

Extended Data for

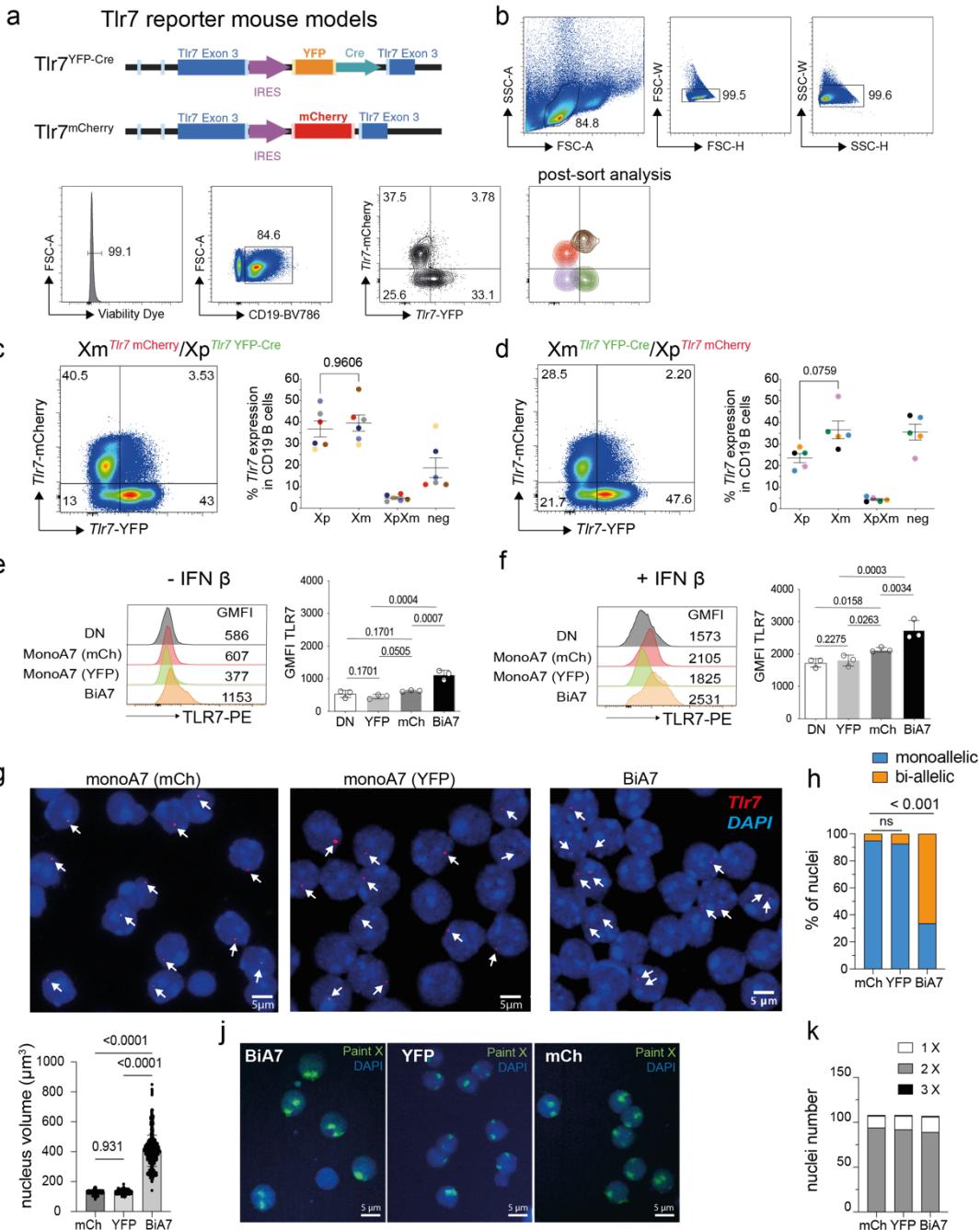
Tlr7-biallelism defines a hyperfunctional state of female B lymphocytes

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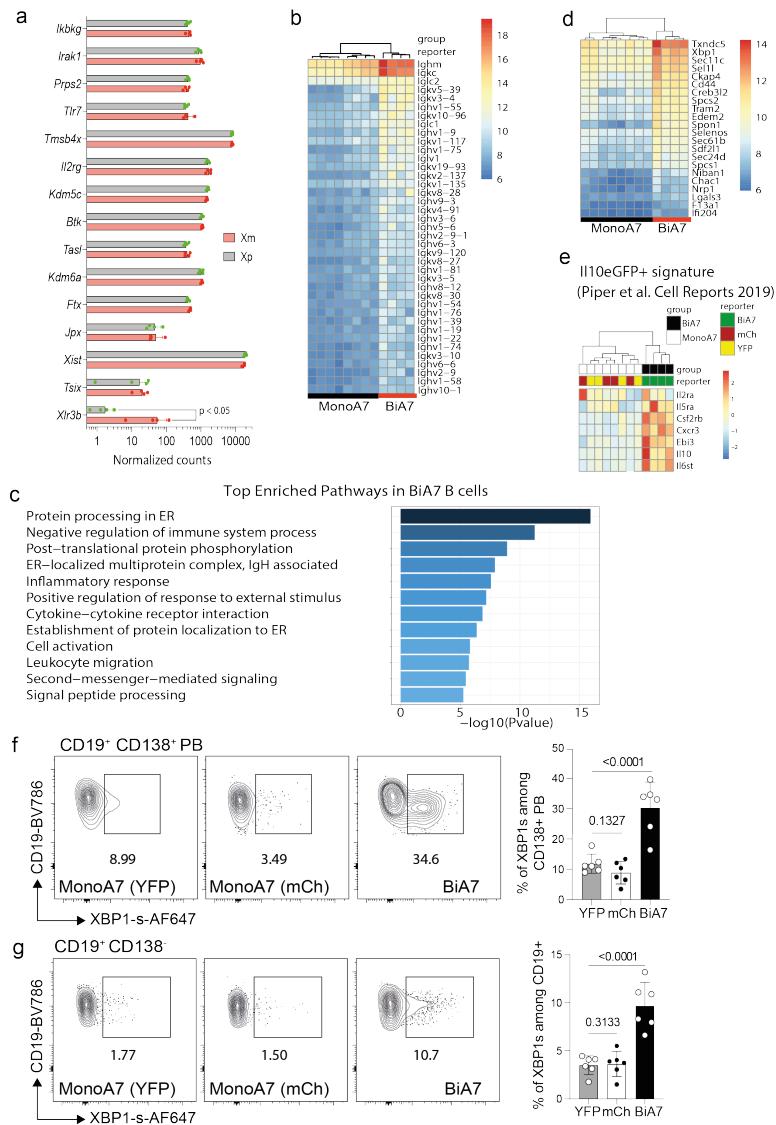
The PDF file includes:

