8 Hz, 2H), 6.88 (d, *J* = 8.8 Hz, 2H), 4.77 (d, *J* = 12.0 Hz, 1H), 4.66 (d, *J* = 11.9 Hz, 1H), 3.76 (s, 3H), 3.05 (ddd, *J* = 13.5, 11.4, 6.4 Hz, 1H), 2.58 (ddd, *J* = 13.7, 8.7, 5.5 Hz, 1H), 2.48 (ddd, *J* = 13.0, 8.7, 6.4 Hz, 1H), 2.13 (ddd, *J* = 13.0, 11.4, 5.5 Hz, 1H), 1.10 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 170.9, 159.4, 147.0, 127.6, 126.4, 114.0, 94.3, 74.5, 66.9, 62.2, 55.2, 26.1, 25.4, 13.0.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1810, 1732, 1614, 1516, 1445, 1379, 1315, 1278, 1252, 1178, 1093.

HRMS (ESI⁺) calcd. for $\text{C}_{16}\text{H}_{16}\text{Cl}_3\text{NO}_4\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 414.0037, found: 414.0030.

1-(4-Methoxyphenyl)-4-methyl-2-propionyl-2-azabicyclo[2.2.0]hexan-3-one (6w)



Prepared according to **General Procedure D** using **2w** (280 mg, 1.03 mmol, 1.00 equiv), CH_3CN (10.3 mL), and $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbbpy})](\text{PF}_6)$ (11.5 mg, 0.010 mmol, 1 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 25%), affording **6w** (232 mg, 0.85 mmol, 83%) as a colourless oil.^A

Notes: (A) The crude ^1H NMR spectrum indicated a 20:1 molar ratio of **6w**/**5w**.

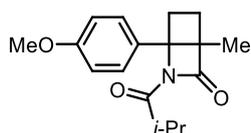
^1H NMR (500 MHz, CDCl_3) δ 7.16 (d, J = 8.9 Hz, 2H), 6.89 (d, J = 8.8 Hz, 2H), 3.76 (s, 3H), 2.97 (ddd, J = 12.9, 11.4, 6.0 Hz, 1H), 2.88 – 2.72 (m, 2H), 2.50 (ddd, J = 12.8, 8.7, 5.3 Hz, 1H), 2.44 (ddd, J = 12.8, 8.7, 6.0 Hz, 1H), 2.15 (ddd, J = 12.8, 11.4, 5.3 Hz, 1H), 1.17 (t, J = 7.5 Hz, 3H), 1.08 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 172.0, 171.5, 159.2, 127.3, 127.1, 114.0, 65.5, 60.6, 55.2, 30.5, 26.4, 26.3, 12.9, 8.0.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1772, 1699, 1614, 1515, 1461, 1371, 1249, 1178, 1031.

HRMS (ESI⁺) calcd. for $\text{C}_{16}\text{H}_{19}\text{NO}_3\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 296.1257, found: 296.1271.

2-Isobutyryl-1-(4-methoxyphenyl)-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (6x)



Prepared according to **General Procedure D** using **2x** (275 mg, 0.96 mmol, 1.00 equiv), CH_3CN (9.6 mL), and $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbbpy})](\text{PF}_6)$ (10.8 mg, 9.6 μmol , 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **6x** (183 mg, 0.64 mmol, 67%) as a colourless oil.^A

Notes: (A) The crude ^1H NMR spectrum indicated an 18:1 molar ratio of **6x**/**5x**.

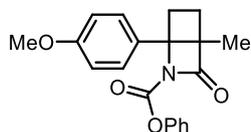
^1H NMR (500 MHz, CDCl_3) δ 7.15 (d, $J = 8.8$ Hz, 2H), 6.89 (d, $J = 8.8$ Hz, 2H), 3.77 (s, 3H), 3.34 (hept, $J = 6.9$ Hz, 1H), 3.04 – 2.91 (m, 1H), 2.52 – 2.41 (m, 2H), 2.21 – 2.12 (m, 1H), 1.25 (d, $J = 7.0$ Hz, 3H), 1.17 (d, $J = 6.8$ Hz, 3H), 1.09 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 175.3, 171.2, 159.2, 127.3, 127.2, 114.1, 65.4, 60.4, 55.3, 35.1, 26.5, 26.3, 19.1, 17.4, 13.0.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1773, 1698, 1614, 1516, 1462, 1386, 1248, 1179, 1032.

HRMS (ESI $^+$) calcd. for $\text{C}_{17}\text{H}_{21}\text{NO}_3\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 310.1414, found: 310.1427.

Phenyl 1-(4-methoxyphenyl)-4-methyl-3-oxo-2-azabicyclo[2.2.0]hexane-2-carboxylate (**6y**)



Prepared according to **General Procedure D** using **2y** (337 mg, 1.00 mmol, 1.00 equiv), CH_3CN (10.0 mL), and $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbbpy})](\text{PF}_6)$ (11.2 mg, 10.0 μmol , 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **6y** (232 mg, 0.69 mmol, 69%) as a white solid.^A

Notes: (A) The crude ^1H NMR spectrum indicated a 16:1 molar ratio of **6y**/**5y**.

M.P.: 143-145 $^\circ\text{C}$

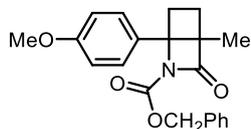
^1H NMR (500 MHz, CDCl_3) δ 7.34 – 7.28 (m, 4H), 7.18 (t, $J = 7.4$ Hz, 1H), 7.11 (d, $J = 8.1$ Hz, 2H), 6.95 (d, $J = 8.9$ Hz, 2H), 3.78 (s, 3H), 3.07 (ddd, $J = 13.6, 11.4, 6.3$ Hz, 1H), 2.66 (ddd, $J = 13.8, 8.7, 5.6$ Hz, 1H), 2.56 (ddd, $J = 13.0, 8.7, 6.3$ Hz, 1H), 2.18 (ddd, $J = 13.0, 11.4, 5.6$ Hz, 1H), 1.15 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 170.8, 159.3, 149.9, 147.0, 129.3, 127.5, 126.7, 126.1, 121.2, 114.1, 66.7, 61.8, 55.2, 26.4, 25.5, 13.0.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1806, 1737, 1614, 1516, 1320, 1276, 1253, 1180, 1024.

HRMS (ESI $^+$) calcd. for $\text{C}_{20}\text{H}_{19}\text{NO}_4\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 360.1206, found: 360.1222.

Benzyl 1-(4-methoxyphenyl)-4-methyl-3-oxo-2-azabicyclo[2.2.0]hexane-2-carboxylate (**6z**)



Prepared according to **General Procedure D** using **2z** (365 mg, 1.04 mmol, 1.00 equiv), CH₃CN (10.4 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.7 mg, 10.4 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 25%), affording **6z** (191 mg, 0.54 mmol, 52%) as a white solid.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 10:1 molar ratio of **6z**/**5z**.

M.P.: 137-139 °C

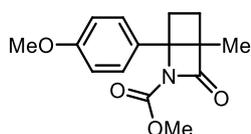
¹H NMR (500 MHz, CDCl₃) δ 7.31 – 7.27 (m, 3H), 7.24 – 7.19 (m, 4H), 6.90 (d, *J* = 8.9 Hz, 2H), 5.23 (d, *J* = 12.5 Hz, 1H), 5.18 (d, *J* = 12.5 Hz, 1H), 3.80 (s, 3H), 3.06 – 2.96 (m, 1H), 2.55 – 2.43 (m, 2H), 2.15 – 2.08 (m, 1H), 1.10 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.9, 159.3, 148.8, 135.1, 128.4, 128.2, 127.8, 127.5, 126.9, 114.0, 67.6, 66.4, 61.6, 55.2, 26.3, 25.5, 13.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1803, 1713, 1613, 1516, 1455, 1387, 1314, 1283, 1252, 1207, 1180.

HRMS (ESI⁺) calcd. for C₂₁H₂₁NO₄Na⁺ [M+Na]⁺: 374.1363, found: 374.1382.

Methyl 1-(4-methoxyphenyl)-4-methyl-3-oxo-2-azabicyclo[2.2.0]hexane-2-carboxylate (6aa**)**



Prepared according to **General Procedure D** using **2aa** (285 mg, 1.04 mmol, 1.00 equiv), CH₃CN (10.4 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.7 mg, 10.4 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 25%), affording **6aa** (173 mg, 0.63 mmol, 61%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 10:1 molar ratio of **6aa**/**5aa**.

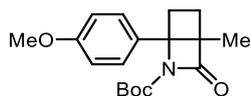
¹H NMR (500 MHz, CDCl₃) δ 7.20 (d, *J* = 8.8 Hz, 2H), 6.90 (d, *J* = 8.8 Hz, 2H), 3.78 (s, 3H), 3.77 (s, 3H), 2.99 (ddd, *J* = 13.1, 11.4, 6.0 Hz, 1H), 2.51 (ddd, *J* = 13.3, 8.7, 5.4 Hz, 1H), 2.45 (ddd, *J* = 12.9, 8.7, 6.1 Hz, 1H), 2.12 (ddd, *J* = 13.0, 11.5, 5.4 Hz, 1H), 1.07 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.9, 159.3, 149.6, 127.5, 126.9, 114.1, 66.2, 61.4, 55.2, 53.3, 26.3, 25.7, 13.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1790, 1729, 1614, 1516, 1438, 1320, 1278, 1252, 1178, 1030.

HRMS (ESI⁺) calcd. for C₁₅H₁₇NO₄Na⁺ [M+Na]⁺: 298.1050, found: 298.1064.

***tert*-Butyl 1-(4-methoxyphenyl)-4-methyl-3-oxo-2-azabicyclo[2.2.0]hexane-2-carboxylate (6ab)**



Prepared according to **General Procedure D** using **2ab** (2.63 g, 8.30 mmol, 1.00 equiv), CH₃CN (83.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (46.6 mg, 41.5 μmol, 0.50 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **6ab** (1.09 g, 3.44 mmol, 41%) as a colourless oil and **5ab** (240 mg, 0.76 mmol, 9%) as a white solid.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 4.5:1 molar ratio of **6ab**/**5ab**.

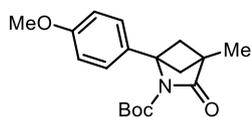
¹H NMR (500 MHz, CDCl₃) δ 7.17 (d, *J* = 8.9 Hz, 2H), 6.88 (d, *J* = 8.8 Hz, 2H), 3.77 (s, 3H), 3.01 – 2.91 (m, 1H), 2.49 – 2.39 (m, 2H), 2.11 – 2.03 (m, 1H), 1.36 (s, 9H), 1.04 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.4, 159.1, 147.7, 127.5, 127.4, 113.8, 82.9, 66.2, 61.1, 55.2, 27.9, 26.3, 25.5, 13.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1796, 1718, 1614, 1516, 1457, 1370, 1321, 1251, 1152, 1035.

HRMS (ESI⁺) calcd. for C₁₈H₂₃NO₄Na⁺ [M+Na]⁺: 340.1519, found: 340.1507.

***tert*-Butyl 1-(4-methoxyphenyl)-4-methyl-3-oxo-2-azabicyclo[2.1.1]hexane-2-carboxylate (5ab)**



M.P.: 96-98 °C

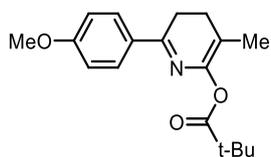
¹H NMR (500 MHz, CDCl₃) δ 7.14 (d, *J* = 8.8 Hz, 2H), 6.86 (d, *J* = 8.8 Hz, 2H), 3.78 (s, 3H), 2.68 (dd, *J* = 4.6, 2.1 Hz, 2H), 2.34 (dd, *J* = 4.6, 2.1 Hz, 2H), 1.31 (s, 3H), 1.14 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 179.1, 159.4, 149.7, 129.5, 127.9, 113.6, 81.7, 68.4, 55.3, 54.9, 49.6, 27.7, 13.2.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1786, 1722, 1702, 1615, 1519, 1455, 1360, 1291, 1246, 1157, 1032.

HRMS (ESI⁺) calcd. for C₁₈H₂₃NO₄Na⁺ [M+Na]⁺: 340.1519, found: 340.1508.

6-(4-Methoxyphenyl)-3-methyl-4,5-dihydropyridin-2-yl pivalate (9)



Prepared according to **General Procedure D** using **2ac** (315 mg, 1.05 mmol, 1.00 equiv), CH₃CN (10.5 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (11.7 mg, 10.4 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 25%), affording **9** (193 mg, 0.64 mmol, 61%) as a colourless oil.

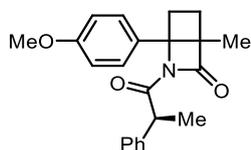
¹H NMR (500 MHz, CD₂Cl₂) δ 7.87 (d, *J* = 8.9 Hz, 2H), 6.93 (d, *J* = 8.9 Hz, 2H), 3.84 (s, 3H), 2.77 (t, *J* = 9.8 Hz, 2H), 2.36-2.28 (m, 2H), 1.73 (s, 3H), 1.36 (s, 9H).

¹³C NMR (126 MHz, CD₂Cl₂) δ 176.8, 162.7, 162.0, 145.9, 130.9, 128.8, 114.0, 108.3, 55.7, 39.2, 27.4, 26.1, 24.3, 15.6.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1745, 1607, 1512, 1461, 1276, 1259, 1116, 1029.

HRMS (ESI⁺) calcd. for C₁₈H₂₄NO₃⁺ [M+H]⁺: 302.1751, found: 302.1743.

(1*S*,4*S*)-1-(4-methoxyphenyl)-4-methyl-2-((*S*)-2-phenylpropanoyl)-2-azabicyclo[2.2.0] hexan-3-one & (1*R*,4*R*)-1-(4-methoxyphenyl)-4-methyl-2-((*S*)-2-phenylpropanoyl)-2-azabicyclo[2.2.0]hexan-3-one (6ad** and **6ad'**)**



Diastereoisomer 1

Prepared according to **General Procedure D** using **2ad** (151 mg, 0.43 mmol, 1.00 equiv), CH₃CN (4.3 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (4.8 mg, 4.3 μmol, 1 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 25%) and CH₂Cl₂/pentane (0% to 100%), affording **6ad** (41 mg, 0.12 mmol, 27%) as a colourless oil and **6ad'** (40.3 mg, 0.12 mmol, 27%) as a colourless oil.^A

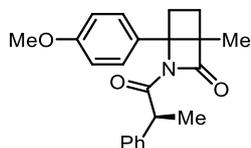
Notes: (A) The crude ¹H NMR spectrum indicated an 18:1 molar ratio of (**6ad**+**6ad'**)/(**5ab**+**5ad'**). The specific identity of the diastereomers could not be confirmed so no assignment has been made.

¹H NMR (500 MHz, CDCl₃) δ 7.42 – 7.39 (m, 2H), 7.35 – 7.31 (m, 2H), 7.29 – 7.25 (m, 1H), 7.20 (d, *J* = 8.8 Hz, 2H), 6.92 (d, *J* = 8.7 Hz, 2H), 4.64 (q, *J* = 7.0 Hz, 1H), 3.82 (s, 3H), 2.87 (dt, *J* = 13.5, 8.7 Hz, 1H), 2.20 (ddd, *J* = 13.7, 7.7, 6.4 Hz, 1H), 2.01-1.96 (m, 2H), 1.52 (d, *J* = 7.1 Hz, 3H), 1.07 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 172.4, 171.5, 159.3, 139.7, 128.7, 128.2, 127.5, 127.4, 114.2, 66.0, 60.6, 55.4, 45.9, 26.4, 26.1, 18.9, 12.9.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1777, 1700, 1517, 1454, 1358, 1275, 1260, 1179.

HRMS (ESI⁺) calcd. for $\text{C}_{22}\text{H}_{23}\text{NO}_3\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 372.1570, found: 372.1568.



Diastereoisomer 2

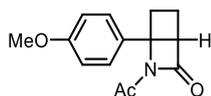
^1H NMR (500 MHz, CDCl_3) δ 7.39 – 7.27 (m, 5H), 6.80 (d, J = 8.9 Hz, 2H), 6.74 (d, J = 8.8 Hz, 2H), 4.60 (q, J = 7.3 Hz, 1H), 3.76 (s, 3H), 3.02 (ddd, J = 13.0, 11.4, 6.1 Hz, 1H), 2.56 (dt, J = 12.8, 6.9 Hz, 1H), 2.49 (ddd, J = 12.7, 8.6, 6.1 Hz, 1H), 2.14 (ddd, J = 12.7, 11.4, 5.4 Hz, 1H), 1.48 (d, J = 7.0 Hz, 3H), 0.97 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 172.0, 171.1, 159.2, 140.4, 128.8, 128.2, 127.4, 127.2, 126.9, 114.0, 65.6, 60.7, 55.3, 46.6, 26.5, 26.0, 17.9, 13.0.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1775, 1701, 1516, 1453, 1356, 1275, 1260, 1179.

HRMS (ESI⁺) calcd. for $\text{C}_{22}\text{H}_{23}\text{NO}_3\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 372.1570, found: 372.1566.

2-Acetyl-1-(4-methoxyphenyl)-2-azabicyclo[2.2.0]hexan-3-one (6ae)



Prepared according to **General Procedure D** using **2ae** (220 mg, 0.90 mmol, 1.00 equiv), CH_3CN (9.0 mL), and $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbbpy})](\text{PF}_6)$ (10.0 mg, 9.0 μmol , 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **6ae** (120 mg, 0.49 mmol, 55%) as a colourless oil and **5ae** (17 mg, 0.07 mmol, 8%) as a white solid.^A

Notes: (A) The crude ^1H NMR spectrum indicated a 5:1 molar ratio of **6ae**/**5ae**.

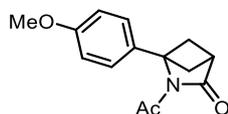
^1H NMR (500 MHz, CDCl_3) δ 7.28 (d, J = 8.8 Hz, 2H), 6.87 (d, J = 8.8 Hz, 2H), 3.76 (s, 3H), 3.61 (dd, J = 9.4, 2.9 Hz, 1H), 2.90 (ddd, J = 13.3, 11.4, 6.6 Hz, 1H), 2.68 (ddd, J = 13.3, 8.6, 5.8 Hz, 1H), 2.63 – 2.53 (m, 1H), 2.39 (s, 3H), 2.26 (dddd, J = 13.1, 8.6, 6.6, 2.9 Hz, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 168.7, 167.9, 159.3, 129.0, 127.4, 113.8, 63.1, 55.2, 54.3, 28.9, 24.3, 18.6.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1771, 1699, 1613, 1516, 1463, 1371, 1288, 1250, 1211, 1178, 1032.

HRMS (ESI⁺) calcd. for C₁₄H₁₅NO₃Na⁺ [M+Na]⁺: 268.0944, found: 268.0937.

2-Acetyl-1-(4-methoxyphenyl)-2-azabicyclo[2.1.1]hexan-3-one (**5ae**)



M.P.: 86-88 °C

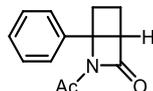
¹H NMR (500 MHz, CDCl₃) δ 7.19 (d, *J* = 8.7 Hz, 2H), 6.89 (d, *J* = 8.7 Hz, 2H), 3.81 (s, 3H), 3.02 (t, *J* = 2.7 Hz, 1H), 2.76-2.71 (m, 2H), 2.61-2.55 (m, 2H), 2.38 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 178.8, 169.6, 159.5, 129.3, 127.7, 113.7, 72.7, 55.3, 50.6, 44.4, 26.3.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1759, 1693, 1614, 1519, 1369, 1348, 1289, 1246, 1177, 1126, 1036.

HRMS (ESI⁺) calcd. for C₁₄H₁₅NO₃Na⁺ [M+Na]⁺: 268.0944, found: 268.09443.

2-Acetyl-1-phenyl-2-azabicyclo[2.2.0]hexan-3-one (**6af**)



Prepared according to **General Procedure D** using **2af** (150 mg, 0.70 mmol, 1.00 equiv), CH₃CN (7.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (7.8 mg, 7.0 μ mol, 1 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **6af** (77 mg, 0.36 mmol, 51%) as a colourless oil and **5af** (38 mg, 0.18 mmol, 25%) as a white solid.^{A,B}

Notes: (A) The crude ¹H NMR spectrum indicated a 2:1 molar ratio of **6af**/**5af**.

(B) When HFIP was used as the solvent, **6af** was isolated in 58% yield. The crude ¹H NMR spectrum indicated a 5:1 molar ratio of **6af**:**5af**.

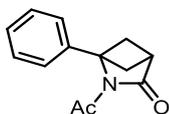
¹H NMR (500 MHz, CDCl₃) δ 7.374 (s, 2H), 7.366 (s, 2H), 7.34 – 7.26 (m, 1H), 3.66 (dd, *J* = 9.5, 2.9 Hz, 1H), 2.97 (ddd, *J* = 13.3, 11.4, 6.6 Hz, 1H), 2.72 (ddd, *J* = 13.4, 8.6, 5.8 Hz, 1H), 2.63 (dddd, *J* = 13.1, 11.4, 9.5, 5.8 Hz, 1H), 2.43 (s, 3H), 2.31 (dddd, *J* = 13.1, 8.6, 6.6, 2.9 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 168.6, 168.0, 137.0, 128.6, 128.2, 126.2, 63.4, 54.6, 28.8, 24.4, 18.8.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1773, 1700, 1371, 1285, 1210, 1183, 997.

HRMS (ESI⁺) calcd. for C₁₃H₁₃NO₂Na⁺ [M+Na]⁺: 238.0838, found: 238.0839.

2-Acetyl-1-phenyl-2-azabicyclo[2.1.1]hexan-3-one (5af)



M.P.: 94-96 °C

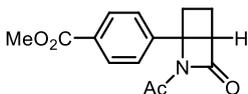
¹H NMR (500 MHz, CDCl₃) δ 7.39 – 7.32 (m, 3H), 7.29 – 7.27 (m, 2H), 3.04 (t, *J* = 2.7 Hz, 1H), 2.78-2.74 (m, 2H), 2.63-2.60 (m, 2H), 2.39 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 178.7, 169.5, 137.1, 128.3, 128.2, 126.3, 72.7, 50.6, 44.4, 26.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1760, 1678, 1450, 1375, 1305, 1198, 1127, 1088, 985.

HRMS (ESI⁺) calcd. for C₁₃H₁₃NO₂Na⁺ [M+Na]⁺: 238.0838, found: 238.0837.

Methyl 4-(2-acetyl-3-oxo-2-azabicyclo[2.2.0]hexan-1-yl)benzoate (6ag)



Prepared according to **General Procedure D** using **2ag** (185 mg, 0.68 mmol, 1.00 equiv), CH₃CN (6.8 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (7.6 mg, 6.8 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 40%), affording **6ag** as a colourless oil (43 mg, 0.16 mmol, 23%) and **5ag** (92 mg, 0.34 mmol, 50%) as a white solid.^{A,B}

Notes: (A) The crude ¹H NMR spectrum indicated a 1:2 molar ratio of **6ag/5ag**.

(B) When HFIP was used as the solvent, **6ag** was isolated in 32% yield. The crude ¹H NMR spectrum indicated a 1.3:1 molar ratio of **6ag/5ag**.

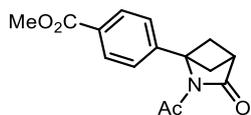
¹H NMR (500 MHz, CDCl₃) δ 8.02 (d, *J* = 8.4 Hz, 2H), 7.42 (d, *J* = 8.5 Hz, 2H), 3.90 (s, 3H), 3.68 (dd, *J* = 9.5, 2.9 Hz, 1H), 3.02 – 2.94 (m, 1H), 2.75 – 2.62 (m, 2H), 2.43 (s, 3H), 2.37 – 2.29 (m, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 168.2, 168.0, 166.6, 141.9, 129.94, 129.91, 126.3, 63.0, 54.9, 52.3, 28.8, 24.4, 18.9.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1778, 1702, 1613, 1435, 1409, 1372, 1275, 1185, 1108, 1019.

HRMS (ESI⁺) calcd. for C₁₅H₁₅NO₄Na⁺ [M+Na]⁺: 296.0893, found: 296.0891.

Methyl 4-(2-acetyl-3-oxo-2-azabicyclo[2.1.1]hexan-1-yl)benzoate (5ag)



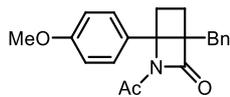
M.P.: 107-109 °C

¹H NMR (500 MHz, CDCl₃) δ 8.01 (d, *J* = 8.4 Hz, 2H), 7.31 (d, *J* = 8.4 Hz, 2H), 3.89 (s, 3H), 3.03 (t, *J* = 2.8 Hz, 1H), 2.76-2.72 (m, 2H), 2.64-2.58 (m, 2H), 2.36 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 178.1, 169.4, 166.6, 141.9, 129.7, 129.5, 126.2, 71.9, 52.2, 50.4, 44.3, 25.8.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1768, 1696, 1613, 1436, 1371, 1347, 1276, 1195, 1113. **HRMS** (ESI⁺) calcd. for C₁₅H₁₅NO₄Na⁺ [M+Na]⁺: 296.0893, found: 296.0893.

2-Acetyl-4-benzyl-1-(4-methoxyphenyl)-2-azabicyclo[2.2.0]hexan-3-one (6ah)



Prepared according to **General Procedure D** using **2ah** (595 mg, 1.78 mmol, 1.00 equiv), CH₃CN (17.8 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (19.9 mg, 17.7 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **6ah** (422 mg, 1.26 mmol, 71%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a >25:1 molar ratio of **6ah/5ah**.

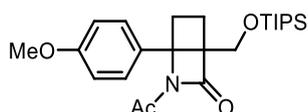
¹H NMR (500 MHz, CDCl₃) δ 7.24 – 7.19 (m, 5H), 7.00 – 6.95 (m, 4H), 3.86 (s, 3H), 2.97 (ddd, *J* = 13.6, 11.3, 6.6 Hz, 1H), 2.93 (d, *J* = 14.8 Hz, 1H), 2.77 (d, *J* = 15.0 Hz, 1H), 2.55 – 2.48 (m, 4H), 2.36 – 2.23 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 171.1, 168.0, 159.4, 135.4, 129.7, 128.3, 127.6, 126.8, 126.5, 114.2, 66.1, 64.4, 55.4, 33.8, 26.1, 24.5, 23.6.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1773, 1702, 1614, 1515, 1454, 1371, 1276, 1253, 1179, 1029.

HRMS (ESI⁺) calcd. for C₂₁H₂₁NO₃Na⁺ [M+Na]⁺: 358.1414, found: 358.1410.

2-acetyl-1-(4-methoxyphenyl)-4-(((triisopropylsilyl)oxy)methyl)-2-azabicyclo[2.2.0] hexan-3-one (6ai)



Prepared according to **General Procedure D** using **2ai** (530 mg, 1.23 mmol, 1.00 equiv), CH₃CN (12.3 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (13.8 mg, 12.3 μmol, 1.0 mol%). The crude reaction mixture was purified by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (0% to 50%) and ethyl acetate/CH₂Cl₂ (0% to 10%), affording **6ai** (372 mg, 0.86 mmol, 70%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a >25:1 molar ratio of **6ai/5ai**.

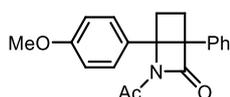
¹H NMR (500 MHz, CDCl₃) δ 7.27 (d, *J* = 8.8 Hz, 2H), 6.86 (d, *J* = 8.8 Hz, 2H), 3.84 (d, *J* = 11.6 Hz, 1H), 3.78 (d, *J* = 11.6 Hz, 1H), 3.76 (s, 3H), 3.01 – 2.92 (m, 1H), 2.58 – 2.44 (m, 5H), 2.28 – 2.21 (m, 1H), 0.98 – 0.86 (m, 21H).

¹³C NMR (126 MHz, CDCl₃) δ 169.6, 168.0, 159.3, 127.8, 127.1, 113.9, 66.9, 66.5, 58.5, 55.4, 27.1, 24.6, 20.1, 17.84, 17.79, 11.7.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2942, 2865, 1777, 1706, 1615, 1516, 1462, 1371, 1307, 1276, 1179.

HRMS (ESI⁺) calcd. for C₂₄H₃₈NO₄Si⁺ [M+H]⁺: 432.2565, found: 432.2586.

2-Acetyl-1-(4-methoxyphenyl)-4-phenyl-2-azabicyclo[2.2.0]hexan-3-one (**6aj**)



Prepared according to **General Procedure D** using **2aj** (500 mg, 1.56 mmol, 1.00 equiv), CH₃CN (15.6 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (17.5 mg, 15.6 μmol, 1.0 mol%). The crude reaction mixture was purified by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 40%) and ethyl acetate/CH₂Cl₂ (0% to 10%), affording **6aj** (168 mg, 0.52 mmol, 34%) as a pale-yellow oil and **5aj** (211 mg, 0.66 mmol, 42%) as a pale-yellow solid.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 1:1.2 molar ratio of **6aj/5aj**.

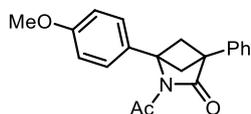
¹H NMR (500 MHz, CDCl₃) δ 7.25 – 7.16 (m, 5H), 7.06 (d, *J* = 8.8 Hz, 2H), 6.77 (d, *J* = 8.8 Hz, 2H), 3.72 (s, 3H), 3.18 (ddd, *J* = 13.1, 10.0, 7.7 Hz, 1H), 2.82 – 2.77 (m, 2H), 2.72 (ddd, *J* = 13.1, 8.3, 5.6 Hz, 1H), 2.53 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 169.8, 168.2, 159.2, 133.6, 128.6, 127.9, 127.8, 126.9, 126.7, 113.8, 68.6, 67.7, 55.2, 27.6, 27.1, 24.7.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1774, 1707, 1612, 1516, 1448, 1372, 1276, 1259, 1179.

HRMS (ESI⁺) calcd. for C₂₀H₂₀NO₃⁺ [M+H]⁺: 322.1438, found: 322.1434.

2-acetyl-1-(4-methoxyphenyl)-4-phenyl-2-azabicyclo[2.1.1]hexan-3-one (5aj)



M.P.: 134-136 °C

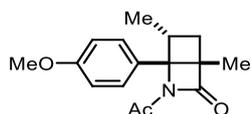
¹H NMR (500 MHz, CDCl₃) δ 7.45 (t, J = 7.4 Hz, 2H), 7.40 – 7.36 (m, 1H), 7.34 – 7.29 (m, 4H), 6.95 (d, J = 8.7 Hz, 2H), 3.84 (s, 3H), 3.17 (dd, J = 4.2, 2.5 Hz, 2H), 2.79 (dd, J = 4.2, 2.3 Hz, 2H), 2.45 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 177.6, 169.4, 159.5, 134.5, 128.8, 128.6, 128.0, 127.8, 127.0, 113.7, 67.6, 56.2, 55.3, 54.0, 26.2.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1766, 1707, 1612, 1516, 1448, 1372, 1276, 1259, 1179.

HRMS (ESI⁺) calcd. for C₂₀H₁₉NO₃Na⁺ [M+Na]⁺: 344.1257, found: 344.1254.

(±)-(1*S*,4*S*,6*R*)-2-acetyl-1-(4-methoxyphenyl)-4,6-dimethyl-2-azabicyclo[2.2.0]hexan-3-one (6ak)



Prepared according to **General Procedure D** using **2ak** (510 mg, 1.87 mmol, 1.00 equiv), CH₃CN (18.7 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (21.0 mg, 18.7 μ mol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of pentane/ethyl acetate (0% to 35%), affording **6ak** (340 mg, 1.25 mmol, 67%) as a colourless oil.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 7:1 molar ratio of **6ak**/**5ak**.

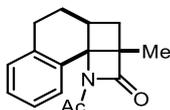
¹H NMR (500 MHz, CDCl₃) δ 7.18 (d, J = 8.8 Hz, 2H), 6.90 (d, J = 8.8 Hz, 2H), 3.77 (s, 3H), 3.47 (dp, J = 10.5, 7.1 Hz, 1H), 2.46 (s, 3H), 2.21 (dd, J = 12.9, 10.6 Hz, 1H), 1.99 (dd, J = 12.9, 7.2 Hz, 1H), 1.17 (d, J = 7.0 Hz, 3H), 1.06 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 172.2, 168.3, 159.2, 127.3, 127.0, 114.1, 68.9, 57.5, 55.2, 33.1, 32.7, 24.5, 16.8, 12.4.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1771, 1704, 1612, 1515, 1445, 1371, 1275, 1179, 1033.

HRMS (ESI⁺) calcd. for C₁₆H₁₉NO₃Na⁺ [M+Na]⁺: 296.1257, found: 296.1246.

(±)-(2a*S*,3a*S*,9b*S*)-1-acetyl-2a-methyl-3,3a,4,5-tetrahydro-1*H*-naphtho[1',2':1,4] cyclobuta[1,2-b]azet-2(2a*H*)-one (6al)



Prepared according to **General Procedure D** using **2al** (465 mg, 1.82 mmol, 1.00 equiv), CH₃CN (18.2 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (20.0 mg, 17.8 μmol, 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (0% to 40%), affording **6al** (246 mg, 0.96 mmol, 53%) as a colourless oil and **5al** (78 mg, 0.31 mmol, 17%) as a white solid.^A

Notes: (A) The crude ¹H NMR spectrum indicated a 3:1 molar ratio of **6al**/**5al**.

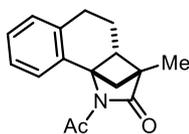
¹H NMR (500 MHz, CDCl₃) δ 7.22 – 7.17 (m, 3H), 7.09 – 7.05 (m, 1H), 2.93 – 2.87 (m, 1H), 2.85 (ddd, *J* = 15.6, 6.8, 3.7 Hz, 1H), 2.74 – 2.66 (m, 1H), 2.64 (dd, *J* = 13.2, 8.1 Hz, 1H), 2.47 (s, 3H), 2.02 (dddd, *J* = 13.5, 10.2, 6.7, 3.8 Hz, 1H), 1.84 (dd, *J* = 13.3, 5.5 Hz, 1H), 1.79 (dddd, *J* = 13.4, 6.8, 5.4, 3.9 Hz, 1H), 1.05 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.7, 167.8, 139.3, 130.3, 128.5, 127.7, 126.7, 124.9, 64.5, 60.2, 36.1, 31.9, 28.0, 27.4, 24.4, 13.2.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1774, 1701, 1457, 1371, 1308, 1276, 1199, 962.

HRMS (ESI⁺) calcd. for C₁₆H₁₇NO₂Na⁺ [M+Na]⁺: 278.1152, found: 278.1151.

(±)-(3*R*,3a*R*,9b*S*)-1-acetyl-3-methyl-3,3a,4,5-tetrahydro-3,9b-methanobenzo[*g*]indol-2(1*H*)-one (5al)



M.P.: 114-116 °C

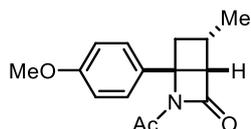
¹H NMR (500 MHz, CDCl₃) δ 7.19 – 7.15 (m, 3H), 7.14 – 7.10 (m, 1H), 2.95 – 2.87 (m, 1H), 2.82 (dt, *J* = 16.2, 3.7 Hz, 1H), 2.77 – 2.71 (m, 1H), 2.68 (d, *J* = 8.2 Hz, 1H), 2.49 (s, 3H), 2.44 (dd, *J* = 8.2, 7.0 Hz, 1H), 2.15 – 2.09 (m, 2H), 1.26 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 181.3, 169.7, 136.2, 132.3, 129.2, 127.6, 127.5, 126.0, 66.4, 61.1, 54.0, 52.1, 28.3, 26.0, 22.2, 10.8.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1758, 1696, 1447, 1361, 1276, 1261, 1182, 1146.

HRMS (ESI⁺) calcd. for $\text{C}_{16}\text{H}_{17}\text{NO}_2\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 278.1152, found: 278.1155.

(±)-(1*R*,4*S*,5*S*)-2-acetyl-1-(4-methoxyphenyl)-5-methyl-2-azabicyclo[2.2.0]hexan-3-one (6am)



Prepared according to **General Procedure D** using **2am** (135 mg, 0.52 mmol, 1.00 equiv), CH_3CN (5.2 mL), and $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbbpy})](\text{PF}_6)$ (5.8 mg, 5.2 μmol , 1.0 mol%). The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of pentane/ethyl acetate (0% to 40%), affording **6am** (31 mg, 0.12 mmol, 23%) as a white solid and **5am** (60 mg, 0.23 mmol, 44%) as a white solid.^{A,B}

Notes: (A) The crude ^1H NMR spectrum indicated a 1:2.2 molar ratio of **6am**/**5am**.

(B) When HFIP was used as the solvent, **6am** was isolated in 27% yield. The crude ^1H NMR spectrum indicated a 1:1 molar ratio of **6am**:**5am**.

M.P.: 55-57 °C

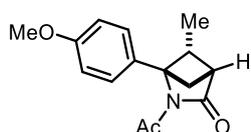
^1H NMR (500 MHz, CDCl_3) δ 7.30 (d, J = 8.8 Hz, 2H), 6.89 (d, J = 8.8 Hz, 2H), 3.80 (s, 3H), 3.27 (d, J = 2.4 Hz, 1H), 2.93 (dd, J = 13.4, 7.7 Hz, 1H), 2.75-2.66 (m, 1H), 2.44-2.38 (m, 4H), 1.33 (d, J = 7.0 Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 168.2, 168.2, 159.5, 129.6, 127.6, 114.0, 61.3, 60.9, 55.4, 37.6, 28.5, 24.5, 22.2.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1774, 1700, 1613, 1516, 1457, 1371, 1277, 1259, 1179, 1027.

HRMS (ESI⁺) calcd. for $\text{C}_{15}\text{H}_{17}\text{NO}_3\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 282.1101, found: 282.1090.

(±)-(1*S*,4*R*,5*R*)-2-acetyl-1-(4-methoxyphenyl)-5-methyl-2-azabicyclo[2.1.1]hexan-3-one (5am)



M.P.: 106-108 °C

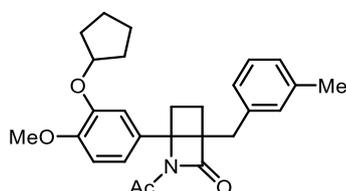
¹H NMR (500 MHz, CDCl₃) δ 7.17 (d, *J* = 8.7 Hz, 2H), 6.88 (d, *J* = 8.7 Hz, 2H), 3.80 (s, 3H), 2.94 (qd, *J* = 6.4, 2.8 Hz, 1H), 2.89 (t, *J* = 2.7 Hz, 1H), 2.64 (d, *J* = 7.4 Hz, 1H), 2.43 (s, 3H), 2.39 (dd, *J* = 7.3, 2.6 Hz, 1H), 1.25 (d, *J* = 6.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 178.4, 170.3, 159.3, 129.4, 127.5, 113.6, 74.5, 56.6, 55.3, 49.3, 48.3, 26.0, 11.9.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1760, 1694, 1614, 1519, 1462, 1364, 1276, 1177, 1030.

HRMS (ESI⁺) calcd. for C₁₅H₁₇NO₃Na⁺ [M+Na]⁺: 282.1101, found: 282.1091.

2-Acetyl-1-(3-(cyclopentyloxy)-4-methoxyphenyl)-4-(3-methylbenzyl)-2-azabicyclo[2.2.0]hexan-3-one (6an)



Prepared according to **General Procedure D** using **2an** (1.69 g, 3.90 mmol, 1.00 equiv), CH₃CN (39.0 mL), and [Ir(dF(CF₃)ppy)₂(dtbbpy)](PF₆) (43.8 mg, 39.0 μmol, 1.0 mol%). The crude reaction mixture was purified by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient elution of Et₂O/pentane (0% to 60%) and pentane/ethyl acetate (0% to 30%), affording **6an** (1.02 g, 2.36 mmol, 60%) as a white solid.^A

Notes: (A) The crude ¹H NMR spectrum indicated a >25:1 molar ratio of **6an**/**5an**.

M.P.: 89-91 °C

¹H NMR (400 MHz, CDCl₃) δ 7.07 (t, *J* = 7.5 Hz, 1H), 6.98 – 6.94 (m, 1H), 6.89 (d, *J* = 8.4 Hz, 1H), 6.81 (dd, *J* = 8.4, 2.2 Hz, 1H), 6.77 (d, *J* = 7.5 Hz, 1H), 6.68 (d, *J* = 2.1 Hz, 1H), 6.58 (s, 1H), 4.72-4.66 (m, 1H), 3.86 (s, 3H), 2.90 (ddd, *J* = 13.5, 11.3, 6.7 Hz, 1H), 2.85 (d, *J* = 14.9 Hz, 1H), 2.76 (d, *J* = 14.8 Hz, 1H), 2.50 – 2.42 (m, 4H), 2.34 – 2.22 (m, 2H), 2.20 (s, 3H), 1.91 – 1.81 (m, 6H), 1.66 – 1.55 (m, 2H).

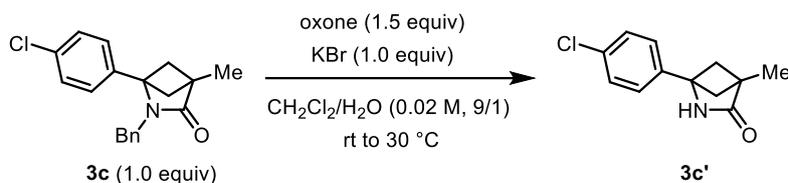
¹³C NMR (126 MHz, CDCl₃) δ 171.2, 167.8, 149.9, 147.7, 137.7, 135.3, 130.6, 128.0, 127.2, 127.2, 126.7, 119.0, 113.2, 111.9, 80.5, 66.3, 64.6, 56.1, 33.6, 32.9, 32.7, 26.0, 24.4, 24.1, 23.5, 21.3.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 2955, 1773, 1705, 1516, 1419, 1371, 1276, 1254.

HRMS (ESI⁺) calcd. for C₂₇H₃₂NO₄⁺ [M+H]⁺: 434.2326, found: 434.2311.

2.5 Product Derivatisation Studies

1-(4-Chlorophenyl)-4-methyl-2-azabicyclo[2.1.1]hexan-3-one (**3c'**)



An oven-dried 50 mL round-bottom flask charged with **3c** (62 mg, 0.20 mmol, 1.0 equiv), KBr (33.8 mg, 0.20 mmol, 1.0 equiv) and oxone (92 mg, 0.30 mmol, 1.5 equiv) were sealed and subjected to three cycles of evacuation and nitrogen backfilling. CH₂Cl₂ (9.0 mL) and H₂O (1.0 mL) were added at room temperature, and the mixture was stirred at 30 °C for 7 h. The reaction was quenched with sat. aq. NaHCO₃ solution (50 mL) and extracted with CH₂Cl₂ (3×30 mL). The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 60%), affording **3c'** (35 mg, 0.16 mmol, 79%) as a white solid.

M.P.: 149-151 °C

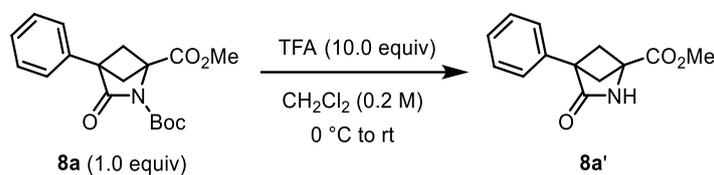
¹H NMR (500 MHz, CDCl₃) δ 7.35 (d, *J* = 8.5 Hz, 2H), 7.24 (d, *J* = 8.5 Hz, 2H), 5.69 (s, 1H), 2.69 (dd, *J* = 3.9, 2.0 Hz, 2H), 2.49 (d, *J* = 4.9 Hz, 2H), 1.33 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 185.8, 135.4, 134.4, 129.1, 127.5, 63.4, 59.4, 50.6, 13.3.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 3186, 1720, 1600, 1496, 1400, 1378, 1245, 1129, 1088.

HRMS (ESI⁺) calcd. for C₁₂H₁₃ClNO⁺ [M+H]⁺: 222.0680, found: 222.0675.

Methyl 3-oxo-4-phenyl-2-azabicyclo[2.1.1]hexane-1-carboxylate (**8a'**)



To an oven-dried 100 mL round-bottom flask containing a solution of **8a** (1.32 g, 4.00 mmol, 1.00 equiv) in CH₂Cl₂ (20.0 mL, 0.2 M), was added TFA (3.00 mL, 40.0 mmol, 10.0 equiv) at 0 °C. The mixture was then stirred at room temperature for 1 h before being concentrated *in vacuo* and diluted with ethyl acetate (50 mL). Then, the mixture was basified to pH 10-11 using 1 M aq. NaOH solution and the aqueous phase were extracted with ethyl acetate (2×50 mL). The combined organic layers

were dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 100%), affording **8a'** (561 mg, 2.43 mmol, 61%) as a white solid.

M.P.: 116-118 °C

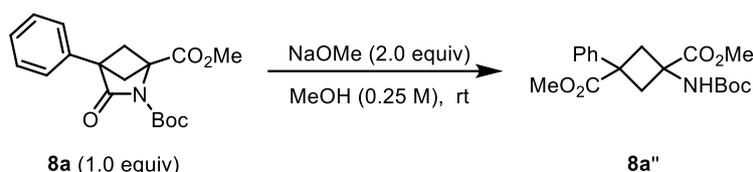
^1H NMR (500 MHz, CDCl_3) δ 7.40 – 7.35 (m, 2H), 7.33 – 7.27 (m, 3H), 6.38 (s, 1H), 3.82 (s, 3H), 3.05 – 2.97 (m, 4H).

^{13}C NMR (126 MHz, CDCl_3) δ 180.8, 167.6, 134.9, 128.5, 127.8, 126.9, 60.3, 58.7, 58.0, 52.7.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1731, 1438, 1293, 1266, 1209, 1154, 1140, 1101.

HRMS (ESI^+) calcd. for $\text{C}_{13}\text{H}_{14}\text{NO}_3^+$ $[\text{M}+\text{H}]^+$: 232.0968, found: 232.0957.

Dimethyl 1-((tert-butoxycarbonyl)amino)-3-phenylcyclobutane-1,3-dicarboxylate (**8a''**)



An oven-dried 25 mL round-bottom flask charged with **8a** (180 mg, 0.54 mmol, 1.00 equiv) was sealed and subjected to three cycles of evacuation and nitrogen backfilling. Sodium methoxide solution (0.5 M in MeOH, 2.17 mL, 1.08 mmol, 2.00 equiv) was added dropwise at room temperature, and the mixture was stirred at this temperature for 1 h. Then the reaction was quenched with brine (50 mL) and extracted with CH_2Cl_2 (3 \times 30 mL). The combined organic layers were dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **8a''** (197 mg, 0.54 mmol, 100%) as a white solid.

M.P.: 153-155 °C

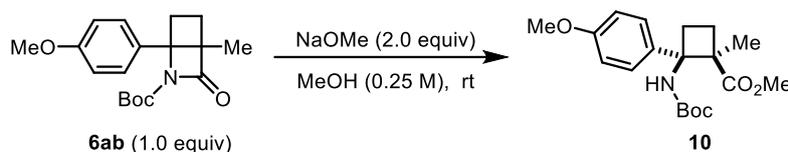
^1H NMR (500 MHz, CDCl_3) δ 7.33 – 7.29 (m, 2H), 7.27 – 7.20 (m, 3H), 5.30 (s, 1H), 3.65 (s, 3H), 3.62 (s, 3H), 3.11 (s, 4H), 1.43 (s, 9H).

^{13}C NMR (126 MHz, CDCl_3) δ 175.8, 173.0, 155.1, 142.2, 128.5, 127.1, 126.0, 80.2, 55.0, 52.7, 52.5, 47.7, 41.2, 28.3.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1721, 1495, 1448, 1367, 1275, 1260, 1215, 1158, 1096.

HRMS (ESI^+) calcd. for $\text{C}_{19}\text{H}_{25}\text{NO}_6\text{Na}^+$ $[\text{M}+\text{Na}]^+$: 386.1574, found: 386.1582.

(±)-Methyl (1*R*,2*R*)-2-((*tert*-butoxycarbonyl)amino)-2-(4-methoxyphenyl)-1-methylcyclobutane-1-carboxylate (10**)**



An oven-dried 50 mL round-bottom flask charged with **6ab** (160 mg, 0.50 mmol, 1.00 equiv) was sealed and subjected to three cycles of evacuation and nitrogen backfilling. Sodium methoxide solution (0.5 M in MeOH, 2.0 mL, 1.0 mmol, 2.0 equiv) was added dropwise at room temperature, and the mixture was stirred at this temperature for 2 h. Then, the reaction was quenched with brine (50 mL) and extracted with CH₂Cl₂ (3 × 30 mL). The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **10** (162 mg, 0.46 mmol, 93%) as a colourless oil.

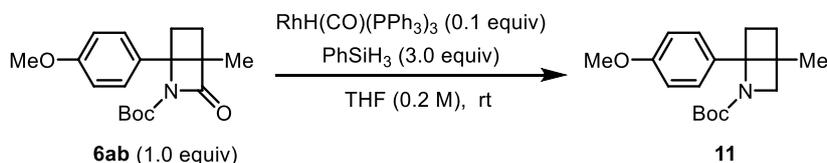
¹H NMR (500 MHz, CDCl₃) δ 7.31 (d, *J* = 8.8 Hz, 2H), 6.82 (d, *J* = 8.8 Hz, 2H), 6.04 (br. s, 1H), 3.76 (s, 3H), 3.75 (s, 3H), 2.72 (dt, *J* = 11.6, 8.7 Hz, 1H), 2.63 (dt, *J* = 10.9, 8.9 Hz, 1H), 2.50 (br. s, 1H), 1.69 (ddd, *J* = 11.0, 8.5, 3.4 Hz, 1H), 1.25 (s, 9H), 0.94 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 176.3, 158.3, 154.7, 134.3, 128.1, 113.2, 79.3, 64.0, 55.2, 52.1, 50.5, 28.3, 27.2, 24.8, 22.3.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1714, 1612, 1513, 1462, 1366, 1275, 1260, 1153, 1120, 1062.

HRMS (ESI⁺) calcd. for C₁₉H₂₇NO₅Na⁺ [M+Na]⁺: 372.1781, found: 372.1784.

***tert*-Butyl 1-(4-methoxyphenyl)-4-methyl-2-azabicyclo[2.2.0]hexane-2-carboxylate (**11**)**



To an oven-dried 25 mL round-bottom flask equipped with a magnetic stir bar was added **6ab** (160 mg, 0.50 mmol, 1.0 equiv) and RhH(CO)(PPh₃)₃ (46.0 mg, 50.0 μmol, 0.10 equiv) under an argon atmosphere in a glovebox. Anhydrous THF (2.5 mL, 0.2 M) and phenylsilane (PhSiH₃, 0.18 mL, 1.5 mmol, 3.0 equiv) were added and the vessel was removed from the glovebox. The reaction was stirred at room temperature overnight under a nitrogen atmosphere. After completion, the crude reaction mixture was concentrated under reduced pressure and purified by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient

elution of ethyl acetate/pentane (0% to 30%) and ethyl acetate/CH₂Cl₂ (0% to 10%). The desired product **11** (64 mg, 0.21 mmol, 42%) was obtained as a colourless oil.^A

Notes: (A) Two sets of peaks were observed in the ¹H and ¹³C NMR spectra due to carbamate rotamers.

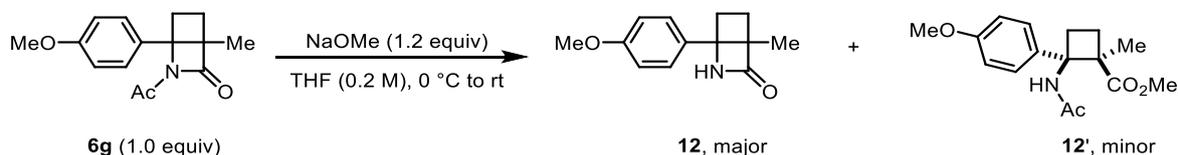
¹H NMR (500 MHz, CDCl₃) δ 7.25 – 7.20 (m, 2H, *overlapping*), 6.91 – 6.86 (m, 2H, *overlapping*), 4.13 (d, *J* = 8.5 Hz, 1H, *major*), 4.10 (d, *J* = 8.5 Hz, 1H, *minor*), 3.94-3.87 (m, 1H, *overlapping*), 3.81 (s, 3H, *major*), 3.79 (s, 3H, *minor*), 2.86-2.76 (m, 1H, *overlapping*), 2.72 – 2.64 (m, 1H, *minor*), 2.54 (ddd, *J* = 13.2, 9.0, 6.0 Hz, 1H, *major*), 2.41-2.28 (m, 1H, *overlapping*), 2.12-1.97 (m, 1H, *overlapping*), 1.47 (s, 9H, *minor*), 1.26 (s, 9H, *major*), 0.93 (s, 3H, *major*), 0.91 (s, 3H, *minor*).

¹³C NMR (126 MHz, CDCl₃) δ 158.62 (*minor*), 158.58 (*major*), 155.9 (*major*), 155.1 (*minor*), 131.5 (*major*), 131.2 (*minor*), 127.6 (*major*), 127.4 (*minor*), 113.8 (*minor*), 113.4 (*major*), 79.5 (*minor*), 79.3 (*major*), 75.2 (*major*), 74.7 (*minor*), 62.7 (*minor*), 61.1 (*major*), 55.3 (*overlapping*), 43.0 (*major*), 42.8 (*minor*), 30.8 (*minor*), 30.6 (*major*), 28.9 (*overlapping*), 28.7 (*minor*), 28.5 (*major*), 19.3 (*overlapping*).

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1693, 1613, 1513, 1455, 1384, 1364, 1275, 1260, 1176, 1147, 1035.

HRMS (ESI⁺) calcd. for C₁₈H₂₅NO₃Na⁺ [M+Na]⁺: 326.1727, found: 326.1732.

1-(4-Methoxyphenyl)-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (**12**)



An oven-dried 50 mL round-bottom flask charged with **6g** (430 mg, 1.66 mmol, 1.00 equiv) was sealed and subjected to three cycles of evacuation and nitrogen backfilling. Anhydrous THF (8.30 mL, 0.2 M) was then added, and the reaction mixture was cooled to 0 °C. Sodium methoxide solution (0.5 M in MeOH, 4.0 mL, 2.0 mmol, 1.2 equiv) was added dropwise at 0 °C, and the mixture was stirred at this temperature for 45 minutes. Then, the reaction was allowed to warm to room temperature and quenched with brine (60 mL) and extracted with CH₂Cl₂ (3 × 40 mL). The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 60%), affording **12** (231 mg, 1.06 mmol, 64%) as a white solid and **12'** (106 mg, 0.36 mmol, 22%) as a colourless oil.

M.P.: 126-128 °C

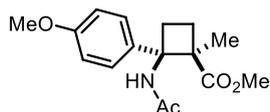
¹H NMR (500 MHz, CDCl₃) δ 7.27 (d, *J* = 8.9 Hz, 2H), 6.92 (d, *J* = 8.8 Hz, 2H), 6.26 (s, 1H), 3.82 (s, 3H), 2.98 – 2.87 (m, 1H), 2.38 – 2.35 (m, 1H), 2.34 – 2.30 (m, 1H), 2.05 – 1.99 (m, 1H), 1.04 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 174.6, 159.2, 129.5, 127.7, 114.1, 63.4, 62.6, 55.4, 29.6, 24.2, 13.6.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1741, 1612, 1515, 1463, 1276, 1260, 1179, 1032.

HRMS (ESI⁺) calcd. for C₁₃H₁₆NO₂⁺ [M+H]⁺: 218.1176, found: 218.1179.

(±)-Methyl (1*R*,2*R*)-2-acetamido-2-(4-methoxyphenyl)-1-methylcyclobutane-1-carboxylate (12')



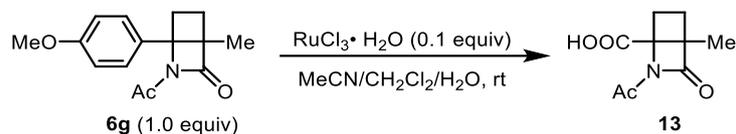
¹H NMR (500 MHz, CDCl₃) δ 7.34 – 7.30 (m, 3H), 6.83 (d, *J* = 8.8 Hz, 2H), 3.79 (s, 3H), 3.77 (s, 3H), 2.83 – 2.74 (m, 1H), 2.67 – 2.55 (m, 2H), 1.88 (s, 3H), 1.78 – 1.71 (m, 1H), 1.00 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 177.0, 169.5, 158.4, 134.0, 128.2, 113.4, 64.4, 55.3, 52.3, 50.2, 28.0, 24.9, 24.4, 22.3.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1716, 1682, 1611, 1513, 1460, 1369, 1275, 1260, 1181, 1148, 1035.

HRMS (ESI⁺) calcd. for C₁₆H₂₁NO₄Na⁺ [M+Na]⁺: 314.1363, found: 314.1367.

2-Acetyl-4-methyl-3-oxo-2-azabicyclo[2.2.0]hexane-1-carboxylic acid (13)



In an oven-dried 250 mL round-bottom flask, **6g** (990 mg, 3.82 mmol, 1.00 equiv) was dissolved in anhydrous MeCN (20 mL), CH₂Cl₂ (20 mL) and H₂O (30 mL) at room temperature. Ruthenium(III) chloride hydrate (86 mg, 0.38 mmol, 0.10 equiv) was then added, followed by NaIO₄ (16.4 g, 76.4 mmol, 20.0 equiv). The reaction mixture was stirred at room temperature overnight until completion of the reaction (TLC). The reaction was quenched with 1 M aqueous HCl solution (80 mL) and extracted with ethyl acetate three times (3 × 50 mL). The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The crude reaction mixture was purified by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 100%) and ethyl acetate/CH₂Cl₂ (0% to 50%), affording **13** (625 mg, 3.17 mmol, 83%) as a white solid.

M.P.: 138-140 °C

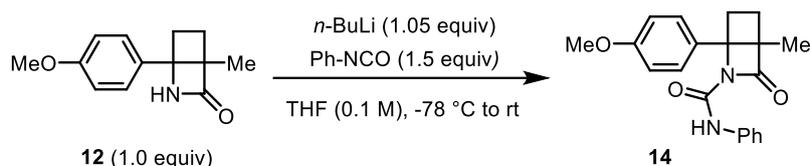
¹H NMR (500 MHz, CDCl₃) δ 11.42 (s, 1H), 3.06 – 2.94 (m, 1H), 2.49-2.37 (m, 4H), 2.29 – 2.16 (m, 2H), 1.38 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 172.3, 168.8, 167.6, 61.8, 61.7, 26.2, 23.8, 23.3, 13.2.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1783, 1708, 1376, 1318, 1276, 1261, 1194, 1106.

HRMS (Nanospray) calcd. for C₉H₁₀NO₄ [M-H]⁻: 196.0615, found: 196.0614.

1-(4-Methoxyphenyl)-4-methyl-3-oxo-N-phenyl-2-azabicyclo[2.2.0]hexane-2-carboxamide (**14**)



An oven-dried 25 mL round-bottom flask charged with **12** (44 mg, 0.20 mmol, 1.0 equiv) was sealed and subjected to three cycles of evacuation and nitrogen backfilling. Anhydrous THF (2.0 mL, 0.1 M) was then added, and the reaction mixture was cooled to -78 °C. *n*-BuLi (2.5 M in hexane, 84 μ L, 0.21 mmol, 1.05 equiv) was added dropwise at -78 °C, and the mixture was stirred at this temperature for 5 minutes. Phenyl isocyanate (32 μ L, 0.30 mmol, 1.5 equiv) was subsequently added at -78 °C, and the reaction was allowed to warm to room temperature over 1 hour. The reaction was then quenched with brine (50 mL) and extracted with ethyl acetate (3 \times 30 mL). The combined organic layers were dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 30%), affording **14** (60 mg, 0.18 mmol, 89%) as a white solid.

M.P.: 124-126 °C

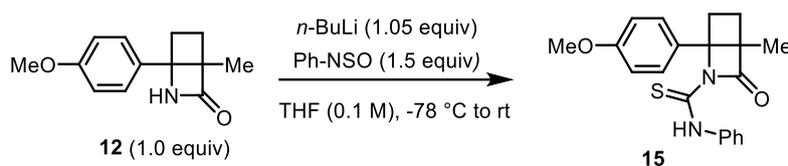
¹H NMR (500 MHz, CDCl₃) δ 8.80 (s, 1H), 7.57 – 7.50 (m, 2H), 7.34 – 7.27 (m, 4H), 7.10 (tt, *J* = 7.2, 1.2 Hz, 1H), 6.94 (d, *J* = 8.9 Hz, 2H), 3.81 (s, 3H), 3.05 (ddd, *J* = 13.7, 11.3, 6.3 Hz, 1H), 2.68 (ddd, *J* = 13.8, 8.7, 5.9 Hz, 1H), 2.51 (ddd, *J* = 13.0, 8.7, 6.3 Hz, 1H), 2.22 (ddd, *J* = 13.1, 11.3, 5.8 Hz, 1H), 1.15 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 173.9, 159.4, 147.8, 137.4, 129.2, 127.6, 127.1, 124.2, 119.6, 114.2, 67.2, 61.6, 55.4, 26.8, 26.2, 13.1.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1752, 1710, 1602, 1546, 1446, 1252, 1180.

HRMS (ESI⁺) calcd. for C₂₀H₂₁N₂O₃⁺ [M+H]⁺: 337.1547, found: 337.1546.

1-(4-Methoxyphenyl)-4-methyl-3-oxo-N-phenyl-2-azabicyclo[2.2.0]hexane-2-carbothioamide (**15**)



An oven-dried 25 mL round-bottom flask charged with **12** (44 mg, 0.20 mmol, 1.0 equiv) was sealed and subjected to three cycles of evacuation and nitrogen backfilling. Anhydrous THF (2.0 mL, 0.1 M) was then added, and the reaction mixture was cooled to -78°C . *n*-BuLi (2.5 M in hexane, 84 μL , 0.21 mmol, 1.05 equiv) was added dropwise at -78°C , and the mixture was stirred at this temperature for 5 minutes. Phenyl isothiocyanate (36 μL , 0.30 mmol, 1.5 equiv) was subsequently added at -78°C , and the reaction was allowed to warm to room temperature over 1 hour. The reaction was then quenched with brine (50 mL) and extracted with ethyl acetate (3 \times 30 mL). The combined organic layers were dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 35%), affording **15** (55 mg, 0.16 mmol, 78%) as a white solid.

M.P.: 70-72 $^\circ\text{C}$

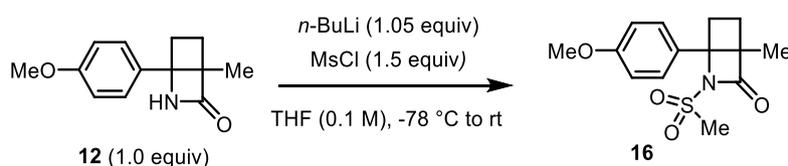
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 10.58 (s, 1H), 7.75 – 7.72 (m, 2H), 7.41 – 7.37 (m, 2H), 7.29 (d, $J = 8.9$ Hz, 2H), 7.25 – 7.21 (m, 1H), 6.94 (d, $J = 8.8$ Hz, 2H), 3.82 (s, 3H), 3.09 (ddd, $J = 14.0, 11.4, 6.4$ Hz, 1H), 2.92 (ddd, $J = 14.0, 8.8, 6.0$ Hz, 1H), 2.58 (ddd, $J = 13.0, 8.8, 6.4$ Hz, 1H), 2.27 (ddd, $J = 13.1, 11.3, 6.0$ Hz, 1H), 1.12 (s, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 174.9, 172.5, 159.2, 137.6, 128.9, 127.8, 126.7, 126.4, 123.5, 114.0, 69.6, 60.6, 55.3, 27.3, 25.6, 12.9.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1741, 1601, 1515, 1374, 1312, 1250, 1227, 1178, 1031.

HRMS (ESI^+) calcd. for $\text{C}_{20}\text{H}_{21}\text{N}_2\text{O}_2\text{S}^+$ $[\text{M}+\text{H}]^+$: 353.1318, found: 353.1317.

1-(4-Methoxyphenyl)-4-methyl-2-(methylsulfonyl)-2-azabicyclo[2.2.0]hexan-3-one (**16**)



An oven-dried 25 mL round-bottom flask charged with **12** (44 mg, 0.20 mmol, 1.0 equiv) was sealed and subjected to three cycles of evacuation and nitrogen backfilling. Anhydrous THF (2.0 mL, 0.1 M) was then added, and the reaction mixture was cooled to -78°C . *n*-BuLi (2.5 M in hexane, 84 μL , 0.21 mmol, 1.05 equiv) was added dropwise at -78°C , and the mixture was stirred at this temperature for

5 minutes. Methanesulfonyl chloride (23 μ L, 0.30 mmol, 1.5 equiv) was subsequently added at -78 $^{\circ}$ C, and the reaction was allowed to warm to room temperature over 1 hour. The reaction was then quenched with brine (50 mL) and extracted with ethyl acetate (3×30 mL). The combined organic layers were dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 50%), affording **16** (32 mg, 0.11 mmol, 54%) as a colourless oil.

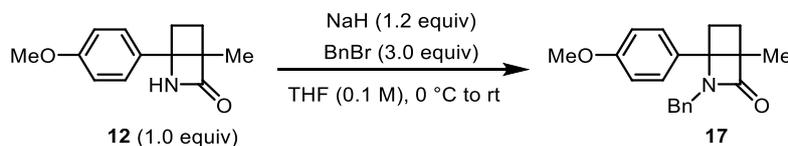
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.29 (d, $J = 8.8$ Hz, 2H), 6.95 (d, $J = 8.8$ Hz, 2H), 3.82 (s, 3H), 3.08 (ddd, $J = 13.9, 11.4, 6.8$ Hz, 1H), 3.02 (s, 3H), 2.87 (ddd, $J = 14.0, 8.7, 5.4$ Hz, 1H), 2.58 (ddd, $J = 13.1, 8.7, 6.8$ Hz, 1H), 2.16 (ddd, $J = 13.1, 11.4, 5.4$ Hz, 1H), 1.20 (s, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 170.8, 159.9, 128.2, 126.3, 114.4, 70.9, 62.9, 55.4, 43.3, 26.6, 25.5, 13.3.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1780, 1676, 1601, 1513, 1349, 1245, 1151, 1028.

HRMS (ESI $^+$) calcd. for $\text{C}_{14}\text{H}_{17}\text{NO}_4\text{SNa}^+$ $[\text{M}+\text{Na}]^+$: 318.0770, found: 318.0770.

2-Benzyl-1-(4-methoxyphenyl)-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (**17**)



An oven-dried 25 mL round-bottom flask charged with **12** (44 mg, 0.20 mmol, 1.0 equiv) was sealed and subjected to three cycles of evacuation and nitrogen backfilling. Anhydrous THF (2.0 mL, 0.1 M) was then added, and the reaction mixture was cooled to 0 $^{\circ}$ C. NaH (60% dispersion in mineral oil, 9.6 mg, 0.24 mmol, 1.2 equiv) was added at 0 $^{\circ}$ C, and the mixture was stirred at this temperature for 30 minutes. Benzoyl bromide (71 μ L, 0.60 mmol, 3.0 equiv) was subsequently added at 0 $^{\circ}$ C, and the reaction was allowed to warm to room temperature over 1 hour. The reaction was then quenched with brine (50 mL) and extracted with ethyl acetate three times (3×30 mL). The combined organic layers were dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 40%), affording **17** (50 mg, 0.16 mmol, 81%) as a colourless oil.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.27 – 7.18 (m, 5H), 7.10 (d, $J = 8.8$ Hz, 2H), 6.87 (d, $J = 8.8$ Hz, 2H), 4.59 (d, $J = 14.8$ Hz, 1H), 4.02 (d, $J = 14.8$ Hz, 1H), 3.80 (s, 3H), 2.67 (ddd, $J = 13.0, 11.0, 6.2$ Hz,

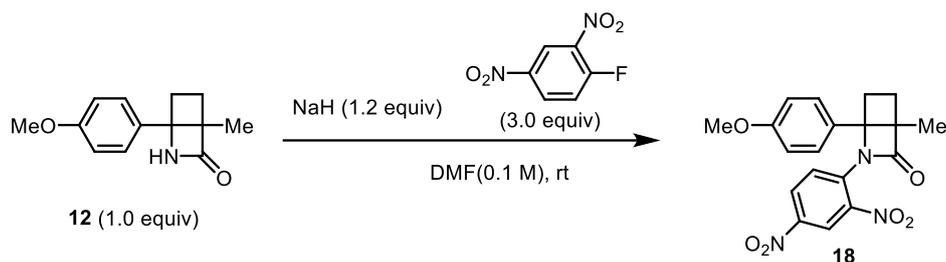
1H), 2.26 (ddd, $J = 12.5, 8.6, 6.2$ Hz, 1H), 1.88 (ddd, $J = 12.6, 11.0, 5.5$ Hz, 1H), 1.78 (ddd, $J = 13.0, 8.6, 5.6$ Hz, 1H), 1.06 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 173.5, 159.2, 136.6, 129.0, 128.7, 128.2, 127.7, 127.6, 114.1, 67.3, 62.5, 55.4, 44.2, 25.4, 24.8, 13.3.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1733, 1611, 1513, 1454, 1297, 1250, 1177, 1030.

HRMS (ESI⁺) calcd. for $\text{C}_{20}\text{H}_{22}\text{NO}_2^+$ $[\text{M}+\text{H}]^+$: 308.1645, found: 308.1642.

2-(2,4-Dinitrophenyl)-1-(4-methoxyphenyl)-4-methyl-2-azabicyclo[2.2.0]hexan-3-one (18)



An oven-dried 25 mL round-bottom flask charged with **12** (65 mg, 0.30 mmol, 1.0 equiv) was sealed and subjected to three cycles of evacuation and nitrogen backfilling. Anhydrous DMF (3.0 mL, 0.1 M) was then added at room temperature, followed by the addition of NaH (60% dispersion in mineral oil, 14.4 mg, 0.360 mmol, 1.2 equiv). The mixture was stirred at room temperature for 30 minutes. 1-Fluoro-2,4-dinitrobenzene (113 μL , 0.90 mmol, 3.0 equiv) was then added and the reaction was stirred at room temperature for 1 hour. The reaction was then quenched with brine (50 mL) and extracted with ethyl acetate (3×30 mL). The combined organic layers were dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude reaction mixture was purified by two successive flash chromatography separations using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 50%) and ethyl acetate/ CH_2Cl_2 (0% to 10%), affording **18** (88 mg, 0.23 mmol, 77%) as a pale-yellow solid.

M.P.: 140-142 $^\circ\text{C}$

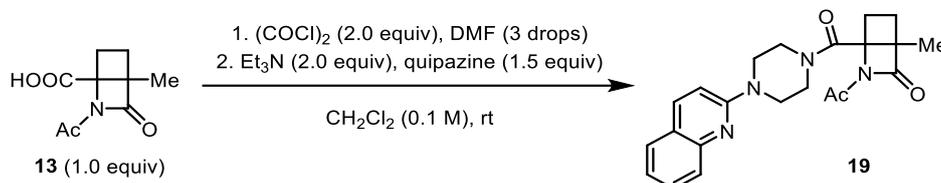
^1H NMR (500 MHz, CDCl_3) δ 8.67 (d, $J = 2.5$ Hz, 1H), 8.25 (dd, $J = 9.0, 2.5$ Hz, 1H), 7.36 (d, $J = 8.9$ Hz, 2H), 7.04 (d, $J = 9.0$ Hz, 1H), 6.95 (d, $J = 8.8$ Hz, 2H), 3.80 (s, 3H), 3.26 (ddd, $J = 13.7, 11.4, 5.8$ Hz, 1H), 2.66 (ddd, $J = 13.6, 8.9, 6.4$ Hz, 1H), 2.51 (ddd, $J = 13.0, 8.9, 5.7$ Hz, 1H), 2.20 (ddd, $J = 13.1, 11.3, 6.3$ Hz, 1H), 1.15 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 170.0, 160.0, 142.7, 141.9, 133.7, 128.2, 127.9, 125.1, 121.9, 120.1, 114.8, 68.8, 63.0, 55.4, 25.74, 25.66, 13.3.

IR (ATR): $\tilde{\nu}$ (cm^{-1}) = 1762, 1600, 1515, 1334, 1253, 1180, 1137, 1030.

HRMS (ESI⁺) calcd. for C₁₉H₁₈N₃O₆⁺ [M+H]⁺: 384.1190, found: 384.1190.

2-Acetyl-4-methyl-1-(4-(quinolin-2-yl)piperazine-1-carbonyl)-2-azabicyclo[2.2.0]hexan-3-one (19)



An oven-dried 25 mL round-bottom flask containing **13** (78 mg, 0.40 mmol, 1.0 equiv) was sealed and subjected to three N₂ evacuation/refill cycles before pre-sparged anhydrous CH₂Cl₂ (4.0 mL, 0.1 M) was added. Oxalyl chloride (68 μL, 0.80 mmol, 2.0 equiv) was then added at room temperature, followed by the addition of DMF (3 drops). The reaction mixture was stirred at room temperature for 1 h. The reaction was then concentrated under reduced pressure to afford the crude acyl chloride, which was used directly for the next step without further purification.

The crude acyl chloride was re-dissolved in anhydrous CH₂Cl₂ (4.0 mL, 0.1 M) at room temperature. Et₃N (112 μL, 0.80 mmol, 2.0 equiv) was then added, followed by the addition of quipazine (150 mg, 0.60 mmol, 1.5 equiv). The reaction mixture was stirred at room temperature for 1 h until complete (TLC). The reaction was then quenched with brine (50 mL) and extracted with CH₂Cl₂ (3 × 30 mL). The combined extracts were dried over anhydrous MgSO₄ and concentrated *in vacuo*. The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/CH₂Cl₂ (0% to 50%), affording **19** (87 mg, 0.22 mmol, 55%) as a white solid.

M.P.: 189-191 °C

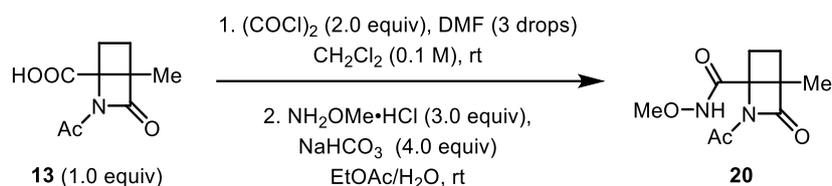
¹H NMR (500 MHz, CDCl₃) δ 7.91 (d, *J* = 9.0 Hz, 1H), 7.73 (s, 1H), 7.60 (dd, *J* = 8.1, 1.5 Hz, 1H), 7.54 (ddd, *J* = 8.5, 7.0, 1.5 Hz, 1H), 7.24 (ddd, *J* = 8.0, 7.2, 1.4 Hz, 1H), 6.95 (d, *J* = 9.1 Hz, 1H), 4.14 – 3.91 (m, 4H), 3.72 – 3.35 (m, 4H), 2.78 (ddd, *J* = 13.0, 11.4, 7.1 Hz, 1H), 2.51 (ddd, *J* = 13.0, 8.3, 4.8 Hz, 1H), 2.42 (s, 3H), 2.36 (ddd, *J* = 12.9, 8.3, 7.0 Hz, 1H), 2.07 (ddd, *J* = 12.9, 11.4, 4.8 Hz, 1H), 1.48 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.3, 168.1, 166.3, 156.9, 147.6, 137.9, 129.8, 127.3, 126.7, 123.2, 122.9, 109.6, 62.1, 45.5, 44.9, 42.6, 27.9, 25.9, 24.1, 14.0.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1781, 1701, 1635, 1603, 1506, 1428, 1376, 1302, 1276, 1230.

HRMS (ESI⁺) calcd. for C₂₂H₂₅N₄O₃⁺ [M+H]⁺: 393.1921, found: 393.1911.

2-Acetyl-*N*-methoxy-4-methyl-3-oxo-2-azabicyclo[2.2.0]hexane-1-carboxamide (20)



An oven-dried 25 mL round-bottom flask containing **13** (60 mg, 0.30 mmol, 1.0 equiv) was sealed and subjected to three N₂ evacuation/refill cycles before pre-sparged anhydrous CH₂Cl₂ (3.0 mL, 0.1 M) was added. Oxalyl chloride (51 μL, 0.60 mmol, 2.0 equiv) was then added at room temperature, followed by the addition of DMF (3 drops). The reaction mixture was stirred at room temperature for 1 h. The reaction was then concentrated under reduced pressure to afford the crude acyl chloride which was used directly for the next step without further purification.

In another oven-dried 25 mL round-bottom flask, methoxyamine hydrochloride (76 mg, 0.90 mmol, 3.0 equiv) was added to a biphasic mixture of NaHCO₃ (101 mg, 1.20 mmol, 4.00 equiv) in EtOAc/H₂O (2:1, 3.0 mL). The mixture was stirred at room temperature for 5 minutes and then transferred to a round-bottom flask containing the crude acyl chloride. The reaction mixture was then stirred at room temperature for 1 h until complete (TLC). The reaction was then quenched with brine (50 mL) and extracted with CH₂Cl₂ (3 × 30 mL). The combined extracts were dried over anhydrous MgSO₄ and concentrated *in vacuo*. The crude reaction mixture was purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 65%), affording **20** (56 mg, 0.25 mmol, 83%) as a white solid.

M.P.: 100-102 °C

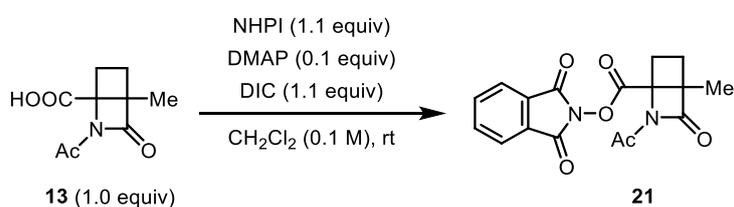
¹H NMR (500 MHz, CDCl₃) δ 9.51 (s, 1H), 3.72 (s, 3H), 2.91 (ddd, *J* = 13.4, 11.3, 6.7 Hz, 1H), 2.45 (s, 3H), 2.37 (ddd, *J* = 12.9, 8.6, 6.7 Hz, 1H), 2.26 (ddd, *J* = 13.0, 11.3, 5.6 Hz, 1H), 2.18 (ddd, *J* = 13.4, 8.6, 5.7 Hz, 1H), 1.39 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.2, 169.9, 165.2, 64.5, 62.6, 61.3, 26.3, 25.2, 24.1, 13.3.

IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1785, 1709, 1659, 1504, 1374, 1312, 1276, 1195, 1065.

HRMS (ESI⁺) calcd. for C₁₀H₁₅N₂O₄⁺ [M+H]⁺: 227.1026, found: 227.1026.

1,3-dioxoisindolin-2-yl 2-acetyl-4-methyl-3-oxo-2-azabicyclo[2.2.0]hexane-1-carboxylate (**21**)



In an oven-dried 25 mL round-bottom flask carboxylic acid **16** (60 mg, 0.30 mmol, 1.0 equiv), *N*-hydroxyphthalimide (NHPI, 54 mg, 0.33 mmol, 1.1 equiv) and DMAP (3.6 mg, 0.03 mmol, 0.10 equiv) were dissolved in anhydrous CH₂Cl₂ (3.0 mL, 0.1 M) at room temperature. *N,N'*-Diisopropylcarbodiimide (DIC, 51 μL, 0.33 mmol, 1.1 equiv) was then added and the reaction was stirred at room temperature for 1 h. The crude product was directly purified using a Biotage automated flash chromatography system with a gradient elution of ethyl acetate/pentane (0% to 50%), affording **21** (89 mg, 0.26 mmol, 87%) as a colourless oil.

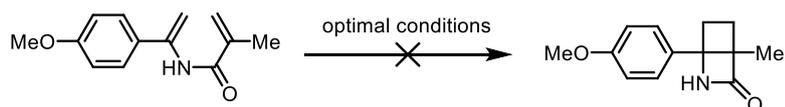
¹H NMR (500 MHz, CDCl₃) δ 7.86 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.79 (dd, *J* = 5.5, 3.1 Hz, 2H), 3.15 – 3.07 (m, 1H), 2.52 – 2.46 (m, 4H), 2.40 – 2.31 (m, 2H), 1.56 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 167.9, 166.7, 163.8, 161.3, 135.0, 128.8, 124.2, 63.2, 60.1, 26.3, 23.8, 23.8, 13.5.

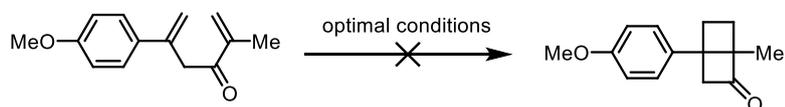
IR (ATR): $\tilde{\nu}$ (cm⁻¹) = 1785, 1742, 1712, 1372, 1316, 1275, 1260, 1185, 1074.

HRMS (ESI⁺) calcd. for C₁₇H₁₄N₂O₆Na⁺ [M+Na]⁺: 365.0744, found: 365.0738.

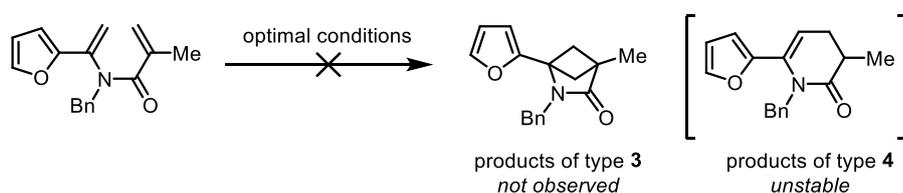
2.6 Unsuccessful Substrates



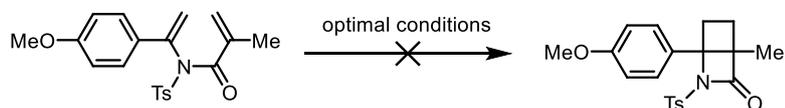
Crude ^1H NMR analysis after 1.5 h showed a significant amount of unreacted starting material, whereas after 24 h, the spectrum appeared complex and the desired product could not be identified.



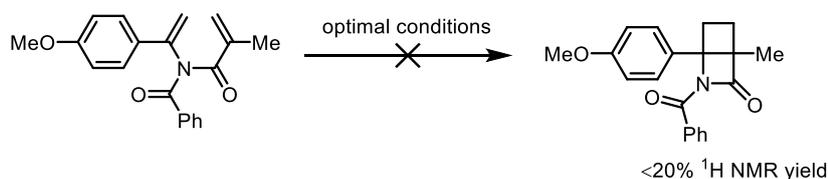
No product formation was detected, and the crude ^1H NMR spectrum after 1.5 h exhibited significant complexity.



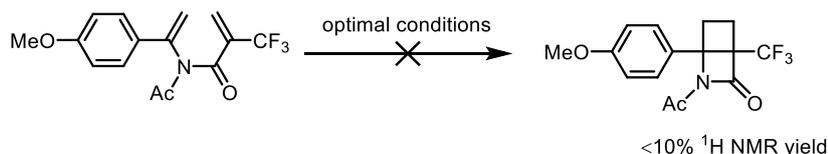
No product formation was detected, and the crude ^1H NMR spectrum after 1.5 h exhibited significant complexity. This may be due to the electron-rich nature of the furan moiety favouring the 6-*endo*-trig pathway, leading predominantly to the formation of the corresponding six-membered-ring product 4. However, due to the photoinstability of 4 under the reaction conditions, no identifiable species were observed in the reaction mixture.



No product formation was detected, and the crude ^1H NMR spectrum after 1.5 h exhibited significant complexity.



The crude ^1H NMR spectrum after 1.5 h showed that the desired product was present in less than 20% yield.



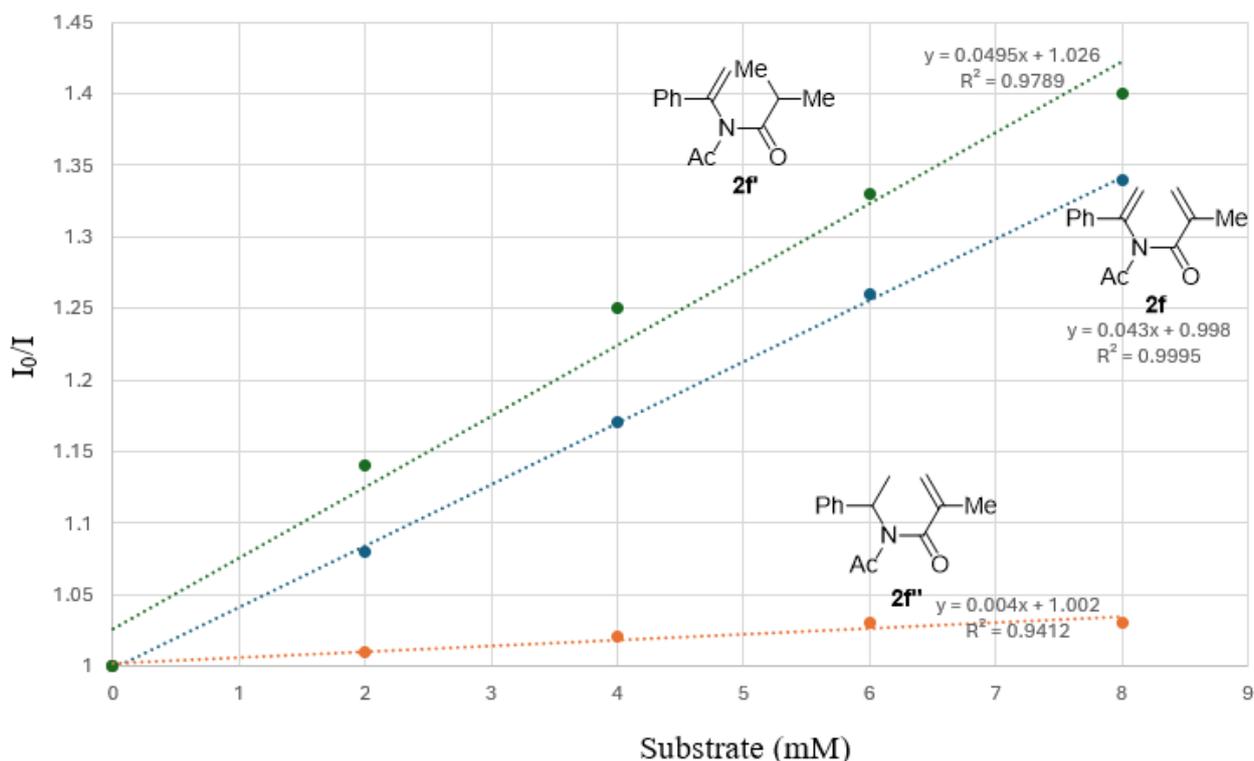
The crude ^1H NMR spectrum after 1.5 h showed that the desired product was present in less than 10% yield.

Supplementary Figure S2. Unsuccessful substrates

3. Stern-Volmer Quenching Studies

All samples were prepared using stock solutions of $(\text{Ir}[\text{dF}(\text{CF}_3)(\text{ppy})]_2)(\text{dtbpy})\text{PF}_6$ (0.10 mM), **2f** (0.01 M), **2f'** (0.01 M) or **2f''** (0.01 M) in dry MeCN. To a volumetric flask was added $(\text{Ir}[\text{dF}(\text{CF}_3)(\text{ppy})]_2)(\text{dtbpy})\text{PF}_6$ (200 μL , 0.10 mM) and the respective amount of quencher. The volume was then adjusted to 3 mL with dry MeCN. The solution was transferred to a 1-cm quartz cuvette and degassed by sparging with nitrogen gas for 15 min. Emission spectra were recorded using a FluoroMax spectrofluorometer with an excitation wavelength of 420 nm. The emission intensities for the Stern-Volmer analysis were observed at 472 nm. The ratio of I_0/I was plotted as a function of the quencher concentration (I_0 : emission intensity of $(\text{Ir}[\text{dF}(\text{CF}_3)(\text{ppy})]_2)(\text{dtbpy})\text{PF}_6$ without quencher; I : emission intensity of $(\text{Ir}[\text{dF}(\text{CF}_3)(\text{ppy})]_2)(\text{dtbpy})\text{PF}_6$ in the presence of quencher). The Stern-Volmer analysis shows that $(\text{Ir}[\text{dF}(\text{CF}_3)(\text{ppy})]_2)(\text{dtbpy})\text{PF}_6$ is only efficiently quenched by the styrene moiety in **2f** or **2f'**, while the enone moiety (**2f''**) does not quench the photocatalyst.

Stern-Volmer Quenching Reaction



Supplementary Figure S3. Stern-Volmer quenching study

4. Crystallographic Data (X-Ray)

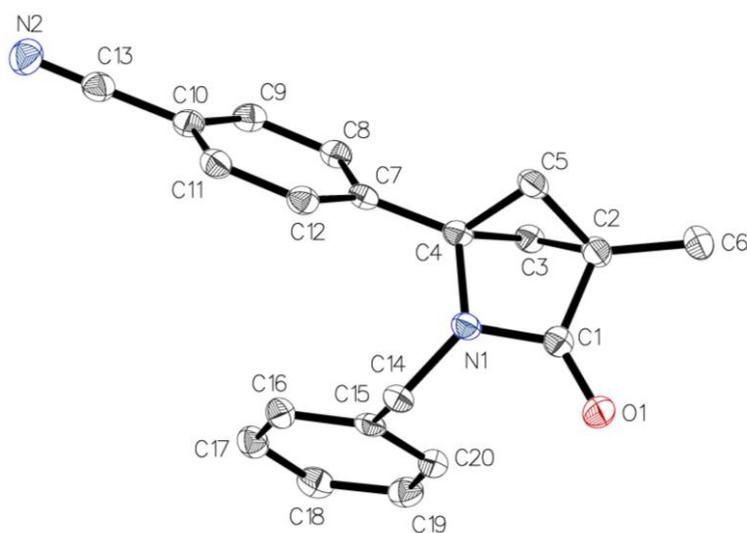
4.1 Structure Report for 3d

Crystallographic data are presented in the Tables below. A single crystal of **3d** was coated in perfluoropolyether oil and mounted on a glass fibre. X-ray measurements were made using a Bruker D8 Venture dual source kappa CPAD diffractometer with Cu K_{α} radiation ($\lambda = 1.54184 \text{ \AA}$).²¹ Intensities were integrated from several series of exposures, each exposure covering 0.5° in ω or ϕ . Absorption corrections were applied, based on multiple and symmetry-equivalent measurements.²² The structure was solved by direct methods and refined by least squares on weighted F^2 values for all reflections (see **Table S3**).²³

All non-hydrogen atoms were assigned anisotropic displacement parameters and refined without positional constraints. All hydrogen atoms were constrained to ideal geometries and were assigned isotropic displacement parameters equal to 1.2 times that of their parent atom.

Refinement proceeded smoothly to give the residuals shown in **Table S3**. Complex neutral-atom scattering factors were used.²⁴

The absolute configuration of the molecule has been established with a Flack/Parsons parameter value of 0.02(9).²⁵



Supplementary Figure S4: Molecular structure of **3d** with thermal ellipsoids shown at the 50% probability level; hydrogen atoms have been omitted for clarity

Supplementary Table S3. Crystal data and structure refinement for **3d**.

Identification code	cu_ZZX652Cu2_0m	
Empirical formula	C ₂₀ H ₁₈ N ₂ O	
Formula weight	302.36	
Temperature	108(2) K	
Wavelength	1.54184 Å	
Crystal system	Monoclinic	
Space group	C2	
Unit cell dimensions	$a = 17.4943(12)$ Å	$\alpha = 90^\circ$
	$b = 7.3922(5)$ Å	$\beta = 103.709(2)^\circ$
	$c = 12.5448(8)$ Å	$\gamma = 90^\circ$
Volume	1576.09(18) Å ³	
Z	4	
Density (calculated)	1.274 Mg/m ³	
Absorption coefficient	0.624 mm ⁻¹	
$F(000)$	640	
Crystal size	0.320 x 0.240 x 0.160 mm	
θ range for data collection	3.627 to 68.742°	
Index ranges	-21 ≤ h ≤ 20, -8 ≤ k ≤ 8, -15 ≤ l ≤ 15	
Reflections collected	11966	
Independent reflections	2821 [$R_{int} = 0.0313$]	
Completeness to $\theta = 67.684^\circ$	99.0 %	
Absorption correction	Numerical	
Max. and min. transmission	1.0000 and 0.7772	
Refinement method	Full-matrix least-squares on F^2	
Data / restraints / parameters	2821 / 1 / 210	
Goodness-of-fit on F^2	$S = 1.078$	
R indices [for 2779 reflections with $I > 2\sigma(I)$]	$R_1 = 0.0317, wR_2 = 0.0765$	
R indices (for all 2821 data)	$R_1 = 0.0329, wR_2 = 0.0771$	
Weighting scheme	$w^{-1} = \sigma^2(F_o^2) + (aP)^2 + (bP)$, where $P = [\max(F_o^2, 0) + 2F_c^2]/3$ $a = 0.0487, b = 0.3852$	
Absolute structure (Flack) parameter	0.02(9)	
Extinction coefficient	0.0032(5)	
Largest diff. peak and hole	0.274 and -0.206 eÅ ⁻³	

Supplementary Table S4. Bond lengths [Å] and angles [°] for **3d**.

C(1)-O(1)	1.224(2)
C(1)-N(1)	1.369(2)
C(1)-C(2)	1.527(3)
C(2)-C(6)	1.501(3)
C(2)-C(5)	1.562(3)
C(2)-C(3)	1.571(3)
C(3)-C(4)	1.562(2)
C(4)-C(7)	1.487(2)
C(4)-N(1)	1.501(2)
C(4)-C(5)	1.549(3)
C(7)-C(8)	1.391(3)
C(7)-C(12)	1.400(2)
C(8)-C(9)	1.382(3)
C(9)-C(10)	1.399(3)
C(10)-C(11)	1.394(3)
C(10)-C(13)	1.435(2)
C(11)-C(12)	1.382(3)
C(13)-N(2)	1.152(3)
C(14)-N(1)	1.456(2)
C(14)-C(15)	1.518(2)
C(15)-C(16)	1.391(3)
C(15)-C(20)	1.397(2)
C(16)-C(17)	1.391(3)
C(17)-C(18)	1.390(3)
C(18)-C(19)	1.390(3)
C(19)-C(20)	1.382(3)
O(1)-C(1)-N(1)	127.64(17)
O(1)-C(1)-C(2)	129.23(17)
N(1)-C(1)-C(2)	103.08(15)
C(6)-C(2)-C(1)	117.65(16)
C(6)-C(2)-C(5)	124.64(17)
C(1)-C(2)-C(5)	99.99(14)
C(6)-C(2)-C(3)	123.83(16)
C(1)-C(2)-C(3)	97.93(14)
C(5)-C(2)-C(3)	85.60(13)
C(4)-C(3)-C(2)	81.62(13)
C(7)-C(4)-N(1)	115.98(14)

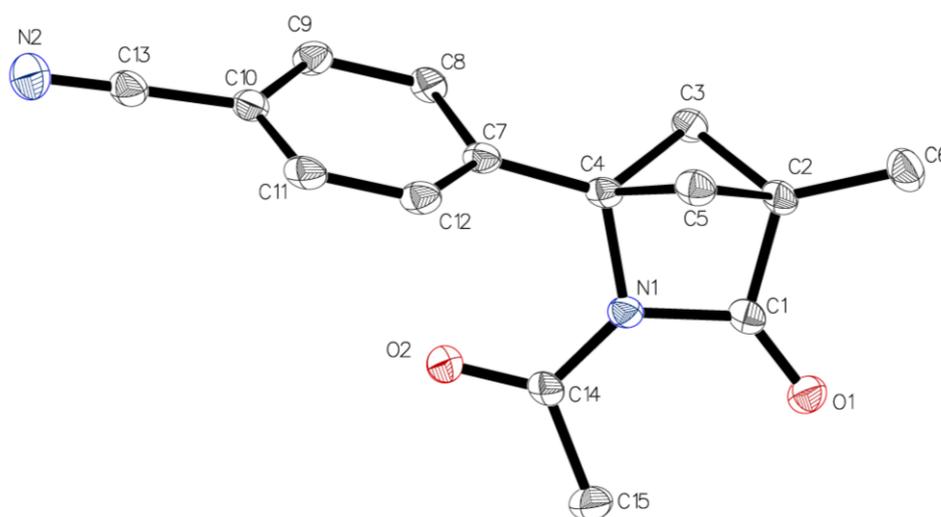
C(7)-C(4)-C(5)	121.93(15)
N(1)-C(4)-C(5)	99.25(14)
C(7)-C(4)-C(3)	125.50(14)
N(1)-C(4)-C(3)	101.60(13)
C(5)-C(4)-C(3)	86.32(13)
C(4)-C(5)-C(2)	82.31(13)
C(8)-C(7)-C(12)	118.98(17)
C(8)-C(7)-C(4)	120.09(15)
C(12)-C(7)-C(4)	120.85(15)
C(9)-C(8)-C(7)	120.78(16)
C(8)-C(9)-C(10)	119.65(16)
C(11)-C(10)-C(9)	120.25(16)
C(11)-C(10)-C(13)	120.46(16)
C(9)-C(10)-C(13)	119.29(16)
C(12)-C(11)-C(10)	119.39(16)
C(11)-C(12)-C(7)	120.92(17)
N(2)-C(13)-C(10)	178.6(2)
N(1)-C(14)-C(15)	114.84(14)
C(16)-C(15)-C(20)	118.68(16)
C(16)-C(15)-C(14)	120.44(15)
C(20)-C(15)-C(14)	120.84(16)
C(17)-C(16)-C(15)	120.80(17)
C(18)-C(17)-C(16)	119.85(17)
C(19)-C(18)-C(17)	119.72(17)
C(20)-C(19)-C(18)	120.18(16)
C(19)-C(20)-C(15)	120.73(17)
C(1)-N(1)-C(14)	123.15(15)
C(1)-N(1)-C(4)	102.64(14)
C(14)-N(1)-C(4)	126.14(14)

4.2 Structure Report for 5p

Crystallographic data are presented in the Tables below. A single crystal of **5p** was coated in perfluoropolyether oil and mounted on a glass fibre. X-ray measurements were made using a Bruker D8 Venture dual source kappa CPAD diffractometer with Mo K_{α} radiation ($\lambda = 0.71073 \text{ \AA}$).²¹ Intensities were integrated from several series of exposures, each exposure covering 0.5° in ω or ϕ . Absorption corrections were applied, based on multiple and symmetry-equivalent measurements.²² The structure was solved by direct methods and refined by least squares on weighted F^2 values for all reflections (see **Table S5**).²³

All non-hydrogen atoms were assigned anisotropic displacement parameters and refined without positional constraints. The positions of the methyl hydrogen atoms were assigned by a rotating group refinement with fixed, idealised C-H distances. All other hydrogen atoms were constrained to ideal geometries. All hydrogen atoms were assigned isotropic displacement parameters equal to 1.5 times (methyl hydrogen atoms) or 1.2 times (all other hydrogen atoms) that of their parent atom.

Refinement proceeded smoothly to give the residuals shown in **Table S5**. Complex neutral-atom scattering factors were used.²⁴



Supplementary Figure S5: Molecular structure of **5p** with thermal ellipsoids shown at the 50% probability level; hydrogen atoms have been omitted for clarity

Supplementary Table S5. Crystal data and structure refinement for **5p**.

Identification code	zzx752_2	
Empirical formula	C ₁₅ H ₁₄ N ₂ O ₂	
Formula weight	254.28	
Temperature	100(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P2 ₁ /n	
Unit cell dimensions	$a = 7.5188(6)$ Å	$\alpha = 90^\circ$
	$b = 12.0570(10)$ Å	$\beta = 95.497(3)^\circ$
	$c = 14.2757(12)$ Å	$\gamma = 90^\circ$
Volume	1288.20(18) Å ³	
Z	4	
Density (calculated)	1.311 Mg/m ³	
Absorption coefficient	0.089 mm ⁻¹	
$F(000)$	536	
Crystal size	0.590 x 0.330 x 0.230 mm	
θ range for data collection	2.215 to 28.343°	
Index ranges	-10 ≤ h ≤ 10, -15 ≤ k ≤ 16, -18 ≤ l ≤ 19	
Reflections collected	13926	
Independent reflections	3165 [$R_{int} = 0.0961$]	
Completeness to $\theta = 25.242^\circ$	98.9 %	
Absorption correction	Numerical	
Max. and min. transmission	1.0000 and 0.5363	
Refinement method	Full-matrix least-squares on F^2	
Data / restraints / parameters	3165 / 0 / 174	
Goodness-of-fit on F^2	$S = 1.076$	
R indices [for 2224 reflections with $I > 2\sigma(I)$]	$R_1 = 0.0887$, $wR_2 = 0.2660$	
R indices (for all 3165 data)	$R_1 = 0.1174$, $wR_2 = 0.2908$	
Weighting scheme	$w^{-1} = \sigma^2(F_o^2) + (aP)^2$, where $P = [\max(F_o^2, 0) + 2F_c^2]/3$ $a = 0.1986$	
Largest diff. peak and hole	0.530 and -0.528 eÅ ⁻³	

Supplementary Table S6. Bond lengths [Å] and angles [°] for **5p**

C(1)-O(1)	1.206(3)
C(1)-N(1)	1.414(3)
C(1)-C(2)	1.521(3)
C(2)-C(6)	1.500(4)
C(2)-C(3)	1.562(4)
C(2)-C(5)	1.563(3)
C(3)-C(4)	1.548(3)
C(4)-C(7)	1.490(3)
C(4)-N(1)	1.511(3)
C(4)-C(5)	1.547(4)
C(7)-C(8)	1.389(4)
C(7)-C(12)	1.401(3)
C(8)-C(9)	1.388(4)
C(9)-C(10)	1.393(4)
C(10)-C(11)	1.399(4)
C(10)-C(13)	1.437(4)
C(11)-C(12)	1.379(4)
C(13)-N(2)	1.152(4)
C(14)-O(2)	1.215(3)
C(14)-N(1)	1.389(3)
C(14)-C(15)	1.501(3)
O(1)-C(1)-N(1)	128.2(2)
O(1)-C(1)-C(2)	129.9(2)
N(1)-C(1)-C(2)	101.87(19)
C(6)-C(2)-C(1)	116.9(2)
C(6)-C(2)-C(3)	123.7(2)
C(1)-C(2)-C(3)	99.7(2)
C(6)-C(2)-C(5)	124.8(2)
C(1)-C(2)-C(5)	98.54(19)
C(3)-C(2)-C(5)	86.34(19)
C(4)-C(3)-C(2)	82.33(18)
C(7)-C(4)-N(1)	116.94(19)
C(7)-C(4)-C(5)	123.21(19)
N(1)-C(4)-C(5)	99.23(19)
C(7)-C(4)-C(3)	123.8(2)
N(1)-C(4)-C(3)	99.97(17)
C(5)-C(4)-C(3)	87.39(19)

C(4)-C(5)-C(2)	82.32(18)
C(8)-C(7)-C(12)	119.4(2)
C(8)-C(7)-C(4)	120.3(2)
C(12)-C(7)-C(4)	120.2(2)
C(9)-C(8)-C(7)	120.8(2)
C(8)-C(9)-C(10)	119.0(2)
C(9)-C(10)-C(11)	120.6(2)
C(9)-C(10)-C(13)	119.5(3)
C(11)-C(10)-C(13)	119.8(2)
C(12)-C(11)-C(10)	119.6(2)
C(11)-C(12)-C(7)	120.2(3)
N(2)-C(13)-C(10)	178.8(2)
O(2)-C(14)-N(1)	121.3(2)
O(2)-C(14)-C(15)	121.8(2)
N(1)-C(14)-C(15)	116.9(2)
C(14)-N(1)-C(1)	128.6(2)
C(14)-N(1)-C(4)	128.0(2)
C(1)-N(1)-C(4)	102.23(18)

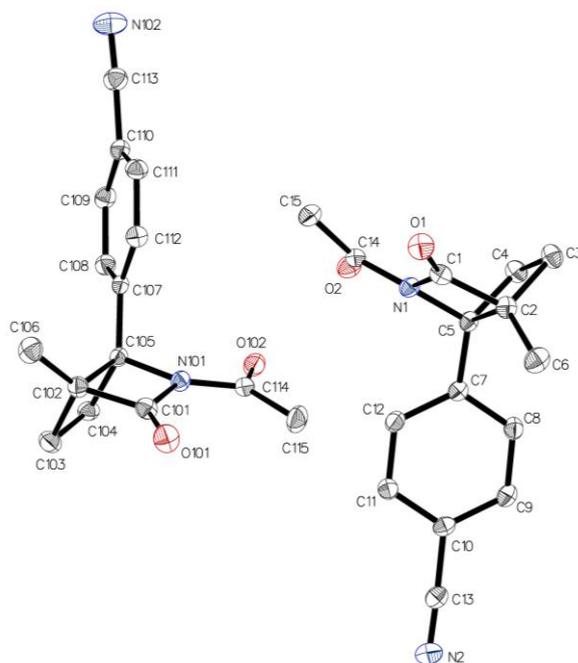
4.3 Structure Report for 6p

Crystallographic data are presented in the Tables below. A single crystal of **6p** was coated in perfluoropolyether oil and mounted on a glass fibre. X-ray measurements were made using a Bruker D8 Venture dual source kappa CPAD diffractometer with Mo K_{α} radiation ($\lambda = 0.71073 \text{ \AA}$).²¹ Intensities were integrated from several series of exposures, each exposure covering 0.5° in ω or ϕ . Absorption corrections were applied, based on multiple and symmetry-equivalent measurements.²² The structure was solved by direct methods and refined by least squares on weighted F^2 values for all reflections (see **Table S7**).²³

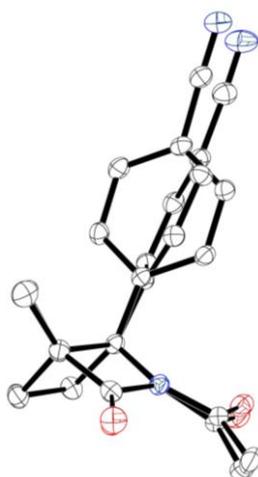
All non-hydrogen atoms were assigned anisotropic displacement parameters and refined without positional constraints. The positions of the methyl hydrogen atoms were assigned by a rotating group refinement with fixed, idealised C-H distances. All other hydrogen atoms were constrained to ideal geometries. All hydrogen atoms were assigned isotropic displacement parameters equal to 1.5 times (methyl hydrogen atoms) or 1.2 times (all other hydrogen atoms) that of their parent atom.

Refinement proceeded smoothly to give the residuals shown in **Table S7**. Complex neutral-atom scattering factors were used.²⁴

The unit cell contains two distinct molecules (shown in **Figure S6**) with identical stereochemistry. An overlay of the two molecules (shown in **Figure S7**) shows that they mainly differ in the conformation of the cyanophenyl ring with the N1-C5-C7-C8 torsion angle differing from its N101-C105-C107-C108 counterpart by $58.59(16)^{\circ}$ with an additional slight difference in the positions of the acyl substituent of the N1-C5 ring, presumably accounted for by packing forces within the structure.

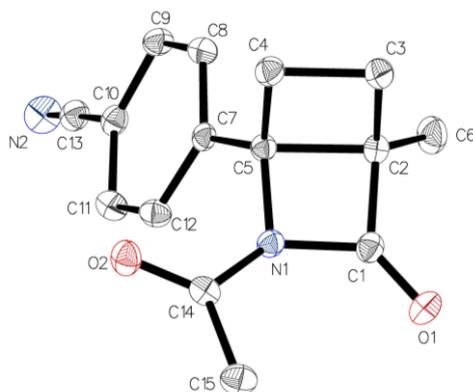


Supplementary Figure S6: Molecular structures of the two independent molecules found in the unit cell for **6p** with thermal ellipsoids shown at the 50% probability level; hydrogen atoms have been omitted for clarity.



Supplementary Figure S7: Overlay of the two independent molecules found in the unit cell for **6p** with thermal ellipsoids shown at the 50% probability level; hydrogen atoms have been omitted for clarity.

The molecular structure of the first molecule within the unit cell is shown in **Figure S8**. Within the four-membered C2-C5 ring, the C2-C3 bond is slightly [0.015(3) Å] longer than those seen for C3-C4 and C4-C5, whilst the C2-C5 bond that links the bridgehead atoms is appreciably [0.047(2) Å] longer than the C4-C5 bond.



Supplementary Figure S8: Molecular structure of one of the two independent molecules found in the unit cell for **6p** with thermal ellipsoids shown at the 50% probability level; hydrogen atoms have been omitted for clarity.

Supplementary Table S7. Crystal data and structure refinement for **6p**

Identification code	mo_ZZX752_1_0m	
Empirical formula	C ₁₅ H ₁₄ N ₂ O ₂	
Formula weight	254.28	
Temperature	100(2) K	
Wavelength	0.71073 Å	
Crystal system	Triclinic	
Space group	P-1	
Unit cell dimensions	a = 9.696(3) Å	α = 84.369(7)°.
	b = 12.448(4) Å	β = 68.207(7)°.
	c = 12.746(4) Å	γ = 68.928(7)°.
Volume	1331.8(7) Å ³	
Z	4	
Density (calculated)	1.268 Mg/m ³	
Absorption coefficient	0.086 mm ⁻¹	
F(000)	536	
Crystal size	0.960 x 0.520 x 0.190 mm ³	
Theta range for data collection	1.755 to 28.271°.	
Index ranges	-12 ≤ h ≤ 12, -16 ≤ k ≤ 16, -16 ≤ l ≤ 16	
Reflections collected	28961	
Independent reflections	6407 [R _{int} = 0.0495]	
Completeness to theta = 25.242°	99.2 %	
Absorption correction	Numerical	
Max. and min. transmission	1.0000 and 0.8452	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	6407 / 0 / 347	
Goodness-of-fit on F ²	1.079	
Final R indices [I > 2σ(I)]	R ₁ = 0.0472, wR ₂ = 0.1115	
R indices (all 6407 data)	R ₁ = 0.0557, wR ₂ = 0.1181	
Weighting scheme	w ⁻¹ = σ ² (F _o ²) + (aP) ² + (bP), where P = [max(F _o ² , 0) + 2F _c ²]/3 a = 0.0577, b = 0.5294	
Largest diff. peak and hole	0.407 and -0.346 e.Å ⁻³	

Supplementary Table S8. Bond lengths [Å] and angles [°] for **6p**

C(1)-O(1)	1.2056(16)
C(1)-N(1)	1.4112(17)
C(1)-C(2)	1.5247(18)
C(2)-C(6)	1.5106(18)
C(2)-C(3)	1.5676(18)
C(2)-C(5)	1.5990(18)
C(3)-C(4)	1.5528(19)
C(3)-H(3A)	0.9900
C(3)-H(3B)	0.9900
C(4)-C(5)	1.5523(17)
C(4)-H(4A)	0.9900
C(4)-H(4B)	0.9900
C(5)-N(1)	1.4867(15)
C(5)-C(7)	1.4952(16)
C(6)-H(6A)	0.9800
C(6)-H(6B)	0.9800
C(6)-H(6C)	0.9800
C(7)-C(8)	1.3992(17)
C(7)-C(12)	1.4003(17)
C(8)-C(9)	1.3884(17)
C(8)-H(8)	0.9500
C(9)-C(10)	1.3997(18)
C(9)-H(9)	0.9500
C(10)-C(11)	1.3973(17)
C(10)-C(13)	1.4461(17)
C(11)-C(12)	1.3866(17)
C(11)-H(11)	0.9500
C(12)-H(12)	0.9500
C(13)-N(2)	1.1507(17)
C(14)-O(2)	1.2160(16)
C(14)-N(1)	1.3971(16)
C(14)-C(15)	1.5036(18)
C(15)-H(15A)	0.9800
C(15)-H(15B)	0.9800
C(15)-H(15C)	0.9800
C(101)-O(101)	1.2073(16)
C(101)-N(101)	1.4080(16)
C(101)-C(102)	1.5280(18)

C(102)-C(106)	1.5084(18)
C(102)-C(103)	1.5661(18)
C(102)-C(105)	1.5987(17)
C(103)-C(104)	1.5537(18)
C(103)-H(10C)	0.9900
C(103)-H(10D)	0.9900
C(104)-C(105)	1.5485(17)
C(104)-H(10A)	0.9900
C(104)-H(10B)	0.9900
C(105)-N(101)	1.4895(16)
C(105)-C(107)	1.5036(16)
C(106)-H(10E)	0.9800
C(106)-H(10F)	0.9800
C(106)-H(10G)	0.9800
C(107)-C(108)	1.3959(18)
C(107)-C(112)	1.4036(17)
C(108)-C(109)	1.3960(17)
C(108)-H(108)	0.9500
C(109)-C(110)	1.4020(19)
C(109)-H(109)	0.9500
C(110)-C(111)	1.398(2)
C(110)-C(113)	1.4507(17)
C(111)-C(112)	1.3895(17)
C(111)-H(111)	0.9500
C(112)-H(112)	0.9500
C(113)-N(102)	1.1488(18)
C(114)-O(102)	1.2178(16)
C(114)-N(101)	1.3927(16)
C(114)-C(115)	1.5048(18)
C(115)-H(11A)	0.9800
C(115)-H(11B)	0.9800
C(115)-H(11C)	0.9800
O(1)-C(1)-N(1)	131.31(12)
O(1)-C(1)-C(2)	136.16(12)
N(1)-C(1)-C(2)	92.51(9)
C(6)-C(2)-C(1)	118.59(11)
C(6)-C(2)-C(3)	120.37(11)
C(1)-C(2)-C(3)	110.72(10)
C(6)-C(2)-C(5)	124.92(11)
C(1)-C(2)-C(5)	86.00(9)

C(3)-C(2)-C(5)	88.40(9)
C(4)-C(3)-C(2)	91.05(9)
C(4)-C(3)-H(3A)	113.5
C(2)-C(3)-H(3A)	113.5
C(4)-C(3)-H(3B)	113.5
C(2)-C(3)-H(3B)	113.5
H(3A)-C(3)-H(3B)	110.8
C(5)-C(4)-C(3)	90.63(9)
C(5)-C(4)-H(4A)	113.5
C(3)-C(4)-H(4A)	113.5
C(5)-C(4)-H(4B)	113.5
C(3)-C(4)-H(4B)	113.5
H(4A)-C(4)-H(4B)	110.8
N(1)-C(5)-C(7)	116.63(10)
N(1)-C(5)-C(4)	111.59(9)
C(7)-C(5)-C(4)	122.85(10)
N(1)-C(5)-C(2)	86.84(9)
C(7)-C(5)-C(2)	120.76(10)
C(4)-C(5)-C(2)	89.90(9)
C(2)-C(6)-H(6A)	109.5
C(2)-C(6)-H(6B)	109.5
H(6A)-C(6)-H(6B)	109.5
C(2)-C(6)-H(6C)	109.5
H(6A)-C(6)-H(6C)	109.5
H(6B)-C(6)-H(6C)	109.5
C(8)-C(7)-C(12)	119.00(11)
C(8)-C(7)-C(5)	119.40(11)
C(12)-C(7)-C(5)	121.39(10)
C(9)-C(8)-C(7)	120.81(11)
C(9)-C(8)-H(8)	119.6
C(7)-C(8)-H(8)	119.6
C(8)-C(9)-C(10)	119.42(11)
C(8)-C(9)-H(9)	120.3
C(10)-C(9)-H(9)	120.3
C(11)-C(10)-C(9)	120.28(11)
C(11)-C(10)-C(13)	118.91(11)
C(9)-C(10)-C(13)	120.78(11)
C(12)-C(11)-C(10)	119.69(11)
C(12)-C(11)-H(11)	120.2
C(10)-C(11)-H(11)	120.2

C(11)-C(12)-C(7)	120.65(11)
C(11)-C(12)-H(12)	119.7
C(7)-C(12)-H(12)	119.7
N(2)-C(13)-C(10)	177.77(13)
O(2)-C(14)-N(1)	118.99(11)
O(2)-C(14)-C(15)	124.88(12)
N(1)-C(14)-C(15)	116.11(11)
C(14)-C(15)-H(15A)	109.5
C(14)-C(15)-H(15B)	109.5
H(15A)-C(15)-H(15B)	109.5
C(14)-C(15)-H(15C)	109.5
H(15A)-C(15)-H(15C)	109.5
H(15B)-C(15)-H(15C)	109.5
C(14)-N(1)-C(1)	132.98(10)
C(14)-N(1)-C(5)	128.16(10)
C(1)-N(1)-C(5)	94.64(9)
O(101)-C(101)-N(101)	131.57(12)
O(101)-C(101)-C(102)	136.30(12)
N(101)-C(101)-C(102)	92.11(10)
C(106)-C(102)-C(101)	118.59(12)
C(106)-C(102)-C(103)	119.67(11)
C(101)-C(102)-C(103)	109.89(10)
C(106)-C(102)-C(105)	126.92(10)
C(101)-C(102)-C(105)	86.32(9)
C(103)-C(102)-C(105)	88.07(9)
C(104)-C(103)-C(102)	91.30(9)
C(104)-C(103)-H(10C)	113.4
C(102)-C(103)-H(10C)	113.4
C(104)-C(103)-H(10D)	113.4
C(102)-C(103)-H(10D)	113.4
H(10C)-C(103)-H(10D)	110.7
C(105)-C(104)-C(103)	90.33(10)
C(105)-C(104)-H(10A)	113.6
C(103)-C(104)-H(10A)	113.6
C(105)-C(104)-H(10B)	113.6
C(103)-C(104)-H(10B)	113.6
H(10A)-C(104)-H(10B)	110.9
N(101)-C(105)-C(107)	114.59(10)
N(101)-C(105)-C(104)	111.74(9)
C(107)-C(105)-C(104)	121.79(11)

N(101)-C(105)-C(102)	86.41(9)
C(107)-C(105)-C(102)	125.29(10)
C(104)-C(105)-C(102)	90.27(9)
C(102)-C(106)-H(10E)	109.5
C(102)-C(106)-H(10F)	109.5
H(10E)-C(106)-H(10F)	109.5
C(102)-C(106)-H(10G)	109.5
H(10E)-C(106)-H(10G)	109.5
H(10F)-C(106)-H(10G)	109.5
C(108)-C(107)-C(112)	119.25(11)
C(108)-C(107)-C(105)	121.14(11)
C(112)-C(107)-C(105)	119.61(11)
C(107)-C(108)-C(109)	120.72(12)
C(107)-C(108)-H(108)	119.6
C(109)-C(108)-H(108)	119.6
C(108)-C(109)-C(110)	119.32(12)
C(108)-C(109)-H(109)	120.3
C(110)-C(109)-H(109)	120.3
C(111)-C(110)-C(109)	120.38(11)
C(111)-C(110)-C(113)	119.34(12)
C(109)-C(110)-C(113)	120.24(12)
C(112)-C(111)-C(110)	119.69(12)
C(112)-C(111)-H(111)	120.2
C(110)-C(111)-H(111)	120.2
C(111)-C(112)-C(107)	120.59(12)
C(111)-C(112)-H(112)	119.7
C(107)-C(112)-H(112)	119.7
N(102)-C(113)-C(110)	177.20(14)
O(102)-C(114)-N(101)	118.50(11)
O(102)-C(114)-C(115)	124.94(12)
N(101)-C(114)-C(115)	116.55(12)
C(114)-C(115)-H(11A)	109.5
C(114)-C(115)-H(11B)	109.5
H(11A)-C(115)-H(11B)	109.5
C(114)-C(115)-H(11C)	109.5
H(11A)-C(115)-H(11C)	109.5
H(11B)-C(115)-H(11C)	109.5
C(114)-N(101)-C(101)	135.89(11)
C(114)-N(101)-C(105)	126.90(10)
C(101)-N(101)-C(105)	95.15(9)

Supplementary Table S9. Torsion angles [°] for **6p**.

O(1)-C(1)-C(2)-C(6)	53.2(2)
N(1)-C(1)-C(2)-C(6)	-128.40(12)
O(1)-C(1)-C(2)-C(3)	-92.10(17)
N(1)-C(1)-C(2)-C(3)	86.35(11)
O(1)-C(1)-C(2)-C(5)	-178.87(15)
N(1)-C(1)-C(2)-C(5)	-0.42(8)
C(6)-C(2)-C(3)-C(4)	129.21(12)
C(1)-C(2)-C(3)-C(4)	-86.25(11)
C(5)-C(2)-C(3)-C(4)	-1.13(9)
C(2)-C(3)-C(4)-C(5)	1.16(9)
C(3)-C(4)-C(5)-N(1)	85.42(11)
C(3)-C(4)-C(5)-C(7)	-128.56(12)
C(3)-C(4)-C(5)-C(2)	-1.14(9)
C(6)-C(2)-C(5)-N(1)	122.83(12)
C(1)-C(2)-C(5)-N(1)	0.40(8)
C(3)-C(2)-C(5)-N(1)	-110.50(9)
C(6)-C(2)-C(5)-C(7)	3.52(18)
C(1)-C(2)-C(5)-C(7)	-118.91(11)
C(3)-C(2)-C(5)-C(7)	130.19(11)
C(6)-C(2)-C(5)-C(4)	-125.54(13)
C(1)-C(2)-C(5)-C(4)	112.03(9)
C(3)-C(2)-C(5)-C(4)	1.13(9)
N(1)-C(5)-C(7)-C(8)	-172.97(11)
C(4)-C(5)-C(7)-C(8)	42.57(17)
C(2)-C(5)-C(7)-C(8)	-69.87(15)
N(1)-C(5)-C(7)-C(12)	1.66(17)
C(4)-C(5)-C(7)-C(12)	-142.80(12)
C(2)-C(5)-C(7)-C(12)	104.76(14)
C(12)-C(7)-C(8)-C(9)	-3.07(19)
C(5)-C(7)-C(8)-C(9)	171.69(12)
C(7)-C(8)-C(9)-C(10)	-0.45(19)
C(8)-C(9)-C(10)-C(11)	3.10(19)
C(8)-C(9)-C(10)-C(13)	-174.59(12)
C(9)-C(10)-C(11)-C(12)	-2.18(19)
C(13)-C(10)-C(11)-C(12)	175.55(12)
C(10)-C(11)-C(12)-C(7)	-1.4(2)
C(8)-C(7)-C(12)-C(11)	4.01(19)
C(5)-C(7)-C(12)-C(11)	-170.65(12)

O(2)-C(14)-N(1)-C(1)	160.89(12)
C(15)-C(14)-N(1)-C(1)	-17.57(18)
O(2)-C(14)-N(1)-C(5)	10.09(18)
C(15)-C(14)-N(1)-C(5)	-168.36(11)
O(1)-C(1)-N(1)-C(14)	21.7(2)
C(2)-C(1)-N(1)-C(14)	-156.91(12)
O(1)-C(1)-N(1)-C(5)	179.02(13)
C(2)-C(1)-N(1)-C(5)	0.45(9)
C(7)-C(5)-N(1)-C(14)	-78.37(15)
C(4)-C(5)-N(1)-C(14)	69.94(15)
C(2)-C(5)-N(1)-C(14)	158.58(11)
C(7)-C(5)-N(1)-C(1)	122.61(11)
C(4)-C(5)-N(1)-C(1)	-89.07(11)
C(2)-C(5)-N(1)-C(1)	-0.43(9)
O(101)-C(101)-C(102)-C(106)	51.3(2)
N(101)-C(101)-C(102)-C(106)	-130.18(11)
O(101)-C(101)-C(102)-C(103)	-91.51(18)
N(101)-C(101)-C(102)-C(103)	87.01(11)
O(101)-C(101)-C(102)-C(105)	-178.12(16)
N(101)-C(101)-C(102)-C(105)	0.40(9)
C(106)-C(102)-C(103)-C(104)	130.97(12)
C(101)-C(102)-C(103)-C(104)	-86.69(11)
C(105)-C(102)-C(103)-C(104)	-1.30(9)
C(102)-C(103)-C(104)-C(105)	1.34(9)
C(103)-C(104)-C(105)-N(101)	84.93(11)
C(103)-C(104)-C(105)-C(107)	-134.36(11)
C(103)-C(104)-C(105)-C(102)	-1.31(9)
C(106)-C(102)-C(105)-N(101)	123.09(13)
C(101)-C(102)-C(105)-N(101)	-0.38(8)
C(103)-C(102)-C(105)-N(101)	-110.46(9)
C(106)-C(102)-C(105)-C(107)	5.3(2)
C(101)-C(102)-C(105)-C(107)	-118.18(12)
C(103)-C(102)-C(105)-C(107)	131.74(12)
C(106)-C(102)-C(105)-C(104)	-125.15(13)
C(101)-C(102)-C(105)-C(104)	111.38(9)
C(103)-C(102)-C(105)-C(104)	1.30(9)
N(101)-C(105)-C(107)-C(108)	128.44(12)
C(104)-C(105)-C(107)-C(108)	-11.26(17)
C(102)-C(105)-C(107)-C(108)	-127.71(13)
N(101)-C(105)-C(107)-C(112)	-51.71(14)

C(104)-C(105)-C(107)-C(112)	168.59(11)
C(102)-C(105)-C(107)-C(112)	52.15(16)
C(112)-C(107)-C(108)-C(109)	-1.66(17)
C(105)-C(107)-C(108)-C(109)	178.20(11)
C(107)-C(108)-C(109)-C(110)	-0.23(18)
C(108)-C(109)-C(110)-C(111)	1.96(18)
C(108)-C(109)-C(110)-C(113)	-176.03(11)
C(109)-C(110)-C(111)-C(112)	-1.76(18)
C(113)-C(110)-C(111)-C(112)	176.25(11)
C(110)-C(111)-C(112)-C(107)	-0.17(18)
C(108)-C(107)-C(112)-C(111)	1.87(18)
C(105)-C(107)-C(112)-C(111)	-177.99(11)
O(102)-C(114)-N(101)-C(101)	166.89(13)
C(115)-C(114)-N(101)-C(101)	-14.4(2)
O(102)-C(114)-N(101)-C(105)	7.50(18)
C(115)-C(114)-N(101)-C(105)	-173.77(11)
O(101)-C(101)-N(101)-C(114)	14.6(2)
C(102)-C(101)-N(101)-C(114)	-164.02(14)
O(101)-C(101)-N(101)-C(105)	178.20(14)
C(102)-C(101)-N(101)-C(105)	-0.43(9)
C(107)-C(105)-N(101)-C(114)	-66.39(15)
C(104)-C(105)-N(101)-C(114)	77.32(14)
C(102)-C(105)-N(101)-C(114)	166.17(11)
C(107)-C(105)-N(101)-C(101)	127.85(11)
C(104)-C(105)-N(101)-C(101)	-88.44(11)
C(102)-C(105)-N(101)-C(101)	0.41(9)

Supplementary Table S10. Crystal data and structure refinement for **8a**.

Identification code	zzx606	
Empirical formula	C ₁₈ H ₂₁ NO ₅	
Formula weight	331.36	
Temperature	100(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P2 ₁ /n	
Unit cell dimensions	$a = 5.986(8)$ Å	$\alpha = 90^\circ$
	$b = 30.35(4)$ Å	$\beta = 98.61(3)^\circ$
	$c = 9.570(14)$ Å	$\gamma = 90^\circ$
Volume	1719(4) Å ³	
Z	4	
Density (calculated)	1.280 Mg/m ³	
Absorption coefficient	0.094 mm ⁻¹	
$F(000)$	704	
Crystal size	0.200 x 0.100 x 0.020 mm	
θ range for data collection	2.255 to 24.998°	
Index ranges	-7 ≤ h ≤ 7, -36 ≤ k ≤ 36, -11 ≤ l ≤ 11	
Reflections collected	31906	
Independent reflections	3026 [$R_{int} = 0.0839$]	
Completeness to $\theta = 24.998^\circ$	100.0 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.9590 and 0.8116	
Refinement method	Full-matrix least-squares on F^2	
Data / restraints / parameters	3026 / 0 / 221	
Goodness-of-fit on F^2	$S = 1.076$	
R indices [for 2236 reflections with $I > 2\sigma(I)$]	$R_1 = 0.0488$, $wR_2 = 0.1262$	
R indices (for all 3026 data)	$R_1 = 0.0725$, $wR_2 = 0.1408$	
Weighting scheme	$w^{-1} = \sigma^2(F_o^2) + (aP)^2 + (bP)$, where $P = [\max(F_o^2, 0) + 2F_c^2]/3$ $a = 0.0759$, $b = 0.3963$	
Largest diff. peak and hole	0.211 and -0.418 eÅ ⁻³	

Supplementary Table S11. Bond lengths [Å] and angles [°] for **8a**.

C(1)-C(2)	1.545(3)
C(1)-N(1)	1.427(3)
C(1)-O(3)	1.215(3)
C(2)-C(3)	1.576(3)
C(2)-C(5)	1.589(3)
C(2)-C(11)	1.500(3)
C(3)-H(3A)	0.9900
C(3)-H(3B)	0.9900
C(3)-C(4)	1.559(3)
C(4)-C(5)	1.549(3)
C(4)-C(17)	1.513(4)
C(4)-N(1)	1.509(3)
C(5)-H(5A)	0.9900
C(5)-H(5B)	0.9900
C(6)-N(1)	1.403(3)
C(6)-O(1)	1.218(3)
C(6)-O(2)	1.333(3)
C(7)-C(8)	1.525(3)
C(7)-C(9)	1.521(3)
C(7)-C(10)	1.531(4)
C(7)-O(2)	1.506(3)
C(8)-H(8A)	0.9800
C(8)-H(8B)	0.9800
C(8)-H(8C)	0.9800
C(9)-H(9A)	0.9800
C(9)-H(9B)	0.9800
C(9)-H(9C)	0.9800
C(10)-H(10A)	0.9800
C(10)-H(10B)	0.9800
C(10)-H(10C)	0.9800
C(11)-C(12)	1.401(3)
C(11)-C(16)	1.404(3)
C(12)-H(12)	0.9500
C(12)-C(13)	1.395(3)
C(13)-H(13)	0.9500
C(13)-C(14)	1.399(3)
C(14)-H(14)	0.9500
C(14)-C(15)	1.396(4)

C(15)-H(15)	0.9500
C(15)-C(16)	1.394(3)
C(16)-H(16)	0.9500
C(17)-O(4)	1.210(3)
C(17)-O(5)	1.348(3)
C(18)-H(18A)	0.9800
C(18)-H(18B)	0.9800
C(18)-H(18C)	0.9800
C(18)-O(5)	1.463(3)

N(1)-C(1)-C(2)	101.4(2)
O(3)-C(1)-C(2)	130.44(19)
O(3)-C(1)-N(1)	128.10(19)
C(1)-C(2)-C(3)	99.68(18)
C(1)-C(2)-C(5)	98.88(18)
C(3)-C(2)-C(5)	85.60(17)
C(11)-C(2)-C(1)	118.2(2)
C(11)-C(2)-C(3)	124.41(18)
C(11)-C(2)-C(5)	122.91(18)
C(2)-C(3)-H(3A)	115.0
C(2)-C(3)-H(3B)	115.0
H(3A)-C(3)-H(3B)	112.0
C(4)-C(3)-C(2)	81.95(15)
C(4)-C(3)-H(3A)	115.0
C(4)-C(3)-H(3B)	115.0
C(5)-C(4)-C(3)	87.60(15)
C(17)-C(4)-C(3)	122.84(18)
C(17)-C(4)-C(5)	122.99(18)
N(1)-C(4)-C(3)	98.95(16)
N(1)-C(4)-C(5)	102.59(19)
N(1)-C(4)-C(17)	116.13(18)
C(2)-C(5)-H(5A)	115.0
C(2)-C(5)-H(5B)	115.0
C(4)-C(5)-C(2)	81.85(16)
C(4)-C(5)-H(5A)	115.0
C(4)-C(5)-H(5B)	115.0
H(5A)-C(5)-H(5B)	112.0
O(1)-C(6)-N(1)	122.2(2)
O(1)-C(6)-O(2)	128.07(19)
O(2)-C(6)-N(1)	109.67(18)

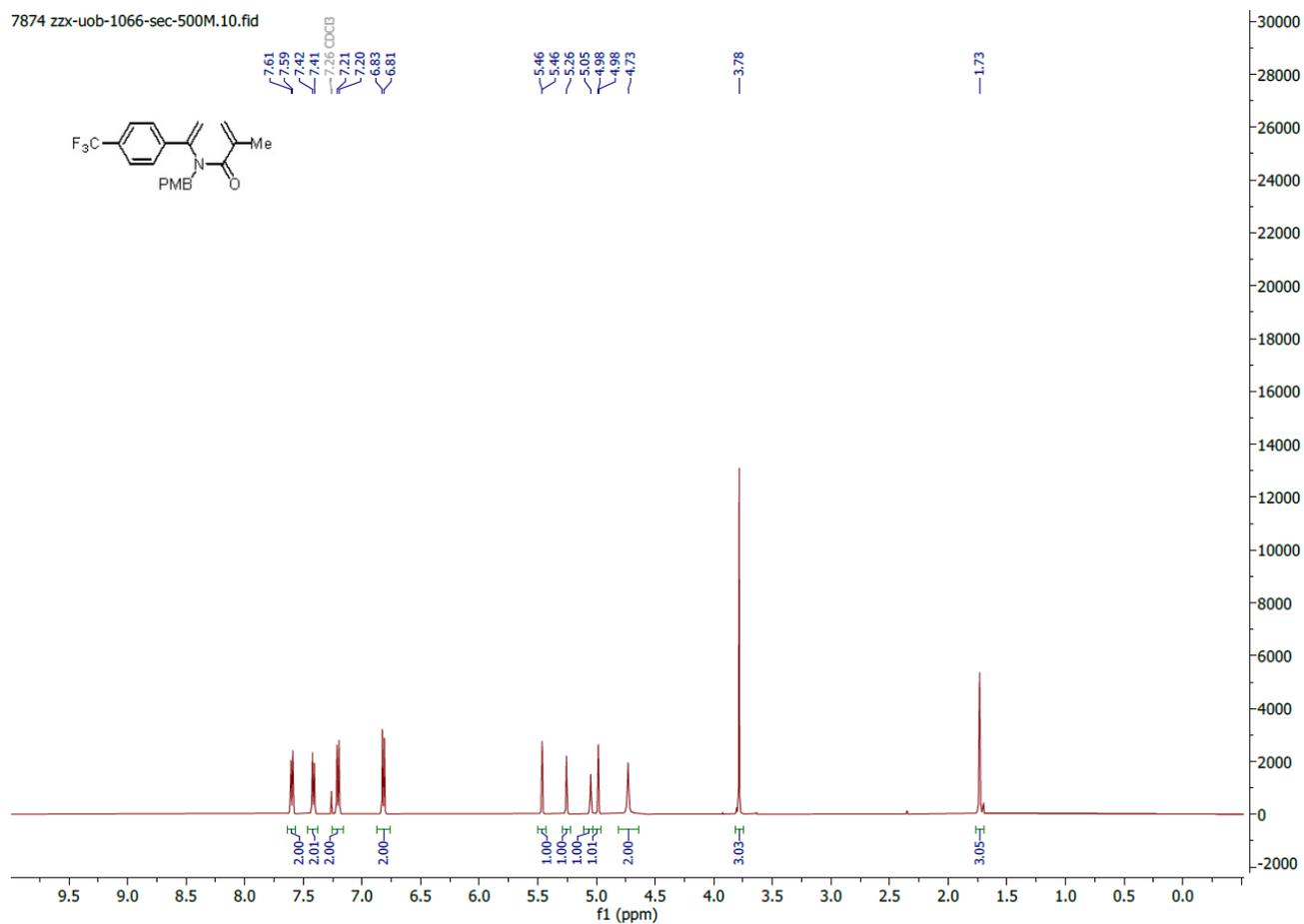
C(8)-C(7)-C(10)	112.1(2)
C(9)-C(7)-C(8)	113.0(2)
C(9)-C(7)-C(10)	110.3(2)
O(2)-C(7)-C(8)	108.47(19)
O(2)-C(7)-C(9)	110.52(19)
O(2)-C(7)-C(10)	101.81(18)
C(7)-C(8)-H(8A)	109.5
C(7)-C(8)-H(8B)	109.5
C(7)-C(8)-H(8C)	109.5
H(8A)-C(8)-H(8B)	109.5
H(8A)-C(8)-H(8C)	109.5
H(8B)-C(8)-H(8C)	109.5
C(7)-C(9)-H(9A)	109.5
C(7)-C(9)-H(9B)	109.5
C(7)-C(9)-H(9C)	109.5
H(9A)-C(9)-H(9B)	109.5
H(9A)-C(9)-H(9C)	109.5
H(9B)-C(9)-H(9C)	109.5
C(7)-C(10)-H(10A)	109.5
C(7)-C(10)-H(10B)	109.5
C(7)-C(10)-H(10C)	109.5
H(10A)-C(10)-H(10B)	109.5
H(10A)-C(10)-H(10C)	109.5
H(10B)-C(10)-H(10C)	109.5
C(12)-C(11)-C(2)	121.45(19)
C(12)-C(11)-C(16)	118.86(18)
C(16)-C(11)-C(2)	119.68(19)
C(11)-C(12)-H(12)	119.8
C(13)-C(12)-C(11)	120.4(2)
C(13)-C(12)-H(12)	119.8
C(12)-C(13)-H(13)	119.9
C(12)-C(13)-C(14)	120.3(2)
C(14)-C(13)-H(13)	119.9
C(13)-C(14)-H(14)	120.2
C(15)-C(14)-C(13)	119.61(19)
C(15)-C(14)-H(14)	120.2
C(14)-C(15)-H(15)	119.9
C(16)-C(15)-C(14)	120.1(2)
C(16)-C(15)-H(15)	119.9
C(11)-C(16)-H(16)	119.7

C(15)-C(16)-C(11)	120.7(2)
C(15)-C(16)-H(16)	119.7
O(4)-C(17)-C(4)	123.85(19)
O(4)-C(17)-O(5)	125.4(2)
O(5)-C(17)-C(4)	110.70(17)
H(18A)-C(18)-H(18B)	109.5
H(18A)-C(18)-H(18C)	109.5
H(18B)-C(18)-H(18C)	109.5
O(5)-C(18)-H(18A)	109.5
O(5)-C(18)-H(18B)	109.5
O(5)-C(18)-H(18C)	109.5
C(1)-N(1)-C(4)	102.27(18)
C(6)-N(1)-C(1)	128.19(19)
C(6)-N(1)-C(4)	124.02(17)
C(6)-O(2)-C(7)	121.02(16)
C(17)-O(5)-C(18)	115.34(16)

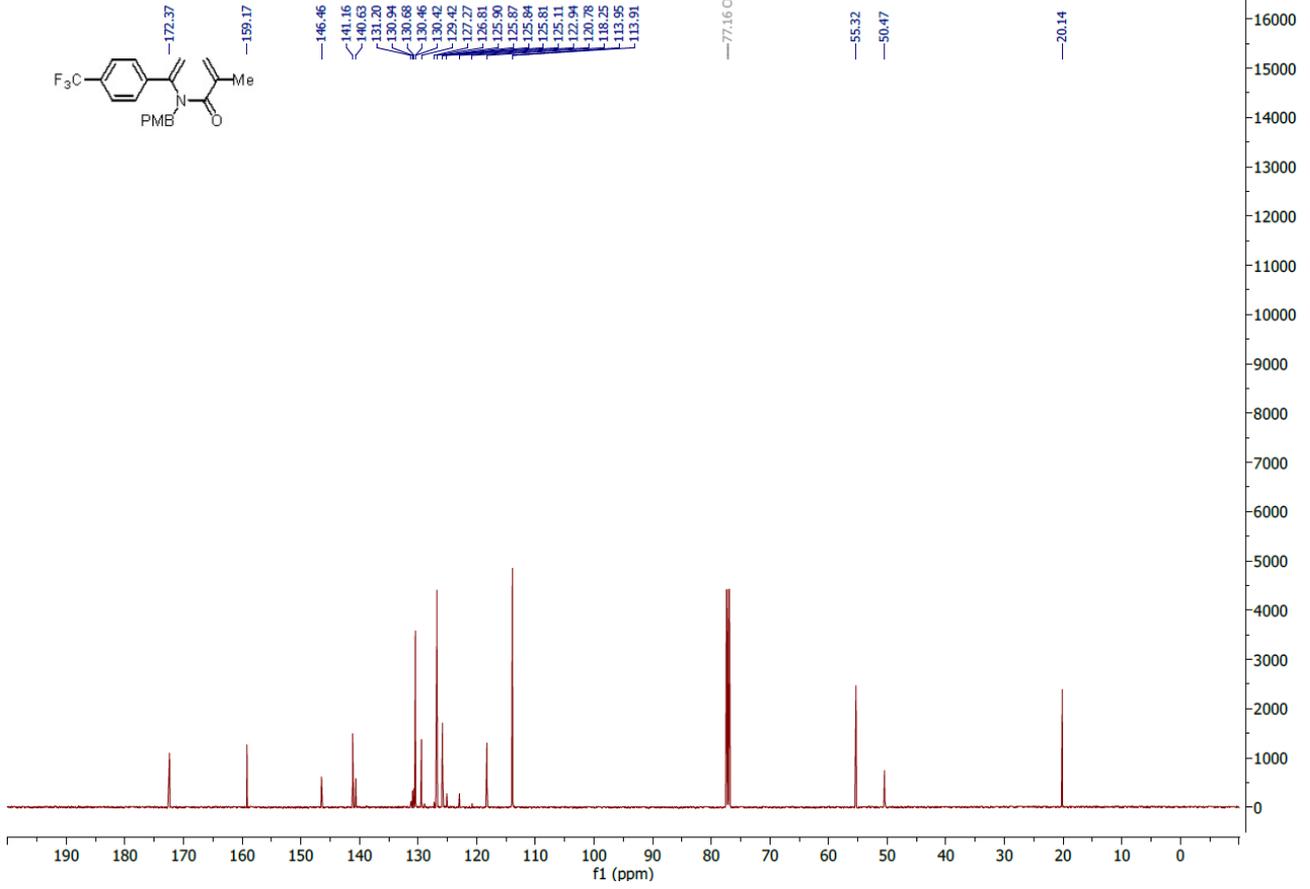
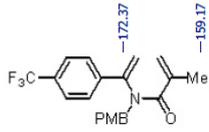
5. NMR Spectra

5.1 NMR Spectra of Cyclisation Precursors (1)

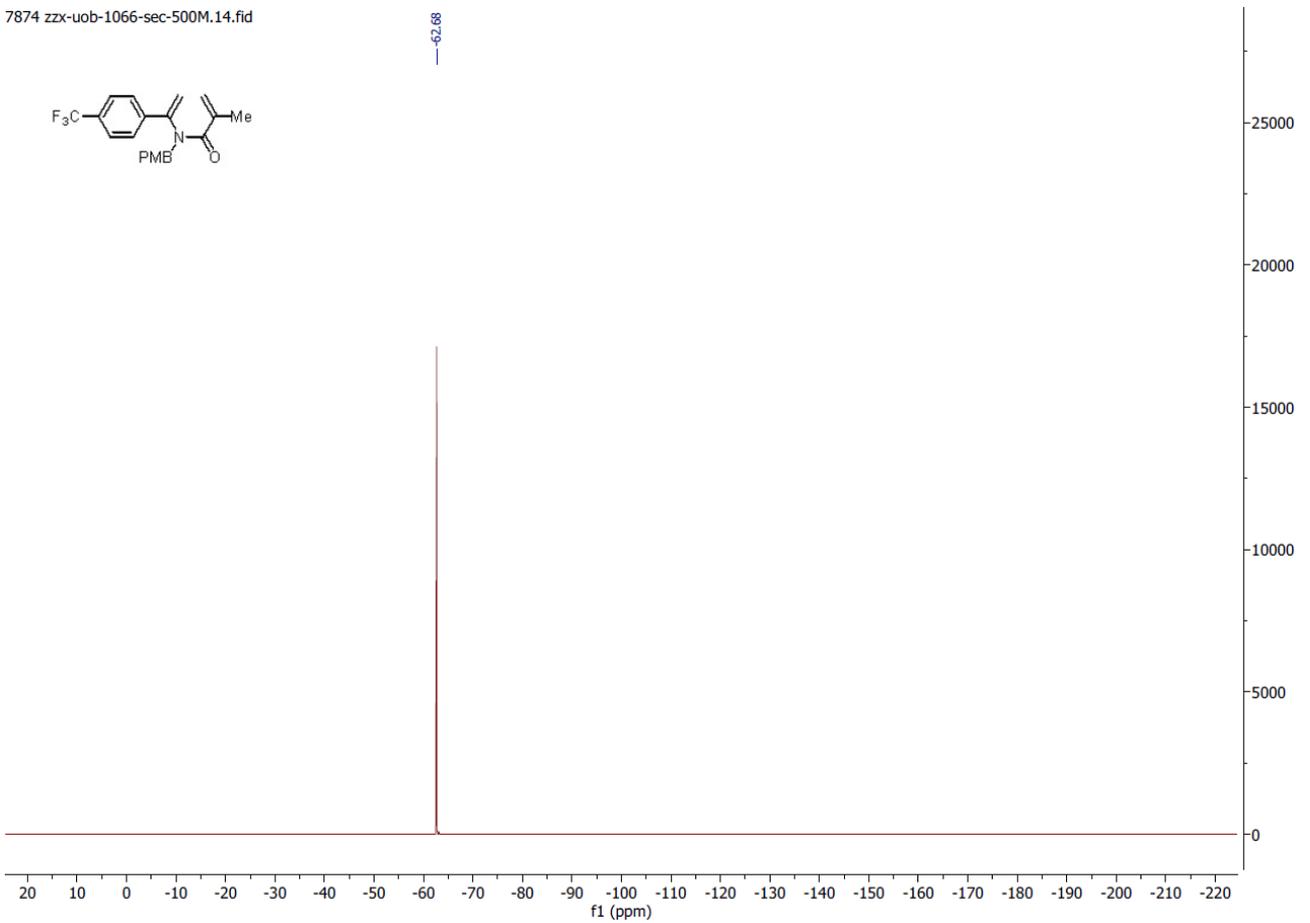
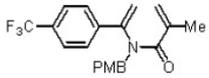
Compound 1a



7874 zzx-uob-1066-sec-500M.11.fid

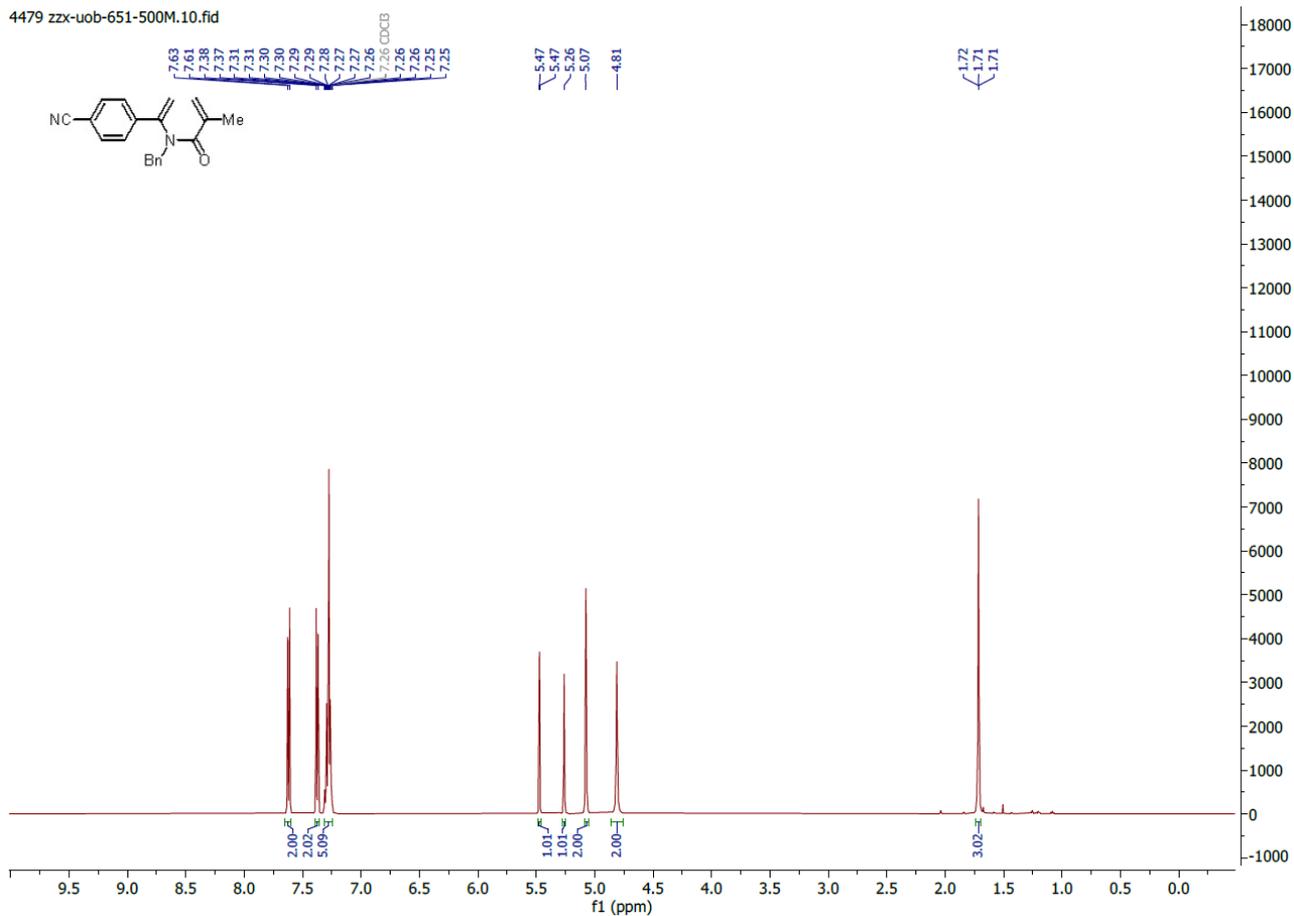


7874 zzx-uob-1066-sec-500M.14.fid

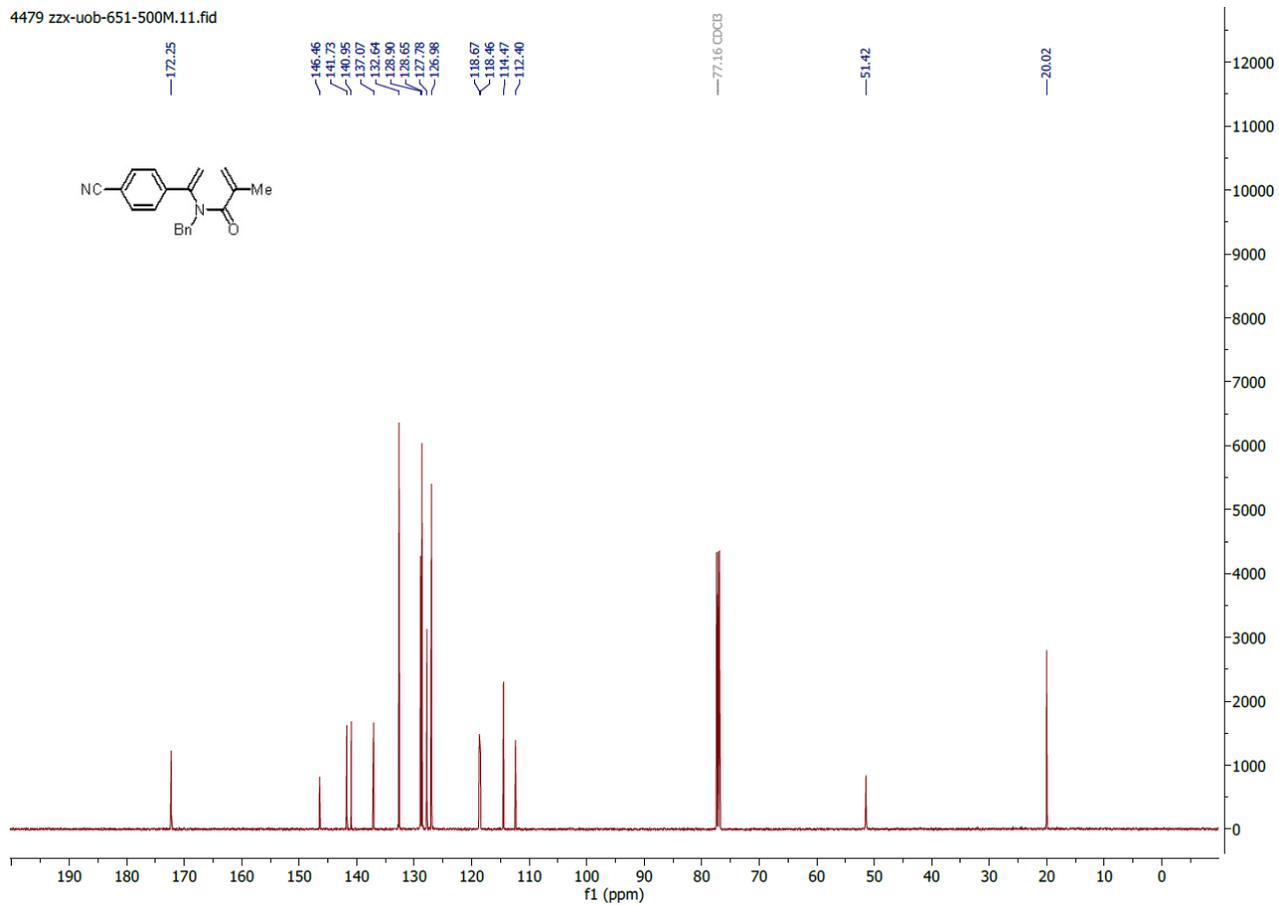


Compound 1d

4479 zzx-uob-651-500M.10.fid

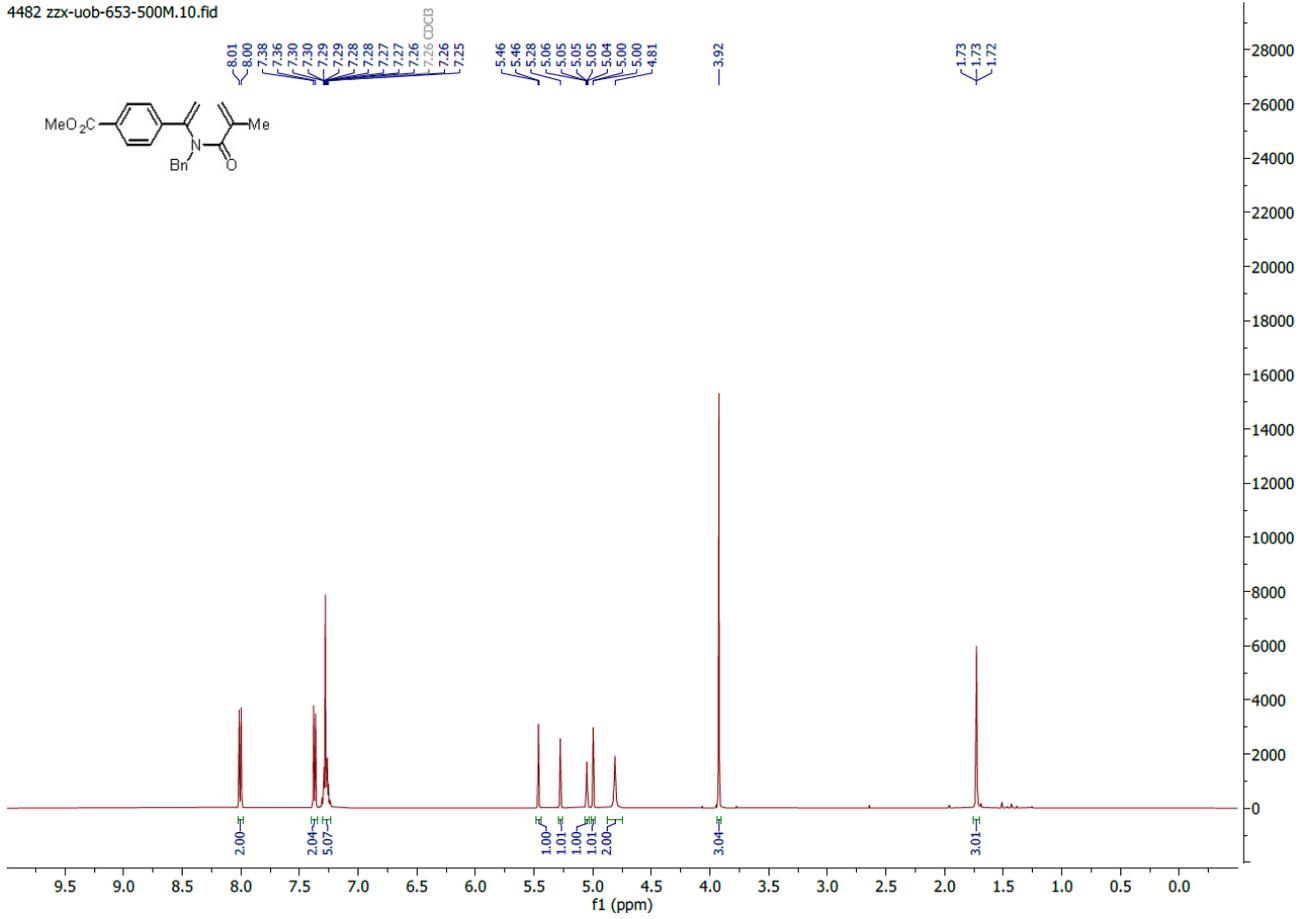


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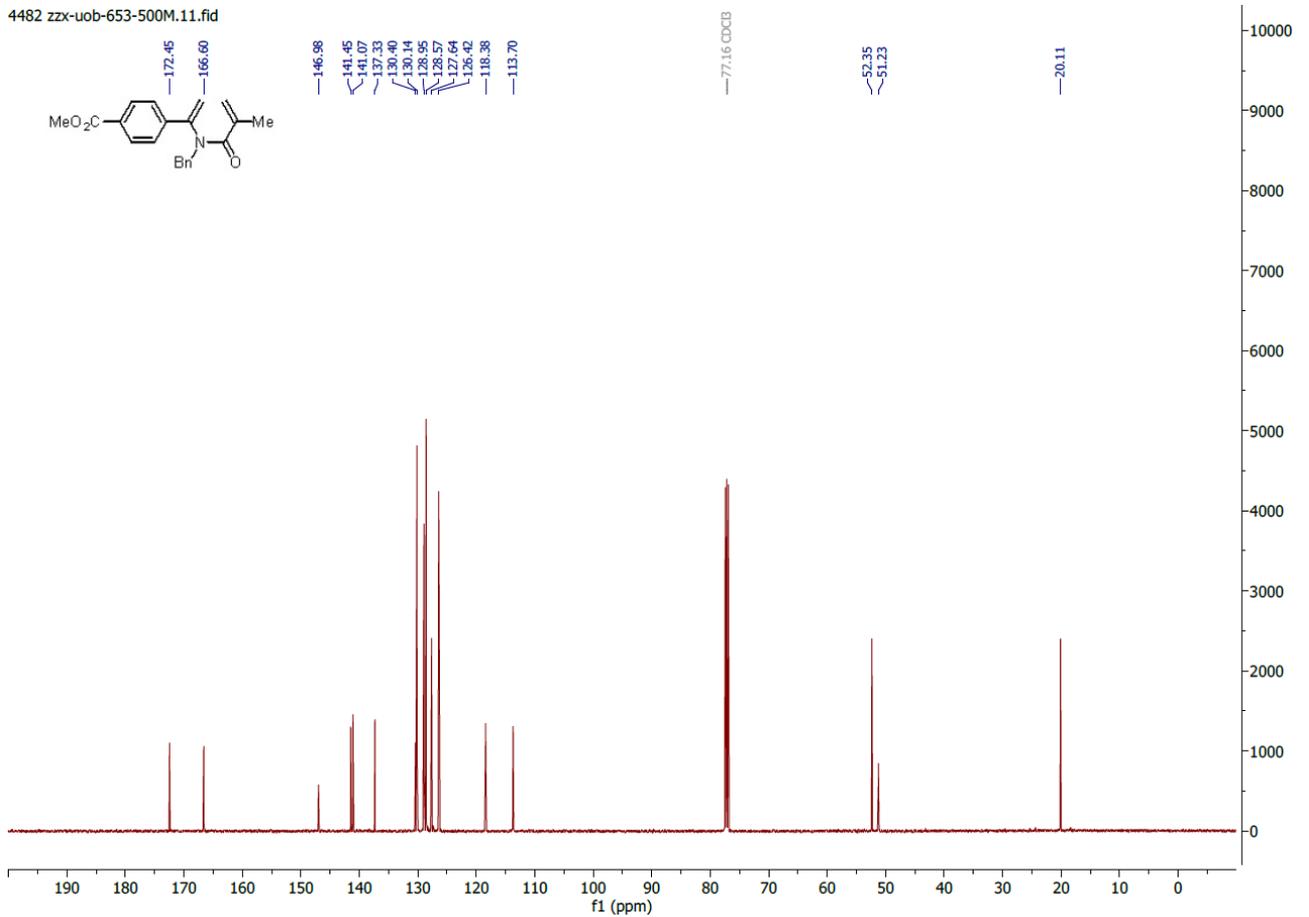


Compound 1e

4482 zzx-uob-653-500M.10.fid

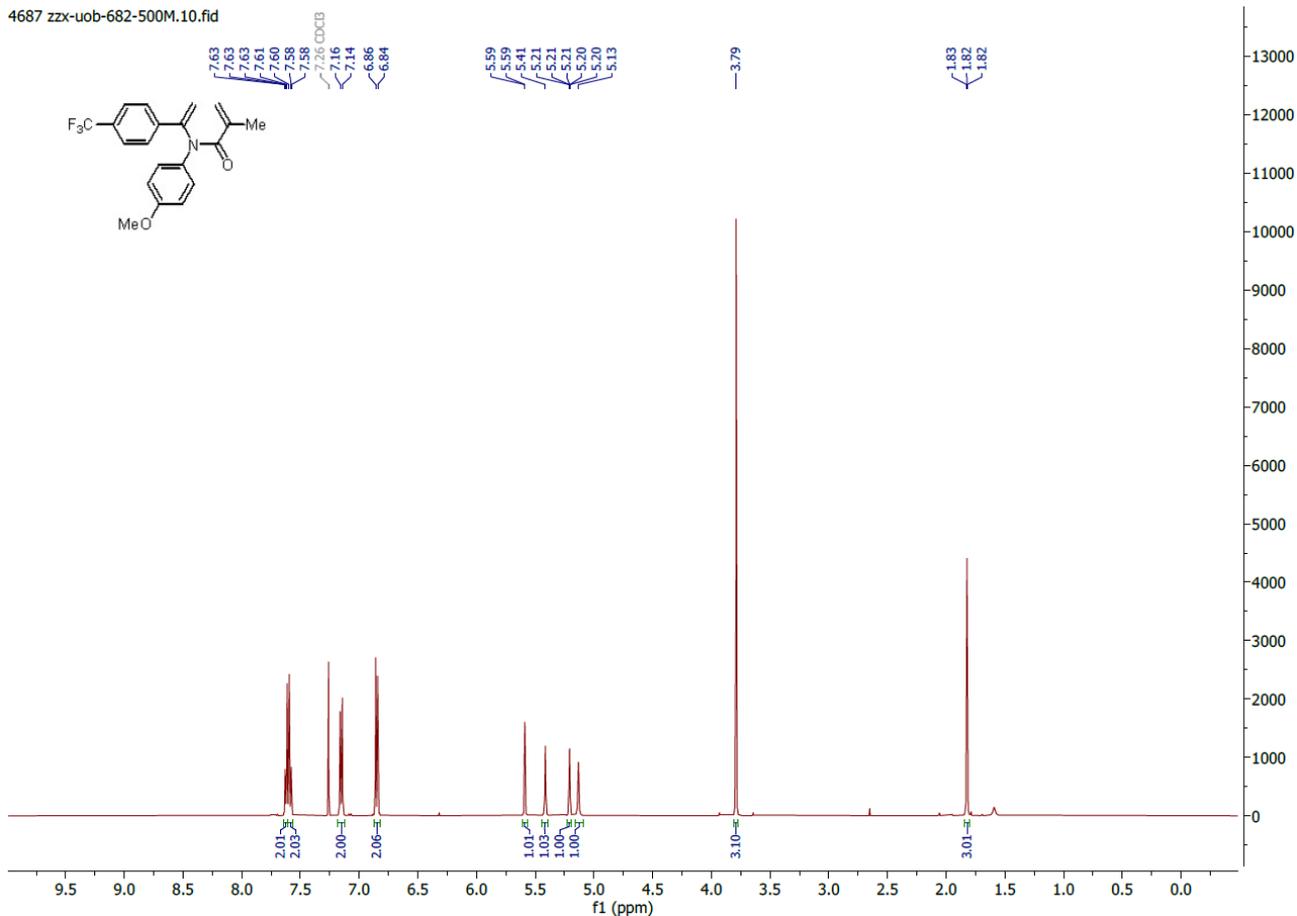


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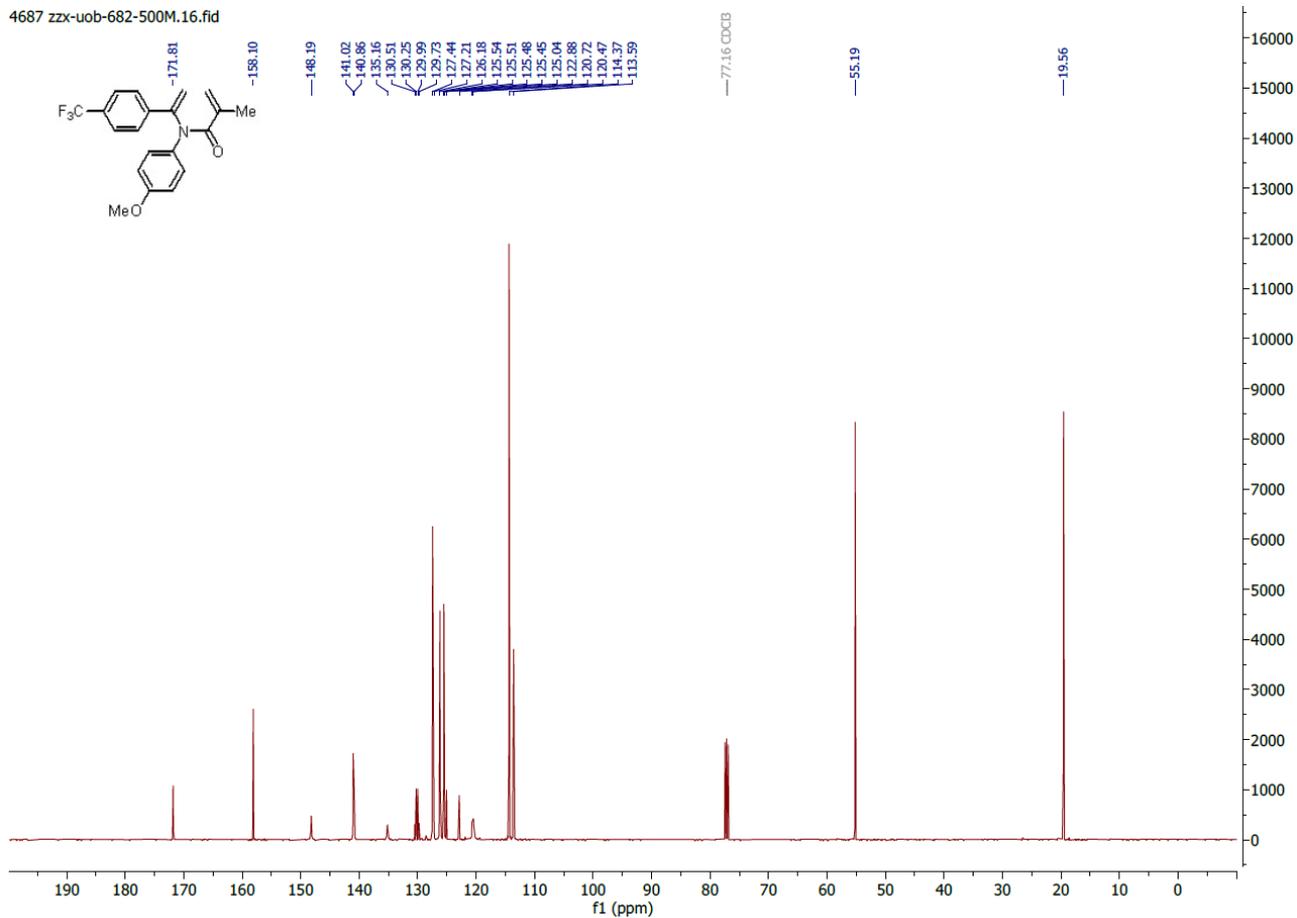


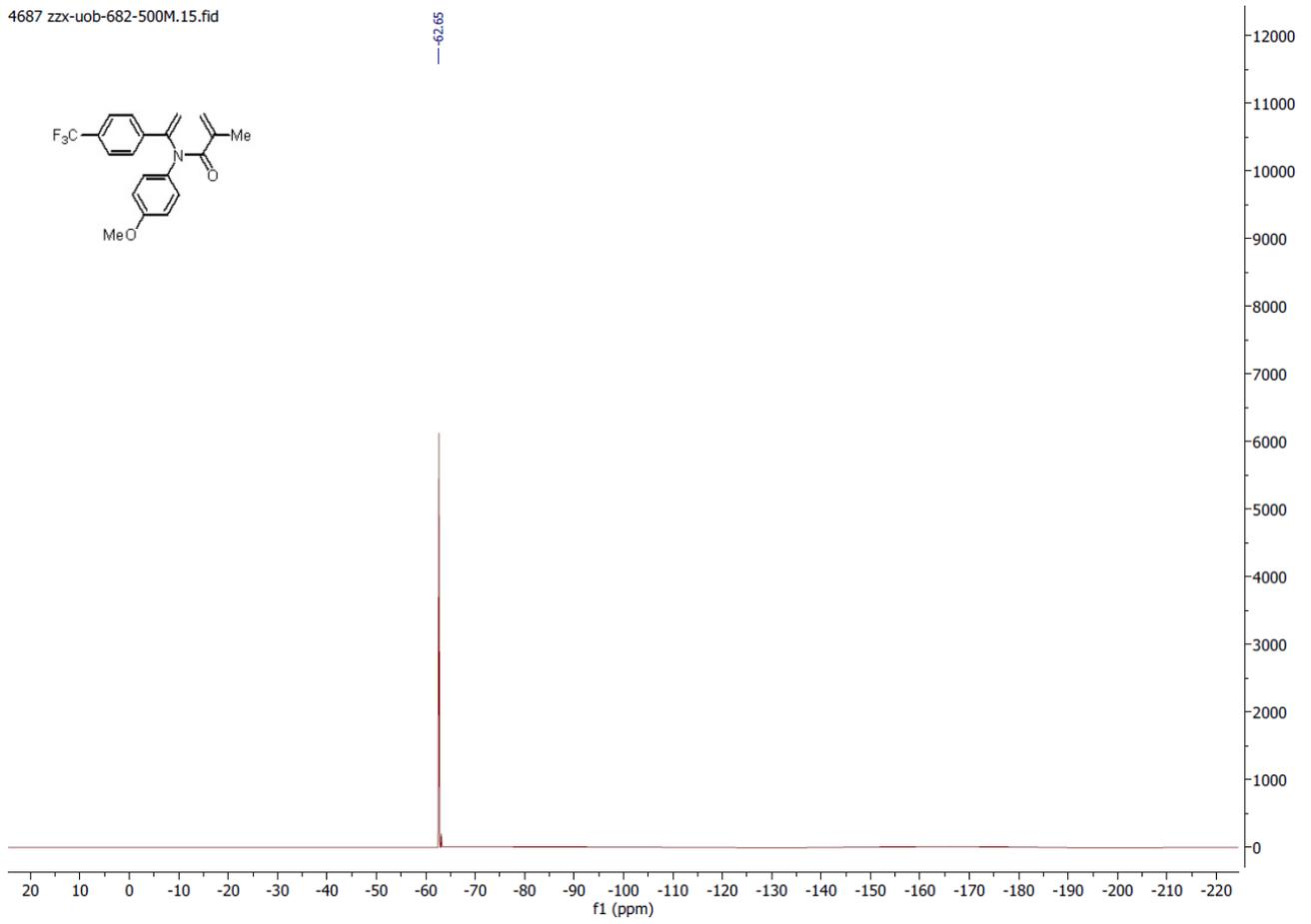
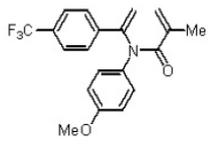
Compound 1j

4687 zzx-uob-682-500M.10.fid



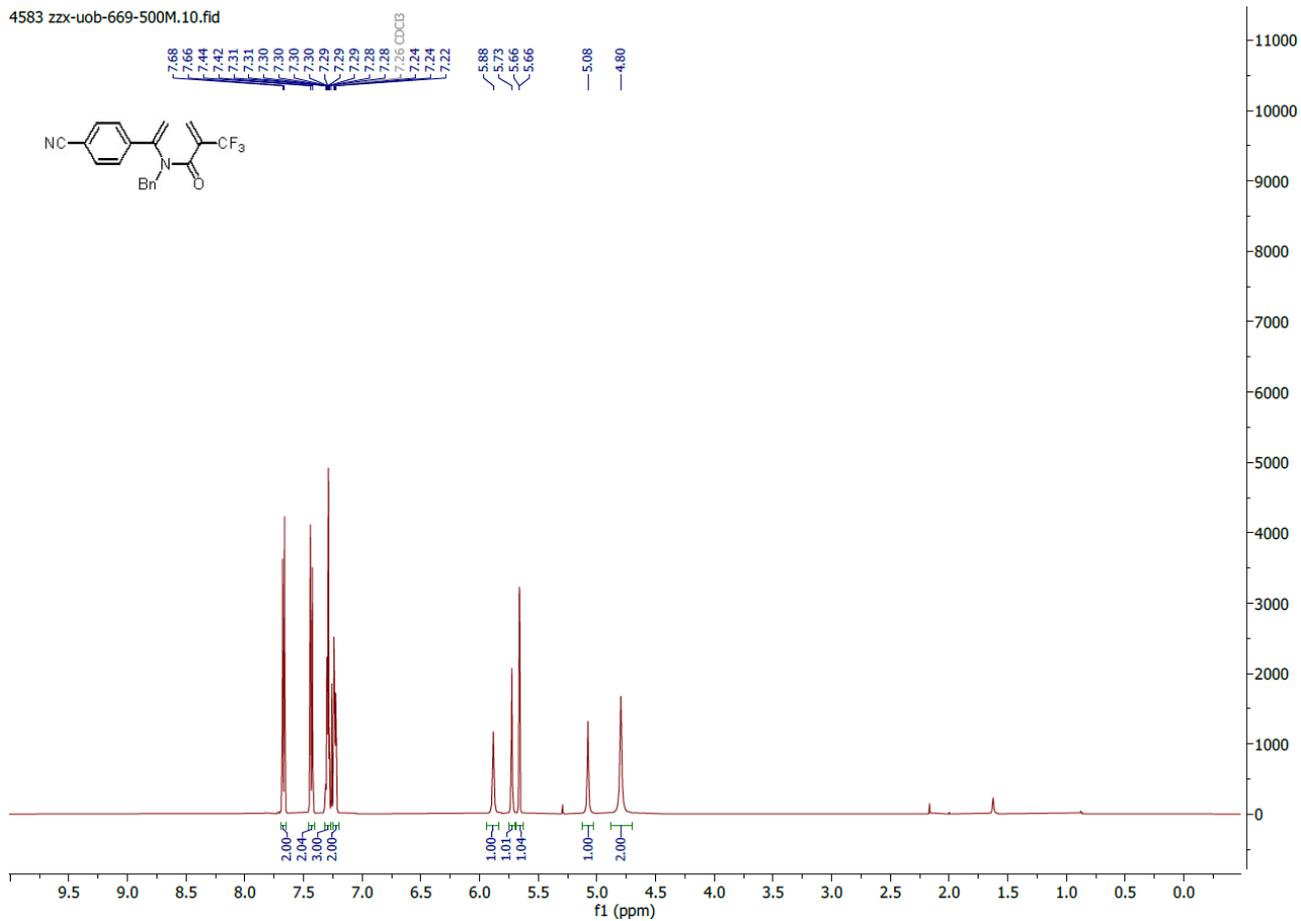
4687 zzx-uob-682-500M.16.fid



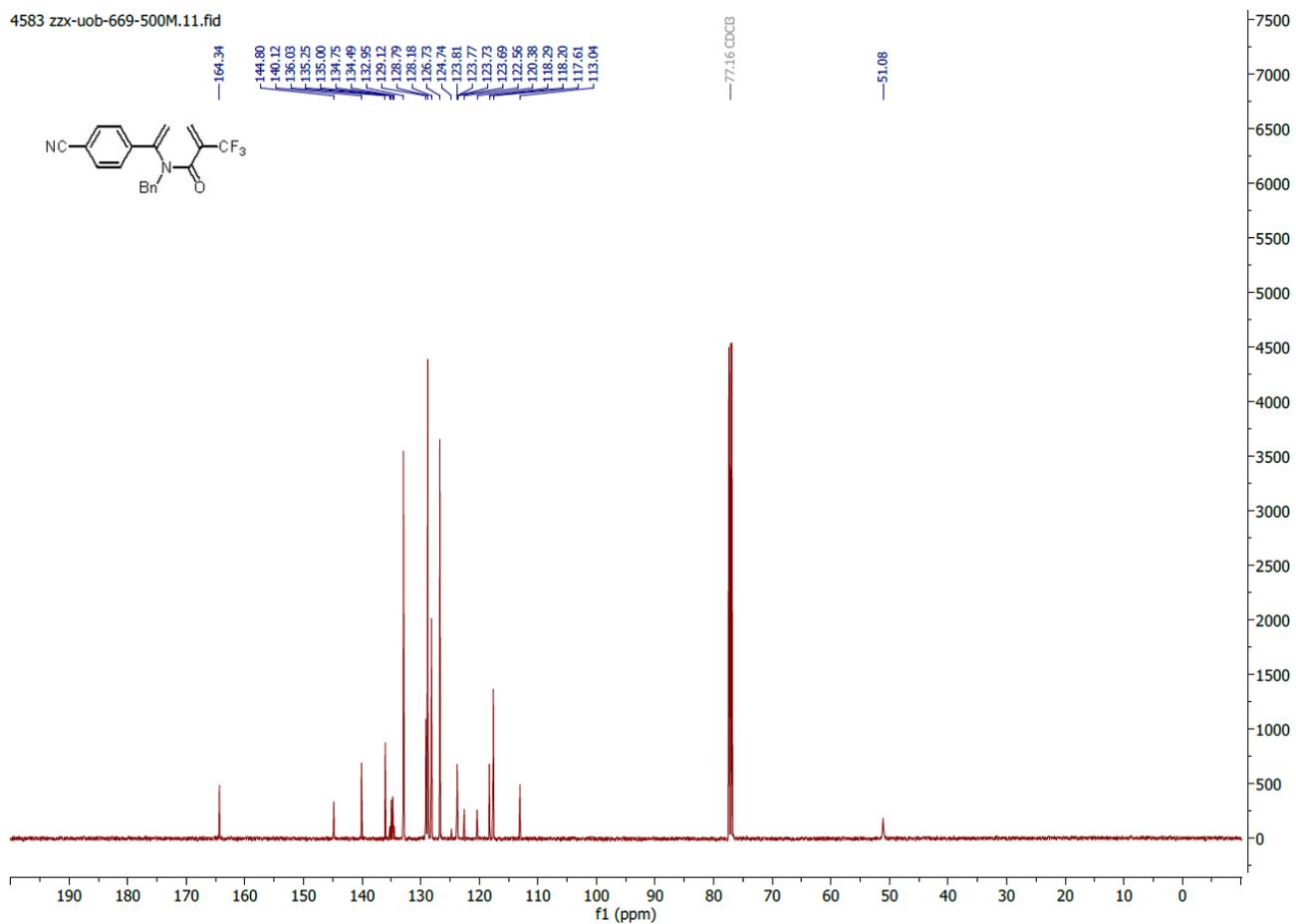


Compound 1k

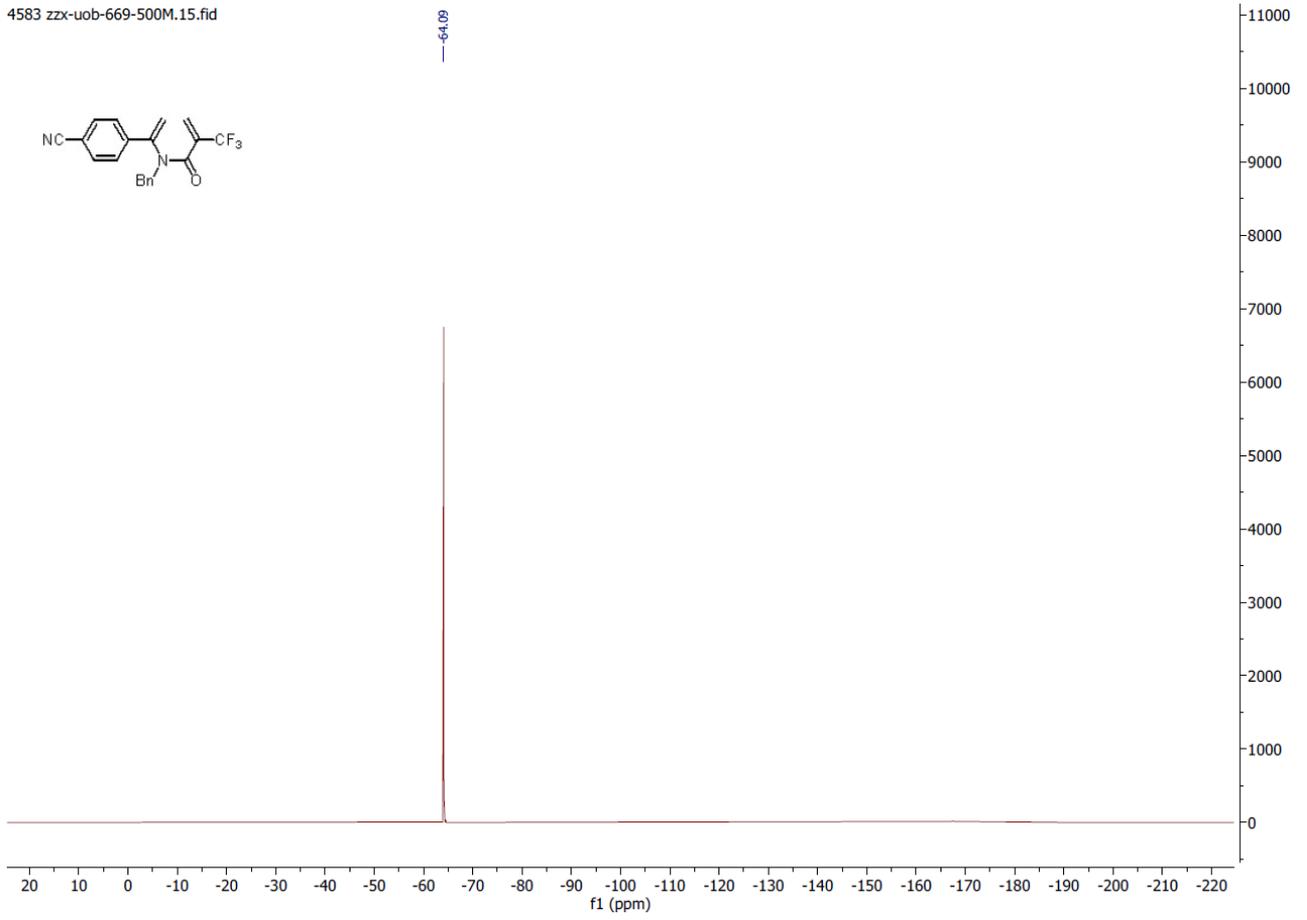
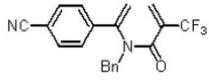
4583 zzx-uob-669-500M.10.fid



4583 zzx-uob-669-500M.11.fid

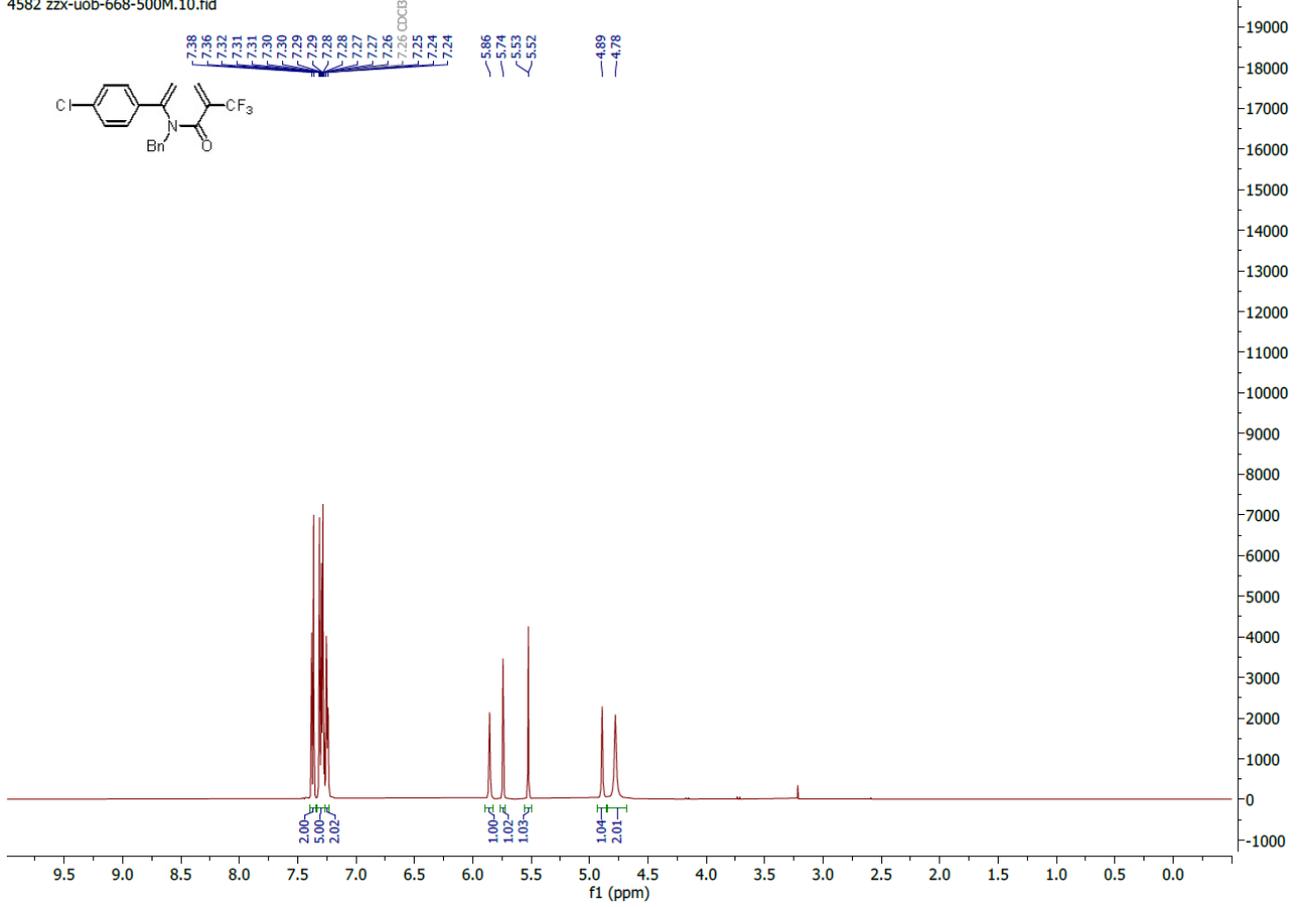
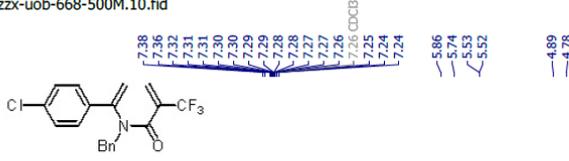


