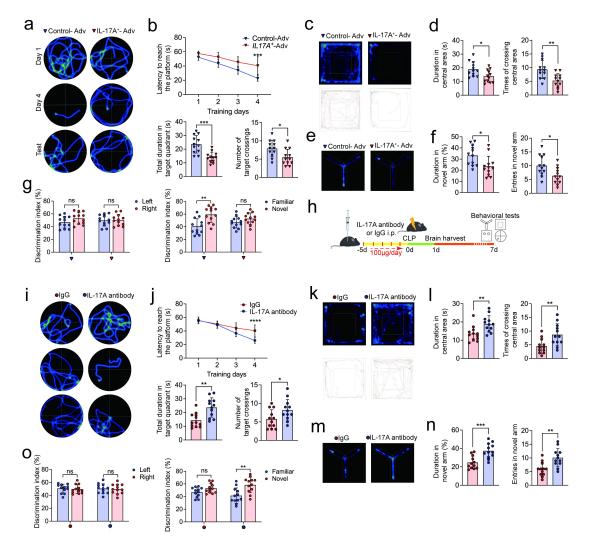


Extended Data Fig. 1.  $\gamma\delta$  T17 cells damage microglial mitochondria and induced SAE.

a Representative flow cytometry (FCM) plots showing CD45<sup>+</sup>  $\gamma\delta$  T cells (CD45<sup>+</sup> CD3<sup>+</sup>  $\gamma\delta$  TCR<sup>+</sup>) in the brain of sham-operated and CLP-treated mice. **b** FCM gating strategy of Kaede red  $\gamma\delta$  T17 subgroups. **c** Representative FCM image showing IL-17A<sup>+</sup>  $\gamma\delta$  T cells (CD45<sup>+</sup> CD3<sup>+</sup>  $\gamma\delta$  TCR<sup>+</sup> IL-17A<sup>+</sup>) in the meninges. Graph displaying the percentage of IL-17A<sup>+</sup> cells among  $\gamma\delta$  T cells in meninges (n = 6). **d** FCM analysis to assess the purity of primary microglial cells. **e** FCM analysis to assess the purity of primary  $\gamma\delta$  T cells. Representative images (**f**) and statistical analysis (**g**) of open field tests (n = 12–13). **h** Statistical analysis of novel object recognition tests (n = 12–13). **i** Treatment schedule. BV2 cells treated with ethidium bromide for 28 days to deplete

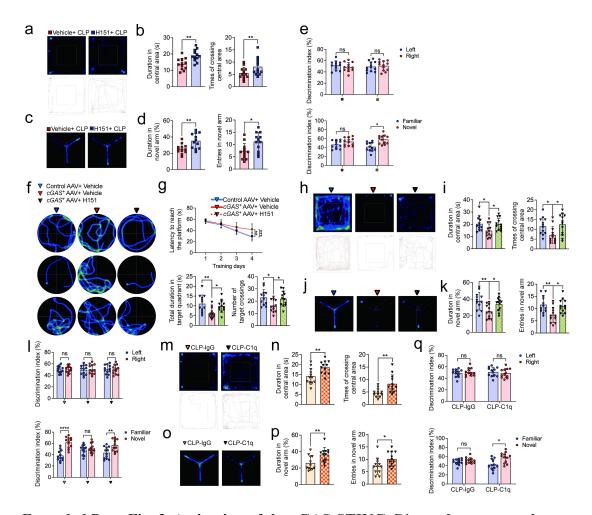
mitochondria, generating BV2 $^{p0}$  cells. Western blot (**j**) and quantification (**k**) of cGAS and STING in BV2 and BV2 $^{p0}$  cells treated with IL-17A and LPS (n = 6). Data are shown as mean  $\pm$  SD.  $^*P$  < 0.05,  $^{**}P$  < 0.01,  $^{***}P$  < 0.001. ns, not significant. P values were calculated using one-way ANOVA followed by Šídák's multiple comparisons test (c, g), two-way ANOVA followed by Šídák's multiple comparisons test (h) and two-sided Student's unpaired t-tests (k).



Extended Data Fig. 2. IL-17A exacerbates SAE.

