

Supplementary data

Table S1. List of nonsynonymous SNPs identified in the fifth-passage strain

Chromosome	Gene	Nucleotide change	Amino acid change	Chromosome	Gene	Nucleotide change	Amino acid change
II	<i>POP4</i>	A ⁴¹ →C	Lys ¹⁴ →Thr		<i>IRC20</i>	G ¹⁴⁸ →A	Gly ⁵⁰ →Ser
	(<i>YBR257W</i>)	G ³¹⁴ →A	Arg ¹⁰⁵ →Lys		(<i>YLR247C</i>)		
V	<i>DDI1</i>	G ¹⁸² →A	Arg ⁶¹ →Lys		<i>CTS1</i>	C ¹³⁰⁰ →A	Leu ⁴³⁴ →Ile
	(<i>YER143W</i>)				(<i>YLR286C</i>)		
	<i>NEL1</i>	G ¹²³⁴ →A	Gly ⁴¹² →Arg	XII	<i>YLR287C</i>	G ¹⁰⁰¹ →A	Cys ³³⁴ →Tyr
	(<i>YHR035W</i>)						
	<i>BRL1</i>	G ⁷⁸⁸ →A	Arg ²⁶³ →Lys		<i>MEC3</i>	G ⁵²⁶ →A	Asp ¹⁷⁶ →Asn
	(<i>YHR036W</i>)				(<i>YLR288C</i>)		
VIII	<i>SMF2</i>	T ³⁸⁸ →C	Leu ¹³⁰ →Phe		<i>YLR297W</i>	G ¹⁸⁵ →C	Arg ⁶² →Pro
	(<i>YHR050W</i>)						
	<i>RRP3</i>	G ⁶⁰⁷ →A	Val ²⁰³ →Ile		<i>IMH1</i>	C ²⁴⁸⁶ →G	Ala ⁸²⁹ →Gly
	(<i>YHR065C</i>)				(<i>YLR309C</i>)		
	<i>SPL2</i>	C ⁴ →G	Arg ² →Gly	XIII	<i>YML131W</i>	G ⁸²⁵ →T	Glu ²⁷⁵ →Asp
	(<i>YHR136C</i>)						
	<i>YIL092W</i>	G ²⁶⁴ →A	Met ⁸⁸ →Ile		<i>SWT21</i>	G ⁵⁸⁰ →A	Val ¹⁹⁴ →Met
		G ¹⁰⁸⁶ →A	Met ³⁶² →Ile		(<i>YNL187W</i>)		
IX	<i>FYV10</i>	G ⁴⁶⁵ →C	Lys ¹⁵⁵ →Asn	XIV	<i>CNM67</i>	G ³³⁸ →A	Gly ¹¹³ →Asp
	(<i>YIL097W</i>)				(<i>YNL225C</i>)		
	<i>CSM2</i>	G ⁵³⁰ →A	Arg ¹⁷⁷ →Lys		<i>JJJ1</i>	C ¹⁶²¹ →T	Pro ⁵⁴¹ →Ser
	(<i>YIL132C</i>)				(<i>YNL227C</i>)		
	<i>SLN1</i>	GG ²²¹⁷ →AA	Asp ⁷⁴⁰ →Asn	XV	<i>TAT2</i>	C ¹⁴⁸ →A	His ⁵⁰ →Asn

X	<i>(YIL147C)</i>	G ³⁴¹⁶ →A	Ser ¹¹³⁹ →Asn
	<i>MLP2</i>	A ⁷⁴ →G	Lys ²⁵ →Arg
	<i>(YIL149C)</i>	G ²⁴²⁶ →A	Gly ⁸⁰⁹ →Asp
	<i>MDE1</i>	G ³⁰¹ →A	Asp ¹⁰¹ →Asn
	<i>(YJR024C)</i>	G ³⁵⁵ →C	Val ¹¹⁹ →Leu
	<i>DAN1</i>	C ²¹⁰ →G	Asn ⁷⁰ →Lys
XII	<i>GRC3</i>	G ⁵¹¹ →A	Asp ¹⁷¹ →Asn
	<i>(YLL035W)</i>		
	<i>YLR230W</i>	C ²²⁶ →G	Pro ⁷⁶ →Ala

