

Electro-induced C-H/S-H Cross-Coupling for the Functionalization/Macrocyclization of Cysteine-Containing Peptides

Fang Xiang,^a Jia Deng,^b Xiaotong Bu,^a Youjun Zhou,^a Jianrong Zhang,^a Yue Weng,^a and Meng Gao^{*a}

Supporting Information

TABLE OF CONTENTS

1. General Information	- 2 -
2. General Experimental Procedures.....	- 3 -
2.1 Synthesis of starting materials dipeptides.....	- 3 -
2.2. Reaction optimization.....	- 4 -
2.3 Gram-scale synthesis of 1aa at 5 mmol scale.....	- 5 -
3. Mechanistic and control experiments.	- 6 -
3.1 Control experiments.....	- 6 -
3.2 General procedure for cyclic voltammetry (CV)	- 8 -
4. Bioactive molecules scope and characterization.....	- 10 -
4.1 Dipeptides scope and characterization	- 10 -
4.2 Polypeptide scope and characterization.....	- 20 -

4.3 Enamine and azole scope and characterization	- 27 -
4.4 Electrochemical peptides functionalization	- 40 -
4.5 Electrochemical peptides cyclization	- 45 -
5. References	- 59 -
6. NMR spectra of all products.....	- 60 -
7. X-ray single-crystal data.....	- 138 -

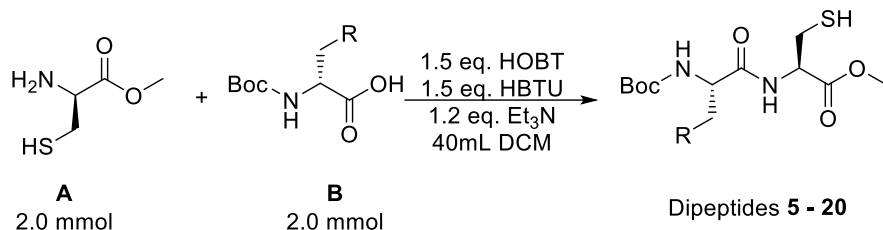
1. General Information

Unless otherwise stated, analytical grade solvents and commercially available reagents were used without further purification. All solvents were analytical reagent or better and were degassed prior to use. The instrument for electrolysis was dual display potentiostat (DJS-292B) (made in China). The anode electrode is platinum plate electrodes (Φ 6mm) and the cathode electrode is platinum plate electrodes (15 mm×15 mm×0.3 mm). Thin layer chromatography (TLC) employed glass 0.25 mm silica gel plates. Flash chromatography columns were packed with 200-300 mesh silica gel in petroleum (boiling point is between 60-90°C). Gradient flash chromatography was conducted eluting with a continuous gradient from petroleum to the indicated solvent, and they are listed as volume/volume ratios. High resolution mass spectra (HRMS) for polypeptides were measured with an Agilent 6224 instrument and accurate masses were reported for the molecular ion + Hydrogen (M+H) or molecular ion + Sodium (M+Na). The ^1H , ^{13}C and ^{19}F NMR spectra were recorded on a Bruker Advance III (400 MHz)

spectrometers with tetramethylsilane as an internal standard. All chemical shifts (δ) are reported in ppm and coupling constants (J) in Hz. For ^1H NMR, chemical shifts (δ) were given in ppm relatives to internal standard (TMS at 0 ppm, CDCl_3 at 7.26 ppm, $\text{MeOH-}d_4$ at 3.31 ppm, DMSO- d_6 at 2.50 ppm). For ^{13}C -NMR, chemical shifts (δ) were reported in ppm using solvent as internal standard (CDCl_3 at 77.00 ppm, $\text{MeOH-}d_4$ at 49.00 ppm, DMSO- d_6 at 39.50 ppm). GC-MS spectra were recorded on a Varian GC-MS 3900-2100T and Shimadzu GCMS-QP2010SE. All the polypeptide precursors used in this article were provided by the PeptiL and Pharmatech CO., LTD.

2. General Experimental Procedures

2.1 Synthesis of starting materials dipeptides^{[1][2]}



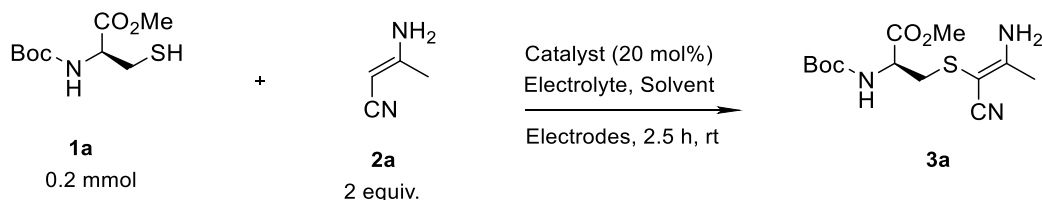
To a solution of L-Cysteine methyl ester hydrochloride **A** (343 mg, 2.0 mmol, 1.0 equiv.) in 40 mL CH_2Cl_2 was added HOBT (1-hydroxybenzotriazole) (3.0 mmol), HBTU (O-benzotriazole-*N,N,N'*, *N'*-tetramethyluronium-hexafluorophosphate) (3.0 mmol) and triethylamine (2.4 mmol). The mixture was stirred for 30 min at room temperature, and then, peptide **B** (2.0 mmol) was added to the solution. The reaction was stirred overnight. After regular workup, the reaction mixture washed by saturated NaHCO_3 solution (40 mL x 3), 2 M hydrochloric acid solution (40 mL x 3) and H_2O (40 mL x 3). The organic layers were combined, dried over Na_2SO_4 , and

concentrated. The resulting crude product was purified by flash chromatography (DCM / MeOH) to afford corresponding dipeptides **5-20**.

2.2. Reaction optimization

In an oven-dried undivided three-necked bottle (25 mL) equipped with a stir bar, cysteine residue (0.2 mmol), 3-Aminocrotononitrile(0.4 mmol), ⁿBu₄NBr (0.2 mmol) and NiBr₂•diglyme (0.04 mmol) were combined and added. Then, solvent (7 mL) were injected into the tubes via syringes. The bottle was equipped with platinum plate (15 mm×15 mm×0.3 mm) as the anode and platinum plate (15 mm×15 mm×0.3 mm) as the cathode. The reaction mixture was stirred and electrolysis at constant current under room temperature. When the reaction was finished, the solvent was removed by reduced pressure and the crude product was purified by flash column chromatography on silica gel (eluent: petroleum ether/ethyl acetate= 1:1). A summary of optimization results is presented in **Table S1** below.

Table S1. Investigation of the reaction conditions

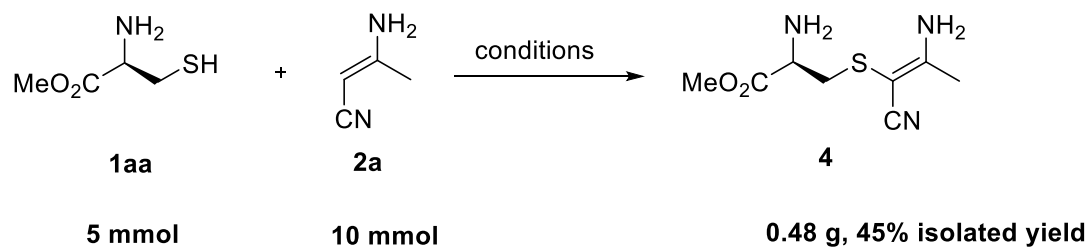


Entry	Electrodes	Electrolyte	Solvent	Catalyst	Halogen source	Current	Yield /%
1	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN	/	/	12 mA	trace
2	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN	/	MgCl ₂ •6 H ₂ O	12 mA	8
3	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN	/	NaCl	12 mA	14
4	Pt(+)/Pt(-)	ⁿ Bu ₄ NCl	CH ₃ CN	/	ⁿ Bu ₄ NCl	12 mA	17
5	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₂ Cl ₂	/	CH ₂ Cl ₂	12 mA	25
6	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	DCE	/	DCE	12 mA	12

7	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	/	CHCl ₃	12 mA	30
8	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHBr ₃ = 6/1	/	CHBr ₃	12 mA	27
9	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	Fe(Cp) ₂	CHCl ₃	12 mA	38
10	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	Mn(OTf) ₂	CHCl ₃	12 mA	57
11	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	Ni(acac) ₂	CHCl ₃	12 mA	42
12	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	Ni(cod) ₂	CHCl ₃	12 mA	47
13	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	12 mA	80
14	Pt(+)/Pt(-)	ⁿ Bu ₄ NI	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	12 mA	70
15	Pt(+)/Pt(-)	ⁿ Bu ₄ NBF ₄	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	12 mA	22
16	Pt(+)/Pt(-)	ⁿ Bu ₄ NPF ₆	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	12 mA	35
17	Pt(+)/Pt(-)	ⁿ Bu ₄ NCl	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	12 mA	68
18	C(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	12 mA	65
19	Pt(+)/Ni(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	12 mA	42
20	Pt(+)/Pb(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	12 mA	25
21	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	5 mA	36
22	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	8 mA	56
23	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	15 mA	72
24	Pt(+)/Pt(-)	ⁿ Bu ₄ NBr	CH ₃ CN/CHCl ₃ = 6/1	NiBr ₂ •diglyme	CHCl ₃	/	n.d.

^aReaction conditions: Undivided cell, Pt anode (15 mm×15 mm×0.3 mm), Pt cathode (15 mm×15 mm×0.3 mm), **1a** (0.2 mmol), **2a** (2 equiv.), ⁿBu₄NBr (1 equiv.), NiBr₂•diglyme (0.2 equiv.), 6 mL MeCN, 1 mL CHCl₃, air, rt, 12 mA, 2.5 h. Yields of isolated products are shown. N.D. = Not Detected.

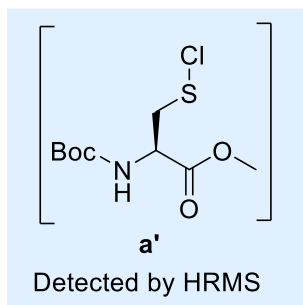
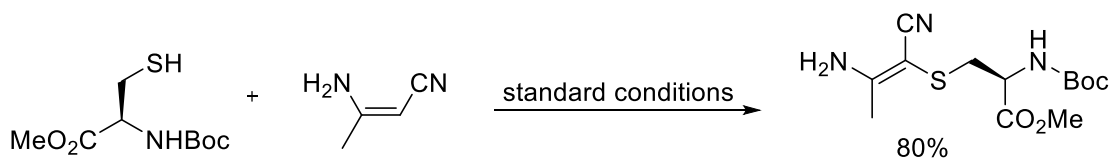
2.3 Gram-scale synthesis of **1aa** at 5 mmol scale



^bReaction conditions: Undivided cell, Pt anode(15 mm×15 mm×0.3 mm), Pt cathode(15 mm×15 mm×0.3 mm), **1aa** (5 mmol), **2a** (10 mmol), ⁿBu₄NBr (5 mmol), NiBr₂•diglyme (1 mmol), K₂CO₃ (5 mmol), 18 mL MeCN, 3 mL CHCl₃, air, rt, 12 mA, 12 h. ^bIsolated yields.

3. Mechanistic and control experiments.

3.1 Control experiments



Detected by HRMS of **a'** calcd for C₉H₁₆ClNO₄S [M+H]⁺ : 270.05613; Found: 270.05611

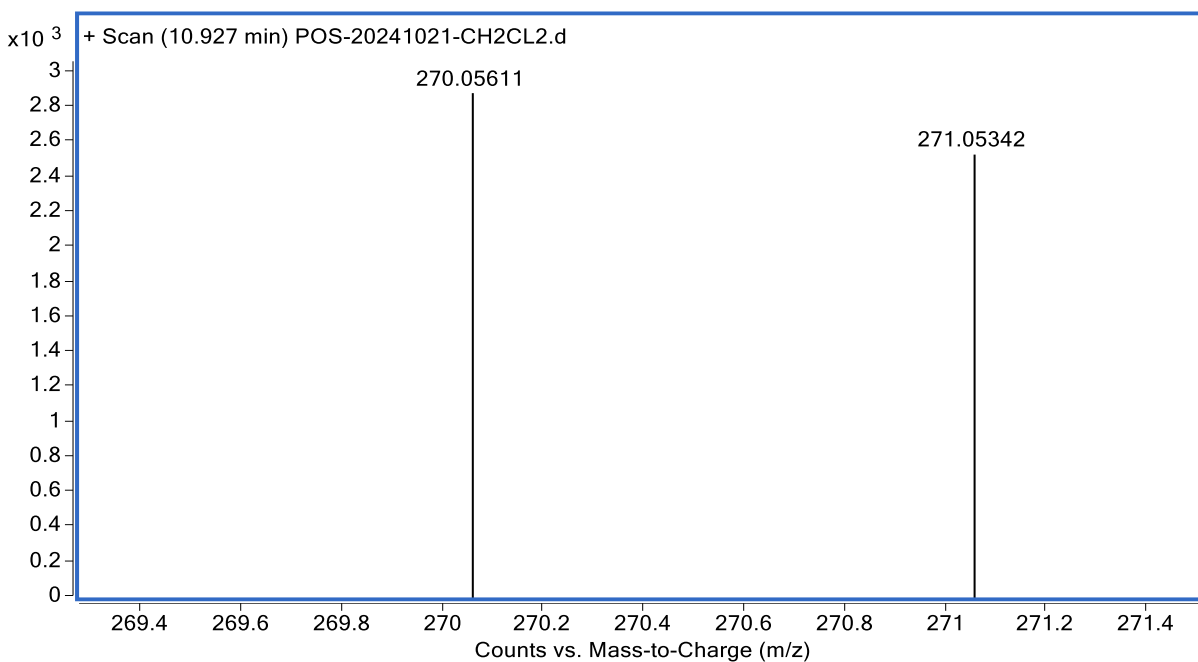
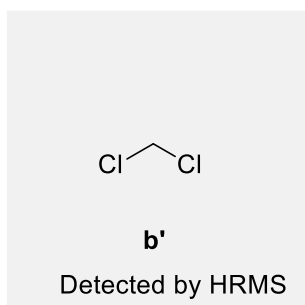


Figure S1. The HRMS spectra of compound **a'**



Detected by HRMS of **b'** calcd for CH₂Cl₂ [M+H]⁺ : 84.96063; Found: 84.96082

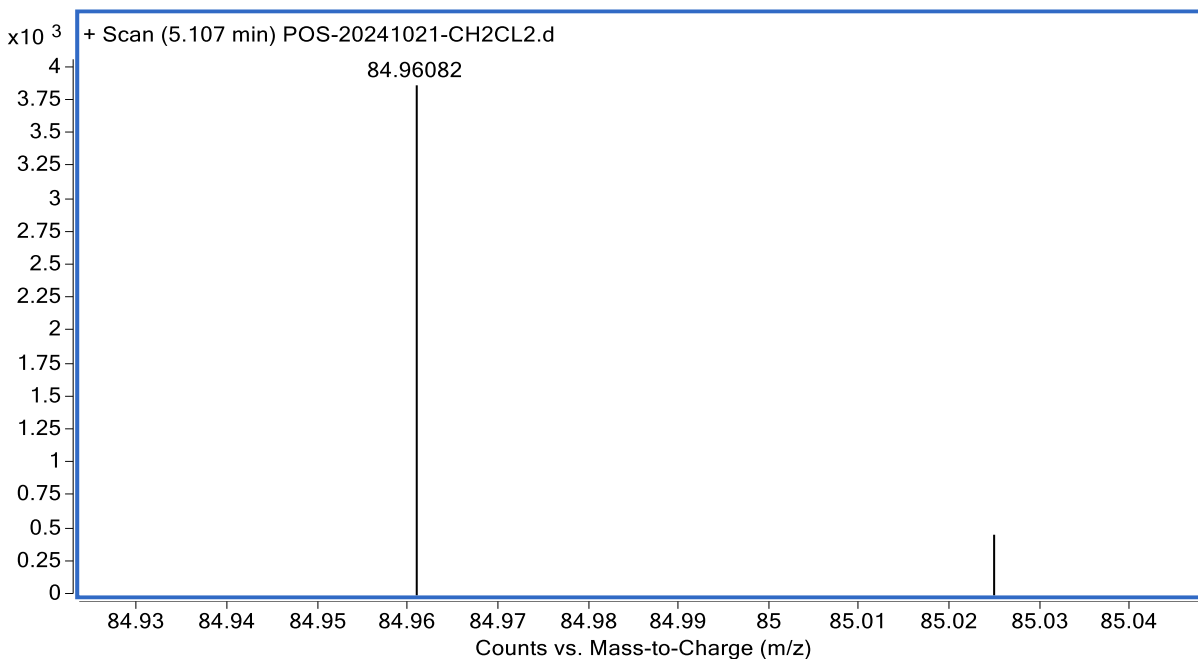
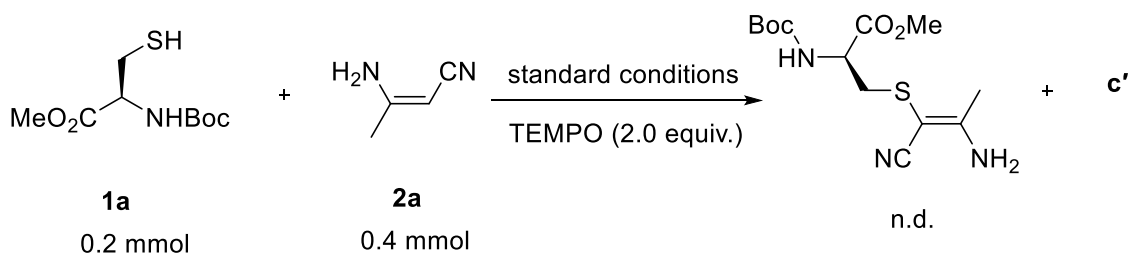
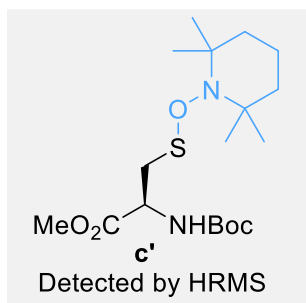


Figure S2. The HRMS spectra of compound **b'**



Reaction conditions: Undivided cell, Pt anode(15 mm × 15 mm × 0.3 mm), Pt cathode(15 mm × 15 mm × 0.3 mm), **1a** (0.2 mmol), **2a** (2 equiv.), ⁿBu₄NBr (1 equiv.), NiBr₂•diglyme (0.2 equiv.), TEMPO(2.0 equiv.), 6 mL MeCN, 1 mL CHCl₃, air, rt, 12 mA, 2.5 h.



Detected by HRMS of **c'** calcd for C₁₈H₃₄N₂O₅S [M+H]⁺ : 391.22612; Found: 391.22618

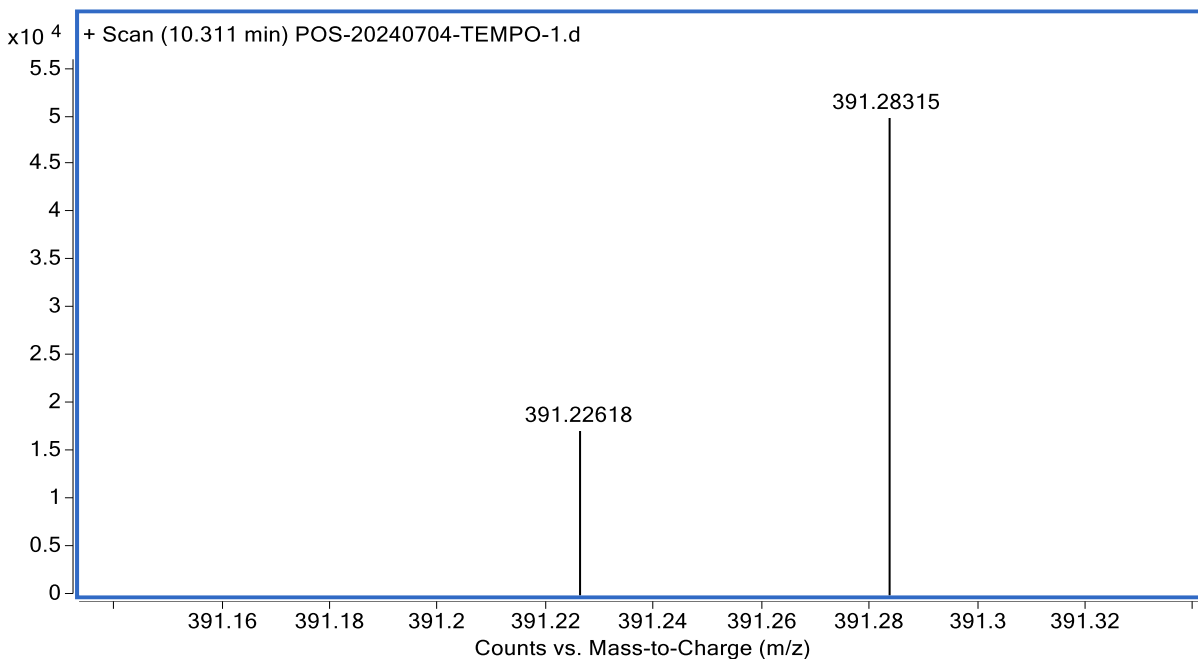


Figure S3. The HRMS spectra of compound **c'**

3.2 General procedure for cyclic voltammetry (CV)

Cyclic voltammetry was performed in a three-electrode cell connected to a schlenk line at room temperature. The working electrode was a steady glassy carbon disk electrode, the counter electrode was a platinum wire. The reference was an Ag/AgCl electrode submerged in saturated aqueous KCl solution and separated from a reaction by a salt bridge. 8 mL of CH₃CN containing 0.02 M ⁿBu₄NBF₄ were poured into the electrochemical cell in all experiments. The scan rate is 0.1 V/s. The positive scan range was from -2.0 V to 2.0 V and 0 V to 2.0 V.

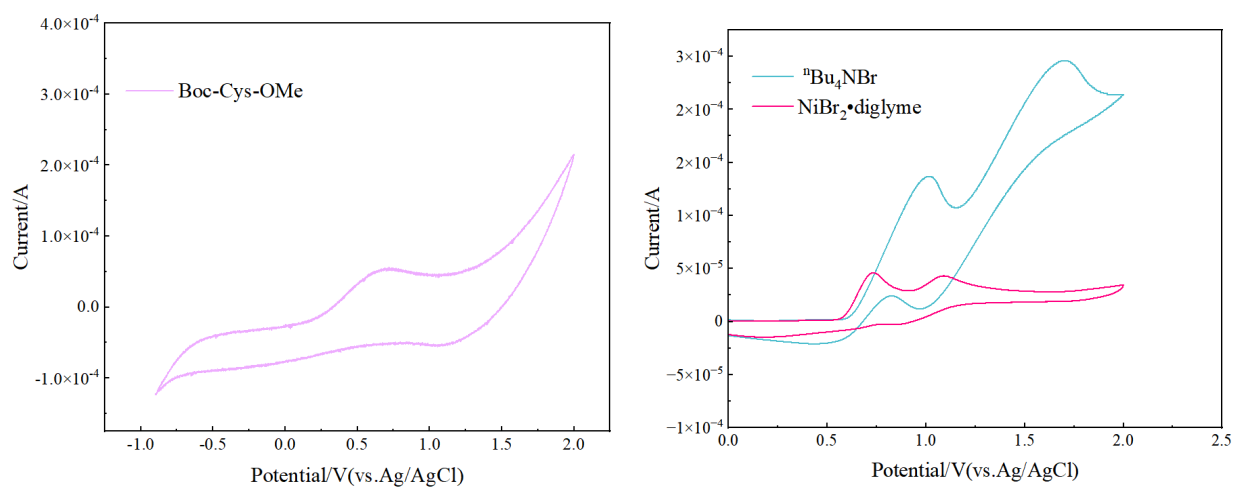


Figure S4. Cyclic voltammograms of substrate **Boc-Cys-OMe**, **$n\text{Bu}_4\text{NBr}$** , **$\text{NiBr}_2\cdot\text{diglyme}$**

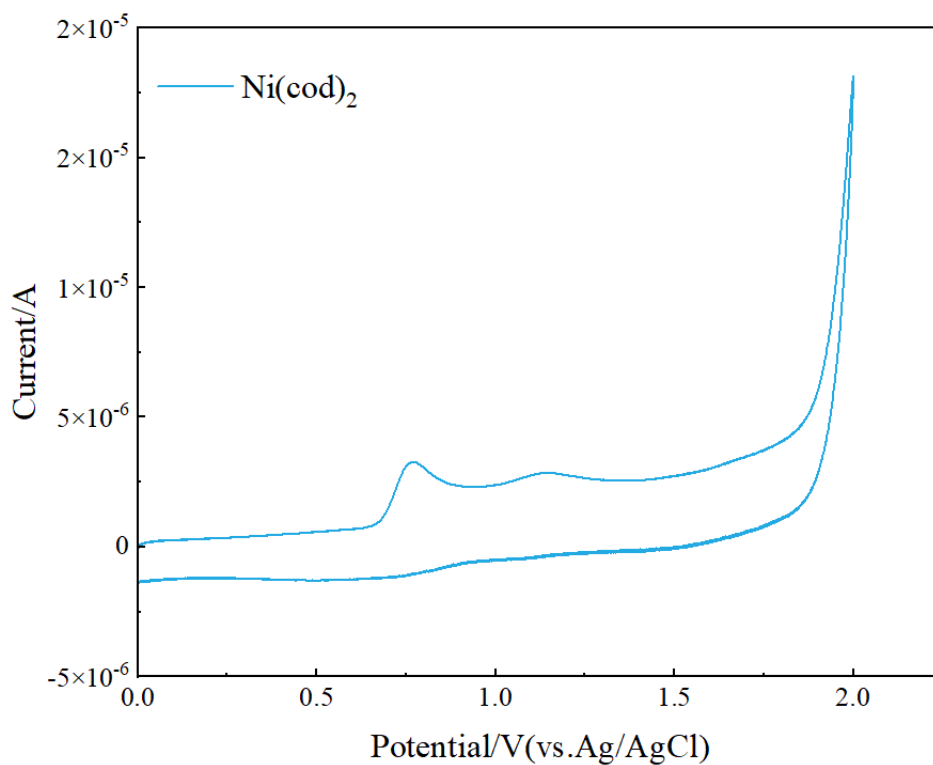
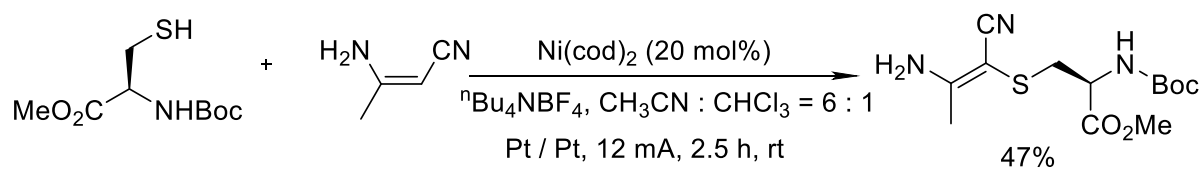
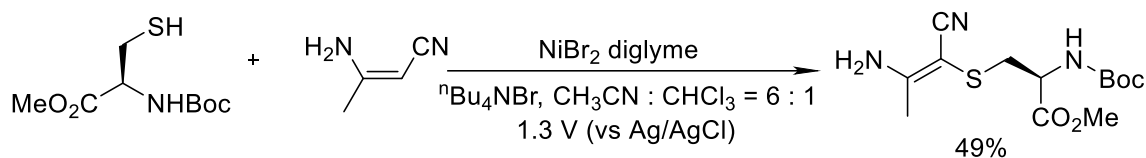


Figure S5. Cyclic voltammograms of substrate **Ni(cod)_2**

To avoid the interference of bromide ions in the CV experiment, we choose to use Ni(cod)_2

instead of $\text{NiBr}_2 \cdot \text{diglyme}$, and ${}^n\text{Bu}_4\text{NBF}_4$ instead of ${}^n\text{Bu}_4\text{NBr}$. The $\text{Ni}(\text{cod})_2$ displayed a dual anodic wave with a peak potential of 0.6V and 1.3V, which we attributed to the $\text{Ni}^0/\text{Ni}^{\text{I}}$ and $\text{Ni}^{\text{I}}/\text{Ni}^{\text{II}}$ redox. Thus, a controlled potential electrolysis was carried out. When the potential of the anode was maintained at 1.3 V, the corresponding product was obtained in 49% yield.

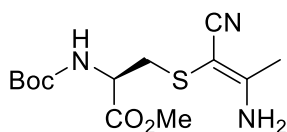


4. Bioactive molecules scope and characterization

4.1 Dipeptides scope and characterization

General procedure for bioconjugated product (3): In an oven-dried undivided three-necked bottle (25 mL) equipped with a stir bar, cysteine residue (0.2 mmol), 3-Aminocrotononitrile (0.4 mmol), ${}^n\text{Bu}_4\text{NBr}$ (0.2 mmol) and $\text{NiBr}_2 \cdot \text{diglyme}$ (0.04 mmol) were combined and added. Then, CH_3CN (6 mL), CHCl_3 (1 mL) were injected into the tubes via syringes. The bottle was equipped with platinum plate (15 mm×15 mm×0.3 mm) as the anode and platinum plate (15 mm×15 mm×0.3 mm) as the cathode. The reaction mixture was stirred and electrolysis at a constant current of 12 mA under room temperature for 2.5 h. After completion of the reaction, as indicated by TLC and LC-MS, the pure product (yield: 80%, 50.4 mg) was obtained by flash column chromatography on silica gel (eluent: petroleum ether/ethyl acetate= 1:1).

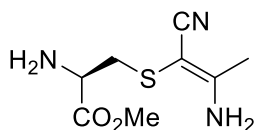
Detailed descriptions for products:



3

Methyl (Z)-S-(2-amino-1-cyanoprop-1-en-1-yl)-N-(tert-butoxycarbonyl)-L-cysteinate (3):

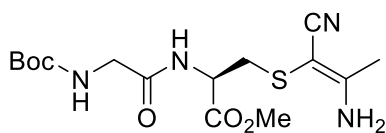
yellow solid (Yield: 80%, 50.41 mg), ^1H NMR (400 MHz, Chloroform-d) δ 5.36 (d, J = 8.3 Hz, 1H), 4.46 (d, J = 6.4 Hz, 1H), 3.75 (s, 3H), 3.12 – 3.03 (m, 1H), 2.92 (dd, J = 14.1, 6.7 Hz, 1H), 2.17 (s, 3H), 1.43 (s, 9H). ^{13}C NMR (151 MHz, CDCl_3) δ 171.27, 164.26, 155.36, 121.23, 80.38, 66.58, 53.66, 52.72, 37.98, 28.28, 20.26. HRMS (ESI) calcd. for $(\text{M}+\text{Na})^+$ $\text{C}_{13}\text{H}_{21}\text{N}_3\text{O}_4\text{S}$: 338.1145 found, 338.1141.



4

Methyl (Z)-S-(2-amino-1-cyanoprop-1-en-1-yl)-L-cysteinate (4): yellow solid (Yield: 45%,

0.48 g), ^1H NMR (400 MHz, Chloroform-d) δ 4.29 (dt, J = 7.9, 2.9 Hz, 1H), 3.83 (s, 3H), 3.12 (dd, J = 12.7, 3.2 Hz, 1H), 2.81 (dd, J = 12.7, 7.9 Hz, 1H), 2.20 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 169.80, 148.93, 119.00, 65.55, 55.23, 53.31, 25.62, 21.24. HRMS (ESI) calcd. for $(\text{M}+\text{H})^+$ $\text{C}_8\text{H}_{13}\text{N}_3\text{O}_2\text{S}$: 216.0801 found, 216.0803.

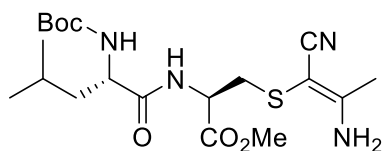


5

Methyl (Z)-S-(2-amino-1-cyanoprop-1-en-1-yl)-N-((tert-butoxycarbonyl)glycyl)-L-cysteinate (5): brown oil (Yield: 70%, 52.10 mg), ^1H NMR (400 MHz, Chloroform-d) δ 7.33 (d, J = 7.8

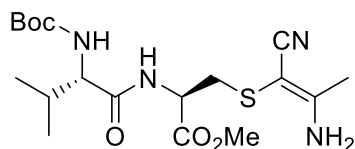
Hz, 1H), 5.72 (s, 1H), 4.84 – 4.56 (m, 1H), 3.92 – 3.80 (m, 2H), 3.77 (s, 3H), 3.16 – 2.94 (m, 2H), 2.18 (s, 3H), 1.46 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 170.82, 170.31, 165.02, 156.40,

121.67, 80.49, 65.36, 52.86, 52.02, 44.32, 37.18, 28.33, 20.23. HRMS (ESI) cald. for (M+Na)⁺ C₁₅H₂₄N₄O₅S: 395.1359 found, 395.1359.



6

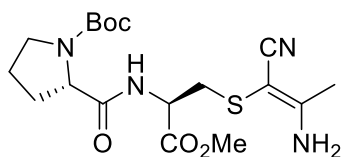
Methyl S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-N-((tert-butoxycarbonyl)-L-leucyl)-L-cysteinate (6): yellow solid (Yield: 45%, 38.53 mg), ¹H NMR (400 MHz, Chloroform-d) δ 7.10 (d, *J* = 7.6 Hz, 1H), 5.30 – 5.10 (m, 1H), 4.81 – 4.63 (m, 1H), 4.15 – 4.06 (m, 1H), 3.73 (s, 3H), 3.05 – 2.93 (m, 2H), 2.13 (s, 3H), 1.69 – 1.59 (m, 2H), 1.52 – 1.46 (m, 1H), 1.41 (s, 9H), 0.90 (t, *J* = 6.4 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 172.68, 170.60, 164.21, 155.88, 121.14, 80.40, 66.89, 52.77, 52.14, 40.64, 37.23, 28.36, 24.77, 22.93, 21.84, 20.37. HRMS (ESI) cald. for (M+Na)⁺ C₁₉H₃₂N₄O₅S: 451.1985 found, 451.1950.



7

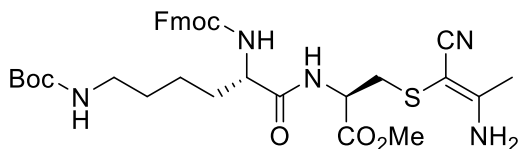
Methyl S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-N-((tert-butoxycarbonyl)-L-valyl)-L-cysteinate (7): yellow solid (Yield: 40%, 33.14 mg), ¹H NMR (400 MHz, Chloroform-d) δ 7.04 (d, *J* = 7.6 Hz, 1H), 5.27 (q, *J* = 3.7, 2.9 Hz, 1H), 4.68 (q, *J* = 6.2 Hz, 1H), 3.94 (dd, *J* = 8.8, 6.4 Hz, 1H), 3.74 (d, *J* = 3.5 Hz, 3H), 3.04 (dt, *J* = 18.5, 8.4 Hz, 2H), 2.14 (s, 3H), 1.41 (s, 9H), 1.00 – 0.89 (m, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 172.15, 170.68, 164.73, 156.12, 121.46, 80.16, 66.02,

60.18, 52.83, 52.17, 37.23, 30.49, 28.36, 20.30, 19.31, 17.91. HRMS (ESI) calcd. for (M+H)⁺ C₁₈H₃₀N₄O₅S: 415.2009 found, 415.2009.



8

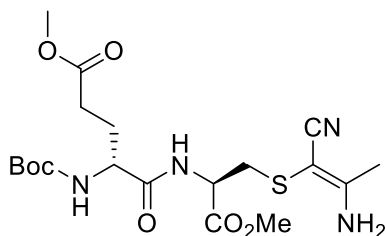
Tert-butyl (S)-2-(((R)-3-(((Z)-2-amino-1-cyanoprop-1-en-1-yl)thio)-1-methoxy-1-oxoprop-2-yl)carbamoyl)pyrrolidine-1-carboxylate (8): brown oil (Yield: 70%, 57.70 mg), ¹H NMR (400 MHz, Chloroform-d) δ 4.79 – 4.49 (m, 1H), 4.25 (d, *J* = 7.1 Hz, 1H), 3.70 (s, 3H), 3.49 – 3.25 (m, 2H), 2.96 (qd, *J* = 14.1, 11.9, 6.6 Hz, 2H), 2.31 – 2.15 (m, 1H), 2.12 (s, 3H), 2.04 – 1.66 (m, 3H), 1.40 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 172.54, 170.70, 164.93, 155.63, 121.38, 80.56, 59.90, 53.50, 52.66, 51.70, 47.18, 37.17, 28.41, 24.50, 20.32. HRMS (ESI) calcd. for (M+H)⁺ C₁₈H₂₈N₄O₅S: 413.1853 found, 413.1854.



9

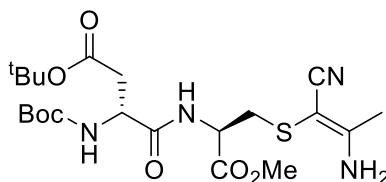
Methyl N-(N2-(((9H-fluoren-9-yl)methoxy)carbonyl)-N6-(tert-butoxycarbonyl)-L-lysyl)-S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-L-cysteinate(9): yellow solid (Yield: 42%, 55.88 mg), ¹H NMR (400 MHz, Chloroform-d) δ 7.75 (d, *J* = 7.5 Hz, 2H), 7.59 (d, *J* = 7.5 Hz, 2H), 7.41 – 7.29 (m, 2H), 7.22 – 7.04 (m, 1H), 5.81 (d, *J* = 8.0 Hz, 2H), 4.74 (dq, *J* = 12.9, 5.7 Hz, 2H), 4.38 (d, *J* = 7.1 Hz, 2H), 4.21 (t, *J* = 7.0 Hz, 2H), 3.73 (s, 3H), 3.18 – 2.98 (m, 4H), 2.12 (s, 3H), 1.96 – 1.87 (m, 1H), 1.74 – 1.65 (m, 1H), 1.50 (q, *J* = 7.5, 5.9 Hz, 4H), 1.43 (s, 9H). ¹³C

NMR (101 MHz, CDCl₃) δ 172.09, 170.59, 164.57, 156.35, 143.75, 141.29, 127.78, 127.13, 125.12, 121.41, 120.01, 79.32, 67.26, 66.28, 54.88, 52.77, 52.15, 50.64, 47.11, 39.92, 37.05, 31.64, 29.50, 28.45, 22.45, 20.20. HRMS (ESI) cald. for (M+H)⁺ C₃₄H₄₃N₅O₇S: 666.2956 found,666.2956.



10

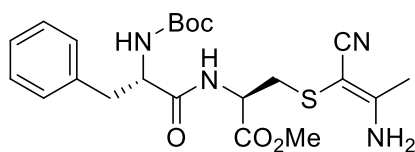
Methyl (R)-5-(((R)-3-(((Z)-2-amino-1-cyanoprop-1-en-1-yl)thio)-1-methoxy-1-oxopropan-2-yl)amino)-4-((tert-butoxycarbonyl)amino)-5-oxopentanoate (10): yellow solid (Yield: 71%, 65.06 mg), ¹H NMR (400 MHz, Chloroform-d) δ 7.45 – 7.00 (m, 1H), 5.63 (dd, *J* = 55.0, 9.0 Hz, 1H), 4.91 – 4.64 (m, 1H), 4.37 – 4.12 (m, 1H), 3.77 (s, 3H), 3.69 (s, 3H), 3.22 – 2.96 (m, 2H), 2.49 (q, *J* = 6.8 Hz, 2H), 2.18 (s, 3H), 1.96 (dt, *J* = 14.5, 7.6 Hz, 2H), 1.45 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 173.98, 171.90, 170.61, 164.62, 155.88, 121.38, 80.39, 66.19, 53.75, 52.87, 52.00, 37.06, 30.19, 28.35, 27.51, 20.35. HRMS (ESI) cald. for (M+Na)⁺ C₁₉H₃₀N₄O₇S: 481.1727 found,481.1724.



11

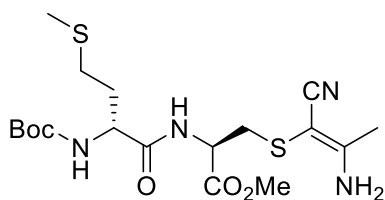
Tert-butyl (R)-4-(((R)-3-(((Z)-2-amino-1-cyanoprop-1-en-1-yl)thio)-1-methoxy-1-oxopropa

n-2-yl)amino)-3-((tert-butoxycarbonyl)amino)-4-oxobutanoate (11): light yellow oil (Yield: 61%, 59.31 mg), ^1H NMR (400 MHz, Chloroform- d) δ 7.40 (d, J = 7.8 Hz, 1H), 5.92 (d, J = 8.5 Hz, 1H), 4.67 (t, J = 6.6 Hz, 1H), 4.58 – 4.44 (m, 1H), 3.76 (s, 3H), 3.07 (td, J = 12.2, 10.3, 5.5 Hz, 2H), 2.83 – 2.68 (m, 2H), 2.18 (s, 3H), 1.47 (s, 9H), 1.45 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.30, 170.74, 170.46, 164.93, 155.68, 121.46, 81.71, 80.49, 65.33, 53.52, 52.72, 52.09, 51.01, 37.05, 28.30, 27.97, 20.16. HRMS (ESI) cald. for $(\text{M}+\text{Na})^+$ $\text{C}_{21}\text{H}_{34}\text{N}_4\text{O}_7\text{S}$: 509.2040, found, 509.2015.



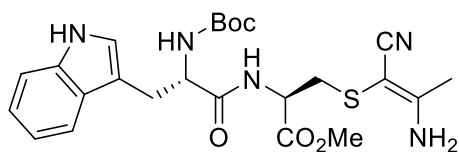
12

Methyl S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-N-((tert-butoxycarbonyl)-L-phenylalanyl)-L-cysteinate (12): light yellow solid (Yield: 75%, 69.33 mg), ^1H NMR (400 MHz, Chloroform- d) δ 7.35 – 7.19 (m, 5H), 6.89 (d, J = 7.8 Hz, 1H), 5.14 (d, J = 7.5 Hz, 1H), 4.61 (s, 1H), 4.37 (q, J = 7.2 Hz, 1H), 3.74 (s, 3H), 3.15 (dd, J = 14.2, 6.3 Hz, 1H), 3.04 (p, J = 9.8, 7.8 Hz, 2H), 2.93 – 2.72 (m, 1H), 2.16 (s, 3H), 1.40 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 172.03, 170.81, 164.88, 155.74, 136.32, 129.29, 128.82, 127.16, 121.26, 80.77, 65.11, 55.94, 52.83, 51.73, 50.63, 37.78, 36.95, 28.31, 20.40. HRMS (ESI) cald. for $(\text{M}+\text{H})^+$ $\text{C}_{22}\text{H}_{30}\text{N}_4\text{O}_5\text{S}$: 463.2009, found, 463.2007.



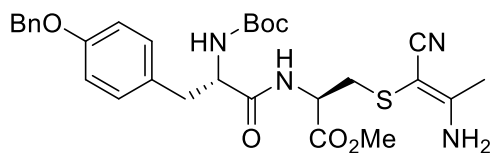
13

Methyl S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-N-((tert-butoxycarbonyl)-D-methionyl)-L-cysteinate(13): light yellow solid (Yield: 73%, 65.14 mg), ^1H NMR (400 MHz, Chloroform-d) δ 7.13 (d, J = 7.7 Hz, 1H), 5.45 (d, J = 8.2 Hz, 1H), 4.88 – 4.56 (m, 1H), 4.30 (t, J = 7.4 Hz, 1H), 3.77 (d, J = 5.7 Hz, 3H), 3.18 – 3.00 (m, 2H), 2.60 (t, J = 7.2 Hz, 2H), 2.18 (s, 3H), 2.11 (d, J = 3.4 Hz, 4H), 1.96 (dd, J = 14.3, 7.2 Hz, 1H), 1.45 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 172.00, 170.61, 164.56, 155.86, 121.36, 80.53, 66.57, 53.70, 52.89, 52.34, 37.27, 31.37, 30.33, 28.45, 20.41, 15.39. HRMS (ESI) calcd. for $(\text{M}+\text{Na})^+$ $\text{C}_{18}\text{H}_{30}\text{N}_4\text{O}_5\text{S}_2$: 469.1549, found, 469.1545.



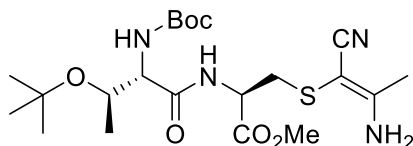
14

Methyl S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-N-((tert-butoxycarbonyl)-L-tryptophyl)-L-cysteinate (14): light yellow solid (Yield: 40%, 40.10 mg), ^1H NMR (400 MHz, Chloroform-d) δ 8.51 (s, 1H), 7.64 (d, J = 8.0 Hz, 1H), 7.36 (d, J = 8.1 Hz, 1H), 7.22 – 7.05 (m, 4H), 6.77 (d, J = 7.8 Hz, 1H), 4.74 – 4.64 (m, 1H), 4.46 (q, J = 7.0 Hz, 1H), 3.69 (s, 3H), 3.34 – 3.19 (m, 2H), 2.92 (dd, J = 13.5, 5.9 Hz, 2H), 2.00 (s, 3H), 1.42 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 172.06, 170.55, 164.51, 155.73, 136.33, 127.56, 123.69, 122.22, 121.48, 119.72, 118.74, 111.52, 110.17, 80.44, 66.27, 55.43, 52.89, 52.36, 37.46, 28.41, 28.02, 20.20. HRMS (ESI) calcd. for $(\text{M}+\text{Na})^+$ $\text{C}_{24}\text{H}_{31}\text{N}_5\text{O}_5\text{S}$: 524.1938, found, 524.1931.



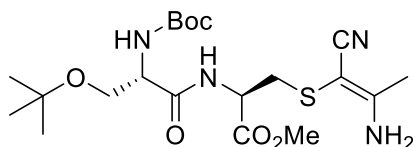
15

Methyl S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-N-((S)-3-(4-(benzyloxy)phenyl)-2-((tert-butoxycarbonyl)amino)propanoyl)-L-cysteinate (15): light yellow solid (Yield: 47%, 53.41 mg), ^1H NMR (400 MHz, Chloroform- d) δ 7.42 – 7.27 (m, 5H), 7.12 (d, J = 8.5 Hz, 2H), 7.02 (s, 1H), 6.93 – 6.87 (m, 2H), 5.28 – 5.23 (m, 1H), 5.02 (s, 2H), 4.68 (s, 1H), 4.34 (d, J = 7.2 Hz, 1H), 3.73 (s, 3H), 3.13 – 2.91 (m, 4H), 2.13 (s, 3H), 1.39 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 172.00, 170.51, 164.89, 157.80, 155.74, 136.95, 130.39, 128.76, 128.62, 128.02, 127.51, 121.80, 114.99, 80.41, 69.98, 65.93, 56.01, 52.88, 52.36, 37.63, 36.92, 28.33, 20.31. HRMS (ESI) calcd. for $(\text{M}+\text{H})^+$ $\text{C}_{29}\text{H}_{36}\text{N}_4\text{O}_6\text{S}$: 569.2428, found, 569.2428.



16

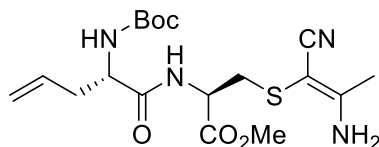
Methyl S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-N-(N-(tert-butoxycarbonyl)-O-(tert-butyl)-L-threonyl)-L-cysteinate (16): light yellow solid (Yield: 55%, 51.95 mg), ^1H NMR (400 MHz, Chloroform- d) δ 7.97 – 7.77 (m, 1H), 5.60 – 5.52 (m, 1H), 4.71 (d, J = 6.5 Hz, 1H), 4.22 – 4.09 (m, 2H), 3.77 (s, 3H), 3.09 (q, J = 6.4, 4.8 Hz, 2H), 2.18 (s, 3H), 1.46 (s, 9H), 1.31 (s, 9H), 1.12 (d, J = 6.1 Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 170.26, 164.17, 155.52, 120.92, 79.77, 75.65, 67.22, 66.32, 58.32, 52.59, 52.10, 37.19, 28.35, 20.26, 16.97. HRMS (ESI) calcd. for $(\text{M}+\text{Na})^+$ $\text{C}_{21}\text{H}_{36}\text{N}_4\text{O}_6\text{S}$: 495.2247, found, 495.2200.



17

Methyl S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-N-(N-(tert-butoxycarbonyl)-O-(tert-butyl)

-L-seryl)-L-cysteinate(17): light yellow solid (Yield: 40%, 36.66 mg), ^1H NMR (400 MHz, Chloroform-d) δ 7.45 (d, J = 8.3 Hz, 1H), 5.50 – 5.39 (m, 1H), 4.87 – 4.64 (m, 1H), 4.31 – 4.10 (m, 1H), 3.80 (d, J = 4.9 Hz, 1H), 3.77 (s, 3H), 3.46 (t, J = 7.8 Hz, 1H), 3.05 (ddd, J = 52.3, 14.1, 5.6 Hz, 2H), 2.19 (s, 3H), 1.47 (s, 9H), 1.22 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.05, 170.50, 164.41, 155.61, 121.16, 80.32, 74.21, 66.48, 61.62, 54.50, 52.76, 52.36, 37.94, 28.31, 27.38, 20.35. ^{13}C NMR (101 MHz, Chloroform-d) δ 165.89, 165.75, 165.45, 147.78, 147.58, 140.18, 132.22, 130.46, 130.16, 125.80, 123.83, 123.02, 75.96, 68.47, 68.39, 53.49, 53.35, 27.95, 25.70. HRMS (ESI) cald. for $(\text{M}+\text{H})^+$ $\text{C}_{20}\text{H}_{34}\text{N}_4\text{O}_6\text{S}$: 459.2271, found, 459.2277.



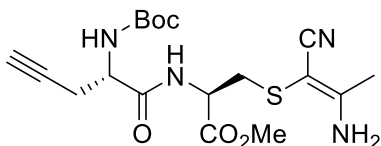
18

Methyl S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-N-((S)-2-((tert-butoxycarbonyl)amino)pent-

4-enoyl)-L-cysteinate (18): yellow solid (Yield: 75%, 61.83 mg), ^1H NMR (400 MHz, Chloroform-d) δ 7.11 (d, J = 7.7 Hz, 1H), 5.84 – 5.73 (m, 1H), 5.26 – 5.11 (m, 3H), 4.83 – 4.64 (m, 1H), 4.20 (q, J = 7.1 Hz, 1H), 3.77 (s, 3H), 3.06 (tt, J = 14.2, 8.0 Hz, 2H), 2.53 (dp, J = 29.1, 7.5 Hz, 2H), 2.18 (s, 3H), 1.45 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.84, 170.52, 164.51,

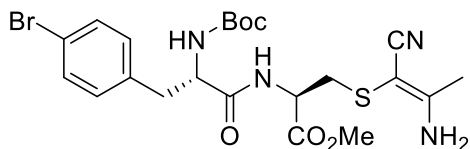
155.73, 132.92, 121.33, 119.09, 80.44, 66.42, 54.08, 52.75, 52.26, 37.52, 36.26, 28.32, 20.29.

HRMS (ESI) cald. for (M+K)⁺ C₁₈H₂₈N₄O₅S: 451.1412, found, 451.1412.



19

Methyl S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-N-((S)-2-((tert-butoxycarbonyl)amino)pent-4-ynoyl)-L-cysteinate (19): light yellow solid (Yield: 53%, 43.48 mg), ¹H NMR (400 MHz, Chloroform-d) δ 7.24 (d, *J* = 7.7 Hz, 1H), 5.55 – 5.46 (m, 1H), 4.79 – 4.68 (m, 1H), 4.40 – 4.28 (m, 1H), 3.78 (s, 3H), 3.07 (tt, *J* = 14.2, 8.0 Hz, 2H), 2.73 (qdd, *J* = 16.6, 6.2, 2.5 Hz, 2H), 2.19 (s, 3H), 2.13 (s, 1H), 1.47 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 170.59, 164.49, 155.58, 121.30, 80.77, 79.26, 71.82, 66.39, 53.43, 52.81, 52.37, 37.44, 28.30, 22.04, 20.28. HRMS (ESI) cald. for (M+H)⁺ C₁₈H₂₆N₄O₅S: 411.1696, found, 411.1696.

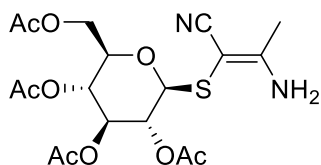


20

Methyl S-((Z)-2-amino-1-cyanoprop-1-en-1-yl)-N-((S)-3-(4-bromophenyl)-2-((tert-butoxycarbonyl)amino)propanoyl)-L-cysteinate (20): light yellow solid (Yield: 53%, 57.25 mg), ¹H NMR (400 MHz, Chloroform-d) δ 7.47 – 7.43 (m, 2H), 7.11 (d, *J* = 8.2 Hz, 2H), 6.83 (d, *J* = 7.7 Hz, 1H), 4.95 (d, *J* = 7.3 Hz, 1H), 4.60 (s, 1H), 4.34 (q, *J* = 7.0 Hz, 1H), 3.77 (s, 3H), 3.12 (dd, *J* = 14.0, 6.0 Hz, 2H), 3.02 (dd, *J* = 14.1, 7.3 Hz, 1H), 2.91 – 2.76 (m, 1H), 2.18 (s, 3H), 1.41 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 171.69, 170.70, 164.66, 155.68, 135.38, 131.80, 131.08,

120.99, 80.89, 65.58, 55.69, 52.87, 51.85, 37.07, 28.29, 20.42. HRMS (ESI) cald. for (M+H)⁺

C₂₂H₂₉BrN₄O₅S: 541.1114,found, 541.1113.



21

(2R,3R,4S,5R,6S)-2-(acetoxymethyl)-6-(((Z)-2-amino-1-cyanoprop-1-en-1-yl)thio)tetrahydr

o-2H-pyran-3,4,5-triyl triacetate(21): yellow solid (Yield: 35%, 31.08 mg), ¹H NMR (400

MHz, Chloroform-d) δ 5.21 (t, *J* = 9.3 Hz, 1H), 5.07 (t, *J* = 9.7 Hz, 1H), 4.99 – 4.93 (m, 1H),

4.57 (d, *J* = 10.1 Hz, 1H), 4.20 – 4.15 (m, 2H), 3.70 – 3.65 (m, 1H), 2.25 (s, 3H), 2.11 (s, 3H),

2.09 (s, 3H), 2.04 (s, 3H), 2.01 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.66, 170.18, 169.62,

169.43, 166.66, 121.17, 87.23, 76.13, 73.82, 69.66, 68.01, 63.62, 61.79, 20.81, 20.80, 20.65,

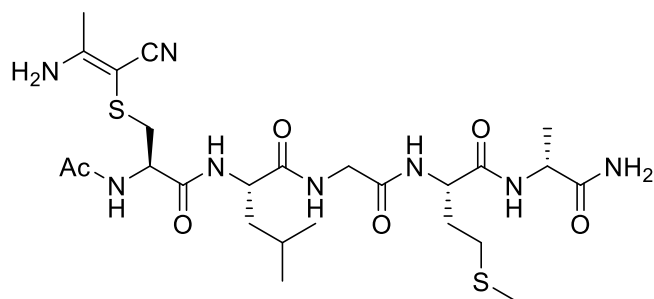
20.61, 20.60. HRMS (ESI) cald. for (M+H)⁺ C₁₈H₂₄N₂O₉S: 445.1275,found, 445.1241.

4.2 Polypeptide scope and characterization

The procedure for the bioconjugated product (22): In an oven-dried undivided three-necked bottle (15 mL) equipped with a stir bar, polypeptides (0.01 mmol), 3-Aminocrotononitrile (2 equiv.), ⁿBu₄NBr (2 equiv.), NiBr₂•diglyme (20 mol%), MeCN (6 mL), CHCl₃ (1 mL) were combined and added. The bottle was equipped platinum plate (10 mm×10 mm×0.3 mm) as the anode and platinum plate (10 mm×10 mm×0.3 mm) as the cathode. The reaction mixture was stirred and electrolysis at constant current of 4 mA under room temperature for 15 min. After completion of the reaction, the solution was analyzed by TOF-LC/MS spectroscopy. The reaction was analyzed by reversed-phase high-performance liquid chromatography (HPLC) on

an Agilent Zorbax SB-Aq 5 μ m column with a length of 250 mm. A binary high-pressure gradient mode was employed with a total flow rate of 0.5 mL/min, and the concentration of buffer solution B was 100%. The elution time was 20 minutes. HPLC analysis used buffers A (water) and B (acetonitrile + 0.1% TFA). Conversion reported as a % conversion as determined.

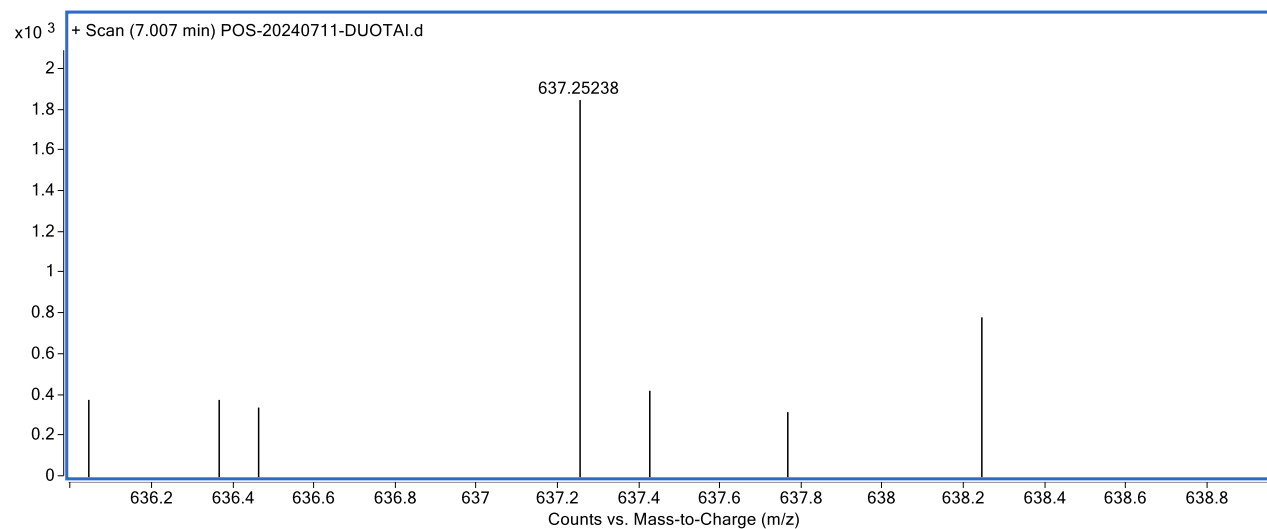
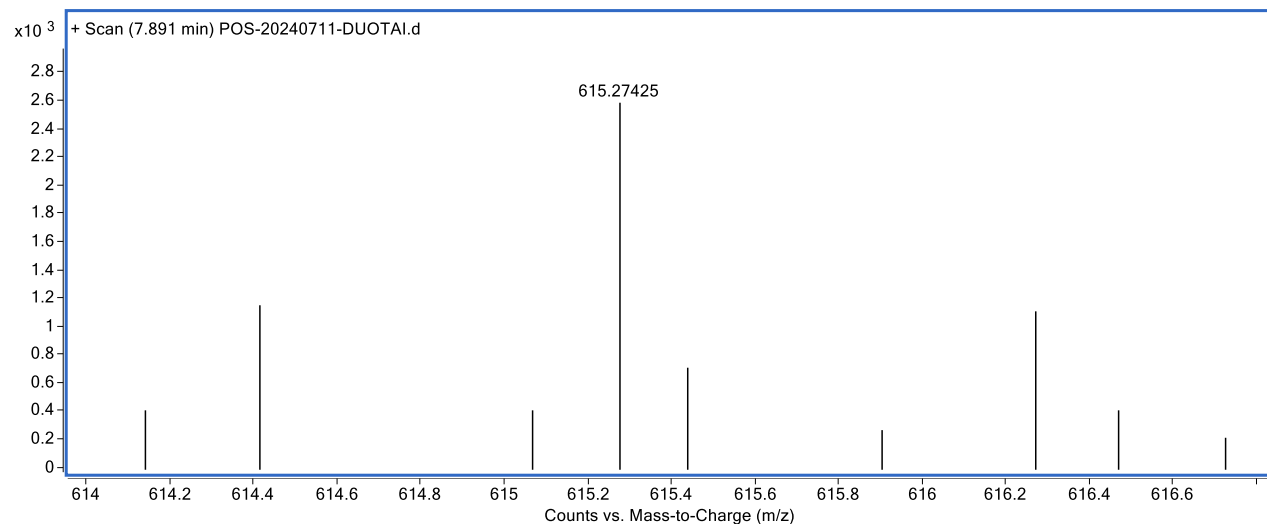
bioconjugated product 22 :



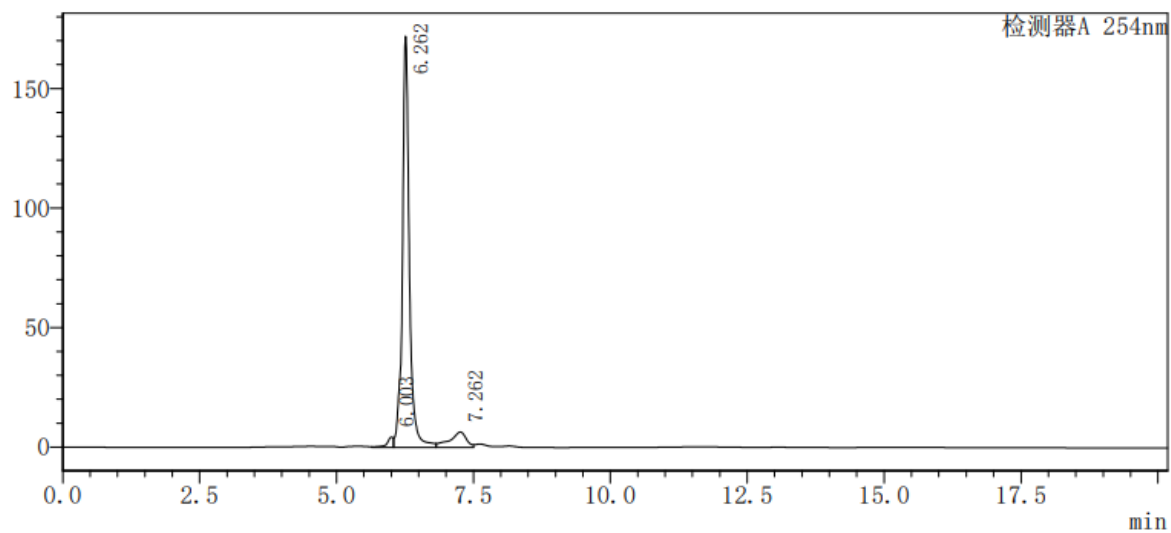
HPLC: >99% conversion.