Representative images (a) and statistical analysis (b) of Morris water maze (n = 12). Representative images (c) and statistical analysis (d) of open field tests (n = 12). Representative images (e) and statistical analysis (f) of Y-maze tests (n = 12). g Statistical analysis of novel object recognition tests (n = 12). h Treatment schedule. IL-17A neutralizing antibody or IgG (100  $\mu$ g/day) was administered via intraperitoneal injection for 5 days, with four doses given prior to the cecum ligation and puncture (CLP) challenge and one dose immediately after. Brain tissues were collected 1 day post-CLP, and behavioral testing was conducted 7 days post-CLP. Representative images (i) and statistical analysis (j) of the Morris water maze (n = 12).

Representative images (**k**) and statistical analysis (**l**) of open field tests (n = 12). Representative images (**m**) and statistical analysis (**n**) of Y-maze tests (n = 12). **o**Statistical analysis of novel object recognition tests (n = 12). Data are shown as mean  $\pm$  SD. \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001. ns, not significant. P values were calculated using two-sided Student's unpaired t-tests (b, d, f, j, l, n) and multiple unpaired t-tests (g, o).



Extended Data Fig. 3. Activation of the cGAS-STING-C1q pathway exacerbates cognitive dysfunction.

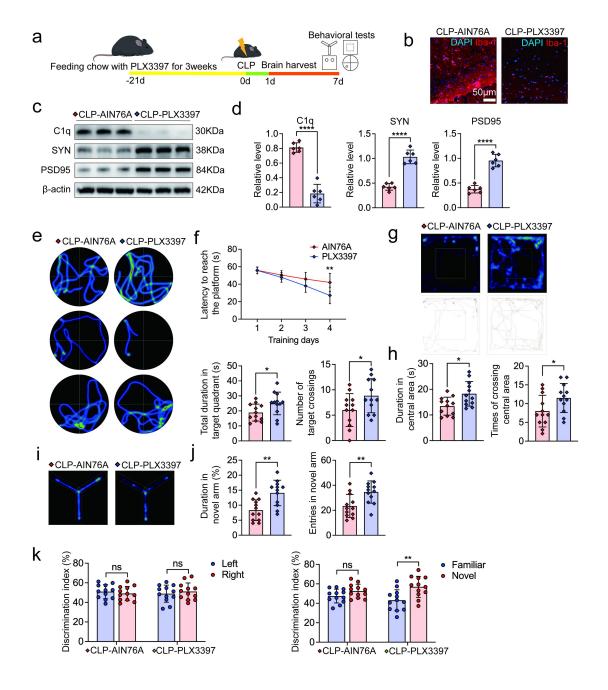
Representative images (a) and statistical analysis (b) of open field tests (n = 12).

Representative images (c) and statistical analysis (d) of Y-maze tests (n = 12). e

Statistical analysis of novel object recognition tests (n = 12). Representative images (f) and statistical analysis (g) of the Morris water maze (n = 12). Representative images

(h) and statistical analysis (i) of open field tests (n = 12). Representative images (j) and statistical analysis (k) of Y-maze tests (n = 12). I Statistical analysis of novel object recognition tests (n = 12). Representative images (m) and statistical analysis (n) of open field tests (n = 12). Representative images (o) and statistical analysis (p) of Y-maze tests (n = 12). q Statistical analysis of novel object recognition tests (n = 12).

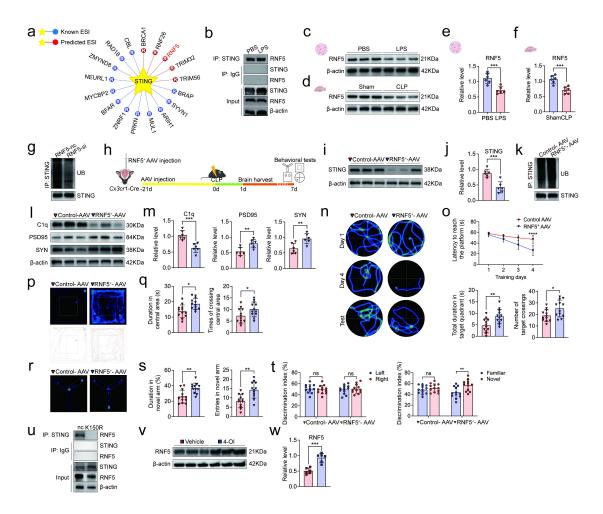
Data are shown as mean  $\pm$  SD. \*P < 0.05, \*\*P < 0.01, \*\*\*\*P < 0.0001. ns, not significant. P values were calculated using two-sided Student's unpaired t-tests (b, d, n, p), multiple unpaired t-tests (e, q), one-way ANOVA followed by Šídák's multiple comparisons test (g, i, k) and two-way ANOVA followed by Šídák's multiple comparisons test (l).