<i>(YOL020W)</i>	A ¹⁵²⁶ →G	Asp ⁵⁰⁹ →Gly
<i>MDM38</i>	G ¹³⁶⁰ →A	Glu ⁴⁵⁴ →Lys
<i>(YOL027C)</i>		
<i>YOL036W</i>	C ⁸⁵⁵ →G	Asn ²⁸⁵ →Lys
<i>INP54</i>	A ³⁴⁵ →T	Lys ¹¹⁵ →Asn
<i>(YOL065C)</i>		
<i>ATG34</i>	G ¹¹⁵³ →A	Val ³⁸⁵ →Met
<i>(YOL083W)</i>		
<i>PHM7</i>	G ¹¹²⁶ →A	Gly ³⁷⁶ →Ser
<i>(YOL084W)</i>		

Table S2. *Saccharomyces cerevisiae* strains and plasmids used in this study

Name	Description	Reference
Strains		
D452-2	<i>MATa, leu2, his3, ura3 and can1</i>	[1]
POP4_1*	D452-2 with the <i>POP4</i> ^{K14T} mutation	In this study
POP4_2*	D452-2 with the <i>POP4</i> ^{R105K} mutation	In this study
DDI1*	D452-2 with the <i>DDI1</i> ^{R61K} mutation	In this study
RRP3*	D452-2 with the <i>RRP3</i> ^{V203I} mutation	In this study
MDE1*	D452-2 with the <i>MDE1</i> ^{D101N, V119L} mutation	In this study
GRC3*	D452-2 with the <i>GRC3</i> ^{D171N} mutation	In this study
SWT21*	D452-2 with the <i>SWT21</i> ^{V194M} mutation	In this study
JJJ1*	D452-2 with the <i>JJJ1</i> ^{P541S} mutation	In this study
TAT2_1*	D452-2 with the <i>TAT2</i> ^{H50N} mutation	In this study
TAT2_2*	D452-2 with the <i>TAT2</i> ^{D509G} mutation	In this study
ATG34*	D452-2 with the <i>ATG34</i> ^{V385M} mutation	In this study
Plasmids		
pCas9_AUR	Aur ^R , p414- <i>TEF1p-Cas9-CYC1t</i> , modified Cas9 expression plasmid, Amp ^R	[2]
pgRNA- <i>TRP1</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gTRP1-SUP4t</i> , Amp ^R	[3]
pgRNA- <i>POP4_1</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gPOP4_1-SUP4t</i> , Amp ^R	In this study
pgRNA- <i>POP4_2</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gPOP4_2-SUP4t</i> , Amp ^R	In this study
pgRNA- <i>DDI1</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gDDI1-SUP4t</i> , Amp ^R	In this study
pgRNA- <i>RRP3</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gRRP3-SUP4t</i> , Amp ^R	In this study
pgRNA- <i>MDE1</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gMDE1-SUP4t</i> , Amp ^R	In this study
pgRNA- <i>GRC3</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gGRC3-SUP4t</i> , Amp ^R	In this study
pgRNA- <i>SWT21</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gSWT21-SUP4t</i> , Amp ^R	In this study

