Supplementary Information for

2	Combining same-target drugs exhibits additive efficacy with minimal toxicity
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46 1.1 Supplementary evidence for the additive effects of DNA TOPO II inhibitors

1.1.1 Animal experiments

1.1.1.1 Survival details in the 8-drug ascites tumour study

Mice began to die from day 6 after inoculation, and all mice died in three groups (Model, epirubicin, and pirarubicin) by day 10. The onset of death occurred on day 13 in the combination-drug group (Cdg), whereas the onset of death occurred on days 6–8 in the other groups. No deaths occurred in the Cdg by day 12; however, in the single-drug groups (Sdgs) only one mouse survived in the aclarubicin group by day 12. Between days 13–19, all mice in the Cdg died, whereas all 90 mice died between days 6–14 in the Sdgs and the Model group (Extended Data Table 3).

1.1.1.2 Tumour inhibition rate in 4-drug solid tumour study

Extended Data Fig. 2 shows tumour mass index, tumour inhibition rate (TIR), and tumour volume results. The TIR of the 4-drug-Cdg was 59%, which was higher than the TIRs for the doxorubicin (11.5%), etoposide (19.5%), idarubicin (10.6%), and amsacrine (7.7%) groups (all P < 0.01) and tended to be higher than the High-dose doxorubicin [Dox (H)] group (49.2%, P > 0.05). Based on the efficacy criteria, which require a TIR greater than 50%⁶⁷ or 58%⁶⁵, treatments in the single-drug groups were ineffective. The dose of each drug in this experiment was used at 1/10 of its 50% lethal dose (LD₅₀) in both the Sdgs and Cdg, except the Dox (H) group, which had a dose of 1/2 of the LD₅₀ (Extended Data Table 4).

1.1.2 Cell experiments

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1.1.2.1 Additive effects of four inhibitors on HepG2 cells

I. Four inhibitors in different combinations

i. Four inhibitors with similar IC₅₀ values

HepG2 cells were used in these experiments. Four inhibitors, including doxorubicin, daunorubicin, idarubicin, and amsacrine, at concentrations (Supplementary Table 8) corresponding to their IC_{18} were used in the single-drug treatments. The inhibition rates (16.0%, 18.9%, 18.9%, and 17.2%) in the single-drug treatments were close to the expected inhibition rate of 18% (Fig. 4d). The inhibition rate (18.9%) of the four inhibitors combined at 4.5% inhibitor concentration (IC_{4.5}) was closed to the individual inhibitors at IC₁₈, indicating the four inhibitors had an additive effect. The inhibition rate of the combination increased (Fig. 4d) as the concentrations (Supplementary Table 8) of the inhibitors in the combination increased to IC₉, IC₁₈, and IC₃₆. The combination of the four inhibitors at IC_{4.5} achieved an inhibition rate of 18.9%, which is close to the expected inhibition rate of 18% if the inhibitors induced an additive effect, which was the sum of the individual 4.5% rates. Combining the four inhibitors at IC₉ achieved an inhibition rate of 32.9%, which was close to the sum (36%) of the four inhibitors at 9%. These two experiments indicated that the four inhibitors in the combination produced an additive effect, and each of the four inhibitors contributed to the additive effect. However, the combination of the four inhibitors at IC₁₈ had an inhibition rate of only 55.6% instead of the expected inhibition rate (72%, 4 × 18%) if the 4 inhibitors elicited an additive effect. The combination of the four inhibitors at IC₃₆ had an inhibition rate of 83% instead of the expected inhibition rate of 144% (which is impossible to reach) if the inhibitors elicited an additive effect. These results indicated

that the four inhibitors exhibit additive inhibitory effects on the proliferation of HepG2 cells, but the additive effects became saturated at higher concentrations. The differences between the single-drug treatment at IC_{18} and the combination treatments of the four inhibitors combined at IC_{18} were significant (P < 0.05), indicating that combination treatment significantly improved the inhibitory activity compared with the single-drug treatments (Fig. 4d).

ii. Four inhibitors with large differences in IC50 values

The effects of doxorubicin, etoposide, daunorubicin, and podophyllotoxin with large differences in IC₅₀ values, which represent large differences in their chemical structures (Supplementary Fig. 1), on HepG2 cell proliferation were examined. As shown in Fig. 4e, the inhibition rates (18.3% and 18.4%) of etoposide and daunorubicin were close to the expected inhibition rate (18%). The inhibition rates (20.5%, 33.6%, 60.5%, and 87.4%) of the combination of the four inhibitors increased (Fig. 4e) as the concentrations (Supplementary Table 9) of the individual inhibitors in the combination increased (IC_{4.5}, IC₉, IC₁₈, and IC₃₆).

The results indicate that the four inhibitors with large differences in IC₅₀ values also elicited additive effects. The inhibition rate was significantly higher for the IC₁₈ combination group compared with the inhibition rates for etoposide and daunorubicin used separately at IC₁₈ (P < 0.05).

II. Proportionally reducing the concentration of the four inhibitors in the combinations

The concentrations of inhibitors (doxorubicin, etoposide, daunorubicin, and podophyllotoxin) in the 4-in-1 combinations were proportionally reduced from the IC₃₆ concentrations (Supplementary Table 10). In HepG2 cells, the inhibition rates (45.6% and 43.3%) of the single inhibitor group (doxorubicin and etoposide at IC₃₆) were close to the expected inhibition rates (36%). As the inhibitor combination at IC₃₆ was diluted (IC₃₆ × 1, IC₃₆ × 0.75,

IC₃₆ × 0.5, IC₃₆ × 0.25, and IC₃₆ × 0.125), the inhibition rates (93.7%, 73.6%, 64.2%, 46.1%, and 37.5%; Fig. 4g) decreased proportionally. The difference between the combination group (IC₃₆) and the doxorubicin or etoposide group at IC₃₆ was significant (P < 0.05).

1.1.2.2 The additive effects of the six-inhibitor combination on HepG2 cells

The additive effects of combining doxorubicin, etoposide, daunorubicin, idarubicin, amsacrine, and ellipticine at IC_{18} concentrations (Supplementary Table 5) on HepG2 cell proliferation were assessed. The inhibition rates (16.3%–23.6%) of single-drug treatments were close to the expected inhibition rates (18%), while the inhibition rate of the combination treatment was 87.4% (Fig. 4b). Theoretically, combining 6 inhibitors at IC_{18} should result in a theoretical inhibition rate of 108% (6 × 18% = 108%). Based on the actual inhibition rates of the six inhibitors in the combination, the calculated inhibition rate should be 123.2% (22.6% + 19.0% + 23.6% + 21.2% + 16.3% + 20.5% = 123.2%). Although both the theoretical (108%) and calculated (123.2%) inhibition rates are impossible to reach, the inhibition rate of the 6-in-1 combination reached 87.4%, demonstrating that combining the six inhibitors has an additive effect but the additive effect is restricted and becomes saturated at higher concentrations. The inhibition rates for the single-drug treatments at IC_{18} and the combination 6-in-1 inhibitor at IC_{18} for each inhibitor were significantly different (P < 0.05), indicating the inhibitory activity of the combination treatment was significantly better than the single-drug treatments.

1.1.2.3 Antitumour activity of combinations with different numbers of inhibitors at the same inhibition rates in HepG2 cells

The inhibition rates of the six-drug combinations were significantly higher than the inhibition rates of the four-drug combinations (Fig. 4f and Supplementary Table 11) when combined at the same inhibition rates. Thus, the additive inhibitory effects of the combination treatments are

related to both the number and concentration of inhibitors in the combination. The combination of four inhibitors exhibited an additive inhibitory effect on HepG2 cell proliferation, and the inhibition rates (63.2% at IC₉ or 87.4% at IC₁₈) of the six-drug combinations (doxorubicin, etoposide, daunorubicin, idarubicin, amsacrine, and ellipticine) were higher than the inhibition rates for the corresponding four-drug combinations (33.6% and 32.9% at IC₉ and 60.5% and 55.6% at IC₁₈) (Fig. 4f and Supplementary Table 11). Thus, the inhibition rate increased as the number of inhibitors in the combination or the concentration of the inhibitors increased (Fig. 4f).

1.1.2.4 The additive effects of six inhibitors on U87, HL-60, BxPC-3, and Hela cells

The effects of combinations of doxorubicin, etoposide, daunorubicin, idarubicin, amsacrine, and ellipticine at IC_{4.5}, IC₉, IC₁₈, and IC₃₆ concentrations (Supplementary Table 6) were assessed in U87, HL-60, BxPC-3, and Hela cells. The methods were similar to the experiments in HepG2 cells. As shown in Fig. 4c, the additive inhibitory effects of combination treatments on U87, HL-60, BxPC-3, and Hela cells were similar to the effects on HepG2 cells. Combining the six inhibitors significantly increased the inhibitory effects on cell proliferation compared with the single-drug treatments at the same concentrations in all cell lines. The combination (Comb IC_{4.5}) of six inhibitors at IC_{4.5} achieved inhibition rates of 28.3%, 27.6%, 30.1%, and 25.8% in U87, HL-60, BxPC-3, and Hela cells, respectively (Fig. 4c); the inhibition rates were similar to the expected inhibition rate of 27% (6 × 4.5%). Thus, the six inhibitors in the combination produced an additive effect, with each of the six inhibitors contributing to the additive effect. However, the combination (Comb IC₉) of six inhibitors at IC₉ achieved an inhibition rate of 36.4%–46.5% (Fig. 4c), which was slightly lower than the expected rate of 54% (6 × 9%). The combination (Comb IC₁₈) of six inhibitors at IC₁₈ exhibited an inhibition rate of only 47.5%–76.8% (Fig. 4c), which did not reach the expected inhibition rate of 108% (6 × 18%), which is impossible to reach. The

combination (Comb IC₃₆) of six inhibitors at IC₃₆ had an inhibition rate of 59.2%–94.2% (Fig. 4c) instead of the expected rate of 216% (also impossible to achieve). These results were similar to the four-drug combinations, indicating that the six-drug combinations exhibited additive inhibitory effects on cancer cell proliferation in multiple cancer cell lines. However, the additive effects become saturated at higher concentrations.

1.1.2.5 Dose-response curves and calculation of the combination index (CI)

SRB assays were used to evaluate cell proliferation. The inhibitory effects of doxorubicin, etoposide, daunorubicin, idarubicin, amsacrine, and ellipticine on HepG2, U87, HL-60, BxPC-3, and Hela cell proliferation were determined, and the dose-response curves were plotted (Supplementary Fig. 2). The corresponding IC₅₀ values (Supplementary Table 7) were obtained for the subsequent determination of dosing concentrations. For example, the concentrations of each of the six inhibitors corresponding to their 4.5% inhibitory rates against HepG2, U87, HL-60, BxPC-3, and Hela cells were obtained from the dose-response curves and these concentrations were used for the six inhibitors in combination 1 (Comb IC_{4.5}; Fig. 4c and Supplementary Table 6). Similarly, the concentrations corresponding to the inhibitory rates at 9%, 18%, and 36% were obtained from the dose-response curves and used in combinations 2, 3, and 4 (Comb IC₉, Comb IC₁₈, and Comb IC₃₆; Fig. 4 b,c, and Supplementary Tables 5 and 6). All combination groups 1–4 were also tested in U87 cells; the inhibition rates were 28.3%, 36.4%, 47.5%, and 63.2% and the CI values were calculated (Supplementary Table 17).

The results from the experiments on the cell lines showed that the combined use of the six

The results from the experiments on the cell lines showed that the combined use of the six TOPO II inhibitors at low concentrations exhibited additive inhibitory effects in tumour cell lines. The CI values of these combinations were all between 0.92 and 1.01 (Supplementary Table 17), suggesting that the combined effects of the 6 TOPO II inhibitors were mainly additive. This

result was consistent with a previous study showing that two-drug combinations of amsacrine with the TOPO II inhibitors doxorubicin, etoposide, or daunorubicin exhibited additive effects while combining amsacrine with the TOPO II inhibitor mitoxantrone exhibited a synergistic effect in human T-cell leukemia cells⁶⁸. These results indicated that the interactions of most drugs targeting TOPO II are additive.

1.1.3 Enzyme experiments

1.1.3.1 TOPO II-mediated kDNA decatenation assay

In a 1% agarose gel, kDNA mostly remains in the loading well. TOPO II disrupts the interlinks in kDNA and liberates small DNA minicircles, which can migrate into the agarose gel (Fig. 4h and Supplementary Tables 12 and 13). Lane D of Fig. 4h contained the kDNA control, which did not migrate. Lane T contained a mixture of kDNA and TOPO II; only a small amount of kDNA remained in the loading well, indicating that the majority of the kDNA was enzymatically digested by TOPO II to generate DNA minicircles after decatenation. Thus, TOPO II can enzymatically degrade kDNA in the absence of any inhibitor. Lanes 1–3 were loaded with mixtures of kDNA, TOPO II, and doxorubicin at low, medium, and high doses (Supplementary Table 12). As the inhibitor concentration increased, the amount of migrating DNA minicircles decreased. Analysis of the electrophoretic pattern showed that the low, medium, and high doses of doxorubicin inhibited TOPO II activity by 9.1%, 15.6%, and 88.7%, respectively (Supplementary Table 13). Lanes 6–8 and 9–11 contained kDNA treated with TOPO II and low, medium, and high doses of daunorubicin and idarubicin, respectively. The inhibitory effects of the two inhibitors on TOPO II activity were also dose-dependent, with inhibition rates of 0.8%, 11.9%, 87.1%, and 7.1%, 11.5%, 86.1% for daunorubicin and idarubicin, respectively. The high

dose of each inhibitor was considered the positive control because they almost completely inhibited TOPO II. Lanes 4–5, 12–13, and 14–15 correspond to 2 doses (low and medium doses) of etoposide, amsacrine, and ellipticine, respectively. The inhibition rates for etoposide, amsacrine, and ellipticine were 8.0% and 16.6%, 8.1% and 19.5%, and 11.6% and 22.2%, respectively. Lanes 16–17 (Fig. 4h) showed the effects of the 6-in-1 combinations of doxorubicin, etoposide, daunorubicin, idarubicin, amsacrine, and ellipticine at low and medium doses, respectively. The low-dose combination included the individual inhibitors at their low doses, and the medium-dose combination included the individual inhibitors at their medium doses (Supplementary Table 12). The corresponding inhibition rates were 17.8% and 84.2%, respectively (Supplementary Table 13). Thus, the inhibitory effects of the combinations on TOPO II activity were dose-dependent. The medium dose was more effective than the single inhibitor treatments. The inhibitory effects of the six individual TOPO II inhibitors were much lower than the inhibitory effects of the combined inhibitors at the corresponding dose. These results confirm the additive effect of combining multiple drugs targeting TOPO II.

1.1.3.2 TOPO II-mediated DNA uncoiling assay

In the TOPO II-mediated DNA uncoiling assay, supercoiled DNA (SC) and relaxed DNA (RLX) are distinguished by their different gel migration rates (Supplementary Tables 12 and 14, and Fig. 4i). In Fig. 4i, Lane D shows the pBR322 control, which exhibits the SC band properties of the negatively supercoiled structure. Lane T shows pBR322 treated with TOPO II; the SC band is not detectable, indicating that the majority of the DNA was uncoiled and transformed into RLX by TOPO II. Lane 1 was loaded with a mixture of pBR322, TOPO II, and medium-dose etoposide. The absence of the SC signal and the presence of the RLX signal in the lane indicates that a majority of the DNA was uncoiled by TOPO II. Thus, the medium-dose

etoposide did not completely inhibit TOPO II (inhibition rate 11.6%; Supplementary Table 14). Lane 2 shows a mixture of pBR322, TOPO II, and the combination consisting of the six TOPO II inhibitors (doxorubicin, etoposide, daunorubicin, idarubicin, amsacrine, and ellipticine) at medium doses (Supplementary Table 12). Only the SC band appeared in this lane, and no RLX band was visible (Fig. 4i). This result indicates that the combination drug significantly inhibited TOPO II-mediated uncoiling of the DNA, with an inhibition rate of 89.2% (Supplementary Table 14). Lane 3 was loaded with a mixture of pBR322, TOPO II, and high-dose etoposide. Only the SC band was detected (Fig. 4i), indicating that high-dose etoposide inhibited the ability of TOPO II to uncoil the supercoiled structure; the corresponding inhibition rate was 81.7% (Supplementary Table 14). These results indicate that etoposide did not exhibit a significant inhibitory effect at the medium dose, but the combination of six TOPO II inhibitors at medium doses significantly inhibited TOPO II. Thus, the combination suppressed TOPO II better than the single-drug treatments, confirming the additive effect on the same target.

