

Defying the oxidative-addition prerequisite in cross-coupling through artful single-atom catalysts

Jiwei Shi^{1,2,3,12}, Gang Wang^{4,12}, Duanshuai Tian^{1,12}, Xiao Hai^{5}, Rongwei Meng^{1,2,3,12}, Yifan Xu⁶, Yu Teng^{1,2}, Lu Ma⁷, Shibo Xi⁸, Youqing Yang¹, Xin Zhou¹, Xingjie Fu^{1,2,3}, Hengyu Li¹, Qilong Cai¹, Peng He¹, Huihui Lin^{1,8}, Jinxing Chen¹, Jiali Li¹, Jinghan Li¹, Qian He⁹, Qian-Hong Yang^{2,3}, Jun Li^{4,10}, Dongshuang Wu^{6*}, Yang-Gang Wang^{4*}, Jie Wu^{1*} and Jiong Lu^{1,11*}*

¹Department of Chemistry, National University of Singapore, 3 Science Drive 3, Singapore 117543, Singapore.

²Joint School of National University of Singapore and Tianjin University, International Campus of Tianjin University, Binhai New City, Fuzhou 350207, China.

³Nanoyang Group, Tianjin Key Laboratory of Advanced Carbon and Electrochemical Energy Storage, School of Chemical Engineering and Technology, National Industry-Education Integration Platform of Energy Storage, and Collaborative Innovation Center of Chemical Science and Engineering (Tianjin), Tianjin University, Tianjin, 300072, China.

⁴Department of Chemistry and Guangdong Provincial Key Laboratory of Catalysis, Southern University of Science and Technology, Shenzhen 518055, Guangdong, China.

⁵School of Materials Science and Engineering, Peking University, Beijing, P.R. China.

⁶School of Materials and Engineering, Nanyang Technological University, Singapore, 639798 Singapore.

⁷National Synchrotron Light Source II Brookhaven National Lab Upton, NY 11973, USA.

⁸Institute of Sustainability for Chemicals, Energy and Environment (ISCE2), Agency for Science, Technology and Research (A*STAR), 1 Pesek Road Jurong Island, Singapore 627833, Republic of Singapore.

⁹Department of Material Science and Engineering, College of Design and Engineering,
National University of Singapore, 9 Engineering Drive 1, EA #03-09, 117575, Singapore.

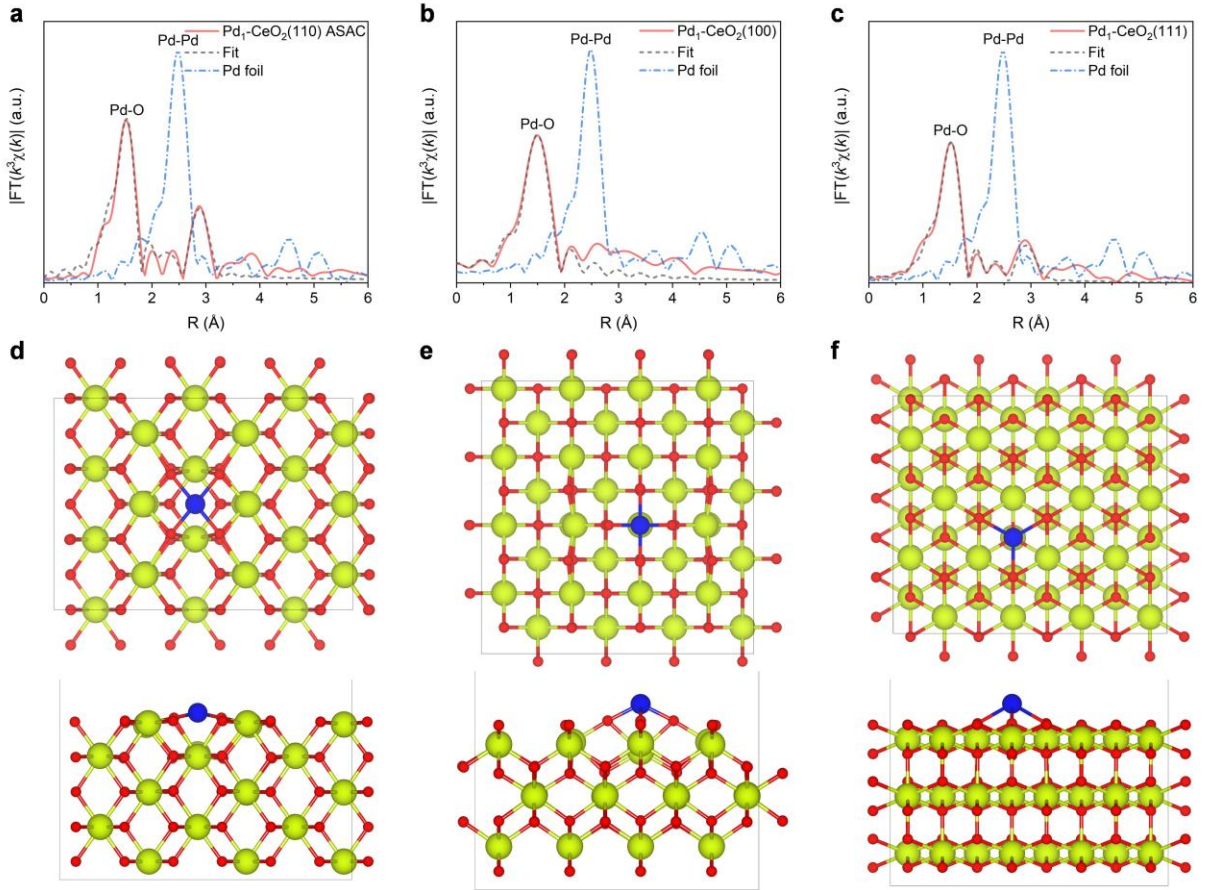
¹⁰Department of Chemistry and Engineering Research Center of Advanced Rare-Earth
Materials of Ministry of Education, Tsinghua University, Beijing, China.

¹¹National University of Singapore (Suzhou) Research Institute, No. 377 Linqun Street,
215123 Suzhou, Jiangsu, China.

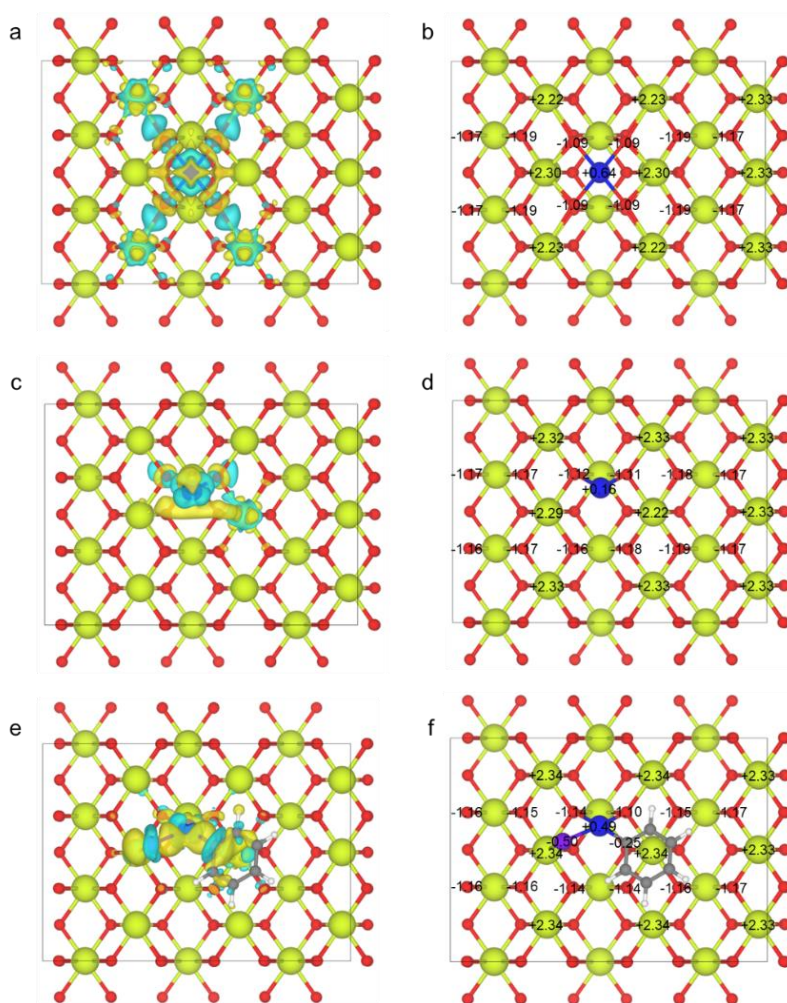
¹²These authors contributed equally: Jiwei Shi, Gang Wang, Duanshuai Tian, Rongwei Meng

*Corresponding author. Email: xiaohai@pku.edu.cn; dongshuang.wu@ntu.edu.sg;
wangyg@sustech.edu.cn; chmjie@nus.edu.sg; chmluj@nus.edu.sg

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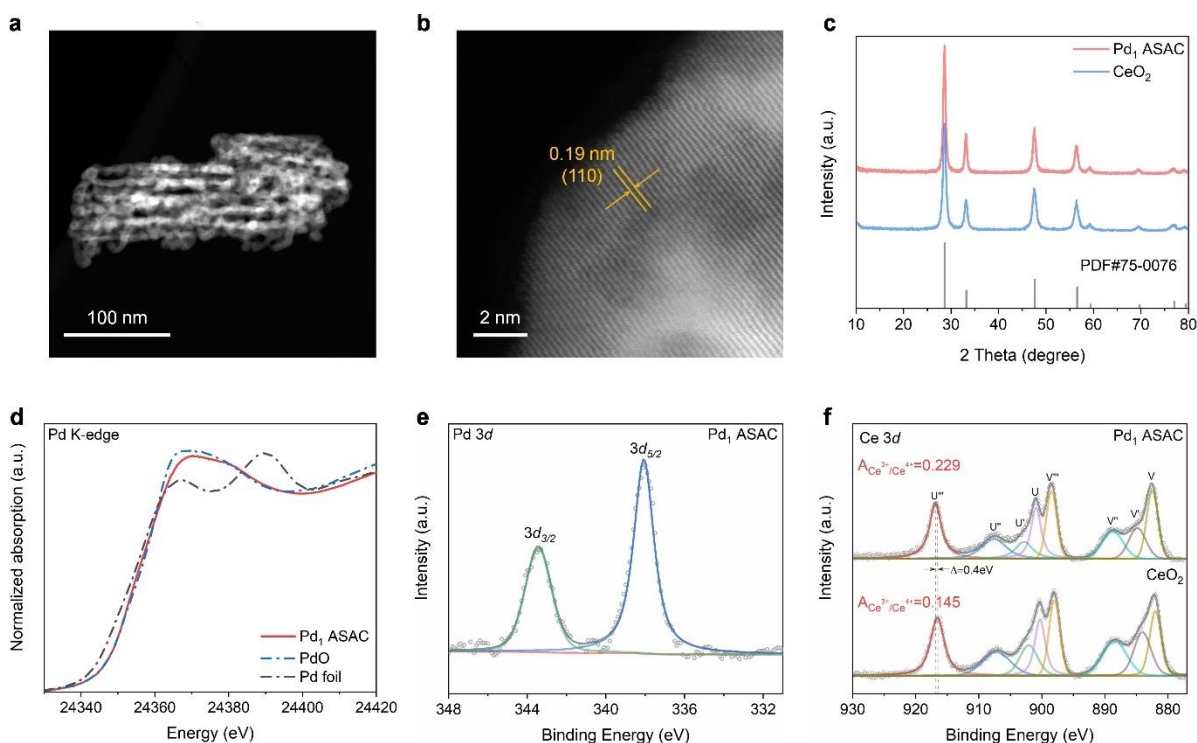


Supplementary Fig. 1 | The experimental and modelled structures of Pd single atoms on facet-dependent CeO₂. Fourier-transformed EXAFS spectra of Pd₁-CeO₂(110) ASAC (a), Pd₁-CeO₂(100) (b), and Pd₁-CeO₂(111) (c). Top views and side views of Pd SACs on CeO₂ with different crystal planes: Pd₁-CeO₂(110) ASAC (d), Pd₁-CeO₂(100) (e), Pd₁-CeO₂(111) (f). The red, yellow, and blue spheres represent oxygen, cerium, and palladium, respectively.

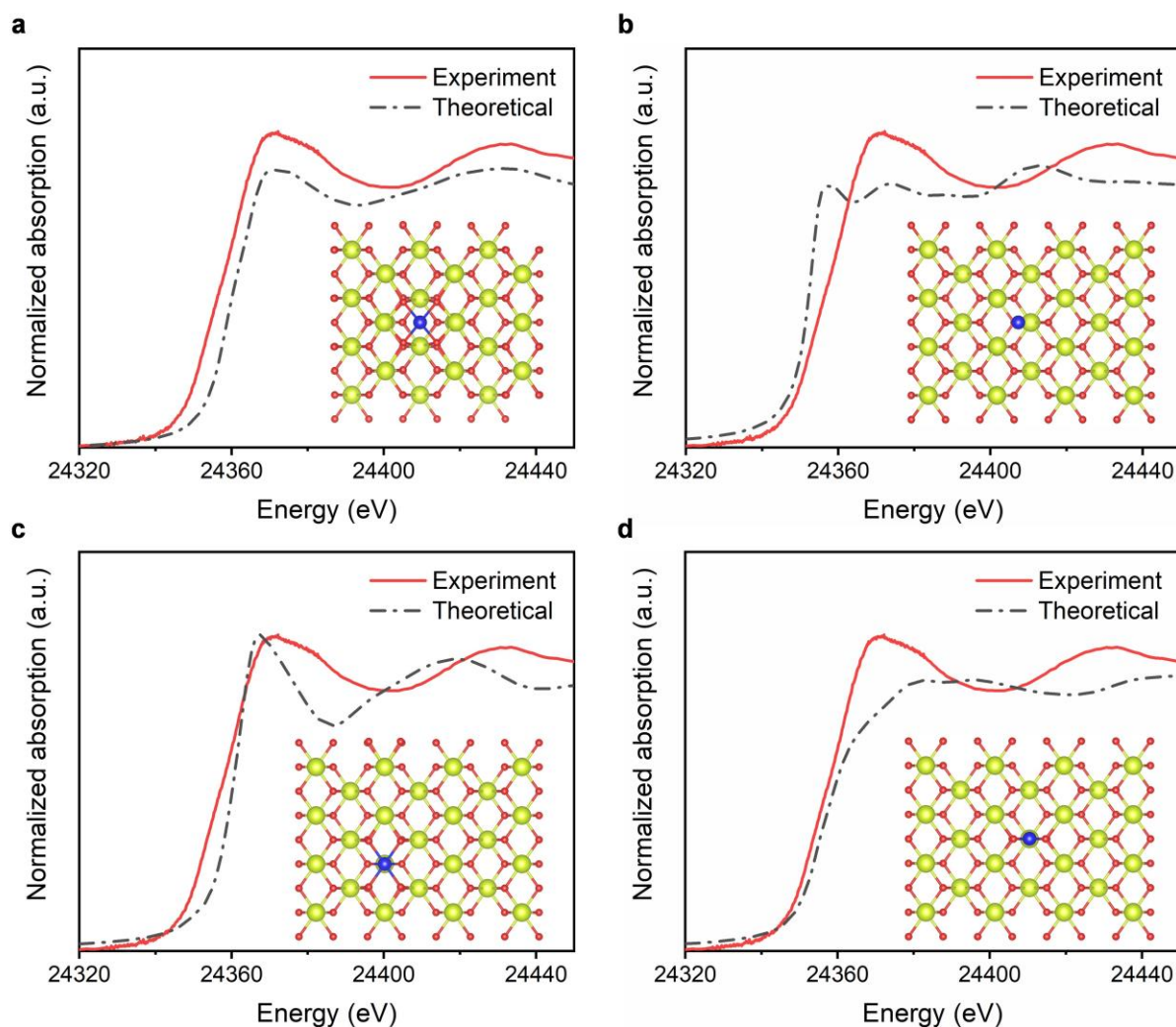


Supplementary Fig. 2 | Charge density differences of Pd₁ ASACs catalyst. The white, grey, purple, red, yellow, and blue spheres represent hydrogen, carbon, bromine, oxygen, cerium, and palladium atoms, respectively.

The Pd in the initially four-coordinated Pd₁ ASACs catalyst is in a high-valence oxidized state, with electrons predominantly transferred to the Ce and O atoms in the Pd-O-Ce linkage (**Supplementary Fig. 2a, b**). During the reaction, the Pd-O bond is cleaved, forming a bicoordinated structure, and the Pd valence state is reduced from +0.64 to +0.16 as electrons transfer back from the Ce and O atoms to the Pd atom (**Supplementary Fig. 2c, d**). Following oxidative addition, the dissociation of bromobenzene results in the bromide ion and phenyl ring gaining 0.5 and 0.25 electrons, respectively, leading to Pd oxidation from +0.16 to +0.49. Concurrently, additional electrons are transferred from the Ce-O-Pd linkage to the Pd atom (**Supplementary Fig. 2e, f**). Thus, the continuous electron transfer between the Pd atom and the O and Ce atoms in the Pd-O-Ce bond ensures the stabilization of the Pd valence state throughout the reaction.

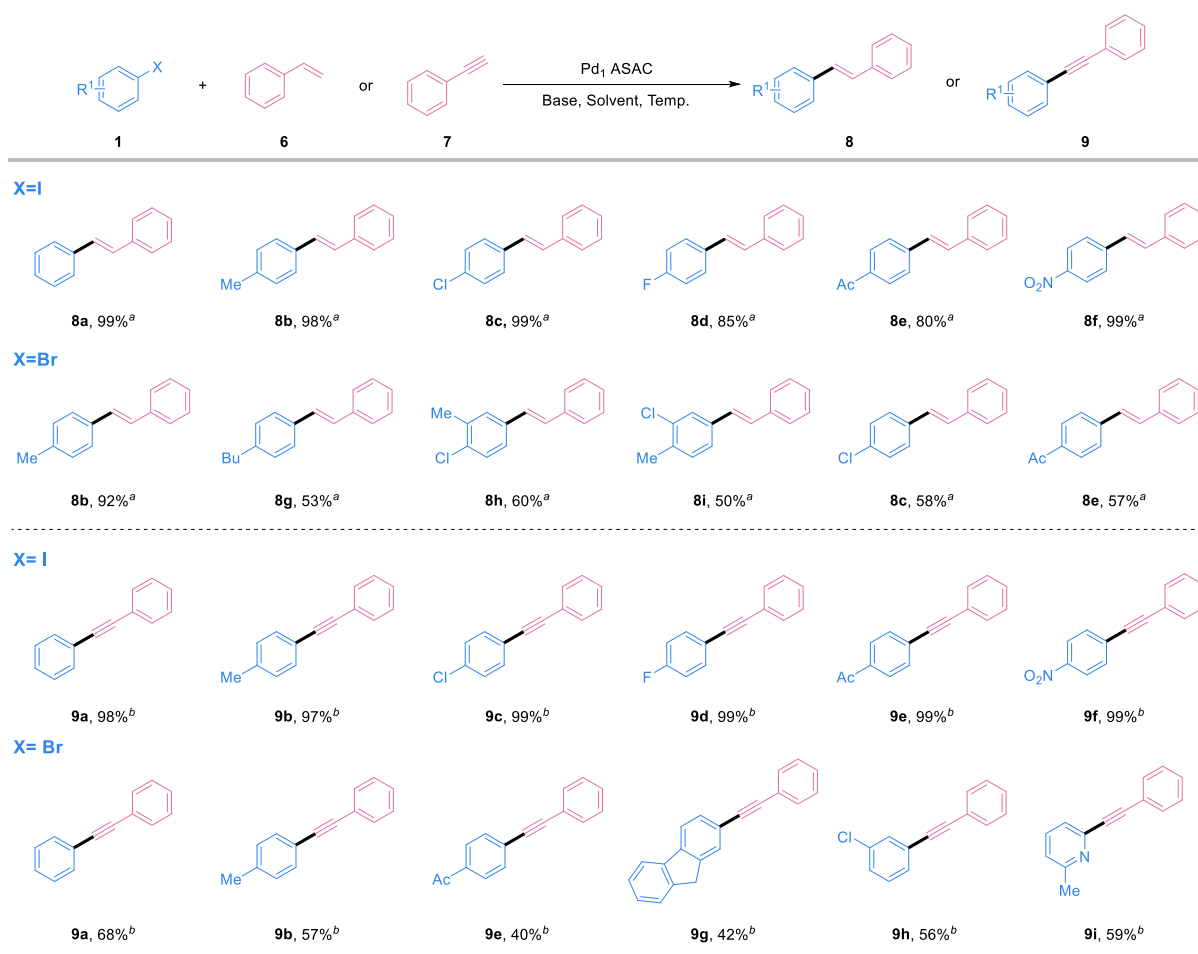


Supplementary Fig. 3 | Characterization of Pd₁ ASAC. **a**, TEM image of Pd₁ ASAC. **b**, Atomic-resolution ADF-STEM image of Pd₁ ASAC. **c**, XRD patterns of Pd₁ ASAC. **d**, Pd K-edge XANES spectra of Pd₁ ASAC and references (Pd foil and PdO). **e**, Pd 3d XPS spectra of Pd₁ ASAC. **f**, Ce 3d XPS spectra of Pd₁ ASAC and CeO₂. The peaks V' and U' correspond to the signals of Ce³⁺ species, and the others are assigned to the Ce⁴⁺ species.



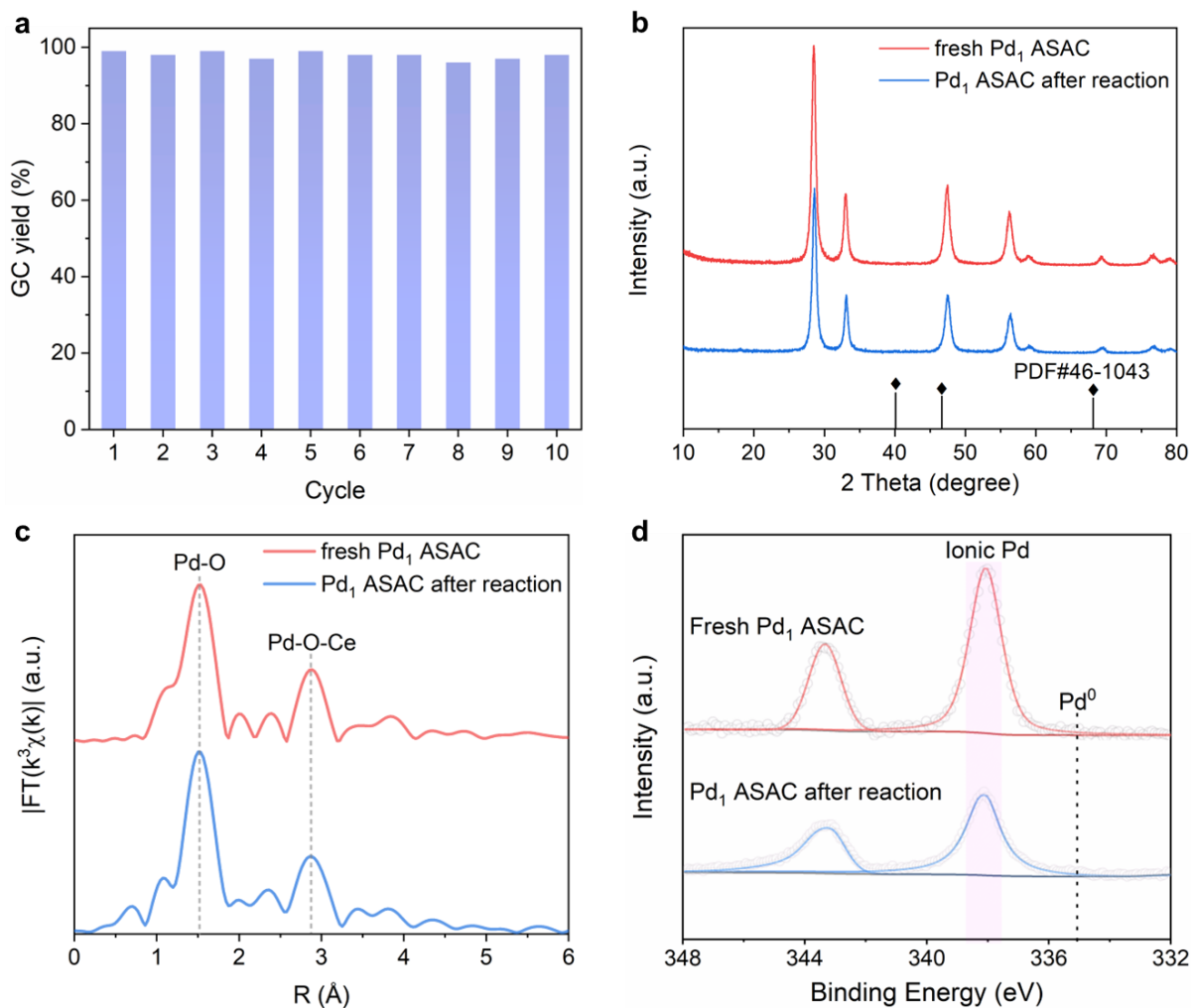
Supplementary Fig. 4 | Comparison of the experimental and modelled Pd K-edge XANES spectra.

a, Pd single-atom is anchored on the $\text{CeO}_2(110)$ facet in a 4-coordinated manner with the surface oxygen atoms. **b**, Pd single-atom is anchored on the oxygen vacancy on the $\text{CeO}_2(110)$ facet. **c**, Pd single-atom is anchored on the Ce vacancy on the $\text{CeO}_2(110)$ facet. **d**, Pd single-atom is anchored on the $\text{CeO}_2(110)$ facet in a 2-coordinated manner with the surface oxygen atoms. The corresponding DFT-modelled atomic structures are shown in the insets. The red, yellow, and blue spheres represent oxygen, cerium, and palladium, respectively.

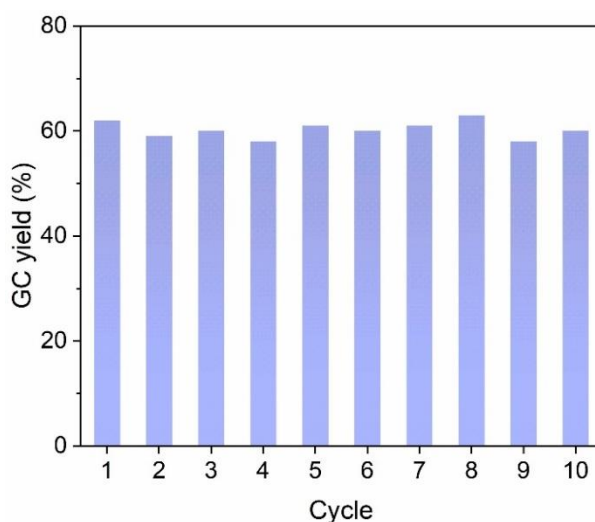


Supplementary Fig. 5 | Substrate scope of Pd₁ ASAC catalyzed Heck and Sonogashira cross-coupling reactions.

Conditions: a: **1** (0.5 mmol), **6** (1 mmol), Pd₁ ASAC (5 mg, 0.35 mol%), K₂CO₃ (1.5 mmol), EtOH/H₂O (2 mL: 2 mL), 100 °C, 10 h, isolated yield. b: **1** (0.5 mmol), **7** (0.75 mmol), Pd₁ ASAC (5 mg, 0.35 mol%), K₂CO₃ (3 mmol), EtOH/H₂O (3.5 mL: 0.5 mL), 100 °C, 10 h, isolated yield.

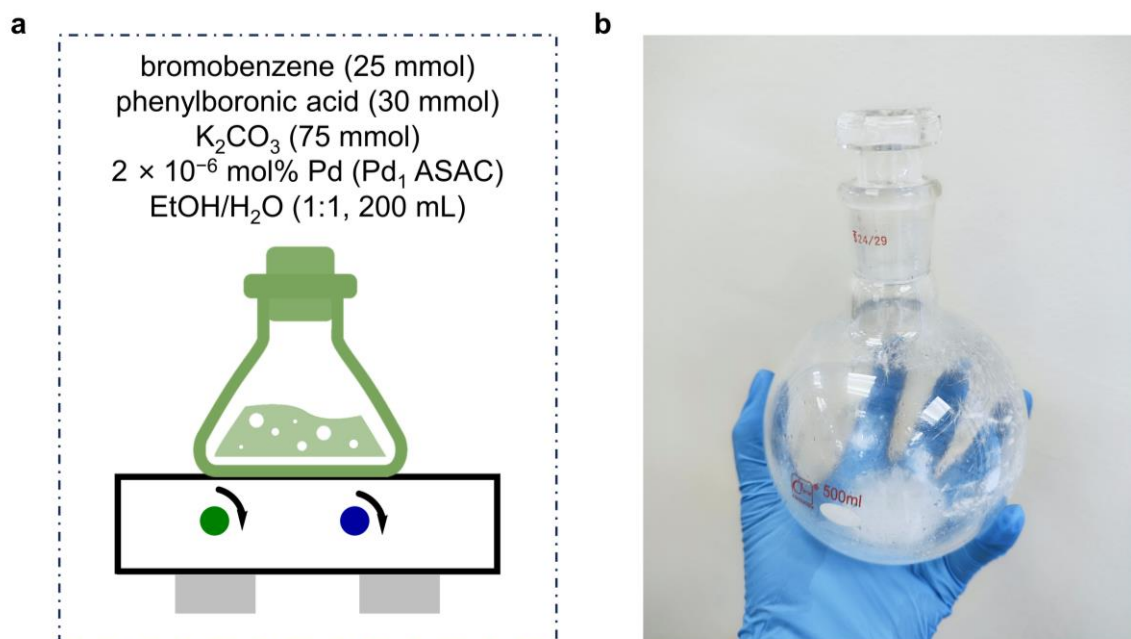


Supplementary Fig. 6 | Stability of Pd₁ ASAC in Suzuki cross-coupling. **a**, Cycling test for the Suzuki cross-coupling reaction using 4-bromotoluene and phenylboronic acid over Pd₁ ASAC. **b**, XRD patterns, **c**, Fourier transformed EXAFS spectra and **d**, XPS spectra of the fresh Pd₁ ASAC and the Pd₁ ASAC recovered after 10 reaction cycles.



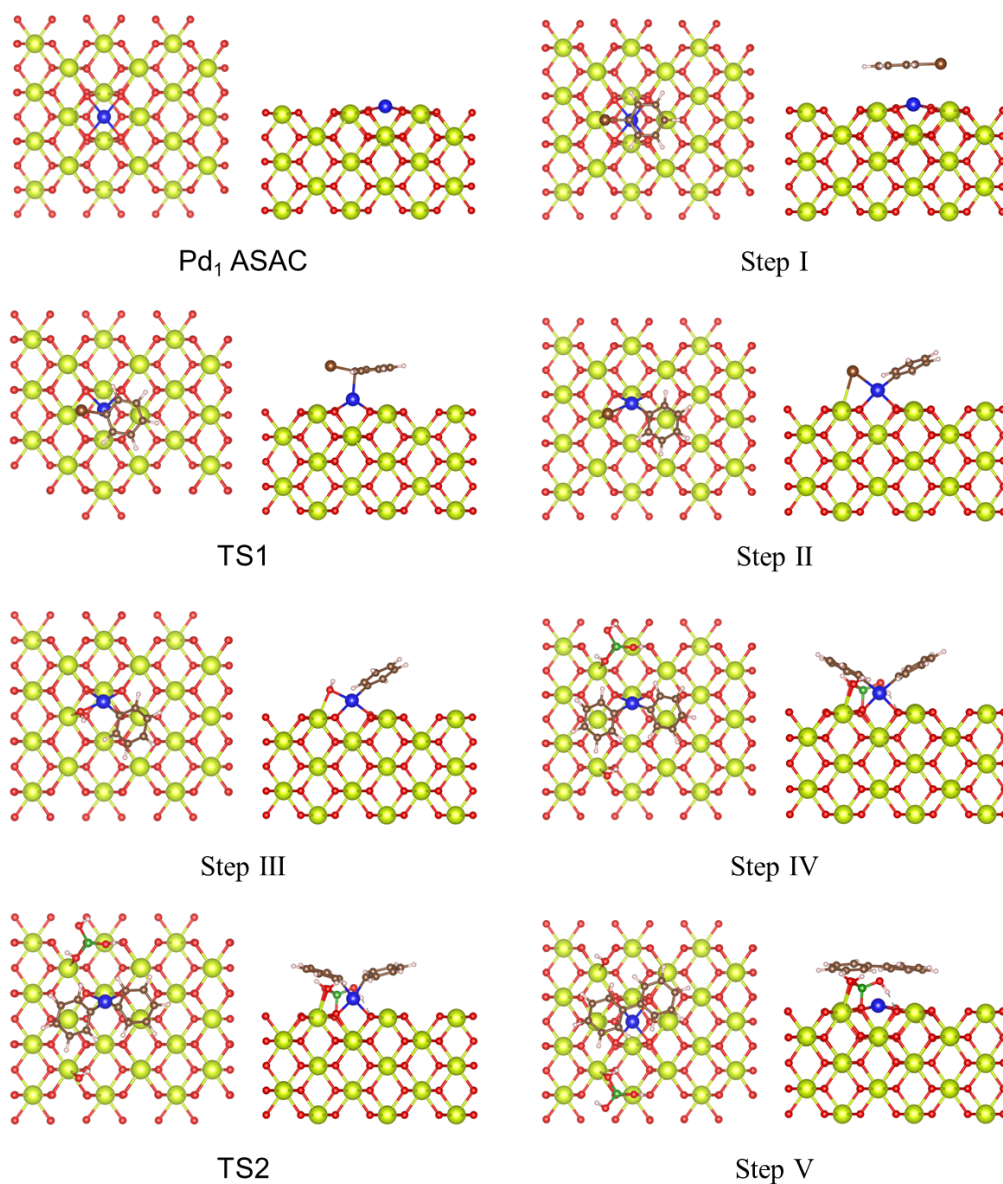
Supplementary Fig. 7 | Cycling test for the Suzuki cross-coupling reaction using 4-bromotoluene and phenylboronic acid over Pd1 ASAC under low-conversion conditions.

Reaction conditions: 4-Bromotoluene (5 mmol), phenylboronic acid (6 mmol), K_2CO_3 (15 mmol), catalyst, EtOH/ H_2O (20 mL/20 mL) were sequentially added to the round-bottomed flask. The flask was heated and stirred in a 100 °C oil bath for 10 min. The products were extracted with DCM, and the yield was calculated by GC analysis. The catalyst was isolated by centrifugation, washed with EtOH/ H_2O , and dried at 80 °C overnight for the next cycle. To avoid the decrease in efficiency caused by the loss of catalyst during operation, it is necessary to keep the dosage ratio of catalyst, substrate, solvent, and base constant in each cycle.

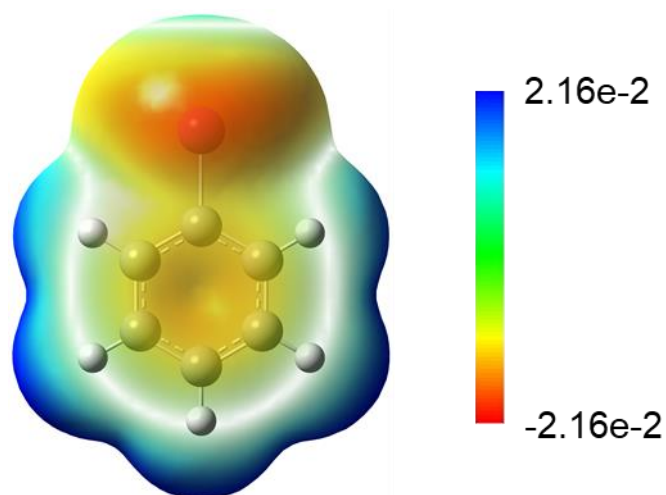


Supplementary Fig. 8 | Turnover number (TON) of Pd_I ASAC in cross-coupling. **a**, Specific conditions of the TON test experiment. **b**, Photograph of the experimental product biphenyl. $\text{TON} = \text{mole of converted aryl halides/mole of Pd catalyst}$

Experimental procedure is shown in Supplementary Fig. 8a : Bromobenzene (25 mmol), phenylboronic acid (30 mmol), K_2CO_3 (75 mmol), Pd_I ASAC (2×10^{-6} mol%, according to Pd), ethanol (100 mL), and H_2O (100 mL) were sequentially added to the flask. The flask was placed in an oil bath preheated to 80 °C, and stirring in an ambient atmosphere, yielding biphenyl (Supplementary Fig. 8b) in approximately 90% yield.

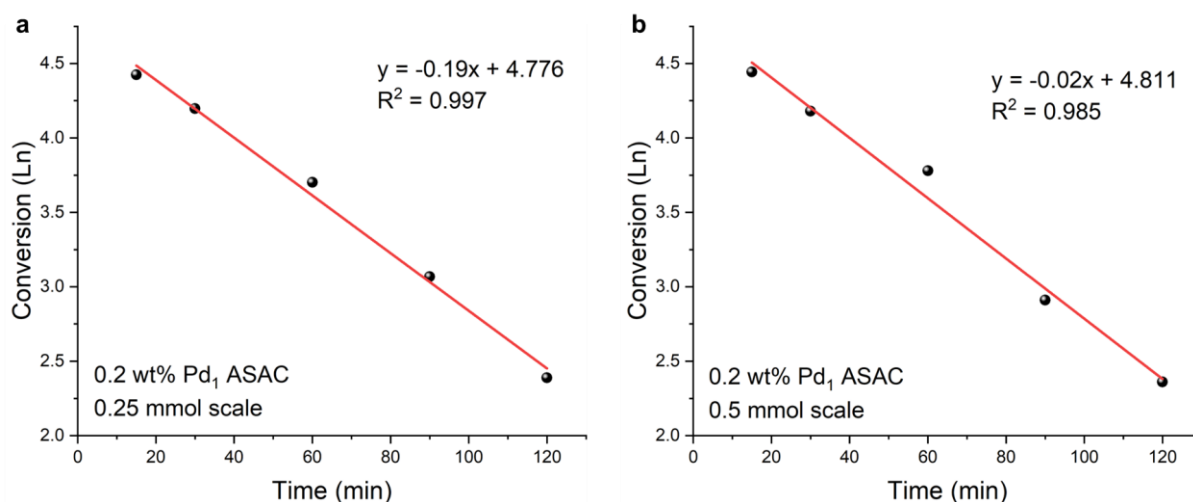


Supplementary Fig. 9 | Atomic structure of Pd₁ ASAC calculated by DFT. Top and side views of the optimized structure of Pd₁ ASAC model in all reaction steps.



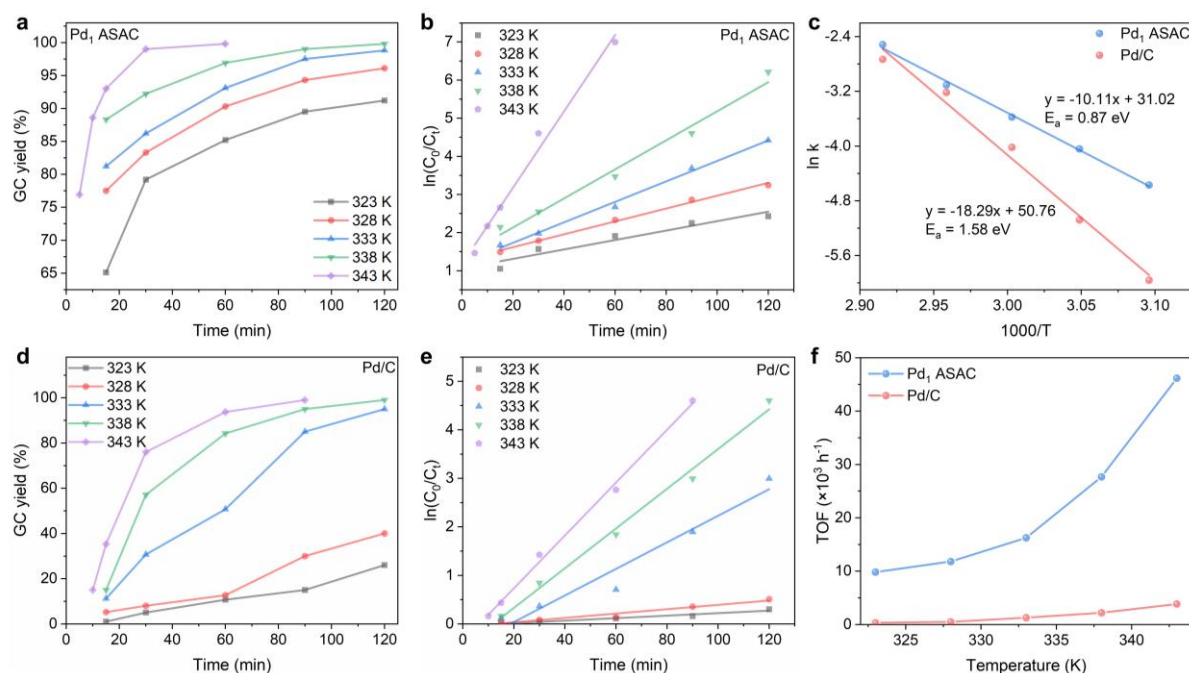
Supplementary Fig. 10 | Electrostatic potential diagram of bromobenzene.

According to the electrostatic potential diagram of bromobenzene, it can be seen that the charges of the molecule tend to gather around the Br atom. As a result, the Br ion after bromobenzene dissociation is adsorbed on the electrophilic Ce site, and the phenyl group is adsorbed on the nucleophilic Pd site.



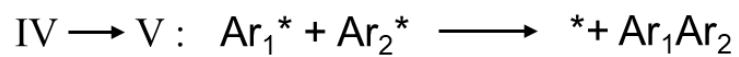
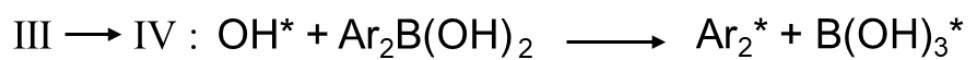
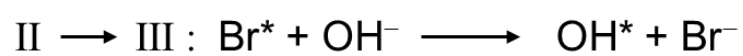
Supplementary Fig. 11 | Kinetic studies of Pd₁ ASAC catalysed Suzuki cross-coupling. The plots of the natural logarithm of the concentration of aryl halide versus time: initial concentration of bromobenzene $C_0 = 0.25$ M (**a**), $C_0 = 0.5$ M (**b**). All reactions were conducted with 0.1 mol% Pd.

Experimental procedure: In a capped reaction tube, sequentially add 0.25 mmol ($C_0 = 0.25$ M) or 0.5 mmol ($C_0 = 0.5$ M) of bromobenzene, 0.6 mmol of phenylboronic acid, 1.5 mmol of K₂CO₃, Pd₁ ASAC (0.1 mol% Pd, according to Pd), 2 mL of ethanol, 2 mL of H₂O, and 0.5 mmol of decane (as an internal standard for GC analysis). Place the reaction tube in an oil bath at 25 °C and stir under ambient atmosphere for the desired time (10-120 min). After the reaction, remove the cap and transfer a portion of the solution into a vial. Extract with DCM and analyze the conversion of reactants via gas chromatography. Under optimized reaction conditions, the natural logarithm of the bromobenzene concentration versus time plot is linear, indicating a first-order reaction with respect to the aryl halide. This suggests that the aryl halide is involved in the rate-determining step, consistent with DFT studies.



Supplementary Fig. 12 | Catalytic performance for the Suzuki reaction of bromobenzene and phenylboronic acid. Time-dependent yield of biphenyl over Pd₁ ASAC (a) and Pd/C (d). The corresponding $\ln(C_0/C_t)$ -time curves for Suzuki coupling reactions over Pd₁ ASAC (b) and Pd/C (e). c, Arrhenius plots of the reaction over Pd₁ ASAC and commercial Pd/C. f, A TOF comparison of Pd₁ ASAC and commercial Pd/C.

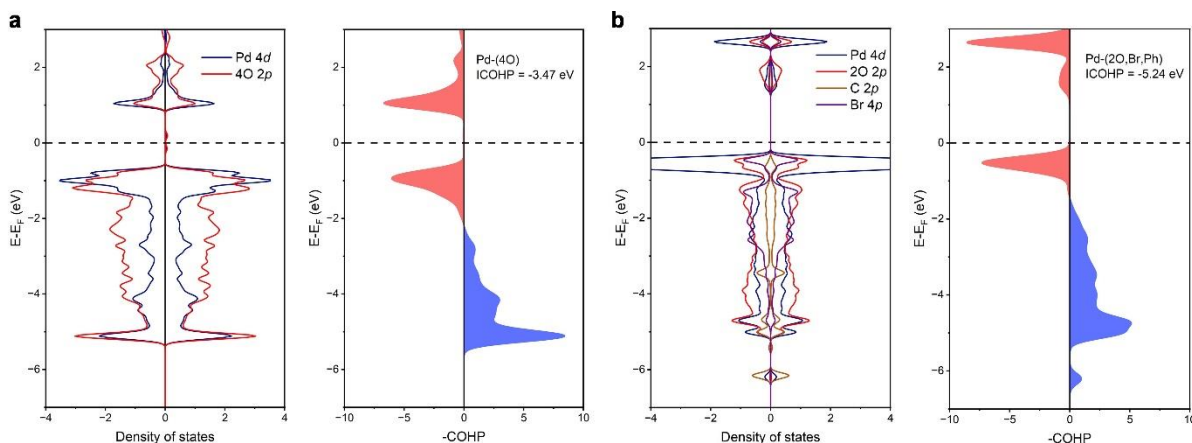
To further study the reaction kinetics of Pd₁ ASAC, we investigated the coupling reaction of bromobenzene and phenylboronic acid at different temperatures (323, 328, 333, 338, and 343 K) (Supplementary Fig. 12a) and compared them with commercial Pd/C (Supplementary Fig. 12d). The relationship between $\ln(C_0/C_t)$ and time is consistent with first-order reaction kinetics (Supplementary Fig. 12b and e). The rate constant (k) of the reaction can be calculated from the slope of the $\ln(C_0/C_t)$ -time curve, and then the apparent activation energy can be calculated by plotting $\ln k$ as a function of $1000/T$ in an Arrhenius plot (Supplementary Fig. 12c). The results demonstrate that the activation energy of Pd₁ ASAC-catalyzed reaction is much lower than that of commercial Pd/C. Notably, Pd₁ ASAC is able to exhibit a significant turnover frequency (TOF) of 46,149 h⁻¹ at 343 K (Supplementary Fig. 12f), outperforming all listed catalysts by about an order of magnitude or more in the Suzuki cross-coupling reactions (Supplementary Table 3).



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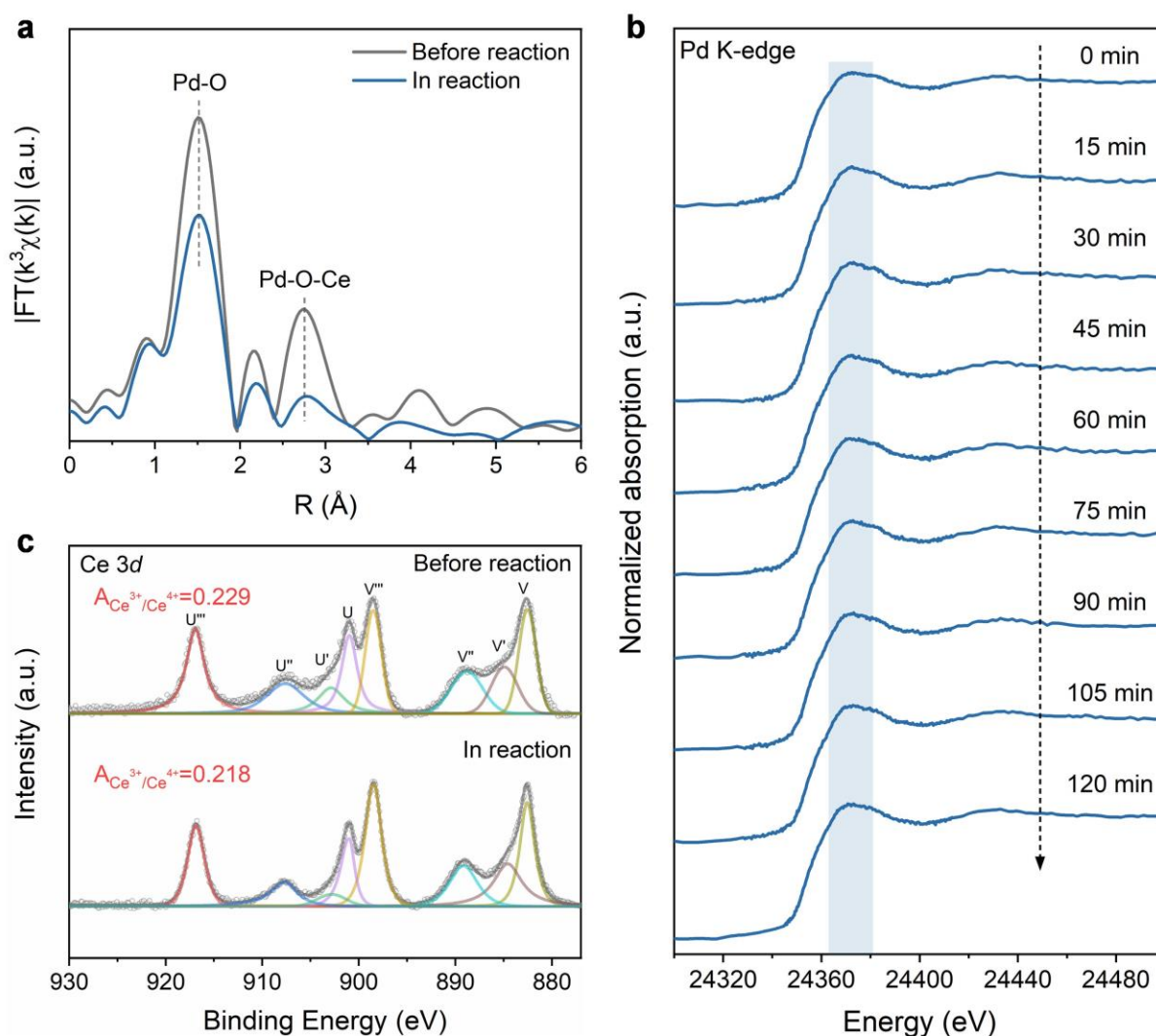
154 **Supplementary Fig. 13** | The chemical reaction equation of the entire reaction process.

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Supplementary Fig. 14 | Electronic structure of Pd₁ ASAC with different coordination configurations. **a**, DOS and ICOHP of the initial Pd-(4O) coordination structure. **b**, DOS and ICOHP of the Pd-(2O, Br, Ph) coordination structure after oxidative addition. The square-planar coordinated Pd-O bonds have obvious occupation of the antibonding orbital near E_F , while the occupation of the antibonding orbital near E_F is significantly weakened in the new tetra-coordinated structure formed by the dissociation of bromobenzene. The calculated ICOHP also confirms that Pd center has stronger bond interaction after bromobenzene dissociation (-5.24 eV vs. -3.47 eV), which is believed to be the driving force for the opening of Pd-O bonds.

The Pd d -orbital center of the four-coordinated PdO₄ motif lies further below E_F and once two Pd-O bonds are opened to form new four-coordinated structures, the Pd d -orbital centers shift significantly closer to E_F . Therefore, the Pd-adsorbate interactions are enhanced and show better catalytic activity. As a result, the Pd₁ ASAC can adaptively regulate the Pd electronic structure to facilitate catalysis through dynamic coordination configuration tuning during the reaction. The $5d$ -orbitals of adjacent Ce revealed a slight decrease in the integrated occupied state below E_F and a gradual increase in empty states, with these empty states shifting toward E_F . All these observations indicate the accumulation of positive charge over Ce as the reaction proceeds.

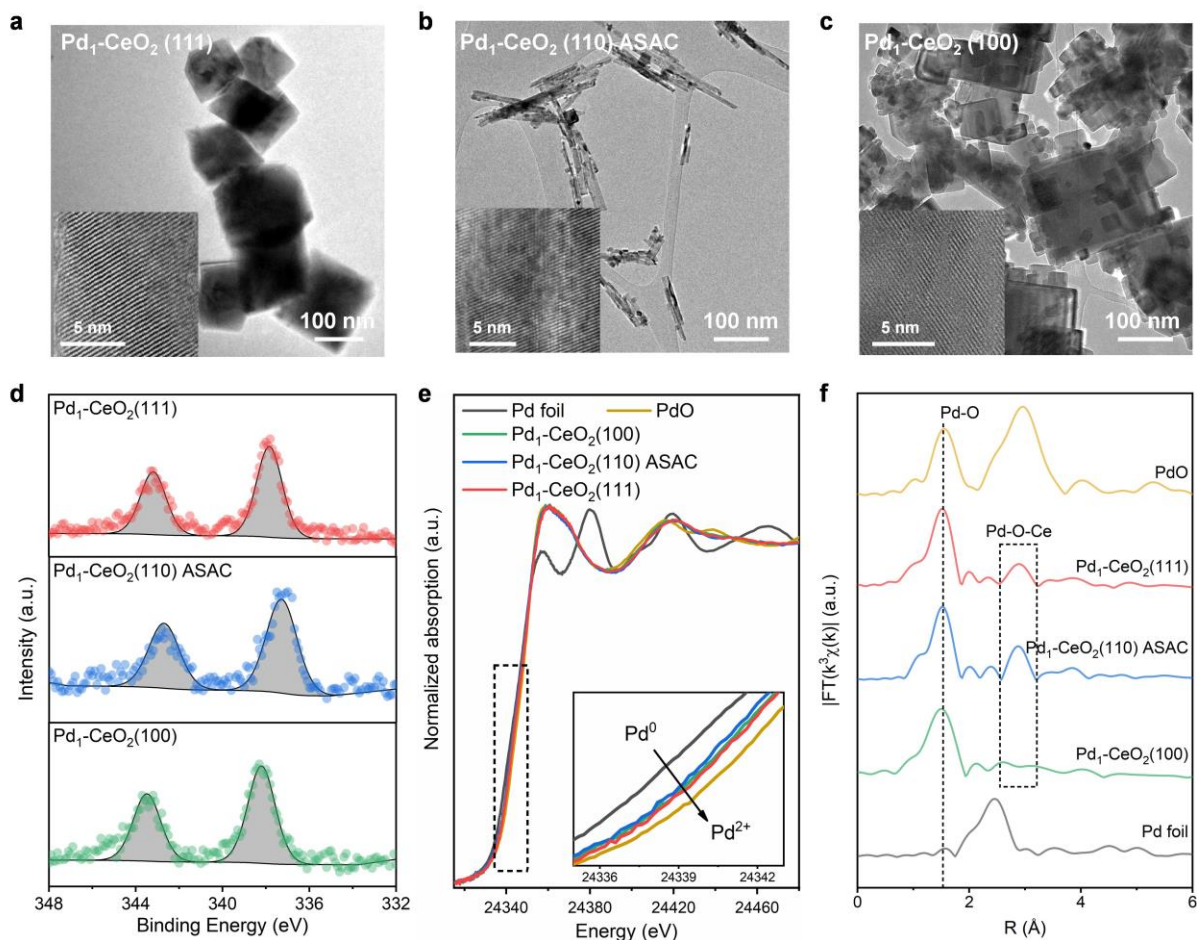


Supplementary Fig. 15 | Operando XANES and ex situ XPS spectra during Suzuki cross-

coupling reaction. a, Fourier-transformed EXAFS spectra of Pd₁ ASAC collected before and during the Suzuki cross-coupling reaction. **b**, Operando Pd K-edge XANES of Pd₁ ASAC recorded at different times in the Suzuki cross-coupling reaction. **c**, Ex situ XPS spectra of Pd₁ ASAC measured before and in the Suzuki cross-coupling reaction. The peaks V' and U' correspond to the signals of Ce³⁺ species, and the others are assigned to the Ce⁴⁺ species.

The changes in the chemical state of Pd₁ ASAC during the Suzuki cross-coupling reaction can be observed from the in situ Pd K-edge XANES. Throughout the reaction, there are no significant peak shifts or intensity changes, indicating that the valence state of Pd does not undergo substantial changes. In the Fourier-transformed EXAFS spectra, the intensity of the main peaks associated with the first coordination shell path Pd-O and the second coordination shell path Pd-O-Ce decreases, indicating a

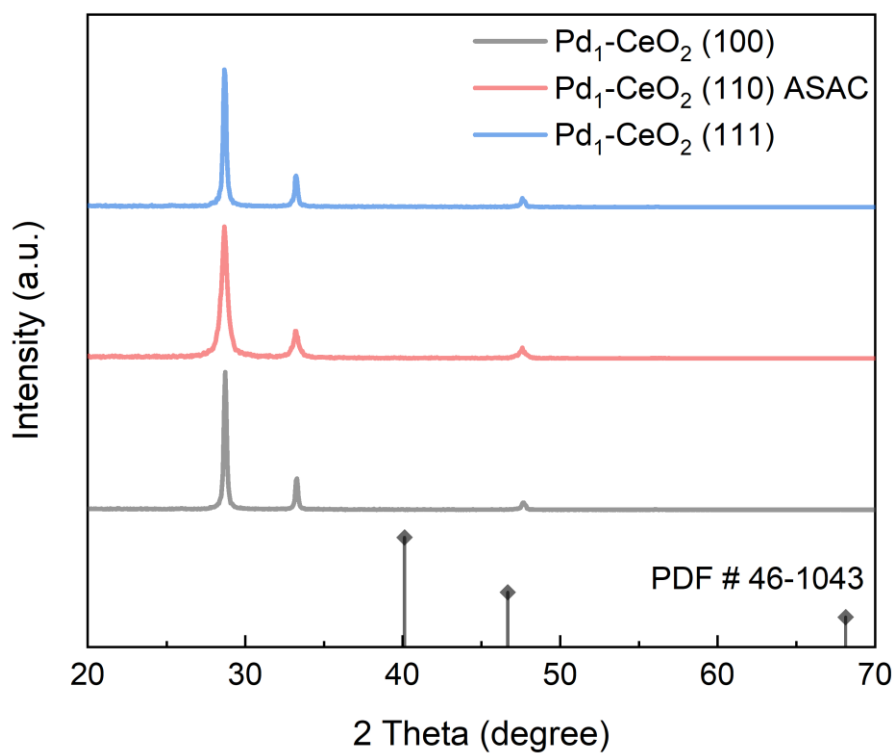
reduction in the coordination number of Pd-O. This demonstrates that during the reaction, Pd₁ ASAC transfers from the initial Pd-O₄ planar four-coordination structure through the opening of two Pd-O bonds, with the Pd atom being pulled up and bonded by dissociated phenyl and bromide ions, forming a new four-coordinated configuration, consistent with DFT calculations. Ex situ XPS results showed a slight increase in Ce³⁺/Ce⁴⁺ ratio during the reaction, which may be due to the transfer of electrons from Ce atoms to Pd atoms in ASAC during the oxidative addition process, thus ensuring that the valence state of Pd remains constant.



Supplementary Fig. 16 | characterizations of Pd₁ anchored on facet-dependent CeO₂. TEM

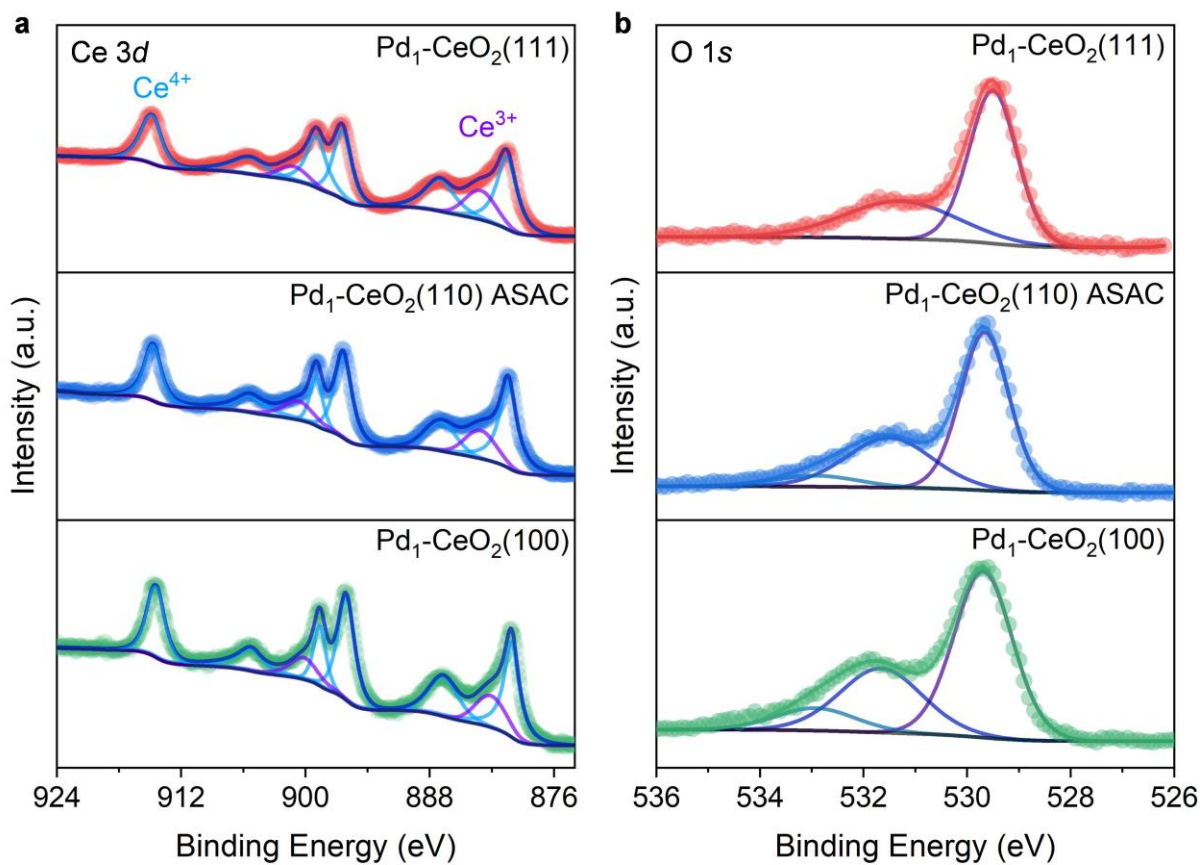
images of Pd₁-CeO₂(111) (**a**), Pd₁-CeO₂(110) ASAC (**b**), Pd₁-CeO₂(100) (**c**), inserts are their HRTEM images respectively. **d**, Pd 3d XPS spectra, **e**, Pd K-edge XANES spectra, **f**, Pd K-edge Fourier transformed EXAFS spectra of Pd single atoms on facet-dependent CeO₂.

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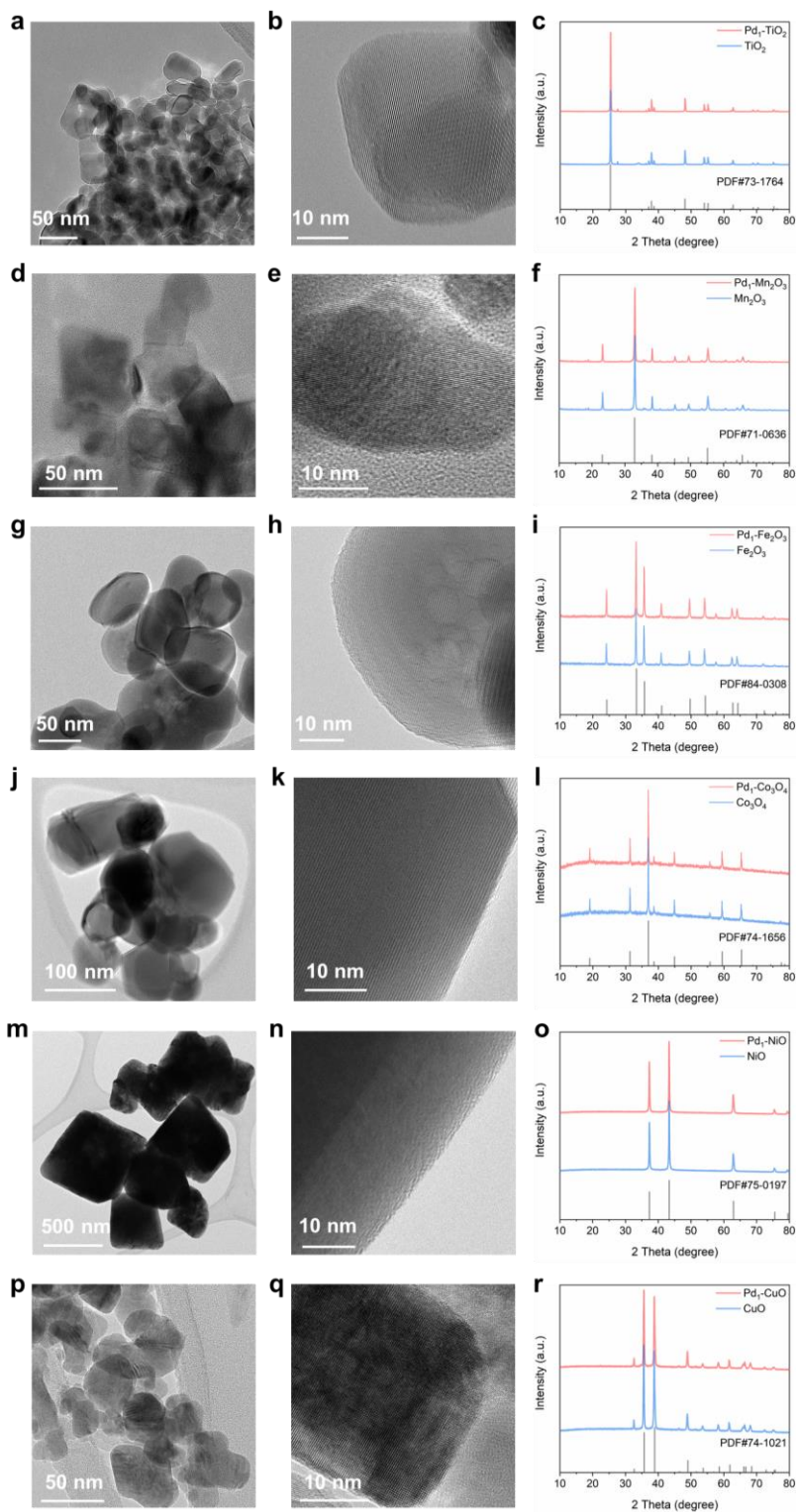


204 **Supplementary Fig. 17** | XRD patterns of Pd₁-CeO₂(100), Pd₁-CeO₂(110) ASAC, and Pd₁-CeO₂(111).

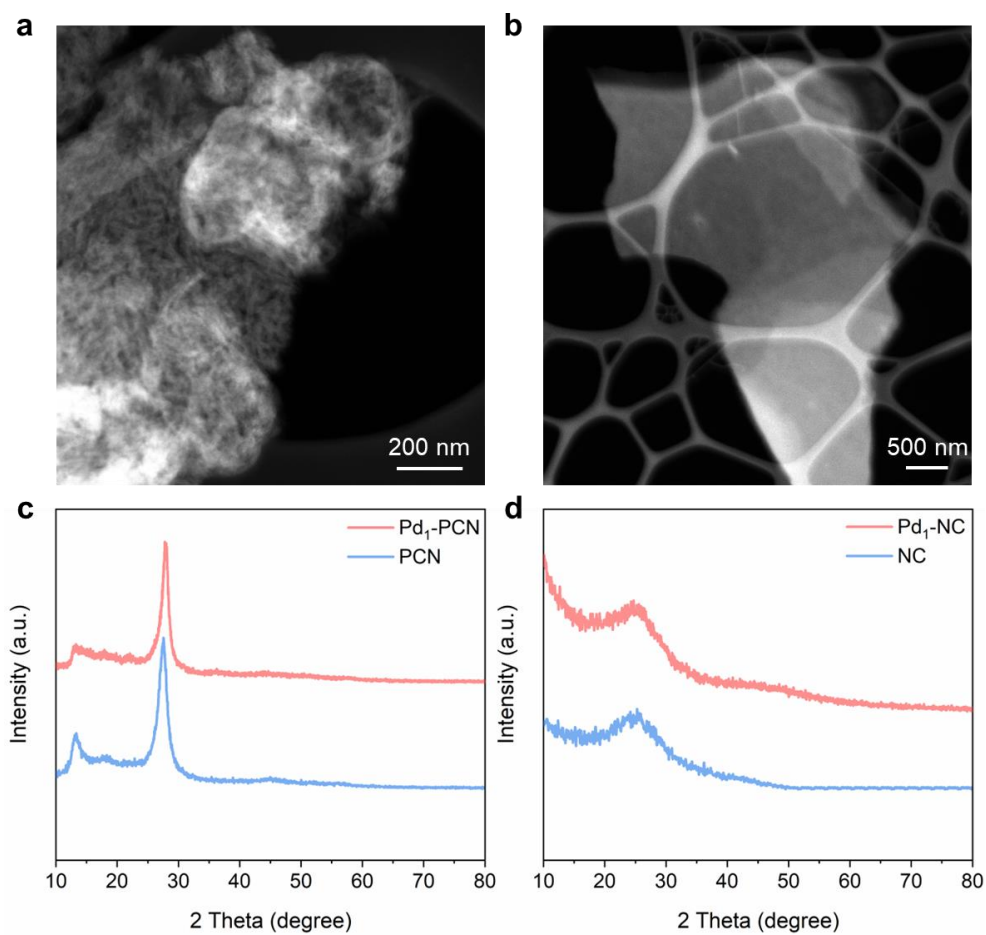
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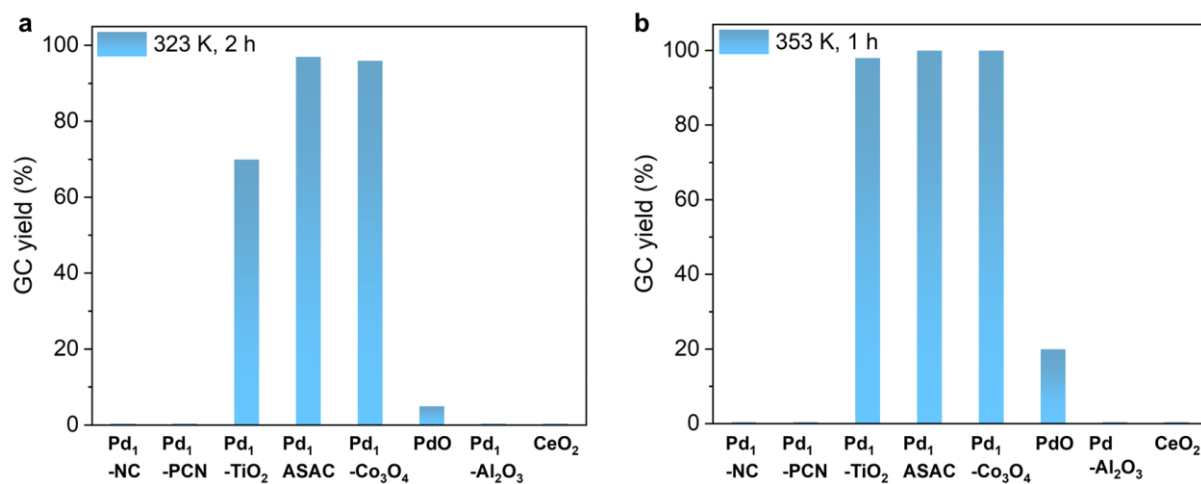
Supplementary Fig. 18 | Characterization of catalysts. **a**, Ce 3d XPS spectra, **b**, O 1s XPS spectra of Pd single atoms on facet-dependent CeO_2 .



211 **Supplementary Fig. 19** | Characterization of catalysts. Low-magnification TEM images of Pd₁-TiO₂
 212 (a), Pd₁-Mn₂O₃ (d), Pd₁-Fe₂O₃ (g), Pd₁-Co₃O₄ (j), Pd₁-NiO (m), and Pd₁- CuO (p). High-resolution
 213 TEM images of Pd₁-TiO₂ (b), Pd₁-Mn₂O₃ (e), Pd₁-Fe₂O₃ (h), Pd₁-Co₃O₄ (k), Pd₁-NiO (n), and Pd₁- CuO
 214 (q). XRD patterns of Pd₁-TiO₂ (c), Pd₁-Mn₂O₃ (f), Pd₁-Fe₂O₃ (i), Pd₁-Co₃O₄ (l), Pd₁-NiO (o), and Pd₁-
 215 CuO (r).



Supplementary Fig. 20 | Characterization of catalysts. TEM images of Pd₁-PCN (a), Pd₁-NC (b). XRD patterns of Pd₁-PCN (c), Pd₁-NC (d).



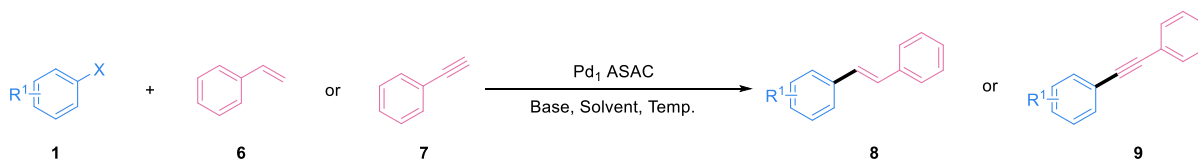
Supplementary Fig. 21 | Yield of biphenyl for reactions at 323 K (a) and 353 K (b).



Supplementary Fig. 22 | The schematic illustration of Suzuki cross-coupling reactions.

General procedures: Compounds **1**, **2**, or **3**, Pd₁ ASAC, and the base were dissolved in the specified solvent. The reaction mixture was stirred at the designated temperature for the required duration. After completion, the solvent was evaporated under reduced pressure, and the residue was re-dissolved in a mixture of ethyl acetate and water. The organic phase was separated and dried over anhydrous sodium sulfate. The solvent was again evaporated under reduced pressure, yielding the crude product mixture. Further purification by silica gel column chromatography afforded compound **4** or **5**.

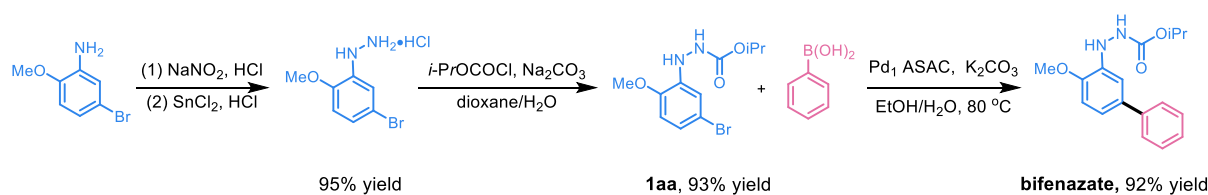
Different conditions for diverse substrates, refer to Fig. 3: Conditions: a: **1** (0.5 mmol), **2** (ArB(OH)₂, 0.6 mmol), Pd₁ ASAC (5 mg, 0.35 mol%), K₂CO₃ (1.5 mmol), EtOH/H₂O (2 mL: 2 mL), 80 °C, 10 h. b: **1** (0.5 mmol), **2** (ArB(OH)₂, 0.6 mmol), Pd₁ ASAC (5 mg, 0.35 mol%), K₂CO₃ (1.5 mmol), EtOH/H₂O (2 mL: 2 mL), 100 °C, 20 h. c: **1** (0.5 mmol), **2** (ArB(OH)₂, 0.6 mmol), Pd₁ ASAC (5 mg, 0.35 mol%), K₂CO₃ (1.5 mmol), THF/H₂O (2 mL: 2 mL), 100 °C, 24 h. d: **1** (0.2 mmol), **2** (ArBF₃K, 0.3 mmol), KHCO₃ (0.6 mmol), Pd₁ ASAC (10.5 mg, 0.35 mol%), EtOH/H₂O (1.5 mL: 0.5 mL), 100 °C, 24 h. e: **1** (0.5 mmol), **2** (ArB(OH)₂, 1.2 mmol), Pd₁ ASAC (5 mg, 0.35 mol%), K₂CO₃ (3 mmol), EtOH/H₂O (2 mL: 2 mL), 80 °C, 10 h, isolated yield. f: **1** (0.2 mmol), **3** (0.24 mmol), K₃PO₄ (0.6 mmol), Pd₁ ASAC (10.5 mg, 0.35 mol%), EtOH/H₂O (1.6 mL: 0.4 mL), 80 °C, 24 h. g: **1** (0.2 mmol), **3** (0.24 mmol), K₃PO₄ (0.6 mmol), Pd₁ ASAC (10.5 mg, 0.35 mol%), EtOH/H₂O (1.6 mL: 0.4 mL), 80 °C, 63 h, isolated yield.



Supplementary Fig. 23 | The schematic illustration of the Heck and Sonogashira cross-coupling reactions.

General procedures: Compounds **1**, **6**, or **7**, Pd₁ ASAC, and the base were dissolved in the specified solvent. The reaction mixture was stirred at the designated temperature for the required duration. After completion, the solvent was evaporated under reduced pressure, and the residue was re-dissolved in a mixture of ethyl acetate and water. The organic phase was separated and dried over anhydrous sodium sulfate. The solvent was again evaporated under reduced pressure, yielding the crude product mixture. Further purification by silica gel column chromatography afforded compound **8** or **9**.

