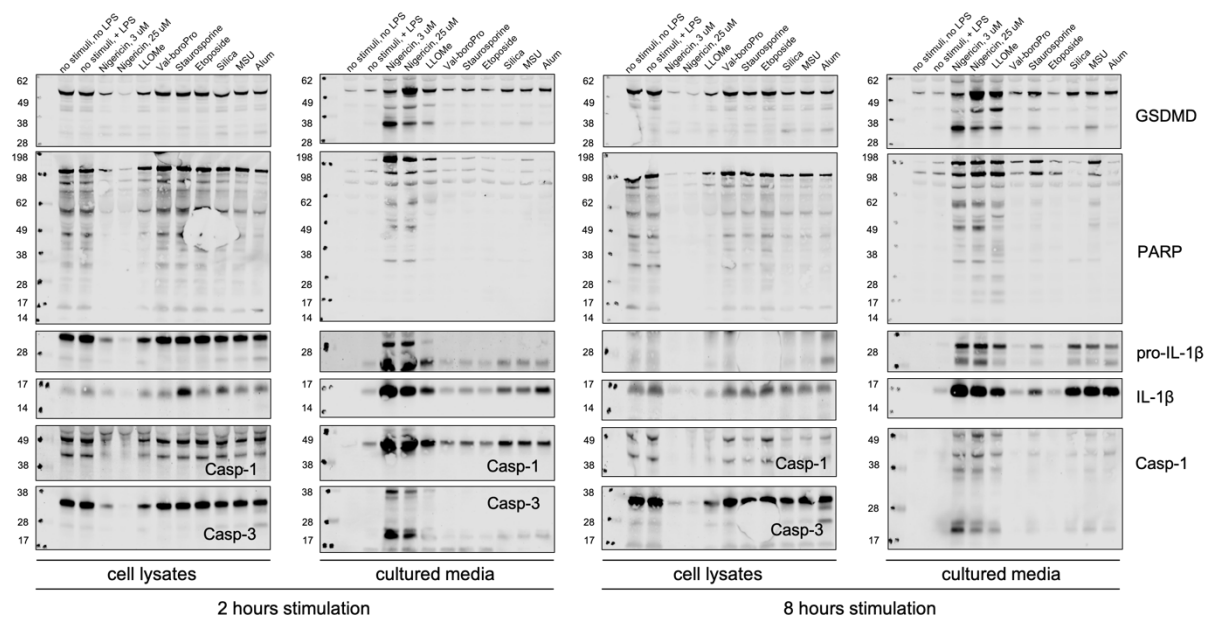


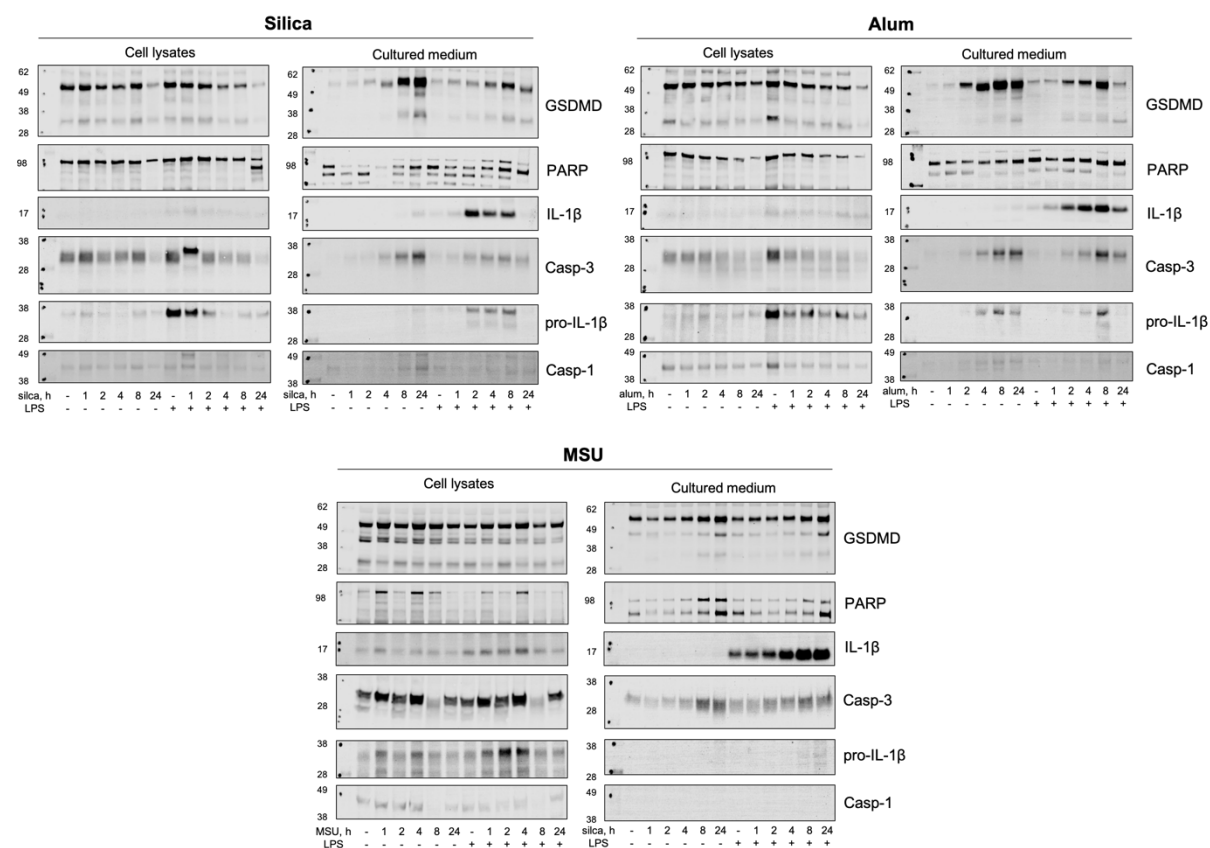
Non-redundant roles of cathepsins in promoting necrosis of human macrophages

