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2 **Supporting Information for:**

3 **Biomolecular condensation using *de novo* designed globular proteins**

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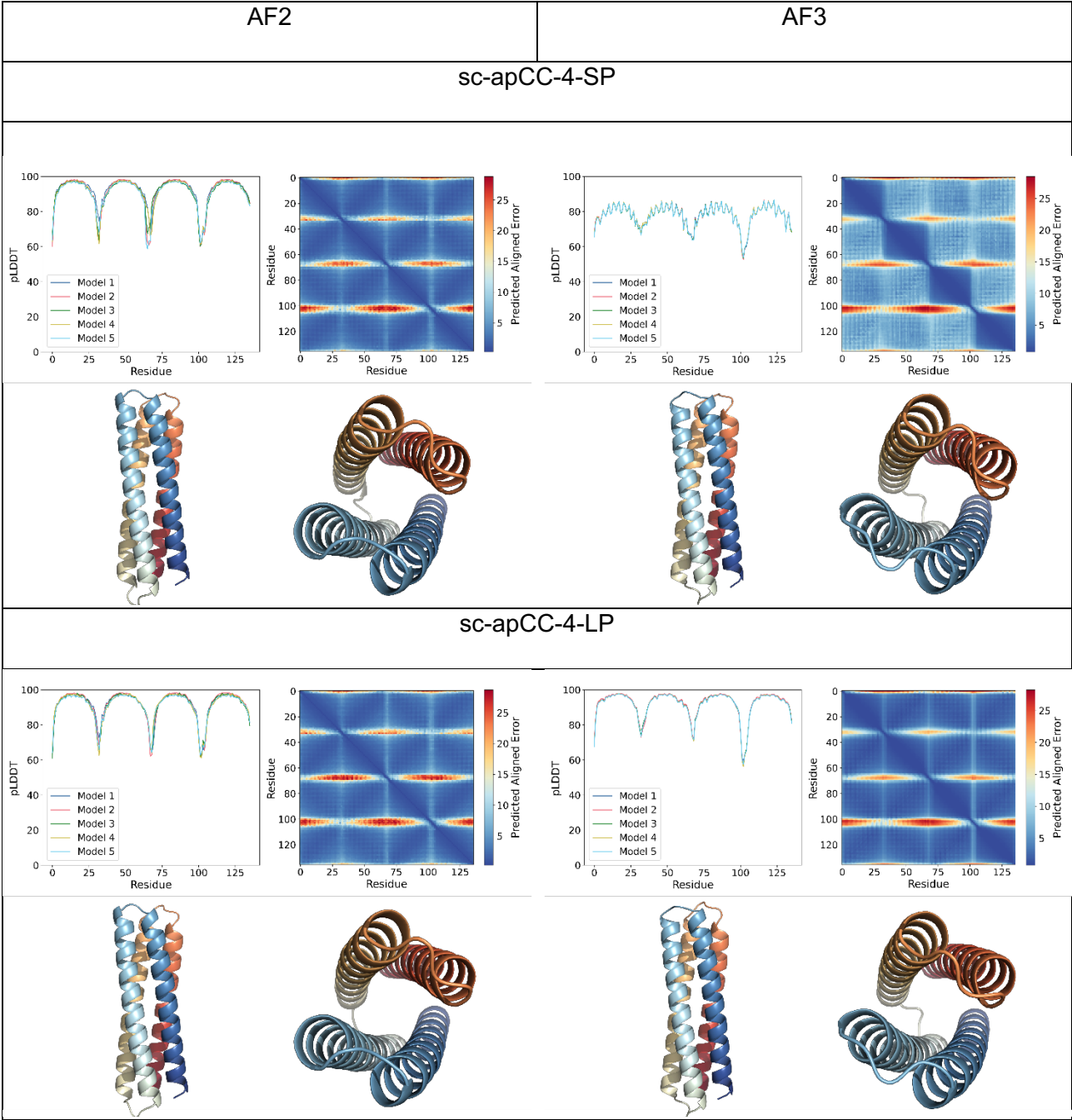
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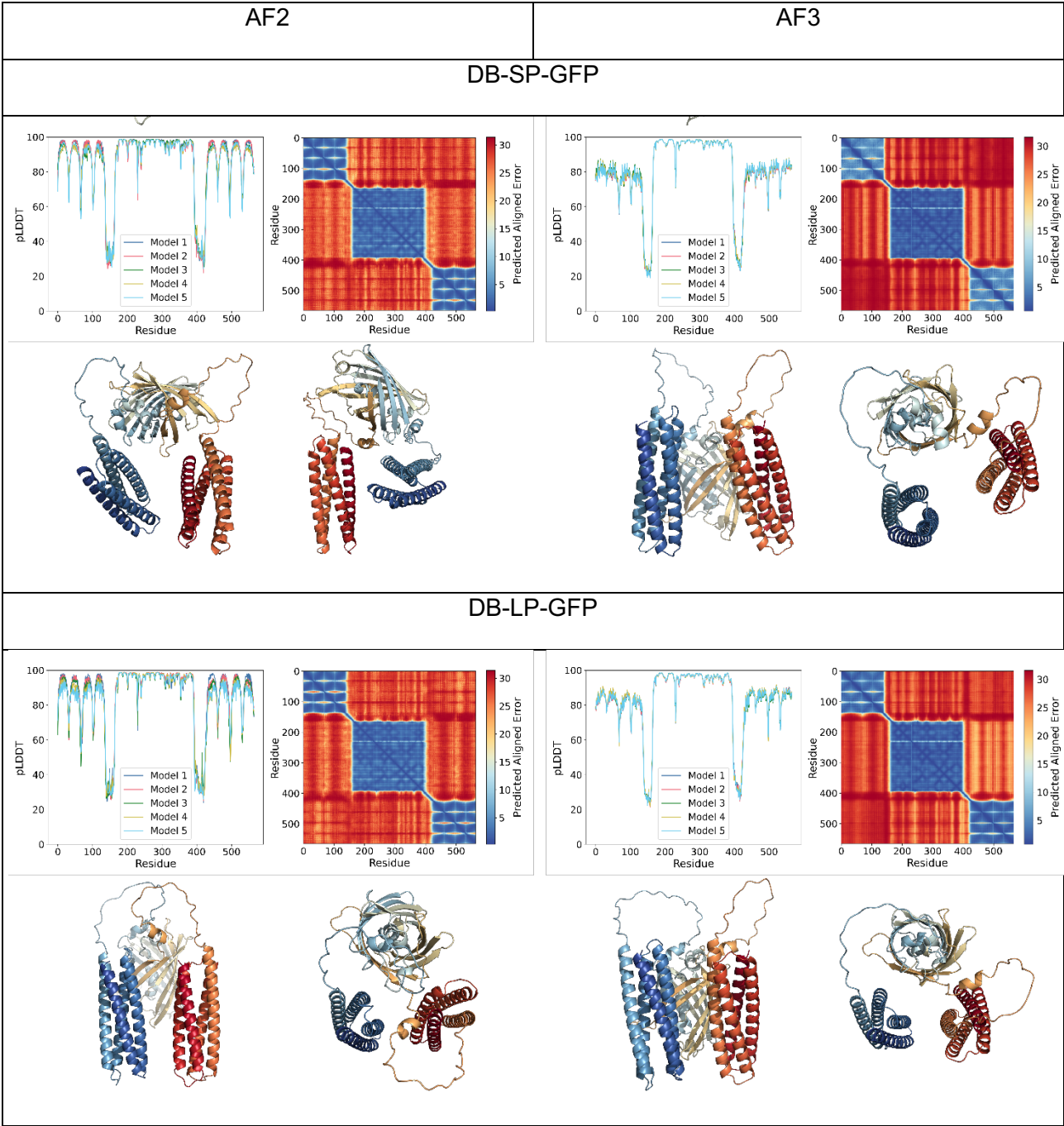
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25	KIF5C : mCherry	MADPAECSIKVMCRFRPLNEAEILRGDKFIPKFKGEETVVIGQGKPYVFDRVLPNNTTQ EQVYNACAKQIVKDVLEGYNGTIFAYGQTSSGKTHTMEGKLHDPQLMGIIIPRIAHDIFD HIYSMDENLEFHIKVSIFYEIIYLDKIRDLLDVSKTNLAVHEDKNRVPYVKGCTERFVSSP EEVMDVIDEGKANRHVAVTNMNEHSSRSHSIFLINIKQENVETEEKLSGKLYLVDLAGS EKVSKTGAEGAVLDEAKNINKSLSALGNVISALAEGTKTHVPYRDSKMTRILQDSLGGN CRTTIVICCSPSVFNEAETKSTLMFGQRAKTIKNTVSVNLELTAEEWKKKYEKEKEKNK ALKSVIQHLEVELNRWRNGEAVPEDEQISAKDQKNLEPCDNTPIIDNITPVTGGSGSGS GGTMVSKGEEDNMAIIKEFMRFKVHMEGSVNGHEFEIEGEGEGRPYEGTQTAKLKVTKG GPLPFAWDILSPQFMYGSKAYVKHPADIPDYLLKLSFPEGFKWERVMNFEDGGVVTVTQD SSLQDGEFIYKVKLRGTNFPDGPVMQKKTMGWEASSERMYPEDGALKGEIKQRLKLKD GGHYDAEVKTTYKAKKPVQLPGAYNVNIKLDITSHNEDYTIVEQYERAEGRHSTGGMDE LYK