Different conditions for diverse substrates: a: **1** (0.5 mmol), **6** (1 mmol), Pd₁ ASAC (5.3 mg, 0.07 mol%), K₂CO₃ (1.5 mmol), EtOH/H₂O (2 mL: 2 mL), 100 °C, 10 h, isolated yield. b: **1** (0.5 mmol), **7** (0.75 mmol), Pd₁ ASAC (5.3 mg, 0.07 mol%), K₂CO₃ (3 mmol), EtOH/H₂O (3.5 mL: 0.5 mL), 100 °C, 10 h, isolated yield.



Supplementary Fig. 24 | Synthesis of bifentazate.

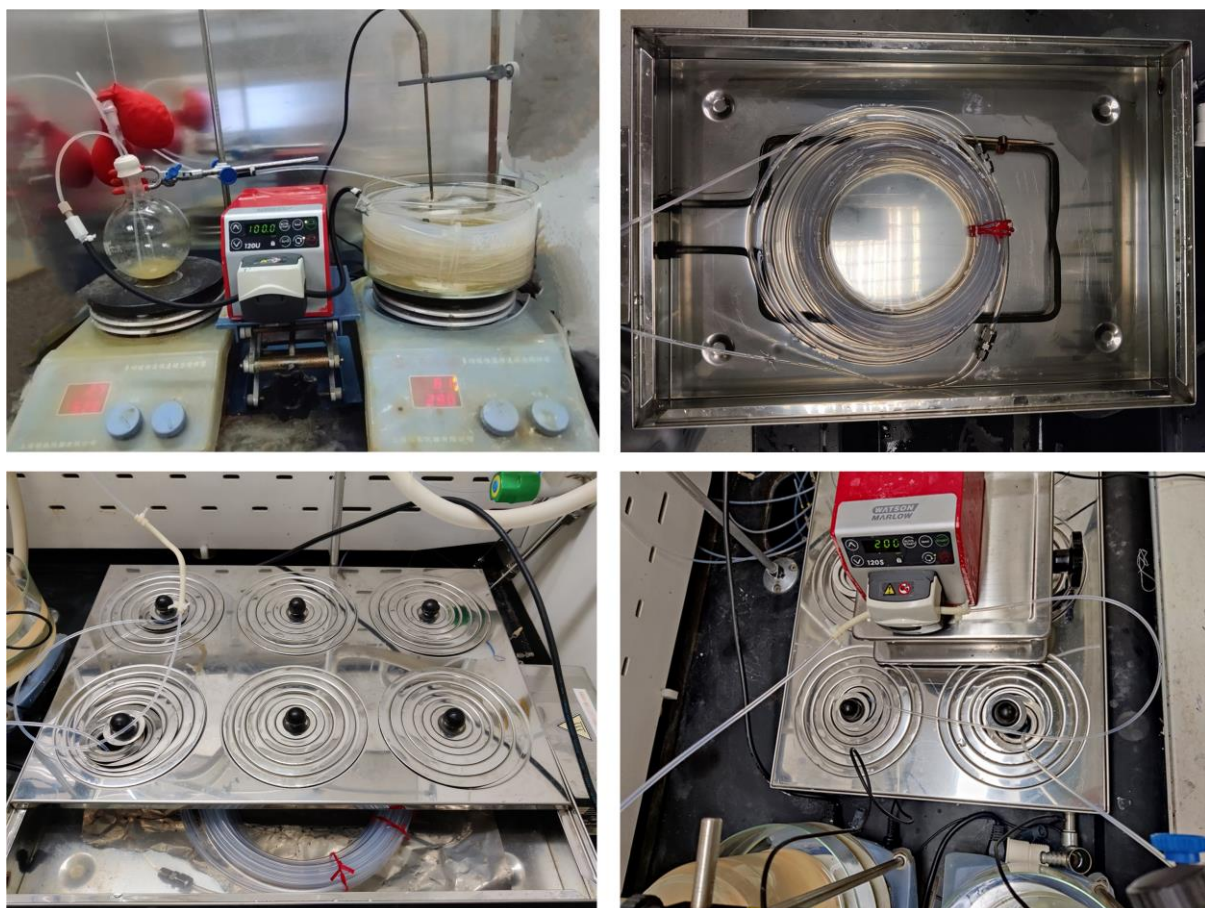
Procedure for the preparation of 5-bromo-2-methoxyphenylhydrazine hydro-chloride: The reaction mixture was maintained at 0 °C during this procedure. 5-bromo-2-methoxyaniline (2.00 g, 10.0 mmol) was added to vigorously stirred concentrated HCl (17 mL) and aged for 10 min. A solution of NaNO₂ (691 mg, 10.0 mmol) in distilled water (4 mL) was added dropwise over 10 min and the mixture was stirred for an additional 15 min. A solution of SnCl₂·H₂O (498 mg, 22.0 mol) in concentrated HCl (5 mL) was added dropwise. The reaction mixture was stirred for 30 min and filtered. The product was dried in vacuo overnight to afford 5-bromo-2-methoxyphenylhydrazine hydrochloride as a brown solid (2.38 g, 95% yield).

Procedure for the preparation of compound **1aa**: A solution of 5-bromo-2-methoxyphenylhydrazine hydrochloride (2.00 g, 7.90 mmol) and triethylamine (2.28 mL, 16.6 mmol) in CH₂Cl₂ (16 mL) was cooled to 0 °C and a solution of isopropyl chloroformate (0.93 mL, 8.08 mmol) in CH₂Cl₂ (8 mL) was added dropwise at a rate that maintained a temperature below 0 °C. When the addition was complete, the reaction mixture was allowed to warm to room temperature. The reaction mixture was stirred for 3h and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (n-hexane/EtOAc, 2:1) to afford isopropyl 3-(5-bromo-2-methoxyphenyl)carbazate as a colorless solid (2.21 g, 93% yield).

Procedure for the preparation of bifentazate: In a 20-mL round-bottom flask with a stir bar, added isopropyl 3-(5-bromo-2-methoxyphenyl)carbazate (151 mg, 0.5 mmol), phenylboronic acid (92 mg, 0.75 mmol), K₂CO₃ (207 mg, 1.5 mmol), Pd₁ ASAC (5 mg, 0.35 mol%), H₂O (2 mL), and EtOH (2 mL) and the system was sealed with a septum. The mixture was stirred at 80 °C for 12 h. The mixture was diluted with H₂O (50 mL) and EtOAc (50 mL), and filtered through a Celite pad. The filtrate was separated into two layers and the aqueous layer was extracted with EtOAc (2 × 50 mL). The combined organic layers were washed with brine (50 mL), dried over Na₂SO₄, and concentrated in vacuo. The

279 residue was purified by flash column chromatography on silica gel (n-hexane/EtOAc, 50:1 to 5:1) to
280 afford bifenazate as a yellow solid (138 mg, 92% yield).

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Supplementary Fig. 25 | High-rate circulation flow devices for evaluating catalytic performance. Reaction tubing, PFA tubing reactor (outer diameter (O.D.) = 4.8 mm, inner diameter (I.D.) = 3.2 mm, volume (V) = 240 mL) Contherm stainless steel water bath; load 2300W; set temperature: 80 ° C. **d**, Watson-Marlow 100 series cased pump; 120S control, drive with 114DV Pumphead; 4 rollers, Max. operating pressure 2bar, Max.; Tygon® Norprene® Peristaltic Pump.



Conditions optimization:

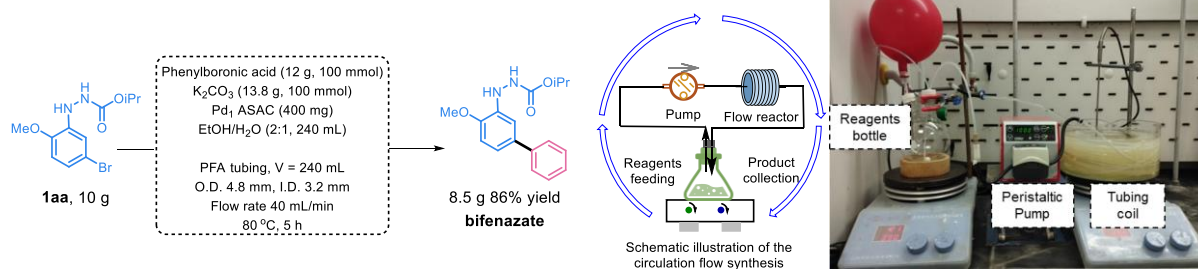
Entries	Pd-CeO ₂ (X mg)	Equivalents of boronic acid and base	Flow rate	Solvent	Temp.	Conversion
1 ^[a]	20	1.5	10 mL/min	EtOH/H ₂ O (1:1)	80°C.	0
2 ^[a]	50	1.5	10 mL/min	EtOH/H ₂ O (1:1)	80°C.	30%
3 ^[b]	50	3	10 mL/min	EtOH/H ₂ O (1:1)	80°C.	70% (block)
4 ^[a]	50	3	10 mL/min	THF/H ₂ O (1:1)	80°C.	0
5 ^[c]	50	3	10 mL/min	EtOH/H ₂ O (2:1)	80°C.	100% (91% yield)

Conditions: [a] Reaction time = 12 h, [b] Reaction time = 3 h, [c] Reaction time = 5 h.

Supplementary Fig. 26 | Conditions optimization for flow synthesis.

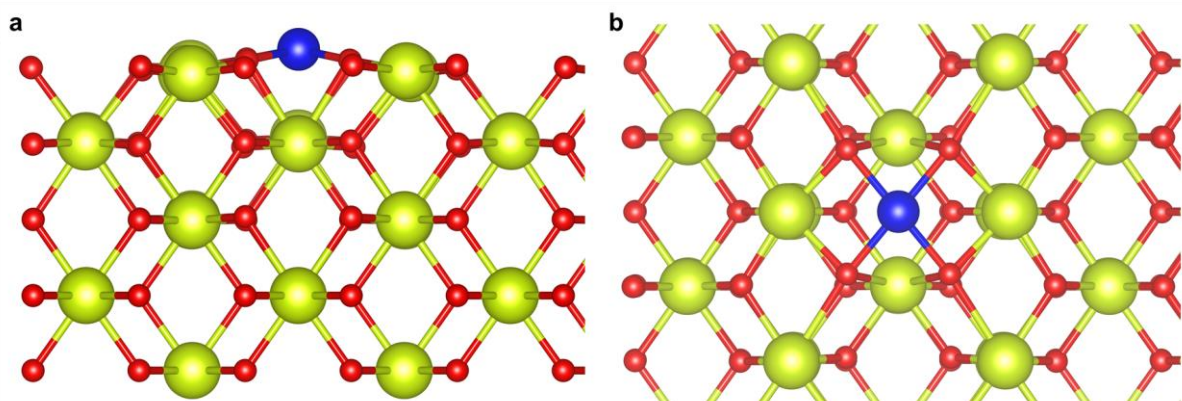
Blockage of the reaction tube is a limiting factor for heterogeneous catalysts in flow synthesis experiments. As illustrated in **Supplementary Fig. 26**, at 1 g scale, insufficient feeding of the catalyst and phenylboronic acid results in low reaction conversion. However, increasing the amounts of catalysts and phenylboronic acid leads to micro-tubing reactor blocking. To address this issue, we optimized the flow rate (10 mL/min), catalyst feeding (50 mg), K₂CO₃ and boronic acid (3 equiv.), and solvent ratio (EtOH/H₂O, 2:1), achieving both sufficient reagent solubility and high reaction yields. Consequently, this modified condition was adopted for circulation flow synthesis.

Note: When we performed a larger scale (10 g) synthesis, we further increased the amount of catalyst (400 mg) to achieve efficient reaction and flow rate (40 mL/min) to avoid clogging.

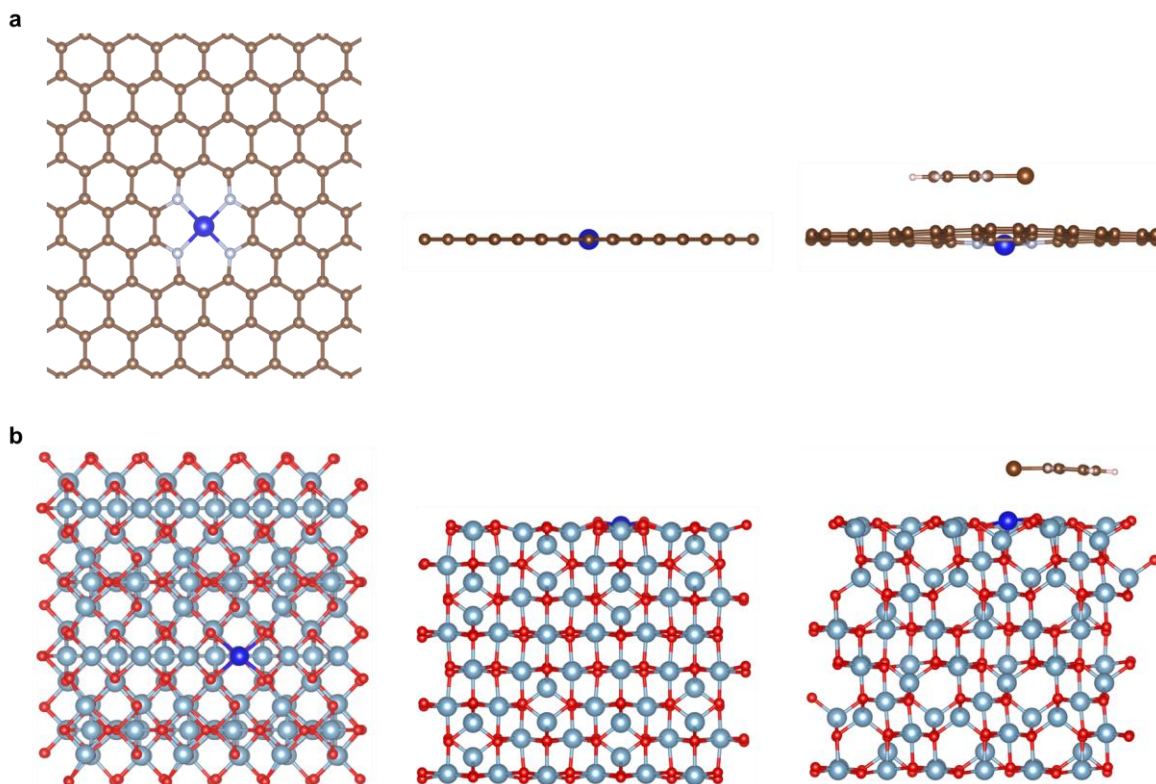


Supplementary Fig. 27 | Large-scale circulated flow synthesis of bifenzate. High-purity perfluoroalkoxy polymer (HPFA) tubing and fittings were purchased from IDEX Health & Science Technologies. The peristaltic pump was purchased from Watson-Marlow.

Substrate **1aa** was mixed with 400 mg Pd_1 ASAC, 12 g phenylboronic acid and 13.8 g potassium carbonate in 240 mL of EtOH/ H_2O (2:1) to create a slurry. The slurry was continuously pumped at 40 mL/min using a peristaltic pump and directed to the PFA tubing reactor (outer diameter (O.D.) = 4.8 mm, inner diameter (I.D.) = 3.2 mm, volume (V) = 240 mL) which was heated to 80 °C with the heating module. The reaction mixture was recirculated to the original reservoir until the reaction was completed to afford bifenzate in 86% isolated yield.



Supplementary Fig. 28 | Structure of Pd₁ ASAC calculated by DFT. Side (a) and top (b) views of the optimized structure of Pd₁ ASAC model. The red, yellow, and blue spheres represent oxygen, cerium, and palladium, respectively.



Supplementary Fig. 29 | Structural models of Pd₁-NC (a) and Pd₁-Al₂O₃ (b) and optimized structures in the oxidative addition step calculated by DFT.

Reducible support is essential for the design of highly active catalysts for cross-coupling reactions, as it functions as electron reservoirs to address the high reaction energy barriers associated with the bivalent elevation at a single metal site. Moreover, achieving a dynamic and reversible coordination transition between Pd single atoms and the support is essential to regulate the electronic structure of the metal center during the reaction. However, in the case of NC, the Pd-N bonds lack the ability to undergo dynamic transitions during the reaction, resulting in catalytic inactivity. In addition, carbon is known to be a conduct but not to be a reducible support that has the excellent capability to accept/donor the electrons. As illustrated in **Supplementary Fig. 29a**, the dissociation of the C-Br bond in bromobenzene was adsorbed on Pd₁-NC, followed by structural optimization. However, the system reverted to the initial bromobenzene configuration, which was also shown in the case of Al₂O₃ (**Supplementary Fig. 29b**), demonstrating that single atom Pd on a non-reducible support is incapable of activating bromobenzene (**Supplementary Video 2 and 3**).

331 **Supplementary Table 1** | The formation energies of Pd single-atom on different crystal planes of CeO₂.

Crystal planes	E _{form} (eV)
Pd ₁ -CeO ₂ (110) ASAC	−1.37
Pd ₁ -CeO ₂ (100)	−1.66
Pd ₁ -CeO ₂ (111)	0.84
Pd ₁ -NC	−2.76
Pd ₁ -Al ₂ O ₃	−5.52

332

333




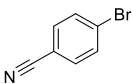

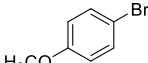
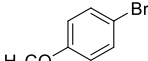








334 **Supplementary Table 2** | Results of EXAFS fittings of catalysts.



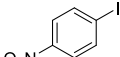




Sample	Shell	<i>N</i>	<i>R</i> (Å)	σ^2 (10 ⁻³ Å ²)	R factor
Pd ₁ ASAC	Pd-O	4.0 ± 0.5	2.01 ± 0.01	0.0024 ± 0.0013	0.026
	Pd-Ce	3.9 ± 1.3	3.14 ± 0.02	0.0068 ± 0.0018	
Pd ₁ -CeO ₂ (110) ASAC	Pd-O	4.0±0.5	1.99±0.01	0.0014±0.0008	0.024
	Pd-Ce	3.7±1.5	3.15±0.02	0.0055±0.0019	
Pd ₁ -CeO ₂ (100)	Pd-O	4.1±0.6	1.99±0.01	0.0012±0.0009	0.007
Pd ₁ -CeO ₂ (111)	Pd-O	3.9±0.3	1.99±0.01	0.0010±0.0009	0.019
	Pd-Ce	3.4±1.0	3.18±0.02	0.0081±0.0026	
Pd ₁ -NC	Pd-N	4.3±0.2	1.95±0.02	0.0032±0.0009	0.009
Pd ₁ -Al ₂ O ₃	Pd-O	3.9±0.5	2.19±0.01	0.0061±0.0009	0.029

335 *N*, coordination number; *R*, distance between absorbing and backscattering atoms; σ^2 , Debye-Waller
336 factor to account for thermal and structural disorders; R factor as a measure of the goodness of fit.

337

Supplementary Table 3 | Comparison of turnover frequencies of reported Pd catalysts in Suzuki cross-coupling reaction.

Entry	Catalyst	Substrate	Conditions	Reaction mode	TOF (h ⁻¹)	Ref.
1	C ₁₈ H ₂₅ C ₁₂ N ₃ O ₄ Pd		333 K/24 h	homogeneous	2.0	[1]
2	C ₂₄ H ₄₅ NP ₂ Pd		338 K/16 h	homogeneous	3.2	[2]
3	(PPh ₃) ₂ PdCl ₂		383 K/24 h	homogeneous	8.2	[3]
4	Pd(OAc) ₂		333 K/6 h	homogeneous	15	[4]
5	Pd/C		393 K/2 h	heterogeneous	10000	[5]
6	Pd/CeO ₂		393 K/4 h	heterogeneous	4100	[5]
7	Pd/Graphene		353 K/20 h	heterogeneous	57	[6]
8	Pd/NMC		323 K/2 h	heterogeneous	137	[7]
9	Pd _{0.10} /g-C ₃ N ₄		298 K/30 min	heterogeneous	120.93	[8]
10	Pd@mZ-x-H		333 K/30 min	heterogeneous	4050	[9]
11	Pd@CMK-3		333 K/5 min	heterogeneous	2800	[10]
12	Pd(0)-MCM-41		353 K/12 h	heterogeneous	6990	[11]
13	Pd/Pyr-GDY		353 K/5 h	heterogeneous	18000	[12]
14	Pd ₁ /UiO-66-NH ₂		333 K/2 h	heterogeneous (SAC)	13043	[13]
15	Pd-ECN		393 K/10 min	heterogeneous (SAC)	549	[14]

16	Pd-SAs/3DOM-CeO ₂		298 min	K/15	heterogeneous (SAC)	520.01	[15]
17	Pd-N ₃ C ₁		373 min	K/24 h	heterogeneous (SAC)	4.2	[16]
18	Pd ₁ /C ₃ N ₄ /rGO		333 min	K/60	heterogeneous (SAC)	362.37	[17]
19	Pd/C		323 min	K/2 h	heterogeneous	325	This work
20	Pd ₁ ASAC		298 min	K/15	heterogeneous (SAC)	2820	This work
21	Pd ₁ ASAC		323 min	K/15	heterogeneous (SAC)	9820	This work
22	Pd ₁ ASAC		343 min	K/5	heterogeneous (SAC)	46149	This work

TOF = [(mole of converted aryl halides / mole of Pd catalyst) / reaction time].

Supplementary Table 4 | Comparison of Pd content in catalysts before and after reaction.

Catalyst	Pd content (wt.%)		
	Before reaction	After reaction (80 °C)	After reaction (50 °C)
Pd ₁ -TiO ₂	0.23	0.22	0.22
Pd ₁ -Mn ₂ O ₃	0.36	0.16	0.33
		0.14	0.35
Pd ₁ -Fe ₂ O ₃	0.29	0.29	0.28
Pd ₁ -Co ₃ O ₄	0.29	0.29	0.28
		0.30	0.29
Pd ₁ -NiO	0.32	0.19	0.12
		0.21	0.21
Pd ₁ -CuO	0.32	0.08	0.10
		0.09	0.11
Pd ₁ ASAC	0.27	0.27	0.28
	0.93	0.94	0.93

Experimental procedure: Bromobenzene (0.5 mmol), phenylboronic acid (0.6 mmol), K₂CO₃ (1.5 mmol), Pd₁ ASAC (0.07 mol%, according to Pd), ethanol (2 mL), and H₂O (2 mL) were sequentially added to the screw-top reaction tube. The reaction tube was placed in an oil bath preheated to 80 °C, and stirring in an ambient atmosphere. After the reaction, the catalyst was filtered out, and fresh phenylboronic acid and base were added to the filtrate for a subsequent reaction. GC-MS analysis detected 0% conversion of the new substrate, indicating no leaching of Pd ions into the filtrate. This finding was supported by ICP-AES analysis, which showed that the Pd content in the catalyst remained virtually unchanged before and after the reaction, with no detectable Pd ions in the solution

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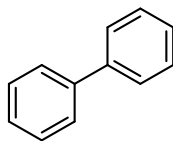
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Supplementary References

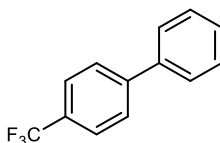
1. Steeples, E., Kelling, A., Schilde, U. & Esposito, D. Amino acid-derived N-heterocyclic carbene palladium complexes for aqueous phase Suzuki-Miyaura couplings. *New J. Chem.* **40**, 4922-4930 (2016).
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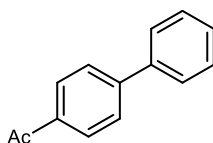
Analysis of the NMR data



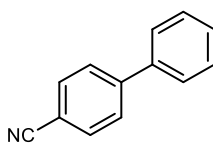
1,1'-biphenyl (4a). The compound was prepared in 99% yield when X=I, Br and Cl. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.65-7.56 (m, 4H), 7.45 (dd, J = 8.4, 7.0 Hz, 4H), 7.39-7.31 (m, 2H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 141.21, 128.72, 127.14.



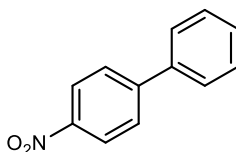
4-(trifluoromethyl)-1,1'-biphenyl (4b). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.70 (s, 4H), 7.62-7.60 (m, 2H), 7.49-7.45 (m, 2H), 7.41-7.33 (m, 1H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 144.71, 139.75, 128.96, 128.73, 128.16, 127.40, 127.26, 127.15, 125.67. ^{19}F NMR (471 MHz, Chloroform-*d*) δ [ppm] -62.39.



1-([1,1'-biphenyl]-4-yl) ethan-1-one (4c). The compound was prepared in 98% yield when X=I and Br. ^1H NMR (400 MHz, Chloroform-*d*) δ [ppm] 8.07-8.01 (m, 2H), 7.72-7.67 (m, 2H), 7.66-7.60 (m, 2H), 7.52-7.44 (m, 2H), 7.44-7.38 (m, 1H), 2.64 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 197.74, 145.75, 139.84, 135.82, 128.93, 128.89, 128.20, 127.24, 127.19, 26.64.

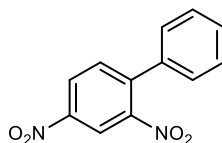


[1,1'-biphenyl]-4-carbonitrile (4d). The compound was prepared in 99% yield when X=I, Br and Cl. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.78-7.71 (m, 2H), 7.71-7.66 (m, 2H), 7.61-7.57 (m, 2H), 7.49 (dd, J = 10.4, 4.8 Hz, 2H), 7.45-7.41 (m, 1H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 145.68, 139.18, 132.59, 129.10, 128.64, 127.73, 127.22, 118.93, 110.91.

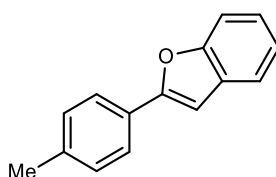


4-nitro-1,1'-biphenyl (4e). The compound was prepared in 99%, 99% and 93% yield when X=I, Br and Cl, respectively. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 8.30 (d, J = 8.9 Hz, 2H), 7.79-7.70 (m, 2H), 7.67-7.59 (m, 2H), 7.55-7.47 (m, 2H), 7.47-7.42 (m, 1H); ^{13}C NMR (125 MHz, Chloroform-

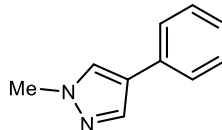
d) δ [ppm] 147.60, 147.05, 138.74, 129.12, 128.88, 127.77, 127.35, 124.07.



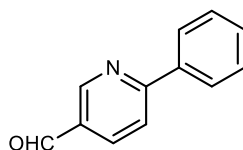
2,4-dinitro-1,1'-biphenyl (4f). The compound was prepared in 99% yield when X=I and Cl. **¹H NMR** (500 MHz, Chloroform-*d*) δ [ppm] 8.74-8.68 (m, 1H), 8.47 (ddd, *J* = 8.5, 2.3, 1.1 Hz, 1H), 7.73-7.64 (m, 1H), 7.53-7.44 (m, 3H), 7.40-7.30 (m, 2H); **¹³C NMR** (125 MHz, Chloroform-*d*) δ [ppm] 149.03, 146.80, 142.21, 135.16, 133.19, 129.52, 129.05, 127.63, 126.44, 119.67.



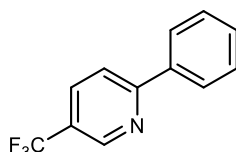
2-(p-tolyl) benzofuran (4g). The compound was prepared in 99% yield when X=I and Br. **¹H NMR** (500 MHz, Chloroform-*d*) δ [ppm] 7.87 (d, *J* = 8.2 Hz, 2H), 7.69-7.60 (m, 2H), 7.41-7.30 (m, 4H), 7.06 (s, 1H), 2.50 (s, 3H); **¹³C NMR** (125 MHz, Chloroform-*d*) δ [ppm] 156.16, 154.74, 138.54, 129.45, 129.32, 127.72, 124.85, 123.96, 122.82, 120.71, 111.05, 100.53, 21.34.



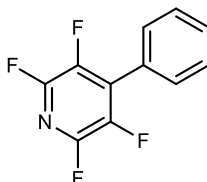
1-methyl-4-phenyl-1H-pyrazole (4h). The compound was prepared in 89% yield. **¹H NMR** (500 MHz, Chloroform-*d*) δ [ppm] 7.76 (d, *J* = 0.8 Hz, 1H), 7.60 (d, *J* = 0.7 Hz, 1H), 7.50 – 7.44 (m, 2H), 7.40 – 7.32 (m, 2H), 7.25 – 7.19 (m, 1H), 3.94 (s, 3H); **¹³C NMR** (125 MHz, Chloroform-*d*) δ [ppm] 136.73, 132.61, 128.82, 126.86, 126.32, 125.48, 123.23, 39.07.



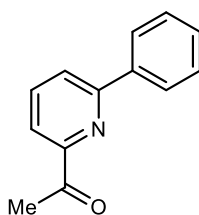
6-phenylnicotinaldehyde (4i). The compound was prepared in 93% yield. **¹H NMR** (500 MHz, Chloroform-*d*) δ [ppm] 10.13 (s, 1H), 9.13 (d, *J* = 2.1 Hz, 1H), 8.22 (dd, *J* = 8.2, 2.2 Hz, 1H), 8.11-8.05 (m, 2H), 7.90 (d, *J* = 8.2 Hz, 1H), 7.55-7.47 (m, 3H); **¹³C NMR** (125 MHz, Chloroform-*d*) δ [ppm] 190.44, 162.18, 152.37, 137.94, 136.50, 130.36, 129.82, 128.97, 127.52, 120.58.



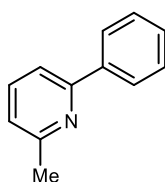
2-phenyl-5-(trifluoromethyl) pyridine (4j). The compound was prepared in 89% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 8.98-8.93 (m, 1H), 8.07-8.01 (m, 2H), 7.98 (dd, $J = 8.3, 2.4$ Hz, 1H), 7.85 (d, $J = 8.3$ Hz, 1H), 7.55-7.45 (m, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 160.66, 146.60, 137.93, 133.92, 130.04, 128.95, 127.25, 124.94, 124.82, 124.67, 122.66, 119.94. ^{19}F NMR (471 MHz, Chloroform-*d*) δ [ppm] -62.24.



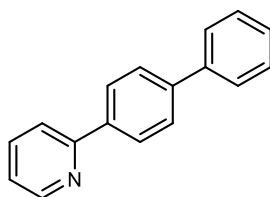
2,3,5,6-tetrafluoro-4-phenylpyridine (4k). The compound was prepared in 92% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.54-7.47 (m, 5H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 145.41, 143.42, 141.30, 139.31, 136.49, 136.26, 134.26, 129.84, 129.82, 127.07. ^{19}F NMR (471 MHz, Chloroform-*d*) δ [ppm] -94.28, -94.34 (d, $J = 6.9$ Hz), -94.39, -145.48 – -145.51 (m), -145.54 – -145.57 (m).



1-(6-phenylpyridin-2-yl) ethan-1-one (4l). The compound was prepared in 98% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 8.10 (dd, $J = 7.2, 1.7$ Hz, 2H), 7.96 (dd, $J = 7.5, 1.2$ Hz, 1H), 7.92-7.82 (m, 2H), 7.51 (dd, $J = 8.3, 6.5$ Hz, 2H), 7.47-7.42 (m, 1H), 2.82 (s, 3H); ^{13}C NMR 200.41, 156.28, 153.23, 138.25, 137.50, 129.35, 128.74, 126.76, 123.30, 119.65, 25.66.

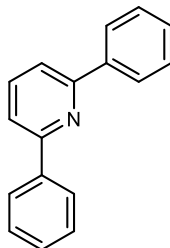


2-methyl-6-phenylpyridine (4m). The compound was prepared in 91% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 8.09-7.95 (m, 2H), 7.62 (t, $J = 7.7$ Hz, 1H), 7.54-7.45 (m, 3H), 7.42 (d, $J = 7.4$ Hz, 1H), 7.09 (d, $J = 7.6$ Hz, 1H), 2.65 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 158.20, 156.82, 139.63, 136.77, 128.59, 128.56, 126.89, 121.48, 117.50, 24.63.

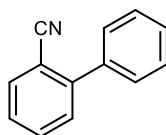


2-([1,1'-biphenyl]-4-yl) pyridine (4n). The compound was prepared in 99% yield. ^1H NMR (500 MHz,

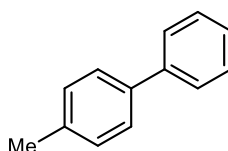
Chloroform-*d*) δ [ppm] 8.81-8.67 (m, 1H), 8.19-8.05 (m, 2H), 7.80-7.74 (m, 3H), 7.74-7.65 (m, 3H), 7.48 (dd, *J* = 8.4, 7.0 Hz, 2H), 7.44-7.35 (m, 1H), 7.27-7.20 (m, 1H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 156.82, 149.54, 141.60, 140.42, 138.06, 136.71, 128.73, 127.43, 127.35, 127.21, 126.98, 122.02, 120.35.



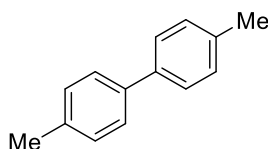
2,6-diphenylpyridine (4o). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 8.23-8.11 (m, 4H), 7.83 (td, *J* = 7.6, 3.0 Hz, 1H), 7.71 (td, *J* = 7.6, 3.5 Hz, 2H), 7.52 (qd, *J* = 7.5, 2.3 Hz, 4H), 7.47-7.41 (m, 2H); ^{13}C NMR 156.81, 139.47, 137.46, 128.95, 128.66, 126.97, 118.62.



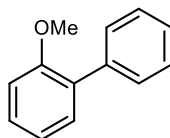
[1,1'-biphenyl]-2-carbonitrile (4p). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.77 (dd, *J* = 7.8, 1.3 Hz, 1H), 7.65 (td, *J* = 7.7, 1.3 Hz, 1H), 7.59 – 7.55 (m, 2H), 7.54 – 7.42 (m, 5H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 145.42, 138.06, 133.69, 132.76, 130.02, 128.69, 128.66, 127.48, 118.67, 111.20.



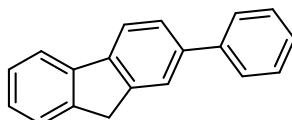
4-methyl-1,1'-biphenyl (4q). The compound was prepared in 99% yield when X=I and Br. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.59 (dd, *J* = 8.2, 1.3 Hz, 2H), 7.50 (d, *J* = 8.1 Hz, 2H), 7.43 (t, *J* = 7.7 Hz, 2H), 7.37-.31 (m, 1H), 7.28-7.24 (m, 2H), 2.41 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 141.13, 138.33, 136.99, 129.45, 128.68, 126.97, 126.94, 21.08.



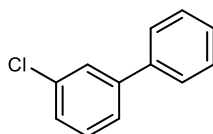
4,4'-dimethyl-1,1'-biphenyl (4r). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.63-7.55 (m, 4H), 7.38-7.30 (m, 4H), 2.49 (s, 6H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 138.24, 136.62, 129.39, 126.76, 21.04.



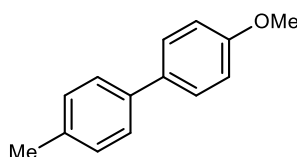
2-methoxy-1,1'-biphenyl (4s). The compound was prepared in 80% yield. ^1H NMR (400 MHz, Chloroform-*d*) δ [ppm] 7.60-7.54 (m, 2H), 7.48-7.41 (m, 2H), 7.39-7.32 (m, 3H), 7.11-6.99 (m, 2H), 3.84 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 156.42, 138.51, 130.86, 130.69, 129.51, 128.57, 127.94, 126.87, 120.79, 111.19, 55.50.



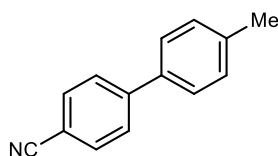
2-phenyl-9H-fluorene (4t). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.93-7.75 (m, 3H), 7.67 (d, J = 8.1 Hz, 2H), 7.63 (d, J = 7.7 Hz, 1H), 7.57 (d, J = 7.4 Hz, 1H), 7.47 (dd, J = 8.3, 7.1 Hz, 2H), 7.43-7.29 (m, 3H), 3.98 (s, 2H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 143.85, 143.43, 141.46, 141.36, 140.89, 139.83, 128.74, 127.15, 127.08, 126.79, 126.70, 125.97, 125.02, 123.77, 120.08, 119.93, 36.98.



3-chloro-1,1'-biphenyl (4u). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.68-7.59 (m, 3H), 7.50 (ddd, J = 9.9, 6.8, 1.8 Hz, 3H), 7.47-7.34 (m, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 142.99, 139.72, 134.59, 129.92, 128.84, 127.80, 127.22, 127.20, 127.04, 125.23.

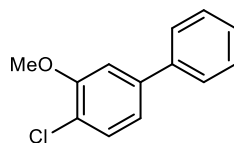


4-methoxy-4'-methyl-1,1'-biphenyl (4v). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.59 (d, J = 8.8 Hz, 2H), 7.53 (d, J = 8.2 Hz, 2H), 7.30 (d, J = 7.5 Hz, 2H), 7.04 (d, J = 8.8 Hz, 2H), 3.90 (s, 3H), 2.46 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 158.86, 137.88, 136.25, 133.64, 129.38, 127.87, 126.50, 114.08, 55.21, 20.99.

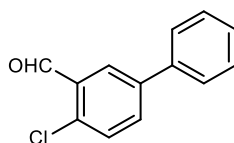


2-methyl-6-(phenylethynyl) pyridine (4w). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.76-7.61 (m, 4H), 7.50 (d, J = 8.1 Hz, 2H), 7.30 (d, J = 8.0 Hz, 2H), 2.42 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 145.48, 138.65, 136.13, 132.45, 129.74, 127.34,

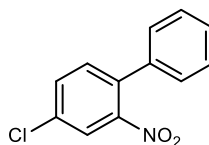
126.95, 118.96, 110.42, 21.10.



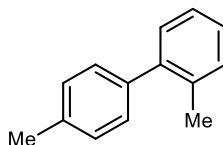
4-chloro-3-methoxy-1,1'-biphenyl (4x). The compound was prepared in 90% yield. **¹H NMR** (400 MHz, Chloroform-*d*) δ [ppm] 7.52-7.47 (m, 2H), 7.45-7.38 (m, 2H), 7.37-7.32 (m, 1H), 7.27-7.23 (m, 1H), 7.03 (dd, *J* = 8.1, 2.0 Hz, 1H), 6.98 (d, *J* = 2.0 Hz, 1H), 3.81 (s, 3H); **¹³C NMR** (125 MHz, Chloroform-*d*) δ [ppm] 156.97, 137.46, 133.88, 131.50, 129.38, 129.22, 128.06, 127.19, 120.81, 111.85, 55.75.



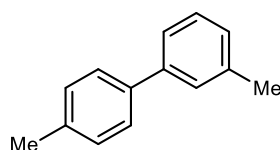
4-chloro-[1,1'-biphenyl]-3-carbaldehyde (4y). The compound was prepared in 75% yield. **¹H NMR** (400 MHz, Chloroform-*d*) δ [ppm] 10.53 (s, 1H), 8.15 (d, *J* = 2.4 Hz, 1H), 7.76 (dd, *J* = 8.3, 2.4 Hz, 1H), 7.63-7.57 (m, 2H), 7.53 (d, *J* = 8.3 Hz, 1H), 7.50-7.43 (m, 2H), 7.43-7.36 (m, 1H); **¹³C NMR** (125 MHz, Chloroform-*d*) δ [ppm] 189.78, 140.52, 138.61, 136.82, 133.47, 132.53, 130.98, 129.04, 128.23, 127.62, 126.93.



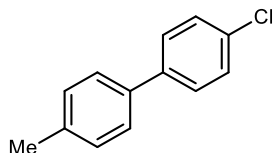
4-chloro-2-nitro-1,1'-biphenyl (4z). The compound was prepared in 81% yield. **¹H NMR** (500 MHz, Chloroform-*d*) δ [ppm] 7.86 (dd, *J* = 3.4, 2.3 Hz, 1H), 7.68 (d, *J* = 8.6 Hz, 1H), 7.60 (dd, *J* = 8.3, 2.2 Hz, 1H), 7.44-7.41 (m, 2H), 7.41-7.38 (m, 1H), 7.30-7.25 (m, 2H); **¹³C NMR** (125 MHz, Chloroform-*d*) δ [ppm] 135.94, 133.31, 132.99, 132.36, 128.79, 128.54, 127.80, 125.77, 124.20.



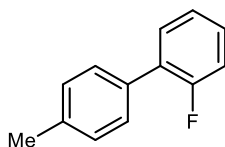
2,4'-dimethyl-1,1'-biphenyl (4aa). The compound was prepared in 99% yield. **¹H NMR** (500 MHz, Chloroform-*d*) δ [ppm] 7.42-7.22 (m, 8H), 2.56-2.43 (m, 3H), 2.42-2.26 (m, 3H); **¹³C NMR** (125 MHz, Chloroform-*d*) δ [ppm] 141.84, 138.99, 136.31, 135.34, 130.23, 129.81, 129.03, 128.73, 127.02, 125.70, 21.14, 20.48.



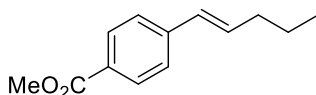
3,4'-dimethyl-1,1'-biphenyl (4ab). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.60 (d, *J* = 8.2 Hz, 2H), 7.50 (d, *J* = 12.4 Hz, 2H), 7.43 (t, *J* = 7.6 Hz, 1H), 7.35 (d, *J* = 7.8 Hz, 2H), 7.26 (t, *J* = 8.1 Hz, 1H), 2.53 (s, 3H), 2.50 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 141.11, 138.42, 138.19, 136.83, 129.37, 128.58, 127.73, 127.67, 126.95, 124.04, 21.51, 21.05.



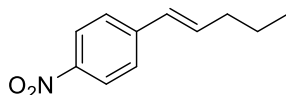
4-chloro-4'-methyl-1,1'-biphenyl (4ac). The compound was prepared in 99% yield when X=I and Br. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.49 (ddd, *J* = 18.6, 8.0, 1.8 Hz, 3H), 7.38 – 7.28 (m, 3H), 7.27 – 7.15 (m, 2H), 2.46 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 139.53, 137.37, 137.04, 132.98, 129.55, 128.80, 128.12, 126.75, 21.06.



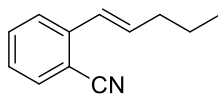
2-fluoro-4'-methyl-1,1'-biphenyl (4ad). The compound was prepared in 99% yield when X=I and Br. ^1H NMR (400 MHz, Chloroform-*d*) δ [ppm] 7.50-7.40 (m, 3H), 7.34-7.24 (m, 3H), 7.23-7.11 (m, 2H), 2.42 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 159.77 (d, *J* = 245.8 Hz), 137.46, 132.86, 130.65 (d, *J* = 3.5 Hz), 129.14, 128.86 (d, *J* = 2.7 Hz), 128.67, 128.60, 124.26 (d, *J* = 3.6 Hz), 116.02 (d, *J* = 22.7 Hz), 21.18. ^{19}F NMR (377 MHz, Chloroform-*d*) δ [ppm] -118.05.



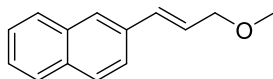
methyl (E)-4-(pent-1-en-1-yl) benzoate (5a). The compound was prepared in 93% yield. ^1H NMR (400 MHz, Chloroform-*d*) δ [ppm] 8.00-7.91 (m, 2H), 7.41-7.36 (m, 2H), 6.46-6.30 (m, 2H), 3.90 (s, 3H), 2.22 (td, *J* = 7.4, 5.9 Hz, 2H), 1.57-1.46 (m, 2H), 0.96 (t, *J* = 7.4 Hz, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 167.01, 142.45, 133.98, 129.86, 129.16, 128.21, 125.73, 51.97, 35.18, 22.33, 13.72.



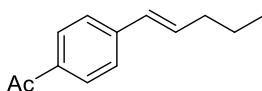
(E)-1-nitro-4-(pent-1-en-1-yl) benzene (5b). The compound was prepared in 93% yield. ^1H NMR (400 MHz, Chloroform-*d*) δ [ppm] 8.20-8.10 (m, 2H), 7.47-7.42 (m, 2H), 6.50-6.36 (m, 2H), 2.32-2.15 (m, 2H), 1.58-1.47 (m, 2H), 0.97 (t, *J* = 7.4 Hz, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 146.38, 144.42, 136.40, 128.24, 126.30, 123.94, 35.22, 22.16, 13.70.



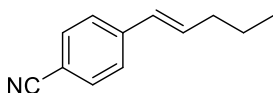
(E)-2-(pent-1-en-1-yl) benzonitrile (5c). The compound was prepared in 88% yield. ¹H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.64-7.56 (m, 2H), 7.53-7.48 (m, 1H), 7.28-7.24 (m, 1H), 6.79-6.71 (m, 1H), 6.50-6.40 (m, 1H), 2.27 (ddd, *J* = 14.6, 7.2, 1.5 Hz, 2H), 1.56-1.50 (m, 2H), 0.98 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 141.24, 136.60, 132.87, 132.56, 126.87, 126.08, 125.36, 118.11, 110.47, 35.19, 22.23, 13.68.



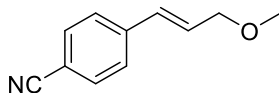
(E)-2-(3-methoxyprop-1-en-1-yl) naphthalene (5d). The compound was prepared in 50% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ [ppm] 7.84-7.77 (m, 3H), 7.75 (s, 1H), 7.62 (dd, *J* = 8.6, 1.7 Hz, 1H), 7.53-7.40 (m, 2H), 6.79 (d, *J* = 16.0 Hz, 1H), 6.42 (dt, *J* = 15.9, 6.0 Hz, 1H), 4.16 (dd, *J* = 6.0, 1.4 Hz, 2H), 3.43 (s, 3H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 134.16, 133.54, 133.01, 132.49, 128.17, 127.96, 127.63, 126.44, 126.32, 126.23, 125.87, 123.55, 73.14, 58.04.



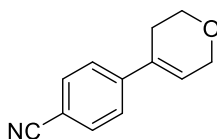
(E)-1-(4-(pent-1-en-1-yl) phenyl) ethan-1-one (5e). The compound was prepared in 99% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ [ppm] 7.94 – 7.84 (m, 2H), 7.45 – 7.37 (m, 2H), 6.47 – 6.25 (m, 2H), 2.58 (s, 3H), 2.22 (td, *J* = 7.4, 5.8 Hz, 2H), 1.52 (dq, *J* = 14.6, 7.4 Hz, 2H), 0.96 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 197.61, 142.66, 135.40, 134.31, 129.09, 128.73, 125.91, 35.21, 26.53, 22.31, 13.73.



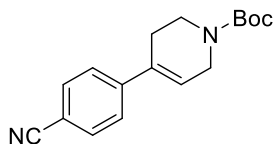
(E)-4-(pent-1-en-1-yl) benzonitrile (5f). The compound was prepared in 70% yield. ¹H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.61-7.51 (m, 2H), 7.46-7.34 (m, 2H), 6.45-6.29 (m, 2H), 2.26-2.19 (m, 2H), 1.57-1.45 (m, 2H), 0.96 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 142.38, 135.30, 132.28, 128.55, 126.33, 119.12, 109.87, 35.11, 22.19, 13.67.



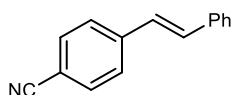
(E)-4-(3-methoxyprop-1-en-1-yl) benzonitrile (5g). The compound was prepared in 81% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ [ppm] 7.62-7.54 (m, 2H), 7.47-7.42 (m, 2H), 6.68-6.54 (m, 1H), 6.39 (dt, *J* = 16.0, 5.5 Hz, 1H), 4.11 (dd, *J* = 5.5, 1.6 Hz, 2H), 3.40 (s, 3H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 141.20, 132.34, 130.13, 129.93, 126.83, 118.86, 110.74, 72.44, 58.31.



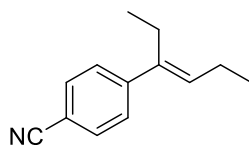
4-(3,6-dihydro-2H-pyran-4-yl) benzonitrile (5h). The compound was prepared in 87% yield. ¹H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.65-7.56 (m, 2H), 7.50-7.43 (m, 2H), 6.27 (dq, *J* = 4.5, 1.5 Hz, 1H), 4.34 (q, *J* = 2.8 Hz, 2H), 3.93 (t, *J* = 5.4 Hz, 2H), 2.50 (ddq, *J* = 5.4, 4.5, 2.7 Hz, 2H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 144.44, 132.89, 132.24, 125.89, 125.16, 118.87, 110.61, 65.71, 64.12, 26.75.



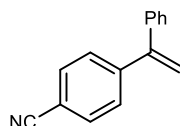
tert-butyl 4-(4-cyanophenyl)-3,6-dihydropyridine-1(2H)-carboxylate (5i). The compound was prepared in 99% yield. ¹H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.63-7.54 (m, 2H), 7.44 (d, *J* = 8.4 Hz, 2H), 6.15 (s, 1H), 4.30-3.95 (m, 2H), 3.63 (t, *J* = 5.7 Hz, 2H), 2.64-2.36 (m, 2H), 1.47 (s, 9H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 154.61, 144.87, 134.11, 132.18, 125.37, 124.19, , 118.80, 110.53, 79.83, 43.60, 28.36, 26.97.



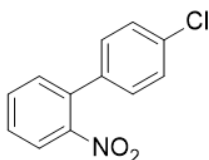
(E)-4-styrylbenzonitrile (5j). The compound was prepared in 99% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ [ppm] 7.67-7.60 (m, 2H), 7.60-7.51 (m, 4H), 7.43-7.36 (m, 2H), 7.36-7.29 (m, 1H), 7.21 (d, *J* = 16.3 Hz, 1H), 7.09 (d, *J* = 16.3 Hz, 1H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 141.75, 136.21, 132.41, 132.33, 128.79, 128.58, 126.85, 126.79, 126.64, 118.97, 110.48.



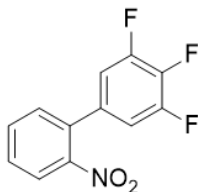
(E)-4-(hex-3-en-3-yl) benzonitrile (5k). The compound was prepared in 99% yield. ¹H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.62 – 7.51 (m, 2H), 7.47 – 7.39 (m, 2H), 5.74 (t, *J* = 7.3 Hz, 1H), 2.51 (q, *J* = 7.5 Hz, 2H), 2.23 (p, *J* = 7.5 Hz, 2H), 1.07 (t, *J* = 7.5 Hz, 3H), 0.97 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 147.64, 139.71, 133.11, 132.00, 126.74, 119.16, 109.79, 22.42, 21.81, 14.16, 13.51..



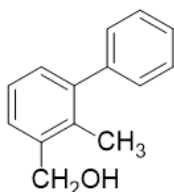
4-(1-phenylvinyl) benzonitrile (5l). The compound was prepared in 99% yield. ¹H NMR (400 MHz, Chloroform-*d*) δ [ppm] 7.66-7.59 (m, 2H), 7.48-7.41 (m, 2H), 7.40-7.32 (m, 3H), 7.32-7.24 (m, 2H), 5.57 (dd, *J* = 17.3, 0.7 Hz, 2H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 148.64, 146.02, 140.14, 132.02, 128.79, 128.41, 128.21, 128.10, 118.81, 116.68, 111.28.



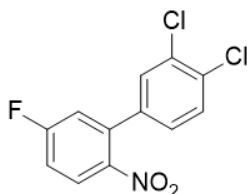
4'-chloro-2-nitro-1,1'-biphenyl (4ae). ^1H NMR (500 MHz, CDCl_3) δ 7.91 (dd, $J = 8.1, 1.3$ Hz, 1H), 7.65 (td, $J = 7.5, 1.3$ Hz, 1H), 7.53 (ddd, $J = 8.0, 7.4, 1.5$ Hz, 1H), 7.47 – 7.35 (m, 3H), 7.28 (d, $J = 8.3$ Hz, 2H). ^{13}C NMR (126 MHz, CDCl_3) δ 149.09, 135.94, 135.23, 134.48, 132.53, 131.87, 129.30, 128.93, 128.60, 124.29.



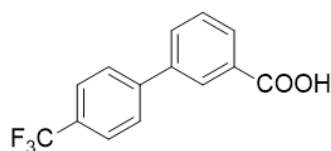
3',4',5'-trifluoro-2-nitro-1,1'-biphenyl (4af). ^1H NMR (500 MHz, CDCl_3) δ 7.96 (dd, $J = 8.1, 1.3$ Hz, 1H), 7.69 (td, $J = 7.6, 1.3$ Hz, 1H), 7.62 – 7.56 (m, 1H), 7.41 (dd, $J = 7.6, 1.5$ Hz, 1H), 7.04 – 6.92 (m, 2H). ^{13}C NMR (126 MHz, CDCl_3) δ 151.14 (ddd, $J = 251.3, 10.1, 4.2$ Hz), 148.72, 139.78 (td, $J = 251.3, 15.0$ Hz), 133.54, 132.82, 131.67, 129.41, 124.54, 112.61 (d, $J = 22.5$ Hz), 112.61 (d, $J = 11.2$ Hz). ^{19}F NMR (471 MHz, CDCl_3) δ -133.43 (d, $J = 20.1$ Hz), -160.54 (t, $J = 20.8$ Hz).



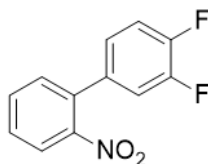
(2-methyl-[1,1'-biphenyl]-3-yl) methanol (4ag). ^1H NMR (500 MHz, CDCl_3) δ 7.48 – 7.42 (m, 3H), 7.40 – 7.36 (m, 1H), 7.35 – 7.27 (m, 3H), 7.24 (dd, $J = 7.6, 1.6$ Hz, 1H), 4.80 (s, 2H), 2.28 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 142.89, 142.07, 139.25, 133.62, 129.61, 129.40, 128.09, 126.83, 126.78, 125.62, 64.09, 15.91.



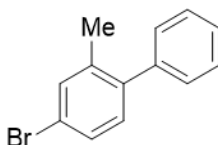
3',4'-dichloro-5-fluoro-2-nitro-1,1'-biphenyl (4ah). ^1H NMR (500 MHz, CDCl_3) δ 8.04 (dd, $J = 9.0, 5.0$ Hz, 1H), 7.53 (d, $J = 8.2$ Hz, 1H), 7.44 (d, $J = 2.1$ Hz, 1H), 7.24 (ddd, $J = 9.0, 7.3, 2.7$ Hz, 1H), 7.15 (dd, $J = 8.2, 2.1$ Hz, 1H), 7.12 (dd, $J = 8.5, 2.8$ Hz, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 164.10 (d, $J = 258$ Hz), 144.74, 137.32 (d, $J = 10$ Hz), 136.55, 133.14 (d, $J = 22.5$ Hz), 130.73, 129.69, 127.43 (d, $J = 8.8$ Hz), 127.13, 118.89 (d, $J = 23.8$ Hz), 116.02 (d, $J = 23.2$ Hz). ^{19}F NMR (471 MHz, CDCl_3) δ -103.57.



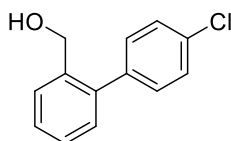
4'-(trifluoromethyl)-[1,1'-biphenyl]-3-carboxylic acid (4ai). ^1H NMR (500 MHz, Methanol- d_4) δ 8.34 (t, J = 1.8 Hz, 1H), 8.09 (dt, J = 7.8, 1.4 Hz, 1H), 7.94 (ddd, J = 7.8, 2.0, 1.1 Hz, 1H), 7.92 – 7.86 (m, 2H), 7.80 (m, 2H), 7.63 (t, J = 7.7 Hz, 1H). ^{13}C NMR (126 MHz, Methanol- d_4) δ 168.03, 143.82, 139.76, 135.44, 131.45, 131.28, 129.39 (q, J = 32.5 Hz), 129.09, 128.95, 127.93, 127.28, 125.53 (q, J = 3.8 Hz), 124.37 (q, J = 270 Hz). ^{19}F NMR (471 MHz, CDCl_3) δ -64.02.



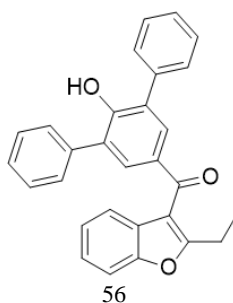
3',4'-difluoro-2-nitro-1,1'-biphenyl (4aj). ^1H NMR (500 MHz, CDCl_3) δ 7.92 (dd, J = 8.1, 1.3 Hz, 1H), 7.66 (td, J = 7.5, 1.3 Hz, 1H), 7.55 (td, J = 7.8, 1.4 Hz, 1H), 7.43 (dd, J = 7.8, 1.4 Hz, 1H), 7.30 – 7.15 (m, 2H), 7.07 – 7.04 (m, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 150.44 (dd, J = 248.8, 12.5 Hz), 150.16 (dd, J = 247.5, 12.5 Hz), 149.00, 134.35 (d, J = 3.8 Hz), 134.29, 132.62, 131.85, 128.93, 124.34, 124.29 (d, J = 3.8 Hz), 117.63 (d, 17.5 Hz), 117.34 (d, 17.5 Hz). ^{19}F NMR (471 MHz, CDCl_3) δ -136.93 (d, J = 21.0 Hz), -138.15 (d, J = 21.6 Hz).



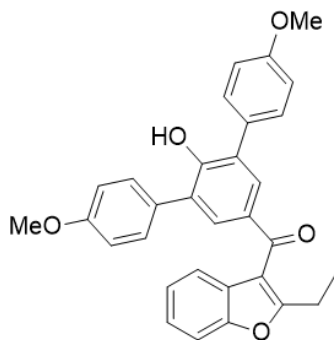
4-bromo-2-methyl-1,1'-biphenyl (4ak). ^1H NMR (500 MHz, CDCl_3) δ 7.48 – 7.43 (m, 3H), 7.42 – 7.37 (m, 2H), 7.34 – 7.30 (m, 2H), 7.13 (d, J = 8.1 Hz, 1H), 2.28 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 140.89, 140.79, 137.67, 133.02, 131.33, 129.04, 128.82, 128.24, 127.15, 121.09, 20.34.



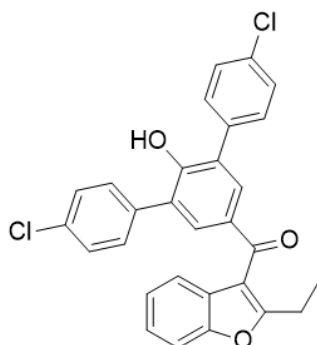
(4'-chloro-[1,1'-biphenyl]-2-yl) methanol (4al). ^1H NMR (400 MHz, Chloroform- d) δ [ppm] 7.57-7.53 (m, 1H), 7.43-7.34 (m, 4H), 7.34-7.29 (m, 2H), 7.28-7.23 (m, 1H), 4.57 (s, 2H), 1.94 (s, 1H); ^{13}C NMR (125 MHz, Chloroform- d) δ [ppm] 140.08, 138.98, 137.82, 133.31, 130.45, 129.91, 128.58, 128.35, 127.98, 127.76, 62.90.



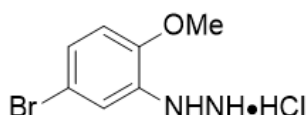
(2-ethylbenzofuran-3-yl) (2'-hydroxy-[1,1':3',1''-terphenyl]-5'-yl) methanone (4am). ¹H NMR (500 MHz, CDCl₃) δ 7.88 (s, 2H), 7.61 – 7.57 (m, 5H), 7.54 – 7.48 (m, 5H), 7.45 – 7.41 (m, 2H), 7.35 – 7.26 (m, 2H), 6.00 (s, 1H), 3.02 (q, J = 7.5 Hz, 2H), 1.38 (t, J = 7.5 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 190.49, 165.66, 153.65, 136.46, 131.95, 131.71, 129.32, 129.06, 128.92, 128.75, 128.20, 127.21, 124.37, 123.44, 121.32, 116.15, 111.06, 21.88, 12.44.



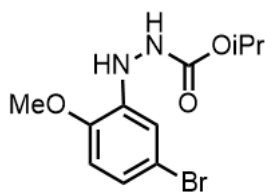
(2-ethylbenzofuran-3-yl) (2'-hydroxy-4,4''-dimethoxy-[1,1':3',1''-terphenyl]-5'-yl) methanone (4an). ¹H NMR (500 MHz, CDCl₃) δ 7.83 (s, 2H), 7.62 – 7.58 (m, 1H), 7.54 – 7.49 (m, 4H), 7.39 – 7.25 (m, 2H), 7.08 – 7.00 (m, 4H), 6.03 (s, 1H), 3.86 (s, 6H), 3.01 (q, J = 7.6 Hz, 2H), 1.38 (t, J = 7.5 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 190.66, 165.59, 159.50, 153.88, 153.70, 131.80, 131.29, 130.51, 128.68, 128.52, 127.26, 124.35, 123.42, 121.35, 116.20, 116.09, 114.48, 111.05, 55.36, 21.87, 12.44.



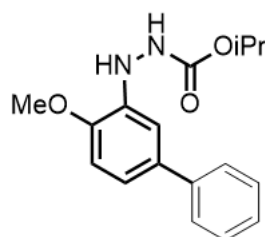
(4,4''-dichloro-2'-hydroxy-[1,1':3',1''-terphenyl]-5'-yl) (2-ethylbenzofuran-3-yl) methanone (4ao). ¹H NMR (500 MHz, CDCl₃) δ 7.83 (s, 2H), 7.53 – 7.43 (m, 10H), 7.32 (ddd, J = 8.5, 7.3, 1.4 Hz, 1H), 7.28 – 7.23 (m, 1H), 3.00 (q, J = 7.5 Hz, 2H), 1.36 (t, J = 7.5 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 190.16, 165.98, 153.71, 153.38, 134.67, 134.42, 132.16, 131.80, 130.66, 129.30, 127.92, 127.05, 124.47, 123.46, 121.17, 115.96, 111.18, 21.86, 12.40.



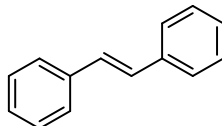
(5-bromo-2-methoxyphenyl) hydrazine. ¹H NMR (500 MHz, Methanol-d₄) δ 7.31 – 7.16 (m, 2H), 6.98 (d, J = 8.6 Hz, 1H), 3.92 (s, 3H). ¹³C NMR (126 MHz, Methanol-d₄) δ 148.22, 134.99, 126.04, 117.85, 112.46, 55.23.



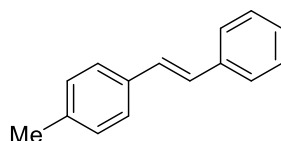
isopropyl 2-(5-bromo-2-methoxyphenyl) hydrazine-1-carboxylate (1aa). ^1H NMR (500 MHz, CDCl_3) δ 6.98 (d, J = 2.4 Hz, 1H), 6.94 (dd, J = 8.5, 2.4 Hz, 1H), 6.67 (d, J = 8.5 Hz, 1H), 6.44 (s, 1H), 6.25 (s, 1H), 4.99 (p, J = 6.2 Hz, 1H), 3.85 (s, 3H), 1.42 – 1.04 (m, 6H). ^{13}C NMR (126 MHz, CDCl_3) δ 156.50, 145.96, 138.91, 122.69, 114.95, 113.60, 111.52, 69.86, 55.77, 22.03.



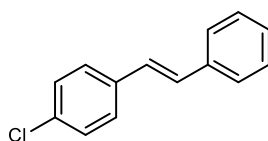
isopropyl 2-(4-methoxy-[1,1'-biphenyl]-3-yl) hydrazine-1-carboxylate (bifenazate). ^1H NMR (500 MHz, CDCl_3) δ 7.58 – 7.54 (m, 2H), 7.42 (dd, J = 8.5, 7.0 Hz, 2H), 7.35 – 7.30 (m, 1H), 7.12 (d, J = 2.2 Hz, 1H), 7.10 (dd, J = 8.2, 2.3 Hz, 1H), 6.91 (d, J = 8.3 Hz, 1H), 6.46 (s, 1H), 6.32 (s, 1H), 4.99 (p, J = 6.2 Hz, 1H), 3.92 (s, 3H), 1.41 – 1.10 (m, 6H). ^{13}C NMR (126 MHz, CDCl_3) δ 156.72, 146.64, 141.40, 137.75, 134.36, 128.63, 126.92, 126.67, 119.17, 111.11, 110.49, 69.62, 55.75, 22.06.



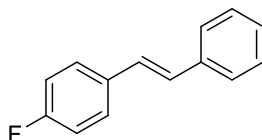
(E)-1,2-diphenylethene (8a). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform- d) δ [ppm] 7.54 (dd, J = 8.2, 1.3 Hz, 4H), 7.38 (dd, J = 8.5, 6.9 Hz, 4H), 7.30-7.26 (m, 2H), 7.13 (s, 2H); ^{13}C NMR (125 MHz, Chloroform- d) δ [ppm] 137.29, 128.65, 127.59, 126.48.



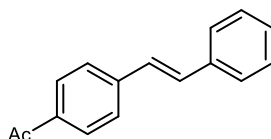
(E)-1-methyl-4-styrylbenzene (8b). The compound was prepared in 97% and 92% yield when $X=\text{I}$ and Br , respectively. ^1H NMR (400 MHz, Chloroform- d) δ [ppm] 7.55-7.50 (m, 2H), 7.47-7.41 (m, 2H), 7.41-7.35 (m, 2H), 7.30-7.24 (m, 1H), 7.20 (d, J = 7.9 Hz, 2H), 7.16-7.05 (m, 2H), 2.39 (s, 3H); ^{13}C NMR (125 MHz, Chloroform- d) δ [ppm] 137.49, 134.52, 129.37, 128.62, 128.60, 127.67, 127.38, 126.41, 126.37, 21.23.



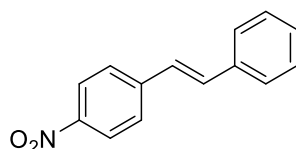
(E)-1-chloro-4-styrylbenzene (8c). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.56-7.50 (m, 2H), 7.45 (d, J = 8.5 Hz, 2H), 7.39 (dd, J = 8.4, 6.9 Hz, 2H), 7.36-7.32 (m, 2H), 7.32-7.28 (m, 1H), 7.12-7.04 (m, 2H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 136.92, 135.78, 133.10, 129.25, 128.78, 128.68, 127.81, 127.61, 127.29, 126.50.



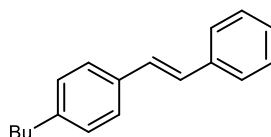
(E)-1-fluoro-4-styrylbenzene (8d). The compound was prepared in 85% yield. ^1H NMR (400 MHz, Chloroform-*d*) δ [ppm] 7.55-7.46 (m, 4H), 7.42-7.34 (m, 2H), 7.32-7.26 (m, 1H), 7.13-6.98 (m, 4H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 162.31 (d, J = 245.7 Hz), 137.14, 133.48 (d, J = 3.2 Hz), 128.68, 128.46 (d, J = 2.1 Hz), 127.96 (d, J = 7.9 Hz), 127.64, 127.44, 126.42, 115.59 (d, J = 21.5 Hz). ^{19}F NMR (377 MHz, Chloroform-*d*) δ [ppm] -114.20.



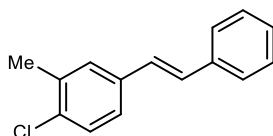
(E)-1-(4-styrylphenyl) ethan-1-one (8e). The compound was prepared in 80% yield. ^1H NMR (400 MHz, Chloroform-*d*) δ 7.98-7.93 (m, 2H), 7.61-7.50 (m, 4H), 7.42-7.34 (m, 2H), 7.33-7.28 (m, 1H), 7.23 (d, J = 16.4 Hz, 1H), 7.13 (d, J = 16.4 Hz, 1H), 2.61 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 197.48, 141.99, 136.67, 135.92, 131.45, 128.86, 128.78, 128.30, 127.42, 126.80, 126.48, 26.57.



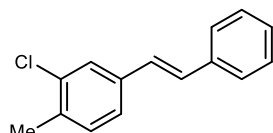
(E)-1-nitro-4-styrylbenzene (8f). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 8.28-8.12 (m, 2H), 7.68-7.59 (m, 2H), 7.59-7.51 (m, 2H), 7.45-7.37 (m, 2H), 7.37-7.31 (m, 1H), 7.27 (d, J = 16.4 Hz, 1H), 7.14 (d, J = 16.3 Hz, 1H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 146.71, 143.80, 136.13, 133.26, 128.86, 128.81, 126.98, 126.81, 126.23, 124.10.



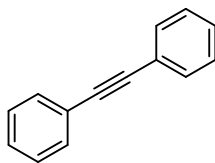
(E)-1-butyl-4-styrylbenzene (8g). The compound was prepared in 53% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.55-7.47 (m, 2H), 7.43 (d, J = 8.1 Hz, 2H), 7.39-7.30 (m, 2H), 7.28-7.22 (m, 1H), 7.18 (d, J = 8.1 Hz, 2H), 7.13-7.04 (m, 2H), 2.67-2.58 (m, 2H), 1.63 (ddd, J = 15.3, 11.0, 7.5 Hz, 2H), 1.39 (dq, J = 14.7, 7.4 Hz, 2H), 0.95 (t, J = 7.4 Hz, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 142.62, 137.66, 134.87, 128.76, 128.64, 127.84, 127.38, 126.45, 126.42, 35.42, 33.53, 22.34, 13.90.



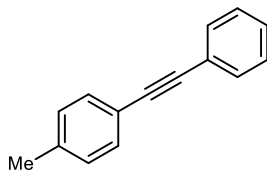
(E)-1-chloro-2-methyl-4-styrylbenzene (8h). The compound was prepared in 60% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.56-7.47 (m, 3H), 7.37 (t, J = 7.6 Hz, 2H), 7.32-7.24 (m, 2H), 7.21 (d, J = 7.8 Hz, 1H), 7.04 (q, J = 16.3 Hz, 2H), 2.38 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 137.00, 136.71, 135.20, 134.69, 131.08, 129.07, 128.70, 127.79, 127.18, 126.77, 126.52, 124.72, 19.83.



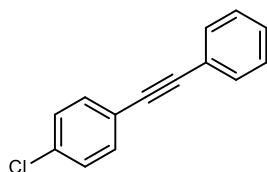
(E)-2-chloro-1-methyl-4-styrylbenzene (8i). The compound was prepared in 50% yield. ^1H NMR (400 MHz, Chloroform-*d*) δ [ppm] 7.54-7.48 (m, 3H), 7.42-7.33 (m, 2H), 7.32-7.26 (m, 2H), 7.23-7.19 (m, 1H), 7.11-6.98 (m, 2H), 2.39 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 136.99, 136.70, 135.19, 134.68, 131.07, 129.06, 128.69, 127.78, 127.16, 126.76, 126.51, 124.71, 19.82.



1,2-diphenylethyne (9a). The compound was prepared in 98% and 68% yield when X=I and Br, respectively. ^1H NMR (400 MHz, Chloroform-*d*) δ [ppm] 7.60-7.50 (m, 2H), 7.43-7.30 (m, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 131.59, 128.32, 128.23, 123.25, 89.35.

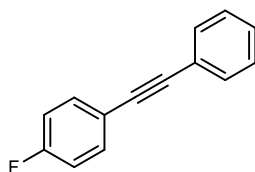


1-methyl-4-(phenylethynyl) benzene (9b). The compound was prepared in 97% and 57% yield when X=I and Br, respectively. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.55-7.51 (m, 2H), 7.43 (d, J = 8.1 Hz, 2H), 7.37-7.30 (m, 3H), 7.16 (d, J = 7.9 Hz, 2H), 2.37 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 138.38, 131.53, 131.48, 129.10, 128.30, 128.06, 123.46, 120.17, 89.53, 88.69, 21.50.

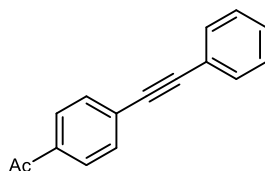


1-chloro-4-(phenylethynyl) benzene (9c). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.59-7.50 (m, 2H), 7.50-7.44 (m, 2H), 7.40-7.29 (m, 5H); ^{13}C NMR (125

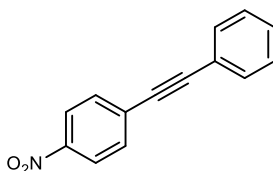
MHz, Chloroform-*d*) δ [ppm] 134.23, 132.78, 131.57, 128.66, 128.46, 128.37, 122.90, 121.76, 90.30, 88.22.



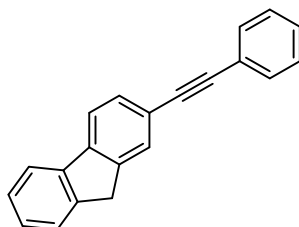
1-fluoro-4-(phenylethynyl)benzene (9d). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 7.57-7.50 (m, 4H), 7.39-7.32 (m, 3H), 7.09-7.02 (m, 2H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 162.48 (d, J = 248.1 Hz), 133.46 (d, J = 8.3 Hz), 131.54, 128.36, 128.32, 123.07, 119.36 (d, J = 3.3 Hz), 115.63 (d, J = 22.0 Hz), 89.02, 88.26. ^{19}F NMR (377 MHz, Chloroform-*d*) δ [ppm] -110.94.



1-(4-(phenylethynyl)phenyl)ethan-1-one (9e). The compound was prepared in 99% and 56% yield when $X=\text{I}$ and Br , respectively. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 8.01-7.88 (m, 2H), 7.65-7.58 (m, 2H), 7.58-7.53 (m, 2H), 7.41-7.33 (m, 3H), 2.61 (s, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 197.28, 136.14, 131.70, 131.65, 128.78, 128.41, 128.23, 128.16, 122.61, 92.68, 88.57, 26.59.



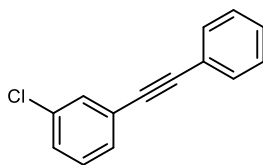
1-nitro-4-(phenylethynyl)benzene (9f). The compound was prepared in 99% yield. ^1H NMR (500 MHz, Chloroform-*d*) δ [ppm] 8.27-8.17 (m, 2H), 7.71-7.63 (m, 2H), 7.61-7.51 (m, 2H), 7.44-7.34 (m, 3H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 146.95, 132.24, 131.82, 130.24, 129.26, 128.52, 123.61, 122.07, 94.68, 87.52.



2-(phenylethynyl)-9H-fluorene (9g). The compound was prepared in 42% yield. ^1H NMR (400 MHz, Chloroform-*d*) δ [ppm] 7.81-7.74 (m, 2H), 7.74-7.70 (m, 1H), 7.62-7.51 (m, 4H), 7.43-7.30 (m, 5H), 3.92 (s, 2H); ^{13}C NMR (125 MHz, Chloroform-*d*) δ [ppm] 143.56, 143.19, 141.88, 141.09, 131.56, 130.50, 128.35, 128.16, 128.13, 127.15, 126.91, 125.09, 123.45, 121.27, 120.18, 119.78, 90.15, 89.37,

1022 36.74.

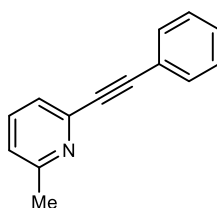
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1025 **1-chloro-3-(phenylethynyl) benzene (9h).** The compound was prepared in 56% yield. ¹H NMR (500
1026 MHz, Chloroform-*d*) δ [ppm] 7.55-7.50 (m, 3H), 7.41 (dt, *J* = 7.3, 1.4 Hz, 1H), 7.39-7.33 (m, 3H), 7.33-
1027 7.27 (m, 2H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 134.22, 131.69, 131.47, 129.71, 129.54,
1028 128.60, 128.49, 128.40, 125.08, 122.82, 90.57, 87.93.

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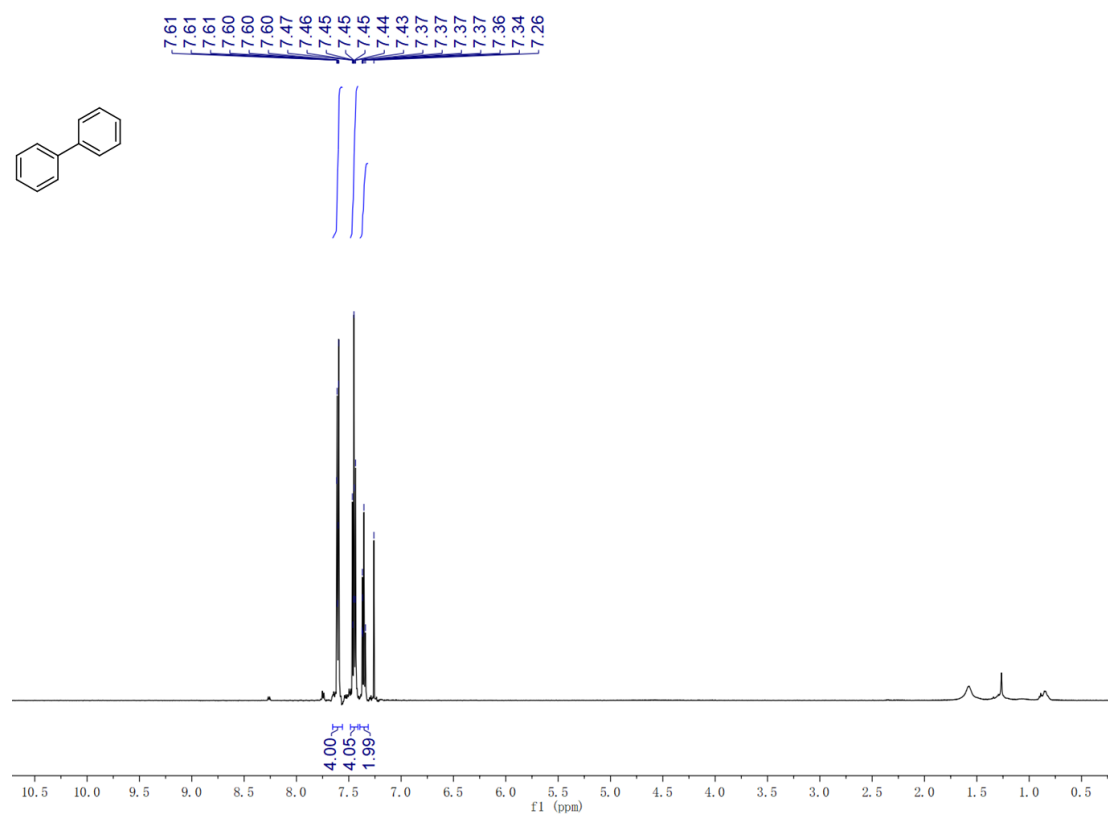
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1031 **2-methyl-6-(phenylethynyl) pyridine (9i).** The compound was prepared in 59% yield. ¹H NMR (500
1032 MHz, Chloroform-*d*) δ [ppm] 8.01-7.88 (m, 2H), 7.65-7.58 (m, 2H), 7.58-7.53 (m, 2H), 7.41-7.33 (m,
1033 3H), 2.61 (s, 3H); ¹³C NMR (125 MHz, Chloroform-*d*) δ [ppm] 158.85, 142.50, 136.61, 132.09, 128.90,
1034 128.32, 124.45, 122.69, 122.33, 89.32, 88.51, 24.42.