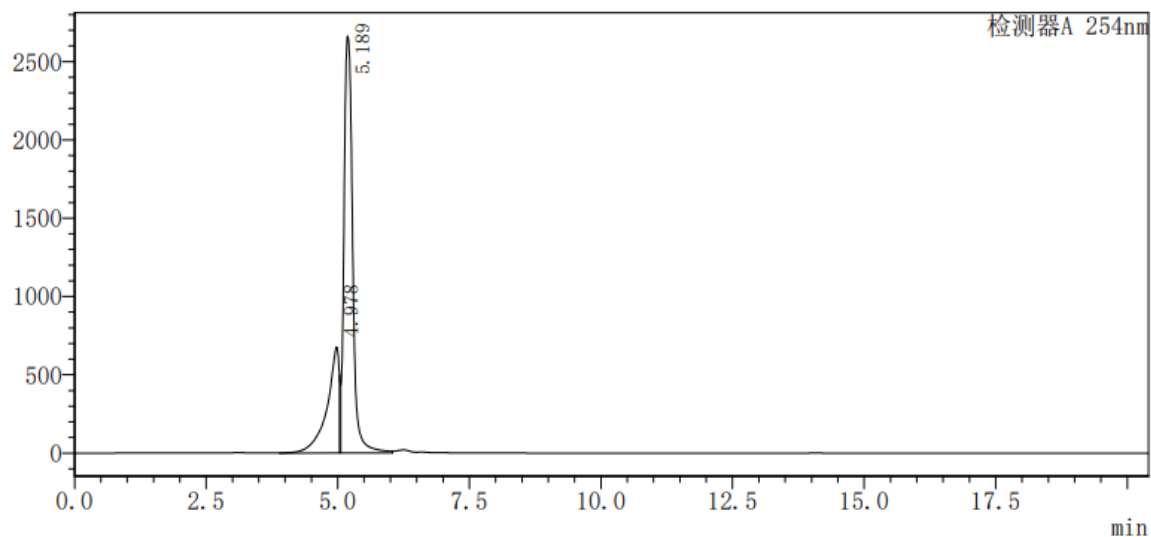
After the reaction finished, there is a peak that elutes at 100% buffer B (acetonitrile + 0.1% TFA) with a retention time of 5.189 min. Polypeptide is a peak that elutes at 100% buffer B (acetonitrile + 0.1% TFA) with a retention time of 6.262 min.

HRMS (ESI-TOF)calcd for $C_{25}H_{42}N_8O_6S_2$, $[M+H]^+$, 615.27415, found 615.27425.calcd for $C_{25}H_{42}N_8O_6S_2$, $[M+Na]^+$, 637.25609, found 637.25238.

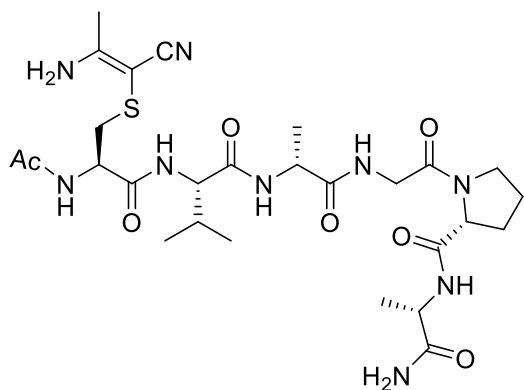


HPLC Spectra:





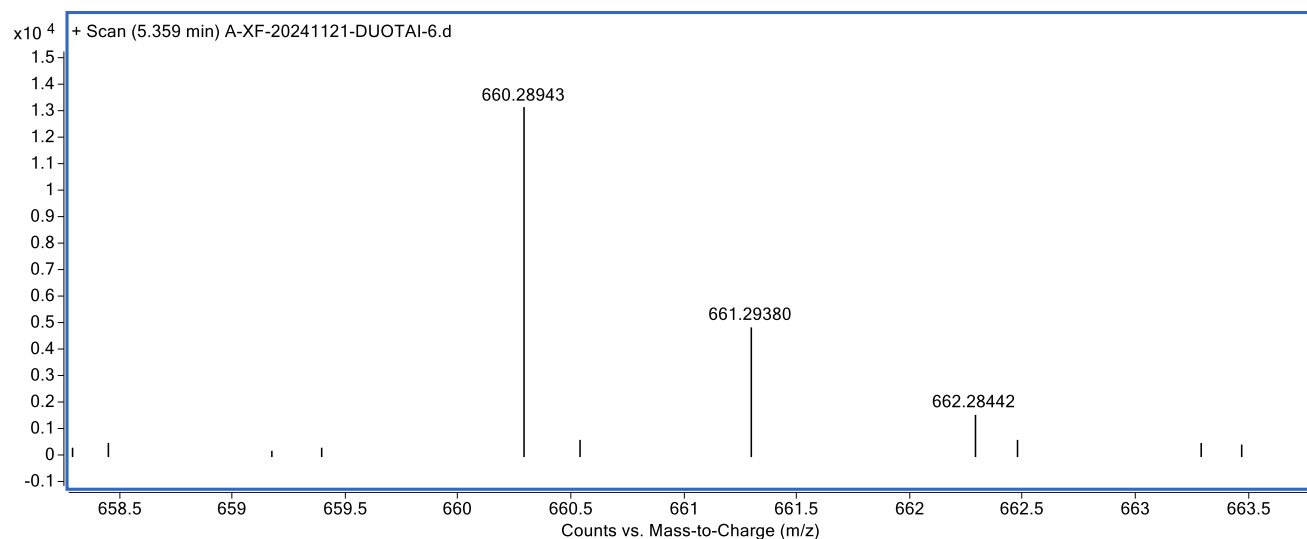
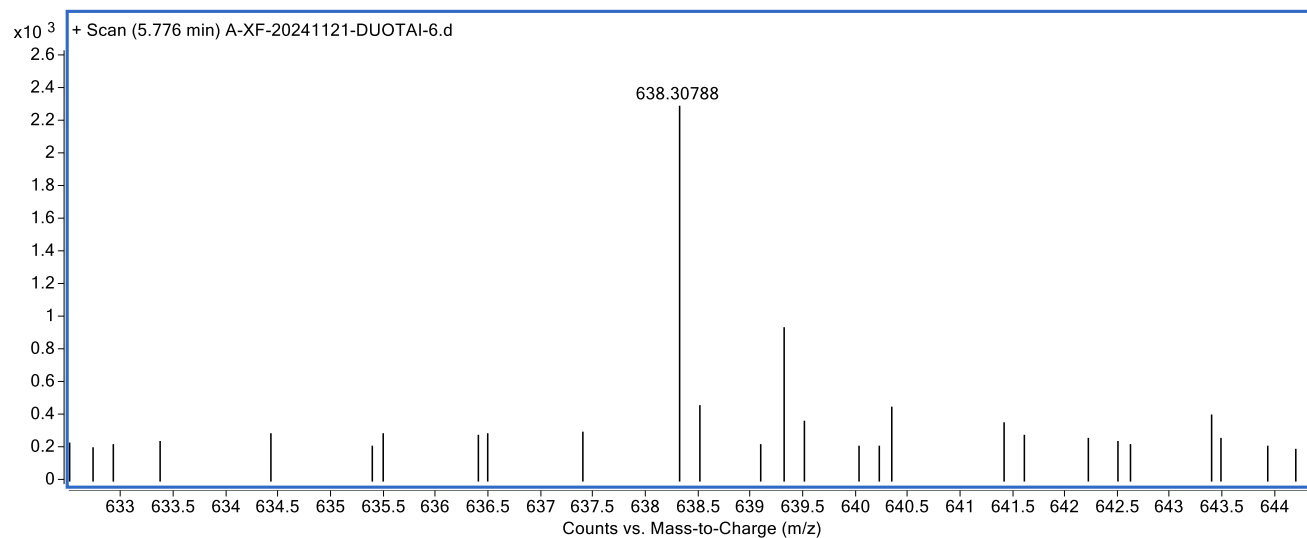
bioconjugated product 23 :



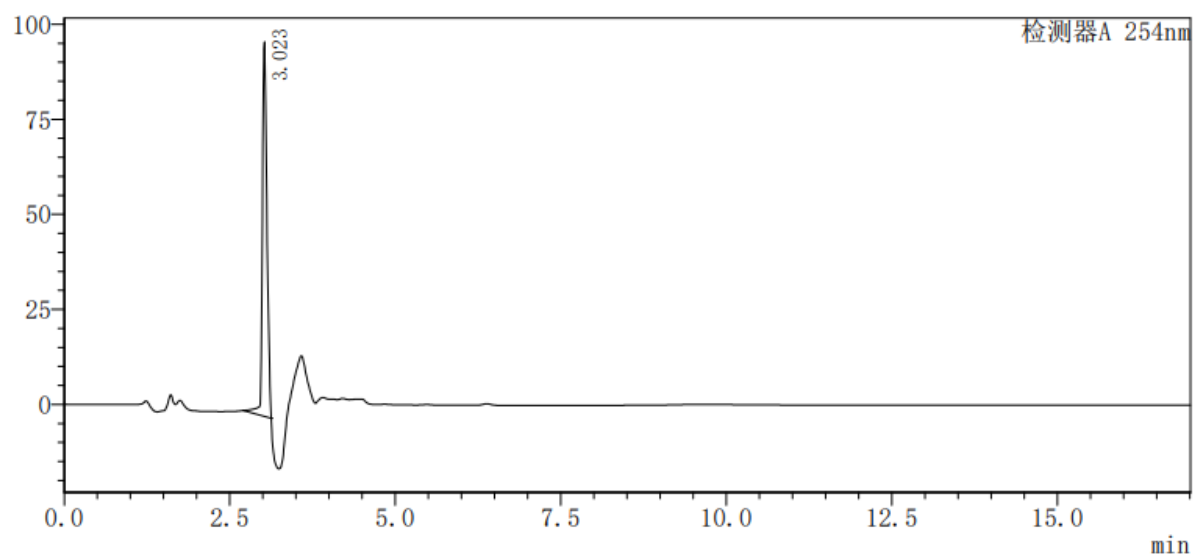
HPLC : 70% conversion.

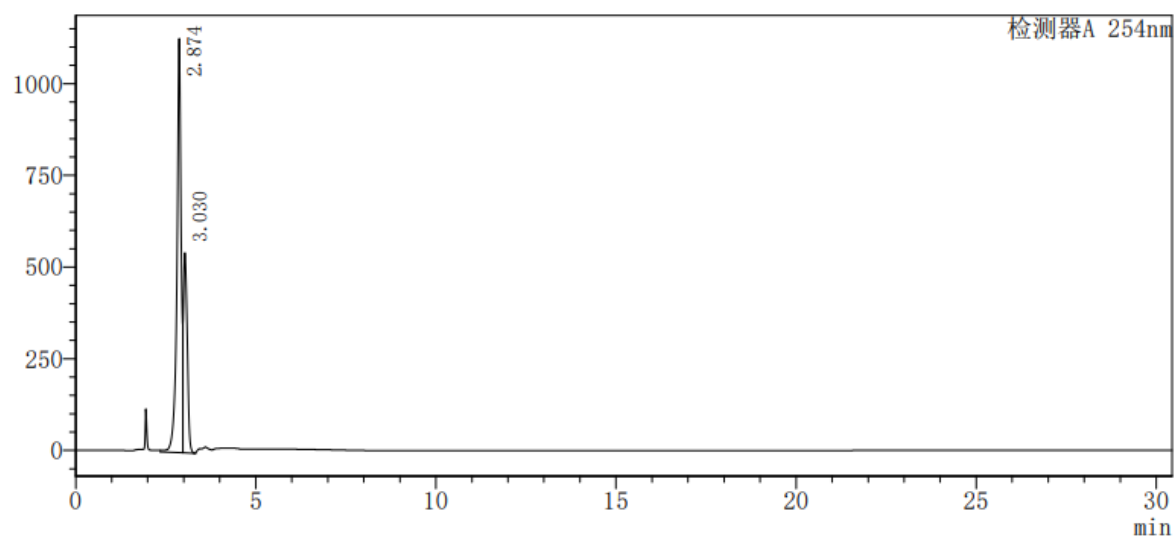
After the reaction finished, there is a peak that elutes at 100% buffer B (acetonitrile + 0.1% TFA) with a retention time of 3.023 min. Polypeptide is a peak that elutes at 100% buffer B (acetonitrile + 0.1% TFA) with a retention time of 2.874 min.

HRMS (ESI-TOF) calcd for C₂₇H₄₃N₉O₇S, [M+H]⁺, 638.30789, found 638.30788. calcd for C₂₇H₄₃N₉O₇S, [M+Na]⁺, 660.28984, found 660.28943.

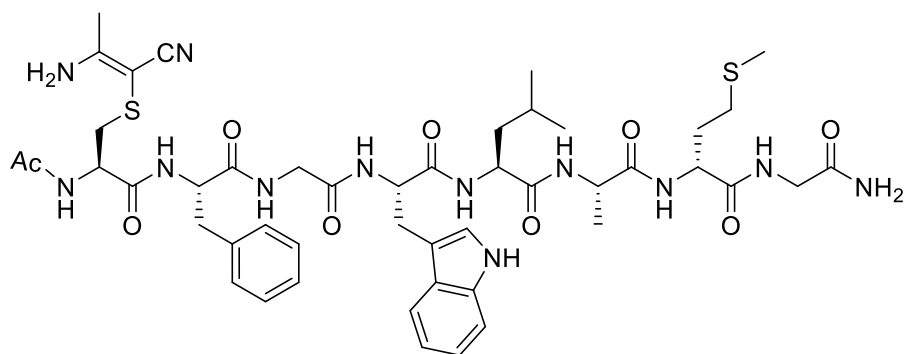


HPLC Spectra:





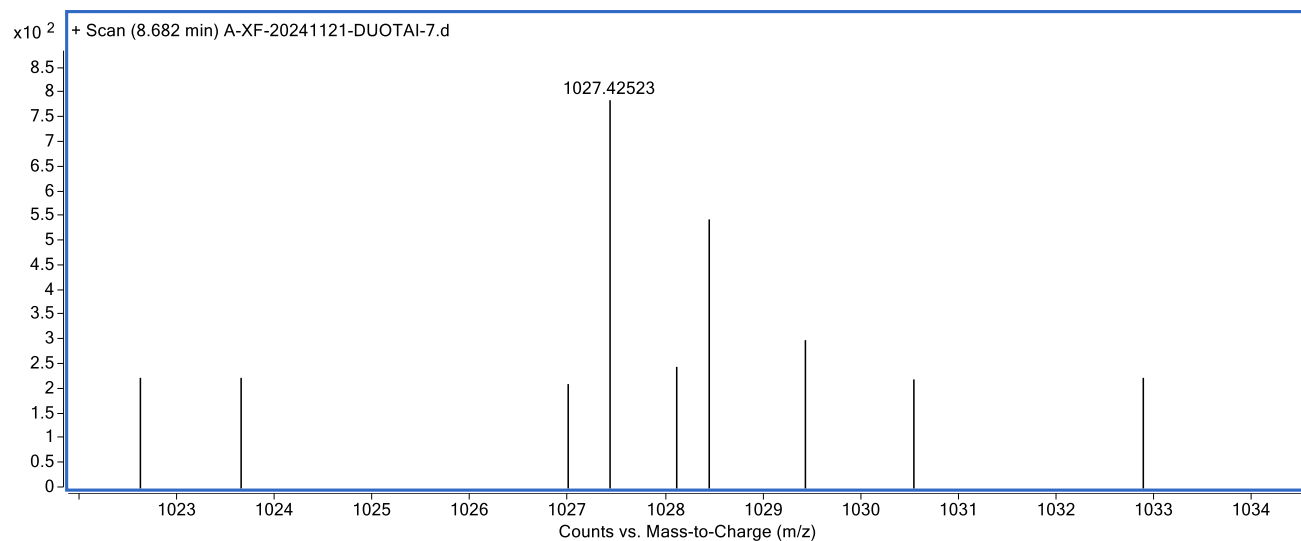
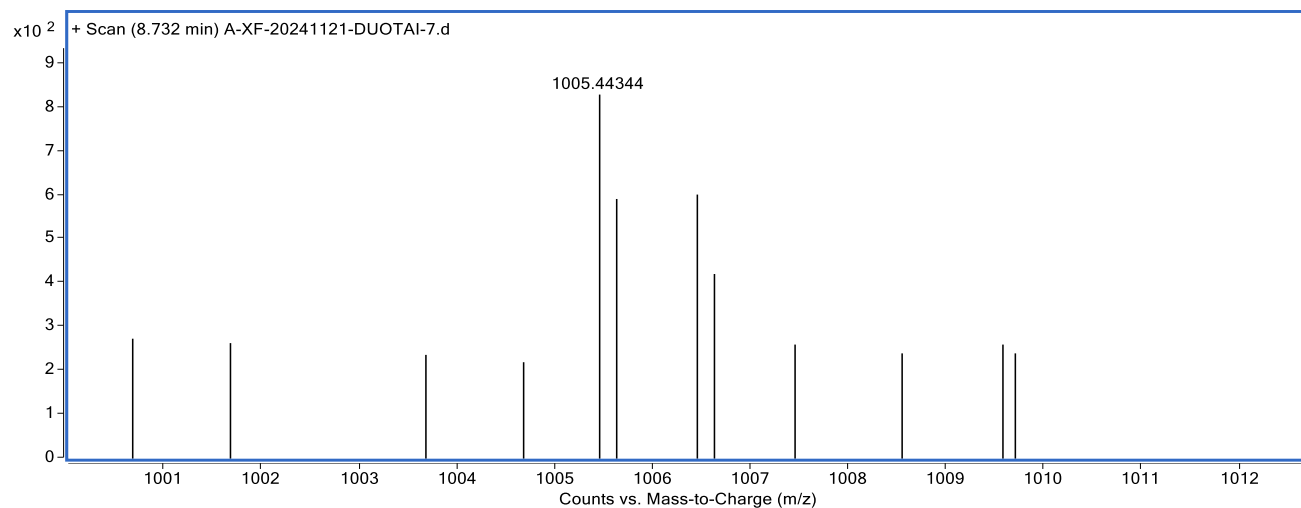
bioconjugated product 24 :



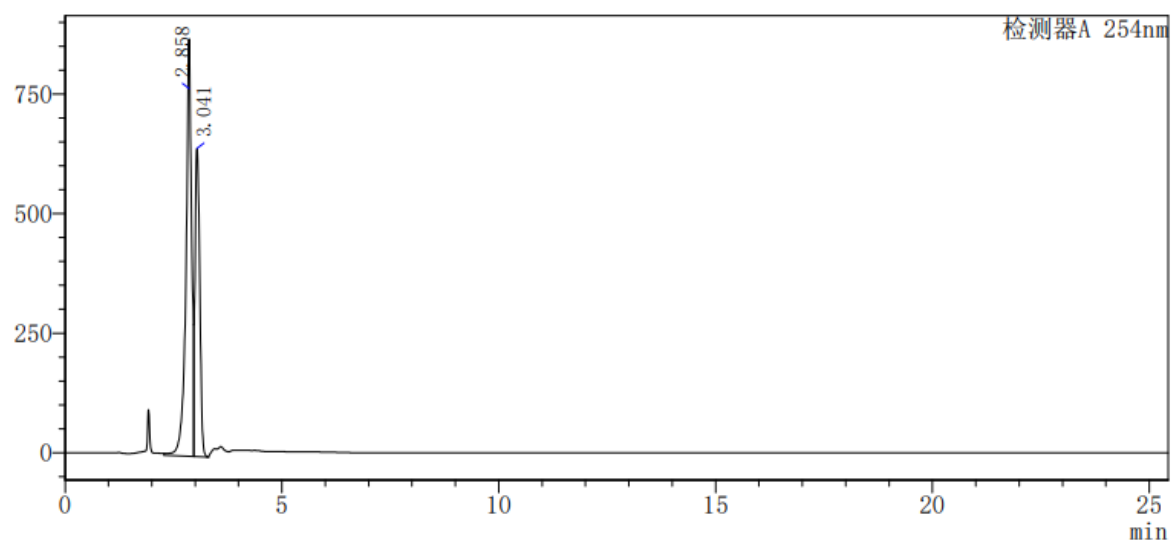
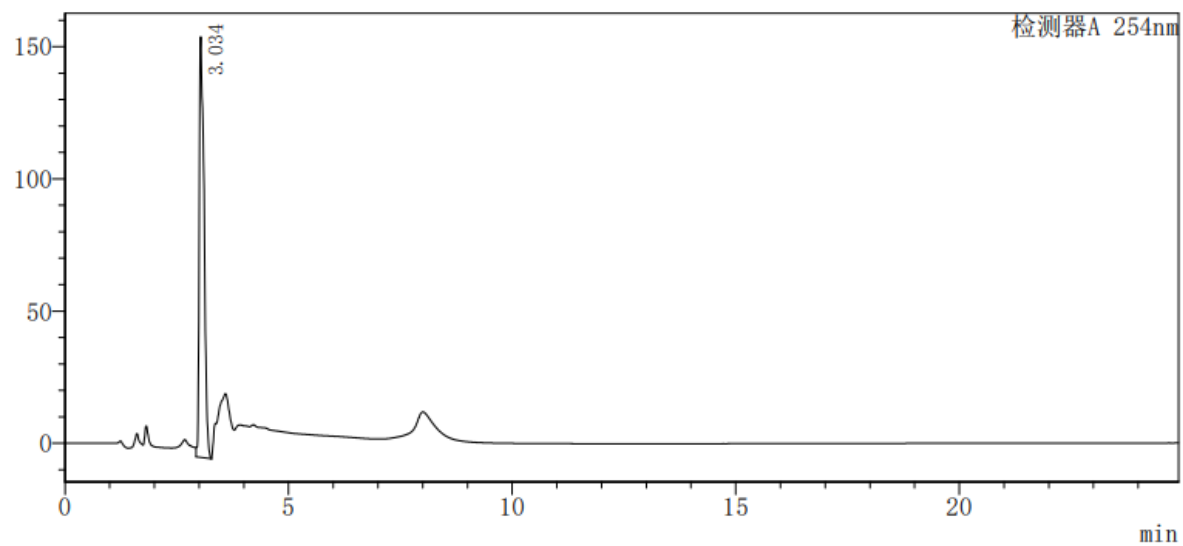
HPLC: 62% conversion.

After the reaction finished, there is a peak that elutes at 100% buffer B (acetonitrile + 0.1% TFA) with a retention time of 3.034 min. Polypeptide is a peak that elutes at 100% buffer B (acetonitrile + 0.1% TFA) with a retention time of 2.858 min.

HRMS (ESI-TOF) calcd for $C_{47}H_{64}N_{12}O_9S_2$, $[M+H]^+$, 1005.44334, found 1005.44344. calcd for $C_{47}H_{64}N_{12}O_9S_2$, $[M+Na]^+$, 1027.42528, found 1027.42523.

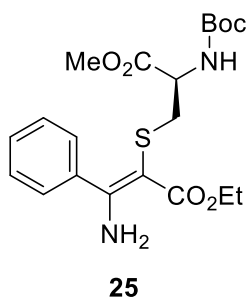


HPLC Spectra:

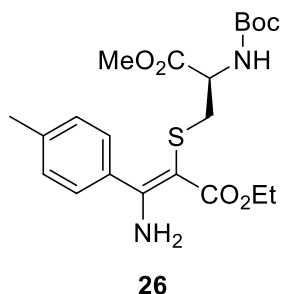


4.3 Enamine and azole scope and characterization

Detailed descriptions for products:

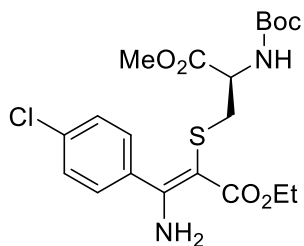


Ethyl (R,E)-3-amino-2-((2-((tert-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)thio)-3-phenylacrylate (25): yellow solid (Yield: 71%, 60.23 mg), ^1H NMR (400 MHz, Chloroform-d) δ 9.10 (s, 1H), 7.42 (d, J = 2.2 Hz, 5H), 5.31 (d, J = 9.2 Hz, 2H), 4.33 – 4.23 (m, 3H), 3.64 (s, 3H), 3.10 (dd, J = 13.9, 3.9 Hz, 1H), 2.46 (dd, J = 13.9, 5.3 Hz, 1H), 1.43 (d, J = 8.3 Hz, 12H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.69, 170.33, 168.57, 155.35, 138.09, 129.44, 128.18, 127.93, 84.45, 79.64, 60.48, 53.47, 52.60, 51.84, 37.58, 28.32, 14.41. HRMS (ESI) calcd. for $(\text{M}+\text{H})^+$ $\text{C}_{20}\text{H}_{28}\text{N}_2\text{O}_6\text{S}$: 425.1740, found, 425.1732.



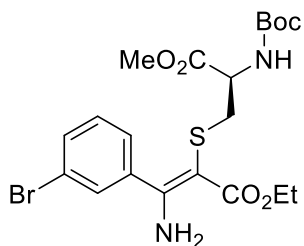
Ethyl (R,E)-3-amino-2-((2-((tert-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)thio)-3-(p-tolyl)acrylate (26): yellow oil (Yield: 66%, 57.84 mg), ^1H NMR (400 MHz, Chloroform-d) δ 9.10 (s, 1H), 7.31 (d, J = 8.1 Hz, 2H), 7.23 (d, J = 7.9 Hz, 2H), 5.29 (d, J = 8.5 Hz, 2H), 4.36 – 4.22 (m, 3H), 3.65 (s, 3H), 3.14 (dd, J = 13.8, 3.7 Hz, 1H), 2.45 (dd, J = 13.9, 5.4 Hz, 1H), 2.39 (s, 3H), 1.41 (d, J = 3.7 Hz, 12H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.73, 170.37, 168.76,

155.40, 139.50, 135.30, 128.86, 127.91, 84.25, 79.60, 60.46, 52.63, 51.79, 37.53, 28.34, 21.43, 14.44. HRMS (ESI) cald. for (M+H)⁺ C₂₁H₃₀N₂O₆S: 439.1897, found, 439.1897.



27

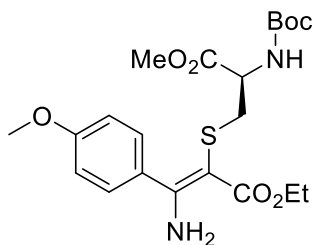
Ethyl (R,E)-3-amino-2-((2-((tert-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)thio)-3-(4-chlorophenyl)acrylate (27): light yellow oil (Yield: 61%, 55.89 mg), ¹H NMR (400 MHz, Chloroform-d) δ 9.08 (s, 1H), 7.41 (t, *J* = 7.6 Hz, 4H), 5.36 – 5.08 (m, 2H), 4.37 – 4.20 (m, 3H), 3.65 (s, 3H), 3.14 (dd, *J* = 13.9, 3.6 Hz, 1H), 2.48 (dd, *J* = 13.9, 5.3 Hz, 1H), 1.43 (d, *J* = 7.3 Hz, 12H). ¹³C NMR (101 MHz, CDCl₃) δ 171.65, 170.13, 167.17, 155.29, 136.35, 135.48, 129.57, 128.46, 84.73, 79.78, 60.58, 52.66, 51.72, 37.34, 28.33, 14.40. HRMS (ESI) cald. for (M+H)⁺ C₂₀H₂₇ClN₂O₆S: 459.1351, found, 459.1351.



28

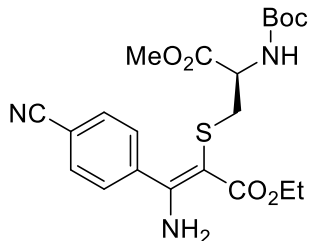
Ethyl (R,E)-3-amino-3-(3-bromophenyl)-2-((2-((tert-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)thio)acrylate (28): light yellow solid (Yield: 68%, 68.27 mg), ¹H NMR (400 MHz, Chloroform-d) δ 9.08 (s, 1H), 7.57 (dt, *J* = 5.7, 1.7 Hz, 2H), 7.40 – 7.29 (m, 2H), 5.47 – 5.14 (m, 2H), 4.40 – 4.20 (m, 3H), 3.66 (s, 3H), 3.12 (dd, *J* = 13.9, 3.9 Hz, 1H), 2.50 (dd, *J* = 13.9, 5.3 Hz,

1H), 1.43 (d, $J = 5.3$ Hz, 12H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.63, 170.12, 166.60, 155.28, 139.83, 132.48, 130.99, 129.78, 126.70, 122.07, 85.14, 79.78, 60.64, 52.66, 51.86, 37.65, 28.31, 14.39. HRMS (ESI) cald. for $(\text{M}+\text{H})^+$ $\text{C}_{20}\text{H}_{27}\text{BrN}_2\text{O}_6\text{S}$: 503.0846, found, 503.0838.



29

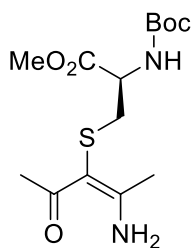
Ethyl (R,E)-3-amino-2-((2-((tert-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)thio)-3-(4-methoxyphenyl)acrylate (29): yellow oil (Yield: 84%, 76.30 mg), ^1H NMR (400 MHz, Chloroform- d) δ 9.13 (s, 1H), 7.41 – 7.35 (m, 2H), 6.96 – 6.92 (m, 2H), 5.39 – 5.19 (m, 2H), 4.34 – 4.22 (m, 3H), 3.84 (s, 3H), 3.65 (s, 3H), 3.13 (dd, $J = 13.9, 3.7$ Hz, 1H), 2.46 (dd, $J = 13.9, 5.4$ Hz, 1H), 1.43 (d, $J = 5.5$ Hz, 12H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.67, 170.44, 168.43, 160.44, 155.35, 130.28, 129.69, 113.40, 84.04, 79.58, 60.40, 55.26, 52.61, 51.75, 37.47, 28.29, 14.43. HRMS (ESI) cald. for $(\text{M}+\text{H})^+$ $\text{C}_{21}\text{H}_{30}\text{N}_2\text{O}_7\text{S}$: 455.1846, found, 455.1885.



30

Ethyl (R,E)-3-amino-2-((2-((tert-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)thio)-3-(4-cyanophenyl)acrylate (30): yellow solid (Yield: 50%, 44.92 mg), ^1H NMR (400 MHz, Chloroform- d) δ 9.08 (s, 1H), 7.74 (d, $J = 8.3$ Hz, 2H), 7.57 (d, $J = 8.0$ Hz, 2H), 5.42 – 5.02 (m,

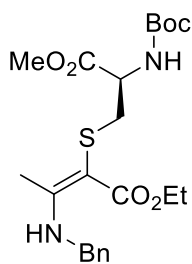
2H), 4.39 – 4.24 (m, 3H), 3.66 (s, 3H), 3.11 (dd, $J = 13.9, 3.8$ Hz, 1H), 2.51 (dd, $J = 13.9, 5.3$ Hz, 1H), 1.43 (d, $J = 7.4$ Hz, 12H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.61, 169.88, 165.95, 155.19, 142.24, 132.05, 129.01, 118.17, 113.26, 85.46, 79.96, 60.79, 52.72, 51.80, 37.38, 28.35, 14.36. HRMS (ESI) calcd. for $(\text{M}+\text{H})^+$ $\text{C}_{21}\text{H}_{27}\text{N}_3\text{O}_6\text{S}$: 450.1693, found, 450.1696.



31

Methyl (E)-S-(2-amino-4-oxopent-2-en-3-yl)-N-(tert-butoxycarbonyl)-L-cysteinate (31):

light yellow solid (Yield: 62%, 41.22 mg), ^1H NMR (400 MHz, Chloroform- d) δ 10.91 – 10.70 (m, 1H), 6.14 (s, 1H), 5.41 (d, $J = 8.0$ Hz, 1H), 4.43 (d, $J = 7.3$ Hz, 1H), 3.72 (s, 3H), 2.98 – 2.83 (m, 2H), 2.43 (s, 3H), 2.30 (s, 3H), 1.45 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 200.14, 171.65, 168.64, 155.22, 127.66, 95.91, 80.18, 52.81, 52.59, 38.99, 28.57, 28.27, 23.32. HRMS (ESI) calcd. for $(\text{M}+\text{H})^+$ $\text{C}_{14}\text{H}_{24}\text{N}_2\text{O}_5\text{S}$: 333.1478, found, 333.1473.



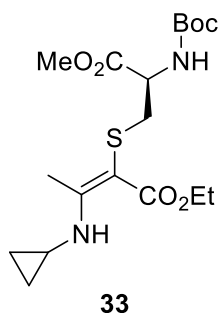
32

Ethyl (R,E)-3-(benzylamino)-2-((2-((tert-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)

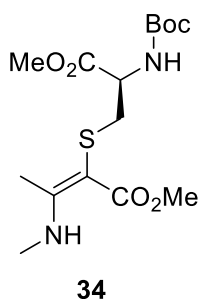
thio)but-2-enoate (32): yellow oil (Yield: 45%, 40.70 mg), ^1H NMR (400 MHz, Chloroform- d)

δ 10.40 (t, $J = 6.0$ Hz, 1H), 7.35 (dd, $J = 8.0, 6.6$ Hz, 2H), 7.30 – 7.25 (m, 3H), 5.80 (d, $J = 8.5$

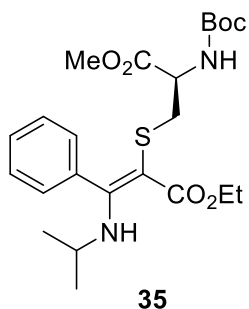
Hz, 1H), 4.47 (t, $J = 7.0$ Hz, 3H), 4.21 (qd, $J = 7.1, 1.1$ Hz, 2H), 3.63 (s, 3H), 3.16 (dd, $J = 13.8, 4.7$ Hz, 1H), 2.75 (dd, $J = 13.8, 4.9$ Hz, 1H), 2.40 (s, 3H), 1.45 (s, 9H), 1.38 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.96, 170.87, 169.12, 155.47, 137.50, 128.94, 127.68, 126.98, 82.72, 79.84, 60.12, 52.47, 48.22, 38.11, 28.35, 17.55, 14.43. HRMS (ESI) calcd. for $(\text{M}+\text{H})^+$ $\text{C}_{22}\text{H}_{32}\text{N}_2\text{O}_6\text{S}$: 453.2053, found, 453.2053.



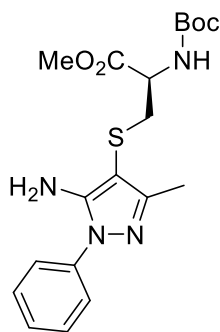
Ethyl (R,E)-2-((2-((tert-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)thio)-3-(cyclopropylamino)but-2-enoate (33): yellow oil (Yield: 71%, 57.11 mg), ^1H NMR (400 MHz, Chloroform- d) δ 9.96 (s, 1H), 5.80 (d, $J = 8.5$ Hz, 1H), 4.44 (dt, $J = 9.0, 4.8$ Hz, 1H), 4.19 (q, $J = 7.1$ Hz, 2H), 3.64 (s, 3H), 3.16 (dd, $J = 13.8, 4.7$ Hz, 1H), 2.74 (dd, $J = 13.8, 4.9$ Hz, 1H), 2.64 – 2.59 (m, 1H), 2.51 (s, 3H), 1.46 (s, 9H), 1.37 (t, $J = 7.1$ Hz, 3H), 0.82 (td, $J = 6.8, 4.9$ Hz, 2H), 0.68 – 0.59 (m, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.89, 171.17, 170.65, 155.43, 82.21, 79.79, 60.02, 52.27, 37.79, 28.31, 25.88, 18.31, 14.41, 7.95. HRMS (ESI) calcd. for $(\text{M}+\text{Na})^+$ $\text{C}_{18}\text{H}_{30}\text{N}_2\text{O}_6\text{S}$: 425.1716, found, 425.1714.



Methyl (R,E)-2-((2-((tert-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)thio)-3-(methylamino)but-2-enoate (34): yellow oil (Yield: 65%, 47.08 mg), ^1H NMR (400 MHz, Chloroform- d) δ 10.07 – 9.91 (m, 1H), 5.82 (d, J = 8.4 Hz, 1H), 4.40 (dt, J = 9.2, 5.0 Hz, 1H), 3.76 (s, 3H), 3.66 (s, 3H), 3.10 (dd, J = 13.8, 5.1 Hz, 1H), 2.98 (d, J = 5.1 Hz, 3H), 2.73 (dd, J = 13.8, 4.9 Hz, 1H), 2.38 (s, 3H), 1.46 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.97, 171.21, 170.31, 155.53, 81.60, 79.75, 52.41, 51.30, 38.36, 30.91, 28.33, 17.21. HRMS (ESI) cald. for $(\text{M}+\text{Na})^+$ $\text{C}_{15}\text{H}_{26}\text{N}_2\text{O}_6\text{S}$: 385.1403, found, 385.1410.



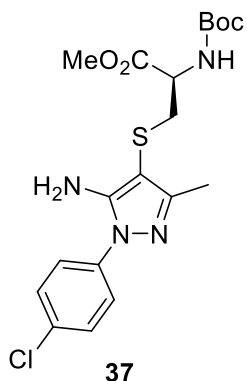
Ethyl (R,E)-2-((2-((tert-butoxycarbonyl)amino)-3-methoxy-3-oxopropyl)thio)-3-(isopropylamino)-3-phenylacrylate (35): light yellow solid (Yield: 64%, 59.67 mg), ^1H NMR (400 MHz, Chloroform- d) δ 9.89 (d, J = 9.6 Hz, 1H), 7.50 – 7.43 (m, 3H), 7.31 (d, J = 7.2 Hz, 1H), 7.24 (d, J = 6.7 Hz, 1H), 5.60 (d, J = 8.8 Hz, 1H), 4.37 (dt, J = 9.0, 4.6 Hz, 1H), 4.32 – 4.23 (m, 2H), 3.67 (s, 3H), 3.17 (dt, J = 9.5, 6.4 Hz, 1H), 3.05 (dd, J = 13.8, 3.9 Hz, 1H), 2.38 (dd, J = 13.8, 5.2 Hz, 1H), 1.45 (d, J = 9.4 Hz, 12H), 1.08 (t, J = 6.6 Hz, 6H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.89, 171.05, 169.97, 155.50, 135.39, 128.73, 128.43, 128.27, 127.41, 81.76, 79.63, 60.21, 52.57, 51.84, 47.08, 38.15, 28.37, 24.06, 14.46. HRMS (ESI) cald. for $(\text{M}+\text{H})^+$ $\text{C}_{23}\text{H}_{34}\text{N}_2\text{O}_6\text{S}$: 467.2210, found, 467.2207.



36

Methyl S-(5-amino-3-methyl-1-phenyl-1H-pyrazol-4-yl)-N-(tert-butoxycarbonyl)-L-cystein

ate (36): yellow oil (Yield: 72%, 58.49 mg), ^1H NMR (400 MHz, Chloroform-d) δ 7.54 (d, J = 7.4 Hz, 2H), 7.47 (t, J = 7.8 Hz, 2H), 7.34 (t, J = 7.4 Hz, 1H), 5.52 (d, J = 8.5 Hz, 1H), 4.51 (s, 2H), 3.68 (s, 3H), 3.05 – 2.87 (m, 2H), 2.29 (s, 3H), 1.43 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.50, 155.41, 152.44, 148.47, 138.55, 129.58, 127.43, 123.56, 89.39, 80.28, 53.78, 52.53, 38.80, 28.30, 12.33. HRMS (ESI) cald. for $(\text{M}+\text{H})^+$ $\text{C}_{19}\text{H}_{26}\text{N}_4\text{O}_4\text{S}$: 407.1747, found, 407.1769.

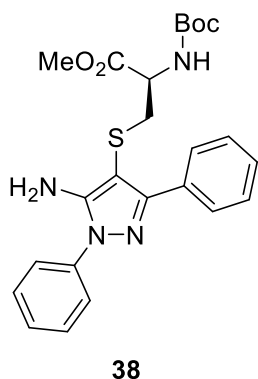


37

Methyl S-(5-amino-1-(4-chlorophenyl)-3-methyl-1H-pyrazol-4-yl)-N-(tert-butoxycarbonyl)-

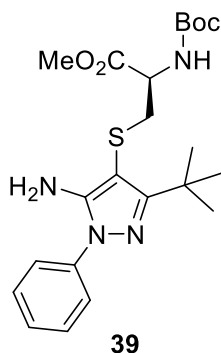
L-cysteinate (37) : yellow oil ((Yield: 62%, 54.58 mg), ^1H NMR (400 MHz, Chloroform-d) δ 7.51 (d, J = 8.7 Hz, 2H), 7.43 (d, J = 8.7 Hz, 2H), 5.49 (d, J = 8.5 Hz, 1H), 4.55 – 4.47 (m, 2H), 3.68 (s, 3H), 2.93 (qd, J = 13.9, 5.3 Hz, 2H), 2.27 (s, 3H), 1.44 (d, J = 11.3 Hz, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.45, 155.40, 152.80, 148.60, 137.13, 132.93, 129.68, 124.62, 89.96,

80.33, 53.79, 52.55, 38.84, 28.29, 12.30. HRMS (ESI) cald. for (M+H)⁺ C₁₉H₂₅ClN₄O₄S: 441.1357,found, 441.1313.



Methyl S-(5-amino-1,3-diphenyl-1H-pyrazol-4-yl)-N-(tert-butoxycarbonyl)-L-cysteinate

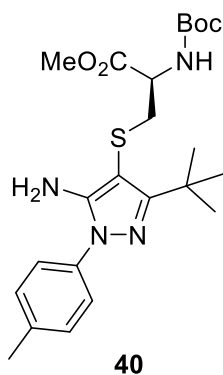
(38) : yellow oil (Yield: 45%, 42.14 mg), ¹H NMR (400 MHz, Chloroform-d) δ 8.09 (d, *J* = 7.6 Hz, 2H), 7.63 (d, *J* = 7.8 Hz, 2H), 7.51 – 7.32 (m, 6H), 5.50 (d, *J* = 8.4 Hz, 1H), 4.65 – 4.56 (m, 1H), 4.50 – 4.42 (m, 1H), 3.74 (s, 1H), 3.60 (s, 3H), 2.94 (d, *J* = 5.3 Hz, 2H), 1.43 (d, *J* = 17.0 Hz, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 171.35, 155.34, 152.32, 149.48, 138.53, 132.93, 129.63, 128.34, 128.28, 127.73, 127.52, 123.77, 88.31, 80.17, 53.58, 52.48, 38.84, 28.32. HRMS (ESI) cald. for (M+K)⁺ C₂₄H₂₈N₄O₄S: 507.1462,found, 507.1427.



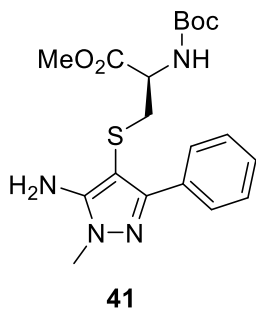
Methyl S-(5-amino-3-(tert-butyl)-1-phenyl-1H-pyrazol-4-yl)-N-(tert-butoxycarbonyl)-L-cysteinate

(39) : brown oil (Yield: 40%, 35.86 mg), ¹H NMR (400 MHz, Chloroform-d) δ 8.09 (d, *J* = 7.6 Hz, 2H), 7.63 (d, *J* = 7.8 Hz, 2H), 7.51 – 7.32 (m, 6H), 5.50 (d, *J* = 8.4 Hz, 1H), 4.65 –

4.56 (m, 1H), 4.50 – 4.42 (m, 1H), 3.60 (s, 3H), 2.94 (d, $J = 5.3$ Hz, 2H), 1.43 (d, $J = 17.0$ Hz, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.70, 162.05, 155.43, 149.60, 138.75, 129.53, 127.19, 123.52, 87.82, 80.23, 54.26, 52.62, 40.59, 33.71, 29.59, 28.31. HRMS (ESI) calcd. for $(\text{M}+\text{Na})^+$ $\text{C}_{22}\text{H}_{32}\text{N}_4\text{O}_4\text{S}$: 471.2036, found, 471.2024.

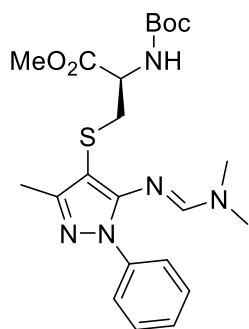


Methyl S-(5-amino-3-(tert-butyl)-1-(p-tolyl)-1H-pyrazol-4-yl)-N-(tert-butoxycarbonyl)-L-cysteinate (40) : brown oil (Yield: 53%, 48.99 mg), ^1H NMR (400 MHz, Chloroform- d) δ 7.43 (d, $J = 8.1$ Hz, 2H), 7.24 (d, $J = 8.1$ Hz, 2H), 5.75 (d, $J = 8.7$ Hz, 1H), 4.70 – 4.40 (m, 3H), 3.74 (s, 3H), 3.02 (dd, $J = 13.6, 4.1$ Hz, 1H), 2.93 (dd, $J = 13.7, 6.4$ Hz, 1H), 2.37 (s, 3H), 1.43 (s, 9H), 1.41 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.71, 161.76, 155.42, 149.48, 137.11, 136.27, 130.02, 123.53, 87.44, 80.16, 54.25, 52.57, 40.51, 33.67, 29.62, 28.31, 21.08. HRMS (ESI) calcd. for $(\text{M}+\text{Na})^+$ $\text{C}_{23}\text{H}_{34}\text{N}_4\text{O}_4\text{S}$: 485.2193, found, 485.2197.



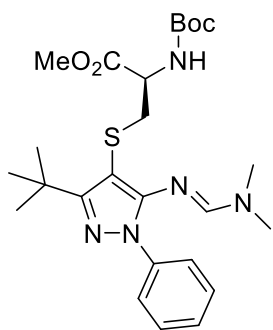
Methyl S-(5-amino-1-methyl-3-phenyl-1H-pyrazol-4-yl)-N-(tert-butoxycarbonyl)-L-cysteinate

ate (41) : brown oil (Yield: 39%, 31.68 mg), ^1H NMR (400 MHz, Chloroform- d) δ 8.04 – 7.94 (m, 2H), 7.43 – 7.37 (m, 2H), 7.36 – 7.30 (m, 1H), 5.42 – 5.21 (m, 1H), 4.46 – 4.35 (m, 1H), 4.25 (s, 2H), 3.70 (s, 3H), 3.60 (s, 3H), 2.90 – 2.80 (m, 2H), 1.43 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.42, 155.32, 150.92, 149.54, 133.18, 128.33, 127.97, 127.27, 88.17, 80.18, 53.49, 52.43, 39.18, 35.02, 28.30. HRMS (ESI) cald. for $(\text{M}+\text{H})^+$ $\text{C}_{19}\text{H}_{26}\text{N}_4\text{O}_4\text{S}$: 407.1747, found, 407.1746.



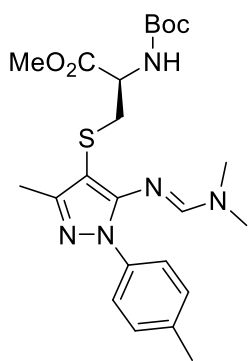
42

Methyl (E)-N-(tert-butoxycarbonyl)-S-(5-(((dimethylamino)methylene)amino)-3-methyl-1-phenyl-1H-pyrazol-4-yl)-L-cysteinate (42) : light yellow oil (Yield: 63%, 58.11 mg), ^1H NMR (400 MHz, Chloroform- d) δ 7.81 – 7.60 (m, 3H), 7.28 (t, J = 7.7 Hz, 2H), 7.12 (t, J = 7.3 Hz, 1H), 5.85 (d, J = 8.9 Hz, 1H), 4.47 (dt, J = 8.8, 4.3 Hz, 1H), 3.53 (s, 3H), 3.10 (dd, J = 14.0, 4.0 Hz, 1H), 3.04 (s, 3H), 2.95 (s, 3H), 2.69 (dd, J = 14.0, 4.3 Hz, 1H), 2.25 (s, 3H), 1.32 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.03, 156.35, 155.42, 152.53, 152.13, 139.93, 128.36, 125.78, 123.19, 95.16, 79.66, 53.52, 52.26, 40.29, 38.15, 34.38, 28.32, 12.70. HRMS (ESI) cald. for $(\text{M}+\text{K})^+$ $\text{C}_{22}\text{H}_{31}\text{N}_5\text{O}_4\text{S}$: 500.1728, found, 500.1723.



43

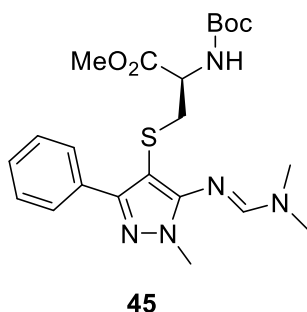
Methyl (E)-N-(tert-butoxycarbonyl)-S-(3-(tert-butyl)-5-(((dimethylamino)methylene)amino)-1-phenyl-1H-pyrazol-4-yl)-L-cysteinate (43) : light yellow solid (Yield: 50%, 50.33 mg), ^1H NMR (400 MHz, Chloroform- d) δ 7.84 (d, J = 7.9 Hz, 2H), 7.51 (s, 1H), 7.35 (t, J = 7.8 Hz, 2H), 7.18 (t, J = 7.4 Hz, 1H), 6.69 (d, J = 9.5 Hz, 1H), 4.74 – 4.49 (m, 1H), 3.69 (s, 3H), 3.25 (dd, J = 14.0, 4.5 Hz, 1H), 3.19 (s, 3H), 3.01 (s, 3H), 2.69 (dd, J = 14.0, 3.5 Hz, 1H), 1.47 (s, 9H), 1.41 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.38, 161.56, 157.11, 155.83, 153.91, 140.09, 128.38, 125.60, 123.10, 95.10, 79.43, 53.97, 52.16, 40.77, 40.29, 34.41, 33.98, 29.66, 28.40. HRMS (ESI) cald. for $(\text{M}+\text{Na})^+$ $\text{C}_{25}\text{H}_{37}\text{N}_5\text{O}_4\text{S}$:526.2458, found, 526.2415.



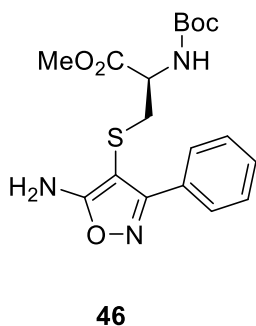
44

Methyl (E)-N-(tert-butoxycarbonyl)-S-(5-(((dimethylamino)methylene)amino)-3-methyl-1-(p-tolyl)-1H-pyrazol-4-yl)-L-cysteinate (44) : colorless oil (Yield: 40%, 38.02 mg), ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 8.10 (s, 1H), 7.63 (d, J = 7.2 Hz, 2H), 7.19 (d, J = 6.7 Hz, 3H), 4.03 (d,

$J = 9.3$ Hz, 1H), 3.58 (s, 3H), 3.04 (s, 3H), 2.94 (s, 3H), 2.77 (s, 2H), 2.31 (s, 3H), 2.18 (s, 3H), 1.38 (s, 9H). ^{13}C NMR (101 MHz, DMSO) δ 171.95, 156.60, 155.77, 152.79, 151.68, 137.97, 135.08, 129.24, 122.73, 94.30, 78.78, 55.34, 53.84, 52.44, 39.36, 34.24, 28.59, 20.96, 12.94. HRMS (ESI) calcd. for $(\text{M}+\text{Na})^+$ $\text{C}_{23}\text{H}_{33}\text{N}_5\text{O}_4\text{S}$: 498.2145, found, 498.2139.

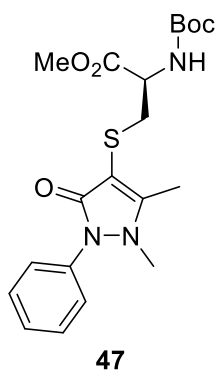


Methyl (E)-N-(tert-butoxycarbonyl)-S-(5-(((dimethylamino)methylene)amino)-1-methyl-3-phenyl-1H-pyrazol-4-yl)-L-cysteinate (45) : yellow oil (Yield: 40%, 36.90 mg), ^1H NMR (400 MHz, Chloroform- d) δ 8.03 – 7.88 (m, 3H), 7.38 – 7.23 (m, 3H), 5.40 (d, $J = 8.8$ Hz, 1H), 4.30 (d, $J = 8.2$ Hz, 1H), 3.66 (s, 3H), 3.50 (s, 3H), 3.06 (d, $J = 16.2$ Hz, 6H), 2.94 (d, $J = 13.2$ Hz, 1H), 2.63 (d, $J = 12.6$ Hz, 1H), 1.34 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 170.99, 156.35, 155.24, 153.10, 151.19, 133.70, 128.22, 127.69, 127.47, 91.63, 79.63, 53.25, 52.18, 40.39, 38.60, 34.98, 34.19, 28.34. HRMS (ESI) calcd. for $(\text{M}+\text{H})^+$ $\text{C}_{22}\text{H}_{31}\text{N}_5\text{O}_4\text{S}$: 462.2169, found, 462.2168.



Methyl S-(5-amino-3-phenylisoxazol-4-yl)-N-(tert-butoxycarbonyl)-L-cysteinate (46) : yellow oil (Yield: 55%, 43.25 mg), ^1H NMR (400 MHz, Chloroform- d) δ 7.45 (q, $J = 3.3$ Hz,

3H), 7.41 – 7.32 (m, 2H), 7.19 – 6.92 (m, 1H), 6.26 – 5.65 (m, 2H), 4.96 – 4.62 (m, 1H), 3.76 (s, 3H), 3.58 (ddd, $J = 14.7, 12.7, 3.5$ Hz, 1H), 3.11 (td, $J = 14.1, 7.5$ Hz, 1H), 1.46 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.40, 165.47, 159.66, 155.36, 130.13, 128.80, 127.92, 85.57, 80.31, 53.67, 52.69, 40.40, 29.70, 28.36. HRMS (ESI) cald. for $(\text{M}+\text{H})^+$ $\text{C}_{18}\text{H}_{23}\text{N}_3\text{O}_5\text{S}$: 394.1431, found, 394.1430.



Methyl N-(tert-butoxycarbonyl)-S-(1,5-dimethyl-3-oxo-2-phenyl-2,3-dihydro-1H-pyrazol-4-yl)-L-cysteinate (47) : yellow oil (Yield: 35%, 29.48 mg), ^1H NMR (400 MHz, Chloroform- d) δ 7.50 – 7.42 (m, 2H), 7.38 – 7.33 (m, 2H), 7.33 – 7.28 (m, 1H), 6.21 (s, 1H), 4.44 (s, 1H), 3.62 (s, 3H), 3.35 – 3.26 (m, 1H), 3.10 – 2.90 (m, 1H), 2.35 (s, 2H), 1.42 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 171.59, 165.12, 158.58, 155.55, 134.67, 129.27, 127.09, 124.36, 99.12, 79.67, 53.03, 52.24, 35.79, 28.33, 12.05. HRMS (ESI) cald. for $(\text{M}+\text{H})^+$ $\text{C}_{20}\text{H}_{27}\text{N}_3\text{O}_5\text{S}$: 422.1744, found, 422.1749.

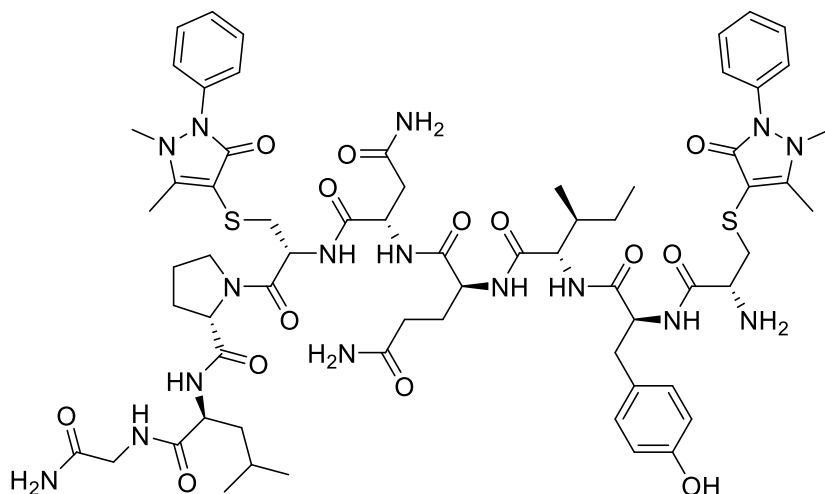
4.4 Electrochemical peptides functionalization^[3]

Bioconjugation of Oxytocin Skeleton:

Synthesis of compound (49) : In an oven-dried undivided three-necked bottle (15 mL) equipped with a stir bar, Oxytocin Skeleton (0.005 mmol), Antipyrine (2 equiv.), $n\text{Bu}_4\text{NBr}$ (2 equiv.),

NiBr₂•diglyme (20 mol%), MeCN (6 mL), CHCl₃ (1 mL), were combined and added. The bottle was equipped platinum plate (10 mm×10 mm×0.3 mm) as the anode and platinum plate (10 mm×10 mm×0.3 mm) as the cathode. The reaction mixture was stirred and electrolysis at constant current of 6 mA under room temperature for 1 h. After completion of the reaction, the solution was analyzed by LC-MS spectroscopy. The reaction was analyzed by reverse phase HPLC using 100% buffer B over 30 minutes on an Agilent Zorbax SB-Aq 5μm column of 250 mm length. HPLC analysis used buffers A (water) and B (acetonitrile + 0.1% TFA). Conversion reported as a % conversion as determined.

