## **EXTENDED DATA TABLES**

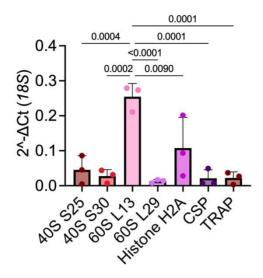
# Extended data table 1 | Plasmodium falciparum homology

	Number of	P. falciparum			
Protein name	patients	homology (%)			
60S L4/L1	7	83			
ETRAMP A5KBH5	7	63			
40S S2	6	97			
Don Juan	6	53			
ETRAMP A5K676	6	43			
40S S11	5	99			
Histone H2A A5K214	5	98			
EF 1-alpha	5	98			
40S S23	5	98			
60S L32	5	94			
60S L29	5	91			
40S S8	5	89			
40S S6	5	89			
40S S30	5	89			
40S S25	5	86			
60S L13	5	80			
Histone H2A A5K7L8	4	97			
60S L37	4	97			
60S L30	4	92			
Alba domain	4	91			
Ubiq/Rib	4	91			
60S L36	4	90			
60S L35	4	81			
60S L23a	4	64			
Unch. A5K8G9	4	33			
Histone H3	3	100			
60S L37a	3	100			
50S S28e	3	97			
60S L44	3	97			
60S L10	3	96 2.7			
60S L8	3	95			
60S L9	3	94			
60S L33-B	3	92			
60S L18a	3 3	92			
60S L13a	3	91			
60S L34-A	3	91			
60S L31	3	90			
60S L17	3	89			
60S L7a	3	84			
PPR_long domain	3	59 43			
Unch. A5K9Y3	3 3	43			
PI3K/PI4K	3	36			
Unch. A5K4L2 Unch. A5K8G8	3	32 0			
	3				
Unch. A5KC18	3	23			

## Extended data table 2 | Primers used for *Plasmodium falciparum* liver stage expression

Primer	Sequence
NF135 40S S25-1F	5'-TCTCCCAGAGCTGTTTACCC-3'
NF135 40S S25-1R	5'-TCATCTTCCAAGTTGTCACACA-3'
NF135 40S S30-1F	5'-ATGGGAAAGGTACATGGATCATTAG-3'
NF135 40S S30-1R	5'-ACTTTGGGACTTTAGGAGTCTGG-3'
NF135 60S L13-1F	5'-AAGAGTTAAAGGGAGCCGGA-3'
NF135 60S L13-1R	5'-ACCTCCAATACCTTTCTTTGCT-3'
NF135 60S L29-1F	5'-ACCGTAAAGCCCATAAGAATGGA-3'
NF135 60S L29-1R	5'-GGATCCAACCCTTTACGCGA-3'
NF135 H2A-1F	5'-TAGGTTCCACAGCAGCAGTT-3'
NF135 H2A-1R	5'-AACACCACCACCAGCAATTG-3'
NF135 CSP-1F	5'-GCCAAATGACCCAAACCGAA-3'
NF135 CSP-1R	5'-GGTTTATTAGCAGAGCCAGGC-3'
NF135 TRAP-1F	5'-AGAGAAACACGTCCACATGG-3'
NF135 TRAP-1R	5'-TCCAGCTATTCCACCTGCAA-3'
18S rRNA (3D7) -1R	5'-GGGTTCTGGGGCGAGTATT-3'
18S rRNA (3D7) -1R	5'-GCCATGCATCACCATCCAAG-3'

### **EXTENDED DATA FIGURE**



Extended data figure 1 | Antigen gene expression in humanized mouse livers infected with *Plasmodium falciparum* (*Pf*). The expression of genes related to antigens found in *P. vivax* immunopeptidomics was analysed in the liver stage of *Pf* infection. Pf 18S rRNA was used as endogenous gene. CSP and TRAP genes were used as positive control. Statistical analyse was performed by one-way ANOVA, with Turkey's multiple comparison test. Actual P values are shown.