Extended Data Fig. 1 to Fig. 10



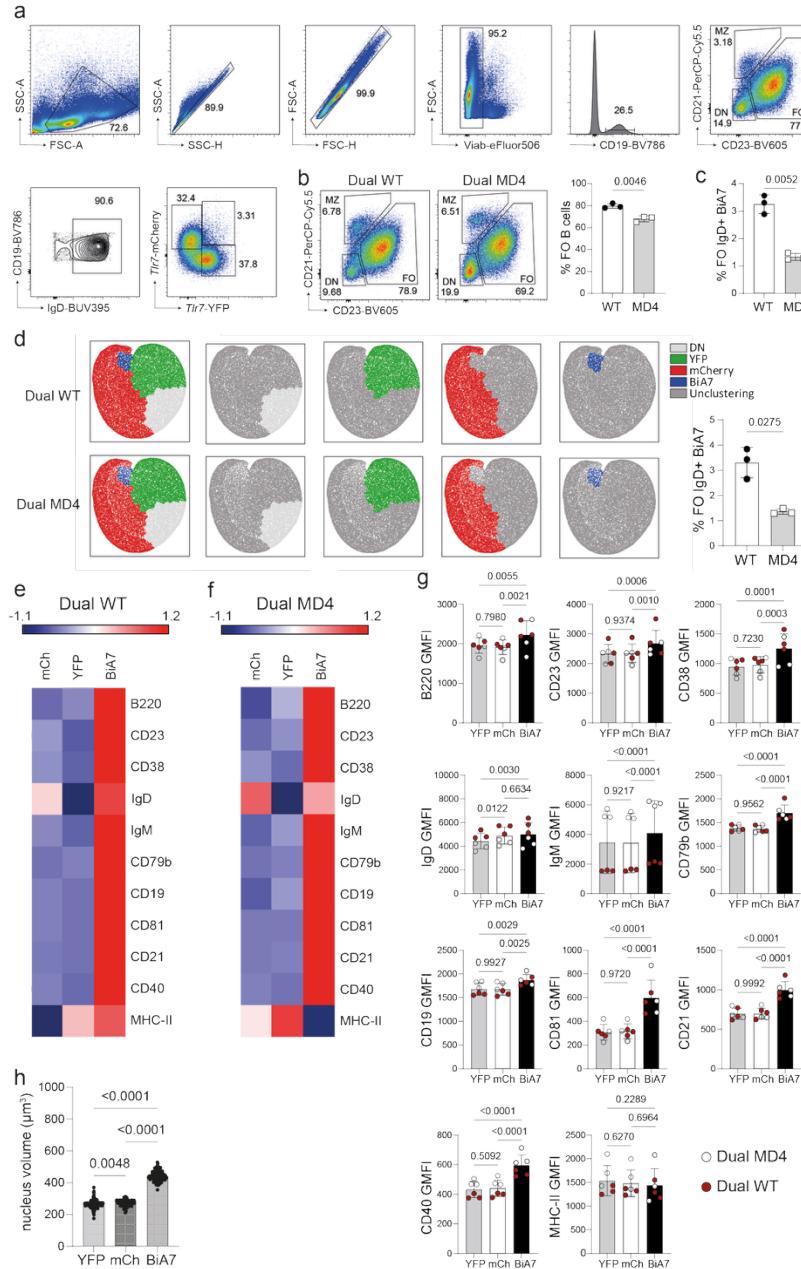
Extended data Figure 1: Characterization of *Tlr7* bi-allelic splenic B cells. (a) Design of *Tlr7*-reporter mice. A reporter cassette including an IRES followed by a fluorescent protein (mCherry or YFP-Cre) was inserted at the 3' end of the endogenous *Tlr7* gene. (b) Gating strategy for cell-sorting of MonoA7 (mCherry or YFP), BiA7 (mCherry⁺/YFP⁺) and double negative (DN) CD19⁺ B cells. Representative flow cytometry purity profile from cell sorting experiment is shown. (c, d) Representative flow cytometry plot and quantification of *Tlr7*-reporter allelic expression from splenic CD19⁺ B cells from Xm^{Tlr7-mCherry}Xp^{Tlr7-YFP-Cre} (c) or the reverse crossing Xm^{Tlr7-YFP-Cre}Xp^{Tlr7-mCherry} (d) female mice (maternal X, Xm; paternal X, Xp). (e,f) GMFI quantification of intracellular TLR7 staining in FACS-sorted DN, MonoA7 and BiA7 CD19⁺ B cells cultured overnight without (e) or with 2 ng/ml IFN- β (f). TLR7 staining expressed as Δ GMFI with isotype control. Quantifications are shown from three independent experiments. (g) Representative RNA FISH images of *Tlr7* primary transcripts.

Nuclei counterstained with DAPI (blue). Scale bars, 5 μ m. (h) Frequency of mono- and bi-allelic RNA FISH signals within the sorted MonoA7 or BiA7 cell populations. (i) Nucleus volume (μ m³) measured by Imaris from MonoA7 and BiA7 B cells. (j) Representative images of paint DNA FISH for the X chromosomes (green). Nuclei counterstained with DAPI (blue). Scale bars, 5 μ m. (k) Quantification of the number of X chromosome territories per nucleus in sorted MonoA7 and BiA7 B cells. Statistical analysis was performed using One-way ANOVA with Tukey multiple-comparison test. For (h) statistical differences were assessed using Fisher's exact test or One-way ANOVA with Tukey multiple-comparison test (i). Exact p values are shown.



