

Polyanionic Receptors for Carboxylates in Water

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1. Synthesis and Characterisation

1.1 General

Commercial reagents were purchased from Sigma–Aldrich, Alfa Aesar or Acros Organics and were used without further purification unless otherwise specified. All air and moisture sensitive manipulations were carried out using standard vacuum line and Schlenk techniques. Solvents for air and moisture sensitive manipulations were obtained from an Anhydrous Engineering Solvent Purification System, distilled and dried over activated molecular sieves, or purchased from Acros Organics.

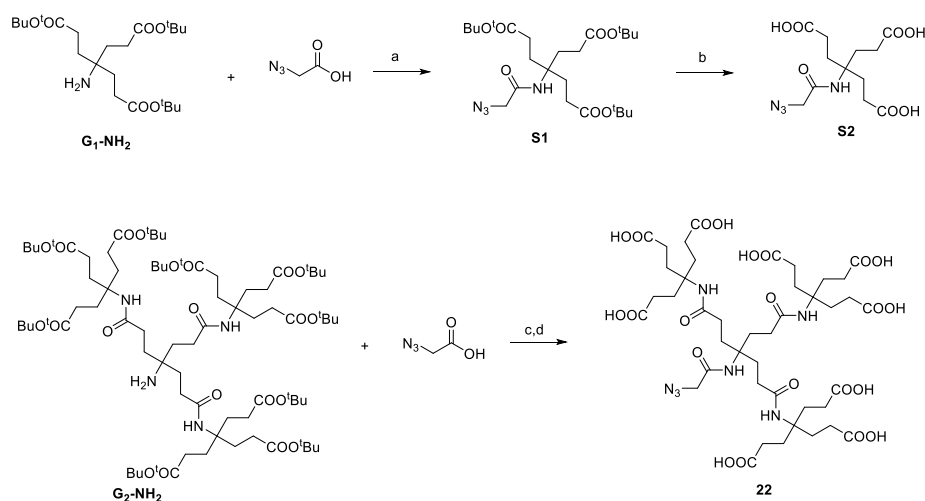
Flash column chromatography was performed on a Biotage® Selekt System using silica (Biotage® Sfär Silica D - 60 µm) or C18 (Biotage® Sfär C18 D - Duo 100 Å 30 µm) columns and a suitable eluent. TLC was performed using aluminium backed TLC plates (Merck-Keisegel 60 F254) and visualised using UV fluorescence (254 or 365 nm) and/or developed using ninhydrin, potassium permanganate or bromocresol green.

NMR spectra were recorded on Varian VNMR 400 MHz, Bruker 400 MHz, Varian VNMRS 500 MHz, Bruker Advance III HD Cryo 500 MHz, Bruker Neo Cryo 600 MHz and Bruker Cryo 700 MHz spectrometers. All spectra were obtained at 298 K. All ¹H and ¹³C NMR chemical shifts are reported relative to the ¹H and ¹³C chemical shifts of the solvent as standard. LRMS (low resolution mass spectrometry) was performed on a Waters 600 Controller with a Waters SQ Detector 2. HRMS (high resolution mass spectrometry) was performed on a Thermo Scientific Orbitrap Elite or a Waters Synapt G2S.

The following starting materials were obtained commercially; diamino acid building blocks Boc-Dap(Z)-OH (**11**), Boc-Dab(Z)-OH (**12**), Fmoc-Dab(Boc)-OH (**S12**) and 1,4-Bis(4-aminophenoxy)benzene (**13**). First and second generation dendritic amine **G₁-NH₂** and **G₂-NH₂** (see Supplementary Scheme 1) were synthesized following the previously reported procedure.¹ The terms G₁ and G₂ are hereafter applied in building blocks or macrocycles which derived from these components, as in “G₁ azido triacid” (**S2**) and “isophthaloyl G₂ tricycle” (**5**). Receptors with protonated COOH side chains are named with ‘-H’ suffixes, as in “dipicolinoyl G₂-acid tricycle” (**6-H**).

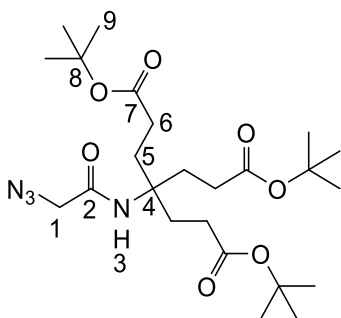
All synthesized molecules were characterized by ¹H and ¹³C NMR spectroscopy in organic solvents to confirm their structure. Macrocycles were further characterized in their operating environments, i.e. DMSO-*d*₆ for receptors **8a**, **9a**, **10a**, **20**, **21**, and D₂O or 9:1 H₂O/D₂O for receptors **8b**, **9b**, **10b**, **5**, **6**.

1.2 Synthesis of azido solubilising groups (**S2** and **22**)



Supplementary Scheme 1. The synthesis of azido solubilising groups **S2** and **22**. a) DIPEA, HBTU, DMF; b) TFA, DCM; c) DIPEA, HBTU, DMF; d) TFA, DCM.

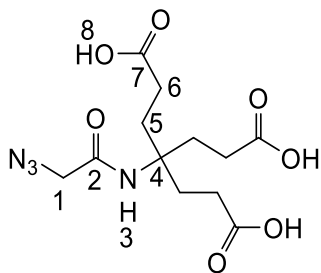
G₁ azido tri-ester (**S1**)



To a solution of the **G₁-NH₂** (400 mg, 0.75 mmol, 1.0 eqv) in THF (20 ml) was added 2-azidoacetic acid (146 mg, 1.44 mmol, 1.5 eqv), HBTU (548 mg, 1.44 mmol, 1.5 eqv) and DIPEA (335 μ L, 1.93 mmol, 2.0 eqv). The mixture was allowed to react at room temperature for 12 hours before removing the solvent *in vacuo*. The residue was purified by column chromatography (EtOAc:hexane = 65:35). The solvent was removed *in vacuo* to give the title compound as a white solid (320 mg, 0.63 mmol, 65 %).

¹H NMR (500 MHz, CDCl₃) δ 6.46 (s, 1H, N3H), 3.86 (s, 2H, C1H), 2.20 (t, J = 5.0 Hz, 6H, C6H), 1.98 (t, J = 4.9 Hz, 6H, C5H), 1.43 (s, 27H, C9H). **¹³C NMR** (126 MHz, CDCl₃) δ 172.7 (C2), 166.0 (C7), 80.9 (C8), 58.0 (C4), 53.1 (C1), 30.0 (C5), 29.8 (C6), 28.2 (C9). The NMR spectra are in accordance with the literature².

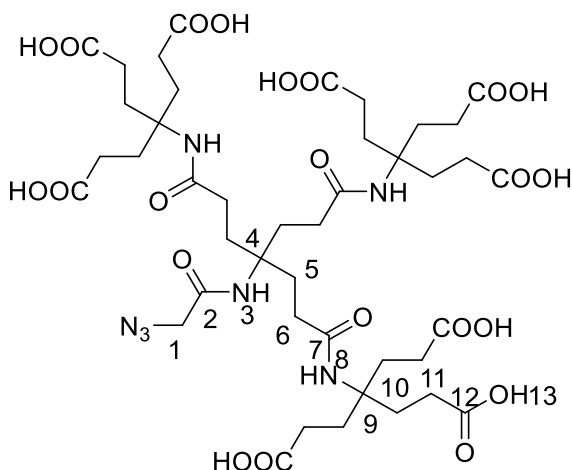
G₁ azido tri-acid (S2)



To a solution of the G₁ azido tri-ester **S1** (50 mg, 0.1 mol, 1.0 eqv) in dry DCM (2 ml) solution was added TFA (1 ml), and the mixture was stirred at 0 °C for 30 min. The solvent was then removed under N₂ flow to give the title compound as a white solid (33 mg, 99 %).

¹H NMR (500 MHz, DMSO-*d*₆) δ 7.44 (s, 1H, N3H), 3.75 (s, 2H, C1H), 2.18 – 2.09 (m, 6H, C6H), 1.89 – 1.79 (m, 6H, C5H). **¹³C NMR** (126 MHz, DMSO-*d*₆) δ 174.3 (C2), 166.9 (C7), 56.9 (C4), 50.8 (C1), 29.0 (C6), 28.0 (C5). **HRMS** for C₁₂H₁₇N₄O₇ [M-H]⁻ Calculated m/z = 329.1097 Found m/z = 329.1090

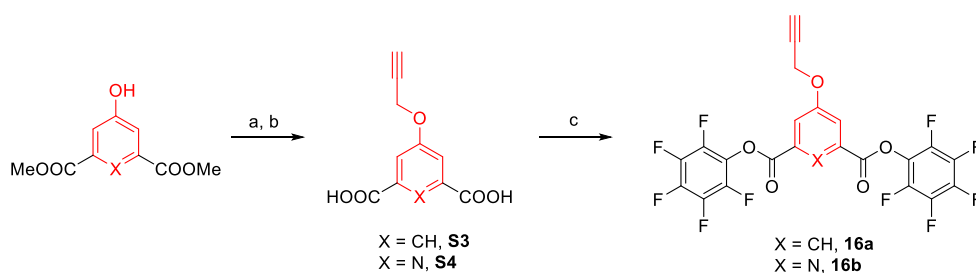
G₂ azido nona-acid (22)



To a solution of **G₂-NH₂** (600 mg, 0.42 mmol, 1.0 eqv) in THF (20 ml) was added 2-azidoacetic acid (63 mg, 0.63 mmol, 1.5 eqv), HBTU (320 mg, 0.63 mmol, 1.5 eqv) and DIPEA (145 μL, 0.83 mmol, 2.0 eqv). The reaction mixture was stirred at room temperature for 12 hours before removing the solvent *in vacuo*. The residue was then purified by column chromatography (EtOAc = 100 %) to give the intermediate nona-ester as a white solid. The solid was stirred for overnight in 25 % TFA DCM solution. Volatiles were then removed under N₂ flow to give the title compound as a white solid (240 mg, 0.24 mmol, 56 %).

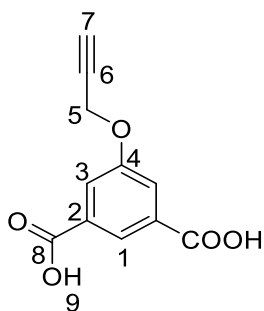
¹H NMR (500 MHz, DMSO) δ 12.04 (s, 9H, O13H), 7.48 (s, 1H, N3H), 7.20 (s, 3H, N8), 3.74 (s, 2H, C1H), 2.15 – 2.06 (m, 18H, C11H), 2.06 – 1.99 (m, 6H, C6H), 1.88 – 1.72 (m, 24H, C5,10H). **¹³C NMR** (126 MHz, DMSO) δ 174.9 (C12), 172.5 (C7), 161.4 (C2), 74.7 (C4), 56.8 (C1), 50.9 (C9), 31.2 (C6), 30.5 (C5), 29.5 (C11), 28.5 (C10). **HRMS** for C₄₂H₆₂N₇O₂₂ [M-H]⁻ Calculated m/z = 1016.3948 Found m/z = 1016.3965.

1.3 Synthesis of activated linker reagents (16a and 16b)



Supplementary Scheme 2. The synthesis of activated ester linkers. a) Propargyl bromide, K_2CO_3 , MeCN; b) NaOH, MeOH; c) DCC, THF.

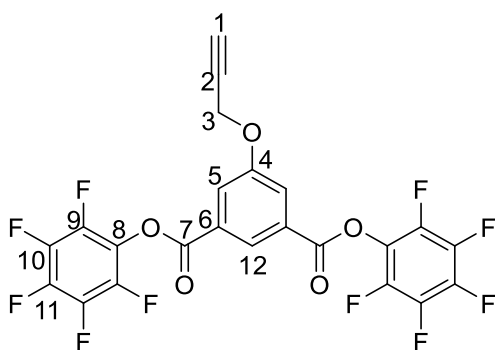
5-propargyloxy isophthalic acid (S3)



Dimethyl 5-(hydroxy)isophthalate (10 g, 54.9 mmol, 1.0 eqv) and potassium carbonate (15 g, 108 mmol, 2.0 eqv) were suspended in acetonitrile (300 mL). Propargyl bromide (80% in PhMe, 5.40 mL, 56.0 mmol, 1.02 eqv) was then added and the reaction refluxed overnight. The solution was then filtered and concentrated under reduced pressure to give a solid which was then dissolved in methanol (150 mL). Sodium hydroxide (5.00 g, 125 mmol, 2.3 eqv) was added to the solution and the mixture was stirred for 2 hours. The solution was then concentrated under reduced pressure. The residue was dissolved in water (100 mL) then acidified to pH 2-3 with aqueous HCl (1 M) and the resulting precipitate collected by filtration. The solid was washed with water and then dried under reduced pressure to give the pure title compound as a colourless solid (10.7 g, 47.8 mmol, 89 %).

1H NMR (500 MHz, $DMSO-d_6$) δ 13.34 (s, 2H, O9H), 8.11 (t, J = 1.4 Hz, 1H, C1H), 7.72 (d, J = 1.4 Hz, 2H, C3H), 4.95 (d, J = 2.4 Hz, 2H, C5H), 3.64 (t, J = 2.3 Hz, 1H, C7H). **^{13}C NMR** (126 MHz, $DMSO-d_6$) δ 166.3 (C8), 157.3 (C4), 132.6 (C2), 122.9 (C1), 119.6 (C3), 79.0 (C6), 78.7 (C7), 56.0 (C5). These data are in accordance with the literature³.

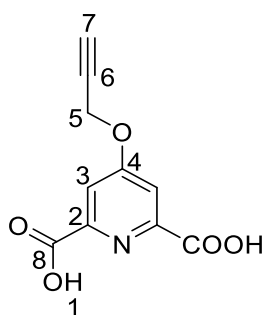
Bis(pentafluorophenyl) 5-(propargyloxy)isophthalate (16a)



DCC (9.80 g, 47.6 mmol, 2.1 eqv) and pentafluorophenol (10.87 g, 54.8 mmol, 2.4 eqv) were dissolved in anhydrous THF (500 mL) and 5-(propargyloxy)isophthalic acid **S3** (5 g, 22.7 mmol, 1.0 eqv) was added in small portions. The reaction was stirred under an inert atmosphere for 18 hours. The solvent was concentrated under reduced pressure, the resultant solid was suspended in diethyl ether and the solid removed by filtration and washed thoroughly with diethyl ether. The filtrate was concentrated under reduced pressure and the crude residue was recrystallized from hot hexane to give the title compound as a colourless solid (9.10 g, 17.9 mmol, 79%).

¹H NMR (400 MHz, CDCl₃) δ 8.64 (t, *J* = 1.5 Hz, 1H, C12H), 8.08 (d, *J* = 1.4 Hz, 2H, C5H), 4.86 (d, *J* = 2.36 Hz, 2H, C3H), 2.60 (t, *J* = 2.53 Hz, 4H, C1H). **¹³C NMR** (101 MHz, CDCl₃) δ 161.31 (C7), 158.2 (C4), 142.2 (C8), 140.5 (C9), 139.0 (C11), 137.3 (C10), 128.8 (C6), 125.7 (C12), 123.4 (C5), 79.9 (C2), 78.8 (C1), 56.6 (C3). **HRMS** for C₂₃H₇F₁₀O₅ [M+H]⁺ Calculated *m/z* = 553.0119 Found *m/z* = 553.0131.

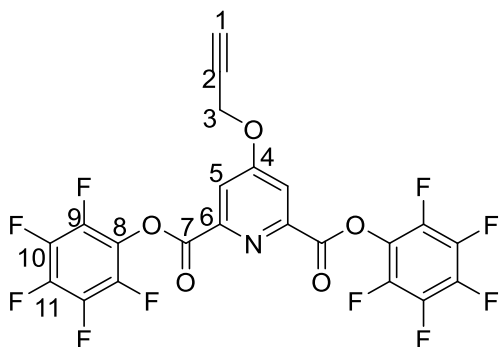
4-(propargyloxy)pyridine-2,6-dicarboxylic acid (S4)



Dimethyl 4-hydroxypyridine-2,6-dicarboxylate (5 g, 27.5 mmol, 1.0 eqv) and potassium carbonate (7.5 g, 54 mmol, 2.0 eqv) were suspended in acetonitrile (100 mL). Propargyl bromide (80% in PhMe, 2.70 mL, 28 mmol, 1.02 eqv) was then added and the reaction refluxed overnight. The solution was then filtered and concentrated under reduced pressure to give a solid which was dissolved in methanol (150 mL). Sodium hydroxide (2.5 g, 62 mmol, 2.3 eqv) was added and the solution stirred for 2 hours. The solution was then concentrated under reduced pressure. The residue was dissolved in water (100 mL) then acidified to pH 2-3 with aqueous HCl (1M) and the resulting precipitate collected by filtration. The solid was washed with water and then dried under reduced pressure to give the title compound as a colourless solid (5.4 g, 23.9 mmol, 87 %).

¹H NMR (500 MHz, DMSO-*d*₆) δ 7.78 (s, 2H, C3H), 5.10 (d, *J* = 2.3 Hz, 2H, C5H), 3.76 (t, *J* = 2.3 Hz, 1H, C7H). **¹³C NMR** (126 MHz, DMSO-*d*₆) δ 165.2 (C8), 149.8 (C3), 114.0 (C3), 79.8 (C6), 77.7 (C7), 56.4 (C5). These data are in accordance with the literature⁴.

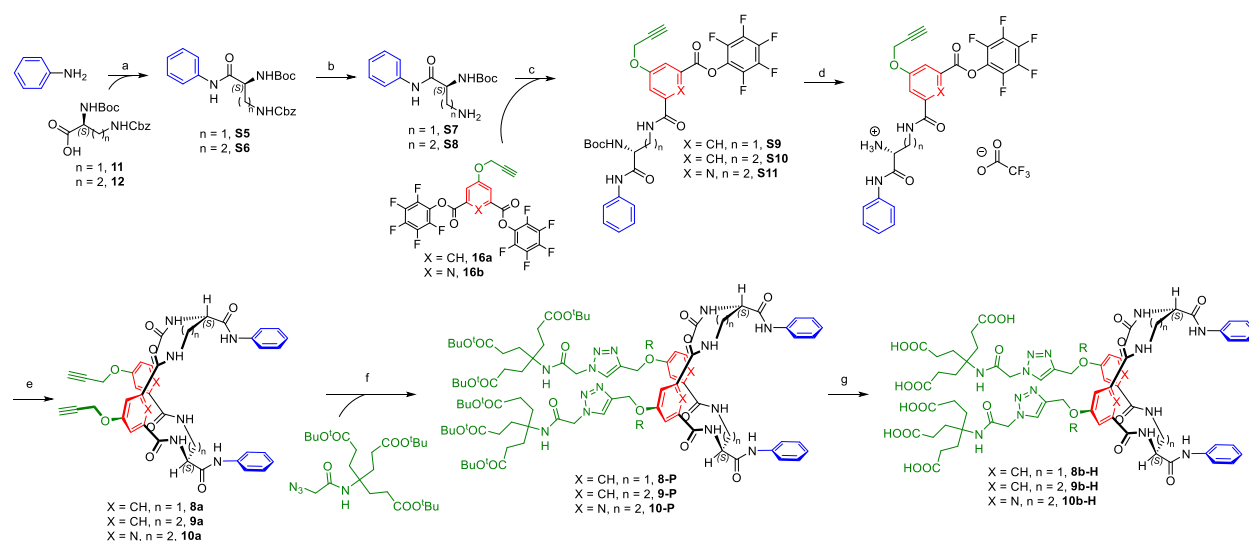
Bis(pentafluorophenyl) 4-(propargyloxy)pyridine-2,6-dicarboxylate (16b)



DCC (4.9 g, 23.8 mmol, 2.1 eqv) and pentafluorophenol (5.4 g, 27.4 mmol, 2.4 eqv) were dissolved in anhydrous THF (200 mL) and 4-(propargyloxy)pyridine-2,6-dicarboxylic acid **S4** (2.5 g, 11.4 mmol, 1.0 eqv) was added in small portions. The reaction was stirred under an inert atmosphere for 18 hours. The solution was evaporated under reduced pressure, the resultant solid was suspended in diethyl ether and the solid removed by filtration and washed thoroughly with diethyl ether. The filtrate was concentrated under reduced pressure and the crude residue was recrystallized from hot hexane to give the title compound as a colourless solid (3.8 g, 7.5 mmol, 66 %).

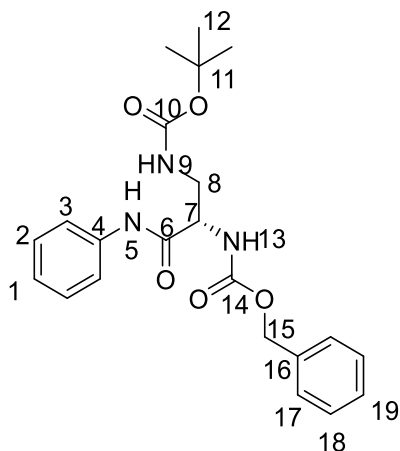
¹H NMR (500 MHz, CDCl₃) δ 8.12 (s, 2H, C5H), 4.96 (d, *J* = 2.4 Hz, 2H, C3H), 2.68 (s, 1H, C1H). **¹³C NMR** (126 MHz, CDCl₃) δ 166.0 (C7), 160.6 (C4), 147.8 (C6), 117.4 (C5), 78.5 (C2), 75.7 (C1), 57.0 (C3). **HRMS** for C₂₂H₆F₁₀NO₅ [M+H]⁺ Calculated *m/z* = 554.0081 Found *m/z* = 554.0077.

1.4 Synthesis of receptors 8a, 8b, 9a, 9b 10a and 10b



Supplementary Scheme 3. Synthesis of receptors **8a**, **9a**, **10a**, **8b-H**, **9b-H** and **10b-H**: a) HBTU, TEA, CH₂Cl₂; b) Pd/C, MeOH, H₂; c) DIPEA, THF; d) TFA, CH₂Cl₂; e) DIPEA, TBACl, THF; f) [Cu(CH₃CN)₄]PF₆, 2,6-lutidine, MeCN; g) TFA, CH₂Cl₂.

Benzyl tert-butyl (3-oxo-3-(phenylamino)propane-1,2-diyl)(S)-dicarbamate (S5)

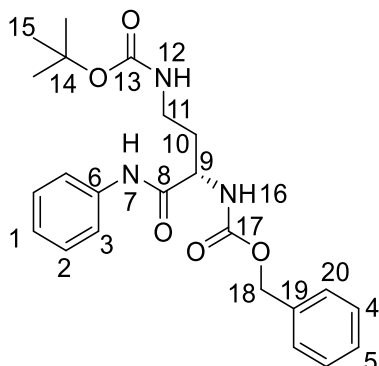


To a solution of commercially available Boc-Dap(Z)-OH **11** (500 mg, 1.48 mmol, 1.0 eqv) in DCM (15 ml) was added TEA (412 μ L, 2.96 mmol, 2.0 eqv) and HBTU (672 mg, 1.77 mmol, 1.2 eqv). The mixture was stirred at room temperature for 20 minutes before adding aniline (202 μ L, 2.22 mmol, 1.5 eqv). The reaction mixture was stirred at room temperature for overnight, then washed with 0.1 M aqueous HCl (2 x 10 ml), followed by brine (1 x 10 ml) and then dried over MgSO₄. The solvent was removed *in vacuo*, and the residue was purified by column chromatography (EtOAc:MeOH = 1:9) to give the title compound (580 mg, 1.4 mmol, 95 %) as a white solid.

¹H NMR (500 MHz, DMSO-*d*₆) δ 9.96 (s, 1H, N5H), 7.61 – 7.55 (m, 2H, C3H), 7.36 – 7.26 (m, 8H, C2,17,18,19,N9H), 7.05 (m, 1H, C1H), 6.89 (d, *J* = 7.9 Hz, 1H, N13H), 5.05 – 4.95 (m, 2H, C15H), 4.19 (q, *J* = 6.7 Hz, 1H, C7H), 3.43 – 3.37 (m, 2H, C8H), 1.38 (s, 9H, C12H). **¹³C NMR** (126 MHz, DMSO-*d*₆) δ 169.0 (C6),

156.3 (C14), 155.2 (C10), 138.9 (C4), 137.1 (C16), 128.6 (C2), 128.3 (C17), 127.7 (C18), 127.6 (C19), 123.3 (C1), 119.5 (C3), 78.4 (C11), 65.3 (C15), 55.3 (C7), 42.2 (C8), 28.2 (C12). **HRMS** for $C_{22}H_{27}N_3O_5Na$ $[M+Na]^+$ Calculated m/z = 436.1848 Found m/z = 436.1869.

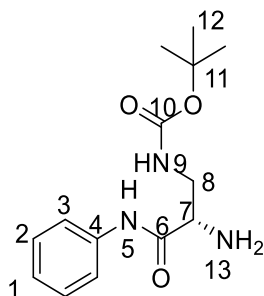
Benzyl *tert*-butyl (4-oxo-4-(phenylamino)butane-1,3-diyl)(*S*)-dicarbamate (S6)



To a solution of commercially available Boc-Dab(Z)-OH **12** (1.0 g, 2.84 mmol, 1.0 eqv) in DCM (15 ml) was added TEA (790 μ L, 5.68 mmol, 2.0 eqv) and HBTU (1294 mg, 3.41 mmol, 1.2 eqv). The above mixture was stirred at room temperature for 20 minutes before adding aniline (396 mg, 4.26 mmol, 1.5 eqv). The reaction mixture was stirred at room temperature overnight, then washed with 0.1 M aqueous HCl (2 x 10 ml), followed by brine (1 x 10 ml) and then dried over $MgSO_4$. The solvent was removed *in vacuo*, and the residue was then purified by reverse phase column chromatography (H_2O :acetone = 20:80) to give the title compound (680 mg, 1.59 mmol, 56%) as a white solid.

1H NMR (400 MHz, $DMSO-d_6$) δ 9.98 (s, 1H, N7H), 7.60 – 7.58 (m, 3H, N16H, C3H), 7.39 – 7.26 (m, 7H, C2,4,5,20H), 7.10 – 7.01 (m, 1H, C1H), 6.74 (t, J = 5.1, 1H, N12H), 5.03 (s, 2H, C18H), 4.17 (q, J = 7.8 Hz, 1H, C9H), 3.03 – 2.98 (m, 2H, C10H), 1.86 – 1.69 (m, 2H, C10H), 1.37 (s, 9H, C15H). **^{13}C NMR** (101 MHz, $DMSO-d_6$) δ 170.6 (C8), 156.0 (C13), 155.5 (C17), 138.8 (C6), 136.9 (C19), 128.7 (C4), 128.3 (C20), 127.8 (C5), 127.7 (C2), 123.4 (C1), 119.2 (C3), 77.7 (C14), 65.5 (C18), 53.4 (C9), 37.0 (C10), 32.0 (C11), 28.2 (C15). **HRMS** for $C_{23}H_{29}N_3O_5Na$ $[M+Na]^+$ Calculated m/z = 450.1999 Found m/z = 450.2003.

***Tert*-butyl (*S*)-(2-amino-3-oxo-3-(phenylamino)propyl)carbamate (S7)**

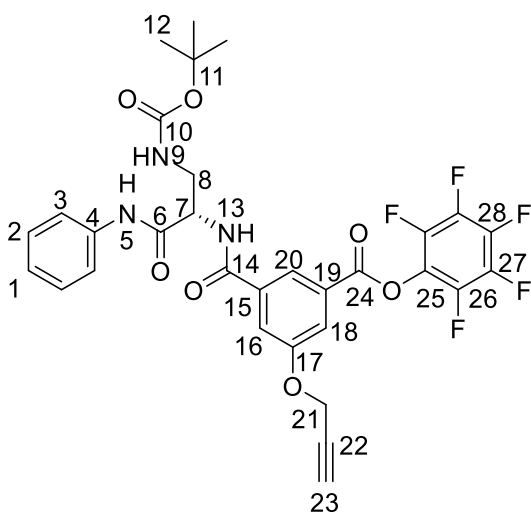


Benzyl *tert*-butyl (3-oxo-3-(phenylamino)propane-1,2-diyl)(*S*)-dicarbamate **S5** (500 mg, 1.21 mmol, 1.0 eqv) and 5 % Pd/C (129 mg, 0.02 mmol, 0.05 eqv) were placed in a three-neck flask, which was then evacuated and backfilled with N_2 3 times. MeOH (20 mL) was added, then the flask was evacuated and backfilled with H_2 3 times. The mixture was allowed to react at room temperature for 12 hours before exchanging H_2 with N_2 . The

reaction mixture was filtered under a N₂ atmosphere through celite and the cake was washed with DCM (30 ml). The solvent was removed *in vacuo* to give the title compound (321 mg, 1.15 mmol, 95 %) as a colourless oil.

¹H NMR (500 MHz, CDCl₃) δ 9.74 (s, 1H, C5H), 7.54 (d, *J* = 7.2 Hz, 2H, C3H), 7.33 – 7.25 (m, 2H, C2H), 7.08 (t, *J* = 7.4 Hz, 1H, C1H), 6.05 (s, 2H, C13H), 4.40 – 4.31 (m, 1H, C7H), 3.33 – 3.25 (m, 1H, C8H), 3.13 – 3.07 (m, 1H, C8H), 1.44 (s, 9H, C12). **¹³C NMR** (126 MHz, CDCl₃) δ 168.8 (C6), 156.7 (C10), 137.8 (C4), 129.1 (C2), 124.6 (C1), 120.2 (C3), 80.9 (C11), 54.6 (C7), 43.8 (C9), 28.4 (C12). **HRMS** for C₁₄H₂₂N₃O₃ [M+H]⁺ Calculated *m/z* = 280.1661 Found *m/z* = 280.1665.

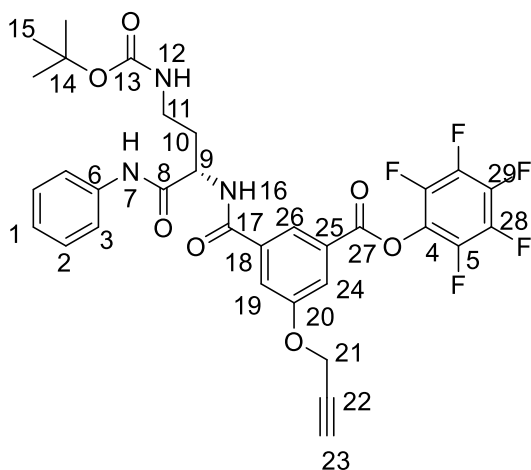
Boc-Dap phenylamino PFP benzoate (S9)



To a solution of bis(pentafluorophenyl) 5-(propargyloxy)isophthalate **16a** (988 mg, 1.79 mmol, 2.5 eqv) in THF (10 ml) was added DIPEA (623 μL, 3.58 mmol, 5.0 eqv). A solution of *tert*-butyl (S)-(2-amino-3-oxo-3-(phenylamino)propyl)carbamate **S7** (200 mg, 716 μmol, 1.0 eqv) in THF (10 ml) was added over 18 hours by a syringe pump. The reaction mixture was allowed to stirred at room temperature for 6 hours after the completion of the addition. The solvent was removed *in vacuo*, and the residue was then purified by column chromatography (hexane:EtOAc = 50:50) to give the title compound (301 mg, 0.46 mmol, 65 %) as a white solid.

¹H NMR (600 MHz, CDCl₃) δ 9.21 (s, 1H, C5H), 8.20 (t, *J* = 1.5 Hz, 1H, C20H), 7.86 – 7.81 (m, 2H, C16H, N9H), 7.76 - 7.70 (m, 1H, C18H), 7.49 – 7.42 (m, 2H, C3H), 7.24 – 7.20 (m, 2H, C2H), 7.09 – 7.03 (m, 1H, C1H), 6.27 (s, 1H, N13H), 4.72 (s, 1H, C21H), 4.59 (s, 1H, C7H) 4.05 – 3.97 (m, 1H, C8H), 3.84 – 3.76 (m, 1H, C8H), 2.53 (t, *J* = 2.4 Hz, 1H, C23H), 1.42 (s, 9H). **¹³C NMR** (151 MHz, CDCl₃) δ 169.1 (C6), 167.4 (C14), 161.7 (C24), 158.1 (C10), 156.9 (C17), 142.1 (25), 140.6 (C26), 139.0 (C27), 137.4 (C4), 137.2 (C28), 136.1 (C15), 129.1 (C2), 128.7 (C19), 124.9 (C1), 122.2 (C20), 120.4 (C3,16), 120.0 (C18), 81.1 (C11), 77.4 (C22), 76.8 (C23), 56.4 (C21), 56.0 (C7), 43.1 (C8), 28.3 (C12). **HRMS** for C₃₁H₂₇F₅N₃O₇ [M+H]⁺ Calculated *m/z* = 648.1769 Found *m/z* = 648.1757.

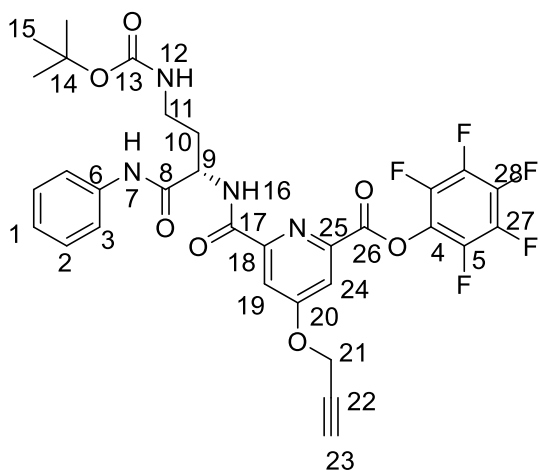
Boc-Dab phenylamino PFP benzoate (S10)



Benzyl *tert*-butyl (4-oxo-4-(phenylamino)butane-1,3-diyl)(*S*)-dicarbamate **S6** (700 mg, 1.17 mmol, 1.0 eqv) and 5 % Pd/C (227 mg, 0.12 mmol mmol, 0.1 eqv) were placed in a three-neck flask, which was then evacuated and backfilled with N₂ 3 times. MeOH (20 mL) was added, then the flask was evacuated and backfilled with H₂ 3 times. The mixture was allowed to react at room temperature for 12 hours before exchanging H₂ with N₂. The reaction mixture was filtered under a N₂ atmosphere through celite and the cake was washed with DCM (30 ml). The solvent was removed *in vacuo* to give the intermediate compound **S8** as a colourless oil. To a solution of bis(pentafluorophenyl) 5-(propargyloxy)isophthalate **16a** (1.61 g, 2.92 mmol, 2.5 eqv) in THF (10 ml) was added DIPEA (1.02 mL, 5.85 mmol, 5.0 eqv). A solution of the intermediate **S8** in THF (10 ml) was added to the above solution over 18 hours by a syringe pump. The mixture was allowed to react at room temperature for 6 hours after the completion of the addition. The solvent was removed *in vacuo*, and the residue was then purified by column chromatography (hexane:EtOAc = 40:60) to give the title compound (619 mg, 0.94 mmol, 80 %) as a white solid.

¹H NMR (600 MHz, CDCl₃) δ 10.03 (s, 1H, N7H), 8.30 (s, 1H, C24H), 7.90 (s, 1H, C26H), 7.82 (m, 2H, C19H, N16H), 7.65 (d, *J* = 8.0 Hz, 2H, C3H), 7.33 (t, *J* = 7.8 Hz, 2H, C2H), 7.12 (t, *J* = 7.3 Hz 1H), 4.86 – 4.82 (m, 1H, C9H), 4.81 (d, *J* = 2.4 Hz, 2H, C21H), 3.66 -3.61 (m, 1H, C11H), 3.15 -3.11 (m, 1H, C11H), 2.57 (t, *J* = 2.6 Hz, C23H), 2.19 – 2.13 (m, 1H, C10H), 2.06 – 1.98 (m, 1H, C10H), 1.47 (s, 9H, C15H). **¹³C NMR** (151 MHz, CDCl₃) δ 169.3 (C8), 165.2 (C17), 161.8 (C27), 158.2 (C13,20), 142.3 (C4), 140.6 (C28), 138.9 (C5), 138.0 (C6), 137.3 (C29), 136.3 (C18), 129.1 (C2), 128.8 (C25), 124.6 (C1), 122.1 (C24), 120.3 (C19), 120.2 (C26), 120.0 (C3), 80.8 (C14), 77.4 (C23), 76.8 (C22), 56.5 (C21), 51.7 (C9), 37.2 (C11), 35.1 (C10), 28.5 (C15). **HRMS** for C₃₂H₂₉F₅N₃O₇ [M+H]⁺ Calculated *m/z* = 662.1920 Found *m/z* = 662.1914.

