

## Supplementary Materials

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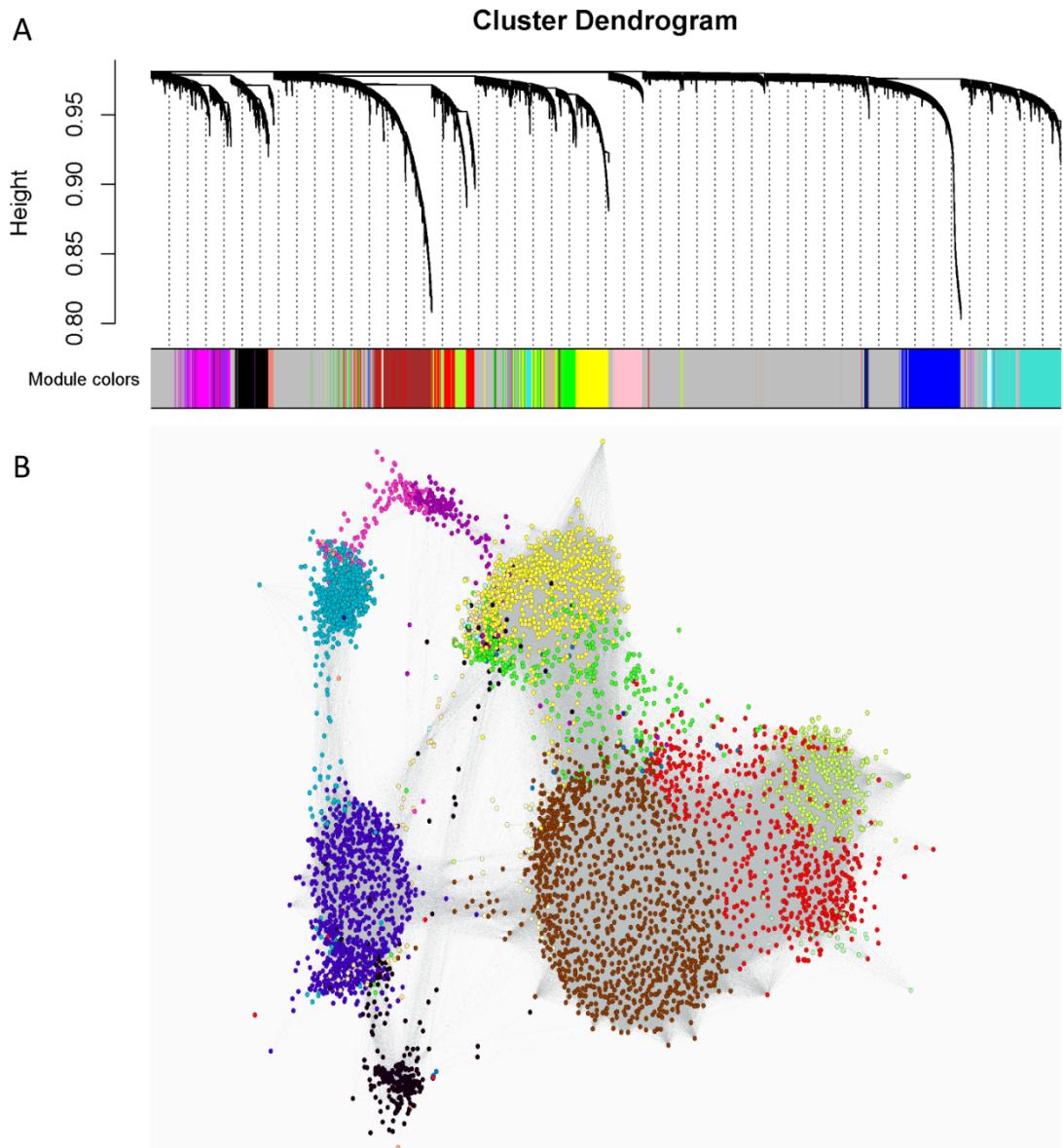
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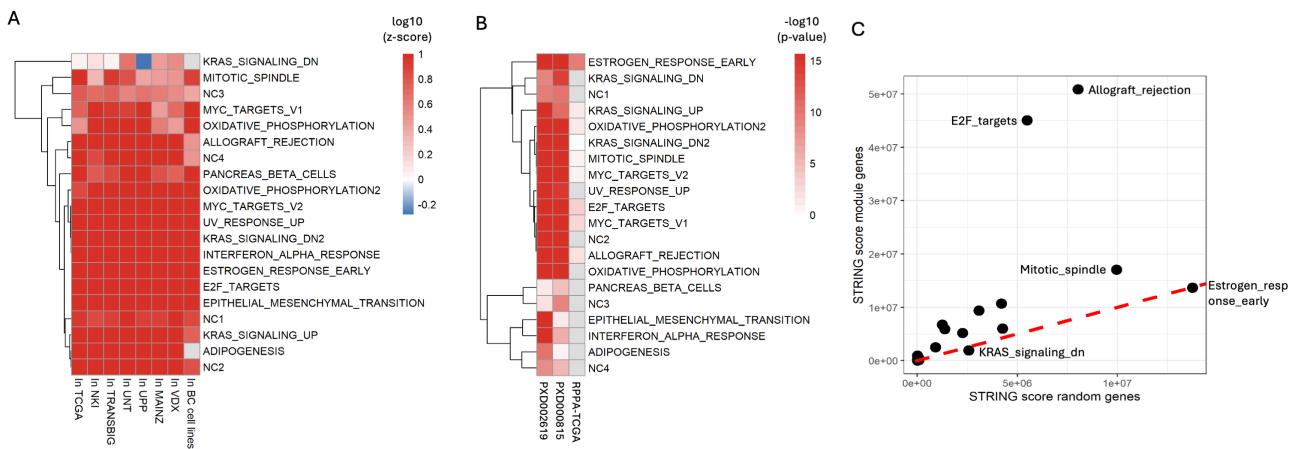
### Supplementary Material and Methods

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**Breast cancer coexpression networks show cross-dataset preservation and reflect protein-level coordination**

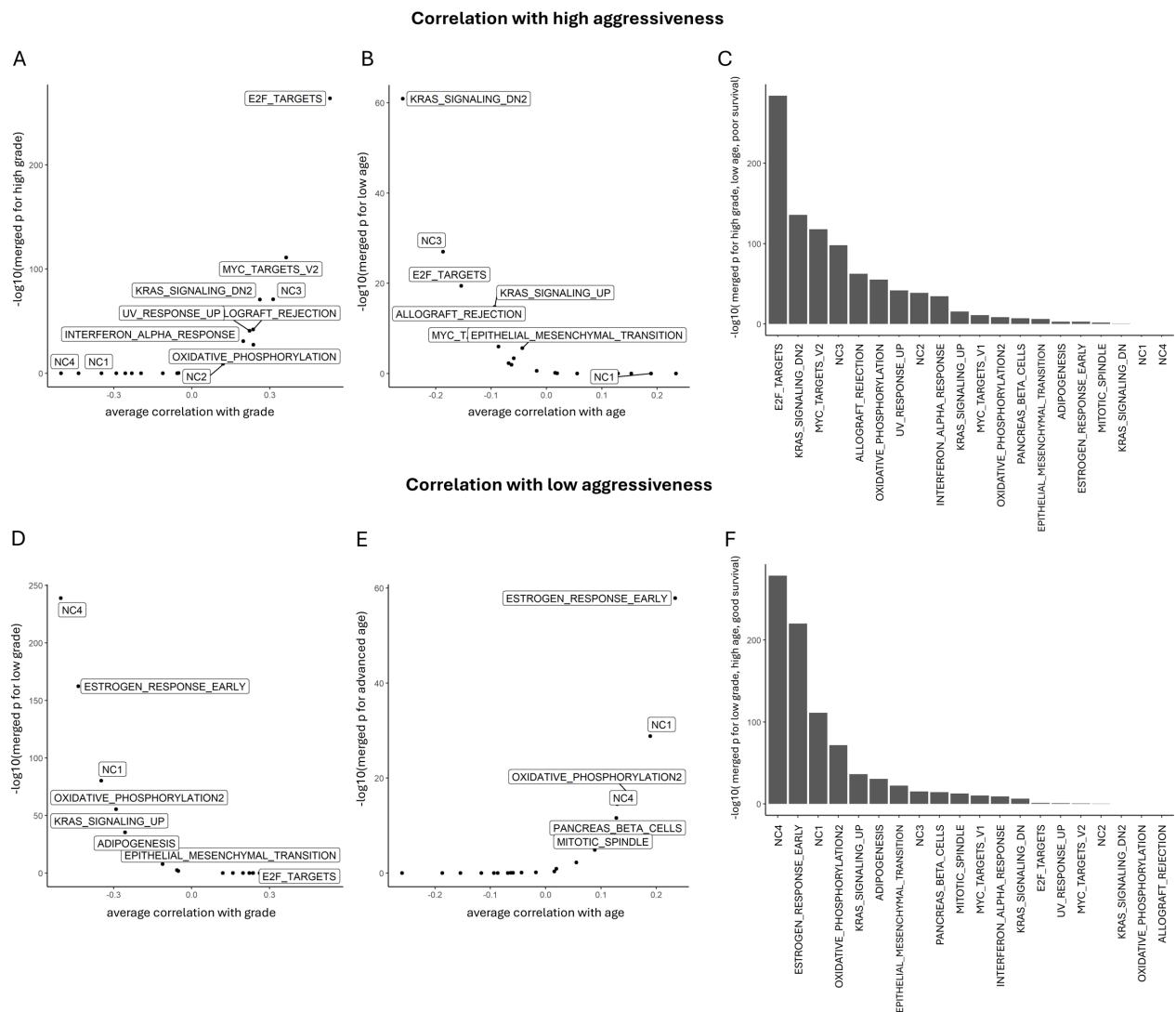


**Supplementary Figure 1. METABRIC gene co-expression network.** (A) Clustering of genes performed with WGCNA, each colour corresponding to a module. (B) Gephi representation of the network, where nodes are genes and edges are connections (only the strongest are represented, Topological Overlap  $\geq 0.05$ ). Different colours correspond to different modules.