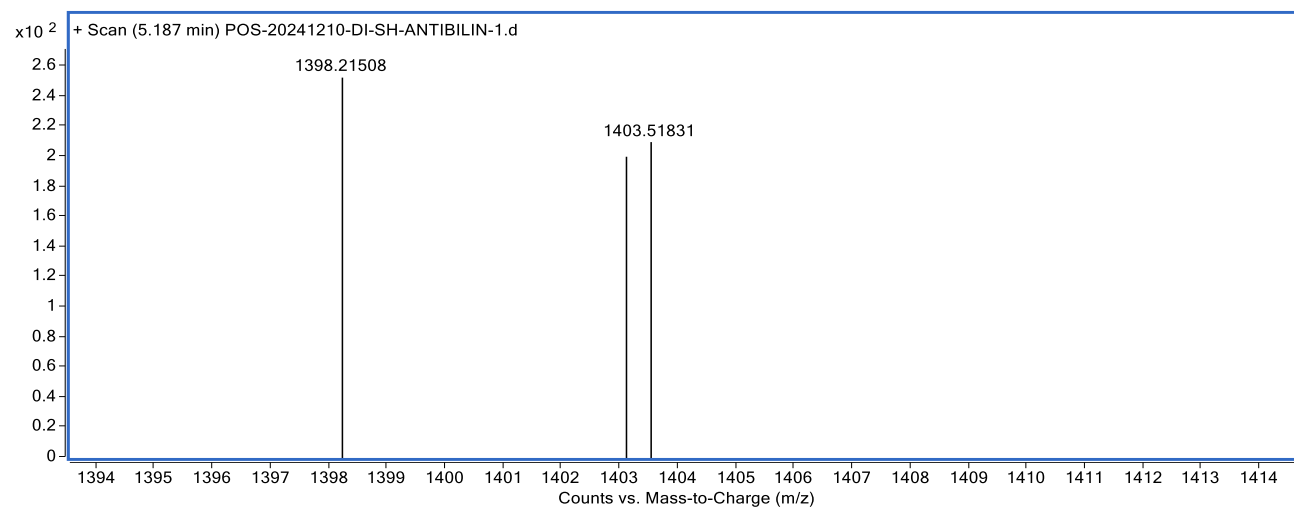
bioconjugated product 49 :



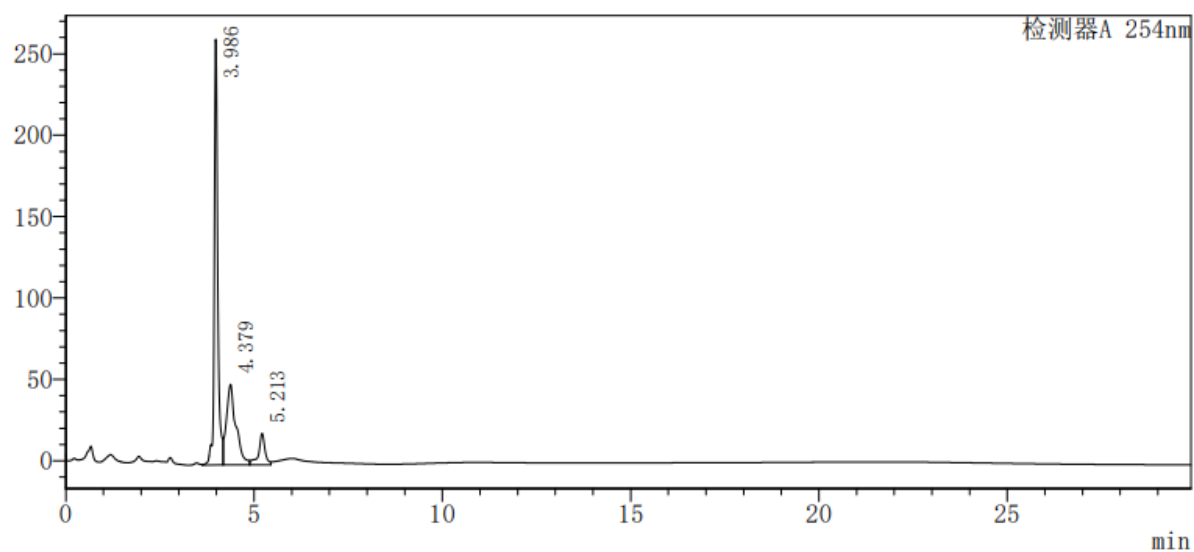
HPLC: 93% conversion.

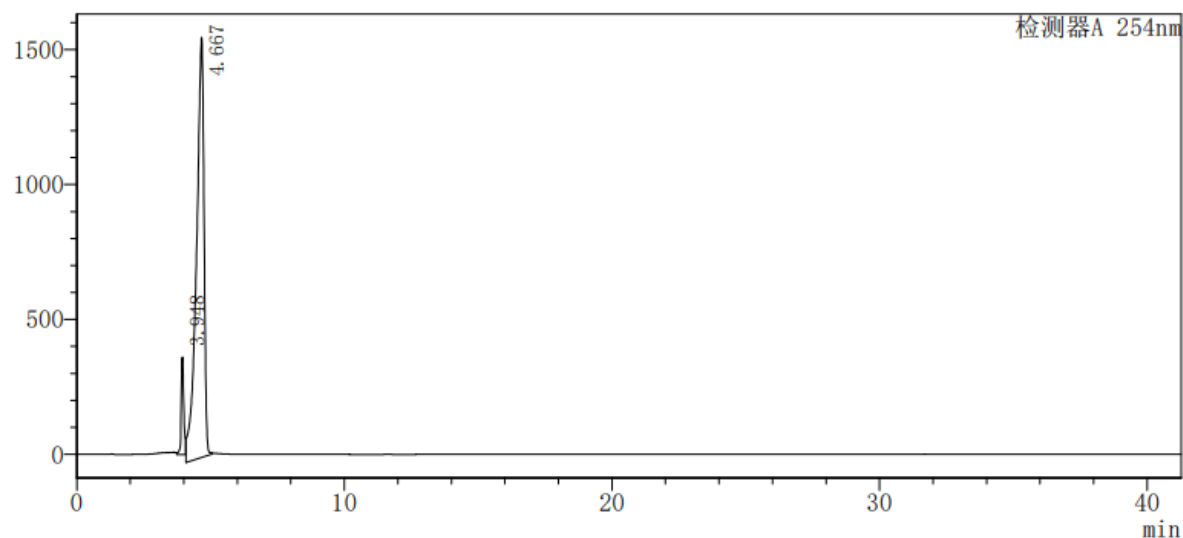
After the reaction finished, there is a peak that elutes at 100% buffer B (acetonitrile + 0.1% TFA) with a retention time of 4.667 min. Polypeptide is a peak that elutes at 100% buffer B (acetonitrile + 0.1% TFA) with a retention time of 3.986 min.

HRMS (ESI-TOF) calcd for C₆₅H₈₈N₁₆O₁₄S₂, [M+Na]⁺, 1403.5999, found 1403.51831.



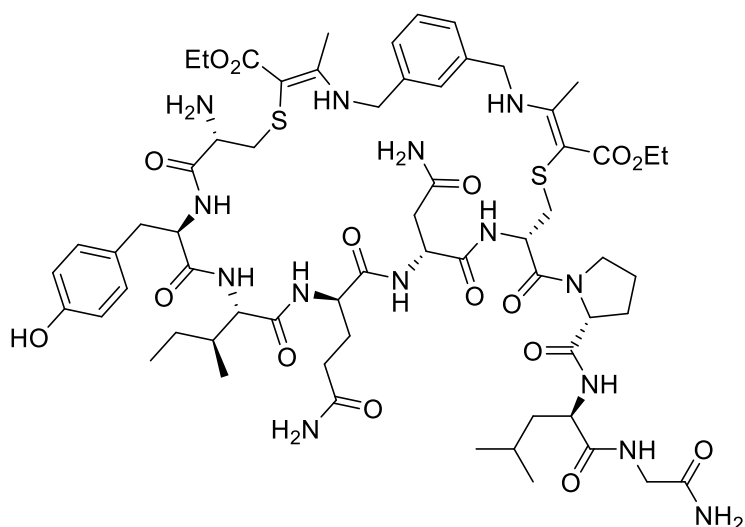
HPLC Spectra:





Synthesis of compound (50) : In an oven-dried undivided three-necked bottle (15 mL) equipped with a stir bar, Oxytocin Skeleton (0.005 mmol), Bisenamine (4 equiv.), $n\text{Bu}_4\text{NBr}$ (2 equiv.), $\text{NiBr}_2 \cdot \text{diglyme}$ (20 mol%), MeCN (6 mL), CHCl_3 (1 mL), were combined and added. The bottle was equipped platinum plate (10 mm×10 mm×0.3 mm) as the anode and platinum plate (10 mm×10 mm×0.3 mm) as the cathode. The reaction mixture was stirred and electrolysis at constant current of 6 mA under room temperature for 1.5 h. After completion of the reaction, the solution was analyzed by LC-MS spectroscopy. The reaction was analyzed by reverse phase HPLC using 100% buffer B over 30 minutes on an Agilent Zorbax SB-Aq 5 μm column of 250 mm length. HPLC analysis used buffers A (water) and B (acetonitrile + 0.1% TFA). Conversion reported as a % conversion as determined.

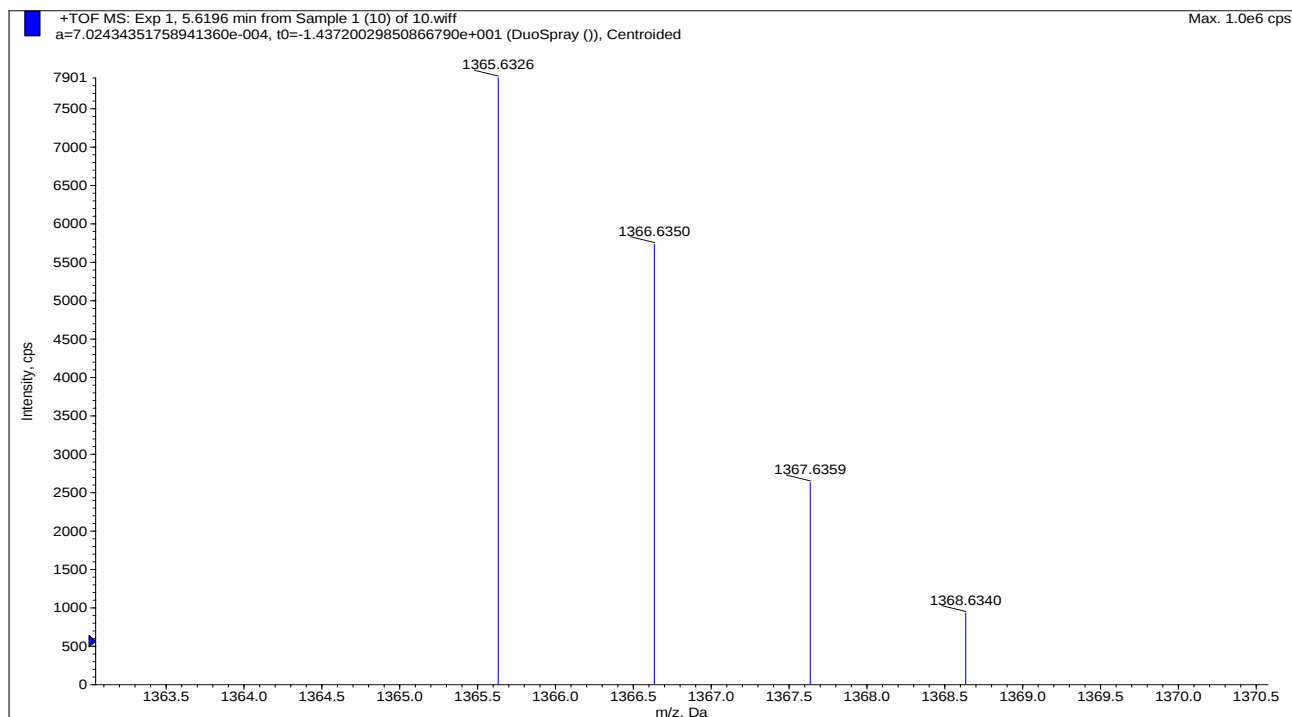
bioconjugated product 50 :



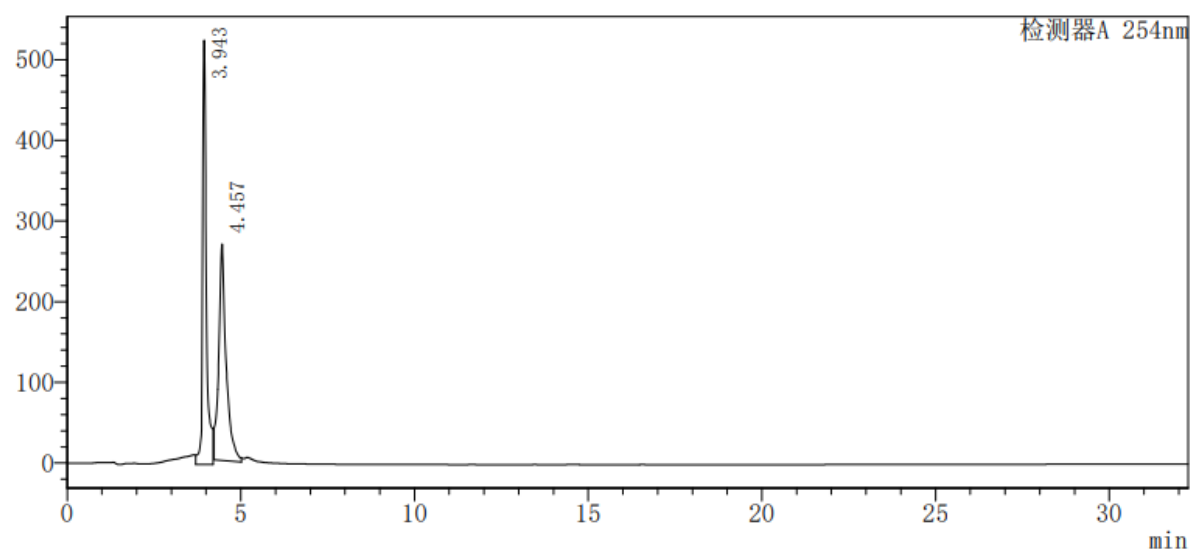
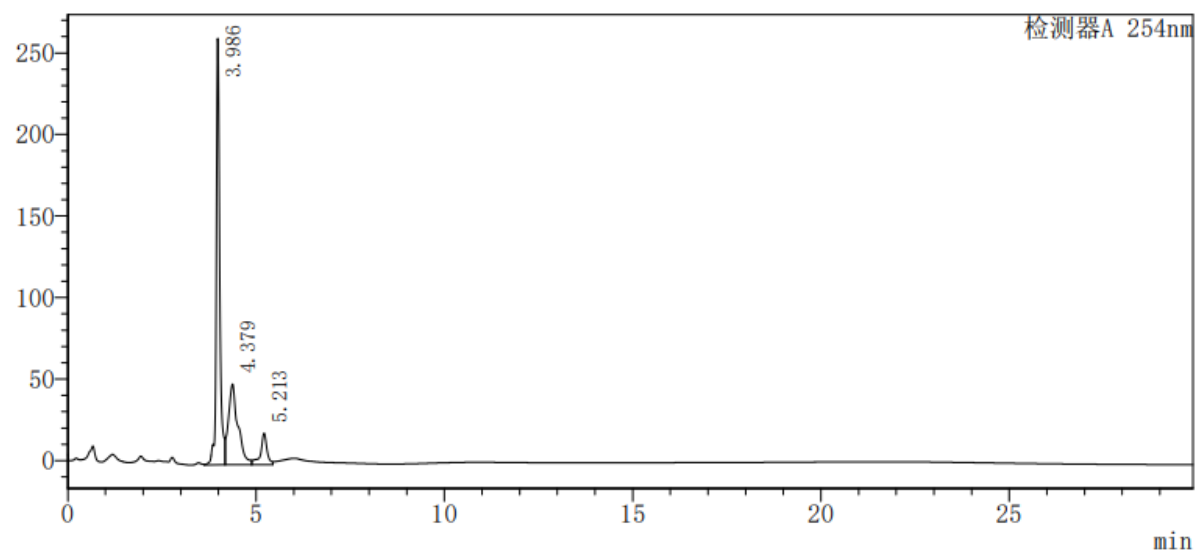
HPLC: 50% conversion.

After the reaction finished, there is a peak that elutes at 100% buffer B (acetonitrile + 0.1% TFA) with a retention time of 4.457 min. Polypeptide is a peak that elutes at 100% buffer B (acetonitrile + 0.1% TFA) with a retention time of 3.986 min.

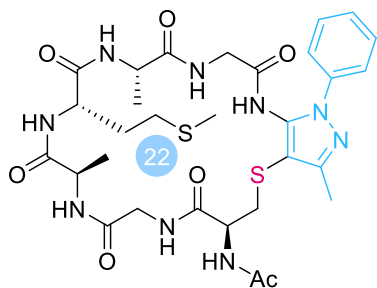
HRMS (ESI-TOF) calcd for $C_{63}H_{92}N_{14}O_{16}S_2$, $[M+H]^+$, 1365.6330, found 1365.6326.



HPLC Spectra:



4.5 Electrochemical peptides cyclization



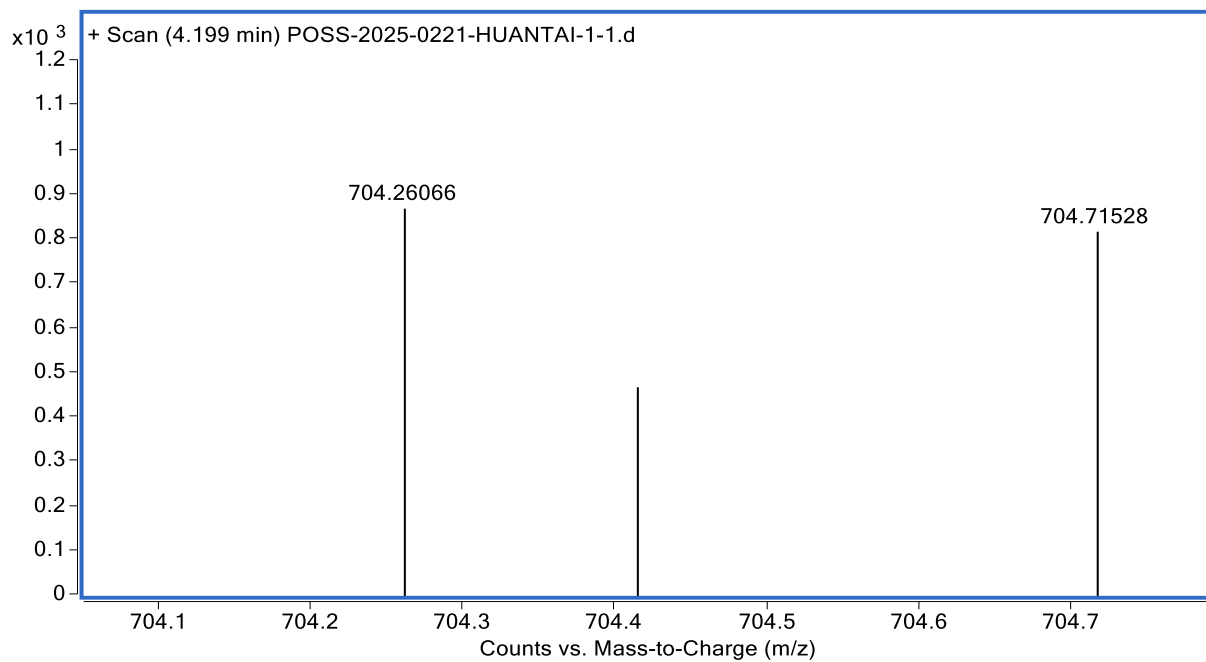
52^o, 85% conv

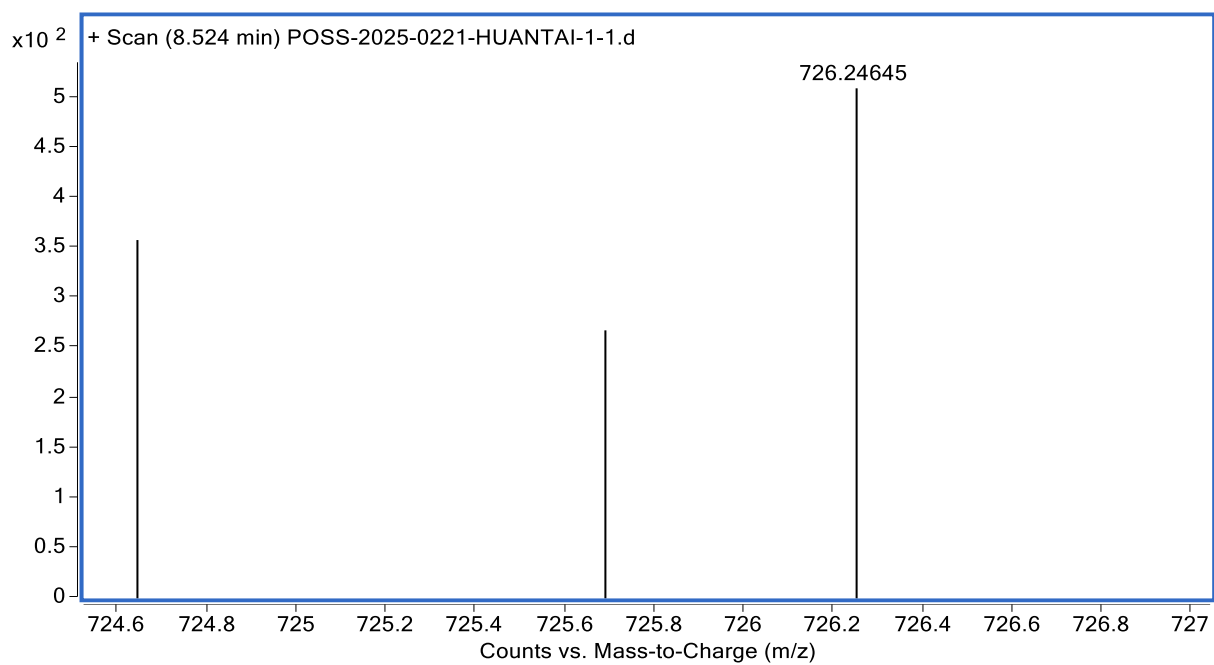
75% (after HPLC purification)

HPLC: 85% conversion.

After the reaction was completed, a peak was observed with a retention time of 5.88 minutes. The polypeptide corresponds to the peak eluting at a retention time of 5.96 minutes.

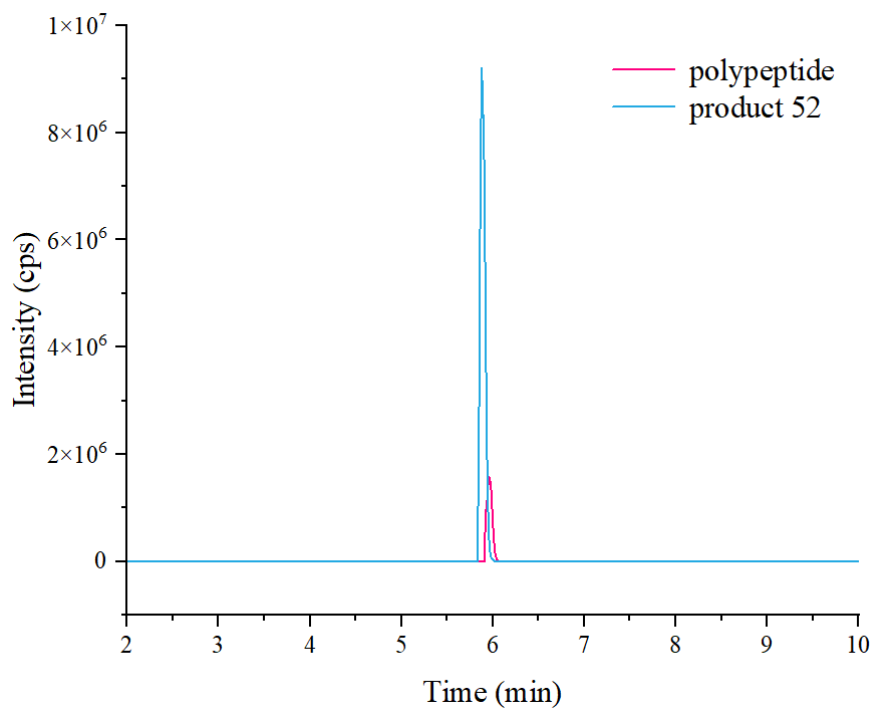
HRMS (ESI-TOF) calcd for $C_{30}H_{41}N_9O_7S_2$, $[M+H]^+$, 704.26431, found 704.26066. calcd for $C_{30}H_{41}N_9O_7S_2$, $[M+Na]^+$, 726.24626, found 726.24645.



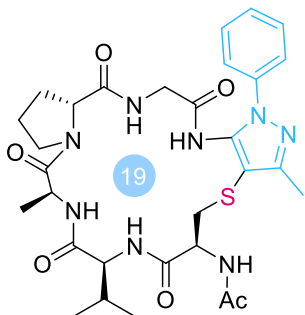
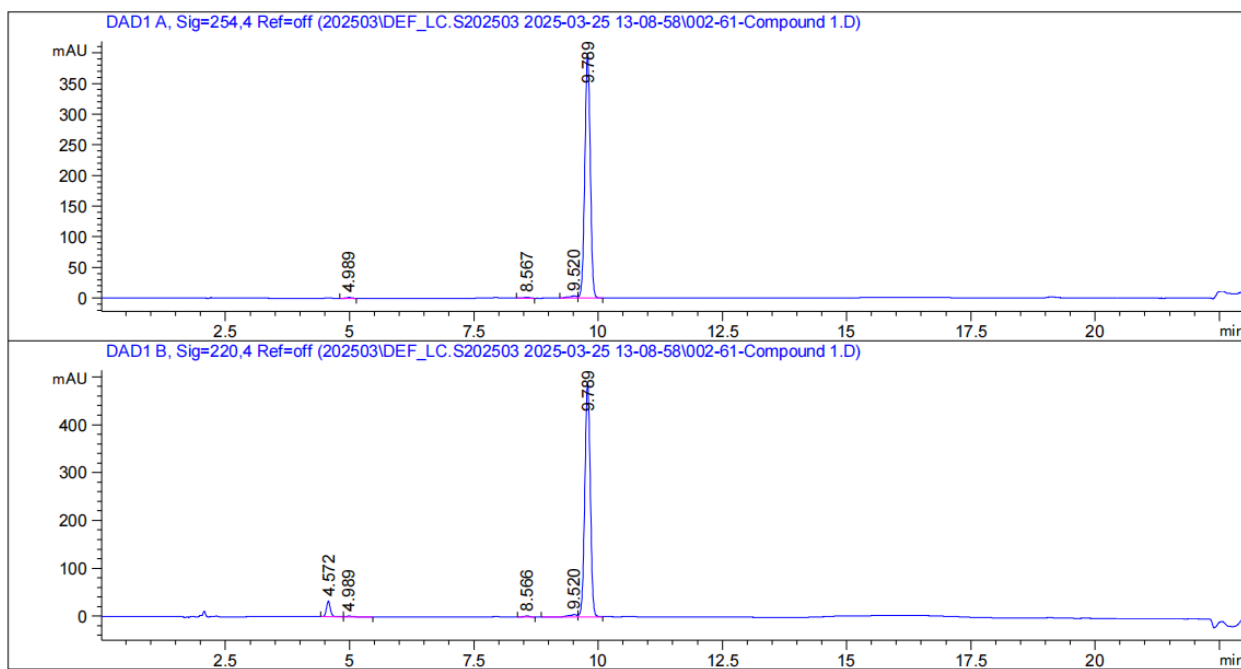


HPLC Spectra:

(1)



(2) After HPLC purification

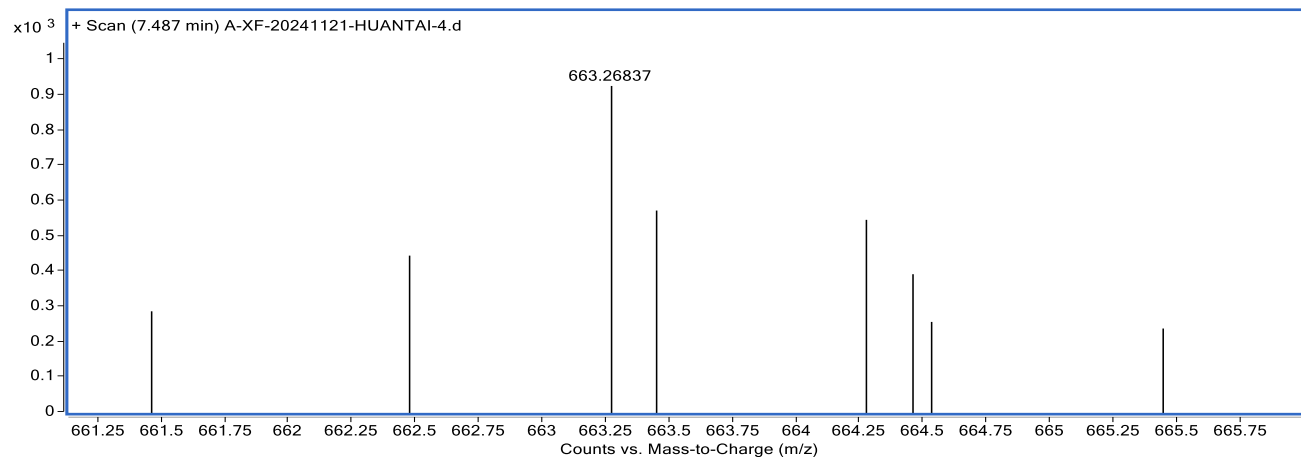
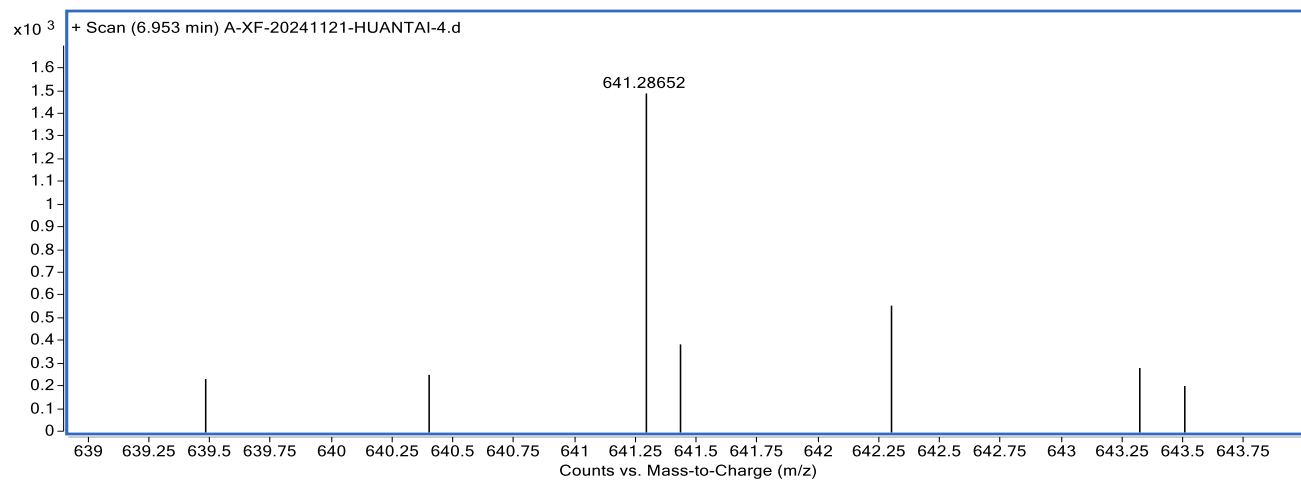


53^e, 32% conv
28% (after HPLC purification)

HPLC: 32% conversion.

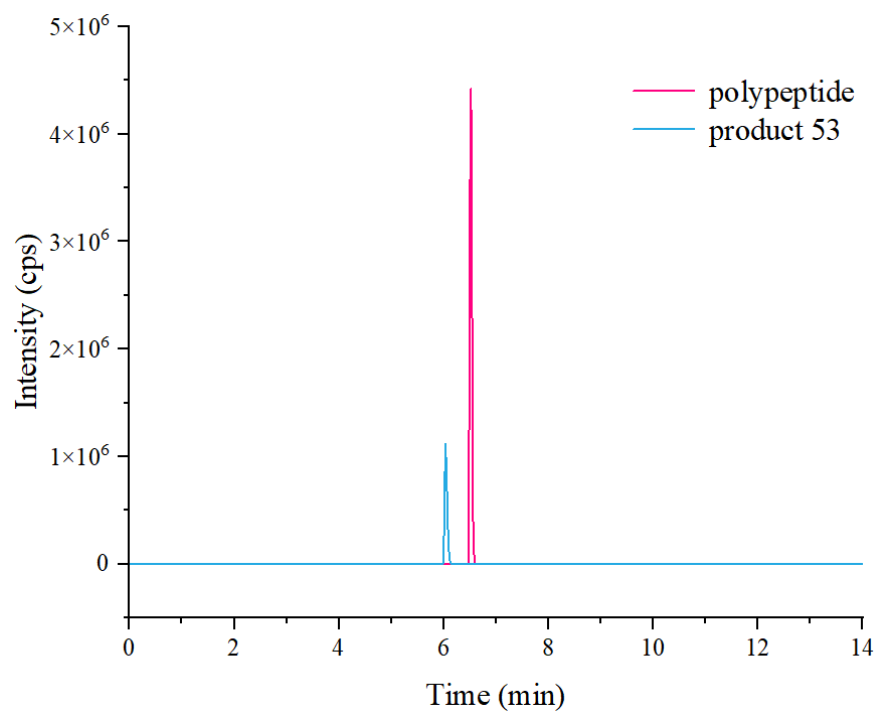
After the reaction was completed, a peak was observed with a retention time of 6.04 minutes. The polypeptide corresponds to the peak eluting at a retention time of 6.52 minutes.

HRMS (ESI-TOF) calcd for $C_{30}H_{40}N_8O_6S$, $[M+H]^+$, 641.28643, found 641.28652. calcd for $C_{30}H_{40}N_8O_6S$, $[M+Na]^+$, 663.26837, found 663.26837.

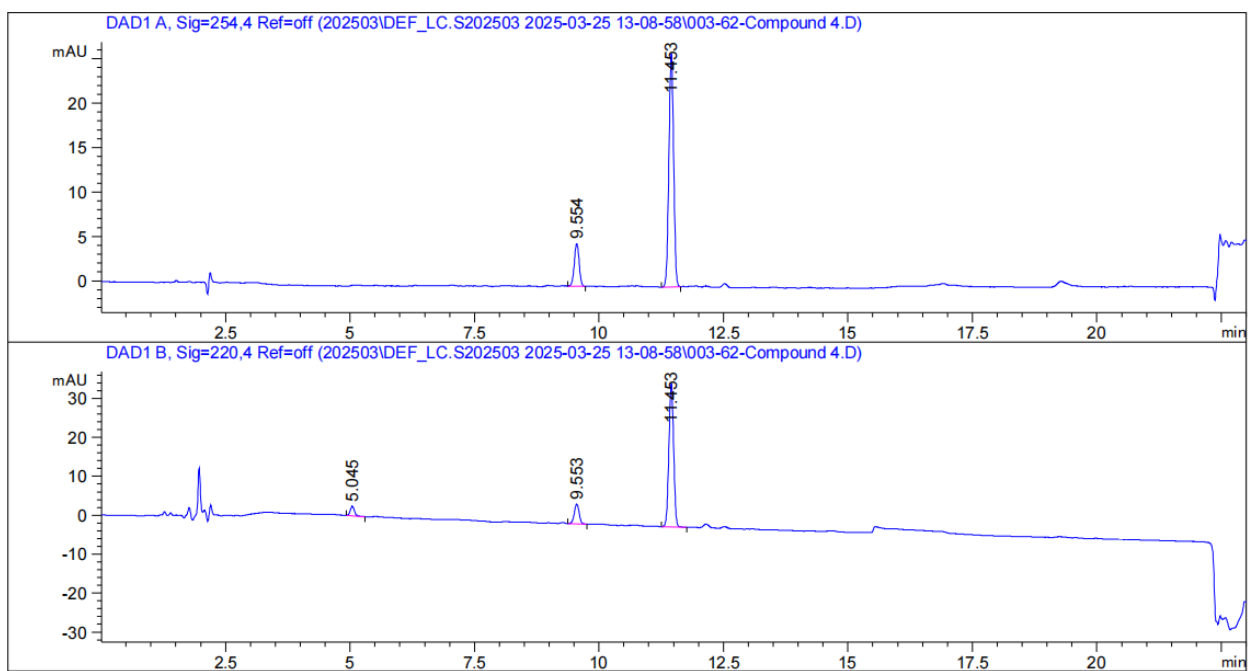


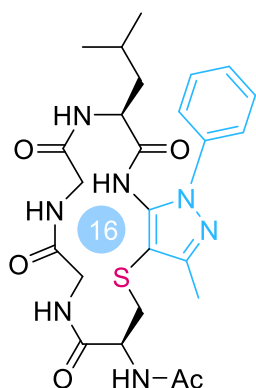
HPLC Spectra:

(1)



(2) After HPLC purification





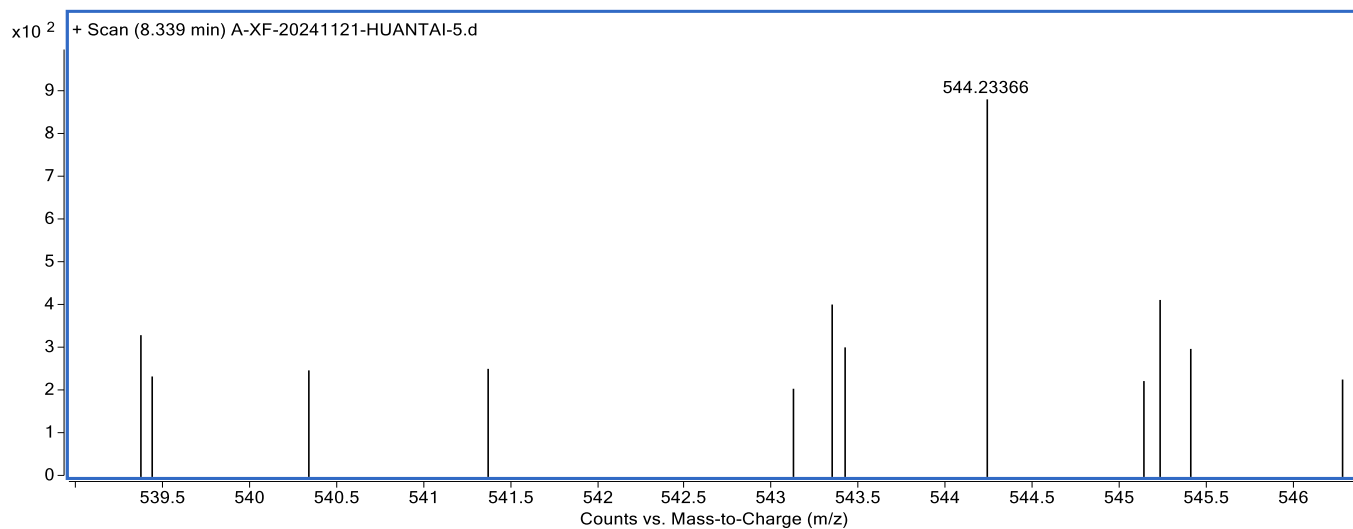
54^e, 98% conv
85% (after HPLC purification)

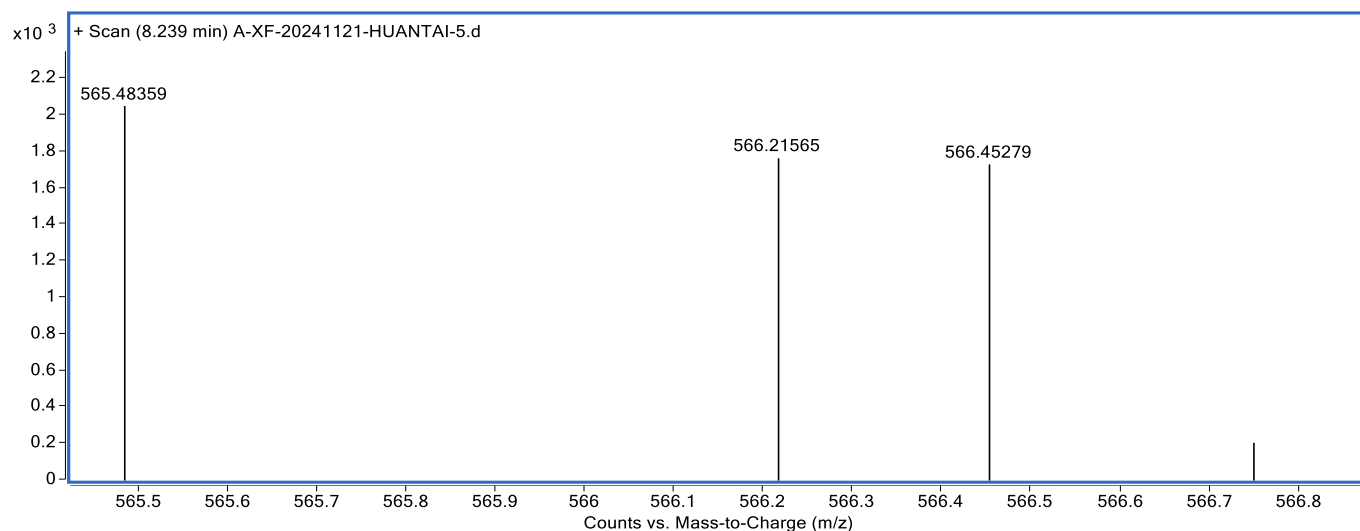
HPLC: 98% conversion.

After the reaction was completed, a peak was observed with a retention time of 6.26 minutes. The polypeptide corresponds to the peak eluting at a retention time of 6.81 minutes.

HRMS (ESI-TOF) calcd for $C_{25}H_{33}N_7O_5S$, $[M+H]^+$, 544.23366, found 544.23366. calcd for

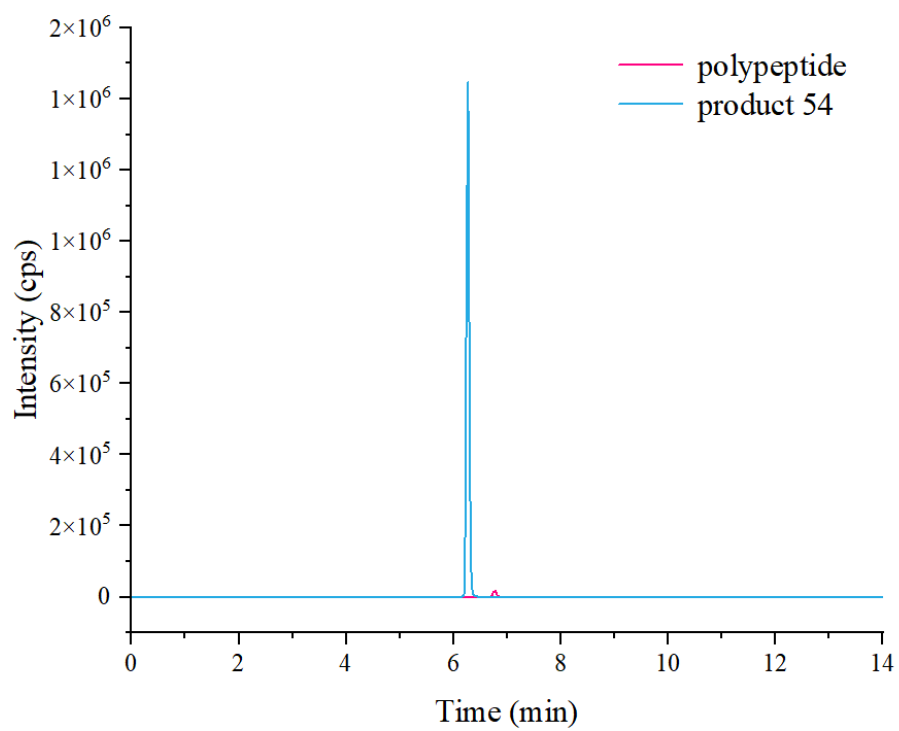
$C_{25}H_{33}N_7O_5S$, $[M+Na]^+$, 566.21561, found 566.21565



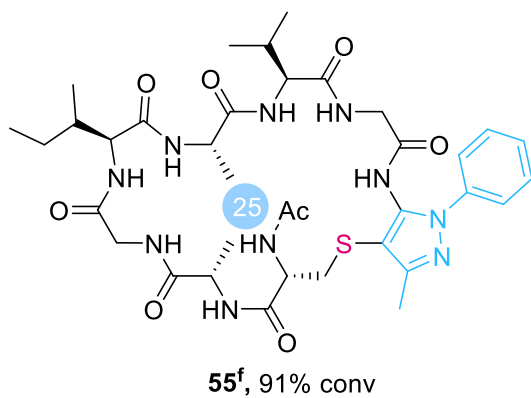
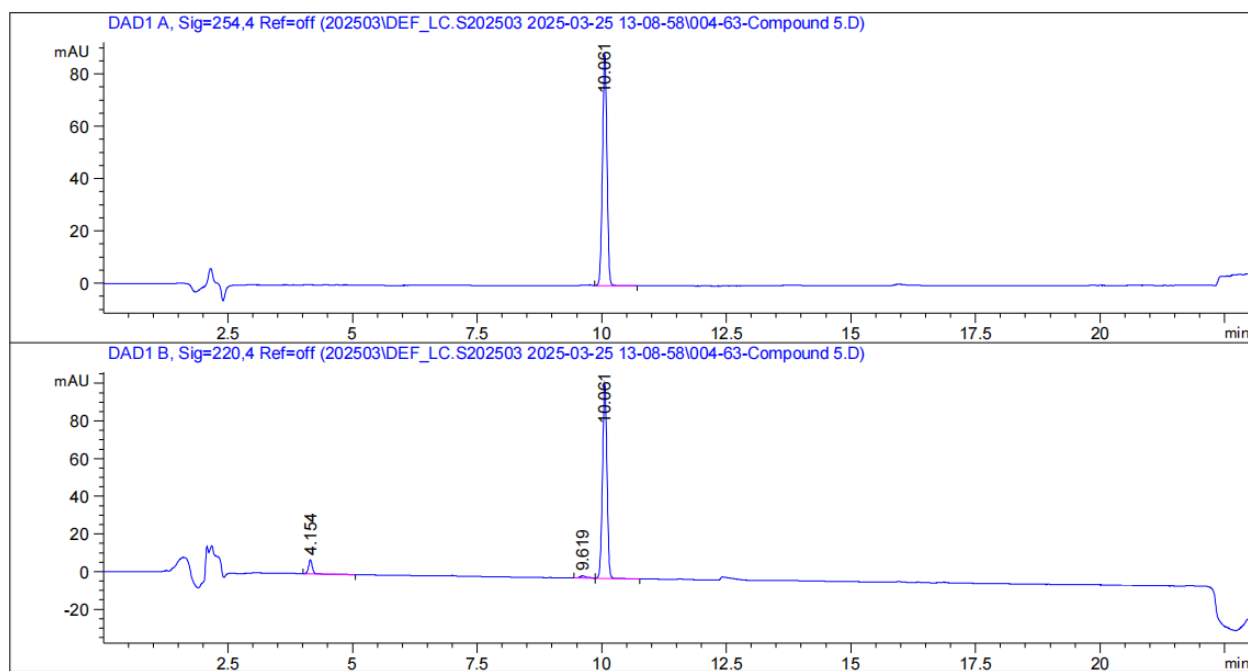


HPLC Spectra:

(1)



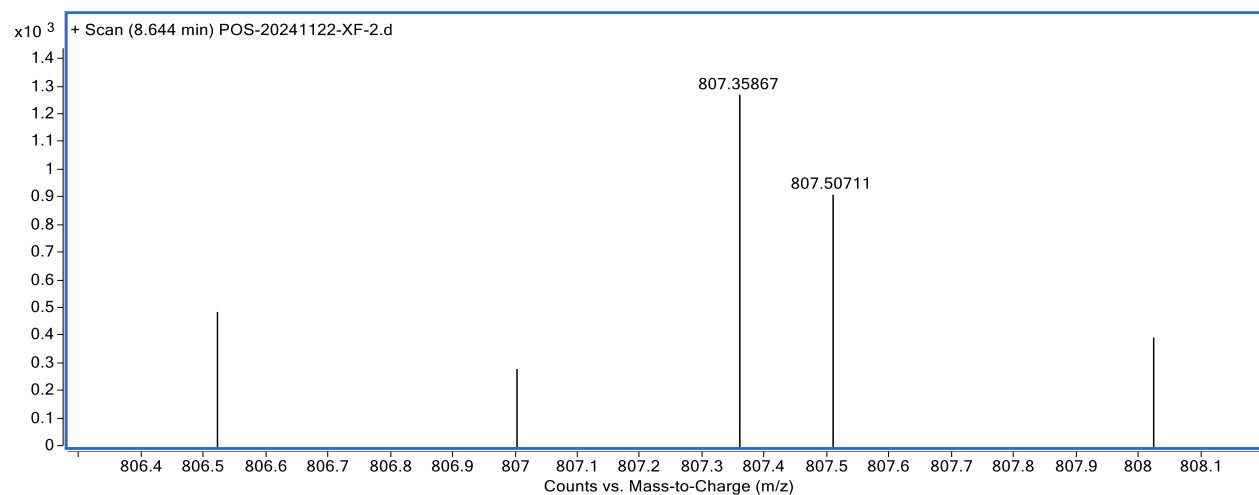
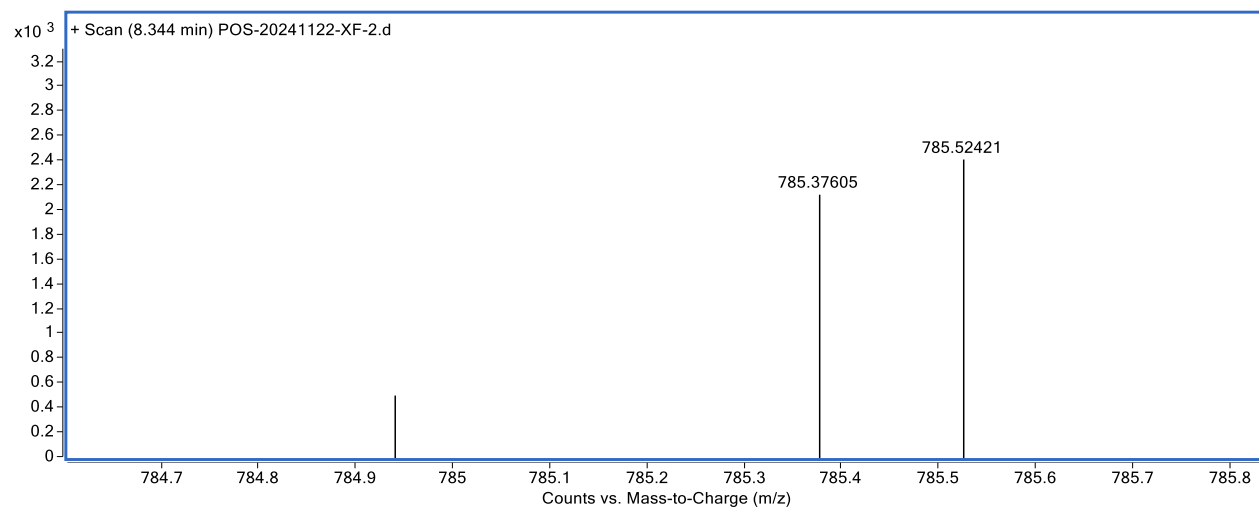
(2) After HPLC purification



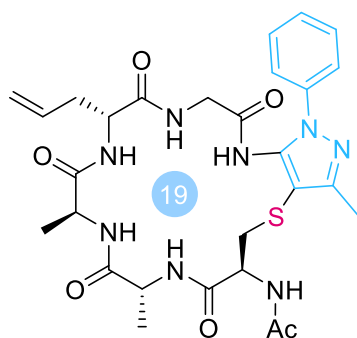
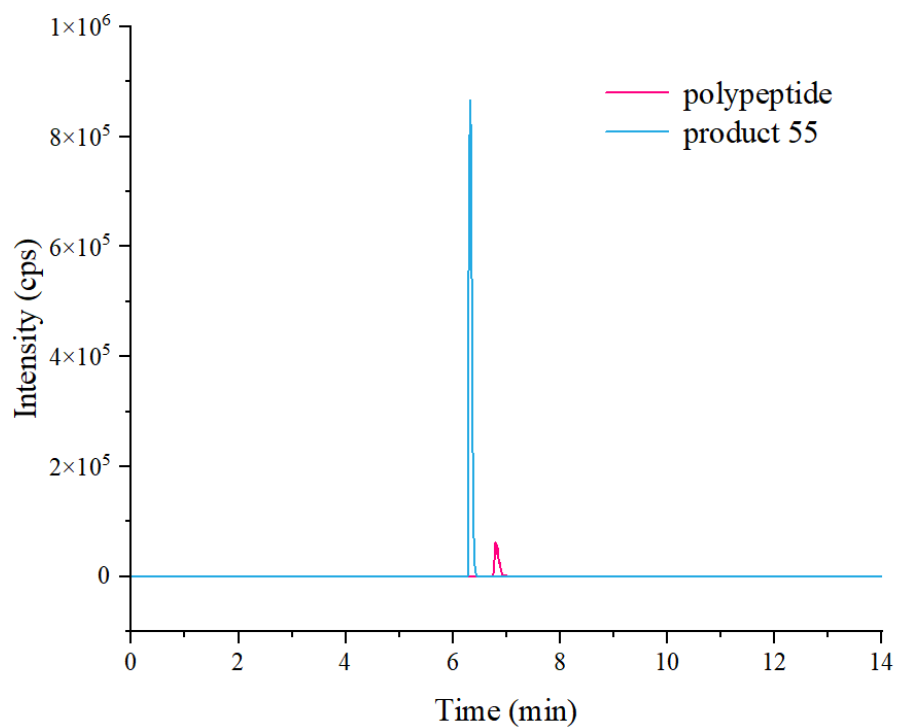
HPLC: 91% conversion.

After the reaction was completed, a peak was observed with a retention time of 6.32 minutes. The polypeptide corresponds to the peak eluting at a retention time of 6.79 minutes.

HRMS (ESI-TOF) calcd for $C_{36}H_{52}N_{10}O_8S$, $[M+H]^+$, 785.37631, found 785.37605. calcd for $C_{36}H_{52}N_{10}O_8S$, $[M+Na]^+$, 807.35825, found 807.35867.



HPLC Spectra:

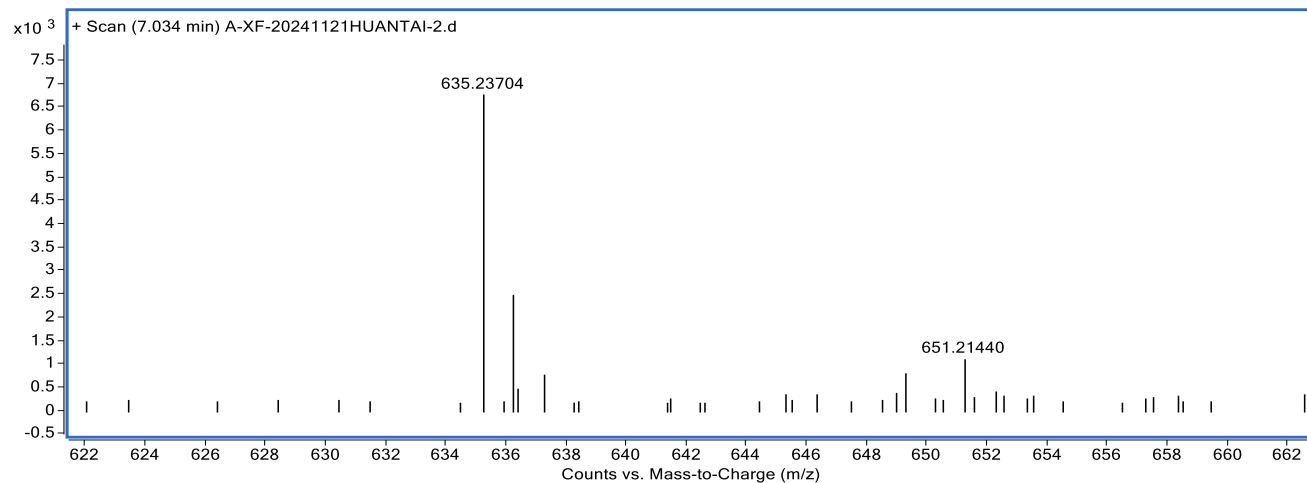
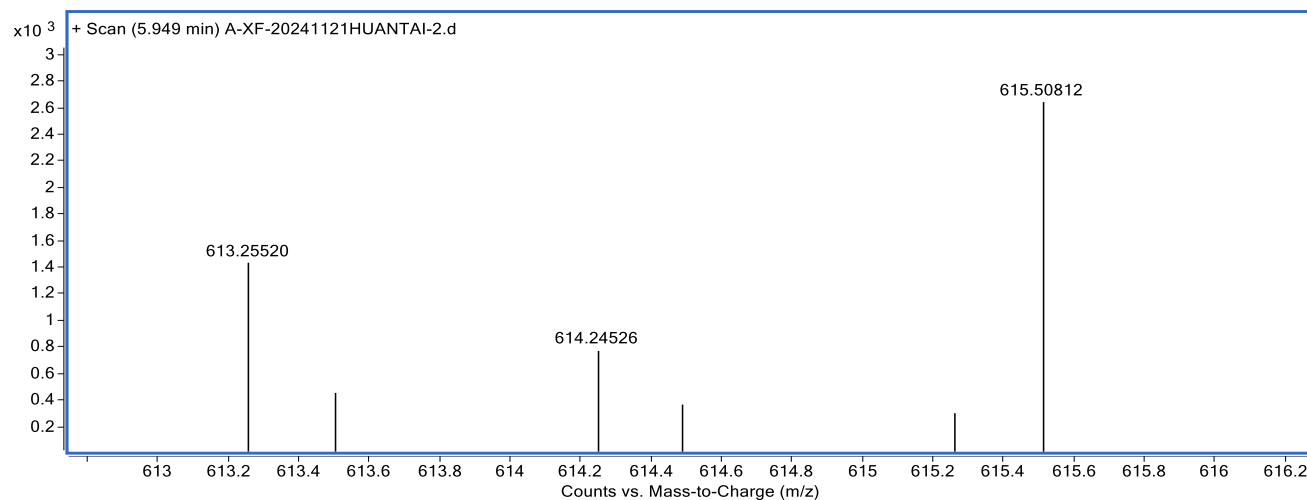


56^f, 60% conv

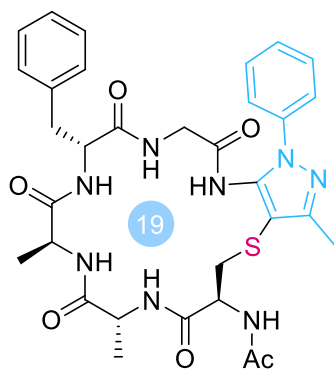
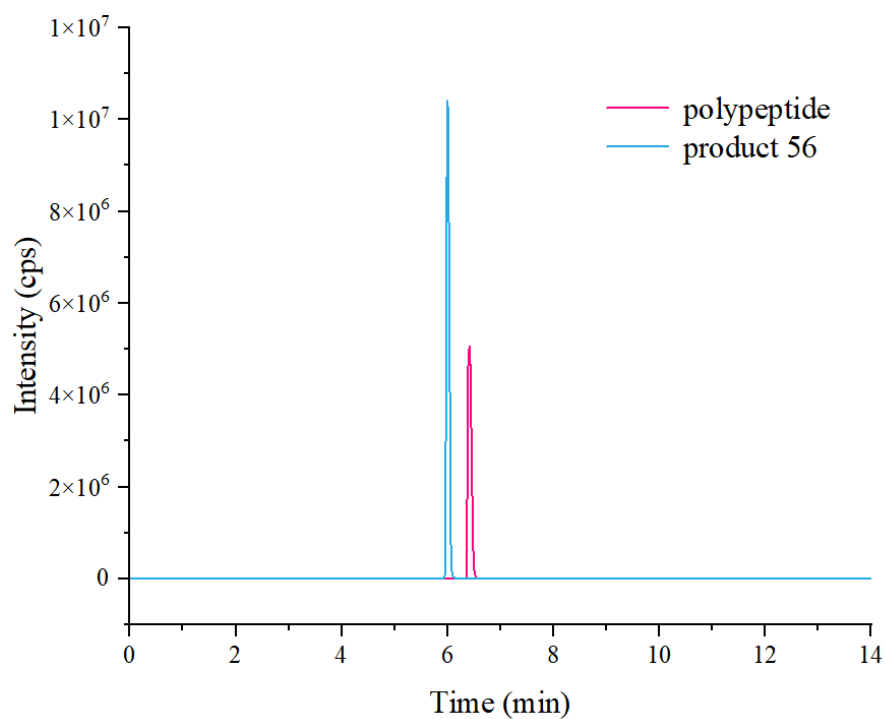
HPLC: 60% conversion.