## Extended data figure 2 | Spectral match validation

Peptide		Biological peptide			Synthetic peptide				Validation		
Sequence	ID	n	[m/z]	z	RT	Peaks Score	[m/z]	z	RT	Peaks Score	(y/n)
AIEPSLAQLAQK	Ubqt/ribos-1	12	635.3643	2	41.42	37.49	634.8646	2	41.64	48.35	Υ
AKAVKKQKKTLKPV	60S L31	14	523.0190	3	21.67	45.30	NA	NA	NA	NA	NA
ANKALLPTAGDD	60S L4/L1-1	12	593.3108	2	34.53	50.03	593.3093	2	34.31	52.84	Υ
APISAGIKKPHR	Hist. H3	12	425.5951	3	25.01	40.59	425.5946	3	23.20	23.34	Υ
AQKIKKKKKLTPA	Don Juan	13	494.6678	3	24.09	36.08	494.6680	3	20.52	41.55	Υ
ASGGVLPNIHNV	Hist. H2A-2	12	589.3206	2	39.78	25.89	589.3203	2	39.89	37.05	Υ
AVKKVGKIVK	60S L32-2	10	357.2537	3	22.97	22.03	357.2534	3	20.62	27.81	Υ
DNNEHVVQEKTV	Unch A5JZN7.2	12	706.3463	2	27.81	53.74	706.3451	2	26.94	49.45	Υ
DNNEHVVQEKTVSF	Unch A5JZN7.1	14	549.2657	3	35.20	61.62	549.2657	3	34.69	47.01	Υ
EDYSPRKV	Unch A5K8G9	8	497.2546	2	26.23	39.03	497.2538	2	24.87	29.67	Υ
GAFGKPNGV	60S L10	9	423.7279	2	30.37	19.14	423.7272	2	29.41	29.60	Υ
GKGKNKEKL	40S S25- 1	9	501.3087	2	20.20	33.76	501.3089	2	19.19	25.14	Υ
GKGKNKEKLNHAVF	40S S25-2	14	523.9681	3	25.17	48.81	523.9667	3	24.09	25.39	Υ
GLNQKQPTKGSNIQ	ETRAMP-2	14	757.4133	2	24.53	37.29	756.9106	2	23.51	45.90	Υ
GPKRATKIRK	40S S6-3	10	385.5873	3	20.09	38.30	385.5876	3	18.95	33.14	Υ
GRIGRYLKKGKYA	Hist. H2A-4	13	378.2308	4	32.42	33.32	378.2307	4	26.84	33.94	Υ
GRIGRYLKKGKYAK	Hist. H2A-1	14	410.2536	4	30.86	33.56	410.2541	4	25.27	17.95	Υ
GSKQVHVV	40S S8-2	8	427.2482	2	24.78	41.76	427.2479	2	23.31	37.69	Υ
GTGRKKGPNSKL	40S S30- 2	12	414.9137	3	20.78	46.04	414.9140	3	19.77	36.32	Υ
GTPIEKLHPI	60S L13-2	10	552.8251	2	34.59	27.43	552.8255	2	34.19	35.45	Υ
GVKKDVAK	40S S6-2	8	422,7661		19.95	34.31	422,7660	2	18.91	26.58	Υ
KKVAAGYKKLTD	ETRAMP-1	12	441.2663		24.33	41.58	441.2662	3	22.62		Υ
KKYKNKKFKPY	60S L35	11	491,2969		27.30	25.41	491.6320	3	23.68	19.14	Υ
KNIKSKNGIGGIPAD	60S L13-3	15	504.6231	3	27.71	31.77	504.6229	3	26.84	16.62	Υ
KSAGADSKSLKKLD	ETRAMP-5	14	483.6093		23.95	34.26	483.2758	3	22.52		Υ
KVPVPPTQAKKPKKN	Hist. H2A-5	15	415.7615		23.19	35.10	415.7620	4	21.87	40.76	Υ
LETYQNMKIQKQTP	40S S2	14	861.4510		32.68	40.56	861.9502	2	31.64	48.98	Y
LGGLNQKQPT	ETRAMP-4	10	528.2960		28.65	37.38	528.2964	2	27.59	39.32	Y
LILRAAIKTK	Unch A5K8J8	10	1126.7635			17.89	376.2605	3	29.11	20.42	N
LNQKQPTKGSNIQ	ETRAMP-3	13	728.4005			25.98	728.4000	2	22.49	41.12	Υ
MYKKVYVID	60S 13a	9	579.8156		32.90	39.80	579.8154	2	31.92	31.61	Y
NKALLPTAGDD	60S L4/L1-2	11	557.7906		34.65	43.43	557.7903	2	35.38	50.46	Y
NQLRPKKKLK	Ubqt/ribos- 2	10	418.2789		21.63	26.12	418.2785	3	19.94	19.81	Y
PKFFKNQRY	60S L29	9	409.8929		28.72	38.08	409.8927	3	27.66	31.37	Y
RLTGGKKKIHKKK	40S S8- 1	13	381.5072			43.53	NA	NA	NA	NA	NA
SDGTGRKKGPNSKL	40S S30- 1	14	482.2673		21.44	48.01	482.2670	3	20.18	37.03	Υ
SFFNSKKIKKGSKS	40S S11	14	529.3069			41.15	397.2319			37.69	Y
SGGVLPNIHNV	Hist. H2A-3	11	553.8022			29.28	533.8016			34.16	Ϋ́
SKNGKNRFIKPKIQ	40S S6- 1	14	415.2520			36.88	415.2518				Ϋ́
SSHAKGIVVEKV	40S S23	12	418.5784			44.55	418.5784				Y
TGRKKGPNSKL	40S S30- 3	11	395.9069			37.56	297.1819		19.67		N
TPIITNKPFG	ETRAMP-6	10	544.3114			36.66	544.3113		38.87	46.35	Y
VITDVGDSDIIKTNE	60S L30	15	809.9135			55.90	809.9135		41.11	55.13	Ϋ́
VSEVTTVEKDE	60S L9	11	618.3045			35.50	618.3038	2	29.67		Ϋ́
YESIEVSKID	60S L13- 1	10	591.7995			40.35	591.7988		37.62		Ϋ́
YGRIFKKKITKK	60S L4/L1-3	12	378.4963			35.21	378.2462				N
n amino acid			0,0.4000	-	_0.00	00.21	0,0.2402		20.02	10.00	

