

Supporting Information for

pH-sensitive Amino Lipid-Driven Pore Formation Enables Endosomal Escape of Lipid Nanoparticles

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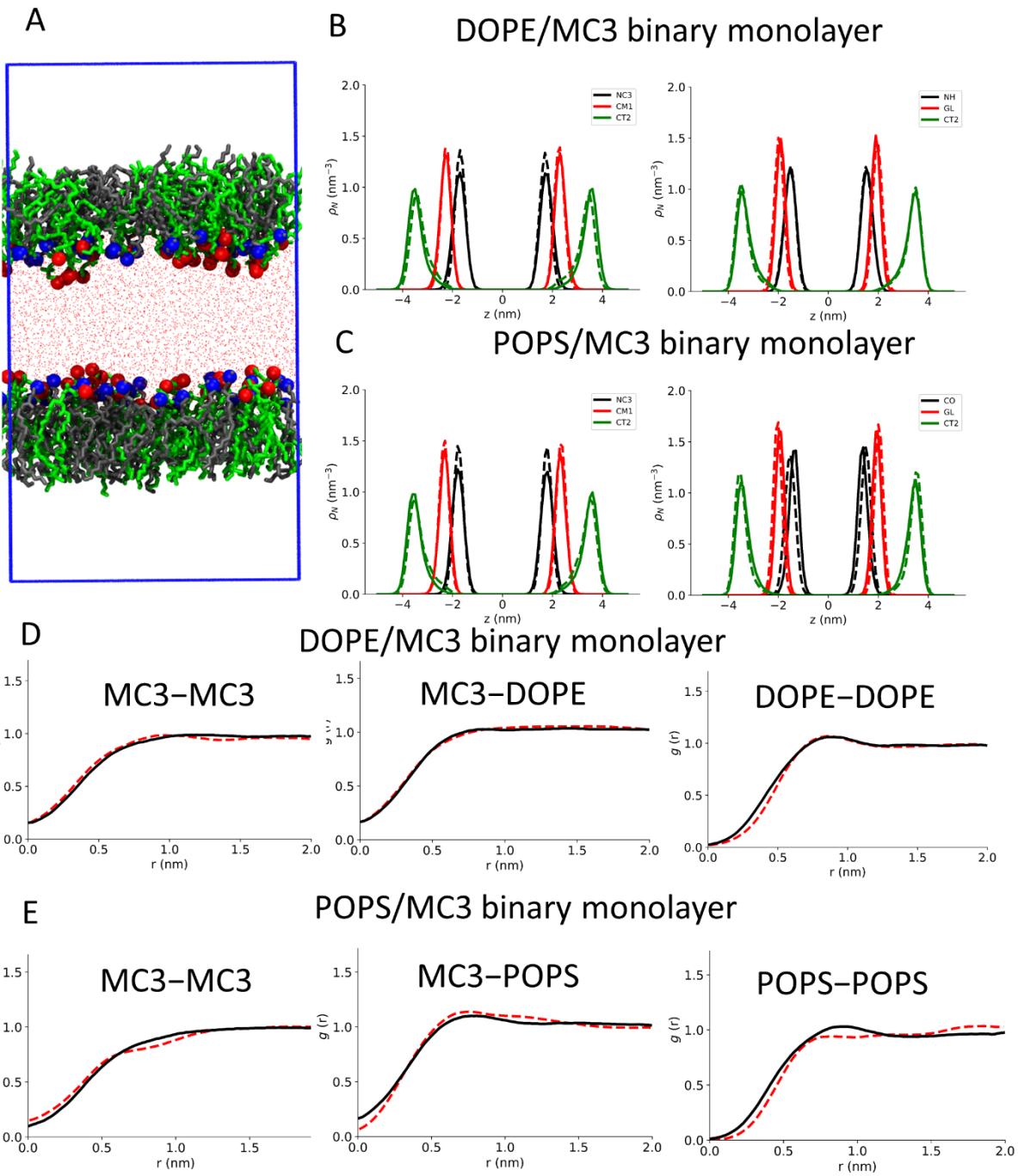