Extended data Figure 2: Characterization of the transcriptional signature of BiA7 B cells. RNA-seq data between BiA7 and MonoA7 B cells (n=4/group). (a) Relative expression of the maternally imprinted gene *Xlr3b* compared to 14 representative X-linked genes from individual mice (n=4). Adjusted p value is shown. (b) Heatmap representing variance-stabilized normalized counts for Ig-related genes identified as differentially regulated between BiA7 and MonoA7 samples and having an average expression greater than 200 normalized counts in at least 3 of the 4 BiA7 samples. (c) Metascape analysis showing the top enriched pathways corresponding to the selected cluster summary terms obtained from 504 non-Ig genes with log2FC > 1, pvalue < 0.05 with background: 12563 non-Ig expressed genes. (d) Heatmap showing the unsupervised clustering of the DEG (log2FC>1, adj. p-value < 0.05).

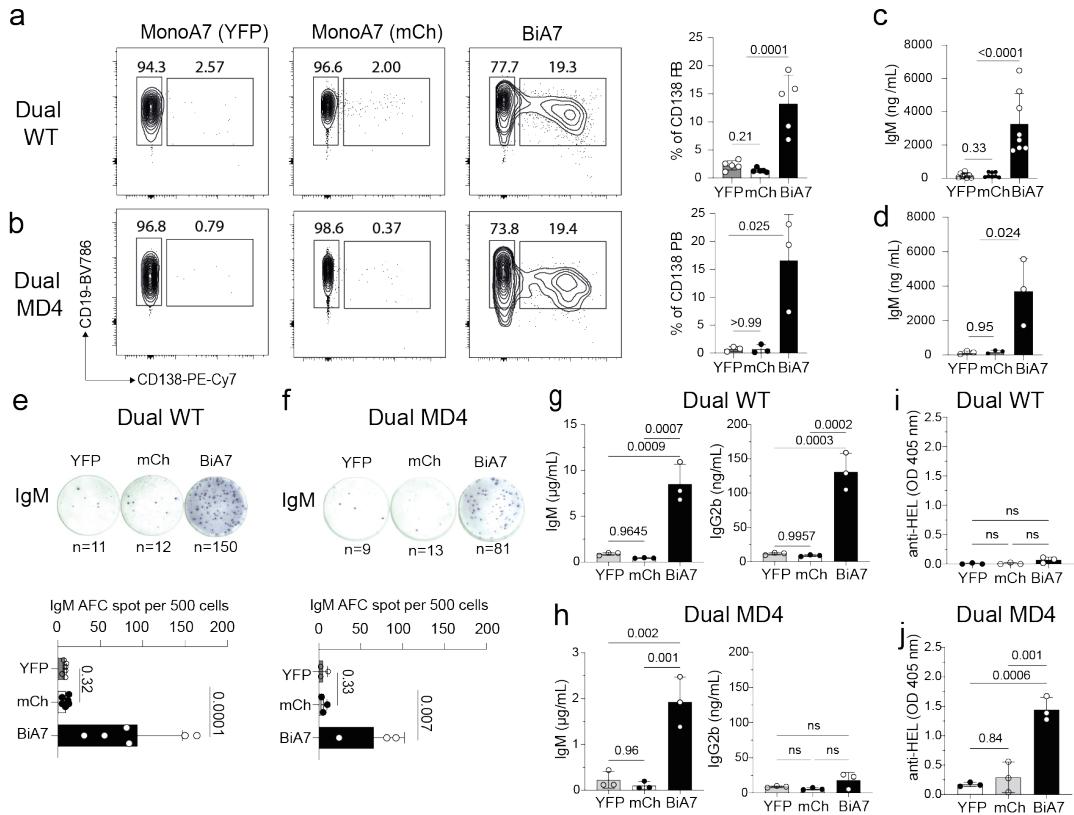
<0.05) from the pathways underlined in bold panel in (c). (e) Heatmap of *Il10eGFP*⁺ signature genes upregulated in BiA7 B cells with $\log_2FC > 1$ adj. p-value <0.05. Heatmap rows ordered based on average expression level in BiA7 samples. (f,g) Intracellular analysis of XBP1s expression on CD19⁺CD138⁺ PB (f) or CD19⁺CD138⁻ B cells (g) from splenic IgD⁺ MonoA7 or BiA7 B cells sorted and stimulated as in Fig. 1g for 3 days. Statistical analysis was performed using One-way ANOVA with Tukey multiple-comparison test. The symbols represent individual mice; the error bars represent the mean±SD.



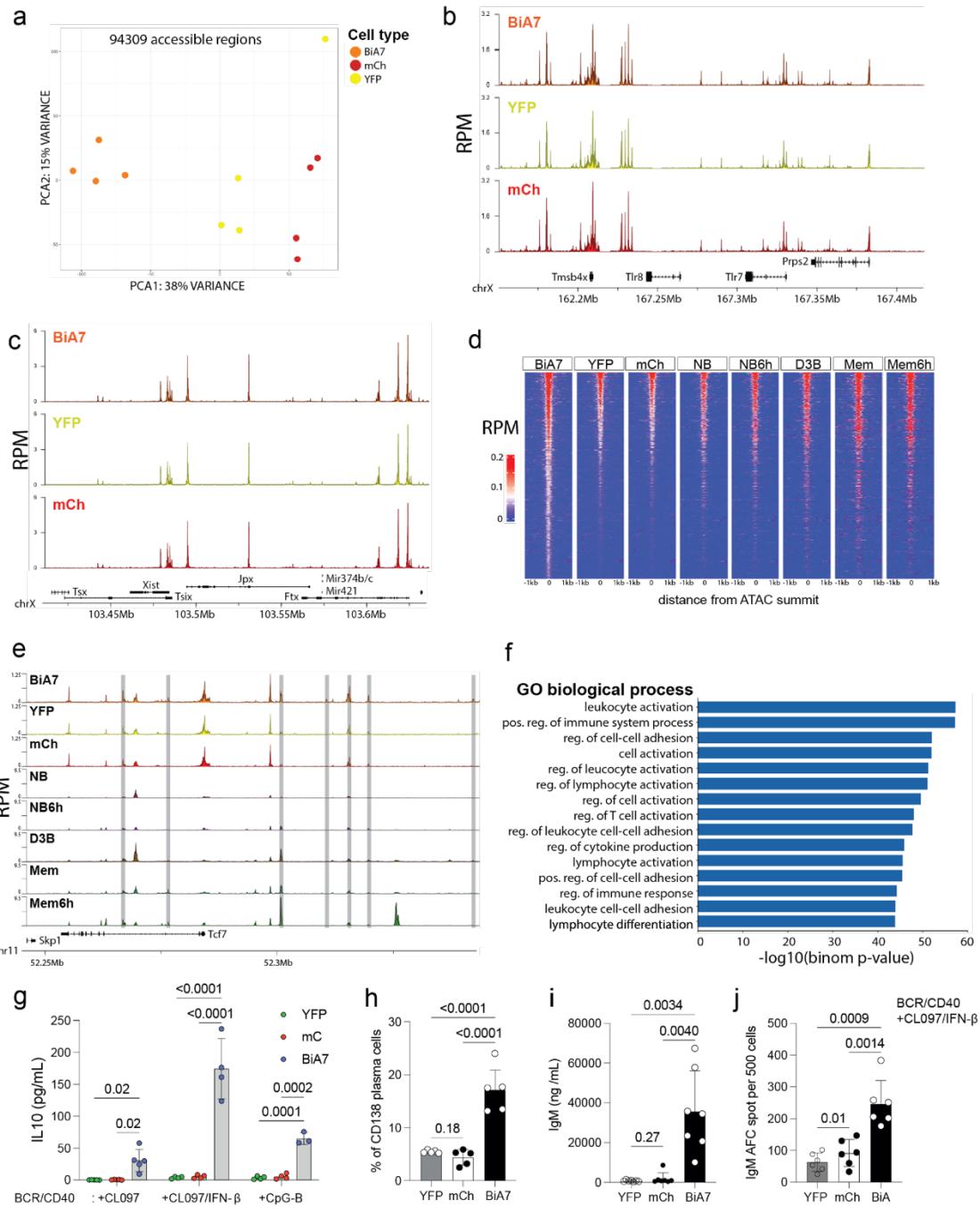
Extended data Figure 3: BCR-specificity does not influence the development of BiA7 B cells.