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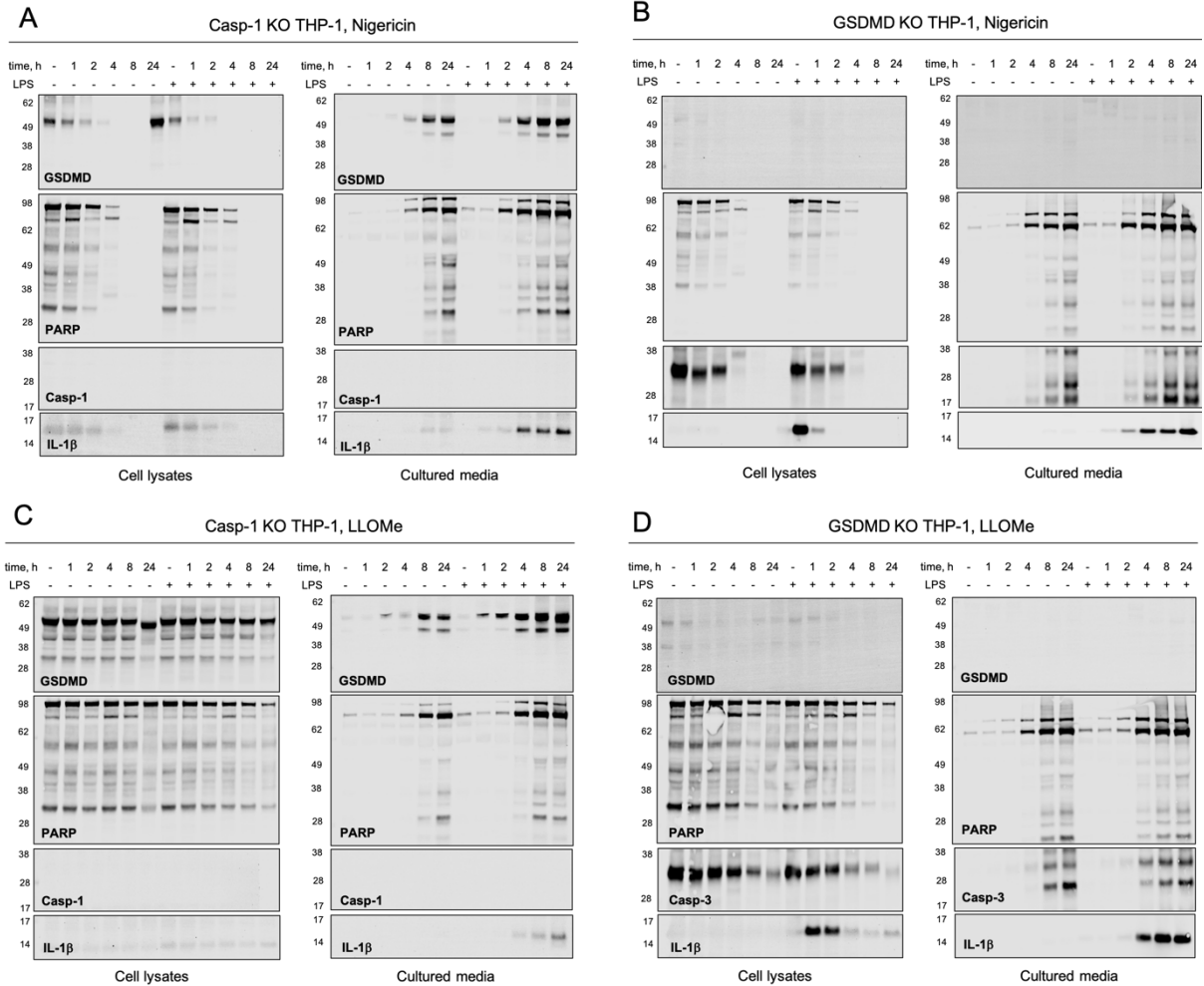
¹Faculty of Chemistry, Wrocław University of Science and Technology, 50-370 Wrocław, Poland; ²SBP Medical Discovery Institute, La Jolla, 92037, CA, USA; ³Jozef Stefan Institute, Ljubljana, 1000, Slovenia; ⁴Faculty of Chemistry and Chemical Engineering, University of Ljubljana, 1000 Ljubljana, Slovenia; ⁵Faculty of Medicine, Wrocław University of Science and Technology, 50-370 Wrocław, Poland; *correspondence: katarzyna.grobocz@gmail.com, marcin.poreba@pwr.edu.pl



