

## Extended Data for

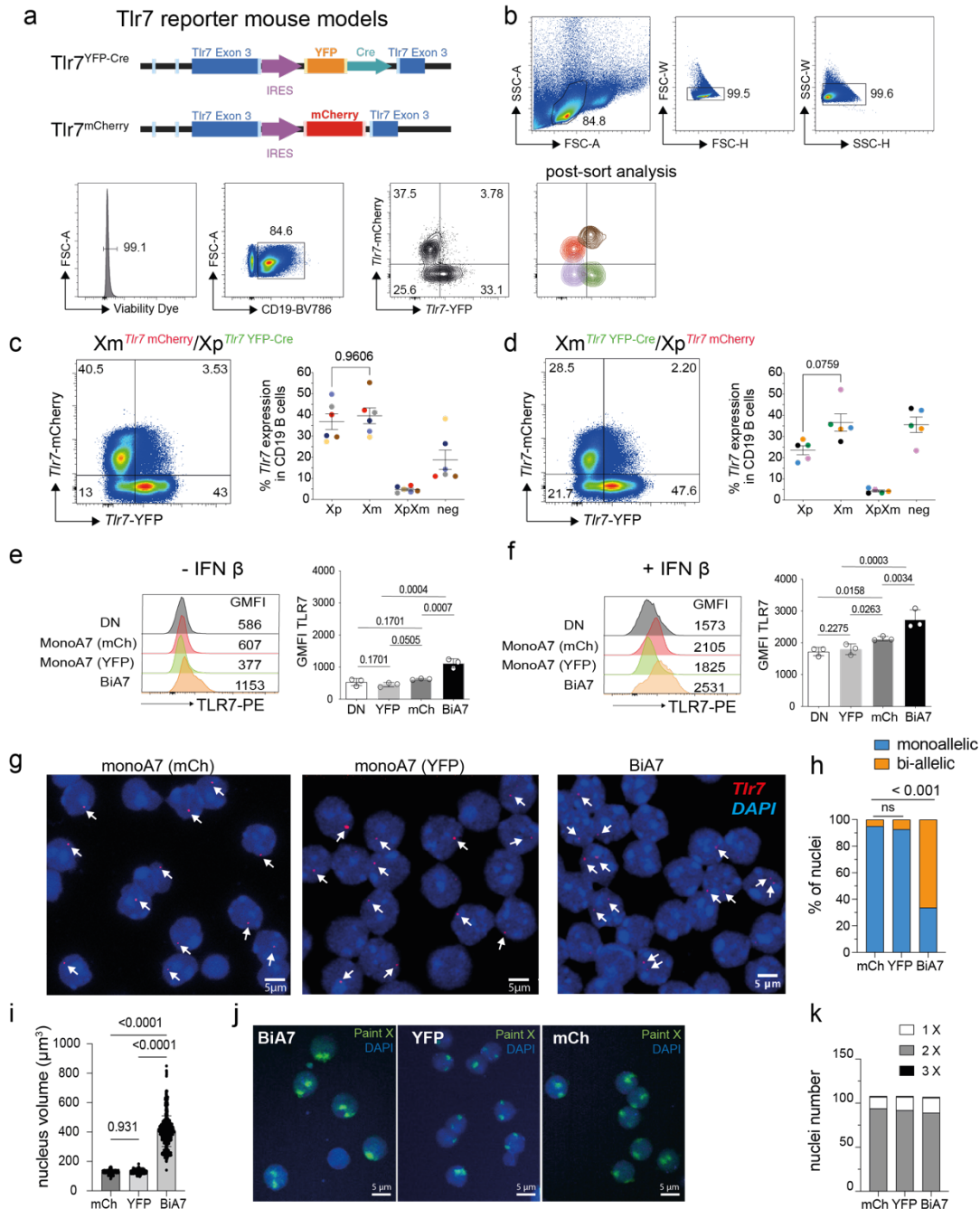
### ***Tlr7*-biallelism defines a hyperfunctional state of female B lymphocytes**

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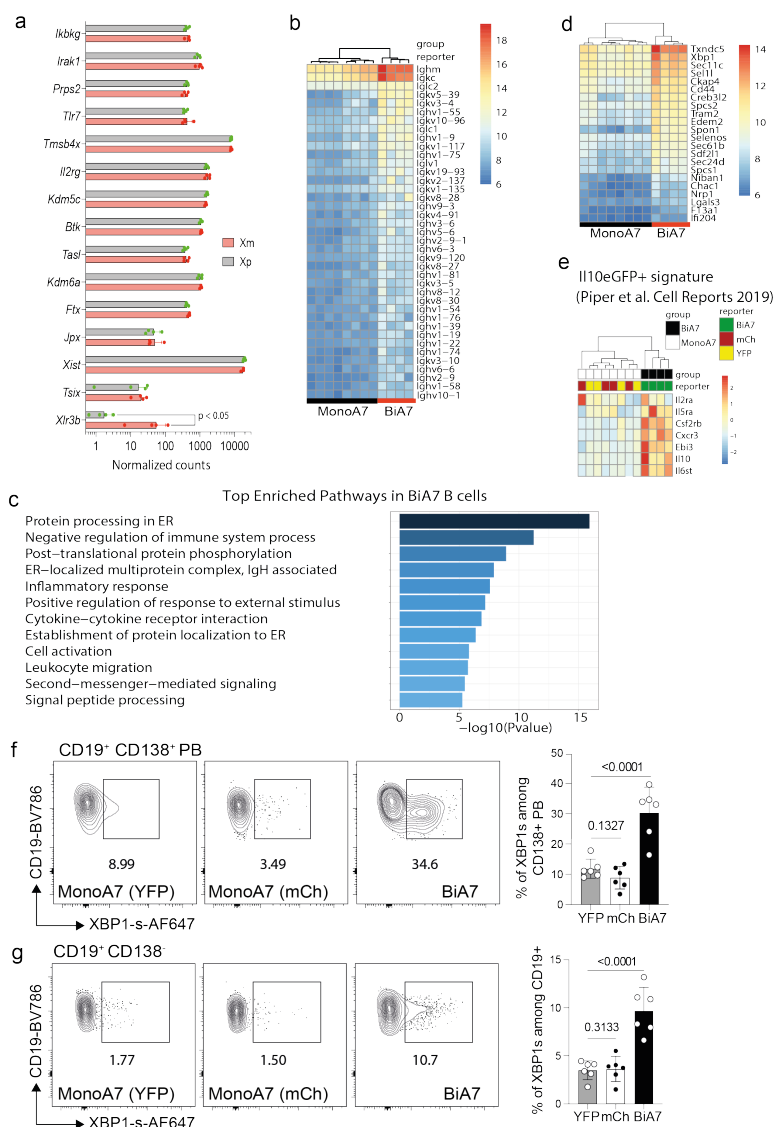
#### **The PDF file includes:**