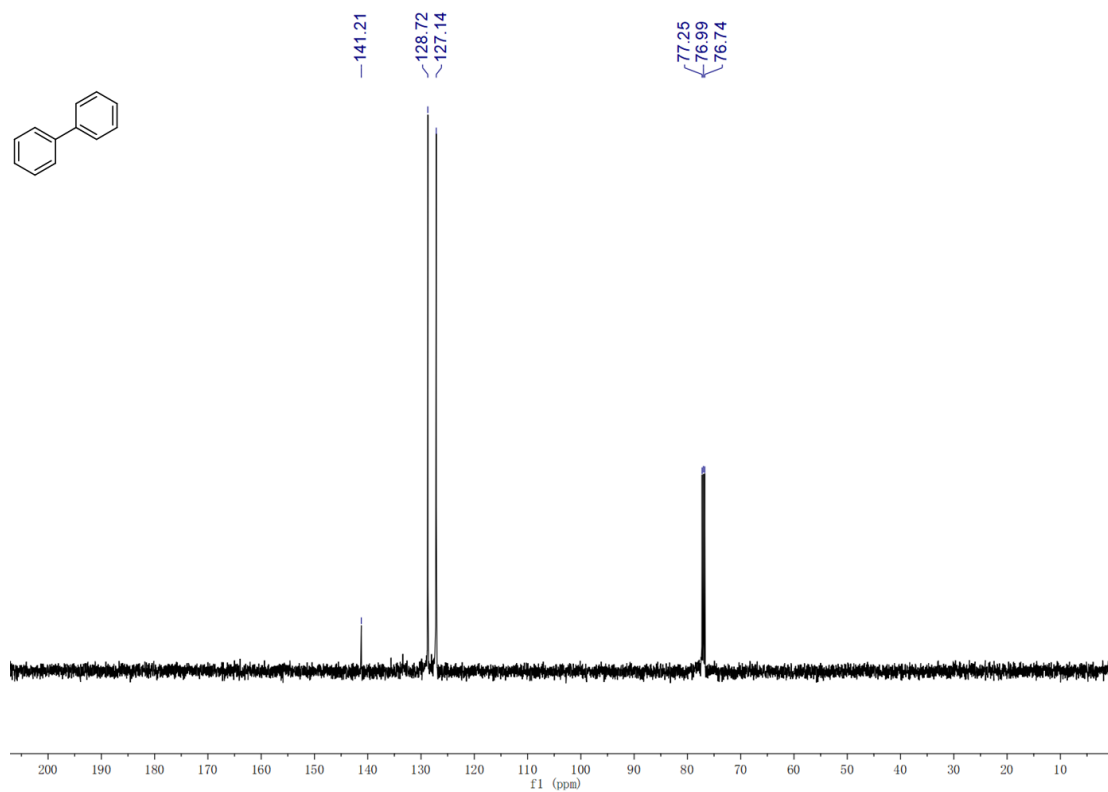
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1036 ¹H and ¹³C-NMR spectra of product 4a.

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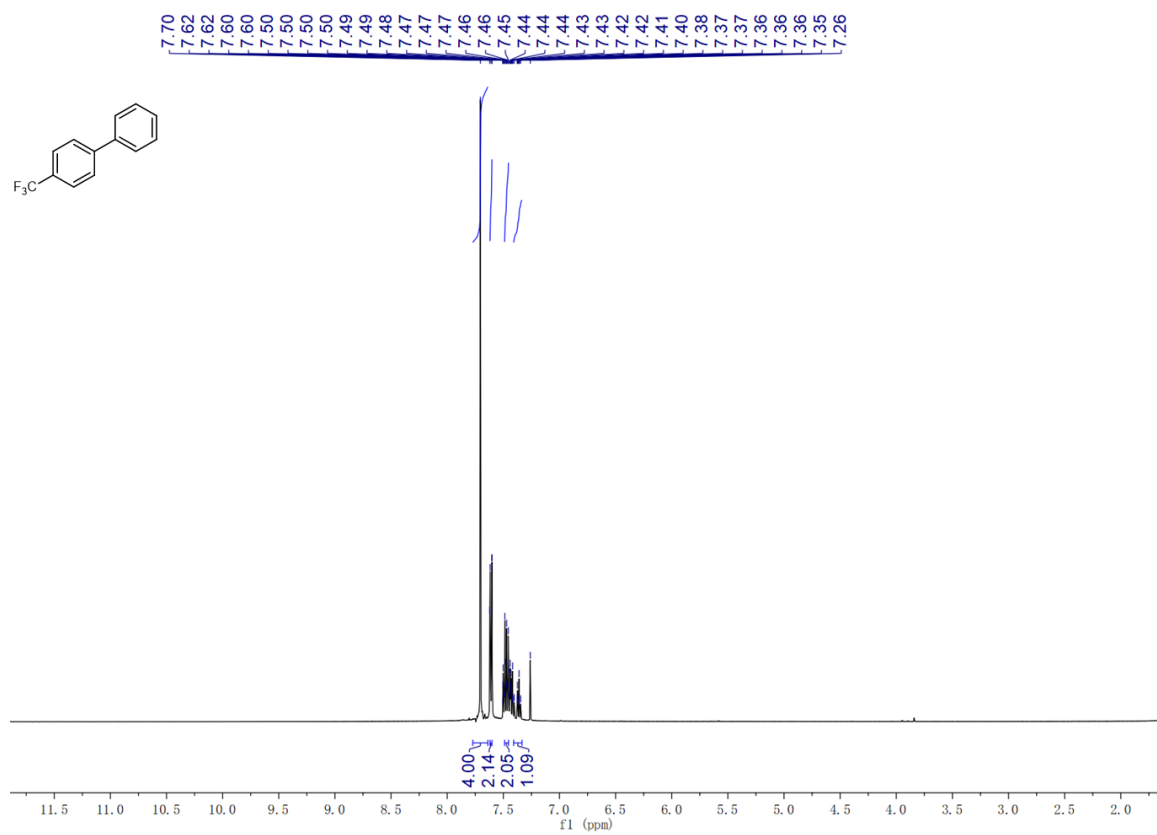
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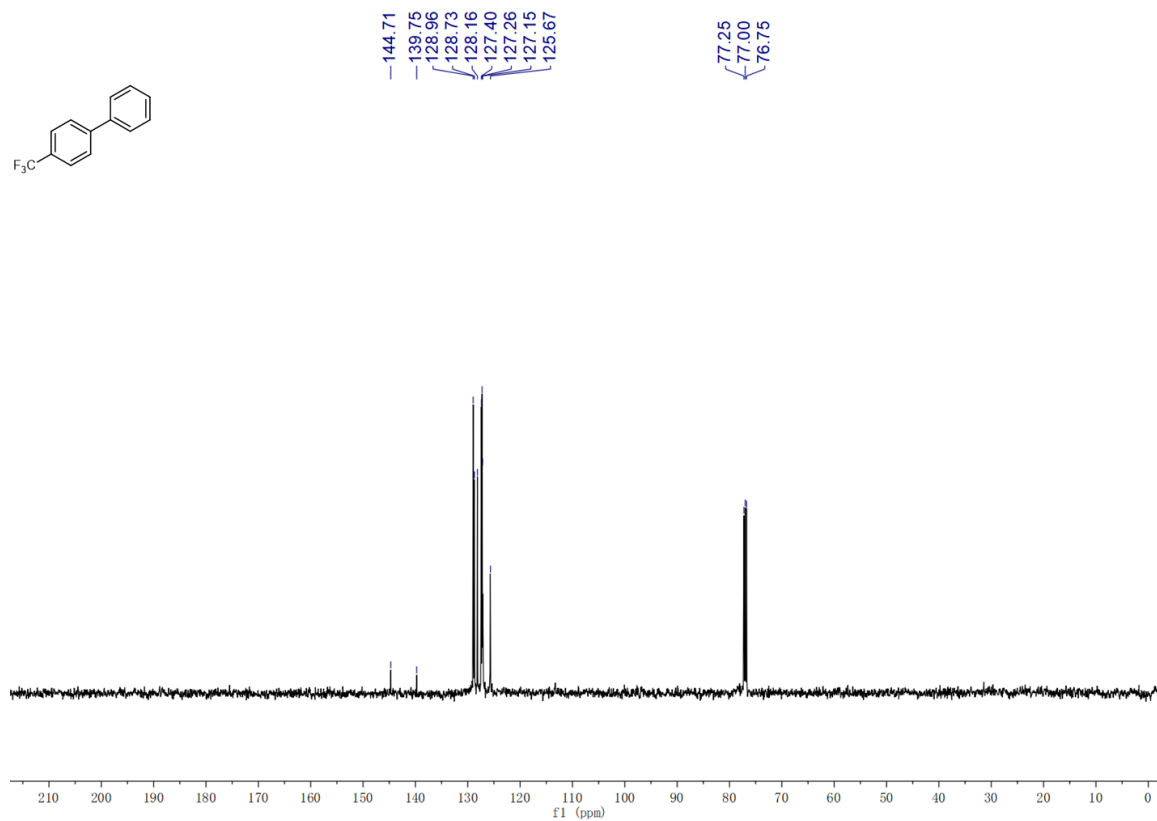
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1042 ¹H and ¹³C, ¹⁹F-NMR spectra of product 4b.

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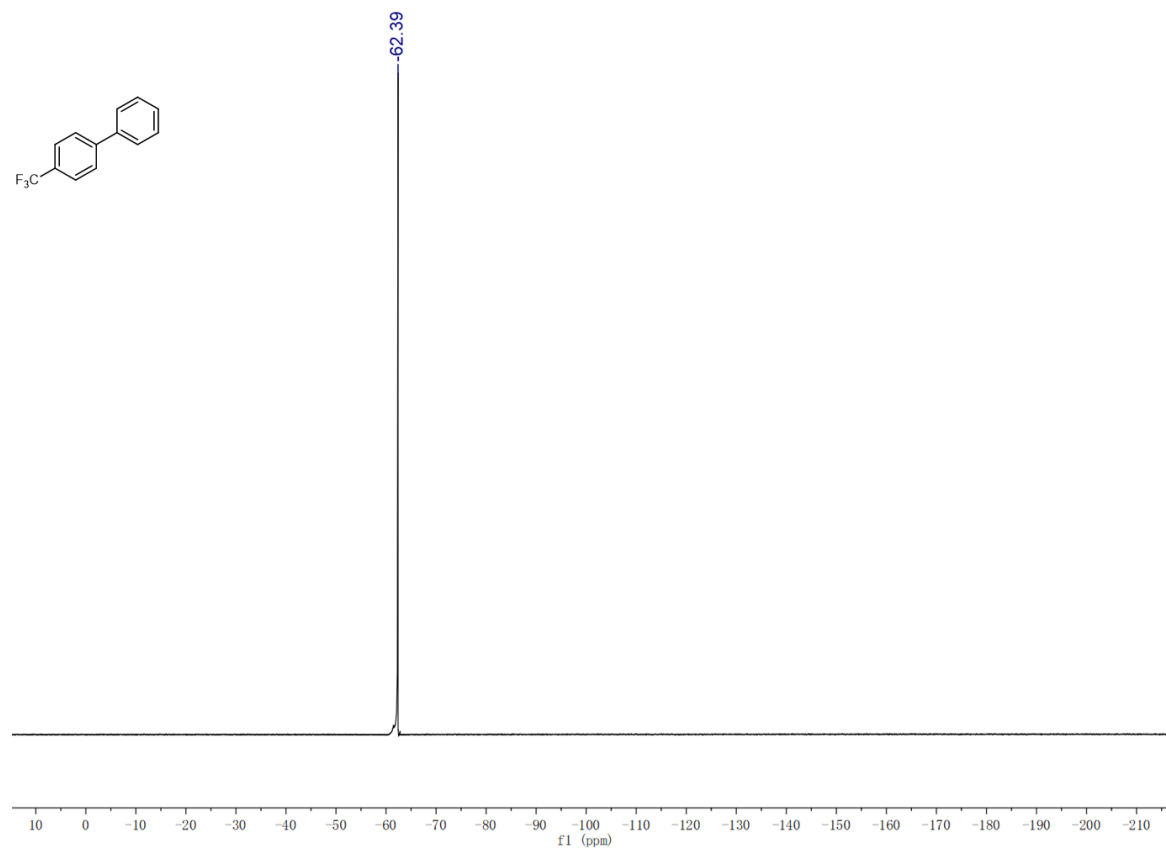


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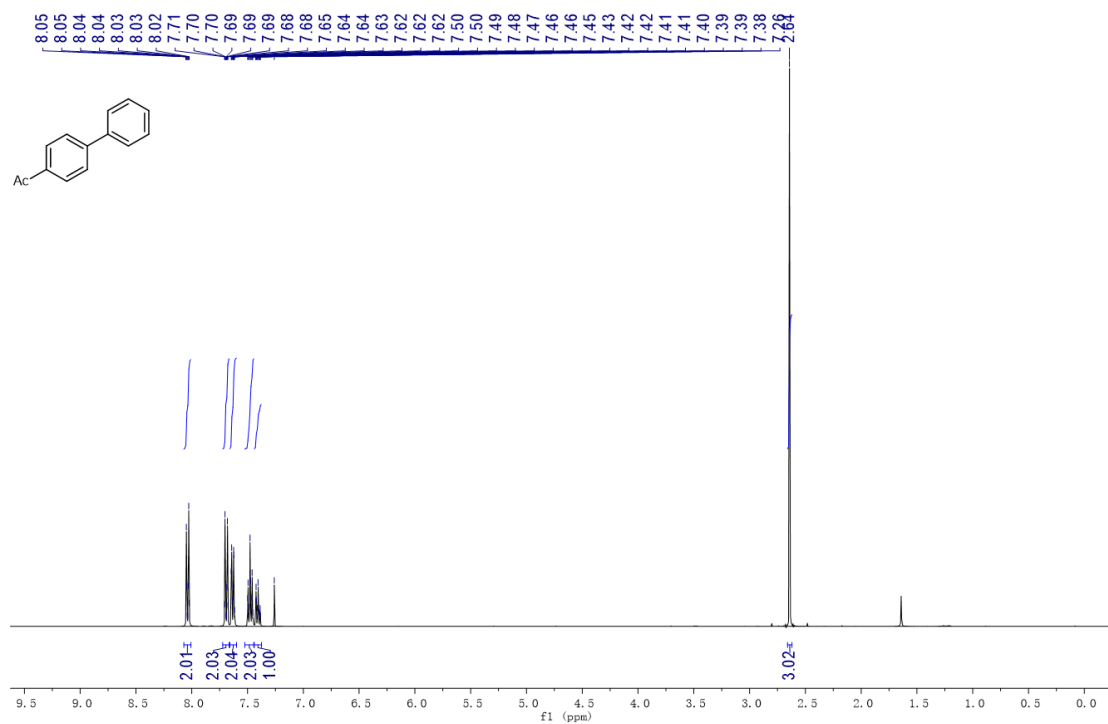


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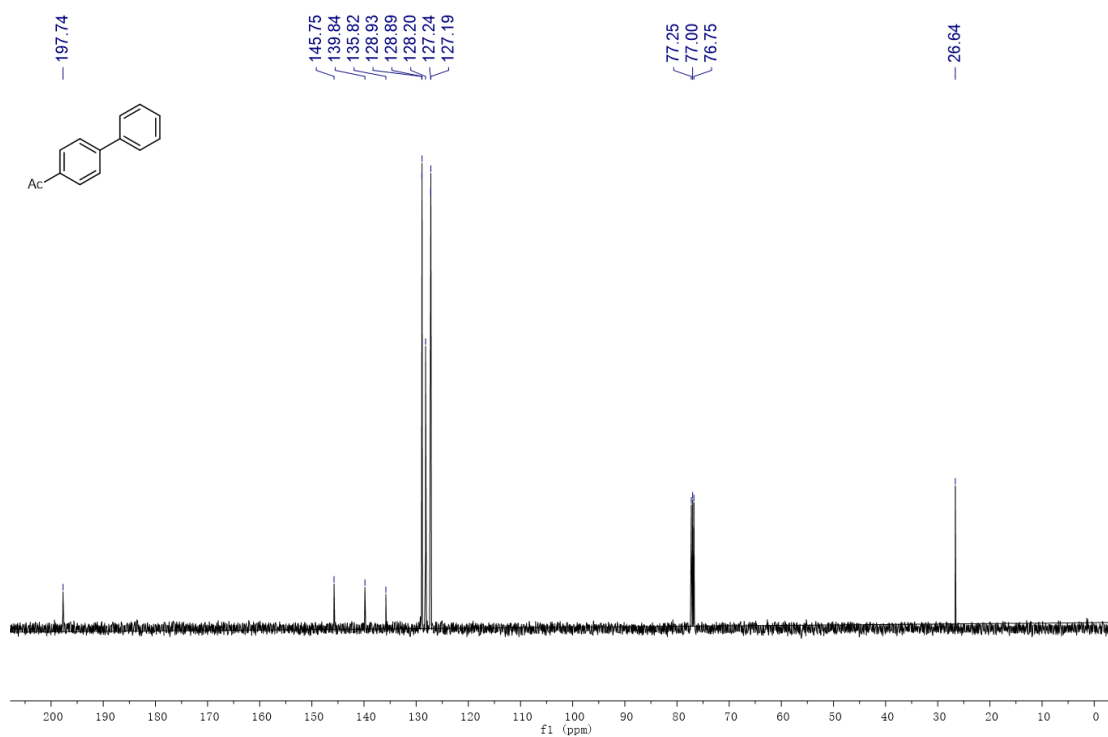
1049 ¹H and ¹³C-NMR spectra of product 4c.

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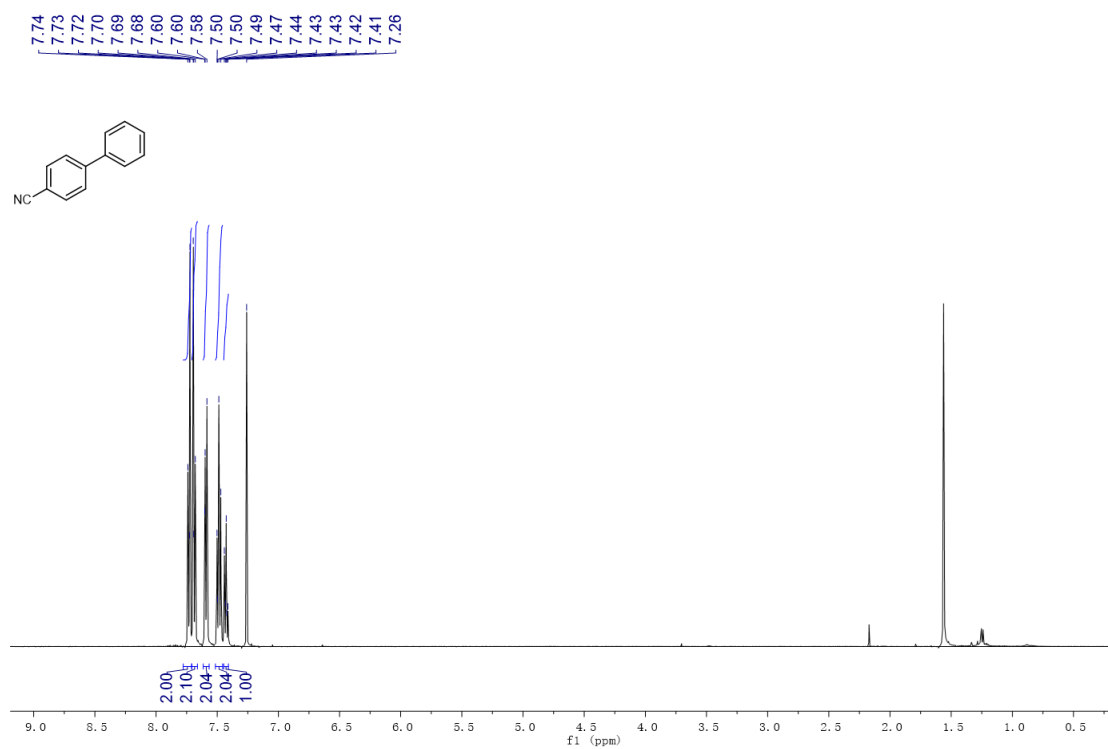


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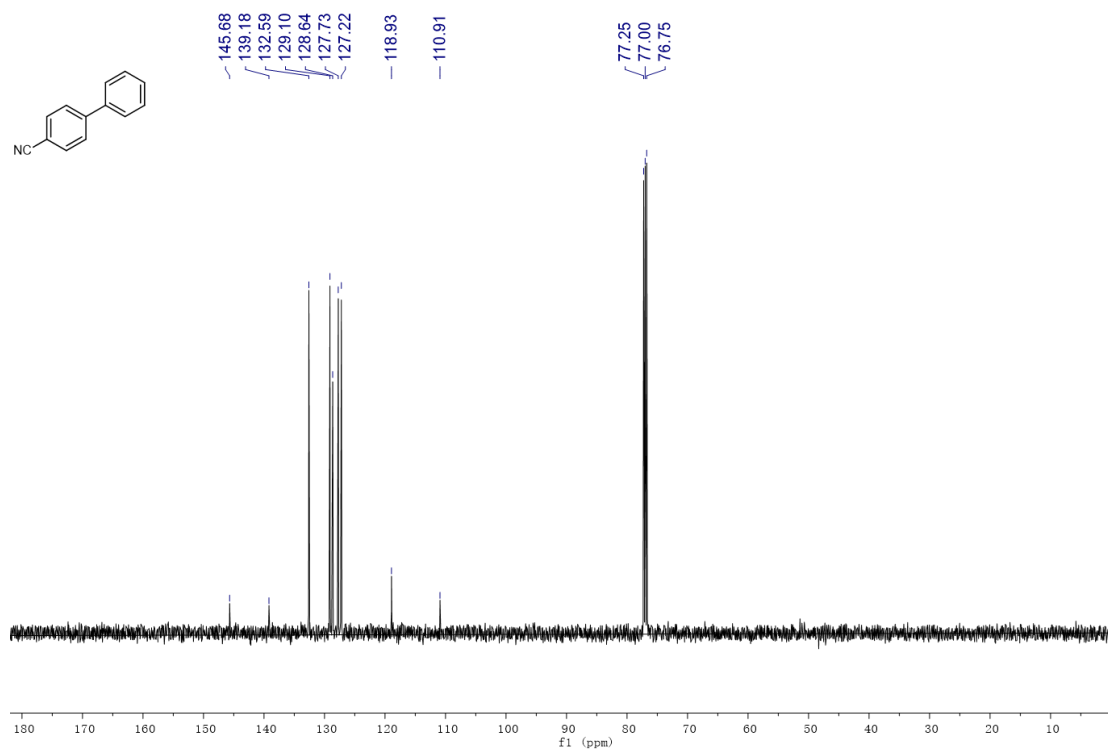
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1055 ¹H and ¹³C-NMR spectra of product 4d.

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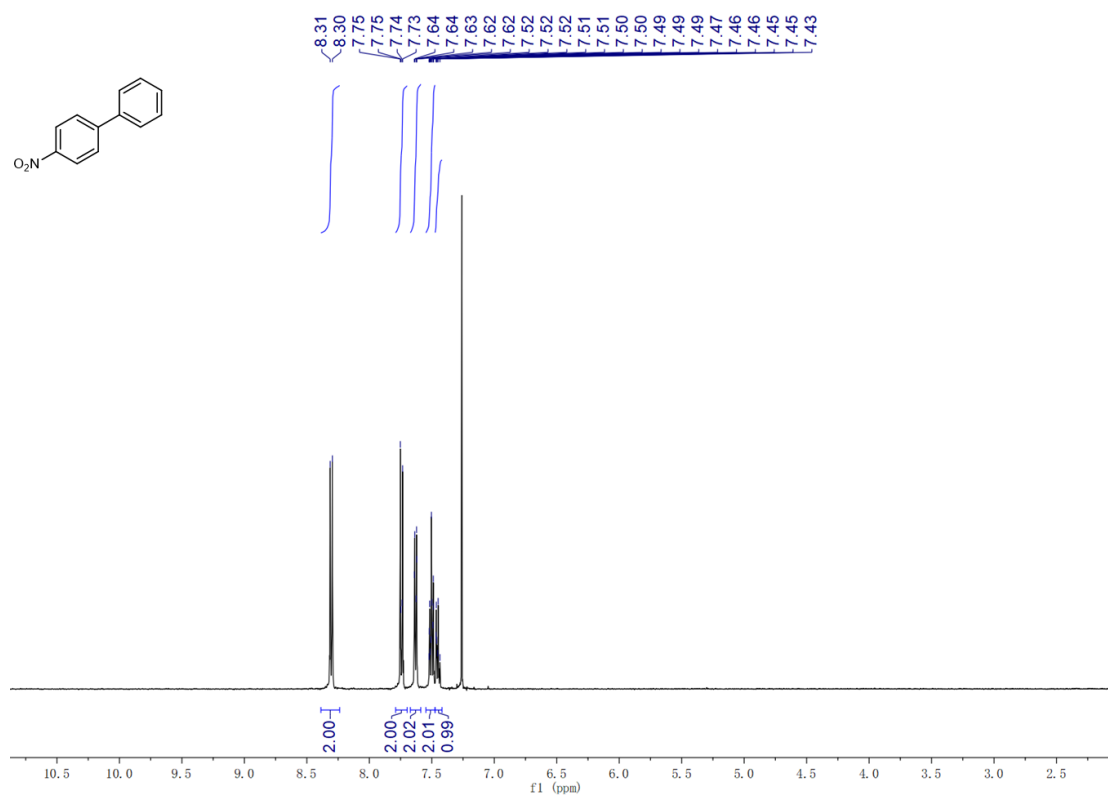
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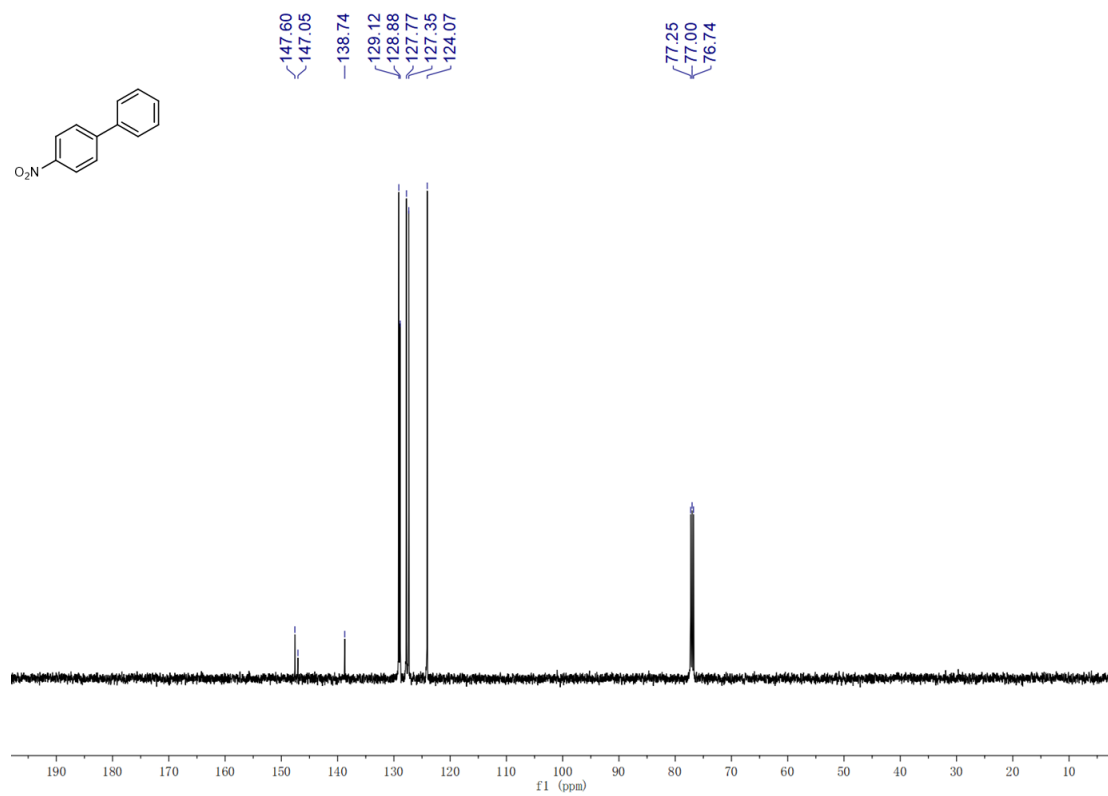
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1061 ¹H and ¹³C-NMR spectra of product 4e.

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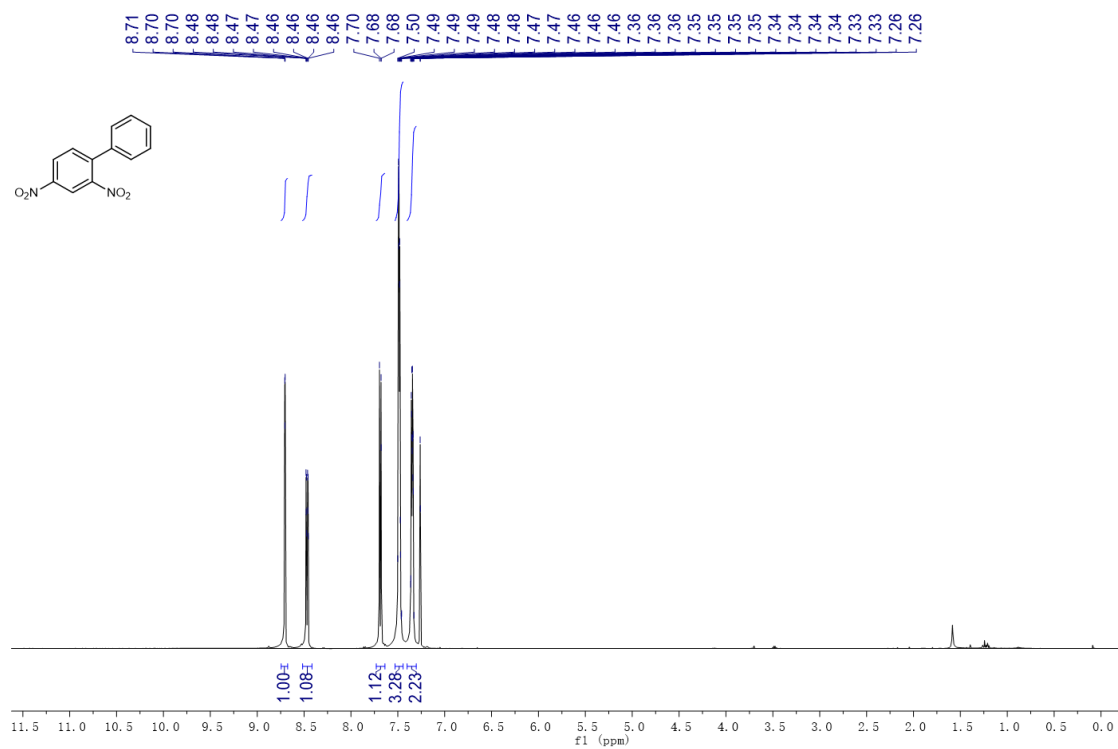


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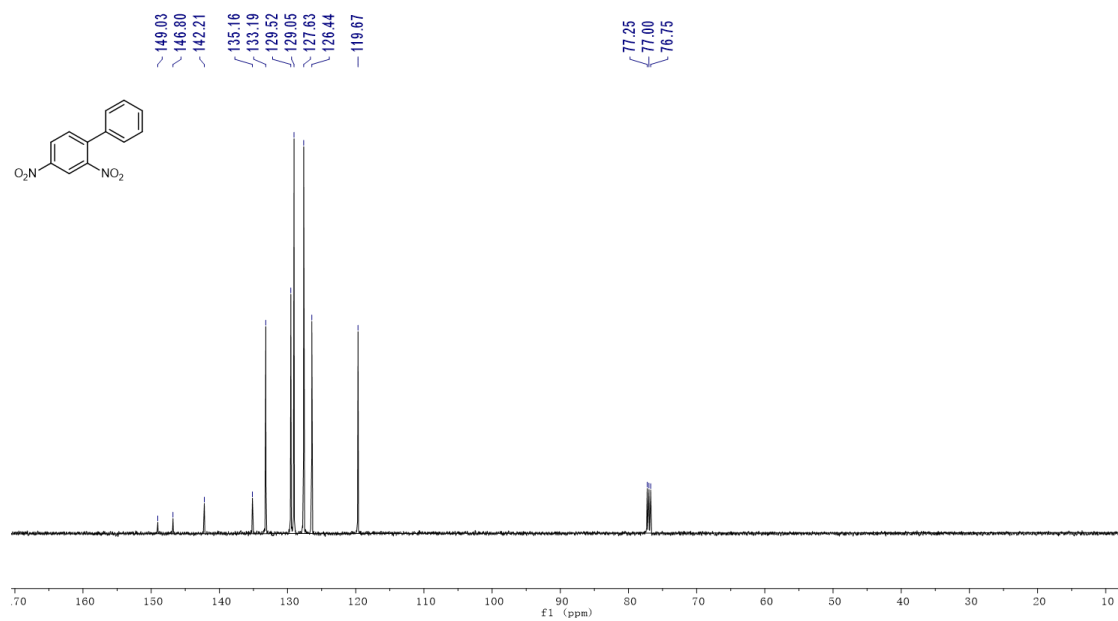
1066 ¹H and ¹³C-NMR spectra of product 4f.

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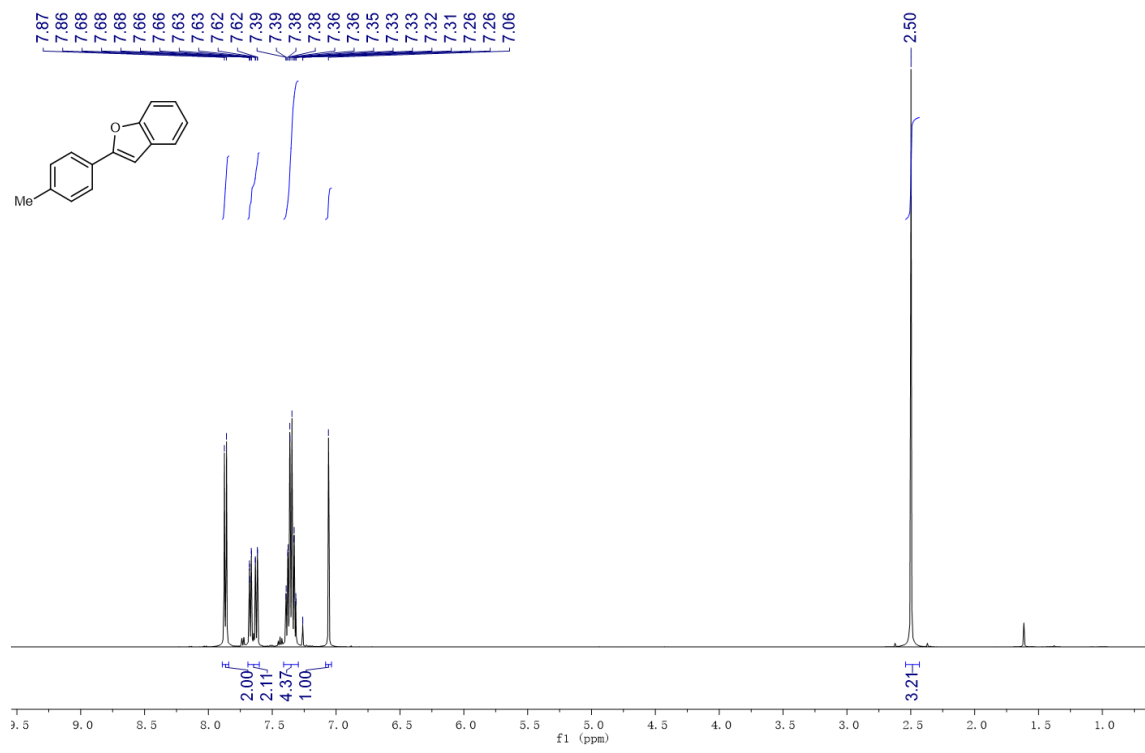


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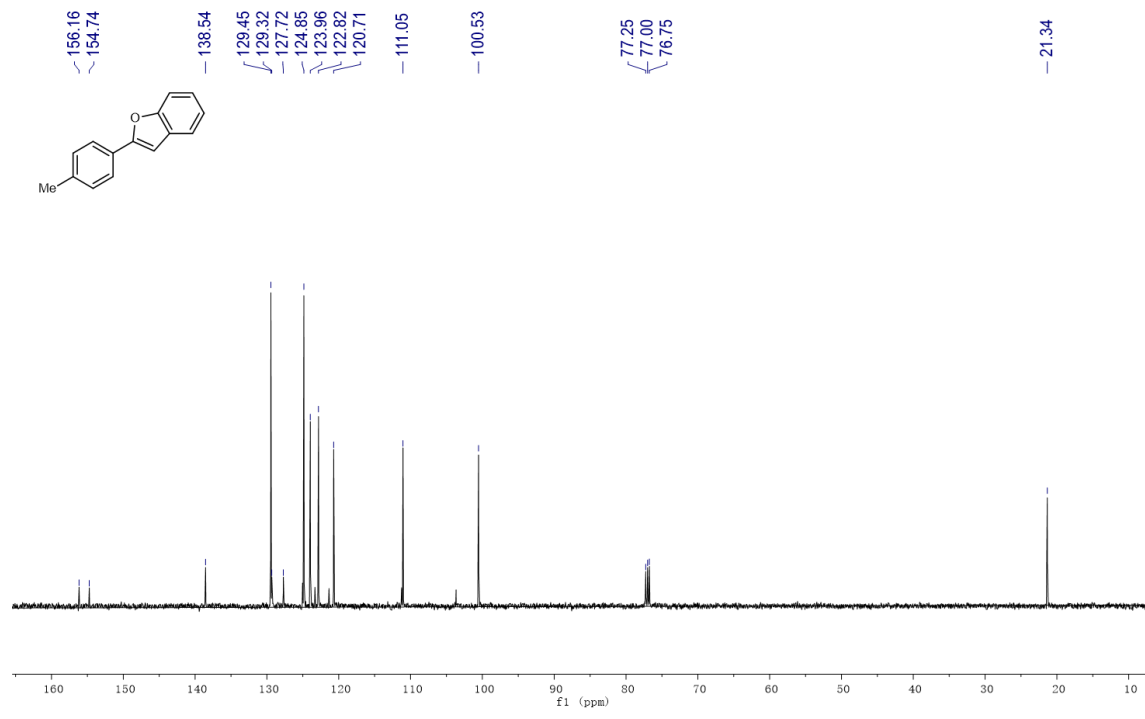
1072 ¹H and ¹³C-NMR spectra of product 4g.

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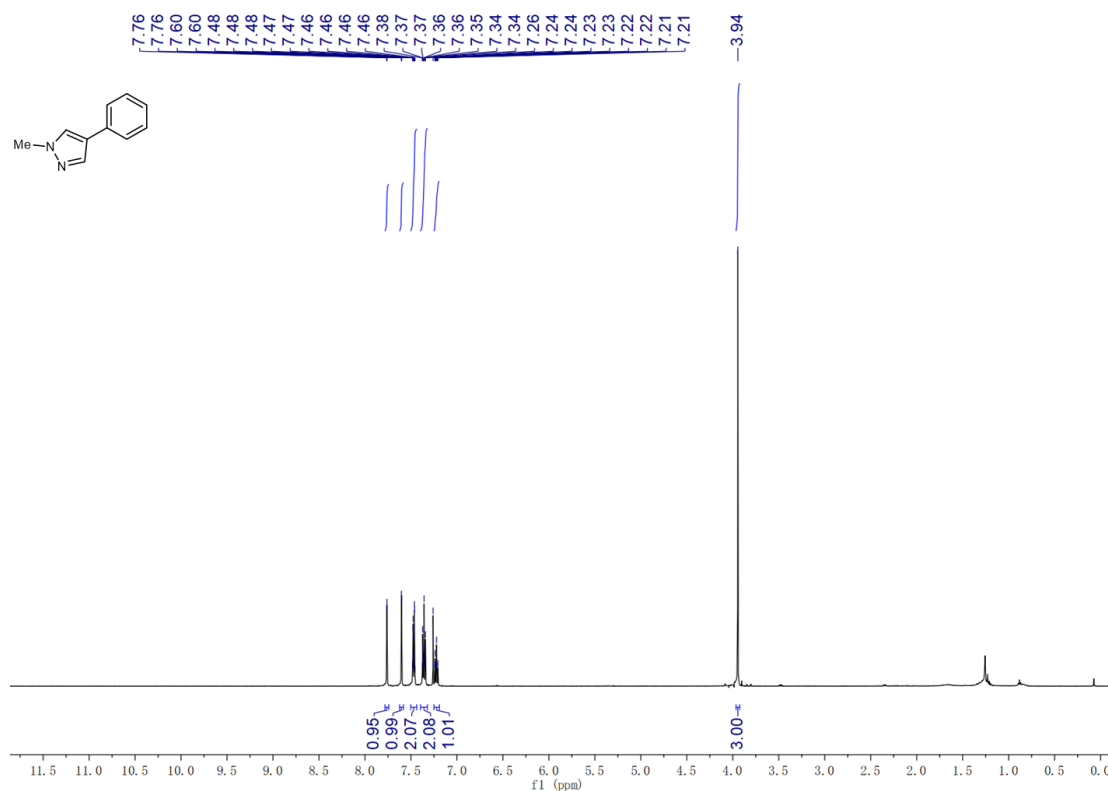
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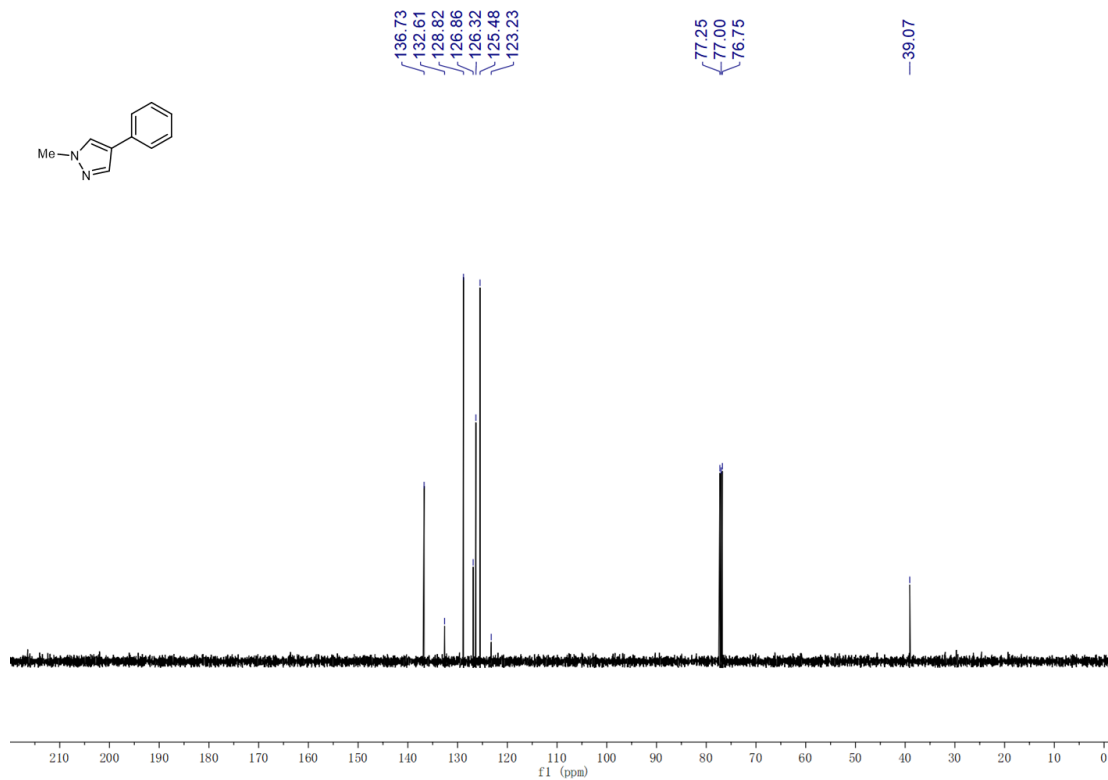
1079 ¹H and ¹³C-spectra of product 4h.

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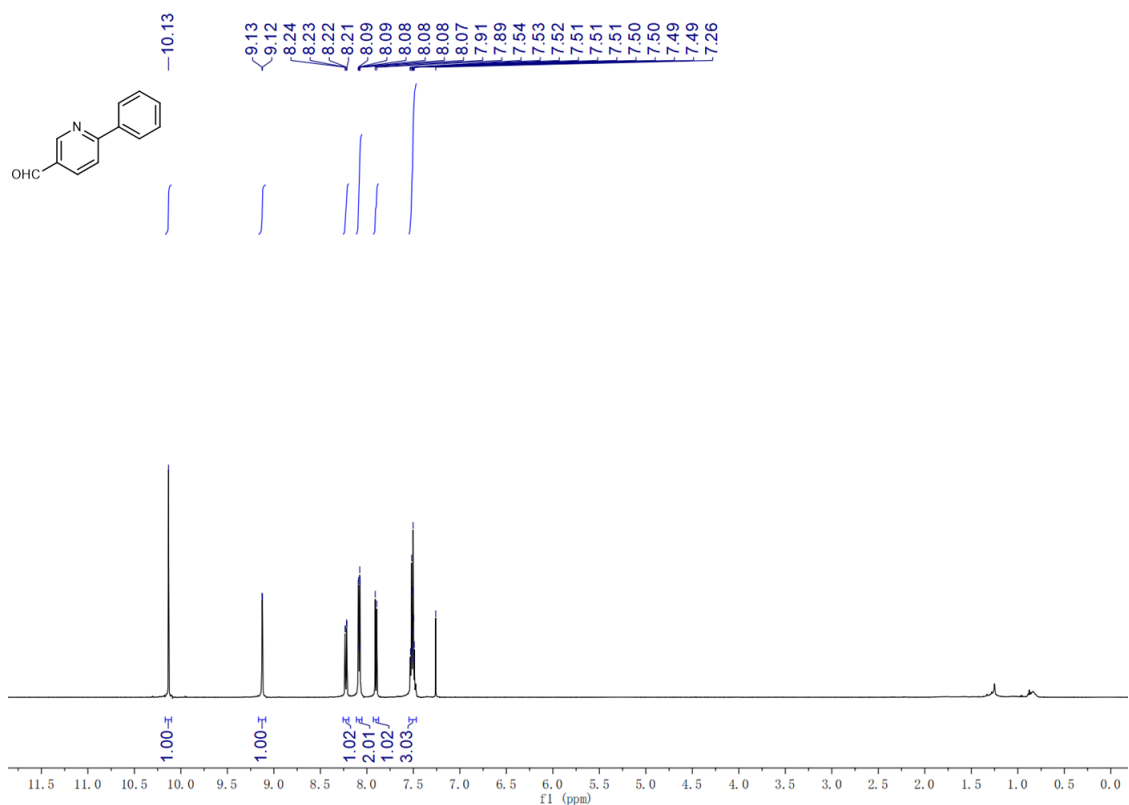
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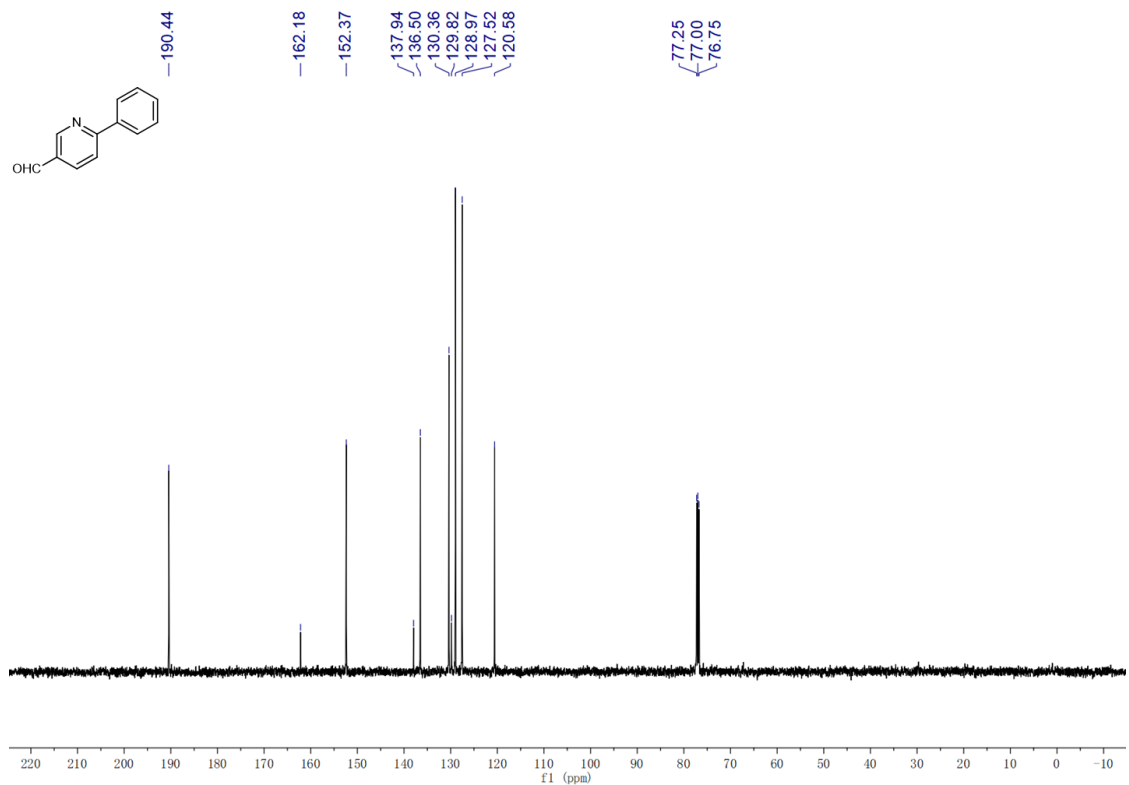
1084 ¹H, ¹³C-NMR spectra of product 4i.

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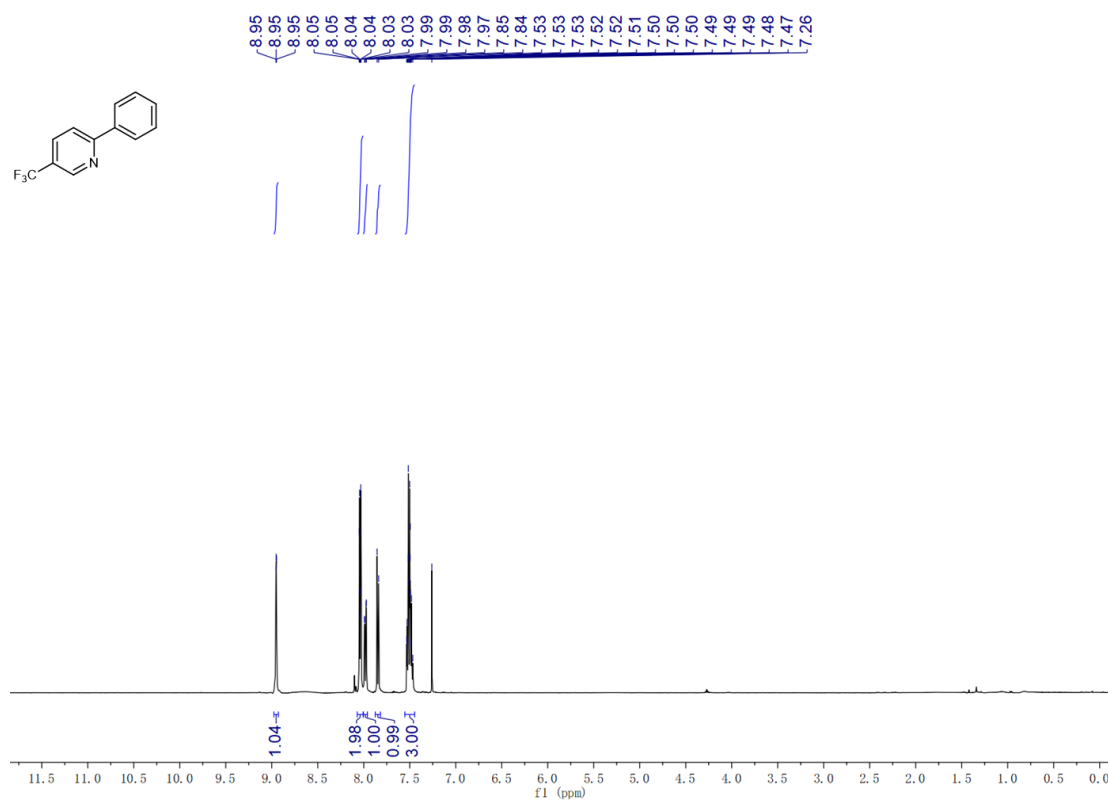


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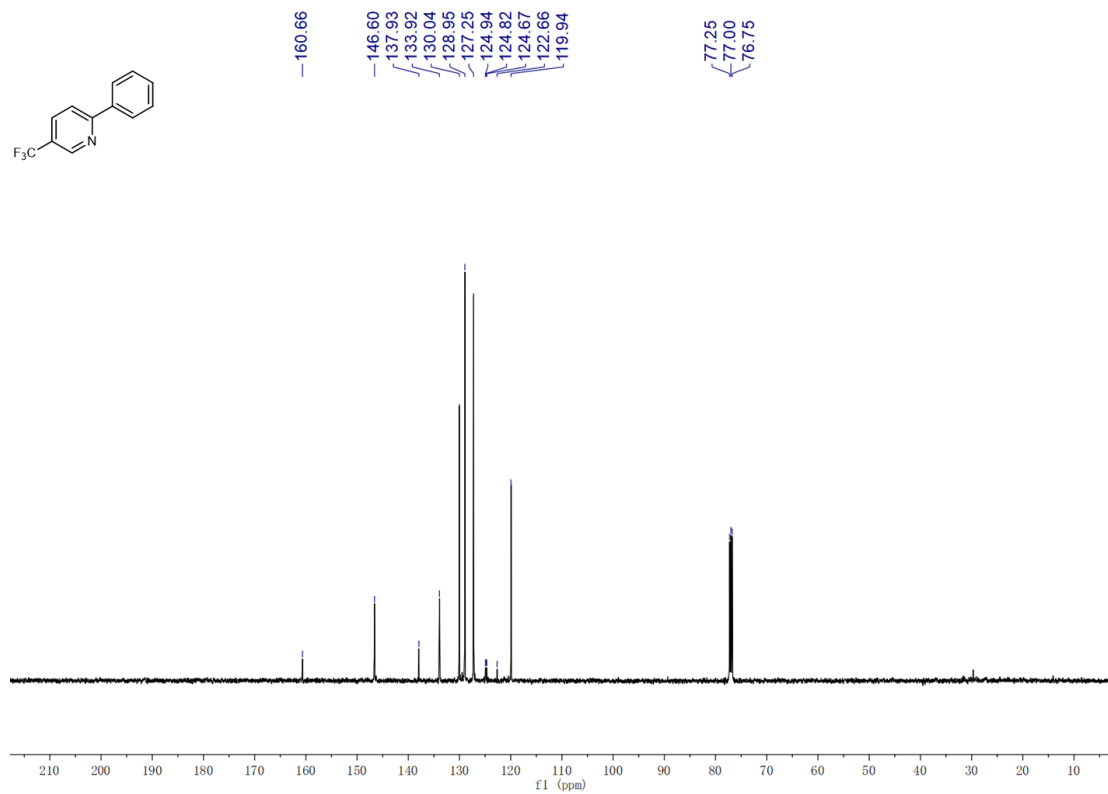
1090 ¹H and ¹³C, ¹⁹F-NMR spectra of product 4j.

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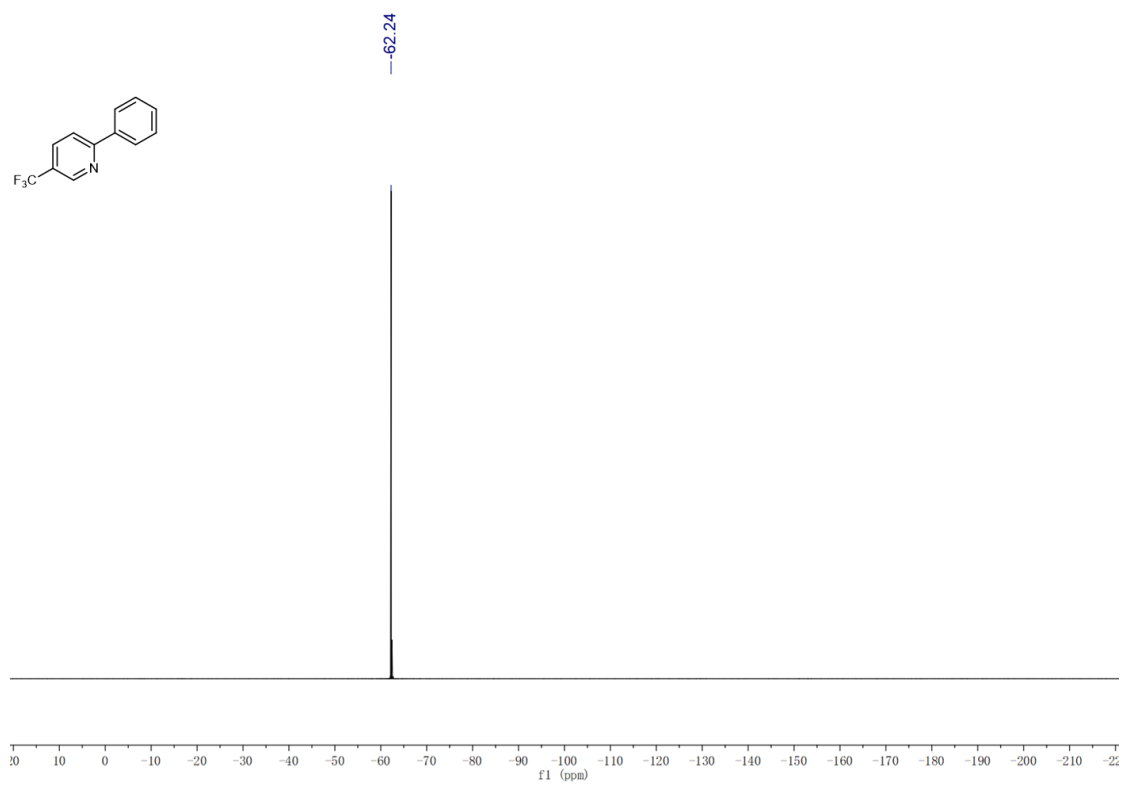
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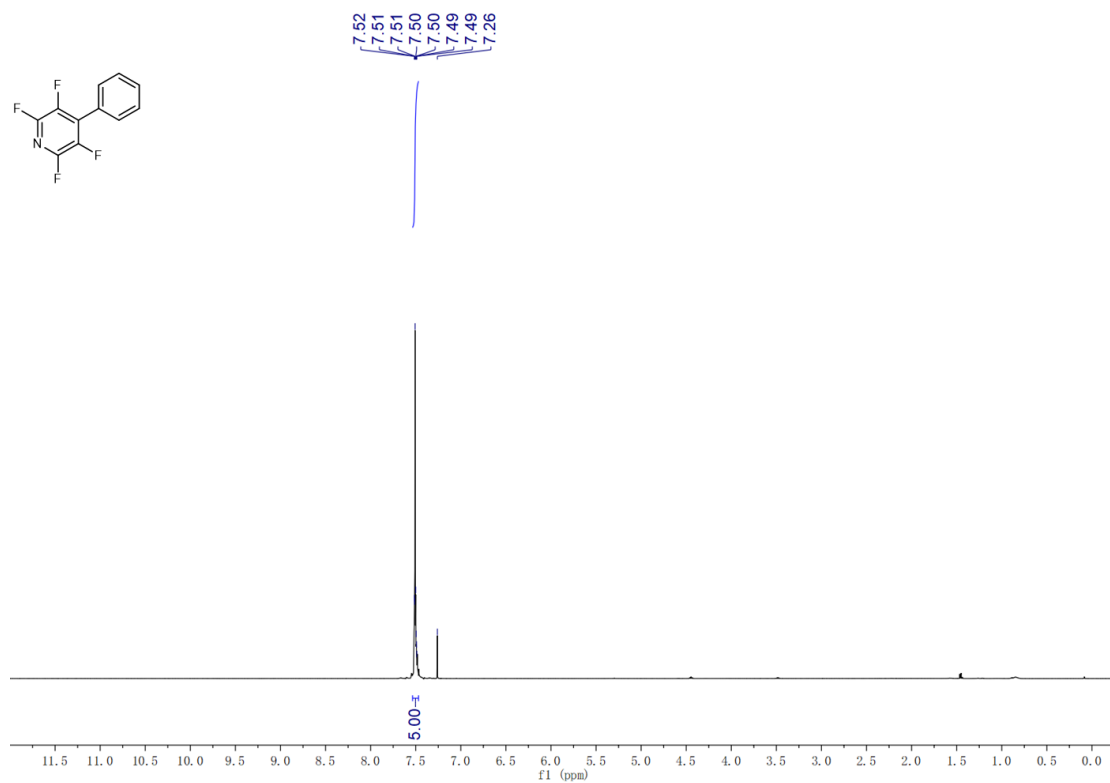
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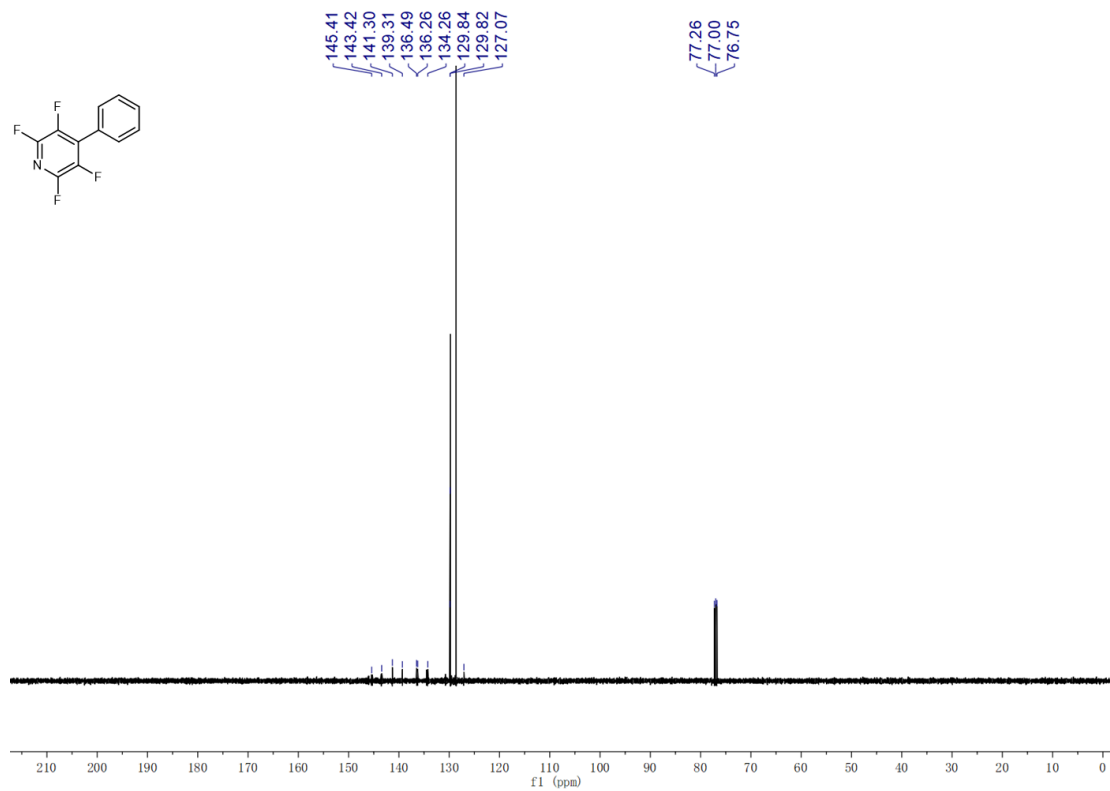
1098 ¹H and ¹³C, ¹⁹F-NMR spectra of product 4k.

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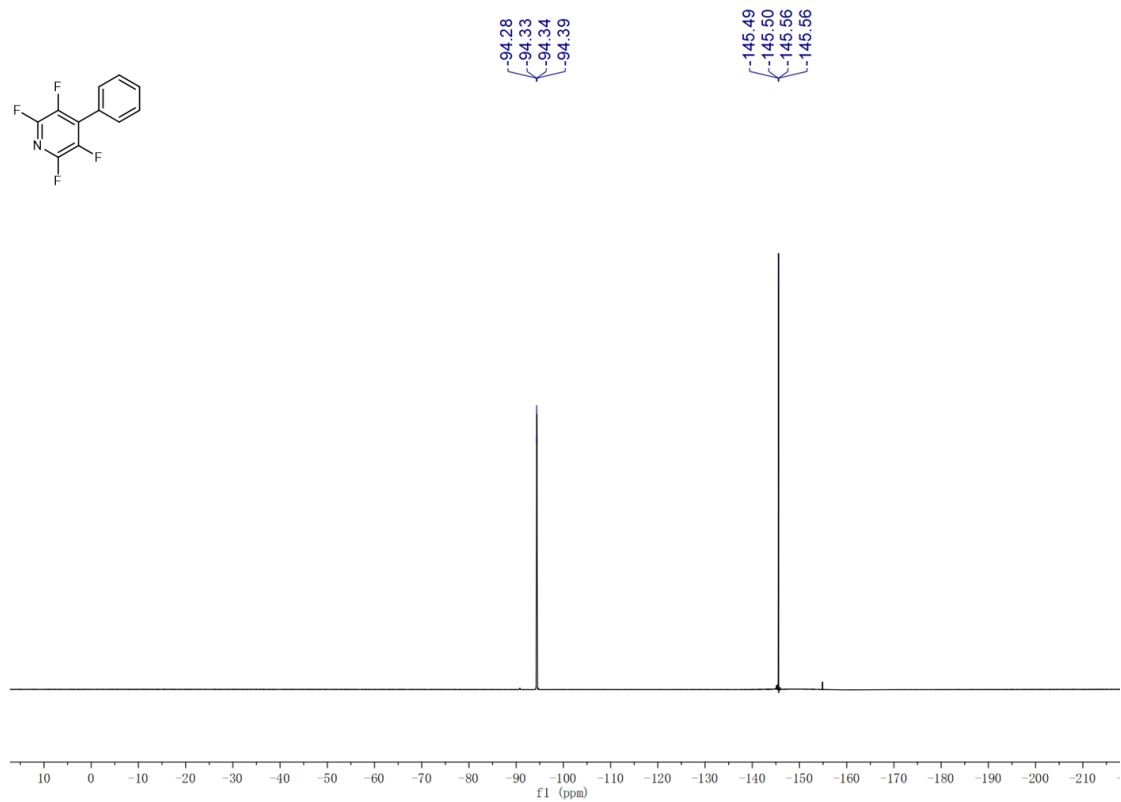
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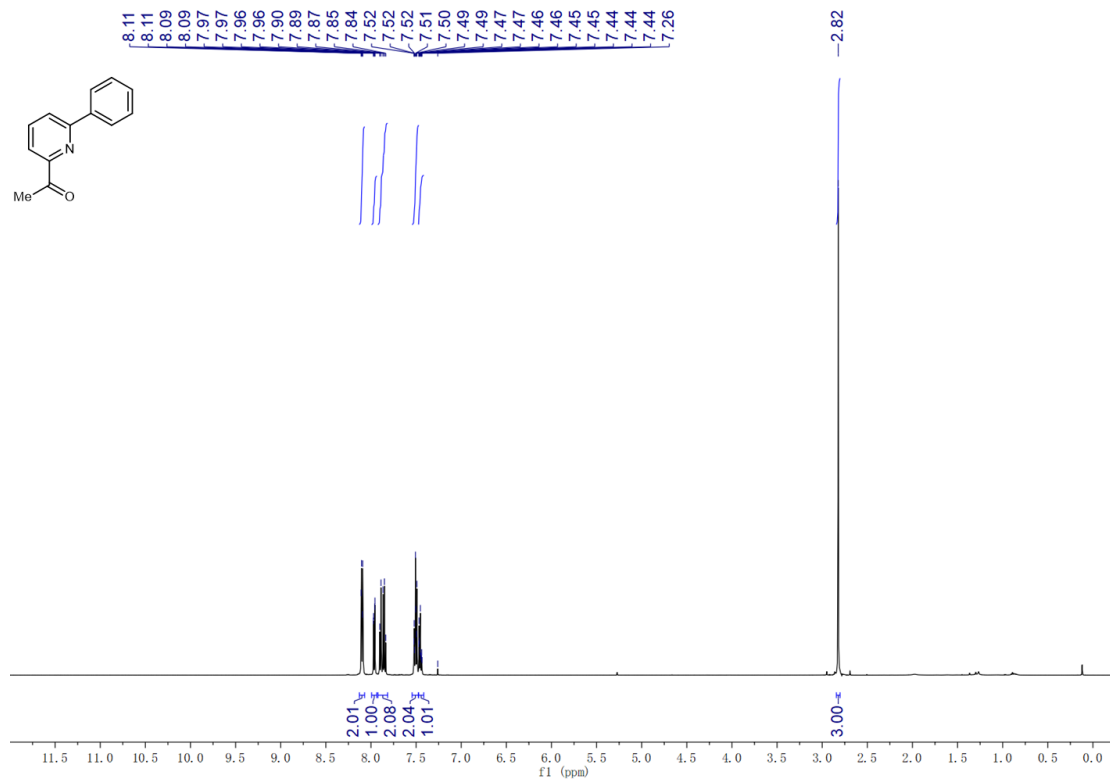


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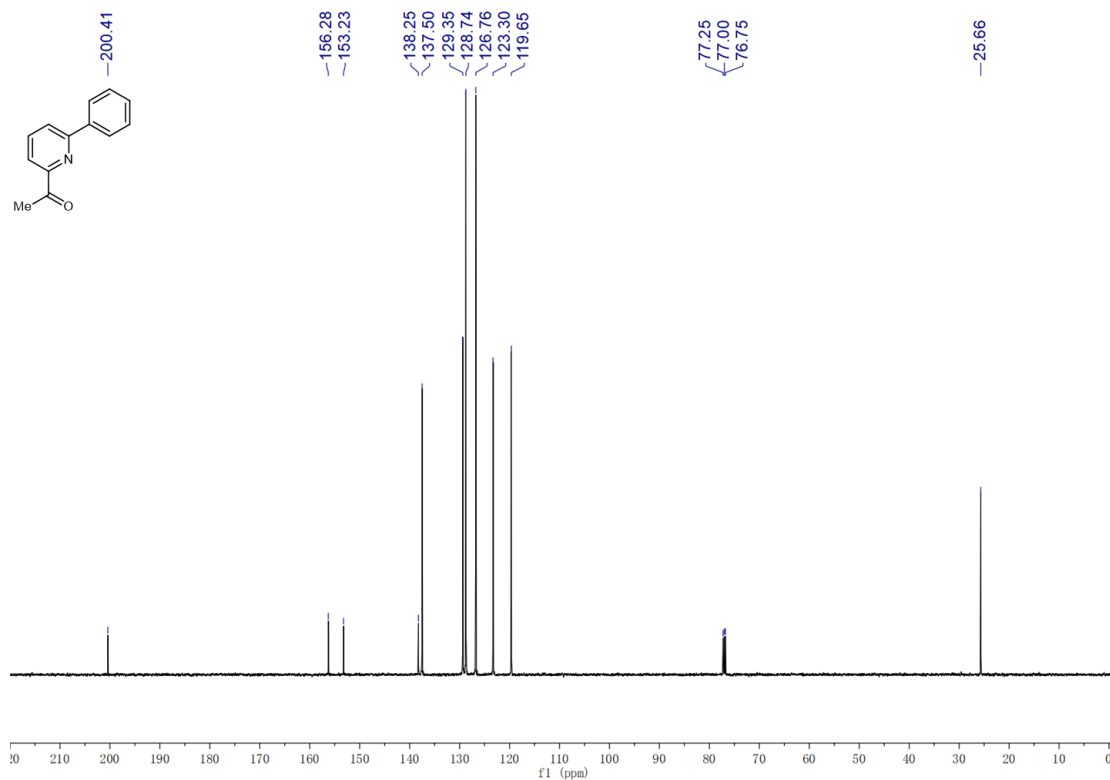
1107 ¹H and ¹³C spectra of product 4l.

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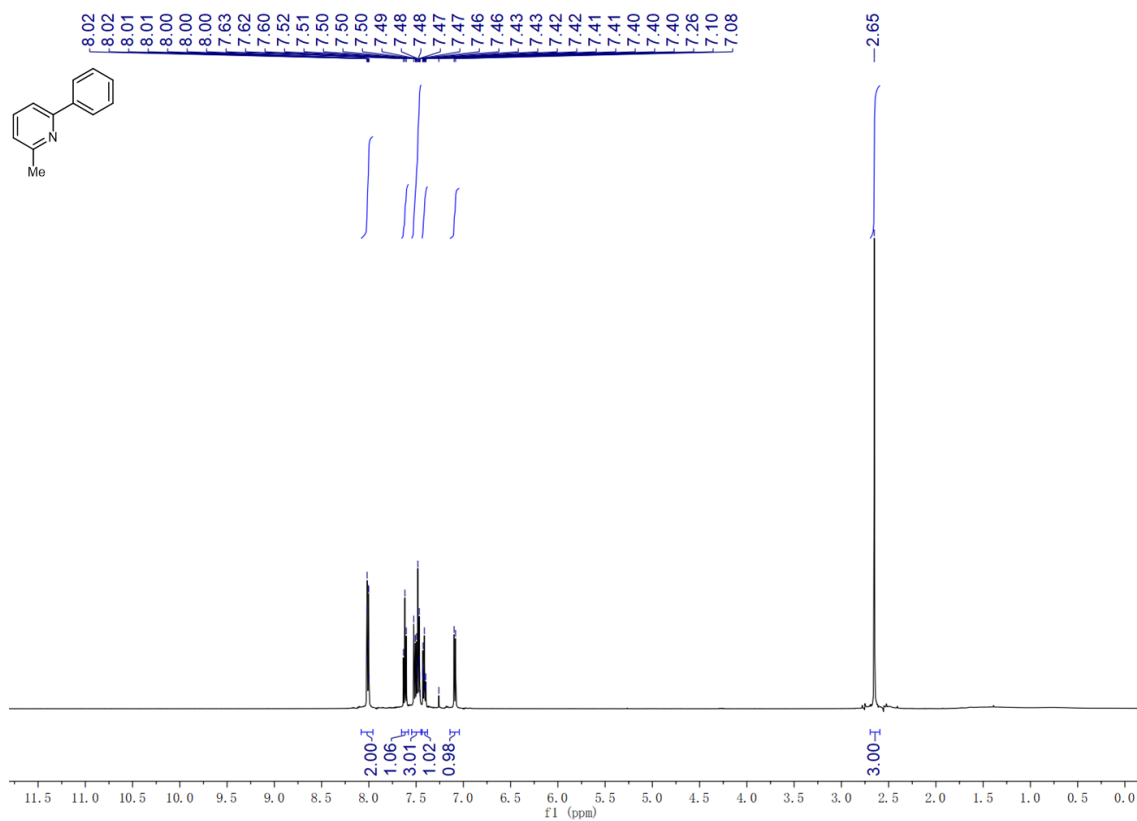
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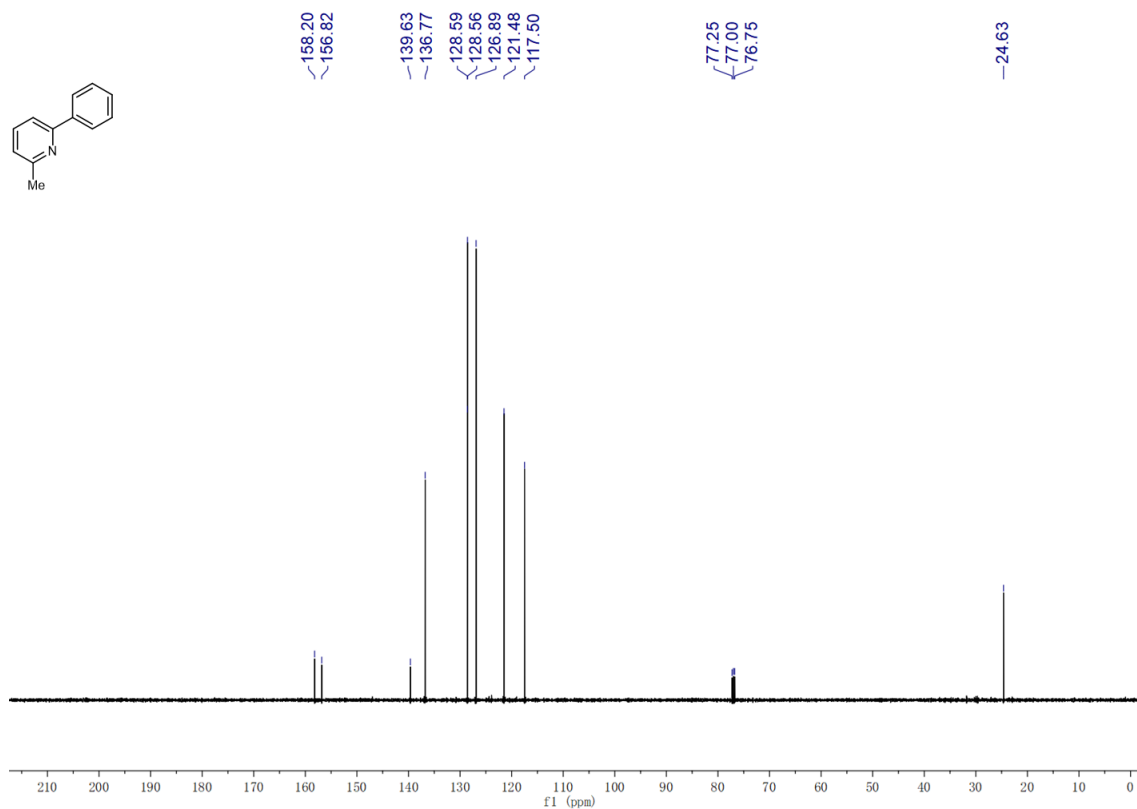
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1114 ¹H and ¹³C-spectra of product 4m.

