

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a	Confirmed
<input type="checkbox"/>	<input checked="" type="checkbox"/> The exact sample size (<i>n</i>) for each experimental group/condition, given as a discrete number and unit of measurement
<input type="checkbox"/>	<input checked="" type="checkbox"/> A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
<input type="checkbox"/>	<input checked="" type="checkbox"/> The statistical test(s) used AND whether they are one- or two-sided <i>Only common tests should be described solely by name; describe more complex techniques in the Methods section.</i>
<input checked="" type="checkbox"/>	<input type="checkbox"/> A description of all covariates tested
<input checked="" type="checkbox"/>	<input type="checkbox"/> A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
<input type="checkbox"/>	<input checked="" type="checkbox"/> A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
<input type="checkbox"/>	<input checked="" type="checkbox"/> For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
<input checked="" type="checkbox"/>	<input type="checkbox"/> For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
<input checked="" type="checkbox"/>	<input type="checkbox"/> For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
<input checked="" type="checkbox"/>	<input type="checkbox"/> Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection	Long and short reads were generated on ONT, PacBio Sequel II and Illumina platform.
Data analysis	Nextdenovo v2.2.0, NextPolish v1.0, Minimap2 v2.17, BWA v0.7.17,LACHESIS, Platanus v1.2.4, Hifiasm v0.14, RepeatMasker v.4.0.7, RepeatModeler v.1.0.11, Augustus v3.2.3, TBLASTN v2.6.0, GENEWISE v2.4.1, PASA v2.3.3, EvidenceModeler v1.1.1, BUSCO v3.0, Maxent 3.4.3, ArcGIS 10.8, OrthoFinder v2.3.11, RAxML v8.2.11, CAFE v5.0, WGDI, BLASTP v2.7.1, ASTRAL v.5.6.2, PAML v4.9e, LTRharvest, LTRdigest, MUSCLE v3.8.31, HISAT2 v2.1.0, StringTie v1.3.3b, R v4.0.3, Bowtie2 v2.4.1, Bismark v0.22.3, MAFFT v7.313, AVID v2.1, VISTA (https://genome.lbl.gov/vista/mvista/submit.shtml)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

All raw sequence data, genome assembly and annotation information have been deposited in the National Genome Data Center (NGDC; <https://bigd.big.ac.cn/bioproject>) under BioProject accession number PRJCA022976.

Research involving human participants, their data, or biological material

Policy information about studies with [human participants or human data](#). See also policy information about [sex, gender \(identity/presentation\), and sexual orientation](#) and [race, ethnicity and racism](#).

Reporting on sex and gender [No human participants or human data used in this study](#)

Reporting on race, ethnicity, or other socially relevant groupings [No human participants or human data used in this study](#)

Population characteristics [No human participants or human data used in this study](#)

Recruitment [No human participants or human data used in this study](#)

Ethics oversight [No human participants or human data used in this study](#)

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

☐ Life sciences ☐ Behavioural & social sciences ☒ Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description	We sequenced and assembled the genomes of 13 species from different genera in the Salicaceae, and used multi-omics data along with molecular methods to study the genome evolutionary history of Salicaceae. This study revealed the types and characteristics of polyploidization events in the Salicaceae, and further explored how different genomic evolutionary patterns following allopolyploidy have driven trait innovation and subsequent adaptive radiation in species.
Research sample	Fresh leaf, flower and fruit samples used in this study were collected: Plant material of <i>C. decandra</i> , <i>D. caffra</i> , <i>S. chinensis</i> , <i>X. longifolia</i> and <i>F. jangomas</i> were collected in XiShuangBanNa Tropical Botanical Garden (Mengla, China), <i>I. orientalis</i> , <i>I. polycarpa</i> and <i>P. deltoides</i> were collected in Chengdu, <i>S. rehderiana</i> was collected in Minya Konka of China, <i>P. euphratica</i> and <i>P. alba</i> var. <i>pyramidalis</i> were collected in Lanzhou, Gansu, <i>S. chaenomeloides</i> was collected in Hanzhong, Shanxi, and <i>Dianyuea turbinata</i> was collected from Dehong, Yunnan.
Sampling strategy	Our sampling was based on the reported phylogenetic relationships within the Salicaceae family. Fresh leaf, flower, and fruit tissues were immediately placed on dry ice or in liquid nitrogen for subsequent sequencing.
Data collection	DNA was extracted from fresh leaf tissues and sequenced on the ONT, PacBio Sequel II and Illumina platforms, generating long and short reads for genome assembly. ATAC-seq libraries and whole genome bisulfite sequencing were performed on the Illumina platform to obtain epigenetic data. RNA was extracted from leaves, flowers, and fruits at different developmental stages, and transcriptome data were obtained through sequencing on the Illumina platform.
Timing and spatial scale	Mature leaf samples of different species were collected for genome sequencing from 2019 to 2023. Flower and fruit samples from <i>I. orientalis</i> , <i>I. polycarpa</i> and <i>P. deltoides</i> were collected at different developmental stages from March to November, and leaves of <i>I. orientalis</i> and <i>I. polycarpa</i> were collected from November to April.
Data exclusions	No data were excluded.

Reproducibility	Two to three replicates were collected for each tissue type of RNA-seq, ATAC-seq, and WGBS. The bootstrapping for phylogenetic analyses based on orthologs in all genomes and different subgenomes was replicated 100 times. The dual-luciferase assay was repeated three to six times to detect the enhancer activity of conserved noncoding elements (CNEs). All attempts at replication were successful.
Randomization	Since this is a genome sequencing study and the data were from a single individual, and leaf, flower and fruit tissues at different developmental stages were collected for transcriptome sequencing, no randomization was applied in this manuscript.
Blinding	Group allocation was not relevant to this study, so blinding was not required for our work.
Did the study involve field work?	<input type="checkbox"/> Yes <input checked="" type="checkbox"/> No

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> Antibodies
<input checked="" type="checkbox"/>	<input type="checkbox"/> Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology
<input checked="" type="checkbox"/>	<input type="checkbox"/> Animals and other organisms
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern
<input type="checkbox"/>	<input checked="" type="checkbox"/> Plants

Methods

n/a	Involved in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

Plants

Seed stocks	Plant material of <i>C. decandra</i> , <i>D. caffra</i> , <i>S. chinensis</i> , <i>X. longifolia</i> and <i>F. jangomas</i> were collected in XiShuangBanNa Tropical Botanical Garden (Mengla, China), <i>I. orientalis</i> , <i>I. polycarpa</i> and <i>P. deltoides</i> were collected in Chengdu, <i>S. rehderiana</i> was collected in Minya Konka of China, <i>P. euphratica</i> and <i>P. alba</i> var. <i>pyramidalis</i> were collected in Lanzhou, Gansu, <i>S. chaenomeloides</i> was collected in Hanzhong, Shanxi, and <i>Dianyuea turbinata</i> was collected from Dehong, Yunnan.. Fresh leaf, flower and fruit tissues were collected and placed on dry ice or liquid nitrogen for subsequent sequencing.
Novel plant genotypes	No novel plant genotypes were produced in this study.
Authentication	No seed stocks were used and no novel plant genotypes were generated in this study.