

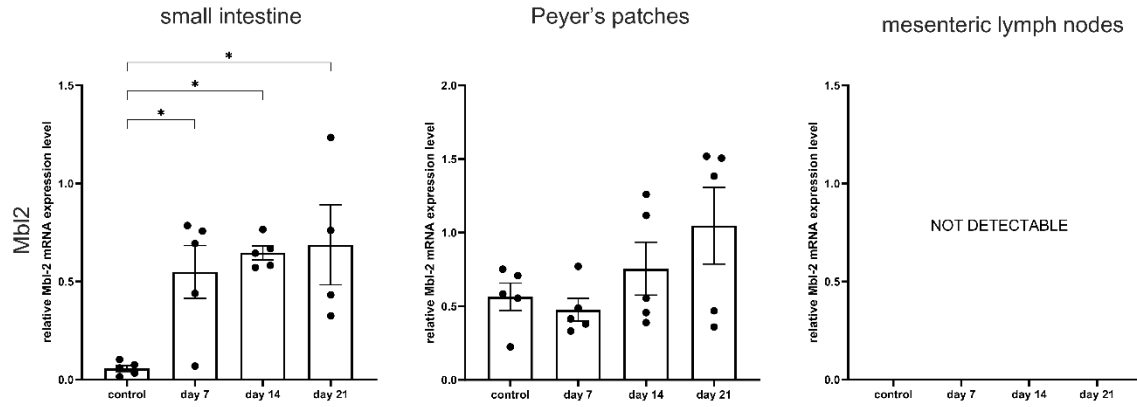
SUPPLEMENTARY INFORMATION

Unravelling the cellular sources and location of IL-17A production during a

***Giardia* infection**

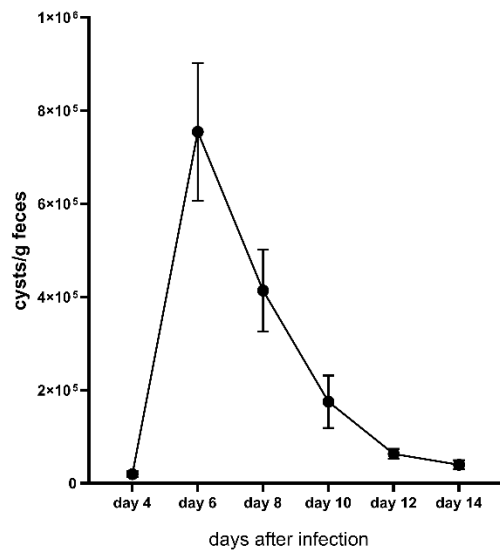
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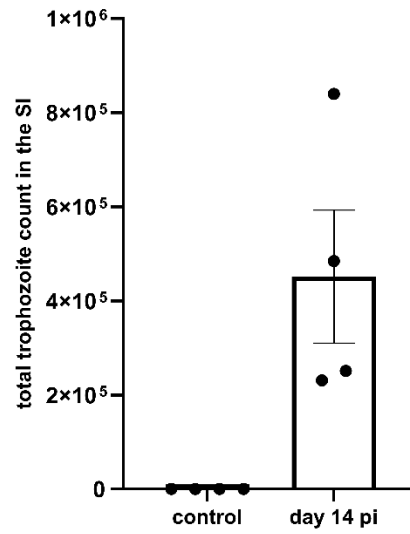


Supplementary Figure S1. Relative Mbl2 mRNA expression levels in the SI, PP and MLN of C57BL/6 mice at day 7, 14 and 21 p.i. as measured by RT-qPCR (n=5). Differences were analyzed using a Kruskal-Wallis test (* $P < 0.05$).