1.2 Supplementary evidence for the reduced toxicity of the 8-drug combination in the 8-

drug solid tumour study

1.2.1 Hepatotoxicity

The liver index of the Model group significantly increased by 28.6% compared with the Blank group (P < 0.01), indicating that the liver was slightly swollen after tumour inoculation (Fig. 3g). The liver index decreased by 12.3% (P < 0.05) in the aclarubicin group and 13.2% (P < 0.05) in the teniposide group compared with the Model group, indicating that the 2 single-drug groups (Sdgs) reversed the liver swelling to some extent. The liver indices in the other groups were not different from the Model group.

Serum albumin (ALB) levels in the Model group tended to be lower than ALB levels in the Blank group. None of the drug groups induced abnormalities in ALB compared with the Model group. The combination-drug group (8-drug-Cdg) tended to reverse the reduction in ALB. Compared with the Cdg (Fig. 3f), ALB was significantly lower in the doxorubicin, amsacrine, teniposide, and epirubicin groups, indicating that the ability of the liver to synthesize proteins was impaired in these 4 Sdgs compared with the Cdg, whereas the Cdg did not impair protein synthesis.

1.2.2 Cardiotoxicity

Cardiac indices (Fig. 3j) were not significantly different when any two of the following groups were compared: the combination-drug, Blank, Model, and single-drug groups. Compared with the Cdg, Serum lactate dehydrogenase (LDH) and creatine kinase (CK) levels were significantly increased in the doxorubicin and idarubicin groups (both P < 0.01 and both P < 0.05, respectively) (Fig. 3h,i). Differences in LDH (5.3; Table 1) and CK (1.9; Table 1) levels in the Cdg compared with the Model group were lower than differences (48.5 and 25.5; Table 1) between the doxorubicin group and the Model group (89.1% and 92.5%, respectively) [Note: (48.5 - 5.3)/48.5 = 89.1%; (25.5 - 1.9)/25.5 = 92.5%]. Differences in LDH and CK levels in the Cdg compared with the Model group were lower than the differences (115.5 and 29.7; Table 1) between the idarubicin group and the Model group (95.4% and 93.6%, respectively). Thus, doxorubicin and idarubicin caused severe myocardial injury and the Cdg reduced myocardial damage by about 90%.

1.2.3 Physical condition

Mice developed visible tumours in their right axilla after tumour cell inoculation. The physical status of mice in the Blank and Model groups was relatively good. Mice in the

combination-drug, Model, and Blank groups were all relatively active and healthy and had sleek shiny coats, with no significant differences between the three groups. However, mice in the Sdgs had dull and ruffled coats and gradually worsening vitality. The mice became increasingly less aggressive when handled. These mice had increased piloerection and consumed less food, and 10%-50% of mice in these groups died during the treatment period (Fig. 3a and Extended Data Table 5). These results indicate that the toxic effects in the Sdgs were more severe than the toxic effects in the Cdg.

1.2.4 Growth status and adverse gastrointestinal effects

Growth status and adverse gastrointestinal effects, which are common toxic effects of anticancer drugs, were assessed using body weight, daily food intake, and daily water intake. On the last day of the experiment, the Cdg exhibited overall good physical condition but body weight (Fig. 3n) declined by 22.2% (P < 0.01) compared with the Model group (Table 1). Body weights (Fig. 3n) in the Sdgs declined, especially in the idarubicin group, compared with the Model group (7.8%–20.3%, P < 0.01), except the amsacrine group (Table 1). A dose was considered toxic if either the recipient died or their body weight dropped by more than 15% from their initial weight^{64,110}. All eight drugs in the Sdgs are toxic, based on the mortality criterion, and the Cdg and four Sdgs (body weight loss: 17.3%, 20.3%, 17.0%, and 17.0% for the Eto, Ida, Acl, and Epi groups, respectively; Table 1) in the present study are toxic, based on the body weight criterion. The average daily food intake per mouse in the Cdg declined by 30.0% compared with the Model group (P < 0.05), suggesting that the body weight loss in the Cdg was due to reduced food intake (Fig. 3o). Mice in the epirubicin and idarubicin groups had the lowest average daily food intake per mouse on the last day of the experiment (35.0%, P < 0.01 and

32.5%, P < 0.05, respectively) compared with the Model group (Table 1). Thus, the epirubicin and idarubicin groups suffered more adverse gastrointestinal effects than the Cdg.

No significant differences in average daily water intake per mouse were detected between the groups (Fig. 3p). The epirubicin and idarubicin groups and the Cdg had the lowest water intake on Day 9 (Fig. 3p and Table 1).

Overall, the data for body weight, daily food intake, and daily water intake suggest that the idarubicin and epirubicin groups and the Cdg experienced moderate adverse effects. As the Cdg demonstrated similar adverse effects to idarubicin and epirubicin groups, the adverse effects of the Cdg may have been predominantly caused by idarubicin and epirubicin in the drug combination. The decrease in body weights in the Cdg was slightly worse than the weight changes in the Sdgs. Daily food intake decreased less in the Cdg compared with the epirubicin and idarubicin groups but more than the other Sdgs.

1.2.5 Myelosuppression

Myelosuppression is a common toxic effect of anticancer drugs. All of the drugs in this study cause varying degrees of myelosuppression (Supplementary Tables 1 and 2). Therefore, a routine blood test was used to evaluate myelosuppression; declines in white cell and platelet counts indicated myelosuppression. No significant differences in white cell and platelet counts were detected between the Model and Blank groups (Fig. 3q,r), indicating that no myelosuppression occurred after tumour inoculation. Mice in the Cdg suffered slight myelosuppression, and more severe myelosuppression was detected in the idarubicin and amsacrine groups compared with the Cdg (Fig. 3q,r, and Table 1). The doxorubicin, pirarubicin, etoposide, and epirubicin groups tended to develop myelosuppression and suffered from drug

toxicity comparable to the Cdg. No significant changes in white cell or platelet counts were detected in the aclarubicin and teniposide groups, indicating less toxicity than the Cdg.

1.2.6 Immune system damage

Immune system damage may occur in the mice in response to the drug intervention (Supplementary Table 2). Thus, damage to the immune system was evaluated using thymus and spleen indices. The thymus (Fig. 3s) and spleen (Fig. 3t) indices increased by 17.6% (P > 0.05) and 93.6% (P < 0.01), respectively, in the Model group compared with the Blank group, indicating that immunity was activated by tumour inoculation. Immune system damage was detected in both the Cdg and Sdgs, but less damage was detected in the Cdg (Fig. 3s,t, and Table 1). Larger changes in the thymus index were detected in seven Sdgs and larger changes in the spleen index were detected in six Sdgs compared with changes in the thymus and spleen indices in the Cdg (Table 1).

1.3 Toxicities of four drugs alone or in combination in solid tumour mouse model

1.3.1 Survival rate (Extended Data Figs. 1c and 3a)

The dose of each drug (doxorubicin, etoposide, idarubicin, and amsacrine; Extended Data Table 4) in both the single-drug groups (Sdgs) and the combination-drug group (4-drug-Cdg) was set at 1/10 of the LD₅₀ for each drug, and each drug was administered in six doses in the 4-drug solid tumour study. The high-dose doxorubicin [Dox (H)] group had a dose (Extended Data Table 4) of 1/2 of the LD₅₀ and was administered in only four doses. Administration was halted after the fourth dose due to severe toxicity, resulting in the death of six mice in the Dox (H) group. Over the 12-day administration period (Extended Data Fig. 1c), no mice died in the Blank, Model, doxorubicin, etoposide, or idarubicin groups, and no differences in survival rates were observed. However, in the other drug groups, 1 to 6 deaths per group occurred starting at day 8

after tumour inoculation. The survival rates were 90% (mortality rate 10%) in the 4-drug-Cdg, 80% (mortality rate 20%) in the amsacrine group, and 40% (mortality rate 60%) in the Dox (H) group (Extended Data Fig. 3a).

1.3.2 Hepatotoxicity (Extended Data Fig. 3b–g)

The liver index increased by 37.7% in the Model group (P < 0.01) compared with the Blank group, indicating mild liver swelling after tumour inoculation. The liver index decreased by 22.3% in the 4-drug-Cdg and increased by 13.2% in the amsacrine group (both P < 0.01) compared with the Model group, indicating abnormal liver function in both groups. No differences in liver index were observed between the other Sdgs and the Model group.

ALT and AST levels increased by 42.4% (P < 0.05) and 187.5% (P < 0.01) and ALB levels decreased by 18.4% (P < 0.05) in the Model group compared with the Blank group, indicating that tumour inoculation induced liver inflammation leading to liver swelling. ALT, AST, and ALP increased by 61.6%, 116.9%, and 49.2% (all P < 0.01), respectively, and TP and ALB decreased by 14.3% and 22.9% (both P < 0.05), respectively, in the Dox (H) group compared with the Model group. No significant differences in the five liver indicators were detected between the doxorubicin, etoposide, idarubicin, and amsacrine groups compared with the Model group. TP decreased by 12.3% in the 4-drug-Cdg compared with the Model group (P < 0.05); no significant differences in the other four indicators were detected between the 4-drug-Cdg and the Model group. ALT, AST, and ALP levels decreased by 34.7%, 49.1%, and 37.2% (all P < 0.01), respectively, while ALB levels increased by 19.1% (P < 0.05) in the 4-drug-Cdg compared with the Dox (H) group. These results suggest much lower liver damage in the 4-drug-Cdg compared with the Dox (H) group. In summary, the Dox (H) group exhibited the most severe damage, the 4-drug-Cdg showed mild liver injury, and none of the other Sdgs exhibited liver damage.

1.3.3 Cardiotoxicity (Extended Data Fig. 3h, i)

No significant differences in the cardiac indices were detected in pairwise comparisons between the 4-drug-Cdg and the Model group and other Sdgs (data not shown). CK increased by 24.2% (P < 0.01) and LDH increased by 15.0% (P > 0.05) in the 4-drug-Cdg compared with the Model group. LDH increased by 83.1% and CK increased by 79.1% in the Dox (H) group compared with the Model group (both P < 0.01), and the increases were significantly higher than the 4-drug-Cdg (both P < 0.01). CK levels were significantly lower in the doxorubicin, etoposide, idarubicin, and amsacrine groups compared with the 4-drug-Cdg (all P < 0.05). In summary, the Dox (H) group exhibited severe myocardial damage, and the 4-drug-Cdg also showed myocardial damage. However, none of the other Sdgs caused myocardial damage.

1.3.4 Nephrotoxicity (Extended Data Fig. 3j–l)

The kidney index, indicating renal impairment, increased by 11.7% in the Dox (H) group compared with the Model group (P < 0.05). No significant differences in kidney index were detected between the other drug groups and the Model group. No significant differences in urea levels were detected between the drug groups and the Model group. The SCr levels, indicating kidney damage, increased by 25.8% in the Dox (H) group compared with the Model group (P < 0.01). No significant differences in SCr levels were detected between the other drug groups and the Model group. In summary, the Dox (H) group exhibited some kidney damage, while none of the other drug groups showed evidence of kidney damage.

1.3.5 Immune system damage (Extended Data Fig. 3m,n)

The thymus index increased by 32.1% (P > 0.05) and the spleen index increased by 88.9% (P < 0.01) in the Model group compared with the Blank group, indicating immune system stimulation and enhanced immunity in response to tumour inoculation. The thymus index

decreased significantly in all drug groups compared with the Model group (P < 0.05 for the amsacrine and Dox (H) groups; P < 0.01 for all others). The thymus index decreased by 48.6%, 24.3%, 51.4%, 29.7%, 24.3%, and 54.1% in the 4-drug-Cdg, doxorubicin, etoposide, idarubicin, amsacrine, and Dox (H) groups, respectively. Overall, the Dox (H) and etoposide groups

exhibited greater thymus indices compared with the 4-drug-Cdg.

Compared with the Model group, the spleen index decreased by 49.0% and 44.1% in the 4-drug-Cdg and Dox (H) group, respectively (both P < 0.01). The spleen index in the amsacrine, doxorubicin, etoposide, and idarubicin groups increased by 22.5% (P < 0.05), 33.3%, 14.7%, and 6.9%, respectively (all P > 0.05), compared with the Model group. In summary, all drug groups experienced some level of immune system damage.

1.3.6 Growth status and adverse gastrointestinal effects (Extended Data Fig. 30,p)

The 4-drug-Cdg exhibited a 25.6% decrease in body weight on day 14 compared with the Model group (P < 0.01). Dox (H) caused an 11.8% decrease in body weight compared with the Model group (P < 0.05). The other Sdgs (excluding the amsacrine group) caused a slight decrease in body weight, but the weight losses were not significantly different from the Model group.

The daily food intake between the 6th and 11th days decreased by 25.0% and 32.5% in the 4-drug-Cdg (3.0 g) and idarubicin group (2.7 g), respectively, compared with the Model group (4.0 g). No significant differences in food intake were detected between the other drug groups and the Model group.

1.3.7 Comparison of toxicities in the 4-drug-Cdg and the single-drug groups in the 8-drug

solid tumour study

The total dose $(1/10 \times 4 \text{ drugs} = 4/10 \text{ LD}_{50}; 4/10 \text{ LD}_{50} \times 6 \text{ injections} = 24/10 \text{ LD}_{50})$ in the 4-

drug-Cdg (doxorubicin, etoposide, idarubicin, and amsacrine combined at 1/10 LD₅₀ each; Extended Data Table 4) was larger than the doses (1/4=2.5/10 LD₅₀; 2.5/10 LD₅₀ \times 5 injections=12.5/10 LD₅₀; Extended Data Table 1) of each drug in the 4 single-drug groups (Sdgs) in the 8-drug solid tumour study. However, the mortality was 10% in the 4-drug-Cdg (Extended Data Fig.3a), which was lower than the mortality (20%–50%; Table 1) in the 4 Sdgs (doxorubicin, etoposide, idarubicin, and amsacrine;). Other indicators of toxicity in the 4-drug-Cdg were also lower than the toxicities of the Sdgs, as shown by the ALT, AST, ALP, LDH, and Scr levels in Fig. 3 for the Sdgs and Extended Data Fig. 3 for the 4-drug-Cdg (The values for the Model groups in the two figures can be used as a baseline for comparison). These data suggest that the 4-drug-Cdg did not elicit an additive effect on toxicity. The tumour inhibition rate (TIR) of the 4-drug-Cdg (59%) was significantly higher than the TIR (24.6%–38.1%) of the Sdgs in the 8drug solid tumour study. Thus, if the doses of each Sdg were increased to achieve a TIR of 59%, the toxicity of each Sdgs (doxorubicin, etoposide, idarubicin, and amsacrine) would be much greater than the toxicity of the 4-drug-Cdg. The toxicities of these anticancer drugs are doselimiting or dose-dependent^{1,9,10,12,13}. Overall, these results indicate that the 4-drug-Cdg elicited a toxicity scattering effect.