4583 zzx-uob-669-500M.15.fid

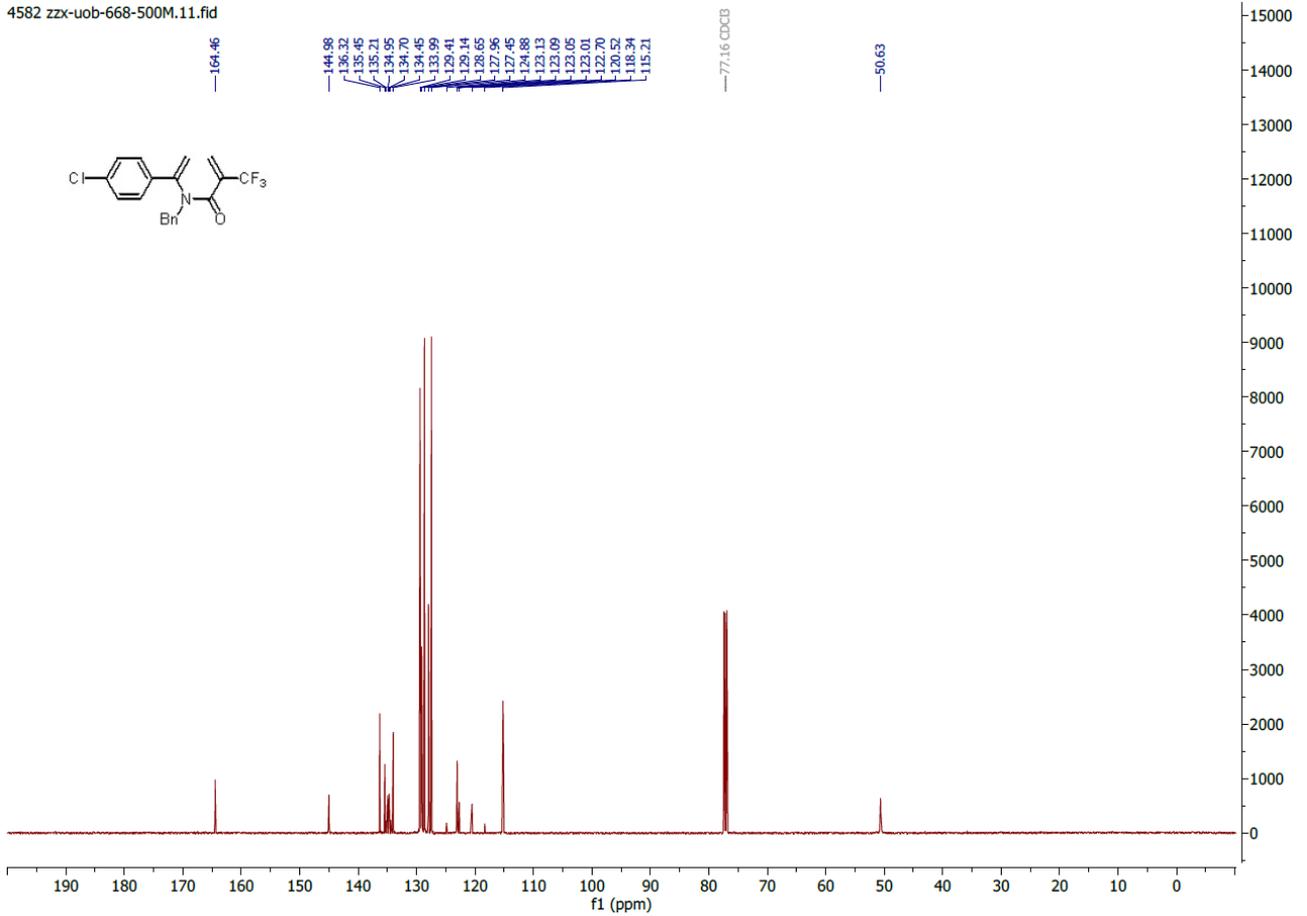
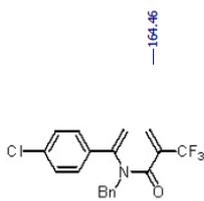


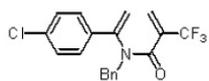
Compound 11

4582 zzx-uob-668-500M.10.fid

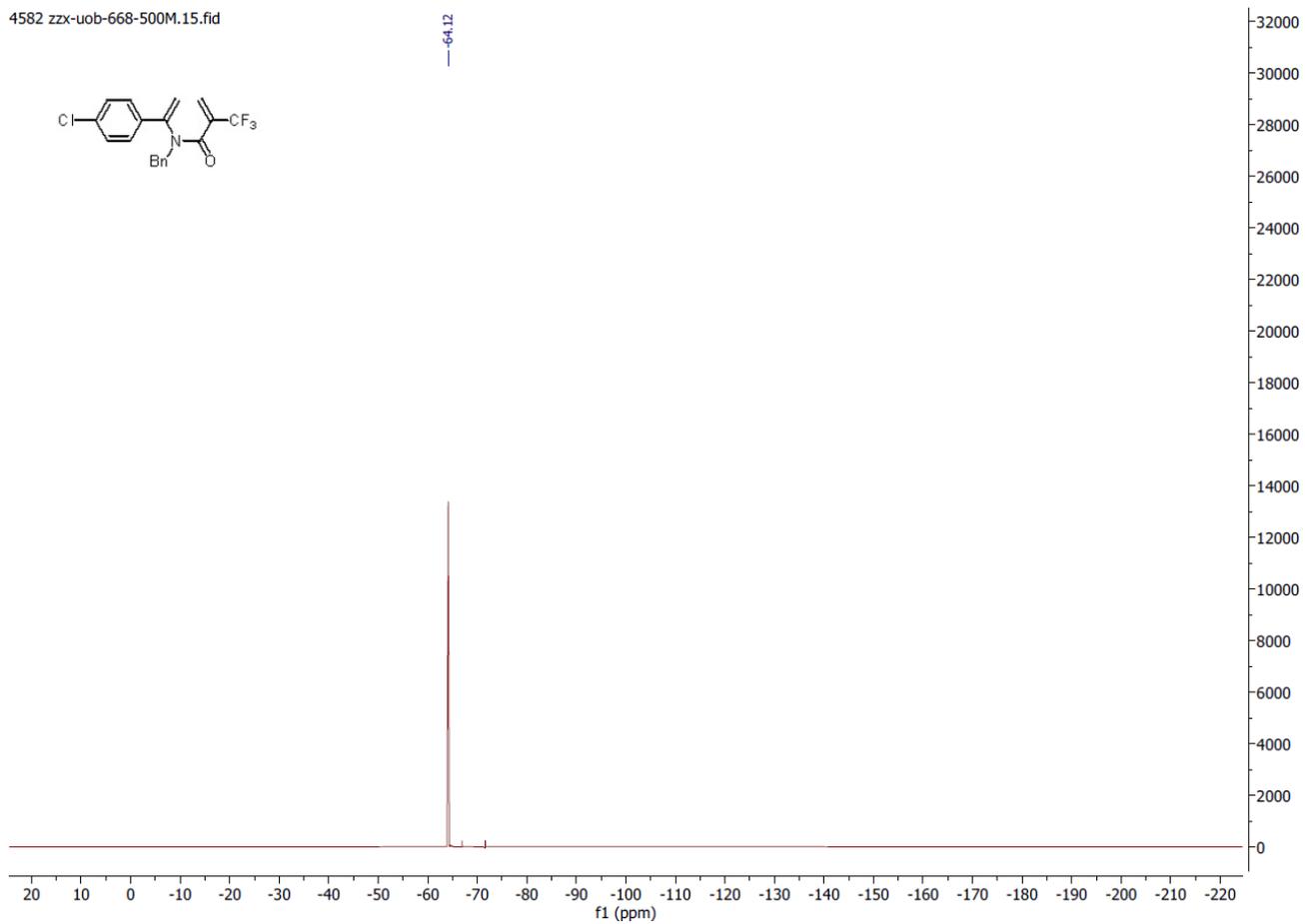


4582 zzx-uob-668-500M.11.fid



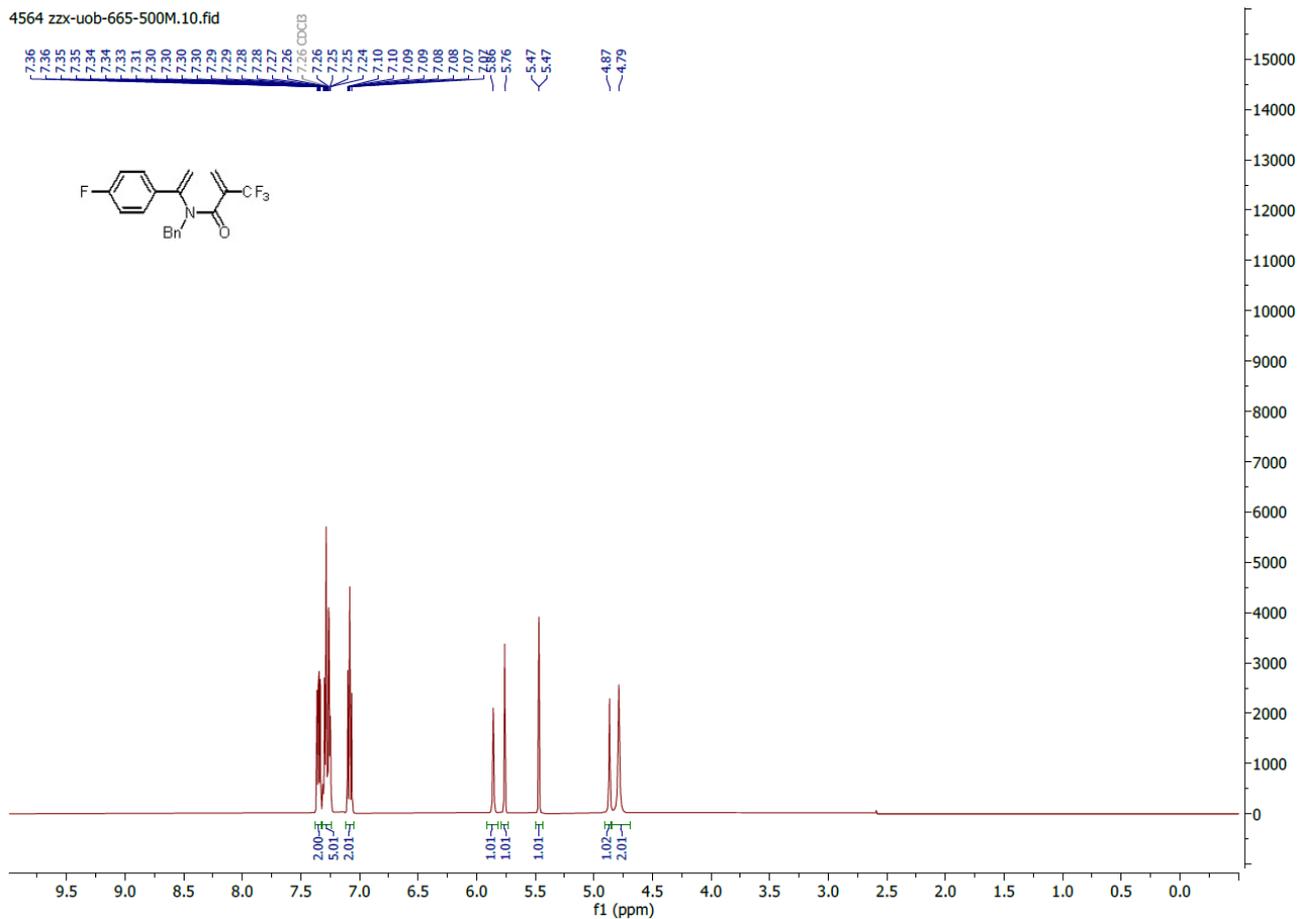


64.12

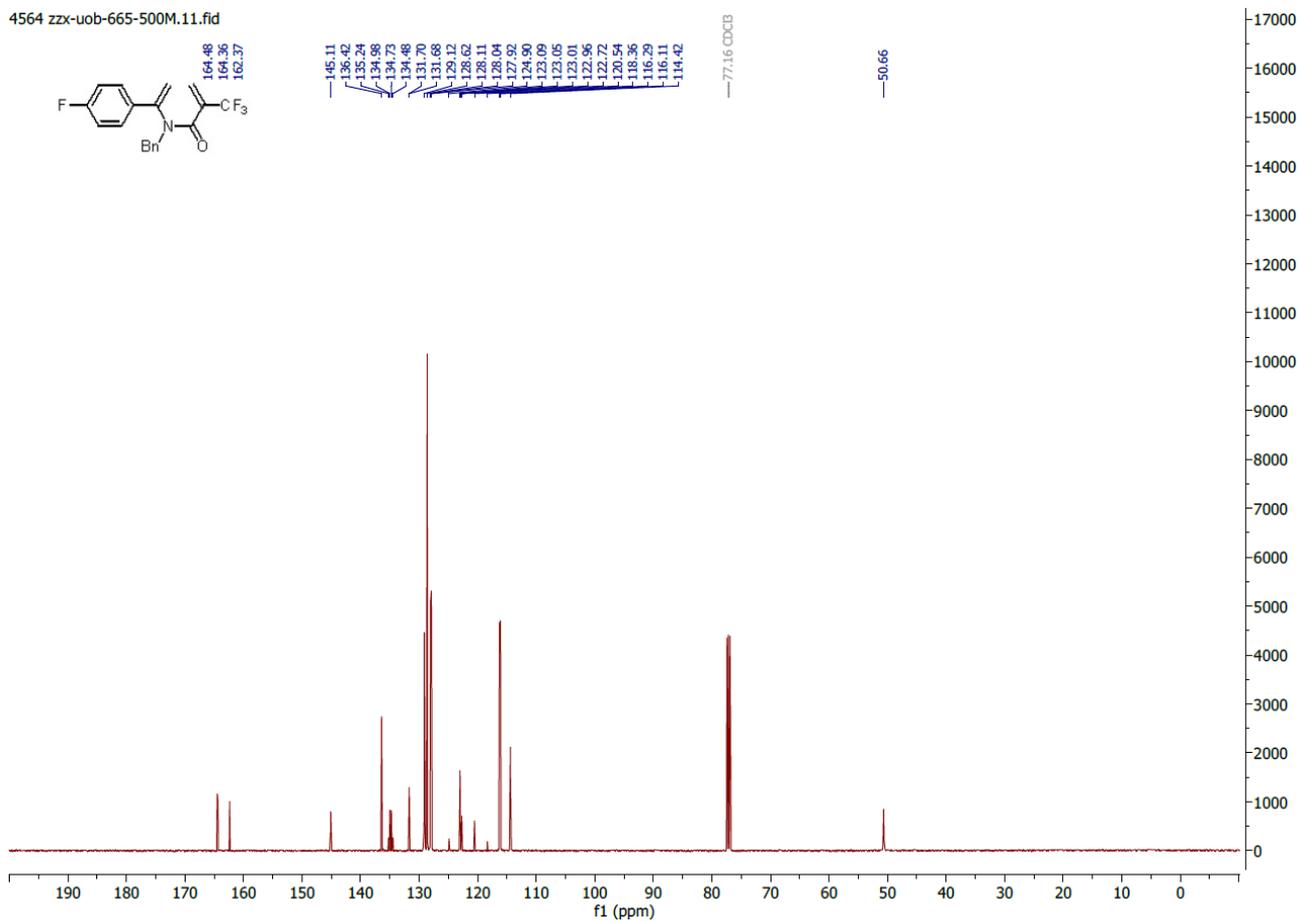


Compound 1m

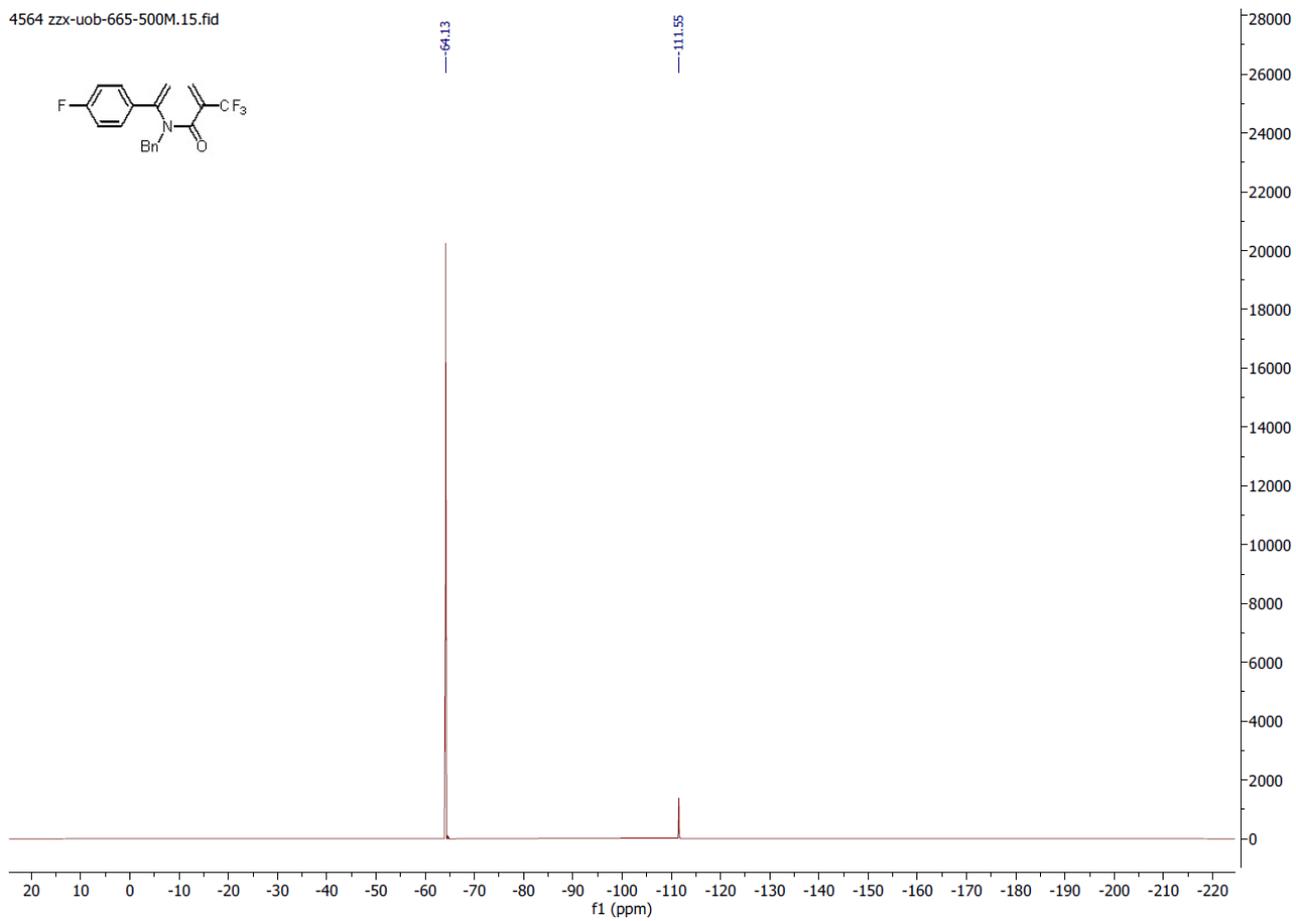
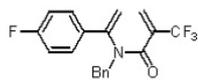
4564 zzx-uob-665-500M.10.fid



4564 zzx-uob-665-500M.11.fid

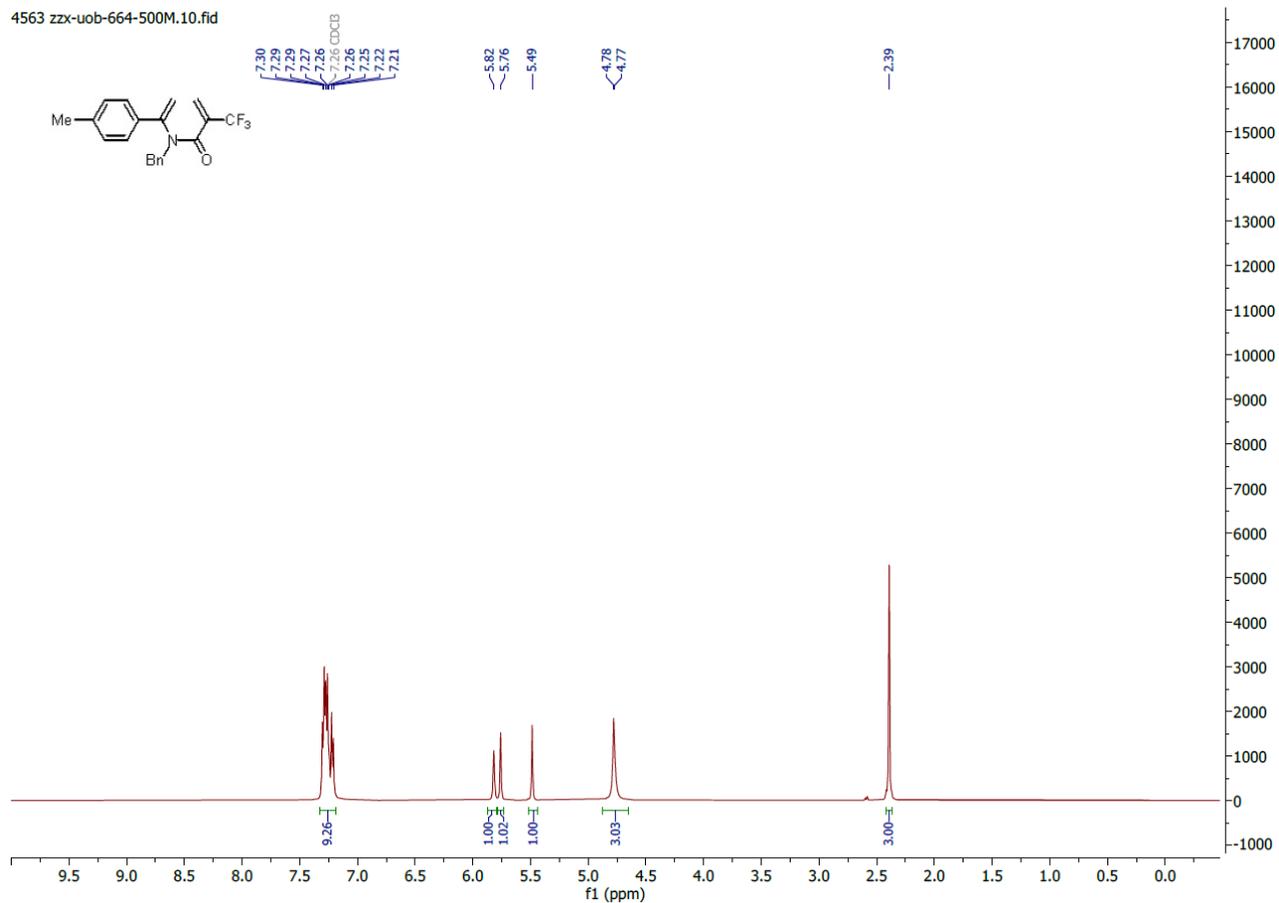


4564 zzx-uob-665-500M.15.fid

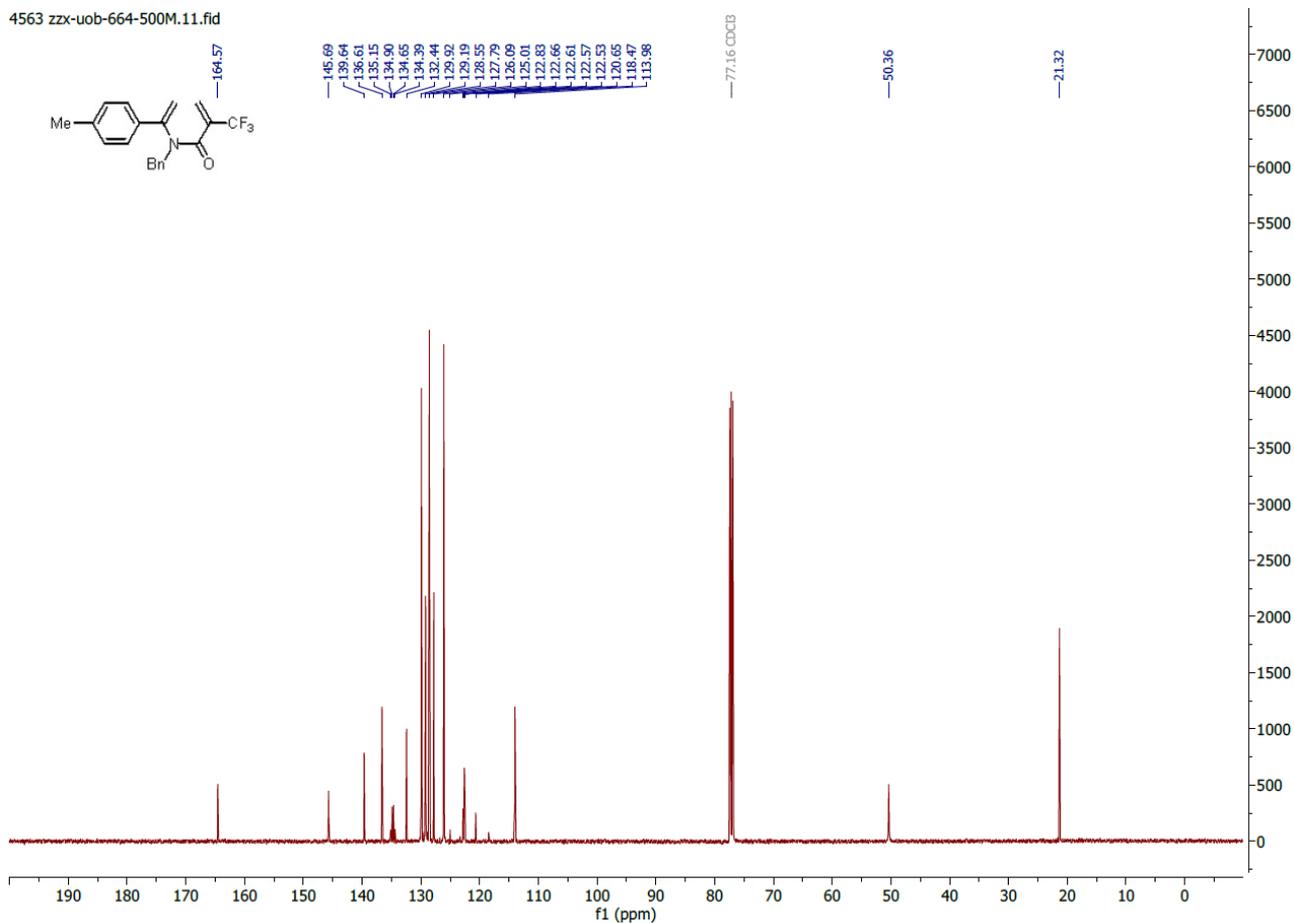


Compound 1o

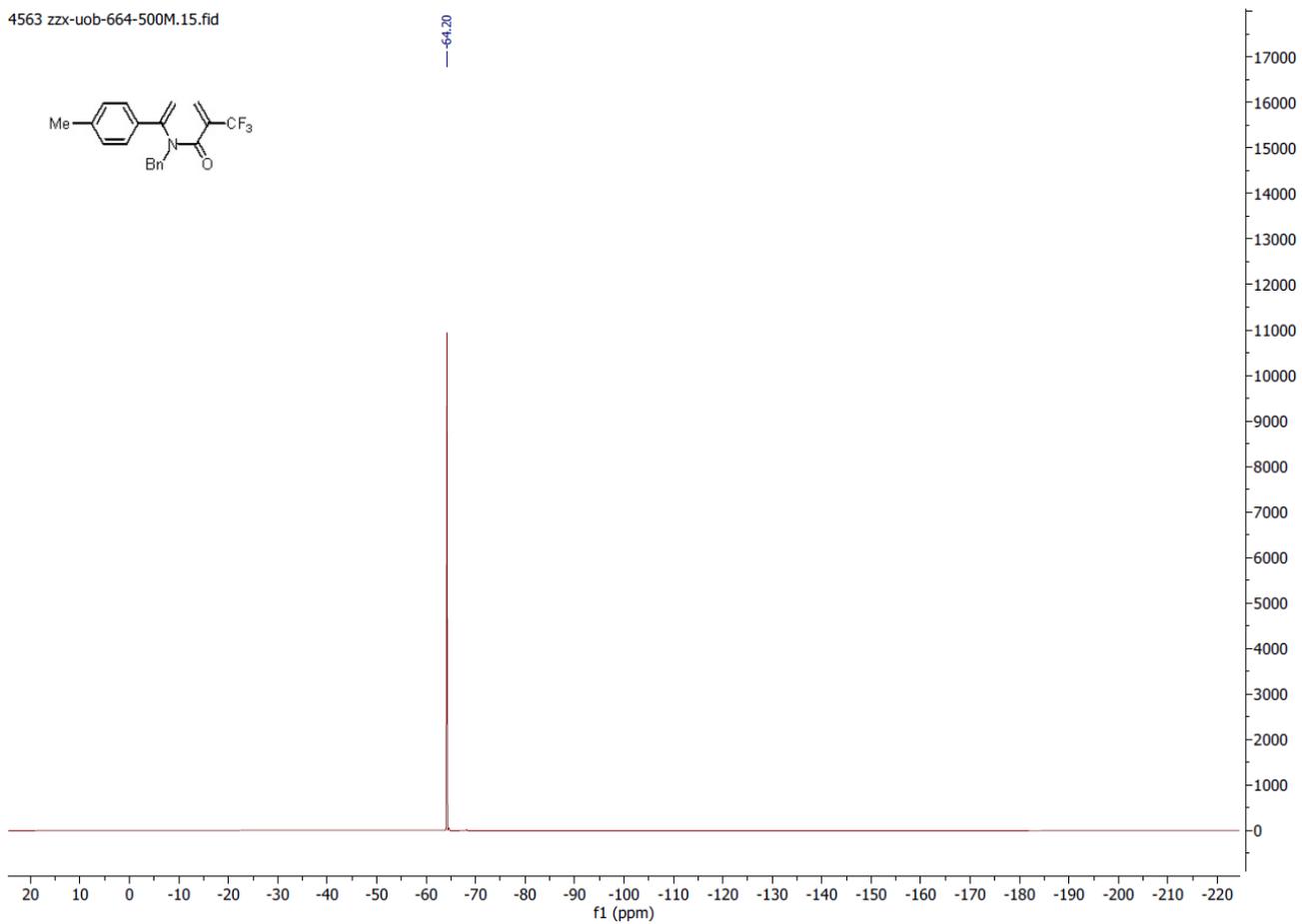
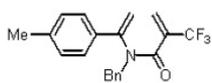
4563 zzx-uob-664-500M.10.fid



4563 zzx-uob-664-500M.11.fid

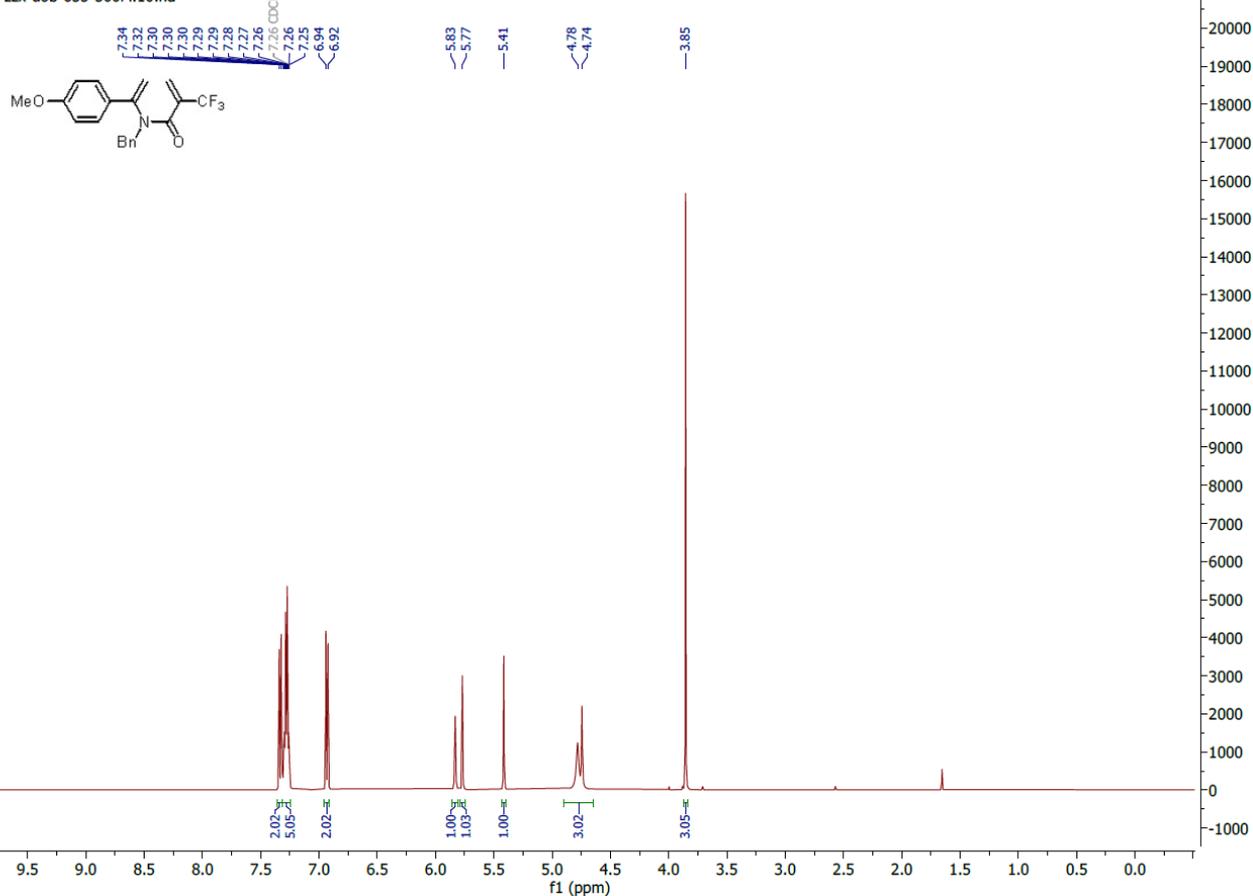


4563 zzx-uob-664-500M.15.fid

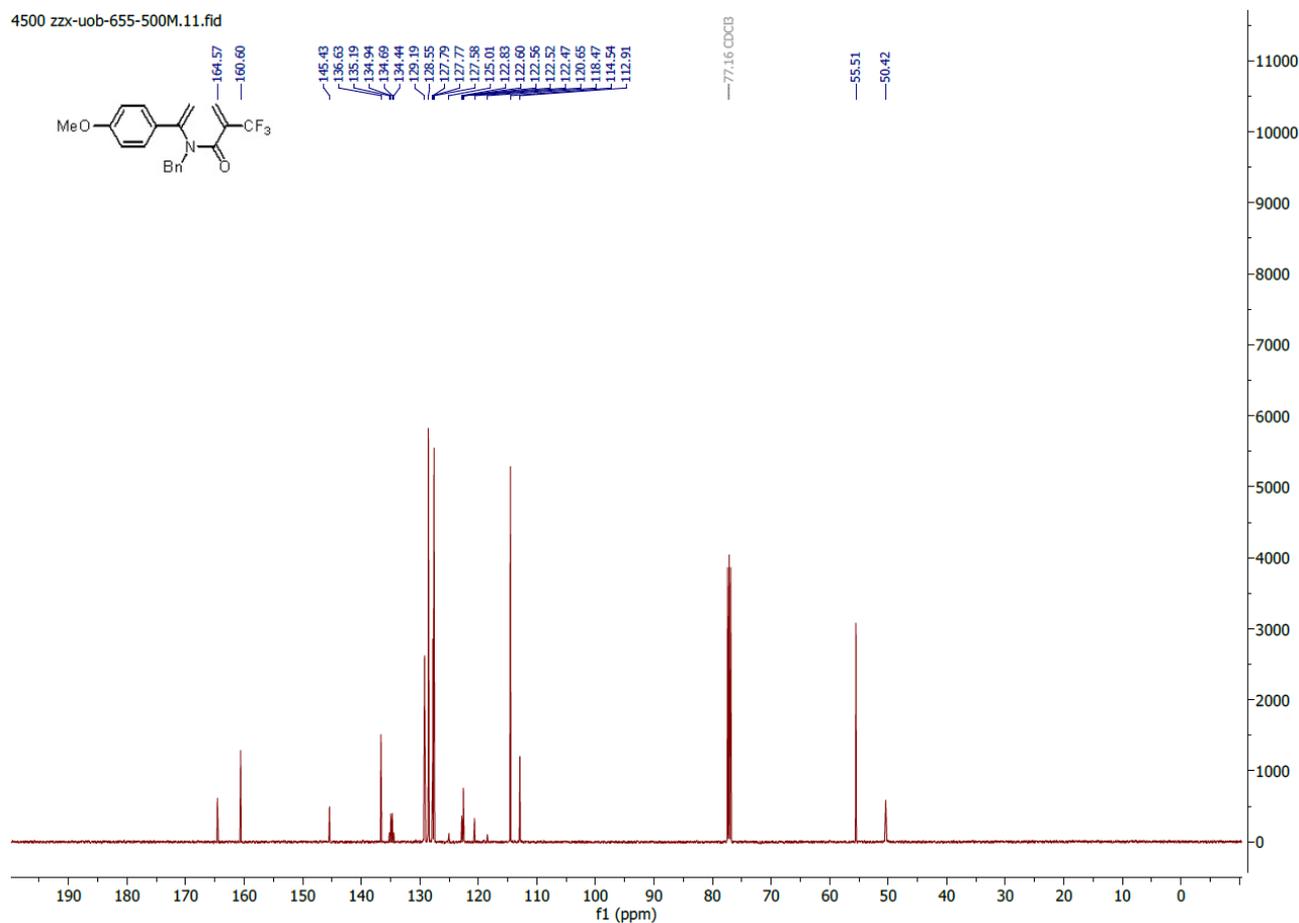


Compound 1p

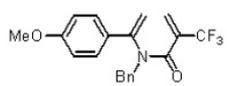
4500 zzx-uob-655-500M.10.fid



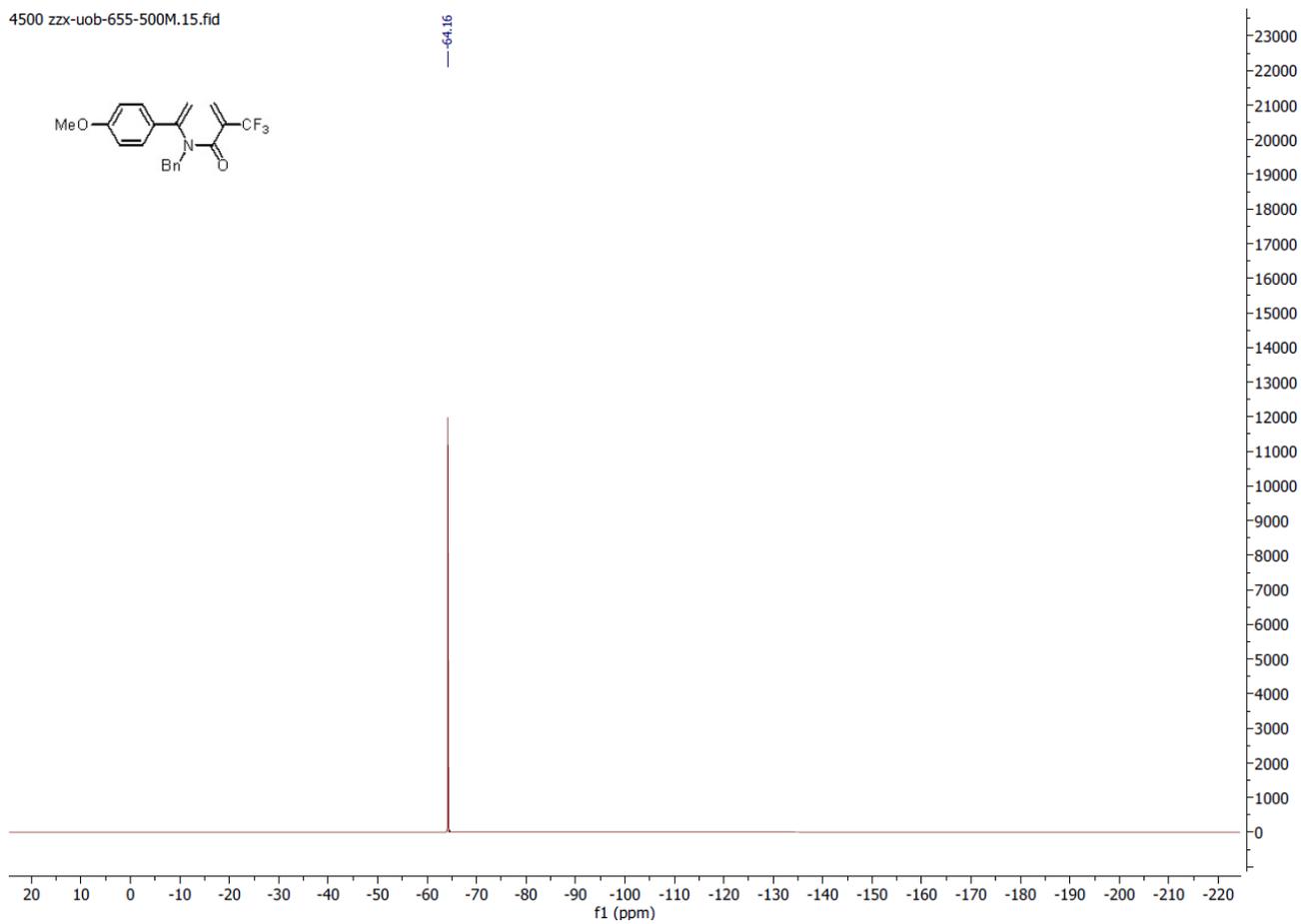
4500 zzx-uob-655-500M.11.fid



4500 zzx-uob-655-500M.15.fid

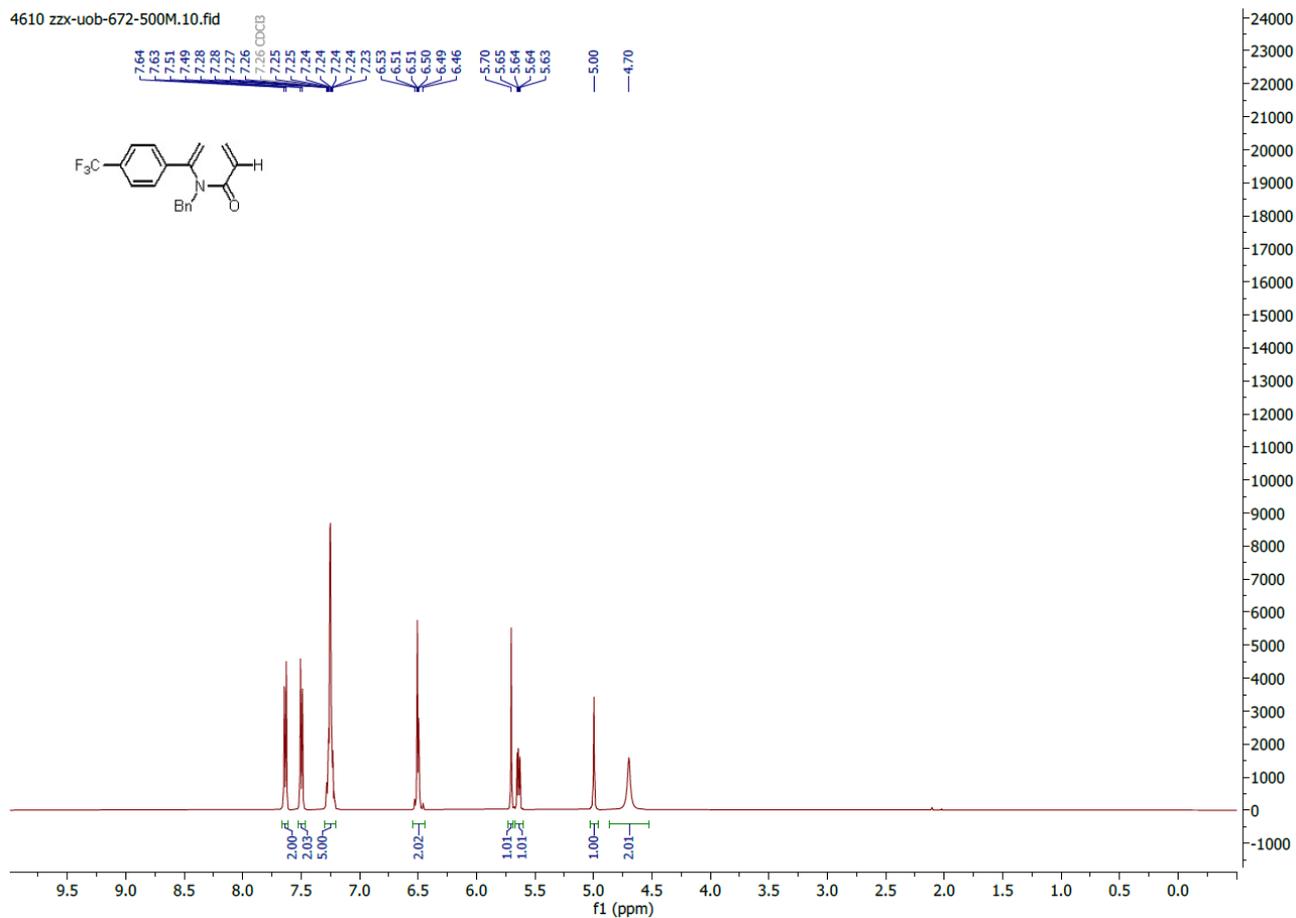
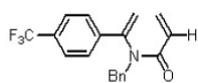


-64.16

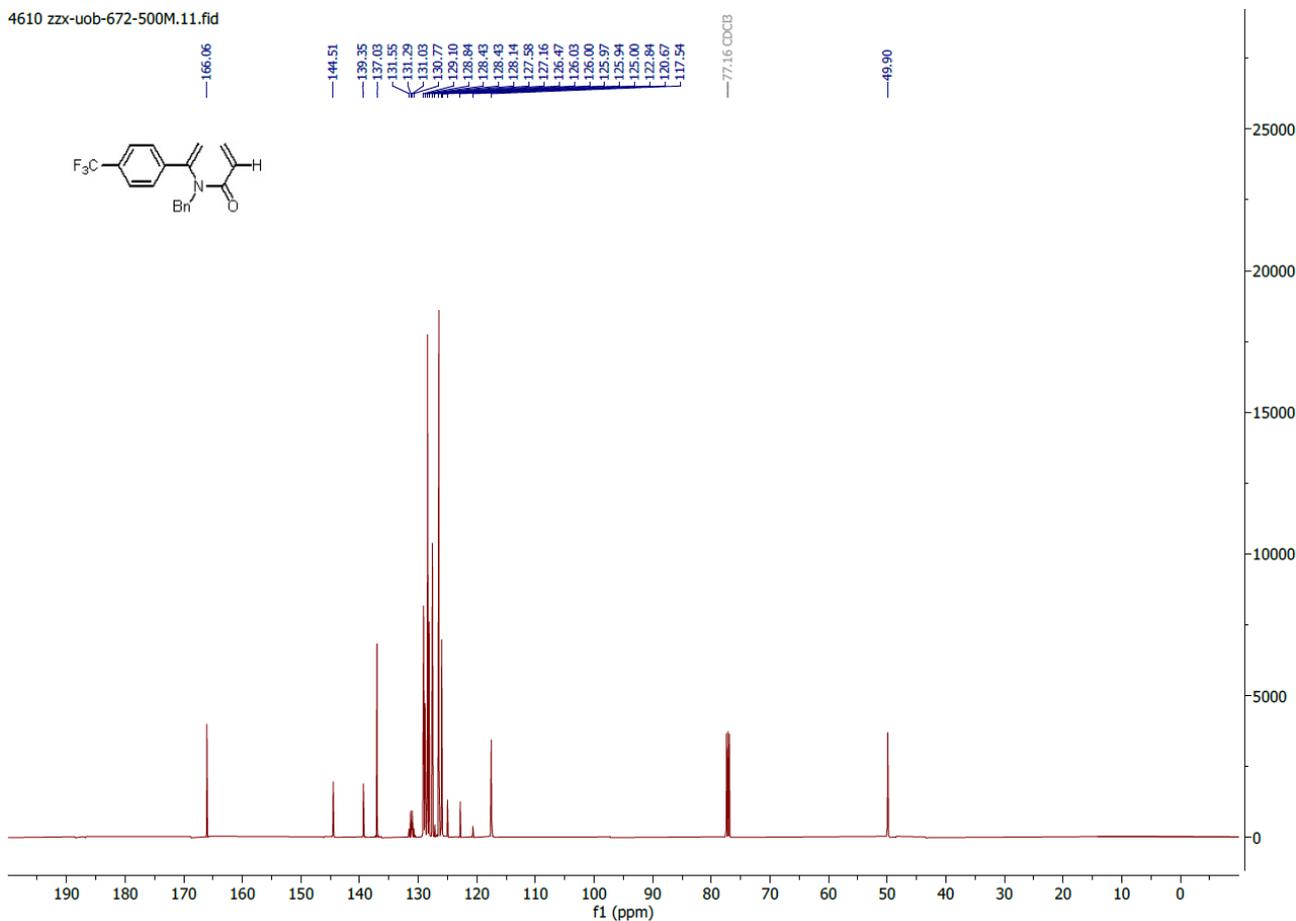
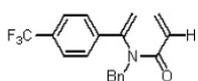


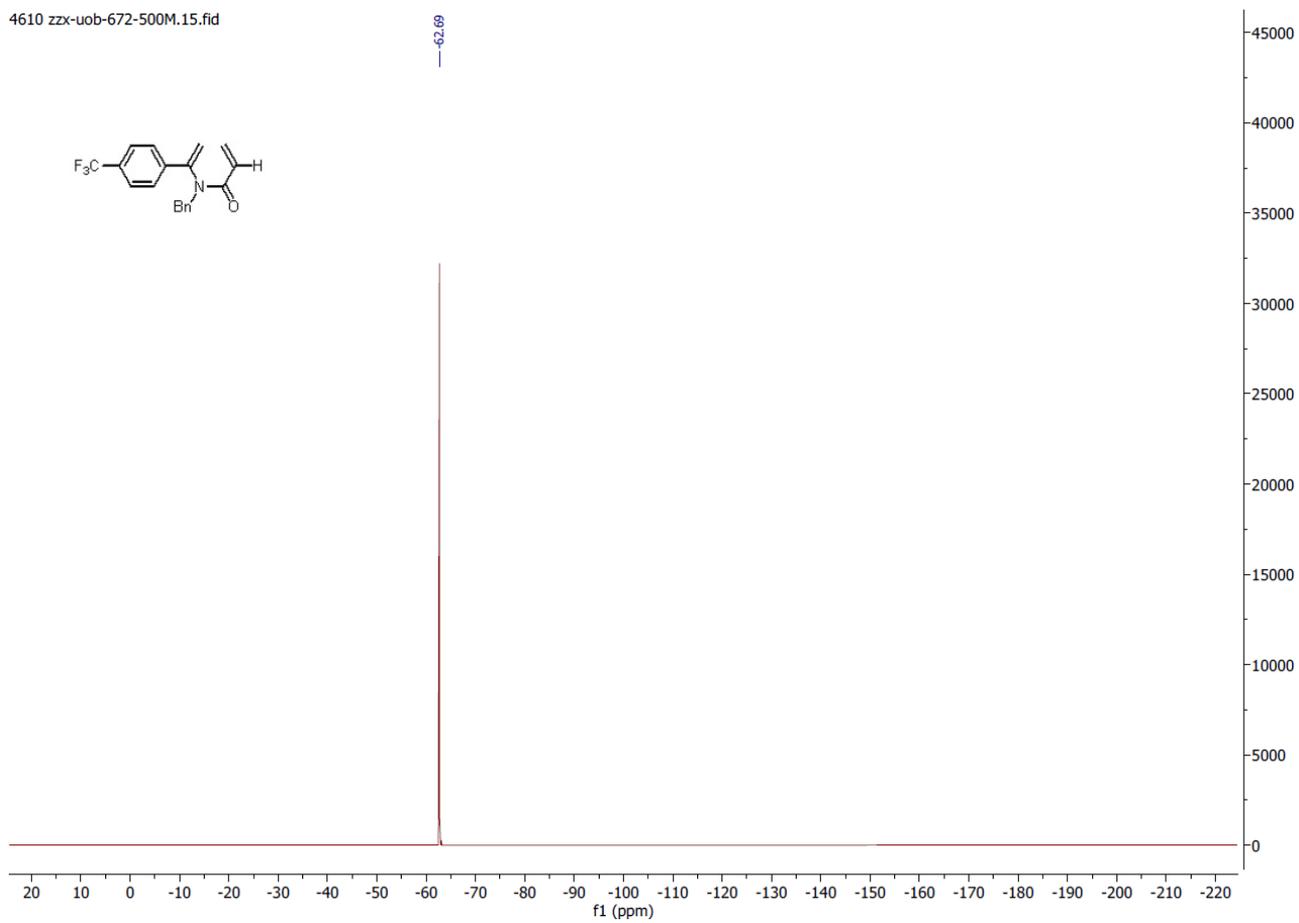
Compound 1q

4610 zzx-uob-672-500M.10.fid



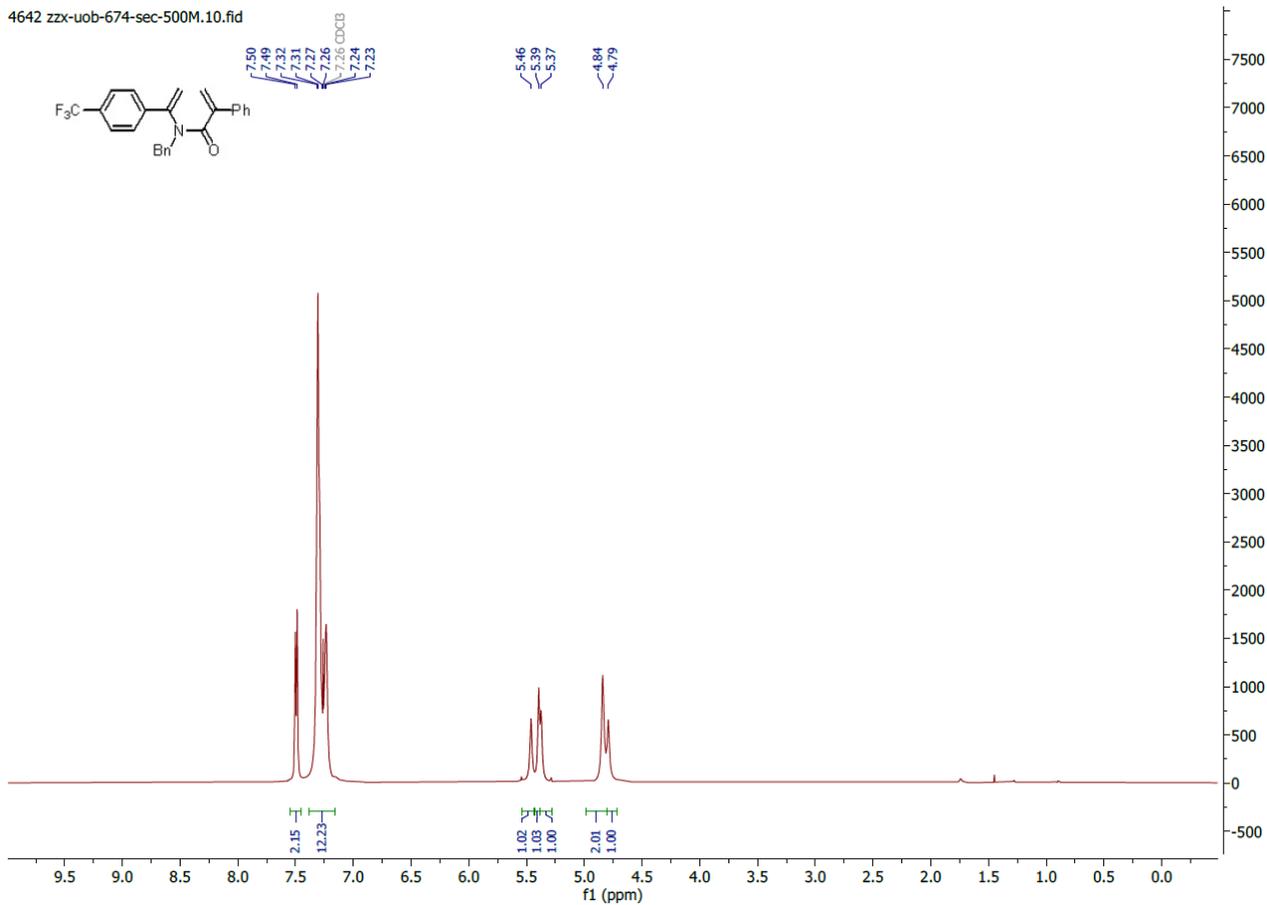
4610 zzx-uob-672-500M.11.fid



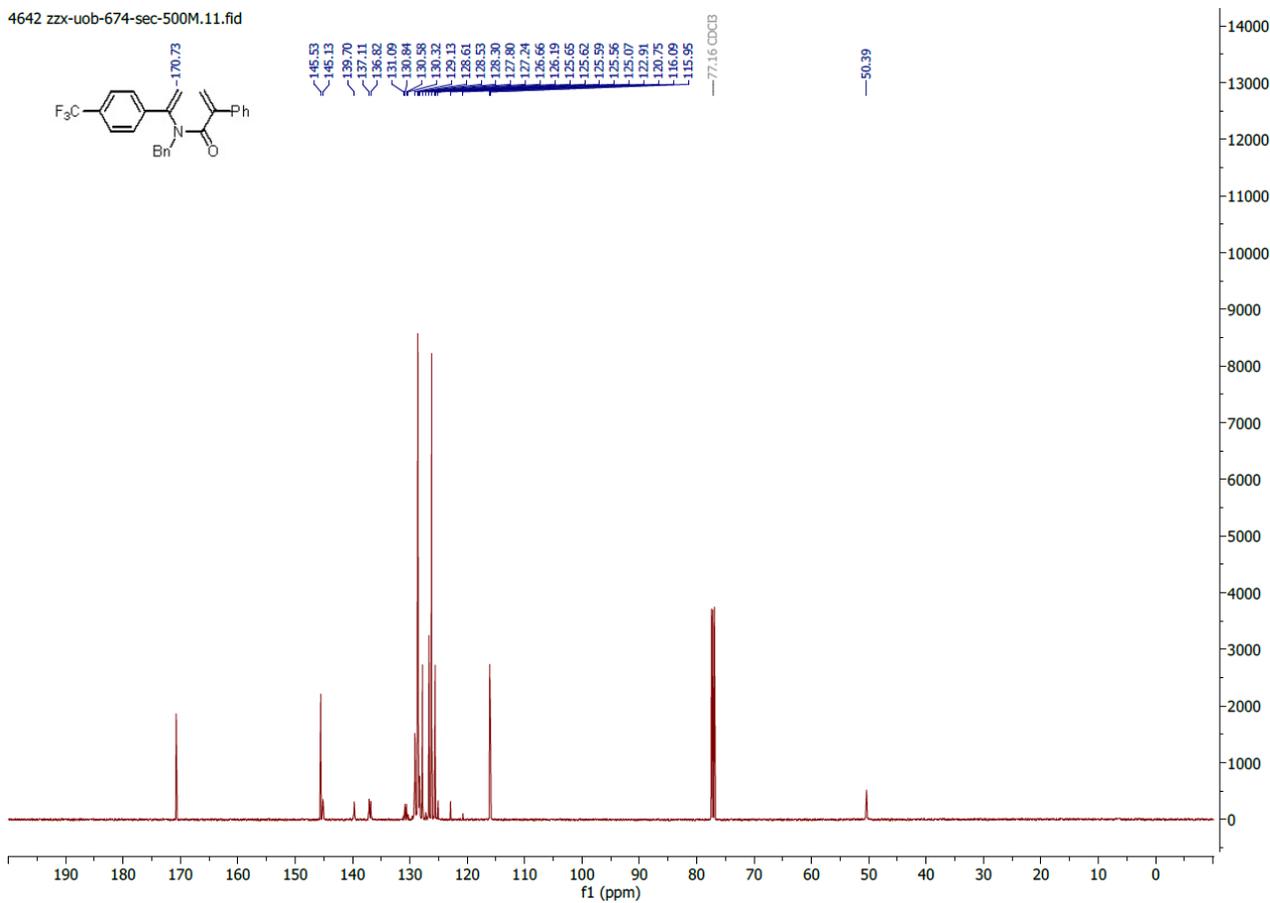


Compound 1r

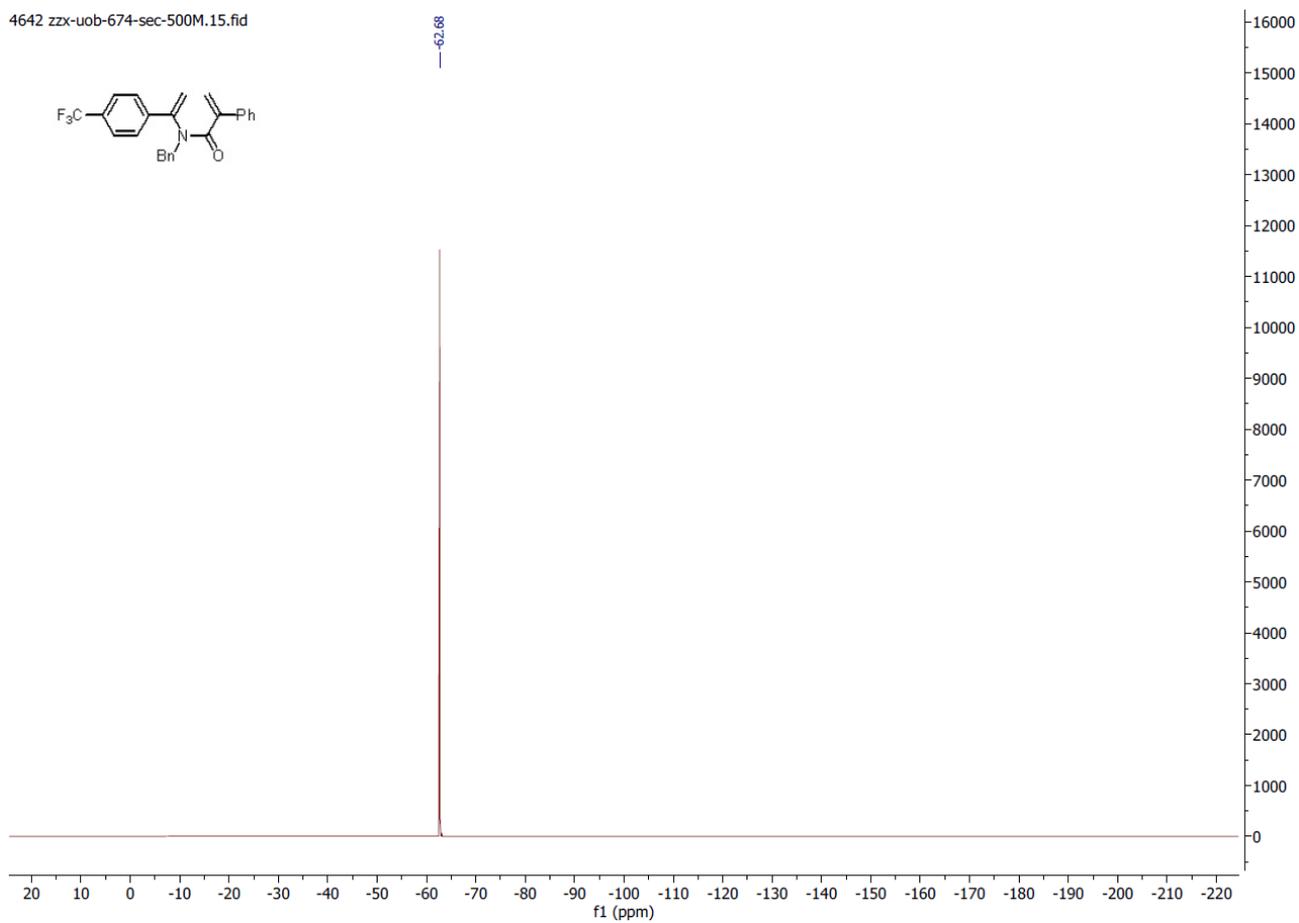
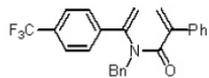
4642 zzx-uob-674-sec-500M.10.fid



4642 zzx-uob-674-sec-500M.11.fid



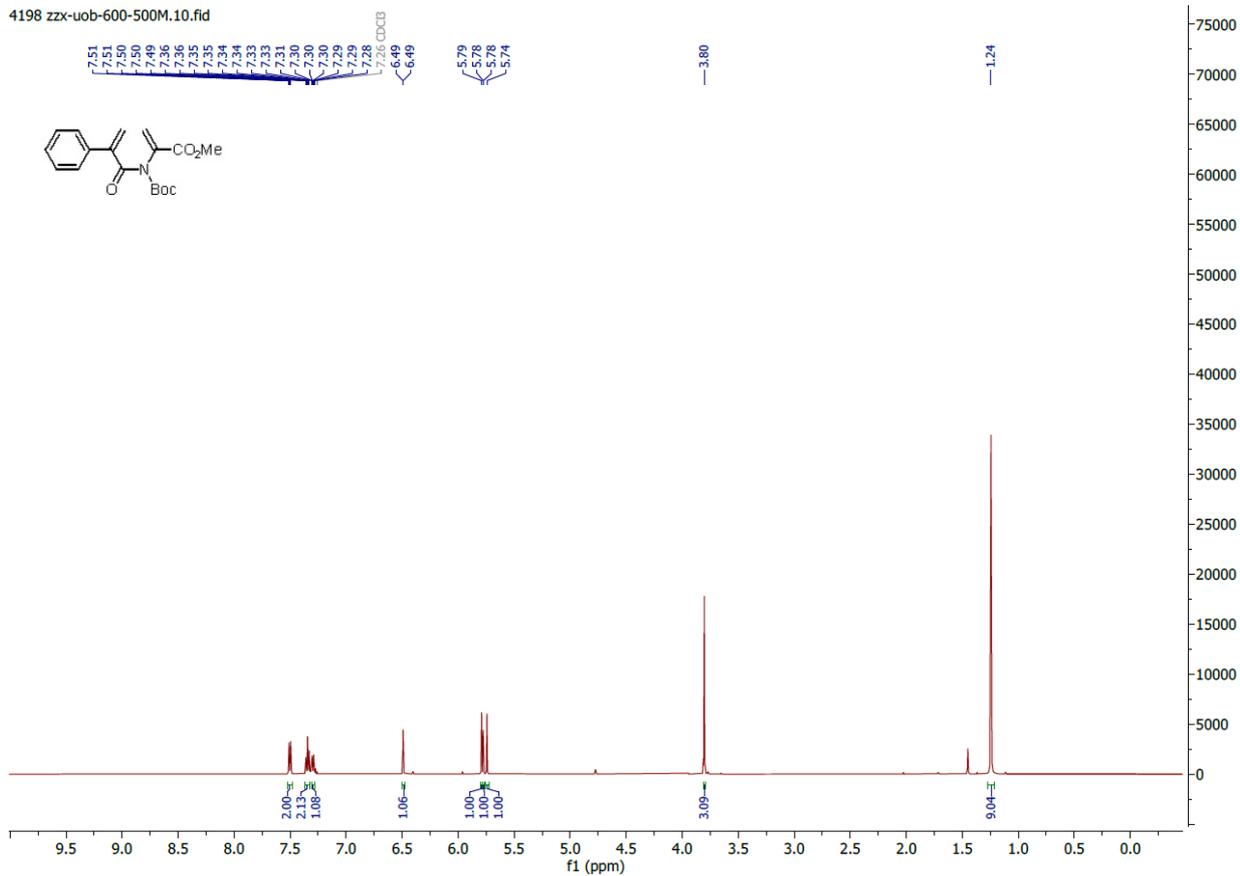
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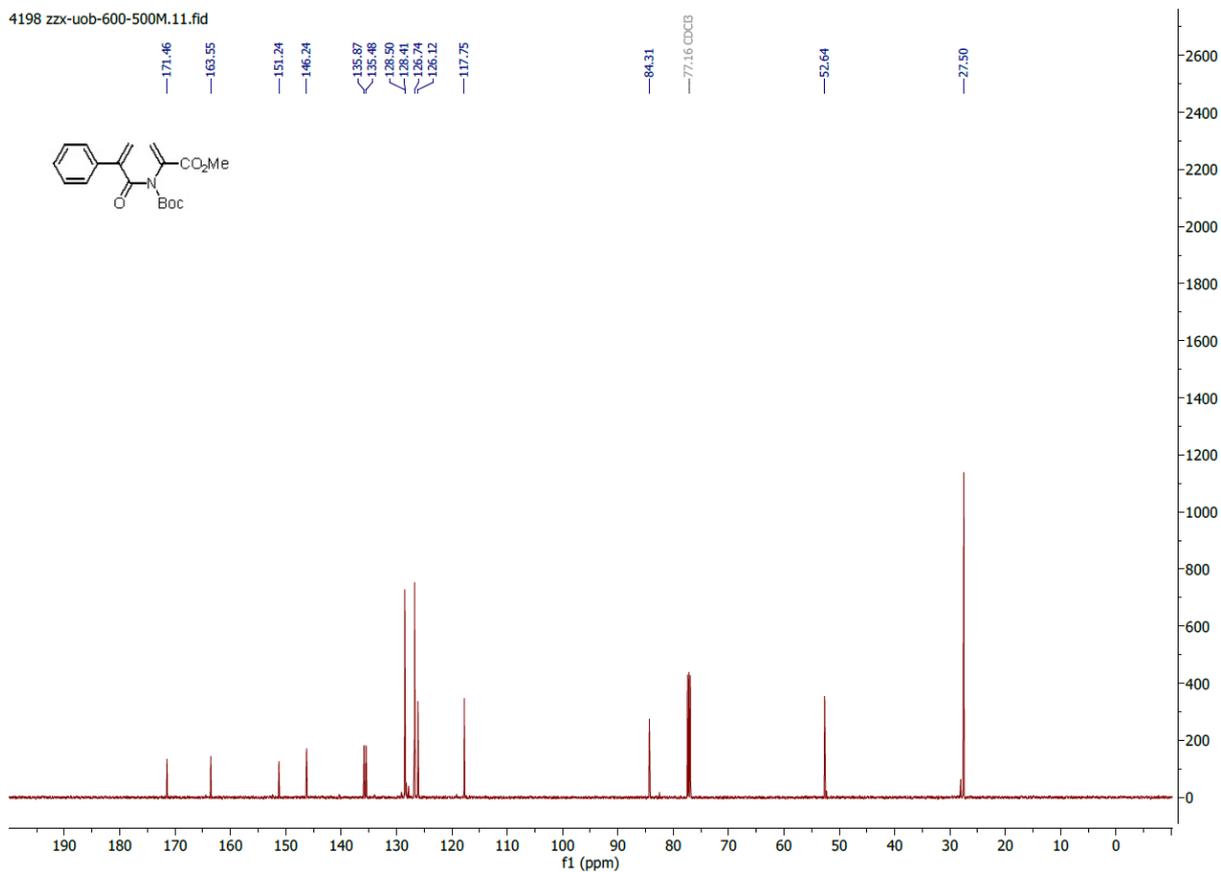
5.2 NMR Spectra of Cyclisation Precursors (7)

Compound 7a

4198 zzx-uob-600-500M.10.fid

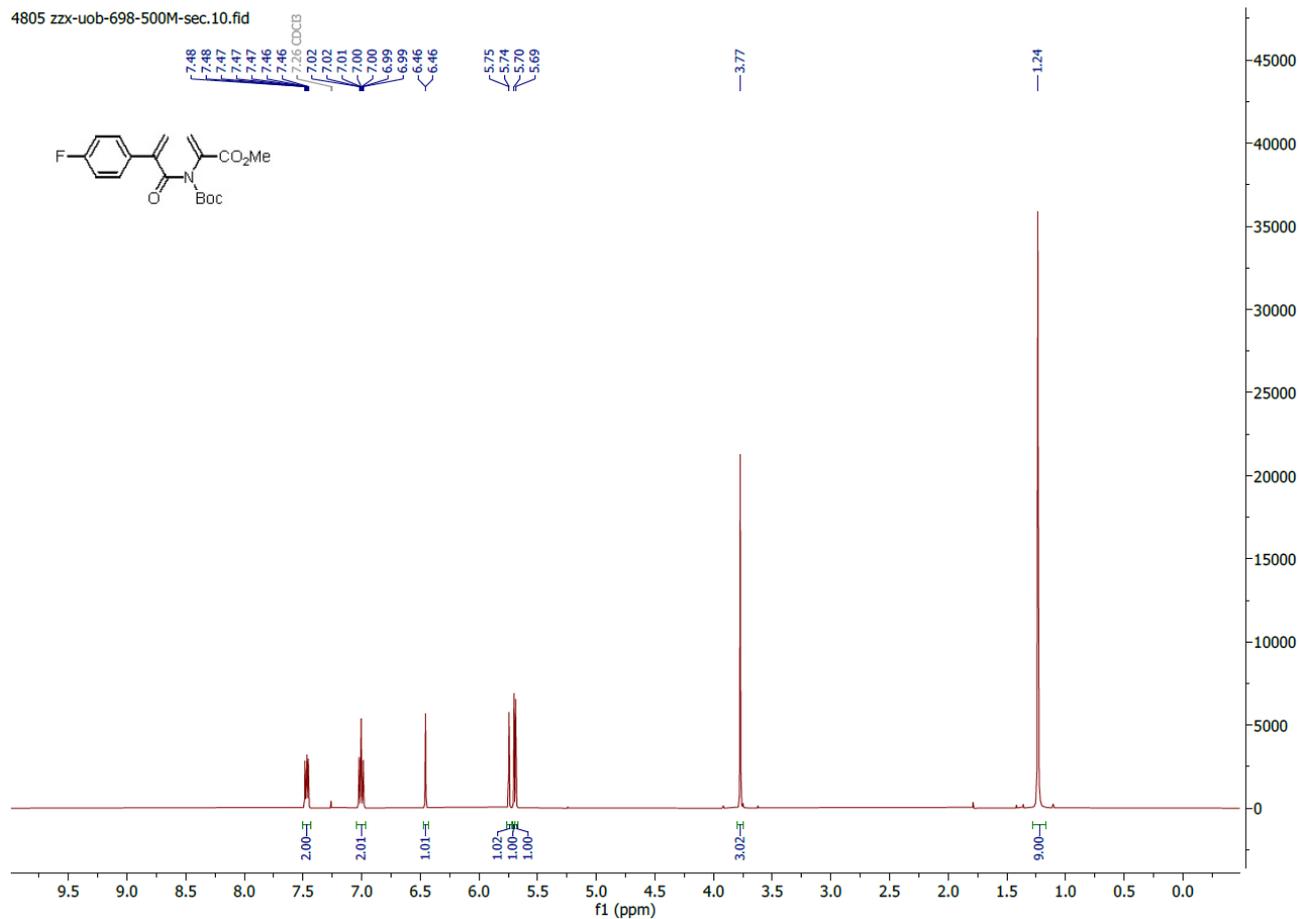


4198 zzx-uob-600-500M.11.fid

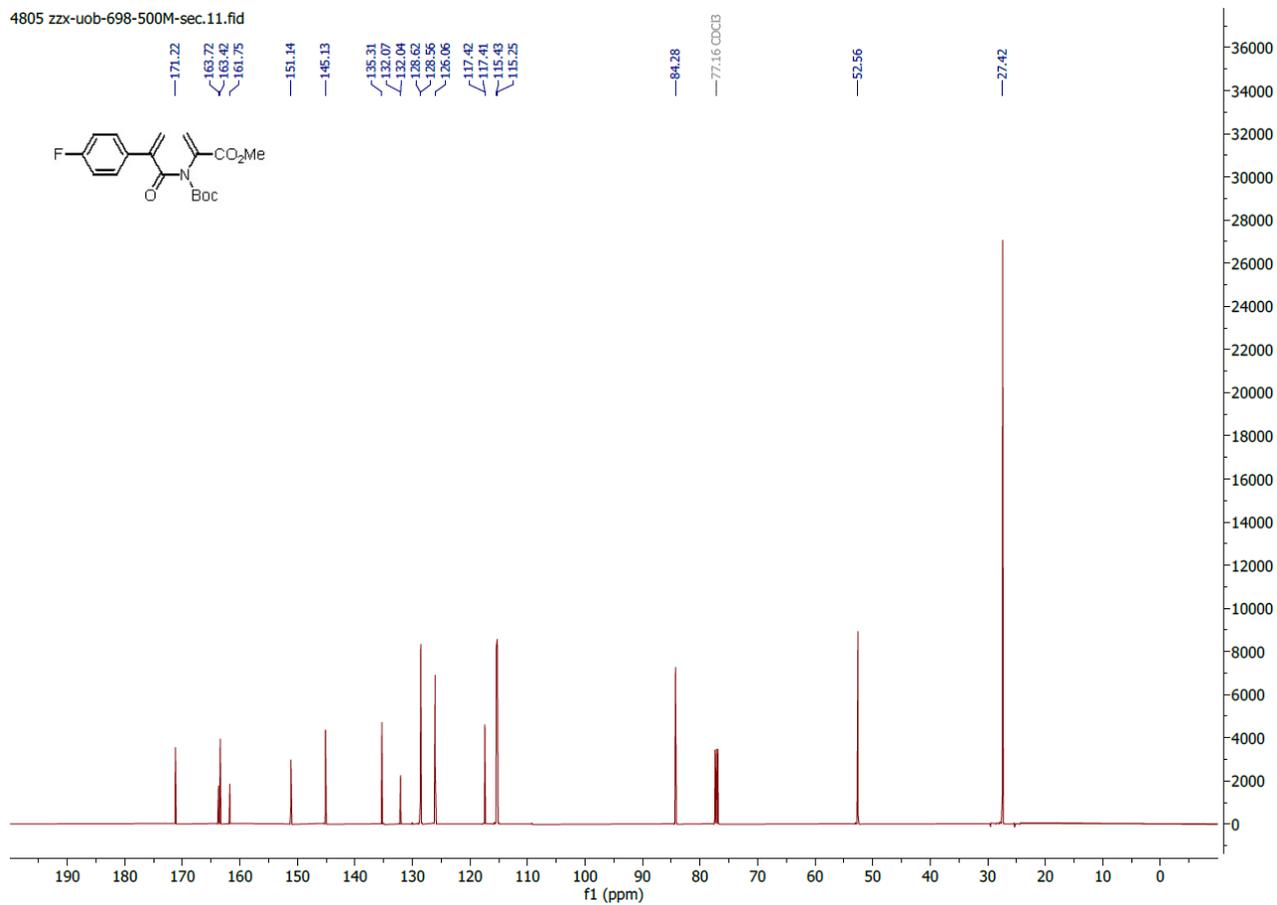


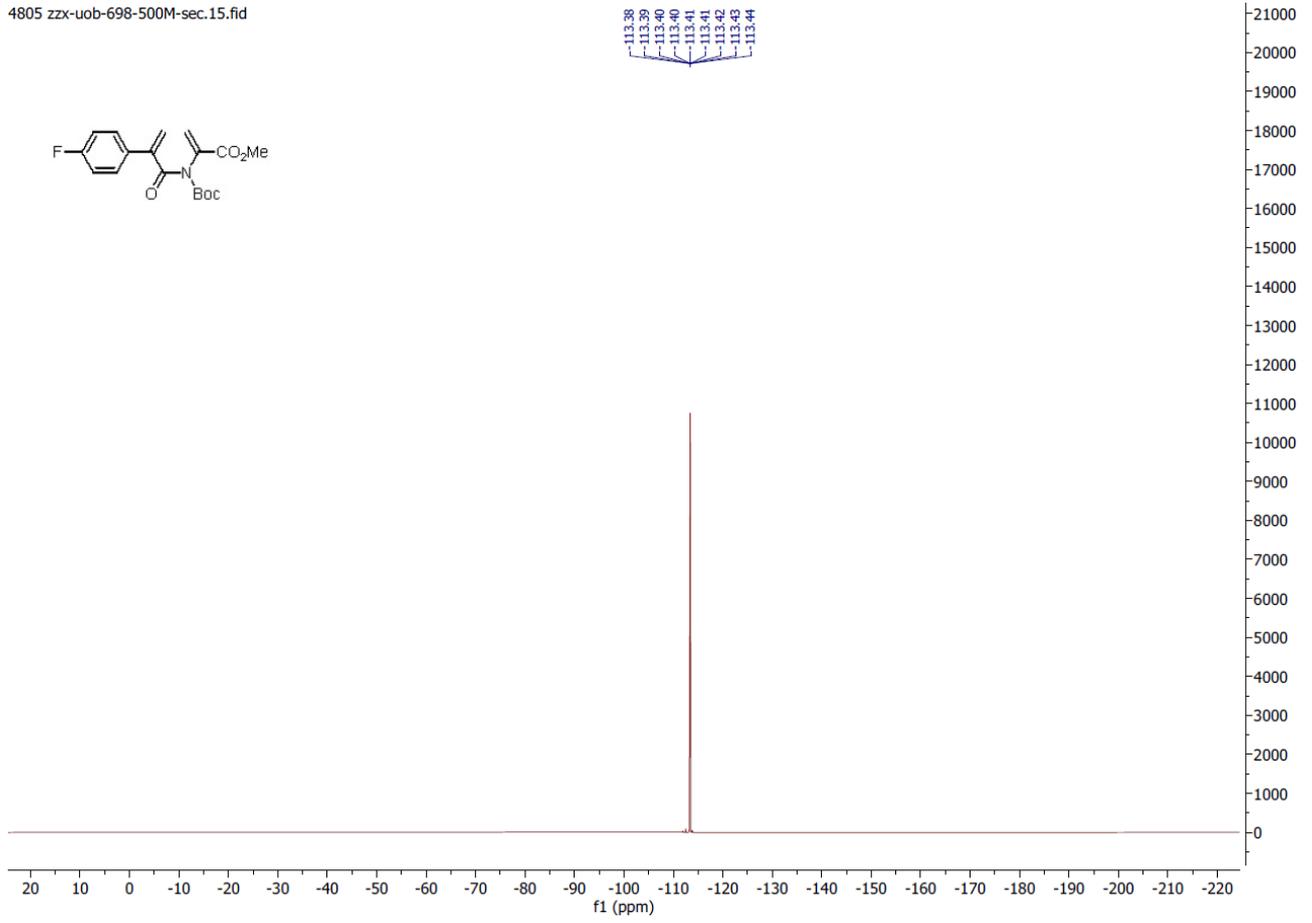
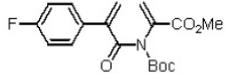
Compound 7b

4805 zzx-uob-698-500M-sec.10.fid



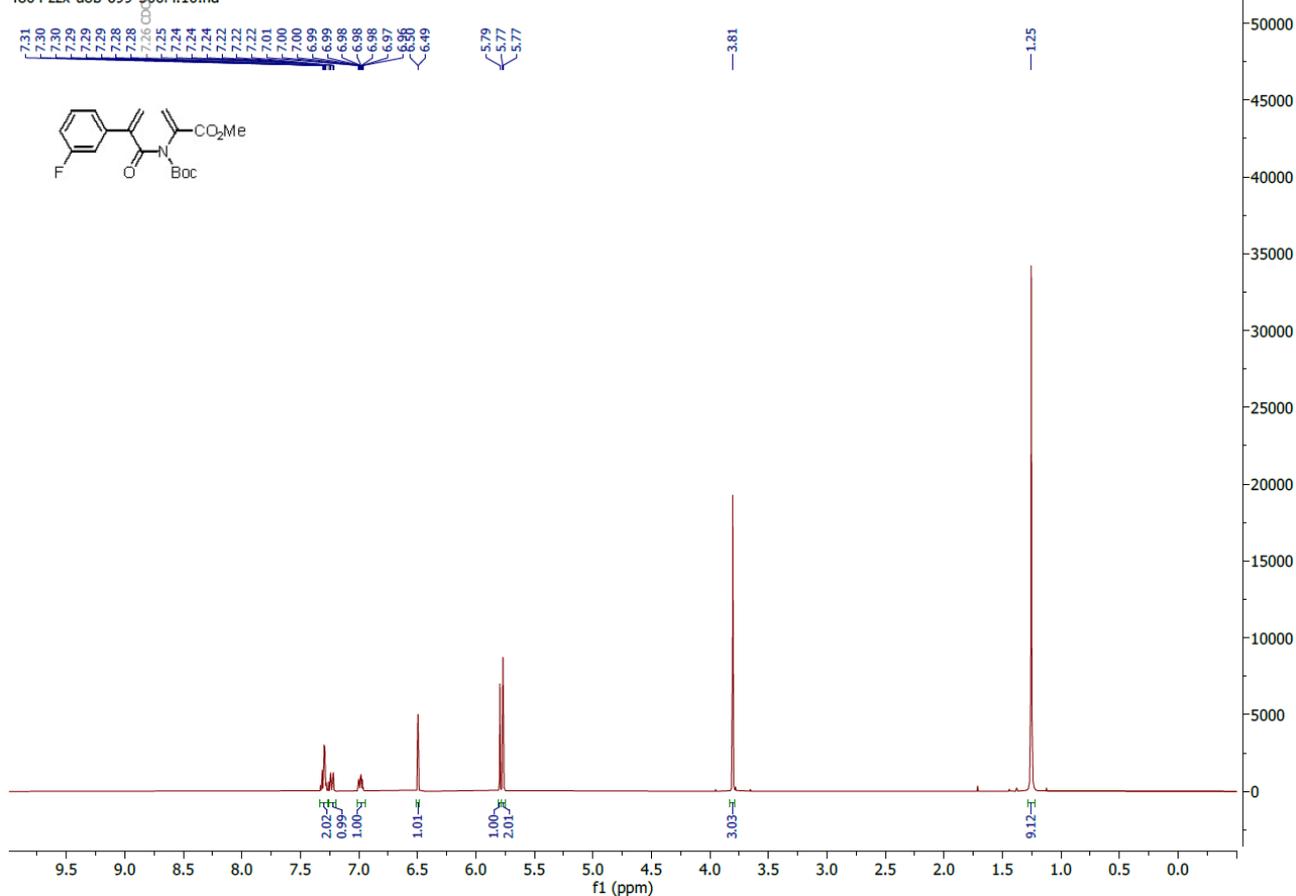
4805 zzx-uob-698-500M-sec.11.fid



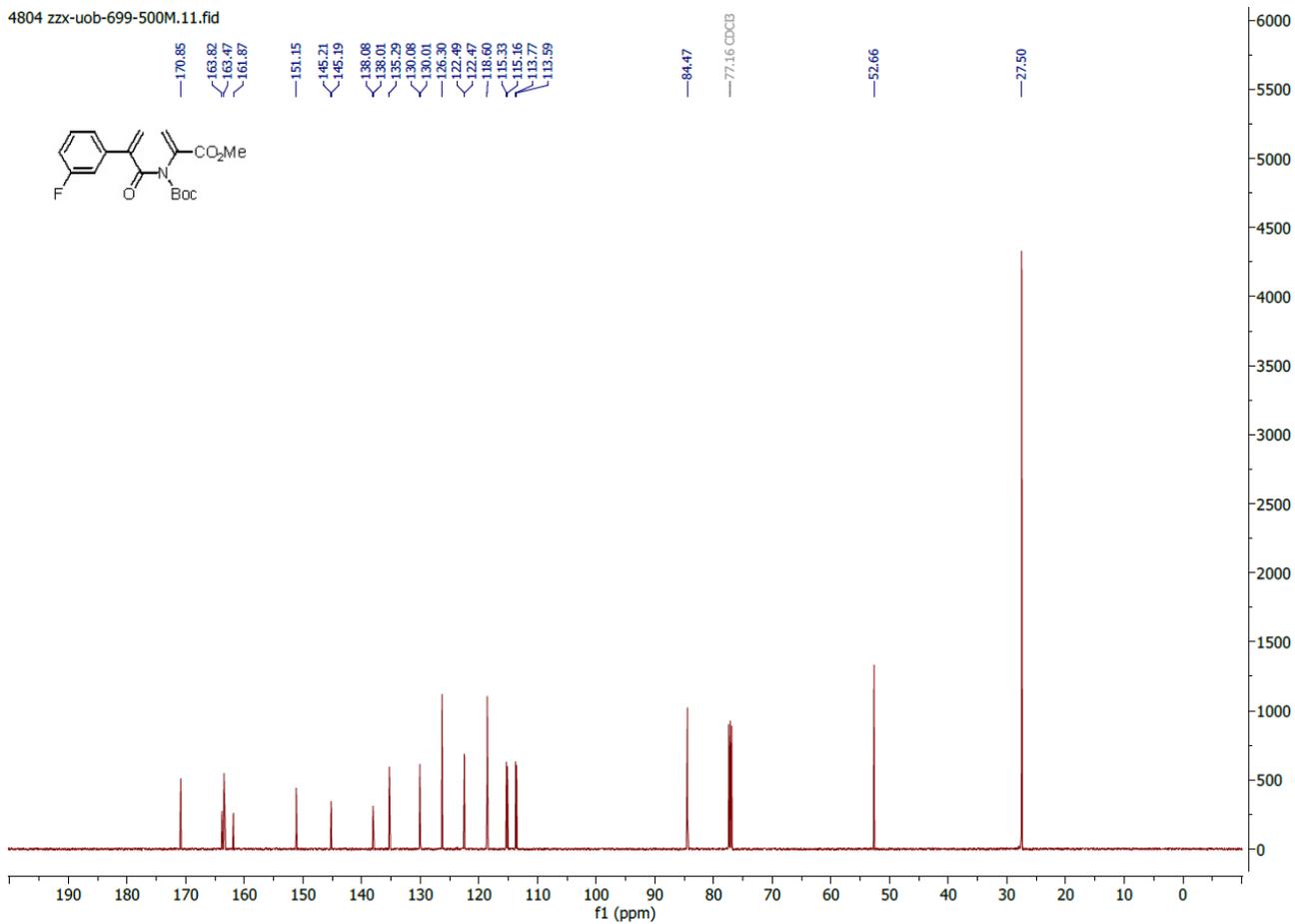


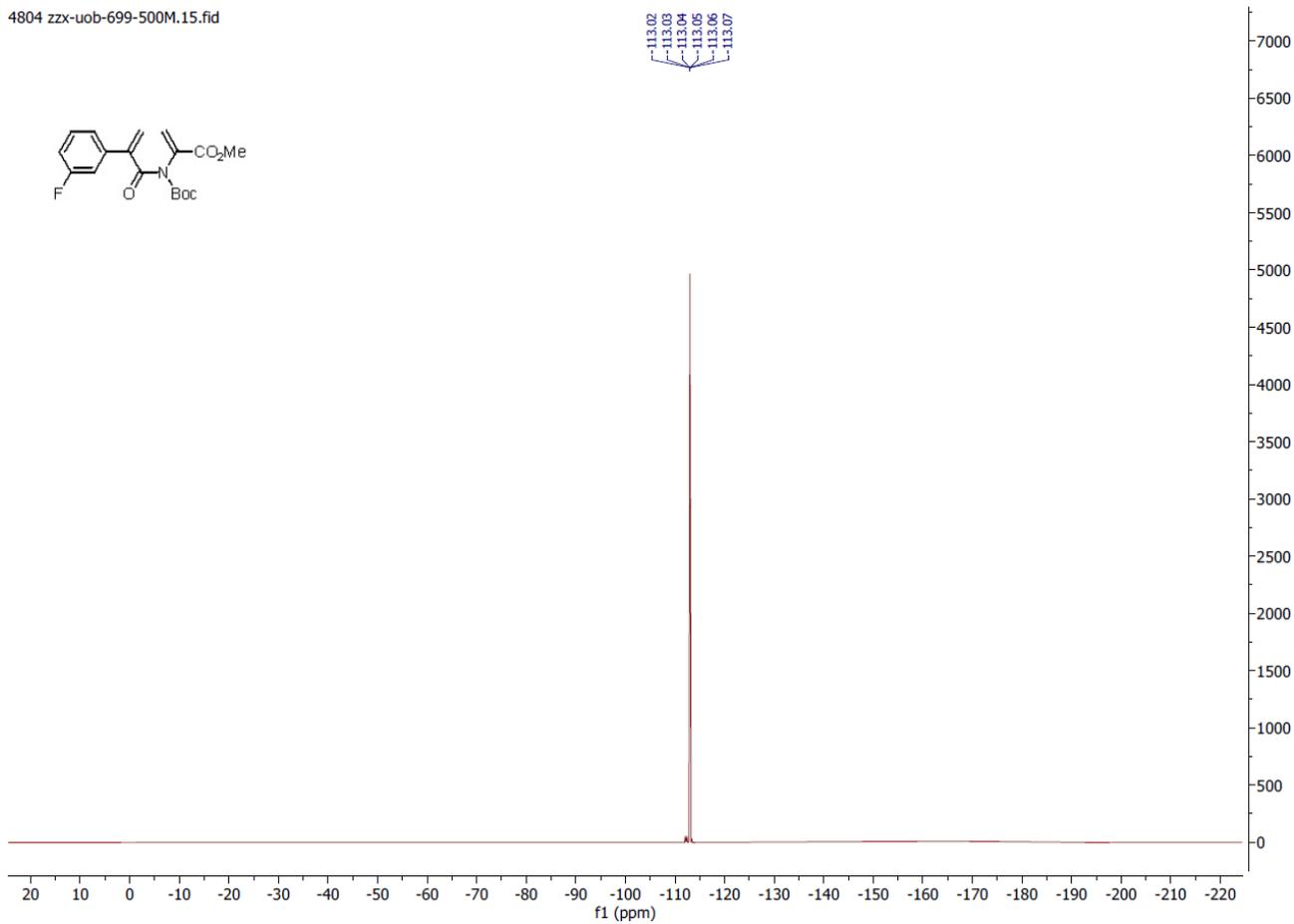
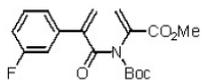
Compound 7c

4804 zzx-uob-699-500M.10.fid



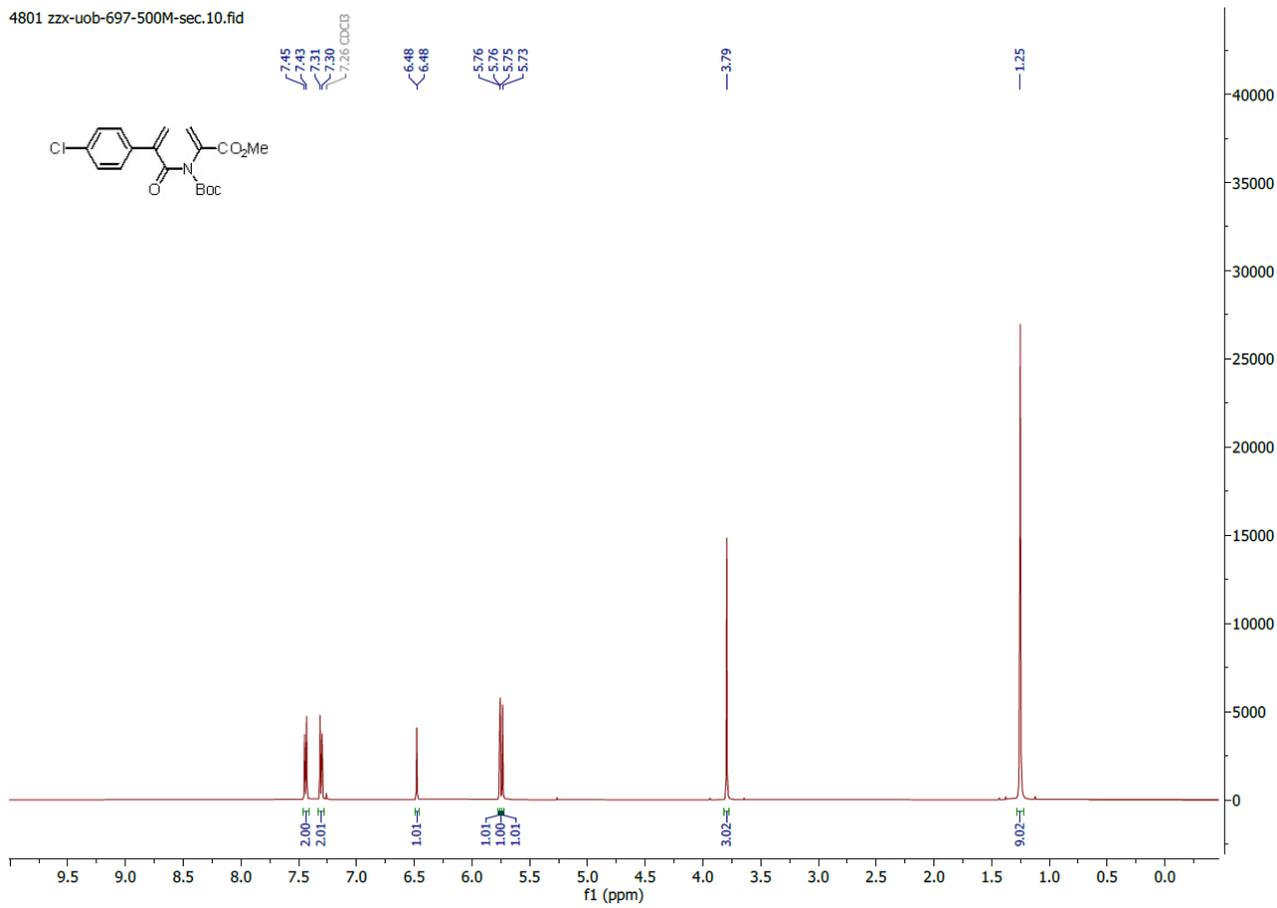
4804 zzx-uob-699-500M.11.fid



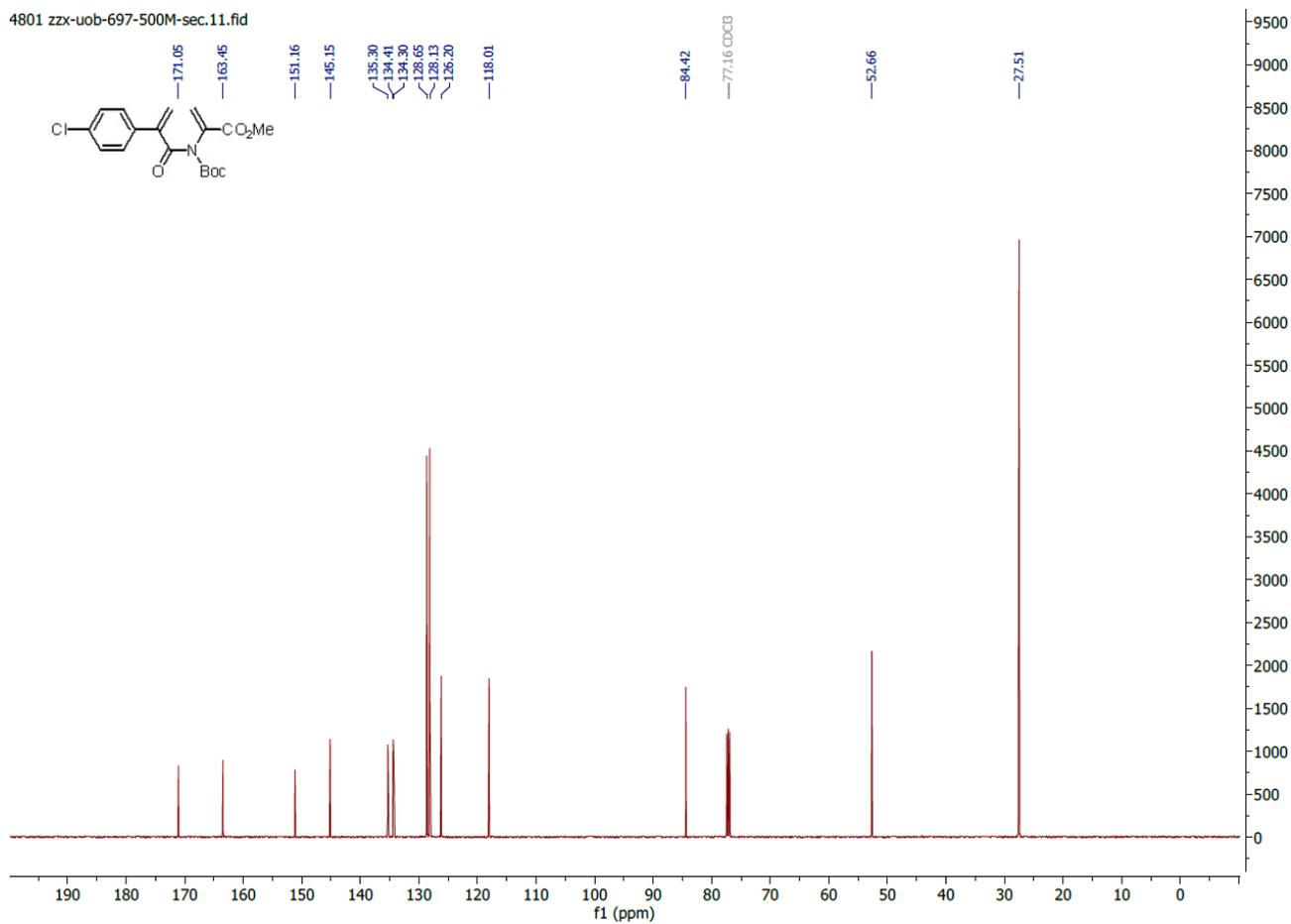


Compound 7d

4801 zzx-uob-697-500M-sec.10.fid

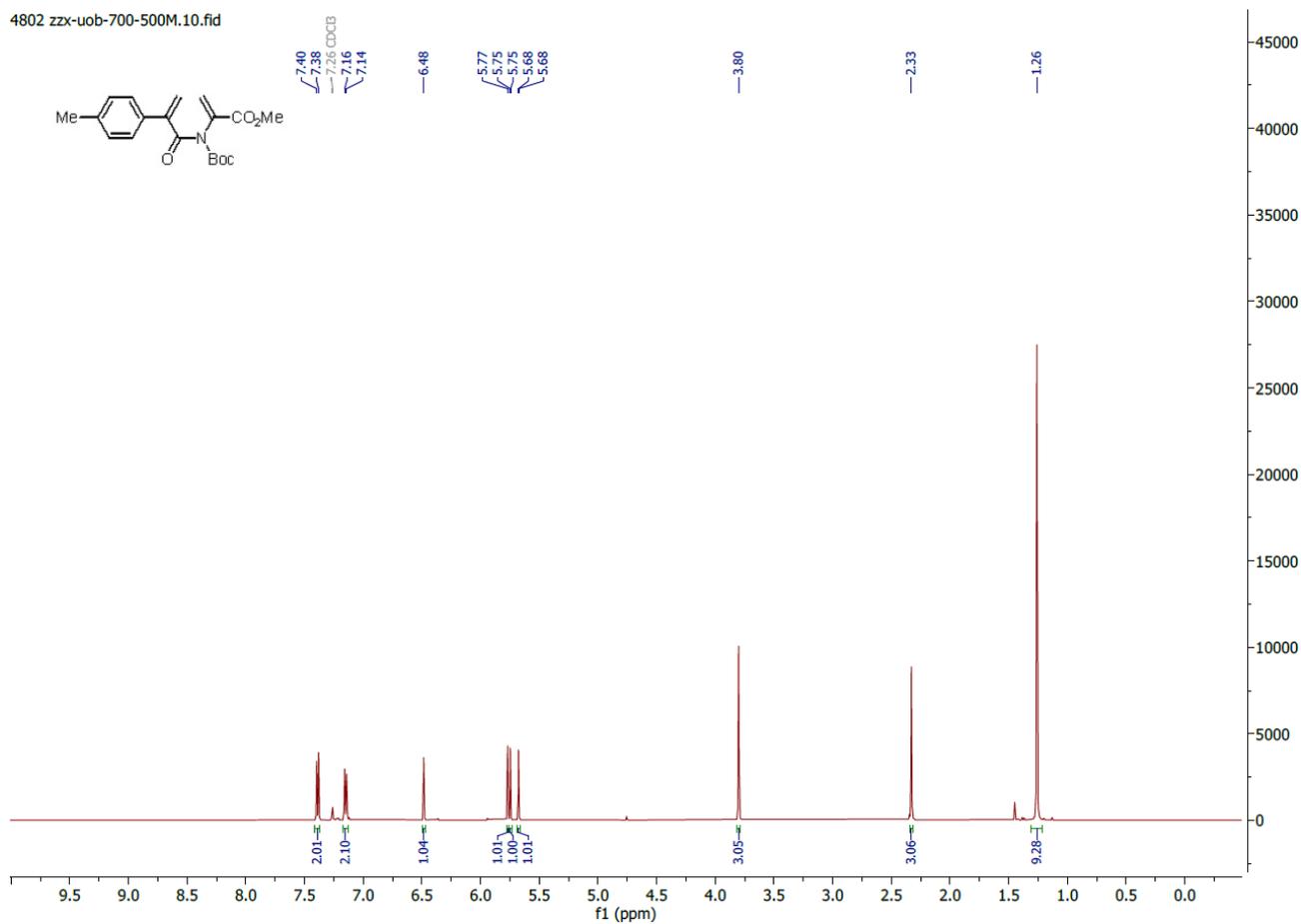
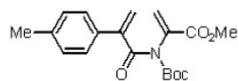


4801 zzx-uob-697-500M-sec.11.fid

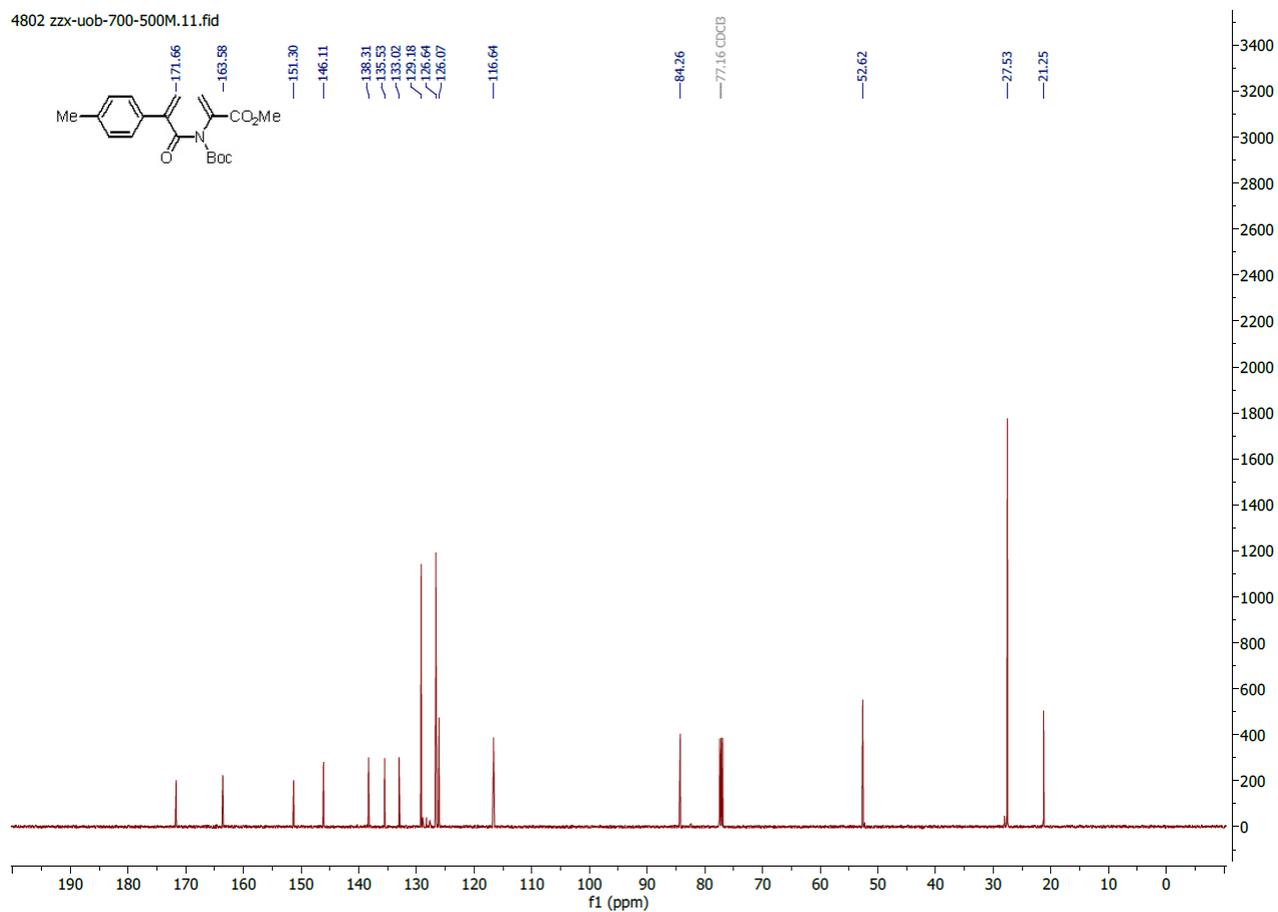
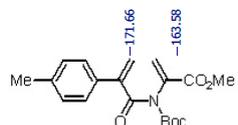


Compound 7e

4802 zzx-uob-700-500M.10.fid

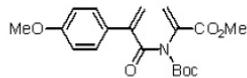


4802 zzx-uob-700-500M.11.fid



Compound 7f

4803 zzx-uob-701-500M.10.fid



7.45
7.43
7.26 CDCl₃

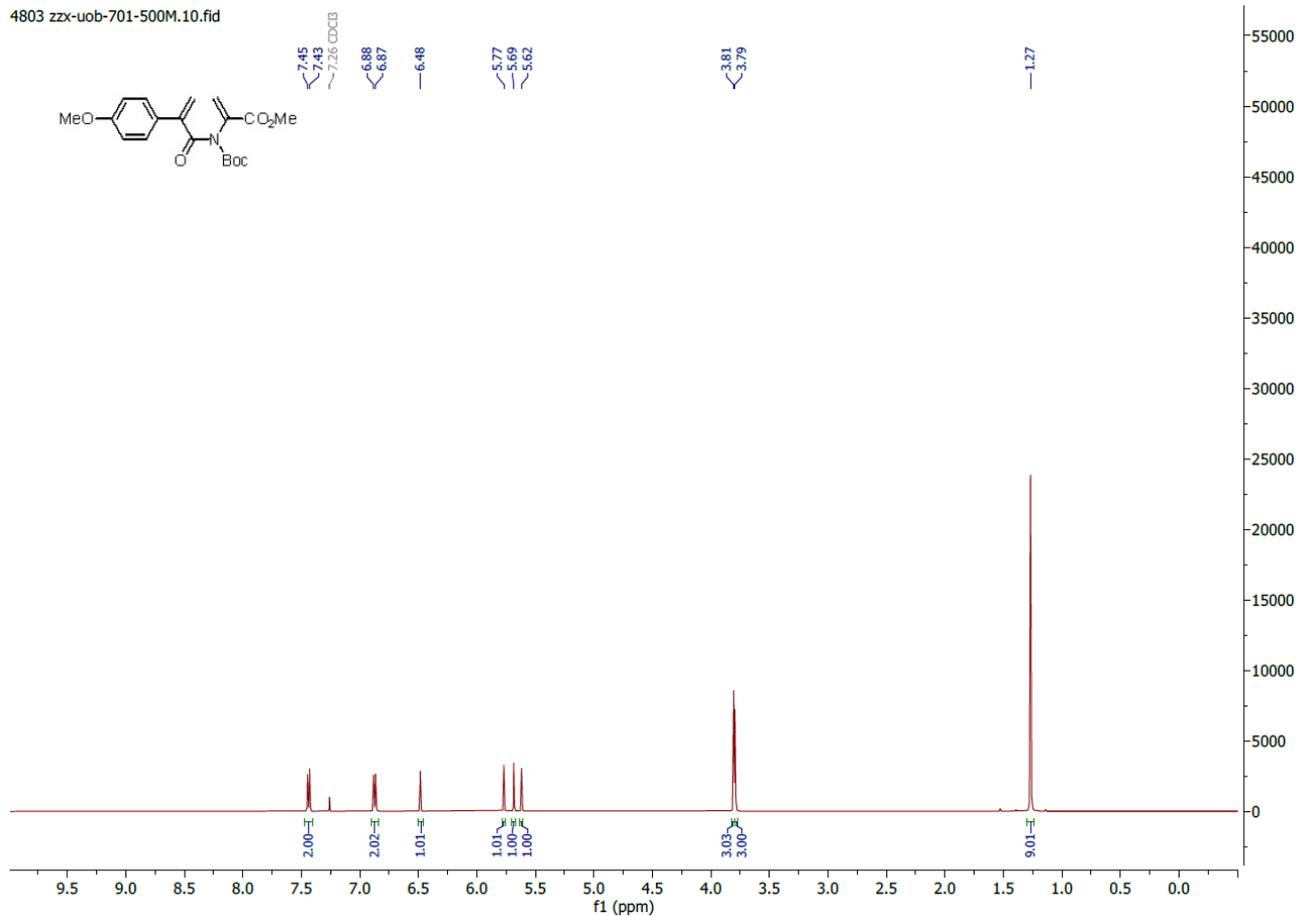
6.88
6.87

6.48

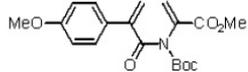
3.77
3.69
3.62

3.81
3.79

1.27



4803 zzx-uob-701-500M.11.fid



171.80

163.61

159.85

151.36

146.70

135.54

128.50

128.07

126.09

115.58

113.93

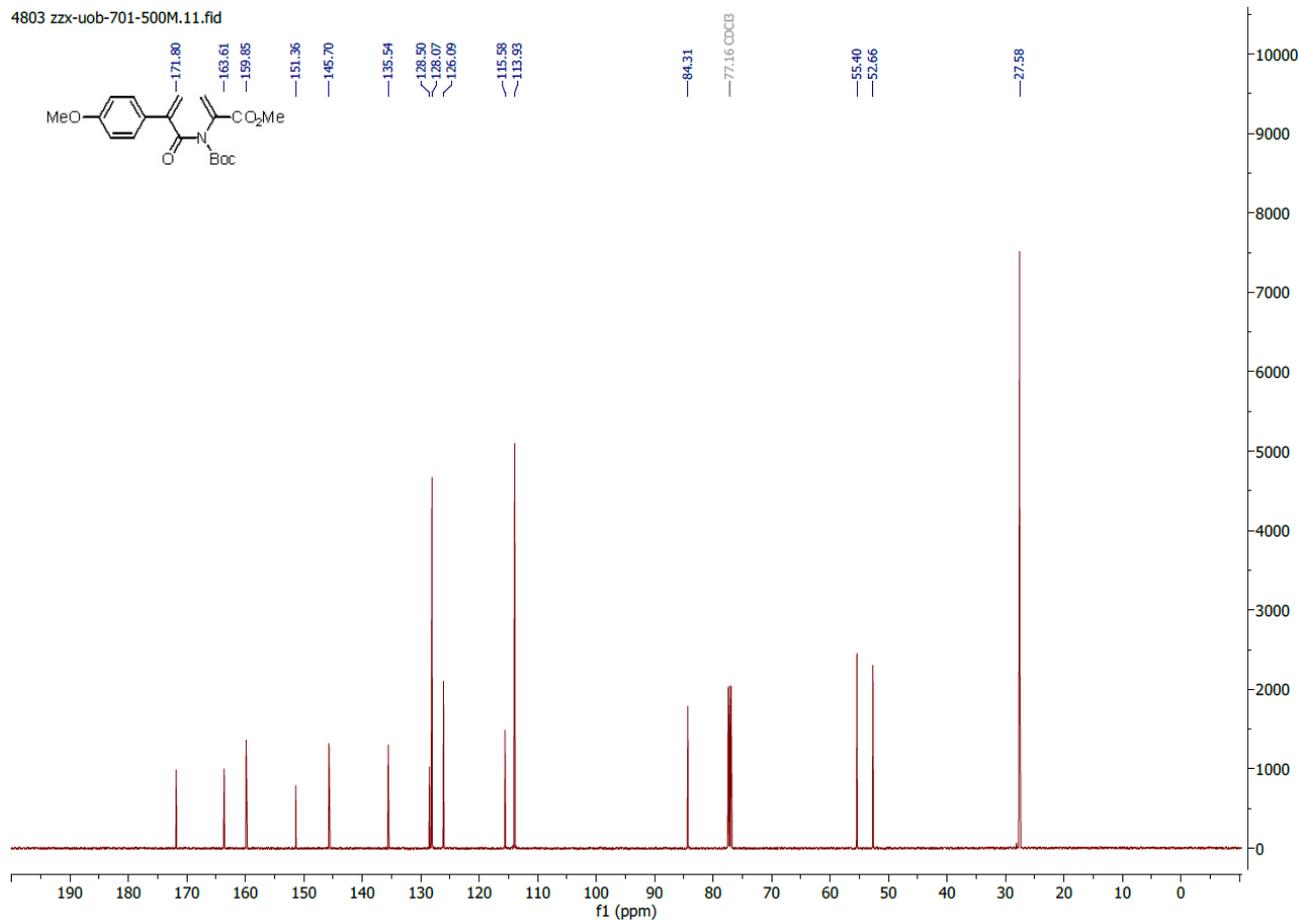
84.31

77.16 CDCl₃

55.40

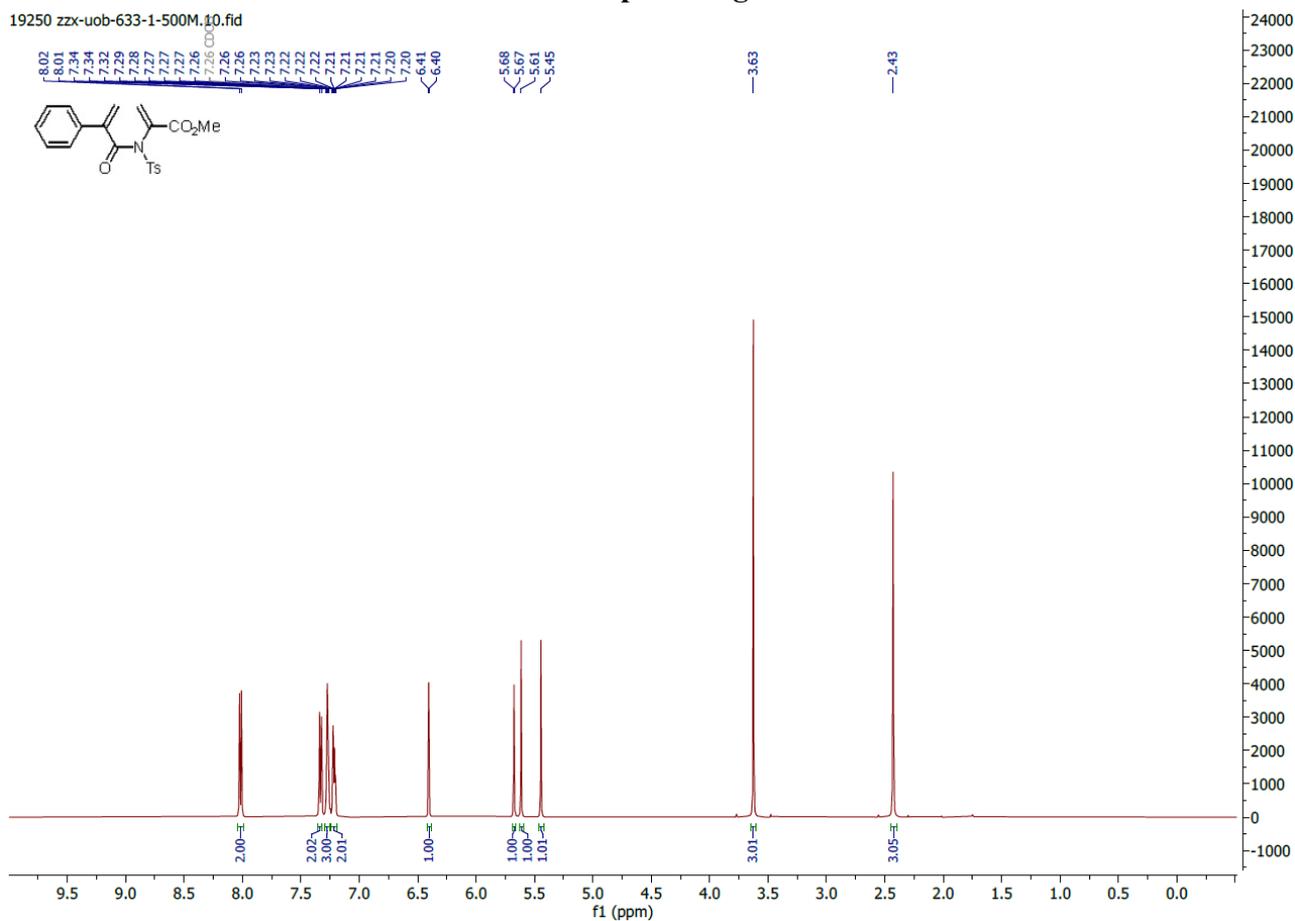
52.66

27.58

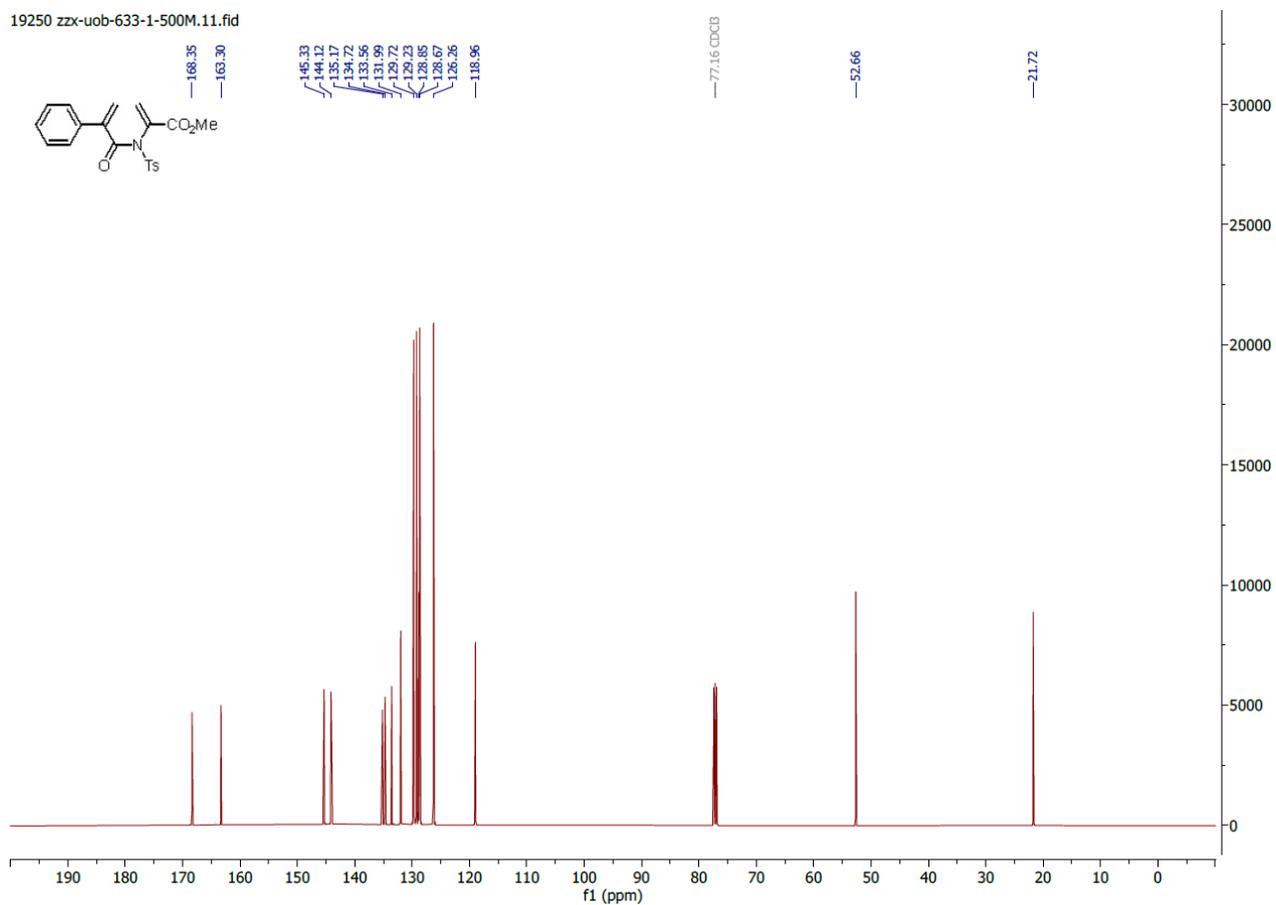


Compound 7g

19250 zzx-uob-633-1-500M.10.fid

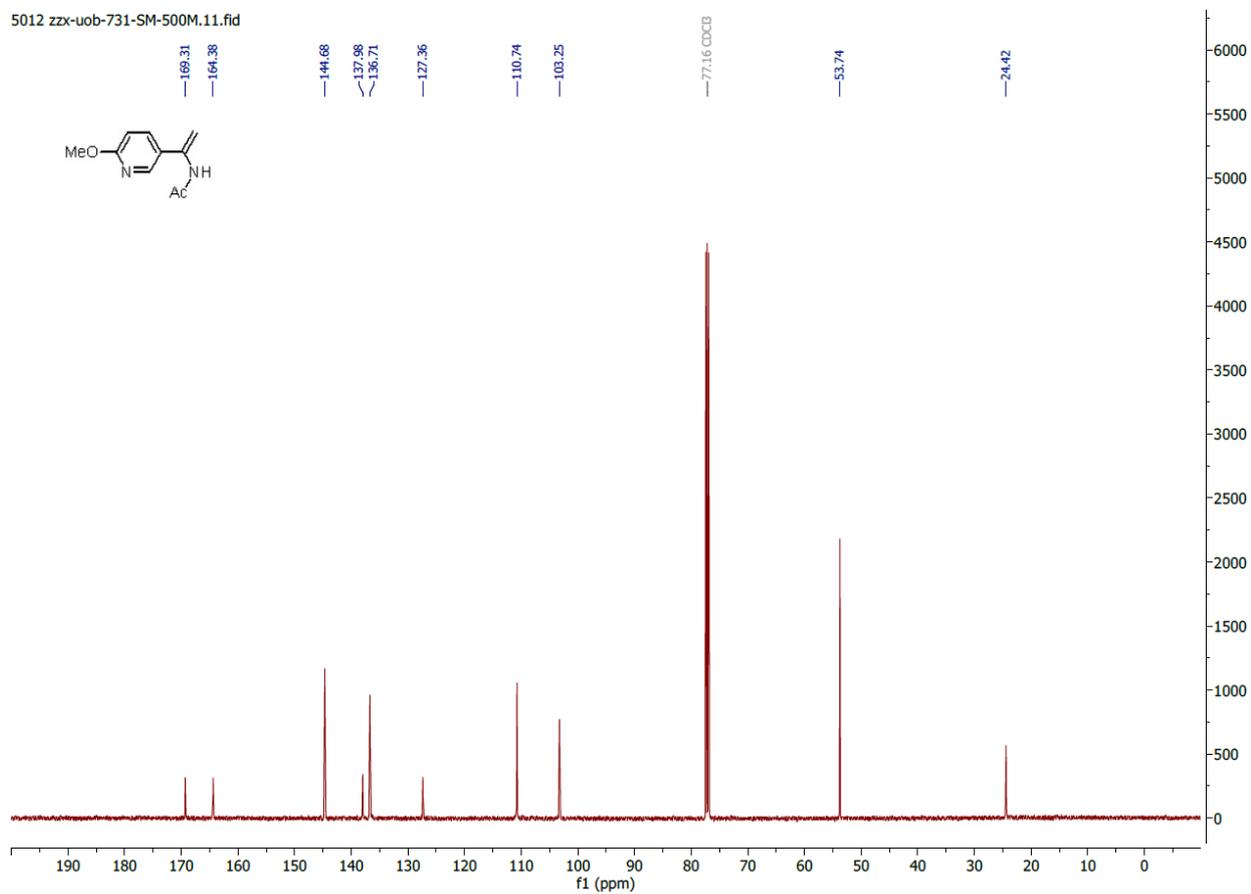
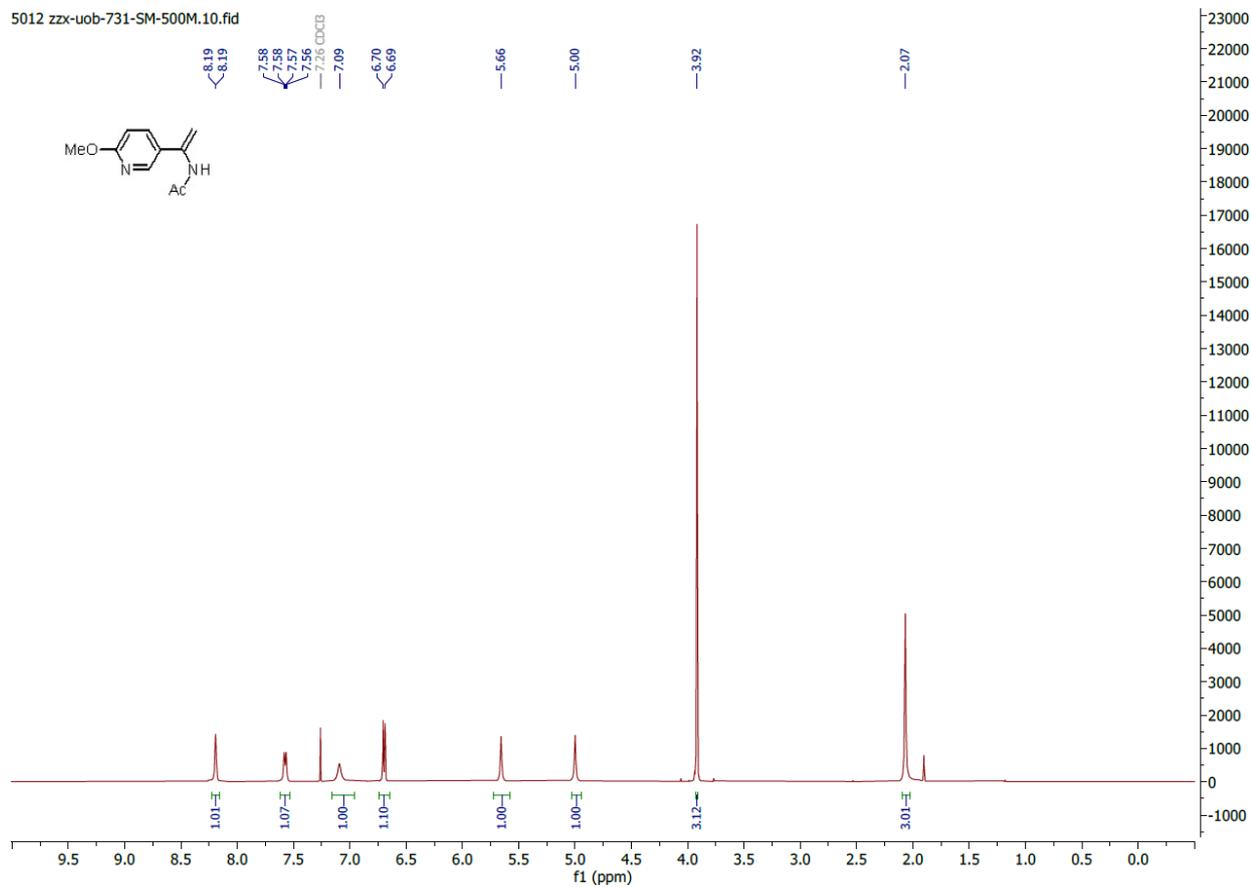


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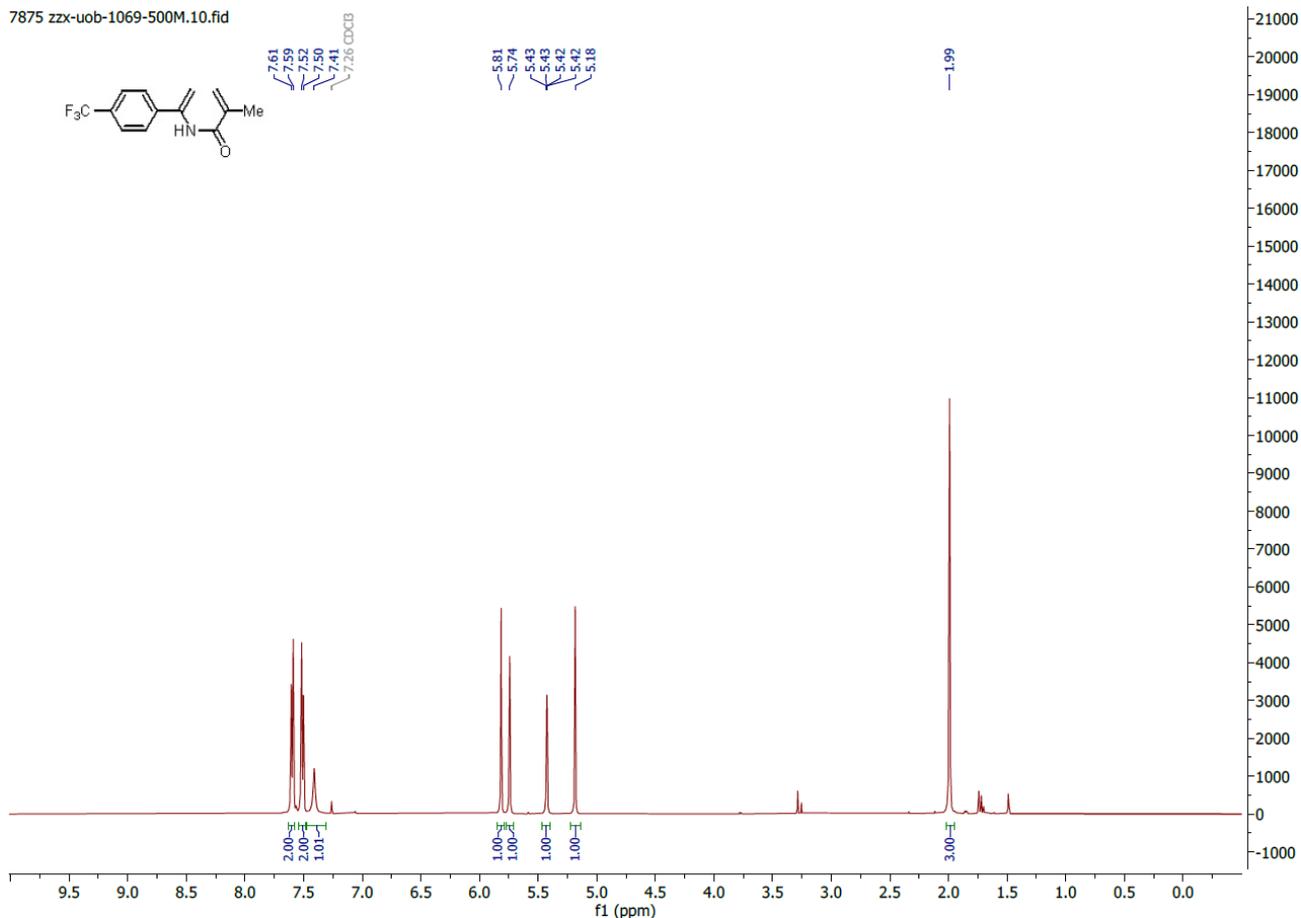
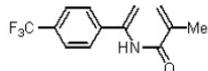
5.3 NMR Spectra of Cyclisation Precursors (2)

Compound S2t

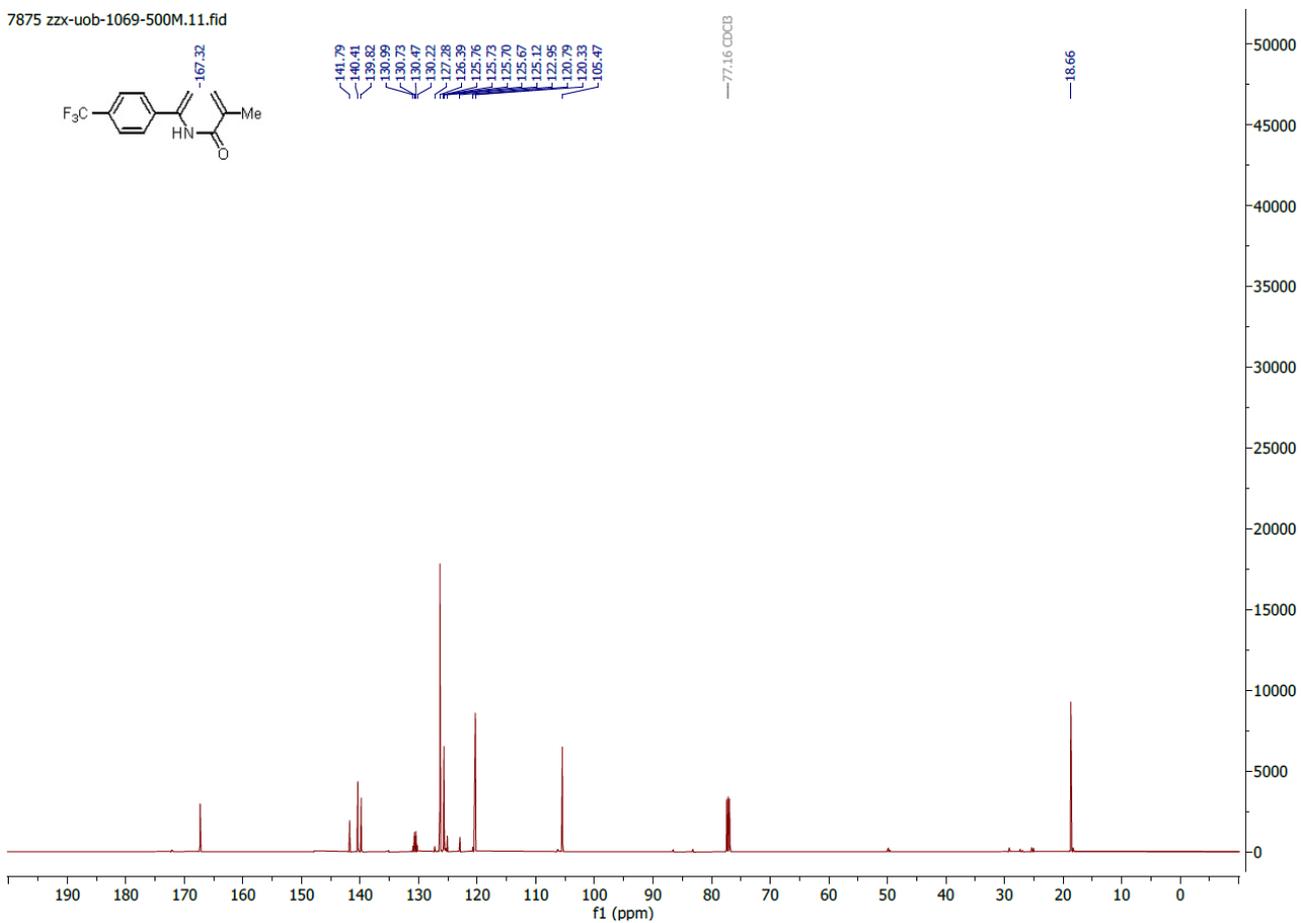
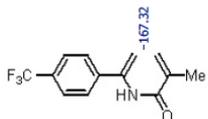


Compound S2a

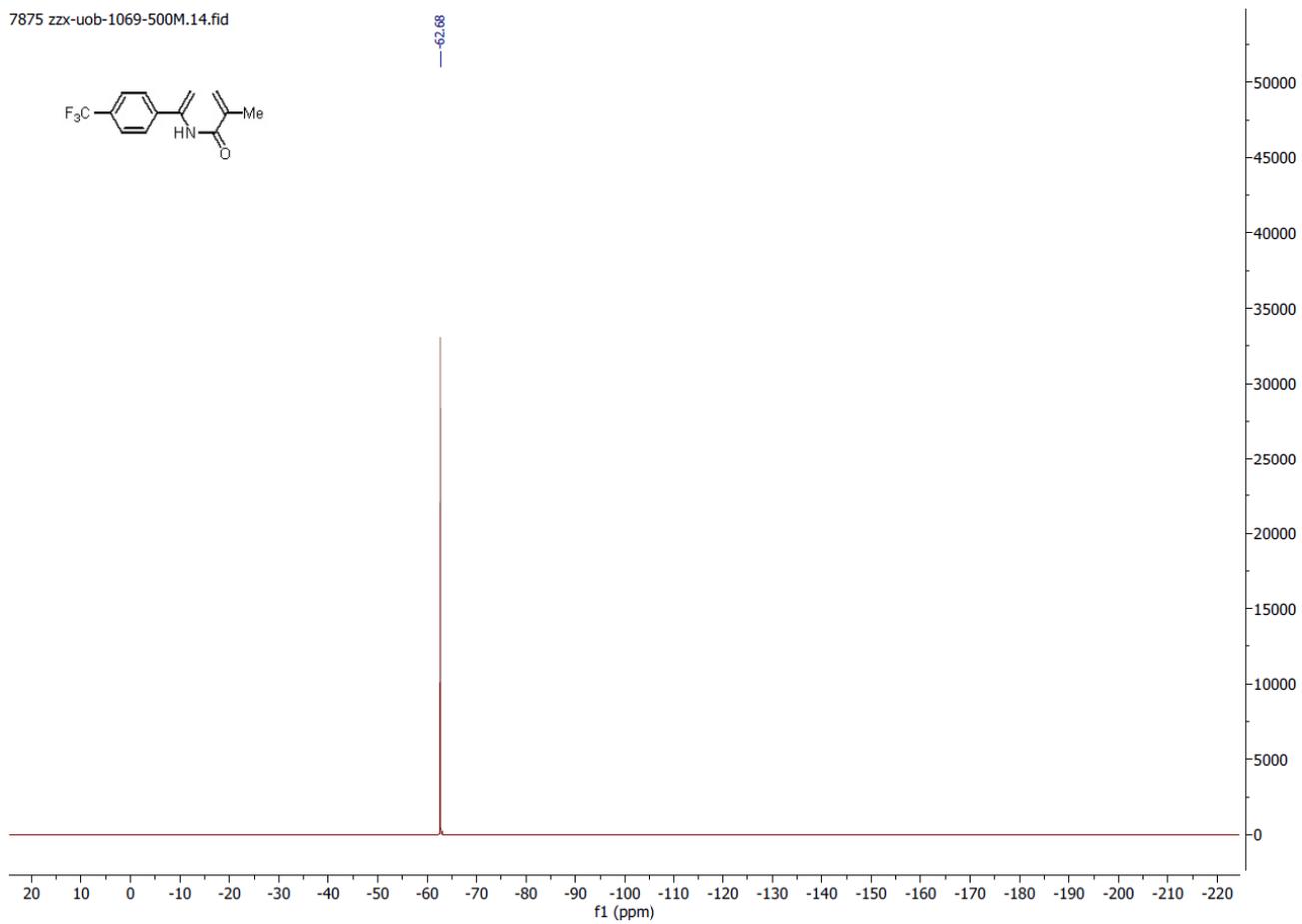
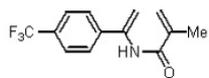
7875 zzx-uob-1069-500M.10.fid



7875 zzx-uob-1069-500M.11.fid

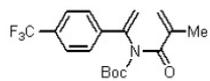


7875 zzx-uob-1069-500M.14.fid

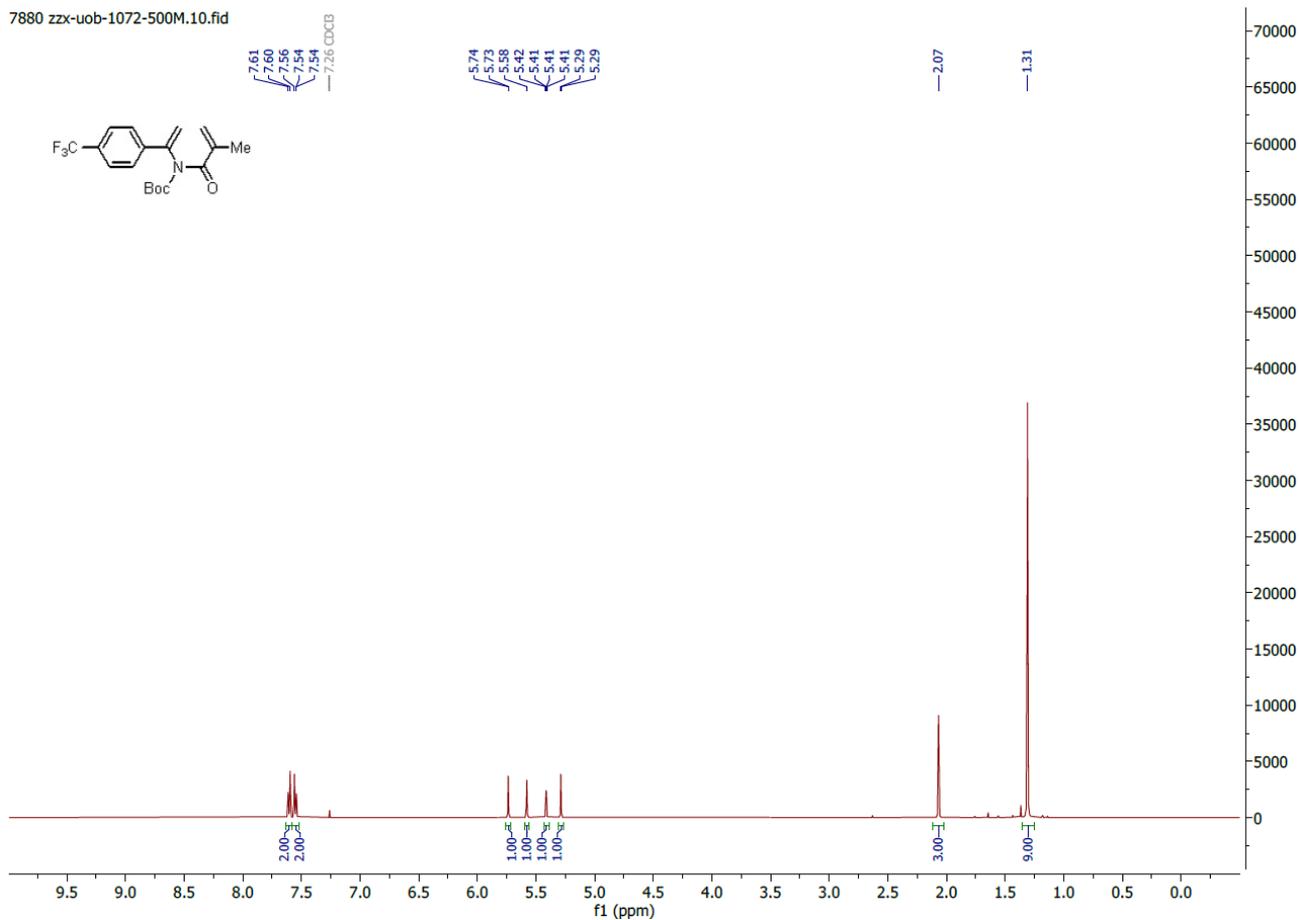


Compound 2a

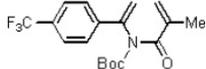
7880 zzx-uob-1072-500M.10.fid



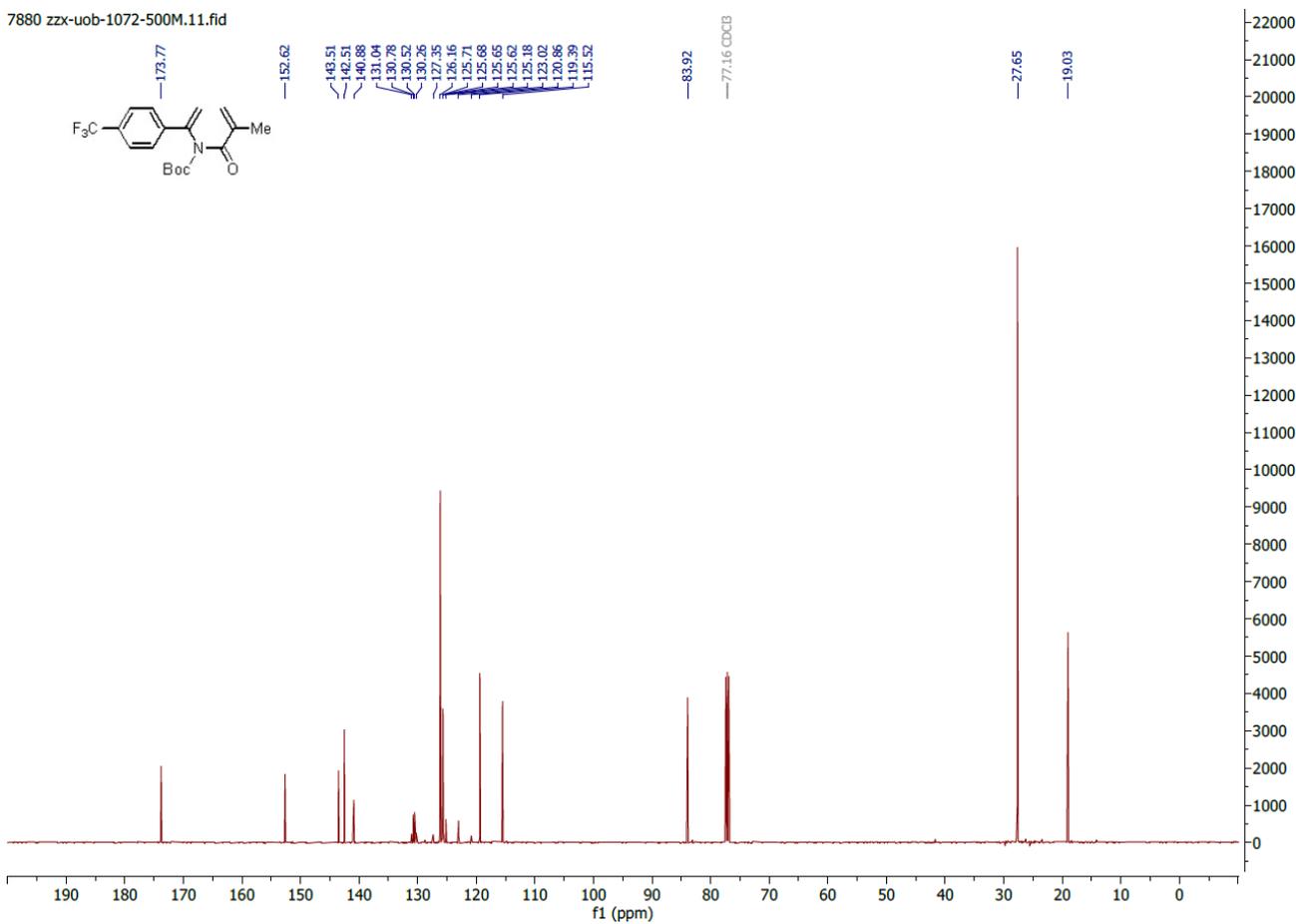
7.61
7.60
7.56
7.54
7.54
7.26 CDCl₃
5.74
5.73
5.58
5.42
5.41
5.41
5.41
5.29
5.29

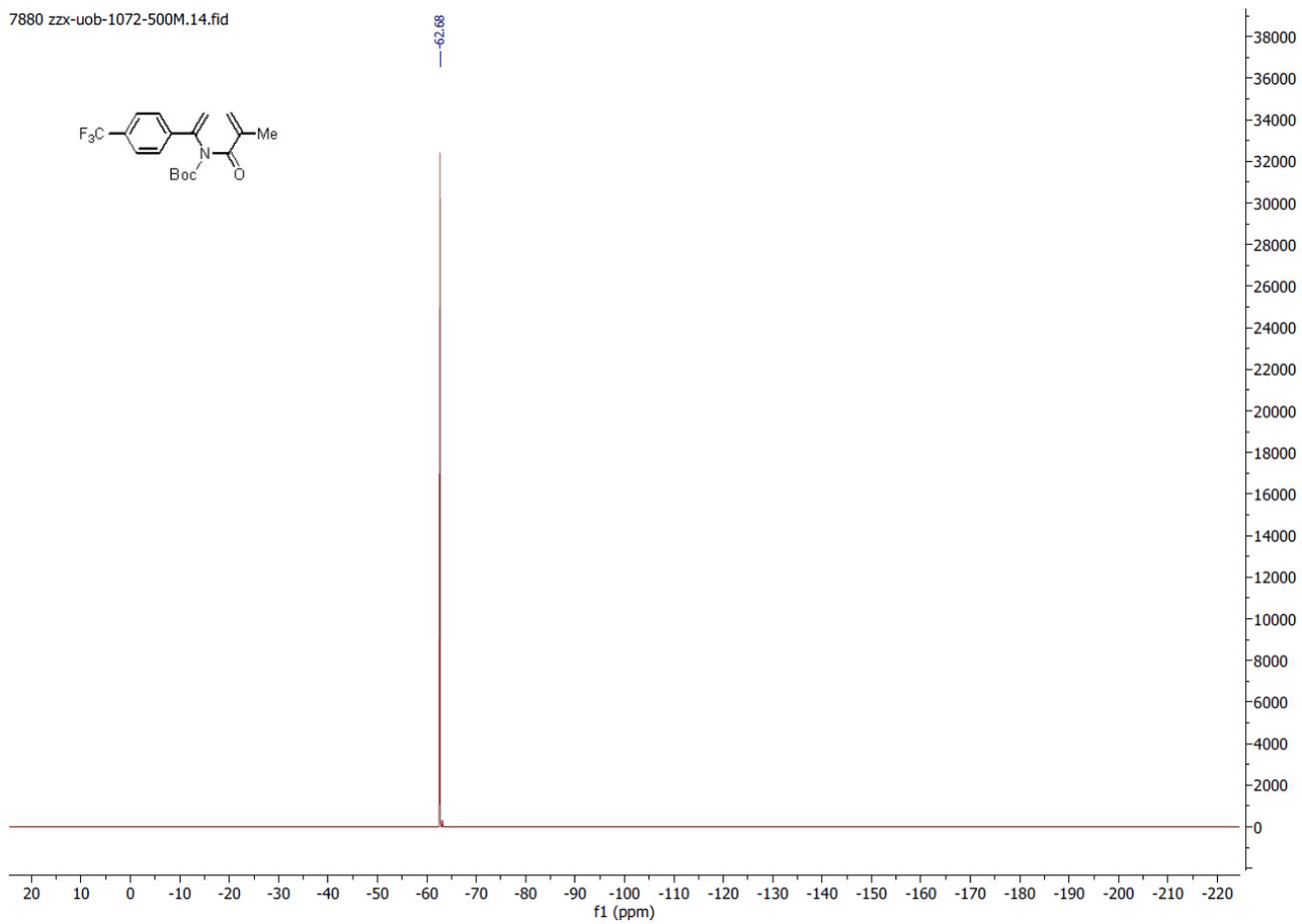
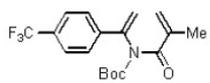


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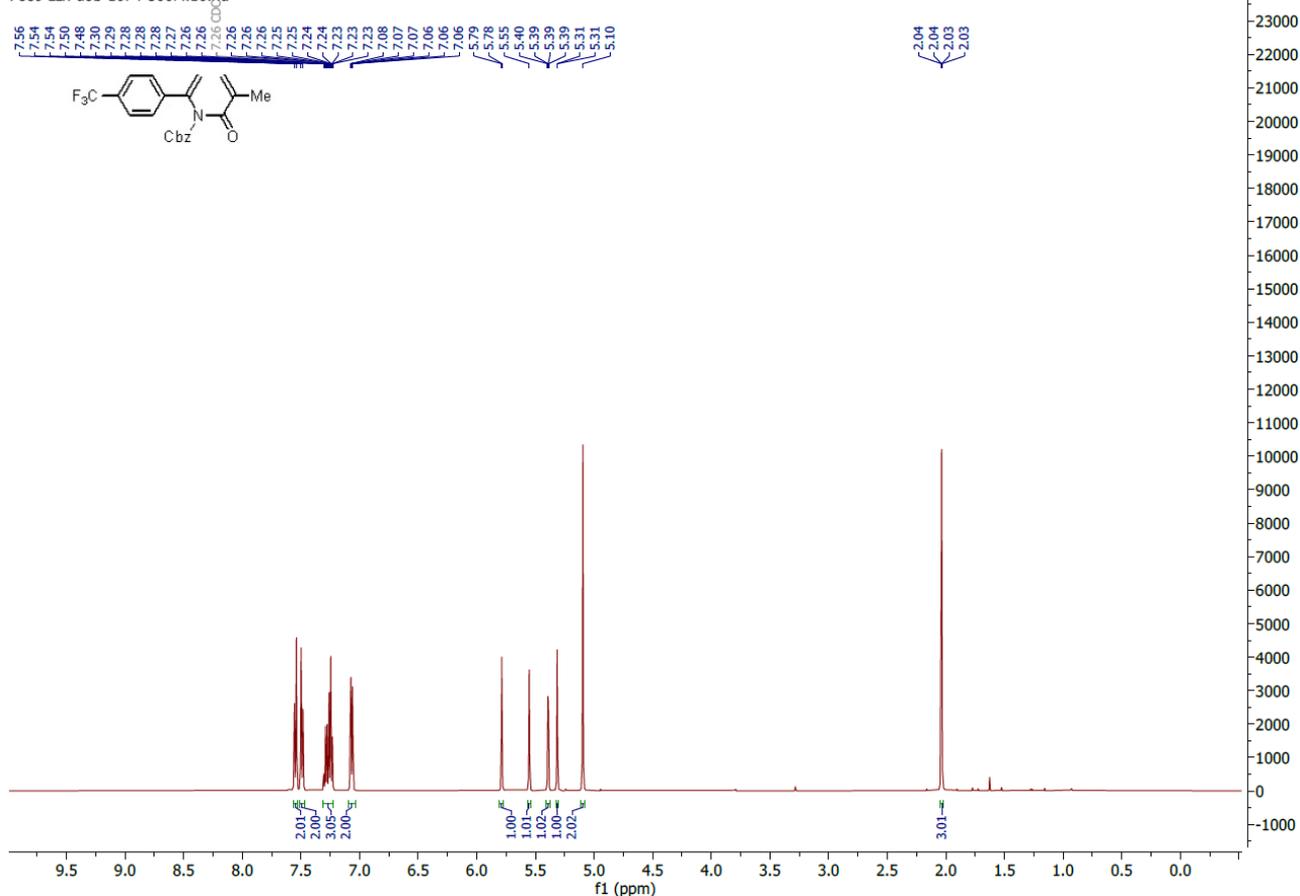
173.77
152.62
143.51
142.51
140.88
131.04
130.78
130.52
130.26
127.35
126.16
125.71
125.68
125.65
125.62
125.18
123.02
120.86
119.29
115.52
83.92
77.16 CDCl₃
27.65
19.03



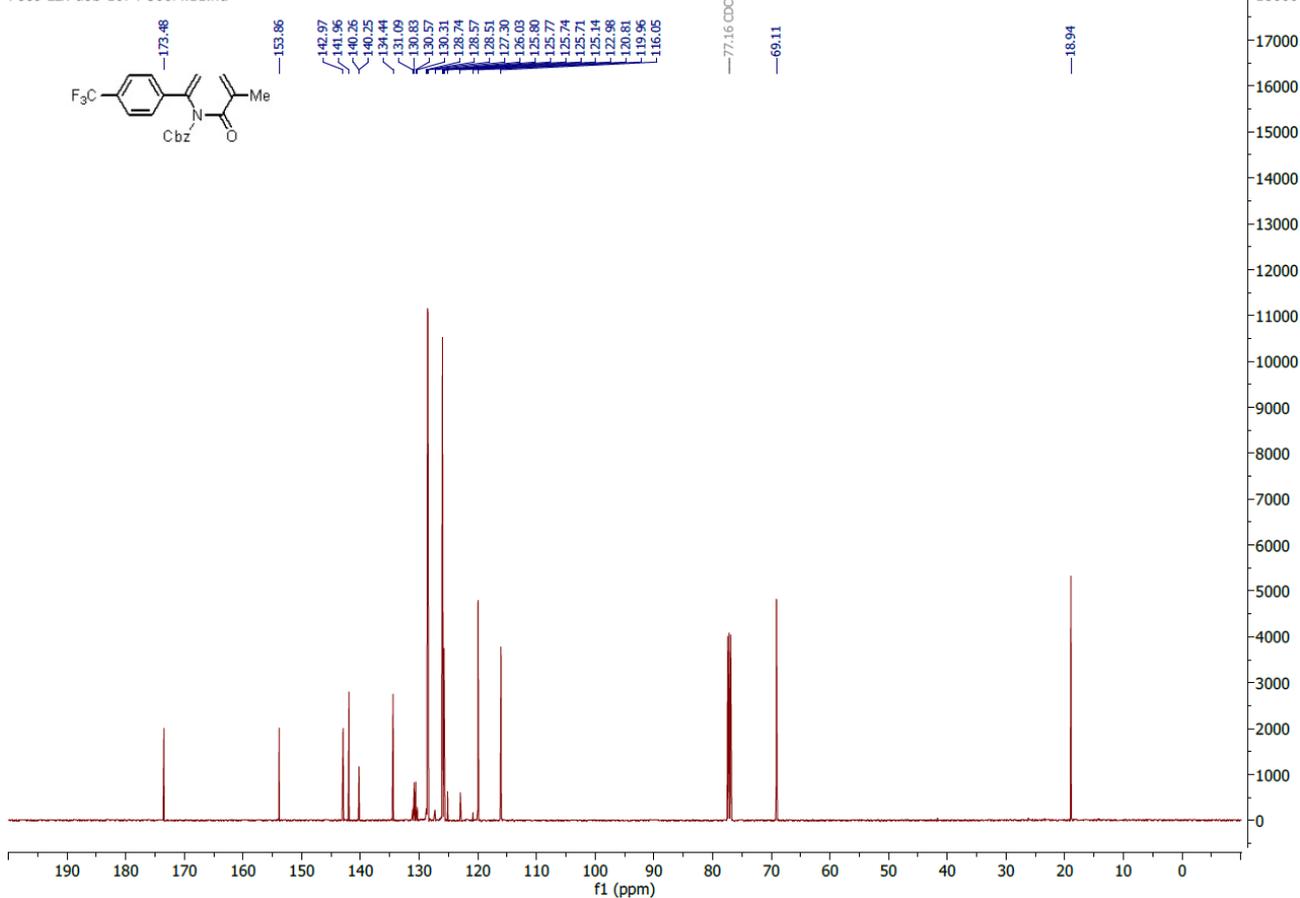


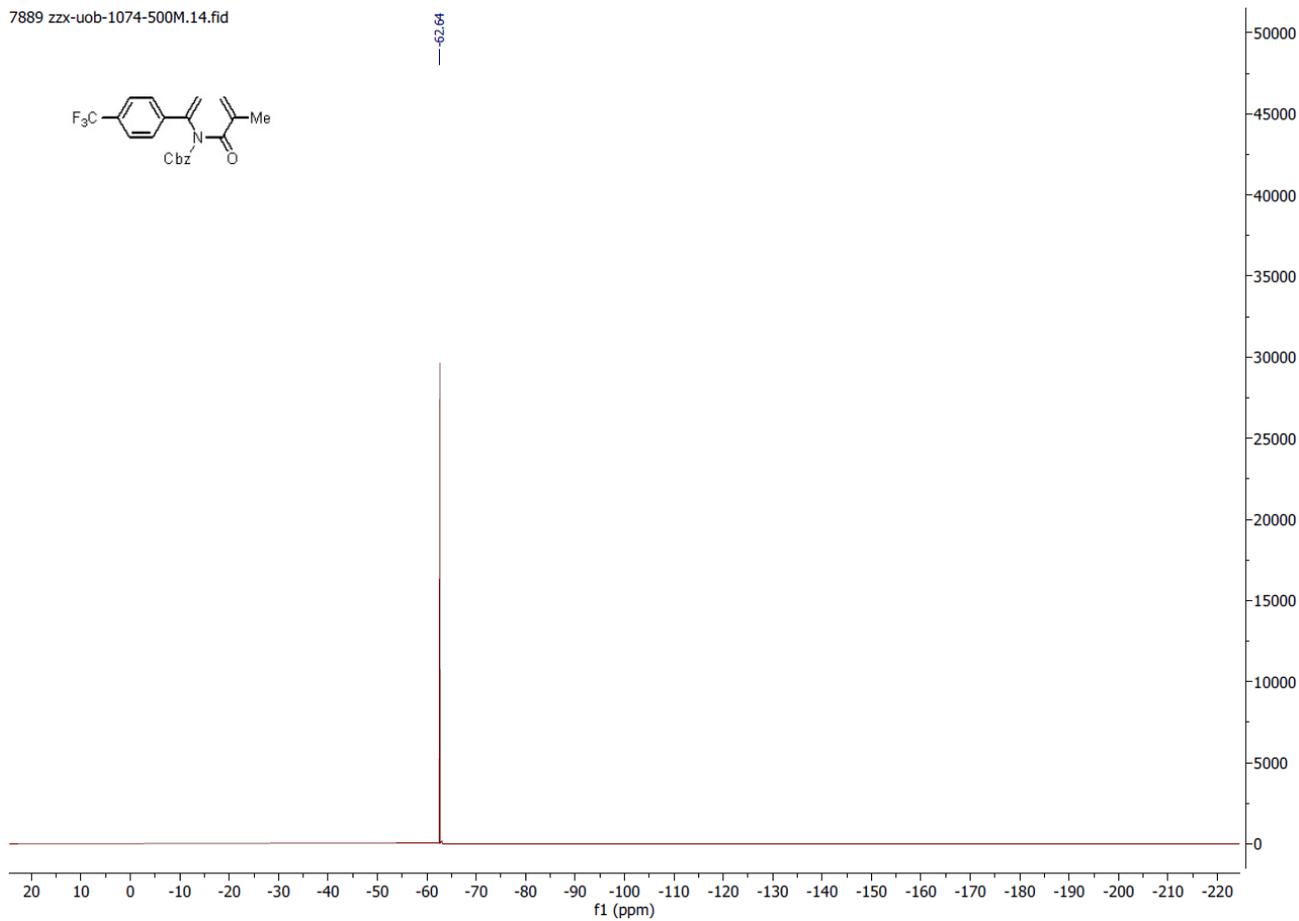
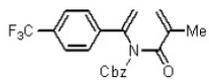
Compound 2b

7889 zzx-uob-1074-500M.10.fid



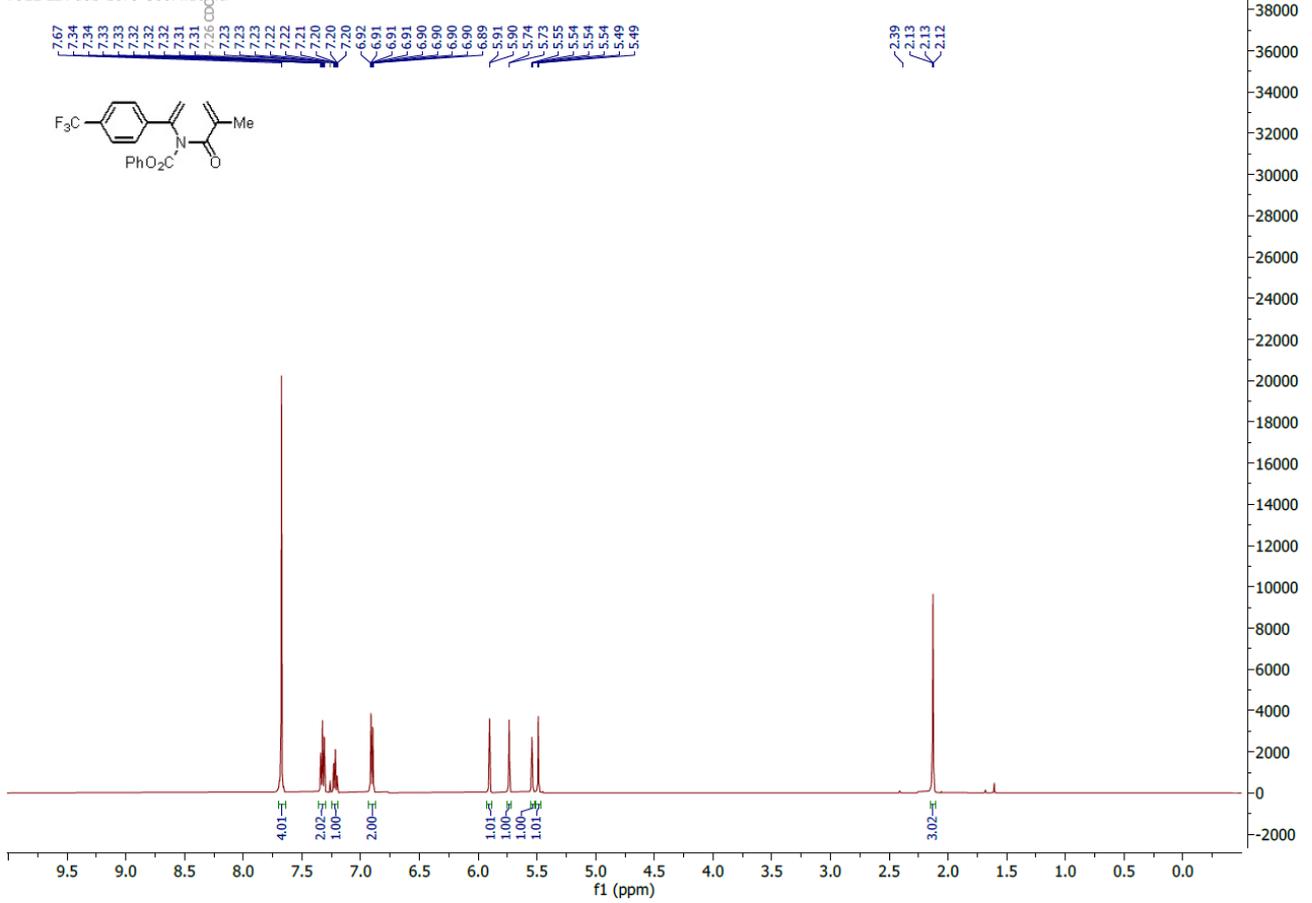
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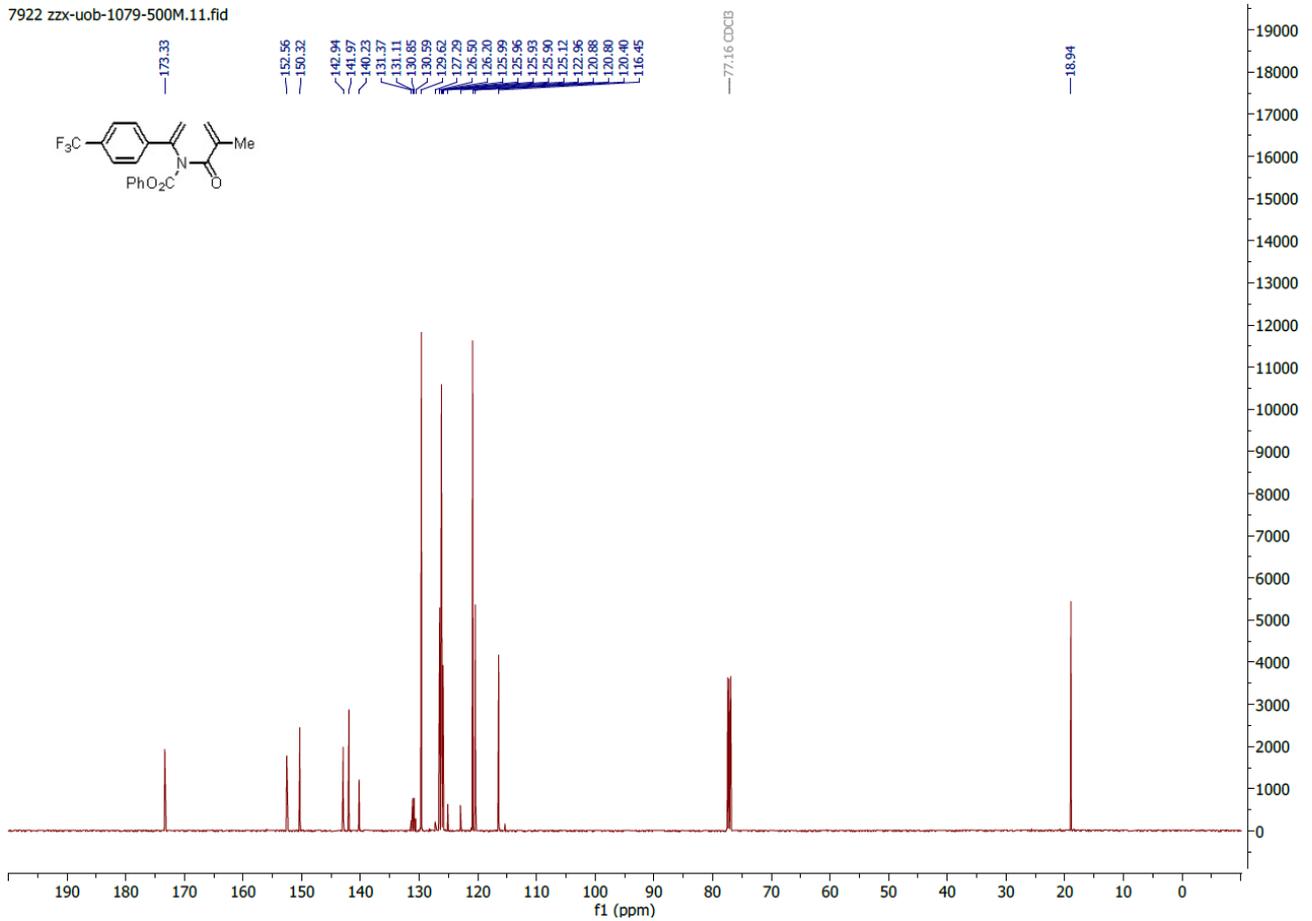


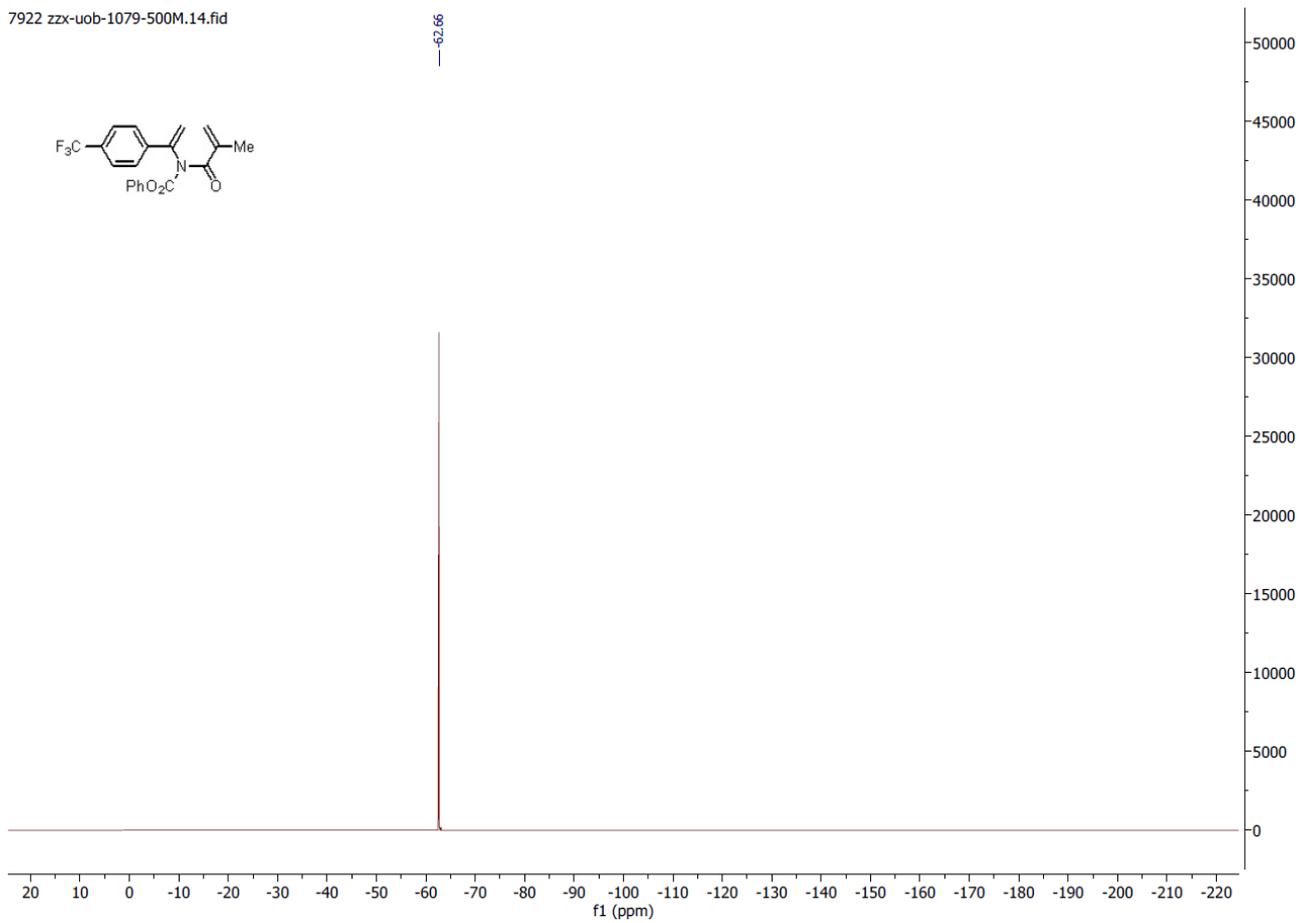
Compound 2c

7922 zzx-uob-1079-500M.10.fid



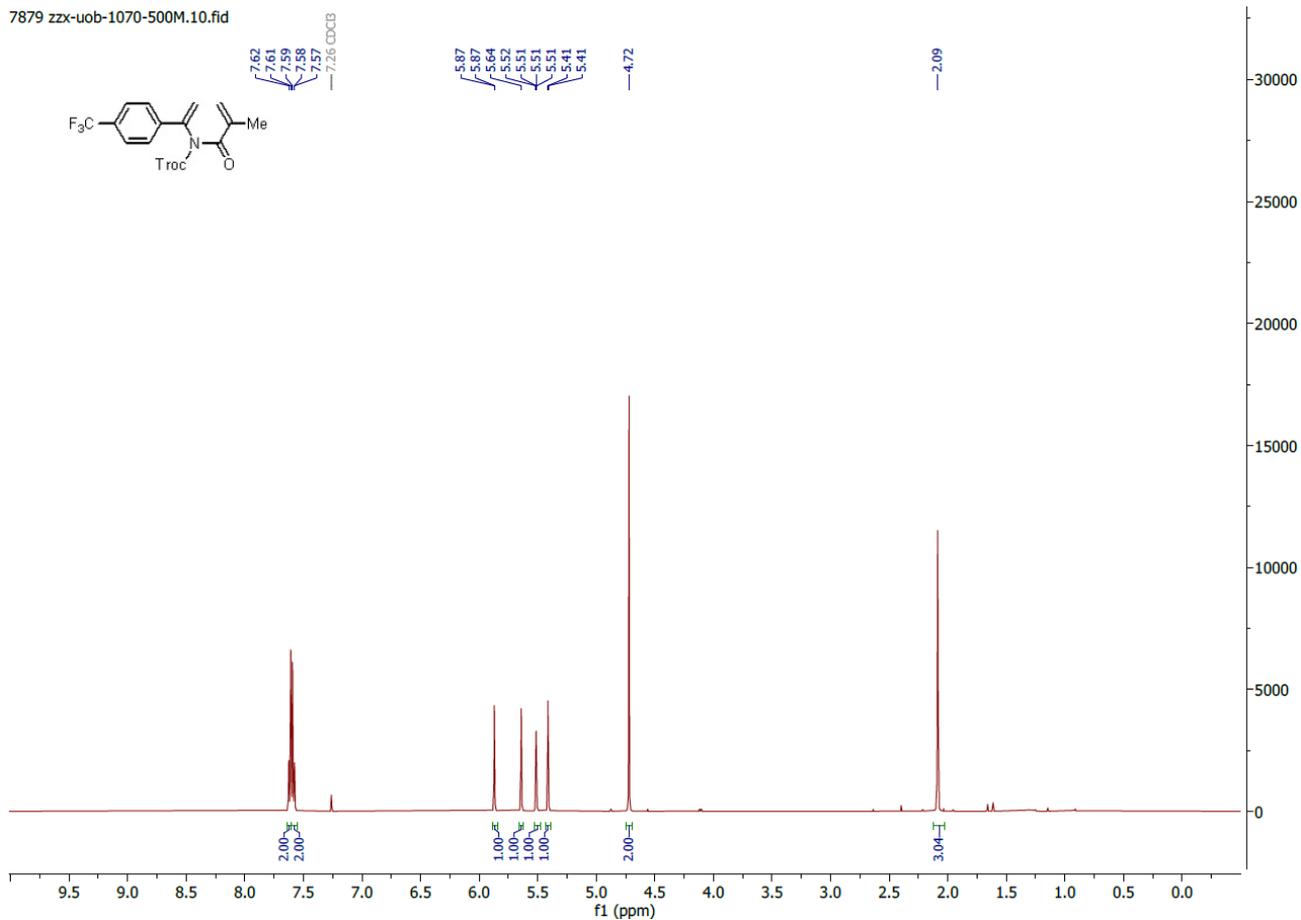
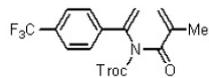
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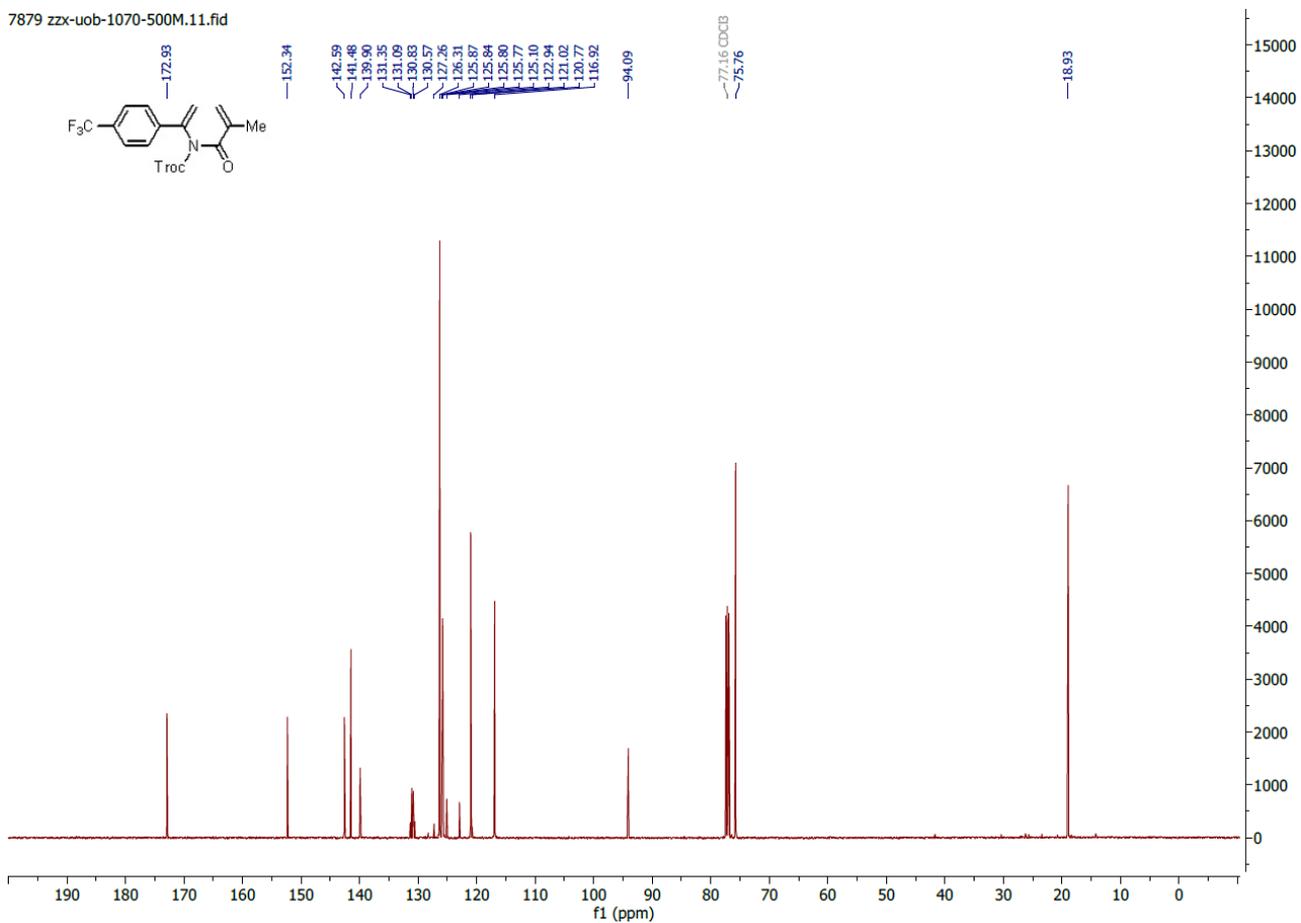
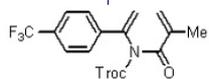