Extended Data Fig. 1 to Fig. 10



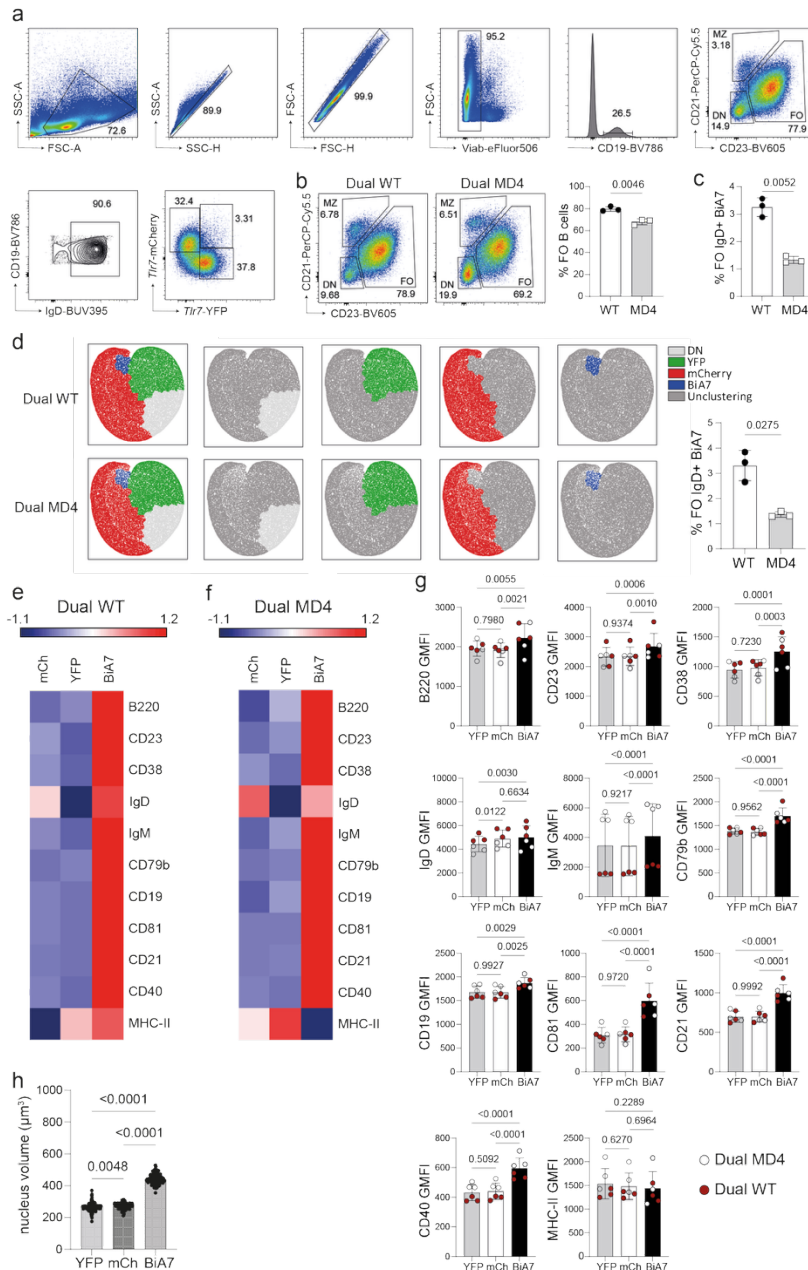
**Extended data Figure 1: Characterization of *Tlr7* bi-allelic splenic B cells.** (a) Design of *Tlr7*-reporter mice. A reporter cassette including an IRES followed by a fluorescent protein (mCherry or YFP-Cre) was inserted at the 3' end of the endogenous *Tlr7* gene. (b) Gating strategy for cell-sorting of MonoA7 (mCherry or YFP), BiA7 (mCherry<sup>+</sup>/YFP<sup>+</sup>) and double negative (DN) CD19<sup>+</sup> B cells. Representative flow cytometry purity profile from cell sorting experiment is shown. (c, d) Representative flow cytometry plot and quantification of *Tlr7*-reporter allelic expression from splenic CD19<sup>+</sup> B cells from  $Xm^{Tlr7\ mCherry}/Xp^{Tlr7\ YFP-Cre}$  (c) or the reverse crossing  $Xm^{Tlr7\ YFP-Cre}/Xp^{Tlr7\ mCherry}$  (d) female mice (maternal X, Xm; paternal X, Xp). (e, f) GMFI quantification of intracellular TLR7 staining in FACS-sorted DN, MonoA7 and BiA7 CD19<sup>+</sup> B cells cultured overnight without (e) or with 2 ng/ml IFN- $\beta$  (f). TLR7 staining expressed as  $\Delta$ GMFI with isotype control. Quantifications are shown from three independent experiments. (g) Representative RNA FISH images of *Tlr7* primary transcripts.

Nuclei counterstained with DAPI (blue). Scale bars, 5 $\mu$ m. **(h)** Frequency of mono- and bi-allelic RNA FISH signals within the sorted MonoA7 or BiA7 cell populations. **(i)** Nucleus volume ( $\mu$ m<sup>3</sup>) measured by Imaris from MonoA7 and BiA7 B cells. **(j)** Representative images of paint DNA FISH for the X chromosomes (green). Nuclei counterstained with DAPI (blue). Scale bars, 5 $\mu$ m. **(k)** Quantification of the number of X chromosome territories per nucleus in sorted MonoA7 and BiA7 B cells. Statistical analysis was performed using One-way ANOVA with Tukey multiple-comparison test. For (h) statistical differences were assessed using Fisher's exact test or One-way ANOVA with Tukey multiple-comparison test (i). Exact p values are shown.