After the reaction was completed, a peak was observed with a retention time of 6.00 minutes. The polypeptide corresponds to the peak eluting at a retention time of 6.41 minutes.

HRMS (ESI-TOF) calcd for $C_{28}H_{36}N_8O_6S$, $[M+H]^+$, 613.25513, found 613.25520. calcd for $C_{28}H_{36}N_8O_6S$, $[M+Na]^+$, 635.23707, found 635.23704.



HPLC Spectra:

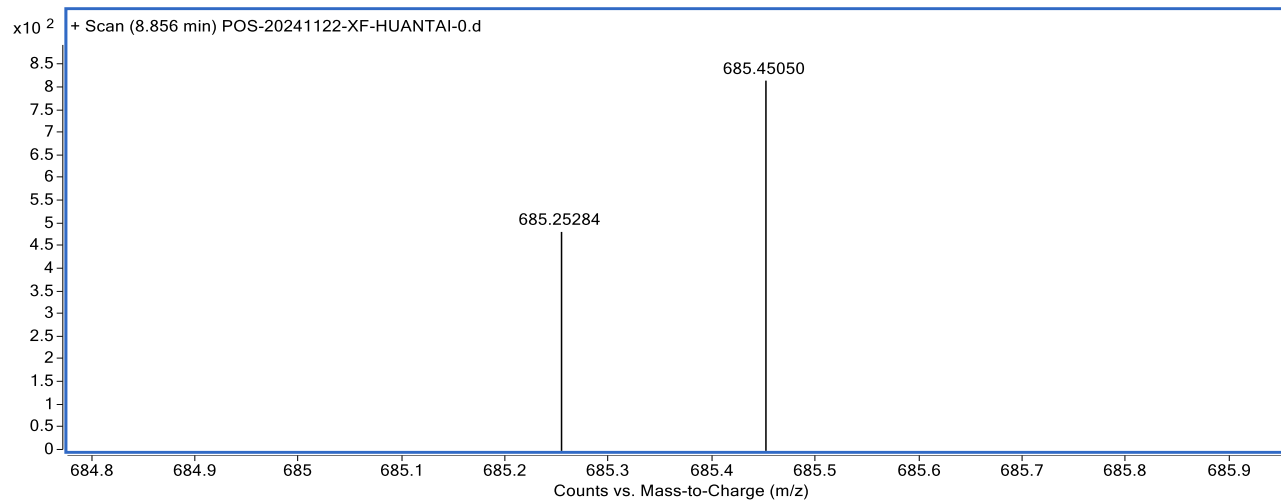
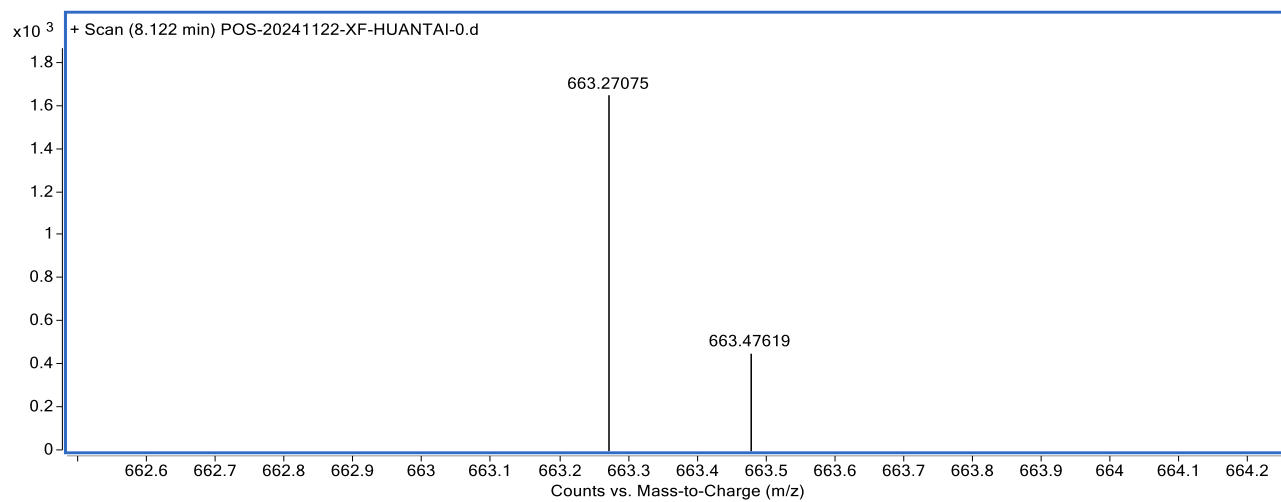


57^f, 50% conv

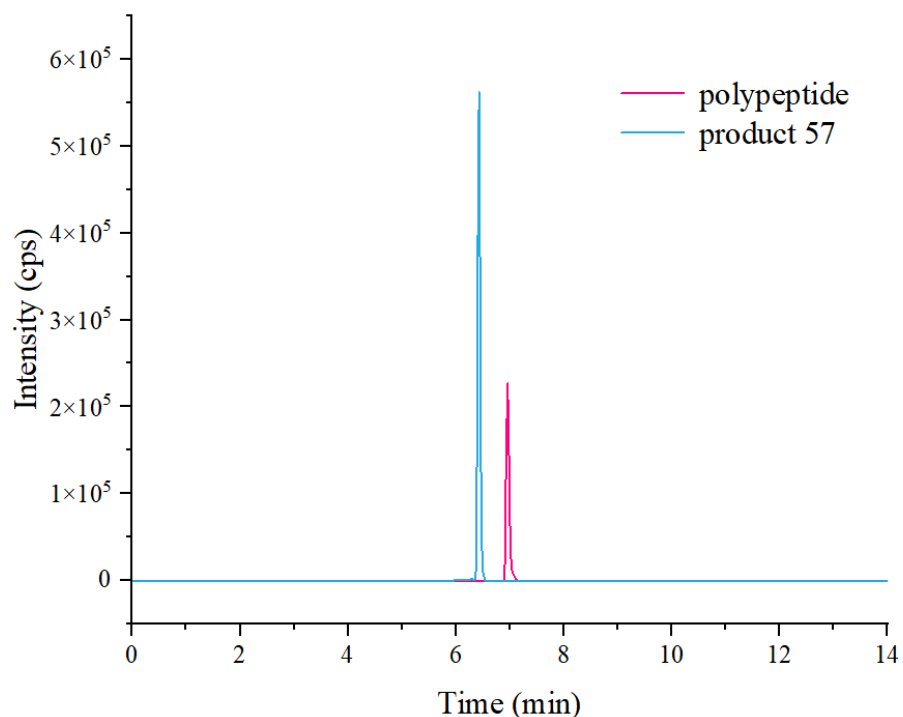
HPLC: 50% conversion.

After the reaction was completed, a peak was observed with a retention time of 6.43 minutes. The polypeptide corresponds to the peak eluting at a retention time of 6.96 minutes.

HRMS (ESI-TOF) calcd for $C_{32}H_{38}N_8O_6S$, $[M+H]^+$, 663.27078, found 663.27075. calcd for $C_{32}H_{38}N_8O_6S$, $[M+Na]^+$, 685.25272, found 685.25284.



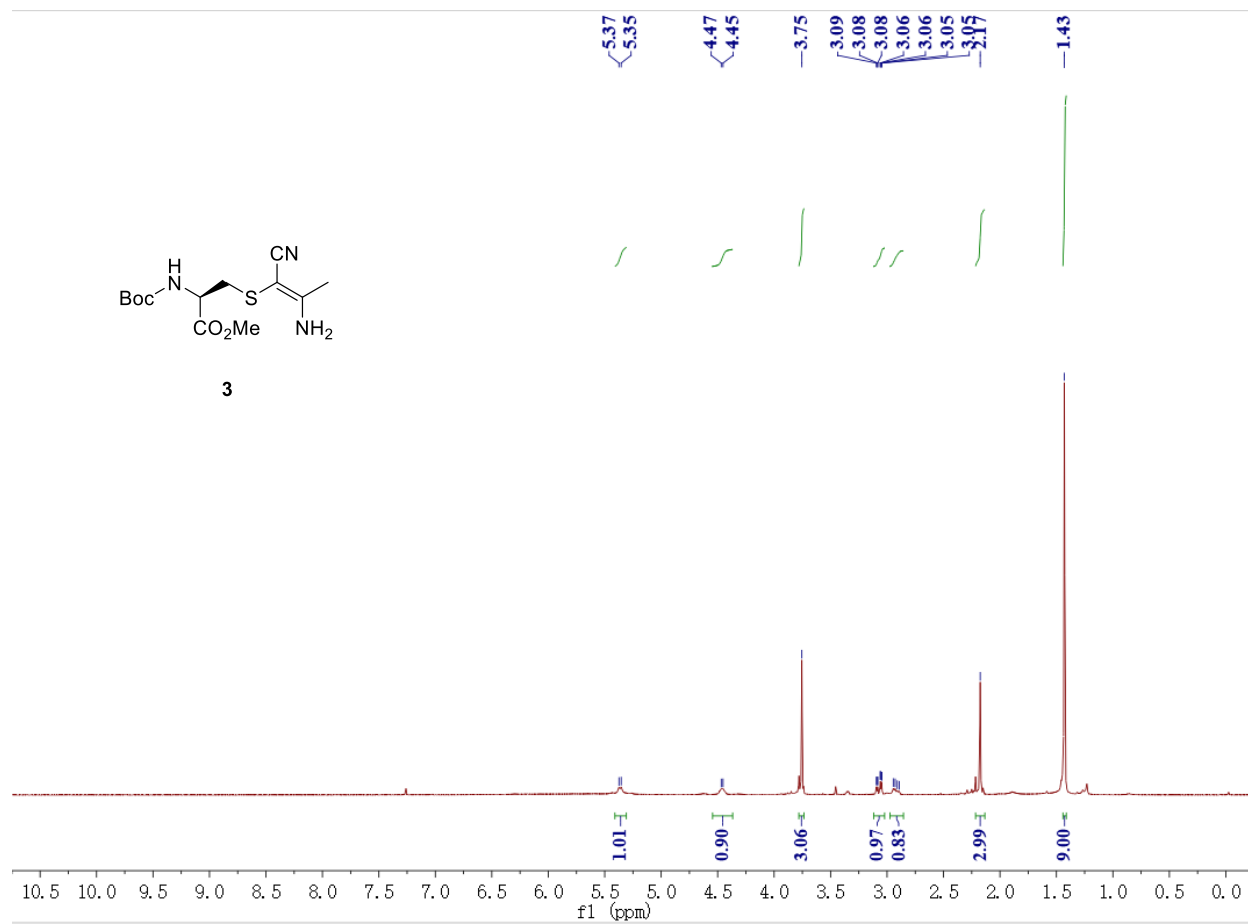
HPLC Spectra:

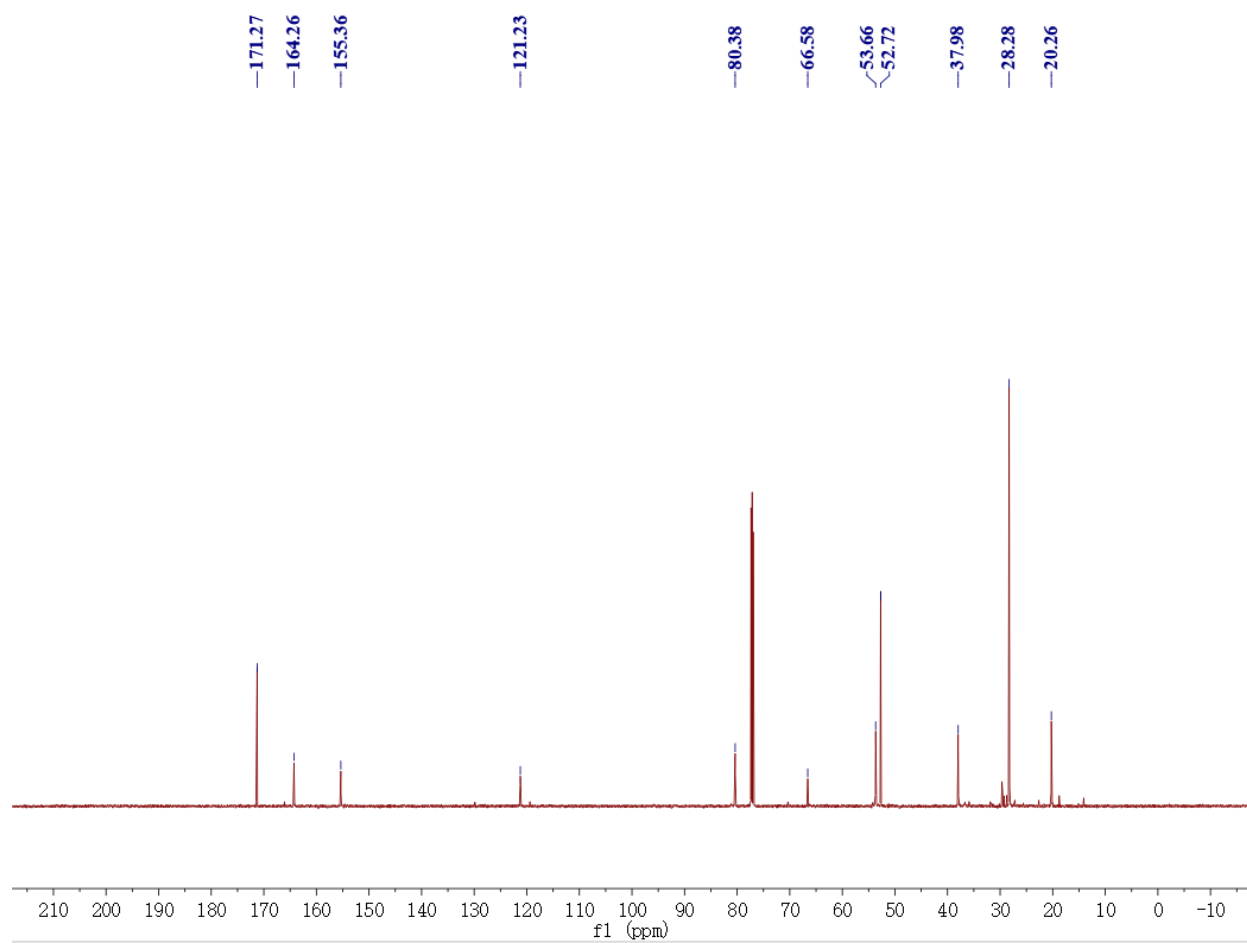


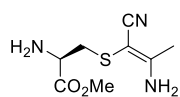
5. References

- [1] R. A. Serwa, J.-M. Swiecicki, D. Homann, C. P. R. Hackenberger, Phosphoramidate-peptide synthesis by solution- and solid-phase Staudinger-phosphite reactions. *J. Pept. Sci.* **2010**, *16*, 563–567.
- [2] Alam J, Keller T H, Loh T P. Functionalization of peptides and proteins by Mukaiyama aldol reaction. *J. Am. Chem. Soc.* **2010**, *132*, 9546-9548.
- [3] Bottecchia C., Erdmann N., Tijssen P.M.A., et al. Batch and Flow Synthesis of Disulfides by Visible-Light-Induced TiO_2 Photocatalysis [J]. *ChemSusChem*, **2016**, *9*(14): 1781-5.

