

Supporting materials

Supporting table: PCR primers.

Genes		Forward (from 5' to 3')	Reverse (from 5' to 3')	NCBI reference seq.
Guinea pig	<i>Ar</i>	CTACTACAACTTTCCGCTACCC	CAGAGTCATCCCTGCTTCATAG	XM_063245991.1
	<i>Esr1</i>	AGCGACAAGGGAAGCATAAC	CTGATCGTGGAGGGTCAAATC	XM_003468423.3
	<i>Actb</i>	AGAGGTATCCTGACCCTGAAA	AGCACTGTGTTGGCATAGAG	NM_001172909.1
	<i>Egf</i>	CACAGCCATCCTTTCCATCT	TACCCGCCAGGACAATAAATC	XM_013150606.3
	<i>Klk1</i>	CGTGCTACAAGGCATCATCT	CTCGTGGTACTCTGTCACTTTG	XM_003465406.4
	<i>Klk2</i>	GGTTCTGTGCCTTGCTCTATTC	CTGGGAATGCTTCTCACACTC	XM_063252406.1
	<i>Ccnd1</i>	TCGATTCCAACGCACTCTTATC	CCTCCCTTCAACACTTCCTAAA	XM_003468321.5
	<i>Ctnnb1</i>	CTGACCTGTAACCCCTCCTTTAG	GCAAGGCTAGGGTCTGATAAA	XM_063260962.1
	<i>Wisp2</i>	CTGTAAACTCATTAGGAGGCTATGT	AATTCAGCCACTCAGTTCTCTC	XM_063248285.1
	<i>Wnt4</i>	CTAAGGCCTAGGGCTGAATAAA	GGATGTCACAATGGCAAGAAG	XM_063235714.1
	<i>Gapdh</i>	ACAGTGACAGCCATTCTTCC	AGCCGAACTCATTGTCATACC	XM_063237559
Mouse	<i>Ar</i>	CATGCACAAGTCTCGGATGTA	GAAGGAGAAGCCCAGGAAATAG	NM_013476.4
	<i>Esr1</i>	CTGGCTGGAGATTCTGATGATT	CGTAGCAAAGGGAGAAAGAGAG	NM_001302531.2
	<i>Actb</i>	GAGGTATCCTGACCCTGAAGTA	GCTCGAAGTCTAGAGCAACATAG	NM_007393.5
	<i>Egf</i>	ACTGGTGTGACACCAAGAGGTC	CCACAGGTGATCCTCAAACACG	NM_010113.4
	<i>Klk1b3</i>	GGATGGAGGCAAAGACACTTGC	TGGTGTAGACGCCTGGCATATC	NM_008693.2
	<i>Klk1b21</i>	GGAGTCCTGTTGAACCCCAACT	TGACCAATCGGTGCTGAGCAGA	NM_010642.3
	<i>Ccnd1</i>	GCAGAAGGAGATTGTGCCATCC	AGGAAGCGGTCCAGGTAGTTCA	NM_007631.3
	<i>Ctnnb1</i>	GTTCGCCTTCATTATGGACTGCC	ATAGCACCTGTTCCCGCAAAG	NM_007614.3
	<i>Wisp2</i>	TGTGTGACCAGGCAGTGATGCA	CAGGCTGTGCTCCAGTTTGGAC	NM_016873.2
	<i>Wnt4</i>	GAGAACTGGAGAAGTGTGGCTG	CTGTGAGAAGGCTACGCCATAG	NM_009523.2
	<i>Gapdh</i>	CATCACTGCCACCCAGAAGACTG	ATGCCAGTGAGCTTCCCGTTCAG	NM_008084.4

Ar, *Esr1* and *Actb* primers are reverse transcription PCR (RT-PCR) primers, and the products span introns. All other primers are quantitative PCR (qPCR) primers. Guinea pig qPCR primer sequences were designed using PrimerQuest tool from Integrated DNA Technologies, Inc. (IDT, Coralville, Iowa, USA). Mouse qPCR primer sequences were obtained from OriGene Technologies, Inc. (Rockville, MD, USA).

