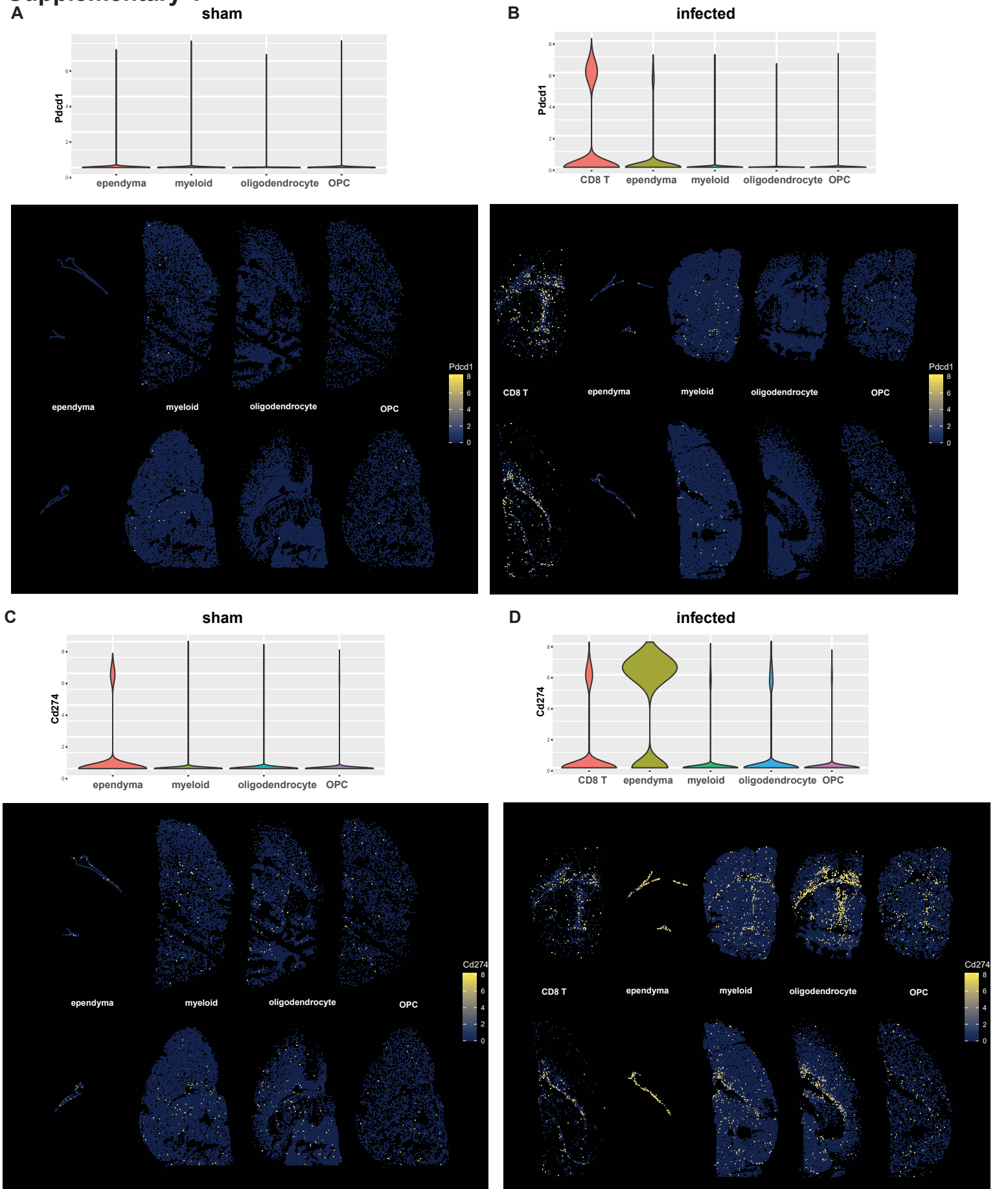
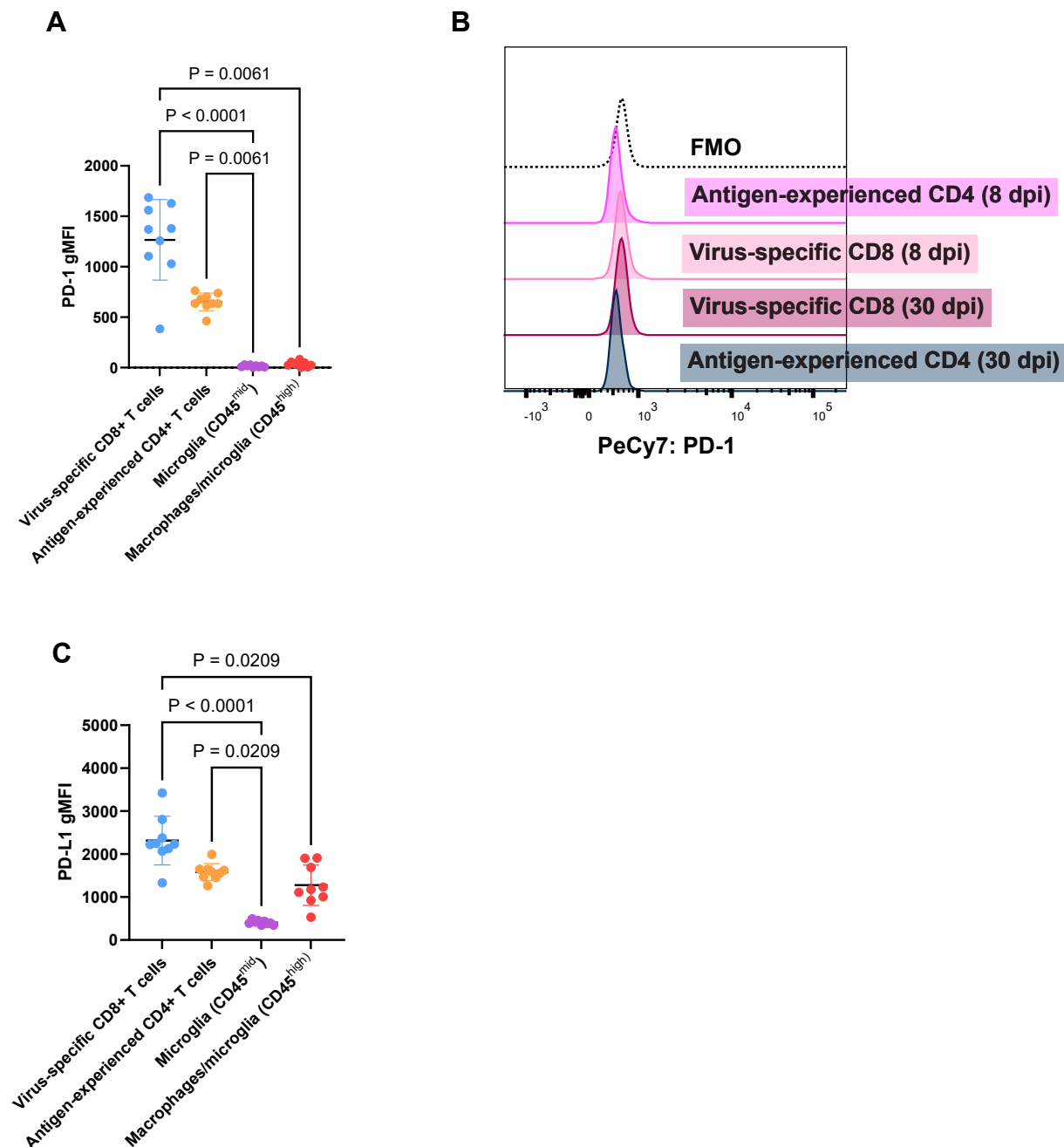


