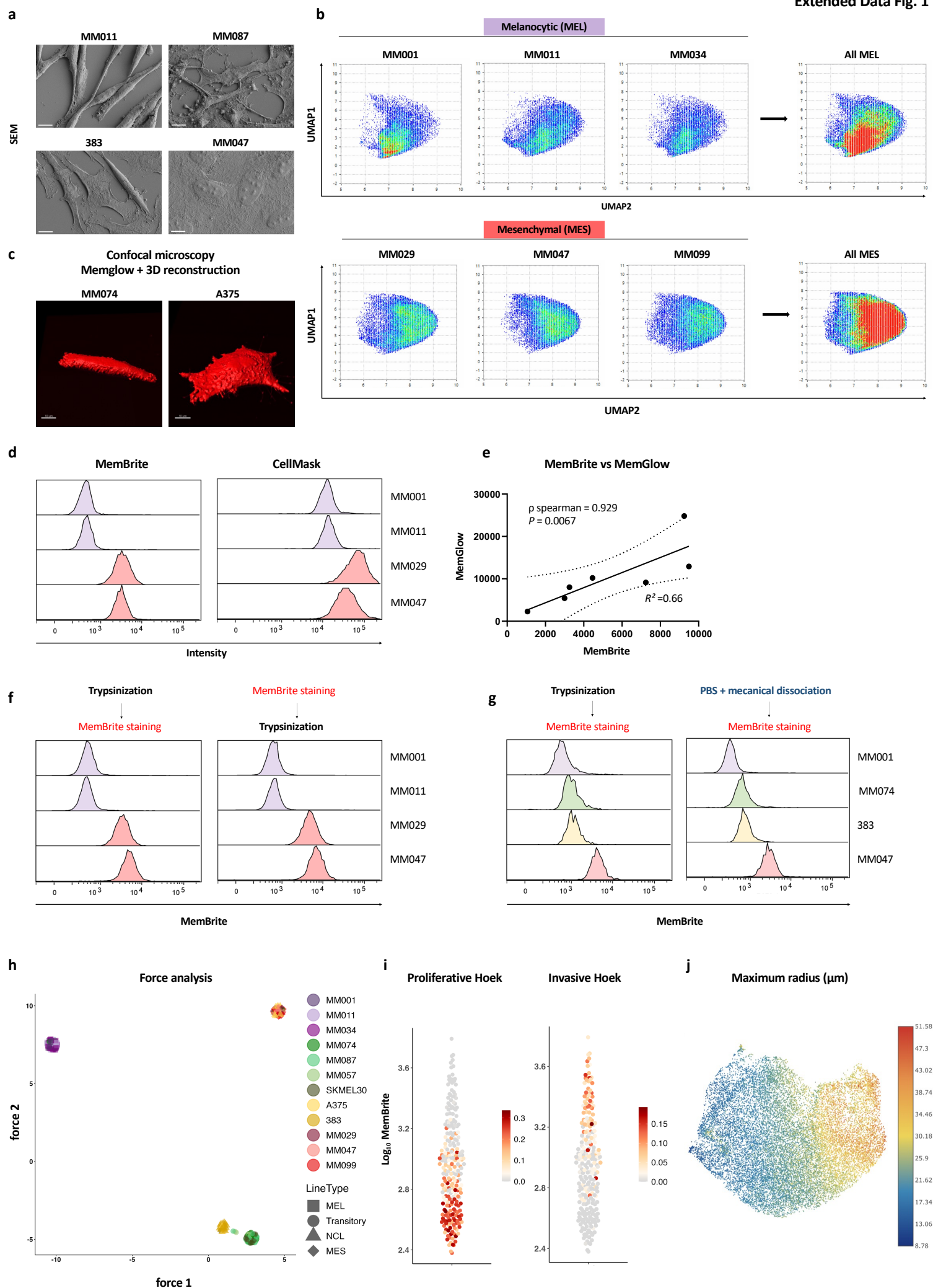
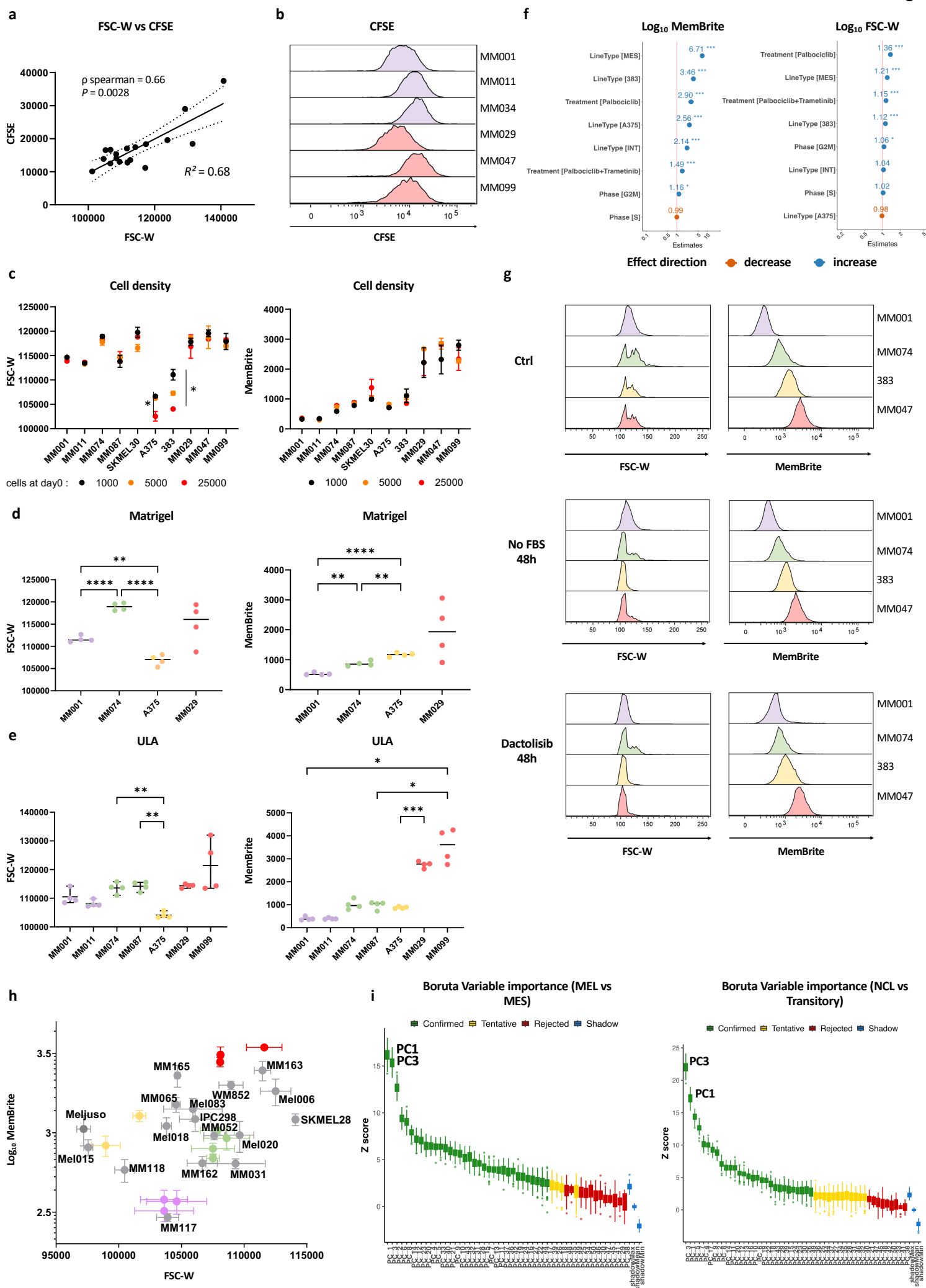