Figure S1. (A) Snapshot of the all-atom (AA) simulation box for the cationic MC3 lipid/DOPE monolayer. Color scheme: grey for MC3, green for DOPE, blue spheres for head-group sites of MC3, red spheres for head-group sites of DOPE, and red dots represent water oxygen. (B, C) Number density profiles of selected CG segments along the monolayer normal of the (B) cationic MC3/DOPE and (C) MC3/POPS binary systems. Solid and dashed lines represent CG- and AA-MD results, respectively. (D, E) Two-dimensional radial distribution functions (2D-RDFs) between lipids in the (D) MC3/DOPE and (E) MC3/POPS binary monolayers, confirming the accuracy of the CG parameters. The red dotted and black solid lines represent the 2D-RDFs obtained from AA- and CG-MD, respectively.

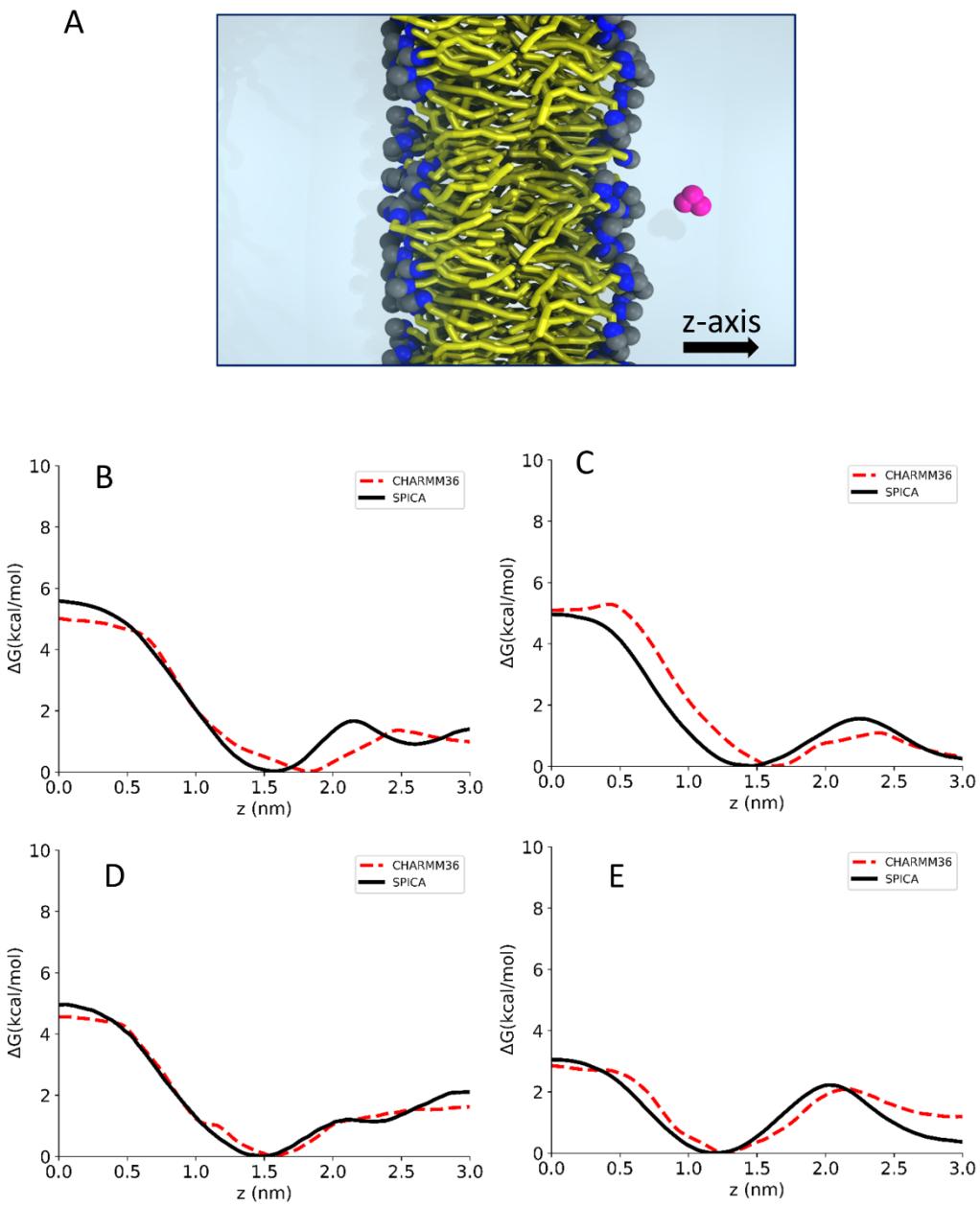