# Supplementary 1



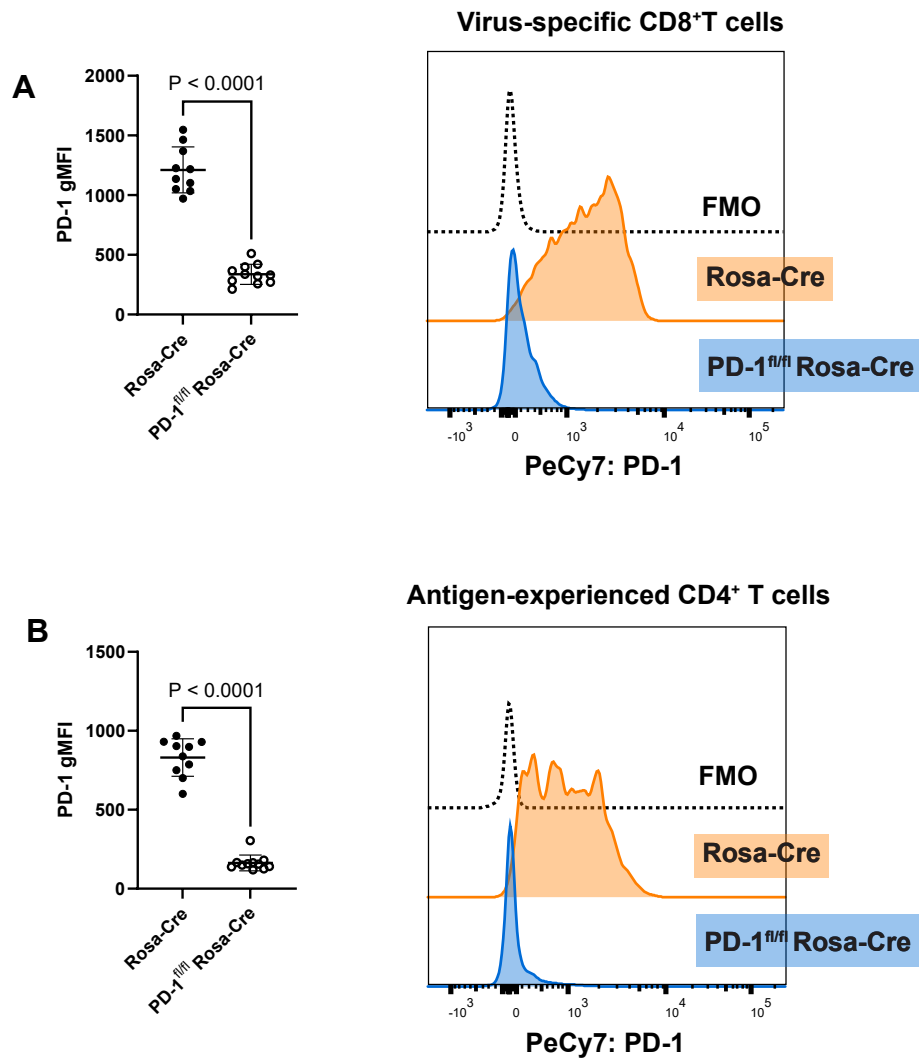
**Supplementary 1. MERFISH reveals spatial expression of *Pdcd1* and *Cd274* in the MuPyV-infected brain.** WT mice were sham-injected with vehicle or infected with MuPyV i.c. Brains were collected at 8 dpi. **A)** Violin plot of *Pdcd1* expression in multiple cell types, along with two representative brain sections, in sham-injected mice. Spots indicate 8 x 8  $\mu\text{m}$  spots, as determined by an image-based segmentation method<sup>22</sup>, and are positioned according to their corresponding x and y coordinates, regardless of z coordinates. Spots are colored according to log-normalized *Pdcd1* transcript expression. **B)** shows *Pdcd1* expression similarly to A) in MuPyV-infected mice. **C)** shows *Cd274* expression in sham-injected mice, while **D)** shows *Cd274* expression in MuPyV-infected mice. n = 2 mice per condition.