Extended data Fig.1. Morphological and transcriptomic characterization of melanoma states

- a**, Scanning electron microscopy (SEM) images of additional melanoma cell lines corresponding to each of the four untreated transcriptomic states.
- b**, UMAP projection of ThinkCyte data showing distinct clustering of melanocytic (n = 3) and mesenchymal (n = 3) different melanoma lines, representative of two independent experiments.
- c**, Representative reconstructed 3D confocal Z-stacks used for single-cell surface quantification (MM074 and A375).
- d**, Comparison of MemBrite and CellMask fluorescence profiles by flow cytometry in melanocytic (MEL: MM001, MM011) and mesenchymal (MES: MM029, MM047) lines.
- e**, Correlation between MemBrite and MemGlow fluorescence intensities measured by flow cytometry (n = 7 lines; Spearman's correlation coefficient (ρ) and linear regression R^2 are shown).
- f**, Comparison of MemBrite staining performed before versus after trypsinization, analyzed by flow cytometry.
- g**, Comparison of enzymatic versus mechanical dissociation prior to MemBrite labeling.
- h**, Force-directed graph layout identifying four main morphological states under untreated conditions.
- i**, Distribution of transcriptomic state-specific gene signature enrichment along MemBrite fluorescence intensity.
- j**, Maximum cell radius (μm) extracted from Deepcell morphological analysis.

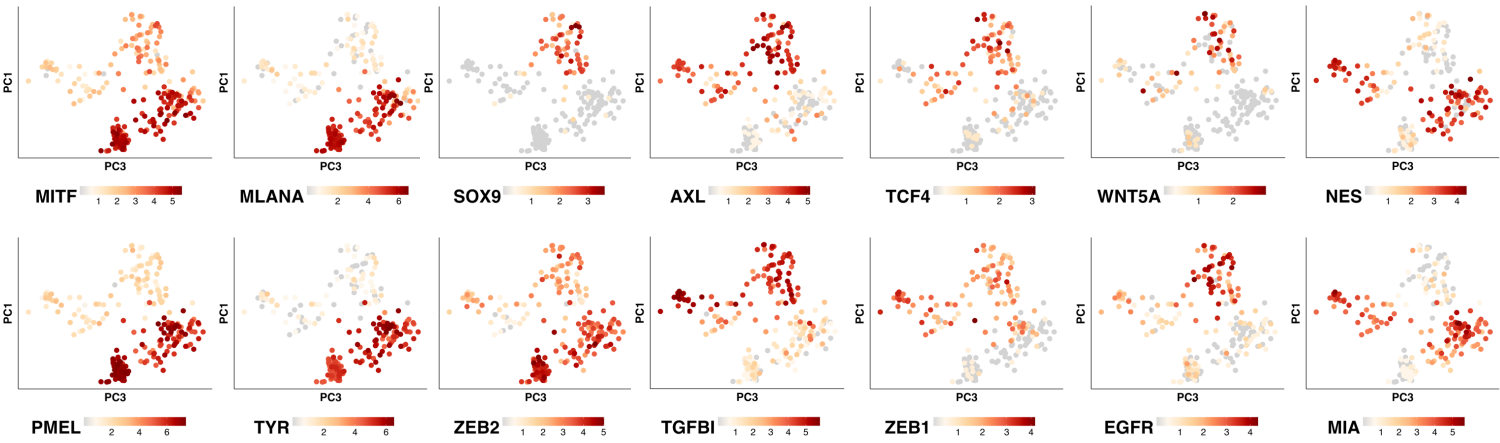


Extended data Fig. 2. Technical parameters influencing cell surface and volume measurements

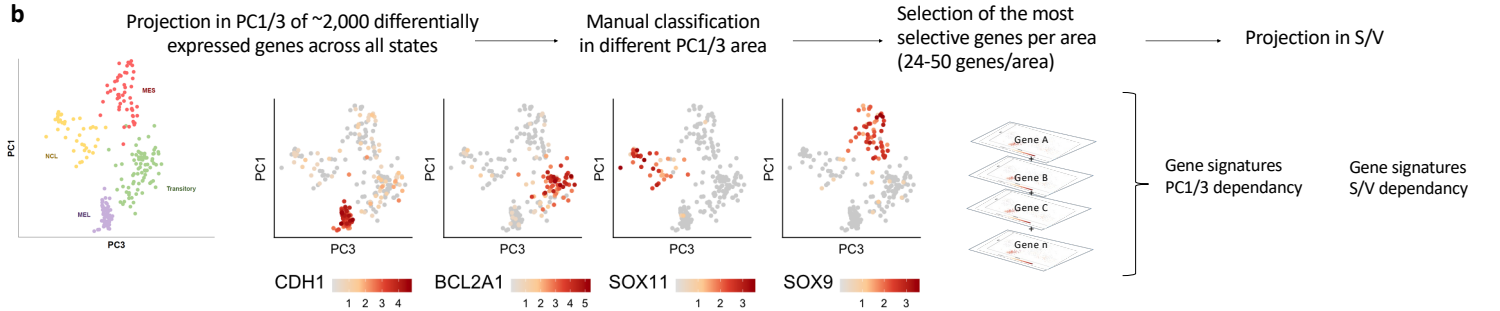
- a**, Correlation between FSC-W and CFSE fluorescence measured by flow cytometry. (n = 18 conditions including untreated and trametinib and/or palbociclib-treated lines).
- b**, CFSE fluorescence intensities between MEL and MES lines.
- c**, Effect of initial plating density (time 0) on FSC-W and MemBrite intensity.
- d**, Influence of Matrigel-based culture versus standard polystyrene on FSC-W and MemBrite intensity.
- e**, Effect of ultra-low attachment (ULA) plates on FSC-W and MemBrite intensity.
- f**, Model estimating the relative contribution of distinct feature classes to FSC-W and MemBrite readouts. Statistical significance for each term was determined using type II Wald tests.
- g**, Representative histograms showing the impact of cell cycle arrest (induced by FBS deprivation, dual PI3K/mTOR inhibition with dactolisib for 48 h, or palbociclib for 24 h) on FSC-W and MemBrite.
- h**, Median MemBrite and FSC-W values across additional melanoma cell lines (n = 12) and PDX-derived lines (n = 5); coloured dots represent reference lines (n = 12).
- i**, Boruta feature-selection analysis combined with principal component analysis (PCA) identifying PC1 and PC3 as the dominant axes discriminating MES/MEL (surface axis) and NCL/transitory (volume axis) states.

Statistical analyses: Welch's ANOVA followed by Dunnett's multiple comparison test was used for panels c-e (*P < 0.05; **P < 0.01; ***P < 0.001; ****P < 0.0001). Spearman's correlation coefficient (ρ) and linear regression R^2 are reported in panel a.