Figure S2. (A) Representative snapshot of the CG-MD simulation performed to obtain the partition-free energy of nucleic acid analogs from an POPS lipid monolayer. Color scheme: yellow for POPC tails, blue and gray spheres for head-group sites of POPS, ice blue for water phase and magenta for thymine. (B, D) Free energy profiles of thymine as a function of distance from the center of the (B) POPS and (D) POPI bilayer along the bilayer normal (z -axis). (C, E) Free energy profiles of deoxyribose as a function of distance from the center of the (C) POPS and (E) POPI bilayer along the bilayer normal (z -axis). Color scheme: red dashed lines represent AA-MD results using the CHARMM36 force field (FF), and black solid lines represent CG-MD results using the optimized SPICA FF.

System II

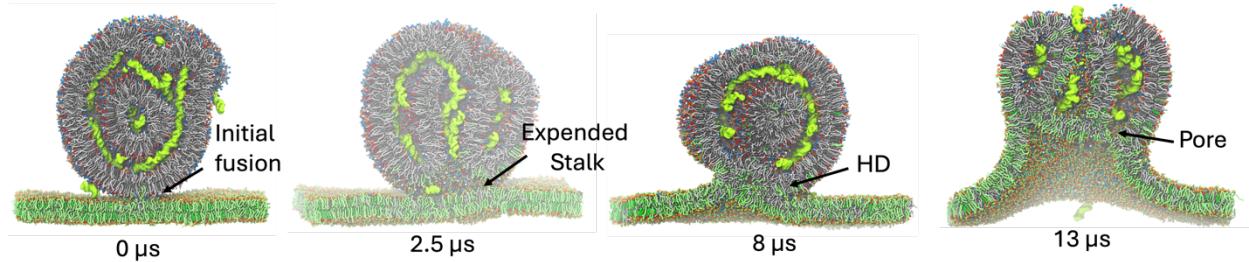
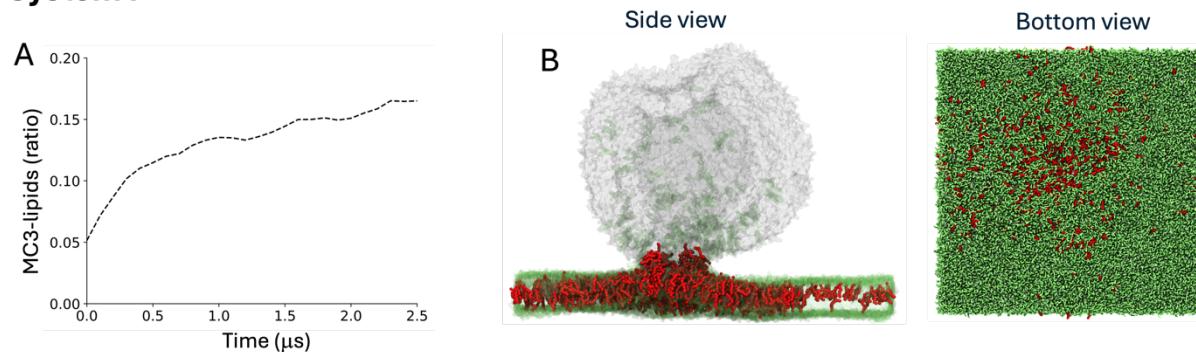