**Extended data Figure 2: Characterization of the transcriptional signature of BiA7 B cells.** RNA-seq data between BiA7 and MonoA7 B cells (n=4/group). **(a)** Relative expression of the maternally imprinted gene *Xlr3b* compared to 14 representative X-linked genes from individual mice (n=4). Adjusted p value is shown. **(b)** Heatmap representing variance-stabilized normalized counts for Ig-related genes identified as differentially regulated between BiA7 and MonoA7 samples and having an average expression greater than 200 normalized counts in at least 3 of the 4 BiA7 samples. **(c)** Metascape analysis showing the top enriched pathways corresponding to the selected cluster summary terms obtained from 504 non-Ig genes with log<sub>2</sub>FC > 1, pvalue < 0.05 with background: 12563 non-Ig expressed genes. **(d)** Heatmap showing the unsupervised clustering of the DEG (log<sub>2</sub>FC > 1, adj. p-value

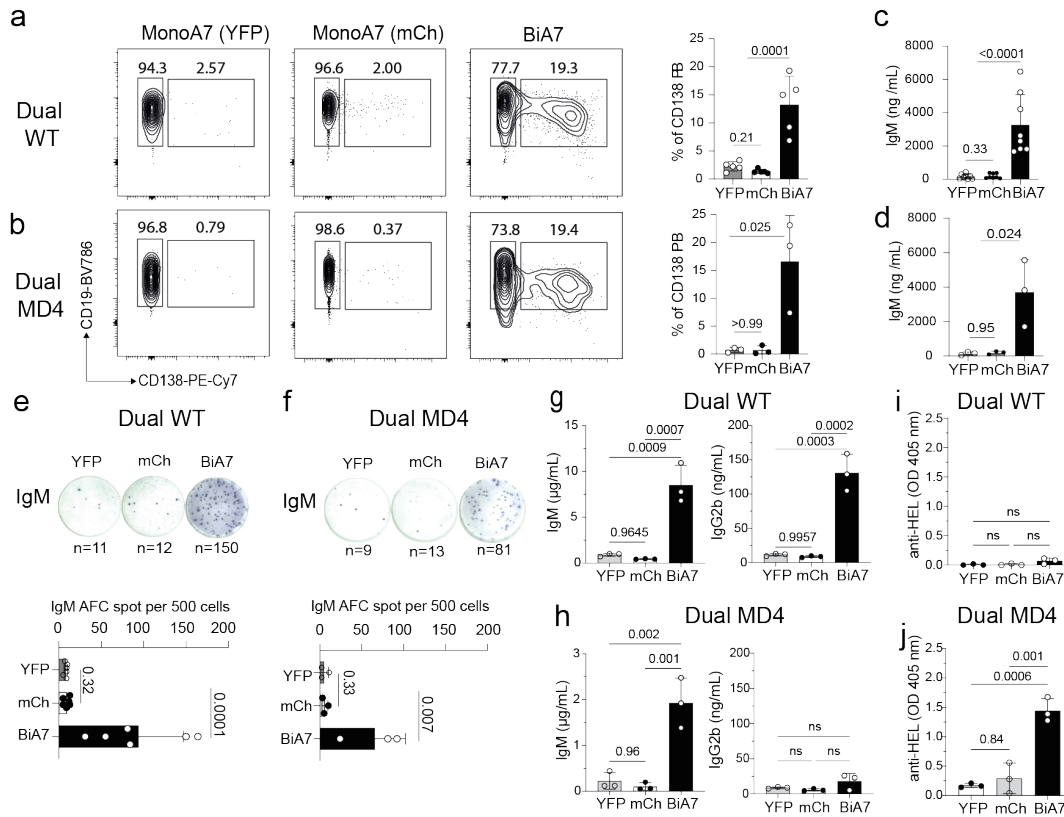
<0.05) from the pathways underlined in bold panel in (c). (e) Heatmap of *Il10*eGFP+ signature genes upregulated in BiA7 B cells with  $\log_2FC > 1$  adj. p-value <0.05. Heatmap rows ordered based on average expression level in BiA7 samples. (f,g) Intracellular analysis of XBP1s expression on CD19+CD138+ PB (f) or CD19+CD138- B cells (g) from splenic IgD+ MonoA7 or BiA7 B cells sorted and stimulated as in Fig. 1g for 3 days. Statistical analysis was performed using One-way ANOVA with Tukey multiple-comparison test. The symbols represent individual mice; the error bars represent the mean  $\pm$  SD.



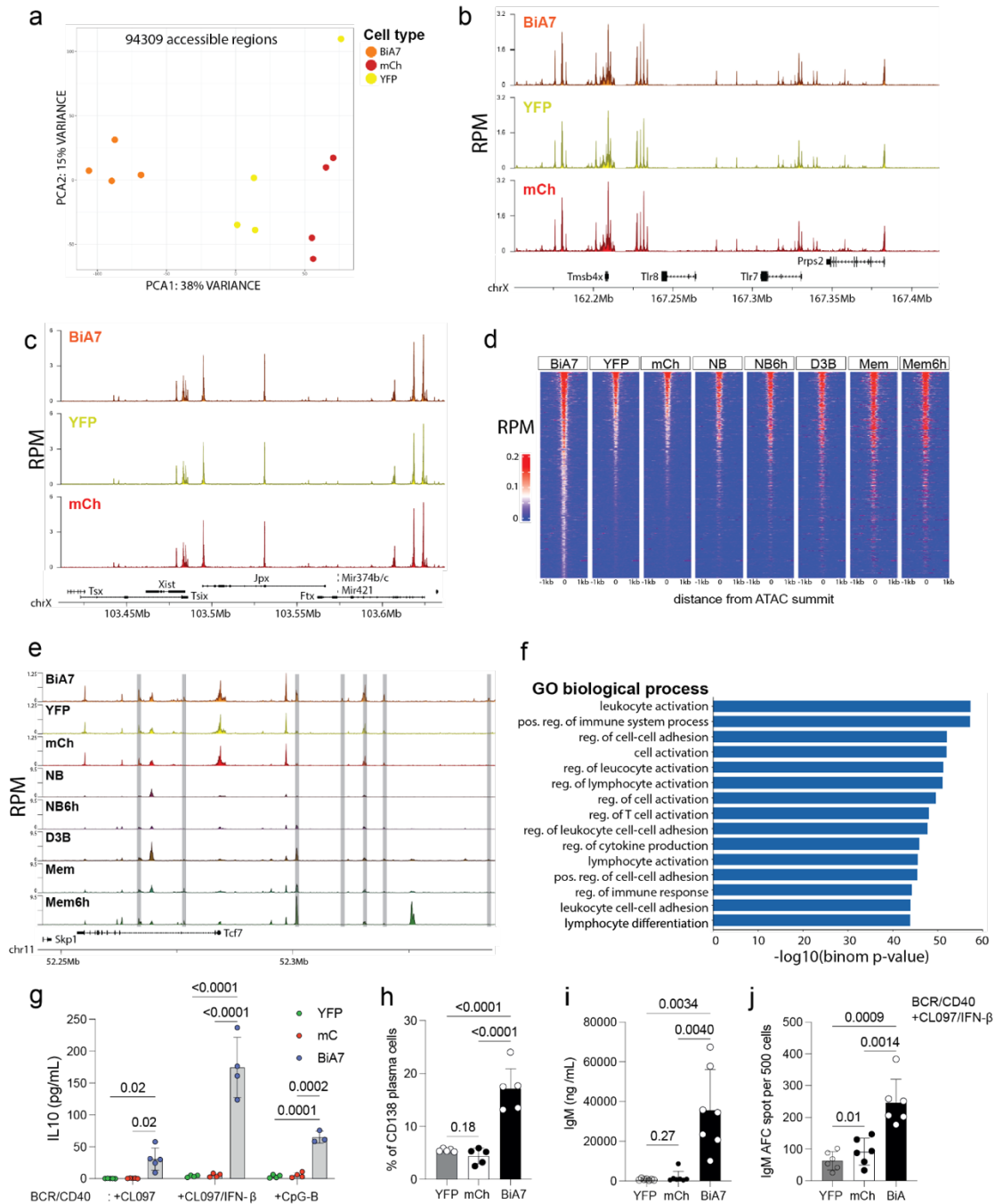
### Extended data Figure 3: BCR-specificity does not influence the development of BiA7 B cells.