Extended Data Fig. 4. Microglia depletion ameliorates SAE.

a Treatment schedule. Mice were fed PLX3397 or AIN76A chow for 3 weeks prior to cecum ligation and puncture (CLP). Brain tissues were collected 1 day post-CLP, and behavioral testing was conducted 7 days post-CLP. **b** Representative immunofluorescence images showing microglia depletion in mice treated with PLX3397. Scale bar, 50 μm. Western blot (**c**) and quantification (**d**) of C1q, PSD95

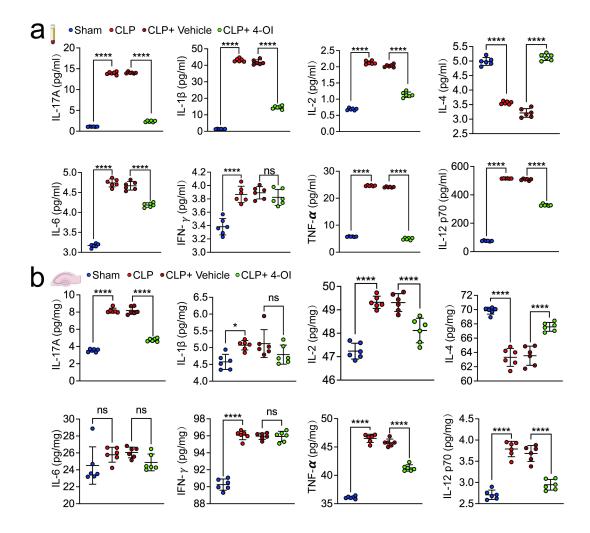
and SYN in hippocampal synaptic proteins of mice fed with PLX3397 and AIN76A (n = 6). Representative images (e) and statistical analysis (f) of the Morris water maze (n = 12). Representative images (g) and statistical analysis (h) of open field tests (n = 12). Representative images (i) and statistical analysis (j) of Y-maze tests (n = 12). k Statistical analysis of novel object recognition tests (n = 12). Data are shown as mean  $\pm$  SD. \*P < 0.05, \*\*P < 0.01, \*\*\*\*\*P < 0.0001. ns, not significant. P values were calculated using two-sided Student's unpaired t-tests (d, f, h, j) and multiple unpaired t-tests (k).



Extended Data Fig. 5. RNF5 enhances STING ubiquitination to mitigate SAE.

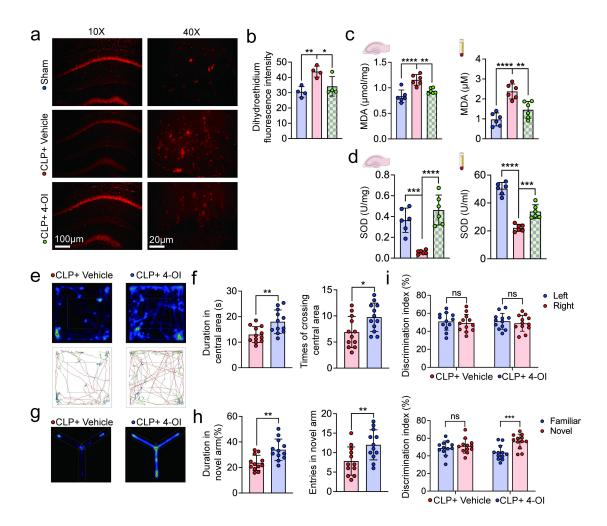
a Predicted E3 ubiquitin ligases for STING from the Ubibrowse database. **b**Co-immunoprecipitation and western blot analysis of the interaction between STING and RNF5 in BV2 cells. Western blot and quantification of RNF in primary microglia (**c**, **e**) and hippocampus (**d**, **f**) of mice (n = 6). **g** Western blot showing STING ubiquitination levels in primary microglia. **h** Treatment schedule. Hippocampal stereotactic injection of RNF5<sup>+</sup> adeno-associated virus (AAV); cecum ligation and puncture (CLP) performed 21 days later. Brain tissues collected 1 day post-CLP, behavioral testing conducted 7 days post-CLP. Western blot (**i**) and quantification (**j**) of STING in the hippocampus of mice treated with control and RNF5<sup>+</sup> AAV (n = 6). **k** Western blot image of STING ubiquitination levels in the hippocampus of mice