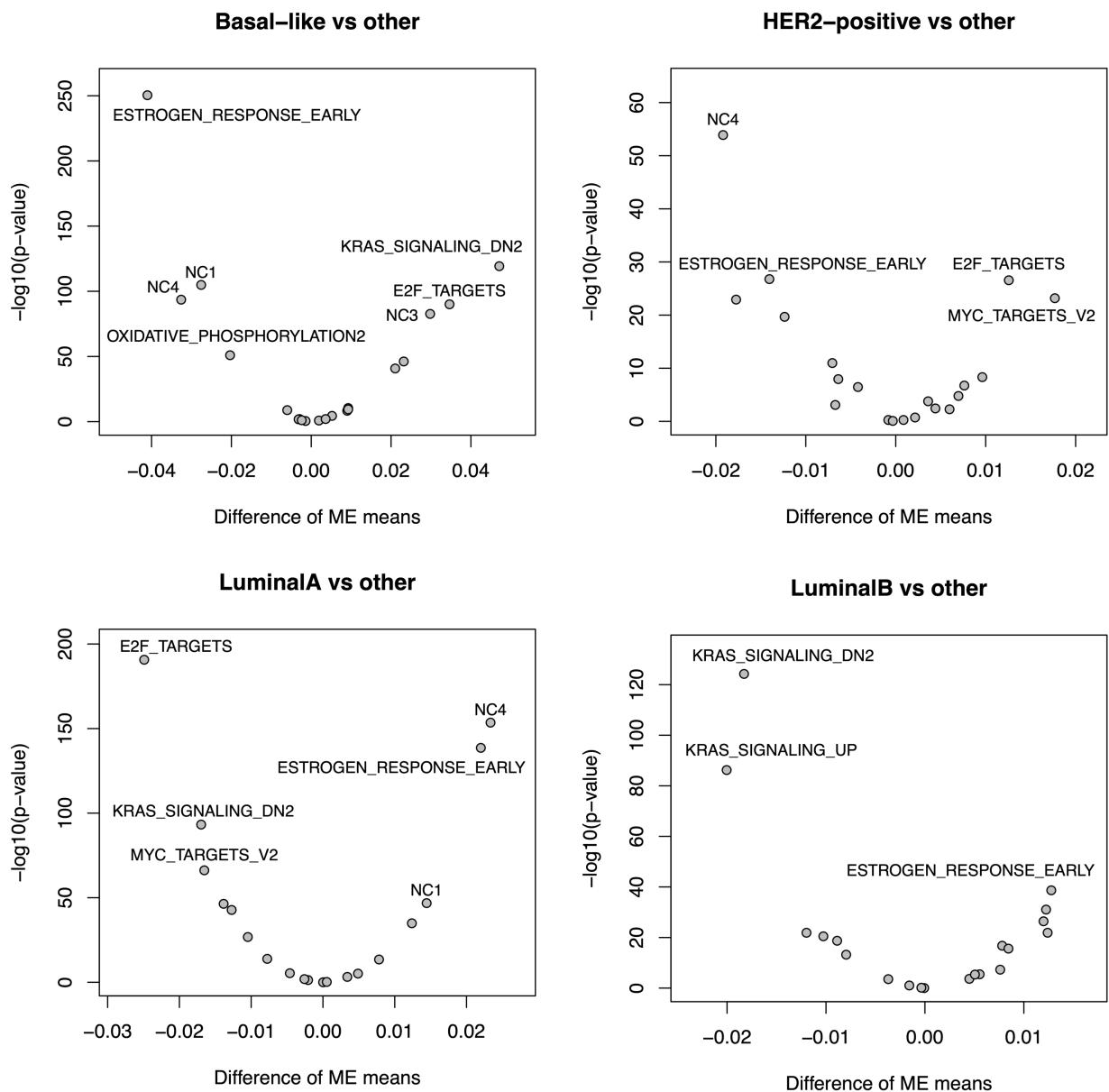


**Supplementary Figure 2. Breast cancer co-expression networks are robust.** **(A)** Heatmap summarizing the z-score of modules' preservation (rows) across 7 independent primary BC transcriptome datasets and one BC cell lines dataset (columns). The colour represents the  $\log_{10}$  of z-scores, with red corresponding to higher preservation. **(B)** Heatmap showing the connectivity of modules (rows) in proteomic datasets (columns). The colour indicates the  $-\log_{10}$  of the p-value for a Kolmogorov-Smirnov test comparing the correlations of modules' genes with those of random genes. **(C)** The proteins encoded by co-expressed genes are more tightly connected than random proteins in the STRING interaction database. Obtained by summing the scores of the STRING database for each pair of interacting genes in one module (y axis) and comparing it with the corresponding measure obtained with an equal number of random genes (x axis). Each dot represents a module. Some relevant modules' names have been included next to the corresponding dot.

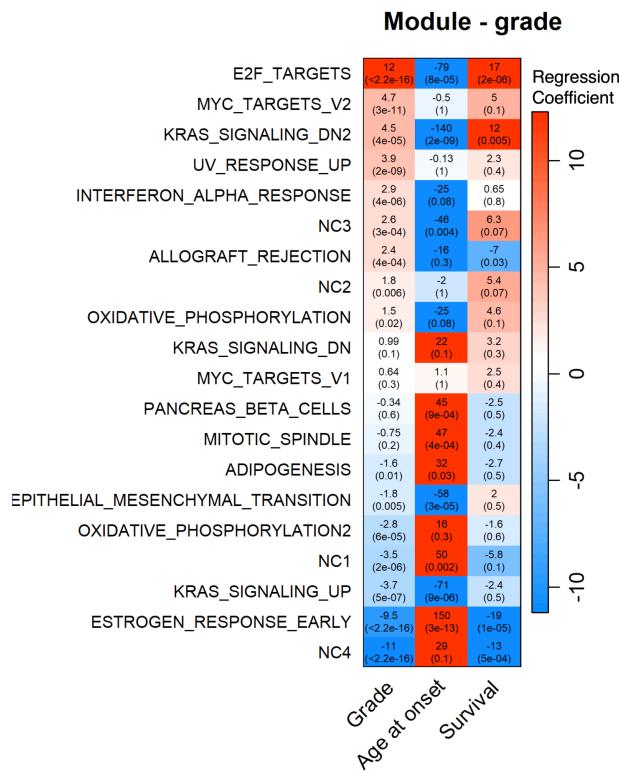
## Coexpression networks are clinically informative



**Supplementary Fig. 3. Summary of modules' relationship with clinical features.** **(A-C)** Modules' correlation with features of high aggressiveness: high grade **(A)**, early onset **(B)**, merged score for high grade, early onset and poor prognosis **(C)**. **(D-F)** Modules' correlation with features of low aggressiveness: low grade **(D)**, late onset **(E)**, merged score for low grade, late onset and good prognosis **(F)**. **(A,B,D,E)** For each clinical feature, p-values were obtained by merging the p-values across all the individual tested datasets with the Fisher's method, considering the direction of the relationship between module's expression and the clinical feature. **(C, F)** for each module, the overall significance for the relationship with grade, age at diagnosis and survival was obtained by multiplying the individual dataset-merged p-values.

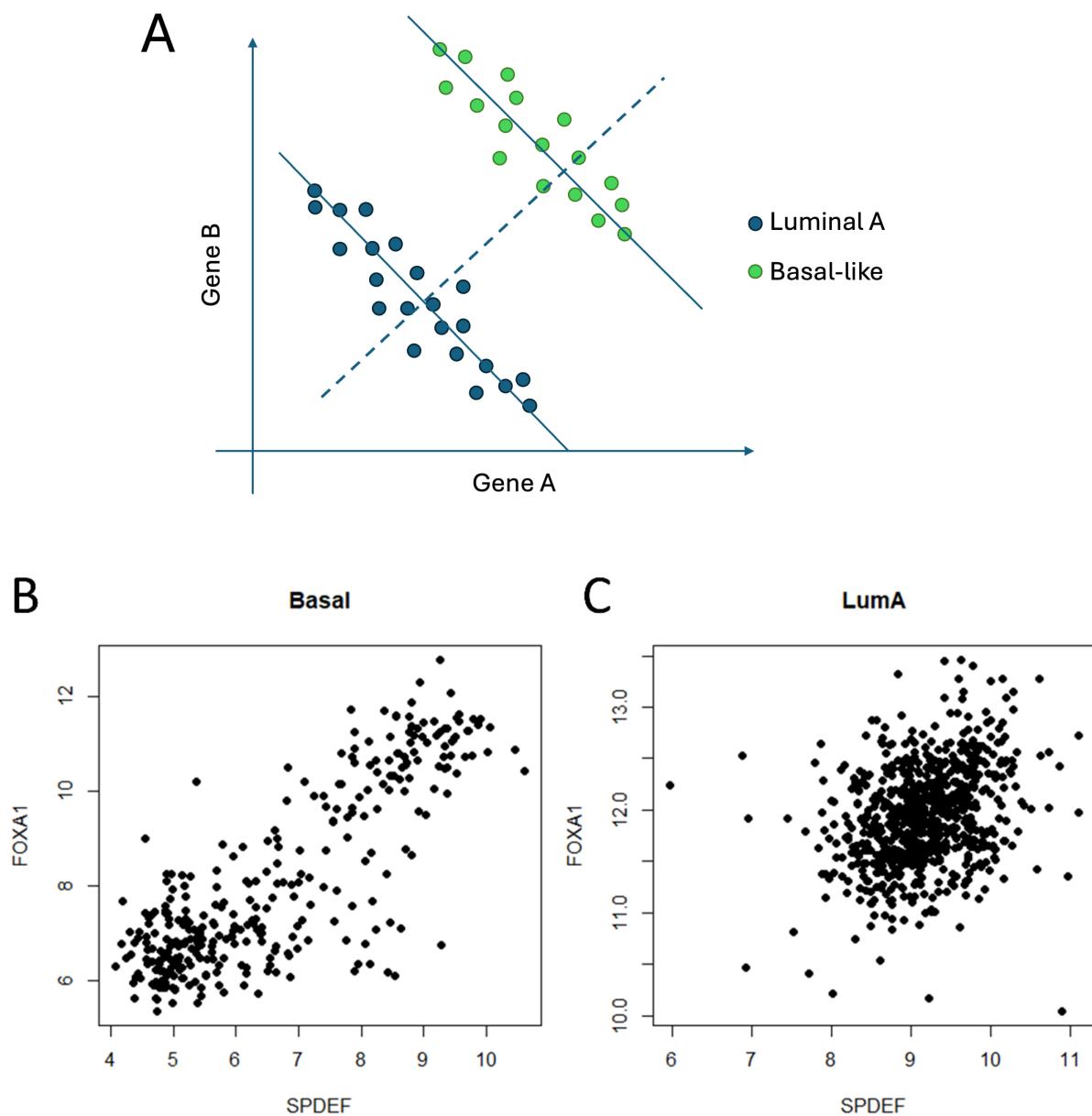


**Supplementary Figure 4. Expression of the modules across BC subtypes.** The expression of each module in a subtype (specified in the plot title) was compared with the expression of the same module in all other subtypes pooled together. The x-axis shows the difference between the module eigengene (ME) between the two groups, the y-axis represents the significance for the comparison. Note the overexpression of the modules *KRAS*\_*signaling\_dn2* and *E2F*\_*targets* in the basal-like tumors, of the *Estrogen*\_*response*\_*early* module in both luminal subtypes, and of the *E2F*\_*targets* in the HER2-positive tumors.  $p\text{-value} < 2.2 \times 10^{-16}$  in all cases.