(a) *Ex vivo* gating strategy on splenocytes from Dual WT and Dual MD4 mice. (b) Representative cytometry plot of DN, MZ and FO splenic B cells profile and the frequency of FO B cells in Dual WT and Dual MD4 mice. (c) Frequency of BiA7 naive FO IgD⁺ B cells in Dual WT and Dual MD4 mice. (d) Reporter-based unsupervised clustering and quantification of BiA7 among FO IgD⁺ cells in Dual WT and Dual MD4 mice. (e,g) Expression of cell surface markers and functional molecules in sorted MonoA7 and BiA7 FO B cells from Dual WT (e) or Dual MD4 (f) mice. (g) Histograms of GMFI of these markers from (e,f). (h) Nucleus volume (μm^3) measured by Imaris in sorted MonoA7 and BiA7 B cells. Statistical analysis was performed using Welch's t test (b,c,d), or One-way ANOVA with Tukey

multiple-comparison test (g,h). The symbols represent individual mice; the error bars represent the mean \pm SD.

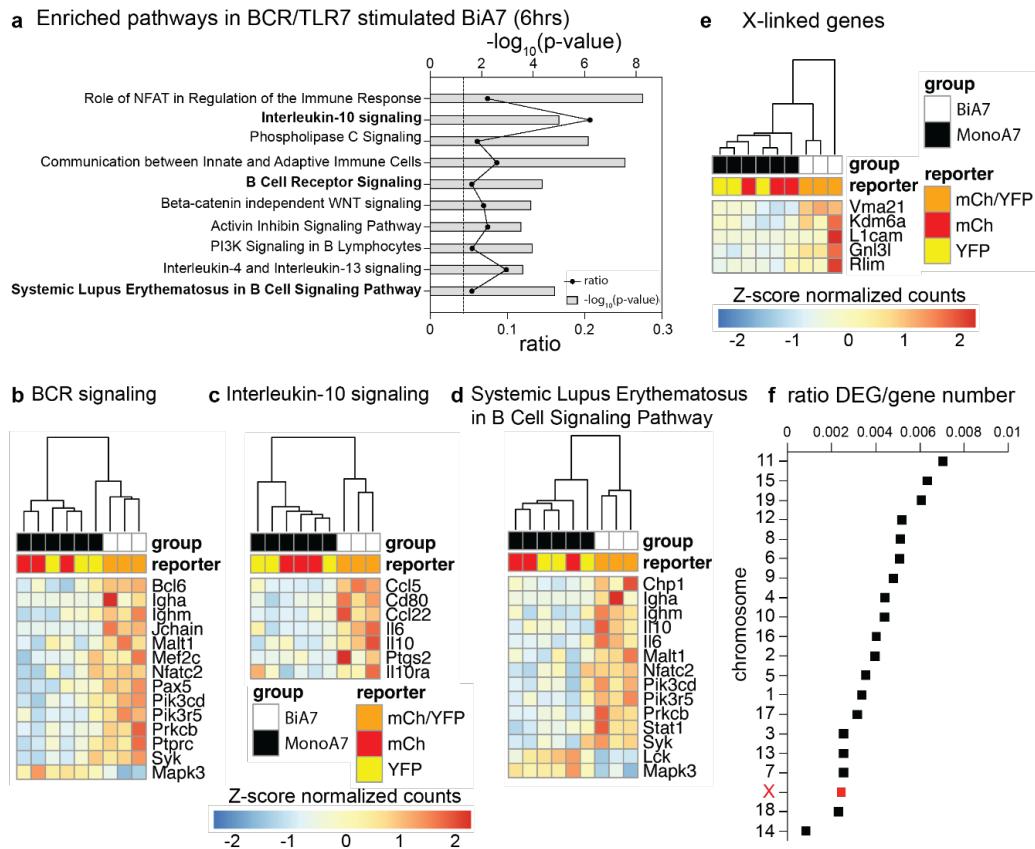


Extended data Figure 4: BiA7 FO B cells with a hyperactive potential developed independently of BCR specificity. (a, b) Representative cytometry plot of CD19⁺CD138⁺ PB on sorted MonoA7 and BiA7 IgD⁺ FO B cells cultured with F(ab')₂ anti-IgM, anti-CD40 and CL097 for 3 days from Dual WT (a) and Dual MD4 (b) mice. Concentration of IgM collected in supernatants of cultured MonoA7 and BiA7 cells from Dual WT (c) and Dual MD4 (d) mice assessed by ELISA. Analysis of antibody forming cells (AFC) secreting IgM antibodies from Dual WT (e) and Dual MD4 (f) assessed by ELISPOT. (g, h) Comparative analysis of the production of IgM, class-switched IgG2b (g, h) and anti-HEL antibody (i, j) between the indicated B cell subsets purified from Dual WT (g, i) or Dual MD4 (h, j) mice. Statistical analyses were performed using a One Way ANOVA and multiple comparison Dunnett's test. The symbols represent individual mice; the error bars represent the mean \pm SD.