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2. Supplementary Discussion

- 2.1 Does the observed toxicity reduction of the 8-drug-Cdg in this study hold significant
- 427 <u>clinical implications? -----Comparison of the toxicities of the 8-drug-Cdg with the clinical</u>
 - toxicities of the eight anticancer drugs
 - Based on the reported clinical toxicities (Supplementary Table 1) of the eight drugs used in the 8-drug solid tumour study, the toxicity reduction in the 8-drug-Cdg is highly significant. For

example, long-term use of doxorubicin causes congestive heart failure with a high mortality rate (Supplementary Table 1), and the incidence of cardiotoxicity positively correlates with the total cumulative dose. When the cumulative dose of doxorubicin reaches 550 mg/m² (being equivalent to 14.9 mg/kg, according to the equation mg/m² = $K_m \times mg/kg$, where $K_m = 60 \text{ kg/}1.62 \text{ m}^2$, APPENDIX B of the FDA document⁶⁹ and other literature^{70–72}) or above, the incidence of congestive heart failure is $7\%^{73}$, $> 18\%^{74}$, or $26\%^{11,18,75}$. When the cumulative dose reaches 700 mg/m² (equivalent to 18.9 mg/kg), the incidence of congestive heart failure is up to 18%⁷³, > 36%⁷⁴, or 48%^{18,75}. A survey by the Paediatric Cardiomyopathy Registry of more than 100 paediatric cardiology centres in North America showed that more than 15% of all patients with cardiomyopathy were previously treated for cancer with anthracyclines (doxorubicin, idarubicin, epirubicin etc.) during childhood or adolescence⁷⁶. In addition, mortality directly related to anthracycline-induced cardiac failure is substantial, with some large series reporting rates of over 20% mortality⁷⁶. Furthermore, the use of doxorubicin at a dose of 25 mg/kg can cause renal, myocardial, and liver damage in rats after 2 days of treatment⁷⁷. If the cumulative dose (2.5) mg/kg × 5 doses = 12.5 mg/kg) of intraperitoneal injections for the doxorubicin group in the present study, which can be considered as the dose by intravenous injection due to the similar bioavailability^{78,79}, is converted to the cumulative dose of intravenous injection for adults, the dose is 37.5 (37.7) mg/m² [being equivalent to 1.02 mg/kg; according to the equations of HED $(mg/m^2) = HED (mg/kg) \times Human K_m = Animal dose (mg/kg) \times (Animal K_m/Human K_m) \times$ Human K_m = Animal dose (mg/kg) × Animal K_m , (where Human K_m = 60 kg/1.62 m²; Animal (mouse) $K_m = 0.02 \text{ kg}/0.007 \text{ m}^2$; HED, human equivalent dose), in APPENDIX B of a FDA document⁶⁹ and other literature^{70–72}]. At this dose, the toxicity including a 20% mortality rate (Table 1) of doxorubicin in mice in our experiments was similar to the toxicity observed in the

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clinical settings in which death, cardiotoxicity, myelosuppression, liver toxicity, and renal damage occurred (Supplementary Table 1). These toxicity responses occurred at the drug dose at which the efficacy in the doxorubicin group was only about 1/2 the efficacy of the 8-drug-Cdg (TIR of the doxorubicin group to the Cdg was 27.0%/54.3% = 49.7%; Fig. 2b). If the doxorubicin group dose was increased to exhibit the same efficacy as the Cdg, the toxicity caused by doxorubicin would be much more severe. We used doxorubicin alone by increasing the dose to 1/2 of its LD₅₀ (the cumulative dose = 5.0 mg/kg × 4 doses = 20.0 mg/kg), resulting in a TIR of 49.2%, but with a 60% mortality rate (see Section 1.3 for the detail). In contrast, the Cdg at this dose (the total dose of the Cdg was 3.02 times the dose used in the doxorubicin group) did not exhibit liver toxicity, cardiotoxicity, or renal toxicity, and no deaths occurred (Table 1). Thus, our drug combination exhibited good anticancer efficacy (TIR being 54.3%) with greatly reduced toxicity.

In this study, the dose of the doxorubicin group (2.5 mg/kg × 5 doses = 12.5 mg/kg in mice,

equivalent to 1.02 mg/kg in humans) was comparable to the clinical dose for doxorubicin (0.68–3.24 mg/kg; Supplementary Table 15). However, this dose did not exhibit significant tumour-inhibitory efficacy (TIR 27.0%; Fig. 2b). In contrast, the TIR (54.3%; Fig. 2b) of the 8-drug-Cdg group was twice TIR of the doxorubicin group, demonstrating effective tumour suppression with almost no detectable toxicity. Therefore, applying this combination of 8 drugs in the clinical setting, which shows a greater therapeutic window, should achieve more desirable tumour treatment effects and significantly reduce the incidence of toxic reactions. Thus, the observed toxicity reduction of the 8-drug-Cdg in this study is clinically significant.

2.2 Why and how is the 'toxicity scattering effect' elicited? Do drugs with different structures that produce distinct toxicity profiles underlie the 'toxicity scattering effect,'

resulting in significantly reduced toxicity when combining multiple drugs with the same

target? ----Relationship between the chemical structures and toxicities of drugs and the

'toxicity scattering effect'

Based on our experimental data and data reported in the literature 11,64,80-82, we found that drugs acting on the same target exhibit variations in their chemical structures (Supplementary Fig. 1), resulting in diverse toxicity profiles, including the types and intensities of toxicities (Table 1, Fig. 3, and Supplementary Tables 1 and 2). These differences suggest varying 'off-target toxicities'. We believe this phenomenon is the basis for the elicitation of the 'toxicity scattering effect' and the significant reduction in toxicity when combining multiple drugs with the same target. The following section presents specific differences in toxicities due to differences in the chemical structure of the related drugs based on our experimental data analysis.

2.2.1 The relationship between the chemical structures and toxicities of the eight drugs in

the present study

The relationship between the chemical structures and toxicity of the eight drugs used in animal experiments in the present study is summarized here based on Fig. 3, Table 1, and Supplementary Fig. 1. The probable basis for the significant reduction in toxicity due to the 'toxicity scattering effect' is described.

The eight drugs used in our animal experiments can be classified into the following three categories based on their chemical structures:

2.2.1.1 Amsacrine is an acridine with a markedly different chemical structure compared with the other seven drugs (Supplementary Fig. 1). Amsacrine did not exhibit heart, liver, or kidney toxicity in our study. However, the other seven drugs caused damage to at least one of these

organs. Amsacrine did not affect body weight, whereas the other seven drugs caused weight loss of 7.8%–20.3% (Table 1).

2.2.1.2 Etoposide and teniposide are podophyllotoxin analogues. Etoposide showed no toxicity to either the heart or liver in our study, whereas the other six drugs (excluding amsacrine) all exhibited cardiac and/or hepatic toxicity. Etoposide exhibited the highest thymus (65.0%) and spleen (40.7%) indices among the eight drugs (Table 1) and were significantly higher than the indices for amsacrine (50.0% and 29.7%, respectively). Etoposide had no significant effect on WBC, but amsacrine increased the WBC by 26.6%. Teniposide increased the liver index by 13.2% but did not affect the spleen index. Thus, teniposide exhibited a different toxicity profile than etoposide, which has no significant effect on liver index (Table 1).

2.2.1.3 Doxorubicin, idarubicin, aclarubicin, epirubicin, and pirarubicin are anthracyclines. Idarubicin had the highest mortality rate (50%), the highest increases in LDH (115.5%), CK (29.7%), ALP (25.2%), and SCr (31.1%) levels, and the greatest decreases in WBC (40.4%), platelet count (35.4%), and body weight (20.3%) among the eight drugs (Table 1). Thus, idarubicin was the most toxic among the eight drugs. The mortality rate in the doxorubicin group was 20%. Doxorubicin did not significantly affect ALP, platelet count, or food intake, but significantly increased urea levels by 31.0%, whereas idarubicin decreased food intake by 32.5% but did not significantly affect urea. Among the eight drugs, only doxorubicin significantly increased ALT (42.9%). Aclarubicin increased AST (44.5%) and urea (42.0%) more than the other seven drugs but did not significantly affect LDH, CK, or WBC. Thus, the toxicities of aclarubicin were different from the toxicities of idarubicin and doxorubicin; doxorubicin increased LDH and CK by 48.5% and 25.5%, respectively, and decreased WBC by 21.3%. Epirubicin decreased food intake the most (35.0%) and was the only drug that decreased TP

levels (14.6%). The mortality rate (10%) and the thymus index (25.0%) for pirarubicin were the lowest among the eight drugs. Pirarubicin induced the lowest body weight loss (7.8%) among the five anthracyclines. Overall, these results indicate significant differences in toxicities among the anthracyclines and the other drugs (Table 1).

2.2.2 Relationship between chemical structures and toxicities of drugs reported in the

literature

2.2.2.1 Different toxicities of doxorubicin and pirarubicin

Treatment (i.v.) with 15 mg/kg pirarubicin and doxorubicin produced 85.07% and 74.74% tumour regression, respectively. Treatment with 15 mg/kg doxorubicin caused 10.6% weight loss in mice, whereas no weight loss was observed after treatment with 15 mg/kg pirarubicin. Treatment with 25 mg/kg doxorubicin caused approximately 67% mortality and the remaining mice lost 26% of their initial body weight, whereas no deaths and less than 5% weight loss were observed in mice treated with 25 mg/kg pirarubicin. All mice died after treatment with 37.5 mg/kg doxorubicin. Mice receiving 37.5 mg/kg pirarubicin showed a > 17% decrease in body weight and no deaths⁶⁴.

2.2.2.2 Different toxicities of doxorubicin and idarubicin

Administration of 15 mg/kg (i.v.) doxorubicin induced left ventricular dysfunction in rats. Idarubicin displayed a much weaker cardiotoxicity compared with doxorubicin; left ventricular functional parameters were altered at 4.5 mg/kg of idarubicin to a significantly lower degree than 15 mg/kg doxorubicin. Treatment with 4.5 mg/kg of idarubicin caused a 26.0% decrease in body weight and treatment with 15 mg/kg doxorubicin caused a 27.8% decrease in body weight. The maximum tolerated doses were 3 mg/kg per injection for doxorubicin and 0.75 mg/kg per injection for idarubicin. The equivalent dose ratio for general toxicity was around 1:4

(idarubicin:doxorubicin), and the equivalent dose ratio for cardiotoxicity was estimated to be about 1:3 (idarubicin:doxorubicin)⁸².

2.2.2.3 Different toxicities of doxorubicin and epirubicin

The risk of congestive heart failure for a cumulative doxorubicin dose of 400 mg/m² is 5%–6%, and the risk increases to over 10% for a cumulative dose of 500 mg/m². For doxorubicin doses over 550 mg/m², approximately 25% of patients develop congestive cardiac failure. A 900 mg/m² cumulative dose of epirubicin is associated with a cumulative congestive heart failure risk of 4%, and the risk increases to 15% for a 1000 mg/m² dose. Thus, doxorubicin incurs a higher risk of congestive heart failure than epirubicin at the same dose¹¹.

2.2.2.4 Different toxicities of etoposide and teniposide

The LD_{10} values for etoposide (VP-16) and teniposide (VM-26) were 9.4 (7.4–11.8) mg/kg daily and 3.4 (2.5–4.5) mg/kg daily, respectively, in mice. The difference in LD_{10} values indicates that the toxicity of these two drugs differs by approximately threefold⁸¹.

2.2.2.5 Different toxicities of the eight drugs in animal experiments in the present study

In the present study, the LD_{50} values for intraperitoneal administration of the eight drugs in mice ranged from 3 to 29.57 mg/kg (Supplementary Table 3). This indicates a significant variation in acute toxicity among these drugs, with the largest difference being 10-fold⁸⁰.

2.2.3 Conclusion for the relationship between the chemical structures and toxicities of

drugs and the 'toxicity scattering effect'

Drugs with different chemical structures acting on the same target exhibit diverse toxicity profiles, including the types and intensities of toxicities. These differences suggest varying 'off-target toxicities'. We believe this phenomenon is the basis for the 'toxicity scattering effect', which significantly reduces toxicity when combining multiple drugs with the same target.

2.3 Why is the anticancer efficacy additive, but the toxicity is not additive? Why do low doses of the 8 drugs have additive effects on cancer cells but not on normal cells?

The simplest explanation is that the off-target effects are non-overlapping, while the ontarget effects overlap. Our enzyme experiments demonstrate that the TOPO II inhibitors exhibit additive effects on the inhibition of TOPO II at low concentrations. We posit that the fundamental principle underlying the reduction in toxicity is the significant decrease in the dose of each drug in the combination. A toxic drug is less toxic if the dose is reduced to 1/8 of the usual dose.

We attribute the additive effects of the 8 drugs to their on-target effects (action on the same efficacy target) (Extended Data Fig.4a). Their binding sites on the protein (TOPO II) should overlap, otherwise, the drugs would not produce an additive effect. The reduced toxicity of the 8-drug combination is likely due to their off-target effects. These off-target binding sites should be located at different locations on the same protein (TOPO II) or on different proteins. Otherwise, the toxicity of the combination drug would not be significantly reduced. In other words, tumour cells are sensitive to all eight drugs; therefore, they exhibit additive on-target effects, as confirmed by our additive effect experiments (Extended Data Fig.4a). However, the sensitivity of the eight drugs varies in normal cells, resulting in only partially additive off-target effects (leading to a toxicity scattering effect) when the drugs are combined (Extended Data Fig.4a).

For example, we consider a scenario where one drug affects normal heart cells, another drug affects normal liver cells, a third affects normal kidney cells, and the remaining five drugs affect normal bone marrow cells. The first three drugs affect specific cells at low concentrations (for each drug: 1/8 of the whole potency of the combination drug; Extended Data Fig.4a), these drugs

do not exert significant toxicity on these cells when combined. Thus, no observable toxicities are observed when these drugs are used together. This is a non-additive effect, resulting in a toxicity scattering effect. On the other hand, the latter five drugs all affect normal bone marrow cells. When combined, the concentration of these drugs adds up (for the five drugs: 5/8 of the whole potency; Extended Data Fig.4a), leading to additive effects on bone marrow suppression. This can be described as a partial additive effect on the same target for the toxicity, resulting in a partial toxicity scattering effect.

We do not have direct evidence for off-target effects. However, as indicated in Supplementary Tables 1 and 2, and Section 2.2.2 above-mentioned, the literature shows that each of the eight drugs has individual toxicities resulting in different toxicity profiles in both humans and animals (for our animal data see Table 1, Fig.3, Extended Data Fig. 3 and Section 2.2.1), although some toxicities may overlap among these drugs. Even when the toxicities are the same, the degree of toxicity varies among these drugs. The toxicity targets must also differ because the toxic effects of each drug differ. In other words, the toxicities of these drugs are not additive (what we call the 'toxicity scattering effect'); thus, these drugs act on different toxicity targets. We believe this is evidence for off-target effects.

2.4 Does combining more drugs with the same target lower the required dose of each drug and increase the likelihood of the 'toxicity scattering effect'?

2.4.1 Impact of the number of the same-target drugs in combination on toxicity in the present study

The 4-drug-Cdg (doxorubicin, etoposide, idarubicin, and amsacrine combined at 1/10 of their LD₅₀) induced a similar tumour inhibition rate (TIR) (59%; Extended Data Fig. 2) in the 4-drug solid tumour study as the 8-drug- Cdg (TIR 54.3%; Fig. 2) in the 8-drug solid tumour study.