Figure S3. Representative snapshots of *System II* illustrate the time evolution of the LNP-EM system during molecular dynamics (MD) simulations. These snapshots demonstrate that the radial expansion of the stalk and the subsequent formation of the hemifusion diaphragm (HD) lead to the initiation of a small fusion pore, similar to what is observed in System I. Color scheme: Different lipid types are shown in distinct colors. Lipid headgroups, tails, and dsDNA molecules are depicted using van der Waals (VDW), Licorice, and QuickSurf (an iso-surface) representation, respectively. For clarity, water and ions are omitted.

System I



System II

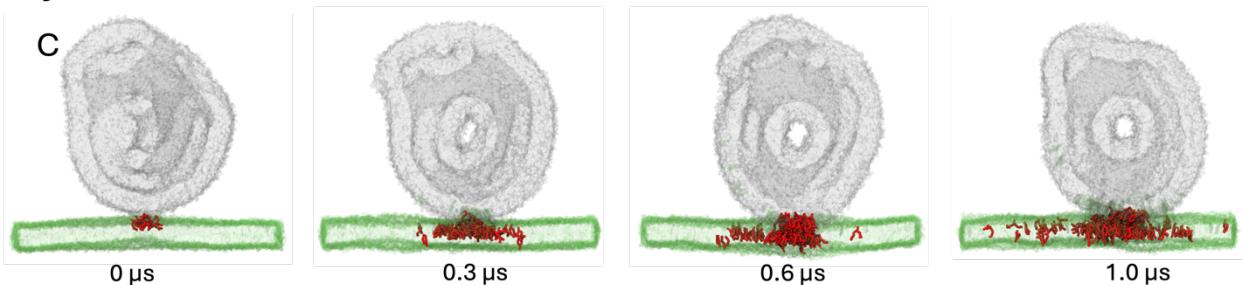


Figure S4. MC3-lipids exhibit rapid migration in the endosomal membrane (EM). (A) Time evolution of the MC3-lipid ratio in EM. The quantitative MC3 lipid ratio was calculated relative to its total number. (B) The cross-sectional view and bottom view of the representative MD simulation snapshot of the LNP-EM complex at $t \sim 0.9 \mu\text{s}$. Color scheme: red for MC3-lipids, gray and green represent the LNP and EM domains, respectively. (C) The time evaluation of the LNP-EM complex simulation in *system II* also highlighted MC3-lipids migration in EM. The color scheme is identical to that of Figure S4B.

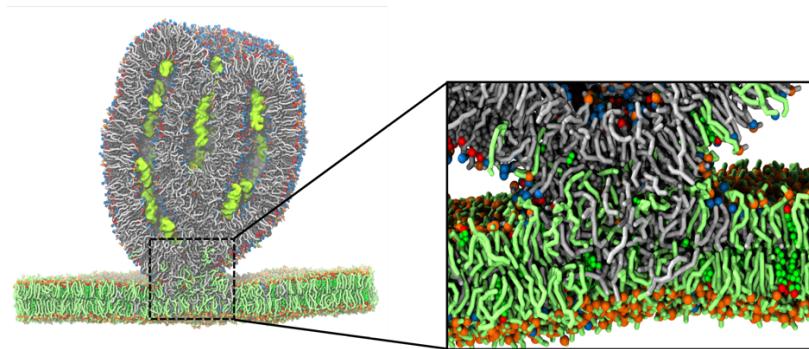


Figure S5. A disrupted stalk compared to conventional lipid bilayer/vesicles and membrane fusion stalk.¹ The cross-sectional view of representative MD simulation snapshot of the LNP-EM complex after fusion at $t \sim 0.25 \mu\text{s}$. Color scheme: Different lipid types are shown in distinct colors. Lipid headgroups, tails, and dsDNA molecules are depicted using van der Waals (VDW), Licorice, and QuickSurf (an iso-surface) representation, respectively. For clarity, water and ions are omitted. Inset: zoom of the fusion region for comparison with previously reported bilayer/vesicle and membrane fusion.¹