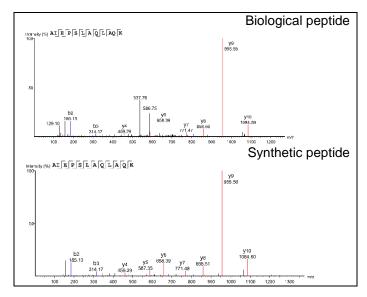
n amino acid length

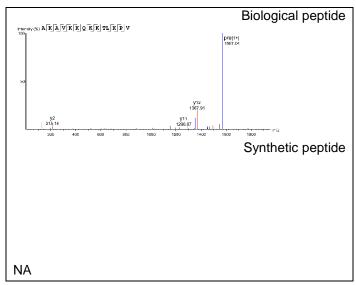
[m/z] Mass over Charge

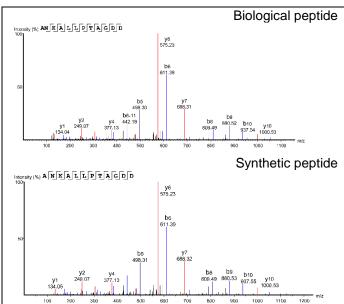
**z** charge state

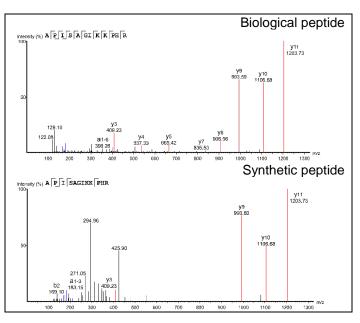
RT Retention Time

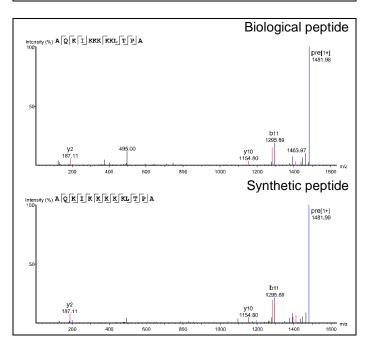
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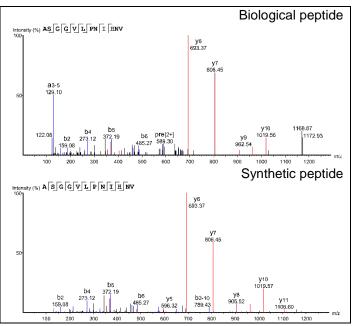