However, the 4-drug-Cdg exhibited a 10% mortality rate, increased CK levels, and decreased TP levels and thymus, spleen, and liver indices (Extended Data Fig. 3 and Extended Data Table 6) compared with the 8-drug-Cdg. No mortality or hepatic, cardiac, or renal toxicities were observed in the 8-drug-Cdg (Fig.3 and Table 1). Thus, the antitumour effects were nearly equivalent, but the toxicity of the 8-drug-Cdg was significantly lower than the toxicity of the 4drug-Cdg. Furthermore, the 4-drug-Cdg exhibited much lower toxicity than doxorubicin at 1/2 of its LD₅₀. The high-dose doxorubicin [Dox (H) group] exhibited a TIR of 49.2%, but a 60% mortality rate, increased levels of ALT, AST, ALP, LDH, CK, SCr, and kidney index, and decreased ALB levels compared with the 4-drug-Cdg (Extended Data Figs. 2 and 3, and Extended Data Table 6). Notably, the Dox (H) group essentially functioned as a 'combination of only one drug'. Therefore, our results indicate that as the number of drugs in the combination increased from 1 to 4 and then to 8, the dose of each drug progressively decreased (from 1/2 LD₅₀ to 1/10 LD₅₀ and then to 1/20 LD₅₀), leading to a corresponding reduction in toxicity. Thus, the more drugs that act on the same target in a combination, the lower the dose required of each drug and the higher the likelihood of a 'toxicity scattering effect'^{38,39}. Increasing the number of drugs with the same target in a combination may progressively lower the toxicity due to the 'toxicity scattering effect'^{38,39}. The 8-drug combination dramatically reduced the mortality rate and achieved a TIR of around 50%, demonstrating a much larger therapeutic window than the single drugs and the 4-drug combination, making this combination the most appropriate anticancer therapy (Extended Data Fig. 4 b,c). 2.4.2 The reason most traditional Chinese medicines (TCMs) have very low toxicity or are non-toxic is that a certain group of active substances of a single TCMs is equivalent to a

combination of numerous constituents that collectively act on the same target.

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Our proposed hypotheses ^{38,39} regarding TCMs 'additive effects' and 'toxicity scattering effects' suggest that a certain group of active substances of a single TCMs include chemical constituents and metabolites that can act on the same target. These compounds can produce an 'additive effect' and a 'toxicity scattering effect.' Each TCMs contains numerous (at least more than ten) compounds with similar chemical structures (including constituents and metabolites) that can produce a same-target 'additive effect', leading to the 'toxicity scattering effect'. Therefore, most TCMs exhibit very low toxicity or are non-toxic. Essentially, a single TCMs is equivalent to a combination of numerous constituents that act on the same target. Thus, "the more drugs in the combination, the lower the dose required for each drug, thereby increasing the likelihood of a 'toxicity scattering effect'."

3. Supplementary Figs. 1 and 2

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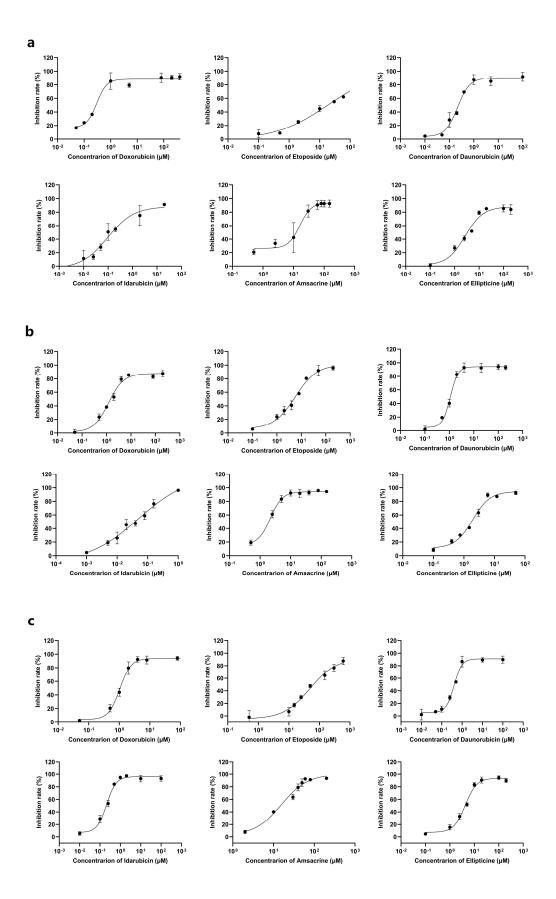
Supplementary Fig. 1. The chemical structures of the 11 anticancer drugs used in this

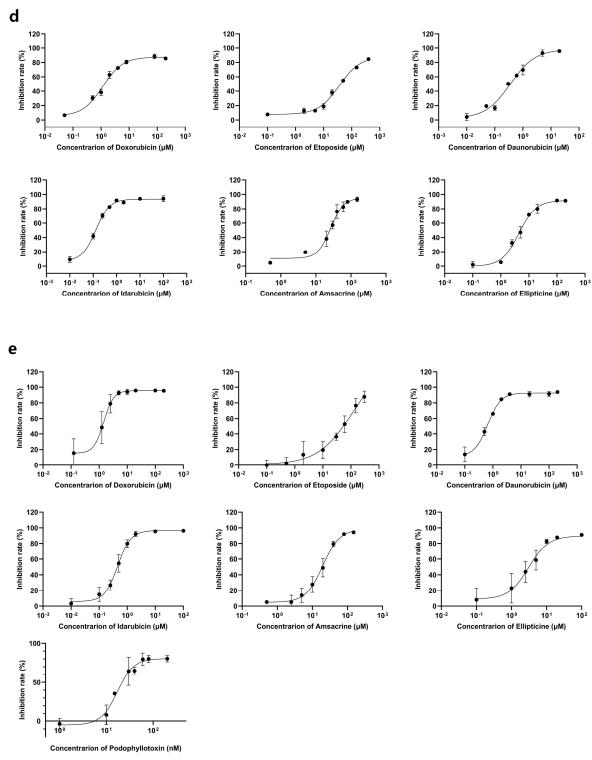
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Podophyllotoxin

(MW: 428.44)

study. All drugs are DNA topoisomerase II inhibitors and marketed anticancer drugs that are used in clinical practice (except ellipticine, which was not used in the animal experiments). The top eight drugs were used in animal experiments, and the top four drugs were also used in cell and enzyme experiments. The bottom three compounds were only used in enzyme and/or cell experiments.





Supplementary Fig. 2. Dose-response curves of seven TOPO II inhibitors in U87 (a), HL-60 (b), BxPC-3 (c), Hela (d), and HepG2 (e) cells. All five cell lines (n = 6) were treated with

- doxorubicin, etoposide, daunorubicin, idarubicin, amsacrine, and ellipticine. Only HepG2 cells were treated
- with podophyllotoxin (n = 6).

4. Supplementary Tables 1–17

Supplementary Table 1. Common toxicities of the TOPO II inhibitors according to the human organs affected

	Dox	Eto	Ida	Ams	Acl	Ten	Epi	Pir
Death	Yes ^{76,83,84}		Yes ⁸⁵		Yes ⁸⁶	Yes ⁸³	Yes ⁸³	
Liver	Hepatotoxicity ^{16,46} ; Changes in transaminase levels ⁸⁷	Hepatotoxicity ⁸⁷	ALP↑/AST↑ ⁸⁷	Hepatotoxicity ⁸⁷ ; ALP↑/AST↑ ⁸⁷	Hepatotoxicity ^{86,88}	Hepatotoxicity ion ^{83,87}	Transaminases↑87	
Heart	Cardiotoxicity ^{1,16,19,46,73,89} ; Cardiomyopathy and congestive ^{11,74,90}	Congestive heart failure ⁸⁷	Cardiotoxicity ^{85,87,90} ; Cardiomyopathy and congestive ⁷⁴ ; LDH† ⁸⁷	Cardiac arrhythmias and congestive heart failure ⁸⁷	Cardiotoxicity ^{74,86,88} ; LDH↑ ⁸⁸	Arrhythmia ^{83,87}	Cardiotoxicity ^{74,83,89,90} ; Congestive heart failure ^{11,87}	Cardiotoxicity ^{33,74}
Kidney	Nephrotoxicity ^{5,16,46}		SCr↑ ⁸⁷	Nephrotoxicity ⁸⁷ ; SCr↑/BUN↑ ⁸⁷		Nephrotoxicity ^{83,87}		
Myelosuppression	Yes ^{87,90} ; WBC↓/PC↓ ⁸³ ; WBC↓ ⁸⁷	Yes ⁹⁰	Yes ⁹⁰ ; WBC↓ ⁸⁵ ; WBC↓/PC↓ ⁸⁷	Yes ⁸⁷ ; WBC↓/PC↓ ⁸⁷	Yes ⁸⁸ ; WBC↓/PC↓ ^{86,88}	Yes ^{83,90} ; WBC↓/PC↓ ^{83,87}	Yes ^{83,90} ; WBC↓/PC↓ ^{83,87}	WBC↓ ³³
Gastrointestinal reactions	Nausea/vomiting ^{16,83,84,87,90} ; Diarrhea ^{83,84,90}	Nausea/vomiting 87,90; Diarrhea ⁹⁰	Nausea/vomiting ^{87,90} ; Diarrhea ⁸⁷	Nausea/vomiting ⁸⁷ ; Diarrhea ⁸⁷	Yes ⁸⁶ ; Nausea/vomiting ^{86,88} ; Diarrhea ⁸⁶	Nausea/vomiting ^{83,87,90} ; Diarrhea ⁸⁷	Nausea/vomiting ^{83,87} ; Diarrhea ^{83,87} ; Bleeding ^{83,87}	Yes ³³ ; Nausea/vomiting ⁸⁴ ; Diarrhea ⁸⁴

An upward arrow (†) indicates an increase in value; a down arrow (†) indicates a decrease in value. Acl, aclarubicin; ALP, alkaline phosphatase; Ams, amsacrine; AST, aspartate aminotransferase; BUN, blood urea nitrogen; Dox, doxorubicin; Epi, epirubicin; Eto, etoposide; Ida, idarubicin; LDH, lactate dehydrogenase; PC, platelet count; Pir, pirarubicin; Ten, teniposide; SCr, creatinine; WBC, white blood cell; Yes, meaning that this toxicity is present.

666 Supplementary Table 2. Drug toxicities in animal models

	Dox	Eto	Ida	Ams	Acla	Ten	Epi	Pir
Death	Yes ^{22,64,82}	Yes ^{67,81,91–93}	Yes ^{82,94}	Yes ⁹⁵	Yes ^{88,96,97}	Yes ^{81,98}	Yes ⁹⁹	Yes ^{41,100–102}
Liver	Hepatotoxicity ^{15,43} ; ALT↑/AST↑ ^{15,43,77} ; AST↑ ²² ; TP↓/ALB↓ ¹⁵	Focal necrosis ⁹¹ ; TP↓/ALP↓ ⁹¹			Hepatotoxicity ⁸⁸	Lesions ^{45,98} ; AST ⁴⁵	Lesions ^{83,99} ; ALT↑/AST↑ ¹⁰³	Lesions ^{33,101} ; ALT↑ ^{41,101,104} ; AST↑ ^{41,101,102,104} ; Liver/body weight↑ ¹⁰¹
Heart	Cardiotoxicity ^{15,43,46,64,82,105-107} ; CK ↑ ²² ; CK-MB↑ ^{15,46,77,105} ; LDH↑ ^{15,22,46,77,105} ; CK-NAC↑ ^{15,46} ; Heart/body weight↑ ¹⁰⁷		Cardiotoxicity ⁸²		Cardiotoxicity ^{96,108}	Lesions ^{45,98} ; LDH↑ ⁴⁵	Cardiotoxicity ^{83,99} ; CK-MB↑ ¹⁰³ ; Heart weight↓/ CKP↑ ⁹⁹	Cardiotoxicity ^{33,41,64,100} ; CK↑/LDH↑ ^{41,102} ; CK-MB↑ ^{41,100} ; LDH↑ ¹⁰⁰
Kidney	Nephrotoxicity ^{15,43} ; BUN↑/SCr↑ ^{15,77}				Congestion ¹⁰⁸		Lesions ⁸³	Nephrotoxicity ⁴¹ ; BUN↑ /SCr↑ ⁴¹
Thymus		Atrophy ^{91,92}			Atrophy ^{88,108}			
Spleen		Atrophy ⁹¹			Atrophy ^{88,108} ; Spleen weight↓ ⁹⁷		Lesions ⁸³	Lesions ³³
Body weight	Weight loss ^{43,64,78,82}	Weight loss ^{91,92}	Weight loss ^{82,94}	Weight loss ⁹⁵	Weight loss ^{88,96,97,108}	Weight loss ⁹⁸	Weight loss ^{83,99,111}	Weight loss ^{33,64,100,101,104,112}
Myelosuppression	Yes ⁴³ ; WBC↓ ^{22,43} ; RBC↓ ⁴³ ; PC↓ ²²	Yes ^{92,93} ; WBC↓ ^{91,93} ; RBC↓ ⁹¹ ; Hypoplasia ⁹¹	WBC↓/PC↓ ⁹⁴ ; LC↓/BMC↓ ¹¹³		Yes ^{88,96,108}	WBC↓ ⁴⁵	Yes ⁸³	WBC↓ ^{33,41,102} ; PC↓ ¹⁰²
Gastrointestinal toxicity	Intestinal mucositis ^{78,106} ; Diarrhea ⁸² ; Gastric gland dilation ¹⁰⁶ ; Vomiting ⁴³	Feed intake \(\psi^91; \) Diarrhea \(\psi^1; \) Bloody stools \(\psi^2; \) Ulcerations \(\psi^93 \)	Intestinal ucositis ¹⁰⁹ ; Diarrhea ^{82,109}		Bleeding ^{88,108} ; Nausea/vomiting ⁸⁸ ; Degeneration ¹⁰⁸		Yes ⁸³ ; Diarrhea, vomiting, and lesions ⁸³	Yes ⁴¹ ; Feed intake↓ ^{100,101,104} ; Gastrointestinal tract injured ⁴¹

An upward arrow (†) indicates an increase in value; a down arrow (↓) indicates a decrease in value. Acl, aclarubicin; ALB, albumin; ALP, alkaline phosphatase; ALT, alanine aminotransferase; Ams, amsacrine; AST, aspartate aminotransferase; BMC, bone marrow cell count; BUN, blood urea nitrogen; CK, creatine kinase; CK-MB, creatine kinase isoenzyme; CK-NAC, N-acetylcysteine activated creatine kinase; CPK, creatine phosphokinase; Dox, doxorubicin; Epi, epirubicin; Eto, etoposide; Ida, idarubicin; LC, leukocyte count; LDH, lactate dehydrogenase; PC, platelet count; Pir, pirarubicin; RBC, red blood cell count; SCr, creatinine; Ten, teniposide; TP, total protein; WBC, white blood cells; Yes, meaning that this toxicity is present.

Supplementary Table 3. LD₅₀ in mice for intraperitoneal injections of the 8 drugs*

Drug	LD ₅₀ (mg/kg)
Doxorubicin	10.70†
Etoposide	64
Idarubicin	3
Amsacrine	15.47†
Aclarubicin	16.10
Teniposide	29.57
Epirubicin (hydrochloride)	10.80‡
Pirarubicin (hydrochloride)	13.90‡

*see reference⁸⁰; †The doses of doxorubicin and amsacrine were calculated using $LD_{50} = 10$ and $LD_{50} = 15$, respectively; ‡ The LD_{50} values of epirubicin hydrochloride and pirarubicin hydrochloride were used as the LD_{50} values of epirubicin and pirarubicin, respectively; no LD_{50} for epirubicin or epirubicin hydrochloride was found for mice, so the LD_{50} for rats was used.