¹Kawamoto S., Klein, M. L., & Shinoda W. Coarse-grained molecular dynamics study of membrane fusion: Curvature effects on free energy barriers along the stalk mechanism, *J. Chem. Phys.* **143**, 243112 (2015)

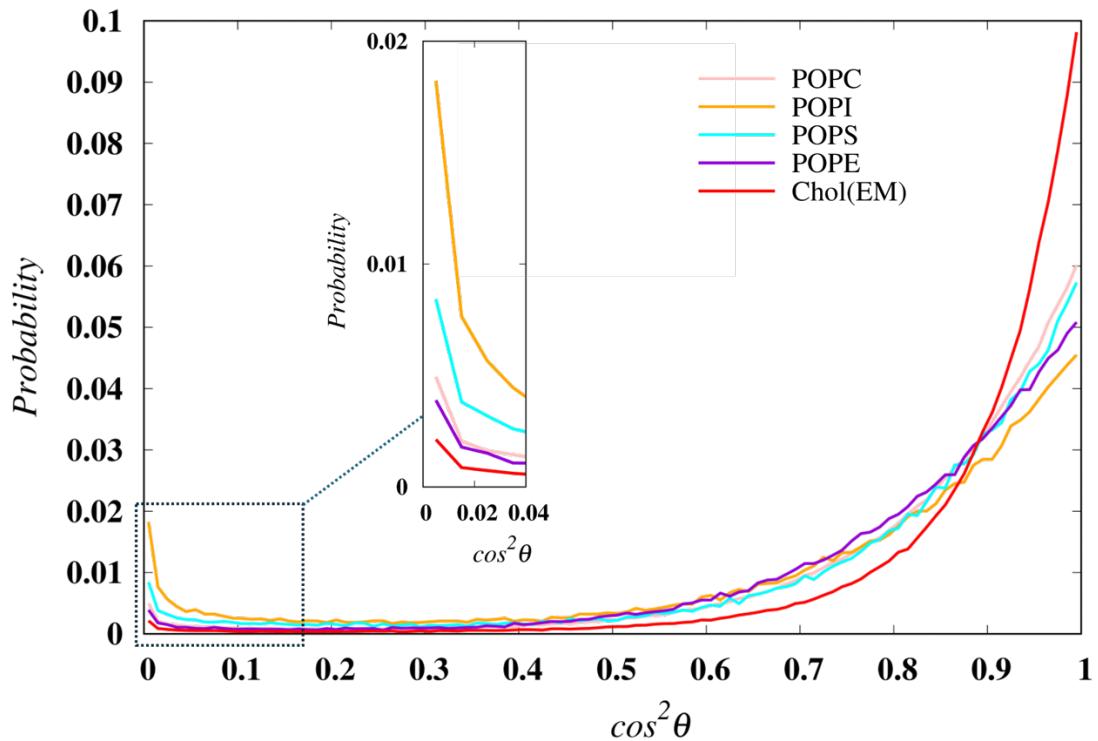


Figure S6. The orientation of lipids in relation to the membrane's normal direction emphasizes their tendency to migrate toward LNPs. Probability distribution of the angle, $\cos^2 \theta$, between the lipid head-to-tail bond vector and the membrane normal. Inset: Distribution of $\cos^2 \theta$ in the range $0 \leq \cos^2 \theta \leq 0.04$.