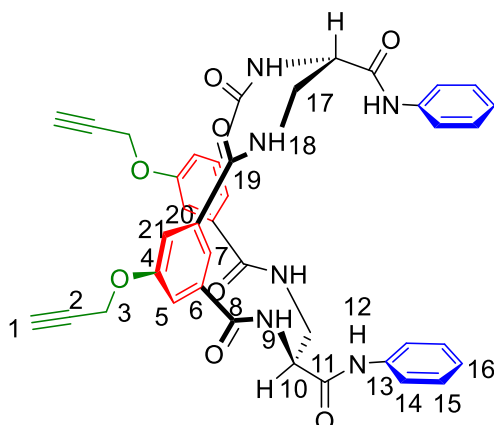
Boc-Dab phenylamino PFP picolinate (S11)



Benzyl *tert*-butyl (4-oxo-4-(phenylamino)butane-1,3-diyl)(*S*)-dicarbamate **S6** (700 mg, 1.17 mmol, 1.0 eqv) and 5 % Pd/C (227 mg, 0.12 mmol mmol, 0.1 eqv) were placed in a three-neck flask, which was then evacuated and backfilled with N₂ 3 times. MeOH (20 mL) was added under N₂ atmosphere, then the flask was evacuated and backfilled with H₂ 3 times. The mixture was allowed to react at room temperature for 12 hours before exchanging H₂ with N₂. The reaction mixture was filtered under a N₂ atmosphere through celite and the cake was washed with DCM (30 ml). The solvent was removed *in vacuo* to give the intermediate compound **S8** as a colourless oil. To a solution of bis(pentafluorophenyl) pyridine-2,6-dicarboxylate (**16b**) (1.61 g, 2.92 mmol, 2.5 eqv) in THF (10 ml) was added DIPEA (1.02 mL, 5.85 mmol, 5.0 eqv). A solution of the intermediate **S8** in THF (10 ml) was added to the above solution over 18 hours by a syringe pump. The mixture was allowed to react at room temperature for 6 hours after the completion of the addition. The solvent was removed *in vacuo*, and the residue was then purified by column chromatography (hexane:EtOAc = 47:53) to give the title compound (317 mg, 0.48 mmol, 41 %) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 9.86 (s, 1H, N7H), 8.89 (s, 1H, N16H), 8.04 (d, *J* = 2.4 Hz, 1H, C19H), 7.95 (d, *J* = 2.5 Hz, 1H, C24H), 7.68 – 7.63 (m, 2H, C3H), 7.30 – 7.33 (m, 2H, C2H), 7.13 – 7.06 (m, 1H, C1H), 5.21 (s, 1H, N12H), 4.88 (d, *J* = 2.4 Hz, 2H, C21H), 3.60 - 3.11 (m, 2H, C11H), 2.62 (t, *J* = 2.4 Hz, 1H, C23H), 2.14 - 2.06 (m, 2H, C10H), 1.45 (s, 9H, C15H). **¹³C NMR** (126 MHz, CDCl₃) δ 168.9 (C8), 166.2 (C17), 163.0 (C26), 160.7 (C13), 157.8 (C20), 152.3 (C18), 145.9 (C25), 142.3 (C4), 140.9 (C27), 139.1 (C5), 138.1 (C6), 137.1 (C28), 129.1 (C2), 124.5 (C1), 120.0 (C3), 116.7 (C24), 112.4 (C19), 80.5 (C14), 77.9 (C23), 76.2 (C22), 56.7 (C21), 51.6 (C9), 37.4 (C11), 35.2 (C10), 28.5 (C15). **HRMS** for C₃₁H₂₈F₅N₄O₇ [M+H]⁺ Calculated *m/z* = 663.1873 Found *m/z* = 663.1876.

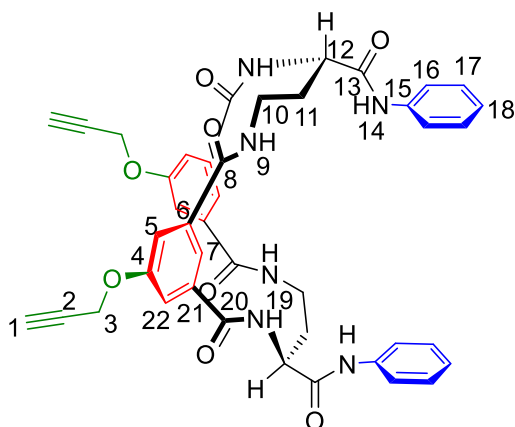
18-membered isophthaloyl macrocycle (8a)



To a solution of Boc-Dap phenylamino PFP benzoate **S9** (100 mg, 0.18 mmol, 1.0 eqv) in DCM (3 ml) was added TFA (1 ml). The reaction was stirred at room temperature for 2 hours before the volatiles were removed by N₂ flow. The residue was dissolved into THF (10 ml) and stored in a syringe for later use. Separately, a solution of DIPEA (317 μ L, 1.82 mmol, 10.0 eqv) and tetrabutylammonium chloride (253 mg, 0.91 mmol, 5.0 eqv) in THF (150 ml) was prepared in a round bottomed flask. The solution in the syringe was added slowly to the round bottomed flask at room temperature over 24 hours. The solution was then stirred at room temperature for 4 hours before the solvent was removed *in vacuo*. The residue was then purified by a combination of reverse (H₂O:acetone = 36:64) and normal phase (EtOAc:MeOH = 95:5) column chromatography to give the title compound (39 mg, 54 μ mol, 59%) as a white solid.

¹H NMR (600 MHz, DMSO-*d*₆) δ 10.23 (s, 1H, N12H), 9.00 (s, 1H, N9H), 8.52 (s, 1H, N18H), 8.15 (s, 1H, C7H), 7.66 – 7.61 (m, 2H, C14H), 7.53 (s, 2H, C5,21H), 7.36 – 7.30 (m, 2H, C15H), 7.11 – 7.05 (m, 1H, C16H), 4.91 (d, *J* = 2.2 Hz, 1H, C3H), 4.70 – 4.62 (m, 1H, C10H), 4.16 – 4.06 (m, 1H, C17H), 3.67 – 3.62 (m, 1H, C17H), 3.59 (t, *J* = 2.4 Hz, 1H, C1H). **¹³C NMR** (151 MHz, MeOD) δ 171.0 (C11), 170.2 (C8), 169.2 (C19), 159.6 (C4), 139.2 (C13), 137.2 (C6), 136.7 (C20), 129.8 (C15), 125.7 (C16), 121.9 (C14), 121.8 (C7), 118.5 (C5), 118.3 (C21), 79.0 (C1), 77.6 (C2), 58.5 (C10), 57.2 (C3), 41.7 (C17). **HRMS** for C₄₀H₃₄N₆O₈Na [M+Na]⁺ Calculated *m/z* = 749.2336 Found *m/z* = 749.2338.

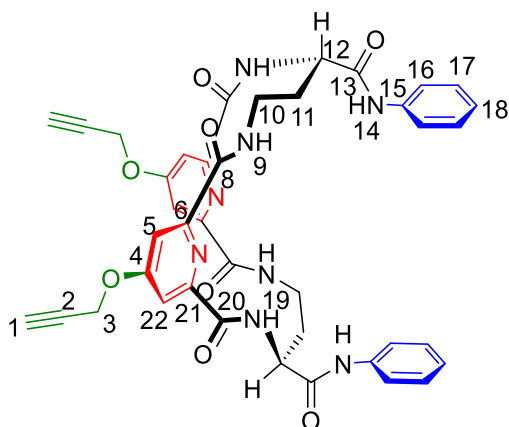
20-membered isophthaloyl macrocycle (9a)



To a solution of Boc-Dab phenylamino PFP benzoate **S10** (100 mg, 0.15 mmol, 1.0 eqv) in DCM (3 ml) was added TFA (1 ml). The reaction was stirred at room temperature for 2 hours before the volatiles were removed by N₂ flow. The residue was dissolved into THF (10 ml) and stored in a syringe for later use. Separately, a solution of DIPEA (262 μ L, 1.5 mmol, 10.0 eqv) and tetrabutylammonium chloride (210 mg, 0.76 mmol, 5.0 eqv) in THF (150 ml) was prepared in a round bottomed flask. The solution in the syringe was added slowly to the round bottomed flask at room temperature over 24 hours. The solution was then stirred at room temperature for 4 hours before the solvent was removed *in vacuo*. The residue was then purified by a combination of reverse (H₂O:acetone = 30:70) and normal phase (EtOAc:MeOH = 85:15) column chromatography to give the title compound (25 mg, 34 μ mol, 45%) as a white solid.

¹H NMR (600 MHz, DMSO-*d*₆) δ 10.07 (s, 1H, N14H), 8.87 (d, *J* = 7.1 Hz, 1H, N19H), 8.68 (t, *J* = 5.8 Hz, 1H, N9H), 8.30 (s, 1H, C7H), 7.62 (d, *J* = 8.2 Hz, 2H, C16H), 7.55 (d, *J* = 7.0, 2H, C5,22H), 7.31 (t, *J* = 7.7 Hz, 2H, C17H), 7.06 (t, *J* = 7.4, 1H, C18H), 4.86 (d, *J* = 2.6 Hz, H, C2H), 4.72 – 4.69 (m, 1H, C12H), 3.68 - 3.63 (m, 1H, C10H), 3.59 (t, *J* = 2.3 Hz, 1H), 3.51 – 3.45 (m, 1H, C10H), 2.32 – 2.12 (m, 2H, C11H). **¹³C NMR** (151 MHz, DMSO-*d*₆) δ 170.5 (C13), 165.3 (C20), 165.0 (C8), 156.9 (C4), 139.0 (C15), 135.3 (C6), 135.0 (C21), 128.7 (C17), 123.4 (C18), 119.4 (C7,16), 116.4 (C5), 116.2 (C22), 78.8 (C2), 78.6 (C1), 55.8 (C3), 54.3 (C12), 37.1 (C10), 31.0 (C11). **HRMS** for C₄₂H₃₉N₆O₈ [M+H]⁺ Calculated *m/z* = 755.2824 Found *m/z* = 755.2818.

20-membered dipicolinoyl macrocycle (10a)



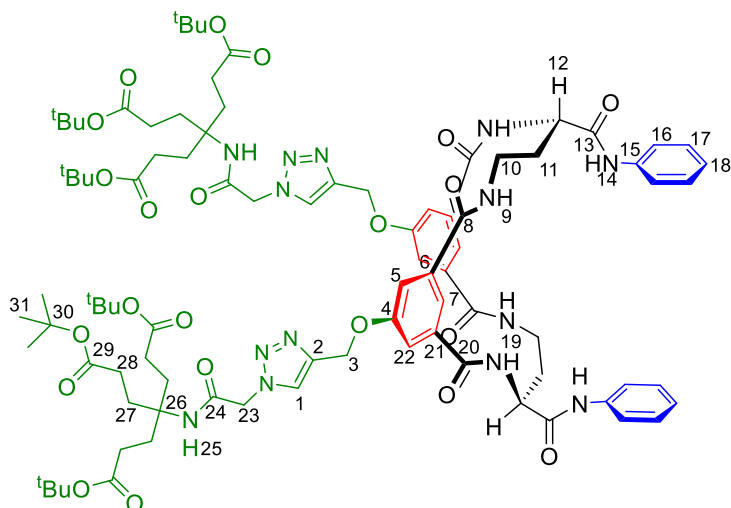
To a solution of Boc-Dab phenylamino PFP picolinate **S11** (100 mg, 0.15 mmol, 1.0 eqv) in DCM (3 ml) was added TFA (1 ml). The reaction was stirred at room temperature for 2 hours before the volatiles were removed by N₂ flow. The residue was dissolved in THF (10 ml) and stored in a syringe for later use. Separately, a solution of, DIPEA (262 μ L, 1.5 mmol, 10.0 eqv) in THF (150 ml) was prepared in a round bottomed flask. The solution in the syringe was added slowly to the round bottomed flask at room temperature over 24 hours. The solution was then stirred at room temperature for 4 hours before the solvent was removed *in vacuo*. The residue was then purified by reverse phase column chromatography (H₂O:acetone = 40:60) to give the title compound (39 mg, 51 μ mol, 68%) as a white solid.

¹H NMR (500 MHz, DMSO-*d*₆) δ 10.22 (s, 1H, N14H), 9.02 (d, *J* = 9.6 Hz, 1H, N19H), 8.92 (d, *J* = 8.2 Hz, 1H, N9H), 7.70 – 7.55 (m, 2H, C16H), 7.39 (d, *J* = 2.6 Hz, 1H, C5H), 7.34 (d, *J* = 2.6 Hz, 1H, C22H), 7.33 – 7.28 (m, 2H, C17H), 7.09 – 7.02 (m, 1H, C18H), 4.89 (d, *J* = 2.2 Hz, 2H, C3H), 4.87 – 4.81 (m, 1H, C12H), 4.35 – 4.28 (m, 1H, C10H), 4.38 – 3.32 (m, 1H, C10H), 3.67 (t, *J* = 2.4 Hz, 1H), 2.54 – 2.45 (m, 1H, C11H), 2.25 – 2.16 (m, 1H, C11H). **¹³C NMR** (126 MHz, DMSO-*d*₆) δ 170.4 (C13), 165.1 (C20), 162.5 (C8), 162.0 (C4), 150.5 (C6), 150.0 (C21), 139.0 (C15), 128.7 (C17), 123.4 (C18), 119.3 (C16), 110.3 (C5), 109.7 (C22), 79.5 (C1), 77.7 (C2), 67.0, 56.1 (C3), 54.5 (C12), 38.2 (C10), 30.7 (C11). **HRMS** for C₄₀H₃₇N₈O₈ [M+H]⁺ Calculated *m/z* = 757.2729 Found *m/z* = 757.2730.

[illegible]

¹H NMR (600 MHz, MeOD) δ 8.09 (s, 1H, C12H), 8.08 (s, 1H, C20H), 7.63 (s, 1H, C14H), 7.59 (d, *J* = 7.9 Hz, 2H, C3H), 7.55 (s, 1H, C16H), 7.31 (t, *J* = 7.8 Hz, 2H, C2H), 7.11 (t, *J* = 7.4 Hz, 1H, C1H), 5.22 (s, 2H, C19H), 5.11 (s, 2H, C22H), 4.79 – 4.74 (m, 1H, C7H), 4.33 (t, *J* = 12.5 Hz, 1H, C8H), 3.72 (d, *J* = 13.8 Hz, 1H, C8H), 2.24 – 2.18 (m, 6H, C27H), 1.97 – 1.91 (m, 6H, C26H), 1.41 (s, 27H, C30H). **¹³C NMR** (151 MHz, MeOD) δ 174.3 (C28), 171.3 (C6), 170.2 (C17), 169.1 (C10), 166.8 (C23), 160.3 (C15), 139.3 (C4), 137.3 (C11), 136.7 (C13), 129.9 (C2), 127.4 (C21), 125.7 (C1), 121.9 (C3), 121.4 (C20), 120.9 (C12), 118.5 (C14), 118.0 (C16), 81.8 (C29), 62.9 (C19), 49.6 (C25), 59.4 (C7), 53.4 (C22), 41.7 (C8), 30.6 (C27), 30.4 (C26), 28.4 (C30). **HRMS** for C₈₈H₁₁₈N₁₄O₂₂Na₂ [M+2Na]²⁺ Calculated *m/z* = 884.4171 Found *m/z* = 884.4164.

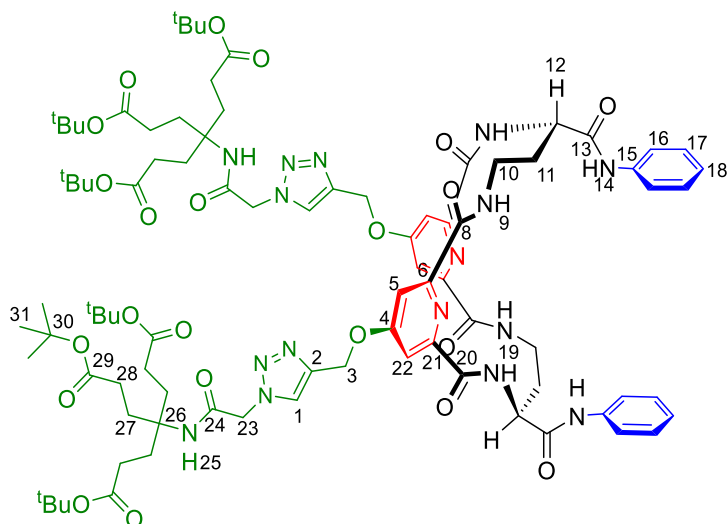
20-membered isophthaloyl G₁-ester macrocycle (9-P)



To a solution of 20-membered isophthaloyl macrocycle **9a** (10 mg, 9.7 μ mol, 1.0 eqv) in de-gassed MeCN (5 ml) was added G₁ azido tri-ester **S1** (15 mg, 29 μ mol, 3.0 eqv) and 2,6-lutidine (10 mg, 11 μ mol, 10.0 eqv). The solution was stirred at room temperature for 15 minutes after which tetrakis(acetonitrile)copper(I) hexafluorophosphate (18 mg, 48 μ mol, 5.0 eqv) was added. The reaction mixture was stirred at 60 °C for overnight then the solvent was removed *in vacuo*. The residue was dissolved in DCM (15 ml) and washed with 0.1 M aqueous HCl (2 x 10 ml). The organic phase was then washed with brine, dried over MgSO₄ and concentrated *in vacuo*. The residue was purified by reverse phase column chromatography (H₂O:acetone = 10:90) to give the title compound (11 mg, 5 μ mol, 53 %) as a white solid.

¹H NMR (500 MHz, Methanol-*d*₄) δ 8.30 (s, 1H, C2H), 8.09 (s, 1H, C7H), 7.68 – 7.42 (m, 4H, C5,22,16H), 7.31 – 7.27 (m, 2H, C17H), 7.09 (d, *J* = 7.6 Hz, 1H, C18H), 5.20 (s, 2H, C3H), 5.14 (s, 2H, C23H), 4.84 – 4.82 (m, 1H, C12H), 3.83 (s, 1H, C10H), 3.42 (s, 1H, C10H), 2.46 – 2.31 (m, 2H, C11H), 2.23 (t, *J* = 8.0 Hz, 6H, C28H), 1.96 (t, *J* = 8.2 Hz, 6H, C27H), 1.42 (s, 27H, C31H). **¹³C NMR** (126 MHz, Methanol-*d*₄) δ 174.3 (C29), 172.4 (C13), 168.4 (C24), 166.8 (C8), 163.1 (C20), 159.7 (C4), 144.4 (C1), 139.5 (C15), 136.5 (C6,21), 129.9 (C17), 127.3 (C7), 125.5 (C18), 121.6 (C16), 121.1 (C2), 118.0 (C5), 117.6 (C22), 81.8(C30), 62.9 (C3), 59.4 (C26), 55.8 (C12), 53.4 (C23), 38.3 (C10), 32.5 (C11), 30.6 (C28), 30.5 (C27), 28.4 (C31). **HRMS** for C₉₀H₁₂₃N₁₄O₂₂ [M+H]⁺ Calculated *m/z* = 1751.8936 Found *m/z* = 1751.8984.

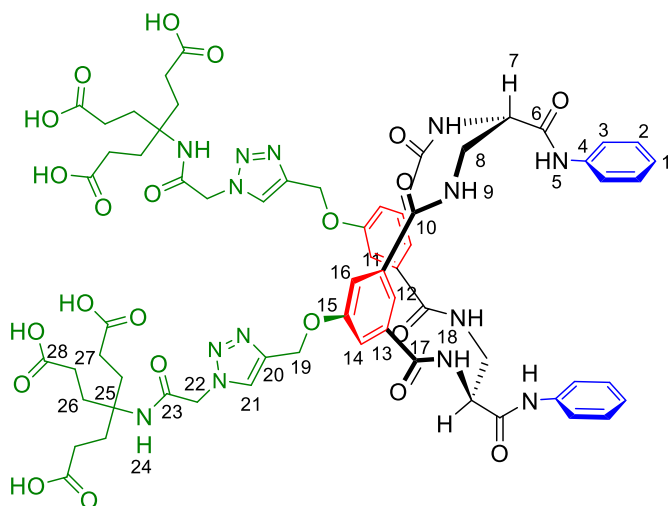
20-membered dipicolinoyl G₁-ester macrocycle (10-P)



To a solution of macrocycle **10a** (10 mg, 9.7 μ mol, 1.0 eqv) in de-gassed MeCN (5 ml) was added G₁ azido tri-ester **S1** (15 mg, 29 μ mol, 3.0 eqv) and 2,6-lutidine (10 mg, 11 μ mol, 10.0 eqv). The solution was stirred at room temperature for 15 minutes after which tetrakis(acetonitrile)copper(I) hexafluorophosphate (18 mg, 48 μ mol, 5.0 eqv) was added. The reaction mixture was stirred at 60 °C for overnight before the solvent was removed *in vacuo*. The residue was dissolved in DCM (15 ml) and washed with 0.1 M aqueous HCl (2 x 10 ml). The organic phase was then washed with brine, dried over MgSO₄ and concentrated *in vacuo*. The residue was purified by reverse phase column chromatography (H₂O:acetone = 6:94) to give the title compound (10 mg, 4.8 μ mol, 48 %) as a white solid.

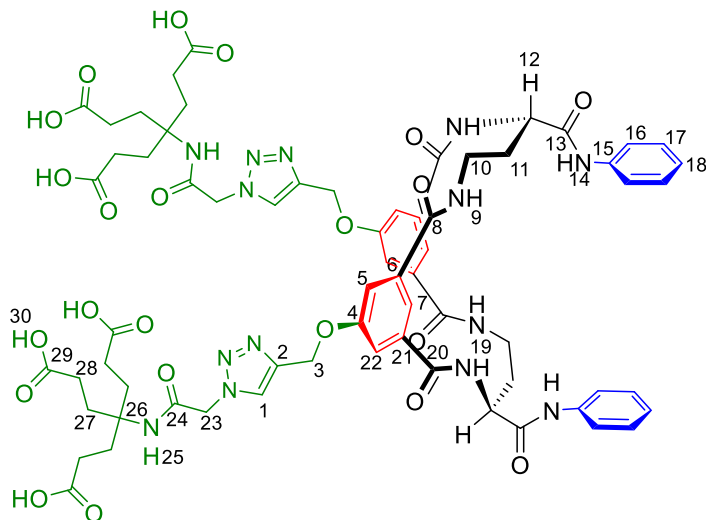
¹H NMR (500 MHz, Methanol-*d*₄) δ 8.18 (s, 1H, C2H), 7.63 – 7.58 (m, 2H, C16H), 7.57 – 7.50 (m, 2H, C5,22H), 7.36 – 7.30 (m, 2H, C17H), 7.16 – 7.09 (m, 1H, C18H), 5.26 (s, 2H, C3H), 5.16 (s, 2H, C23H), 5.10 – 5.04 (m, 1H, C12H), 4.37 (t, *J* = 12.6 Hz, 1H, C10H), 3.43 – 3.37 (m, 1H, C10H), 2.51 – 2.35 (m, 2H, C11H), 2.29 – 2.20 (m, 6H, C28H), 2.00 – 1.93 (m, 6H, C27H), 1.43 (s, 27H, C31H). **¹³C NMR** (126 MHz, Methanol-*d*₄) δ 174.4 (C29), 172.2 (C13), 168.1 (C24), 166.8 (C8), 165.2 (C20), 164.7 (C4), 151.8 (C6), 151.4 (C21), 143.1 (C1), 139.4 (C15), 129.9 (C17), 127.8 (C2), 125.6 (C18), 121.5 (C16), 112.1 (C5), 111.9 (C22), 81.7 (C30), 63.1 (C3), 59.4 (C26), 56.1 (C12), 53.5 (C23), 39.4 (C10), 31.8 (C11), 30.6 (C28), 30.5 (C27), 28.4 (C31). **HRMS** for C₈₈H₁₂₁N₁₆O₂₂ [M+H]⁺ Calculated *m/z* = 1753.8841 Found *m/z* = 1753.8885.

18-membered isophthaloyl G₁-acid macrocycle (8b-H)



To a solution of macrocycle **8-P** (10 mg, 5.7 μ mol) in DCM (3 ml) was added TFA (1 mL). The reaction mixture was stirred at 0 °C for 3 hours before the solvent was removed by N₂ flow to give the title compound (6.8 mg, 4.9 μ mol, 86 %) as a white solid. The compound was characterized in D₂O after deprotonation.

20-membered isophthaloyl G₁-acid macrocycle (9b-H)

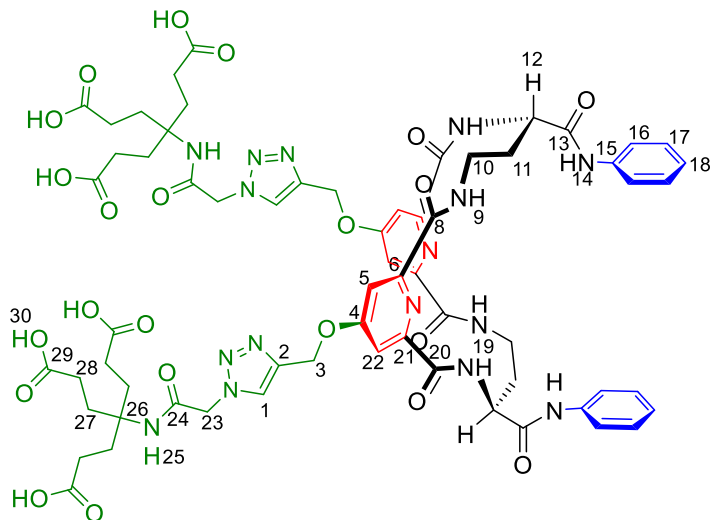


To a solution of macrocycle **9-P** (20 mg, 11.4 μ mol) in DCM (3 ml) was added TFA (1 mL). The reaction mixture was stirred at 0 °C for 3 hours before the volatiles were removed by N₂ flow to give the title compound (13 mg, 9.4 μ mol, 82 %) as a white solid.

¹H NMR (600 MHz, DMSO-*d*₆) δ 12.08 (s, 3H, O30H), 10.07 (s, 1H, N14H), 8.89 (d, *J* = 7.0 Hz, 1H, N19H), 8.79 (t, *J* = 5.4 Hz, 1H, N9H), 8.48 (s, 3H, C7H), 8.14 (s, 1H, C2H), 7.78 (s, 1H, N25), 7.66 – 7.57 (m, 4H, C5,16,22H), 7.30 (t, *J* = 7.8 Hz, 2H, C17H), 7.05 (t, *J* = 7.4 Hz, 1H, C18H), 5.21 (s, 1H, C23H), 5.09 (s, 1H, C3H), 4.71 – 4.65 (m, 1H, C12H), 3.61 (s, 2H, C10H), 2.42 – 2.39 (m, 1H, C11H), 2.22 – 2.10 (m, 7H, C11, 28H), 1.87 – 1.81 (m, 6H, C27H). **¹³C NMR** (151 MHz, DMSO-*d*₆) δ 174.3 (C29), 170.6 (C13), 165.1 (C8), 165.0

(C20), 164.7 (C4), 157.9 (C24), 142.0 (C1), 139.0 (C15), 135.2 (C6), 134.9 (C21), 128.7 (C17), 126.1 (C2), 123.3 (C18), 119.3 (C16), 118.5 (C7), 116.4 (C5), 116.1 (C22), 61.5 (C3), 57.0 (C26), 54.7 (C12), 51.7 (C23), 37.6 (C10), 30.6 (C11), 29.0 (C28), 27.9 (C27). **HRMS** for $C_{66}H_{75}N_{14}O_{22}$ $[M+H]^+$ Calculated m/z = 1415.5180 Found m/z = 1415.5195.

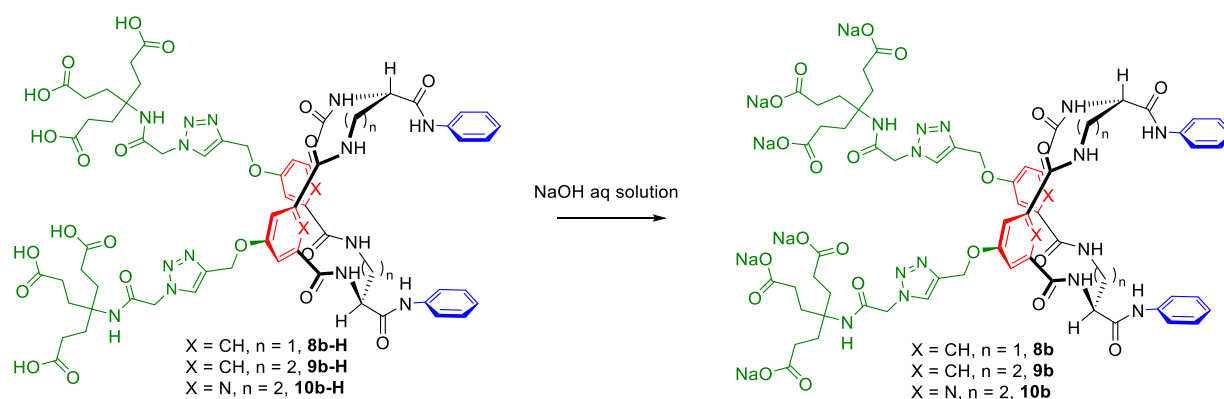
20-membered dipicolinoyl G_1 -acid macrocycle (10b-H)



To a solution of 20-membered dipicolinoyl G_1 -ester macrocycle **10-P** (15 mg, 7.4 μ mol) in DCM (3 ml) was added TFA (1 mL). The reaction mixture was stirred at 0 °C for 3 hours before the volatiles were removed by N_2 flow to give the title compound (12 mg, 6.9 μ mol, 89 %) as a white solid.

1H NMR (500 MHz, $DMSO-d_6$) δ 12.07 (s, 3H, O30H), 10.24 (s, 1H, N14H), 9.19 (d, J = 9.2 Hz, 1H, N9H), 9.06 (d, J = 8.0 Hz, 1H, N19H), 8.14 (s, 1H, C2H), 7.77 (s, 1H, N25H), 7.64 (d, J = 7.7 Hz, 2H, C16H), 7.49 (d, J = 2.6 Hz, 1H, C5H), 7.43 (d, J = 2.6 Hz, 1H, C22H), 7.34 – 7.28 (m, 2H, C17H), 7.08 – 7.03 (m, 1H, C18H), 5.27 (s, 2H, C23H), 5.08 (s, 2H, C3H), 4.83 (t, J = 9.3 Hz, 1H, C12H), 4.33 – 4.24 (m, 1H, C10H), 2.70 – 2.61 (m, 1H, C11H), 2.21 – 2.11 (m, 7H, C11,28H), 1.87 – 1.80 (m, 6H, C27H). **^{13}C NMR** (126 MHz, $DMSO-d_6$) δ 174.3 (C29), 170.5 (C13), 165.9 (C4), 164.6 (C24), 162.6 (C8), 162.3 (C20), 150.7 (C6), 150.1 (C21), 141.1 (C1), 139.1 (C15), 128.7 (C17), 126.5 (C2), 123.1 (C18), 119.3 (C16), 109.6 (C5), 110.2 (C22), 61.5 (C23), 57.0 (C26), 54.7 (C12), 51.5 (C3), 37.9 (C10), 30.2 (C11), 29.0 (C28), 27.9 (C27). **HRMS** for $C_{64}H_{73}N_{16}O_{22}$ $[M+H]^+$ Calculated m/z = 1417.5085 Found m/z = 1417.5082.

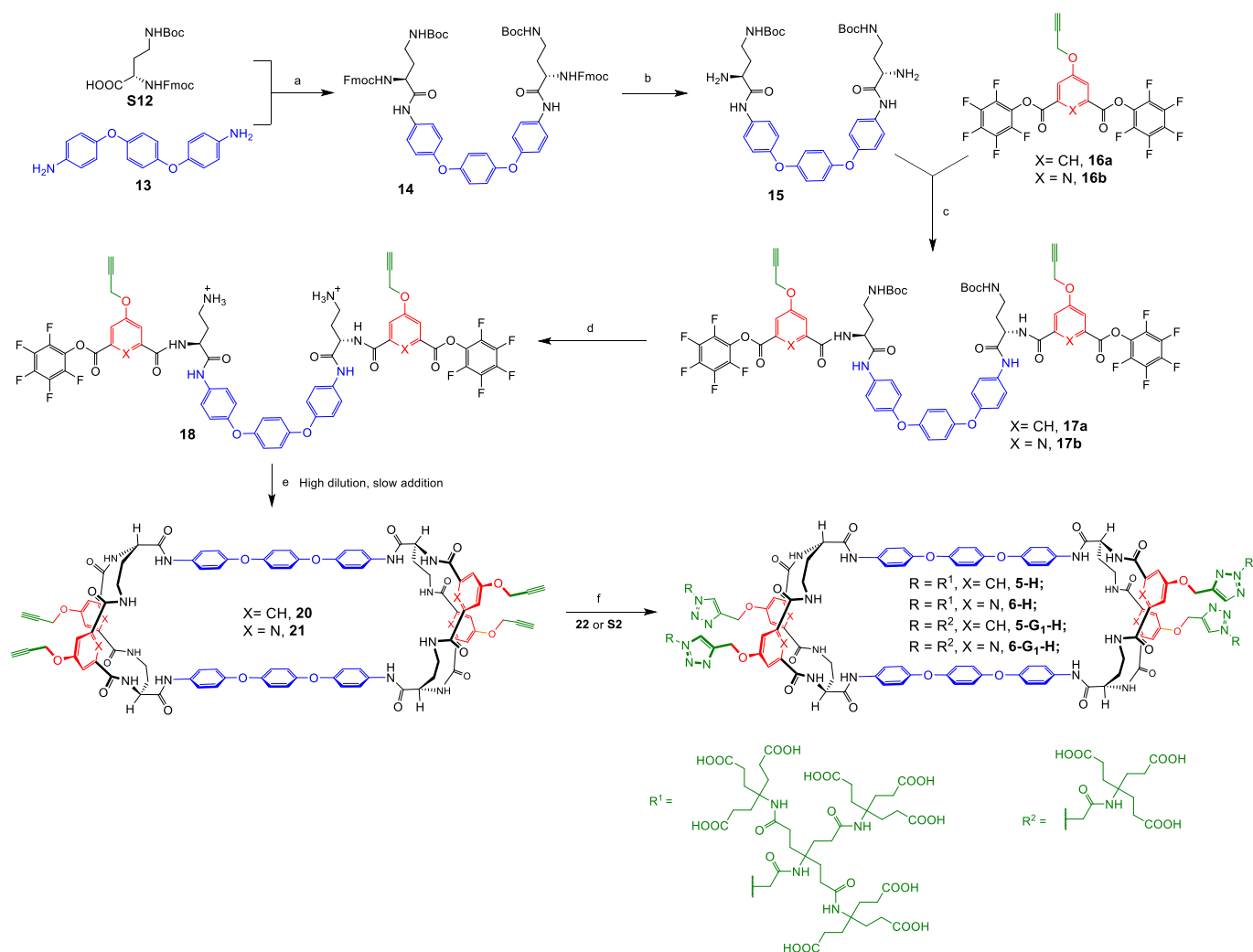
Water soluble 18/20-membered isophthaloyl/dipicolinoyl G₁ macrocycles (**8b**, **9b** and **10b**)



Supplementary Scheme 4. Synthesis of receptors **8b**, **9b** and **10b**.

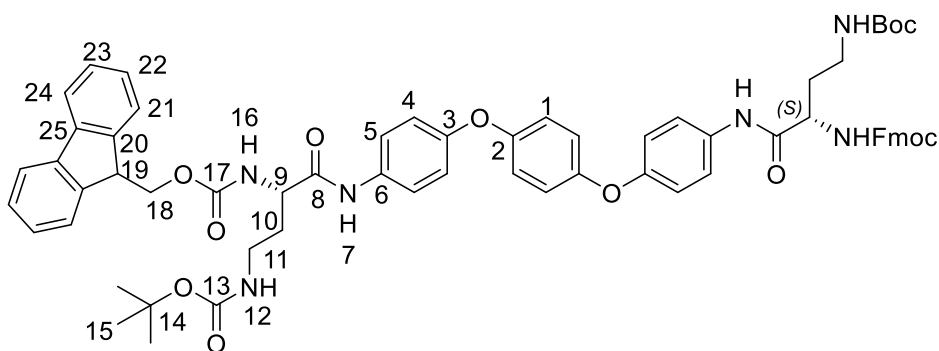
To a suspension of 18/20-membered isophthaloyl/dipicolinoyl G₁-acid macrocycles **8b-H**, **9b-H** or **10b-H** in water (10 ml) was added 10 mM NaOH aq solution to adjust the pH to 7.4. The resulting clear colourless solution was freeze-dried to yield the water soluble 18/20-membered isophthaloyl/dipicolinoyl G₁ macrocycles **8b**, **9b** or **10b** (quant yield) in their sodium form as white solids. Samples for characterisation and further studies were prepared by dissolution in D₂O or 9:1 H₂O/D₂O. The process of freeze-drying and dissolution was found to have no effect on the pH.