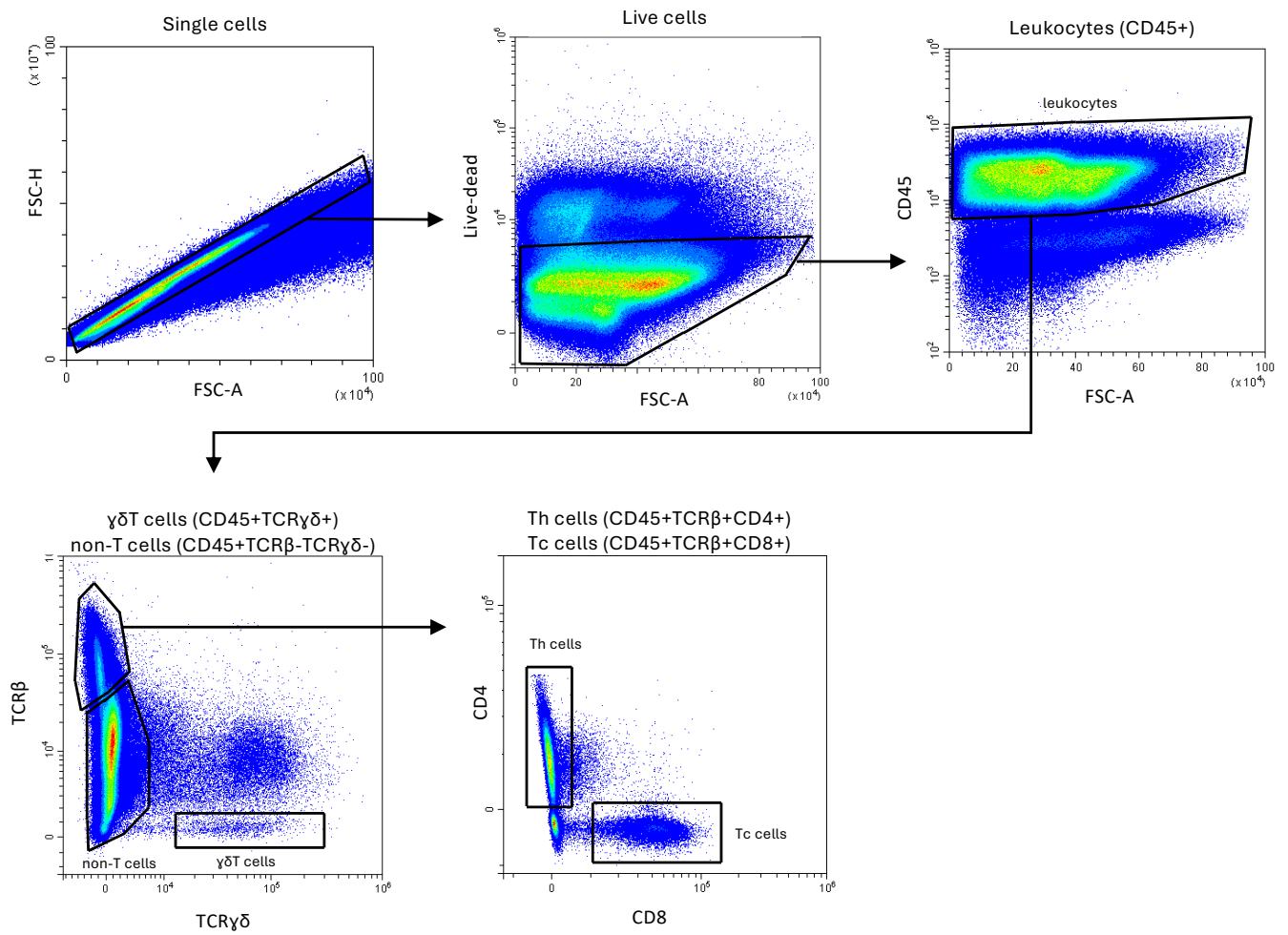
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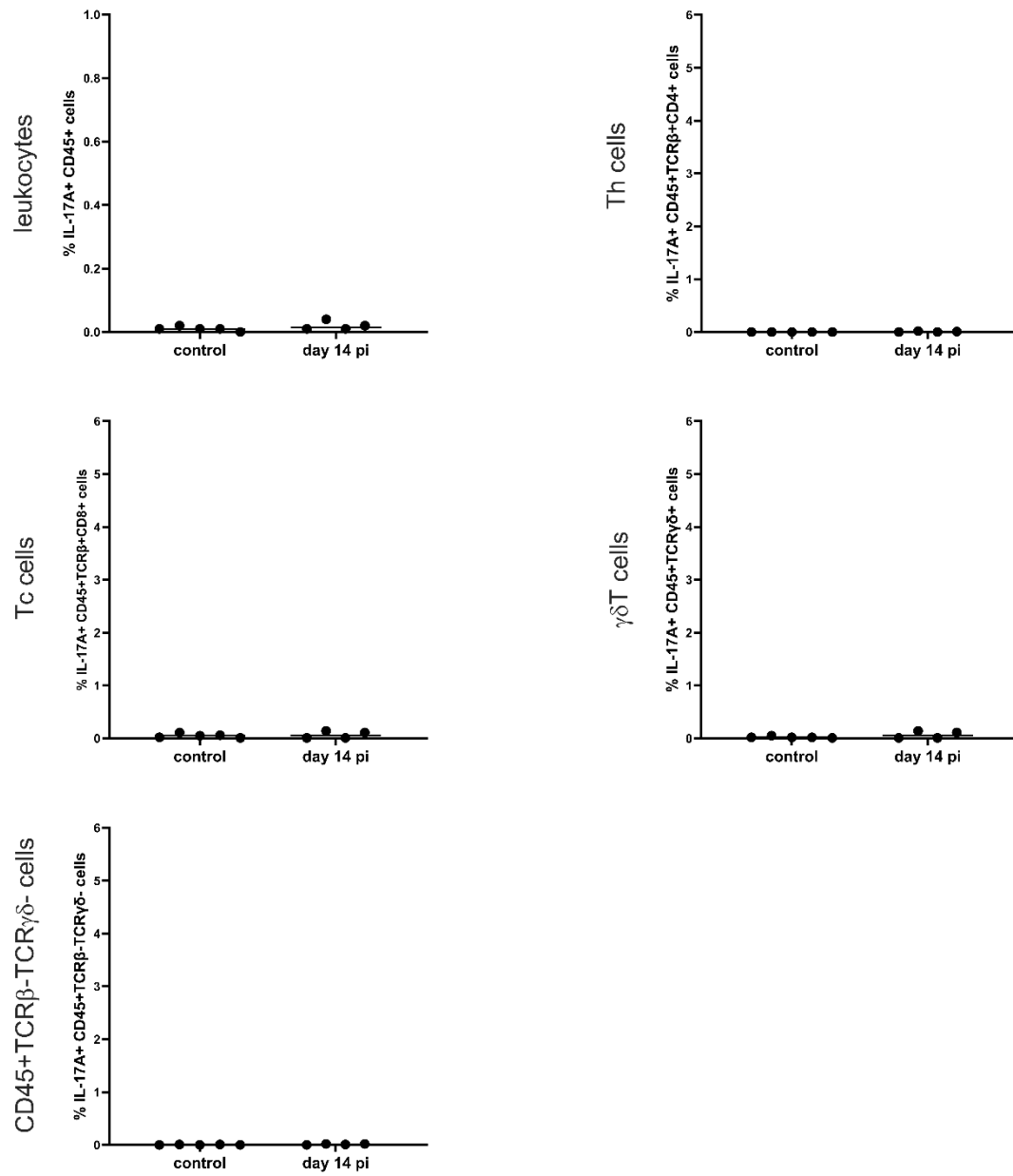
b