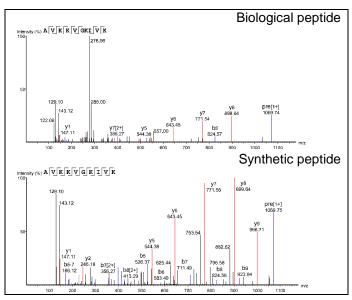


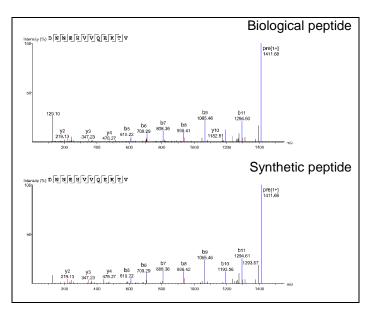


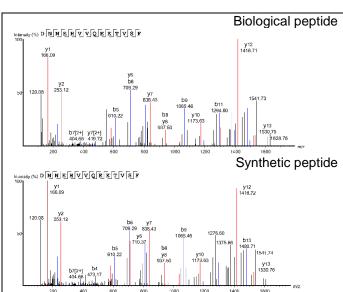


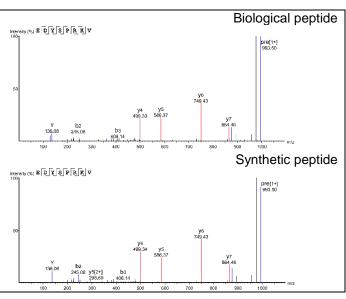


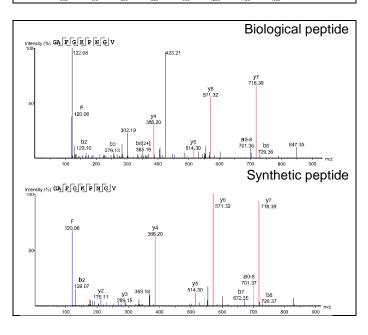


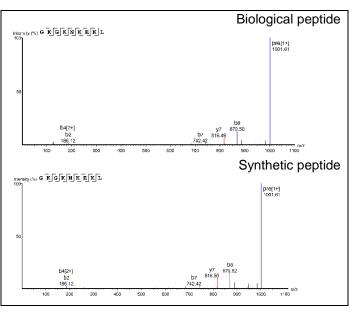


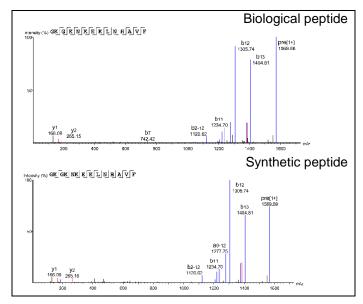


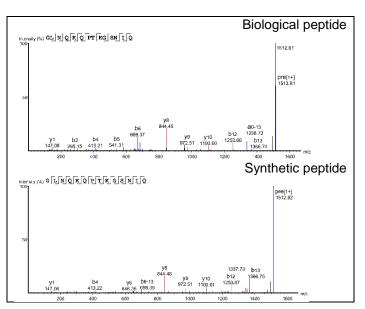


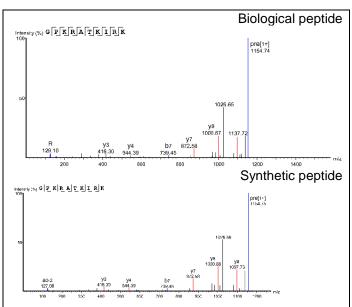


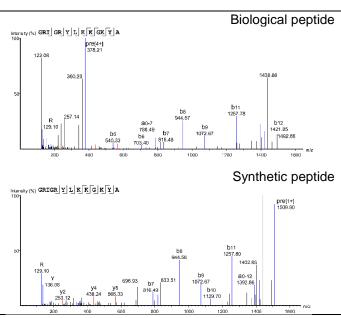


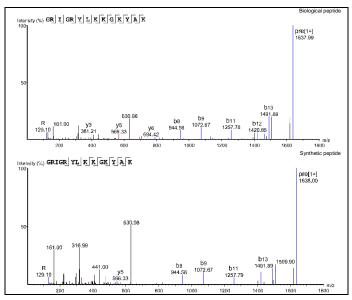


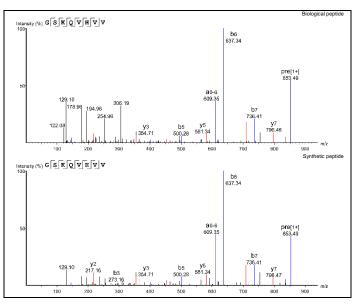


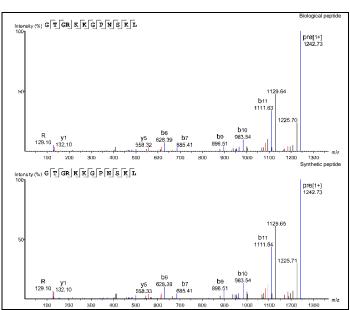


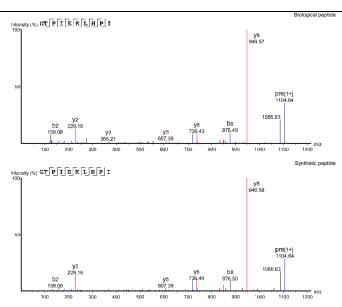


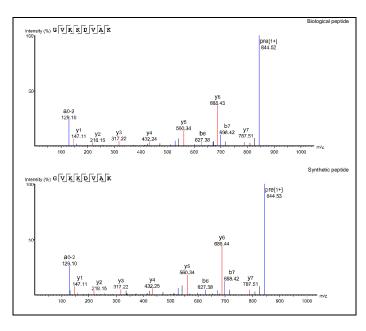


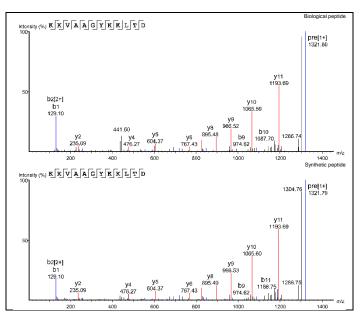