1.5 Synthesis of receptors 5, 6, 20, 21, 5-G₁ and 6-G₁



Supplementary Scheme 5. Synthesis of receptors **5-H**, **6-H**, **20**, **21**, **5-G₁-H** and **6-G₁-H**. a) HBTU, Na₂CO₃, THF; b) NaOH, MeOH, water; c) DIPEA, THF; d) TFA, DCM; e) DIPEA, THF, (X = N); DIPEA, TBACl, THF, (X = CH); f) [(CH₃CN)₄Cu]PF₆, 2,6-lutidine, DMF, MeCN.

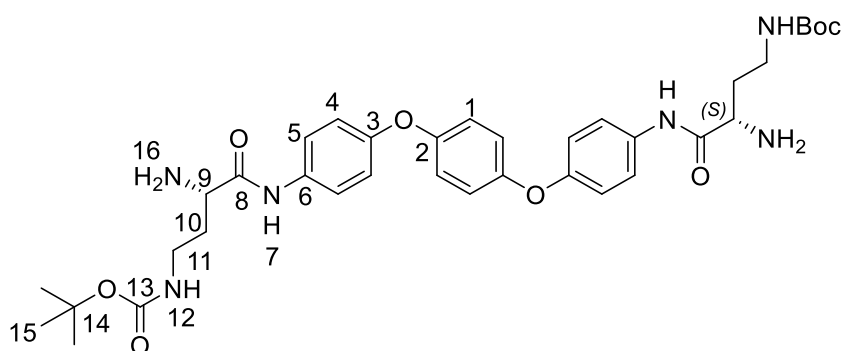
Bis(Fmoc-Dab(Boc)-phenoxyphenyl) diamide (14)



To a solution of commercially available Fmoc-Dab(Boc)-OH **S12** (100 mg, 227 μmol , 2.1 eqv) in dry THF (20 ml) was added HBTU (123 mg, 324 μmol , 3.0 eqv), Na_2CO_3 (57 mg, 540 μmol , 5.0 eqv) and 1,4-Bis(4-aminophenoxy)benzene **13** (32 mg, 108 μmol , 1.0 eqv). The reaction mixture was stirred at 60 $^\circ\text{C}$ for 12 hours before the solvent was removed *in vacuo*. The residue was suspended in water (50 ml), then filtered and washed with water (100 ml). The filtration cake was collected and dried in the oven to give the title compound (147 mg, 106 μmol , 98 %) as a white solid.

^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 10.07 (s, 1H, N7H), 7.89 (d, $J = 7.6$ Hz, 2H, C24H), 7.76 – 7.70 (m, 3H, N16H, C5H), 7.64 – 7.59 (m, 2H, C21H), 7.44 – 7.38 (m, 2H, C23H), 7.36 – 7.29 (m, 2H, C22H), 7.02 – 6.97 (m, 4H, C1, 4H), 6.80 – 6.76 (m, 1H, N12H), 4.32 – 4.20 (m, 3H, C18, 19H), 4.16 (q, $J = 7.7$ Hz, 1H, C9H), 3.02 (q, $J = 6.7$ Hz, 2H, C11H), 1.91 – 1.71 (m, 2H, C10H), 1.37 (s, 9H, C15H). **^{13}C NMR** (126 MHz, $\text{DMSO}-d_6$) δ 170.5 (C8), 164.6 (C17), 156.0 (C13), 155.5 (C2), 152.5 (C3), 143.8 (C20), 140.7 (C25), 134.5 (C6), 127.6 (C22), 127.1 (C23), 125.4 (C5), 120.9 (C21), 120.1 (C24), 119.7 (C1), 118.9 (C4), 77.7 (C14), 65.7 (C18), 53.3 (C9), 46.6 (C19), 37.1 (C11), 32.0 (C10), 28.2 (C15). **HRMS** for $\text{C}_{66}\text{H}_{69}\text{N}_6\text{O}_{12}$ $[\text{M}+\text{H}]^+$ Calculated $m/z = 1137.4960$ Found $m/z = 1137.4971$.

Bis(Boc-Dab-phenoxyphenyl) diamide (15)

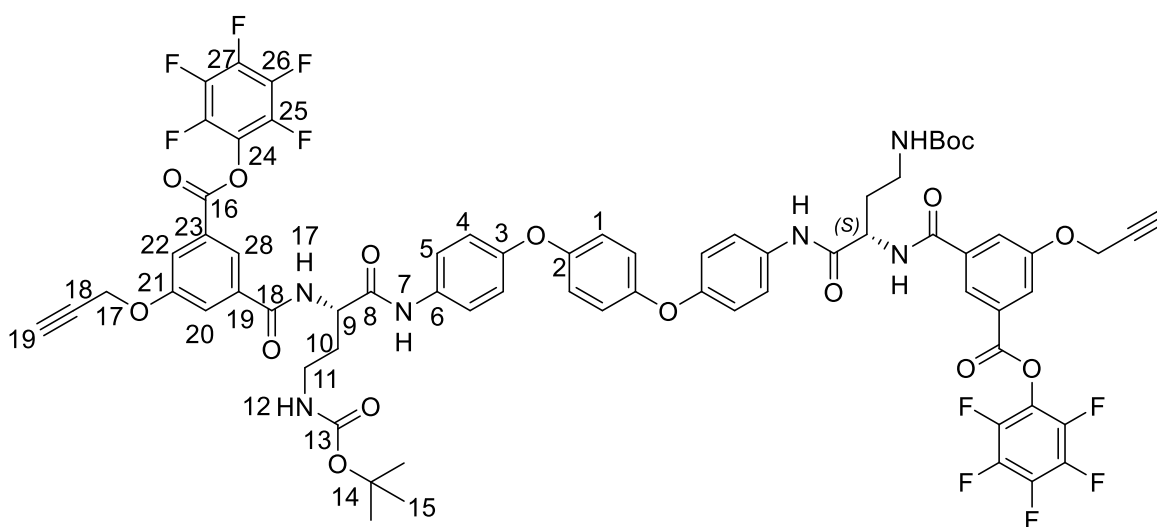


Bis(Fmoc-Dab(Boc)-phenoxyphenyl) diamide **14** (250 mg, 216 μmol , 1.0 eqv) was suspended in MeOH (30 ml) before 1 M NaOH aq solution (10 ml) was then added. The mixture was refluxed for 2 hours then concentrated *in vacuo*. The remaining aqueous phase was then filtered, and the cake was washed with water (3 x 20 ml) and

then hexane (3 x 20 ml). The filtration cake was collected and dried to give the title compound (114 mg, 164 μmol , 76 %) as a colourless gum.

^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 7.67 – 7.60 (m, 2H, C5H), 7.00 – 6.94 (m, 4H, C1,4H), 6.80 (t, J = 5.7 Hz, 1H, N12H), 3.32 – 3.26 (m, 1H, C9H), 3.05 (q, J = 6.7 Hz, 2H, C11H), 1.83 – 1.74 (m, 1H, C10H), 1.56 – 1.48 (m, 1H, C10H), 1.37 (s, 9H, C15H). **^{13}C NMR** (126 MHz, $\text{DMSO}-d_6$) δ 173.6 (C8), 155.6 (C13), 152.6 (C3), 152.4 (C2), 134.6 (C6), 120.8 (C5), 119.7 (C4), 118.8 (C1), 77.5 (C14), 53.5 (C9), 37.2 (C11), 35.0 (C10), 28.3 (C15). **HRMS** for $\text{C}_{36}\text{H}_{49}\text{N}_6\text{O}_8$ $[\text{M}+\text{H}]^+$ Calculated m/z = 693.3612 Found m/z = 693.3600.

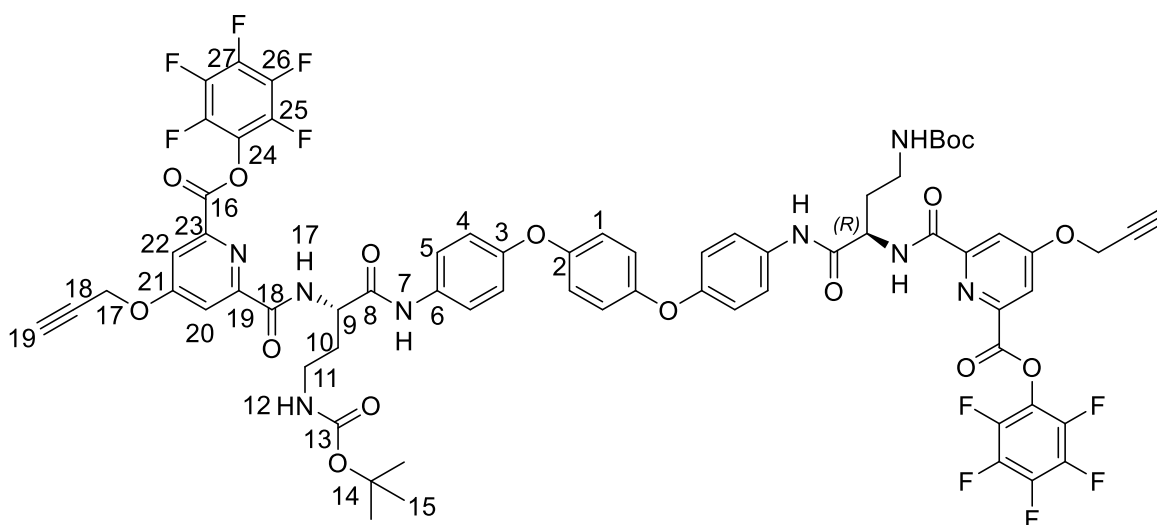
Bis(Boc-Dab phenylamino PFP benzoate) (17a)



To a solution of bis(pentafluorophenyl)-5-(propargyloxy)isophthalate **16a** (799 mg, 1.4 mmol, 5.0 eqv) in THF (30 ml) was added DIPEA (250 μL , 1.4 mmol, 5.0 eqv). A solution of bis(Boc-Dab-phenoxyphenyl) diamide **15** (200 mg, 289 μmol , 1.0 eqv) in THF (10 ml) was added over 1 hour by a syringe pump. The reaction was stirred at room temperature for 4 hours after the completion of the addition. The solvent was removed *in vacuo*, and the residue was then purified by column chromatography (hexane:EtOAc = 10:90) to give the title compound (373 mg, 261 μmol , 90 %) as a white solid.

^1H NMR (500 MHz, CDCl_3) δ 10.08 (s, 1H, N7H), 8.28 (s, 1H, C28H), 7.94 – 7.86 (m, 3H, N17, C20H), 7.81 (s, 1H, C22H), 7.59 (d, J = 8.6 Hz, 2H, C5H), 6.98 – 6.90 (m, 4H, C1,4H), 5.37 – 5.27 (m, 1H, N12H), 4.89 – 4.82 (m, 1H, C9H), 4.79 (s, 2H, C17H), 3.65 – 3.54 (m, 1H, C11H), 3.20 – 3.11 (m, 1H, C11H), 2.56 (t, J = 2.3 Hz, 1H, C19H), 2.19 – 2.00 (m, 2H, C10H), 1.44 (s, 9H, C15H). **^{13}C NMR** (126 MHz, CDCl_3) δ 177.9 (C8), 169.3 (C18), 165.3 (C16), 161.7 (C21), 158.1 (C13), 154.2 (C2), 153.0 (C3), 142.3 (C24), 140.3 (C26), 139.1 (C27), 137.0 (C25), 136.2 (C6), 133.4 (C19), 128.7 (C23), 122.1 (C28), 121.6 (C5), 120.4 (C22), 120.1 (C20, C1), 119.1 (C4), 80.7 (C14), 77.2 (C18), 76.8 (C19), 56.5 (C17), 51.7 (C9), 37.2 (C11), 34.9 (C10), 28.5 (C15). **HRMS** for $\text{C}_{70}\text{H}_{58}\text{F}_{10}\text{N}_6\text{O}_{16}\text{Na}$ $[\text{M}+\text{Na}]^+$ Calculated m/z = 1451.3647 Found m/z = 1451.3622.

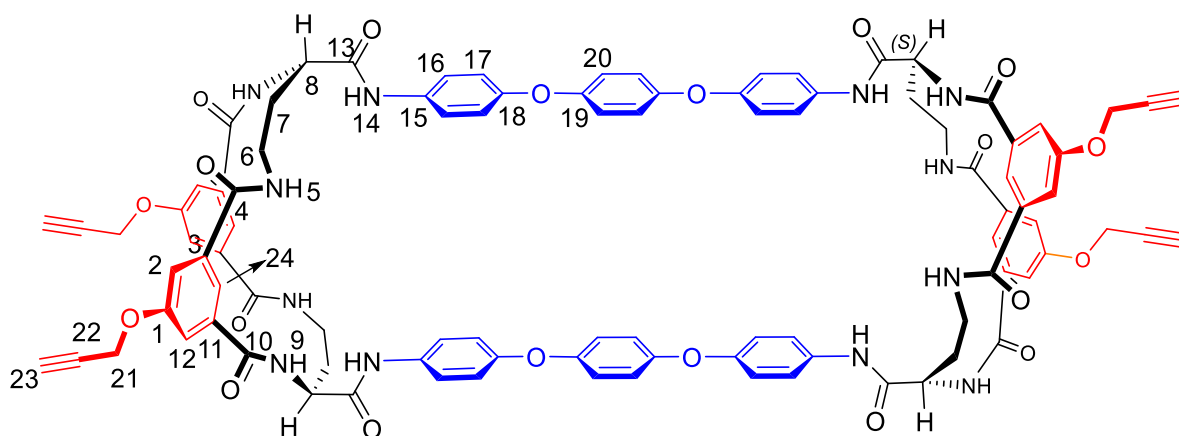
Bis(Boc-Dab phenylamino PFP picolinate) (17b)



To a solution of bis(pentafluorophenyl) 4-(propargyloxy)pyridine-2,6-dicarboxylate **16b** (799 mg, 1.4 mmol, 5.0 eqv) in THF (30 ml) was added bis(Boc-Dab-phenoxypyphenyl) diamide **15** (200 mg, 289 μ mol, 1.0 eqv) THF solution (10 ml) over 1 hour by a syringe pump. The reaction was allowed to stirred at room temperature for 1 hour after the completion of the addition. The solvent was removed *in vacuo*, and the residue was then purified by column chromatography (hexane:EtOAc = 15:85) to give the title compound (339 mg, 237 μ mol, 82 %) as a white solid.

¹H NMR (600 MHz, CDCl₃) δ 9.80 (s, 1H, N7H), 8.85 (d, J = 7.9 Hz, 1H, N17H), 8.05 (d, J = 2.5 Hz, 1H, C20), 7.95 (d, J = 2.5 Hz, 1H, C22), 7.65 – 7.60 (m, 2H, C5H), 6.99 – 6.95 (m, 2H, C4H), 6.94 (s, 2H, C1H), 4.90 (d, J = 2.4 Hz, 2H, C17H), 4.77 (td, J = 9.0, 4.6 Hz, 1H, C9H), 3.61 (ddd, J = 14.5, 10.7, 3.4 Hz, 1H, C11H), 3.12 (dt, J = 14.9, 4.5 Hz, 1H, C11H), 2.63 (t, J = 2.4 Hz, 1H, C19H), 2.16 – 2.10 (m, 1H, C10H), 2.06 – 2.00 (m, 1H, C10H), 1.50 (s, 9H, C15H). **¹³C NMR** (151 MHz, CDCl₃) δ 168.7 (C8), 166.2 (C16), 162.8 (C18), 160.8 (C13), 158.0 (C21), 154.1 (C2), 153.1 (C23), 152.4 (C19), 151.8 (C3), 145.9 (C24), 133.6 (C6), 121.6 (C5), 120.0 (C1), 119.2 (C4), 116.7 (C22), 112.4 (C20), 80.7 (C18), 77.9 (C19), 76.2 (C14), 56.7 (C19), 51.5 (C9), 37.4 (C11), 35.2 (C10), 28.5 (C15). **HRMS** for C₆₈H₅₆F₁₀N₈O₁₆Na [M+Na]⁺ Calculated m/z = 1453.3552 Found m/z = 1453.3544.

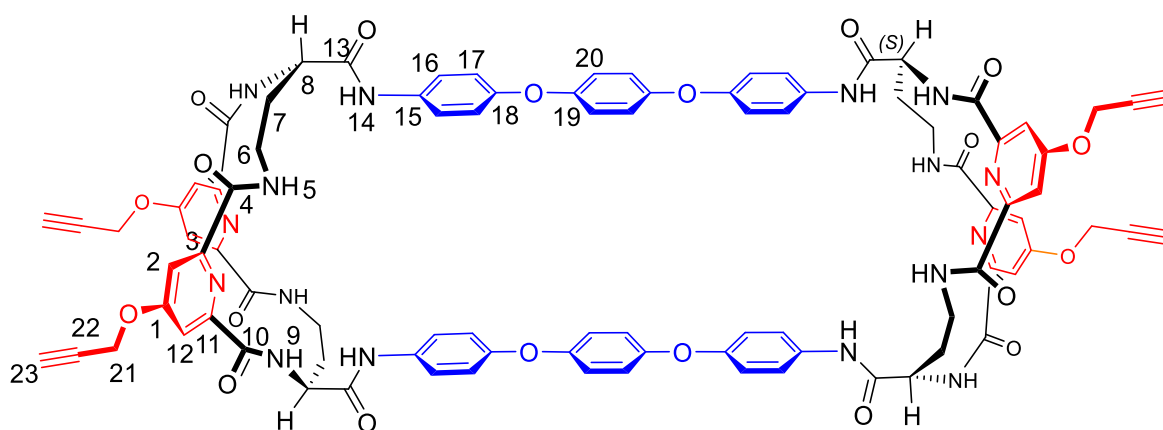
Isophthaloyl tricycle 20



To a solution of bis(Boc-Dab phenylamino PFP benzoate) **17a** (100 mg, 70 μmol , 1.0 eqv) in DCM (3 ml) was added TFA (1 ml). The reaction was stirred at room temperature for 2 hours before the volatiles were removed by N_2 flow. The residue was dissolved in THF (10 ml) and stored in a syringe for later use. Separately, a solution of DIPEA (304 μL , 1.8 mmol, 25.0 eqv) and TBACl (97 mg, 350 μmol , 5.0 eqv) in THF (70 ml) was prepared in a round bottomed flask. The solution in the syringe was added slowly to the round bottomed flask at room temperature over 24 hours. The solution was then stirred at room temperature for 4 hours before the solvent was removed *in vacuo*. The residue was then purified by reverse (H_2O :acetone = 45:56) and normal (DCM:MeOH = 68:32) phase column chromatography to give the title compound (30 mg, 18 μmol , 50%) as a white solid.

^1H NMR (500 MHz, $\text{DMSO}-d_6$) δ 10.13 (s, 1H, N14H), 9.30 (d, J = 4.5 Hz, 1H, N9H), 8.98 (s, 1H, N5H), 8.67 (s, 1H, C24H), 7.63 – 7.55 (m, 6H, C2,12,16H), 7.03 – 6.94 (m, 4H, C17,20H), 4.84 (d, J = 2.3 Hz, 2Hm C21H), 4.61 (s, 1H, C9H), 3.57 (t, J = 2.3 Hz, 1H, C23H), 3.51 (s, 2H, C6H), 2.27 (s, 1H, C7H), 2.18 (s, 1H, C7H). **^{13}C NMR** (126 MHz, $\text{DMSO}-d_6$) δ 170.0 (C13), 165.5 (C4), 164.8 (C10), 157.1 (C1), 152.7 (C19), 152.6 (C18), 134.8 (C3,11), 134.6 (C15), 121.2 (C16), 119.9 (C17), 119.5 (C24), 118.8 (C20), 116.6 (C12), 116.5 (C2), 78.8 (C22), 78.7 (C23), 55.8 (C21), 54.5 (C8), 36.8 (C6), 31.6 (C7). **HRMS** for $\text{C}_{96}\text{H}_{81}\text{N}_{12}\text{O}_{20}$ $[\text{M}+\text{H}]^+$ Calculated m/z = 1721.5690 Found m/z = 1721.5685.

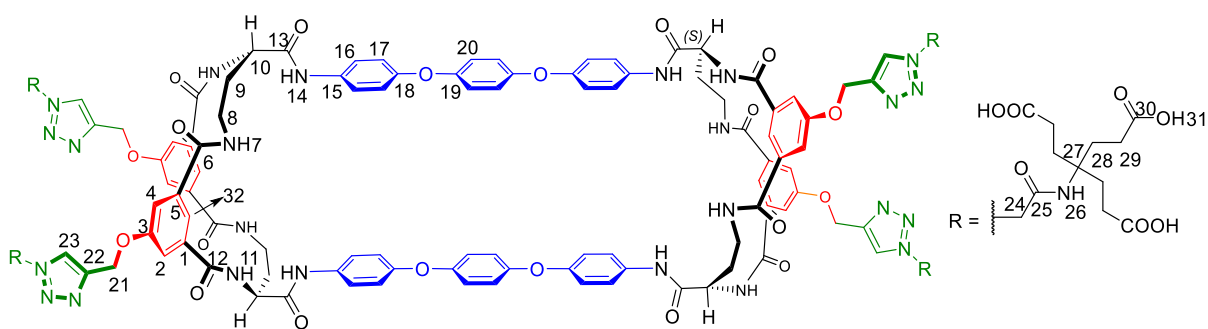
Dipicolinoyl tricycle 21



To a solution of bis(Boc-Dab phenylamino PFP picolinate) **17b** (100 mg, 70 μmol , 1.0 eqv) in DCM (3 ml) was added TFA (1 ml). The reaction was stirred at room temperature for 2 hours before the volatiles were removed by N_2 flow. The residue was dissolved into THF (10 ml) and stored in a syringe for later use. Separately, a solution of DIPEA (304 μL , 1.8 mmol, 25.0 eqv) in THF (70 ml) was prepared in a round bottomed flask. The solution in the syringe was added slowly to the round bottomed flask at room temperature over 24 hours. The solution was stirred at room temperature for 4 hours before the solvent was removed *in vacuo*. The residue was then purified by reverse phase column chromatography (H_2O :acetone = 32:68) to give the title compound (36 mg, 21 μmol , 60%) as a white solid.

^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 10.21 (s, 1H, N13), 8.98 – 8.90 (m, 2H, N5,9H), 7.68 – 7.62 (m, 2H, C16H), 7.39 (d, J = 2.6 Hz, 1H, C2H), 7.36 (d, J = 2.6 Hz, 1H, C12H), 7.03 – 7.00 (m, 2H, C17H), 6.91 (s, 2H, C20H), 4.91 – 4.86 (m, 3H, C21,8H), 4.20 (m, 1H, C6H), 3.67 (t, J = 2.3 Hz, 1H, C23H), 3.33 (m, 1H, C6H), 2.46 (m, 1H, C7H), 2.27 (m, 1H, C7H). **^{13}C NMR** (151 MHz, $\text{DMSO}-d_6$) δ 170.6 (C13), 165.6 (C1), 162.8 (C4), 162.6 (C10), 153.5 (C18), 152.6 (C19), 151.0 (C3), 150.5 (C11), 135.4 (C15), 121.5 (C16), 120.2 (C17), 119.4 (C20), 110.5 (C2), 110.3 (C12), 80.0 (C23), 78.1 (C22), 56.6 (C21), 54.8 (C8), 38.5 (C6), 30.5 (C7). **HRMS** for $\text{C}_{92}\text{H}_{78}\text{N}_{16}\text{O}_{20}$ $[\text{M}+\text{H}]^+$ Calculated m/z = 1725.5500 Found m/z = 1725.5520.

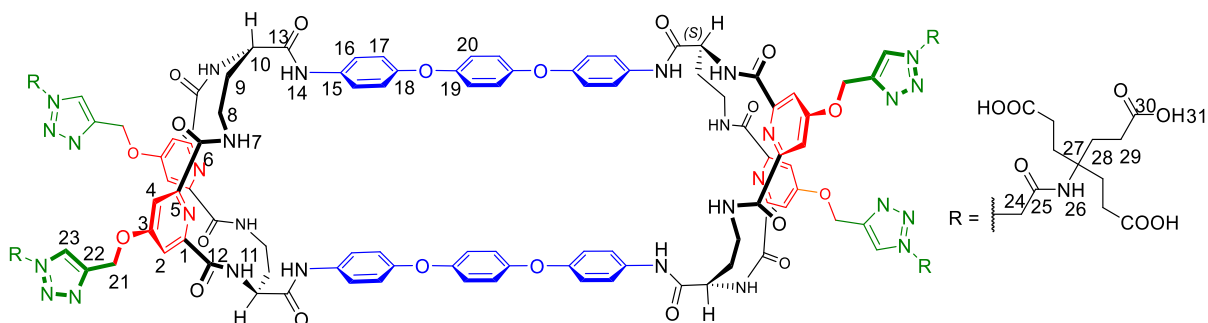
Isophthaloyl G₁-acid tricycle 5-G₁-H



To a solution of tricycle **20** (15.0 mg, 8.7 μ mol, 1.0 eqv) in de-gassed MeCN and DMF mixture (2 ml + 2 ml) was added G₁ azido tri-acid **S2** (14.4 mg, 43.6 μ mol, 5.0 eqv) and 2,6-lutidine (9.3 mg, 87 μ mol, 10.0 eqv). The solution was stirred at room temperature for 15 minutes before tetrakis(acetonitrile)copper(I) hexafluorophosphate (16.2 mg, 43.6 μ mol, 5.0 eqv) was added. The reaction mixture was stirred at 60 °C for overnight before the solvent was removed *in vacuo*. The residue was then purified by reverse phase column chromatography (0.01 M aqueous HCl:acetone = 20:80) to give the title compound (22.5 mg, 7.4 μ mol, 85 %) as a colourless oil.

¹H NMR (600 MHz, DMSO-*d*₆) δ 12.09 (s, 6H, O31H), 10.12 (s, 1H, N14H), 9.11 (s, 1H, N11H), 8.92 (s, 1H, N7H), 8.75 (s, 1H, C32H), 8.12 (s, 1H, C23H), 7.79 (s, 1H, N26H), 7.63 – 7.59 (m, 3H, C17,2H), 7.56 (s, 1H, C4H), 7.01 – 6.97 (m, 4H, C16,20H), 5.17 (s, 2H, C21H), 5.09 (s, 2H, C24H), 4.69 (s, 1H, C10H), 3.68 – 3.65 (m, 1H, C8H), 3.53 – 3.50 (m, 1H, C8H), 2.49 – 2.43 (m, 1H, C9H), 2.21 – 2.14 (m, 7H, C9,29H), 1.88 – 1.81 (m, 6H, C28H). **¹³C NMR** (151 MHz, DMSO-*d*₆) δ 174.3 (C30), 170.0 (C13), 166.1 (C25), 164.7 (C3), 158.1 (C6,12), 152.7 (C18,19), 142.0 (C22), 134.6 (C1,5), 134.4 (C15), 126.1 (C23), 121.4 (C17), 119.8 (C16), 118.9 (C20), 118.1 (C32), 116.7 (C2), 116.2 (C4), 61.5 (C21), 57.0 (C10), 51.8 (C24), 40.4 (C27), 34.2 (C8), 30.4 (C9), 29.1 (C29), 28.0 (C28). **HRMS** for C₁₄₄H₁₄₉N₂₈O₄₈ [M-3H]³⁻ Calculated m/z = 1012.6693 Found m/z = 1012.6707.

Dipicolinoyl G₁-acid tricycle 6-G₁-H

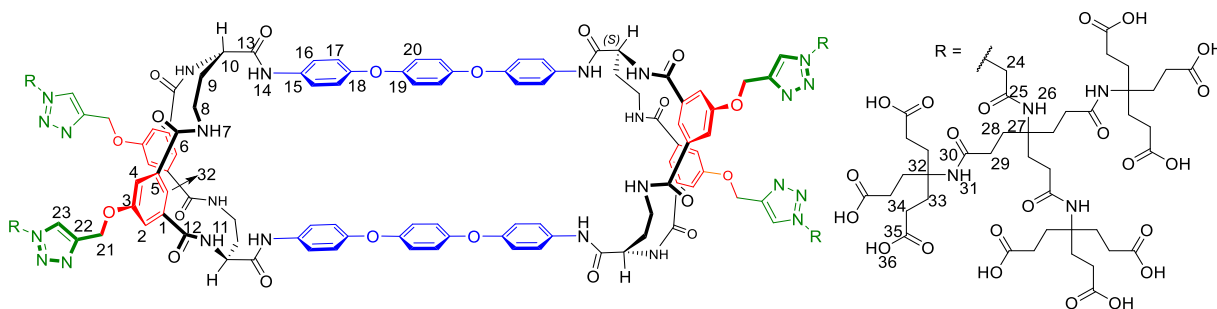


To a solution of tricycle **21** (10.0 mg, 5.8 μ mol, 1.0 eqv) in de-gassed MeCN and DMF mixture (2 ml + 2 ml) was added G₁ azido tri-acid **S2** (9.6 mg, 29.0 μ mol, 5.0 eqv) and 2,6-lutidine (6.2 mg, 58.1 μ mol, 10.0 eqv). The solution was stirred at room temperature for 15 minutes before tetrakis(acetonitrile)copper(I)

hexafluorophosphate (10.8 mg, 29.0 μmol , 5.0 eqv) was added. The reaction mixture was stirred at 60 °C for overnight before the solvent was removed *in vacuo*. The residue was then purified by reverse phase column chromatography (0.01 M aqueous HCl:acetone = 30:70) to give the title compound (13.3 mg, 4.4 μmol , 75 %) as a colourless oil.

^1H NMR (600 MHz, DMSO- d_6) δ 12.08 (s, 6H, O31H), 10.27 (s, 1H, N14H), 9.43 – 9.27 (m, 2H, N7,11H) 8.15 (s, 1H, C23H), 7.84 (s, 1H, N26H), 7.67 (d, J = 8.9 Hz, 2H, C17H), 7.50 (d, J = 2.1 Hz, 1H, C2H), 7.46 (d, J = 2.2 Hz, 1H, C4H), 7.00 (d, J = 8.8 Hz, 2H, C16H), 6.94 (s, 2H, C20H), 5.27 (s, 2H, C21H), 5.09 (s, 2H, C24H), 4.86 (s, 1H, C10H), 4.09 (s, 1H, C8H), 3.50 (s, 1H, C8H), 2.80 (s, 1H, C9H), 2.25 – 2.11 (m, 7H, C9,29H), 1.91 – 1.78 (m, 6H, C28H). **^{13}C NMR** (151 MHz, DMSO- d_6) δ 175.0 (C30), 170.5 (C13), 166.4 (C6,12), 165.1 (C25), 163.0 (C3), 153.4 (C1), 152.7 (C5), 151.3 (C18), 150.8 (C19), 141.6 (C22), 135.4 (C15), 127.0 (C23), 121.6 (C17), 119.9 (C16), 119.6 (C20), 110.6 (C2,4), 70.2 (C27), 62.1 (C21), 57.5 (C8), 55.2 (C10), 52.3 (C24), 40.9 (C9), 29.6 (C28), 28.6 (C29). **HRMS** $\text{C}_{140}\text{H}_{146}\text{N}_{32}\text{O}_{48}$ $[\text{M}-2\text{H}]^{2-}$ Calculated m/z = 1521.9999 Found m/z = 1521.9989.

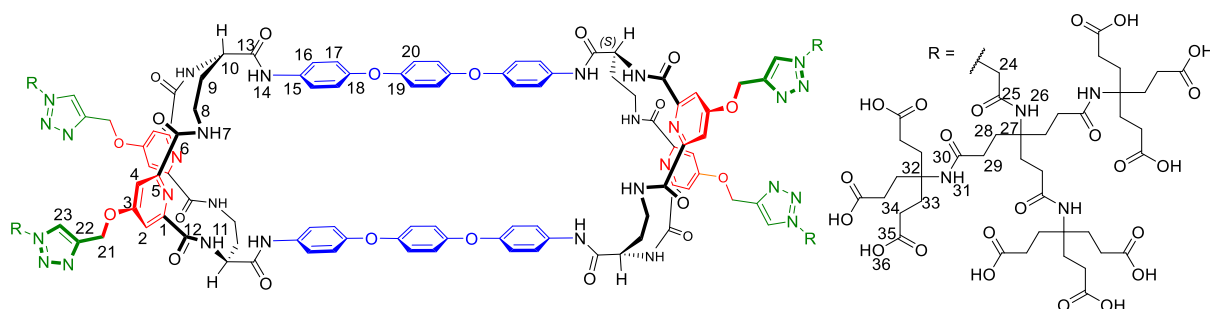
Isophthaloyl G_2 -acid tricycle 5-H



To a solution of isophthaloyl tricycle (15 mg, 8.7 μmol , 1.0 eqv) in de-gassed MeCN and DMF (3 ml + 3 ml) was added G_2 azido tri-acid **22** (53 mg, 52 μmol , 6 eqv) and 2,6-lutidine (10 mg, 52 μL , 10 eqv). The solution was stirred at room temperature for 15 minutes before tetrakis(acetonitrile)copper(I) hexafluorophosphate (19 mg, 52 μmol , 6 eqv) was added. The reaction mixture was stirred at 60 °C for overnight before the solvent was removed *in vacuo*. The residue was then purified by reverse phase column chromatography (0.01 M aqueous HCl:acetone = 40:60) to the title compound (28 mg, 4.8 μmol , 55 %) as a colourless oil.

^1H NMR (500 MHz, DMSO- d_6) δ 12.00 (s, 9H, O36H), 10.12 (s, 1H, N14H), 9.59 (s, 1H, N7H), 9.15 (s, 1H, N11H), 8.84 (s, 1H, C37H), 8.15 (s, 1H, C23H), 7.80 (s, 1H, N26H), 7.77 (s, 1H, C2H), 7.67 (s, 1H, C4H), 7.65 – 7.53 (m, 2H, C16H), 7.20 (s, 3H, N31H), 7.05 – 6.93 (m, 4H, C17,20H), 5.21 (s, 2H, C21H), 5.10 (s, 2H, C24H), 4.57 (s, 1H, C11H), 2.16 – 2.00 (m, 24H, C29,34H), 1.88 – 1.74 (m, 24H, C28,33H). **^{13}C NMR** (151 MHz, DMSO- d_6) δ 174.5 (C35), 172.0 (C30), 170.0 (C13), 165.2 (C25), 164.4 (C6,12), 159.9 (C3), 152.6 (C18,19), 142.0 (C22), 134.8 (C1,5,15), 126.1 (C23), 121.4 (C17), 119.9 (C16), 118.8 (C20), 116.5 (C2,4), 111.7 (C37), 61.4 (C21), 57.6 (C27), 56.3 (C32), 51.8 (C24), 40.4 (C10), 34.2 (C8), 30.9 (C29), 30.4 (C9), 30.1 (C28), 29.0 (C34), 28.1 (C33). **LRMS** for $\text{C}_{264}\text{H}_{329}\text{N}_{40}\text{O}_{108}$ $[\text{M}-3\text{H}]^{3-}$ Calculated m/z = 1930 Found m/z = 1930.