Supplementary Table 4. Literature report of ineffective, effective, or usual doses of 8 anticancer drugs in different mice*

	Dose for i.p. dosing		Dose for i.v. dosing
	Doses* reported: 7.02 (T/C§ 126%), 14.04 (T/C 138%), 28.17 (T/C 152%), 56.25 (T/C 167%), 112.5 (T/C 229%), 225 (T/C 214%), and 450 (T/C 143%) mg/kg ⁶⁵ ; 7.5, 15, 20, and 25 mg/kg ¹⁵ ††;	Doses* in Sdg and Cdg in the present study†: 12.5 mg/kg (TIR 27.0%§) and 2.50 mg/kg Estimated minimum effective dose‡:	Doses* reported: 5 (TIR\$ ca.37%¶) ††† and 10 (TIR ca.54%) mg/kg ¹⁰⁶ ; 10 mg/kg (TIR 29%) ¹¹⁷ ;
Dox	7.8 (ILS§ 25%) and 11.7 (ILS 79%) mg/kg ¹¹⁴ ; 8 mg/kg (TIR§ ca.23%¶) ¹¹⁵ ;	14 mg/kg	10 (none died) and 20 (all mice died) mg/kg ²² ; 20 (all mice died) mg/kg ¹⁰² ;
	9 (TIR by volume ca.50%) mg/kg ⁹⁵ ; 10 mg/kg ¹¹⁶ ; 15 mg/kg ^{43,105} ; 15 and 30 mg/kg ¹⁰⁷	Percentage of dose in Cdg versus the minimum effective dose: 2.5/14=17.9%	15 (MTD; Tumour regression 74.74%), 25 (2/3 rd mice died), and 37.5 (all mice died) mg/kg ⁶⁴
Eto	Reported: 10 (cures § 13%; ILS 242%); 20 (cures 15%, 0%, and 18%; ILS 200%, 188%, and 131%)**; and 40 (cures 81%, 55%, and 27%; ILS -††, -, and 156%)** mg/kg ⁸¹ ; 11 (ILS 8%), 22 (ILS 66%), 33 (ILS 92%), 70 (ILS 105%), 80 (ILS 82%), and 90 (ILS 47%) mg/kg ⁹³ ; 20 (Sd60§ 15%; T/C 183%) and 80 (Sd60 60%; T/C -††) mg/kg ⁹² ; 52, 73, 102, 143 (none died), and 200 (all mice died) mg/kg ⁹² ; 100–400 (T/C 134%–300%), 150 (TIR 92%), and 180 (TIR 91%) mg/kg ⁶⁷	Doses in Sdg and Cdg in the present study†: 80.0 mg/kg (TIR 38.1%) and 16.0 mg/kg Estimated minimum effective dose‡: 82 mg/kg Percentage of dose in Cdg versus the minimum effective dose: 16/82=19.5%	
Ida	Reported: 1 (TIR ca. 7%¶) and 4 mg/kg (TIR ca. 65%¶) ¹¹³ ; 3, 6, and 12 mg/kg ¹¹⁸	Doses in Sdg and Cdg in the present study†: 3.75 mg/kg (TIR 34.5%) and 0.75 mg/kg Estimated minimum effective dose‡: 4.0 mg/kg Percentage of dose in Cdg versus the minimum effective dose: 0.75/4.0=18.8%	Reported: 0.5 (ILS 13%), 1 (ILS 50%), 2 (ILS 113%), 3 (ILS 144%), and 4 (MTD) mg/kg ¹¹⁰ ; 2 mg/kg ¹¹⁹
Ams	Reported: 8, 10, 15, 20, and 24 (TIR by volume ca.27%) mg/kg ^{††*95} ; 26.7 (ILS 38%, 42%, and 78%)§§ and 39.9 (ILS 38%, 42%, and 78%)§§ mg/kg ¹¹⁴ ; 14.4, 24, and 39.9 mg/kg ¹²⁰	Doses in Sdg and Cdg in the present study†: 18.75 mg/kg (TIR 31.1%) and 3.75 mg/kg Estimated minimum effective dose‡: 28 mg/kg Percentage of dose in Cdg versus the minimum effective dose: 3.75/28=13.4%	Reported: 90 (ILS 58% and 72%)§§ mg/kg ¹¹⁴
Acl	Reported: 2.5 (TIR 18%), 5 (TIR 60%), 12.5 (TIR 100%), and 25 (TIR 100%) mg/kg ⁹⁷ ; 2.5–40 mg/kg (T/C 100%) ¹²¹ ;	Doses in Sdg and Cdg in the present study†: 20.15 mg/kg (TIR 24.6%) and 4.03 mg/kg Estimated minimum effective dose‡:	Reported: 20 (ILS 125%, 200%) mg/kg ⁸⁸ ; 20, 200, and 400 (all mice died) mg/kg ⁹⁶

	<u>6 (ILS 8%)</u> and 9 (ILS -) mg/kg ⁹³ ;	23 mg/kg	
	4.8 (ILS 50%), 9 (ILS 43%), 12 (ILS 133%), 20 (ILS 168%, >170%, or 112%), 36 (ILS 113%), 45 (ILS	Percentage of dose in Cdg versus the minimum	
	36%, 81%, 108%, and 122%), and 48 (ILS 80%), 10 (TIR 91%), 11 (TIR 56%), 20 (TIR 24%, 64%, and	effective dose:	
	80%), 28.2 (TIR 47%), 30 (TIR 64%, 67%, and 56%), 36 (TIR 84%), 54 (TIR 100%), 60 (TIR 94%), and	4.03/23=17.5%	
	60 (TIR 74% and 94%) mg/kg¶ ⁸⁸		
		Doses in Sdg and Cdg in the present study†:	
	Reported:	37.0 mg/kg (TIR 29.0%) and 7.40 mg/kg	
	3.75 (cures 13%; ILS 167%); 7.5 (cures 46%, 23%, and 0%; ILS 344%, 175%, and 125%)**; and 15 (cures	Estimated minimum effective dose‡:	Day and de
en	72%, 70%, and 27%; ILS -††, -, and 181%)** mg/kg ⁸¹ ;	42 mg/kg	Reported:
	<u>30 (TIR 48%)</u> and 100 (all mice died) mg/kg ⁹⁸ ;	Percentage of dose in Cdg versus the minimum	20 , 40 , and 60 mg/kg ⁴⁵
	ED $_{50} = 40.46 \text{ mg/kg}^{98}$	effective dose:	
		7.4/42=17.6%	
		Doses in Sdg and Cdg in the present study†:	
ъ	Demonstrate	13.5 mg/kg (TIR 28.0%) and 2.70 mg/kg	
	Reported:	Estimated minimum effective dose‡:	Reported:
Epi	10 and 20 mg/kg ⁹⁹ ;	35 mg/kg	20.5 mg/kg (ca. 0.5 times human dose) ⁸³ ;
	12.5 mg/kg (TIR ca. 25%¶) ¹⁰³ ;	Percentage of dose in Cdg versus the	30 mg/kg (TIR by volume ca.25% \P) ¹²²
	60 mg/kg ¹¹¹	minimum effective dose:	
		2.7/35=7.7%	
		Doses in Sdg and Cdg in the present study†:	Reported:
		17.4 mg/kg (TIR 28.4%) and 3.48 mg/kg	5 (TIR by volume ca. 80%¶) mg/kg ¹¹² ;
		17.4 mg/kg (11K 28.4%) and 5.48 mg/kg	9 (TIR by size ca. 35%¶) mg/kg ¹²³ ;
		Estimated minimum effective dose‡:	10 (TIR by volume ca. 54%¶) mg/kg ³³ ;
Pir	Reported:	19 mg/kg	10 (all mice died) and 20 (all mice died)
11	$8 \text{ mg/kg} (\text{TIR } 40\%)^{115}$	19 Hig/kg	mg/kg ¹⁰² ;
		Developes of deep in Cda versus the	10 (TIR 81.4%), 20 (TIR 90.8%), and 30 (
		Percentage of dose in Cdg versus the	mice died) mg/kg ⁴¹ ;
		minimum effective dose:	15 (Tumour regression 85.07%), 25 (MTD)
		3.48/19=18.3%	and 37.5 (toxic) mg/kg ⁶⁴
\ manco	for the percentage of the dose of each drug in Cdg equivalent to its minimum effective dose:	7.7%–19.5%,	
v range	to the percentage of the dose of each urug in Cug equivalent to its minimum effective dose:	average 16.3%	

^{*} Since the drugs (8 TOPO II inhibitors) used in this study were developed and applied in clinical practice during the early years, the effective anticancer dose of these drugs in animals, especially the effective dose for i.p. in mice with clear and complete information were difficult to find. For example, there was no record of the route of administration and/or the number of doses in some publications. Therefore, the dose information for each drug listed in this table is limited. The minimum effective dose for mouse i.p. was also difficult to determine from the reported doses listed in the table. The doses in the table indicate an accumulated dose in a cycle for each drug in the free-drug form. † The data for each single-drug group (Sdg) and each drug in the combination-drug group (Cdg) in the 8-drug solid tumour study.

[‡] We used the estimated dose to achieve 50% TIR as the minimum effective dose for i.p. in mice, which should be higher than the individual dose of each Sdg because none of the Sdgs had a TIR over 50% in the present study, and because different mouse tumour models^{81,88} or different treatment schedule⁸⁸ may respond differently to the same dose.

[§] A drug was considered to have anticancer activity if it produced T/C of $\geq 125\%$ 67,124, \geq ILS 30% 65, \geq T/C 130%65, \geq TIR 50%67, or \geq TIR 58%65. For ascites tumours, ILS (%) = (T/C -1) ×100, where T and C are the survival

periods (days) for treated and control mice, respectively 65 . For solid tumours: TIR (%) = $(1 - Wt/Wc) \times 100$, where Wt and Wc are the tumour weights in treated and control mice, respectively 65 . Longer survivals than 40 days in L1210 and P388 and 60 days in other ascites tumours with no retention of ascites were considered to be cured; mice without any trace of tumour at the termination of the experiments were considered to be cured 65 .

 $\label{eq:final_state} \textbf{§ These values are estimated by us according to the graphs in the literature} \textbf{§ These values are estimated by us according to the graphs in the literature} \textbf{§ 1.13,115,122,123}.$

** These data were obtained from three different mouse models for both etoposide and teniposide⁸¹.

†† The '-' means that no value was shown for groups in which > 50% of the animals were cured⁸¹.

‡‡ At this dose (24 mg/kg), amsacrine was ineffective against human malignant schwannoma, malignant lymphoma, malignant melanoma, liposarcoma, and neuroblastoma, but was active against testicular cancer95.

§§ These data were obtained from two different mouse models (or dosing schedules) for amsacrine¹¹⁴.

¶ These data were obtained from twelve different mouse ascites models (or dosing schedules) or thirteen different mouse solid (or human xenograft) models for aclarubicin⁸⁸.

††† The underlined dose is the ineffective dose for each drug.

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Acl, aclarubicin; Ams, amsacrine; ca., circa; Cdg, combination-drug group; Dox, doxorubicin; ED₅₀, 50% effective dose; Epi, epirubicin; Eto, etoposide; Ida, idarubicin; ILS, increase in life span (life-prolonging rate, LPR); i.p., intraperitoneal injection; i.v., intravenous injection; mg/kg, mg/kg of body weight; Mod, model; MTD, maximum tolerated dose; Pir, pirarubicin; Sd60 (%), the number of mice surviving to day 60/number of mice in a group; Sdg, single-drug group; T/C, median survival time of dying mice in a drug-treated (T) group divided by the median survival time of untreated mice in a tumour control (C) group; Ten, teniposide; TIR, tumour inhibition rate (by weight).

Supplementary Table 5. Concentrations of the six inhibitors in the additive experiment in

700 Hep G2 cells

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Group	Concentration (µM)
Doxorubicin IC ₁₈ *	0.74
Etoposide IC ₁₈	11.69
Daunorubicin IC ₁₈	0.30
Idarubicin IC ₁₈	0.10
Amsacrine IC ₁₈	4.89
Ellipticine IC ₁₈	0.66
Combination IC ₁₈	18.38

*Individual inhibitors were used as controls. The combination IC₁₈ represented that each inhibitor in the combination was combined at its IC₁₈ [a corresponding concentration of a 18% inhibition rate in its dose-response curve (Supplementary Fig.2) obtained in the previous experiment (i.e., an inhibitor concentration that inhibits cell viability by 18%—henceforth referred as IC₁₈)].

Supplementary Table 6. Concentrations of the six inhibitors in the additive experiment in

U87, HL-60, BxPC-3, and Hela cell lines

-	Concentration (µM)							
Cell	Group	Dox	Eto	Dau	Ida	Ams	Ell	Combination
	Comb IC _{4.5}	0.02	0.08	0.04	0.01	0.12	0.19	0.45
U87	Comb IC ₉	0.04	0.23	0.06	0.01	0.27	0.35	0.96
08/	Comb IC ₁₈	0.07	0.71	0.10	0.03	0.66	0.67	2.25
	Comb IC ₃₆	0.15	2.79	0.17	0.07	1.90	1.48	6.56
	Comb IC _{4.5}	0.04	2.04	0.02	0.01	2.79	0.49	5.39
Hela	Comb IC ₉	0.09	3.79	0.03	0.01	4.29	0.80	9.01
пеіа	Comb IC ₁₈	0.19	7.40	0.07	0.02	6.82	1.37	15.87
	Comb IC ₃₆	0.50	16.27	0.17	0.06	11.79	2.57	31.37
	Comb IC _{4.5}	0.12	0.22	0.27	0.00	0.15	0.10	0.86
HL-60	Comb IC ₉	0.20	0.47	0.37	0.00	0.25	0.19	1.49
ПL-00	Comb IC ₁₈	0.35	1.05	0.52	0.01	0.45	0.39	2.76
	Comb IC ₃₆	0.68	2.71	0.77	0.02	0.87	0.91	5.97
	Comb IC _{4.5}	0.23	2.35	0.06	0.02	1.97	0.58	5.21
BxPC-3	Comb IC ₉	0.32	4.69	0.09	0.04	3.05	0.91	9.10
DAFC-3	Comb IC ₁₈	0.46	9.84	0.15	0.06	4.89	1.46	16.86
	Comb IC ₃₆	0.70	23.60	0.25	0.12	8.53	2.58	35.77

Ams, amsacrine; Comb, combination; Dau, daunorubicin; Dox, doxorubicin; Ell, ellipticine; Eto, etoposide; Ida, idarubicin.

Supplementary Table 7. IC₅₀ values for the seven TOPO II inhibitors in the five cell lines

Cell	Dox (µM)	Eto (μM)	Dau (µM)	Ida (μM)	Ams (µM)	Ell (μM)	Pod (nM)
HepG2	1.59	39.08	0.76	0.40	13.96	2.22	16.52
Hela	0.90	26.39	0.31	0.11	16.49	3.79	
HL-60	1.03	4.84	0.98	0.04	1.32	1.52	
U87	0.23	6.43	0.24	0.12	3.61	2.40	
BxPC-3	0.91	40.23	0.35	0.17	11.98	3.64	

Ams, amsacrine; Dau, daunorubicin; Dox, doxorubicin; Ell, ellipticine; Eto, etoposide; Ida, idarubicin; Pod, Podophyllotoxin

Supplementary Table 8. Concentrations of the four inhibitors with similar IC₅₀ values in

the additive experiment in HepG2 cells

	Concentration (µM))			
Group	Doxorubicin	Daunorubicin	Idarubicin	Amsacrine	Combination
Doxorubicin IC ₁₈ *	0.74				
Daunorubicin IC ₁₈		0.30			
Idarubicin IC ₁₈			0.10		
Amsacrine IC ₁₈				4.89	
Comb IC _{4.5}	0.34	0.12	0.02	1.68	2.16
Comb IC ₉	0.5	0.19	0.05	2.81	3.55
Comb IC ₁₈	0.74	0.30	0.10	4.89	6.03
Comb IC ₃₆	1.19	0.54	0.23	9.38	11.34

*Individual inhibitors were used as controls. Comb, combination.

Supplementary Table 9. Concentrations of the four inhibitors with large IC₅₀ differences

in the additive experiment in HepG2 cells

	Concentration (µM	1)			
Group	Doxorubicin	Etoposide	Daunorubicin	Podophyllotoxin	Combination
Etoposide IC ₁₈ *		11.69			
Daunorubicin IC ₁₈			0.30		
Comb IC _{4.5}	0.34	3.43	0.12	0.006	3.90
Comb IC ₉	0.50	6.20	0.19	0.007	6.90
Comb IC ₁₈	0.74	11.69	0.30	0.010	12.74
Comb IC ₃₆	1.19	24.72	0.54	0.013	26.46

*Etoposide with IC₁₈ and daunorubicin with IC₁₈ were used as controls. Comb, combination.