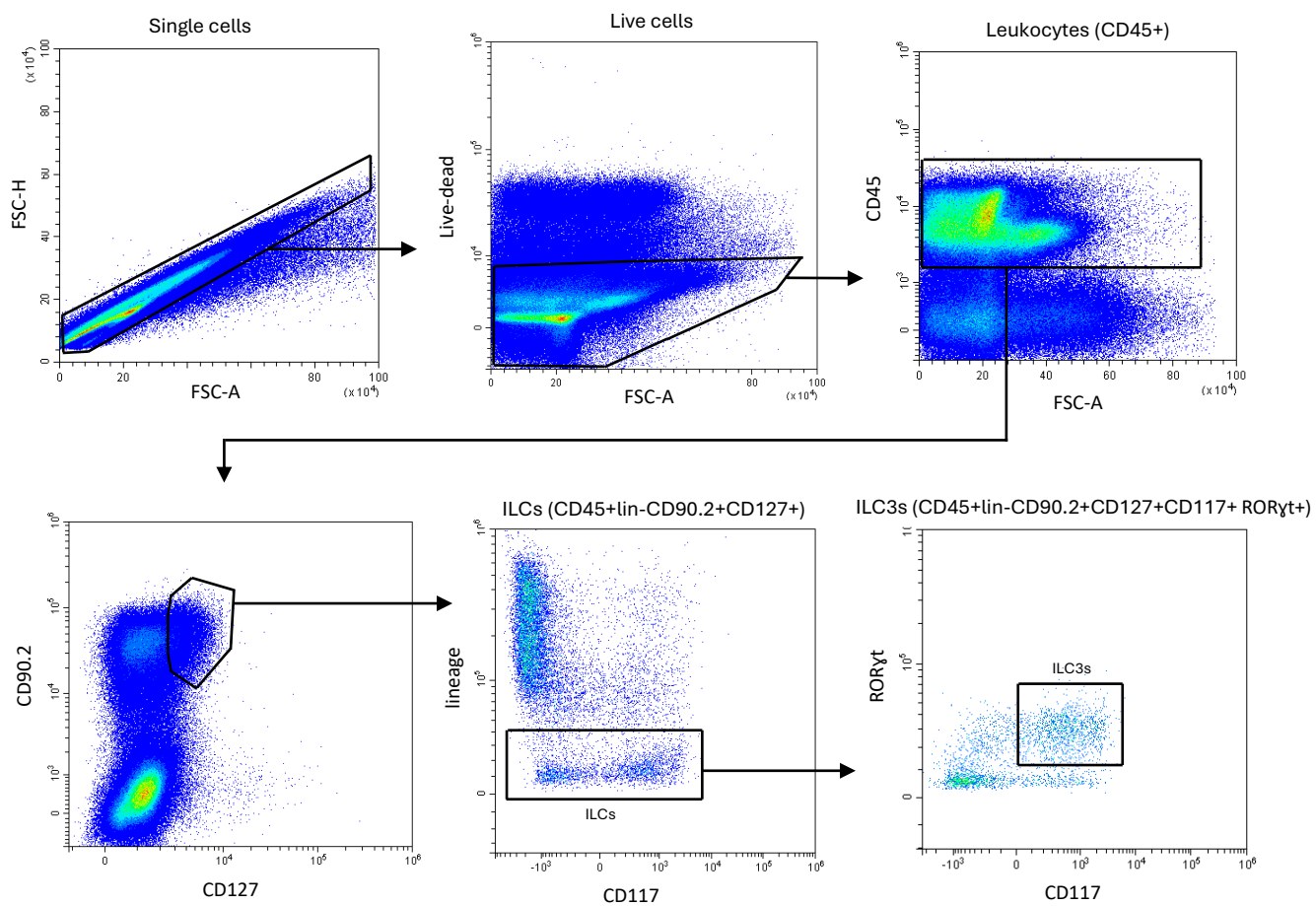
Supplementary Figure S2. Parasitological parameters of the second study in C57BL/6 mice. (a) Fecal cyst counts monitored every 2-3 days until day 14 p.i.. Each point on the graph represents the mean cyst output per gram of feces, with SEM as error bars (n=4). (b) Small intestinal trophozoite counts at day 14 p.i. (n=4).