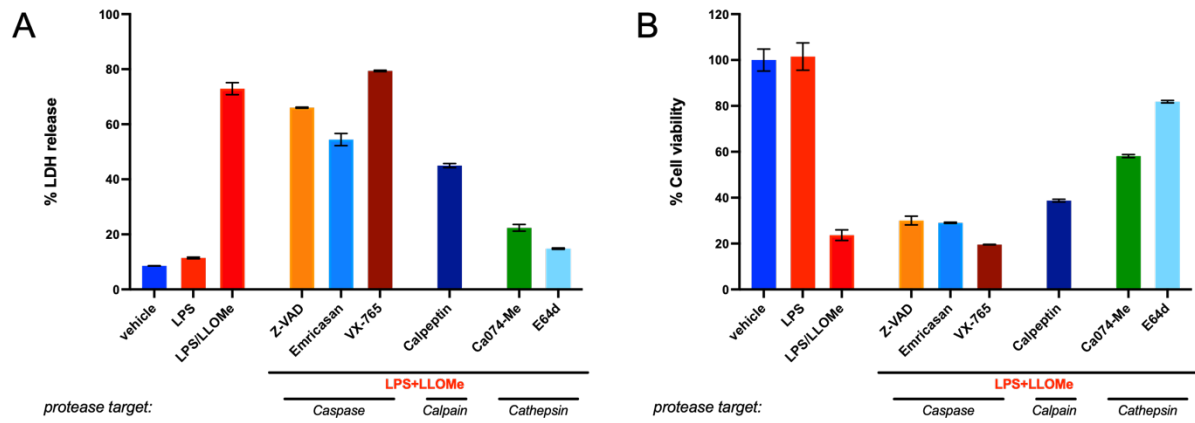
Supplementary Fig. 1 In relation to Figure 1. Cleavage patterns of GSDMD, PARP, IL-1 β , caspase-1, and caspase-3 in LPS-primed THP-1 cells treated with the indicated stimuli: nigericin (3 μ M or 25 μ M), L-leucyl-L-leucine methyl ester (LLOMe), Val-boroPro, staurosporine, etoposide, silica (SiO₂), monosodium urate (MSU), or alum [Al(OH)₃]. Immunoblots of cell lysates and culture supernatants show proteolytic fragments after 2 h and 8 h of stimulation.



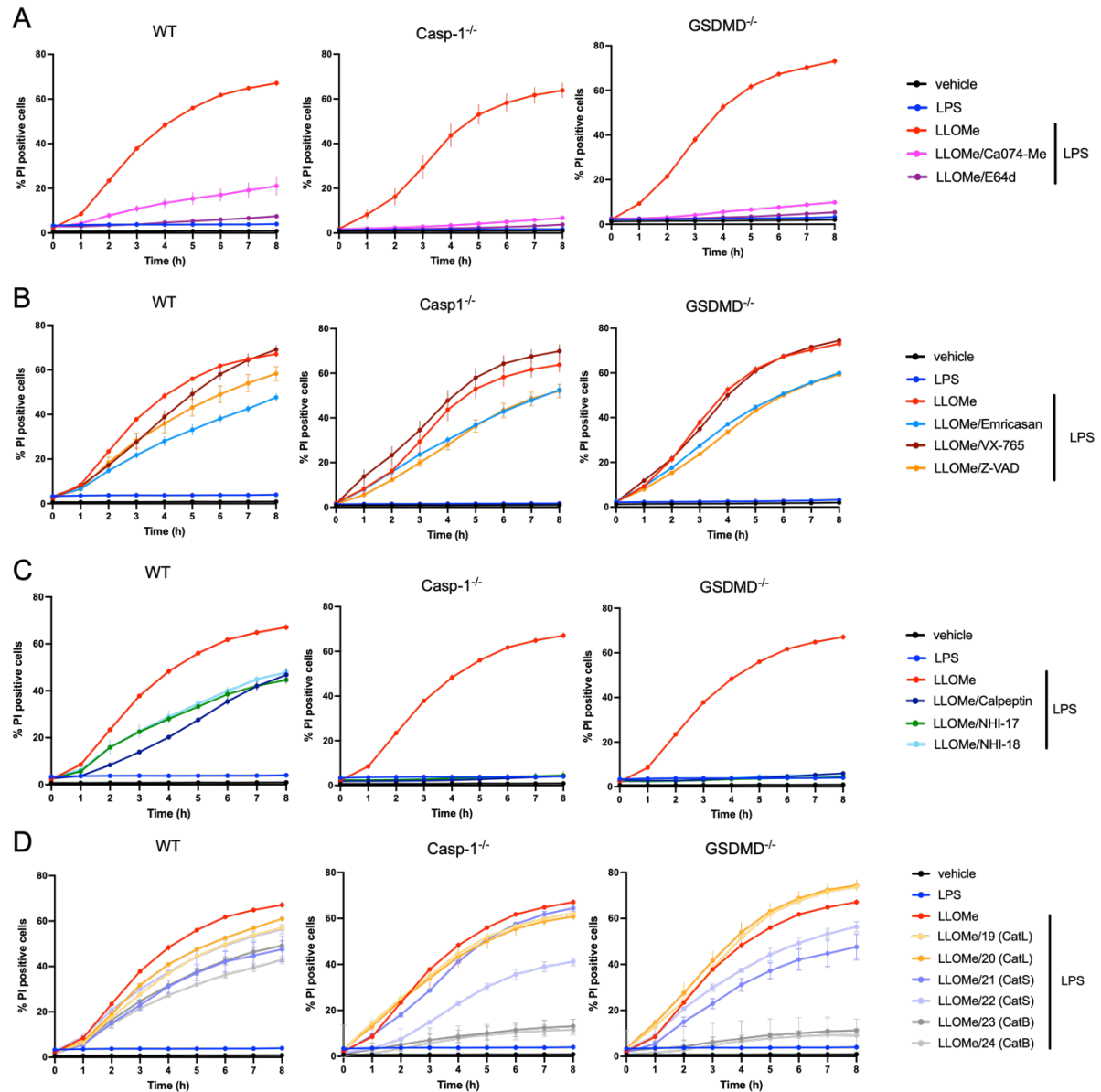
Supplementary Fig. 2 In relation to Figure 1. Cleavage patterns of GSDMD, PARP, (pro)IL-1 β , caspase-1, and caspase-3 in THP-1 cells $\pm$ LPS priming, treated with silica (SiO_2), alum [$\text{Al}(\text{OH})_3$], or monosodium urate (MSU) to model sterile inflammation. Immunoblots of cell lysates and culture supernatants show proteolytic fragments over a time course up to 24 h.