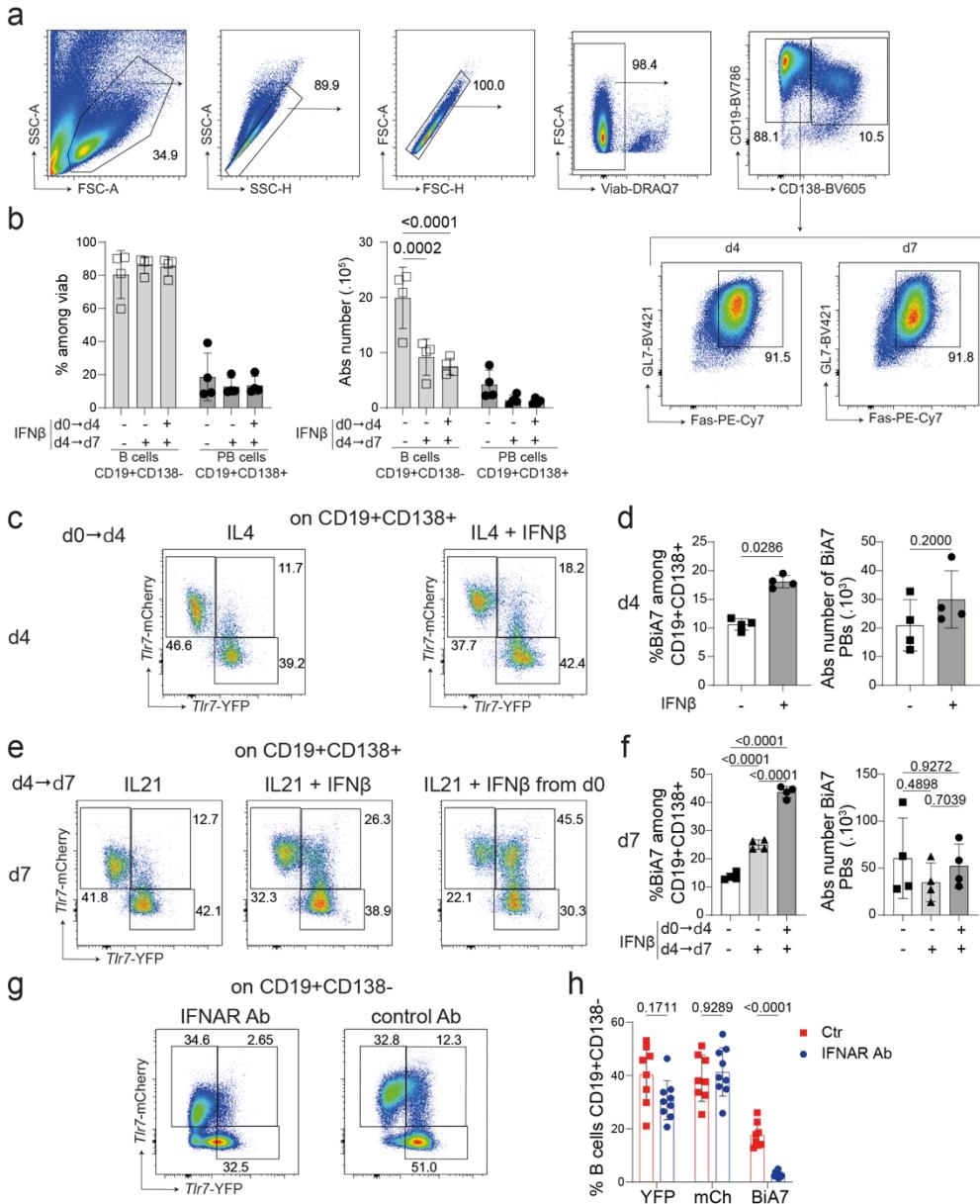


Extended data Figure 5: Accessibility landscape changes in naïve BiA7 B cells. (a) Principal component analysis of chromatin accessibility across 94309 accessible regions between BiA7 and mCh, YFP MonoA7 B cells. (b, c) Chromatin accessibility at the *Tlr7/Tlr8* locus (b) and in the *Xist* region (c) in BiA7 and MonoA7 B cells. (d) Heatmap of chromatin accessibility measured by ATAC-seq across BiA7-specific DARs centered on ATAC-seq summits (+1kb) in BiA7 cells compared to MonoA7 (YFP or mCh) B cells and as well as in naïve (NB), activated (D3B) and memory (Mem) or activated Mem B cells from Q β -VLP immunized mice. (e) Chromatin accessibility at the *Tcf7* locus in the indicated B cell populations. (f) Statistical association of GO biological processes with BiA7-specific DARs. (g) Quantification of IL-10 production in 3 day-culture supernatants of CD19 $^{+}$ IgD $^{+}$ sorted BiA7 and MonoA7 cells stimulated with F(ab') $_{2}$ goat anti-mouse IgM, anti-CD40, and the indicated TLR ligands, either CL097 for TLR7 (alone or in combination with IFN- β) or CpG-B for TLR9. (h-j) Analysis of CD19 $^{+}$ IgD $^{+}$ sorted BiA7 and MonoA7 cells after 3 days of stimulation with F(ab') $_{2}$ goat anti-mouse