Supplementary Table 10. Proportionally reduced concentrations of combined inhibitors in

HepG2 cell proliferation experiments

	Concentration (µM	()			
Group	Doxorubicin	Etoposide	Daunorubicin	Podophyllotoxin	Combination
Doxorubicin IC ₃₆ *	1.19				
Etoposide IC ₃₆		24.72			
Comb IC ₃₆ × 1	1.19	24.72	0.54	0.013	26.46
Comb IC ₃₆ × 0.75	0.89	18.54	0.40	0.01	19.84
Comb IC ₃₆ × 0.5	0.59	12.36	0.27	0.007	13.23
Comb IC ₃₆ × 0.25	0.30	6.18	0.13	0.003	6.61
Comb IC ₃₆ × 0.125	0.15	3.09	0.07	0.002	3.31

^{*} Doxorubicin IC₃₆ and etoposide IC₃₆ were used as controls. Comb, combination.

Supplementary Table 11. Comparison of antitumour activity (inhibition rates) among

combination groups composed of different inhibitors or different numbers of inhibitors in

733 HepG2 cells

Group	Combination of 4 inhibitors †	Combination of 4 inhibitors ‡	Combination of 6 inhibitors§
Comb IC _{4.5}	20.5 ± 11.1	18.9 ± 6.5	
Comb IC ₉	33.6 ± 5.1	32.9 ± 7.5	$63.2 \pm 1.2 *$
Comb IC ₁₈	60.5 ± 1.6	55.6 ± 4.4	$87.4 \pm 6.3*$
Comb IC ₃₆	87.4 ± 2.1	83.6 ± 1.3	

[†] Four inhibitors (doxorubicin, etoposide, daunorubicin, and podophyllotoxin) with large IC₅₀ value differences; ‡ Four inhibitors (doxorubicin, daunorubicin, idarubicin, and amsacrine) with similar IC₅₀ values; § Six inhibitors, including doxorubicin, etoposide, daunorubicin, idarubicin, amsacrine, and ellipticine. Comb, combination. * P < 0.05 compared with the combination of four inhibitors.

Supplementary Table 12. TOPO II assay treatment doses

Group	Low dose (µM)	Medium dose (μM)	High dose (μM)
Doxorubicin	1	10	50
Etoposide	50	100	200
Daunorubicin	1	10	50
Idarubicin	1	10	50
Amsacrine	10	50	
Ellipticine	1	10	
Drug combination	64	190	

TOPO, topoisomerase.

Supplementary Table 13. Gray values (GV) for TOPO II-mediated kDNA decatenation

Lane	Drug	Dose (µM)	GV	Inhibition rate
D	kDNA		971	
T	kDNA + TOPO II		9713	
1	kDNA + TOPO II + doxorubicin (low)	1	8832	9.1%
2	kDNA + TOPO II + doxorubicin (medium)	10	8198	15.6%
3	kDNA + TOPO II + doxorubicin (high)	50	1093	88.7%
4	kDNA + TOPO II + etoposide (low)	50	8940	8.0%
5	kDNA + TOPO II + etoposide (medium)	100	8105	16.6%
6	kDNA + TOPO II + daunorubicin (low)	1	9633	0.8%
7	kDNA + TOPO II + daunorubicin (medium)	10	8560	11.9%
8	kDNA + TOPO II + daunorubicin (high)	50	1249	87.1%
9	kDNA + TOPO II + idarubicin (low)	1	9023	7.1%
10	kDNA + TOPO II + idarubicin (medium)	10	8597	11.5%
11	kDNA + TOPO II + idarubicin (high)	50	1347	86.1%
12	kDNA + TOPO II + amsacrine (low)	10	8931	8.1%
13	kDNA + TOPO II + amsacrine (medium)	50	7819	19.5%
14	kDNA + TOPO II + ellipticine (low)	1	8589	11.6%
15	kDNA + TOPO II + ellipticine (medium)	10	7561	22.2%
16	kDNA + TOPO II + Comb (low)	64	7981	17.8%
17	kDNA + TOPO II + Comb (medium)	190	1531	84.2%

Comb, combination; TOPO, topoisomerase.

749 Supplementary Table 14. Gray values (GV) for TOPO II-mediated DNA uncoiling assay

Lane	Drug	Dose (µM)	GV	Inhibition rate
D	pBR322		651	
T	pBR322 + TOPO II		7510	
1	pBR322 + TOPO II + etoposide (medium)	100	6637	11.6%
2	pBR322 + TOPO II + Comb (medium)	190	814	89.2%
3	pBR322 + TOPO II + etoposide (high)	200	1373	81.7%

Comb, combination; TOPO, topoisomerase.

Supplementary Table 15. Comparison of experimental doses for each drug in 8-drug-Cdg used in the solid tumour mouse

model with clinical doses*

	Total dose† of each	Total dose† of each	Human		Percentage of the dose in 8-drug-	Percentage of the dose in
Drug	drug in 8-drug-Cdg	drug in 8-drug-Cdg	equivalent dose	Reported human dose per cycle (mg/kg, i.v.) ¶	Cdg equivalent to the minimum	8-drug-Cdg equivalent to
	(mg/kg, i.p.)	(mg/kg, i.v.)	(mg/kg, i.v.) §		human dose	the maximum human dos
				$0.68-2.03^{87}$; $0.81-1.08^{61}$; $0.81-3.24^{84}$; 1.08^{17} ; $1.08-2.03^{83}$; 1.35^{18} ;		
Doxorubicin	2.50	2.50	0.193	1.62–2.4390	28.4%	6.0%
				Dosage range#: 0.68-3.24		
D4	16.0	12.64	1.049	$2.70 - 13.51^{87}; 4.32^{17}; 4.73 - 13.51^{90}; 8.11^{125}; 10.14^{86}; 12.97^{126}; 13.51^{127}$	20.00/	7.8%
Etoposide	10.0	13.6‡	1.049	Dosage range#: 2.70–13.51	38.9%	7.8%
Idarubicin	0.75	0.64‡	0.049	$0.27 - 0.41^{118}$; $0.97^{90,127}$; $0.97 - 1.08^{87,128}$; $1.08^{118,129}$	18.1%	4.5%
				Dosage range#: 0.27-1.08	18.1%	
Amsacrine	3.75	2 104	0.246	4.05^{130} ; $8.11-17.03^{131}$; $10.14-16.89^{87}$; 13.51^{132}	6.1%	1.4%
Amsacrine	3./3	3.19‡	0.246	Dosage range#: 4.05-17.03	0.176	1.4/0
Aclarubicin	4.05	3.44‡	0.265	$1.08-2.65^{133}$; $1.62-8.11^{86}$; 2.16^{134}	24.5%	3.3%
Aciarubicin	4.03	3. 44 .	0.203	Dosage range#: 1.08-8.11	24.370	3.370
Teniposide	7.40	6.29‡	0.485	1.35–17.84 ⁸⁷ ; 2.70–12.16 ⁹⁰	35.9%	2.8%
Temposide	7.70	0.27+	0.403	Dosage range#: 1.35–17.48	33.970	2.070
Epirubicin	2.70	2.20÷	0.177	$0.95 - 4.05^{87}$; $2.70 - 3.24^{83,90}$	18.6%	4.4%
Epiruolein	2.70	2.30‡	0.1//	Dosage range#: 0.95-4.05	18.070	T.T/0
Pirarubicin	3.50	2.98‡	0.230	$0.81-2.16^{84}$; 1.62^{125}	28.4%	10.6%
FHAIUDICIII	3.30	2.704	0.230	Dosage range#: 0.81-2.16	20.470	10.070
Average value**					24.9%	5.1%

^{*} The clinical doses for each drug refer to commonly used clinical doses published in literature or on websites.

[†] The total dose represents the cumulative doses of five injections of the combination.

[‡] According to the literature, to convert the intravenous injection (i.v.) dose to intraperitoneal injection (i.p.) dose in mice, the bioavailability of drugs for intraperitoneal injection is usually 80%–85% (the ratio to intravenous injection is 1.18–1.25)¹³⁵; the bioavailability of 5 compounds, including docetaxel, for intraperitoneal injection was 69%, 18%, 94%, 102%, and 106%, with an average of 78% for the 5 compounds¹³⁶; no differences in plasma AUC_{0-6h} between intraperitoneal and intravenous administration routes for doxorubicin^{78,79}; therefore, we assumed that the remaining seven drugs (except doxorubicin) in the table had an i.p. bioavailability of 85% (a ratio of 1.18 to i.v.), and used the equation of the dose for i.v. (mg/kg) = the dose for i.p. (mg/kg) + 1.18, to convert the i.p. doses of the seven drugs to the i.v. doses of the seven drugs.

[§] For estimating the human equivalent dose (HED, mg/kg, i.v.) from the total dose of each drug in the 8-drug-Cdg (mg/kg, i.v.), we used the following equation: HED (mg/kg) = Animal dose (mg/kg) × (Animal Km / Human K_m), (where Human K_m = 60 kg/1.62 m² = 37.04 kg/m²; Animal (mouse) K_m = 0.02 kg/0.007 m² = 2.857 kg/m², in APPENDIX B of a FDA document⁶⁹ and other literature^{70–72}.

 $[\]P \ We \ converted \ published \ clinical \ doses \ from \ surface \ area \ doses \ (mg/m^2, i.v.) \ to \ body \ weight \ doses \ (mg/kg, i.v.) \ based \ on \ the \ equation^{69}: \ Human \ dose \ (mg/kg) = \ Human \ dose \ (mg/m^2)/ \ Human \ K_m = 37.04 \ kg/m^2.$

[#] The dosage range is derived from the combined results of the doses reported in the literature above.

^{**}The values of 24.9% and 5.1% were averaged to 15.0%.

Supplementary Table 16. Comparison of experimental doses for 8 Sdgs in the solid tumour mouse model with clinical doses*

Drug	Total dose† of each drug in 8 Sdgs	Total dose† of each drug in 8 Sdgs	Human equivalent dose	Reported human dose per cycle (mg/kg, i.v.) ¶	Percentage of the dose in Sdg equivalent to the minimum	Percentage of the dose in Sdg equivalent to the	
	(mg/kg, i.p.)	(mg/kg, i.v.)	(mg/kg, i.v.) §		human dose	maximum human dose	
				$0.68-2.03^{87}$; $0.81-1.08^{61}$; $0.81-3.24^{84}$; 1.08^{17} ; $1.08-2.03^{83}$; 1.35^{18} ;			
Doxorubicin	12.50	12.50	0.964	1.62–2.4390	141.8%20	29.8%	
				Dosage range#: 0.68-3.24			
Etoposide	80.0	68.0‡	5.243	$2.70 - 13.51^{87}; 4.32^{17}; 4.73 - 13.51^{90}; 8.11^{125}; 10.14^{86}; 12.97^{126}; 13.51^{127}$	194.2%30	38.8%	
	80.0			Dosage range#: 2.70-13.51	194.27030		
Idarubicin 3.75	2 75	3.19‡	0.246	$0.27 - 0.41^{118}$; $0.97^{90,127}$; $0.97 - 1.08^{87,128}$; $1.08^{118,129}$	91.1%50	22.8%	
	3.73			Dosage range#: 0.27-1.08	91.17030		
Amsacrine	18.75	15.94‡	1.229	4.05^{130} ; $8.11-17.03^{131}$; $10.14-16.89^{87}$; 13.51^{132}	30.3%20	7.2%	
Allisacrille	16.75			Dosage range#: 4.05-17.03	30.37020		
Aclarubicin	20.25	17.21‡	1.327	$1.08 - 2.65^{133}$; $1.62 - 8.11^{86}$; 2.16^{134}	122.9%30	16.4%	
Aciarubiciii	20.23			Dosage range#: 1.08-8.11	122.77030		
Teniposide	37.0	31.45‡	2.425	$1.35-17.84^{87}$; $2.70-12.16^{90}$	179.6%20	13.9%	
				Dosage range#: 1.35–17.48	177.07020		
Epirubicin	13.5	11.48‡	0.885	$0.95 - 4.05^{87}$; $2.70 - 3.24^{83,90}$	93.2%40	21.9%	
Ерпиони				Dosage range#: 0.95-4.05	93.27040		
Pirarubicin	17.5	14.88‡	1.147	$0.81-2.16^{84}$; 1.62^{125}	141.7%10	53.1%	
r narudiciii	17.5			Dosage range#: 0.81-2.16			
Average value**					124.4%	25.5%	

Notes for *, ‡, ¶, and #, see Supplementary Table 15.

[†] The total dose represents the cumulative doses of five injections for each drug in the Sdg.

[§] For estimating the human equivalent dose (HED, mg/kg, i.v.) from the total dose of each drug in the Sdg, see § in Supplementary Table 15.

^{**}The values of 124.4% and 25.5% were averaged to 75.0%.

Supplementary Table 17. Combination index (CI) for U87 cell experiment for combinations of six TOPO II inhibitors

Group	Concentration (µM)						Measured Inhibition (%)	CI value
	Dox	Eto	Dau	Ida	Ams	Ell	Weasured Himblion (70)	
Comb 1 (IC _{4.5})	0.02	0.08	0.04	0.01	0.12	0.19	28.3	0.92
Comb 2 (IC ₉)	0.04	0.23	0.06	0.01	0.27	0.35	36.4	0.97
Comb 3 (IC ₁₈)	0.07	0.71	0.10	0.03	0.66	0.67	47.5	1.01
Comb 4 (IC ₃₆)	0.15	2.79	0.17	0.07	1.9	1.48	63.2	1.01

Ams, amsacrine; Comb, combination; CI, combination index; Dau, daunorubicin; Dox, doxorubicin; Ell, ellipticine; Eto, etoposide; Ida, idarubicin; TOPO, topoisomerase.

