

(Uppercase letters indicate RNA, lowercase letters indicate DNA)

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| Name | Sequences (5' to 3') [modification] | Length |
|--------------------------------|----------------------------------------|--------|
| PSD-95-G4 | GGGGAAAAGGGAGGGAUGGG | 20 |
| Ax647PSD95-G4 | [Alexa Fluor 647]-GGGGAAAAGGGAGGGAUGGG | 20 |
| PSD-95-G4 (10 % 8OG) | ααααAAAAαααAαααAUααα | 20 |
| PSD-95-G4 (20 % 8OG) | ββββAAAAβββAβββAUβββ | 20 |
| PSD-95-G4 (40 % 8OG) | γγγγAAAAγγγAγγγAUγγγ | 20 |
| CaMKIIα-G4 | UGGGGGGGGCGGGUGGGA | 18 |
| Ax647CaMKIIα-G4 | [Alexa Fluor 647]-UGGGGGGGGCGGGUGGGA | 18 |
| CaMKIIα-G4 (10 % 8OG) | UααααααααCαααUαααA | 18 |
| CaMKIIα-G4 (20 % 8OG) | UββββββββCβββUβββA | 18 |
| CaMKIIα-G4 (40 % 8OG) | UγγγγγγγγCγγγUγγγA | 18 |
| C9ORF72 HRE ₄ | ggggccggggccggggccggggcc | 24 |
| Teromere ₄ | gggttagggttagggttagggtta | 24 |
| cMyc | ggggaggggtggggaggggtggggt | 23 |
| UG ₁₂ | UGUGUGUGUGUGUGUGUGUGUGUG | 24 |
| G ₃ UG ₃ | GGGUGGG | 7 |

α = 8-oxoguanine (10 %); RNA G (90 %)

β = 8-oxoguanine (20 %); RNA G (80 %)

γ = 8-oxoguanine (40 %); RNA G (60 %)

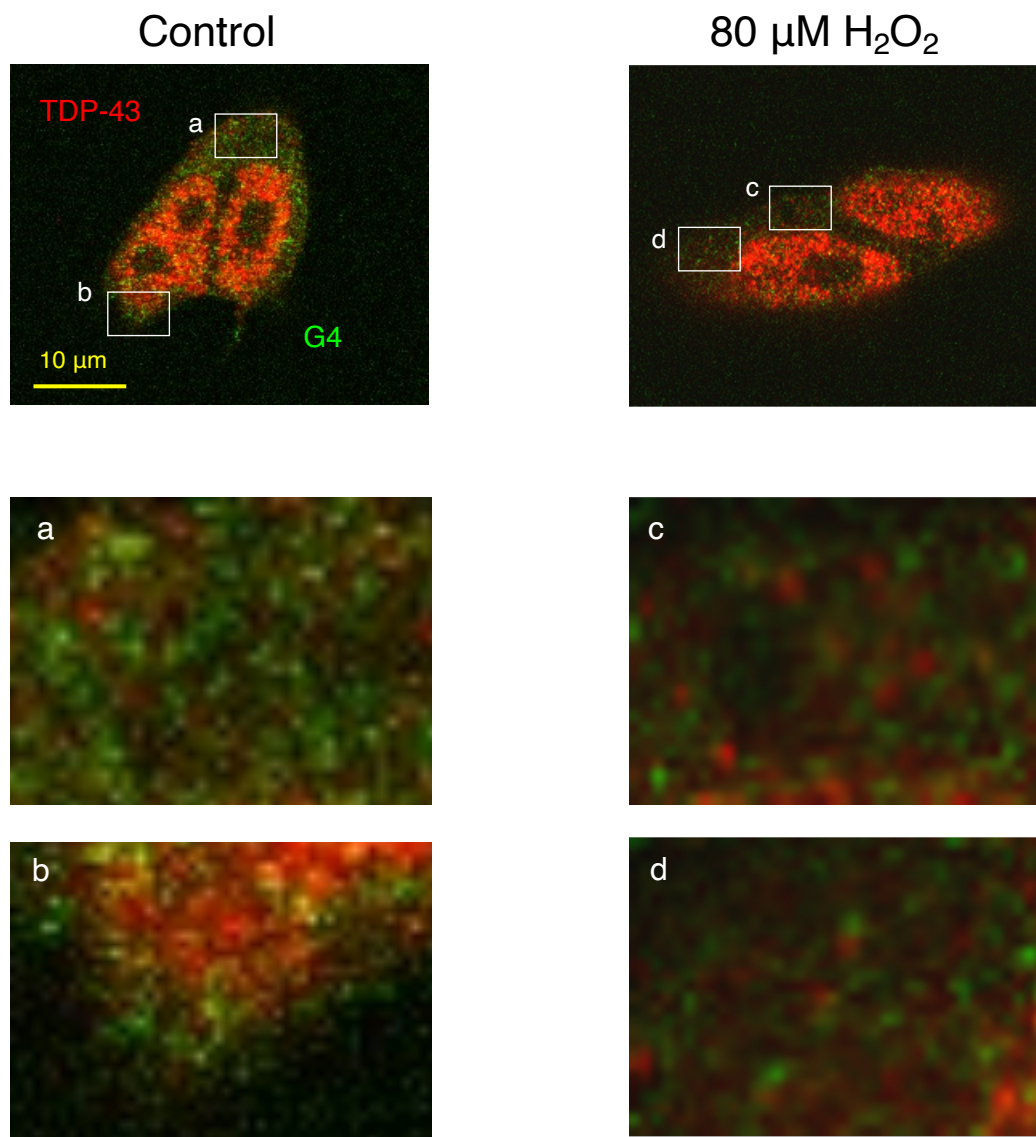


Figure S1. High-magnification image showing the localization of TPD-43 and G4 (related to Fig. 1B).