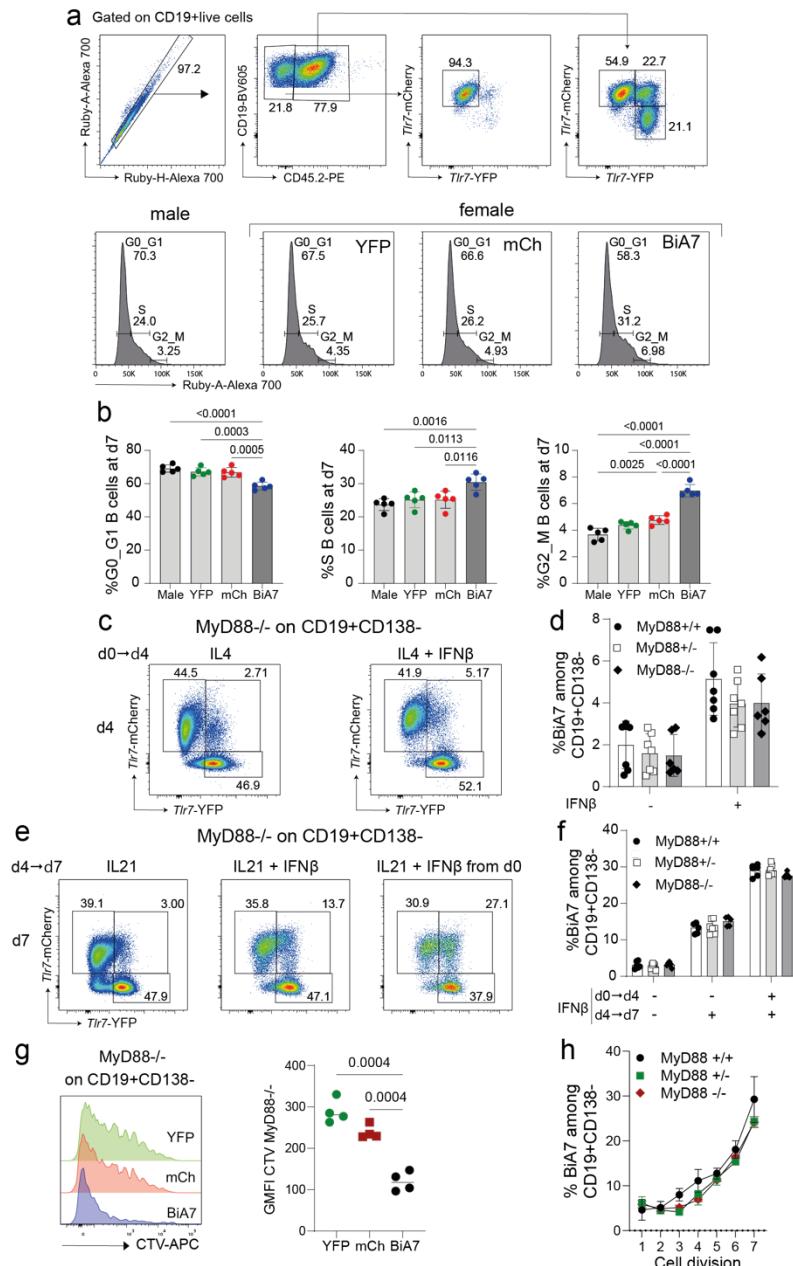
IgM, anti-CD40, CL097 and IFN- β : Frequency of CD19^{int} CD138⁺ PB (h), concentration of IgM (i), and frequency of IgM secreting cells assessed by ELISPOT (j). One-way ANOVA followed by Sidak's multiple comparisons test was used to compare BiA7 cells with the other groups. The symbols represent individual mice; bars represent the mean; the error bars represent the mean \pm SD.



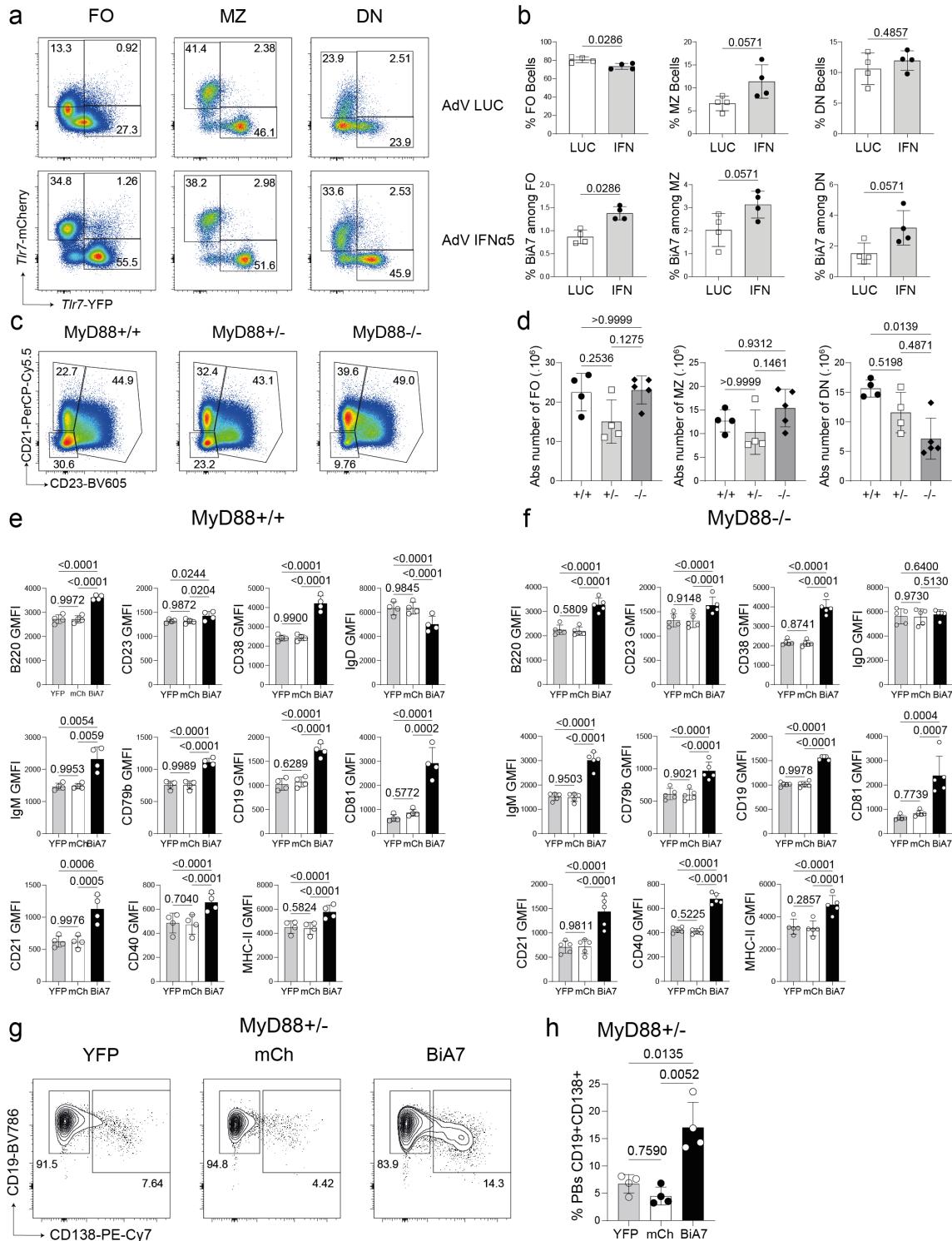
Extended data Figure 6. Early transcriptional changes upon activation of BiA7 B cells. CD19⁺IgD⁺ sorted BiA7 and MonoA7 cells were stimulated with F(ab')₂ goat anti-mouse IgM, anti-CD40, CL097 for 6 hours and processed for RNA-seq analysis. (a) Up-regulated pathways in activated BiA7 B cells identified using ingenuity pathway analysis (IPA, QIAGEN). (b - d) Heatmap representing expression of DEGs from BCR signaling (b), IL10 signaling (c) and from Systemic Lupus Erythematosus in B cell IPA Signaling (d) Pathways in CD19⁺IgD⁺ sorted BiA7 and mCh, YFP MonoA7 activated B cells. (e) expression of X-linked DEGs in BiA7 and MonoA7 activated B cells. (f) DEG/total gene number ratio for each chromosome.



