

Supplementary Figures

Inhibition of the signal peptidase complex blocks cleavage of HTLV-1 ORF1 encoded p12 to p8 and impairs virus transmission

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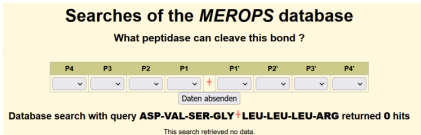
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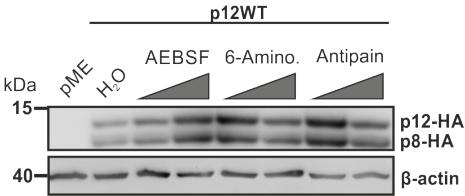
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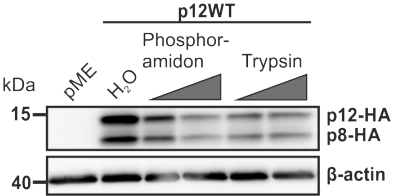
A



B



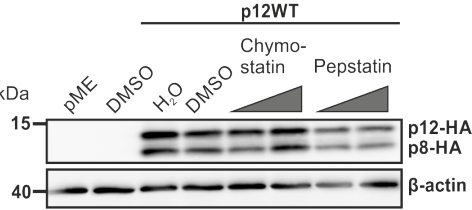
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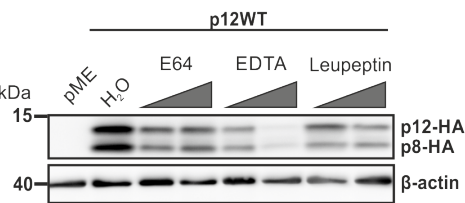
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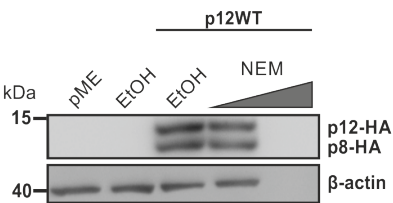
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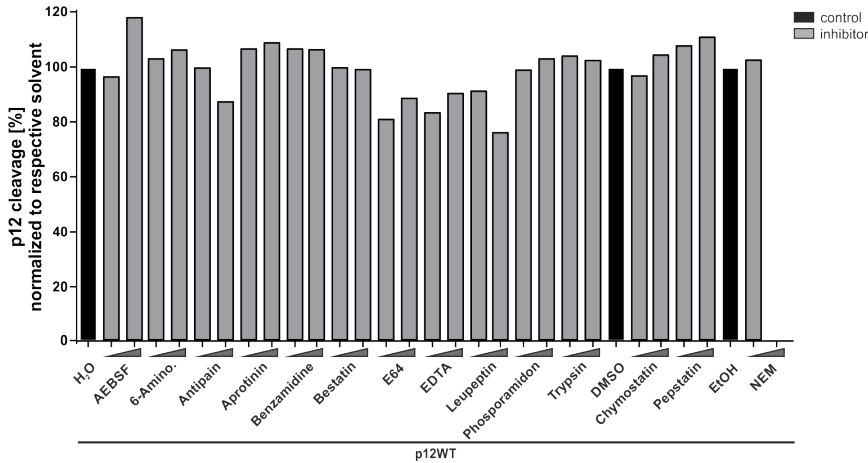
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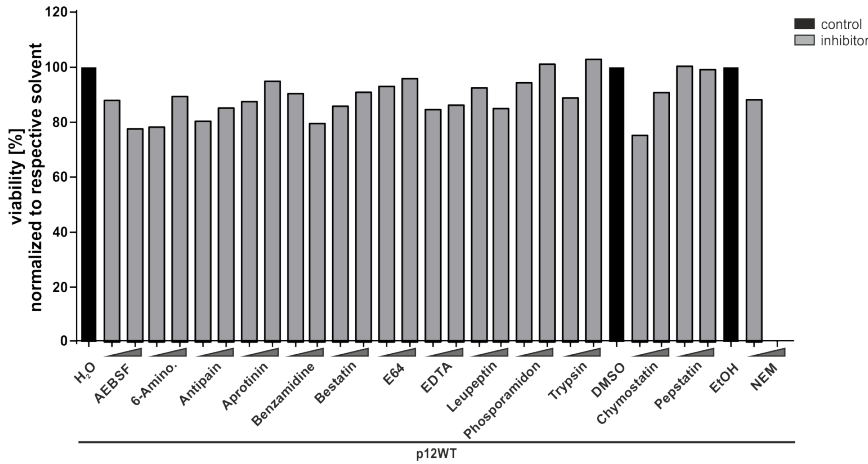
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H