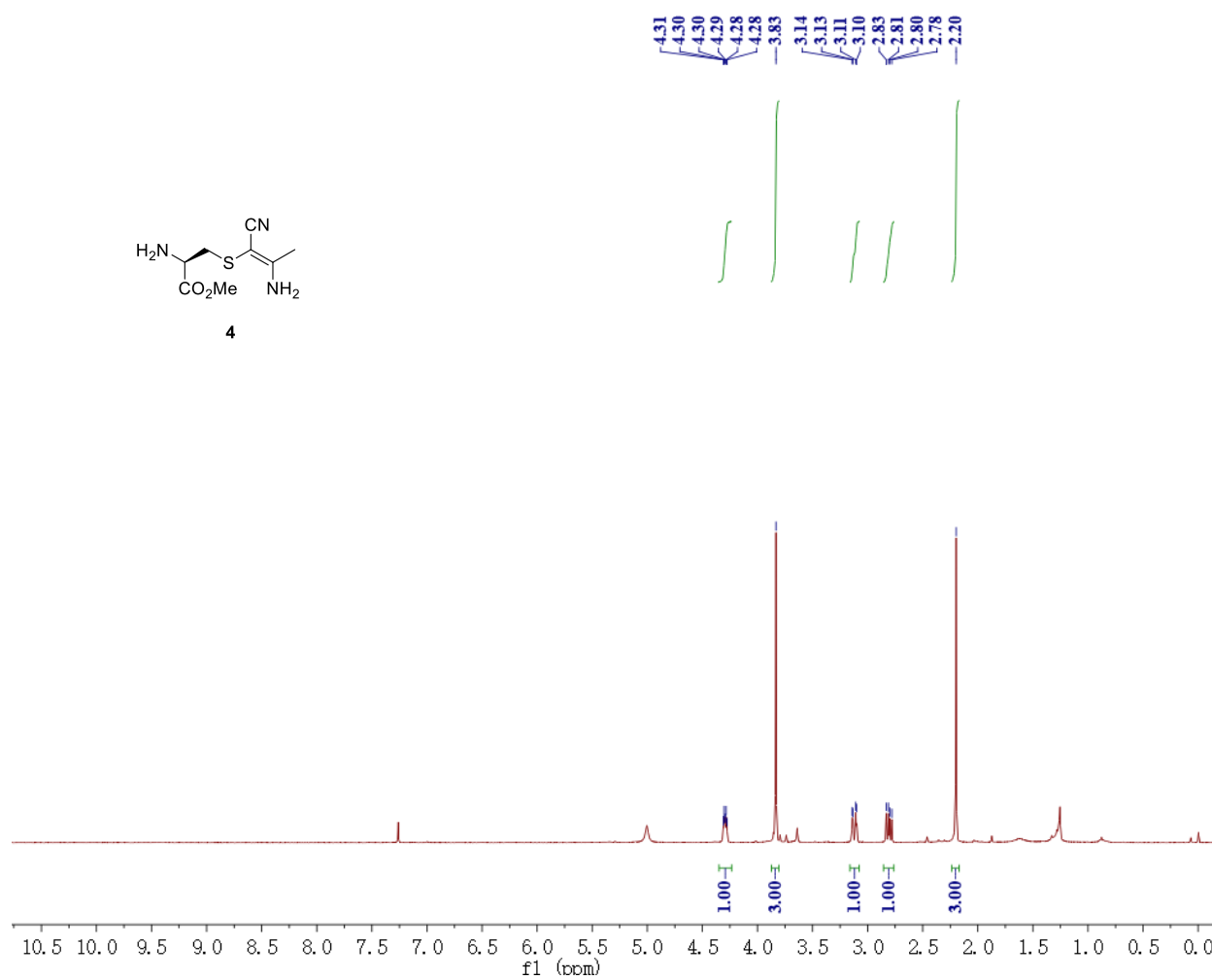
6. NMR spectra of all products

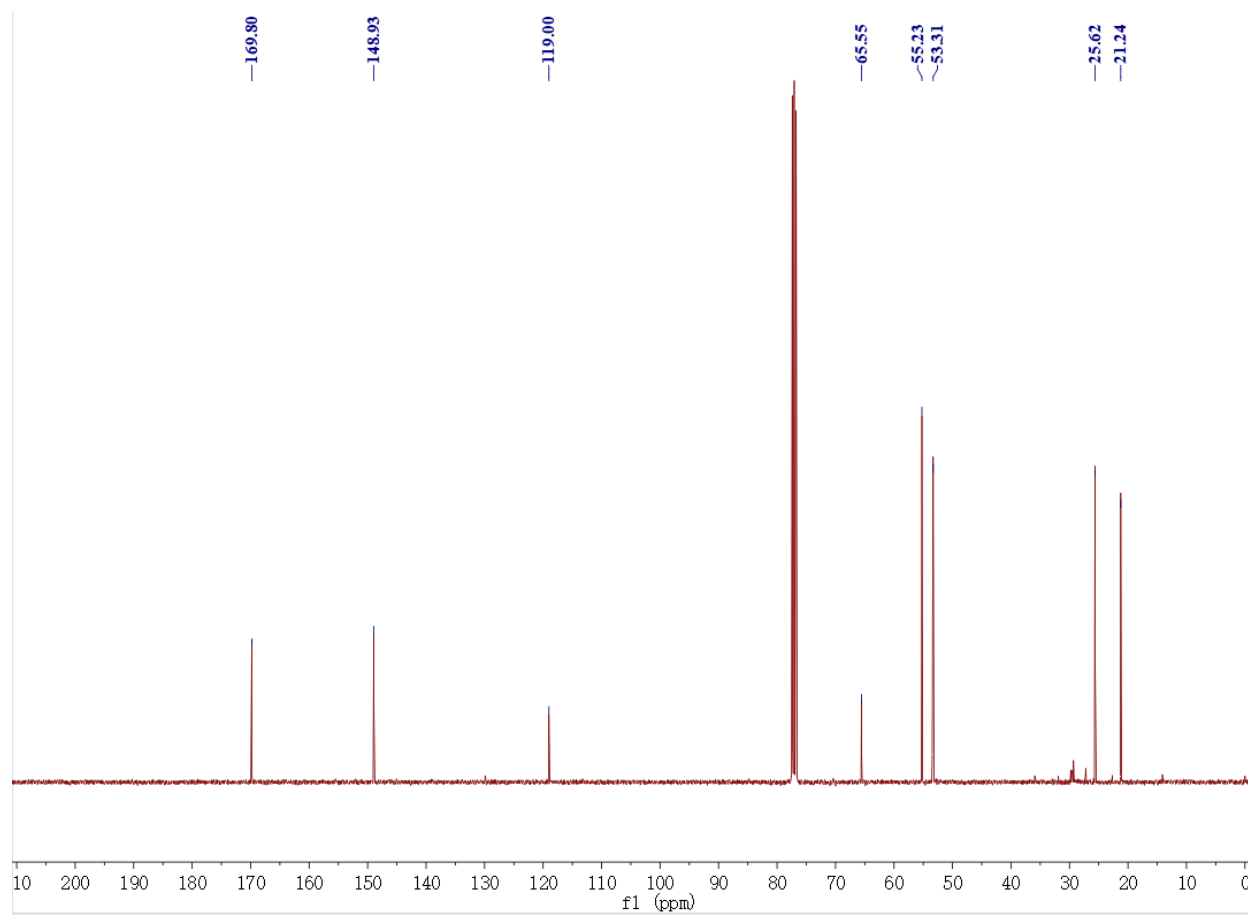


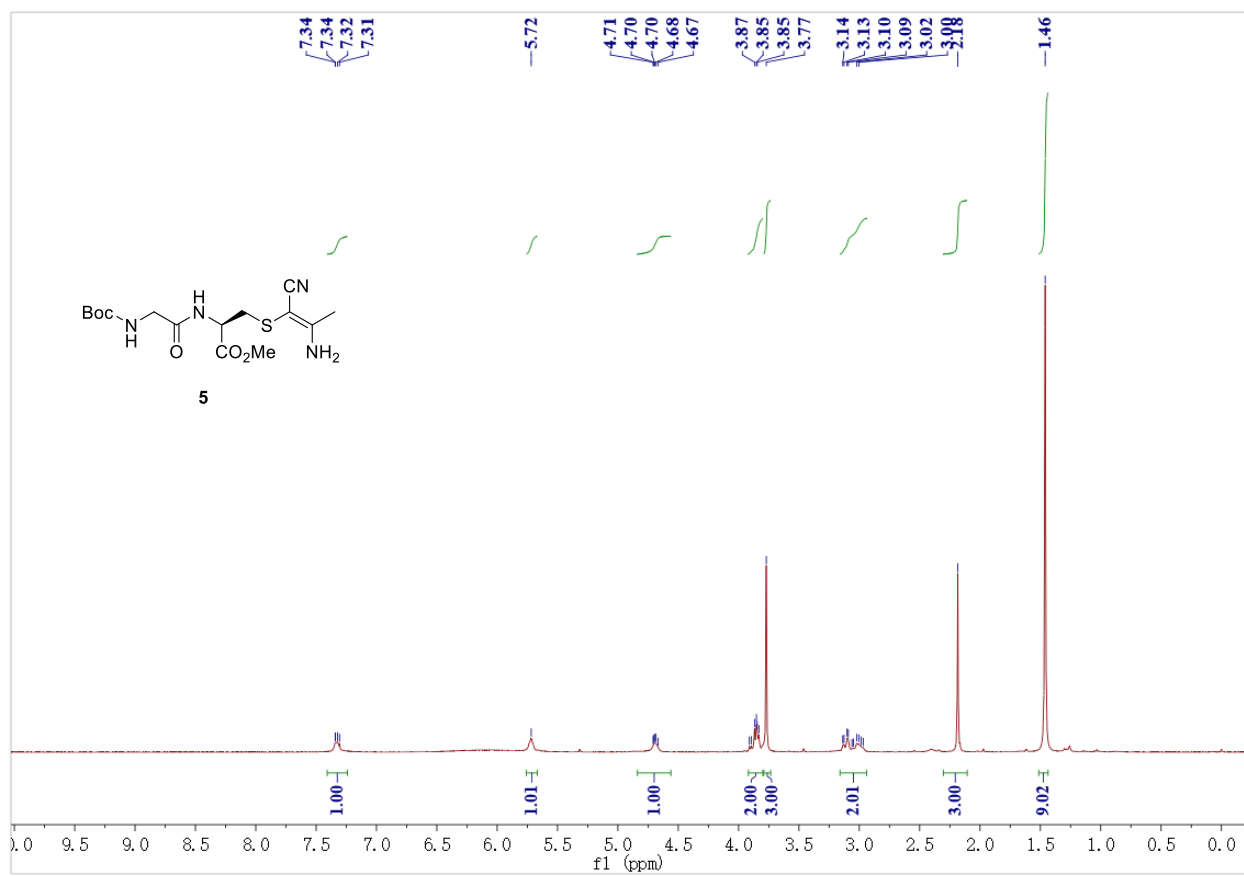


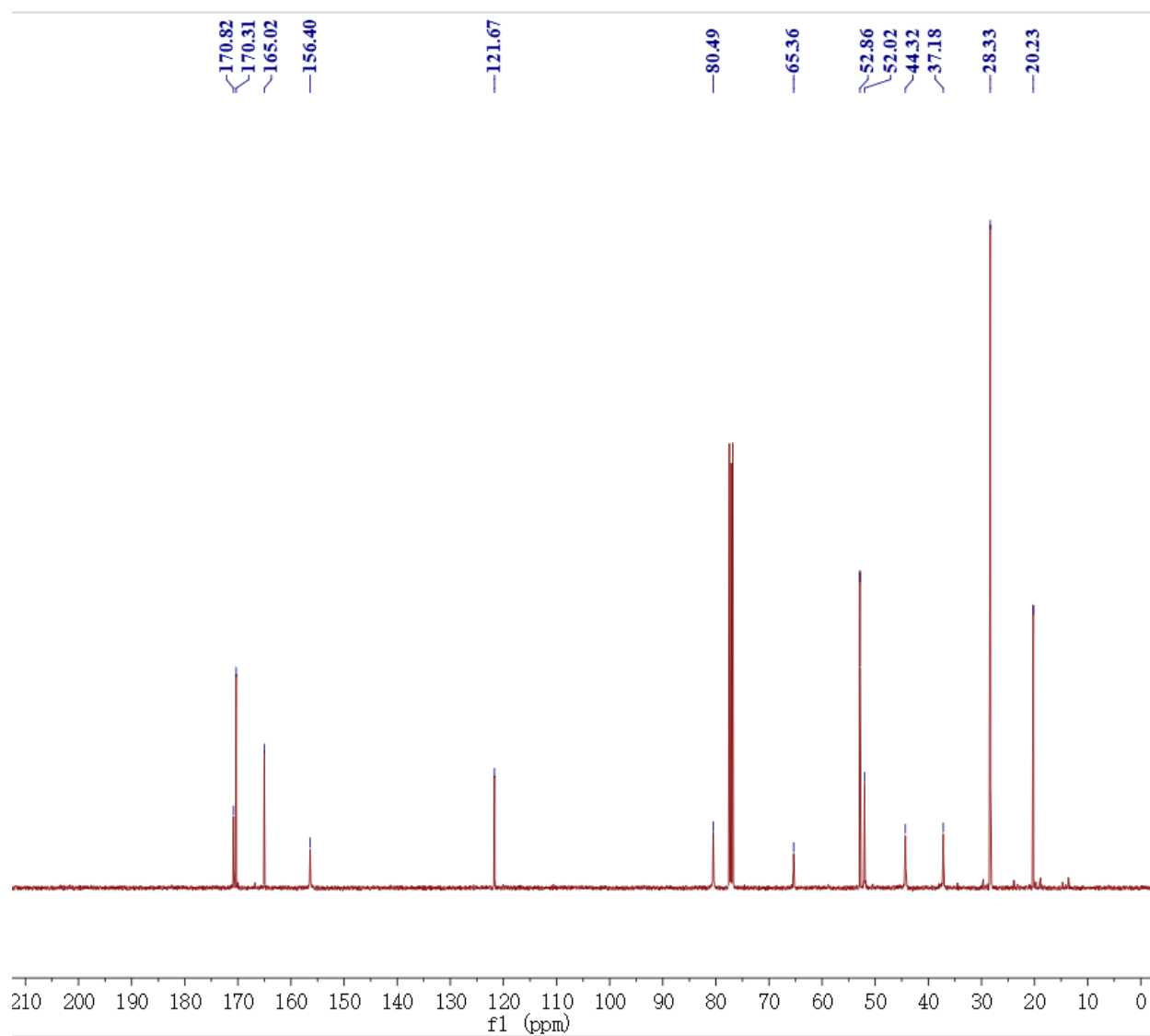


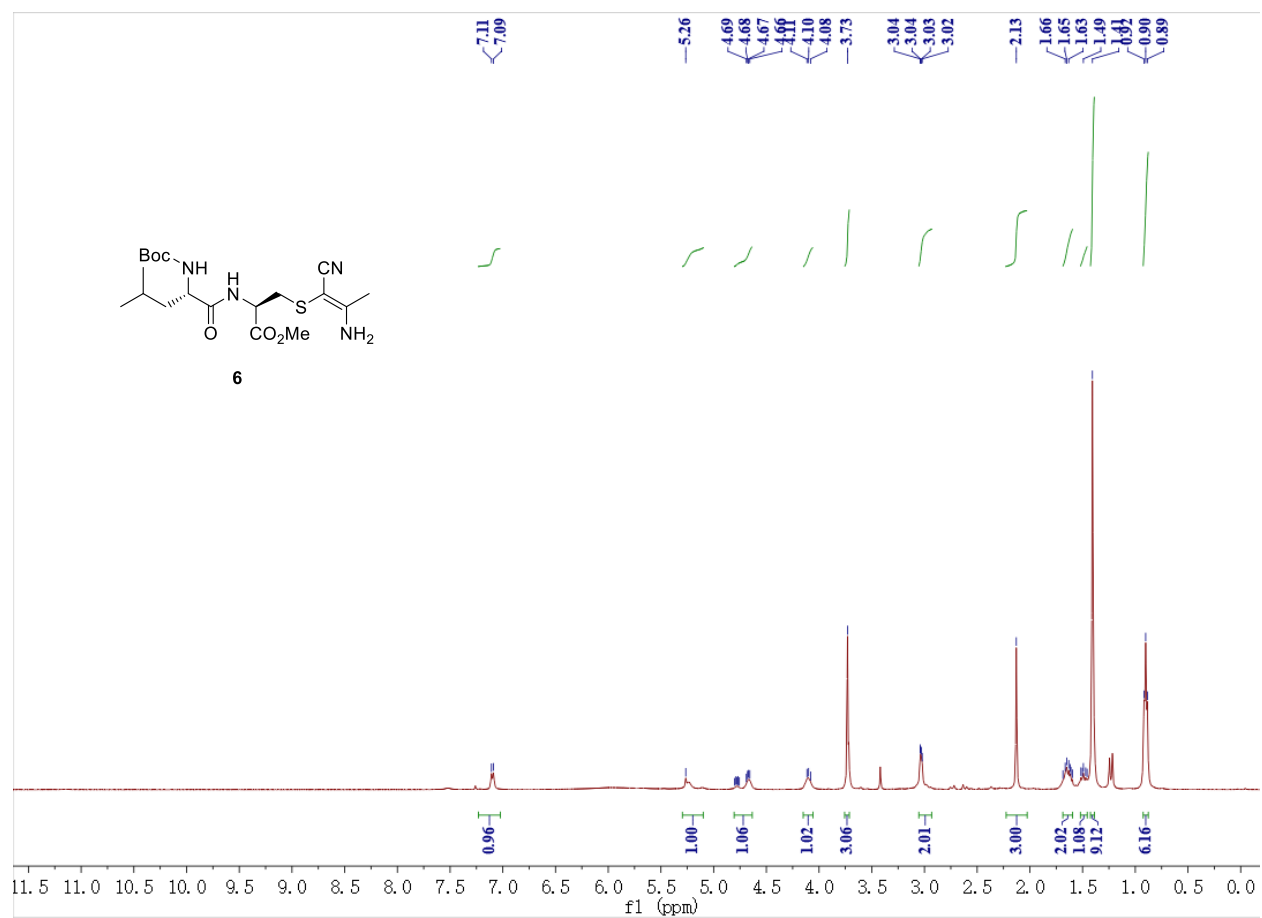
4

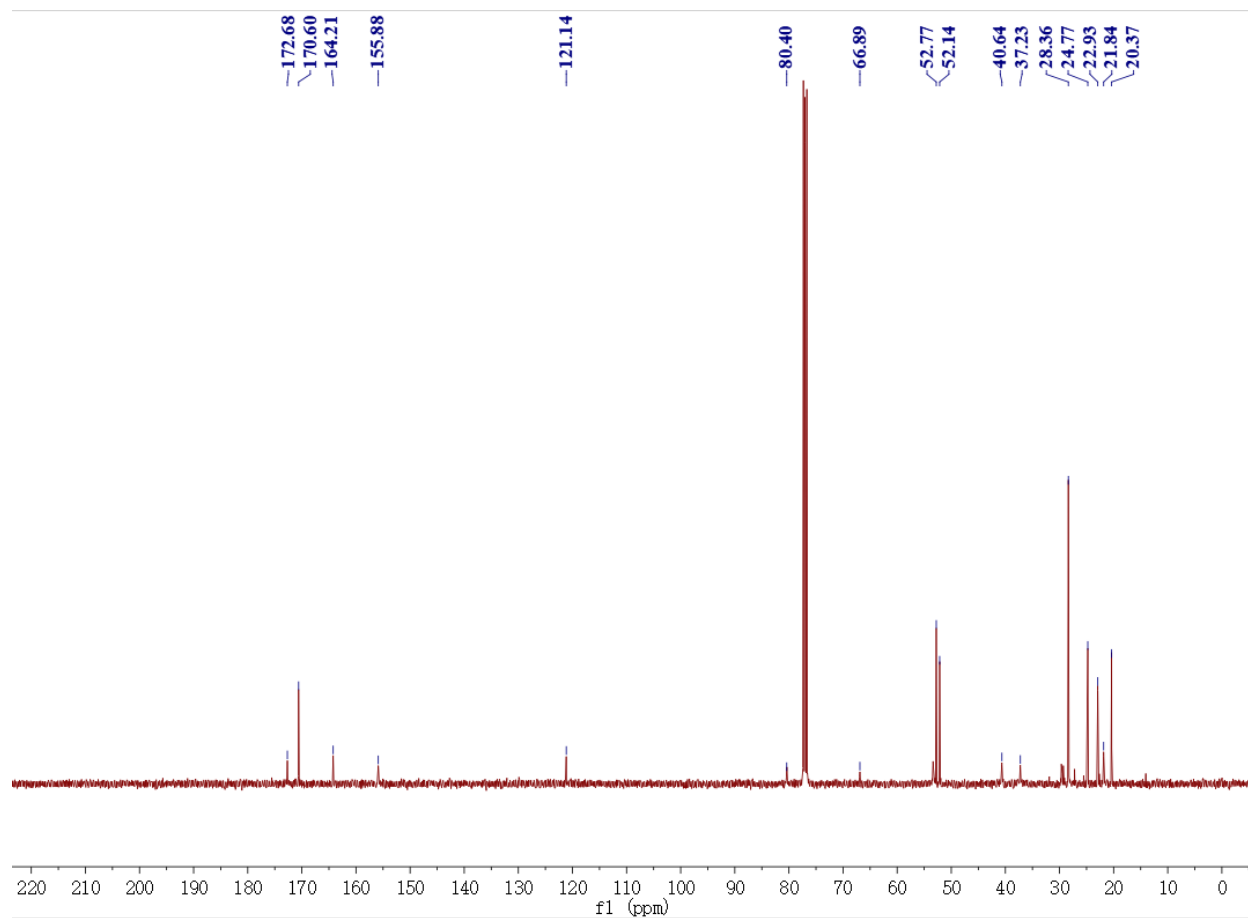


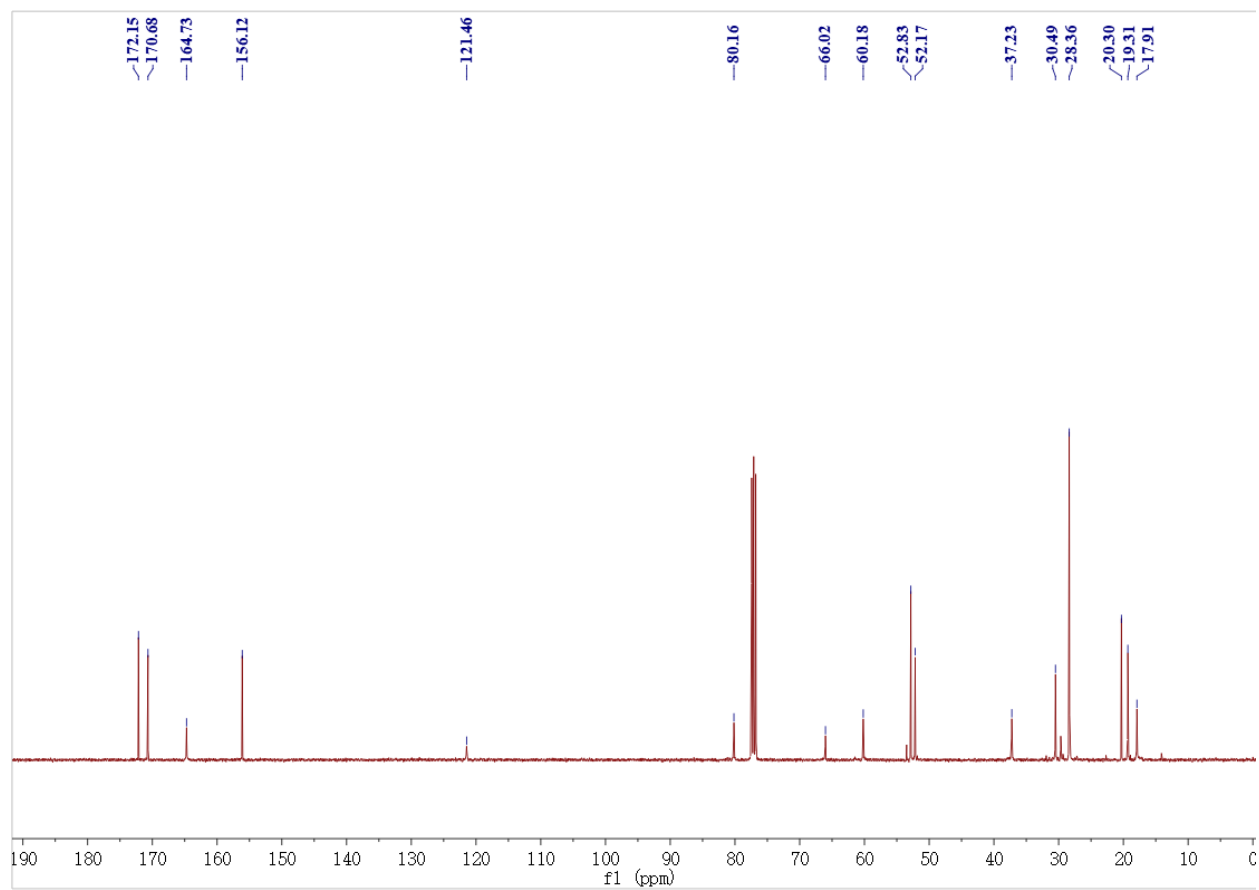


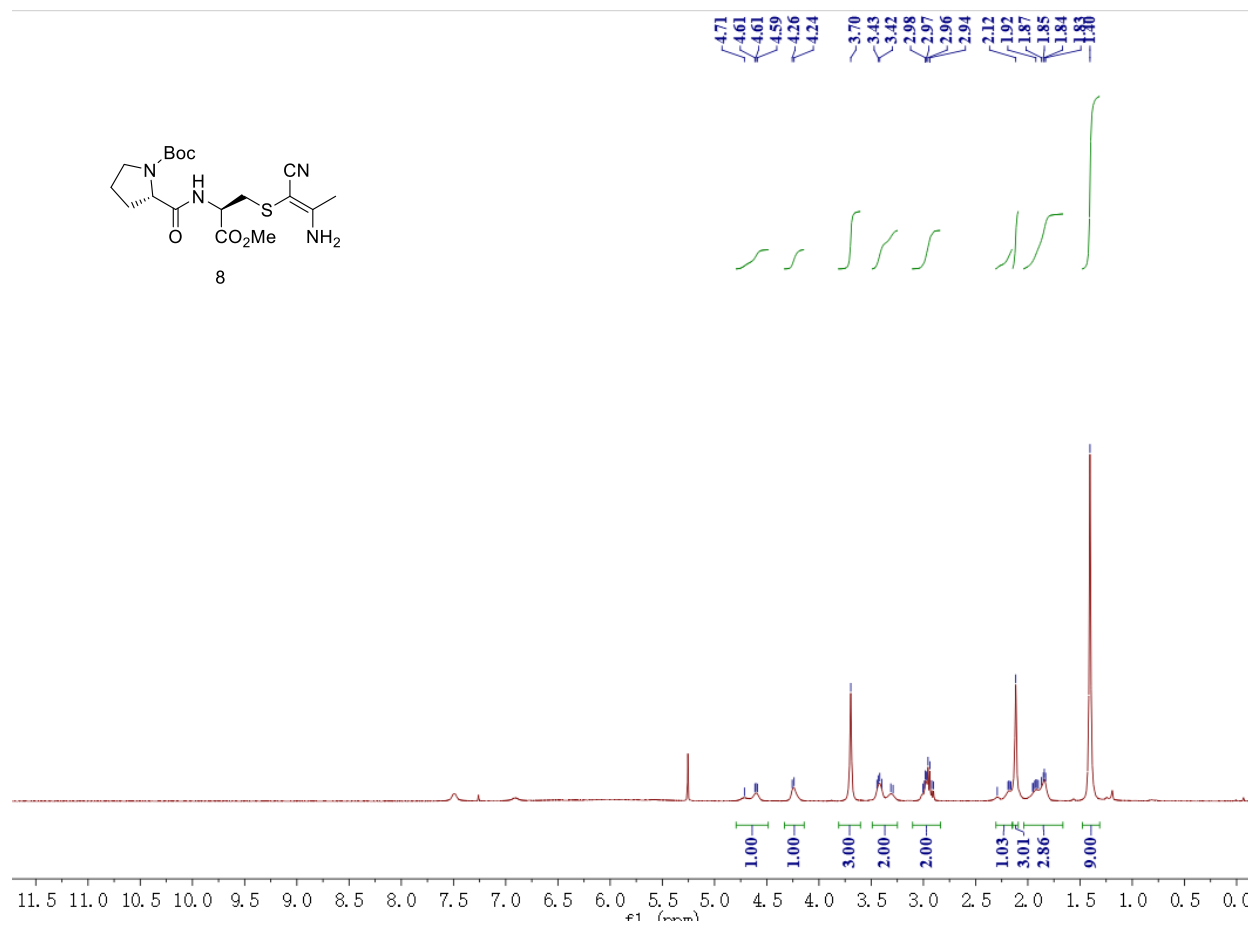


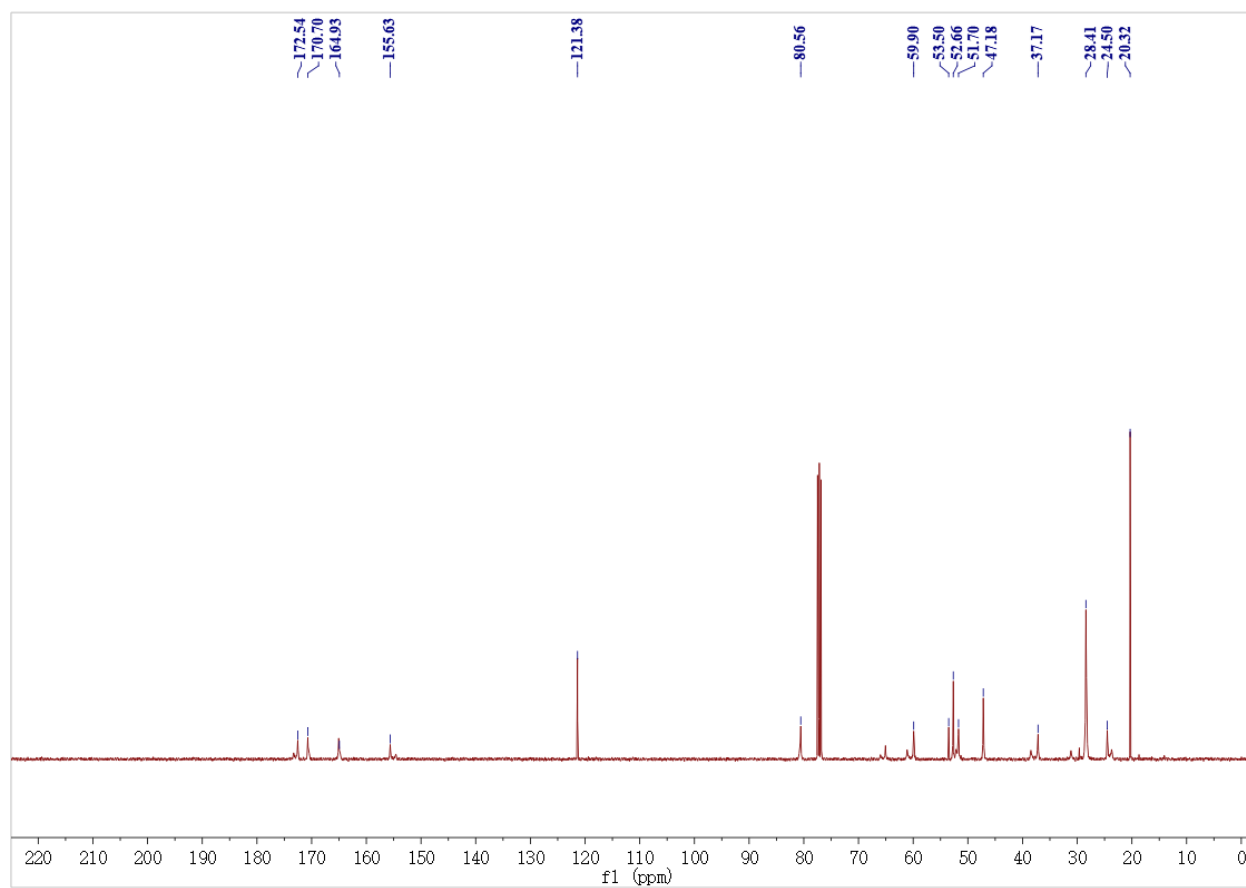


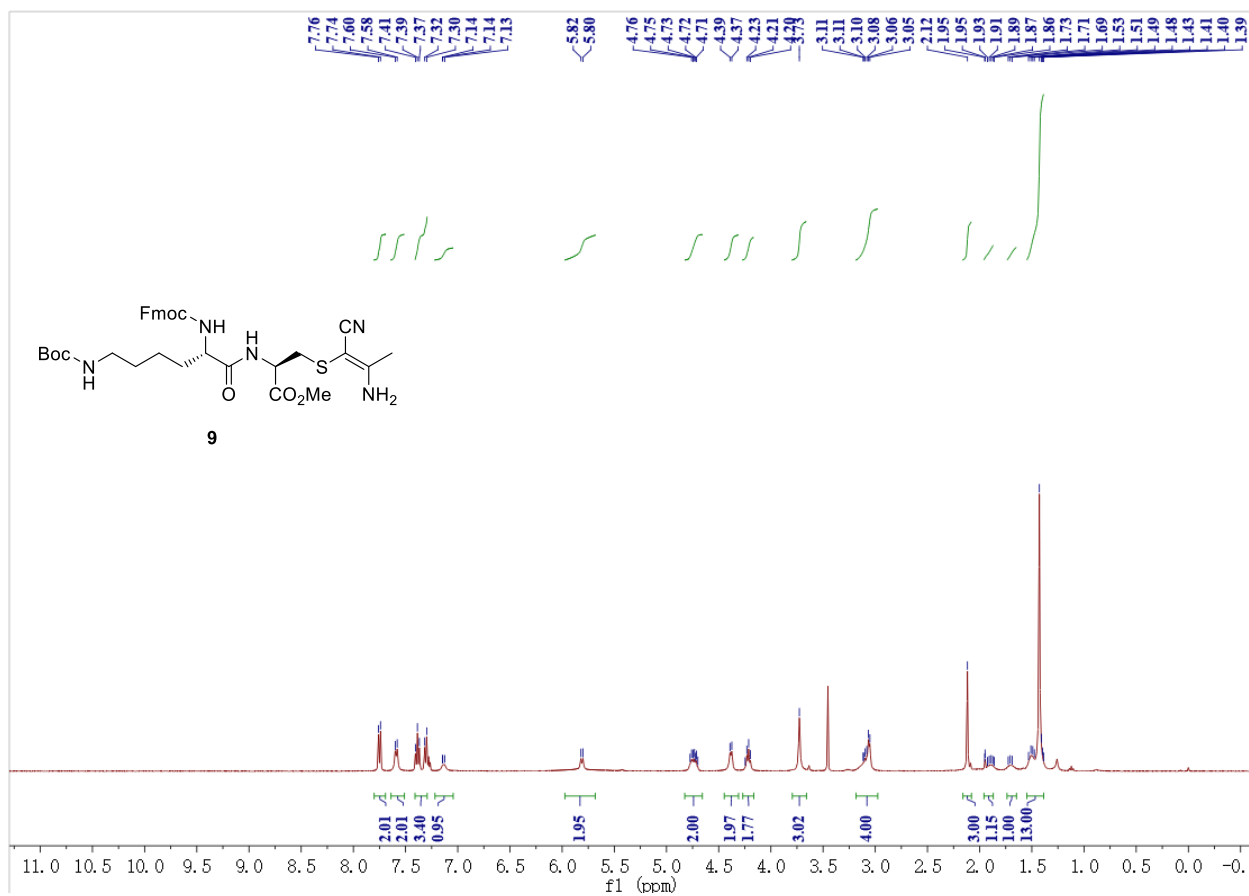


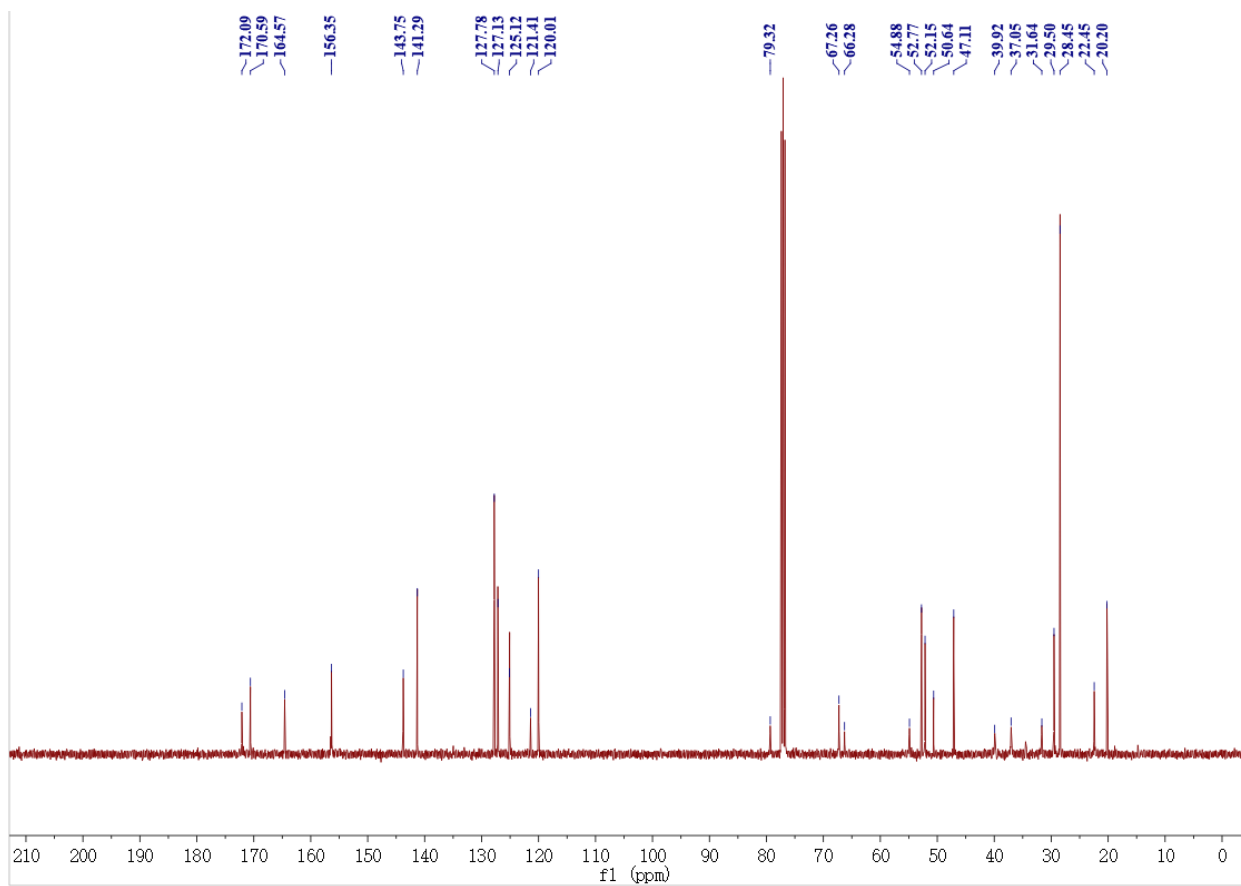


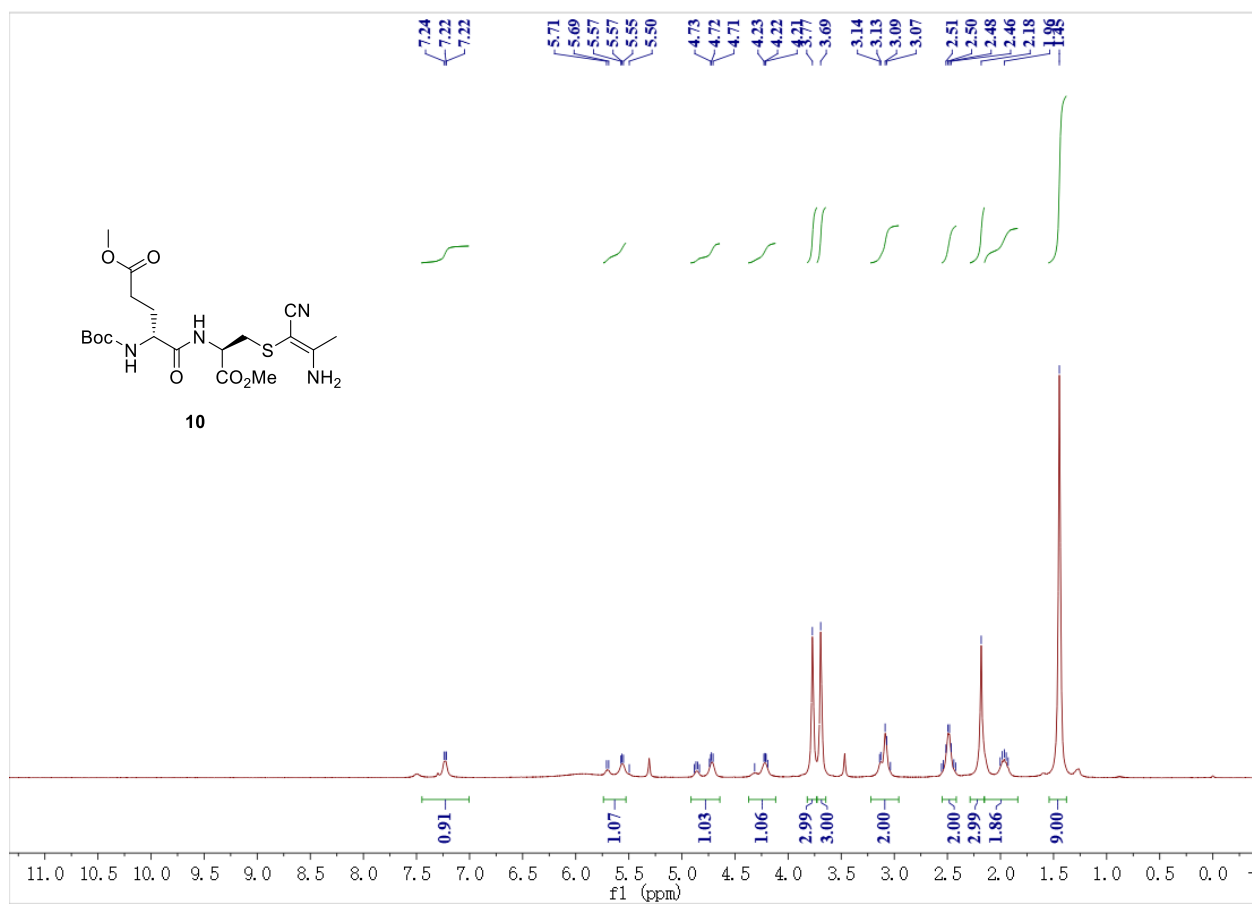


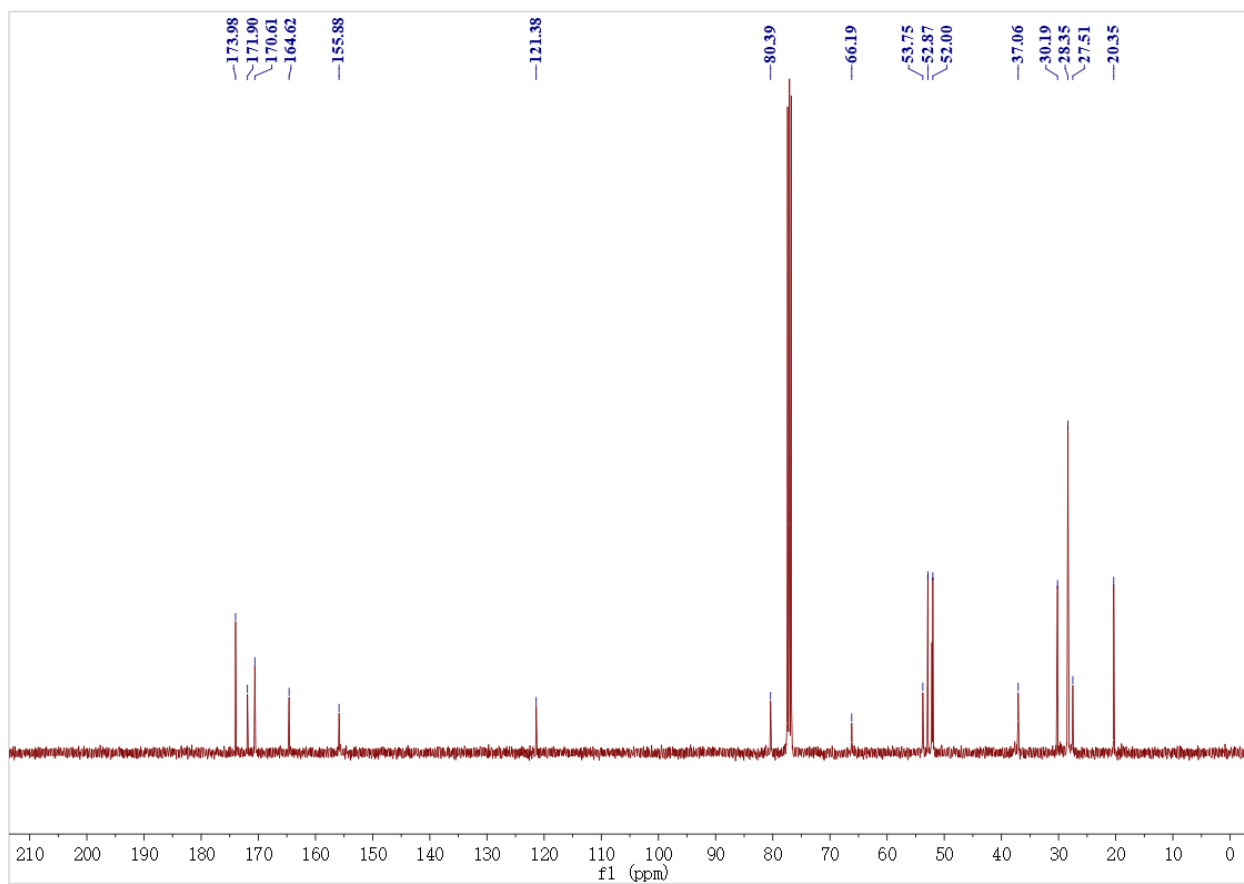


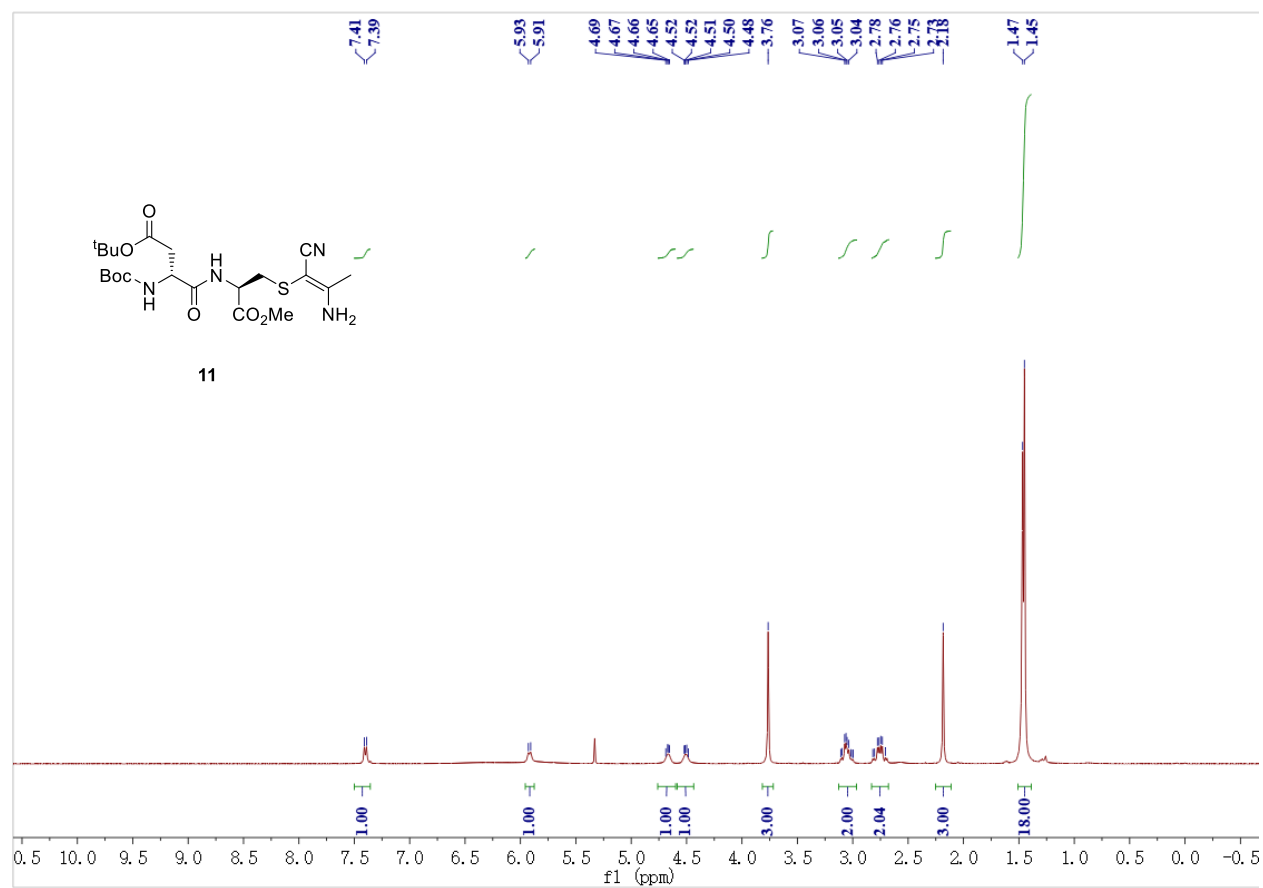


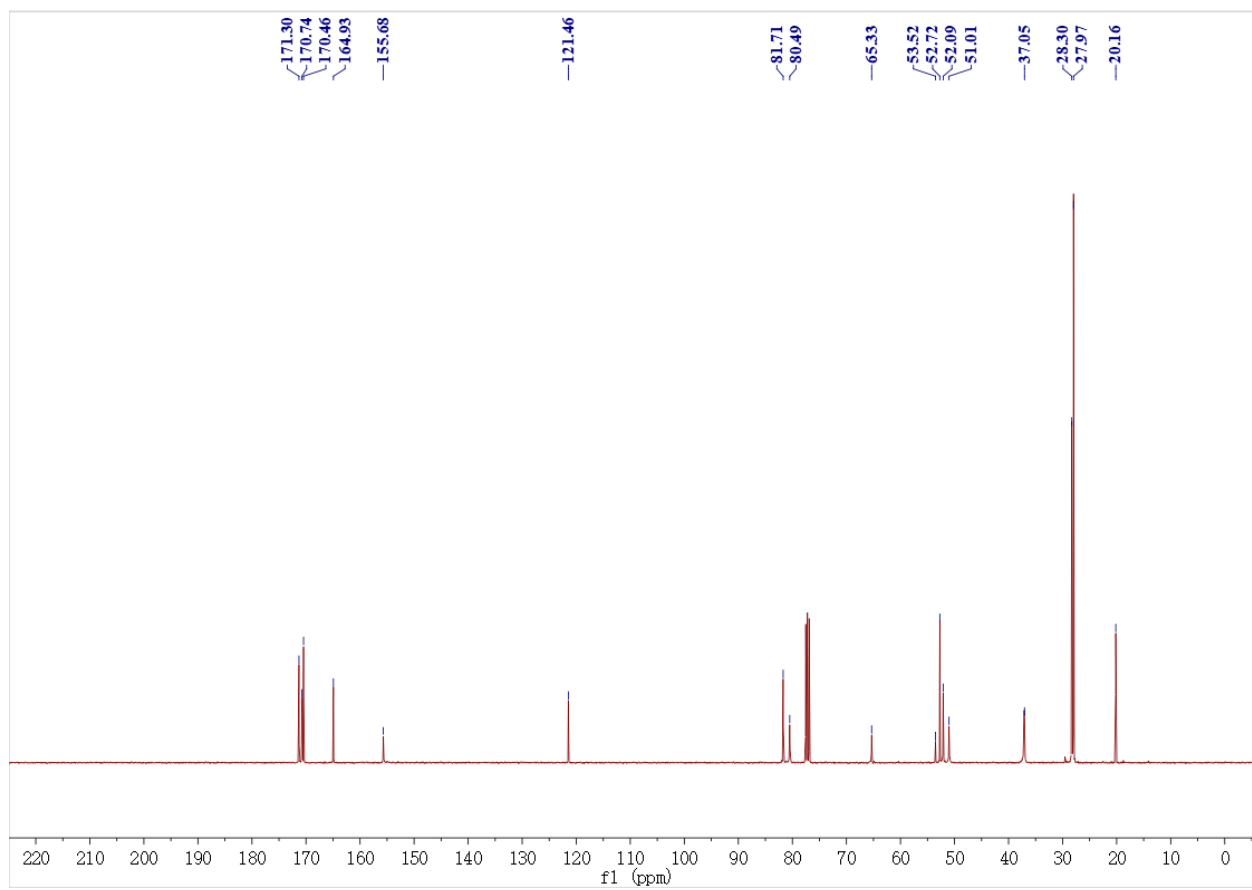


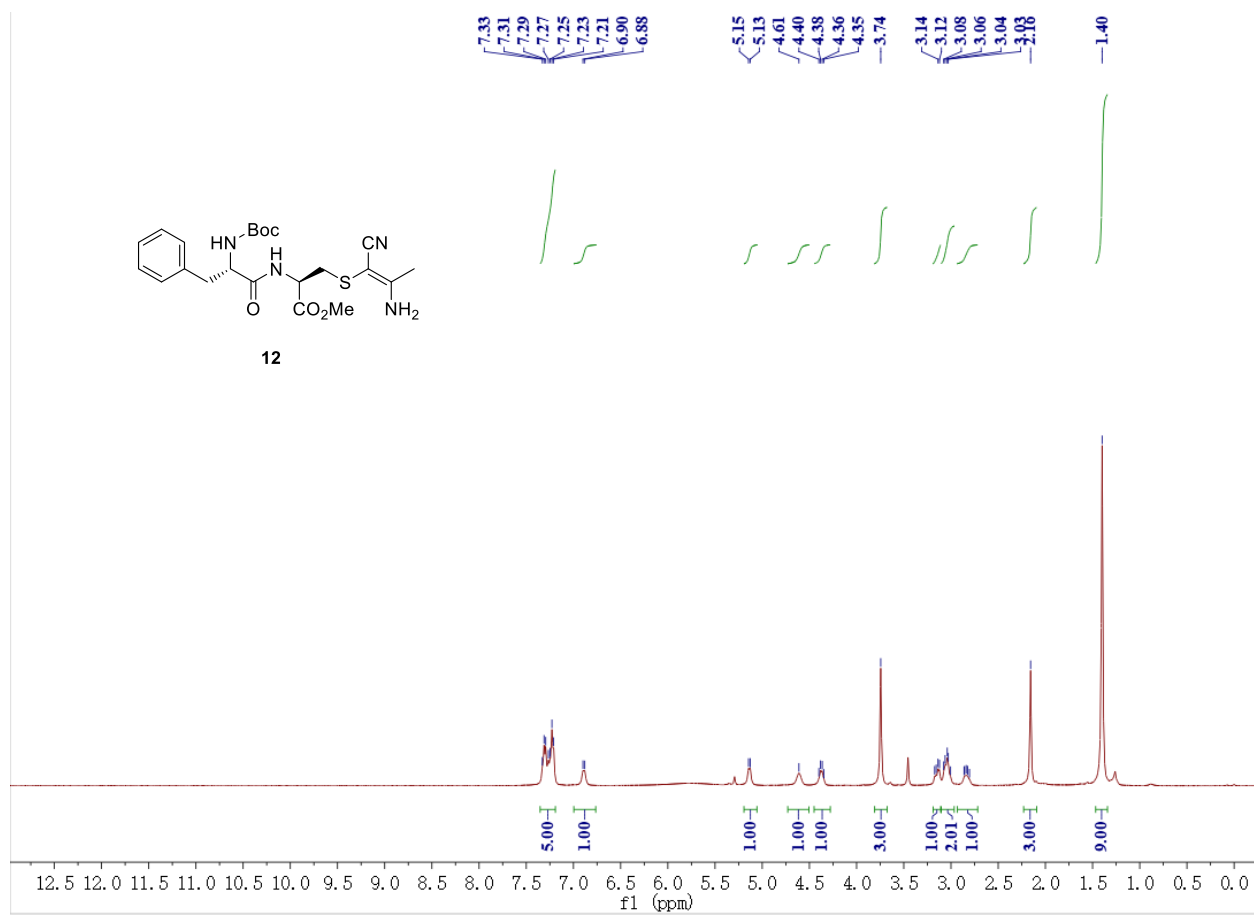


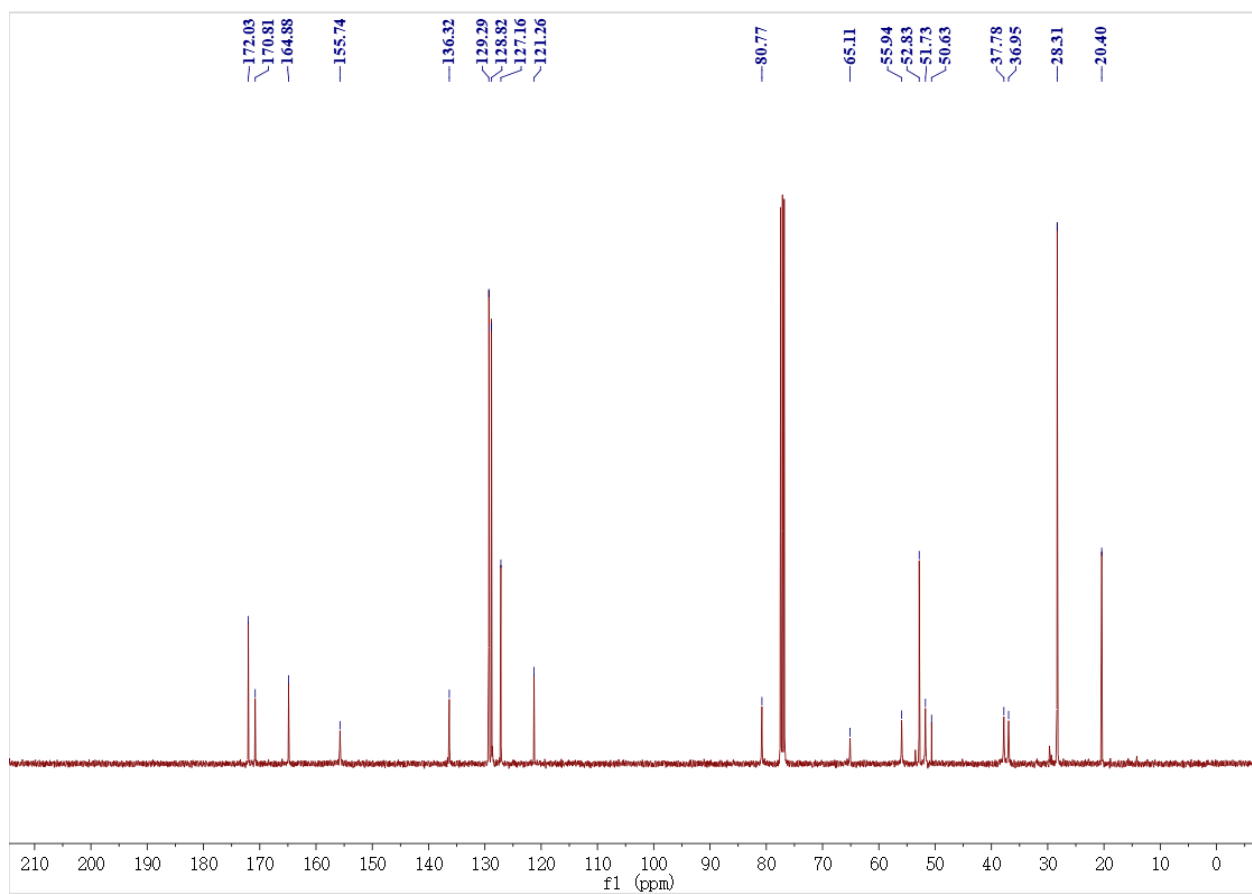


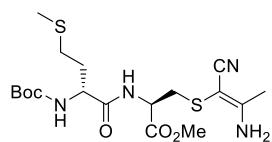




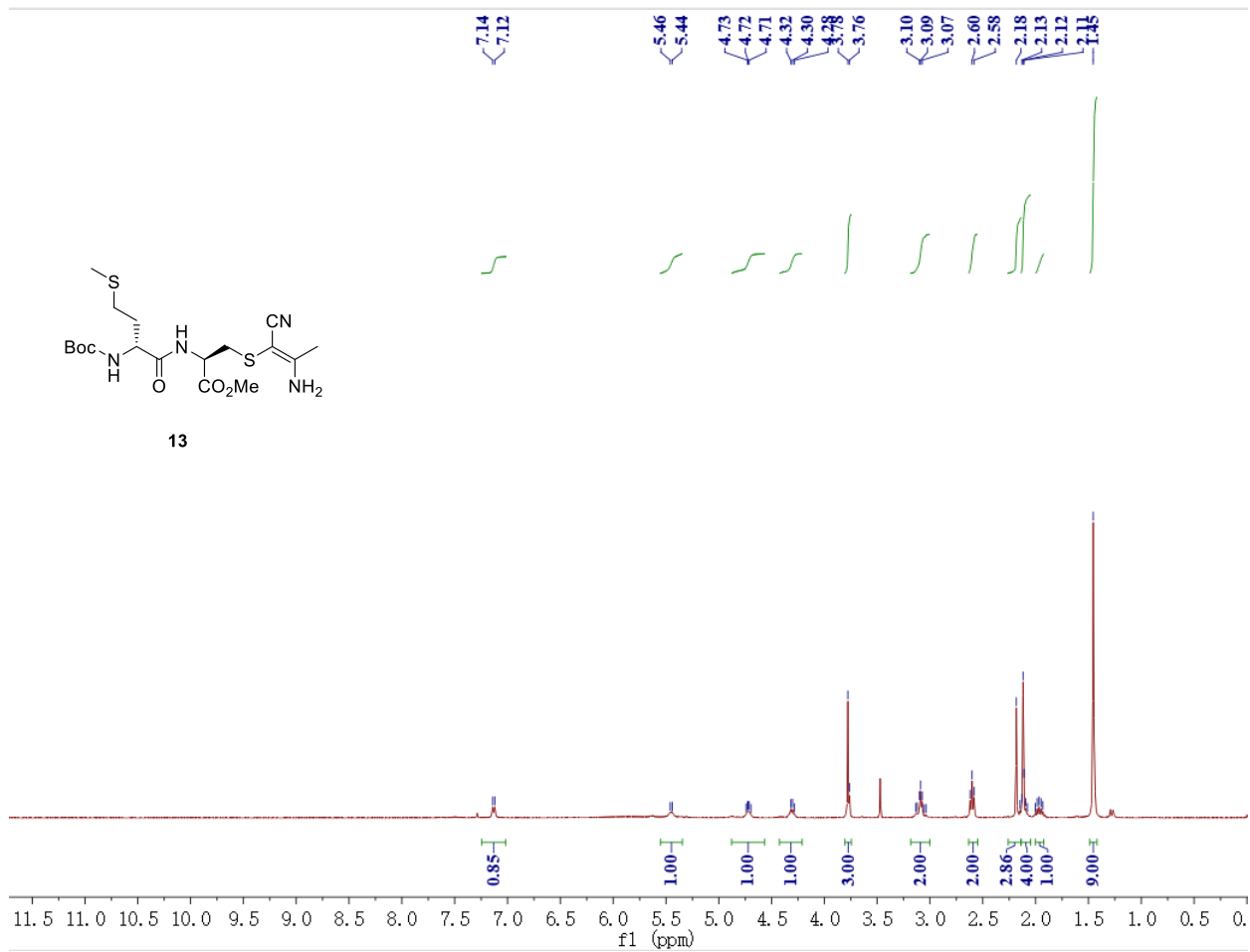


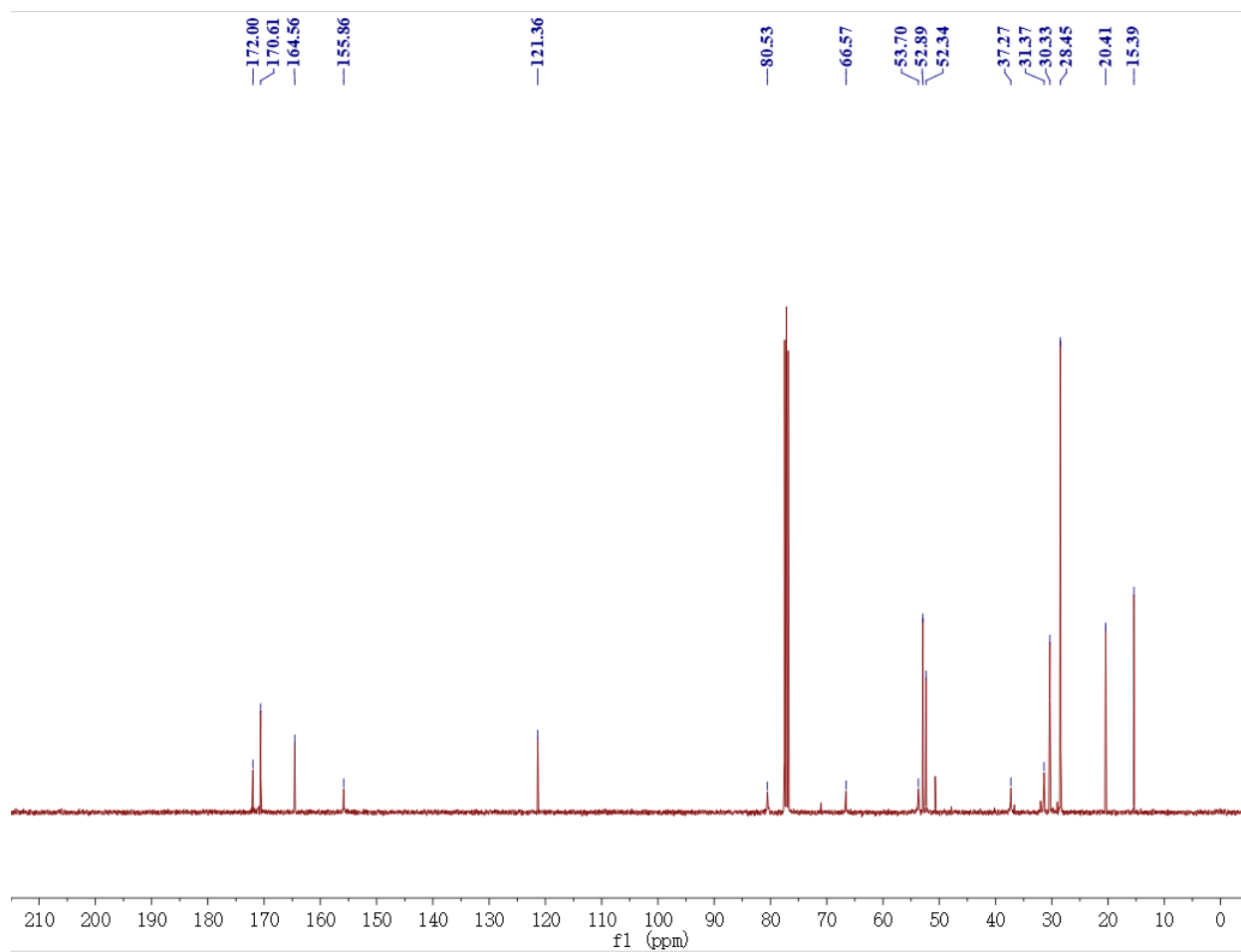


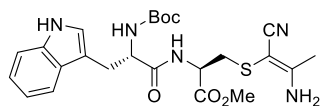




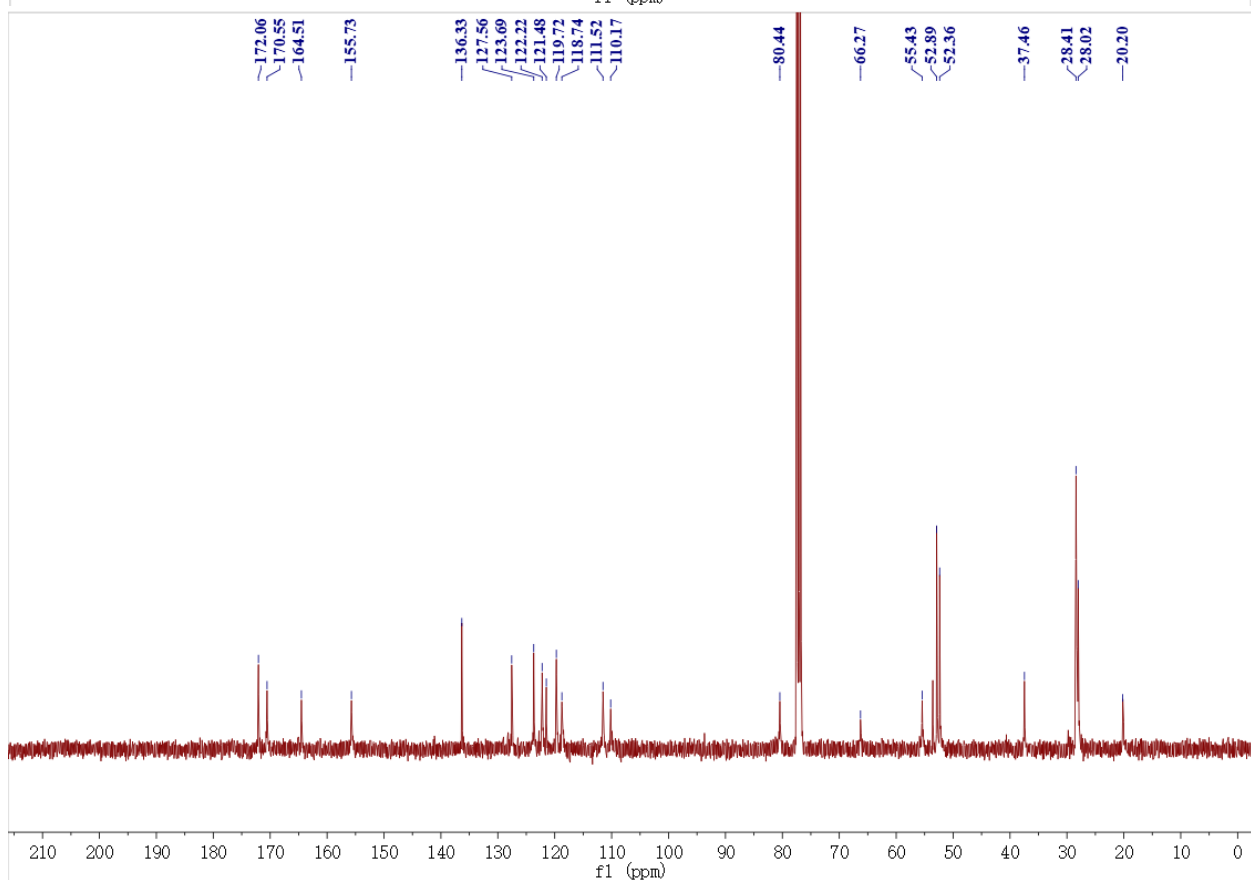
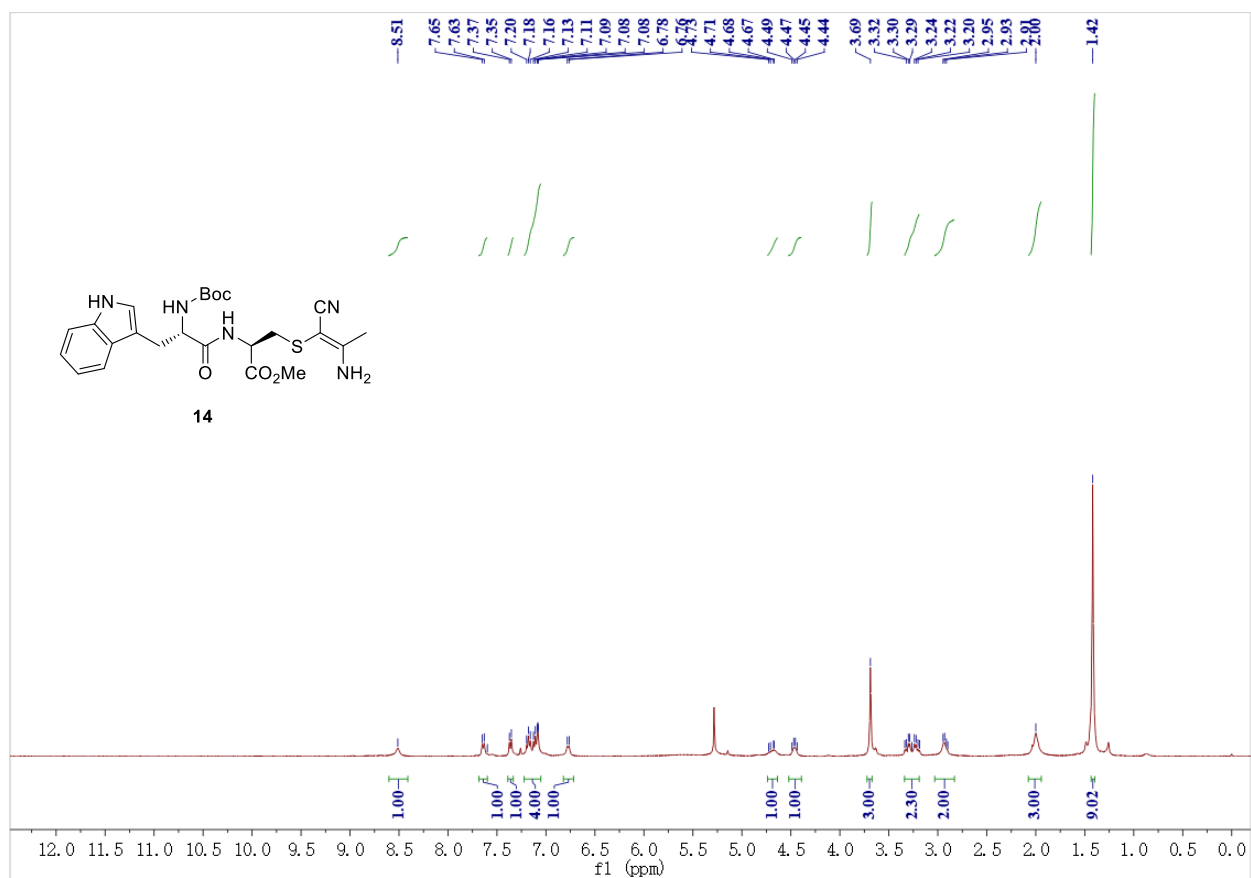
13