Supplementary Fig. 3 In relation to **Figure 2** Cell death signatures in **PMA-differentiated caspase-1- and GSDMD-deficient THP-1 cells**. Western blot analysis showing the processing patterns of GSDMD, PARP, and pro-IL-1 β in response to nigericin (A, B) and LLOMe (C, D) stimulation in caspase-1 KO (A, C) and GSDMD KO (B, D) THP-1 cells. Key features include GSDMD cleavage by caspase-1 (p31 fragment) or by caspase-3 (p45 fragment), and PARP processing via either the canonical apoptotic pathway (caspase-3; p89 and p25 fragments) or non-canonical cleavage by cysteine cathepsins (p55 and p42 fragments). Notably, pro-IL-1 β was processed and secreted as its mature form even in the absence of caspase-1, highlighting the contribution of alternative proteolytic pathways to cytokine activation.

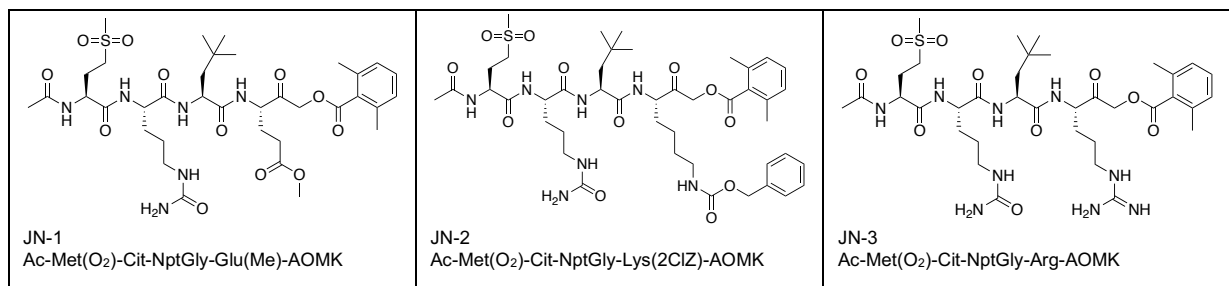


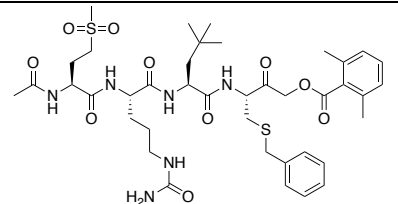
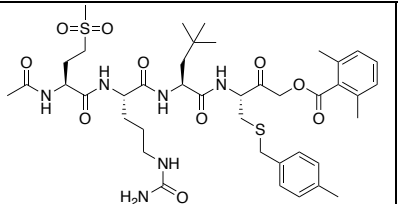
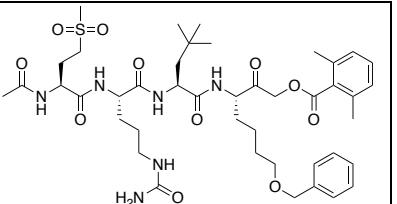
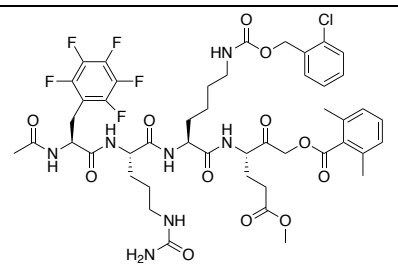
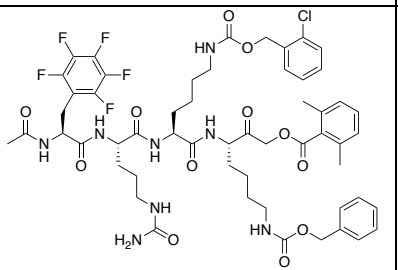
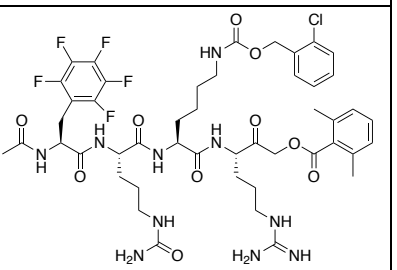
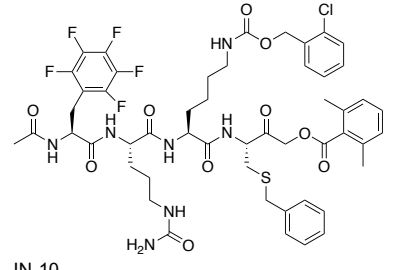
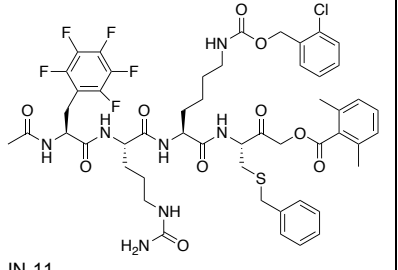
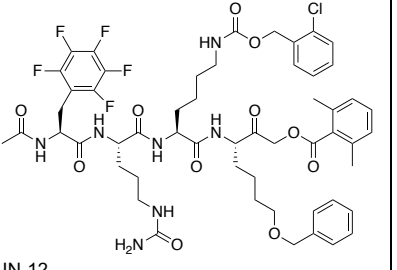
Supplementary Fig. 4 In relation to **Figure 3** LLOMe induces lytic cell death in PMA-primed, LPS-stimulated THP-1 cells. (A) shows LDH release (%), and (B) shows cell viability (%), following stimulation with LPS/LLOMe in the presence or absence of the indicated inhibitors: zVAF-fmk, emricasan, or VX-765 (caspases); calpeptin (calpain-1); and CA074Me or E64d (lysosomal cathepsins).