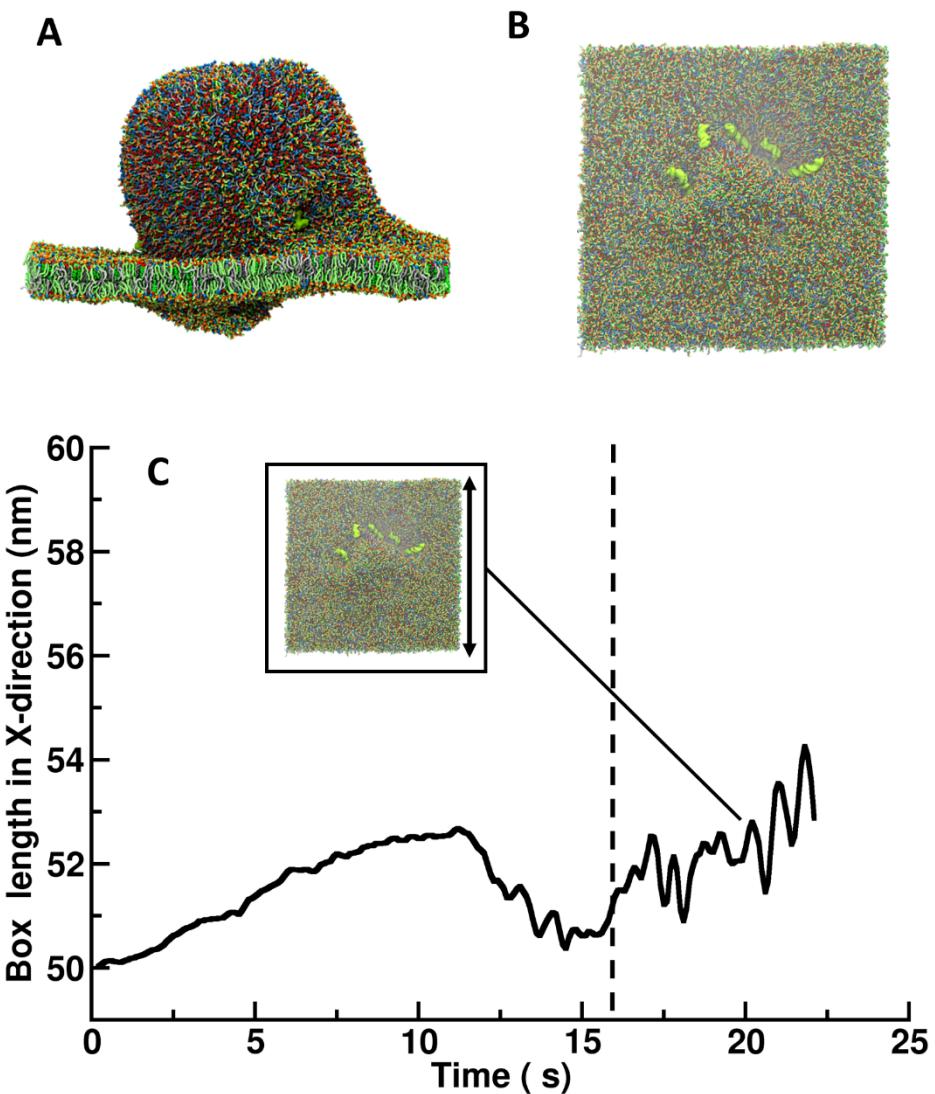


Figure S7. Representative snapshots of the EM-LNP fused complex containing all MC3 lipids in protonated state at 18.4 μ s: (A) side view and (B) top view. (C) Time evolution of the box length in the X-direction during the MD simulations.