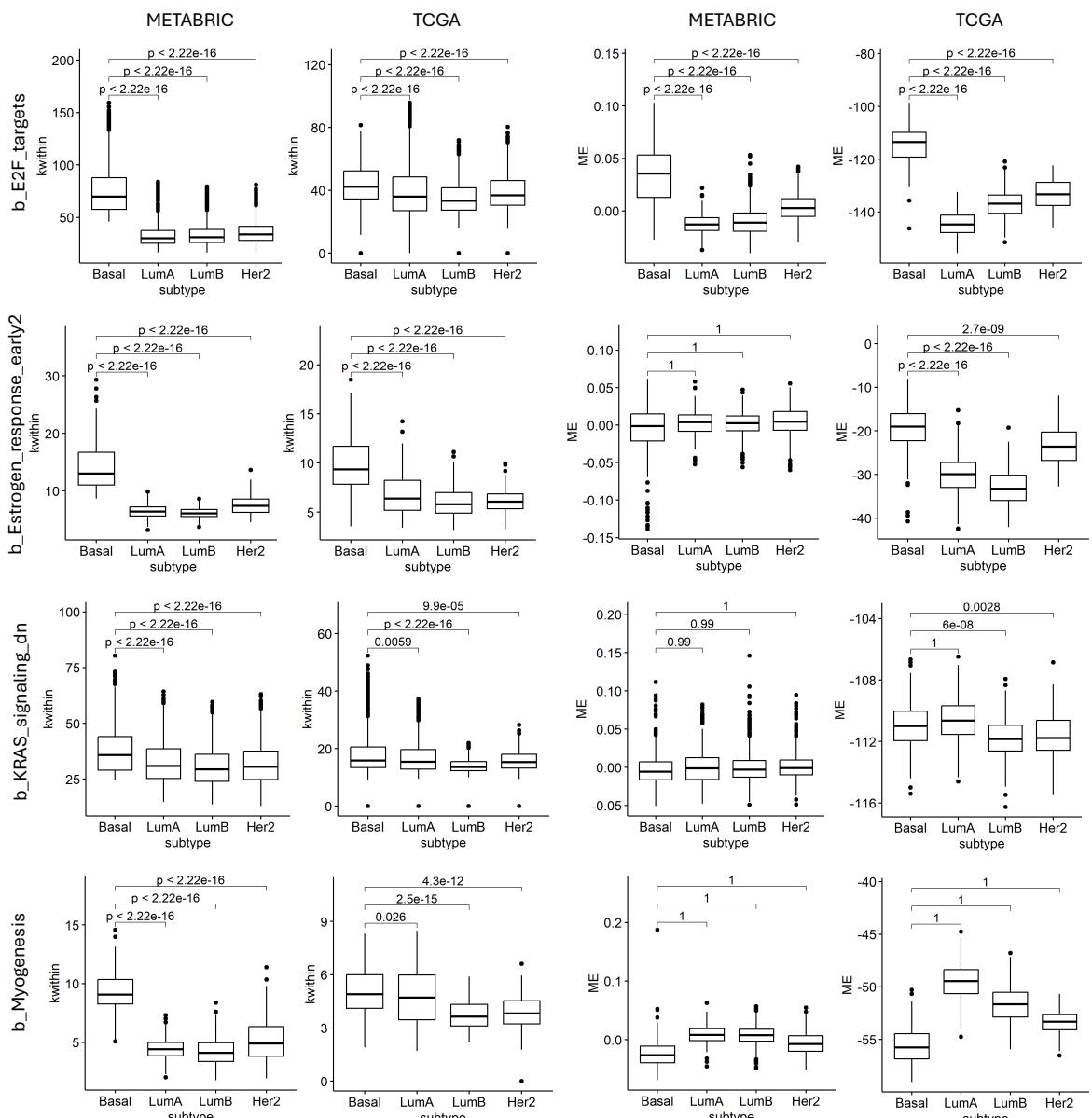
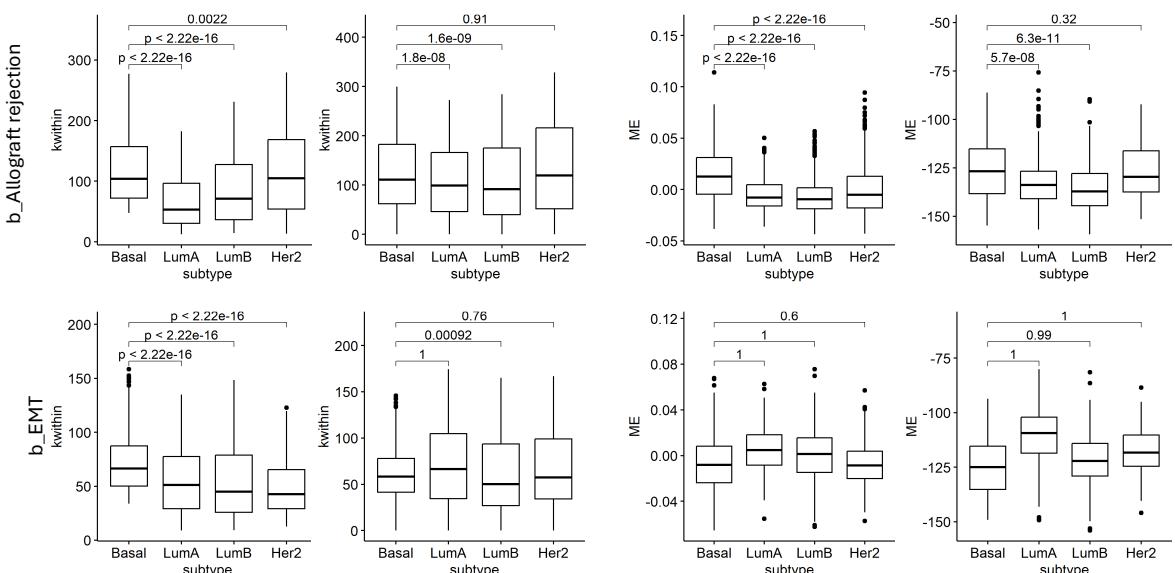


**Supplementary Figure 5. Modules correlate with clinical features independently of tumor subtype.** The heatmap and the enclosed numbers shows regression coefficients and corresponding p-values for module eigengenes (rows) in models with the indicated clinical features (columns) as dependent variables, and tumor subtype as a categorical covariate.

Disentangling subtype effects reveals robust and clinically informative basal-like coexpression modules

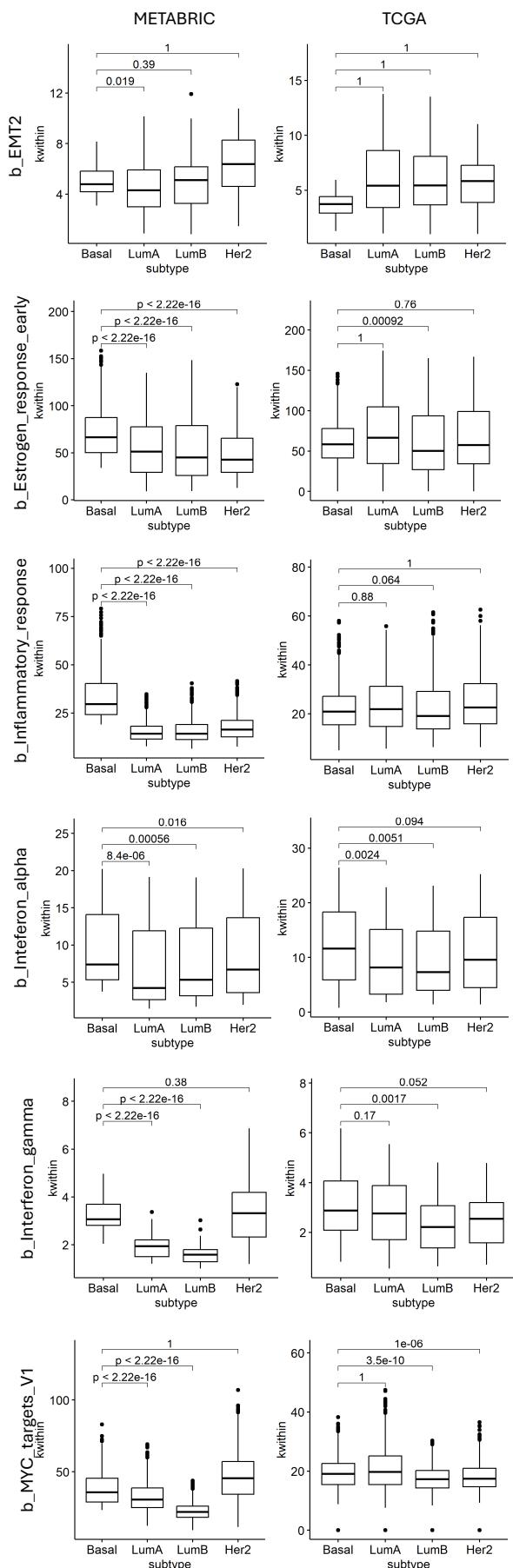


**Supplementary Figure 6. Exemplification of the Simpson's Paradox.** The Simpson's Paradox is a statistical phenomenon where an association between two variables in a dataset emerges, disappears or reverses when the data are divided into more homogeneous subgroups. **(A)** Two ideal genes, known to display overall positive correlation (not shown), are instead negatively correlated in Basal-like or Luminal A subtypes. **(B, C)** The correlation between FOXA1 and SPDEF is strikingly different between Basal-like tumors **(B)** and Luminal A tumors **(C)** in the METABRIC cohort.