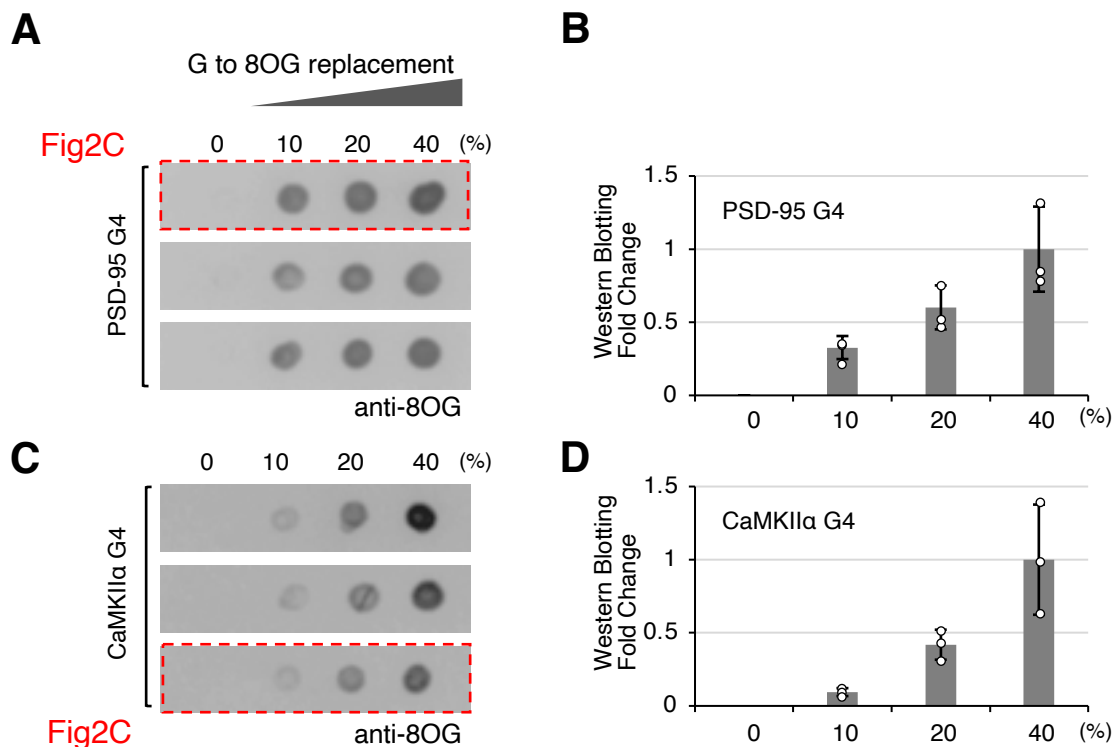


Figure S2. Dot-blot Western blotting of 8OG-modified RNAs. **(A, C)** The nonoxidized and oxidized forms of three types of RNAs (2.5 pmol each) derived from PSD-95-G4 and CaMKIIα-G4 were spotted onto a nylon membrane and analyzed by Western blotting. The regions outlined by the red dashed lines were used for quantification in Fig. 2C. **(B, D)** Experiments were performed in triplicate, and the data are presented as the mean \pm SEM.

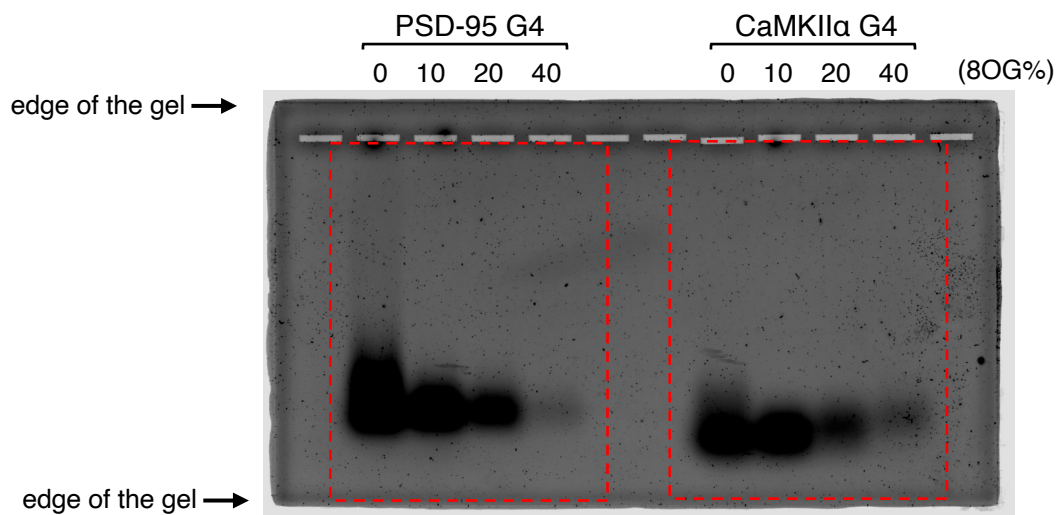


Fig2D

Figure S3. Agarose gel electrophoresis (related to Fig. 2D). The nonoxidized and oxidized forms of the three types of RNAs (25 pmol each) derived from PSD-95-G4 and CaMKIIα-G4 were electrophoresed on a 1% agarose gel containing $0.5 \times$ TBE under native conditions. The gel was stained with the intercalating fluorescent dye SYBR Green II. The regions outlined by the red dashed lines were used for Fig. 2D.