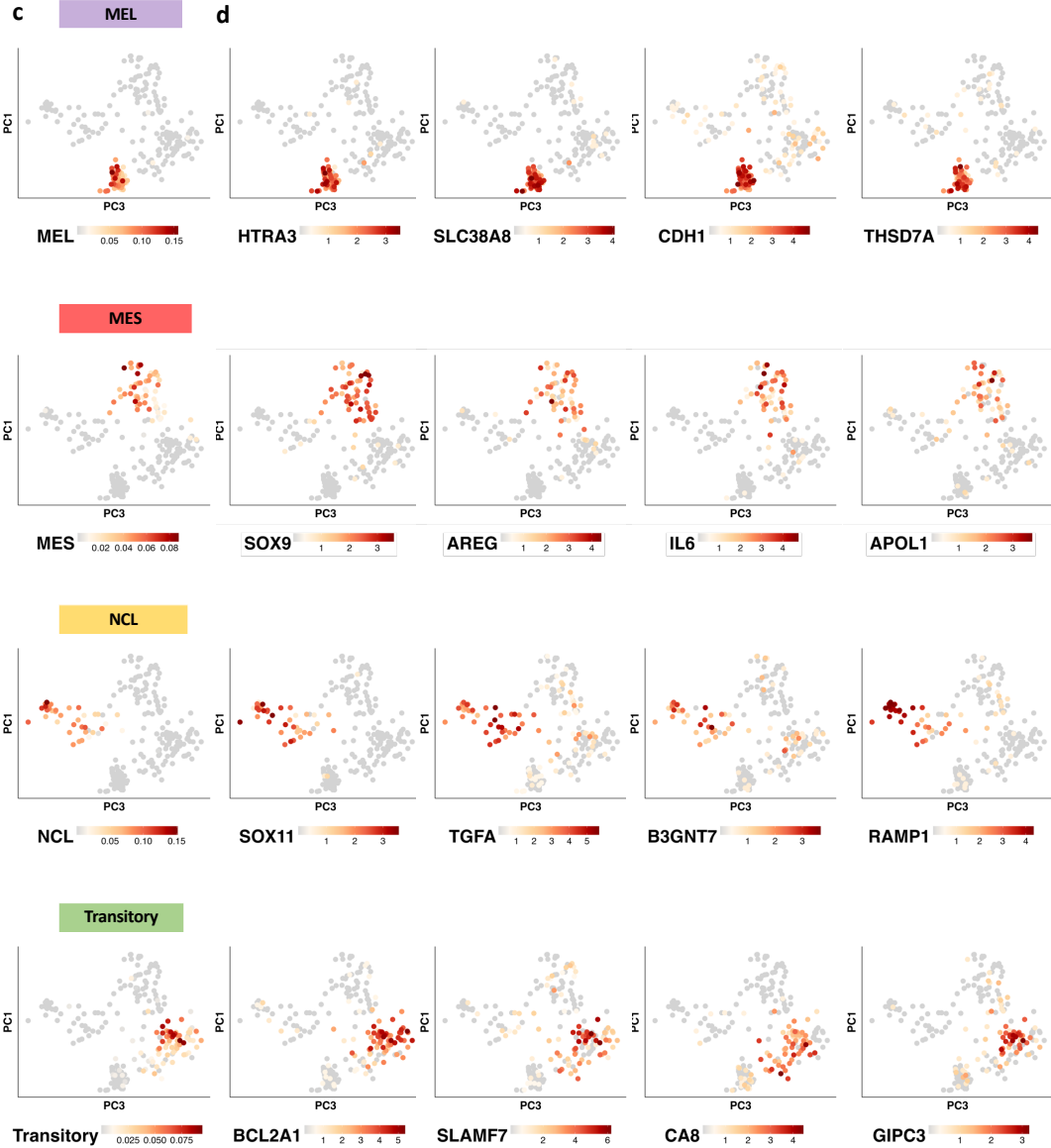
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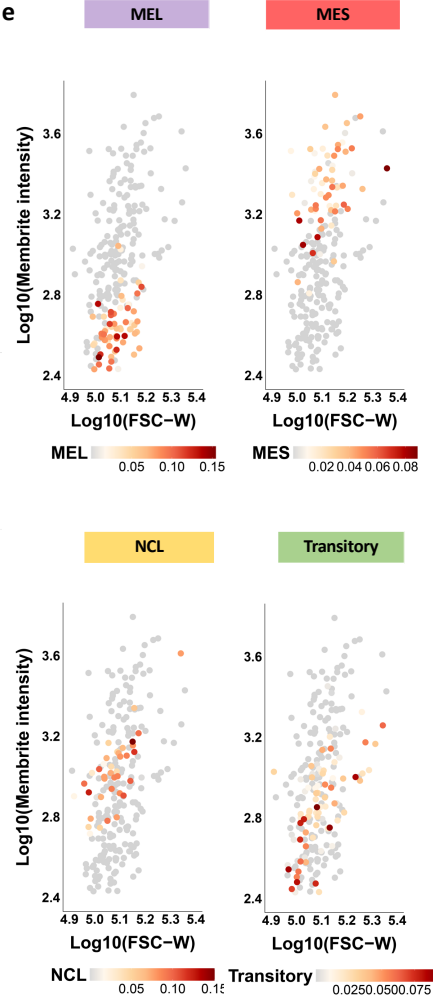
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Extended data Fig. 3. Transcriptomic signatures projected onto the S/V landscape.

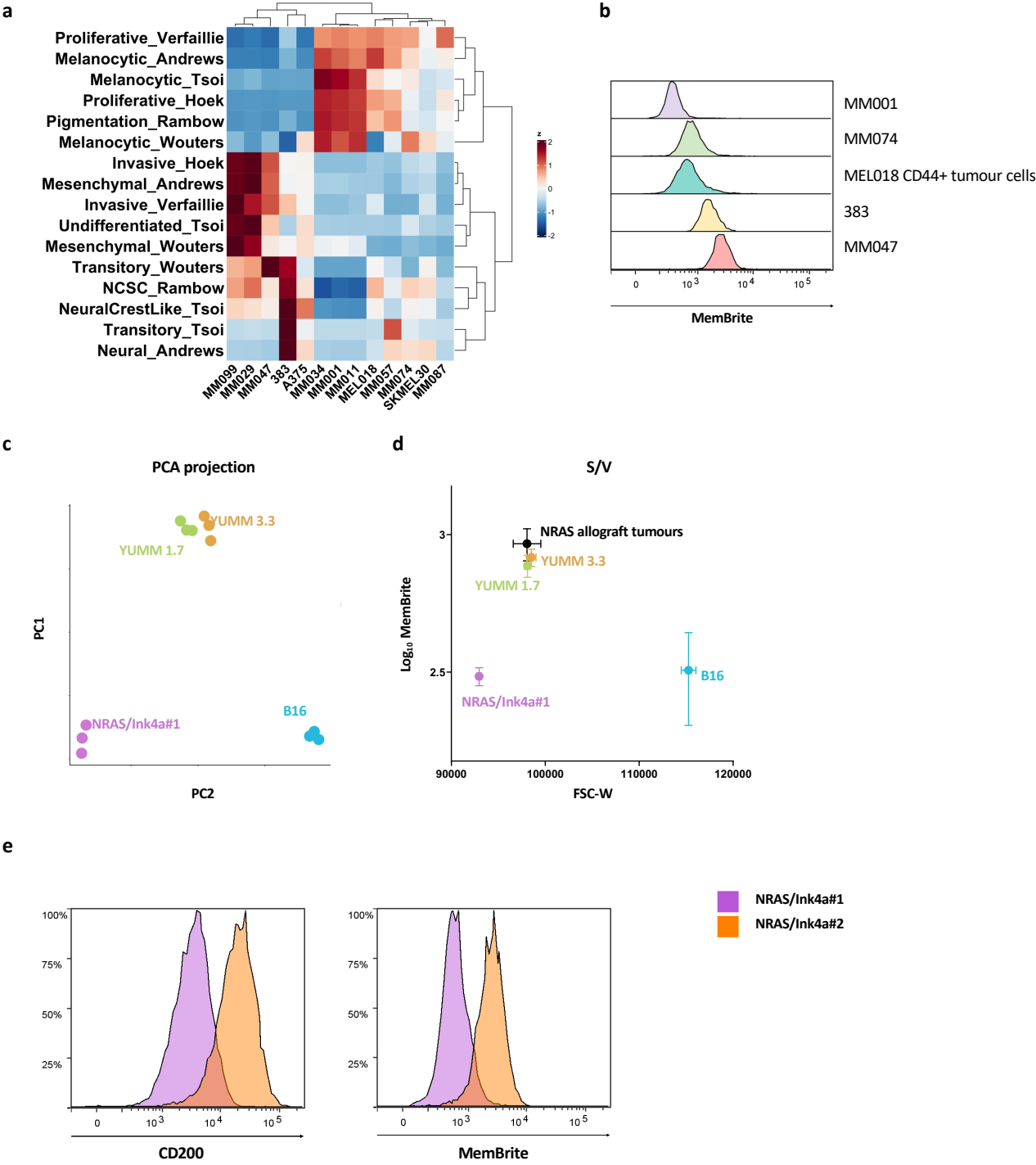
a, Projection of established melanoma state marker genes onto the PC1/PC3 space derived from scRNA-seq data.