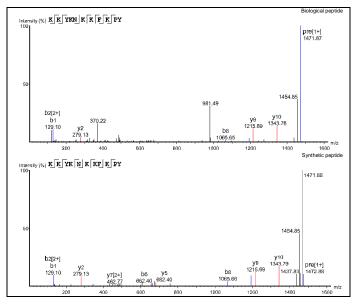


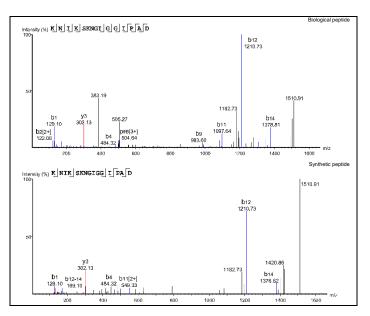


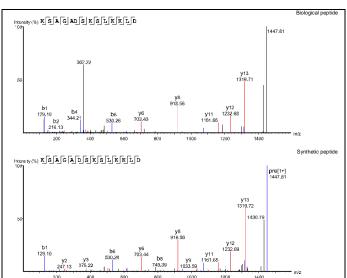


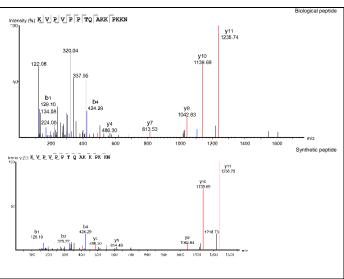


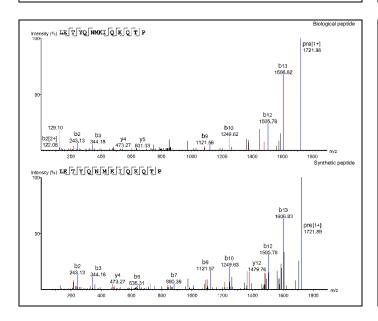


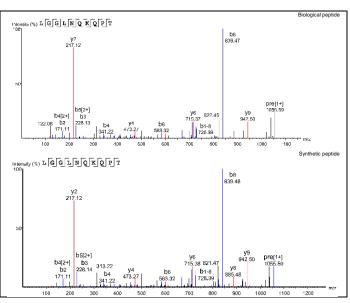


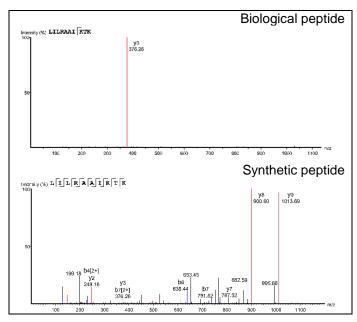


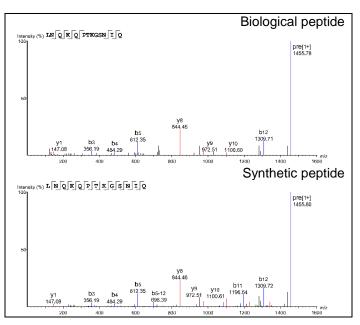


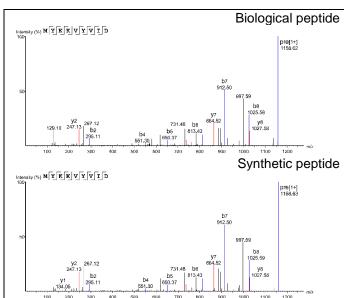


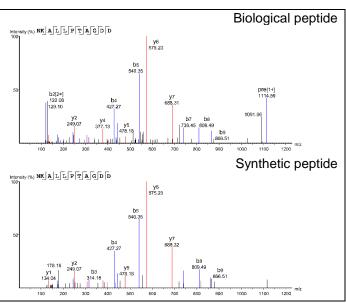


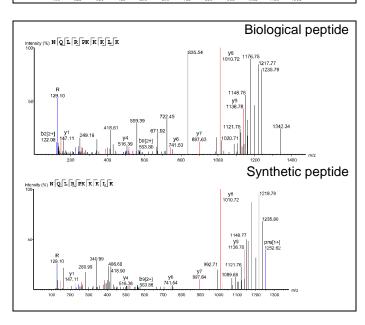


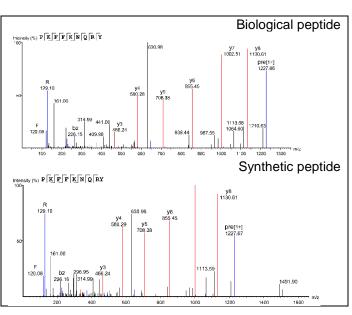


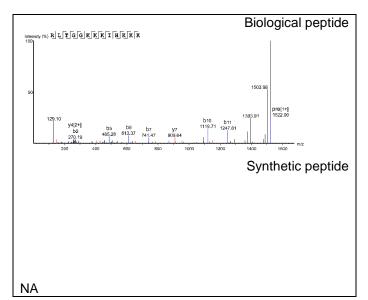


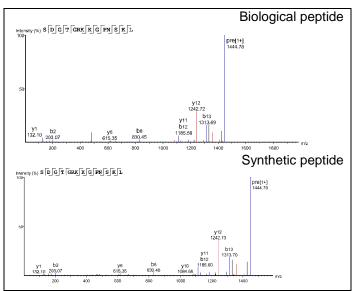


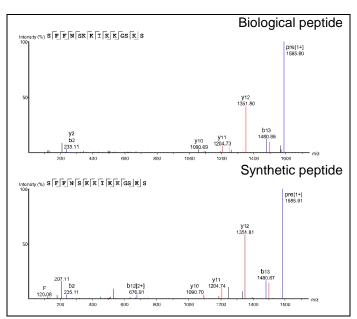


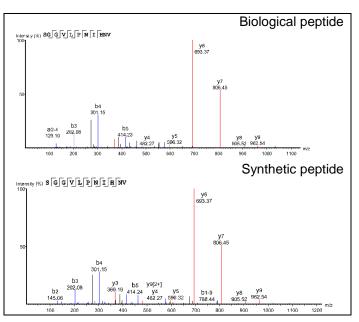


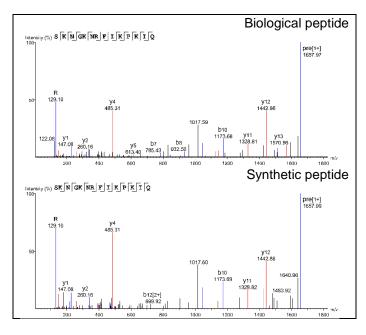