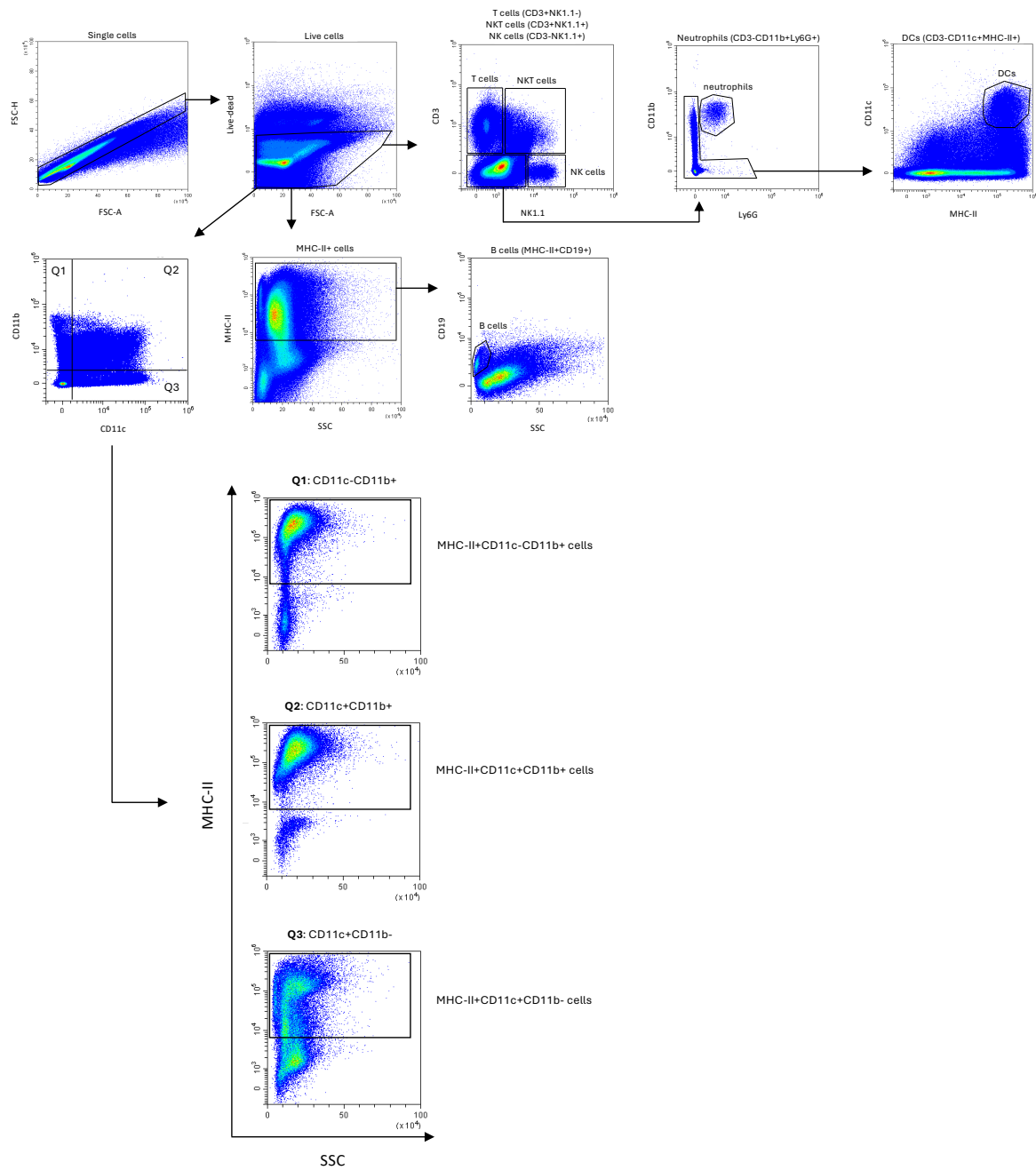
Supplementary Figure S3. Flow cytometry panel 1. Gating strategy for leukocytes, Th cells, Tc cells, $\gamma\delta$ T cells and non-T cells.



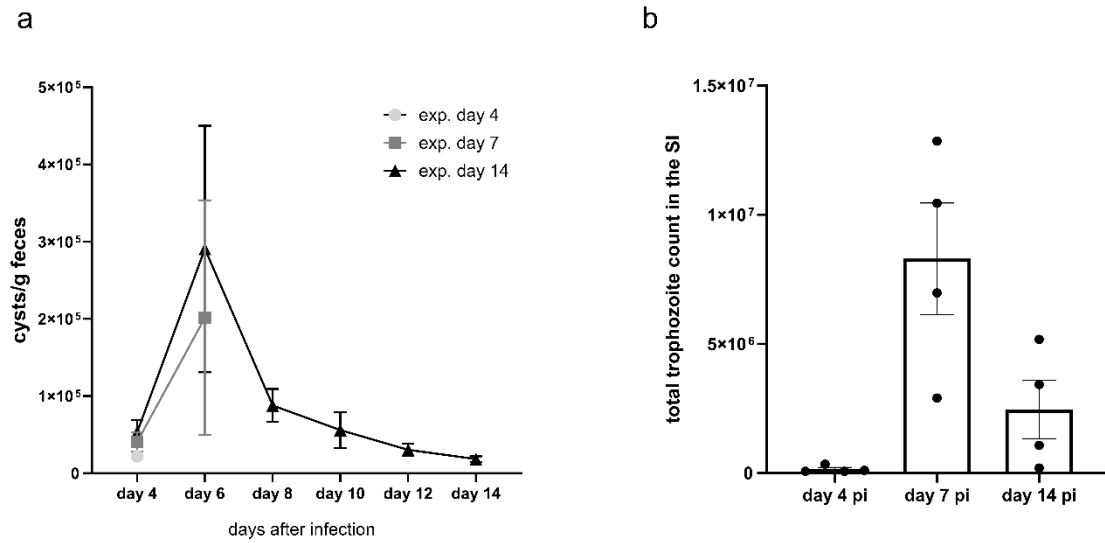
Supplementary Figure S4. IL-17A production by different cell types in the intra-epithelial compartment of *G. muris* infected C57BL/6 mice at day 14 p.i. as measured by flow cytometry, expressed as %IL-17A+ cells of parent population. Differences were analyzed using a Mann-Whitney test.