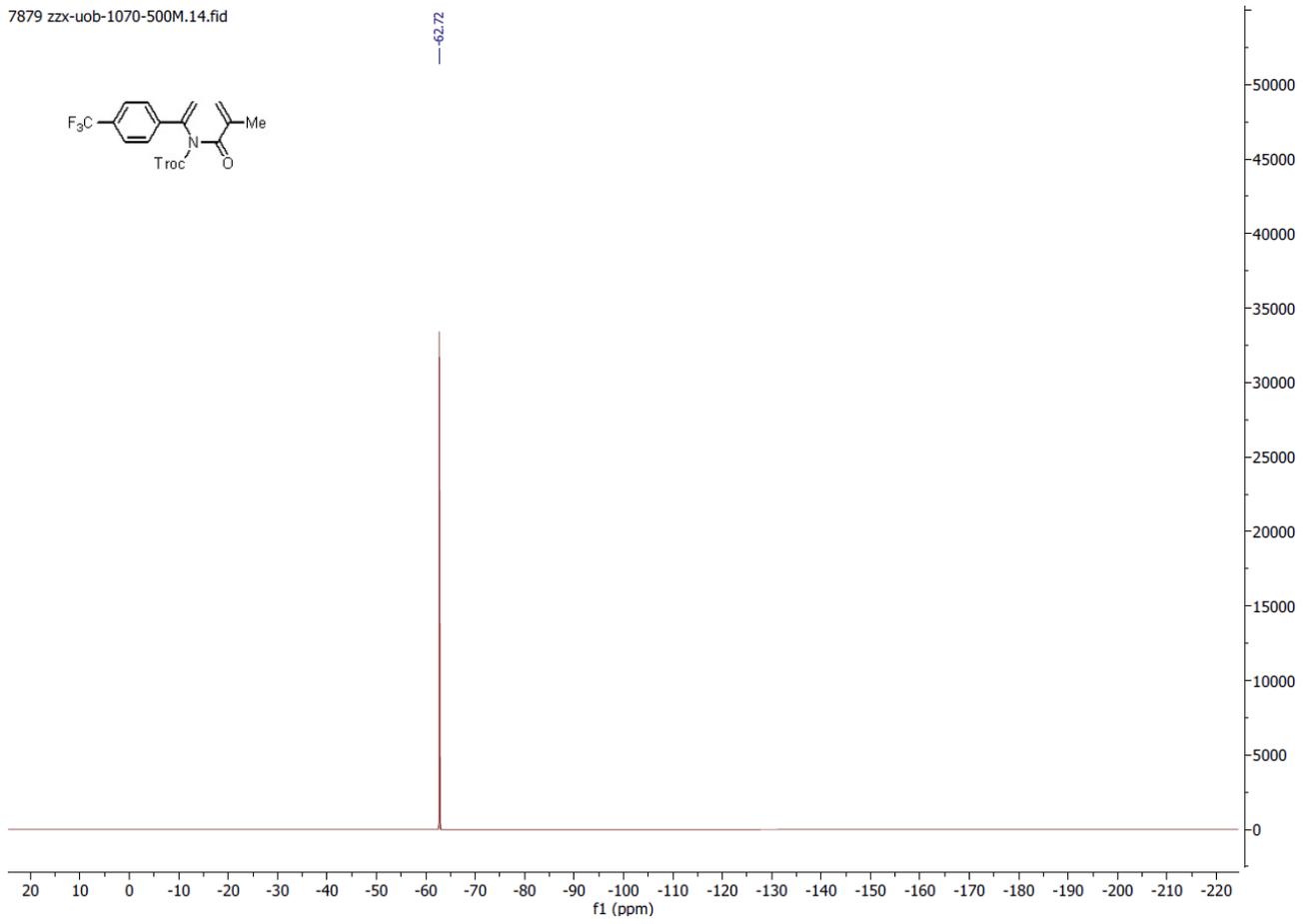
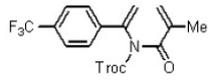
Compound 2e

7879 zzx-uob-1070-500M.10.fid



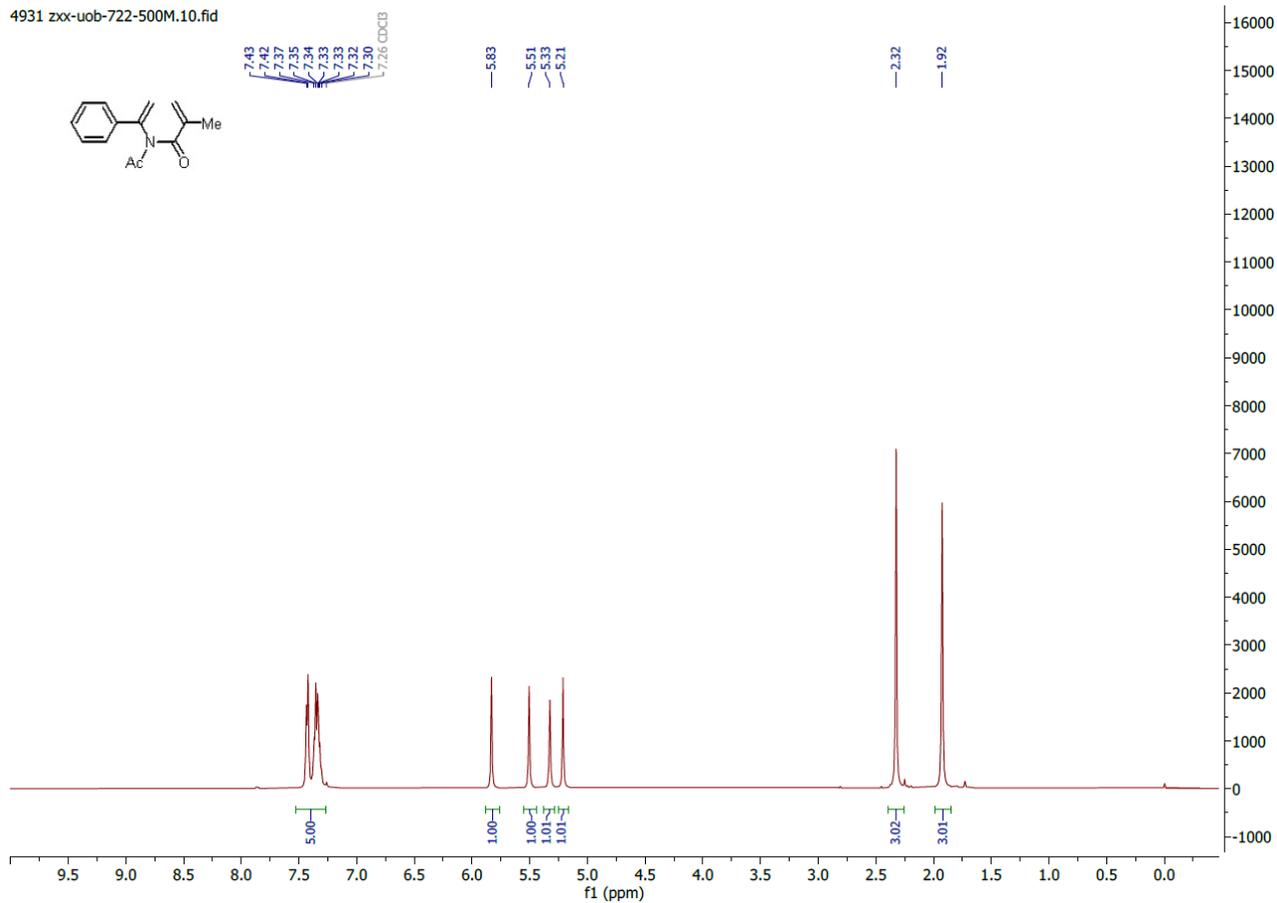
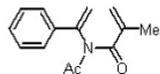
7879 zzx-uob-1070-500M.11.fid



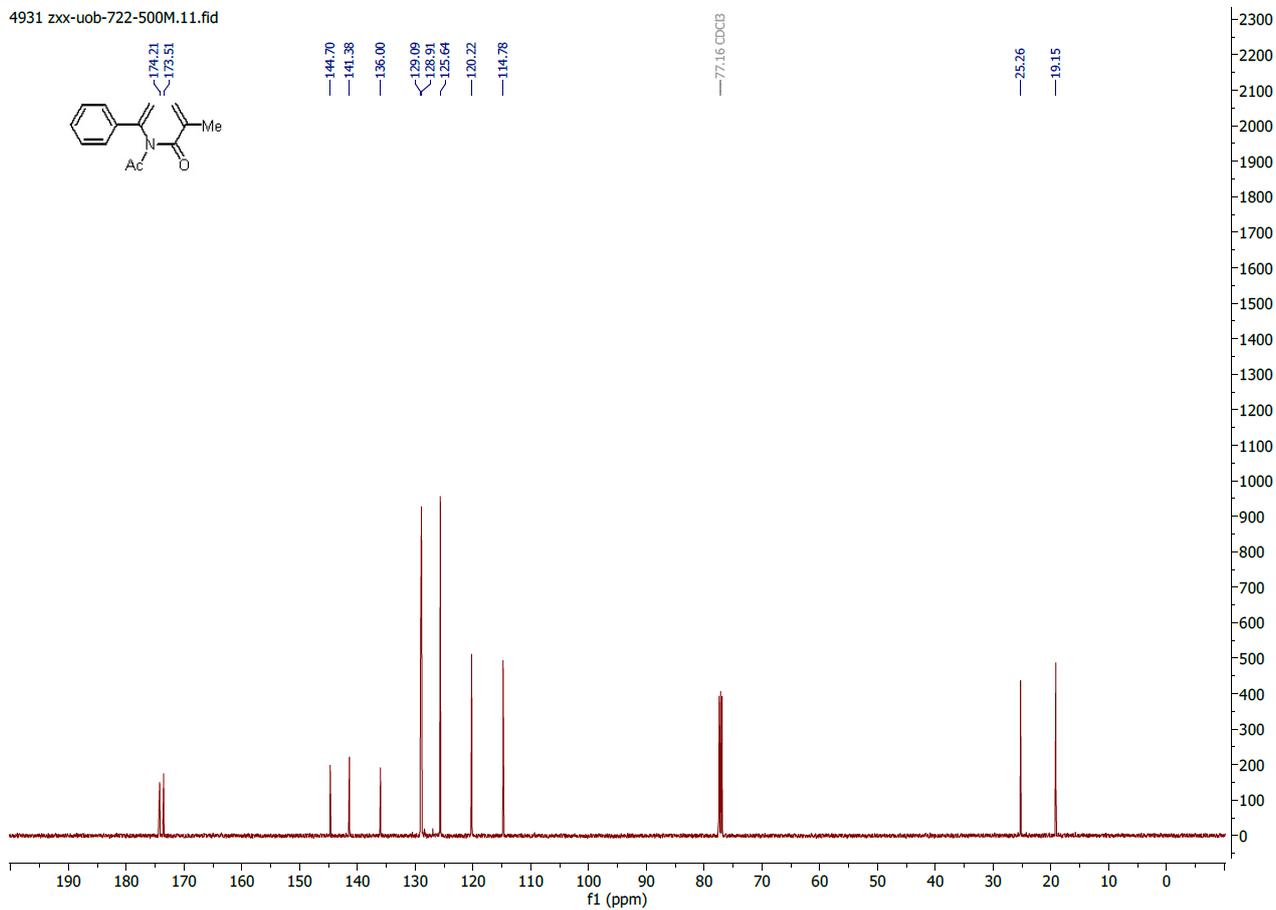
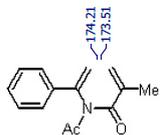


Compound 2f

4931 zxx-uob-722-500M.10.fid

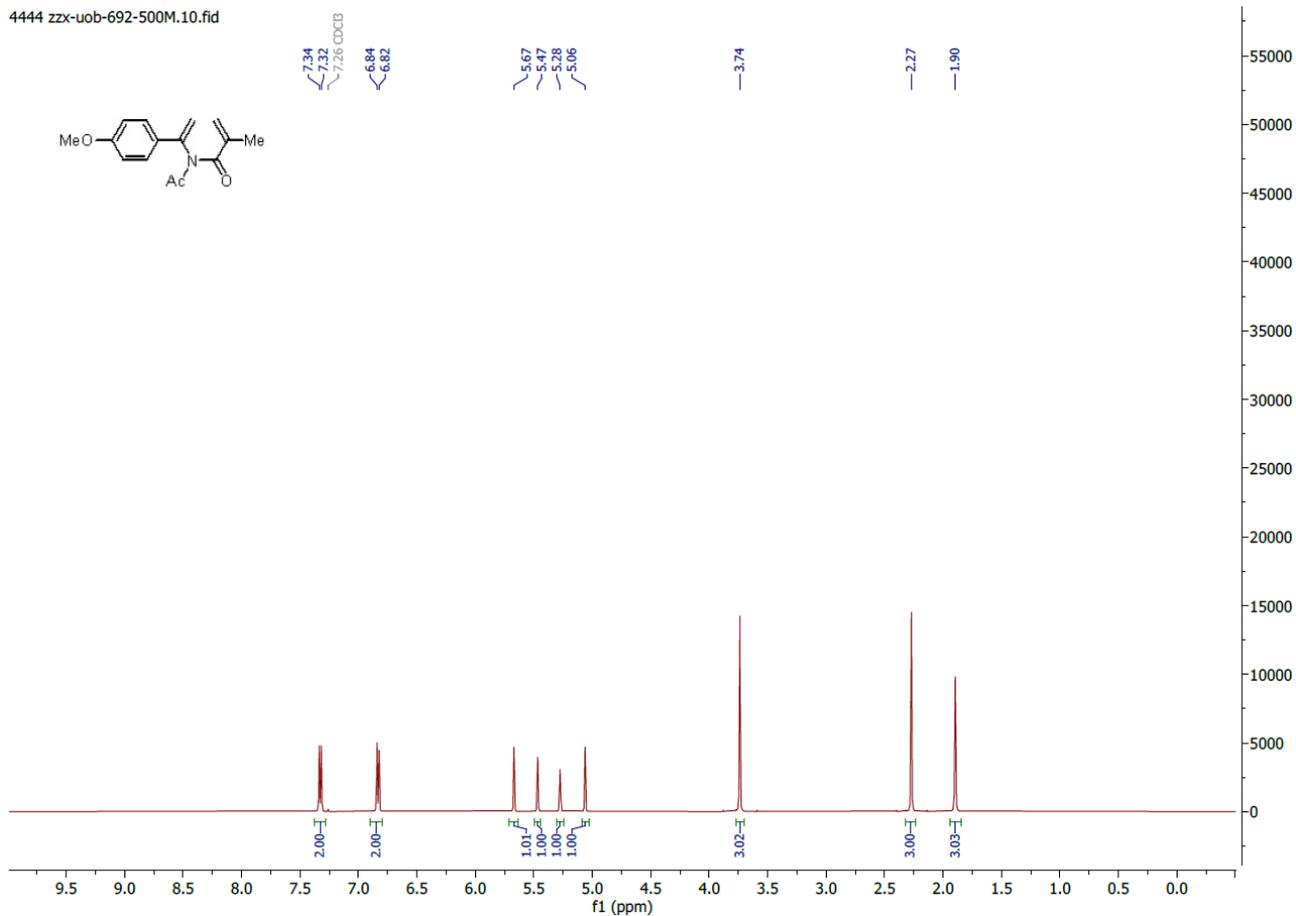


4931 zxx-uob-722-500M.11.fid

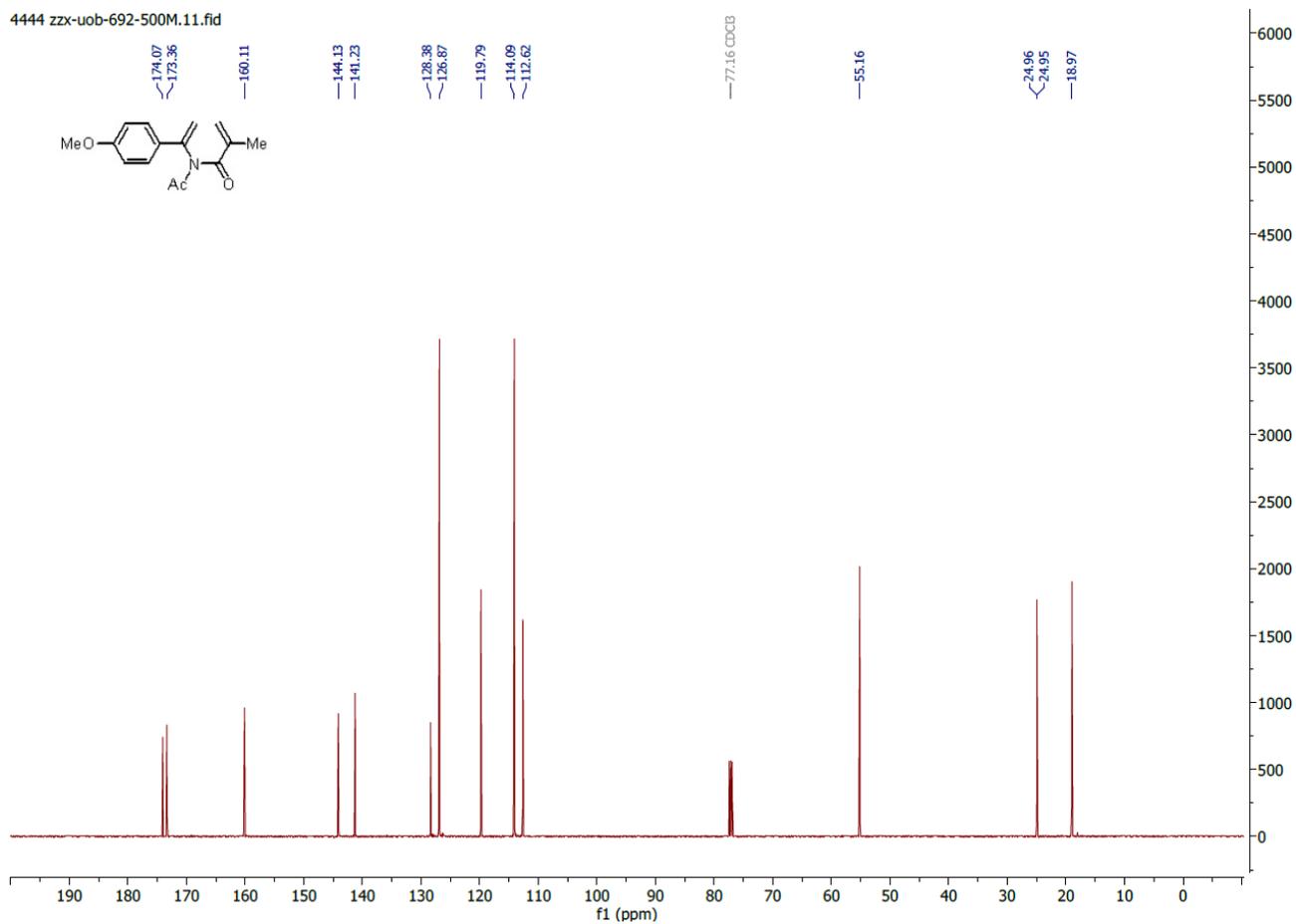


Compound 2g

4444 zzx-uob-692-500M.10.fid

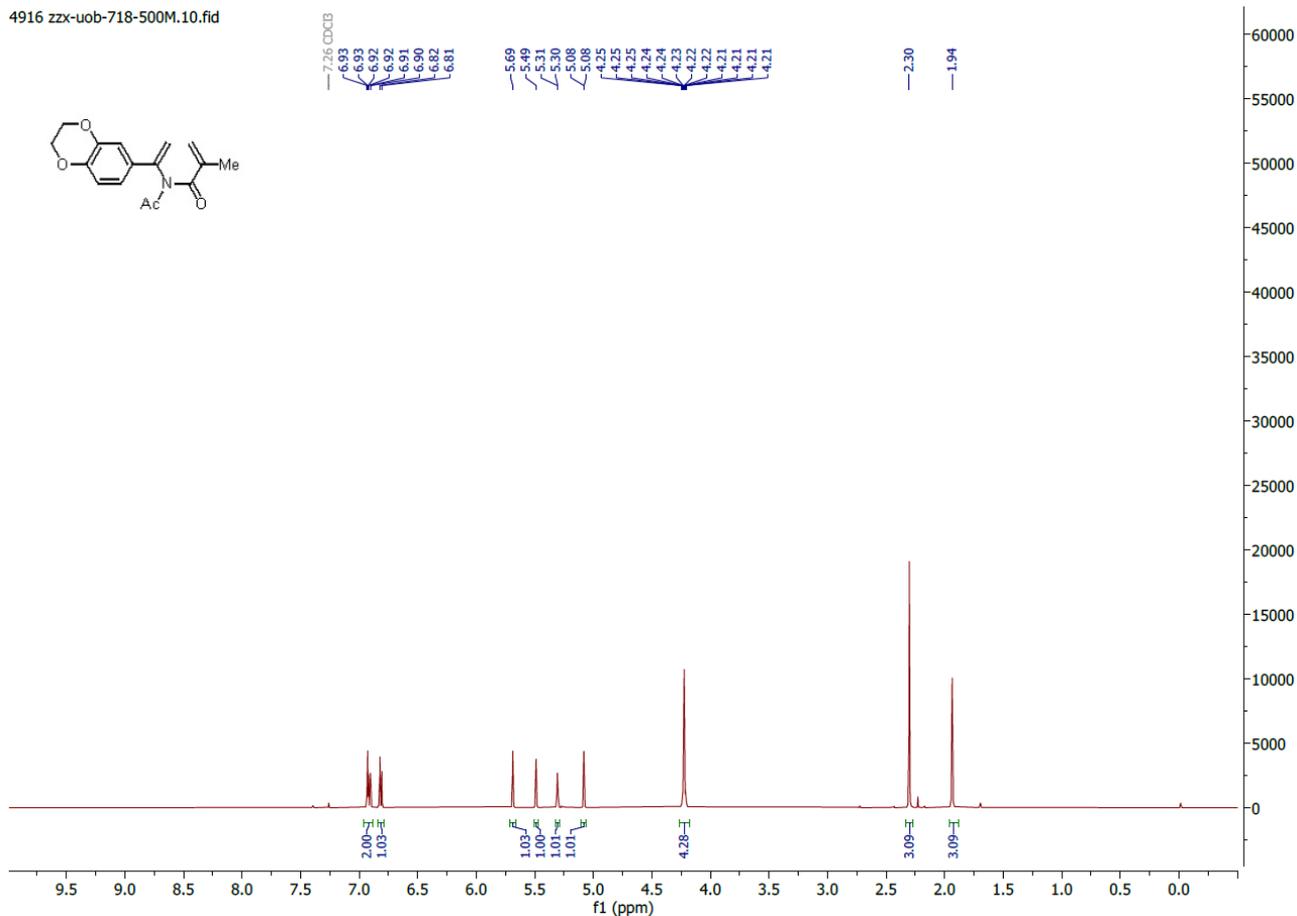
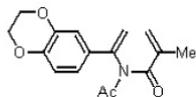


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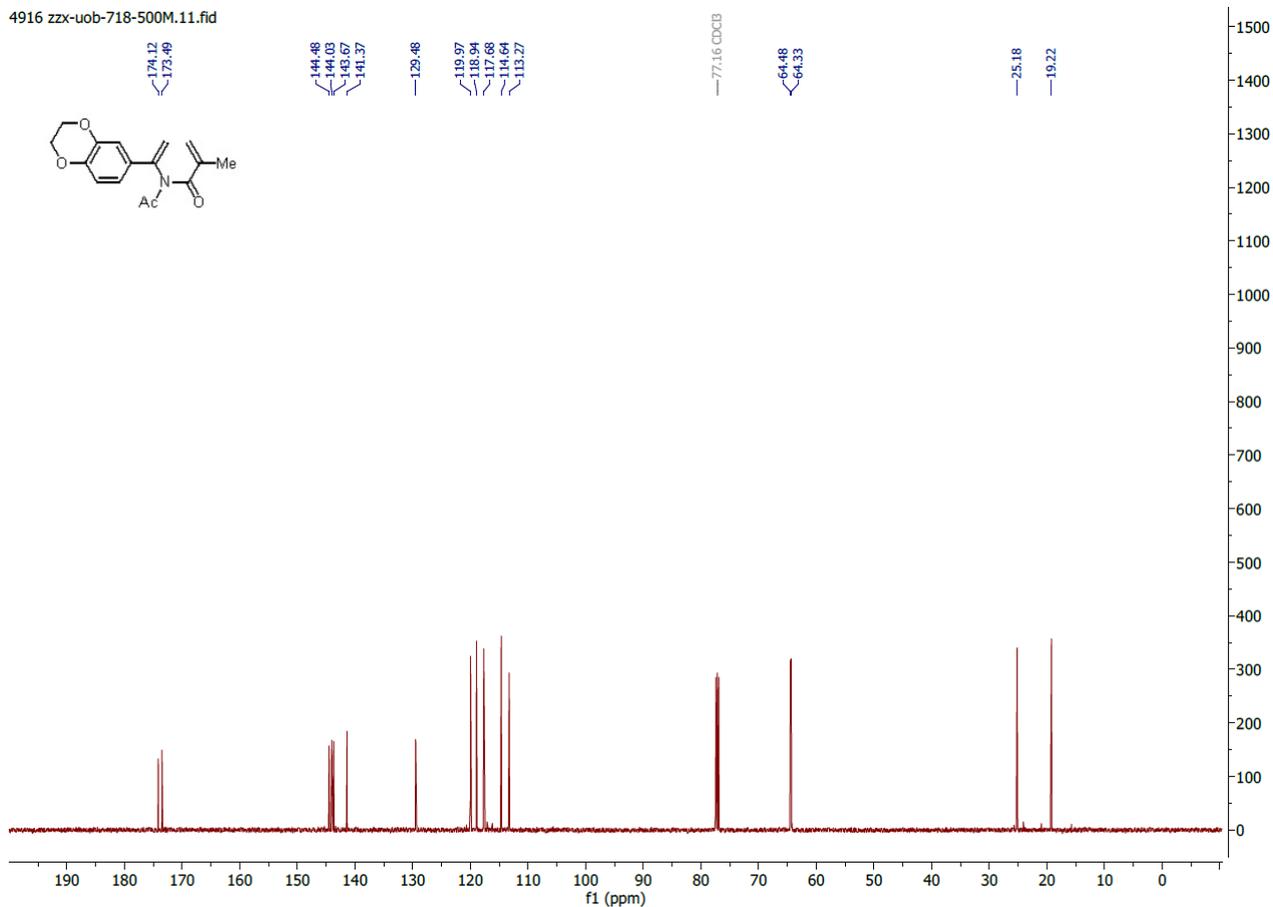
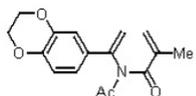


Compound 2h

4916 zzx-uob-718-500M.10.fid

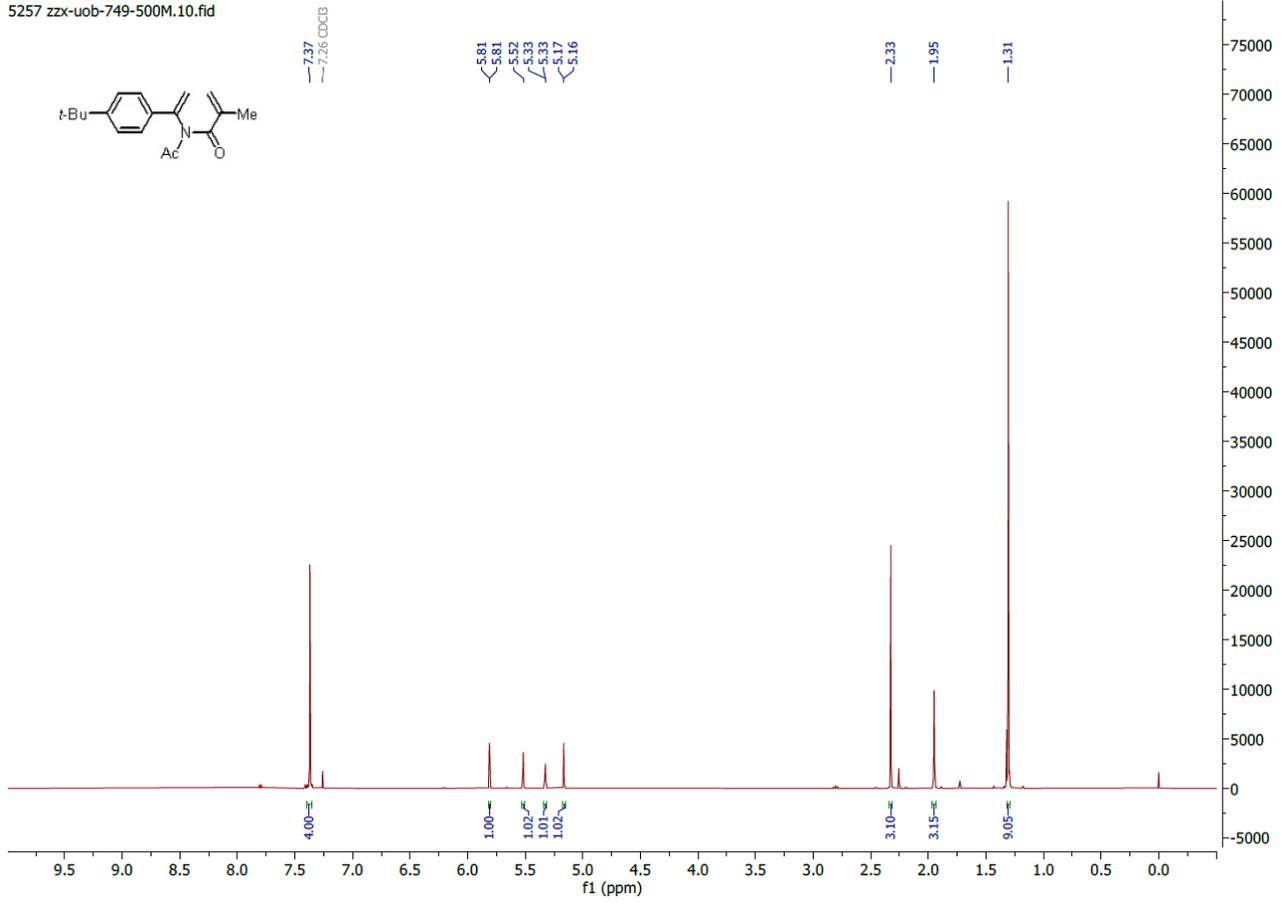


4916 zzx-uob-718-500M.11.fid

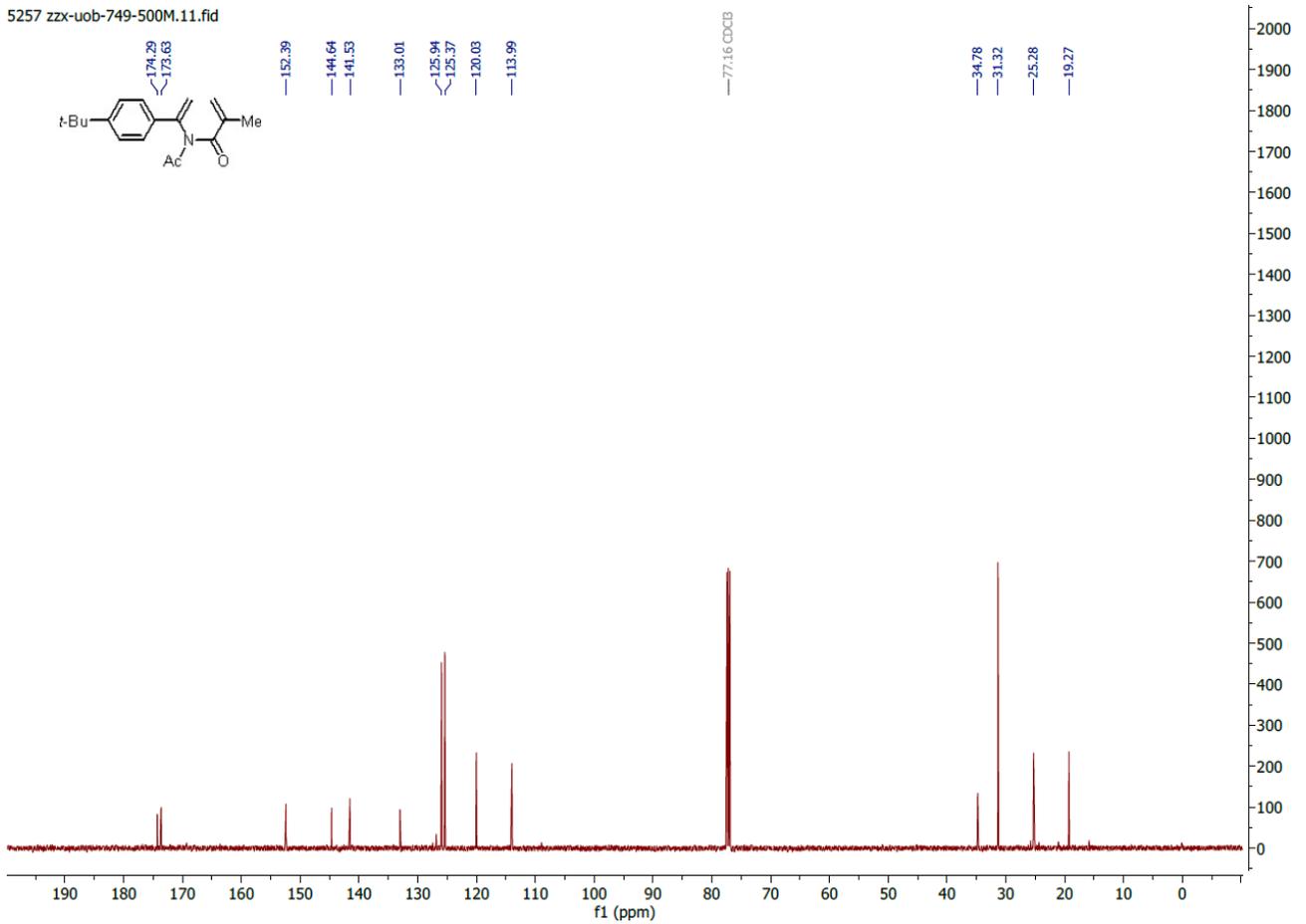


Compound 2i

5257 zzx-uob-749-500M.10.fid

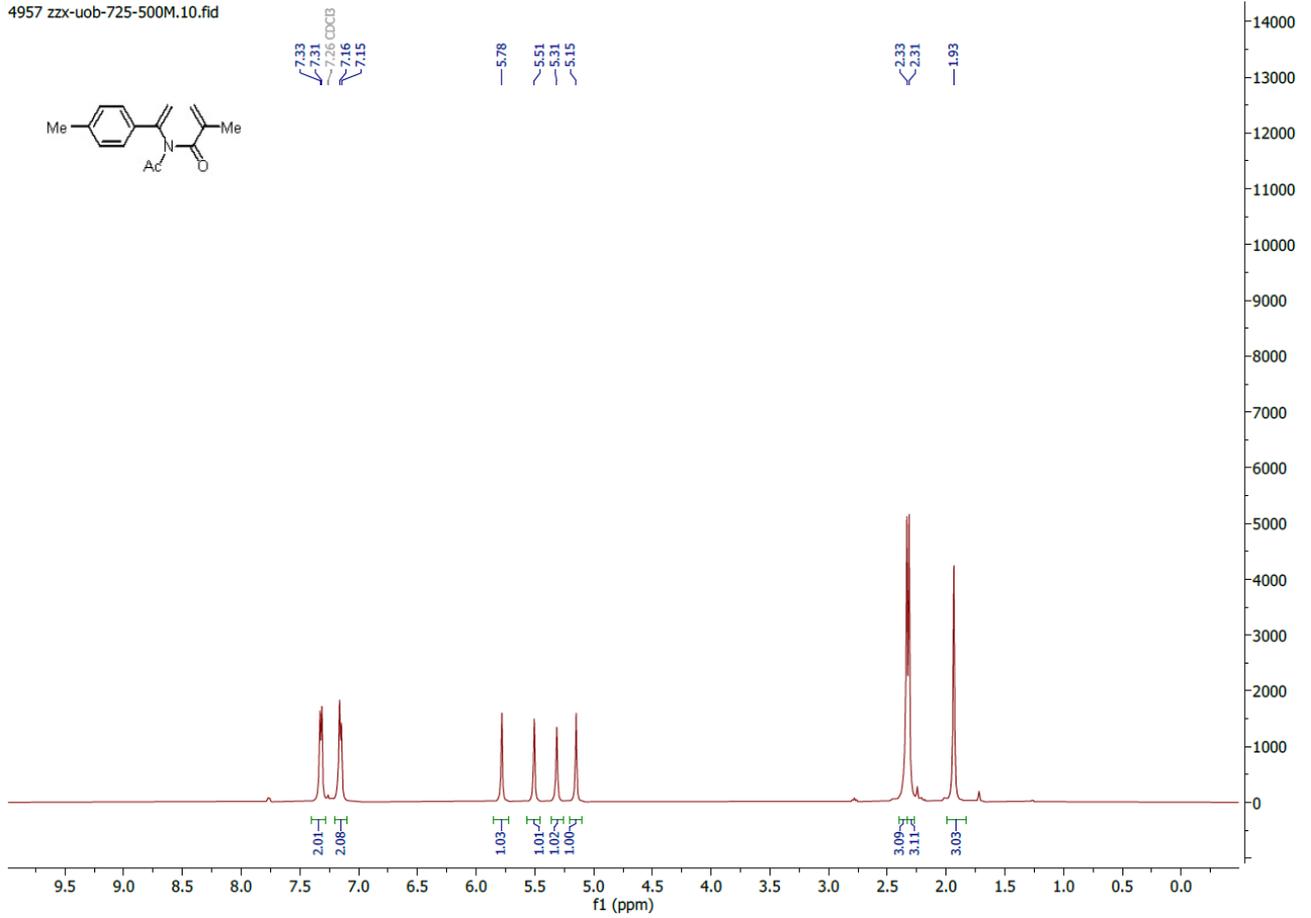
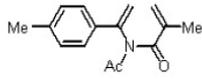


5257 zzx-uob-749-500M.11.fid

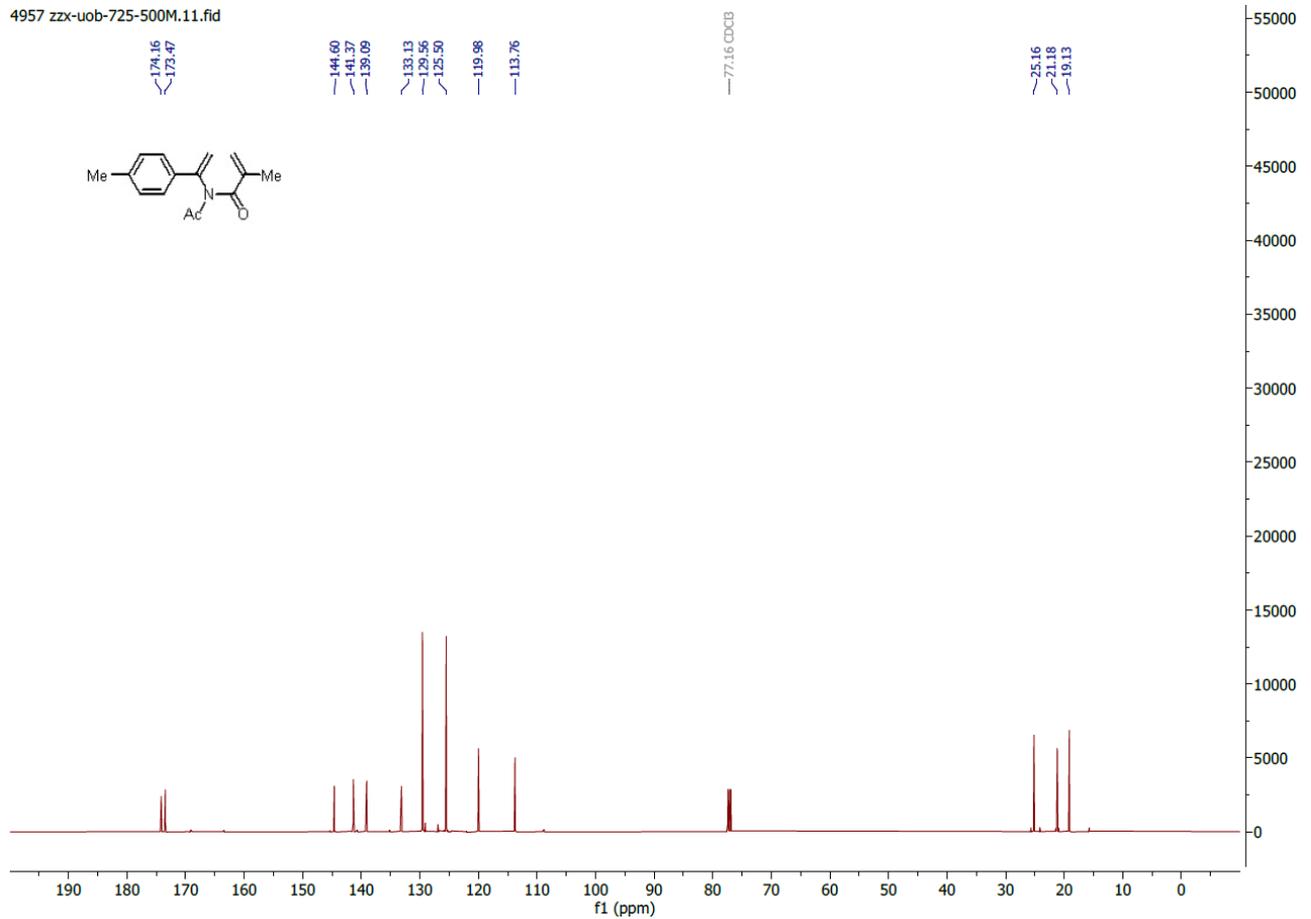
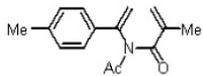


Compound 2j

4957 zzx-uob-725-500M.10.fid

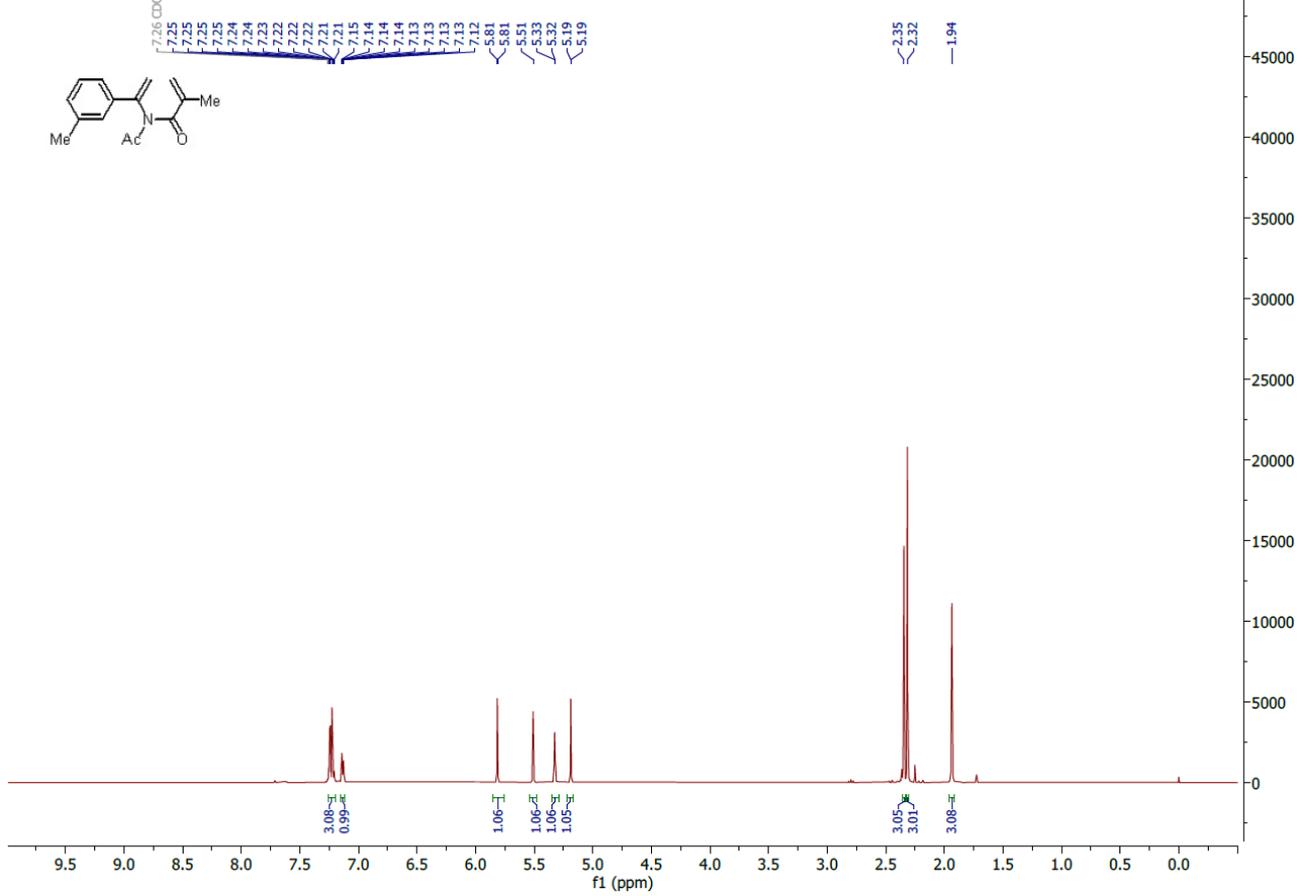


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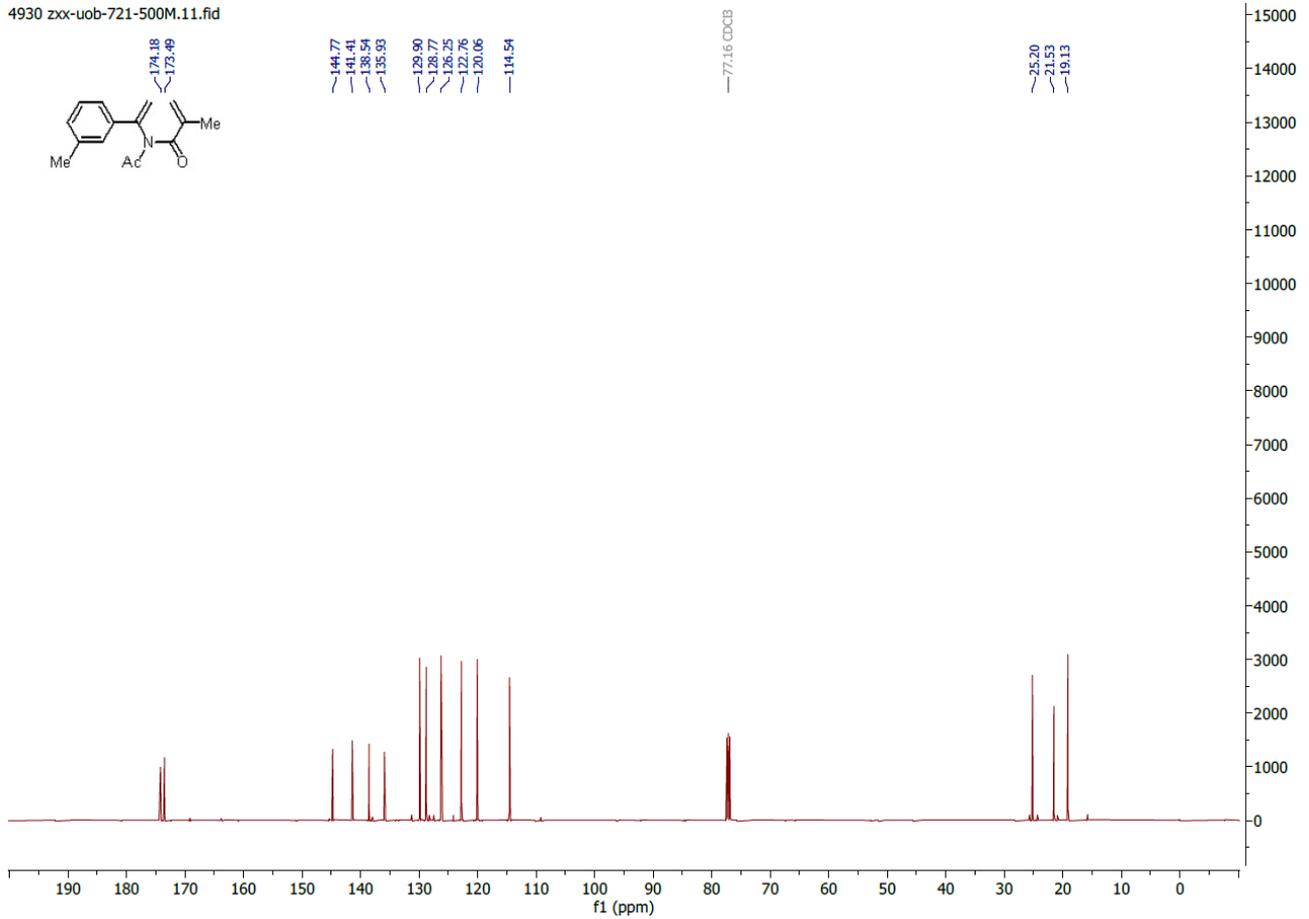


Compound k

4930 zxx-uob-721-500M.10.fid

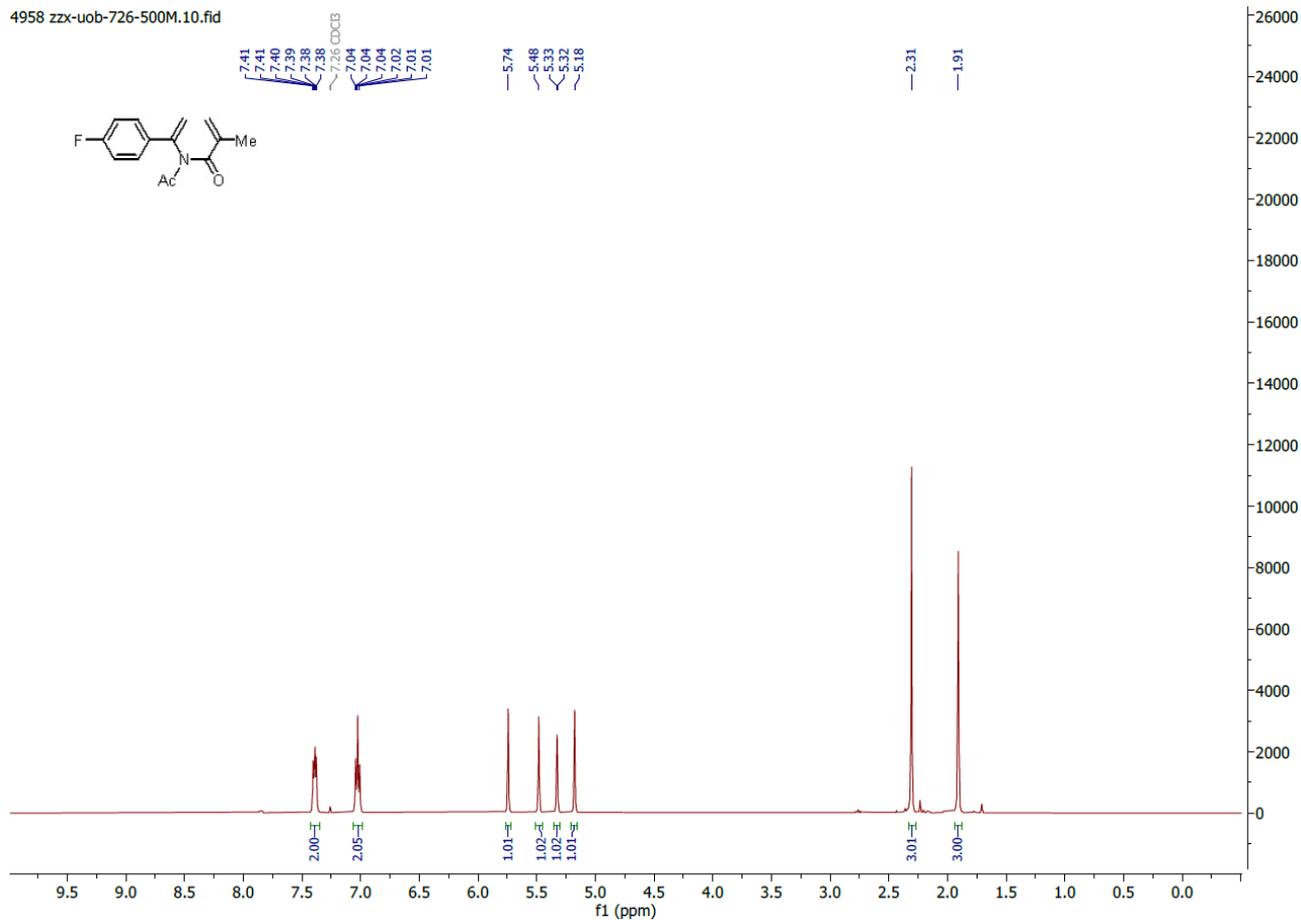


4930 zxx-uob-721-500M.11.fid

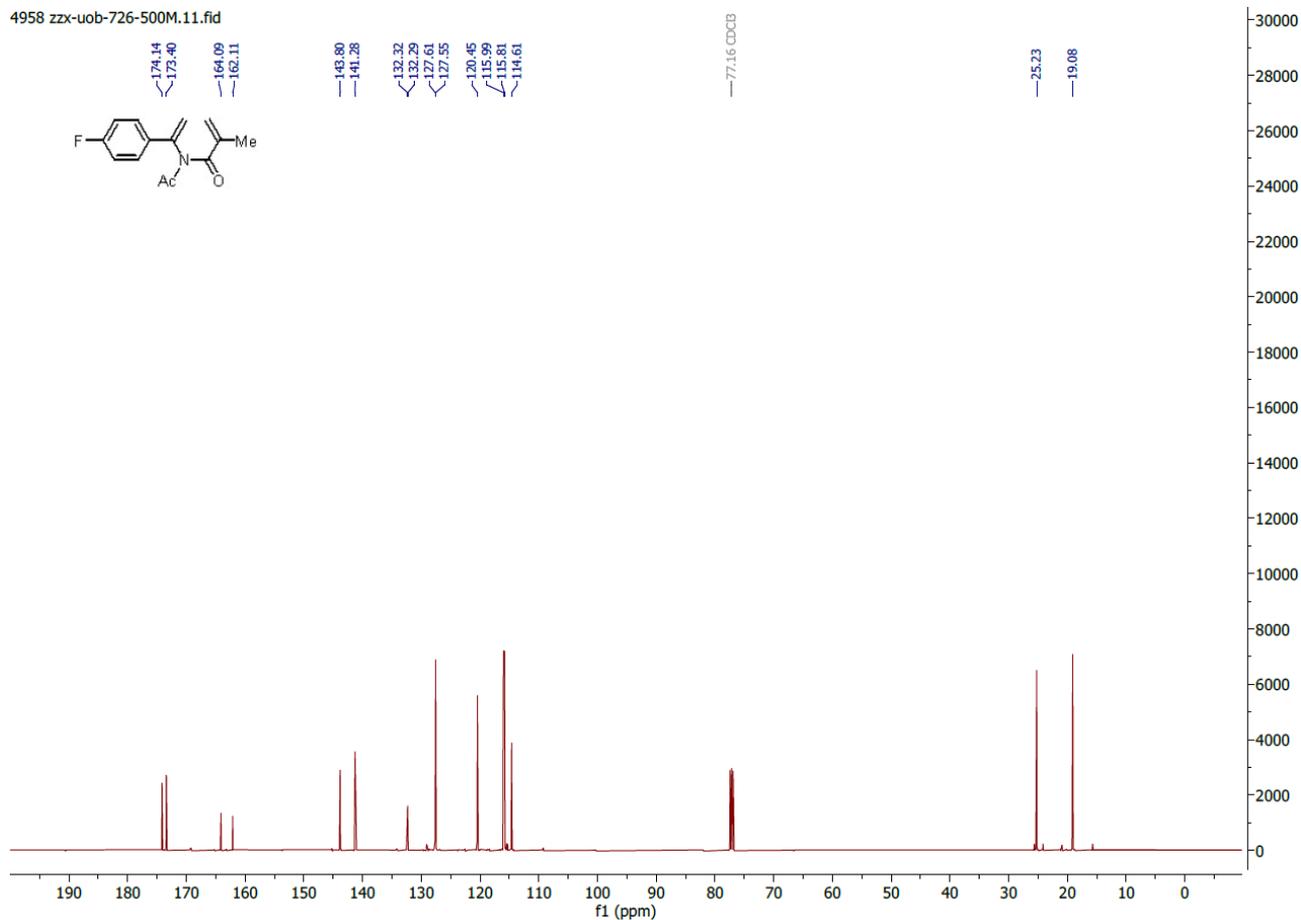


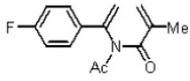
Compound 21

4958 zzx-uob-726-500M.10.fid

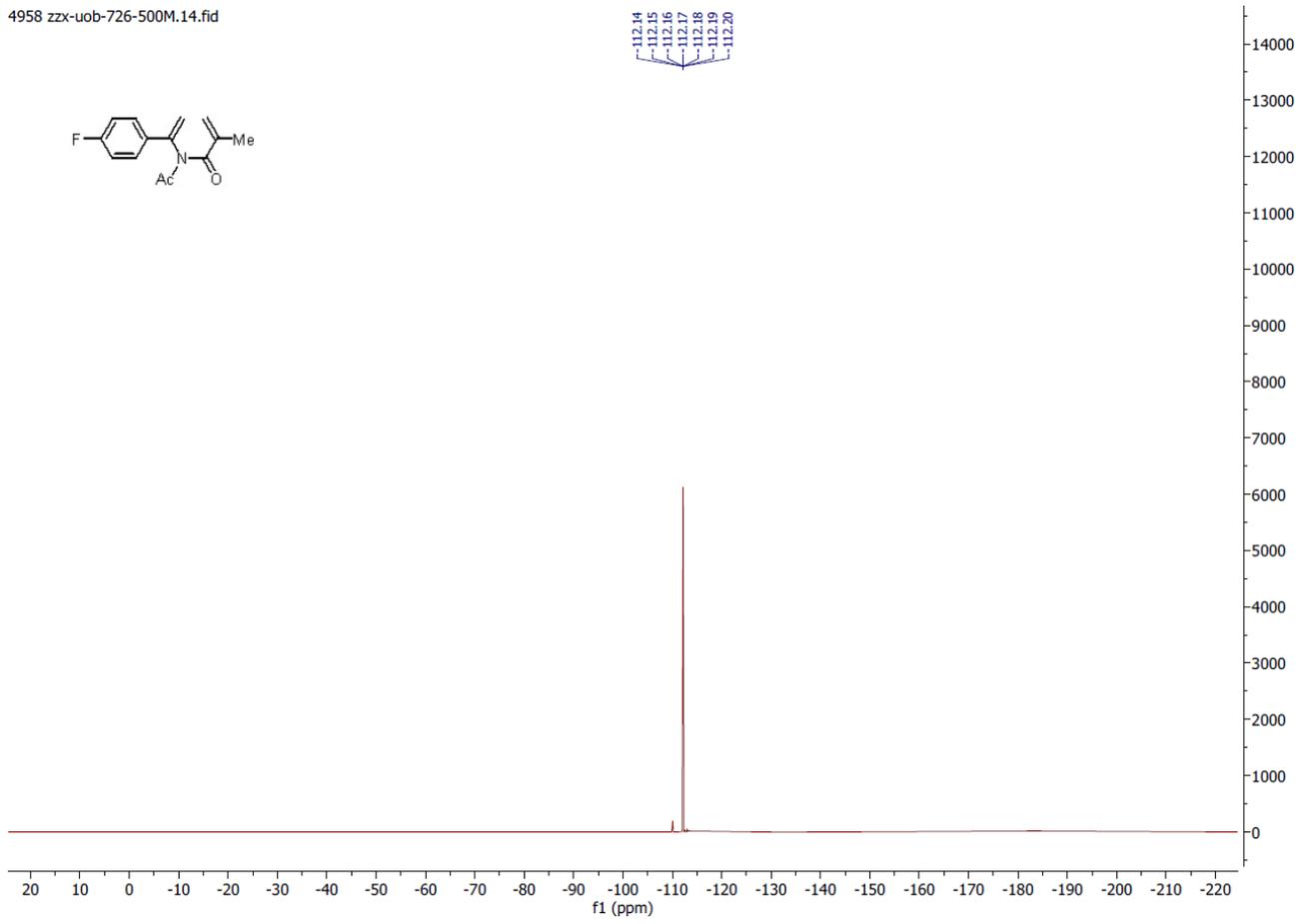


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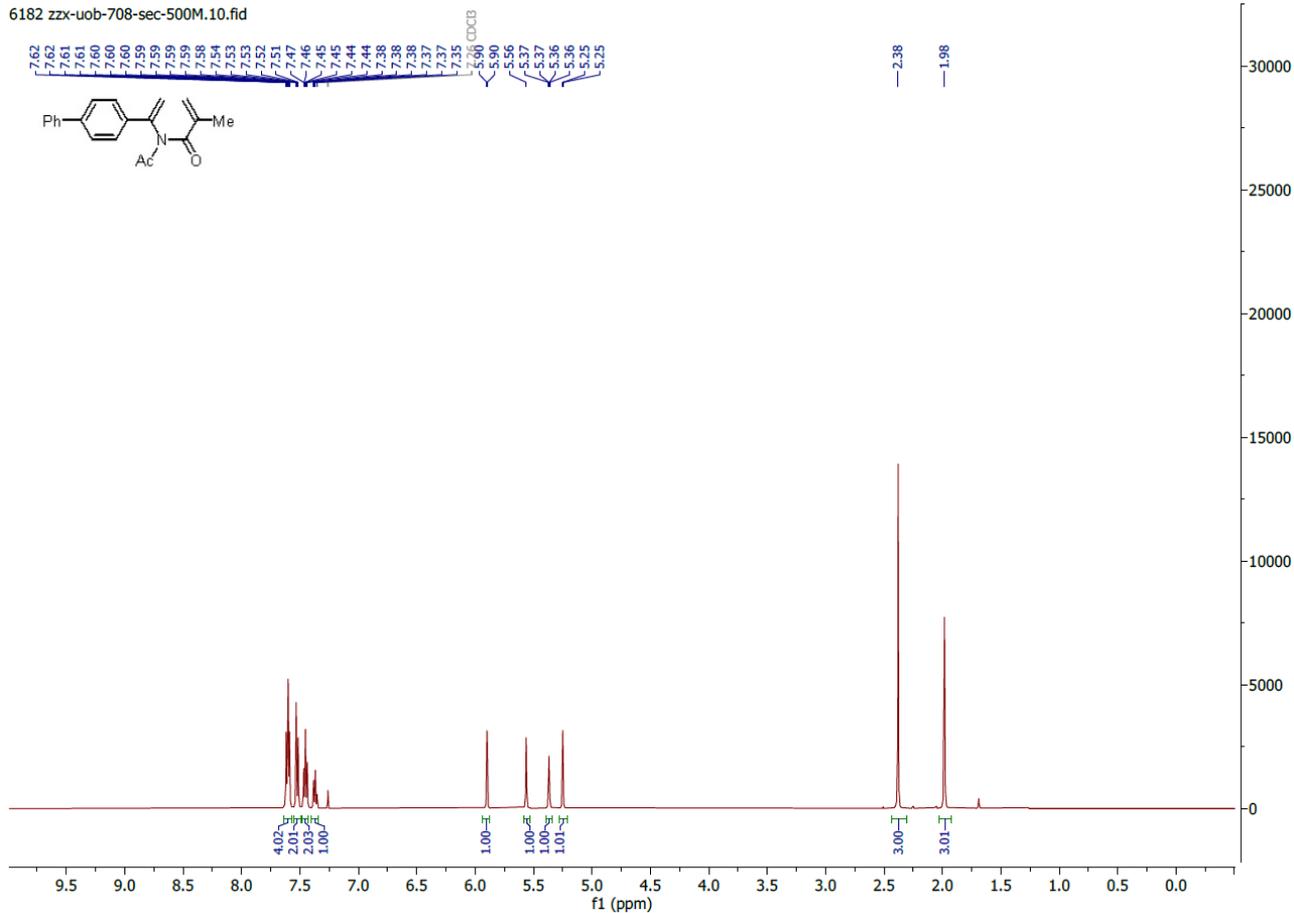


112.14
112.15
112.16
112.17
112.18
112.19
112.20

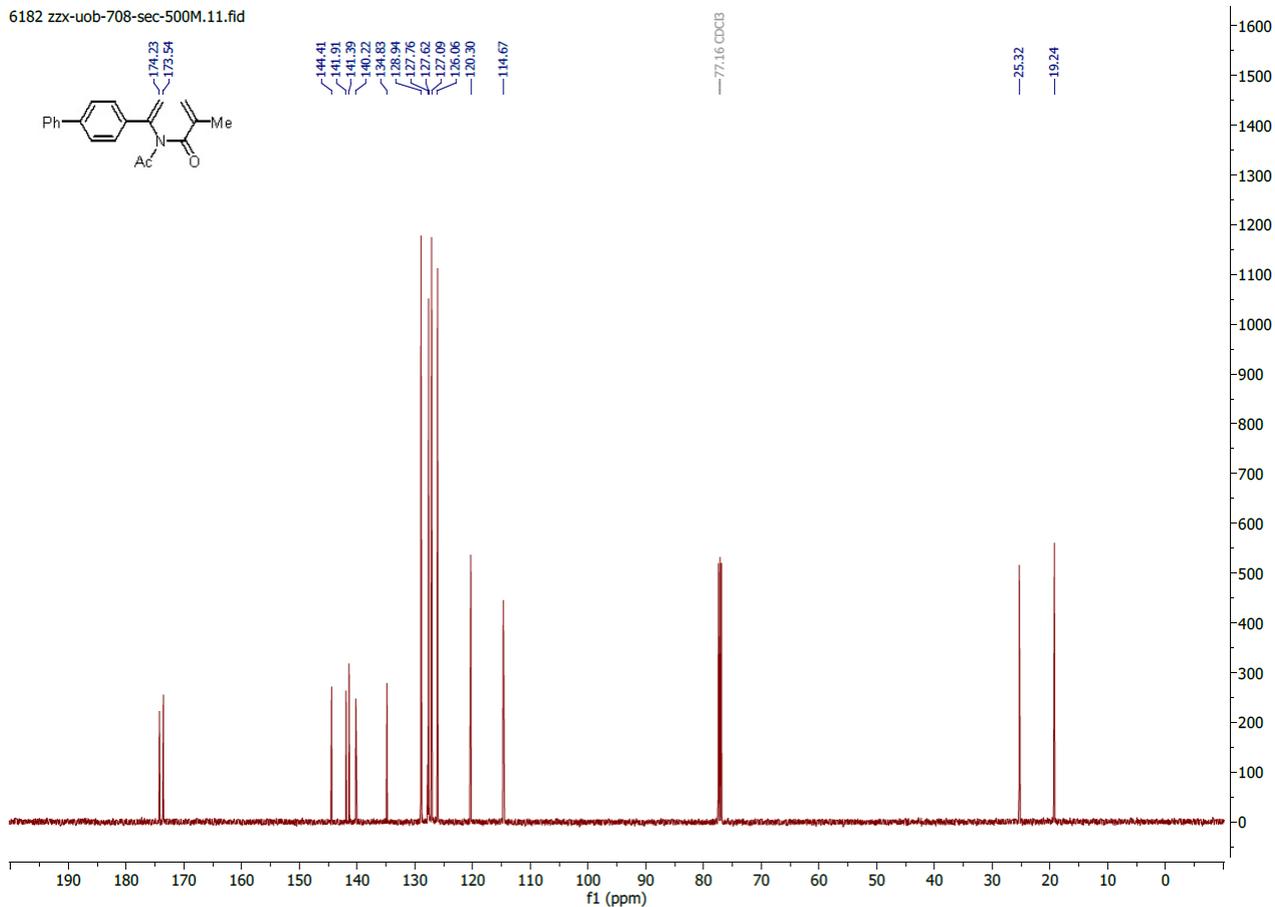


Compound 2m

6182 zzx-uob-708-sec-500M.10.fid

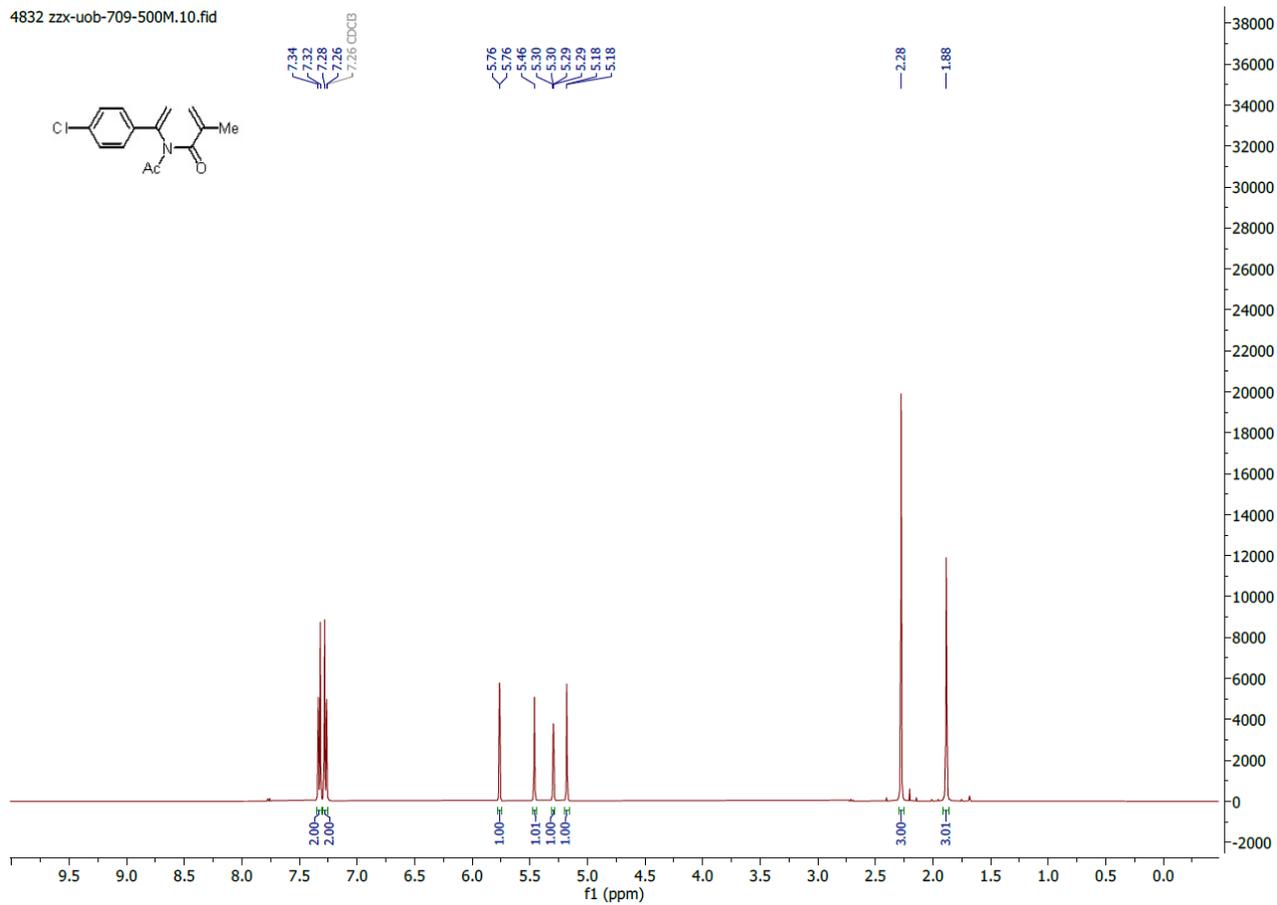
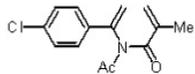


6182 zzx-uob-708-sec-500M.11.fid

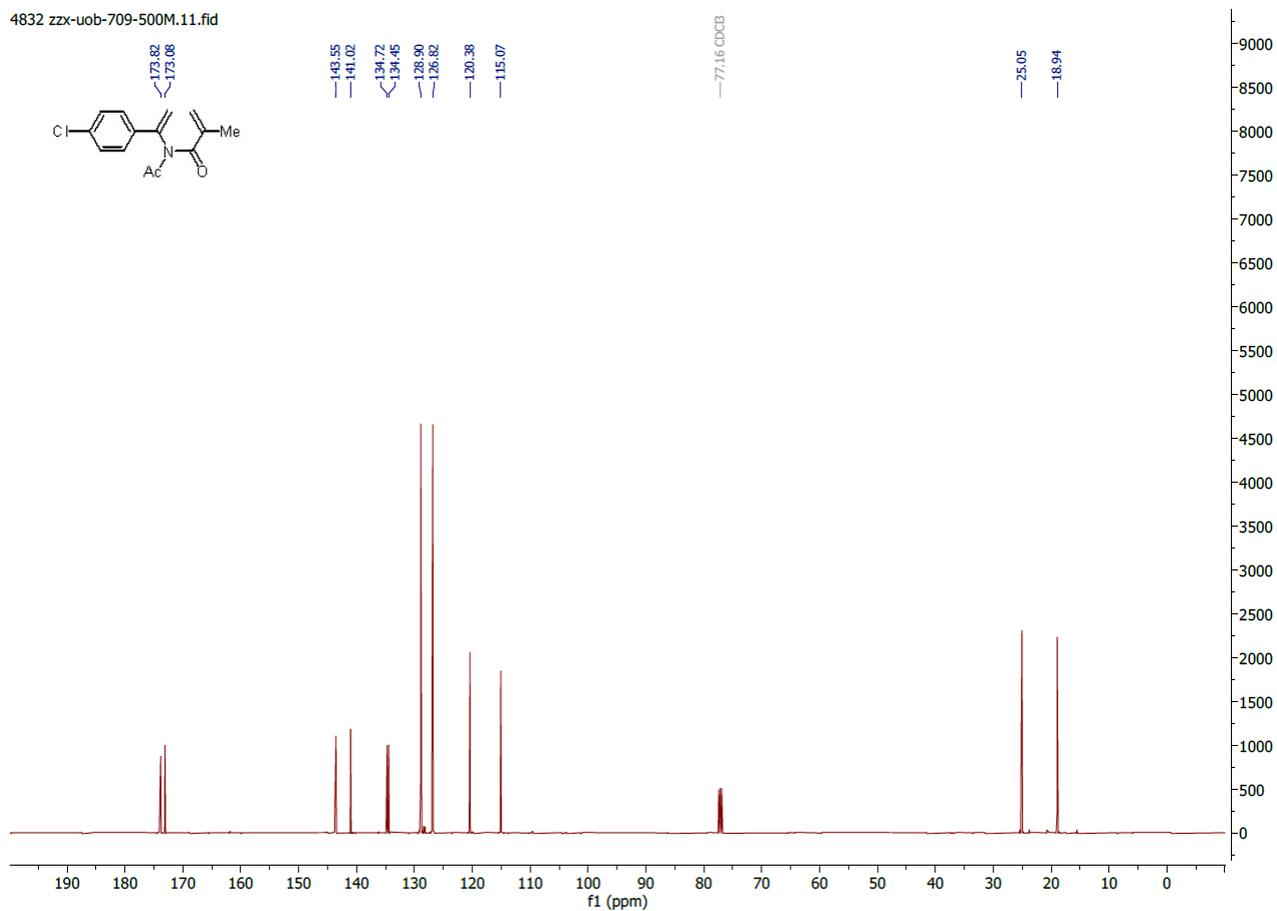
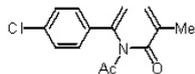


Compound 2n

4832 zzx-uob-709-500M.10.fid

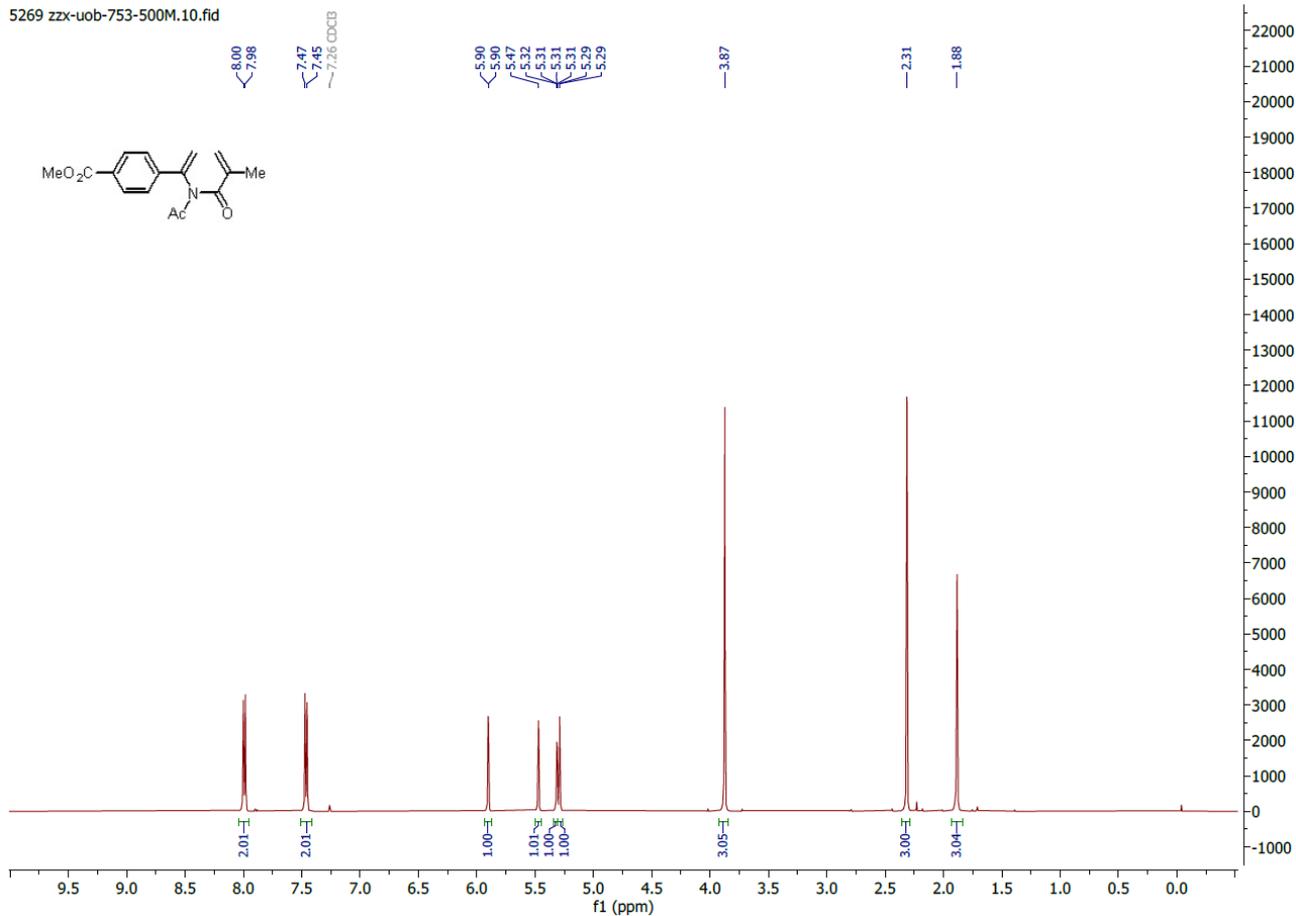


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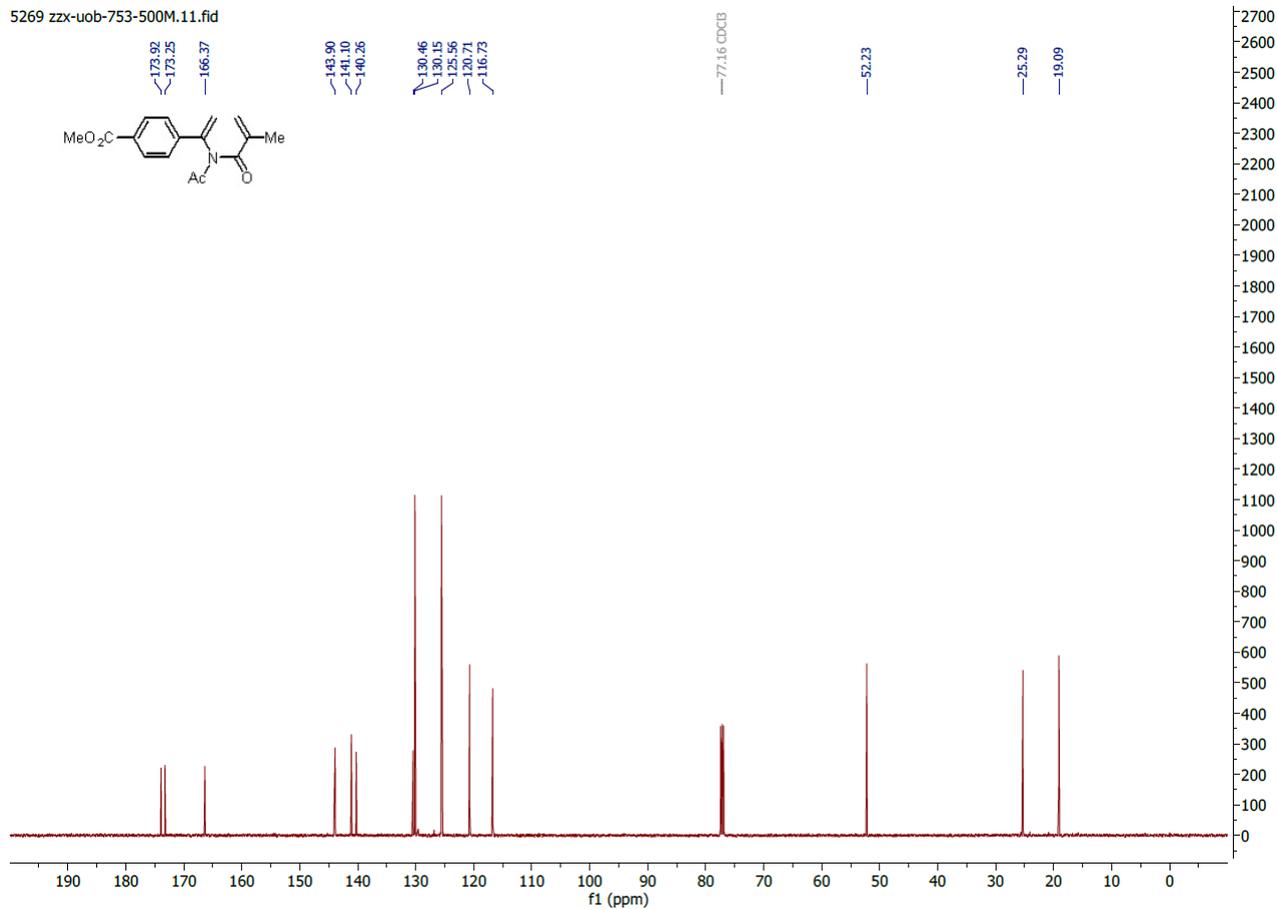


Compound 2o

5269 zzx-uob-753-500M.10.fid

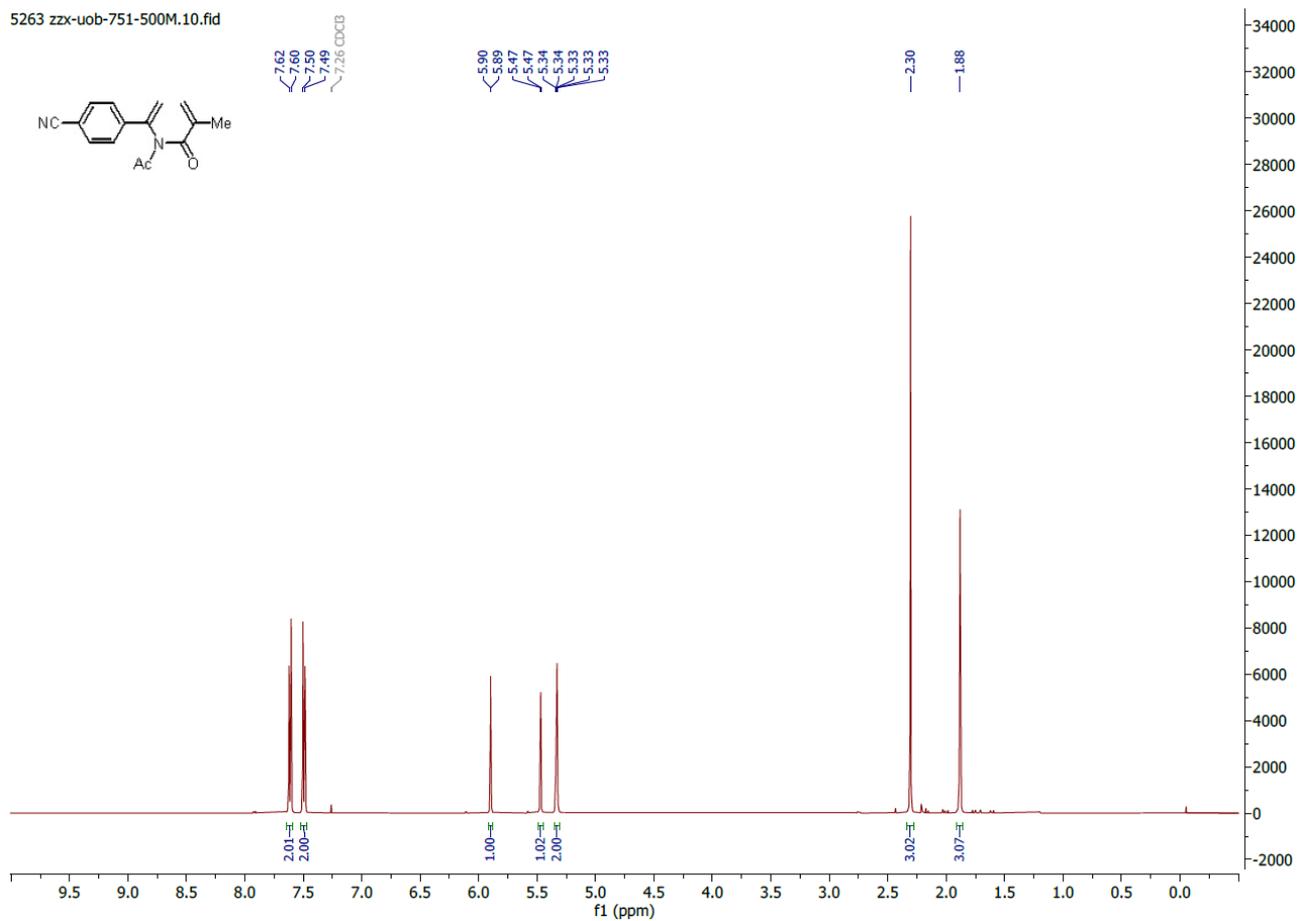


5269 zzx-uob-753-500M.11.fid

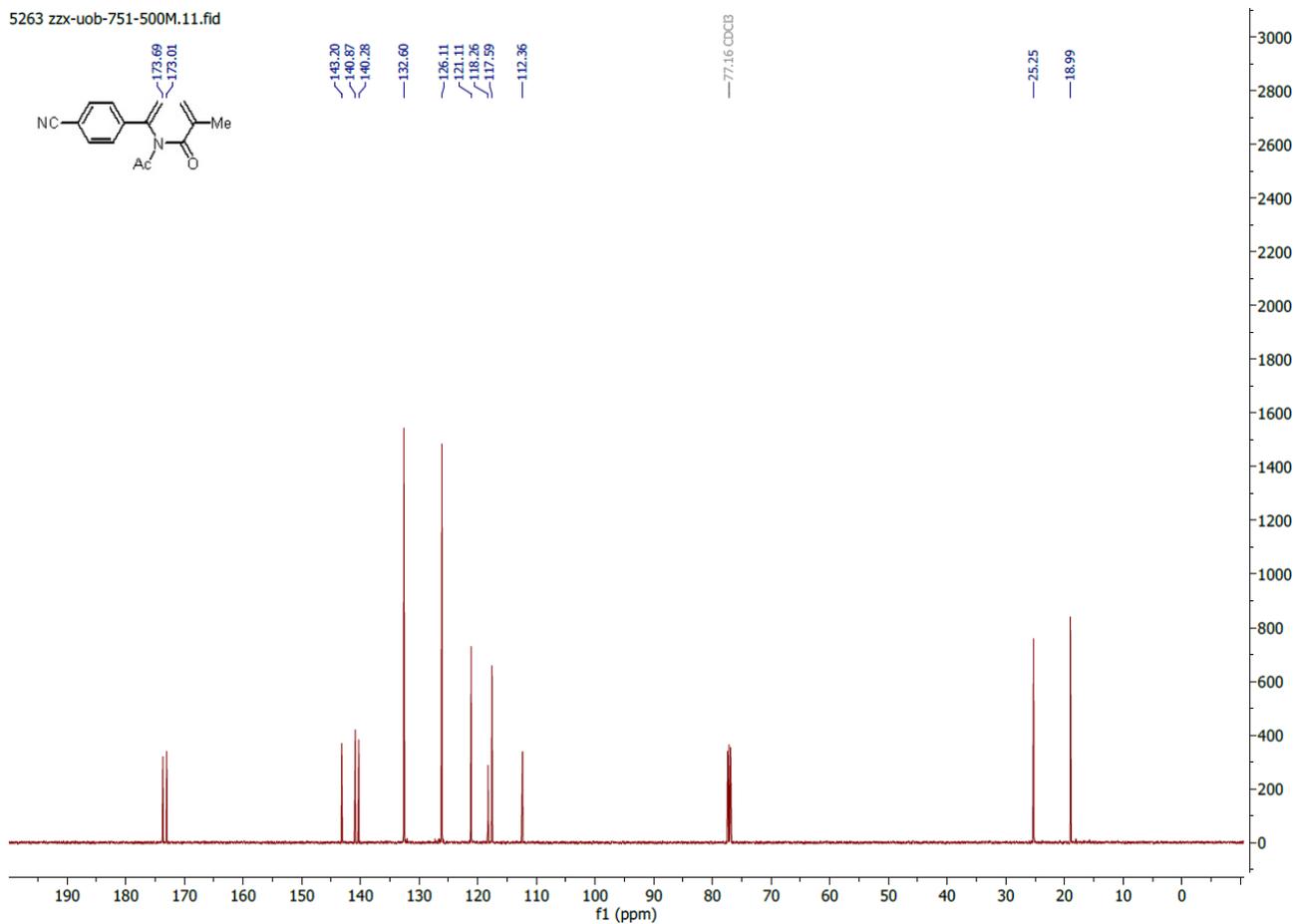


Compound 2p

5263 zzx-uob-751-500M.10.fid

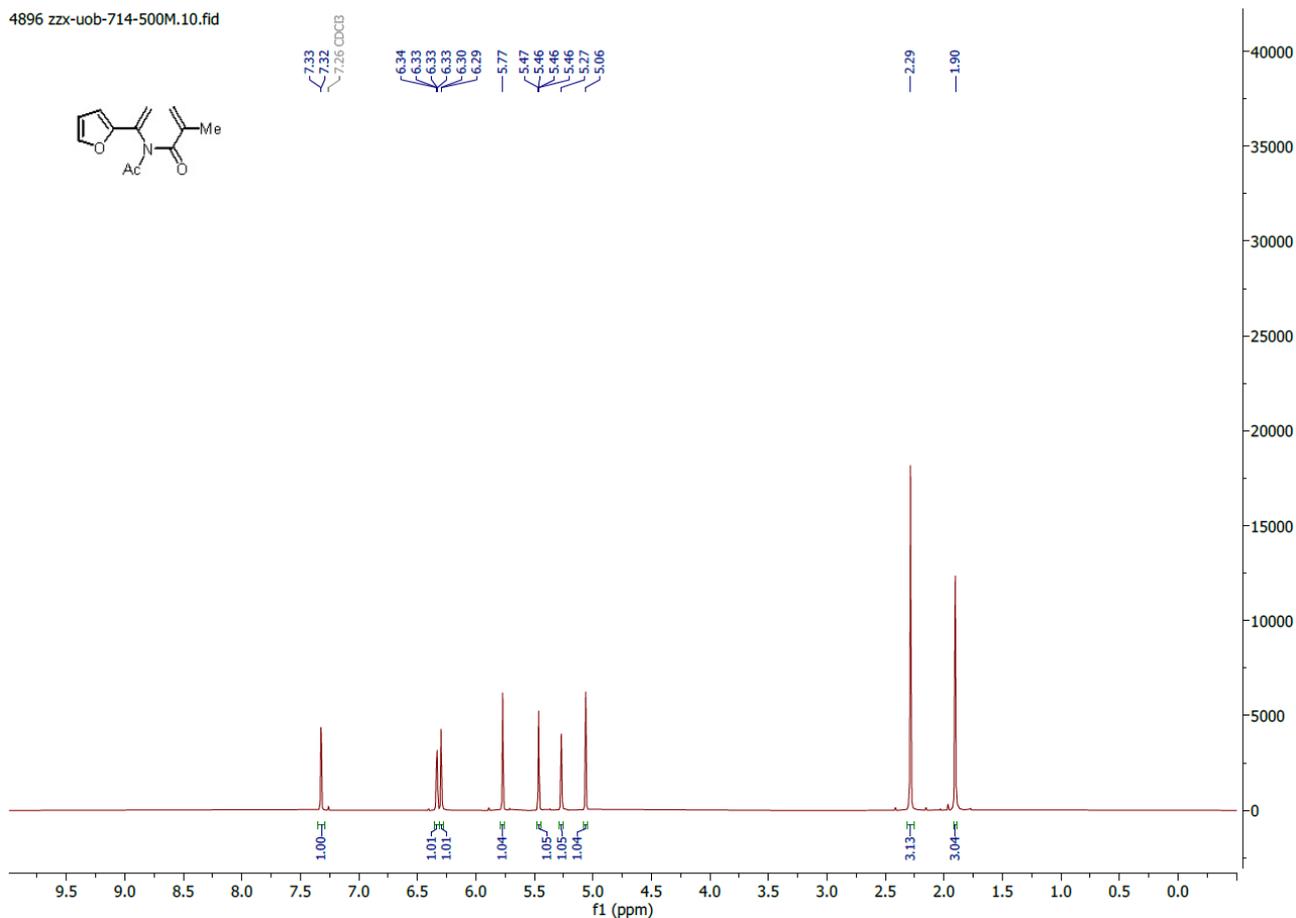
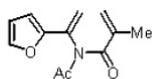


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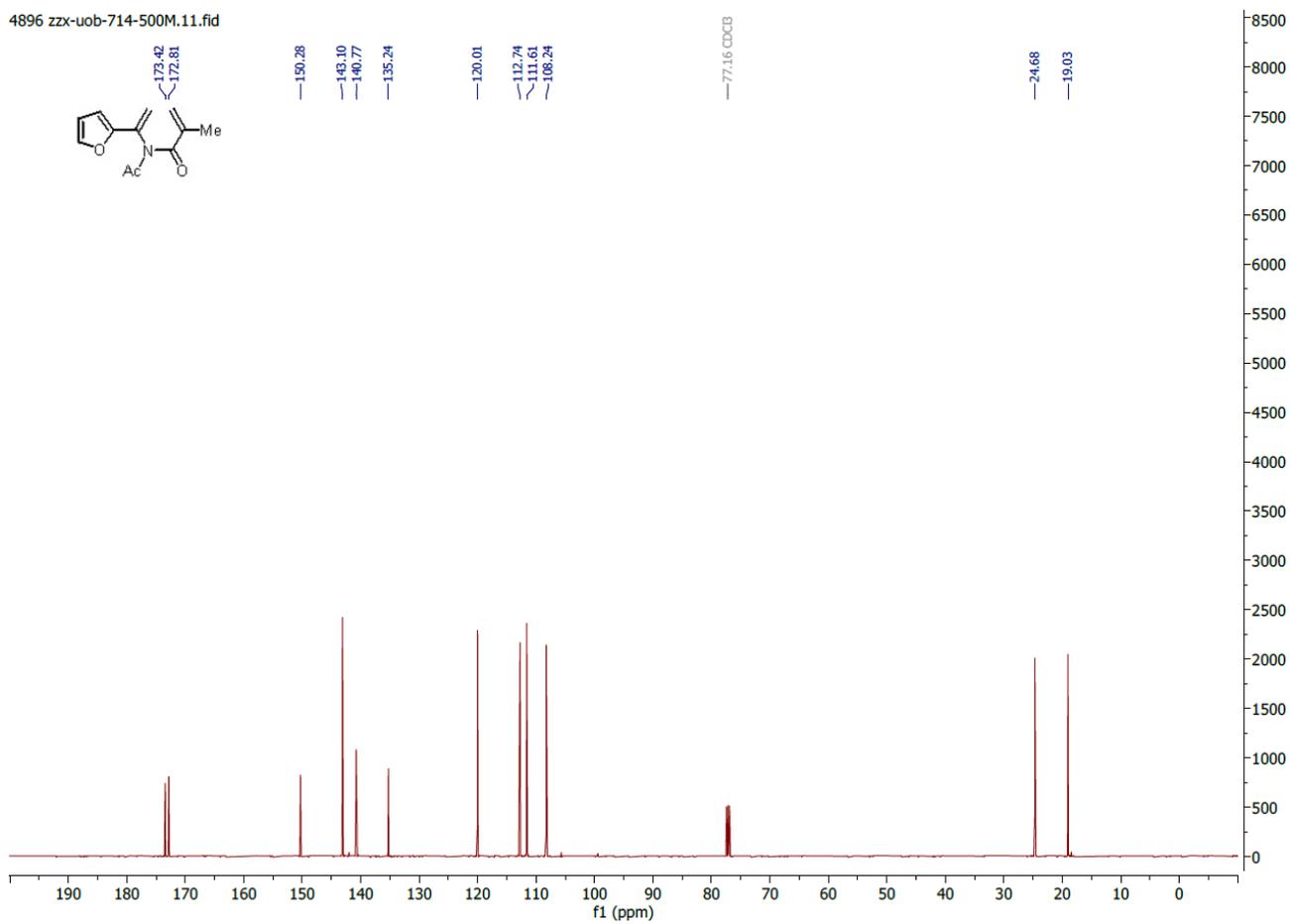
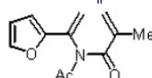


Compound 2q

4896 zzx-uob-714-500M.10.fid

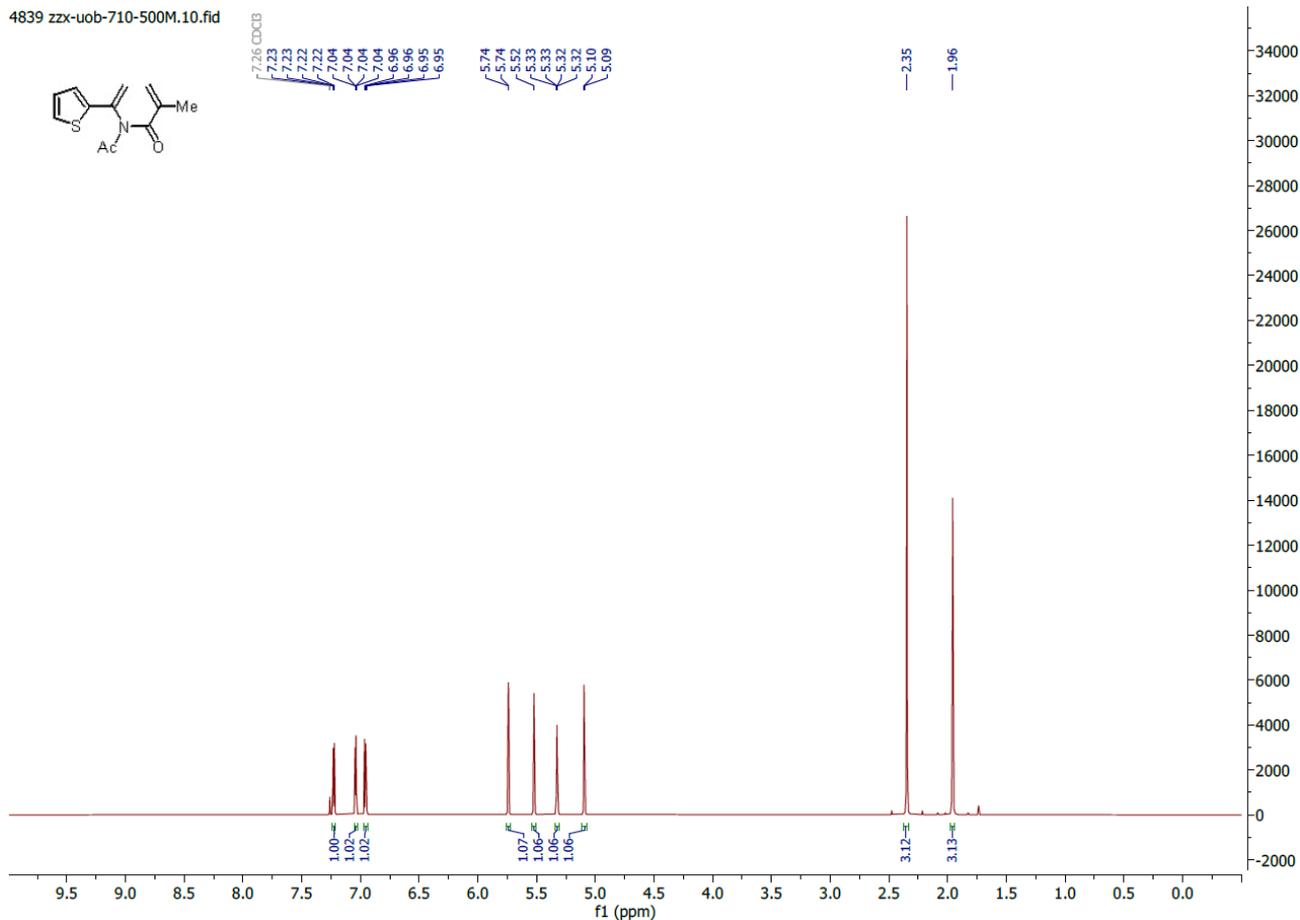
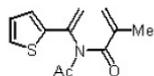


4896 zzx-uob-714-500M.11.fid

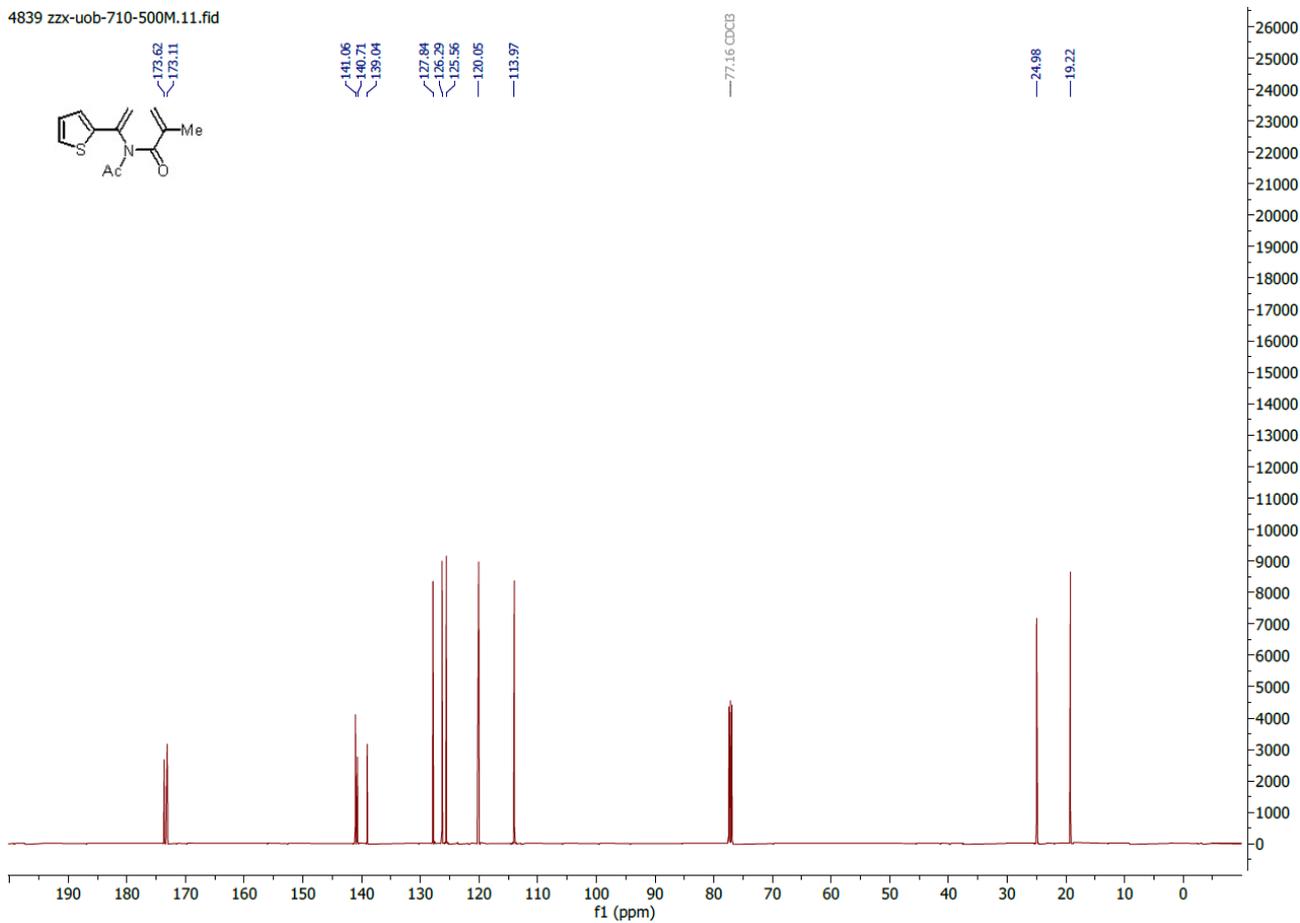
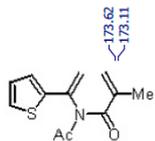


Compound 2r

4839 zzx-uob-710-500M.10.fid

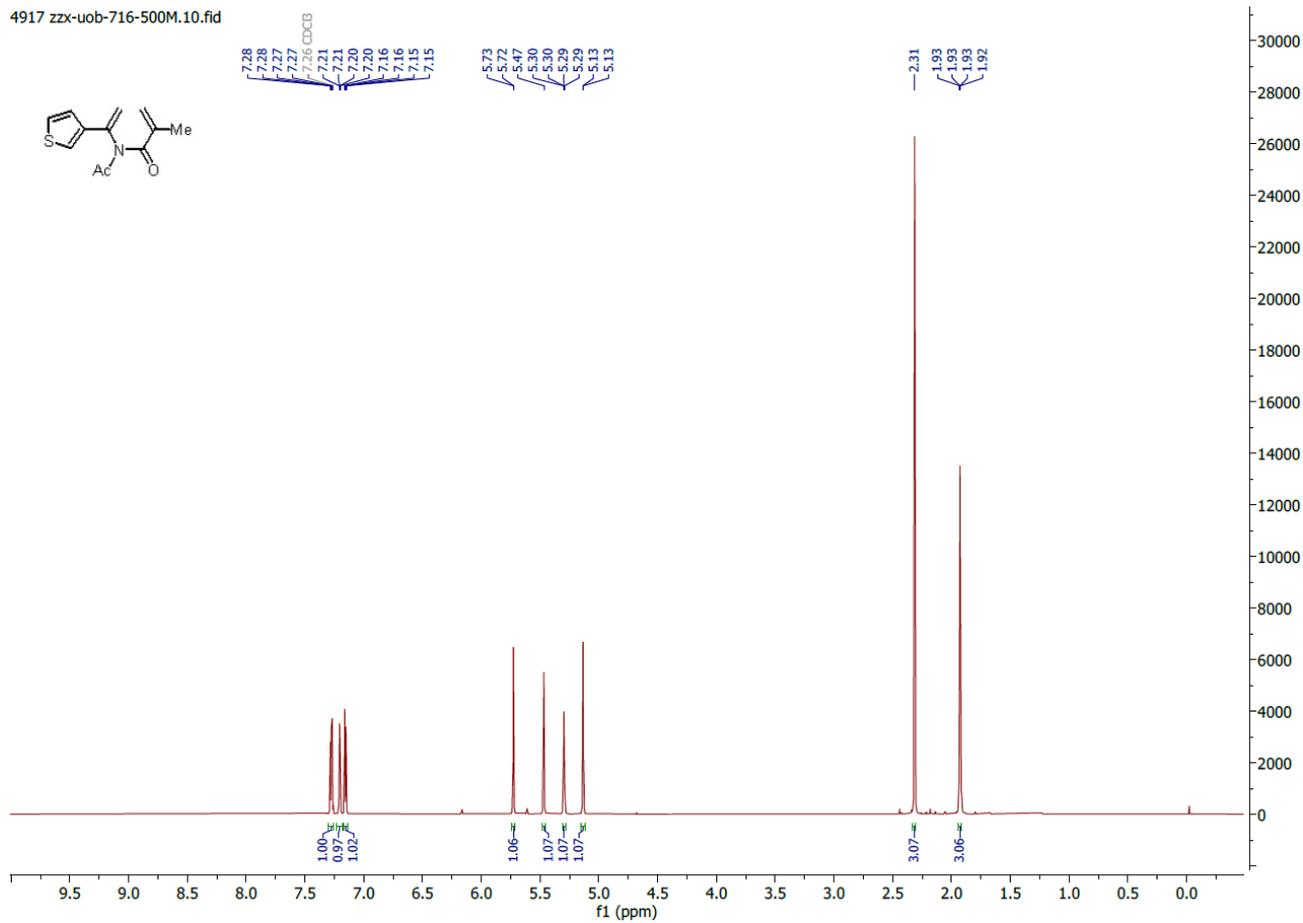
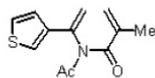


4839 zzx-uob-710-500M.11.fid

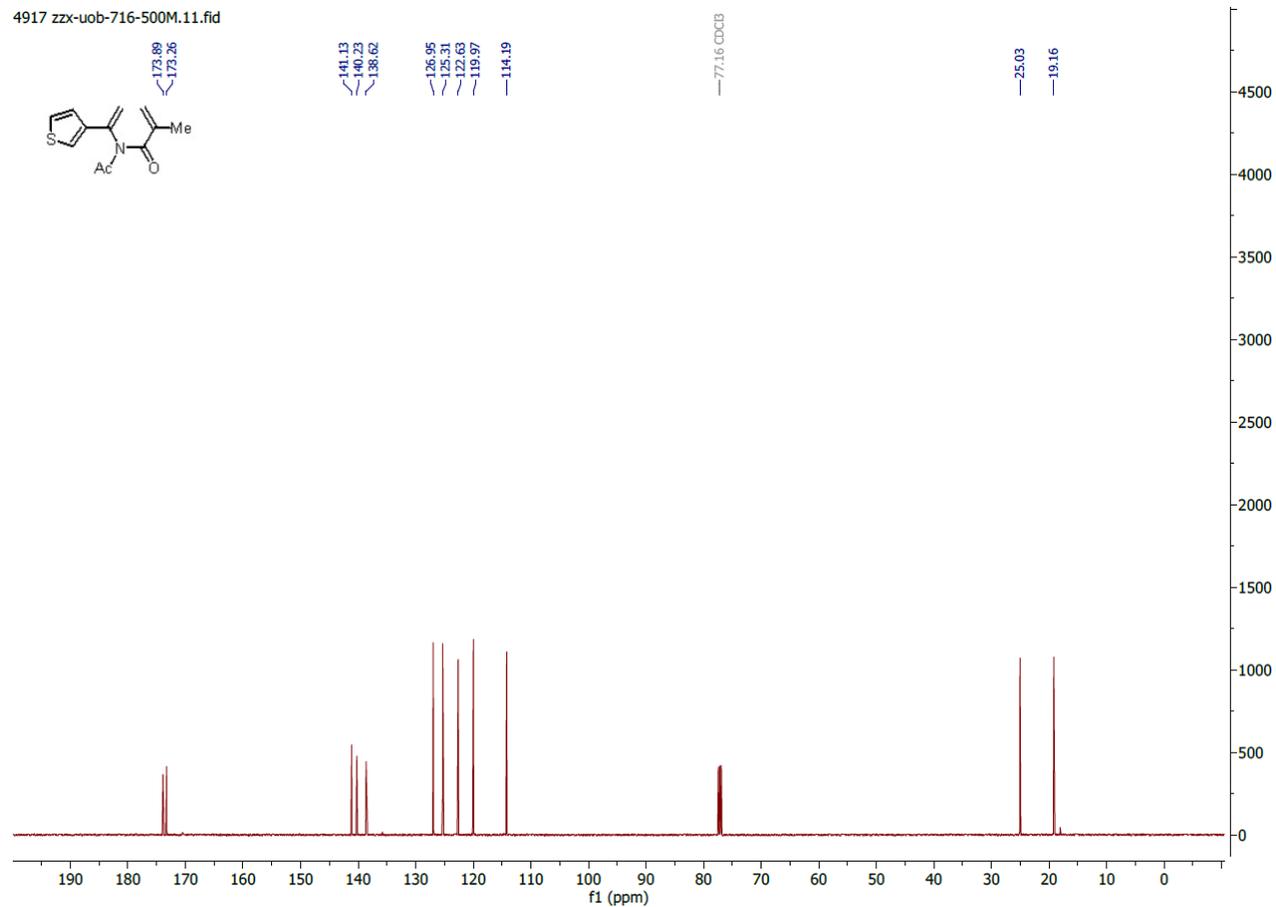
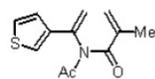


Compound 2s

4917 zzx-uob-716-500M.10.fid

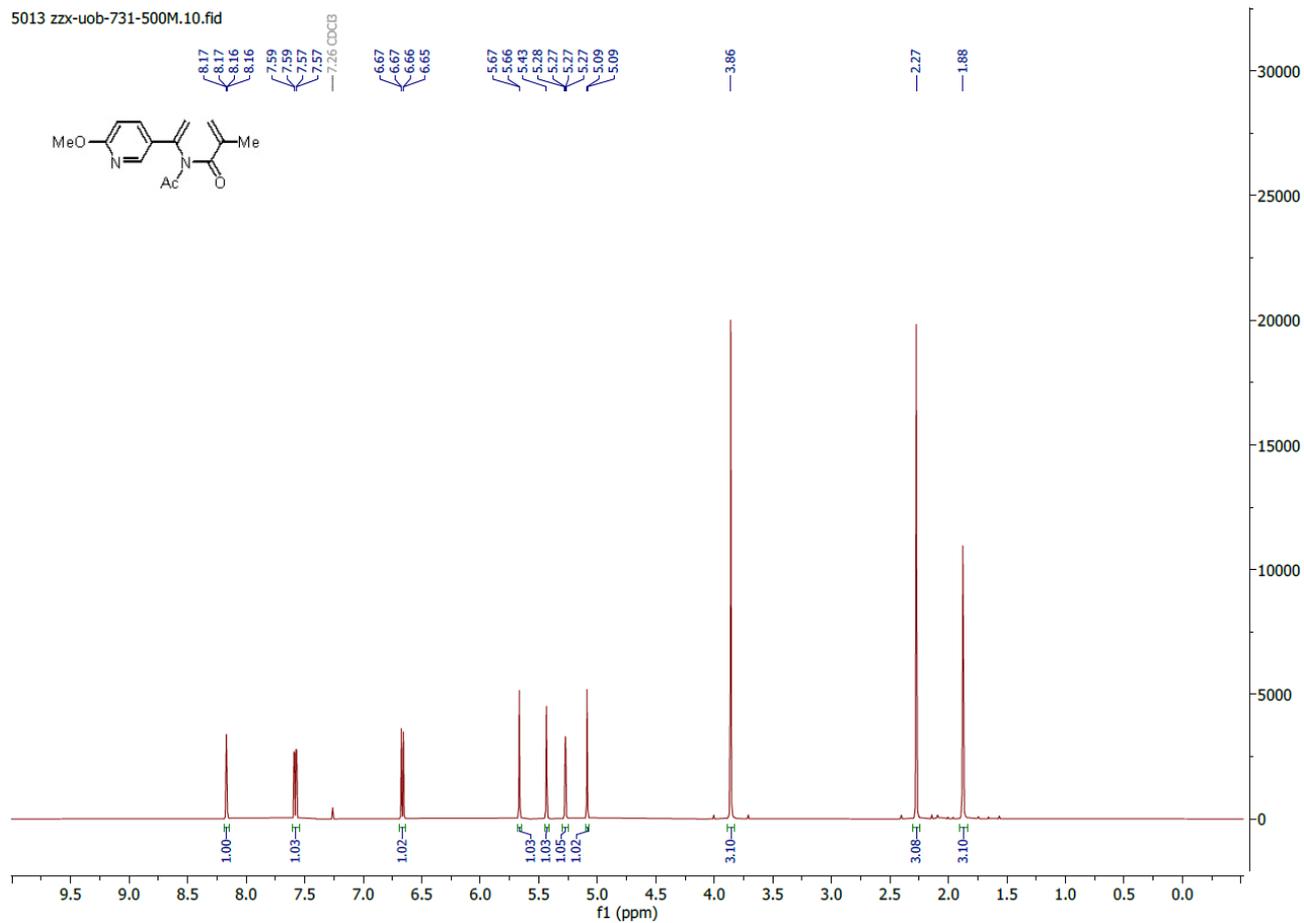


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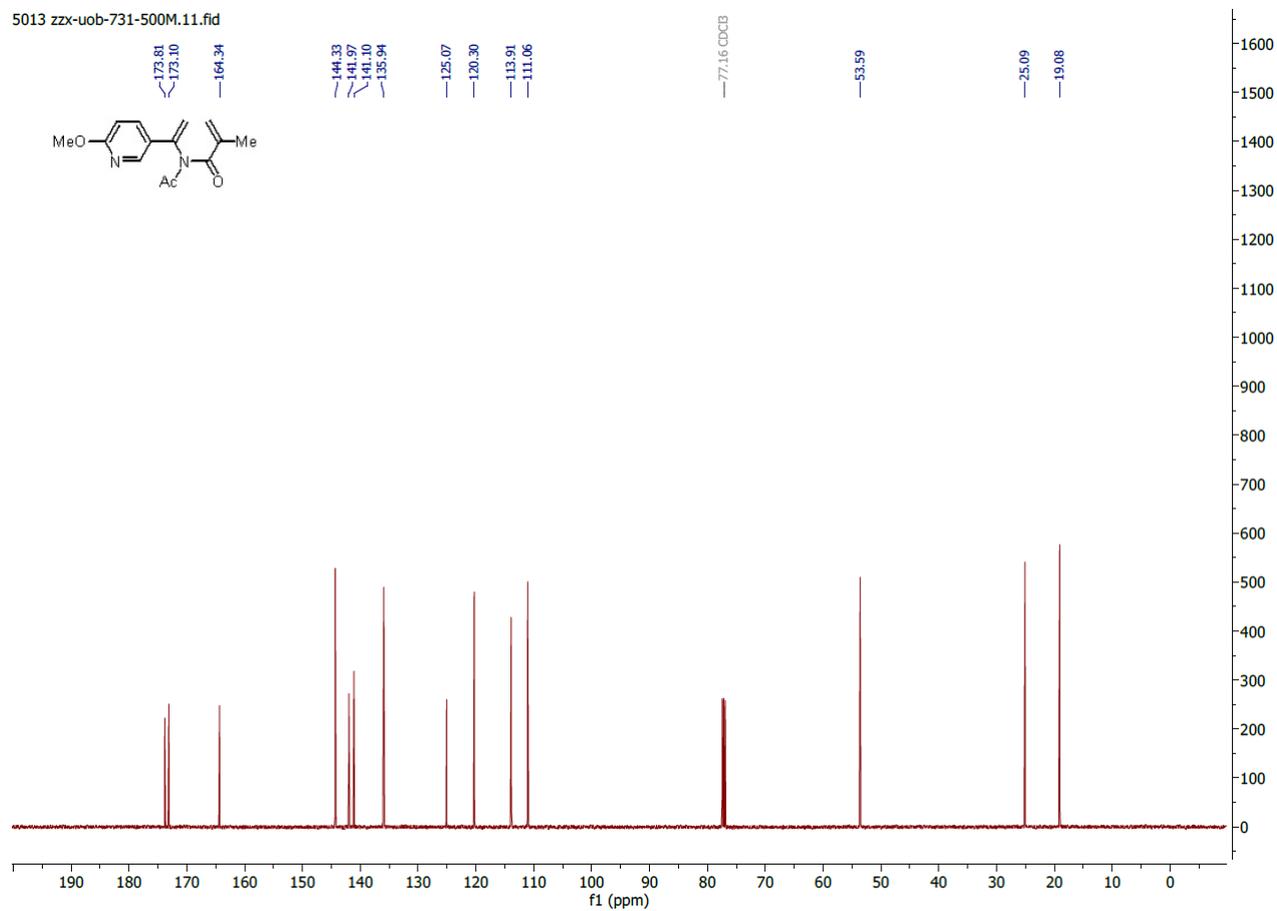


Compound 2t

5013 zzx-uob-731-500M.10.fid

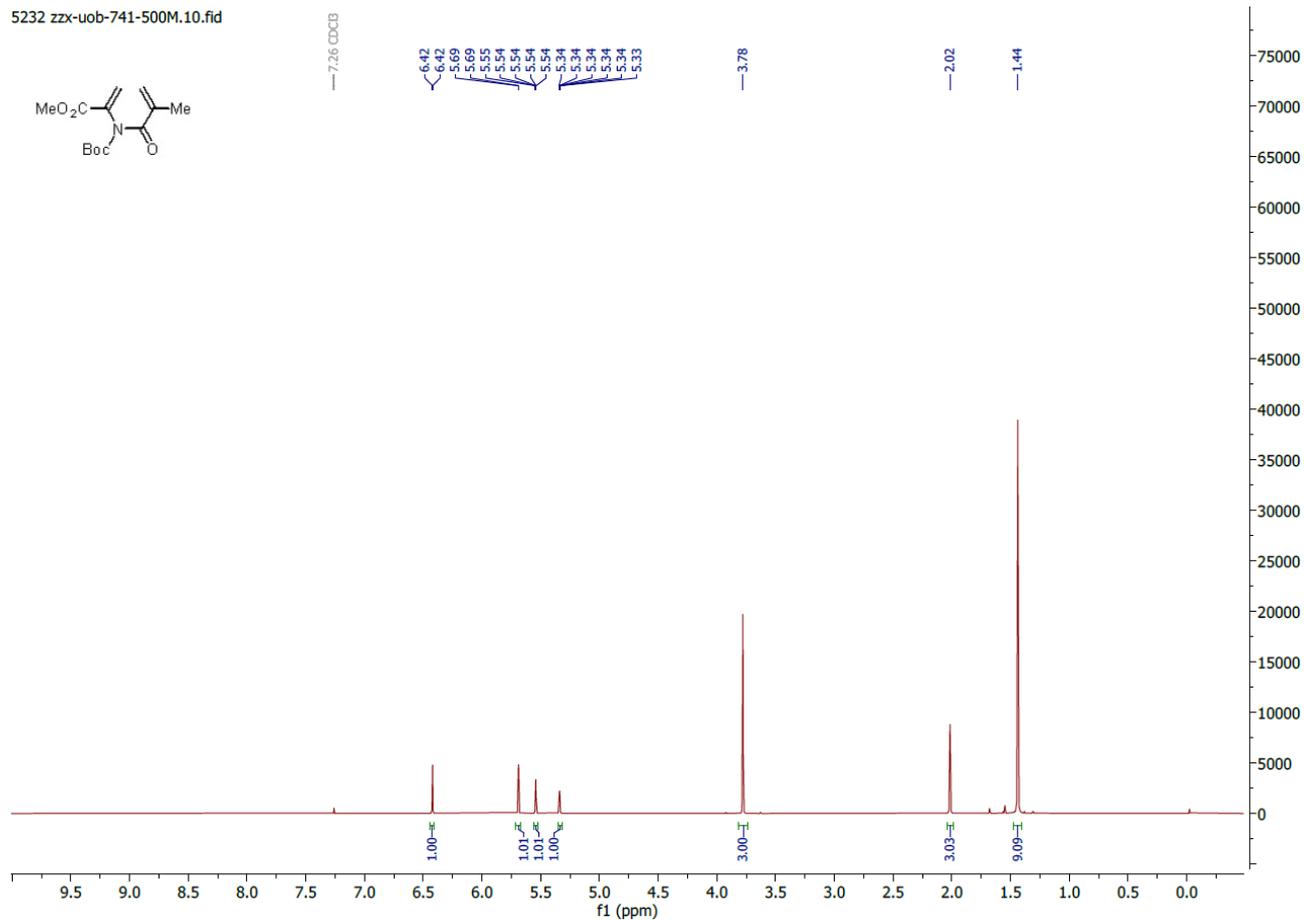
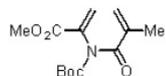


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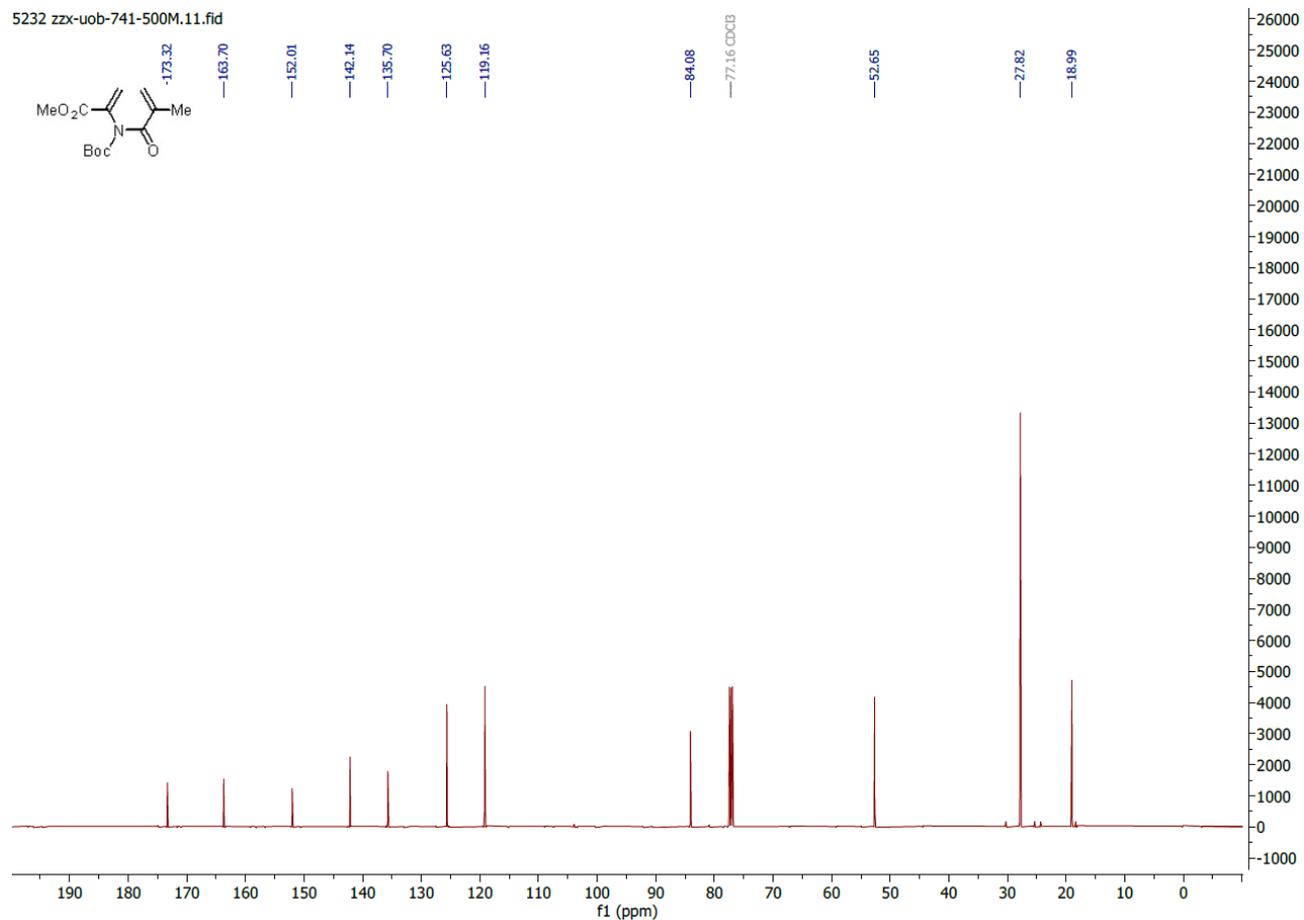
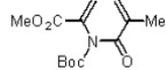


Compound 2u

5232 zzx-uob-741-500M.10.fid

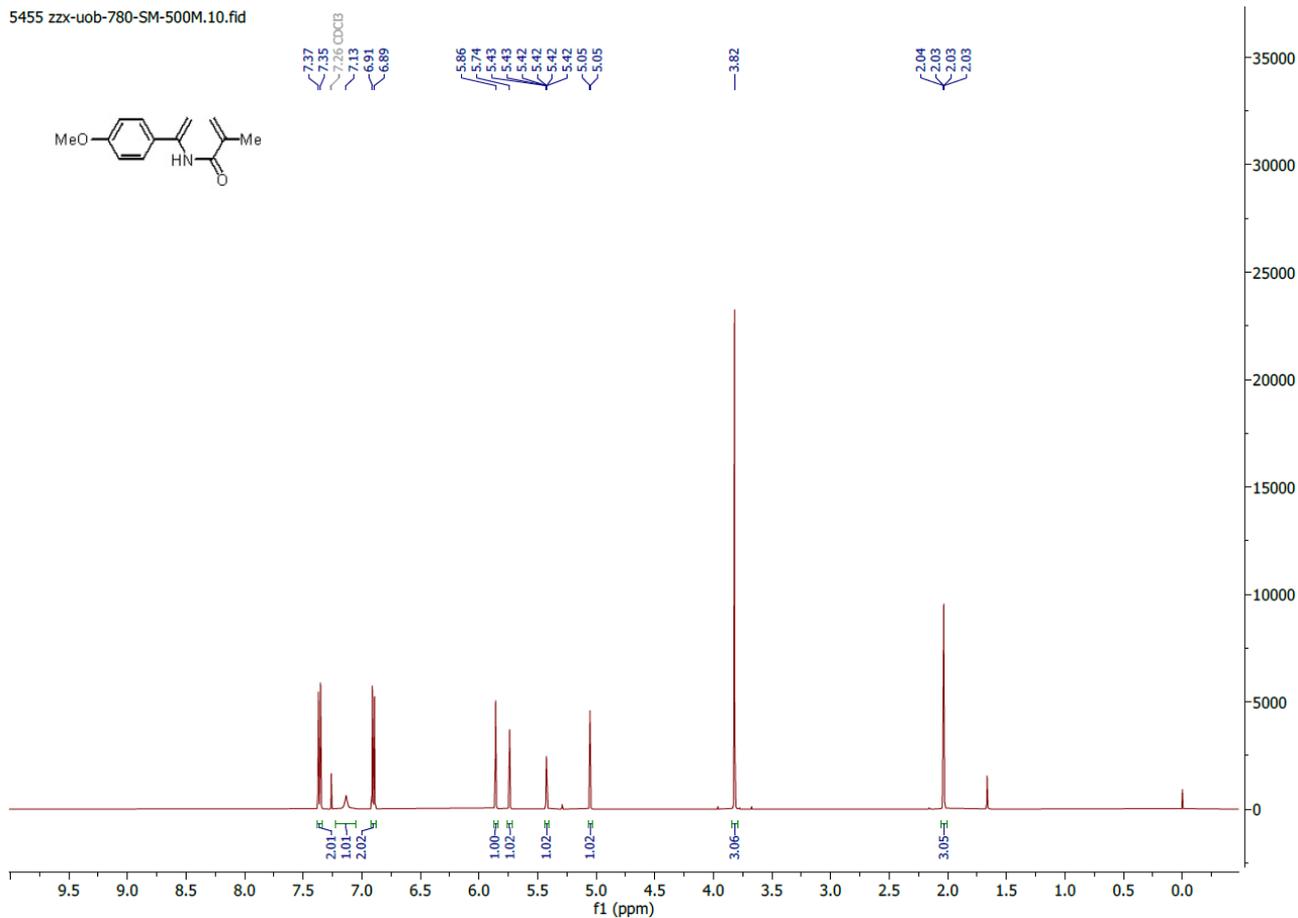
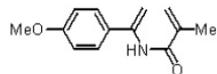


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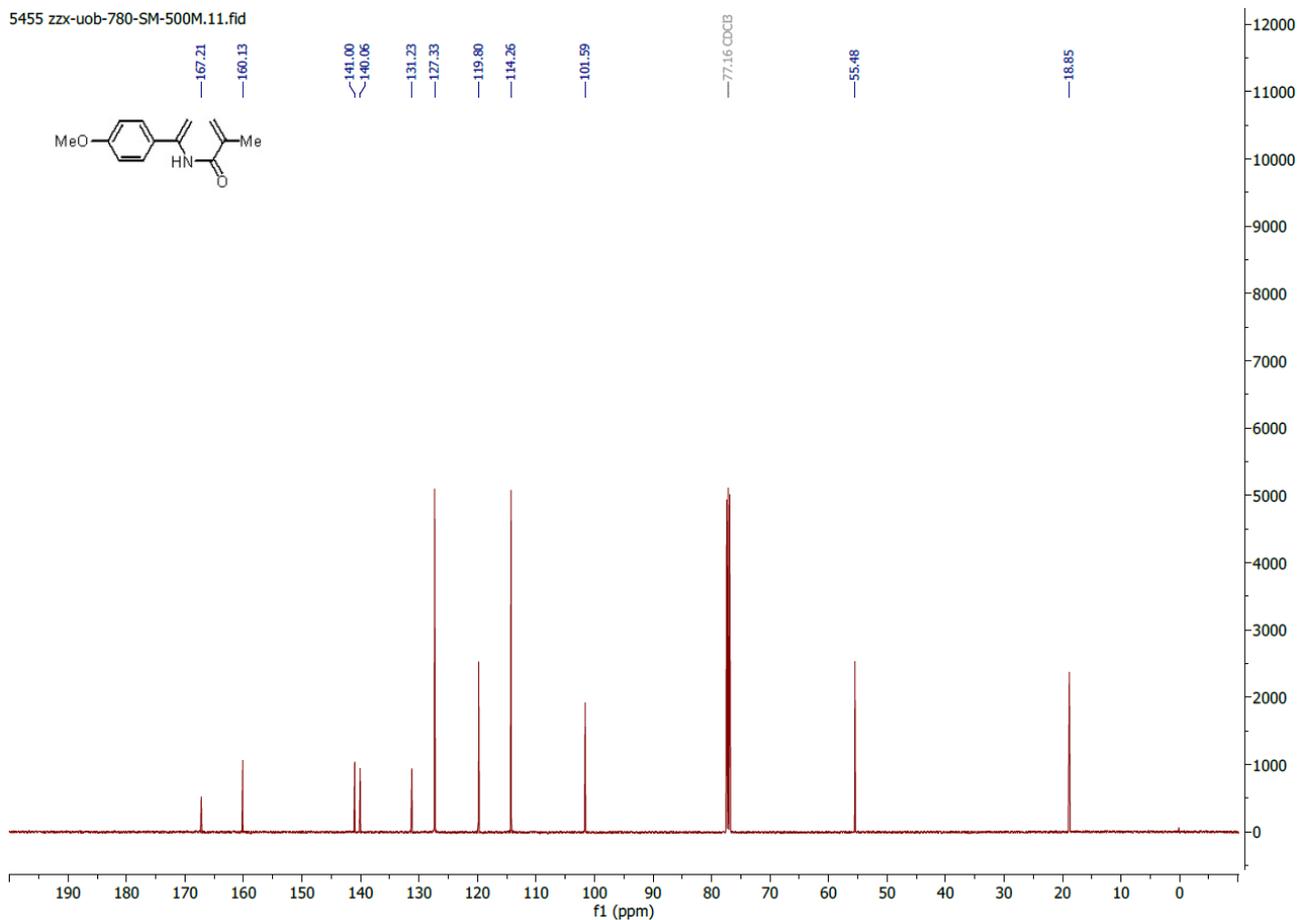
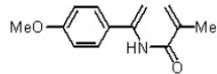


Compound S2v

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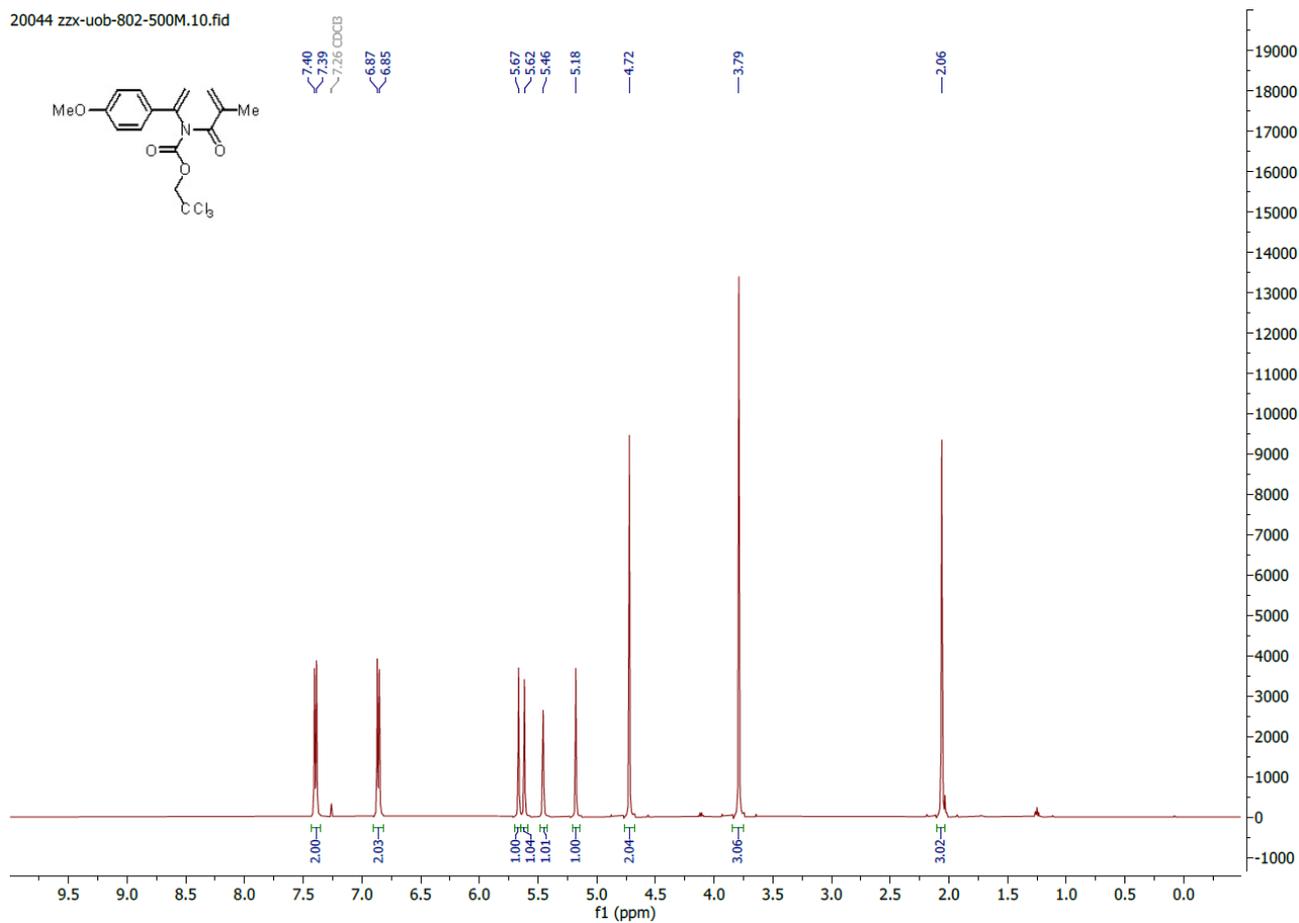
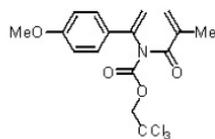


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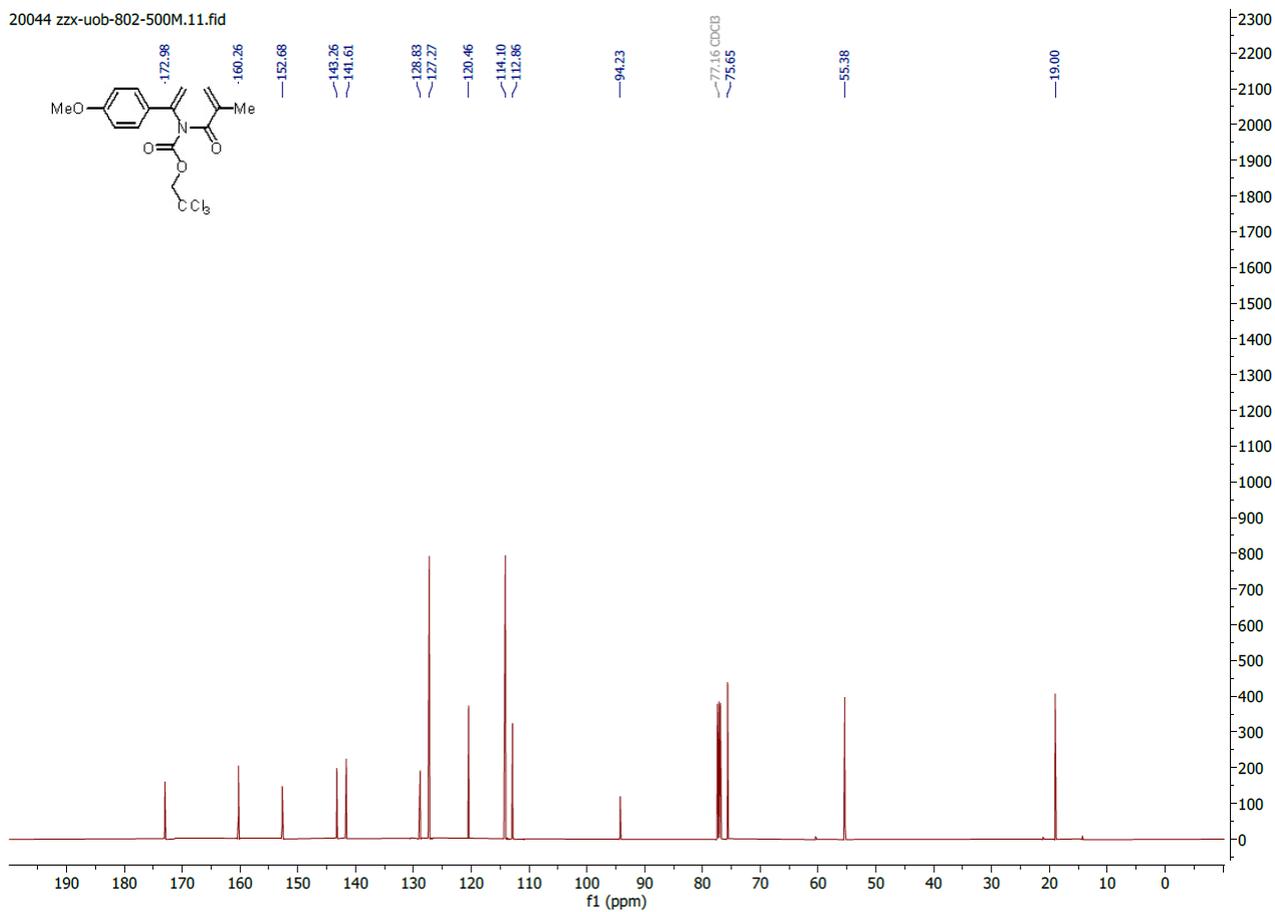
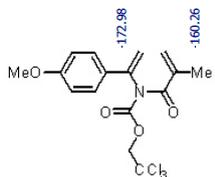


Compound 2v

20044 zzx-uob-802-500M.10.fid

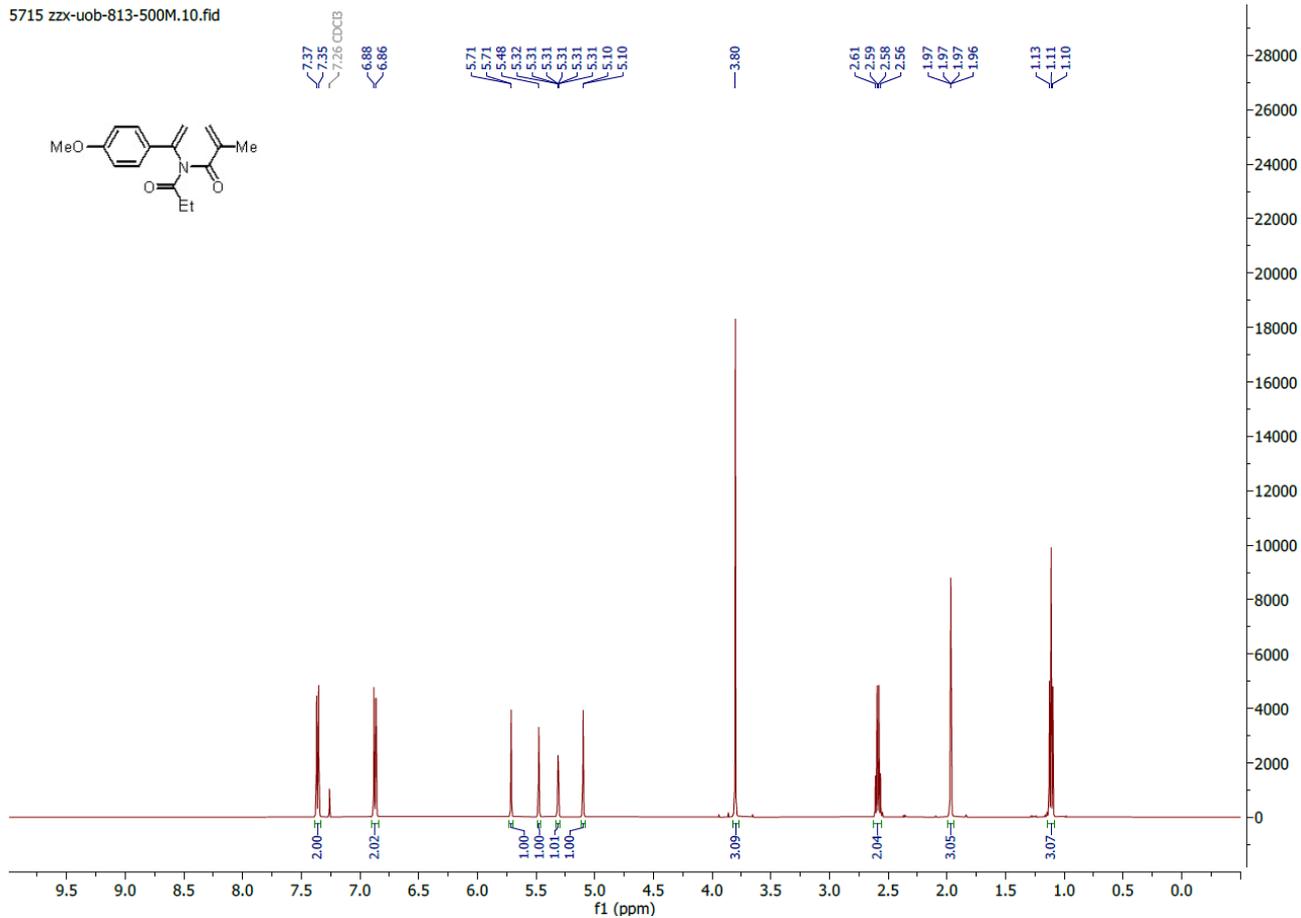
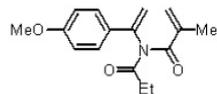


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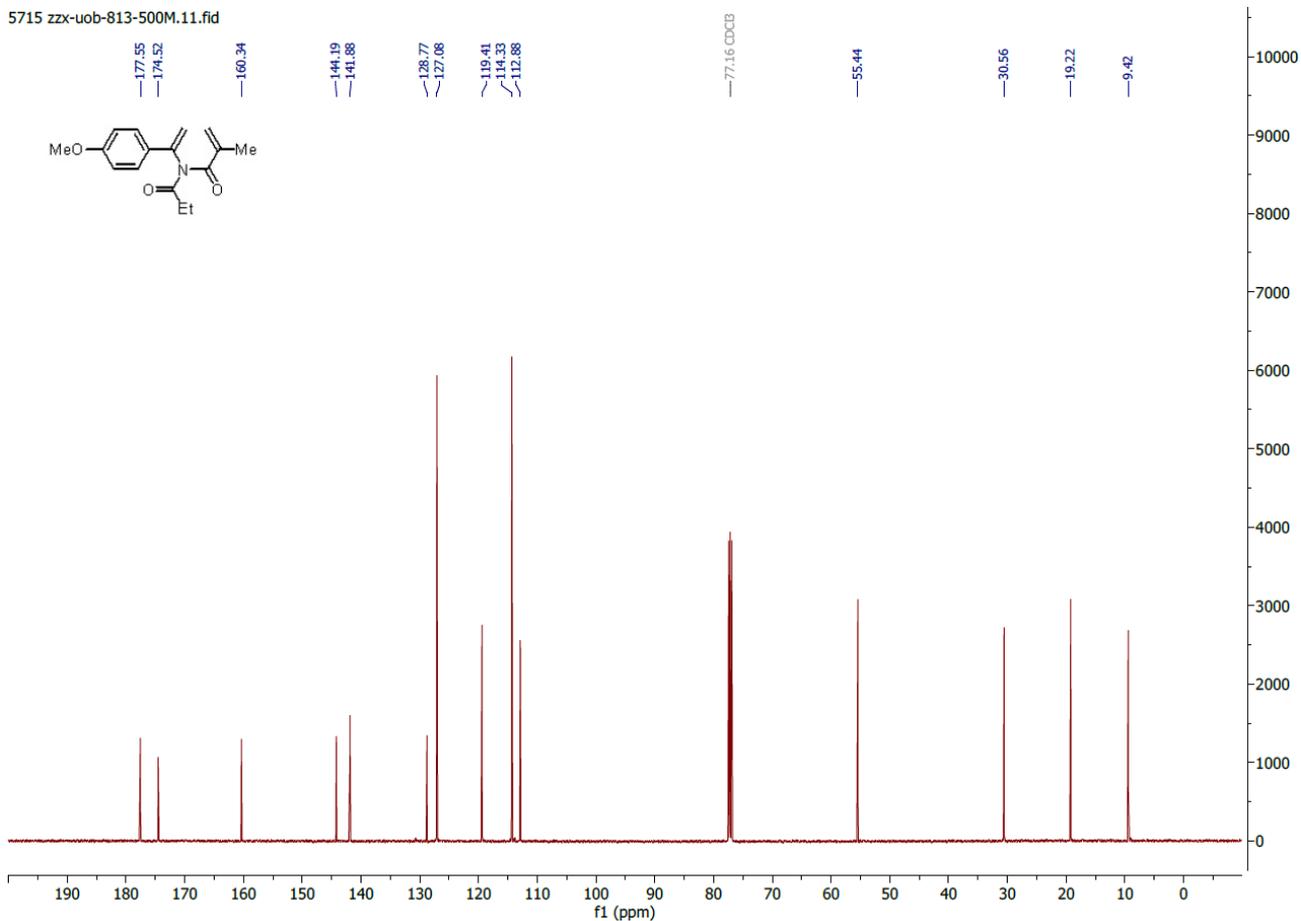
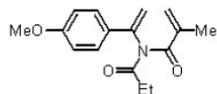


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5715 zzx-uob-813-500M.10.fid

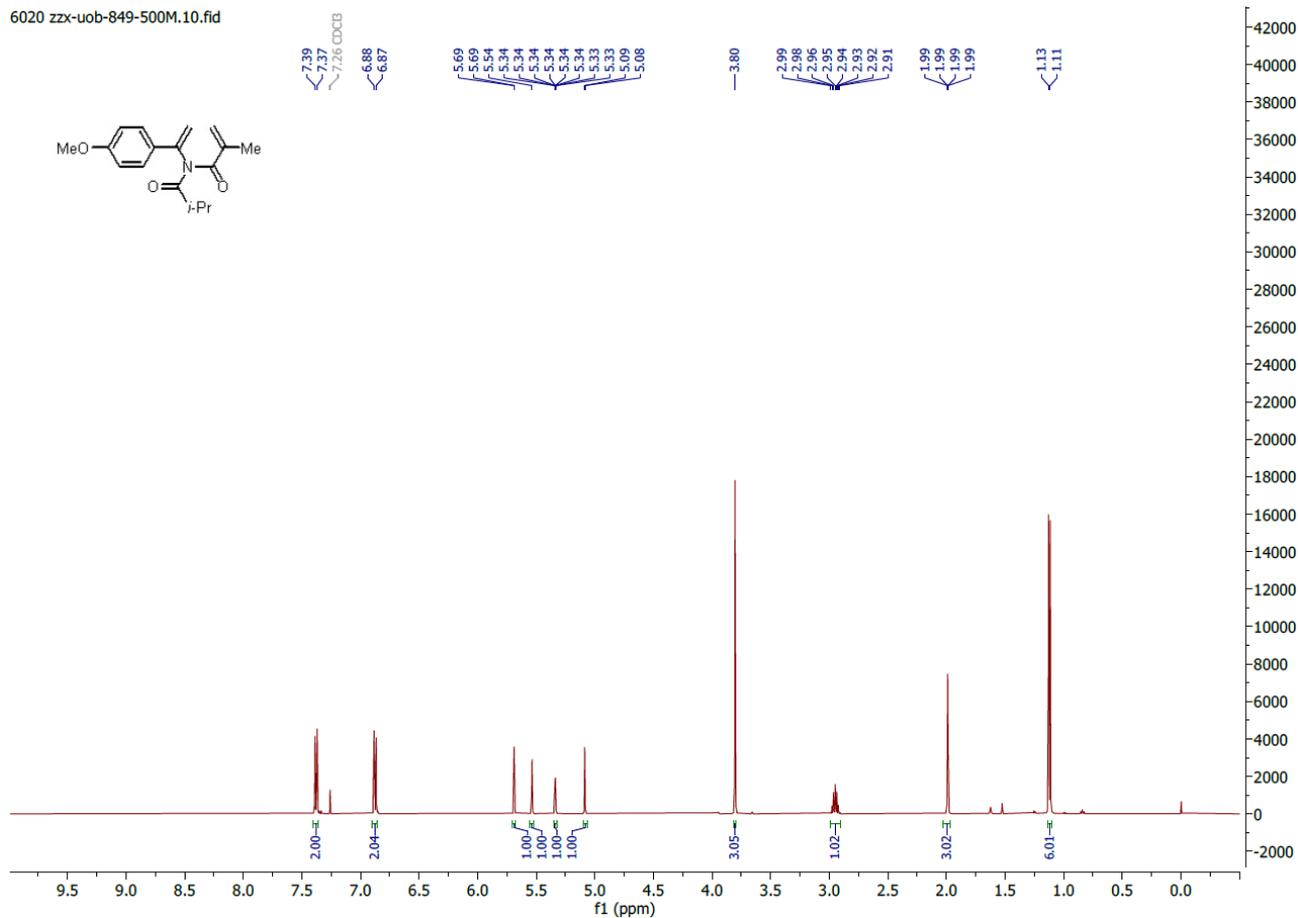
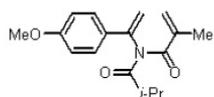


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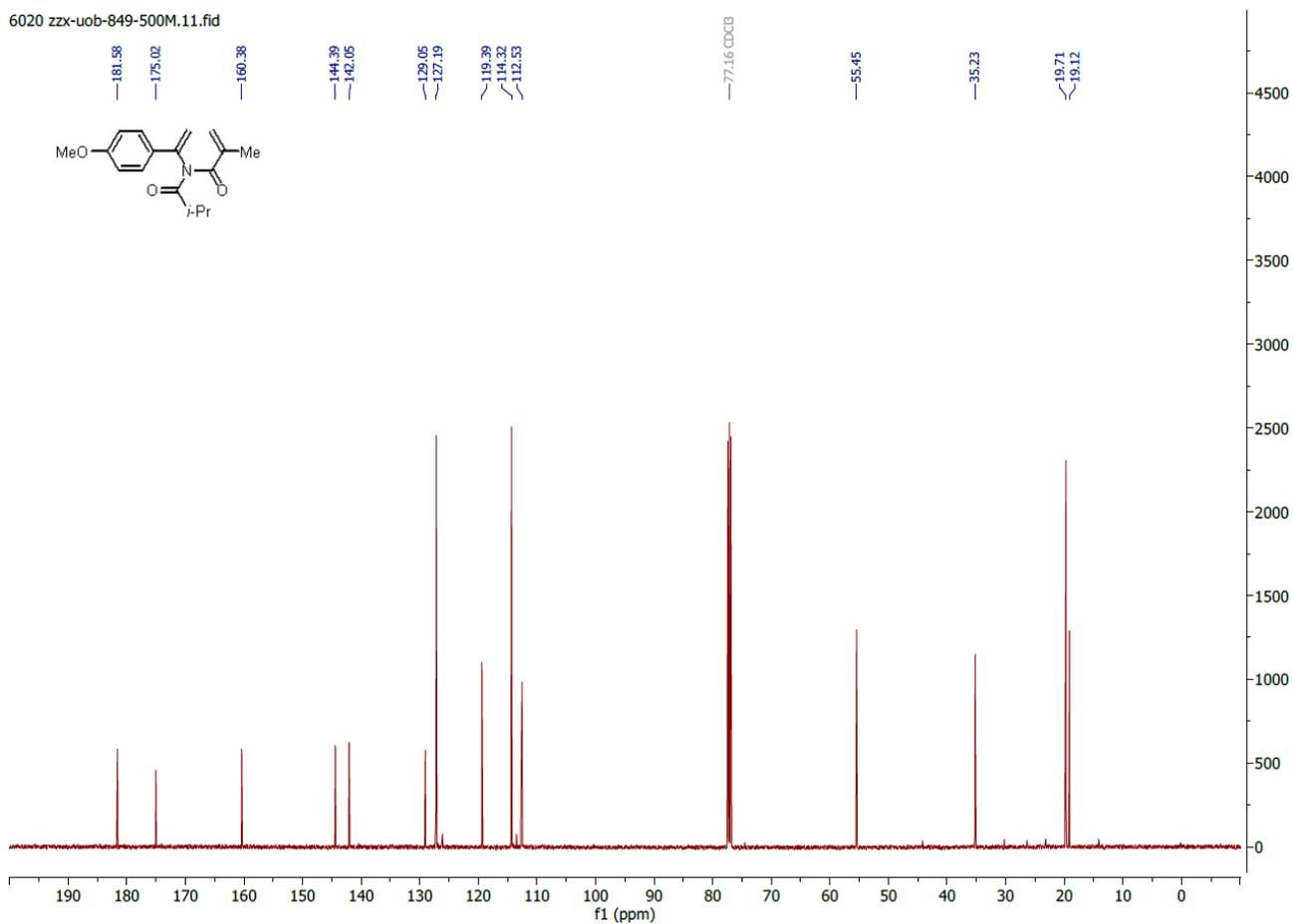
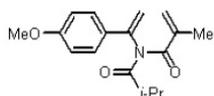


Compound 2x

6020 zzx-uob-849-500M.10.fid

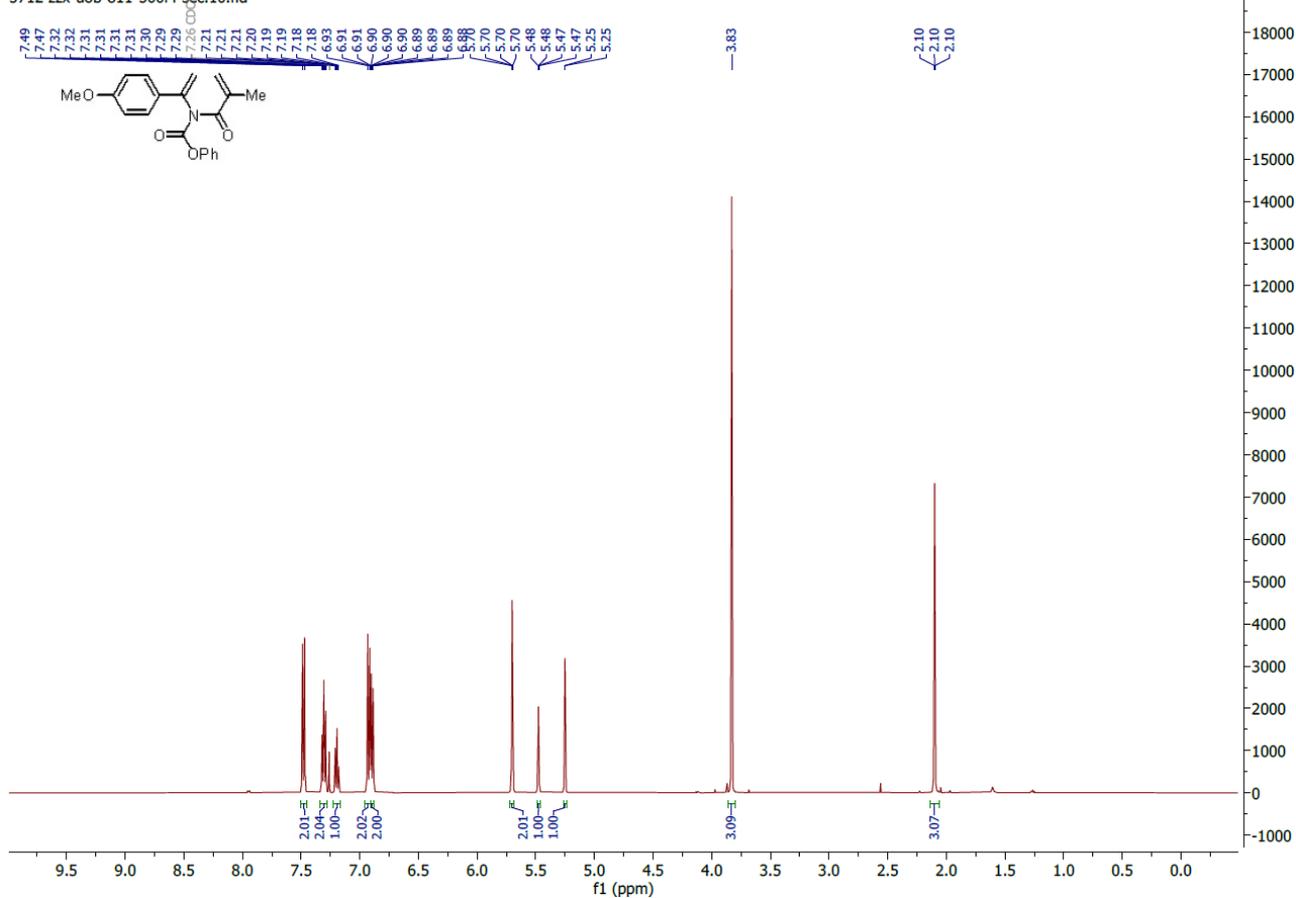


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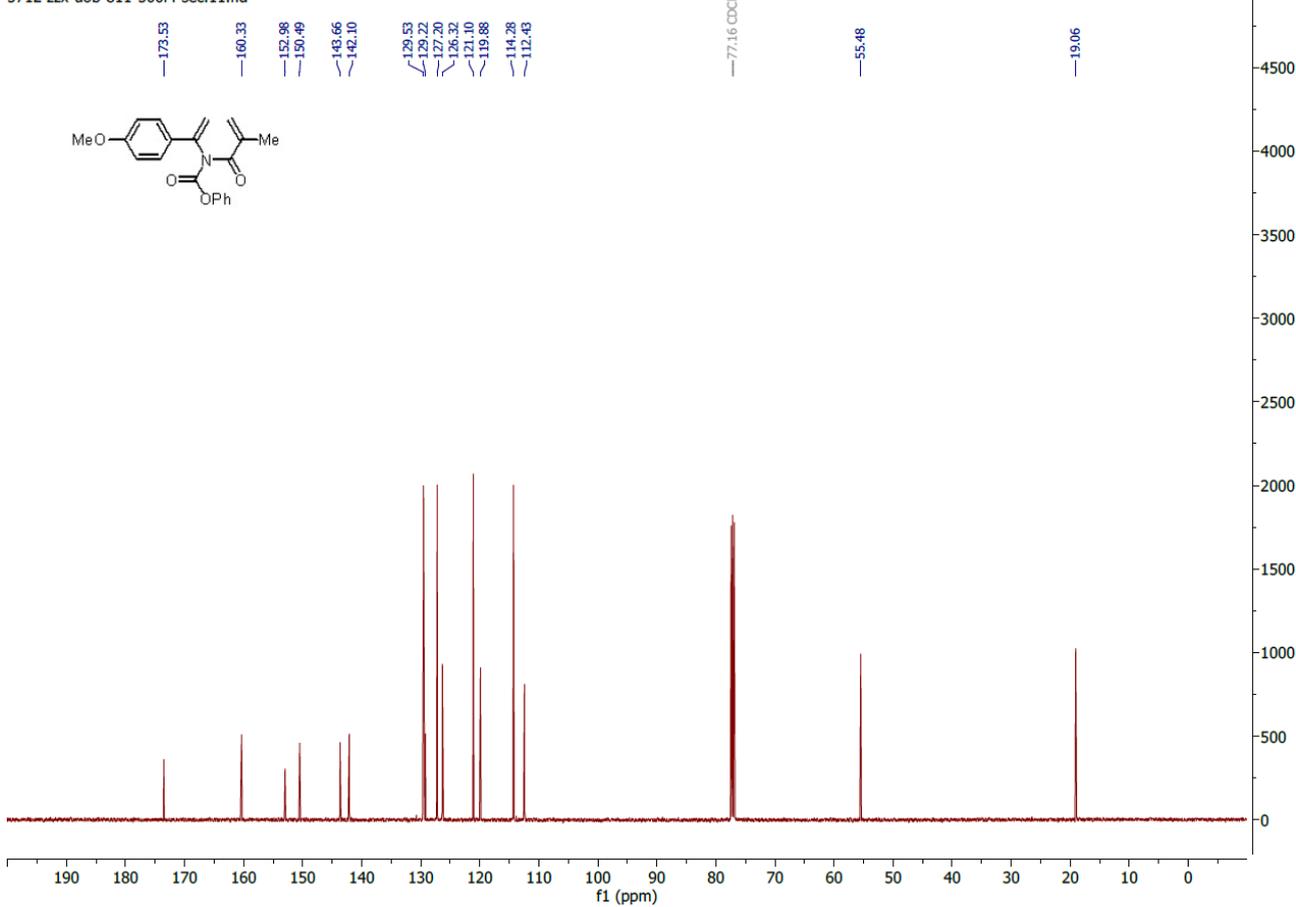


Compound 2y

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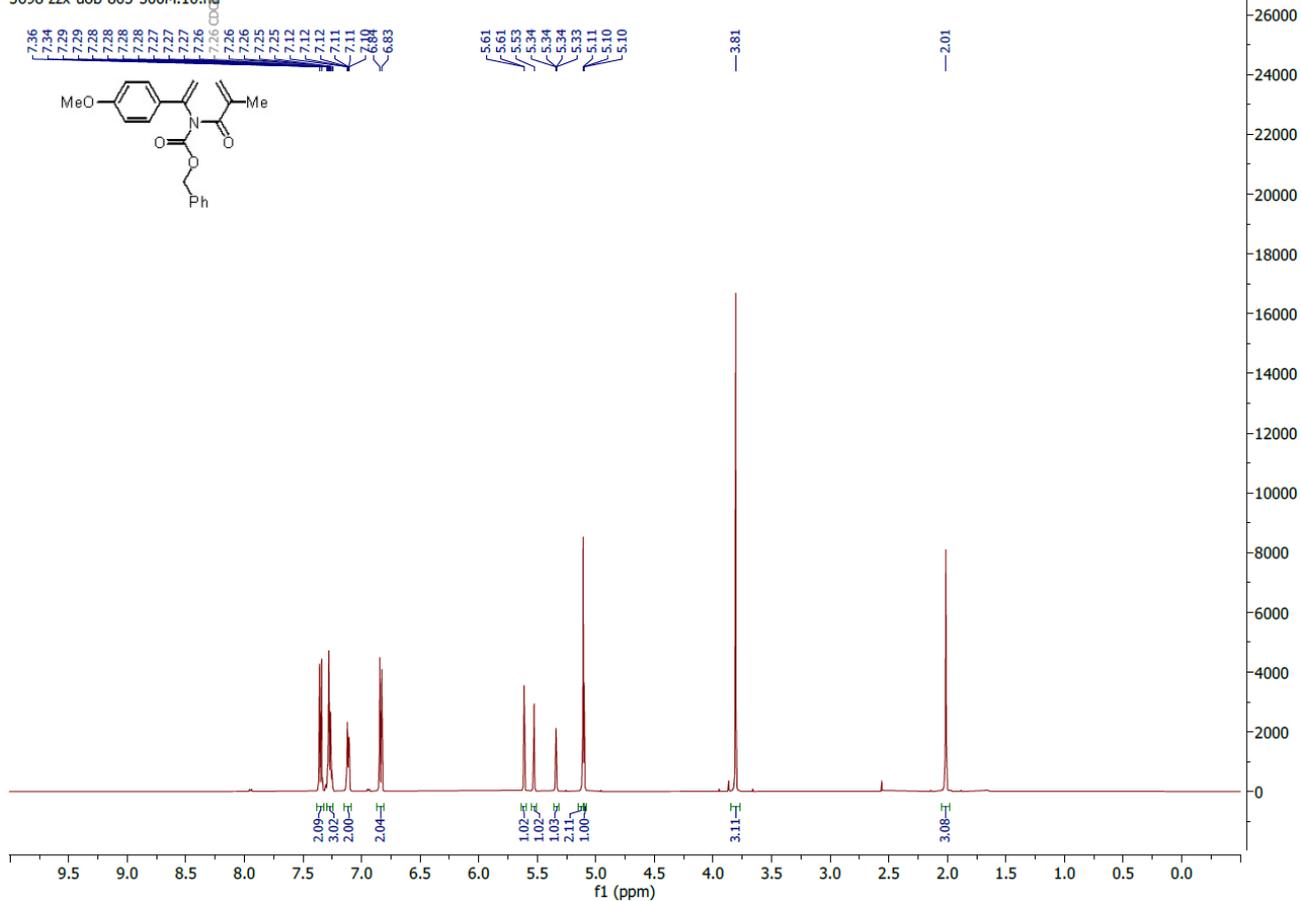


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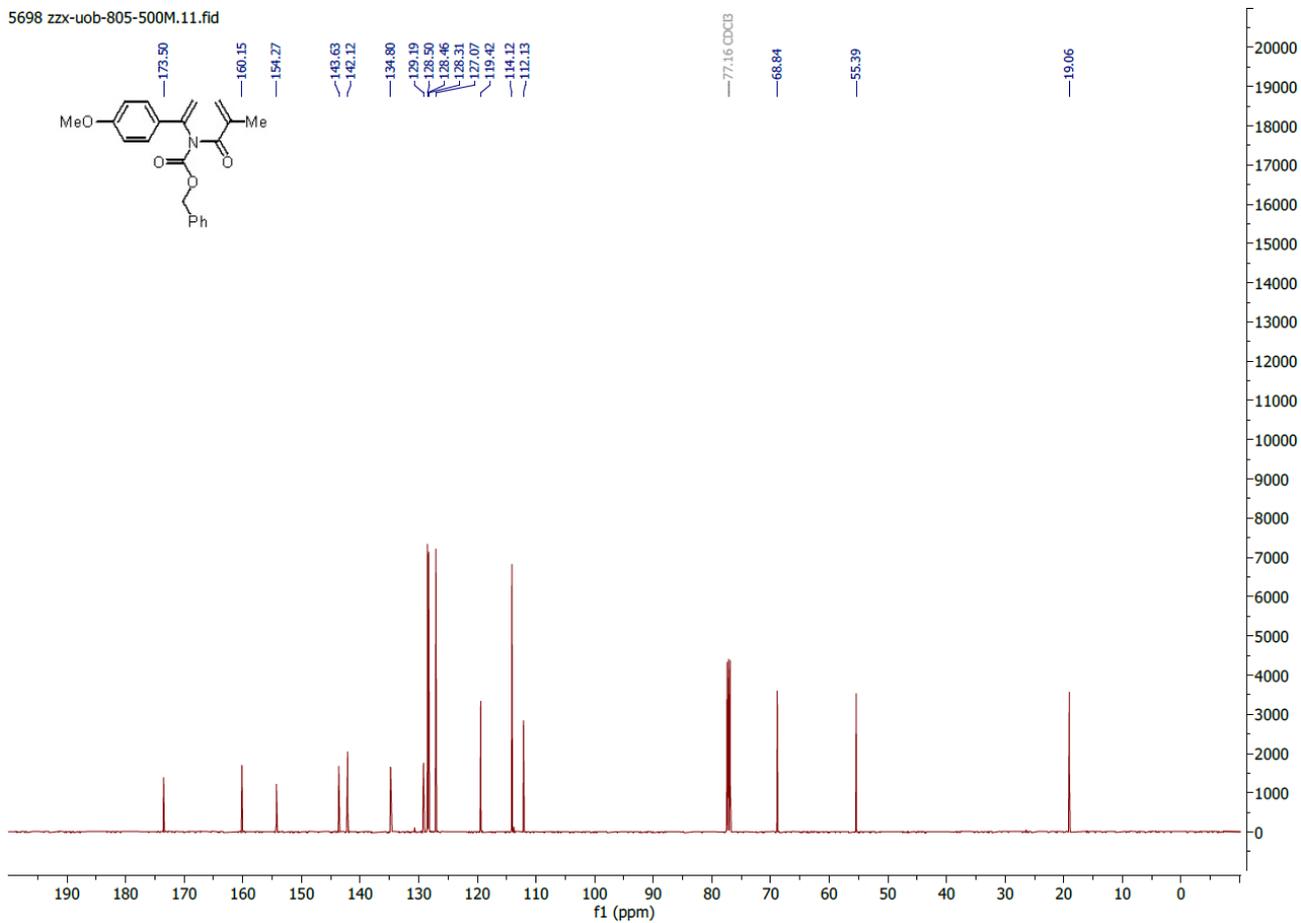


Compound 2z

5698 zzx-uob-805-500M.10.fid

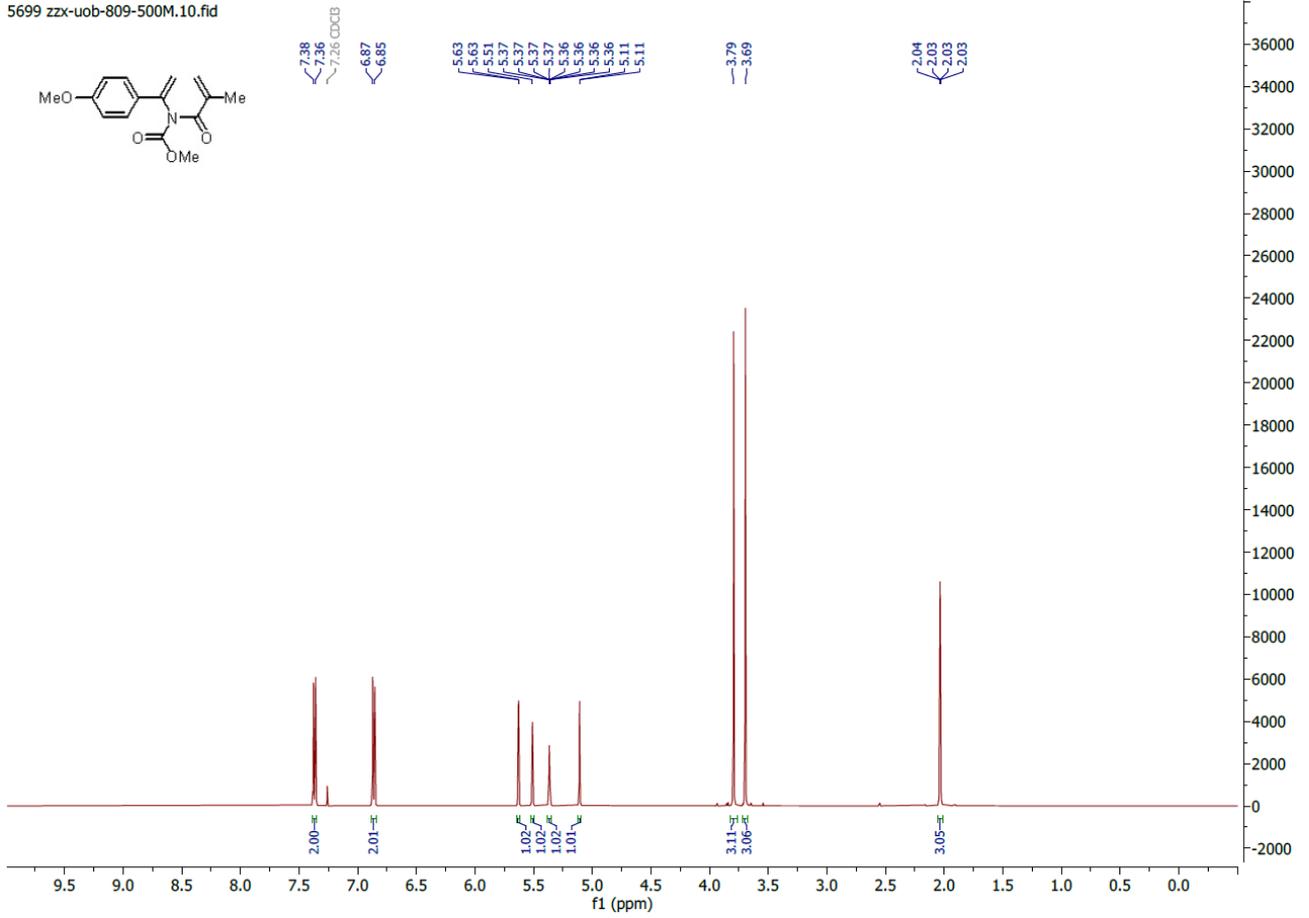


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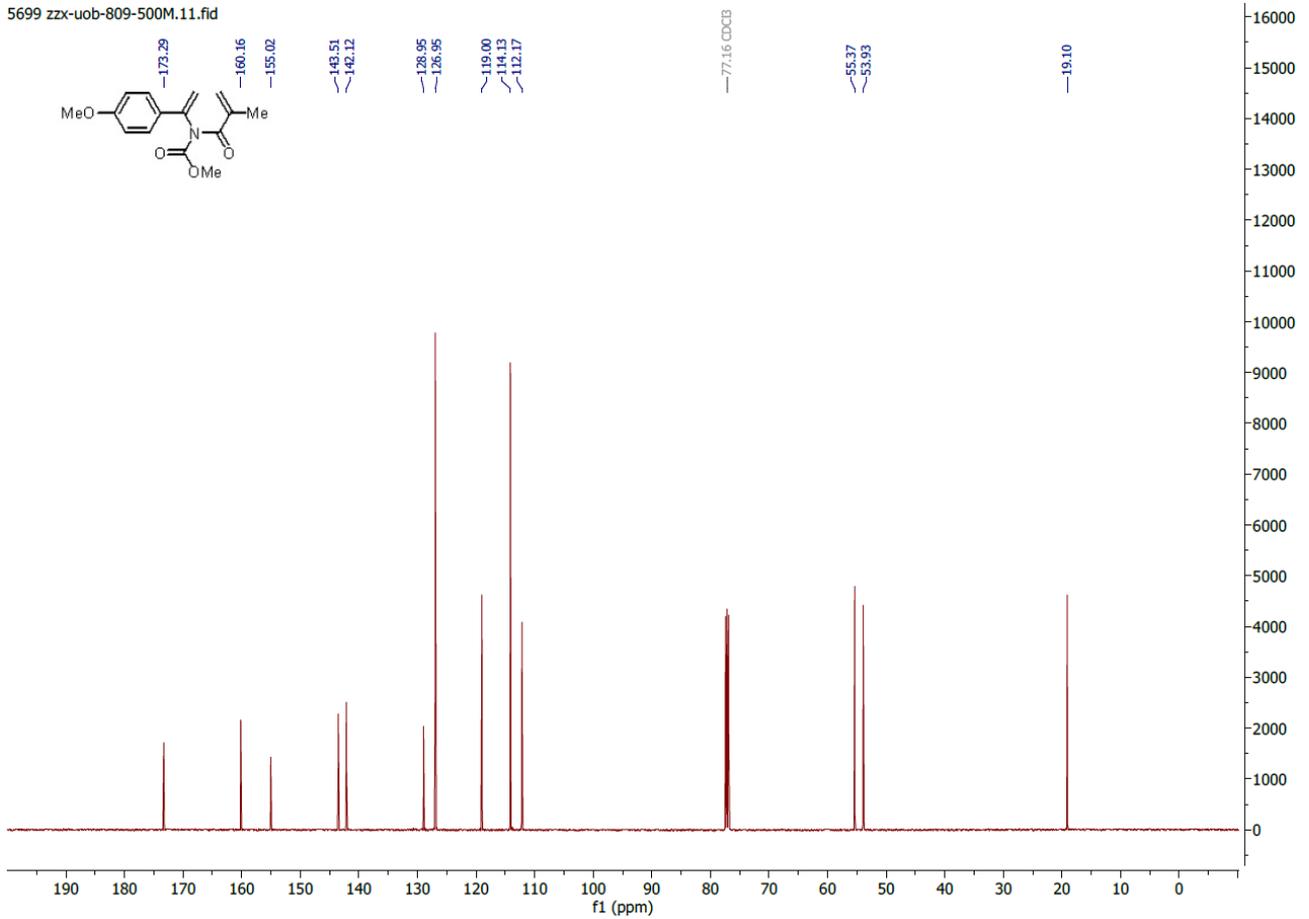


Compound 2aa

5699 zzx-uob-809-500M.10.fid

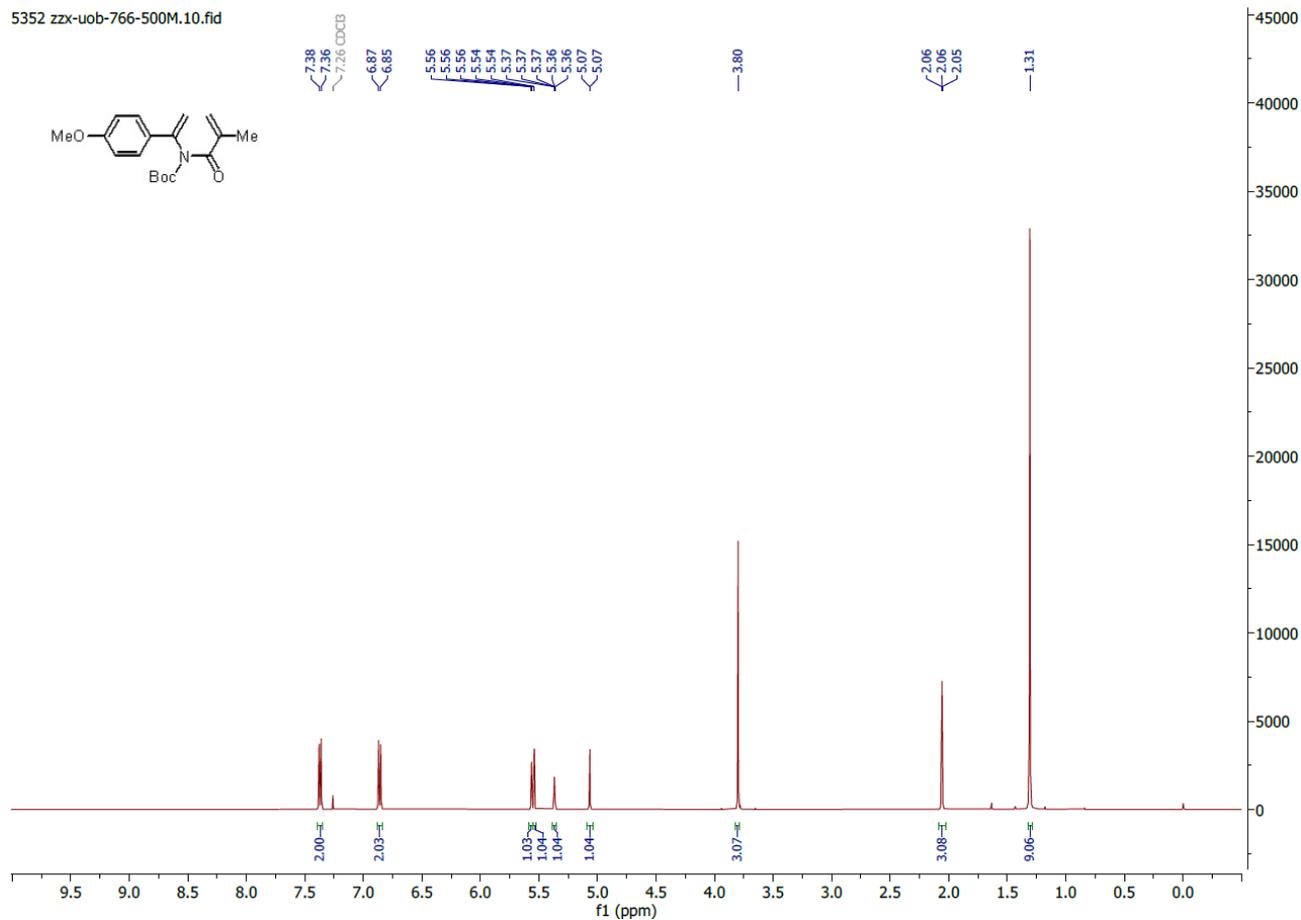
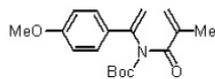