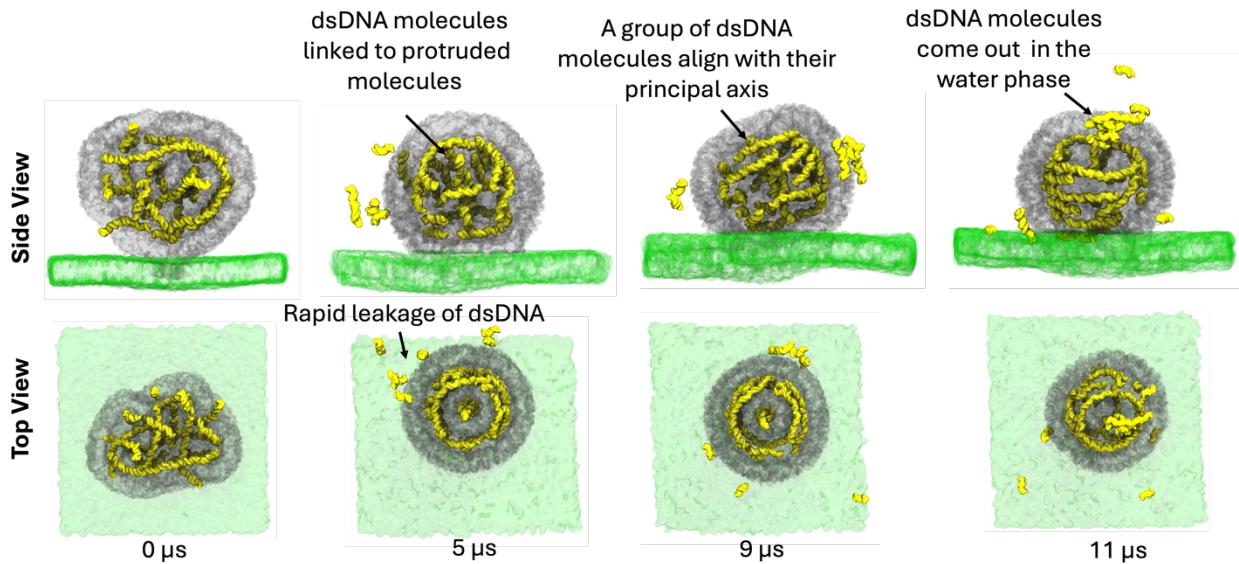


Table S1. Comparison of surface tension (γ , in mN/m) for binary monolayer systems between AA- and CG-MD simulations.

Monolayer system*	γ (AA MD)	γ (CG MD)
cationic MC3/DOPE (1:1)	46 ± 1	44 ± 1
cationic MC3/POPS (1:1)	52 ± 1	50 ± 1
cationic MC3/POPI (1:1)	58 ± 1	55 ± 1

Table S2. Molecular composition of the LNP-EM complex systems simulated in this study.

LNP	pH ~ 4	System I [#]	Molecule	Number
pH ~ 5	System II [#] System III [*] System IV [*]	cationic-MC3	6,400	
		Chol	4,992	
		DSPC	1,284	
		dsDNA	64	
EM	System II [#] System III [*] System IV [*]	cationic-MC3	5,174	
		neutral-MC3	1,226	
		Chol	4,992	
		DSPC	1,284	
		dsDNA	64	
Solvent	WAT	POPC	4,410	
		Chol	2,940	
		POPE	1,078	
		POPI	686	
		POPS	686	
Solvent		WAT	~1,350,000	

[#]Systems I and II that included endosomal membrane (EM) exhibited pore formation. In contrast, ^{*}Systems III and IV with EM did not show any pore formation during a runtime of 20 to 30 μ s. All systems feature explicit coarse-grained (CG) water and sodium/chloride (NaCl) counter-ions. Additionally, Systems II, III, and IV contain physiological ion concentrations of 150 mM NaCl.

Table S3. Number of lipids exchanged between the LNP and the EM at 1.4 μ s

System	Lipids	Total number of molecules	LNP (remained) (moved)	EM (moved) (remained)	Exchanged lipid ratio (%)
LNP	MC3	6400	5574	826	12.9
	Chol (LNP)	4992	4642	350	7.0
	DSPC	1284	1247	37	2.9
EM	POPI	686	89	597	13.0
	POPS	686	78	608	11.4
	POPC	4410	240	4170	5.4
	POPE	1078	54	1024	5.0
	Chol (EM)	2940	61	2879	2.1
Total		11985		10491	