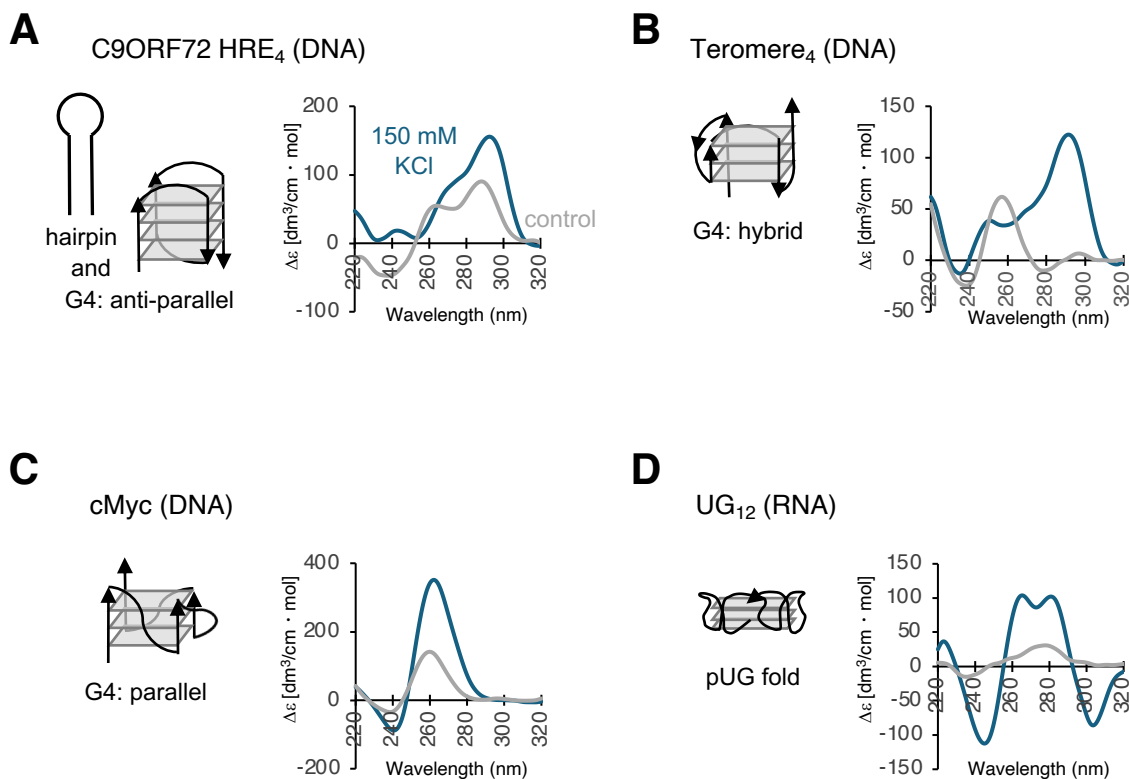


Figure S4. CD spectra of four different conformations. The predicted structures of the synthesized DNA/RNAs were confirmed by CD spectral analysis using 2 μ M oligonucleotides in the presence or absence (control) of 150 mM KCl. The measurements were repeated five times, and the average spectra are shown. **(A)** CD spectrum of C9ORF72 HRE₄ DNA (a mixture of hairpin and antiparallel G4 structures). **(B)** CD spectra of Teromere₄ DNA (hybrid G4). **(C)** CD spectra of cMyc DNA (parallel G4). **(D)** CD spectra of UG₁₂ fold (pUG fold). Traces are shown in the presence and absence of 150 mM KCl.

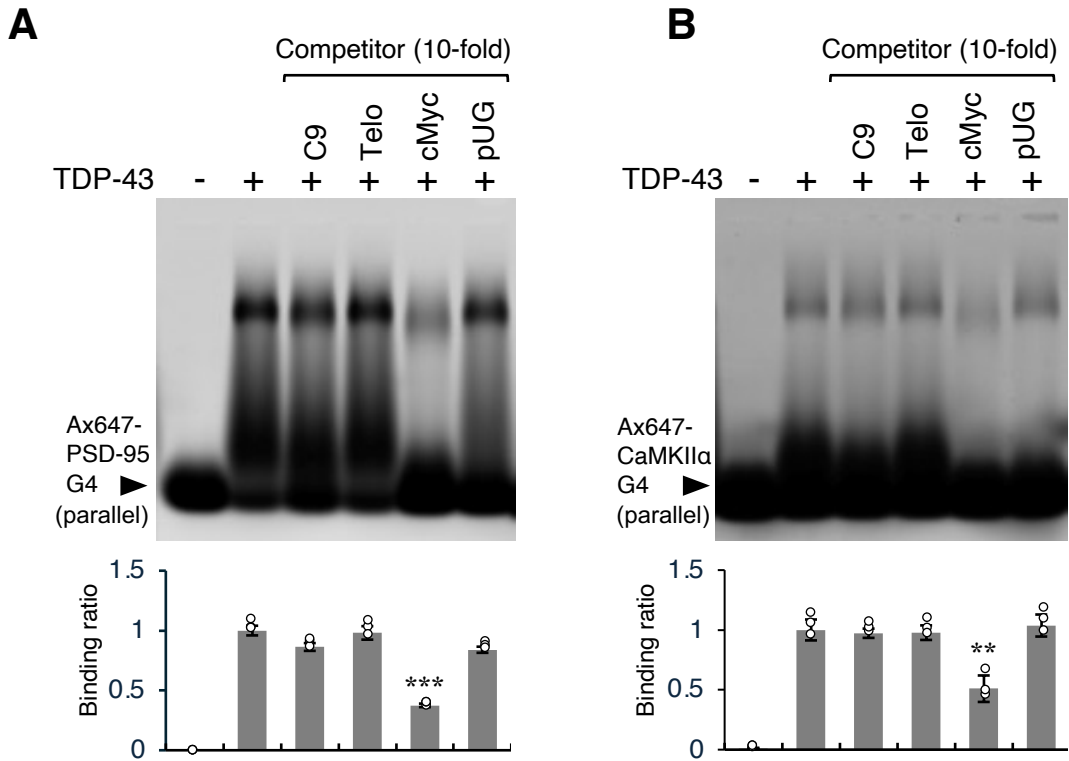


Figure S5. Binding specificity of TDP-43. 0.5 pmol of fluorescently labeled G4 probe (**A**; Ax647-PSD-95-G4 or **B**; Ax647-CaMKIIα-G4) was mixed with TDP-43 (2 pmol) and the indicated unlabeled competitors (10-fold for probe) and electrophoresed under nondenaturing conditions. Each overshifted band was quantified, and the mean and standard error (\pm SEM) were calculated from three independent experiments. The results are shown in the graph, with the y-axis representing the mean \pm SEM. Statistical significance was assessed using a two-tailed Student's *t*-test. ** $P < 0.01$, *** $P < 0.001$.