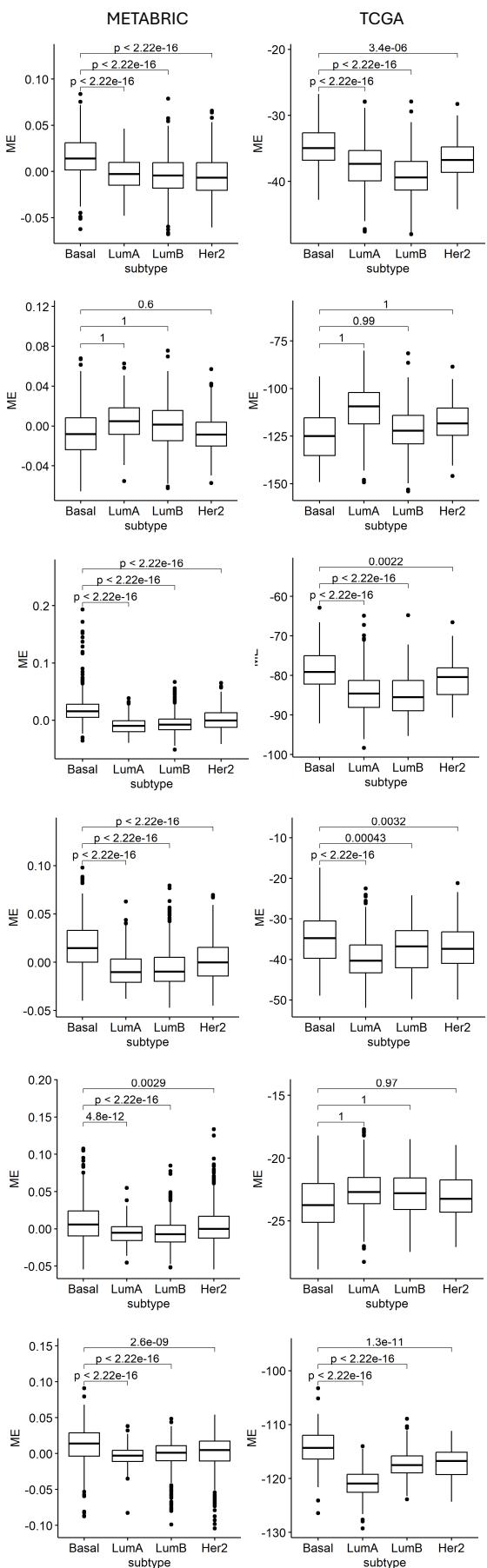
**A****Connectivity****Expression****B**

C

## Connectivity

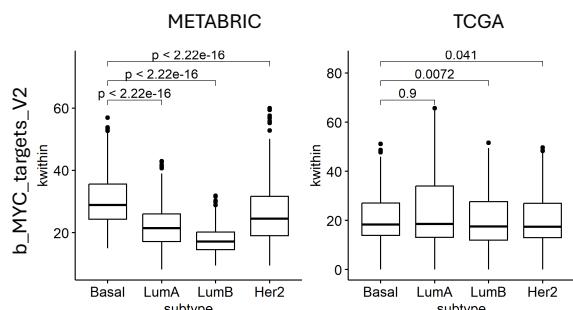


## Expression

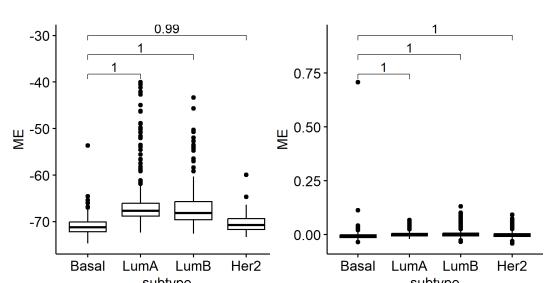
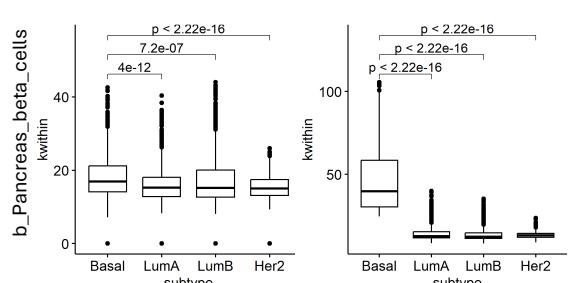
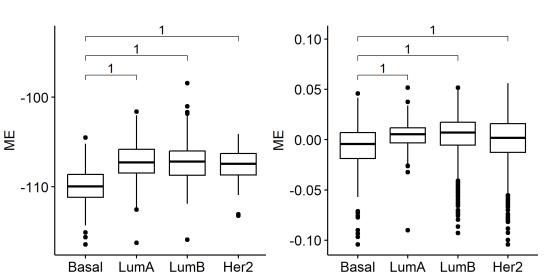
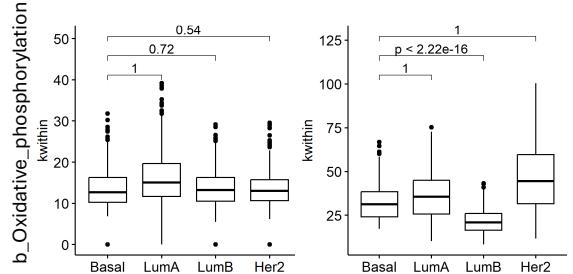
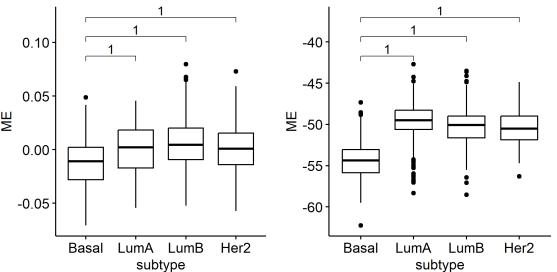
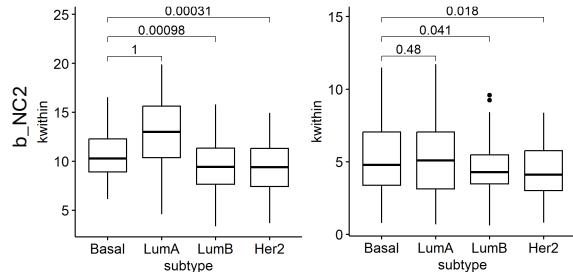
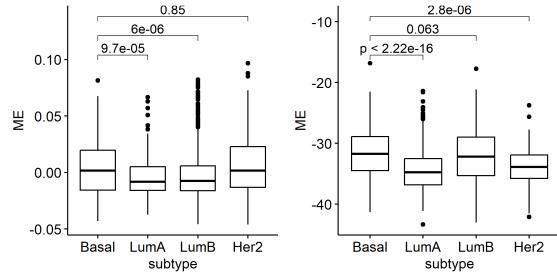
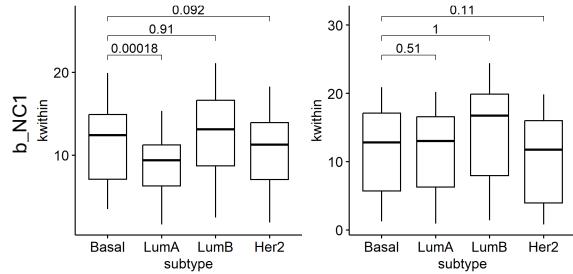
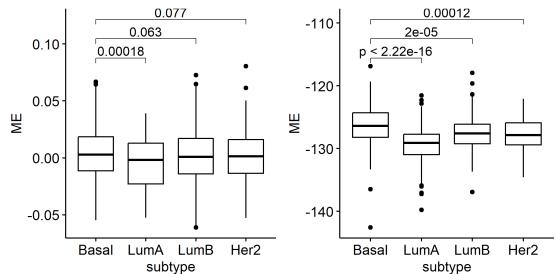
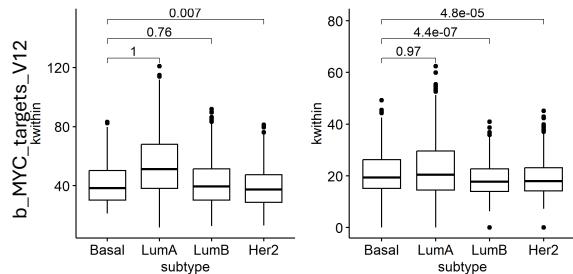
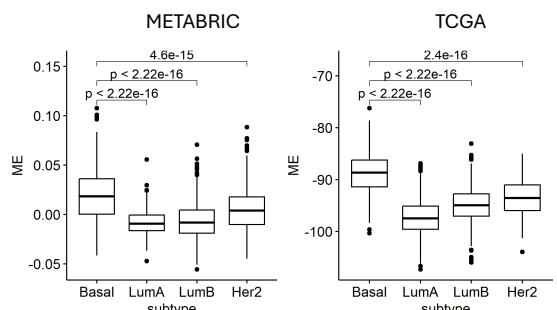