(a) *Ex vivo* gating strategy on splenocytes from Dual WT and Dual MD4 mice. (b) Representative cytometry plot of DN, MZ and FO splenic B cells profile and the frequency of FO B cells in Dual WT and Dual MD4 mice. (c) Frequency of BiA7 naive FO IgD+ B cells in Dual WT and Dual MD4 mice. (d) Reporter-based unsupervised clustering and quantification of BiA7 among FO IgD+ cells in Dual WT and Dual MD4 mice. (e-g) Expression of cell surface markers and functional molecules in sorted MonoA7 and BiA7 FO B cells from Dual WT (e) or Dual MD4 (f) mice. (g) Histograms of GMFI of these markers from (e,f). (h) Nucleus volume ( $\mu\text{m}^3$ ) measured by Imaris in sorted MonoA7 and BiA7 B cells. Statistical analysis was performed using Welch's t test (b,c,d), or One-way ANOVA with Tukey

multiple-comparison test (**g,h**). The symbols represent individual mice; the error bars represent the mean $\pm$ SD.



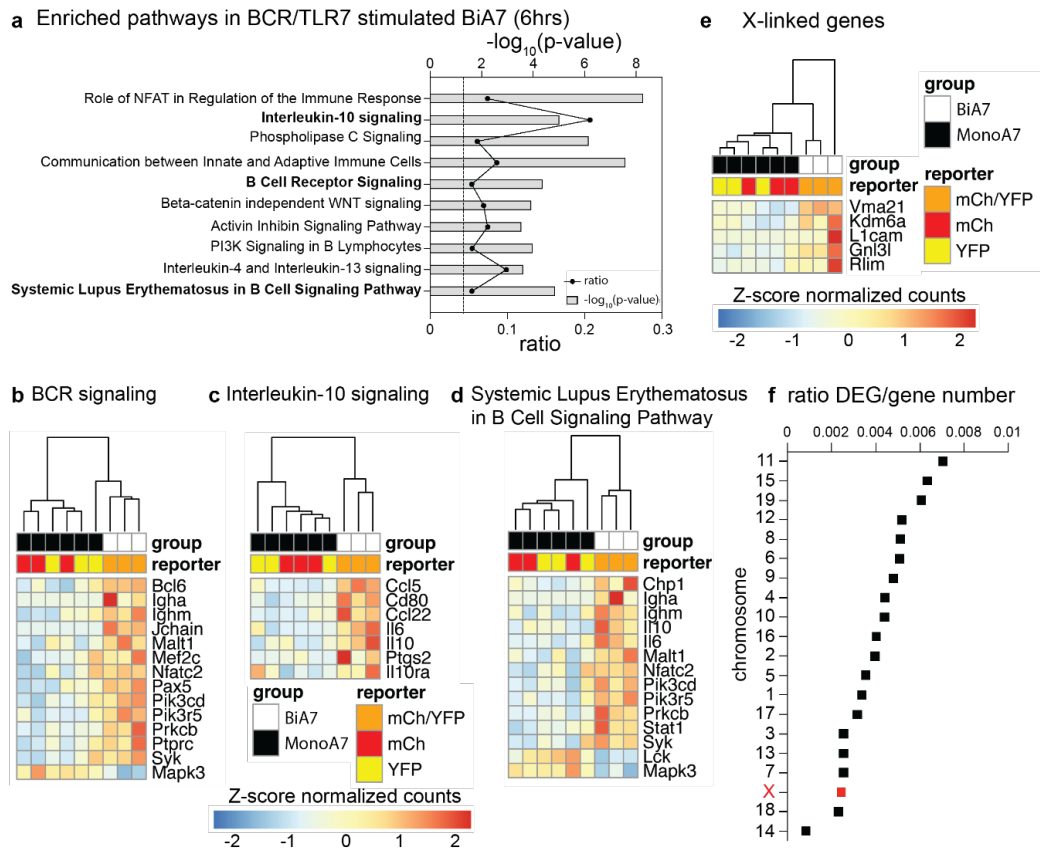
**Extended data Figure 4: BiA7 FO B cells with a hyperactive potential developed independently of BCR specificity.** (**a, b**) Representative cytometry plot of CD19 CD138 staining and frequency of CD19<sup>+</sup>CD138<sup>+</sup> PB on sorted MonoA7 and BiA7 IgD<sup>+</sup> FO B cells cultured with F(ab')<sub>2</sub> anti-IgM, anti-CD40 and CL097 for 3 days from Dual WT (**a**) and Dual MD4 (**b**) mice. Concentration of IgM collected in supernatants of cultured MonoA7 and BiA7 cells from Dual WT (**c**) and Dual MD4 (**d**) mice assessed by ELISA. Analysis of antibody forming cells (AFC) secreting IgM antibodies from Dual WT (**e**) and Dual MD4 (**f**) assessed by ELISPOT. (**g, i**) Comparative analysis of the production of IgM, class-switched IgG2b (**g, h**) and anti-HEL antibody (**i, j**) between the indicated B cell subsets purified from Dual WT (**g, i**) or Dual MD4 (**h, j**) mice. Statistical analyses were performed using a One Way ANOVA and multiple comparison Dunnett's test. The symbols represent individual mice; the error bars represent the mean $\pm$ SD.