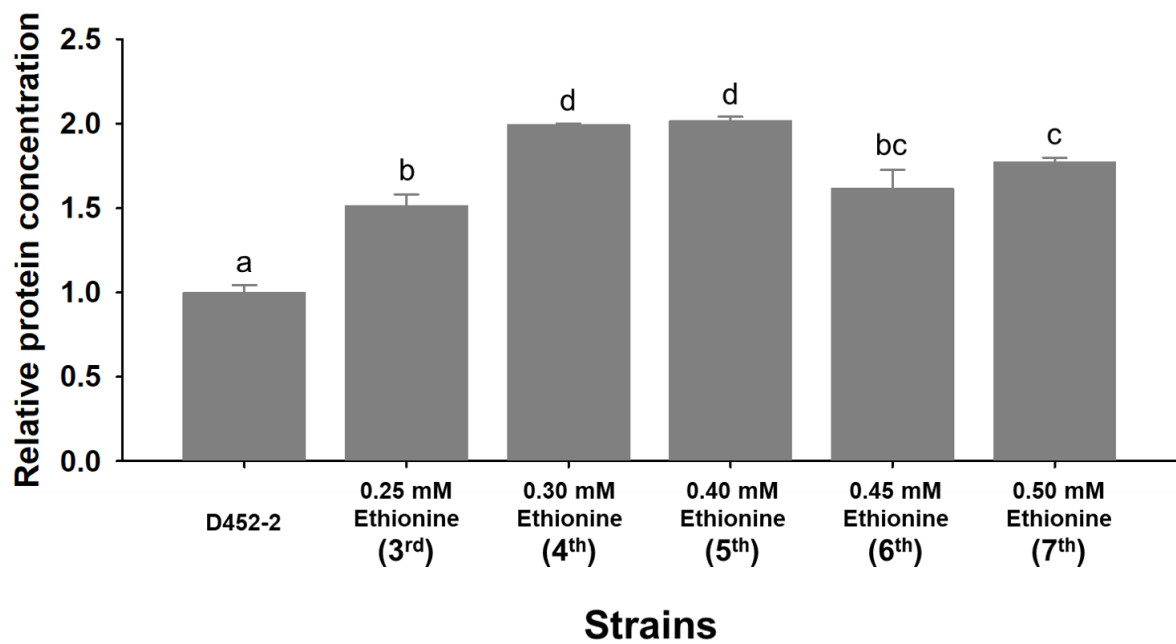
pgRNA- <i>JJJ1</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gJJJ1-SUP4t</i> , Amp ^R	In this study
pgRNA- <i>TAT2_1</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gTAT2_1-SUP4t</i> , Amp ^R	In this study
pgRNA- <i>TAT2_2</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gTAT2_2-SUP4t</i> , Amp ^R	In this study
pgRNA- <i>ATG34</i> -HYB	Hyg ^R , 2 μ origin, <i>SNR52p-gATG34-SUP4t</i> , Amp ^R	In this study