b, Schematic illustrating the strategy used to define transcriptional signatures associated with specific PC1/PC3 regions, which correspond to distinct S/V positions.

c, Mapping of curated gene sets representative of melanocytic (MEL, n = 26 genes), mesenchymal (MES, n = 46), neural crest-like (NCL, n = 24) and transitory (n = 35) states onto the PC1/PC3 space.

d, PC1/PC3 projection of four representative genes from the newly defined transcriptional signatures.

e, Visualization of the resulting state-specific gene signatures projected across the S/V landscape.



Extended data Fig. 4. Morphological and transcriptomic characterization of melanoma states

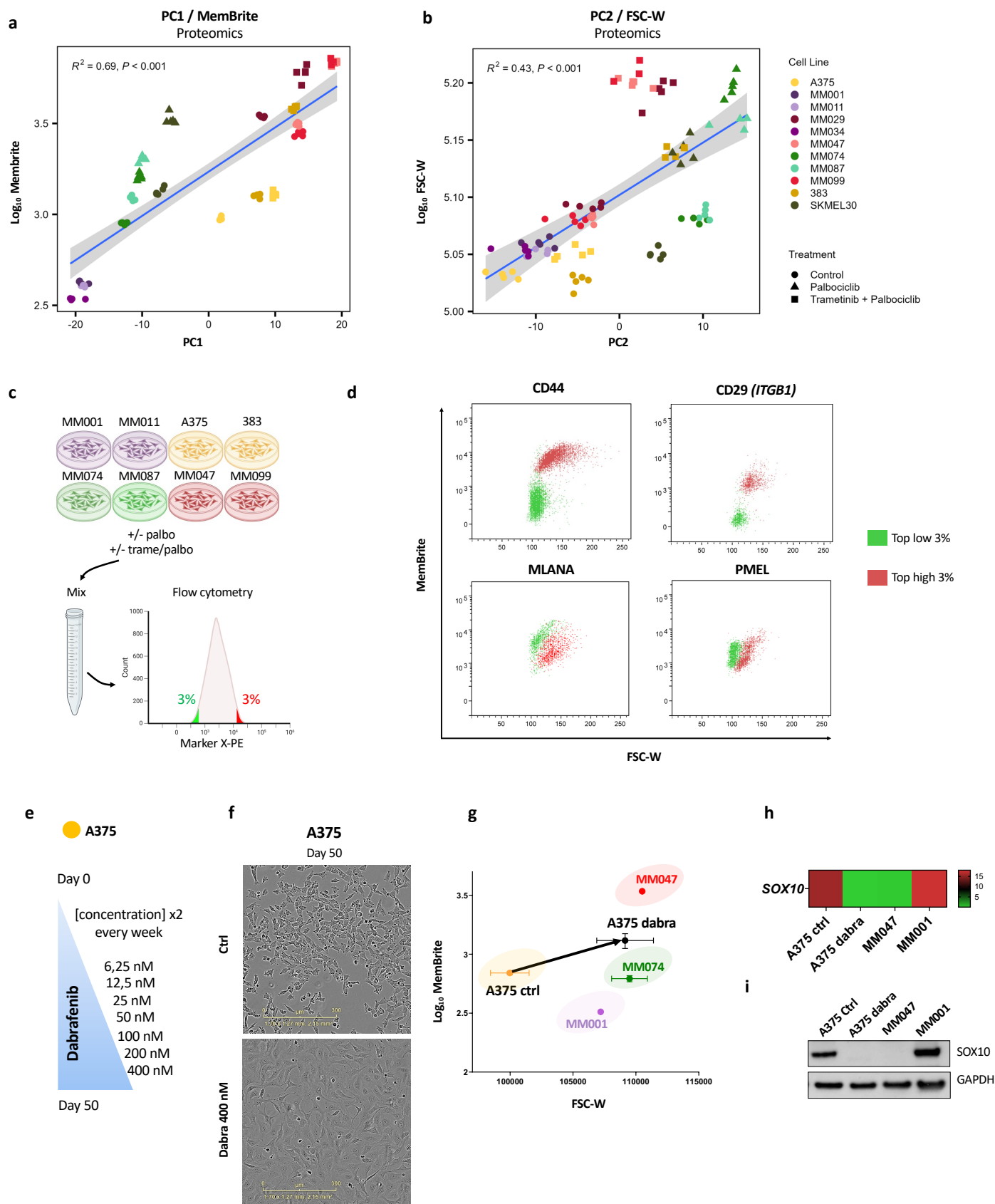
a, Heatmap from scRNA-seq data including the untreated Mel018 PDX sample and additional human melanoma cell lines.

b, Representative flow cytometry histogram of MemBrite fluorescence in CD44⁺ Mel018 tumour cells compared with reference human melanoma lines.

c, Principal component analysis (PC1-PC2) of bulk RNA-seq data from murine melanoma lines (Nras;Ink4a#1, B16, YUMM1.7, and YUMM3.3). Each dot represents a biological replicate.

d, Median log₁₀ MemBrite intensity and FSC-W values in Nras;Ink4a allograft tumours (n=4 biological replicates) compared with reference murine melanoma lines.

e, Differential expression of CD200, a neural crest-like-enriched marker¹⁸, across Nras;Ink4a allograft-derived murine cell lines (Nras;Ink4a #1 and #2).



Extended data Fig. 5. Proteomic profiling reveals patterns consistent with transcriptomic and S/V-based classifications.

a, Linear regression between PC1 from proteomic analysis and $\log_{10}$ MemBrite fluorescence intensity measured from index-sorted cell samples.

b, Linear regression between PC2 from proteomic analysis and FSC-W from index-sorted cell samples.

c, Schematic representation of the workflow used to identify protein markers enriched in specific regions along the S/V continuum.

d, Projection of the top 3% highest and lowest expressing cell populations for selected protein markers across the S/V landscape.

e, Experimental design of a long-term treatment assay (day 0-50) in A375 cells exposed to escalating doses of dabrafenib, with concentrations doubled weekly (adapted from ⁵⁸).