D

## Connectivity



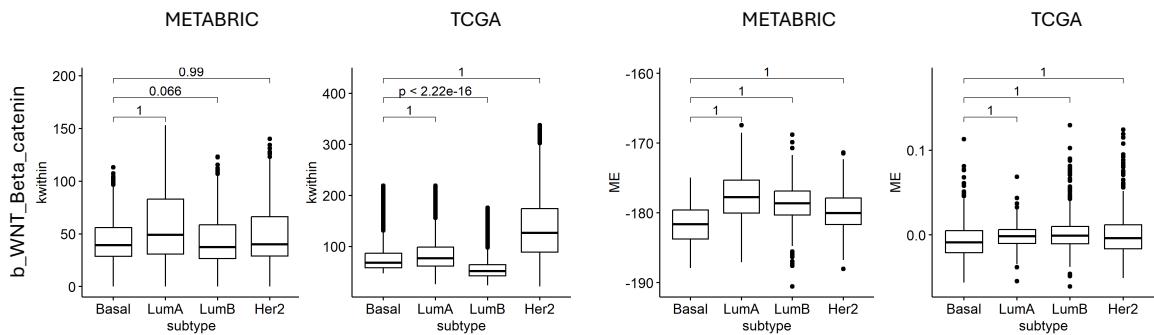
## Expression



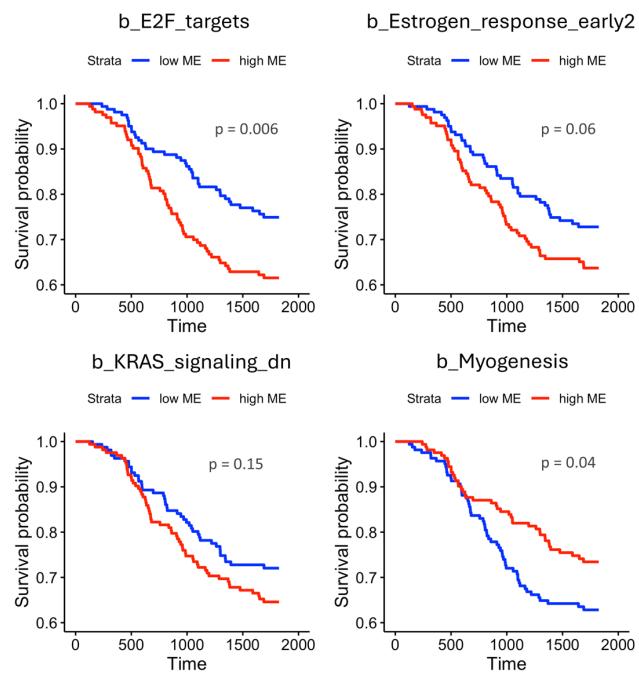
E

## Connectivity

## Expression

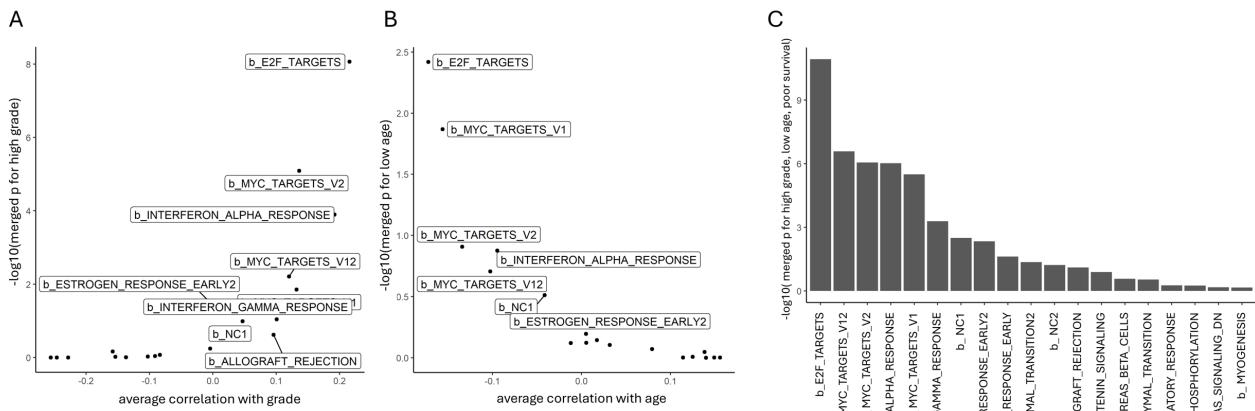


**Supplementary Figure 7. Connectivity and expression of basal-like modules across subtypes.** For each module (rows), intramodular connectivity (kWithin) and Module Eigengene (ME) were computed in both the METABRIC and TCGA databases across subtypes. P-values were obtained with a Wilcoxon rank-sum test. **(A)** shows the four modules displaying significantly higher connectivity/expression in the basal-like subtype. **(B,C,D,E)** all other modules.

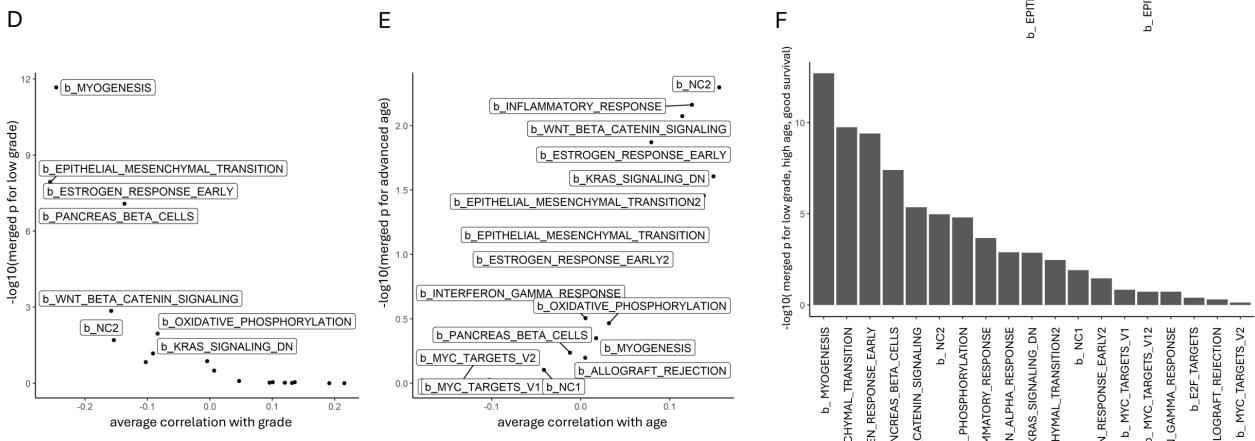


**Supplementary Figure 8. Kaplan-Meier survival curves for basal-like patients stratified by module expression.** Patients were grouped based on the module eigengene (ME) values of modules that show higher intramodular connectivity in basal-like tumours compared to other subtypes.

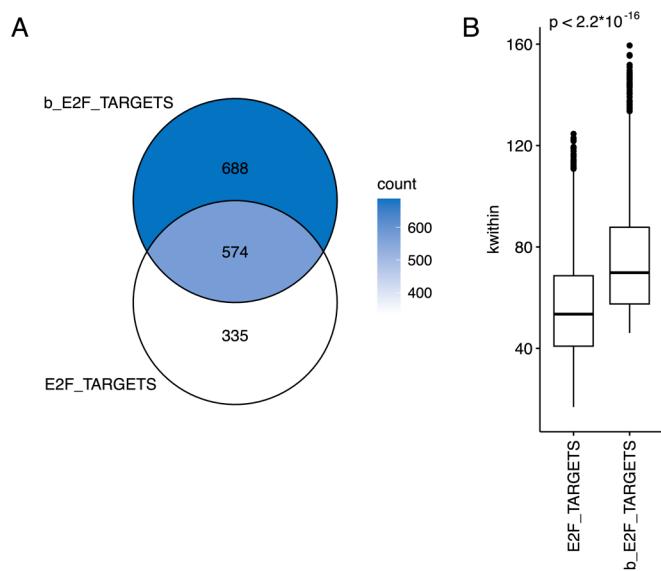
### Correlation with high aggressiveness



### Correlation with low aggressiveness



**Supplementary Figure 9. Summary of basal modules' relationship with clinical features.** **(A-C)** Modules' correlation with features of high aggressiveness: high grade **(A)**, early onset **(B)**, merged score for high grade, early onset and poor prognosis **(C)**. **(D-F)** Modules' correlation with features of low aggressiveness: low grade **(D)**, late onset **(E)**, merged score for low grade, late onset and good prognosis **(F)**. **(A,B,D,E)** For each clinical feature, p-values were obtained by merging the p-values across all the individual tested datasets with the Fisher's method, considering the direction of the relationship between module's expression and the clinical feature. **(C, F)** for each module, the overall significance for the relationship with grade, age at diagnosis and survival was obtained by multiplying the individual dataset-merged p-values. Only basal-like samples were employed in this analysis.



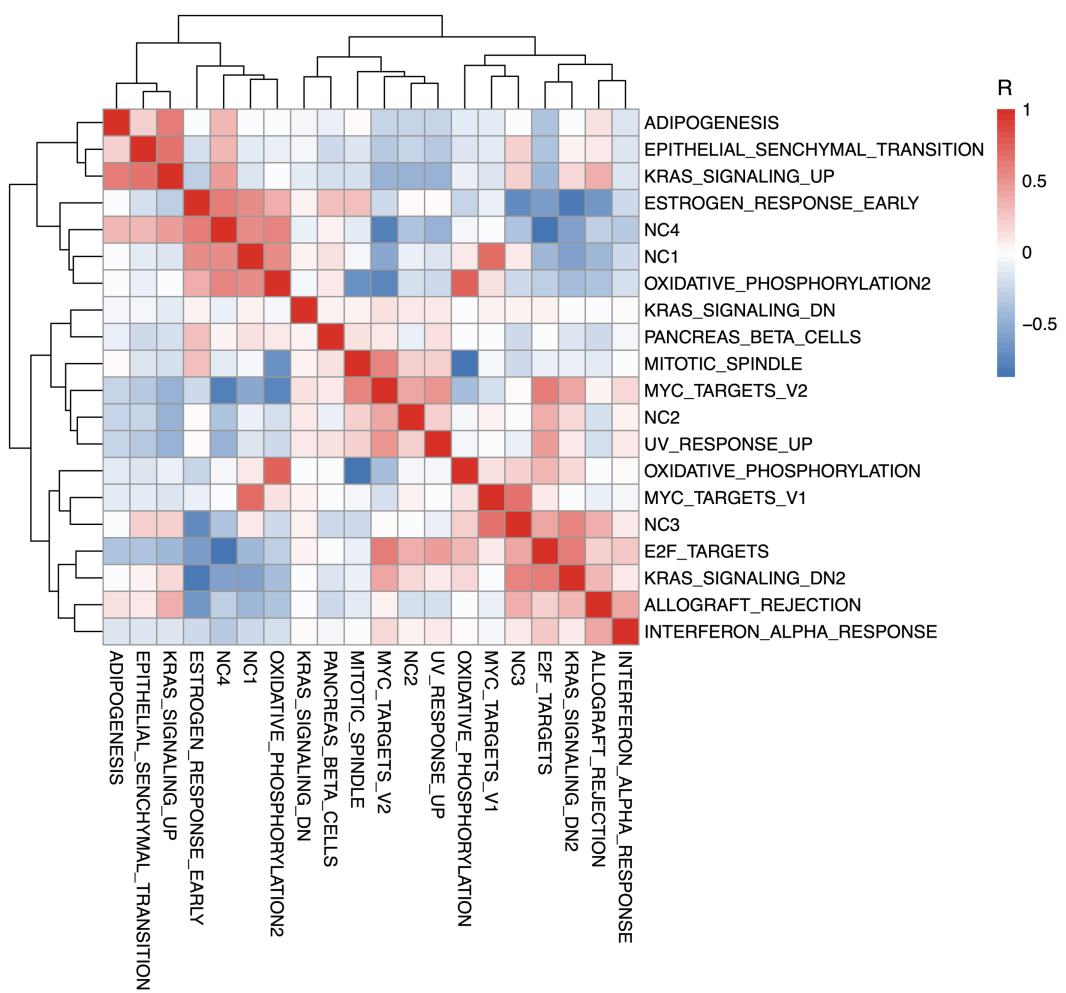
**Supplementary Figure 10. Comparison between global and basal-like-specific *E2F\_targets* modules.** **(A)** Venn diagram showing the overlap between genes in the *E2F\_targets* module (defined using all breast cancer samples) and the *b\_E2F\_targets* module (defined using only basal-like samples). **(B)** Boxplot comparing the intramodular connectivity (kWithin) of genes in each module, computed in basal-like samples. Statistical significance was assessed using a two-sided Wilcoxon test.