Table S3. List of primers used in this study

Name	Description	Amplified gene	Description
TH69	TCTTTTCAAGAAATGCCTGGGTTTTAGAGCTAGAAATAGCAAGTTAAAATA AGG	pgRNA- <i>TRP1</i> -HYB	To construct pgRNA- <i>POP4_1</i> -HYB
TH70	AACCCAGGCATTTCTTGAAAAGAGATCATTTATCTTTCACTGCGGA		
TH71	CGAAAATAAAATTACTGATAGTTTTAGAGCTAGAAATAGCAAGTTAAAATAA GG	pgRNA- <i>TRP1</i> -HYB	To construct pgRNA- <i>POP4_2</i> -HYB
TH72	AACATATCAGTAATTTTATTTTCGGATCATTTATCTTTCACTGCGGA		
TH152	CCTAATCAATAATAAGTCATGTTTTAGAGCTAGAAATAGCAAGTTAAAATAA GG	pgRNA- <i>TRP1</i> -HYB	To construct pgRNA- <i>DDI1</i> -HYB
TH153	AACATGACTTATTATTGATTAGGGATCATTTATCTTTCACTGCGGA		
TH75	AAATGGTCCATTAATCTACCGTTTTAGAGCTAGAAATAGCAAGTTAAAATAA GG	pgRNA- <i>TRP1</i> -HYB	To construct pgRNA- <i>RRP3</i> -HYB
TH76	AACGGTAGATTAATGGACCATTTGATCATTTATCTTTCACTGCGGA		
TH91	AGGAGCATATCACAGCATTTGTTTTAGAGCTAGAAATAGCAAGTTAAAATAA GG	pgRNA- <i>TRP1</i> -HYB	To construct pgRNA- <i>MDE1</i> -HYB
TH92	AACAAATGCTGTGATATGCTCCTGATCATTTATCTTTCACTGCGGA		
TH171	TAGAAGCAGGTCATTTATATGTTTTAGAGCTAGAAATAGCAAGTTAAAATAA GG	pgRNA- <i>TRP1</i> -HYB	To construct pgRNA- <i>GRC3</i> -HYB
TH172	AACATATAAATGACCTGCTTCTAGATCATTTATCTTTCACTGCGGA		
TH146	AGTGCACACCTCGATCGCGGGTTTTAGAGCTAGAAATAGCAAGTTAAAATA AGG	pgRNA- <i>TRP1</i> -HYB	To construct pgRNA- <i>JJJ1</i> -HYB
TH147	AACCCGCGATCGAGGTGTGCACTGATCATTTATCTTTCACTGCGGA		
TH83	TTATGCGAATGAAGTGTTTCGTTTTAGAGCTAGAAATAGCAAGTTAAAATAA GG	pgRNA- <i>TRP1</i> -HYB	To construct pgRNA- <i>SWT21</i> -HYB
TH84	AACGAAACACTTCATTCGCATAAGATCATTTATCTTTCACTGCGGA		
TH85	TCATCTGTCTCGCCTAGAAAGTTTTAGAGCTAGAAATAGCAAGTTAAAATAA GG	pgRNA- <i>TRP1</i> -HYB	To construct pgRNA- <i>ATG34</i> -HYB
TH86	AACCTTCTAGGCGAGACAGATGAGATCATTTATCTTTCACTGCGGA		
TH87	ATGACCACCAACACGGCAAGGTTTTAGAGCTAGAAATAGCAAGTTAAAATA AGG	pgRNA- <i>TRP1</i> -HYB	To construct pgRNA- <i>TAT2_1</i> -HYB
TH88	AACCTTGCCGTGTTGGTGGTCATGATCATTTATCTTTCACTGCGGA		
TH89	CCGCTGGTCCAGCCACCAATGTTTTAGAGCTAGAAATAGCAAGTTAAAATA AGG	pgRNA- <i>TRP1</i> -HYB	To construct pgRNA- <i>TAT2_2</i> -HYB
TH90	AACATTGGTGGCTGGACCAGCGGGATCATTTATCTTTCACTGCGGA		
TH108	GGATAGAACTCAAACGTTTATTAAAGAATGTCTTTTACGAAATGCCTGGA	<i>POP4_1</i> repair DNA	To introduce