Supporting figures

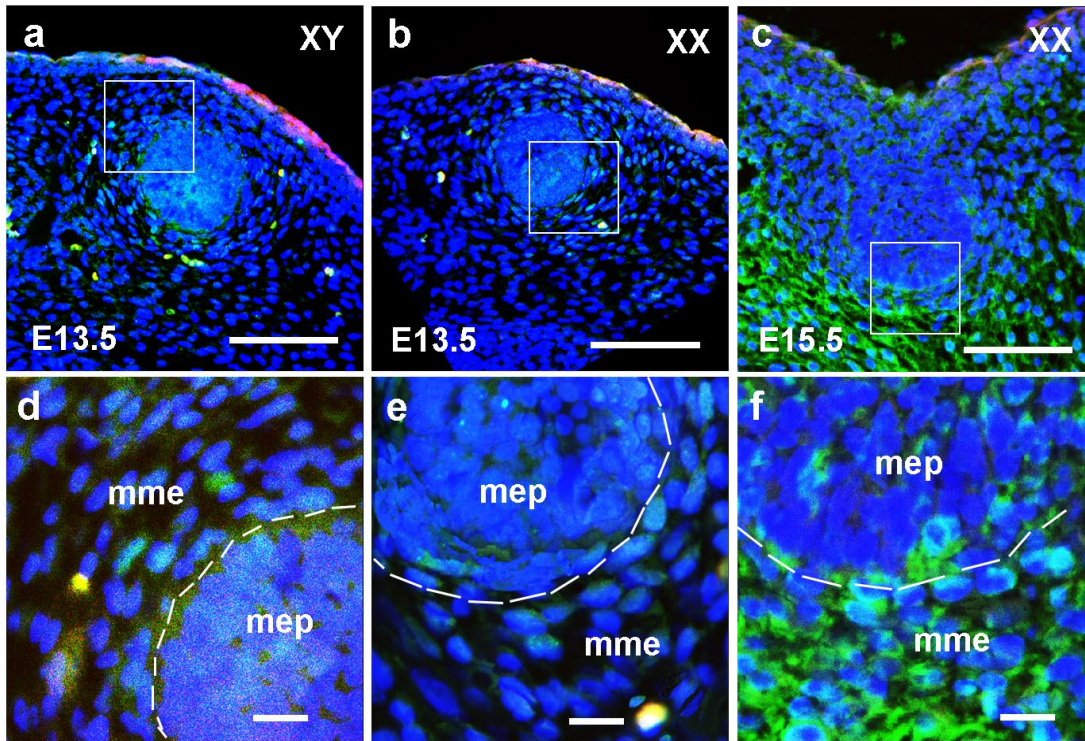


Figure S1. ER α protein localization in developing mammary glands (MGs) of embryonic mice.

(a-c) ER α immunofluorescence in MGs of E13.5 male (a), female (b) and E15.5 (female) mice. ER α signal is green, and Dapi is blue. Red color is non-specific staining. (d-f) are amplified images from square-marked regions in (a-c), respectively. Stages are indicated at bottom left of each image, and sex is indicated by chromosome symbols XY (male) and XX (female) at top right of each image in all supporting figures. Broken white lines in (d-f) indicate boundary of mammary epithelium and mesenchyme. *mep*, mammary epithelium; *mme*, mammary mesenchyme. Scale bars: a-c, 20 μ m; d-f, 5 μ m.

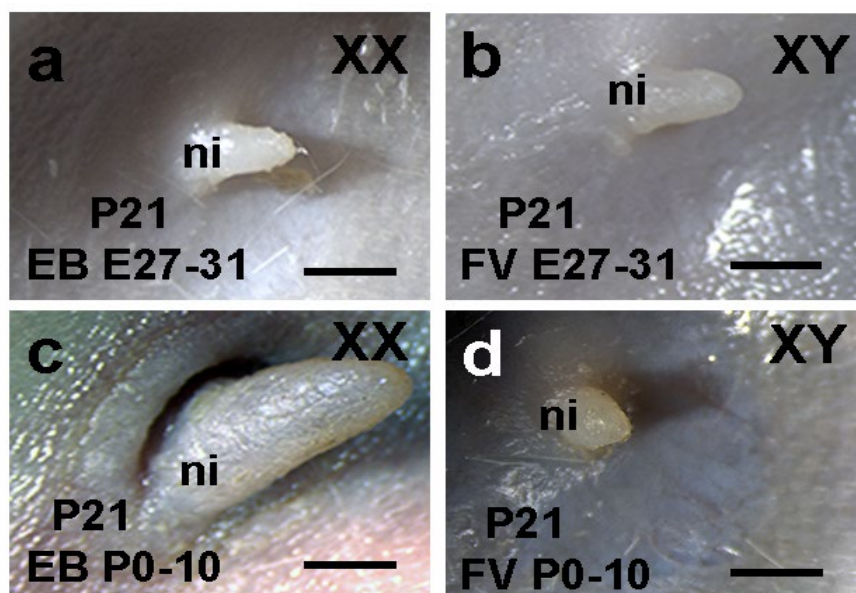


Figure S2. The effect of estrogen signaling on nipple development in guinea pigs.

Images show nipples of prenatally estradiol benzoate (EB)-treated female (A), prenatally fulvestrant (FV)-treated male (B), neonatally EB-treated female (C) and neonatally FV-treated male (D) P21 guinea pigs. Note that neonatal, but not prenatal EB treatment increased nipple size. *ni*, nipple. Scale bars: 2mm.

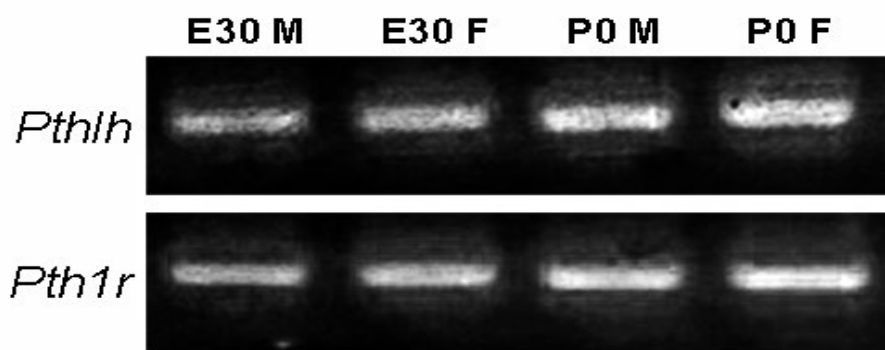


Figure S3. *Pthlh* and *Pth1r* expression were detected in developing MGs of E30 and newborn (P0) guinea pigs of both sexes.

cDNA fragments of *Pthlh* and its receptor gene *Pth1r* were detected using RT-PCR followed by agarose gel electrophoresis and ethidium bromide staining. $n=3$. *M*, male; *F*, female.