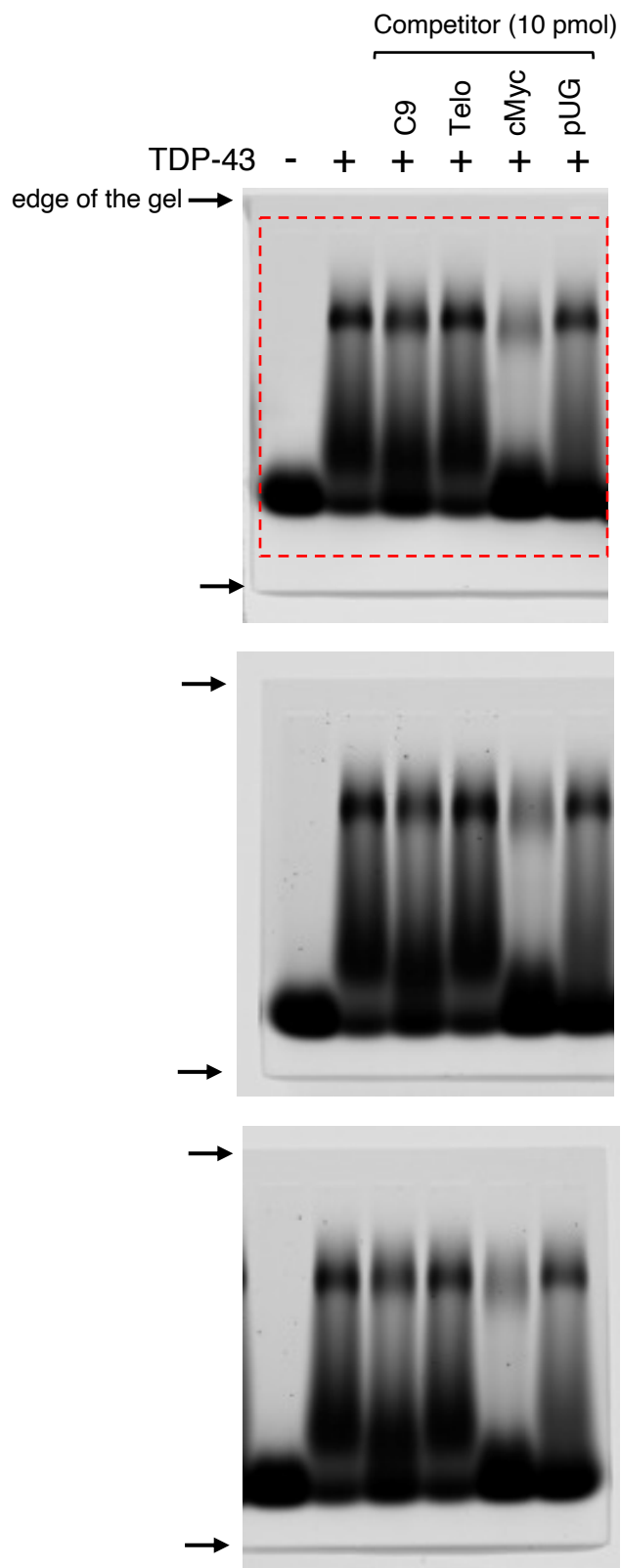


Fig. S5

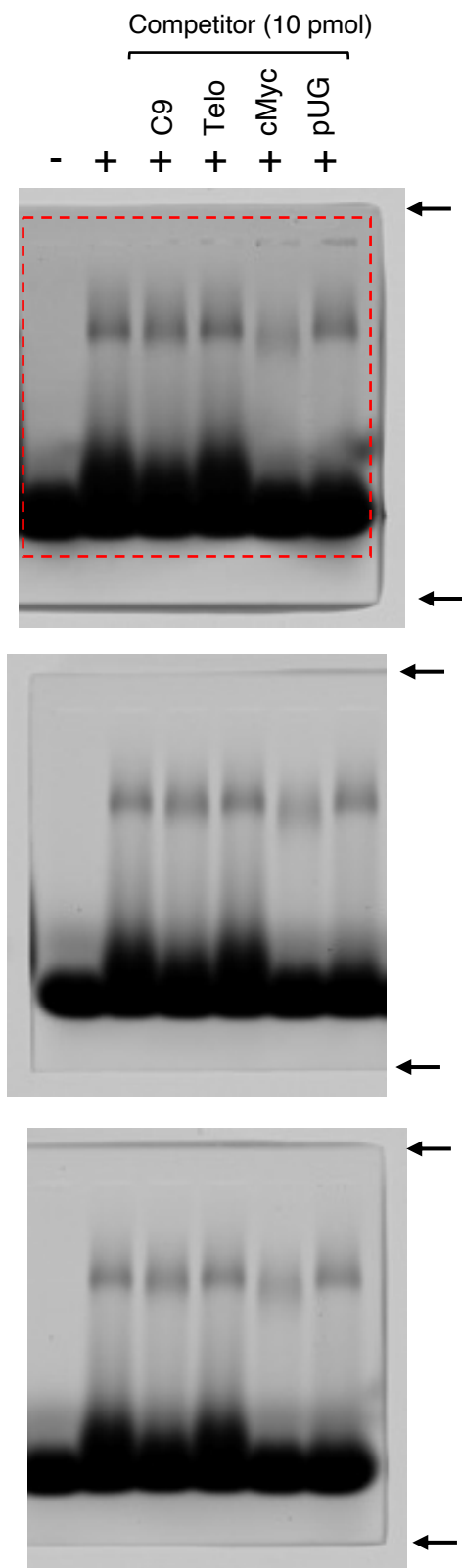


Figure S6. Original images of the gel shift assay (related to Fig. S5). The areas enclosed by the red dashed lines are used for Fig. S5.