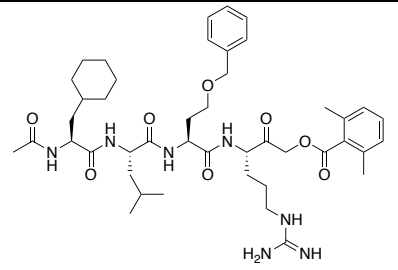
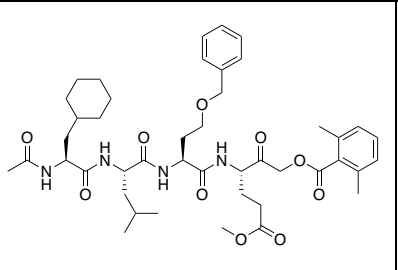
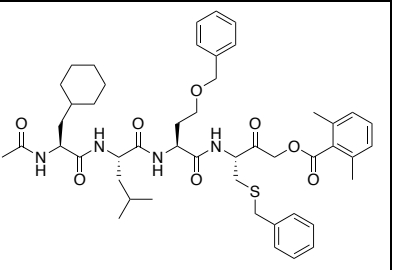
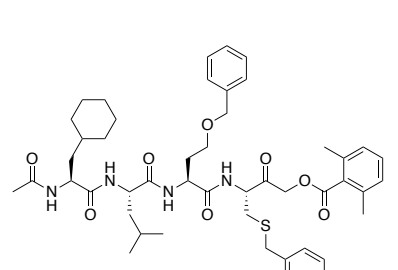
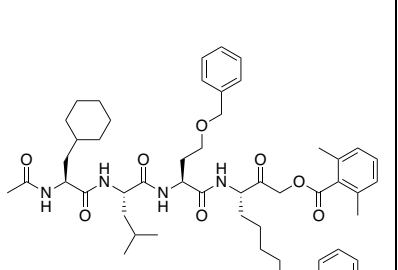
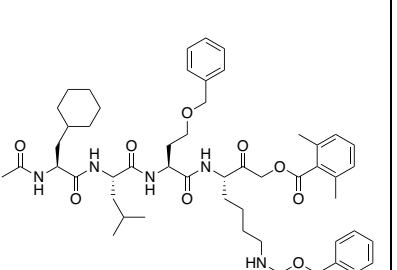
Supplementary Fig. 5 In relations to **Figure 3** Time-course analysis of PI uptake in LPS-primed M0 THP-1 cells treated with LLOMe (500 μ M) in the presence of; (A) pan-cathepsin inhibitors CA-074Me or E64d; (B) caspase inhibitors emricasan, VX-765, or zVAD-fmk; (C) calpeptin; and (D) cathepsin-selective inhibitors (CatL-19, CatL-20, CatS-21, CatS-22, CatB-23, CatB-24).

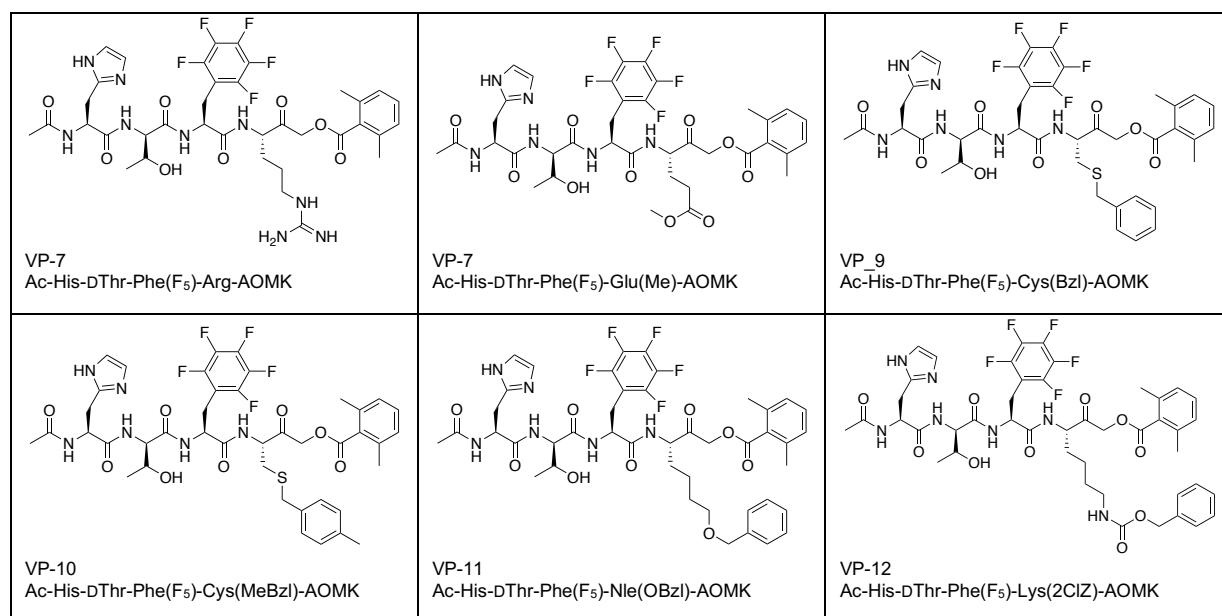
Supplementary Table 1 The structure of cathepsin S inhibitors. In relation to **Figure 3**.