**Highly connected b\_E2F\_targets transcription factors significantly regulate the expression of module's genes.**

**Supplementary Table 1.**

*b\_E2F\_targets* genes belonging to the GO:0003700 - DNA-binding transcription factor activity - Gene Ontology term, ranked by *kWithin*

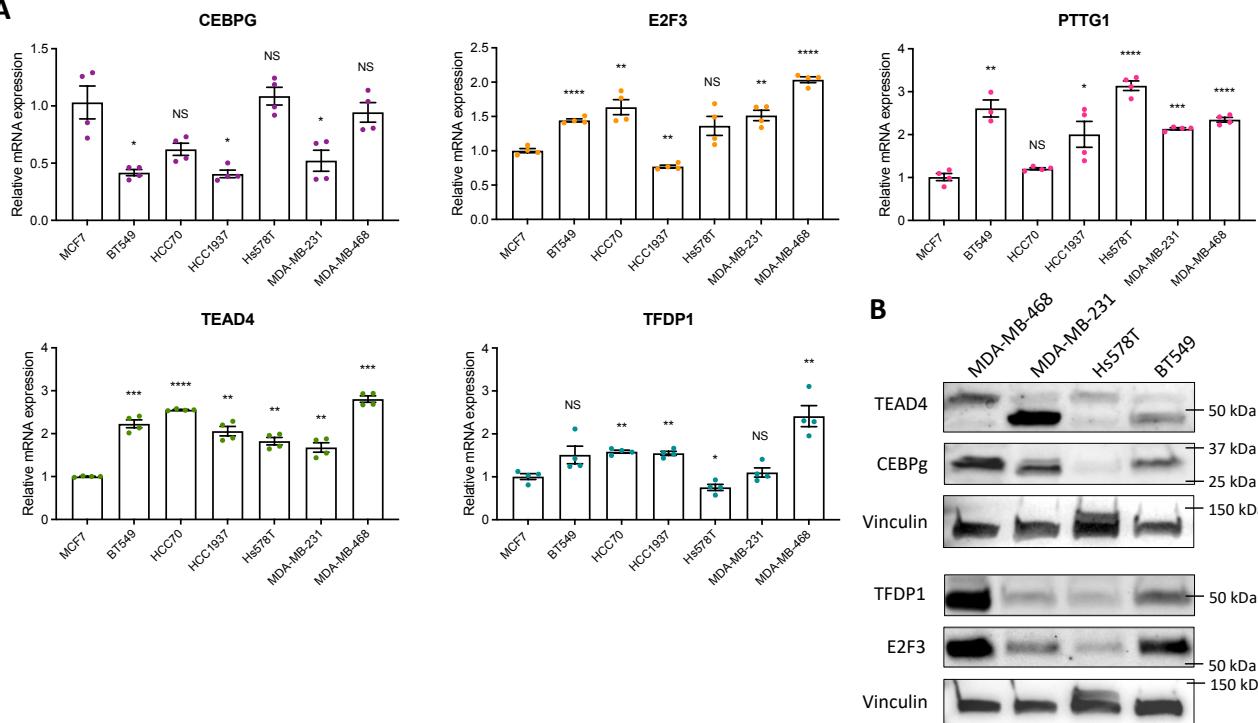
TF	kWithin	rank	dataset
FOXM1	151,849192	4	GSE2222, GSE55204, GSE25741
PTTG1	113,661926	97	GSE48928
TEAD4	111,890624	104	
EZH2	111,377135	106	GSE36939, GSE48979, GSE103242
E2F3	105,938864	137	
FOXC1	104,640658	146	GSE73234
PARP1	101,120351	161	GSE34817
TFDP1	96,5986486	200	
CEBPG	96,2457574	202	
CHCHD3	95,9415737	208	
NOLC1	93,6657614	241	
HMGA1	91,4973555	265	GSE45483, GSE35525
UHRF1	90,0384401	286	
TAF5	90,0092093	287	
SOX9	89,7260918	289	
BOLA1	88,419527	305	
TCF7L1	88,3648149	306	
SSRP1	88,2140726	310	GSE92281
ZNF232	87,4847614	320	
ZNF165	86,9895521	326	GSE63984
ELF5	86,6682148	330	GSE30405
RCOR2	84,5795255	357	
NR2C2	82,2037687	390	
ZBTB5	81,7372596	398	
DNMT1	81,3654839	404	



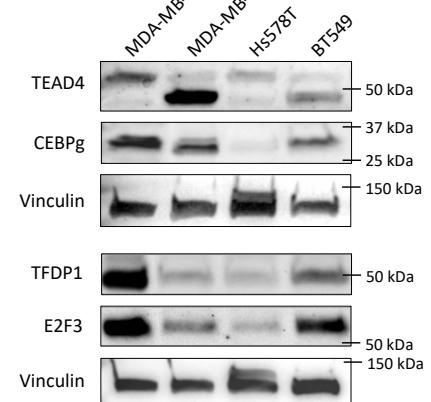
**Supplementary Figure 11. Correlations between basal-like modules.** Heatmap showing Pearson's correlation between module eigengenes, computed across basal-like patients. Hierarchical clustering was based on their eigengene correlation.