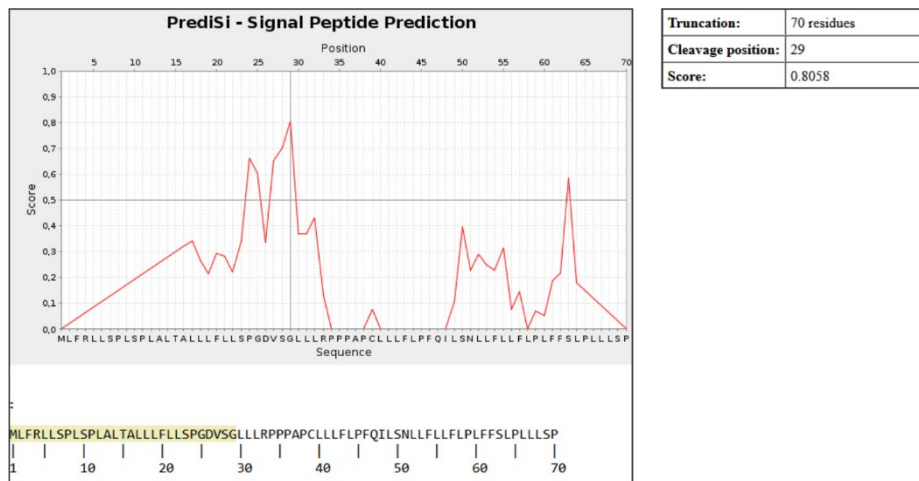


I



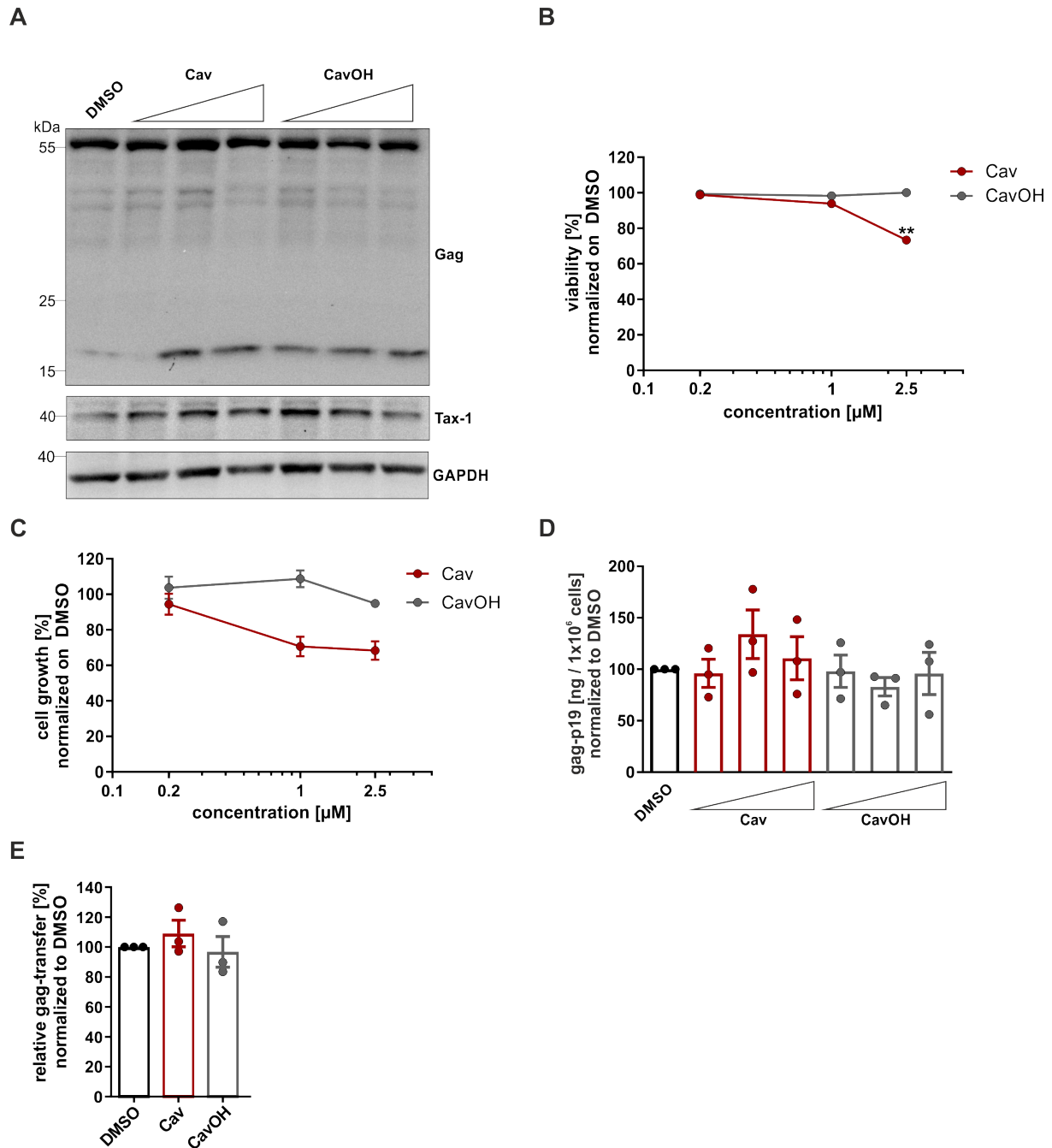
Supplementary Figure 1: Commercial protease inhibitors do not interfere with p12 cleavage in Jurkat.