21 **Supplementary Table 2.** AlphaFold2 and AlphaFold3 predictions of 3D structures for sc-
22 apCC-4 and DB-GFP variants





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26 **Supplementary Table 3.** Literature data for protein dynamics

probe/environment	FRAP recovery half-time (s)	Diff. Coefficient ($\mu m^2/s$)	viscosity (mpa·S)	Source
FUS-GFP/ FUS granules in cytoplasm of HeLa	~ 0.1-1.0		10–100	1
DDX4 ^{YFP} / DDX4 ^{YFP} BCs	~ 2.5	0.3 ± 0.1 (FRAP)		2
DDX4 ^{N1} + DDX4 ^{YFP} / <i>in vitro</i>	~ 60	0.4 ± 0.1 (FRAP)		2
LAF-1 / <i>in vitro</i>	161 ± 42 s	0.010 ± 0.003 (FRAP)		3
LAF-1 + RNA / <i>in vitro</i>	143 ± 28	0.011 ± 0.002 (FRAP)		3
hnRNPA1-Oregon-green + hnRNPA1 / <i>in vitro</i>	3.72 (fast) and 31.6 (slow)			4
hnRNPA1-Oregon-green / stress granules	4.2			4
NPM1 / <i>X. laevis</i> nucleoli	44 ± 6		740 ± 60	5
NPM1 / <i>in vitro</i>	16 ± 1			5
EGFP / cytoplasm of HeLa	NA	56 ± 11 (FCS)	1.93 ± 0.38*	6
EGFP / cytoplasm of HeLa	NA	50.6 (FCS)	2.14*	7

27 * Calculated using the data from literature

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Supplementary Table 4. Summary of fitting the *in vitro* FRAP data for DB-LP-GFP protein.

Conditions: 4 mg/mL protein, Tris 20 mM pH 7.5, NaCl 150 mM, 2% PEG3350, 37 °C.

Model	Parameter	Value	SD	Half-time [s]	R ²	Red. chi ²
Monoeexponential	a	-0.709	0.019	0.6928	0.823	0.002515
	b	1.00	0.05			
	c	0.798	0.003			
Biexponential	a1	-0.64	0.04	0.5311	0.8467	0.002186
	b1	0.58	0.06			
	a2	-0.17	0.04			
	b2	4.09	0.96			
	c	0.815	0.005			
Stretched exponential (Anomalous diffusion)	a	-0.88	0.04	0.4074	0.8387	0.002296
	b	0.73	0.07			
	alpha	0.64	0.04			
	c	0.806	0.003			

Supplementary Table 5. Summary of fitting the *in vivo* FRAP data for DB-LP-GFP protein expressed in HeLa. The measurements performed 24 hours after transfection at 37 °C.