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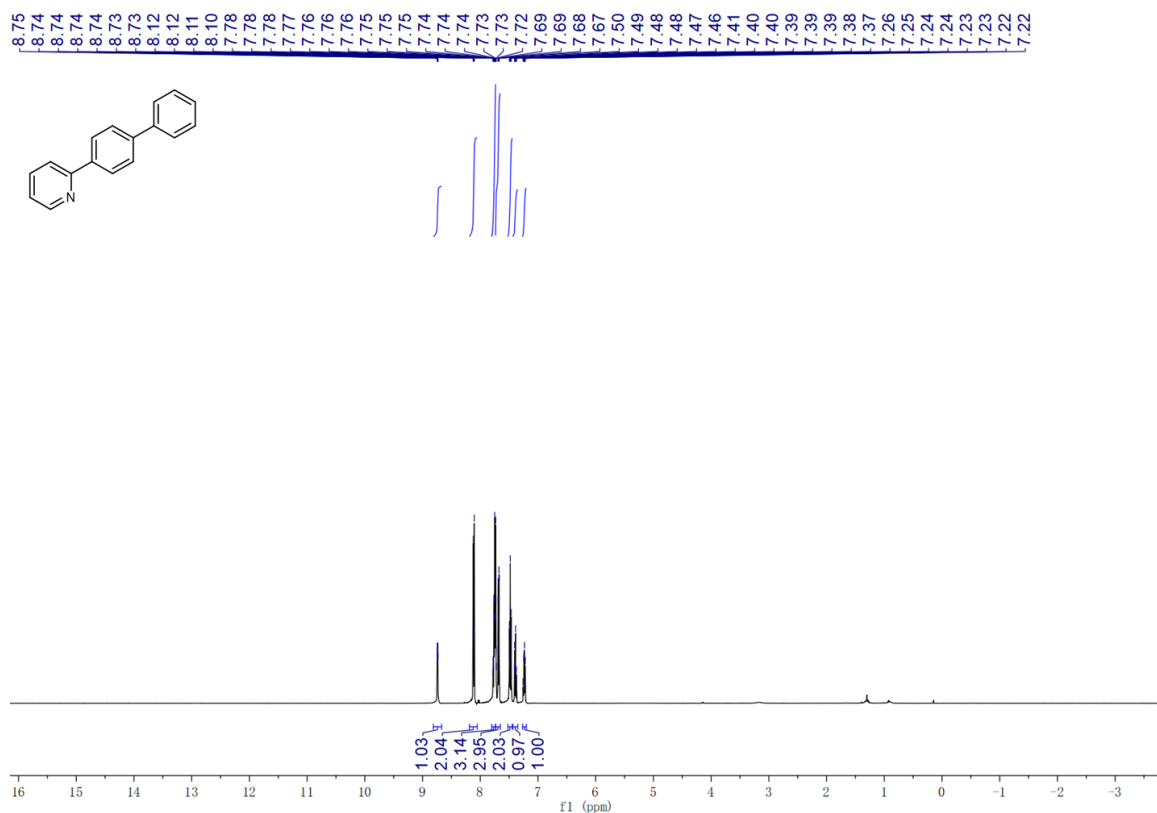


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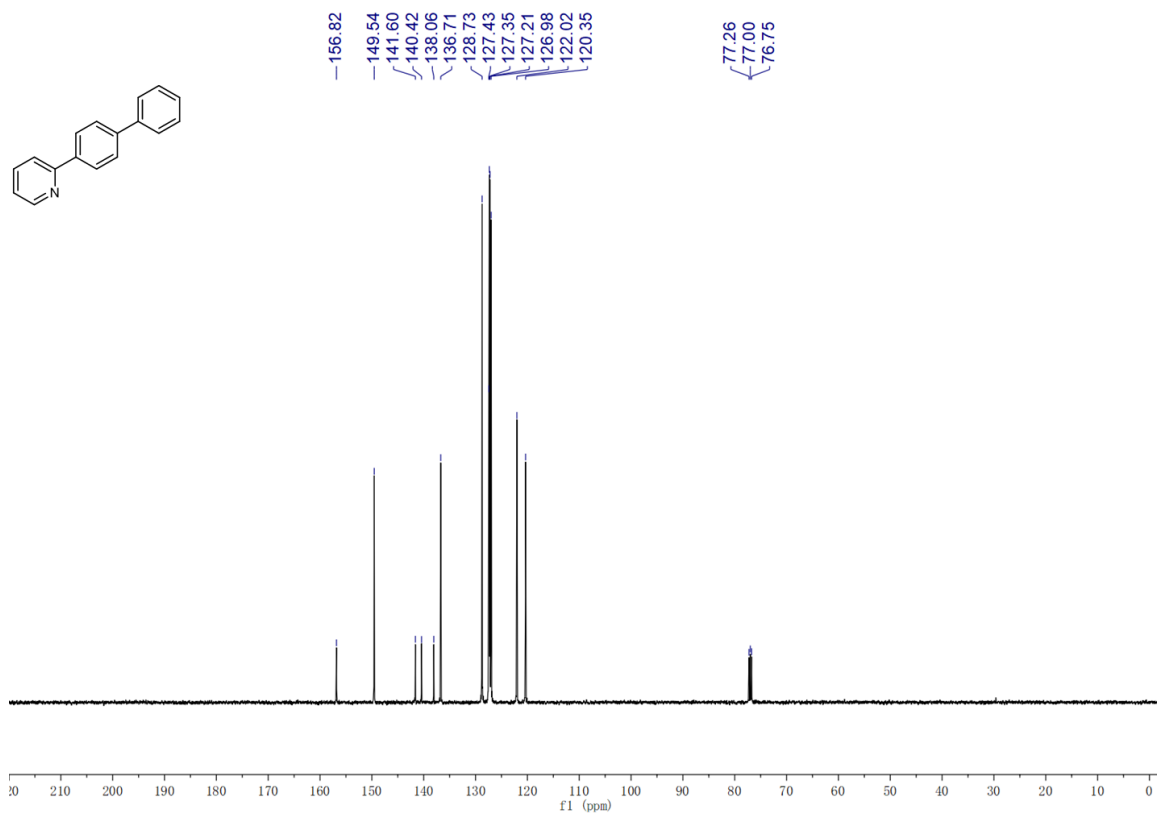
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1119 ¹H and ¹³C-NMR spectra of product 4n.

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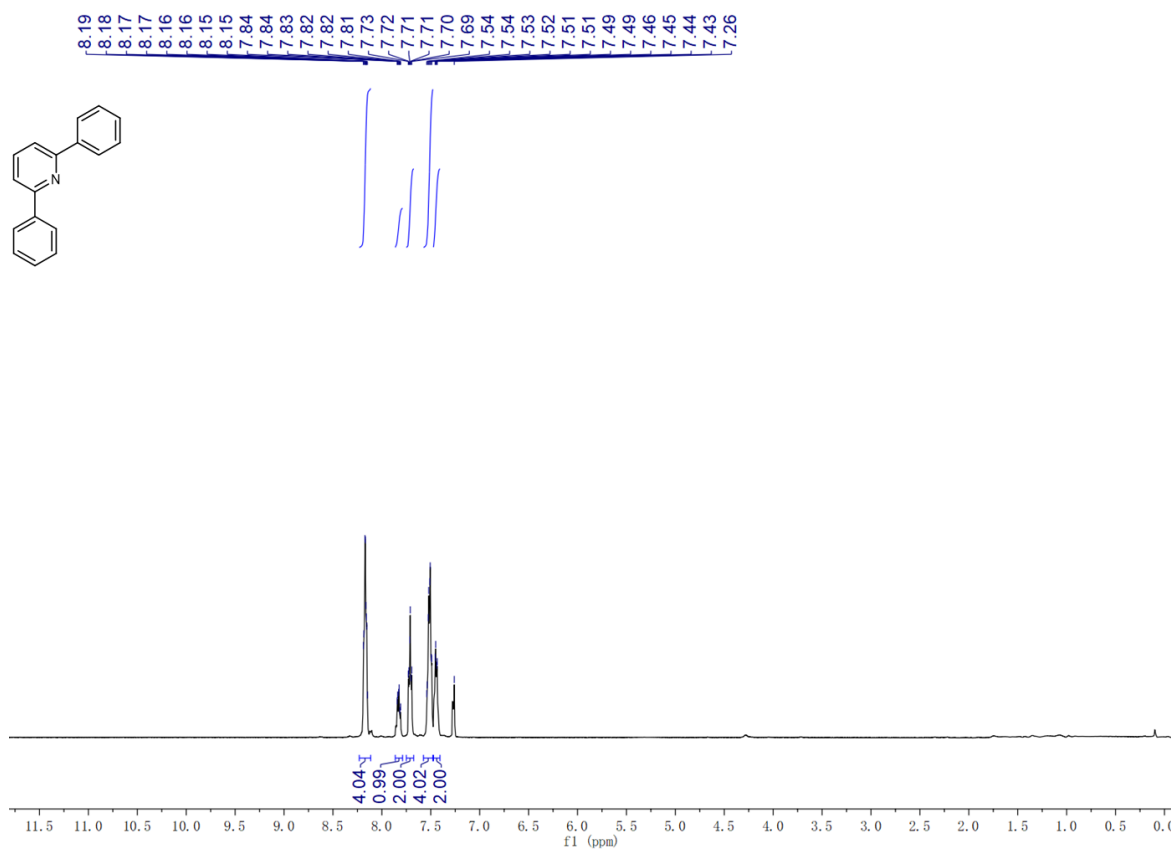


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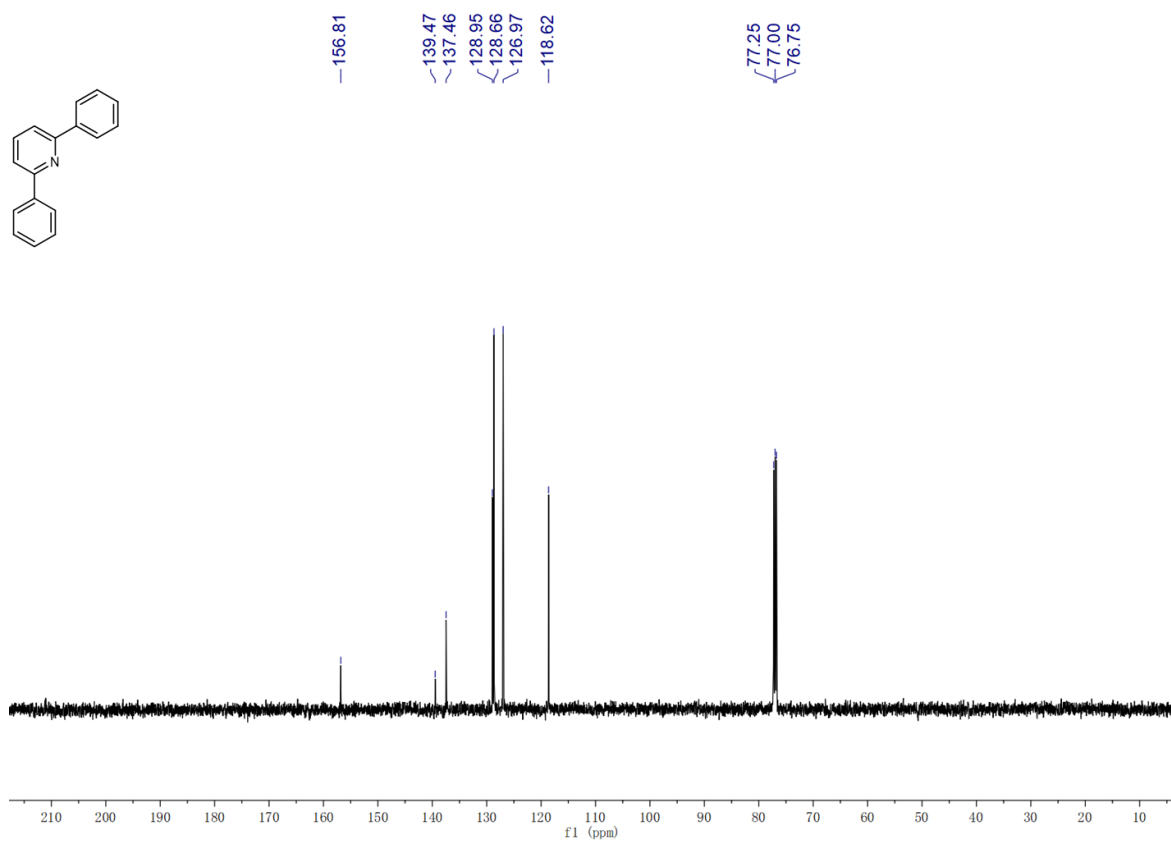
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1124 ¹H and ¹³C spectra of product 4o.

1125



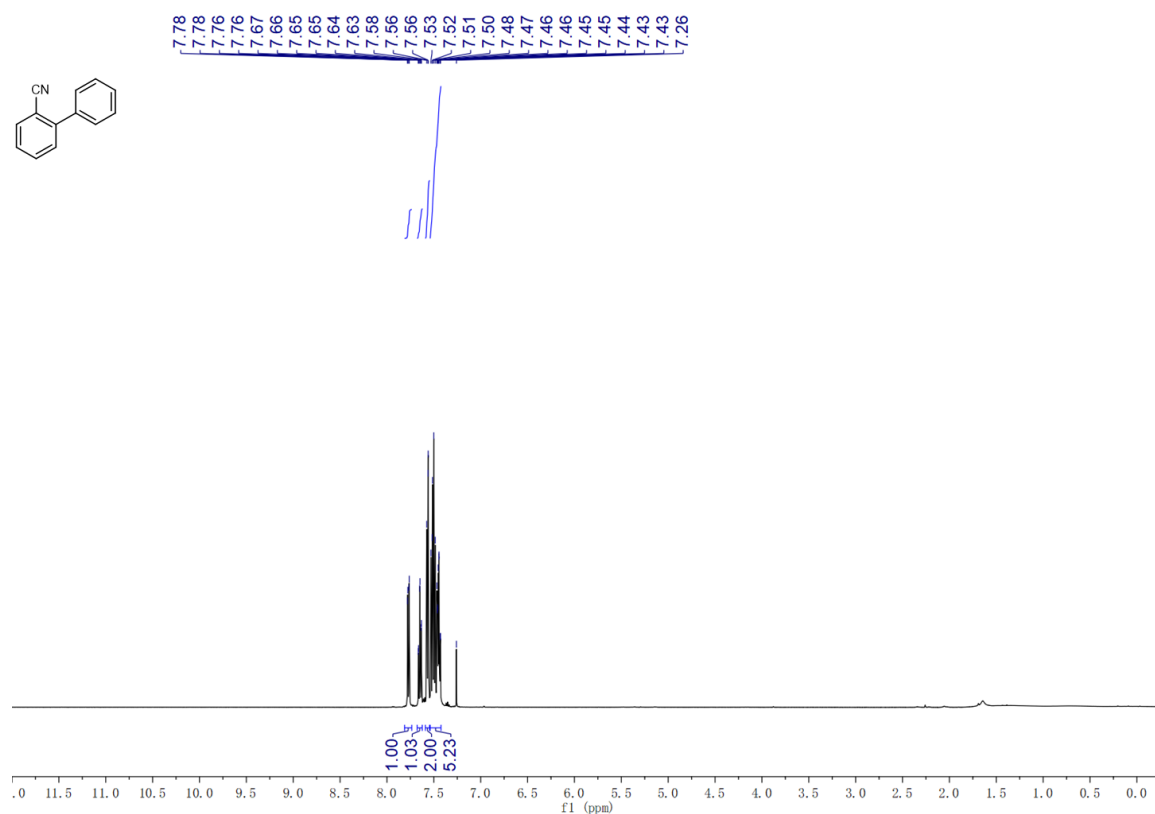
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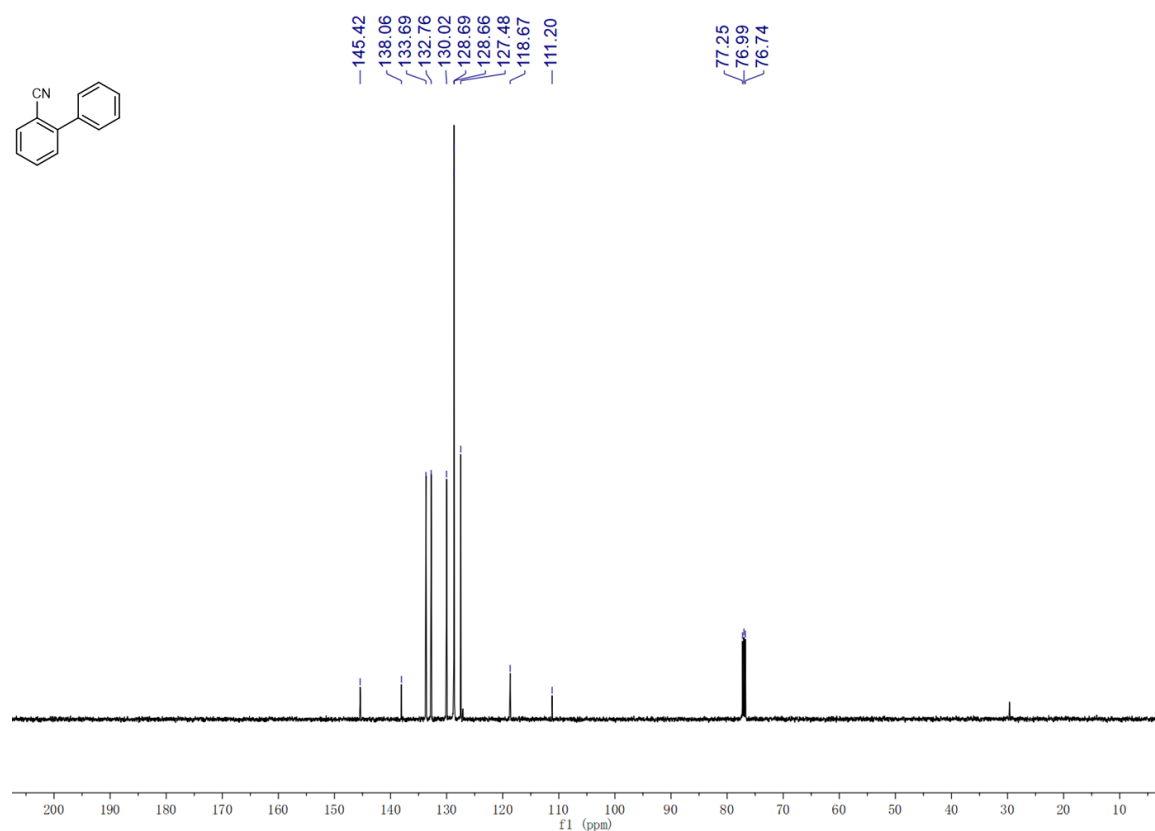
1128 ¹H and ¹³C- NMR spectra of product 4p.

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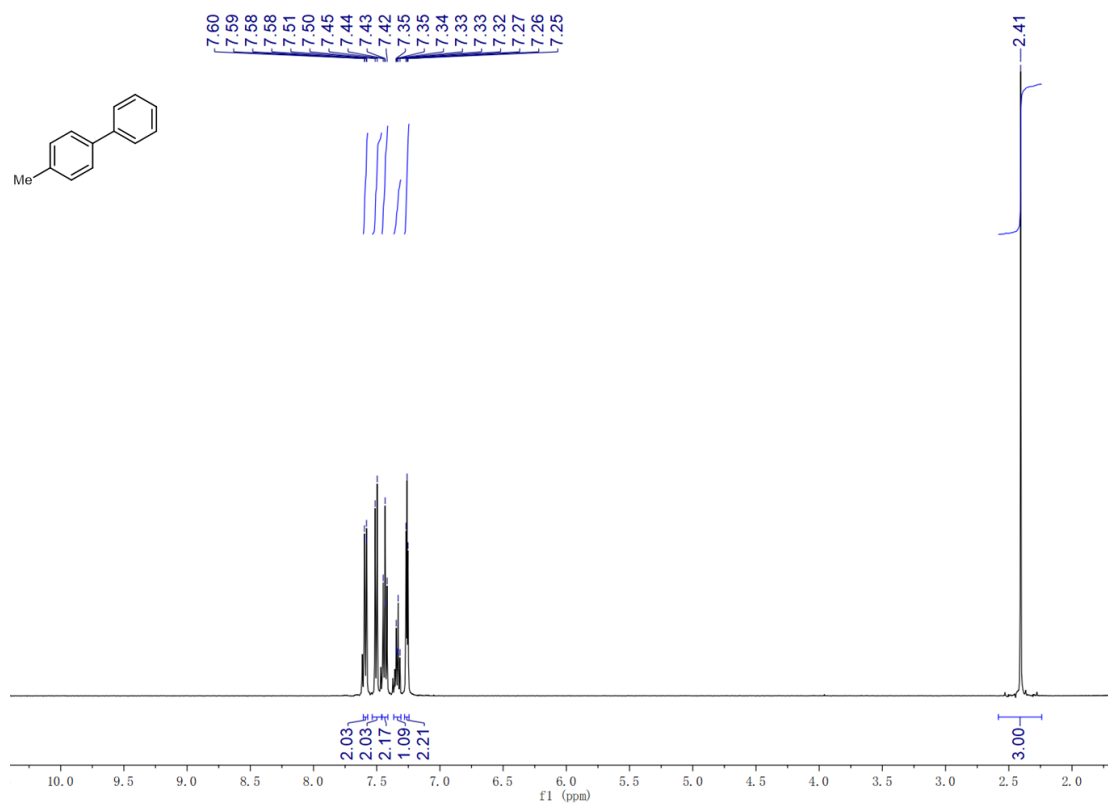
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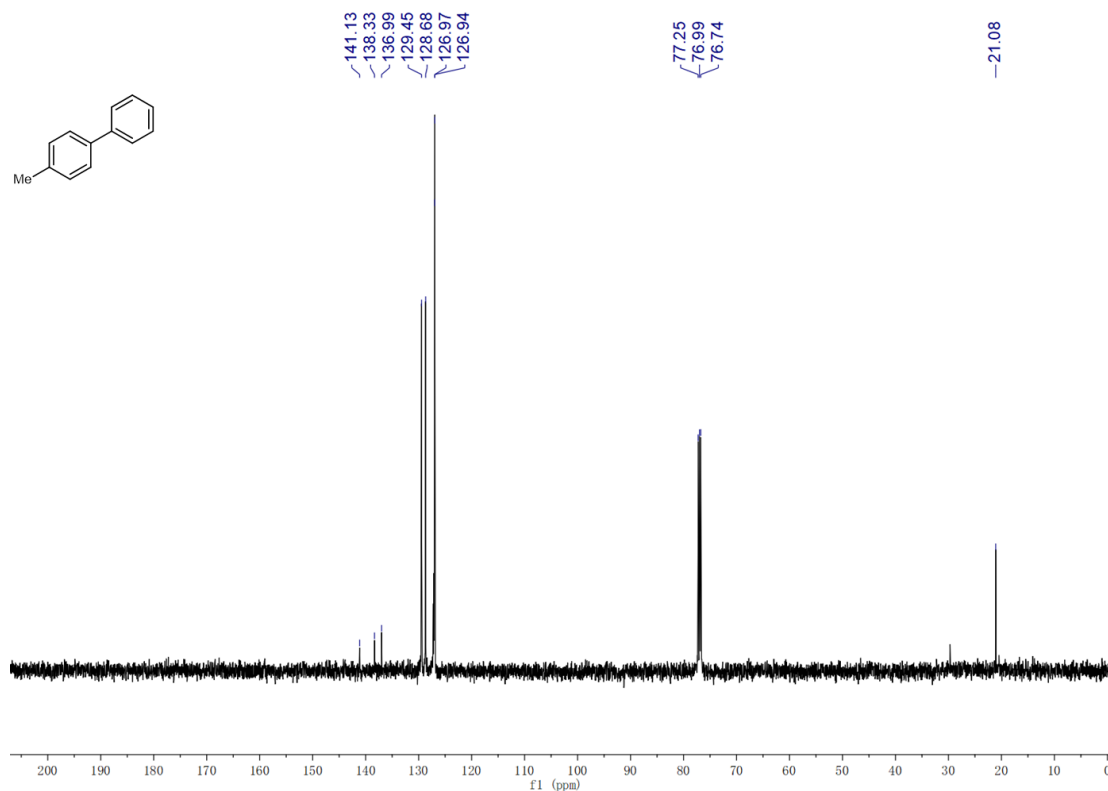


1132 ¹H and ¹³C-NMR spectra of product 4q.

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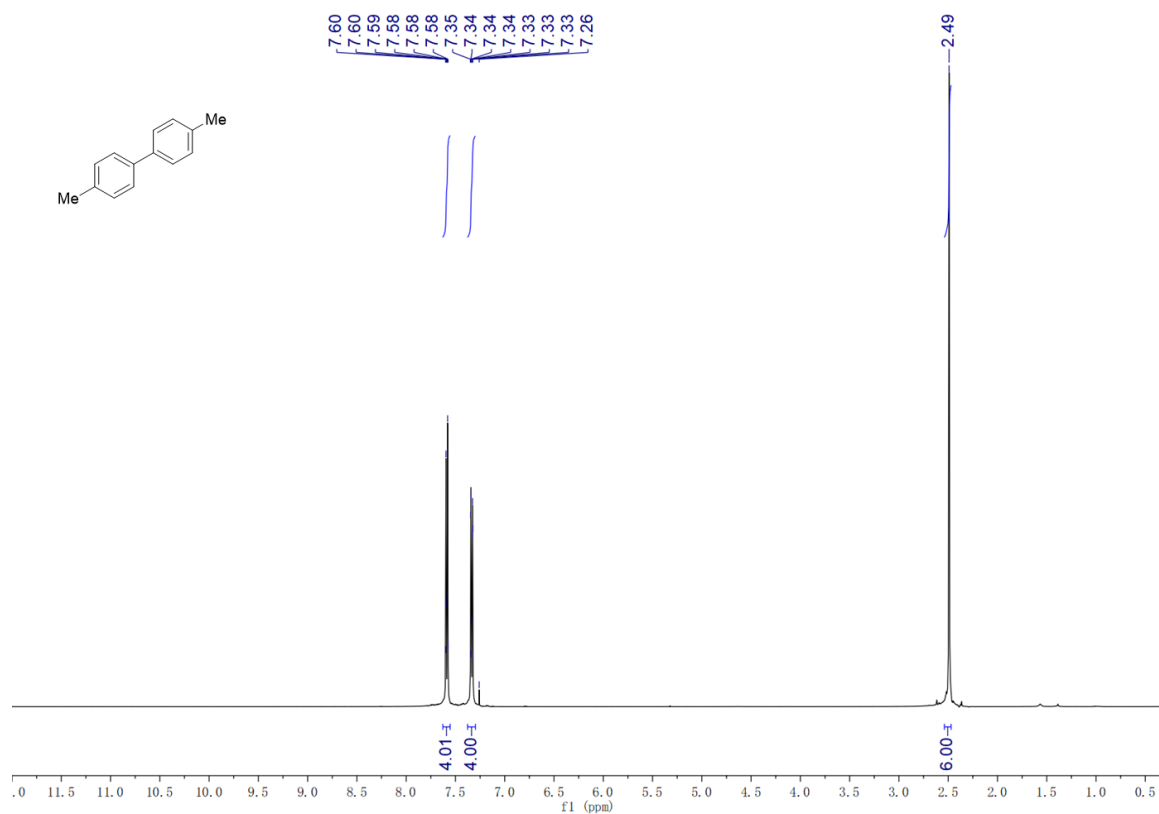


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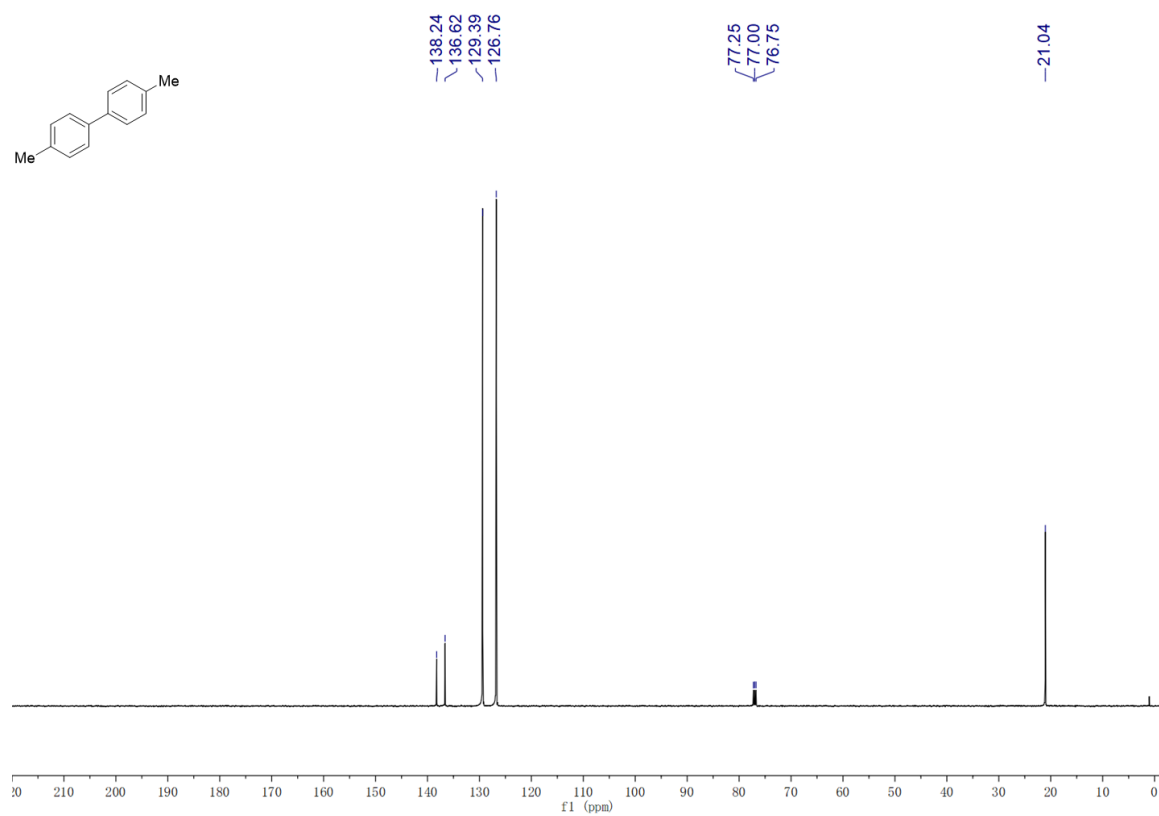
1137 ¹H and ¹³C- NMR spectra of product 4r.

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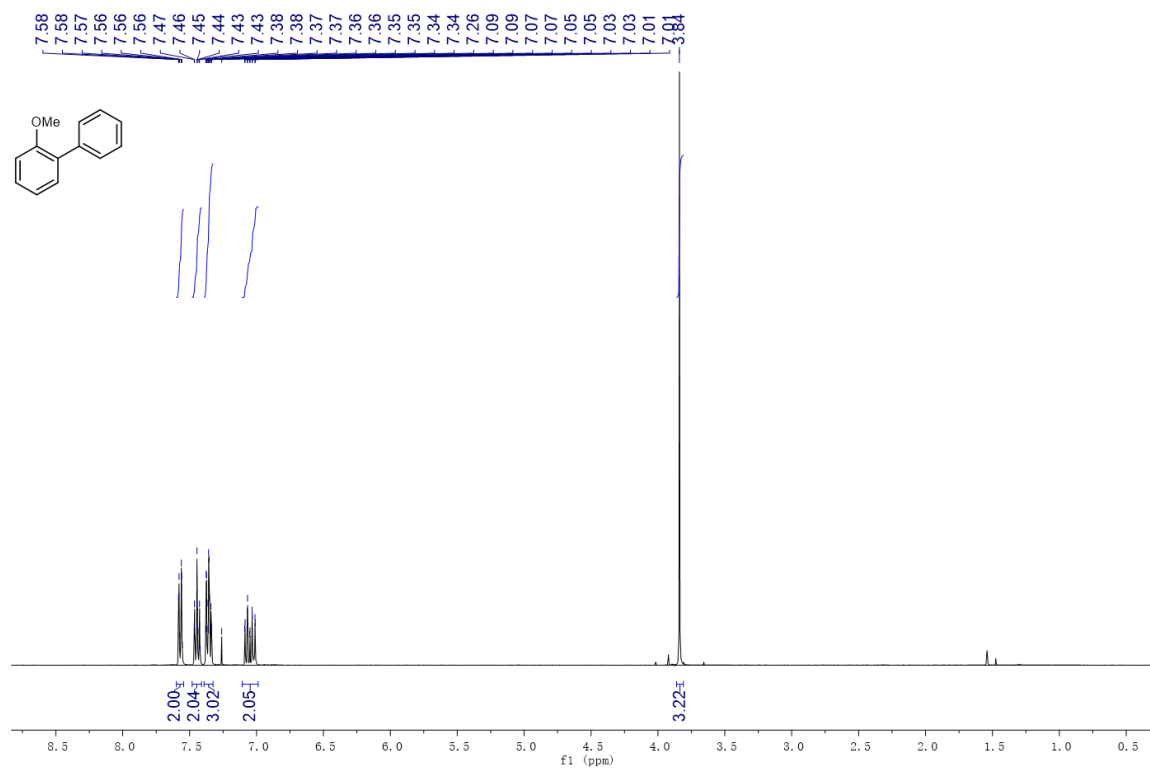
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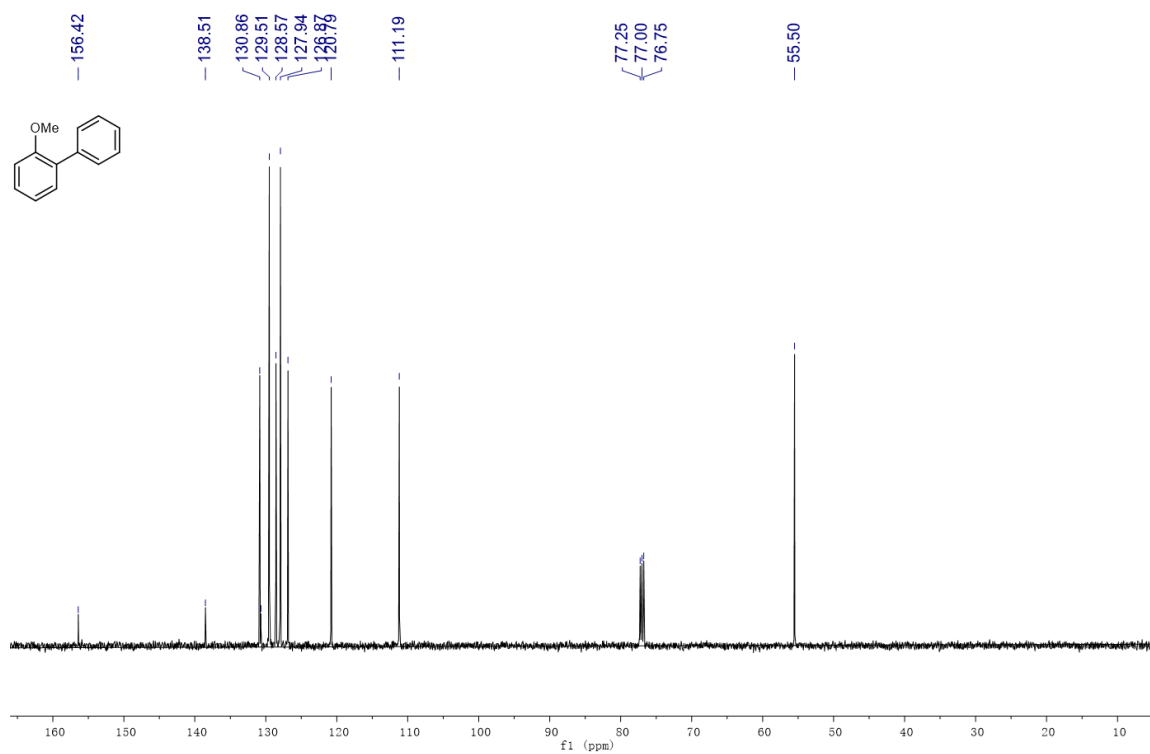
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1142 ¹H and ¹³C-NMR spectra of product 4s.

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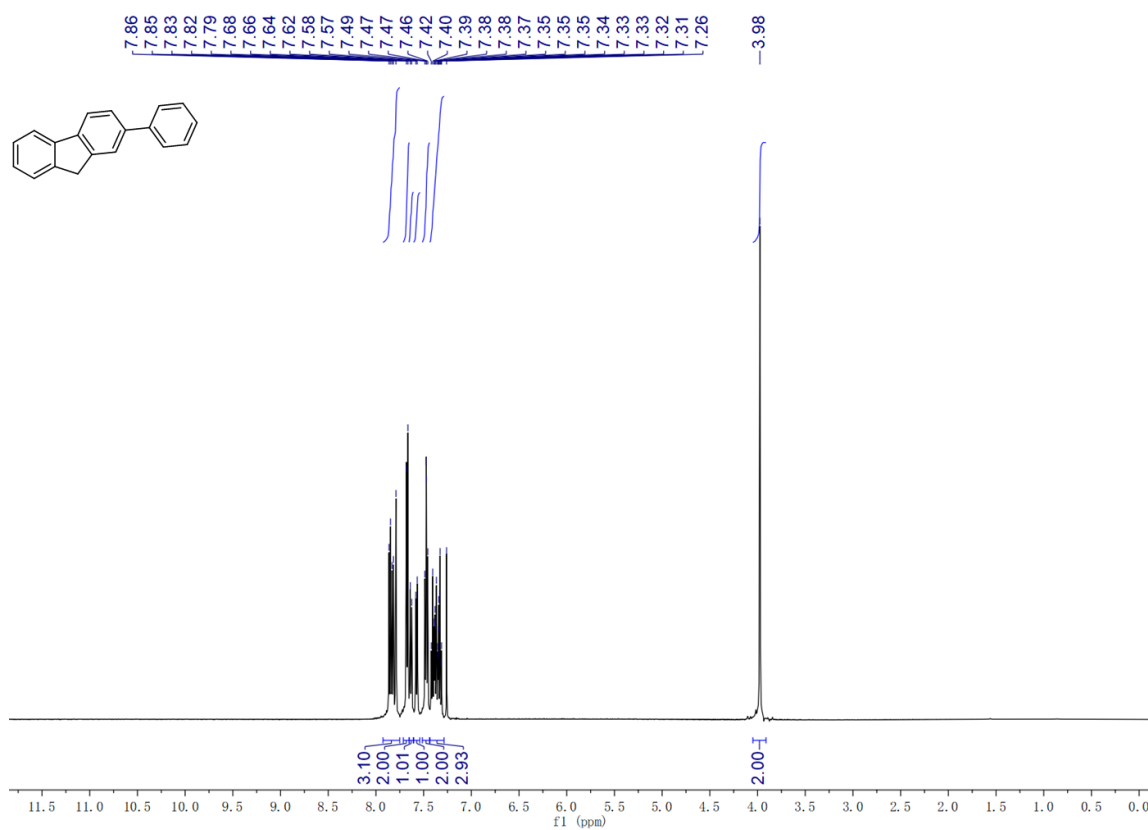


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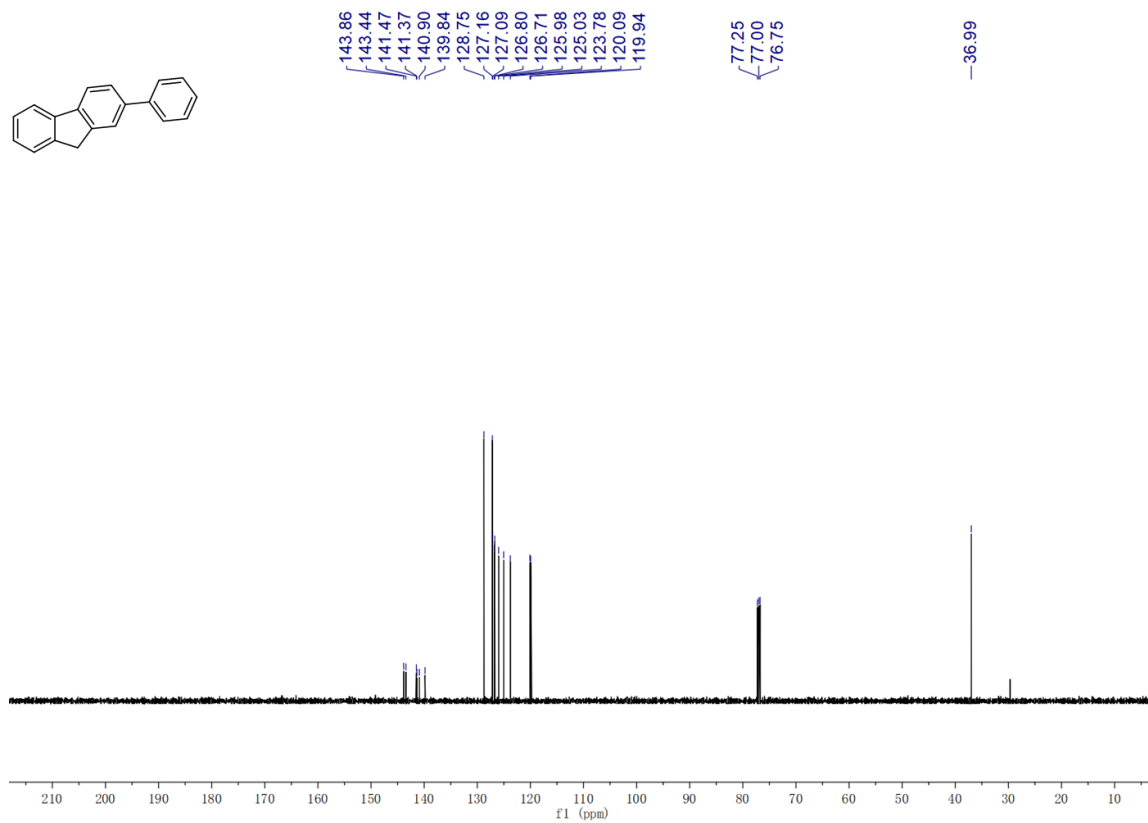
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1147 ¹H and ¹³C-NMR spectra of product 4t.

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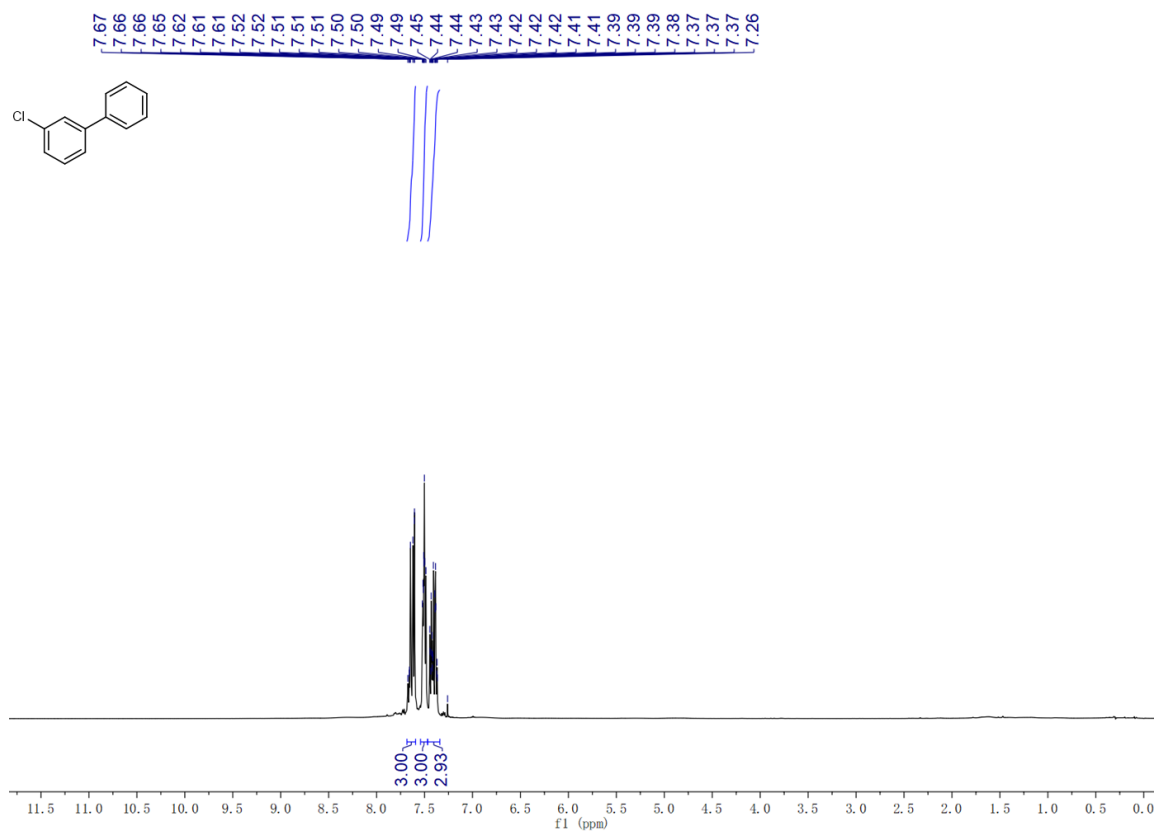


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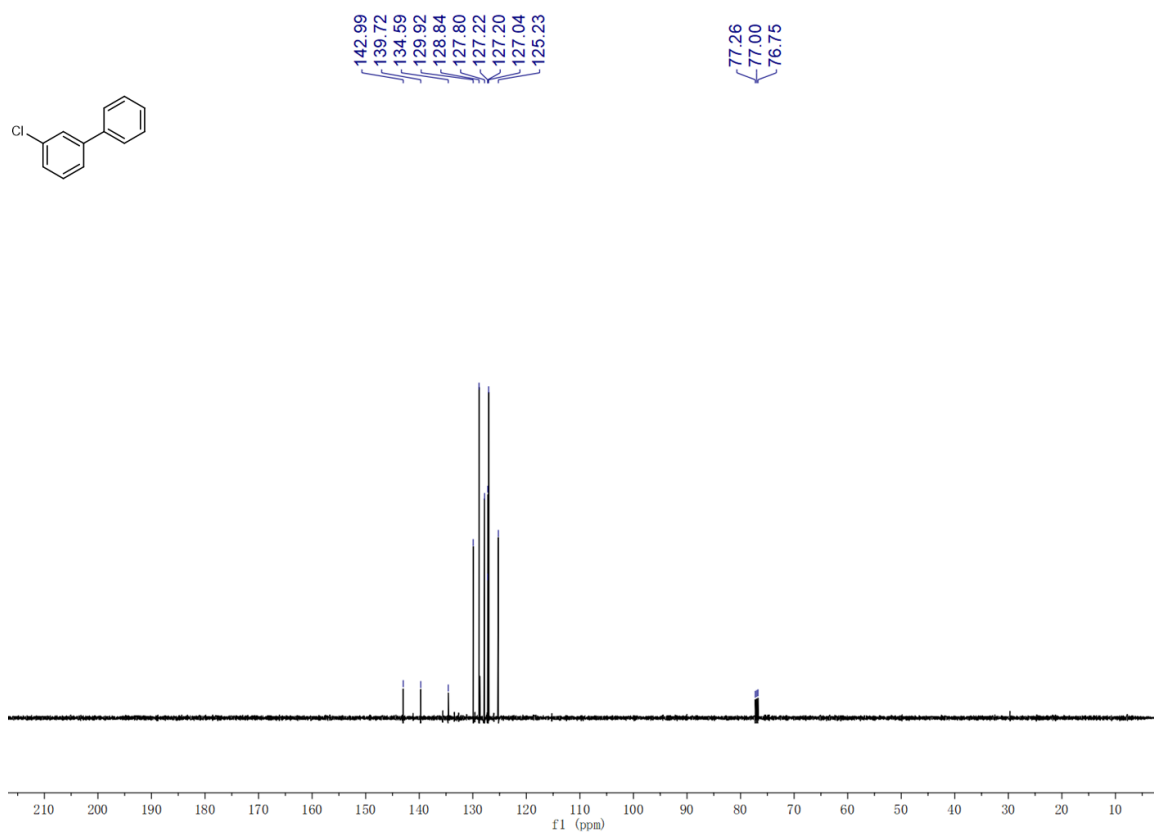
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1152 **¹H and ¹³C-NMR spectra of product 4u.**

1153



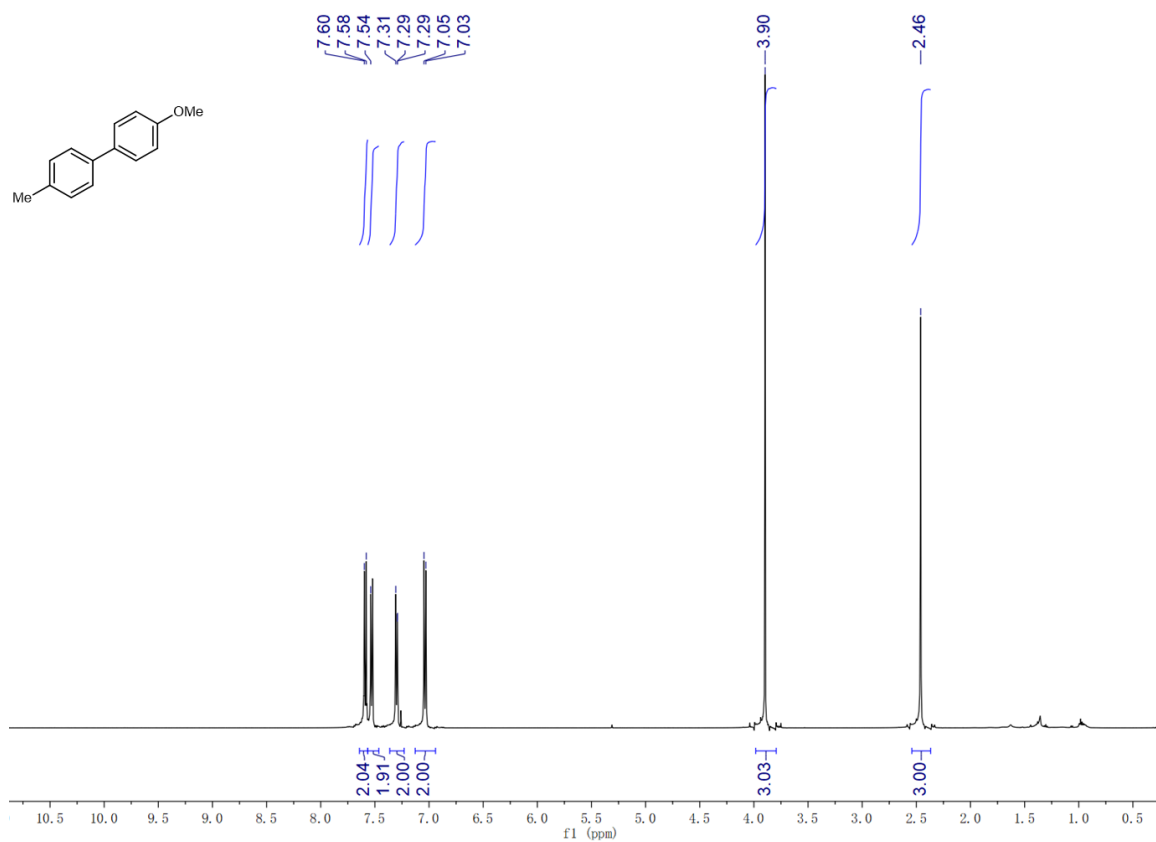
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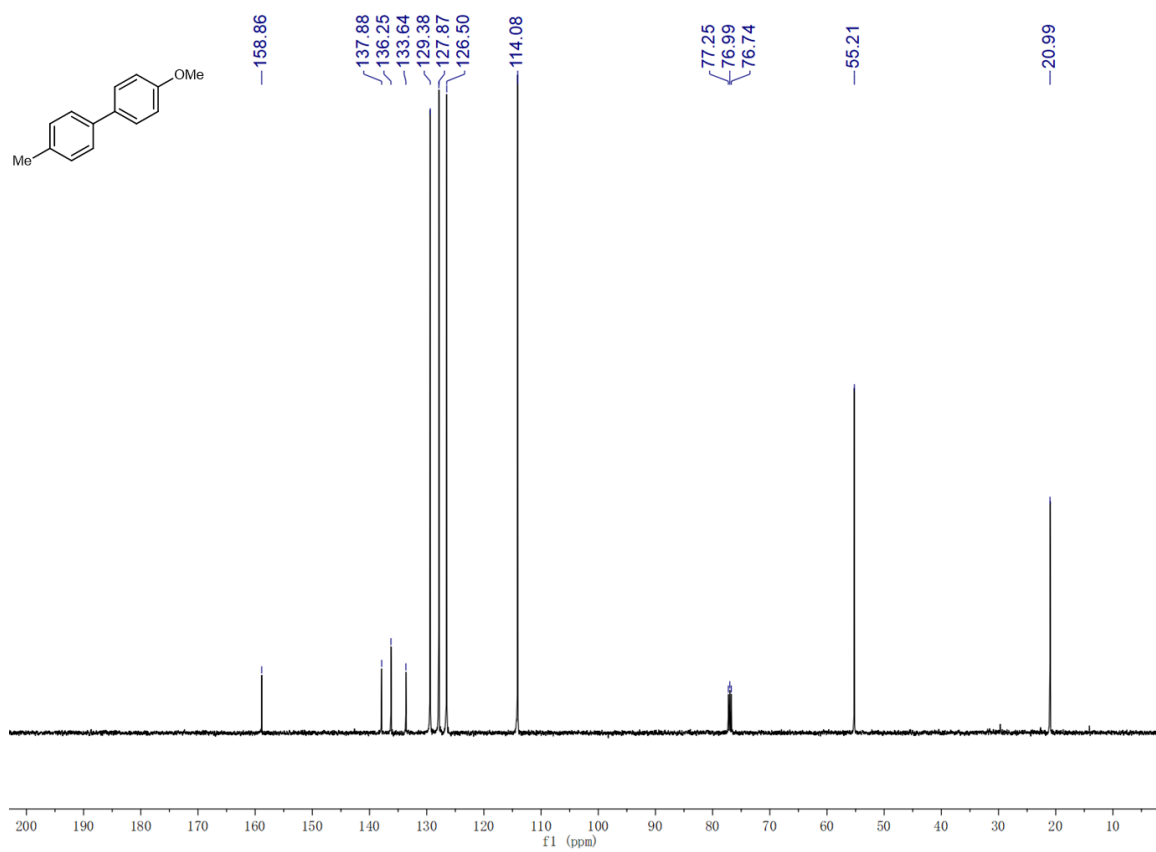
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1156 ¹H and ¹³C- NMR spectra of product 4v.

1157

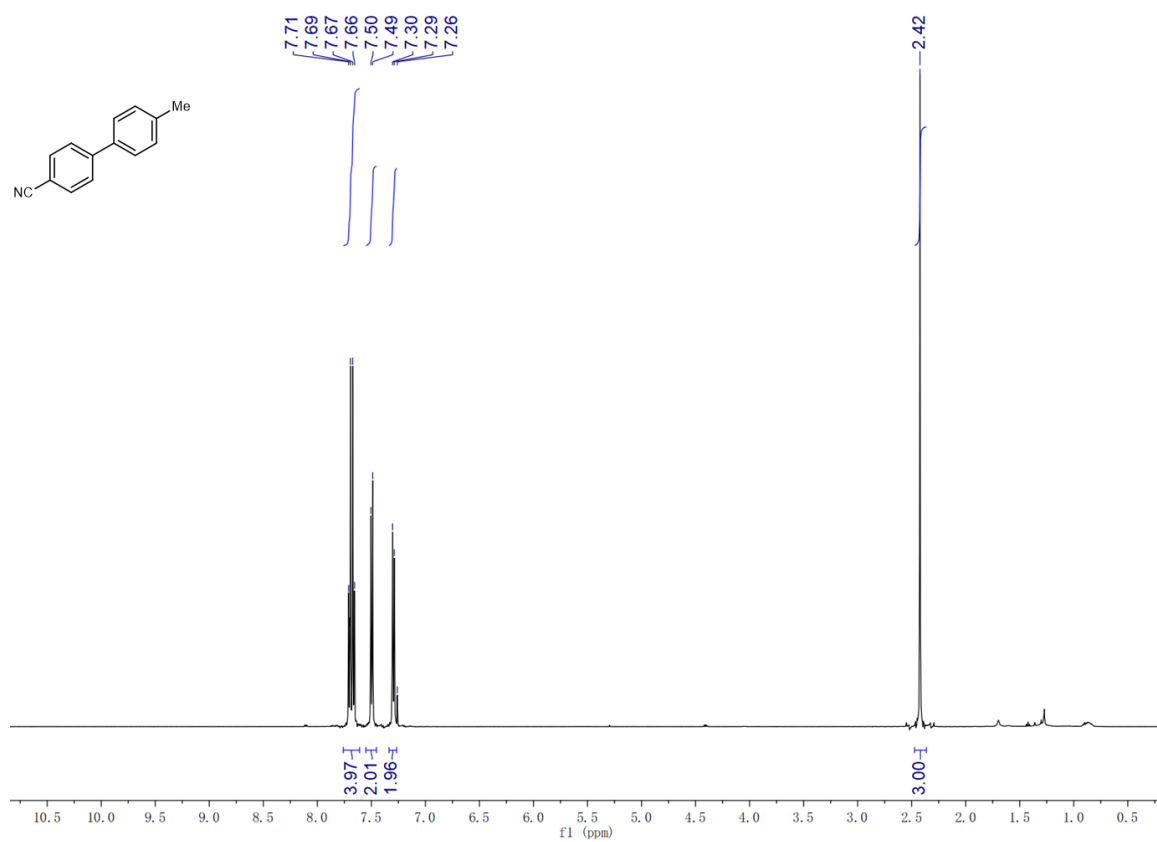


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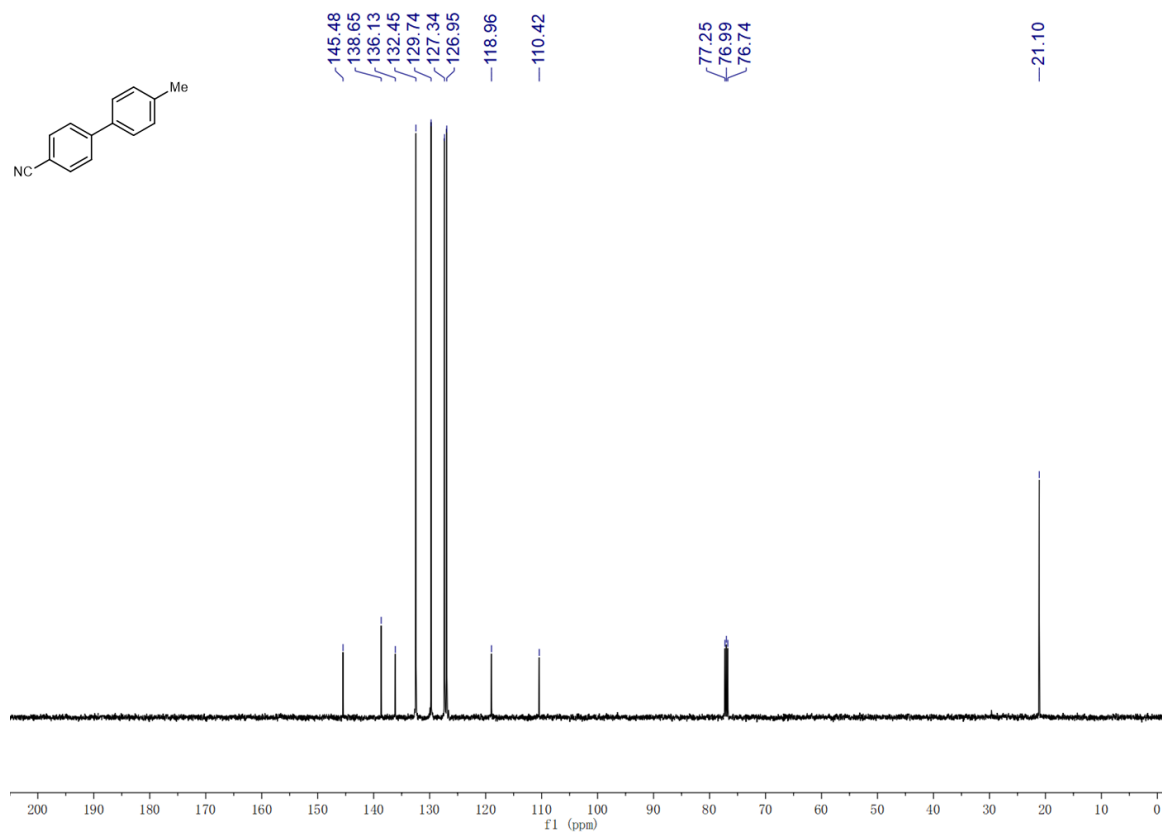
1159 ¹H and ¹³C-NMR spectra of product 4w.

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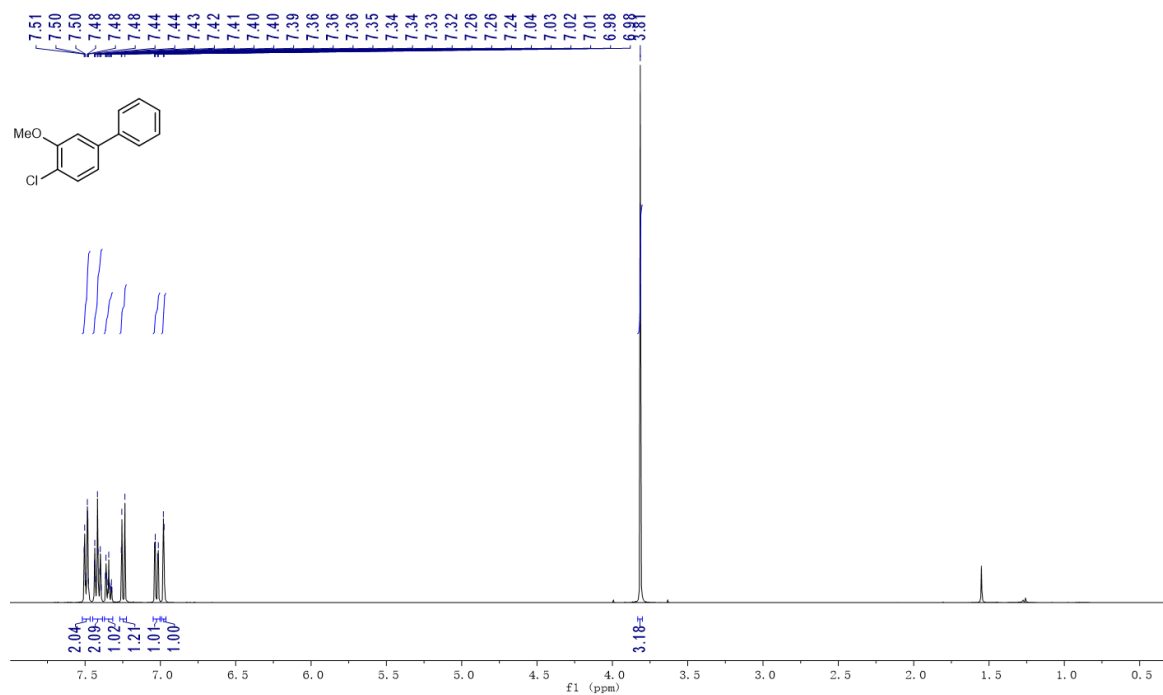
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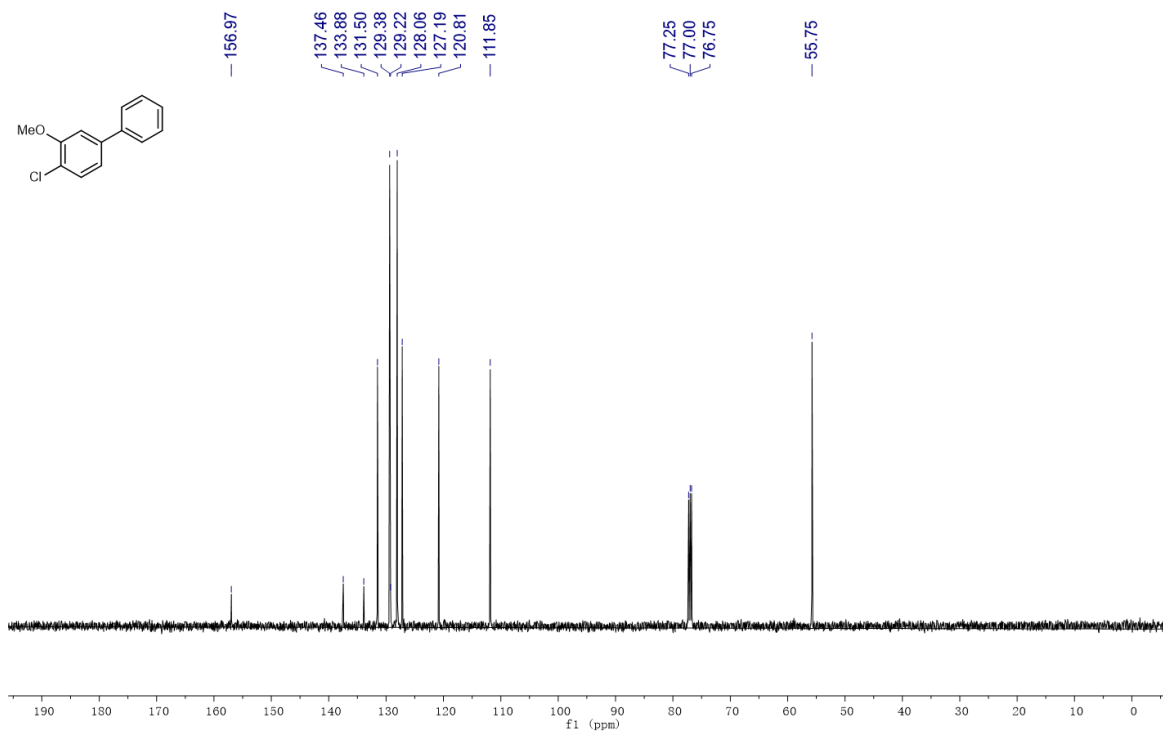


1163 ¹H and ¹³C-NMR spectra of product 4x.

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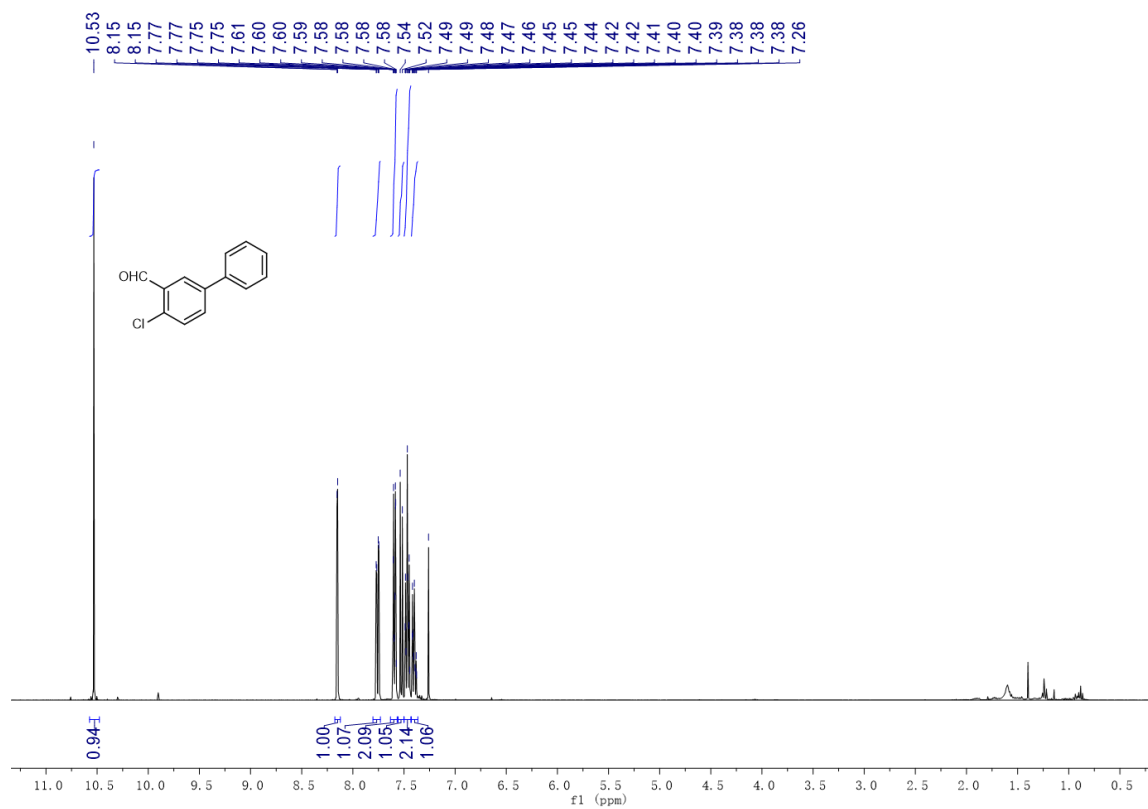


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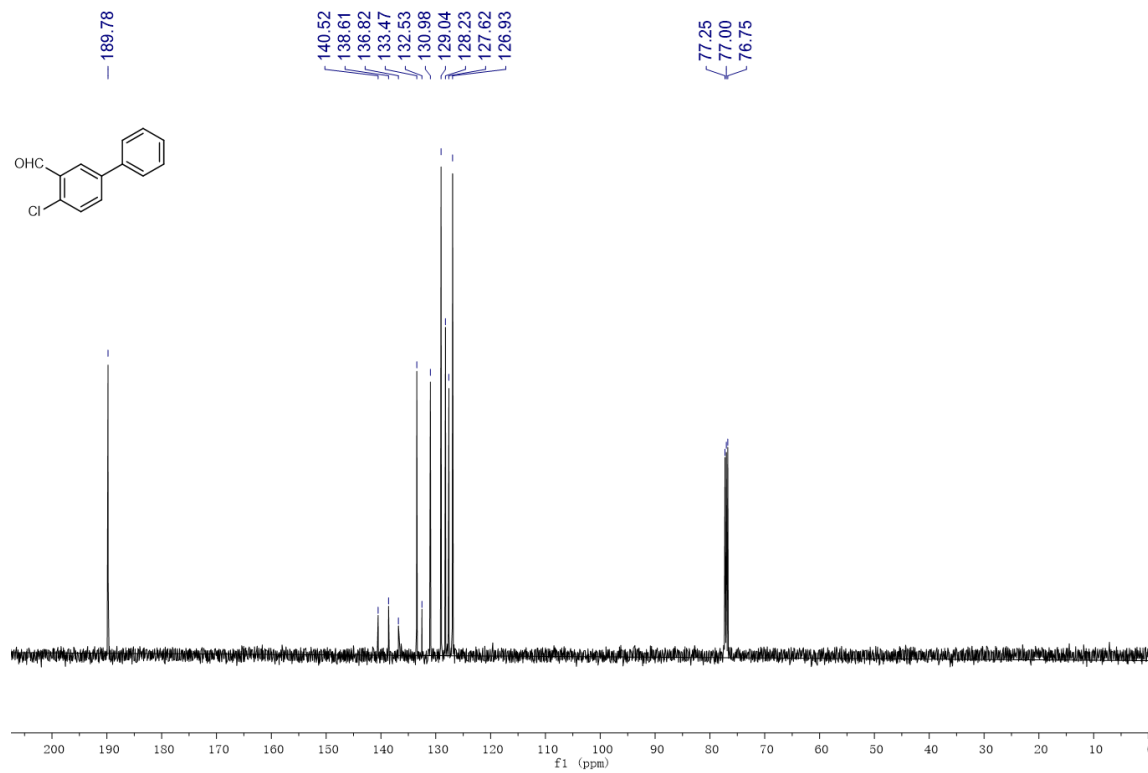
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1168 ¹H and ¹³C-NMR spectra of product 4y.

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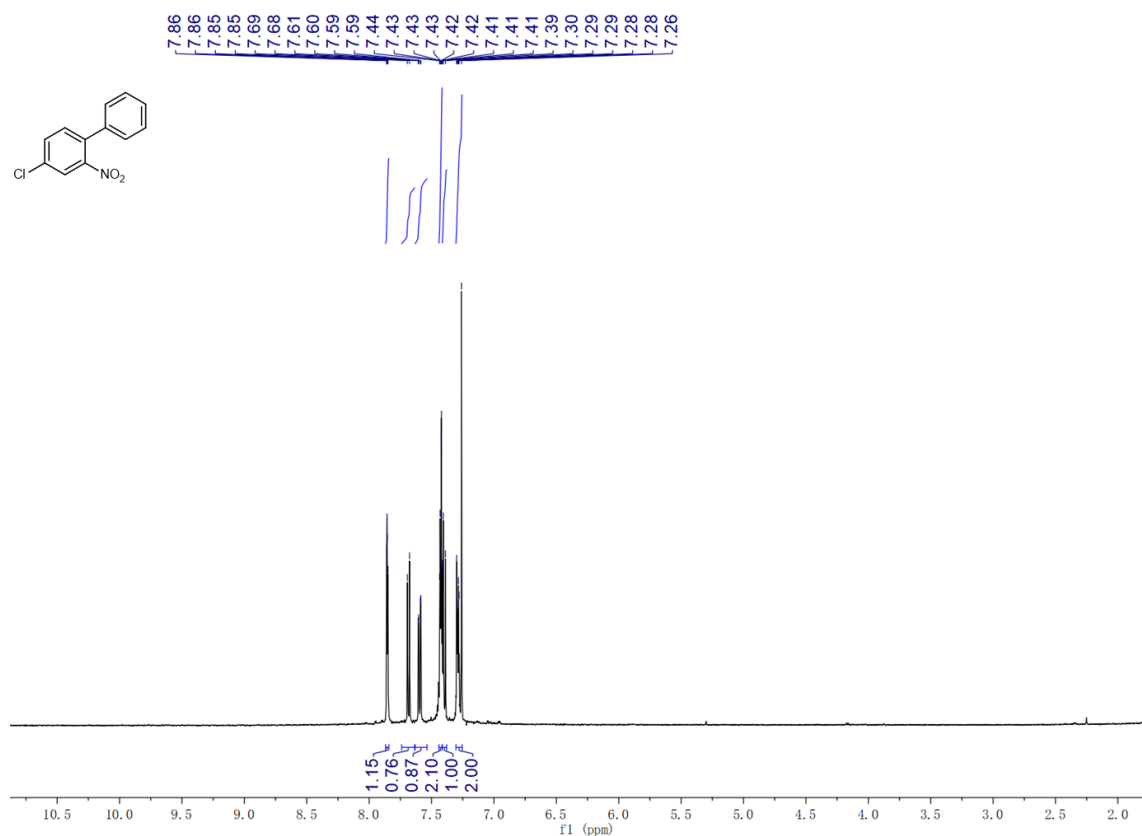
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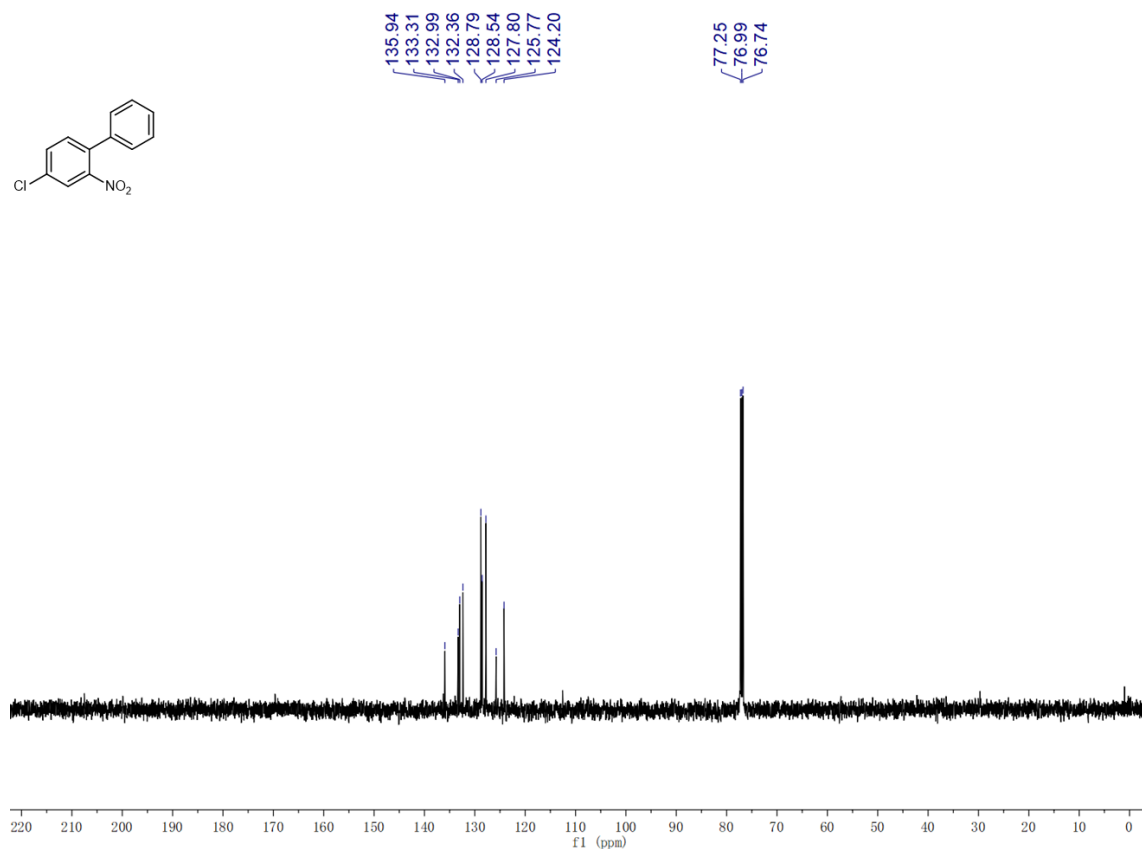
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1174 ¹H and ¹³C-NMR spectra of product 4z.

