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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our Editorial Policies and the Editorial Policy Checklist.

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a	Confirmed	
☐	☒	The exact sample size (*n*) for each experimental group/condition, given as a discrete number and unit of measurement
☐	☒	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
☐	☒	The statistical test(s) used AND whether they are one- or two-sided *Only common tests should be described solely by name; describe more complex techniques in the Methods section.*
☐	☒	A description of all covariates tested
☐	☒	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
☐	☒	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
☐	☒	For null hypothesis testing, the test statistic (e.g. *F*, *t*, *r*) with confidence intervals, effect sizes, degrees of freedom and *P* value noted *Give P values as exact values whenever suitable.*
☒	☐	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
☒	☐	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
☐	☒	Estimates of effect sizes (e.g. Cohen's *d*, Pearson's *r*), indicating how they were calculated

Our web collection on statistics for biologists contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection	No special software was used in this study.
Data analysis	-For mRNA mapping, bowtie2.3.1 and Tophat 2.1.1 were used -For mRNA fpkm calculation Cufflinks 2.1.1 was used -For proteome and phsophoproteome analysis MaxQuant 1.4.0.8 was used -Other customized codes or scripts were uploaded to github repository @ https://github.com/bioexplore/yeastMultiomicsPaperCodes.git

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

RNA-seq raw data was uploaded to NCBI's SRA with the accessing code: PRJNA523289;
Proteome and Phosphoproteome raw data was uploaded to ProteomeXchange's PRIDE Archive with project accession code: PXD012891.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

[✗] Life sciences   [ ] Behavioural & social sciences   [ ] Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	To study how does yeast cell allocate its proteome resources across different specific growth rates, here we chosen nine different specific growth rates and for each specific growth rate we collect one sample. The chosen specific growth rates covered both the almost highest specific growth rate of yeast (0.4 h-1) under carbon-limitation condition and the lowest specific growth rate (0.025 h-1) in reported literature for the same strain used in this work. Additionally, the nine chosen growth rates covered both purely respiratory and respire-fermentative, the two distinct phenotype for S. cerevisiae that is Crabtree positive strain. Based on this, we think we have used the sufficient sample size for supporting conclusions of this study.
Data exclusions	No data exclusion in this study.
Replication	We used triplicate samples that were taken from three distinct bioreactors run under the same condition.
Randomization	This is not relevant, as the same strain was used in this work.
Blinding	This is not relevant, as the same strain was used in this work.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study
✗	Antibodies
✗	Eukaryotic cell lines
✗	Palaeontology and archaeology
✗	Animals and other organisms
✗	Human research participants
✗	Clinical data
✗	Dual use research of concern

Methods

n/a	Involved in the study
✗	ChIP-seq
✗	Flow cytometry
✗	MRI-based neuroimaging