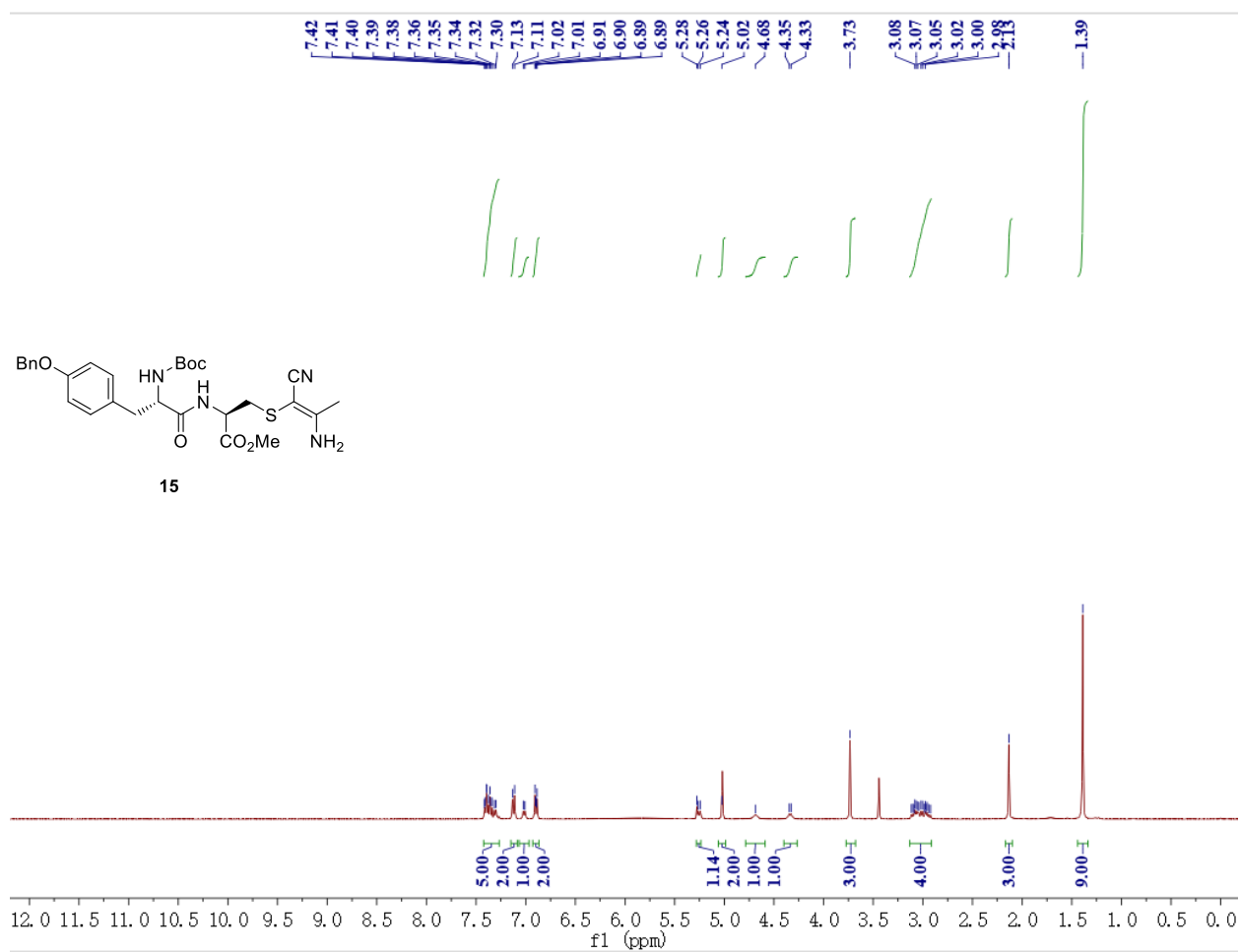


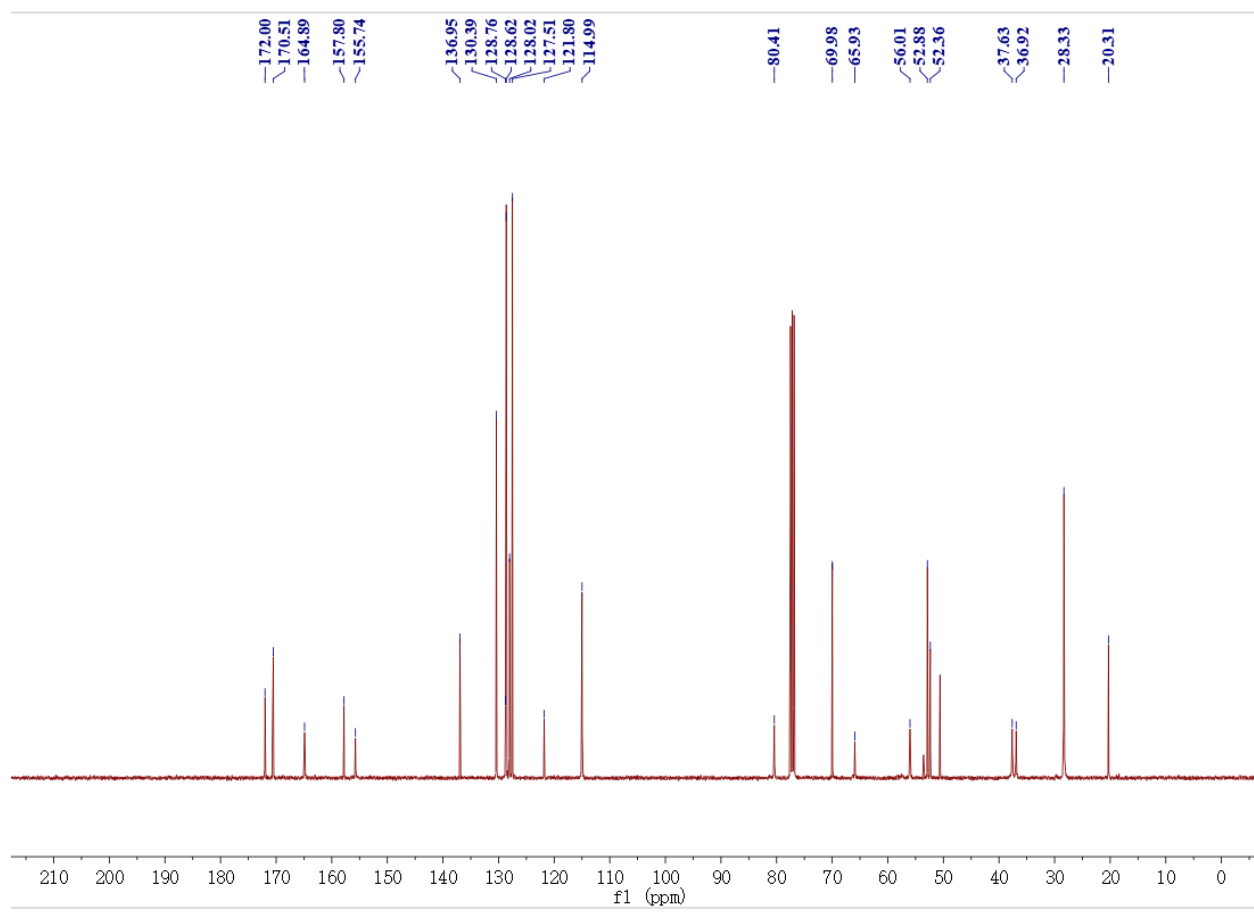


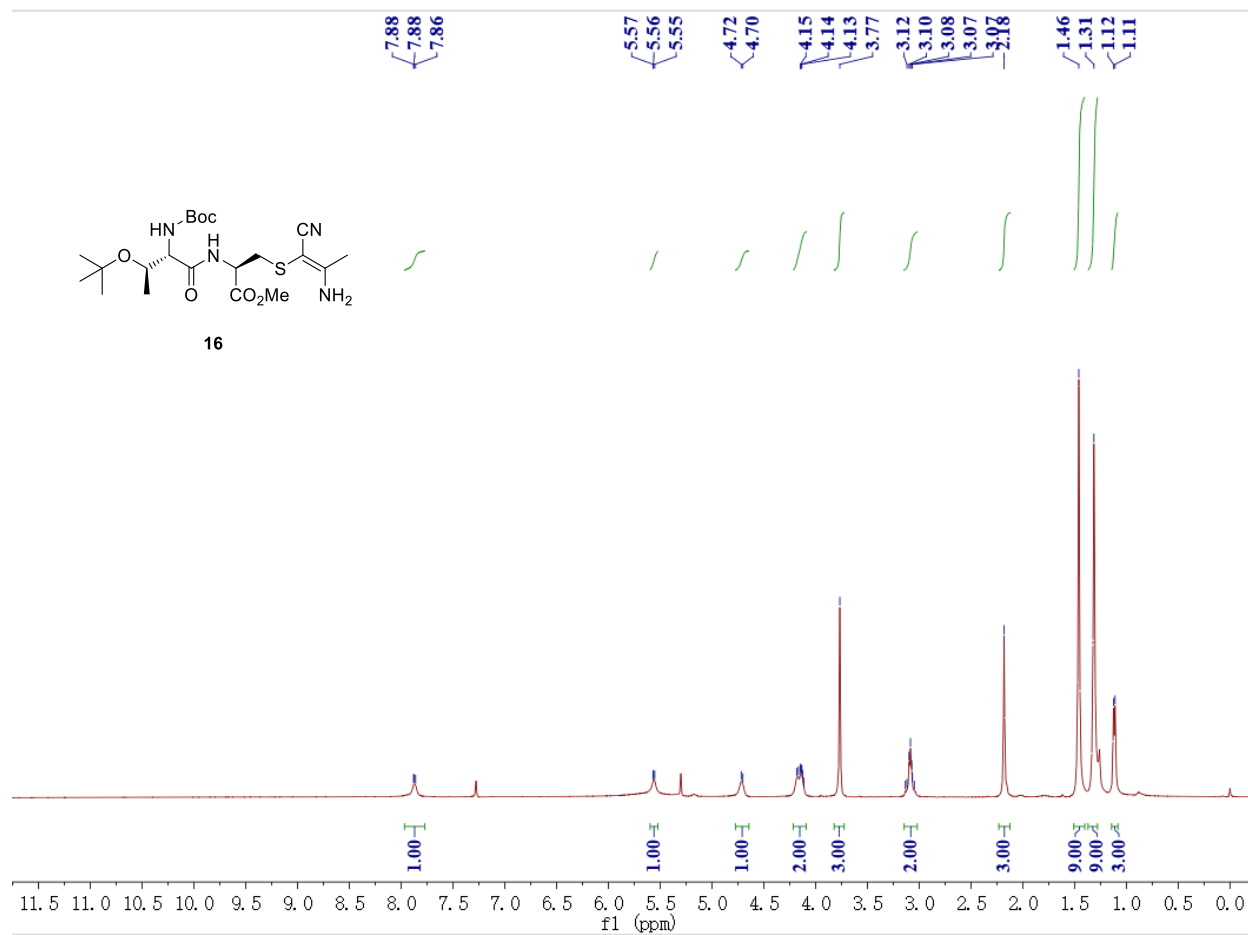


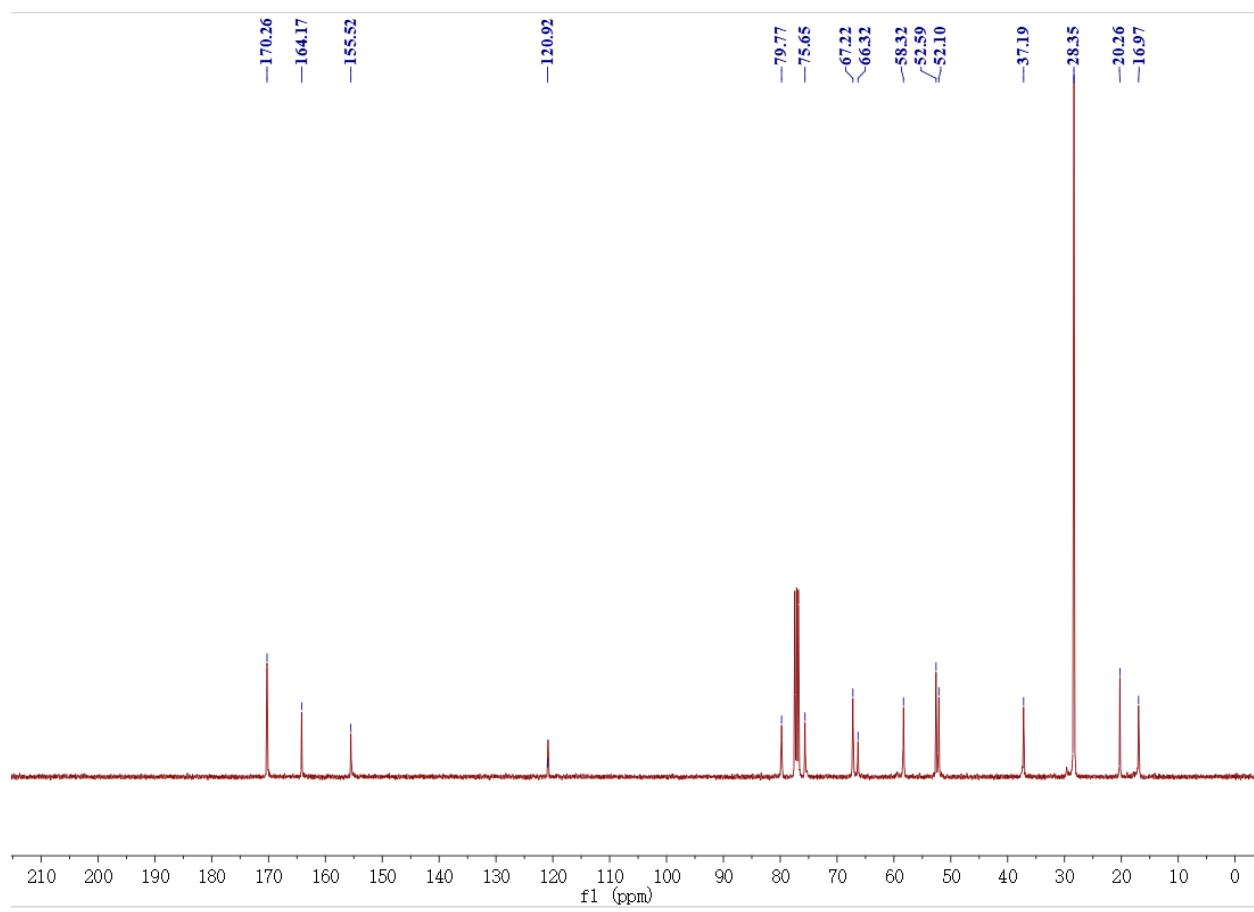
14

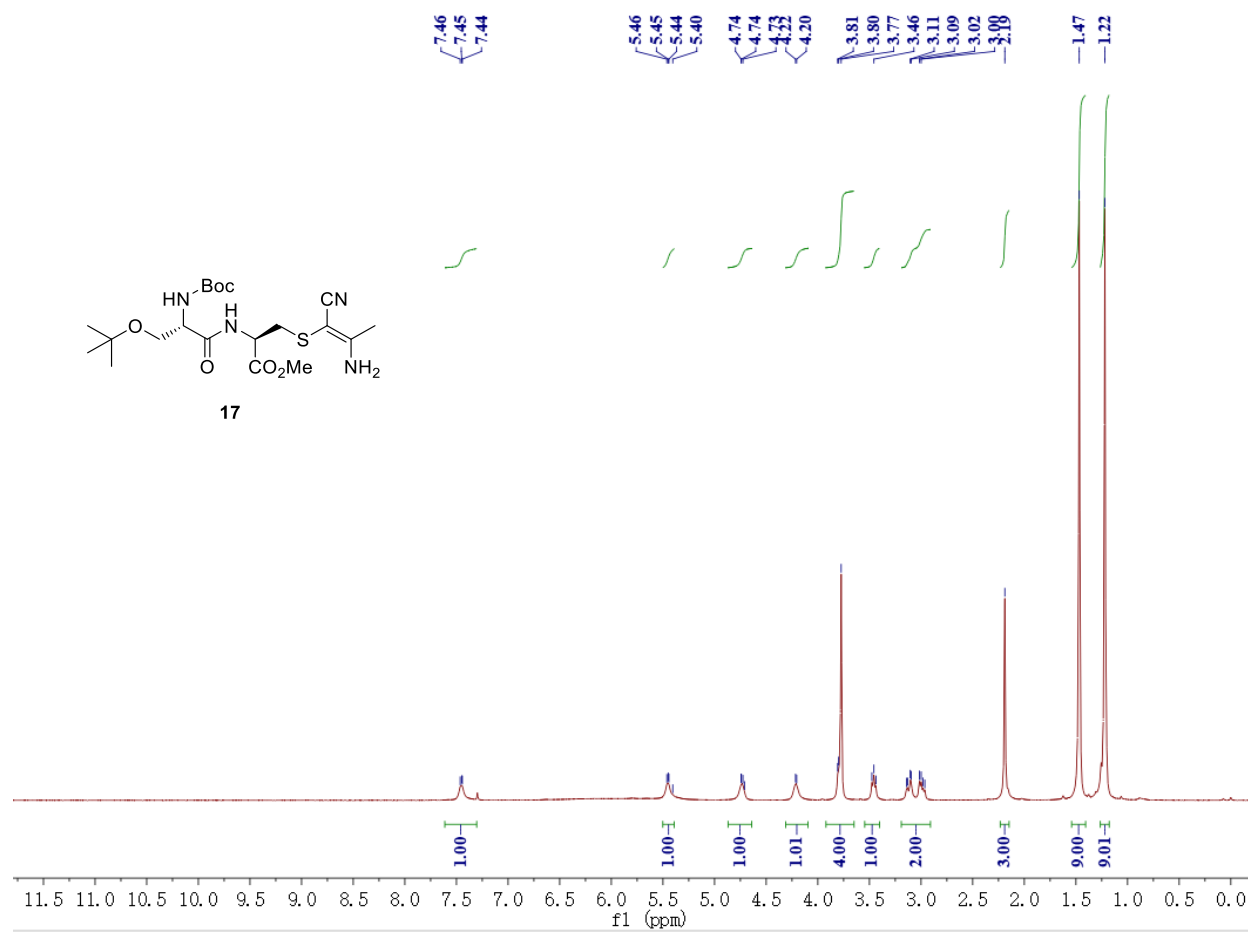


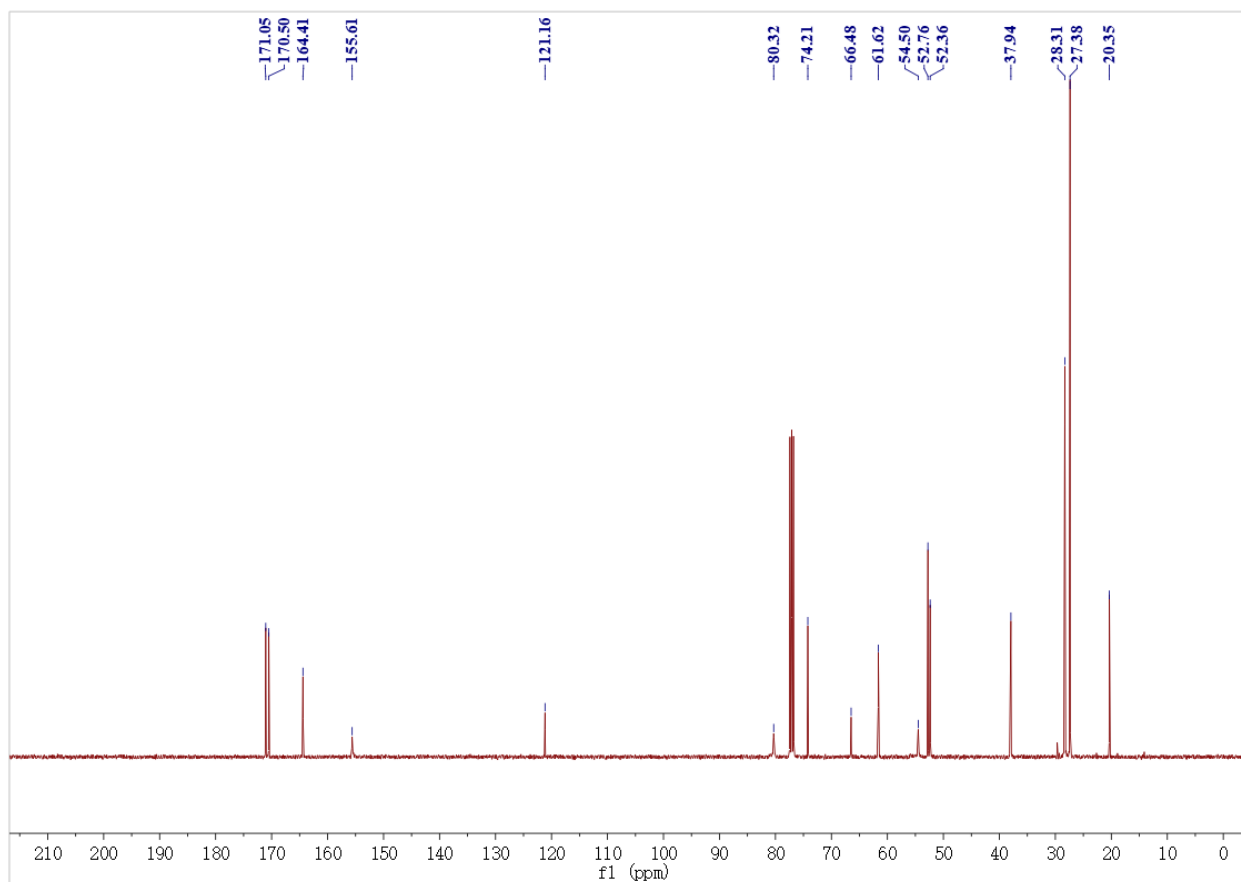


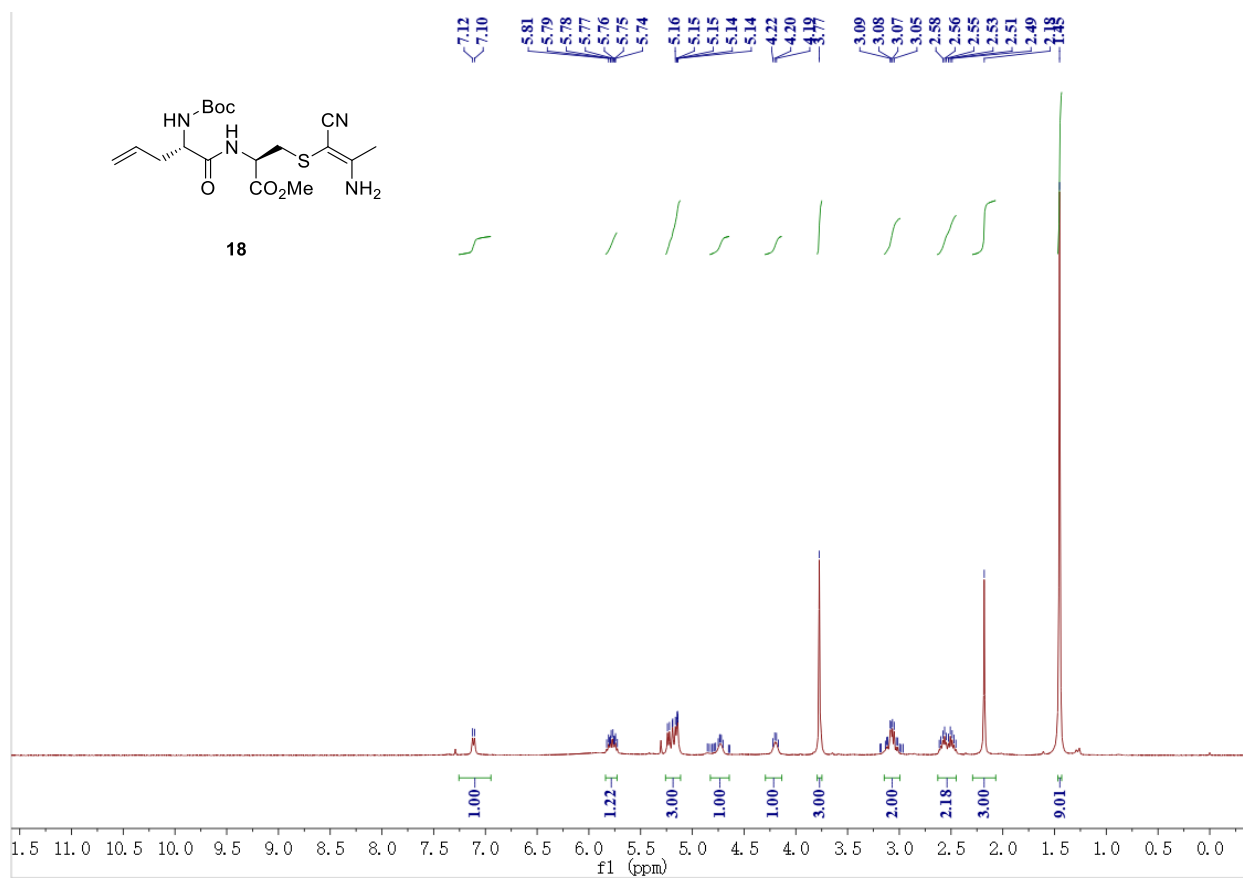


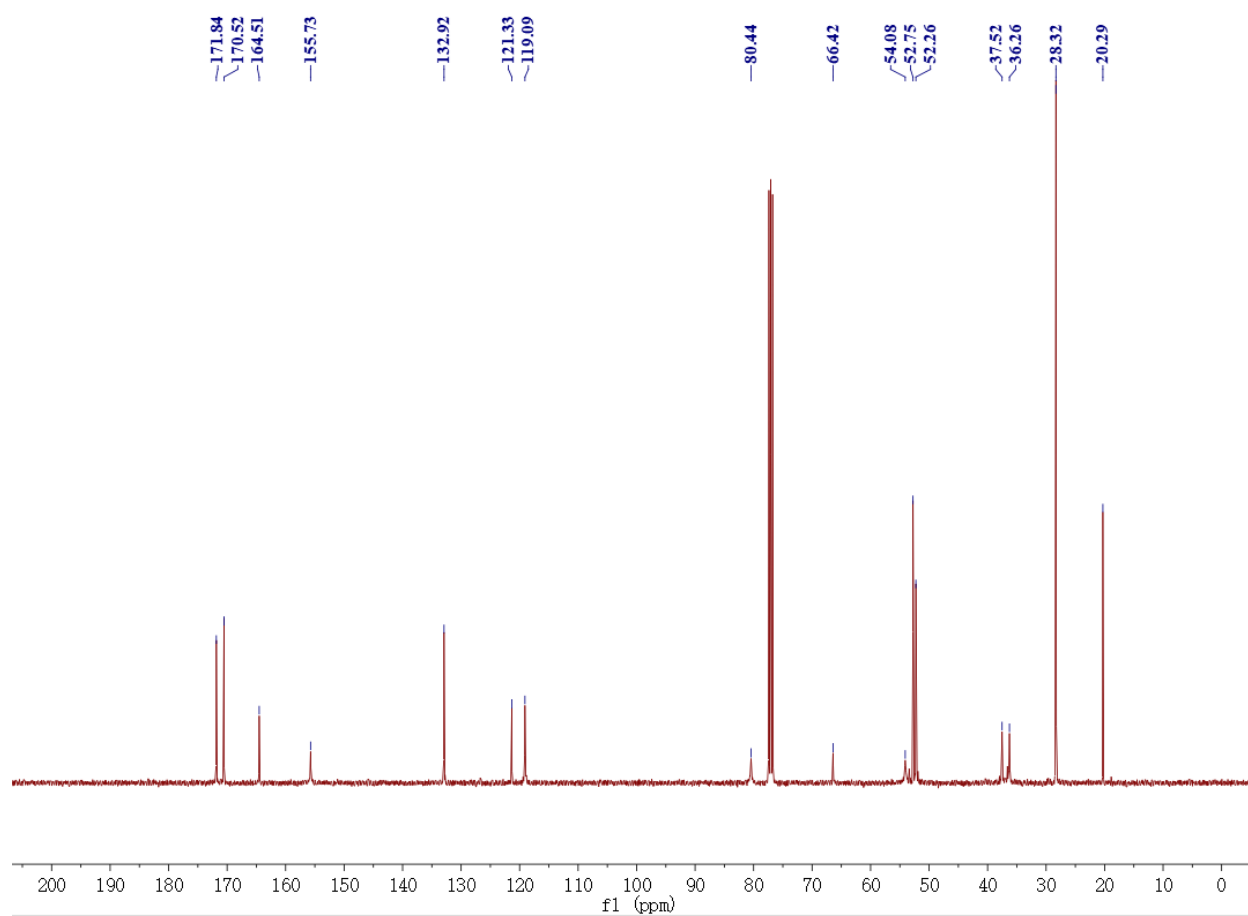


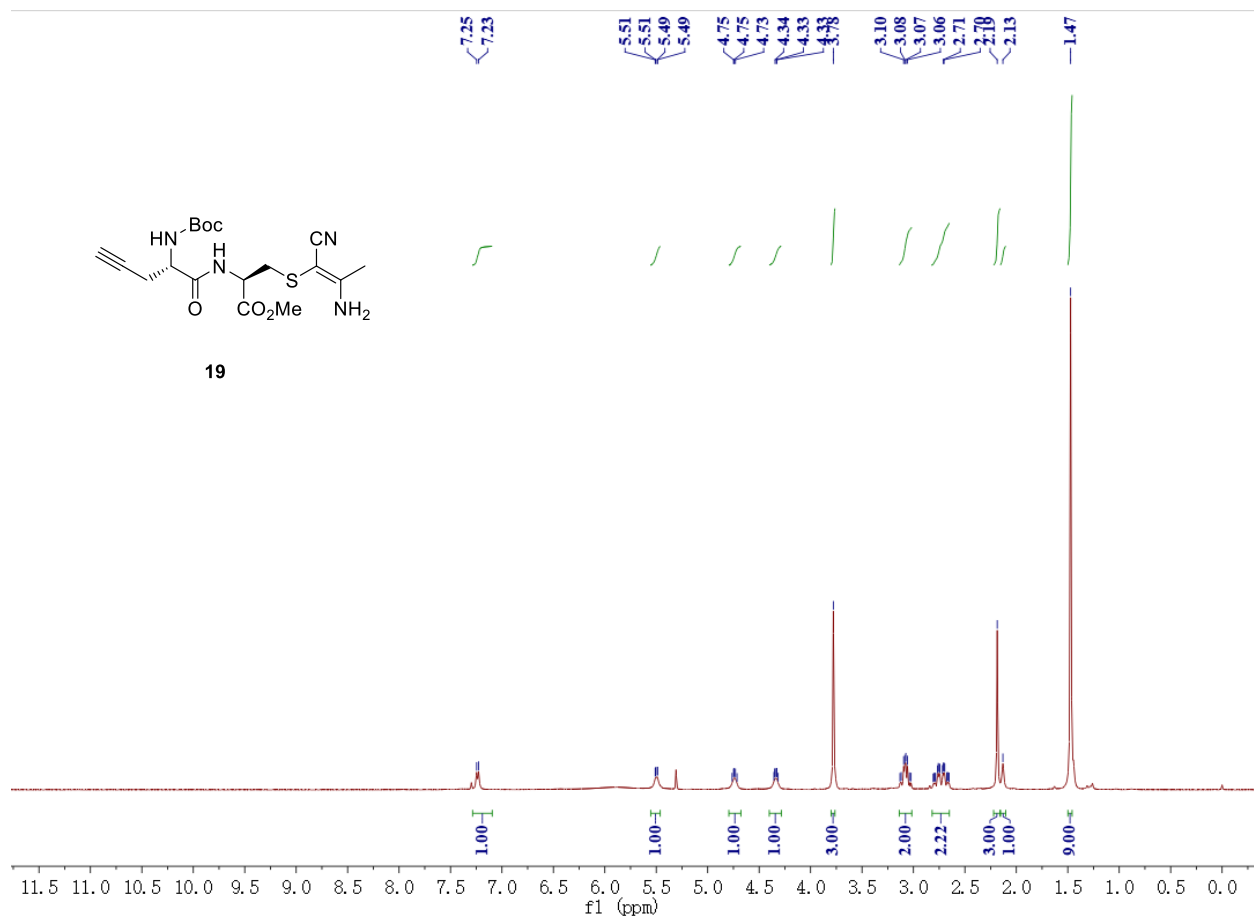


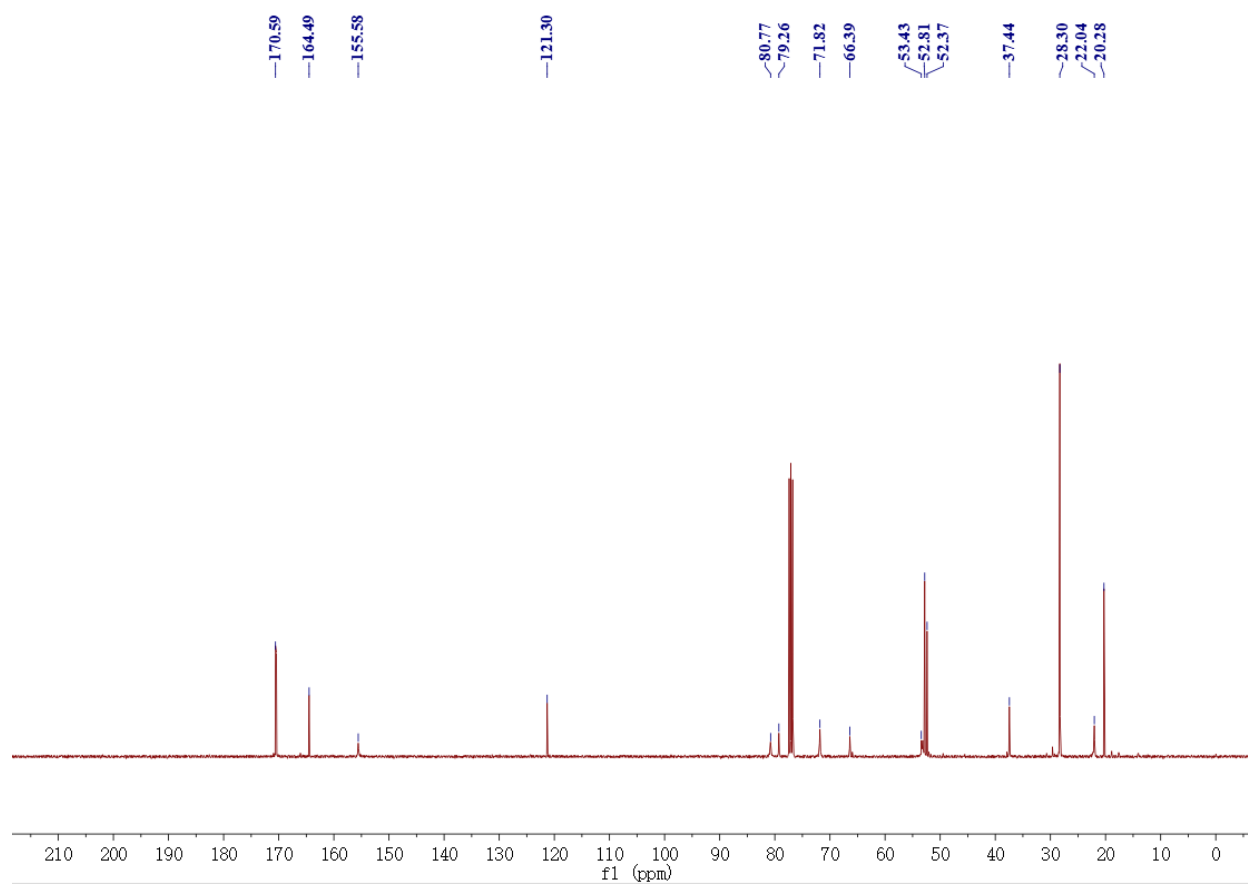


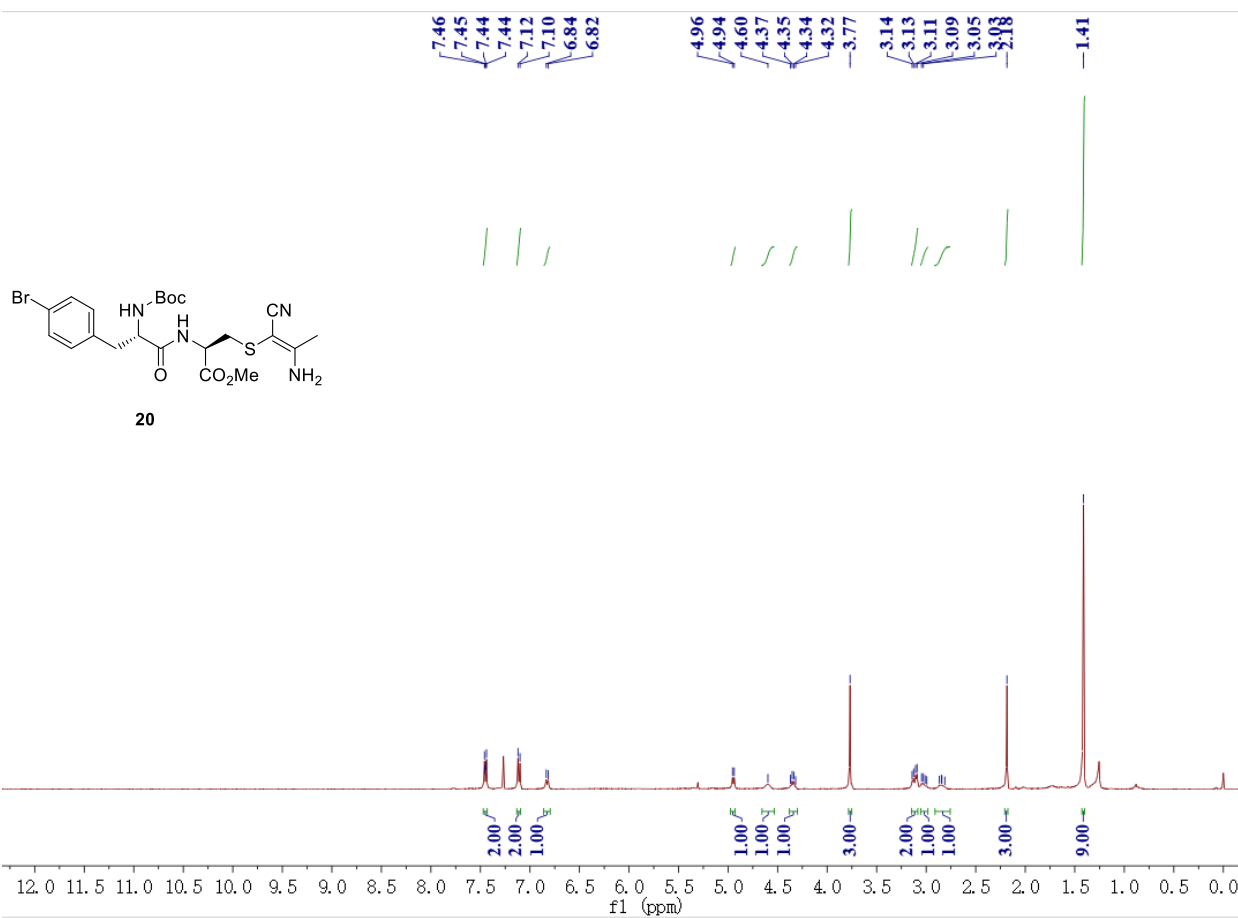


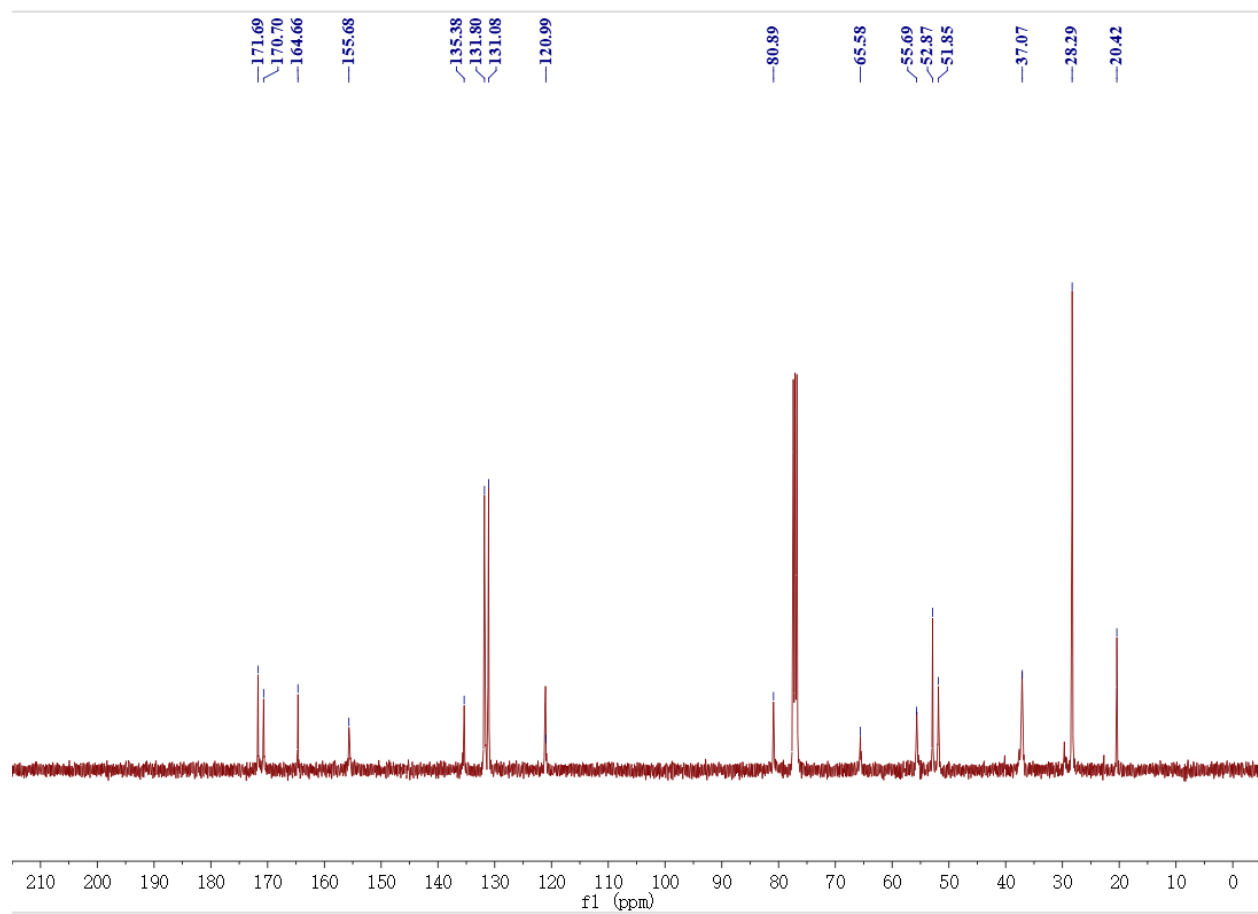


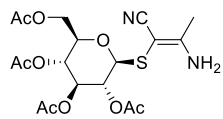




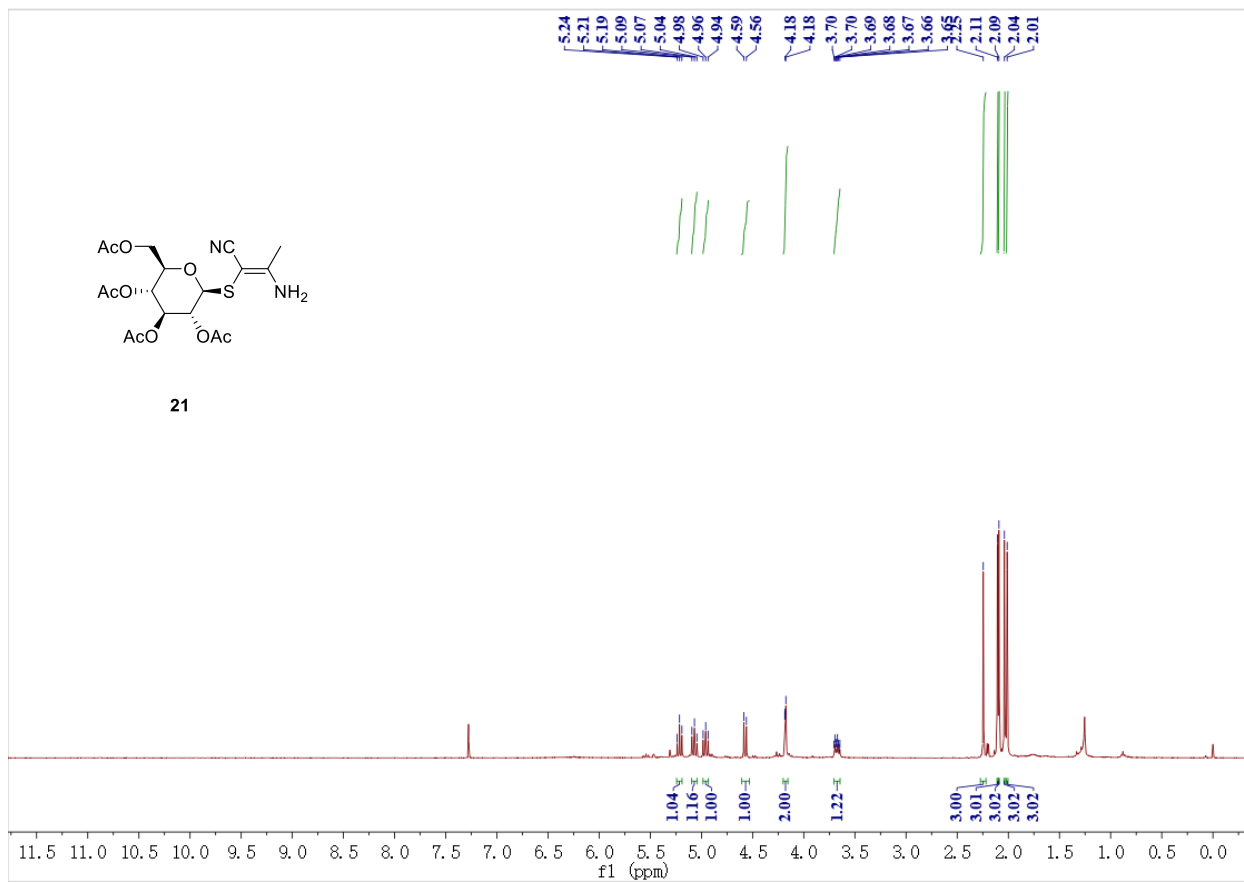


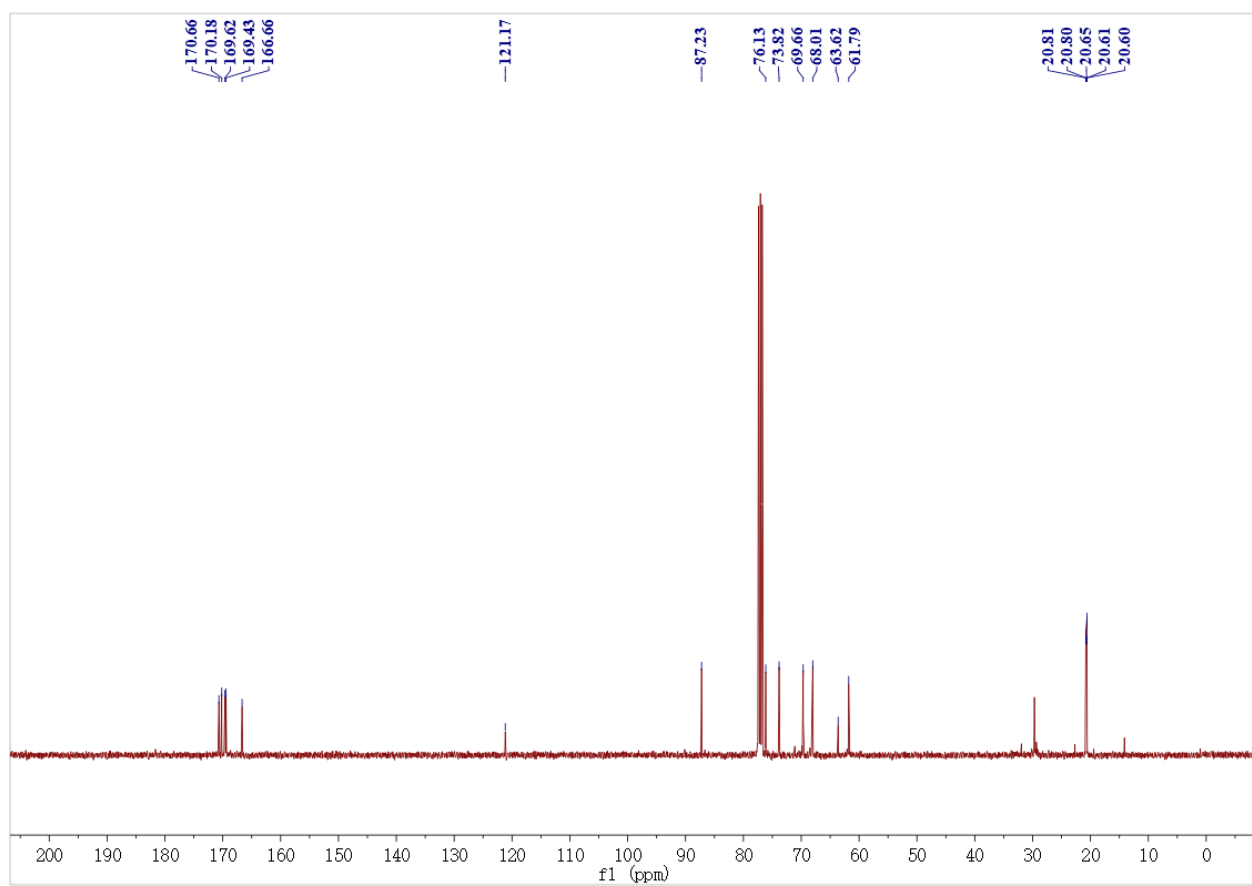


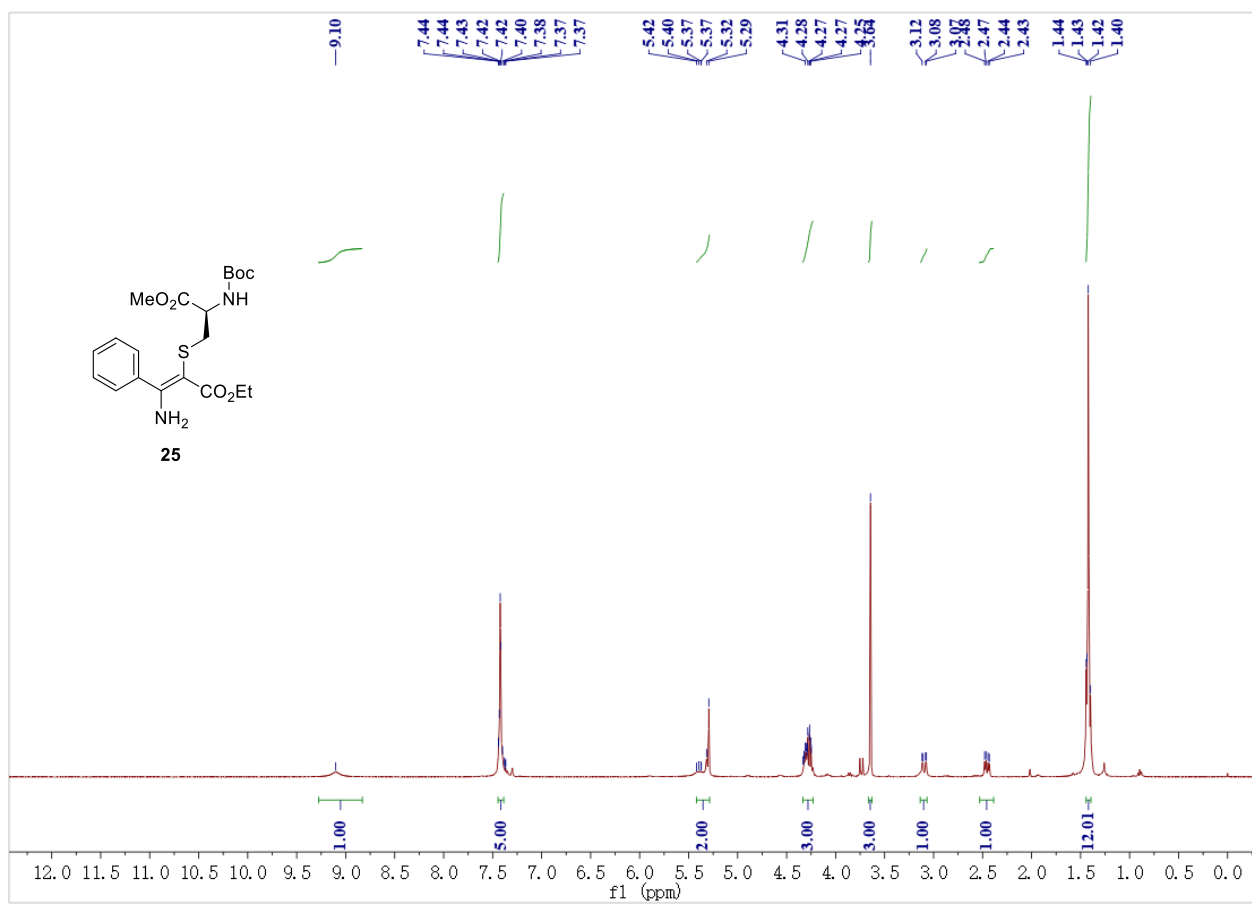


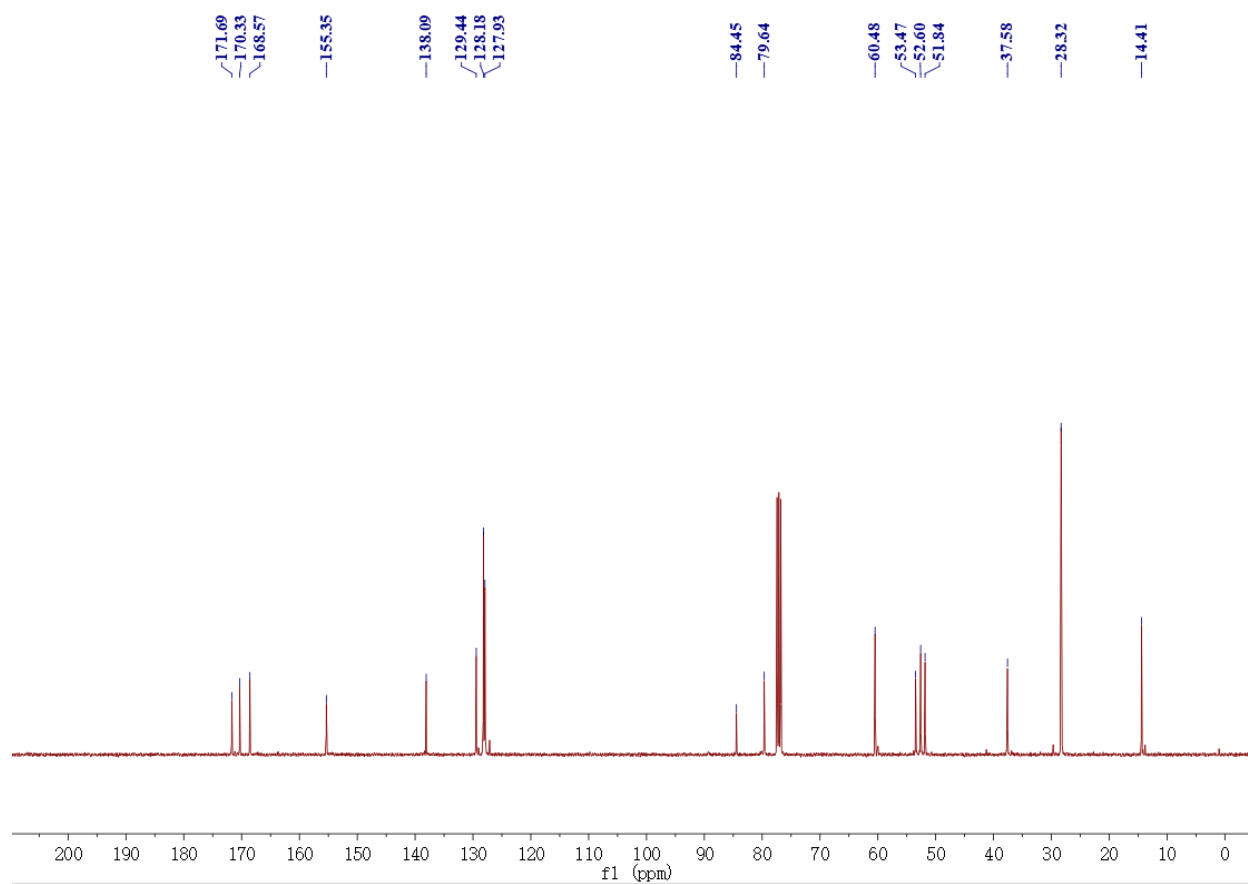


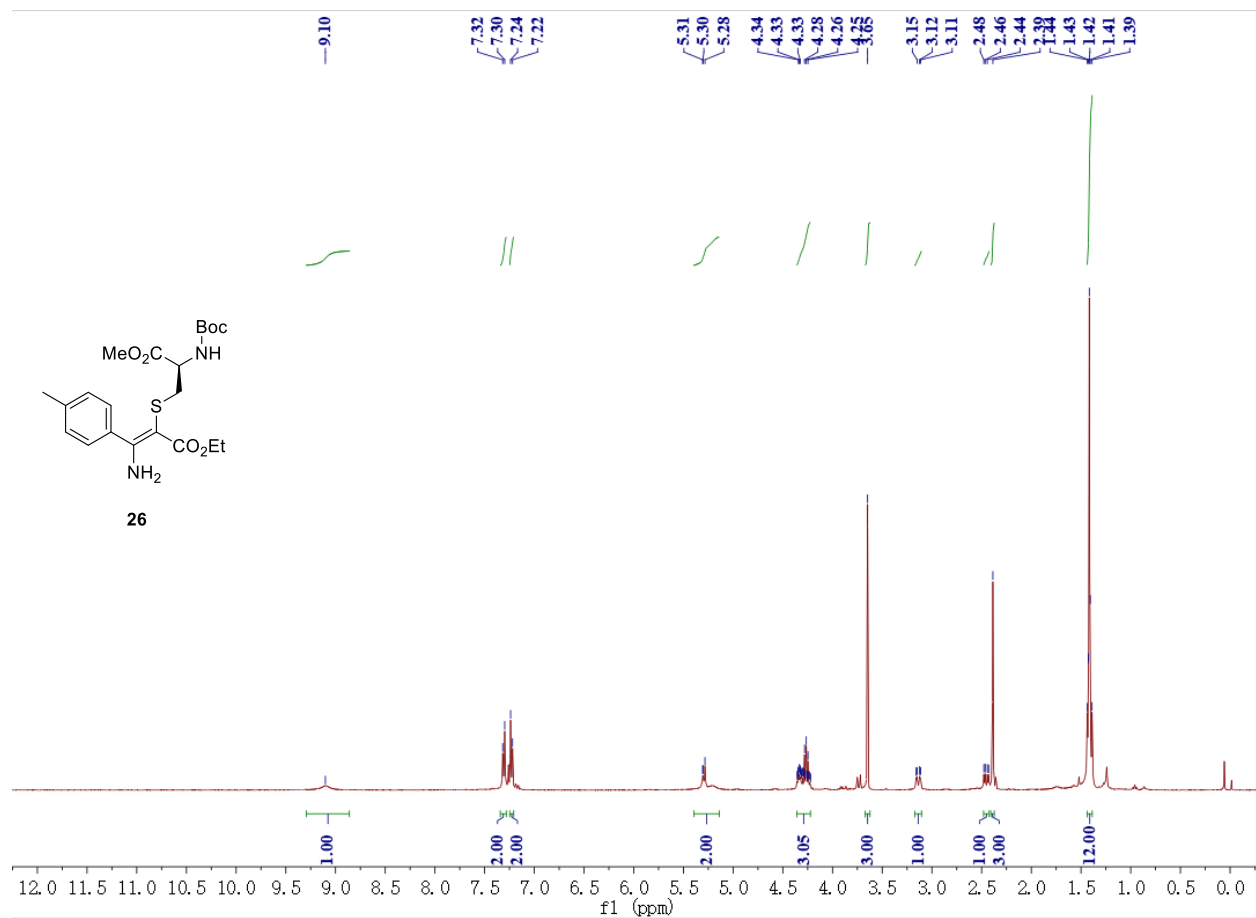
21

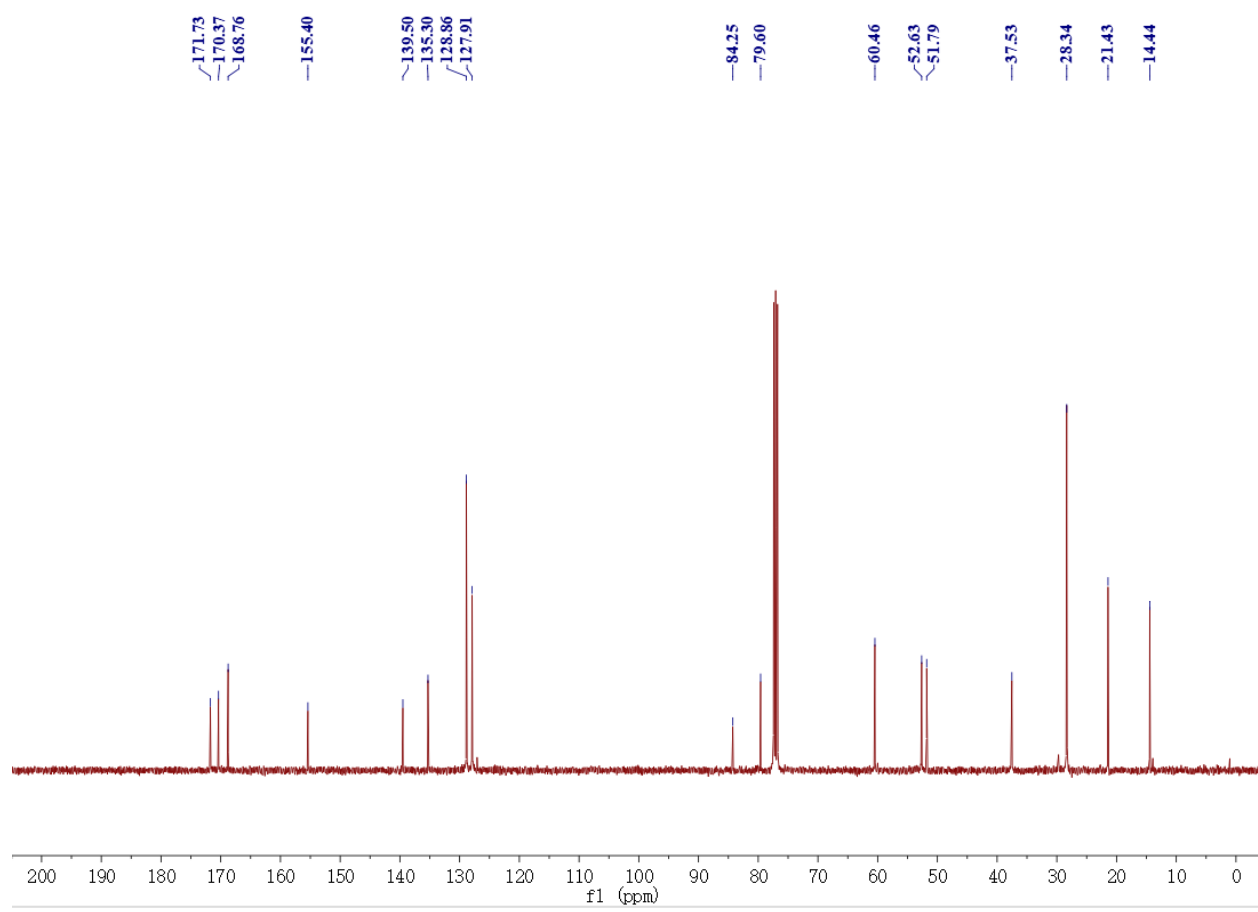


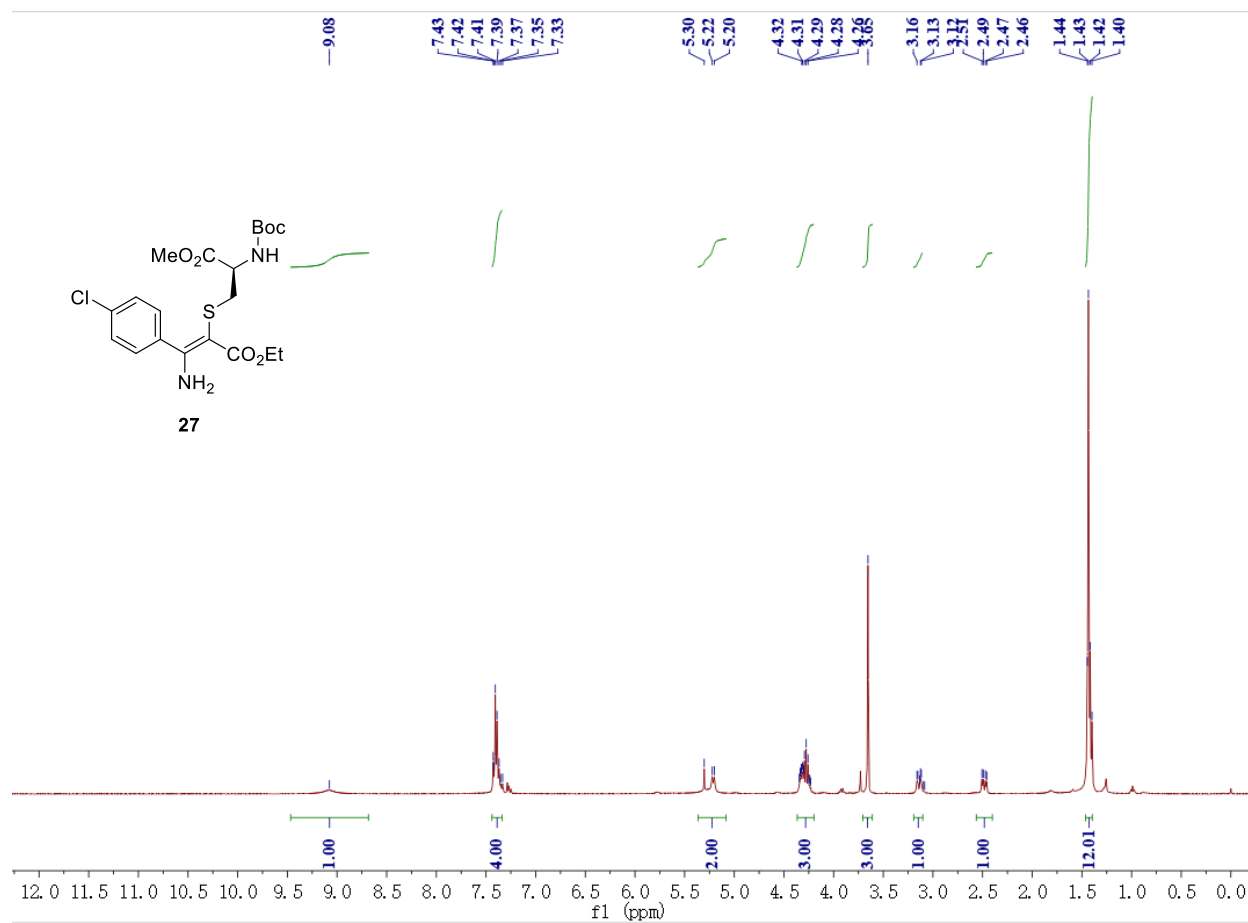


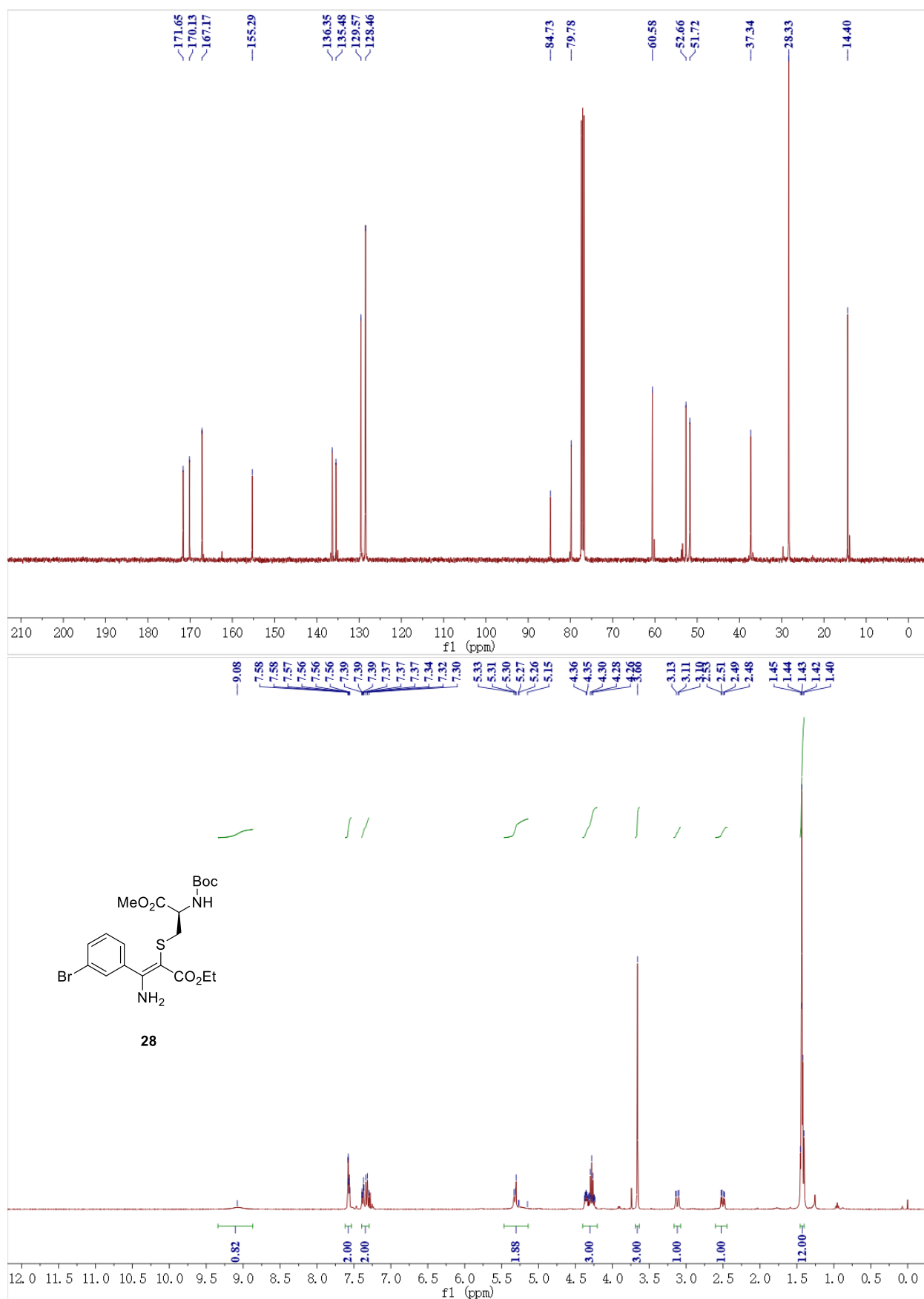


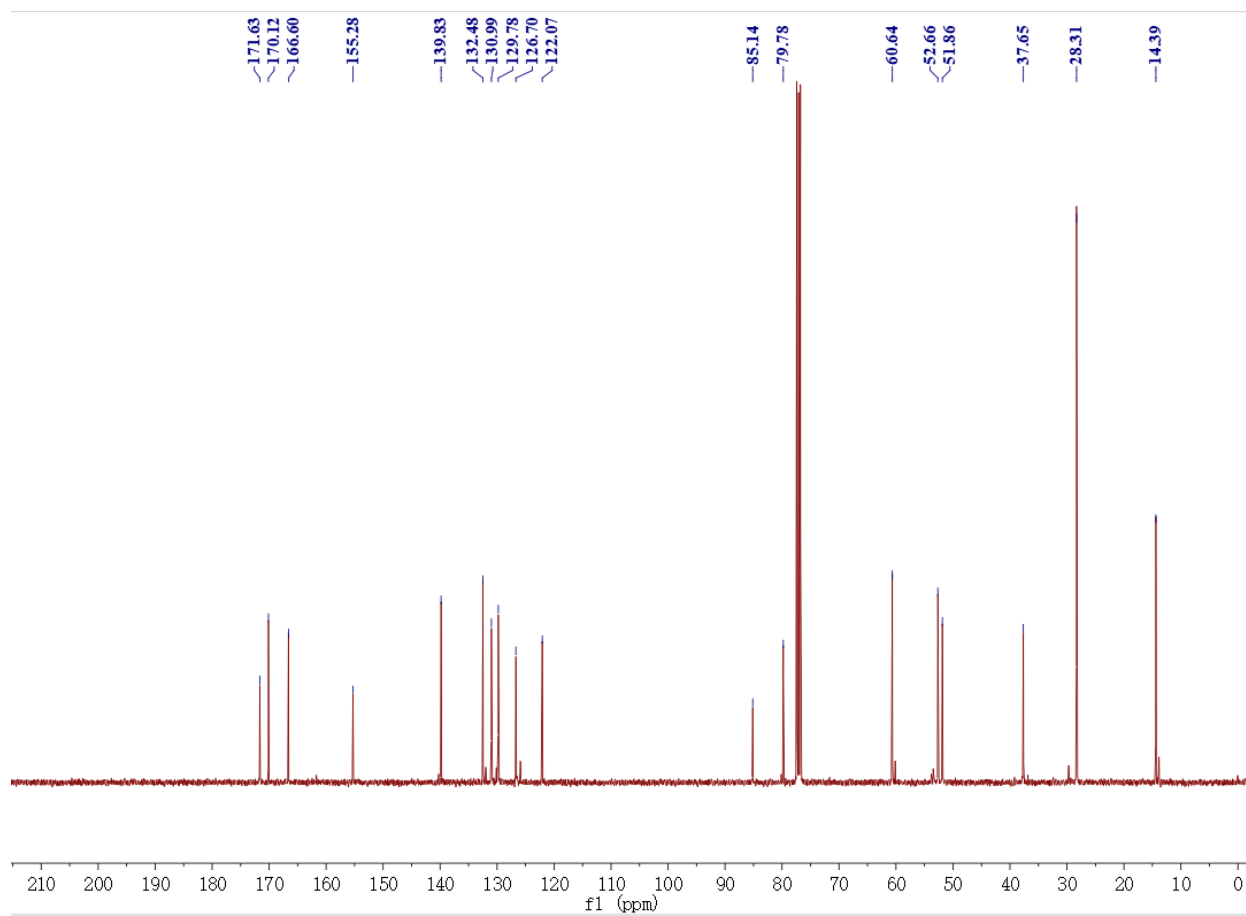


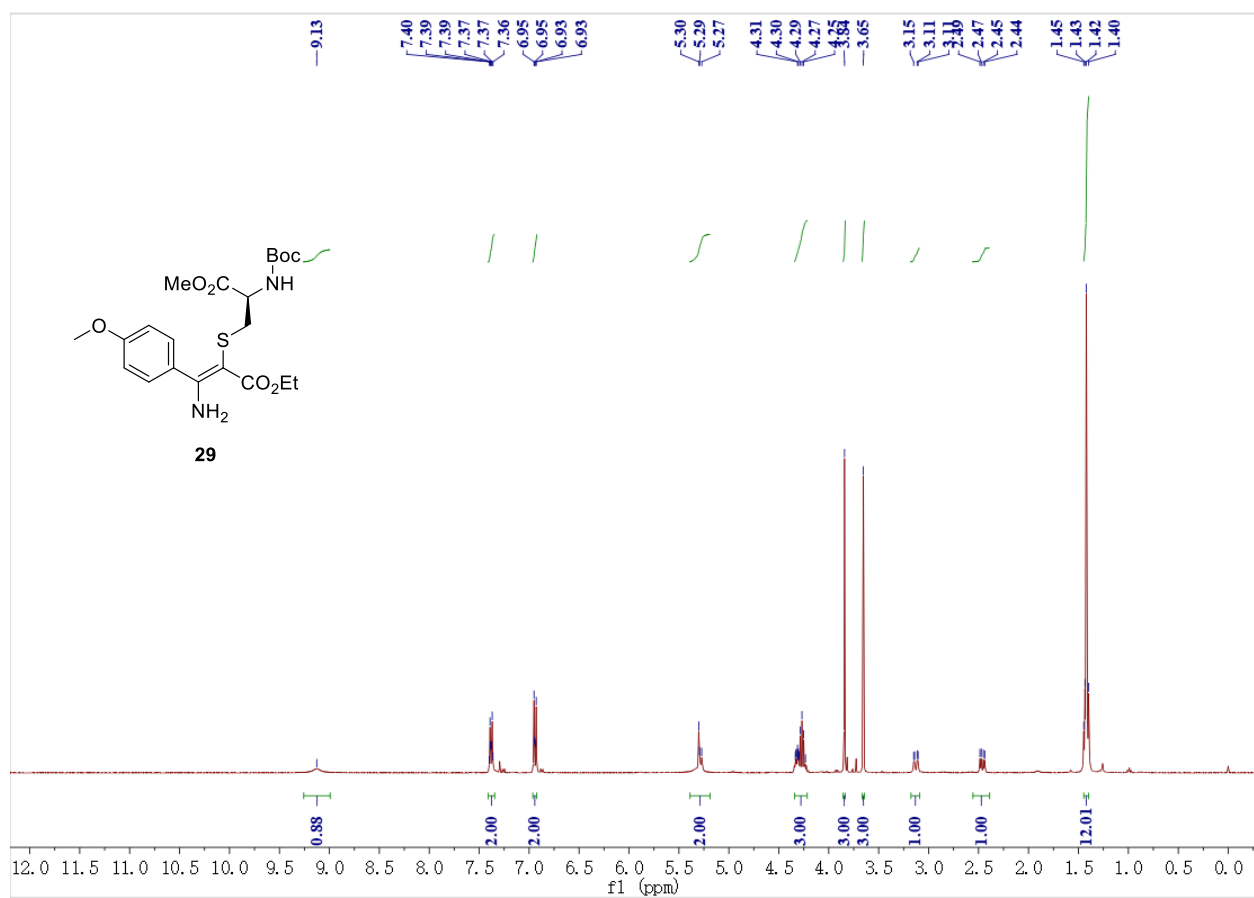


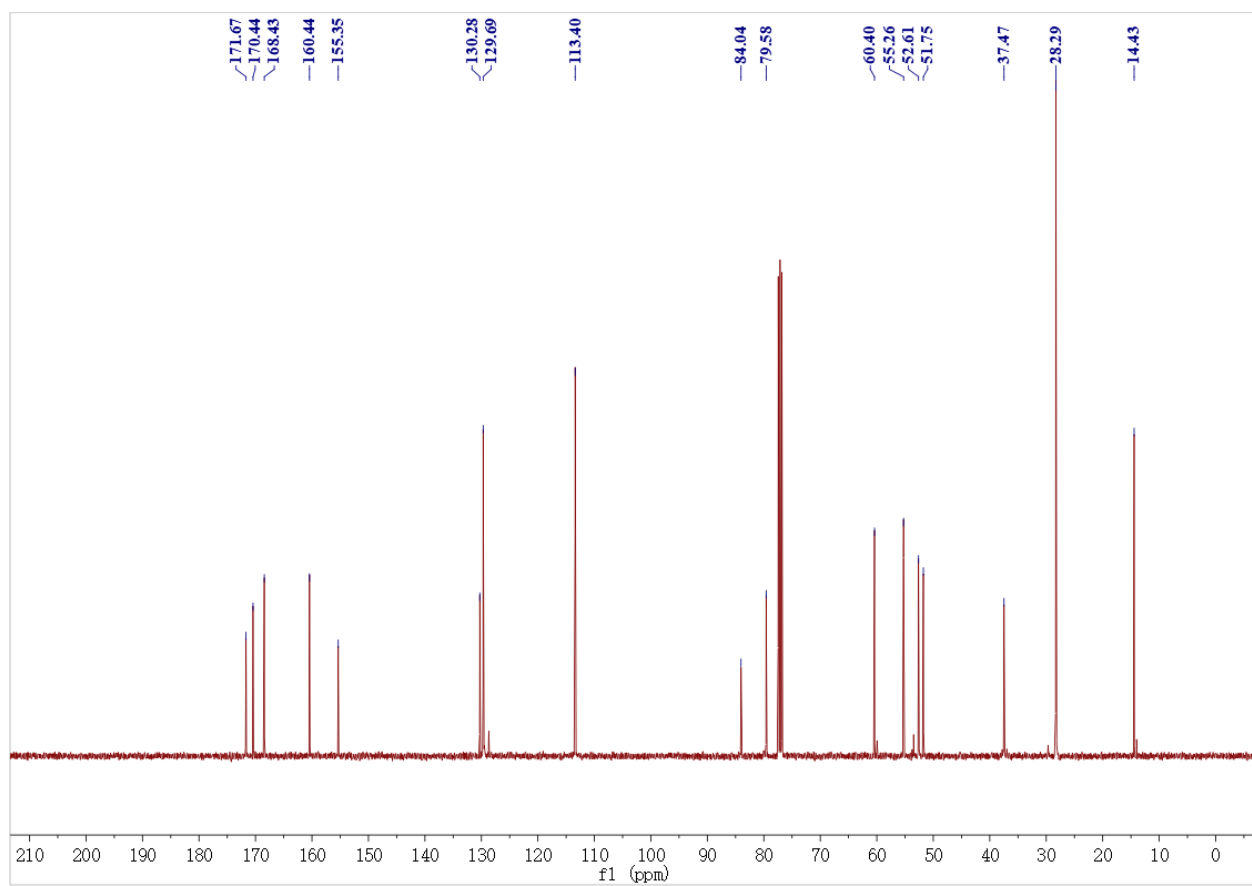


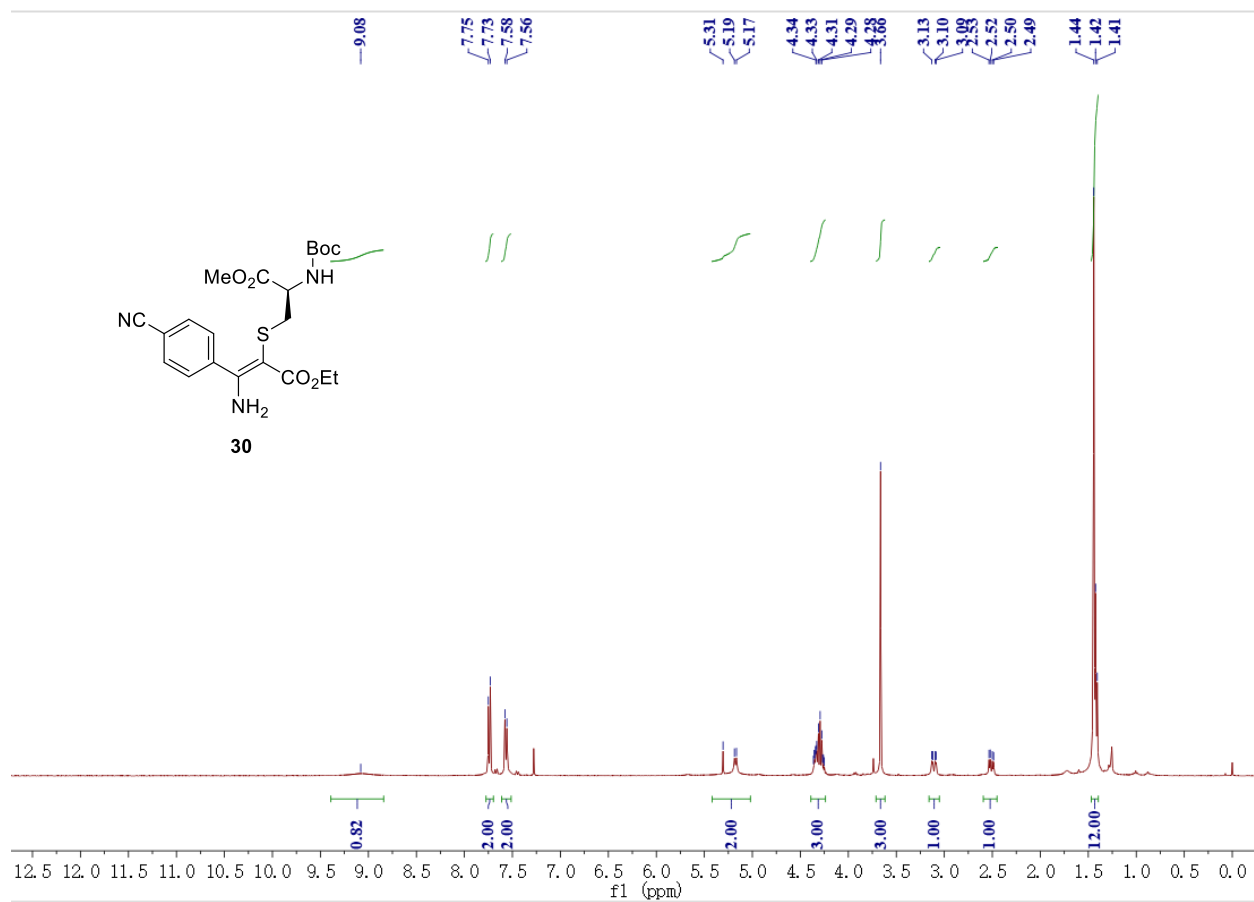


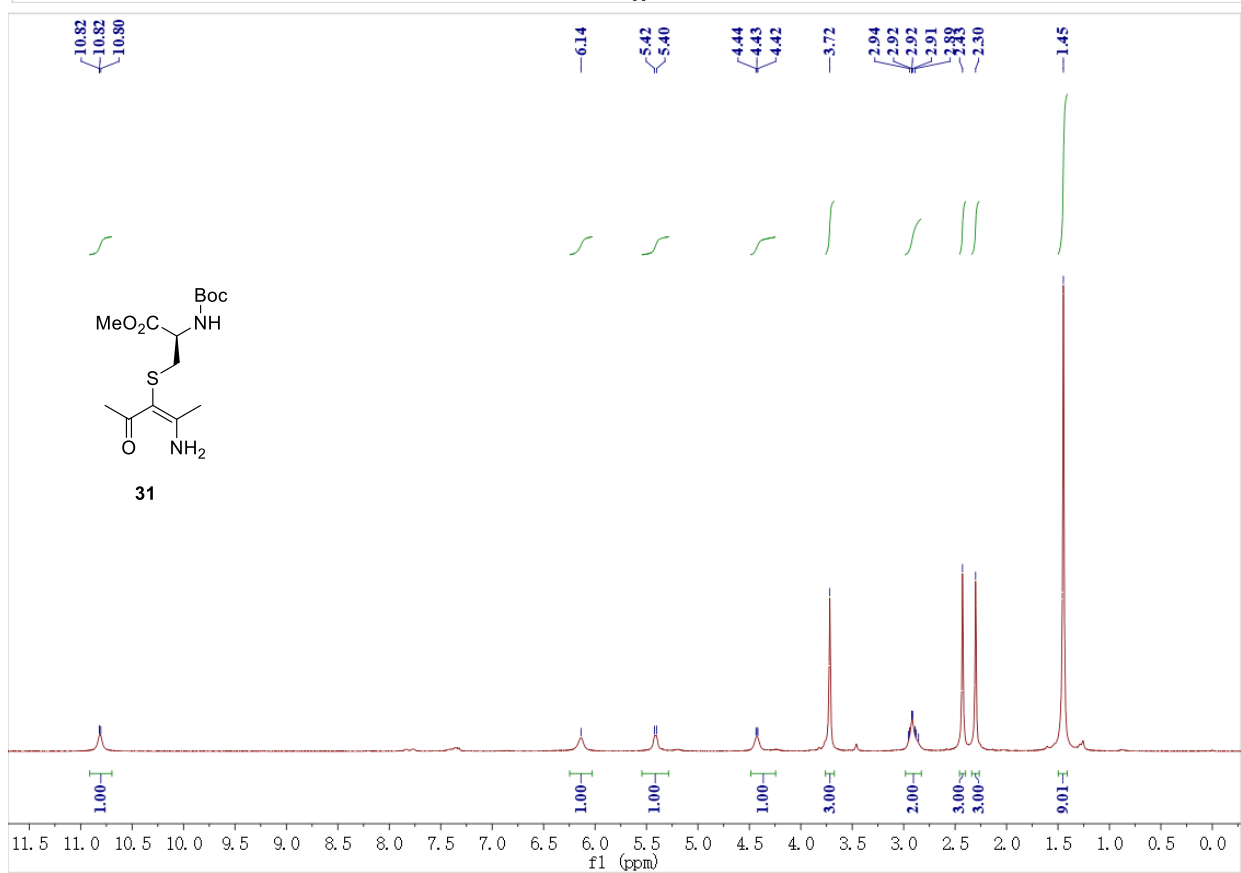
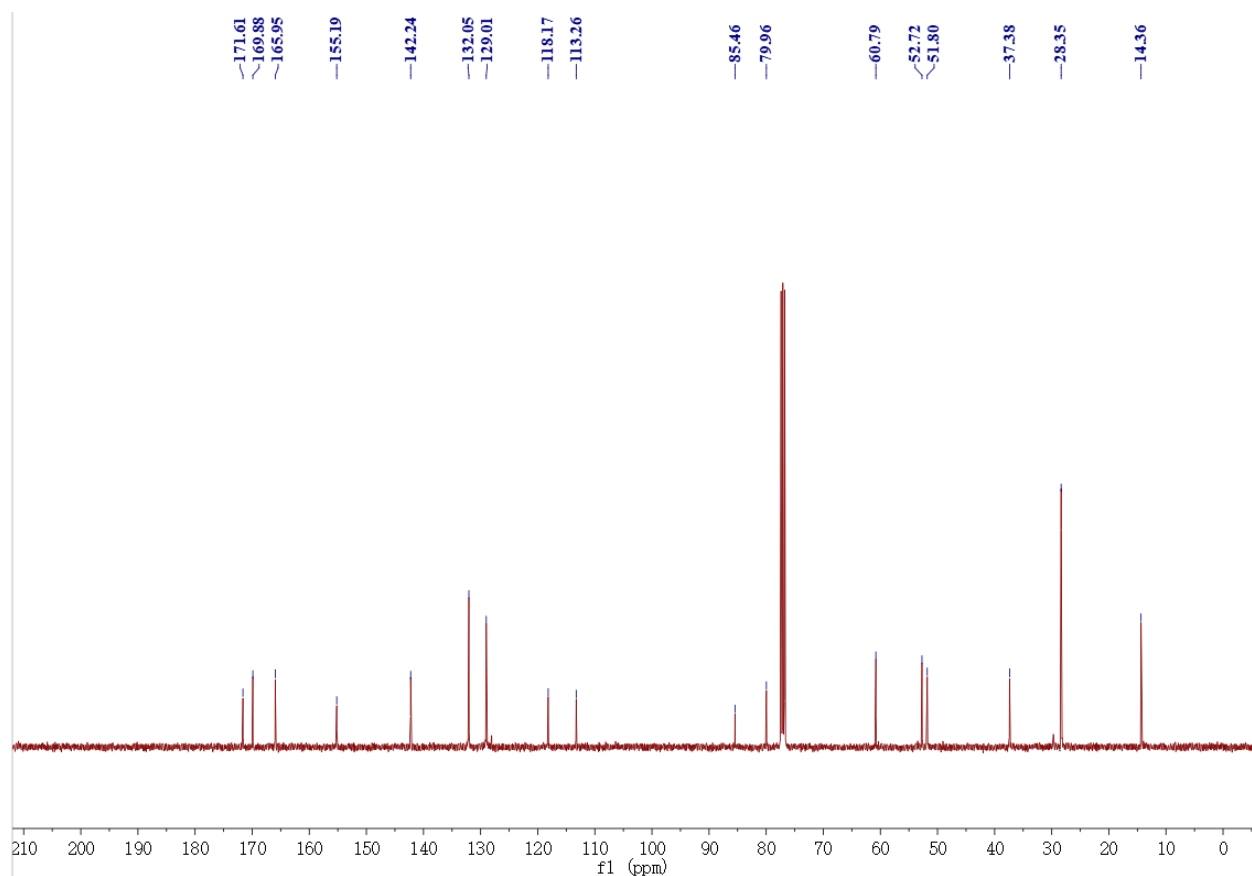