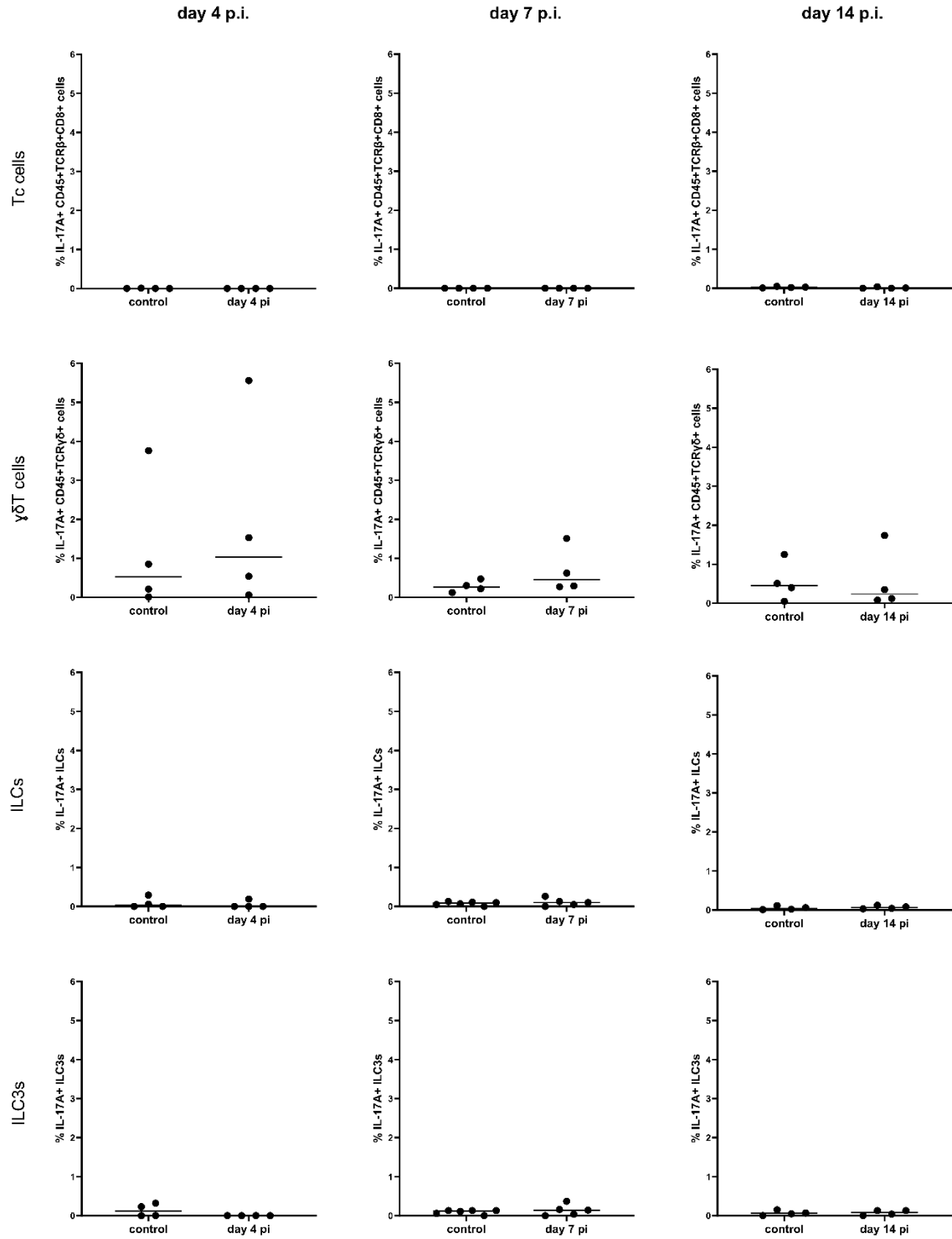
Supplementary Figure S5. Flow cytometry panel 2. Gating strategy for ILCs and ILC3s.



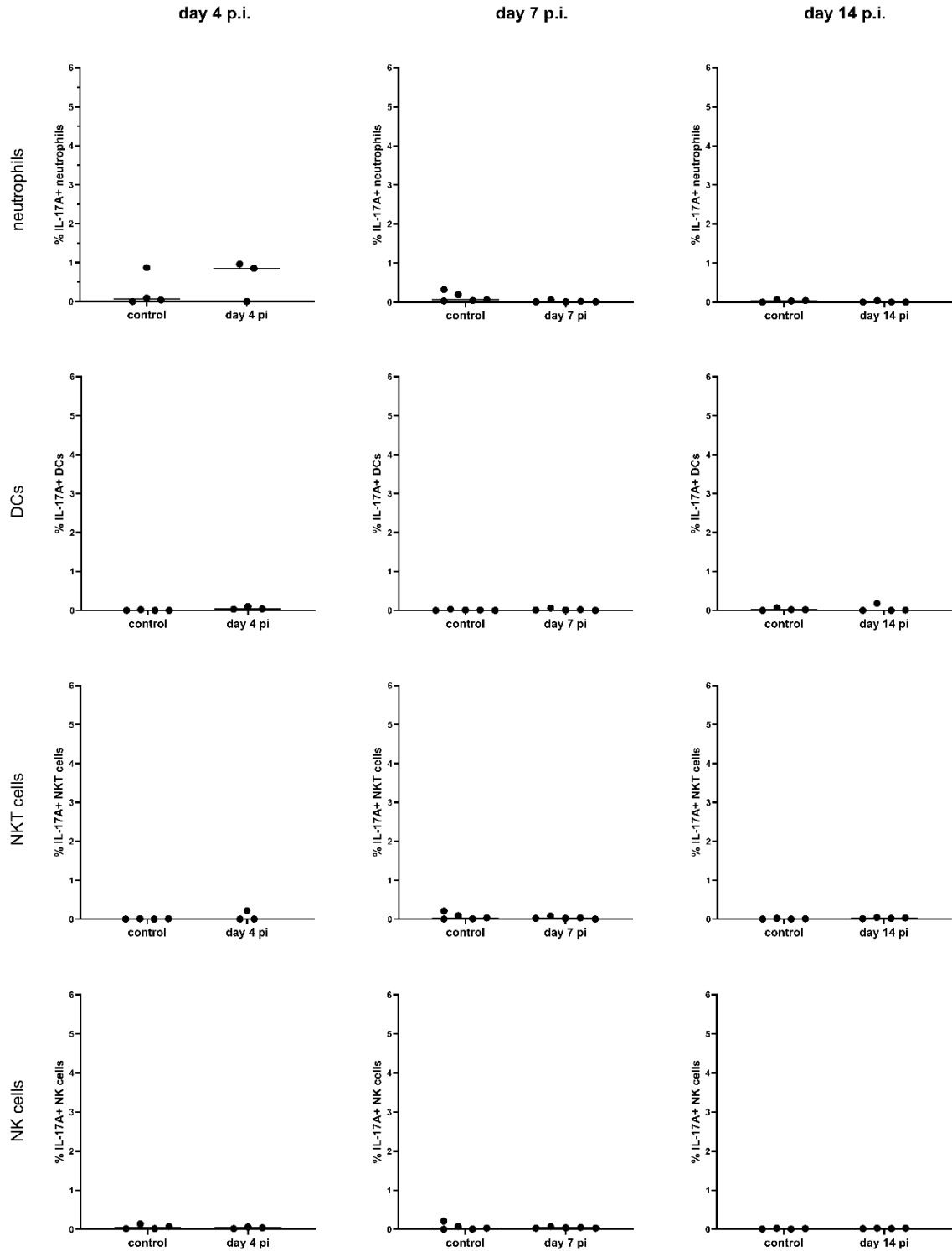
Supplementary Figure S6. Flow cytometry panel 3. Gating strategy for T cells, NKT cells, NK cells, neutrophils, DCs, MHC-II+ cells and MHC-II+CD11c[±]/CD11b[±] cells. Only in TKO mice, phenotyping of B cells was included.