## Supplementary 2



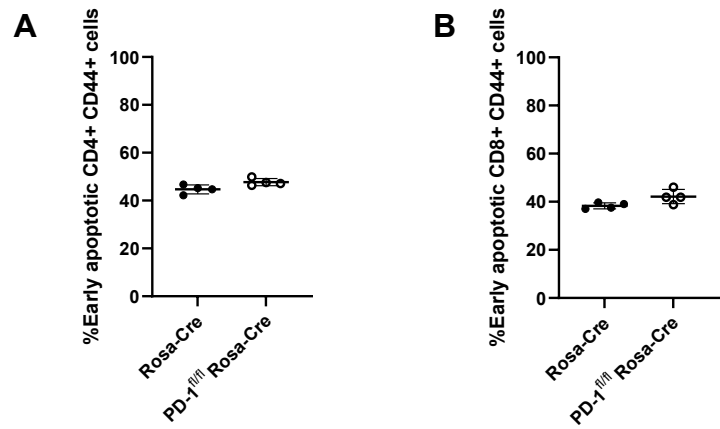
**Supplementary 2. PD-1 and PD-L1 expression are maintained on brain-infiltrating T cells during persistent infection.** WT mice were infected with MuPyV i.c., and at 8 and 30 dpi. Cells were isolated from the brain and blood and analyzed for PD-1 and PD-L1 expression. Gating strategies are specified in Figure 1. **A)** PD-1 expression on different cell types in the brain at 30 dpi. **B)** Representative histogram of PD-1 expression on T cells in the blood at 8 and 30 dpi. **C)** PD-L1 expression on different cell types in the brain at 30 dpi. n = 8 mice for 8 dpi and 9 mice for 30 dpi. Error bars indicate mean  $\pm$  SD. Data are representative of two independent experiments. As each mouse yielded all the immune cell types shown, statistical significance for A) and C) was calculated with the Friedman's test, a nonparametric alternative to the repeated measures one-way ANOVA, with a Dunn's post-test.

## Supplementary 3



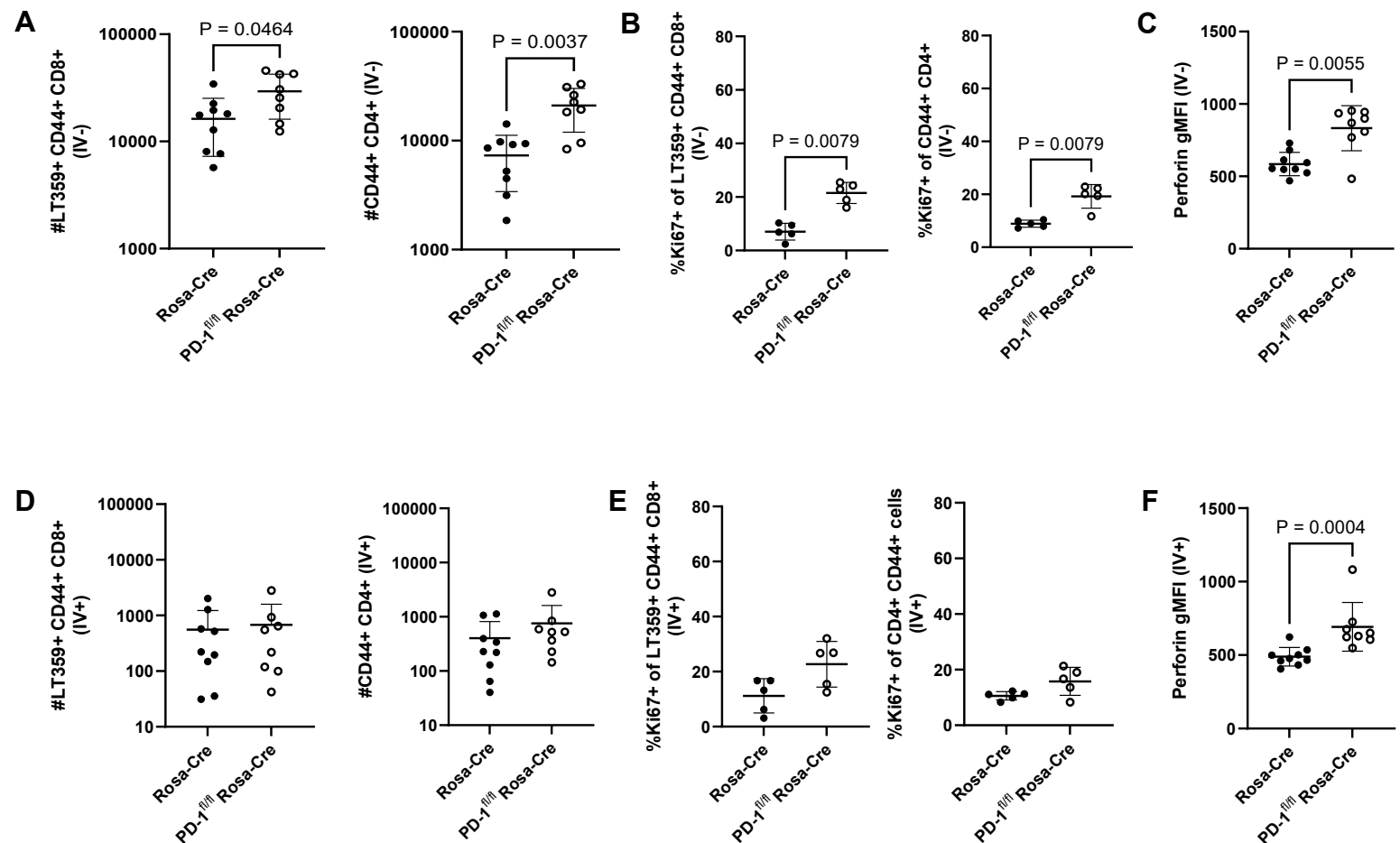
**Supplementary 3. Tamoxifen-induced knockdown of PD-1.** Rosa-Cre controls and PD-1<sup>fl/fl</sup> Rosa-Cre mice were infected with MuPyV i.c., given tamoxifen 8 to 12 dpi, and brains dissected at 15 dpi. **A)** PD-1 expression on brain-infiltrating virus-specific CD8<sup>+</sup> T cells. **B)** PD-1 expression on brain-infiltrating antigen-experienced CD4<sup>+</sup> T cells. n = 10 Rosa-Cre mice and 11 PD-1<sup>fl/fl</sup> Rosa-Cre mice. Error bars are mean ± SD. Data show two combined independent experiments. Statistical significance was determined with a Mann-Whitney U test.

