

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a	Confirmed
<input type="checkbox"/>	<input checked="" type="checkbox"/> The exact sample size (<i>n</i>) for each experimental group/condition, given as a discrete number and unit of measurement
<input type="checkbox"/>	<input checked="" type="checkbox"/> A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
<input type="checkbox"/>	<input checked="" type="checkbox"/> The statistical test(s) used AND whether they are one- or two-sided <i>Only common tests should be described solely by name; describe more complex techniques in the Methods section.</i>
<input checked="" type="checkbox"/>	<input type="checkbox"/> A description of all covariates tested
<input type="checkbox"/>	<input checked="" type="checkbox"/> A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
<input type="checkbox"/>	<input checked="" type="checkbox"/> A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
<input type="checkbox"/>	<input checked="" type="checkbox"/> For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
<input checked="" type="checkbox"/>	<input type="checkbox"/> For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
<input checked="" type="checkbox"/>	<input type="checkbox"/> For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
<input type="checkbox"/>	<input checked="" type="checkbox"/> Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection	FACSCanto (BD Biosciences), Illumina NovaSeq 6000 sequencing platform, Applied Biosystems 7500 Fast Real-Time PCR System, AxioScan (ZEISS), ZEISS Axio Observer
Data analysis	FIJI software, FlowJo V10, Graphpad Prism 8, QuPath v0.5.1, R code (https://github.com/MMdR-lab/Feiyang-triple-therapy.git)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

All data associated with this study are present in the paper or the Supplementary Information. Source data are provided with this paper. The RNA sequencing data used in this study are available in the GSE301844 (reviewer token gzsdcicuvfevpgb).

Research involving human participants, their data, or biological material

Policy information about studies with [human participants or human data](#). See also policy information about [sex, gender \(identity/presentation\), and sexual orientation](#) and [race, ethnicity and racism](#).

Reporting on sex and gender	There is no human participant involved.
Reporting on race, ethnicity, or other socially relevant groupings	There is no human participant involved.
Population characteristics	There is no human participant involved.
Recruitment	There is no human participant involved.
Ethics oversight	There is no human participant involved.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

☒ Life sciences ☐ Behavioural & social sciences ☐ Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	Sample size used determined based on previous experience and similar assays (PMID: 39270021, 37616051). No statistical test was used to pre-determine the sample sizes.
Data exclusions	No data was excluded.
Replication	Each biological experiments were performed in independent replicates. All experiments were repeated at least 3 times, to ensure reproducibility of the results. There were no experiments that we could not replicate independently in this study.
Randomization	In all experiments, mice were randomly assigned to each group and the order in which analysis procedures were performed were done at random.
Blinding	Blinding was not applied to most experiments because data collection and analysis were performed by the same investigators.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study
<input type="checkbox"/>	<input checked="" type="checkbox"/> Antibodies
<input type="checkbox"/>	<input checked="" type="checkbox"/> Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology
<input type="checkbox"/>	<input checked="" type="checkbox"/> Animals and other organisms
<input type="checkbox"/>	<input checked="" type="checkbox"/> Clinical data
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern
<input checked="" type="checkbox"/>	<input type="checkbox"/> Plants

Methods

n/a	Involved in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input type="checkbox"/>	<input checked="" type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

Antibodies

Antibodies used	Phospho-p44/42 MAPK (Erk1/2) (Thr202/Tyr204) (D13.14.4E) XP® Rabbit mAb Cell Signaling Technology #4370 WB, IHC
-----------------	---

Antibodies used

ERK 2 Antibody (C-14) (Santa Cruz, sc-154, WB)
 Phospho-S6 Ribosomal Protein (Ser240/244) (D68F8) XP® Rabbit mAb (Cell Signaling Technology #5364, WB, IHC)
 S6 Ribosomal Protein (54D2) Mouse mAb (Cell Signaling Technology #2317, WB)
 Anti-human cleaved PARP (Asp214) (D64E10) XP® Rabbit mAb (Cell Signaling Technology #5625, WB)
 Anti-mouse cleaved PARP (Asp214) Antibody (Cell Signaling Technology #9544, WB)
 Anti-human ATF-4 (D4B8) Rabbit mAb (Cell Signaling Technology #11815, WB, IHC)
 Anti-mouse ATF-4 antibody (Abcam ab216839, WB, IHC)
 Phospho-eIF2α (Ser51) (D9G8) XP® Rabbit mAb (Cell Signaling Technology #3398, WB)
 eIF2α (L57A5) Mouse mAb (Cell Signaling Technology #2103, WB)
 MTHFD1 Polyclonal antibody (Proteintech 10794-1-AP, WB)
 MTHFD2 Polyclonal antibody (Proteintech 12270-1-AP WB, IHC)
 Phospho-Histone H2A.X (Ser139) Antibody (Cell Signaling Technology #2577, WB)
 Ki-67 (8D5) Mouse mAb (Cell Signaling Technology #9449, IHC)
 Anti-α-actinin Antibody (H-2) (Santa Cruz, sc-17829, WB)
 Monoclonal Anti-β-Actin antibody (clone AC-15) (Sigma-Aldrich #A5441, WB)
 GAPDH (D16H11) XP® Rabbit mAb (Cell Signaling Technology #5174, WB)
 Amersham ECL Mouse IgG, HRP-linked whole Ab (from sheep) (Cytiva NA931V, WB)
 Amersham ECL Rabbit IgG, HRP-linked whole Ab (from donkey) (Cytiva NA934V, WB)
 EnVision+ Single Reagent (HRP, Mouse) (Agilent Dako K4001, IHC)
 EnVision+ Single Reagent (HRP, Rabbit) (Agilent Dako K4003, IHC)