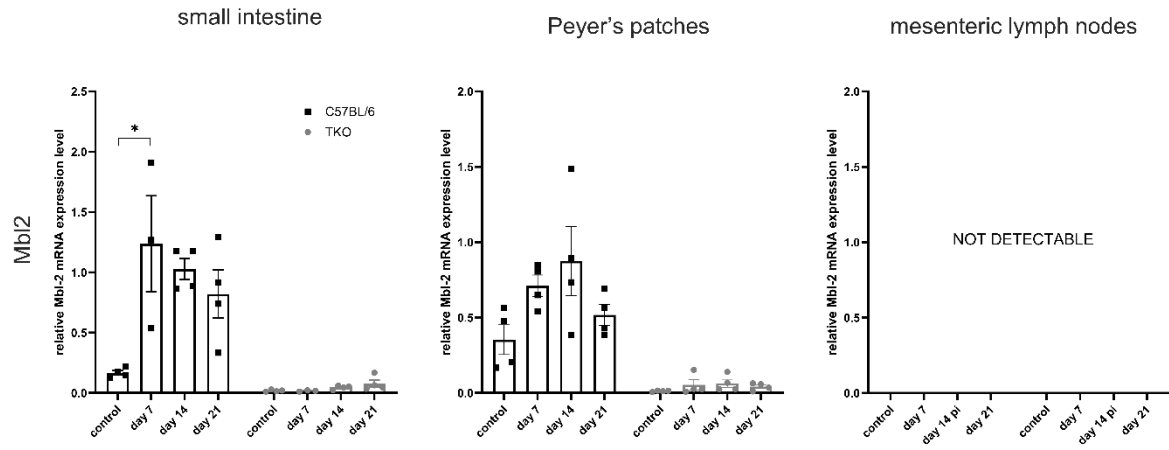
Supplementary Figure S7. Parasitological parameters of the third study in C57BL/6 mice. (a) Fecal cyst counts monitored every 2-3 days until day 4, day 7 and 14 p.i. for each study. Each point on the graph represents the mean cyst output per gram of feces, with SEM as error bars (n=4). (b) Small intestinal trophozoite counts at day 4, 7 and 14 p.i. (n=4).



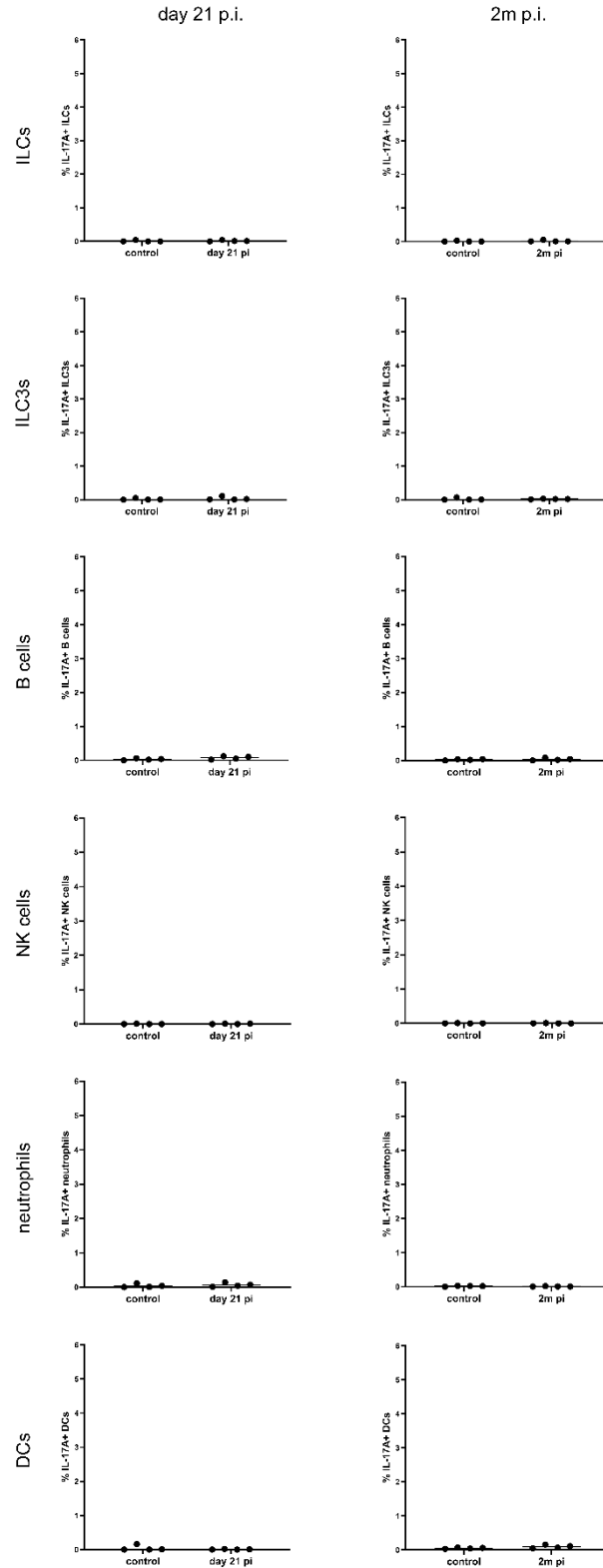
Supplementary Figure S8. IL-17A production by different cell types in the lamina propria compartment of *G. muris* infected C57BL/6 mice at day 4, 7 and 14 p.i. as measured by flow cytometry, expressed as %IL-17A+ cells of parent population. Differences were analyzed using a Mann-Whitney test.