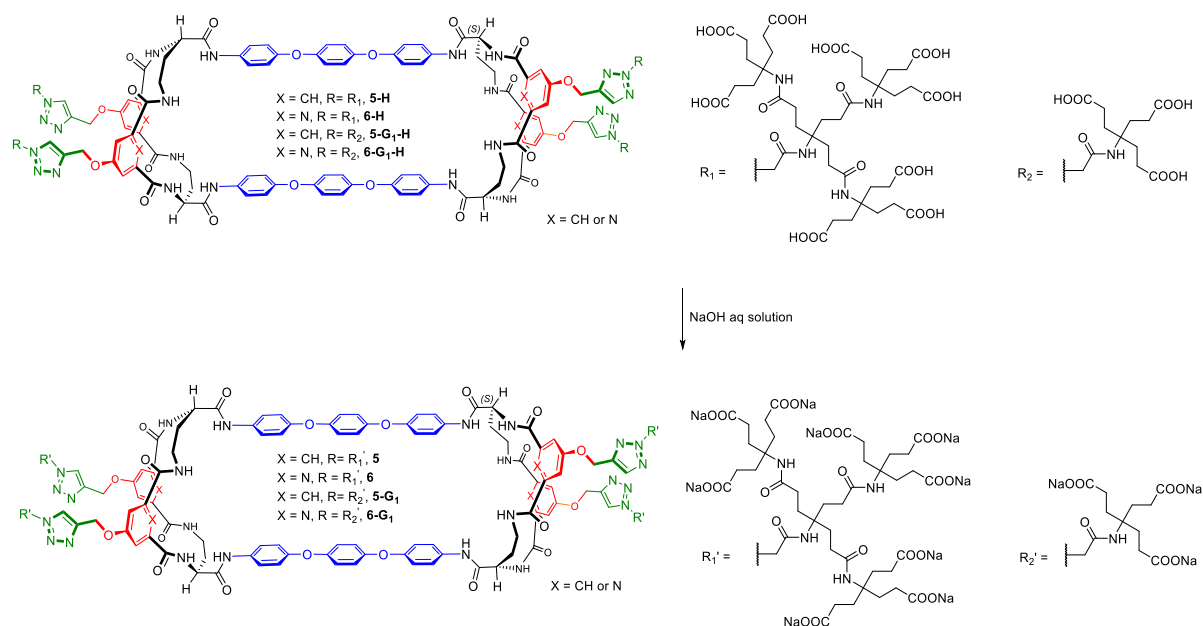
Dipicolinoyl G₂-acid tricycle 6-H



To a solution of dipicolinoyl tricycle **21** (15 mg, 8.7 μmol , 1.0 eqv) in de-gassed MeCN and DMF (3 ml + 3 ml) was added G₂ azido tri-acid **22** (53 mg, 52 μmol , 6 eqv) and 2,6-lutidine (10 mg, 52 μL , 10 eqv). The solution was stirred at room temperature for 15 minutes before tetrakis(acetonitrile)copper(I) hexafluorophosphate (19 mg, 52 μmol , 6 eqv) was added. The reaction mixture was stirred at 60 °C for overnight before the solvent was removed *in vacuo*. The residue was then purified by reverse phase column chromatography (0.01 M aqueous HCl:acetone = 40:60) to give the title compound (23 mg, 3.9 μmol , 45 %) as a colourless oil.

¹H NMR (500 MHz, DMSO-*d*₆) δ 10.30 (s, 1H, N14H), 9.53 (s, 2H, N7,11H), 8.15 (s, 1H, C23H), 7.82 (s, 1H, N26H), 7.67 (d, *J* = 8.7 Hz, 2H, C16H), 7.56 (d, *J* = 10.7 Hz, 2H, C2,4H), 7.24 (s, 3H, N31H), 7.00 (d, *J* = 8.6 Hz, 2H, C17H), 6.95 (s, 2H, C20H), 5.31 (s, 2H, C21H), 5.10 (s, 2H, C24H), 4.85 (s, 1H, N10H), 4.07 – 3.98 (m, 1H, C8H), 2.93 – 2.86 (m, 1H, C8H), 2.17 – 2.02 (m, 26H, C9,28,33H), 1.87 – 1.73 (m, 24H, C29,34H). **¹³C NMR** (126 MHz, DMSO-*d*₆) δ 174.9 (C35), 172.5 (C30), 170.5 (C13), 166.4 (C25), 164.9 (C3), 163.2 (C6), 163.0 (C12), 153.3 (C18), 152.7 (C19), 151.3 (C5), 150.8 (C1), 141.7 (C22), 135.3 (C15), 126.9 (C23), 121.7 (C16), 119.8 (C17), 119.7 (C20), 110.8 (C2,4), 62.0 (C21), 58.0 (C27), 56.8 (C32), 55.5 (C10), 52.3 (C24), 36.3 (C8), 33.8 (C9), 31.4 (C29), 30.6 (C28), 29.5 (C34), 28.5 (C33). **HRMS** C₂₆₀H₃₂₄N₄₄O₁₀₈ [M-4H]⁴⁻ Calculated *m/z* = 1448.2826 Found *m/z* = 1448.2838.

Water soluble isophthaloyl/dipicolinoyl G₁/G₂ tricycles **5**, **6**, **5-G₁** and **6-G₁**

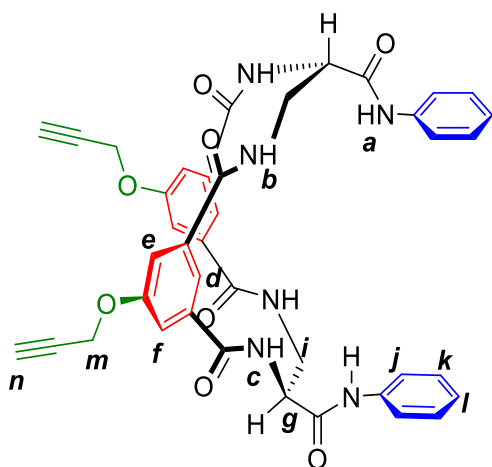


Supplementary Scheme 6. Synthesis of receptors **5**, **6**, **5-G₁** and **6-G₁**.

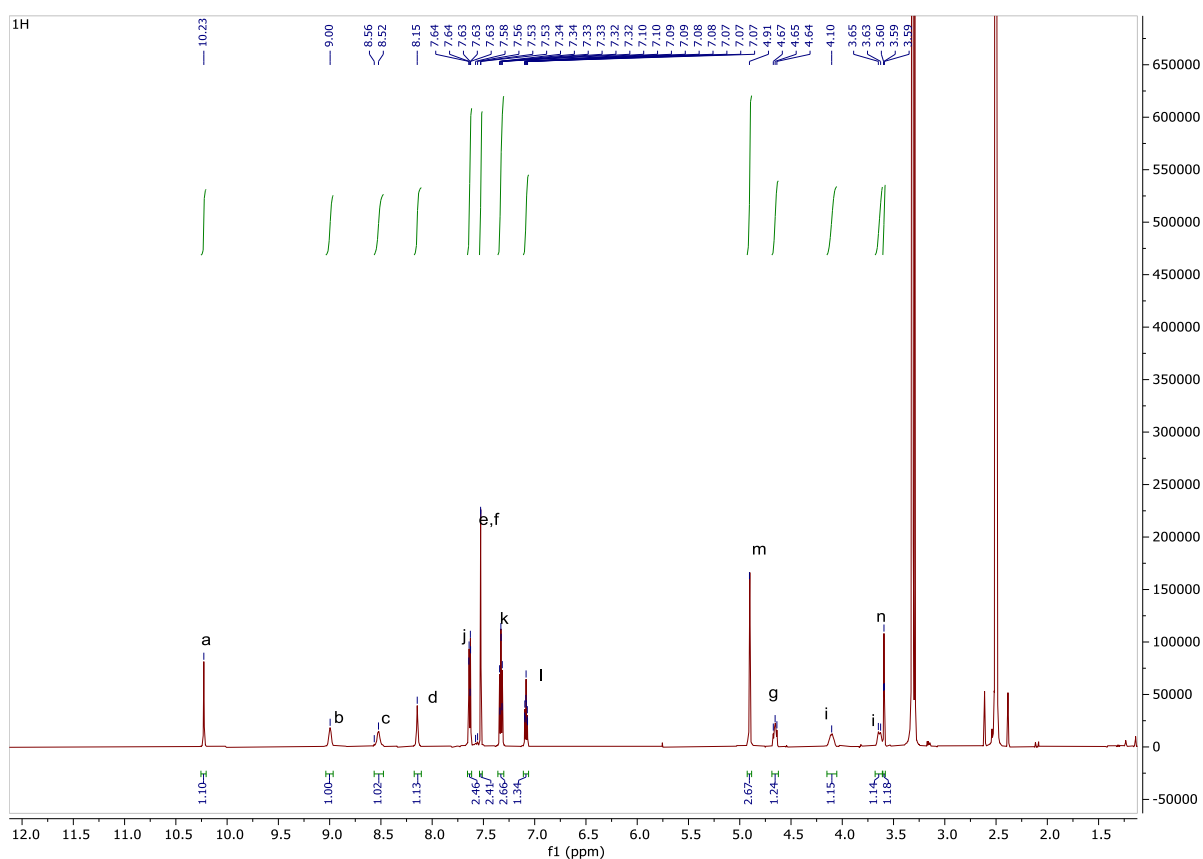
To a suspension of isophthaloyl/dipicolinoyl G₁/G₂ acid tricycle in water (10 ml) was added 10 mM NaOH aq solution to adjust the pH to ~7.4. The resulting solution was freeze-dried to yield the water soluble isophthaloyl/dipicolinoyl G₁/G₂ tricycle (quantitative yield) in their sodium form as white solids. Samples for characterisation and further studies were prepared by dissolution in D₂O or 9:1 H₂O/D₂O. The process of freeze-drying and dissolution was found to have no effect on the pH.

1.6 Characterisation of receptors in their operating environments

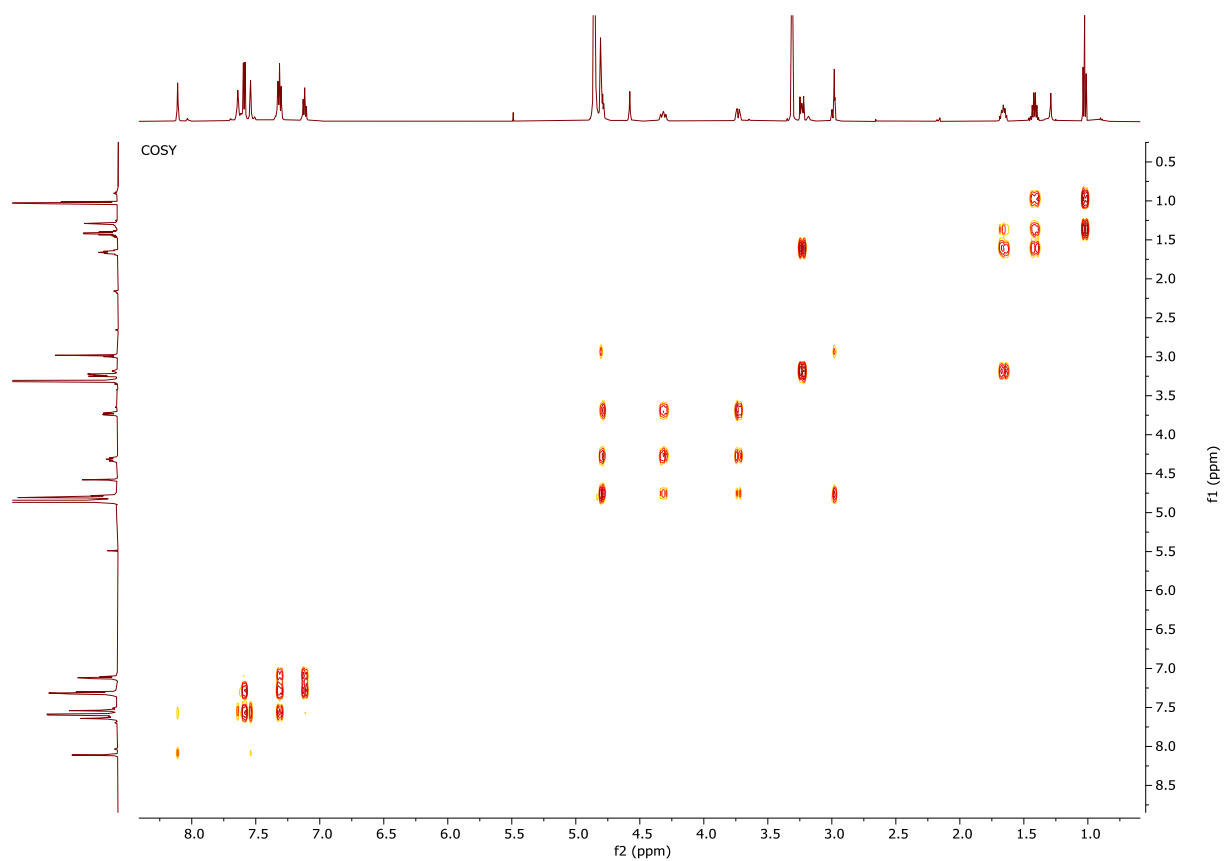
Receptor 8a



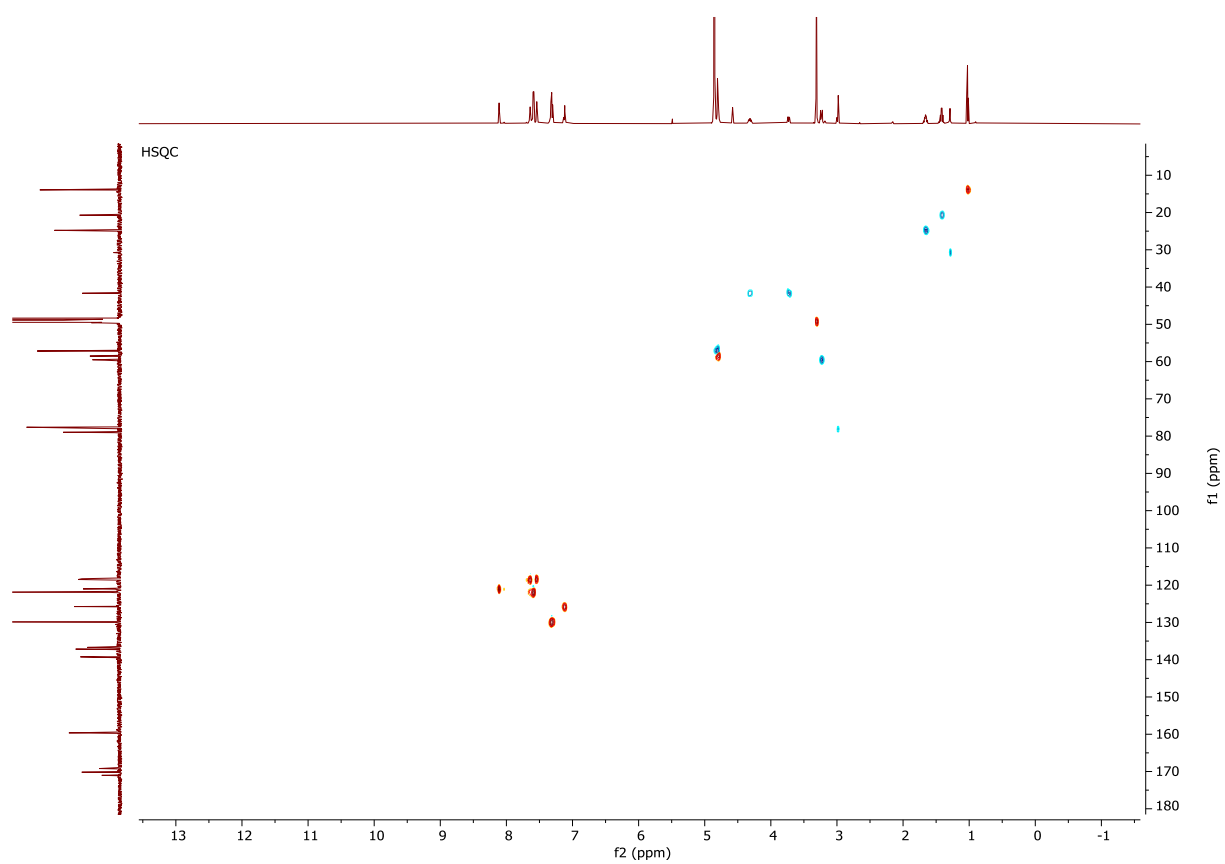
The assignment of the NMR spectrum in DMSO- d_6 was made with the help of 2D COSY and HSQC (see Supplementary Figures below).



Supplementary Figure 1. ^1H NMR spectrum (600 MHz) of receptor **8a** in DMSO- d_6 .

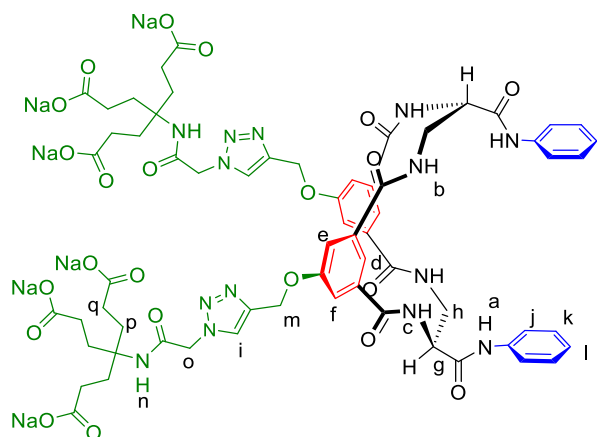


Supplementary Figure 2. 2D COSY NMR spectrum (600 MHz) of receptor **8a** in DMSO- d_6 .

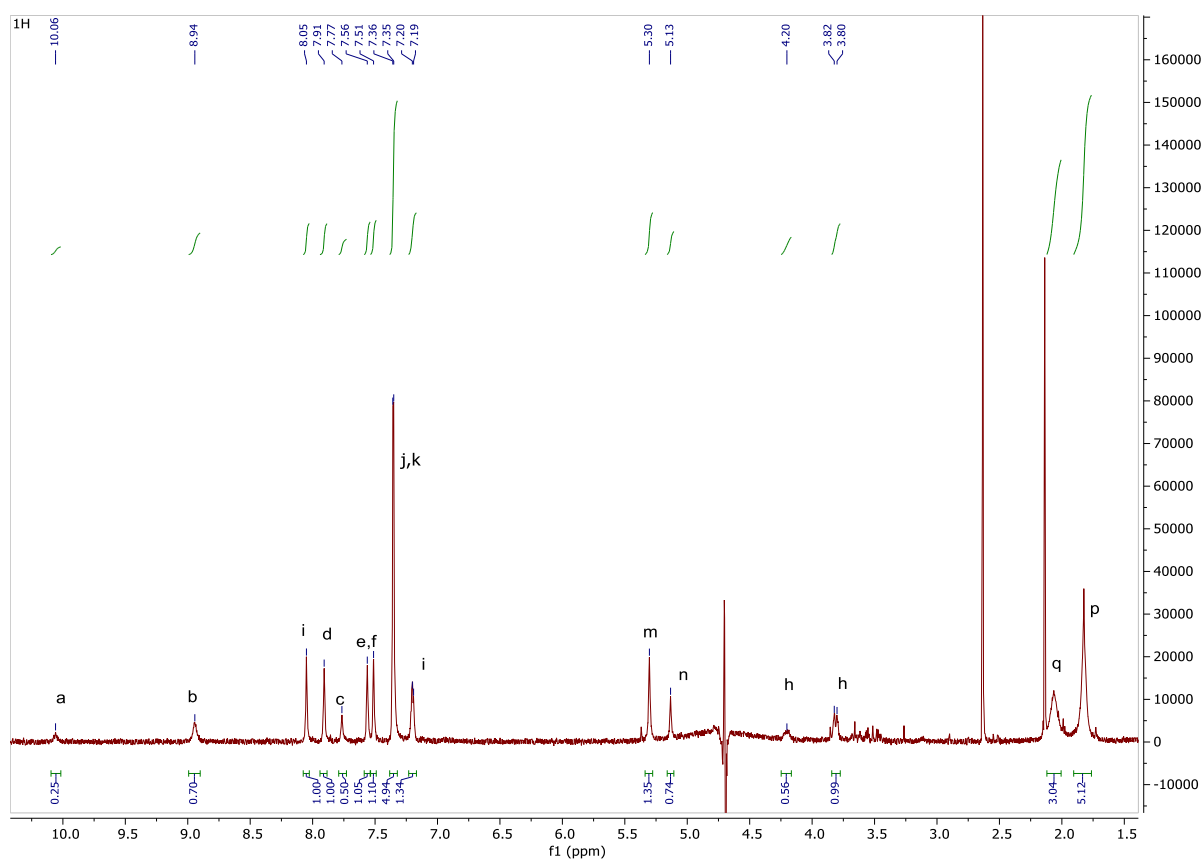


Supplementary Figure 3. 2D HSQC NMR spectrum (600 MHz) of receptor **8a** in DMSO- d_6 .

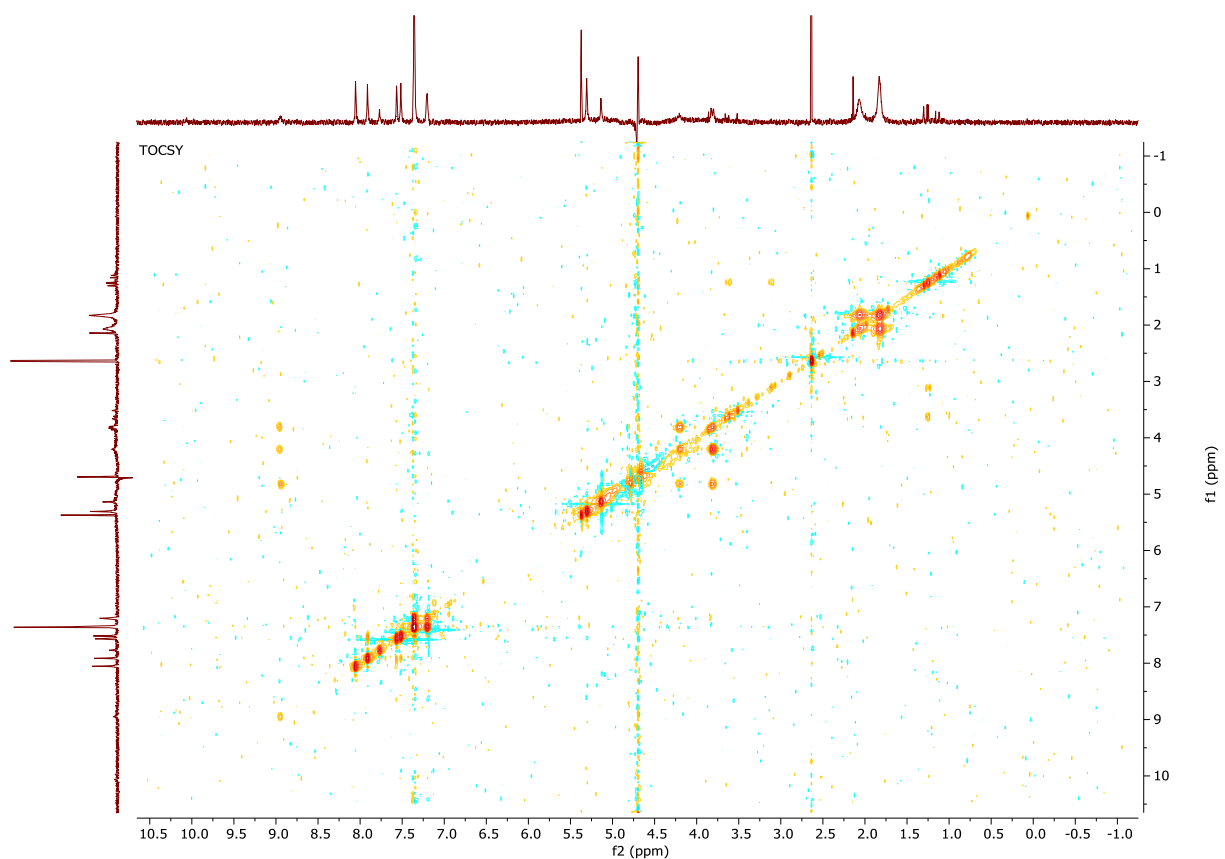
Receptor 8b



The receptor was characterized in H₂O/D₂O. The assignment of the NMR spectrum was made with the help of 2D TOCSY (see Supplementary Figures below)).

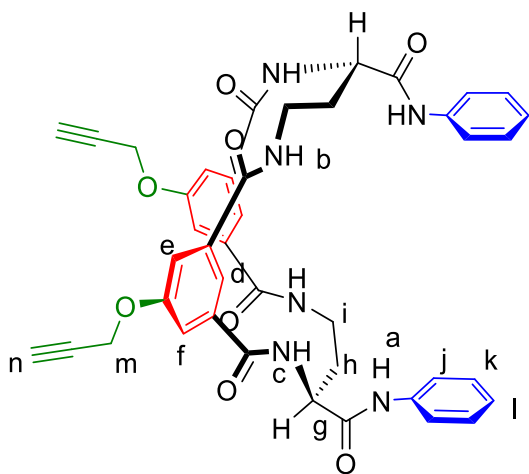


Supplementary Figure 4. ¹H NMR spectrum (600 MHz) of receptor **8b** in 9:1 H₂O/D₂O.

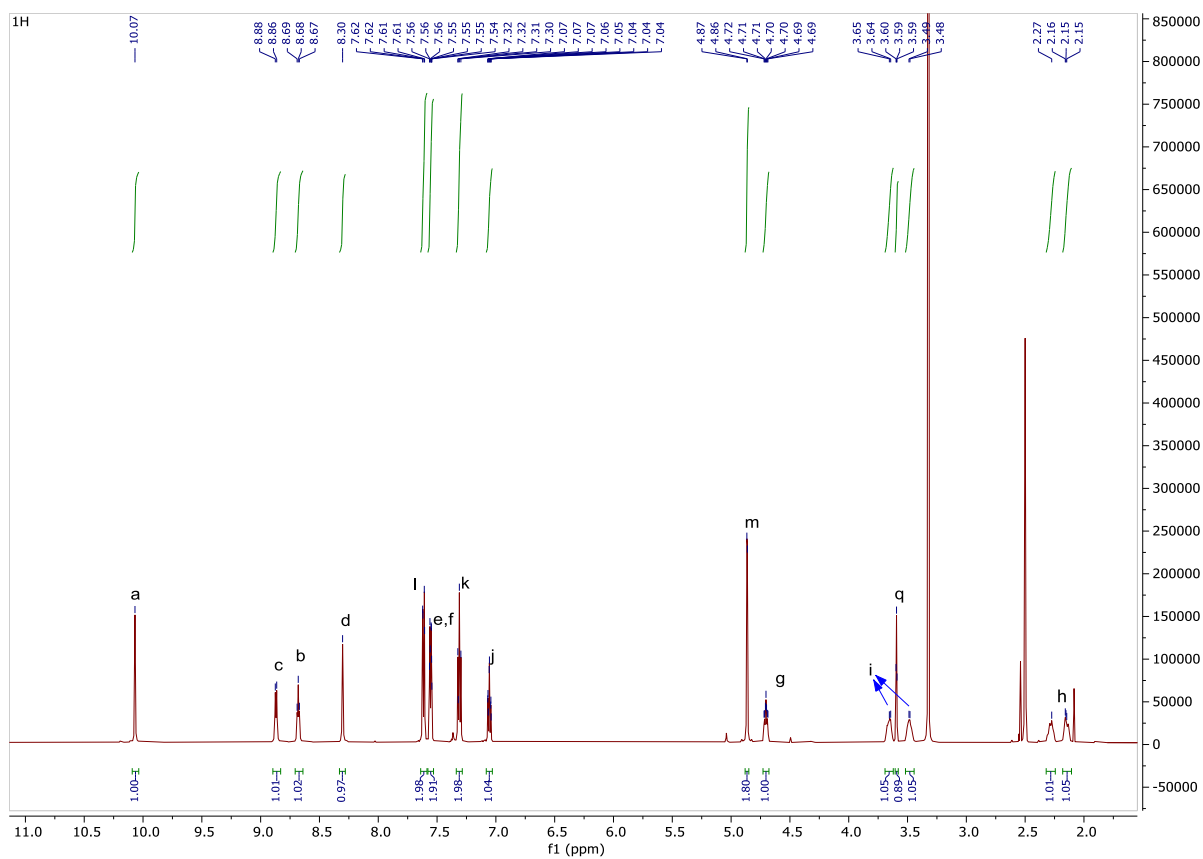


Supplementary Figure 5. 2D TOCSY NMR spectrum (600 MHz) of receptor **8b** in 9:1 H₂O/D₂O.

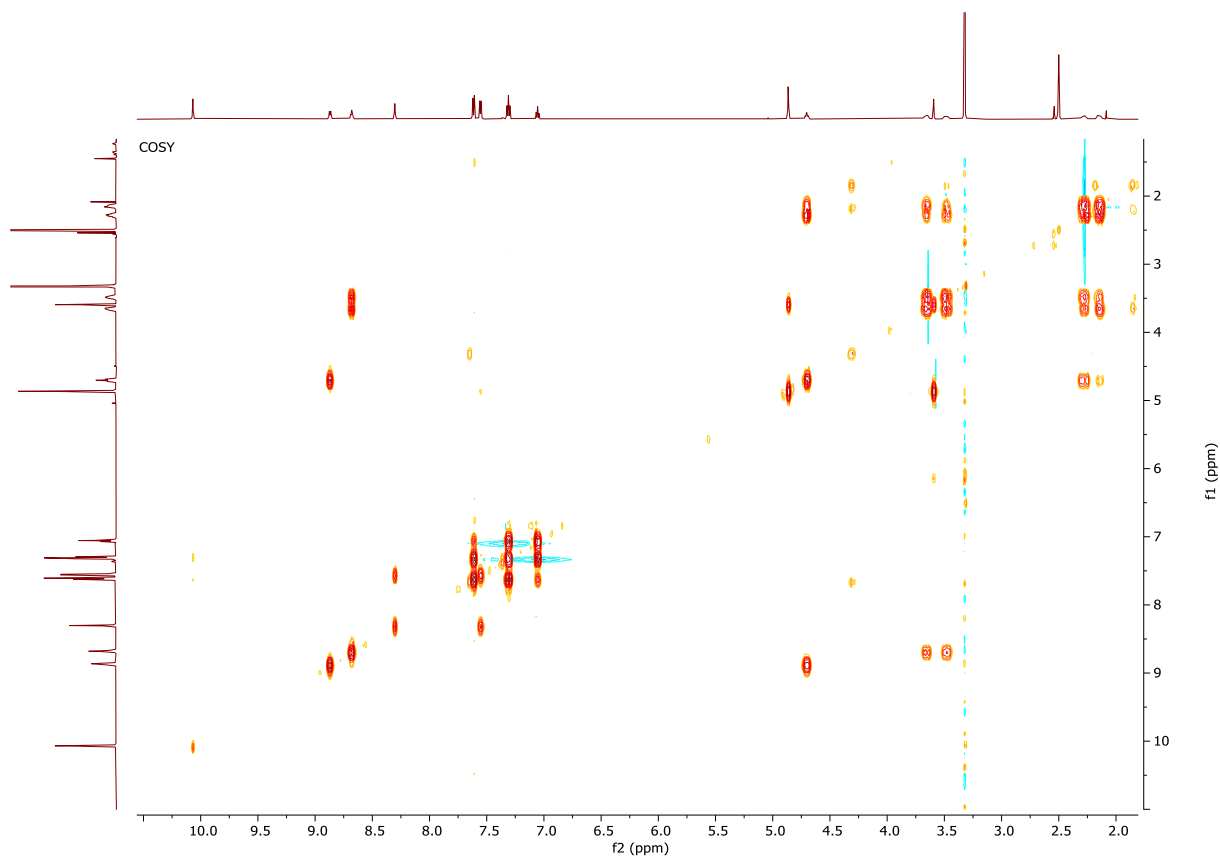
Receptor 9a



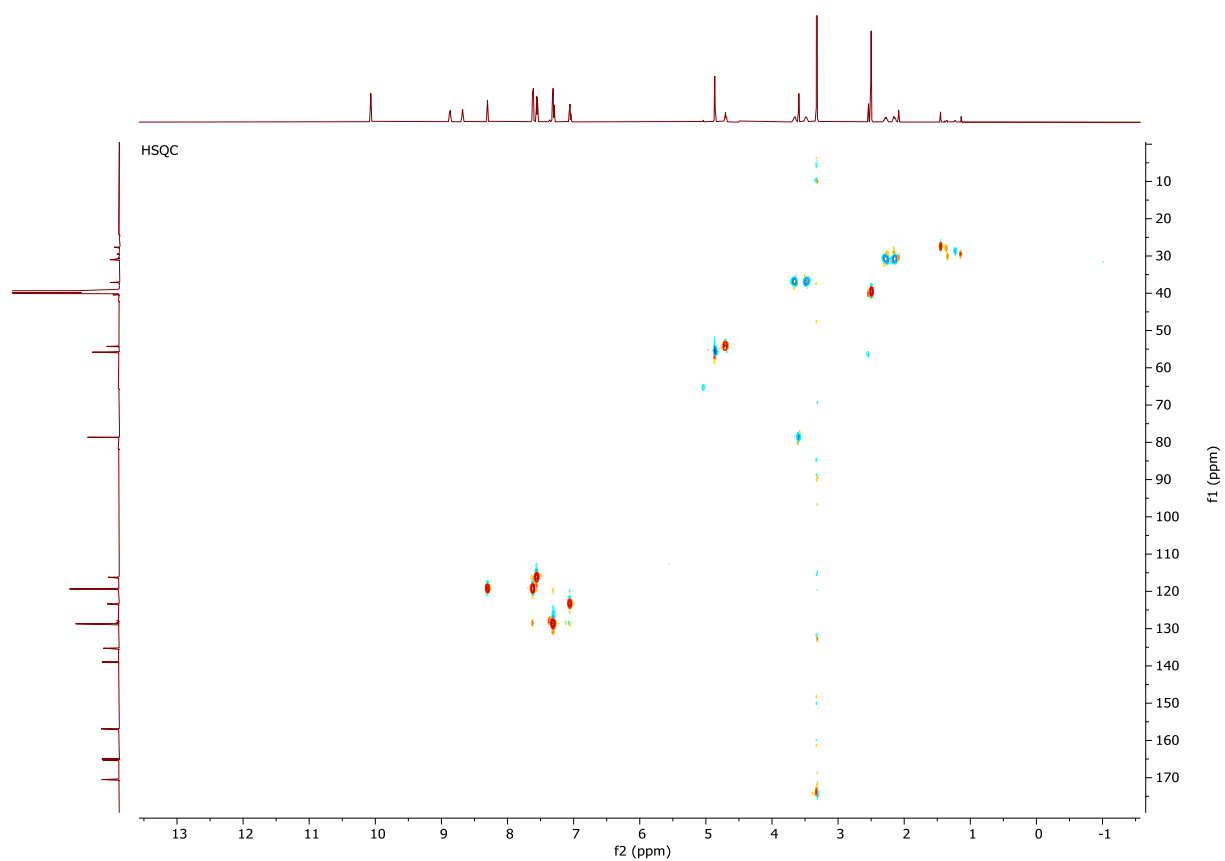
The assignment of the NMR spectrum in DMSO- d_6 was made with the help of 2D COSY and HSQC (see Supplementary Figures below).



Supplementary Figure 6. ¹H NMR spectrum (600 MHz) of receptor **9a** in DMSO-*d*₆.

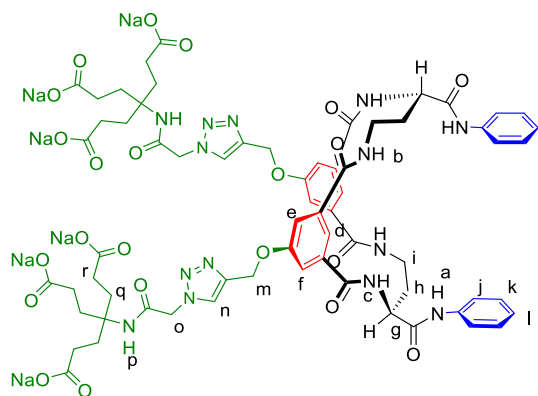


Supplementary Figure 7. 2D COSY NMR spectrum (600 MHz) of receptor **9a** in DMSO-*d*₆.