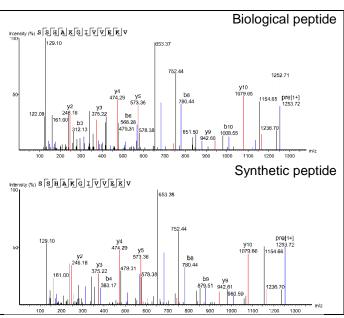


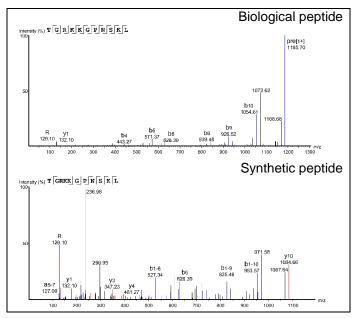


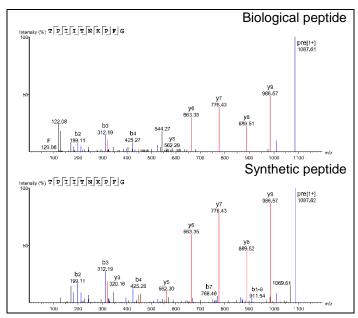


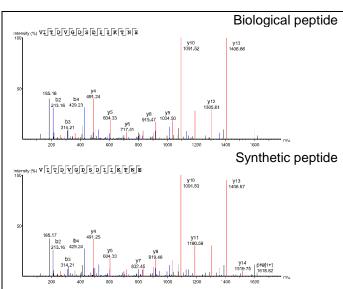


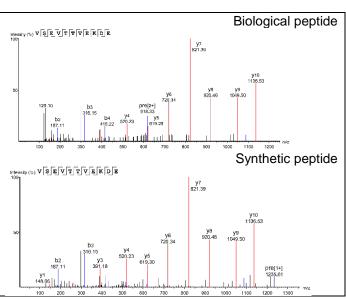


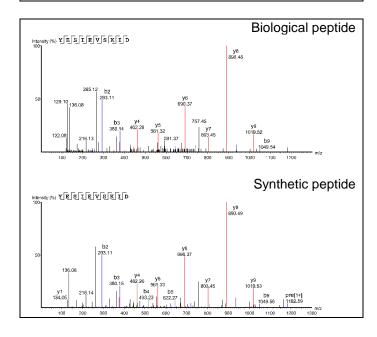


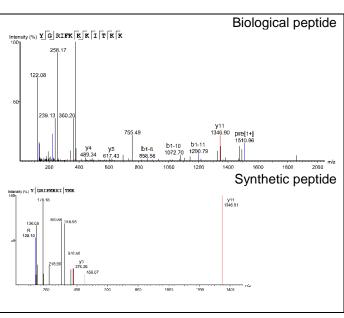


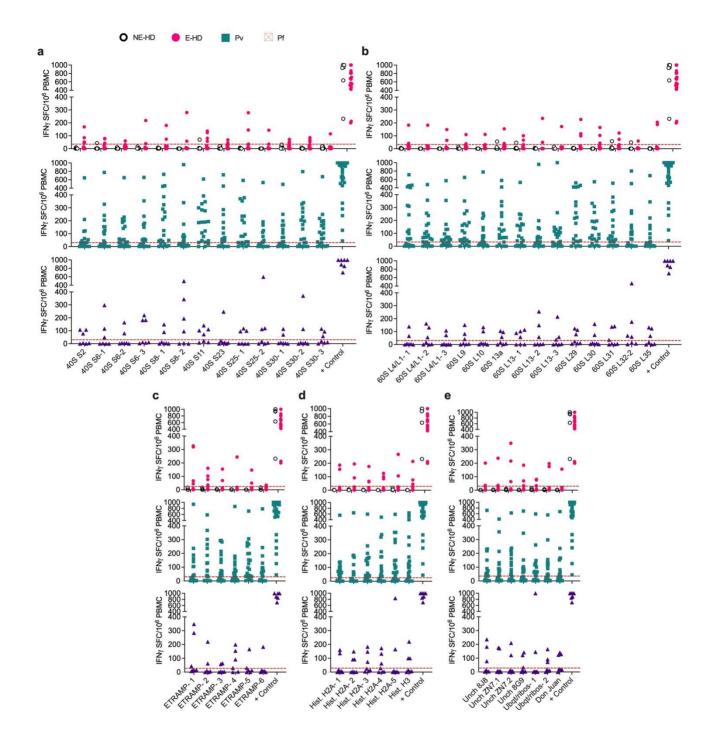




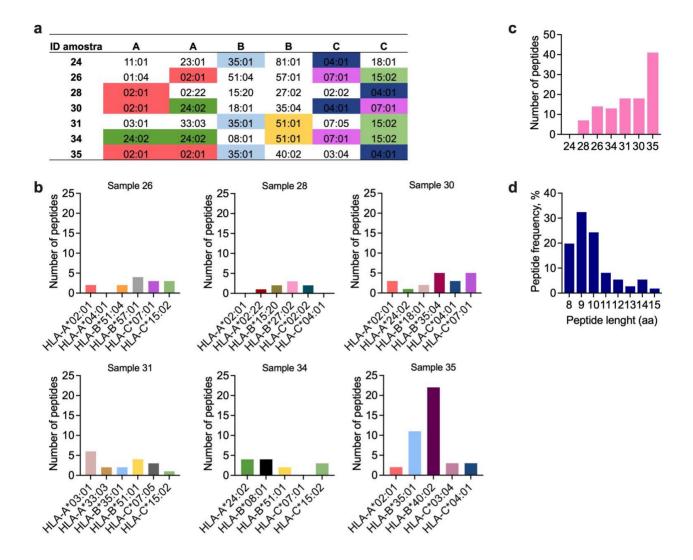




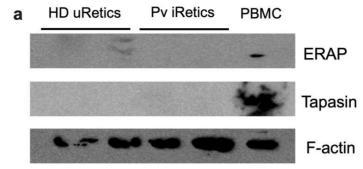


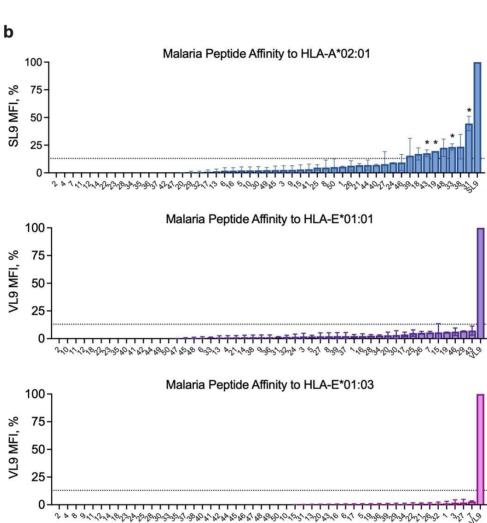


Extended data figure 3 | Peptide validation by ex vivo ELISpot assay. Selected peptides were tested using PBMC isolated from patients infected with Plasmodium vivax (Pv) (n = 24), P. falciparum (Pf) (n=7) and healthy donors from endemic (n = 15) and non-endemic (n = 6) regions for malaria. Cells were stimulated with a, 40S ribosomal protein peptides, b, 60S ribosomal protein peptides, c, ETRAMP peptides, d, Histone peptides, and e, other peptides. Each symbol represents one individual. Green squares are Pv patients, purple triangles are Pf patients, pink circles are endemic healthy donors, and white circles are non-endemic healthy donor. IFN $\gamma$  production was measured by spot counting and the results are expressed as spot-forming cells (SFC) by  $1\times10^6$  PBMC. Positive peptides were considered as responses that induced  $\geq 30$  spots in each patient- above the red dashed line.

