### 1 Non-canonical caspase-8 activation by cathepsin B drives anti-inflammatory

2 human macrophage polarization

3

4 Running title: Cathepsin B-Caspase-8 axis in macrophage polarization

5

- 6 Emeline Kerreneur<sup>1,2‡</sup>, Paul Chaintreuil<sup>1,2</sup>, Chloé Delaby<sup>1,2</sup>, Sonia Boulakirba<sup>1,2</sup>, Maxence
- 7 Bourgoin<sup>1,2,3</sup>, Morgane Fajolles<sup>1,2</sup>, Cécile Favreau<sup>1,2,3</sup>, Adèle Rivault<sup>1,2</sup>, Juba Bennour<sup>1</sup>,
- 8 Nathalie Droin<sup>4,5</sup>, Jean-François Peyron<sup>1</sup>, Johanna Chiche<sup>1,6</sup>, Thomas Cluzeau<sup>1,2,3</sup>, Michaël
- 9 Loschi<sup>1,2,3</sup>, Patrick Auberger<sup>1,2</sup>, Guillaume Robert<sup>1,2</sup> and Arnaud Jacquel<sup>1,2‡</sup>

10

- 11 1- Université Côte d'Azur, INSERM U1065, C3M, Nice, France
- 12 2- Equipe labellisée Association Recherche contre le Cancer, Nice, France
- 13 3- CHU de Nice, Département d'Hématologie Clinique, Nice, France
- 4- INSERM U1187, Gustave Roussy Cancer Center, Villejuif, France
- 5- Faculté de Médecine, Université Paris-Sud, Le Kremlin-Bicêtre, France
- 16 6- Equipe labellisée Ligue Contre le Cancer, Nice, France
- <sup>‡</sup> Corresponding authors

18

- 19 Correspondence: Emeline Kerreneur and Dr. Jacquel Arnaud
- 20 Centre Méditerranéen de Médecine Moléculaire (C3M)
- 21 Inserm U1065, Bâtiment ARCHIMED, 151, route de Saint-Antoine de Ginestière
- 22 BP2 3194, 06204 Nice Cedex 03, France
- 23 Phone: + 33 4 89 15 37 96
- Email: emeline.kerreneur@gmail.com or arnaud.jacquel@univ-cotedazur.fr

# **Supplementary Materials and Methods**

26

25

## 27 <u>L-lactate secretion assay</u>

- 28 100µL of 24H macrophages supernatants and control medium of culture is loaded in triplicates
- 29 into 96-well plate. L-lactate secretion is evaluated on a YSI 2950D Biochemistry Analyzer by
- 30 the difference between 24 vs 0H samples further rationalized to the number of secreting cells
- 31 evaluated by flow cytometry.

32

33

### **Phagocytosis**

- Control macrophages are pre-treated with cytochalasin D ( $1\mu g/mL$ ) for 30 minutes at  $37^{\circ}C$
- 35 and macrophages are then incubated with FITC-beads coated with E. Coli fragments
- 36 (15µg/mL) for 30 minutes at 37°C (Vybrant<sup>™</sup> Phagocytosis Assay kit, Invitrogen V6694).
- 37 Macrophage phagocytic activity is then determined by flow cytometry by subtracting the mean
- 38 fluorescence of macrophages and their control, diluted in trypan blue to eliminate background
- 39 signals.

#### **Supplementary Figure legends**

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

40

Supplementary Fig. 1 Caspase activation during monocyte-to-macrophage differentiation and anti-inflammatory macrophages polarization. a Amino acid sequences of caspase substrates and their putative cleavage sites during human monocyte-tomacrophage differentiation. Human monocytes (d0) are differentiated with CSF-1 for 5 days and then treated with CSF-1 (M0) or polarized with IFN-y + LPS for pro-inflammatory macrophages, IL-4 (M2), IL-4 ± IL-13, IL-6 or IL-10 for anti-inflammatory macrophages for 2 days. **b-d** Enzymatic measurement of non-apoptotic caspase activity in differentiating and polarized macrophages lysates using a specific fluorescent peptide (b: Ac-KWFD-AMC; c: Ac-IETD-AMC; d: Ac-DEVD-AMC). Results are expressed as A.U/min/mg of proteins and represent the mean ± SD of 3 independent experiments performed in triplicates. e-f Enzymatic measurement of recombinant caspase-8 (e) or caspase-3 (f) activity using specific fluorescent peptides (Ac-IETD-AMC, Ac-DEVD-AMC, Ac-NKFD-AMC and Ac-KWFD-AMC). Results are expressed as A.U/min/mg of proteins and represent the mean ± SD of 3 independent experiments performed in triplicates. g Caspase-8, caspase-7 and caspase-3 expression analysis by immunoblotting. HSP60 is used as a loading control. h Enzymatic measurement of non-apoptotic caspase activity using a specific fluorescent peptide (Ac-NKFD-AMC). Results are expressed as A.U/min/mg of proteins and represent the mean ± SD of 3 independent experiments performed in triplicates. ns : p > 0.05; \* p < 0.05; \*\* p < 0.01; \*\*\* p < 0.001 and \*\*\*\* p < 0.0001 according to an ordinary one-way ANOVA.