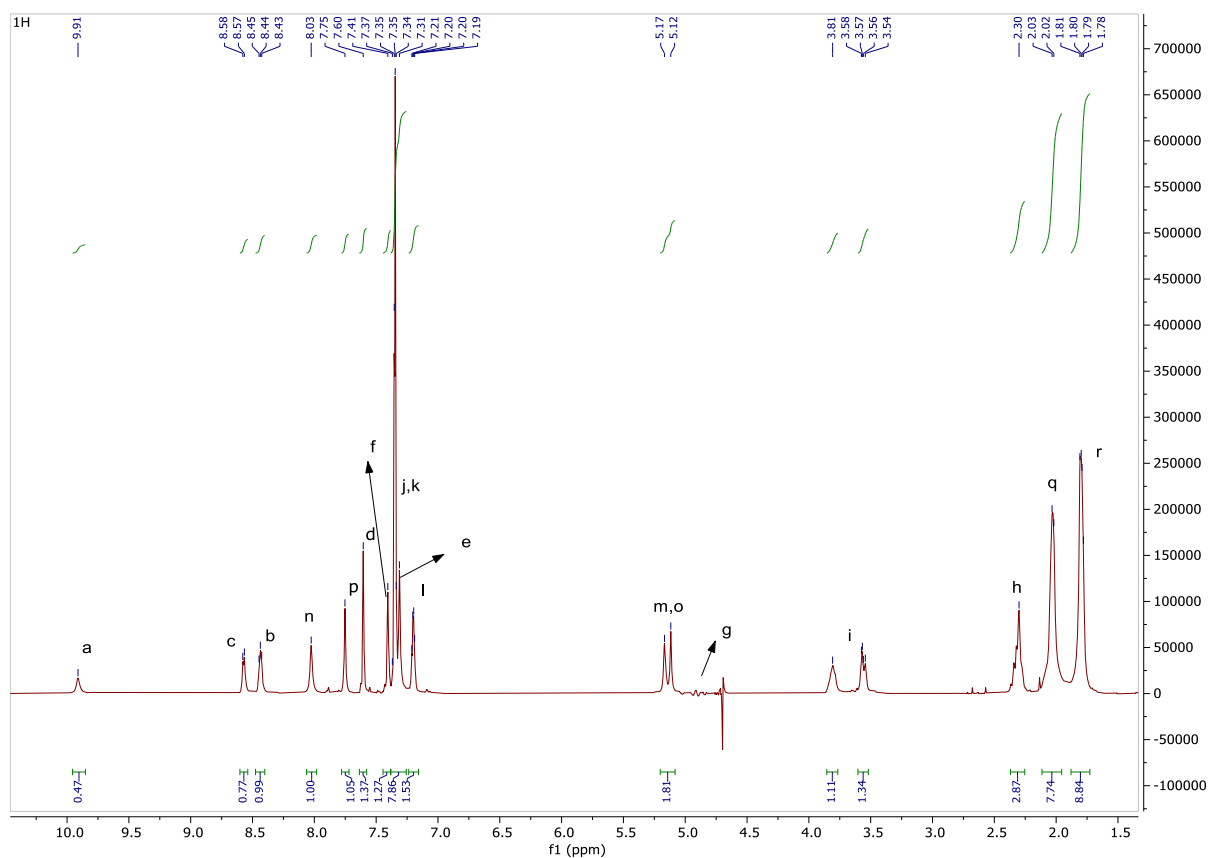


Supplementary Figure 8. 2D HSQC NMR spectrum (600 MHz) of receptor **9a** in DMSO-*d*₆.

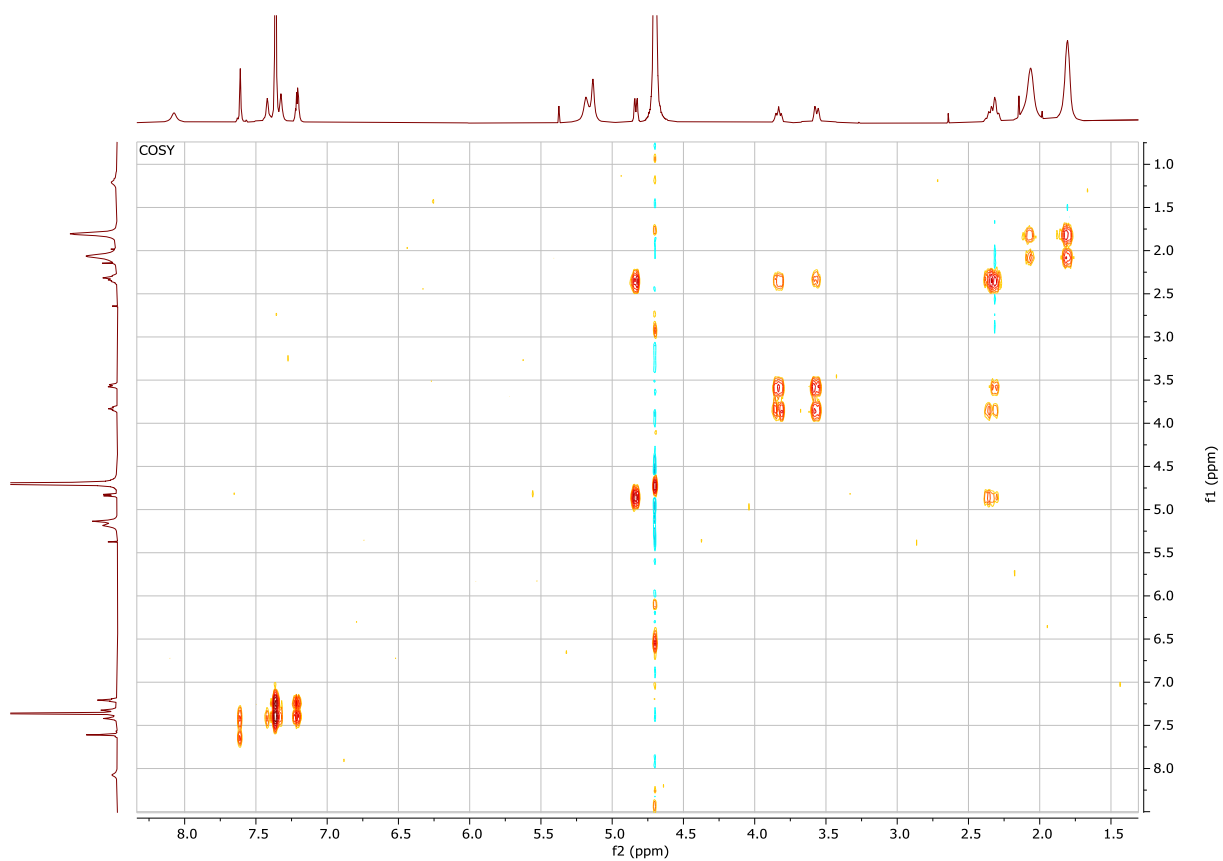
Receptor 9b



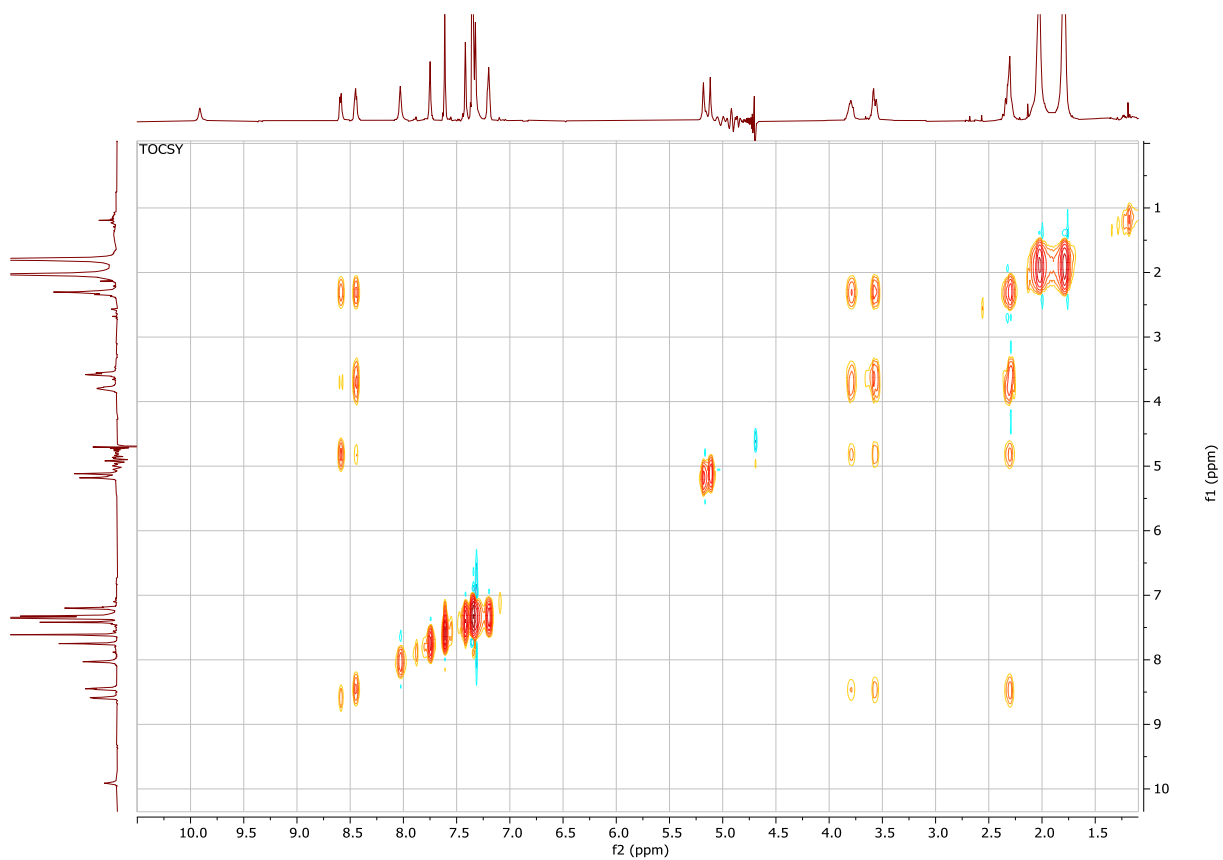
The receptor was characterized in 100 % D₂O and 9:1 H₂O/D₂O. The assignment of the NMR spectrum was made with the help of 2D COSY, TOCSY and HSQC (see Supplementary Figures below).



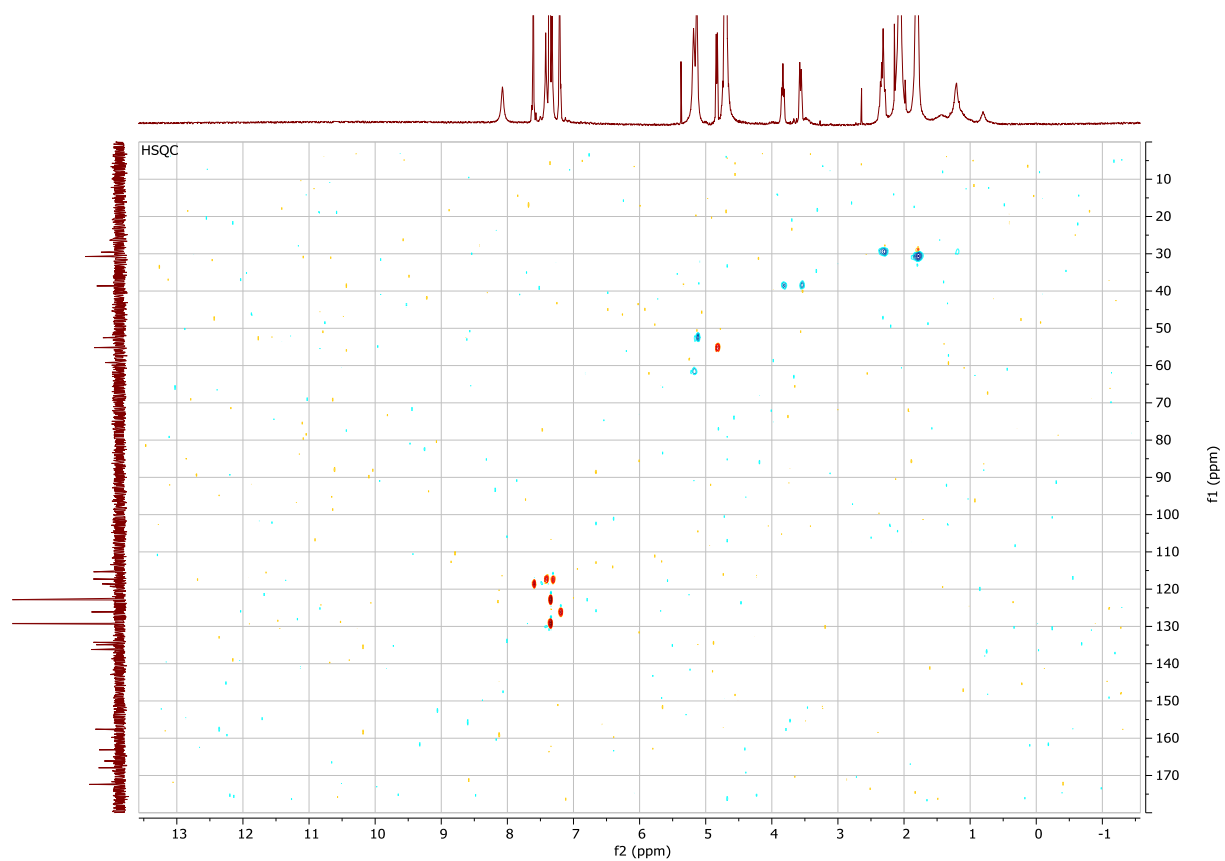
Supplementary Figure 9. ¹H NMR spectrum (600 MHz) of receptor 9b in 9:1 H₂O/D₂O.



Supplementary Figure 10. 2D COSY NMR spectrum (600 MHz) of receptor **9b** in D₂O.

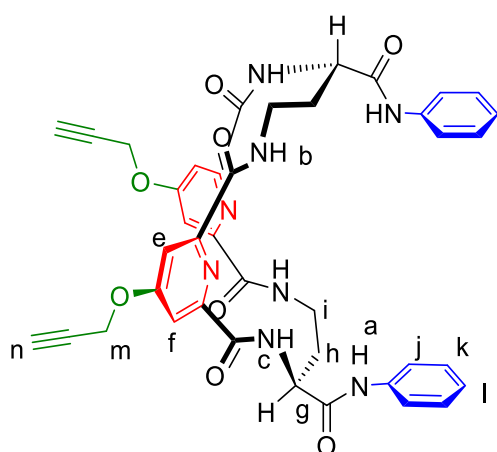


Supplementary Figure 11. 2D TOCSY NMR spectrum (600 MHz) of receptor **9b** in 9:1 H₂O/D₂O.

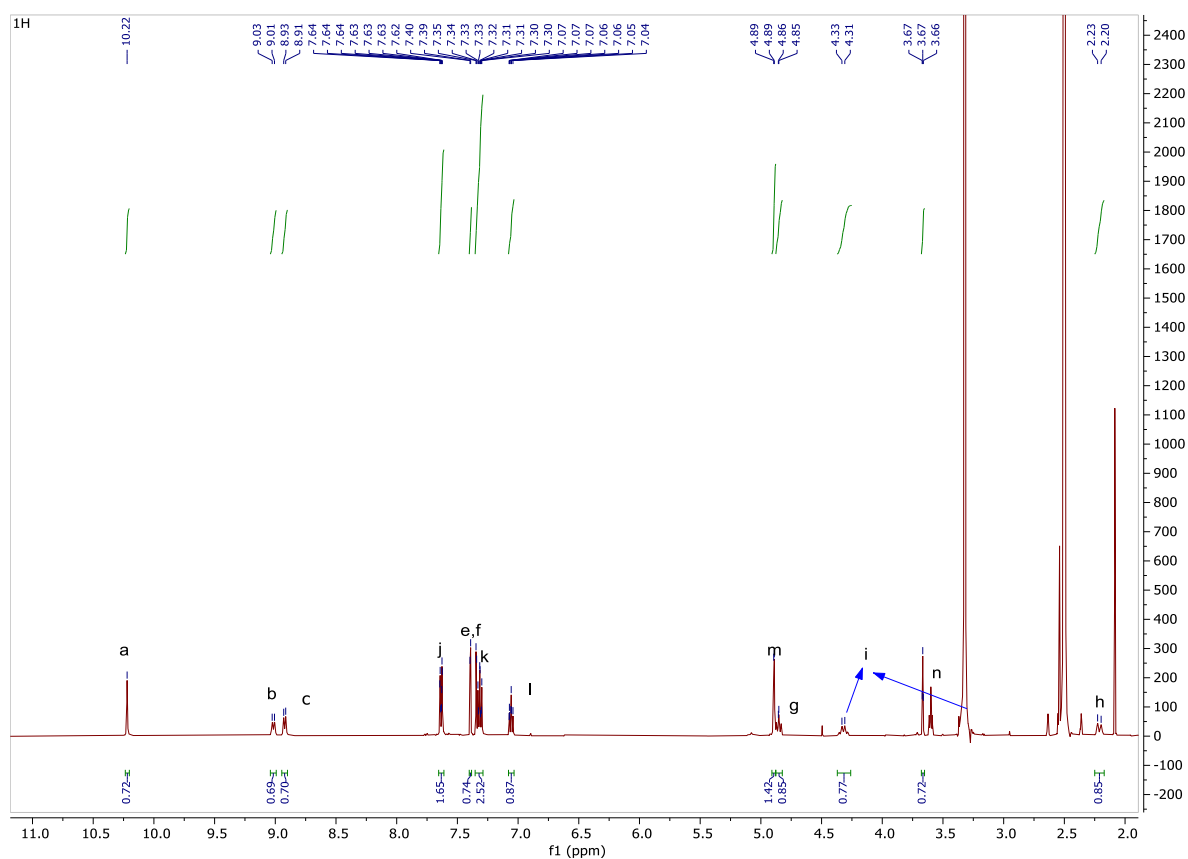


Supplementary Figure 12. 2D HSQC NMR spectrum (600 MHz) of receptor **9b** in D₂O.

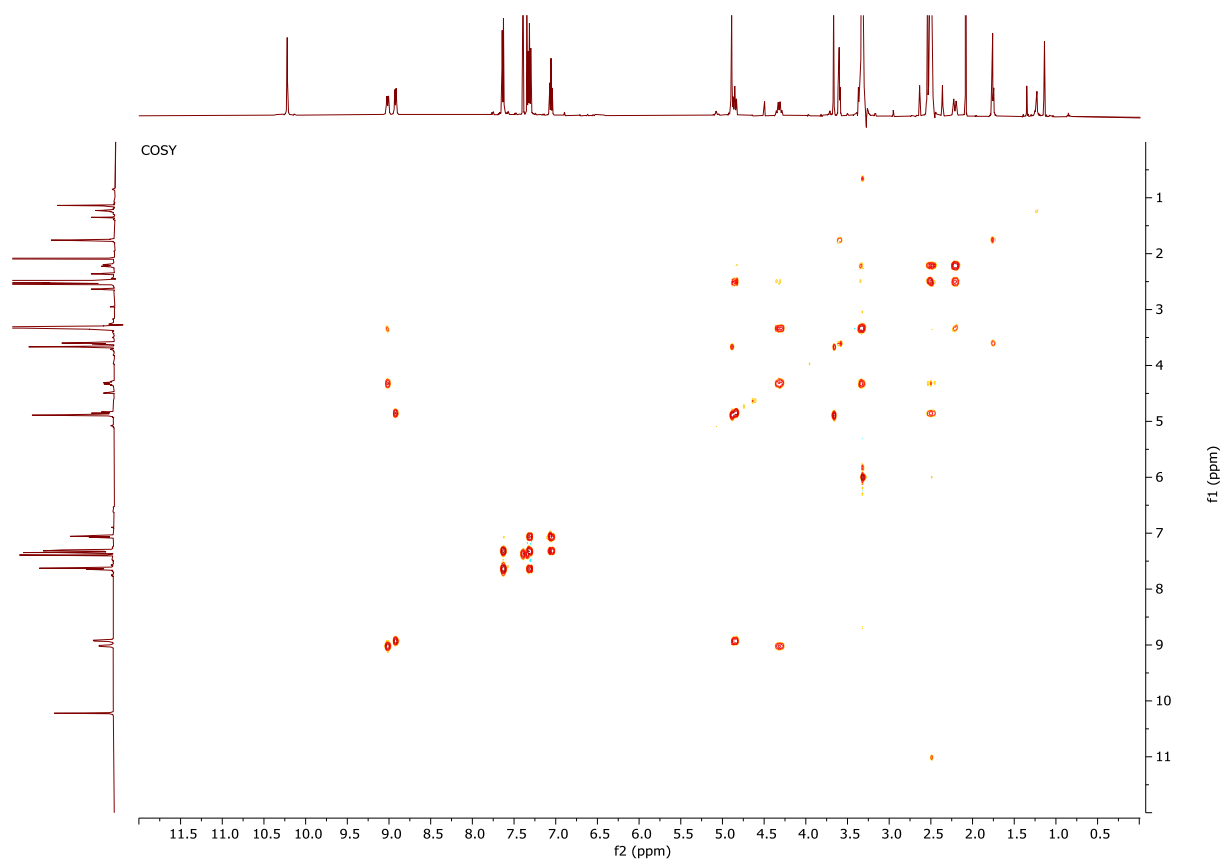
Receptor 10a



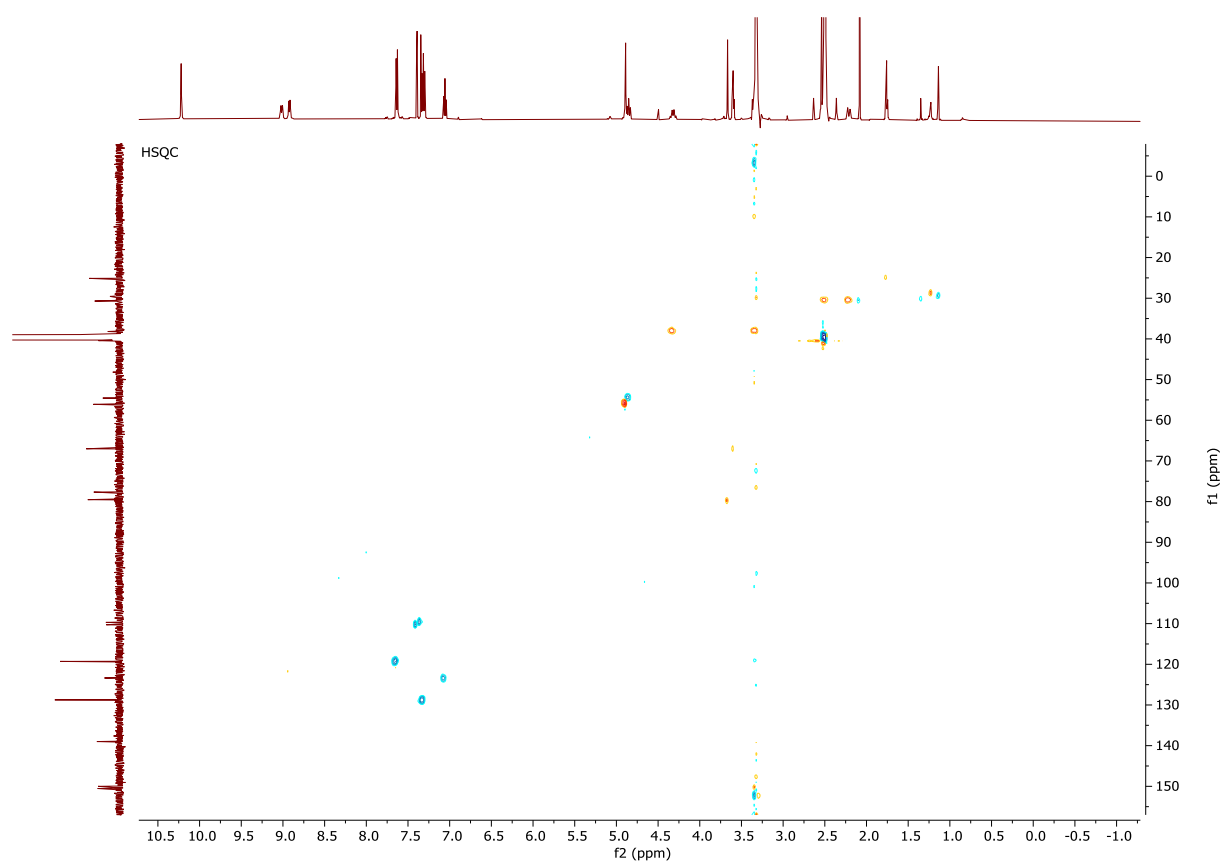
The assignment of the NMR spectrum in DMSO- d_6 was made with the help of 2D COSY and HSQC (see Supplementary Figures below).



Supplementary Figure 13. ^1H NMR spectrum (500 MHz) of receptor **10a** in DMSO- d_6 .

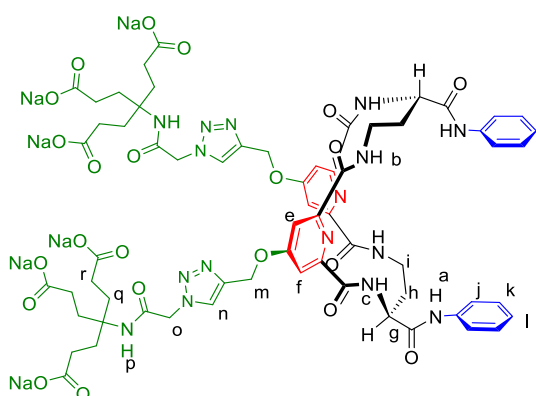


Supplementary Figure 14. 2D COSY NMR spectrum (500 MHz) of receptor **10a** in DMSO- d_6 .

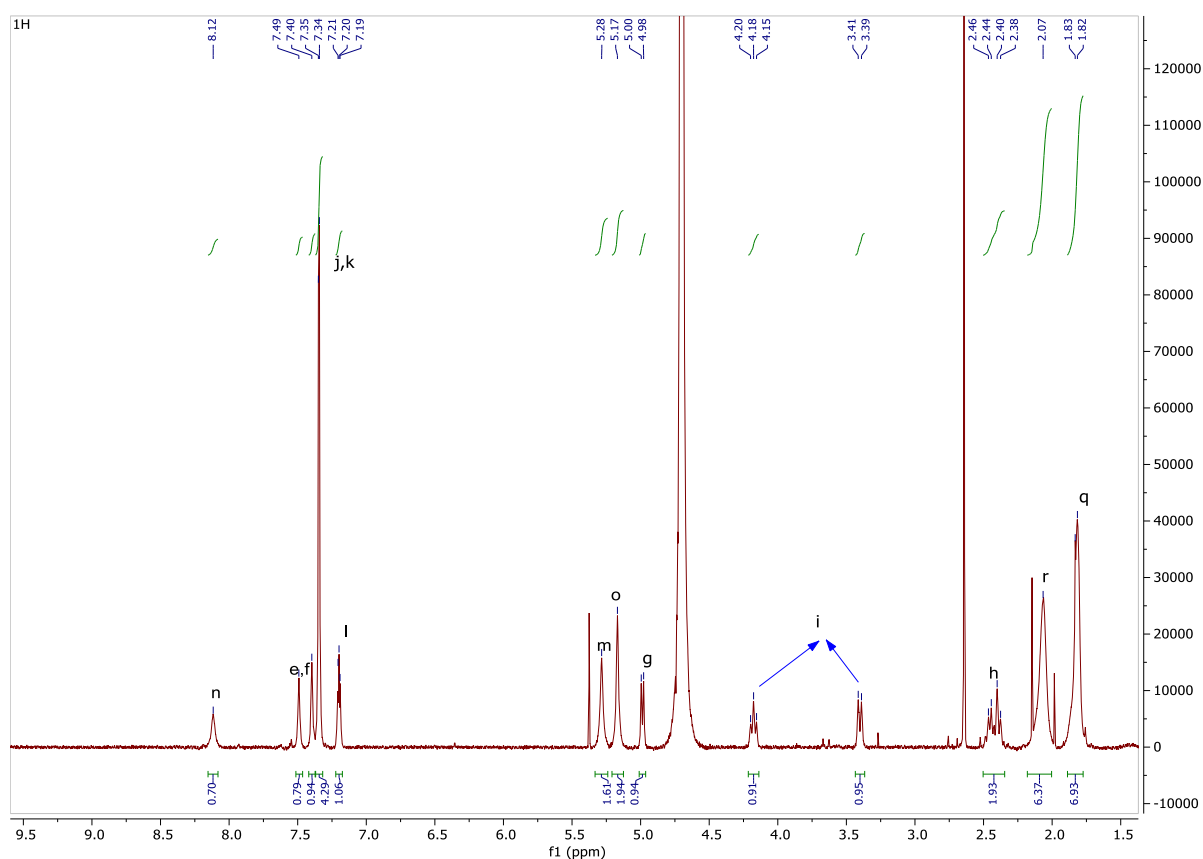


Supplementary Figure 15. 2D HSQC NMR spectrum (500 MHz) of receptor **10a** in DMSO- d_6 .

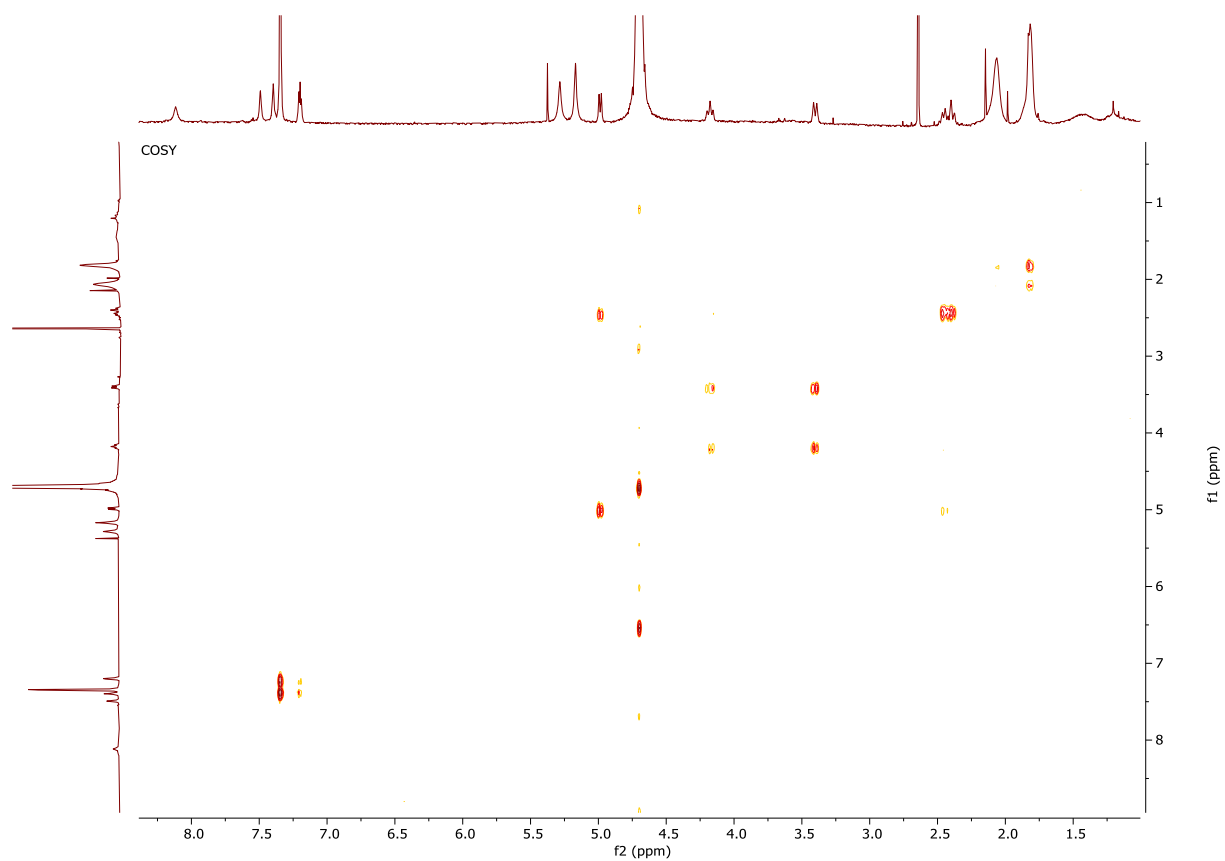
Receptor 10b



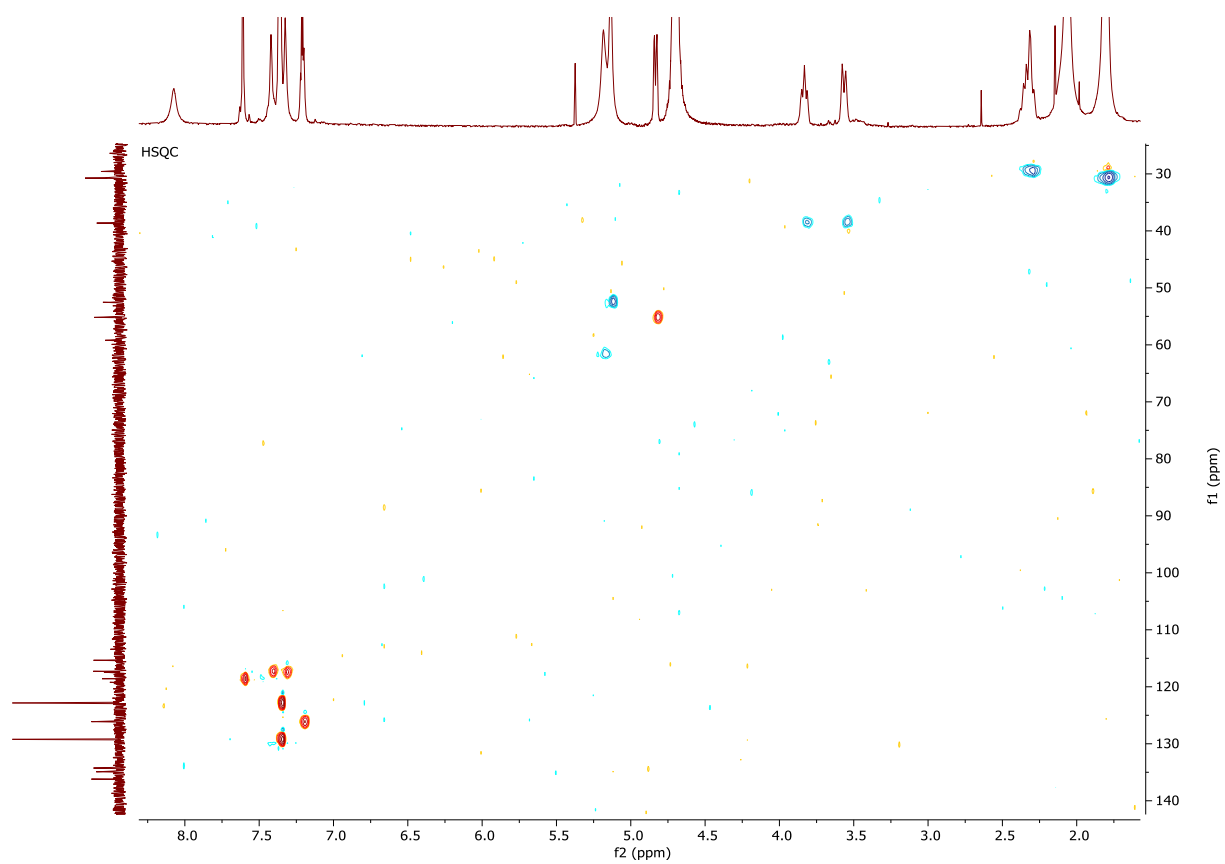
The receptor was characterized in D₂O. The assignment of the NMR spectrum was made with the help of 2D COSY, TOCSY and HSQC (see Supplementary Figures below).



Supplementary Figure 16. ¹H NMR spectrum (600 MHz) of receptor **10b** in D₂O.

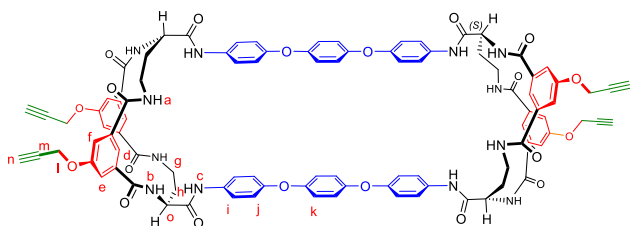


Supplementary Figure 17. 2D COSY NMR spectrum (600 MHz) of receptor **10b** in D₂O.