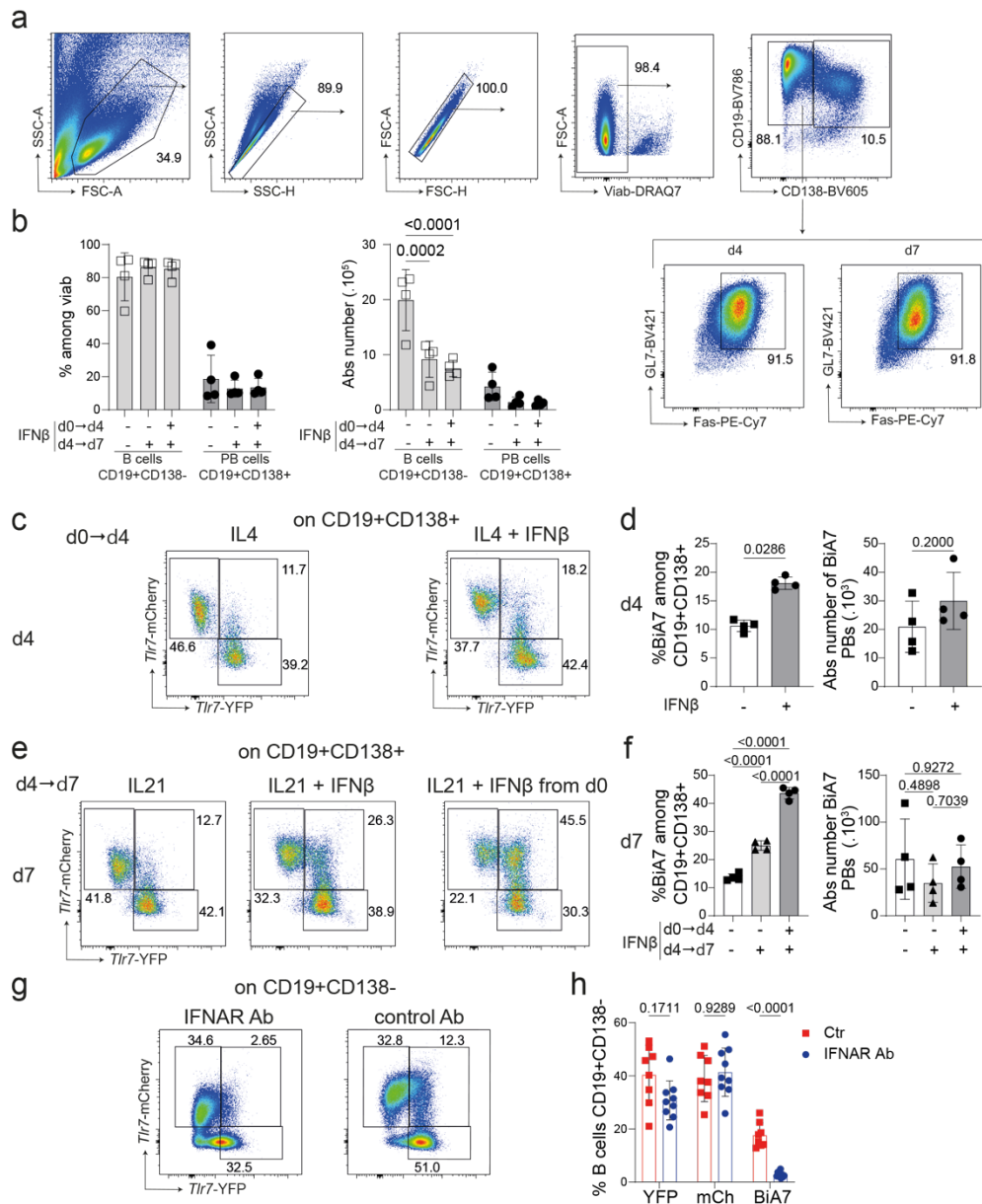
**Extended data Figure 5: Accessibility landscape changes in naive BiA7 B cells.** (a) Principal component analysis of chromatin accessibility across 94309 accessible regions between BiA7 and mCh, YFP MonoA7 B cells. (b, c) Chromatin accessibility at the *Tlr7/Tlr8* locus (b) and in the *Xist* region (c) in BiA7 and MonoA7 B cells. (d) Heatmap of chromatin accessibility measured by ATAC-seq across BiA7-specific DARs centered on ATAC-seq summits (+1kb) in BiA7 cells compared to MonoA7 (YFP or mCh) B cells and as well as in naïve (NB), activated (D3B) and memory (Mem) or activated Mem B cells from Q $\beta$ -VLP immunized mice. (e) Chromatin accessibility at the *Tcf7* locus in the indicated B cell populations. (f) Statistical association of GO biological processes with BiA7-specific DARs. (g) Quantification of IL-10 production in 3 day-culture supernatants of CD19<sup>+</sup>IgD<sup>+</sup> sorted BiA7 and MonoA7 cells stimulated with F(ab')<sub>2</sub> goat anti-mouse IgM, anti-CD40, and the indicated TLR ligands, either CL097 for TLR7 (alone or in combination with IFN- $\beta$ ) or CpG-B for TLR9. (h-j) Analysis of CD19<sup>+</sup>IgD<sup>+</sup> sorted BiA7 and MonoA7 cells after 3 days of stimulation with F(ab')<sub>2</sub> goat anti-mouse



IgM, anti-CD40, CL097 and IFN- $\beta$ : Frequency of CD19<sup>int</sup> CD138<sup>+</sup> PB (h), concentration of IgM (i), and frequency of IgM secreting cells assessed by ELISPOT (j). One-way ANOVA followed by Sidak's multiple comparisons test was used to compare BiA7 cells with the other groups. The symbols represent individual mice; bars represent the mean; the error bars represent the mean $\pm$ SD.