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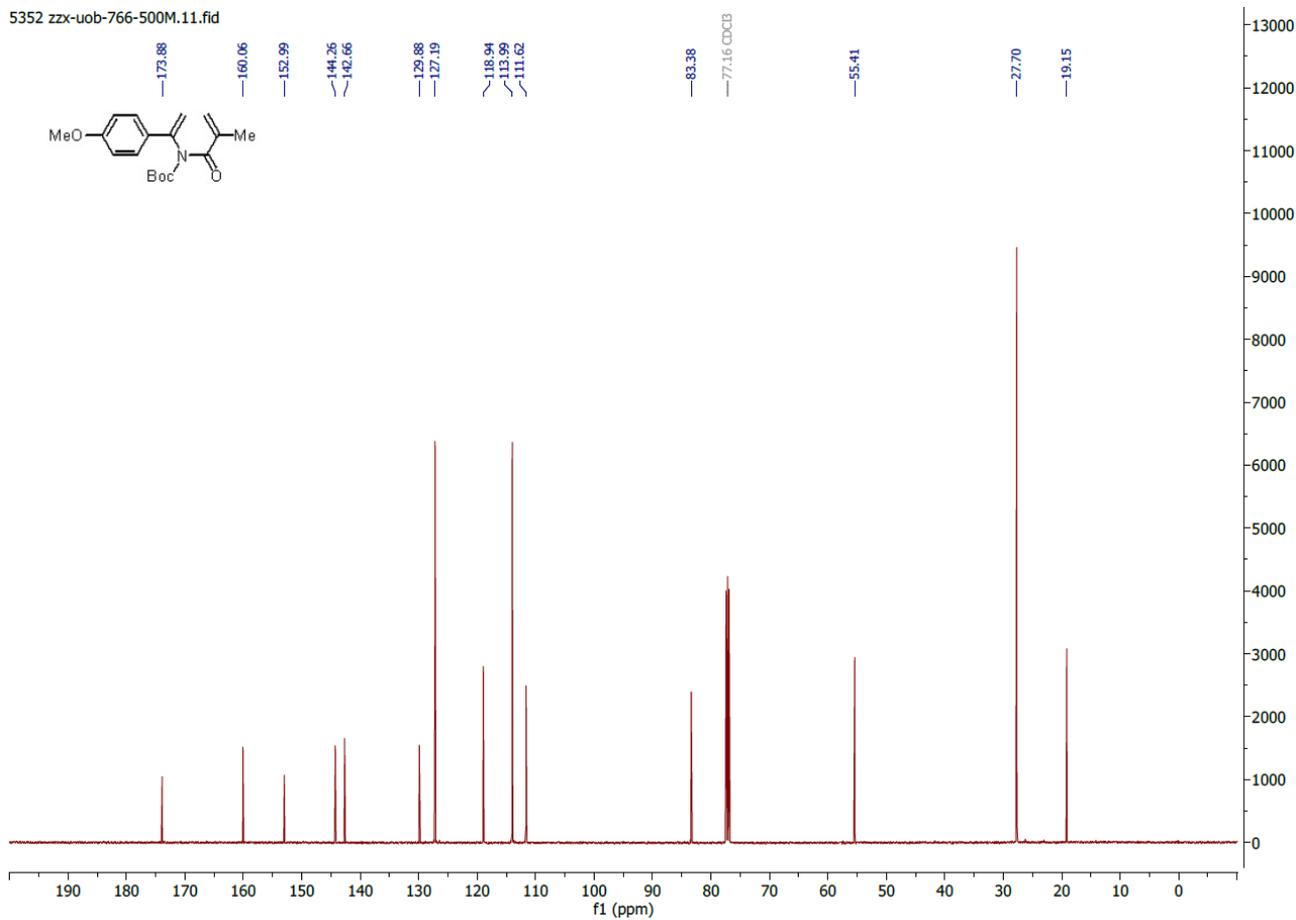
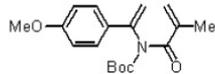


Compound 2ab

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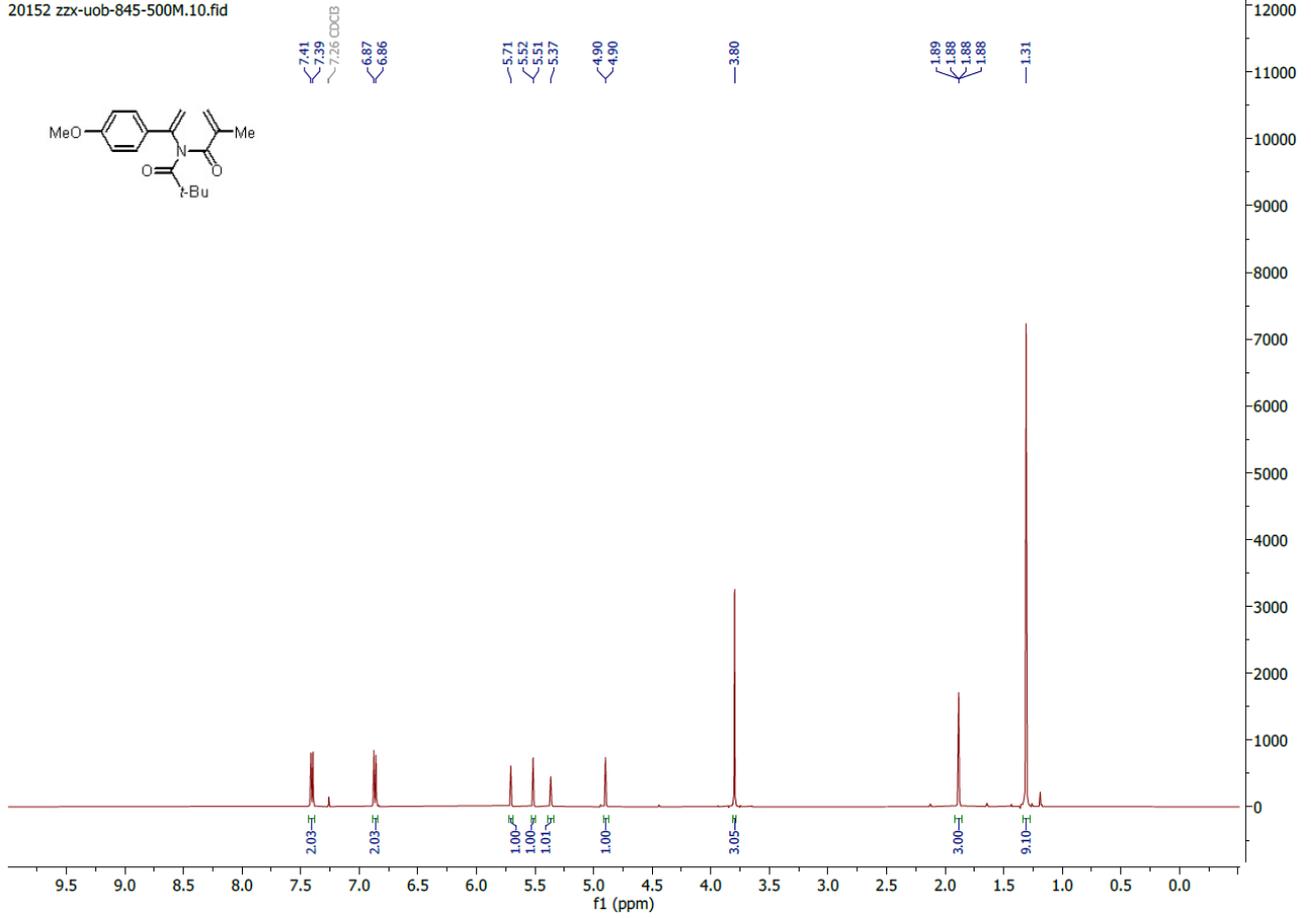


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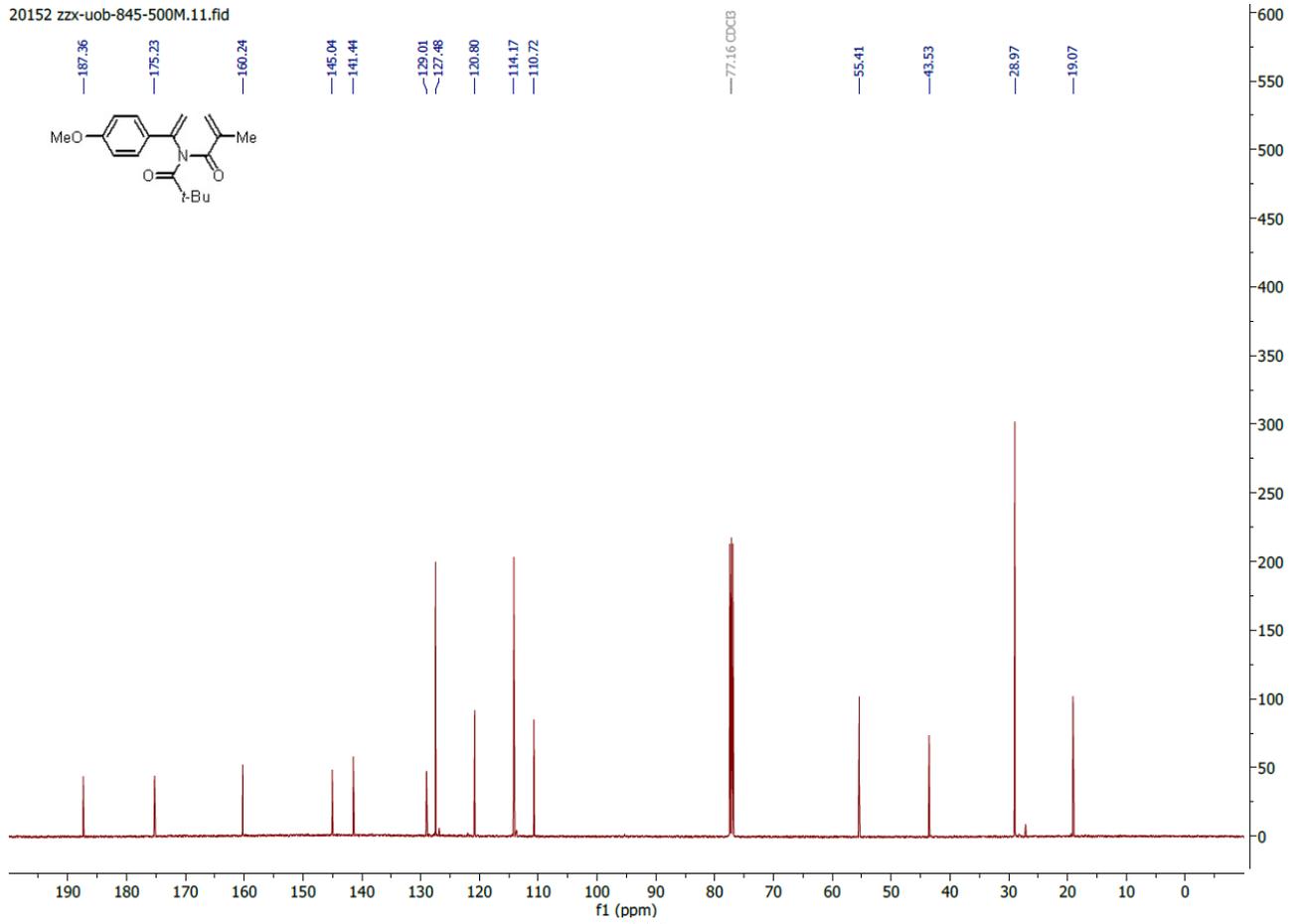


Compound 2ac

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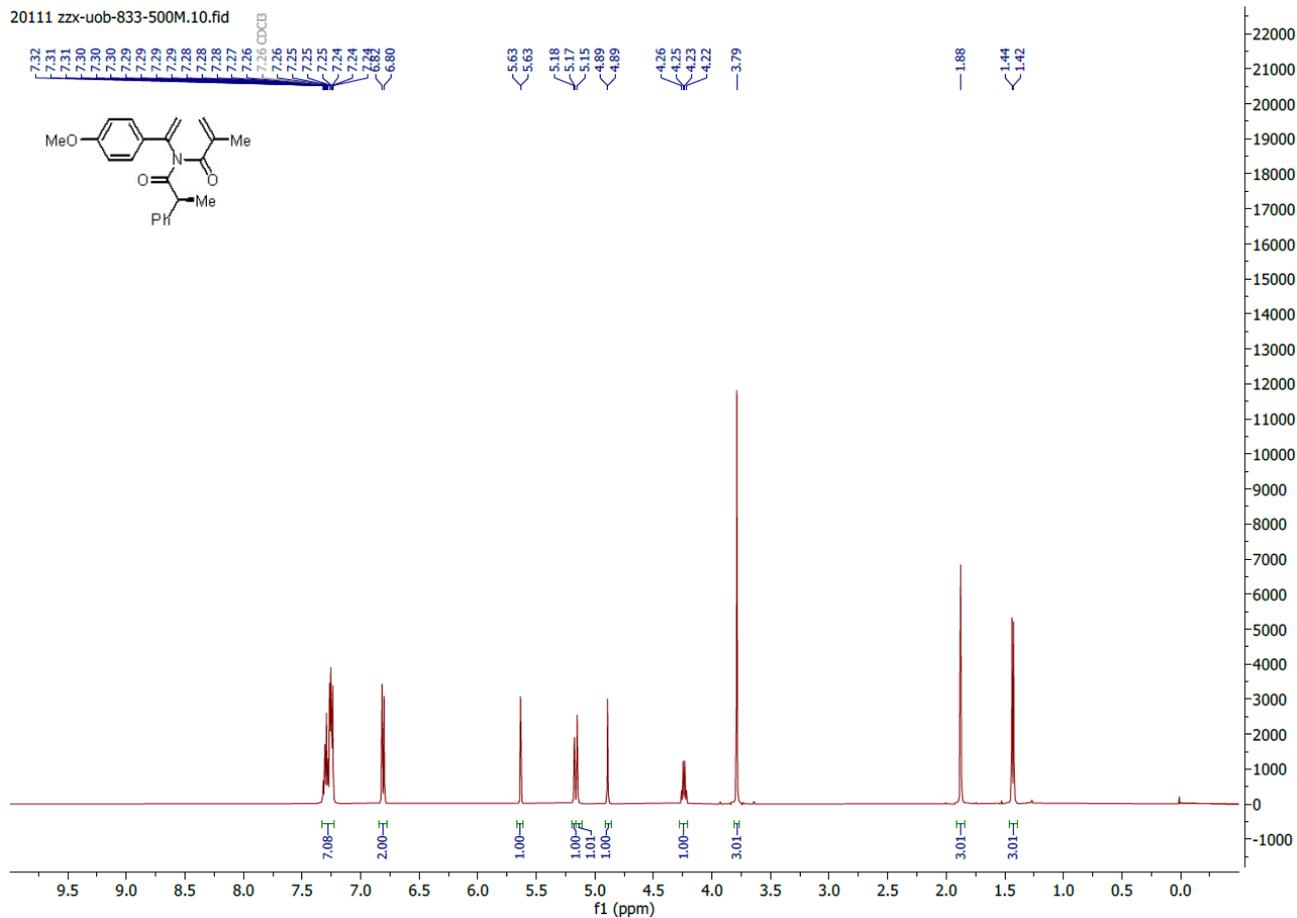


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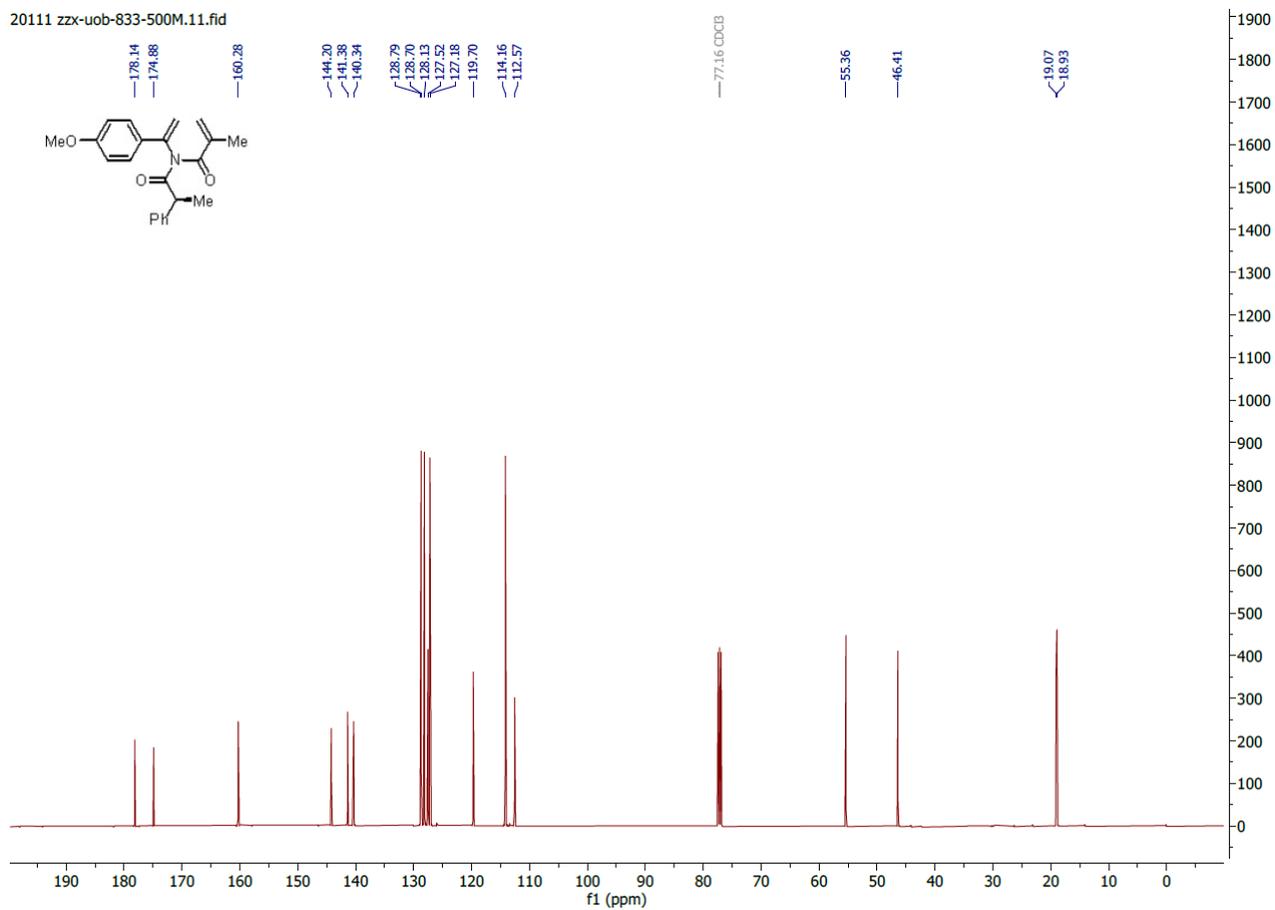


Compound 2ad

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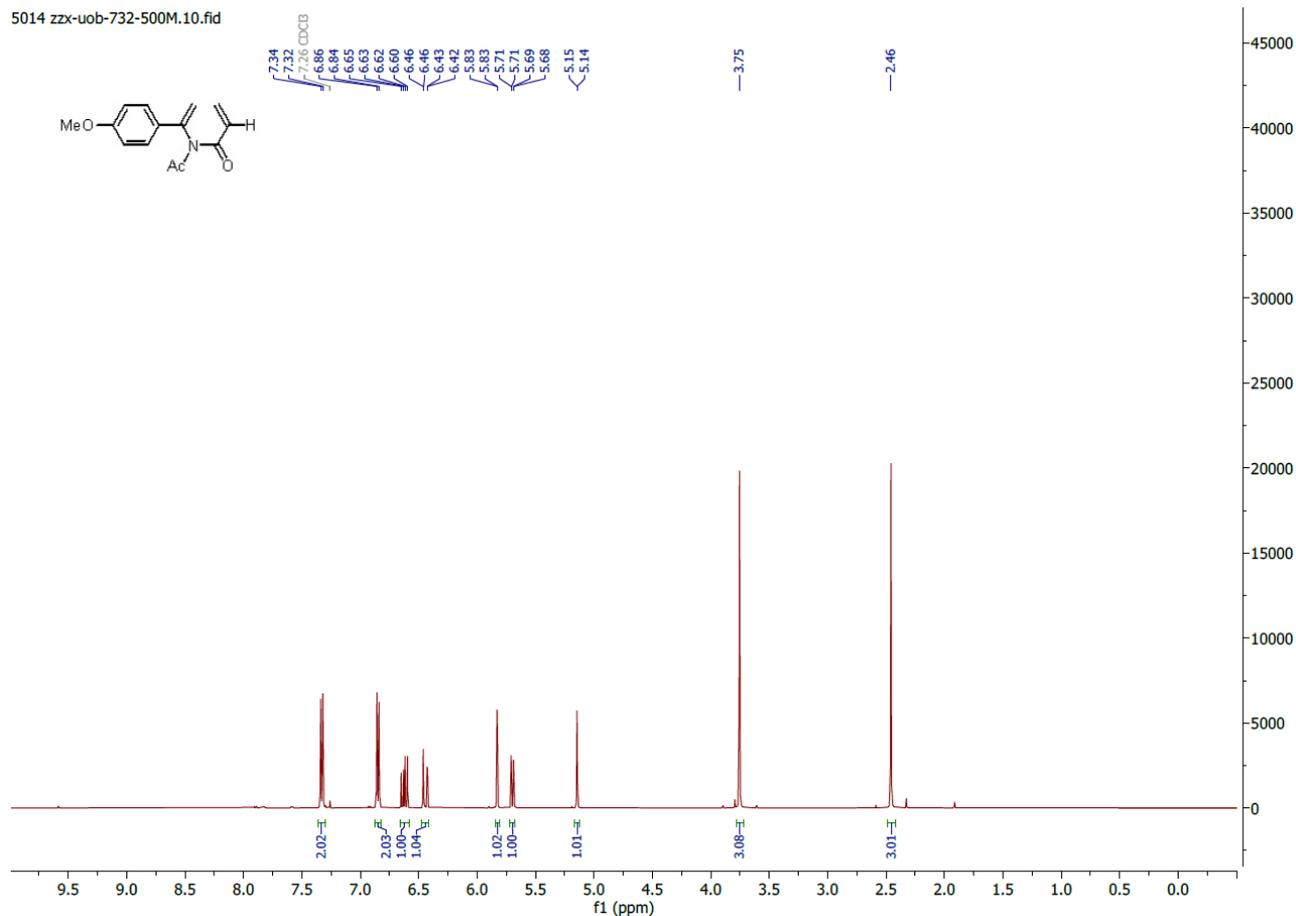
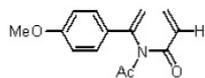


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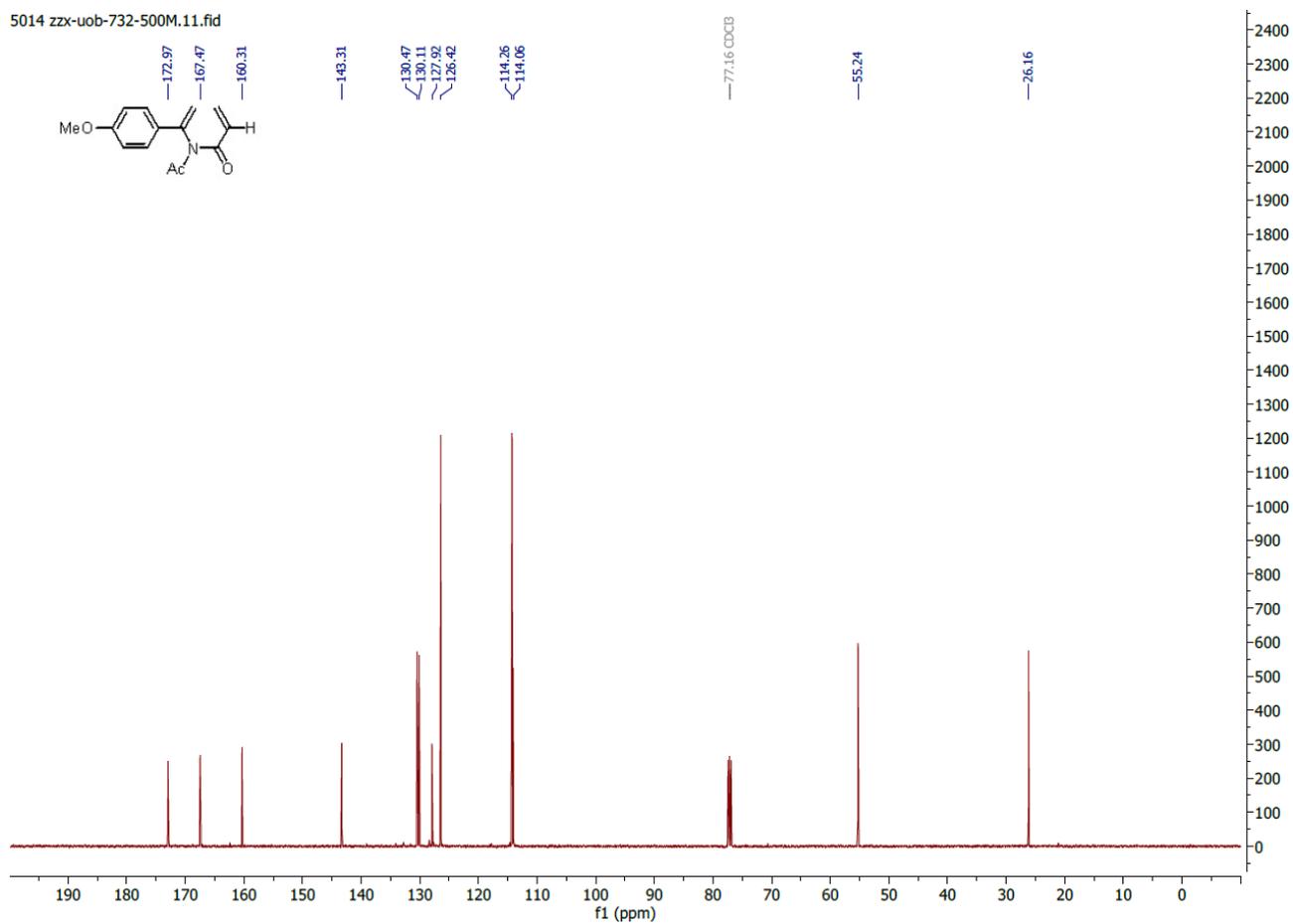
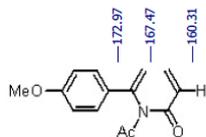


Compound 2ae

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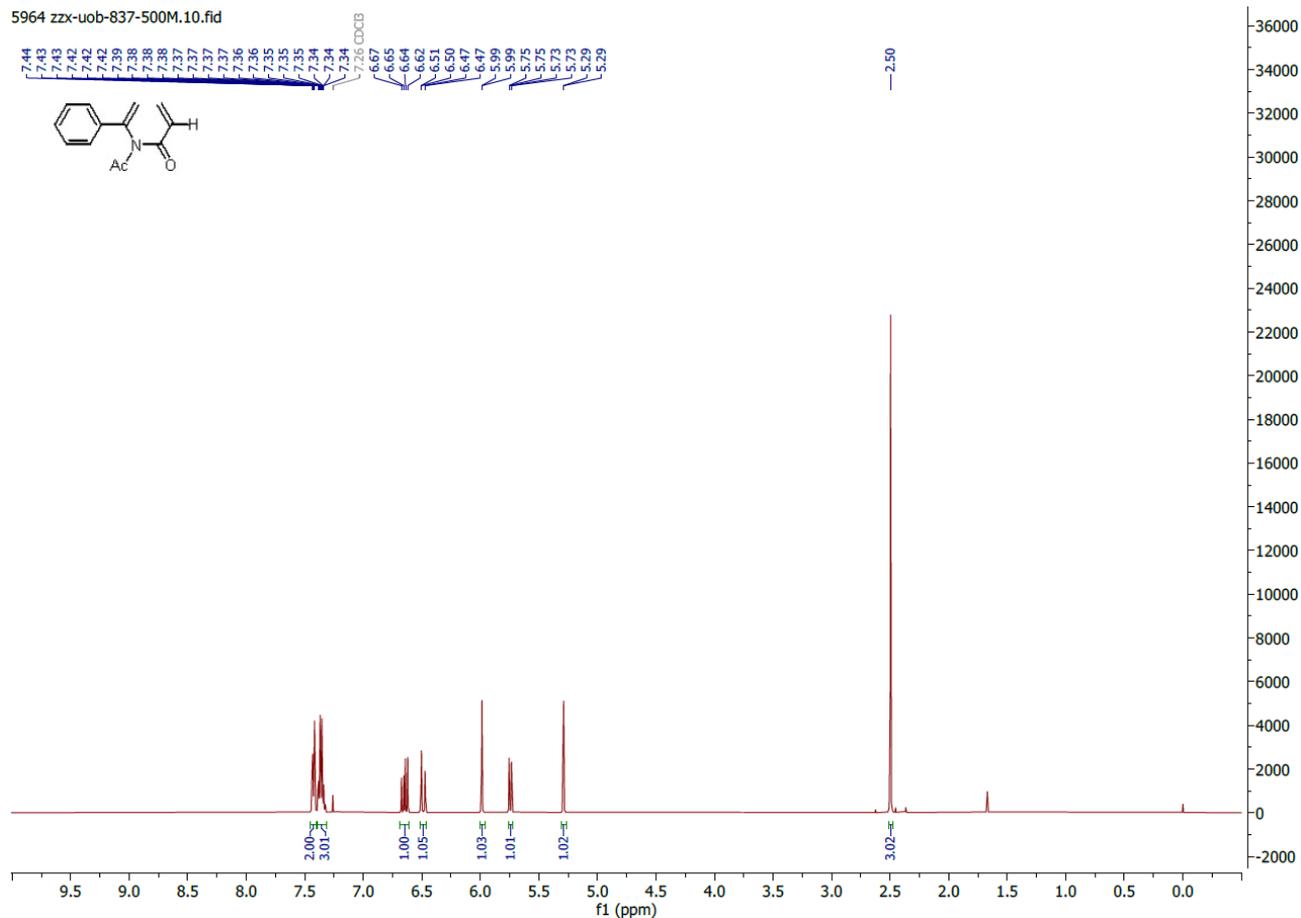


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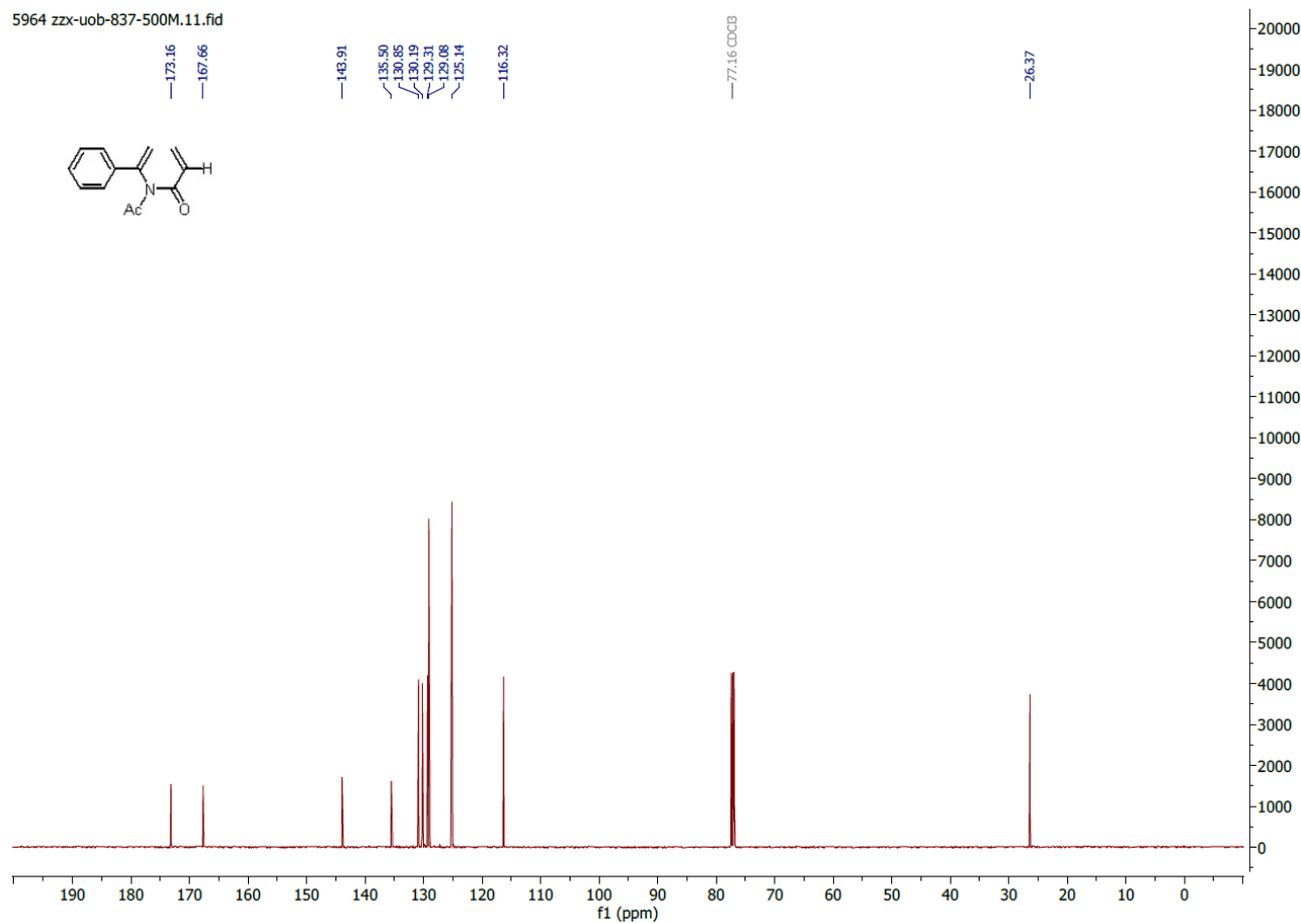


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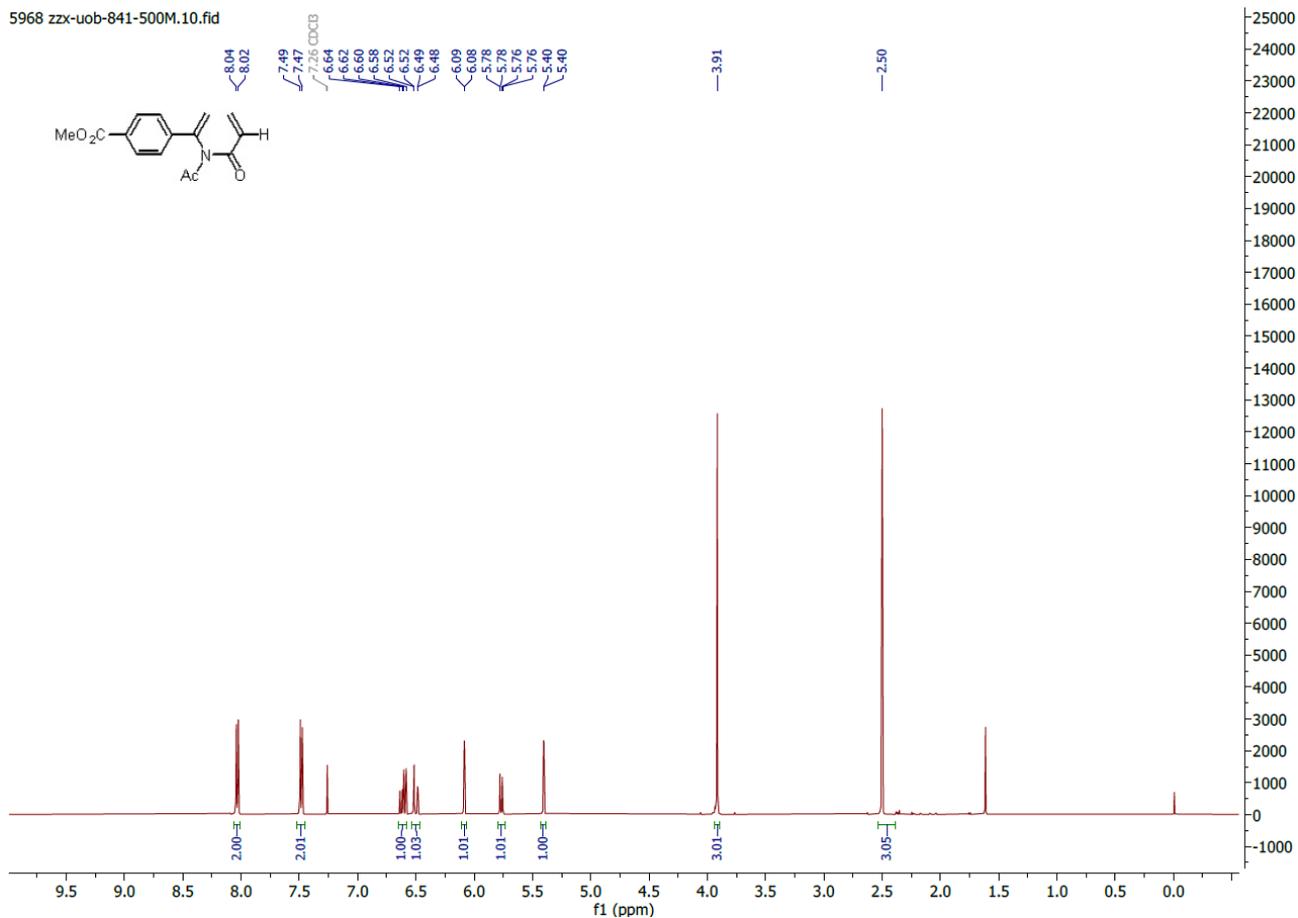


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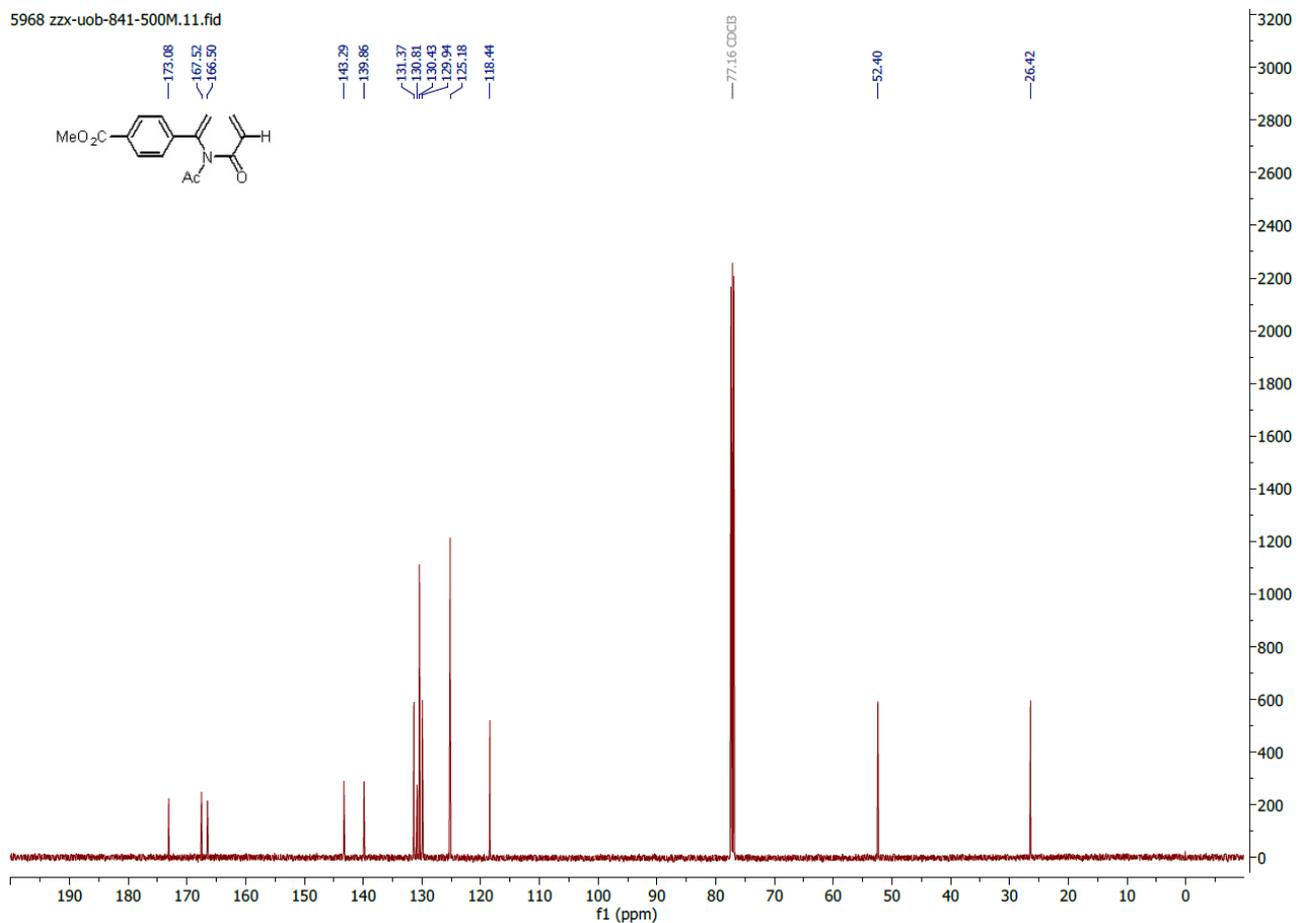


Compound 2ag

5968 zzx-uob-841-500M.10.fid

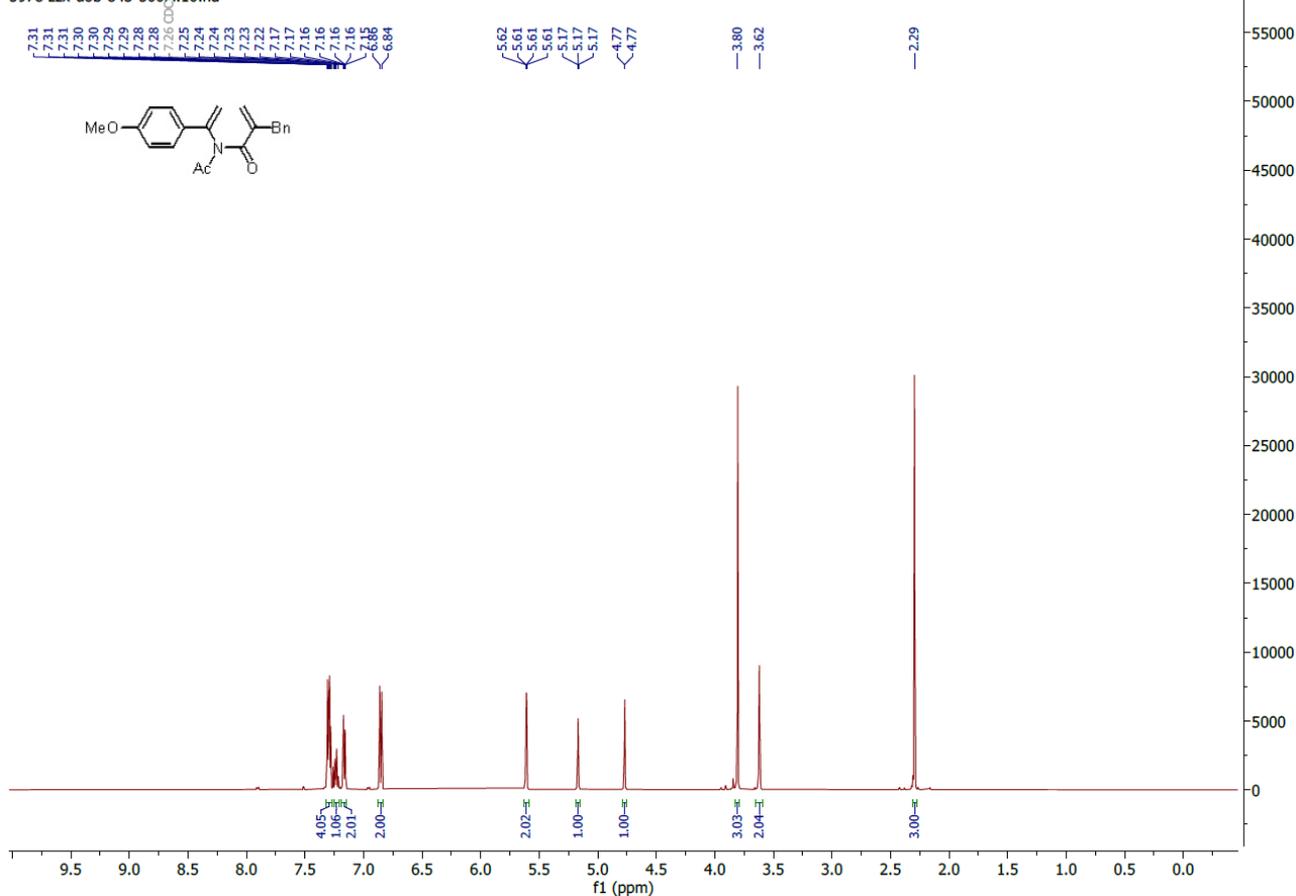


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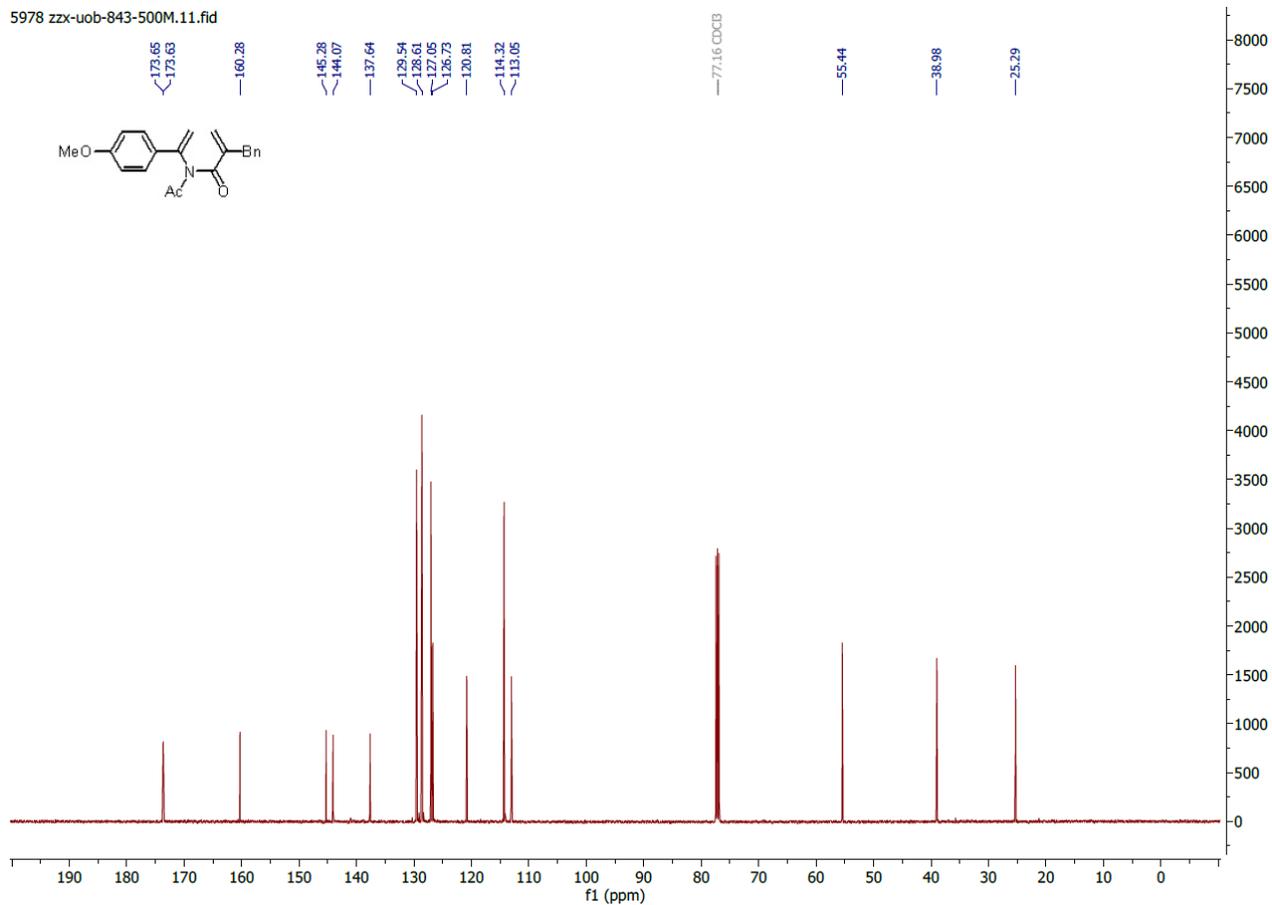


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5978 zzx-uob-843-500M.10.fid

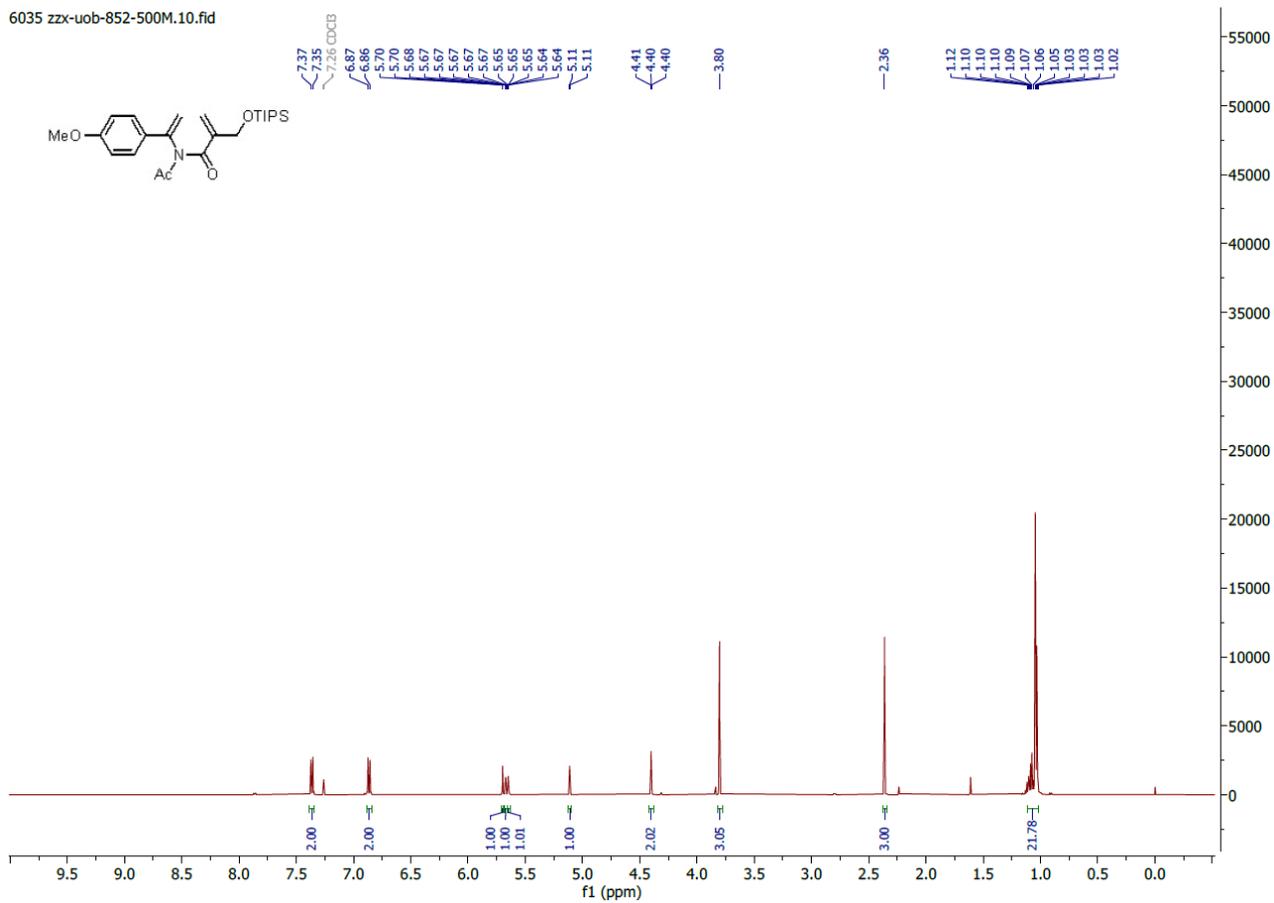
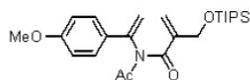


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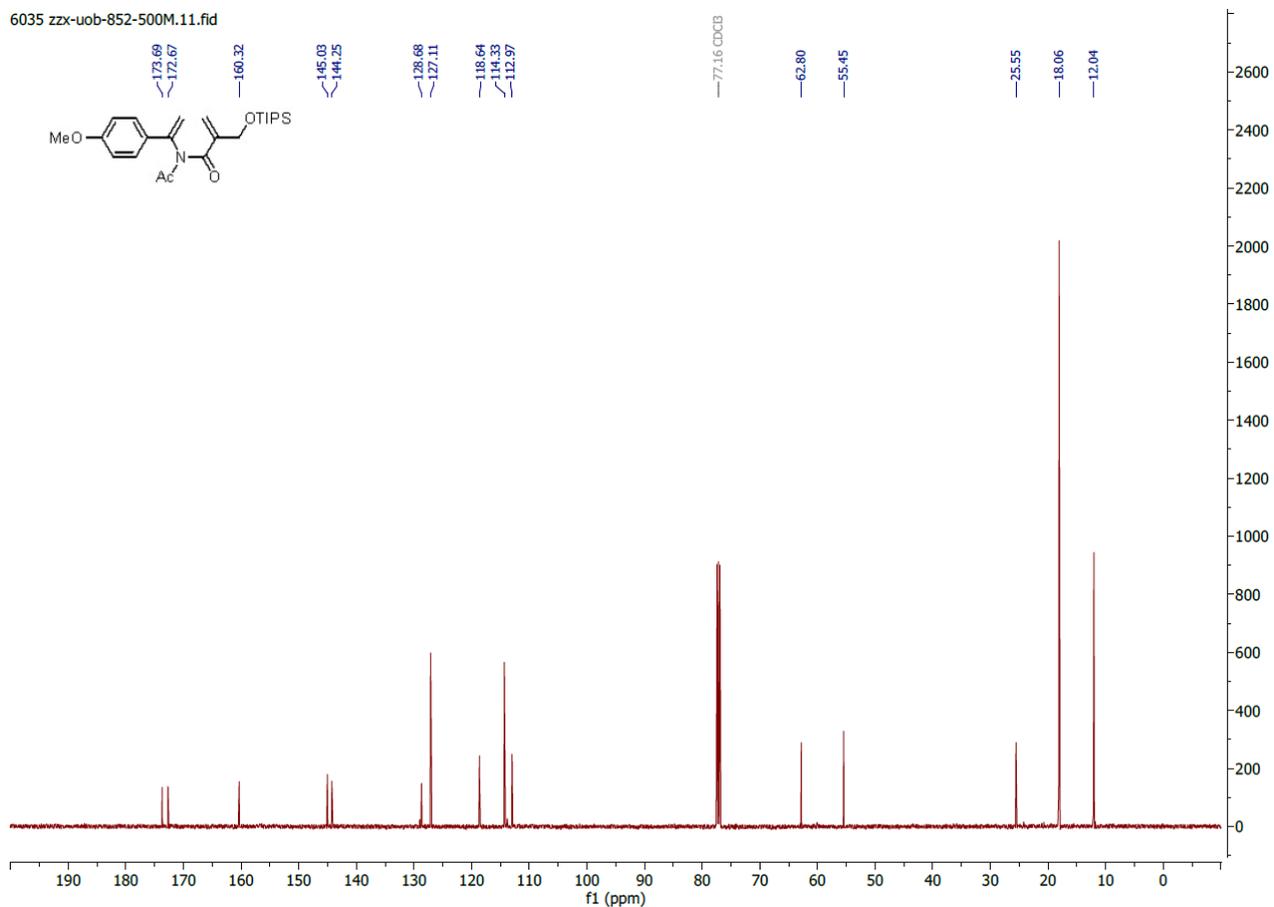
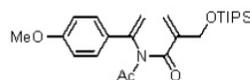


Compound 2ai

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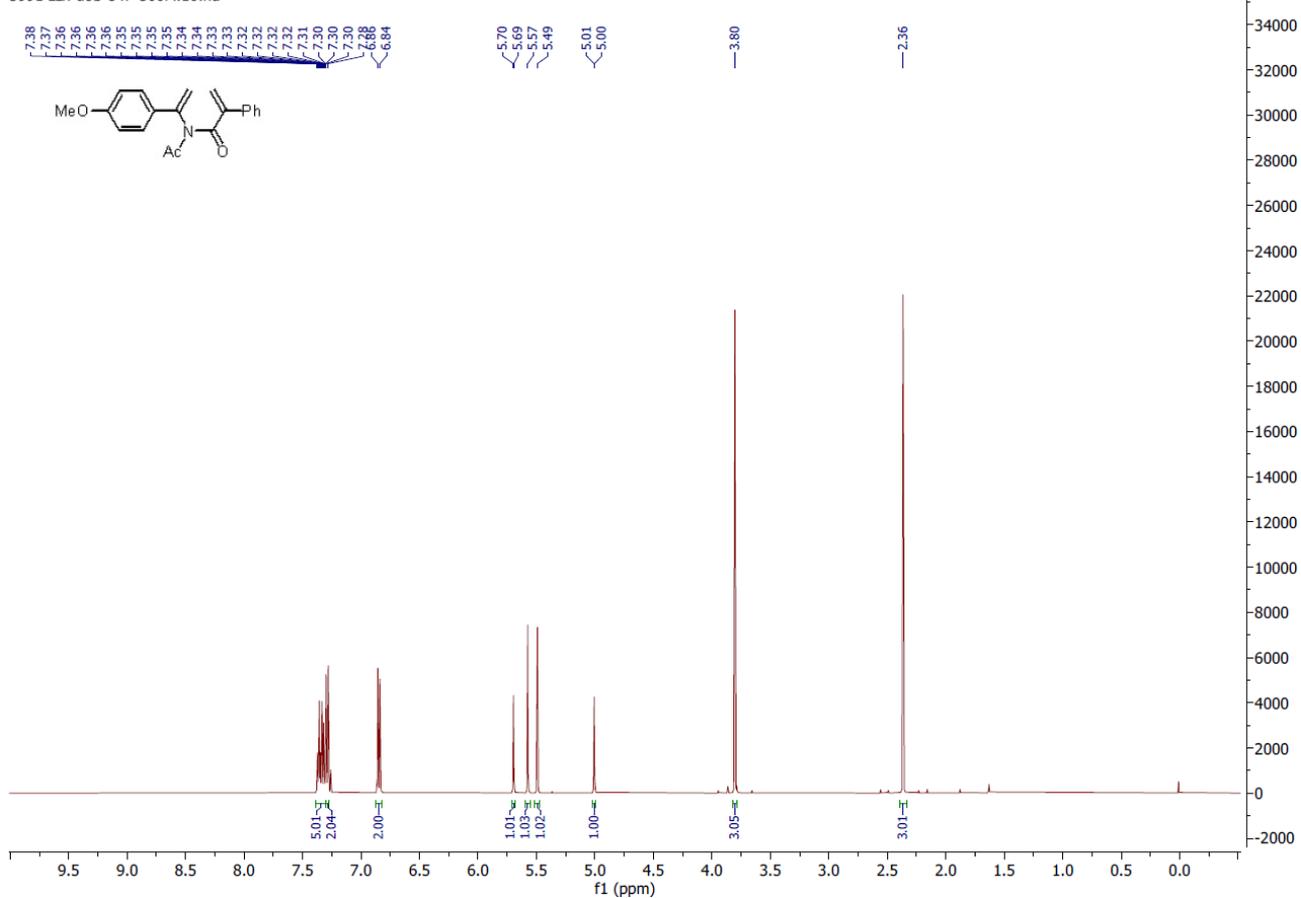


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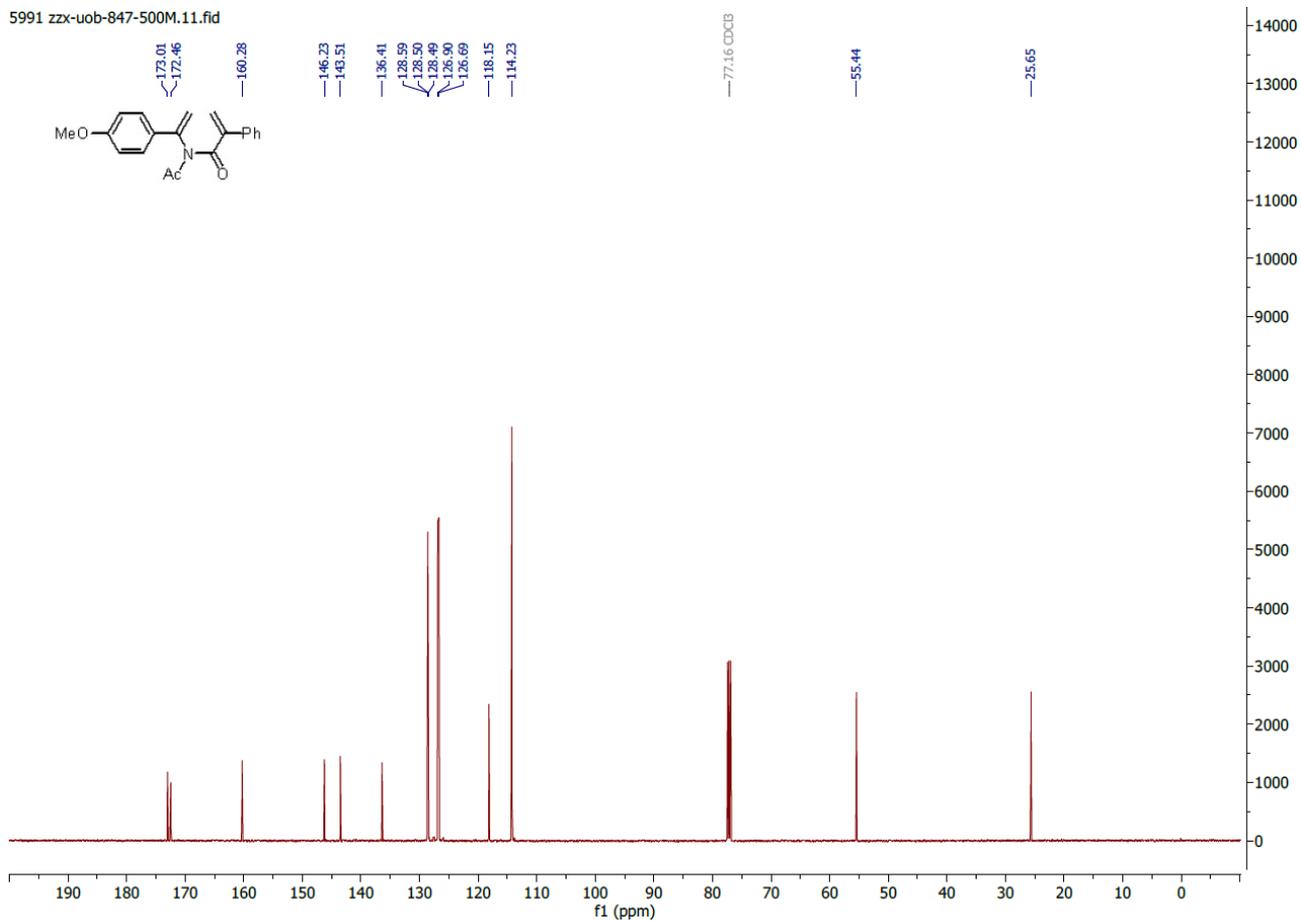


Compound 2aj

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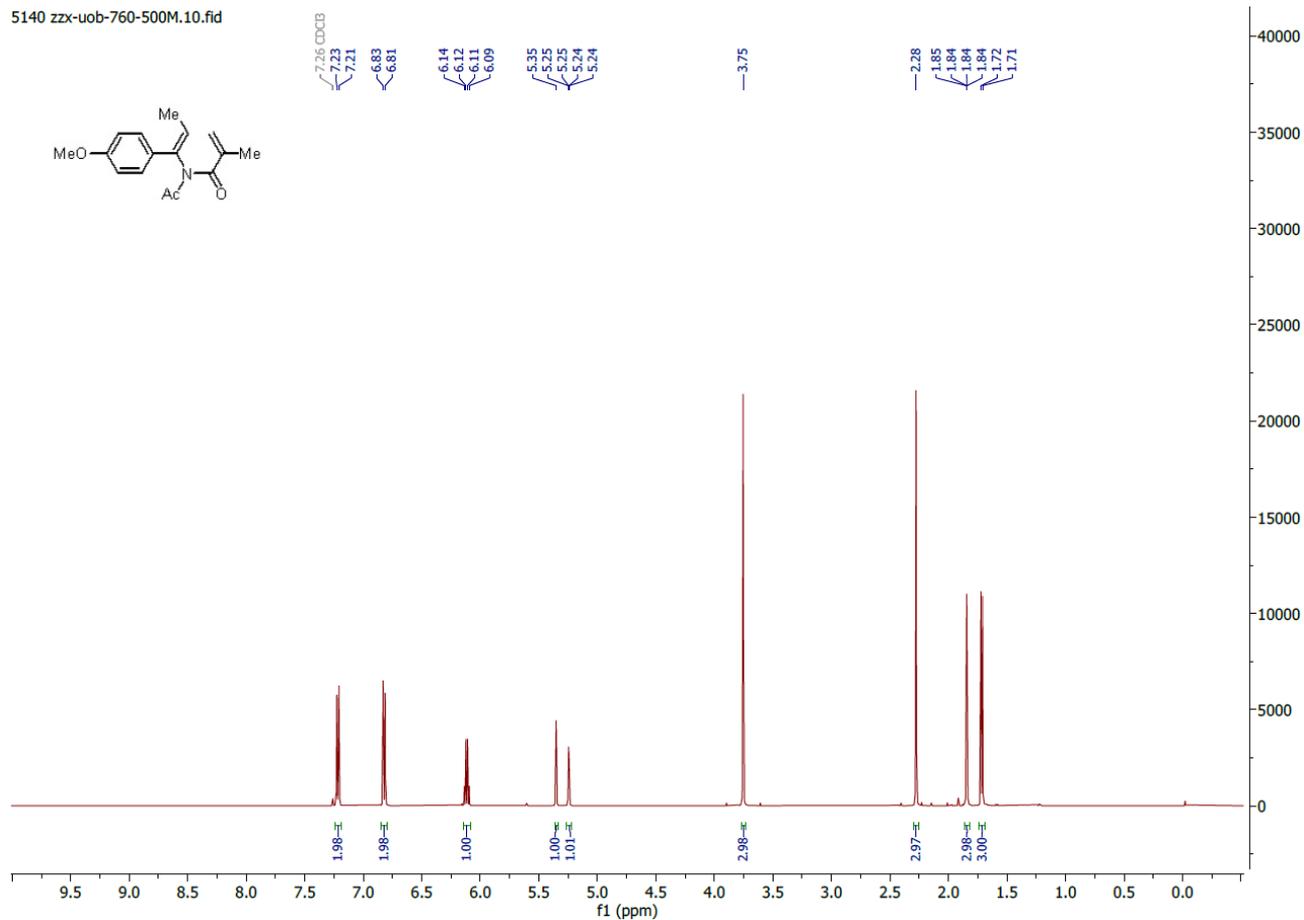
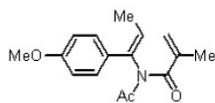


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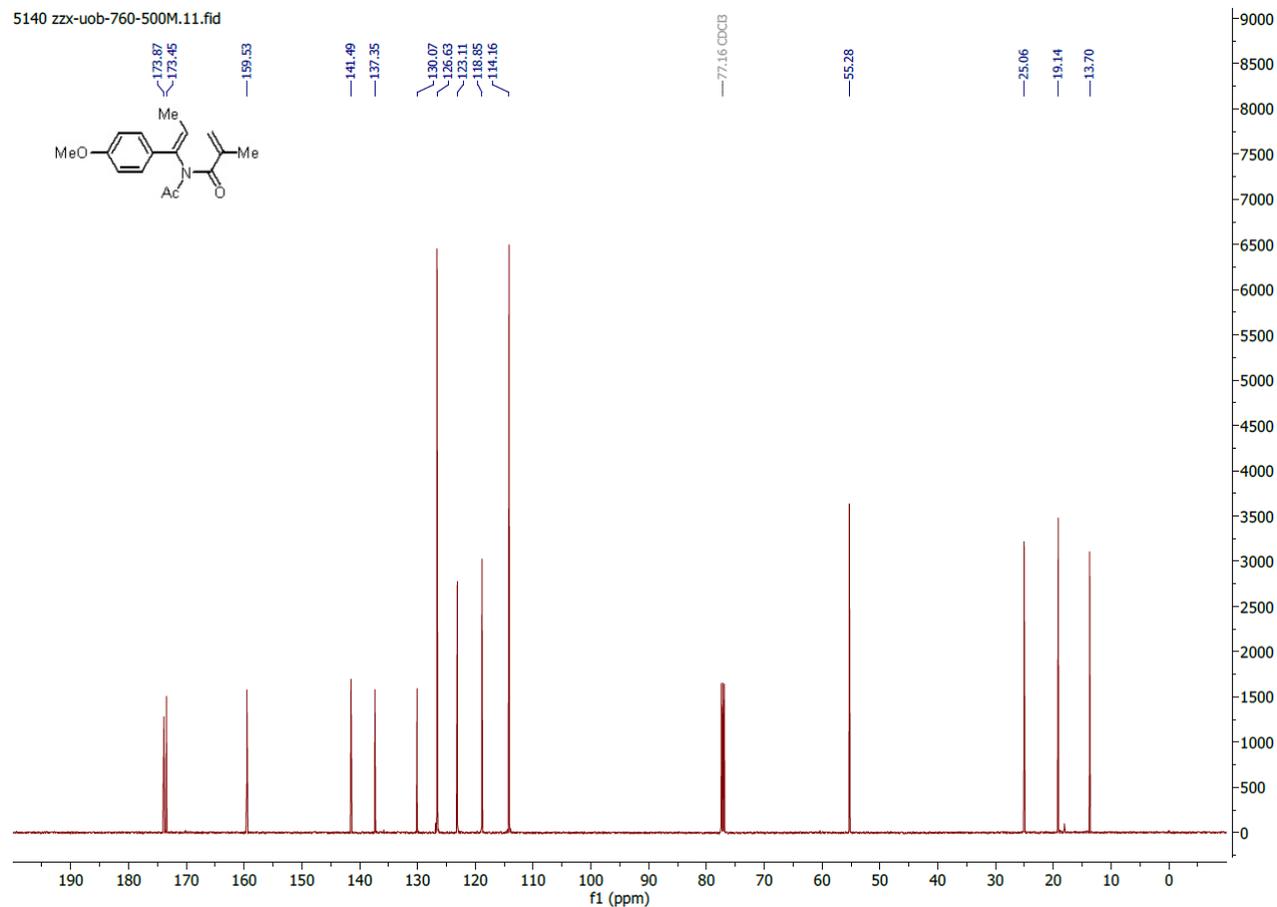
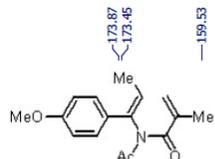


Compound 2ak

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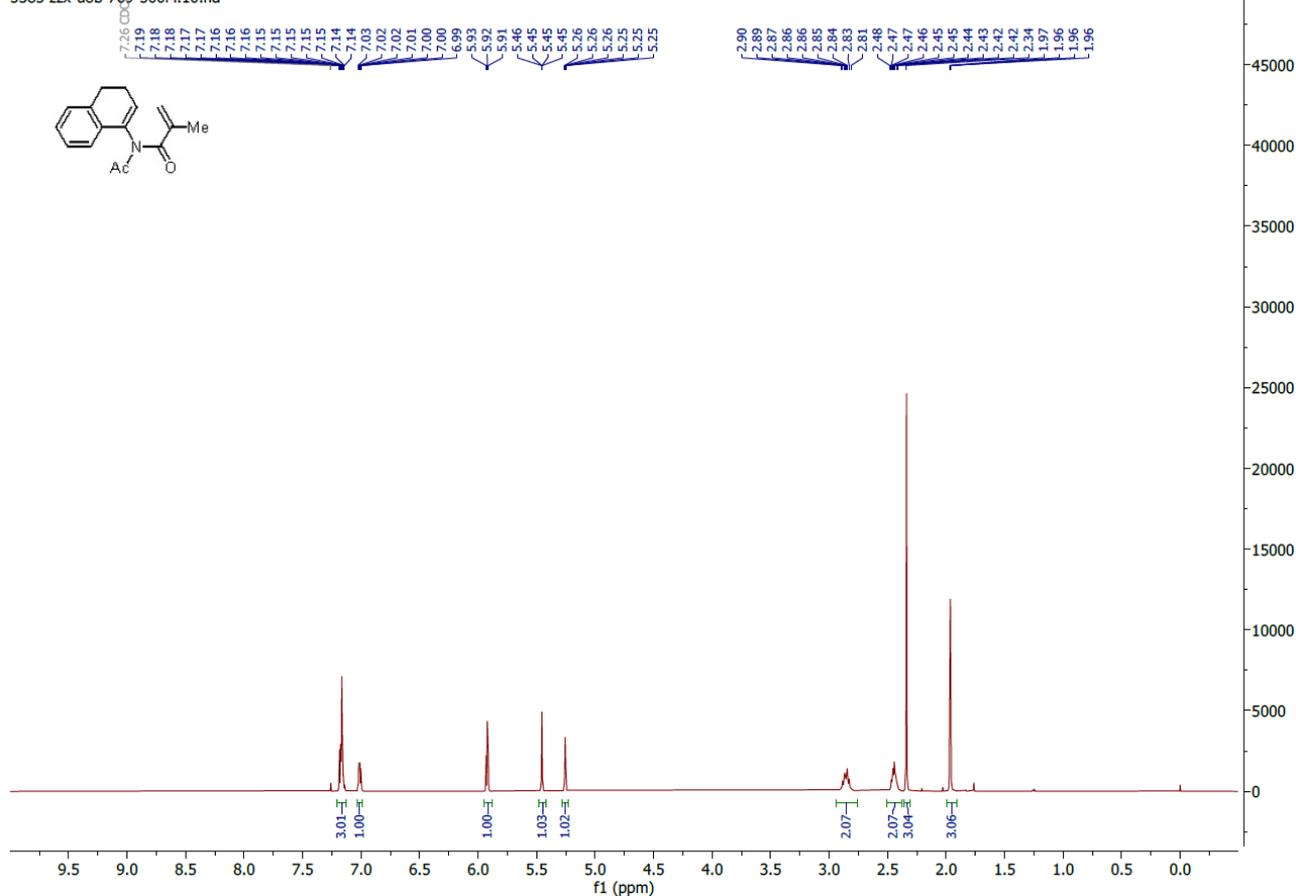


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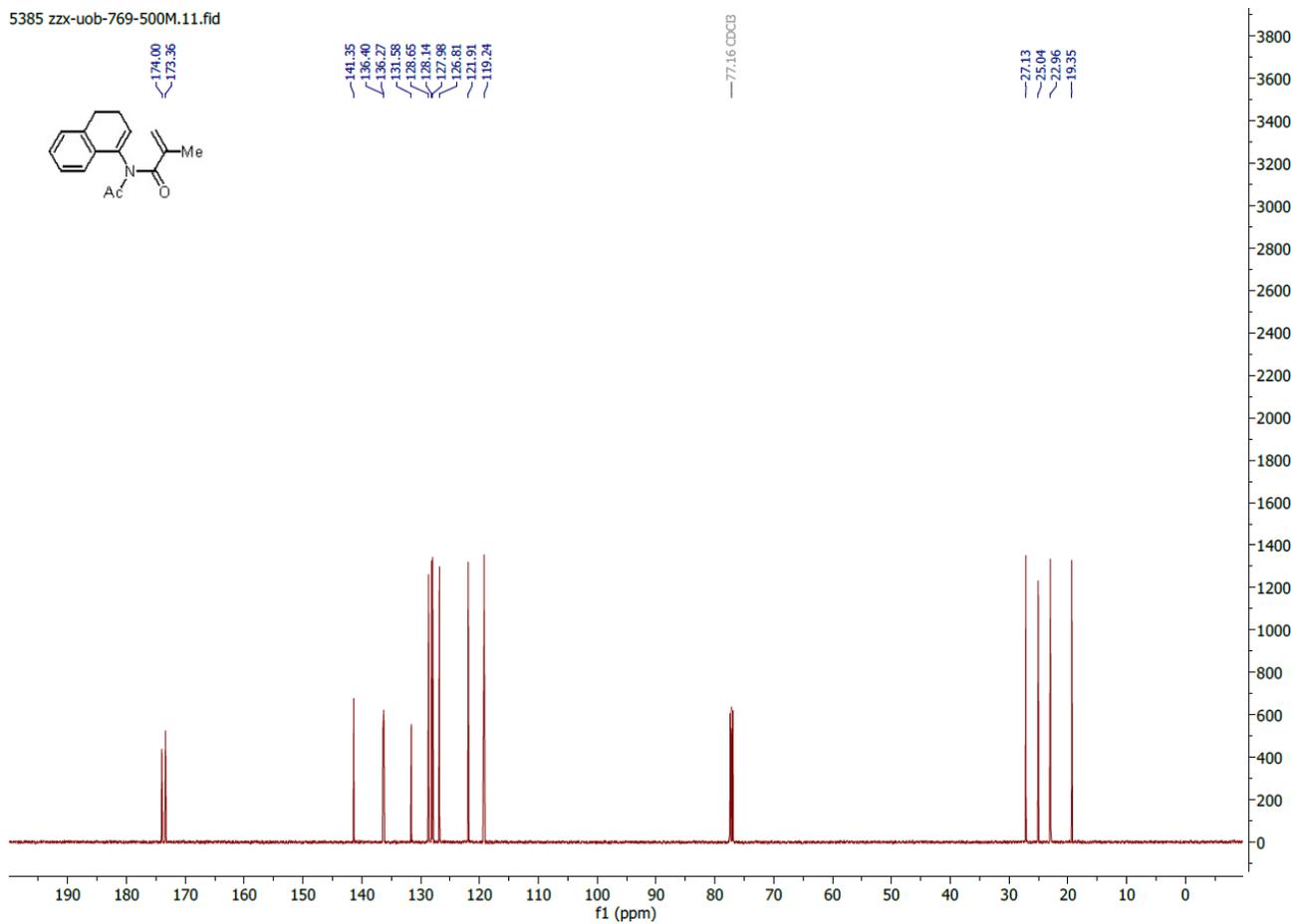


Compound 2a

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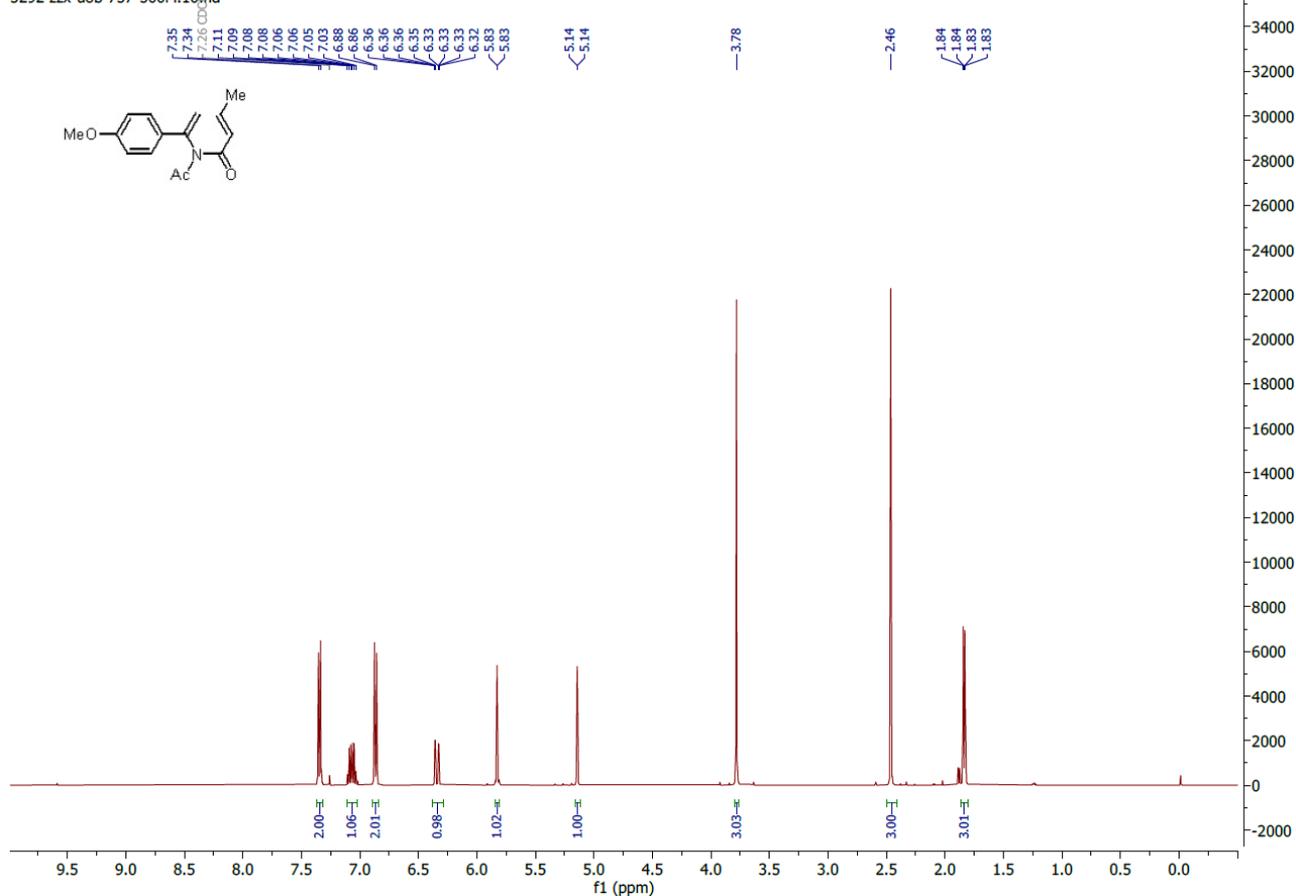


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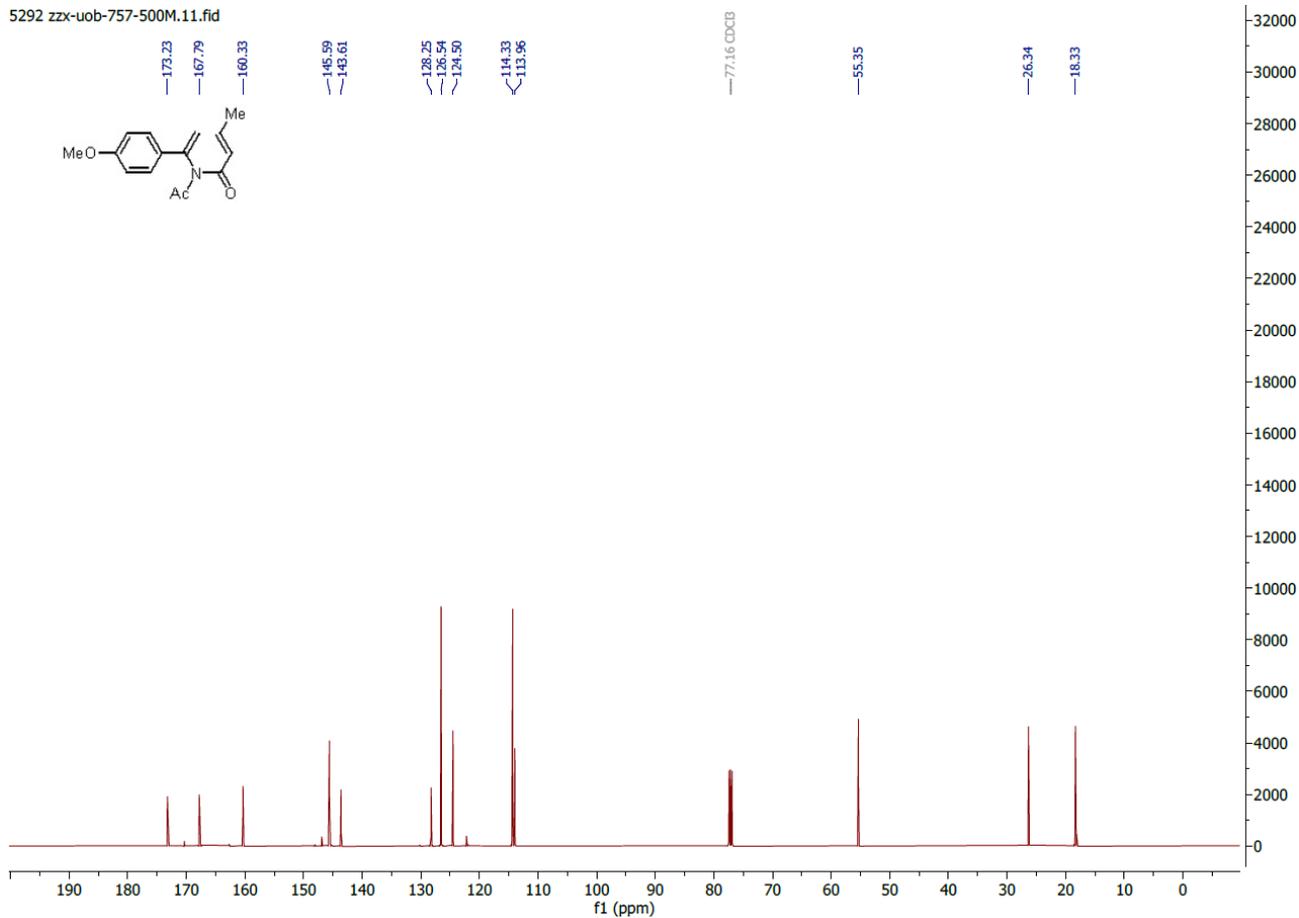


Compound 2am

5292 zzx-uob-757-500M.10.fid

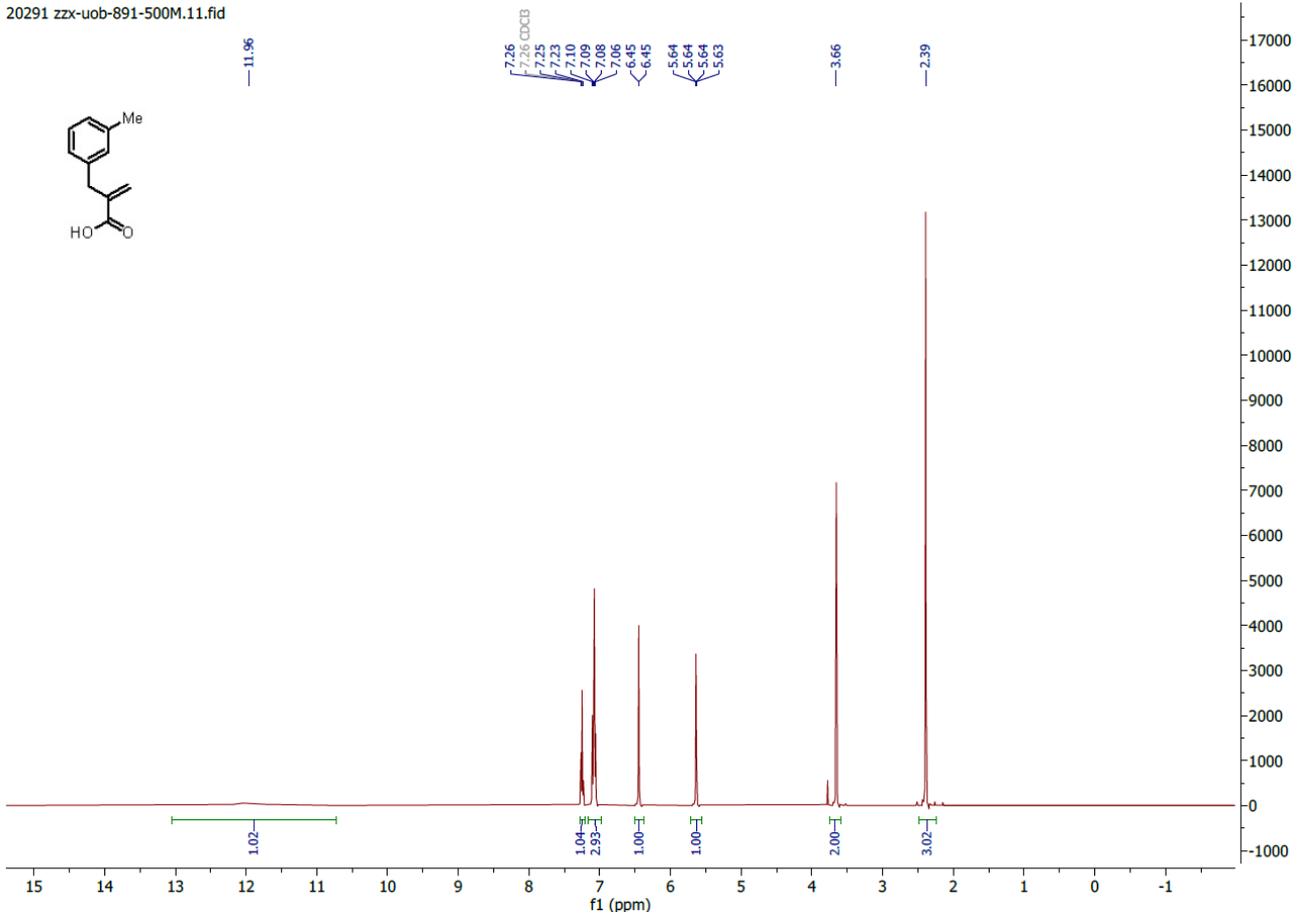
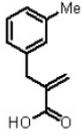


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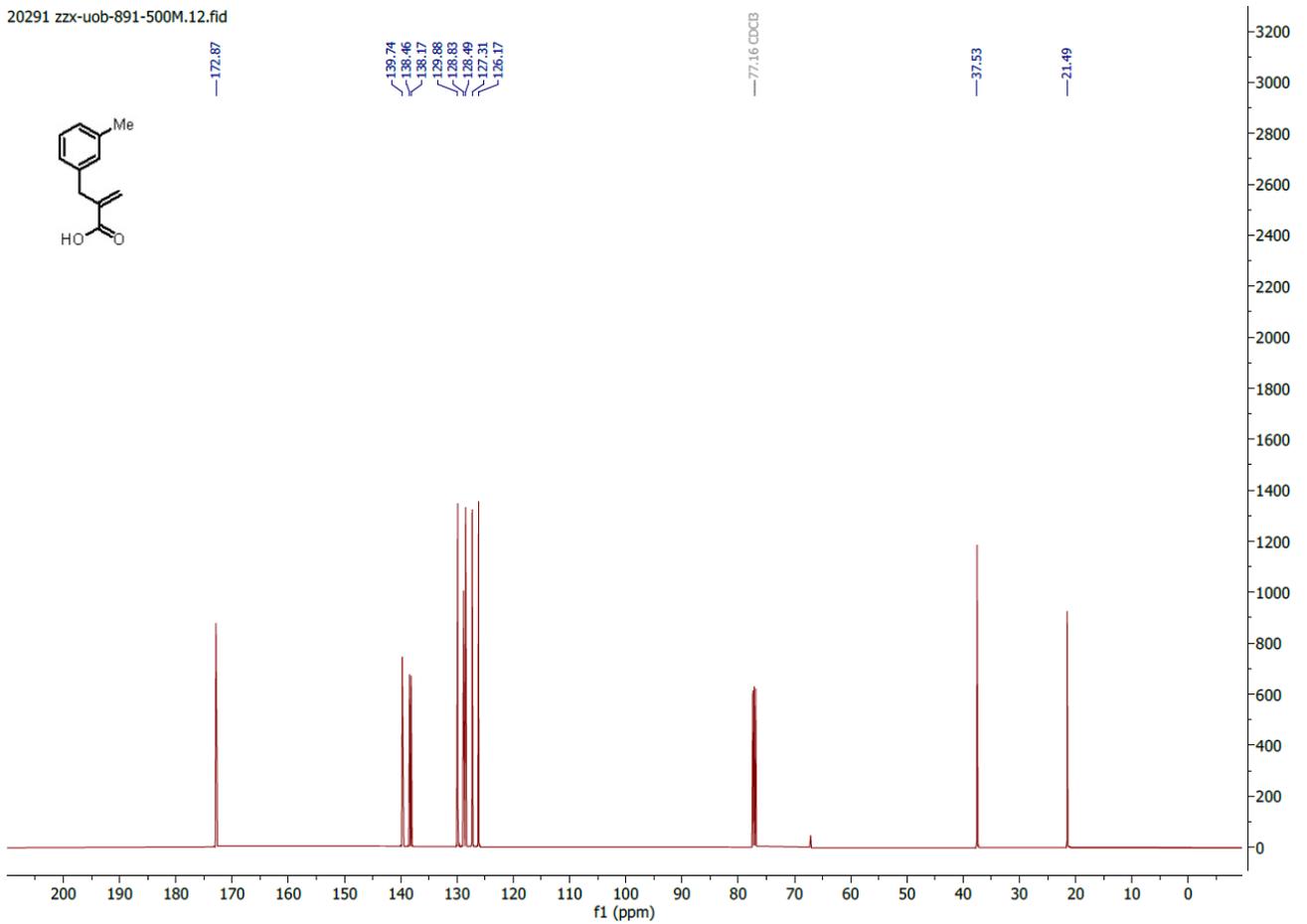
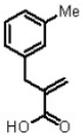


Compound S2an'

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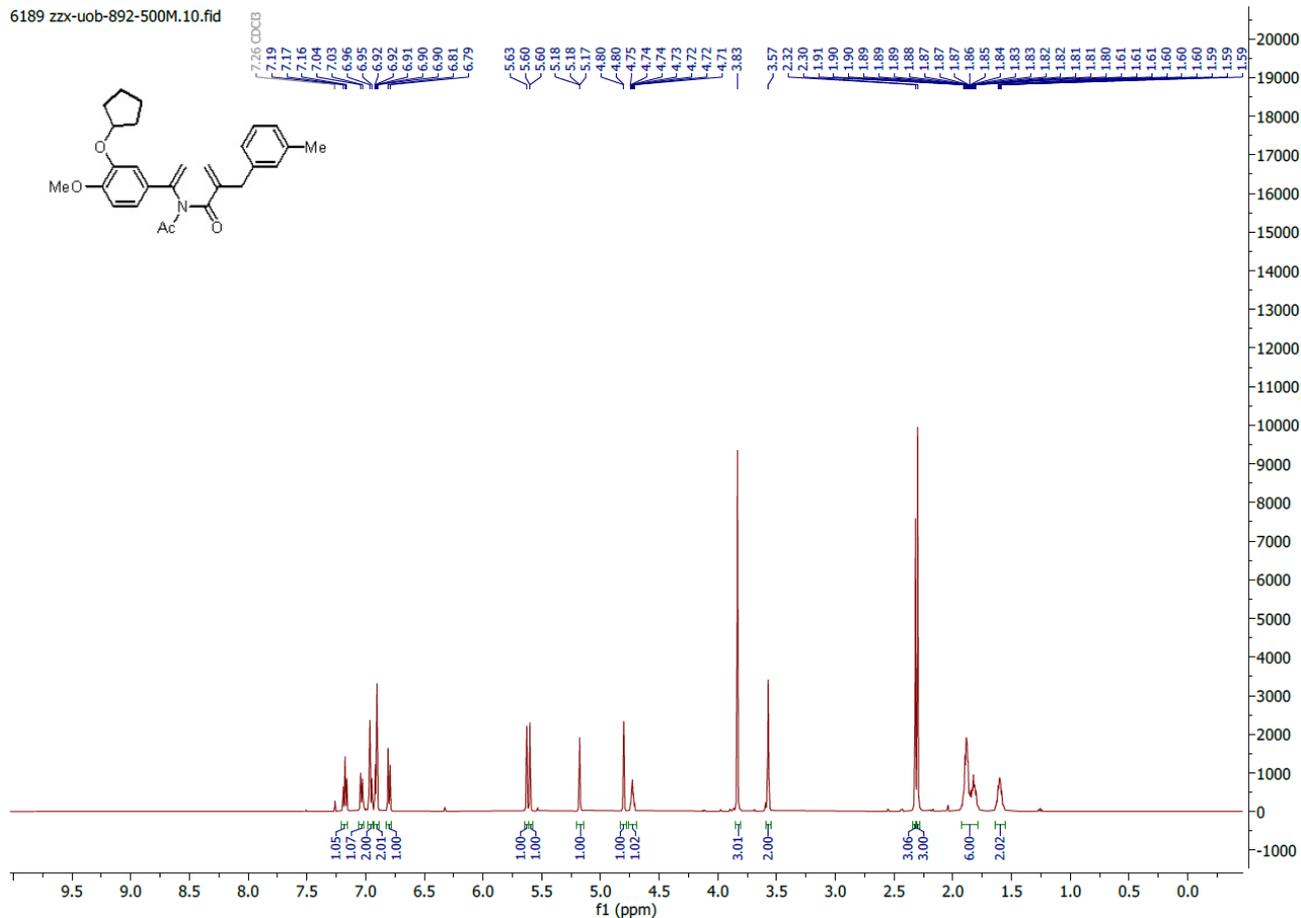


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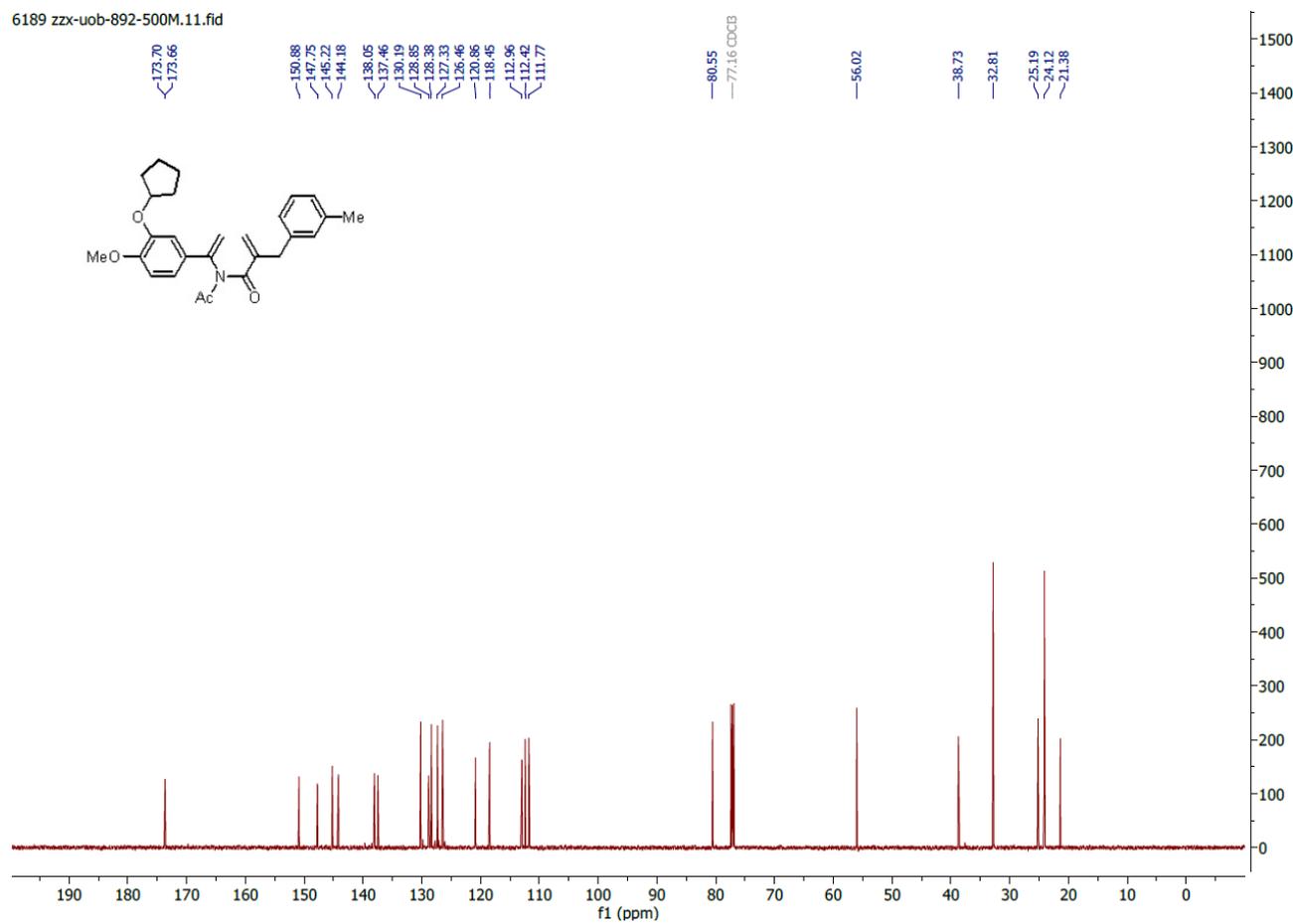


Compound 2a

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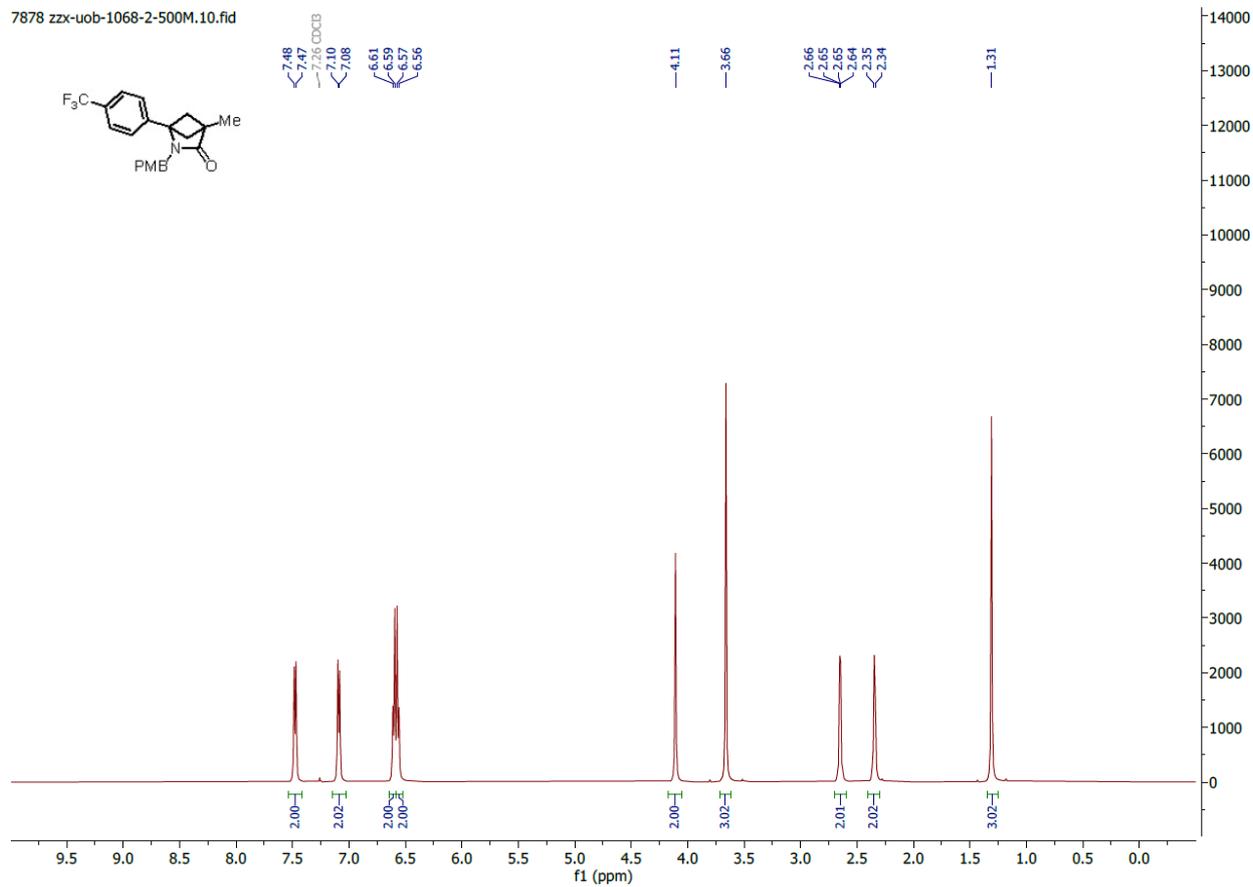
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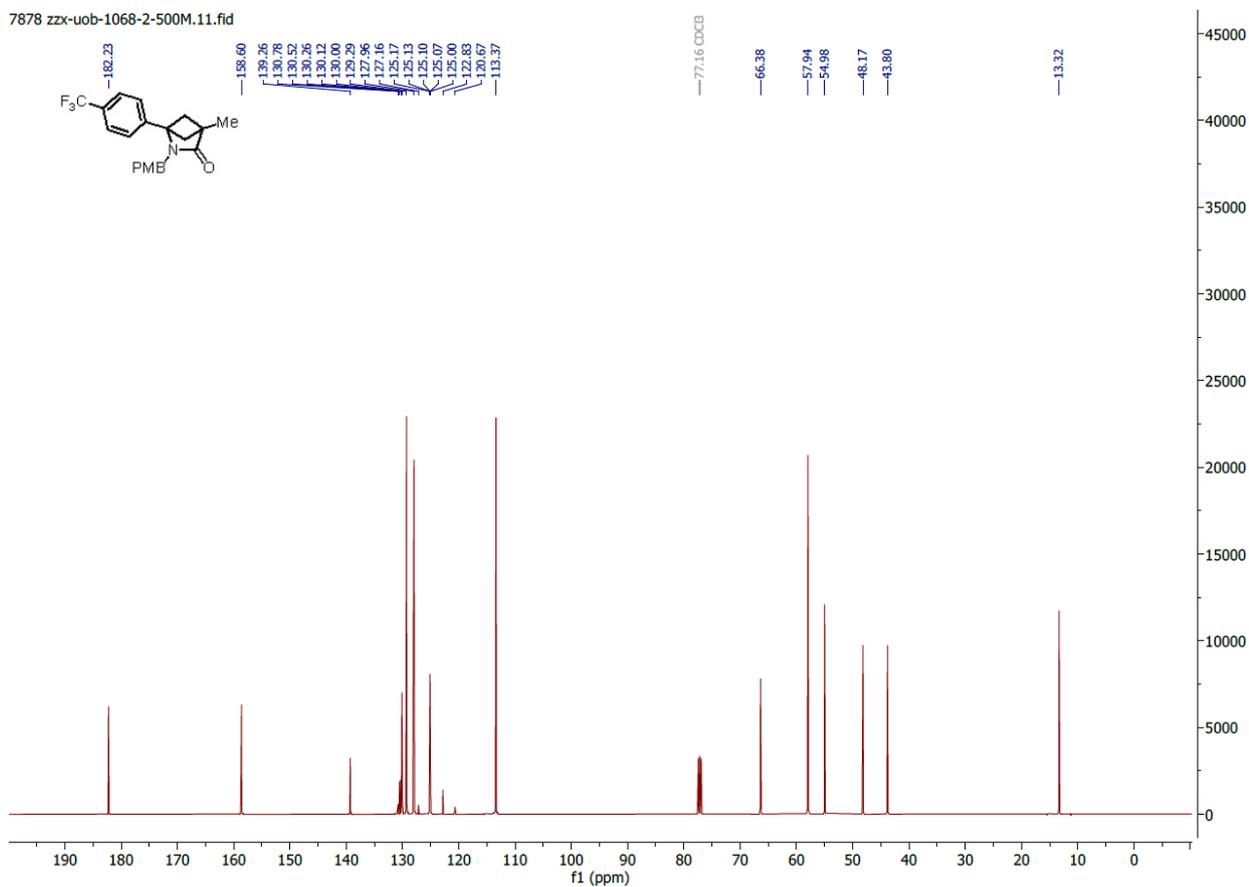
5.4 NMR Spectra of Products (3 and 4)

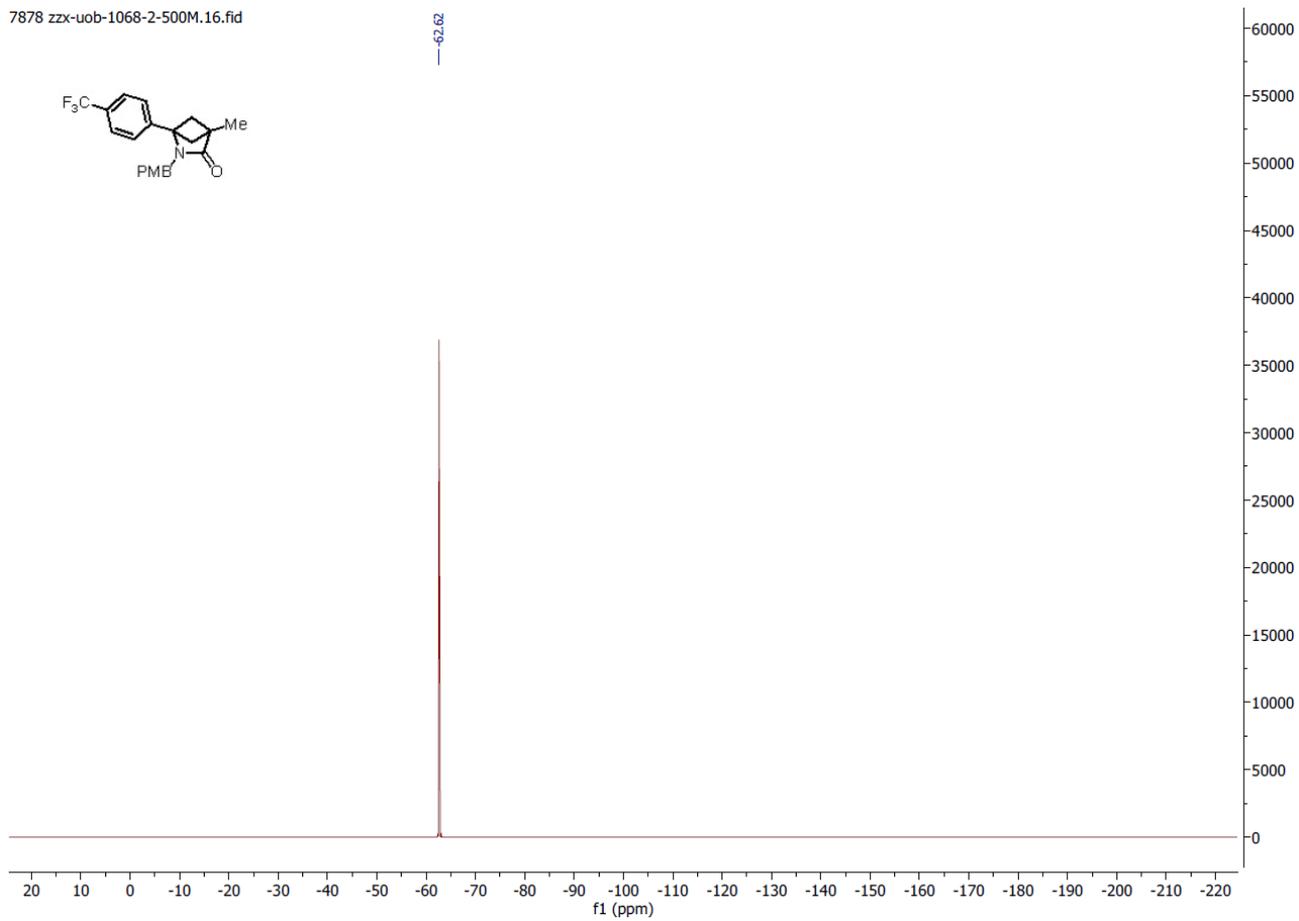
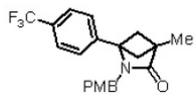
Compound 3a

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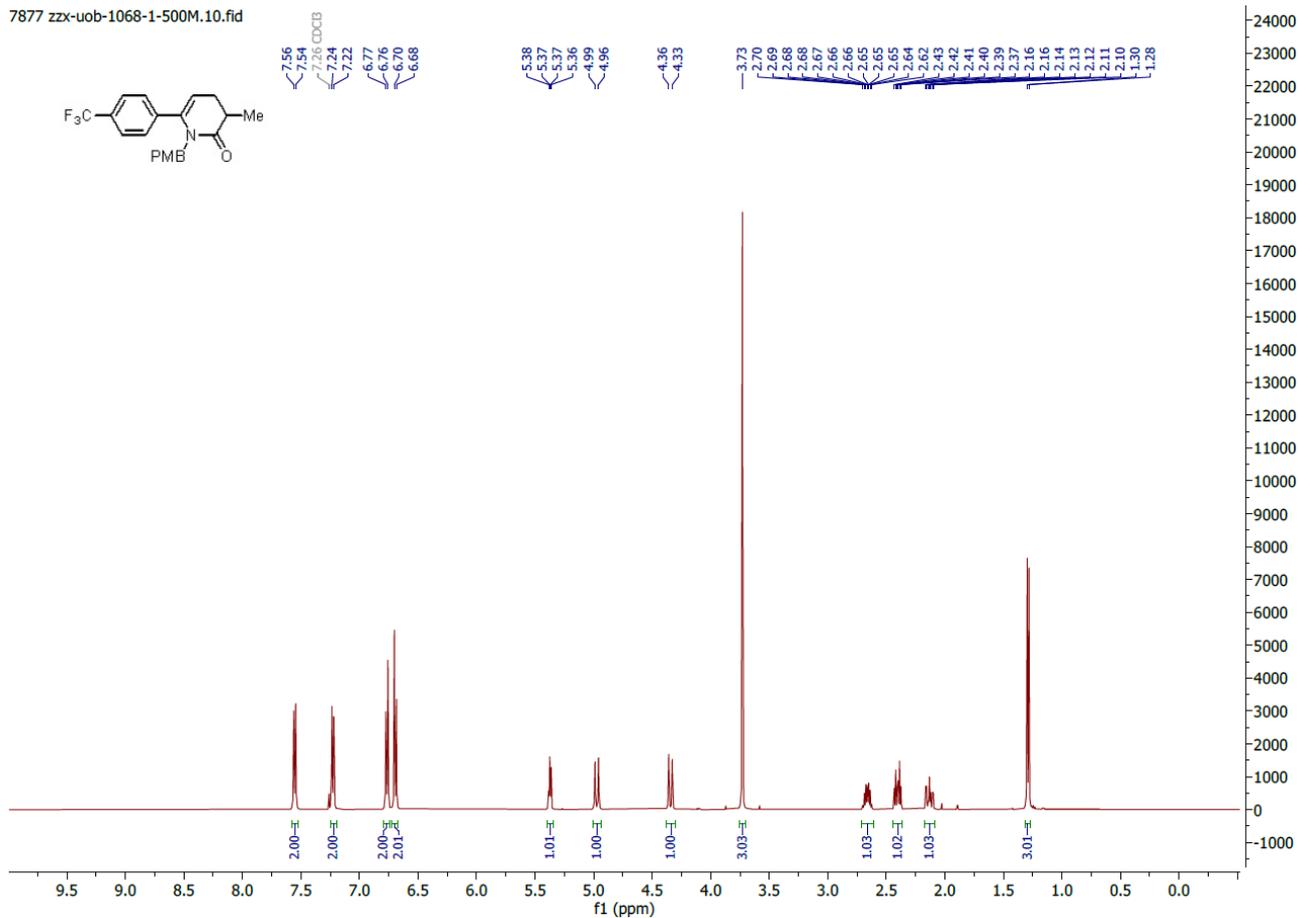
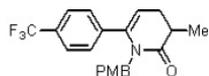
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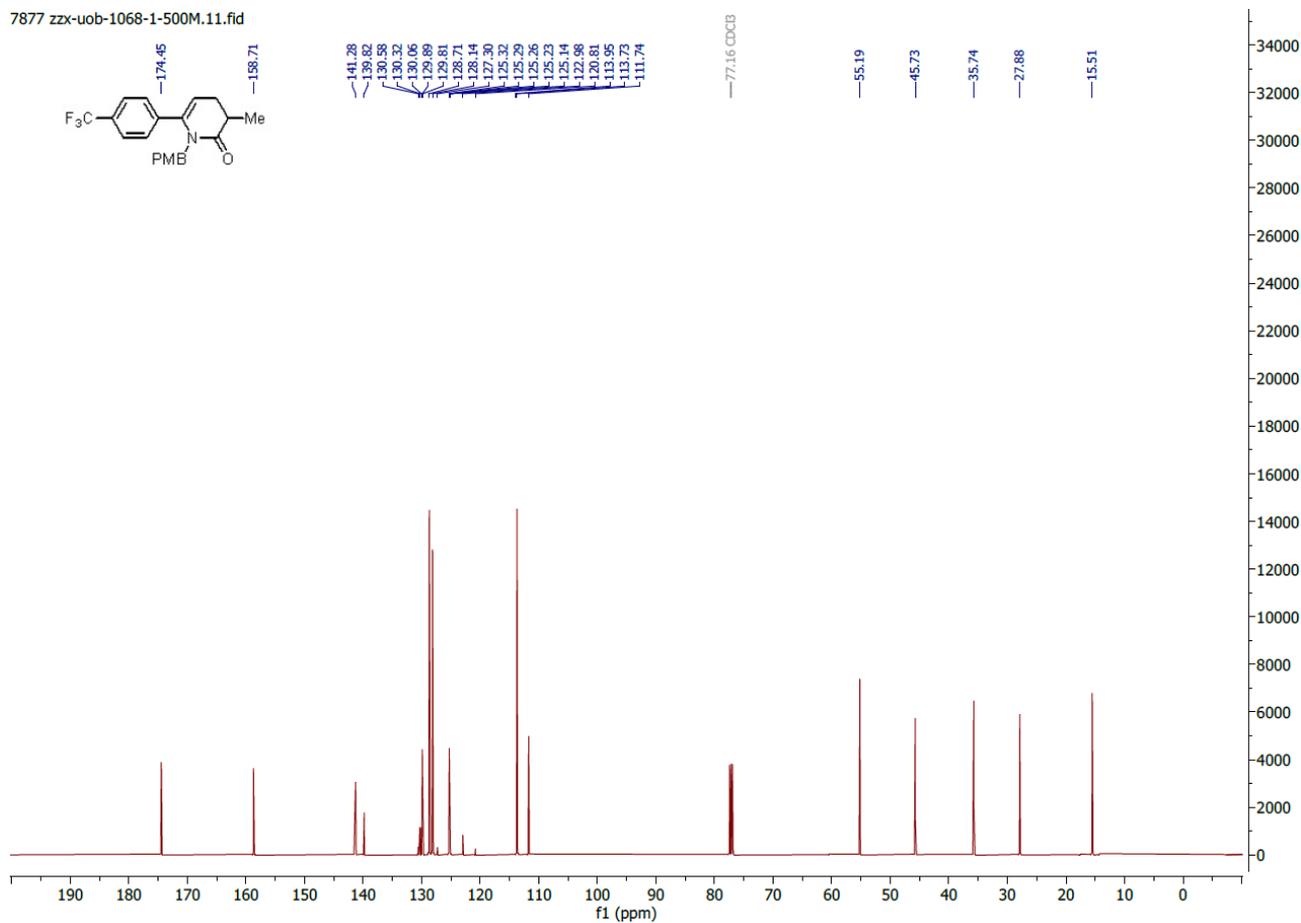
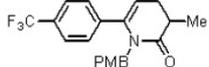


Compound 4a

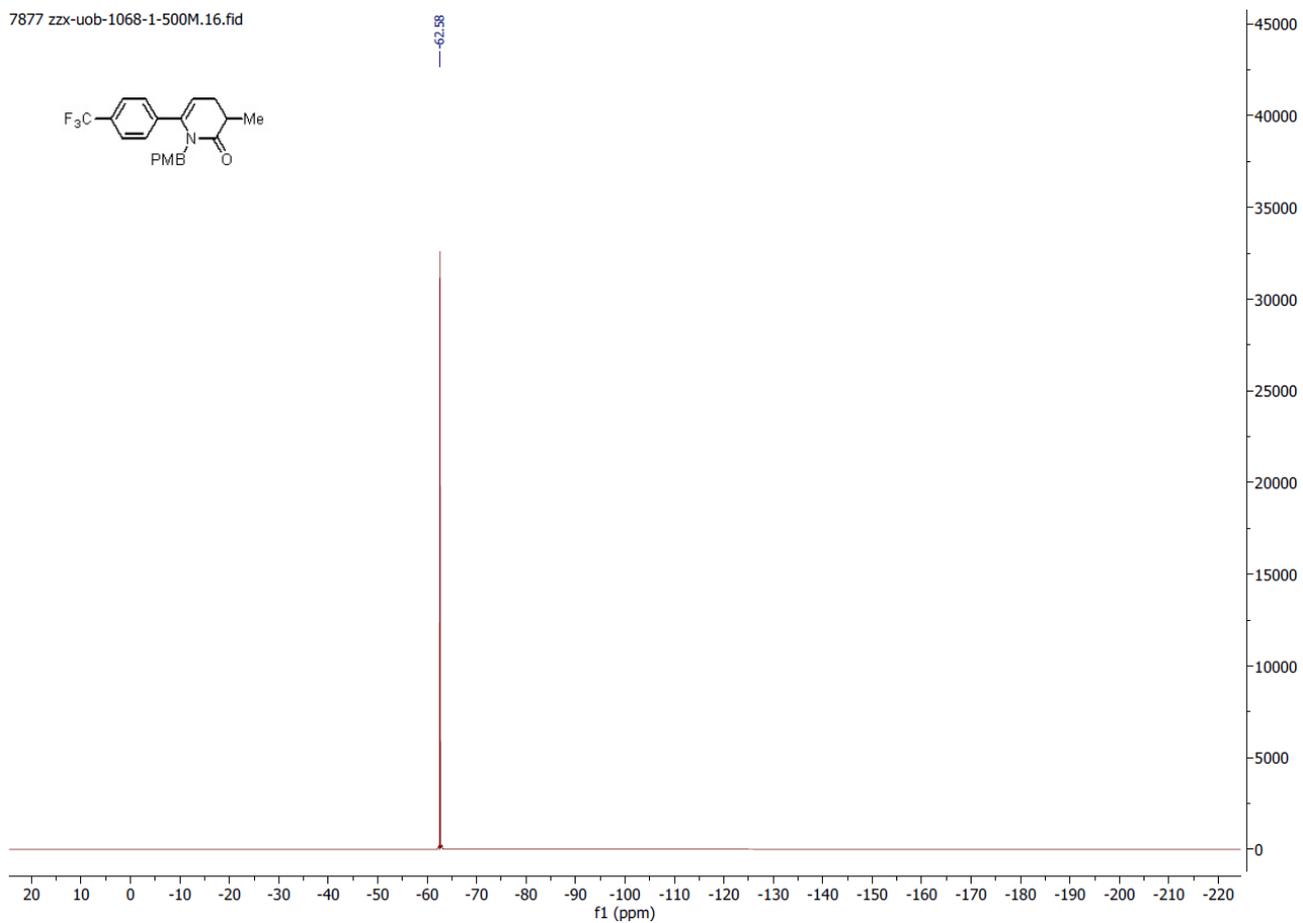
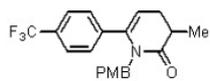
7877 zzx-uob-1068-1-500M.10.fid



7877 zzx-uob-1068-1-500M.11.fid

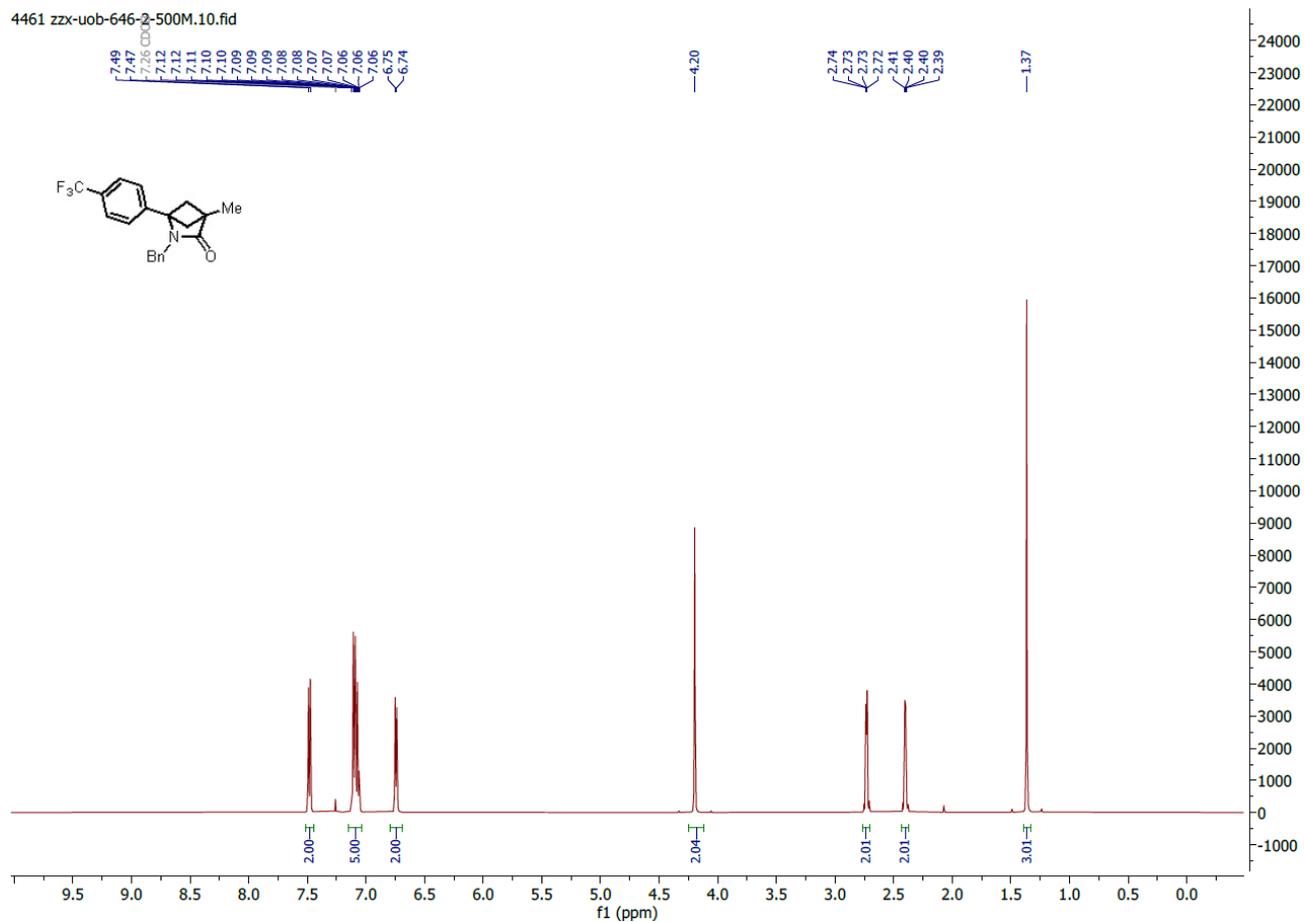


7877 zzx-uob-1068-1-500M.16.fid

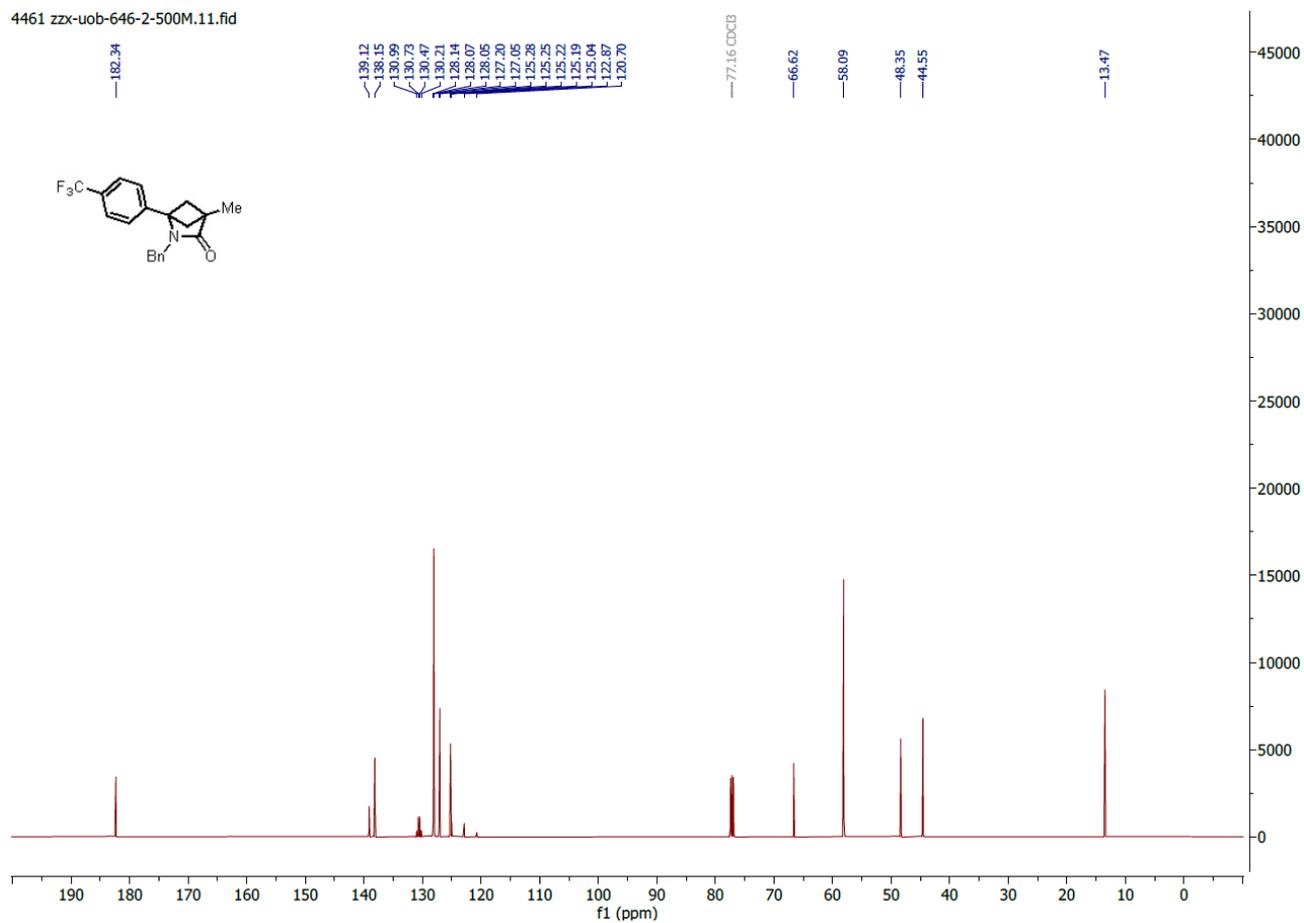


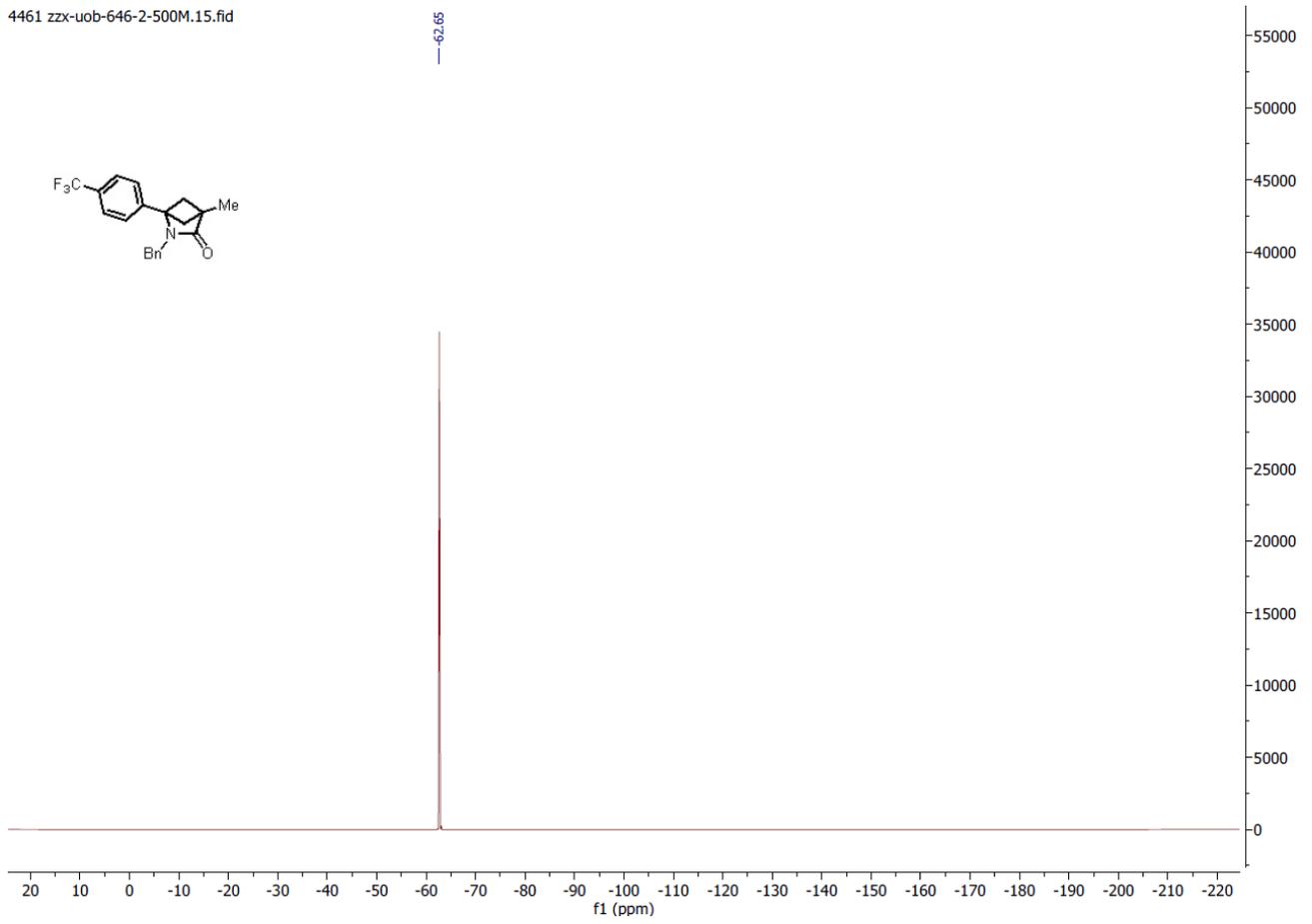
Compound 3b

4461 zzx-uob-646-2-500M.10.fid



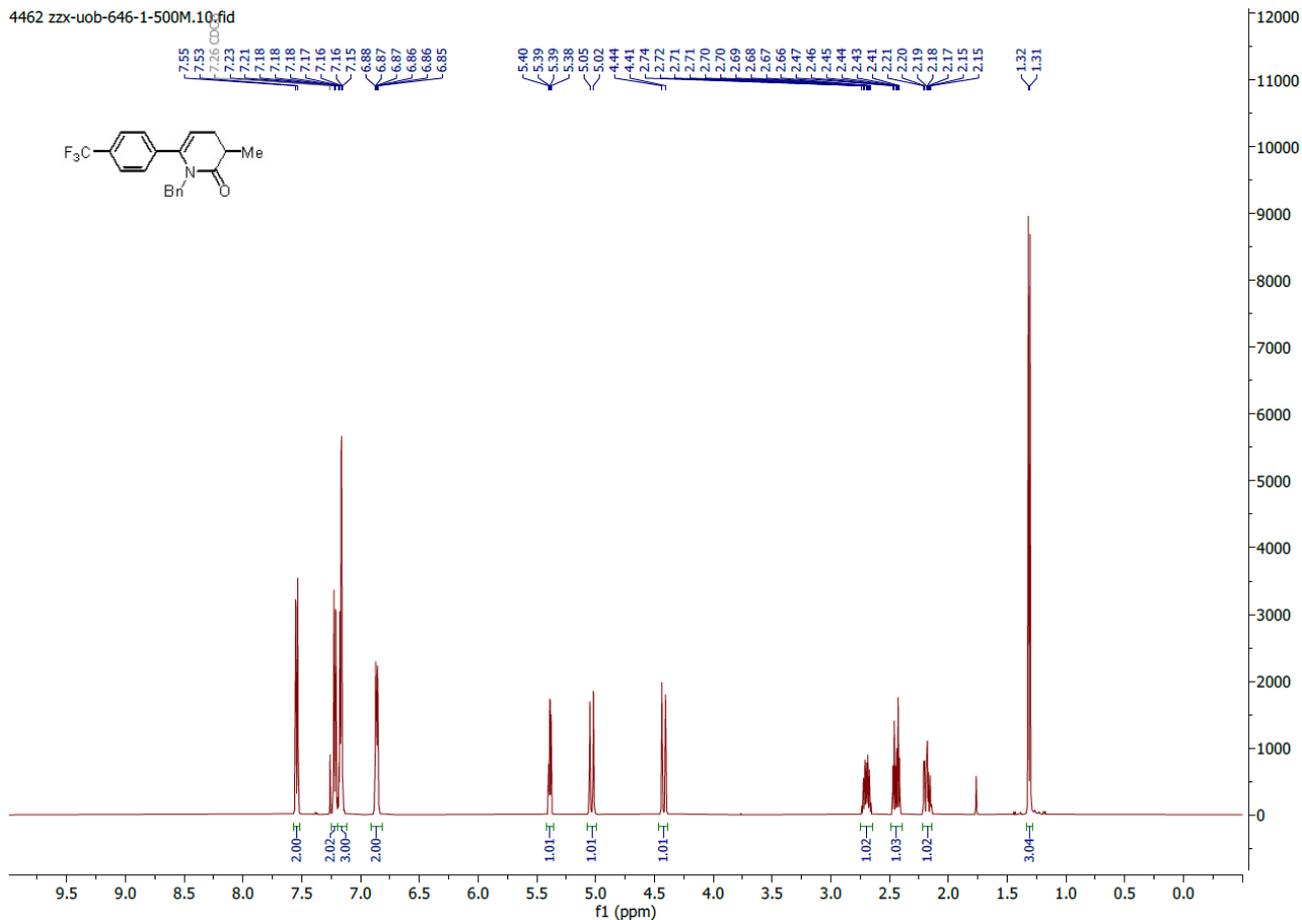
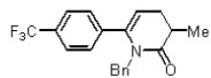
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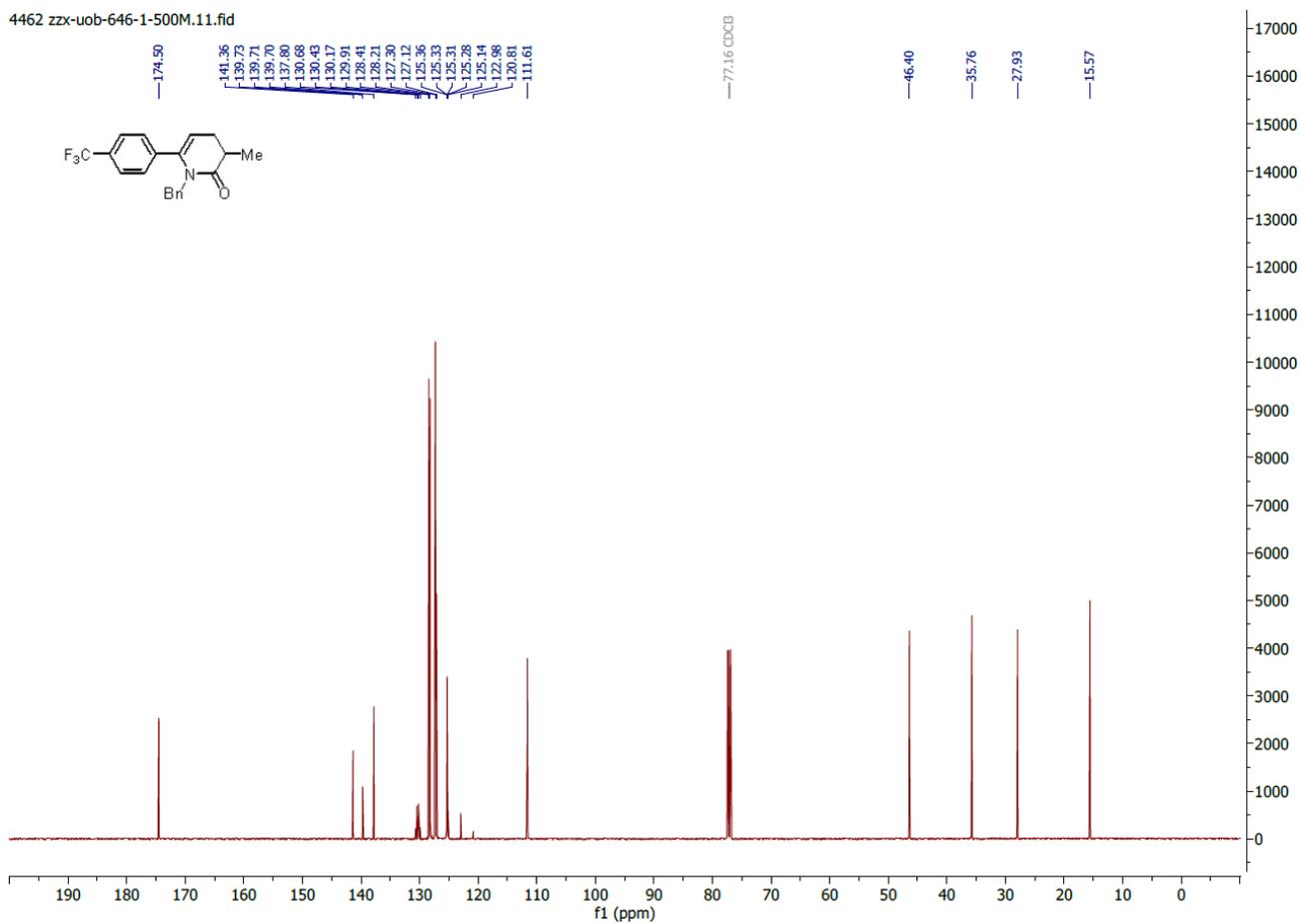
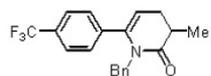


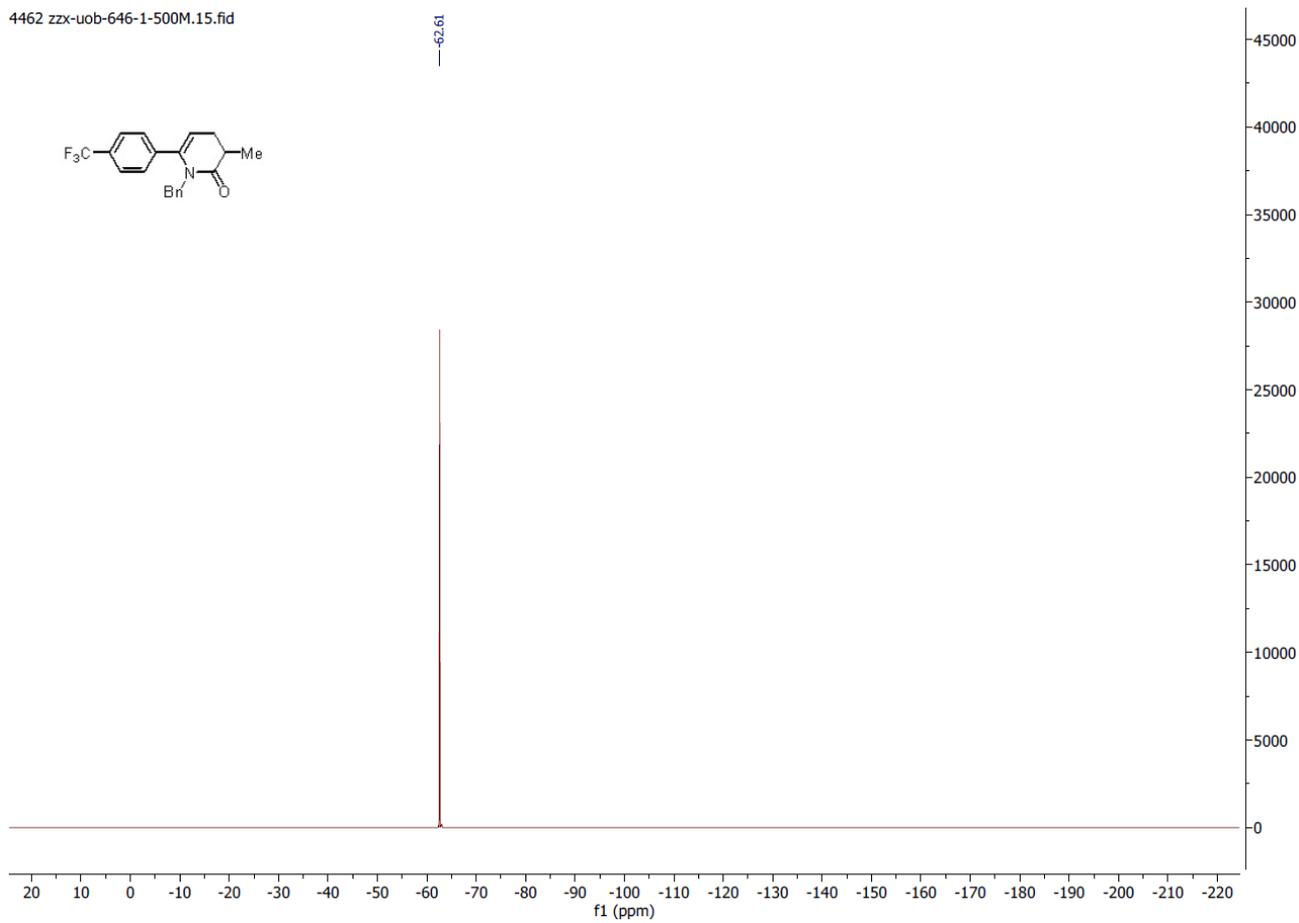
Compound 4b

4462 zzx-uob-646-1-500M.10.fid



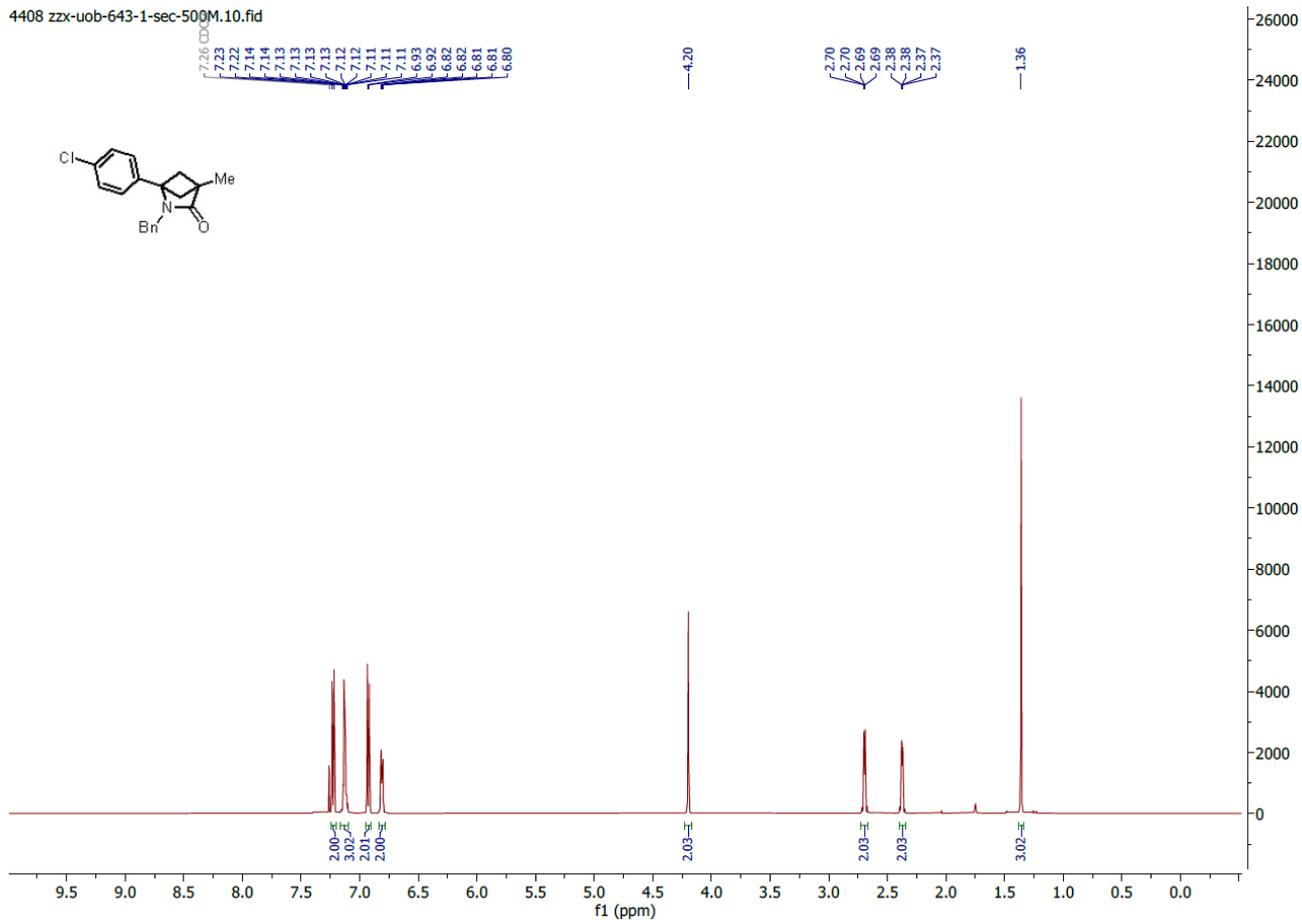
4462 zzx-uob-646-1-500M.11.fid



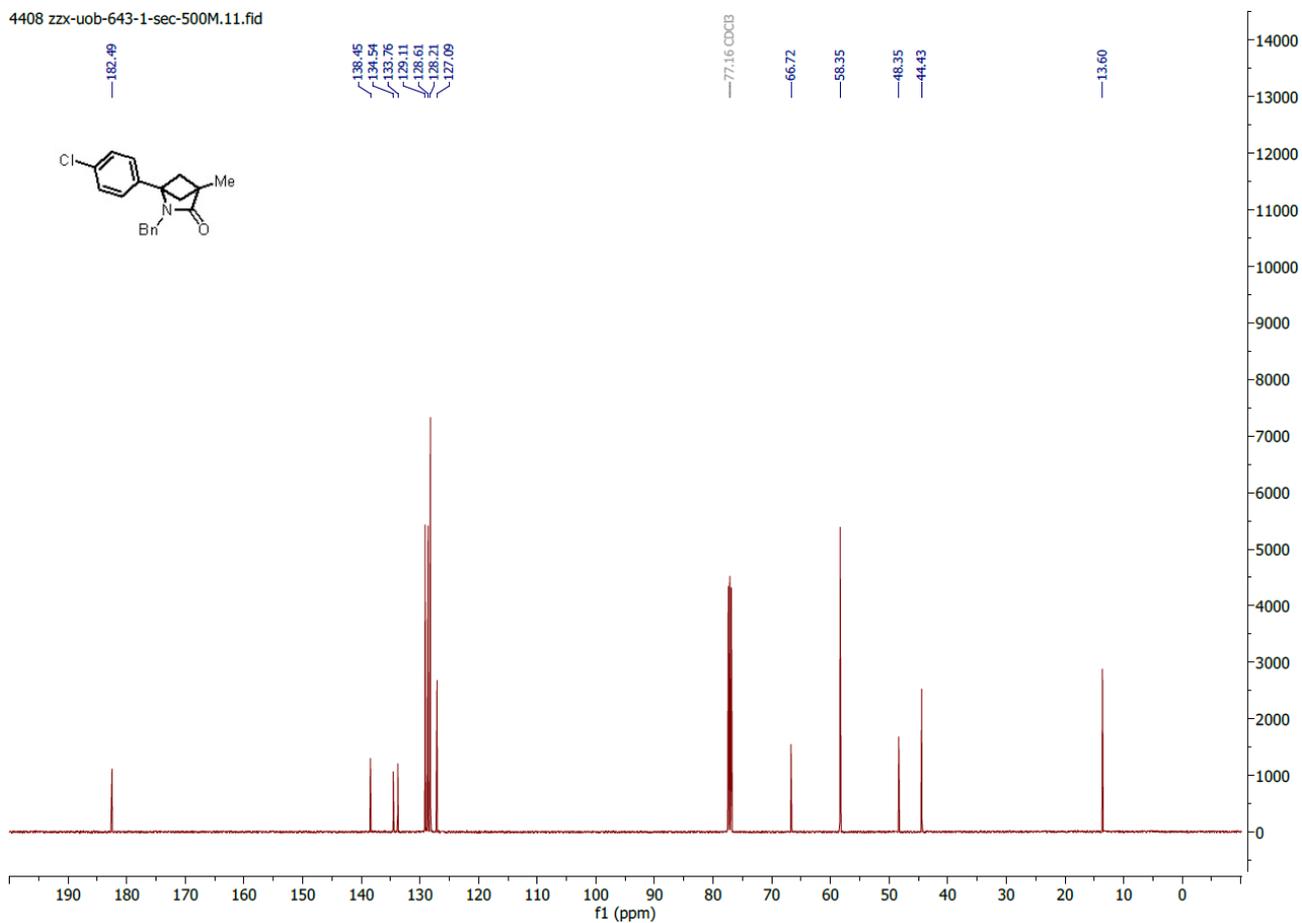


Compound 3c

4408 zzx-uob-643-1-sec-500M.10.fid

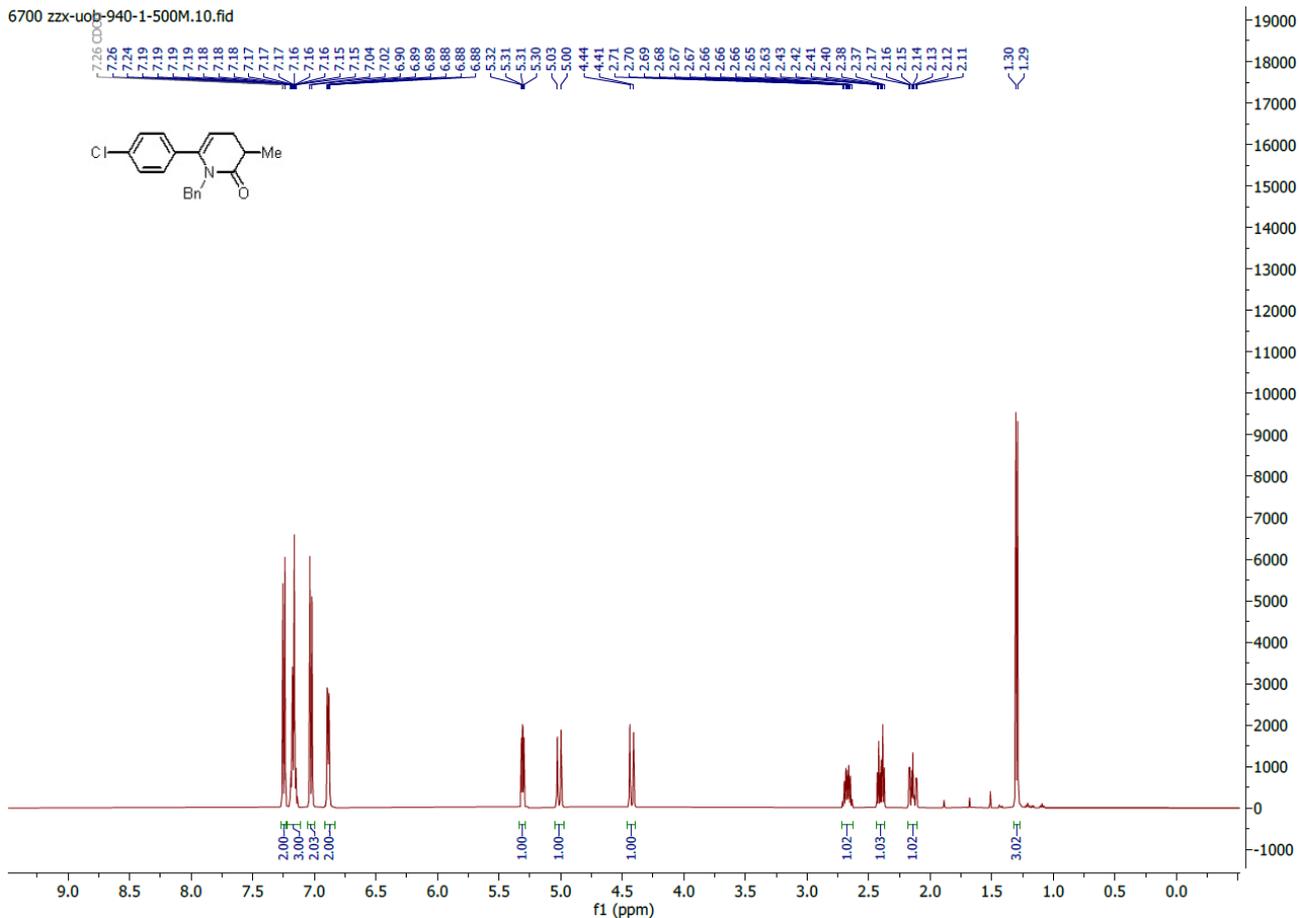


4408 zzx-uob-643-1-sec-500M.11.fid

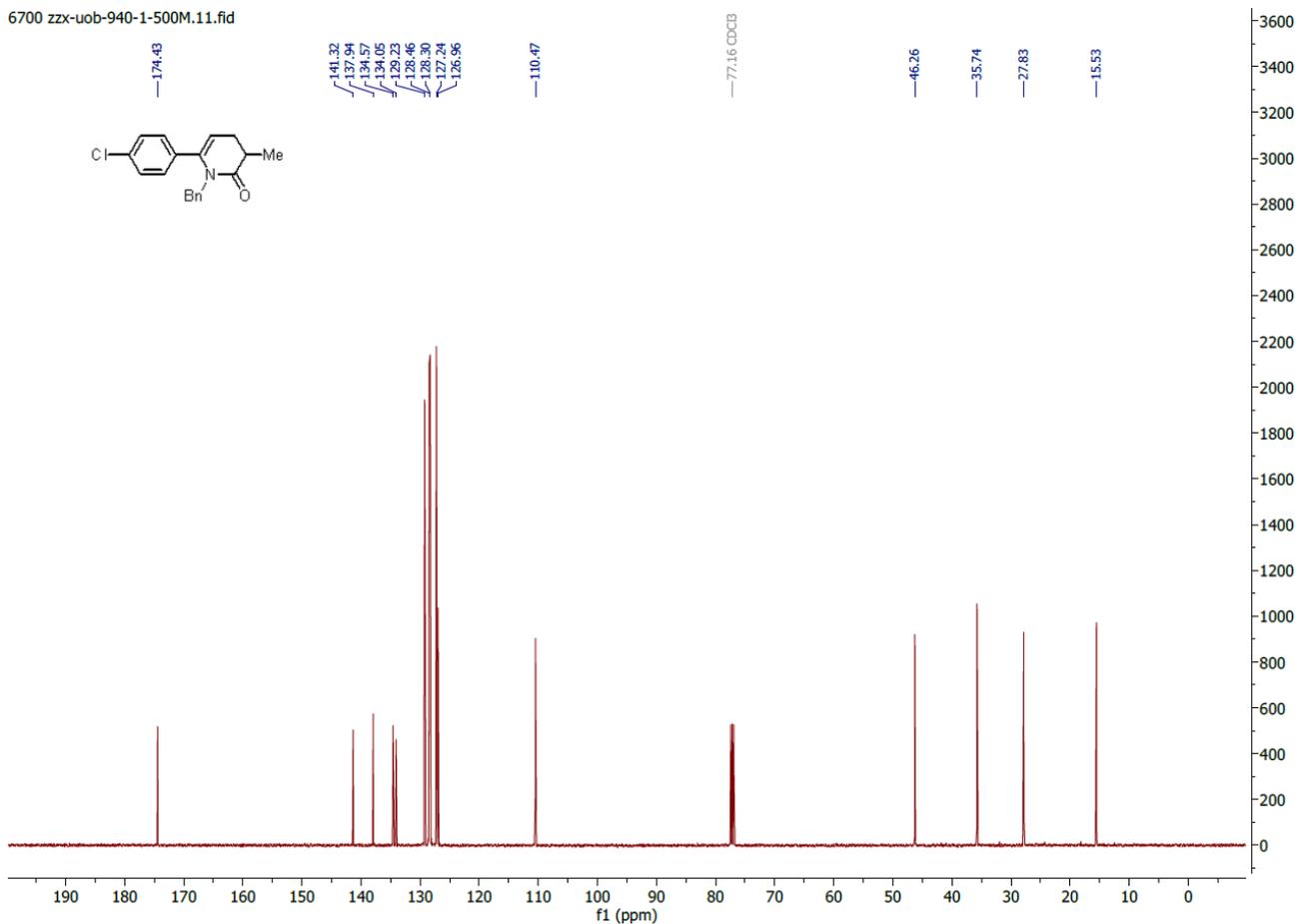


Compound 4c

6700 zzx-uob-940-1-500M.10.fid

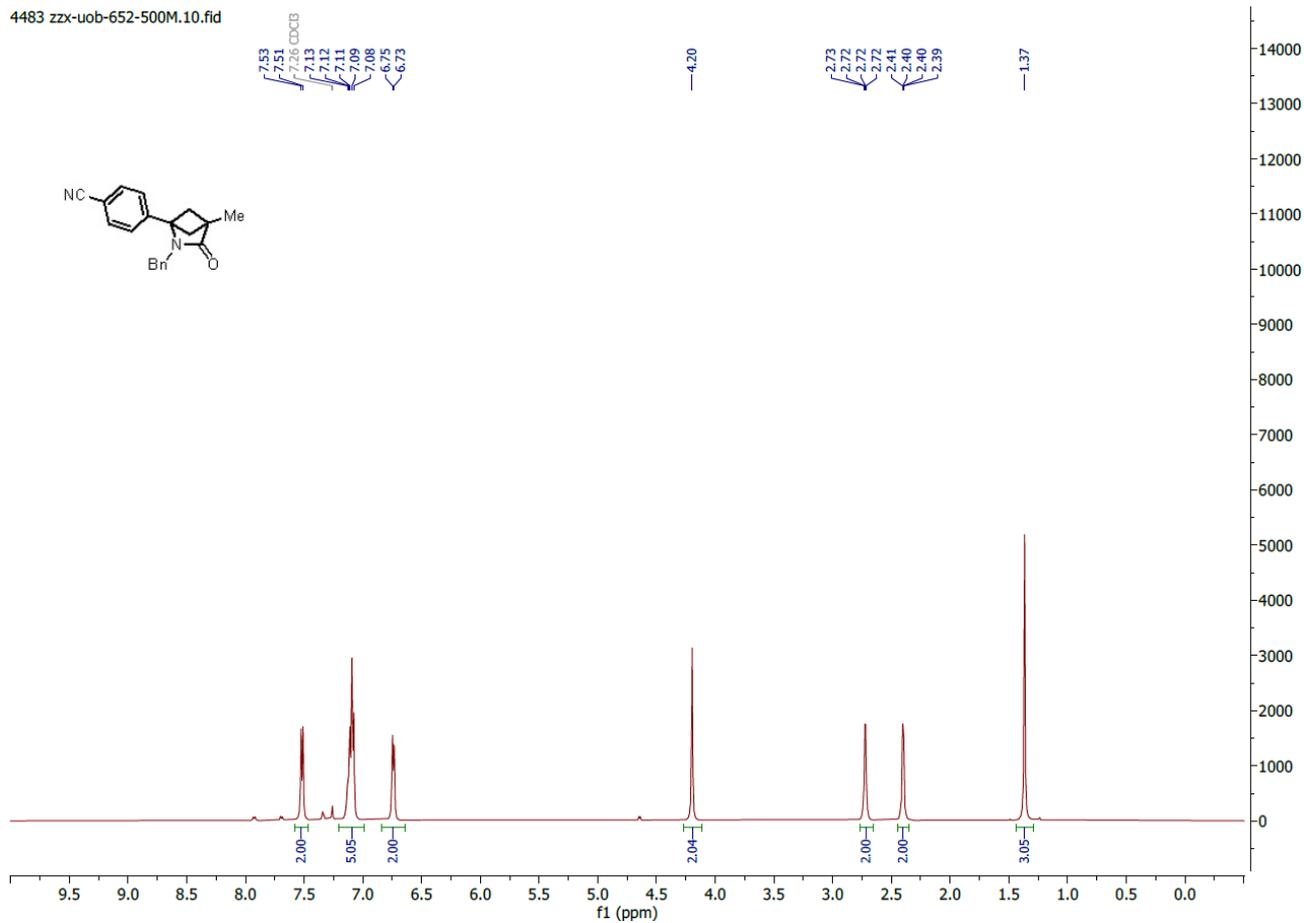


6700 zzx-uob-940-1-500M.11.fid

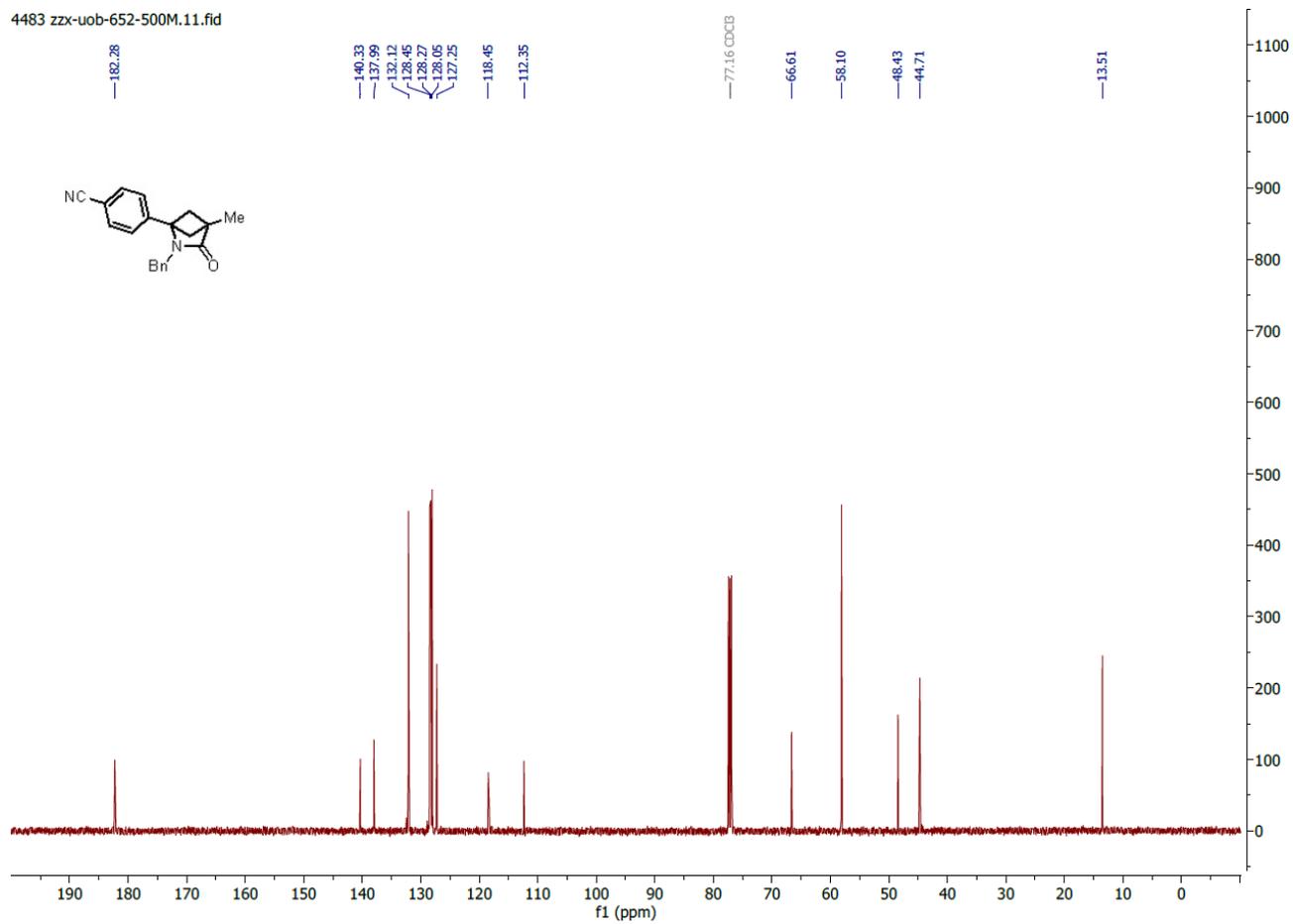


Compound 3d

4483 zzx-uob-652-500M.10.fid

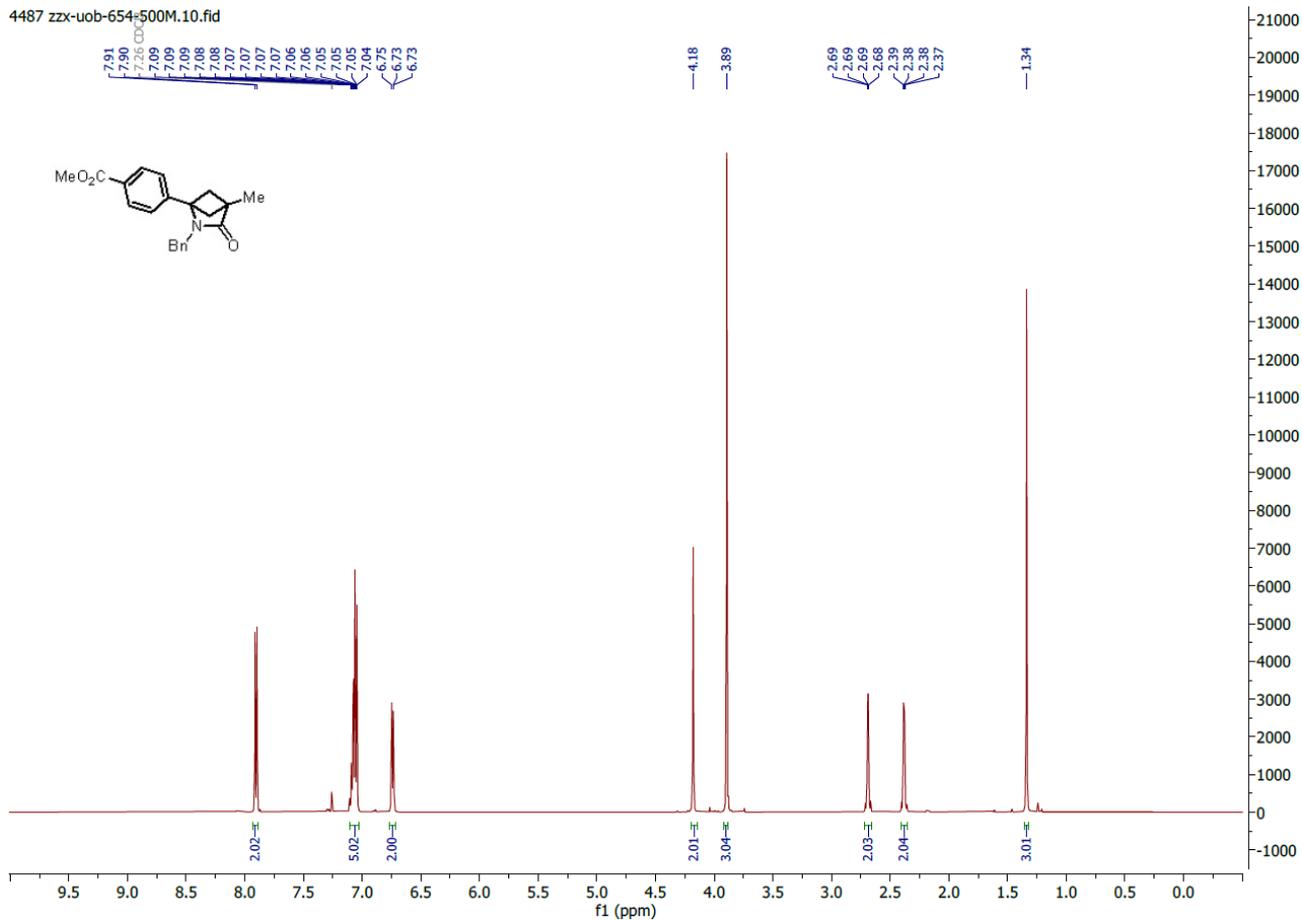


4483 zzx-uob-652-500M.11.fid

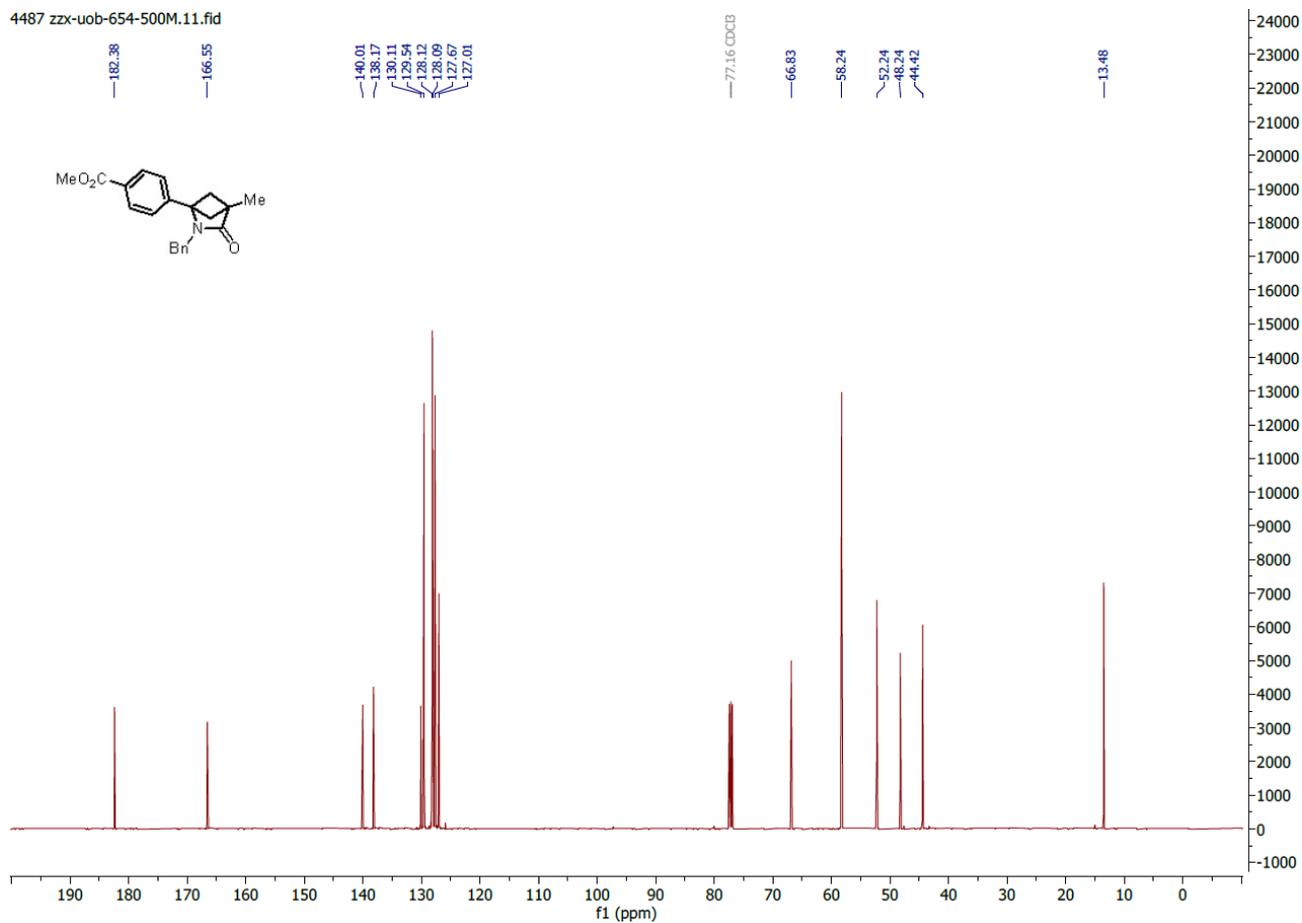


Compound 3e

4487 zzx-uob-654-500M.10.fid

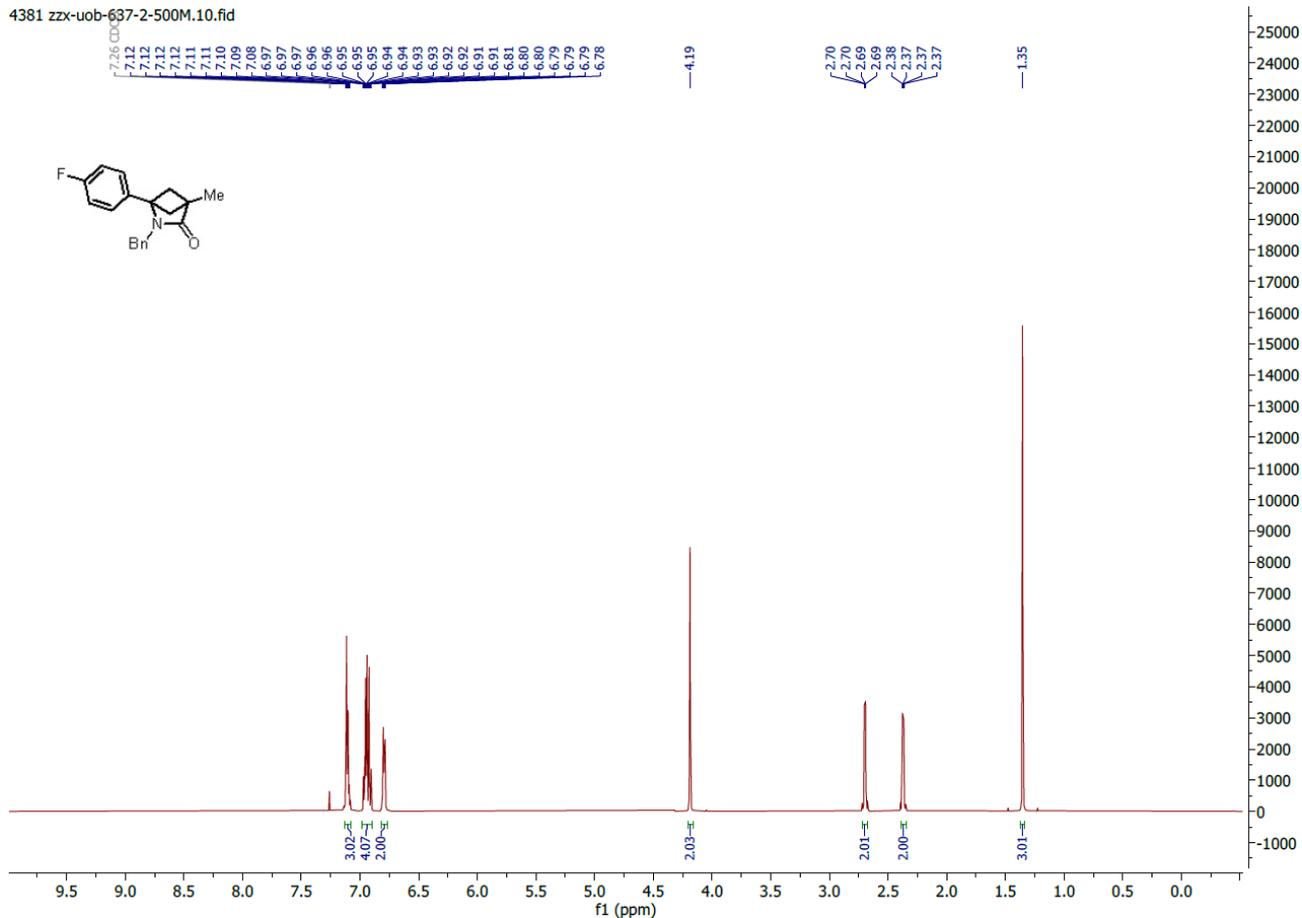


4487 zzx-uob-654-500M.11.fid

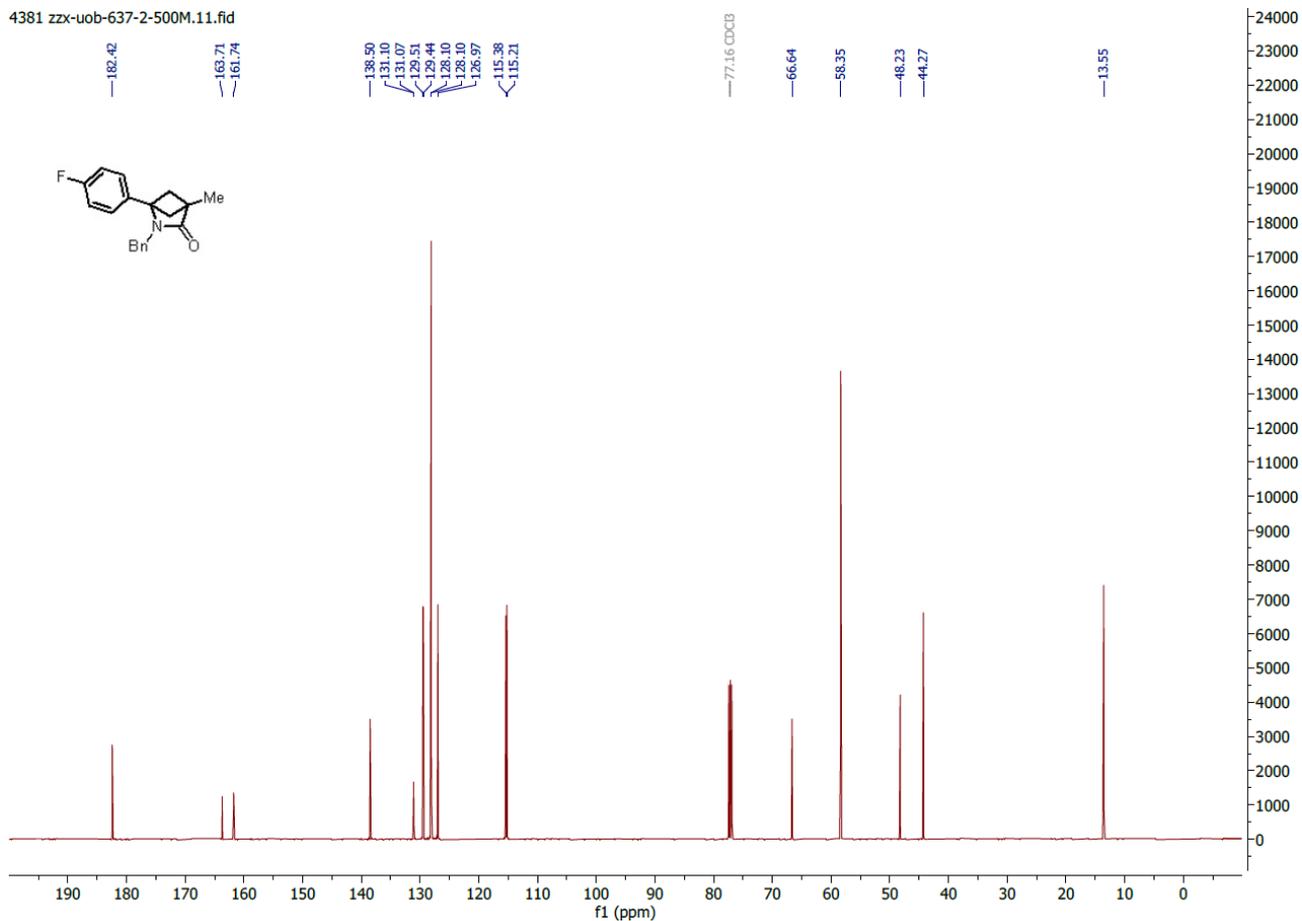


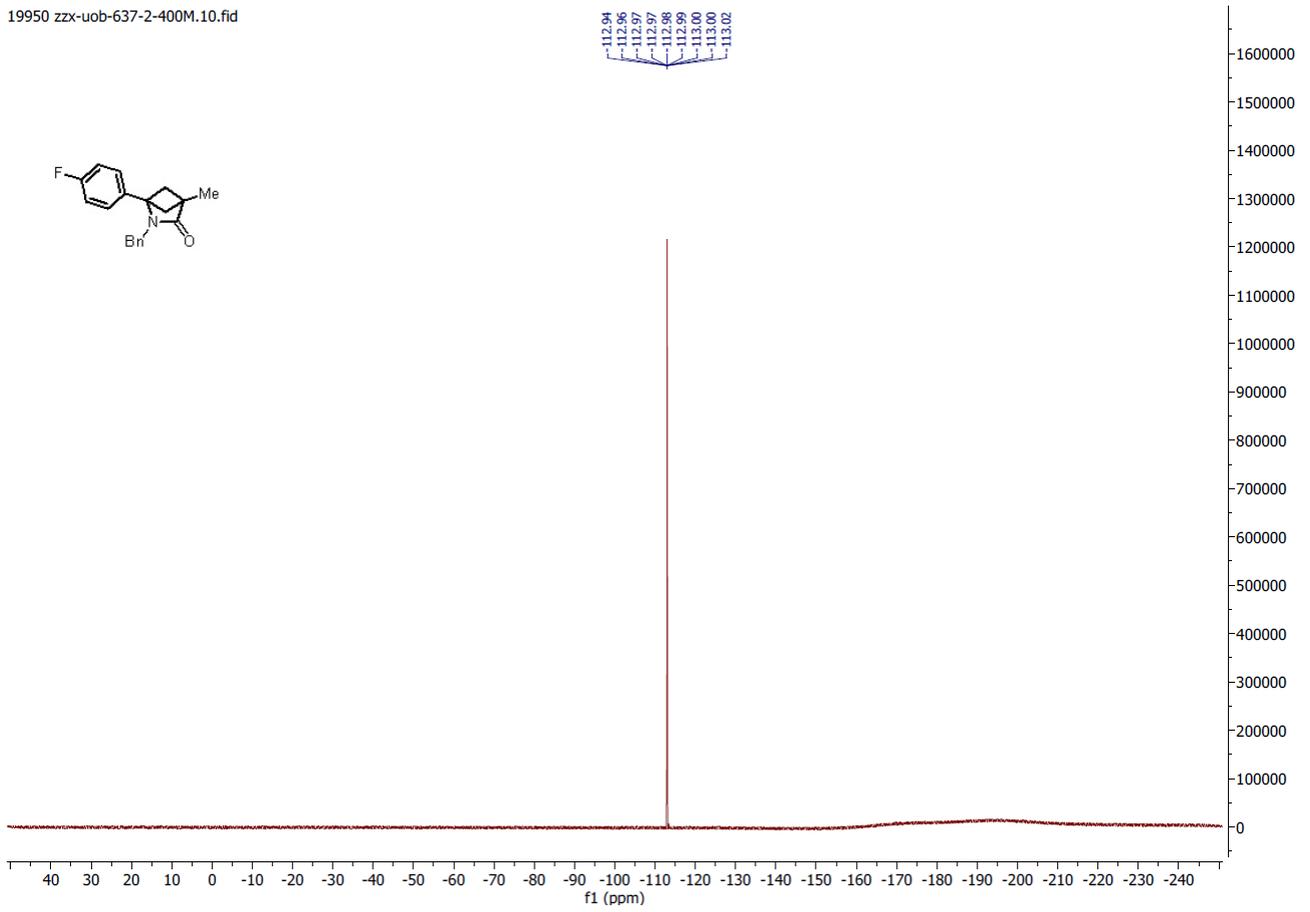
Compound 3f

4381 zzx-uob-637-2-500M.10.fid



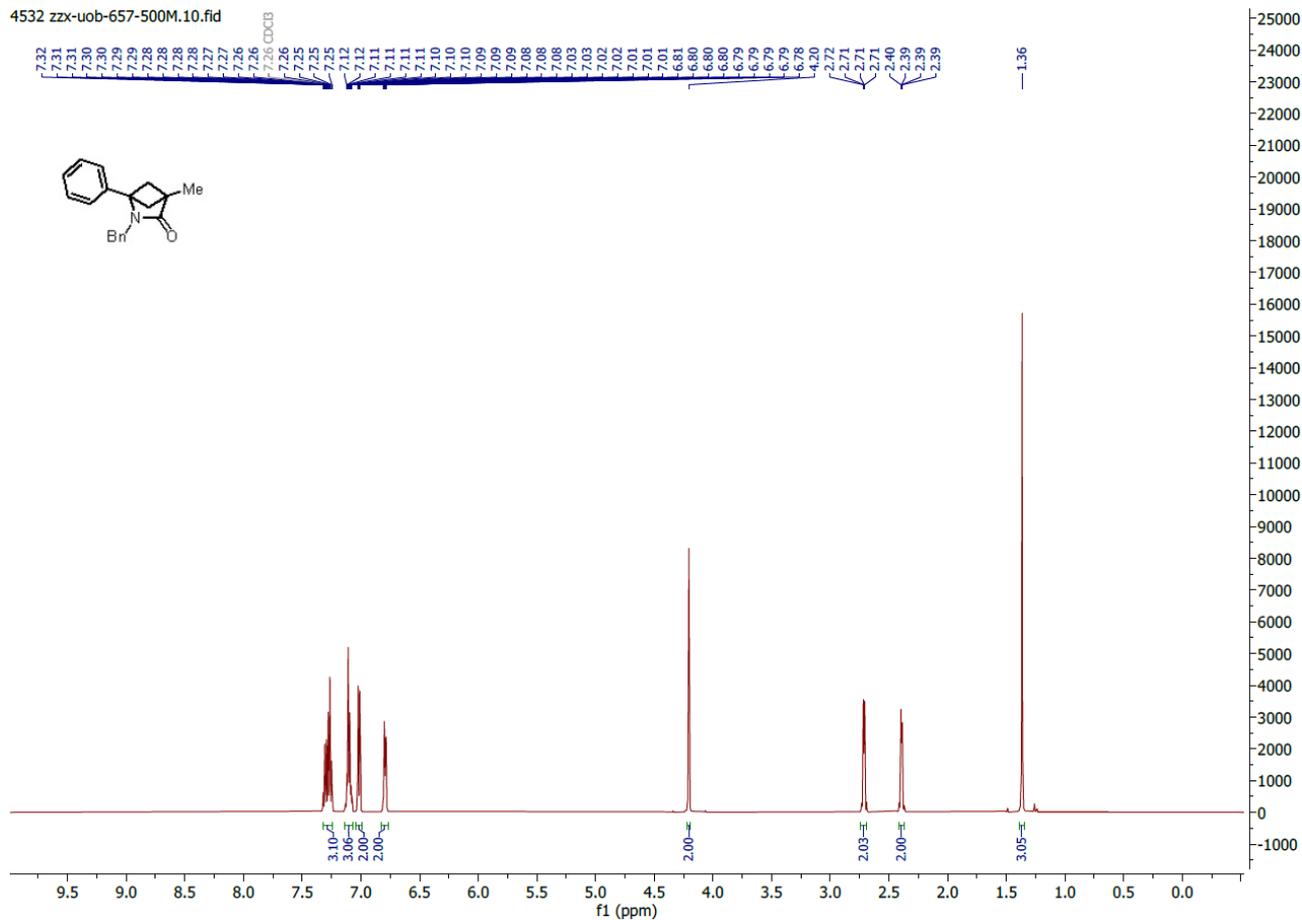
4381 zzx-uob-637-2-500M.11.fid



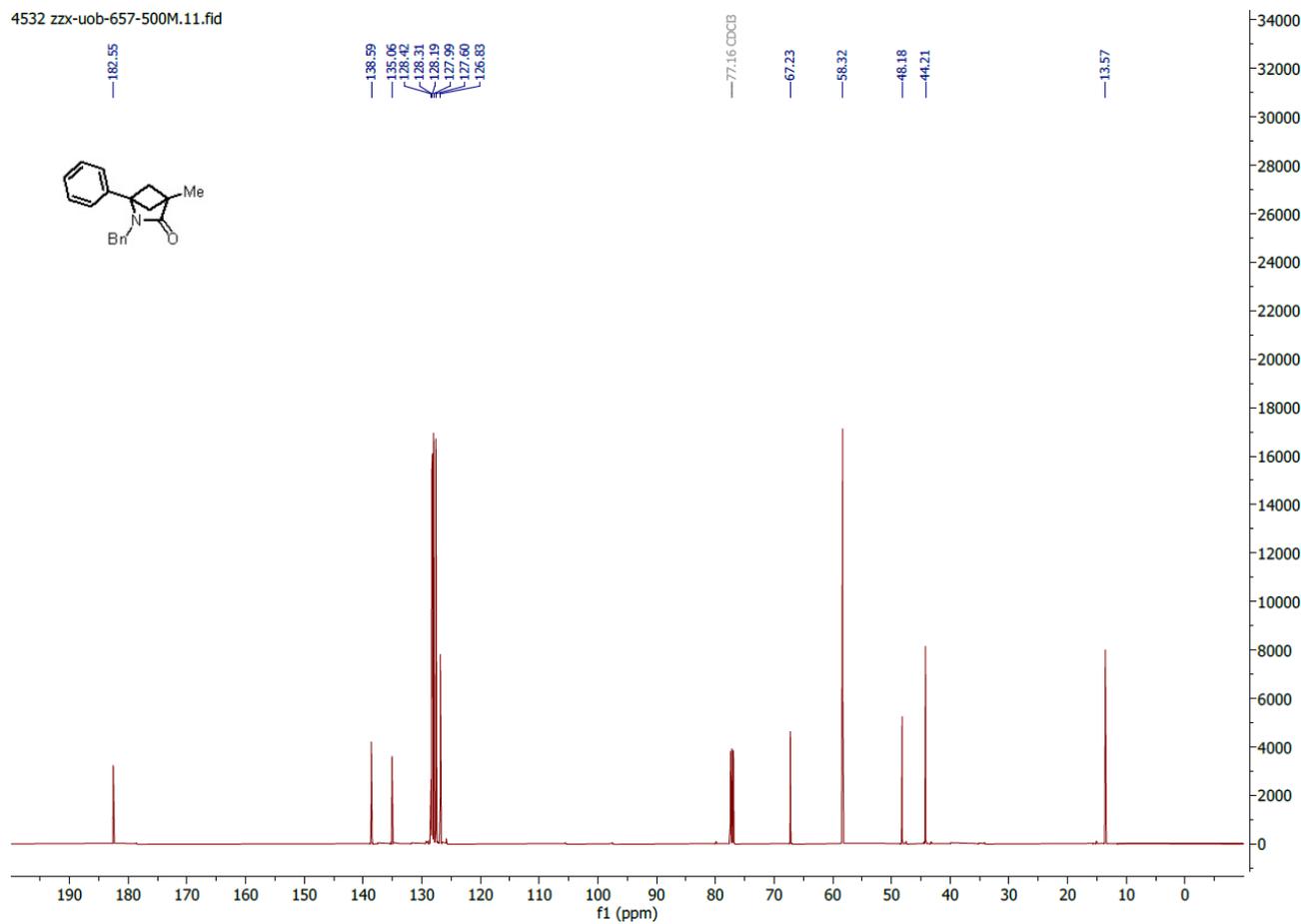


Compound 3g

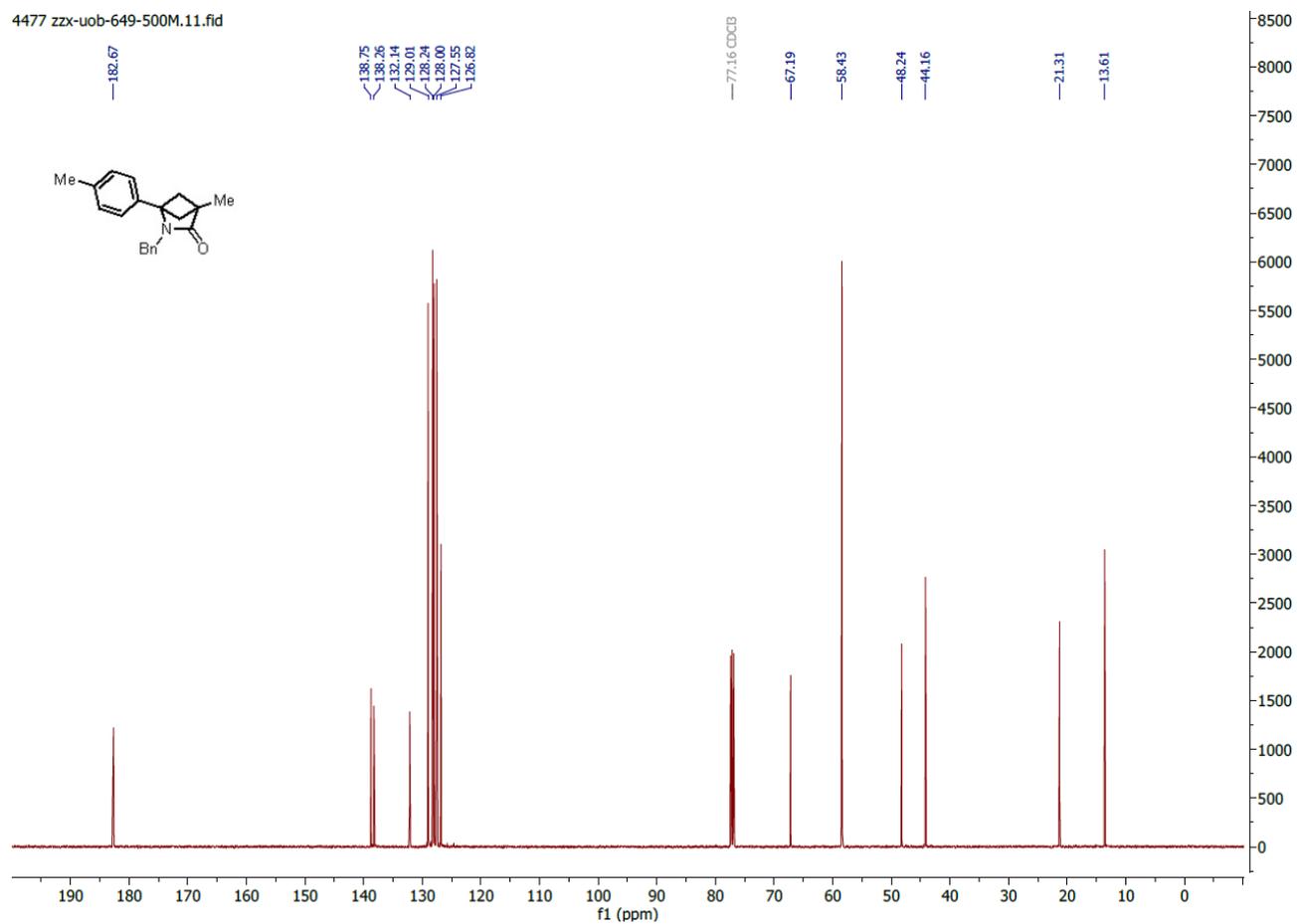
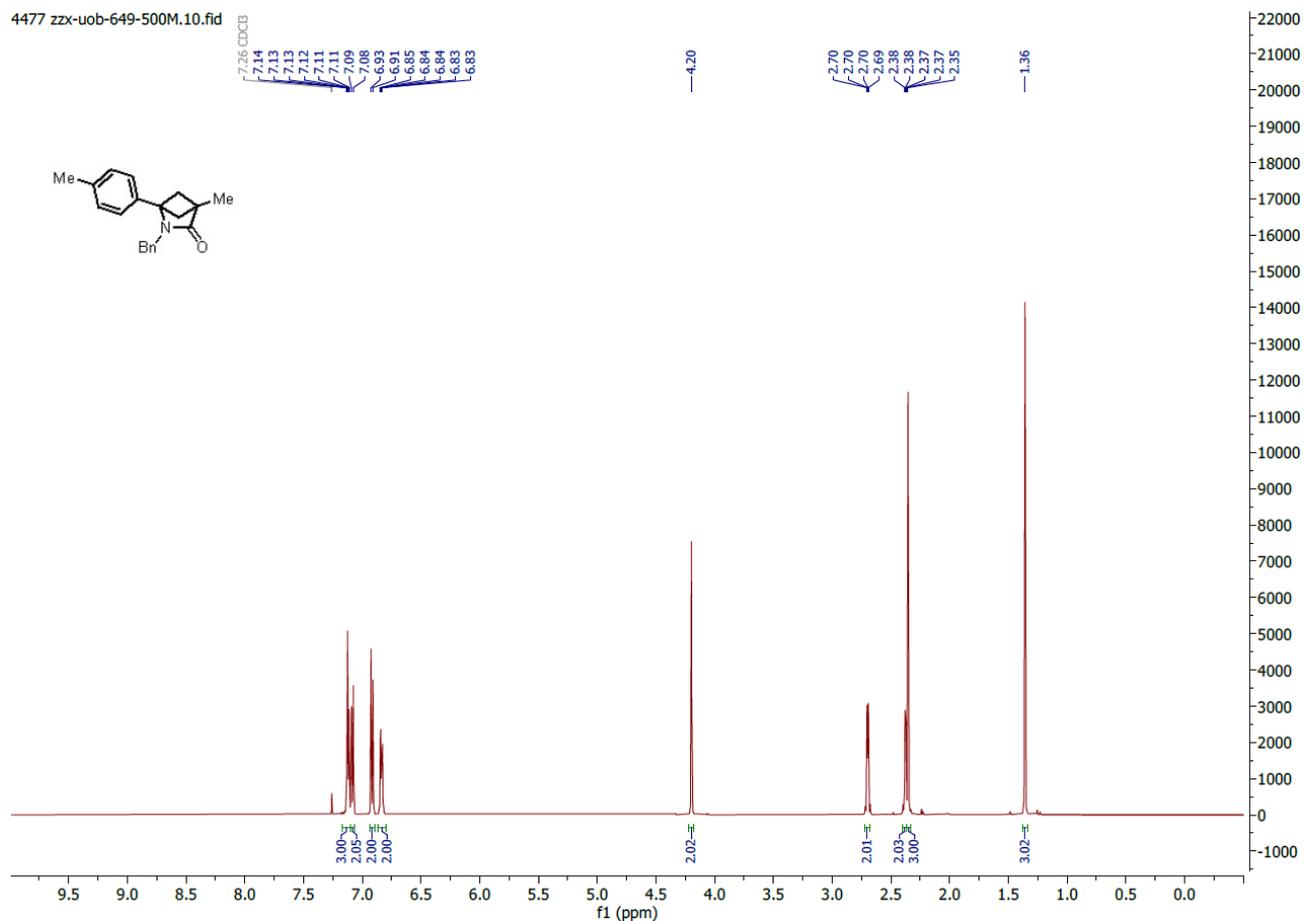
4532 zzx-uob-657-500M.10.fid