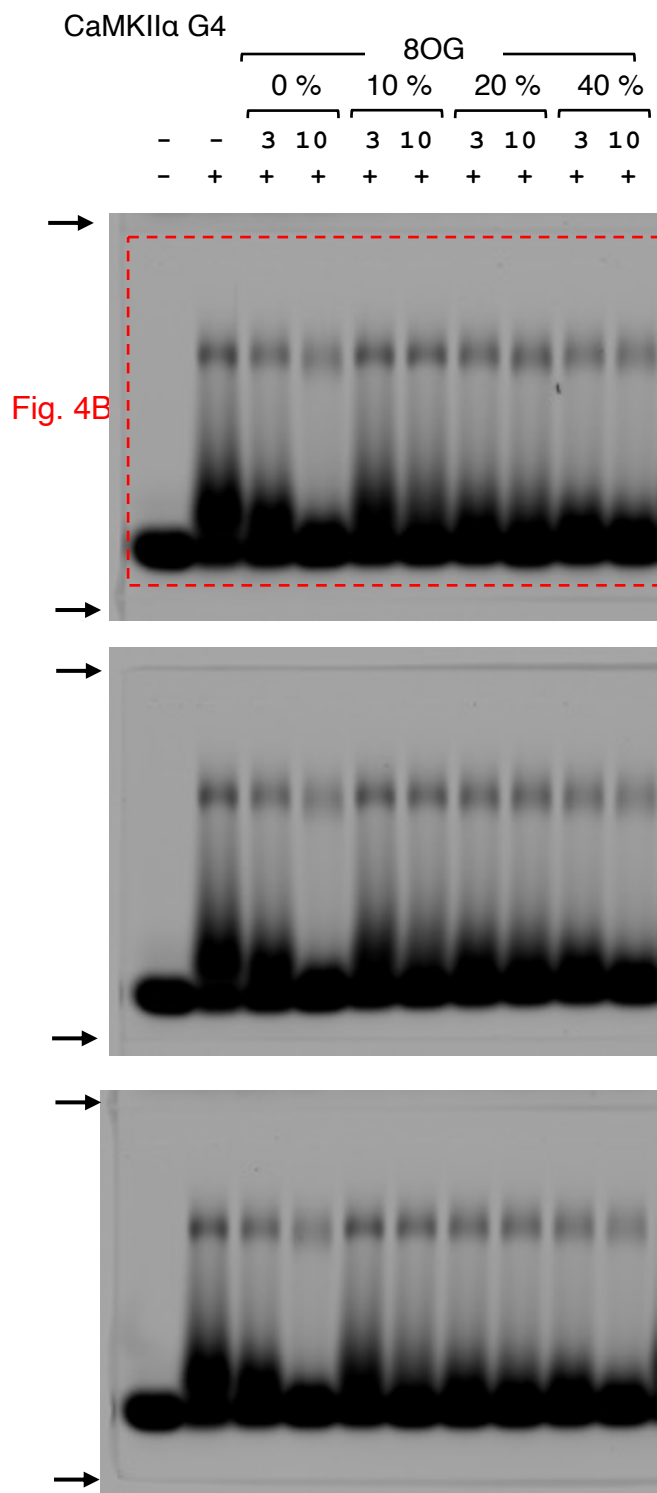
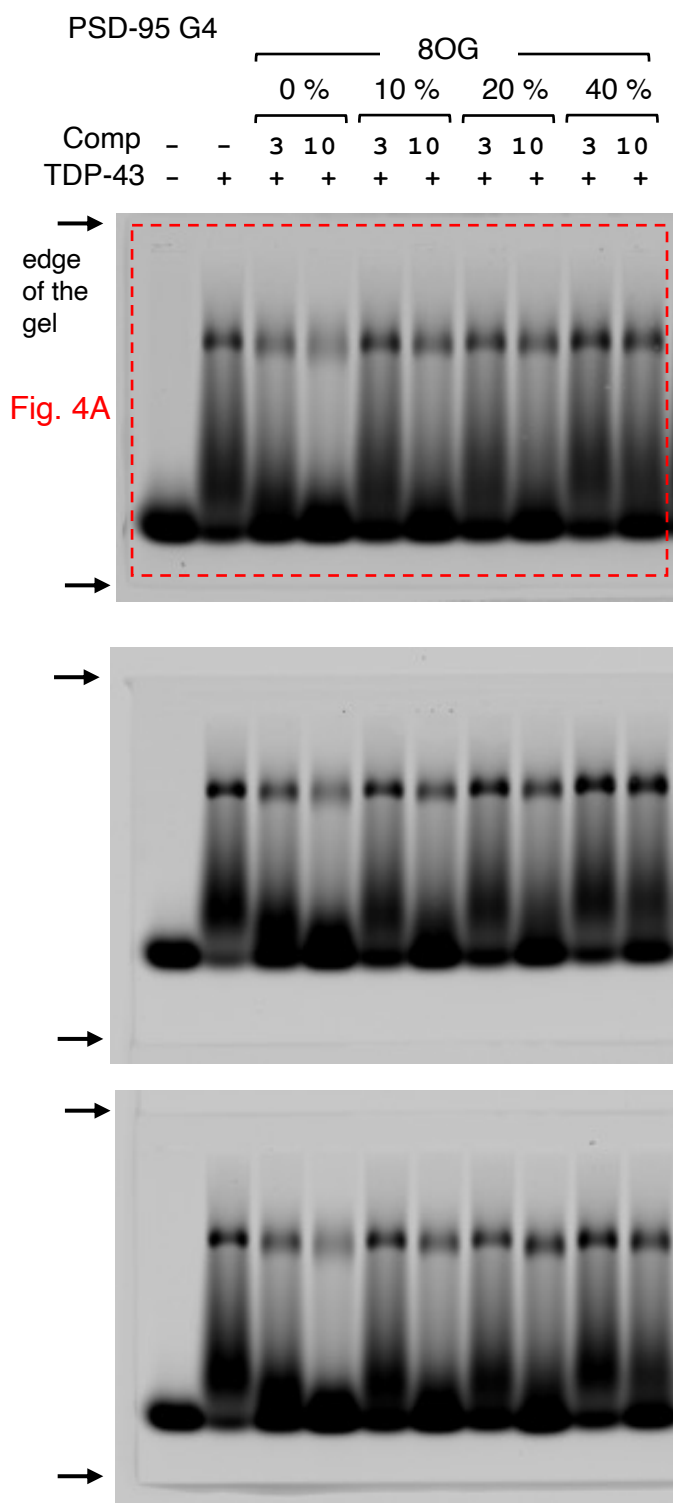