1175



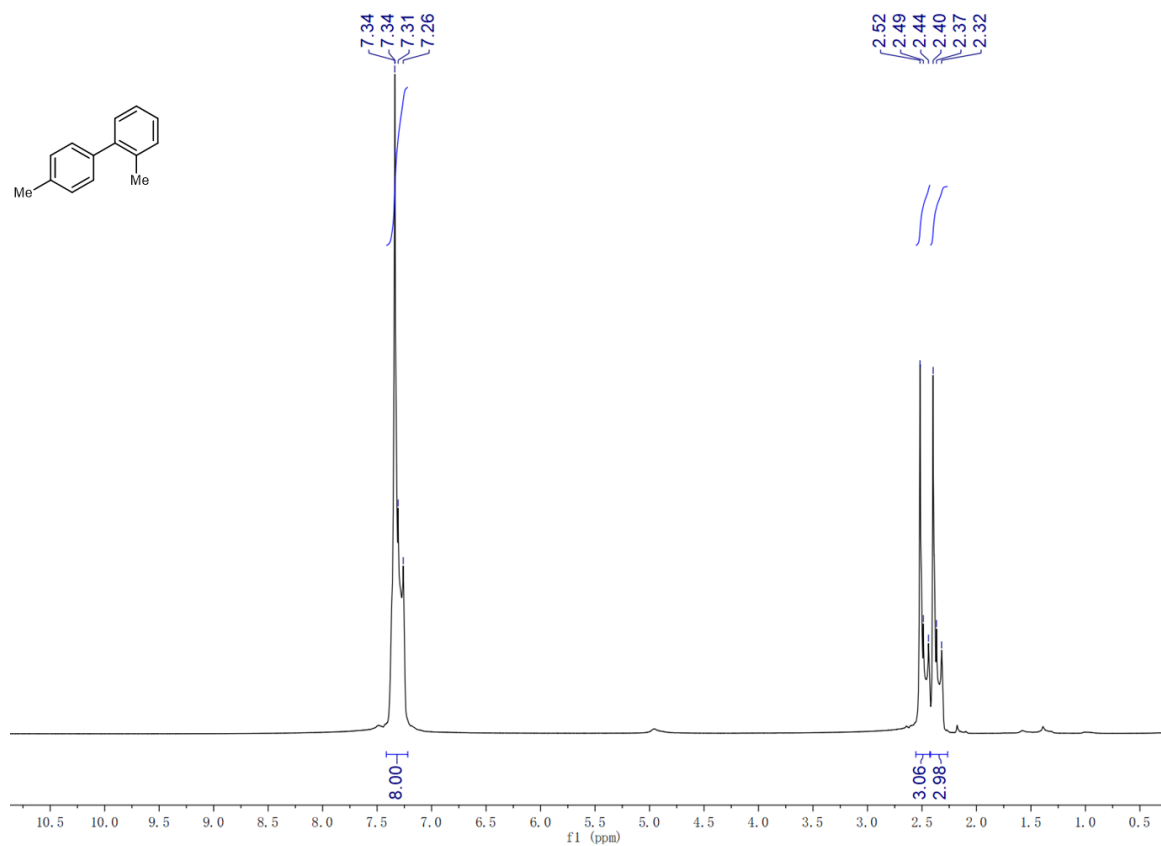
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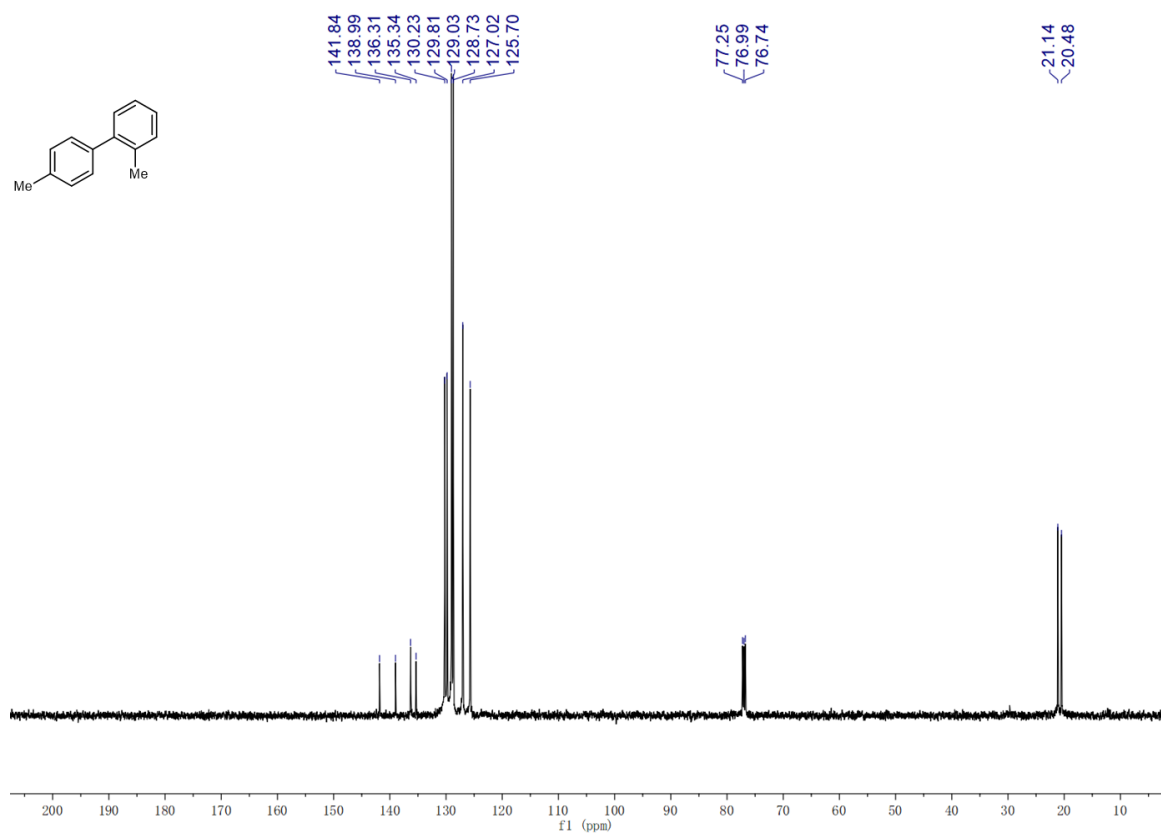
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1178 ¹H and ¹³C-NMR spectra of product 4aa.

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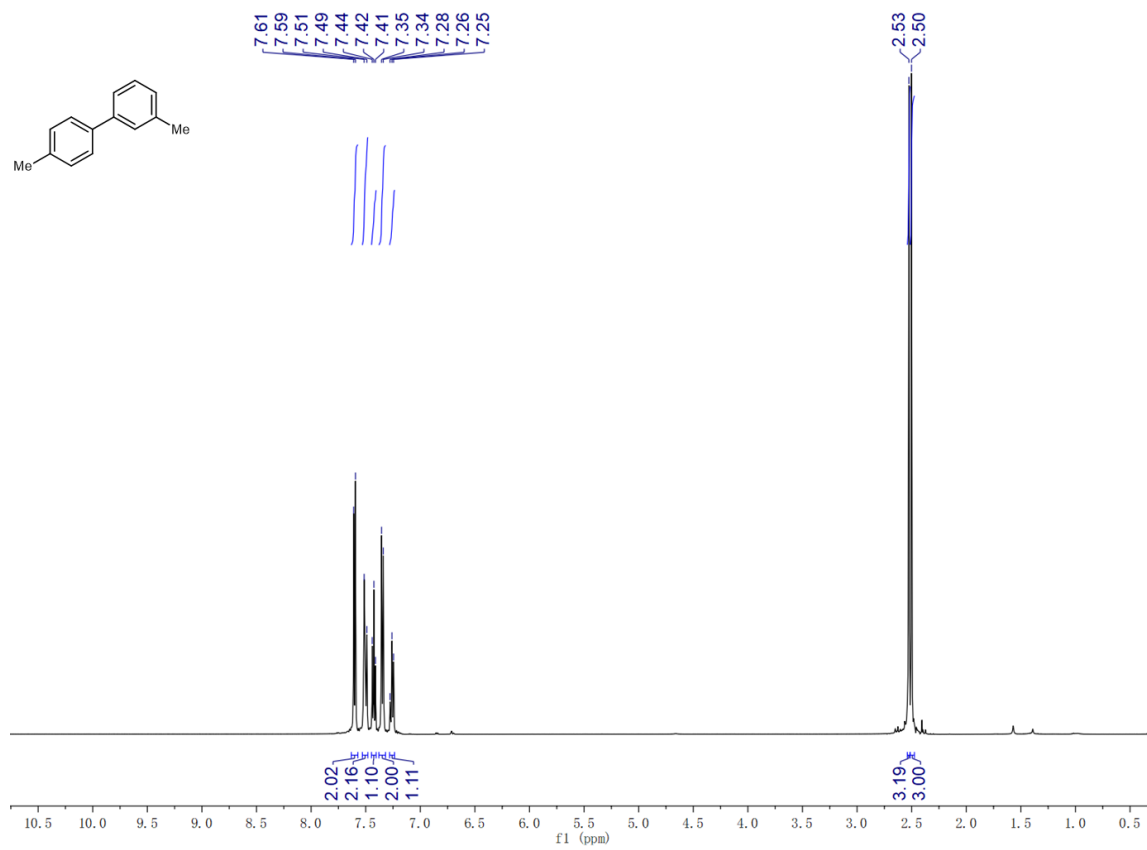


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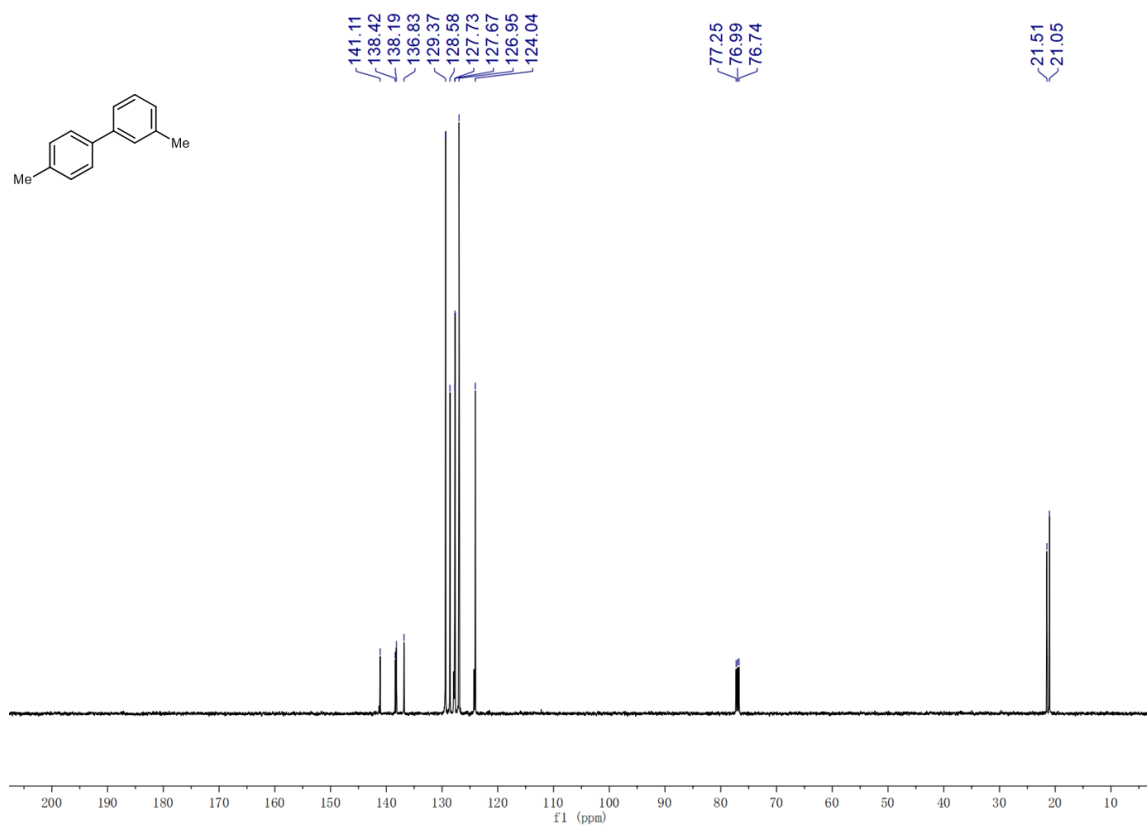


1181 ¹H and ¹³C-NMR spectra of product 4ab.

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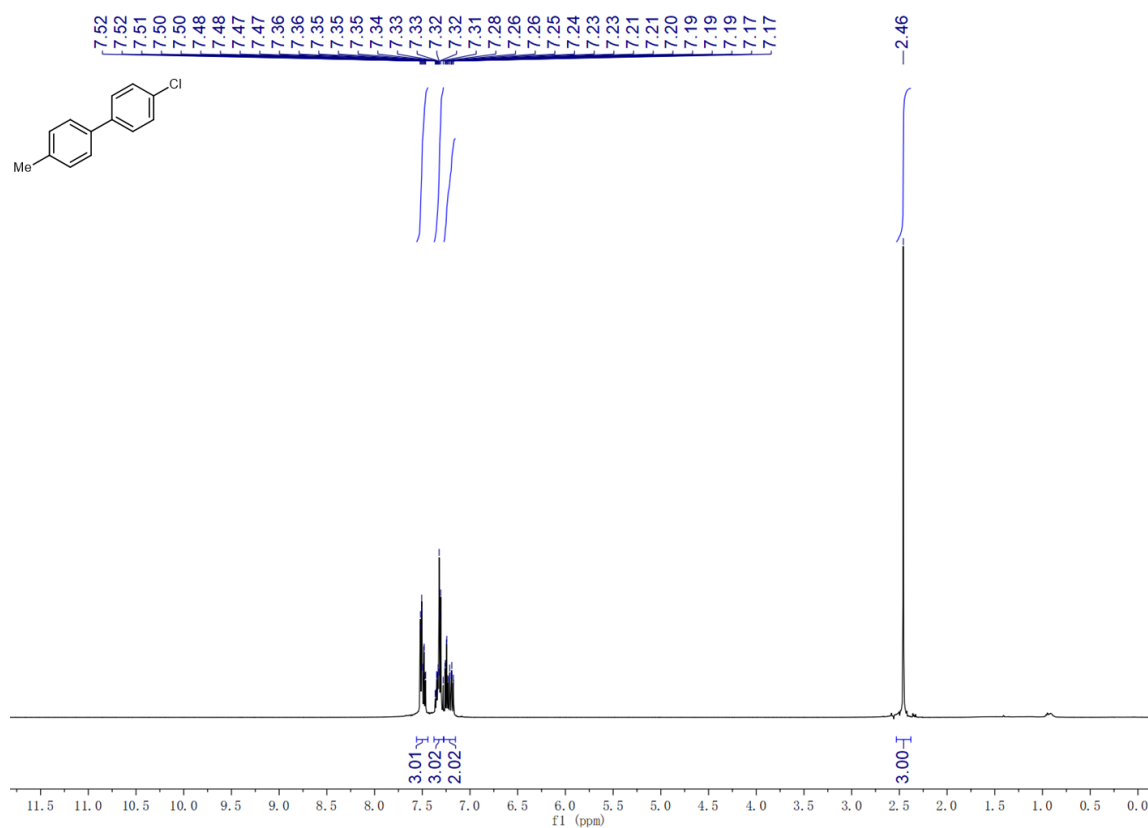
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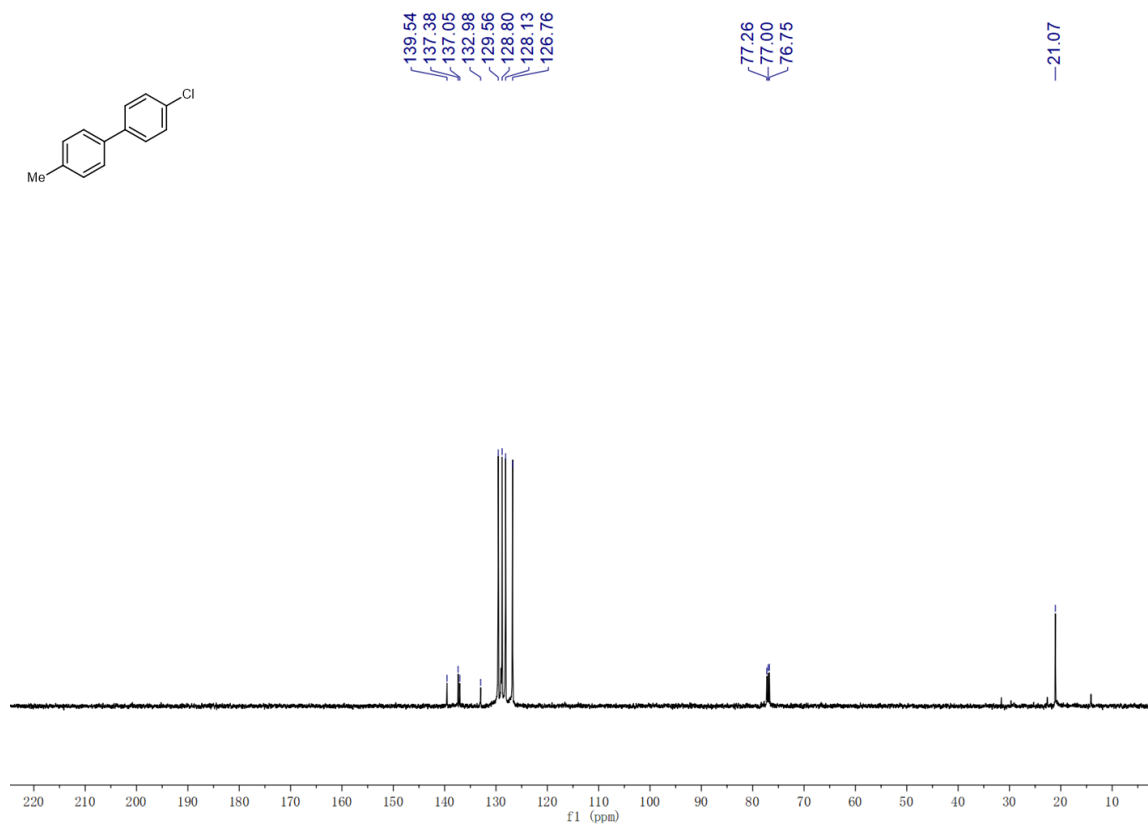
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1185 ¹H and ¹³C-NMR spectra of product 4ac.

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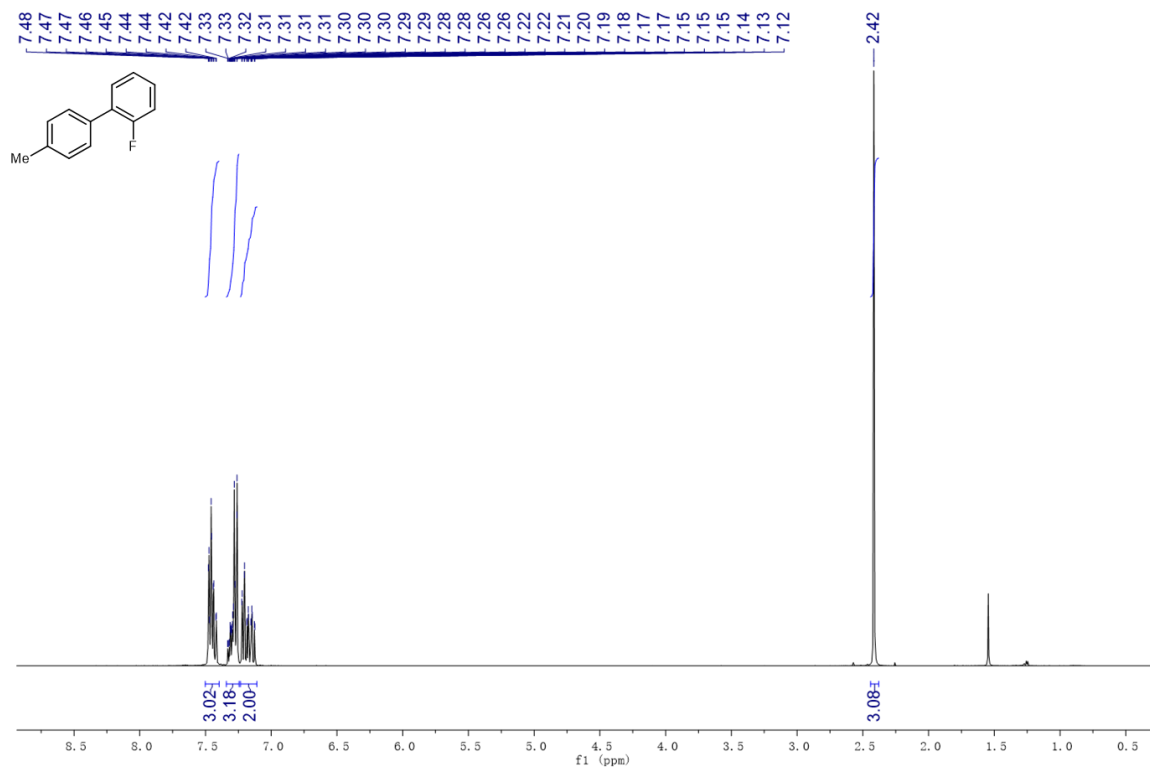


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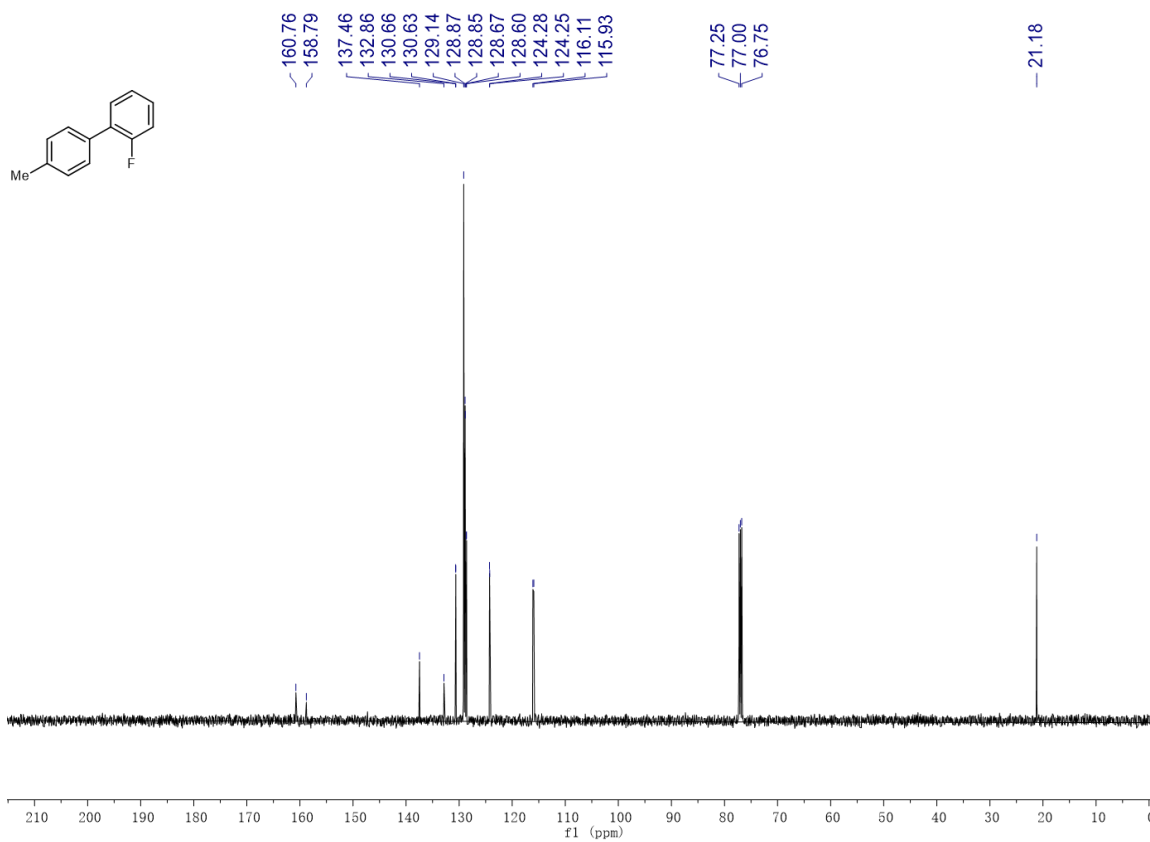
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1190 ¹H and ¹³C, ¹⁹F-NMR spectra of product 4ad.

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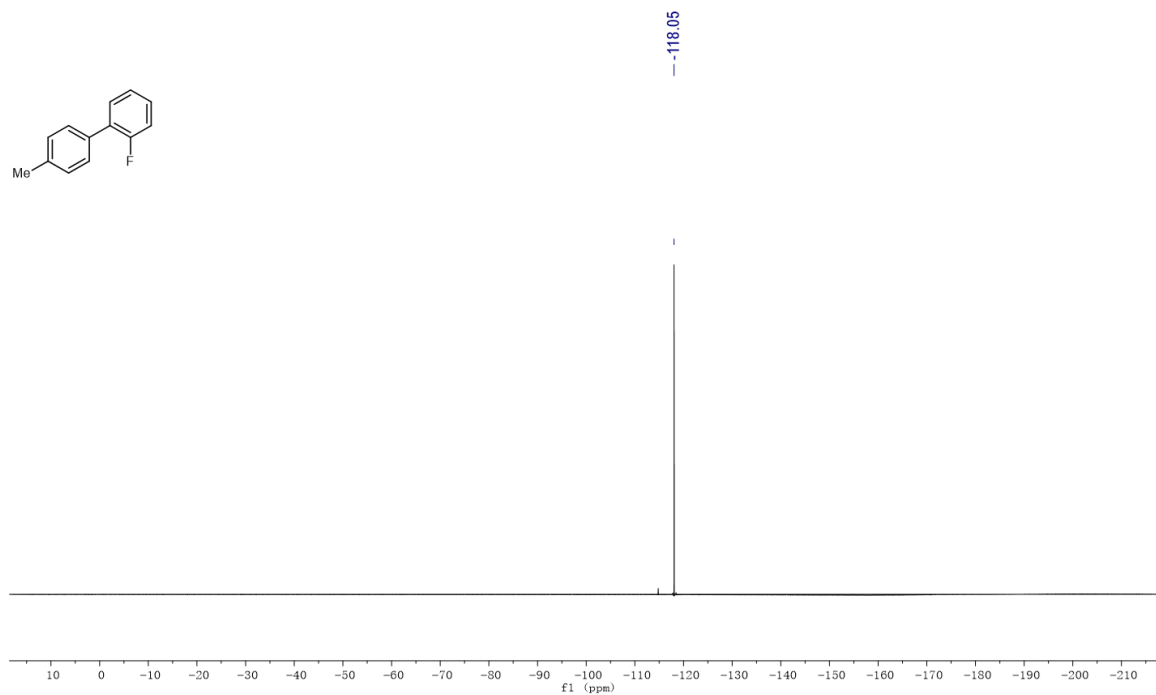
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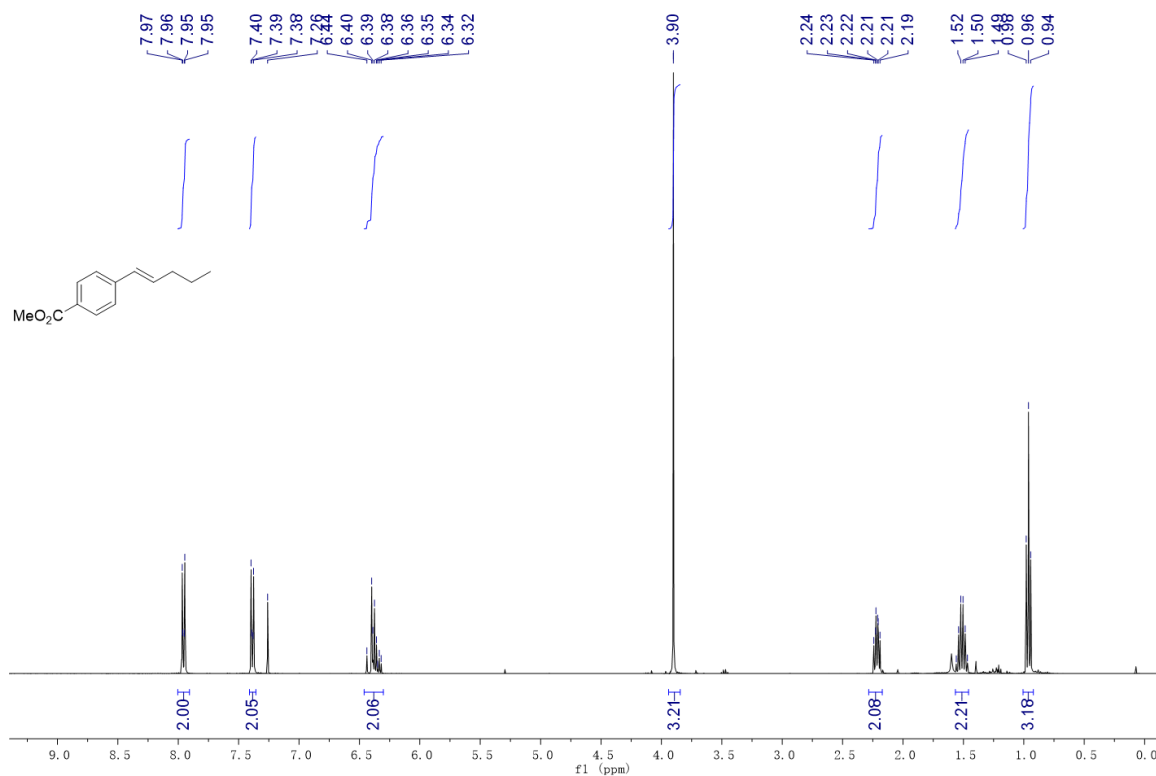


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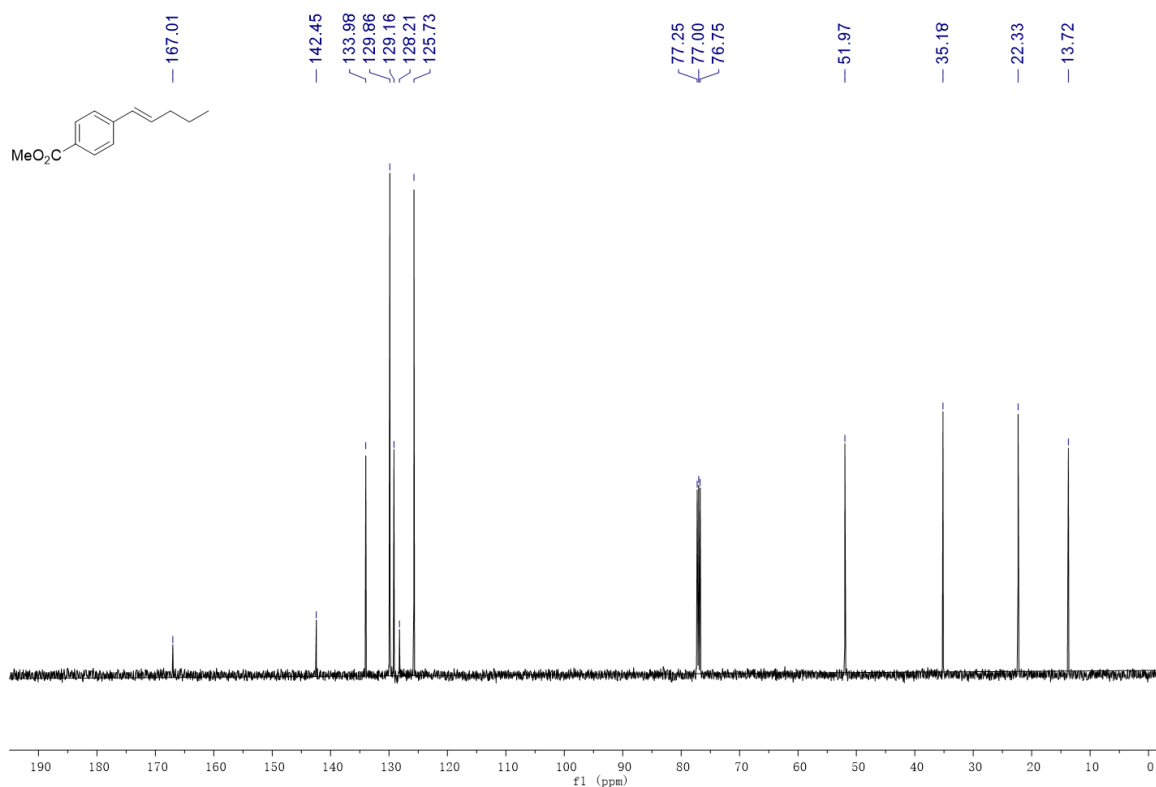
1198 ¹H and ¹³C-NMR spectra of product 5a.

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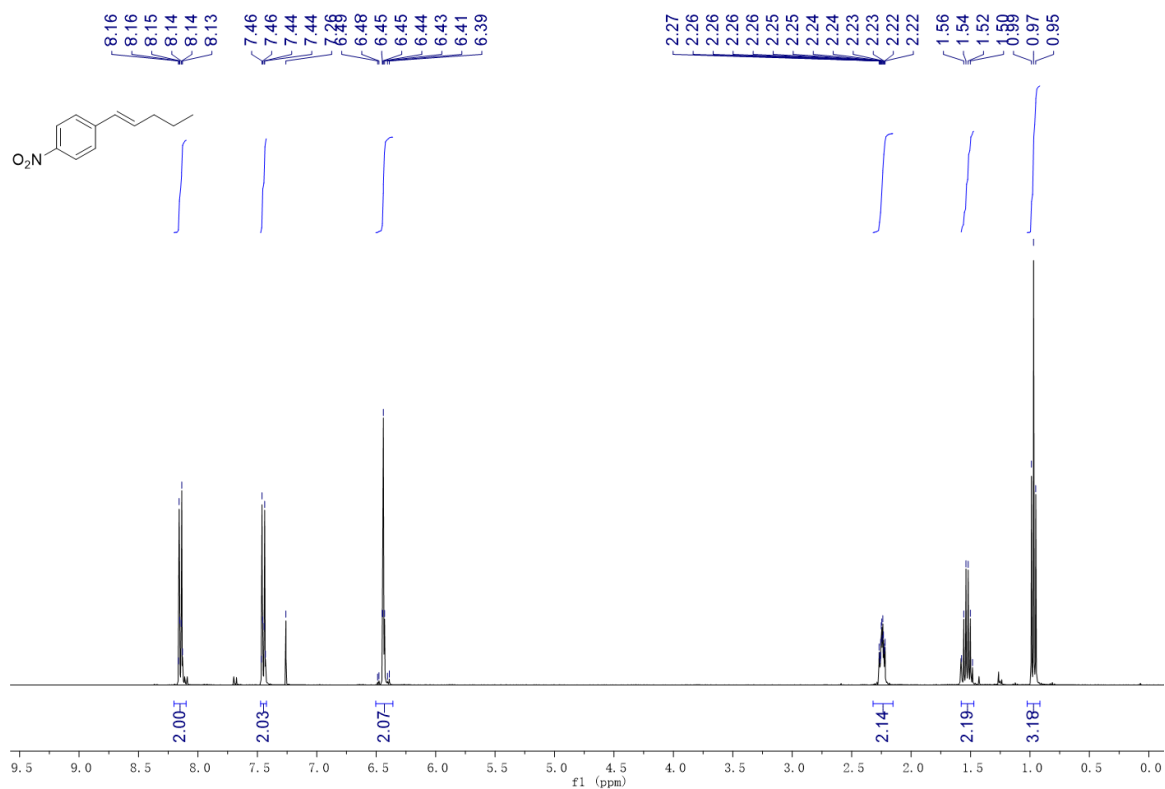
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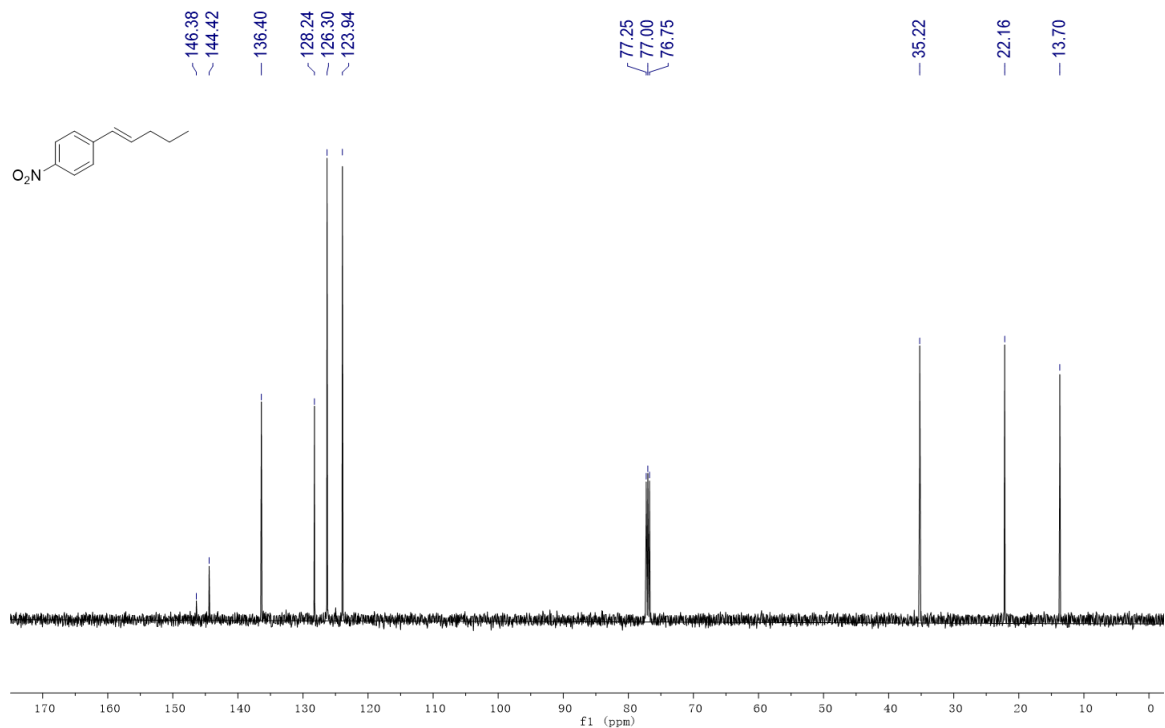
1205 ¹H and ¹³C-NMR spectra of product 5b.

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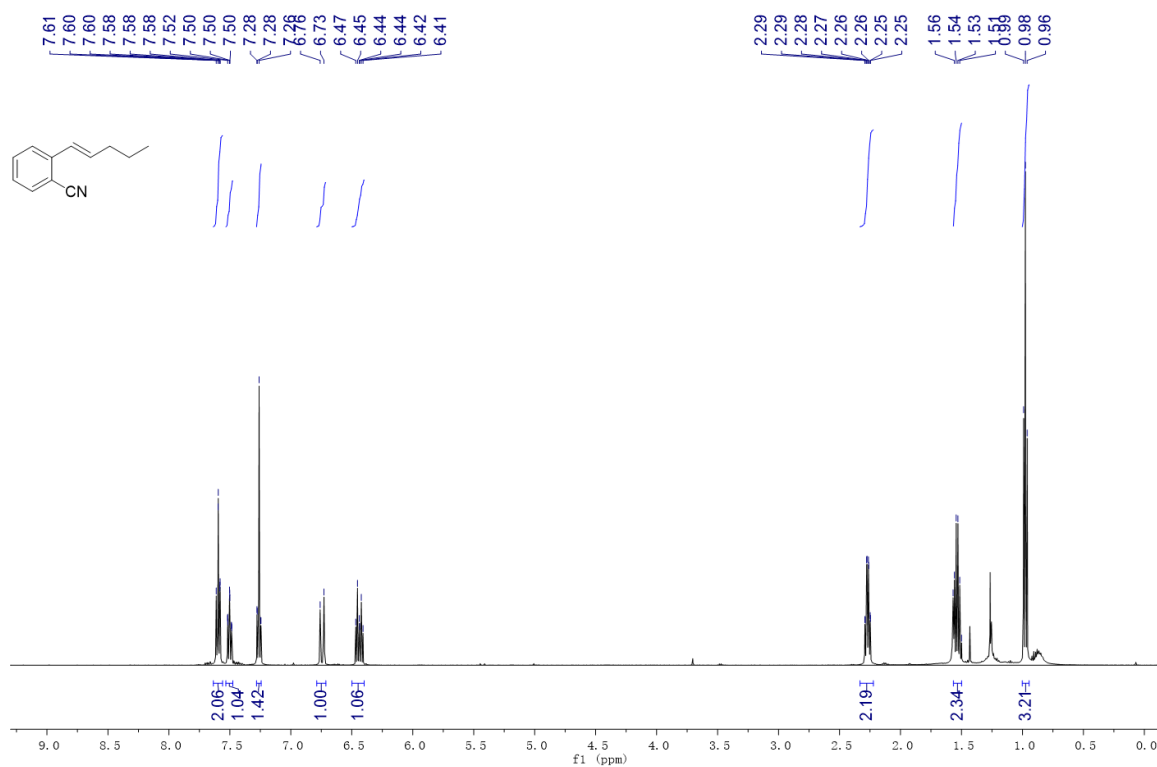


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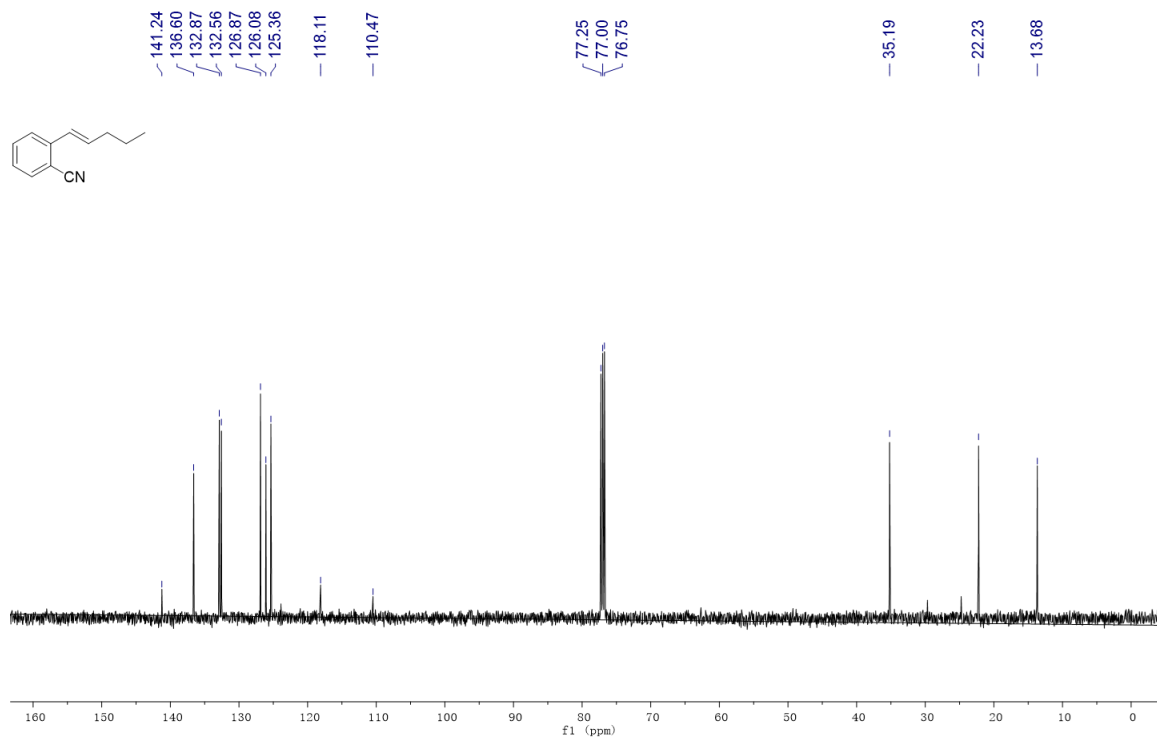
1211 ¹H and ¹³C-NMR spectra of product 5c.

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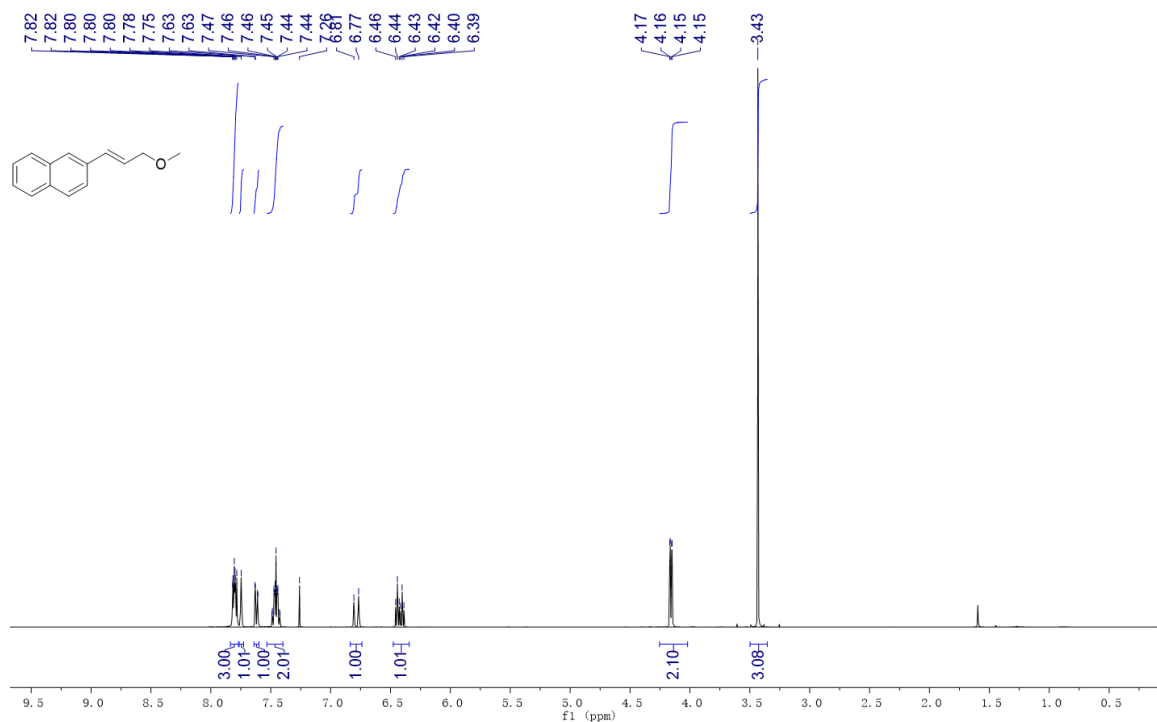


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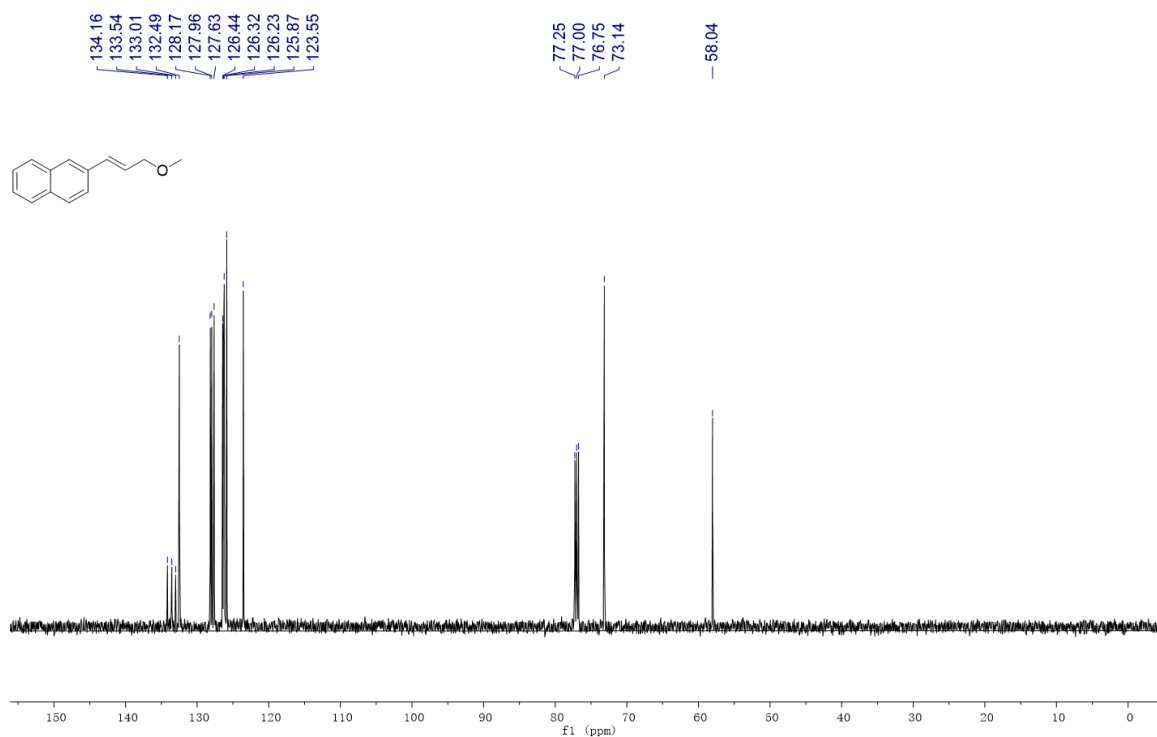
1217 ¹H and ¹³C-NMR spectra of product 5d.

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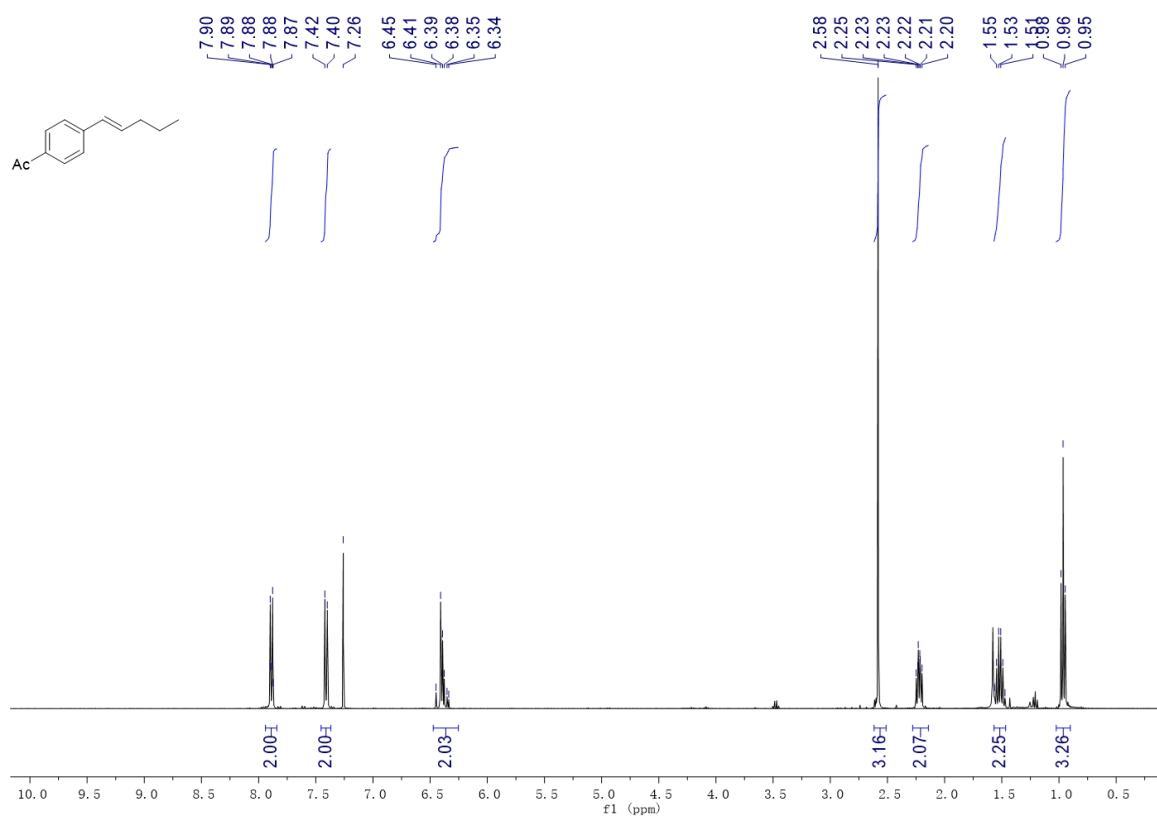
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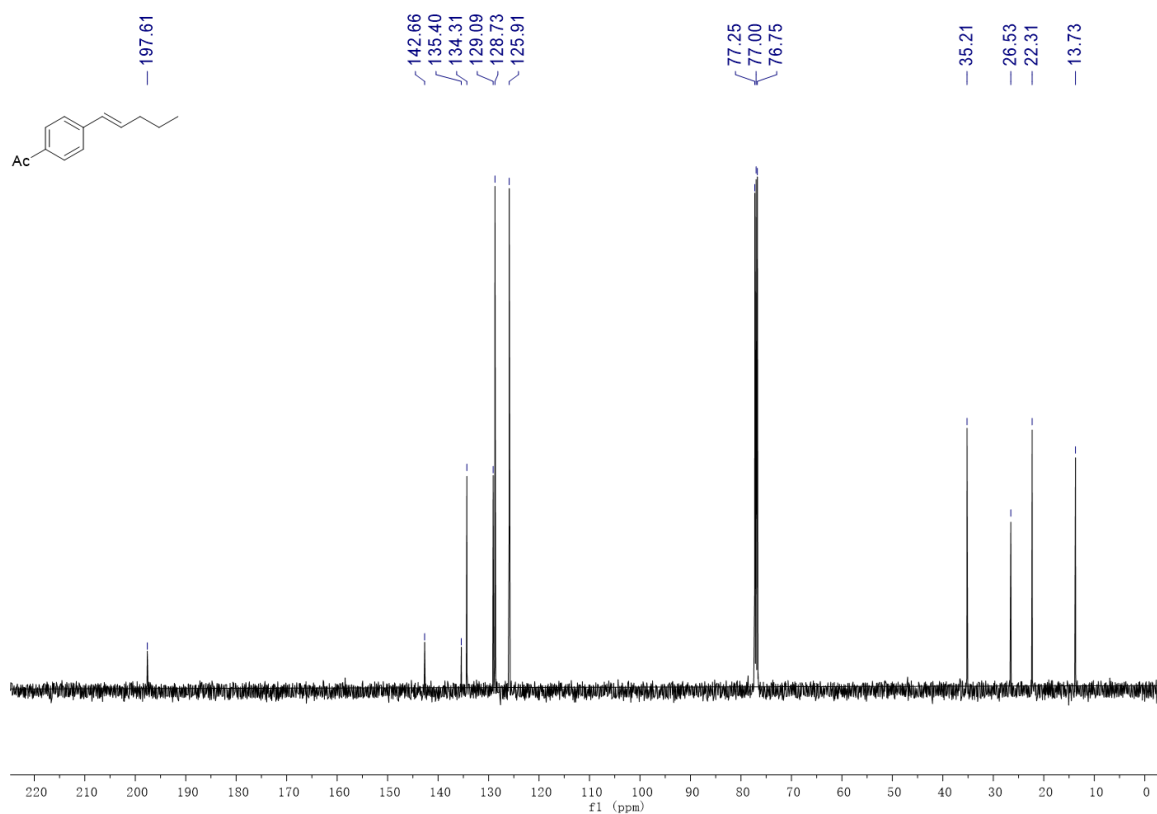
1222 ¹H and ¹³C-NMR spectra of product 5e.

1223



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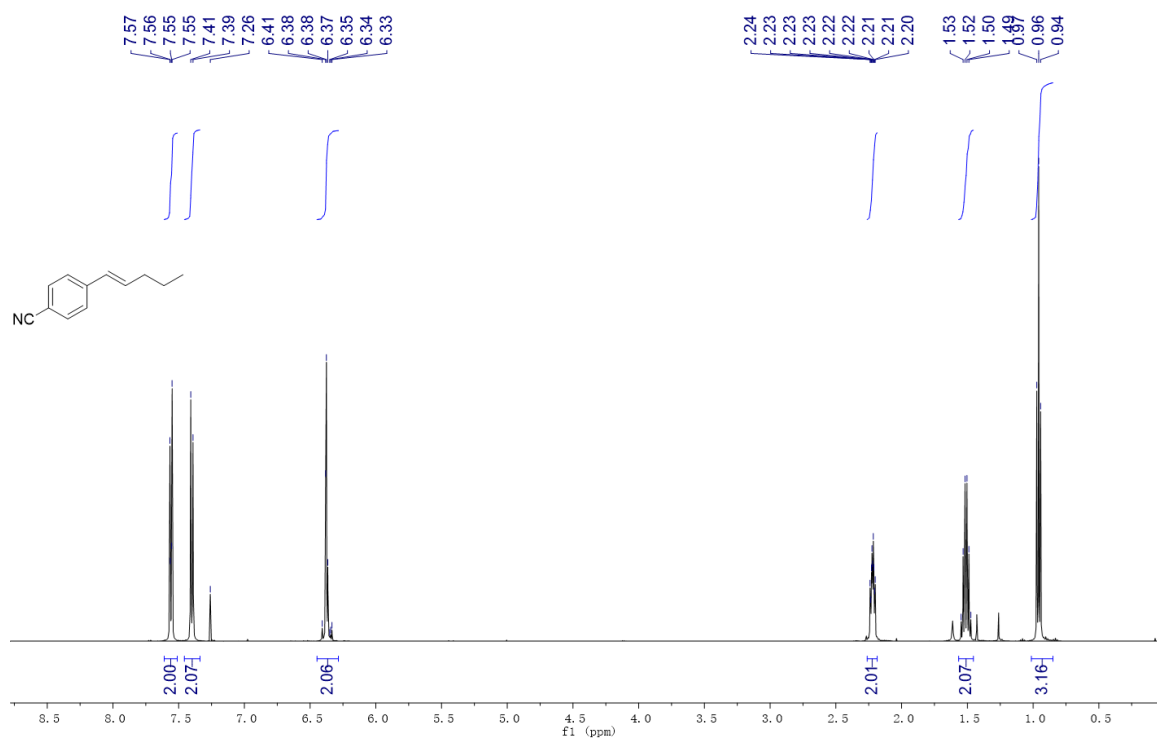
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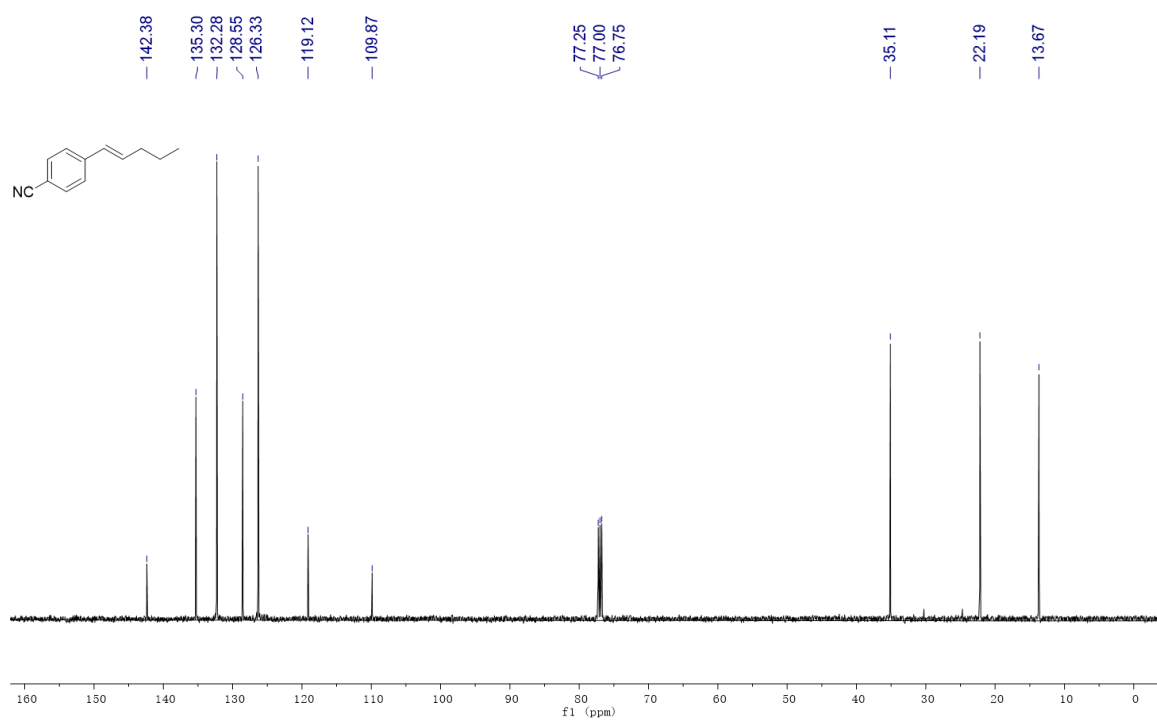
1227 **¹H and ¹³C-NMR spectra of product 5f.**

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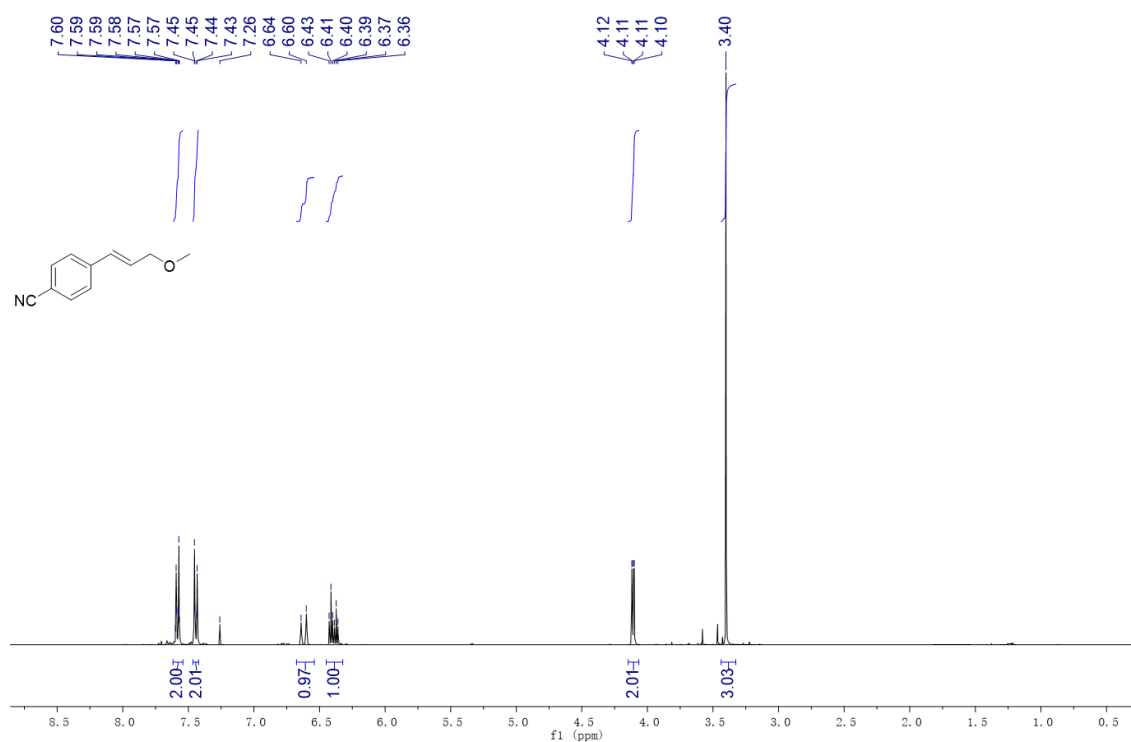
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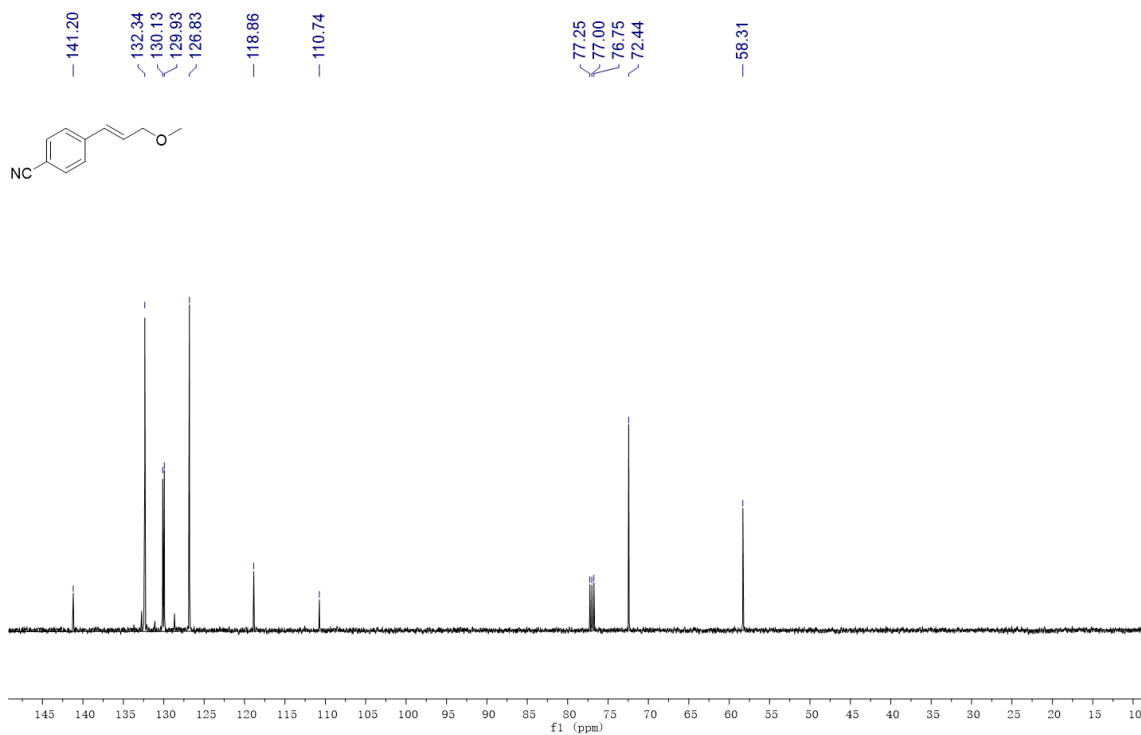
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1233 ¹H and ¹³C-NMR spectra of product 5g.



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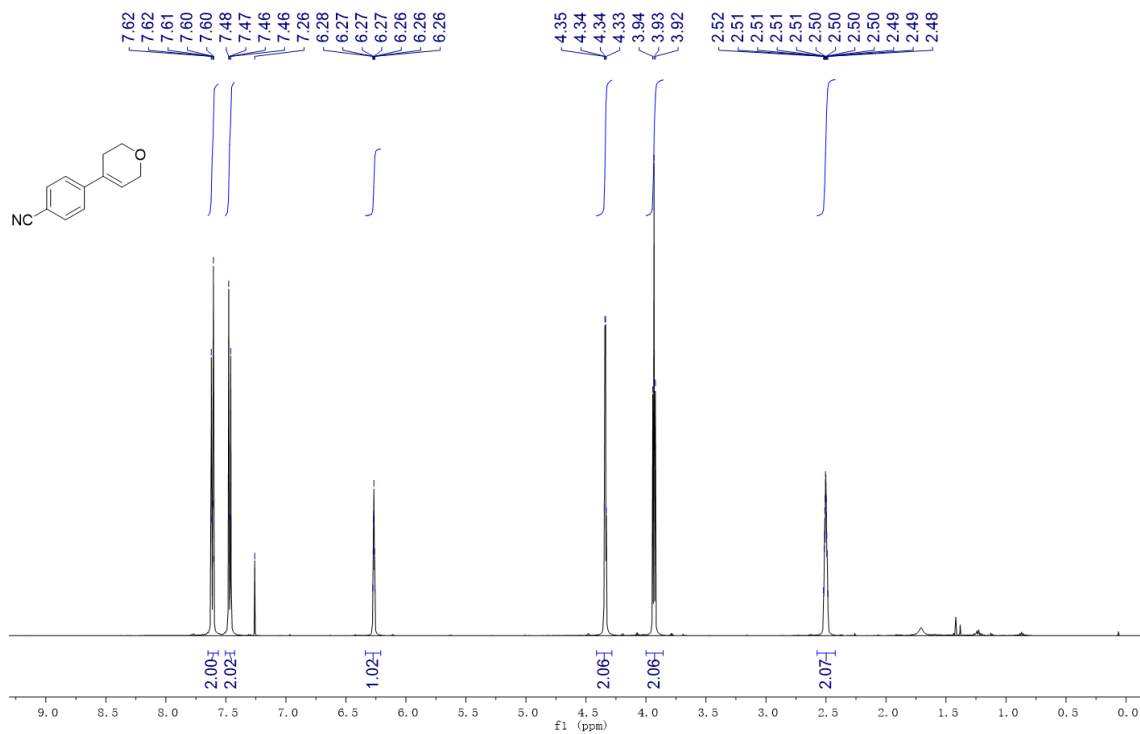


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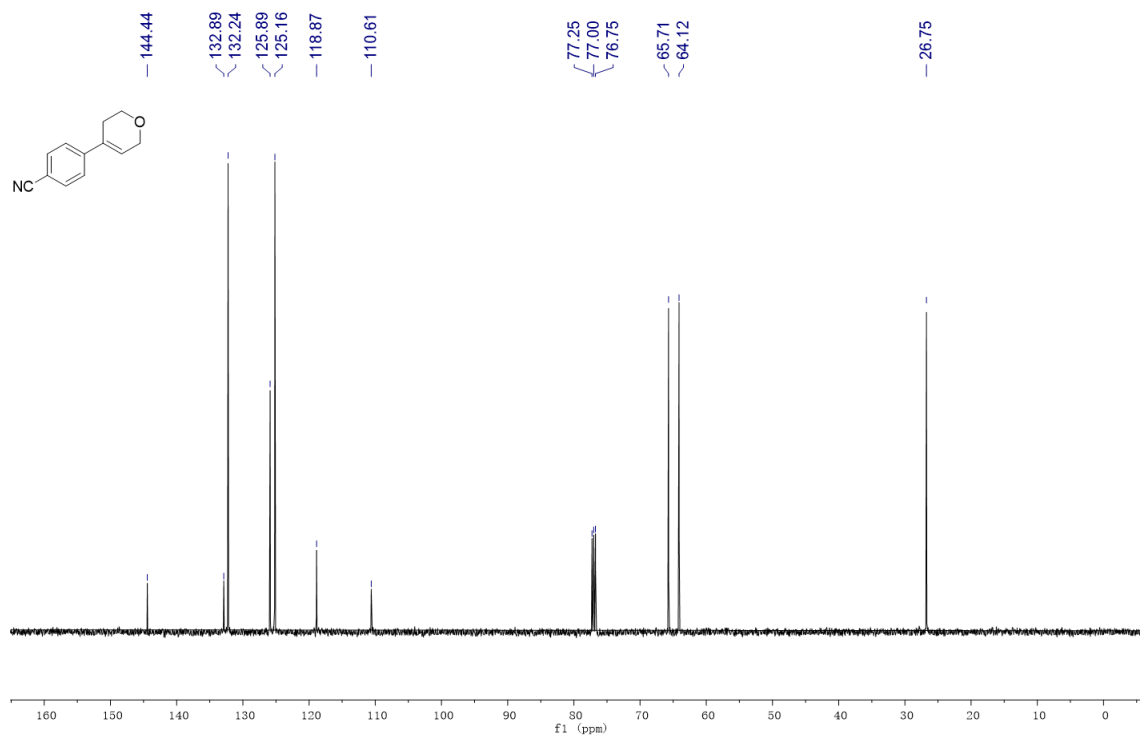
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1237 ¹H and ¹³C-NMR spectra of product 5h.

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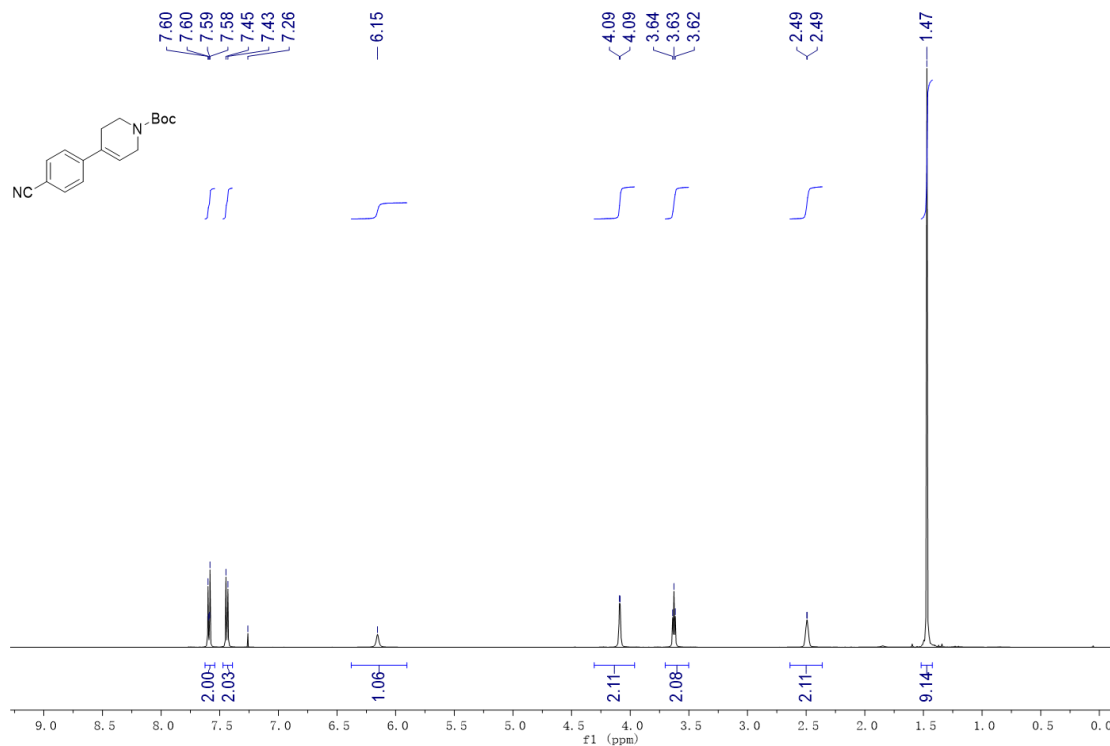


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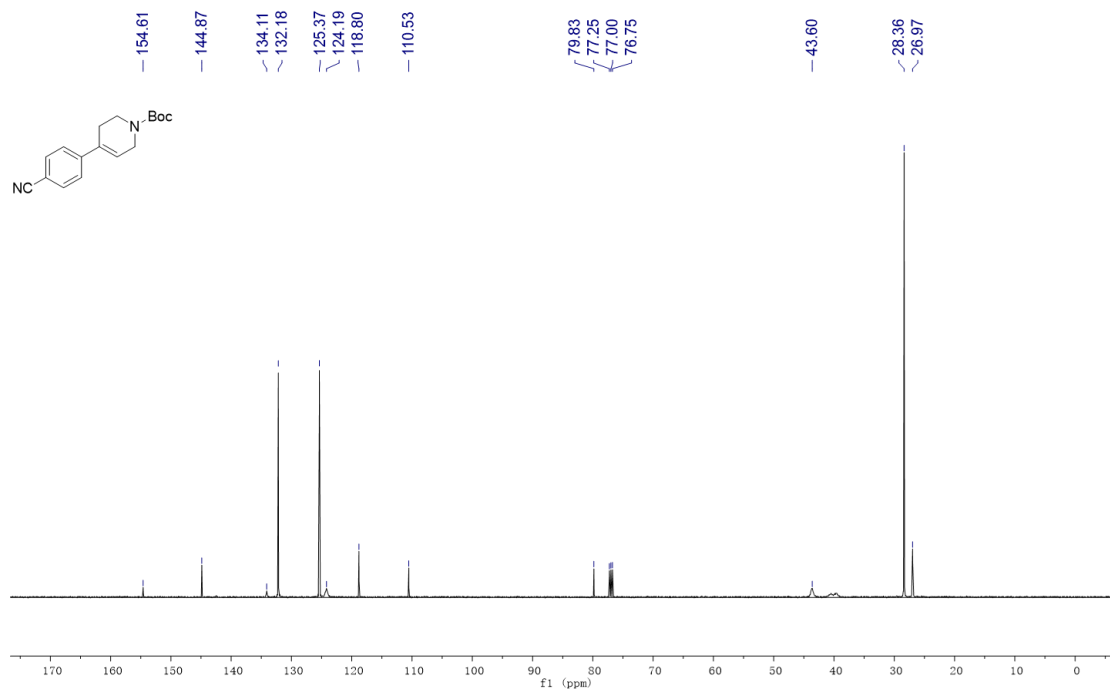
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1242 ¹H and ¹³C-NMR spectra of product 5i.

