

**Additional file 2.** Primer sequences and qRT-PCR conditions.

	<b>Forward</b>	<b>Reverse</b>
<b>18S</b>	TAGATAAAAGGTCGACGCGG	CCCAAAGTCCAAC TACGAGC
<b>PSII-W PGSC0003DMG400020141</b>	GGATGAAAGAATGAGCACAGAAGG	AGACCAAACCAAACCGAAGACT
<b>PSAK PGSC0003DMG400020505</b>	TTGTTGCTGGTAGATTGGGC	GGTCCCCTGTCTGTAAGCC
<b>PSII 5kDa PGSC0003DMG400010498</b>	ACGTCATCTCCATTCTAGTCCA	TCTTATTGCTTTGAATGCGTGG
<b>HSP70 PGSC0003DMG400030089</b>	GACGAGAAAGCAGCAGAAGAC	CCATGCACACACTAATCTCTGG
<b>PSI-H PGSC0003DMG400016504</b>	TCACCCCTACAAC TCCCTTCAGA	AGCCACCTCCCAATATCAAGA
<b>EF-1<math>\alpha</math></b>	ATTGGAAACGGATATGCTCCA	TCCTTACCTGAACGCCTGTCA

18S rRNA gene was used in RT-PCR analysis (standard conditions, Temperature of annealing = 52°C) to control quality of cDNA before qRT-PCR.

qRT-PCR program cycle parameters were set as follows: 50°C for 2 min (stage 1), 95°C for 10 min (stage 2), 95°C for 15 sec and 60°C for 1 min (stage 3 repeated 40 times), 95°C for 15 sec, 60°C for 15 sec and 95°C for 15 sec (stage 4). Melting curve analysis was performed to check specificity of the reactions. The relative expression levels were calculated using the  $2^{-\Delta\Delta CT}$  method (Livak and Schmittgen, 2001). EF-1 $\alpha$  was used as internal control. Data representing three biological replicates and three technical replicates are expressed as mean  $\pm$  standard deviation.

Reference

Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the  $2^{-\Delta\Delta CT}$  method. Methods. 2001;25:402-8.