## Silencing of five TF hubs significantly affects proliferation of BLBC cell lines.

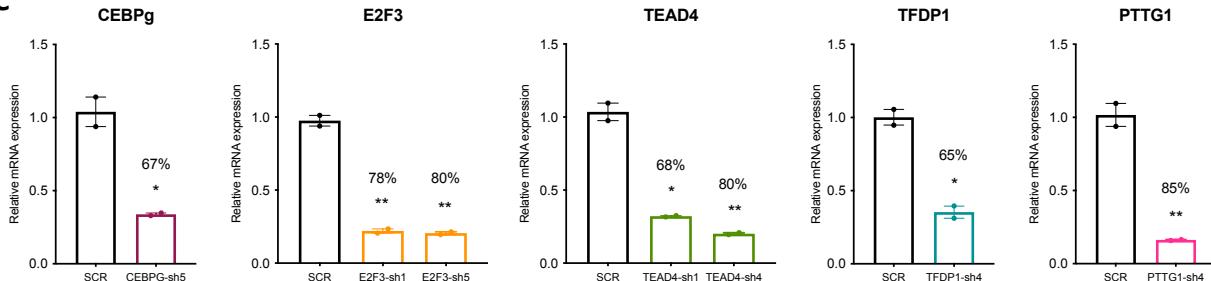
**A**



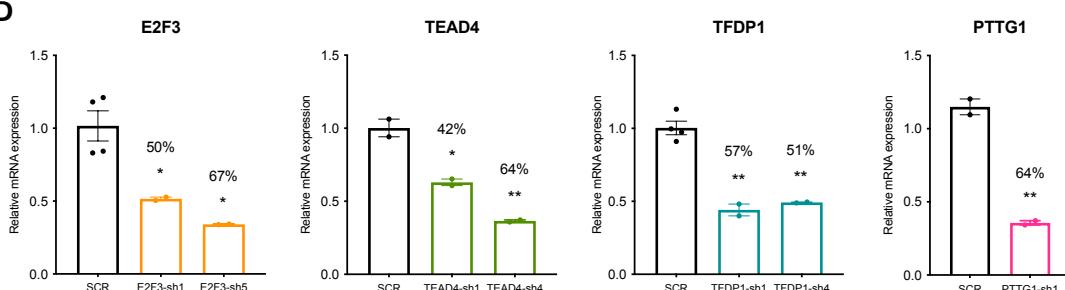
**B**



**C**

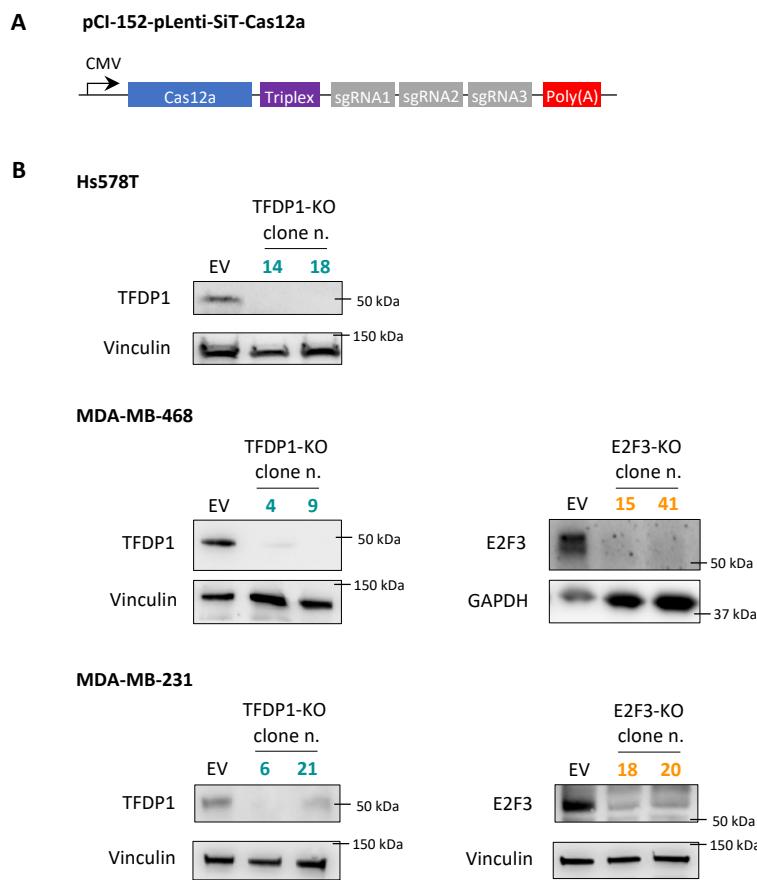


**D**

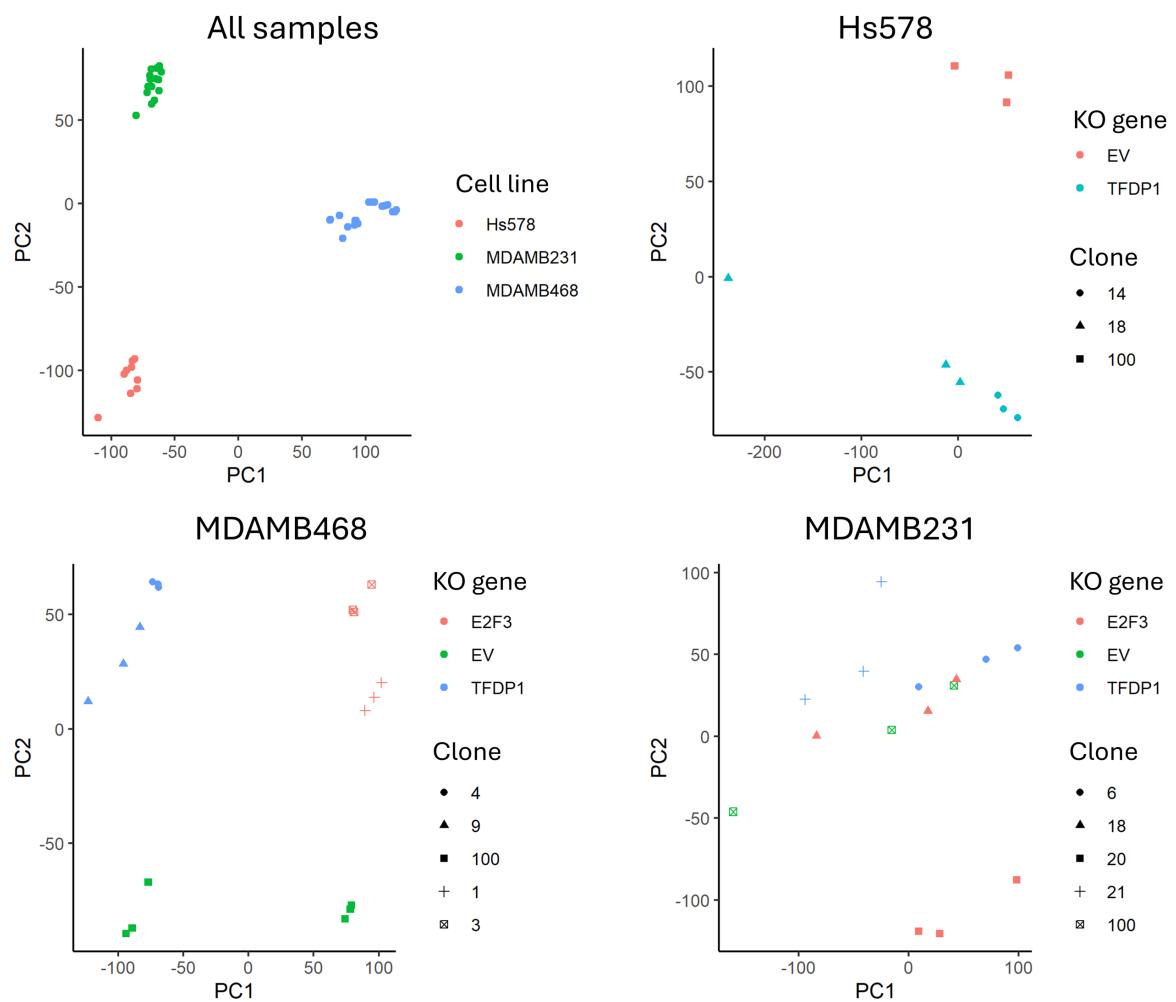


**Supplementary Figure 12. Expression and silencing of *b\_E2F*\_targets TF hubs.** The expression of the indicated genes was measured in the indicated cell lines by RT-qPCR (A), shown as mean  $\pm$  S.E.M relative to the value of MCF7 cells upon normalization to Actin B (n=4 independent experiments), one way ANOVA. \*, p < 0.05; \*\*, p < 0.01, \*\*\*, p < 0.001, \*\*\*\*, p < 0.0001, or by Western blot on whole cell extracts (n=2 independent experiments) (B). TF hubs were silenced by lentiviral-mediated shRNA expression in MDA-MB-468 (C) and -231 (D) cells. Silencing was measured by qRT-PCR and is shown as mean  $\pm$  S.E.M relative to the scrambled control (SCR). Only the best performing shRNAs are shown. (n=4 independent experiments). Unpaired t-test, \*, p < 0.05; \*\*, p < 0.01.

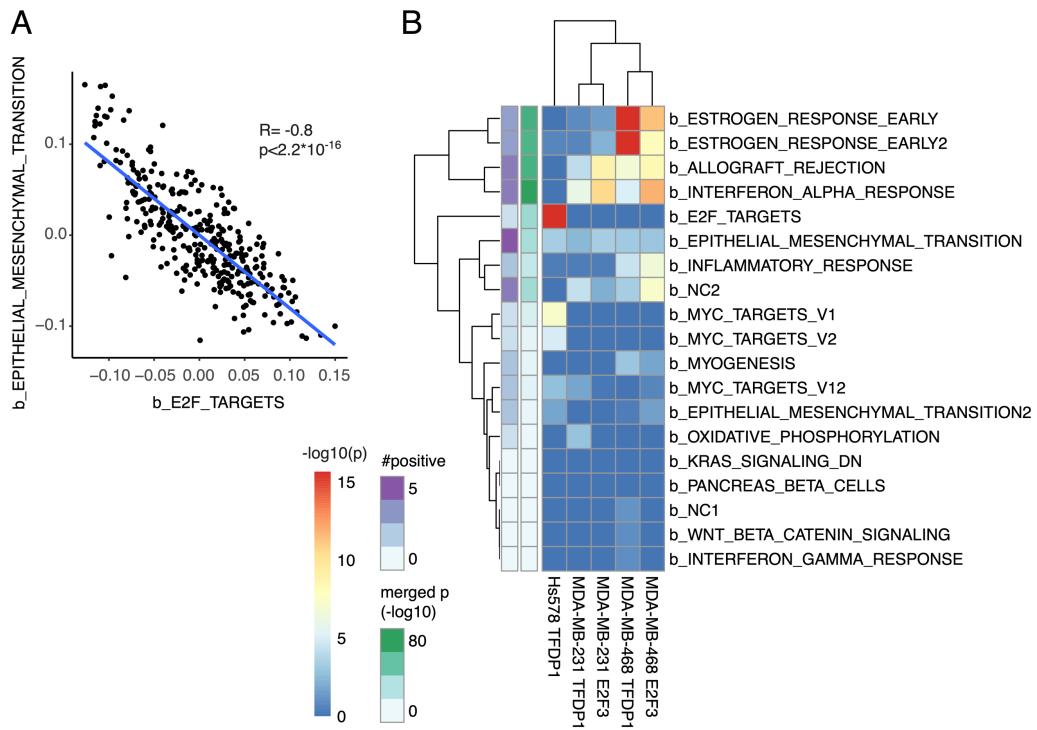
**Phenotypic effects of TF hub genes knockout correlate with transcriptional module disruption.**



**Supplementary Figure 13. Generation of Hs578T, MDA-MB-231 and MDA-MB-468 cells KO for E2F3 and TFDP1. (A)** Schematic representation of vectors encoding for Cas12a (AsCfp1) and carrying a CRISPR array with 3 sgRNAs directed against the target genes. **(B)** Residual protein expression was assessed by Western blot analysis of different clones derived from the indicated cell lines, with the indicated antibodies.



**Supplementary Figure 14. Principal Component Analysis (PCA) of RNA-seq data.** Each point represents a single sample. In the first plot, colors indicate the cell line. In the other plots, samples from each cell line are shown separately, and colors indicate the specific gene knockout or the empty vector control (EV). Clones are indicated by different shapes. Note that MDA-MB-468 E2F3 KO clones and their empty vector were sequenced in a different facility, explaining the PC1 separation from the other samples.



**Supplementary Figure 15. (A)** Scatter plot of *b\_E2F\_targets* and *b\_Epithelial\_mesenchymal\_transition* modules' eigengenes computed from METABRIC basal-like samples. Each point represents a patient, with the x- and y-axes showing the module eigengene of the *b\_E2F\_targets* and *b\_Epithelial\_Mesenchymal\_Transition* modules, respectively. **(B)** Enrichment significance of up-regulated genes upon TF hubs' KO in basal-like BC lines, in-house data.

**Suppl. Table 2. Comparison of all the TF hubs in the E2F\_targets and b\_E2F\_targets modules.**

The top 25% TF hubs in each module are shown in bold. Asterisks indicate genes completely absent from the general module.

Note that 7 of the validated genes (FOXC1, PARP1, SSRP, ELF5, TEAD4, E2F3, TFPD1) would have been missed without analysing subtype-specific networks.

**E2F\_targets**

Gene	kTotal	kWithin	rank
<b>FOXM1</b>	545,8059478	157,2385805	13
<b>PTTG1</b>	518,1612739	146,7355187	27
<b>E2F2</b>	518,25252	131,6658127	53
<b>E2H2</b>	469,5950475	125,0671861	64
<b>UHRF1</b>	447,6648191	117,7752092	82
<b>HMGAI</b>	471,904967	93,27173723	165
<b>E2F7</b>	417,0809894	88,19651424	195
<b>DNMT1</b>	399,0062414	75,66846466	294
<b>NOLC1</b>	398,2846515	75,06320196	301
<b>CHCHD3</b>	401,8862114	71,3488528	337
<b>RCOR2</b>	421,053976	70,59843607	343
<b>CEBPG</b>	427,9626136	66,98986737	385

**b\_E2F\_targets**

Gene	kTotal	kWithin	rank
<b>FOXM1</b>	517,5179103	151,8491924	4
<b>PTTG1</b>	446,7140888	113,6619256	97
<b>TEAD4</b>	464,990407	111,8906237	104
<b>EZH2</b>	432,6551732	111,3771352	106
<b>E2F*</b>	473,9321194	105,9388642	137
<b>FOXC1*</b>	440,7405836	104,6406577	146
<b>PARP1*</b>	479,4244058	101,1203506	161
<b>TFDP1</b>	412,1732918	96,59864862	200
<b>CEBPG</b>	448,8300016	96,24575739	202
<b>CHCHD3</b>	426,7689524	95,94157373	208
<b>NOLC1</b>	422,1877497	93,6657614	241
<b>HMGAI</b>	433,6303252	91,49735551	265
<b>UHRF1</b>	404,7782849	90,03844009	286
<b>TAF5</b>	416,5724709	90,00920929	287
<b>SOX9</b>	421,4073231	89,72609181	289
<b>BOLA1</b>	422,449619	88,41952699	305
<b>TCF7L1</b>	414,9359793	88,36481492	306
<b>SSRP1</b>	460,6987668	88,21407263	310
<b>ZNF232</b>	436,6607204	87,48476136	320
<b>ZNF165</b>	427,9250669	86,9895521	326
<b>ELF5*</b>	393,182212	86,66821477	330
<b>RCOR2</b>	429,3935035	84,57952549	357
<b>NR2C2</b>	399,5668967	82,20376874	390
<b>ZBTB5</b>	391,6089104	81,73725958	398
<b>DNMT1</b>	394,6842731	81,36548393	404