# Supplementary 4



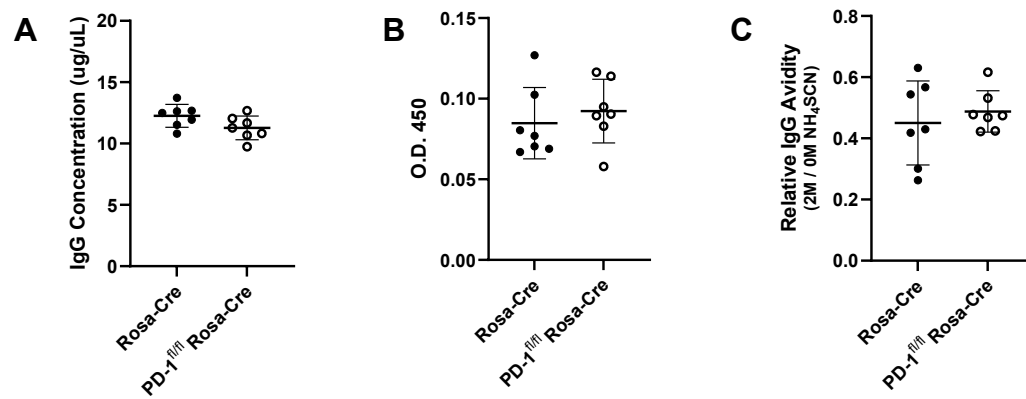
**Supplementary 4. Loss of PD-1 does not impact T cell apoptosis.** Rosa-Cre mice and PD-1<sup>fl/fl</sup> Rosa-Cre mice were treated as in Supplementary 3. Cells were assessed for early apoptosis via positive staining for annexin V and negative expression of viability dye, which stains dead cells. **A)** Frequency of antigen-experienced CD4<sup>+</sup> T cells undergoing early apoptosis. **B)** Frequency of virus-specific CD8<sup>+</sup> T cells going through early apoptosis. n = 4 Rosa-Cre mice and 4 PD-1<sup>fl/fl</sup> Rosa-Cre mice. Statistical significance was calculated with a Mann-Whitney U test. Error bars indicate mean ± SD.

Supplementary 5



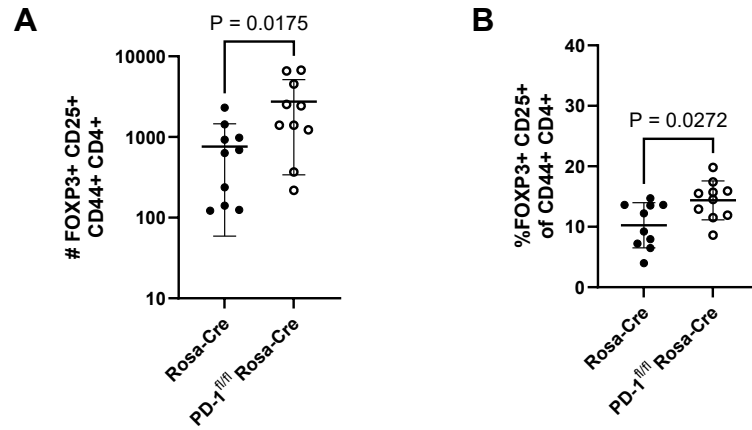
**Supplementary 5. Brain T cell phenotyping and quantification after CD45 antibody IV labeling.** Rosa-Cre mice and PD-1<sup>fl/fl</sup> Rosa-Cre mice were treated as in Supplementary 3 and immediately prior to euthanasia were injected IV with fluorophore-conjugated anti-CD45. **A)** Numbers of brain-infiltrating (IV-) virus-specific CD8<sup>+</sup> and antigen-experienced CD4<sup>+</sup> T cells. **B)** Proportions of IV- T cells positive for proliferation marker Ki67. **C)** Perforin expression in IV- virus-specific CD8<sup>+</sup> T cells. **D)** Numbers of T cells in the intravascular (IV+) compartment. **E)** Frequency of T cells expressing Ki67 in the IV+ compartment. **F)** Perforin expression in IV+ virus-specific CD8<sup>+</sup> T cells. n = 9 Rosa-Cre mice and 8 PD-1<sup>fl/fl</sup> Rosa-Cre mice. Error bars are mean ± SD. Data combined from two independent experiments for all except B) and E), which are representative of two independent experiments. Statistical significance for all graphs was calculated with a Mann-Whitney U test.