 JN-4 Ac-Met(O₂)-Cit-NptGly-Cys(Bzl)-AOMK	 JN-5 Ac-Met(O₂)-Cit-NptGly-Cys(MeBzl)-AOMK	 JN-6 Ac-Met(O₂)-Cit-NptGly-Nle(OBzl)-AOMK
 JN-7 Ac-Phe(F₅)-Cit-Lys(2ClZ)-Glu(Me)-AOMK	 JN-8 Ac-Phe(F₅)-Cit-Lys(2ClZ)-Lys(2ClZ)-AOMK	 JN-9 Ac-Phe(F₅)-Cit-Lys(2ClZ)-Arg-AOMK
 JN-10 Ac-Phe(F₅)-Cit-Lys(2ClZ)-Cys(Bzl)-AOMK	 JN-11 Ac-Phe(F₅)-Cit-Lys(2ClZ)-Cys(MeBzl)-AOMK	 JN-12 Ac-Phe(F₅)-Cit-Lys(2ClZ)-Nle(OBzl)-AOMK

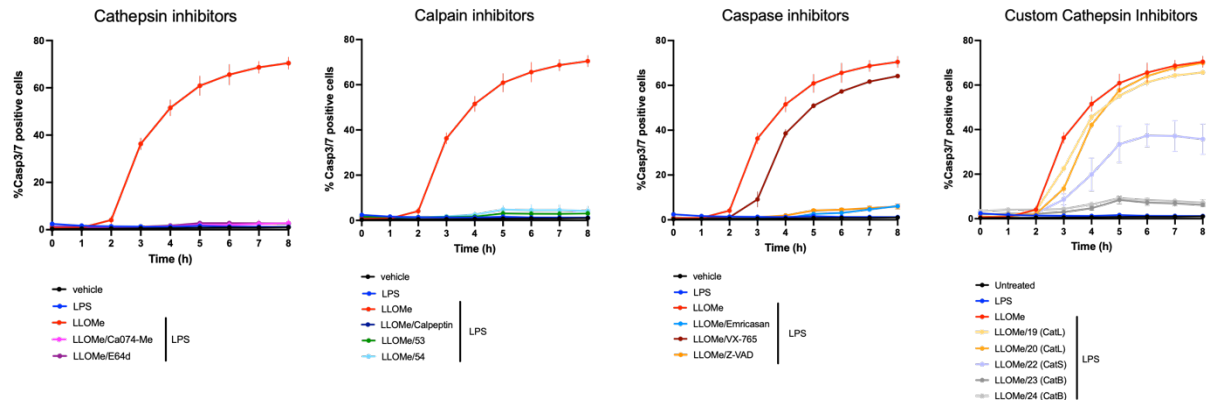
Supplementary Table 2. The structure of cathepsin B and cathepsin L inhibitors. In relation to Figure 3.

 VP-1 Ac-Cha-Leu-hSer(Bzl)-Arg-AOMK	 VP-2 Ac-Cha-Leu-hSer(Bzl)-Glu(Me)-AOMK	 VP-3 Ac-Cha-Leu-hSer(Bzl)-Cys(Bzl)-AOMK
 VP-4 Ac-Cha-Leu-hSer(Bzl)-Cys(MeBzl)-AOMK	 VP-5 Ac-Cha-Leu-hSer(Bzl)-Nle(OBzl)-AOMK	 VP-6 Ac-Cha-Leu-hSer(Bzl)-Lys(2ClZ)-AOMK