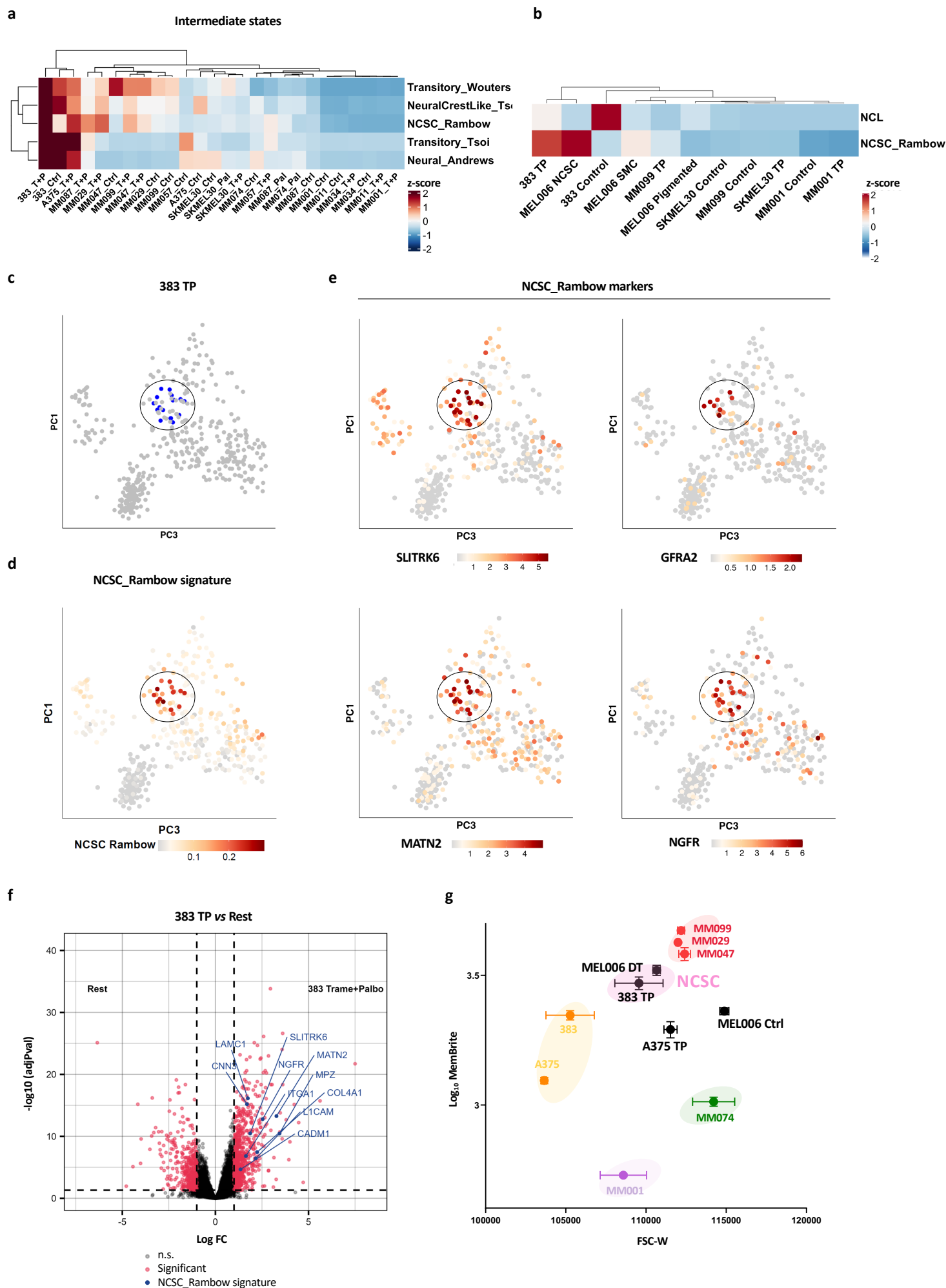
f, Representative incucyte images of A375 cells at day 50 under control and dabrafenib-treated conditions (dabra 400 nM).

g, Median $\log_{10}$ MemBrite intensity and FSC-W values for A375 cells treated with dabrafenib for 50 days ($n = 6$ biological replicates), compared with reference melanoma lines.

h, Quantitative PCR analysis of A375 control and dabrafenib-treated cells compared to MEL and MES reference lines. $n = 3$ biological replicates per condition, each performed in technical triplicate; median values are shown.

i, Western blot analysis of A375 control and dabrafenib-treated cells alongside MEL and MES reference lines, representative of two independent sample preparations.

Illustrations in c were created using BioRender (<https://biorender.com>).



Extended data Fig. 6. Therapy-induced emergence of a NCSC morphostate

a, Heatmap showing enrichment of transitory, neural crest-like (NCL), and neural crest stem-like (NCSC) transcriptional signatures in 383 cells treated with TP.

b, Integrated scRNA-seq analysis combining 383 TP cells, additional melanoma lines, and Mel006 minimal residual disease (MRD) samples collected in vivo at day 20, along with previously defined subpopulations (NCSC, SMC, pigmented)⁶.

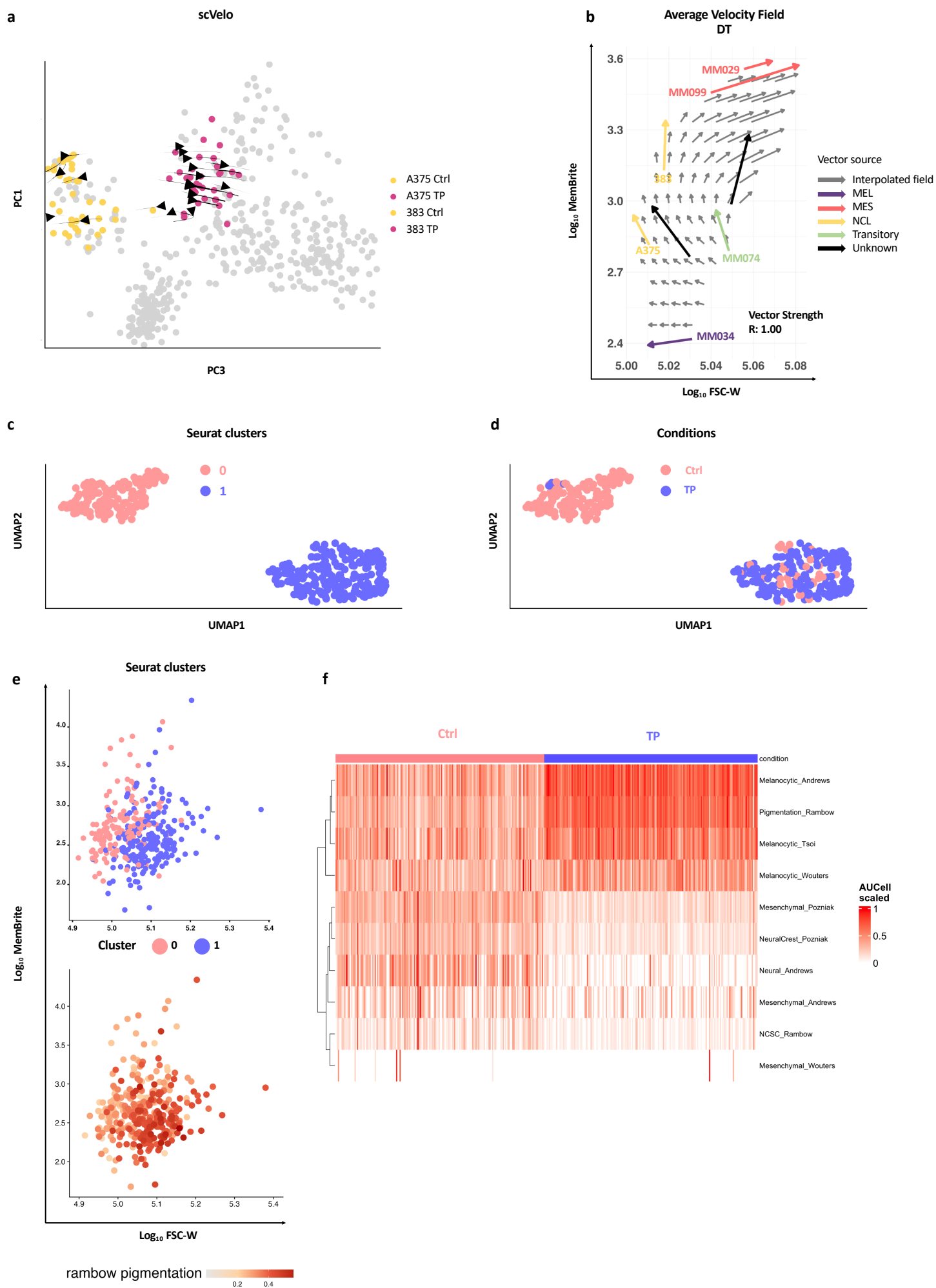
c, PC1-PC3 projection of 383 TP cells across untreated and treated melanoma scRNA-seq datasets.