## Supplementary 6



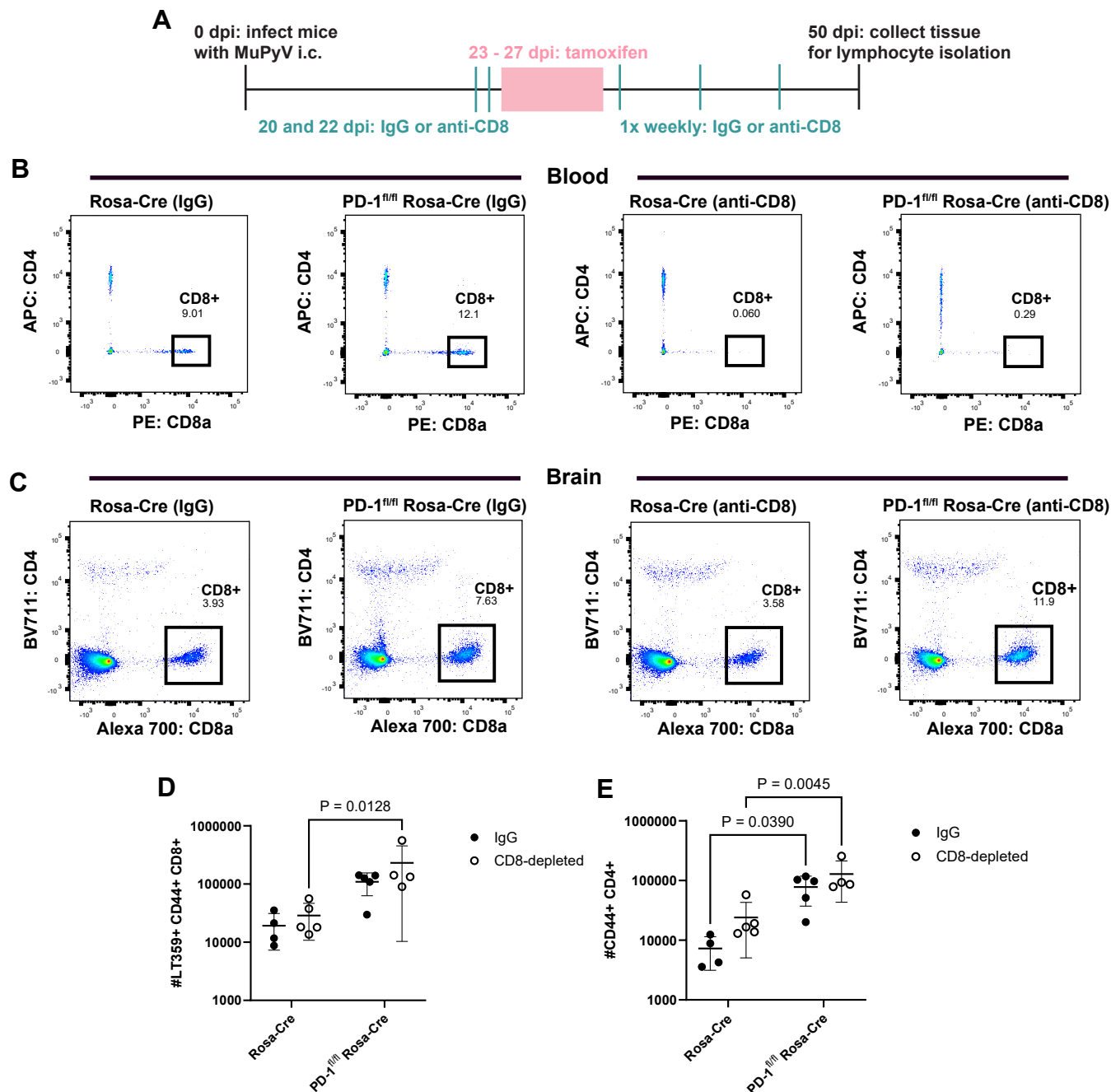
**Supplementary 6. PD-1 does not affect antiviral IgG or IgM.** Sera were collected from Rosa-Cre and PD-1<sup>fl/fl</sup> Rosa-Cre treated as in Supplementary 3, and IgG and IgM were quantified by ELISA. **A)** Concentration of anti-MuPyV IgG. **B)** IgM levels, shown by absorption. **C)** Relative IgG avidity to MuPyV. n = 7 Rosa-Cre mice and 7 PD-1<sup>fl/fl</sup> Rosa-Cre mice. Data combined from two independent experiments. Statistical significance quantified with a Mann-Whitney U test. Error bars show mean  $\pm$  SD.

## Supplementary 7



**Supplementary 7. PD-1 loss modestly increases T<sub>reg</sub> cells in brains of MuPyV-infected mice.** Mice were treated as in Supplementary 3. **A)** Numbers of T<sub>reg</sub> cells, designated as FOXP3<sup>+</sup> CD25<sup>+</sup> CD44<sup>+</sup> CD4<sup>+</sup>. **B)** Proportion of antigen-experienced CD4<sup>+</sup> T cells positive for FOXP3 and CD25. n = 10 Rosa-Cre mice and 10 PD-1<sup>fl/fl</sup> Rosa-Cre mice. Error bars show mean ± SD. Data combined from two independent experiments. Statistical significance calculated with a Mann-Whitney U test.