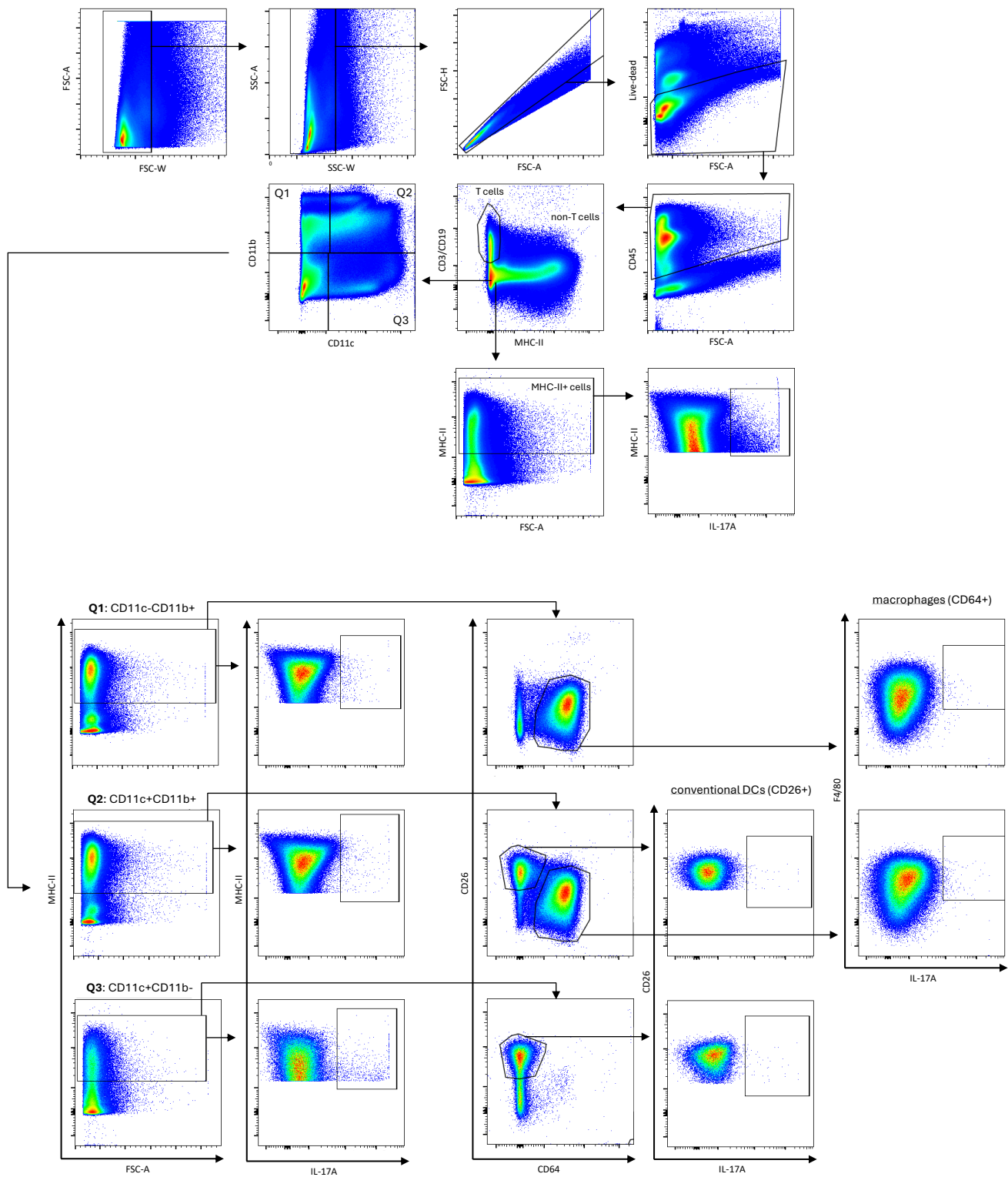
Supplementary Figure S9. IL-17A production by different cell types in the lamina propria compartment of *G. muris* infected C57BL/6 mice at day 4, 7 and 14 p.i. as measured by flow cytometry, expressed as %IL-17A+ cells of parent population. Differences were analyzed using a Mann-Whitney test.