4532 zzx-uob-657-500M.11.fid

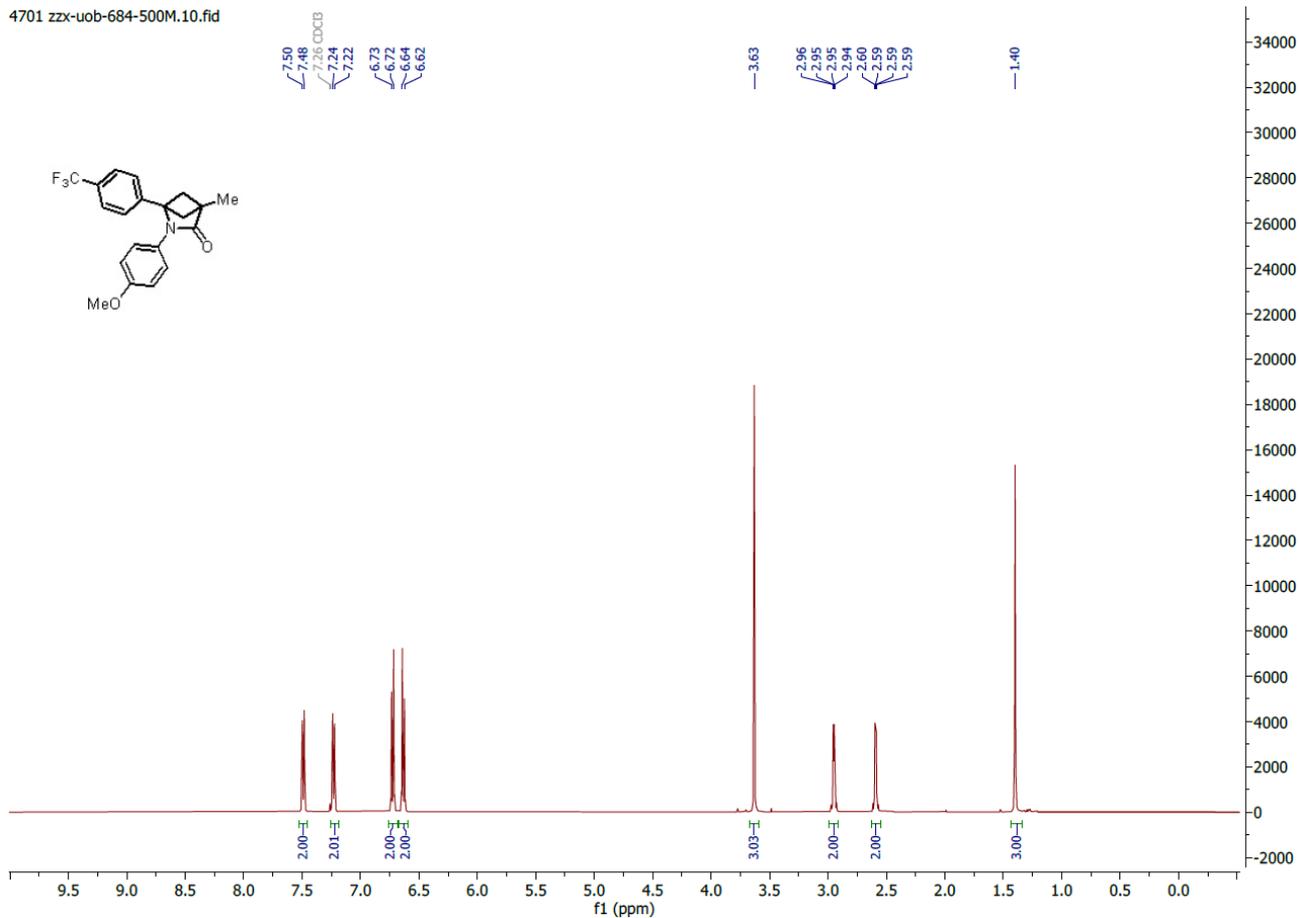


Compound 3h

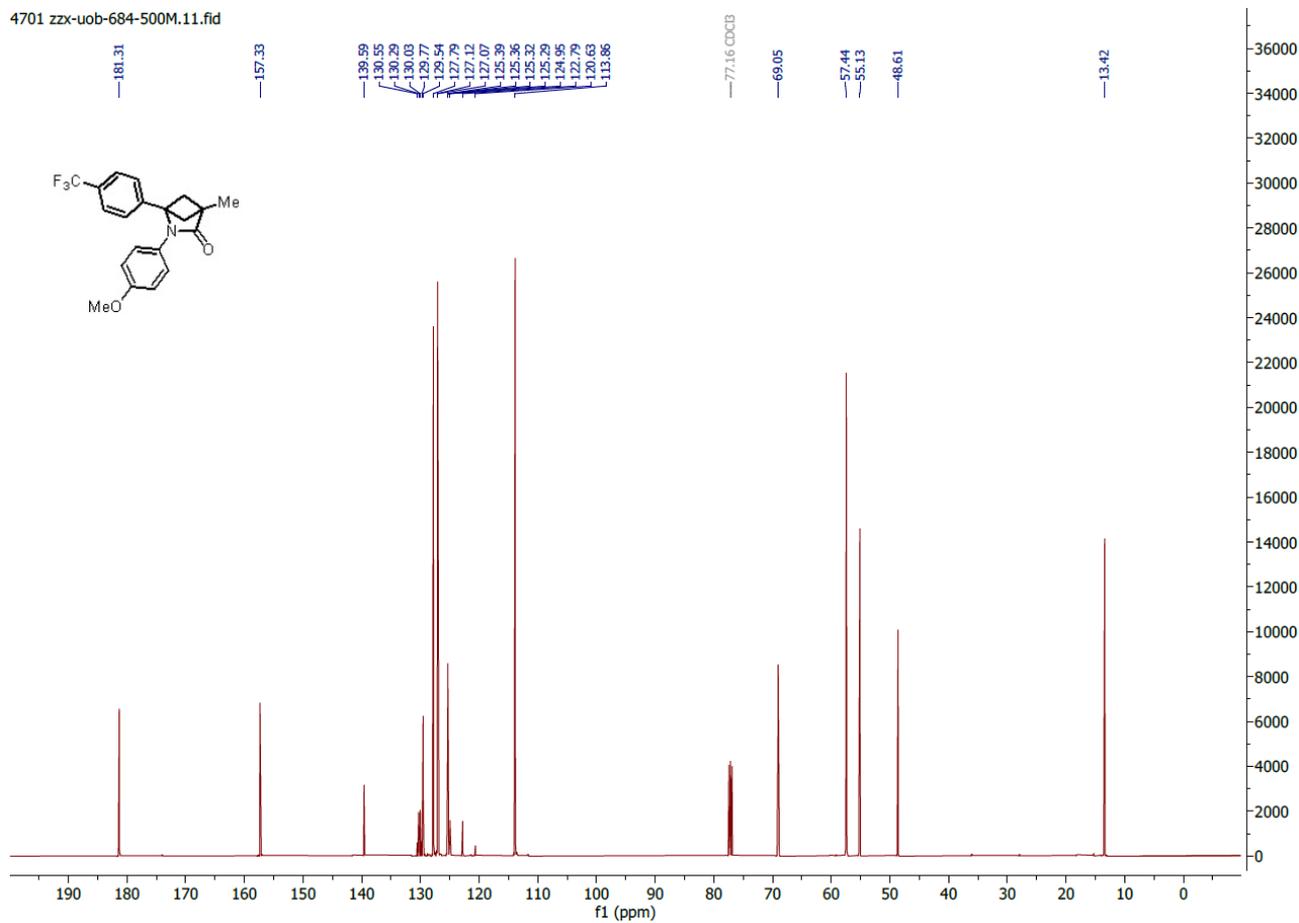


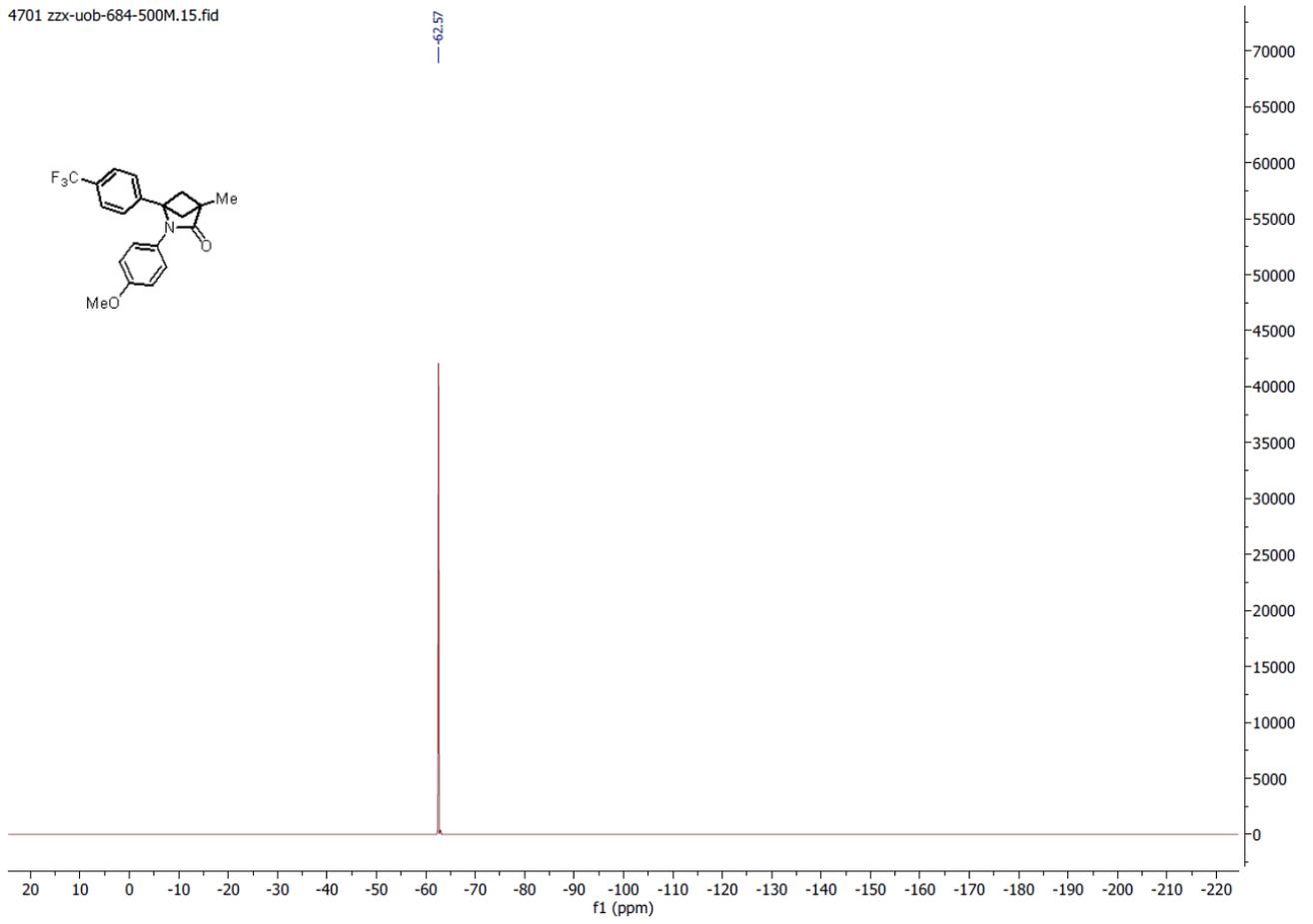
Compound 3j

4701 zzx-uob-684-500M.10.fid



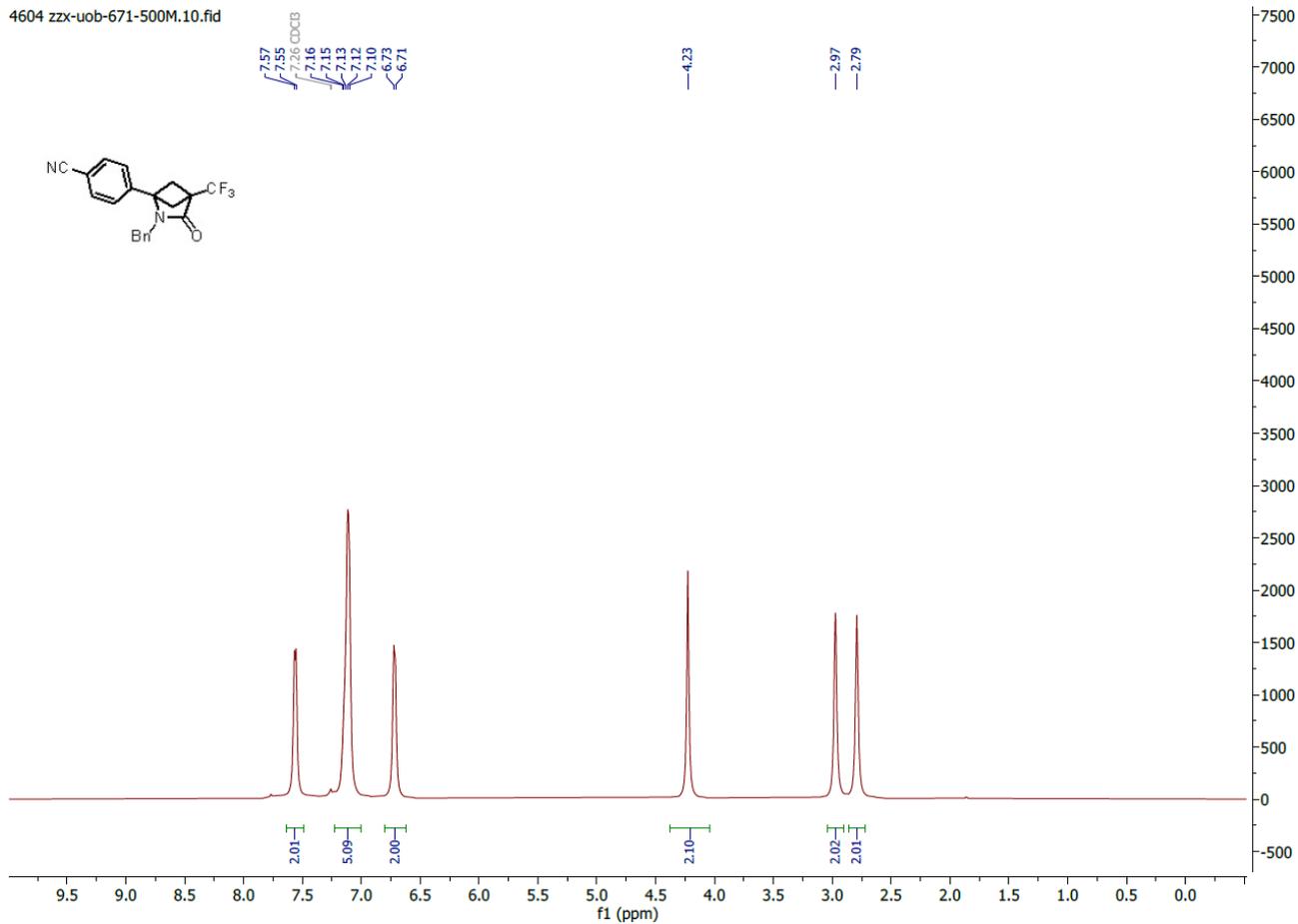
4701 zzx-uob-684-500M.11.fid



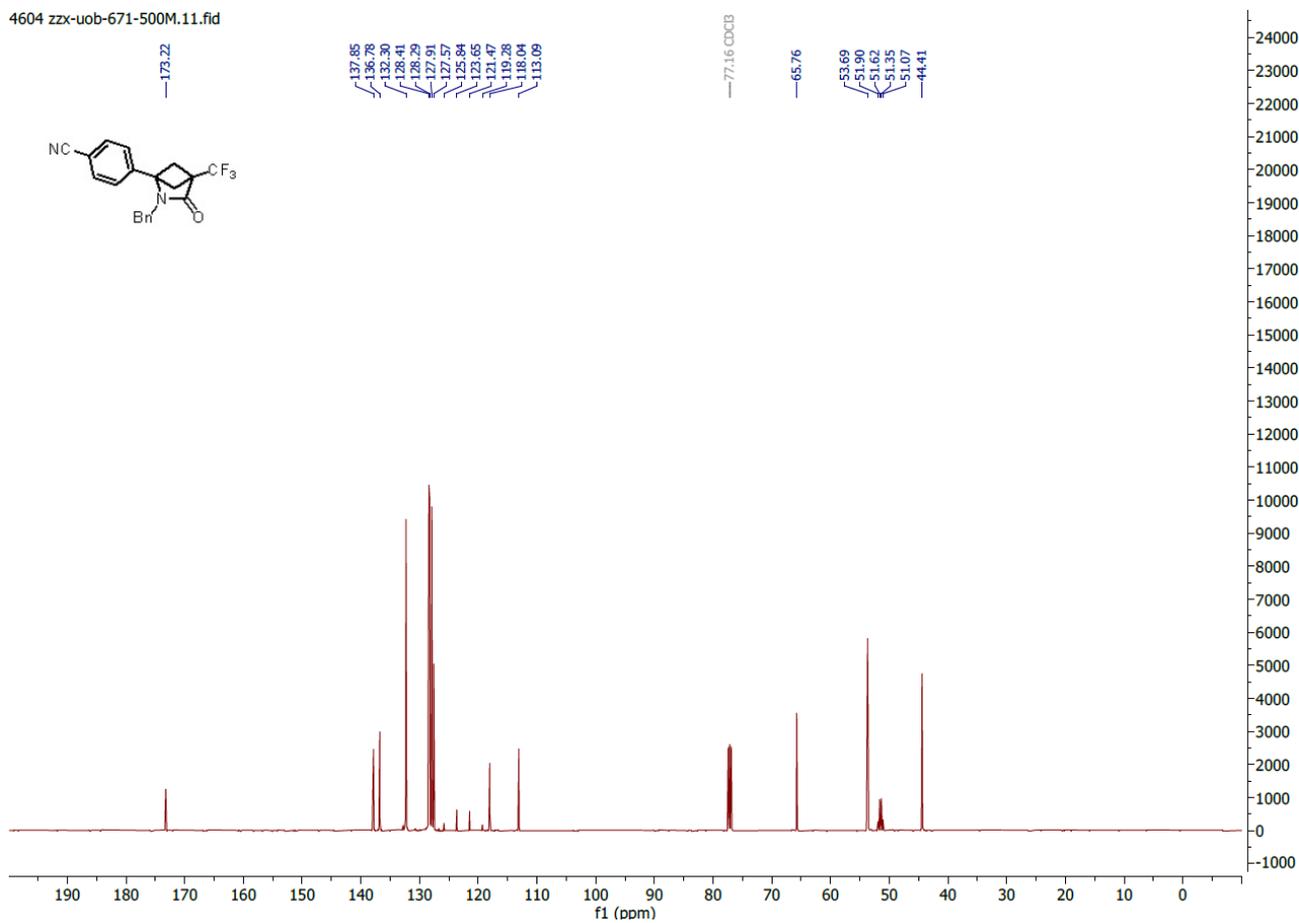


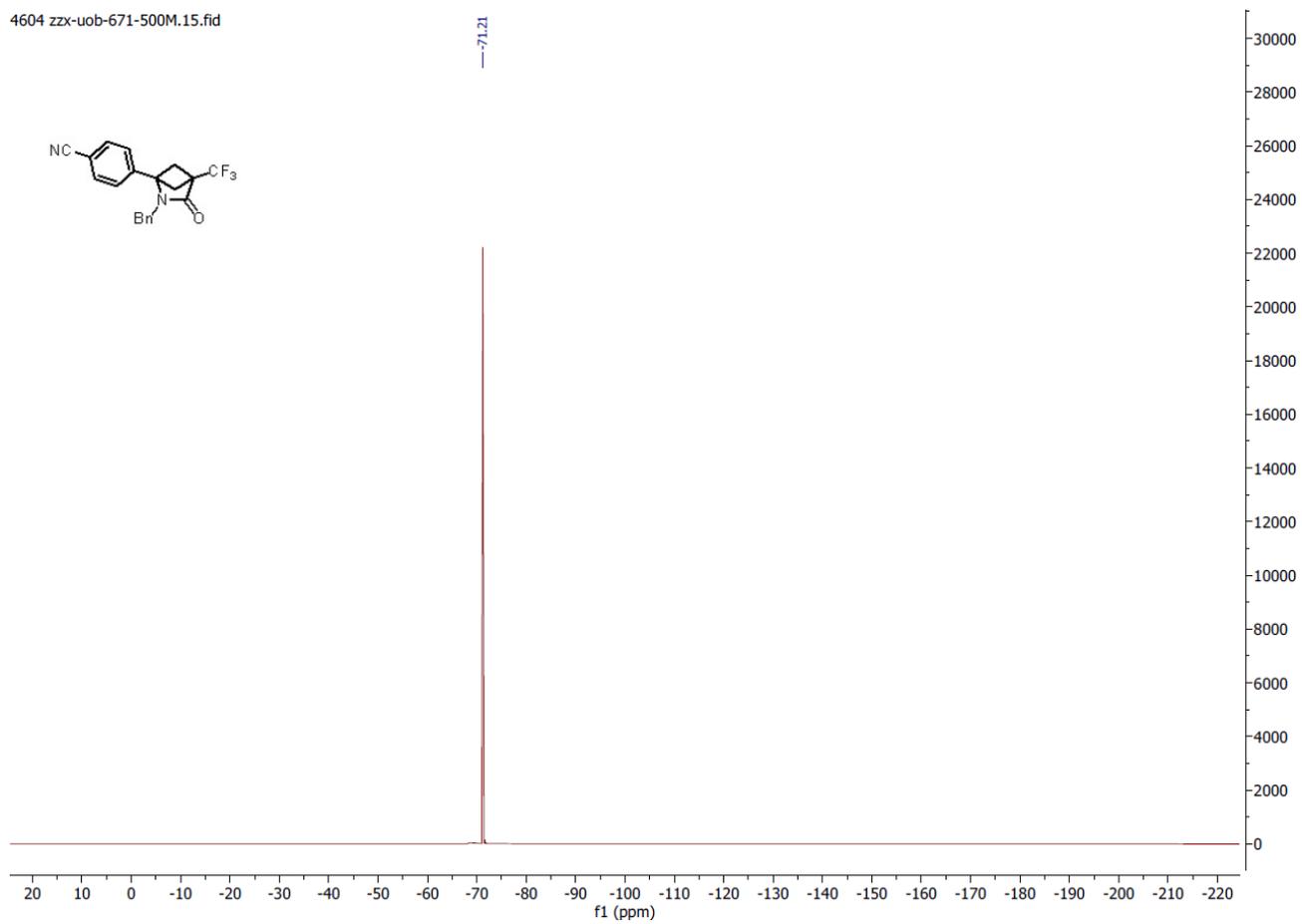
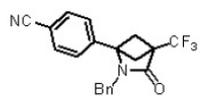
Compound 3k

4604 zzx-uob-671-500M.10.fid



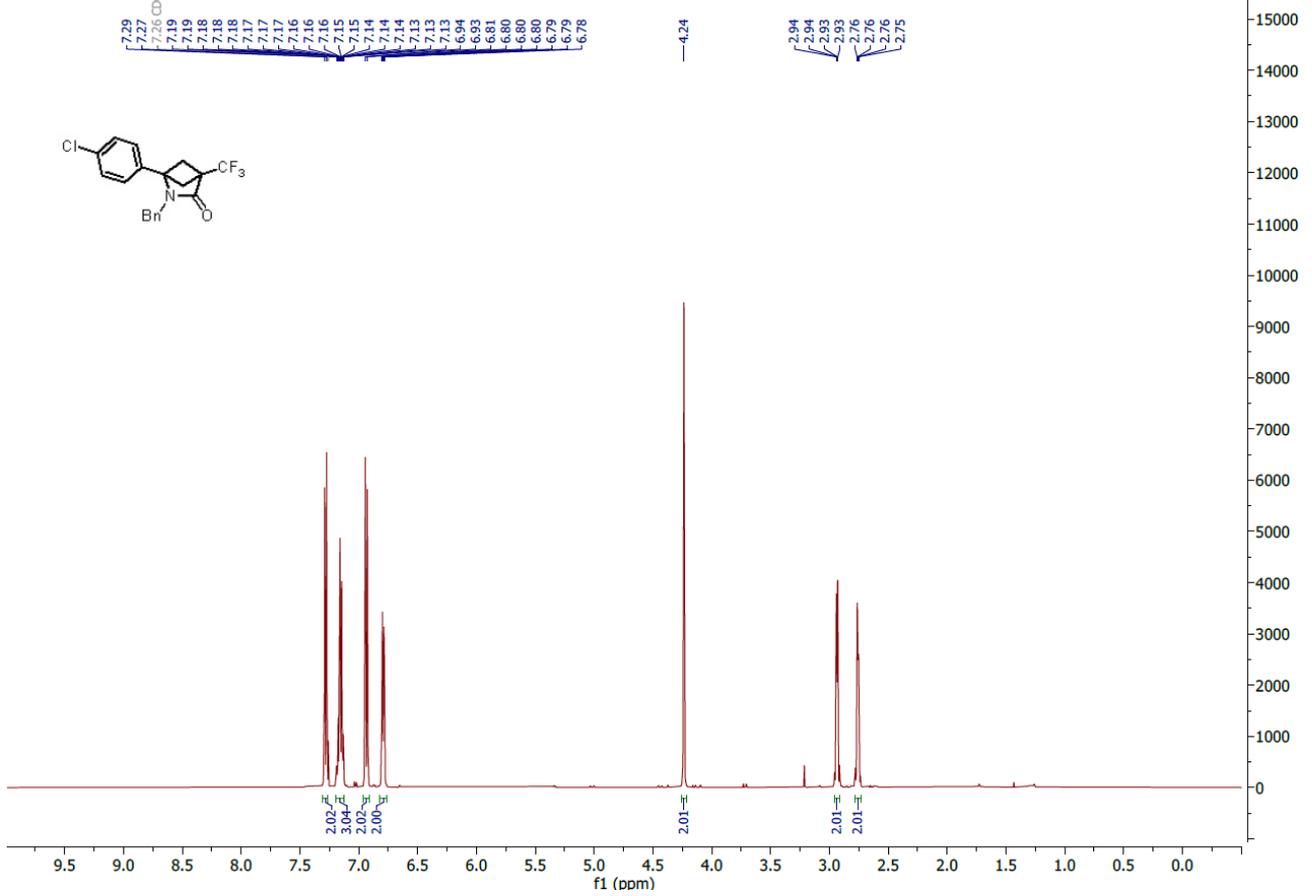
4604 zzx-uob-671-500M.11.fid



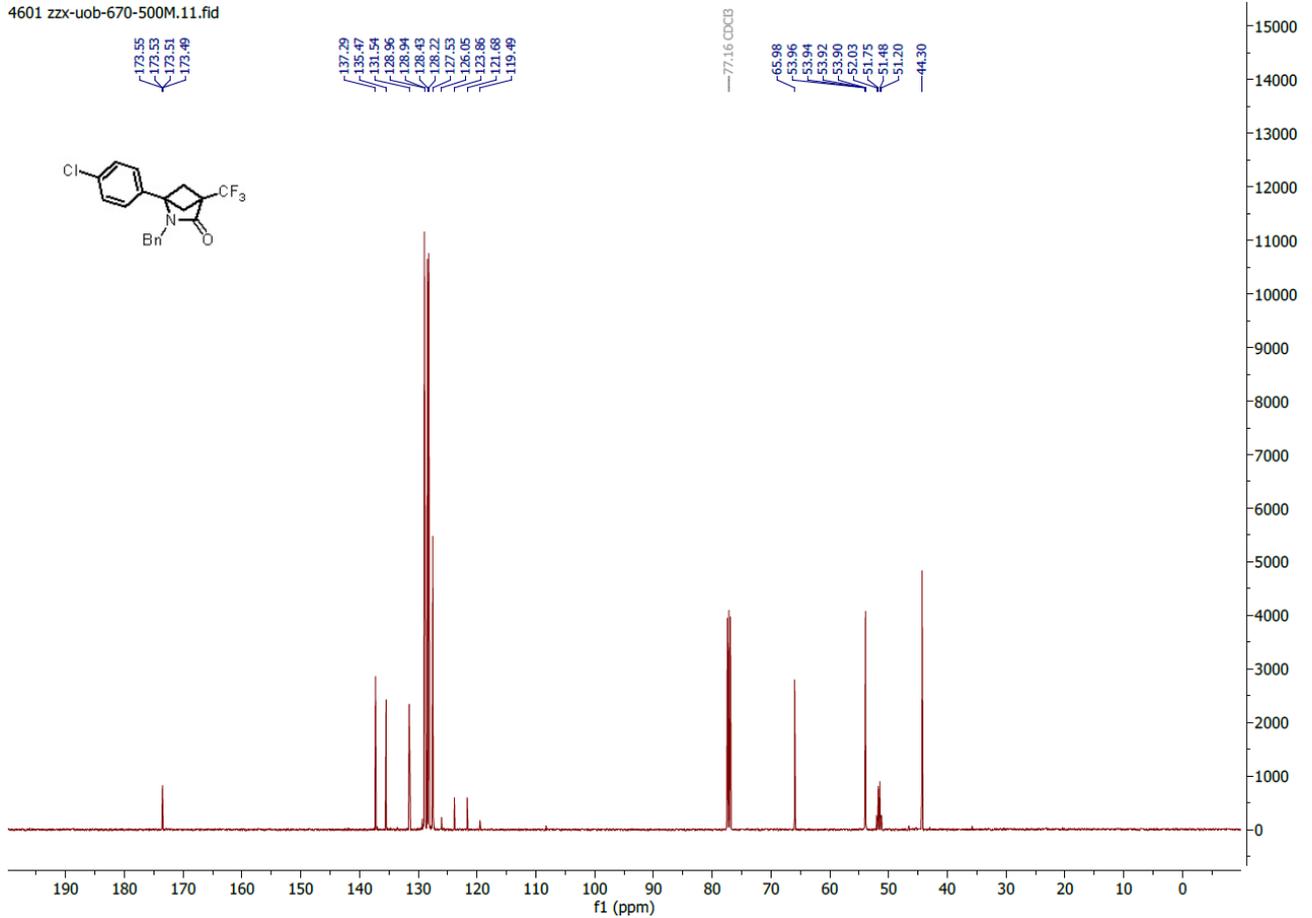


Compound 3l

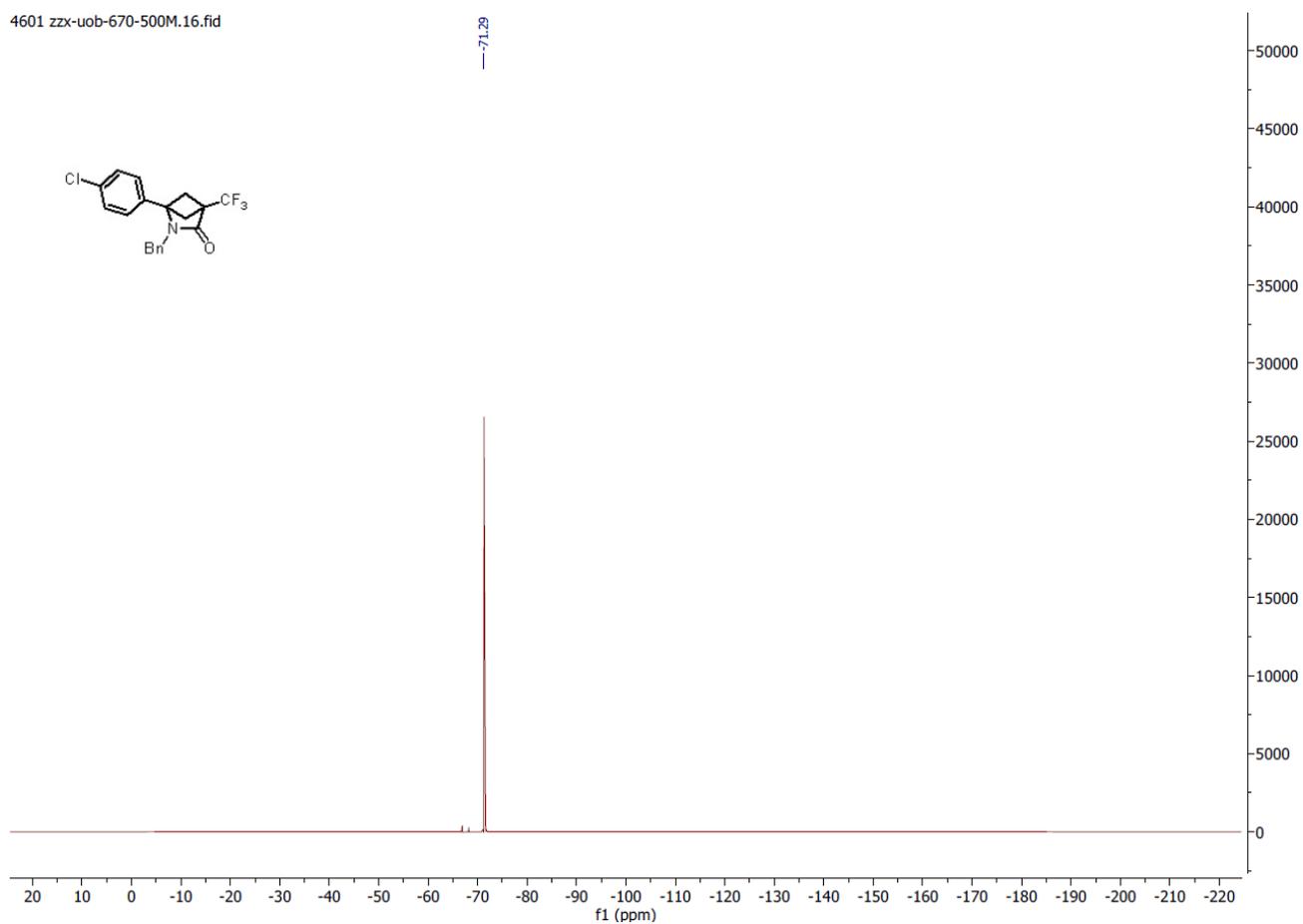
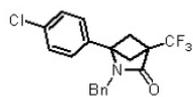
4601 zzx-uob-670-500M.10.fid



4601 zzx-uob-670-500M.11.fid

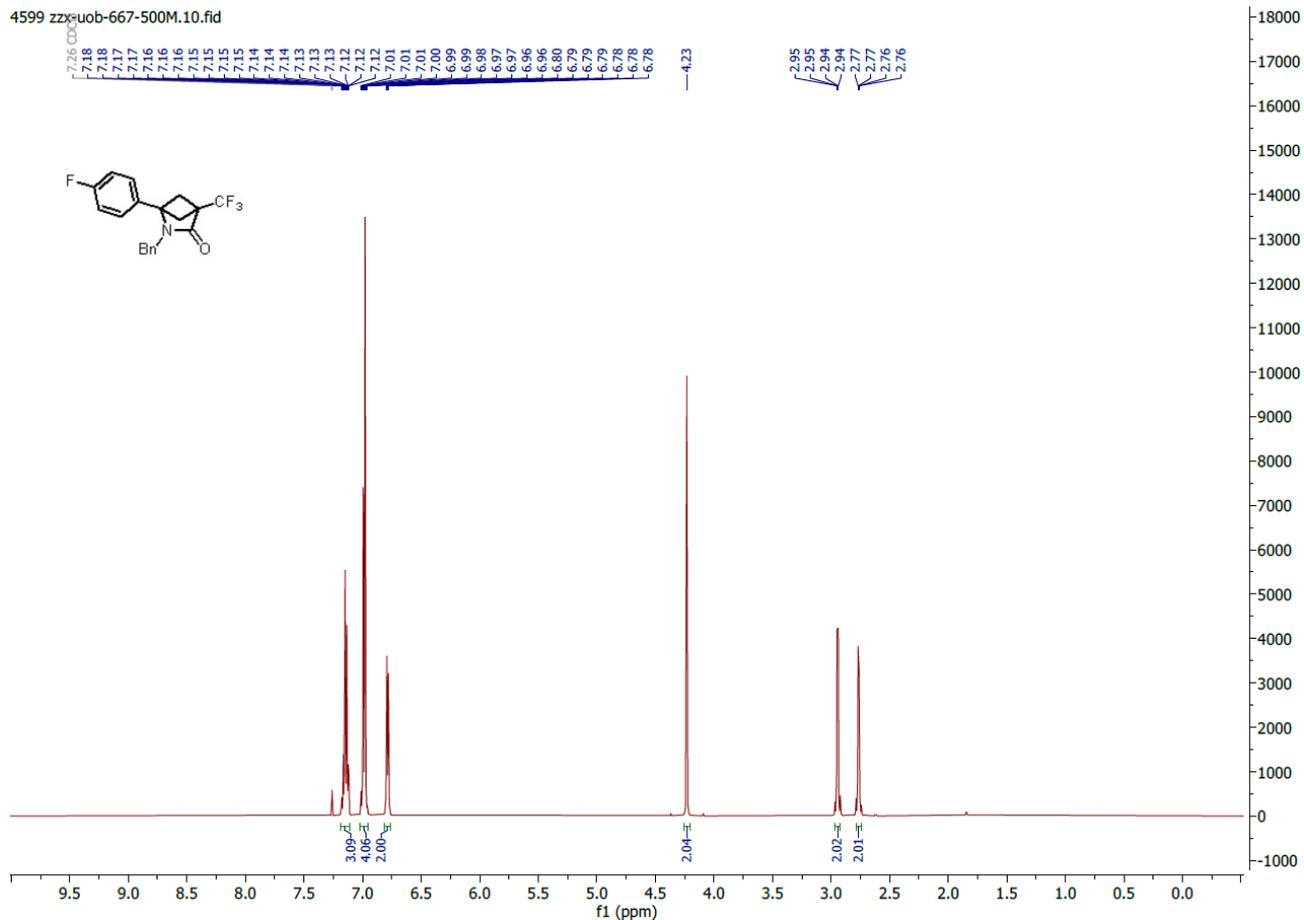


—71.29

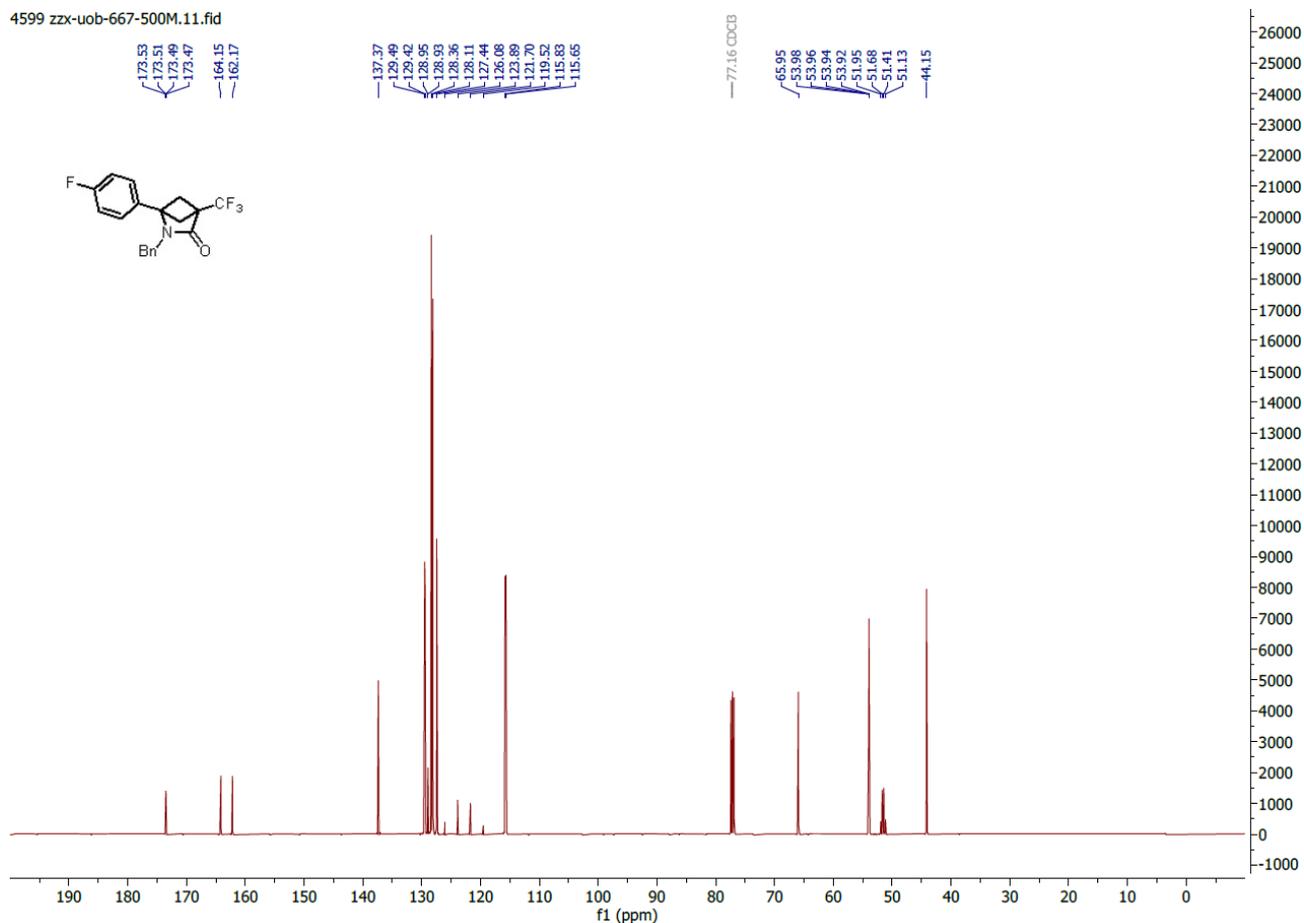


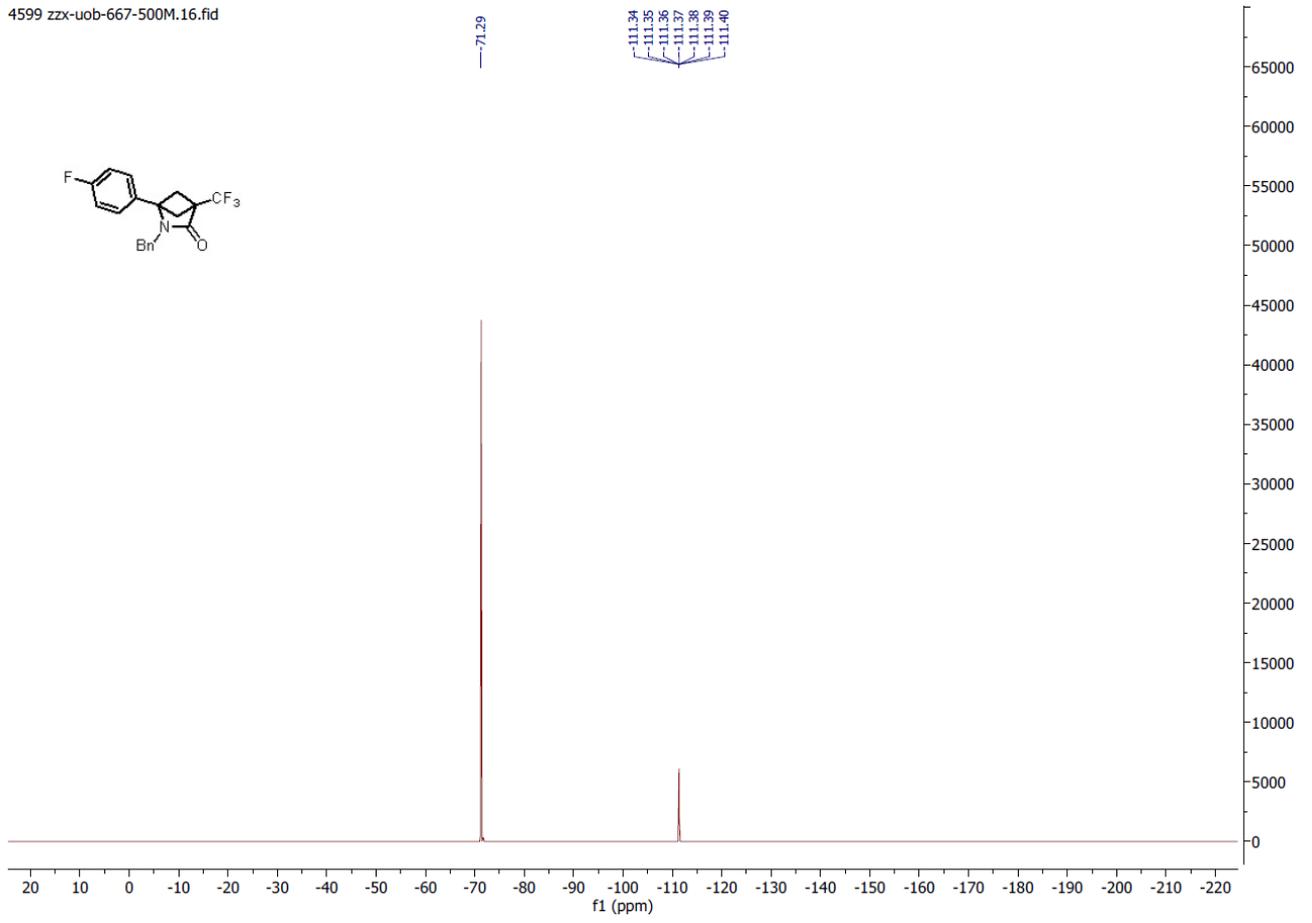
Compound 3m

4599 zzx-uob-667-500M.10.fid



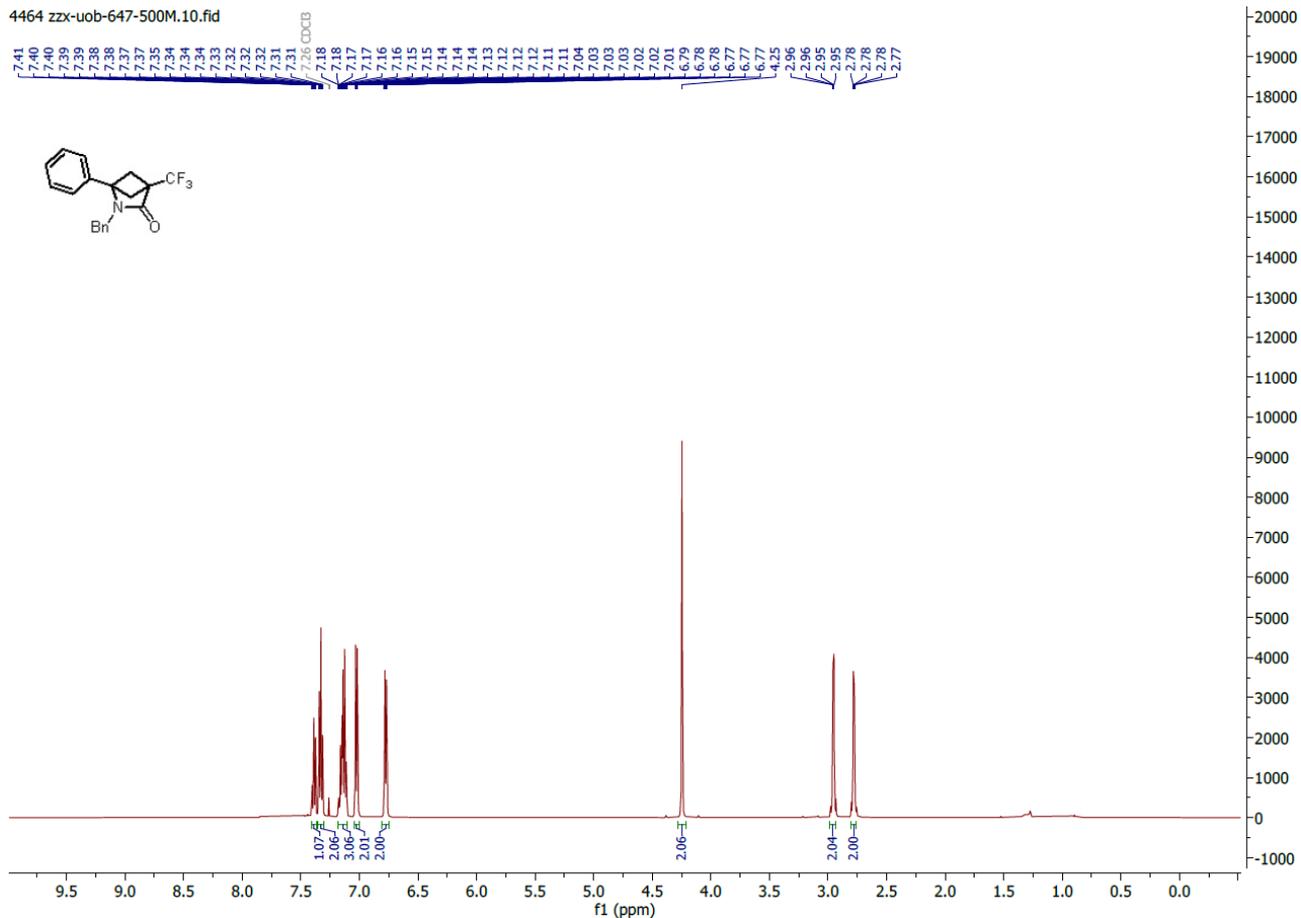
4599 zzx-uob-667-500M.11.fid



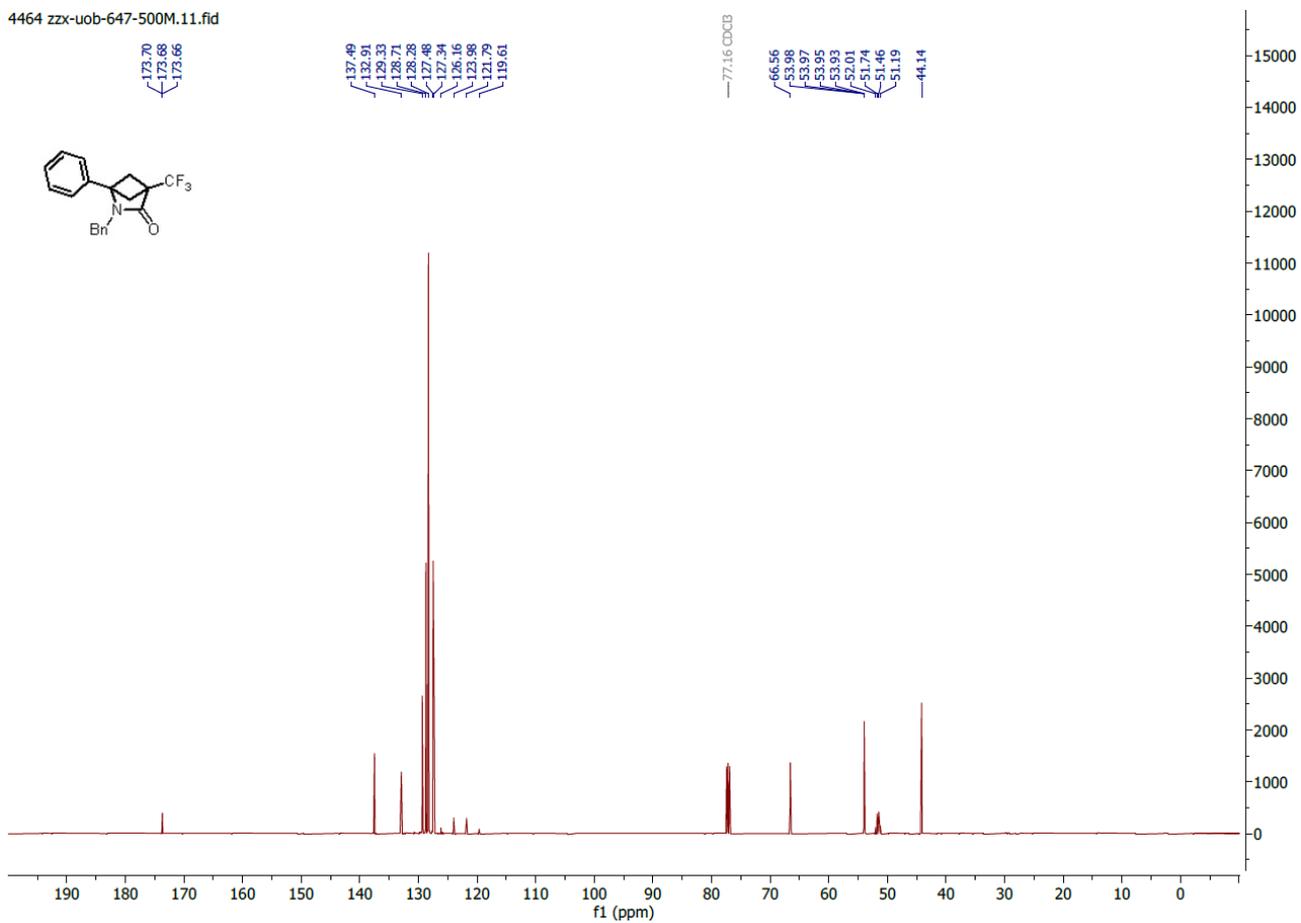


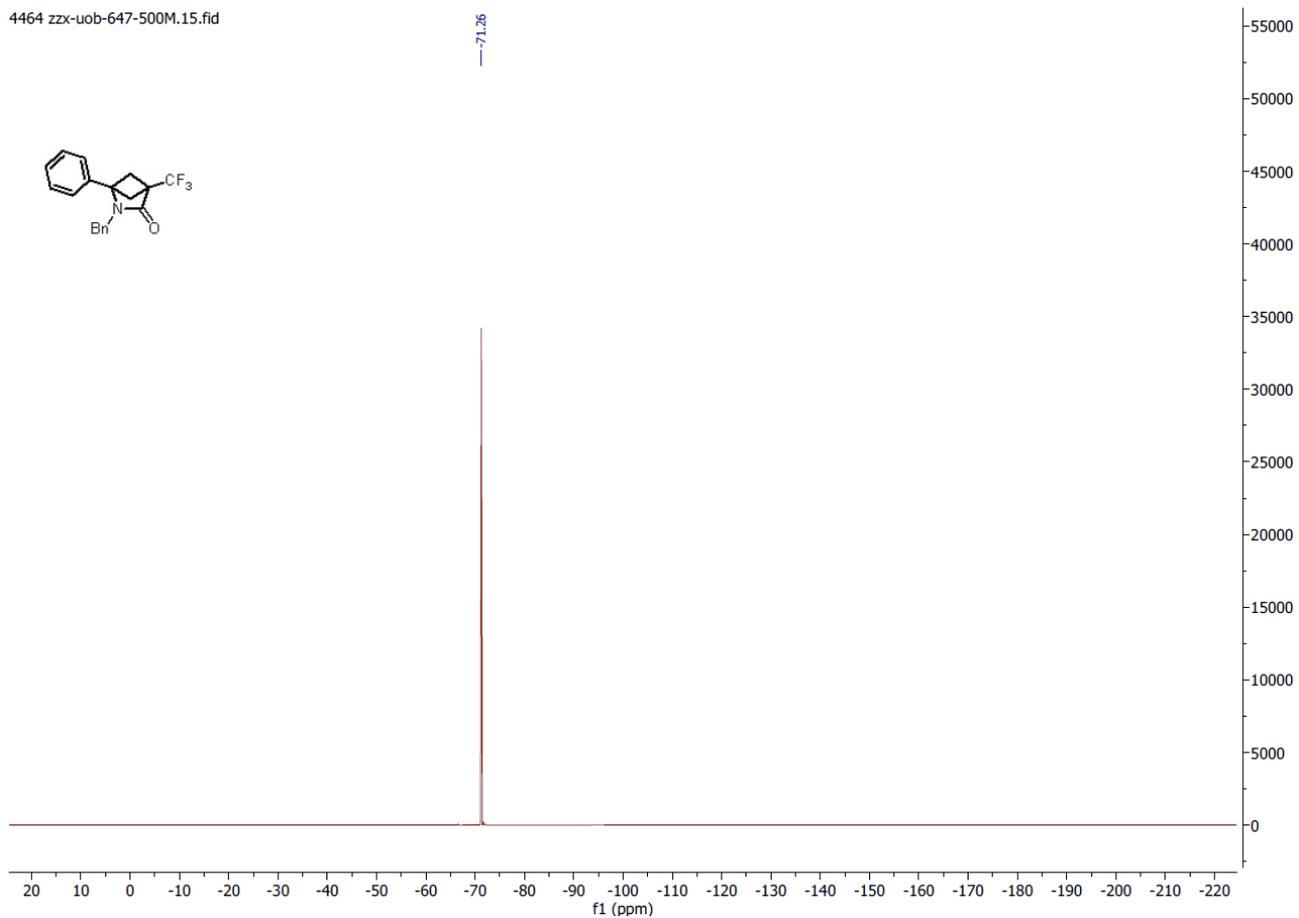
Compound 3n

4464 zzx-uob-647-500M.10.fid



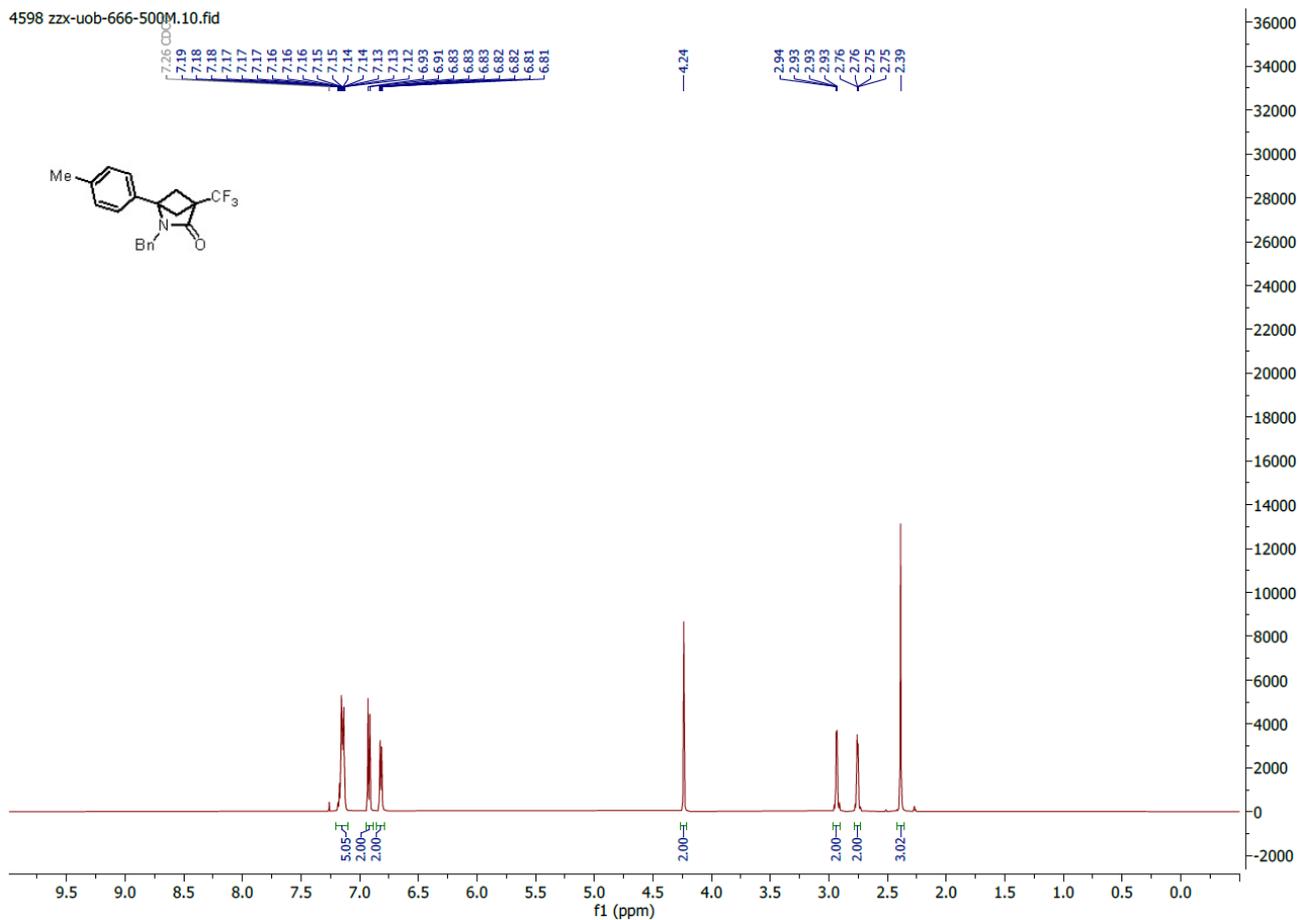
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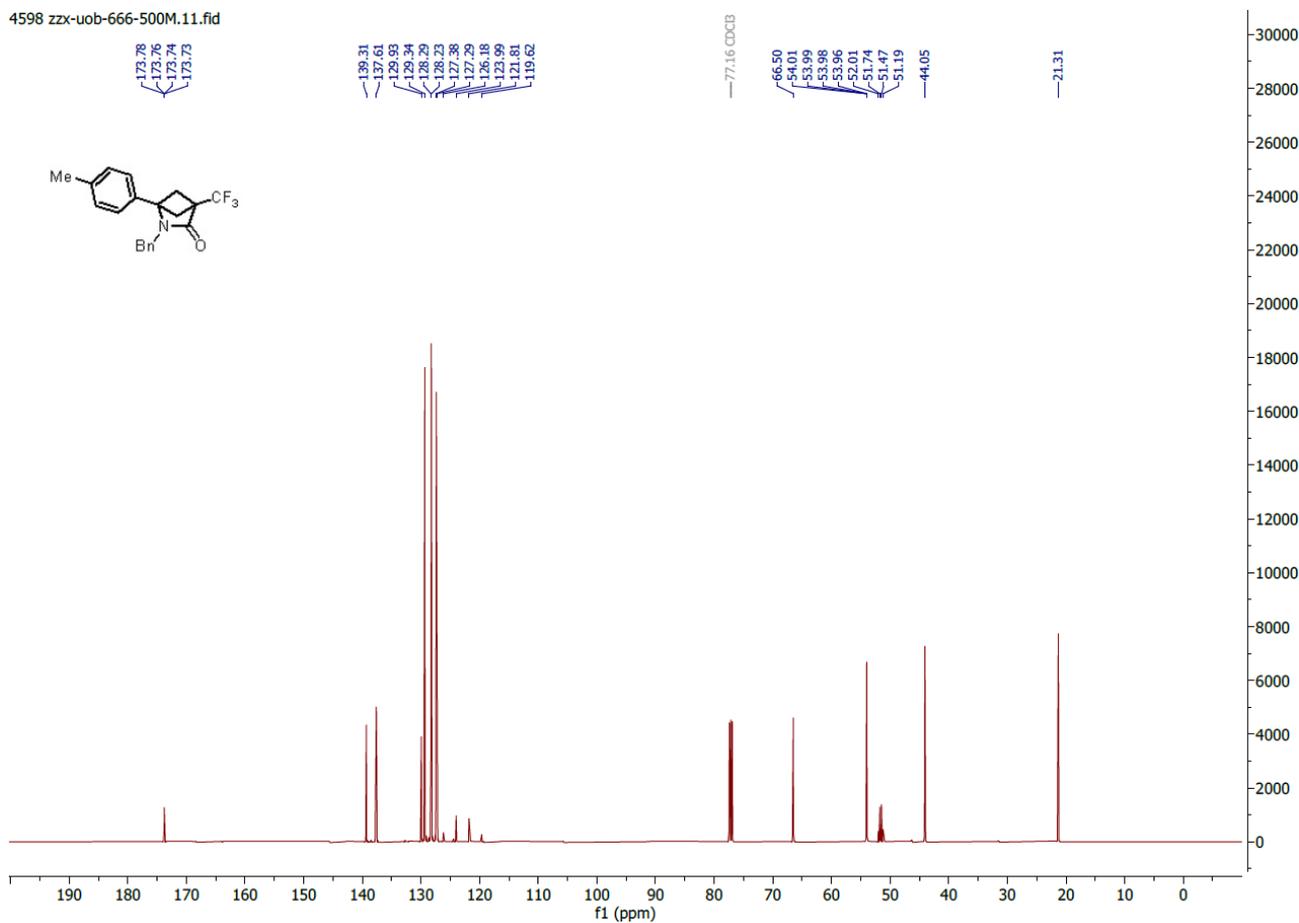


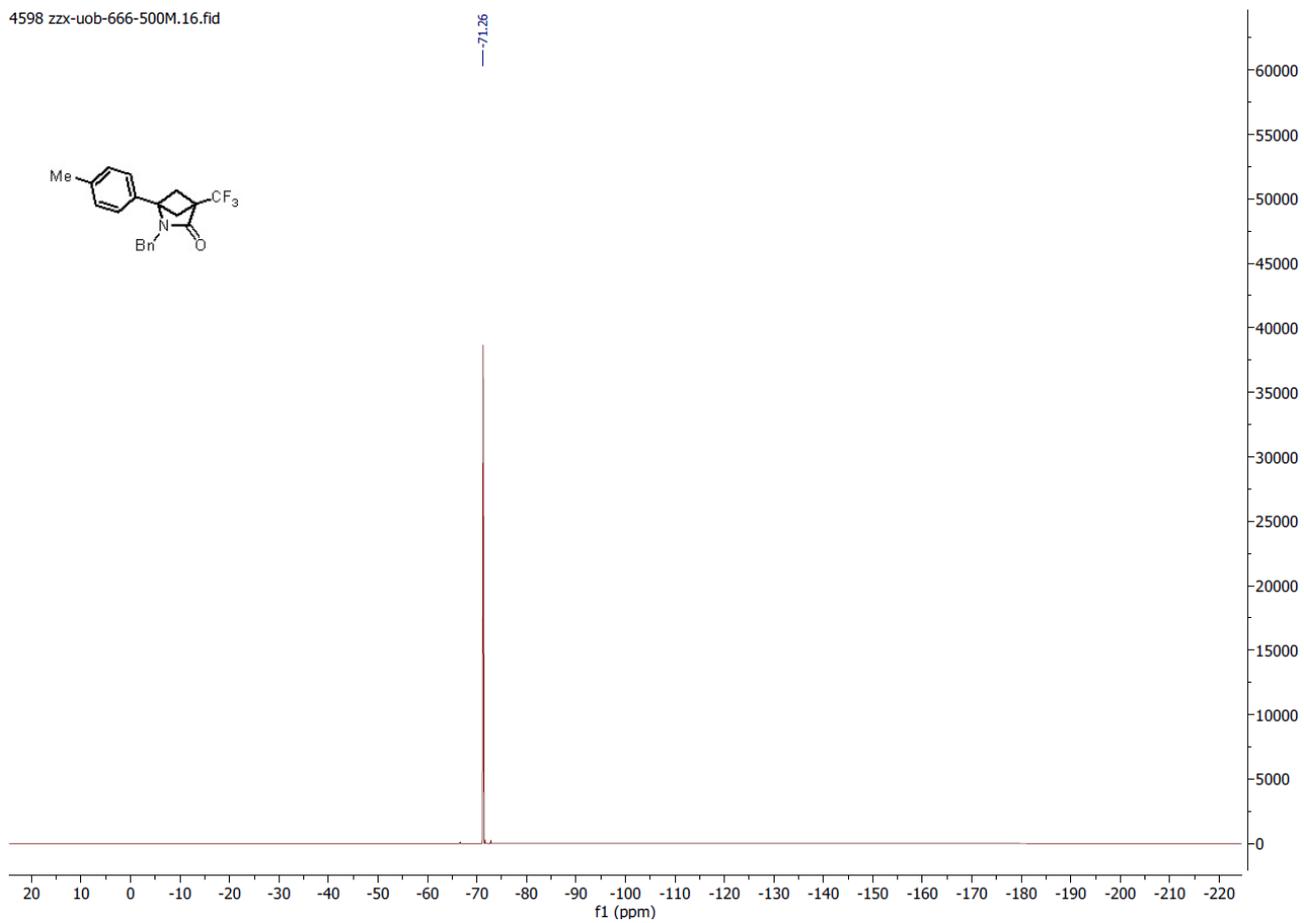
Compound 3o

4598 zzx-uob-666-500M.10.fid



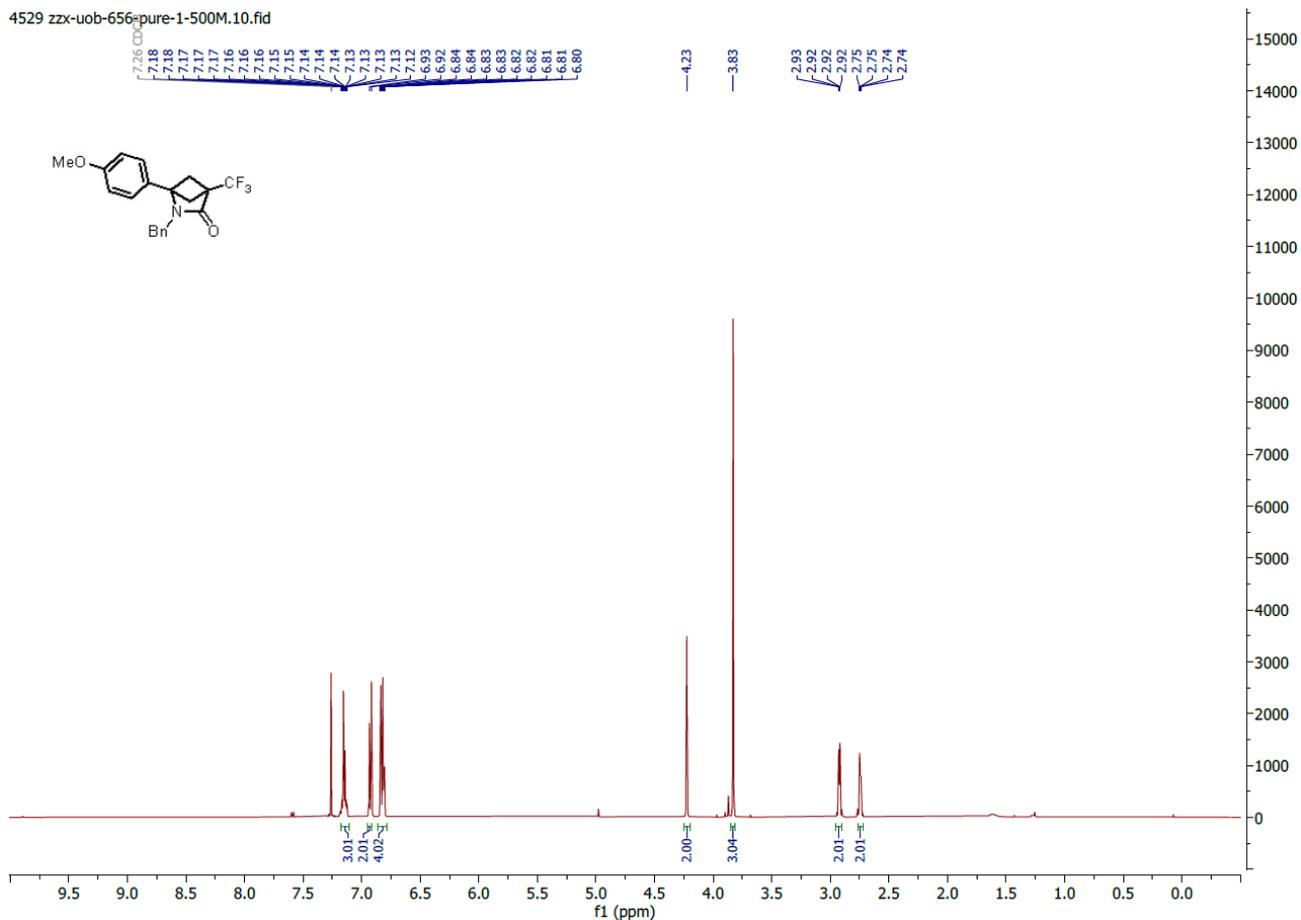
4598 zzx-uob-666-500M.11.fid



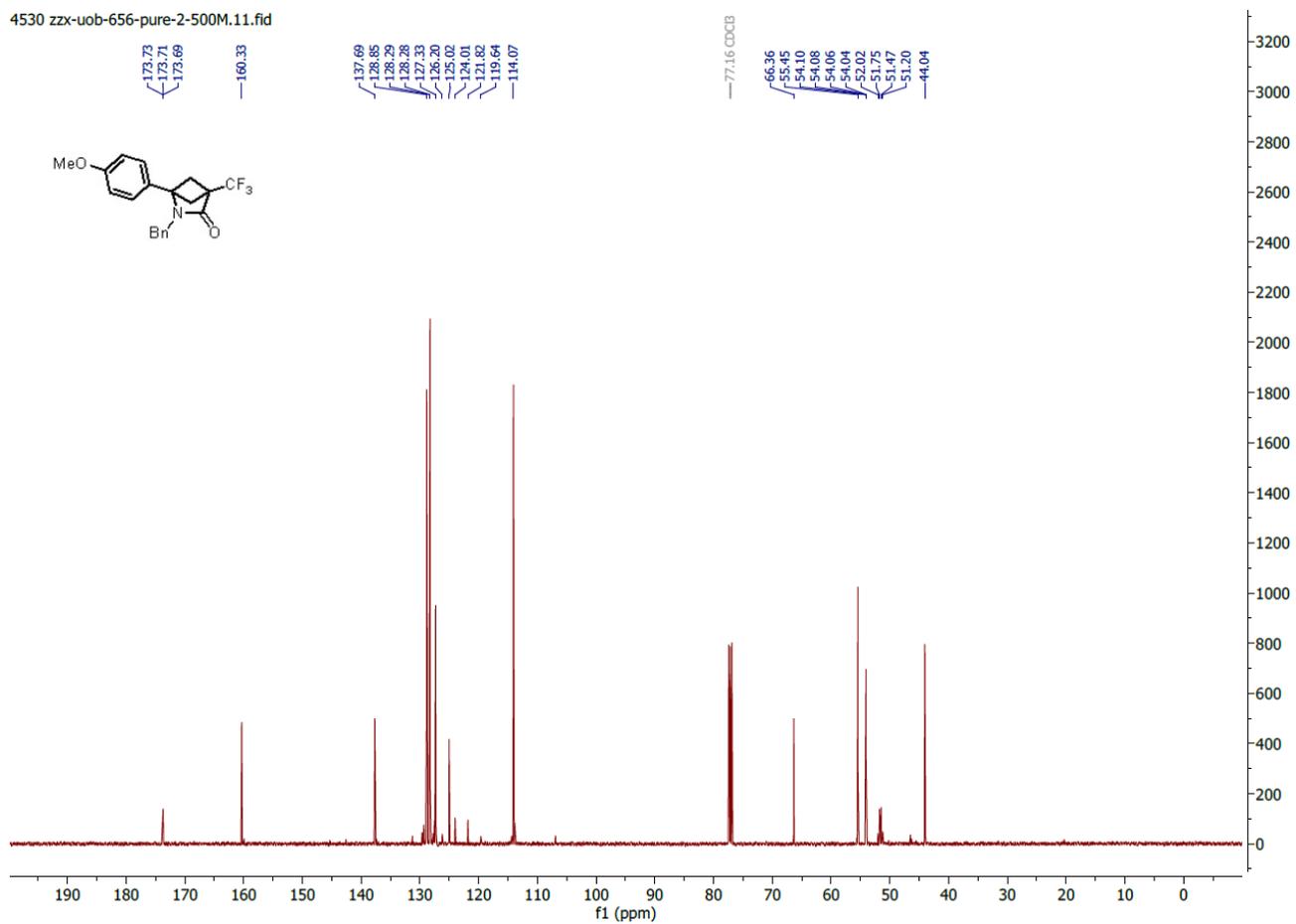


Compound 3p

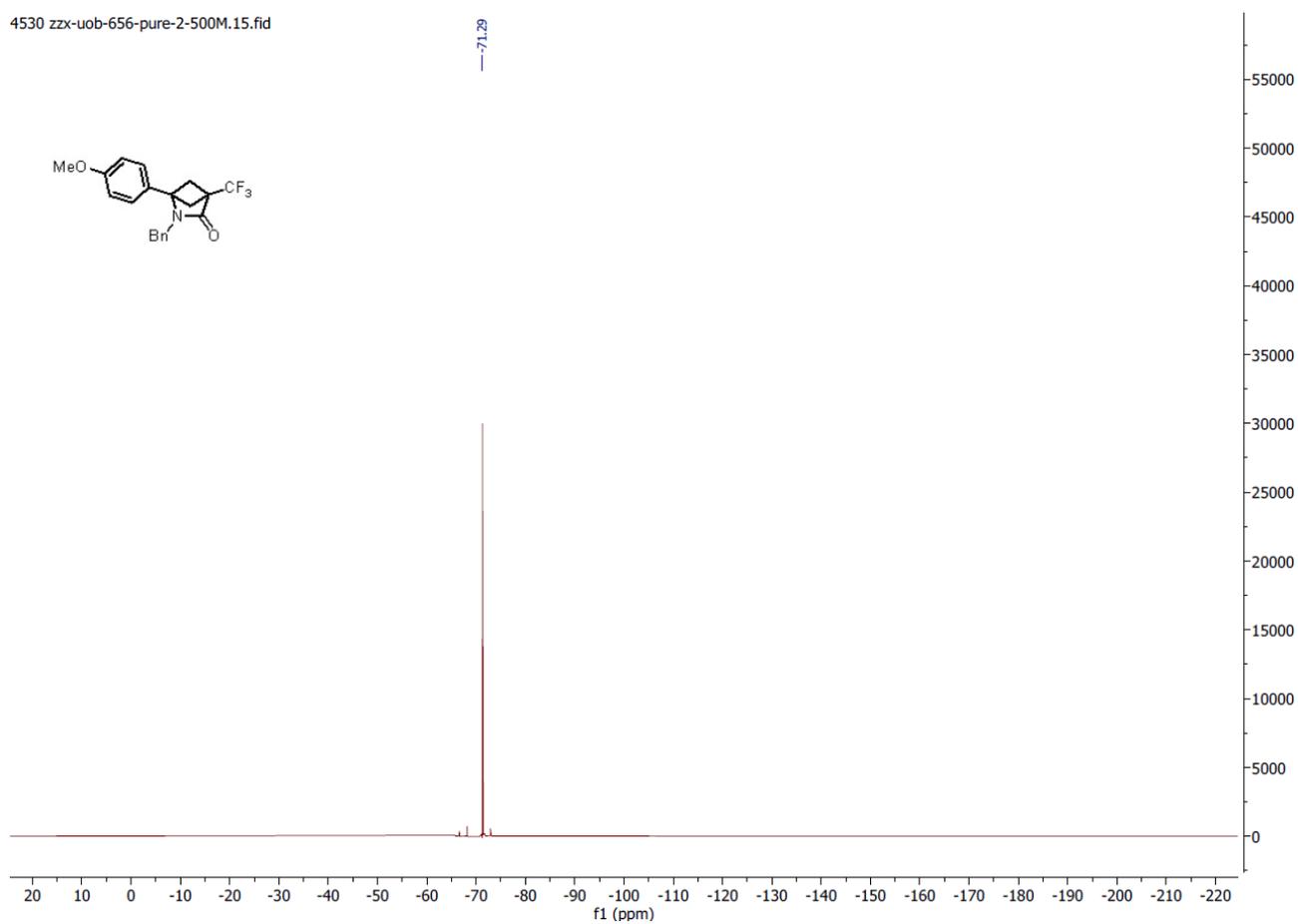
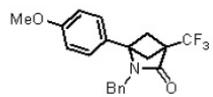
4529 zzx-uob-656-pure-1-500M.10.fid



4530 zzx-uob-656-pure-2-500M.11.fid

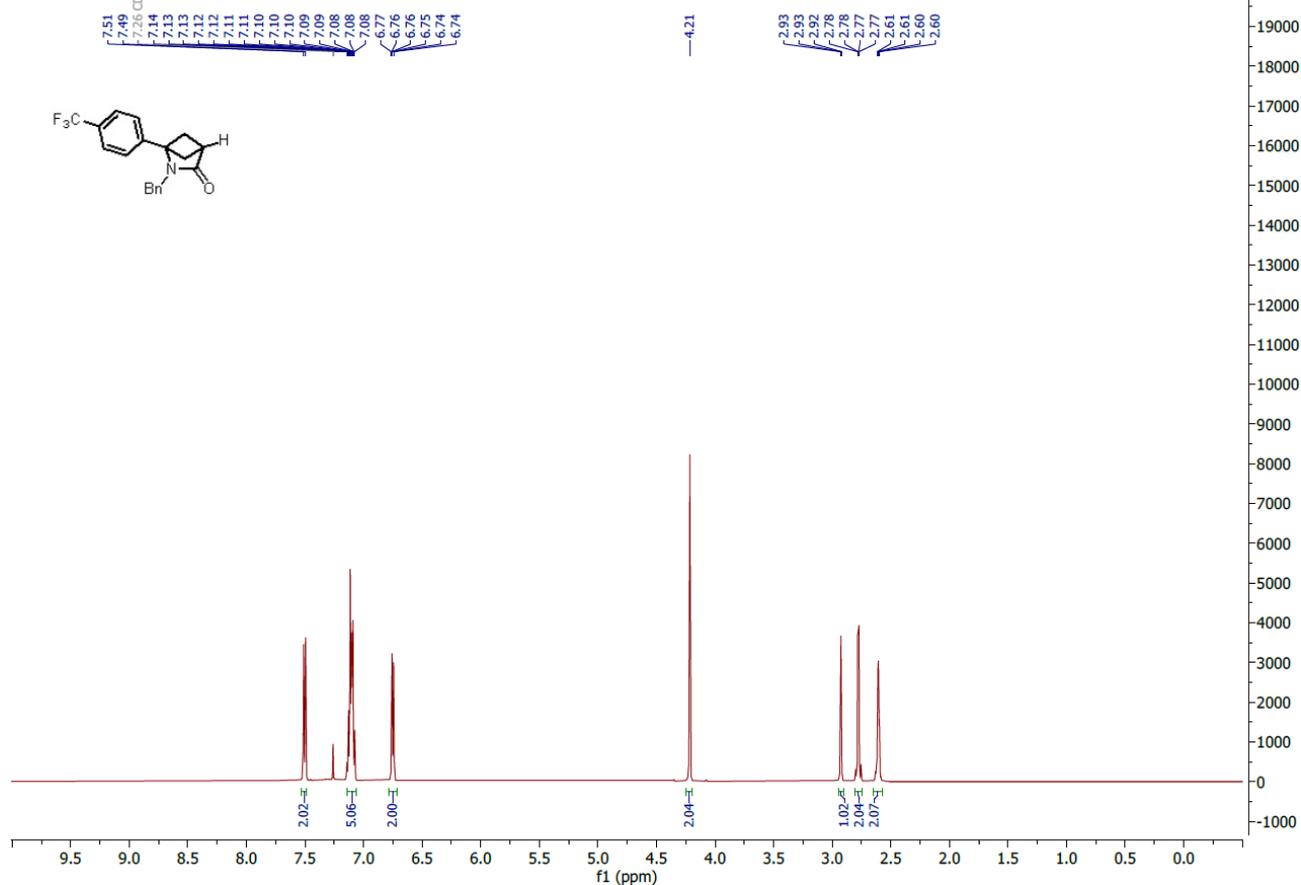


-71.29

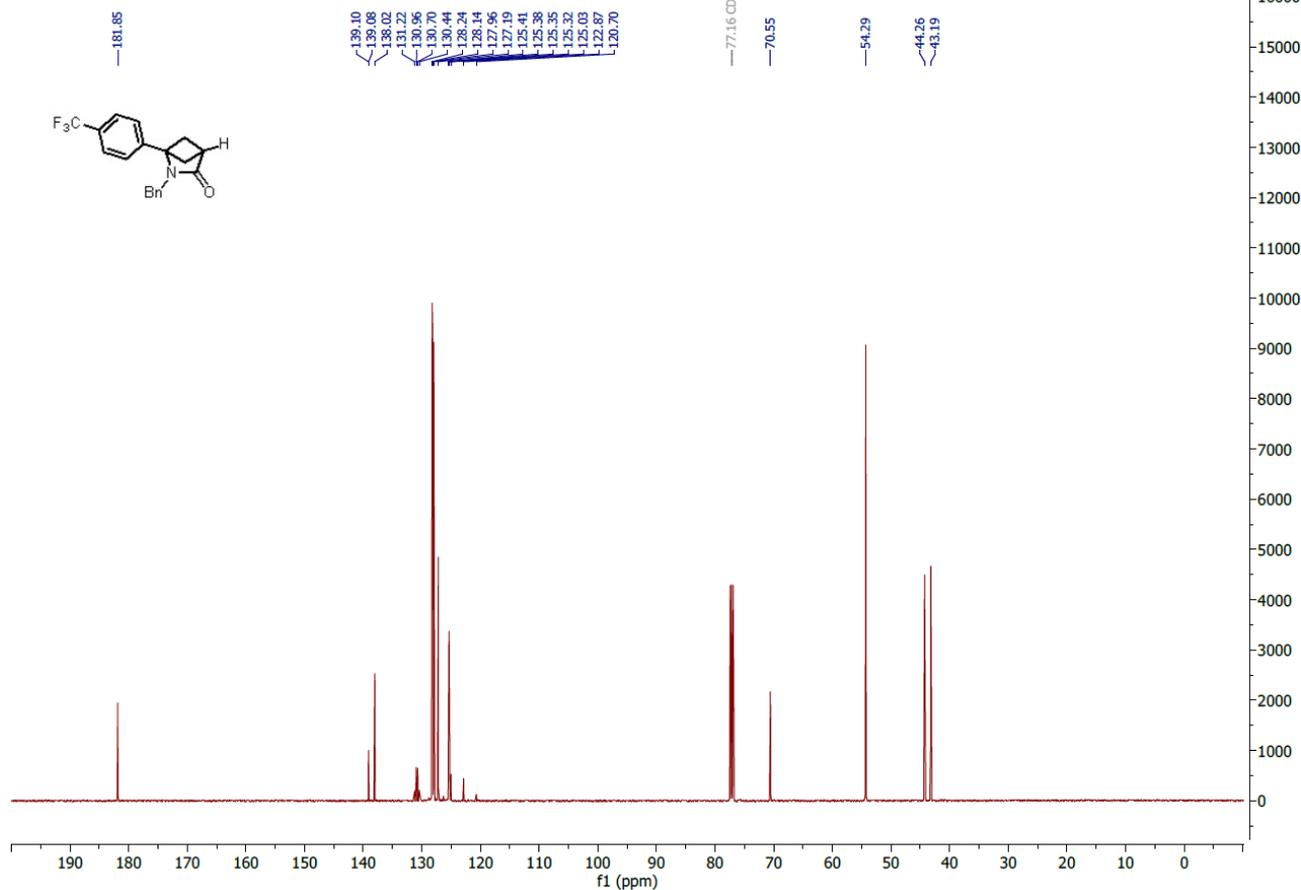


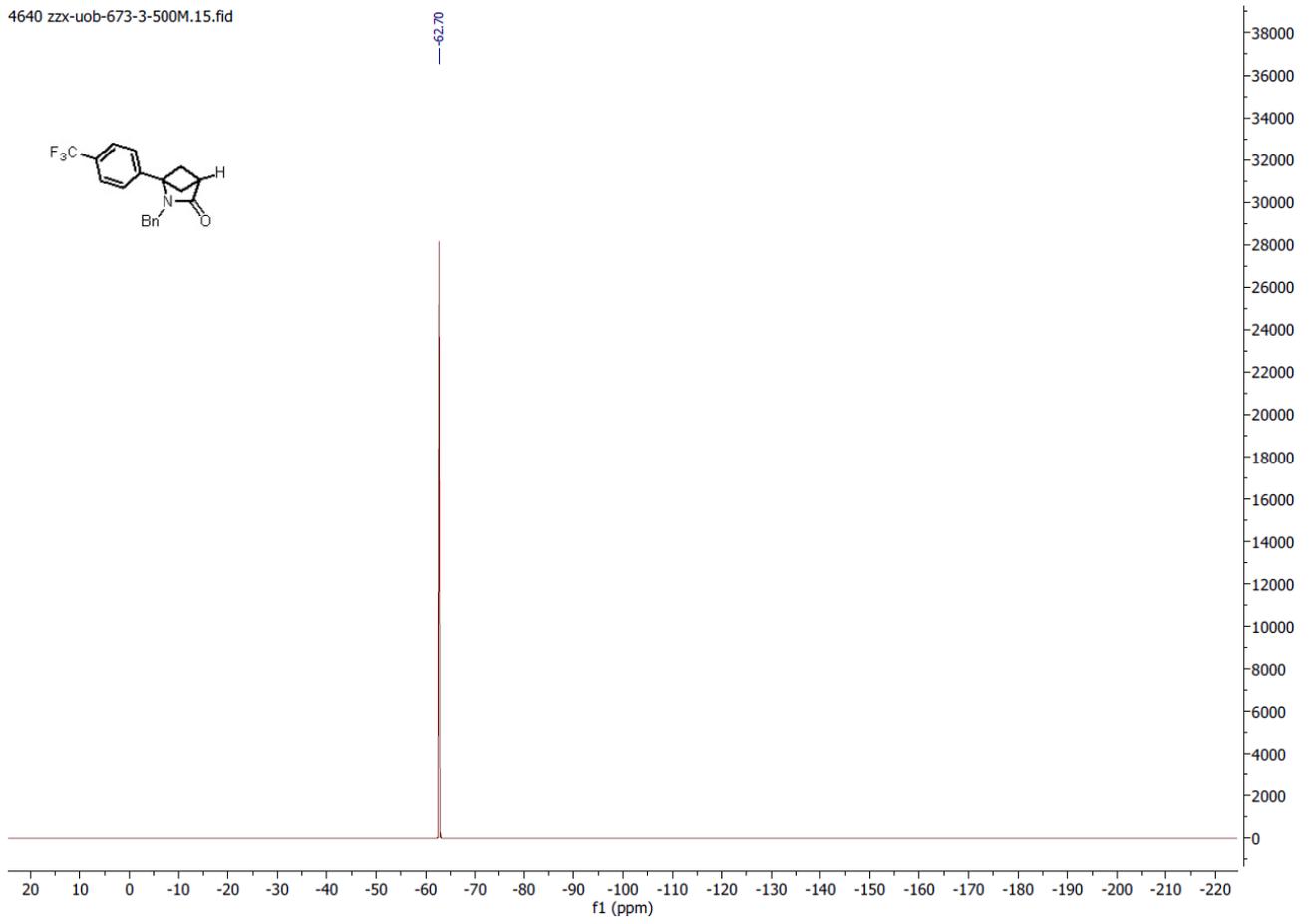
Compound 3q

4640 zzx-uob-673-3-500M.10.fid



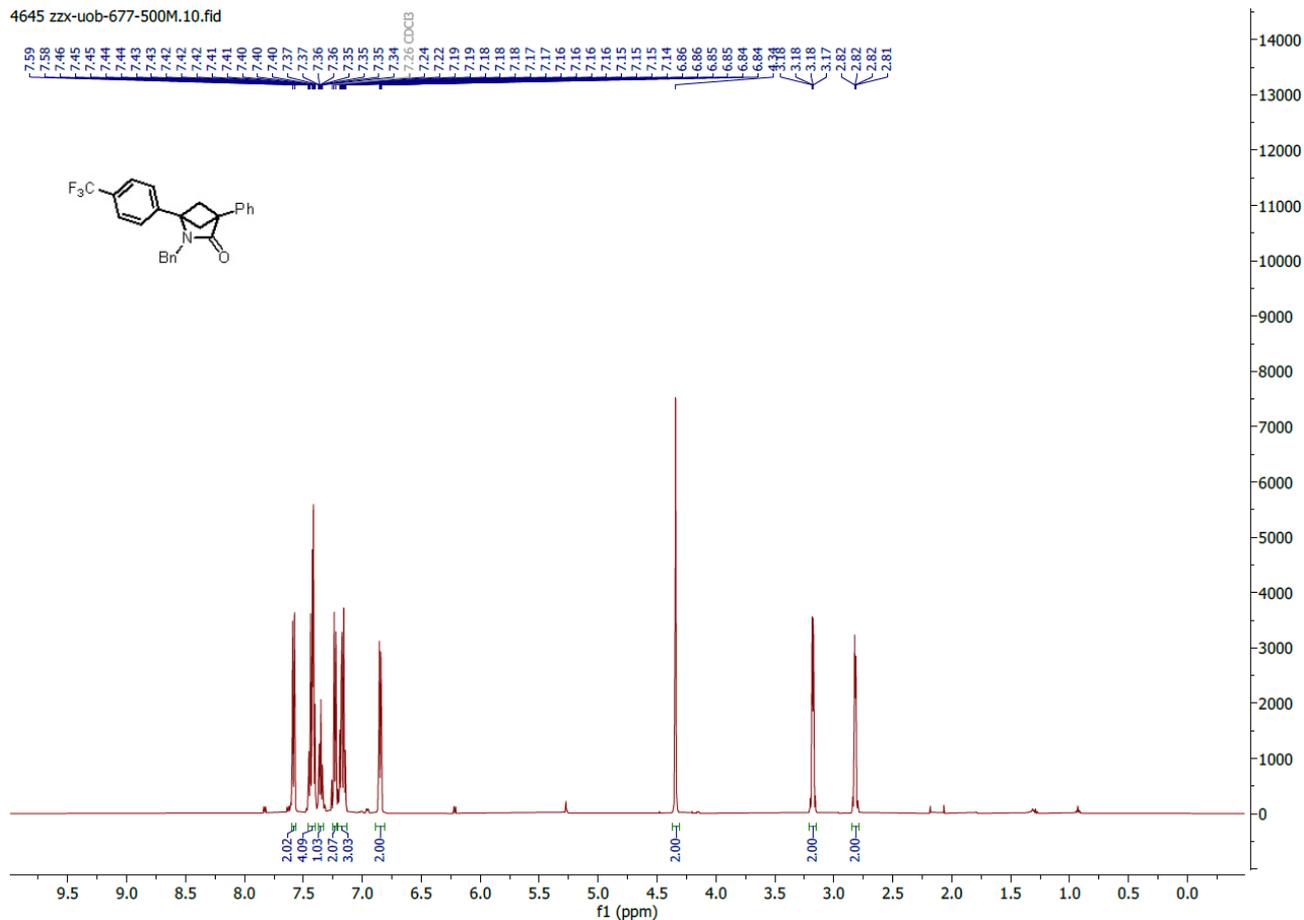
4640 zzx-uob-673-3-500M.11.fid



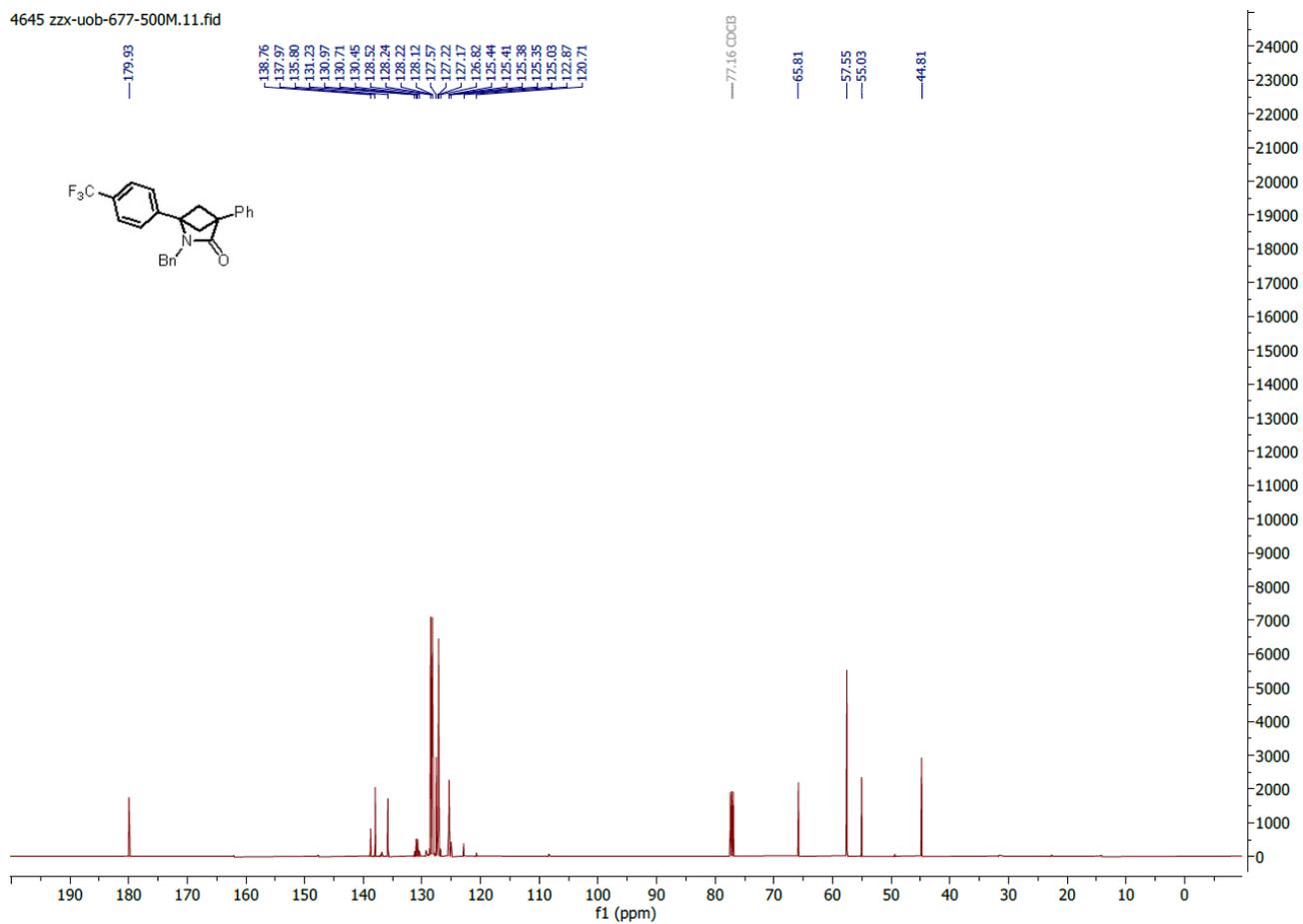


Compound 3r

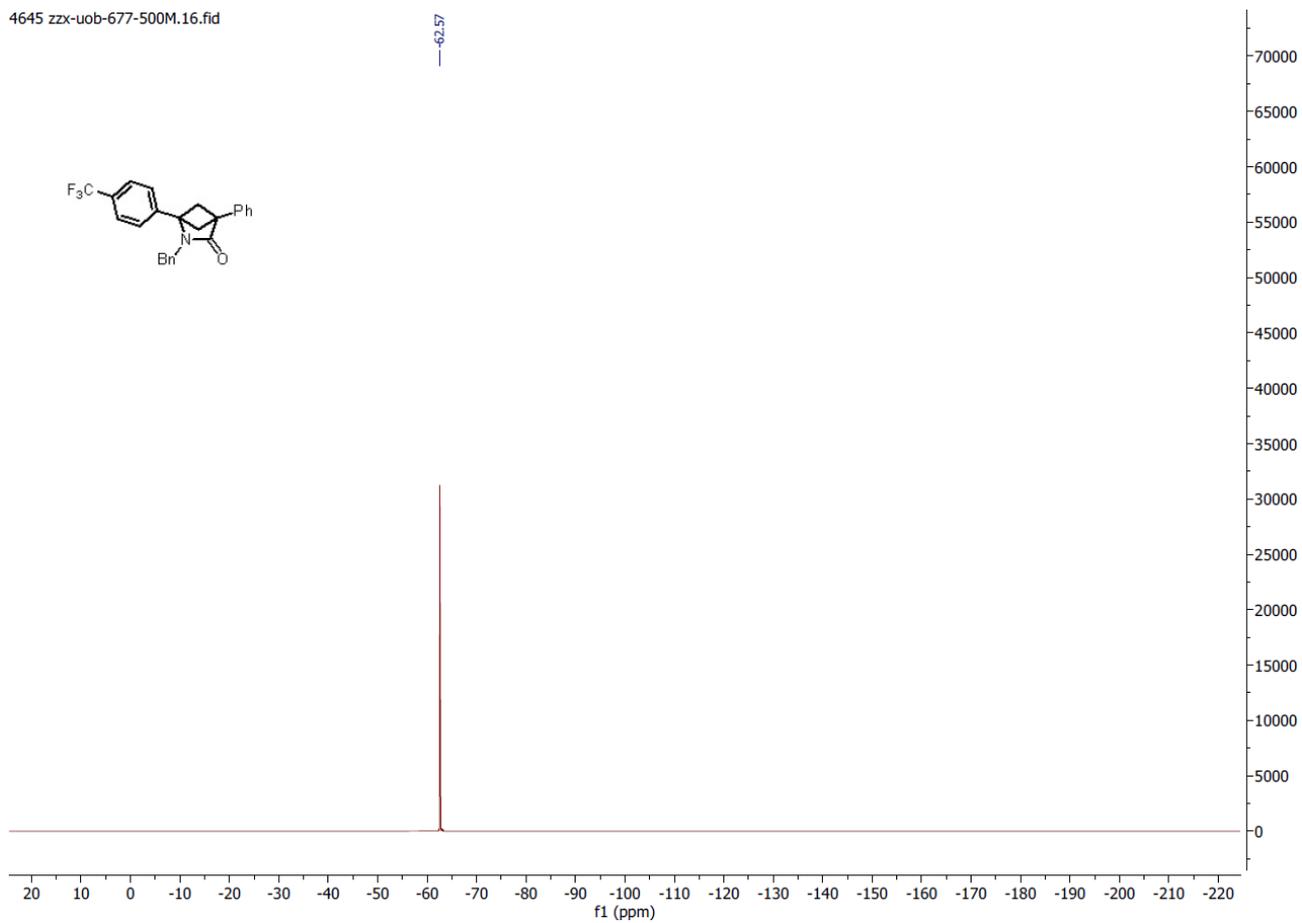
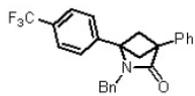
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4645 zzx-uob-677-500M.11.fid

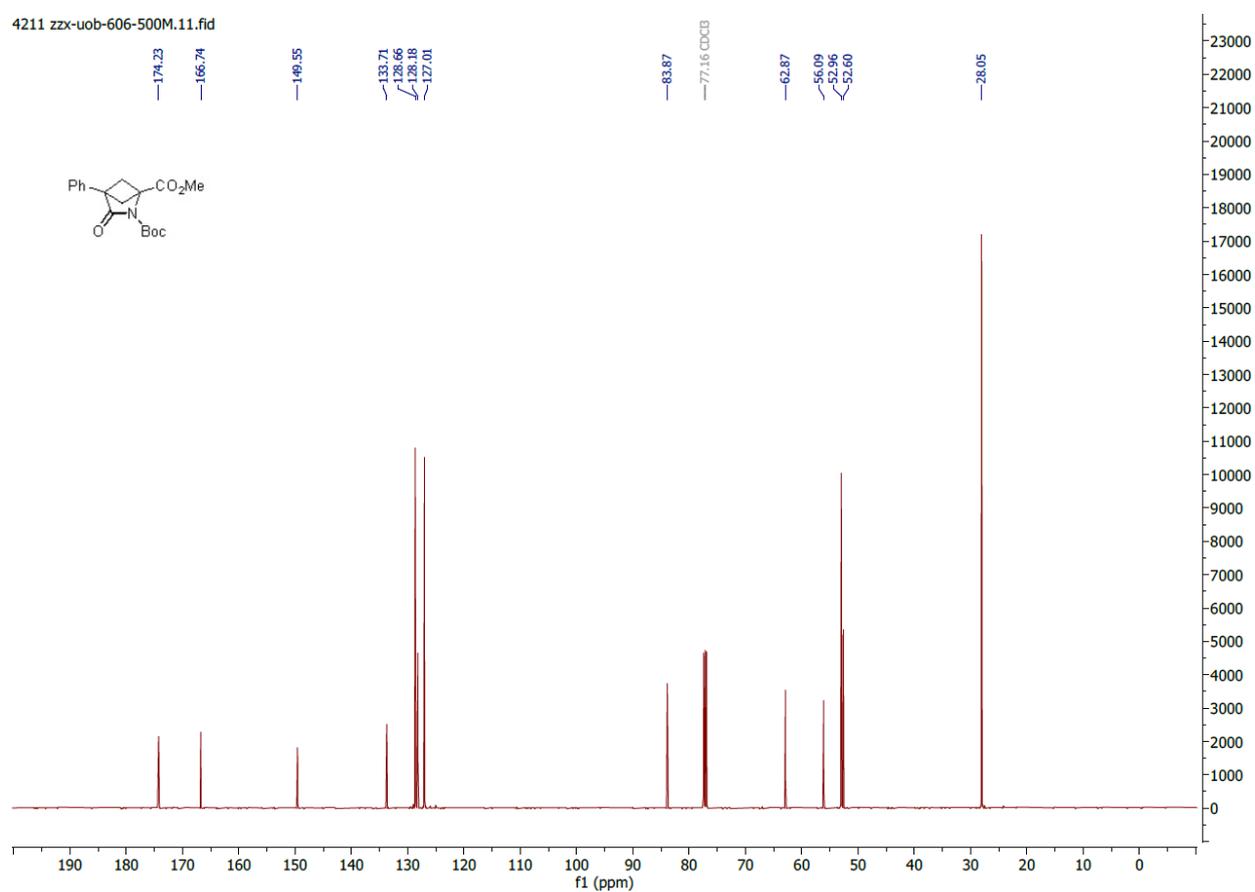
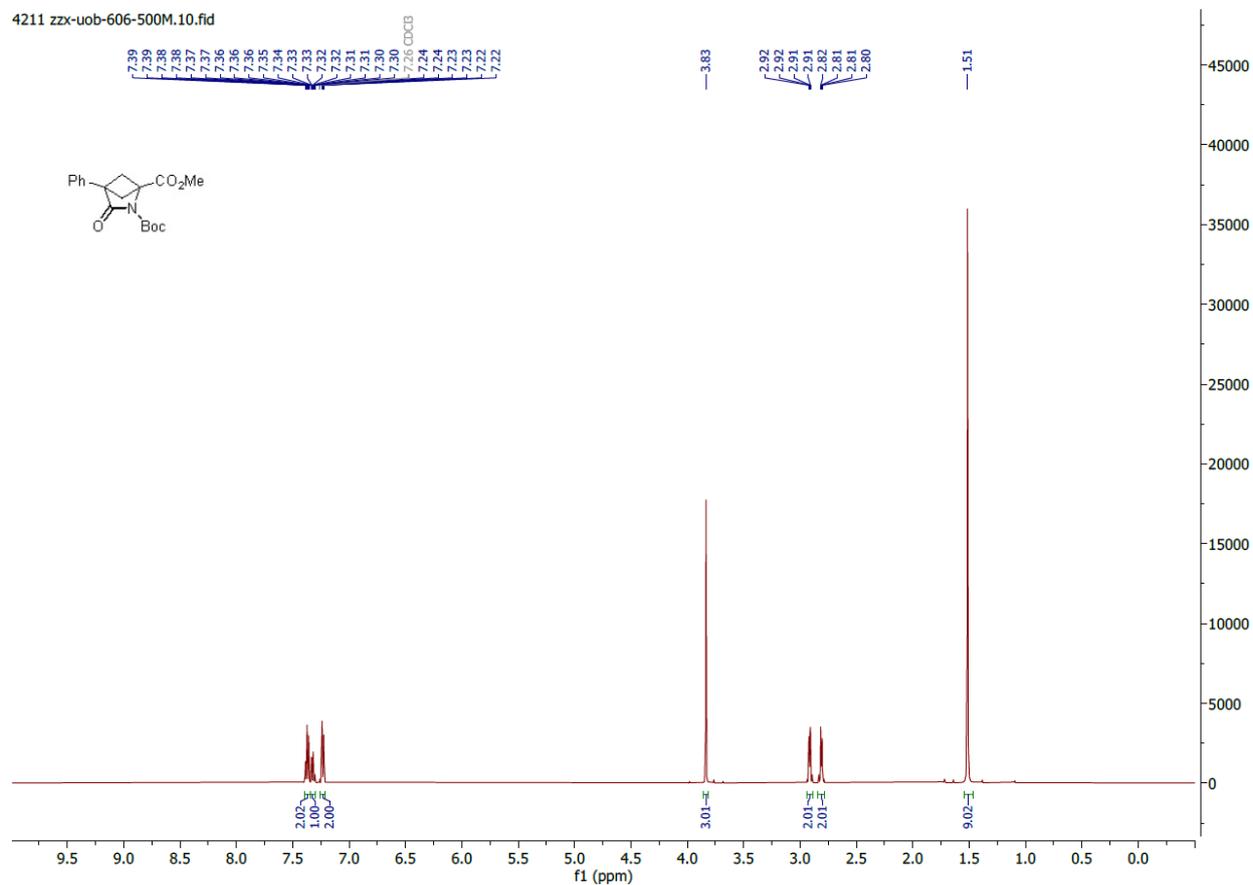


-63.57



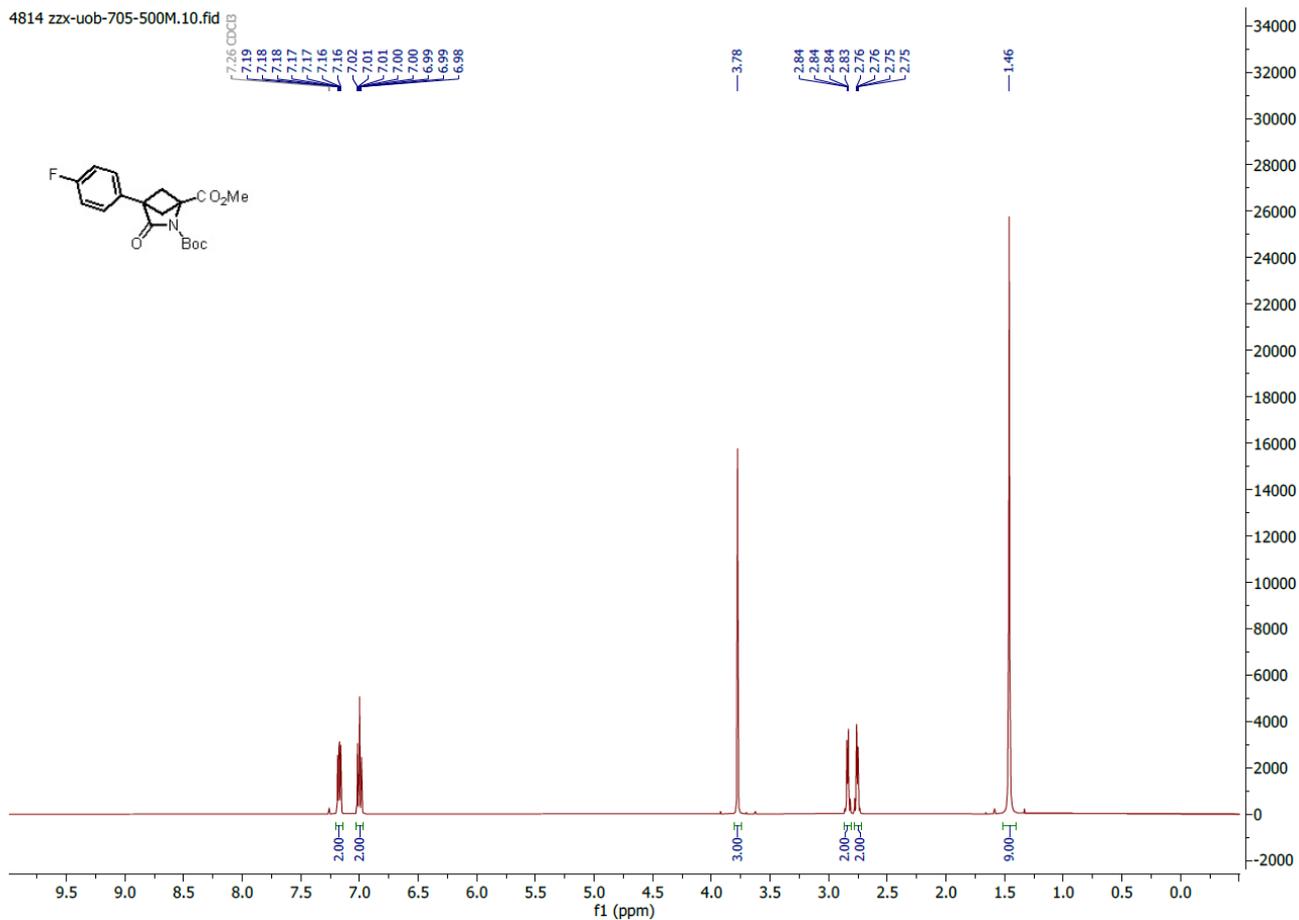
5.5 NMR Spectra of Products (8)

Compound 8a

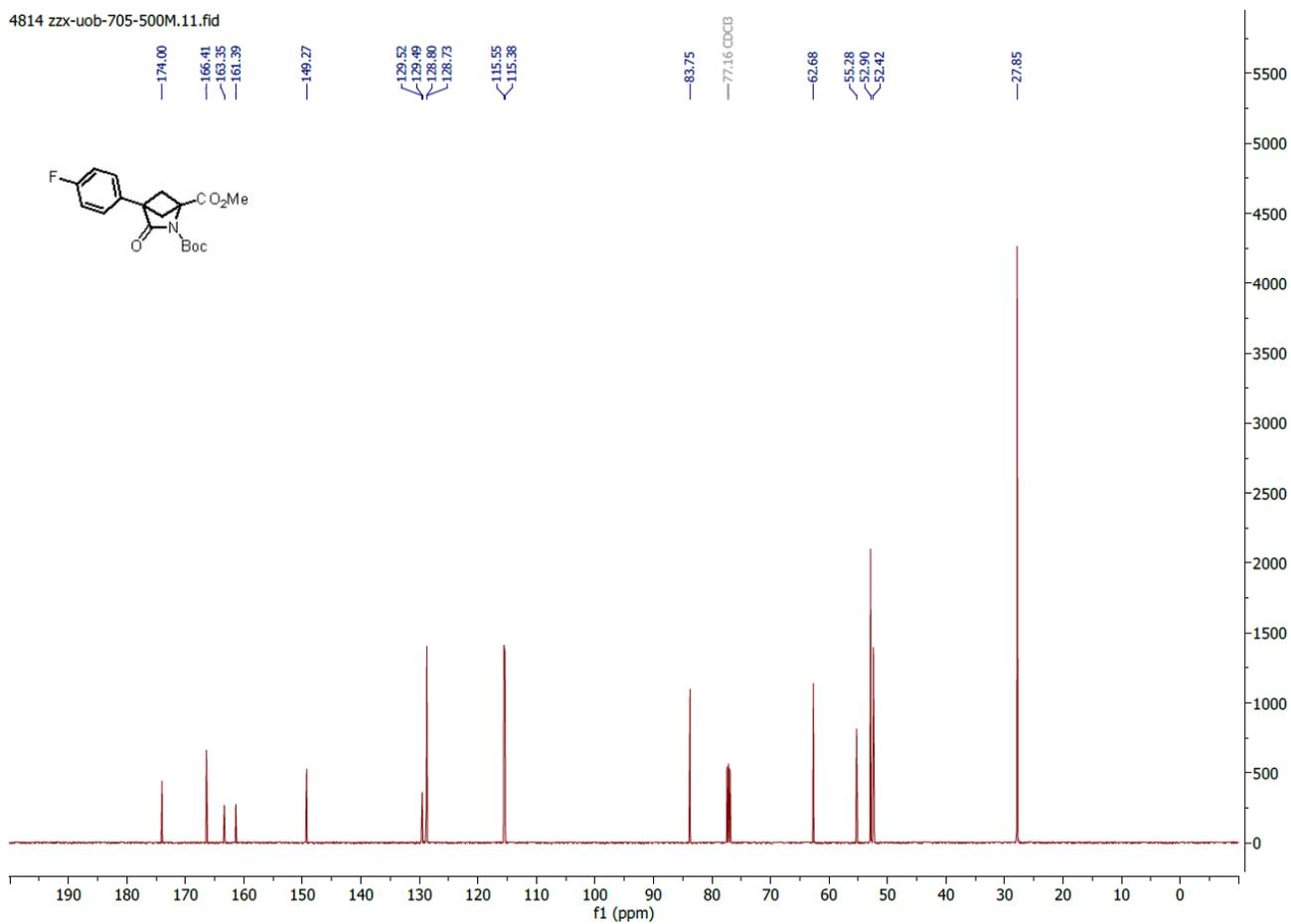


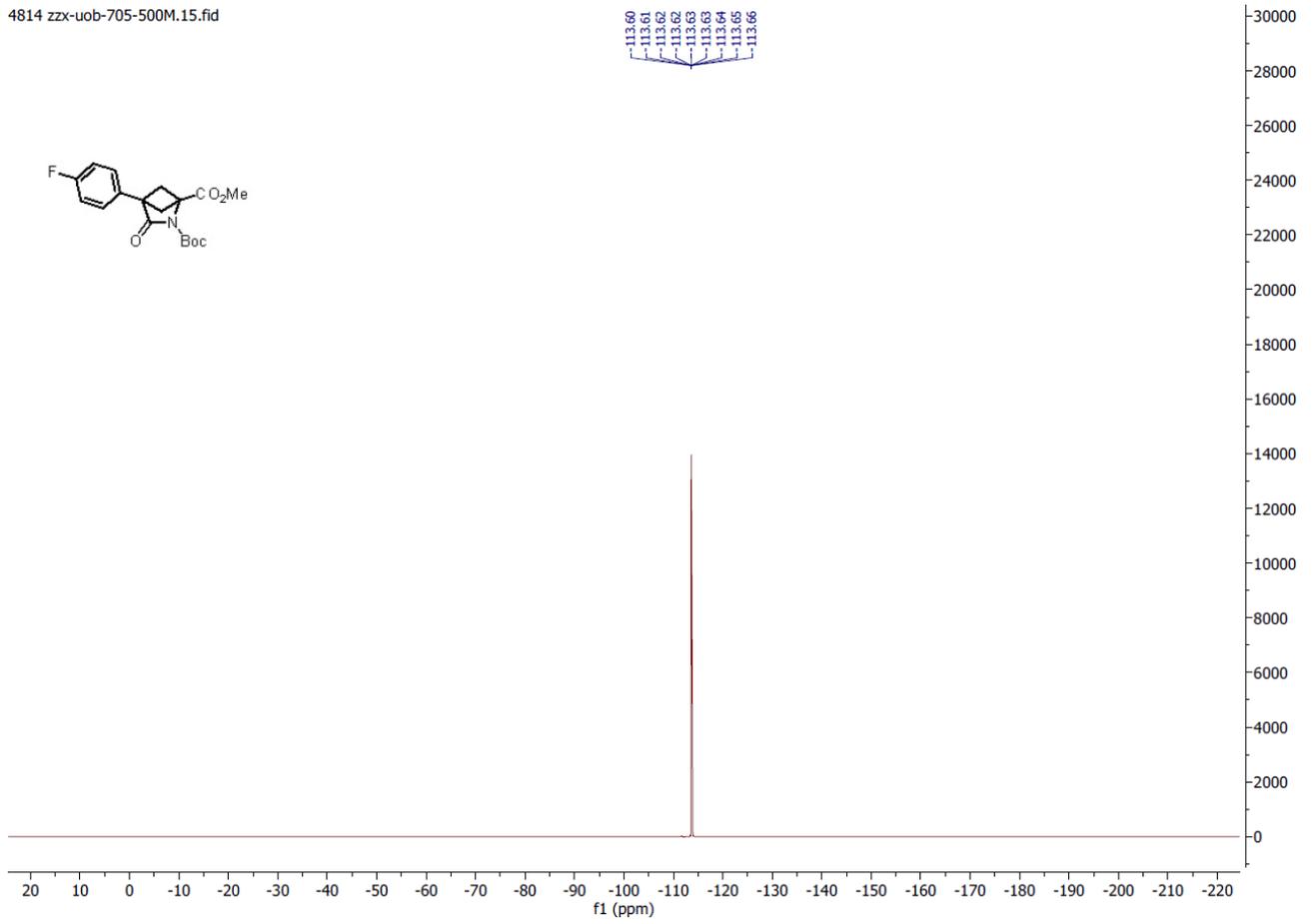
Compound 8b

4814 zzx-uob-705-500M.10.fid



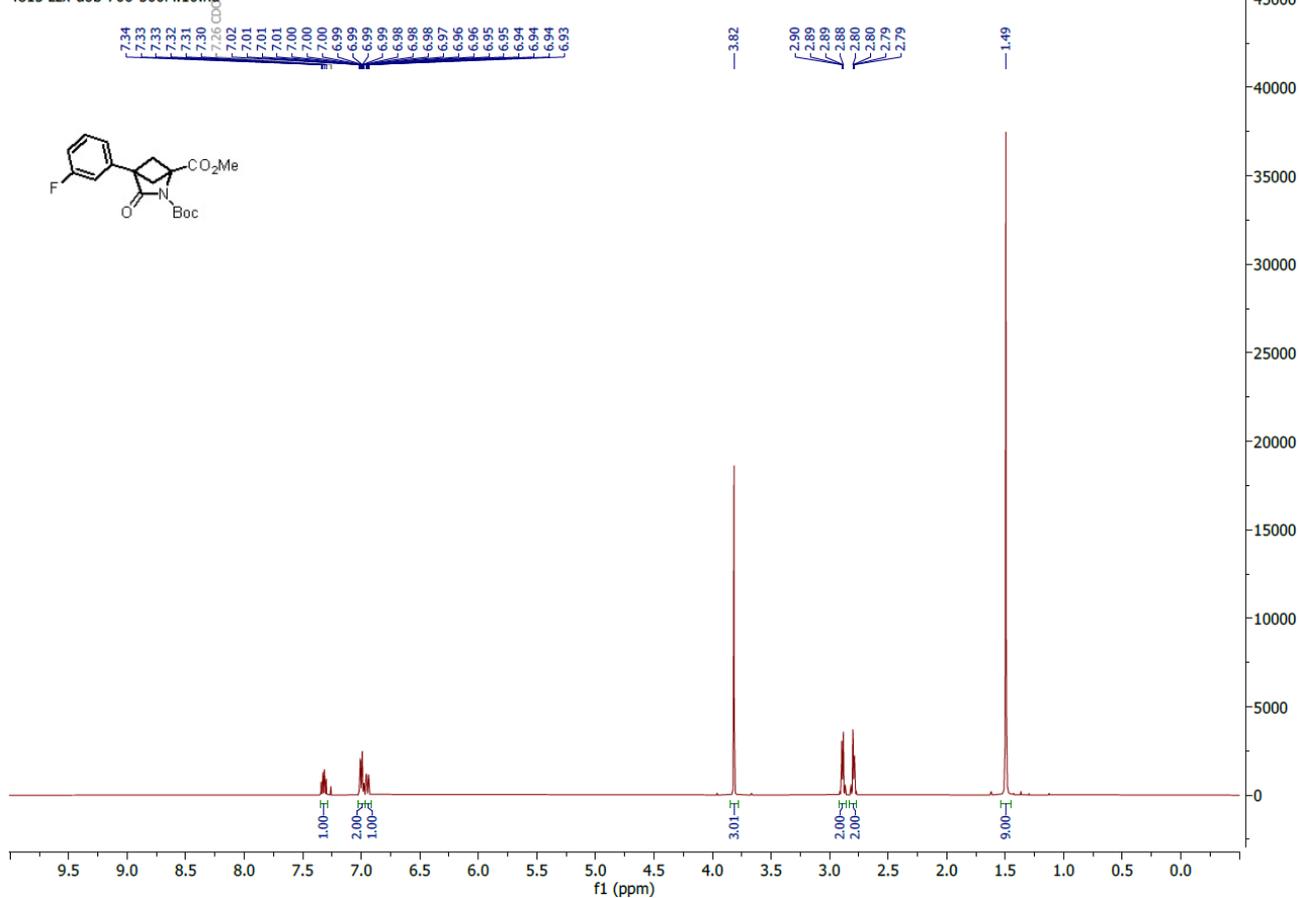
4814 zzx-uob-705-500M.11.fid



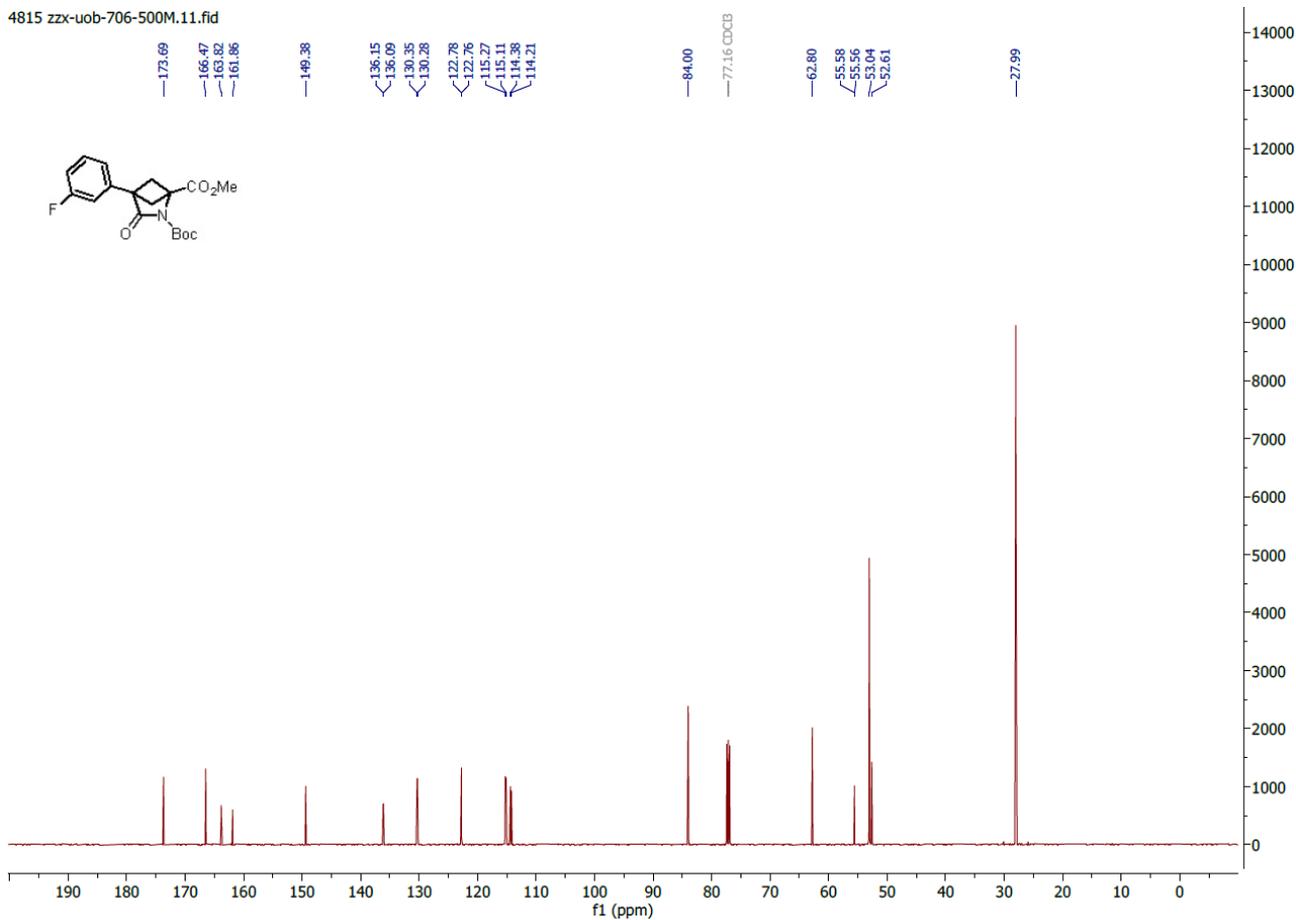


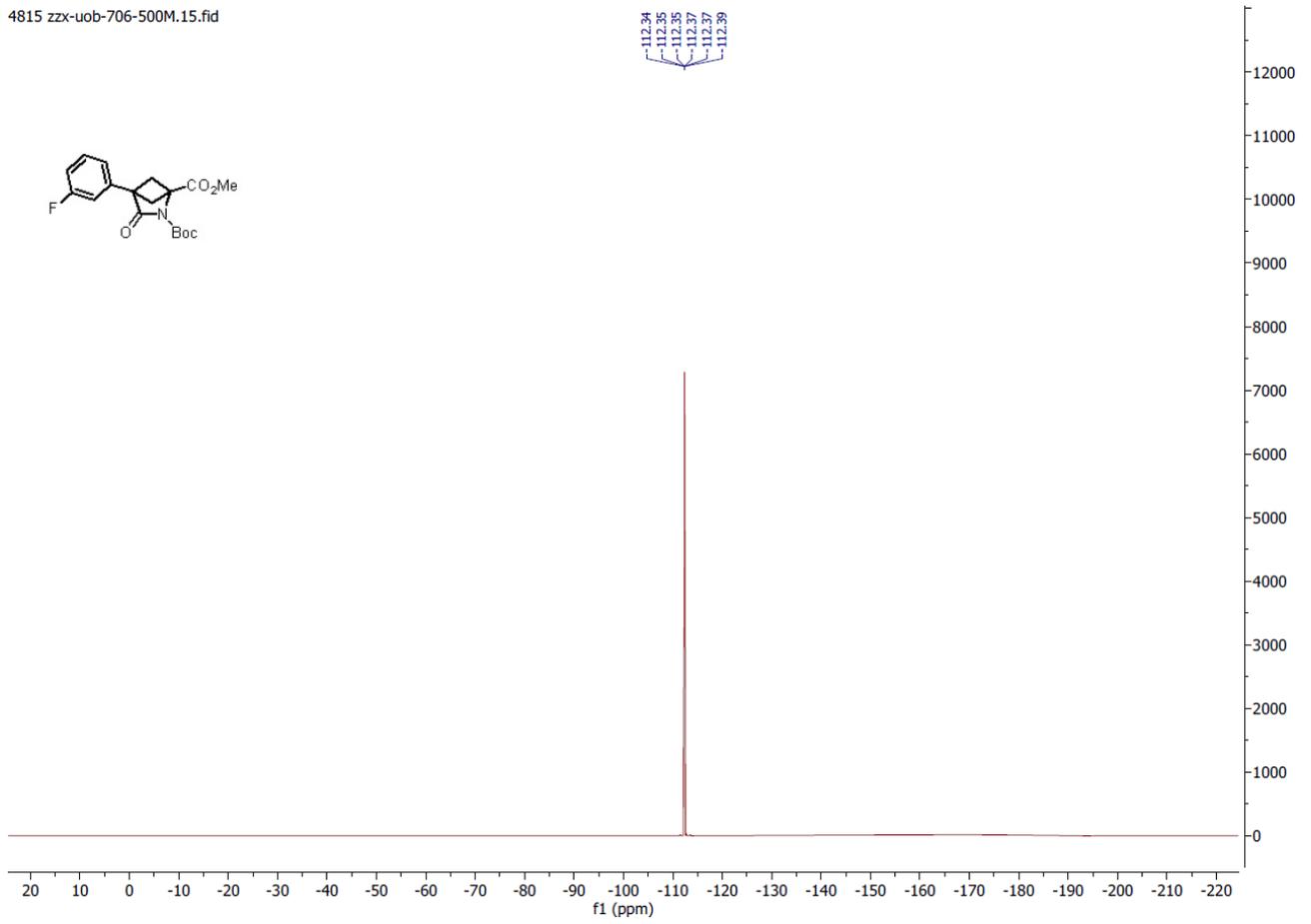
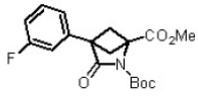
Compound 8c

4815 zzx-uob-706-500M.10.fid



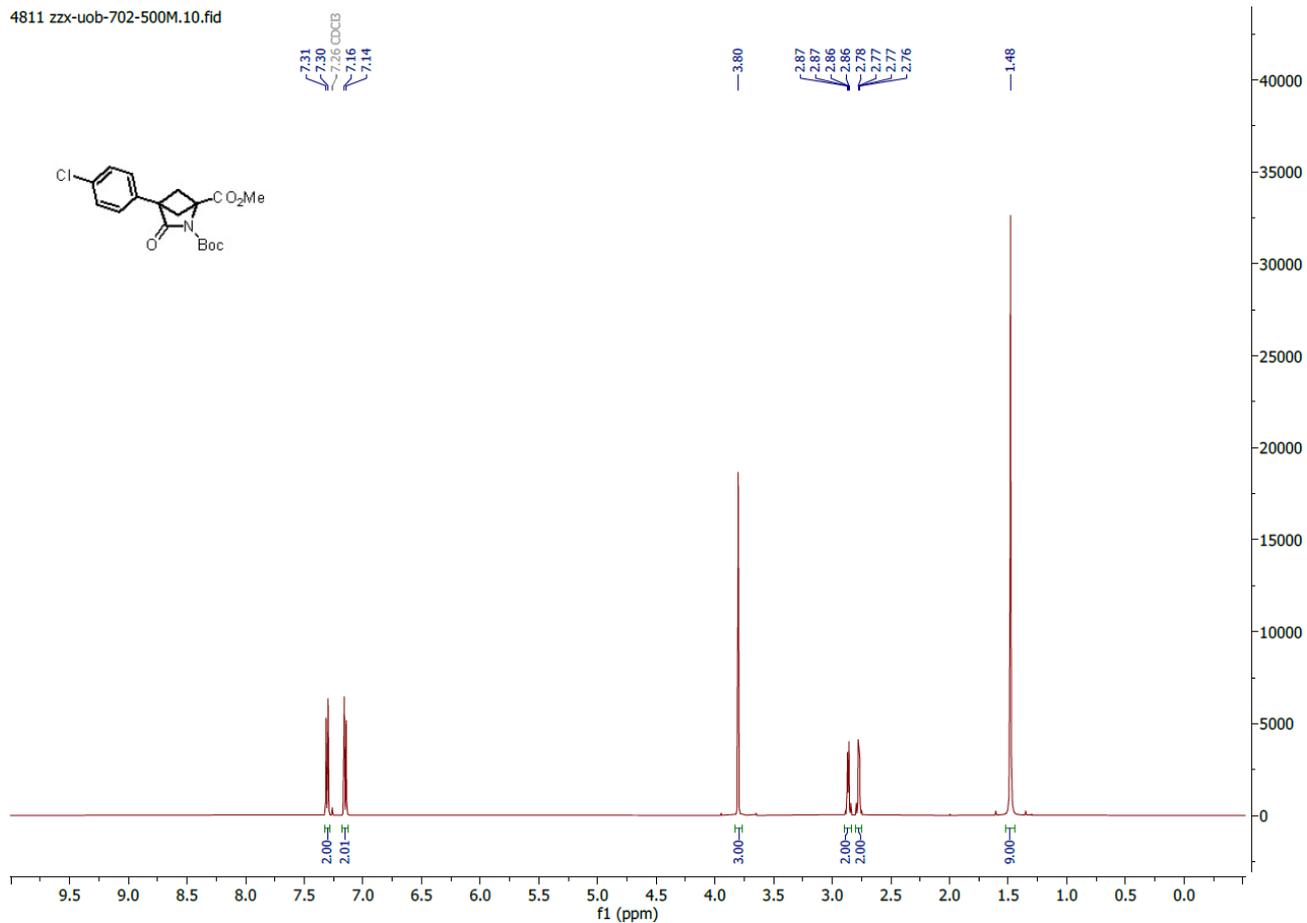
4815 zzx-uob-706-500M.11.fid



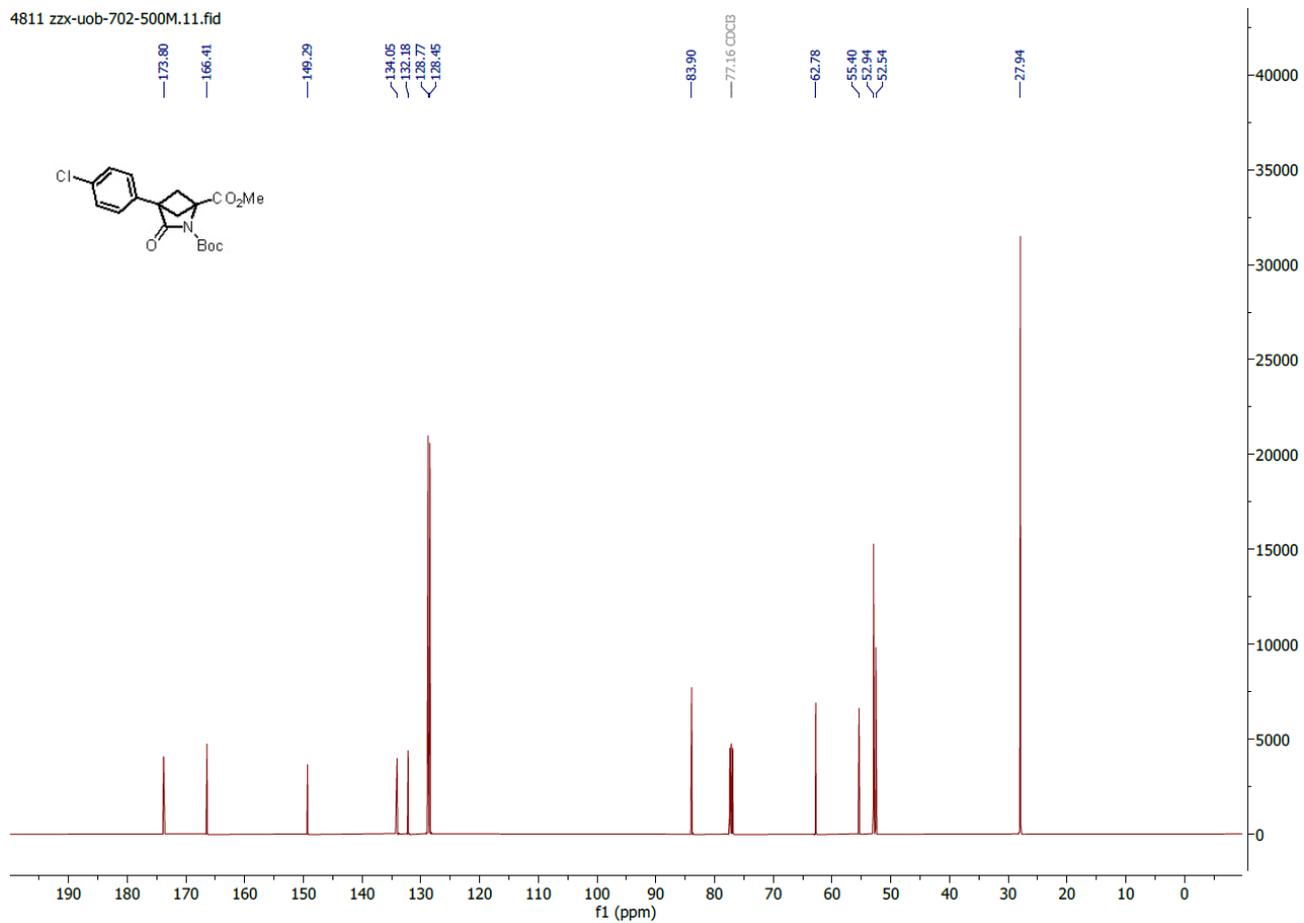


Compound 8d

4811 zzx-uob-702-500M.10.fid

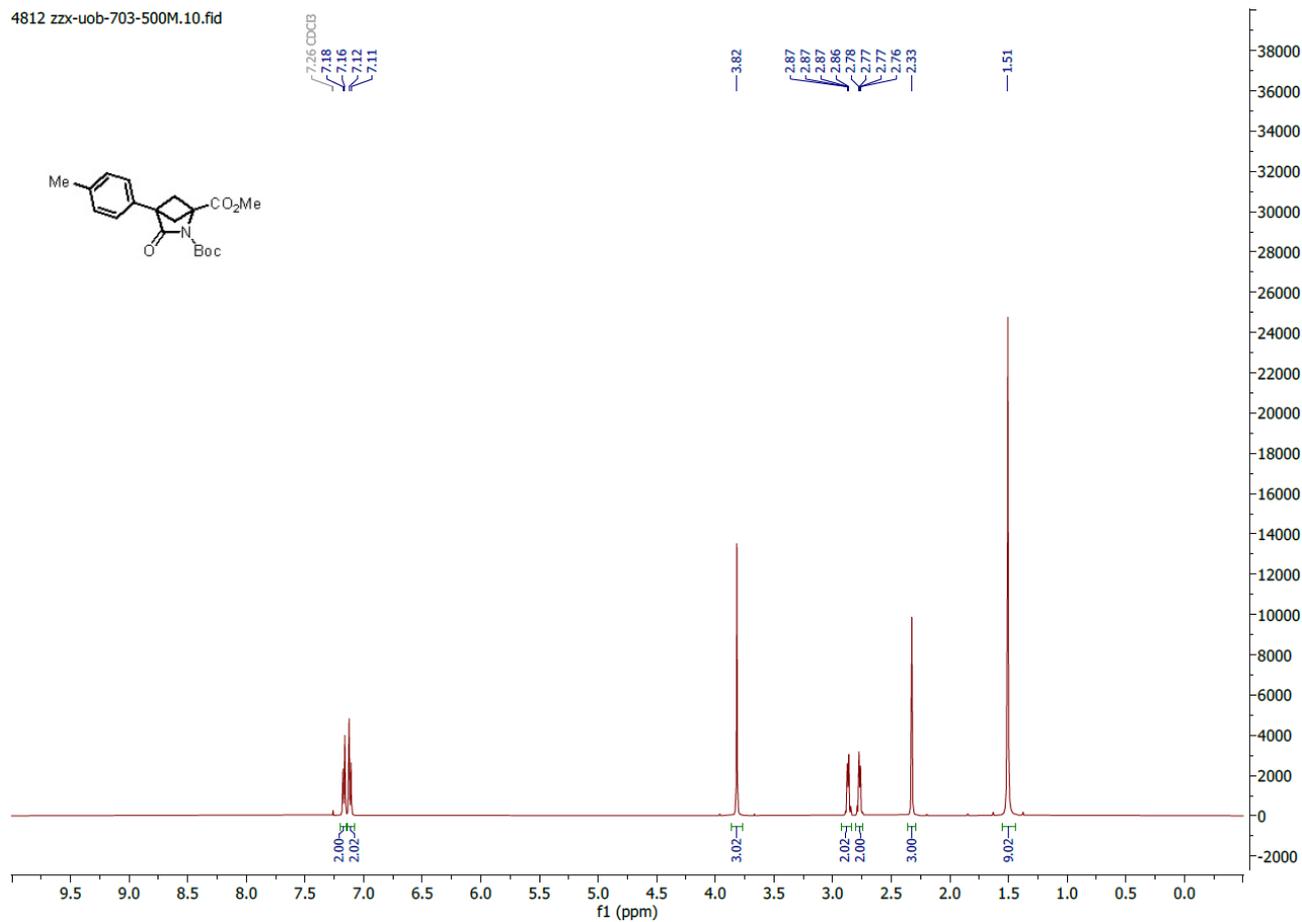


4811 zzx-uob-702-500M.11.fid

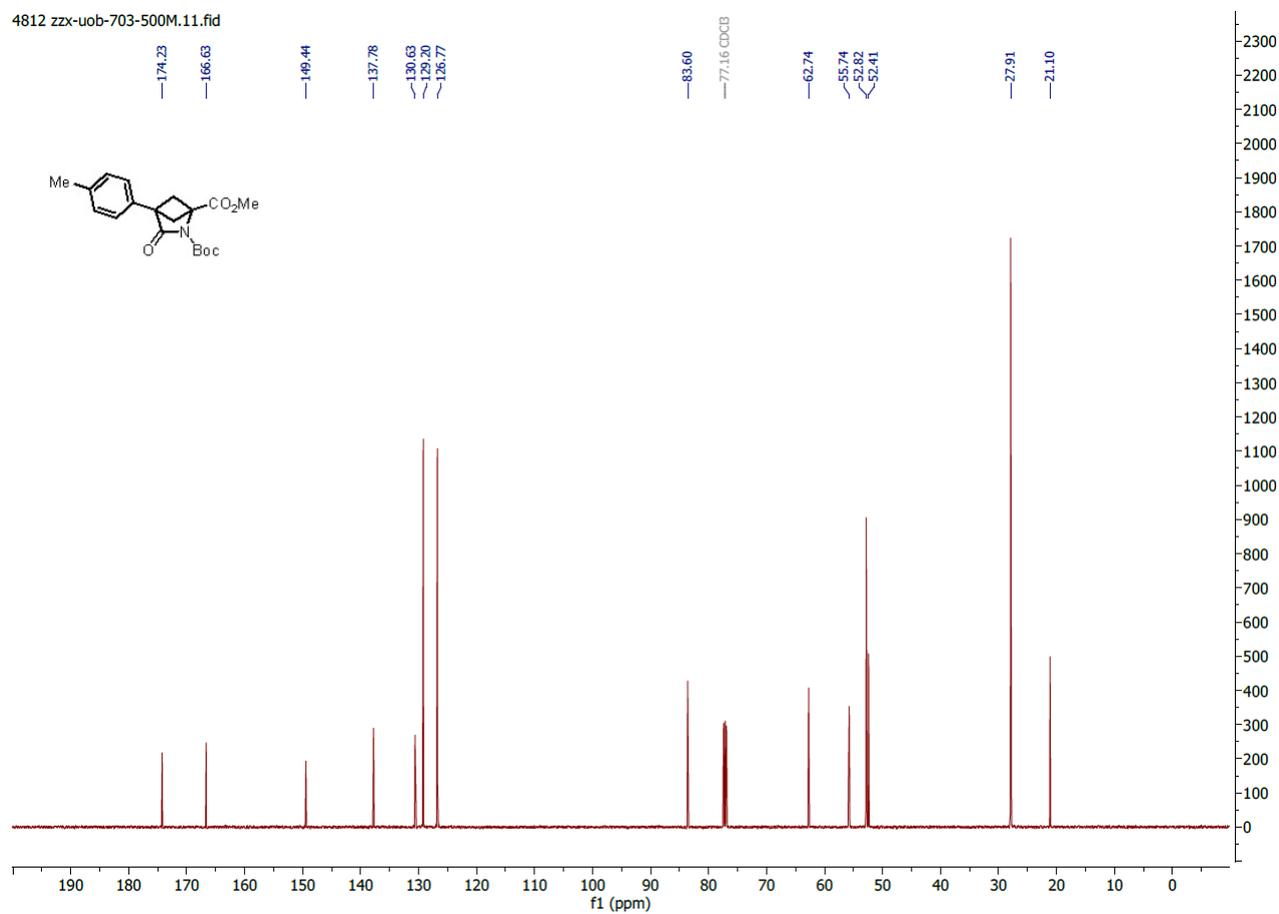


Compound 8e

4812 zzx-uob-703-500M.10.fid

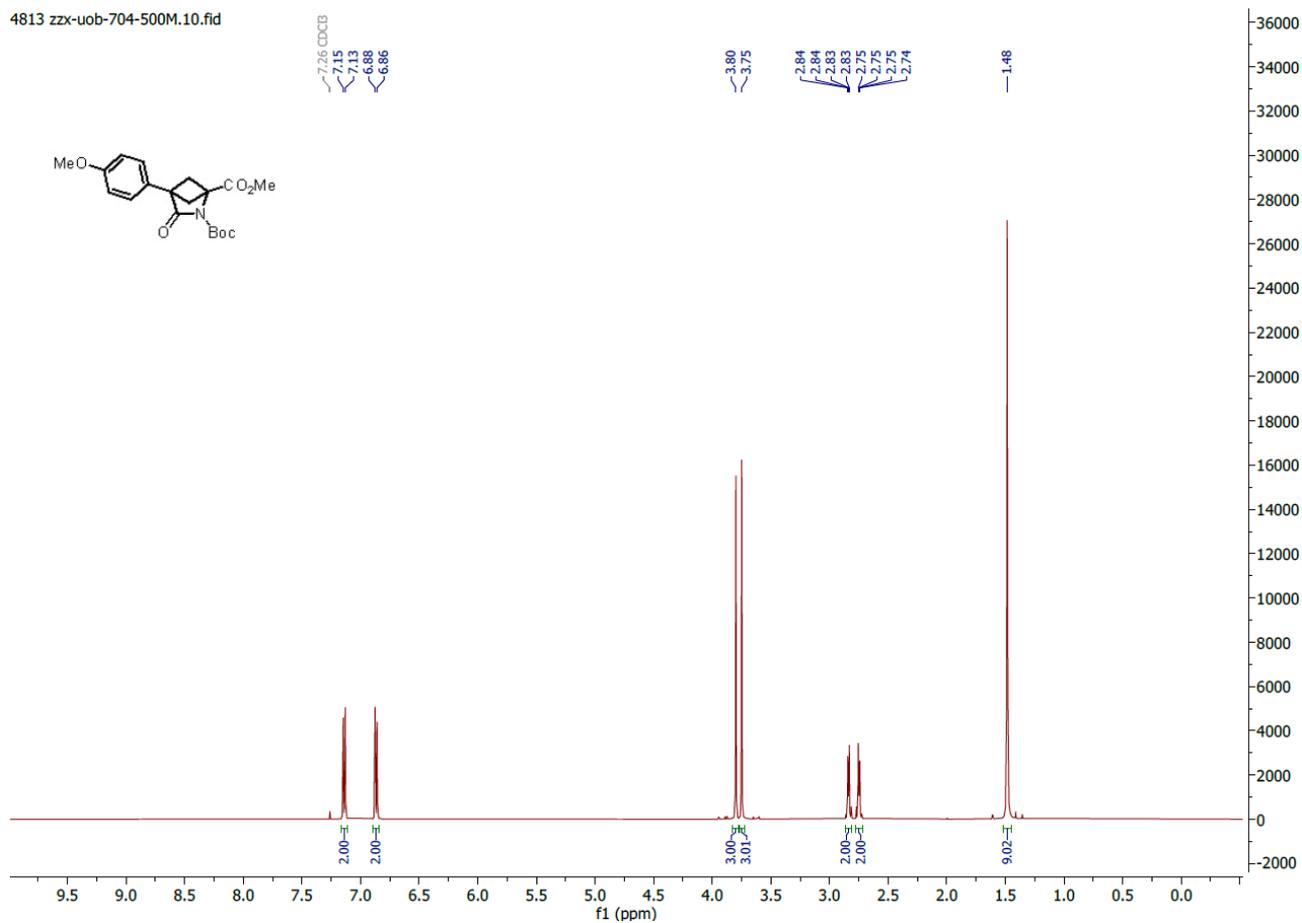


4812 zzx-uob-703-500M.11.fid

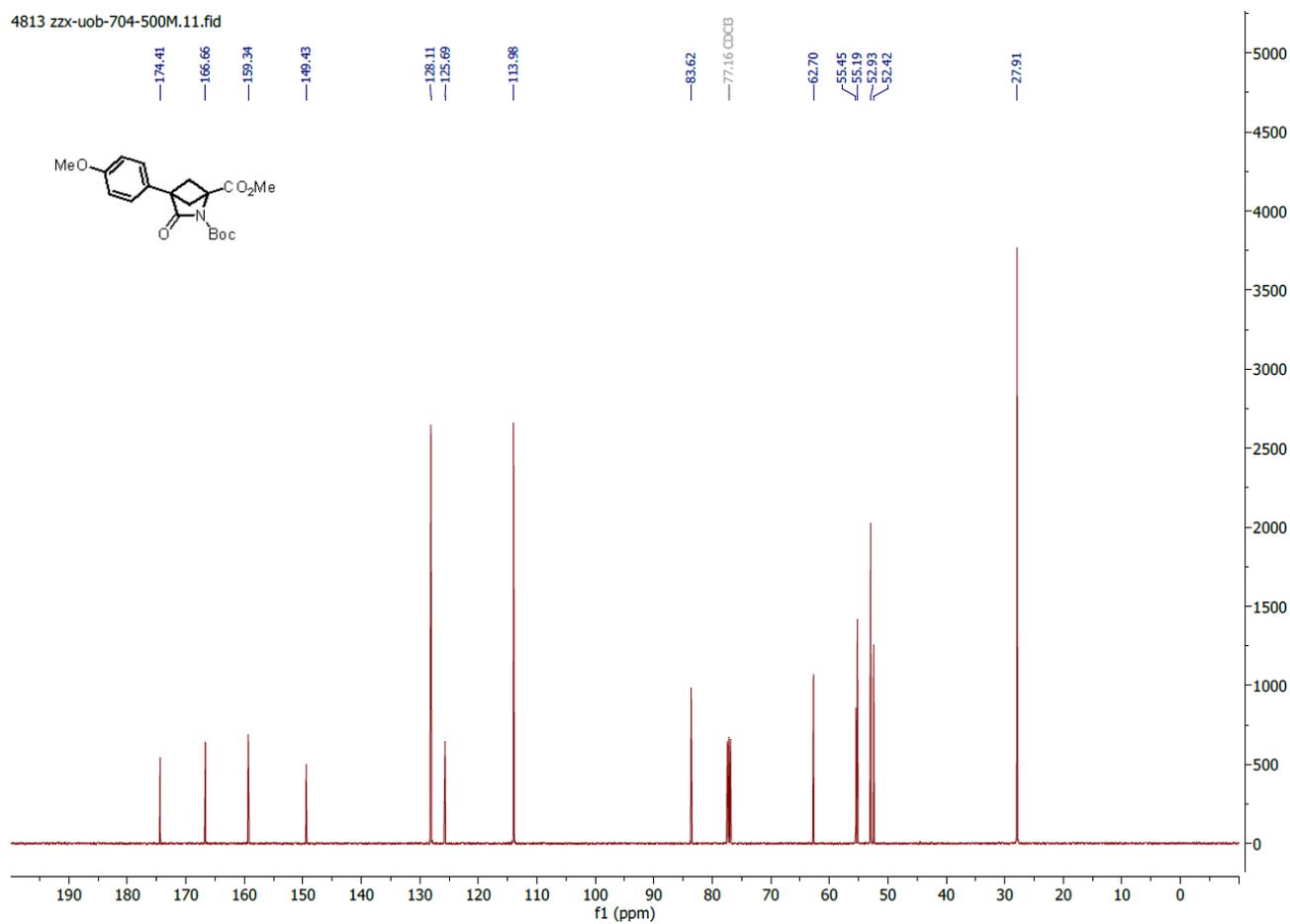


Compound 8f

4813 zzx-uob-704-500M.10.fid



4813 zzx-uob-704-500M.11.fid



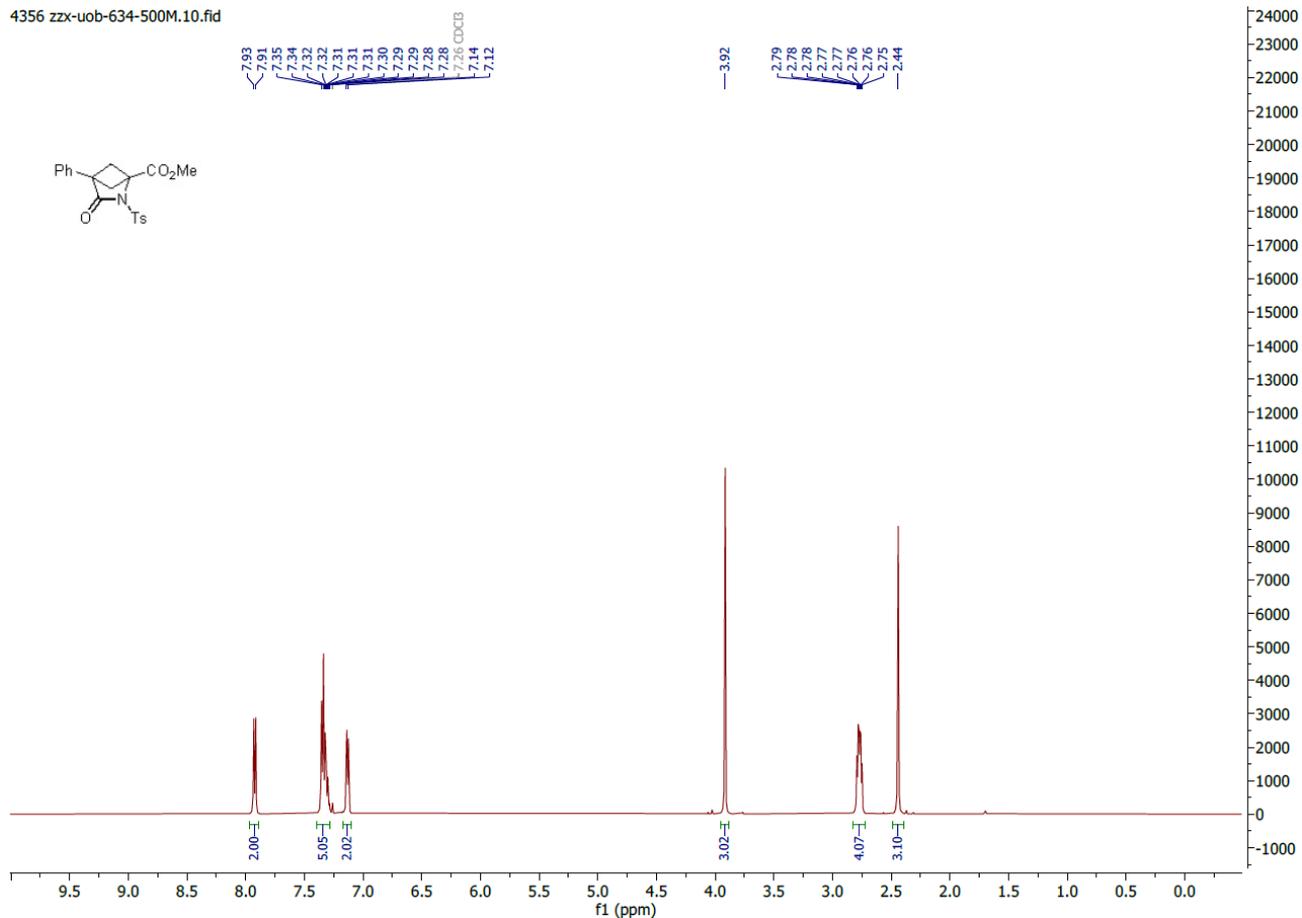
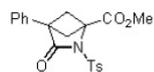
Compound 8g

4356 zzx-uob-634-500M.10.fid

7.93
7.91
7.35
7.34
7.32
7.32
7.31
7.31
7.30
7.30
7.29
7.28
7.28
7.26 CDCl₃
7.14
7.12

3.92

2.79
2.78
2.78
2.77
2.76
2.76
2.75
2.44



4356 zzx-uob-634-500M.11.fid

174.43
166.04

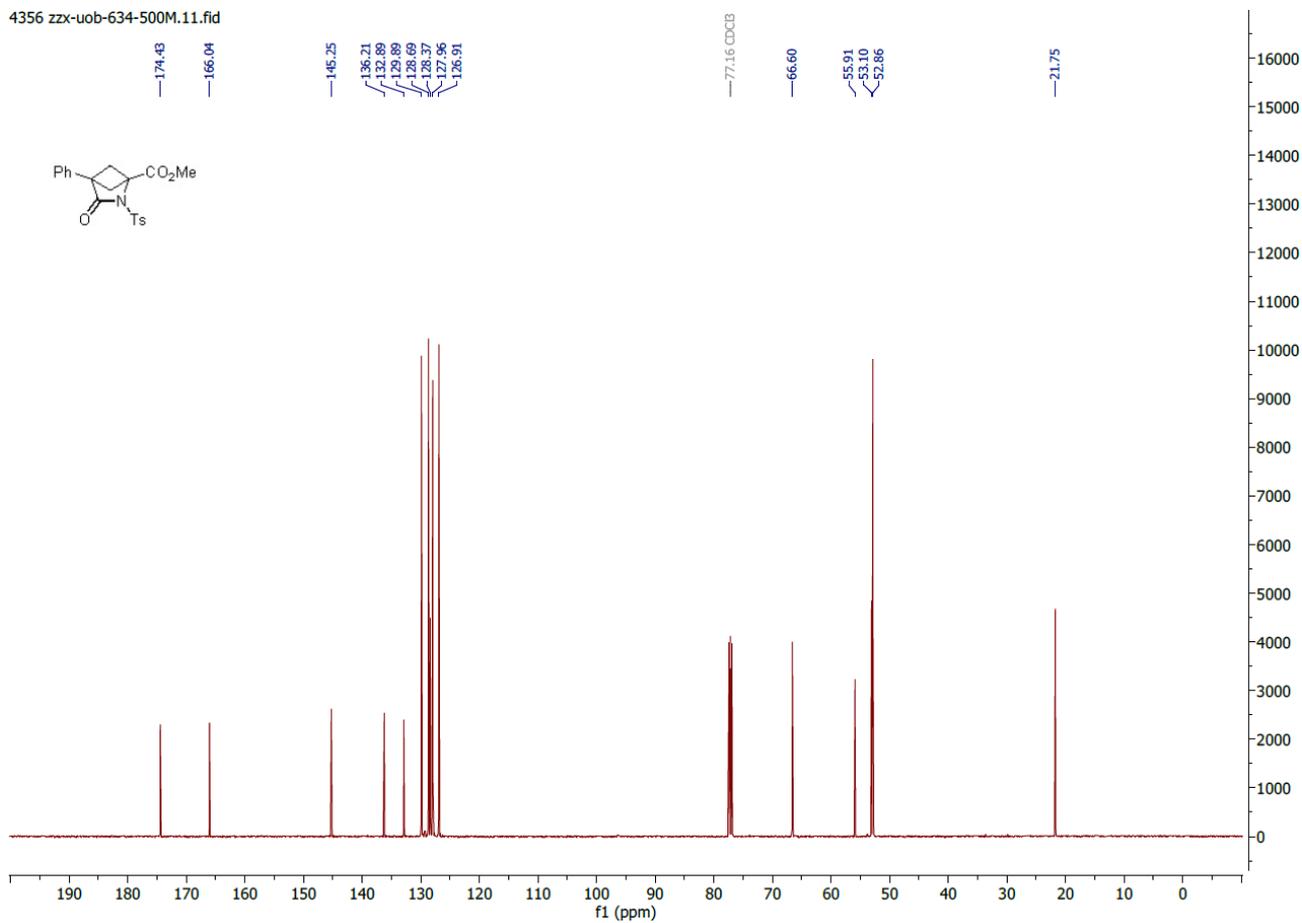
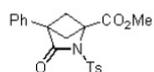
145.25
136.21
132.88
129.88
128.69
128.37
127.96
126.91

77.16 CDCl₃

66.60

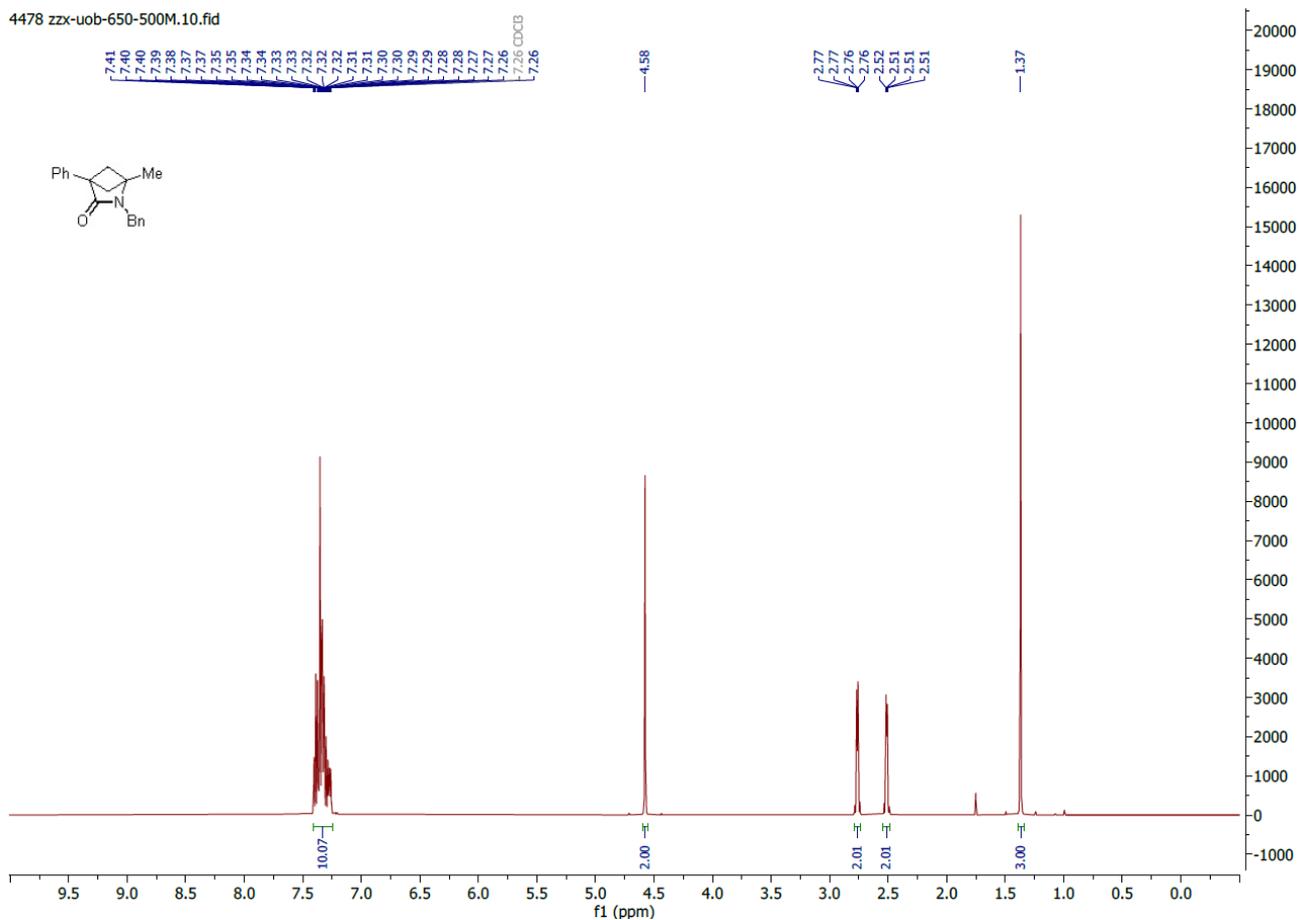
55.91
53.10
52.86

21.75

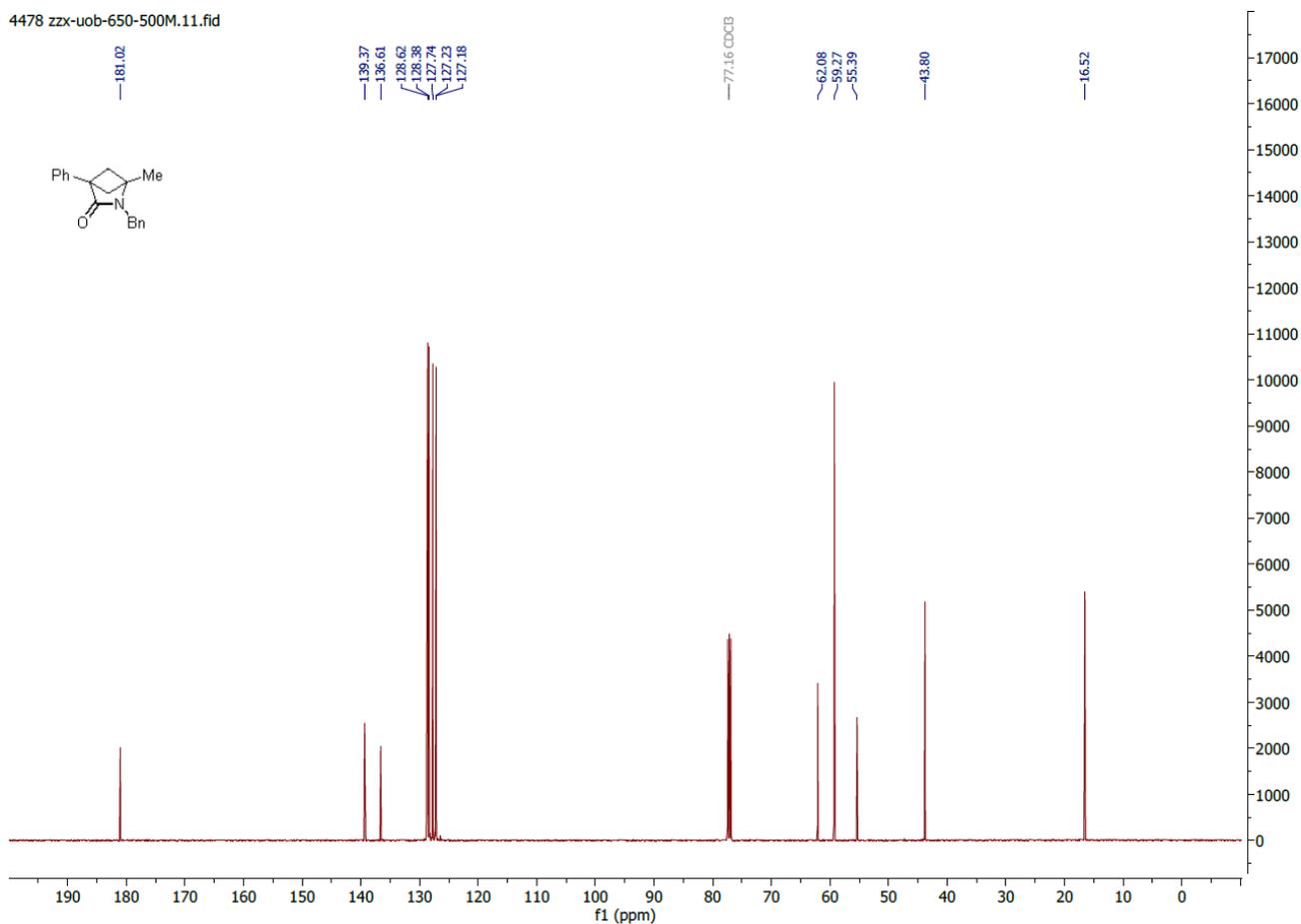


Compound 8h

4478 zzx-uob-650-500M.10.fid



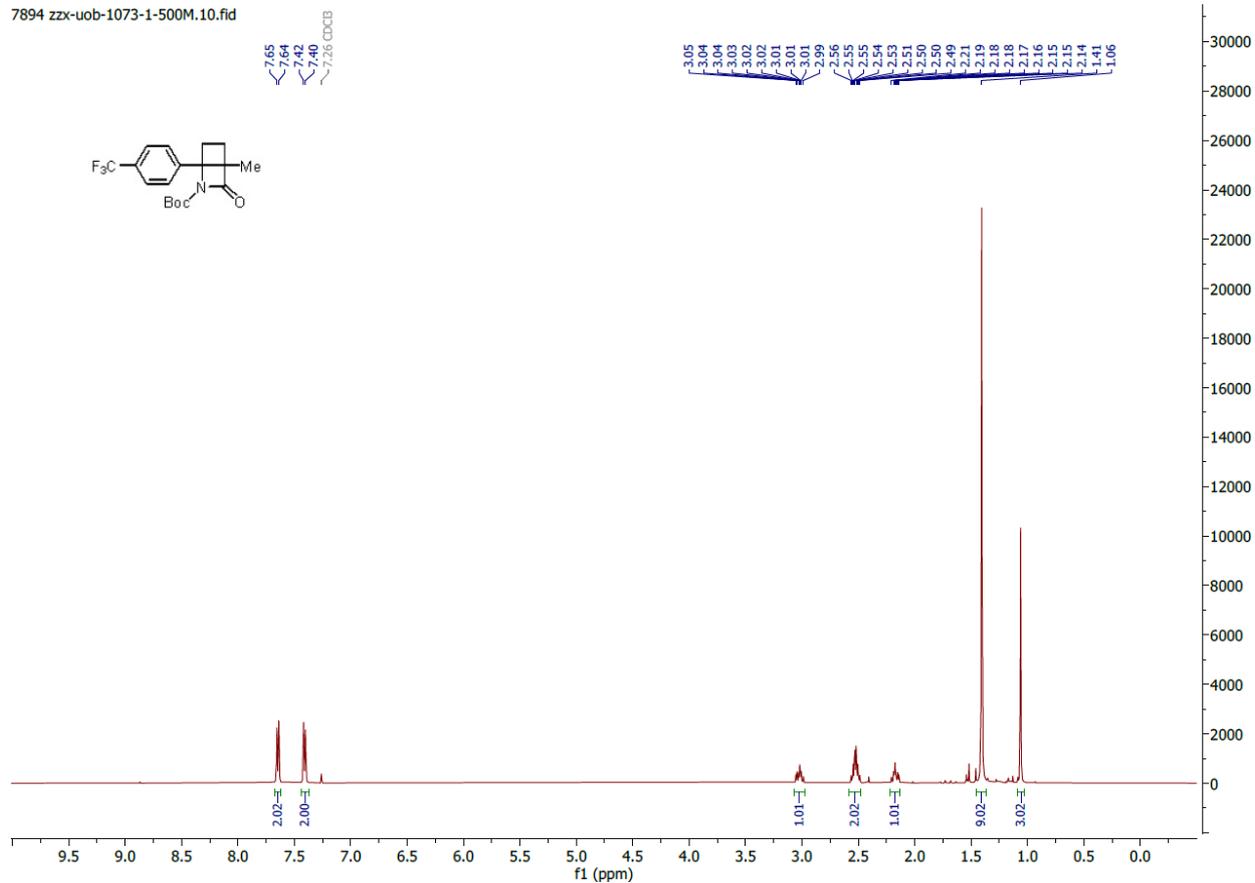
4478 zzx-uob-650-500M.11.fid



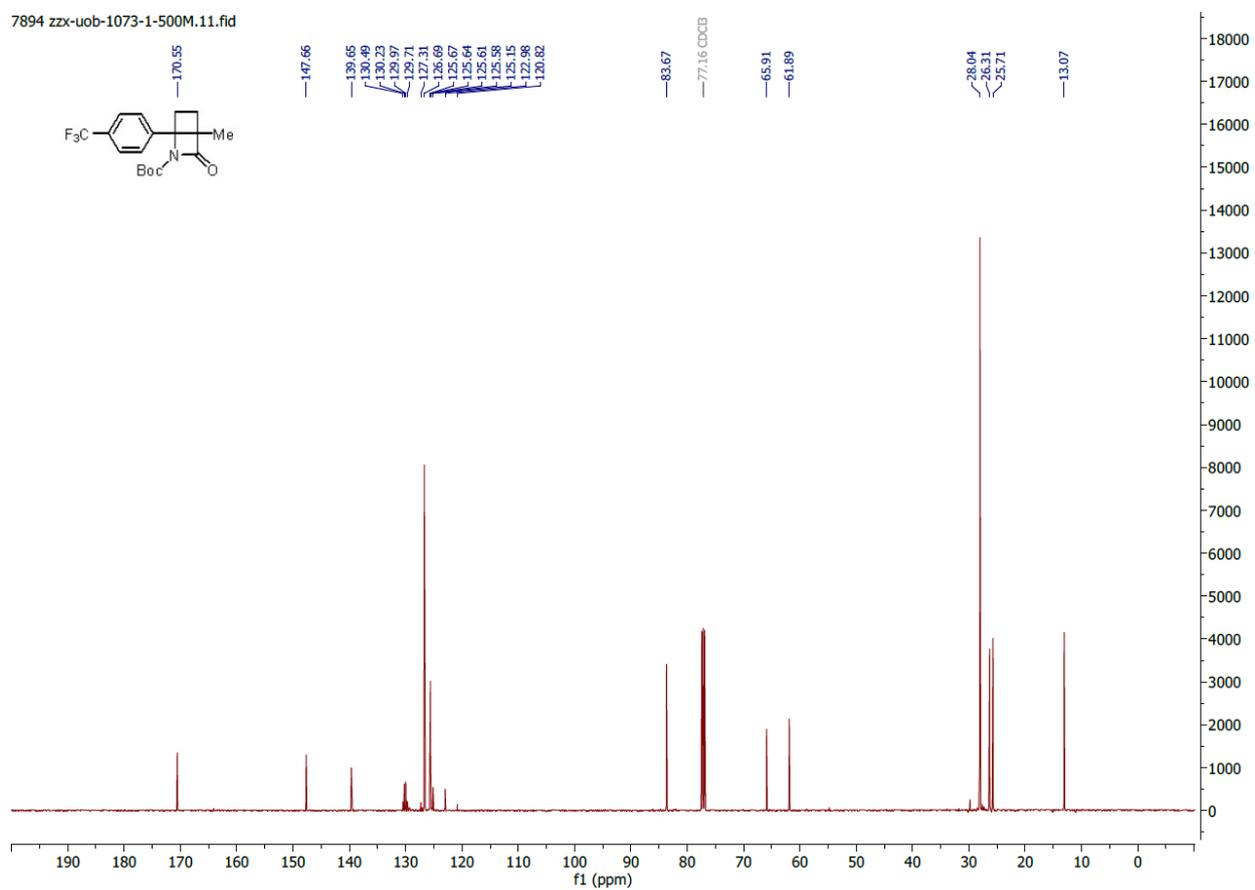
5.6 NMR Spectra of Products (5, 6 and 9)