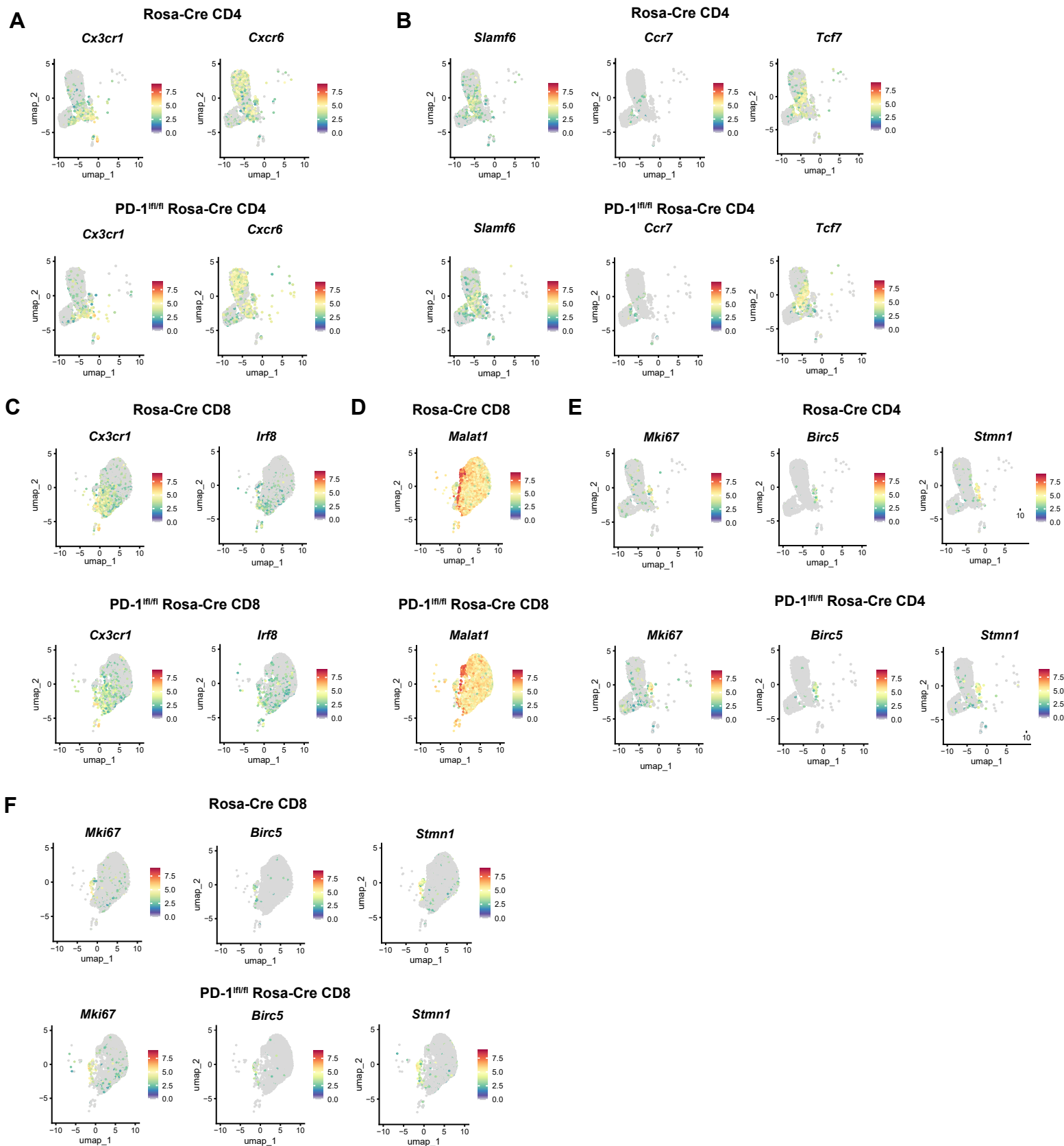
# Supplementary 8



**Supplementary 8. T cells expand in a brain-autonomous manner after loss of PD-1 during persistent infection.** **A)** Experimental setup for antibody-mediated peripheral CD8<sup>+</sup> T cell depletion. Rosa-Cre and PD-1<sup>fl/fl</sup> Rosa-Cre mice were given either IgG control or anti-CD8 antibody prior to tamoxifen administration, then given either IgG or anti-CD8 once a week until endpoint. **B)** Representative flow cytometry plots showing CD8<sup>+</sup> and CD4<sup>+</sup> T cells in blood of IgG-treated and anti-CD8-treated mice. **C)** Representative flow cytometry plots showing CD8<sup>+</sup> and CD4<sup>+</sup> T cells in brains of IgG-treated and anti-CD8-treated mice. **B)** and **C)** were gated from live CD45<sup>+</sup> cells. **D)** Virus-specific CD8<sup>+</sup> T cell and **E)** CD4<sup>+</sup> T cell counts. n = 4 Rosa-Cre mice (rat IgG), 5 Rosa-Cre mice (anti-CD8), 5 PD-1<sup>fl/fl</sup> Rosa-Cre (rat IgG), and 4 PD-1<sup>fl/fl</sup> Rosa-Cre (anti-CD8). Statistical significance for **D)** and **E)** was determined with a two-way ANOVA with uncorrected Fisher's LSD post-test. Error bars indicate mean ± SD.