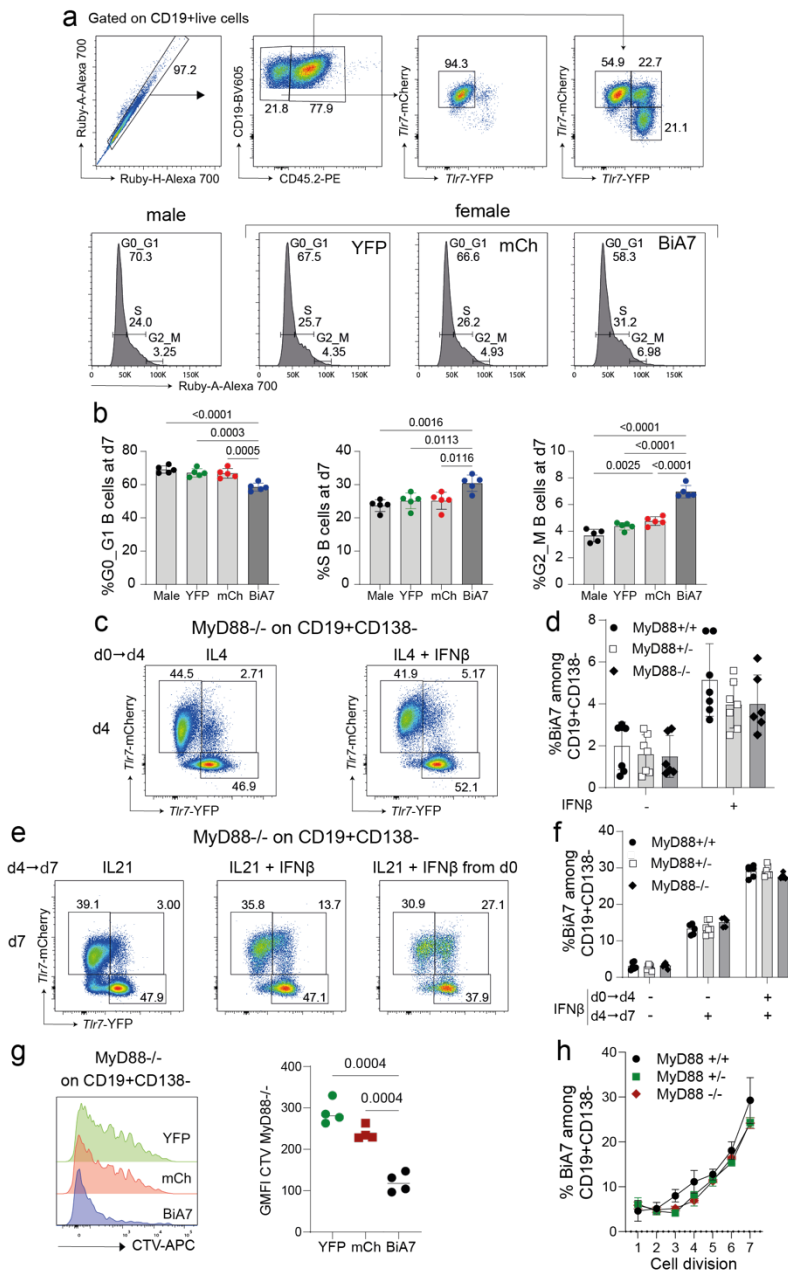


**Extended data Figure 6. Early transcriptional changes upon activation of BiA7 B cells.** CD19<sup>+</sup>IgD<sup>+</sup> sorted BiA7 and MonoA7 cells were stimulated with F(ab')<sub>2</sub> goat anti-mouse IgM, anti-CD40, CL097 for 6 hours and processed for RNA-seq analysis. (a) Up-regulated pathways in activated BiA7 B cells identified using ingenuity pathway analysis (IPA, QIAGEN). (b - d) Heatmap representing expression of DEGs from BCR signaling (b), IL10 signaling (c) and from Systemic Lupus Erythematosus in B cell IPA Signaling (d) Pathways in CD19<sup>+</sup>IgD<sup>+</sup> sorted BiA7 and mCh, YFP MonoA7 activated B cells. (e) expression of X-linked DEGs in BiA7 and MonoA7 activated B cells. (f) DEG/total gene number ratio for each chromosome.