61

62

63

64

65

66

67

Supplementary Fig. 2 CTS enzymatic activity and expression in polarizing macrophages. a Human monocytes (d0) are differentiated with CSF-1 for 5 days and then treated with CSF-1 (M0) or polarized with IFN- $\gamma$  + LPS (M1) or IL-4 (M2) for 2 days. Enzymatic measurement of CTS using a specific fluorescent peptide Ac-FR-AMC. Results are expressed as A.U/min/mg of proteins and represent the mean  $\pm$  SD of 3 independent experiments performed in triplicates. \* p < 0.05 and \*\* p < 0.01 according to an ordinary one-way ANOVA.

**b** Anti-inflammatory polarizing macrophages are transfected with siRNA directed against Luciferase, CTSB, D, L, or S for 2 days. RT-qPCR analysis of CTSB, D, L and S gene expression. Results are expressed as relative expression and represent the mean  $\pm$  SD of 3 independent experiments. \*\* p < 0.01 and \*\*\*\* p < 0.0001 according to a two-tailed unpaired Student's t test.

Supplementary Fig. 3 CTS and non-apoptotic caspase activities in anti-inflammatory macrophages upon pharmacological CTSB or caspase inhibition. Enzymatic measurement of CTSB+L and non-apoptotic caspase activities using a specific fluorescent peptide (Ac-RR-AMC and Ac-NKFD-AMC respectively). Results are expressed as A.U/min/mg of proteins and represent the mean  $\pm$  SD of 3 independent experiments performed in triplicates. ns: p > 0.05; \*p < 0.05 and \*\*p < 0.01 according to a two-tailed unpaired Student's t test.

Supplementary Fig. 4 Phenotype and functional profile of anti-inflammatory macrophages upon CTSB or caspase pharmacological inhibition. Human monocytes are differentiated in macrophages with CSF-1 for 5 days and polarized into anti-inflammatory macrophages during 2 days with IL-4 and further treated with Emricasan or CA-074 for 2 days. **a-b** Enzymatic measurement of CTSB and non-apoptotic caspase activities using specific fluorescent peptides (Ac-RR-AMC and Ac-NKFD-AMC respectively). Results are expressed as A.U/min/mg of proteins and represent the mean  $\pm$  SD of 3 independent experiments performed in triplicates. **c-d** Lactate production analysis. Results are expressed as pg/mL/10<sup>6</sup> cells and represent the mean  $\pm$  SD of 3 independent experiments. **e-f** Phagocytic capacity analysis of macrophages by flow cytometry. Results are expressed as MFI (Mean Fluorescence Intensity) and represent the mean  $\pm$  SD of 3 independent experiments. ns: p > 0.05; \* p < 0.05 and \*\* p < 0.01 according to a two-tailed unpaired Student's t test.

Supplementary Fig. 5 Phenotype and functional profile of anti-inflammatory macrophages upon genetic inhibition of CTSB, caspase-8, caspase-7, caspase-3 or CTSL. Anti-inflammatory macrophages are transfected with siRNA directed against Luciferase (used as a control of transfection), caspase-8, caspase-7, caspase-3, CTSB or CTSL during 2 (a) or 3 days (b, c). a RT-qPCR analysis of CTSB, caspase-8, caspase-7, caspase-3 and CTSL gene expression. Results are expressed as relative expression and represent the mean ± SD of 4 independent experiments. \*\* p < 0.01; \*\*\* p < 0.001 and \*\*\*\* p < 0.0001 according to a two-tailed unpaired Student's t test. b Enzymatic measurement of CTSB and non-apoptotic caspase activities using specific fluorescent peptides (Ac-FR-AMC and Ac-NKFD-AMC respectively). Results are expressed as A.U/min/mg of proteins and represent the mean ± SD of 5 independent experiments performed in triplicates. c Phagocytic capacity analysis by flow cytometry. Results are expressed as MFI (Mean Fluorescence Intensity) and represent the mean ± SD of 3 independent experiments. ns : p > 0.05; \* p < 0.05; \*\* p < 0.01 and \*\*\*\*\* p < 0.0001 according to an ordinary one-way ANOVA.

**Supplementary Fig. 6 Transcriptomic reprogramming of anti-inflammatory macrophages under genetic inhibition of CTSB.** Anti-inflammatory macrophages are transfected with siRNA directed against Luciferase (M2 siLuc), used as a control of transfection, caspase-8 (M2 siC8) or CTSB (M2 siCTSB) for 2 days. **a** RT-qPCR analysis of caspase-8 or CTSB gene expression. Results are expressed as relative expression and represent the mean ± SD of 4 or 3 independent experiments respectively. ns : p > 0.05; \* p < 0.05; \*\* p < 0.01 and \*\*\* p < 0.001 according to according to a two-tailed unpaired Student's t test. **b** Unsupervised 2D projection of the six transcriptome samples principal component analysis. **c** Volcano plot representation of significant dysregulated genes between M2 siCTSB compared to M2 siLuc. **d** Dot plot representation of the top 20 biological processes from gene ontology analysis.