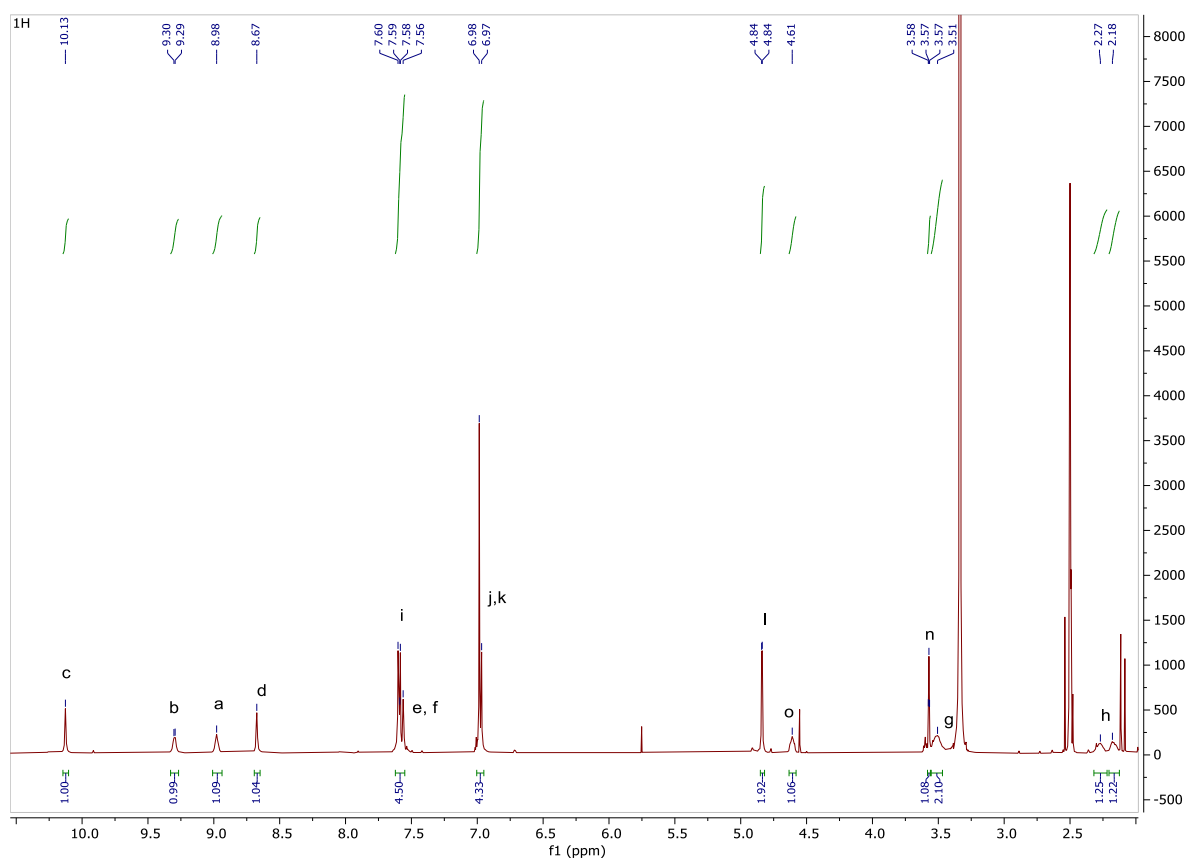


Supplementary Figure 18. 2D HSQC NMR spectrum (600 MHz) of receptor **10b** in D₂O.

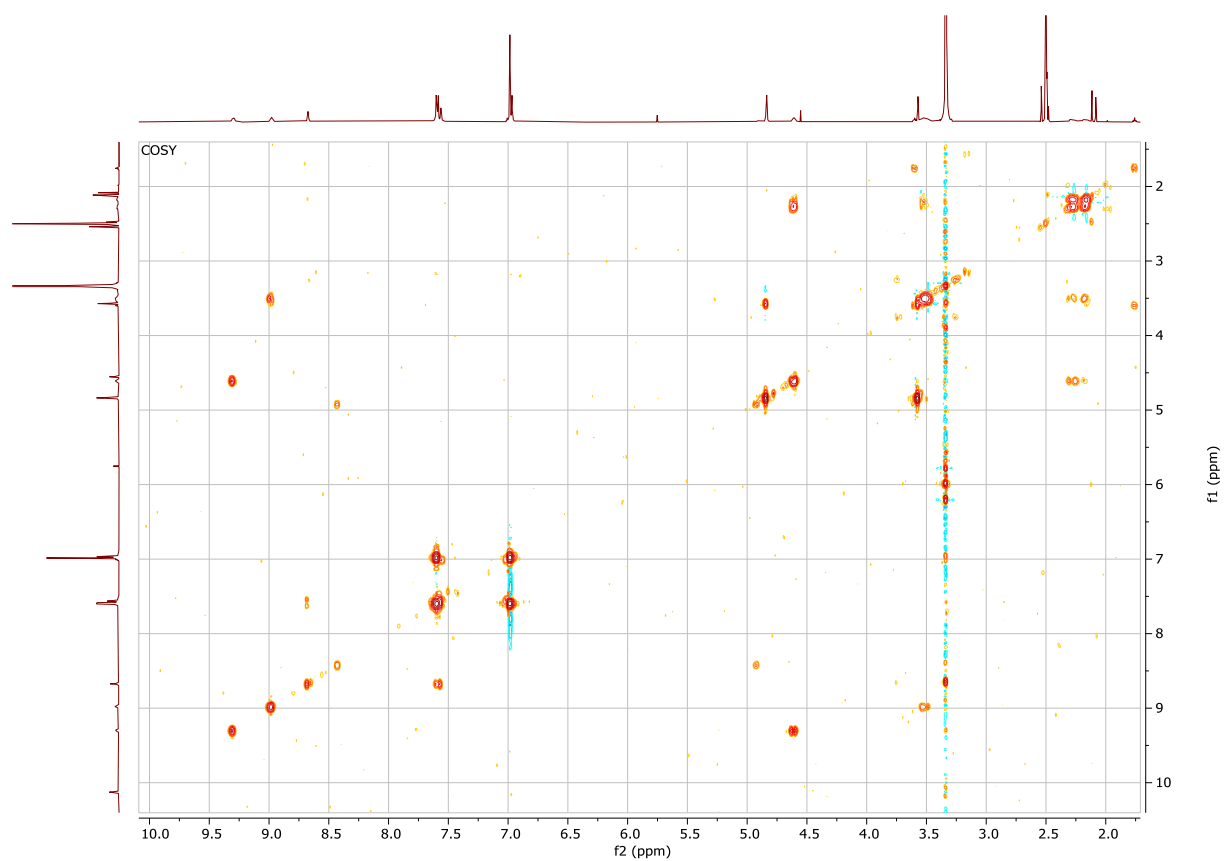
Receptor 20



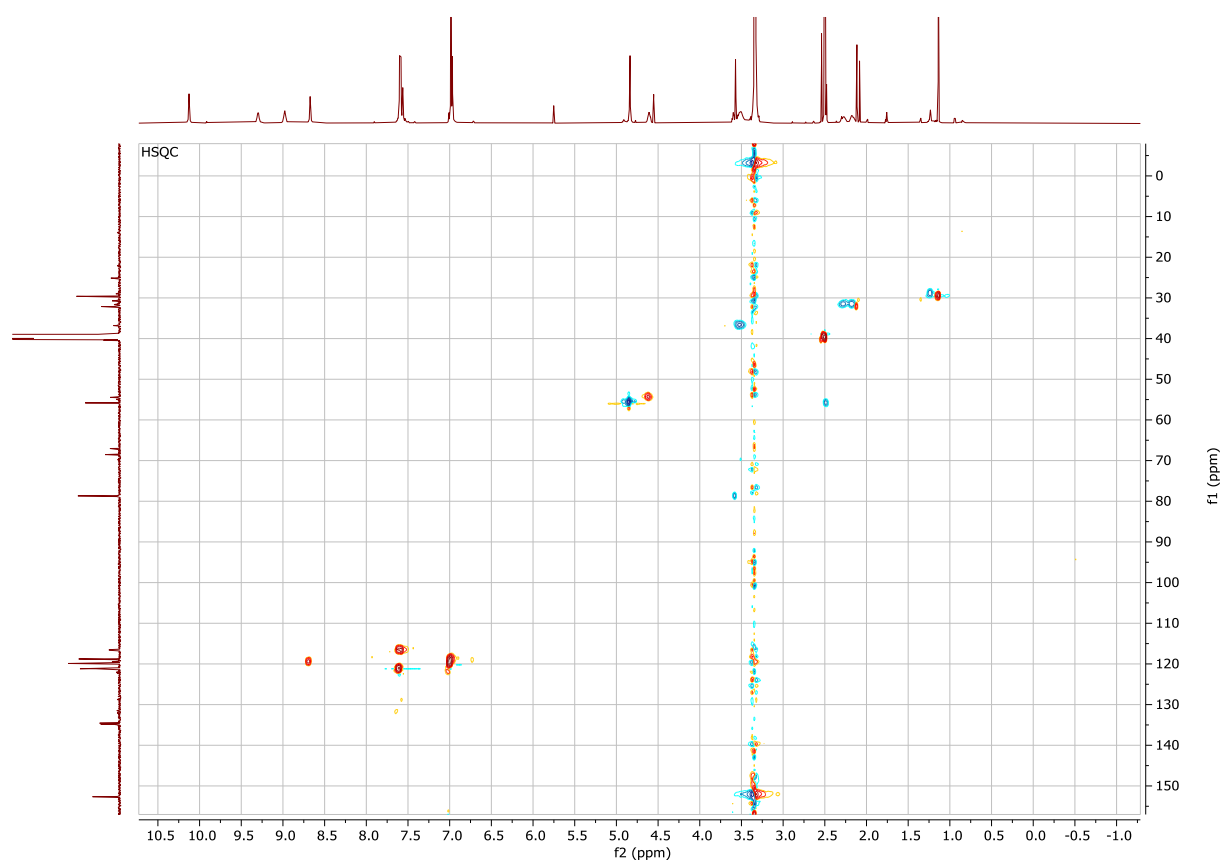
The assignment of the NMR spectrum in DMSO- d^6 was made with the help of 2D COSY and HSQC (see Supplementary Figures below).



Supplementary Figure 19. ^1H NMR spectrum (500 MHz) of receptor 20 in DMSO- d^6 .

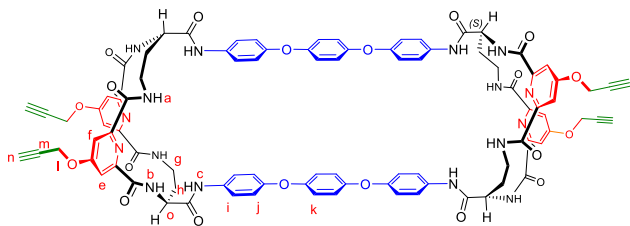


Supplementary Figure 20. 2D COSY NMR spectrum (500 MHz) of receptor **20** in DMSO- d_6 .

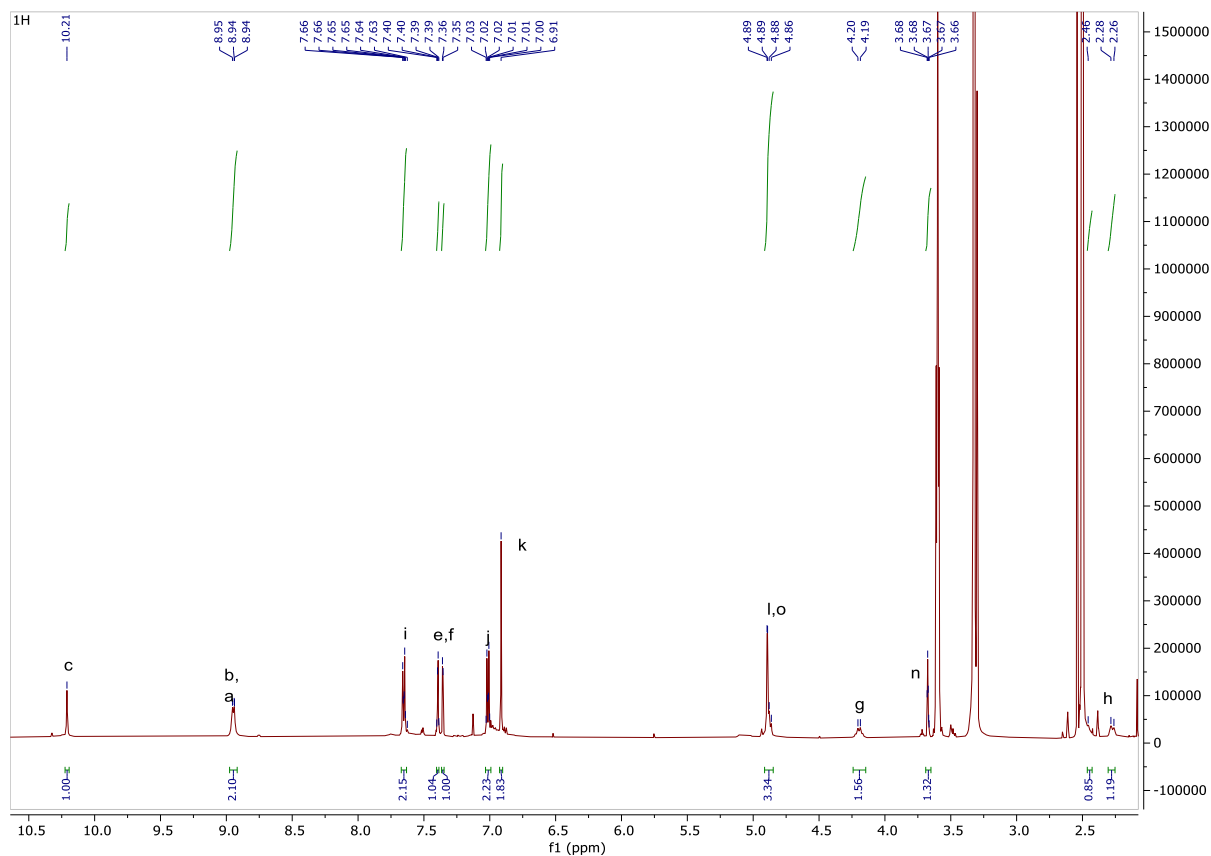


Supplementary Figure 21. 2D HSQC NMR spectrum (500 MHz) of receptor **20** in DMSO- d_6 .

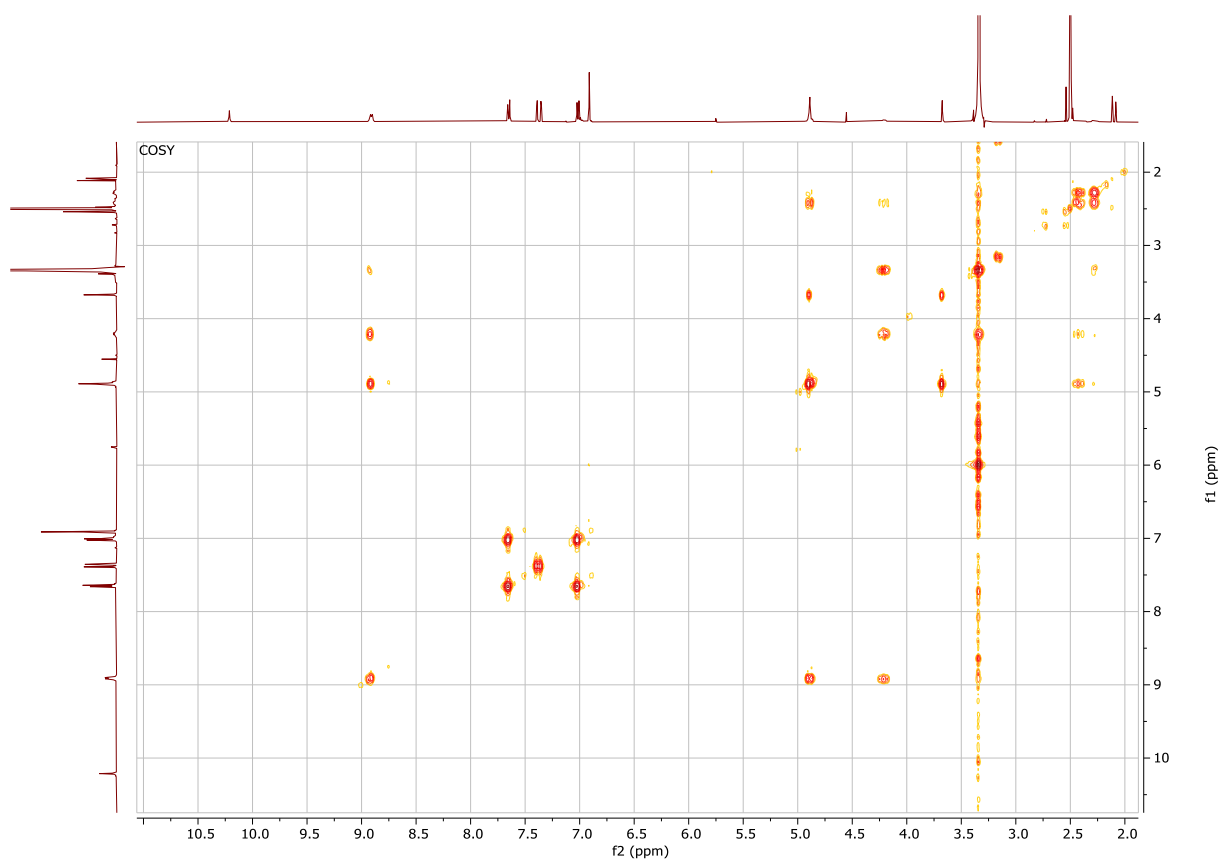
Receptor 21



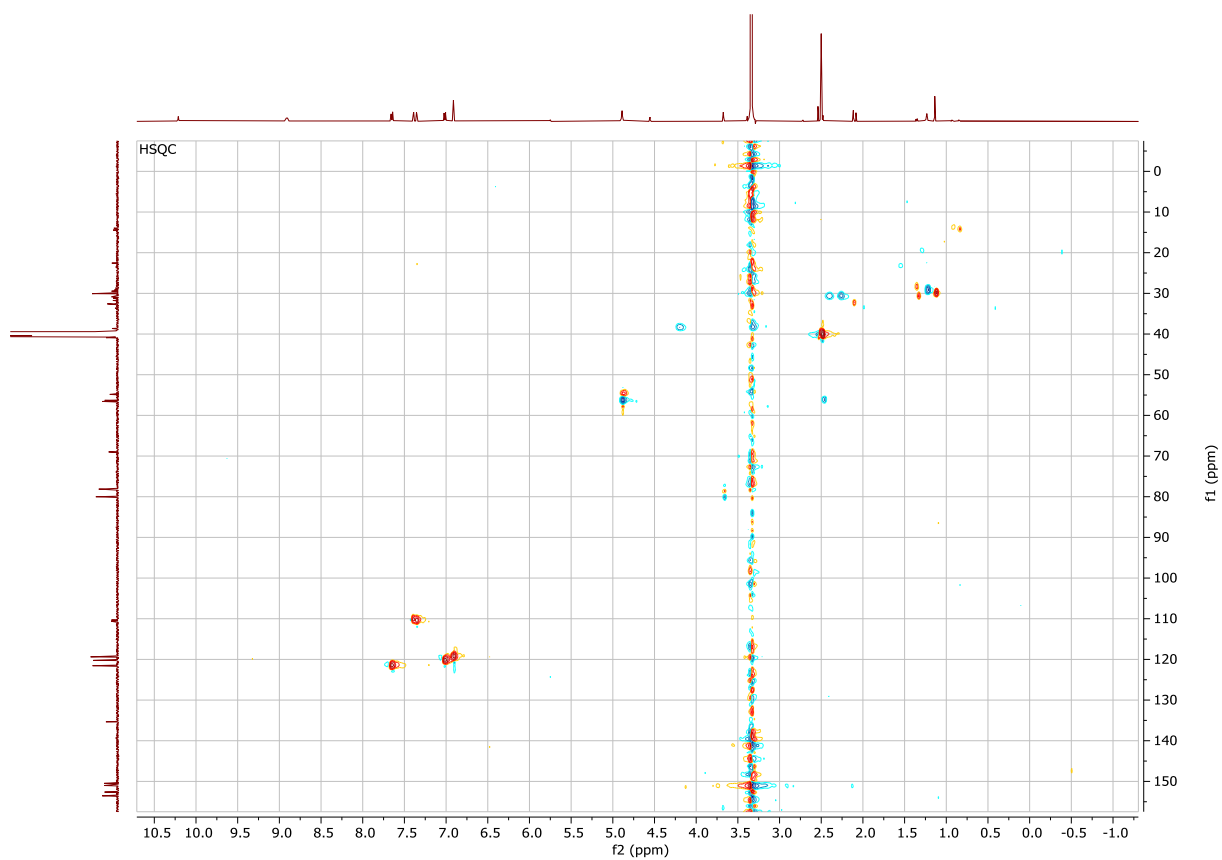
The assignment of the NMR spectrum in DMSO- d^6 was made with the help of 2D COSY and HSQC (see Supplementary Figures below).



Supplementary Figure 22. ^1H NMR spectrum (500 MHz) of receptor **21** in DMSO- d^6 .



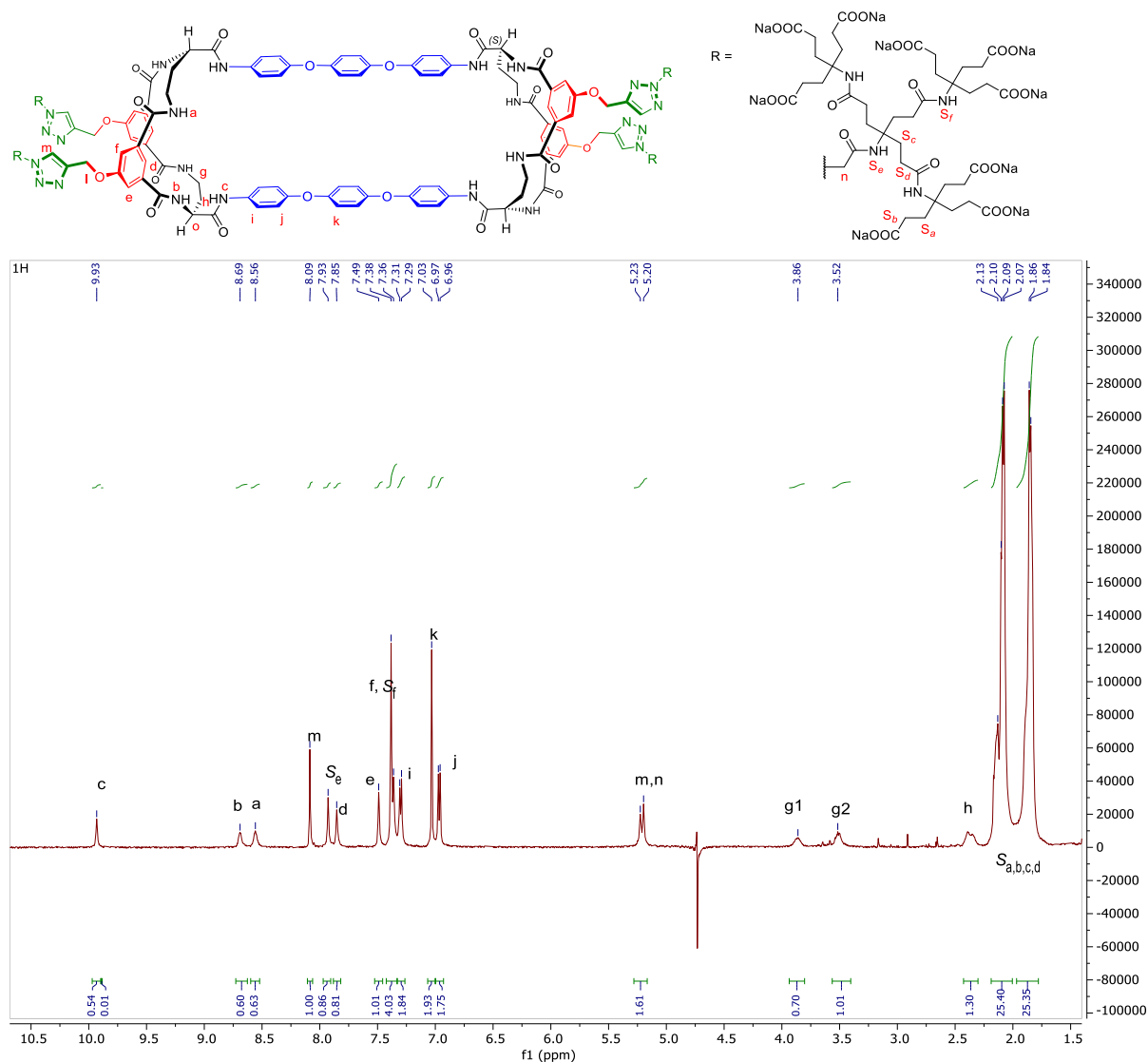
Supplementary Figure 23. 2D COSY NMR spectrum (500 MHz) of receptor **21** in DMSO- d_6 .



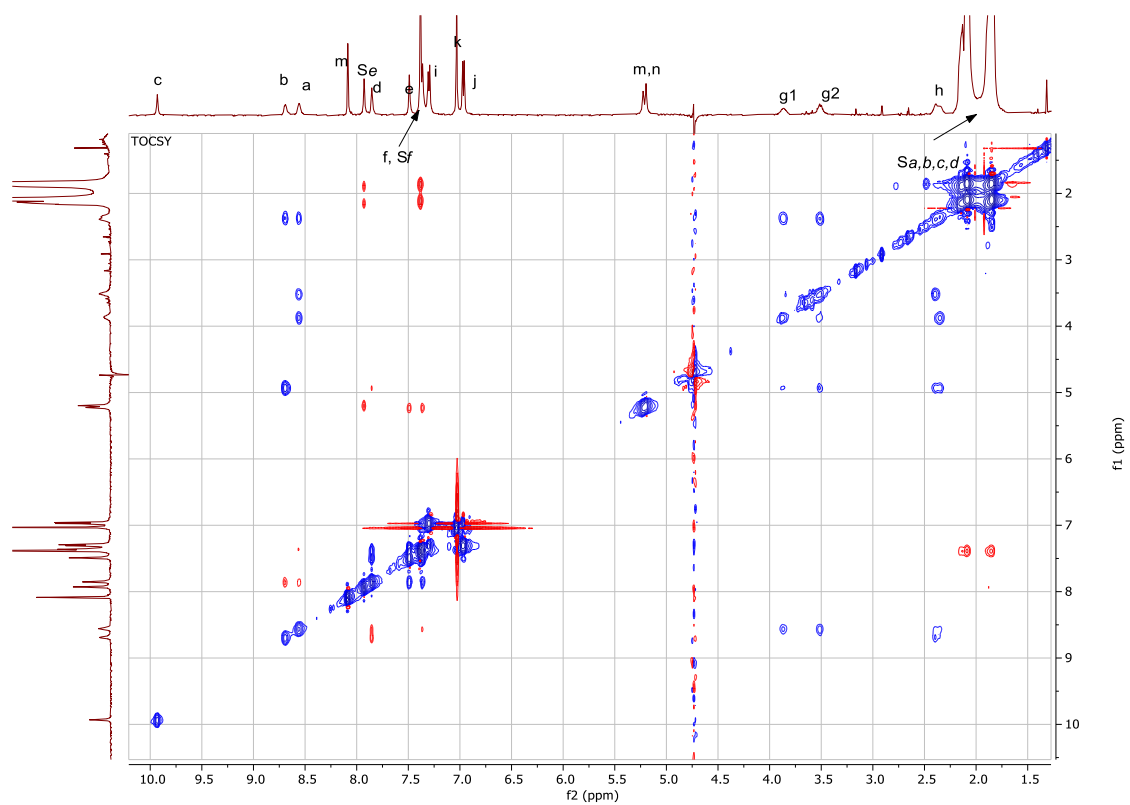
Supplementary Figure 24. 2D HSQC NMR spectrum (500 MHz) of receptor **21** in DMSO- d_6 .

Receptor 5

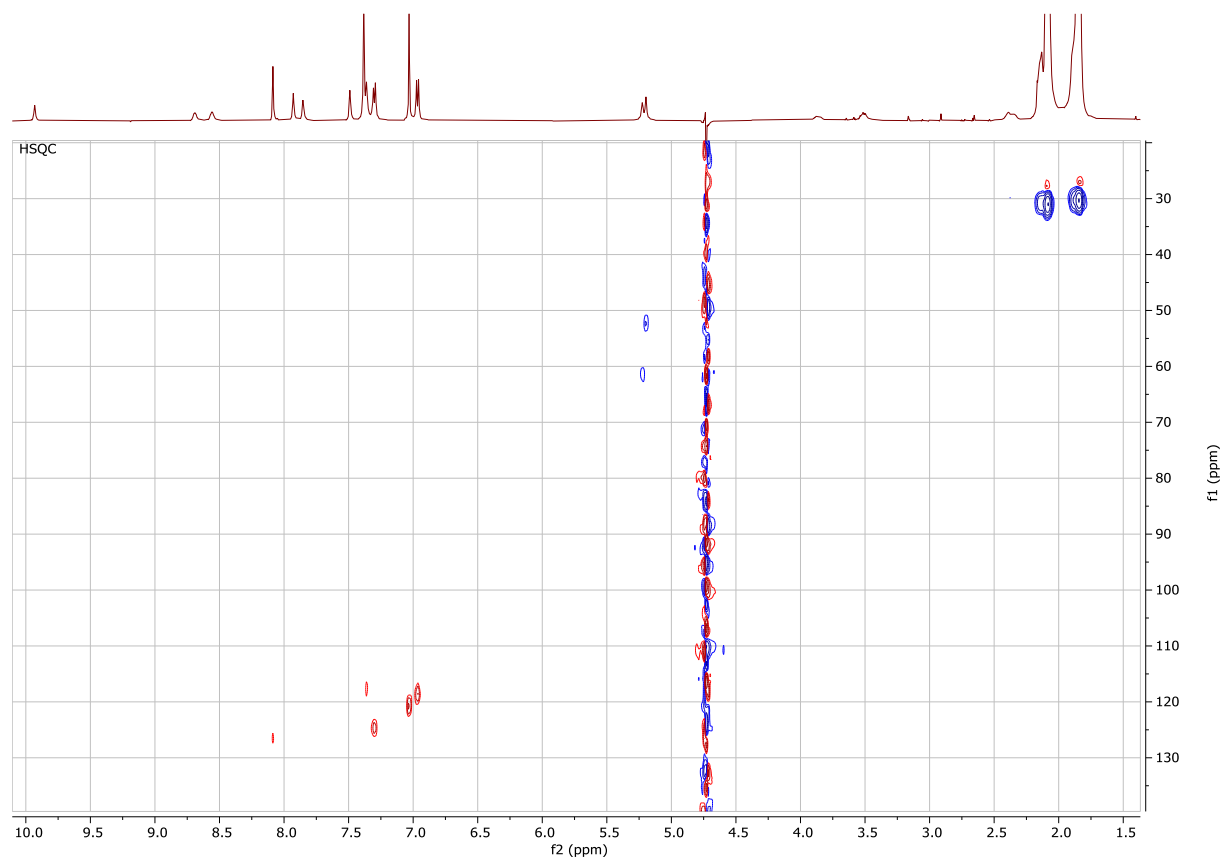
The receptor was characterized in 9:1 H₂O/D₂O. The assignment of the NMR spectrum was made with the help of 2D TOCSY and HSQC (see Supplementary Figures below).



Supplementary Figure 25. ¹H NMR spectrum (600 MHz) of receptor 5 in 9:1 H₂O/D₂O.



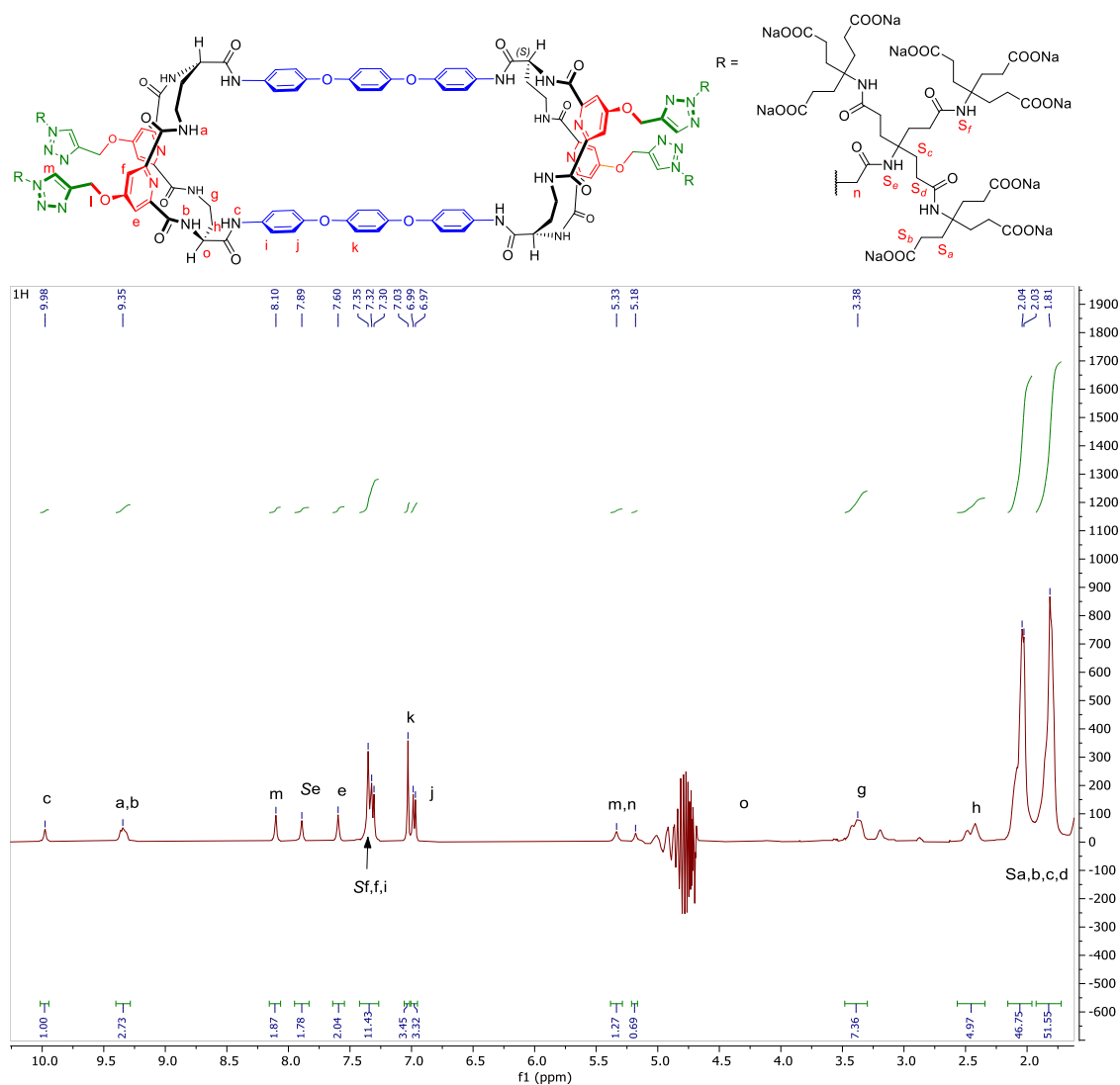
Supplementary Figure 26. 2D TOCSY NMR spectrum (600 MHz) of receptor 5 in 9:1 H₂O/D₂O.



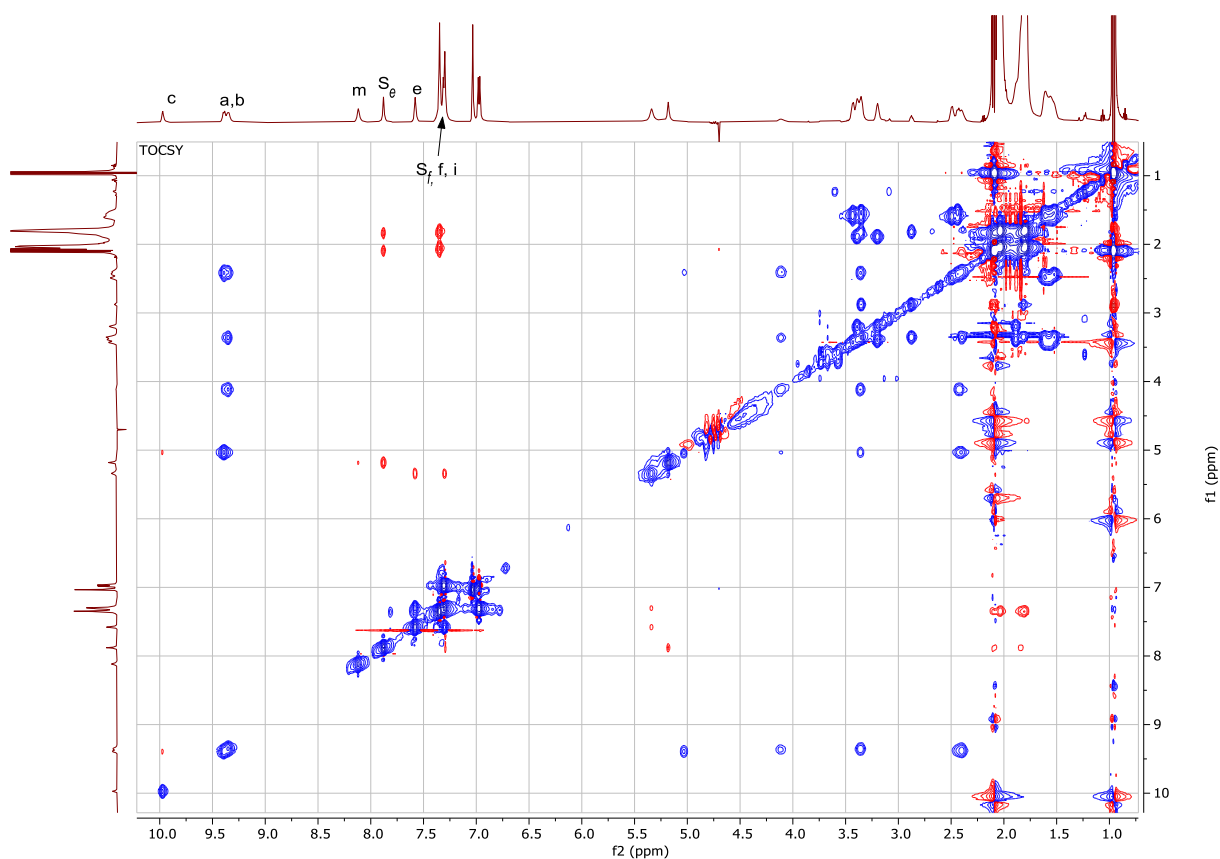
Supplementary Figure 27. 2D HSQC NMR spectrum (600 MHz) of receptor 5 in 9:1 H₂O/D₂O.