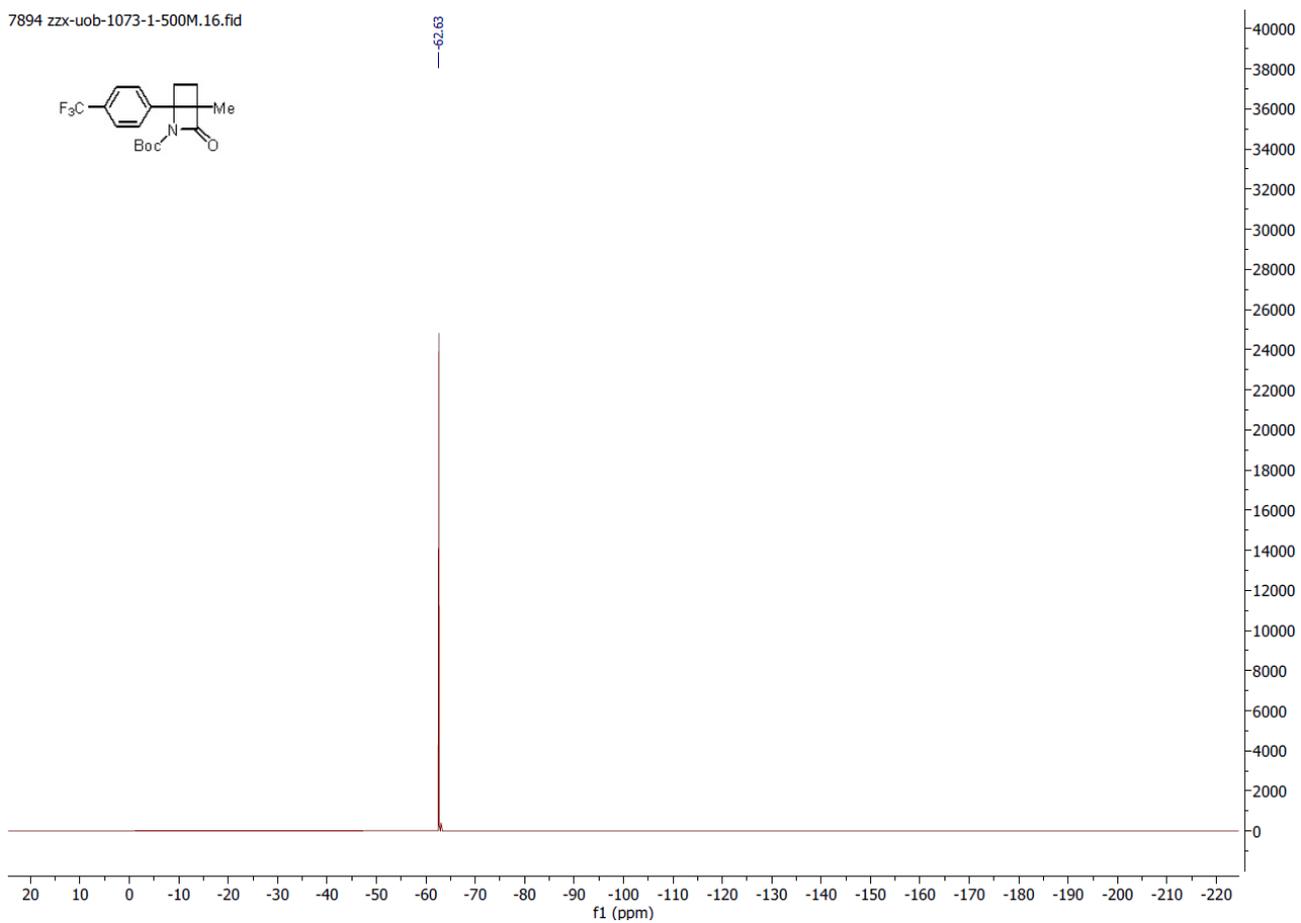
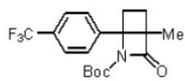
Compound 6a

7894 zzx-uob-1073-1-500M.10.fid



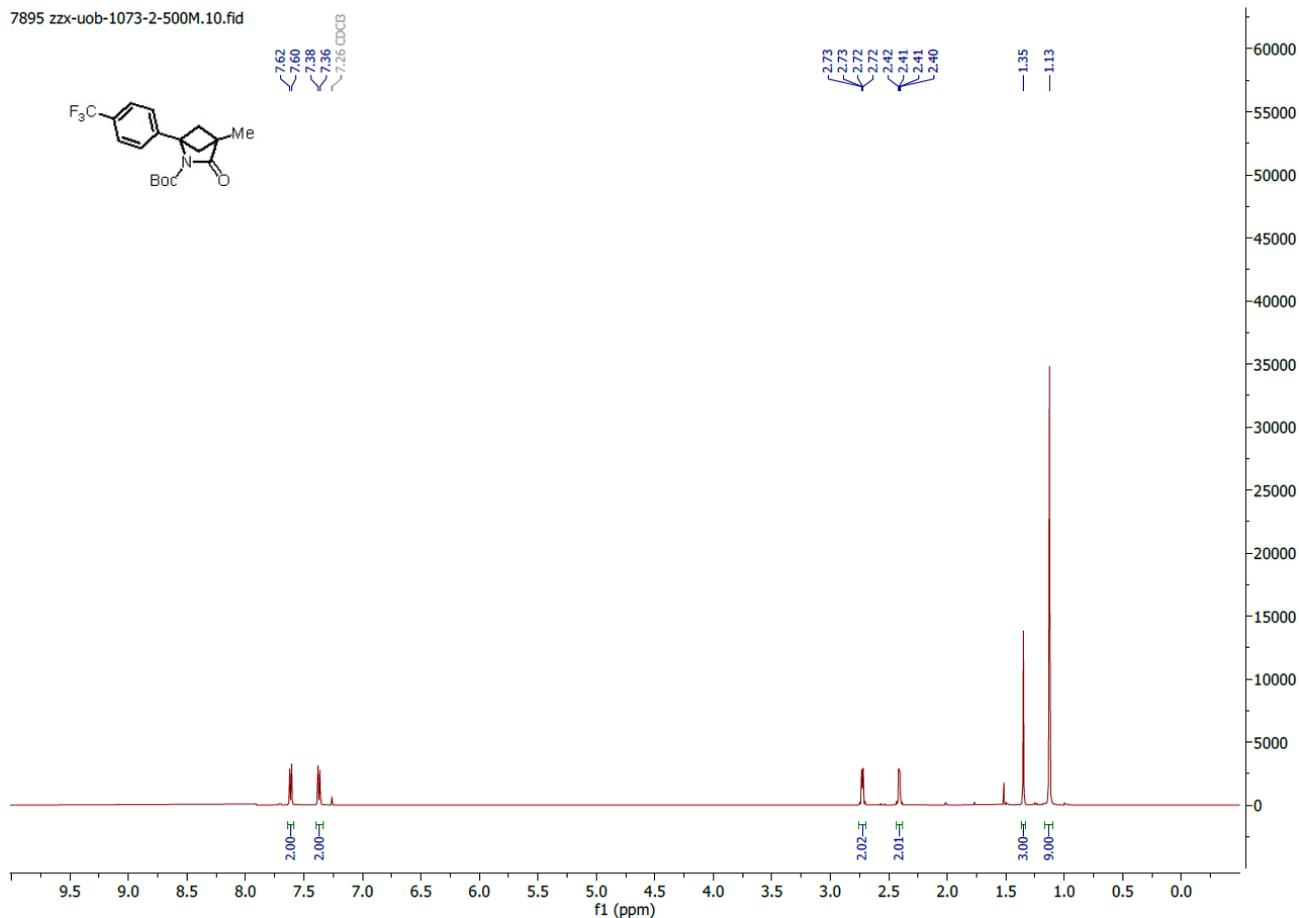
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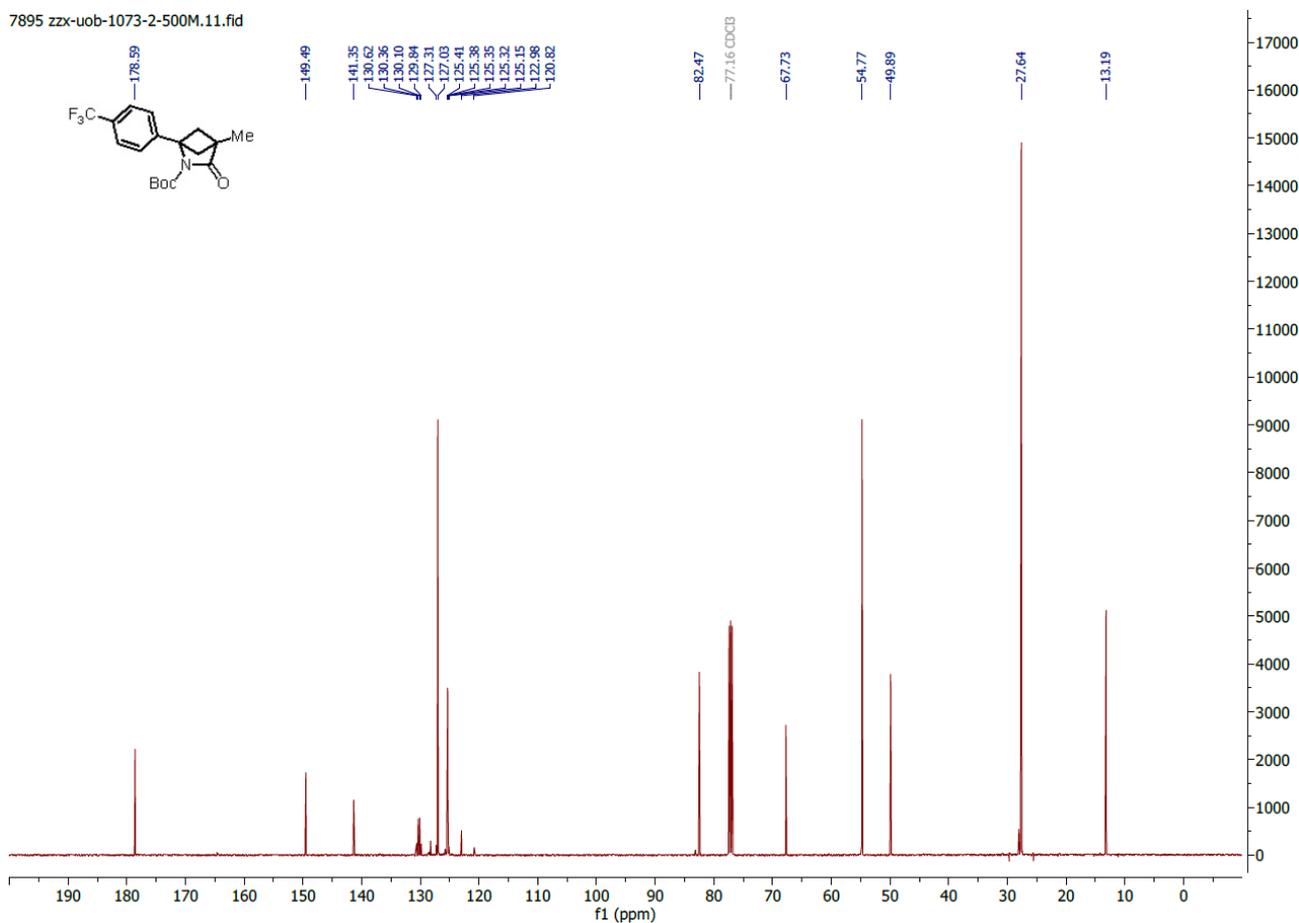


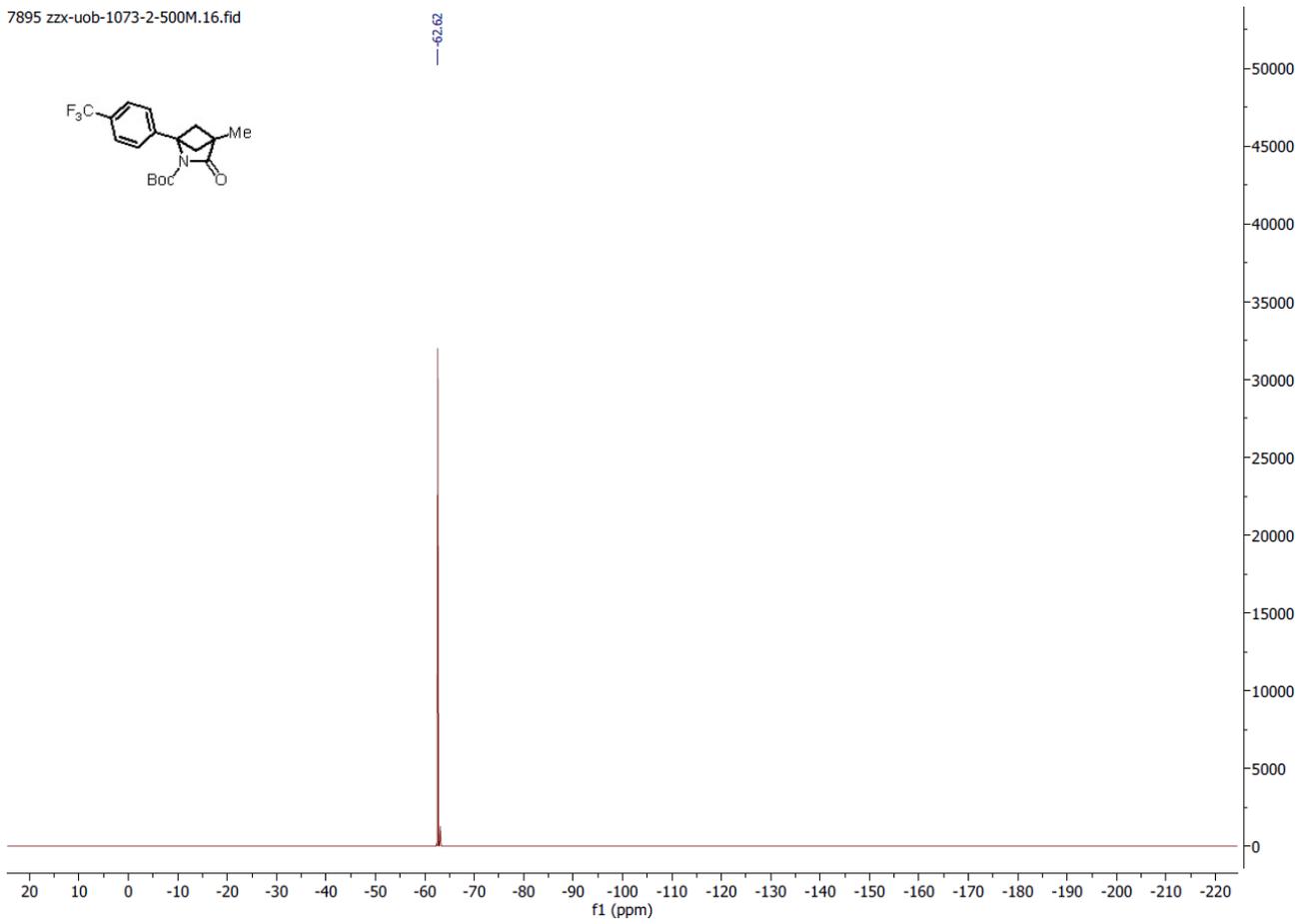
Compound 5a

7895 zzx-uob-1073-2-500M.10.fid



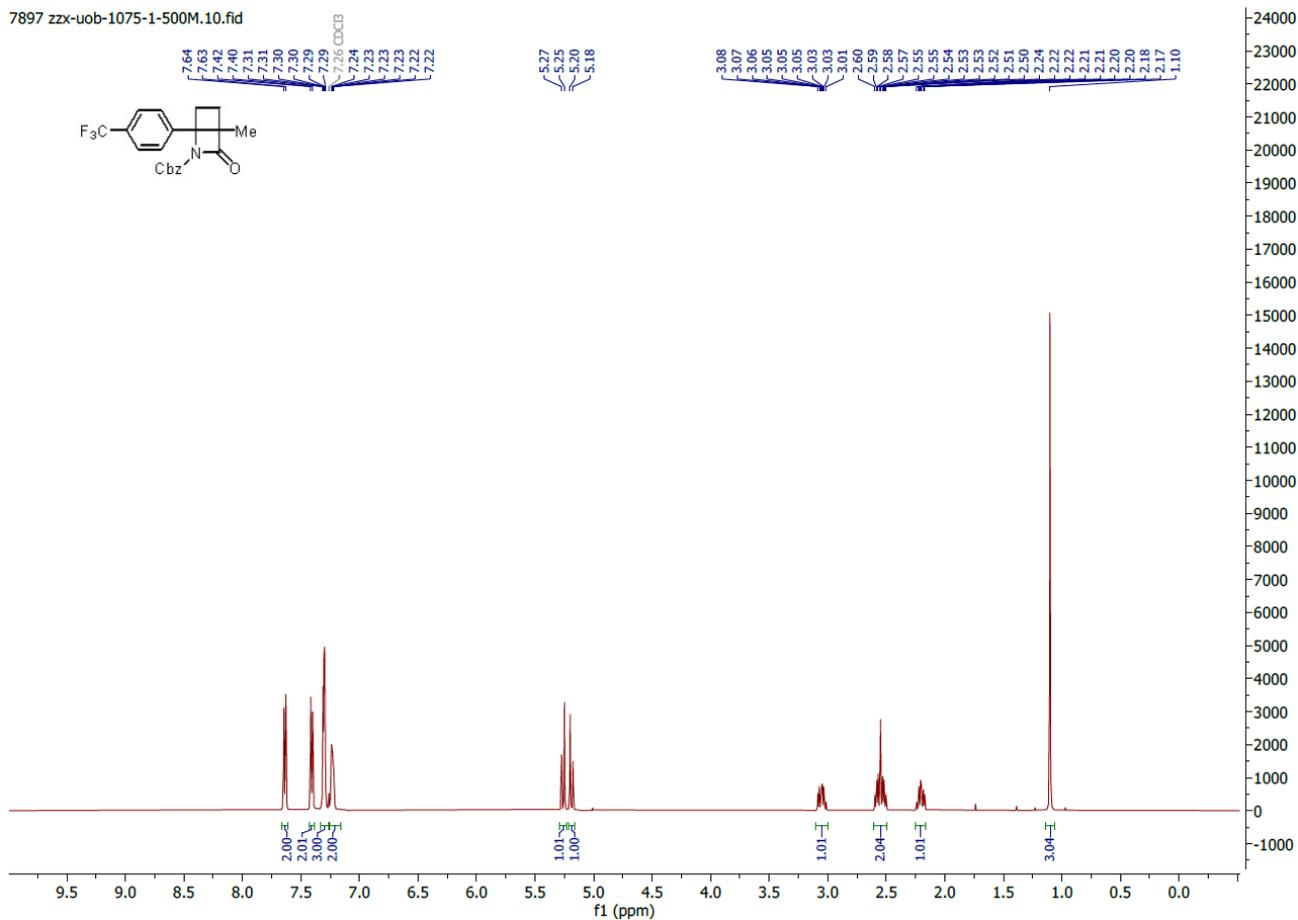
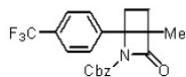
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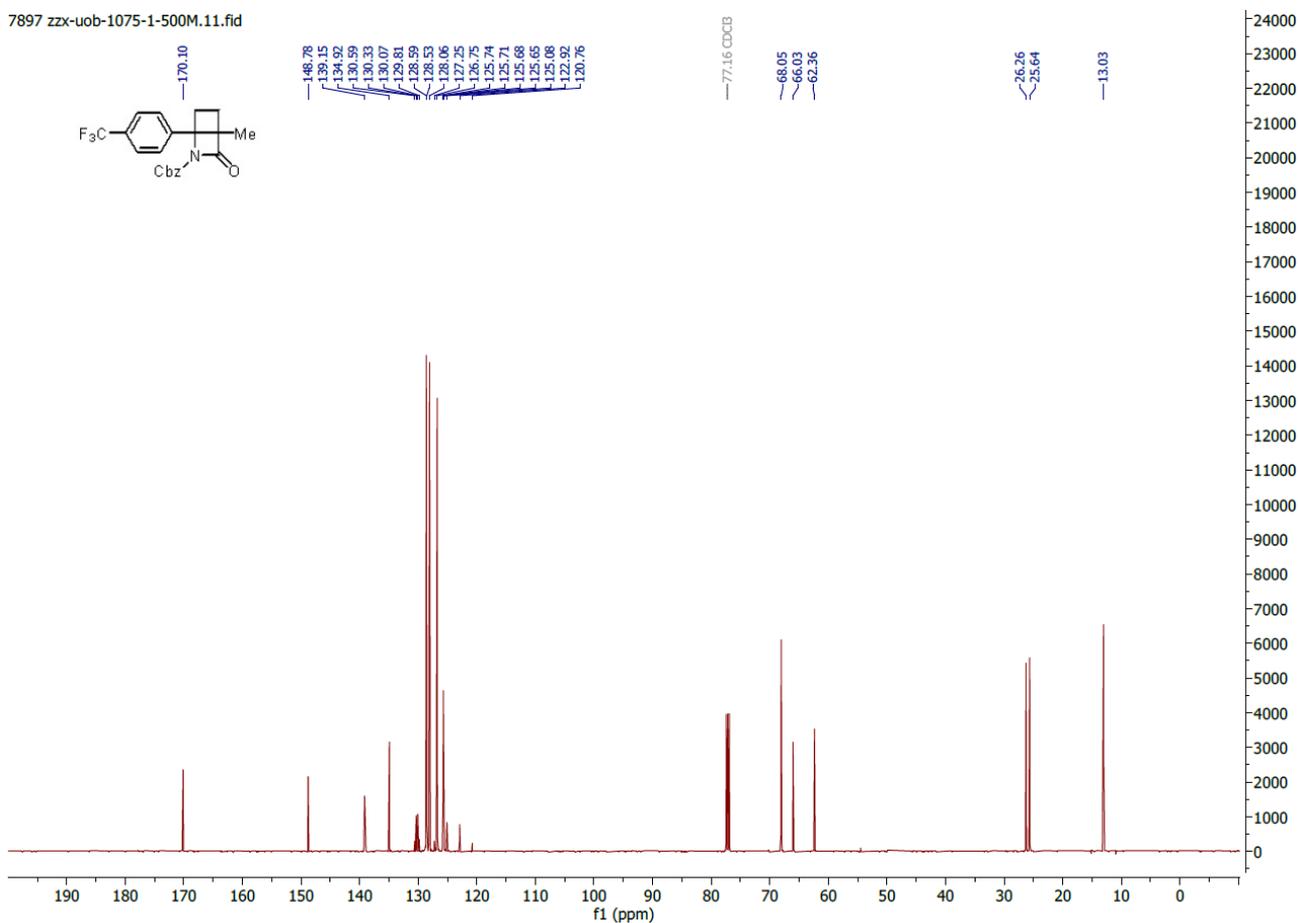
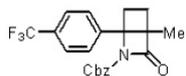


Compound 6b

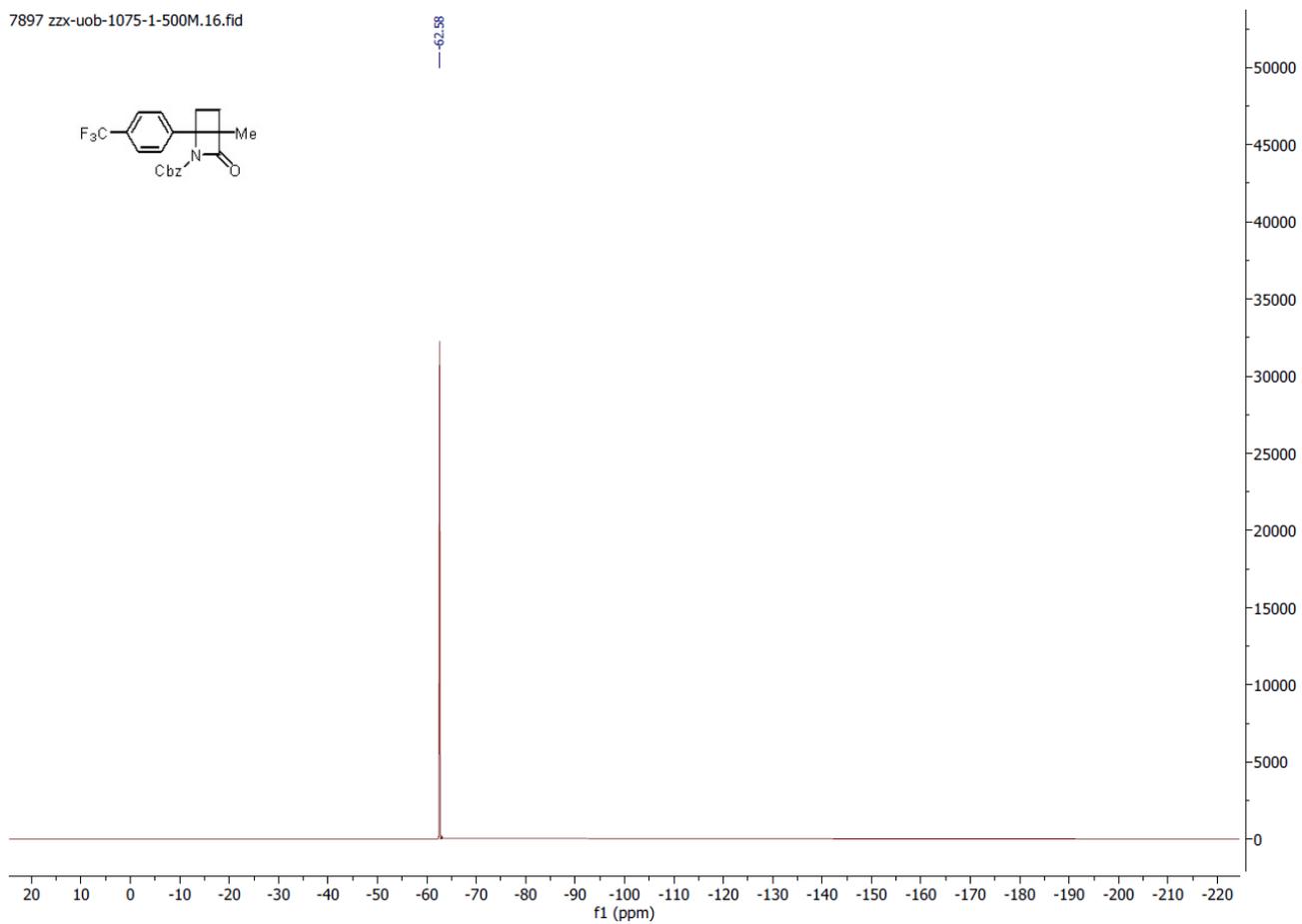
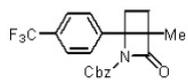
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7897 zzx-uob-1075-1-500M.11.fid

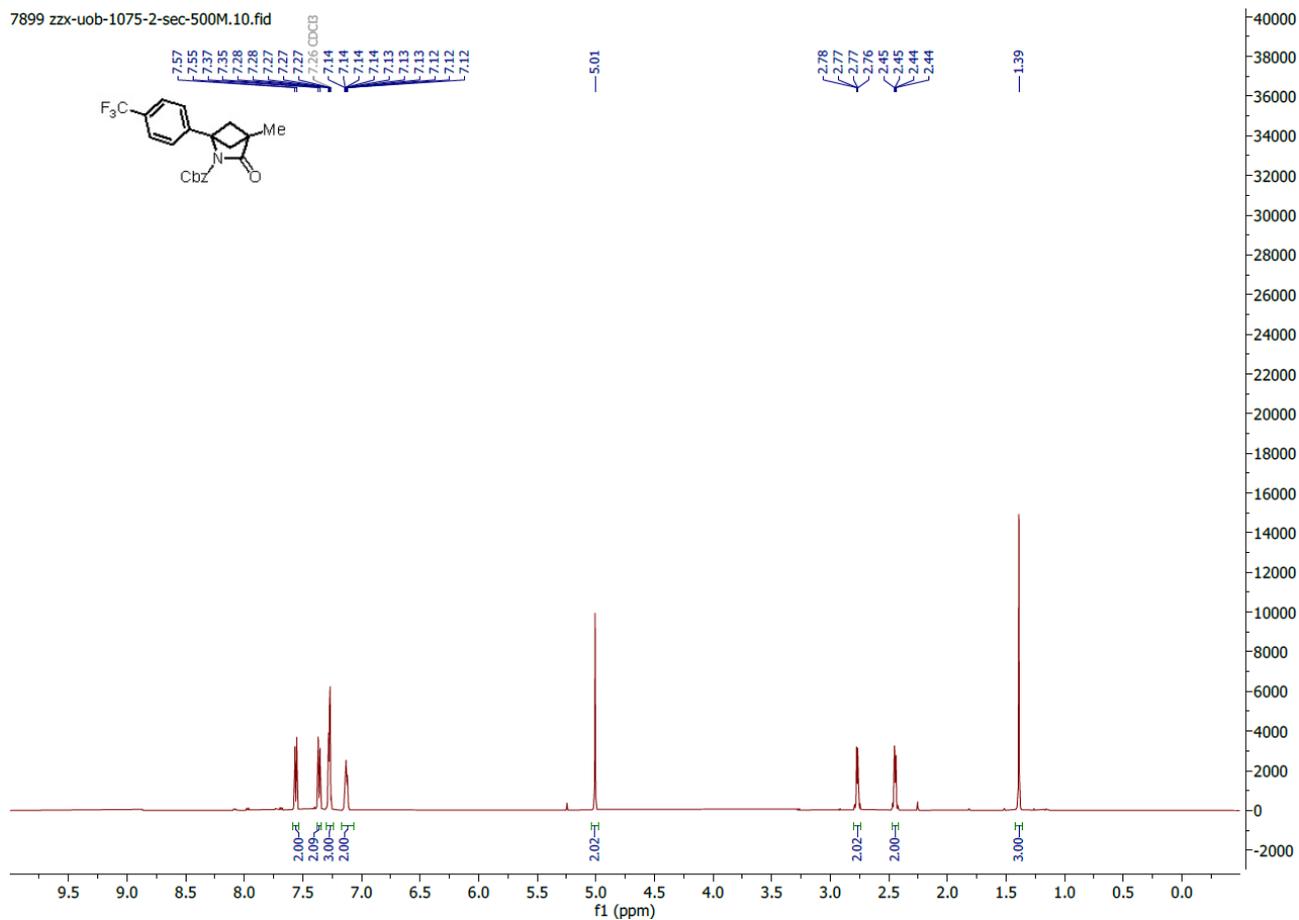


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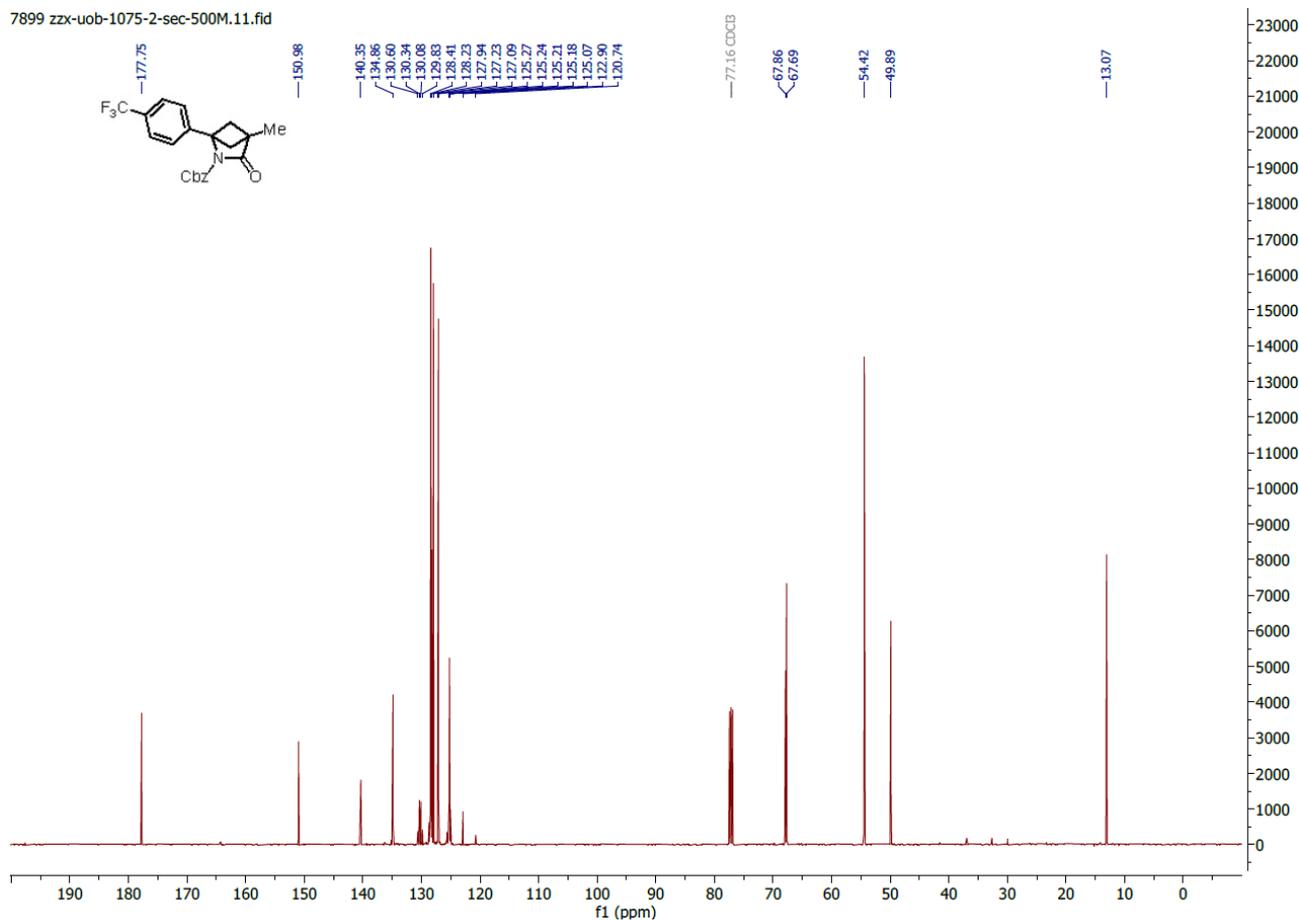


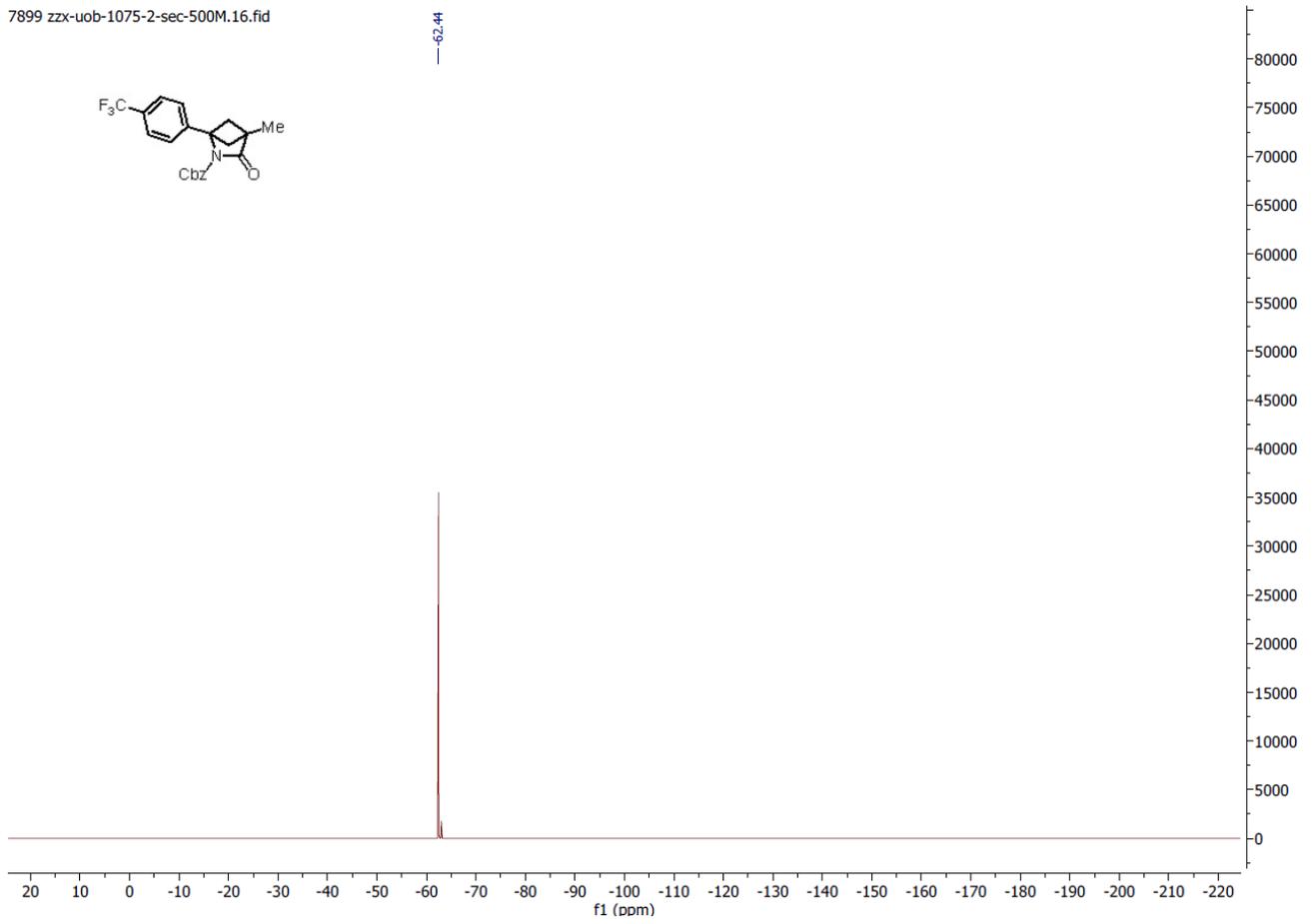
Compound 5b

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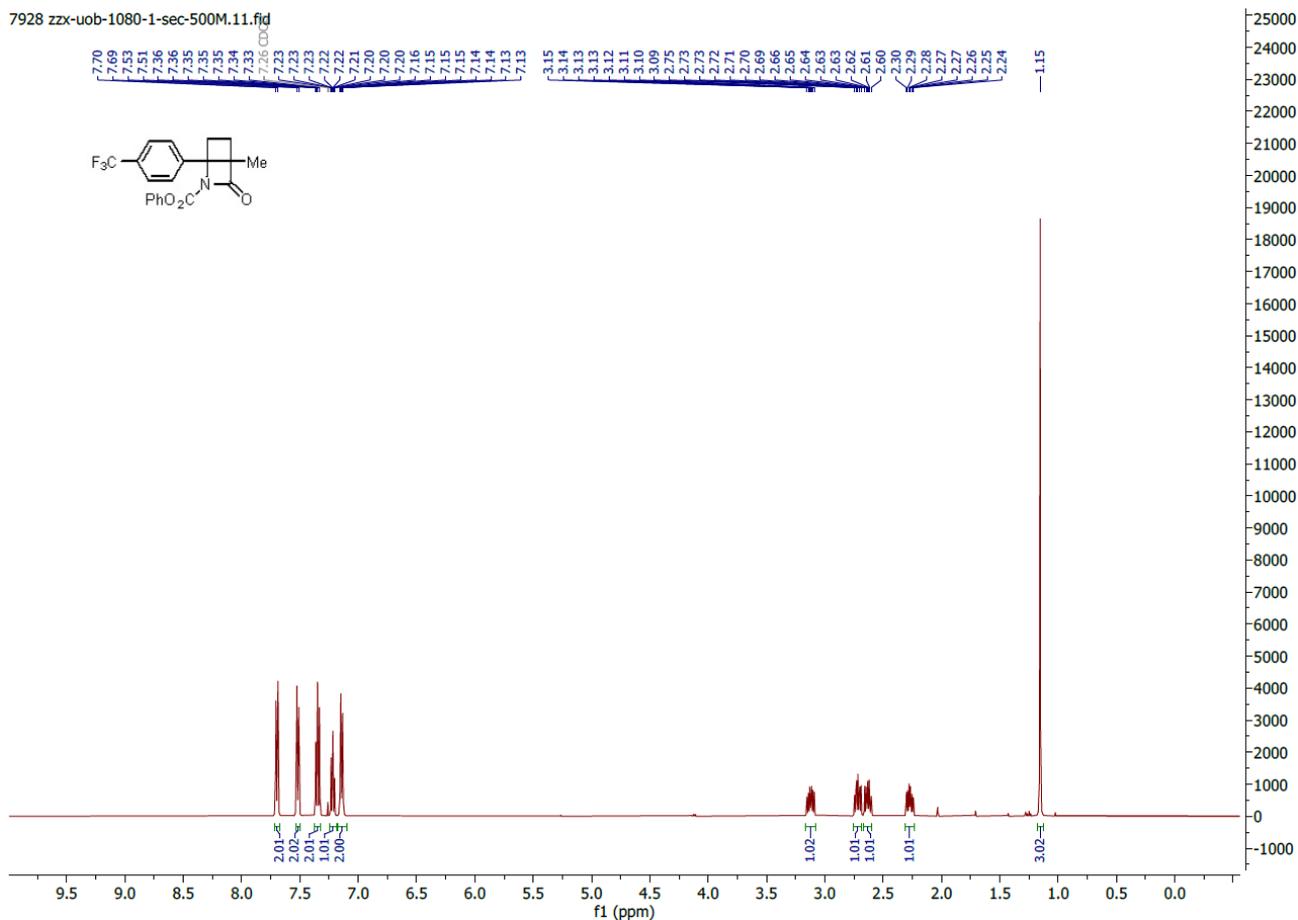
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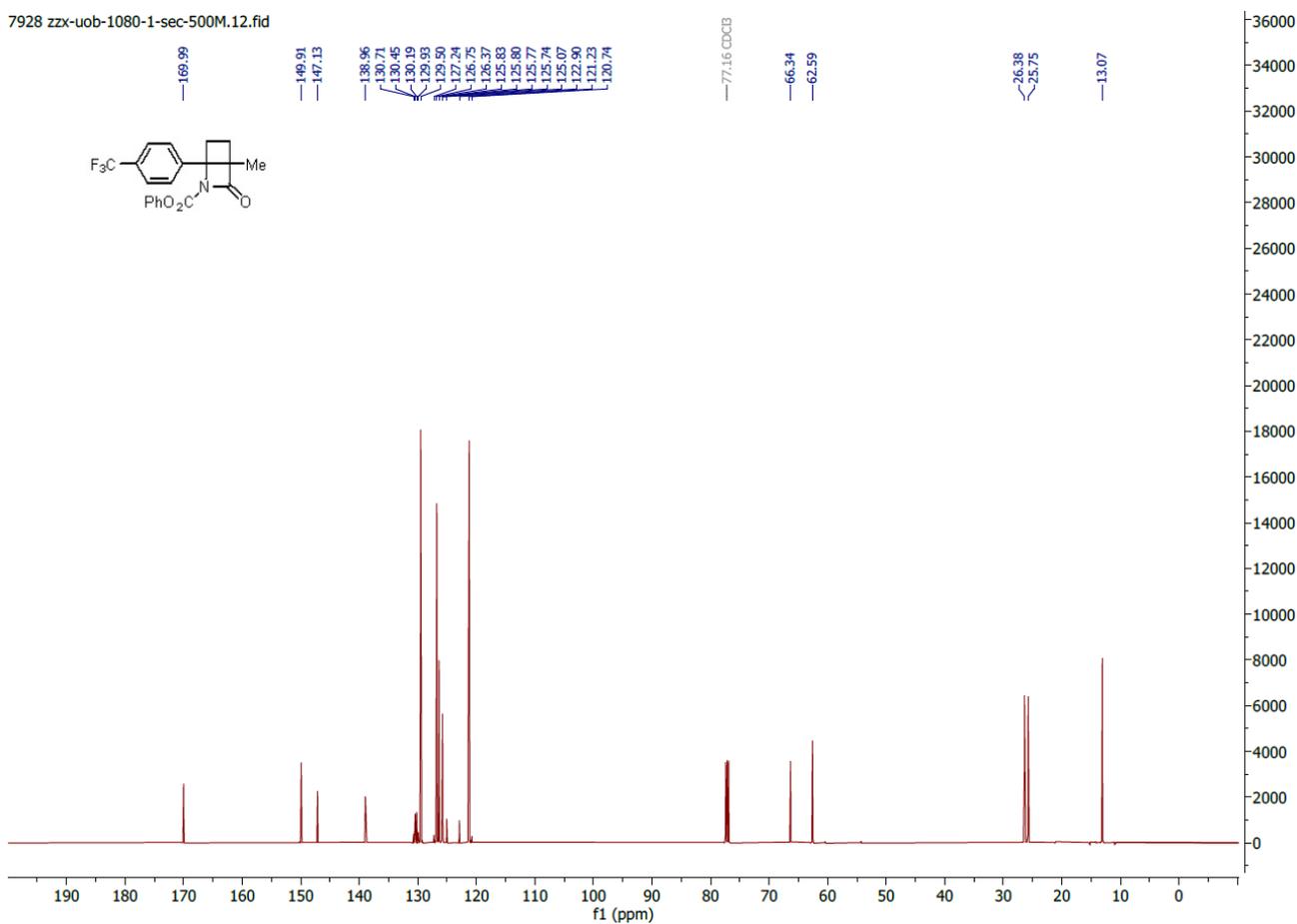


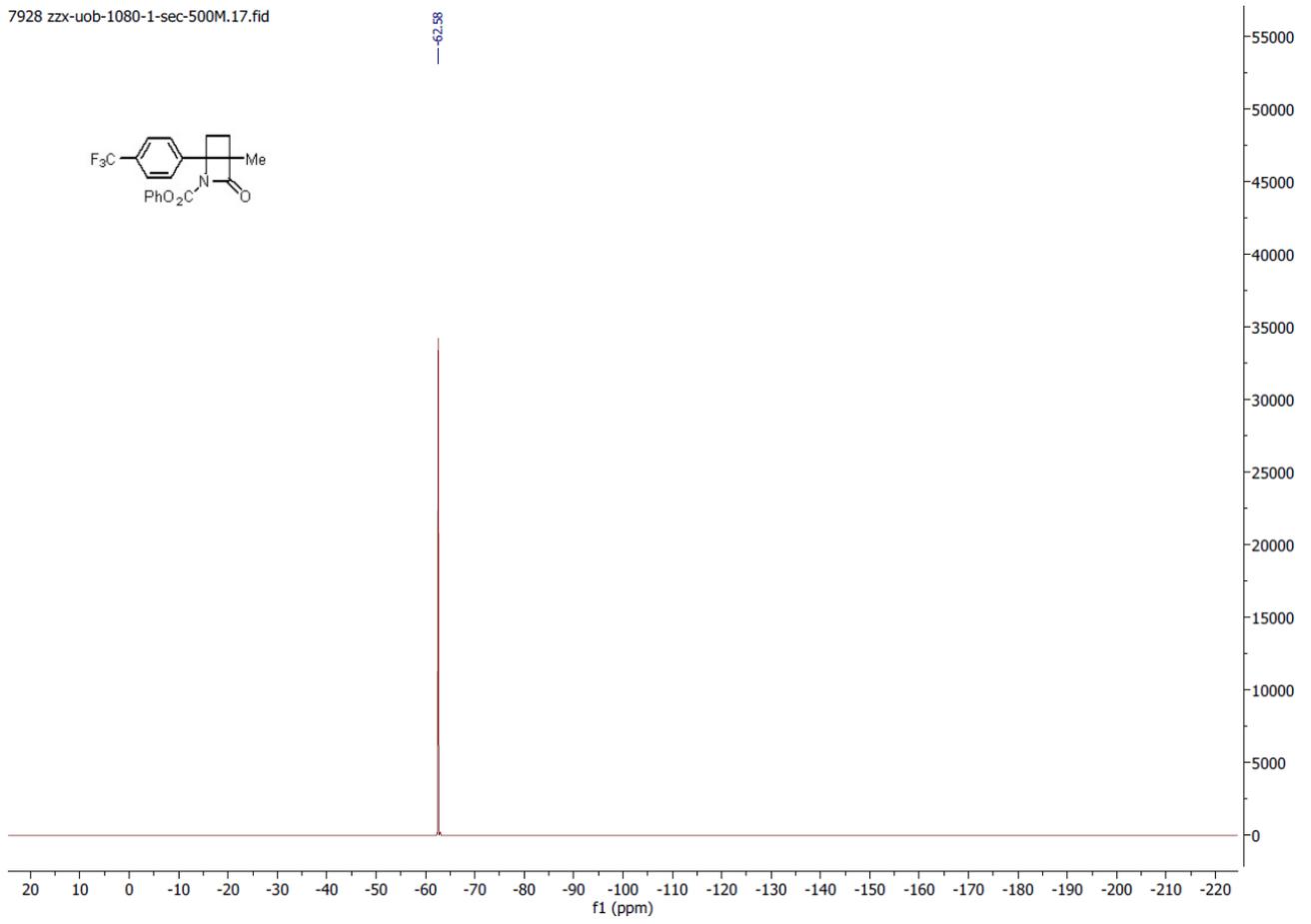
Compound 6c

7928 zzx-uob-1080-1-sec-500M.11.fid



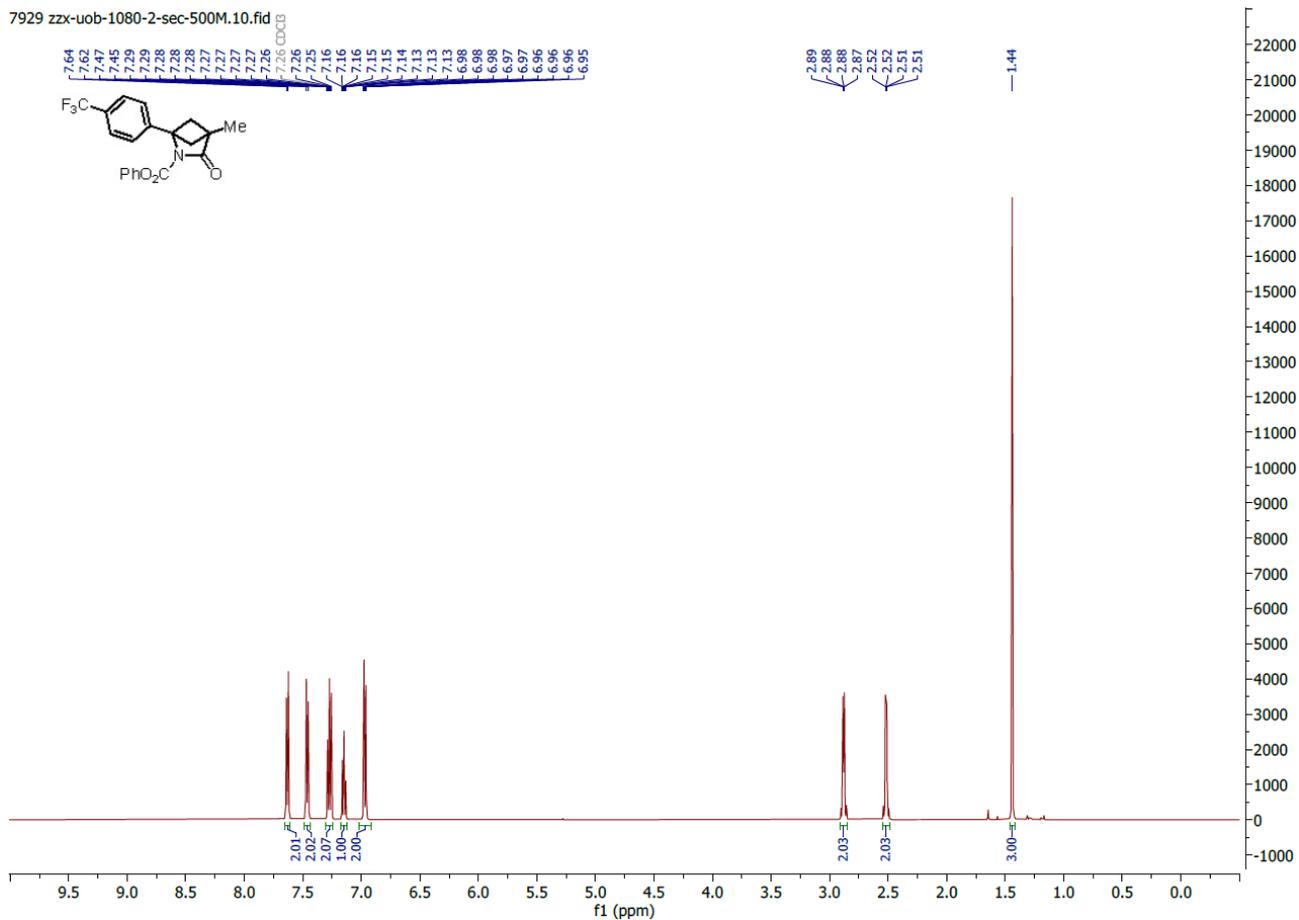
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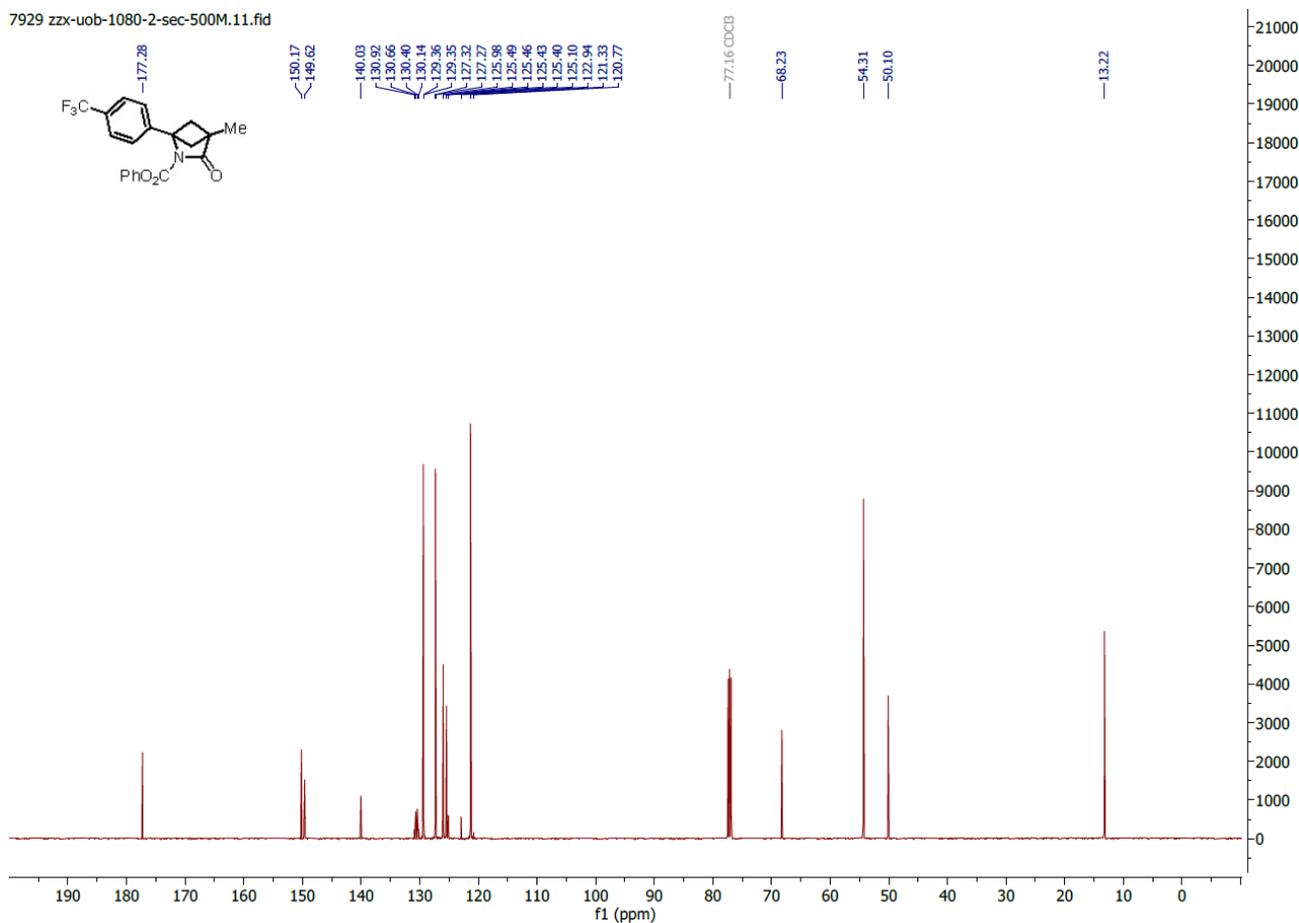


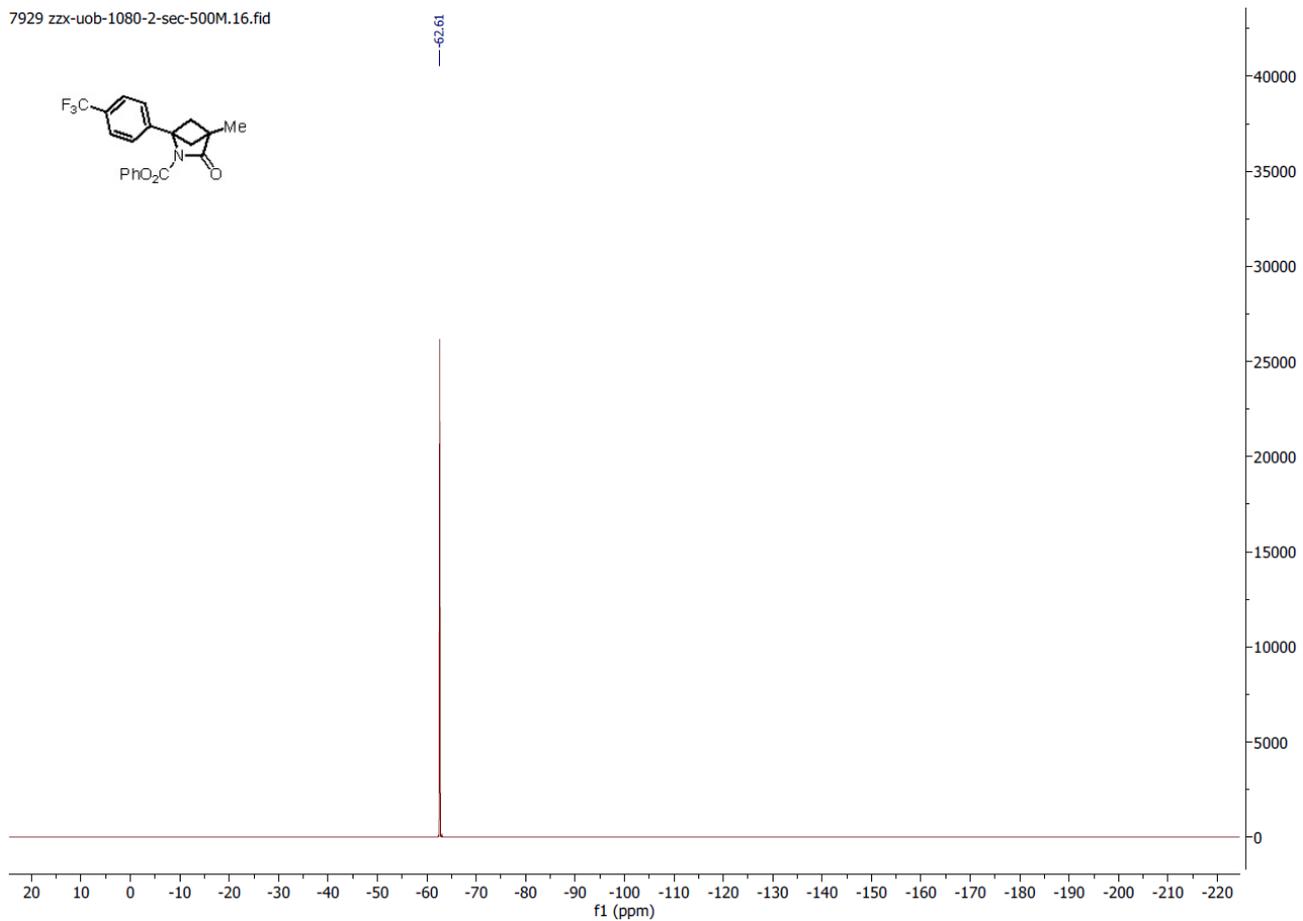
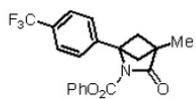
Compound 5c

7929 zzx-uob-1080-2-sec-500M.10.fid



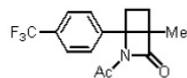
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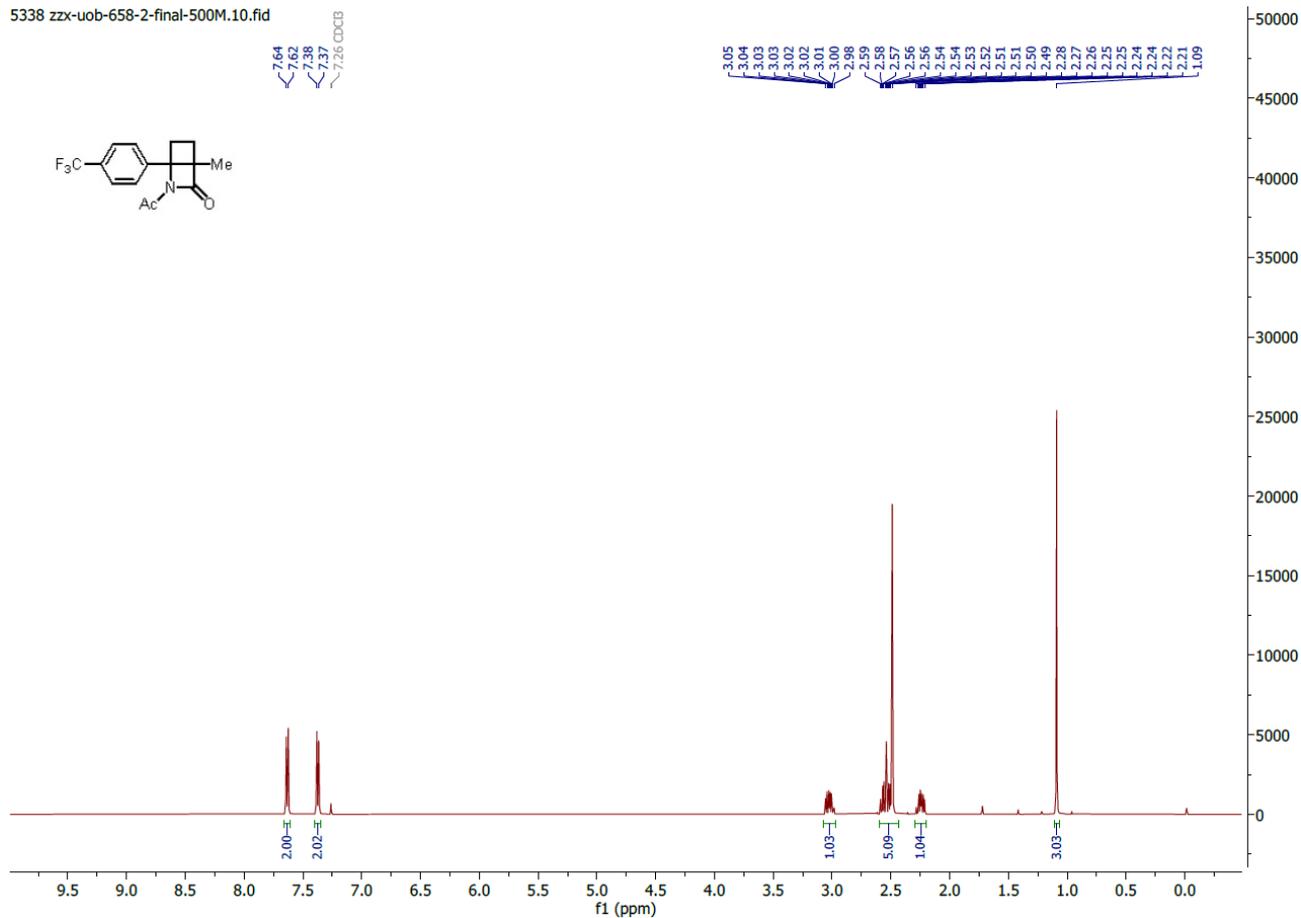
Compound 6d

5338 zzx-uob-658-2-final-500M.10.fid

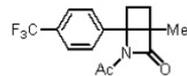


7.64
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7.26 CDCl₃

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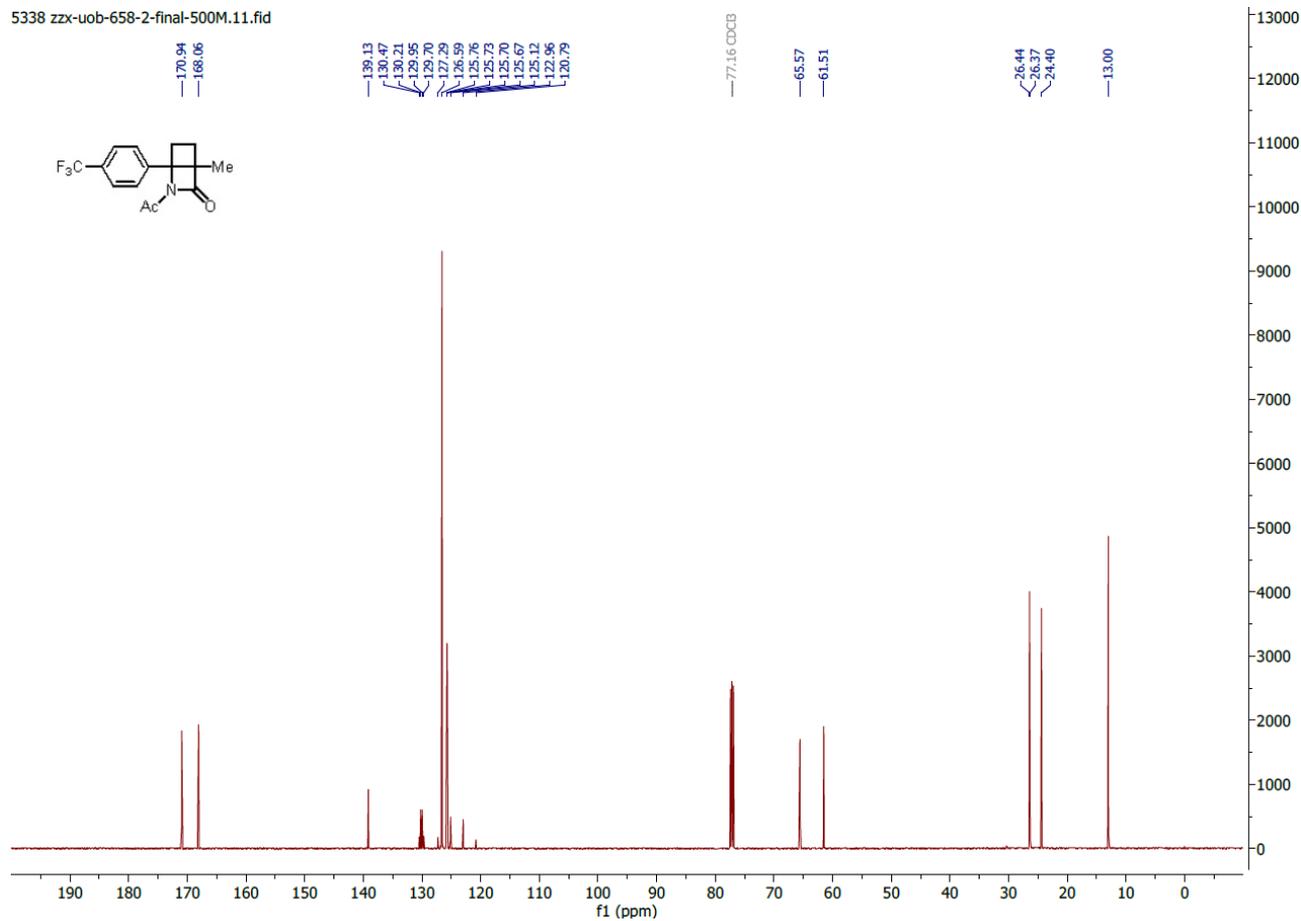
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125.12
122.96
120.79

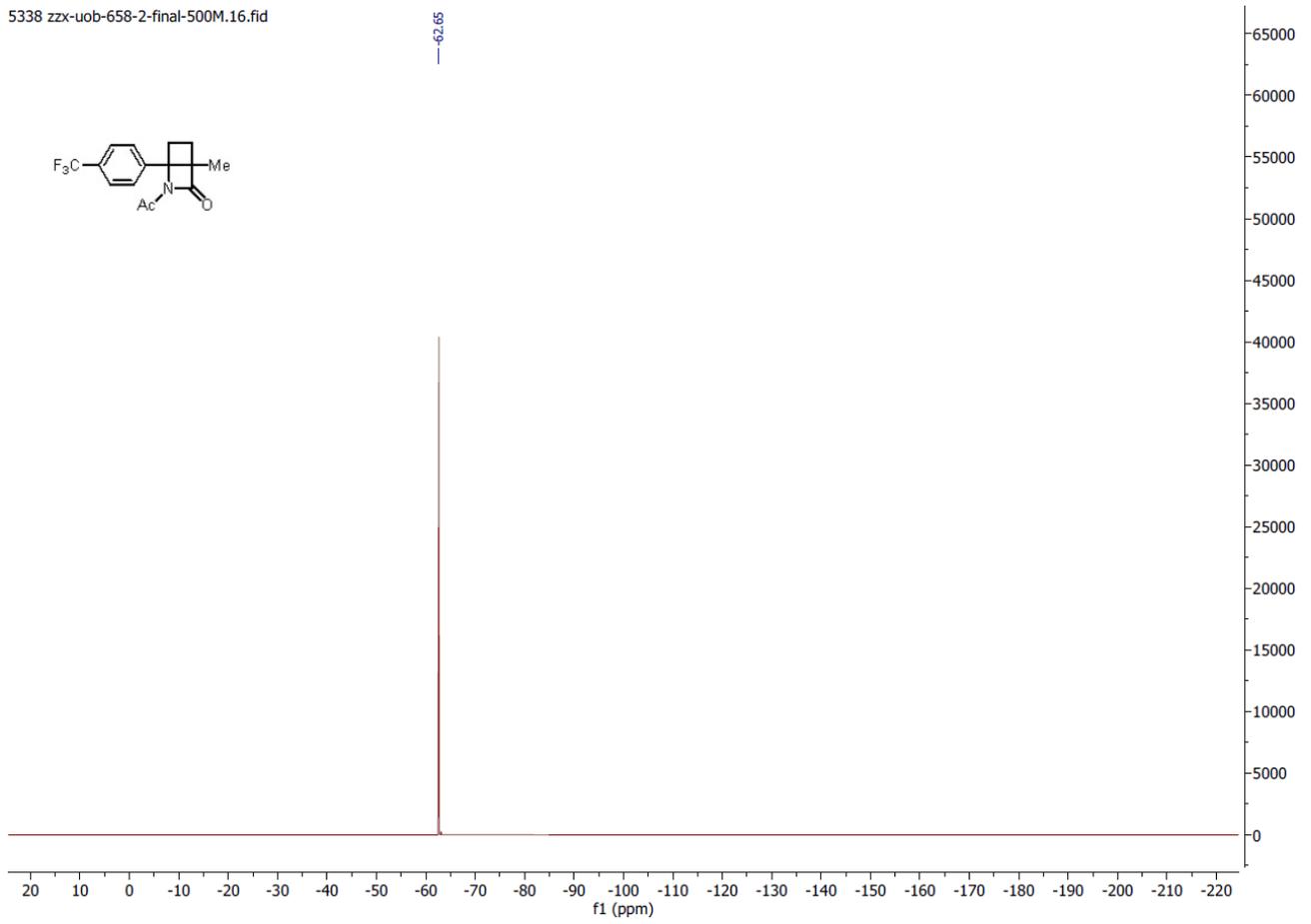
77.16 CDCl₃

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26.44
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24.40

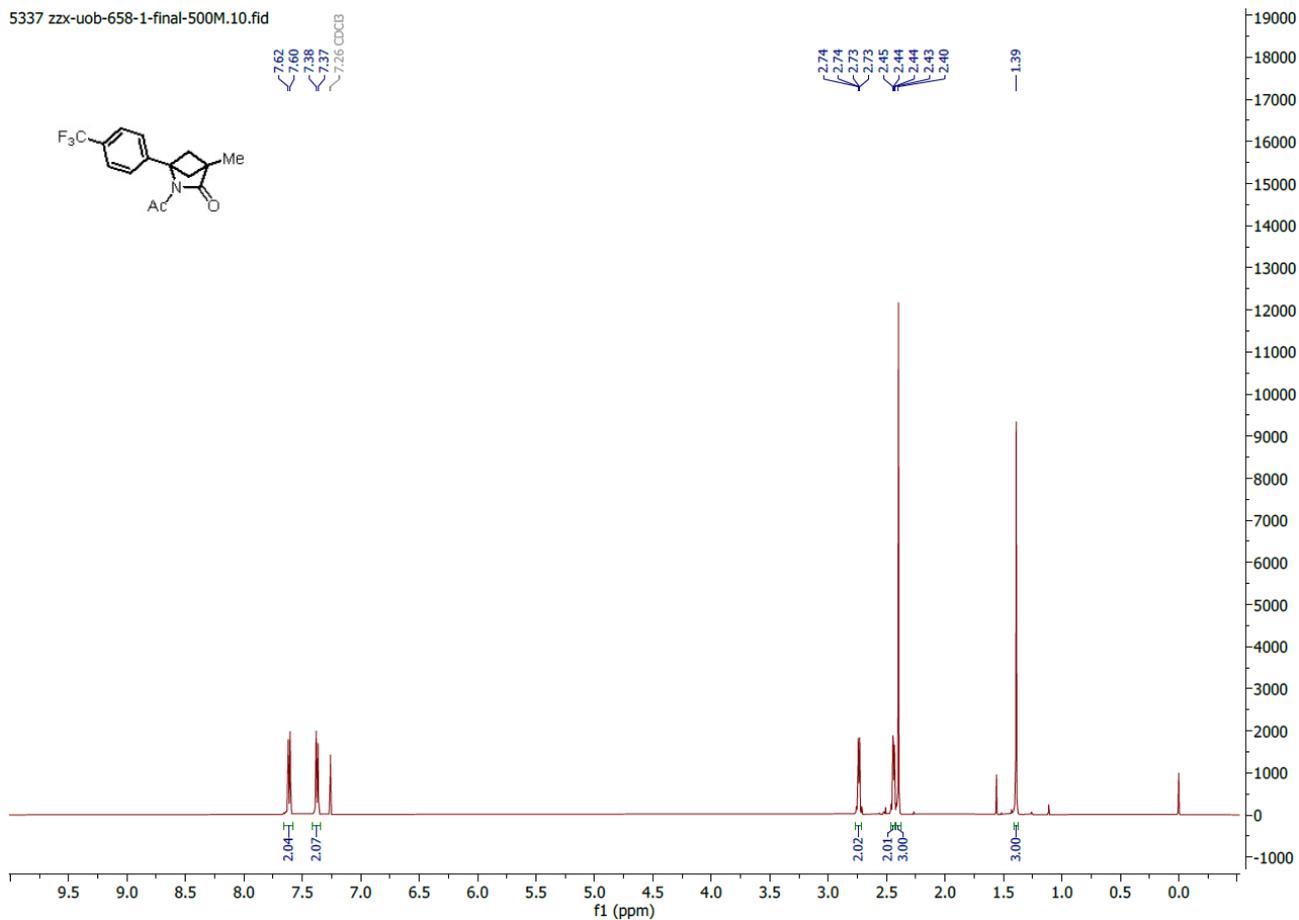
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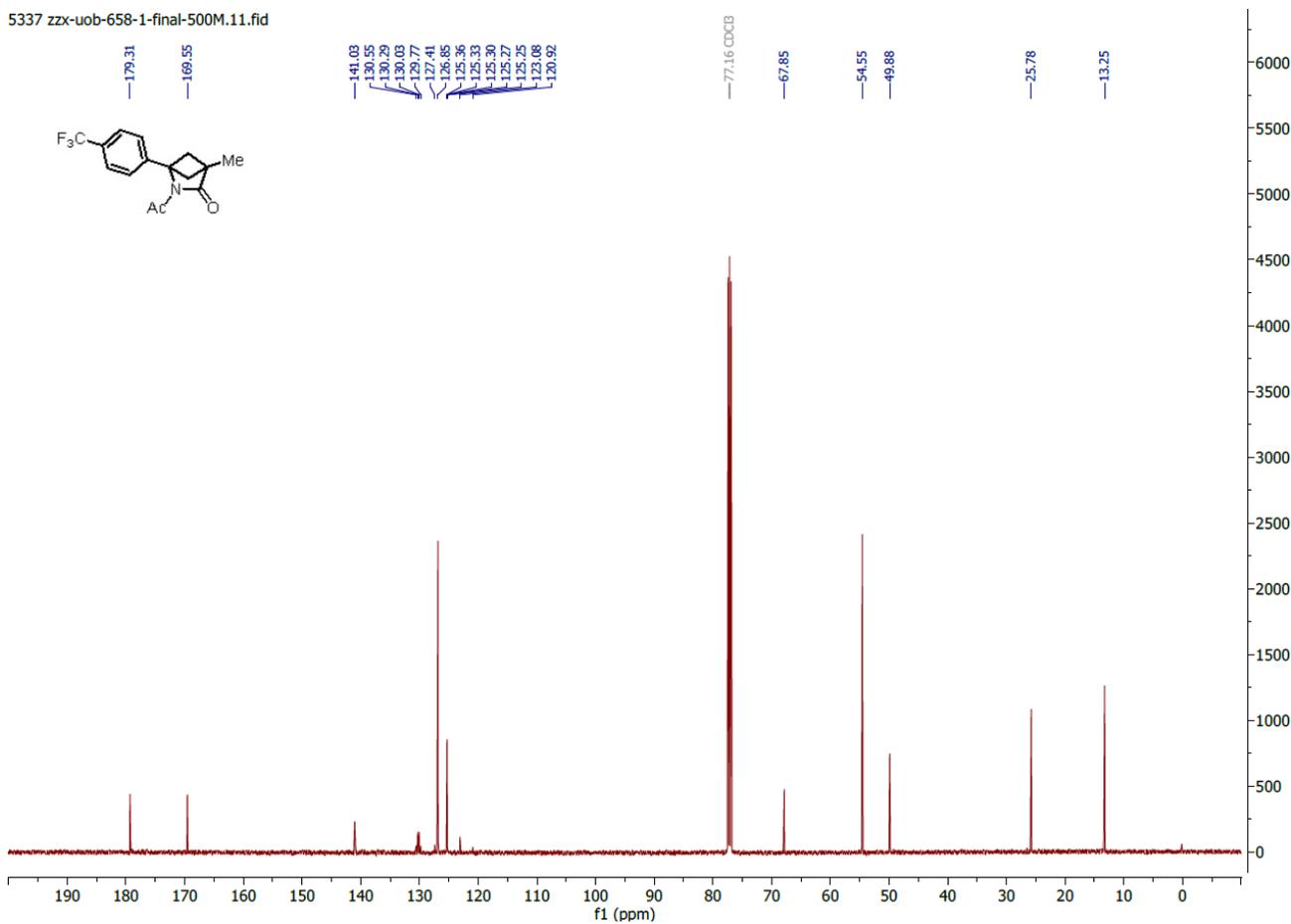


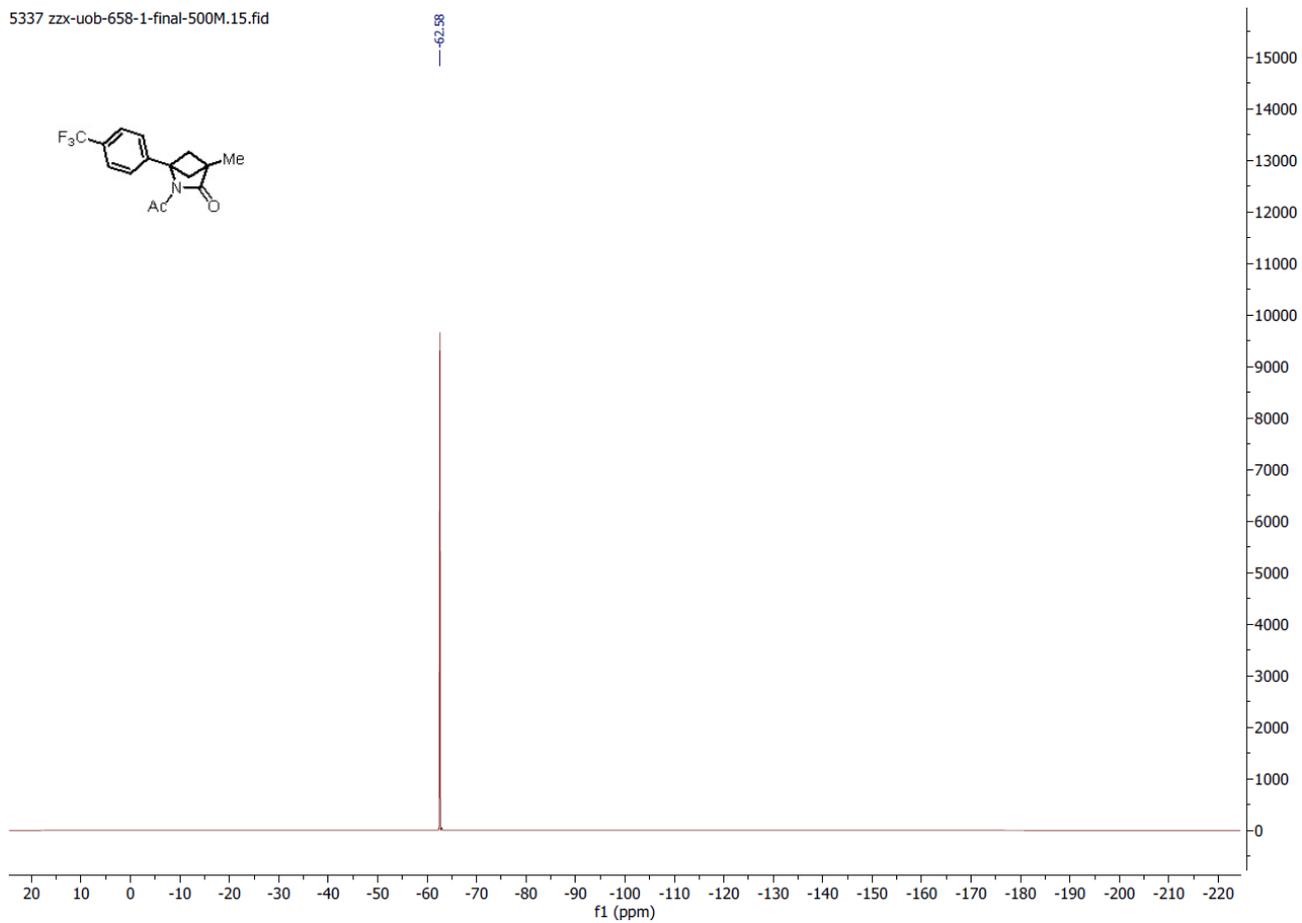
Compound 5d

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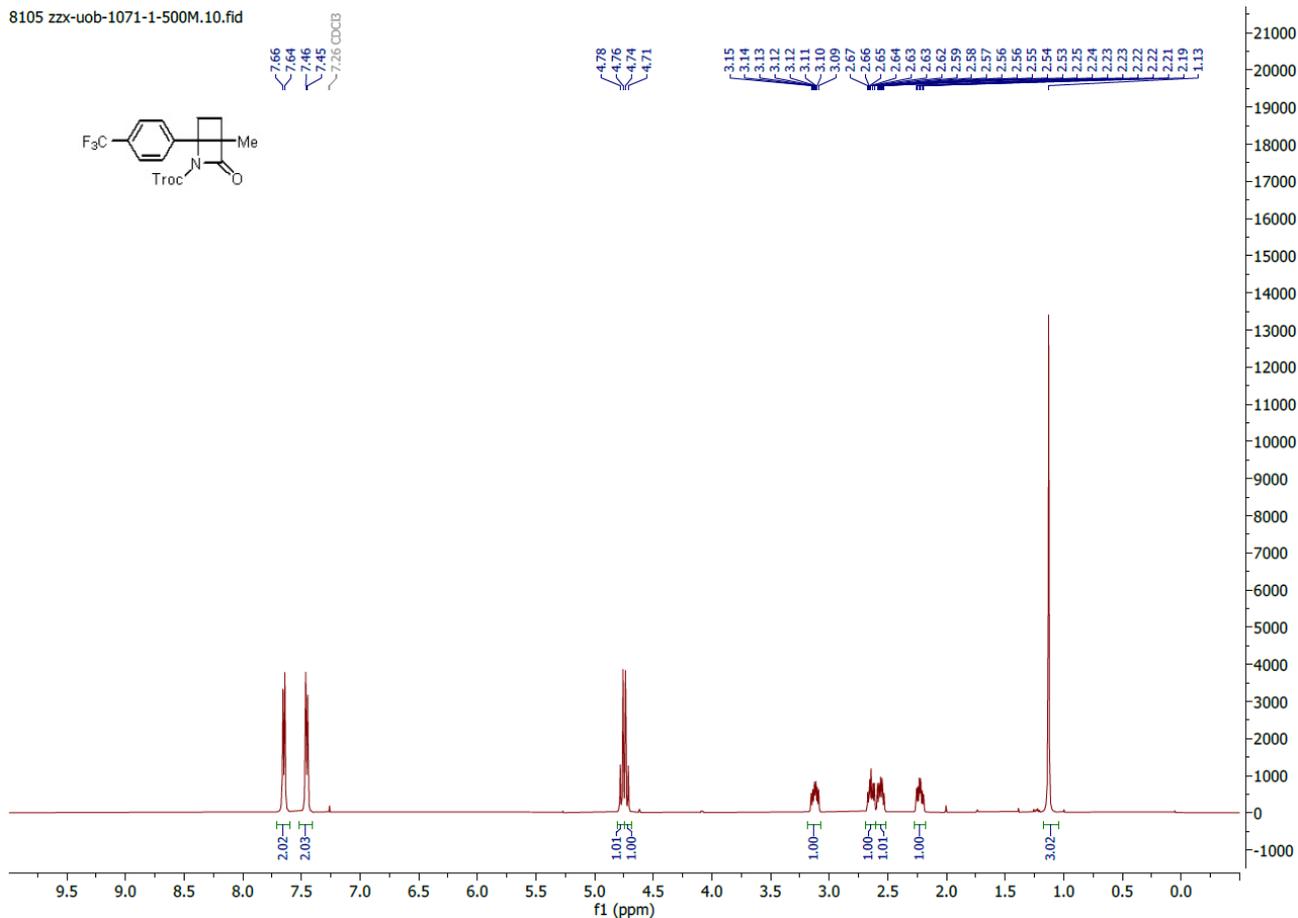
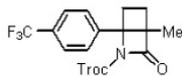
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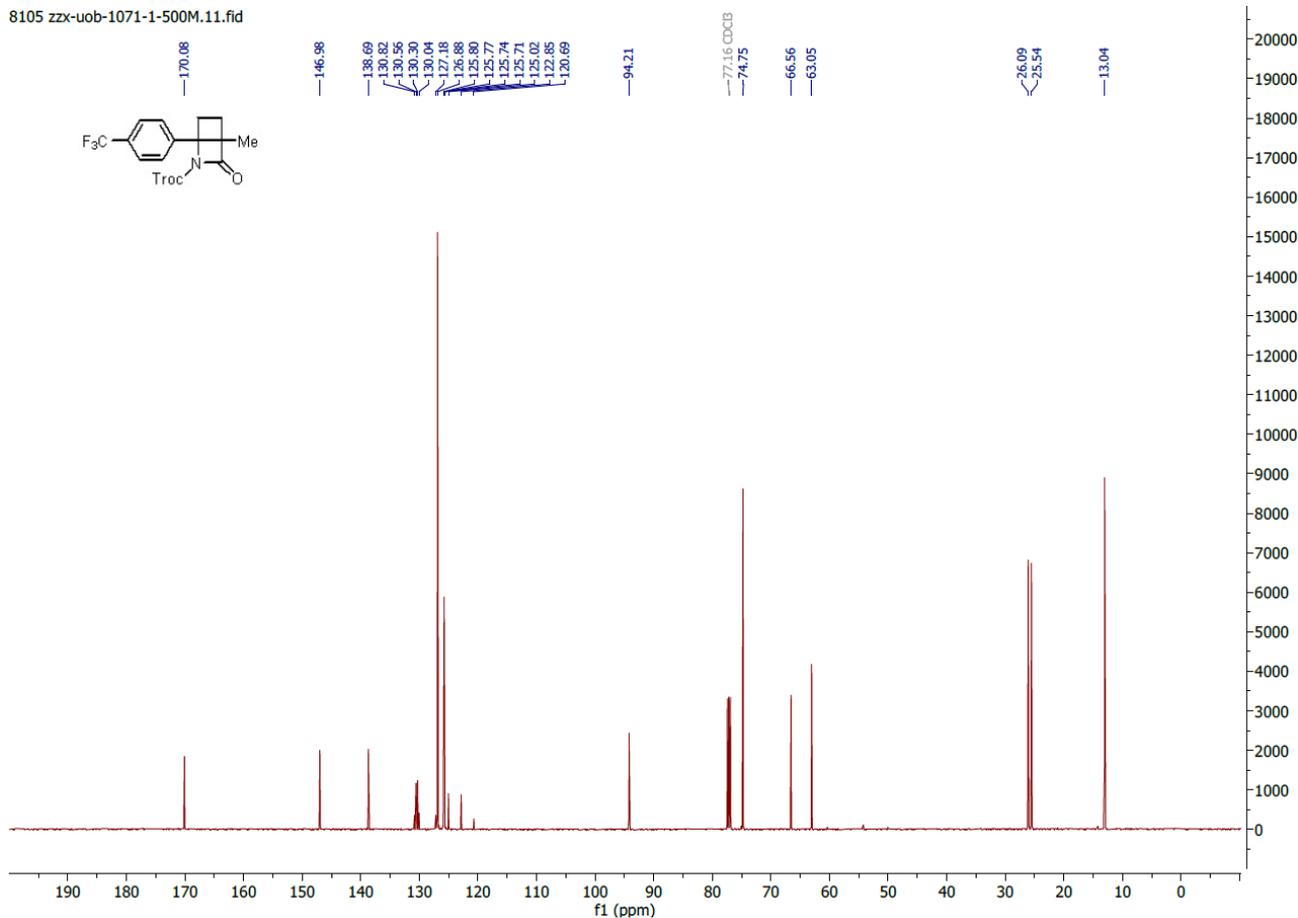
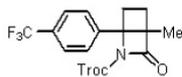


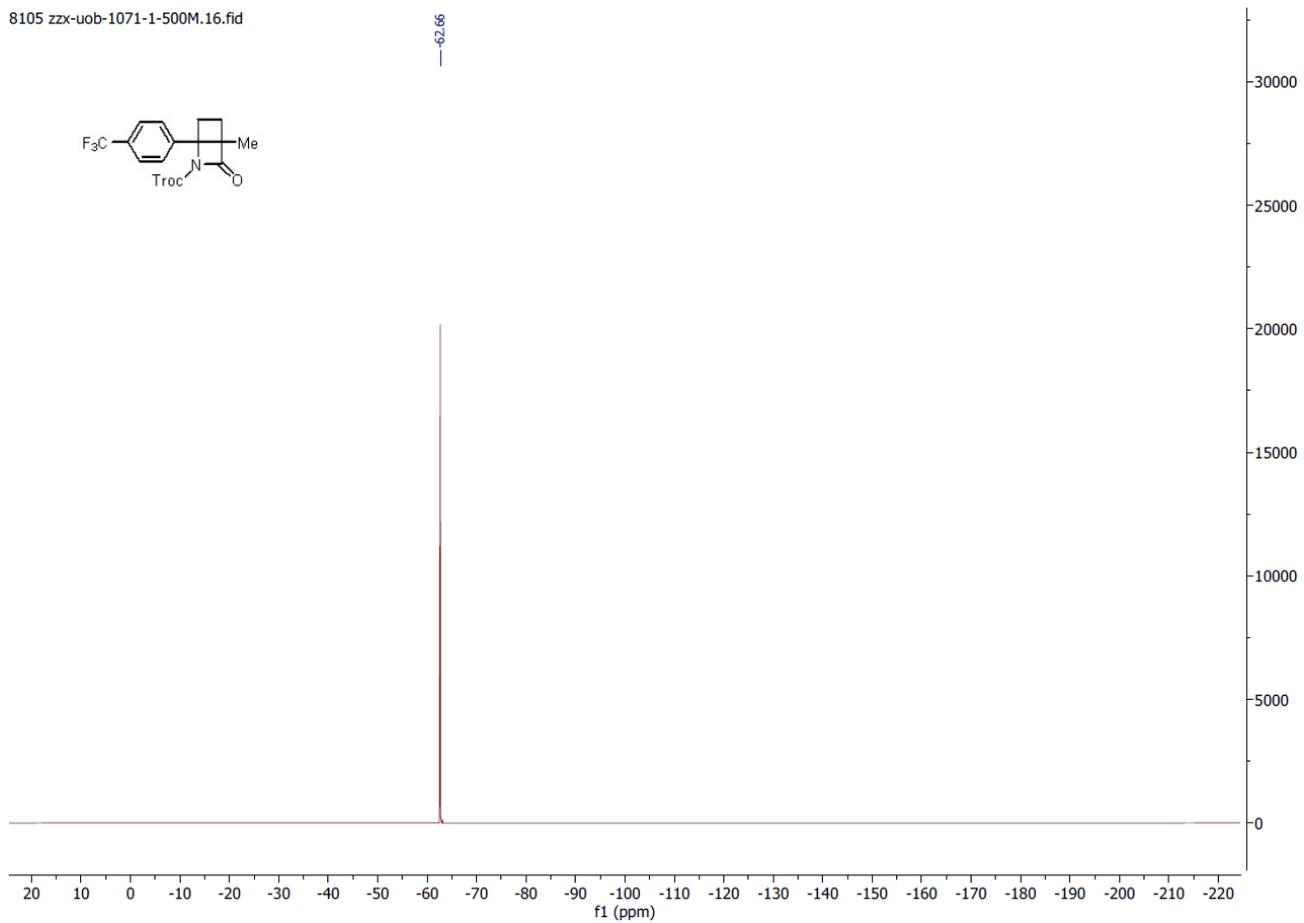
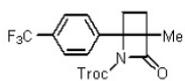
Compound 6e

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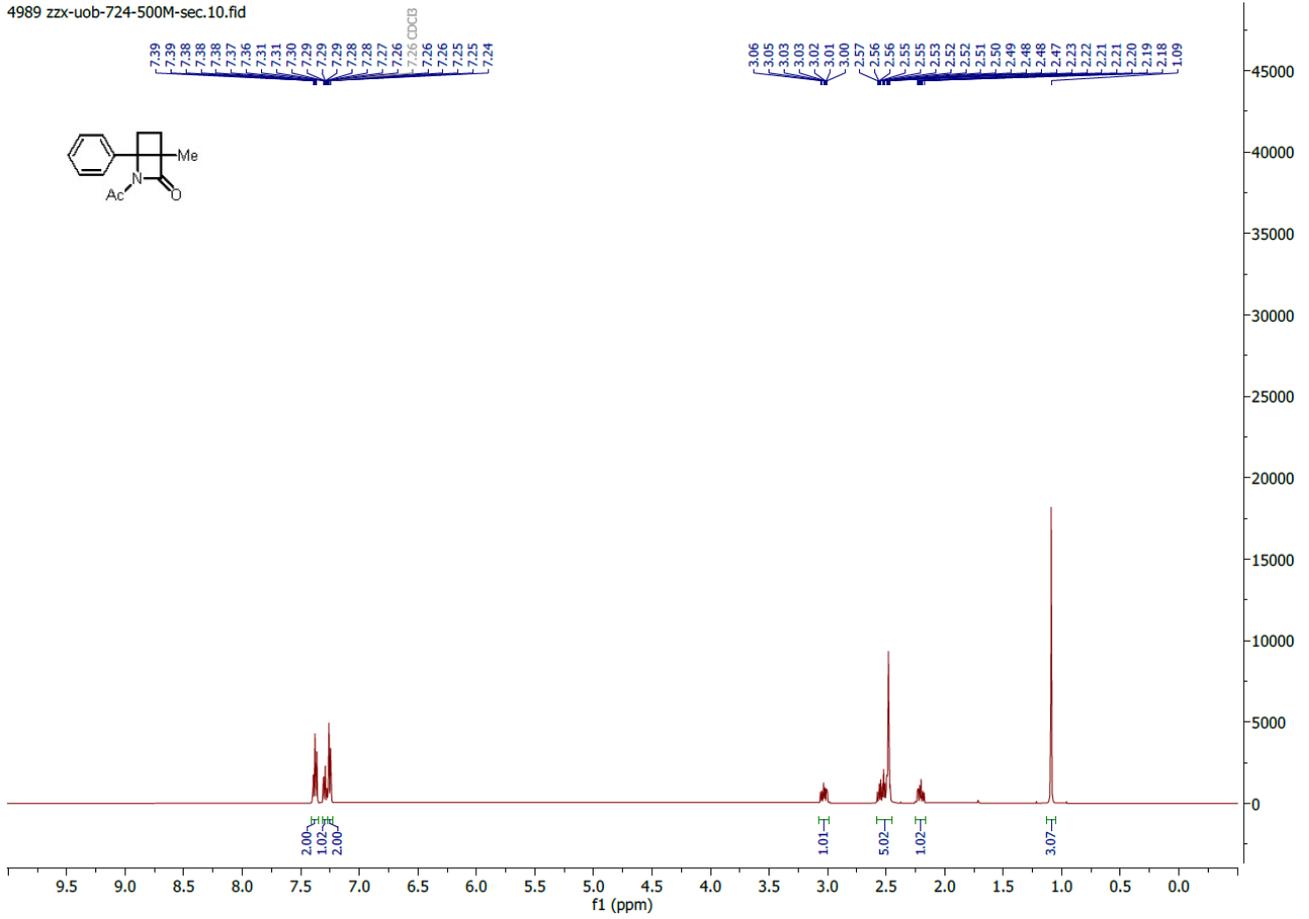
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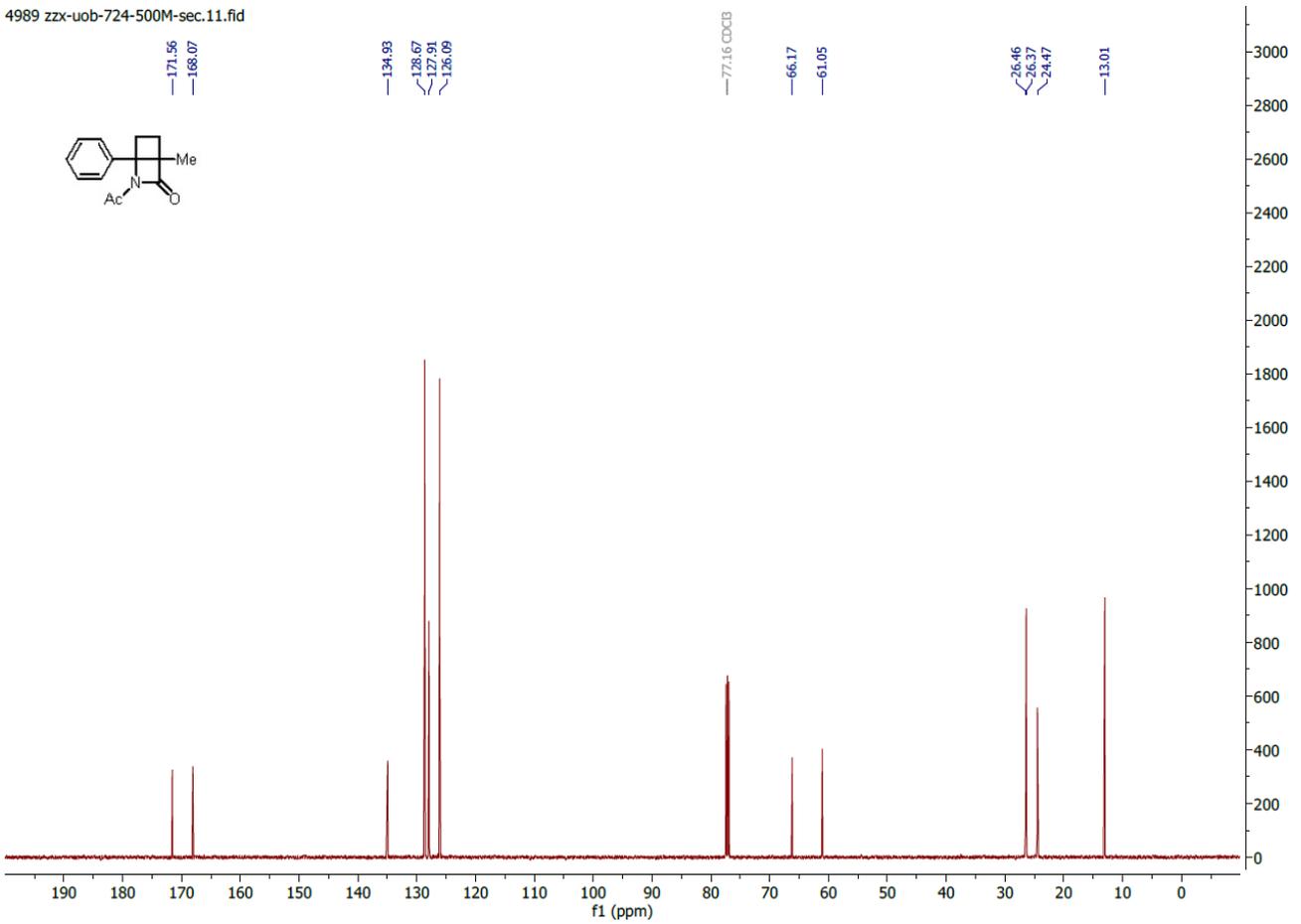


Compound 6f

4989 zzx-uob-724-500M-sec.10.fid

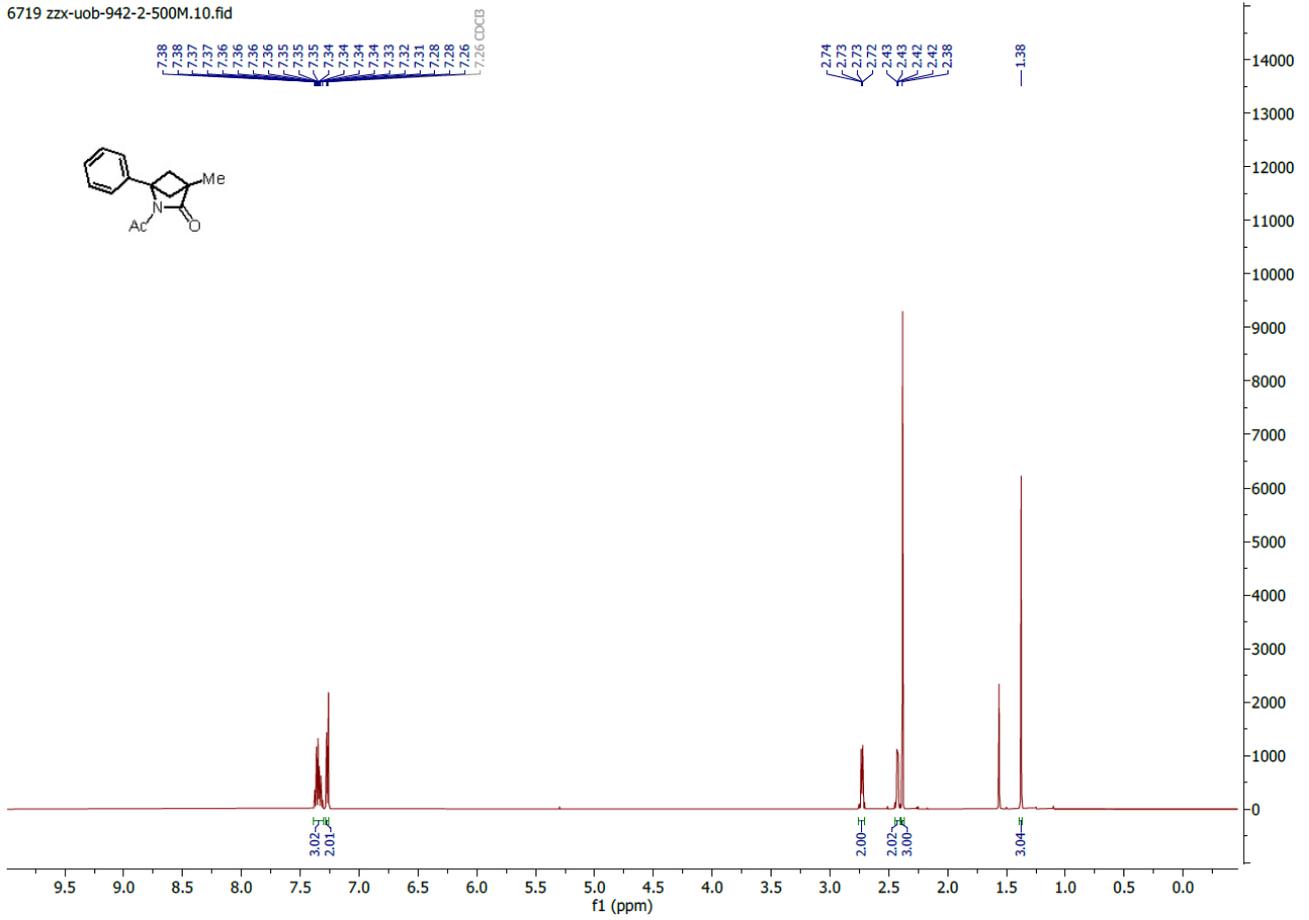


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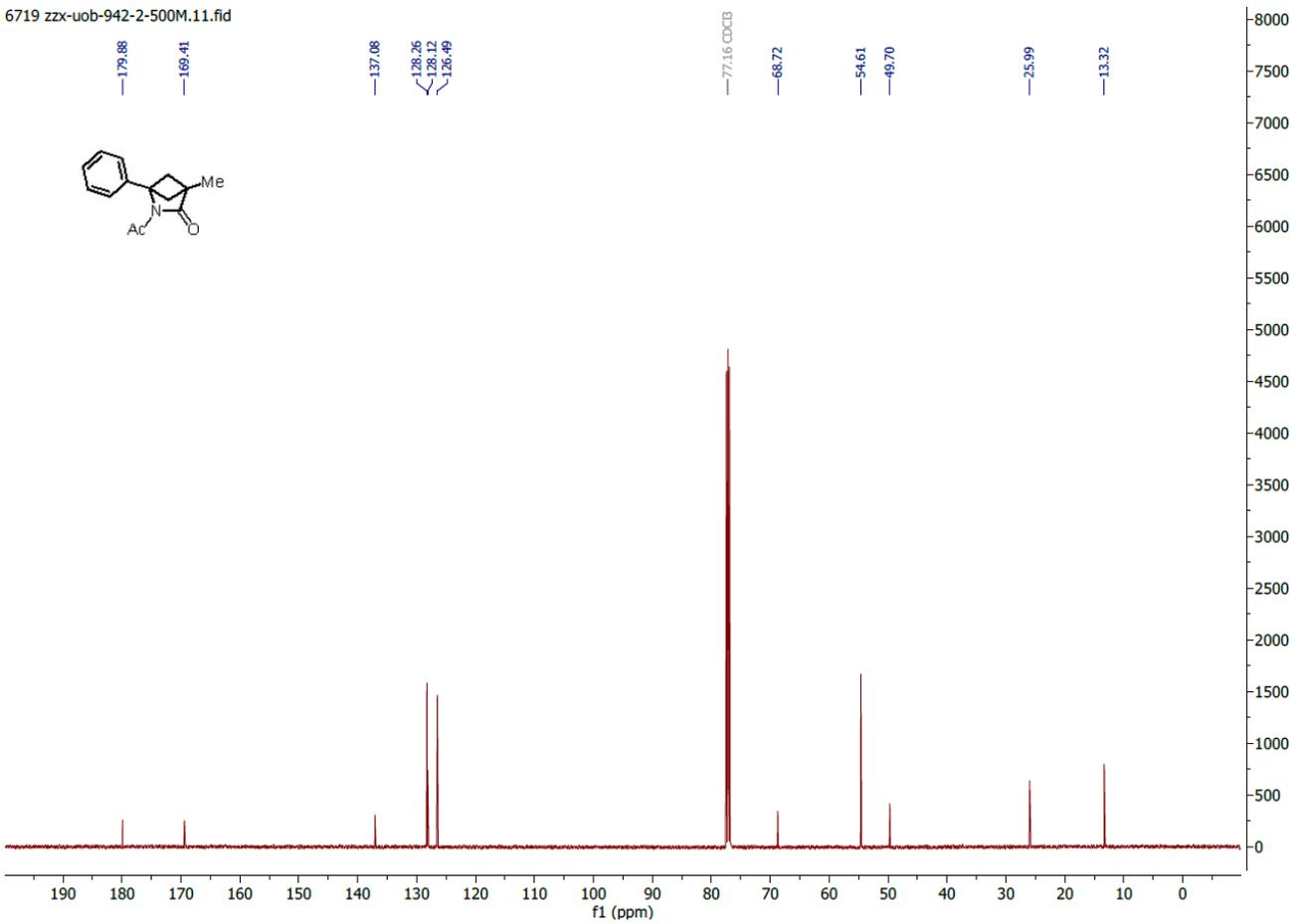


Compound 5f

6719 zzx-uob-942-2-500M.10.fid

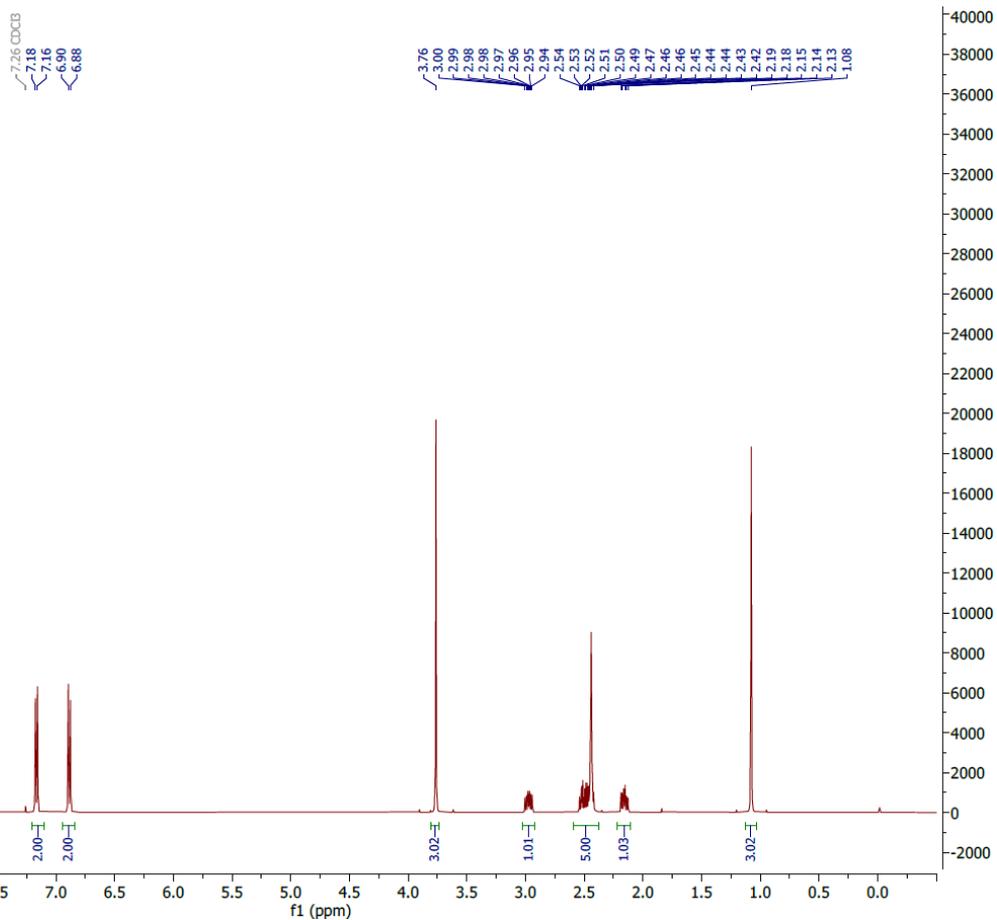
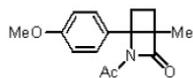


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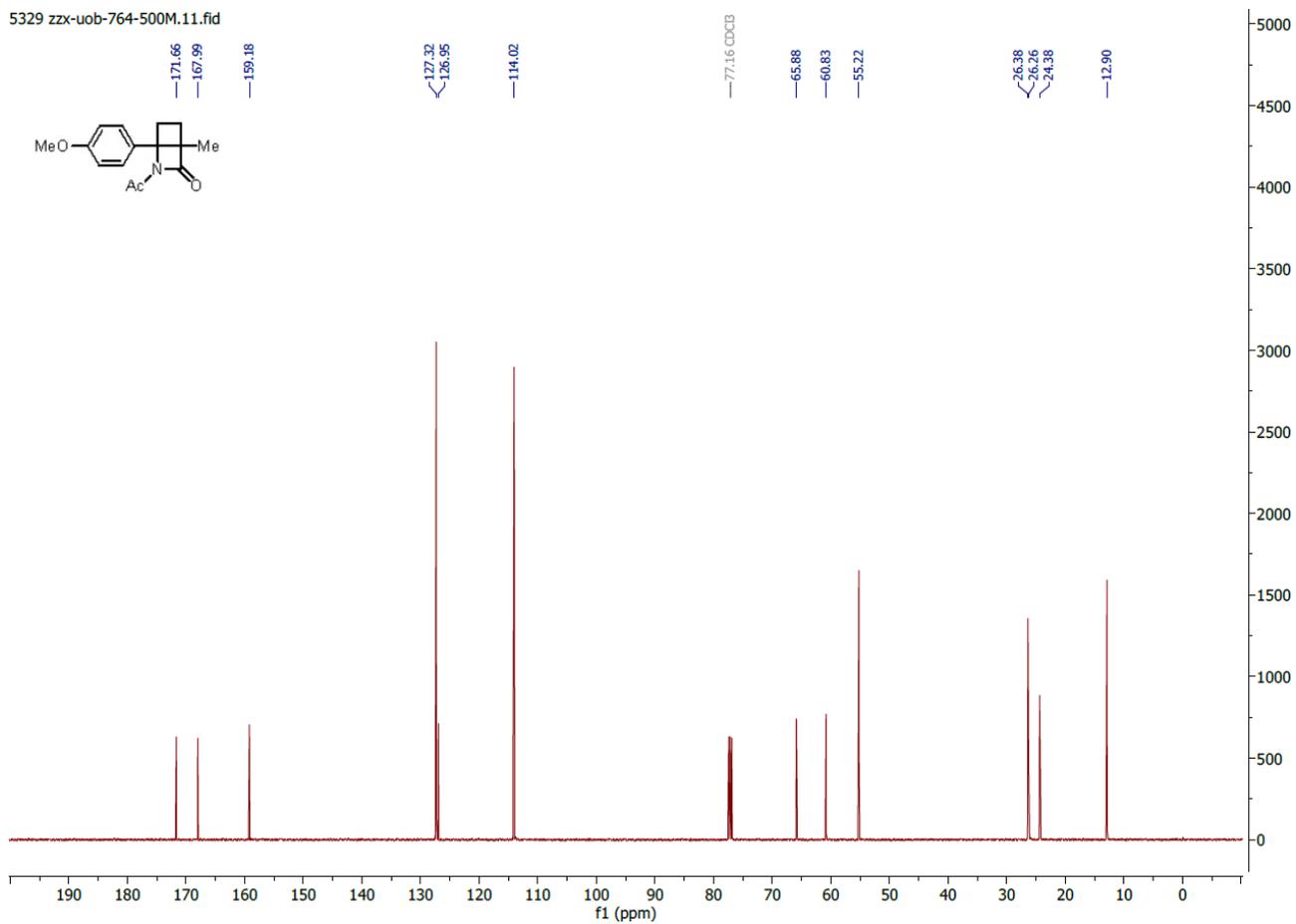
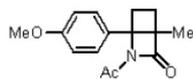


Compound 6g

5329 zzx-uob-764-500M.10.fid

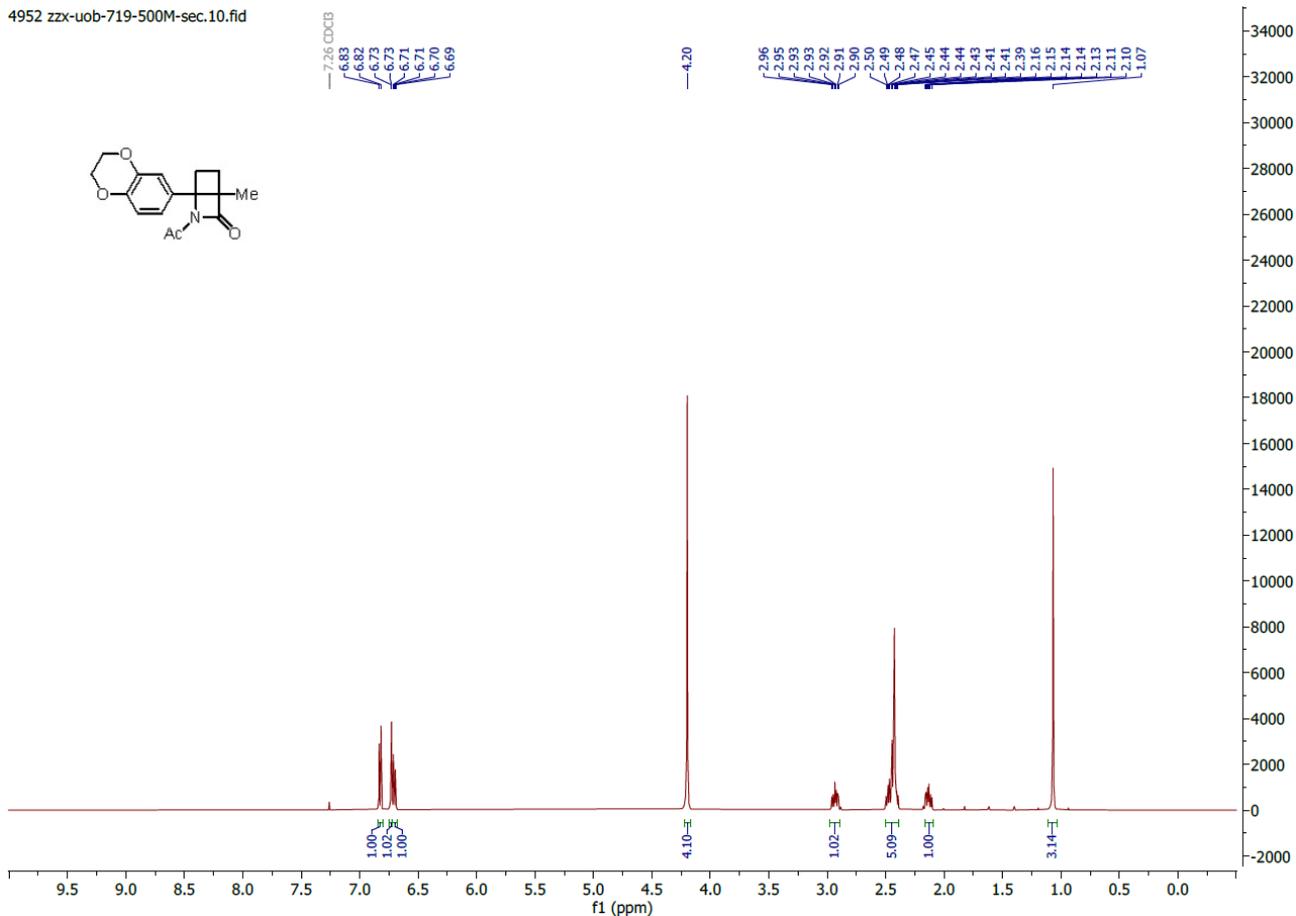


5329 zzx-uob-764-500M.11.fid

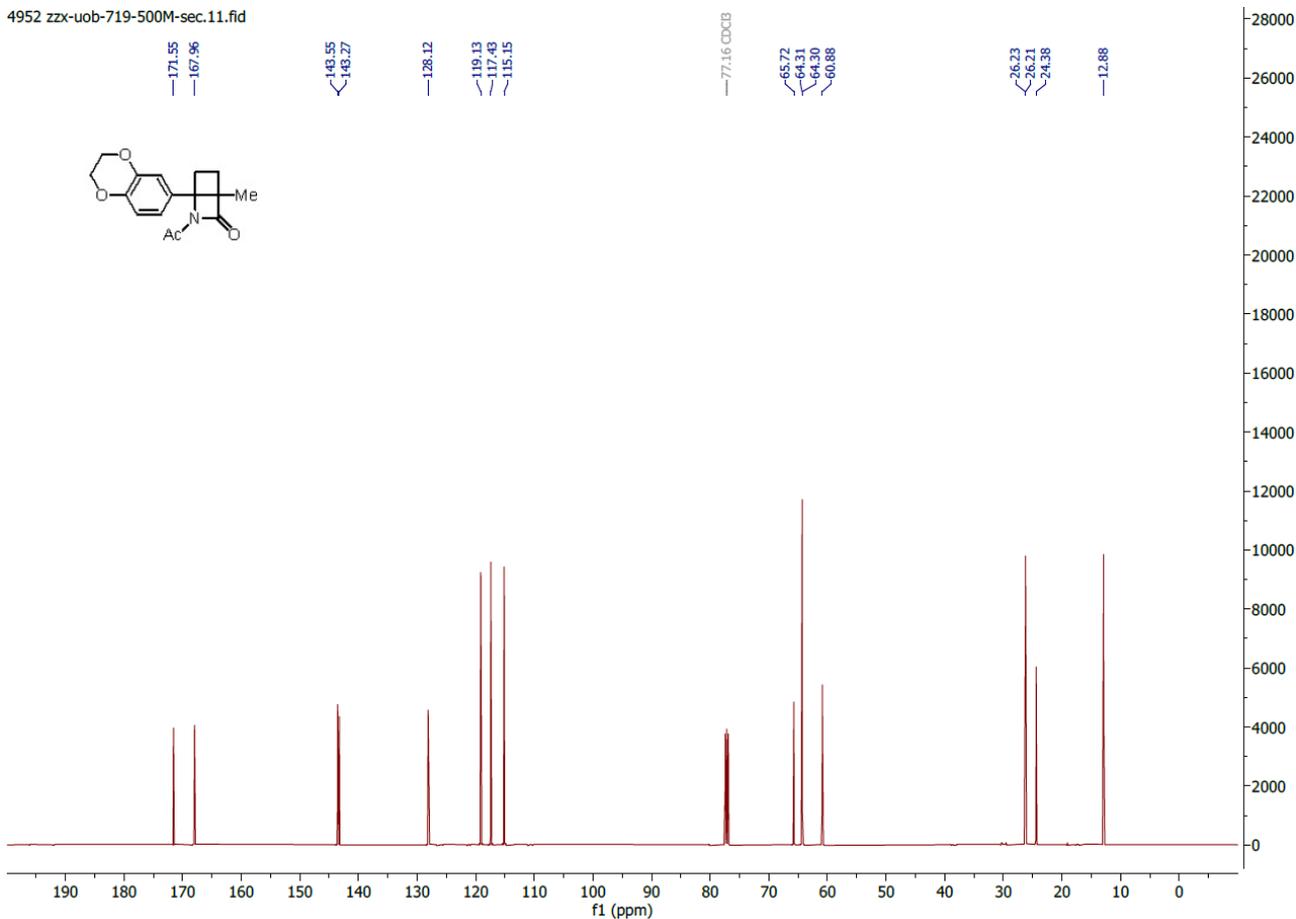


Compound 6h

4952 zzx-uob-719-500M-sec.10.fid

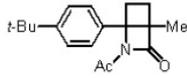


4952 zzx-uob-719-500M-sec.11.fid

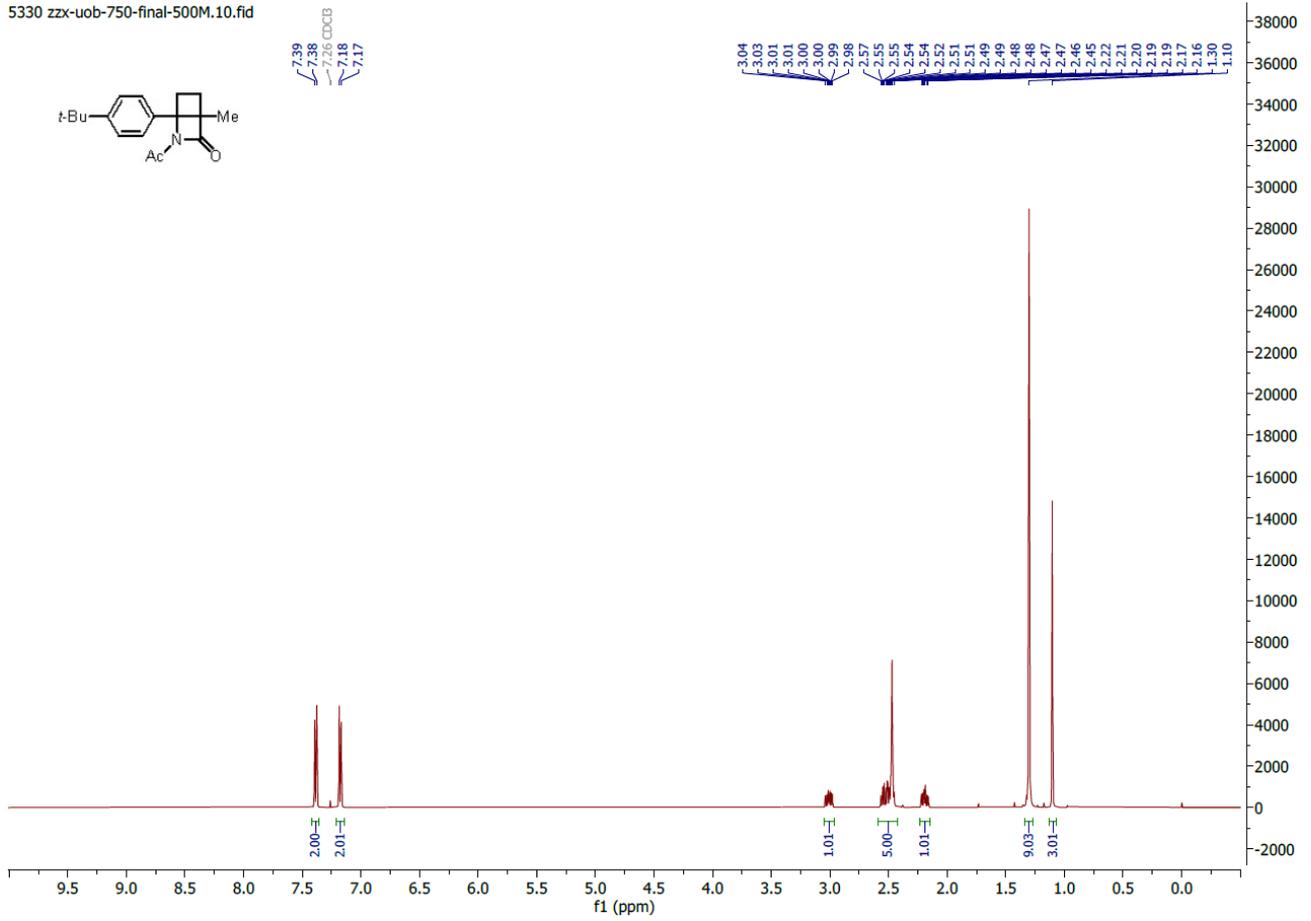


Compound 6i

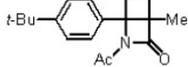
5330 zzx-uob-750-final-500M.10.fid



7.39
7.38
7.26 CDCl₃
7.18
7.17



5330 zzx-uob-750-final-500M.11.fid



171.72
168.08

150.68

131.84

125.82
125.56

77.16 CDCl₃

66.13

60.97

34.55

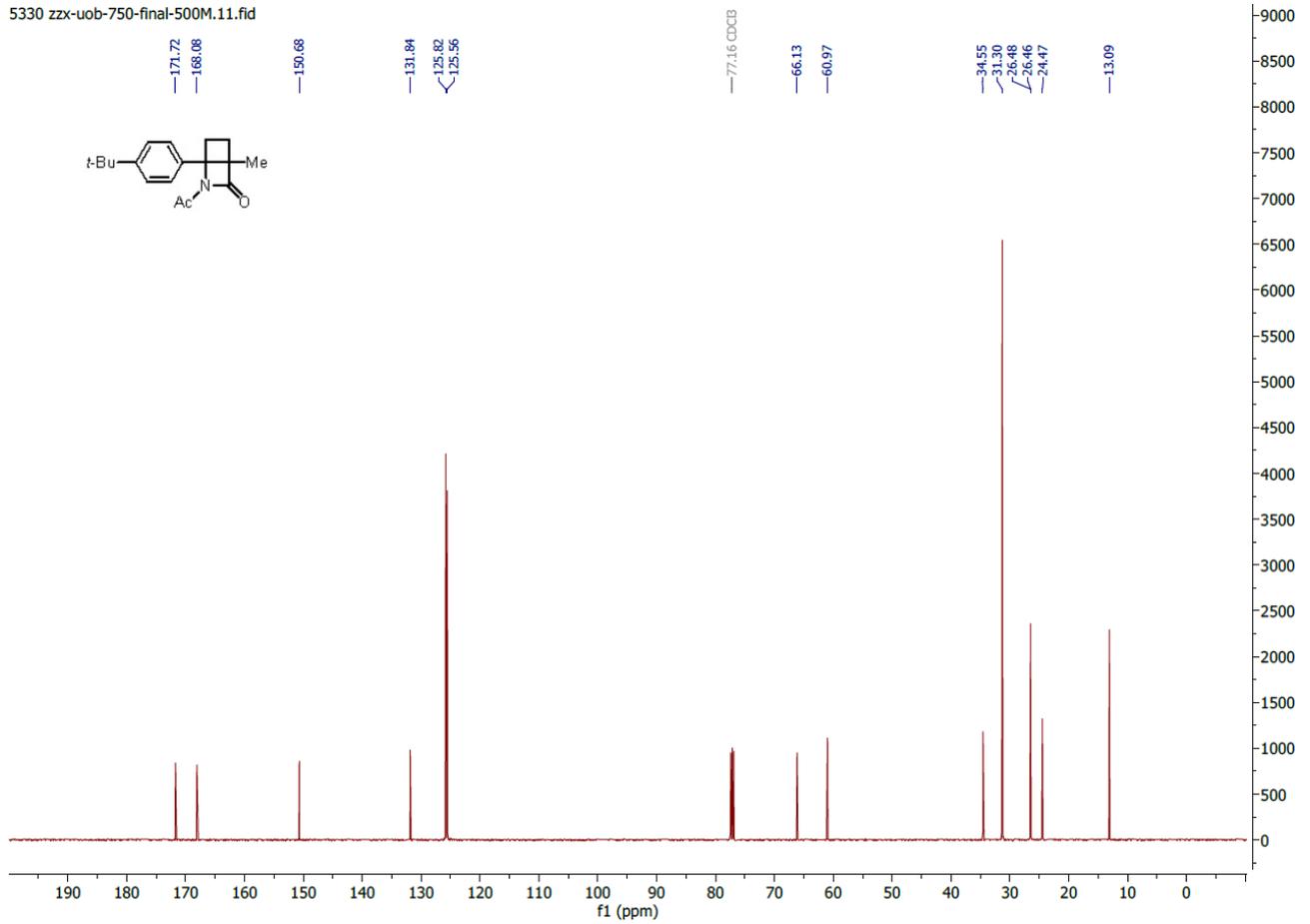
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26.46

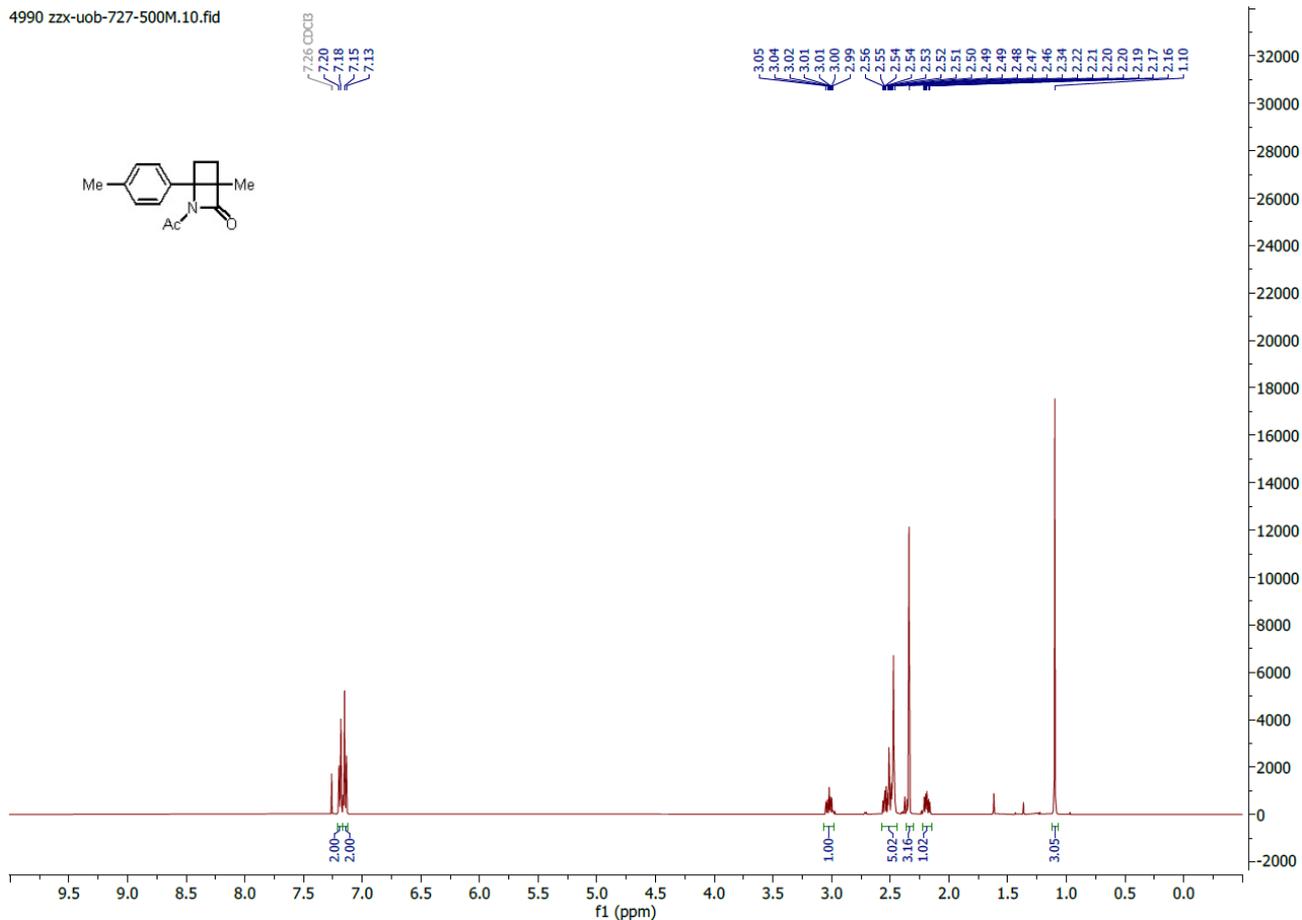
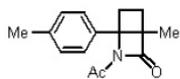
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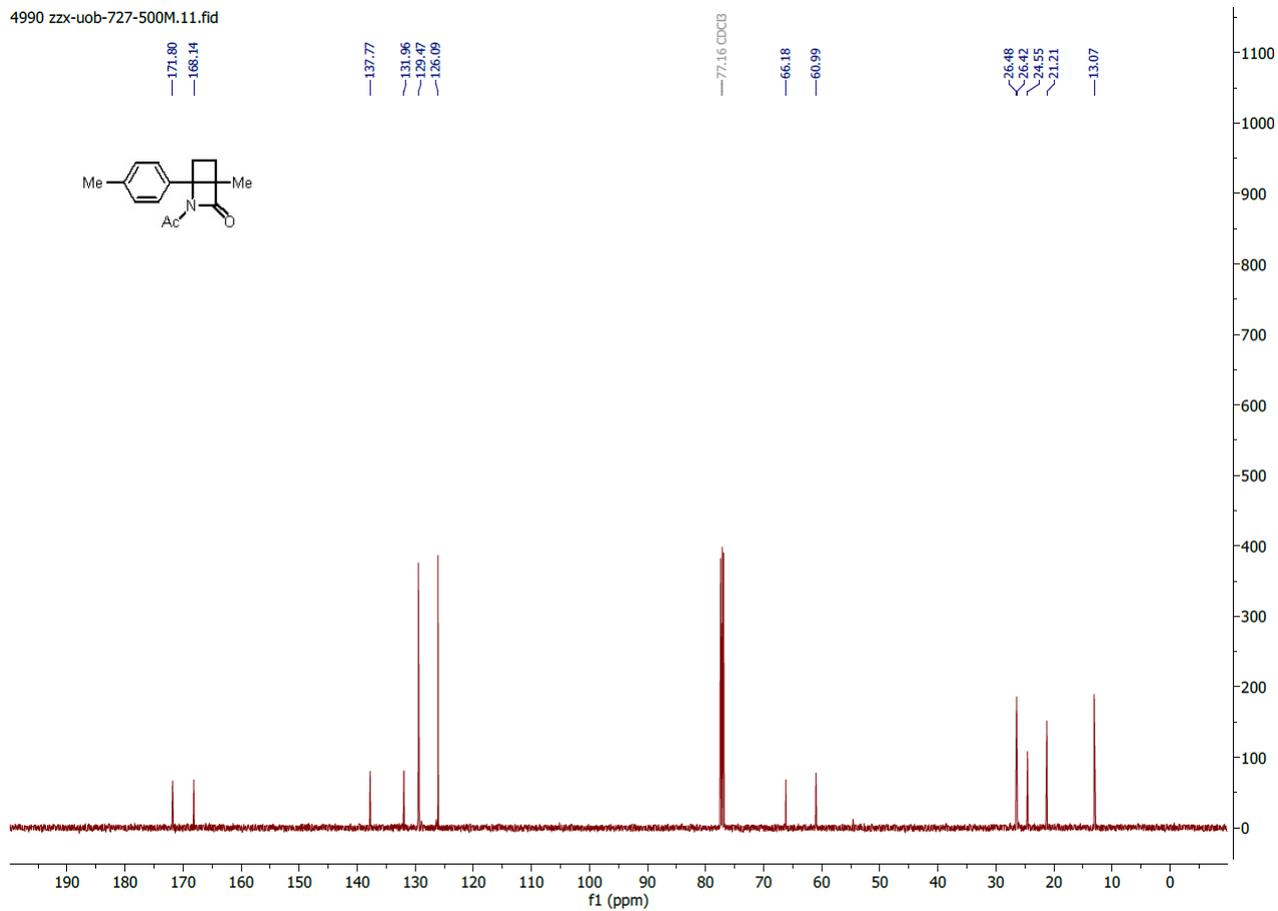
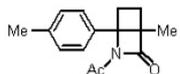


Compound 6j

4990 zzx-uob-727-500M.10.fid

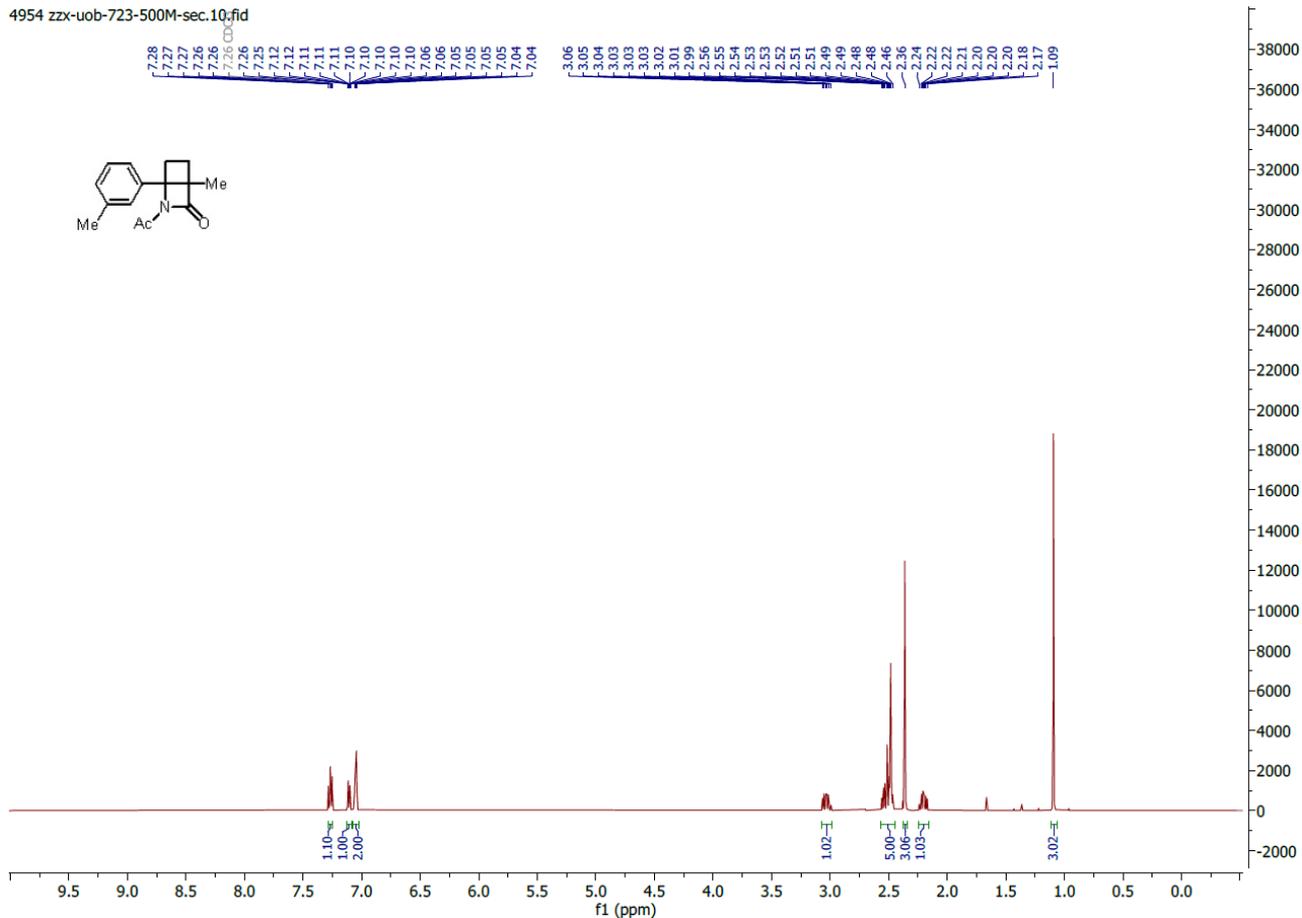


4990 zzx-uob-727-500M.11.fid

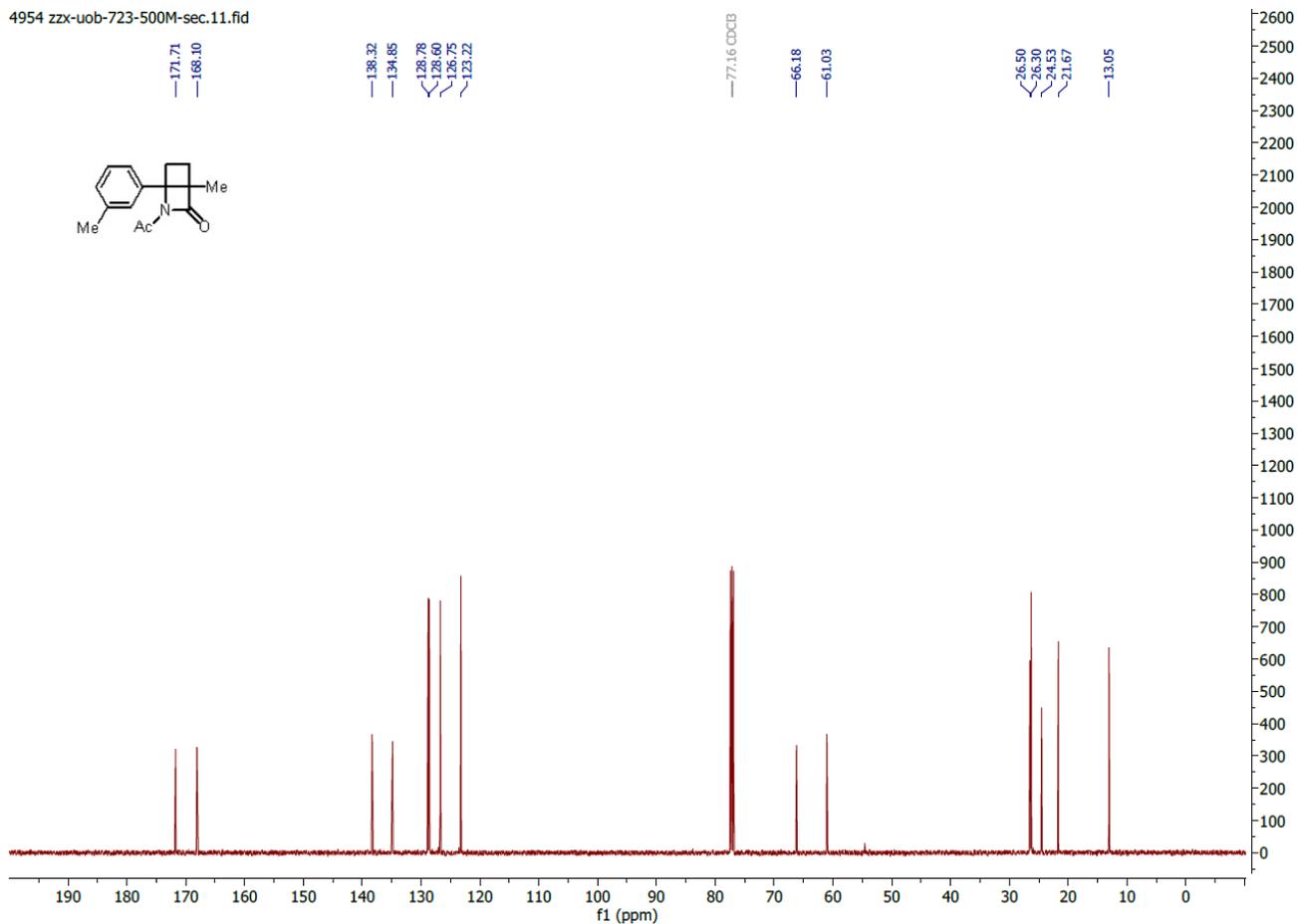


Compound 6k

4954 zzx-uob-723-500M-sec.10.fid

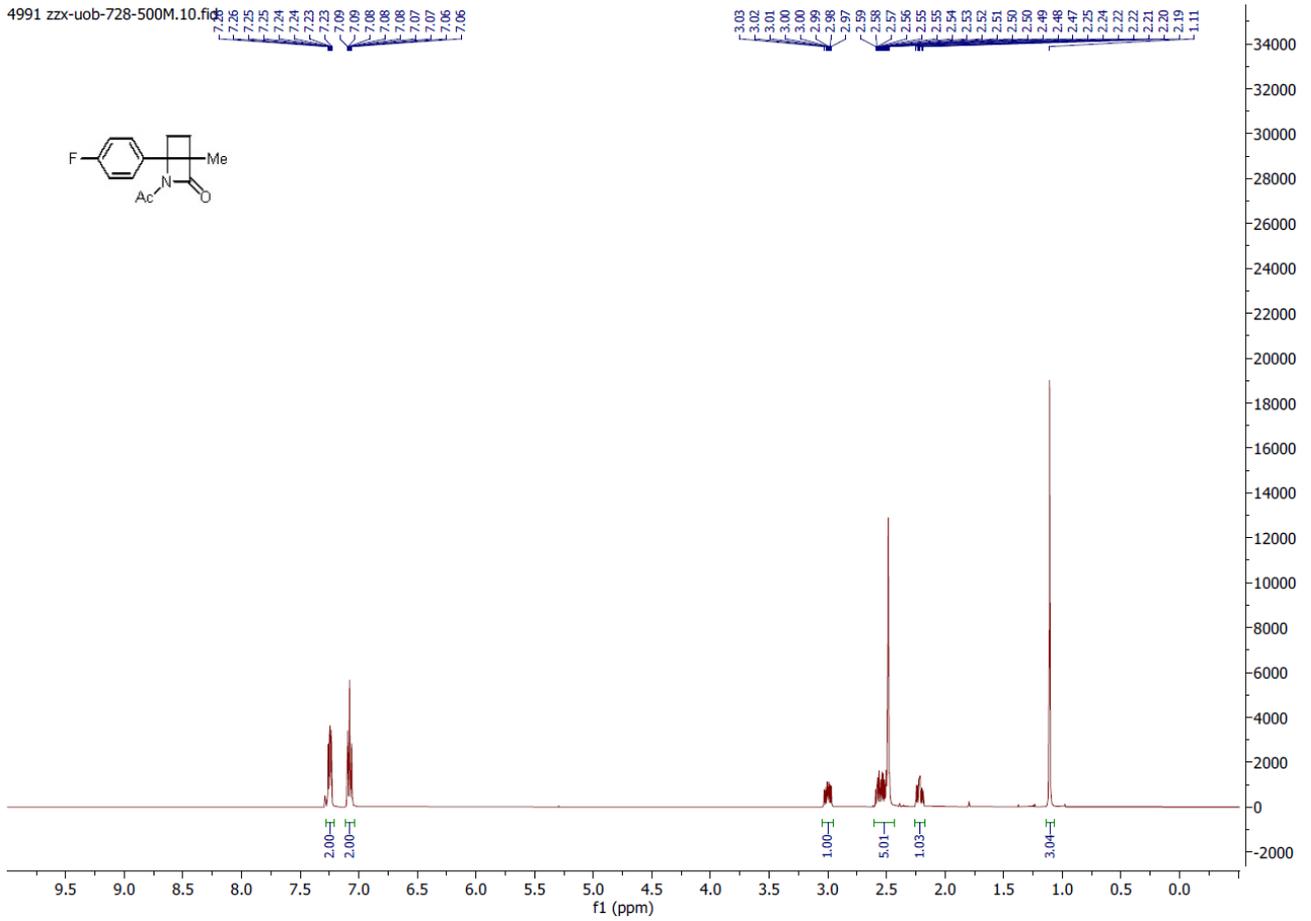


4954 zzx-uob-723-500M-sec.11.fid

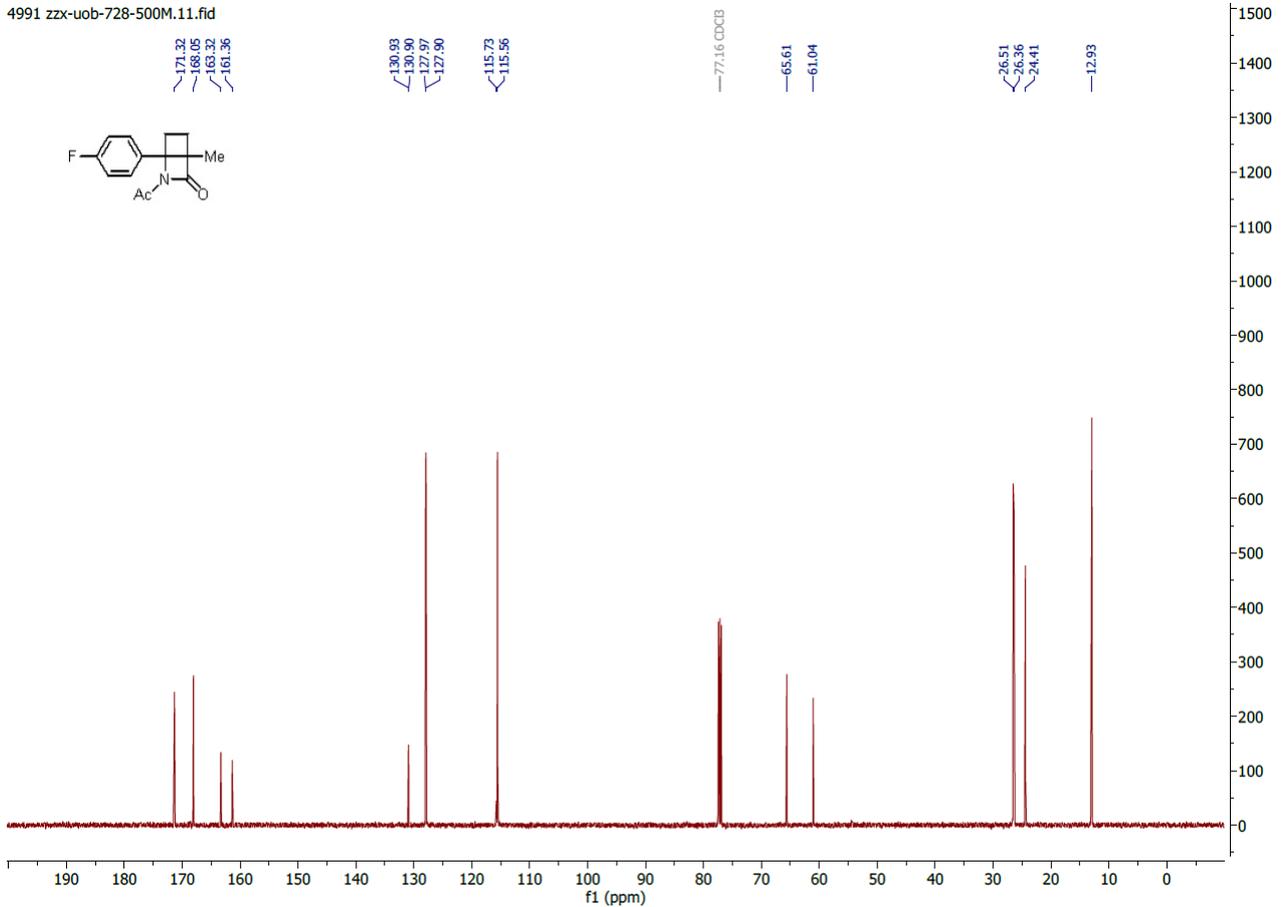


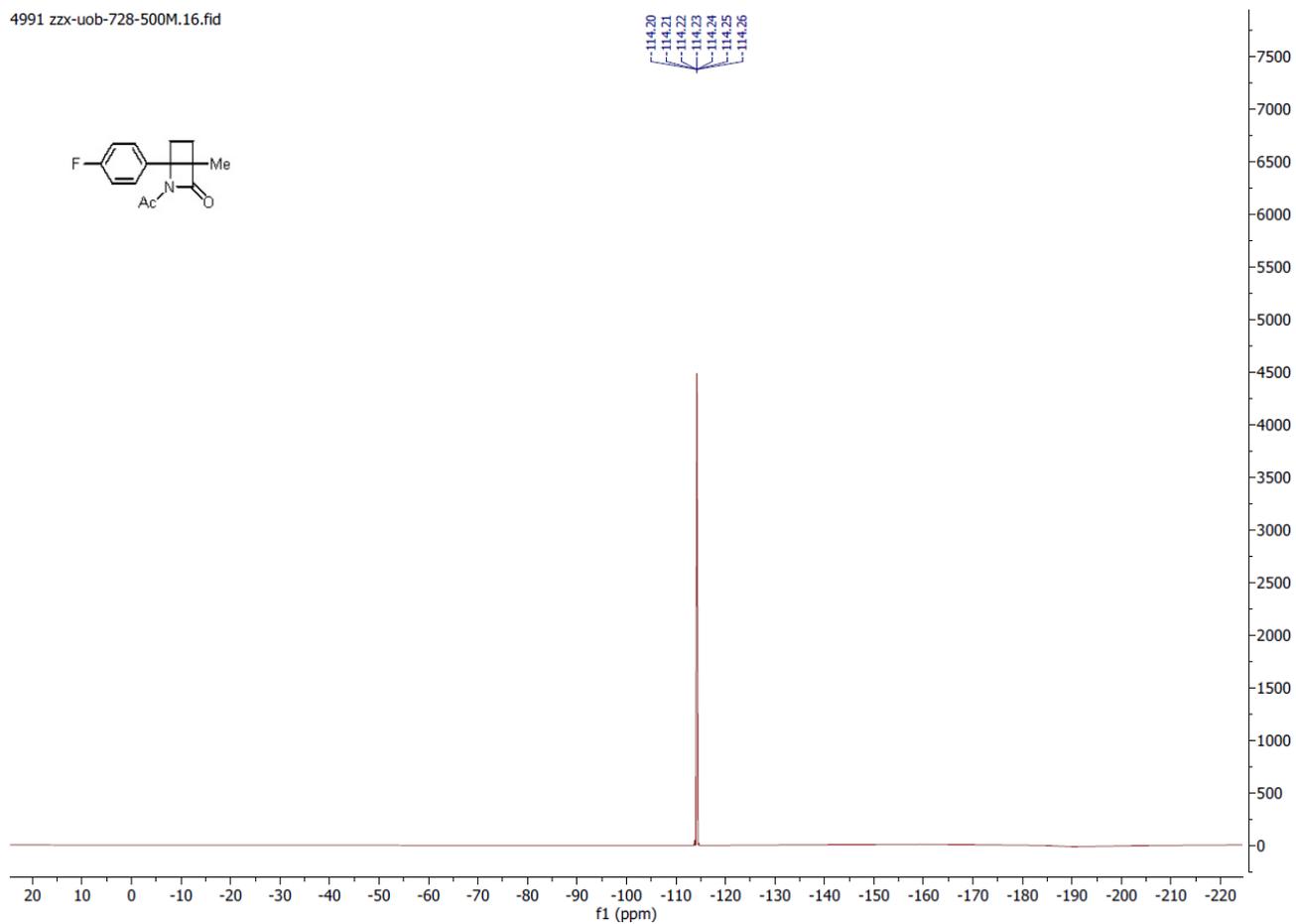
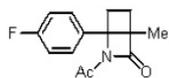
Compound 6l

4991 zzx-uob-728-500M.10.fid



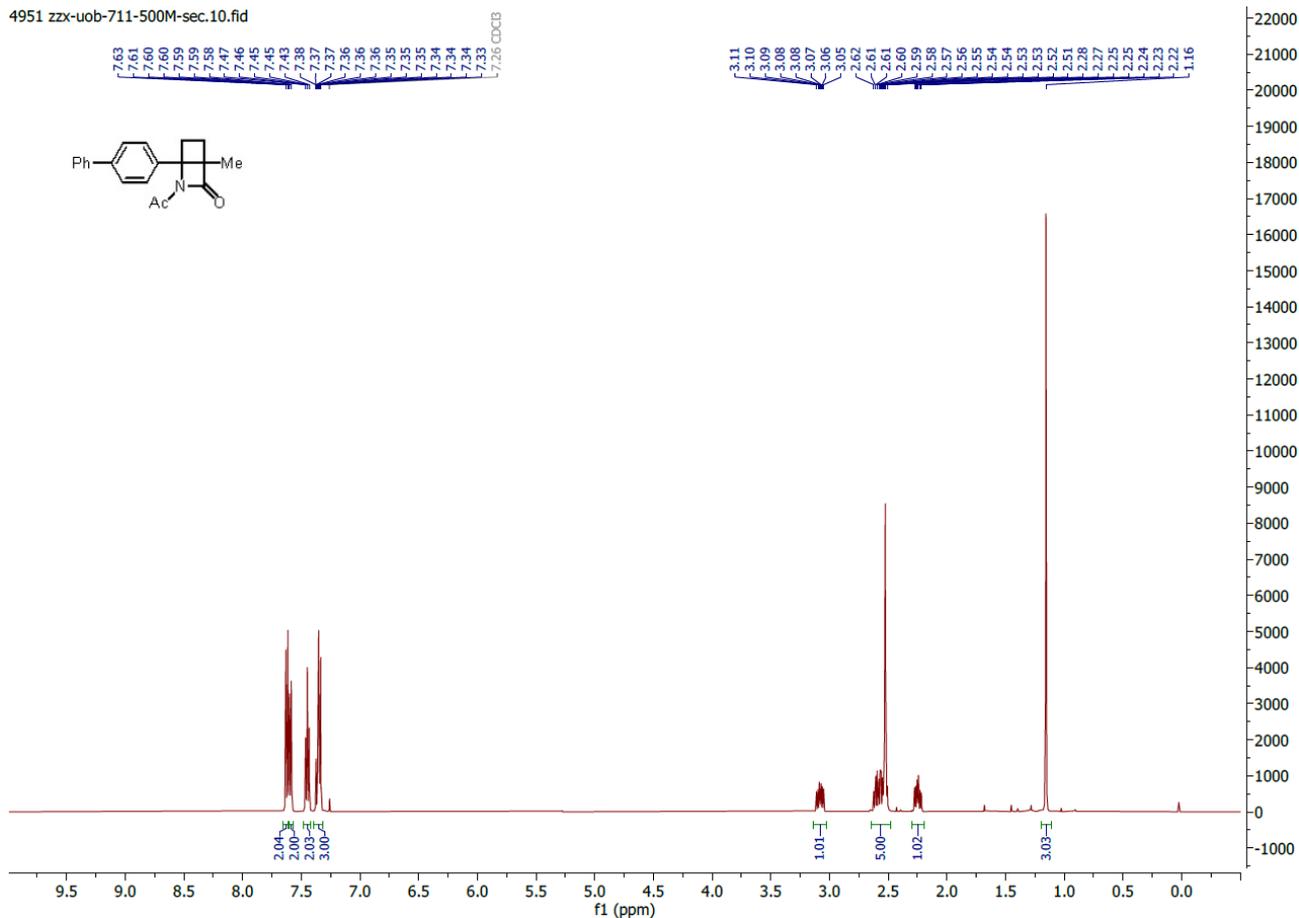
4991 zzx-uob-728-500M.11.fid



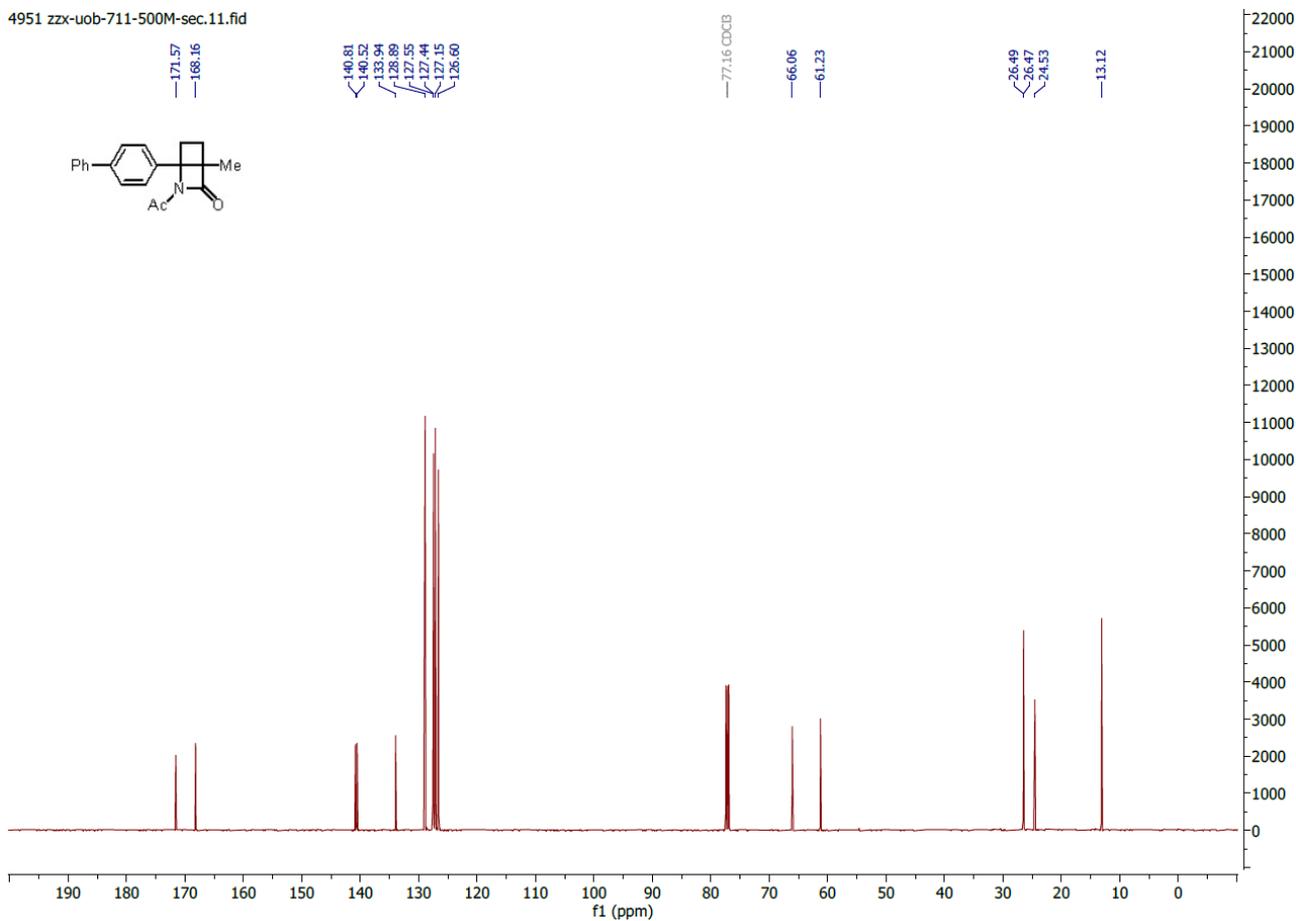


Compound 6m

4951 zzx-uob-711-500M-sec.10.fid

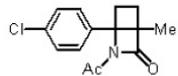


4951 zzx-uob-711-500M-sec.11.fid



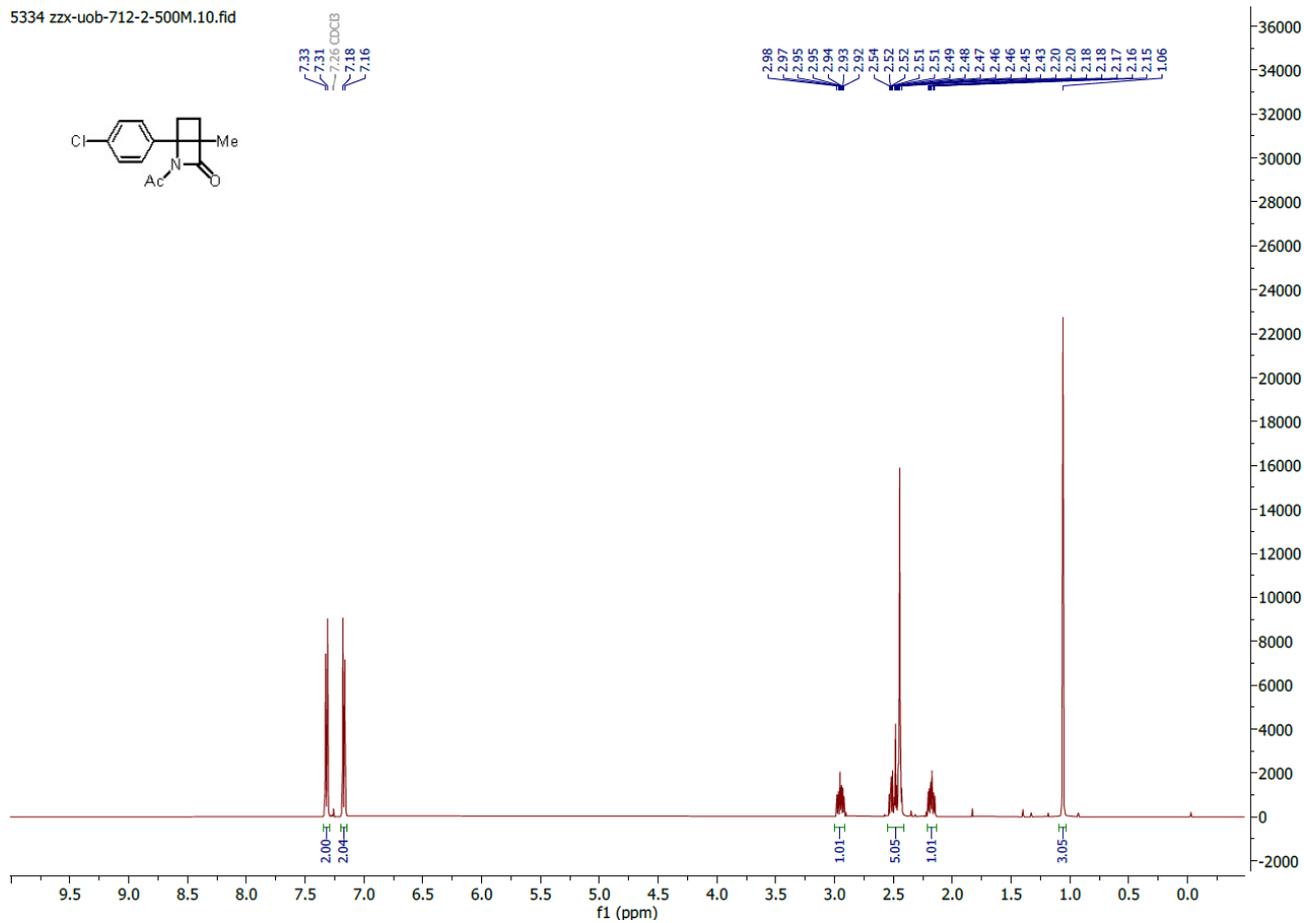
Compound 6n

5334 zzx-uob-712-2-500M.10.fid

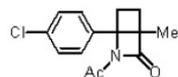


7.33
7.31
7.26 CDCl₃
7.18
7.16

2.98
2.97
2.95
2.94
2.93
2.92
2.54
2.52
2.51
2.49
2.48
2.47
2.46
2.45
2.43
2.20
2.18
2.17
2.16
2.15
1.06



5334 zzx-uob-712-2-500M.11.fid



171.05
167.90

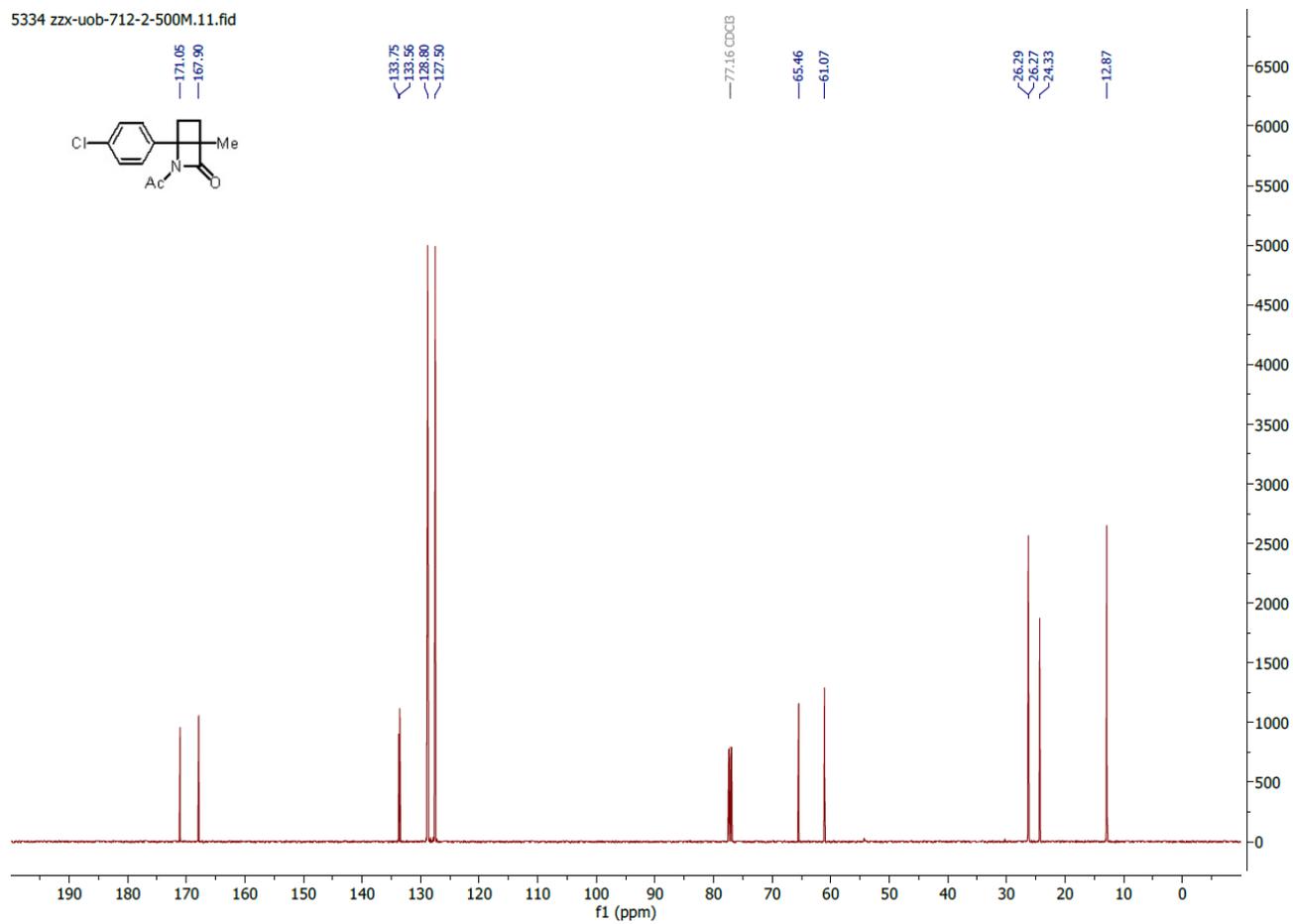
133.75
133.56
128.80
127.50

77.16 CDCl₃

65.46
61.07

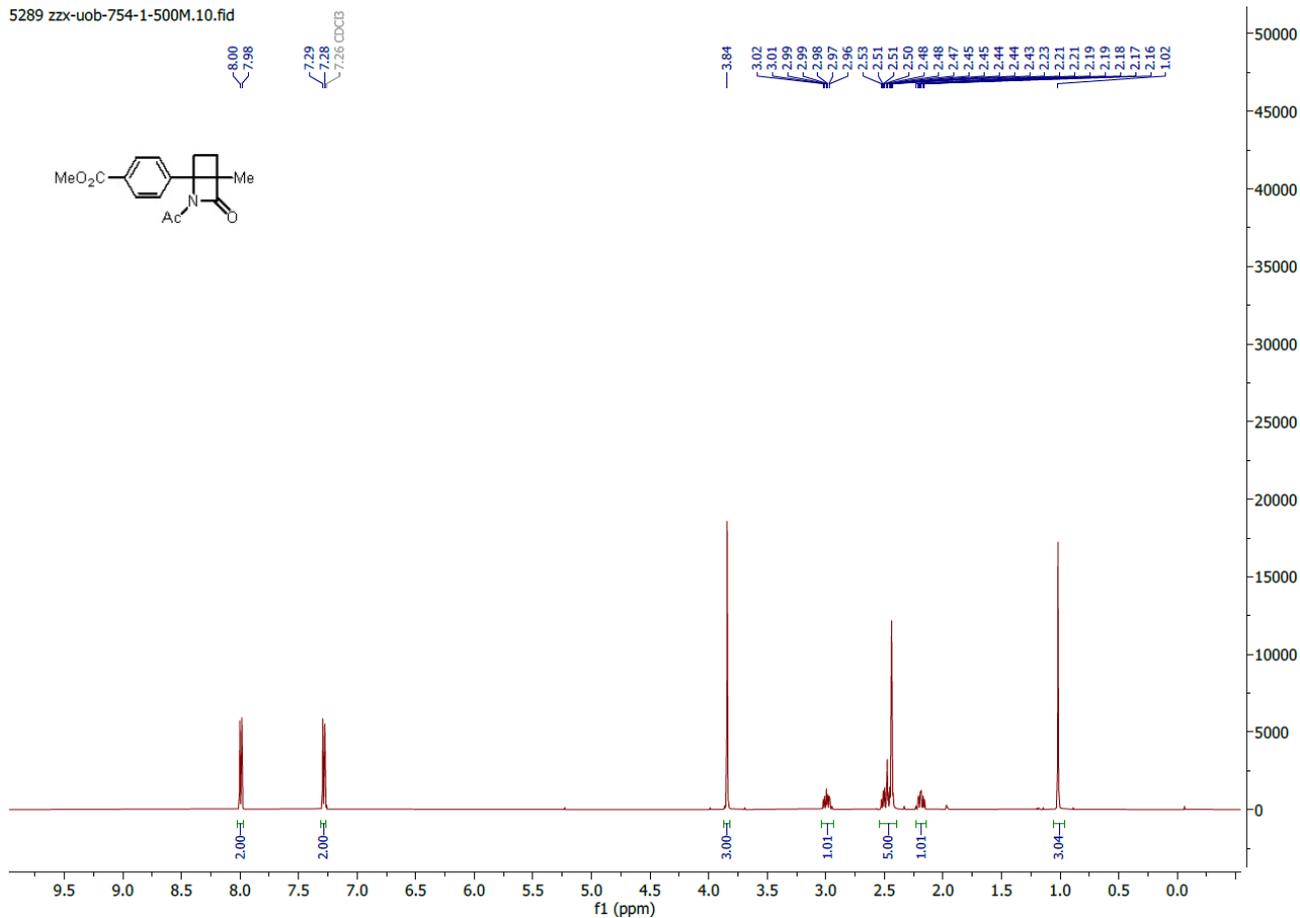
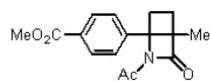
26.29
26.27
24.33