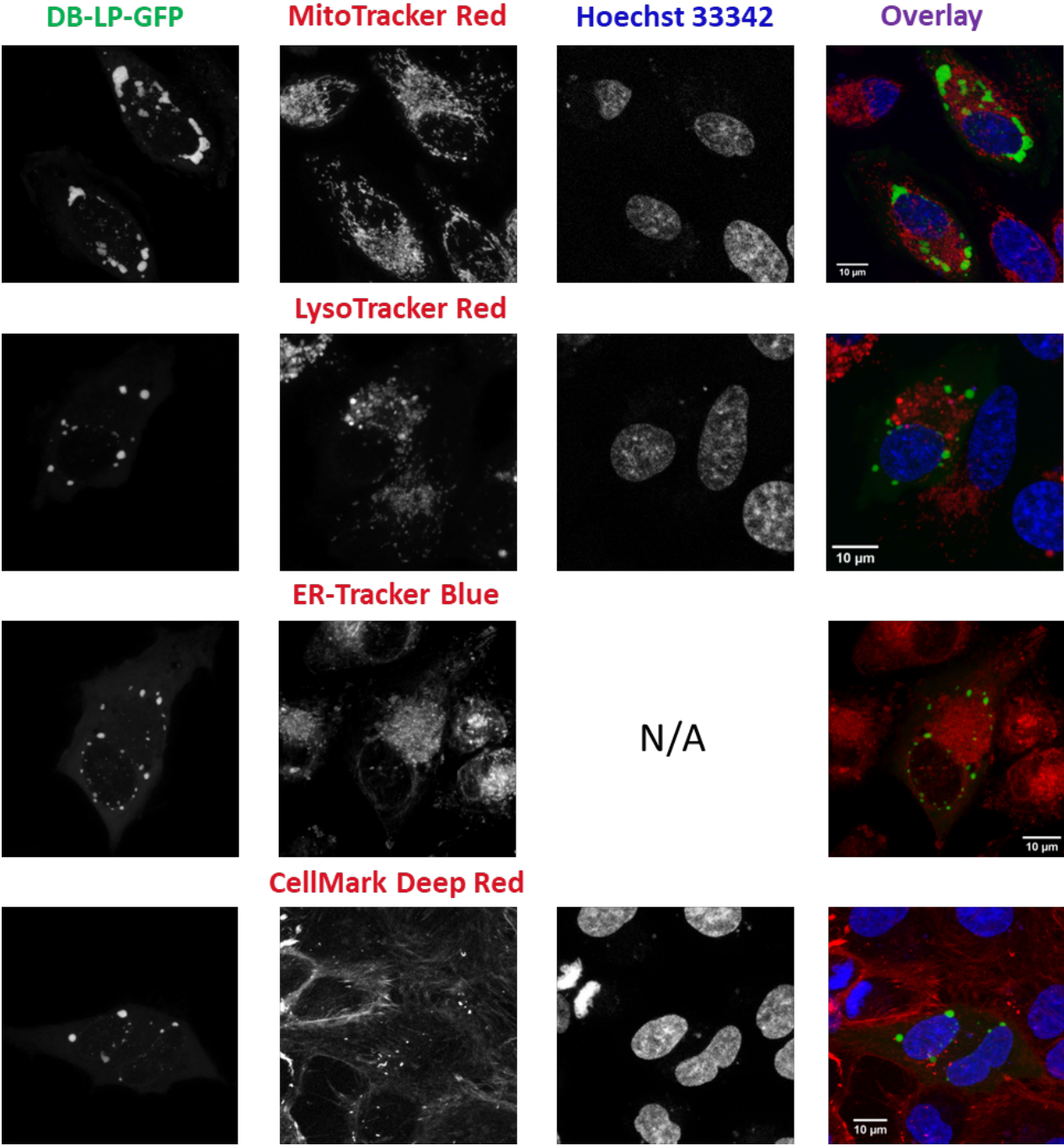
Model	Parameter	Value	SD	Half-time	R ²	Red. chi ²
Monoexp	a	-0.528	0.014	2.266	0.9077	0.002684
	b	3.3	0.3			
	c	0.690	0.009			
Biexp	a1	-0.45	0.03	1.164	0.9232	0.002262
	b1	4.7	0.6			
	a2	-0.27	0.06			
	b2	0.34	0.12			
	c	0.713	0.012			
Stretched Exp	A	0.89	0.16	1.176	0.9222	0.002276
	k	0.38	0.08			
	beta	0.46	0.10			
	C	-0.100	0.110			

38 **Supplementary Table 6.** The results of fitting autocorrelated functions (Fig. 2E) measured
39 for the diffusion of the probe protein using anomalous diffusion model.

Probe environment /	Probe hydrodynamic radius [nm]	Number of molecules (N)	τ_{D1} (slow)	r1(%)	r2(%)	An. factor	Diff. coefficient ($\mu m^2/s$) [fast]	Diff. coefficient ($\mu m^2/s$) [slow]	viscosity (mpaS) [high]
DB-LP-mCherry - / DB-LP-GFP BCs	4.40	6562.51*	22440 $\pm$ 1301	52	48	0.67	72.1 $\pm$ 2.6	0.36 $\pm$ 0.02	143 $\pm$ 9
mCherry / DB-LP-GFP BCs	2.1	48.19	236 $\pm$ 49			0.57	34.3 $\pm$ 7		3.2 $\pm$ 0.7
DB-LP-mCherry / cytoplasm	4.40	12.75	342 $\pm$ 12			0.74	23.7 $\pm$ 0.9		2.18 $\pm$ 0.09
mCherry / cytoplasm	2.1	41.69	122 $\pm$ 10			0.62	66.5 $\pm$ 6		1.63 $\pm$ 0.14