Validation

All antibodies used in this study have been previously validated by the manufacturer, as stated on their associated product websites, and by our own lab in previous experiments.

Eukaryotic cell lines

Policy information about [cell lines and Sex and Gender in Research](#)

Cell line source(s)

BLM (Gifted by Dr. Ian R Watson, MUHC Research Institute)
 WM3406 (Gifted by Dr. April Rose, Lady Davis Institute)
 MeWo (Gifted by Dr. Ian R Watson, MUHC Research Institute)
 HBL (Gifted by Dr. Ghanem Ghanem, Institut Jules Bordet)
 YUGOE (Gifted by Dr. April Rose, Lady Davis Institute)
 WM3623 (Gifted by Dr. April Rose, Lady Davis Institute)
 MaNRAS1007 (Gifted by Dr. Lionel Larue, INSERM, First described in PMID: 31251472)

Authentication

All of the cell lines used were authenticated by short tandem repeat profiling.

Mycoplasma contamination

The cell lines were tested as Mycoplasma negative.

Commonly misidentified lines
(See [ICLAC](#) register)

We did not use commonly misidentified lines.

Animals and other research organisms

Policy information about [studies involving animals](#); ARRIVE guidelines recommended for reporting animal research, and [Sex and Gender in Research](#)

Laboratory animals

C57BL/6N mice (6-8 weeks old); NOD/SCID mice (6-10 weeks old). Mice were housed under 12 hour light/dark cycle at 20-24°C and 45-65% humidity

Wild animals

No wild animals were used in the study.

Reporting on sex

We used male C57BL/6N mice and female NOD/SCID mice since MaNRAS1007 cell is from male mice and WM3406 cell is from female patient.

Field-collected samples

No field-collected samples were used in the study.

Ethics oversight

Animal experiments were conducted according to the regulations established by the Canadian Council of Animal Care, and protocols approved by McGill University Animal Care and Use Committee (#2015-7672).

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Clinical data

Policy information about [clinical studies](#)

All manuscripts should comply with the ICMJE [guidelines for publication of clinical research](#) and a completed [CONSORT checklist](#) must be included with all submissions.

Clinical trial registration

No clinical data is collected in this study.

Study protocol

No clinical data is collected in this study.

Data collection	No clinical data is collected in this study.
Outcomes	No clinical data is collected in this study.

Plants

Seed stocks	No plant is used in this study.
Novel plant genotypes	No plant is used in this study.
Authentication	No plant is used in this study.

Flow Cytometry

Plots

Confirm that:

- ☐ The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).
- ☒ The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).
- ☒ All plots are contour plots with outliers or pseudocolor plots.
- ☒ A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation	Melanoma cells were seeded into 6-well plates at 100,000 cells per well. Following indicated treatments, cells were trypsinized, centrifuged at 240g for 5 min, and washed twice in PBS. For apoptosis detection of non-fixed cells, Alexa Fluor™ 647-Annexin V (Invitrogen™, A23204) and Propidium Iodide (PI) Staining Solution (BD Biosciences, 556463) were diluted in 1xbinding buffer (BD Biosciences, 556454), and subsequently mixed with cells following the manufacturer's instructions.
Instrument	FACSCanto (BD Biosciences)
Software	FlowJo V10
Cell population abundance	Cell population abundance was dependent on the apoptosis level.
Gating strategy	For all gating strategy in this article, we gated with debris exclusion (appropriate FSC-A/SSC-A) and doublets exclusion (FSC-A/FSC-H)

- ☐ Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.