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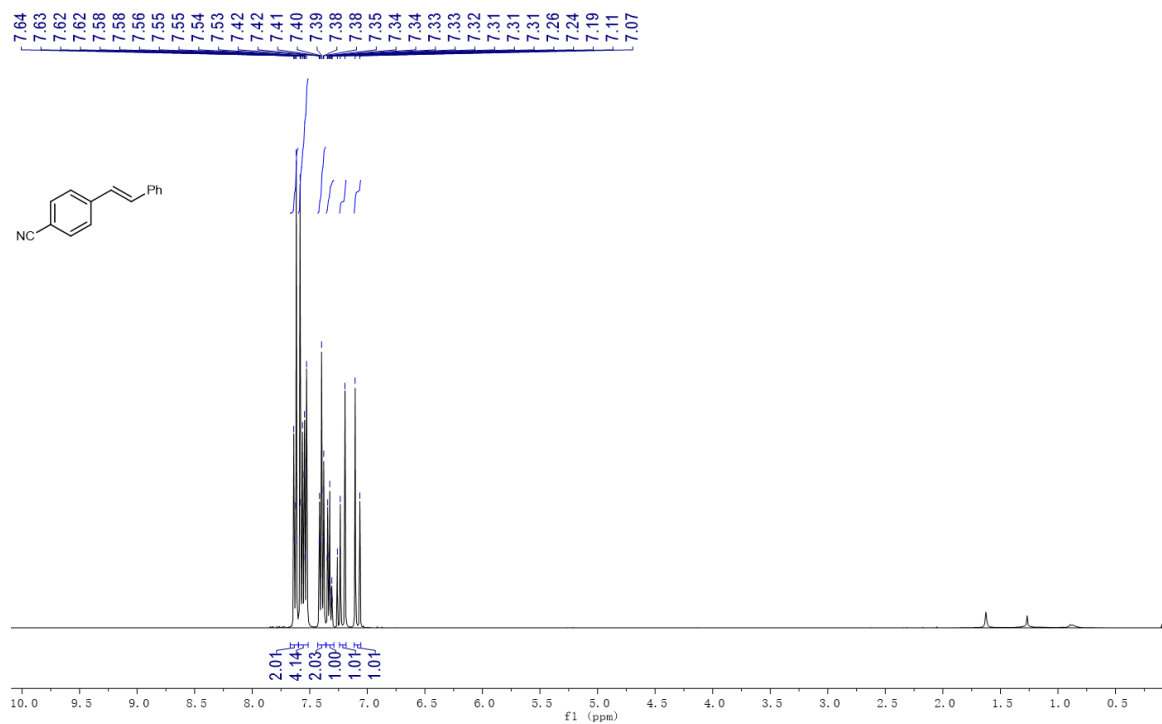


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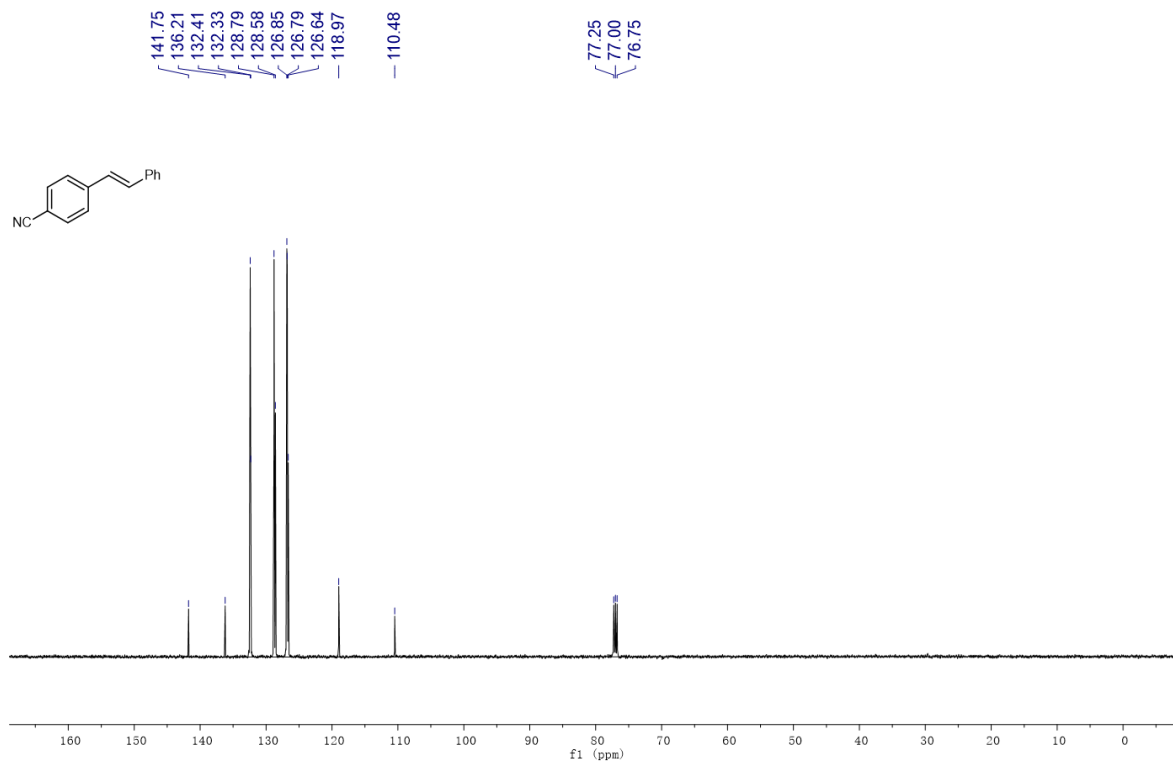
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1247 ¹H and ¹³C-NMR spectra of product 5j.

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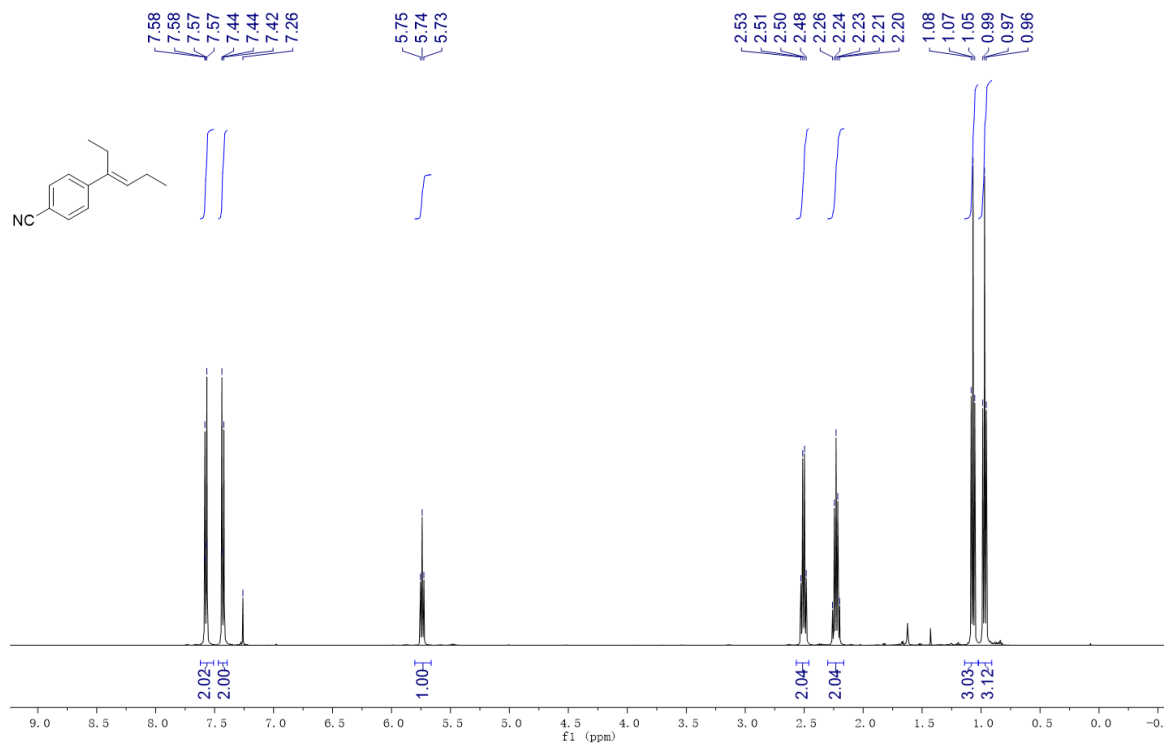


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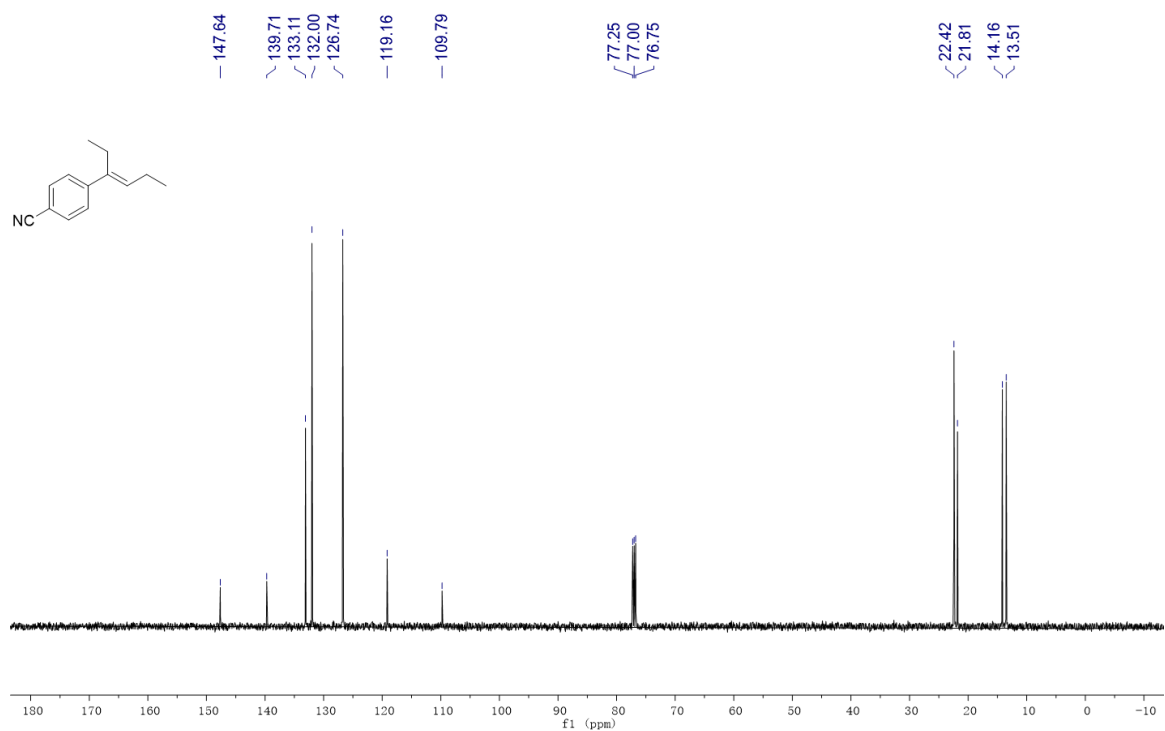
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1252 ¹H and ¹³C-NMR spectra of product 5k.

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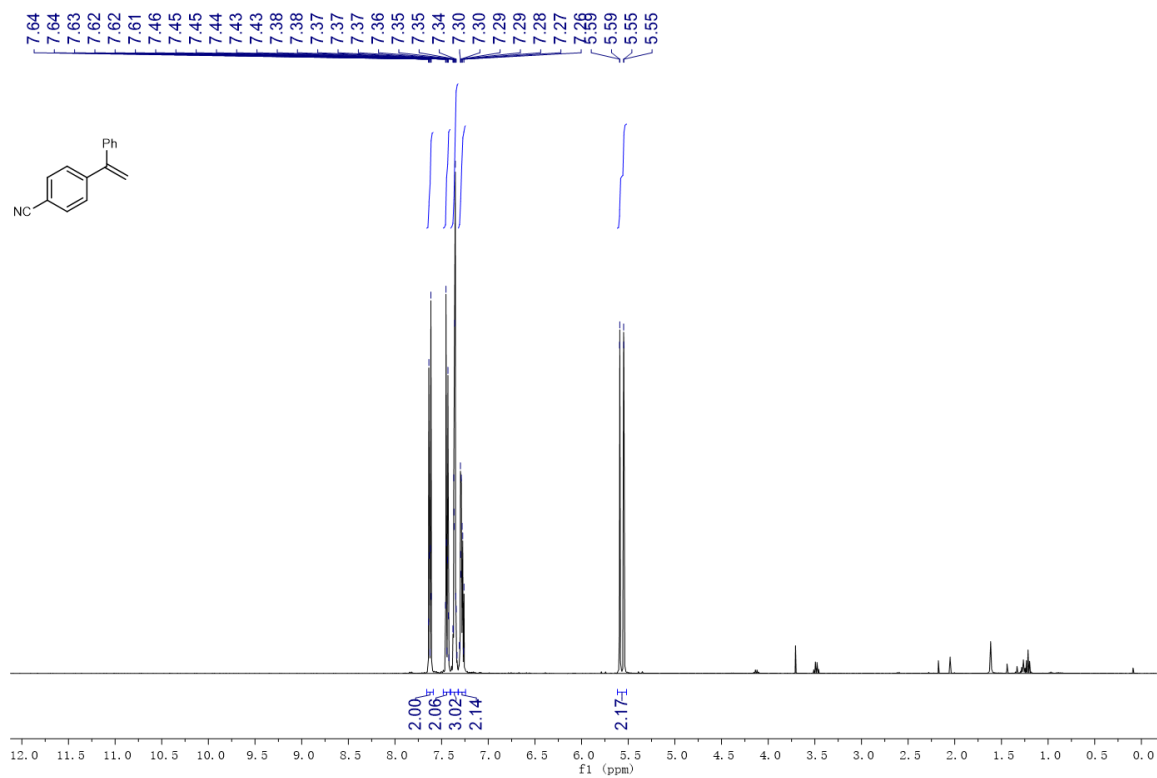


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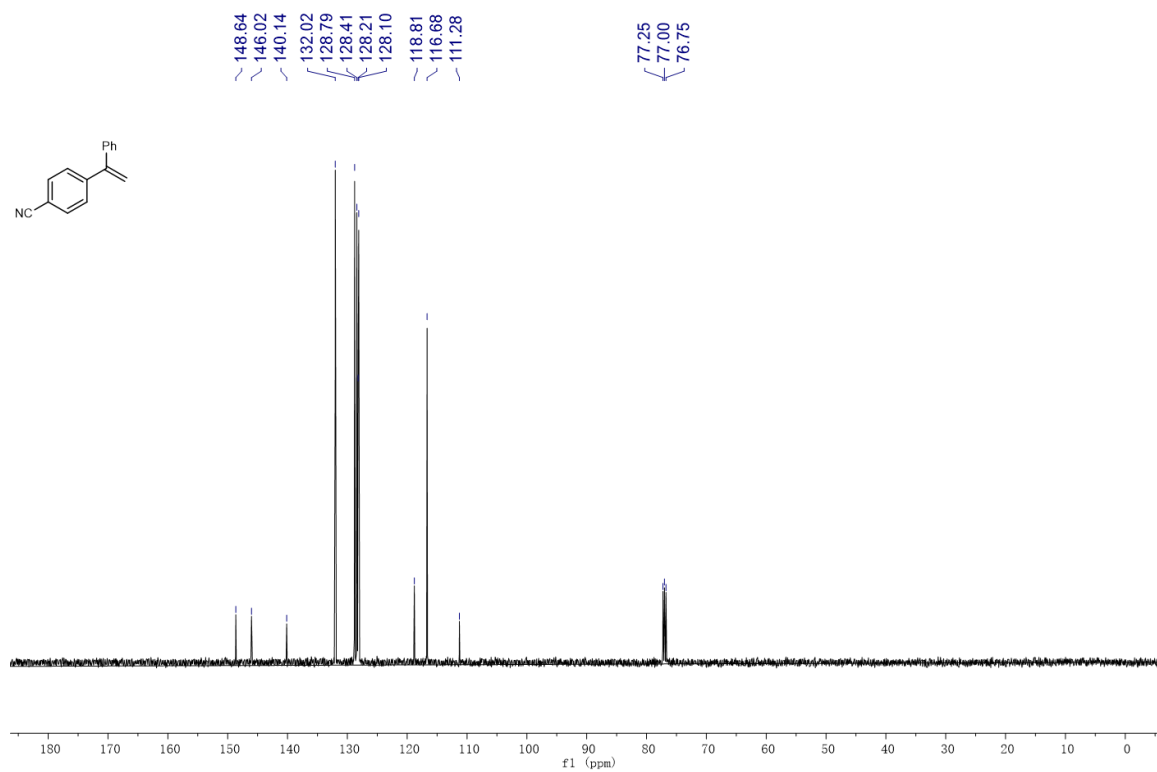
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1257 ¹H and ¹³C-NMR spectra of product 5l.

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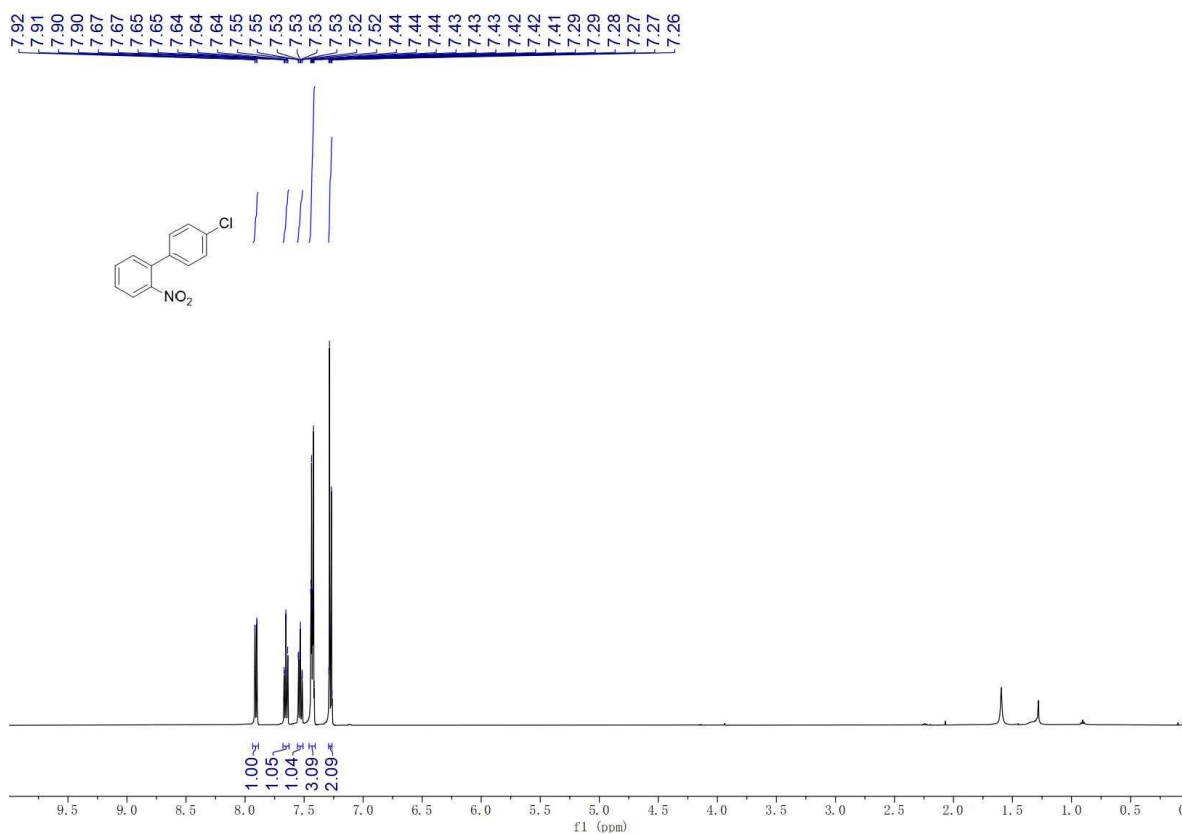


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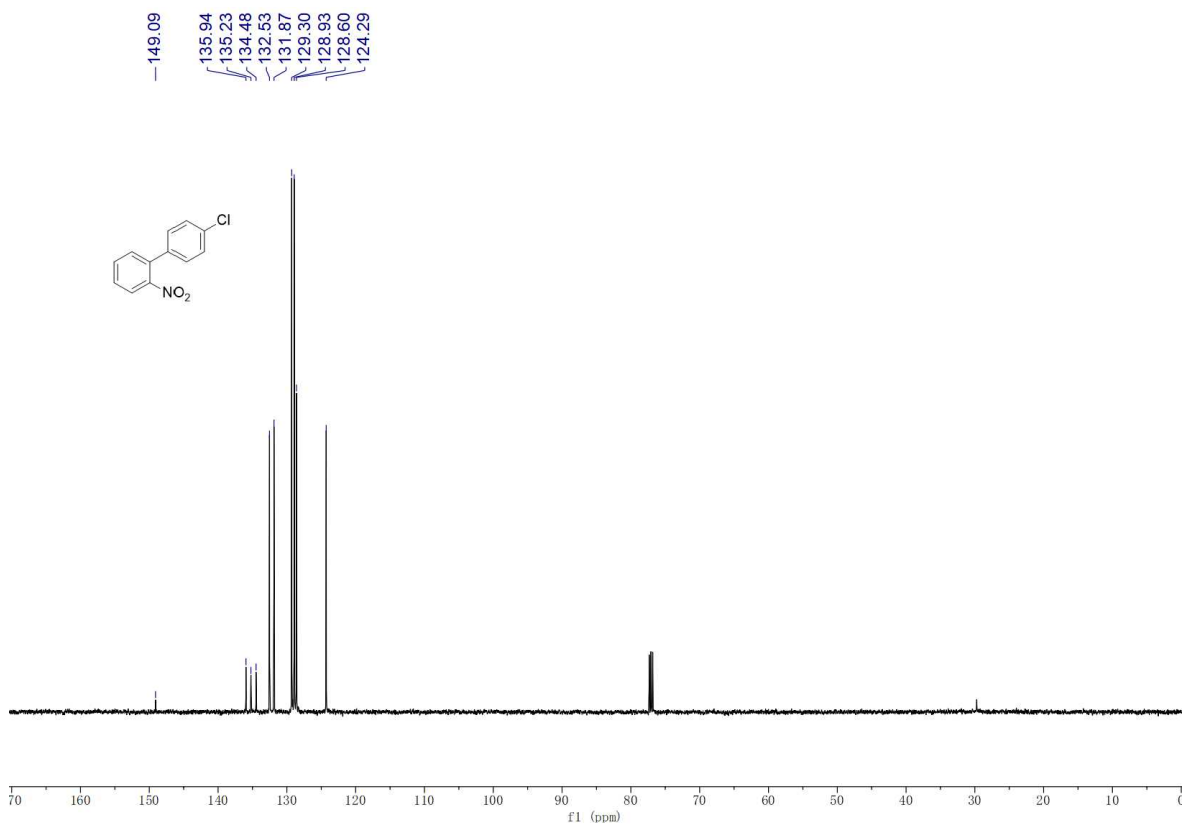
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1262 ¹H, ¹³C-NMR spectra of product 4ae.

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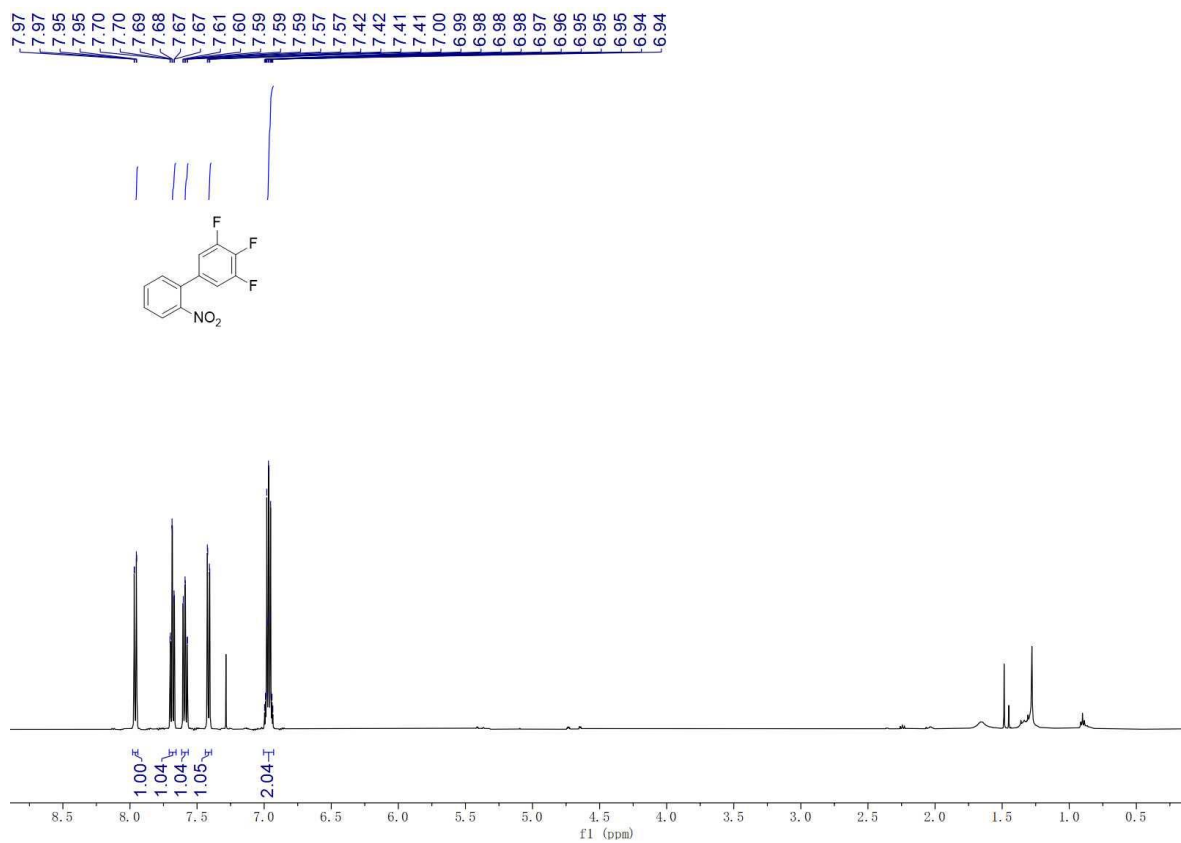
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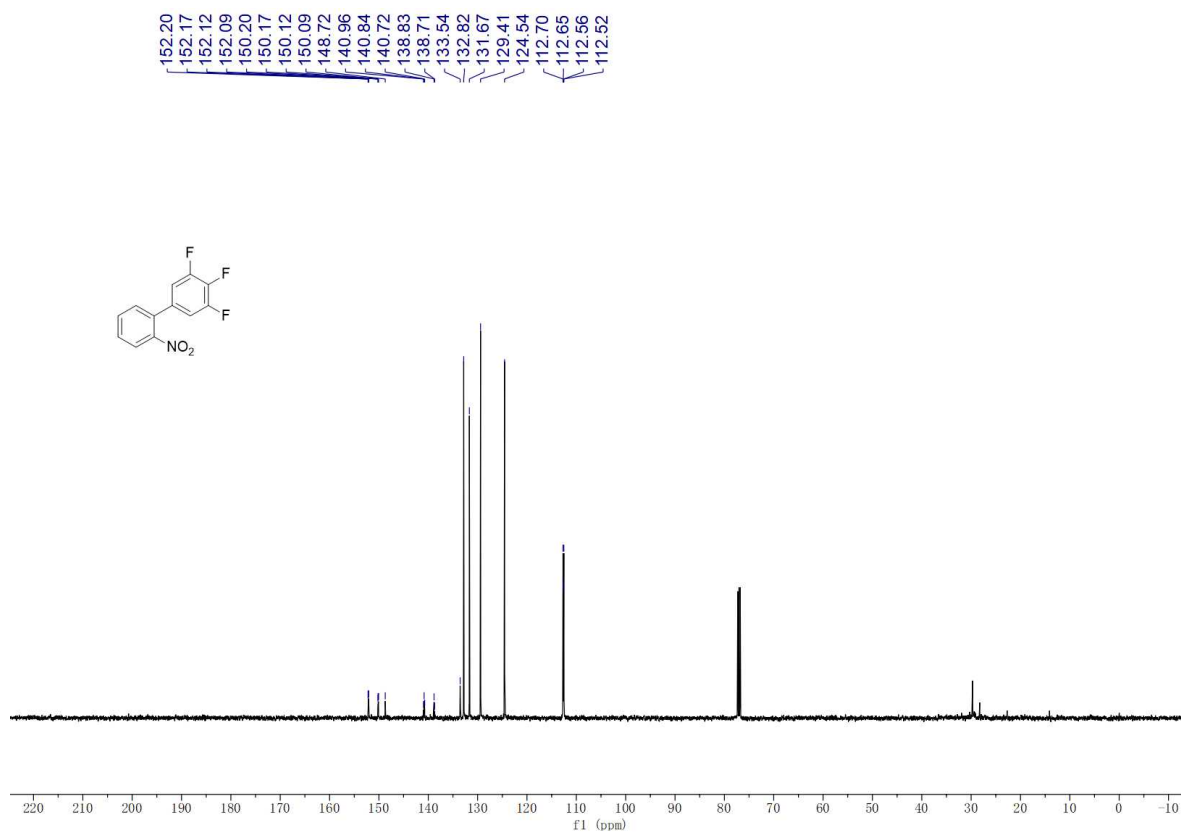
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1266 ¹H, ¹³C and ¹⁹F NMR of product 4af.

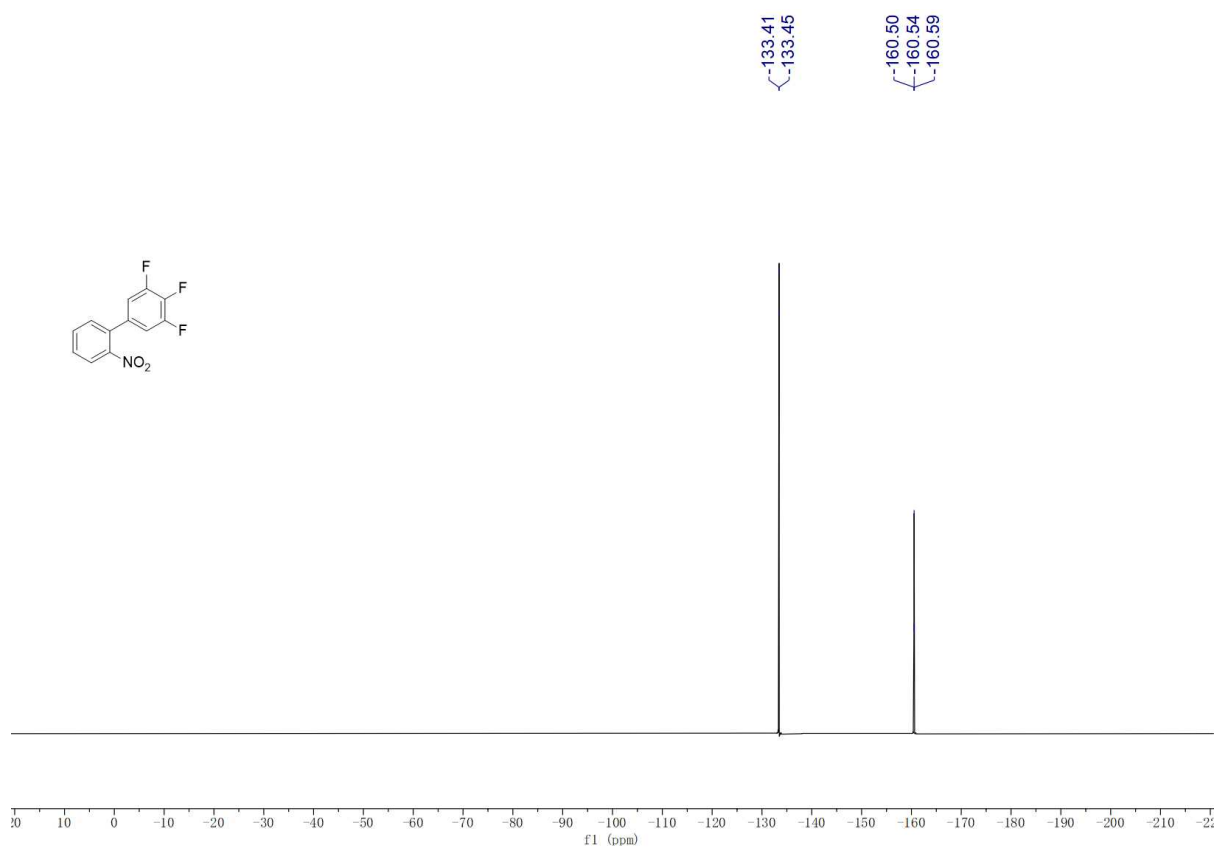
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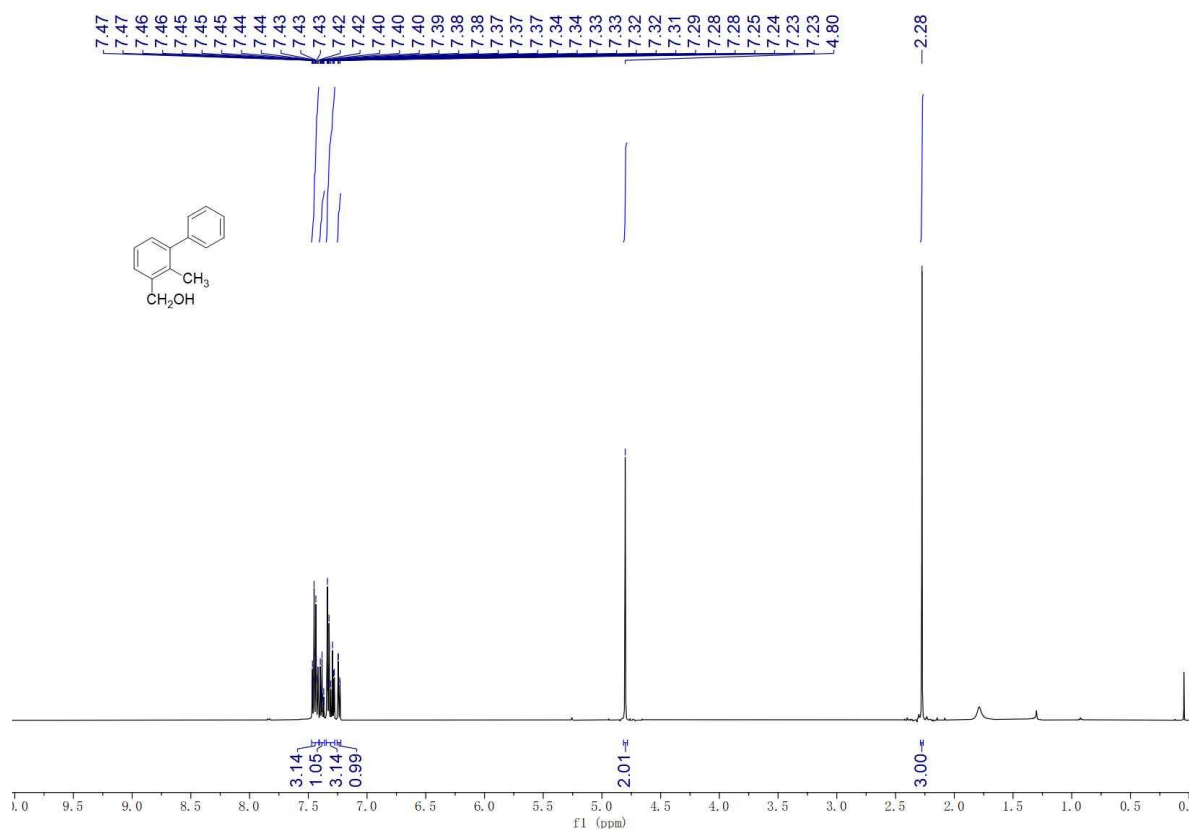


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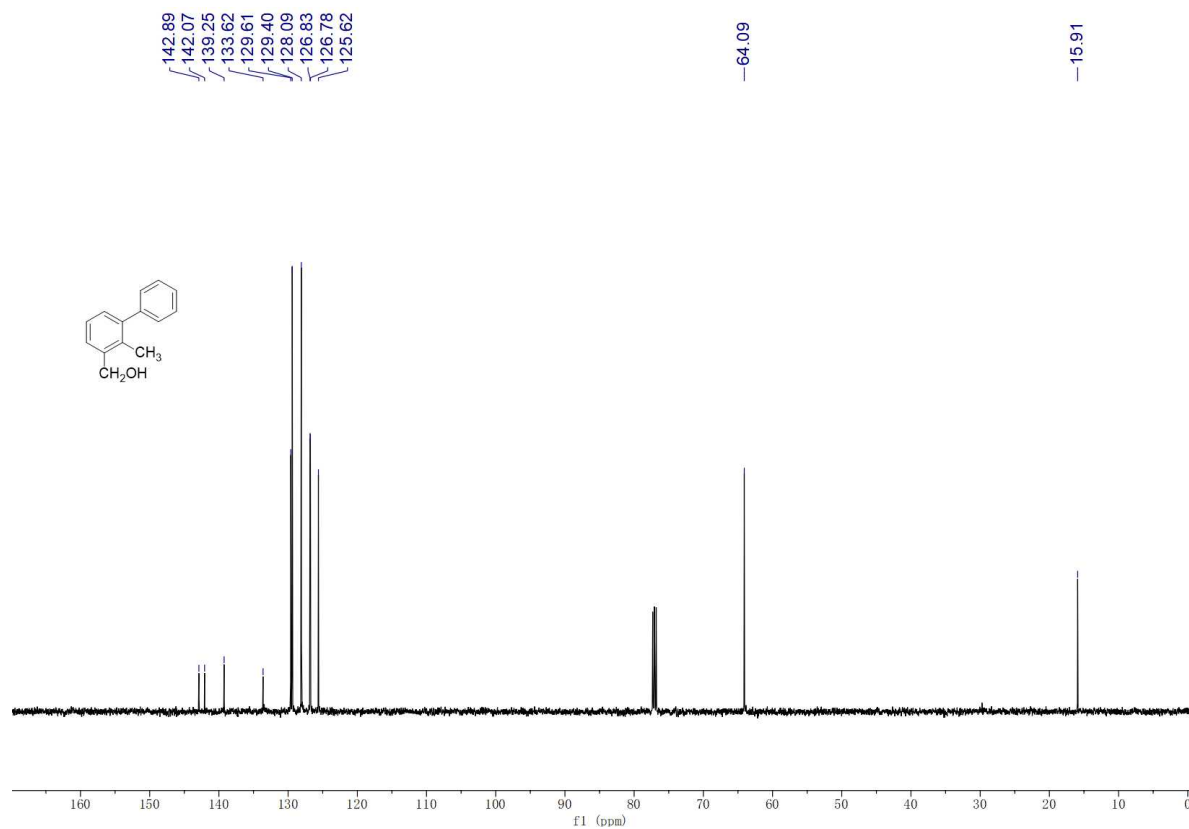
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1272 ¹H and ¹³C spectra of product 4ag.

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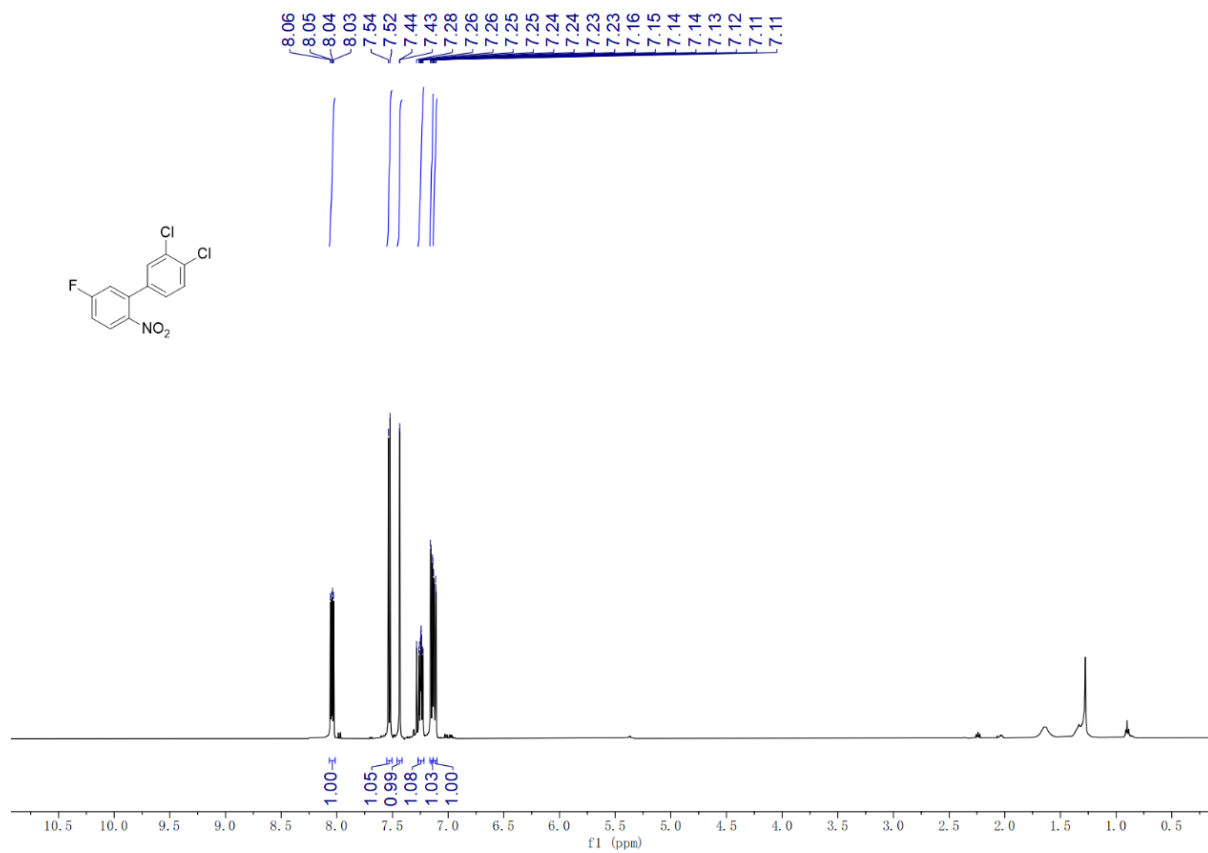
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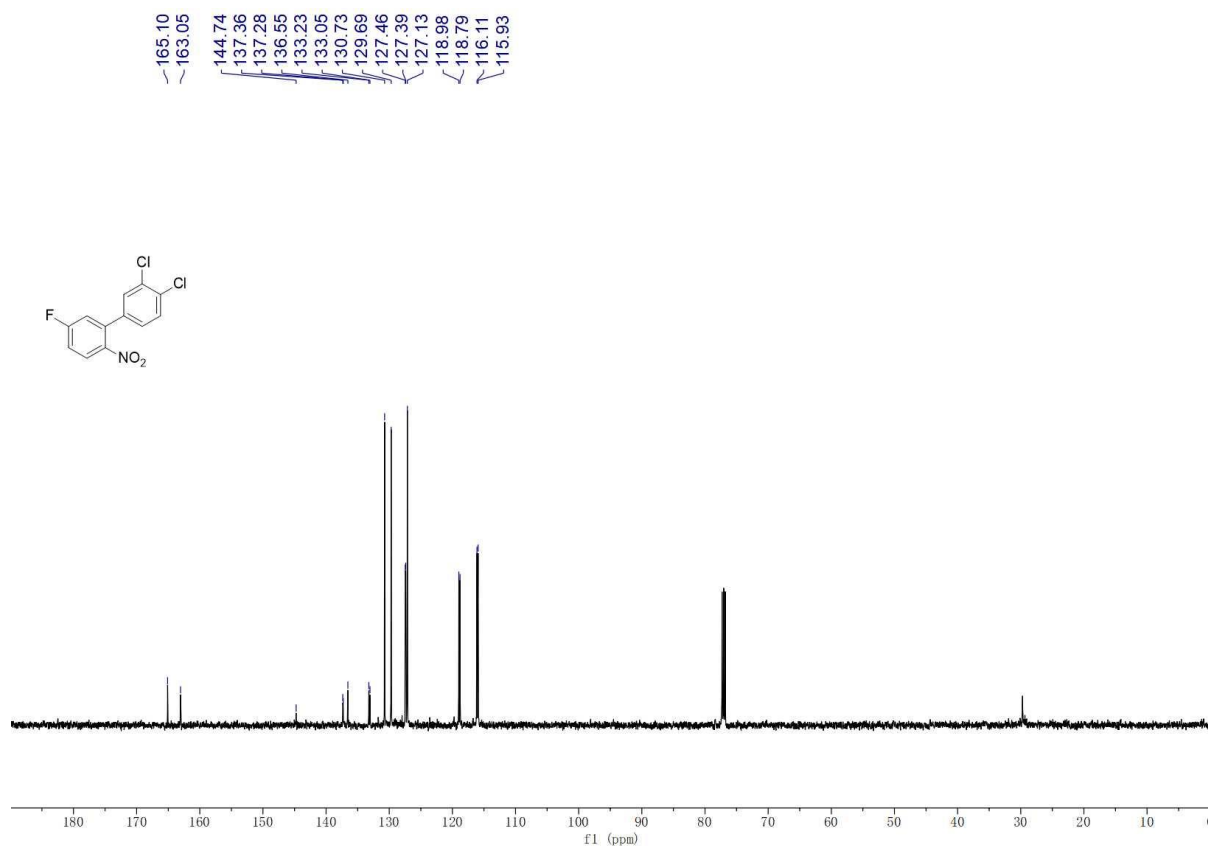
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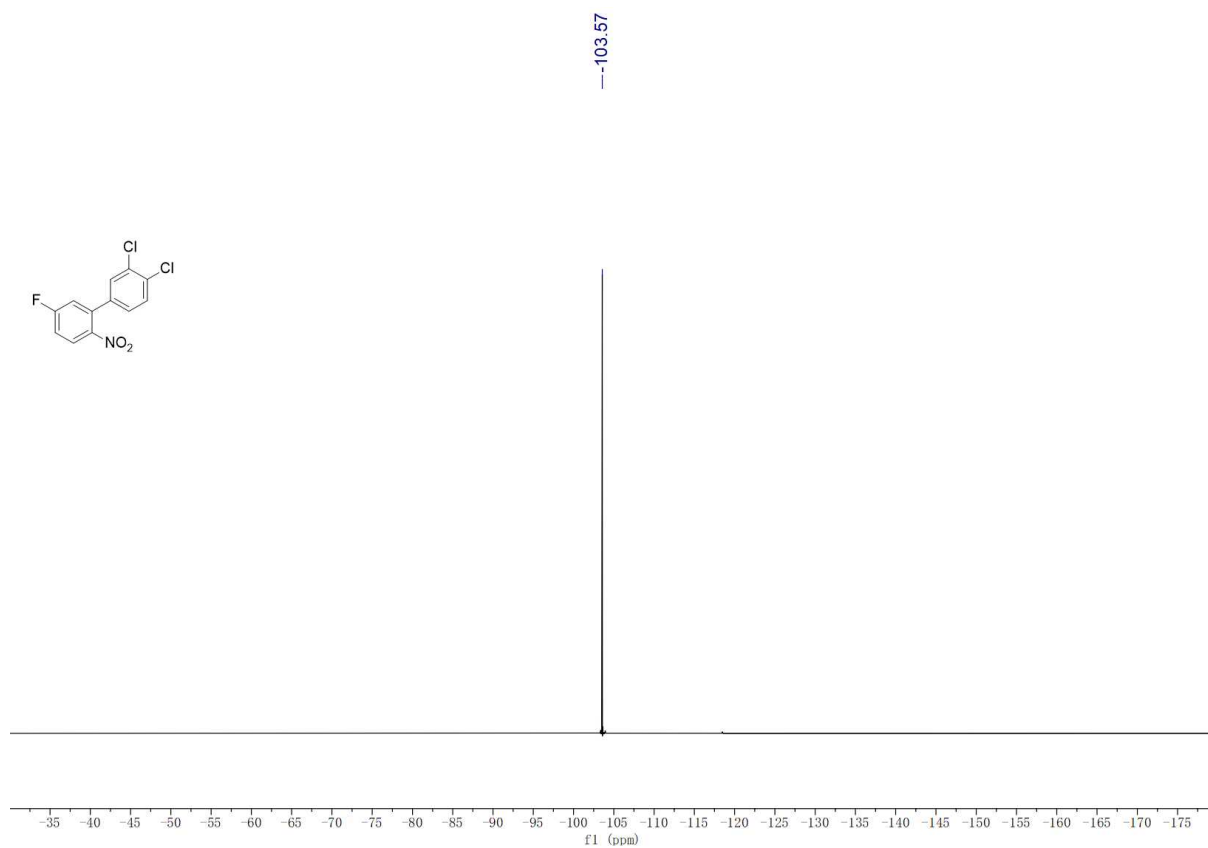
1276 ^1H , ^{13}C and ^{19}F NMR spectra of product 4ah.

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1278



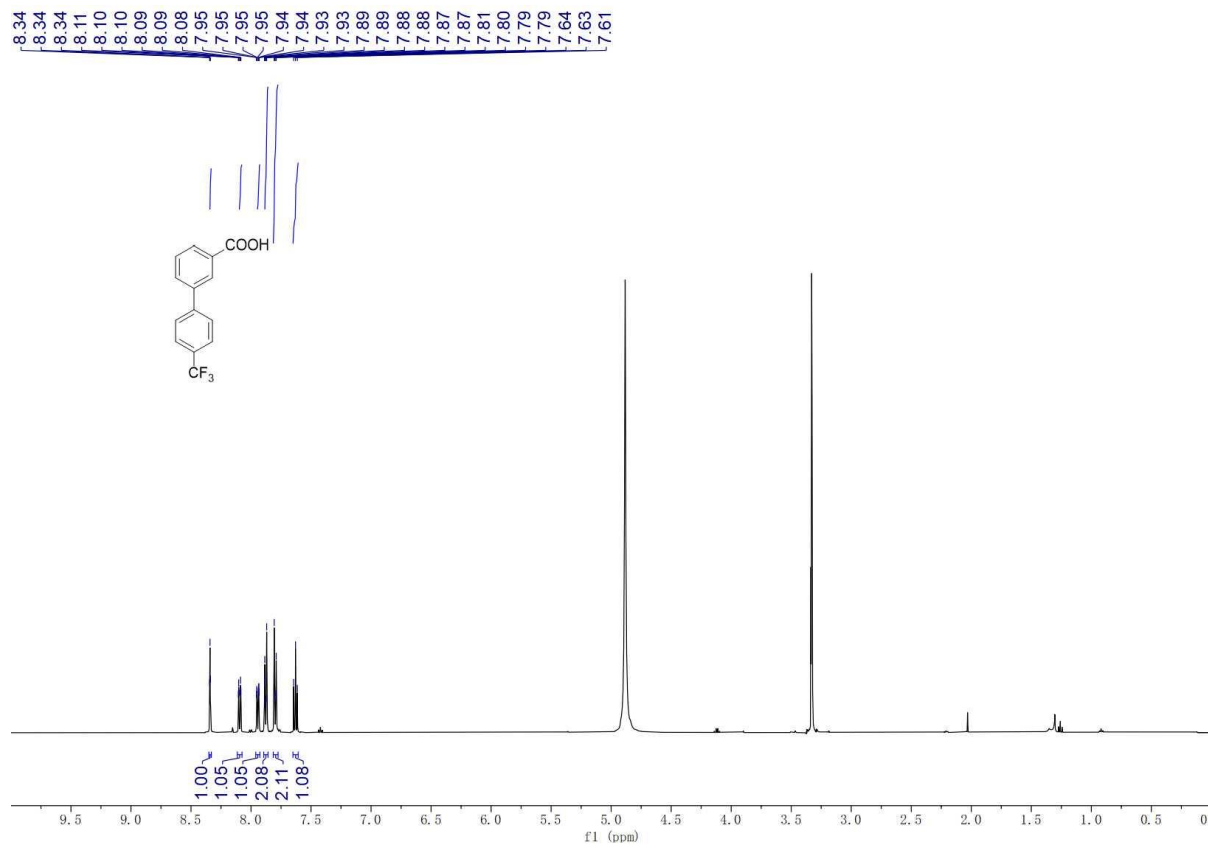


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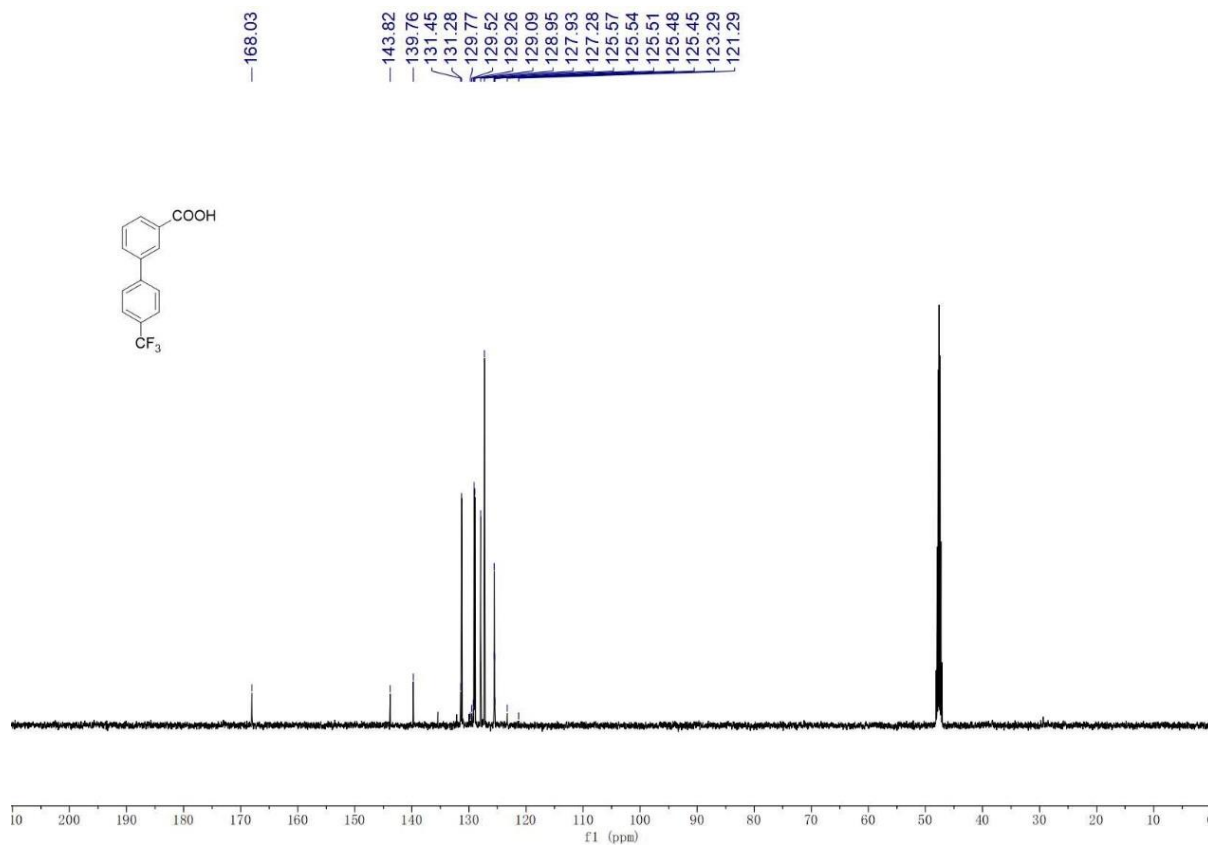
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1281 ^1H , ^{13}C and ^{19}F NMR spectra of product 4ai.

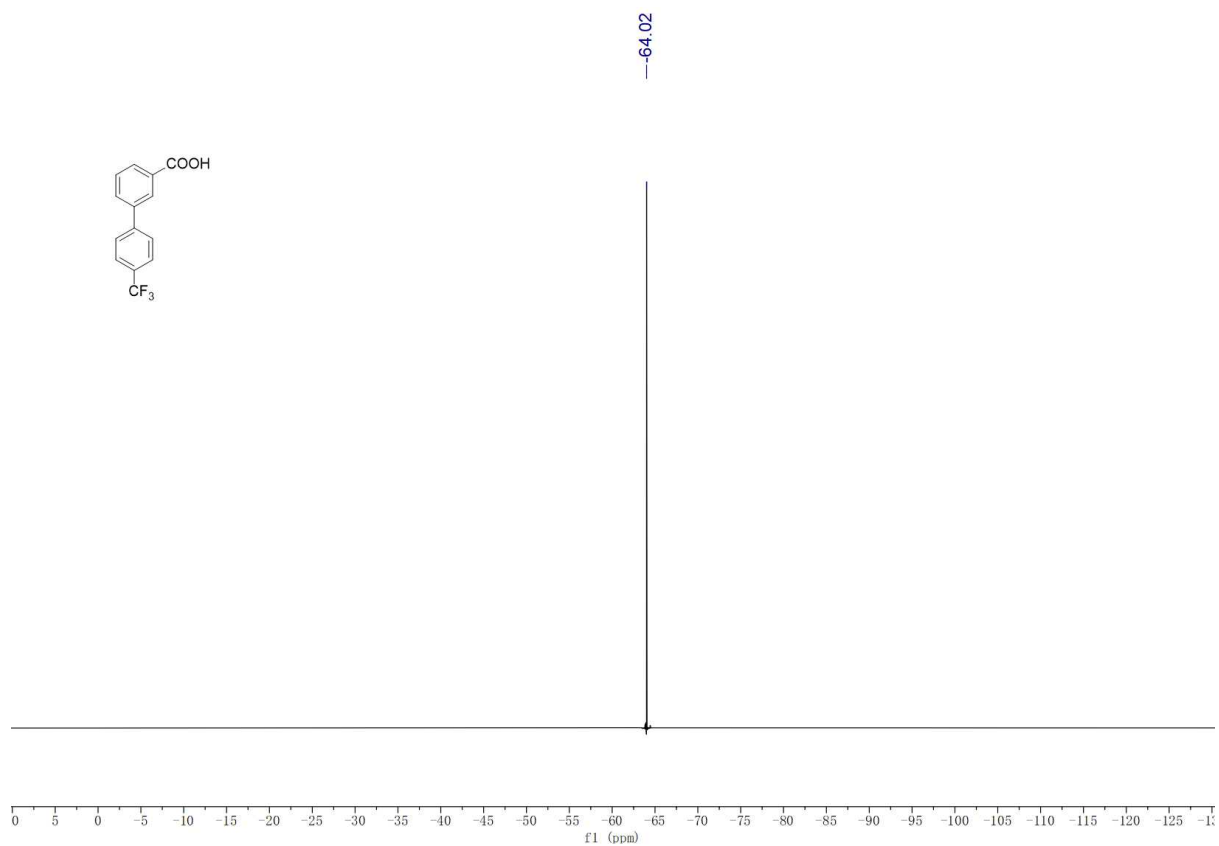
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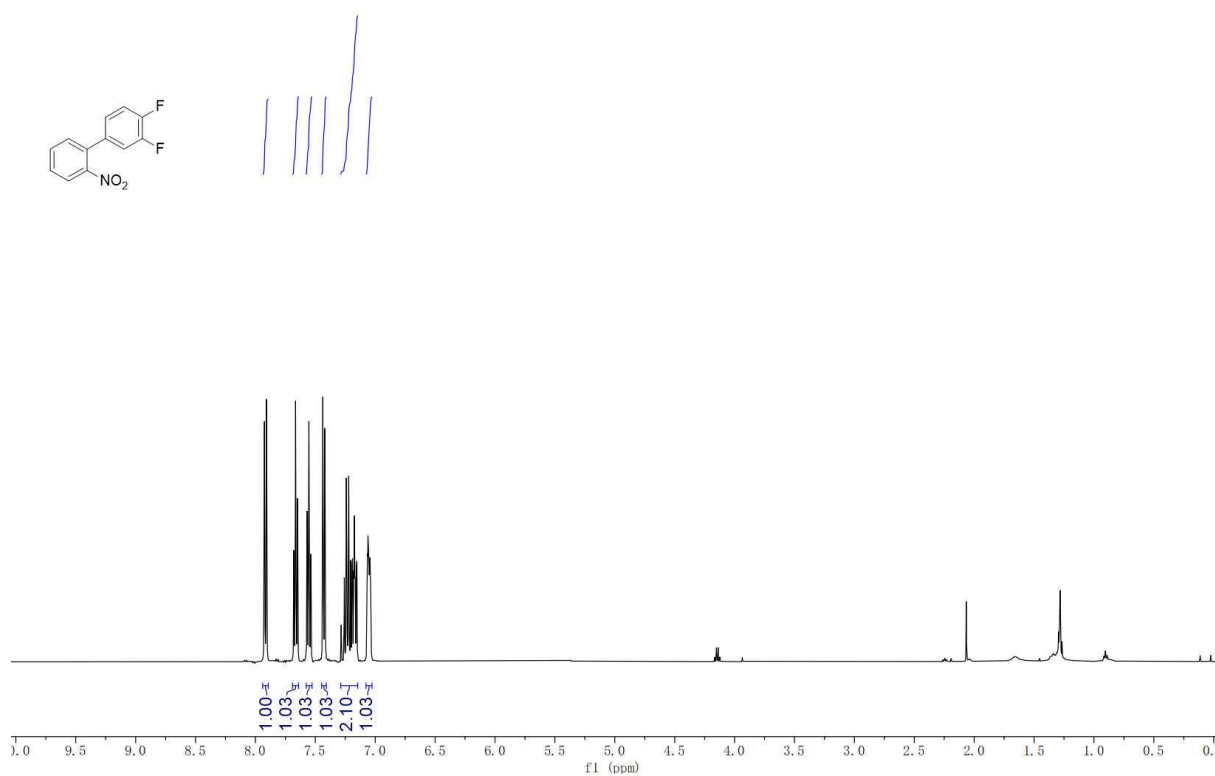


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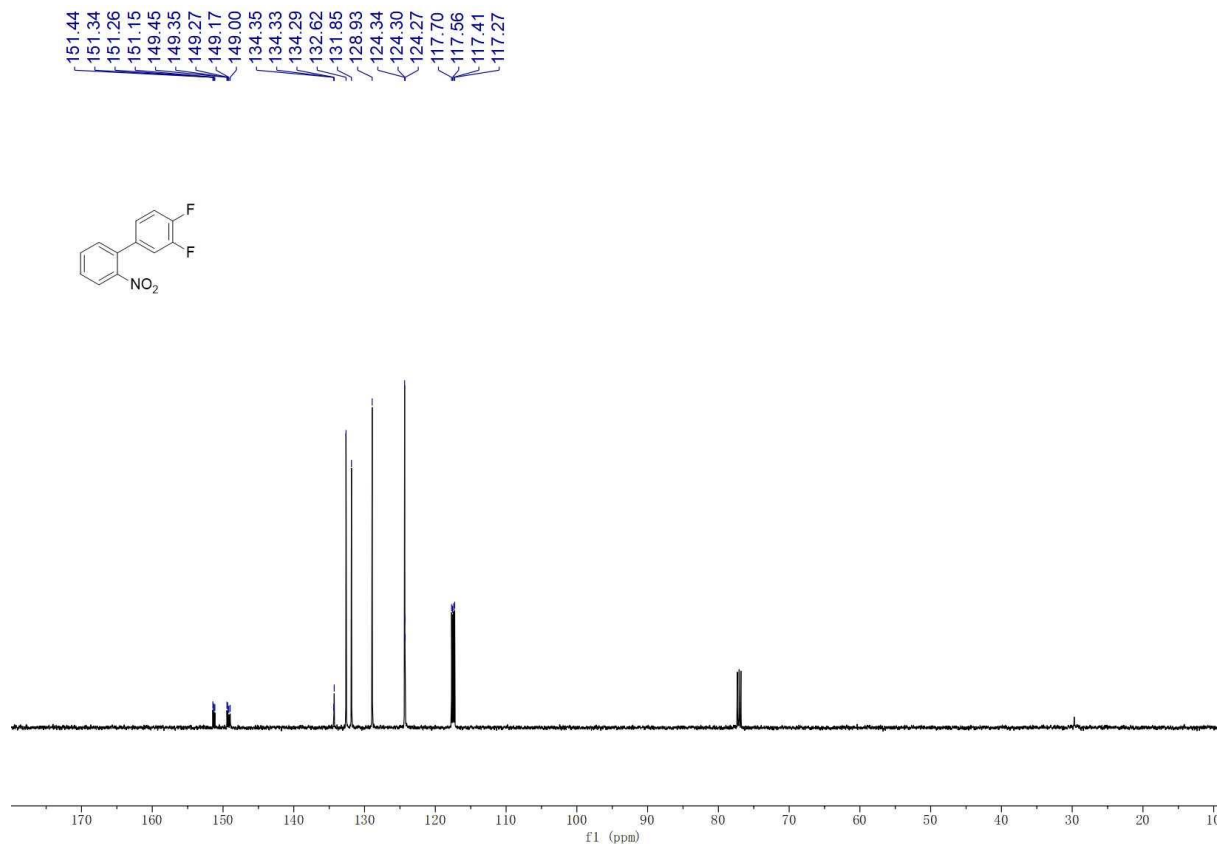
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1287 ^1H , ^{13}C and ^{19}F NMR spectra of product 4aj.

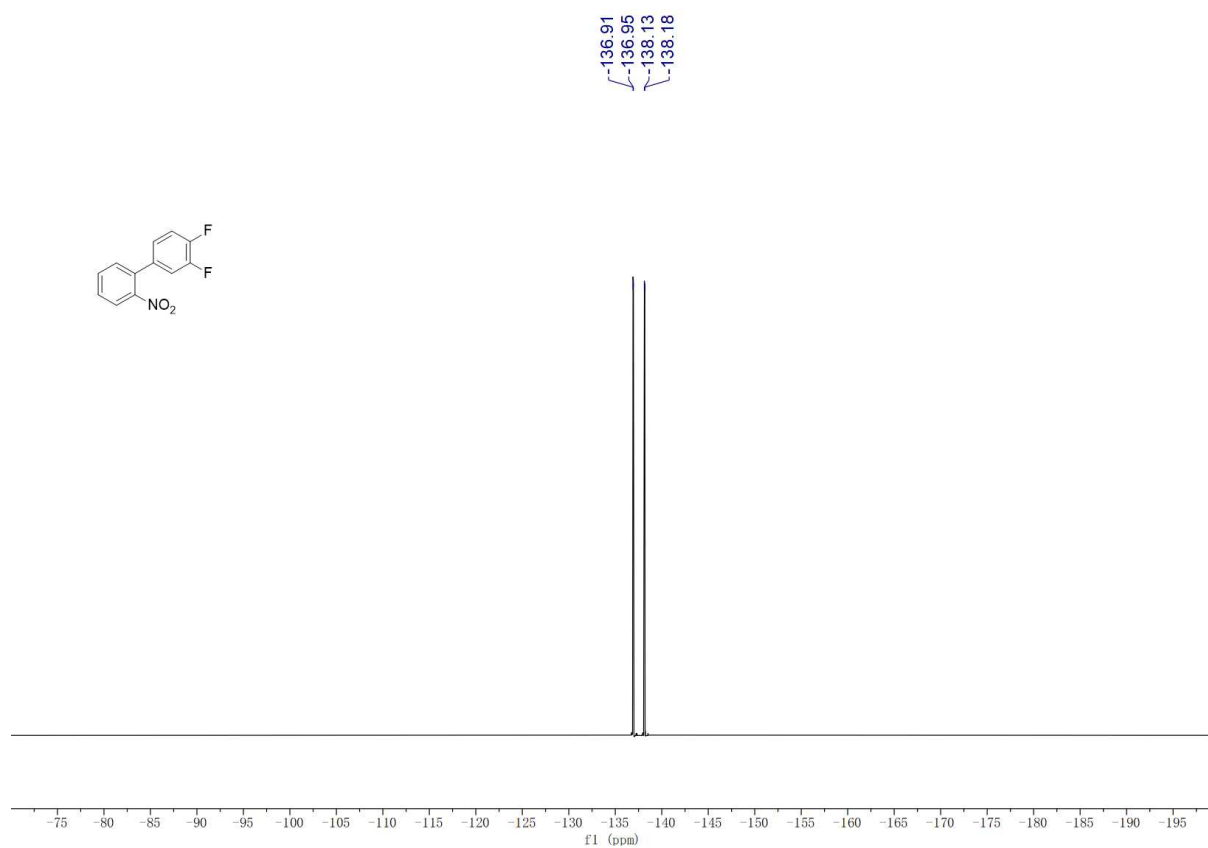
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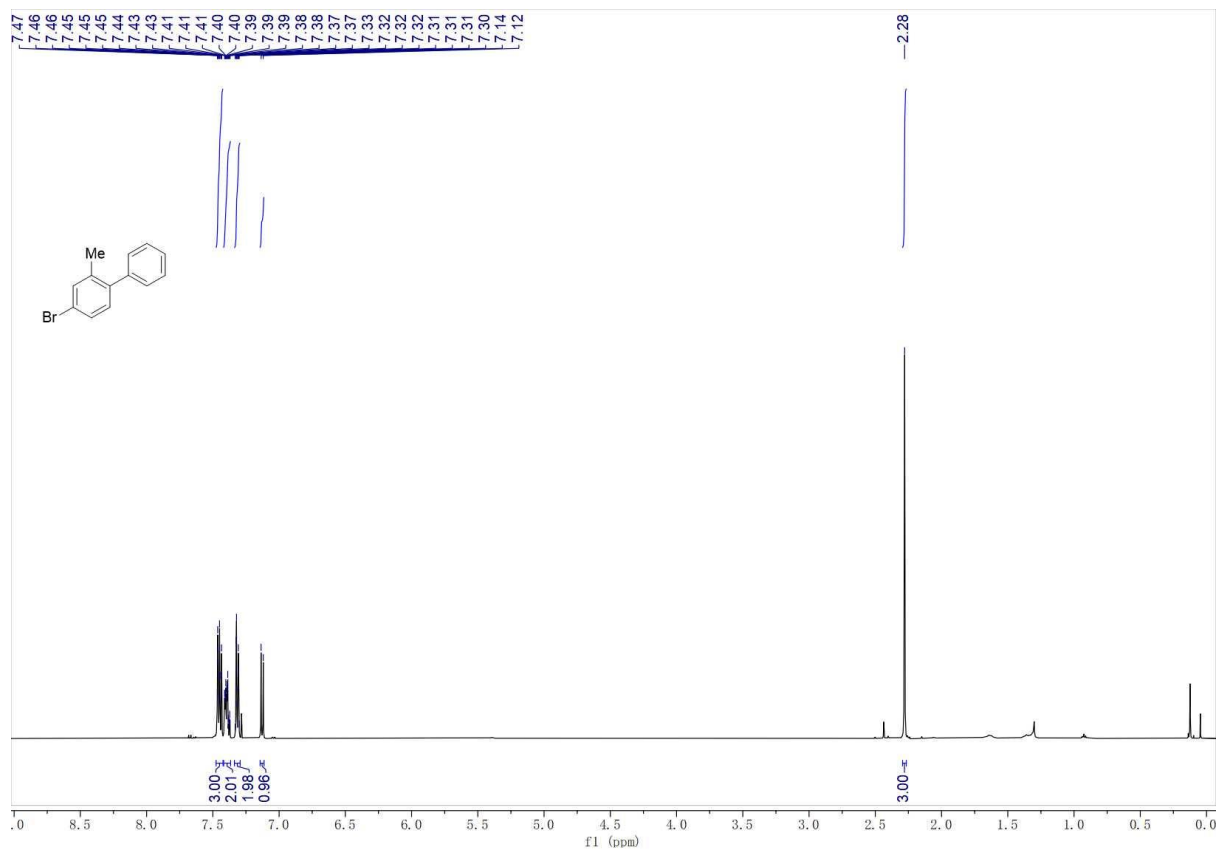


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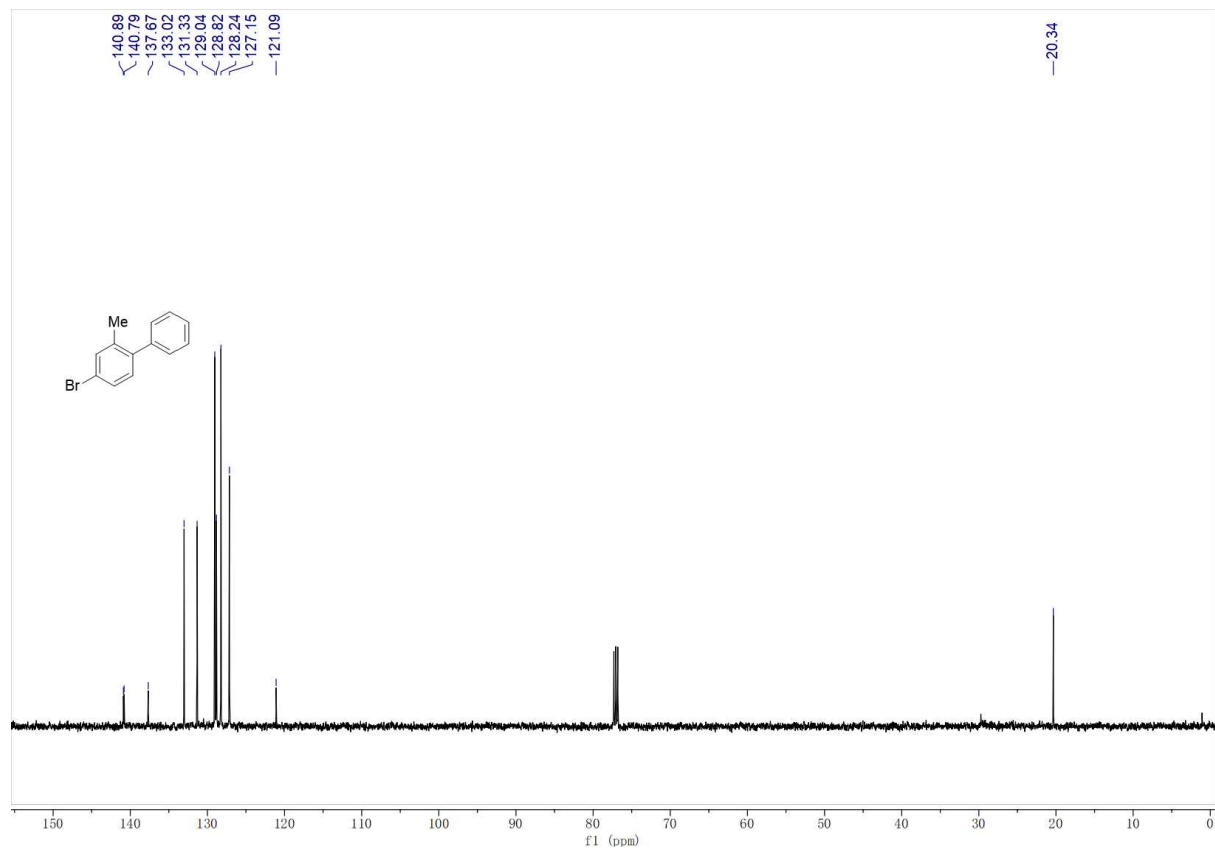
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1293 ¹H and ¹³C NMR spectra of product 4ak.