treated with control and RNF5<sup>+</sup> AAV. Western blot (I) and quantification (**m**) of C1q, PSD95 and SYN in hippocampal synaptic proteins of mice treated with control and RNF5<sup>+</sup> AAV (n = 6). Representative images (**n**) and statistical analysis (**o**) of the Morris water maze (n = 12). Representative images (**p**) and statistical analysis (**q**) of open field tests (n = 12). Representative images (**r**) and statistical analysis (**s**) of Y-maze tests (n = 12). **t** Statistical analysis of novel object recognition tests (n = 12). **u** Co-immunoprecipitation and western blot analysis of the interaction between STING and RNF5 in BV2 cells. Western blot (**v**) and quantification (**w**) of RNF in hippocampus of septic mice received 4-OI or vehicle (n = 6). Data are shown as mean  $\pm$  SD. \*P < 0.05, \*\*P < 0.01, \*\*\*\*P < 0.001, \*\*\*\*P < 0.0001. ns, not significant. P values were calculated using two-sided Student's unpaired t-tests (**e**, f, j, m, q, o, s, w) and multiple unpaired t-tests (t).



Extended Data Fig. 6. Exogenous 4-OI reduced inflammatory cytokine levels.

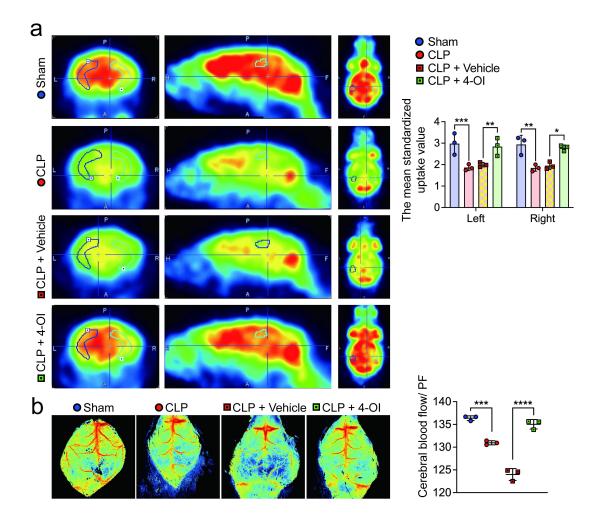
a Plasma inflammatory cytokine concentrations in sham-operated and cecum ligation and puncture (CLP) - treated mice administered 4-Octyl itaconate (4-OI) or vehicle (n = 6). **b** Hippocampal inflammatory cytokine concentrations in sham-operated and CLP-treated mice administered 4-OI or vehicle (n = 6). Data are shown as mean  $\pm$  SD. \*\*\*\*\*P < 0.0001. ns, not significant. P values were calculated using one-way ANOVA followed by Šídák's multiple comparisons test (a, b).



Extended Data Fig. 7. Exogenous 4-OI reduced oxidative stress levels and mitigated SAE.