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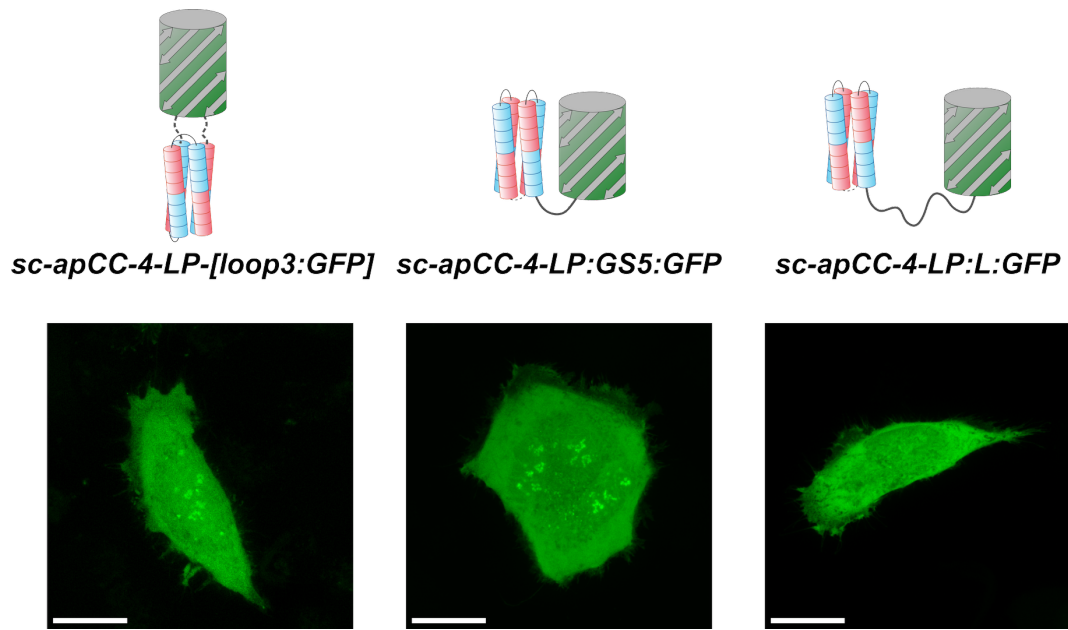
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44 **Supplementary Figure 1.** Live confocal images of HeLa cells in 24 hours of transfection

45 with the DB-LP protein and stained with organelle-specific fluorescent markers. Columns 1-

46 3 correspond to the green, red, and blue channels respectively. The last column is showing

47 the overlay of all channels in respective colours.



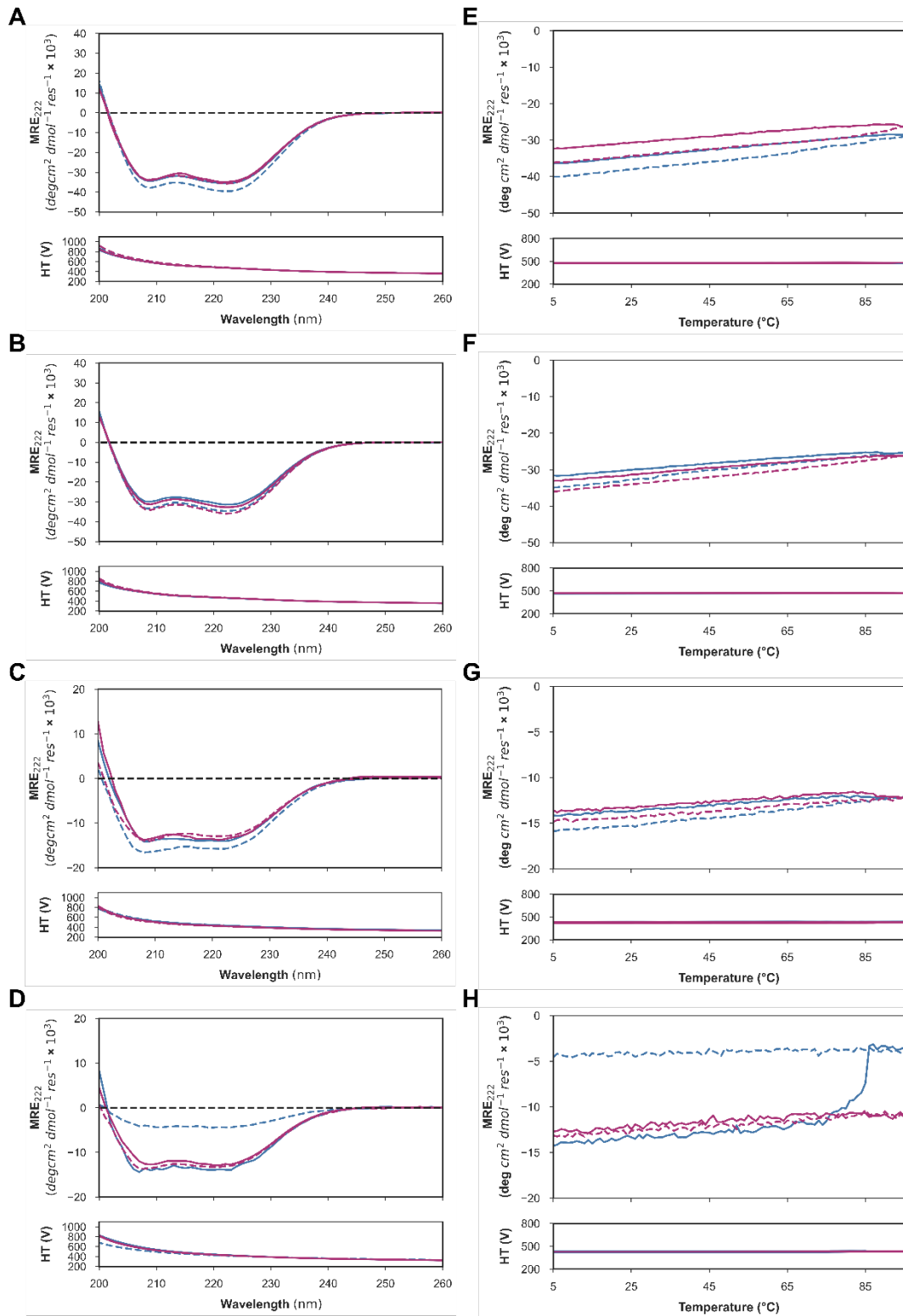
49

50 **Supplementary Figure 2. Both sticker domains are required for BC formation in live**
 51 **cells.** Confocal images of HeLa cells that transiently overexpressed sc-apCC-4-LP fusions
 52 to GFP: the loop fusion (1st column) in which GFP was inserted into the 3rd loop of the sc-
 53 apCC-4-; the linear fusion (2nd column) in which GFP was fused to the C-terminus of sc-
 54 apCC-4-LP with a short 5-residue long flexible GS linker, and another linear fusion with the
 55 same 25-residue long linker as in the dumbbell constructs (3rd column) matching the one in
 56 DB constructs. Scalebars = 20 μ m.