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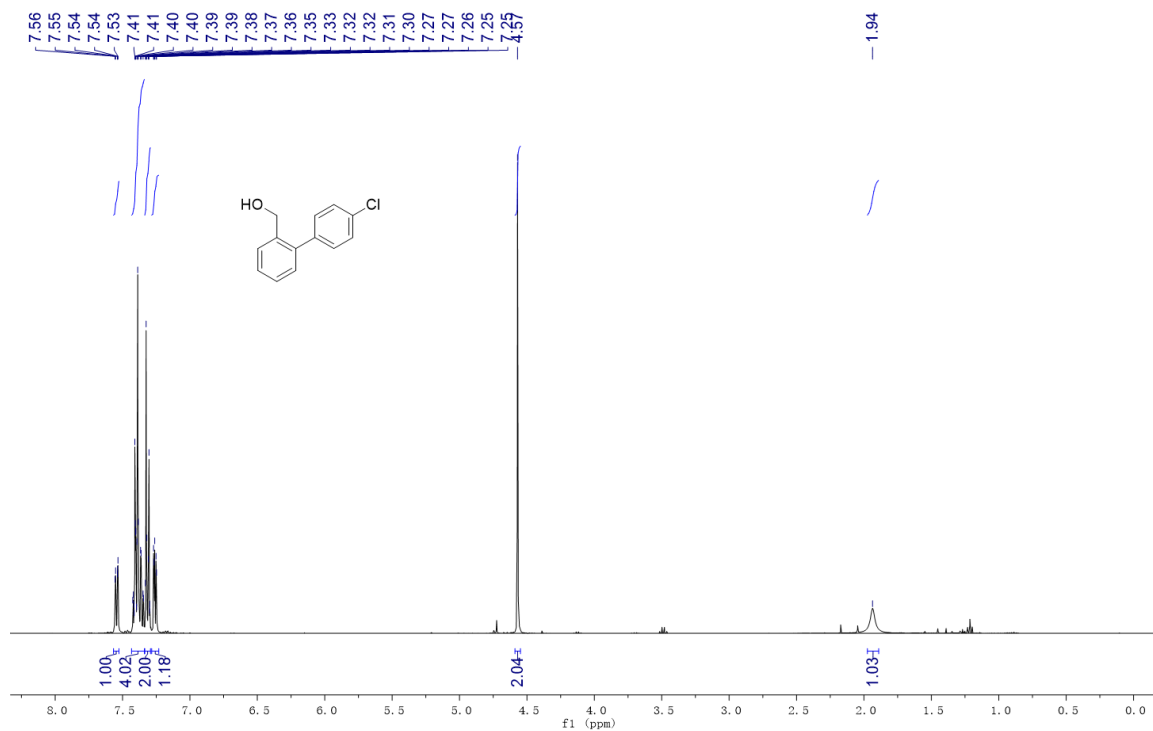
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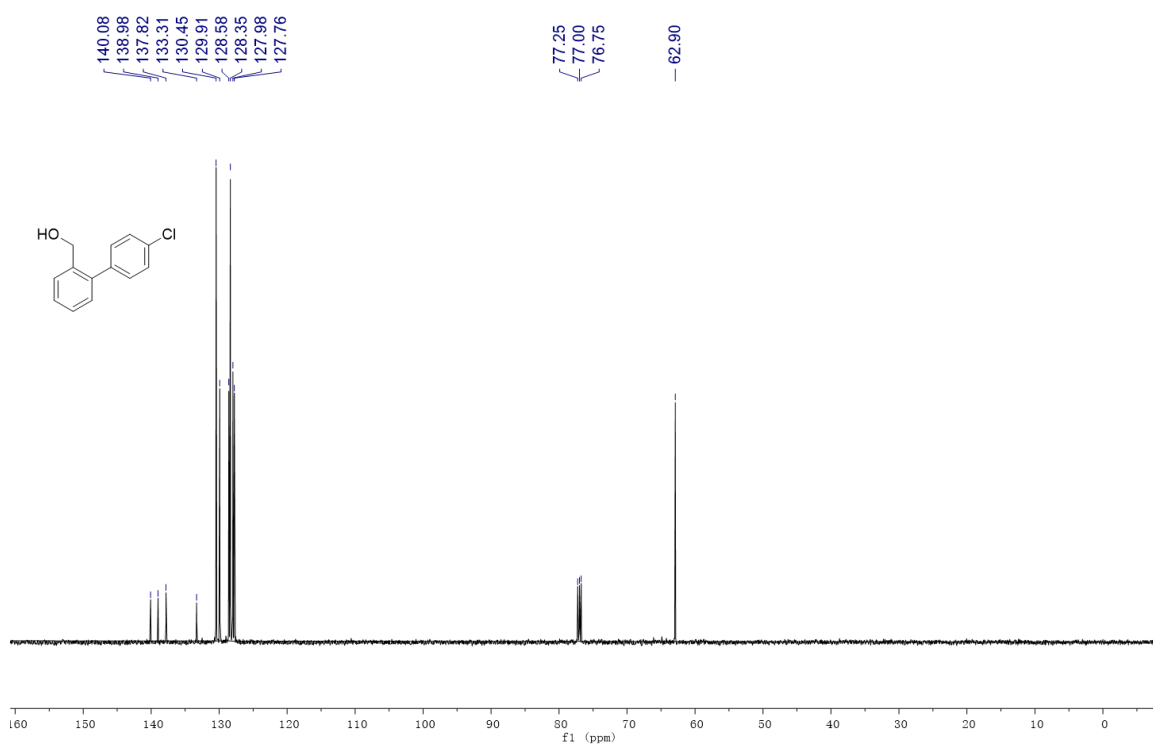
1297 ¹H and ¹³C- NMR spectra of product 4al.

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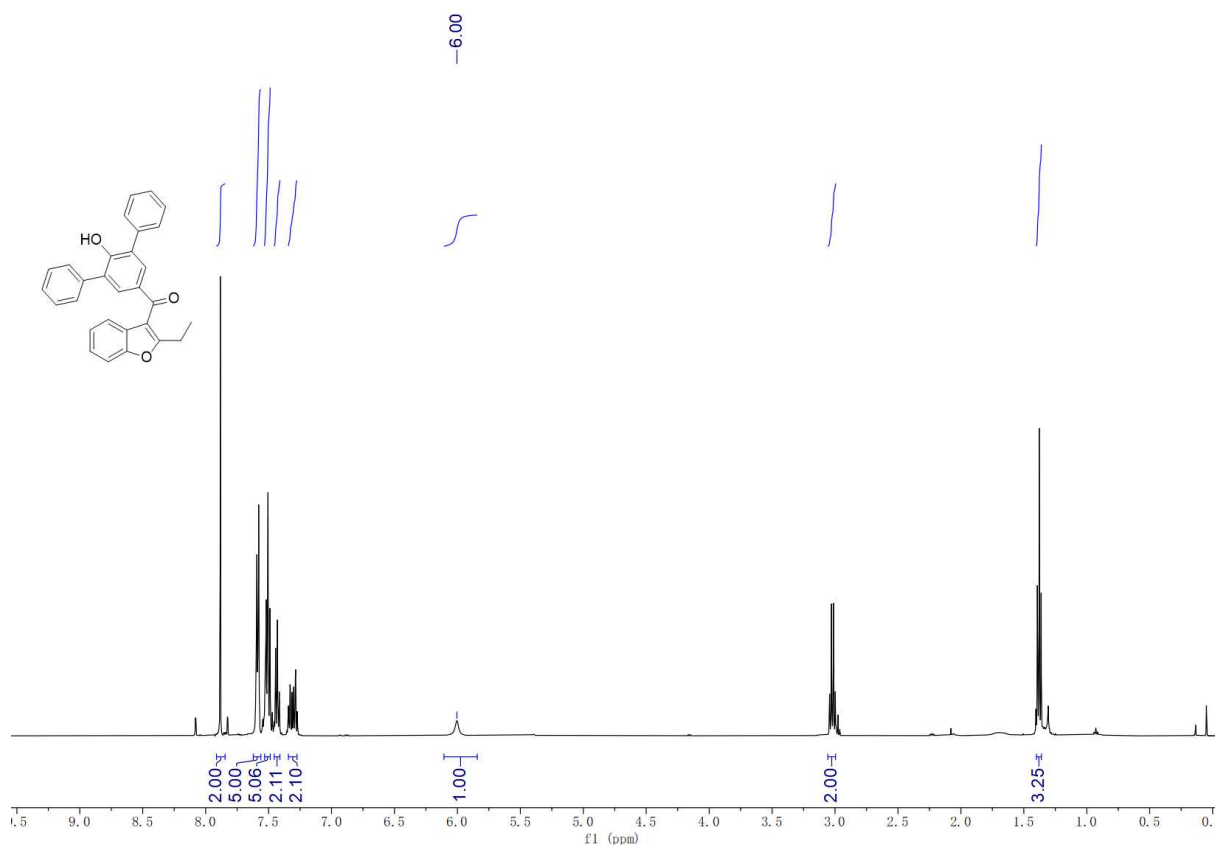
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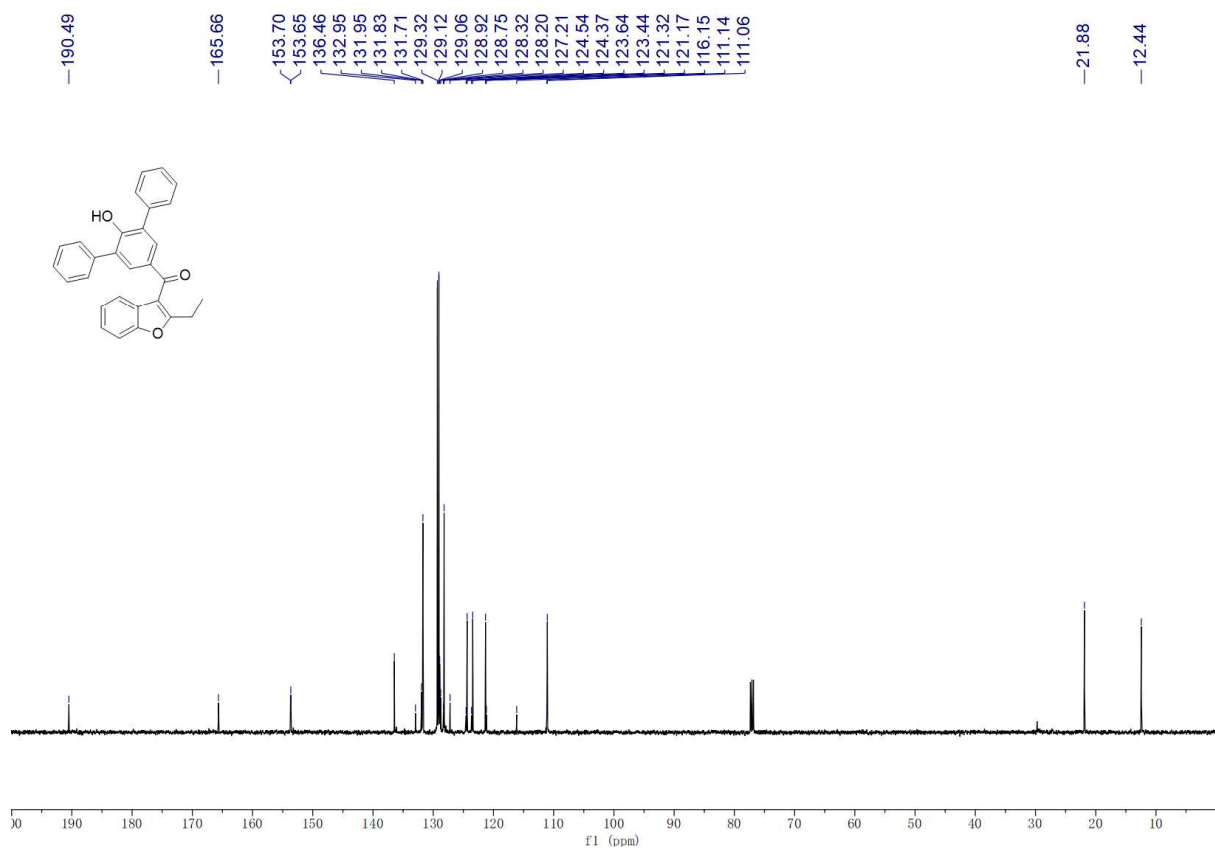
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1302 ¹H and ¹³C spectra of product 4am

1303

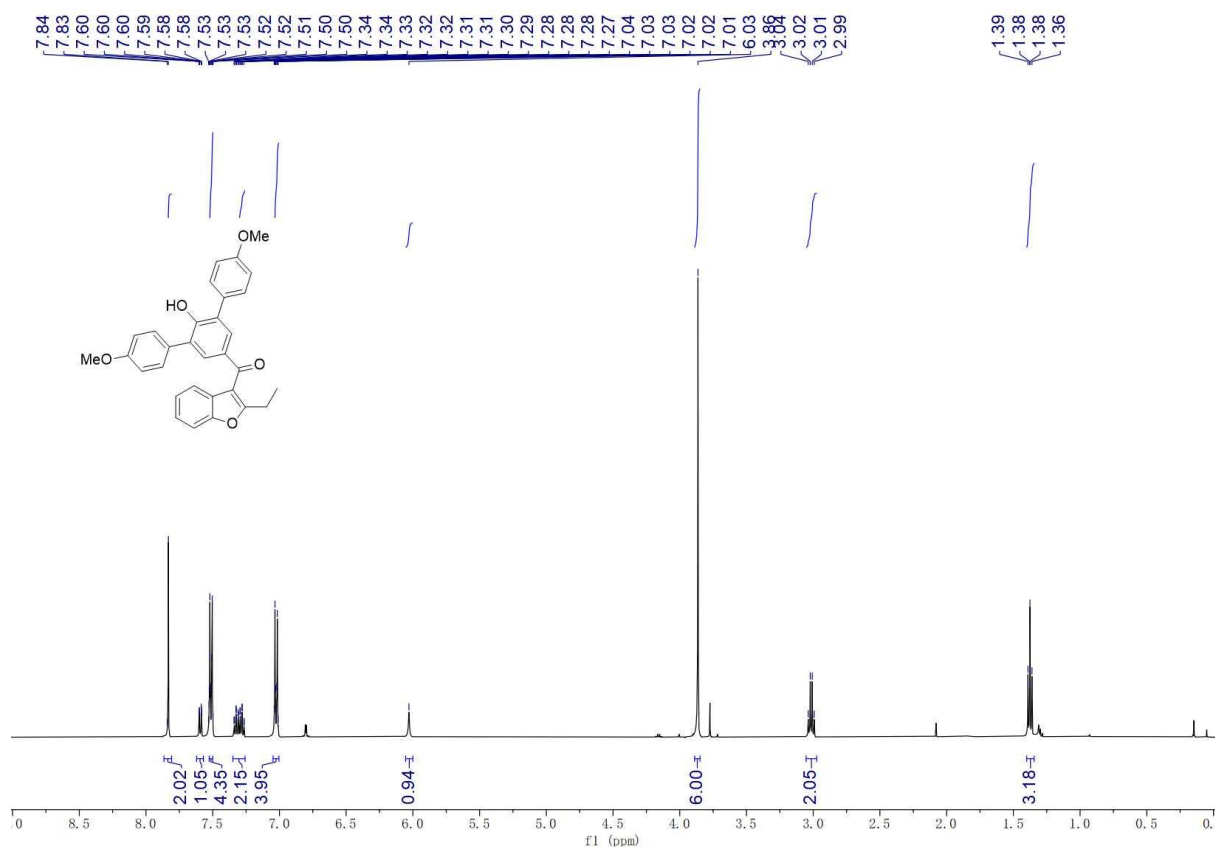


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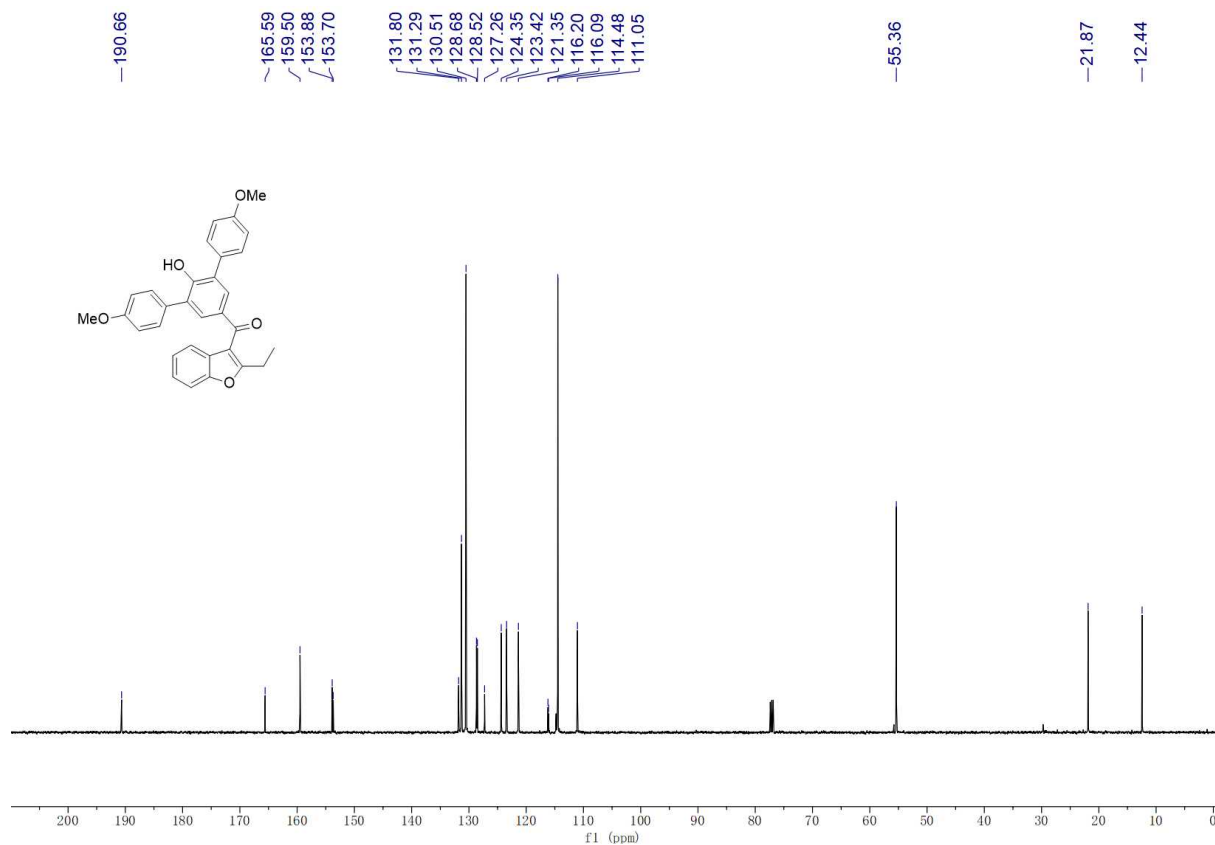


1305 **¹H and ¹³C spectra of product 4an**

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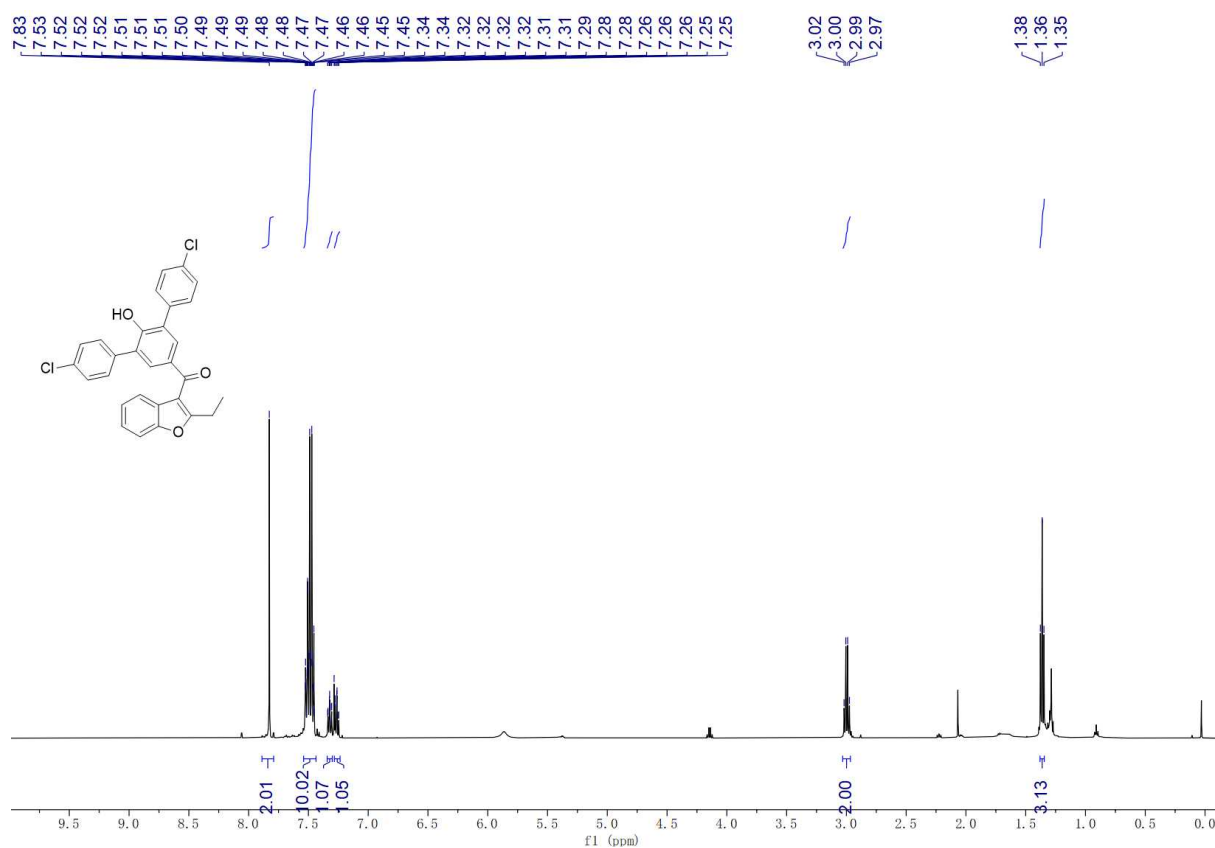
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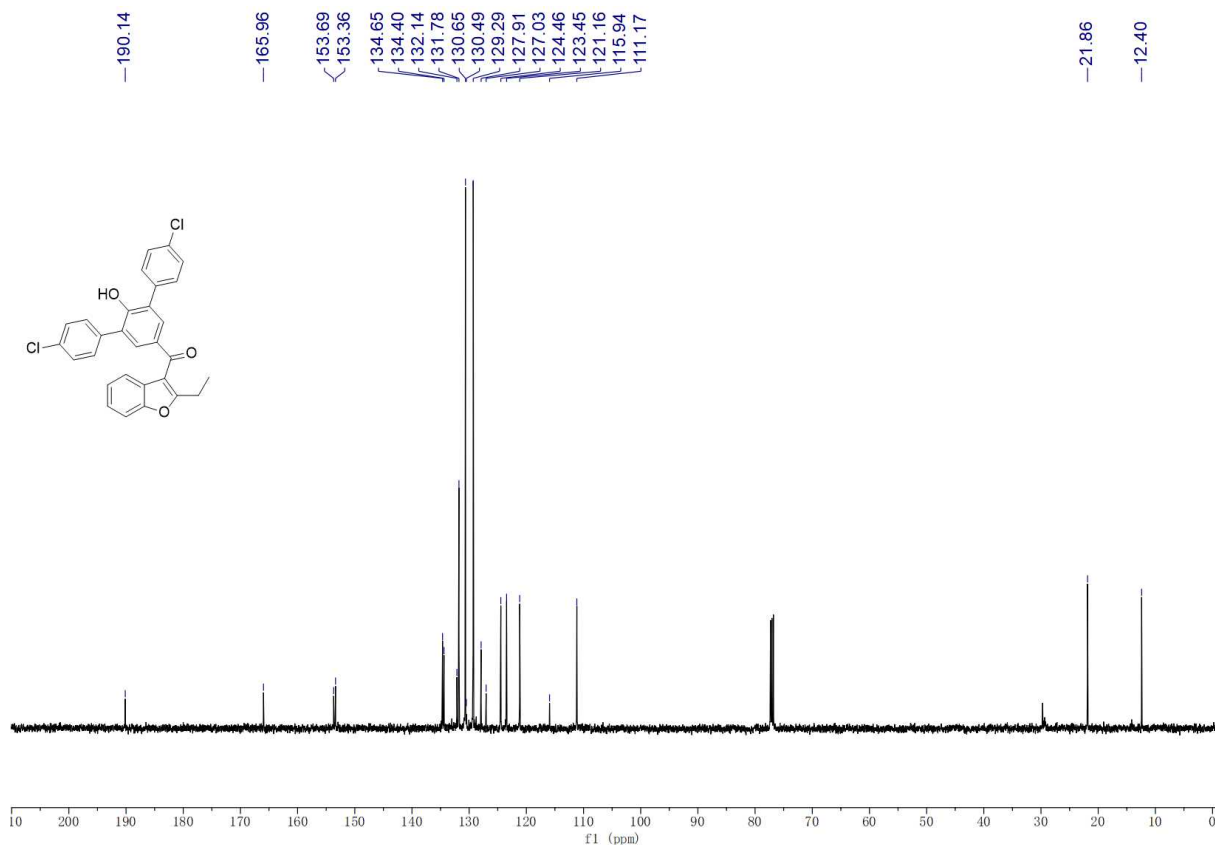
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1309 ¹H and ¹³C spectra of product 4ao

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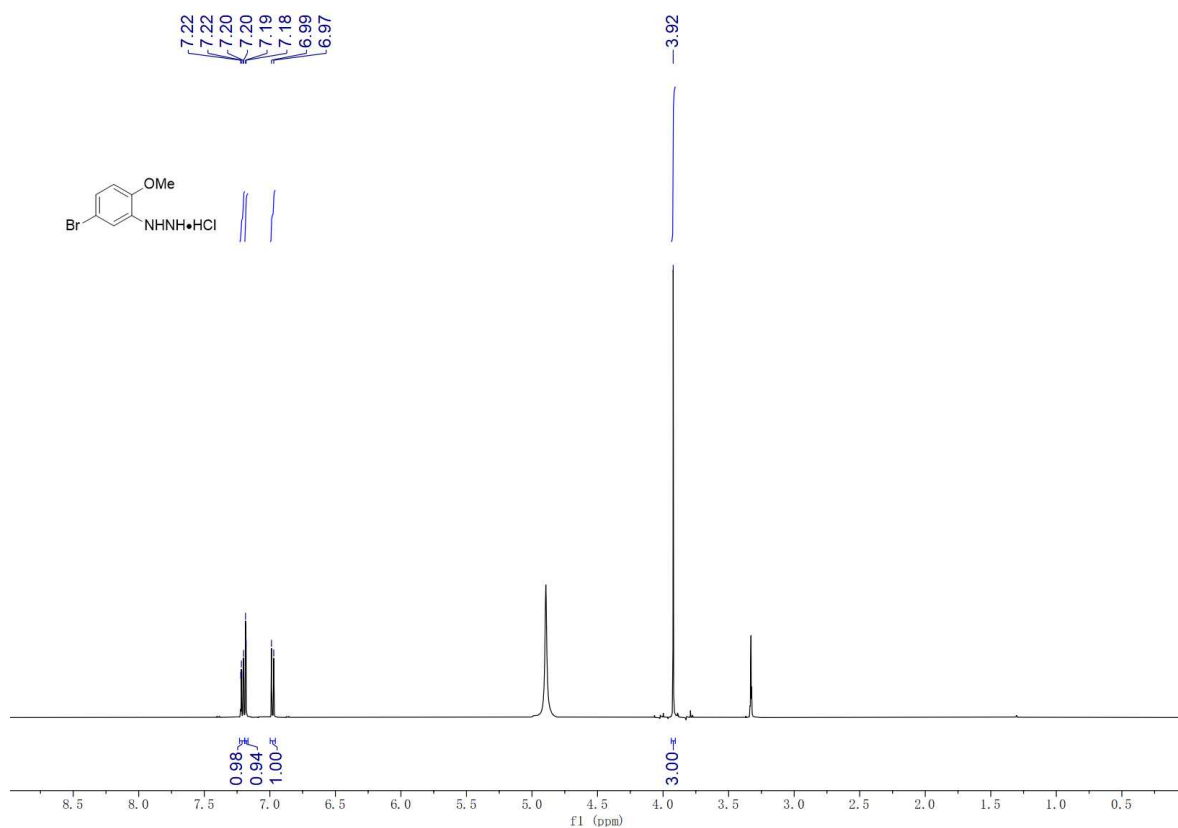
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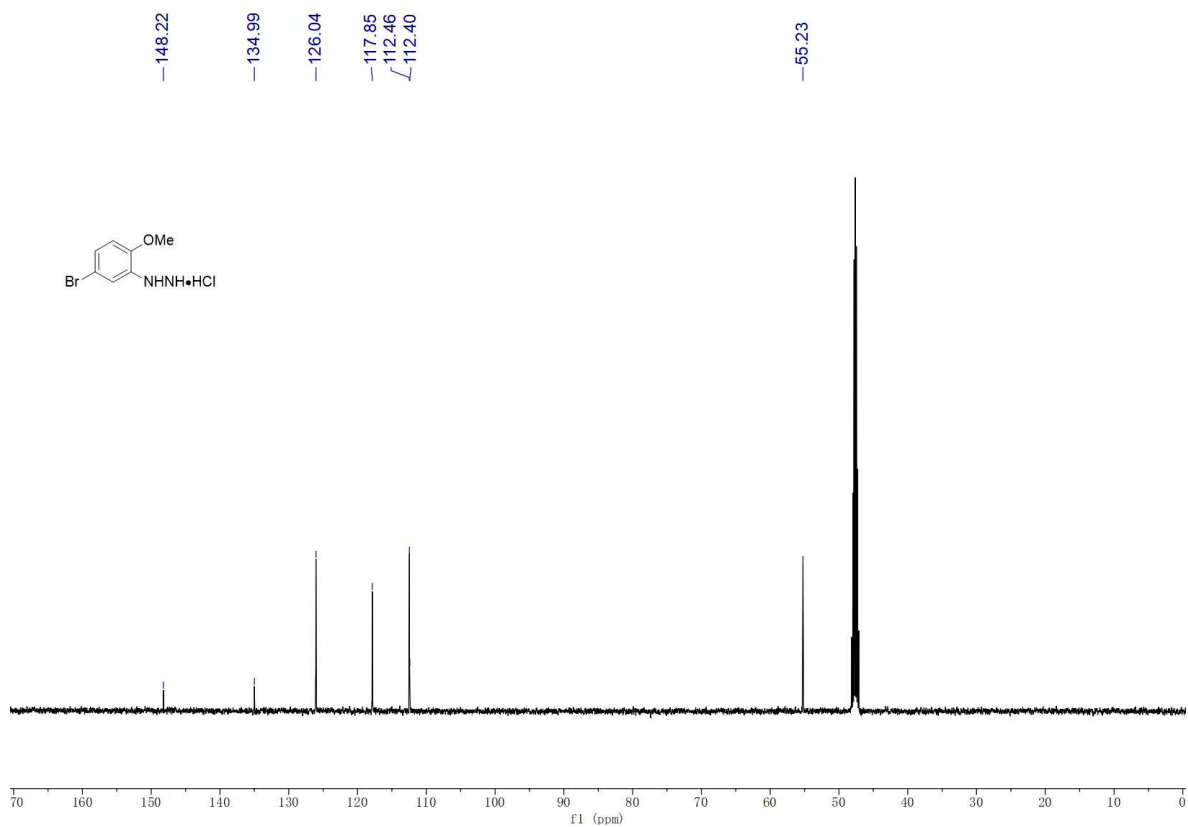
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1313 ¹H and ¹³C- NMR spectra of product.

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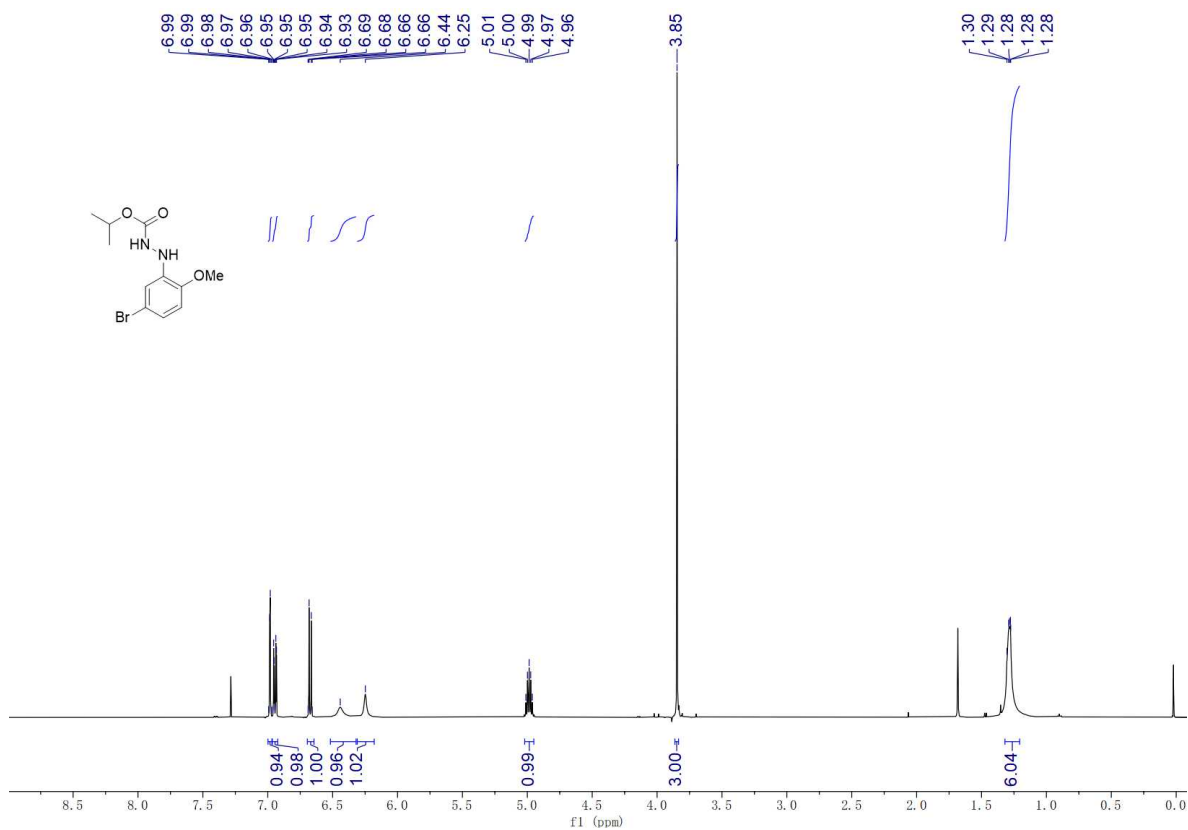
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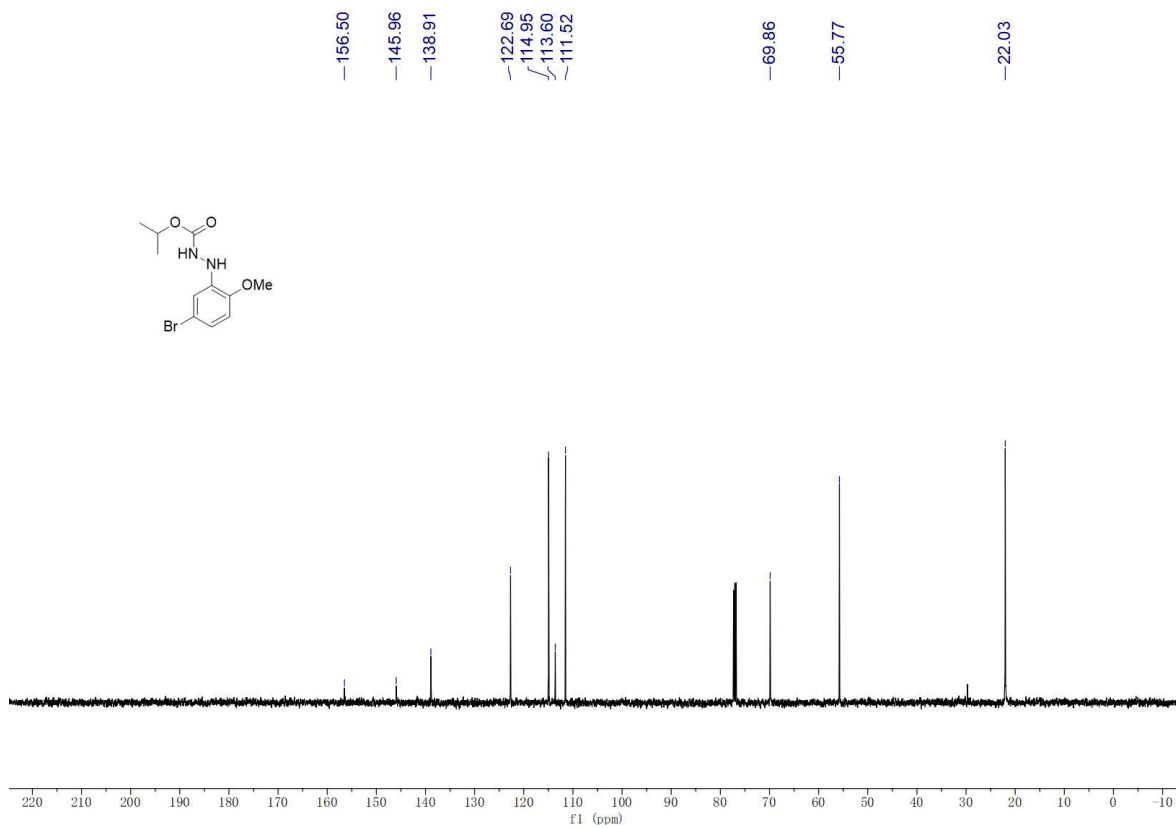
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1317 ¹H and ¹³C spectra of product 1aa

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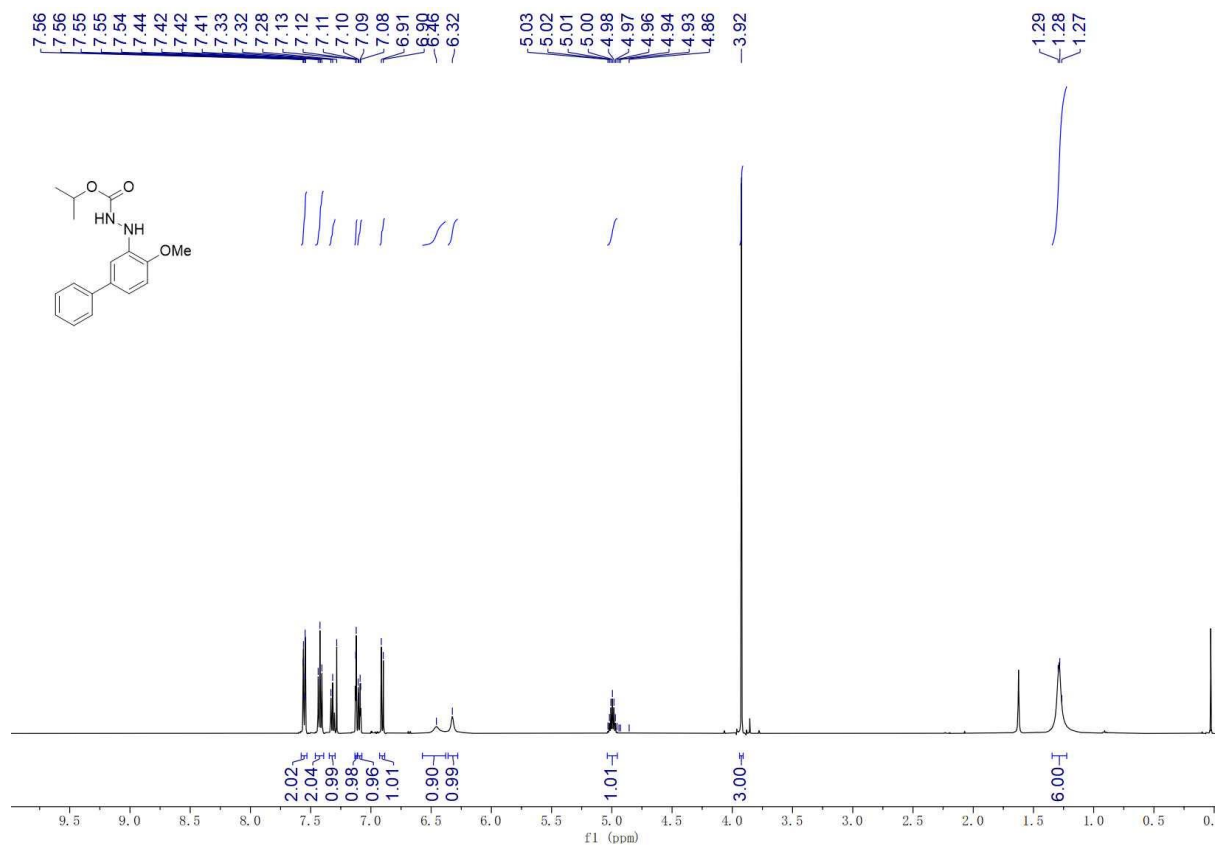
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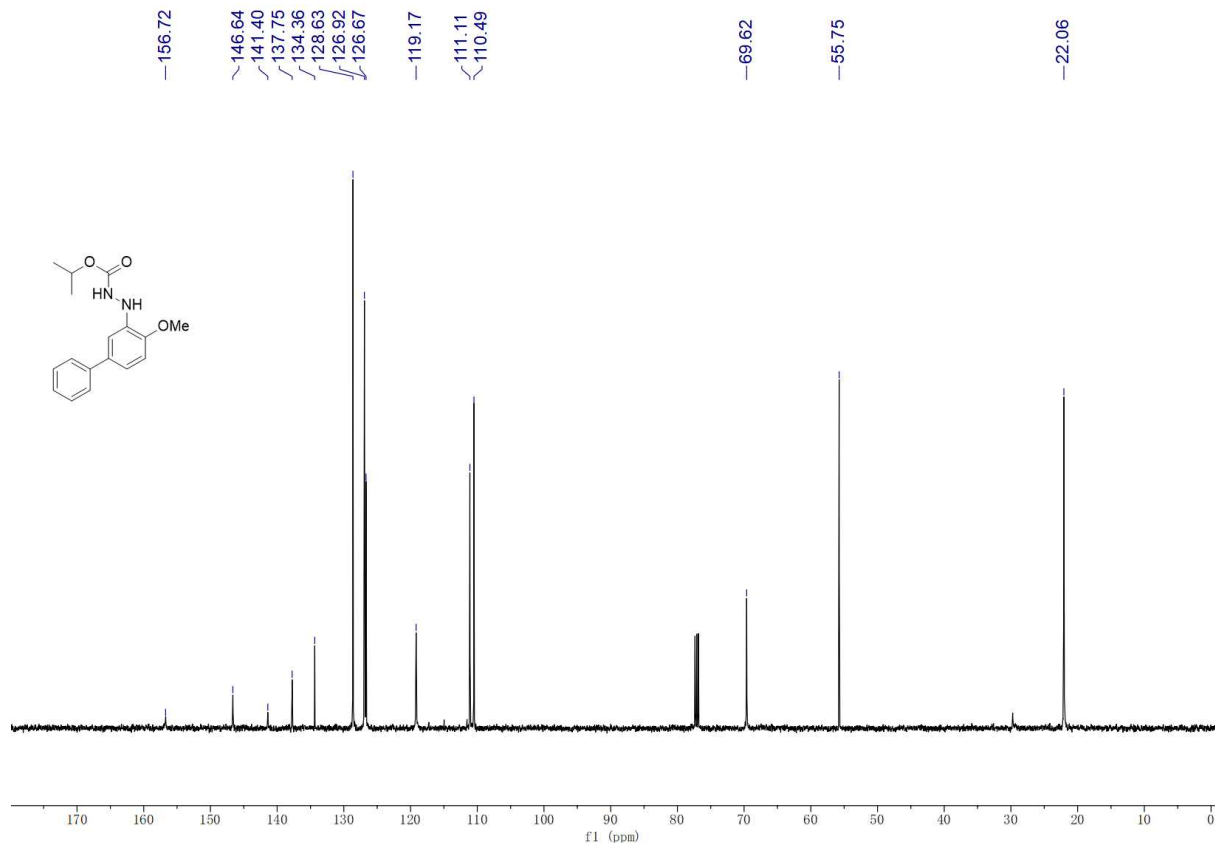
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1321 ¹H and ¹³C spectra of product bifenazate.

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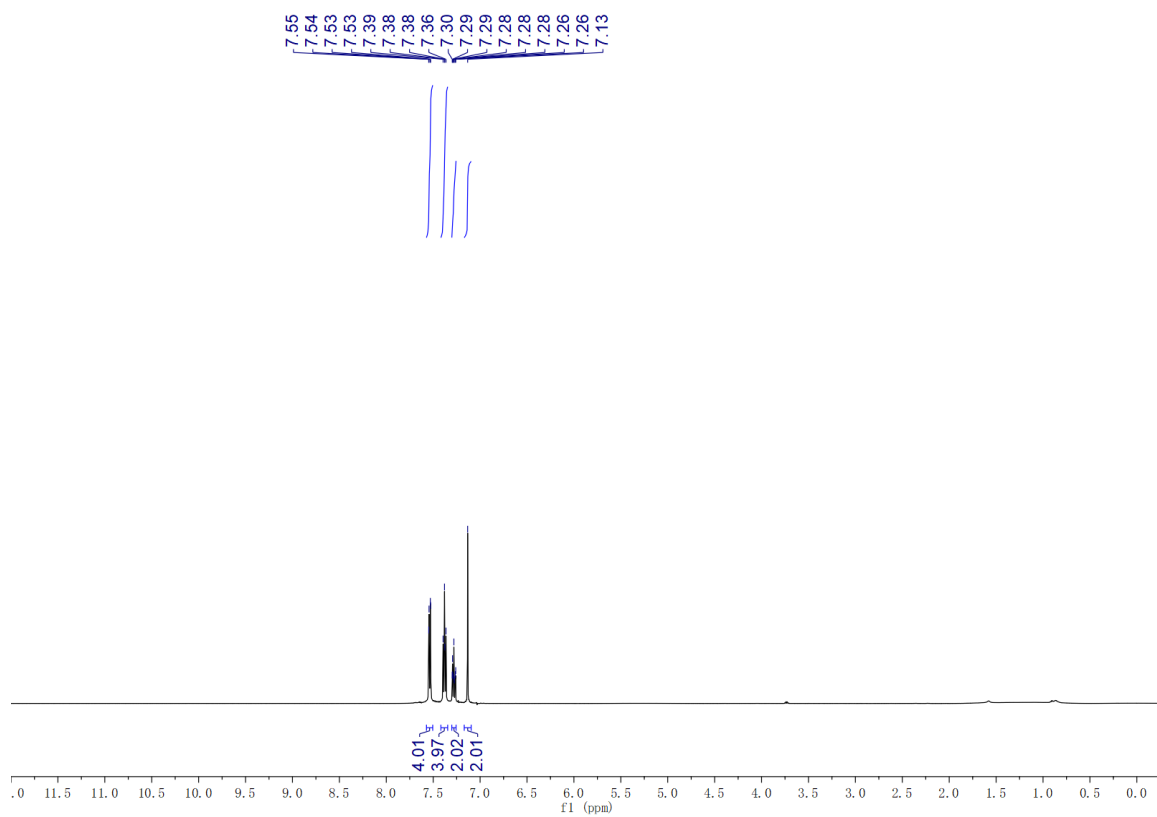
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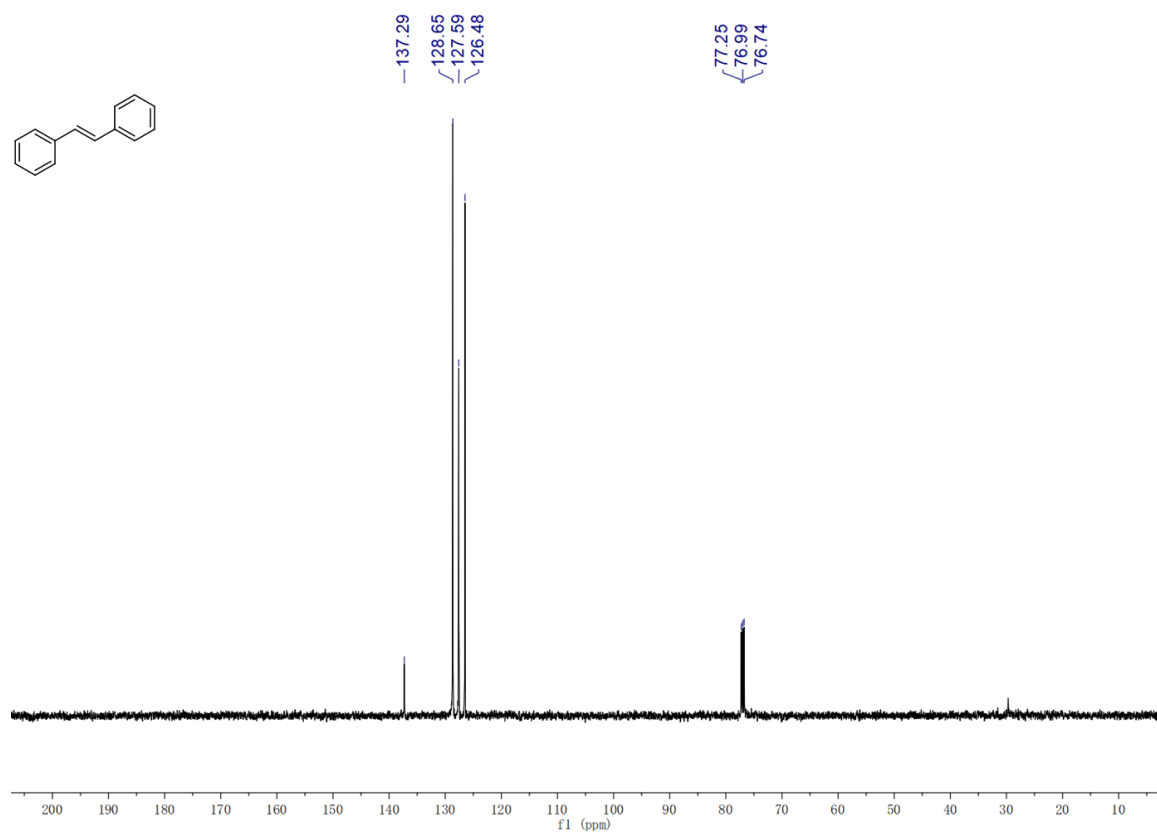
1325 ¹H and ¹³C-NMR spectra of product 8a.

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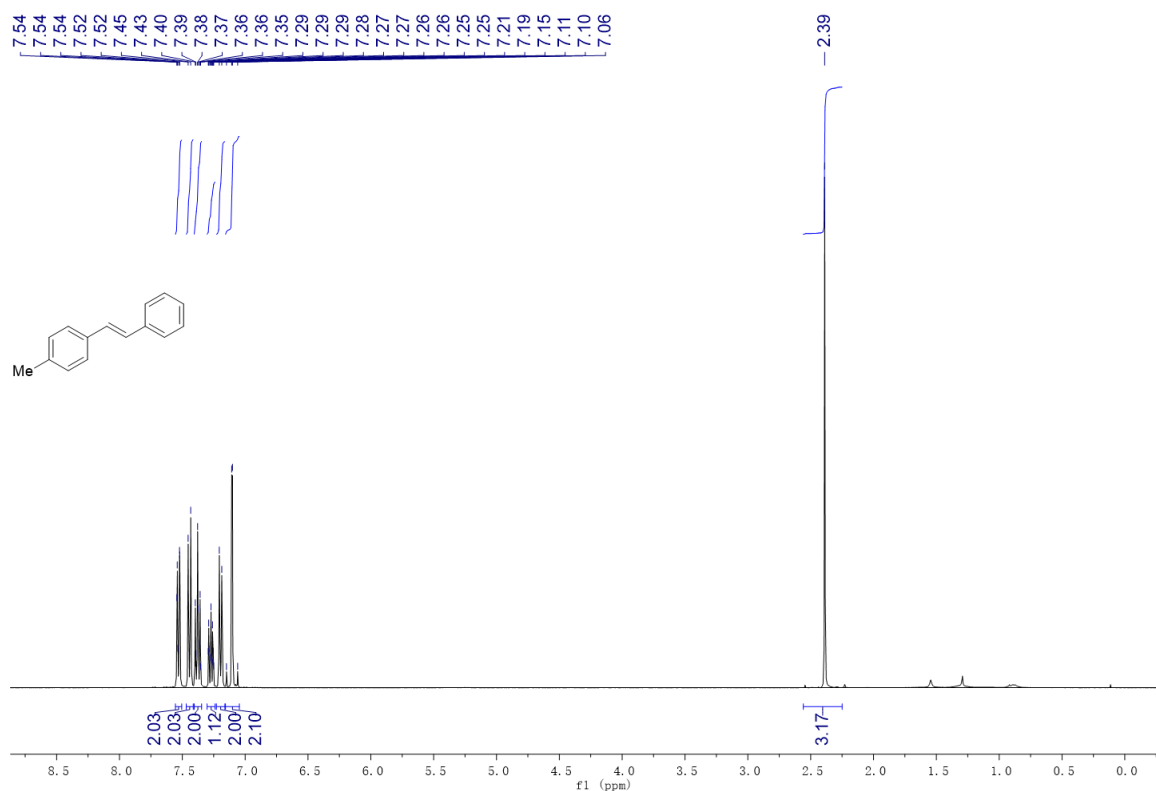
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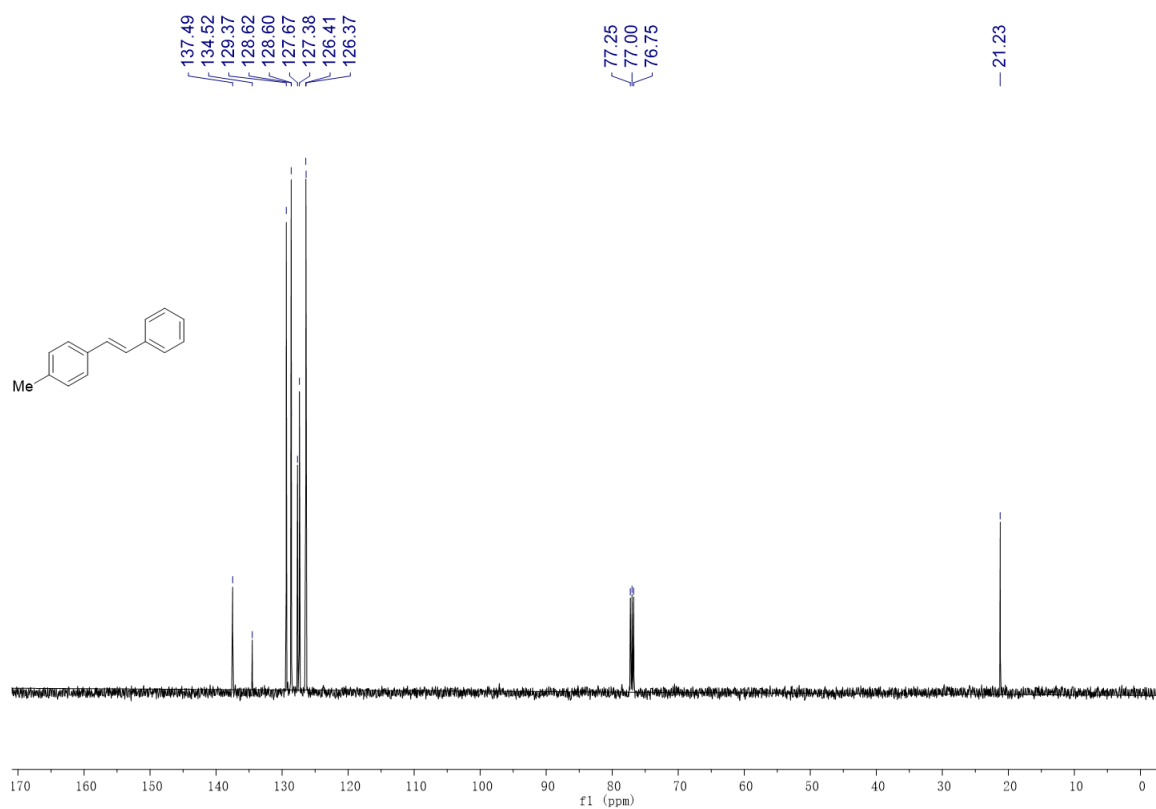
1329 ¹H and ¹³C-NMR spectra of product 8b.

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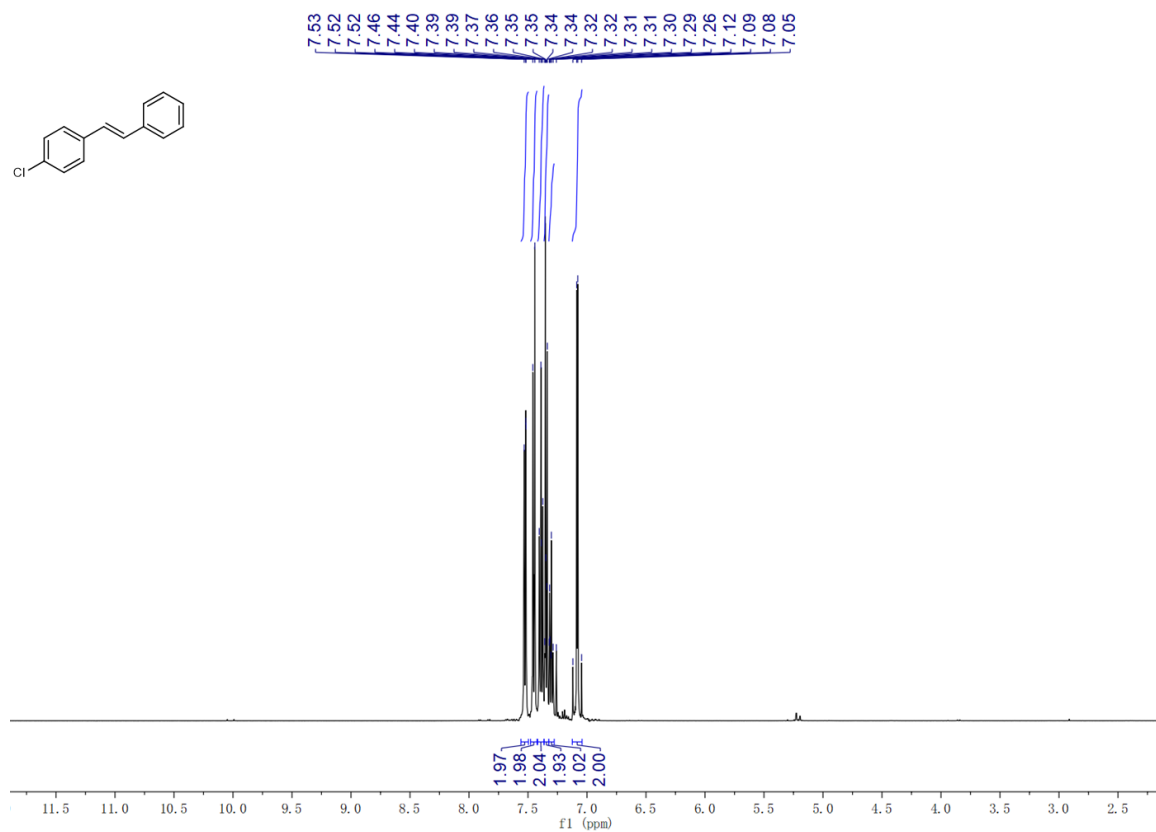


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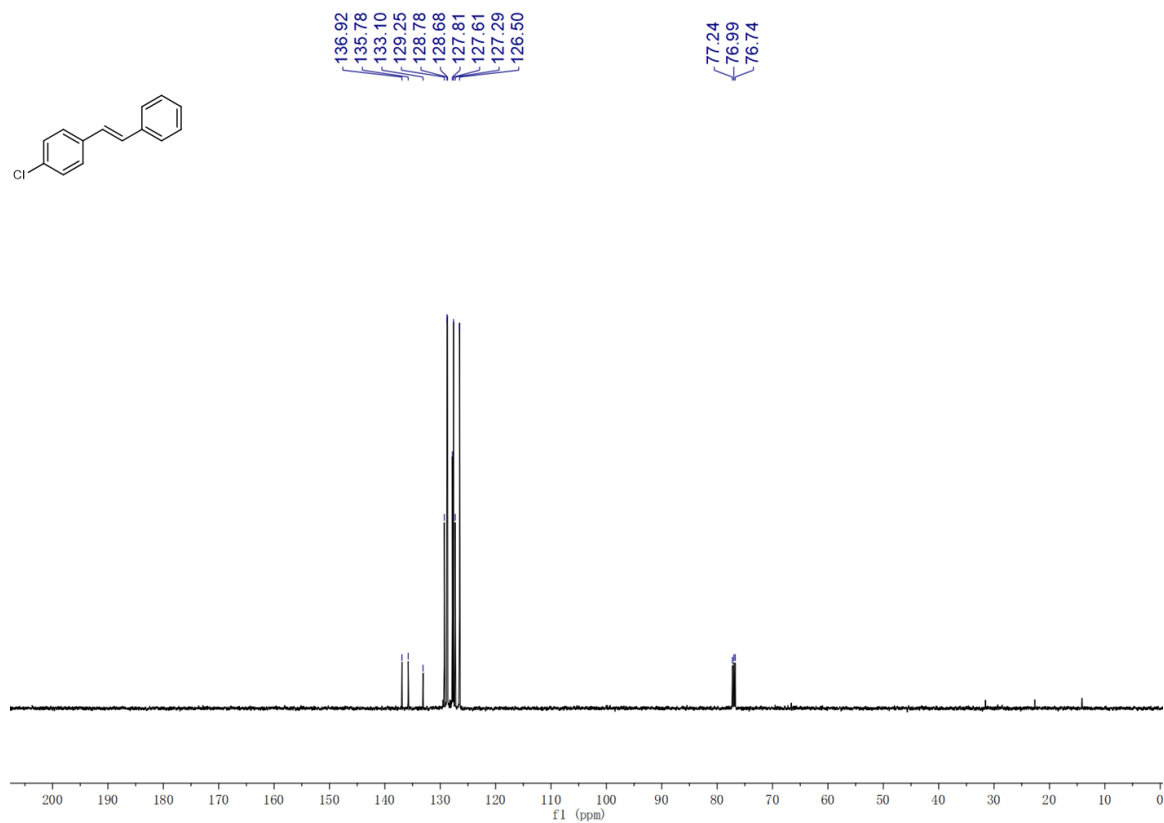
1335 ¹H and ¹³C-NMR spectra of product 8c.

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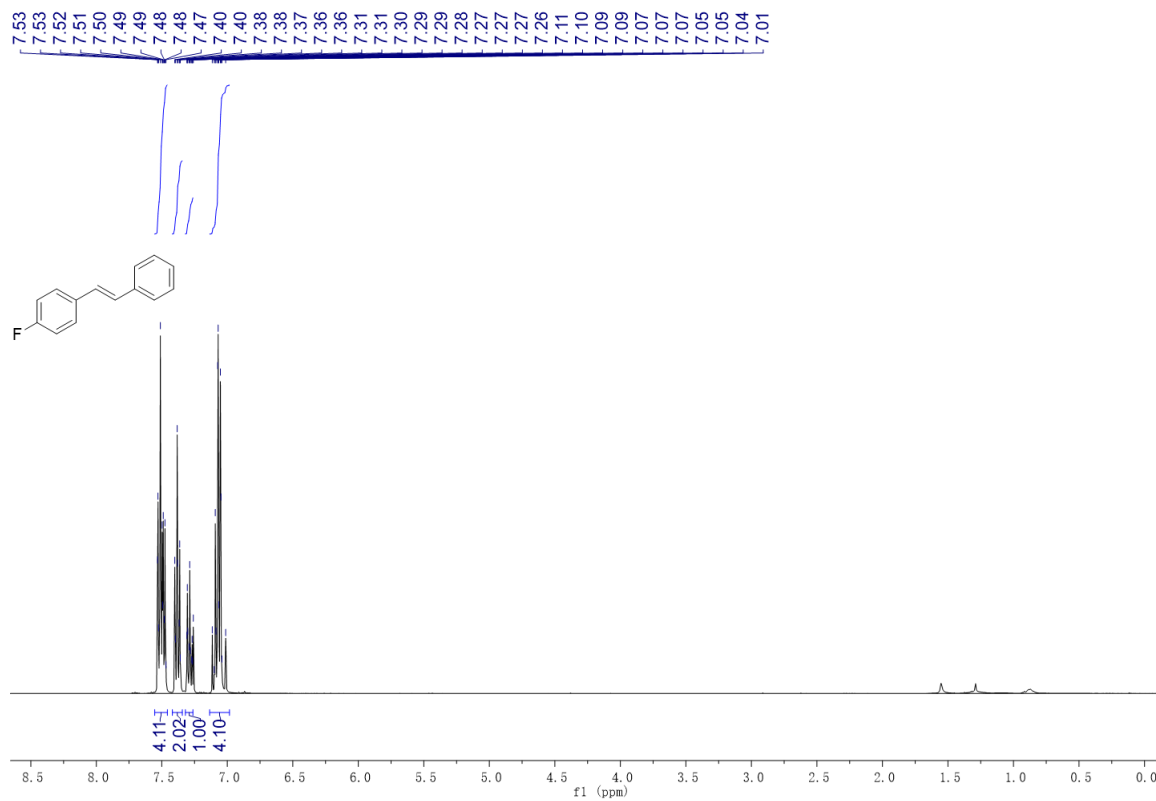
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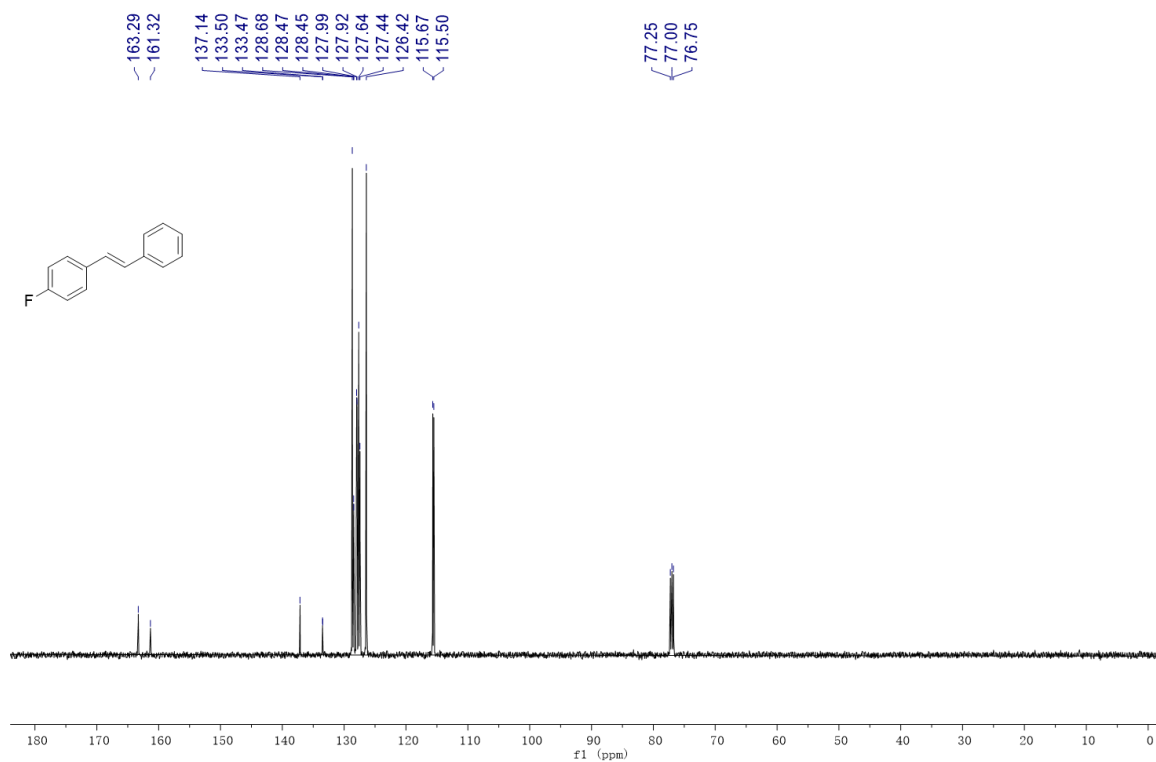
1339 ¹H and ¹³C, ⁹F-NMR spectra of product 8d.

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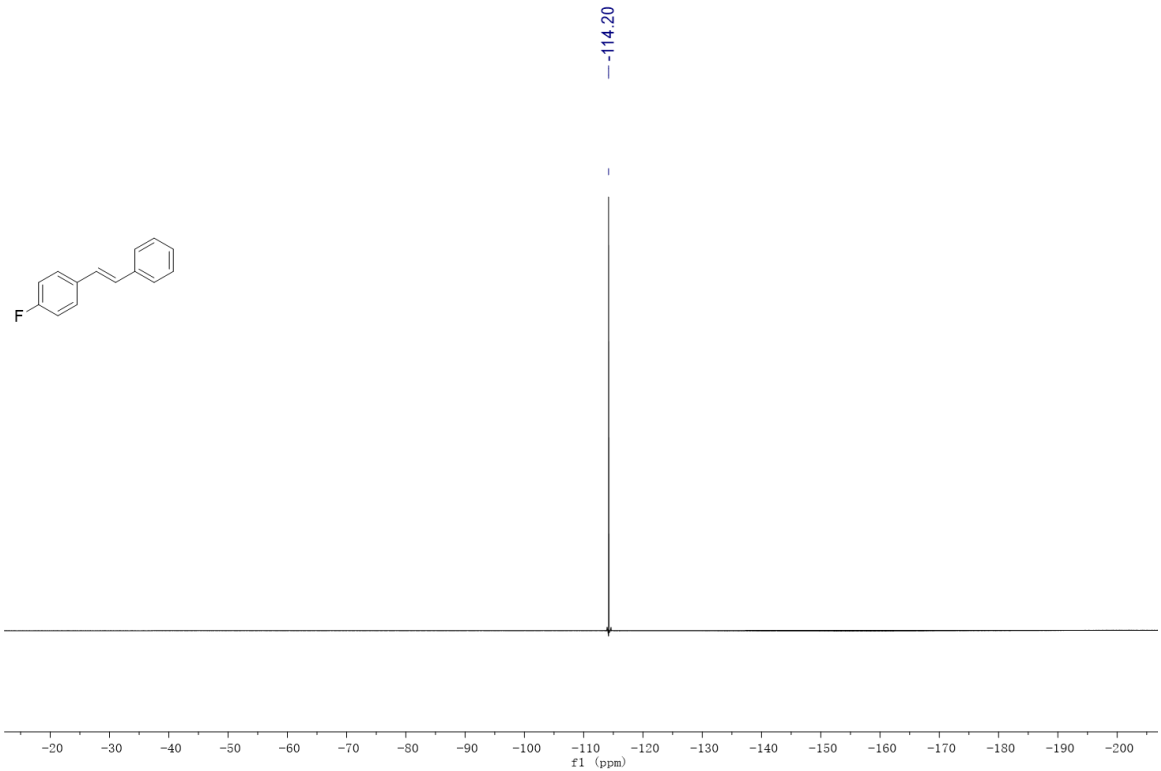


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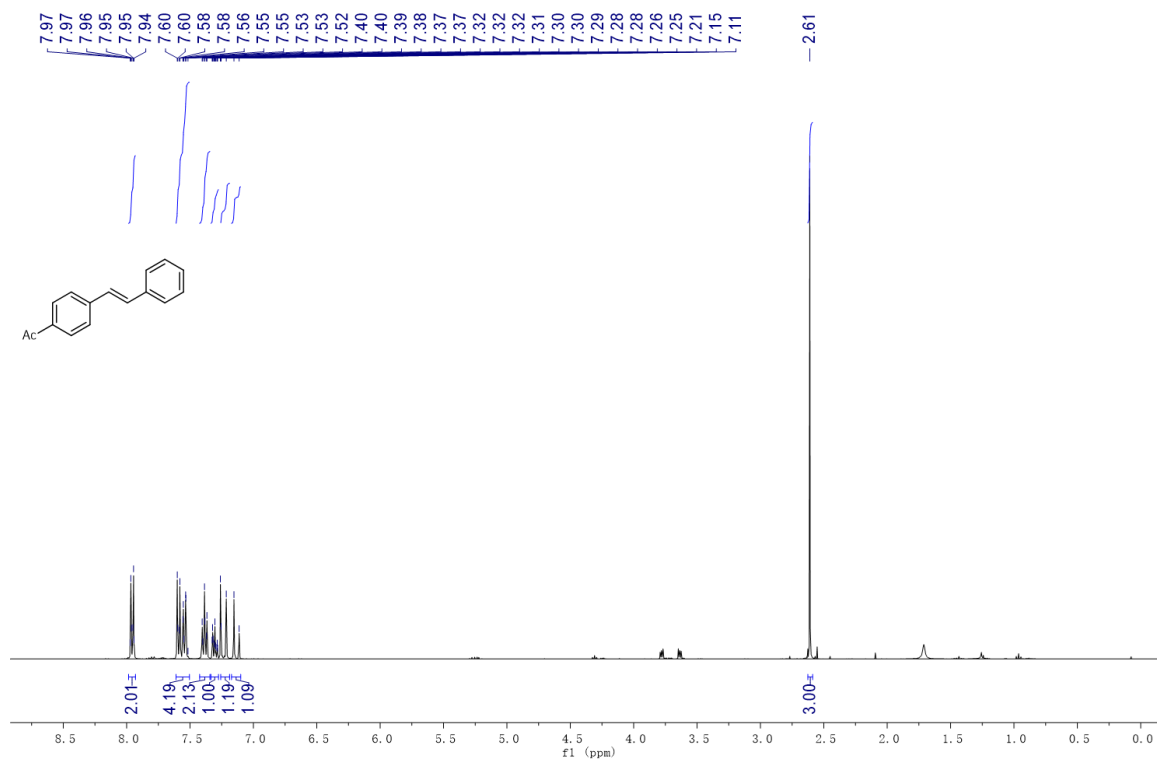


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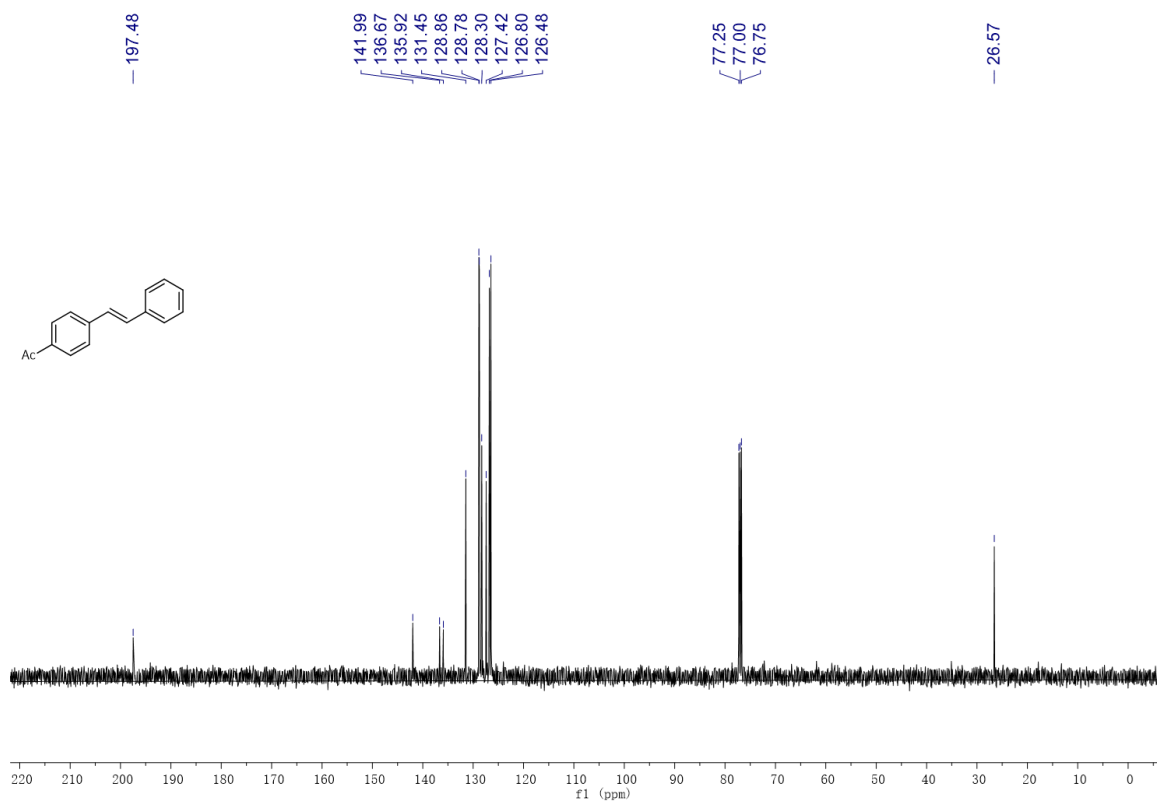
1349 ¹H, ¹³C-NMR spectra of product 8e.