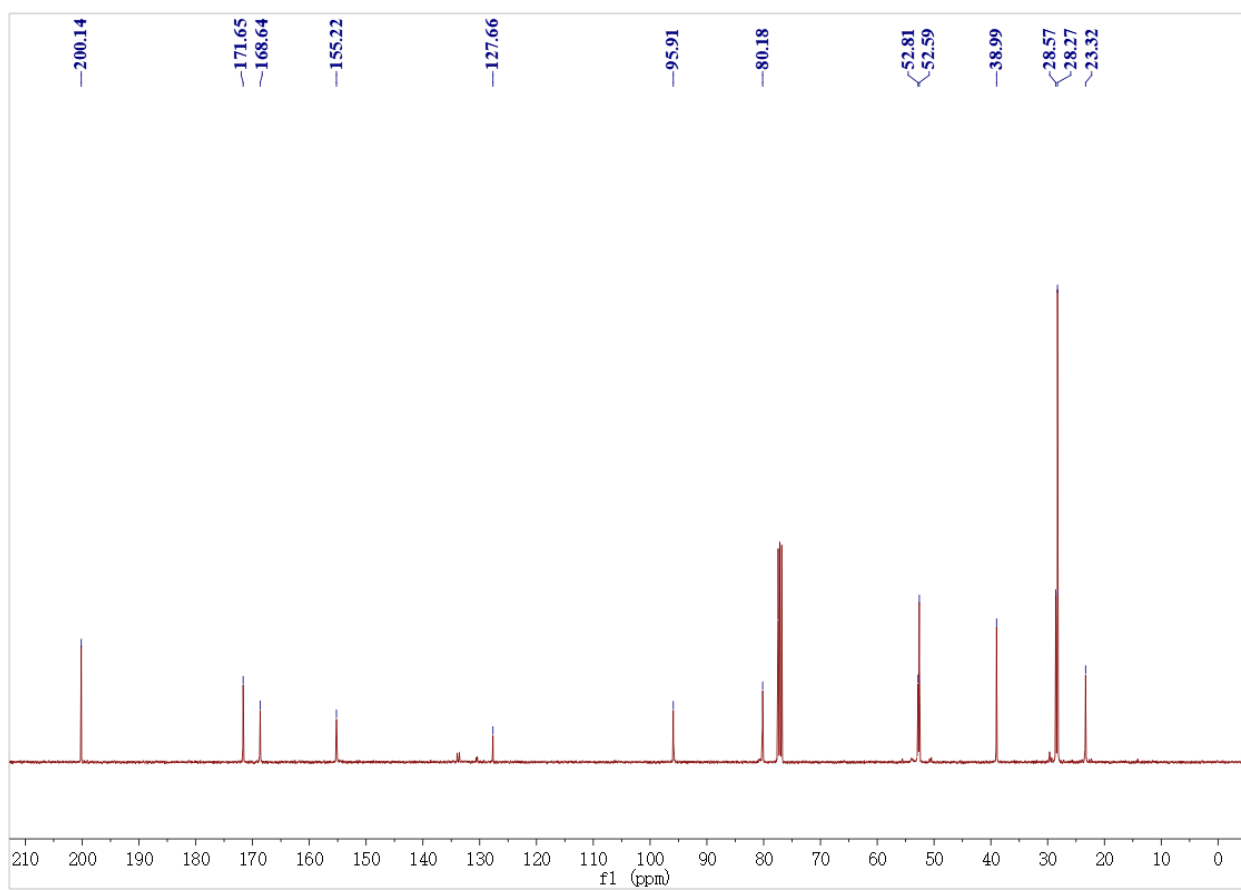


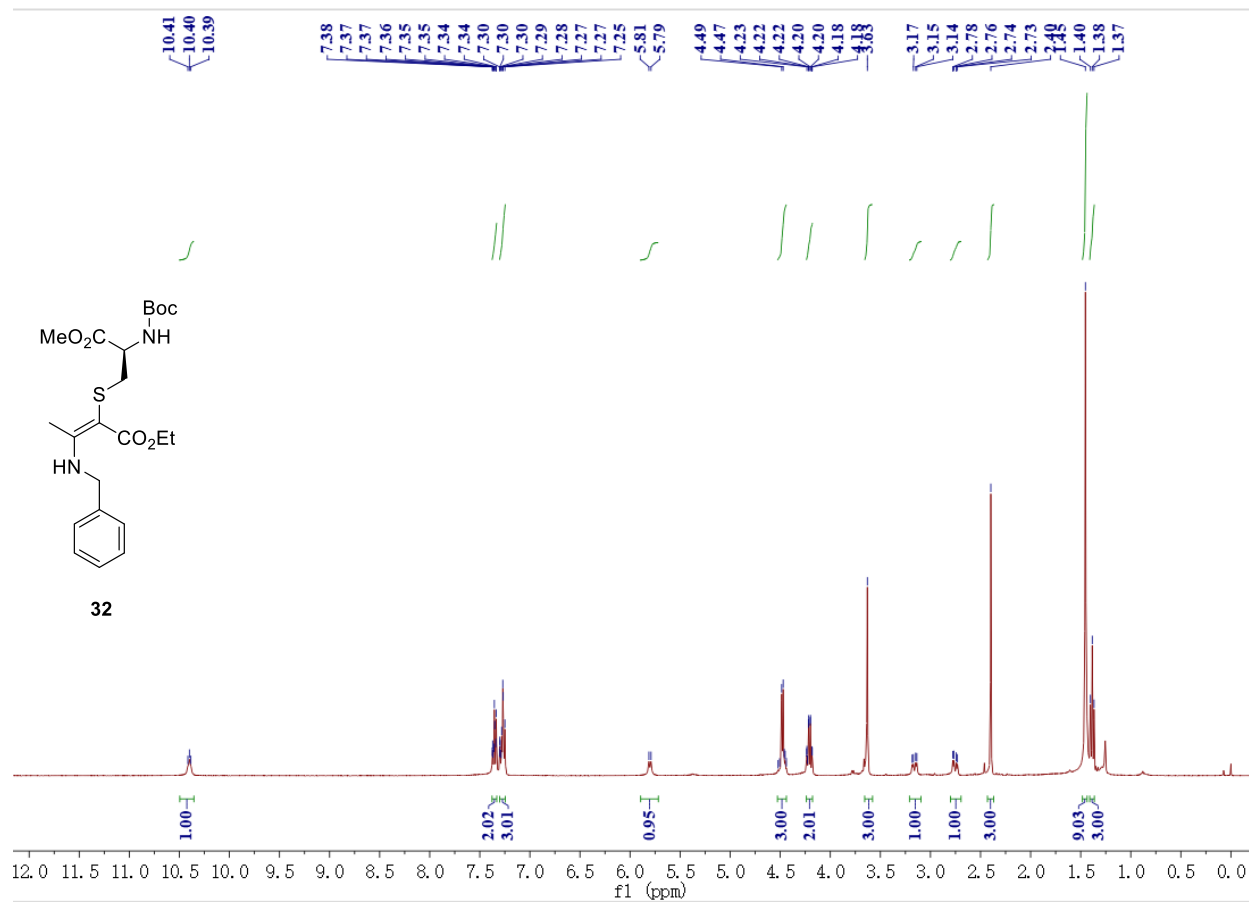


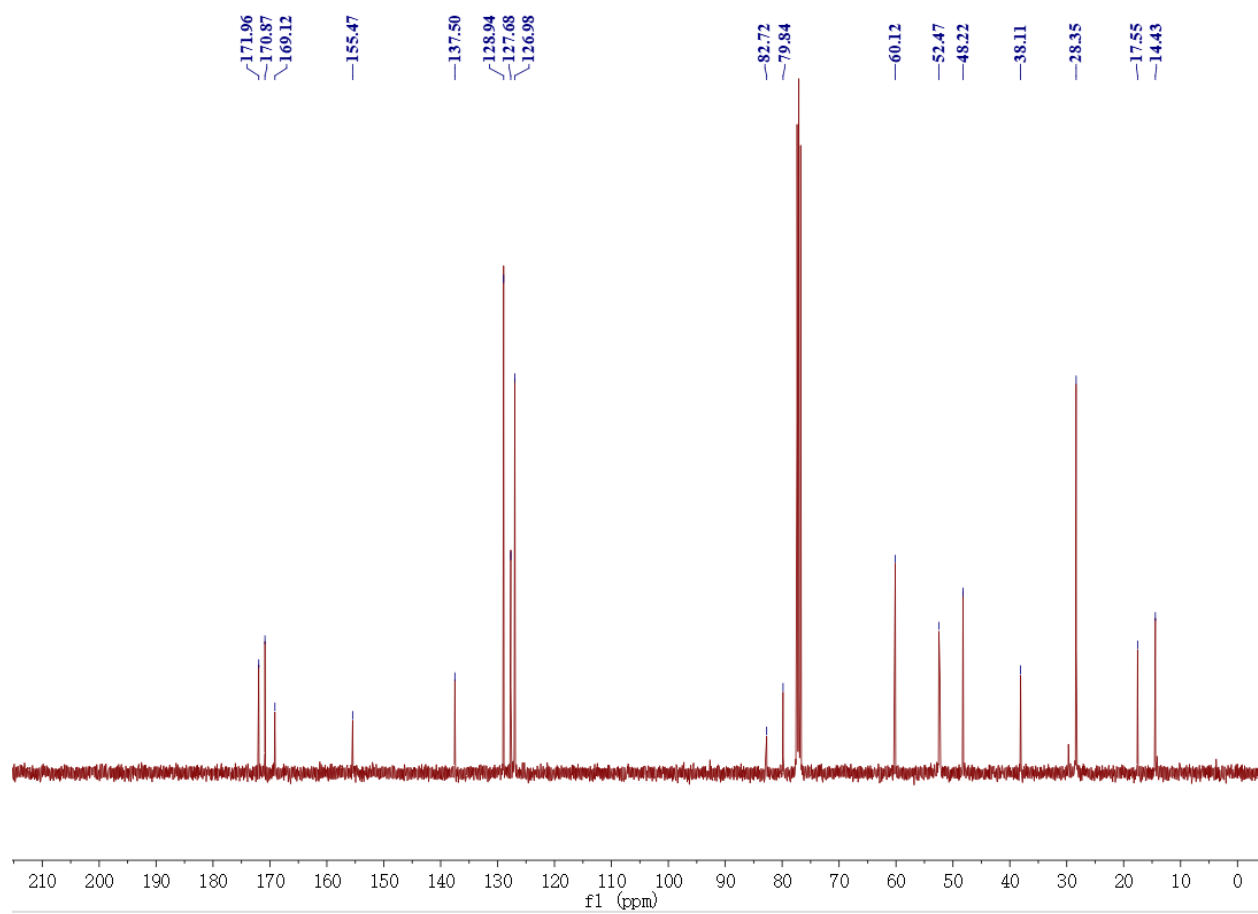


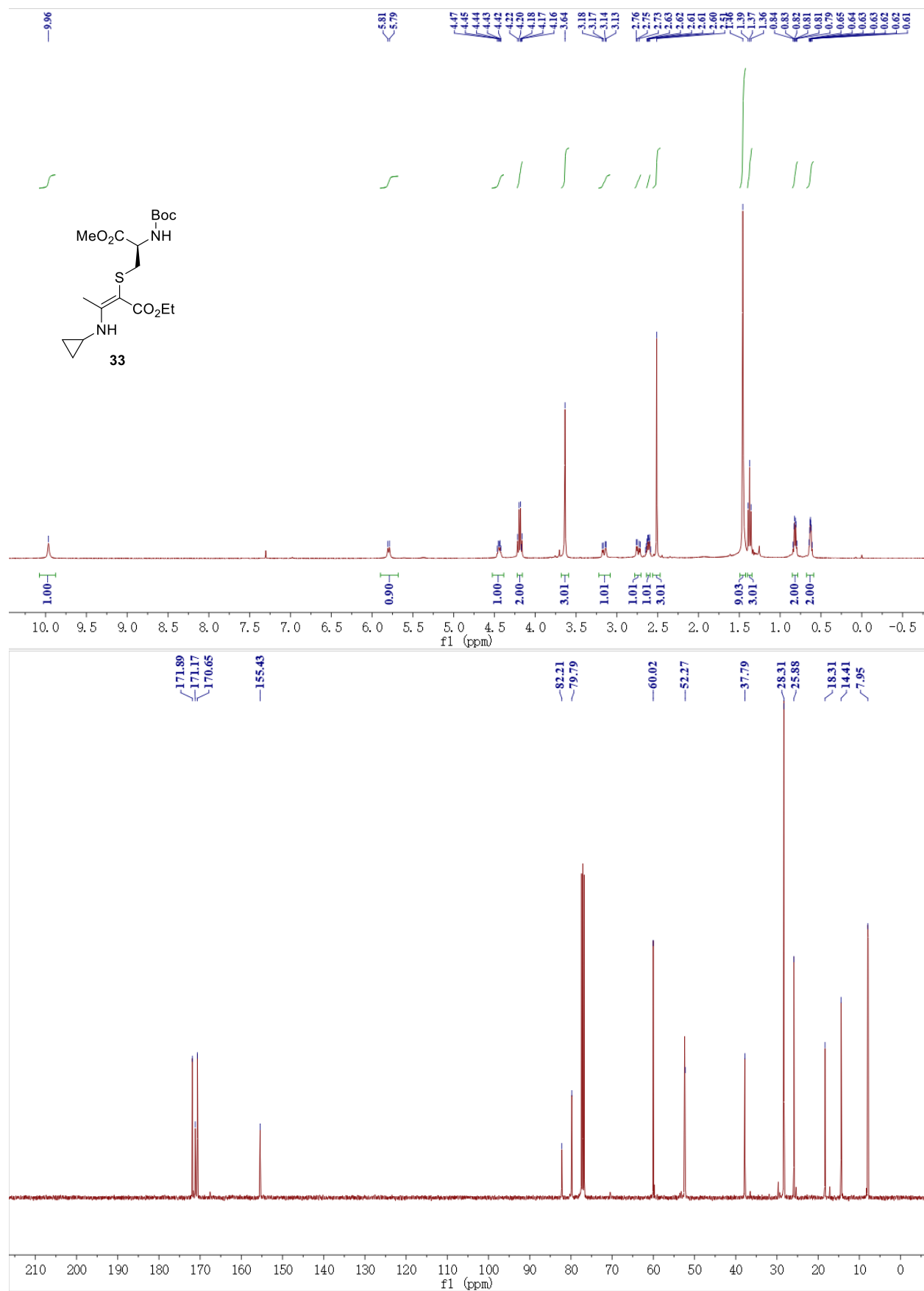


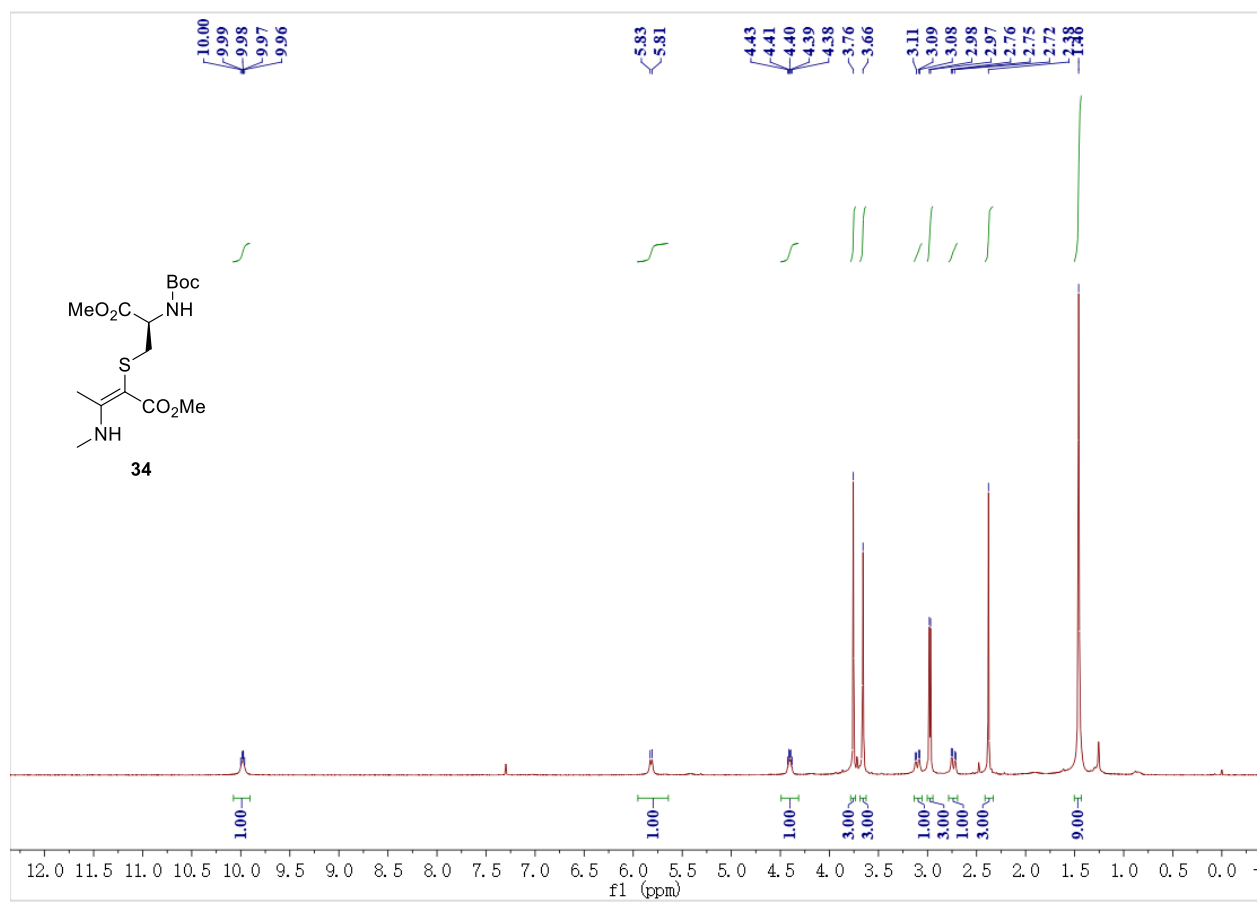


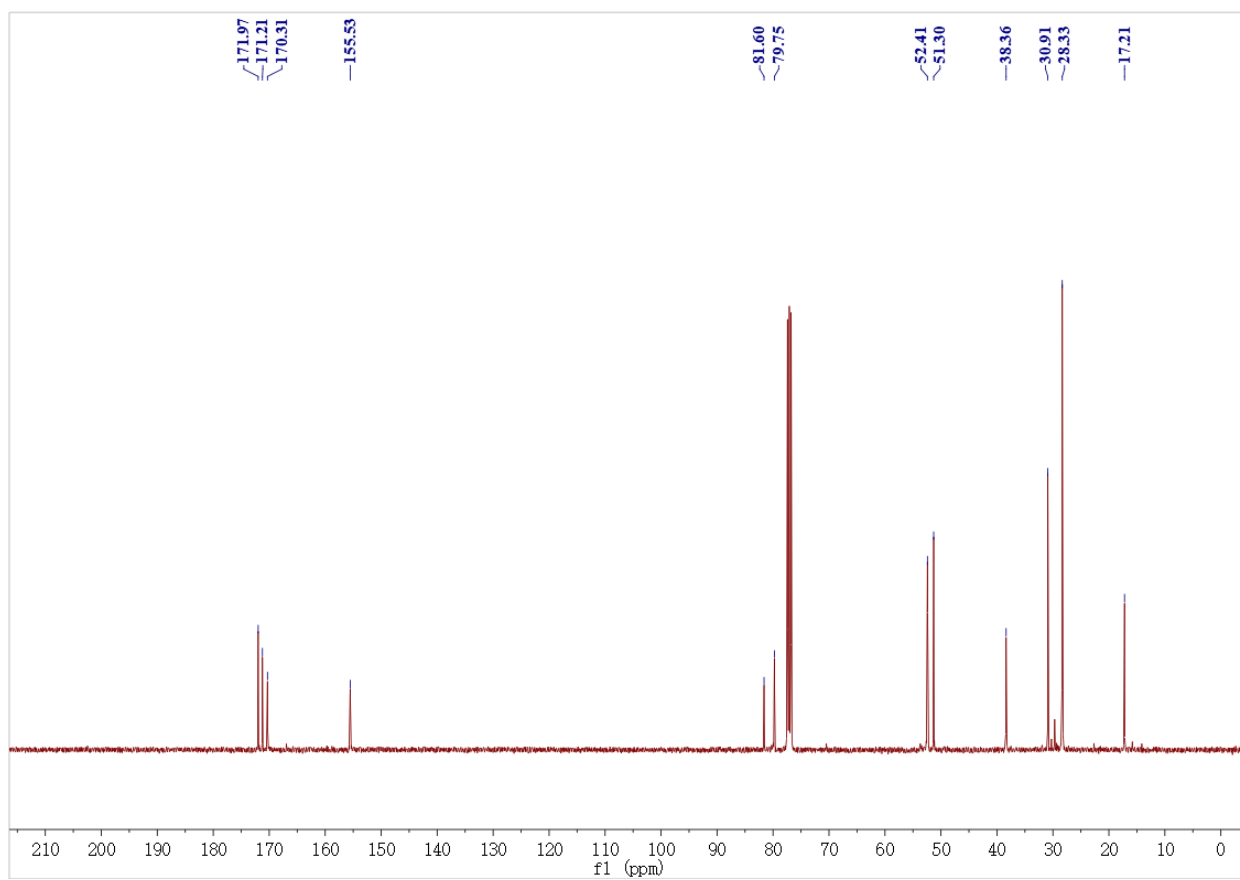


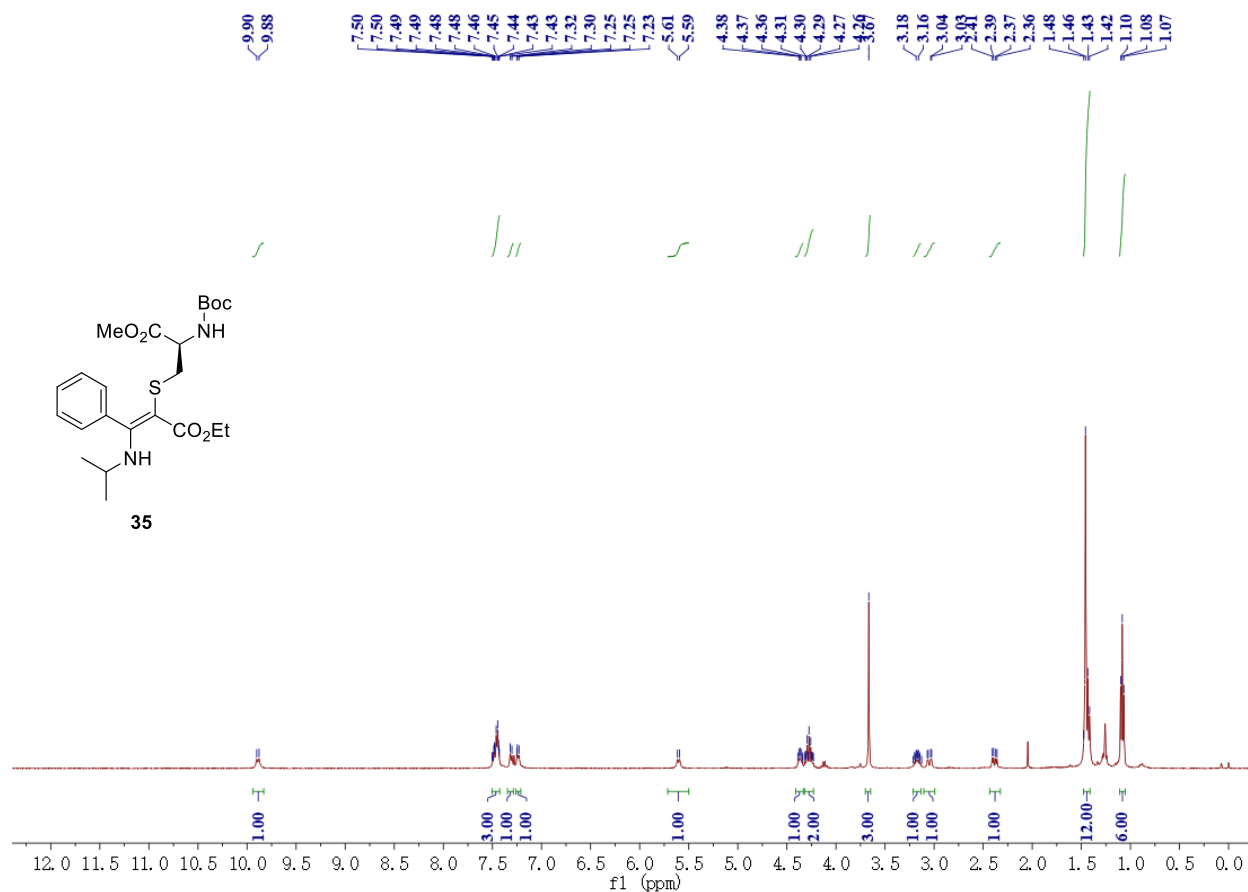


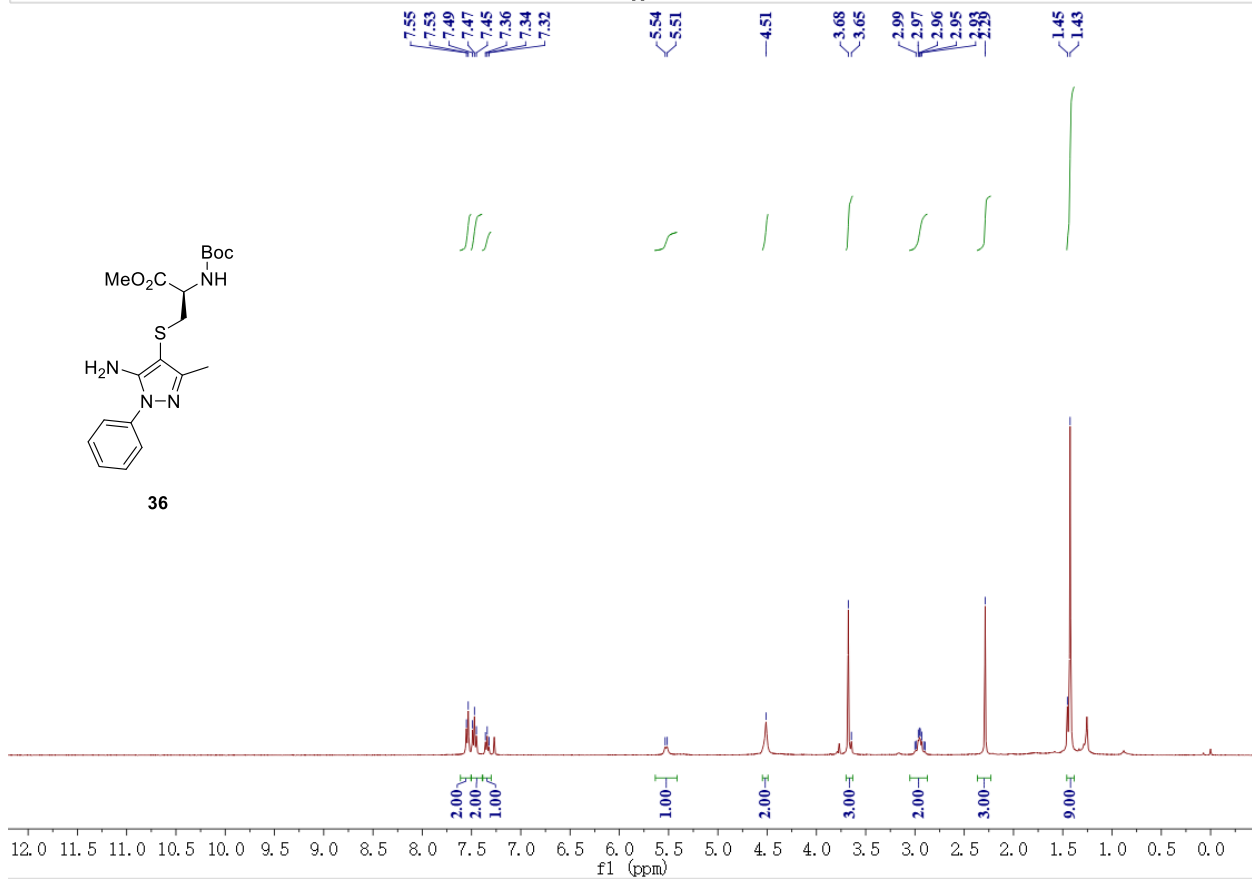
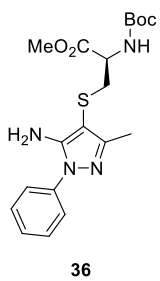
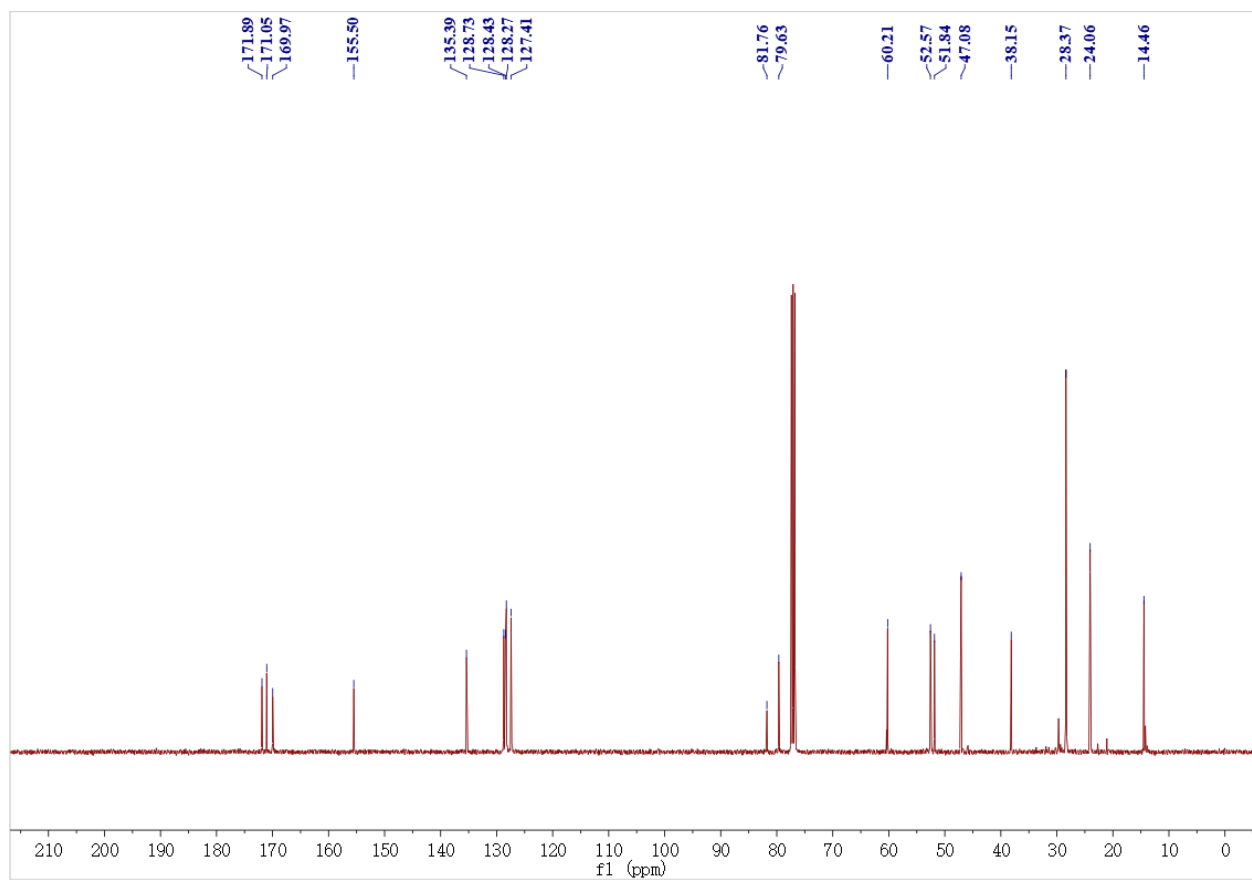