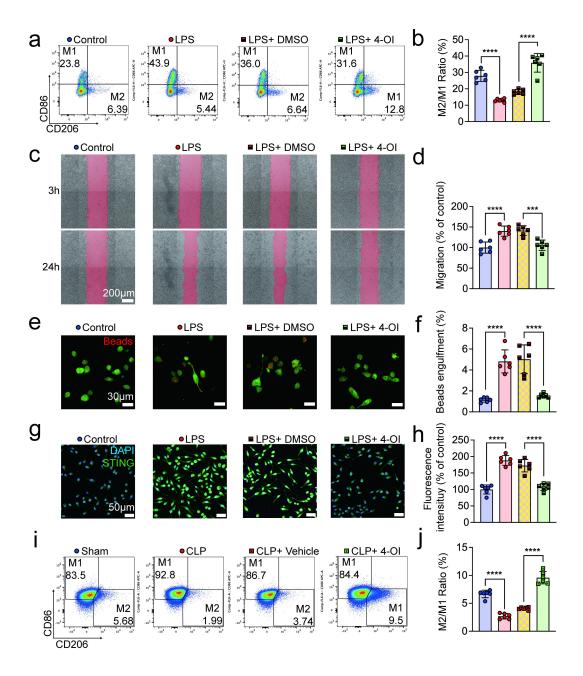
Representative immunofluorescence images (a) and analysis (b) of dihydroethidium (DHE) staining in hippocampus (n = 6). c Levels of malonaldehyde (MDA) in the hippocampus and plasma of mice (n = 6). d Levels of Superoxide dismutase (SOD) in the hippocampus and plasma of mice (n = 6). Representative images (e) and statistical analysis (f) of open field tests (n = 12). Representative images (g) and statistical analysis (h) of Y-maze tests (n = 12). i Statistical analysis of novel object recognition tests (n = 12). Data are shown as mean  $\pm$  SD.  $^*P < 0.05$ ,  $^{**}P < 0.01$ ,  $^{***}P < 0.001$ ,  $^{***}P < 0.001$ . ns, not significant. P values were calculated using one-way ANOVA

followed by Šídák's multiple comparisons test (b-d), two-sided Student's unpaired t-tests (f, h) and multiple unpaired t-tests (i).



Extended Data Fig. 8. Exogenous 4-Octyl itaconate enhances cerebral metabolism and improves cortical blood flow.

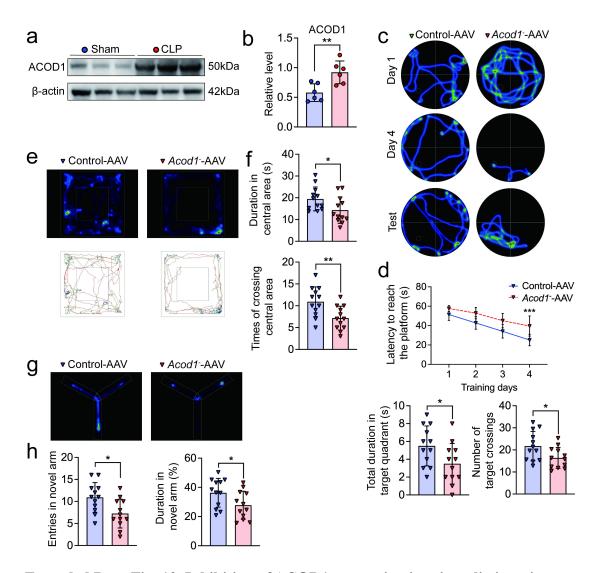
a Representative Positron Emission Tomography (PET) images and analysis of brain glucose metabolism in mice (n = 3). **b** Representative laser speckle imaging and analysis of cortical blood flow in mice (n = 3). Data are shown as mean  $\pm$  SD. \* $^*P$  < 0.05, \* $^*P$  < 0.01, \* $^*P$  < 0.001, \* $^*P$  < 0.0001. P values were calculated using two-way ANOVA followed by Šídák's multiple comparisons test (a) and one-way ANOVA followed by Šídák's multiple comparisons test (b).



Extended Data Fig. 9. Exogenous 4-OI reduces microglial inflammation and phagocytic capacity.

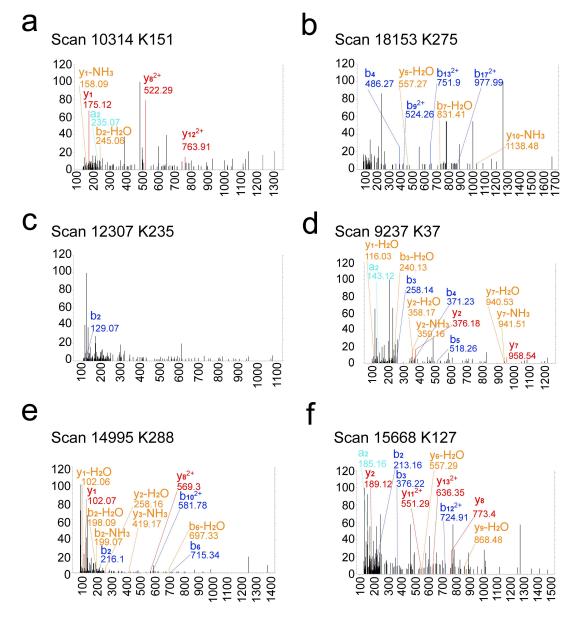
**a, b** Representative flow cytometry (FCM) plots showing M1 (CD86<sup>+</sup>) and M2 (CD206<sup>+</sup>) phenotypes in BV2 cells following lipopolysaccharide (LPS) and 4-Octyl itaconate (4-OI) treatment. Graph displaying the M2/M1 ratio (n = 6). **c, d**Representative images of scratch assay in BV2 cells treated with LPS and 4-OI.
Graph displaying the migration rate (% of control) (n = 6). **e, f** Representative images