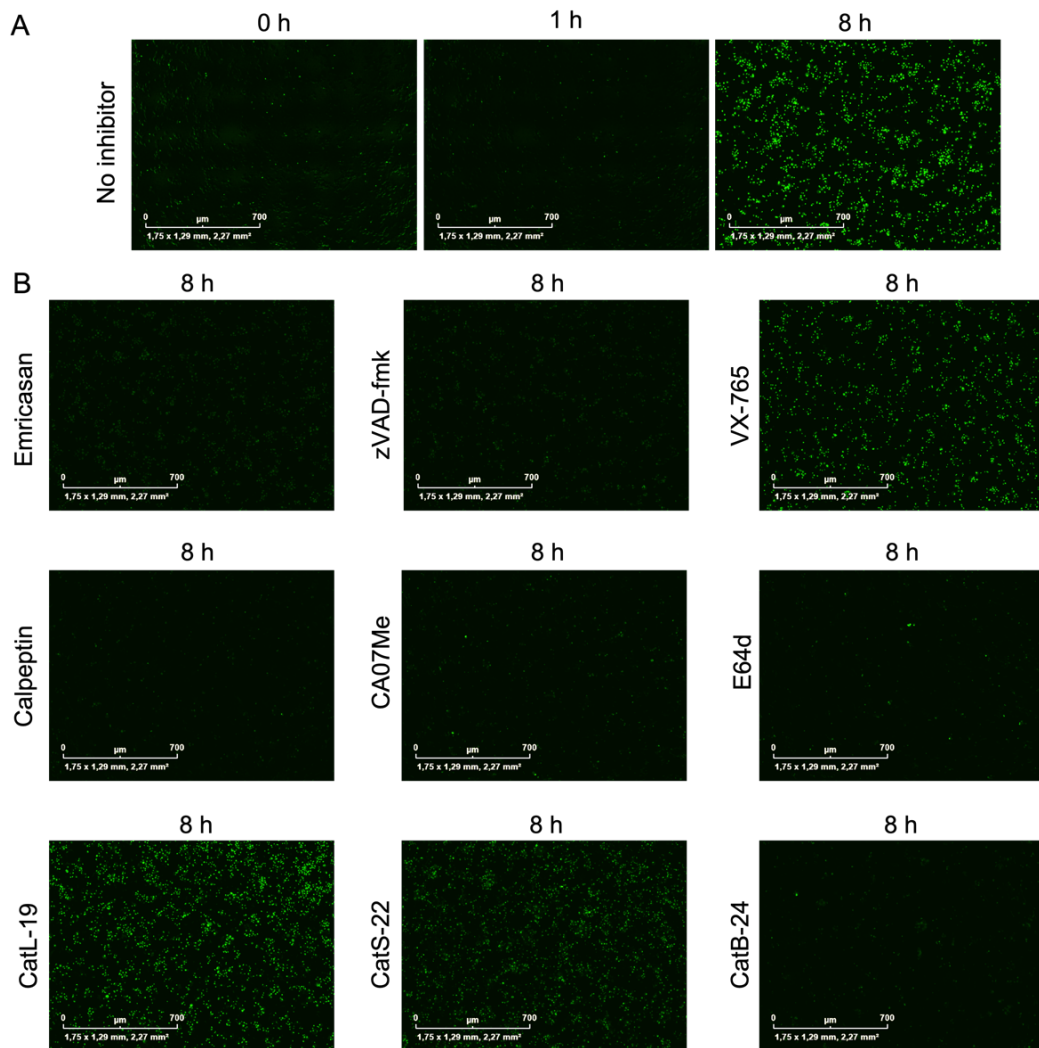


Supplementary Table 3. Kinetic parameters of cathepsin inhibitors. In relation to Figure 3.

P4-P2	P1	Code	$k_{obs}/[I], M^{-1}s^{-1}$		
			Cathepsin S	Cathepsin L	Cathepsin B
Cha-Leu-hSer(Bzl)	Arg	VP-1	3,700	2,700	1,790,000
	Glu(Me)	VP-2	760	rev. < 1,000	883,000
	Cys(Bzl)	VP-3	180	rev. < 1,000	174,000
	Cys(MeBzl)	VP-4	130	rev. < 1,000	106,000
	Nle(OBzl)	VP-5	460	rev. < 1,000	173,000
	Lys(2ClZ)	VP-6	570	rev. < 1,000	2,200
His-DThr-Phe(F ₅)	Arg	VP-7	20	69,900	1,400
	Glu(Me)	VP-8	1,100	26,600	2,300
	Cys(Bzl)	VP-9	710	45,500	3,900
	Cys(MeBzl)	VP-10	220	29,600	480
	Nle(OBzl)	VP-11	700	32,200	42,00
	Lys(2ClZ)	VP-12	900	17,400	830
Met(O ₂)-Cit-NptGly	Glu(Me)	JN-1	392,000	560	7,440
	Lys(2ClZ)	JN-2	269,000	220	5,100
	Arg	JN-3	236,000	4,840	41,500
	Cys(Bzl)	JN-4	443,000	450	22,400
	Cys(MeBzl)	JN-5	344,000	610	10,500
	Nle(OBzl)	JN-6	246,000	650	16,900
Phe(F ₅)-Cit-Lys(2ClZ)	Glu(Me)	JN-7	609,999	870	17,000
	Lys(2ClZ)	JN-8	225,000	350	1,600
	Arg	JN-9	371,000	4,270	148,000
	Cys(Bzl)	JN-10	545,000	620	16,100
	Cys(MeBzl)	JN-11	261,000	320	1,900
	Nle(OBzl)	JN-12	392,000	490	5,160
		E-64	132,000	65,100	81,200