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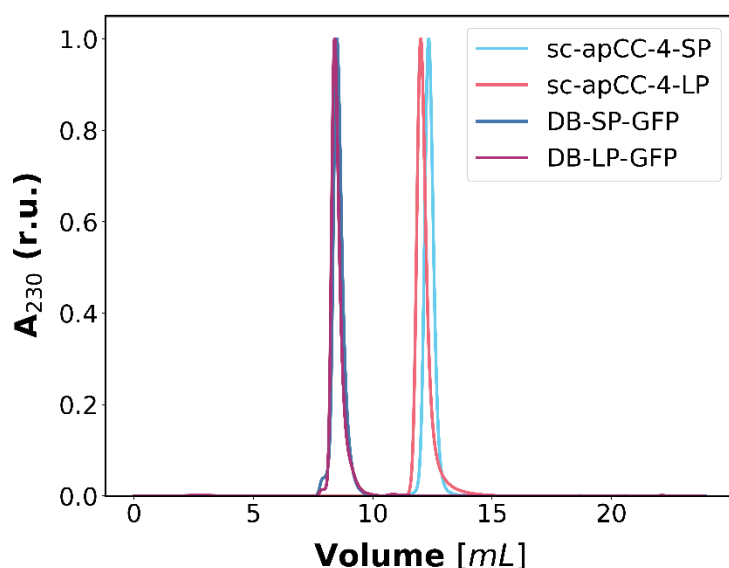
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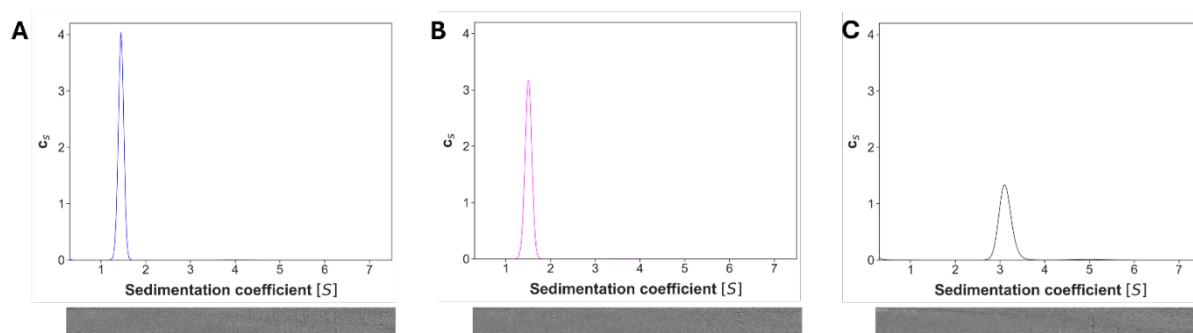
60

61 **Supplementary Figure 3.** CD spectra at 5 °C (A, B, C, D) and thermal response curves at
 62 222 nm (E, F, G, H) of sc-apCC-4-SP (A, E), sc-apCC-4-LP (B, F), DB-SP-GFP (C, G) , and
 63 DB-LP-GFP (D, H) at 150 mM (blue line) and 500 mM (dark red line) NaCl before (solid line)
 64 and after (dashed line) annealing. Conditions: 50 mM Tris pH 7.5.



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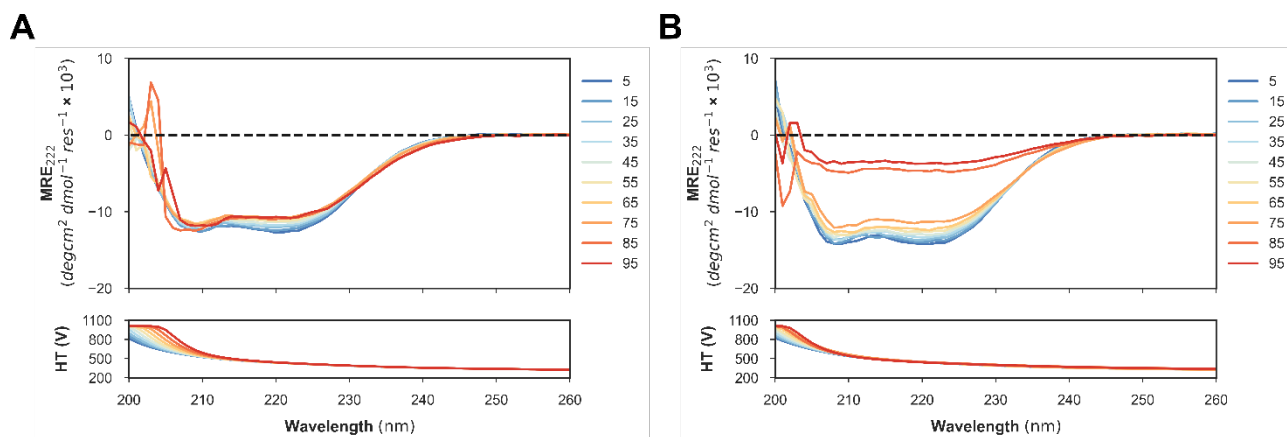
66 **Supplementary Figure 4.** Analytical size-exclusion chromatography traces for the designer
 67 proteins sc-apCC-4-SP, sc-apCC-4-LP, DB-SP-GFP, DB-LP-GFP. Conditions: 50 mM Tris
 68 pH 7.5, 500 mM NaCl, 0.8 mL/min, Sephadex 75pg Increase.



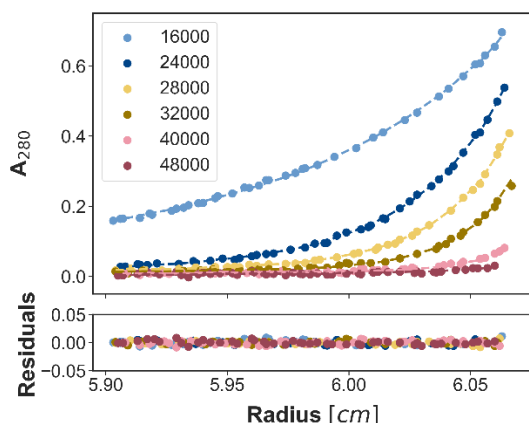
69

70 **Supplementary Figure 5.** Sedimentation velocity (SV) AUC traces of the *de novo* proteins
 71 designed for this study: A) sc-apCC-4-SP, B) sc-apCC-4-LP, C) DB-LP-GFP. Fits returned
 72 weights of 0.98, 1.04, and 1.02× monomer weight, respectively. 80 μ M sc-apCC-4-SP and
 73 sc-apCC-4-LP, 15 μ M DB-LP-GFP. Conditions: 50 mM Tris pH 7.5, 500 mM NaCl, 20 $^{\circ}$ C.