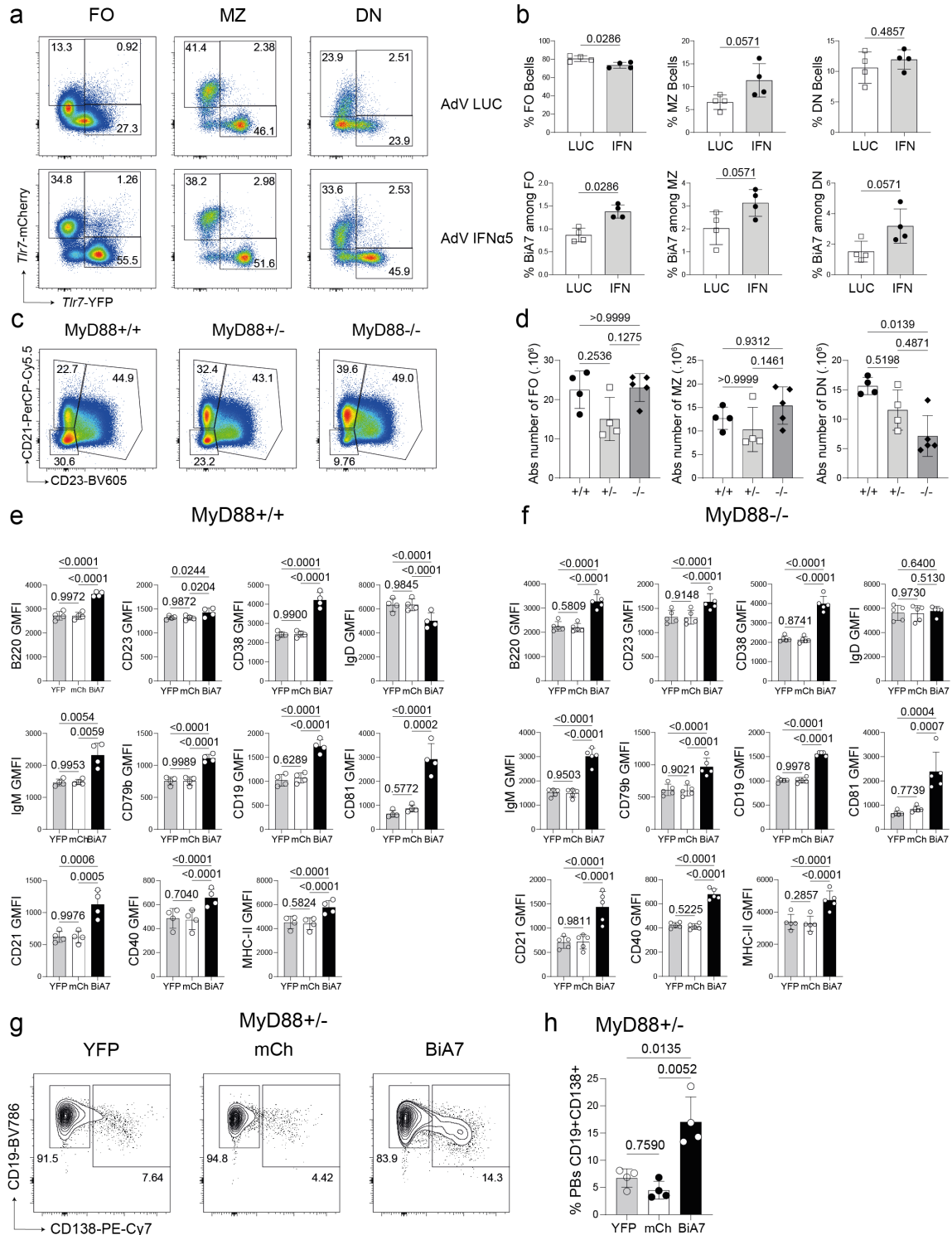


**Extended data Figure 7: IFNAR1-signaling is required for the emergence of BiA7 B cells.** (a) Gating strategy used for iGB cell analysis by flow cytometry. (b) Frequency and absolute number of CD19<sup>+</sup>CD138<sup>-</sup> B cells and CD19<sup>+</sup>CD138<sup>+</sup> PBs in iGB cell culture system with or without IFNβ. (c-f) Representative flow cytometry plots showing *Tlr7*-reporter expression in CD19<sup>+</sup>CD138<sup>+</sup> PBs at d4 (c) and d7 (e) of iGB cell culture with or without IFNβ. Related histograms showing the frequency and the absolute number of BiA7 B cells among PBs at d4 (d) or d7 (f). (g, h) Analysis of naive B cells from iGB culture with IL-4 alone for 4 days, then cultured with IL-21 and IFNβ in presence of IFNAR1 blocking or control antibody for 3 more days. Representative dot plot of *Tlr7*-reporter expression (g) and frequency of reporter expression (h) among CD19<sup>+</sup>CD138<sup>-</sup> or CD19<sup>+</sup>CD138<sup>+</sup> (PBs) cells. Data are representative of at least two independent experiments. Each dot represents an individual female mouse. Data are presented as mean ± SD. For (b), Two-way ANOVA was performed followed by Tukey's multiple-comparisons test. For (d), Mann-Whitney test was used and for (f) One-way ANOVA was performed followed by Tukey's multiple-comparisons test. For (h), two independent experiments were pooled and a two-way ANOVA (Geisser–Greenhouse corrected) followed by Sidak's multiple comparisons test, was performed