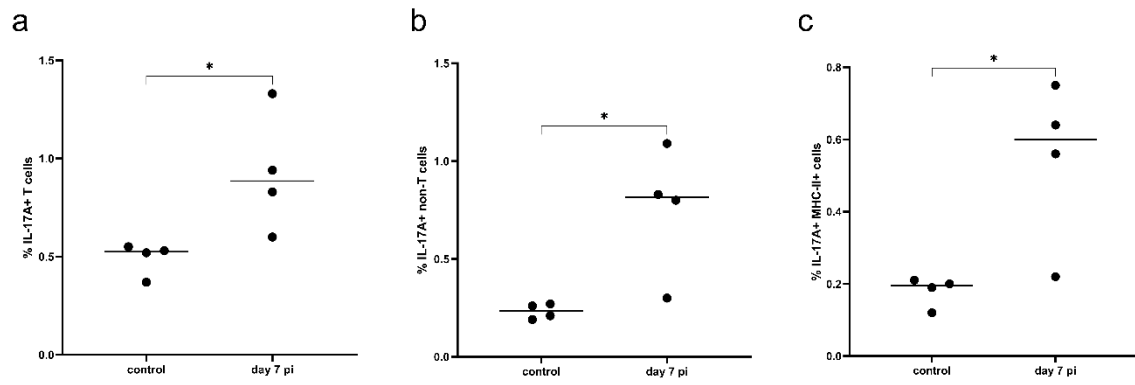
Supplementary Figure S10. Relative Mbl2 mRNA expression levels in the SI, PP and MLN of C57BL/6 and TKO mice at day 7, 14 and 21 p.i. as measured by RT-qPCR (n=4). Differences were analyzed using a Kruskal-Wallis test (* $P < 0.05$).



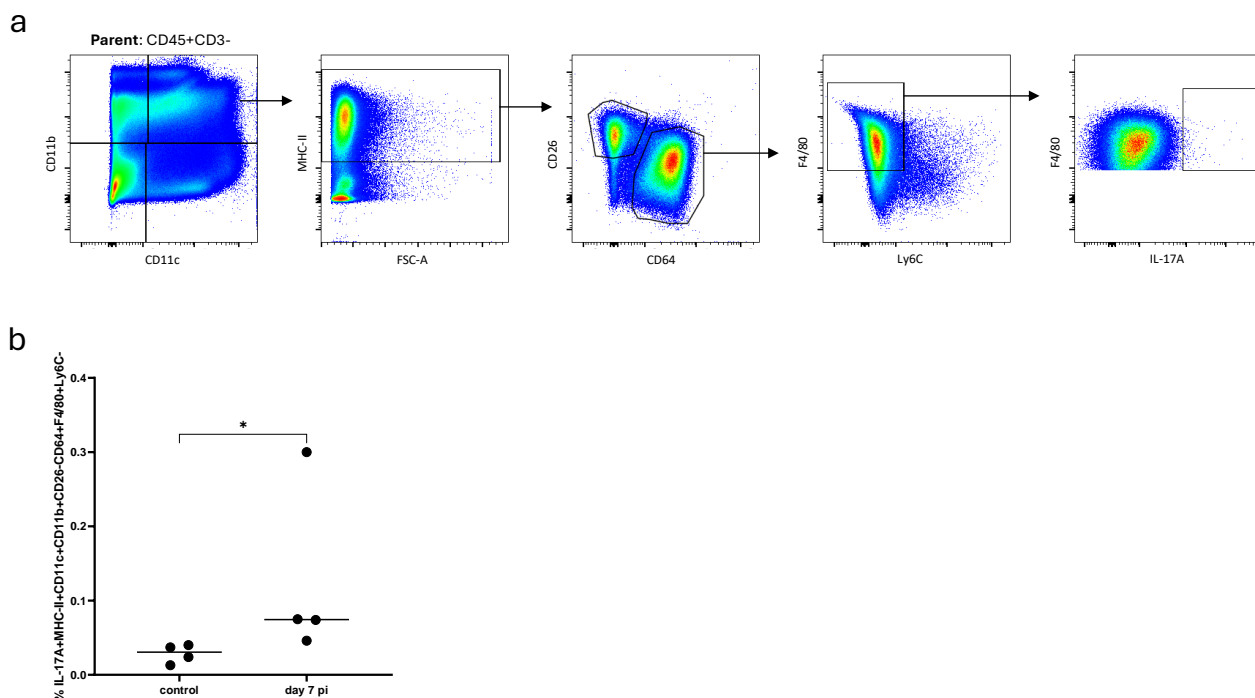
Supplementary Figure S11. IL-17A production by different cell types in the lamina propria compartment of *G. muris* infected TKO mice at day 21 and 2 months p.i. as measured by flow cytometry, expressed as %IL-17A⁺ cells of parent population. Differences were analyzed using a Mann-Whitney test.



Supplementary Figure S12. Panel targeting antigen-presenting cells. Gating strategy for T cells, non-T cells, MHC-II⁺ cells and different subtypes of MHC-II⁺ cells, including conventional DCs and macrophages.



Supplementary Figure S13. IL-17A production by T cells (a), non-T cells (b) and MHC-II+ cells (c) in the lamina propria compartment of *G. muris* infected C57BL/6 mice at day 7 p.i. as measured by flow cytometry, expressed as %IL-17A+ cells of parent population. Differences were analyzed using a Mann-Whitney test (*P < 0.05).



Supplementary Figure S14. (a) Flow cytometry gating strategy for MHC-II+CD11c+CD11b+CD26-CD64+F4/80+Ly6C- cells. (b) IL-17A production by this cell type in the lamina propria compartment of *G. muris* infected C57BL/6 mice at day 7 p.i. as measured by flow cytometry, expressed as %IL-17A+ cells of parent population. Differences were analyzed using a Mann-Whitney test (* $P < 0.05$).