Figure S7. Original images of the gel shift assay (related to Fig. 4). The areas enclosed by the red dashed lines are used for Fig. 4A and B.

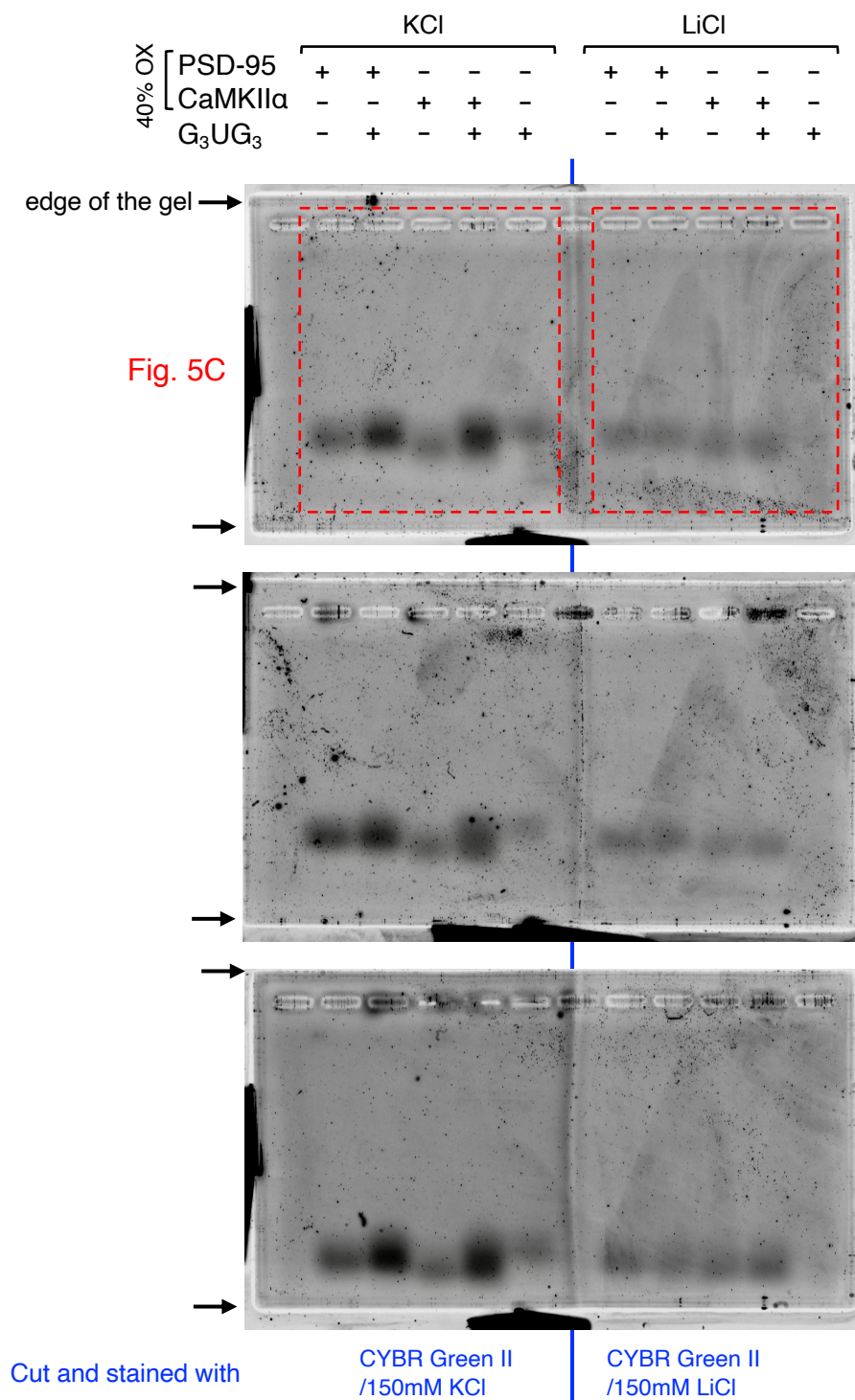


Figure S8. Original images of electrophoresis (related to Fig. 5C). The areas enclosed by the red dashed lines are used for Fig. 5C. The samples containing potassium or lithium were electrophoresed simultaneously on the same gel, cut upon completion, and stained with a CYBR Green II staining solution containing 150 mM KCl or LiCl.