d, Projection of the NCSC_Rambow signature⁶.

e, PC1-PC3 projection of representative genes defining the NCSC transcriptional program as previously described⁶.

f, Proteomic analysis showing enrichment of NCSC_Rambow associated markers in 383 TP cells compared to all other clusters.

g, S/V distributions of 383 TP and Mel006 cells treated with dabrafenib (40 nM) and trametinib (8 nM) (DT) for 4 days. N = 3 biological replicates.



Extended data Fig. 7. Predicting therapy-induced trajectories

a, scVelo analysis based on scRNA-seq data from A375 and 383 cell lines cultured under control or trametinib/palbociclib (TP) conditions for 7 days, projected onto PC1-PC3 space.

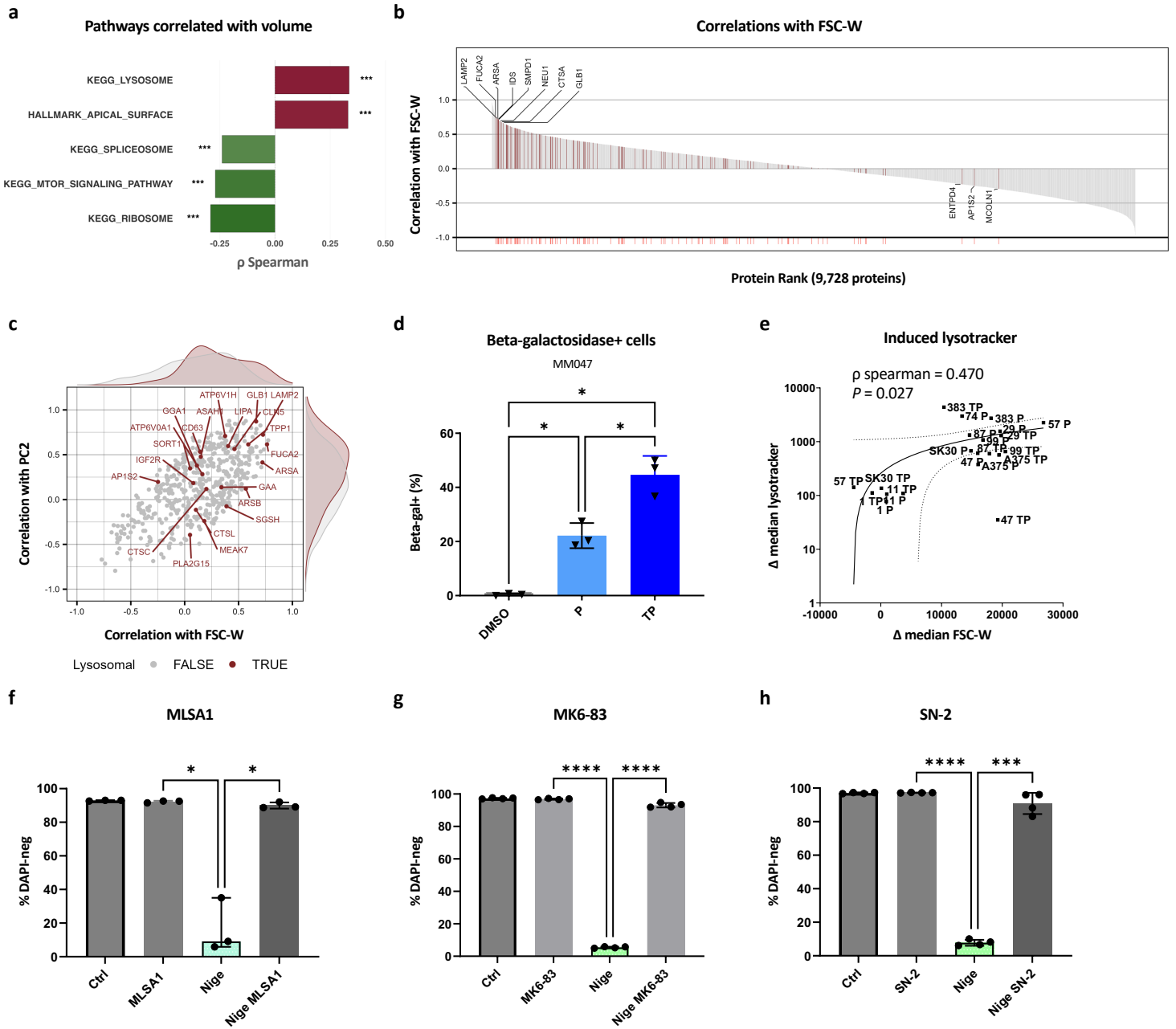
b, Average velocity field under dabrafenib/trametinib (DT; 20/100 nM) treatment for 3 days. Colored arrows represent reference cell lines, while grey arrows indicate interpolated trajectories inferred from reference S/V dynamics.

c, Seurat clustering analysis (clusters 0 and 1) projected onto UMAP1-UMAP2 space.

d, UMAP1-UMAP2 projection comparing control and TP-treated conditions.

e, Projection of the Rambow pigmentation signature within S/V space and across Seurat clusters.