Receptor 6

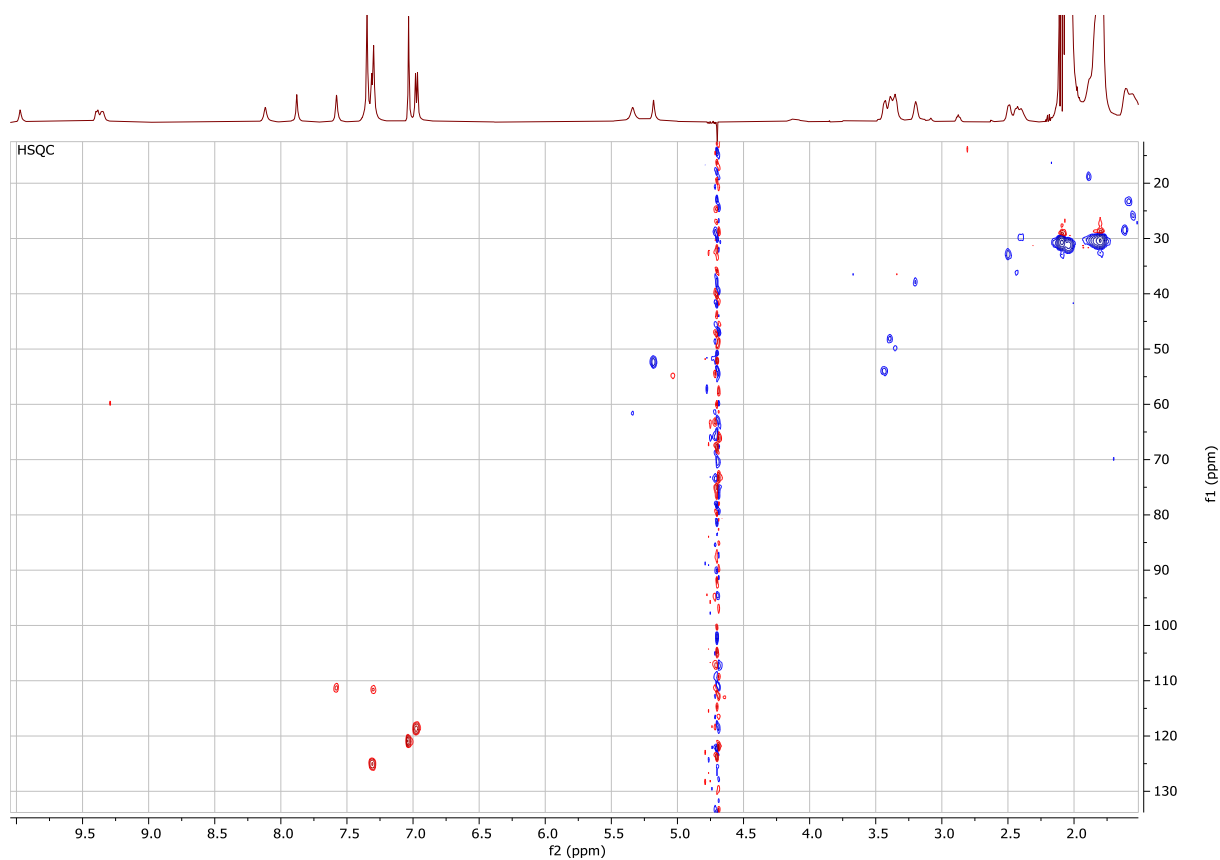
The receptor was characterized in 9:1 H₂O/D₂O. The assignment of the NMR spectrum was made with the help of 2D TOCSY and HSQC (see Supplementary Figures below).



Supplementary Figure 28. ¹H NMR spectrum (600 MHz) of receptor **6** in 9:1 H₂O/D₂O.



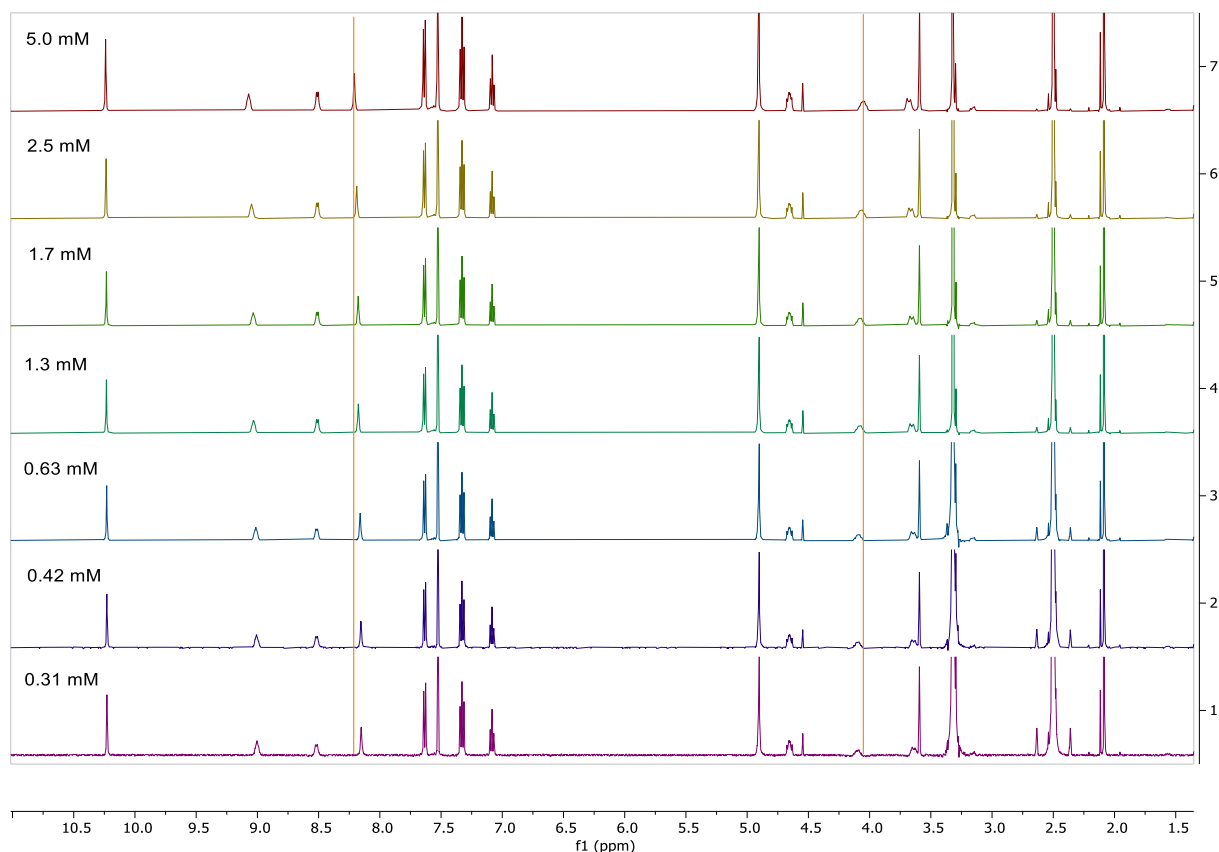
Supplementary Figure 29. 2D TOCSY NMR spectrum (600 MHz) of receptor **6** with sodium propionate in 9:1 H₂O/D₂O. The addition of propionate helps the assignment by resolving peaks between δ 7.20 – 7.40 ppm.



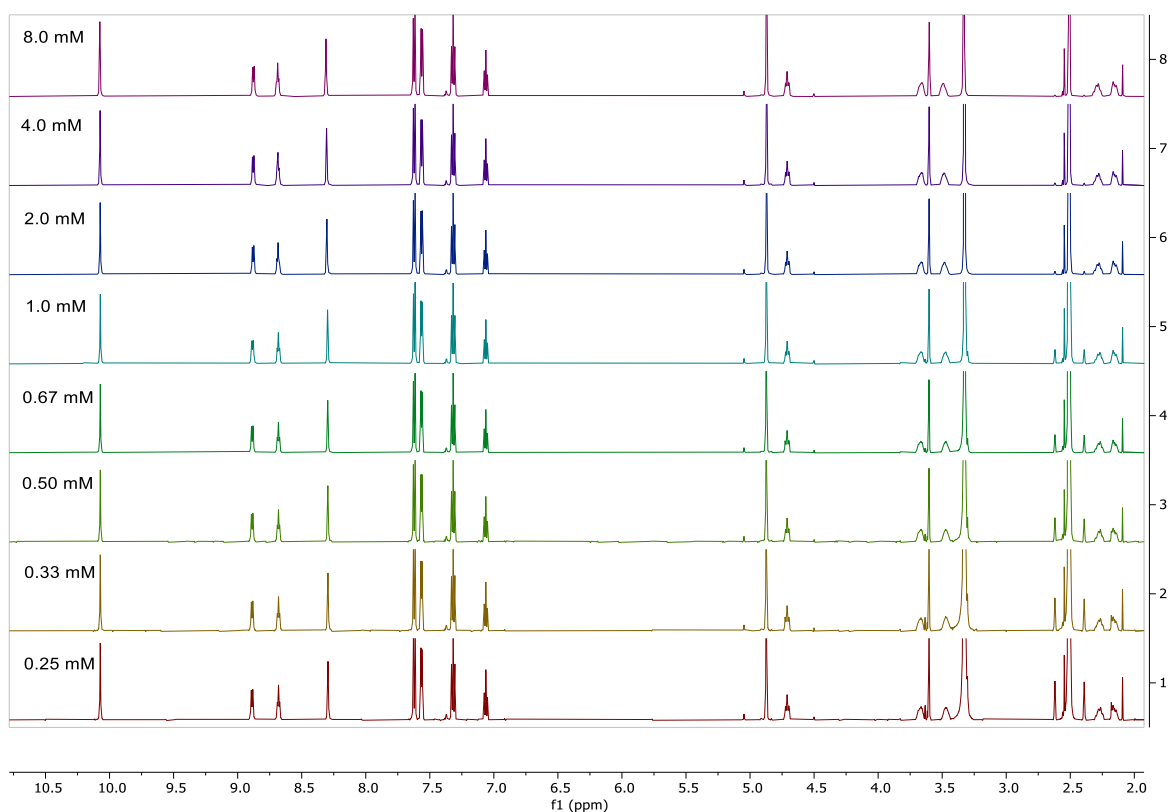
Supplementary Figure 30. 2D HSQC NMR spectrum (600 MHz) of receptor **6** with sodium propionate in 9:1 H₂O/D₂O. The addition of propionate helps the assignment by resolving peaks between δ 7.20 – 7.40 ppm.

1.7 Dilution studies

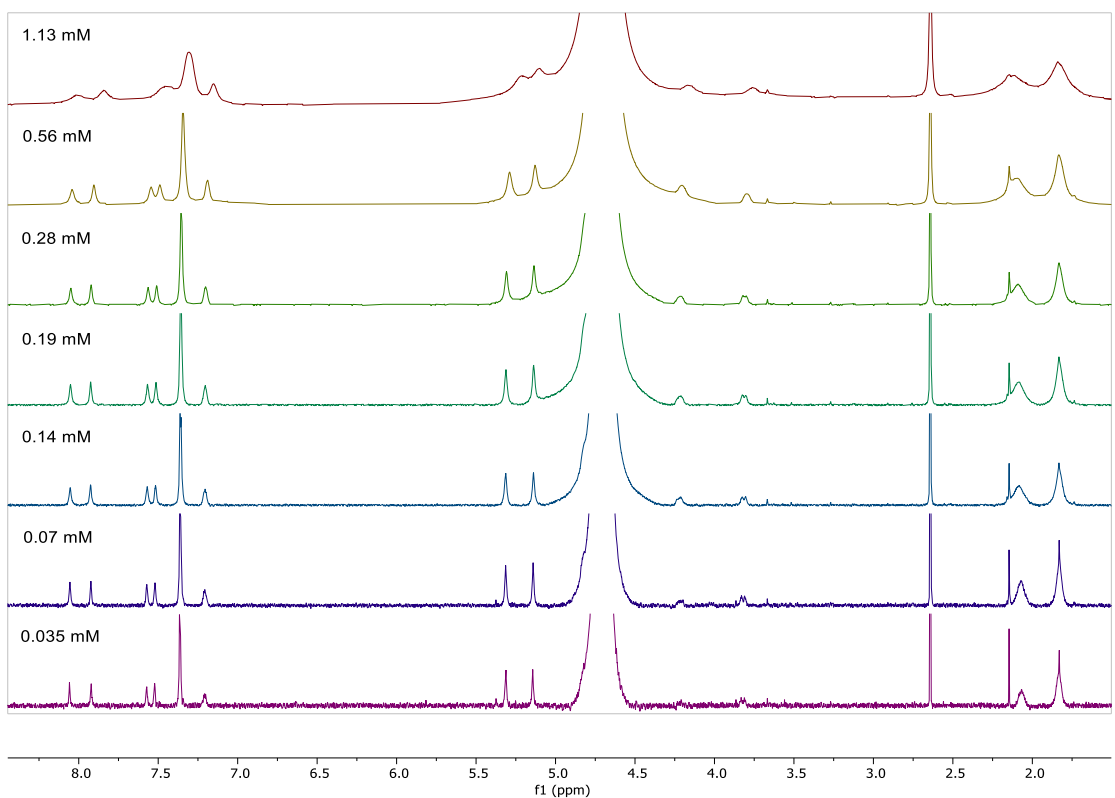
The solid receptors were dissolved in corresponding solvents (DMSO- d_6 or D_2O or 9:1 H_2O/D_2O) to make solutions with specific starting concentrations for each dilution study. 500 μL of the above solution was transferred into an NMR tube and the 1H NMR spectrum was acquired. The receptor solution was then diluted in the NMR tube by adding the same solvent, and a series of 1H NMR spectra was obtained.



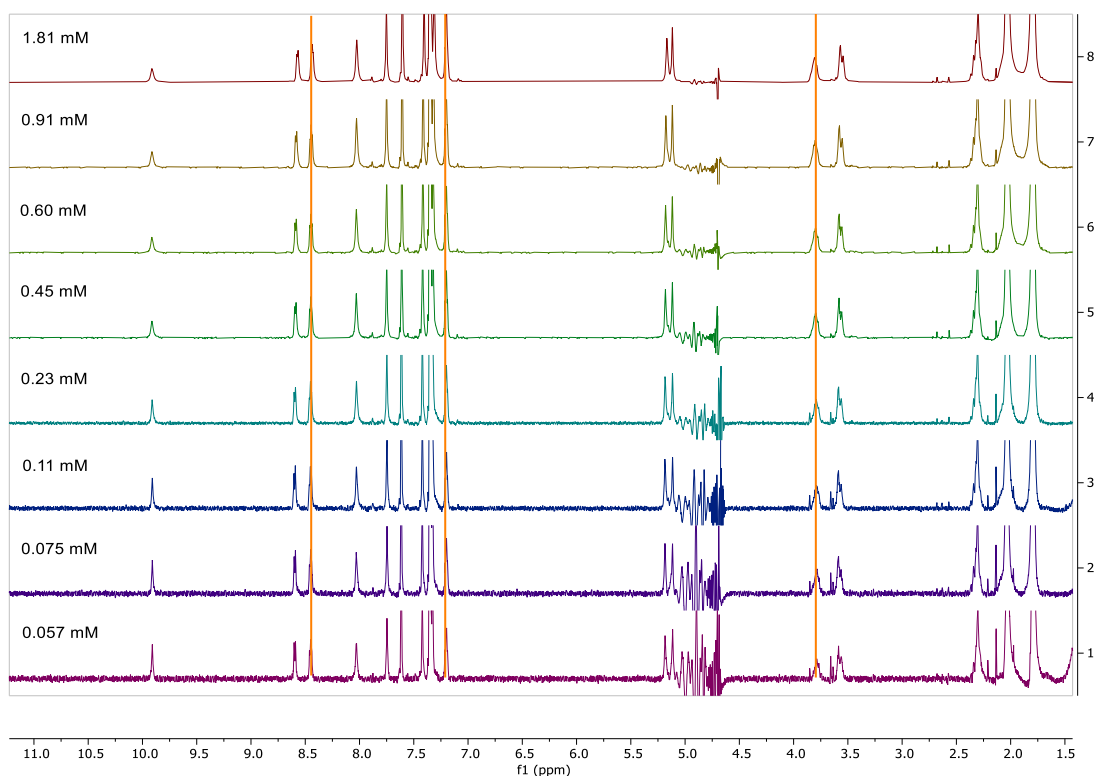
Supplementary Figure 31. 1H NMR spectra showing receptor **8a** at various concentrations in DMSO- d_6 . See yellow reference line for movements of proton d. The receptor is taken to be monomeric below 0.42 mM.



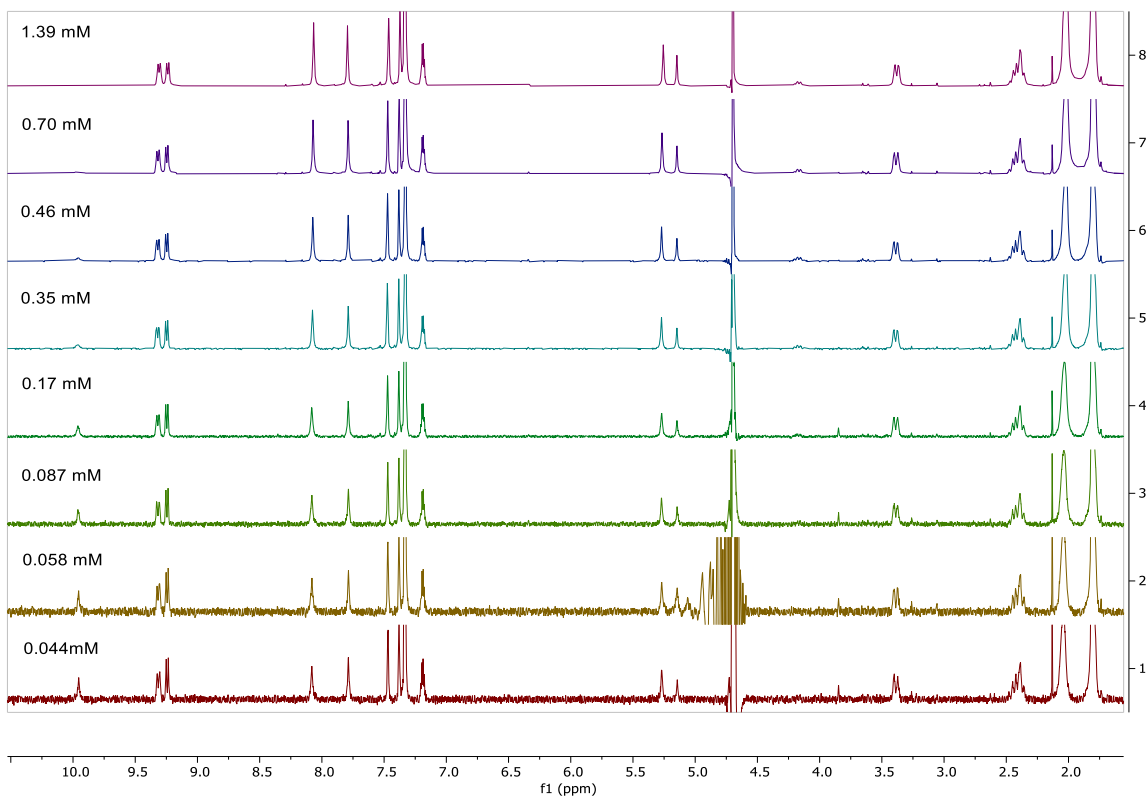
Supplementary Figure 32. ^1H NMR spectra showing receptor **9a** at various concentrations in $\text{DMSO}-d_6$. The receptor is taken to be monomeric below 0.67 mM.



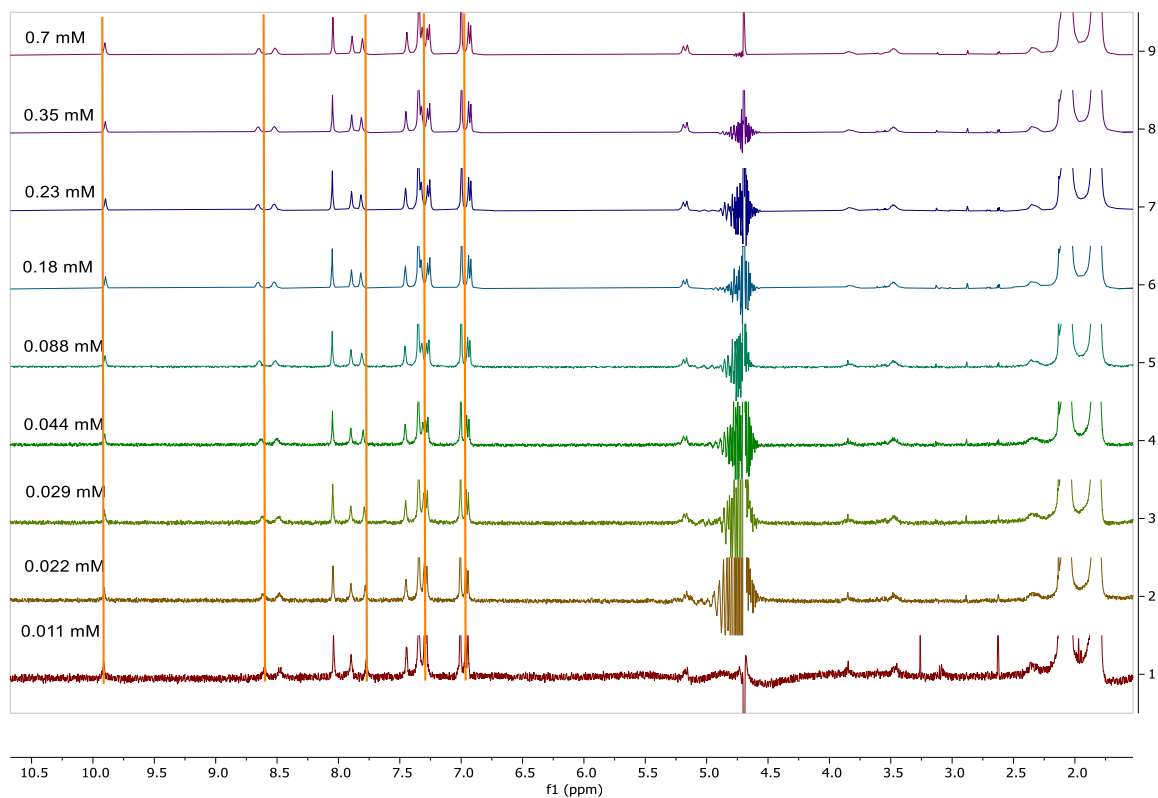
Supplementary Figure 33. ^1H NMR spectra showing receptor **8b** at various concentrations in D_2O . The receptor is taken to be monomeric below 0.07 mM.



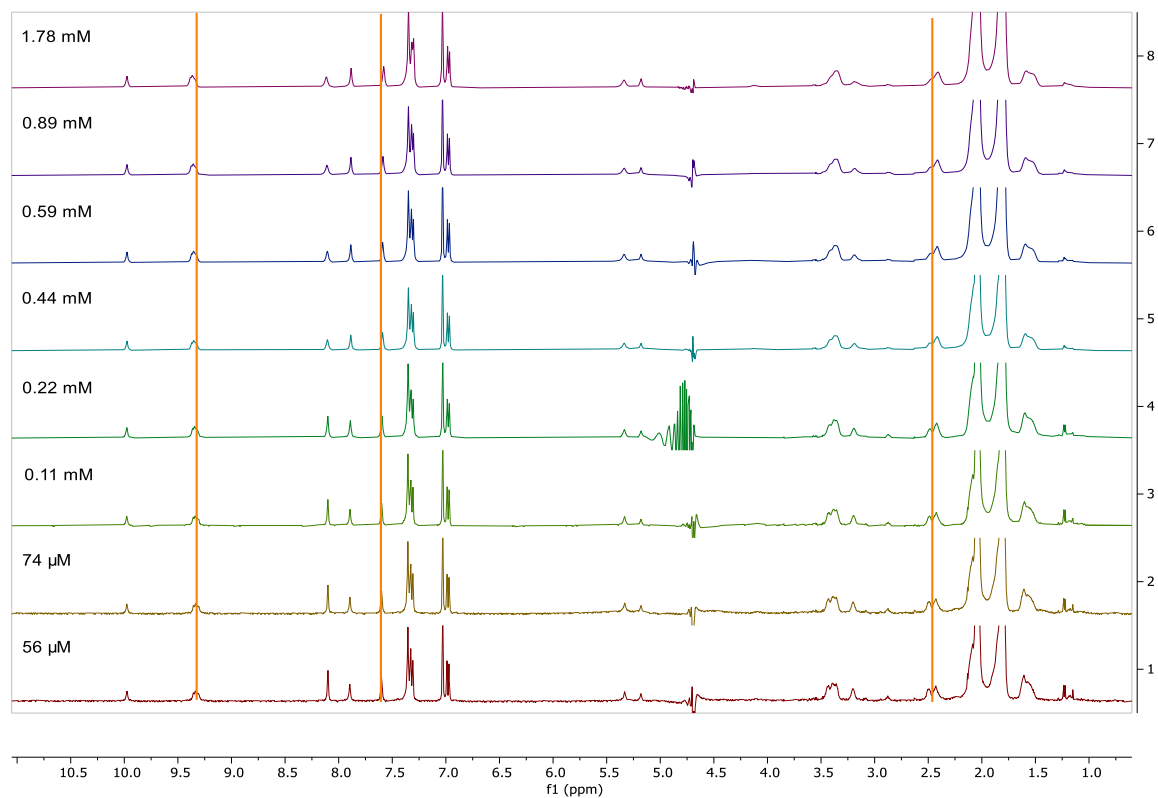
Supplementary Figure 34. ^1H NMR spectra showing receptor **9b** at various concentrations in 9:1 $\text{H}_2\text{O}/\text{D}_2\text{O}$. The receptor is taken to be monomeric below 0.075 mM.



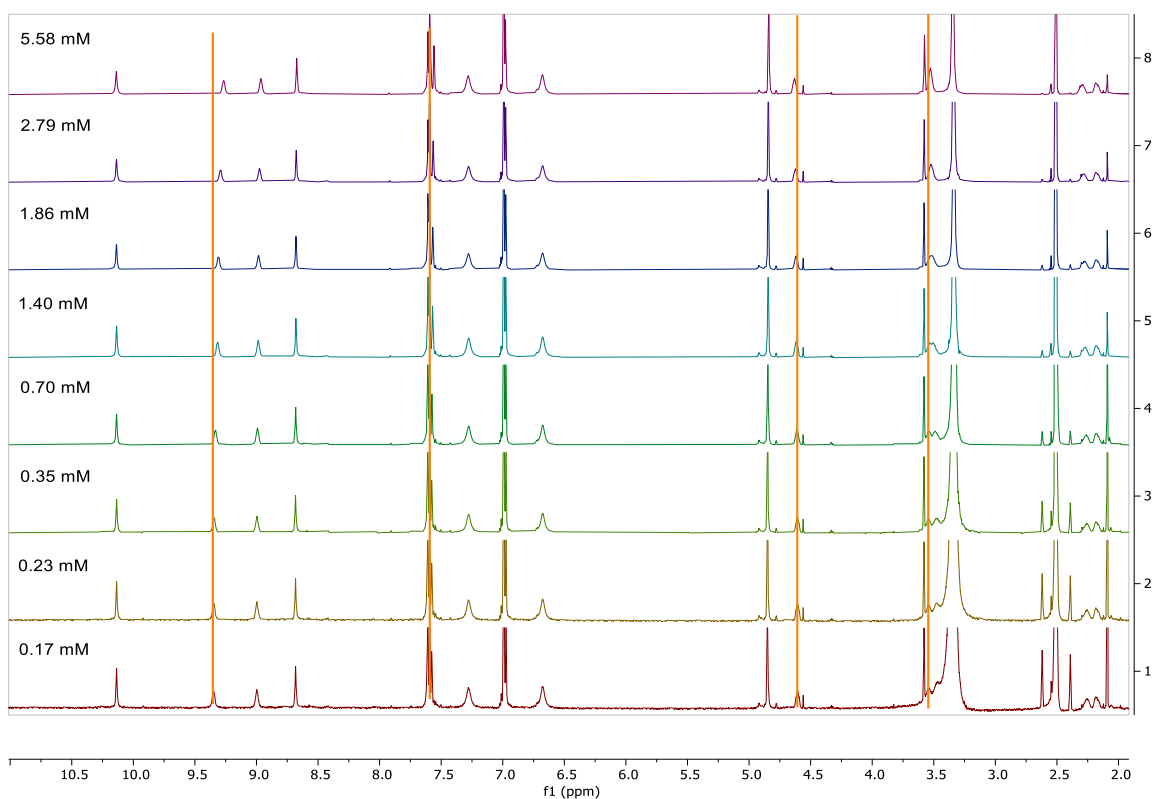
Supplementary Figure 35. ^1H NMR spectra showing receptor **10b** at various concentrations in 9:1 $\text{H}_2\text{O}/\text{D}_2\text{O}$ (Yellow reference line has been put on the spectrum). The receptor is taken to be monomeric below 0.058 mM.



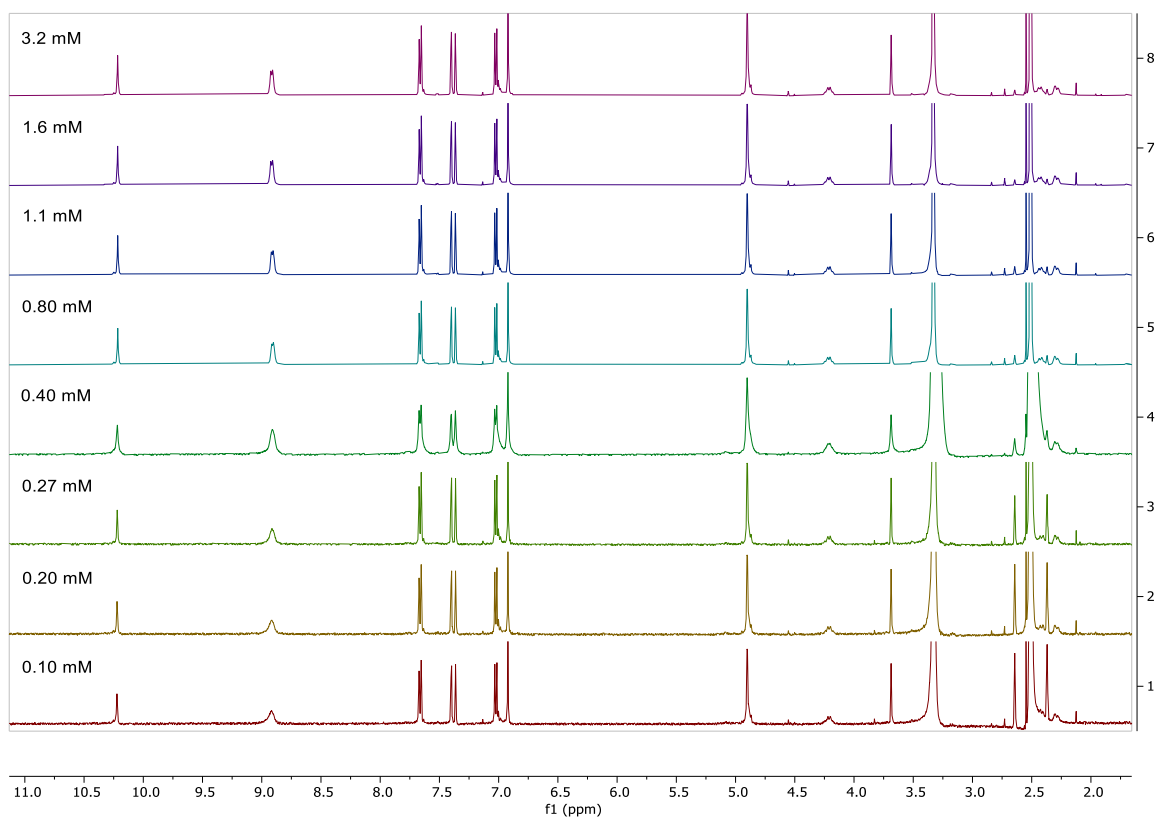
Supplementary Figure 36. ^1H NMR spectra showing receptor **5** at various concentrations in 9:1 $\text{H}_2\text{O}/\text{D}_2\text{O}$ (Yellow reference line has been put on the spectrum). The receptor is taken to be monomeric below 0.022 mM.



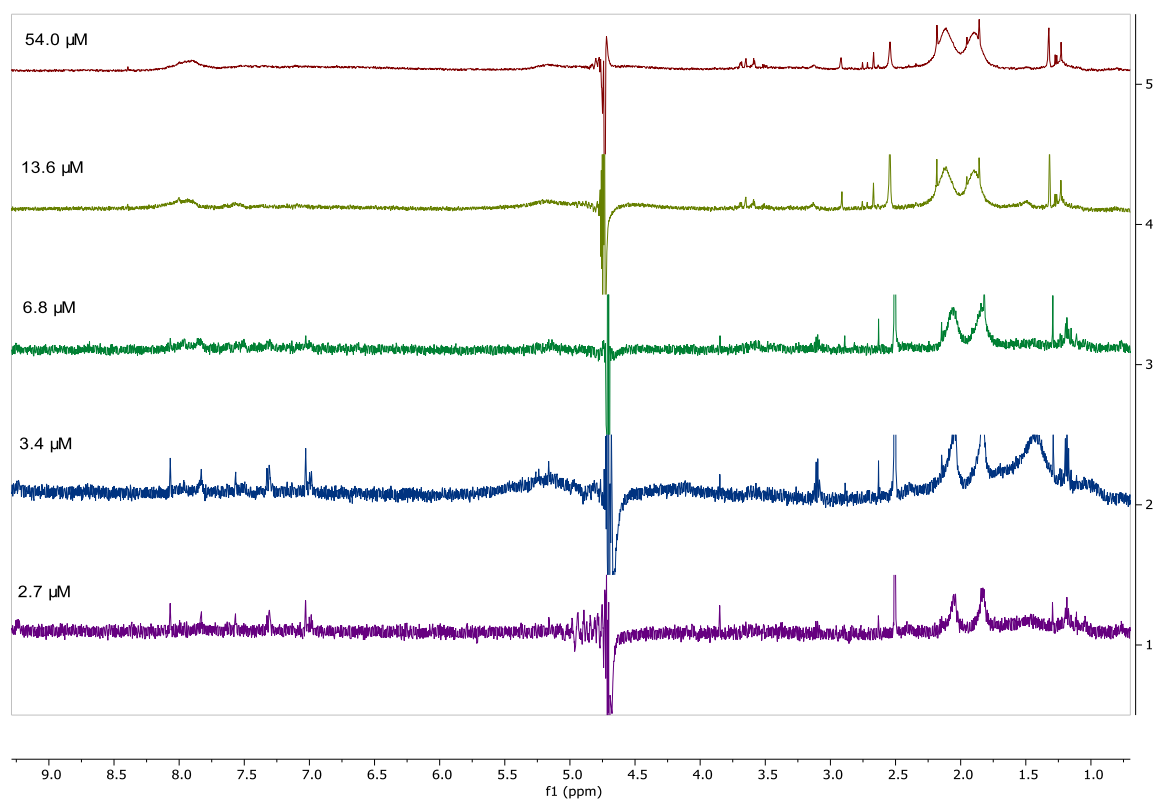
Supplementary Figure 37. ^1H NMR spectra showing receptor **6** at various concentrations in 9:1 $\text{H}_2\text{O}/\text{D}_2\text{O}$ (Yellow reference line has been put on the spectrum). The receptor is taken to be monomeric below 0.074 mM.



Supplementary Figure 38. ^1H NMR spectra showing receptor **20** at various concentrations in $\text{DMSO}-d_6$ (Yellow reference line has been put on the spectrum). The receptor is taken to be monomeric below 0.35 mM.