- 5. Supplementary References (including references 0–66, which have already appeared in the main text and the method section)
- 1. Yazbeck, V. et al. Chapter One An overview of chemotoxicity and radiation toxicity in cancer therapy. in Advances in Cancer
- 782 Research (eds. Gewirtz, D. A. & Fisher, P. B.) vol. 155, 1–27 (2022).
- 2. Gewirtz, D. A. & Fisher, P. B. Preface. in Advances in Cancer Research (eds. Gewirtz, D. A. & Fisher, P. B.) vol. 155, xi-xii
- 784 (2022).
- 785 3. Lakshmikuttyamma, A., Patel, H., Kim, H. & Hajjar, E. Chapter 5 Side effects of antipsychotic agents. in *Side Effects of Drugs*
- 786 Annual (ed. Ray, S. D.) vol. 45, 49–60 (Elsevier, 2023).
- 4. Perucca, P. & Gilliam, F. G. Adverse effects of antiepileptic drugs. *Lancet Neurol.* 11, 792–802 (2012).
- 5. Chen, C., Xie, D., Gewirtz, D. A. & Li, N. Chapter Three Nephrotoxicity in cancer treatment: An update. in *Advances in Cancer*
- 789 Research (eds. Gewirtz, D. A. & Fisher, P. B.) vol. 155, 77–129 (2022).
- 790 6. Rudd, S. G. Targeting pan-essential pathways in cancer with cytotoxic chemotherapy: Challenges and opportunities. *Cancer*
- 791 *Chemother. Pharmacol.* **92**, 241–251 (2023).
- 792 7. Fundytus, A. et al. Access to cancer medicines deemed essential by oncologists in 82 countries: An international, cross-sectional
- 793 survey. *Lancet Oncol.* **22**, 1367–1377 (2021).
- 794 8. WHO Model Lists of Essential Medicines-23rd list (2023).
- 795 https://www.who.int/groups/expert-committee-on-selection-and-use-of-essential-medicines/essential-medicines-lists
- 796 https://www.who.int/publications/i/item/WHO-MHP-HPS-EML-2023.02

- 9. Naaktgeboren, W. R. et al. Efficacy of physical exercise to offset anthracycline-induced cardiotoxicity: A systematic review and
- meta-analysis of clinical and preclinical studies. J. Am. Heart Assoc. 10, e021580 (2021).
- 799 10. Alexander, J. L. et al. Gut microbiota modulation of chemotherapy efficacy and toxicity. Nat. Rev. Gastroenterol. Hepatol. 14,
- 800 356–365 (2017).
- 11. Turner, N., Biganzoli, L. & Di Leo, A. Continued value of adjuvant anthracyclines as treatment for early breast cancer. Lancet
- 802 *Oncol.* **16**, e362-369 (2015).
- 12. Akbarali, H. I., Muchhala, K. H., Jessup, D. K. & Cheatham, S. Chapter Four Chemotherapy induced gastrointestinal toxicities.
- in Advances in Cancer Research (eds. Gewirtz, D. A. & Fisher, P. B.) vol. 155, 131–166 (2022).
- 13. Mauro, A. G., Hunter, K. & Salloum, F. N. Chapter Five Cardiac complications of cancer therapies. in *Advances in Cancer*
- 806 Research (eds. Gewirtz, D. A. & Fisher, P. B.) vol. 155, 167–214 (2022).
- 14. Kahn, A. M., Blenman, K. R. M., Sonis, S. T. & Lustberg, M. B. Chapter Six Strategies to mitigate the toxicity of cancer
- therapeutics. in *Advances in Cancer Research* (eds. Gewirtz, D. A. & Fisher, P. B.) vol. 155, 215–244 (2022).
- 15. Renu, K., Pureti, L. P., Vellingiri, B. & Valsala Gopalakrishnan, A. Toxic effects and molecular mechanism of doxorubicin on
- 810 different organs an update. *Toxin Rev.* **41**, 650–674 (2022).
- 16. Tacar, O., Sriamornsak, P. & Dass, C. R. Doxorubicin: An update on anticancer molecular action, toxicity and novel drug delivery
- 812 systems. *J. Pharm. Pharmacol.* **65**, 157–170 (2013).

- 17. Lakshman, A. et al. Efficacy of VDT PACE-like regimens in treatment of relapsed/refractory multiple myeloma. Am. J. Hematol.
- **93**, 179–186 (2018).
- 18. Swain, S. M., Whaley, F. S. & Ewer, M. S. Congestive heart failure in patients treated with doxorubicin. *Cancer* **97**, 2869–2879
- 816 (2003).
- 19. Agarwal, S. et al. Statins for primary prevention of anthracycline chemotherapy-related cardiac dysfunction: A systematic review
- and meta-analysis of randomized controlled trials. *Am. J. Cardiol.* **206**, 63–66 (2023).
- 20. Fourie Zirkelbach, J. et al. Improving dose-optimization processes used in oncology drug development to minimize toxicity and
- maximize benefit to patients. *J. Clin. Oncol.* **40**, 3489–3500 (2022).
- 21. Scharovsky, O. G., Mainetti, L. E. & Rozados, V. R. Metronomic chemotherapy: Changing the paradigm that more is better. Curr.
- 822 *Oncol.* **16**, 7–15 (2009).
- 22. Greish, K., Sawa, T., Fang, J., Akaike, T. & Maeda, H. SMA-doxorubicin, a new polymeric micellar drug for effective targeting to
- solid tumours. *J. Control. Release* **97**, 219–230 (2004).
- 23. Borchmann, P. et al. PET-guided treatment in patients with advanced-stage Hodgkin's lymphoma (HD18): Final results of an
- open-label, international, randomised phase 3 trial by the German Hodgkin study group. *The Lancet* **390**, 2790–2802 (2017).
- 827 24. Behringer, K. et al. Gonadal function and fertility in survivors after Hodgkin lymphoma treatment within the German Hodgkin
- study group HD13 to HD15 trials. *J. Clin. Oncol.* **31**, 231–9 (2013).

- 25. Pomeroy, A. E., Schmidt, E. V., Sorger, P. K. & Palmer, A. C. Drug independence and the curability of cancer by combination
- 830 chemotherapy. *Trends Cancer* **8**, 915–929 (2022).
- 26. Kerbel, R. S. & Kamen, B. A. The anti-angiogenic basis of metronomic chemotherapy. *Nat. Rev. Cancer* 4, 423–436 (2004).
- 832 27. Mahdy, A. et al. Outcome and toxicity of ifosfamide, carboplatin, and etoposide versus gemcitabine and vinorelbine regimen for
- paediatric patients with relapsed or refractory Hodgkin's lymphoma. Front. Oncol. 13, 1153128 (2023).
- 28. Fudio, S. et al. Anti-cancer drug combinations approved by US FDA from 2011 to 2021: Main design features of clinical trials
- and role of pharmacokinetics. Cancer Chemother. Pharmacol. 90, 285–299 (2022).
- 29. Jia, J. et al. Mechanisms of drug combinations: Interaction and network perspectives. Nat. Rev. Drug Discov. 8, 111–128 (2009).
- 30. Lehár, J. et al. Synergistic drug combinations tend to improve therapeutically relevant selectivity. Nat. Biotechnol. 27, 659–666
- 838 (2009).
- 31. Fernandes Neto, J. M. et al. Multiple low dose therapy as an effective strategy to treat EGFR inhibitor-resistant NSCLC tumours.
- 840 *Nat. Commun.* **11**, 3157 (2020).
- 32. Hu, W. et al. Discovery of novel topoisomerase II inhibitors by medicinal chemistry approaches. J. Med. Chem. 61, 8947–8980
- 842 (2018).
- 843 33. Yi, X. et al. Co-delivery of pirarubicin and paclitaxel by human serum albumin nanoparticles to enhance antitumour effect and
- reduce systemic toxicity in breast cancers. *Mol. Pharm.* **12**, 4085–4098 (2015).

- 34. Motyka, S. *et al.* Podophyllotoxin and its derivatives: Potential anticancer agents of natural origin in cancer chemotherapy.
- 846 Biomed. Pharmacother. 158, 114145 (2023).
- 35. Gilad, Y., Gellerman, G., Lonard, D. M. & O'Malley, B. W. Drug combination in cancer treatment—from cocktails to conjugated
- 848 combinations. *Cancers* **13**, 669 (2021).
- 36. Cai, S.-Q. et al. A combination drug with both 'additive effect' on the efficacy and 'toxicity scattering effect'. China patent
- announcement 2016, CN105288629A,1–26 (2016); Patent application No. 201410234283.6 (2014); Patent No. ZL 2014 1
- 851 0234283.6 (2021).
- 37. Cai, S. Q., Feng, X., Shang, M. Y. & Wang, X. Design idea of 'ultra hypotoxic multidrug' based on the "efficacy theory" of
- traditional Chinese medicines. in *Planta Medica* **82** (S 01), S1-S381 (Georg Thieme Verlag KG, 2016).
- 38. Xu, F., Yang, D., Shang, M., Wang, X. & Cai, S. Effective forms, additive effect, and toxicities scattering effect of
- pharmacodynamic substances of TCMs some reflections evoked by the study on the metabolic disposition of traditional Chinese
- medicines (TCM). World Sci. Technol.-Mod. Tradit. Chin. Med. 12, 688–703 (2014).
- 39. Cai, S., Wang, X., Shang, M., Xu, F. & Liu, G. 'efficacy theory' may help to explain characteristic advantages of traditional
- 858 Chinese medicines. Zhongguo Zhong Yao Za Zhi = Zhongguo Zhongyao Zazhi = China J. Chin. Mater. Medica 40, 3435–3443
- 859 (2015).
- 40. Rudmann, D. G. On-target and off-target-based toxicologic effects. *Toxicol. Pathol.* 41, 310–314 (2013).
- 41. Zhou, J. et al. Novel lipid hybrid albumin nanoparticle greatly lowered toxicity of pirarubicin. Mol. Pharm. 10, 3832–3841 (2013).

- 42. Ozer, J. S. et al. Enhancing the utility of alanine aminotransferase as a reference standard biomarker for drug-induced liver injury.
- 863 Regul. Toxicol. Pharmacol. **56**, 237–246 (2010).
- 43. You, J. et al. A medicinal and edible formula YH0618 ameliorates the toxicity induced by doxorubicin via regulating the
- expression of bax/bcl-2 and FOXO4. *J. Cancer* **10**, 3665–3677 (2019).
- 44. Fan, H. et al. Comparison of liver biomarkers in 288 COVID-19 patients: A mono-centric study in the early phase of pandemic.
- 867 Front. Med. 7, 584888 (2021).
- 45. He, X. et al. Encapsulation of teniposide into albumin nanoparticles with greatly lowered toxicity and enhanced antitumour
- activity. Int. J. Pharm. 487, 250–259 (2015).
- 46. Karale, S., Yamuna, P. & Kamath, J. V. Protective effect of capsaicin against doxorubicin induced cardiotoxicity in experimental
- rats. Indian J. Pharm. Educ. Res. **54**, 95–100 (2019).
- 47. Pujara, N. & Chaudhary, A. Evaluation of creatine kinase and lactate dehydrogenase for liver abnormalities: A biochemical study.
- 873 *Int. J. Adv. Med.* **4**, 1355–1357 (2017).
- 48. Zheng, Q., Wang, H., Hou, W. & Zhang, Y. Use of anti-angiogenic drugs potentially associated with an increase on serum AST,
- LDH, CK, and CK-MB activities in patients with cancer: A retrospective study. Front. Cardiovasc. Med. 8, 755191 (2021).
- 49. Forkasiewicz, A. et al. The usefulness of lactate dehydrogenase measurements in current oncological practice. Cell. Mol. Biol. Lett.
- **25**, 35 (2020).
- 878 50. Albright, R. C. Acute renal failure: A practical update. *Mayo Clin. Proc.* **76**, 67–74 (2001).

- 51. Obermüller, N., Geiger, H., Weipert, C. & Urbschat, A. Current developments in early diagnosis of acute kidney injury. *Int. Urol.*
- 880 *Nephrol.* **46**, 1–7 (2014).
- 52. Sung, H. et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185
- 882 countries. CA. Cancer J. Clin. 71, 209–249 (2021).
- 53. Silva, E., Rajapakse, N. & Kortenkamp, A. Something from "nothing" Eight weak estrogenic chemicals combined at
- concentrations below NOECs produce significant mixture effects. *Environ. Sci. Technol.* **36**, 1751–1756 (2002).
- 885 54. Puts, M. T. E. et al. Medication problems in older, newly diagnosed cancer patients in Canada: How common are they? a
- prospective pilot study. *Drugs Aging* **26**, 519–536 (2009).
- 887 55. McCourt, E. et al. The effect of computerised physician order entry on prescribing errors: An interrupted time-series study at a
- secondary referral hospital in Australia. *Int. J. Med. Inf.* **165**, 104829 (2022).
- 889 56. Maggiore, R. J., Gross, C. P. & Hurria, A. Polypharmacy in older adults with cancer. *The Oncologist* 15, 507–522 (2010).
- 57. Sorensen, L., Stokes, J. A., Purdie, D. M., Woodward, M. & Roberts, M. S. Medication management at home: Medication-related
- risk factors associated with poor health outcomes. Age Ageing **34**, 626–632 (2005).
- 892 58. Beijing Municipal Health Commission. Guidelines for Special Evaluation of Prescriptions in Beijing Medical Institutions (Trial).
- 893 (2012).
- http://www.nhc.gov.cn/zwgkzt/s9968/201212/f76fcb555824429195c131efc24372d2.shtml

- 895 59. National Health Commission of the People's Republic of China. Regulations for the Management of Hospital Prescription Review
- 896 (Trial). (2010).
- 897 http://www.nhc.gov.cn/zwgk/glgf/201306/094ebc83dddc47b5a4a63ebde7224615.shtml
- 898 60. Muchtar, E. et al. First line and salvage therapy with total therapy 3-based treatment for multiple myeloma—an extended single
- septiment to the septiment of the septim
- 900 61. Barlogie, B. et al. Incorporating bortezomib into upfront treatment for multiple myeloma: Early results of total therapy 3. Br. J.
- 901 *Haematol.* **138**, 176–185 (2007).
- 62. Toft, S., Horwitz, H. & Dalhoff, K. P. Long-term mortality after poisoning with antipsychotics. Clin. Toxicol. 55, 267–274 (2017).
- 903 63. Dincer, B. et al. Antipsychotics induced reproductive toxicity by stimulating oxidative stress: A comparative in vivo and in silico
- 904 study. *Chem. Biodivers.* **20**, e202201190 (2023).
- 905 64. Anees, M., Tiwari, S., Mehrotra, N., Kharbanda, S. & Singh, H. Development and evaluation of PLA based hybrid block
- copolymeric nanoparticles for systemic delivery of pirarubicin as an anti-cancer agent. *Int. J. Pharm.* **620**, 121761 (2022).
- 907 65. Kunimoto, T. et al. Antitumour activity of 7-ethyl-10-[4-(1-piperidino)-1-piperidino]carbonyloxy-camptothec in, a novel water-
- soluble derivative of camptothecin, against murine tumours. *Cancer Res.* 47, 5944–5947 (1987).
- 909 66. Zhou, X. et al. Synergistic effects of Chinese herbal medicine: A comprehensive review of methodology and current research.
- 910 Front. Pharmacol. 7, 201 (2016).

- 67. Rose, W. C. et al. Preclinical antitumour activity of a soluble etoposide analog, BMY-40481-30. Invest. New Drugs 8, S25–S32
- 912 (1990).
- 68. Kano, Y. et al. Effects of amsacrine in combination with other anticancer agents in human acute lymphoblastic leukemia cells in
- 914 culture. *Leukemia Res.* **15**, 1059–1066 (1991).
- 915 69. US Food and Drug Administration (USFDA). Guidance for Industry Estimating the Maximum Safe Starting Dose in Initial
- 916 Clinical Trials for Therapeutics in Adult Healthy Volunteers. (2005). Available from:
- 917 https://www.fda.gov/regulatory-information/search-fda-guidance-documents/estimating-maximum-safe-starting-dose-initial-
- 918 clinical-trials-therapeutics-adult-healthy-volunteers
- 919 https://www.fda.gov/media/72309/download
- 70. Reagan-Shaw, S., Nihal, M. & Ahmad, N. Dose translation from animal to human studies revisited. *FASEB J.* **22**, 659–661 (2008).
- 71. Nair, A. B. & Jacob, S. A simple practice guide for dose conversion between animals and human. J. Basic Clin. Pharm. 7, 27–31
- 922 (2016).
- 72. Saadh, M. J., Haddad, M., Dababneh, M. F., Bayan, M. F. & Al-Jaidi, B. A. A guide for estimating the maximum safe starting
- dose and conversion it between animals and humans. Syst. Rev. Pharm. 11, 98–101 (2020).
- 73. Vejpongsa, P. & Yeh, E. T. H. Prevention of anthracycline-induced cardiotoxicity: Challenges and opportunities. J. Am. Coll.
- 926 *Cardiol.* **64**, 938–945 (2014).