# Supplementary 9



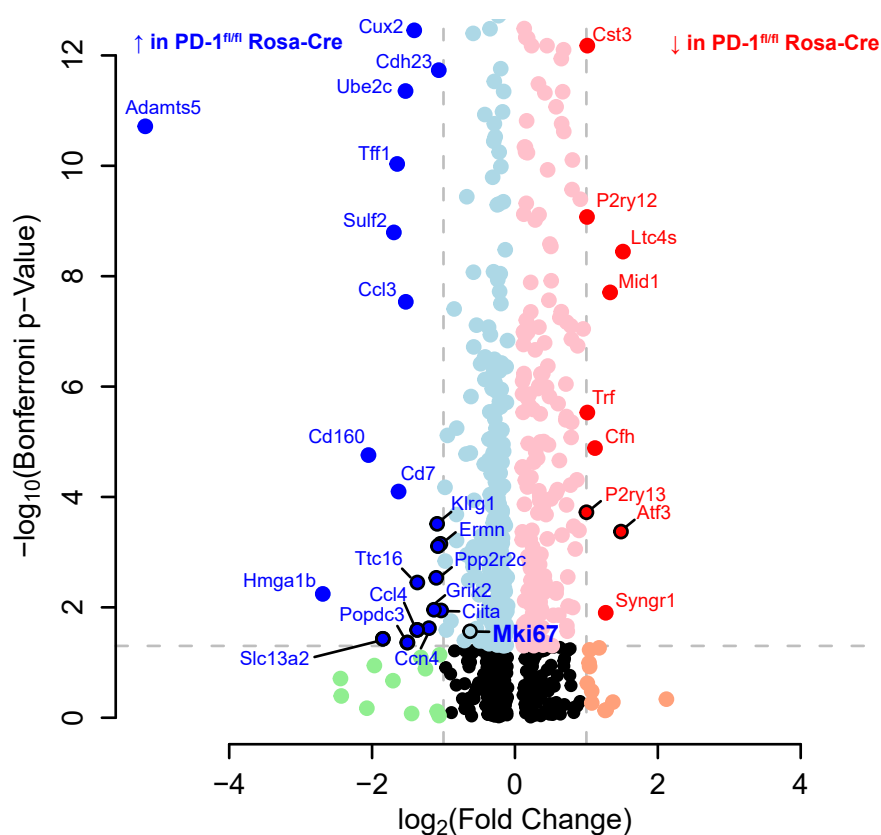
**Supplementary 9. Feature plots of genes in CD4<sup>+</sup> and CD8<sup>+</sup> T cell clusters.** Virus-specific CD8<sup>+</sup> T cells and antigen-experienced CD4<sup>+</sup> T cells were sorted from pooled groups of Rosa-Cre and PD-1<sup>fl/fl</sup> Rosa-Cre mice, and processed for scRNA-seq. **A)** Feature plots for *Cx3cr1* and *Cxcr6* in antigen-experienced CD4<sup>+</sup> T cells. **B)** Feature plots for genes in cluster 6 of CD4<sup>+</sup> T cells. **C)** Genes in cluster 0 of virus-specific CD8<sup>+</sup> T cells. **D)** Feature plots for *Malat1* in cluster 8 of CD8<sup>+</sup> T cells. **E)** Feature plots of genes in cluster 7 in CD4<sup>+</sup> T cells. **F)** Feature plots of genes in cluster 7 in CD8<sup>+</sup> T cells. n = 10 mice per strain.

### Rosa-Cre CD8 vs PD-1<sup>fl/fl</sup> Rosa-Cre CD8

Volcano plot showing differentially expressed genes in PD-1<sup>fl/fl</sup> Rosa-Cre mice. The y-axis represents  $-\log_{10}(\text{Bonferroni } p\text{-Value})$  (0 to 200), and the x-axis represents  $\log_2(\text{Fold Change})$  (-4 to 4). Genes are color-coded: blue for downregulated, red for upregulated, and green for no significant change. Labeled genes include:

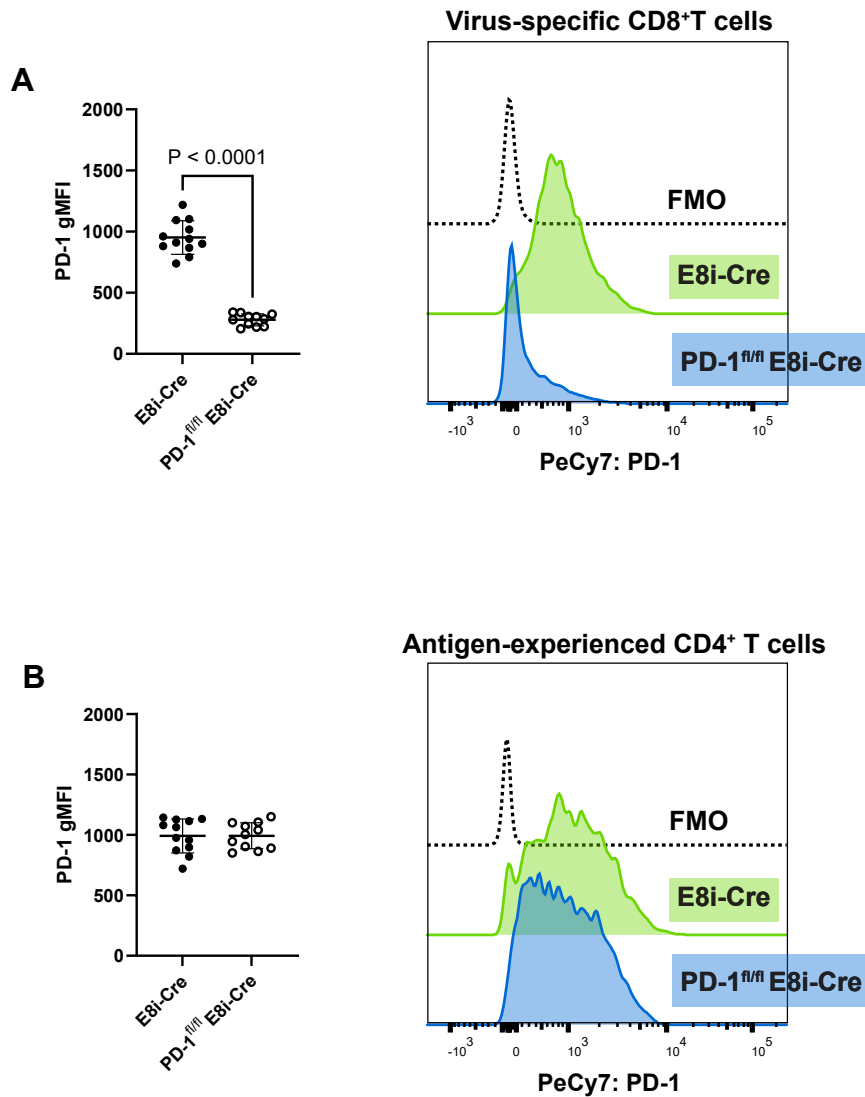
- Downregulated (Blue):** Ppp2r2c, Itgae, Itgb7, Trgc2, Trgv2, Rab4a, Klrk1, Htra3, Trgc4, Klrk1, Pclaf, Gzma, Prf1, Cd244a, Tmem273, Cd200r1, Ikzf4, MKi67.
- Upregulated (Red):** Nr4a1, Tcf7, Cx3cr1, Cd81, Tyrobp, Hexb, Trem2, Csfr1, Uaca, Selenop, Sparc, Cst3, Fcrls, Hmgn1, Trf, Slfn5, Serpine2, Cfh, Siglech, Oasl2, Xcl1, Fcscn1, Ltc4s, Mid1, Trim30c, Slc40a1, Ebf3, Lef1, Timp2.
- No significant change (Green):** Klrk1, Pclaf.