12.87

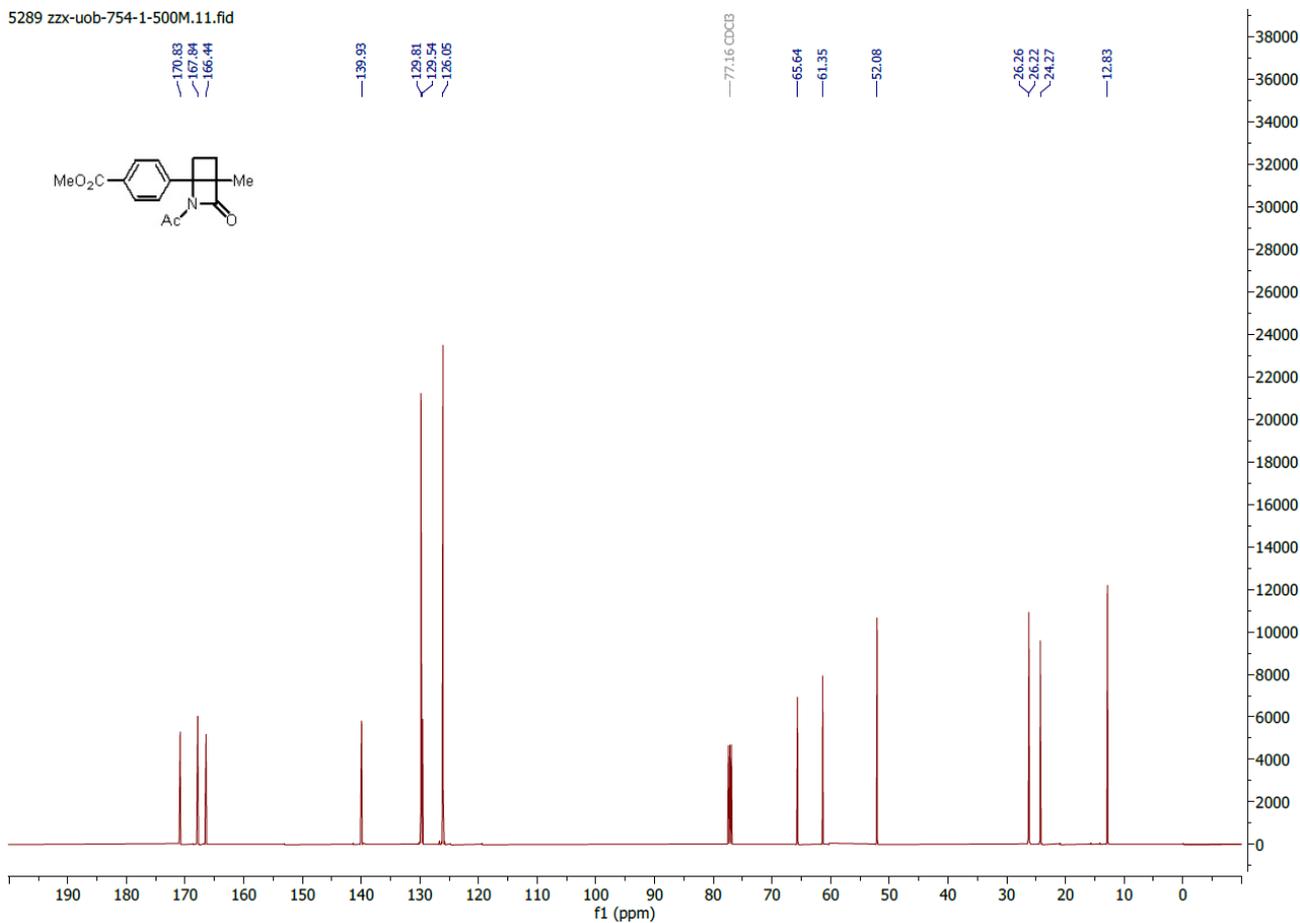
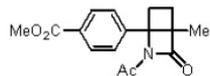


Compound 60

5289 zzx-uob-754-1-500M.10.fid

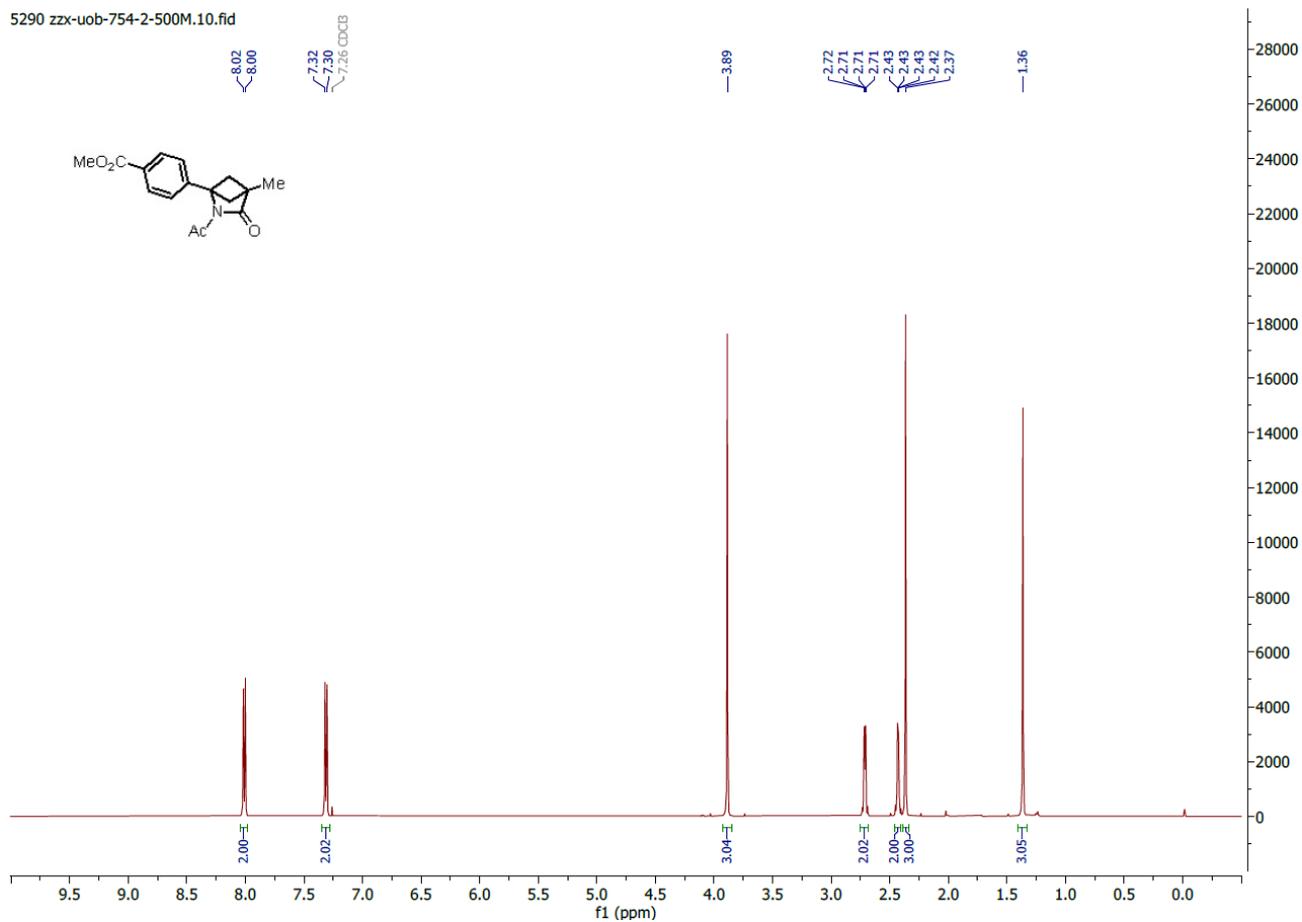


5289 zzx-uob-754-1-500M.11.fid

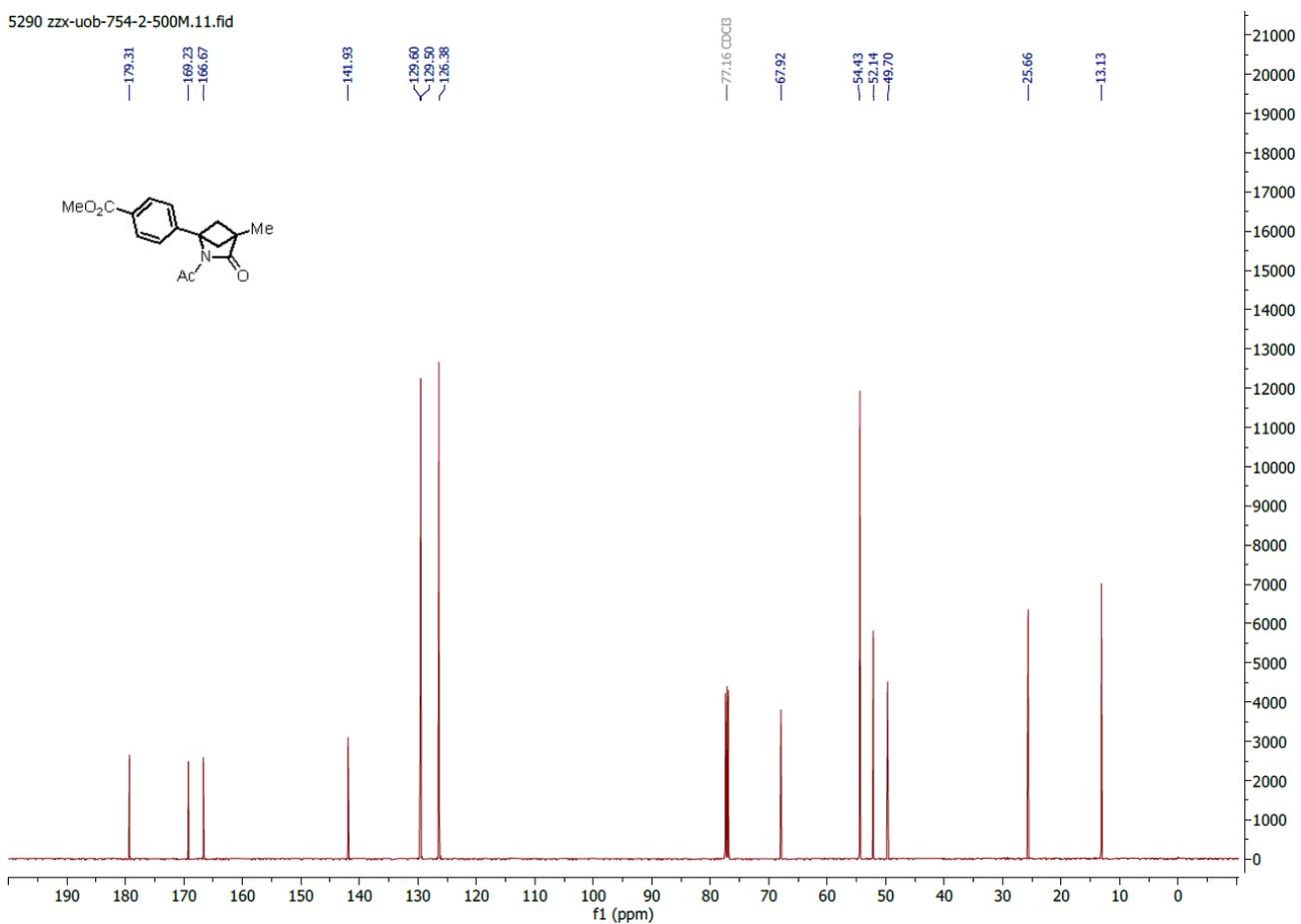


Compound 50

5290 zzx-uob-754-2-500M.10.fid

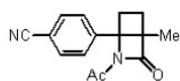


5290 zzx-uob-754-2-500M.11.fid



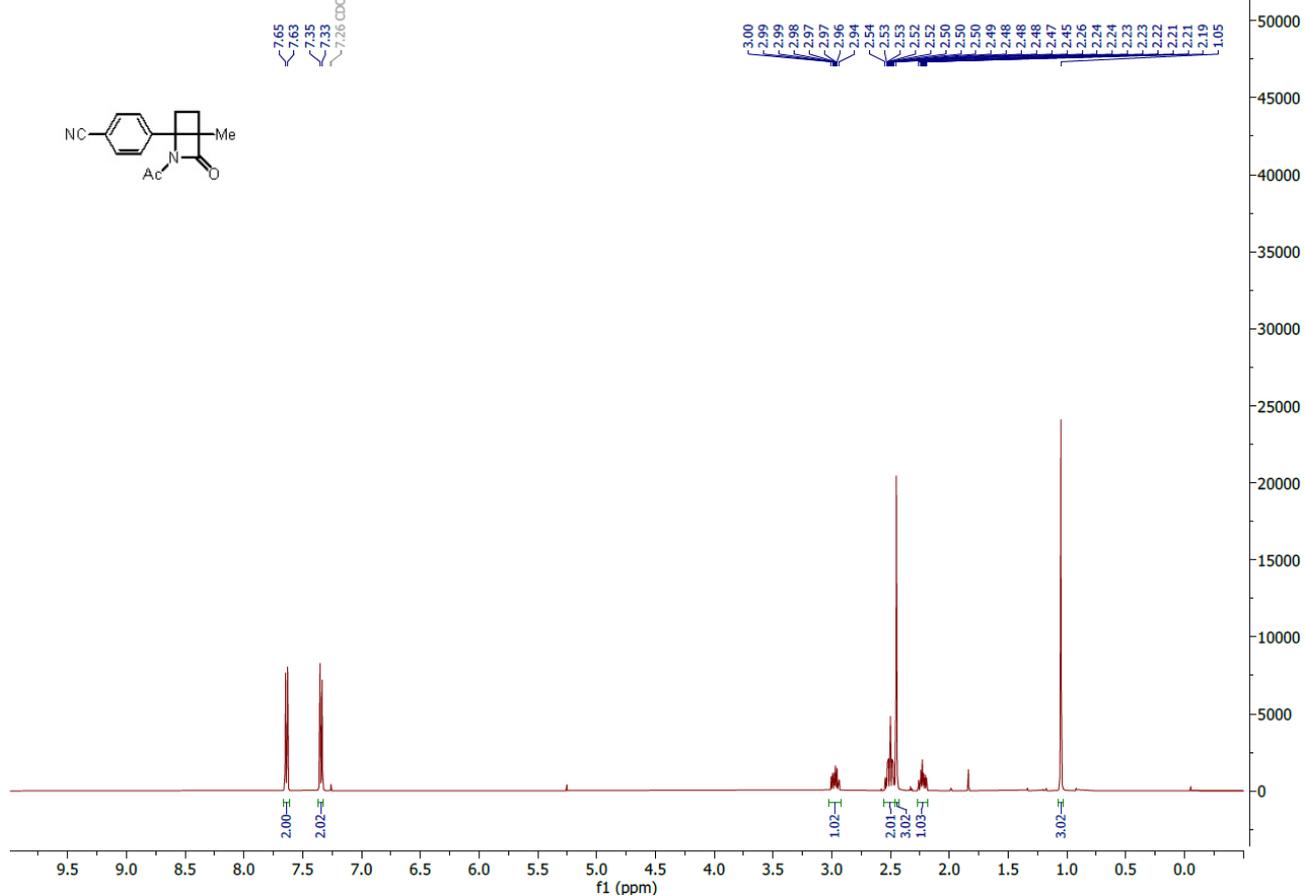
Compound 6p

5287 zzx-uob-752-1-500M.10.fid

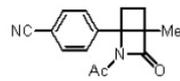


7.65
7.63
7.35
7.33
7.26 CDCl₃

3.00
2.99
2.99
2.98
2.97
2.97
2.96
2.94
2.54
2.53
2.53
2.52
2.50
2.50
2.50
2.48
2.48
2.48
2.47
2.45
2.26
2.24
2.24
2.23
2.23
2.22
2.21
2.21
2.19
1.05



5287 zzx-uob-752-1-500M.11.fid



170.44
167.88

140.30

132.38

126.87

118.39

111.62

77.16 CDCl₃

65.34

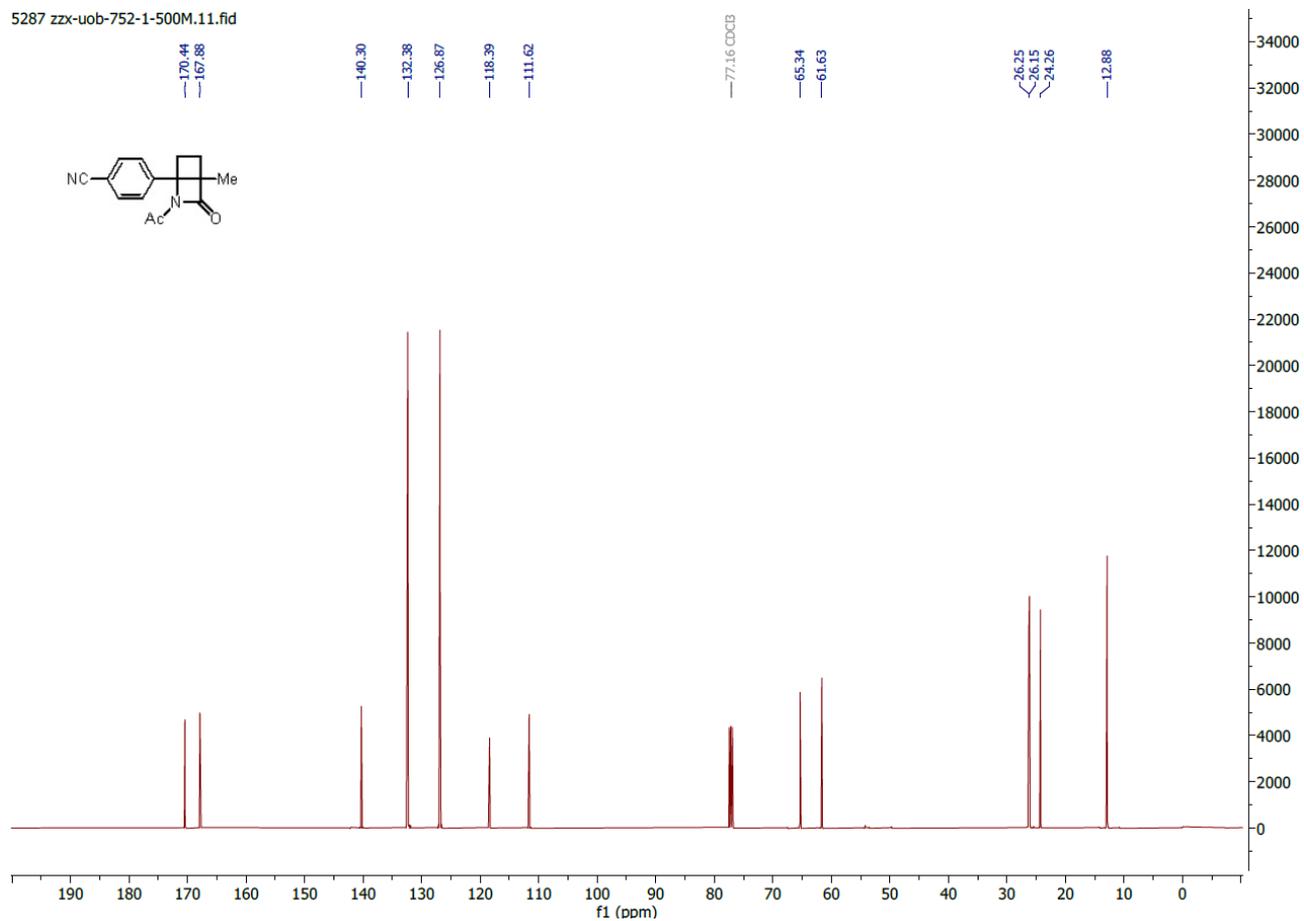
61.63

26.25

26.15

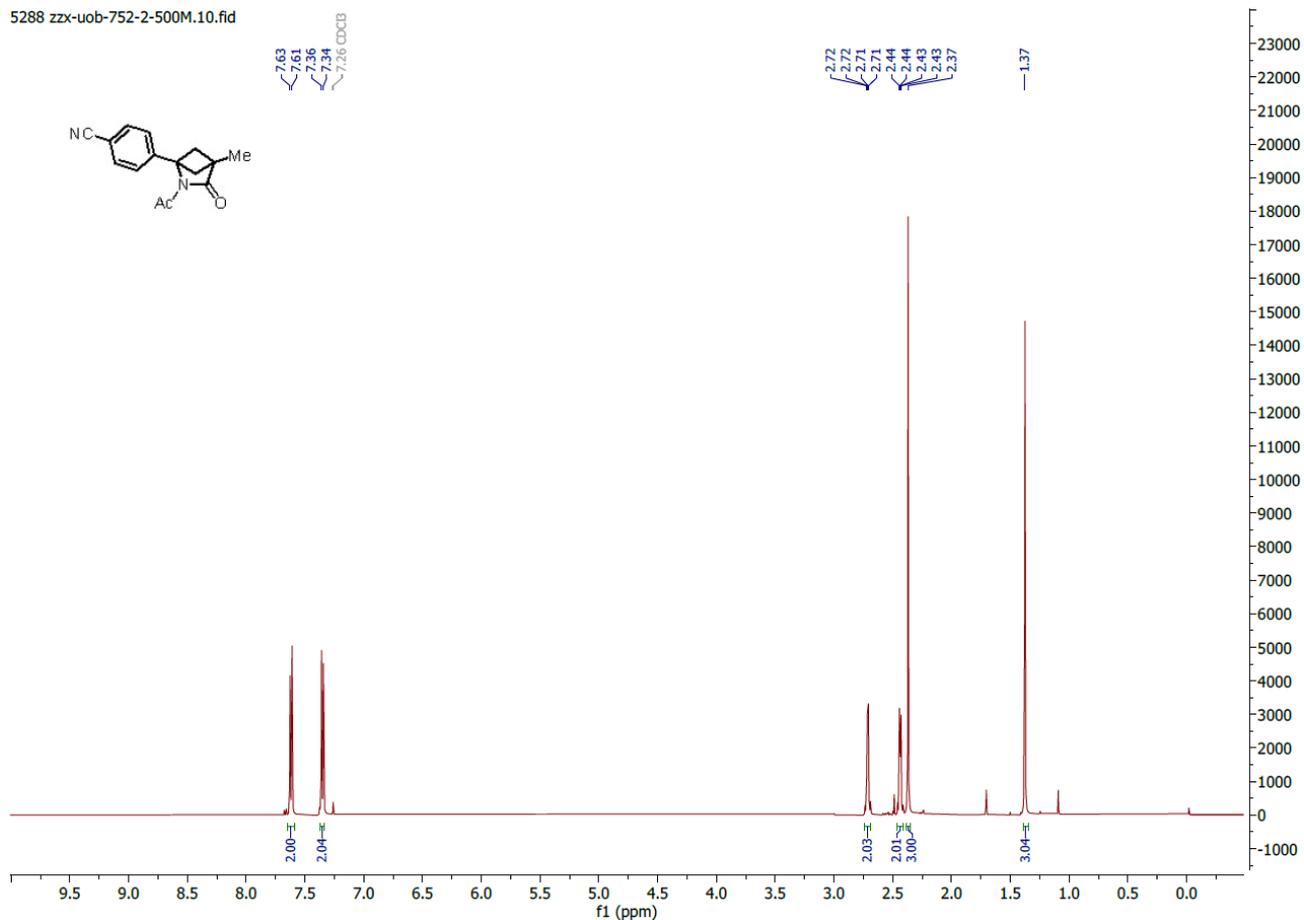
24.26

12.88

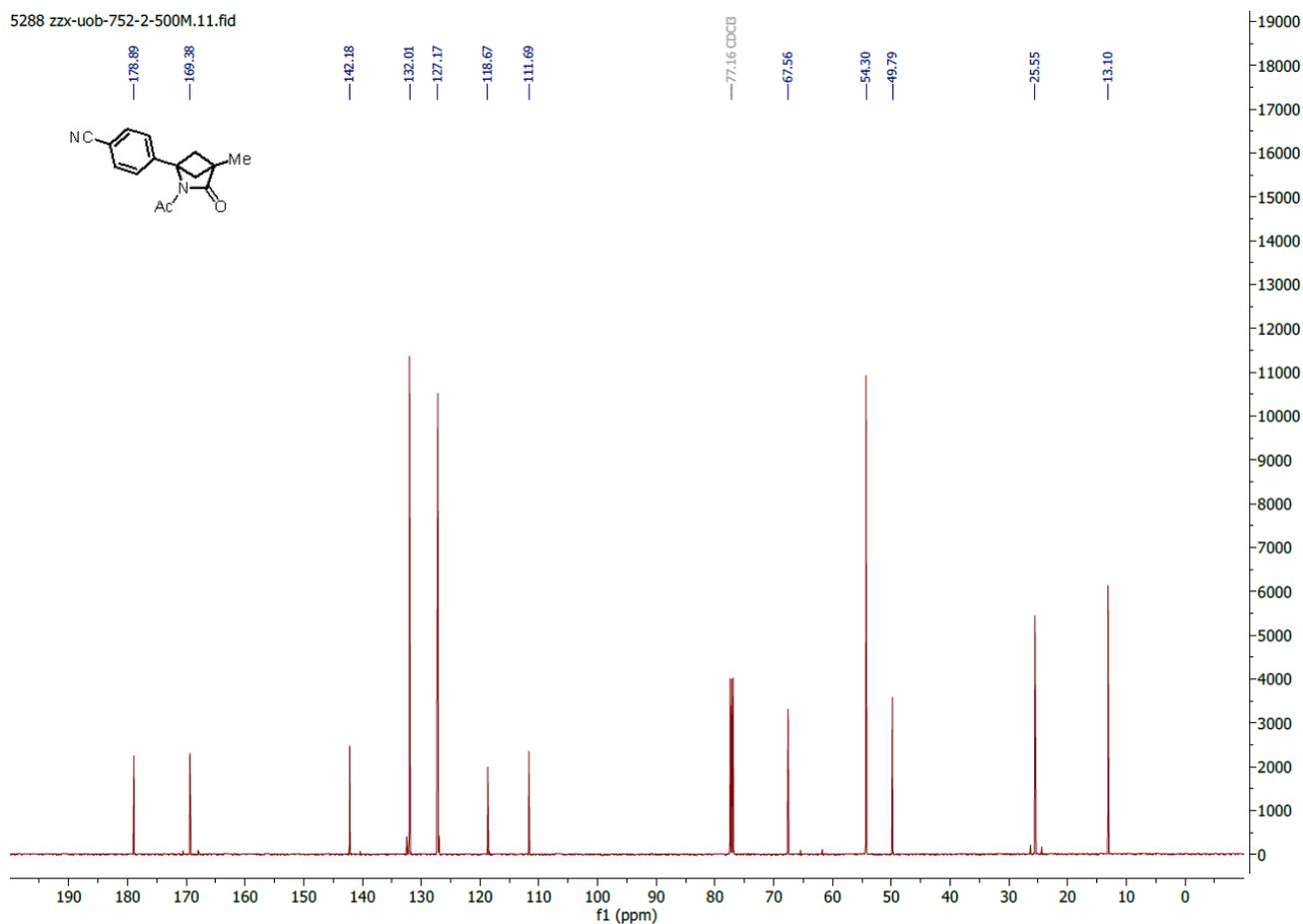


Compound 5p

5288 zzx-uob-752-2-500M.10.fid

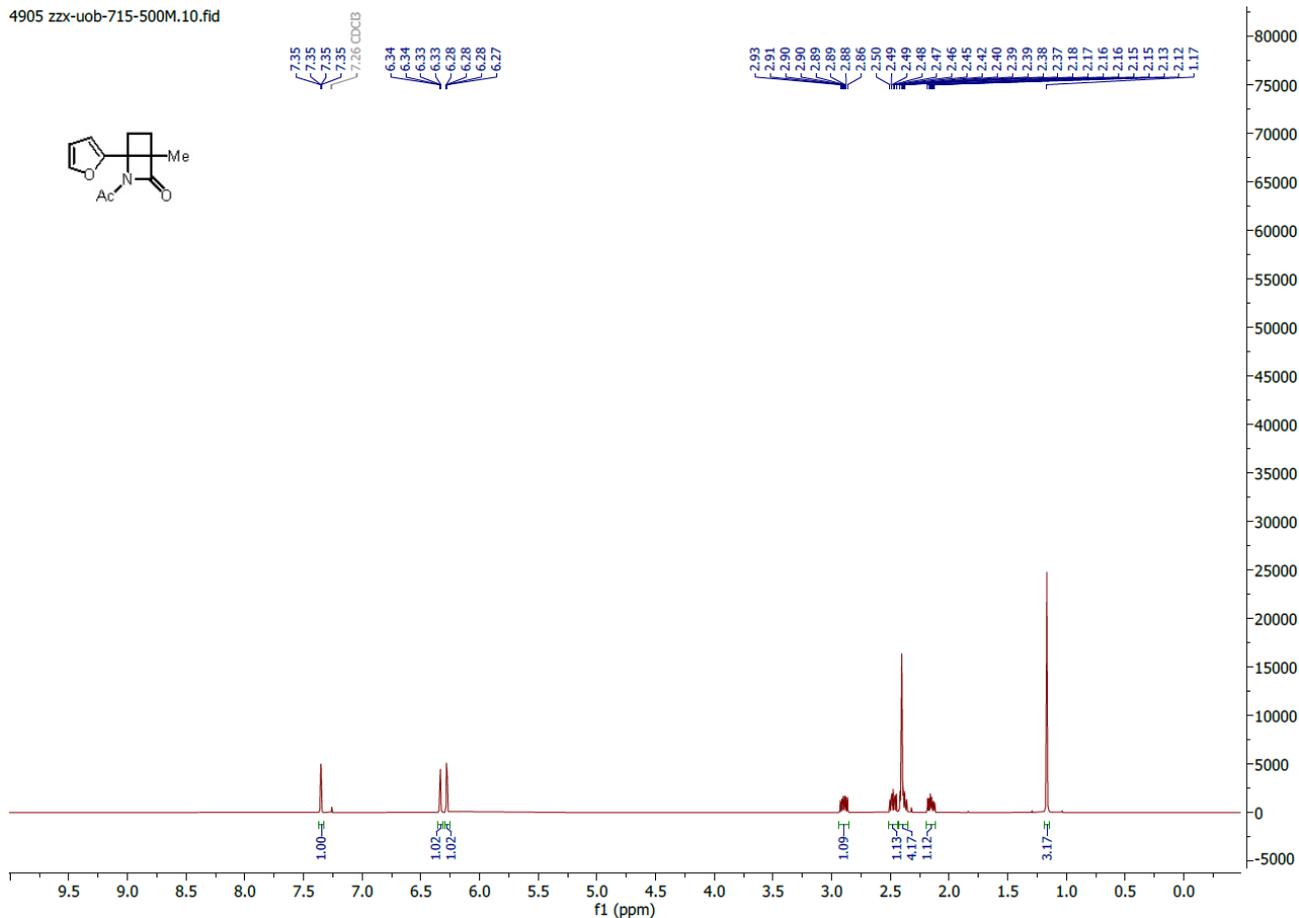
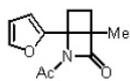


5288 zzx-uob-752-2-500M.11.fid

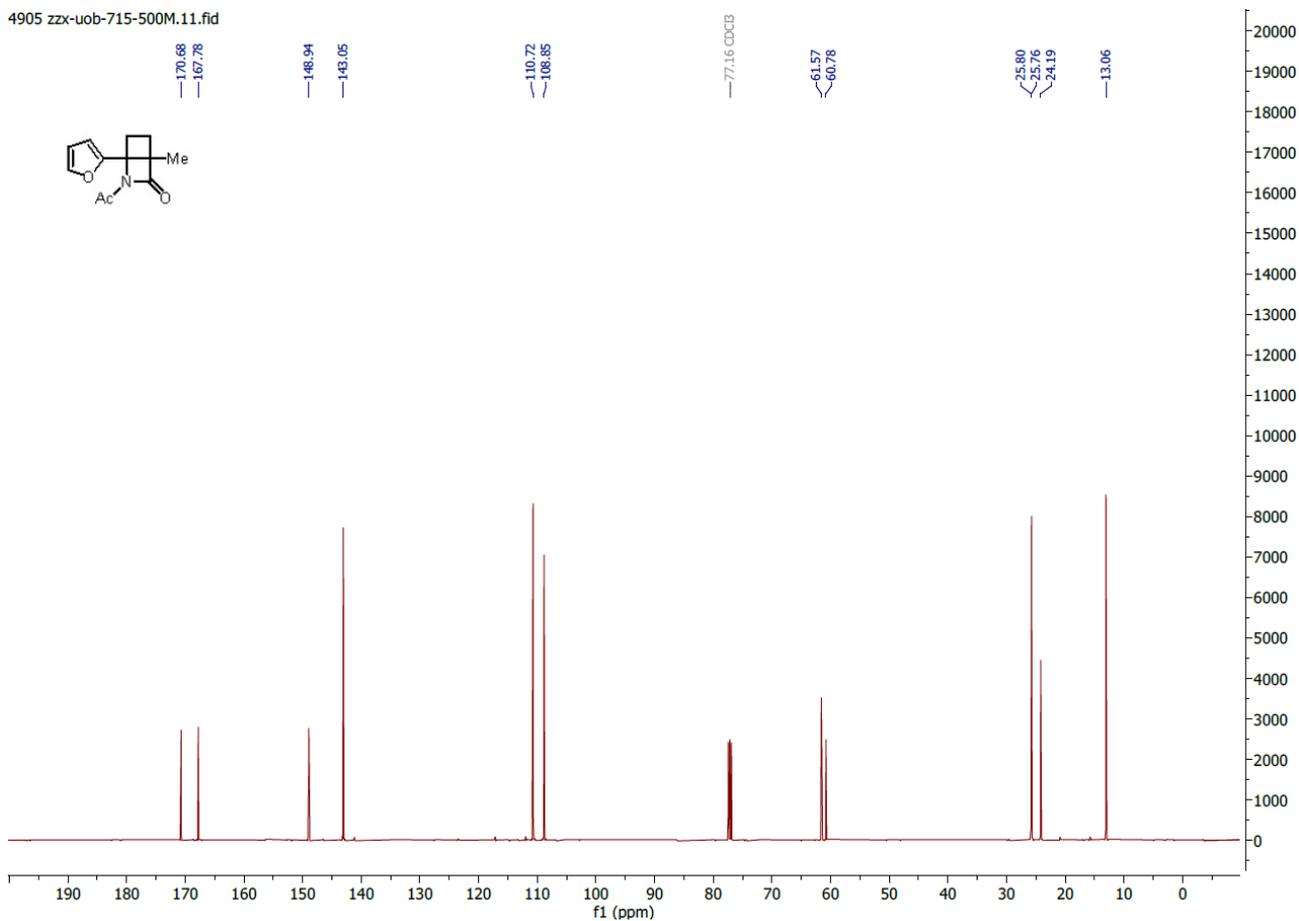
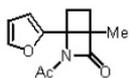


Compound 6q

4905 zzx-uob-715-500M.10.fid

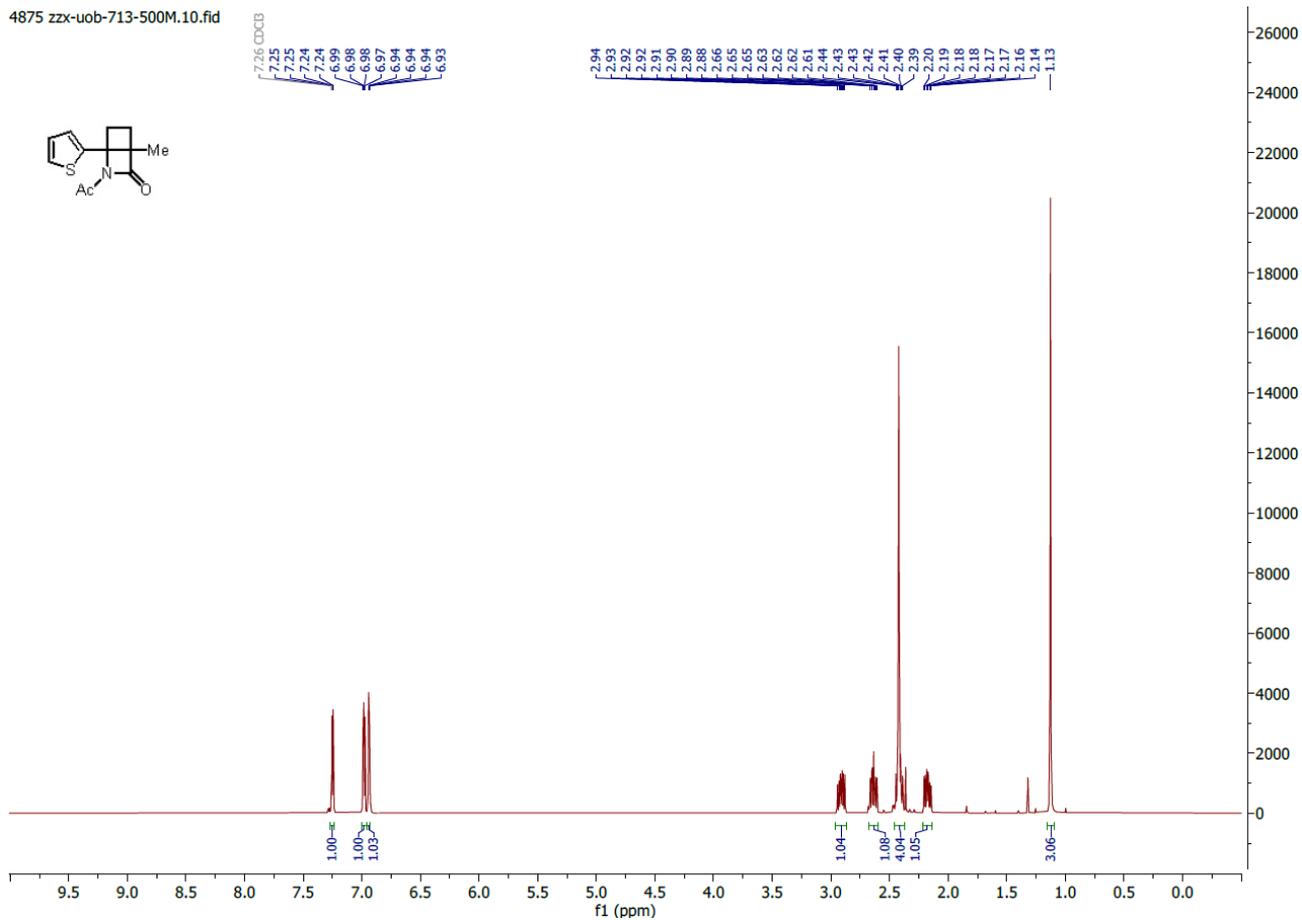


4905 zzx-uob-715-500M.11.fid

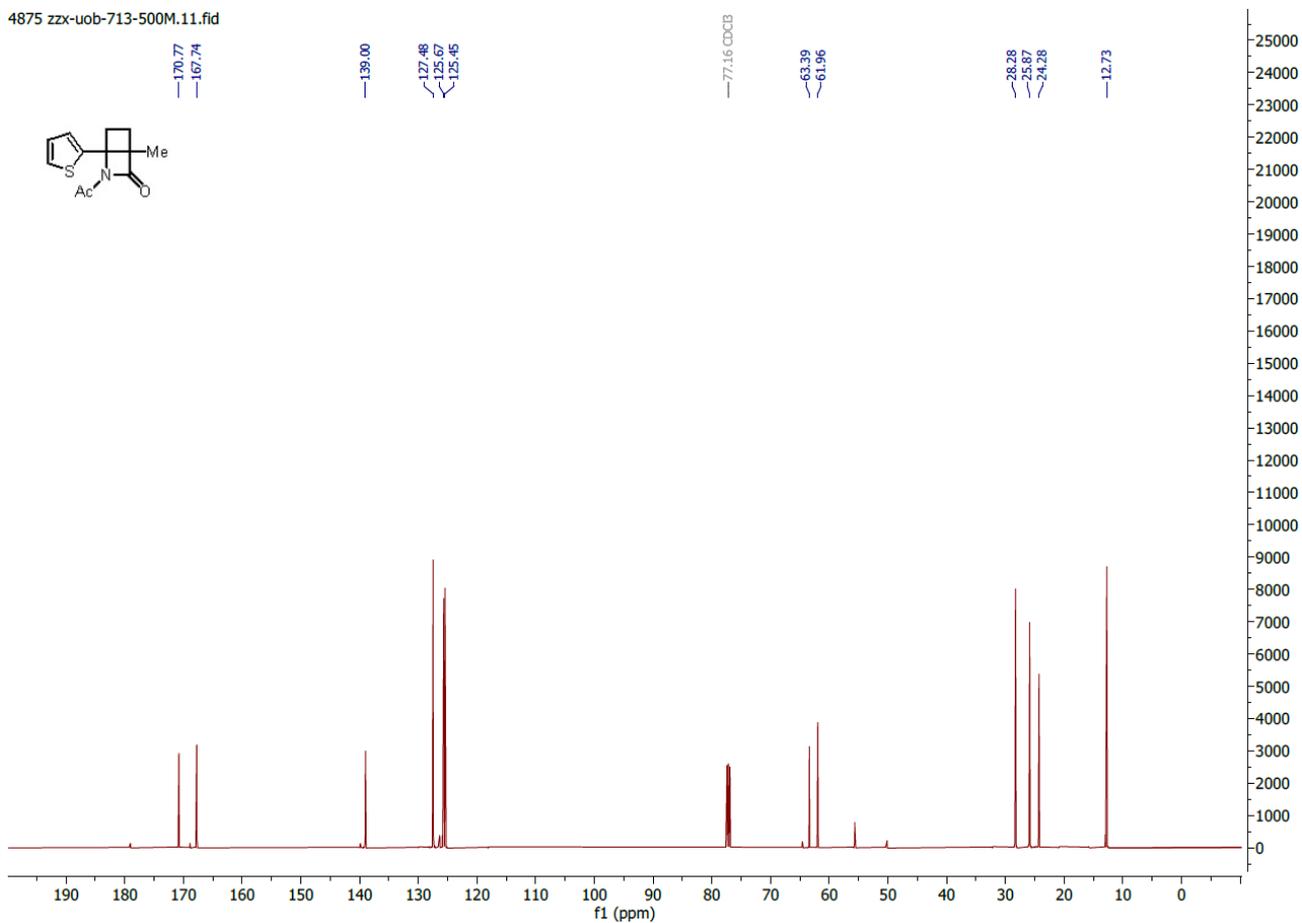


Compound 6r

4875 zzx-uob-713-500M.10.fid

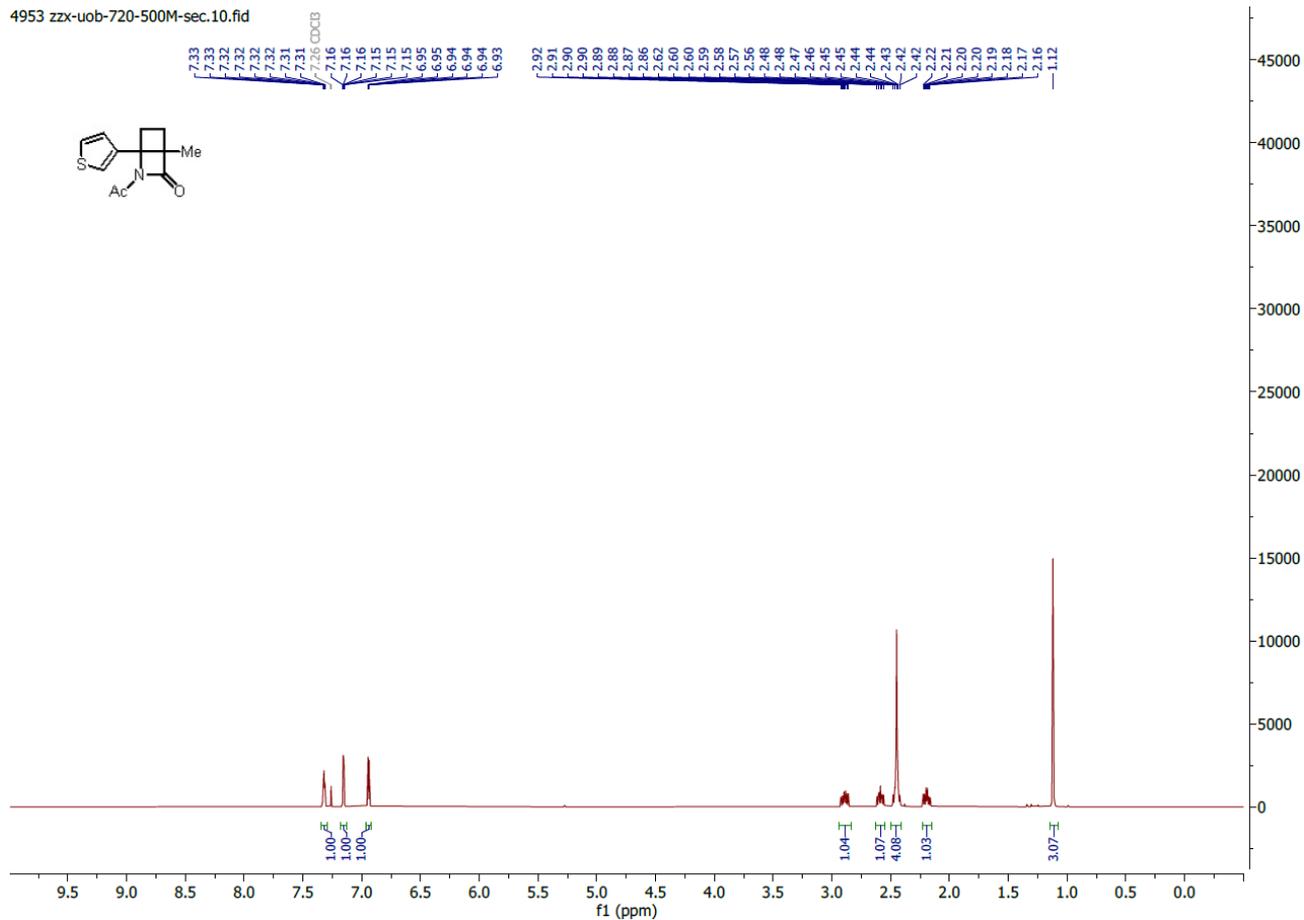
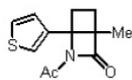


4875 zzx-uob-713-500M.11.fid

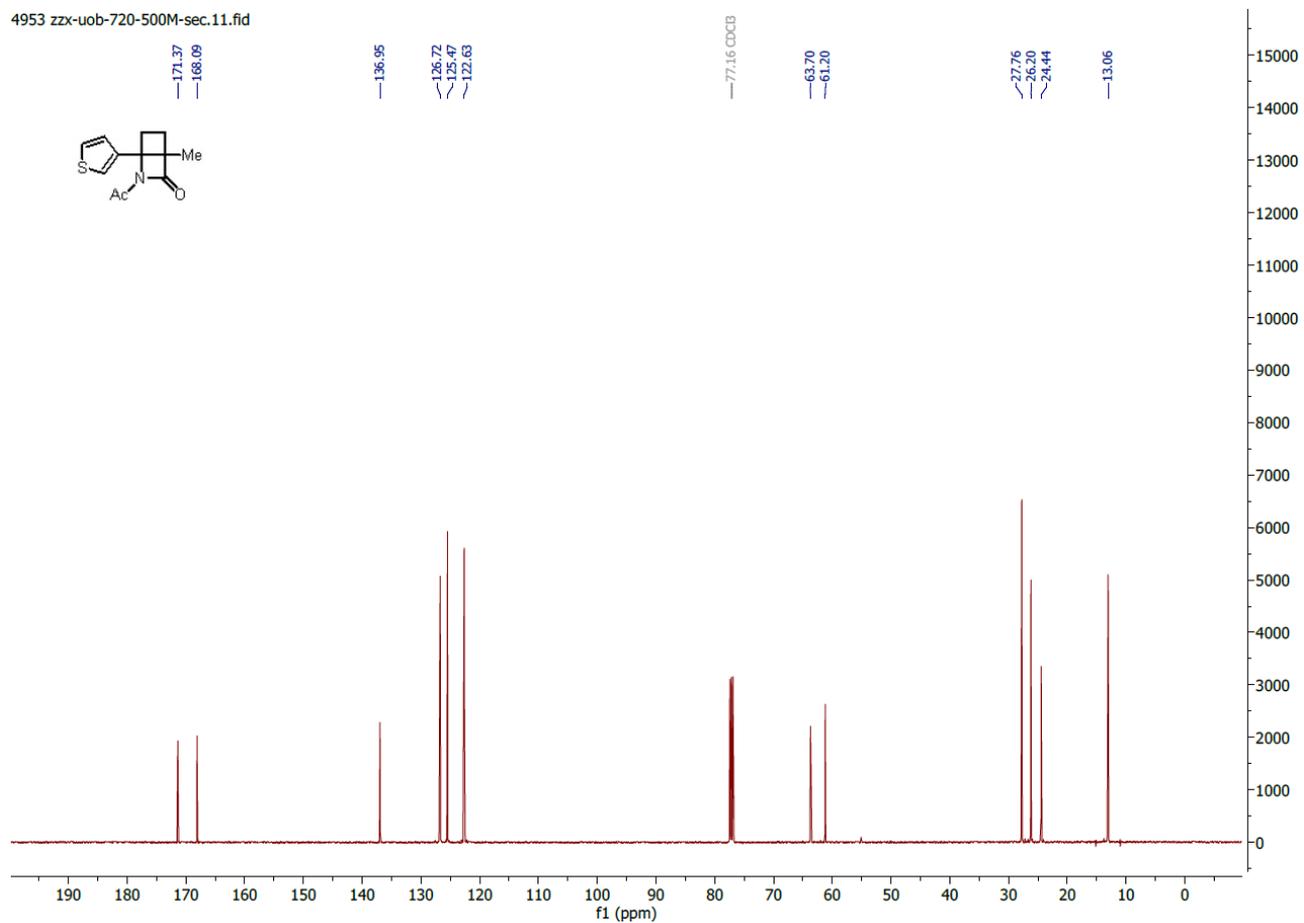
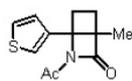


Compound 6s

4953 zzx-uob-720-500M-sec.10.fid

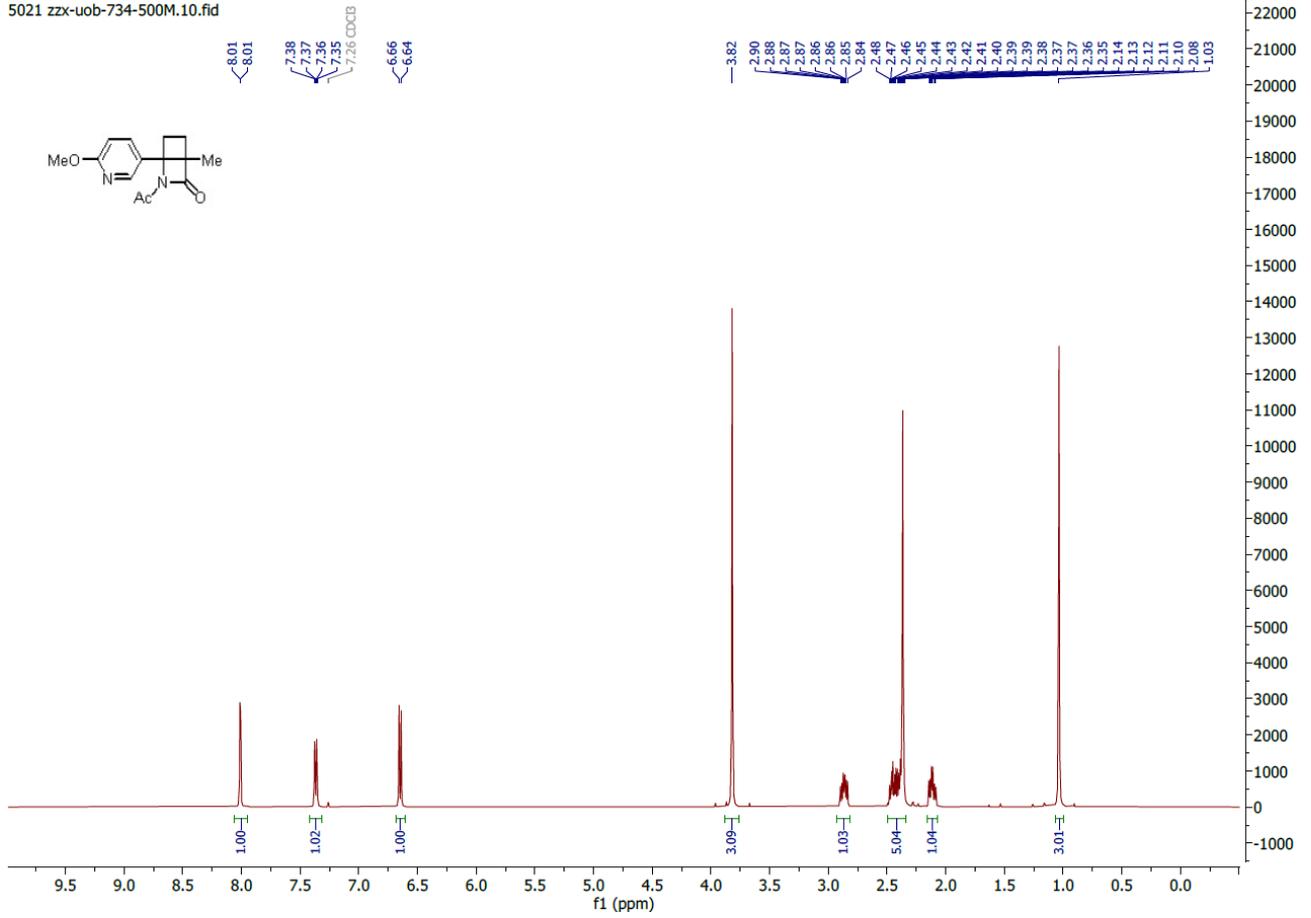


4953 zzx-uob-720-500M-sec.11.fid

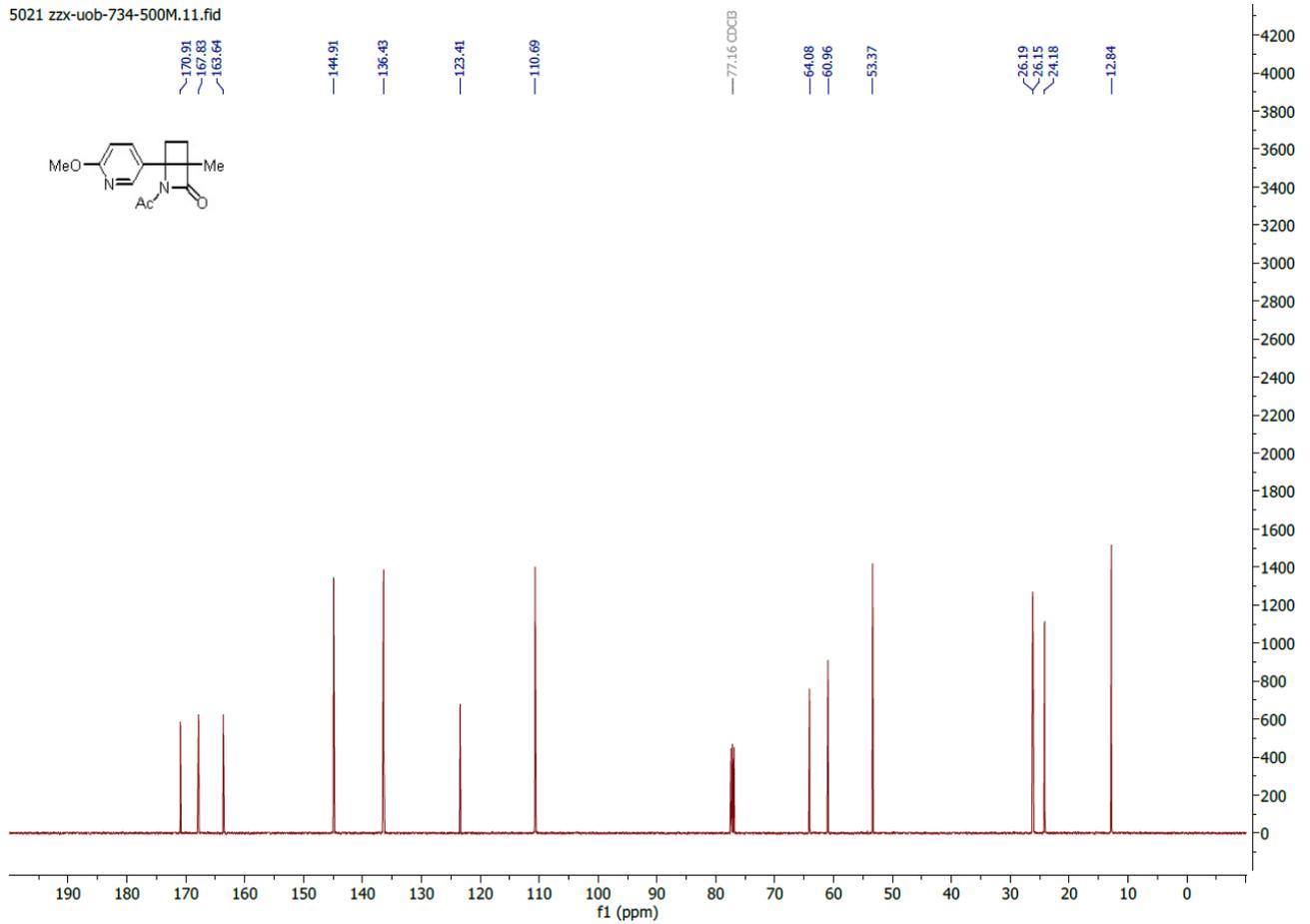


Compound 6t

5021 zzx-uob-734-500M.10.fid

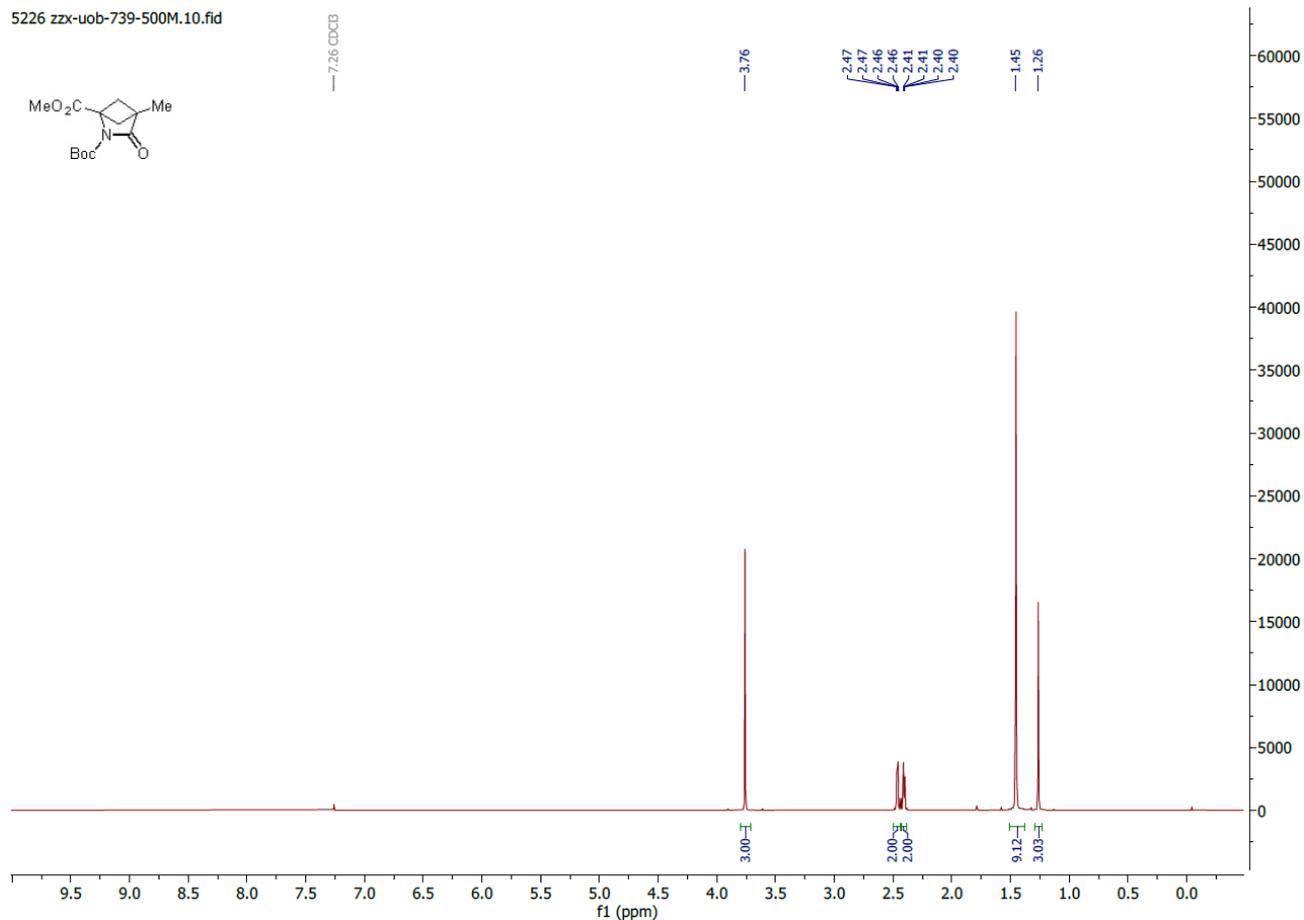


5021 zzx-uob-734-500M.11.fid

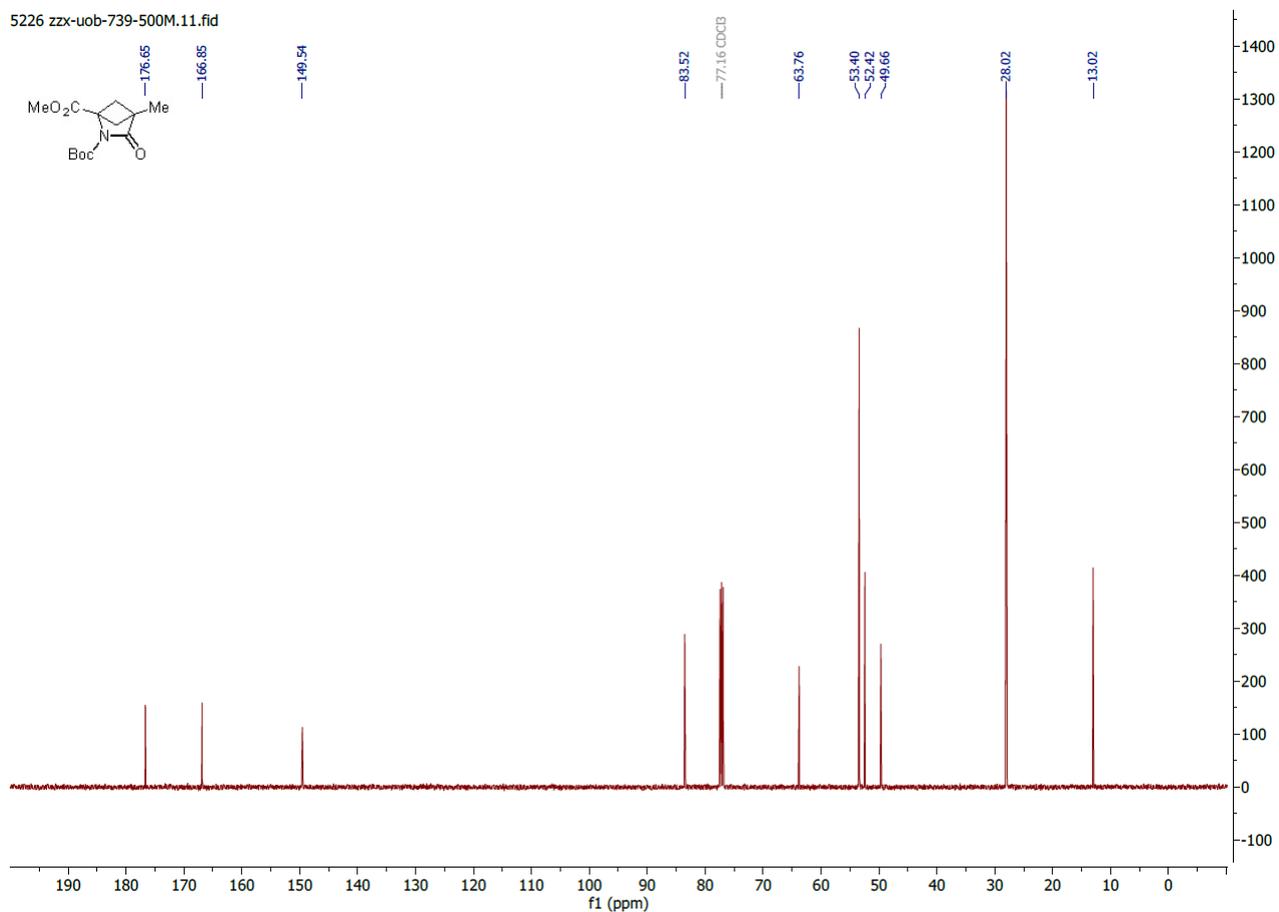


Compound 5u

5226 zzx-uob-739-500M.10.fid

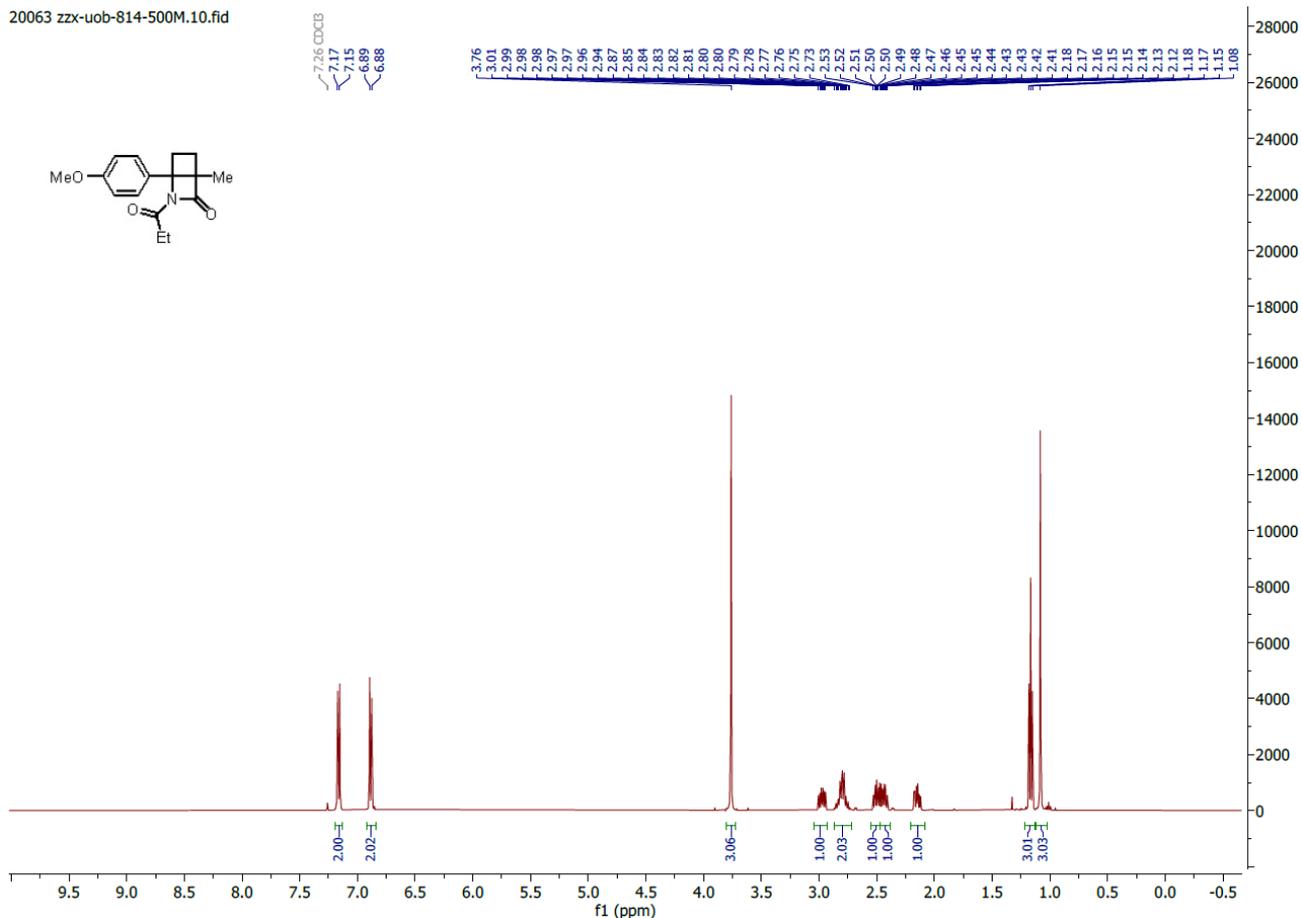


5226 zzx-uob-739-500M.11.fid

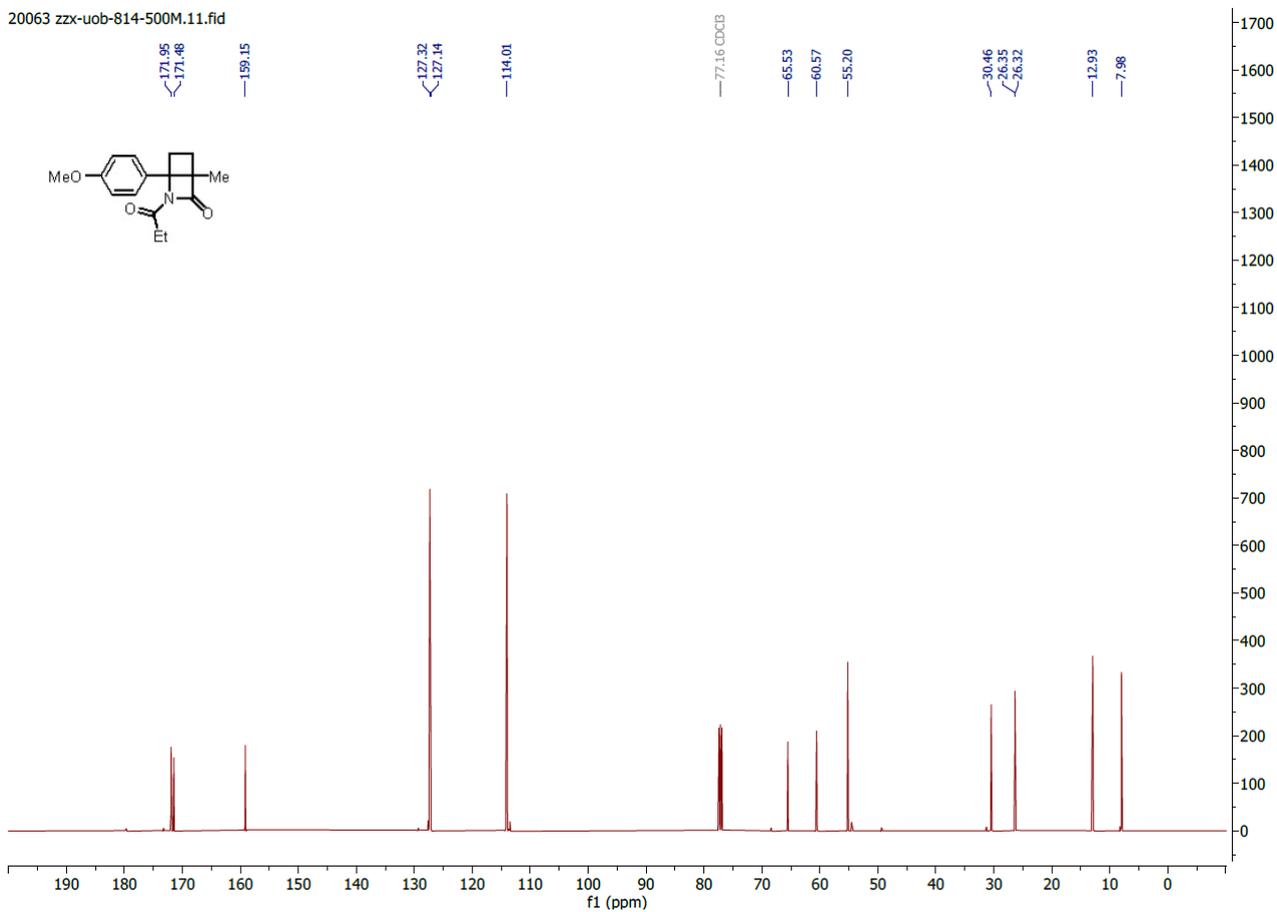


Compound 6w

20063 zzx-uob-814-500M.10.fid

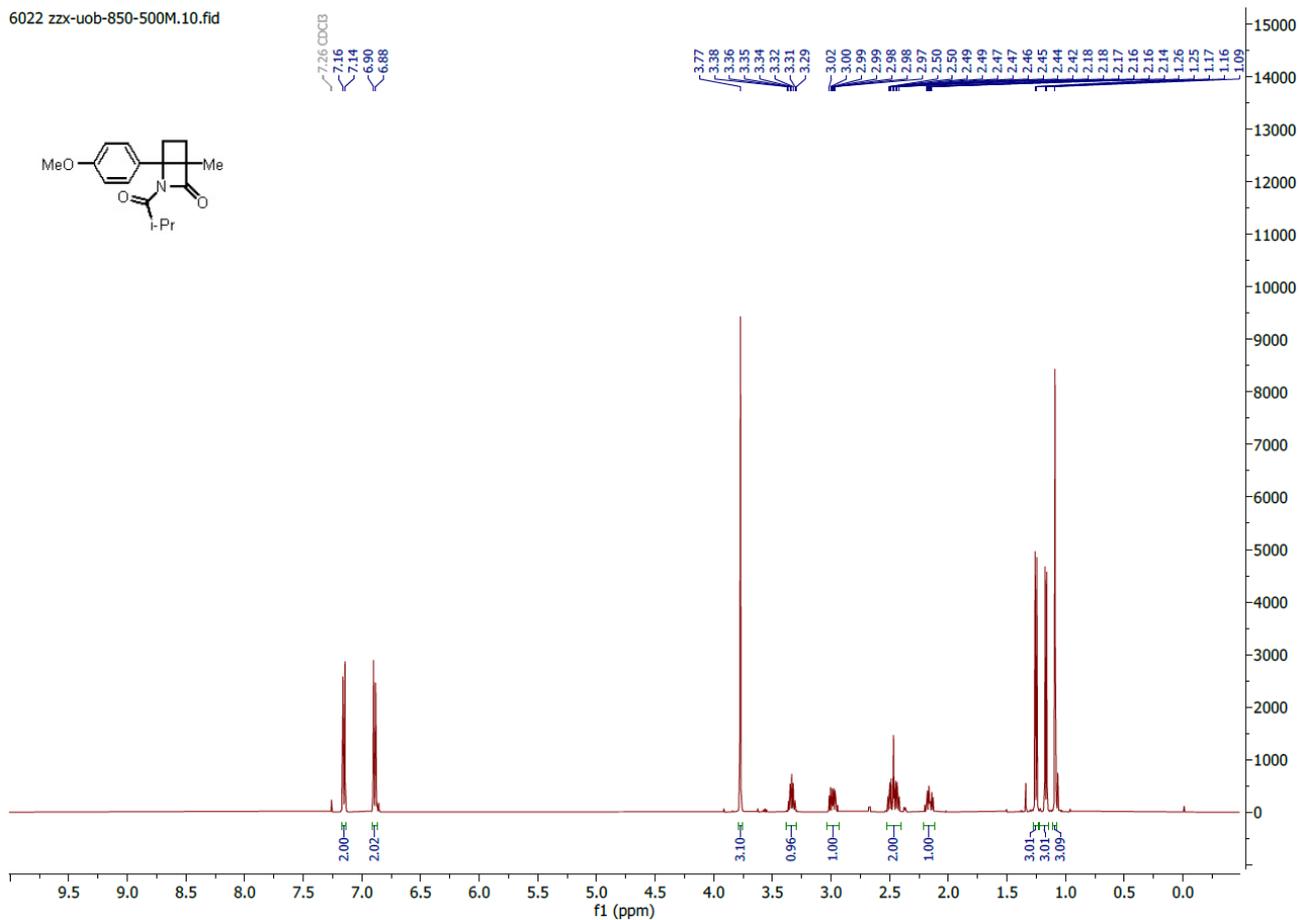
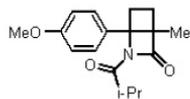


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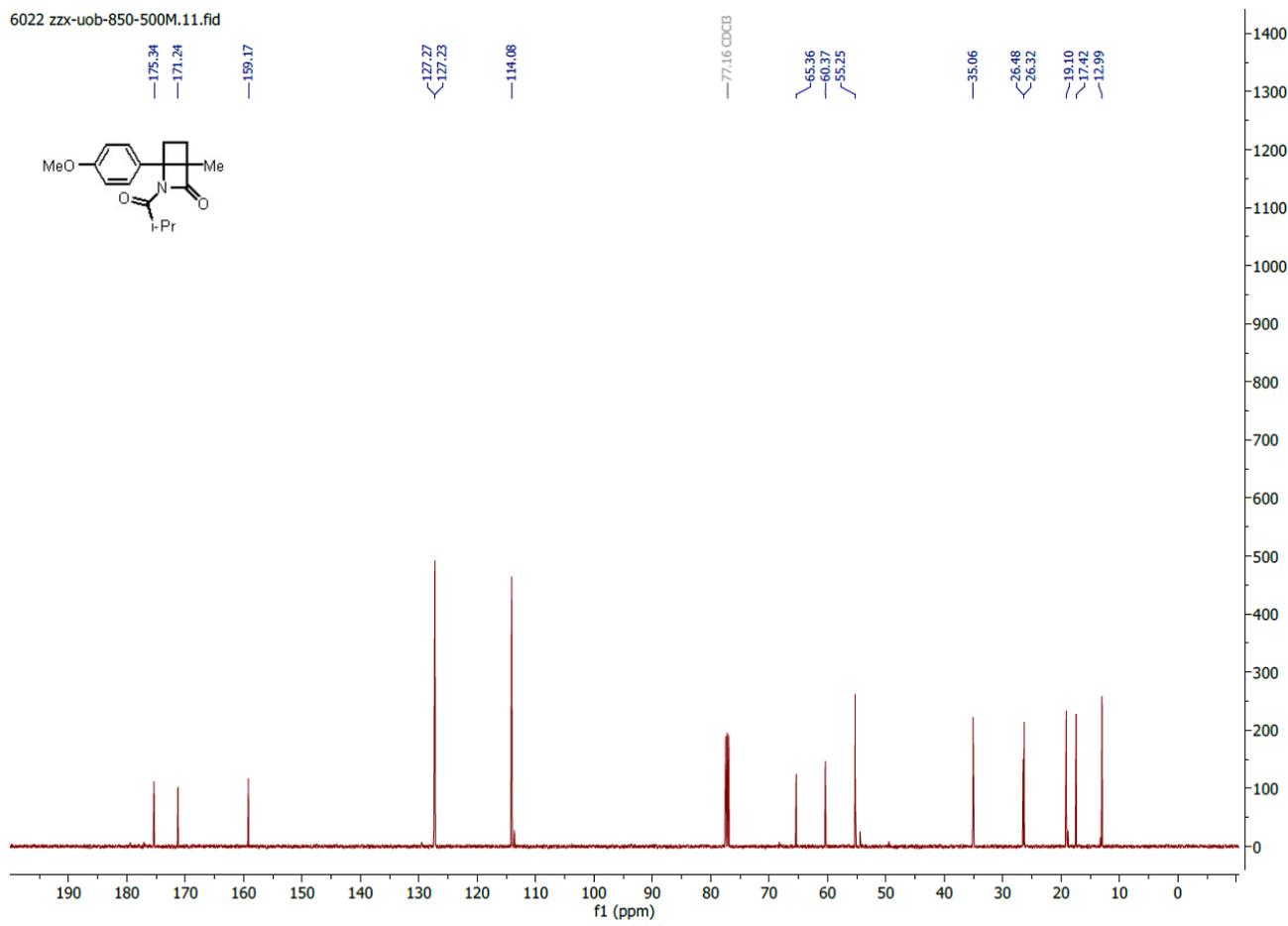
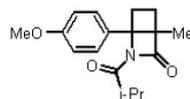


Compound 6x

6022 zzx-uob-850-500M.10.fid

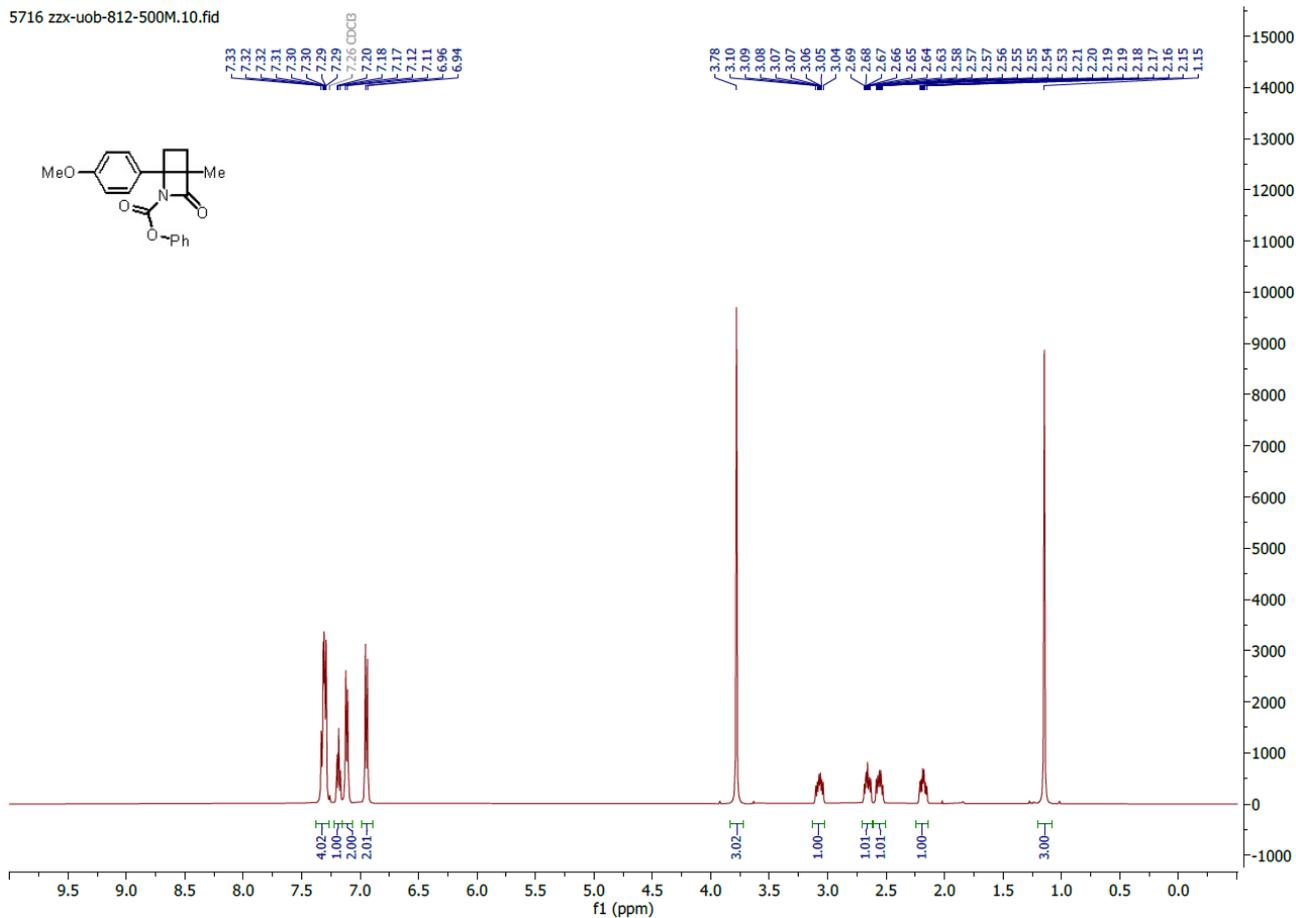


6022 zzx-uob-850-500M.11.fid

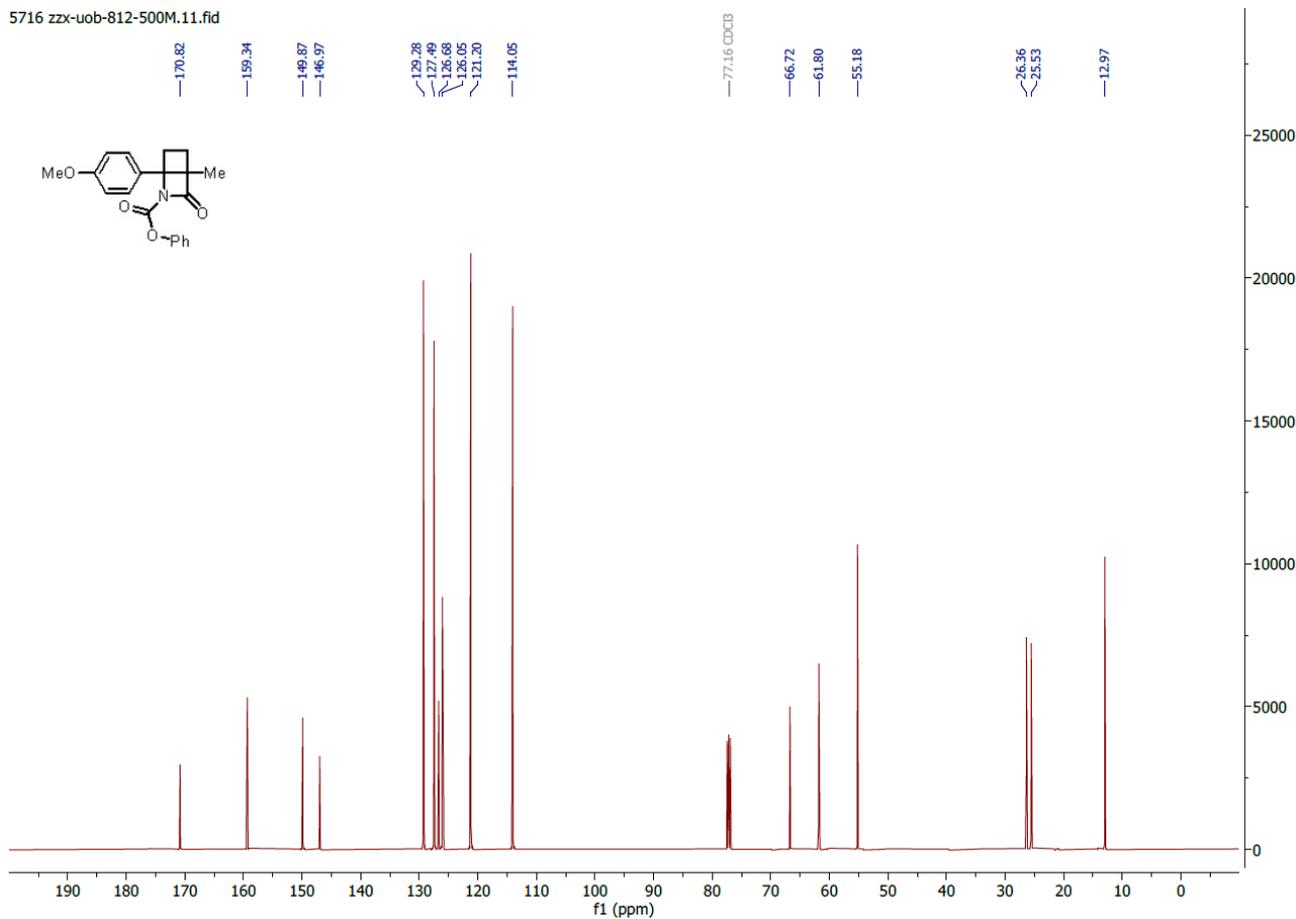


Compound 6y

5716 zzx-uob-812-500M.10.fid

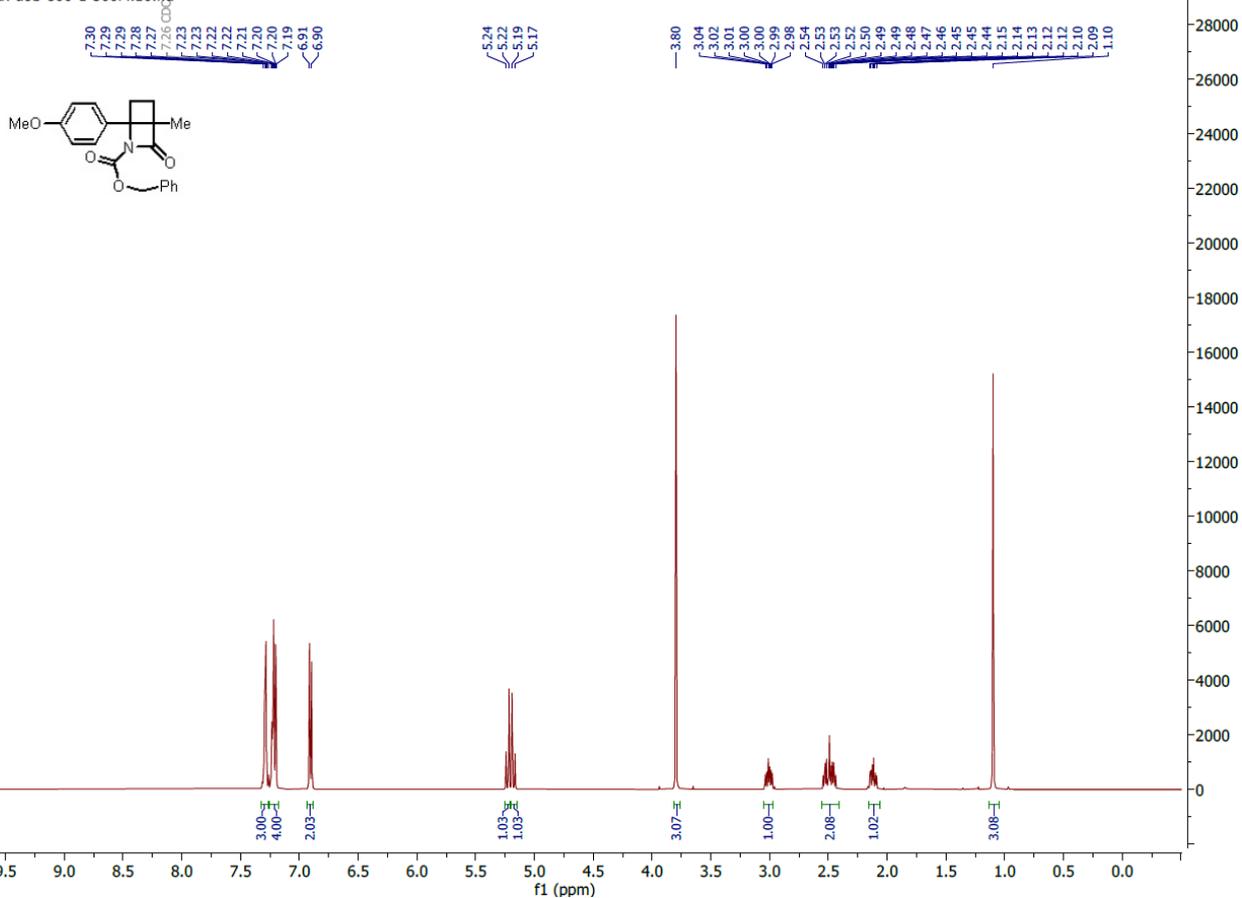


5716 zzx-uob-812-500M.11.fid

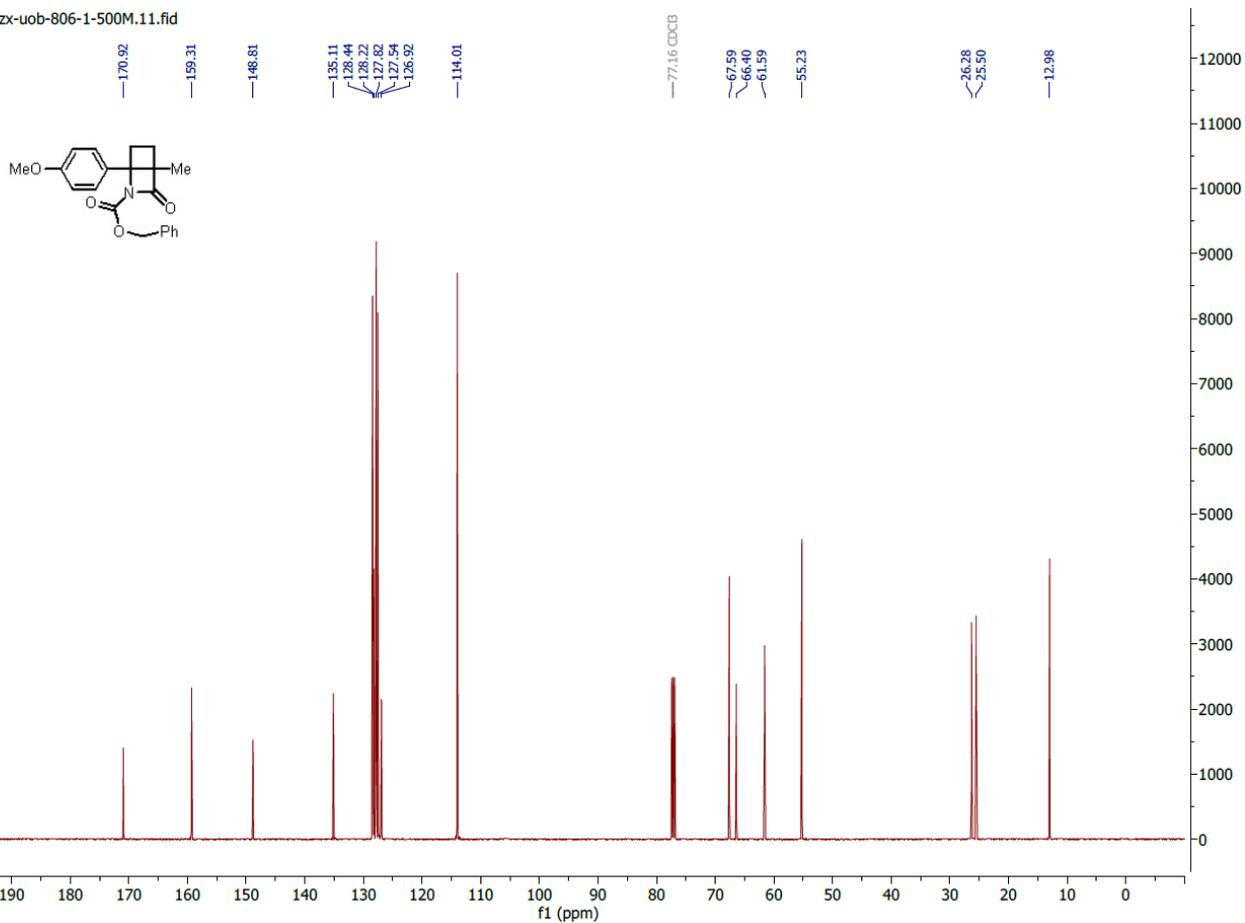


Compound 6z

5710 zzx-uob-806-1-500M.10.fid

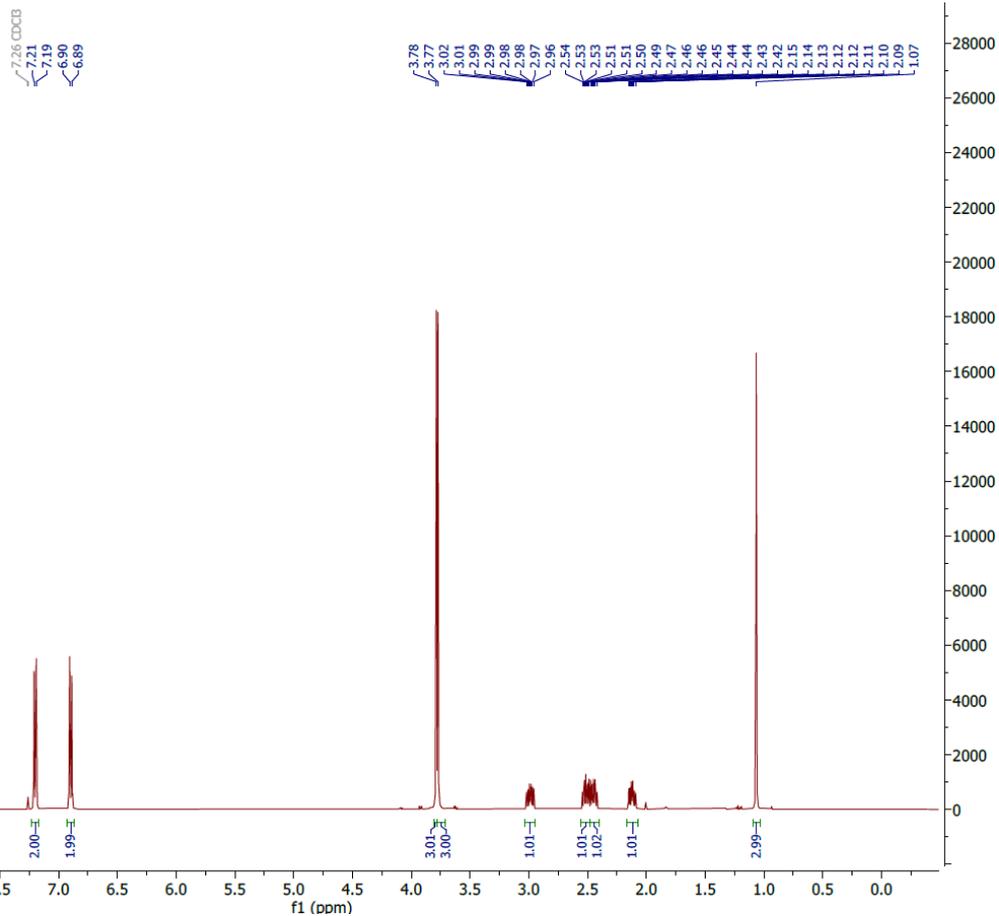
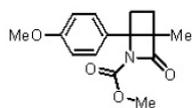


5710 zzx-uob-806-1-500M.11.fid

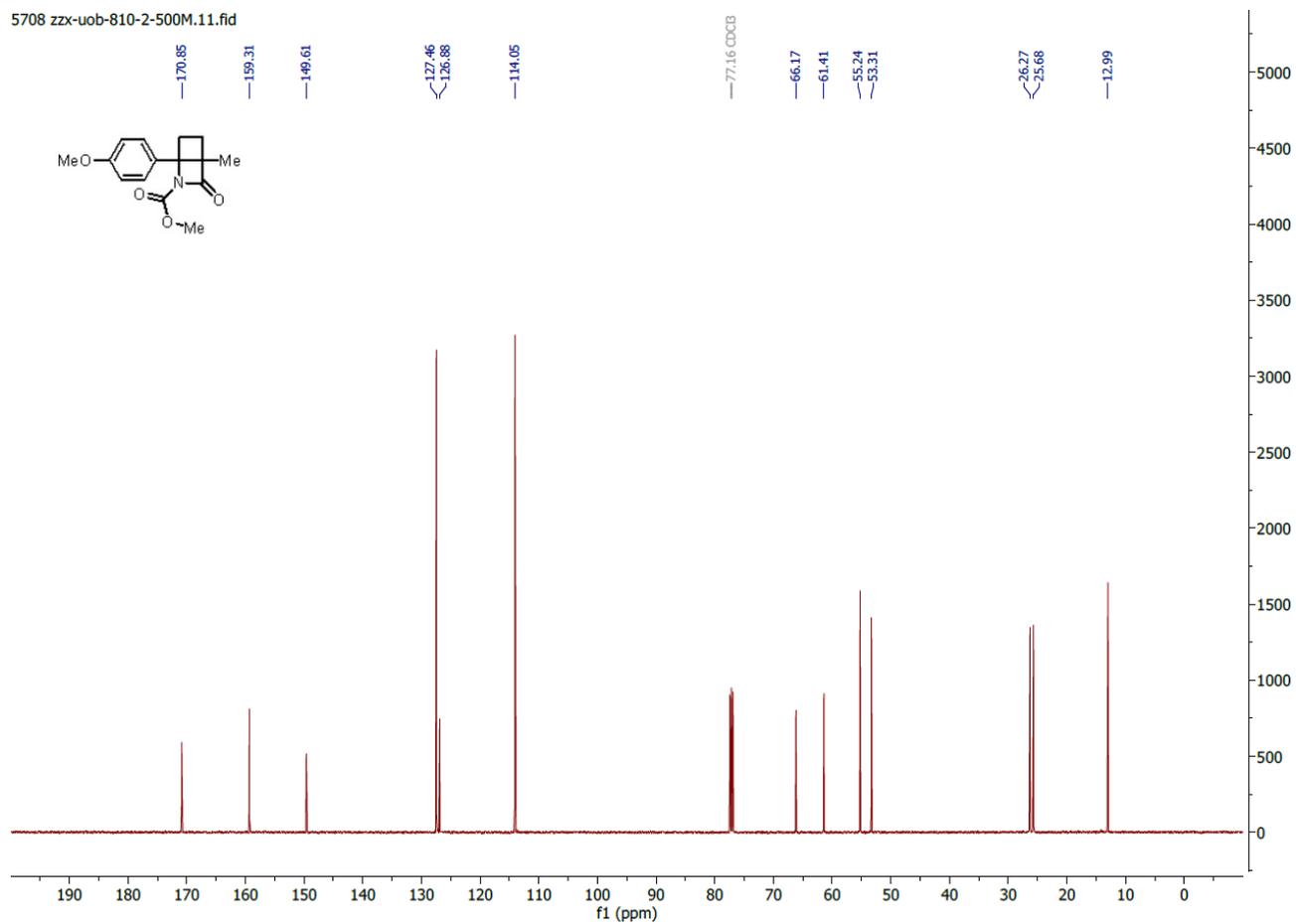
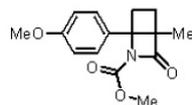


Compound 6aa

5708 zzx-uob-810-2-500M.10.fid

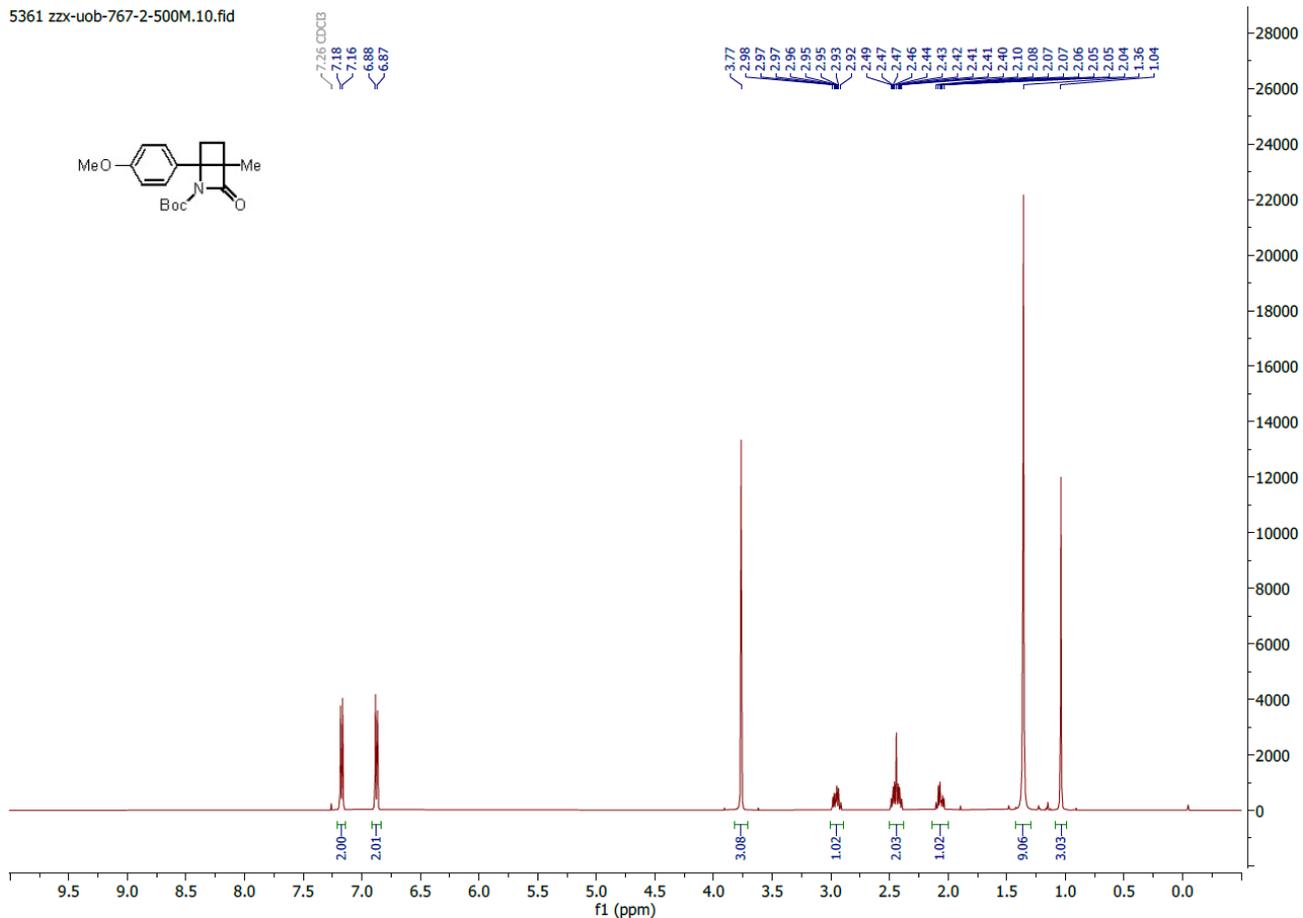


5708 zzx-uob-810-2-500M.11.fid

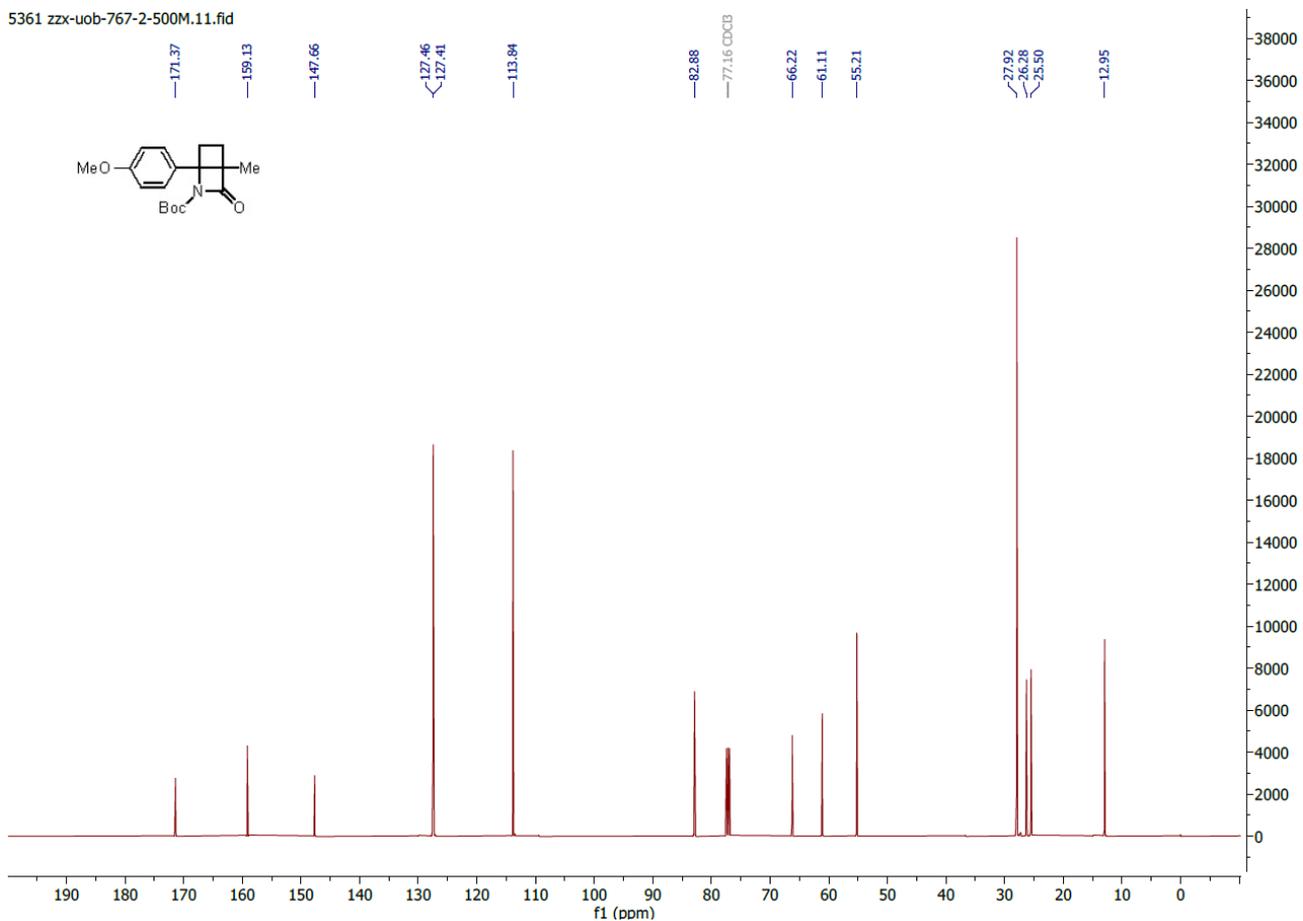


Compound 6ab

5361 zzx-uob-767-2-500M.10.fid

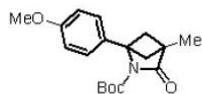


5361 zzx-uob-767-2-500M.11.fid



Compound 5ab

5593 zzx-uob-798-3-sec-500M.10.fid



7.26 CDCl₃
7.15
7.13
6.87
6.85

3.78

2.69
2.69
2.68
2.68
2.34
2.34
2.33
2.33

1.31
1.14

35000

30000

25000

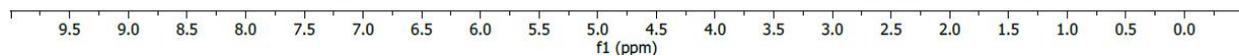
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15000

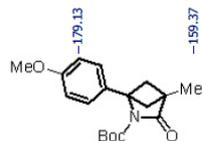
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5000

0



5593 zzx-uob-798-3-sec-500M.11.fid



179.13

159.37

149.65

129.48

127.86

113.63

81.71

77.16 CDCl₃

68.35

55.32

54.88

49.57

27.70

13.22

25000

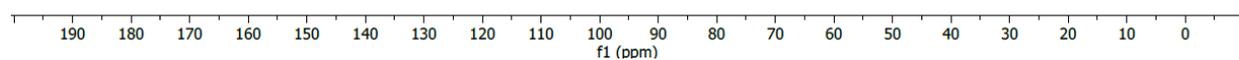
20000

15000

10000

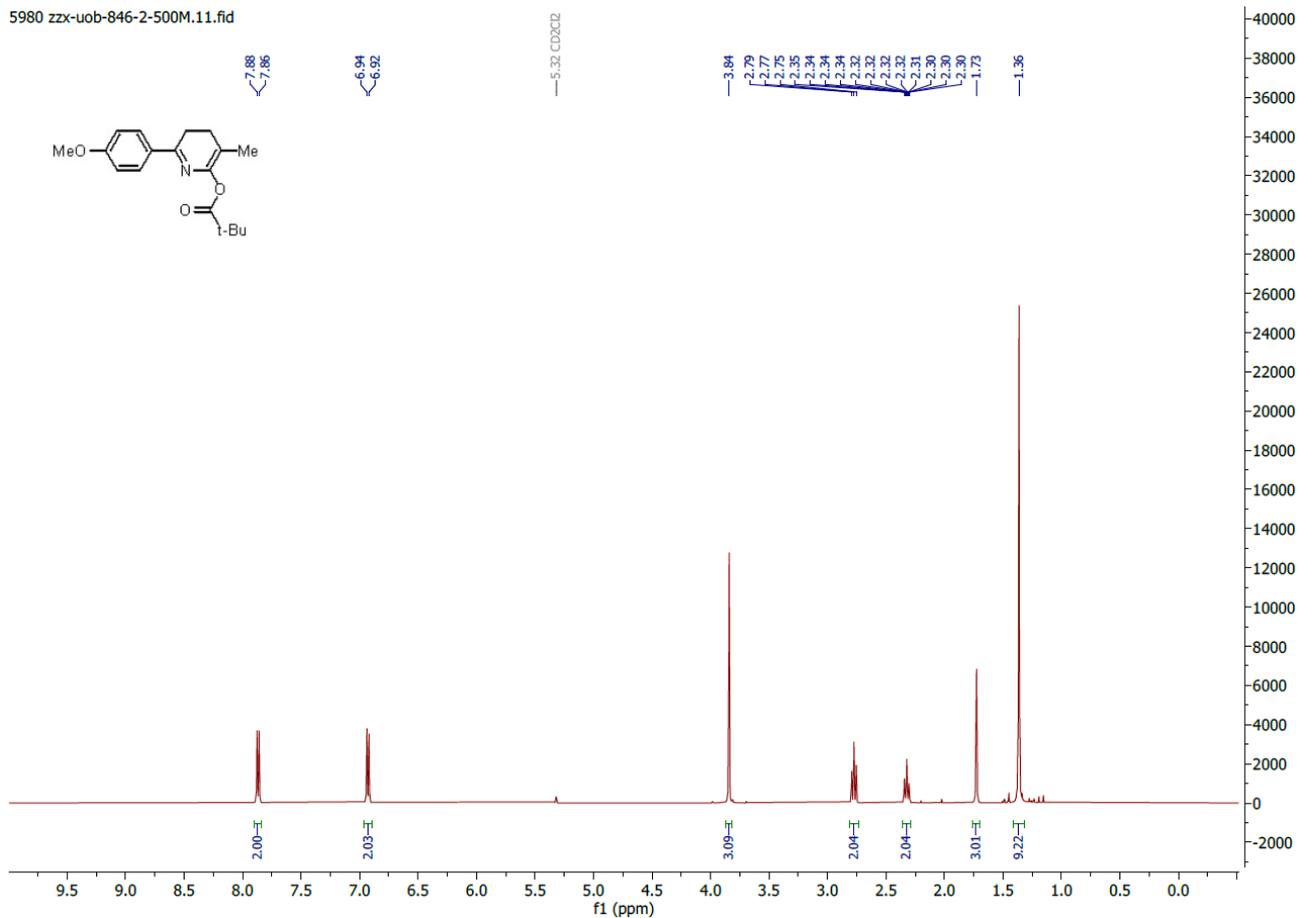
5000

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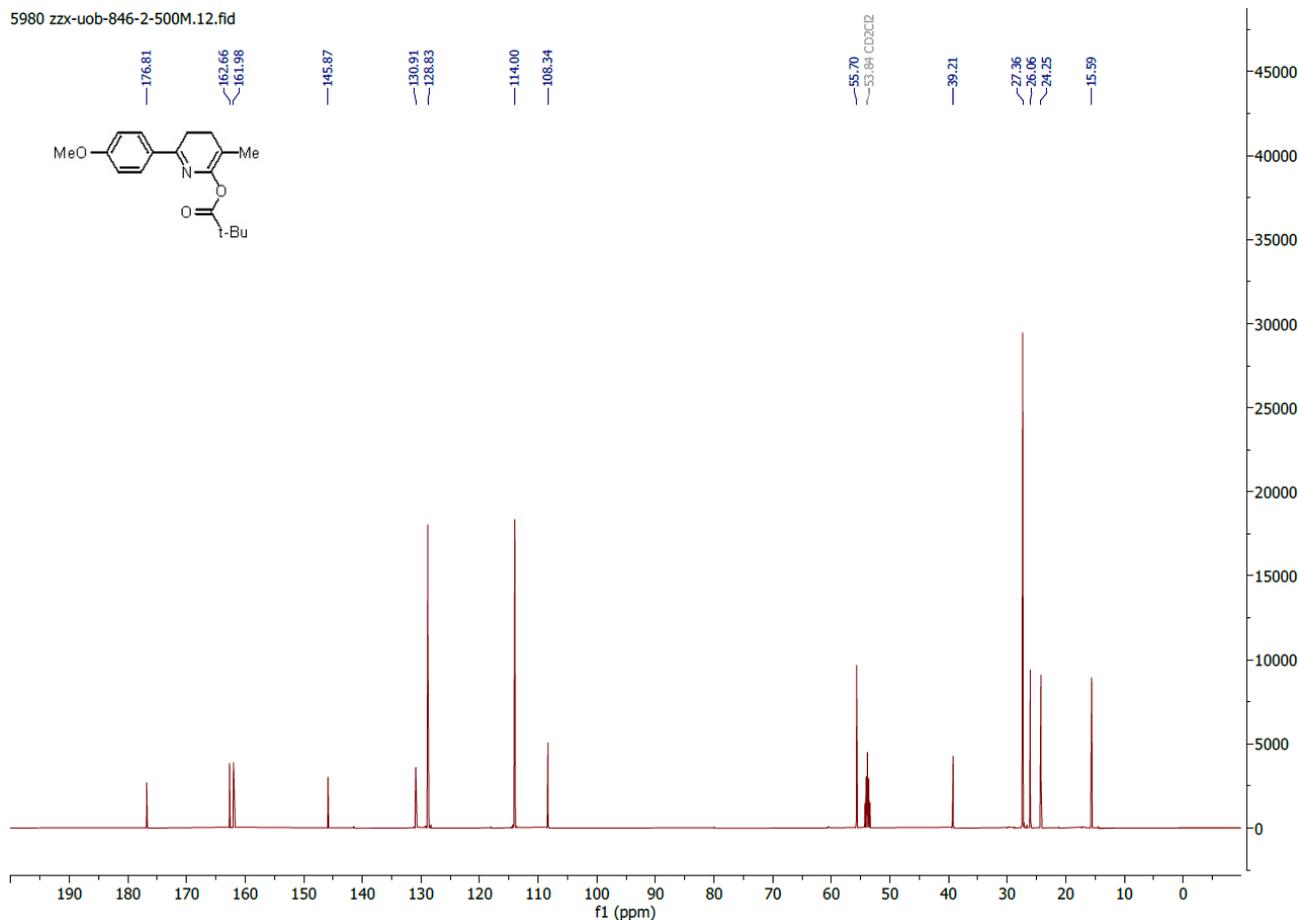


Compound 9

5980 zzx-uob-846-2-500M.11.fid

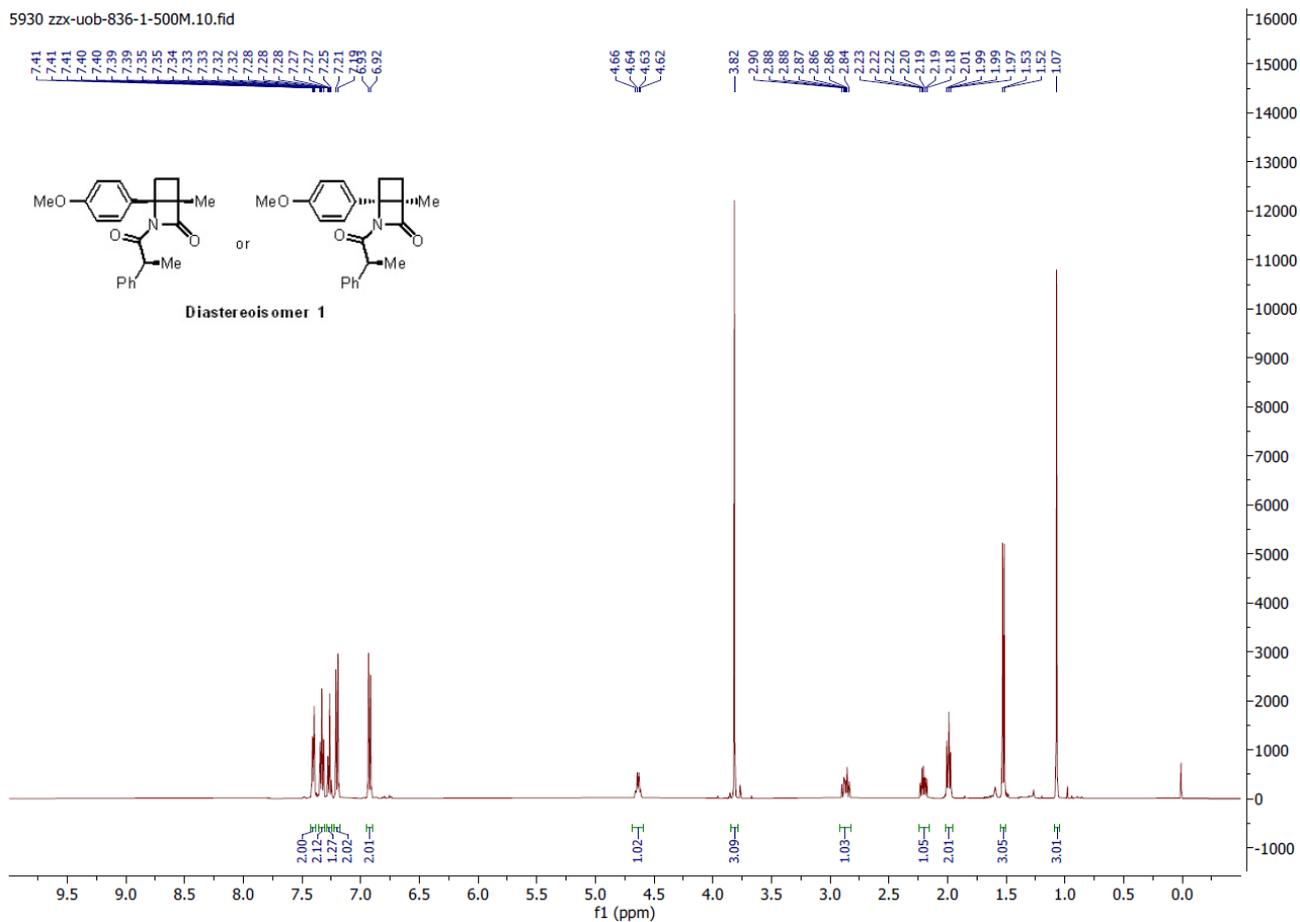


5980 zzx-uob-846-2-500M.12.fid

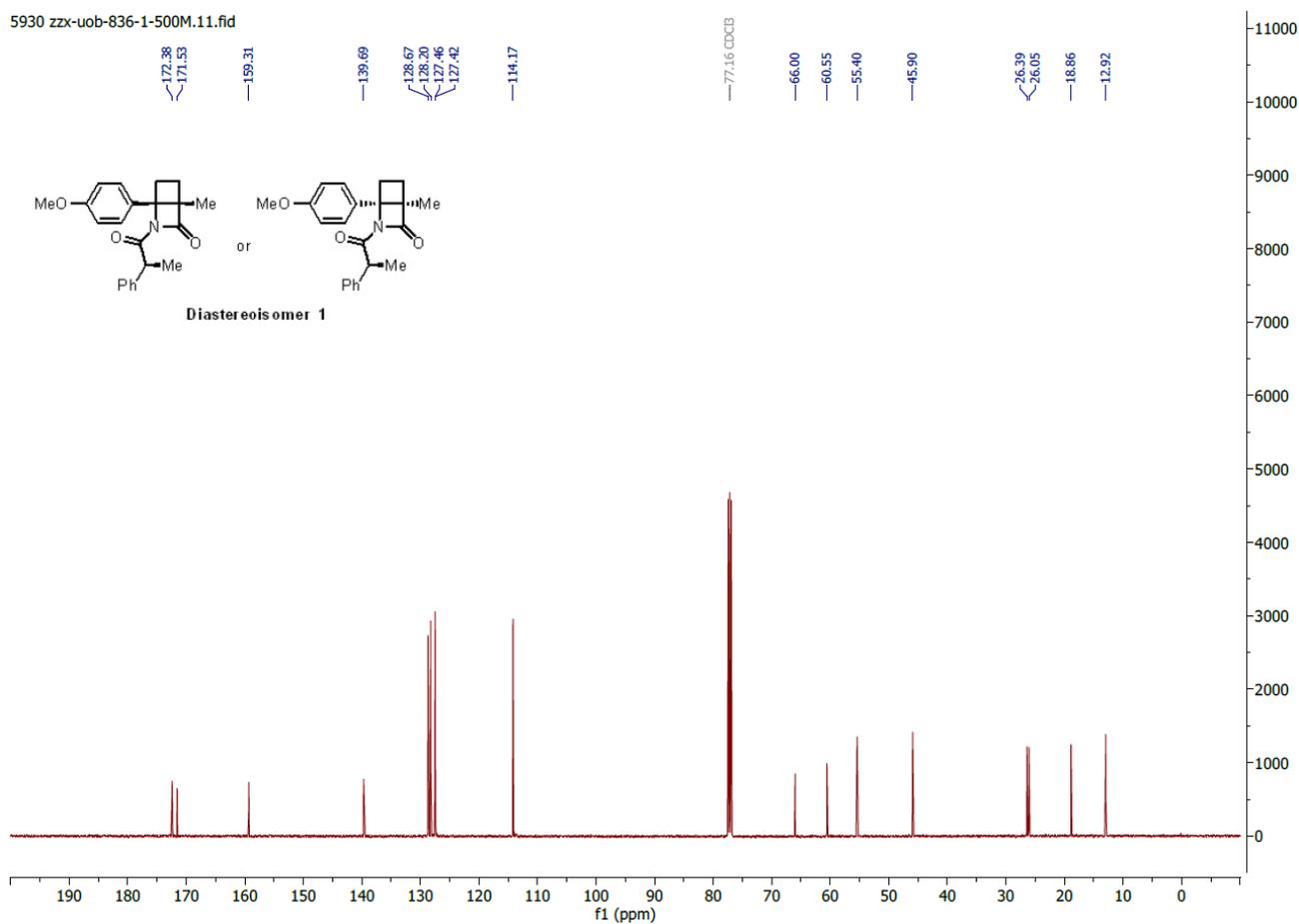


Compound 6ad and 6ad'

5930 zzx-uob-836-1-500M.10.fid

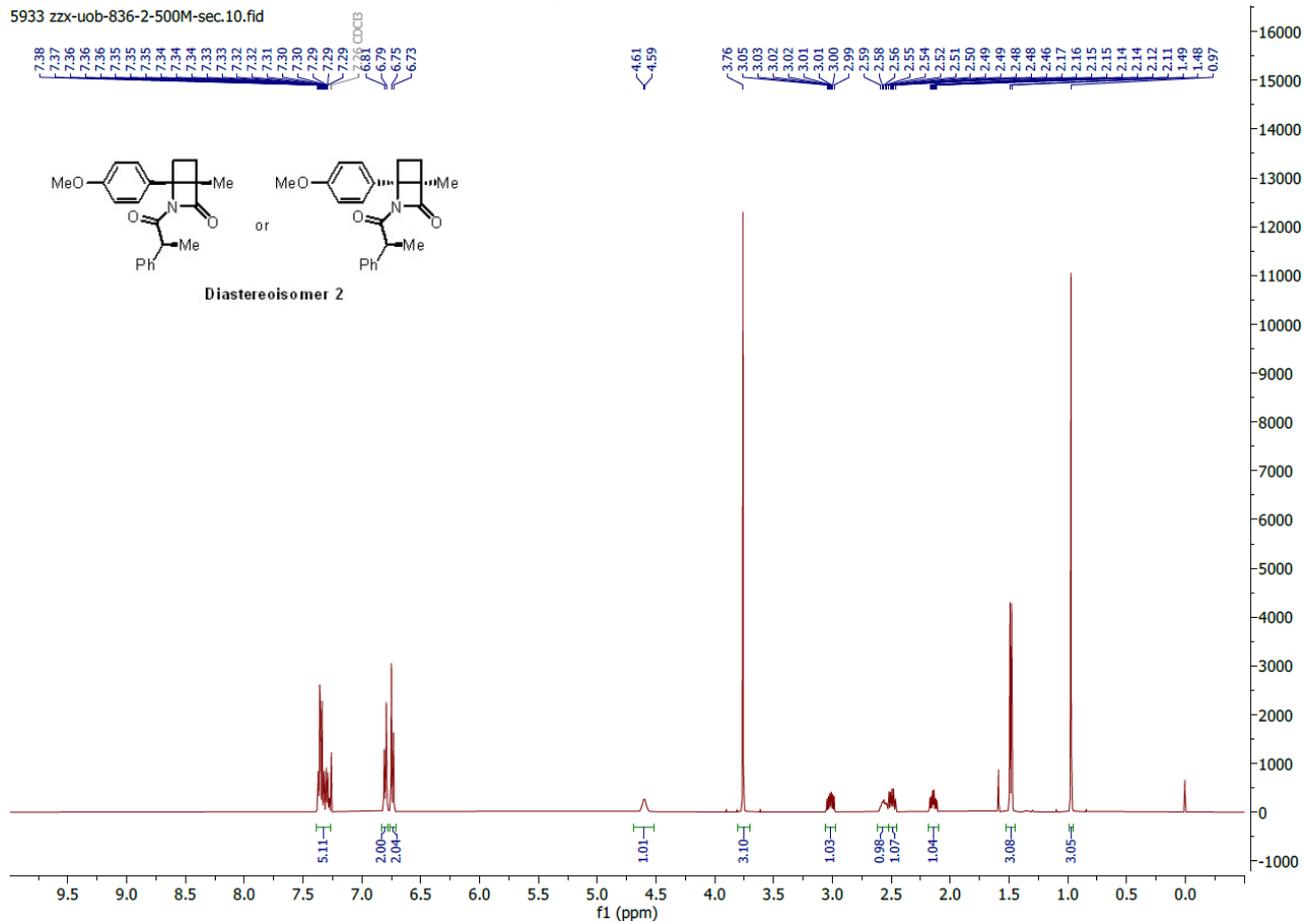


5930 zzx-uob-836-1-500M.11.fid

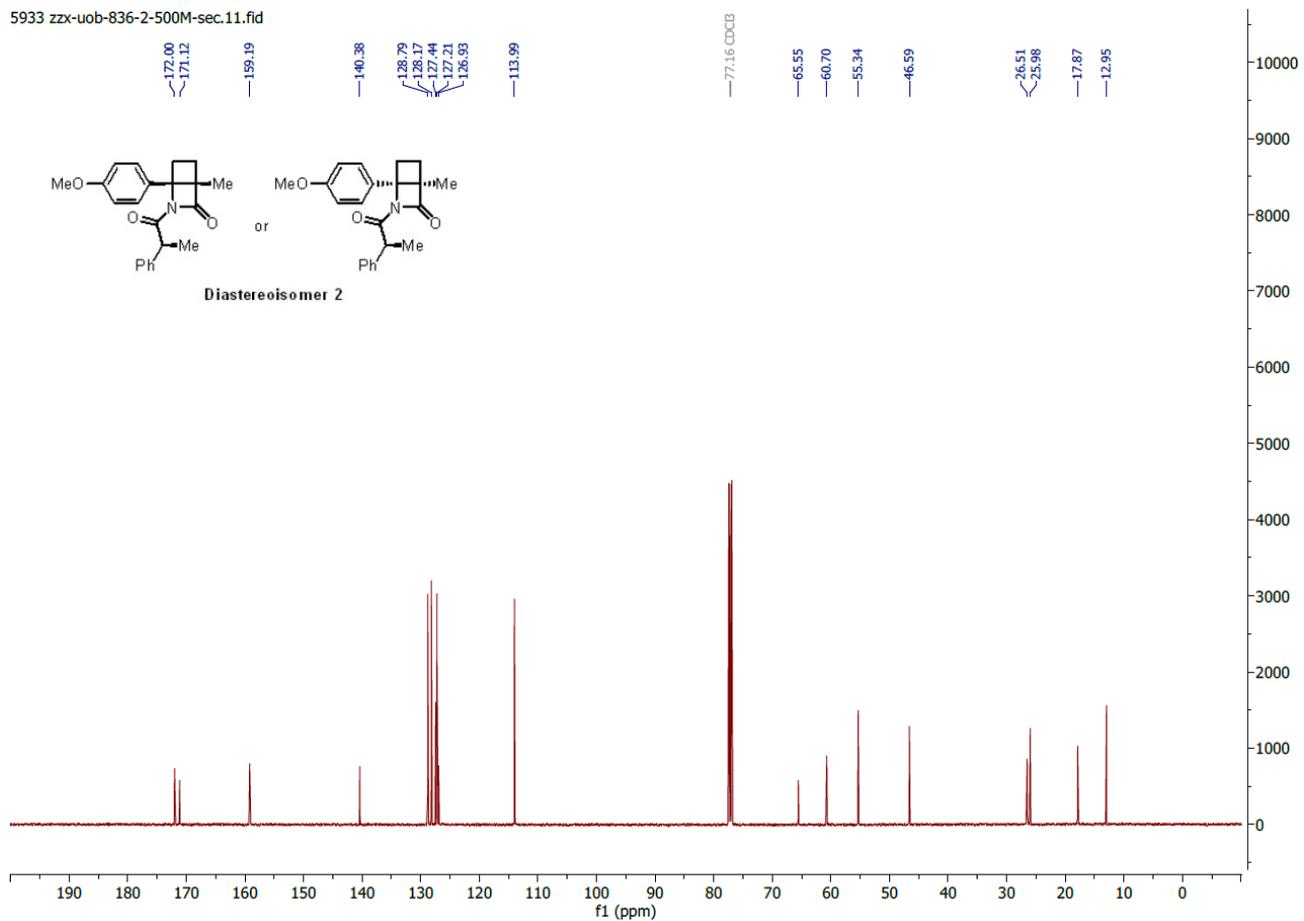


Compound 6ad and 6ad'

5933 zzx-uob-836-2-500M-sec.10.fid

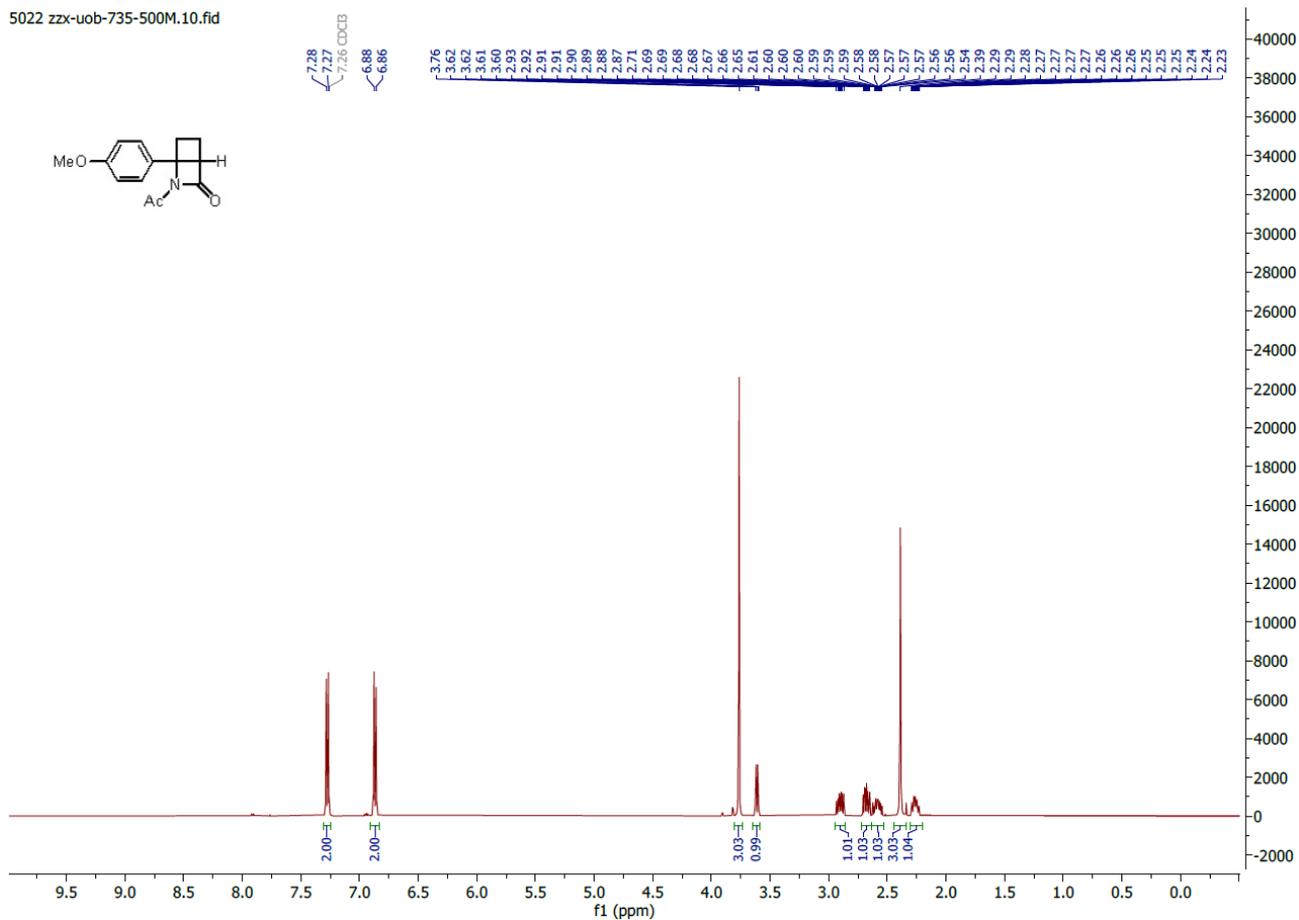
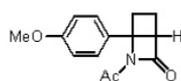


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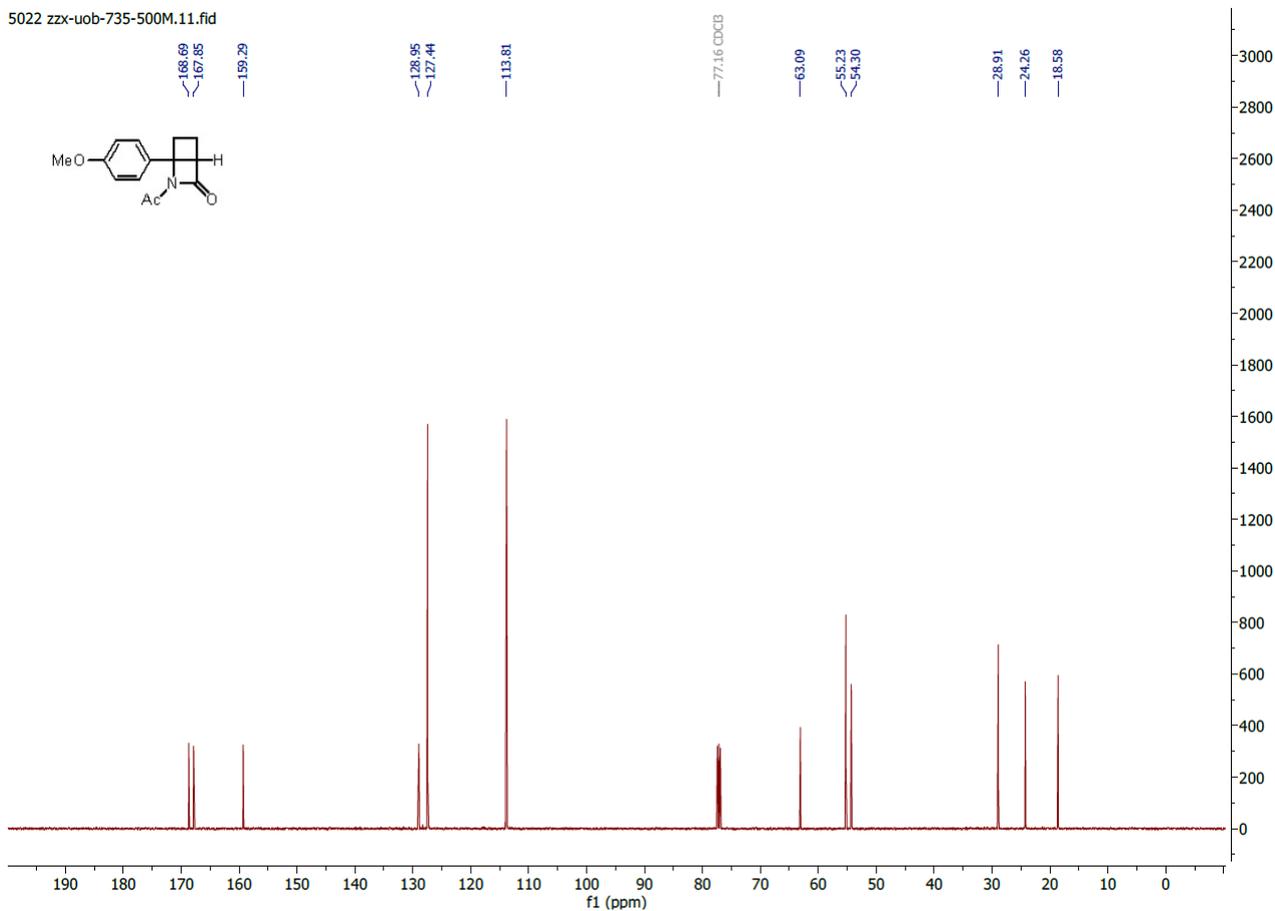
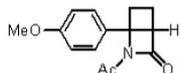


Compound 6ae

5022 zzx-uob-735-500M.10.fid

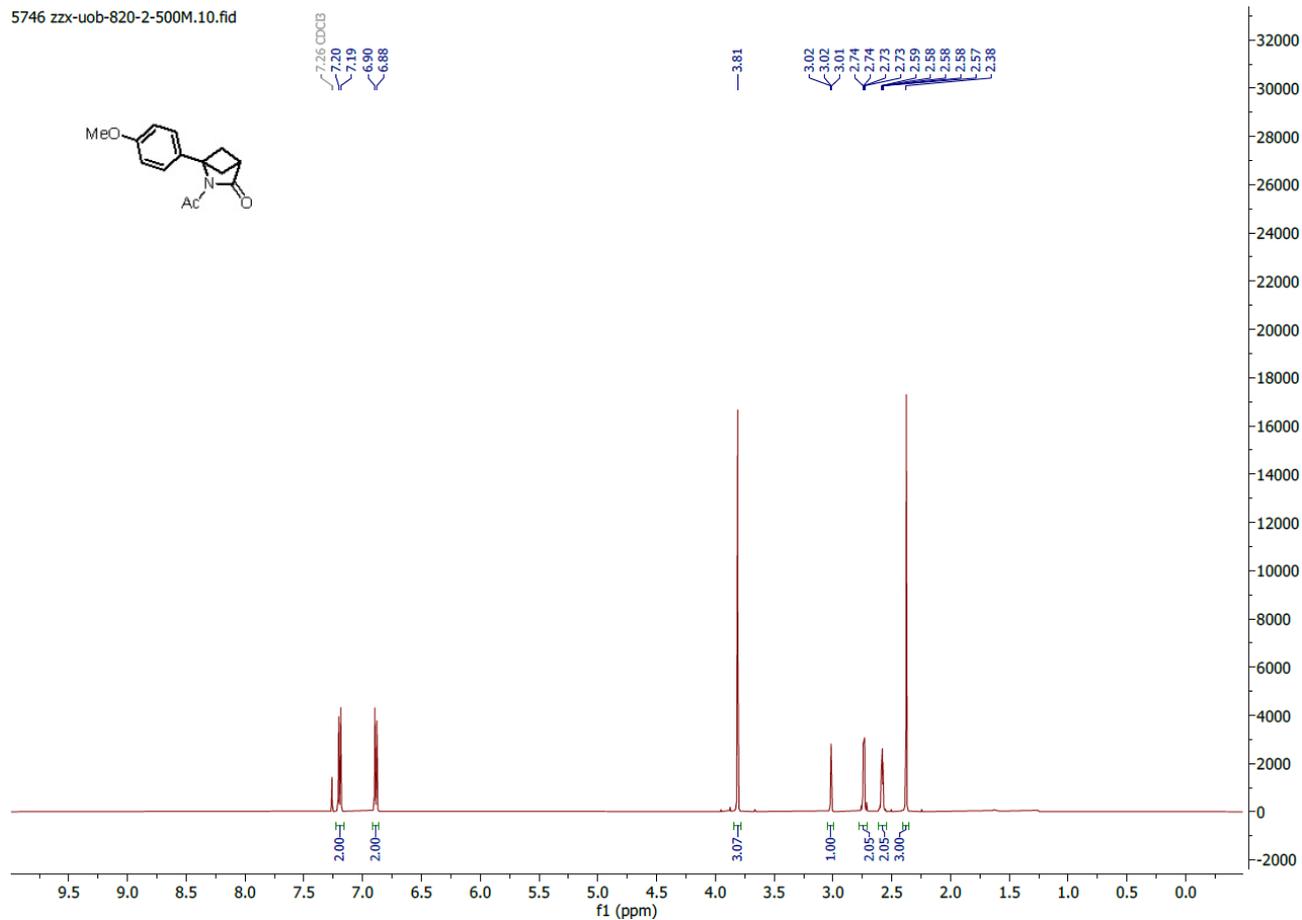


5022 zzx-uob-735-500M.11.fid

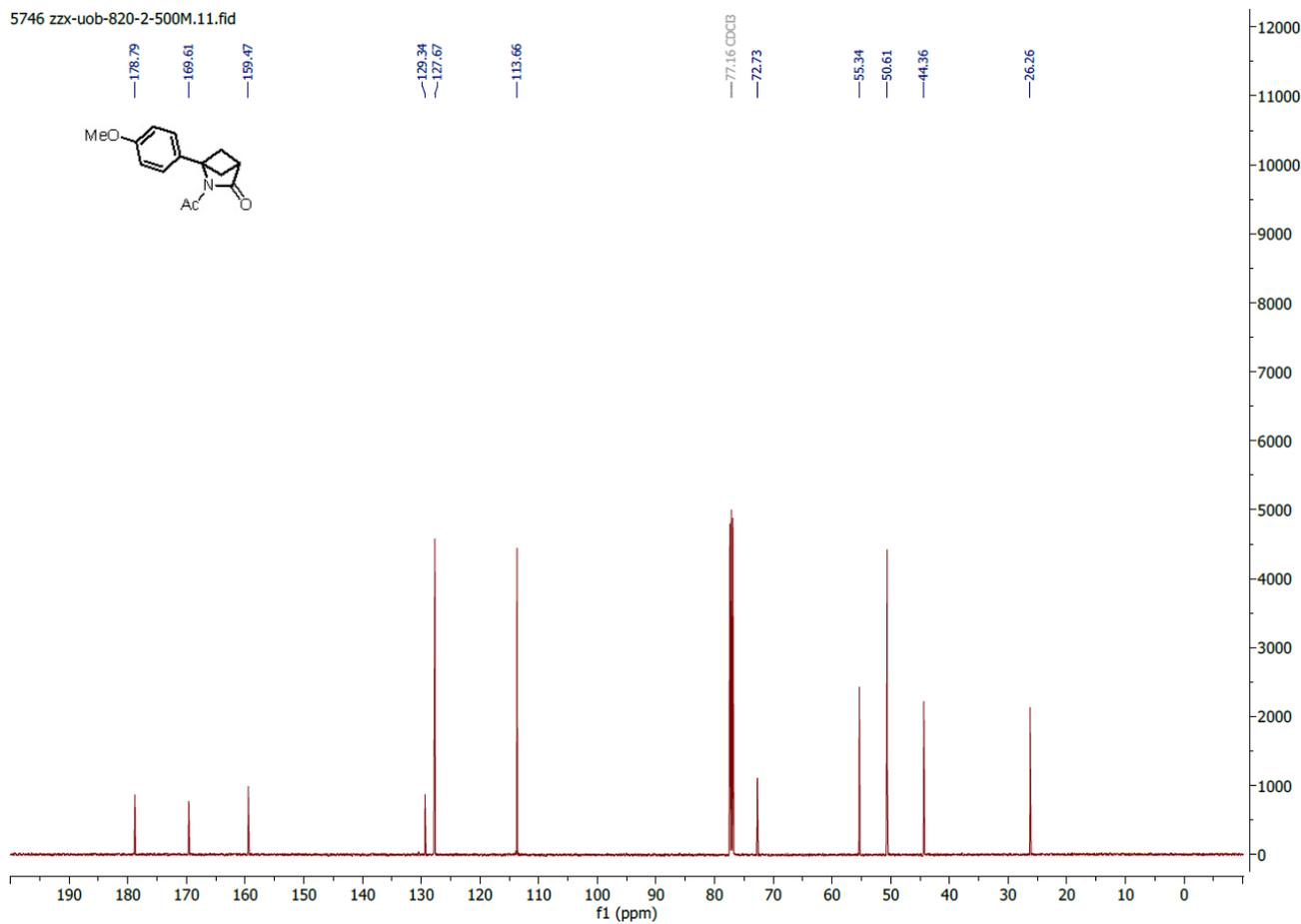


Compound 5ae

5746 zzx-uob-820-2-500M.10.fid

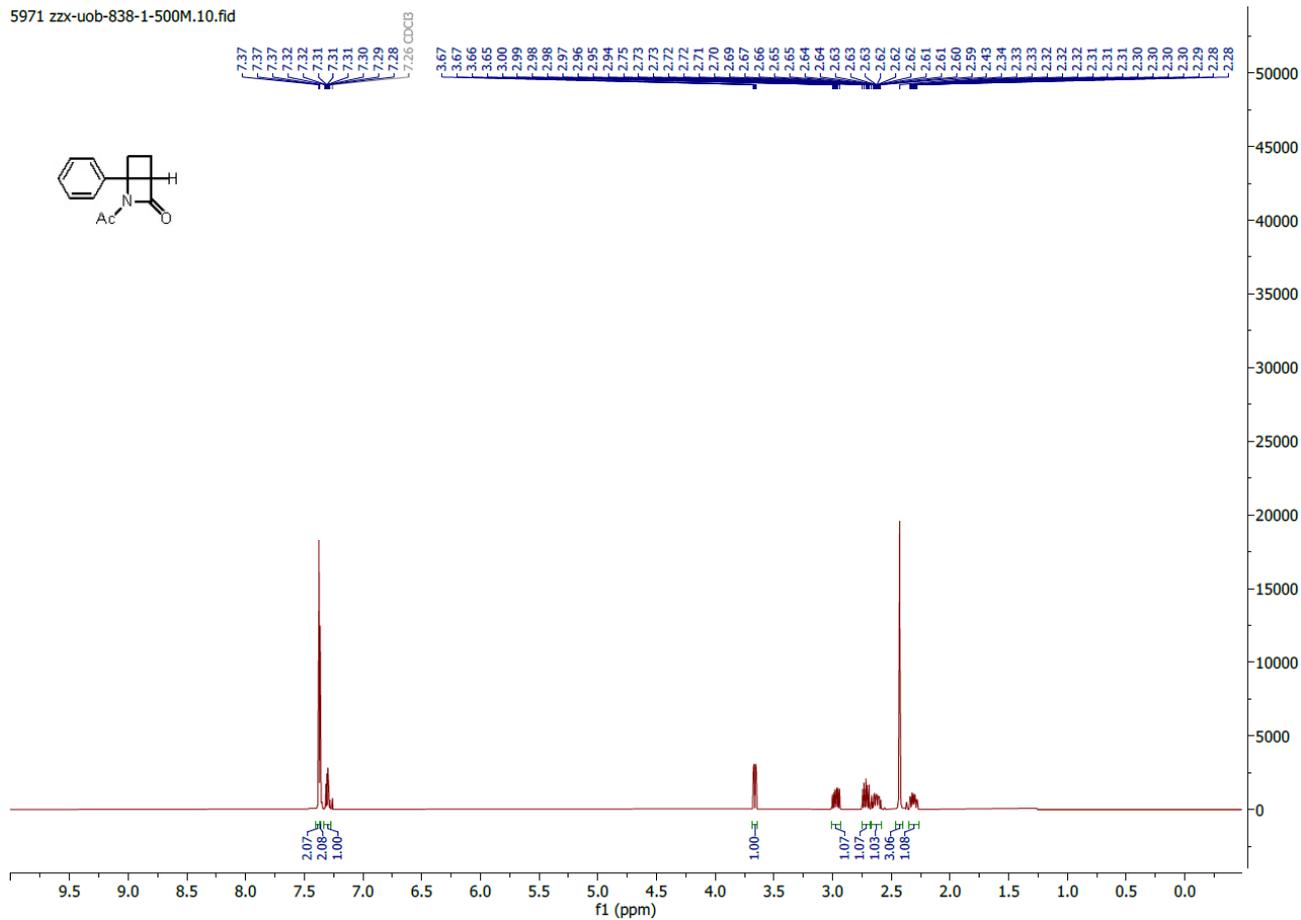


5746 zzx-uob-820-2-500M.11.fid

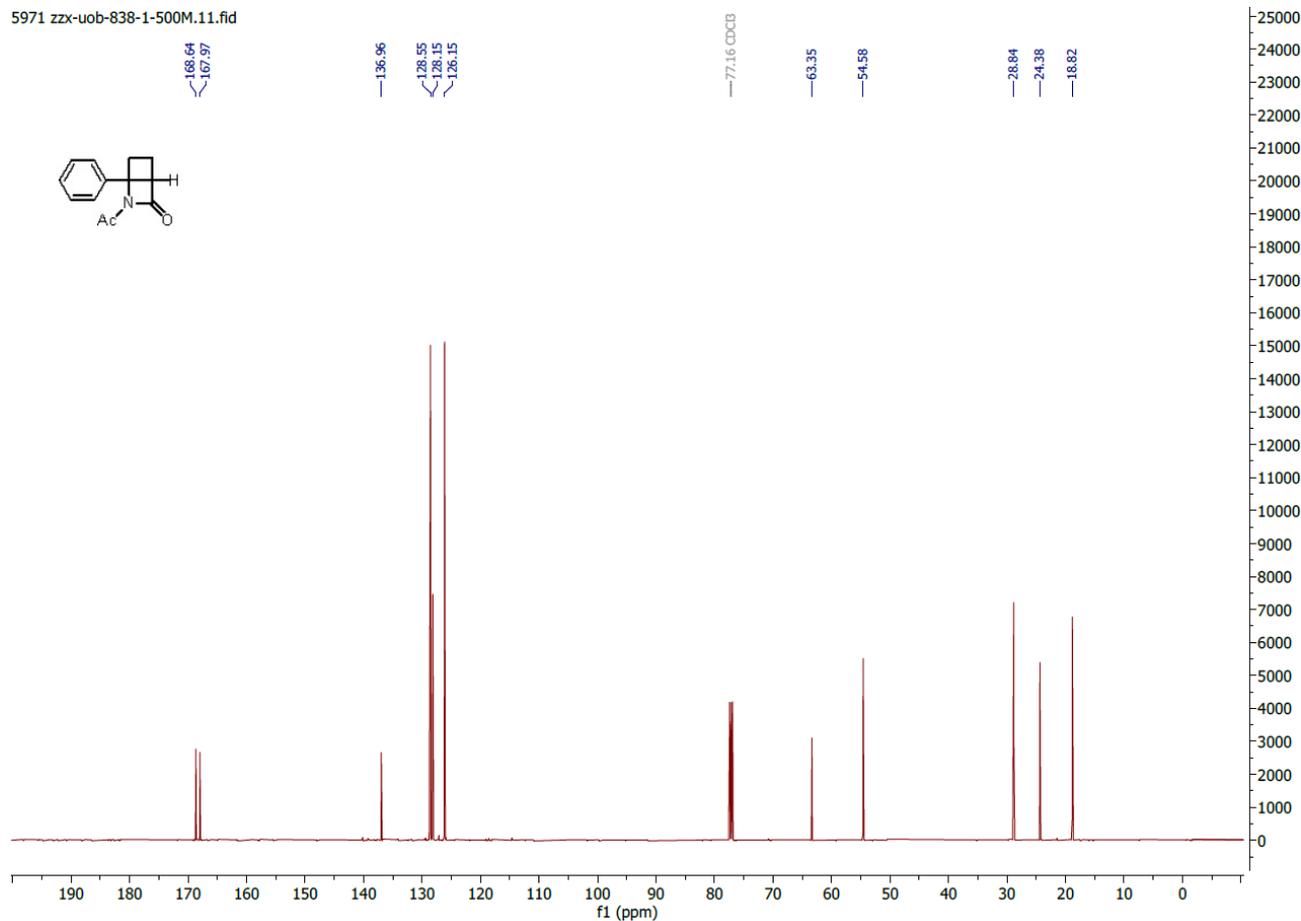


Compound 6af

5971 zzx-uob-838-1-500M.10.fid

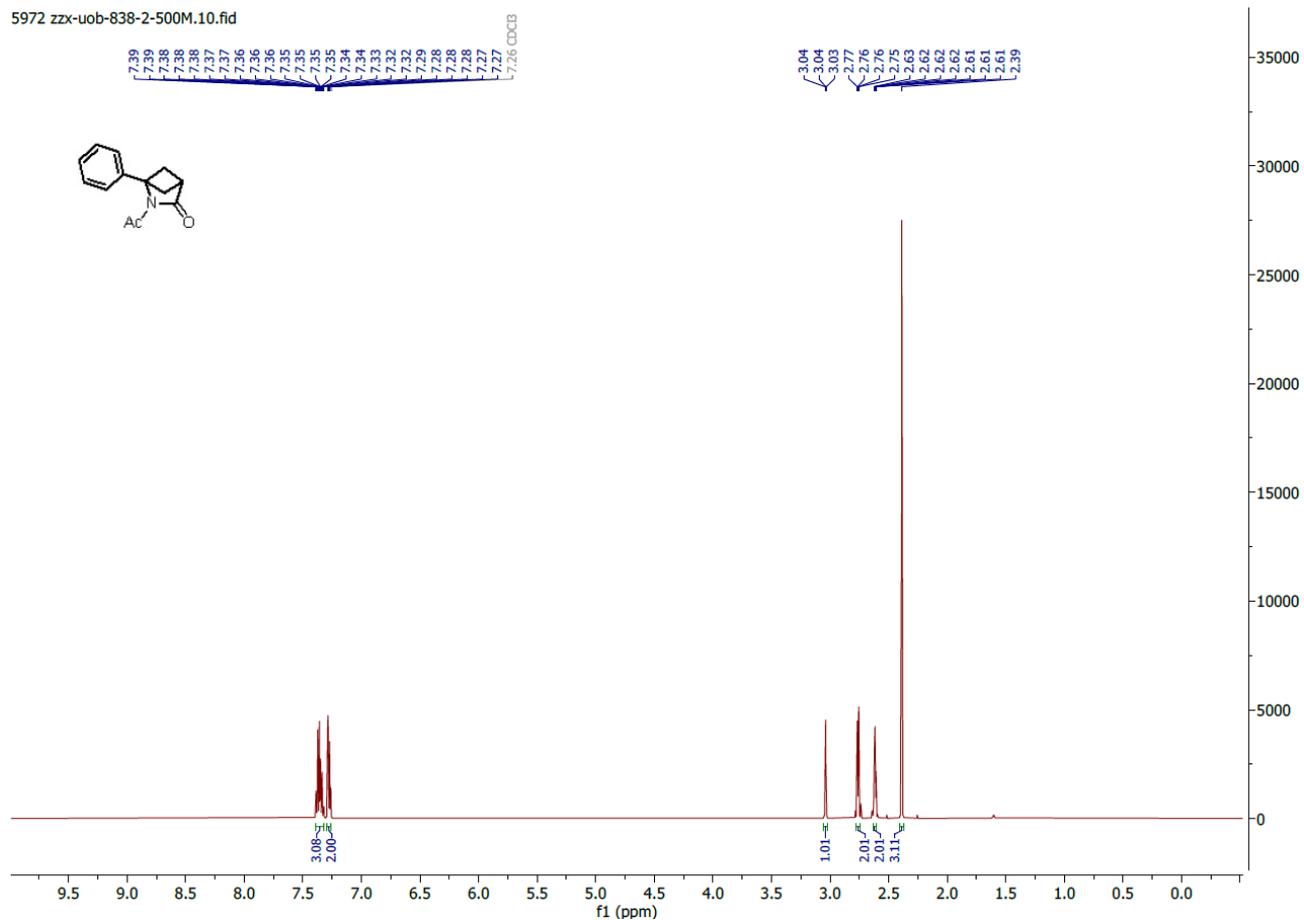


5971 zzx-uob-838-1-500M.11.fid

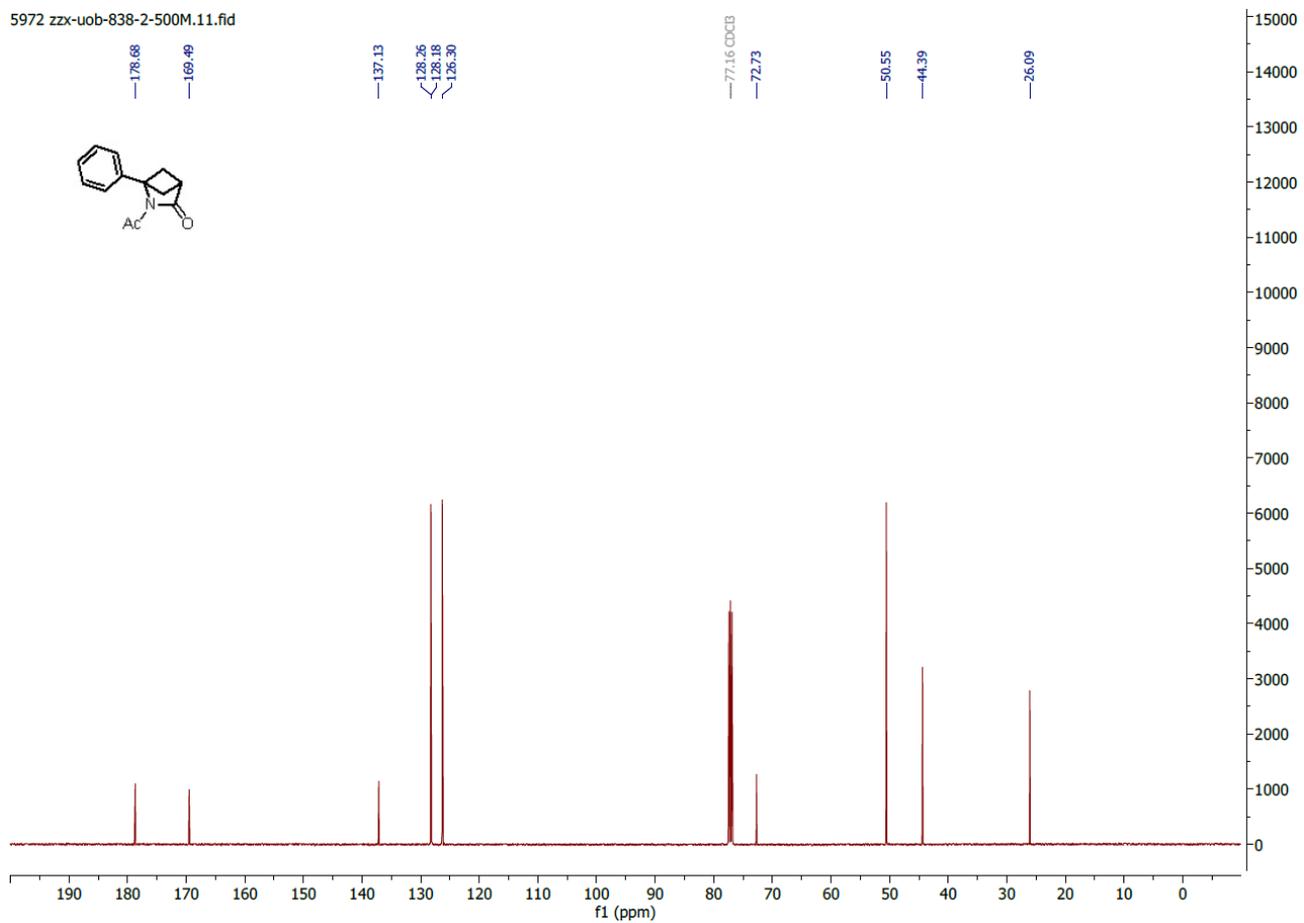


Compound 5af

5972 zzx-uob-838-2-500M.10.fid

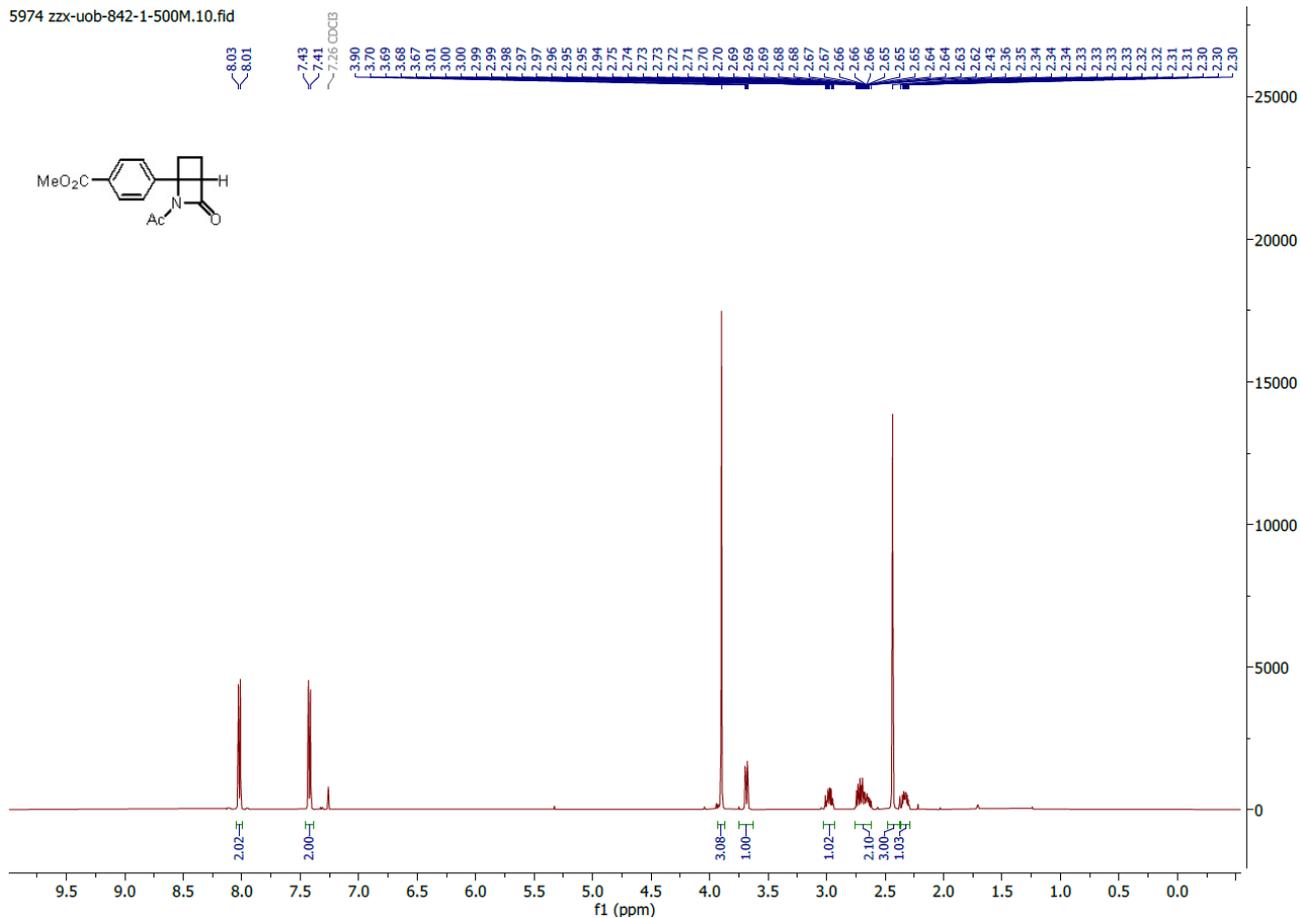


5972 zzx-uob-838-2-500M.11.fid

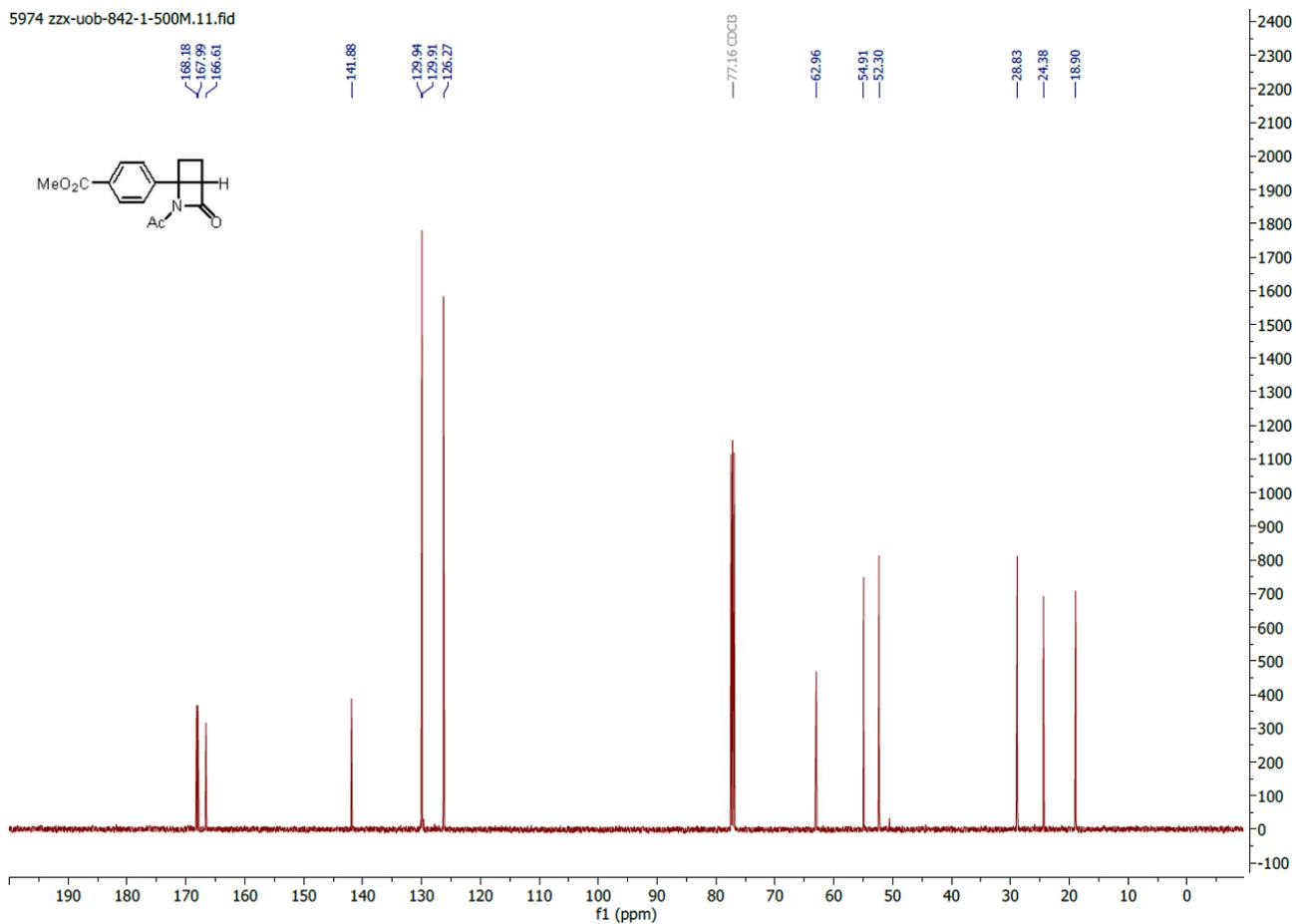


Compound 6ag

5974 zzx-uob-842-1-500M.10.fid

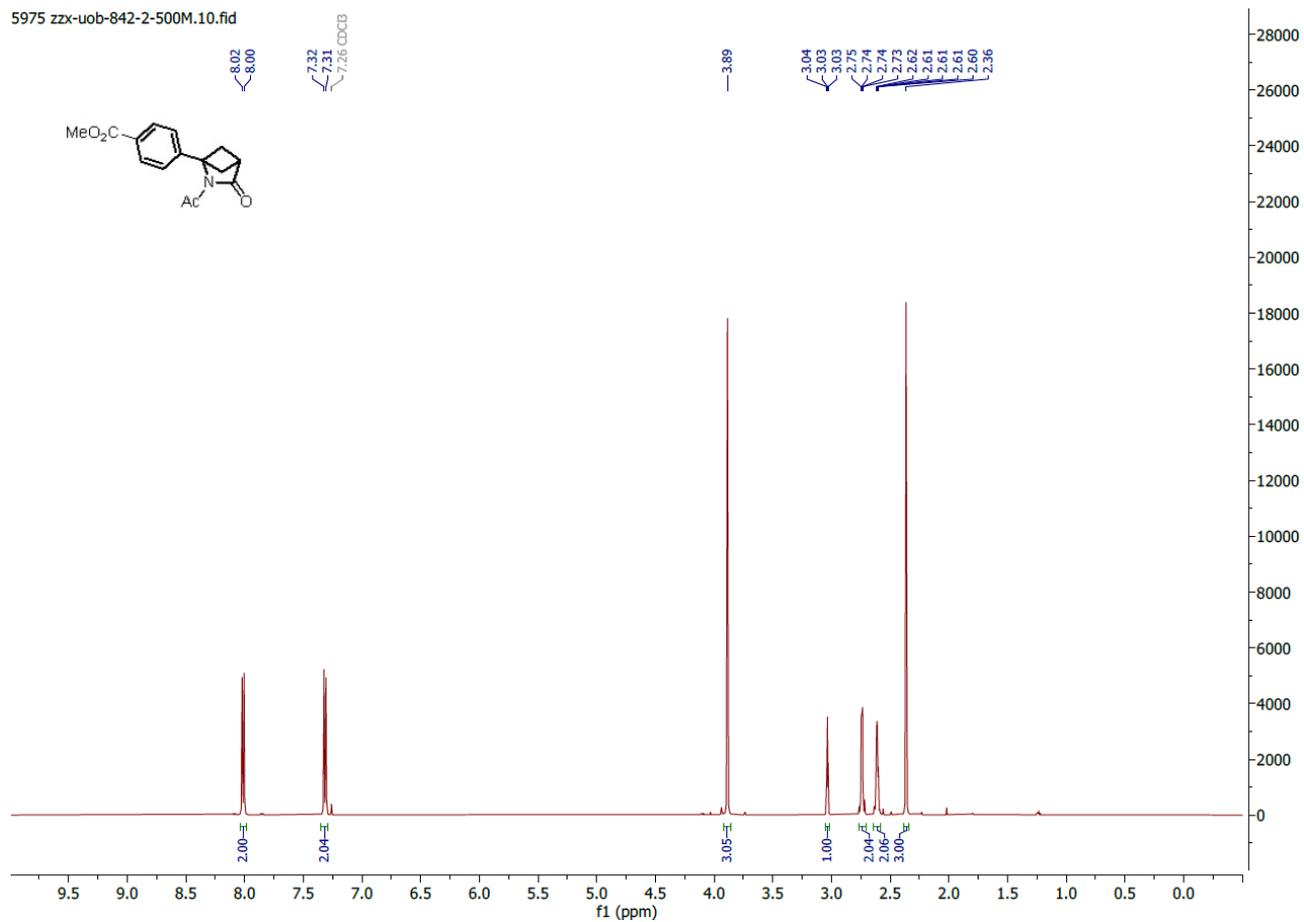


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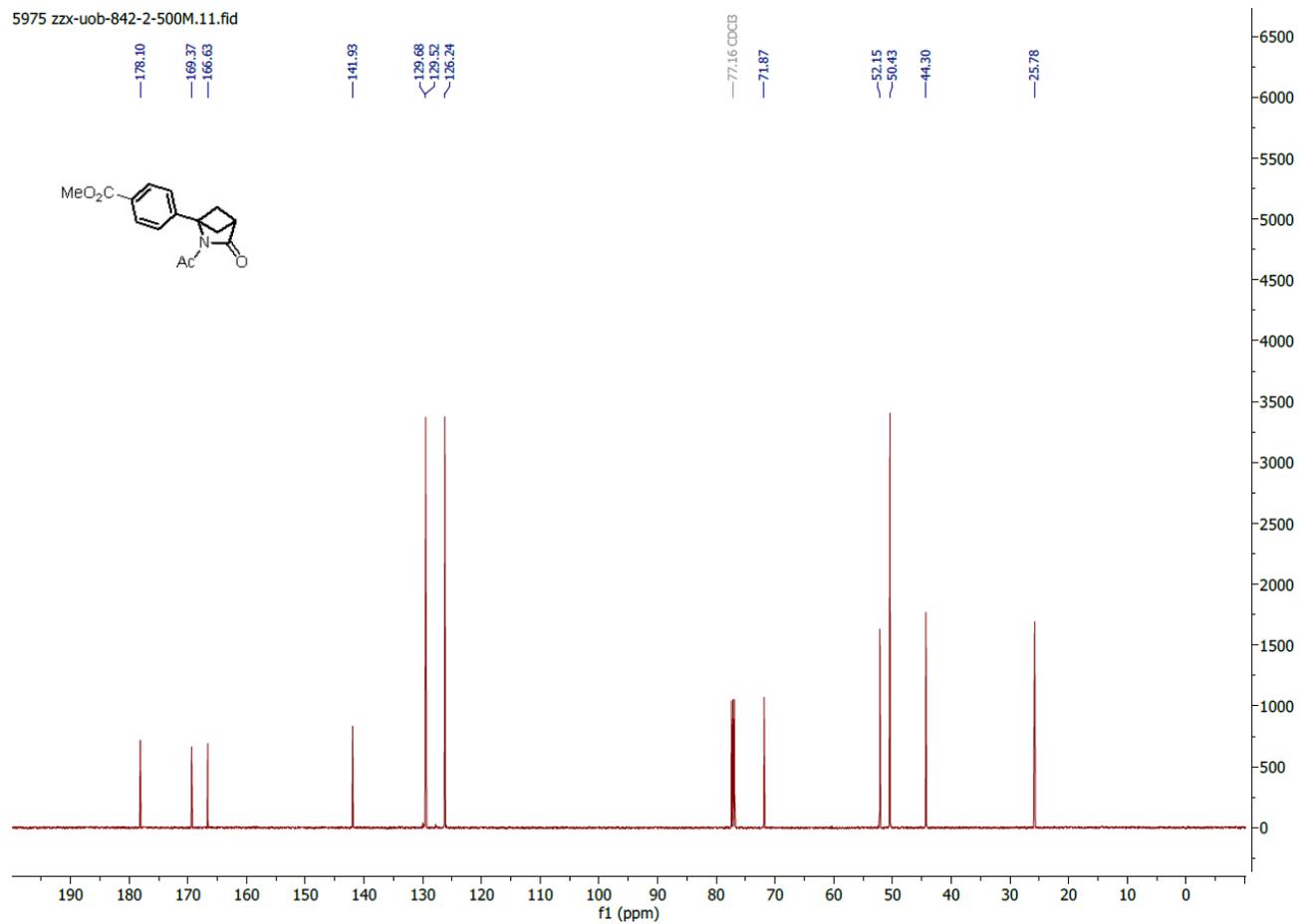


Compound 5ag

5975 zzx-uob-842-2-500M.10.fid

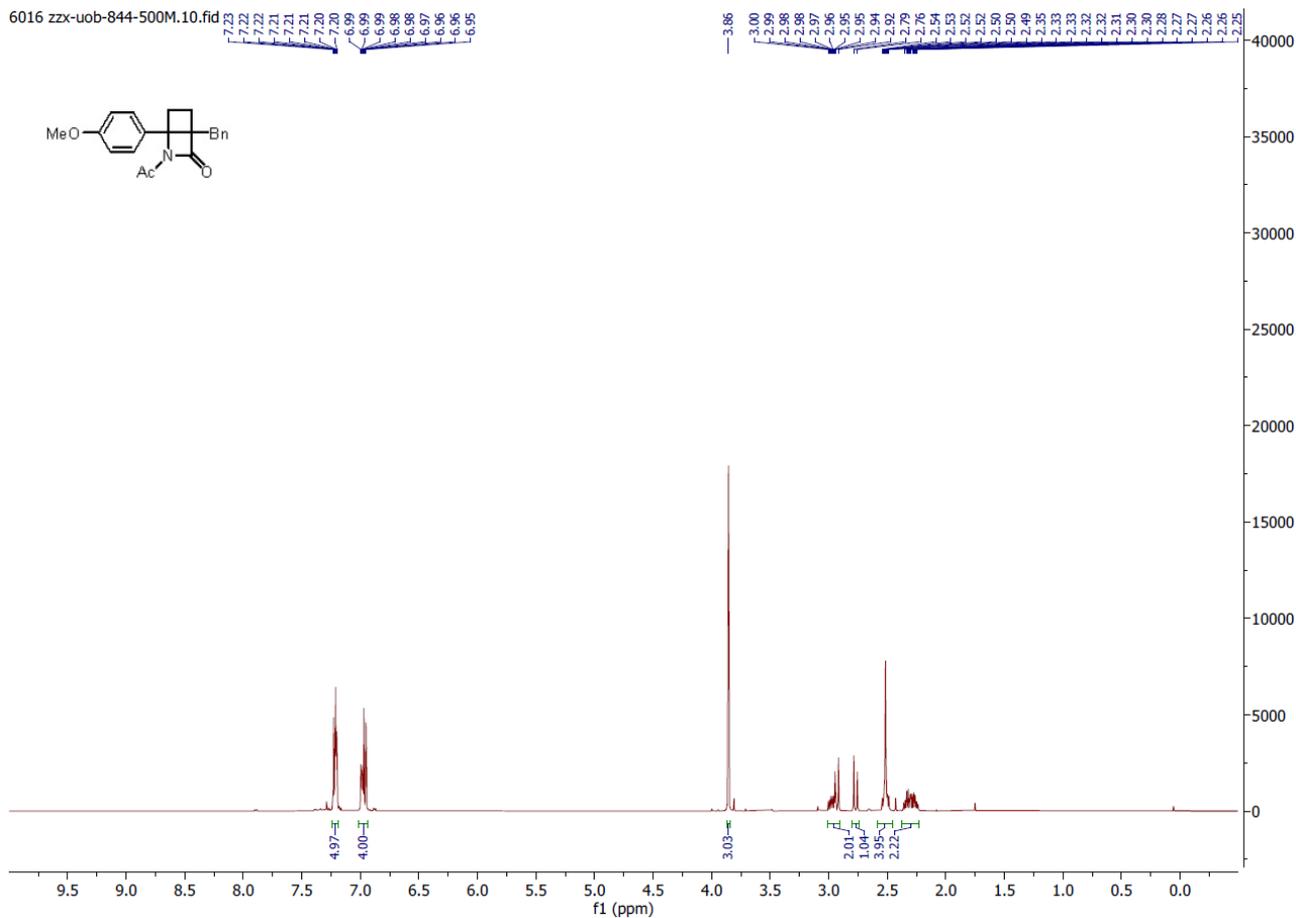
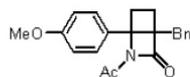


5975 zzx-uob-842-2-500M.11.fid

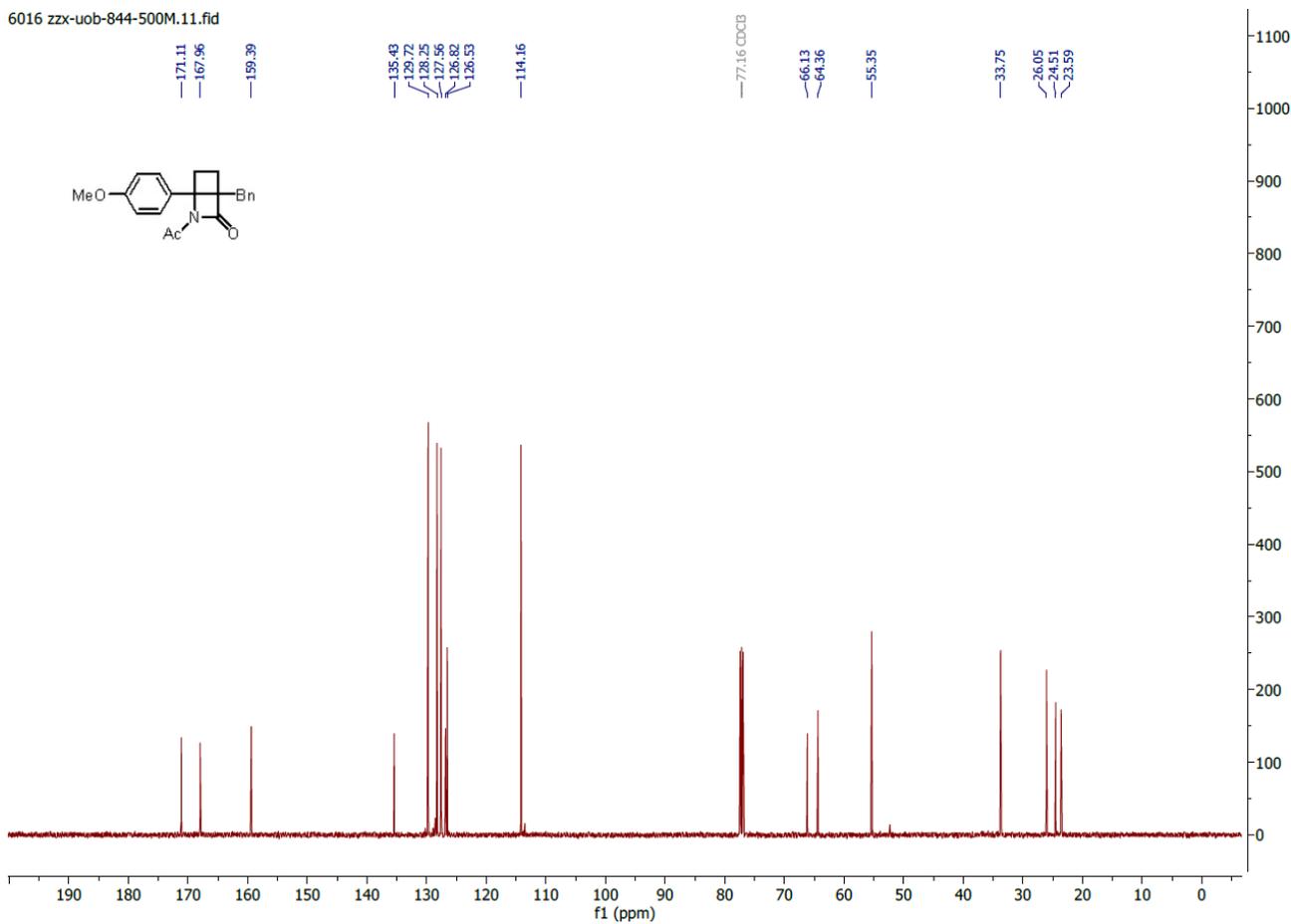
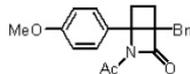