(A) The p12 cleavage site was analysed with the MEROPS data base. **(B-I)** At 4 h prior electroporation, Jurkat T-cells were treated with two concentrations of one of the protease inhibitors (AEBSF: 0.01 mM, 0.1 mM; 6-Aminohexanoic acid: 0.025 mg/ml, 0.25 mg/ml; Antipain dihydrochloride: 1 μ M, 100 μ M; Aprotinin: 10 nM, 800 nM; Benzamidine: 0.5 mM, 4 mM; Bestatin: 4 μ M, 40 μ M; Chymostatin: 10 μ M, 100 μ M; E64: 1 μ M, 10 μ M; EDTA: 0.1 mM, 1 mM; NEM: 0.01 mM, 0.1 mM; Leupeptin: 10 μ M, 100 μ M; Pepstatin: 0.5 μ g/ml, 1 μ g/ml; Phosphoramidon: 1 μ M, 10 μ M; Trypsin: 10 μ g/ml, 100 μ g/ml). Next, cells were electroporated with 50 μ g HA-tagged pME-p12WT or pME and treated with the respective protease inhibitor for 24 h. Cells were lysed with the respective protease inhibitor added to corresponding lysis buffer and analysed via **(B-G)** immunoblotting with anti-HA and anti- β -actin antibodies. **(H)** Densitometric analysis of p12 and p8 specific bands was performed. p12 cleavage was calculated as $p8/(p12+p8)$ and normalized to respective solvent control. The mean of two independent experiments is depicted. **(I)** Cells were stained with propidium iodide (PI). Cell viability was analysed via flow cytometry and normalized to respective solvent control. The mean of two independent experiments is depicted.



Supplementary Figure 2: p12 is predicted to carry a signal peptide using PrediSi.

The p12 wildtype (WT) sequence was analysed with the signal peptide prediction tool PrediSi.



Supplementary Figure 3: Inhibition of p12 cleavage does not affect HTLV-1 cell-to-cell transmission from C91-PL to Jurkat.

(A-D) Chronically HTLV-1 infected C91-PL cells were treated with DMSO or increasing doses (0.2 μM, 1 μM or 2.5 μM) of either Cav or CavOH. After 48 h of treatment, cells were analysed via (A) immunoblotting with anti-Gag, anti-Tax-1 and anti-GAPDH antibodies. One representative of three independent experiments is shown. (B) Cell viability was analysed via flow cytometry. (C) Cell growth was analysed via automated

47 cell counting. **(D)** Viral release was analysed via Gag-p19 ELISA. The means of three
48 independent experiments \pm SEM are depicted. **(E)** C91-PL were treated with DMSO, 1
49 μ M Cav or CavOH for 24 h before co-culture with Jurkat-IRES-zsGreen cells. After 1
50 h, cells were fixed, stained intracellularly against Gag, and analysed via flow cytometry.
51 Quantitative analysis of gag-p19-APC-positive cells normalized to DMSO control was
52 conducted. The means of three independent experiments \pm SEM are depicted. Data
53 were analysed by Kruskal-Wallis test followed by Dunn's multiple comparisons
54 correction. * $p < 0.05$, ** $p < 0.01$.