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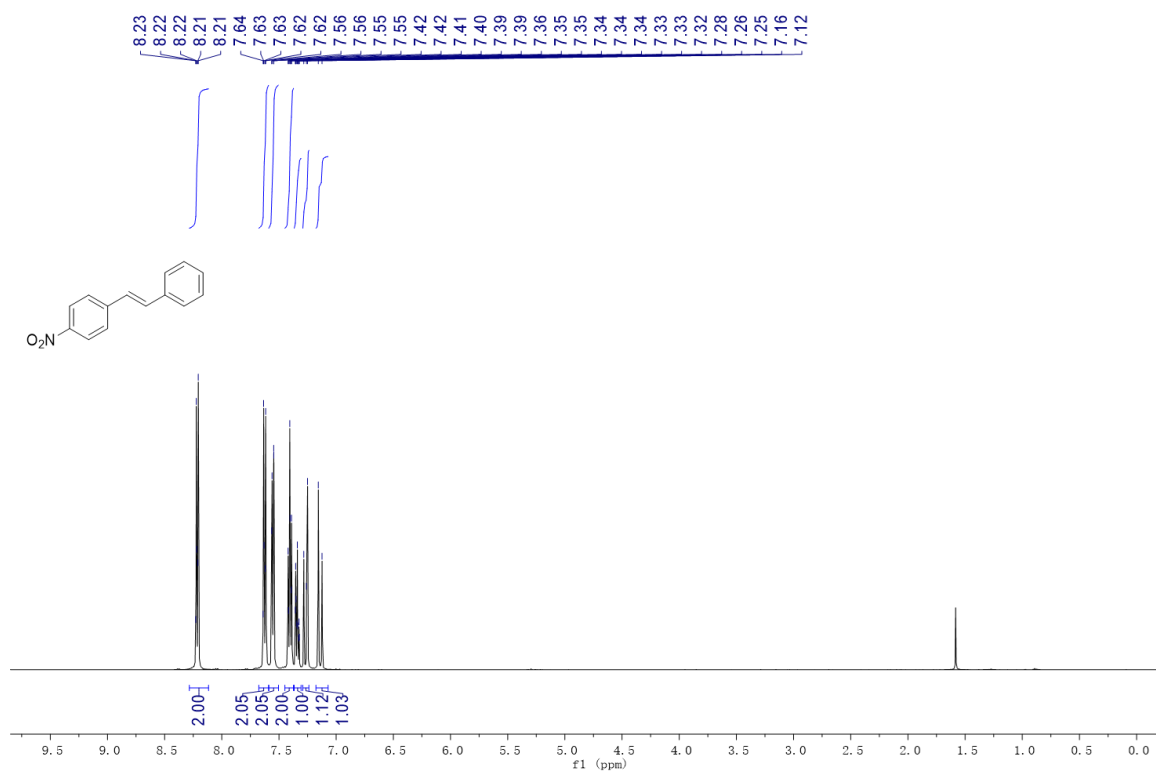


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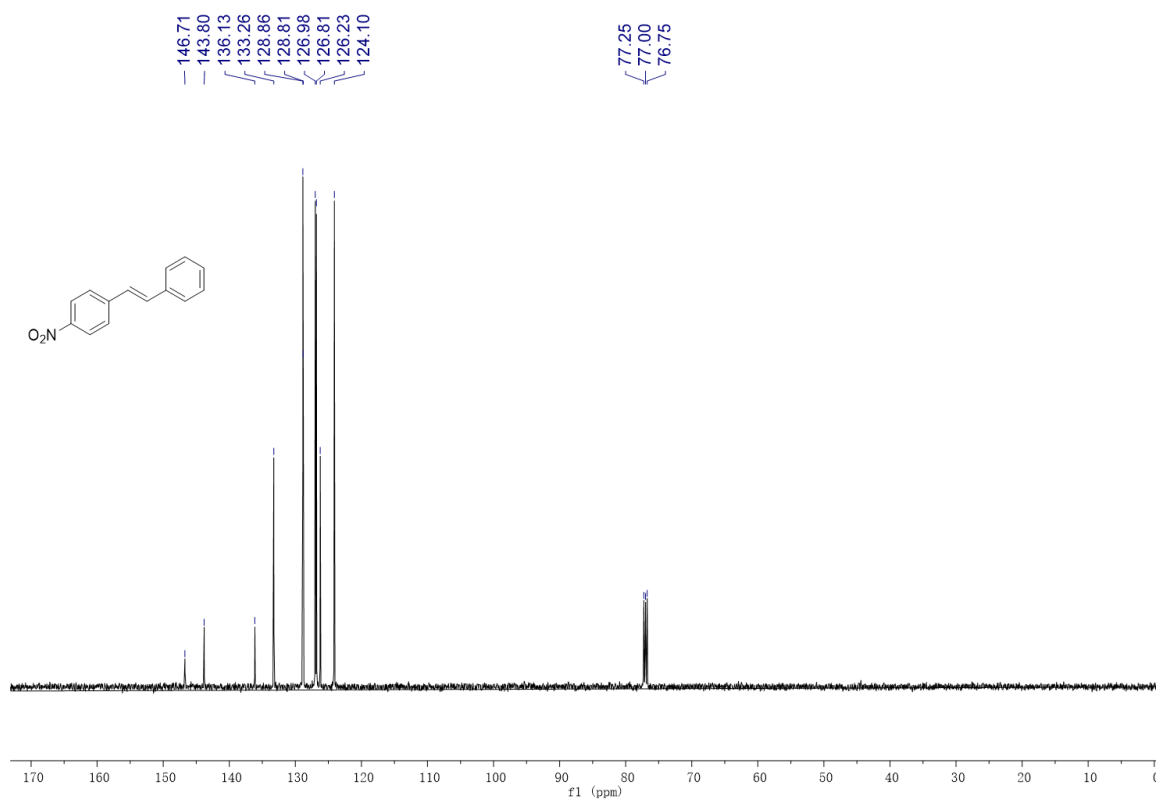
1355 ¹H, ¹³C-NMR spectra of product 8f.

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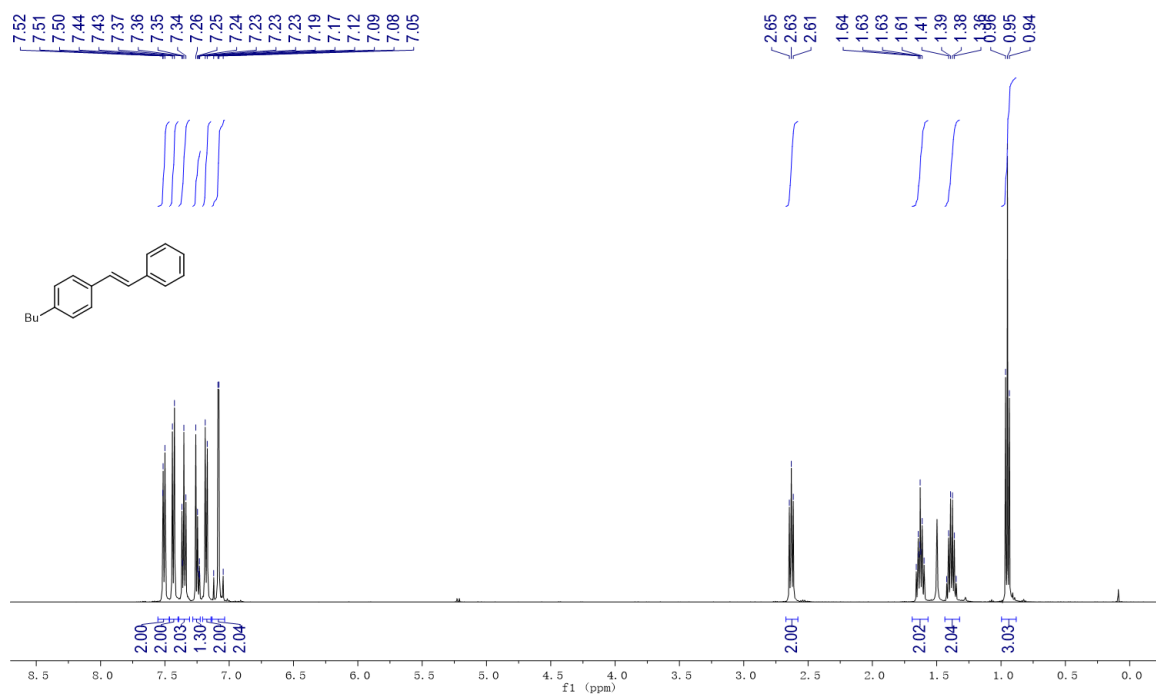


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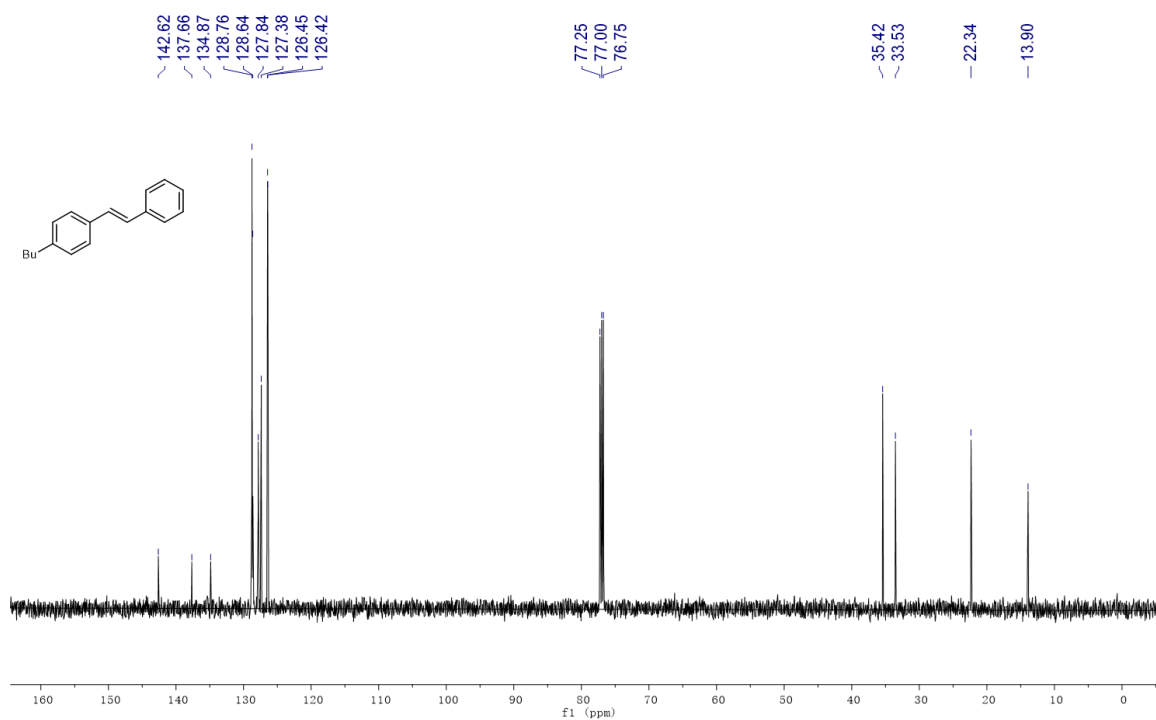
1361 ¹H, ¹³C-NMR spectra of product 8g.

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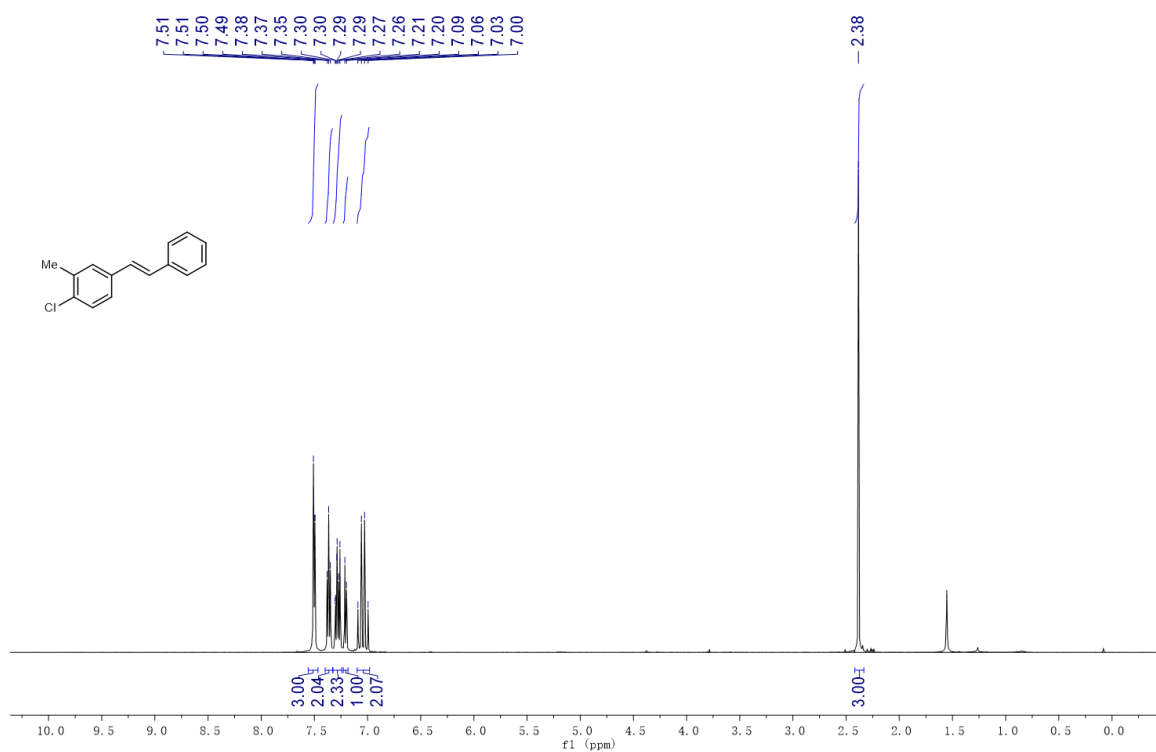


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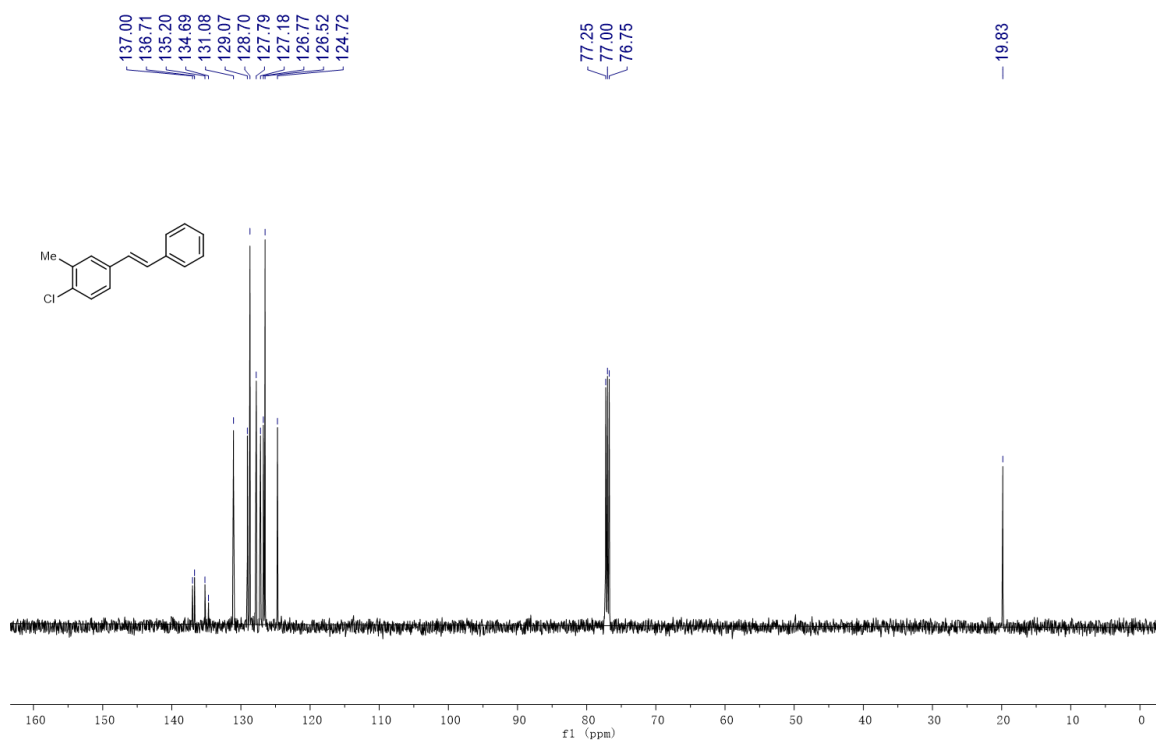
1367 ¹H and ¹³C-NMR spectra of product 8h.

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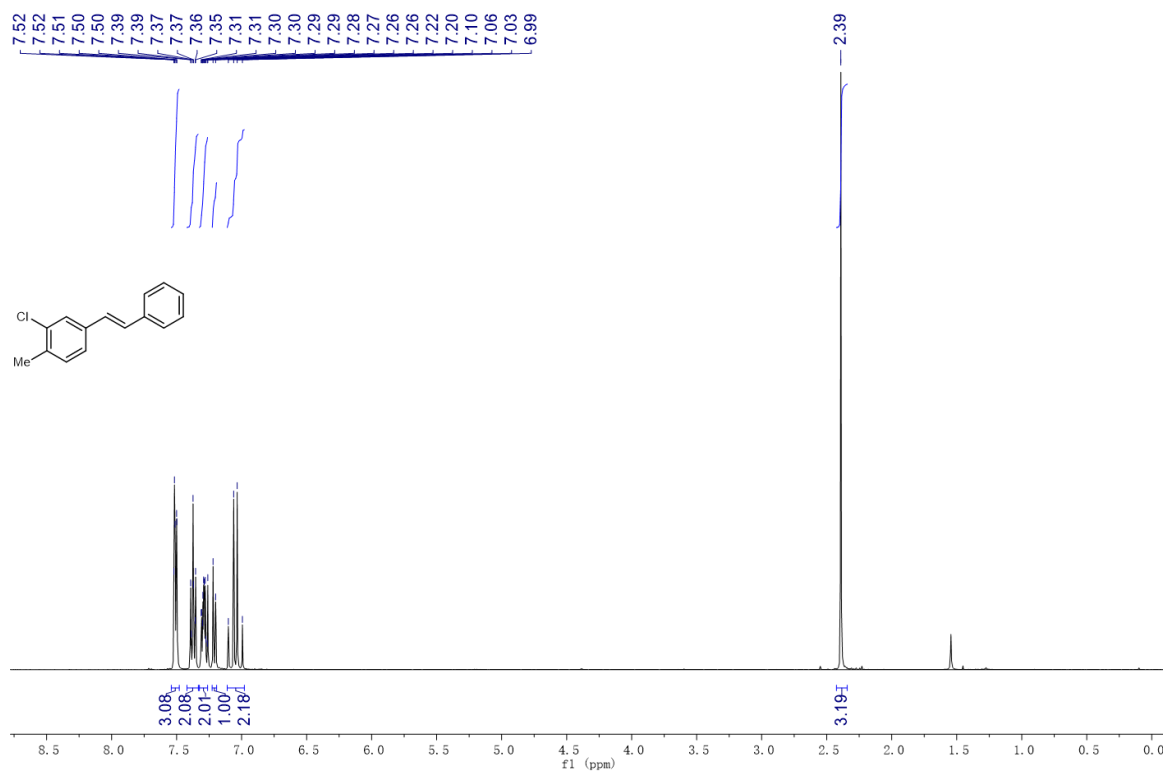


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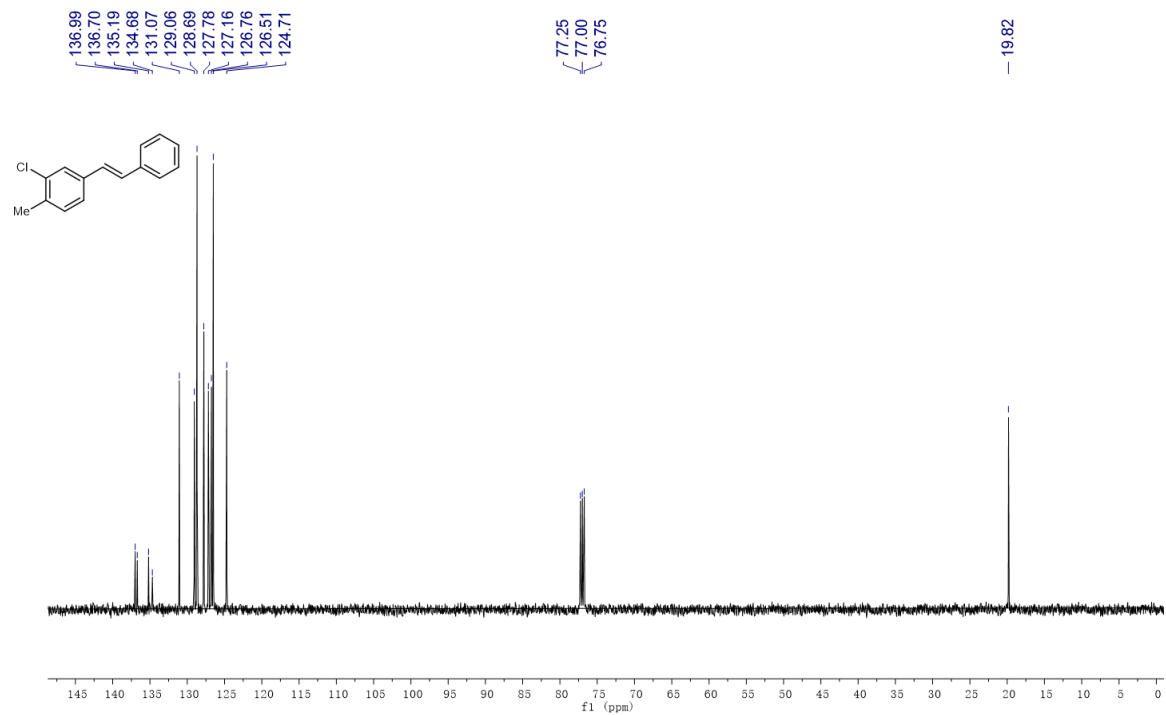
1373 ¹H and ¹³C-NMR spectra of product 8i.

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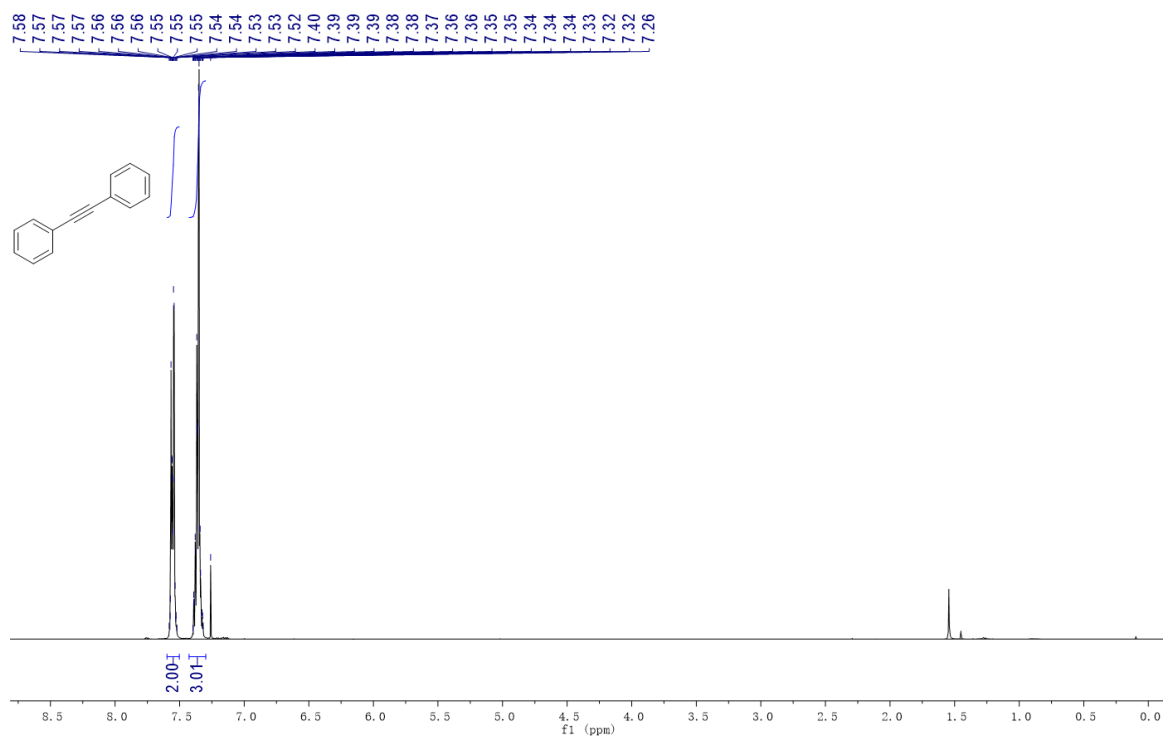


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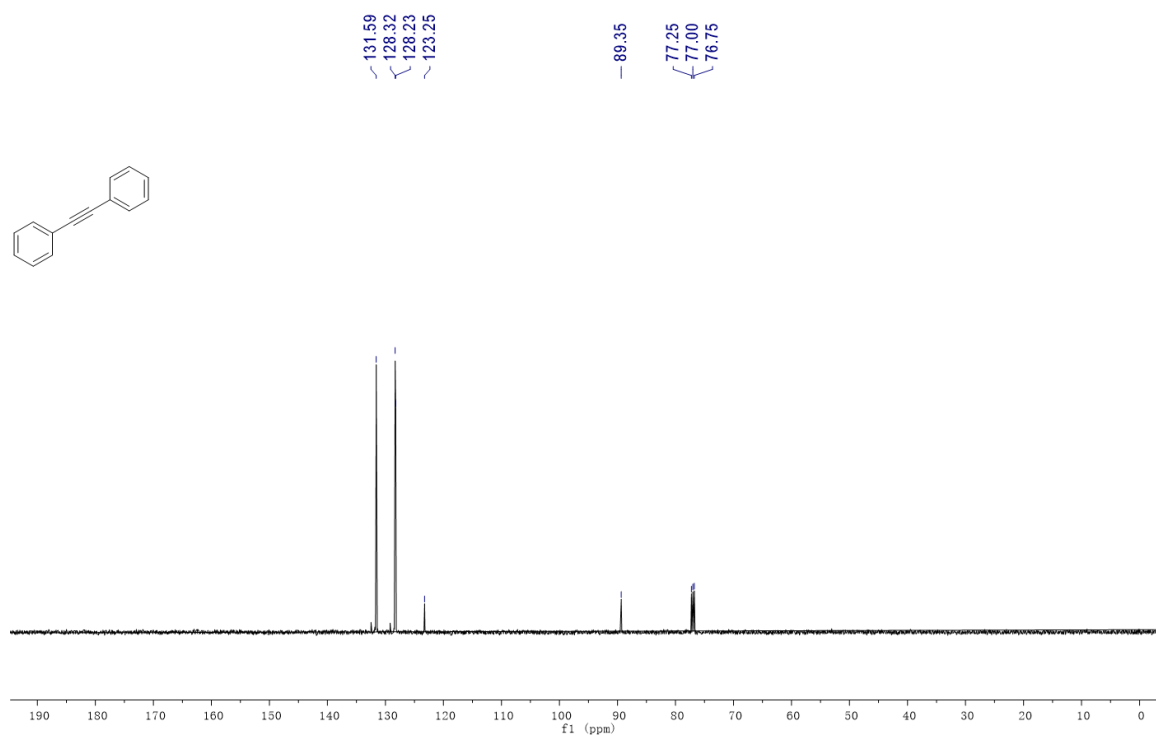
1379 ¹H and ¹³C-NMR spectra of product 9a.

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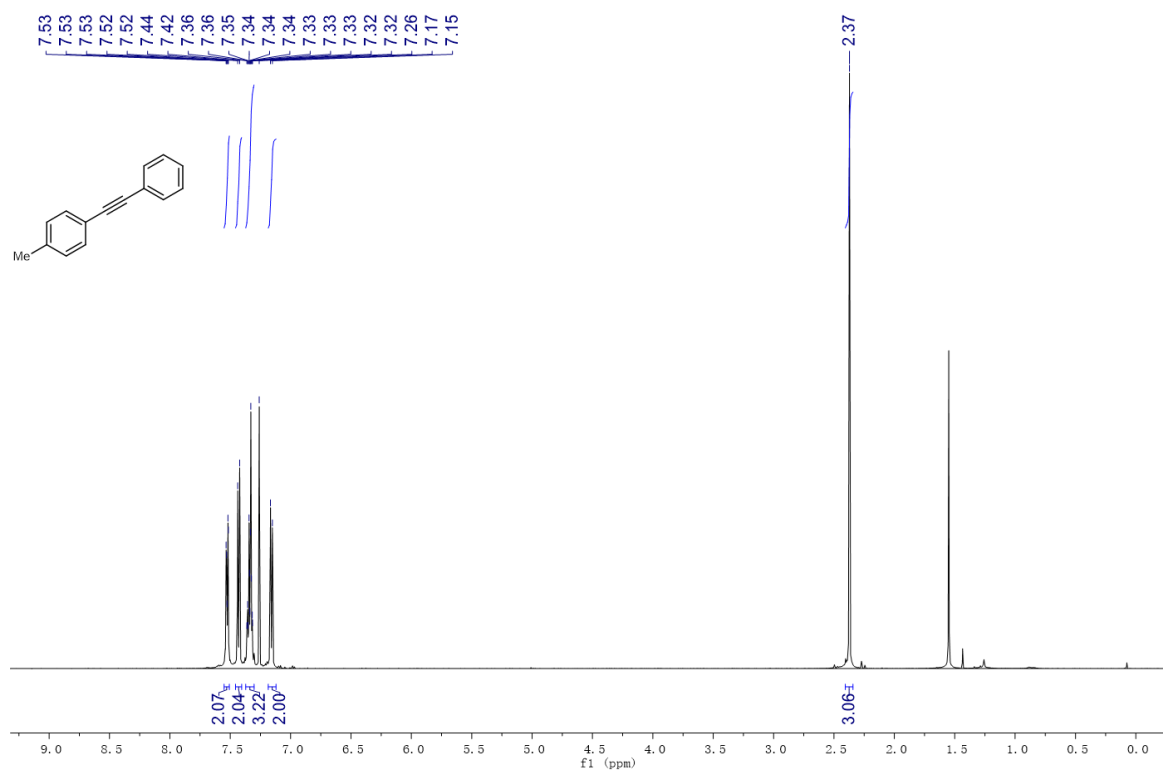


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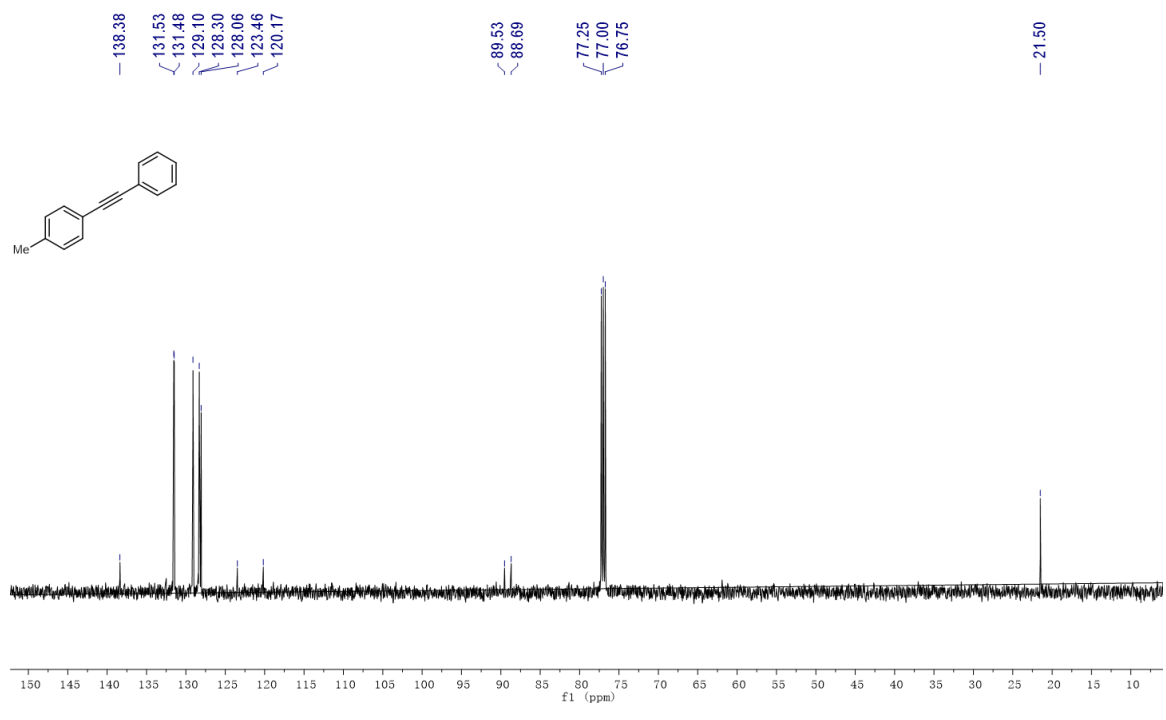
1385 **¹H and ¹³C-NMR spectra of product 9b.**

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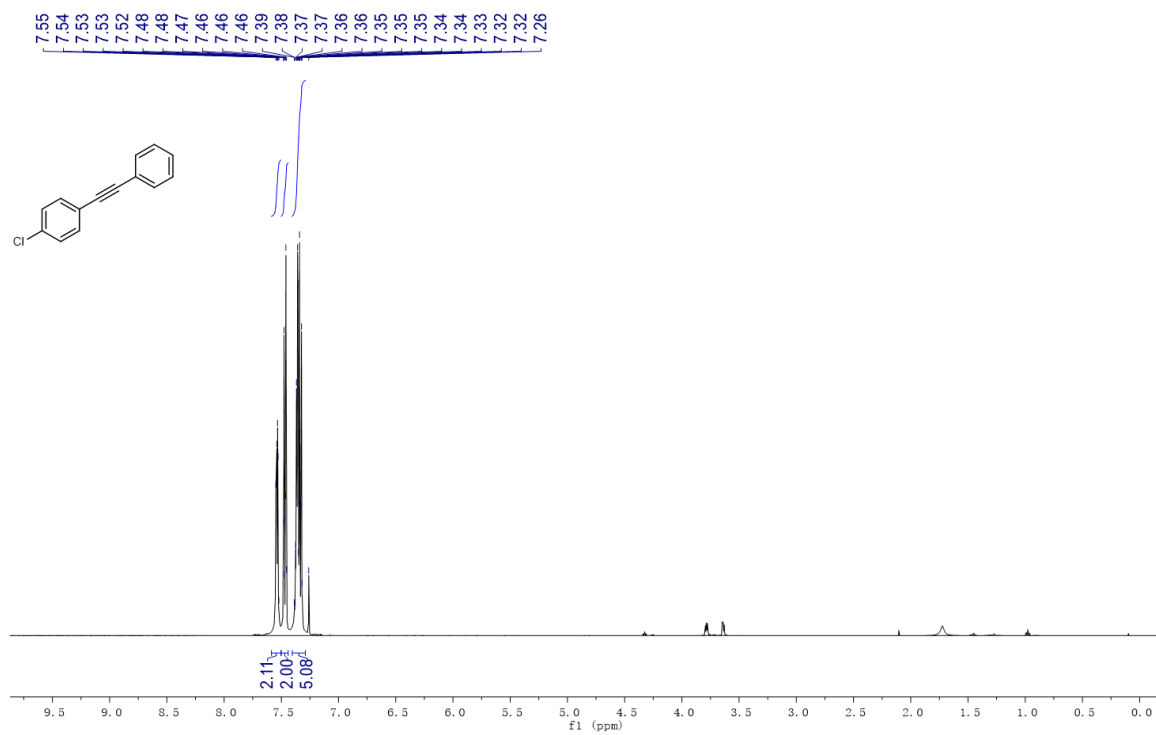


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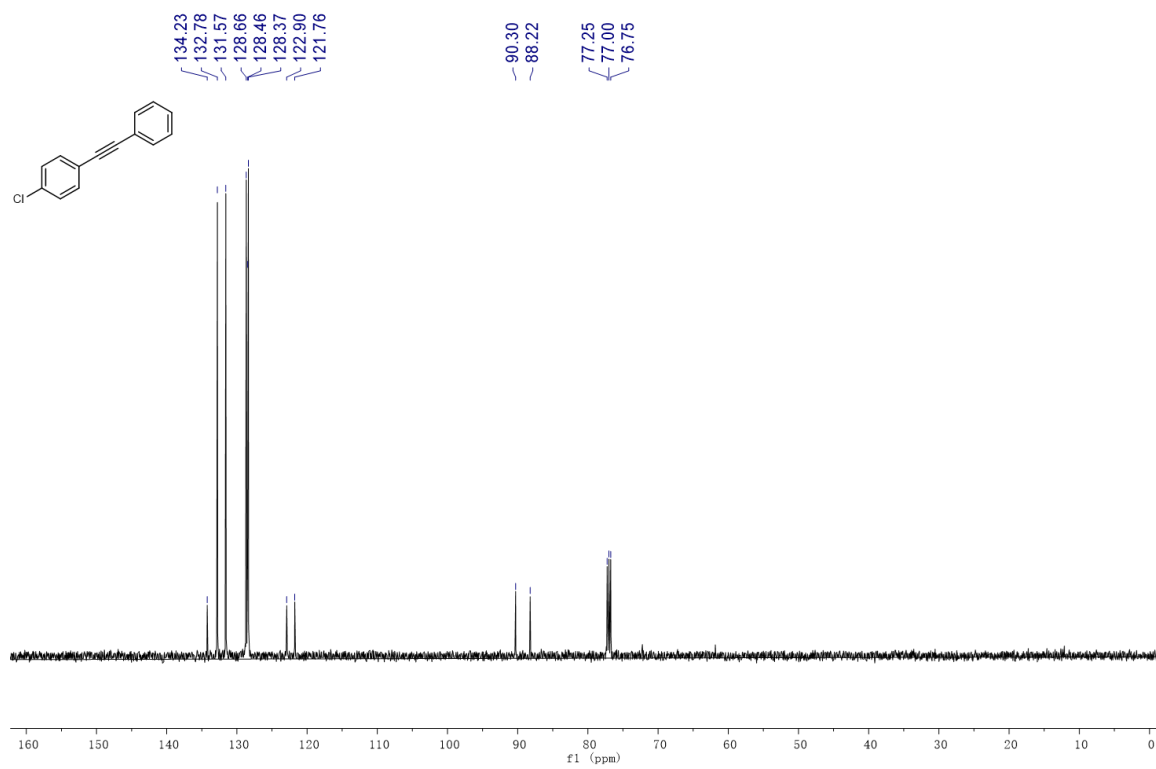
1391 ¹H and ¹³C-NMR spectra of product 9c.

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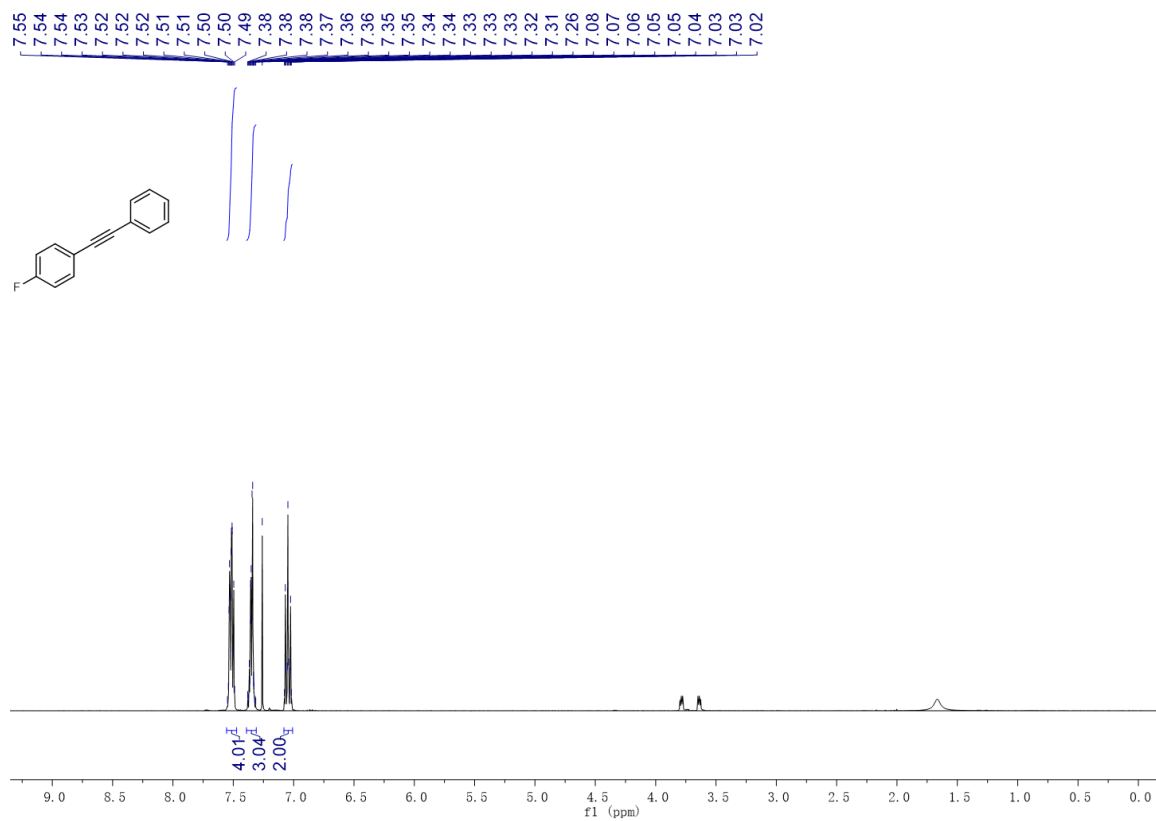


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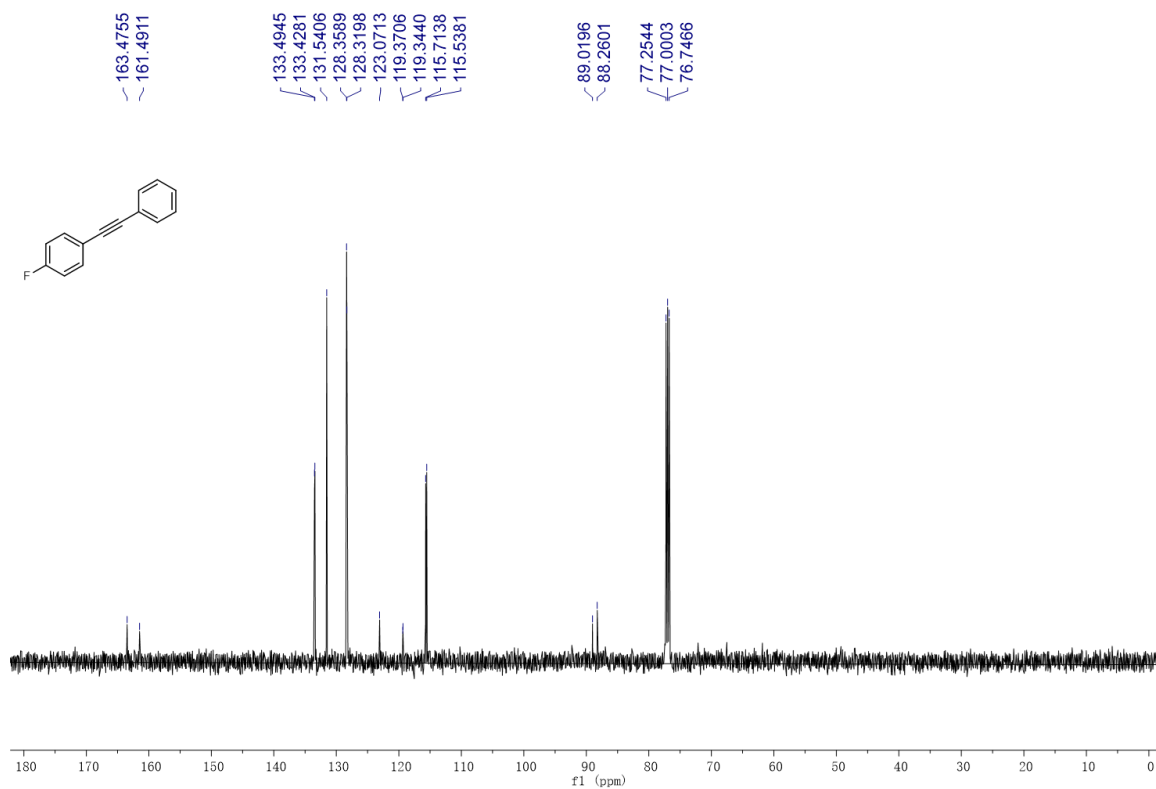
1397 ¹H and ¹³C, ¹⁹F-NMR spectra of product 9d.

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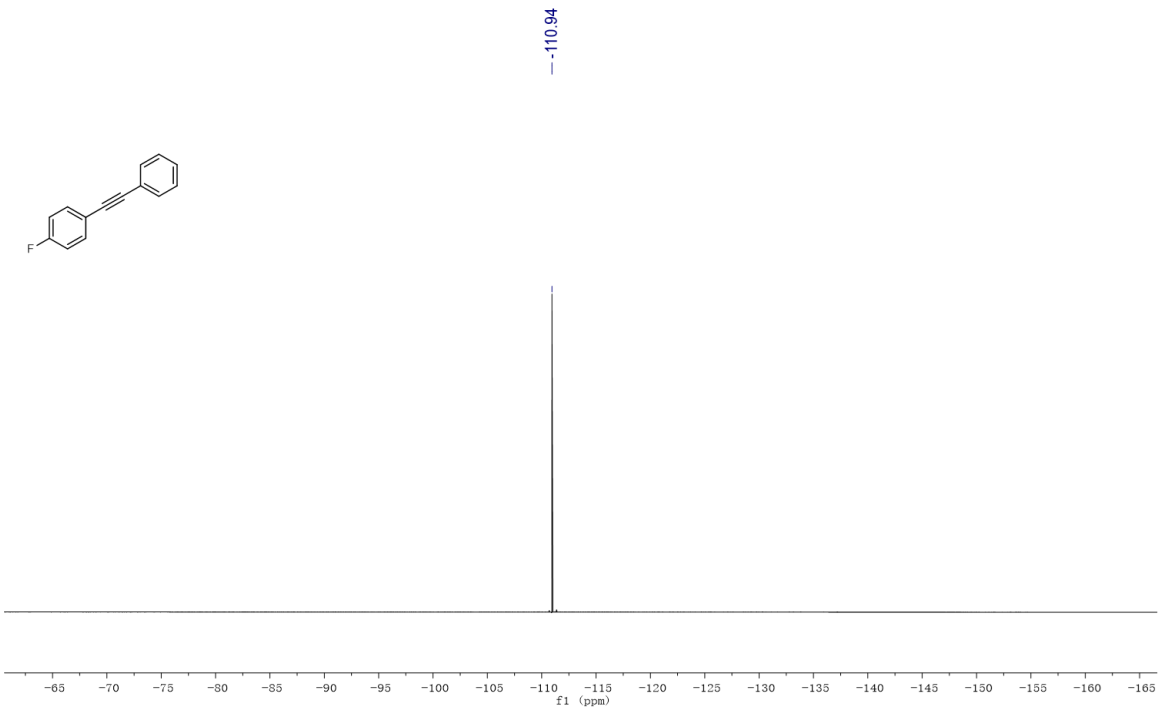
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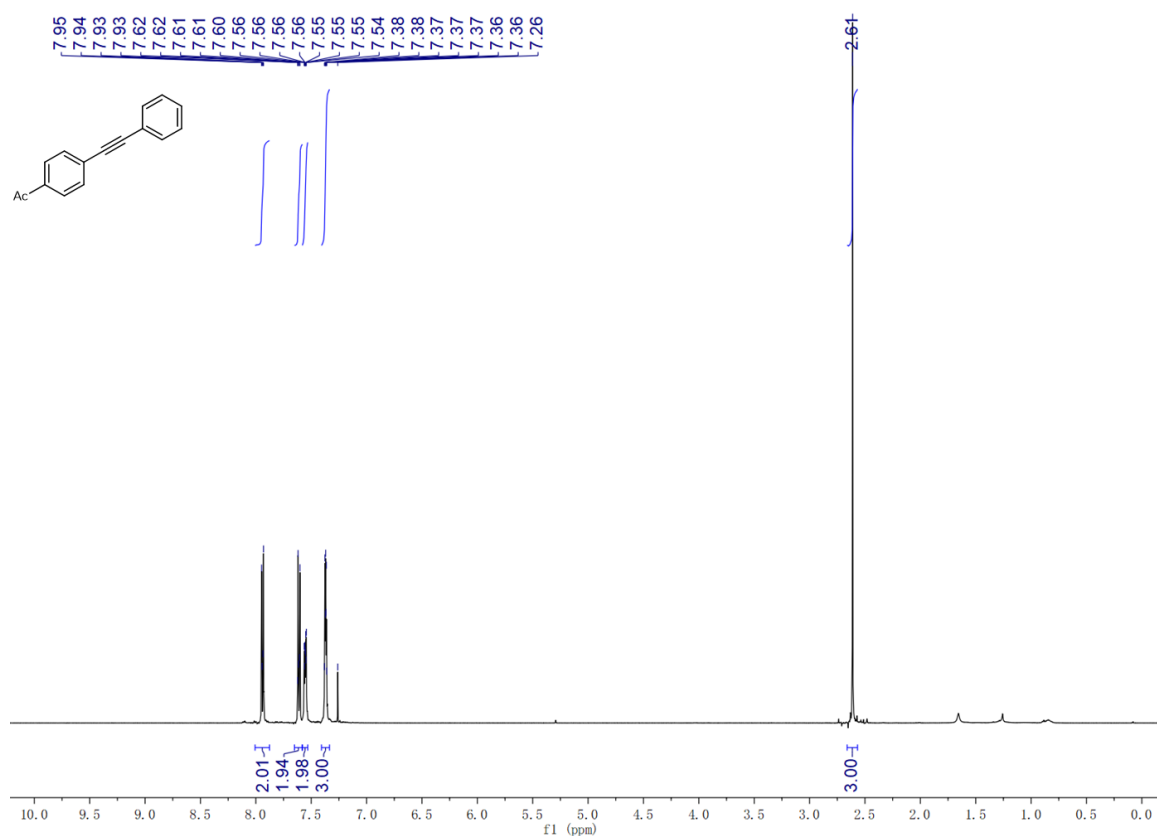
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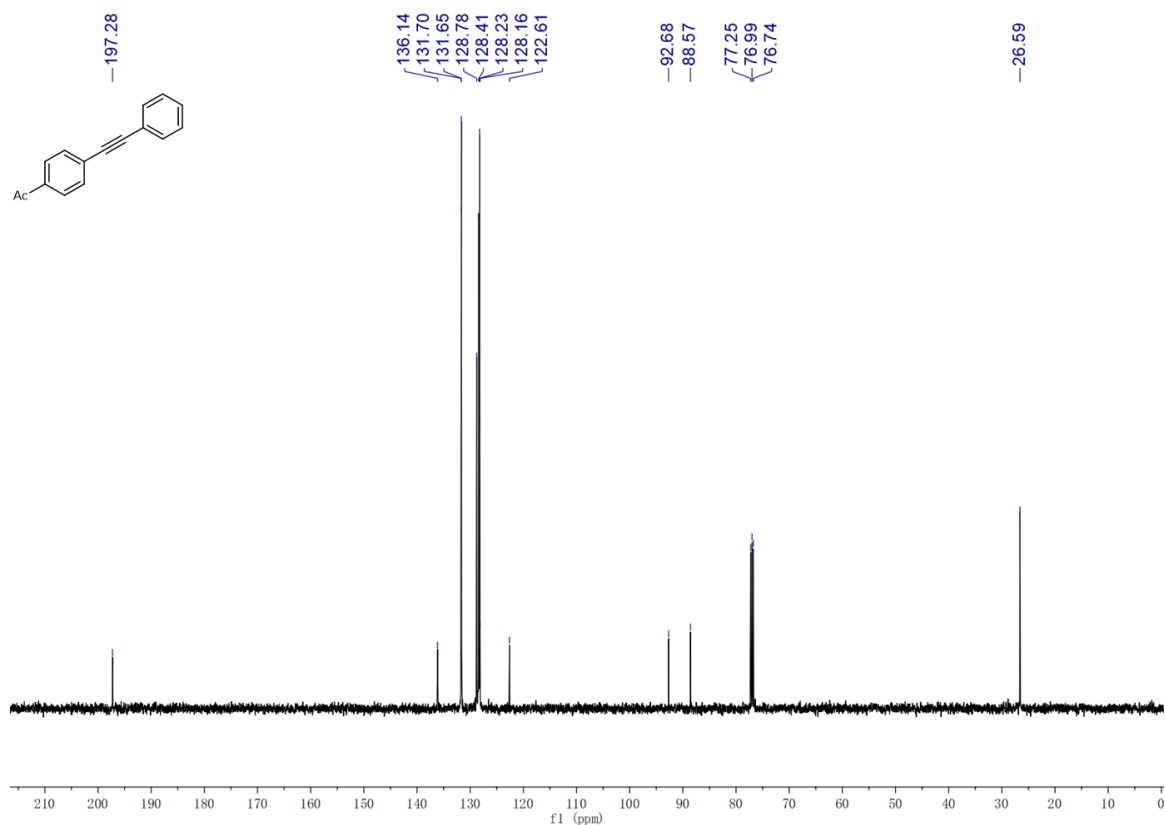
1404 ¹H and ¹³C-NMR spectra of product 9e.

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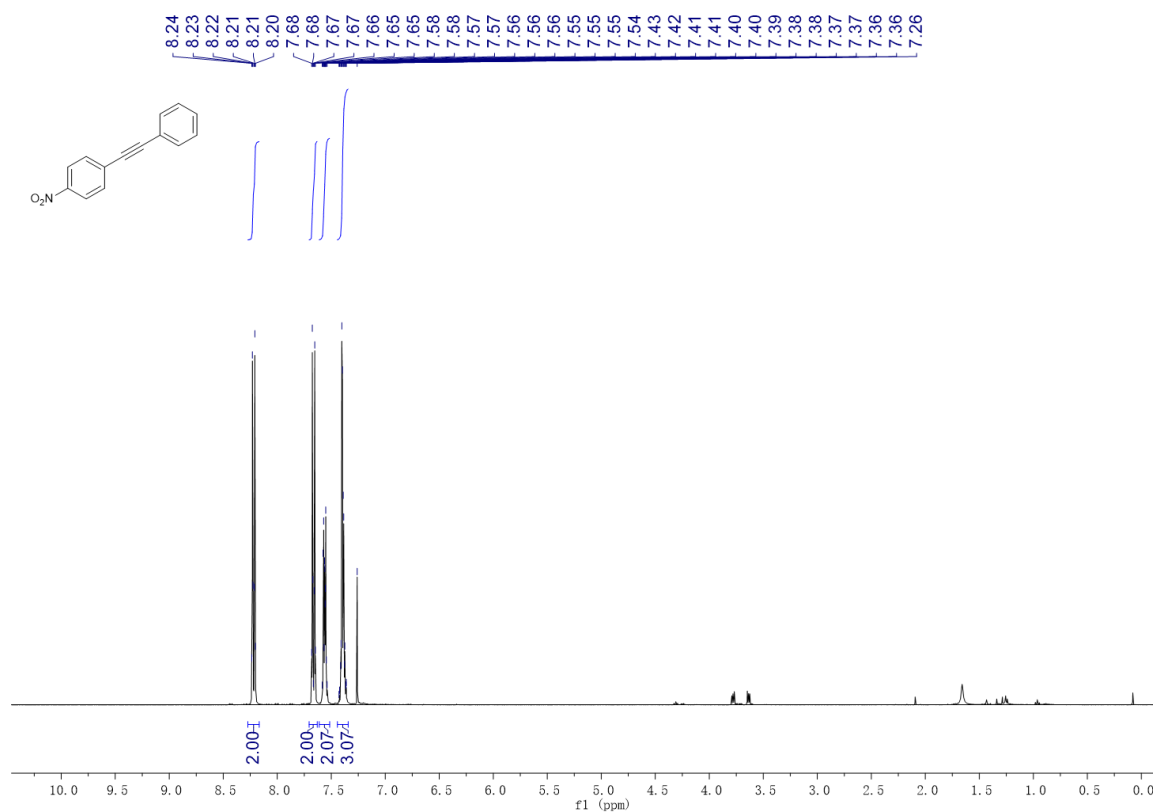
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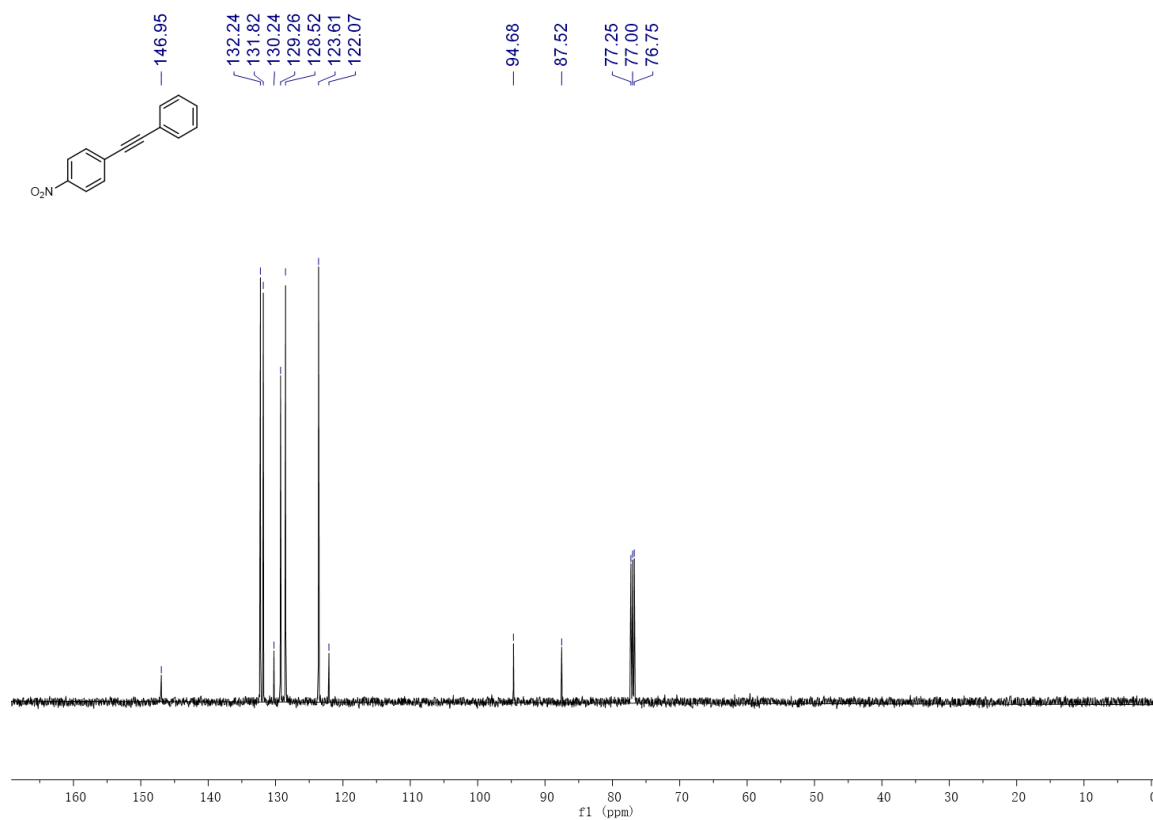
1409 **¹H and ¹³C-NMR spectra of product 9f.**

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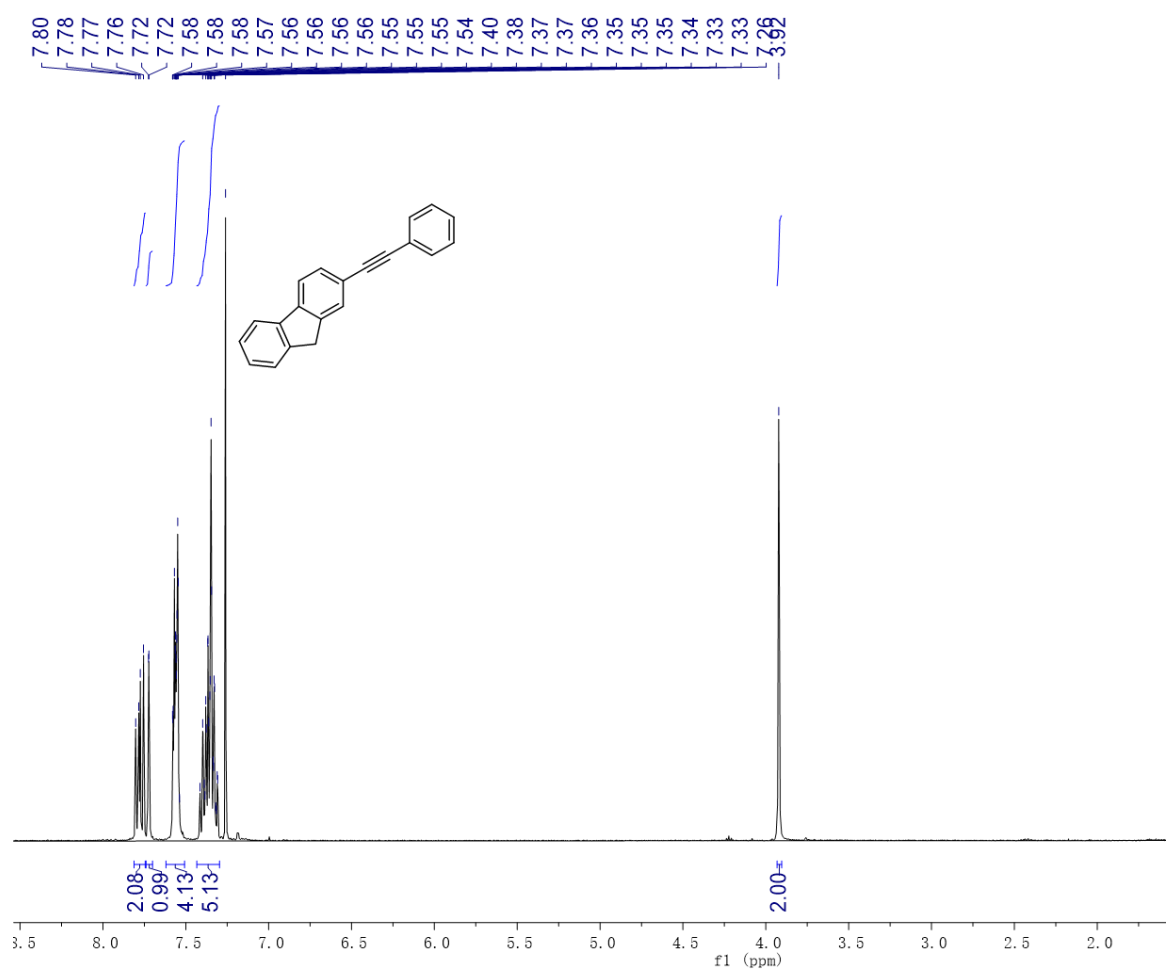
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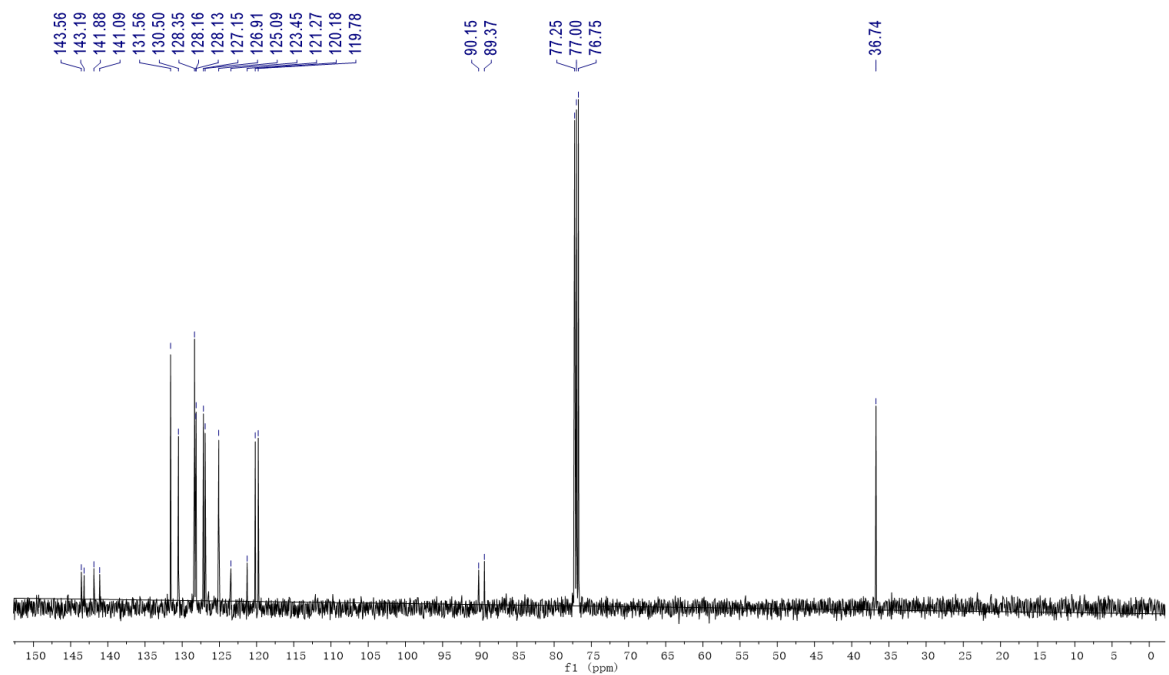
1414 ¹H and ¹³C-NMR spectra of product 9g.

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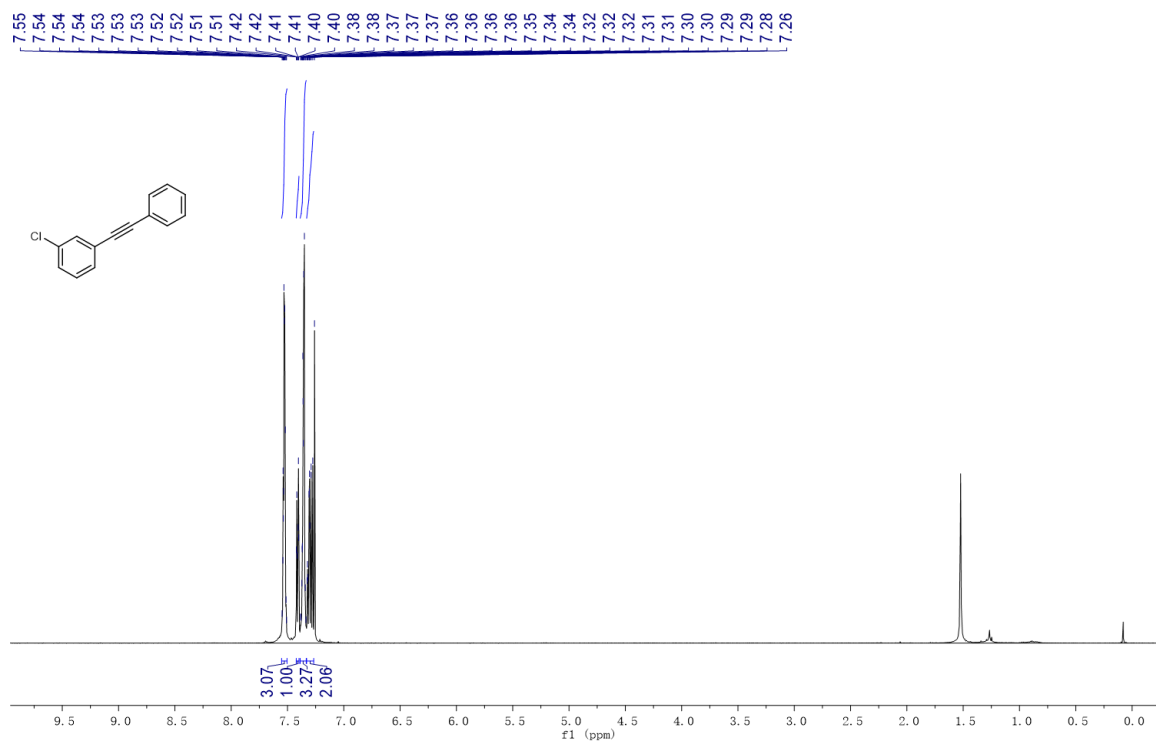
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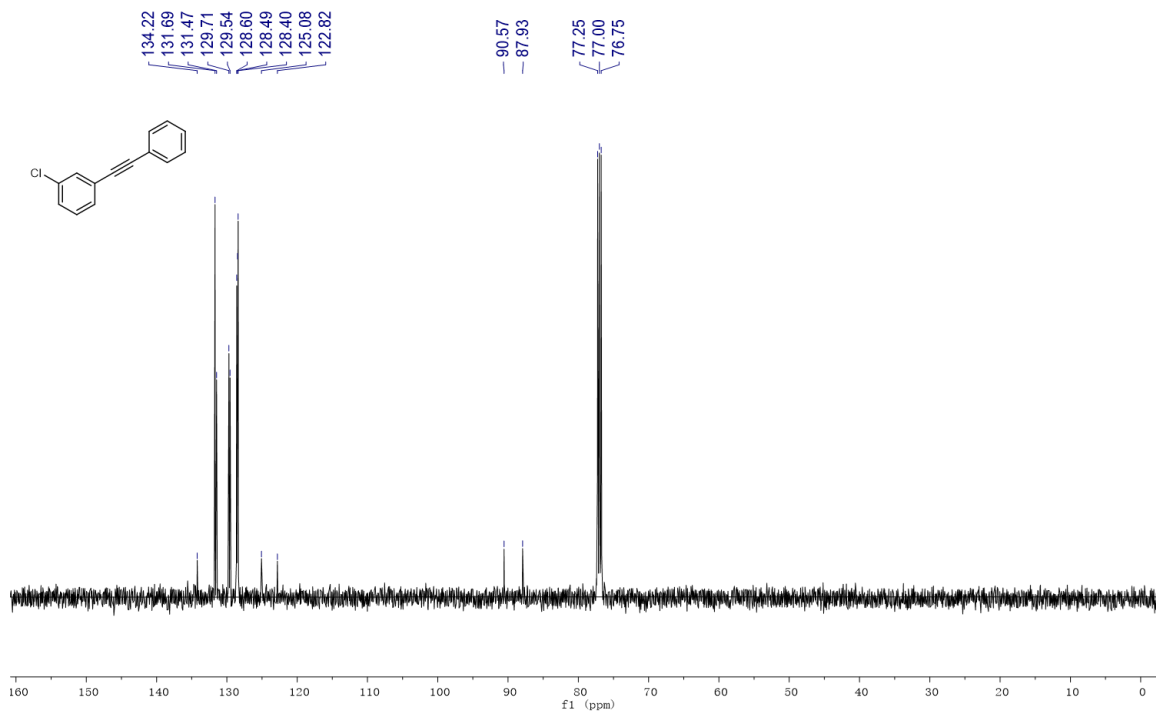
1419 **¹H and ¹³C-NMR spectra of product 9h.**

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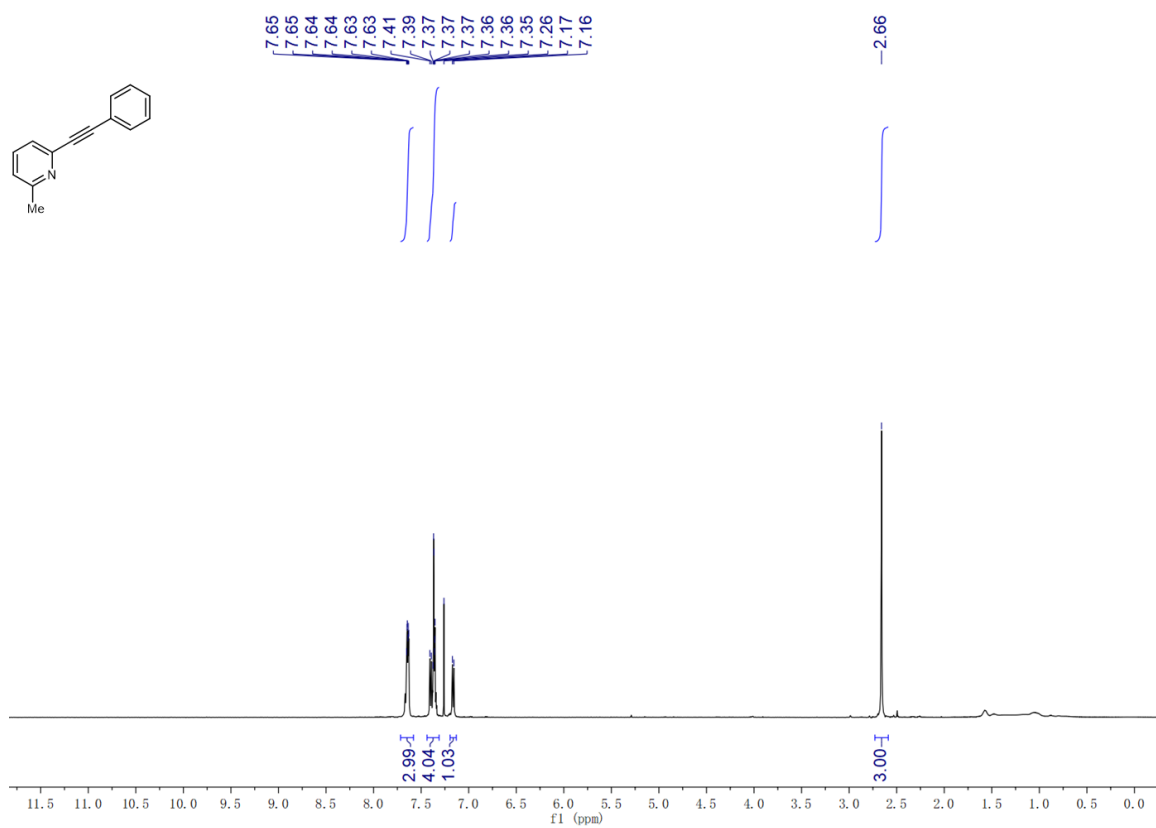


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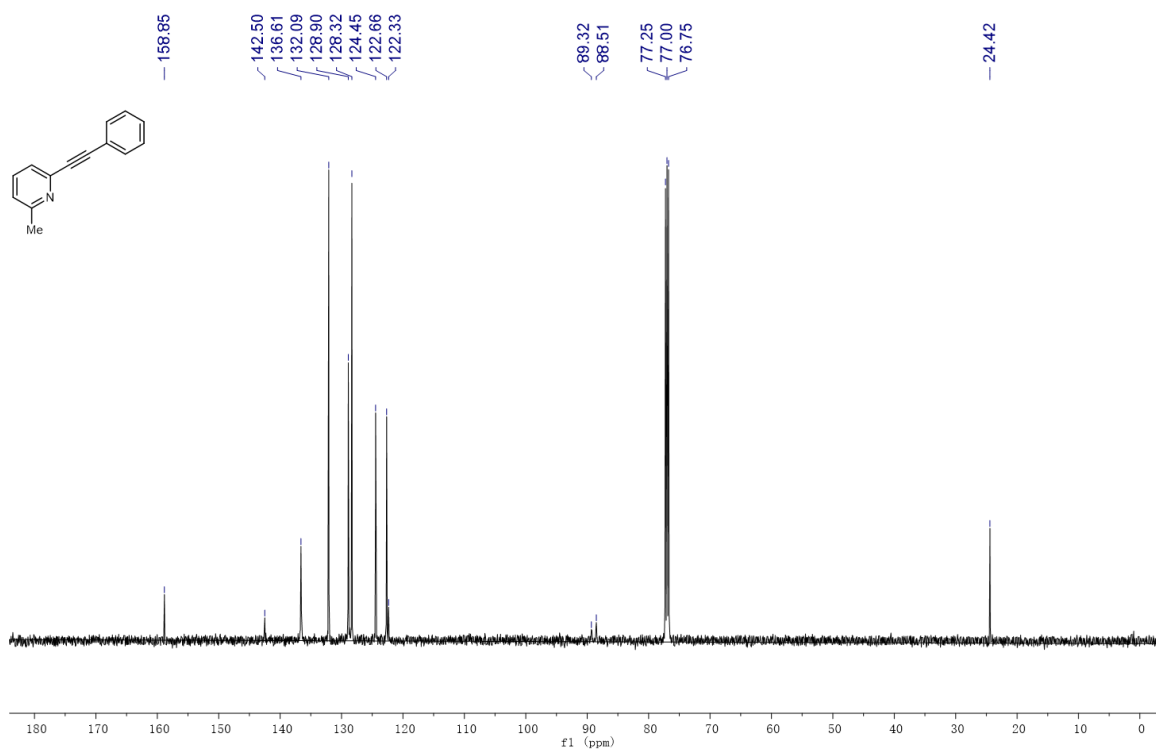
1425 ¹H, ¹³C-NMR spectra of product 9i.

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