Compound 6ah

6016 zzx-uob-844-500M.10.fid

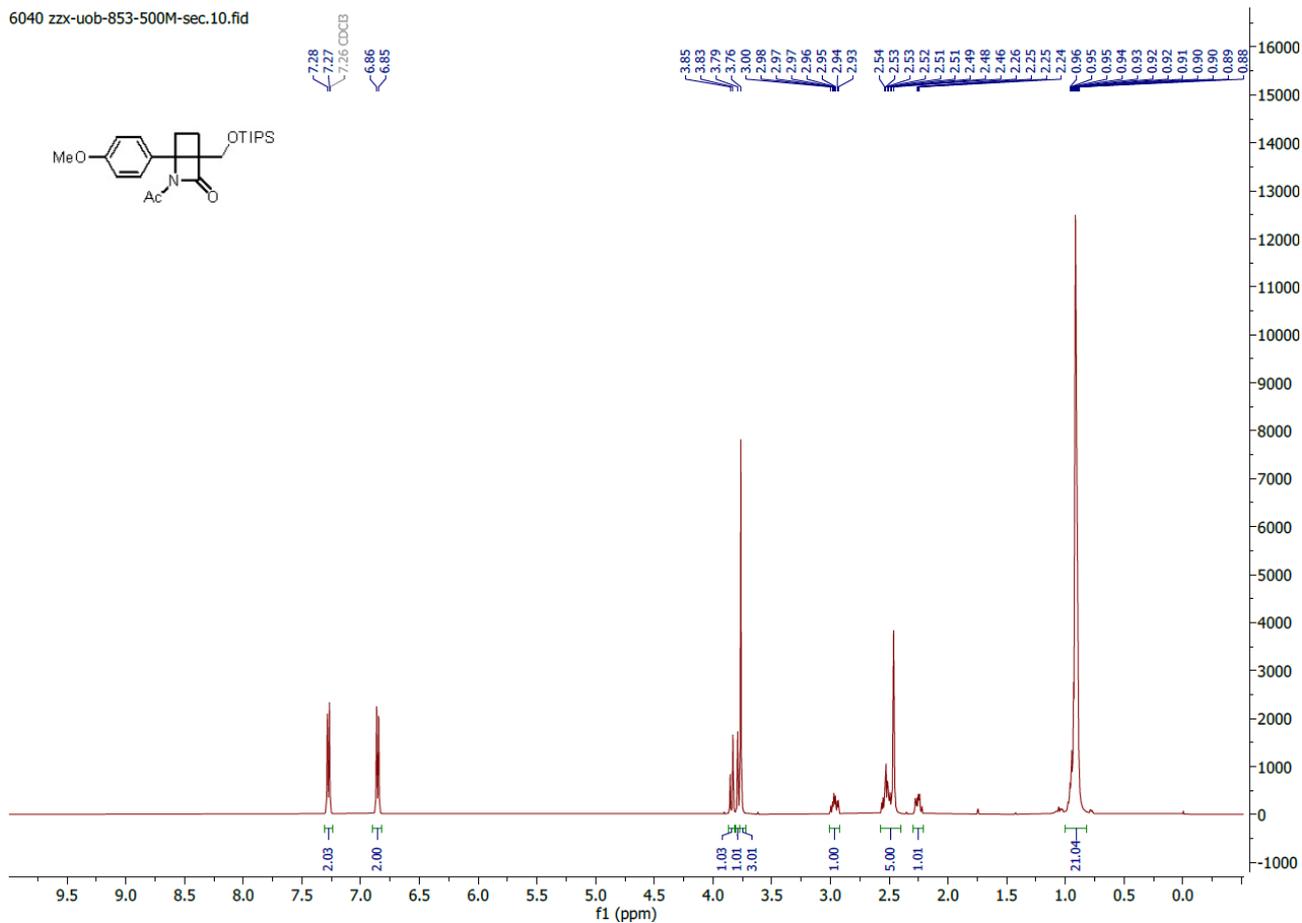
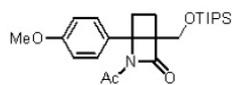


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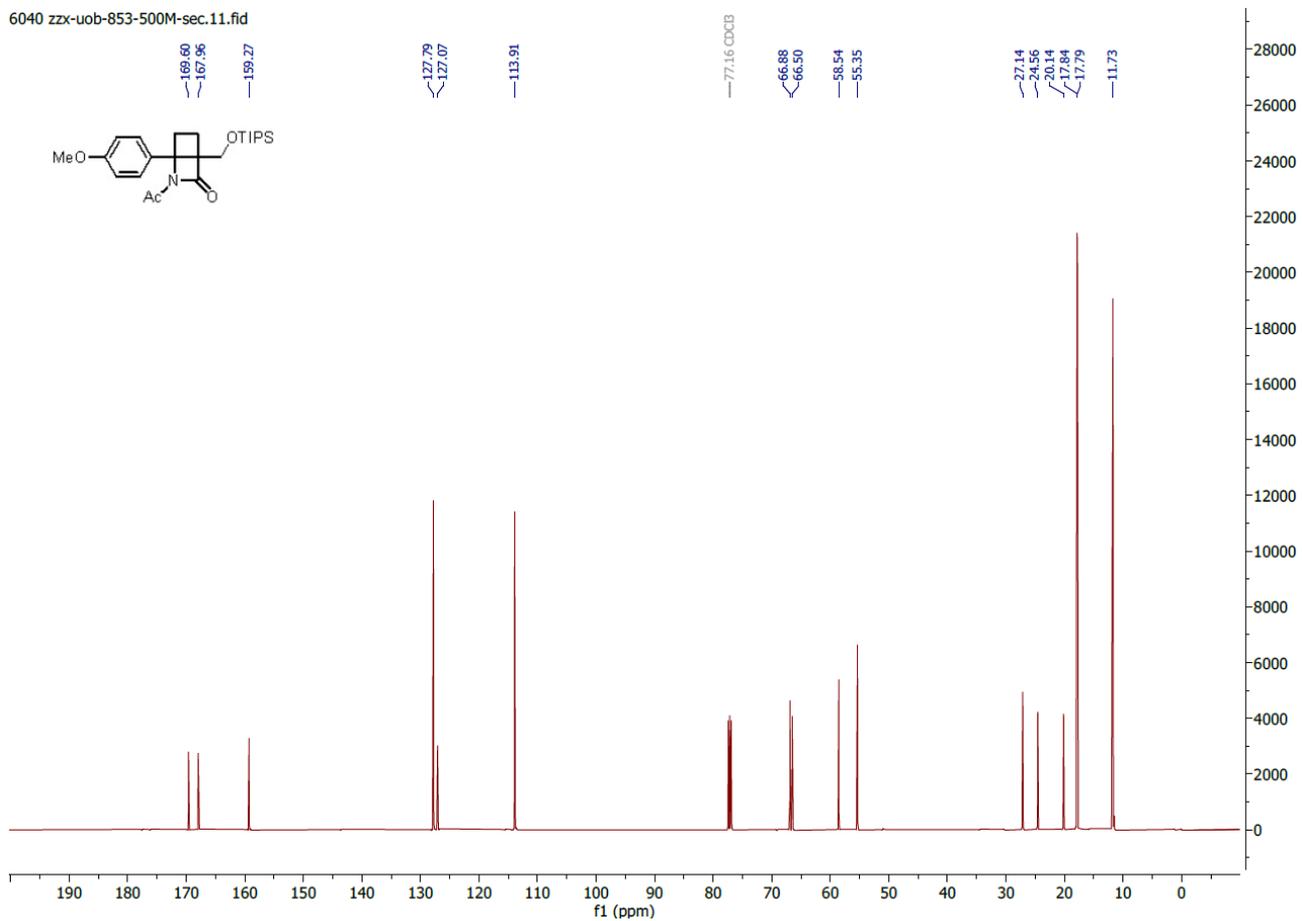
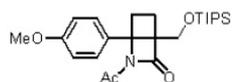


Compound 6ai

6040 zzx-uob-853-500M-sec.10.fid

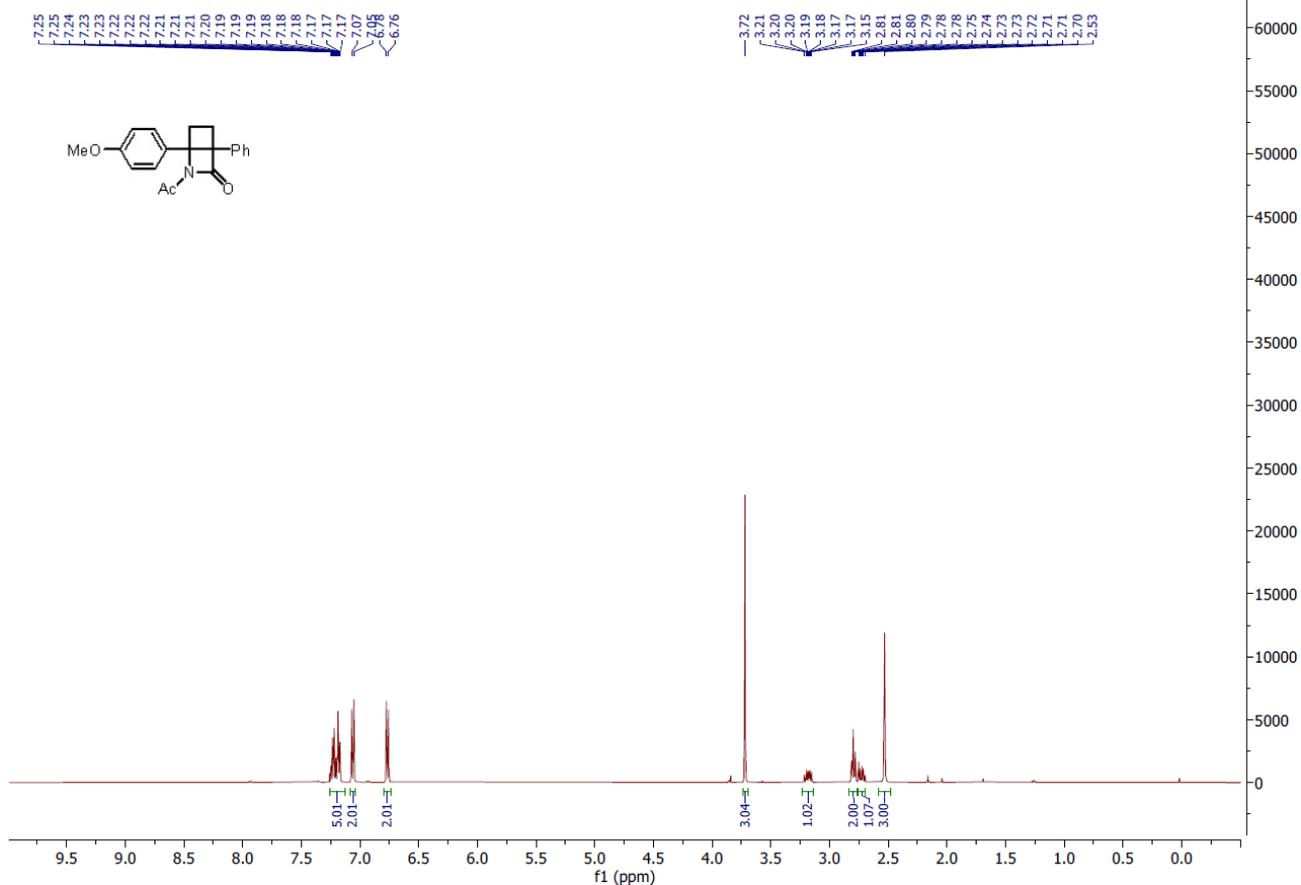


6040 zzx-uob-853-500M-sec.11.fid

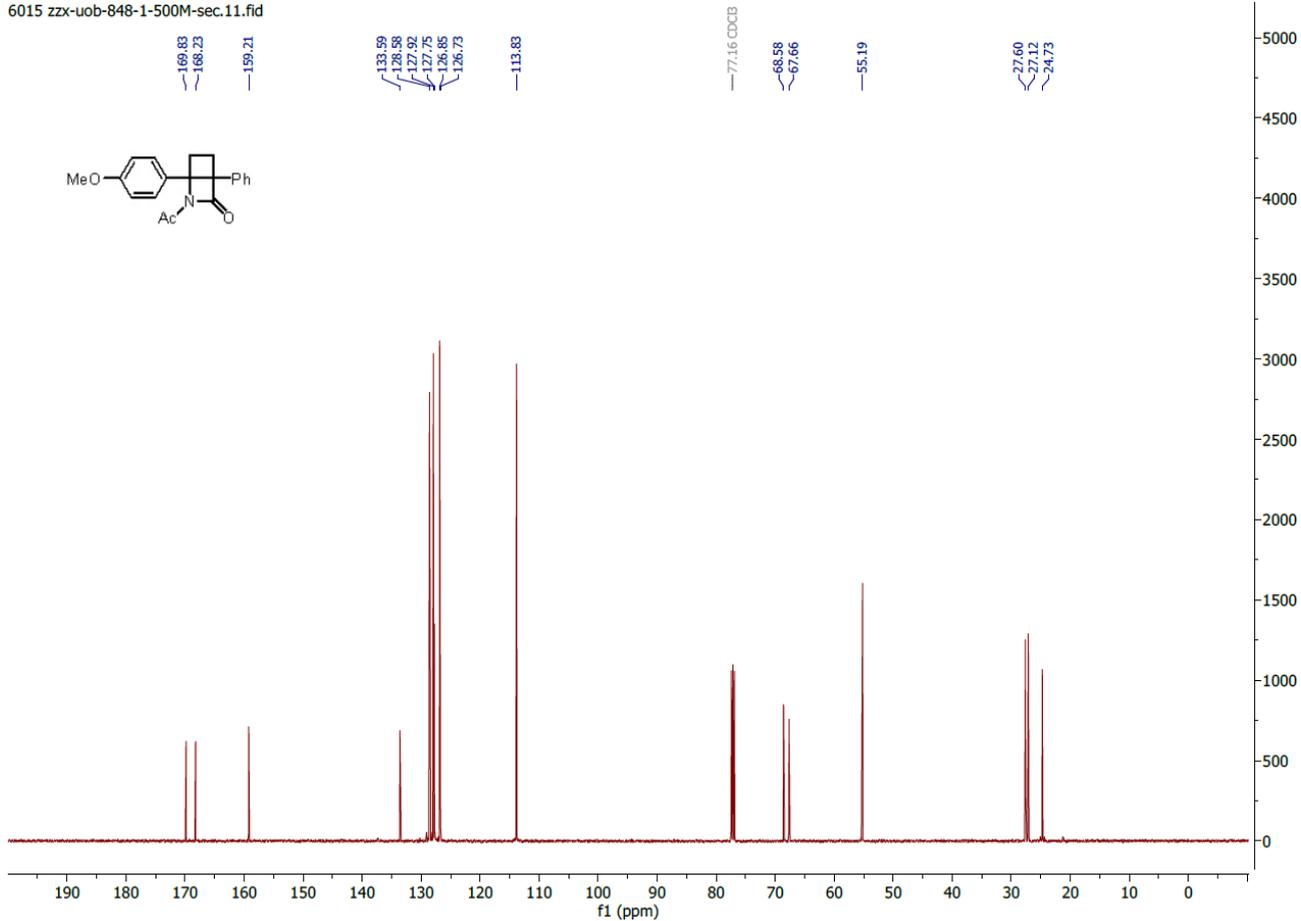


Compound 6aj

6015 zzx-uob-848-1-500M-sec.10.fid

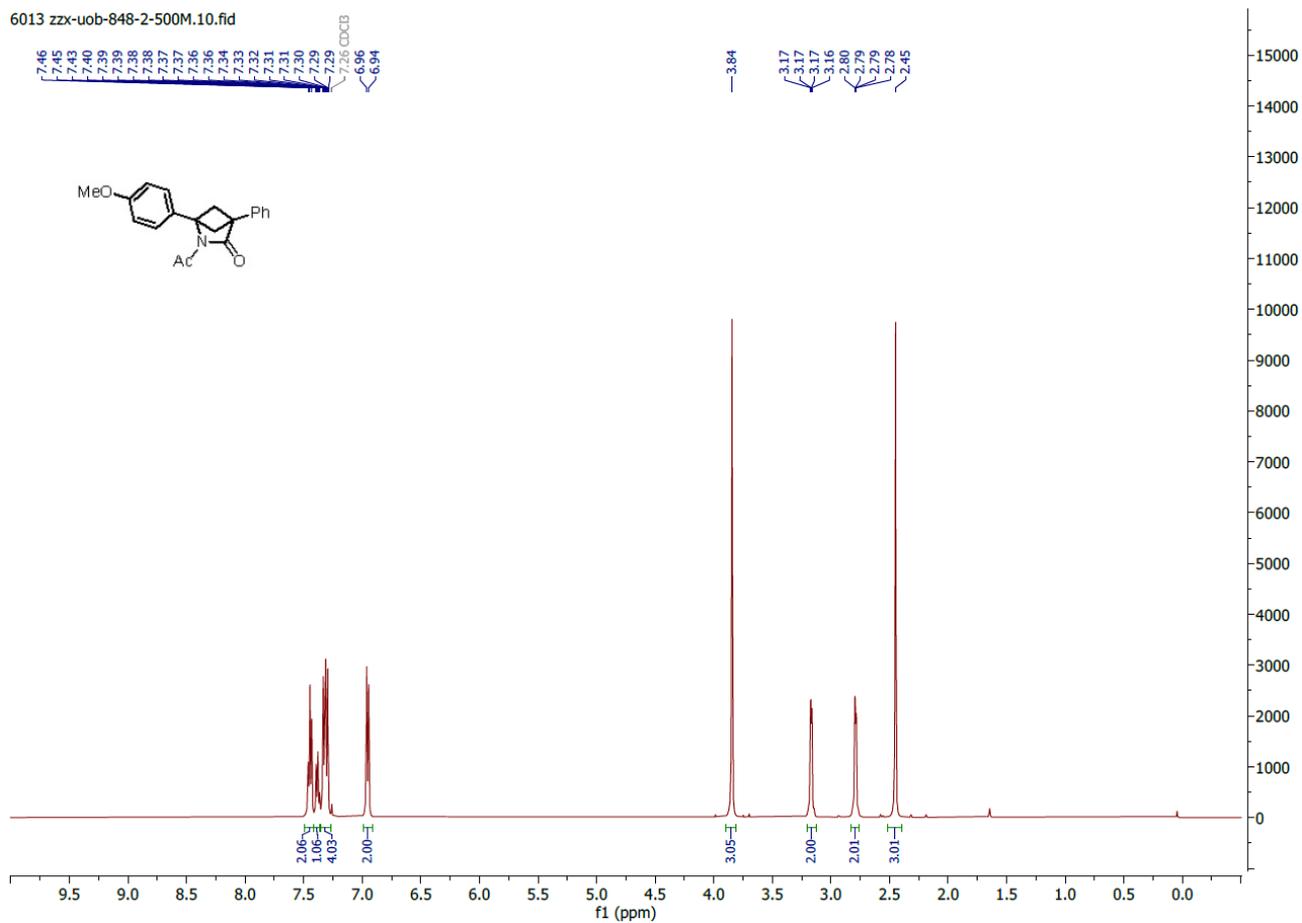


6015 zzx-uob-848-1-500M-sec.11.fid

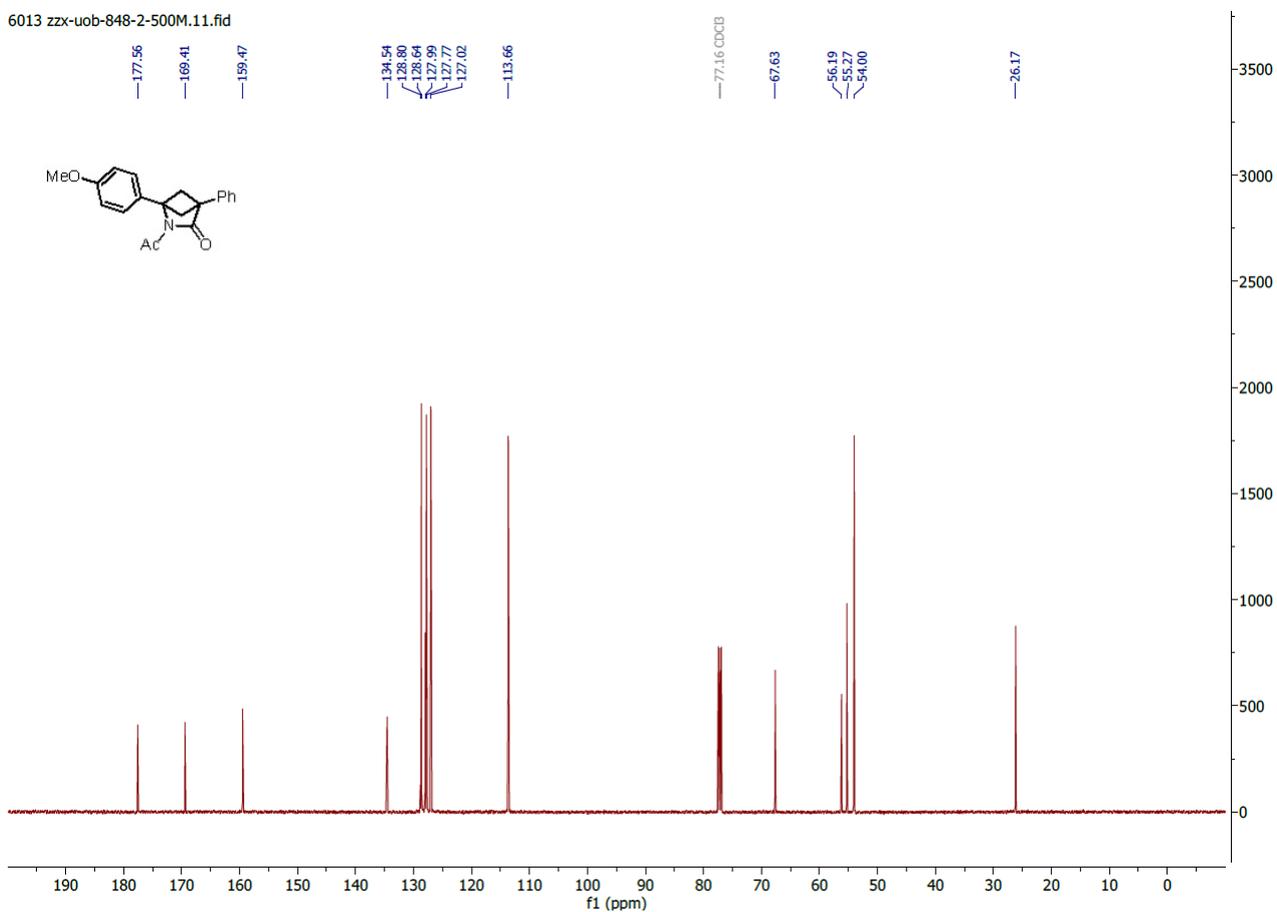


Compound 5aj

6013 zzx-uob-848-2-500M.10.fid

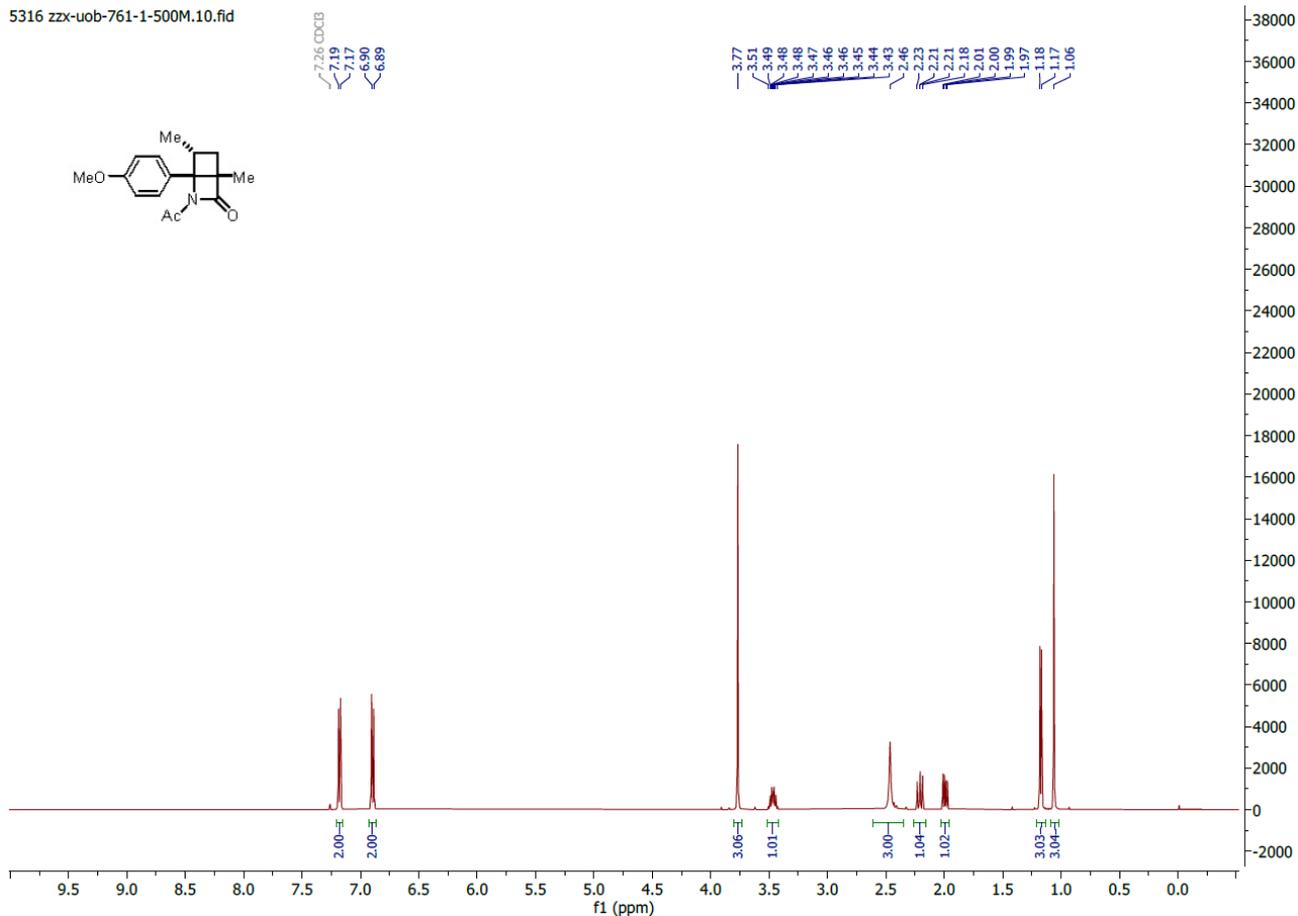
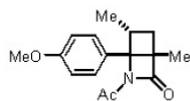


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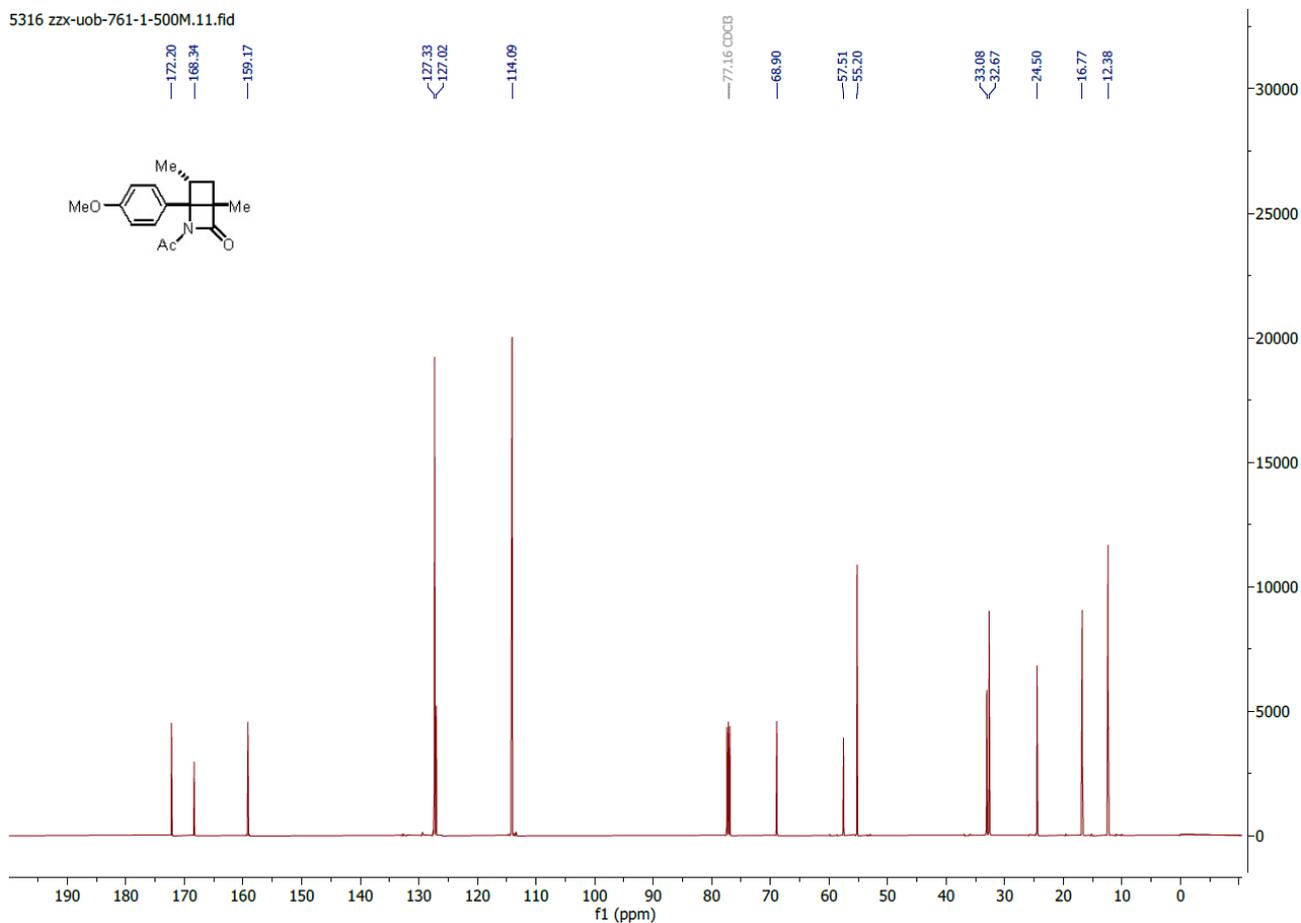
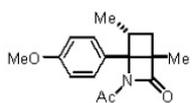


Compound 6ak

5316 zzx-uob-761-1-500M.10.fid

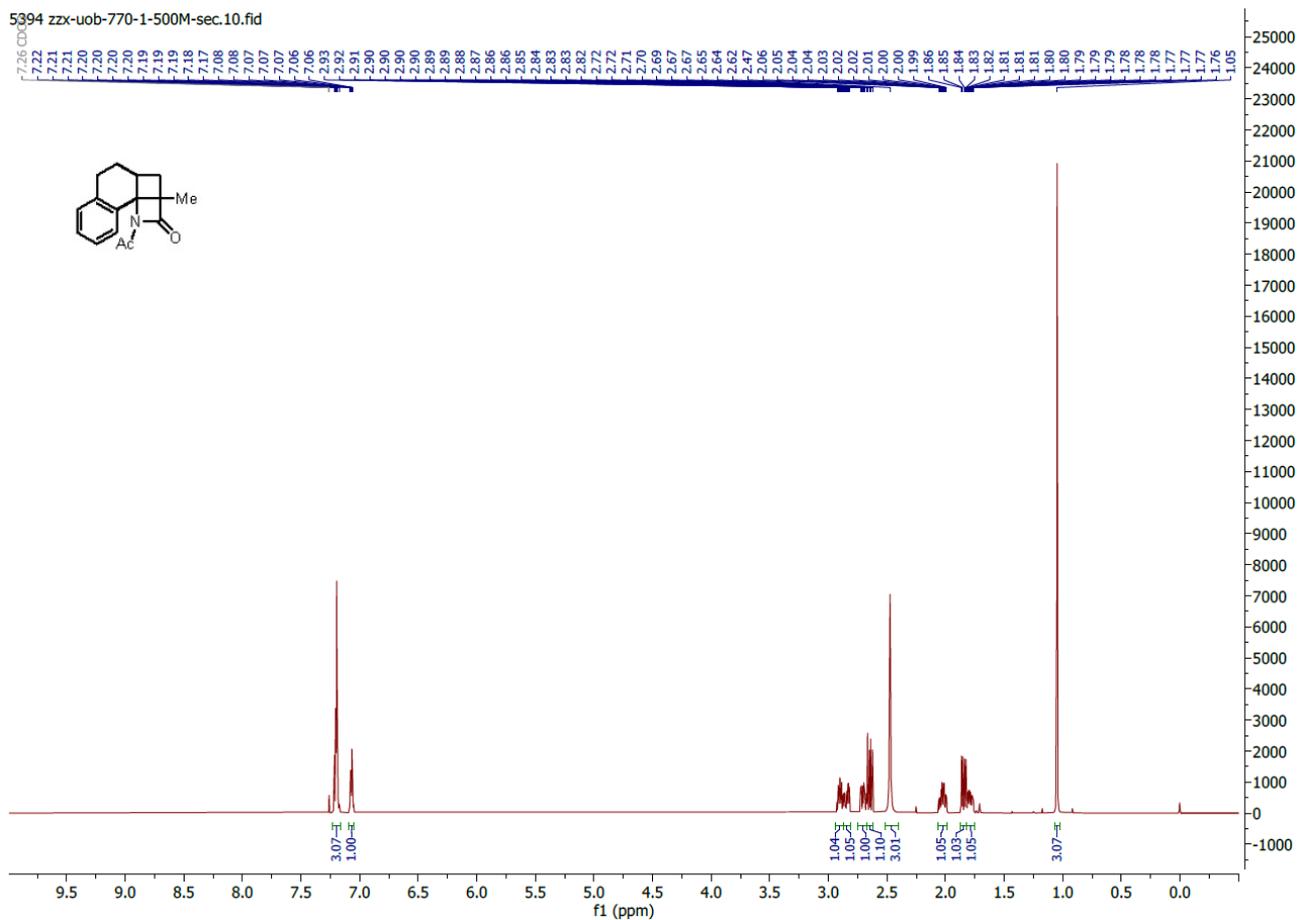


5316 zzx-uob-761-1-500M.11.fid

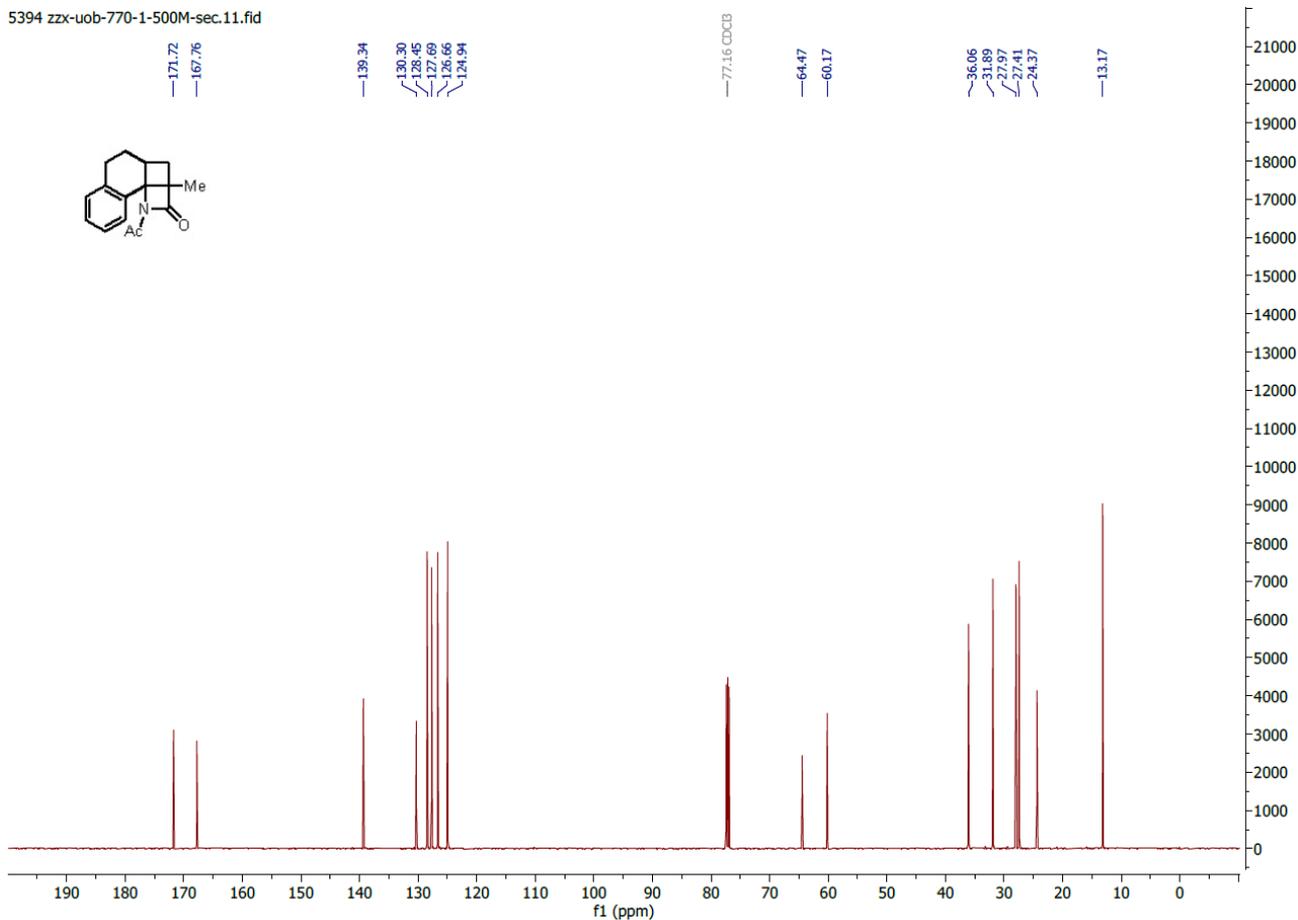


Compound 6al

5394 zzx-uob-770-1-500M-sec.10.fid

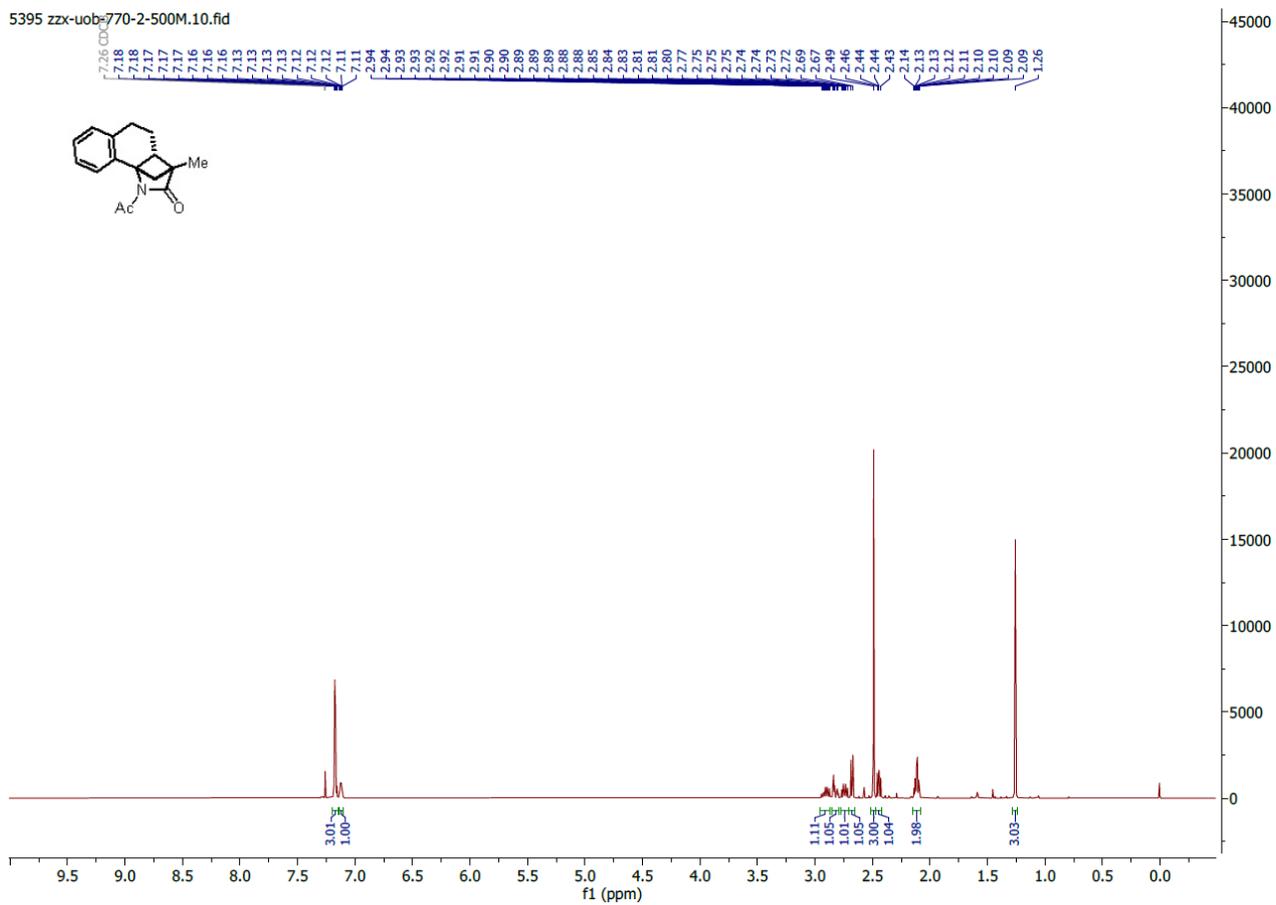


5394 zzx-uob-770-1-500M-sec.11.fid

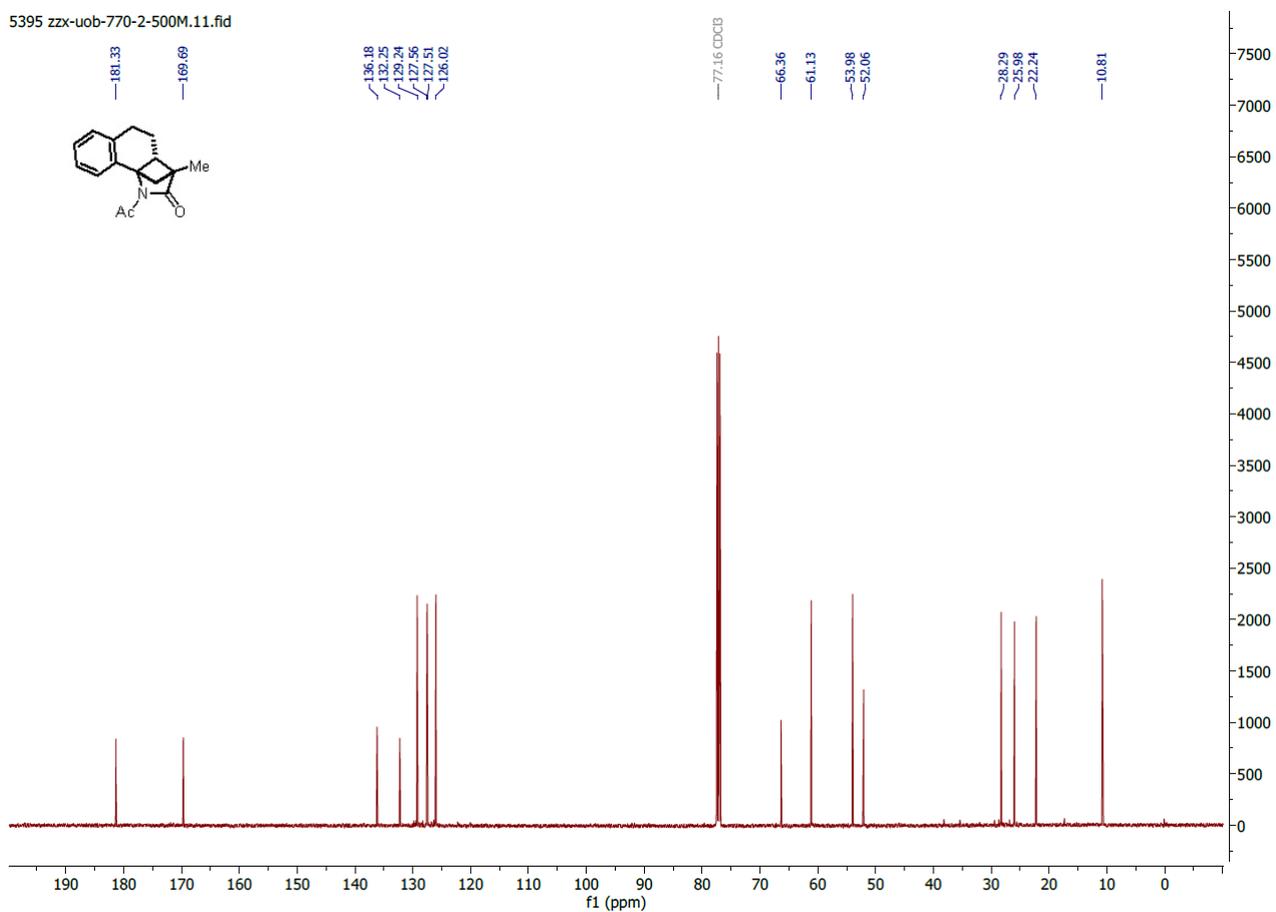


Compound 5aI

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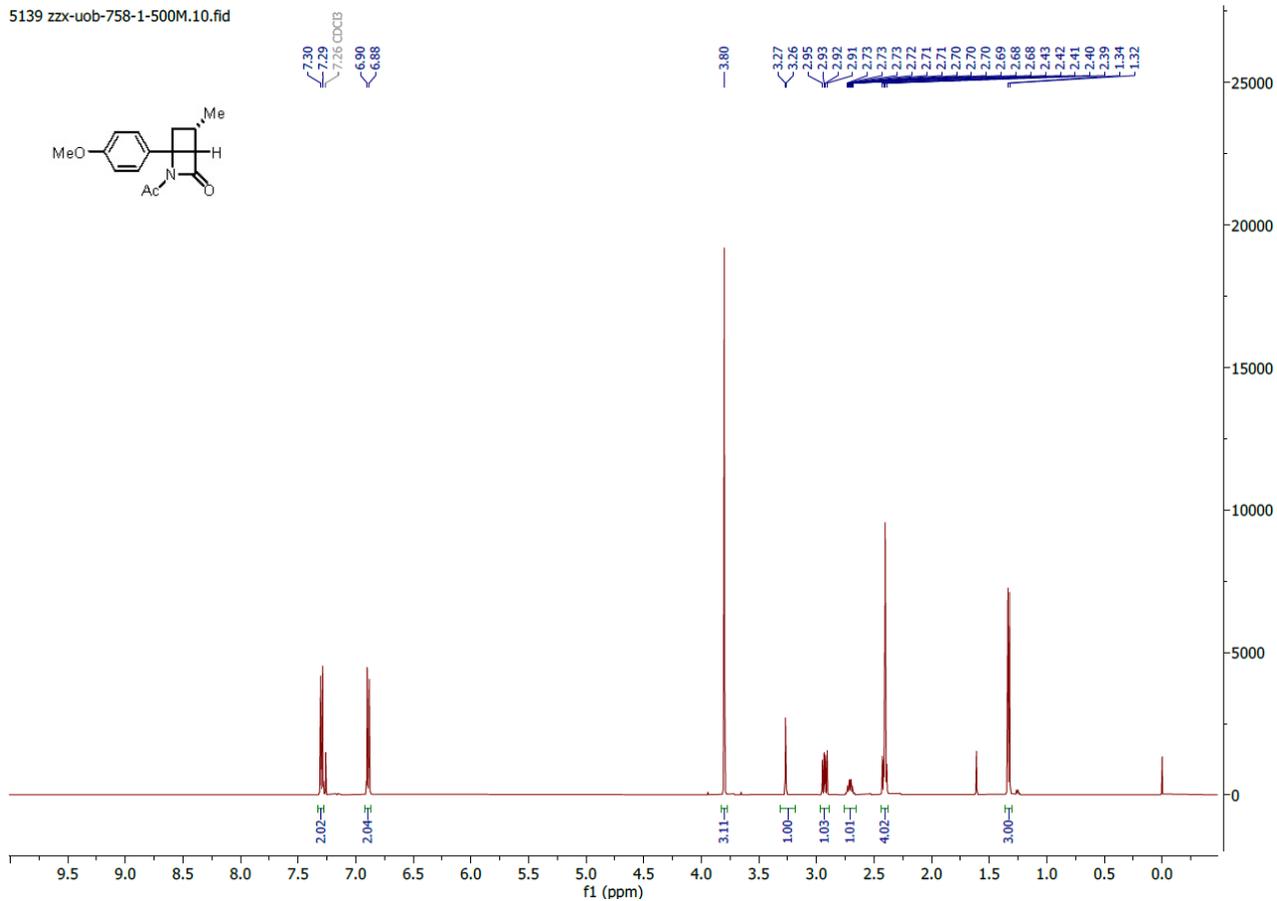
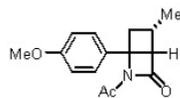


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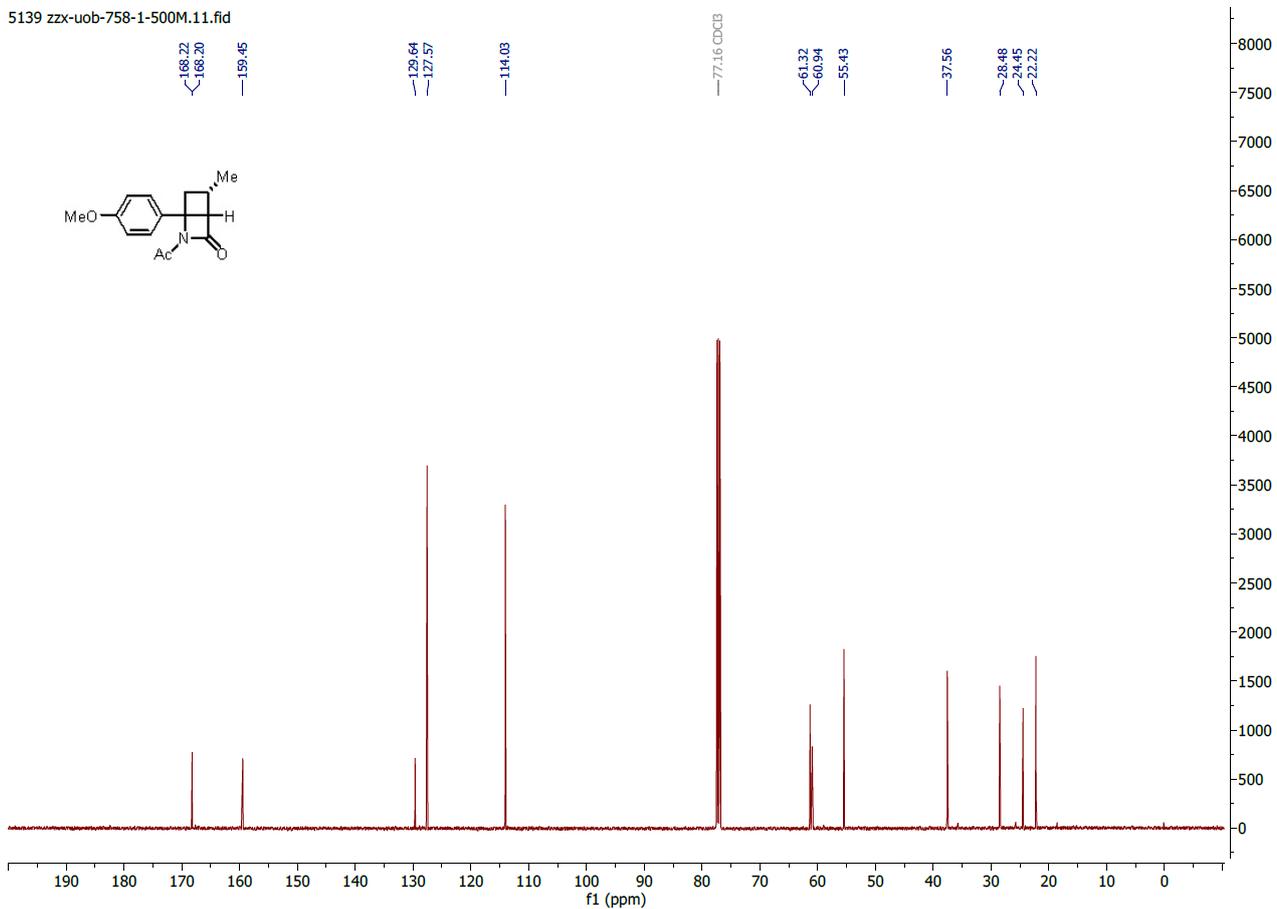
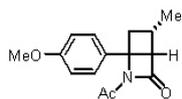


Compound 6am

5139 zzx-uob-758-1-500M.10.fid

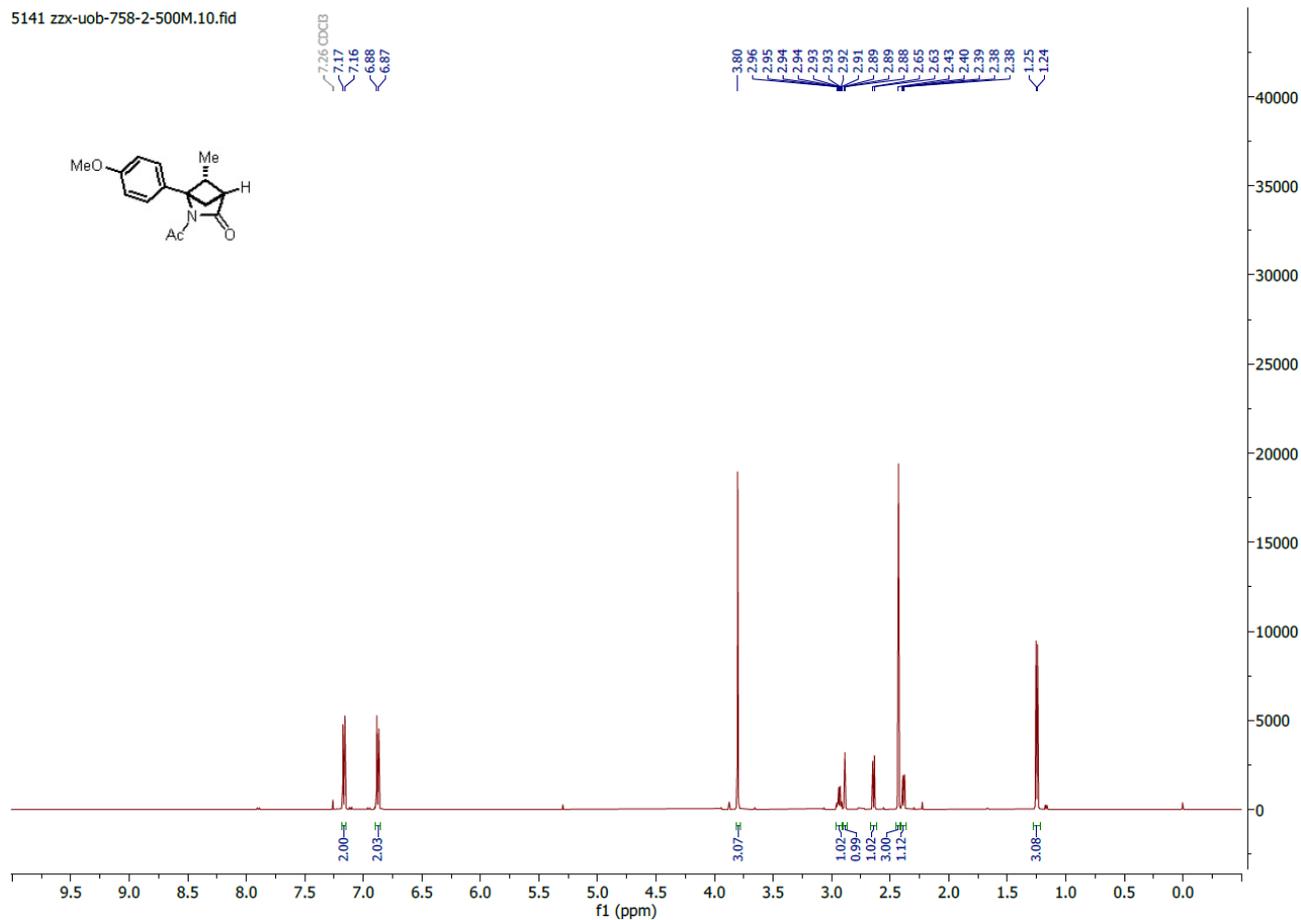


5139 zzx-uob-758-1-500M.11.fid

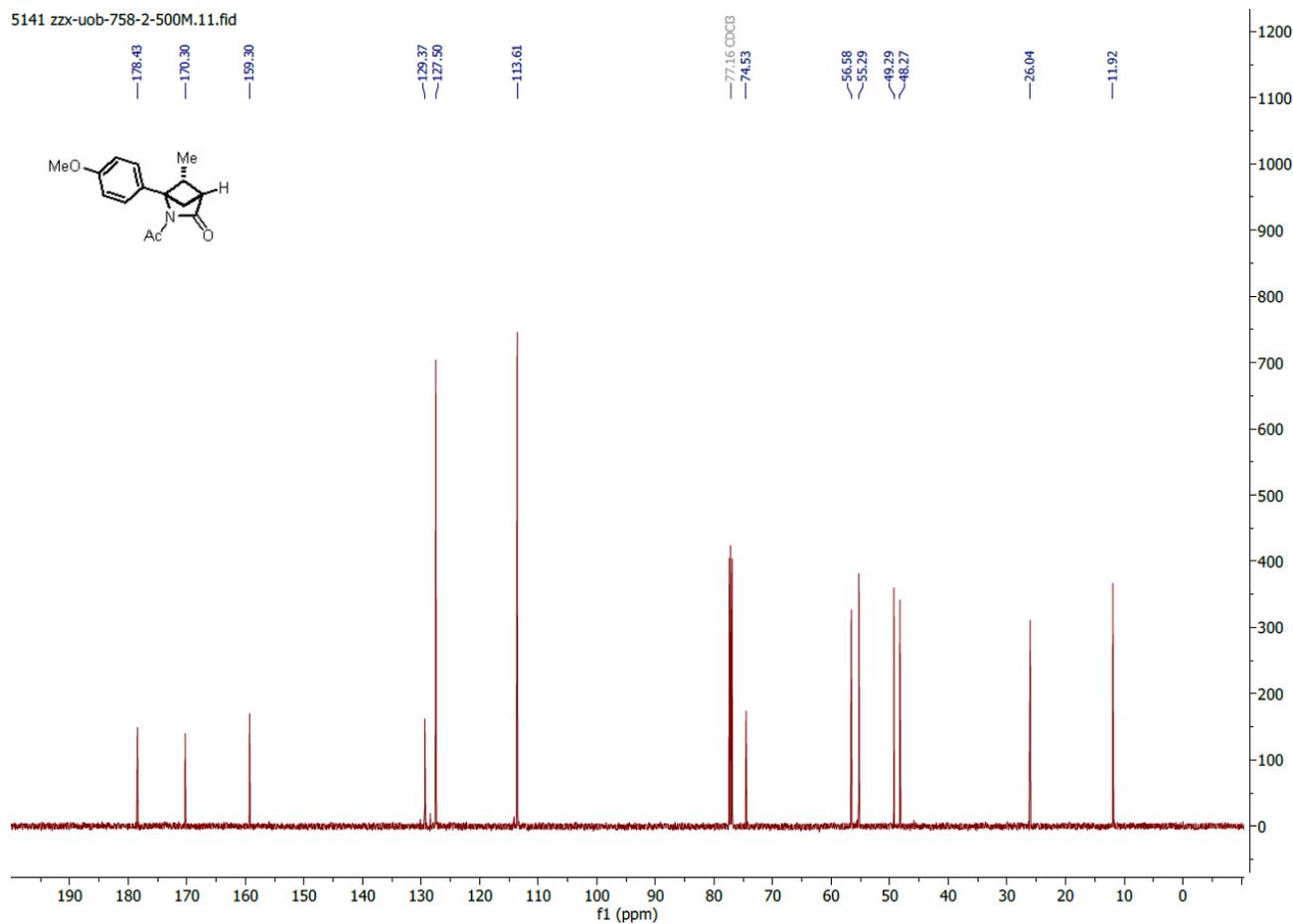


Compound 5am

5141 zzx-uob-758-2-500M.10.fid

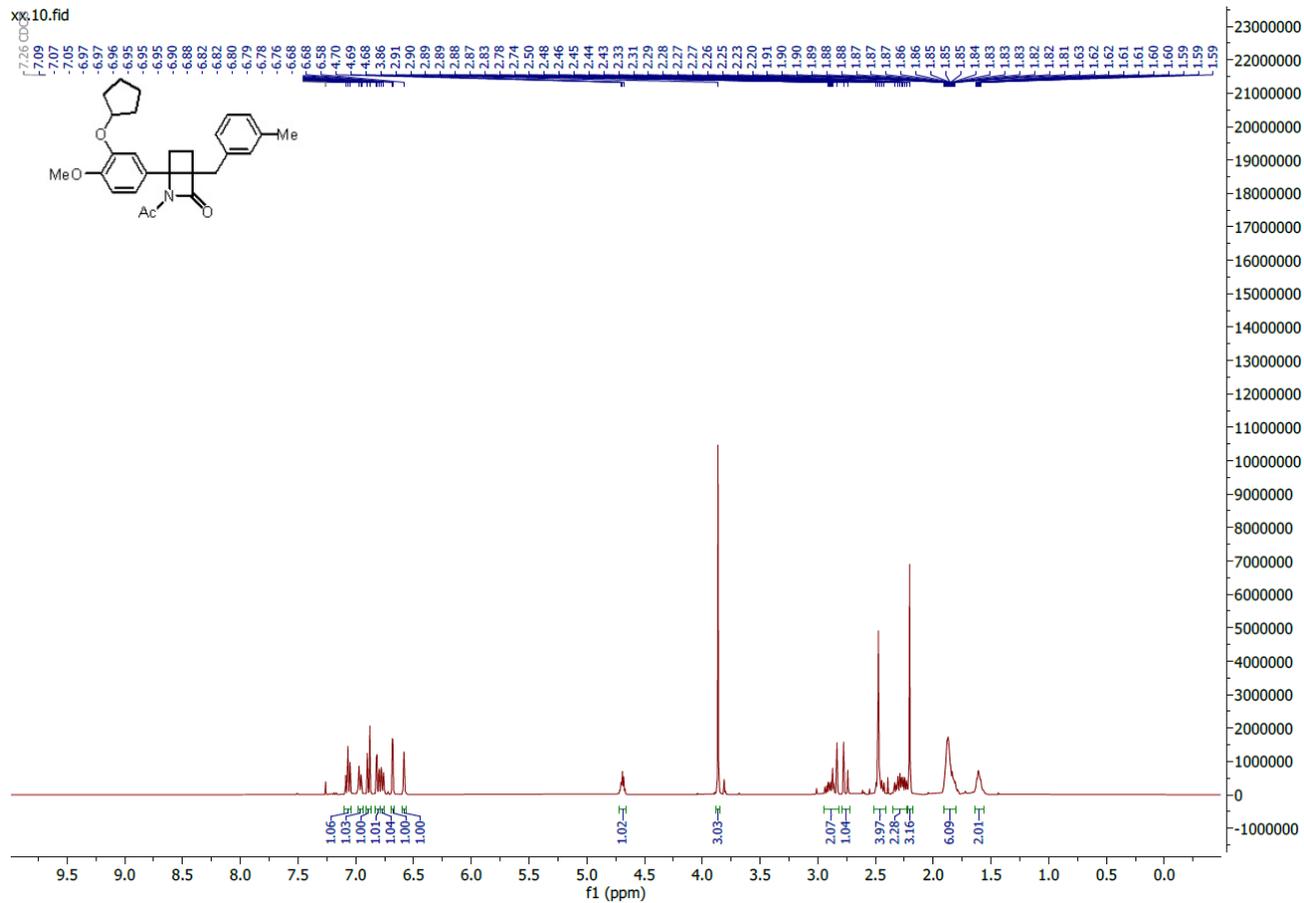


5141 zzx-uob-758-2-500M.11.fid

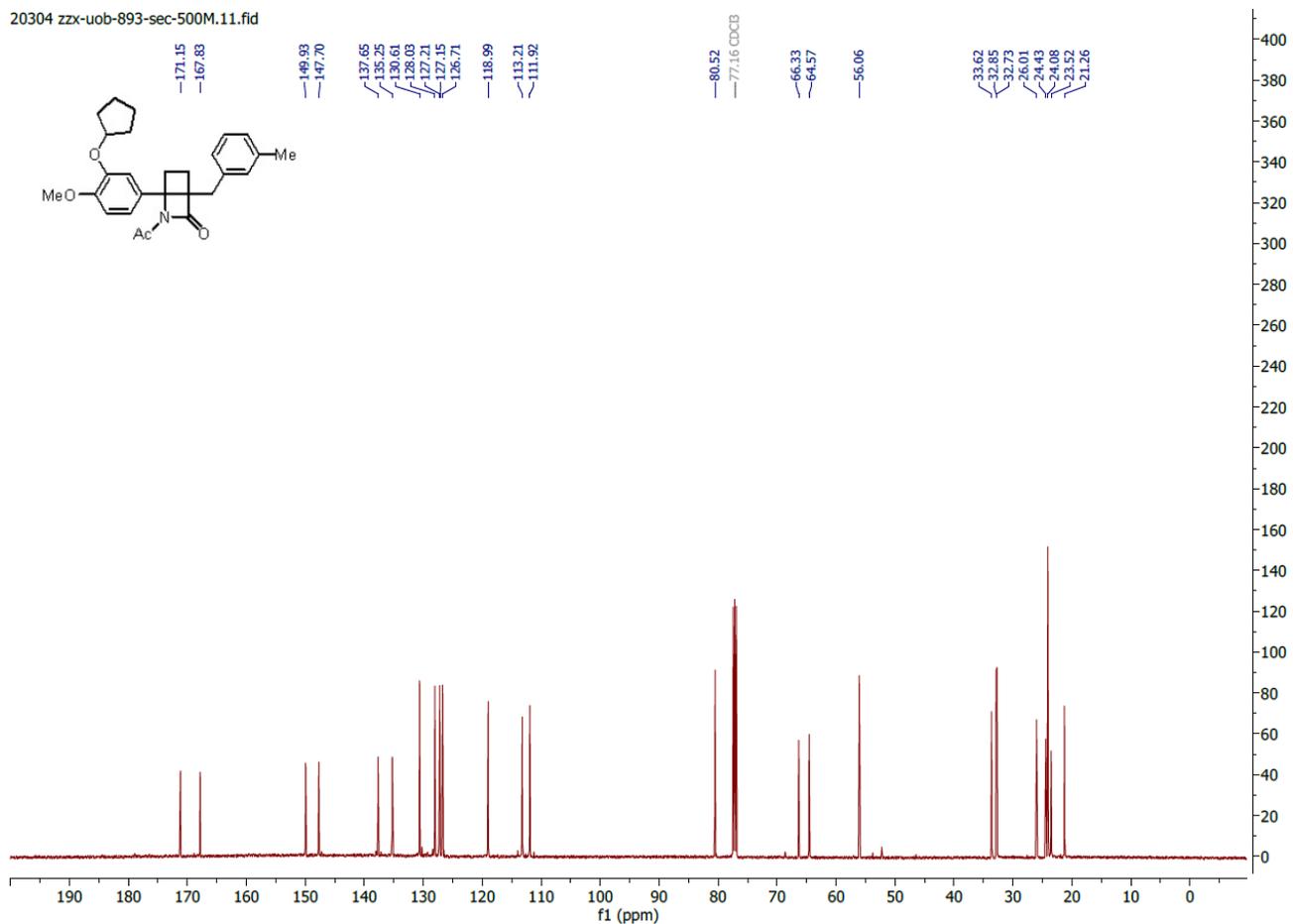


Compound 6an

xx.10.fid



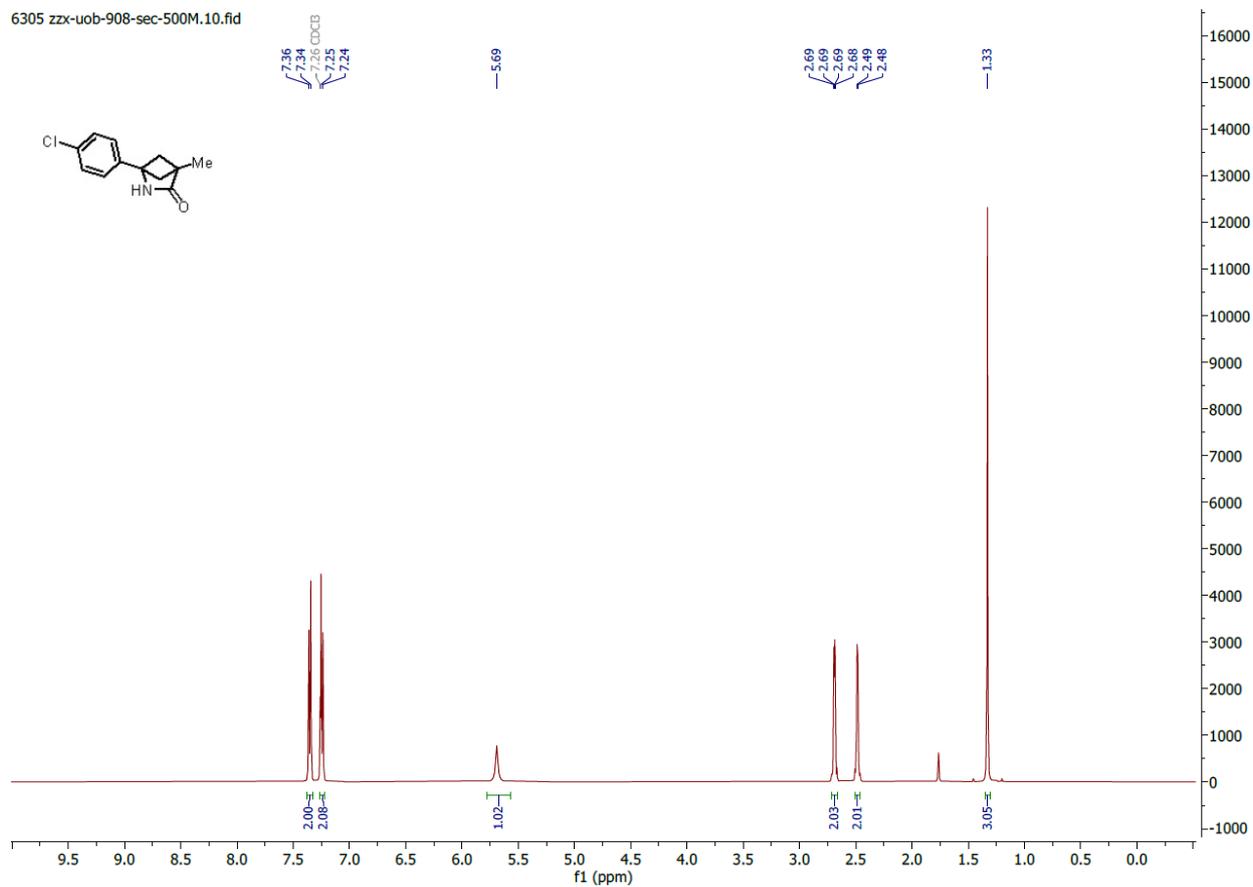
20304 zzx-uob-893-sec-500M.11.fid



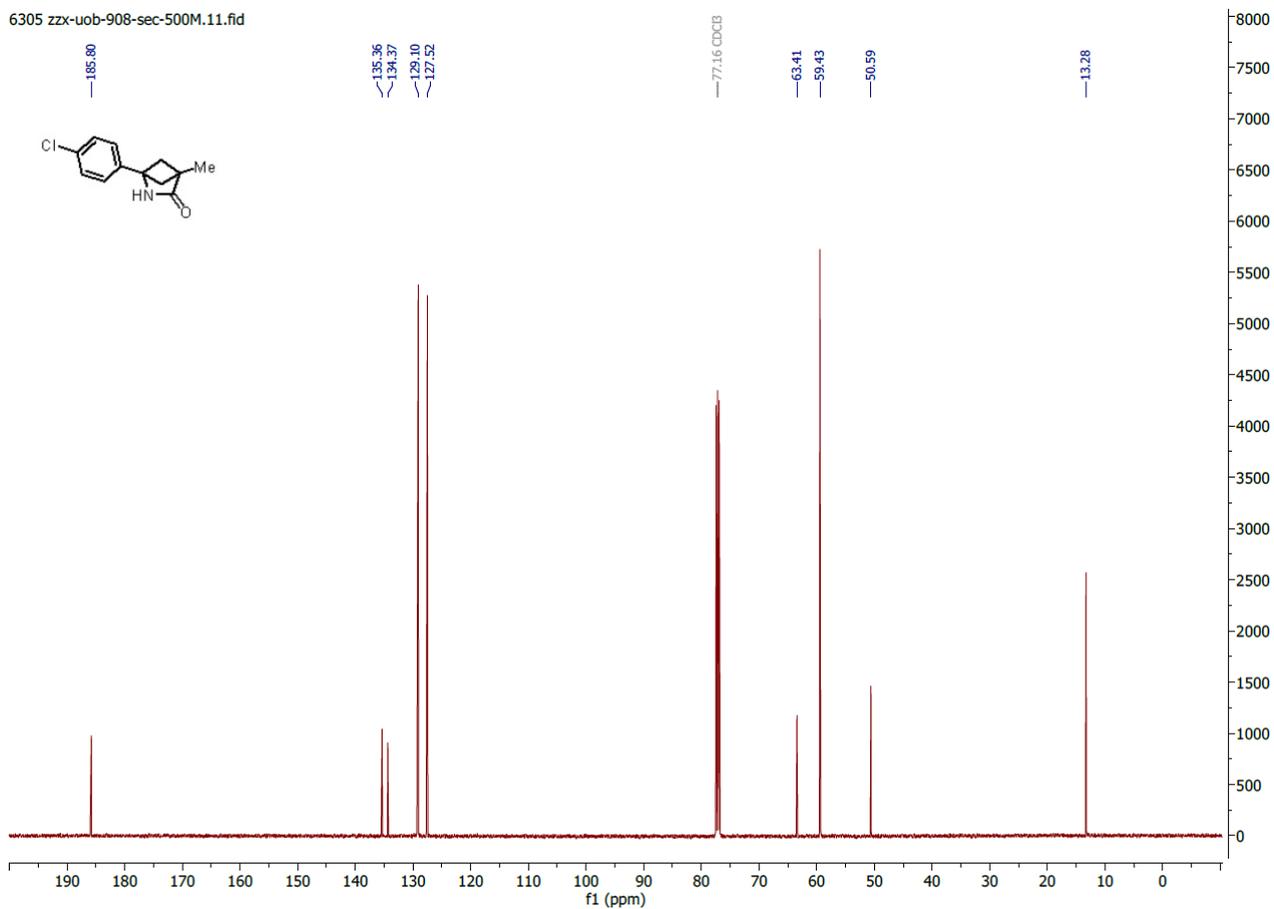
5.7 NMR Spectra of Product Derivatives

Compound 3c'

6305 zzx-uob-908-sec-500M.10.fid

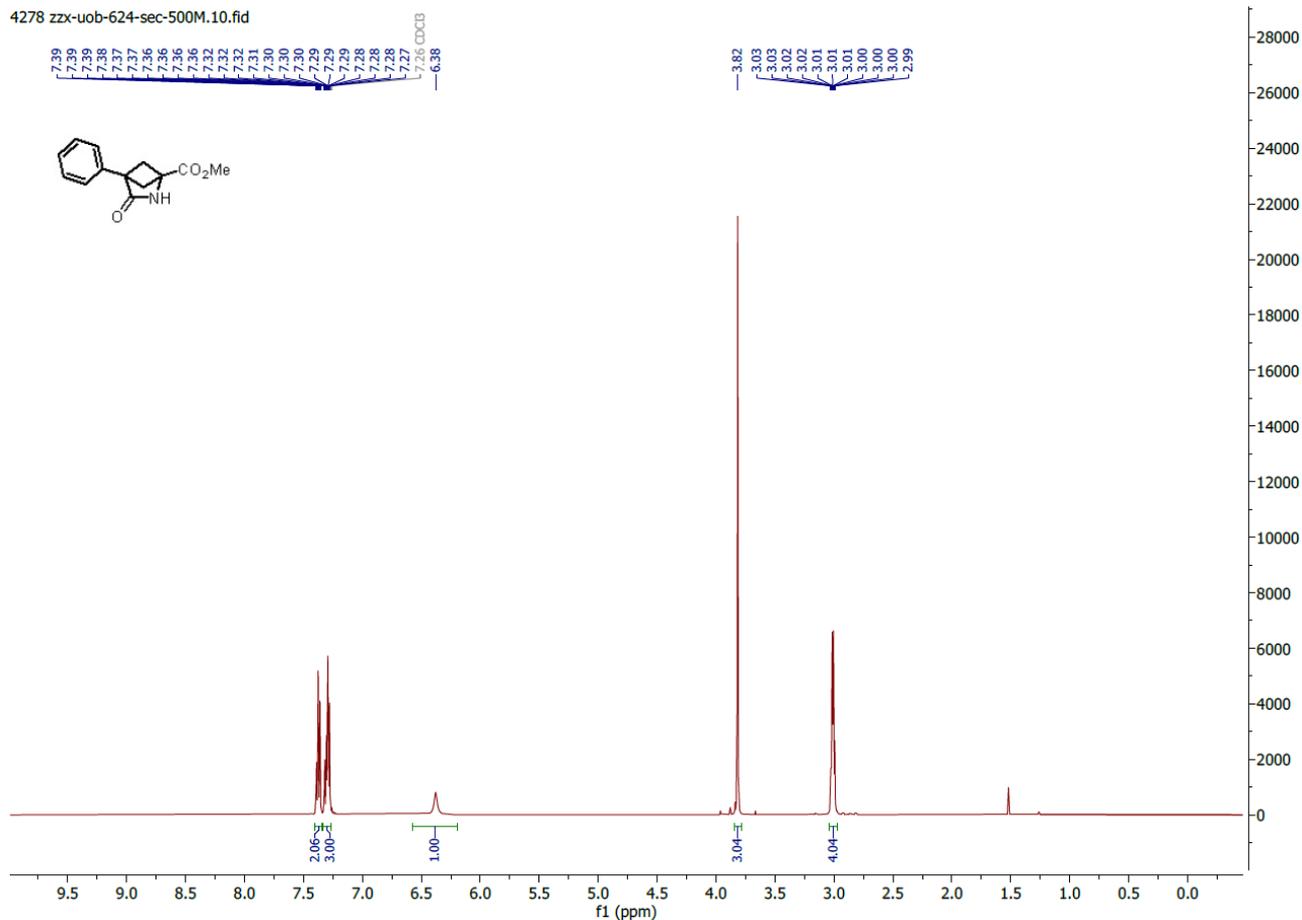


6305 zzx-uob-908-sec-500M.11.fid

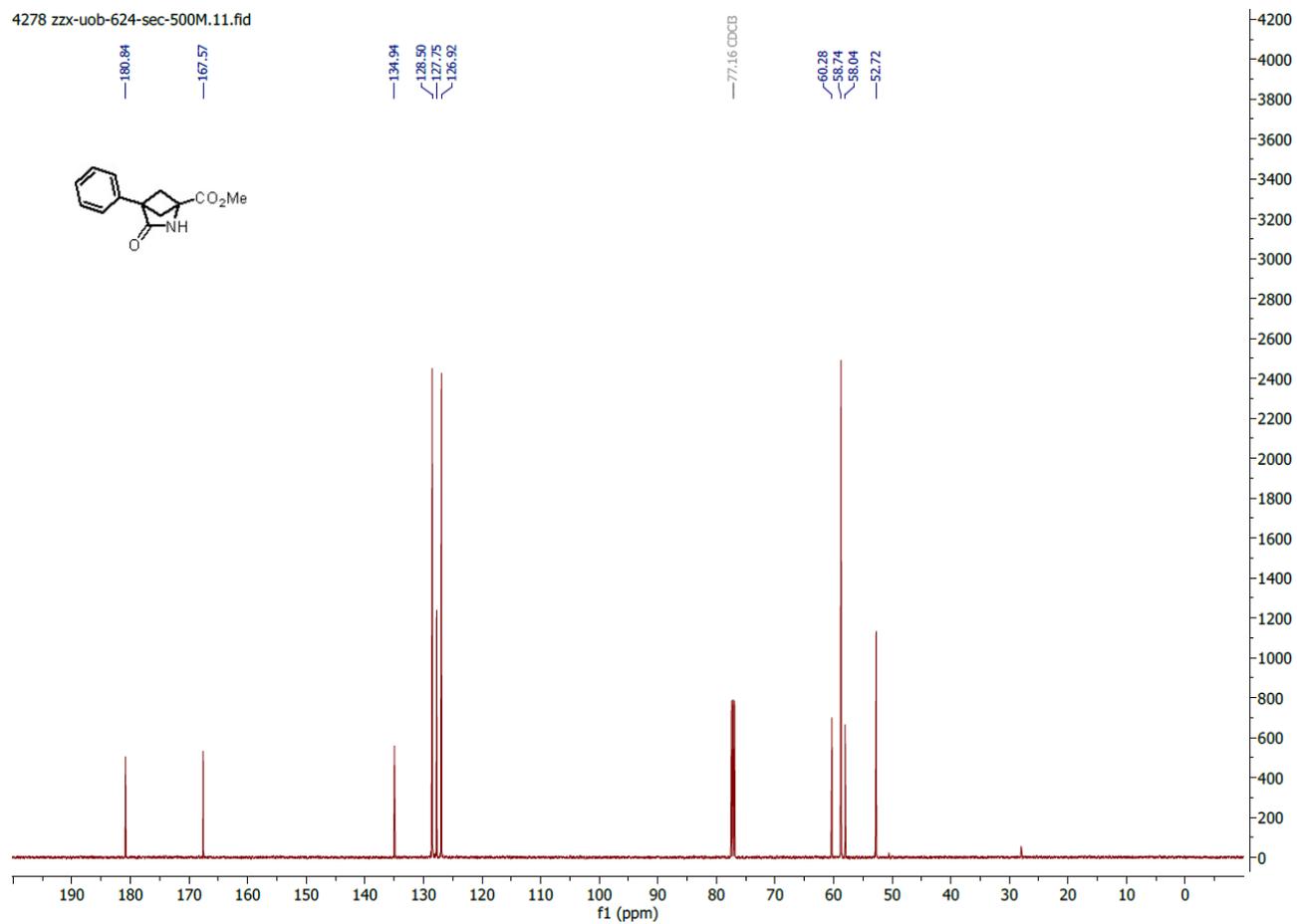


Compound 8a'

4278 zzx-uob-624-sec-500M.10.fid

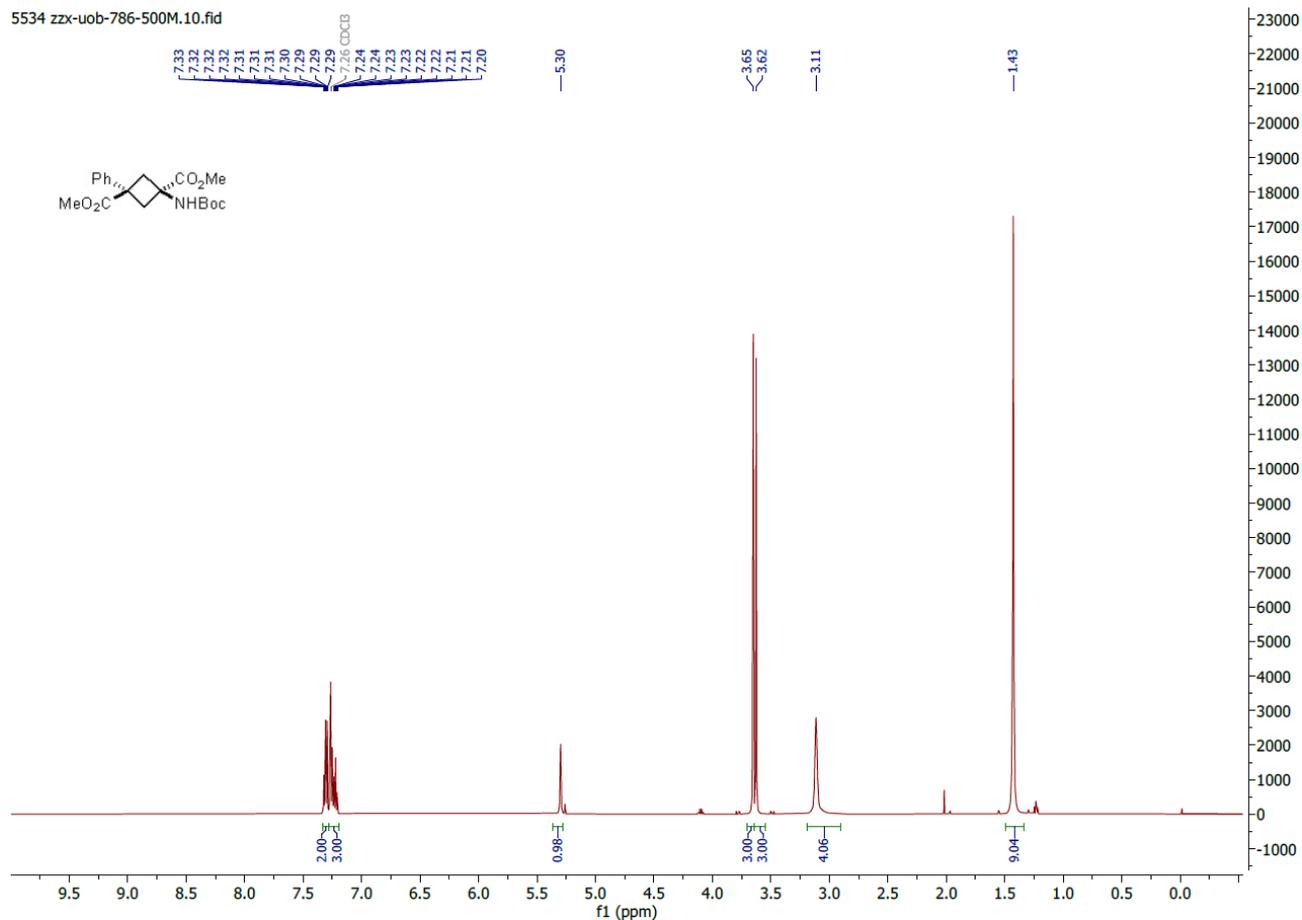


4278 zzx-uob-624-sec-500M.11.fid

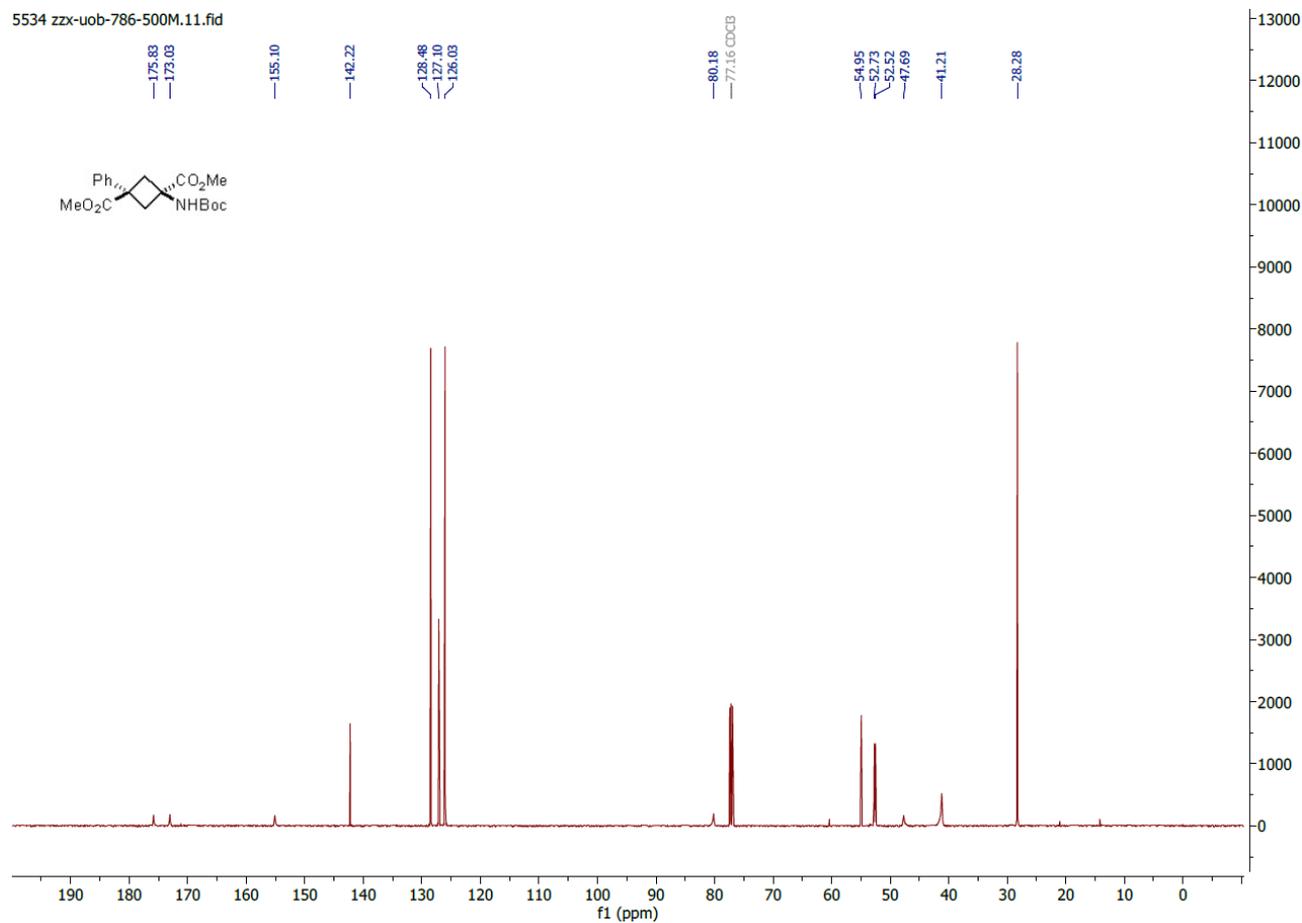


Compound 8a''

5534 zzx-uob-786-500M.10.fid

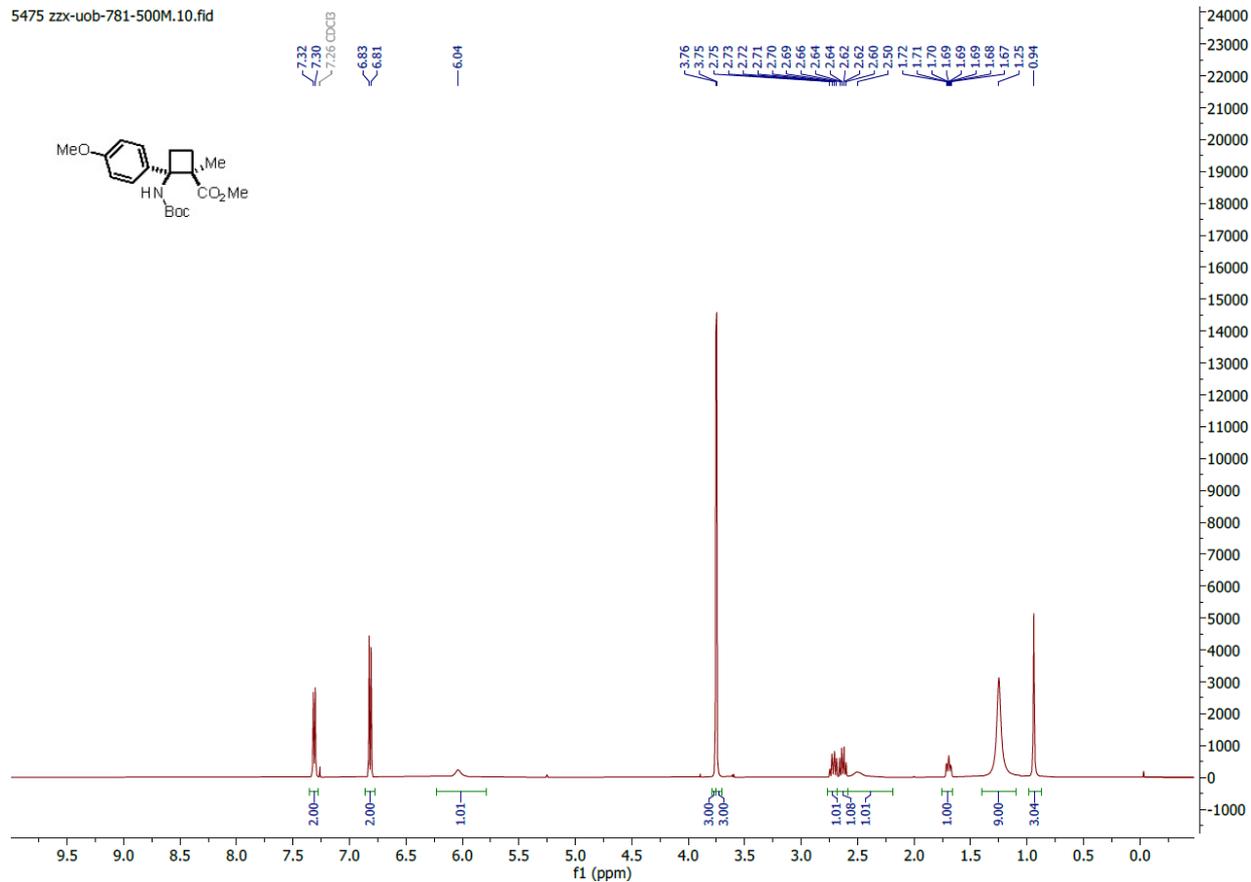


5534 zzx-uob-786-500M.11.fid

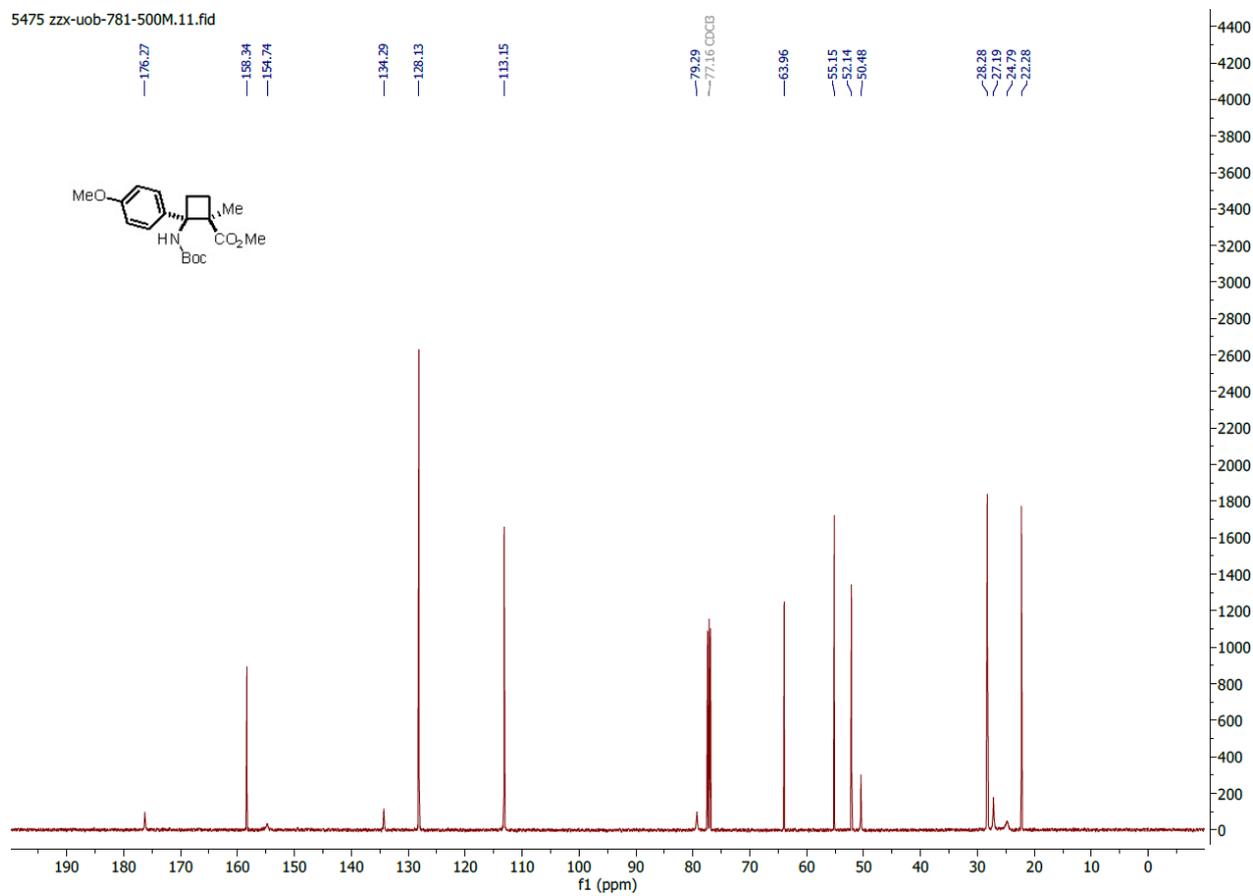


Compound 10

5475 zzx-uob-781-500M.10.fid

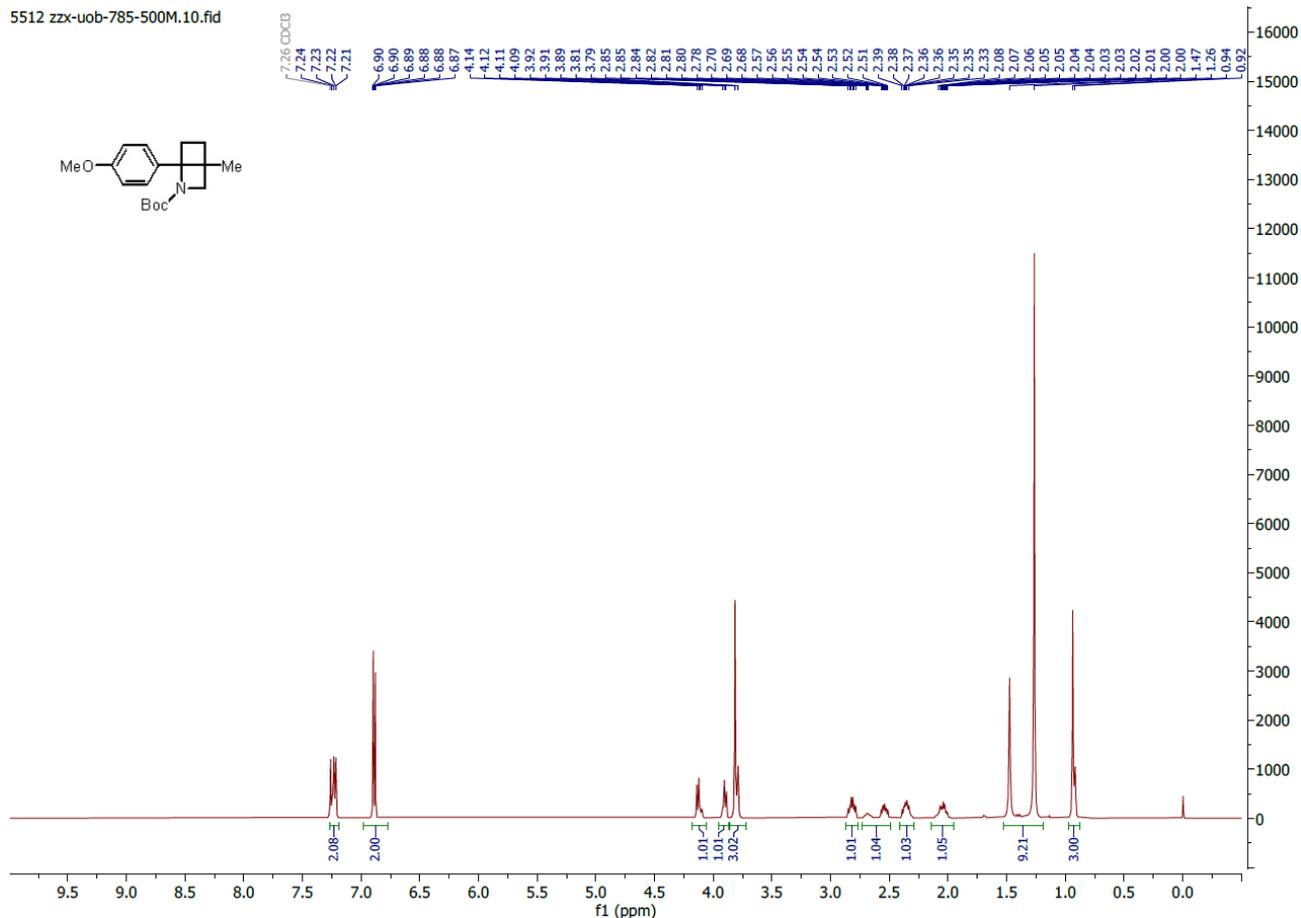
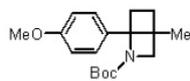


5475 zzx-uob-781-500M.11.fid

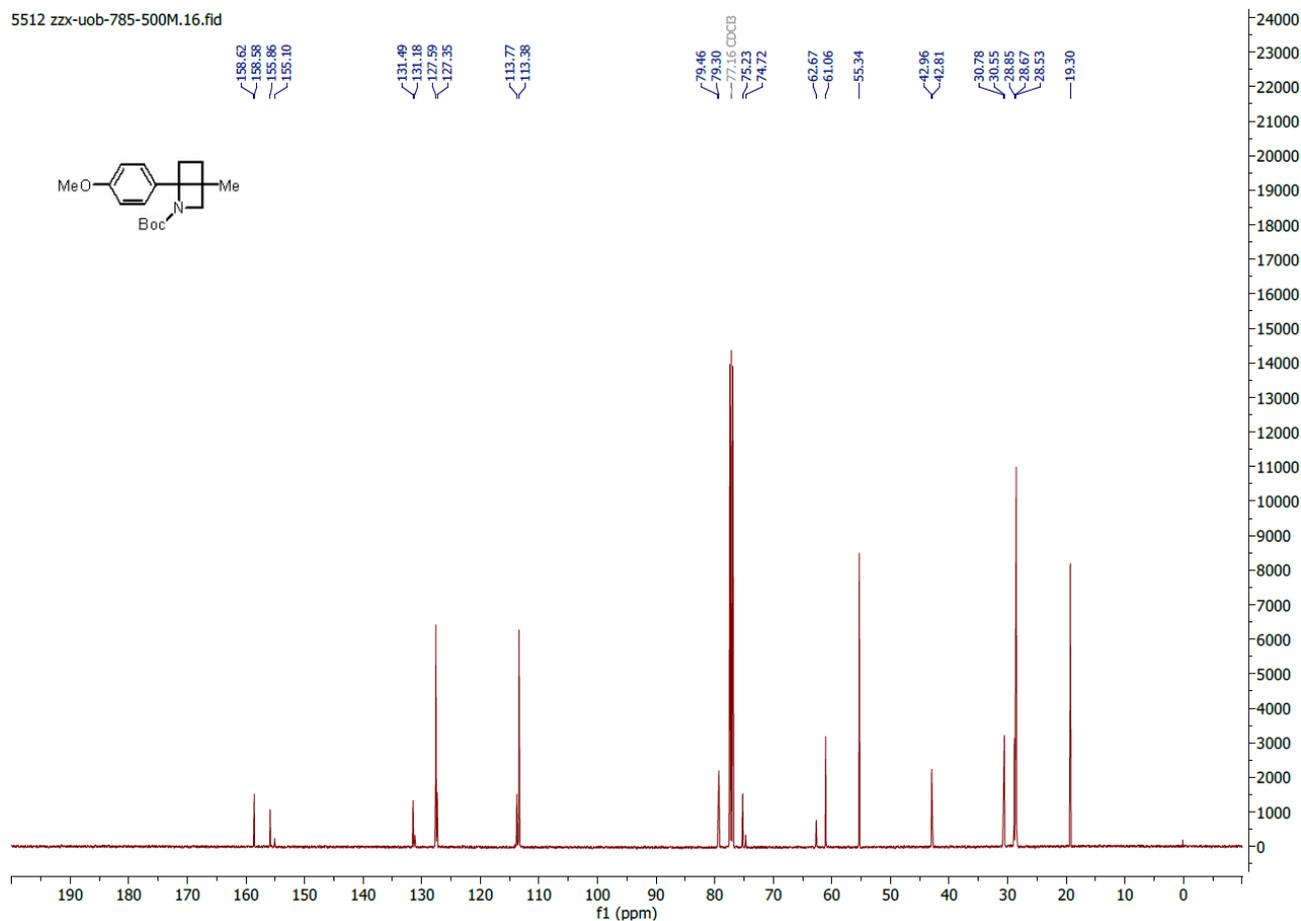
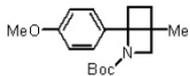


Compound 11

5512 zzx-uob-785-500M.10.fid

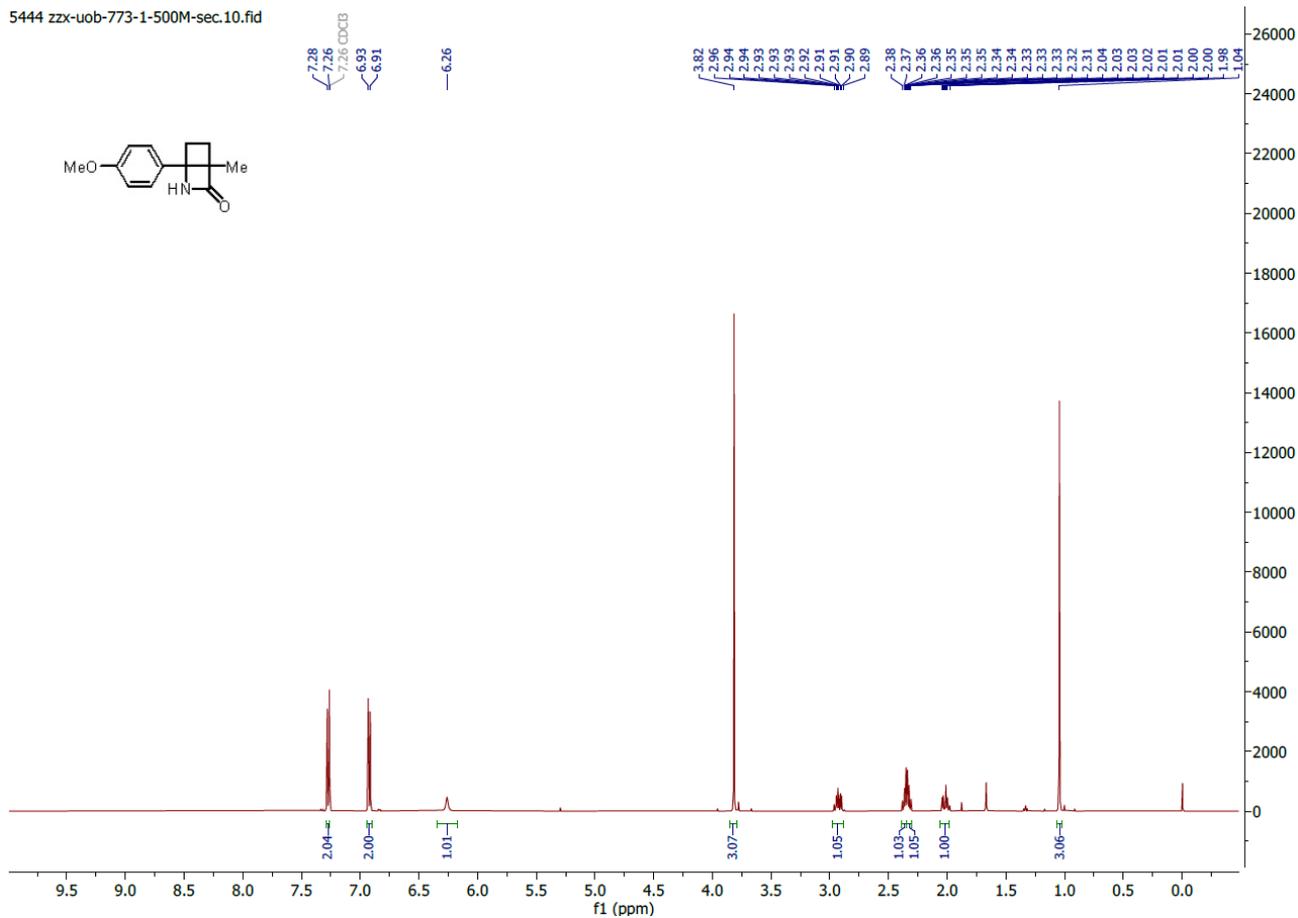
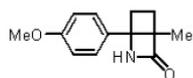


5512 zzx-uob-785-500M.16.fid

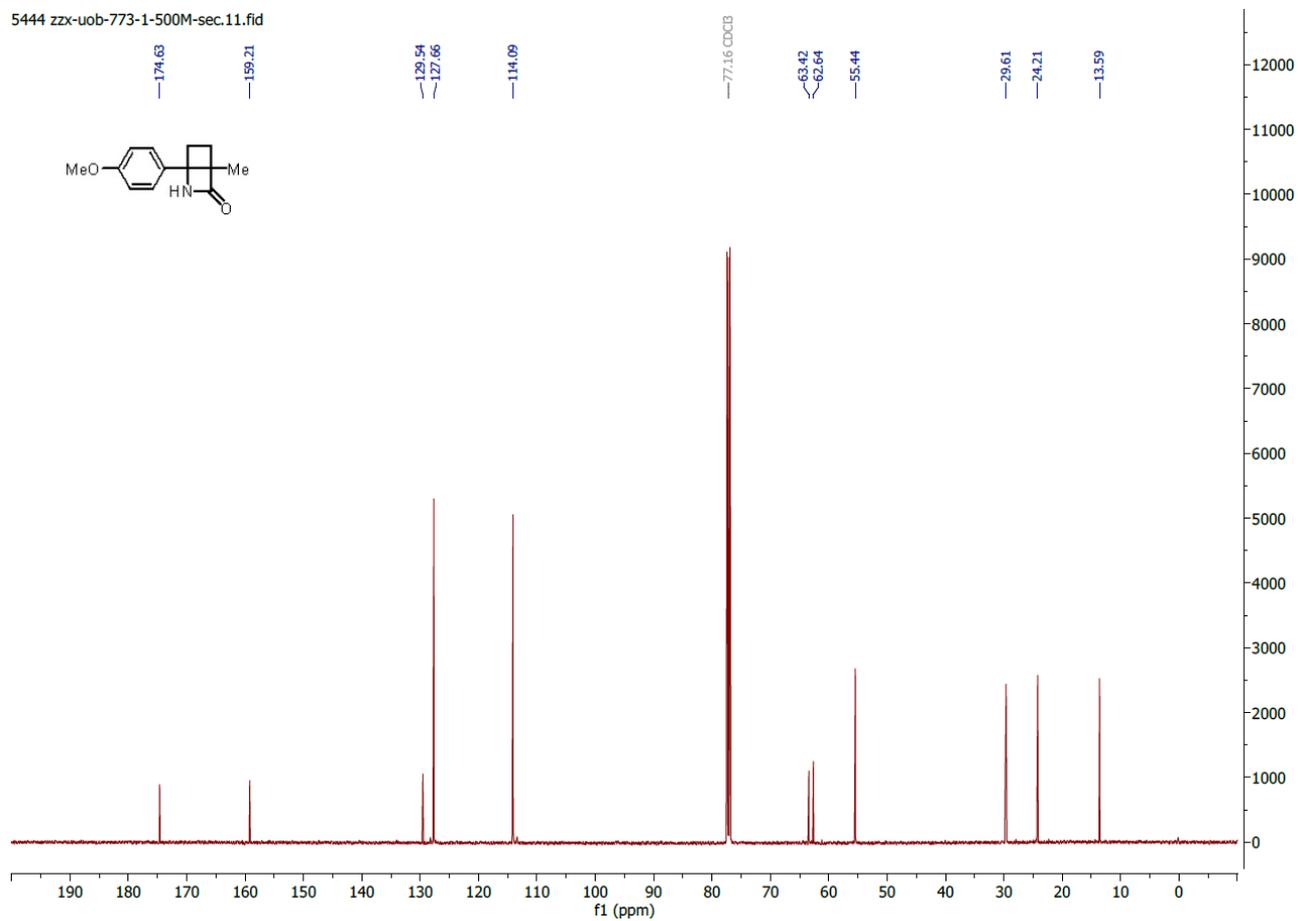
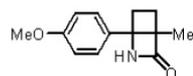


Compound 12

5444 zzx-uob-773-1-500M-sec.10.fid

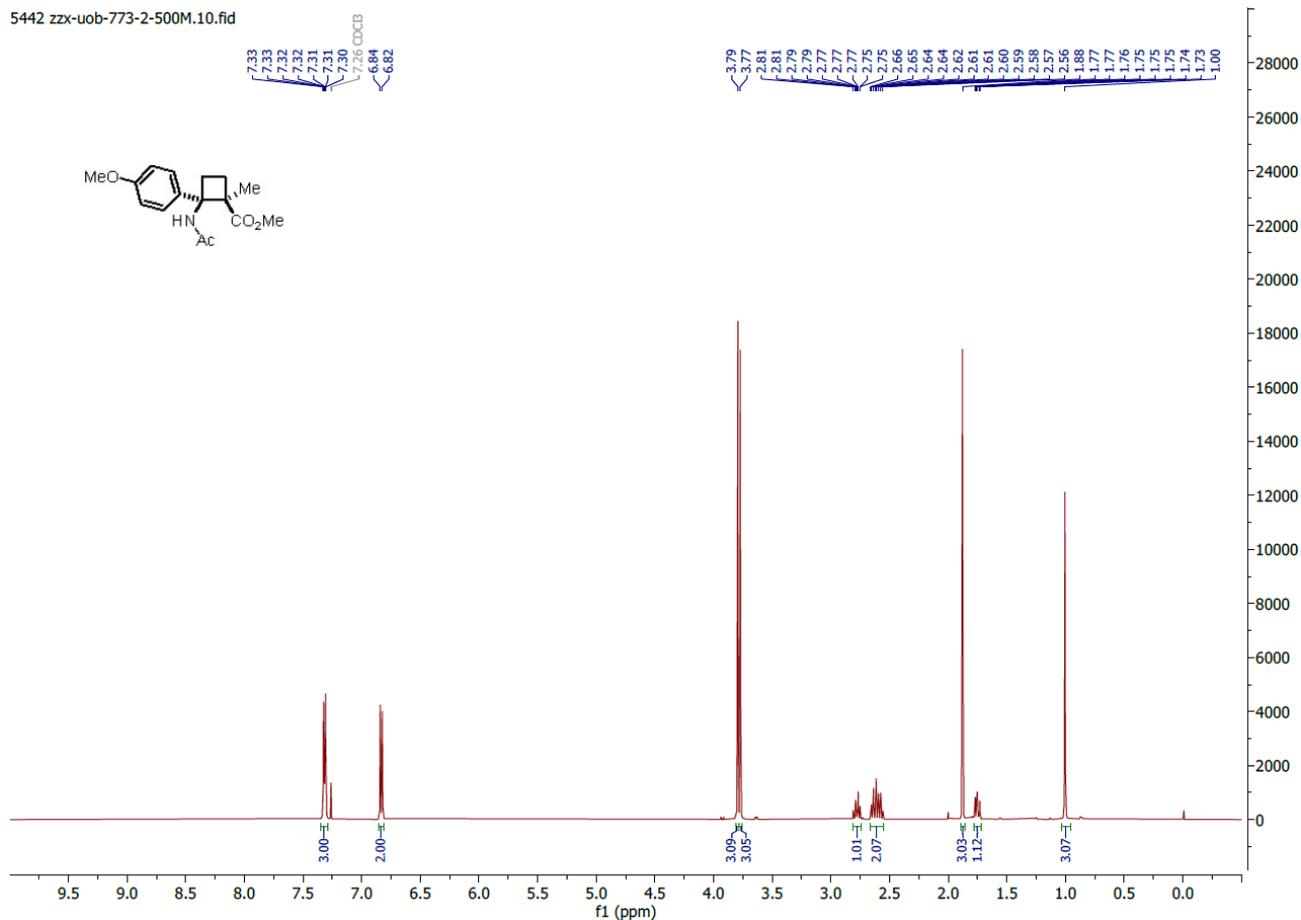


5444 zzx-uob-773-1-500M-sec.11.fid

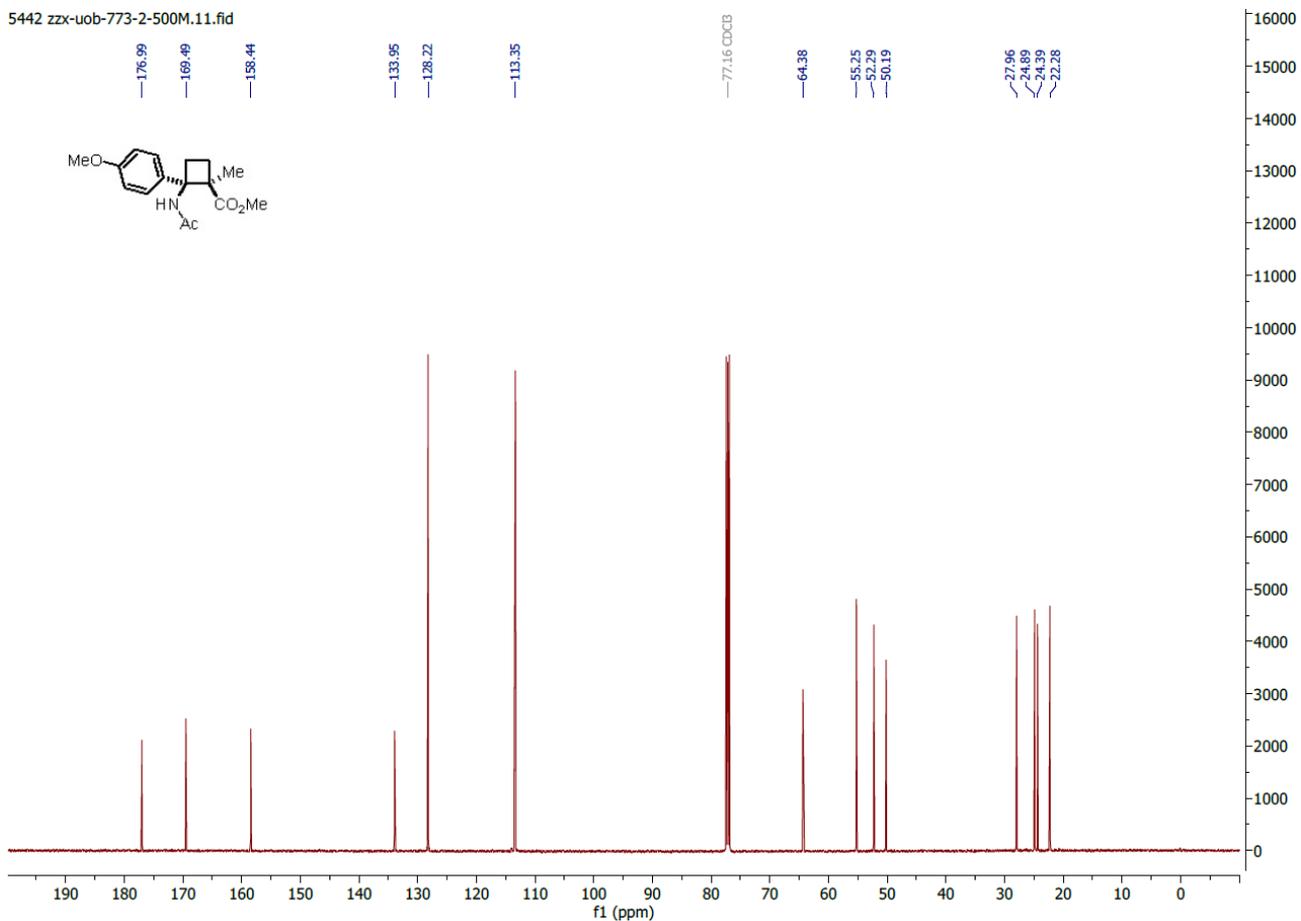


Compound 12'

5442 zzx-uob-773-2-500M.10.fid



5442 zzx-uob-773-2-500M.11.fid

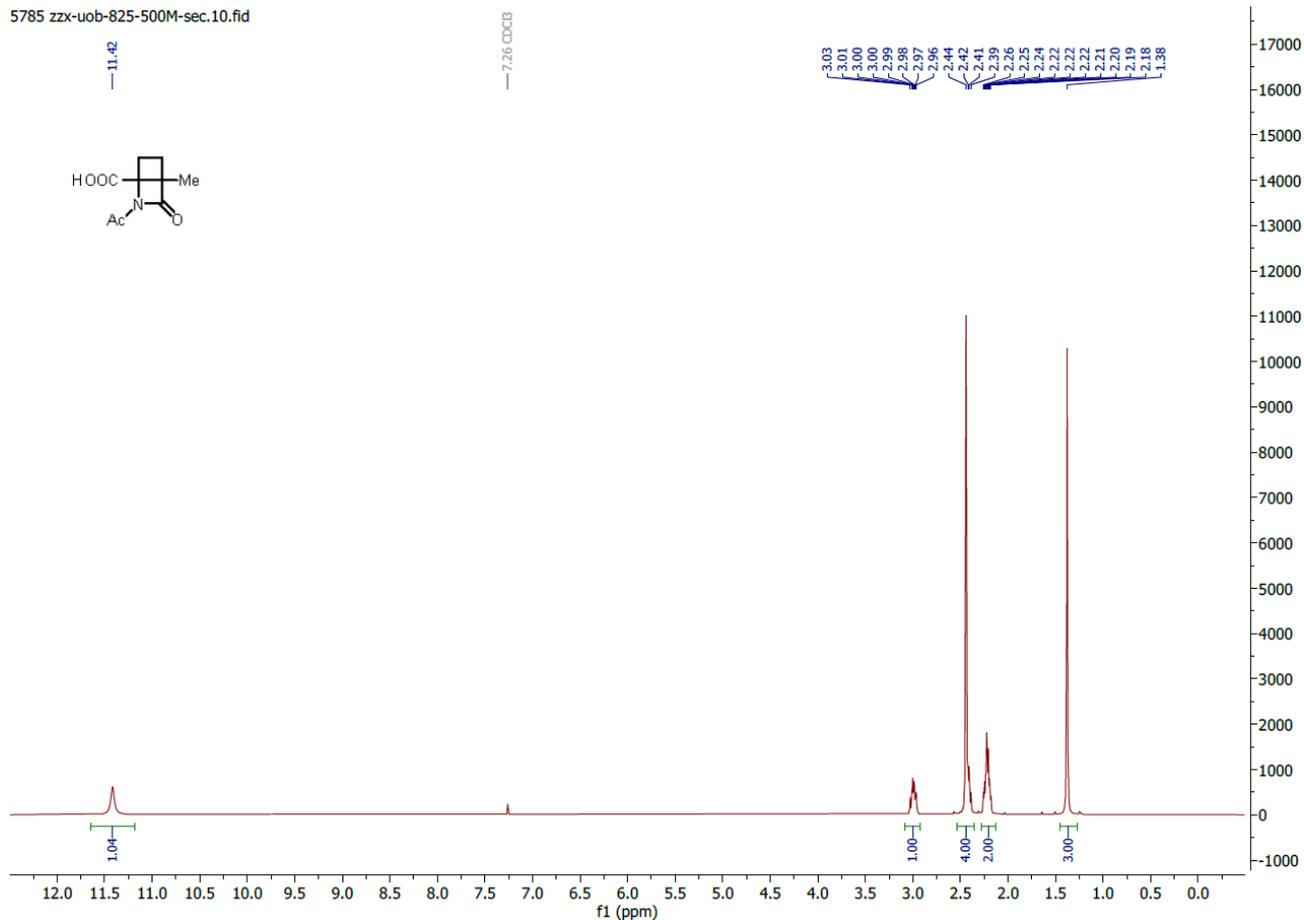
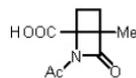


Compound 13

5785 zzx-uob-825-500M-sec.10.fid

11.42

7.26 CDCl₃



5785 zzx-uob-825-500M-sec.11.fid

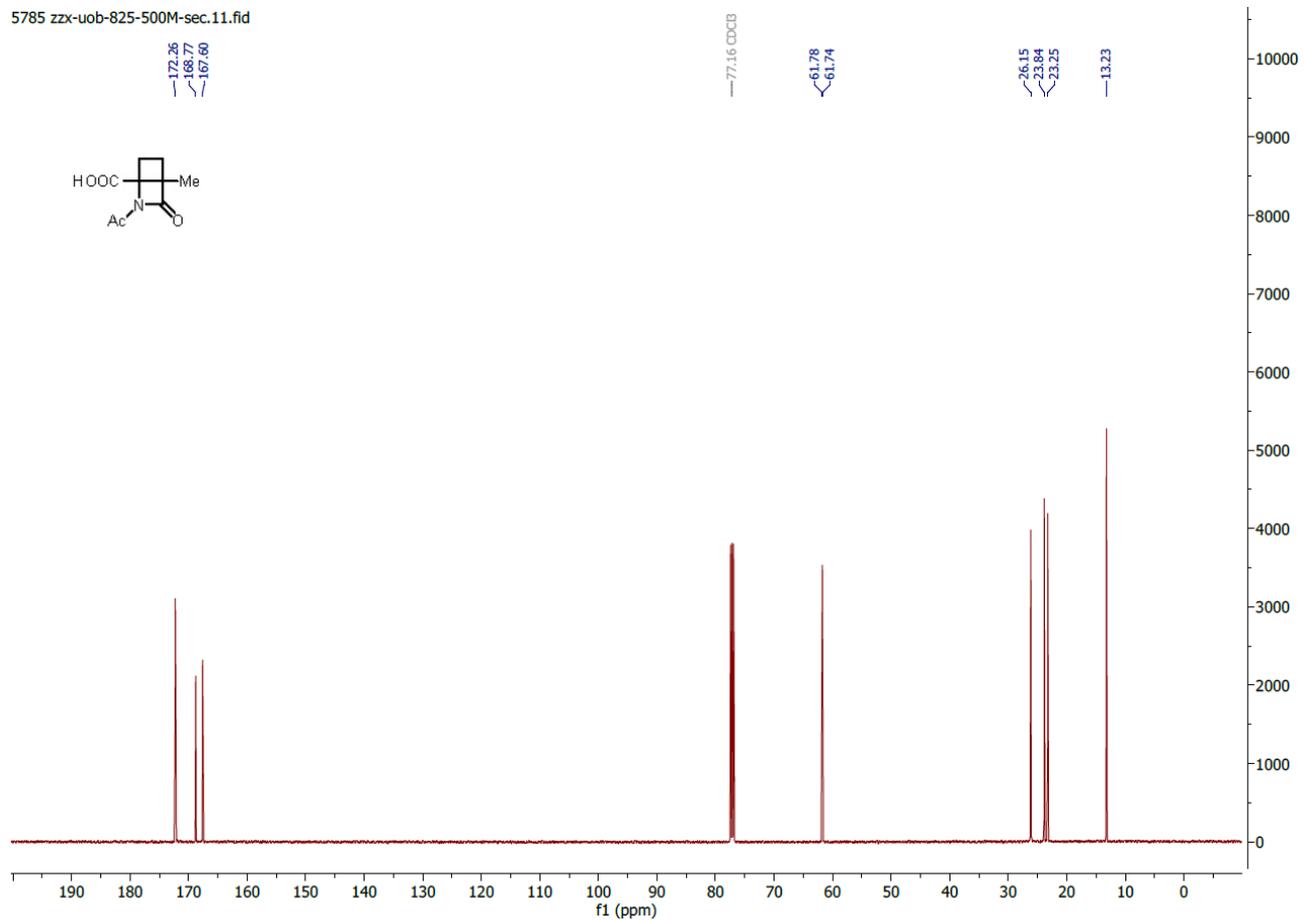
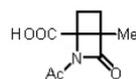
172.26, 168.77, 167.60

77.16 CDCl₃

61.78, 61.74

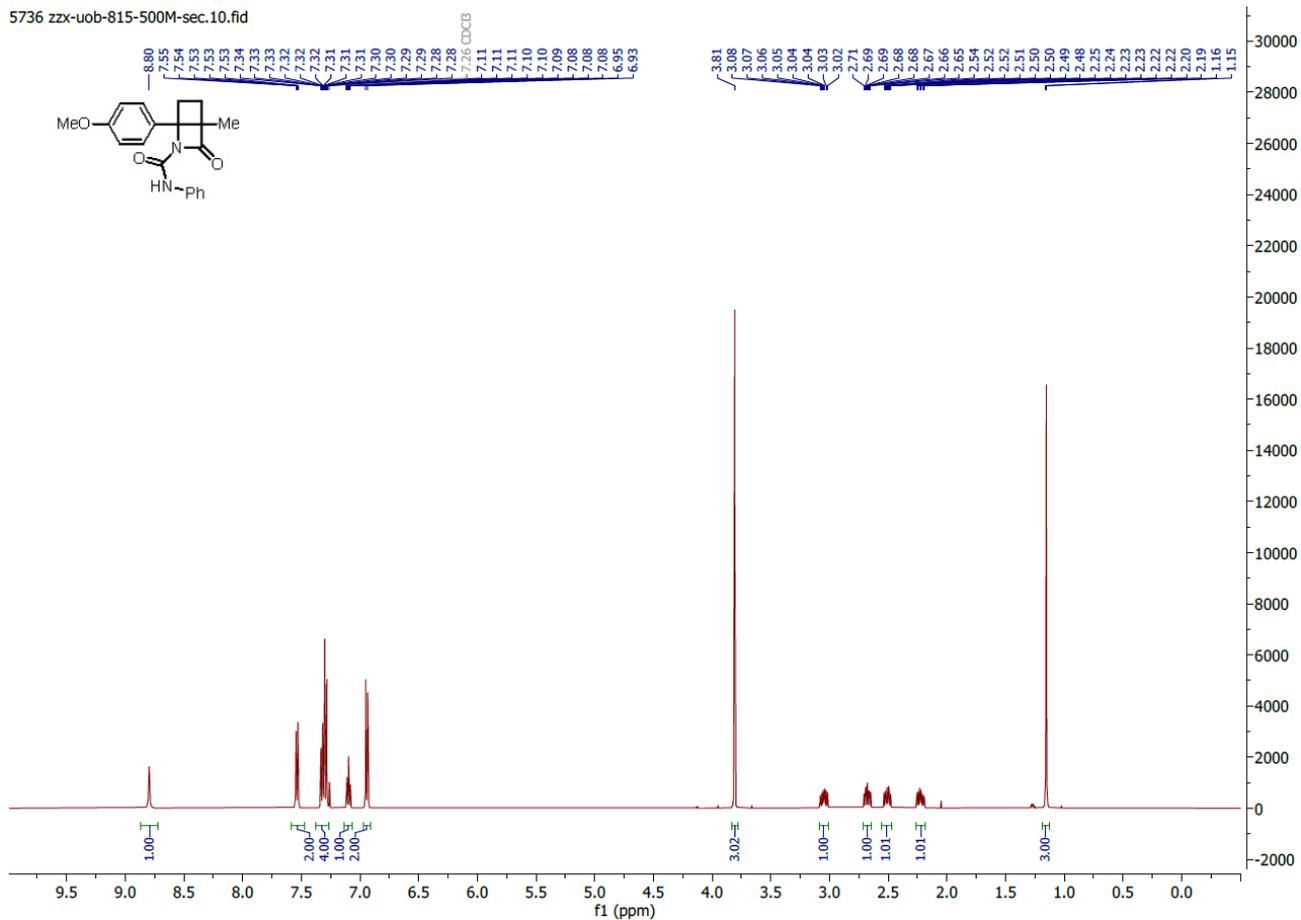
26.15, 25.84, 23.25

13.23

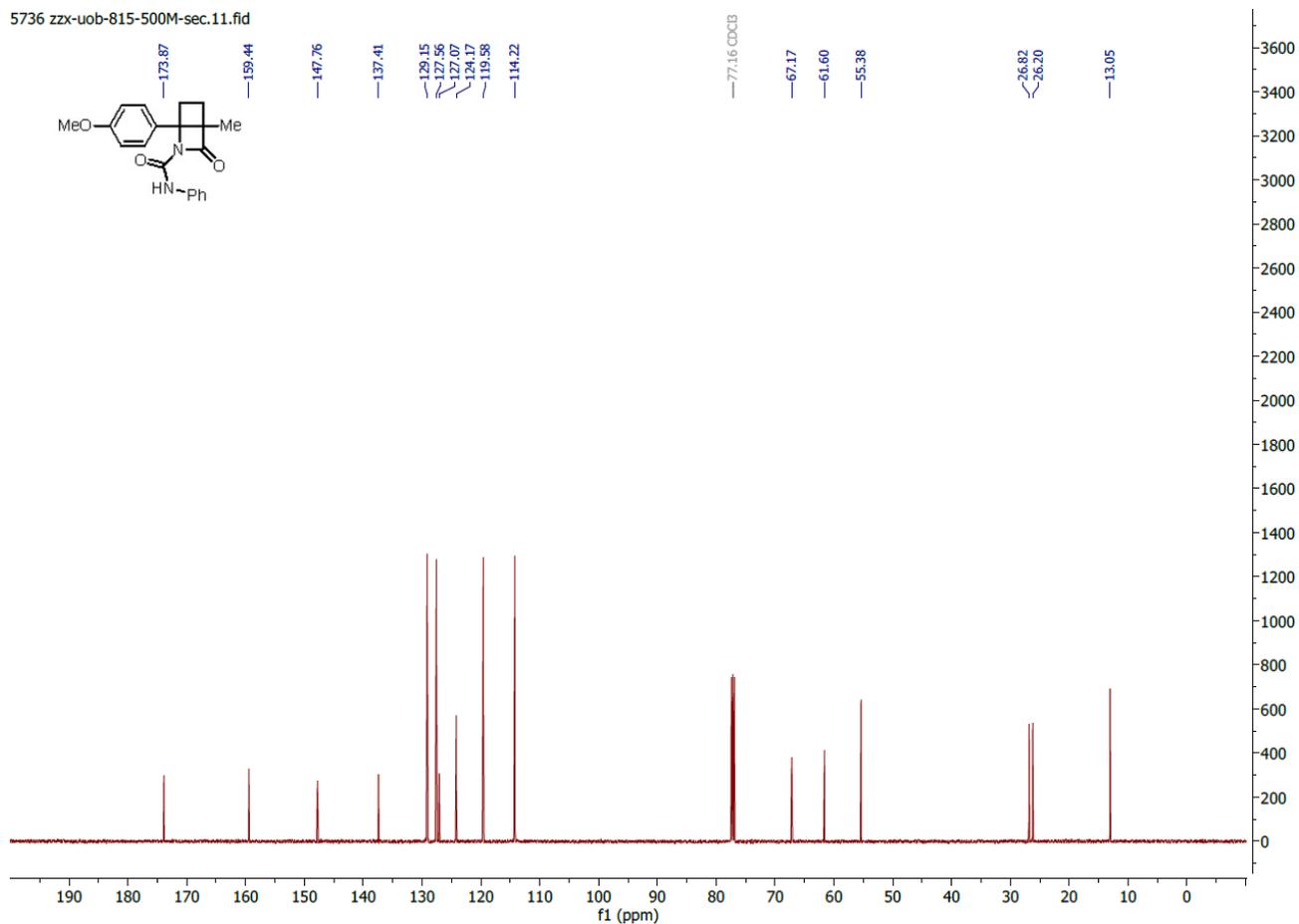


Compound 14

5736 zzx-uob-815-500M-sec.10.fid

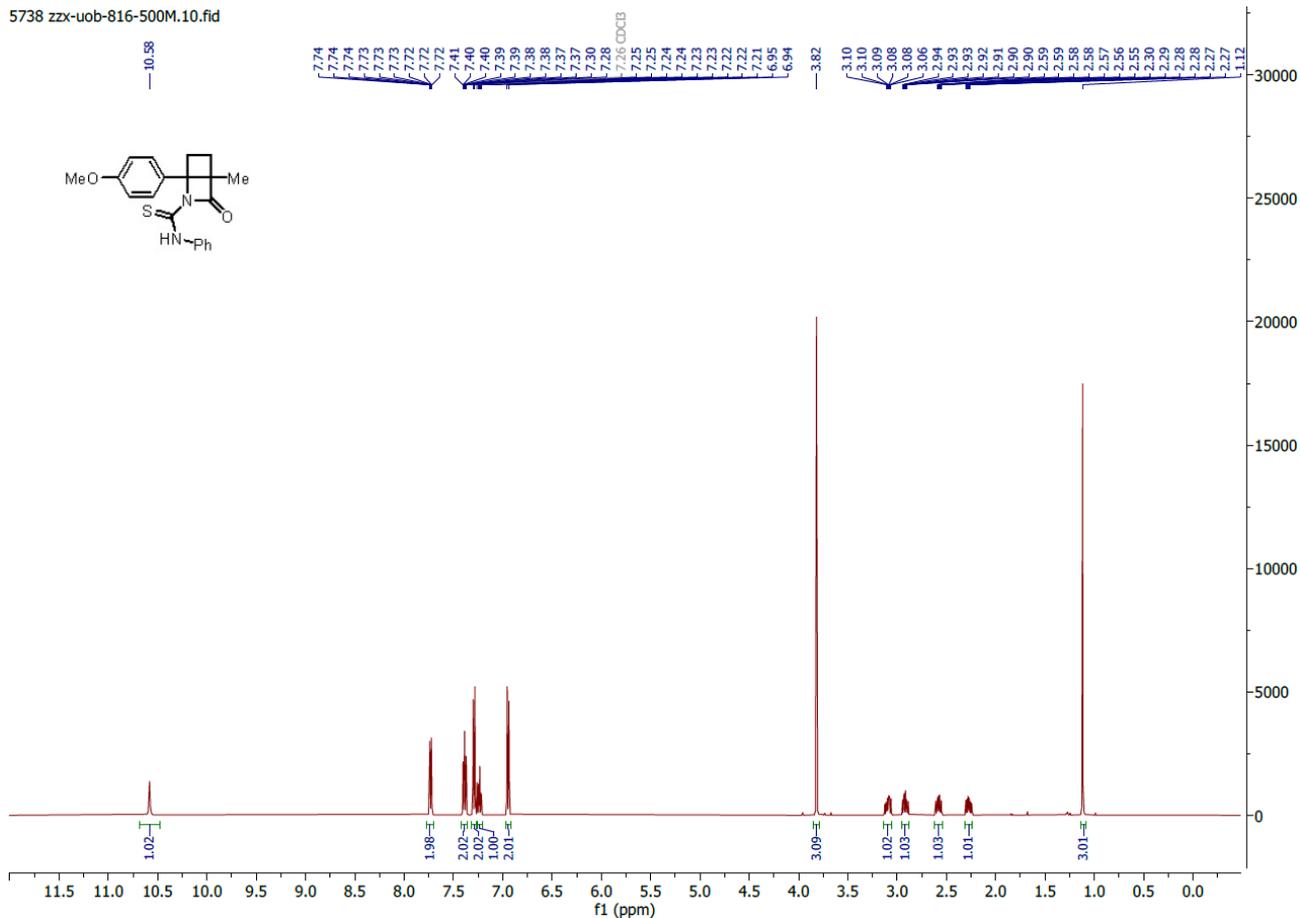
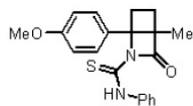


5736 zzx-uob-815-500M-sec.11.fid

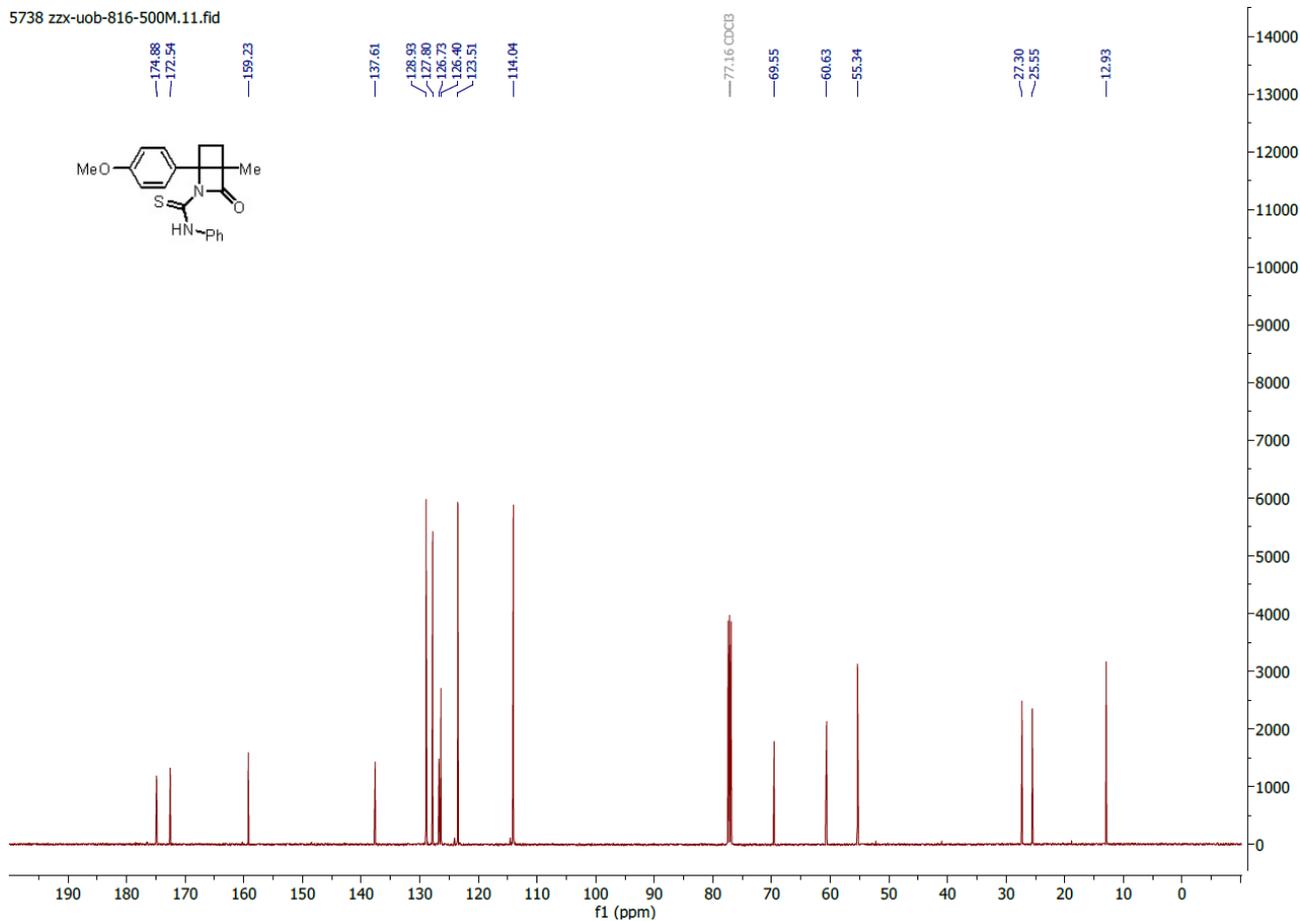
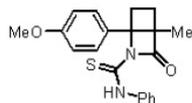


Compound 15

5738 zzx-uob-816-500M.10.fid

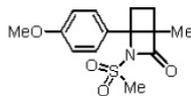


5738 zzx-uob-816-500M.11.fid



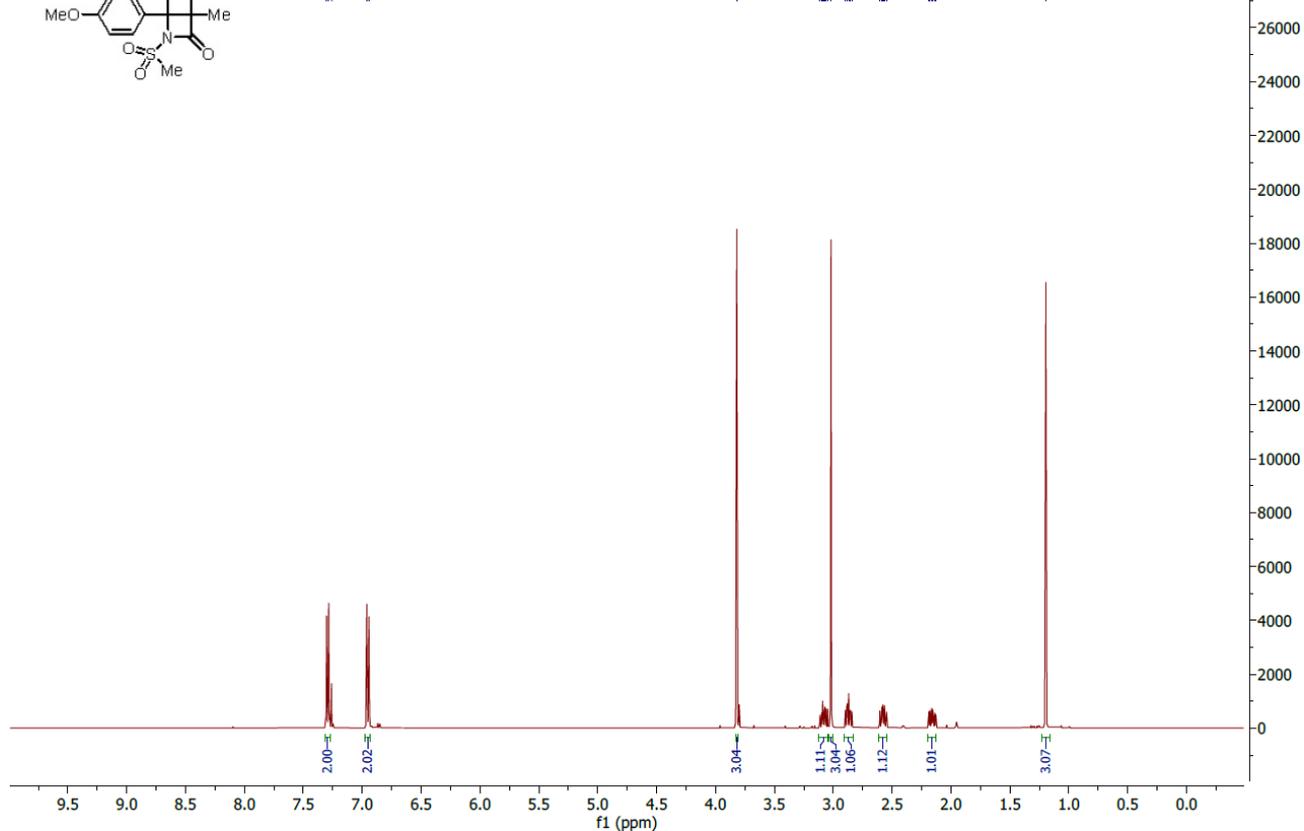
Compound 16

5740 zzx-uob-817-500M.10.fid

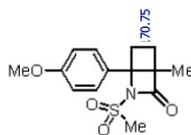


7.30
7.28
7.26 CDCl₃
6.96
6.94

3.82
3.11
3.10
3.09
3.08
3.07
3.06
3.05
3.02
2.99
2.98
2.88
2.87
2.86
2.85
2.84
2.60
2.59
2.59
2.58
2.57
2.56
2.55
2.19
2.18
2.17
2.16
2.16
2.13
2.13
1.20



5740 zzx-uob-817-500M.11.fid



170.75
159.91

128.16
126.32

114.40

77.16 CDCl₃

70.93

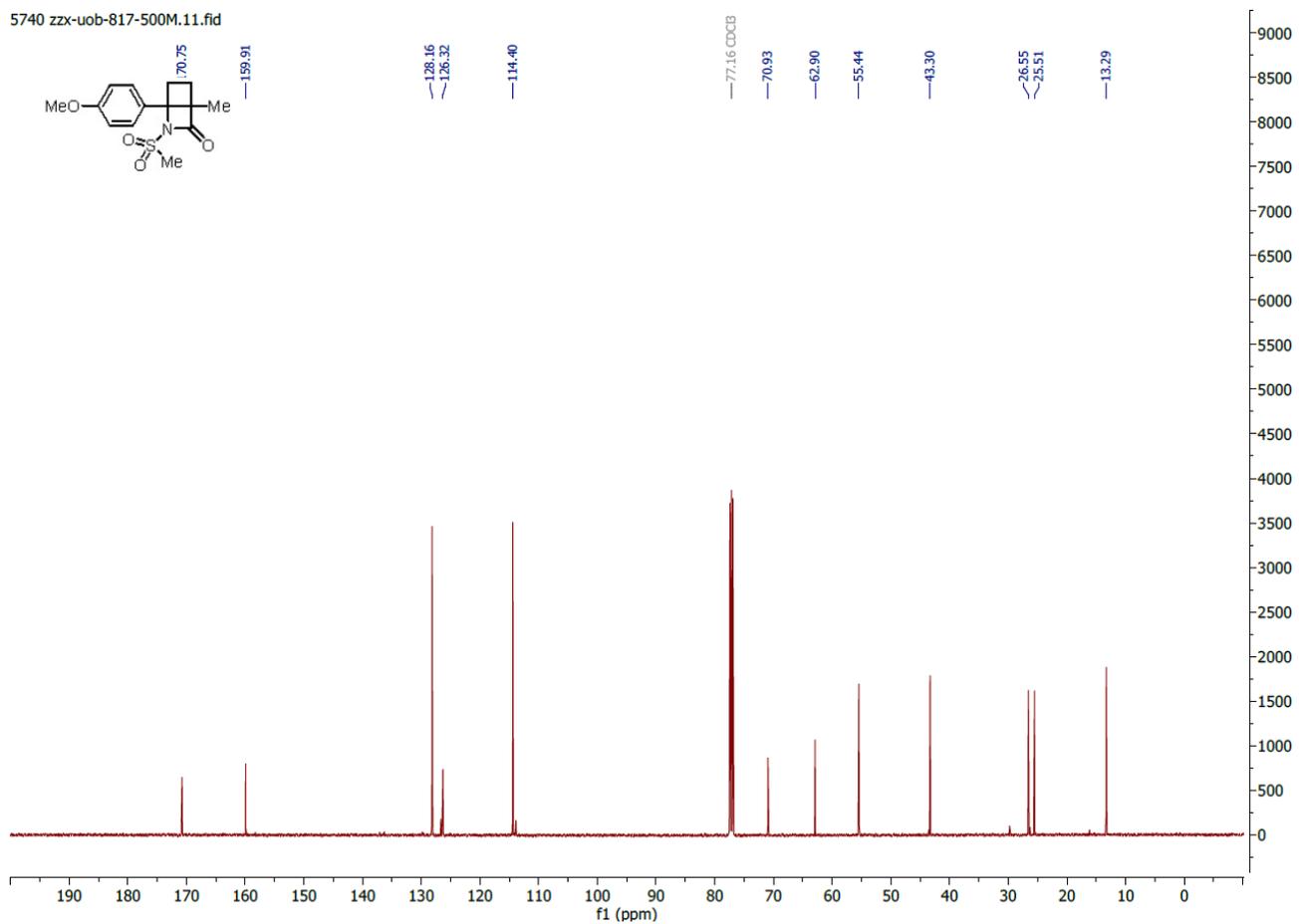
62.90

55.44

43.30

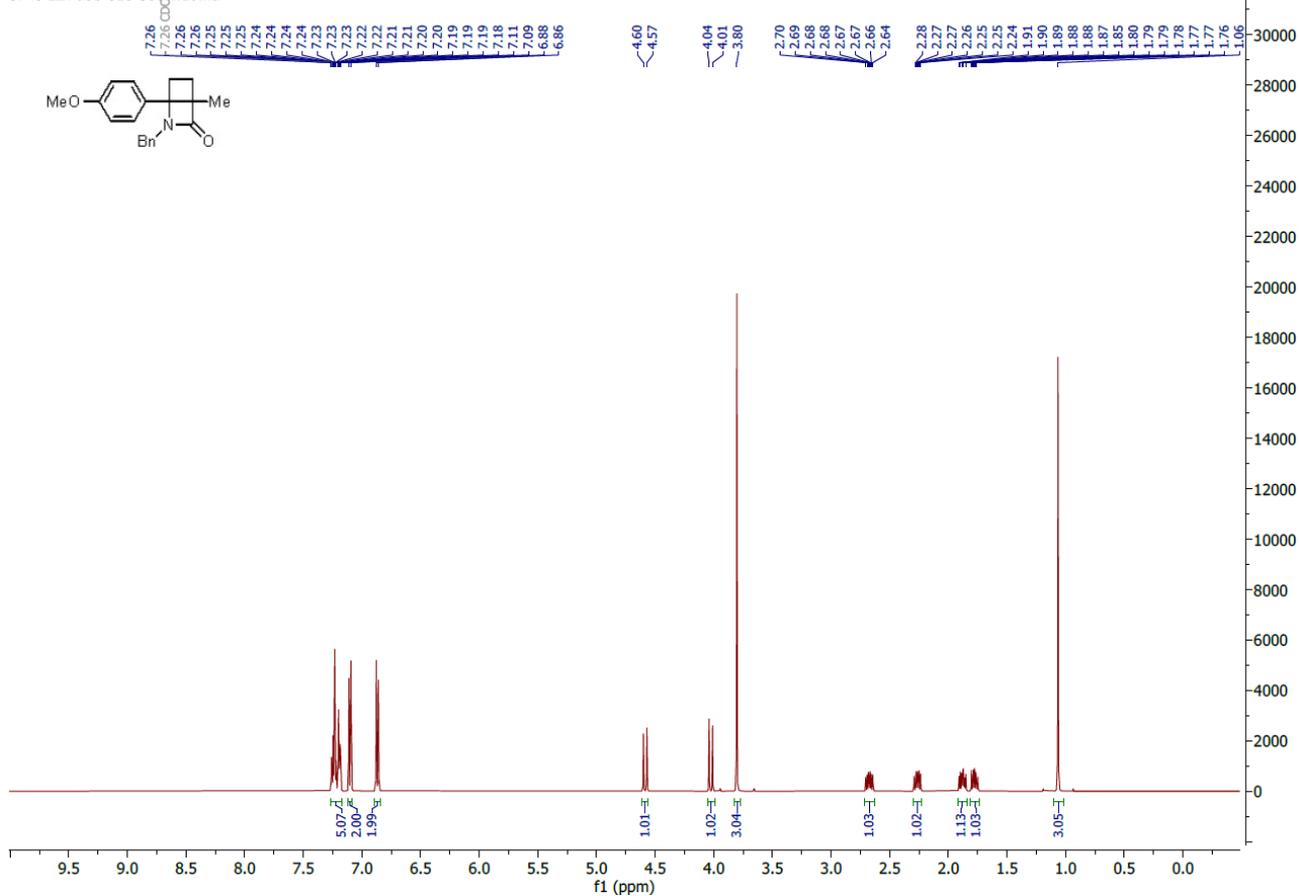
26.55
25.51

13.29

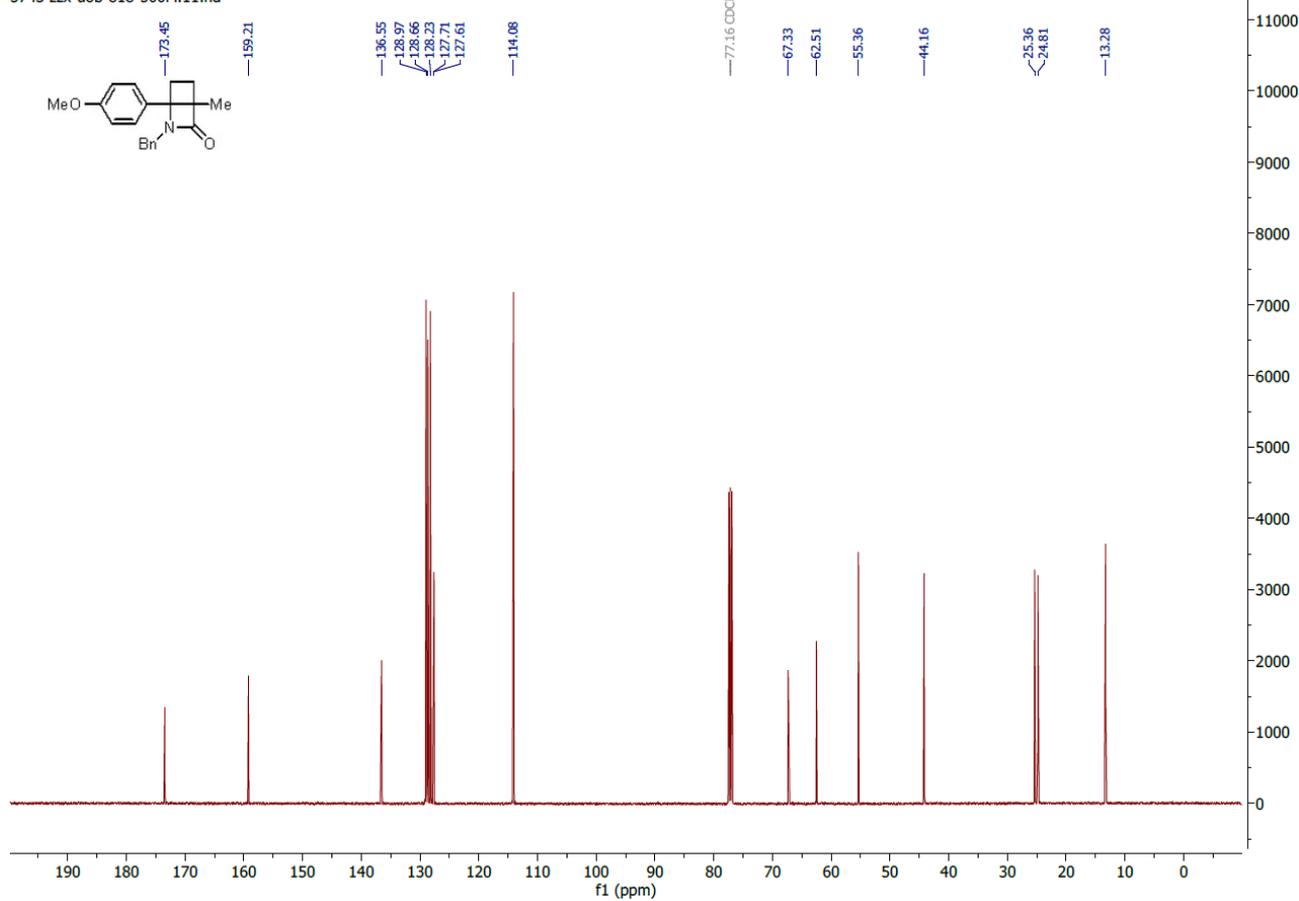


Compound 17

5743 zzx-uob-818-500M.10.fid

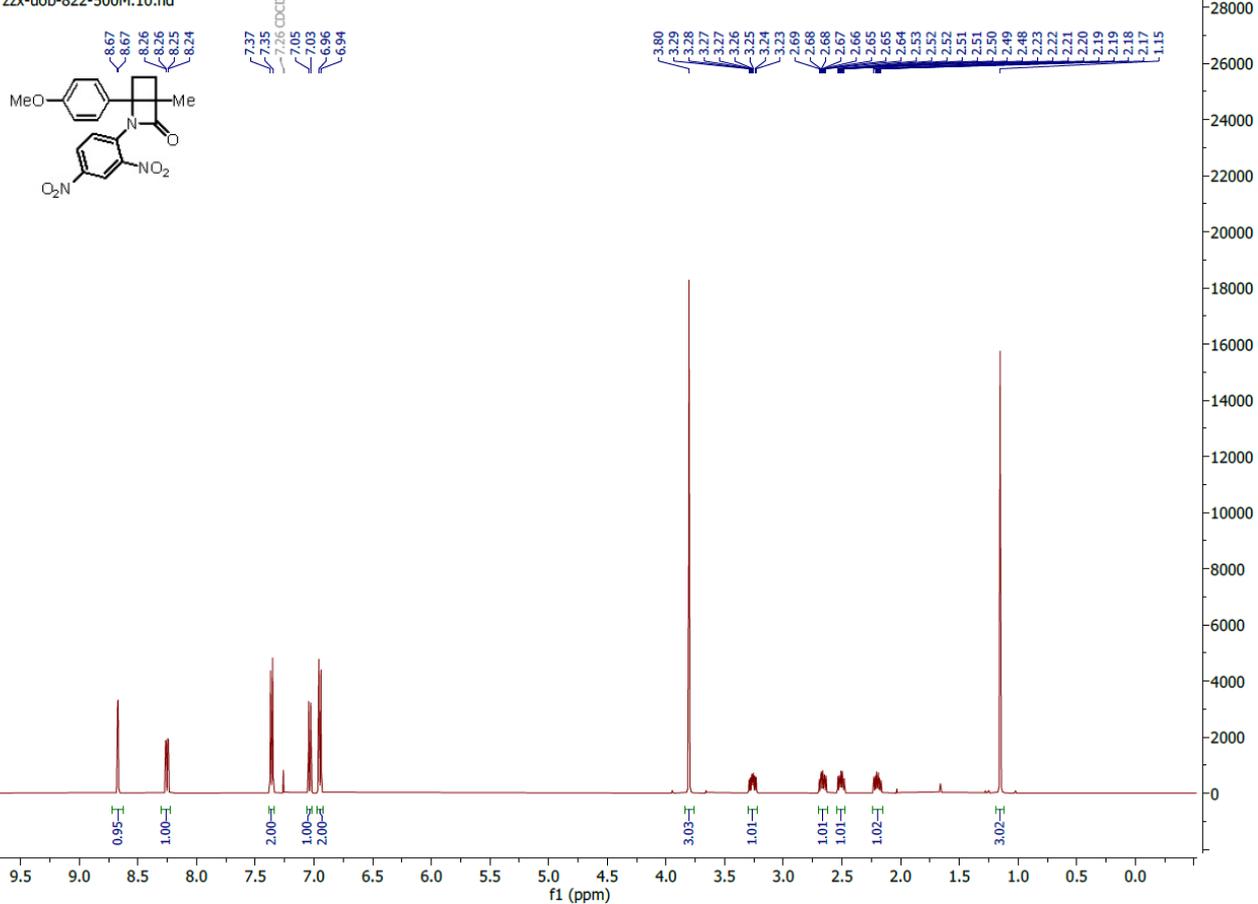


5743 zzx-uob-818-500M.11.fid

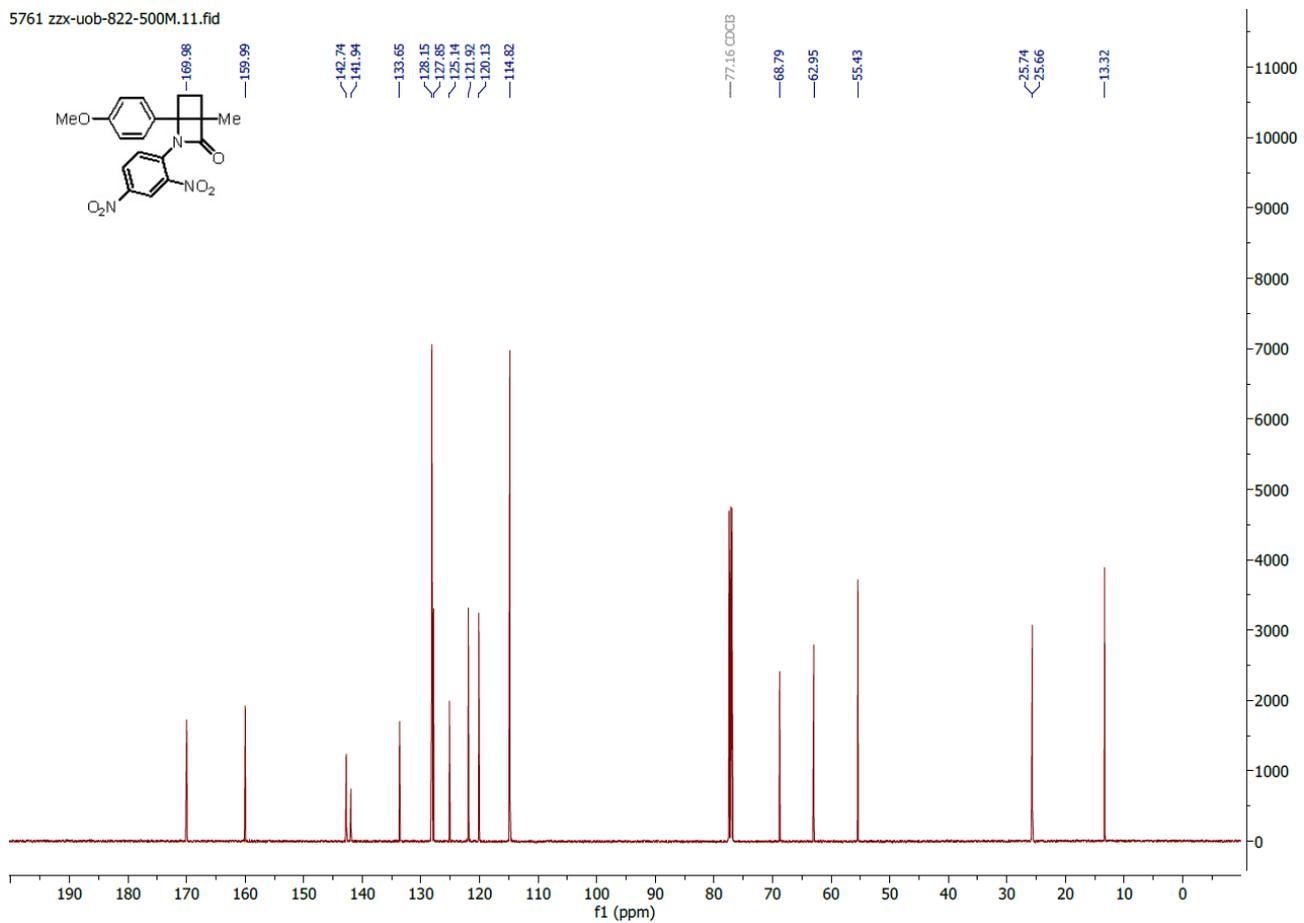


Compound 18

5761 zzx-uob-822-500M.10.fid

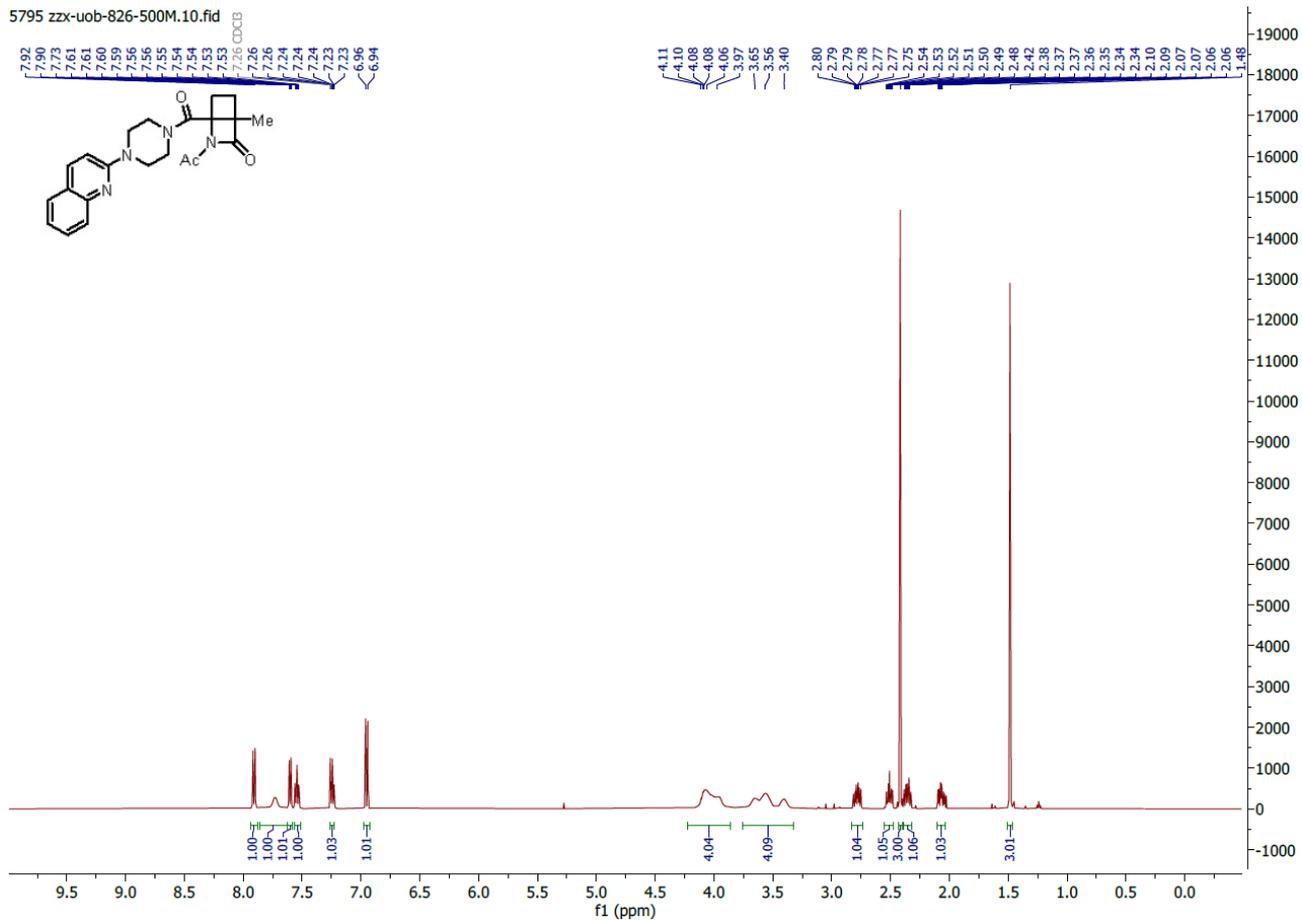


5761 zzx-uob-822-500M.11.fid

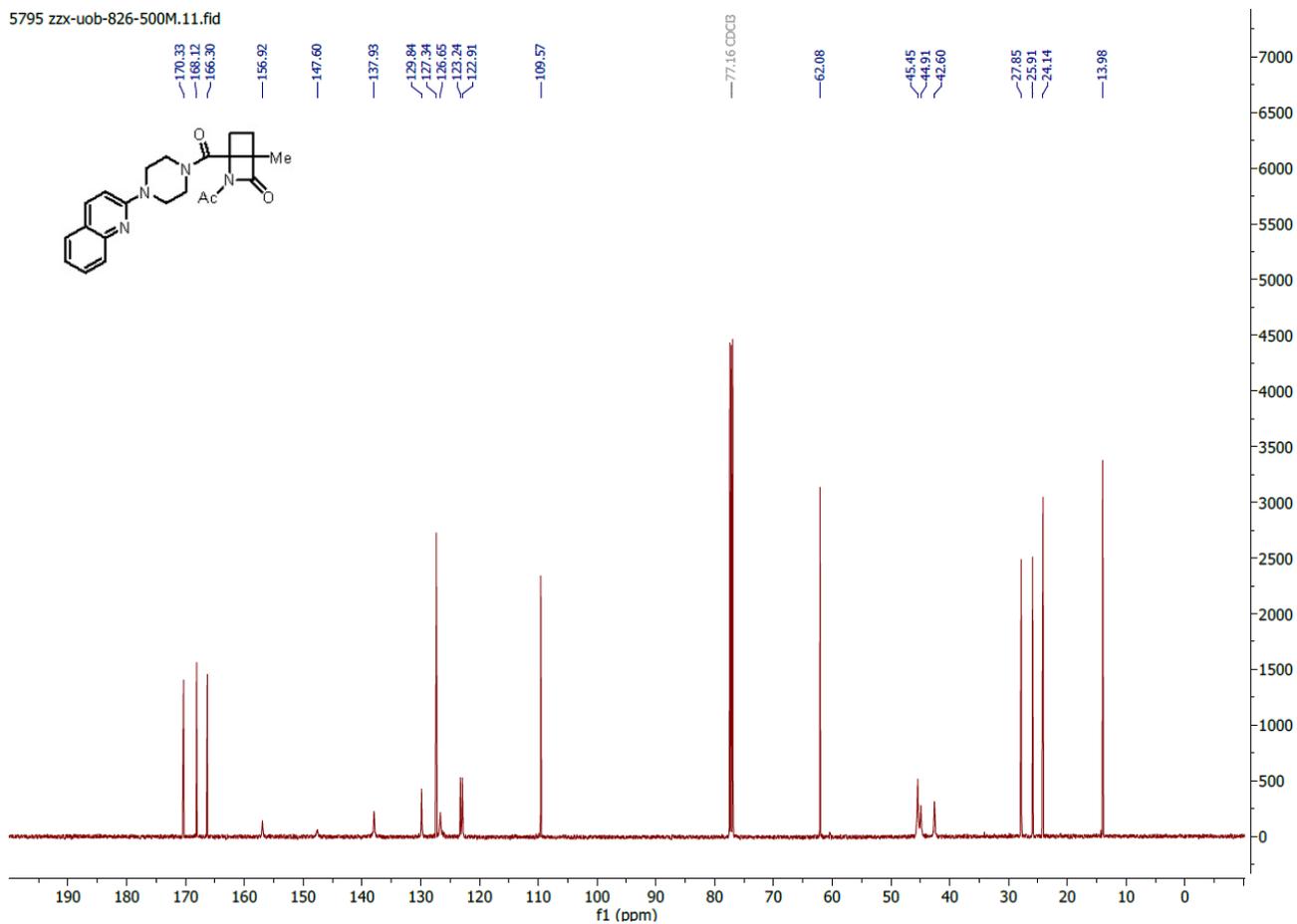


Compound 19

5795 zzx-uob-826-500M.10.fid

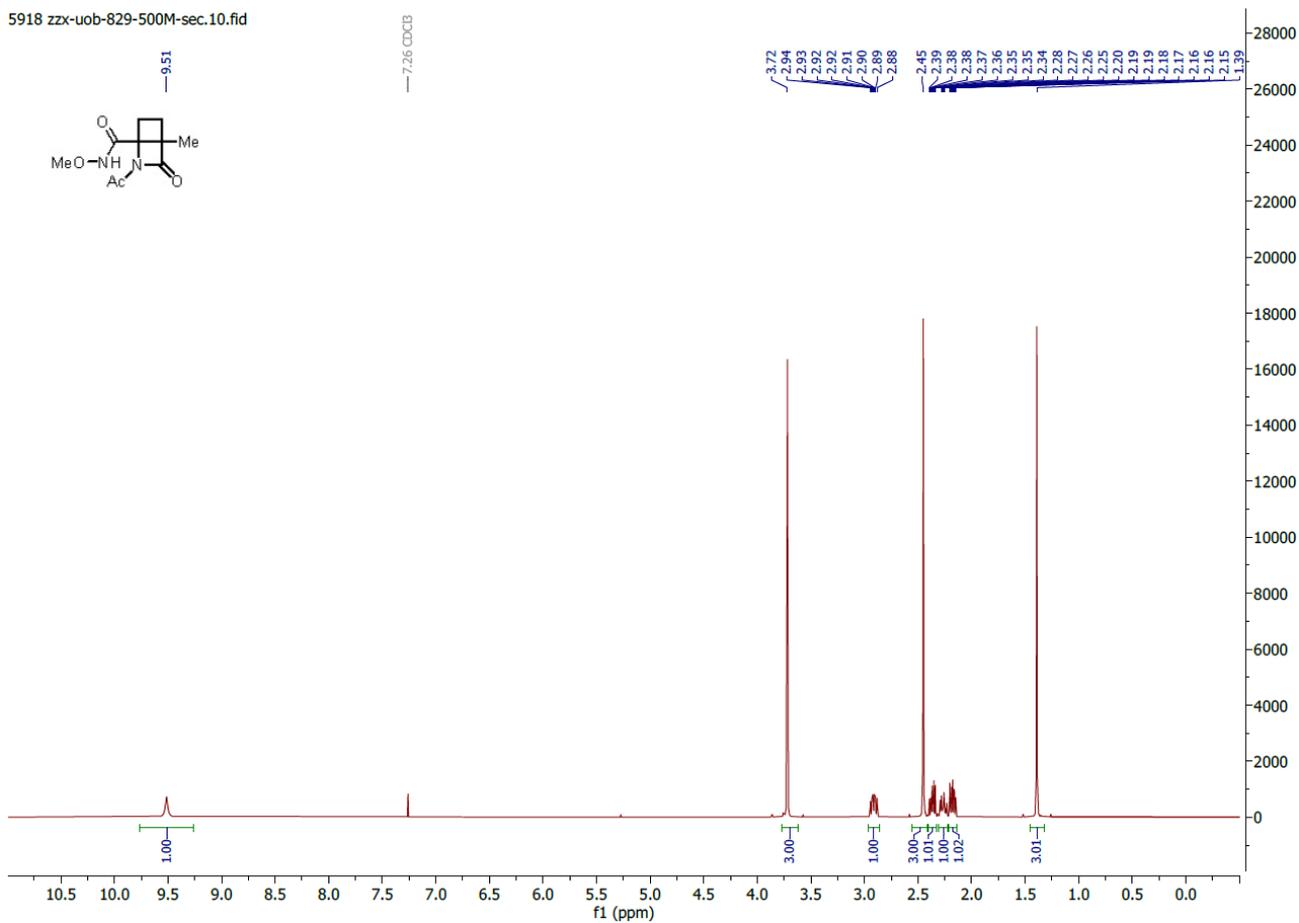


5795 zzx-uob-826-500M.11.fid

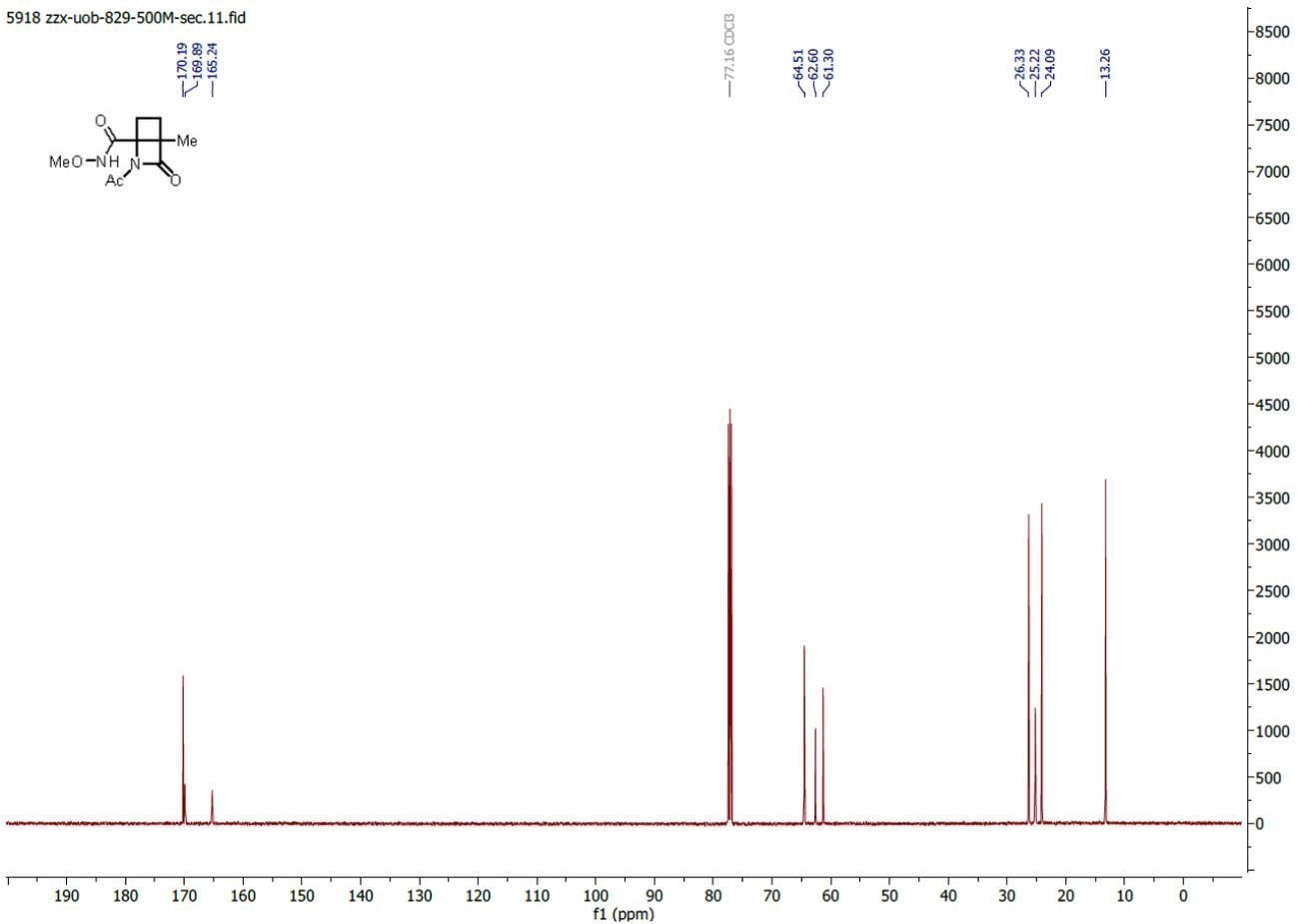


Compound 20

5918 zzx-uob-829-500M-sec.10.fid



5918 zzx-uob-829-500M-sec.11.fid



6. References

- 1 Kunishige, T. & Sawada, D. Efficient and practical synthesis of N-acetyl enamides from ketoximes by unique iron catalytic system. *Tetrahedron Lett.* **60**, 1562-1565 (2019).
- 2 Savarin, C. G. *et al.* Direct N-Acetyl Enamine Formation: Lithium Bromide Mediated Addition of Methyllithium to Nitriles. *Org. Lett.* **8**, 3903-3906 (2006).
- 3 Hu, N. *et al.* Synthesis of Chiral α -Amino Tertiary Boronic Esters by Enantioselective Hydroboration of α -Arylenamides. *J. Am. Chem. Soc.* **137**, 6746-6749 (2015).
- 4 Pak, H. K. *et al.* Synthesis of Enamides by Ruthenium-Catalysed Reaction of Alkyl Azides with Acid Anhydrides in Ionic Liquid. *ChemCatChem* **7**, 4030-4034 (2015).
- 5 Lyu, X., Zhang, J., Kim, D., Seo, S. & Chang, S. Merging NiH Catalysis and Inner-Sphere Metal-Nitrenoid Transfer for Hydroamidation of Alkynes. *J. Am. Chem. Soc.* **143**, 5867-5877 (2021).
- 6 Hu, X. *et al.* Regioselective Photocatalytic Dialkylation/Cyclisation Sequence of 3-Aza-1,5-dienes: Access to 3,4-Dialkylated 4-Pyrrolin-2-ones. *Adv. Synth. Catal.* **364**, 2163-2168 (2022).
- 7 Dai E, *et al.* Copper-Catalysed Cyanoisopropylalkenylation of N-Alkenyl-acrylamides to Give 1,3-Dihydropyrrol-2-ones. *Chin. J. Org. Chem.* **39**, 3524-3531 (2019).
- 8 Suslick, B. A. & Tilley, T. D. Mechanistic Interrogation of Alkyne Hydroarylations Catalysed by Highly Reduced, Single-Component Cobalt Complexes. *J. Am. Chem. Soc.* **142**, 11203-11218 (2020).
- 9 Berne, D., Poli, R., Caillol, S., Ladmiral, V. & Leclerc, E. Combination of Fluorine and Tertiary Amine Activation in Catalyst-Free Thia-Michael Covalent Adaptable Networks. *Macromolecules* **56**, 8260-8274 (2023).
- 10 Bajya, K. R., Maurya, S. K. & Selvakumar, S. Organophotocatalytic Regioselective Silylation/Germylation and Cascade Cyclization of N-Alkenyl α -CF₃ Acrylamides: Access to Densely Functionalized 4-Pyrrolin-2-ones. *Org. Lett.* **26**, 9269-9275 (2024).
- 11 Dong, Q.-R. *et al.* Synthesis of Unsymmetrical Disulfides via Photocatalytic Hydrodisulfuration. *ACS Catal.* **14**, 18237-18246 (2024).
- 12 Li, B. *et al.* Nickel-catalyzed asymmetric hydrogenation for the preparation of α -substituted propionic acids. *Nature Commun.* **15**, 5482 (2024).
- 13 Yang, Q.-L. *et al.* Electrochemistry-Enabled Ir-Catalyzed Vinylic C-H Functionalization. *J. Am. Chem. Soc.* **141**, 18970-18976 (2019).
- 14 Wang, K. *et al.* Highly Efficient and Regioselective Synthesis of N-Acyl Protected 2-Aminoacrylates through a PPh₃-Catalyzed Reaction of Alkyl Propiolates with Amides. *ChemistrySelect* **8**, e202300839 (2023).
- 15 Kobayashi, M., Suda, T., Noguchi, K. & Tanaka, K. Enantioselective Construction of Bridged Multicyclic Skeletons: Intermolecular [2+2+2] Cycloaddition/Intramolecular Diels-Alder Reaction Cascade. *Angew. Chem. Int. Ed.* **50**, 1664-1667 (2011).
- 16 Liu, F. *et al.* Regioselective Radical-Relay Sulfonylation/Cyclization Protocol to Sulfonylated Pyrrolidones under Transition-Metal-Free Conditions. *J. Org. Chem.* **87**, 6137-6145 (2022).
- 17 Huang, K. *et al.* Highly Enantioselective Borane Reduction of Heteroaryl and Heterocyclic Ketoxime Ethers Catalyzed by Novel Spiroborate Ester Derived from Diphenylvalinol: Application to the Synthesis of Nicotine Analogues. *J. Org. Chem.* **73**, 4017-4026 (2008).
- 18 Jiang, B., Zhao, M., Li, S.-S., Xu, Y.-H. & Loh, T.-P. Macrolide Synthesis through Intramolecular Oxidative Cross-Coupling of Alkenes. *Angew. Chem. Int. Ed.* **57**, 555-559 (2018).

- 19 Rao, M. L. N. & Giri, S. Pd-Catalysed Threefold Arylation of Baylis–Hillman Bromides and Acetates with Triarylbismuth Reagents. *Eur. J. Org. Chem.* **2012**, 4580-4589 (2012).
- 20 Brullo, C. *et al.* Design, synthesis, biological evaluation and structural characterisation of novel GEBR library PDE4D inhibitors. *Eur. J. Med. Chem.* **223**, 113638 (2021).
- 21 SAINT integration software version 8.39.0 (Bruker AXS Inc., Madison, WI).
- 22 SADABS version 2016/2 (Bruker AXS Inc., Madison, WI).
- 23 Dolomanov, O. V., Bourhis, L. J., Gildea, R. J., Howard, J. A. K. & Puschmann, H. OLEX2: a complete structure solution, refinement and analysis program. *J. Appl. Crystallogr.* **42**, 339-341 (2009).
- 24 *International Tables for Crystallography*. Vol. C (Wiley, 2016).
- 25 Parsons, S., Flack, H. D. & Wagner, T. Use of intensity quotients and differences in absolute structure refinement. *Acta Crystallographica Section B* **69**, 249-259 (2013).