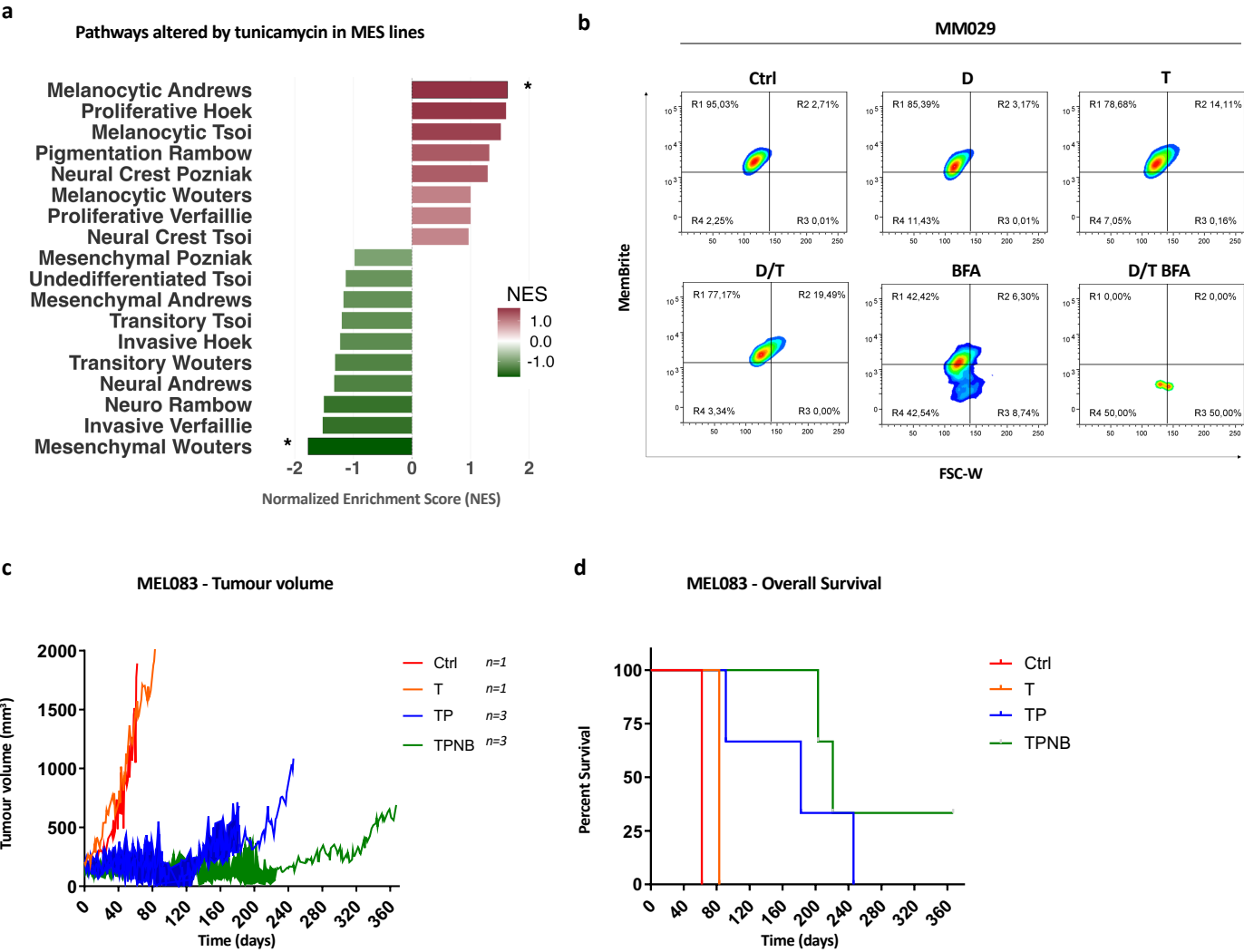
f, Heatmap showing transcriptional signatures differentially activated or repressed between control and TP-treated cells.



Extended Data Fig. 8. Targeting therapy-induced high-volume cells through modulation of lysosomal activity

- a**, ScRNA-seq data identifying top biological processes positively correlated with FSC-W.
- b**, Proteomic ranking of proteins correlated with FSC-W values; lysosomal proteins are highlighted in red.
- c**, Representation of lysosomal proteins showing correlation with both PC2 and FSC-W dimensions.
- d**, β -galactosidase activity in MM047 cells treated with palbociclib or trametinib/palbociclib for 7 days.
- e**, Correlation between changes in Lysotracker signal (Δ median Lysotracker) and corresponding changes in median FSC-W (Δ median FSC-W) under P or TP treatment.
- f**, Rescue of nigericin-induced cell death (% DAP-negative cells) after 12 days of treatment with MLSA1, a TRPML1 agonist.
- g**, Rescue of nigericin-induced cell death (% DAP-negative cells) after 12 days of treatment with MK6-83, a TRPML1 agonist.
- h**, Rescue of nigericin-induced cell death (% DAP-negative cells) after 12 days of treatment with SN-2, a dual TRPML1/TRPML3 agonist.

Statistical analyses: Panel d,f-h: Welch's ANOVA followed by Dunnett's multiple comparison test.



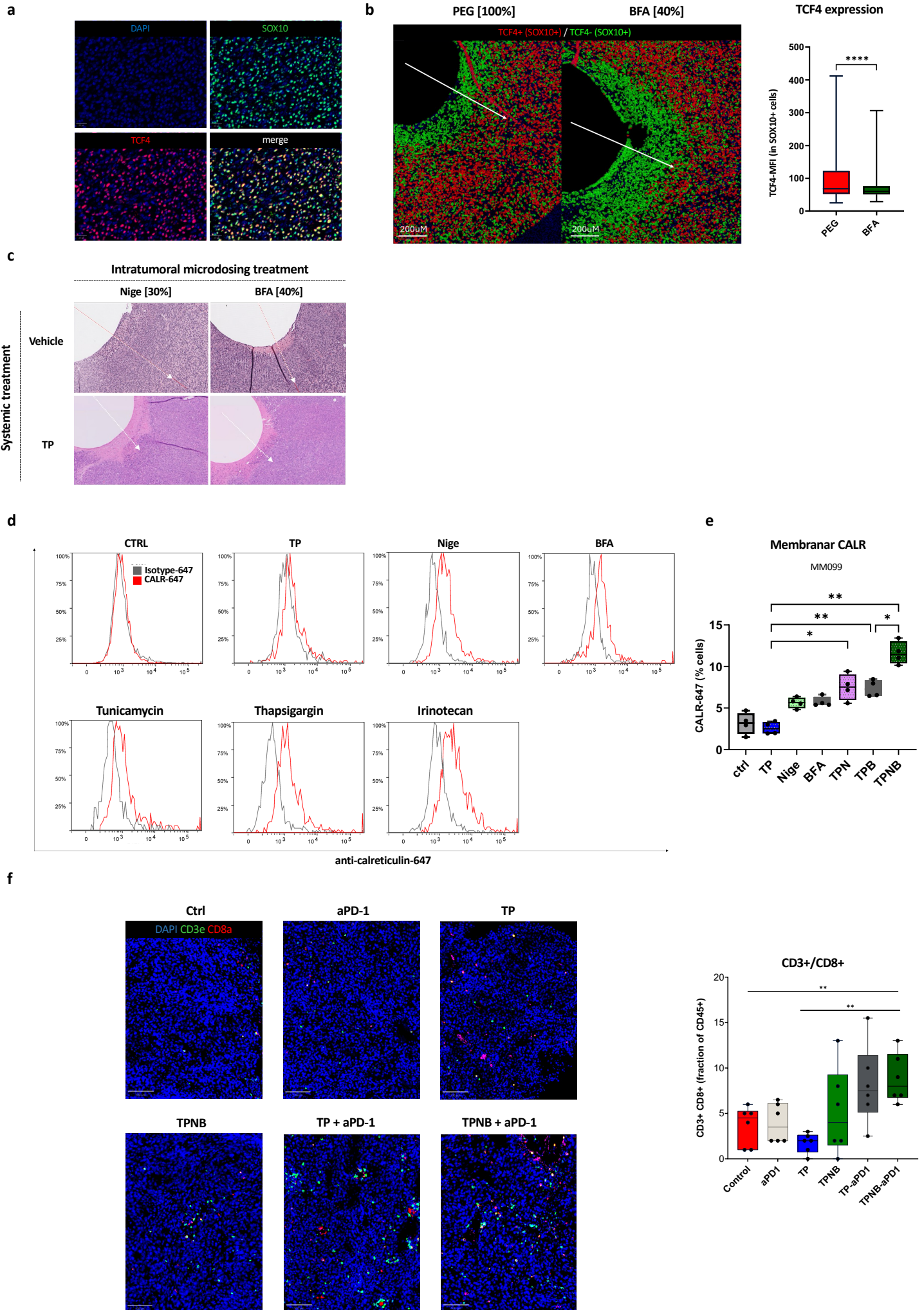
Extended data Fig. 9. Exacerbating therapy responses by decreasing cell surface

a, Bulk RNA-seq analysis showing up- and downregulated pathways across MES lines (MM029, MM047, MM099) following tunicamycin treatment (250 nM, 5 days).