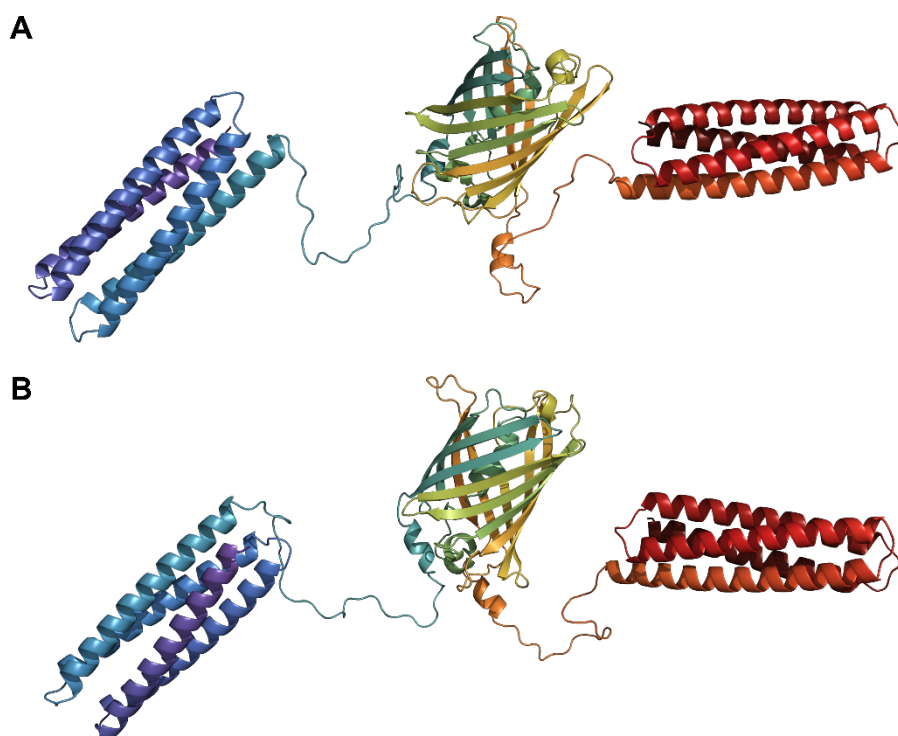
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Supplementary Figure 6. CD spectra of DB-LP-GFP protein at 500 mM (A) and 150 mM (B) NaCl at various temperatures upon heating from 5 to 95 °C. Conditions: 50 mM Tris pH 7.5.

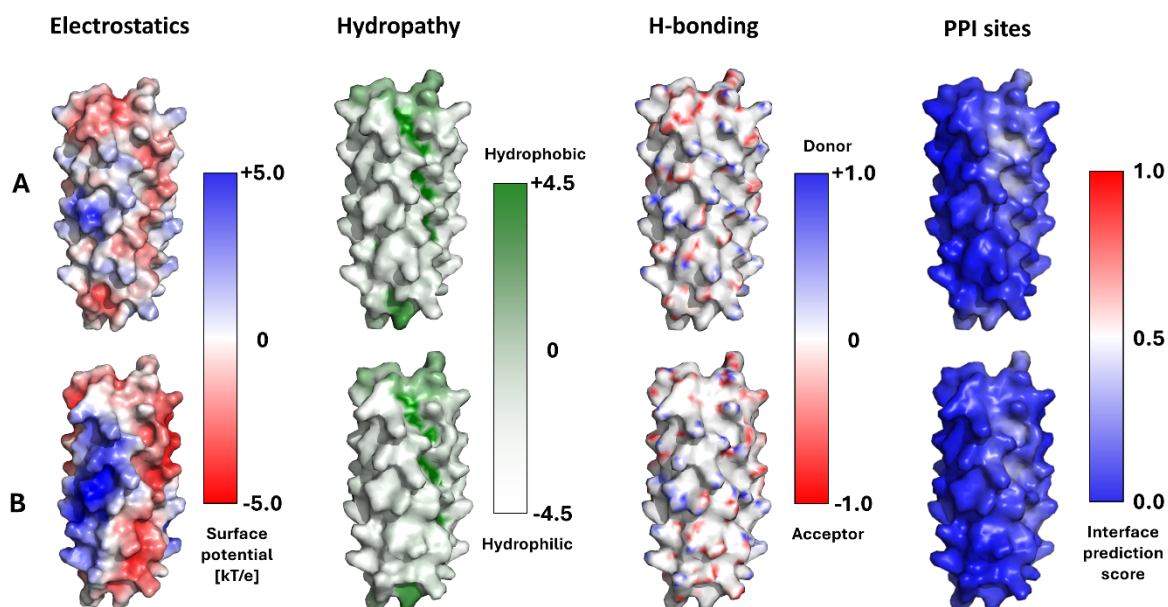


Supplementary Figure 7. Sedimentation equilibrium (SE) AUC data for DB-LP-GFP between 16 and 48 krpm. Fitted single-ideal species model curves are overlaid (dashed lines) and gave a molecular weight of 65.6 kDa which corresponds to 1.03× DB-LP-GFP monomer weight. 15 μM DB-LP-GFP. Conditions: 50 mM Tris pH 7.5, 500 mM NaCl, 20 °C.



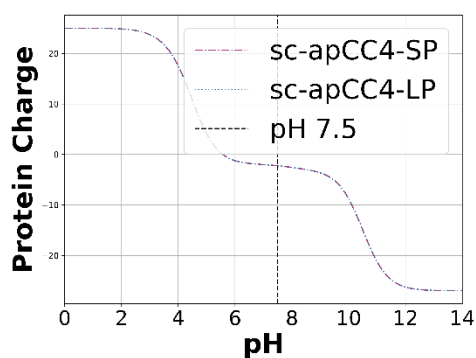
86

87 **Supplementary Figure 8.** The best-fit models for the SAXS data for A) DB-SP-GFP and B)
88 DB-LP-GFP obtained using MultiFoXS (see Methods).