Extended data Figure 7: IFNAR1-signaling is required for the emergence of BiA7 B cells. (a) Gating strategy used for iGB cell analysis by flow cytometry. (b) Frequency and absolute number of CD19⁺CD138⁻ B cells and CD19⁺CD138⁺ PBs in iGB cell culture system with or without IFNβ. (c-f) Representative flow cytometry plots showing *Tlr7*-reporter expression in CD19⁺CD138⁺ PBs at d4 (c) and d7 (e) of iGB cell culture with or without IFNβ. Related histograms showing the frequency and the absolute number of BiA7 B cells among PBs at d4 (d) or d7 (f). (g, h) Analysis of naive B cells from iGB culture with IL-4 alone for 4 days, then cultured with IL-21 and IFNβ in presence of IFNAR1 blocking or control antibody for 3 more days. Representative dot plot of *Tlr7*-reporter expression (g) and frequency of reporter expression (h) among CD19⁺CD138⁻ or CD19⁺CD138⁺ (PBs) cells. Data are representative of at least two independent experiments. Each dot represents an individual female mouse. Data are presented as mean \pm SD. For (b), Two-way ANOVA was performed followed by Tukey's multiple-comparisons test. For (d), Mann-Whitney test was used and for (f) One-way ANOVA was performed followed by Tukey's multiple-comparisons test. For (h), two independent experiments were pooled and a two-way ANOVA (Geisser-Greenhouse corrected) followed by Sidak's multiple comparisons test, was performed