TH109	AGACCCTGA TCTTGAAATCTATTTTCATTAAAGGGCTTTTCAGGGTCTTCCAGGCATTTTCG TGAAAAGA		<i>POP4_1</i> SNP
TH110	ATTGTAAAAAATAACAAAAAATGTCTGAAATTGGCATACGAAAAATAAAA TTACTGATA		
TH111	TCTCCTCTATGTAGTGTAAGGTCTTCTTTATCAGTAATTTTATTTTCGTATG CCAATT	<i>POP4_2</i> repair DNA	To introduce <i>POP4_2</i> SNP
TH154	AACAGAACTCAGTCTTTAAAAGAATTAGGGCTCAAAACGGATGACTTATTA TTGATTAGG		
TH155	ATCTGTTTGAATGGAATTGGAAATCTTACCCCTAATCAATAATAAGTCATCCG TTTTGAG	<i>DDI1</i> repair DNA	To introduce <i>DDI1</i> SNP
TH117	AGACCTGATGAGAAAGCCACATATTATCATTGCTACAACCTGGTAGATTAATG GACCATT		
TH118	CAGTGAAAACCCCTTTGGTATTCTCTAAATGGTCCATTAATCTACCAGTTGTA GCAATGAT	<i>RRP3</i> repair DNA	To introduce <i>RRP3</i> SNP
TH129	CCATTGTTCTTAGCGTGCTACCAAAAGAAAAATGCAGGTGCTATAATACATA CACACTCCTAAAATGCTGTGATATGCTCCTTGC		
TH130	TGTTAGCAATCCTGAACTCATCGCCAAATAGCAAGGAGCATATCACAGCATT TTAGGAGTGTGTATGTATTATAGCACCTGCATT	<i>MDE1</i> repair DNA	To introduce <i>MDE1</i> SNP
TH173	TTTCCATGTGTTCTTCGCATTTTTAATTCTAATCATACGGGCTTGTTAGAAGC AGGTCATTATATCGAG		
TH174	GTTCTTTAGGTTTCCATAGATAATTAACATCTCGATATAAATGACCTGCTTCT AACAAGCCCGTATGATT	<i>GRC3</i> repair DNA	To introduce <i>GRC3</i> SNP
TH123	AAGCTCTCTCAAGAGGATCTCTATTGTGTGGCAGTTATGCGAATGAAATGTT TCAAGTAG		
TH124	TCAGTCTCTCTAGTCGTTGATGACGGCAGTCTACTTGAAACATTTTCATTCGC ATAACTGC	<i>SWT21</i> repair DNA	To introduce <i>SWT21</i> SNP
TH148	AGCACTCCGTCCTTATCGACTCTATCGTCCTCTATGTCTCCAACATCCGCGAT CGAGGTGTGCACT		
TH149	ATTTGACTATCAAATGATTCTCCGCATGTAGTGCACACCTCGATCGCGGAT GTTGGAGACATAGA	<i>JJJ1</i> repair DNA	To introduce <i>JJJ1</i> SNP
TH119	TCCTCATCACACAATTCAAACCTCATCGCATCACGATGATGACAACCAACAC GGCAAGAGA		
TH120	GAAAGAATCAACACATCGCTGGAAAAATTTCTCTTGCCGTGTTGGTTGTC ATCATCGTG	<i>TAT2_1</i> repair DNA	To introduce <i>TAT2_1</i> SNP
TH127	ATCTTGATAGCTCAATTCTATTGTTCACTATGGACCATTGGTGGCTGGACCA GCGGCAAA		
TH128	ATAATTTTGAAAAAATATTTTAGCCCTTTCTTTGCCGCTGGTCCAGCCACCA	<i>TAT2_2</i> repair DNA	To introduce <i>TAT2_2</i> SNP

	ATGGTCCA		
TH125	GCGCTTTCTTCCCCTGATGAATCAAGCATCATGAGCACTACCTTTCTAGGCG AGACAGATGA		
TH126	CTACTTAATGTAGAGCCAGAATTATAAACTTCATCTGTCTCGCCTAGAAAGG TAGTGCTCAT	<i>ATG34</i> repair DNA	To introduce <i>ATG34</i> SNP



Supplementary Figure 1. Comparison of relative protein concentration of parental and evolved strains. Relative protein content was determined using the Bradford assay. Results are the mean of three experiments, and error bars indicate the standard deviation. Different letters represent significantly different means (Tukey's honestly significant difference tests, $p < 0.05$).

Reference

1. Hosaka K, Nikawa J, Kodaki T, Yamashita S. A dominant mutation that alters the regulation of *INO1* expression in *Saccharomyces cerevisiae*. J Biochem. 1992;111(3):352–8.
2. Lee YG, Jin YS, Cha YL, Seo JH. Bioethanol production from cellulosic hydrolysates by engineered industrial *Saccharomyces cerevisiae*. Bioresour Technol. 2017;228:355–61.
3. Zhang GC, Kong II, Kim H, Liu JJ, Cate JH, Jin YS. Construction of a quadruple auxotrophic mutant of an industrial polyploid *Saccharomyces cerevisiae* strain by using RNA-guided Cas9 nuclease. Appl Environ Microbiol. 2014;80(24):7694–701.