**B**



**Supplementary 10. Volcano plots showing differentially expressed genes in CD8<sup>+</sup> and CD4<sup>+</sup> T cells.** Genes in blue are upregulated with loss of PD-1, and genes in red are downregulated with loss of PD-1. Bolded genes indicate ones associated with effector, resident memory-like, or proliferative phenotypes. **A)** Differentially expressed genes in CD8<sup>+</sup> T cells. **B)** Differentially expressed genes in CD4<sup>+</sup> T cells. n = 10 per mice. Thresholds set at log2 fold change below -1 and above 1 and above -log<sub>10</sub>(0.05).

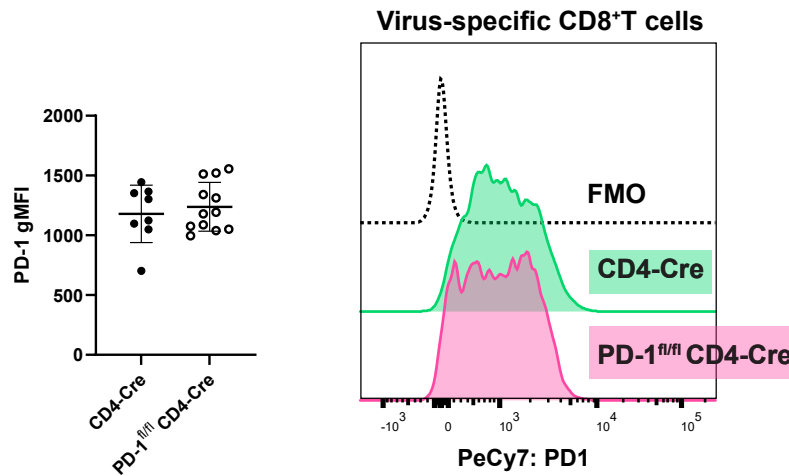
# Supplementary 11



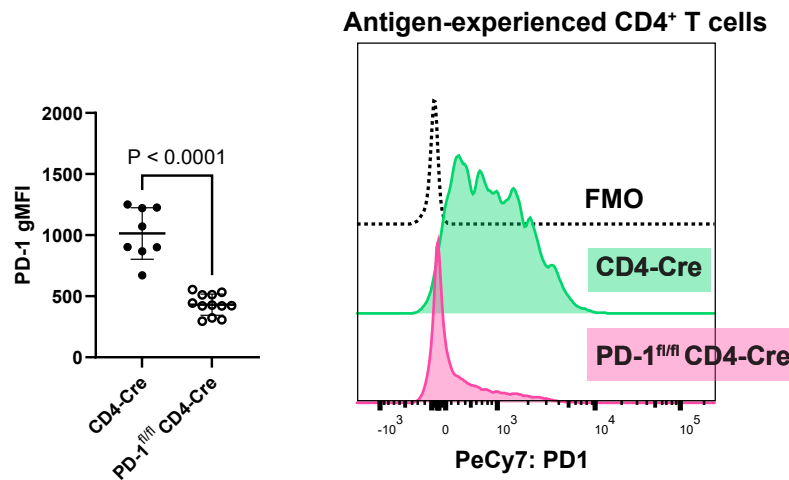
**Supplementary 11. CD8<sup>+</sup> T cell-specific PD-1 knockdown.** E8i-Cre controls and PD-1<sup>fl/fl</sup> E8i-Cre mice were i.c. inoculated with MuPyV, administered tamoxifen at 8 to 12 dpi, and T cells isolated from brains at 15 dpi. **A)** PD-1 expression on virus-specific CD8<sup>+</sup> T cells. **B)** PD-1 expression on antigen-experienced CD4<sup>+</sup> T cells. n = 12 E8i-Cre mice and 11 PD-1<sup>fl/fl</sup> E8i-Cre mice. Data combined from two independent experiments. Statistical significance calculated with a Mann-Whitney U test.

# Supplementary 12

**A**



**B**



**Supplementary 12. CD4<sup>+</sup> T cell-specific PD-1 knockdown.** CD4-Cre mice and PD-1<sup>fl/fl</sup> CD4-Cre mice were infected with MuPyV i.c. and given tamoxifen at 8 to 12 dpi. Brains were phenotyped through flow cytometry at 15 dpi. **A)** PD-1 expression on virus-specific CD8<sup>+</sup> T cells. **B)** PD-1 expression on antigen-experienced CD4<sup>+</sup> T cells. n = 8 CD4-Cre mice and 12 PD-1<sup>fl/fl</sup> CD4-Cre mice. Data combined from two independent experiments. Statistical significance quantified with a Mann-Whitney U test.