Gene	kTotal	kWithin	rank
<b>E2F7</b>	400,8491659	80,10593558	422
<b>ZSCAN20</b>	427,7471509	79,86075991	425
<b>PHF5A</b>	395,2935163	79,29558779	433
<b>YEATS2</b>	406,0701829	78,21482604	461
<b>SOX10</b>	389,7648041	77,9770118	467
<b>ZNF239</b>	403,9111869	77,78196325	469
<b>FUBP1</b>	396,8369653	77,70745143	473
<b>SMARCC1</b>	397,8883862	77,5632953	474
<b>ZNF384</b>	405,9870069	76,93547543	487
<b>WDHD1</b>	385,5216104	76,75213613	494
<b>ZNF670</b>	382,3450882	75,2118989	519
<b>NFYA</b>	382,4027691	75,06933825	520
<b>TGIF2</b>	417,1442289	74,44344408	532
<b>TFCP2L1</b>	394,6230583	74,05010485	541
<b>YBX3</b>	384,5219229	72,35824483	576
<b>VGLL1</b>	387,5594175	72,34639726	578
<b>MAZ</b>	401,513753	72,28084519	580
<b>EN1</b>	399,3617986	71,75996199	588
<b>YBX1</b>	382,3245053	71,23871339	594
<b>TMSB15B</b>	380,9568685	70,92572076	606
<b>OTX1</b>	393,3594442	70,89675465	607
<b>SOX4</b>	401,6418641	69,51332771	644
<b>ZBED4</b>	376,3568408	69,22634404	653
<b>GATA2D2</b>	409,5072559	69,21675457	654
<b>FOXP4</b>	402,3270348	68,90484656	662
<b>NFE2L3</b>	399,5758507	67,69332685	683
<b>TFDP2</b>	382,6879624	67,29559159	697
<b>ZFP82</b>	400,0982005	67,28340886	698
<b>NFXL1</b>	373,4736797	66,88047719	703
<b>TBX19</b>	389,1013931	66,78783836	705
<b>C9orf40</b>	366,3663232	66,11517465	732
<b>AEBP2</b>	380,2555544	65,89111461	740
<b>CEBPB</b>	403,5518432	65,73412403	742
<b>HLTF</b>	377,9463084	65,17978805	755
<b>ZNF286A</b>	378,4517372	64,75248909	766
<b>E2F8</b>	354,9233308	64,68845453	768
<b>ADNP2</b>	373,5173621	64,42152296	773
<b>NONO</b>	387,8471739	63,97066484	781
<b>HCFC1</b>	371,2887376	63,88609397	785
<b>ETV3</b>	368,2194684	62,83174142	811
<b>MLLT10</b>	363,2272514	62,70973976	817
<b>BUD31</b>	411,0265793	61,54055655	850
<b>NKX2-5</b>	361,3111176	61,24424751	855
<b>TCF19</b>	381,1940255	61,19142204	857
<b>NFIB</b>	378,5458887	61,07810058	859
<b>ZNF280C</b>	364,8530416	60,89722564	863
<b>ETV4</b>	380,0937957	59,76683398	895
<b>PLAGL2</b>	349,1219779	59,46128338	901
<b>LMO4</b>	405,0865999	59,06760596	913
<b>TEAD2</b>	391,1747808	58,70543135	920
<b>HR</b>	373,5257837	58,55145997	922
<b>BARX2</b>	355,5920909	57,94898961	942
<b>TAF4</b>	377,8939906	57,33074599	953
<b>STAG1</b>	366,1406773	57,29681045	954
<b>MYNN</b>	367,215472	57,21276879	958
<b>ALX1</b>	362,9987381	56,62261547	977
<b>SCML2</b>	376,7797932	56,48103523	980
<b>SRF</b>	380,4723943	56,23521831	987
<b>ENO1</b>	422,2986428	56,12750106	990
<b>ZNF576</b>	370,1216093	53,1790406	1060
<b>HNRNPAB</b>	365,8886302	52,9682541	1065
<b>ZNF121</b>	353,8446039	52,74373801	1071
<b>NOCT</b>	392,7677661	52,34512882	1080
<b>BMPR1A</b>	353,8991549	52,10447718	1086
<b>POU4F1</b>	352,3619428	51,23862725	1118
<b>LRRKIP2</b>	374,0518174	50,95025584	1124
<b>DMRT1</b>	356,3936143	50,8452056	1127
<b>FOXK2</b>	353,0962496	50,39816632	1142
<b>ZNF519</b>	363,9779497	50,31956502	1147
<b>RLF</b>	359,5718594	50,31559673	1148
<b>KIAA1549</b>	369,3737717	50,07255753	1158
<b>ZNF280B</b>	362,9154775	49,7708106	1169
<b>PPARA</b>	357,3288836	49,46446769	1177
<b>ZNF285</b>	366,2030602	49,44221832	1178
<b>ZSCAN16</b>	371,1125928	48,99338275	1195
<b>TET1</b>	361,6461064	48,38719262	1216
<b>ZNF200</b>	351,8604136	47,61080732	1240

## Supplementary Material and Methods

**Supplementary Table 3.** shRNA, sgRNA, qRT-PCR assay sequences and IDs.

shRNA	clone ID	sequence
CEBPG-sh1	TRCN000003836	CCGGTGGCGACAATGCAGGACAGTACTCGAGTACTGTCCTGCATTGCGCCATTTT
CEBPG-sh2	TRCN000003837	CCGGGCAACGCCAGAGAGGAACAACCTCGAGTTCTCTCGCGTTGCTTTTT
CEBPG-sh3	TRCN000003838	CCGGCCTACCCCTTCAGACTTACTCGAGTAAGCTGGAAAGGGTGAGGTTTT
CEBPG-sh4	TRCN000003839	CCGGCTGACCAAGGAATTAAGTGTACTCGAGTACACTTAATTCTGGTCAGTTTT
CEBPG-sh5	TRCN0000010821	CCGGGATTTGTTCTGAGCATGCACTCGAGTGCATGCTCAAGAAACAAATCTTTT
E2F3-sh1	TRCN0000013803	CCGGCCGCTTACTTCAGGAATCGAGATTCTGAAGAGTAAAGCGGGTTTT
E2F3-sh2	TRCN0000013804	CCGGCCTATTAAAGAAGAAGTCTAAGTGTAGACTCTCTTAATGAGGTTTT
E2F3-sh3	TRCN0000013805	CCGGCCAAACTGTTAGTTGAACTCGAGTTACAACATAACAGTTGGTTTT
E2F3-sh4	TRCN0000013806	CCGGCCTACTCAATAGAGAGCCTACTCGAGTAGGCTCTATTGAGTCAGGTTTT
E2F3-sh5	TRCN0000013807	CCGGCCAACTCAGGACATAGCGATTCTCGAGAACGCTATGCTGAGTTGGTTTT
TEAD4-sh1	TRCN0000015873	CCGGGAAGAGACGTGTGAGCAGGAACTCGAGTTCTGCACACACGTCTCTTTT
TEAD4-sh2	TRCN0000015874	CCGGCCGGATATTGAGCAGAGTTCTCGAGAAACTCTGCTCAATATCCGGGTTTT
TEAD4-sh3	TRCN0000015875	CCGGGAGACAGATATGCTCGATCTCGAGATAGCAGCATACTCTGCTCTTTT
TEAD4-sh4	TRCN0000015876	CCGGCCTTCTCAGAACCTATCGAGATAGGTTGCTGAGAGAAAGGTTTT
TEAD4-sh5	TRCN0000015877	CCGGGCTGTGCATTGCCATGCTTCTCGAGAACGACATAGGCAATGCGACAGCTTTT
PTTG1_sh1	TRCN0000015103	CCGGCCTCAATCAAAGCCTAGATCTCGAGATCAAGGCTTGATTGAAGGTTTT
PTTG1_sh2	TRCN0000015104	CCGGCCTAACCTAAAGCTACTAGAACCTCGAGTTCTAGTAGCTTAGGTAAGGTTTT
PTTG1_sh3	TRCN0000015105	CCGGCCAATCTGTCAGCTCCTCTCGAGAACGGAGACTGCAACAGATTGGTTTT
PTTG1_sh4	TRCN0000015106	CCGGCCCTCAATCCTAGACTTCTCGAGAACAGTCTAGAGGATTGAAGGGTTTT
PTTG1_sh5	TRCN0000015107	CCGGGTGGTGTCAAGGATGGGCTCTCGAGAGGCCATCCTAGAACACCACATTTT
TFDP1-sh1	TRCN0000019880	CCGGCCTACGGCATTCTCCATGAACCTCGAGTTCATGGAGAAATGCCGTAGGTTTT
TFDP1-sh2	TRCN0000019881	CCGGGCTTCATAGACCGAACCTCTCGAGAACGGGTTGGTATGAAGACTTTT
TFDP1-sh3	TRCN0000019882	CCGGCGACAACCACATCTTACCAAACCTCGAGTTGTAAGATGTGTTGCGTTTT
TFDP1-sh4	TRCN0000019883	CCGGGACGATGACTCAACGAGAACCTCGAGATTCTCGTTGAAGTCATCGTCTTTT
TFDP1-sh5	TRCN0000019879	CCGGCCGTTATTGCCGTAGTTCTCGAGAACACTACGGCAAATAAGCGGTTTT
SCR		CCGGCCTAAGTTAACGTCGCCCTCGAGCGAGGGCAGTTAACCTAGGTTTT

sgRNA		sequence
hsE2F3		AGATAAATCTGATATGTAACATAAAATTCTACTCTGAGATTGATATCACCAACGTTCTGGAAAGAATTCTACTCTGAGATGACTATCTGGACTTCGTAGTGCA
hsTFDP1		AGATAAGGAAGCGAACAGGAAAGGAGAACCTACTCTGAGATAAGGTGAAAGACAGAGGAGACTAATTCTACTCTGAGATGCAAATTGCCCTCAAGAACCTGG

gene	cat. n.	qRT-PCR assay ID
CEBPG	4331182	Hs00156454_m1
E2F3	4331182	Hs00605457_m1
FoxM1	4453320	Hs01073586_m1
PTTG1	4331182	Hs00851754_u1
TEAD4	4331182	Hs01125032_m1
TFDP1	4331182	Hs00955491_gH
TFDP1	4331182	Hs00955488_g1
GAPDH	4331182	Hs99999905_m1
B-act	4448484	Hs01060665_g1