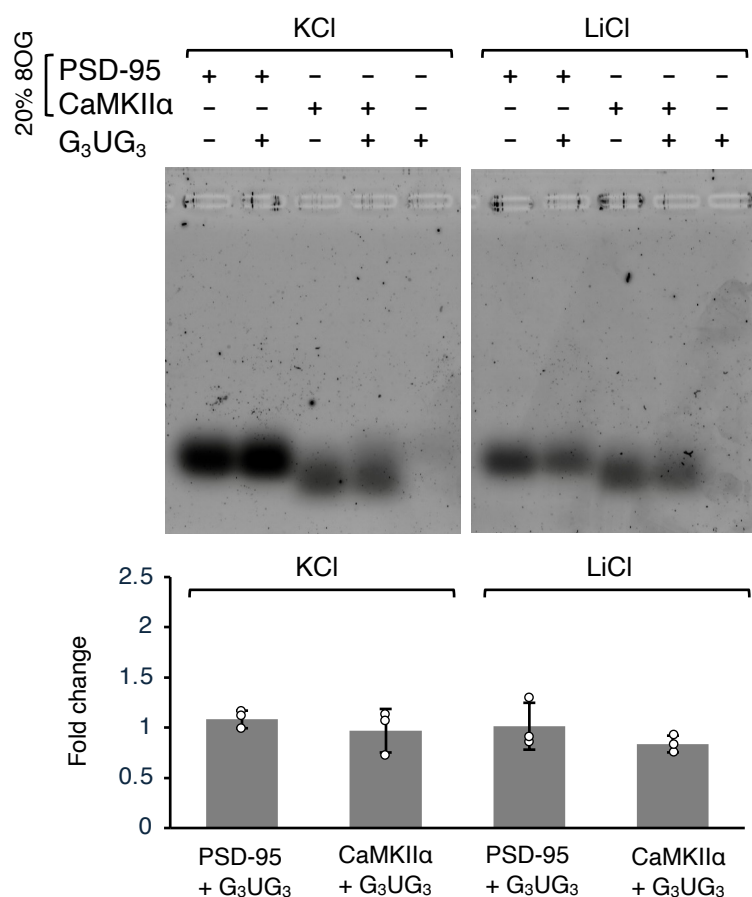


Figure S9. G4 formation assay. The indicated 20% oxidized RNAs (20 pmol) were mixed with the absence or presence of G₃UG₃ (20 pmol) and electrophoresed on 1% agarose gel under native conditions. The gels were stained with the intercalating fluorescent dye SYBR Green II, and the individual signals were quantified. The graph below shows the fold change after the addition of G₃UG₃, which was calculated by subtracting the signal for G₃UG₃ alone from the value for mixing 20% oxidized PSD-95 or CaMKII α RNAs with G₃UG₃. The experiments were performed in triplicate, and the data are presented as the mean \pm SEM.

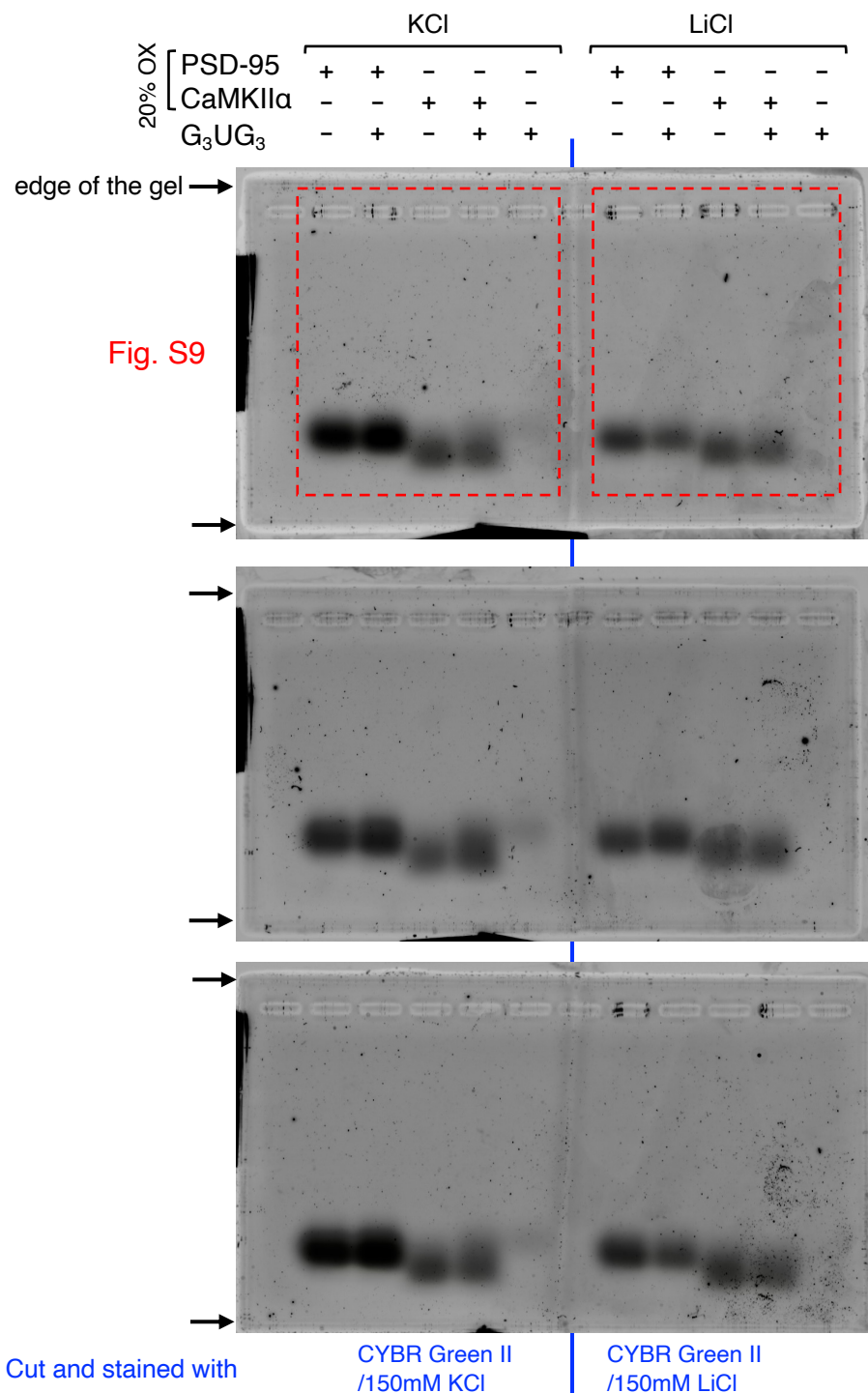


Figure S10. Original images of the gel shift assay (related to Fig. S9). The areas enclosed by the red dashed lines are used for Fig. S9. The samples containing potassium or lithium were electrophoresed simultaneously on the same gel, cut upon completion, and stained with a CYBR Green II staining solution containing 150 mM KCl or LiCl.