of phagocytosis assay in BV2 cells treated with LPS and 4-OI. Graph displaying the beads engulfment (n = 6). **g, h** Representative immunofluorescence images of STING in BV2 cells treated with LPS and 4-OI. Graph displaying the fluorescence intensity (n = 6). **i, j** Representative FCM plots depicting M1 (CD86<sup>+</sup>) and M2 (CD206<sup>+</sup>) microglia following CLP and 4-OI treatment. Graph displaying the ratio of M2/M1 (n = 6). Data are shown as mean  $\pm$  SD. \*\*\*P < 0.001, \*\*\*\*P < 0.0001. P values were calculated using one-way ANOVA followed by Šídák's multiple comparisons test (b, d, f, h, j).



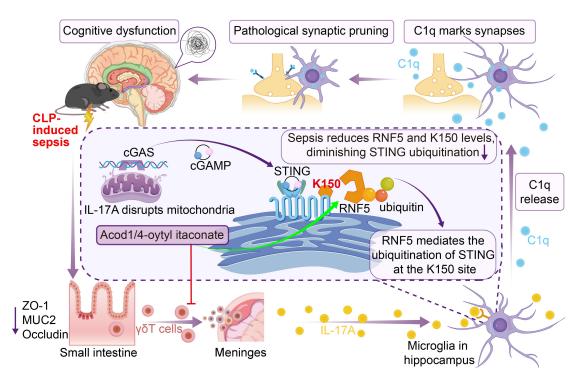
Extended Data Fig. 10. Inhibiting of ACOD1 expression in microglia impaires cognitive function.

Western blot (**a**) and quantification (**b**) of ACOD1 in hippocampus of mice (n = 6). Representative images (**c**) and statistical analysis (**d**) of the Morris water maze (n = 12). Representative images (**e**) and statistical analysis (**f**) of open field tests (n = 12). Representative images (**g**) and statistical analysis (**h**) of Y-maze tests (n = 12). Data are shown as mean  $\pm$  SD. \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001. P values were calculated using two-sided Student's unpaired t-tests (b, d, f, h).



Extended Data Fig. 11. Mass spectrometry identification of STING ubiquitination sites.

Potential ubiquitination sites at lysine 151 (K151) (a), lysine 275 (K275) (b), lysine 235 (K235) (c), lysine 37 (K37) (d), lysine 288 (K288) (e), and lysine 127 (K127) (f) on STING.



Extended Data Fig. 12. Schematic of mechanism.

During sepsis, IL-7R<sup>high</sup> CD8<sup>low</sup>  $\gamma\delta$  T17 cells migrate from the small intestine to the meninges, where they secrete IL-17A. This secretion activates microglia in the hippocampus, leading to impaired oxidative metabolism and mitochondrial damage. The resulting release of mitochondrial deoxyribonucleic acid (mtDNA) triggers the activation of the cGAS-STING signaling pathway. This activation increases the release of complement component C1q by microglia, enhancing synaptic tagging and causing excessive synaptic pruning. Treatment with 4-Octyl Itaconate (4-OI) has been shown to mitigate sepsis-associated encephalopathy (SAE) by inhibiting the migration of IL-7R<sup>high</sup> CD8<sup>low</sup>  $\gamma\delta$  T17 cells to the meninges following sepsis. Additionally, 4-OI promotes the expression of the ubiquitination site at K150 on STING, facilitating K150-dependent STING ubiquitination, thereby alleviating the effects of SAE.