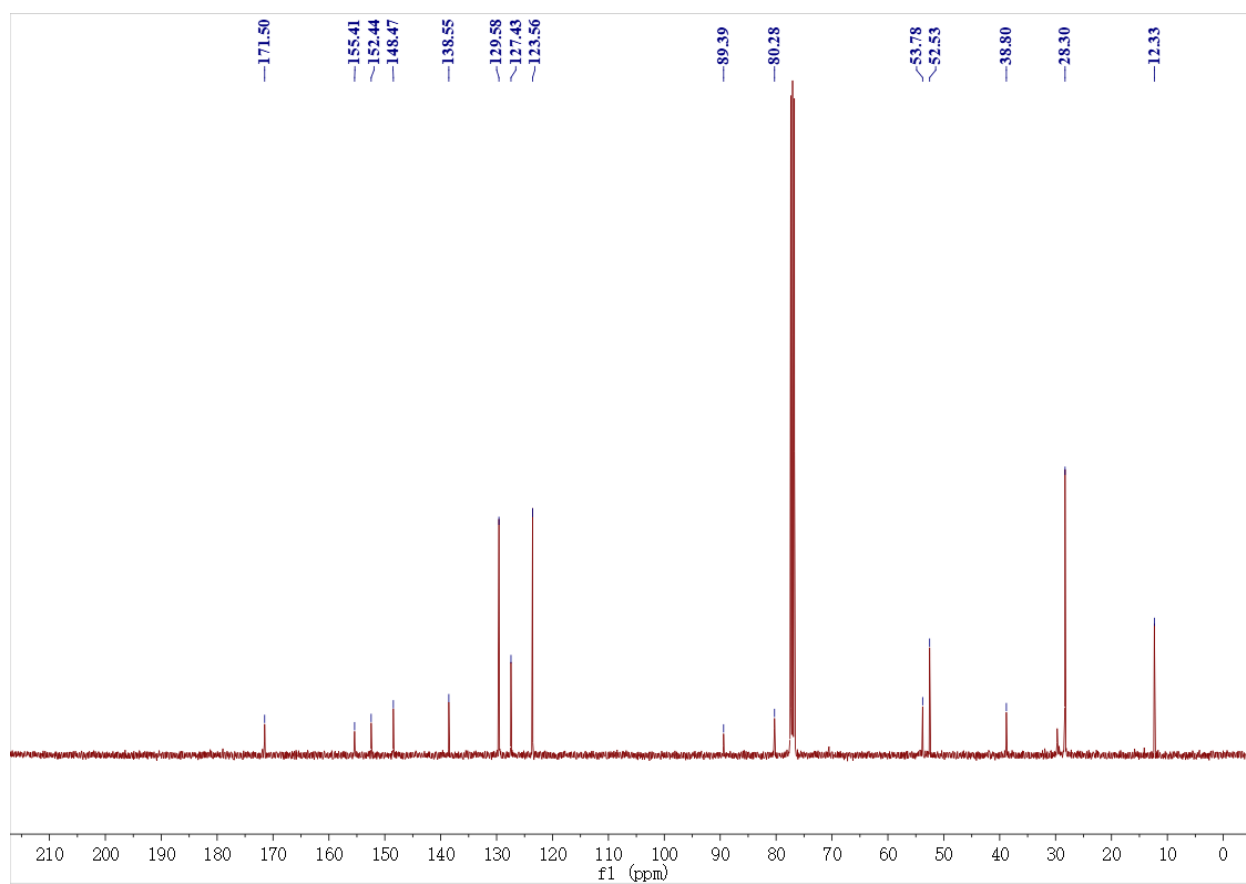


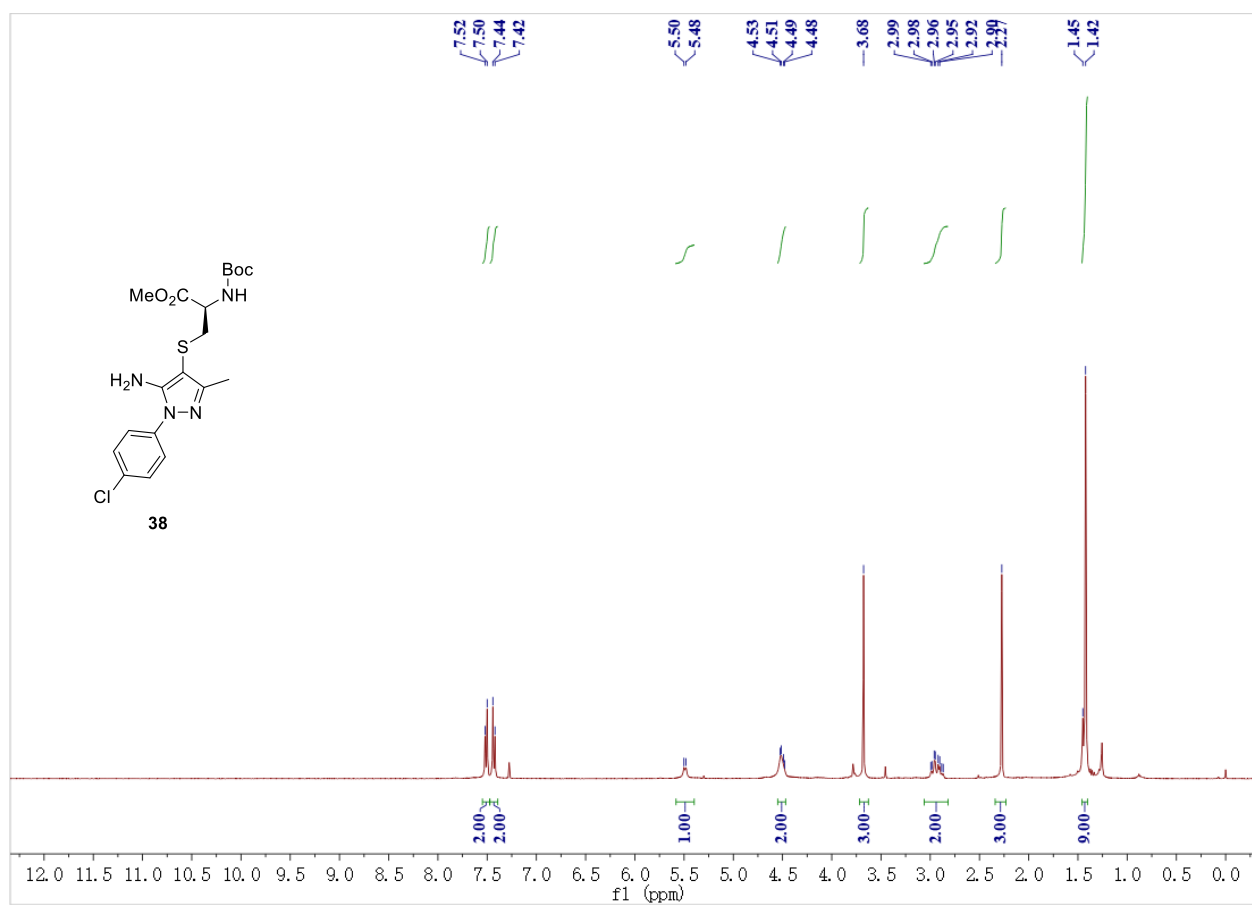


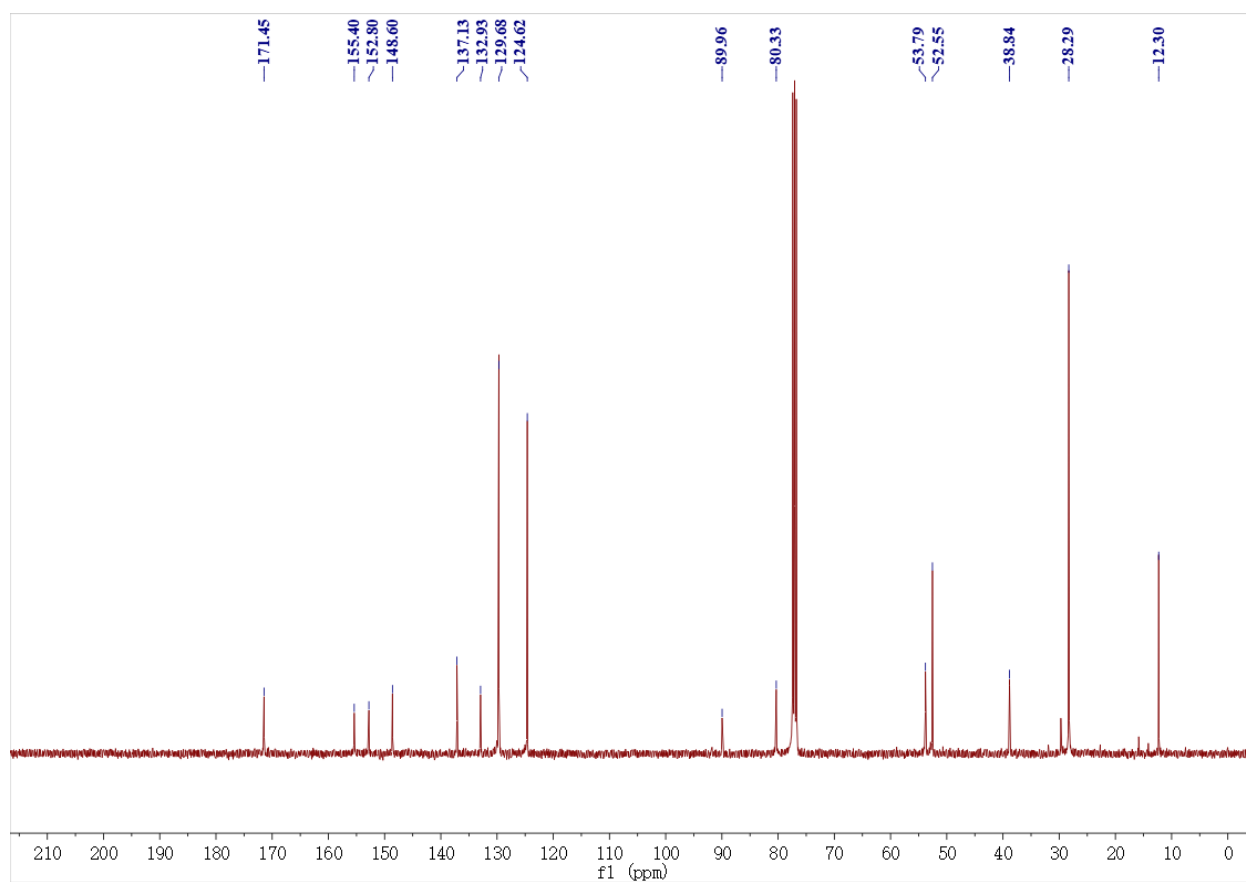


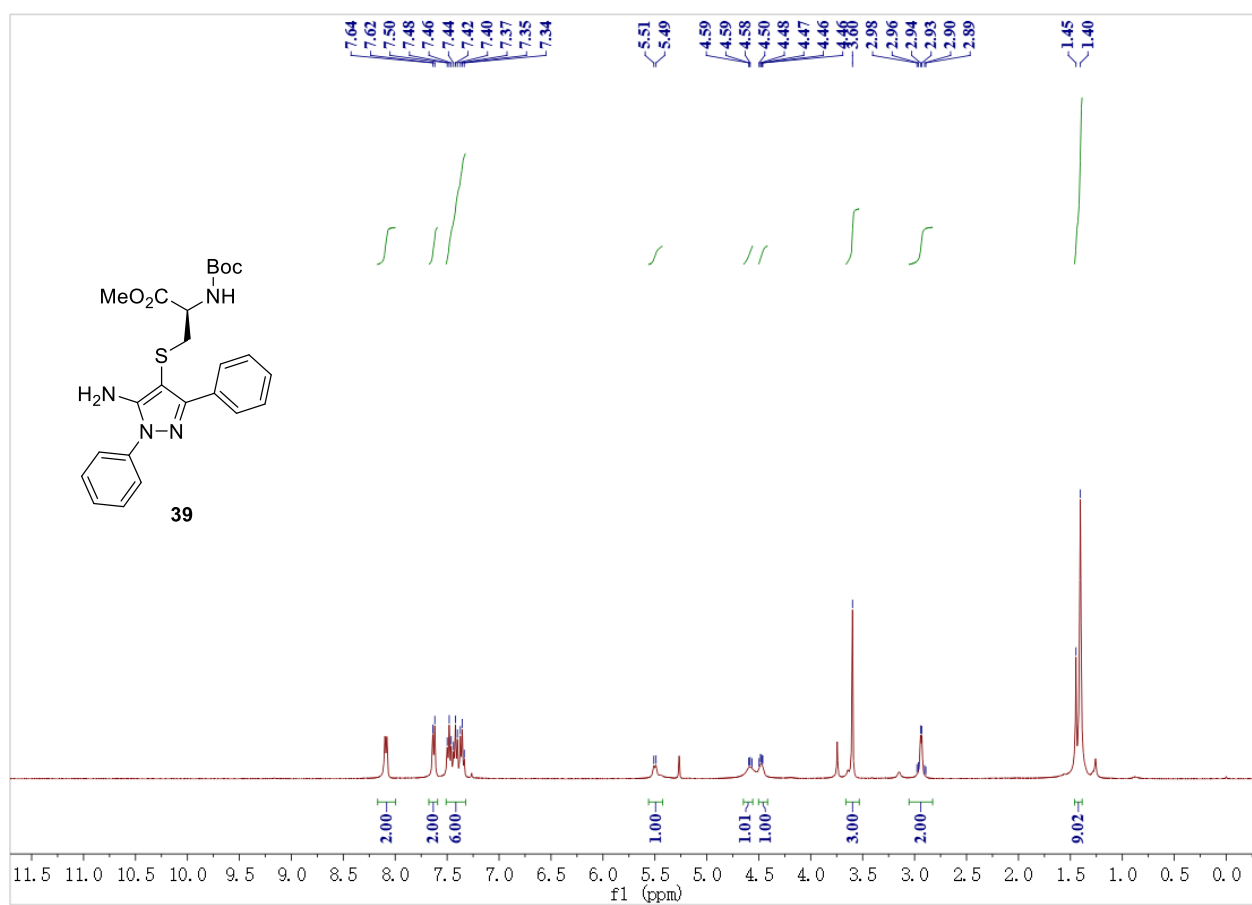


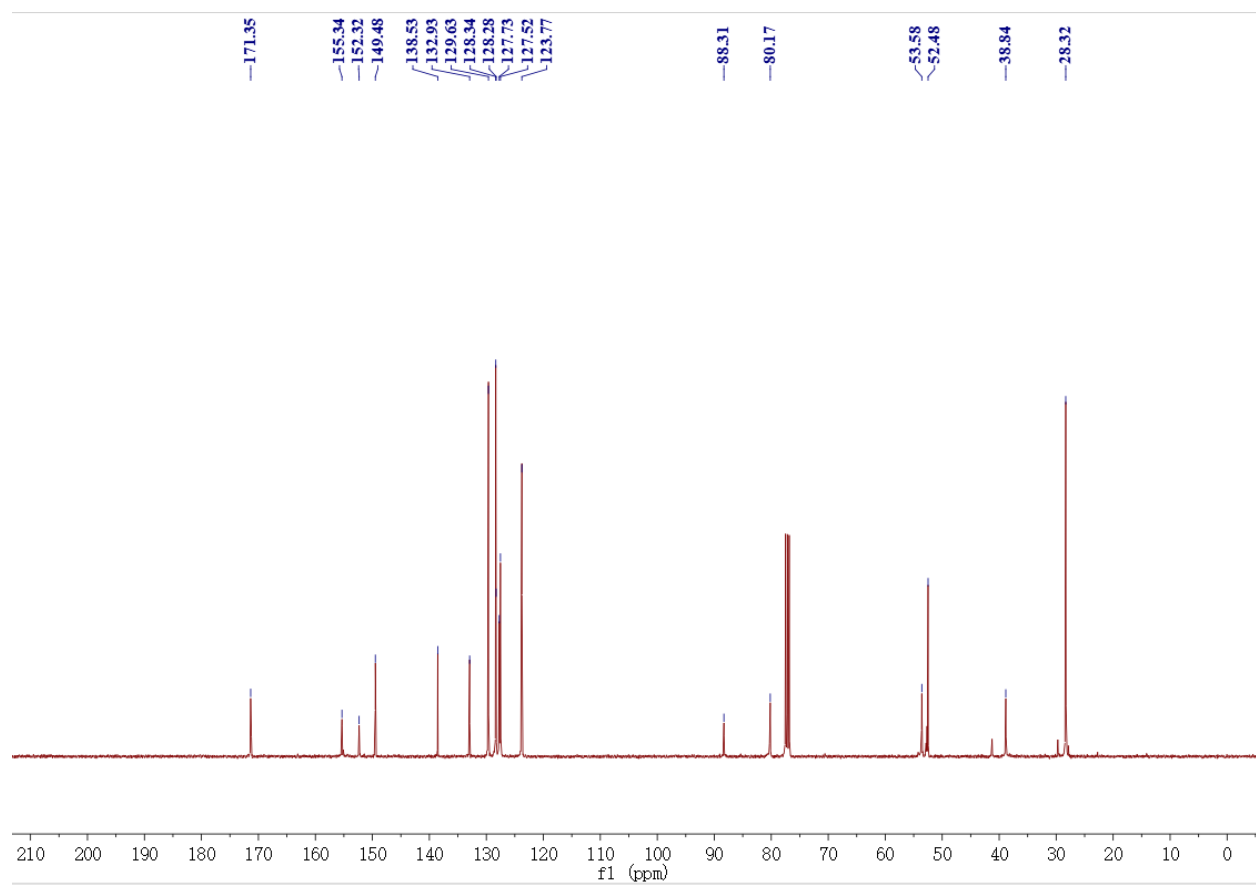


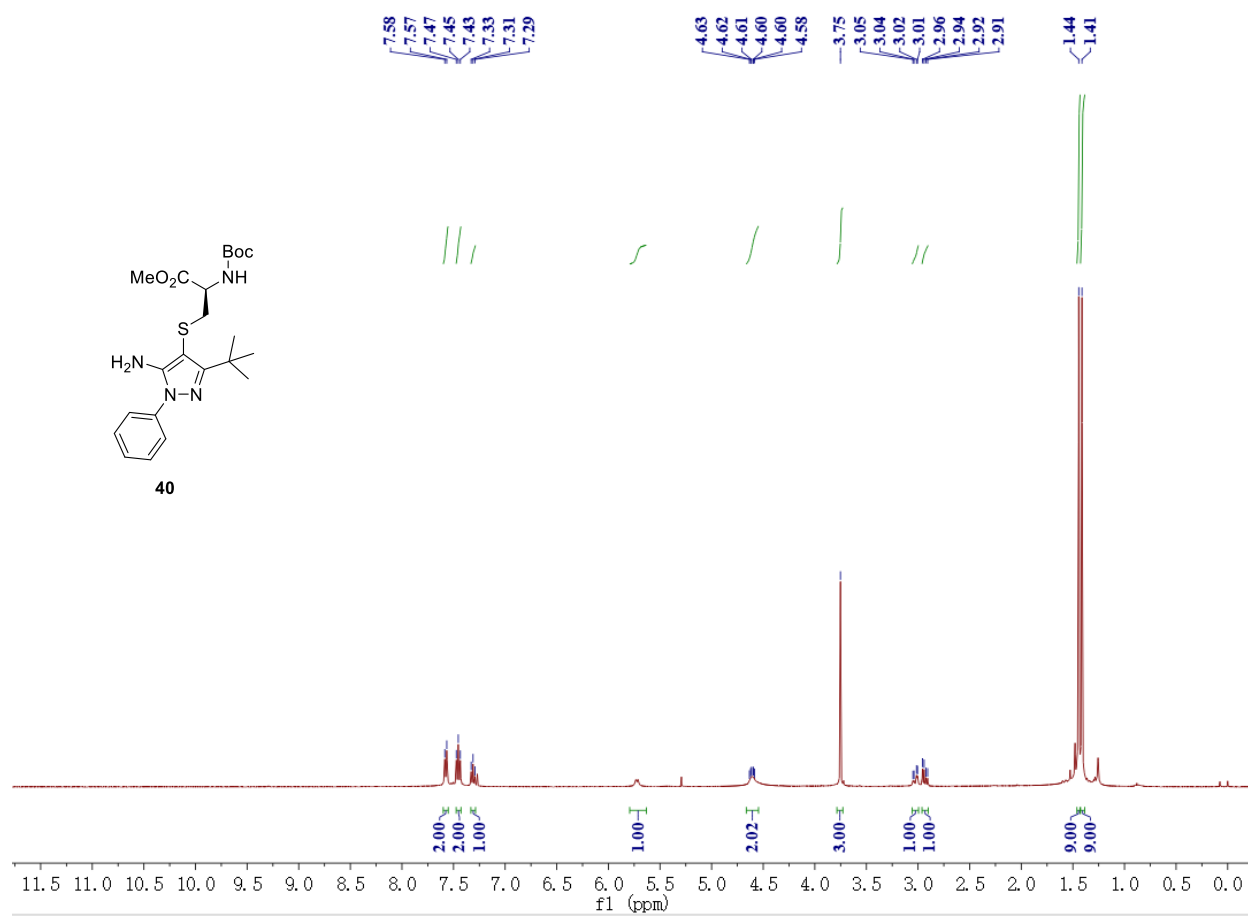


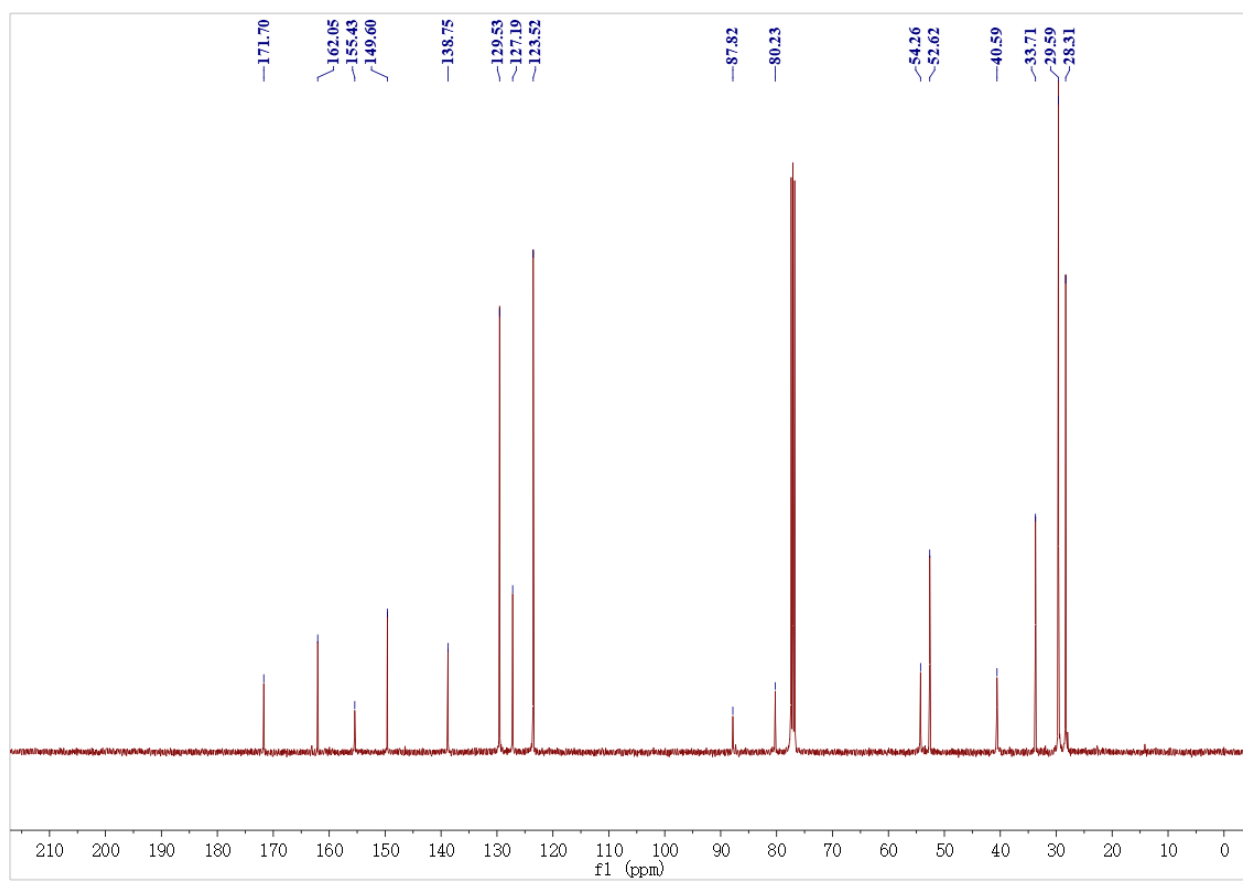


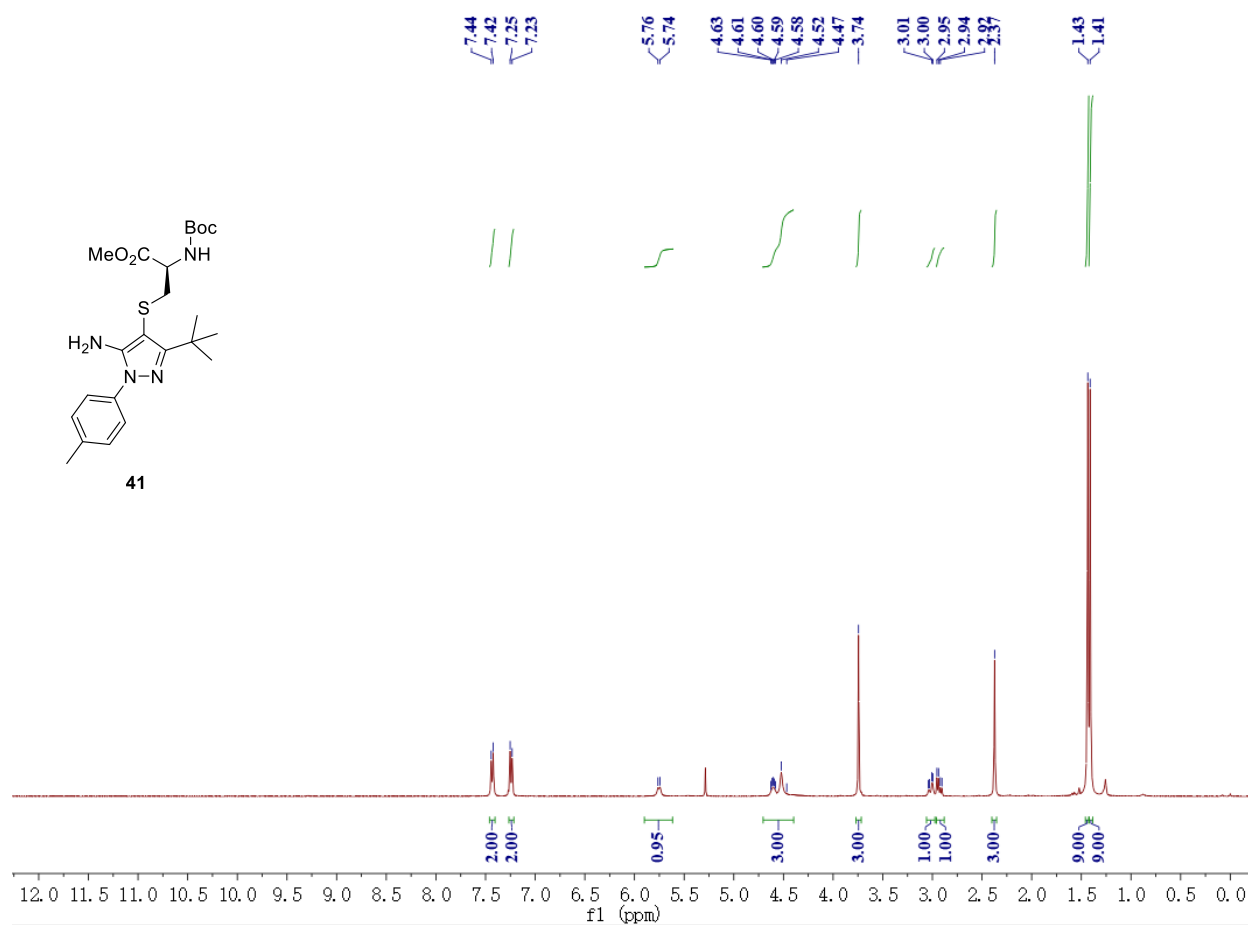


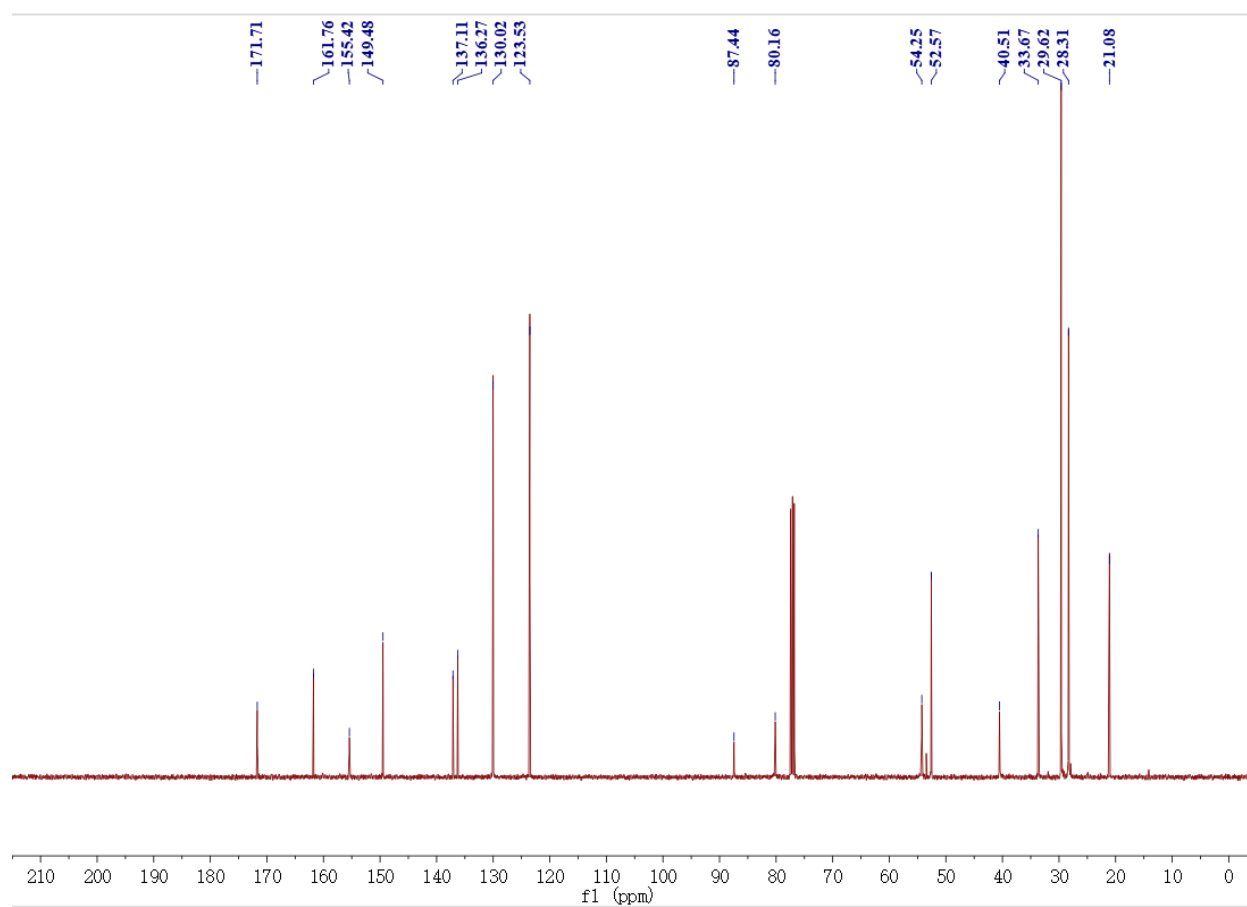


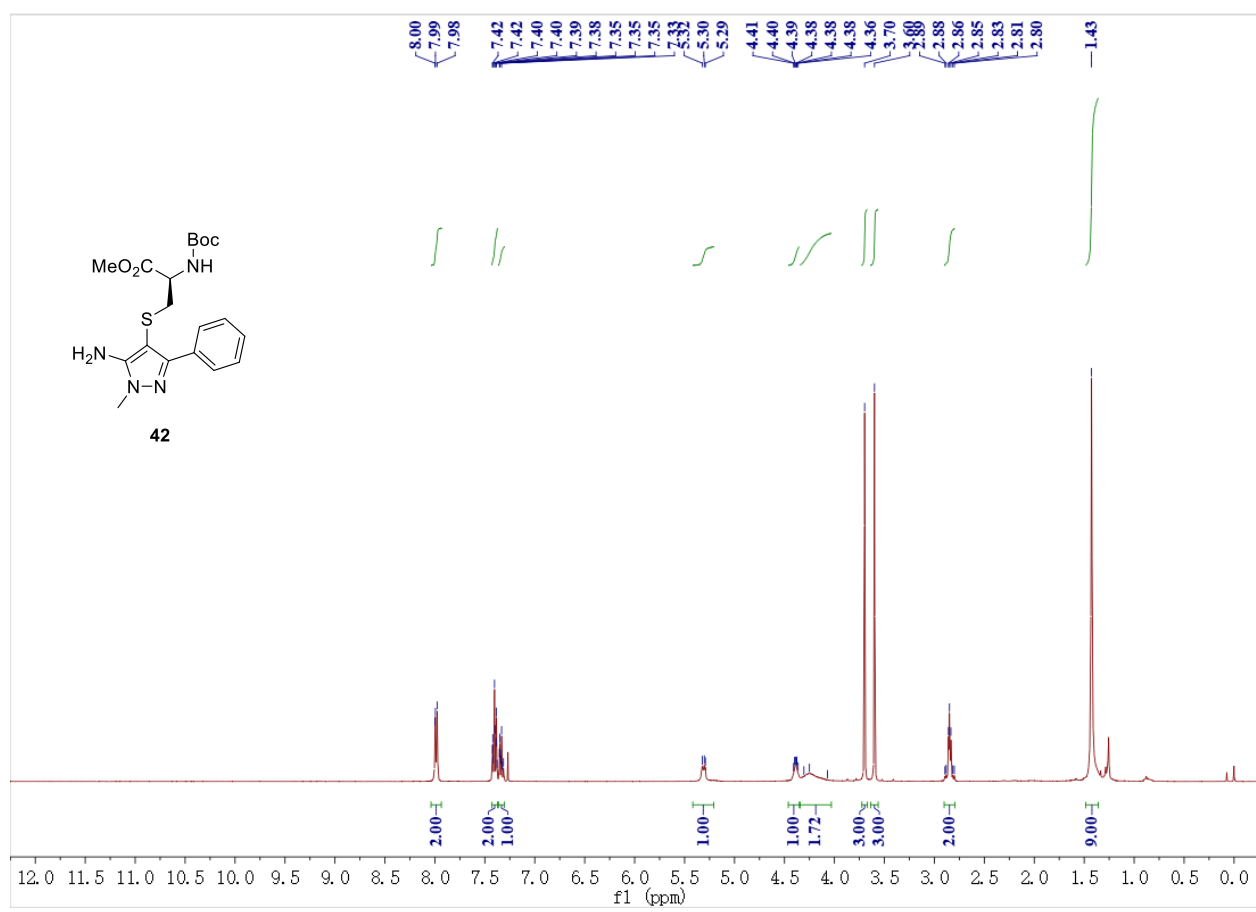


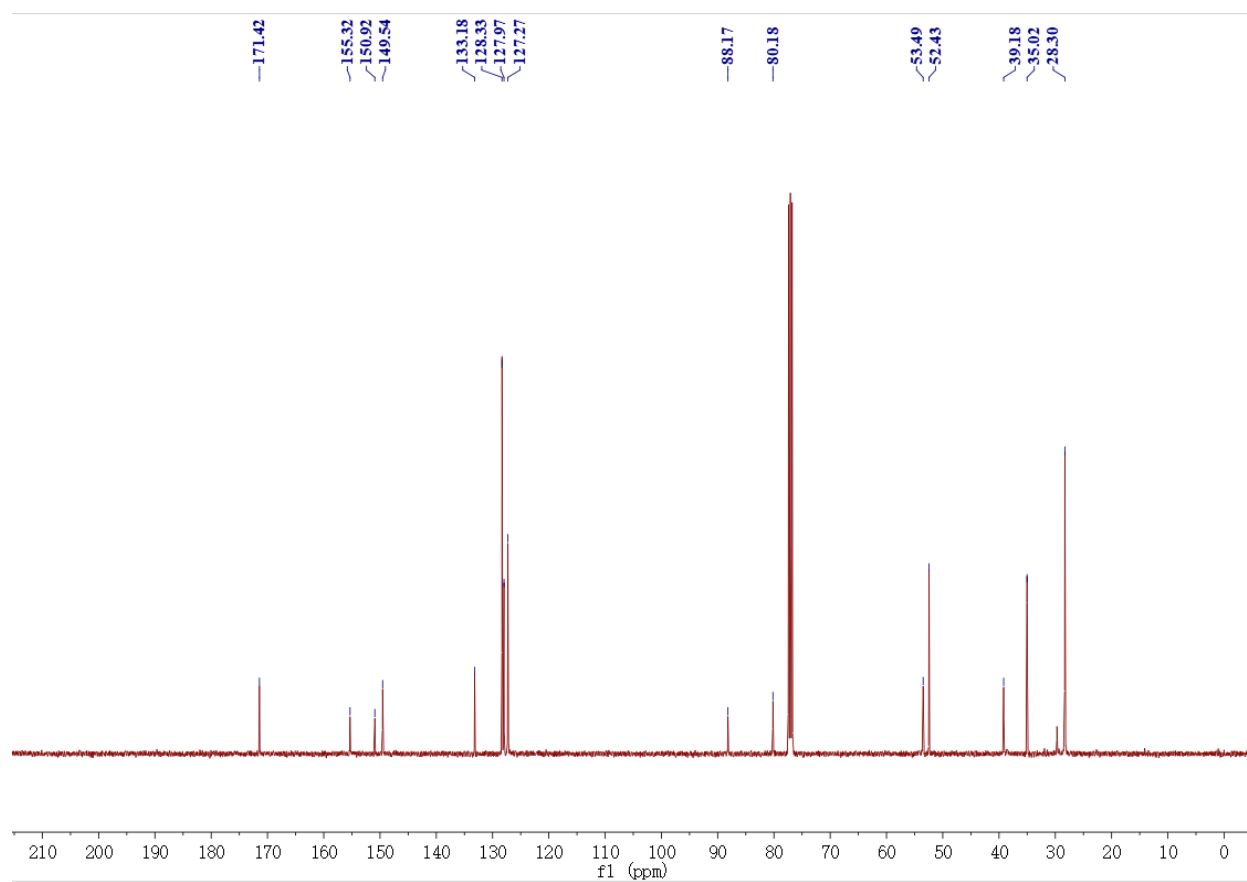


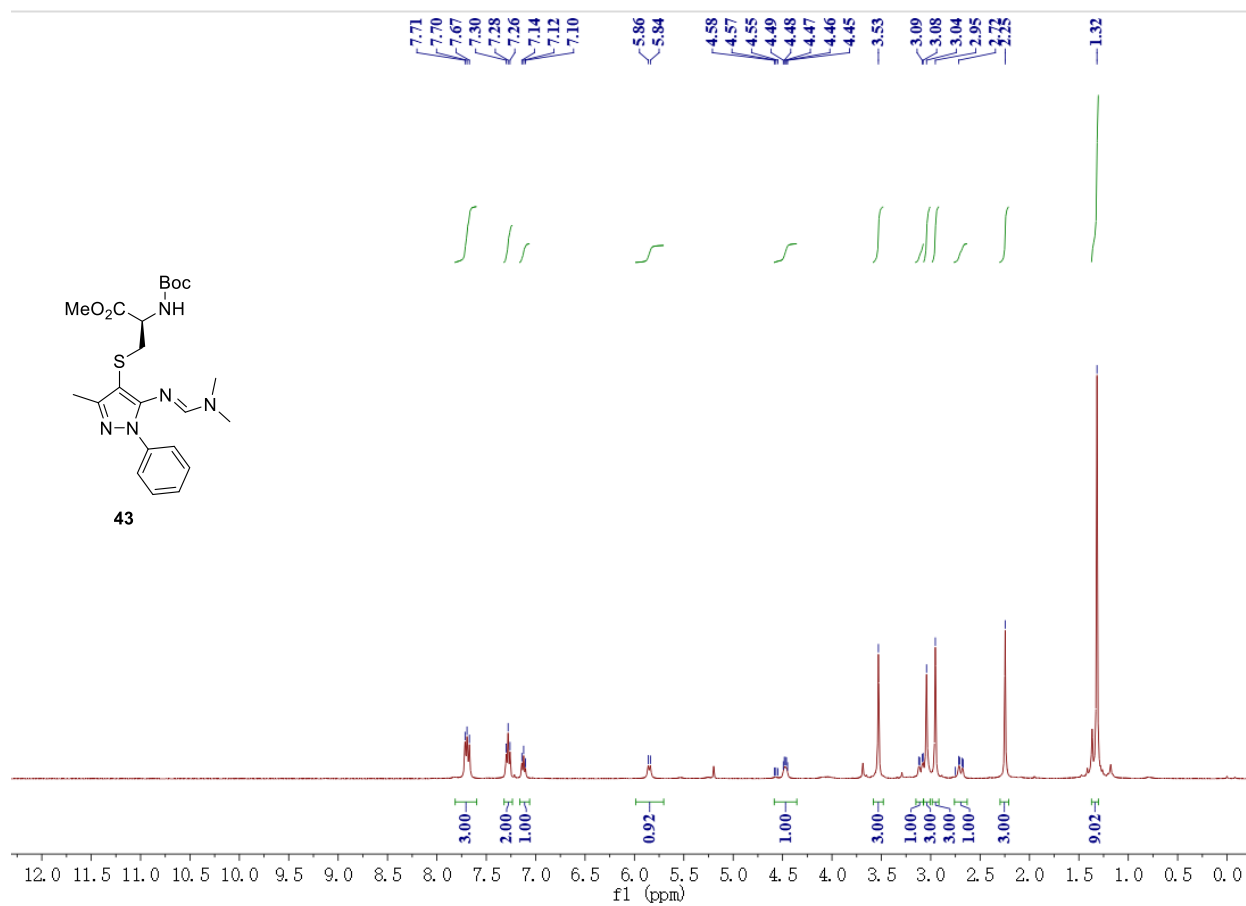


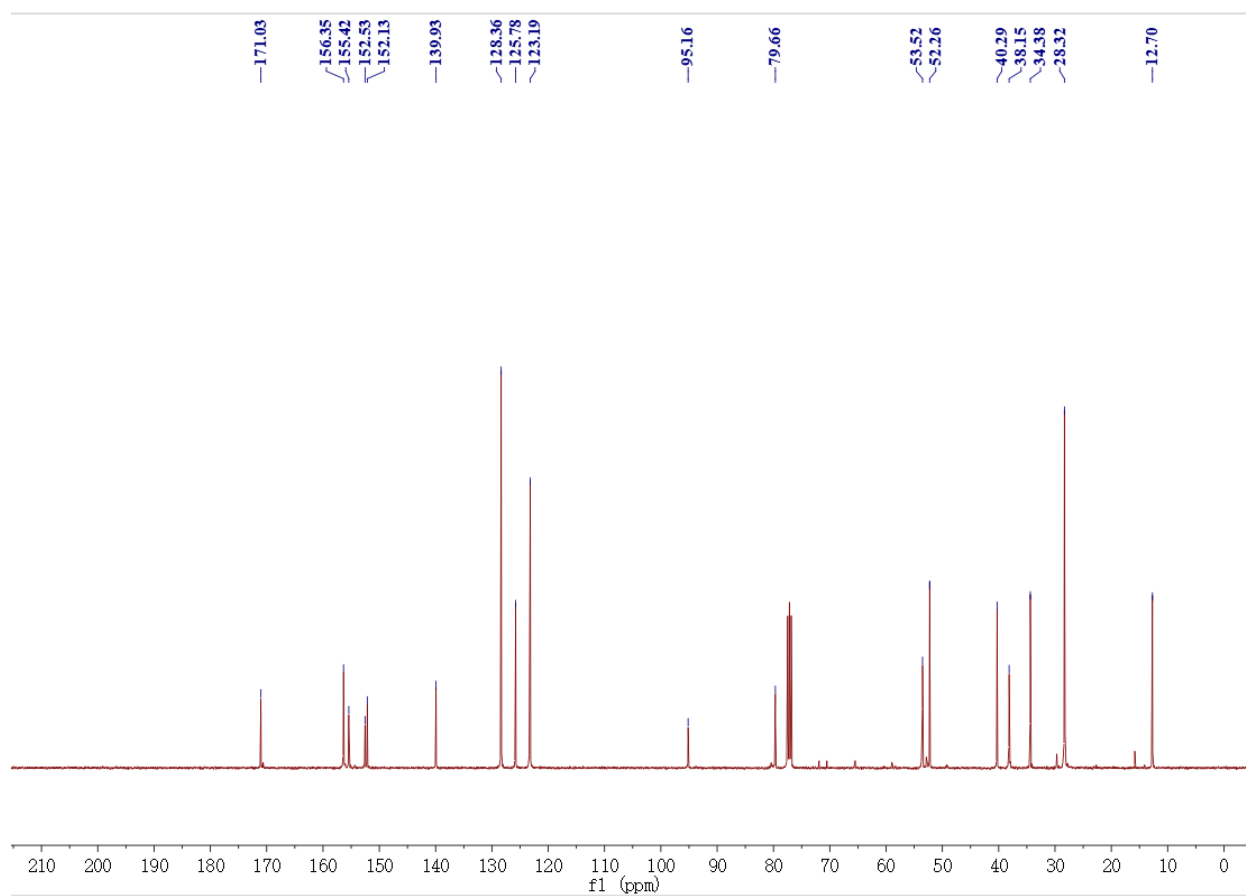


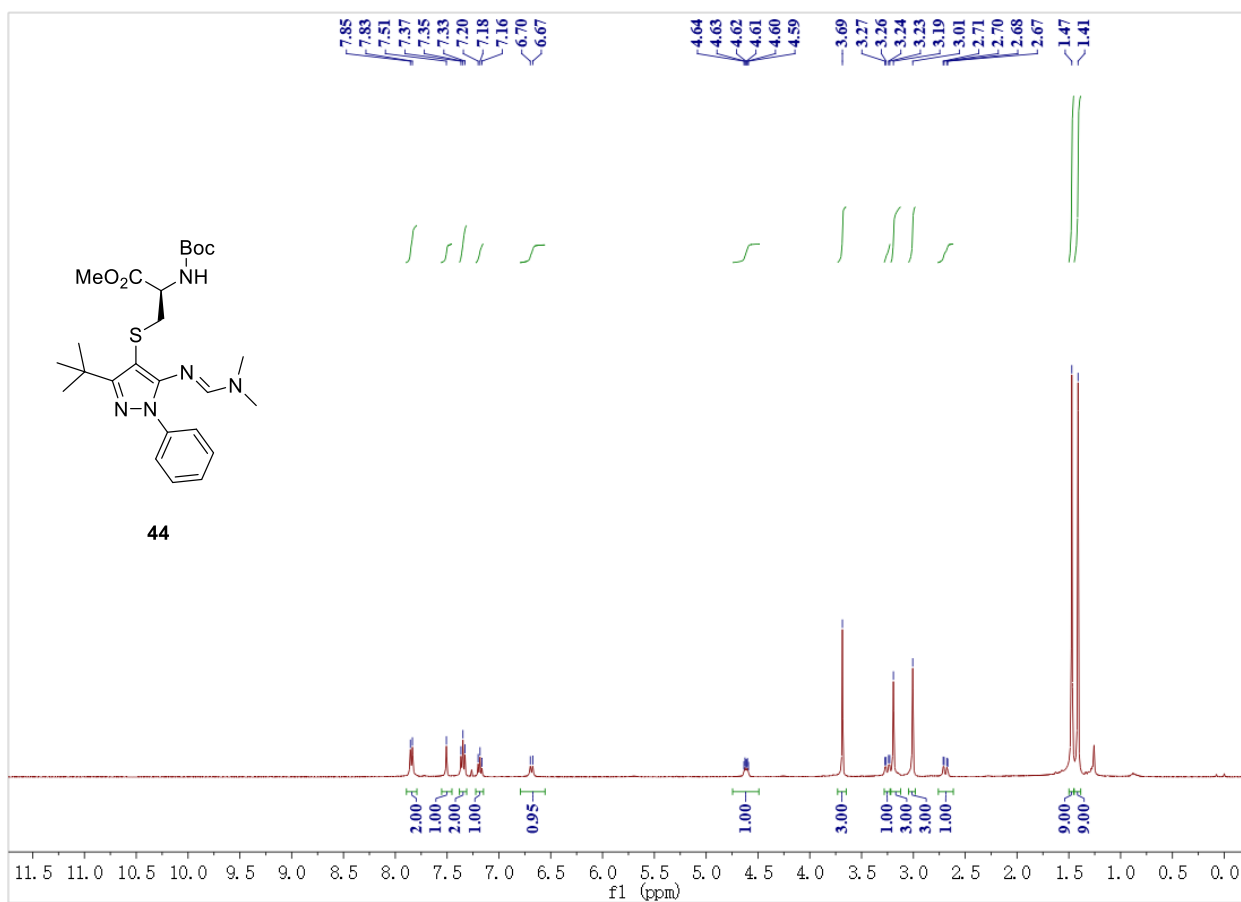


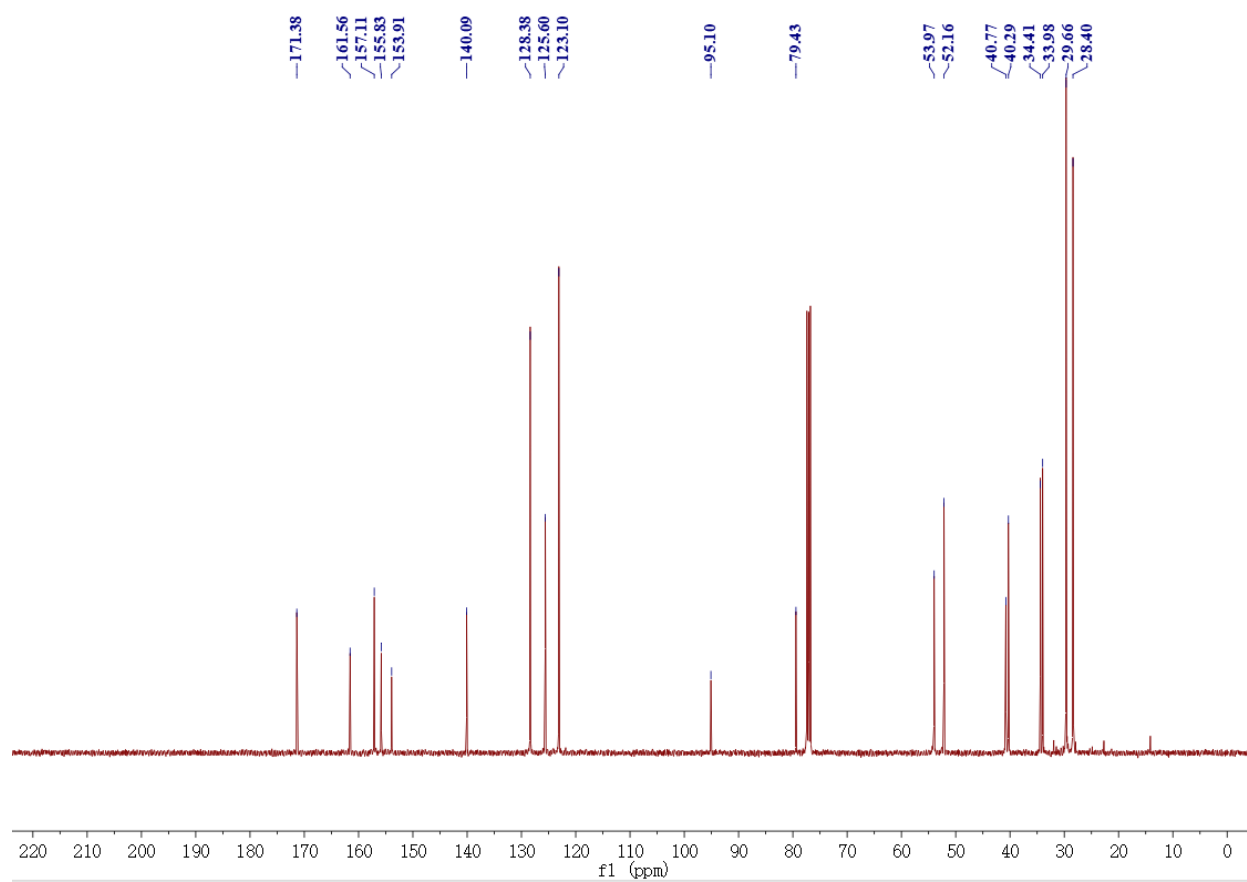


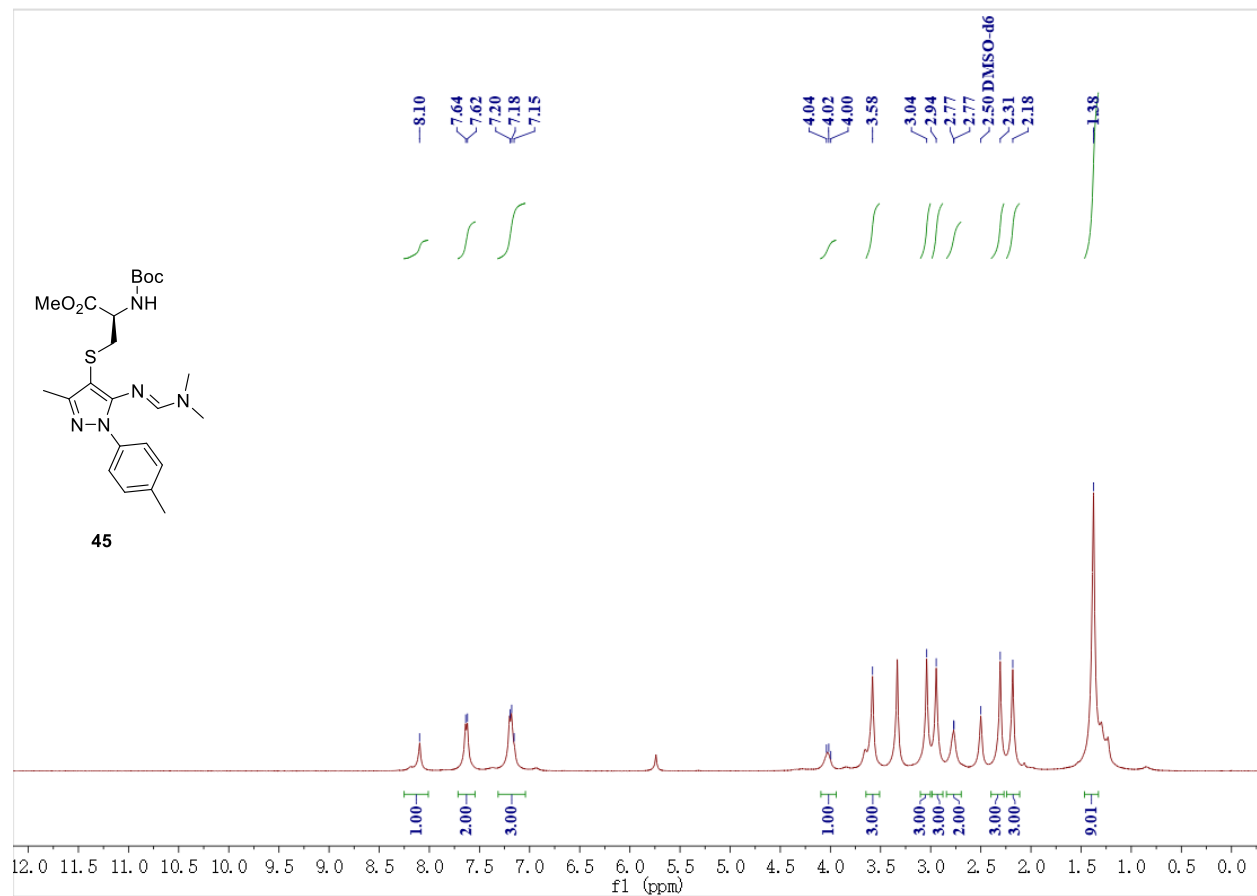


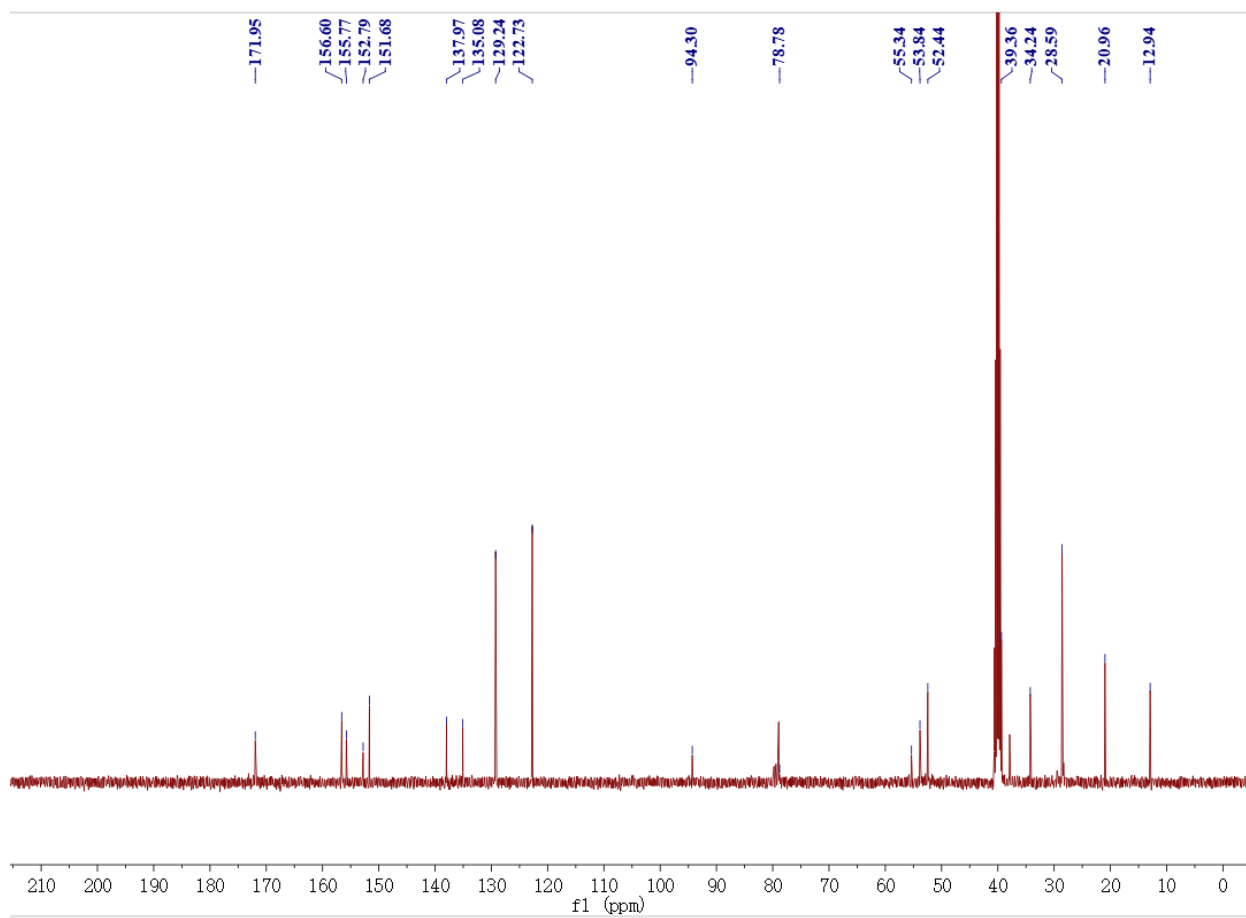


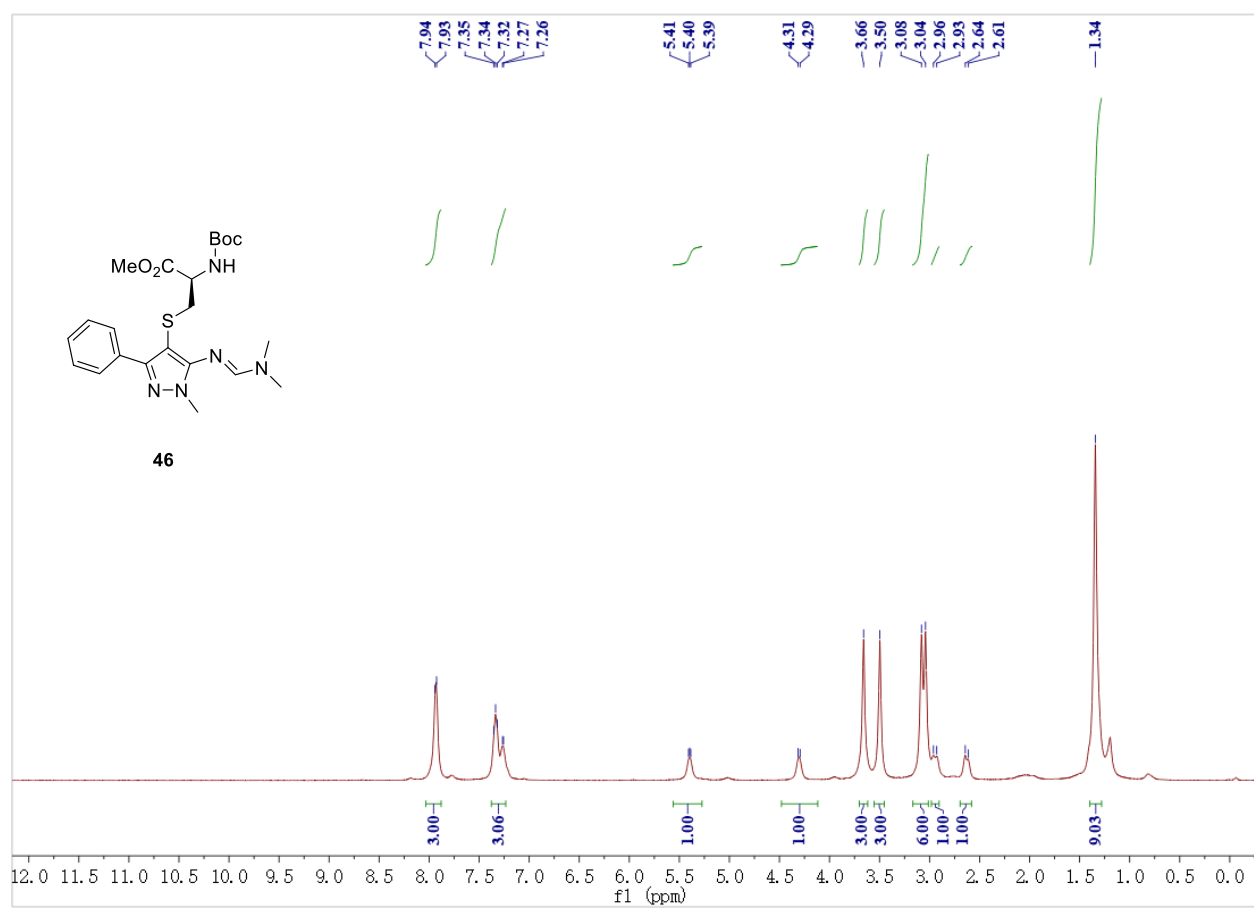


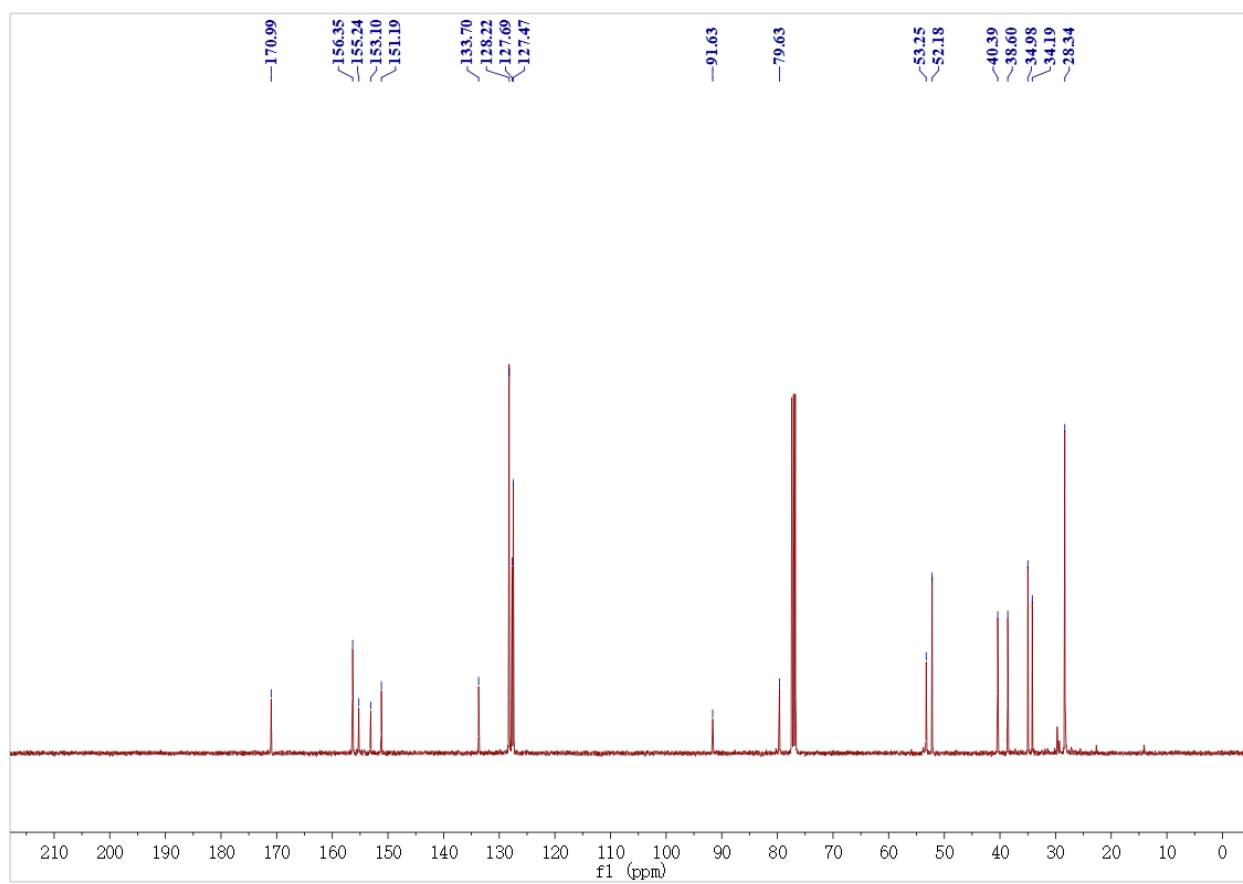


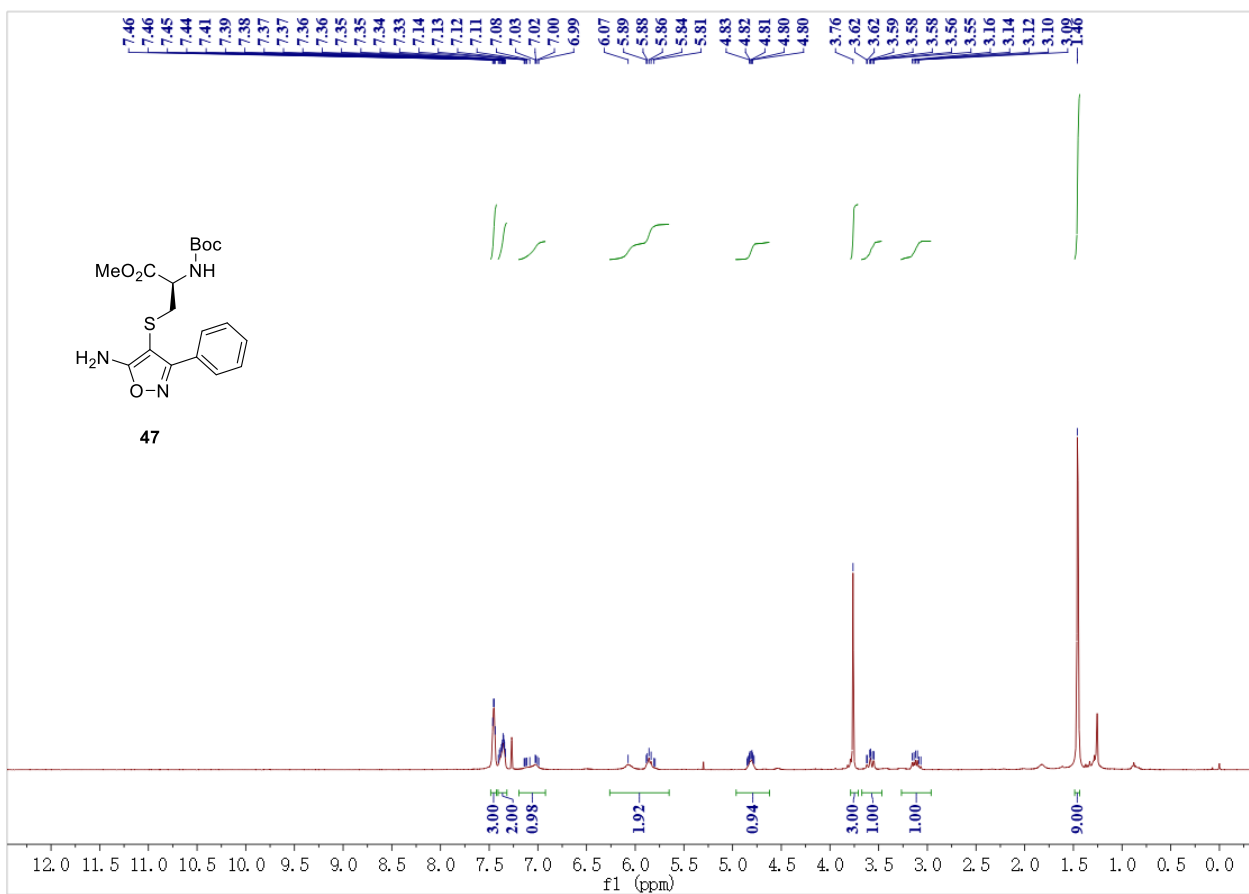


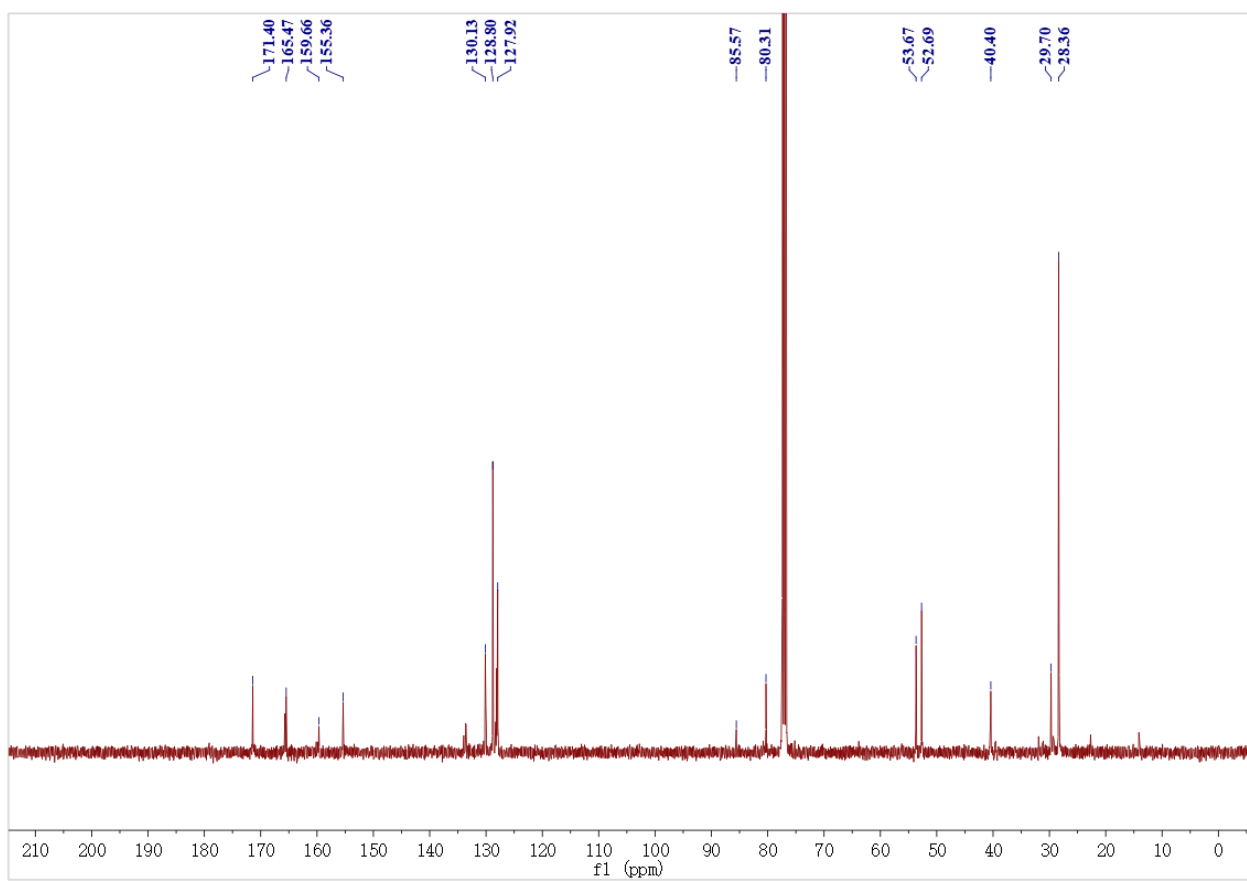


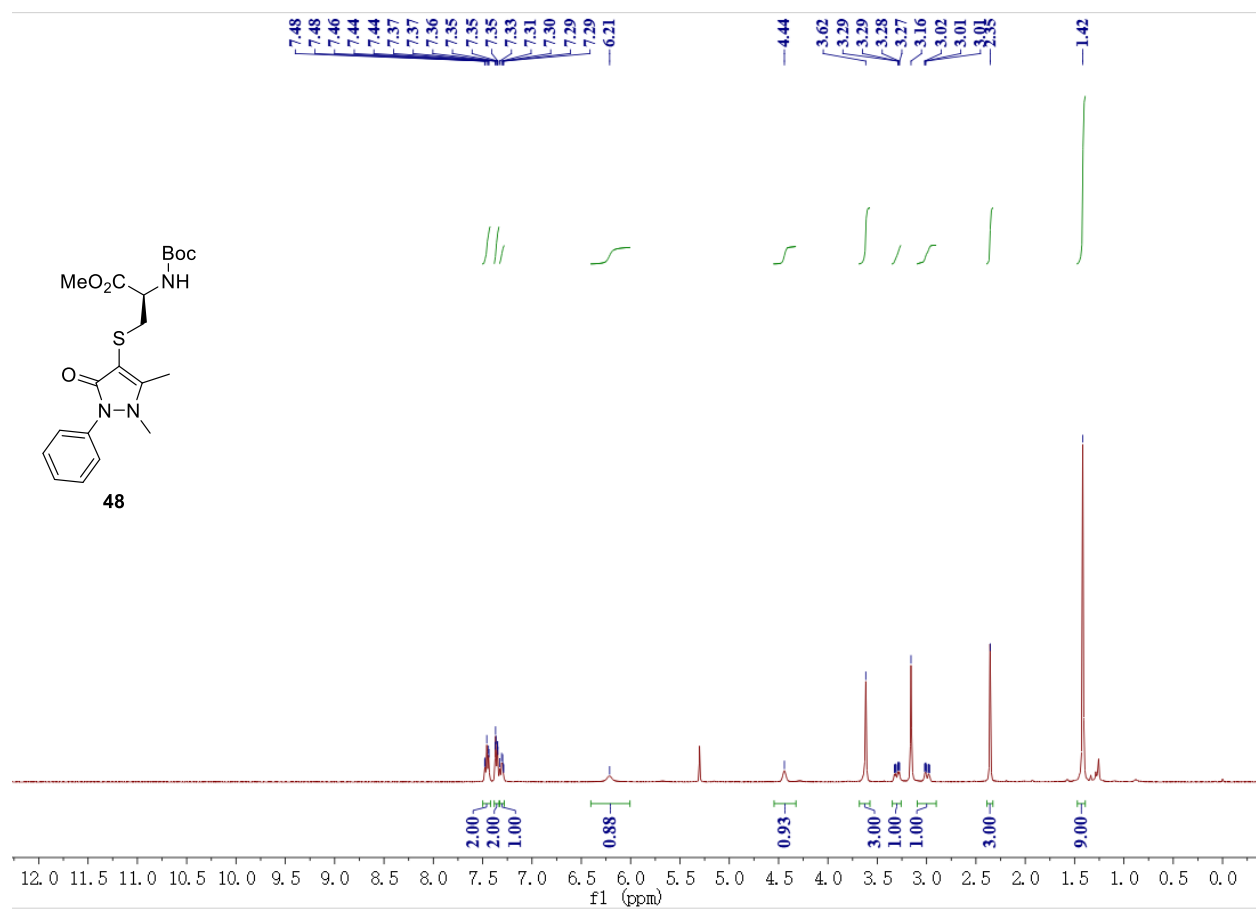


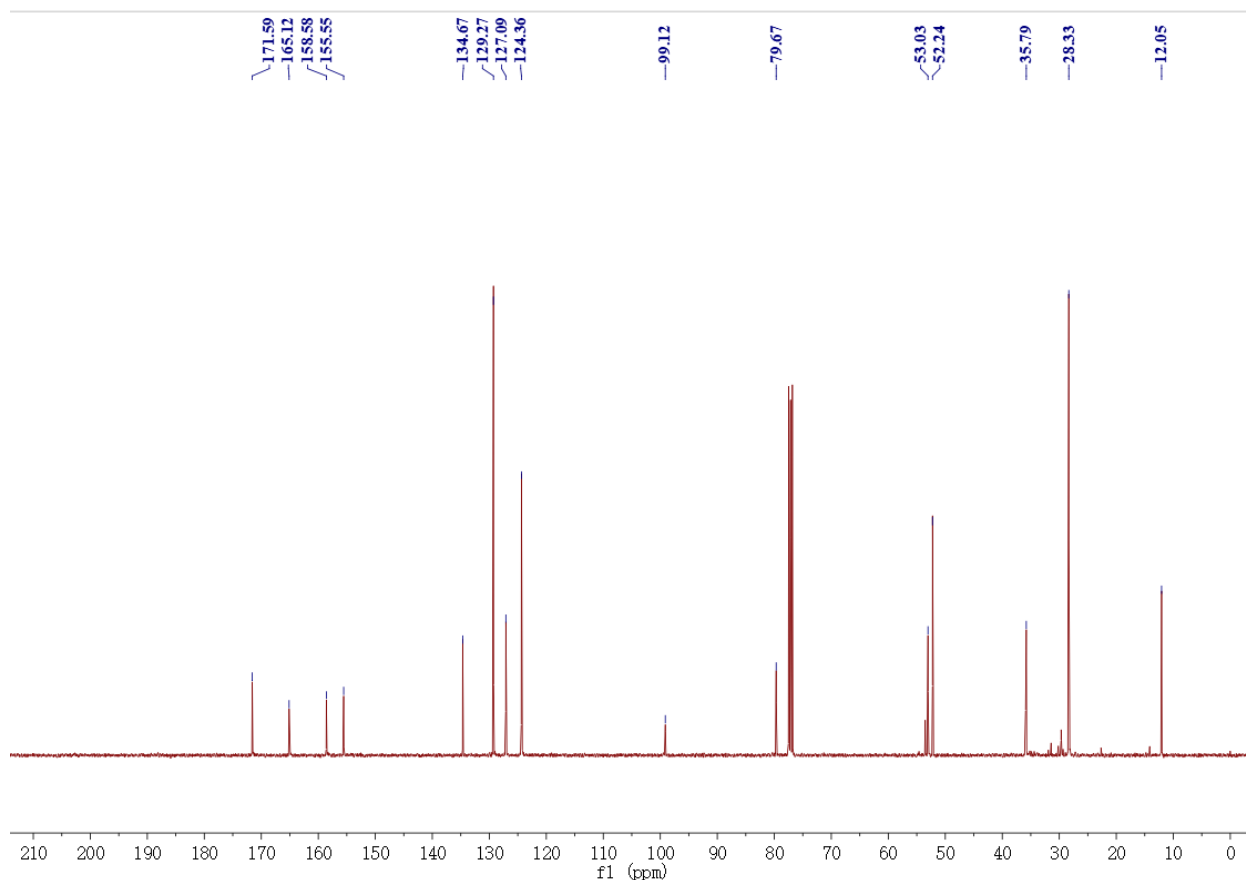




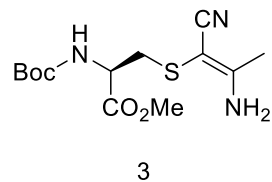
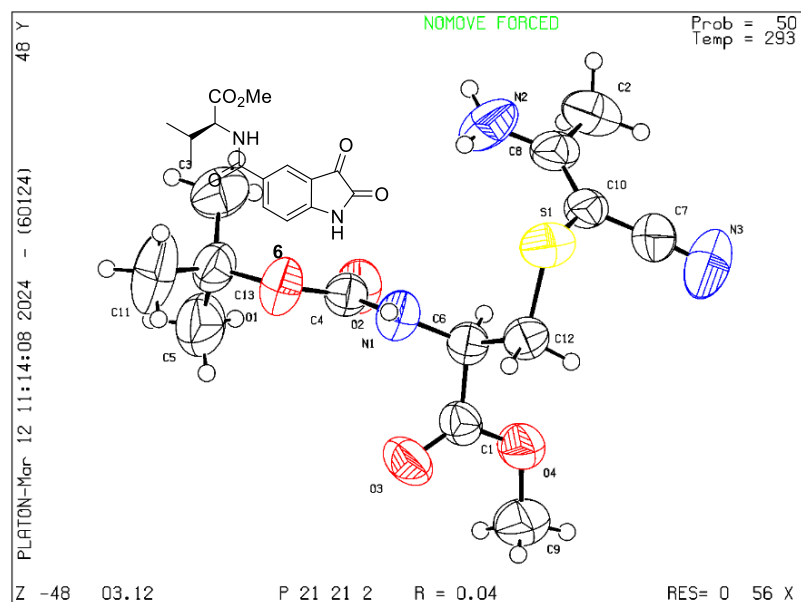








7. X-ray single-crystal data



Datablock:

Bond precision: C-C = 0.0050 Å Wavelength=1.54184
 Cell: a=7.2970(1) b=22.7082(4) c=10.4714(2)
 alpha=90 beta=90 gamma=90
 Temperature: 293 K

	Calculated	Reported
Volume	1735.13(5)	1735.13(5)
Space group	P 21 21 2	P 21 21 2
Hall group	P 2 2ab	P 2 2ab
Moiety formula	C13 H21 N3 O4 S	?
Sum formula	C13 H21 N3 O4 S	C13 H21 N3 O4 S
Mr	315.39	315.39
Dx, g cm ⁻³	1.207	1.207
Z	4	4
Mu (mm ⁻¹)	1.818	1.818
F000	672.0	672.0
F000'	675.32	
h, k, lmax	9, 28, 13	8, 28, 12
Nref	3519[2043]	3453
Tmin, Tmax	0.790, 0.819	
Tmin'	0.790	

Correction method= Not given
 Data completeness= 1.69/0.98 Theta(max)= 73.974
 R(reflections)= 0.0386(3187) wR2(reflections)= 0.1141(3453)
 S = 1.072 Npar= 195

The following ALERTS were generated. Each ALERT has the format
test-name_ALERT_alert-type_alert-level.
 Click on the hyperlinks for more details of the test.

Alert level A

[EXPT005_ALERT_1_A](#) _exptl_crystal_description is missing
 Crystal habit description.
 The following tests will not be performed.
 CRYSR_01
[DIFF003_ALERT_1_A](#) _diffrn_measurement_device_type is missing
 Diffractometer make and type. Replaces _diffrn_measurement_type.
[PLAT183_ALERT_1_A](#) Missing _cell_measurement_reflns_used Value Please Do !
[PLAT184_ALERT_1_A](#) Missing _cell_measurement_theta_min Value Please Do !
[PLAT185_ALERT_1_A](#) Missing _cell_measurement_theta_max Value Please Do !
[PLAT699_ALERT_1_A](#) Missing _exptl_crystal_description Value Please Do !

Alert level B

[PLAT035_ALERT_1_B](#) _chemical_absolute_configuration Info Not Given Please Do !

Alert level C

[PLAT052_ALERT_1_C](#) Info on Absorption Correction Method Not Given Please Do !
[PLAT242_ALERT_2_C](#) Low 'MainMol' Ueq as Compared to Neighbors of C7 Check
And 2 other PLAT242 Alerts
 More ...
[PLAT340_ALERT_3_C](#) Low Bond Precision on C-C Bonds 0.005 Ang.

Alert level G

[PLAT007_ALERT_5_G](#) Number of Unrefined Donor-H Atoms 3 Report
 H1 H2A H2B
[PLAT199_ALERT_1_G](#) Reported _cell_measurement_temperature (K) 293 Check
[PLAT200_ALERT_1_G](#) Reported _diffrn_ambient_temperature (K) 293 Check
[PLAT883_ALERT_1_G](#) No Info/Value for _atom_sites_solution_primary . Please Do !
[PLAT899_ALERT_4_G](#) SHELXL2018 is Deprecated and Succeeded by SHELXL 2019/3 Note
[PLAT912_ALERT_4_G](#) Missing # of FCF Reflections Above Sth/L= 0.600 12 Note
[PLAT969_ALERT_5_G](#) The 'Henn et al.' R-Factor-gap value 3.13 Note
 Predicted wR2: Based on SigI**2 3.64 or SHELX Weight 10.96
[PLAT978_ALERT_2_G](#) Number C-C Bonds with Positive Residual Density. 0 Info

6 **ALERT level A** = Most likely a serious problem - resolve or explain
 1 **ALERT level B** = A potentially serious problem, consider carefully
 5 **ALERT level C** = Check. Ensure it is not caused by an omission or oversight
 8 **ALERT level G** = General information/check it is not something unexpected

11 ALERT type 1 CIF construction/syntax error, inconsistent or missing data
 4 ALERT type 2 Indicator that the structure model may be wrong or deficient
 1 ALERT type 3 Indicator that the structure quality may be low
 2 ALERT type 4 Improvement, methodology, query or suggestion
 2 ALERT type 5 Informative message, check