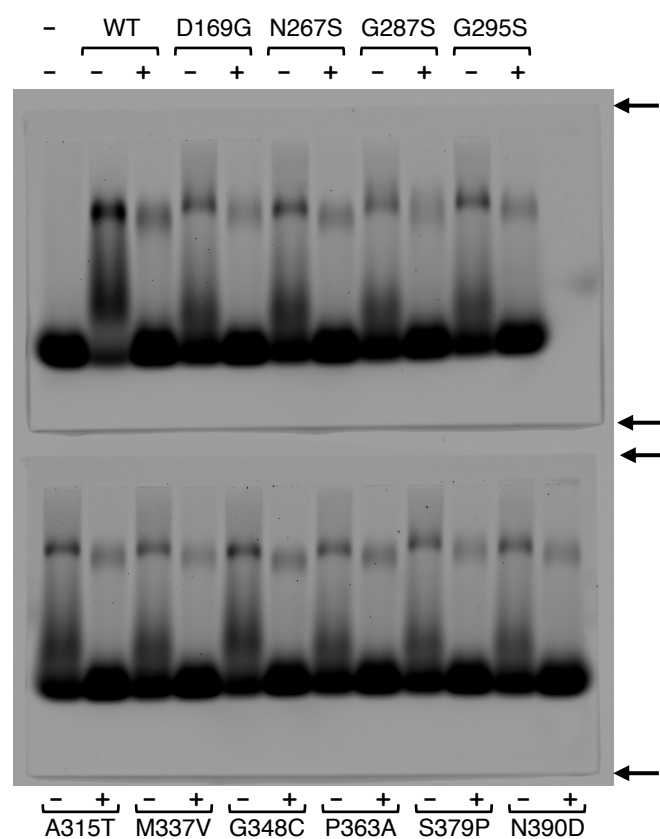
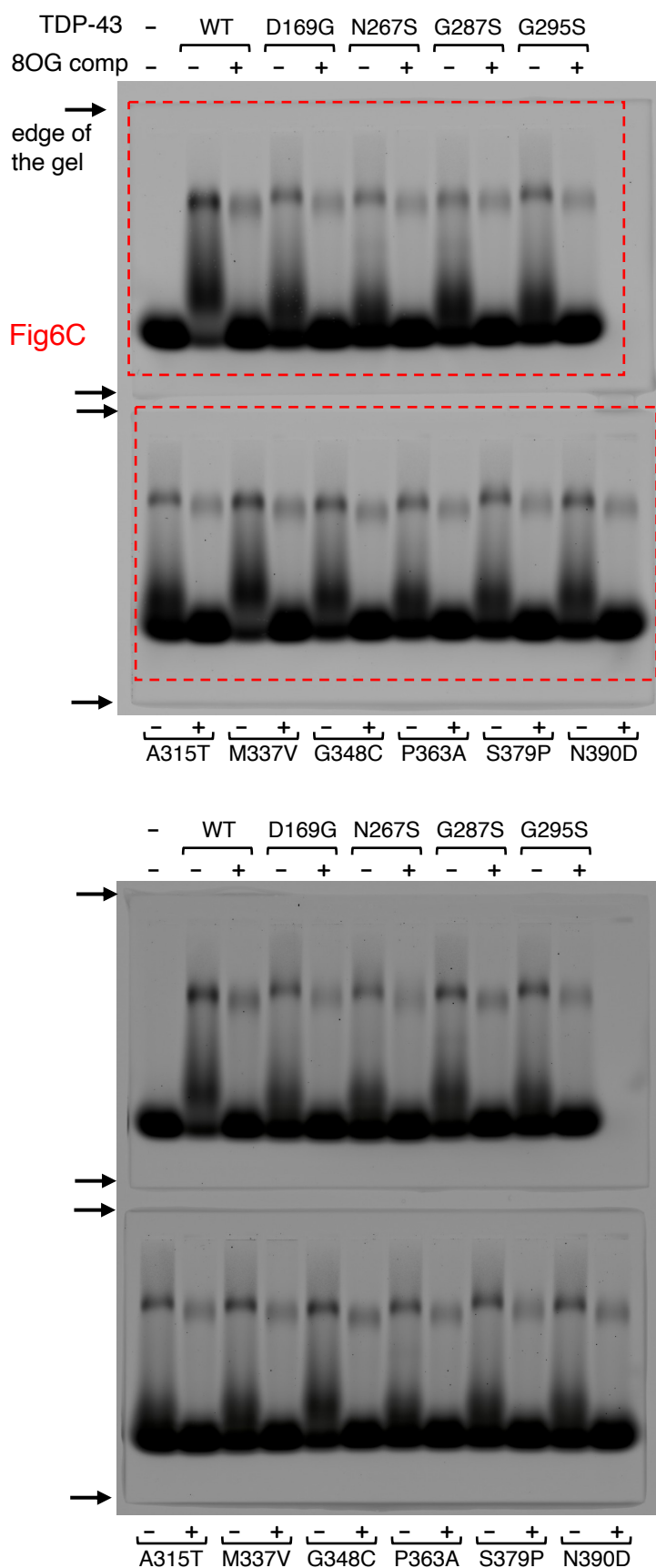


Figure S11. Original images of the gel shift assay (related to Fig. 6). The areas enclosed by the red dashed lines are used for Fig. 6C.