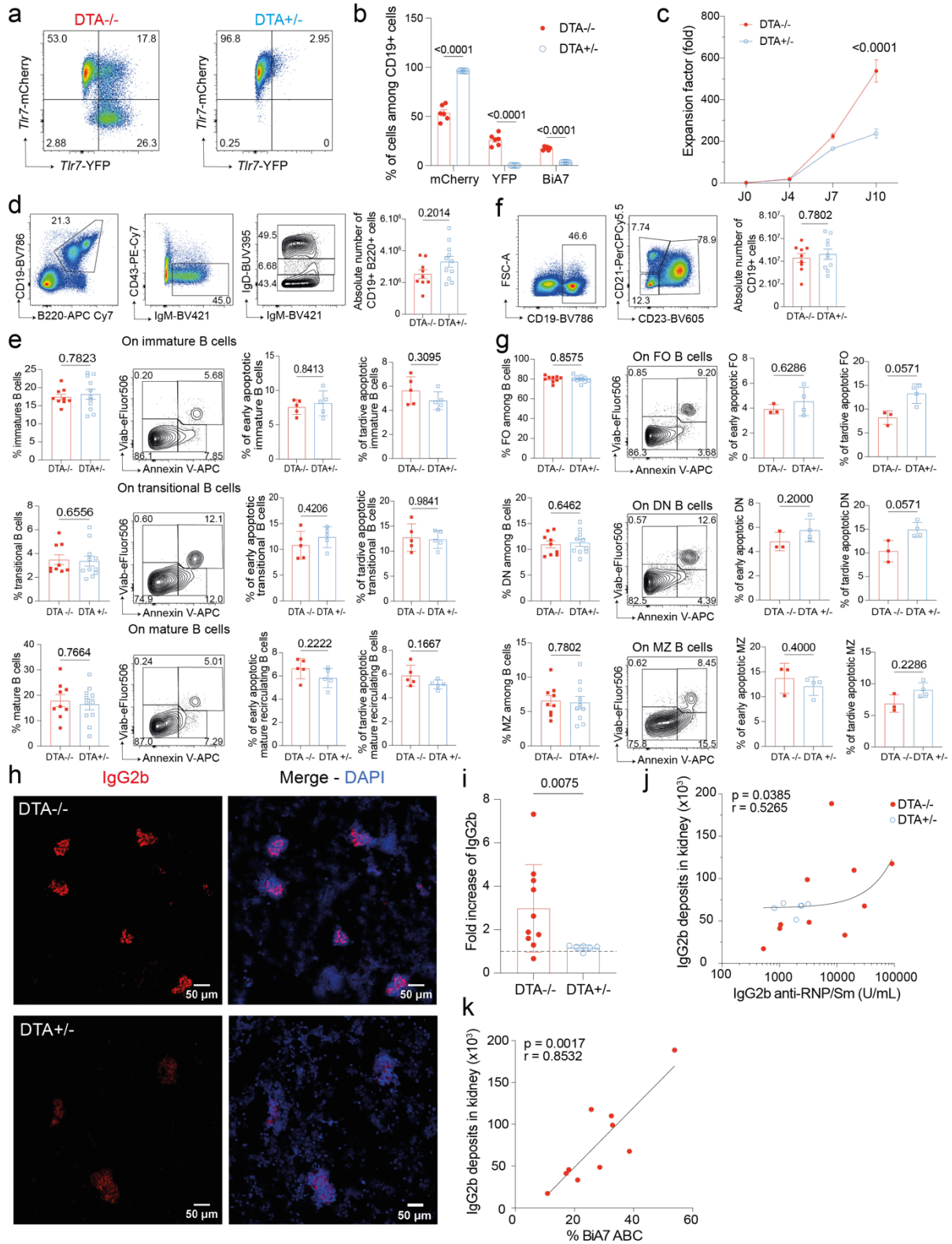


**Extended data Figure 8: MyD88-signaling is dispensable for BiA7 B cell development *in vitro*.** (a,b) Cell cycle analysis with Ruby staining of mix of 1/4 male (CD45.1/CD45.1) and 3/4 female (CD45.1/CD45.2) B cells in iGB cell cultures. Flow cytometry plot showing the gating strategy at d+7 (a). Analysis of DNA content in the indicated B cell using Ruby staining, and frequency of cells in each cell-cycle phases among male, MonoA7 and BiA7 female CD138<sup>-</sup> iGB cells at d+7 (b). (c-f) Analysis of MyD88<sup>+/+</sup>, MyD88<sup>+/-</sup> or MyD88<sup>-/-</sup> purified B cells from dual reporter iGB cultures. Representative dot plots of *Tlr7*-reporter expression gated on CD19<sup>+</sup>CD138<sup>-</sup> B cells and frequency of BiA7 cells at d4 (c,d) or d+7 (e,f). (g) CTV dilution profile and CTV GMFI among CD19<sup>+</sup>CD138<sup>-</sup> B cells from dual reporter MyD88<sup>-/-</sup> mice. (h) Frequency of BiA7 B cells populations over cell division from MyD88<sup>+/+</sup>, MyD88<sup>+/-</sup> and MyD88<sup>-/-</sup> dual reporter mice. Data are representative of at least two independent experiments. Each dot represents an individual mouse. Data are presented as mean ± SD. For (b) One-Way ANOVA was used with Tukey statistical hypothesis. For (d,f), two independent experiments were pooled and Two-way ANOVA was performed followed by Tukey's multiple comparisons test. For (g), paired RM one-way ANOVA (Geisser–Greenhouse corrected) was performed followed by Holm–Sidak multiple comparisons test.



**Extended data Figure 9: Impact of MyD88-deficiency on BiA7 B cell development among splenic B cell subsets.** (a-b) Flow cytometry analysis of spleens from *Tlr7*-dual reporter mice harvested 2 weeks after i.v. injection of  $10^7$  ifu of AdV encoding either type I IFN (IFN- $\alpha$ ) or Luciferase as control. Representative flow cytometry plots showing *Tlr7* reporter expression in FO, MZ and DN B cells subsets (a), frequency of each B cell subsets among CD19<sup>+</sup> cells and of BiA7 among each B cell subsets (b). (c-h) Flow cytometry analysis of splenic B cell subsets from *Rag2*-deficient mice sub-lethally irradiated and reconstituted with BM from either WT, MyD88<sup>+/-</sup> or MyD88<sup>-/-</sup> *Tlr7*-dual reporter mice, and i.v.

injected with AdV-IFN- $\alpha$  for 2 weeks. Representative flow cytometry plots (**c**) and absolute numbers (**d**) of FO, MZ and DN B cells subsets. (**e-f**) Histograms of individual GMFI of B cell markers gated on MonoA7 or BiA7 FO IgD<sup>+</sup> cells in reconstituted-MyD88<sup>+/+</sup> (**e**) or MyD88<sup>-/-</sup> (**f**) mice. (**g,h**) MonoA7 (either YFP or mCh) and BiA7 CD19<sup>+</sup> IgD<sup>+</sup> sorted B cells from MyD88<sup>+/+</sup> *Tlr7*-dual reporter chimeric mice were *in vitro* stimulated for 3 days with F(ab')<sub>2</sub> goat anti-mouse IgM, anti-CD40 and CL097. (**g**) Representative CD19 CD138 staining at day 3 and (**h**) frequency of *in vitro* differentiated CD19<sup>+</sup>CD138<sup>+</sup> PBs. Data are representative of 2-5 independent experiments. Each dot represents an individual female mouse. Data are presented as mean  $\pm$  SD. For (**b**), a Mann-Whitney test was used. For (**d**), a Kruskal-Wallis test followed by Dunn's multiple comparisons test was used. For (**e, f**), a paired one-way ANOVA followed by Tukey's multiple comparisons test and for(**h**), a paired one-way ANOVA followed by Sidak's with Sidak multiple comparisons test was used.



**Extended data Figure 10: Enforced *Tlr7* mono-allelism has no significant impact on B cell development but protects from immunoglobulin deposits in the glomeruli in pristane-induced lupus-like disease. (a-b)** Visualization of *Tlr7*-reporter activity in CD19<sup>+</sup> cells (a) and frequency of MonoA7 and BiA7 among CD19<sup>+</sup> cells (b) from DTA<sup>-/-</sup> and DTA<sup>+/-</sup> dual reporter mice cultured on 40LB stromal cells for 7 days (step1 no IFN $\beta$ ; step 2 with IFN $\beta$ ). (c) Cumulative fold increase in the number of CD19<sup>+</sup> iGB cells after step 1 & 2, and an additional culture with IL-21 and IFN- $\beta$  from day 7 to 10. (d-g) Flow cytometry analysis of bone marrow (d-e) and splenic (f-g) B cell populations from DTA<sup>-/-</sup>

and DTA<sup>+/-</sup> dual reporter mice. **(d)** Gating strategy for immature, transitional and mature B cells and absolute number of CD19<sup>+</sup> B220<sup>+</sup> cells in the bone marrow. **(e)** Frequency of immature, transitional and mature B cells among CD19<sup>+</sup>B220<sup>+</sup>B cells. Representative dot plot of annexin V/viab. dye staining profiles. Frequency of early and tardive apoptotic among immature, transitional and mature B cells. **(f)** Gating strategy of splenic FO, DN and MZ B cells and absolute number of CD19<sup>+</sup> cells. **(g)** Frequency of FO, DN and MZ B cells, representative dot plot of annexin V/viab. dye staining and frequency of early and tardive apoptotic among FO, DN and MZ CD19<sup>+</sup> B cells. **(h,i)** Kidney analysis 7 months post pristane injection of female DTA<sup>-/-</sup> and DTA<sup>+/-</sup> dual *Tlr7*-reporter mice. **(h)** Representative images of kidney sections stained for IgG2b (red). Nuclei counterstained with DAPI (blue). Scale bars, 50µm. **(i)** Fold increase of IgG2b deposits in pristane injected compare to not injected mice. Each symbol represents the mean of 20-160 glomeruli of individual mouse. **(j,k)** Correlation between the frequency of IgG2b deposits in kidney **(j)** and the titer of IgG2b anti-RNP/Sm in pristane-injected DTA<sup>-/-</sup> and DTA<sup>+/-</sup> mice **(j)** and the frequency of BiA7 IgD<sup>-</sup> ABC cells in pristane-injected DTA<sup>-/-</sup> dual reporter mice **(k)**. For (b-g, i), a Mann-Whitney tests were used. Spearman tests were used for correlations (j) and Pearson test for panel (k). The symbols represent individual mouse; bars represent the mean; the error bars represent the mean±SD. Data are representative of two independent experiments.