89

90 **Supplementary Figure 9.** APBS⁸ and MaSIF⁹ predictions for sc-apCC-4-SP and sc-apCC-
 91 4-LP.

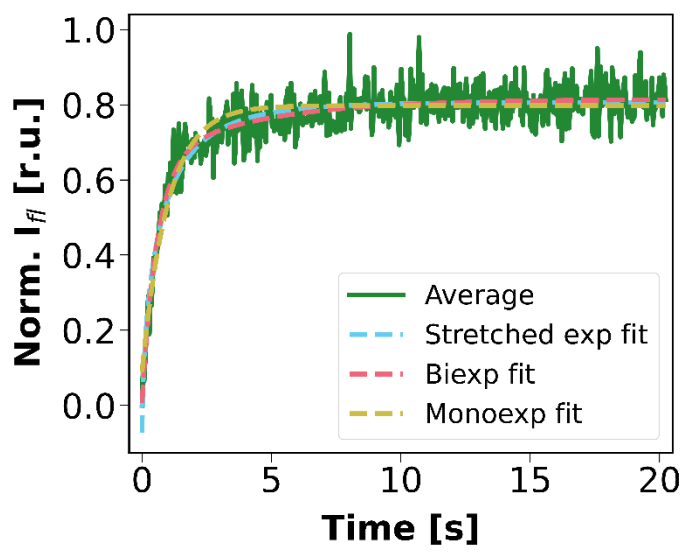


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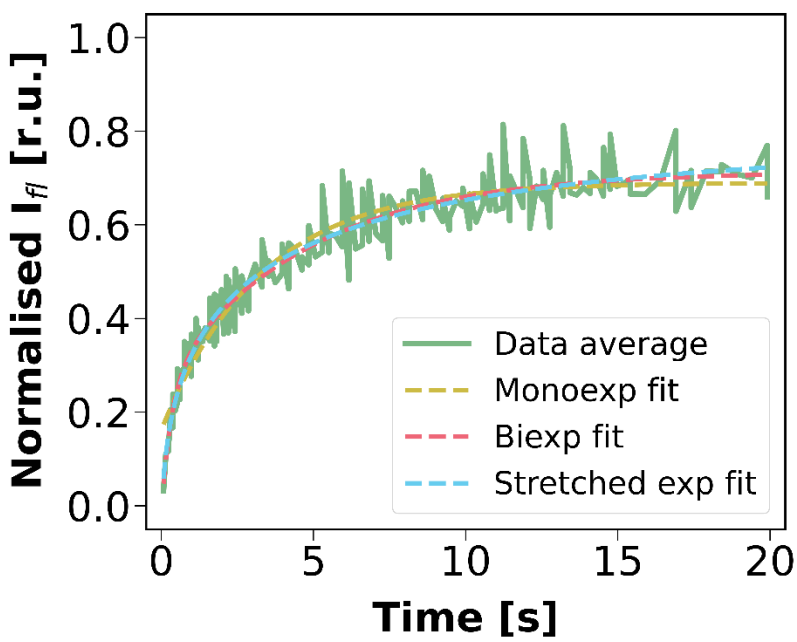
93 **Supplementary Figure 10.** PROPKA 3 prediction of the protein net charge at various pH
 94 values for sc-apCC-4-SP and sc-apCC-4-LP^{10,11}.

95

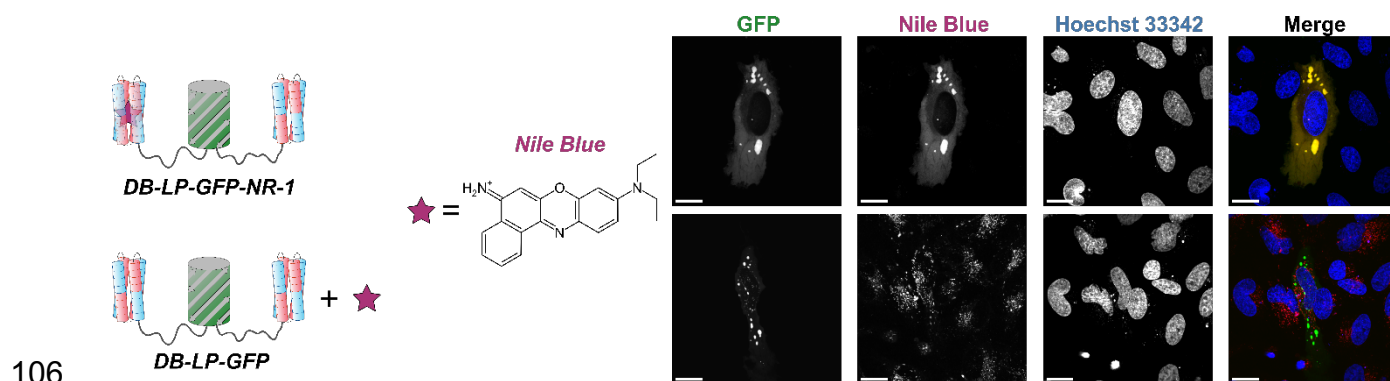
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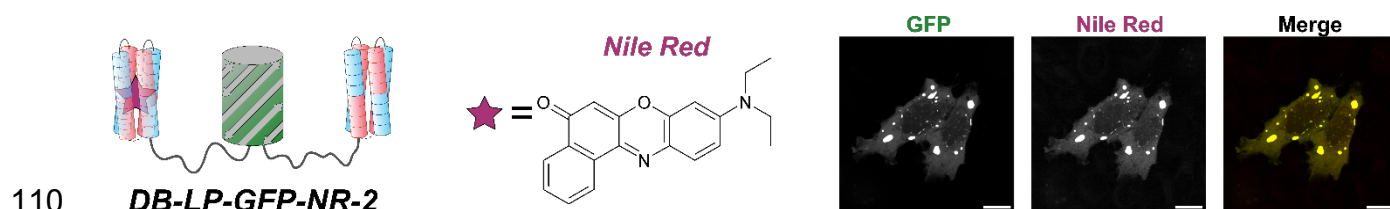
99 **Supplementary Figure 11.** Fitting the *in vitro* FRAP data for DB-LP-GFP protein with mono-
100 , bi- and stretched exponential functions. Conditions: 4 mg/mL protein, Tris 20 mM pH 7.5,
101 NaCl 150 mM, 2% PEG3350, 37 °C.