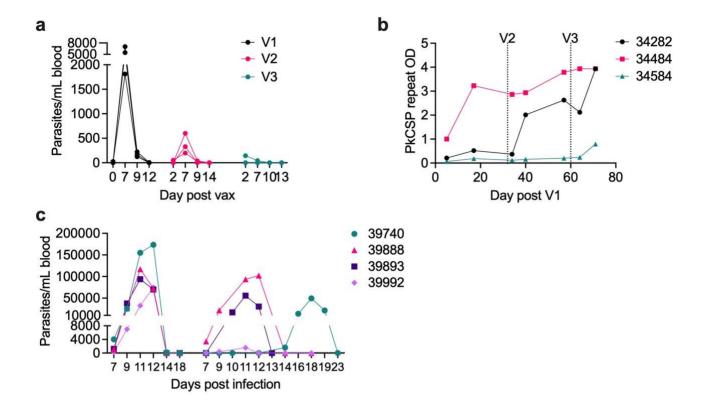


Extended data figure 4 | Plasmodium vivax peptides are presented by different HLAs. a, HLA-ABC genotyping typing of the 7 patients used in the immunopeptidome analysis. b-d, NetMHC bind analysis of peptides identified in each sample to their donor HLA type. b, Number of peptides identified as HLA-ABC binders for each sample. HLA-ABC genotyping of the donor of samples used in the immunopeptidomics assays. Alleles represented with same color mean that they were shared across different samples.

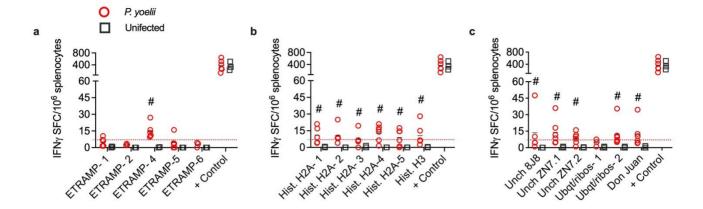




Extended data figure 5 | Tapasin and ERAP1 expression on reticulocytes. Western blot of cell lysates from two healthy donors (HD): uninfected Retics (uRetics) and PBMC; and three *Plasmodium vivax*-infected reticulocytes (Pv iRetics) from three patients were probed for the antigen processing protein Tapasin and Endoplasmic reticulum aminopeptidase 1 (ERAP1). PBMC was used as positive control. F-actin was loaded as endogen protein expression control. *In vitro* binding assay of all selected peptides pulsed in an HLA-A\*02:01, HLA-E\*01:01, and HLA-E\*01:03 cell line, including control peptides. Those peptides marked with asterisk were considered HLA-A: 0201 binders by *in silico* affinity predictions.



Extended data figure 6 Parasitemia during Plasmodium knowlesi (Pk) and P. cynomolgi (Pcy) infection of non-human primates. a, Parasitemia of Pk as measured by qRT-PCR following each immunization. b, Antibodies to Pk CSP repeat region throughout immunization as measured by single dilution ELISA OD value. c, Parasitemia of Pcy as measured by Giemsa-stained thin smears.



Extended data figure 7 Peptide validation by ex vivo IFN $\gamma$  ELISpot in mouse model. Immunogenicity in Plasmodium yoelii (Py) infected mice. Mice were intraperitoneally injected with  $10^5$  Py iRBC. At 12 days post-infection (dpi), mouse splenocytes were isolated and incubated with selected peptides. Cells were stimulated with **a**, ETRAMP peptides, **b**, Histone peptides and **c**, Miscellaneous peptides. Each symbol represents one individual. Red circles are Py infected mice (n=6), and black ones are non-infected mice (n=3). IFN $\gamma$  production was measured by spot counting, and the results are expressed as SFC by  $1\times10^6$  splenocytes. Positive peptides (#) were considered as responses that induced  $\geq 10$  spots in each mouse – above the red dashed line – and P  $\geq 0.05$  by multiple unpaired student t tests.

### **MAIN FIGURES**

- Figure 1 | Peptide identification and immunogenicity validation.
- Figure 2 | Classical and non-classical HLA peptide binders
- Figure 3 | Novel *Plasmodium* antigens are recognized by T cells elicited during *P. knowlesi* and *P. cynomolgi* infection of non-human primates.
- Figure 4 | Antigen validation in the *P. yoelii* malaria experimental model.

### **EXTENDED DATA TABLES**

Extended data table 1 | *Plasmodium falciparum* homology
Extended data table 2 | Primers used for *Plasmodium falciparum* liver stage expression

#### **EXTENDED DATA FIGURES**

Extended data figure 1   Antigen gene expression in humanized mouse livers infected with <i>P</i> .
falciparum.
Extended data figure 2   Spectral match validation
Extended data figure 3   Peptide validation by ex vivo ELISpot assay.
Extended data figure 4   Pv peptides are presented by different HLAs
Extended data figure 5   Tapasin and ERAP1 expression on reticulocytes
Extended data figure 6   Parasitemia during P. knowlesi and P. cynomolgi infection of non-human
primates.
Extended data figure 7   Peptide validation by <i>ex vivo</i> IFNy ELISpot in mouse model.

### **SUPPLEMENTARY INFORMATION (SI)**

Supplementary information 1	Clinical information
Supplementary information 2	1 1
Supplementary information 3	Plasmodium vivax orthologous gene expression in parasite life cycle.
Supplementary information 4	Selected peptides used for in vitro stimulation
Supplementary information 5	HLA binding