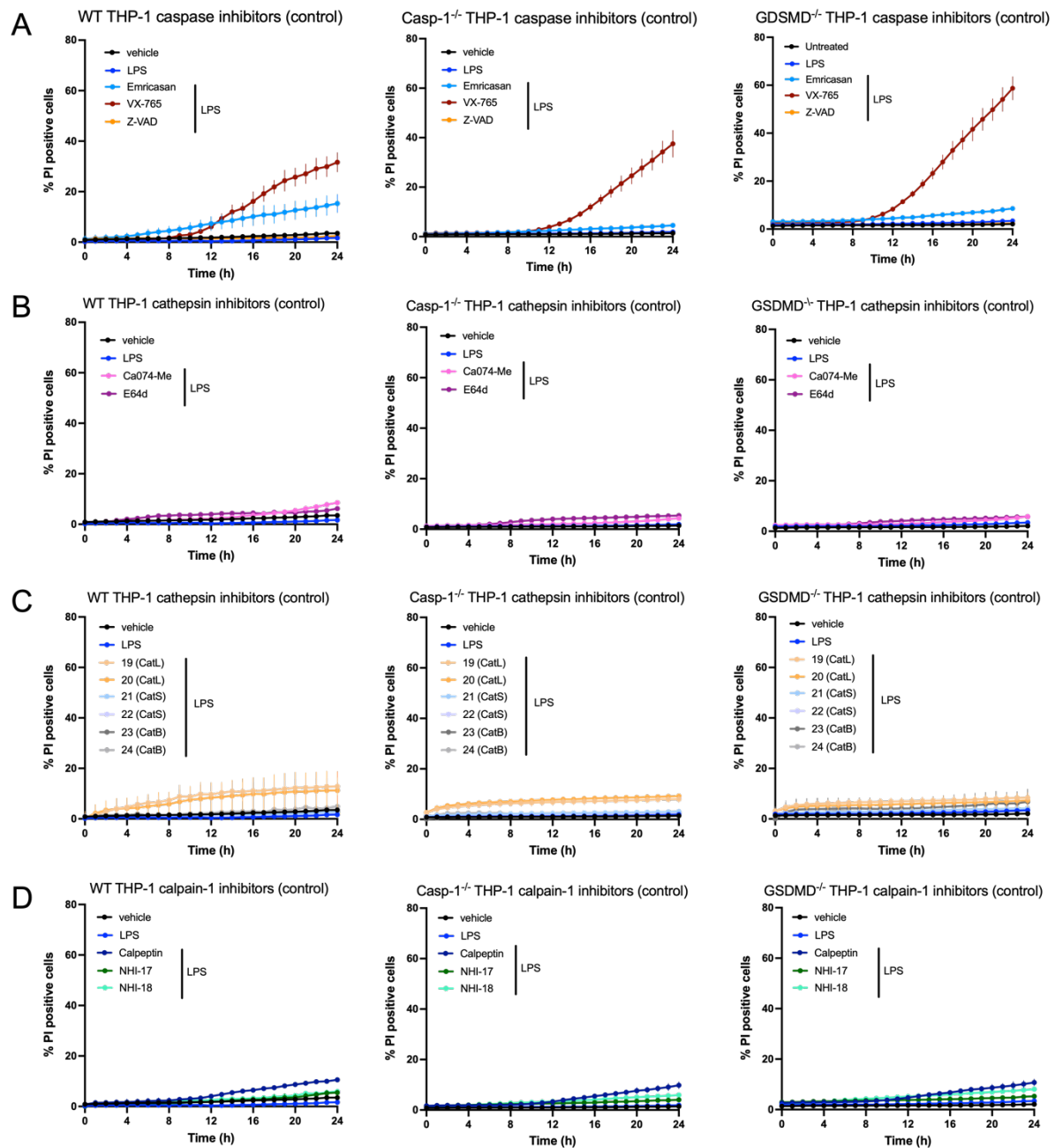


Supplementary Figure 6. In relations to Figure 3 Time-course analysis of DEVD-ase activity in LPS-primed M0 THP-1 cells treated with LLOMe (500 μ M) in the presence of pan-cathepsin inhibitors (CA-074Me or E64d); calpain-1 inhibitors (calpeptin, NHI-17, NHI-18); caspase inhibitors (emricasan, VX-765, zVAD-fmk); and cathepsin-selective inhibitors (CatL-19, CatL-20, CatS-21, CatS-22, CatB-23, CatB-24).

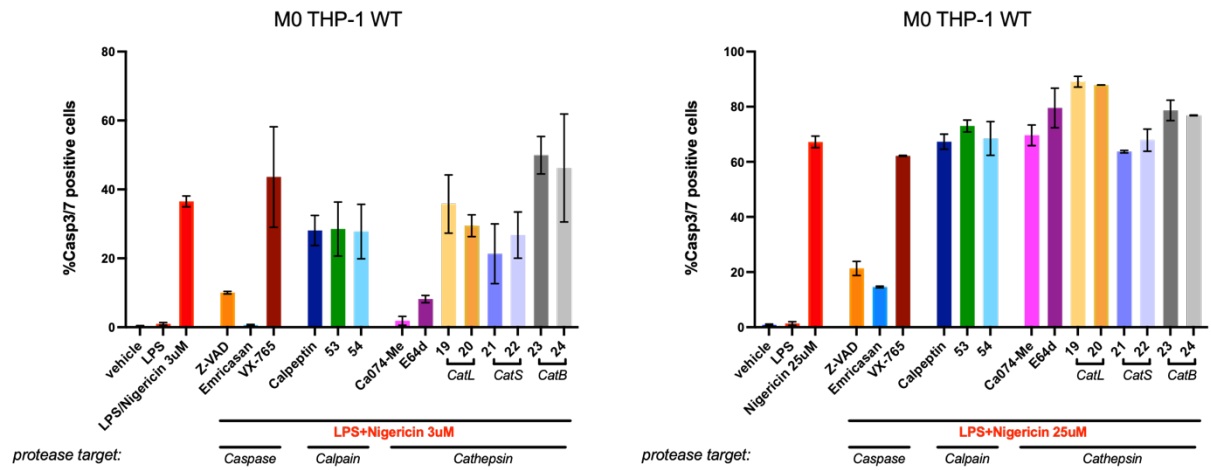


Supplementary Figure 7. Caspase-3/7 activation in LPS/LLOMe-treated WT M0 THP-1 cells. (A) Representative fluorescence images of caspase-3/7 activity in stimulated cells with no inhibitor at 0, 1,

and 8 h. (B) Representative fluorescence images of caspase-3/7 activity after 8 h of stimulation in cells pretreated with the indicated inhibitors (25 μ M, 2 h prior stimulation).



Supplementary Figure 8. In relations to Figure 3. Time-course analysis of PI uptake in LPS-primed M0 THP-1 cells (WT, Casp-1^{-/-}, and GSDMD^{-/-}) without cell death stimulants in the presence of (A) caspase inhibitors (emricasan, VX-765, zVAD-fmk), (B) pan-cathepsin inhibitors (CA-074Me or E64d), (C) cathepsin-selective inhibitors (CatL-19, CatL-20, CatS-21, CatS-22, CatB-23, CatB-24), and calpain-1 inhibitors (calpeptin, NHI-17, NHI-18).



Supplementary Figure 9. In relation to Figure 4. DEVD-ase activity in M0 WT THP-1 cells following 3 μ M and 25 μ M nigericin in the presence of pan-caspase (zVAD, Emricasan), caspase-1 selective (VX-765), calpain-1 (Calpeptin), or cathepsin inhibitors (CA-074Me, E64d, CatL-19, CatL-20, CatS-21, CatS-22, CatB-23, CathB-24).