b, Representative S/V flow cytometry plots of MM029 cells treated with dabrafenib (100 nM), trametinib (20 nM), BFA (25 nM), and in combinations at day 21.

c, Tumour growth kinetics of Mel083 PDX models treated with T, TP, or the TPNB combination (trametinib + palbociclib + nigericin + BFA). The experiment was terminated at day 367.

d, Overall survival of Mel083 PDX-bearing mice treated as in c (Log-rank test).



Extended Data Fig. 10. TPNB promotes Immunogenic cell death

a, Immunofluorescence staining for SOX10 and TCF4 in the YUMM3.3 R^{TT} model.

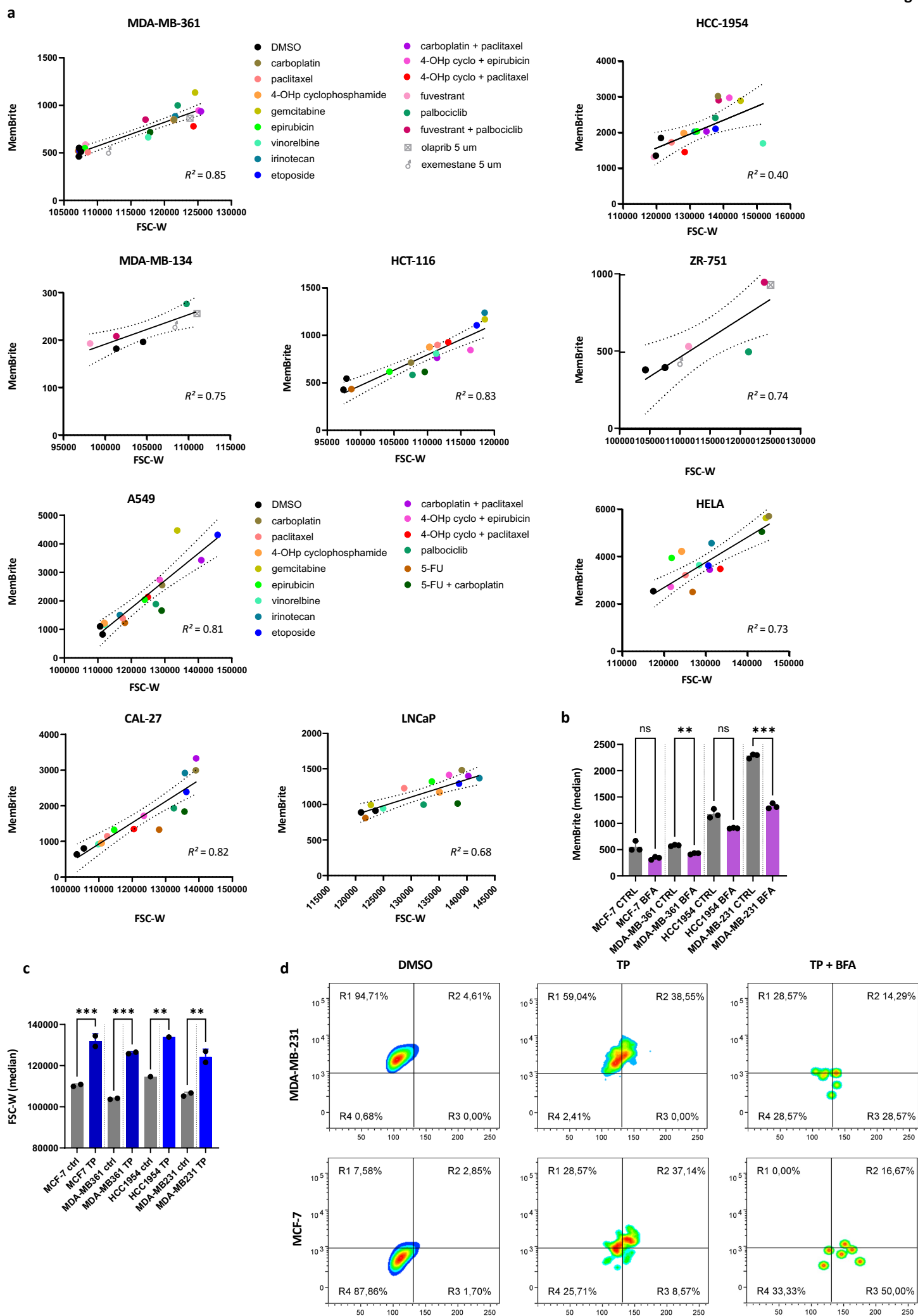
b, Quantification of TCF4 expression within SOX10⁺ cells (SOX10⁺/TCF4⁺ and SOX10⁺/TCF4⁻) in the YUMM3.3 model treated or not with BFA. (two-tailed Welch's t-test).

c, Representative H&E staining images of YUMM3.3 R^{TT} tumours treated systemically with vehicle or TP and the indicated compounds via the IMD, and quantification of necrotic area (n = 2-6 reservoirs per condition).

d, Representative flow cytometry histograms showing membranar calreticulin exposure after 3 days of treatment with TP, nige, BFA, additional UPR inducers (tunicamycin, thapsigargin) and the positive immunogenic cell-death control irinotecan.

e, Quantification of calreticulin exposure at day 6 under the different drug treatments.

f, Representative immunofluorescence images showing CD3⁺/CD8⁺ T-cell infiltration under the indicated treatment conditions, analysed at humane endpoint ($\approx 1000 \text{ mm}^3$), with corresponding quantifications.



Extended data Fig. 11. Chemotherapy rewires S/V in a panel of cancer cell lines

a, Modulation of S/V by standard-of-care chemotherapies across multiple human cancer cell lines, including MDA-MB-361, HCC-1954, MDA-MB-134, HCT116, ZR-751, A549, HeLa, CAL27, and LNCaP. Spearman's ρ and linear regressions are shown.

b, Changes in MemBrite intensity following chemotherapy treatment in MCF-7, MDA-MB-361, HCC-1954, and MDA-MB-231 cell lines.

c, Corresponding modulation of FSC-W under the same conditions as in b.

d, Representative S/V plots of MDA-MB-231 and MCF-7 cells treated with TP (10 nM/375 nM) and BFA (25 nM)

Statistical analyses: Panels b-c: One-way ANOVA followed by multiple-comparison tests (**** $p < 0.0001$; *** $p < 0.001$; ** $p < 0.01$; * $p < 0.05$).