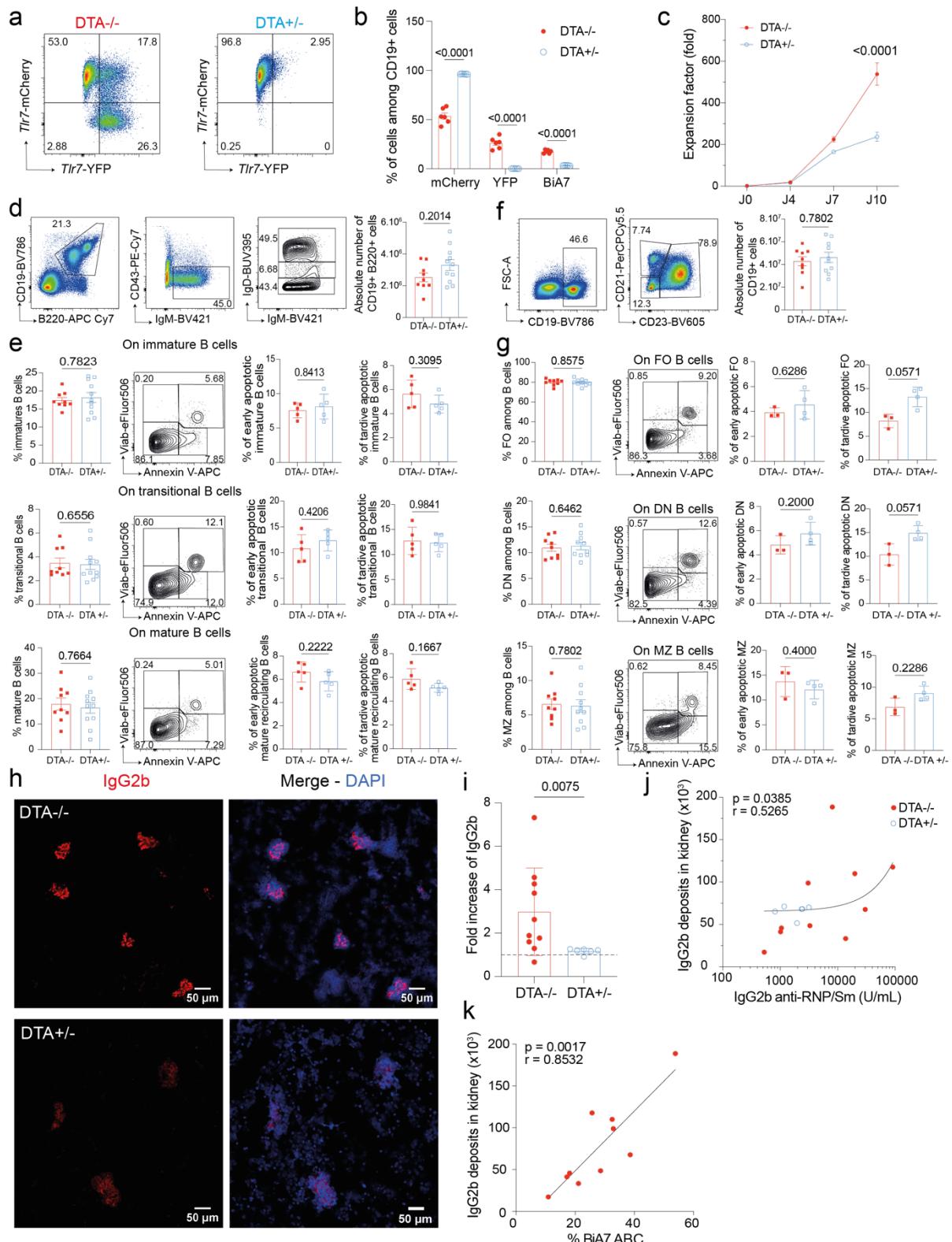


Extended data Figure 8: Myd88-signaling is dispensable for BiA7 B cell development *in vitro*. **(a,b)** Cell cycle analysis with Ruby staining of mix of $\frac{1}{4}$ male (CD45.1/CD45.1) and $\frac{3}{4}$ female (CD45.1/CD45.2) B cells in iGB cell cultures. Flow cytometry plot showing the gating strategy at d+7 **(a)**. Analysis of DNA content in the indicated B cell using Ruby staining, and frequency of cells in each cell-cycle phases among male, MonoA7 and BiA7 female CD138⁺ iGB cells at d+7 **(b)**. **(c-f)** Analysis of MyD88^{+/+}, MyD88⁺⁻ or MyD88^{-/-} purified B cells from dual reporter iGB cultures. Representative dot plots of *Tlr7*-reporter expression gated on CD19⁺CD138⁺ B cells and frequency of BiA7 cells at d4 **(c,d)** or d+7 **(e,f)**. **(g)** CTV dilution profile and CTV GMFI among CD19⁺CD138⁺ B cells from dual reporter MyD88^{-/-} mice. **(h)** Frequency of BiA7 B cells populations over cell division from MyD88^{+/+}, MyD88⁺⁻ and MyD88^{-/-} dual reporter mice. Data are representative of at least two independent experiments. Each dot represents an individual mouse. Data are presented as mean \pm SD. For **(b)** One-Way ANOVA was used with Tukey statistical hypothesis. For **(d,f)**, two independent experiments were pooled and Two-way ANOVA was performed followed by Tukey's multiple comparisons test. For **(g)**, paired RM one-way ANOVA (Geisser–Greenhouse corrected) was performed followed by Holm–Sidak multiple comparisons test.



Extended data Figure 9: Impact of MyD88-deficiency on BiA7 B cell development among splenic B cell subsets. (a-b) Flow cytometry analysis of spleens from *Tlr7*-dual reporter mice harvested 2 weeks after i.v. injection of 10^7 ifu of AdV encoding either type I IFN (IFN- α) or Luciferase as control. Representative flow cytometry plots showing *Tlr7* reporter expression in FO, MZ and DN B cells subsets (a), frequency of each B cell subsets among CD19 $^+$ cells and of BiA7 among each B cell subsets (b). (c-h) Flow cytometry analysis of splenic B cell subsets from *Rag2*-deficient mice sub-lethally irradiated and reconstituted with BM from either WT, MyD88 $^{+/-}$ or MyD88 $^{-/-}$ *Tlr7*-dual reporter mice, and i.v.

injected with AdV-IFN- α for 2 weeks. Representative flow cytometry plots (e) and absolute numbers (d) of FO, MZ and DN B cells subsets. (e-f) Histograms of individual GMFI of B cell markers gated on MonoA7 or BiA7 FO IgD $^+$ cells in reconstituted-MyD88 $^{+/+}$ (e) or MyD88 $^{+/-}$ (f) mice. (g,h) MonoA7 (either YFP or mCh) and BiA7 CD19 $^+$ IgD $^+$ sorted B cells from MyD88 $^{+/-}$ *Tlr7*-dual reporter chimeric mice were *in vitro* stimulated for 3 days with F(ab') $_2$ goat anti-mouse IgM, anti-CD40 and CL097. (g) Representative CD19 CD138 staining at day 3 and (h) frequency of *in vitro* differentiated CD19 $^+$ CD138 $^+$ PBs. Data are representative of 2-5 independent experiments. Each dot represents an individual female mouse. Data are presented as mean \pm SD. For (b), a Mann-Whitney test was used. For (d), a Kruskal-Wallis test followed by Dunn's multiple comparisons test was used. For (e, f), a paired one-way ANOVA followed by Tukey's multiple comparisons test and for(h), a paired one-way ANOVA followed by Sidak's with Sidak multiple comparisons test was used.



Extended data Figure 10: Enforced *Tlr7* mono-allelism has no significant impact on B cell development but protects from immunoglobulin deposits in the glomeruli in pristane-induced lupus-like disease. (a-b) Visualization of *Tlr7*-reporter activity in CD19⁺ cells (a) and frequency of MonoA7 and BiA7 among CD19⁺ cells (b) from DTA^{-/-} and DTA^{+/-} dual reporter mice cultured on 40LB stromal cells for 7 days (step1 no IFN β ; step 2 with IFN β). (c) Cumulative fold increase in the number of CD19⁺ iGB cells after step 1 & 2, and an additional culture with IL-21 and IFN- β from day 7 to 10. (d-g) Flow cytometry analysis of bone marrow (d-e) and splenic (f-g) B cell populations from DTA^{-/-}

and DTA^{+/−} dual reporter mice. (d) Gating strategy for immature, transitional and mature B cells and absolute number of CD19⁺ B220⁺ cells in the bone marrow. (e) Frequency of immature, transitional and mature B cells among CD19⁺ B220⁺ B cells. Representative dot plot of annexin V/viab. dye staining profiles. Frequency of early and tardive apoptotic among immature, transitional and mature B cells. (f) Gating strategy of splenic FO, DN and MZ B cells and absolute number of CD19⁺ cells. (g) Frequency of FO, DN and MZ B cells, representative dot plot of annexin V/viab. dye staining and frequency of early and tardive apoptotic among FO, DN and MZ CD19⁺ B cells. (h,i) Kidney analysis 7 months post pristane injection of female DTA^{+/−} and DTA^{+/−} dual *Tlr7*-reporter mice. (h) Representative images of kidney sections stained for IgG2b (red). Nuclei counterstained with DAPI (blue). Scale bars, 50μm. (i) Fold increase of IgG2b deposits in pristane injected compare to not injected mice. Each symbol represents the mean of 20-160 glomeruli of individual mouse. (j,k) Correlation between the frequency of IgG2b deposits in kidney (j) and the titer of IgG2b anti-RNP/Sm in pristane-injected DTA^{+/−} and DTA^{+/−} mice (j) and the frequency of BiA7 IgD[−] ABC cells in pristane-injected DTA^{+/−} dual reporter mice (k). For (b-g, i), a Mann-Whitney tests were used. Spearman tests were used for correlations (j) and Pearson test for panel (k). The symbols represent individual mouse; bars represent the mean; the error bars represent the mean±SD. Data are representative of two independent experiments.