- 74. Minotti, G., Menna, P., Salvatorelli, E., Cairo, G. & Gianni, L. Anthracyclines: Molecular advances and pharmacologic
- developments in antitumour activity and cardiotoxicity. *Pharmacol. Rev.* **56**, 185–229 (2004).
- 75. Hande, K. R. Topoisomerase II inhibitors. *Update Cancer Ther.* **3**, 13–26 (2008).
- 930 76. Giantris, A., Abdurrahman, L., Hinkle, A., Asselin, B. & Lipshultz, S. E. Anthracycline-induced cardiotoxicity in children and
- 931 young adults. Crit. Rev. Oncol. Hematol. 27, 53–68 (1998).
- 932 77. Saad, S. Y., Najjar, T. A. & Al-Rikabi, A. C. The preventive role of deferoxamine against acute doxorubicin-induced cardiac,
- renal and hepatic toxicity in rats. *Pharmacol. Res.* **43**, 211–218 (2001).
- 78. Kullenberg, F. et al. The progression of doxorubicin-induced intestinal mucositis in rats. Naunyn-Schmiedeberg's Arch.
- 935 *Pharmacol.* **396**, 247–260 (2023).
- 79. Johansen, P. B. Doxorubicin pharmacokinetics after intravenous and intraperitoneal administration in the nude mouse. *Cancer*
- 937 Chemoth. Pharm. 5, 267–270 (1981).
- 80. The values of LD₅₀ in Supplementary Table 3 were obtained from Chemical Toxicity Database:
- http://www.drugfuture.com/toxic/search.aspx/.
- 81. Jensen, P. B. et al. Antitumour activity of the two epipodophyllotoxin derivatives VP-16 and VM-26 in preclinical systems: A
- comparison of in vitro and in vivo drug evaluation. *Cancer Chemother. Pharmcol.* **27**, 194–198 (1990).
- 82. Platel, D., Pouna, P., Bonoron-Adèle, S. & Robert, J. Comparative cardiotoxicity of idarubicin and doxorubicin using the isolated
- perfused rat heart model. Anti-Cancer Drugs 10, 671–676 (1999).

- 944 83. Drugs@FDA: FDA-Approved Drugs, for hydrochloride of doxorubicin, teniposide and epirubicin.
- 945 https://www.accessdata.fda.gov/scripts/cder/daf/index.cfm.
- 84. Shinozaki, T., Watanabe, H., Yanagawa, T., Shirakura, K. & Takagishi, K. Pirarubicin-based versus doxorubicin-based
- osteosarcoma chemotherapy. *Ann. Pharmacother.* **36**, 996–999 (2002).
- 948 85. Owattanapanich, W., Owattanapanich, N., Kungwankiattichai, S., Ungprasert, P. & Ruchutrakool, T. Efficacy and toxicity of
- 949 idarubicin versus high-dose daunorubicin for induction chemotherapy in adult acute myeloid leukemia: A systematic review and
- 950 meta-analysis. Clin. Lymph. Myelom. Leuk. 18, 814-821. (2018).
- 86. Jin, J. et al. Homoharringtonine in combination with cytarabine and aclarubicin resulted in high complete remission rate after the
- first induction therapy in patients with de novo acute myeloid leukemia. *Leukemia* **20**, 1361–1367 (2006).
- 87. A website of the Drug Index on "BC Cancer" website of Provincial Health Services Authority (PHSA) in British Columbians, for
- monographs of doxorubicin, etoposide, idarubicin, amsacrine, teniposide, and epirubicin. http://www.bccancer.bc.ca/health-
- professionals/clinical-resources/cancer-drug-manual/drug-index
- 88. Oki, T., Takeuchi, T., Oka, S. & Umezawa, H. New Anthracycline Antibiotic Aclacinomycin A: Experimental Studies and
- Correlations with Clinical Trials. in *New Drugs in Cancer Chemotherapy* (eds. Carter, S. K., Sakurai, Y. & Umezawa, H.) 21–40
- 958 (Springer, Berlin, Heidelberg, 1981).
- 89. Ryberg, M. et al. New insight into epirubicin cardiac toxicity: Competing risks analysis of 1097 breast cancer patients. JNCI: J.
- 960 *Natl. Cancer Inst.* **100**, 1058–1067 (2008).

- 90. Chemotherapeutic Agents and Their Uses, Dosages, and Toxicities (2016). Cancer Network, for doxorubicin, etoposide,
- idarubicin, teniposide, and epirubicin. https://www.cancernetwork.com/view/chemotherapeutic-agents-and-their-uses-dosages-
- and-toxicities.
- 91. Takahashi, N. et al. TOXICITY STUDIES OF VP 16-213 (II): Oral one-month subacute toxicity in rats. J. Toxicol. Sci. 11, 17-49
- 965 (1986).
- 92. Lee, J. S. et al. Safety and efficacy of intraperitoneal injection of etoposide in oil suspension in mice with peritoneal
- carcinomatosis. Cancer Chemother. Pharmcol. **36**, 211–216 (1995).
- 93. Holm, B., Jensen, P. B., Sehested, M. & Hansen, H. H. In vivo inhibition of etoposide-mediated apoptosis, toxicity, and
- antitumour effect by the topoisomerase II-uncoupling anthracycline aclarubicin. Cancer Chemother. Pharmcol. **34**, 503–508
- 970 (1994).
- 94. Kuhlmann, O., Hofmann, S. & Weiss, M. Pharmacokinetics and toxicity of idarubicin in the rat. Eur. J. Drug Metab.
- 972 *Pharmacokinet.* **26**, 215–219 (2001).
- 95. Sordillo, P. P., Helson, L. & Lesser, M. Evaluation of 4'-(9-acridinylamino) methanesulfon-m-anisidide (m-AMSA, NSC 249992)
- on human tumours in nude mice. Cancer Clin. Trials 3, 385–389 (1980).
- 96. Han, B. et al. Preparation of aclarubicin PLGA nanospheres and related in vitro/in vivo studies. J. Appl. Polym. Sci. 117, 2754—
- 976 2761 (2010).

- 97. Vyadro, M. M. & Navashin, S. M. Antitumour activity of liposomal aclarubicin in vitro and in vivo. Bull. Exp. Biol. Med. 112,
- 978 1634–1636 (1991).
- 98. Cheng, J. et al. Discovery of 4,6-O-thenylidene-β-d-glucopyranoside-(2"-acetamido, 3"-acetyl-di-S-5-fluorobenzothizole/5-
- 980 fluorobenzoxazole)-4'-demethylepipodophyllotoxin as potential less toxic antitumour candidate drugs by reducing DNA damage
- and less inhibition of PI3K. J. Med. Chem. **63**, 2877–2893 (2020).
- 982 99. Santucci, L. et al. Cardiac safety and antitumoral activity of a new nitric oxide derivative of pegylated epirubicin in mice. Anti-
- 983 *Cancer Drugs* **18**, 1081 (2007).
- 984 100. Shi, H. et al. RING finger protein 10 regulates AP-1/meox2 to mediate pirarubicin-induced cardiomyocyte apoptosis. Oxid.
- 985 *Med. Cell. Longevity* **2023**, 7872193 (2023).
- 986 101. Shi, H., Yan, Y., Yang, H., Pu, P. & Tang, H. Schisandrin B diet inhibits oxidative stress to reduce ferroptosis and lipid
- peroxidation to prevent pirarubicin-induced hepatotoxicity. *Biomed Res. Int.* **2022**, 5623555 (2022).
- 988 102. Greish, K., Nagamitsu, A., Fang, J. & Maeda, H. Copoly (styrene-maleic acid)-pirarubicin micelles: High tumour-targeting
- efficiency with little toxicity. *Bioconjugate Chem.* **16**, 230–236 (2005).
- 990 103. Wang, Z.-M. et al. Efficacy and safety of anlotinib, a multikinase angiogenesis inhibitor, in combination with epirubicin in
- preclinical models of soft tissue sarcoma. *Cancer Med.* **9**, 3344–3352 (2020).
- 992 104. Lan, Y., Chen, L., Zhu, J., Li, H. & Lin, Z. Scutellarein regulates the PTEN/AKT/NFκB signaling pathway to reduce
- pirarubicin related liver inflammation. *Int. J. Mol. Med.* **52**, 55 (2023).

- 994 105. Lu, Y. et al. Eupatilin attenuates doxorubicin-induced cardiotoxicity by activating the PI3K-AKT signaling pathway in mice.
- 995 *Mol. Cell. Biochem.* **479**, 869–880 (2024).
- 996 106. Lu, W.-L. et al. A pegylated liposomal platform: Pharmacokinetics, pharmacodynamics, and toxicity in mice using
- 997 doxorubicin as a model drug. *J. Pharmacol. Sci.* **95**, 381–389 (2004).
- 998 107. Ito, T. et al. Beneficial effect of taurine treatment against doxorubicin-induced cardiotoxicity in mice. Adv. Exp. Med. Biol. 643,
- 999 65–74 (2009).
- 1000 108. Hagiwara, A. et al. A new dosage form comprising a suspension of activated carbon particles adsorbing aclarubicin: Toxicity
- in mice. Anti-Cancer Drugs 2, 365–369 (1991).
- 1002 109. Kullenberg, F. et al. Anakinra and dexamethasone treatment of idarubicin-induced mucositis and diarrhoea in rats. Basic Clin.
- 1003 *Pharmacol Toxicol.* **132**, 507–516 (2023).
- 1004 110. Santos, N. D. et al. Substantial increases in idarubicin plasma concentration by liposome encapsulation mediates improved
- antitumor activity. *J. Control. Release* **105**, 89–105 (2005).
- 1006 111. Wang, N. et al. Melatonin protects against epirubicin-induced ovarian damage. J. Reprod. Dev. 66, 19–27 (2020).
- 1007 112. Nakamura, H. et al. Comparison between linear and star-like HPMA conjugated pirarubicin (THP) in pharmacokinetics and
- antitumour activity in tumour bearing mice. Eur. J. Pharm. Biopharm. 90, 90–96 (2015).
- 1009 113. Sadzuka, Y., Sugiyama, T. & Sonobe, T. Improvement of idarubicin induced antitumour activity and bone marrow suppression
- 1010 by theanine, a component of tea. *Cancer Lett.* **158**, 119–124 (2000).

- 1011 114. Baguley, B. C. et al. Synthesis, antitumour activity, and DNA binding properties of a new derivative of amsacrine, N-5-
- dimethyl-9-[(2-methoxy-4-methylsulfonylamino) phenylamino]-4-acridinecarboxamide. Cancer Res. 44, 3245–3251 (1984).
- 1013 115. Sugiyama, T. et al. Membrane transport and antitumour activity of pirarubicin, and comparison with those of doxorubicin. Jpn.
- 1014 J. Cancer Res. 90, 775–780 (1999).
- 1015 116. Silva, R. L. dos S. et al. Protective effect of gallic acid on doxorubicin-induced ovarian toxicity in mouse. Reprod. Toxicol.
- 1016 **115**, 147–156 (2023).
- 1017 117. Gabizon, A., Tzemach, D., Mak, L., Bronstein, M. & Horowitz, A. T. Dose dependency of pharmacokinetics and therapeutic
- efficacy of pegylated liposomal doxorubicin (DOXIL) in murine models. J. Drug Targeting 10, 539–548 (2002).
- 1019 118. Attia, S. M. et al. Dexrazoxane averts idarubicin-evoked genomic damage by regulating gene expression profiling associated
- with the DNA damage-signaling pathway in BALB/c mice. *Toxicol. Sci.* **160**, 161–172 (2017).
- 1021 119. Ruchatz, H. et al. Effect of imatinib on haematopoietic recovery following idarubicin exposure. Leukemia 17, 298–304 (2003).
- 1022 120. Osieka, R., Houchens, D. P., Goldin, A. & Johnson, R. K. Chemotherapy of human colon cancer xenografts in athymic nude
- mice. Cancer **40**, 2640–2650 (1977).
- 1024 121. Etiévant, C. et al. F 11782, a dual inhibitor of topoisomerases I and II with an original mechanism of action in vitro, and
- markedly superior in vivo antitumour activity, relative to three other dual topoisomerase inhibitors, intoplicin, aclarubicin and
- 1026 TAS-103. Cancer Chemother. Pharmcol. 46, 101–113 (2000).

- 1027 Tang, H. et al. Co-delivery of epirubicin and paclitaxel using an estrone-targeted PEGylated liposomal nanoparticle for breast
- 1028 cancer. Int. J. Pharm. 573, 118806 (2020).
- 1029 123. Eskandari, Z. et al. Targeting breast cancer using pirarubicin-loaded vasoactive intestinal peptide grafted sterically stabilized
- micelles. Eur. J. Pharm. Sci. 162, 105830 (2021).
- 1031 124. Rose, W. C., Schurig, J. E., Huftalen, J. B. & Bradner, W. T. Experimental antitumour activity and toxicity of a new
- chemotherapeutic agent, BBM 928A. Cancer Res. 43, 1504–1510 (1983).
- 1033 125. Saito, S. et al. Efficacy of combination-chemotherapy with pirarubicin, ifosfamide, and etoposide for soft tissue sarcoma: A
- single-institution retrospective analysis. *BMC Cancer* **20**, 868 (2020).
- 1035 126. Amadori, S. et al. Mitoxantrone, etoposide, and intermediate-dose cytosine arabinoside (MEC): An effective regimen for
- refractory acute myeloid leukemia in *Acute Leukemias* (eds. Hiddemann, W. et al.) vol. 34, 401–403 (Springer-Verlag, Berlin;
- 1037 1992).
- 1038 127. Haas, R. et al. Induction Therapy for Acute Myelogenous Leukemia with Idarubicin/Cytosine Arabinoside and
- Mitoxantrone/Etoposide in "Response-Adapted" Sequence. in *Acute Leukemias* (eds. Hiddemann, W. et al.) vol. 34, 439–444
- 1040 (Springer, Berlin, Heidelberg, 1992).
- 1041 128. Reiffers, J. et al. Prospective Study Comparing Idarubicin and Daunorubicin in Elderly Patients with Acute Myeloid Leukemia.
- in Acute Leukemias (eds. Hiddemann, W. et al.) vol. 34, 628–631 (Springer Berlin Heidelberg, Berlin, Heidelberg, 1992).

- 1043 129. Weiss, M. A. et al. A single, high dose of idarubicin combined with cytarabine as induction therapy for adult patients with
- recurrent or refractory acute lymphoblastic leukemia. *Cancer* **95**, 581–587 (2002).
- 130. Heinemann, V. & Jehn, U. Intermediate-Dose Arabinoside and Amsacrine: An Effective Regimen in Relapsed and Refractory
- Acute Leukemia. in *Acute Leukemias* (eds. Hiddemann, W. et al.) vol. 34, 404–411 (Springer, Berlin, Heidelberg, 1992).
- 1047 131. Maschmeyer, G., Willborn, K. & Heit, W. Use of AMSA Combination Chemotherapy in Patients with Acute Myelogenous
- Leukemia Unsuitable for Anthracycline or Mitoxantrone Treatment. in *Acute Leukemias* (eds. Hiddemann, W. et al.) vol. 34, 445–
- 1049 449 (Springer, Berlin, Heidelberg, 1992).
- 1050 132. Freund, M. et al. Treatment of High-Risk Relapsing or Refractory AML with M-AMSA and ID-ARAC. in Acute Leukemias
- 1051 (eds. Hiddemann, W. et al.) vol. 34, 450–457 (Springer Berlin Heidelberg, Berlin, Heidelberg, 1992).
- 1052 133. Liu, L. et al. Increasing aclarubicin dose in low-dose cytarabine and aclarubicin in combination with granulocyte colony-
- stimulating factor (CAG regimen) is efficacious as salvage chemotherapy for relapsed/refractory mixed-phenotype acute leukemia.
- 1054 Leuk. Res. 39, 805–811 (2015).
- 1055 134. Zhu, J.-F. et al. Efficacy and safety of decitabine combined with HAAG (homoharringtonine, aclarubicin, low-dose cytarabine
- and G-CSF) for newly diagnosed acute myeloid leukemia. Front. Oncol. 12, (2022).
- 1057 135. Biomedical Research Column on Zhihu website for bioavailability (80%–85%) of drugs after intraperitoneal injection.
- https://zhuanlan.zhihu.com/p/347844560

136. Al Shoyaib, A., Archie, S. R. & Karamyan, V. T. Intraperitoneal route of drug administration: Should it be used in experimental animal studies? *Pharm. Res.* **37**, 12 (2019).