103 **Supplementary Figure 12.** Fitting the *in vivo* FRAP data for DB-LP-GFP protein expressed
104 in HeLa. The measurements performed 24 hours after transfection at 37 °C.



107 **Supplementary Figure 13.** Confocal fluorescence images of live HeLa cells expressing DB-
 108 LP-GFP-NR-1 (top row) and DB-LP-GFP (bottom row) and stained with Hoechst 33342 and
 109 Nile Blue (NB).



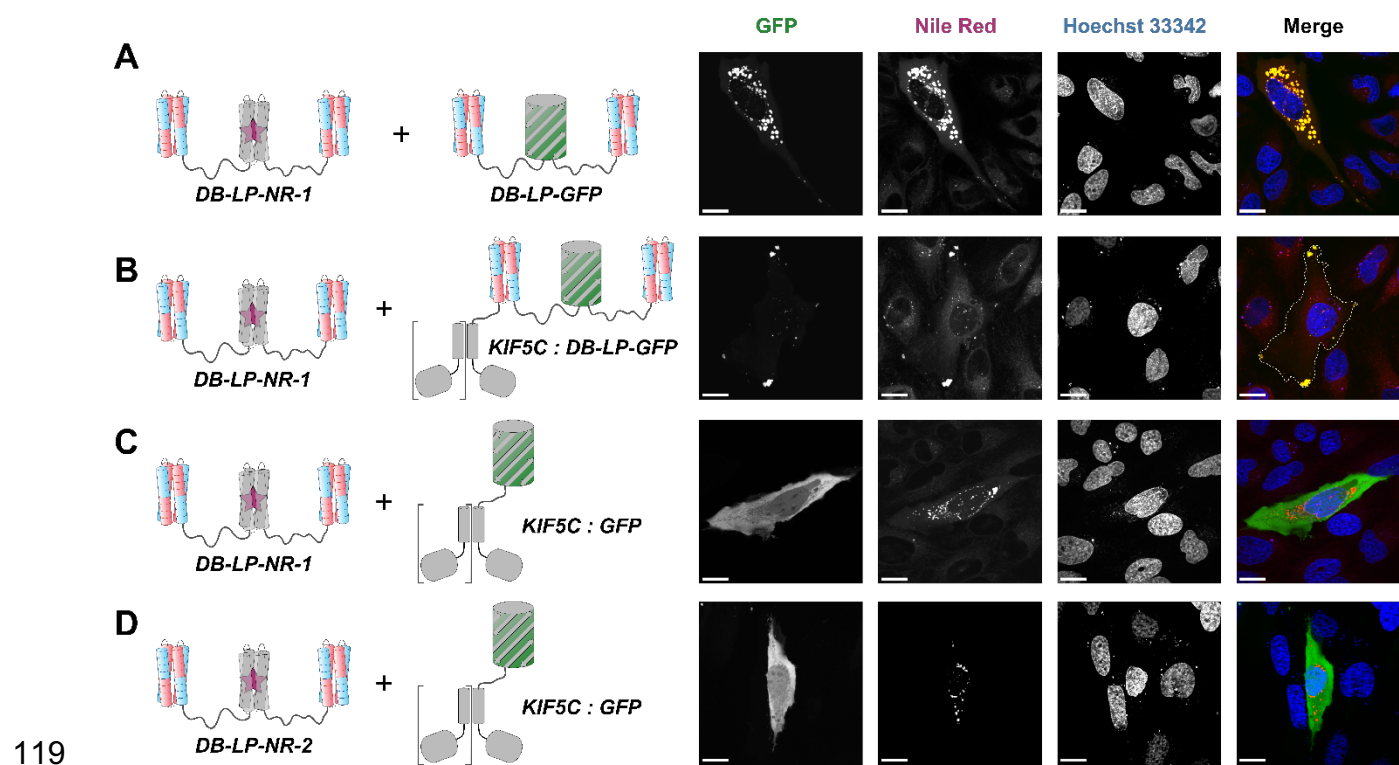
111 **Supplementary Figure 14.** Confocal fluorescence images of live HeLa cells expressing DB-
 112 LP-GFP-NR-2 stained with Nile Red (NR).



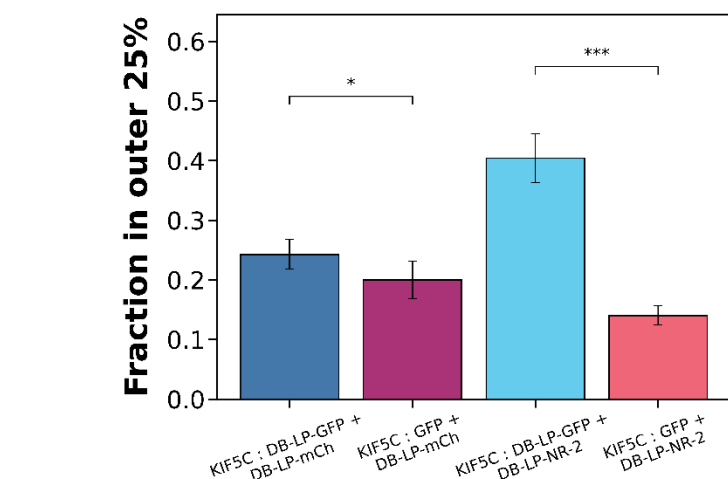
115 **Supplementary Figure 15.** Confocal fluorescence images of live HeLa cells expressing DB-
 116 LP-NR-2 stained with Nile Red (NR).

117

118



120 **Supplementary Figure 16.** Confocal fluorescence images of live HeLa cells co-expressing
 121 DB-LP-NR-1 with A) DB-LP-GFP, and B) KIF5C:DB-LP-GFP, and C) KIF5C:GFP fusions or
 122 D) DB-LP-NR-1 with KIF5C:GFP, and stained with Hoechst 33342 and Nile Red.



124 **Supplementary Figure 17.** Quantification of the distribution of the free DB construct (75%
 125 by DNA) in the presence of an overexpressed motor either fused to a dummy seed (GFP)
 126 or to a full length DB-LP-GFP (25% by DNA). Intensity in the outer 25% of cell was measured
 127 in a minimum of 25 cells.

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