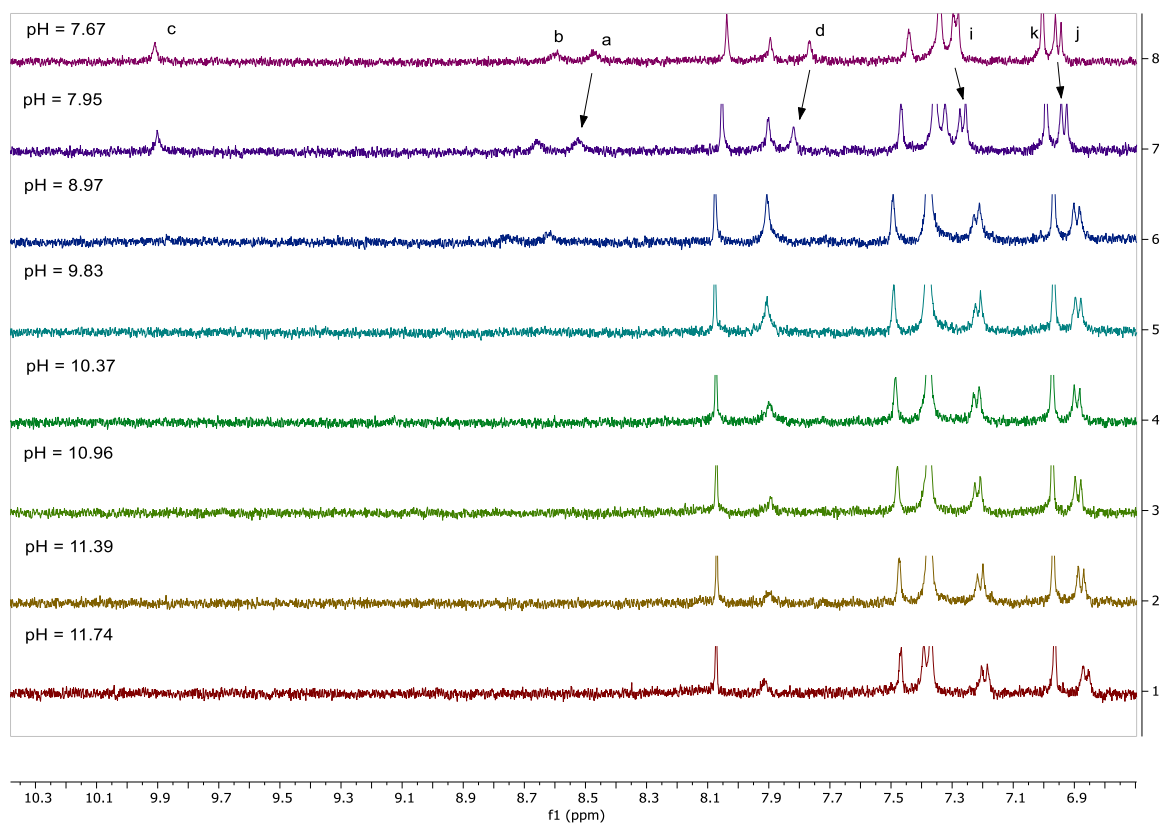
Supplementary Figure 39. ^1H NMR spectra showing receptor **21** at various concentrations in $\text{DMSO}-d_6$. The receptor is taken to be monomeric below 0.40 mM.



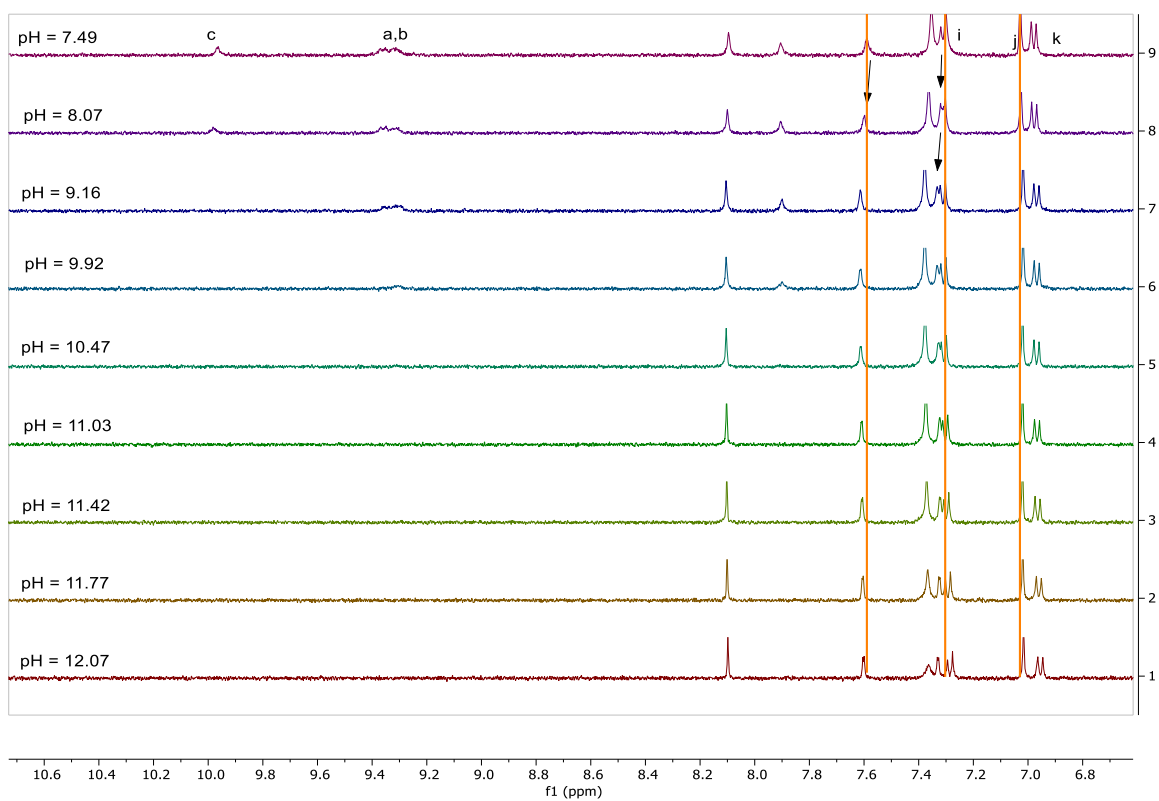
Supplementary Figure 40. ^1H NMR spectra showing receptor **6-G₁** at various concentrations in 9:1 $\text{H}_2\text{O}/\text{D}_2\text{O}$. The monomeric concentration is taken to be below 3.4 μM which is too low for binding studies by ^1H NMR with the equipment available.

1.8 pH titrations of receptors 5 and 6

pH near-neutral solutions of receptors **5** or **6** (21 μM or 22 μM) were made by dissolving the solid receptors into a specific amount to in 9:1 $\text{H}_2\text{O}/\text{D}_2\text{O}$ (typically 1000 μL). Solid NaOH was then dissolved in the above receptor solutions to make 10 mM, 100 mM and 1 M NaOH aqueous solution (100 μL each). The pH titration started by transferring the neutral receptor solution (500 μL) into an NMR tube and adding an aliquot of the prepared NaOH solutions. The NMR tube was shaken after the addition and then the ^1H NMR spectrum was acquired at 298 K. After the acquisition, the titration solution was transferred to a vial for pH measurement before putting it back into the same NMR tube for the next addition.



Supplementary Figure 41. ^1H NMR spectra showing receptor **5** (21 μM) at various pH in 9:1 $\text{H}_2\text{O}/\text{D}_2\text{O}$.



Supplementary Figure 42. ^1H NMR spectra showing receptor **6** (22 μM) at various pH in 9:1 $\text{H}_2\text{O}/\text{D}_2\text{O}$.

2. Binding studies

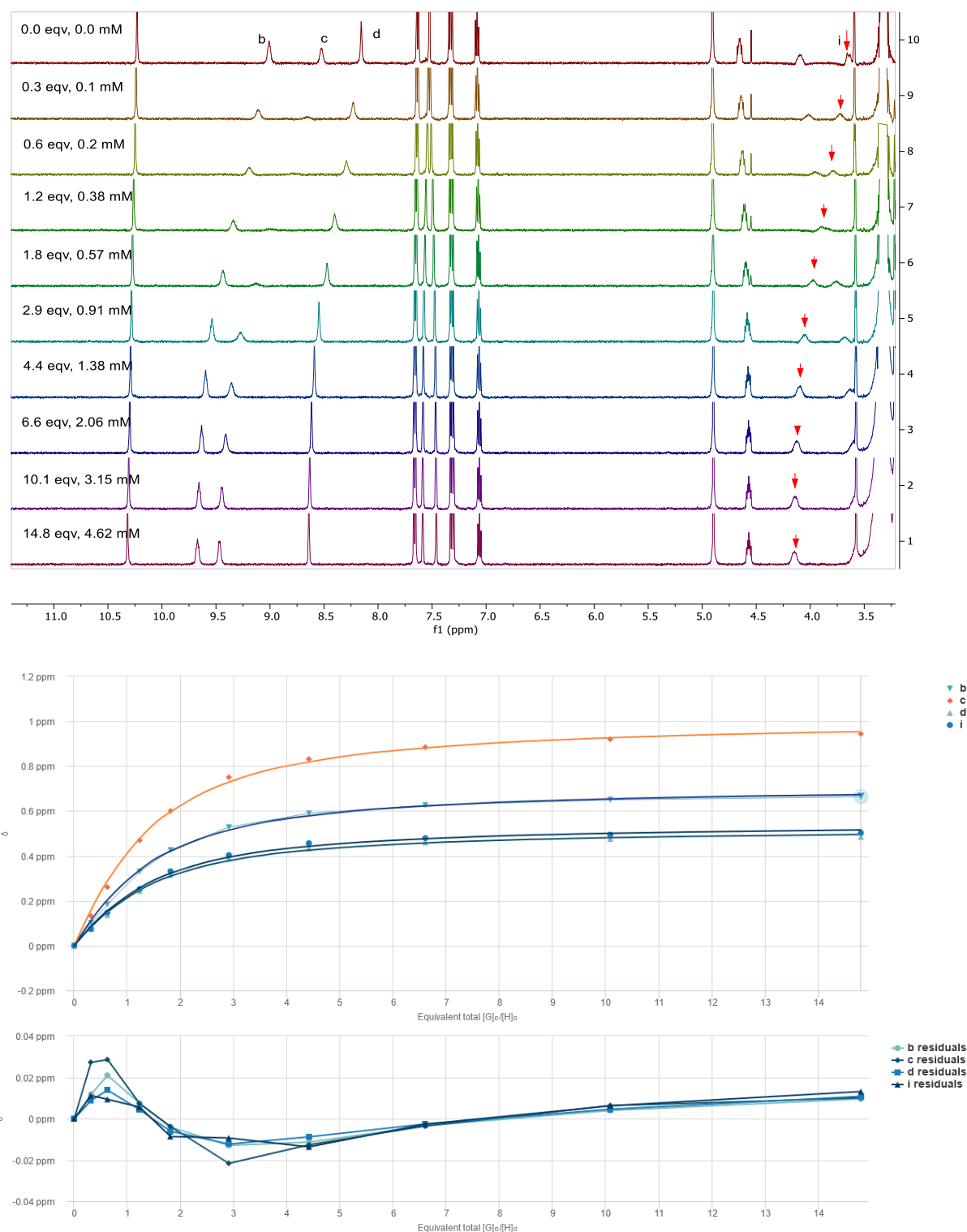
^1H NMR titrations were performed on Bruker Advance III HD Cryo 500 MHz or Bruker Neo Cryo 600 MHz spectrometers.

For titrations in organic solvent ($\text{DMSO-}d^6$), solutions of receptor at known concentrations were prepared in an NMR tube. Aliquots of a solution of receptor at the same concentration and guest were added. The receptor concentration was therefore held constant while the guest concentration was increased. The NMR tube was shaken after each addition and the ^1H NMR spectra acquired at 298 K.

Titrations in aqueous medium were conducted at a constant near-neutral pH in D_2O or 9:1 $\text{H}_2\text{O}/\text{D}_2\text{O}$. The host stock solution was first prepared by dissolving a weighed amount of host solid in D_2O or 9:1 $\text{H}_2\text{O}/\text{D}_2\text{O}$ and the pH was checked to be near-neutral (the pH ranged from 7.35 to 7.50 for different hosts). For carboxylate guests, the guest stock solution was then prepared by dissolving sodium carboxylate salt in water and adjusting pH to match the host solution as closely as possible ($\text{pH}_{\text{guest}} = \text{pH}_{\text{host}} \pm 0.15$) by adding the conjugate acid. Other guest (NaCl , *L*- and *D*-Ala, *L*-Ala-*L*-Ala) stock solutions were prepared by dissolving the compound in water and confirming the neutrality of pH. Before each aqueous binding experiment, two solutions for titration were prepared using the above near-neutral stock solutions. The first consisted of 550 μL of host solution at a concentration below the association threshold (ranging from 10 μM to 133 μM for different hosts). The second solution consisted of 500 μL of host, at the same concentration as the first solution, mixed with guest (50 mM to 500 mM). Both solutions were made in 1.5 mL vials, and the near-neutral pH was confirmed (ranging from 7.30 to 7.65 for different host and host-guest solutions). During the titration, the host solution (500 μL) was transferred to an NMR tube and aliquots of host-guest mixture were added precisely using Gilson pipettes. This method holds the receptor concentration constant while the guest concentration was increased, with the pH staying near-neutral. The NMR tube was shaken after each addition and the ^1H NMR spectra acquired at 298 K. At the end of each titration, the solution in the NMR tube was transferred into a vial and the pH was measured again confirming negligible changes ($\Delta\text{pH} < \pm 0.1$) during the experiment.

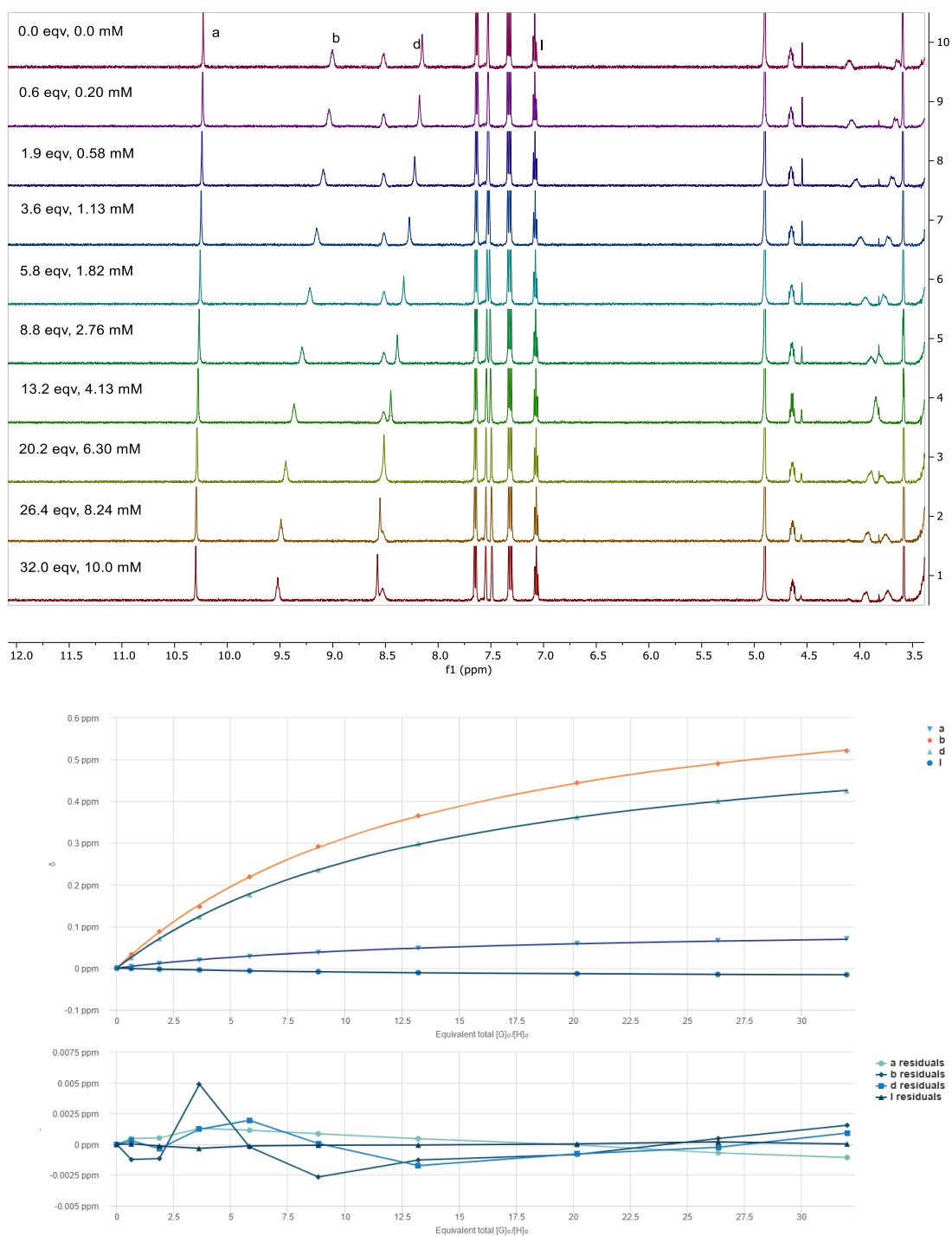
Association constants were determined by monitoring the change in chemical shift ($\Delta\delta$) for selected protons belonging to the receptor species. The $\Delta\delta$ values were analysed to give binding constants K_a using Bindfit⁵, employing the 1:1 binding model for monomeric receptors **8** – **10** and the 1:2 non-cooperative binding models for dimeric receptors **20**, **21**, **5** and **6**.

Receptor 8a & TBA acetate (DMSO- d_6)



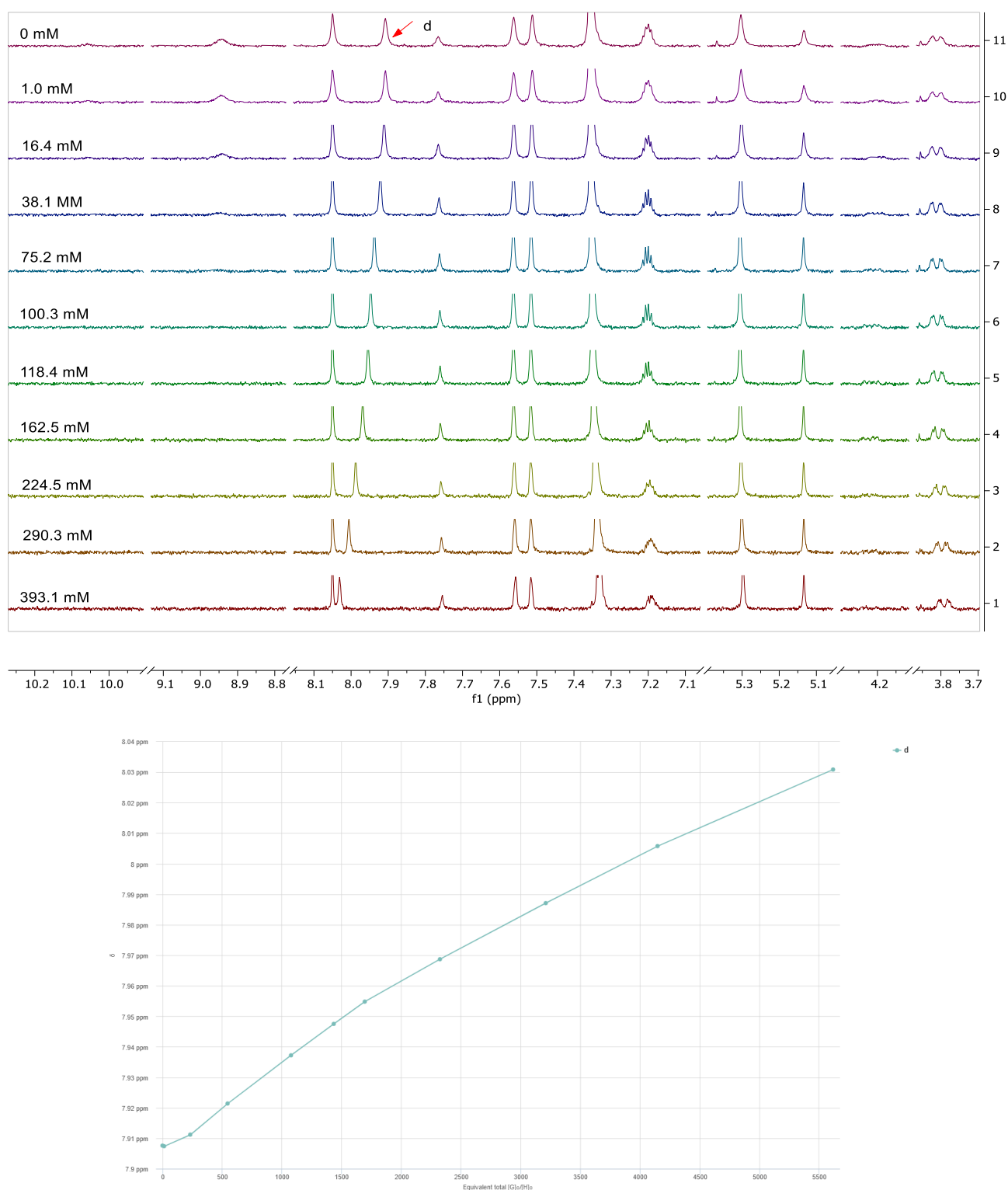
Supplementary Figure 43. Top). ^1H NMR spectra (500 MHz, DMSO- d_6) for receptor **8a** (0.31 mM) titrated with a combined solution of tetrabutylammonium acetate (20 mM) and receptor **8a** (0.31 mM). The equivalents and concentrations of guest added are listed in the graph. Bottom). Global fitting of the binding isotherms (protons b, c, d and i) from Bindfit to a 1:1 model $K_a = 3759 \text{ M}^{-1}$ ($\pm 4.0 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/33a29e26-f8dc-47dd-9593-130fa22860e7>

Receptor 8a & TBA chloride (DMSO- d_6)



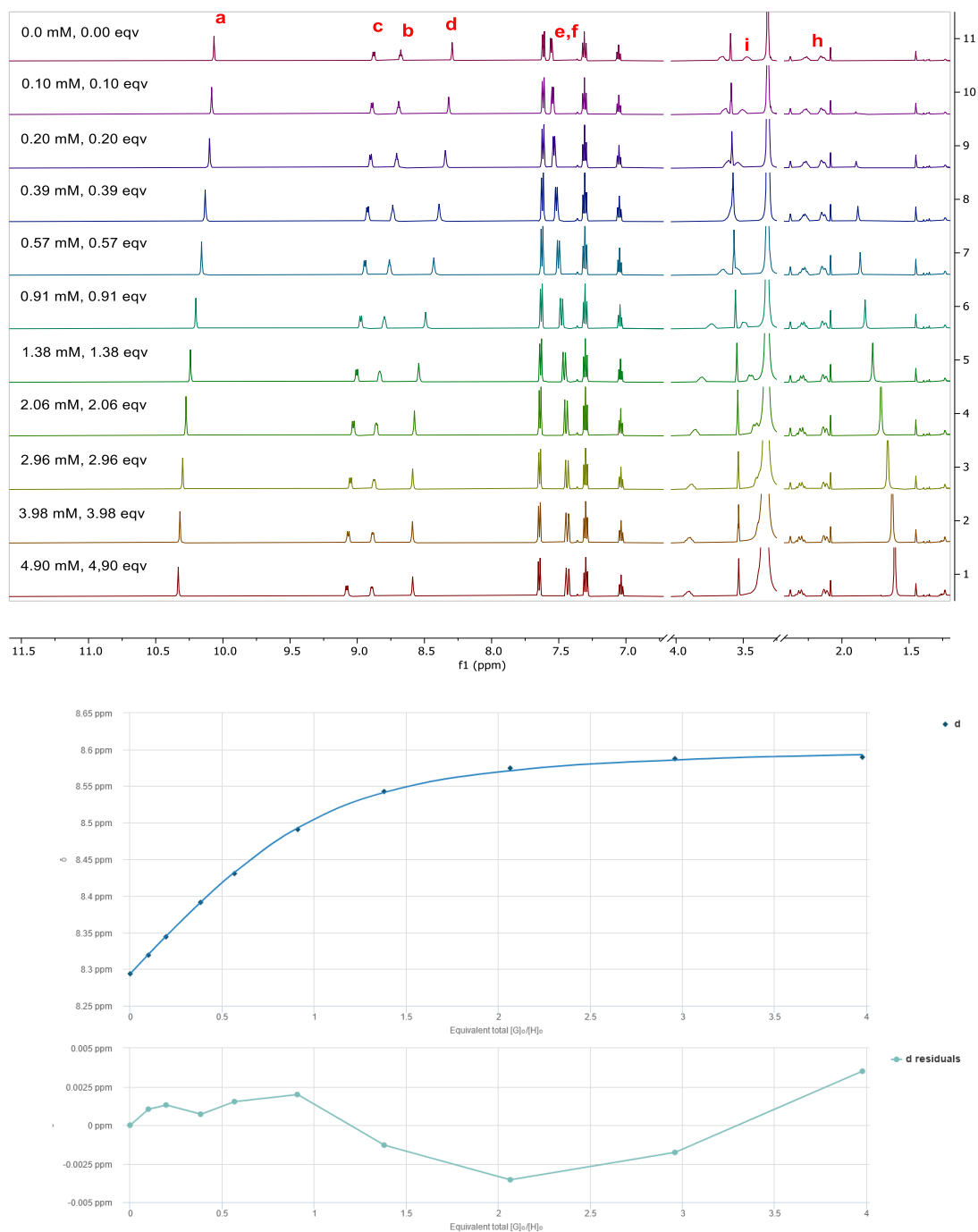
Supplementary Figure 44. Top). ^1H NMR spectra (500 MHz, DMSO- d_6) for receptor **8a** (0.31 mM) titrated with a combined solution of tetrabutylammonium chloride (20 mM) and receptor **8a** (0.31 mM). The equivalents and concentrations of guest added are listed in the graph. Bottom). Global fitting of the binding isotherms (protons a, b, d and l) from Bindfit to a 1:1 model $K_a = 240 \text{ M}^{-1}$ ($\pm 0.5 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/bc5f579a-5564-48df-bf4a-105830f6d889>

Receptor 8b & chloride (H₂O/D₂O)



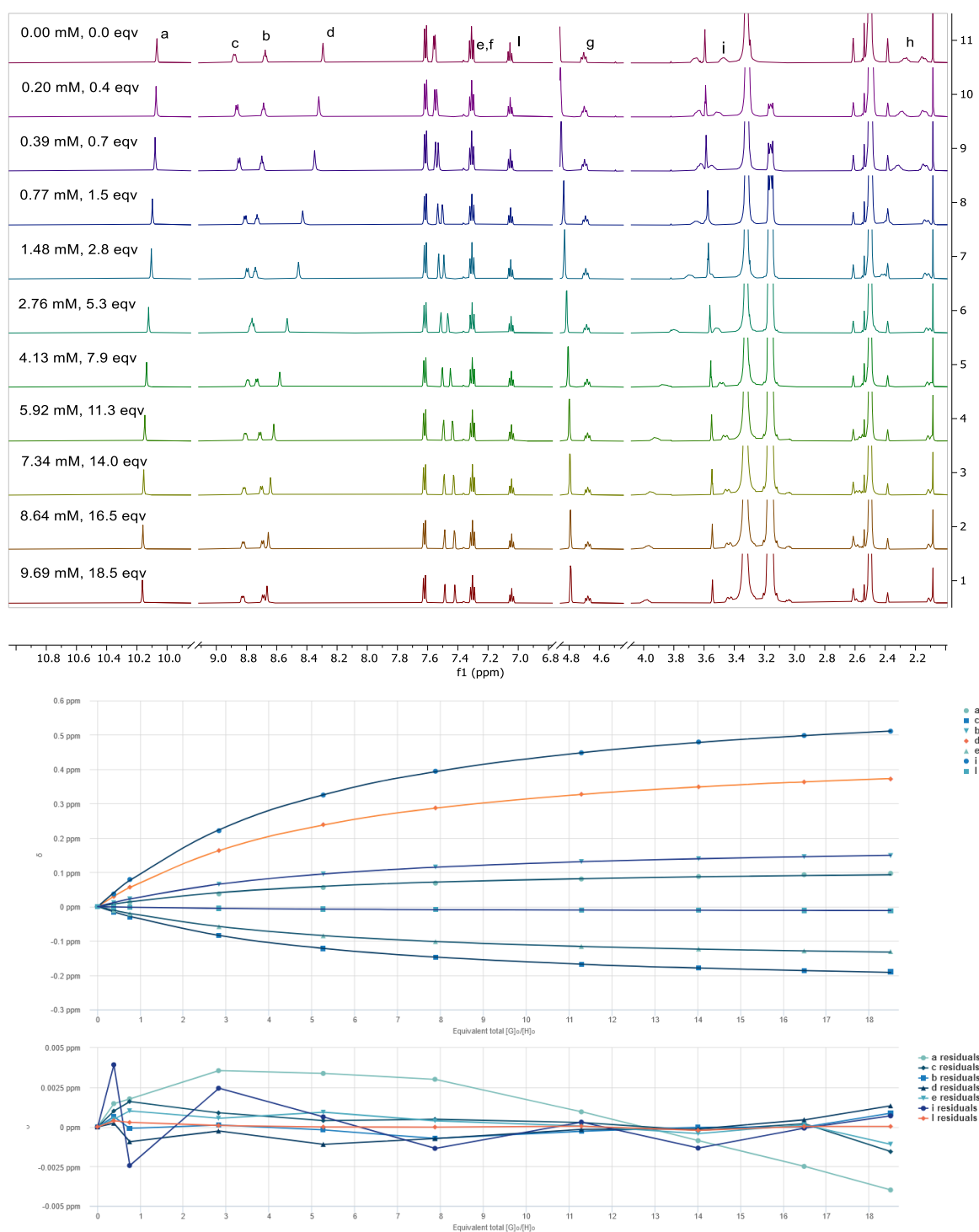
Supplementary Figure 45. Top). ¹H NMR spectra (600 MHz, 1:1 H₂O/D₂O) for receptor **8b** (0.07 mM) titrated with a combined solution of sodium chloride (800 mM) and receptor **8b** (0.07 mM). The equivalents and concentrations of guest added are listed in the graph. pH = 7.60 at the end of titration. Bottom) The chemical shift changes from proton d which cannot be fitted into any model.

Receptor 9a & TBA acetate (DMSO- d_6)



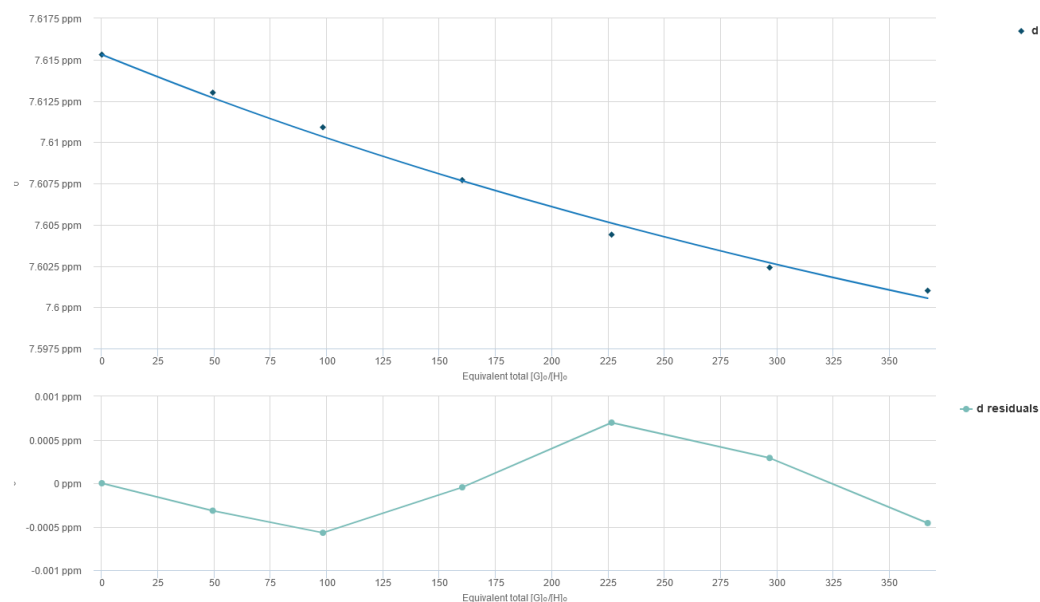
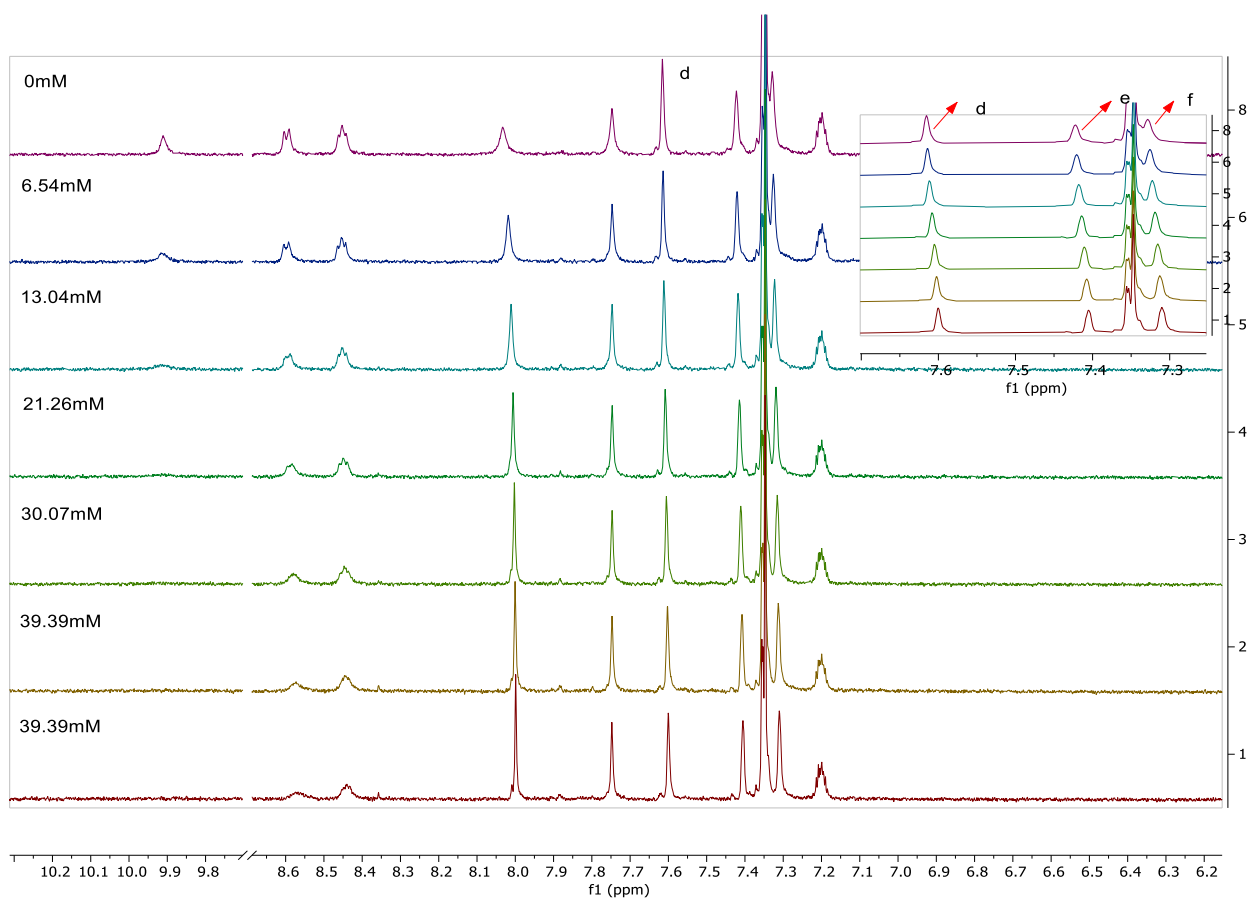
Supplementary Figure 46. Top). ^1H NMR spectra (600 MHz, DMSO- d_6) for receptor **9a** (1 mM) titrated with a combined solution of tetrabutylammonium acetate (10 mM) and receptor **9a** (1 mM). The equivalents and concentrations of guest added are listed in the graph; Bottom). Fitting of the binding isotherm (proton d) in Bindfit to a 1:1 model $K_a = 6173 \text{ M}^{-1}$ ($\pm 7.1 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/11429b47-ec19-4284-be58-35df7f8fb354>

Receptor 9a & TBA chloride (DMSO- d_6)



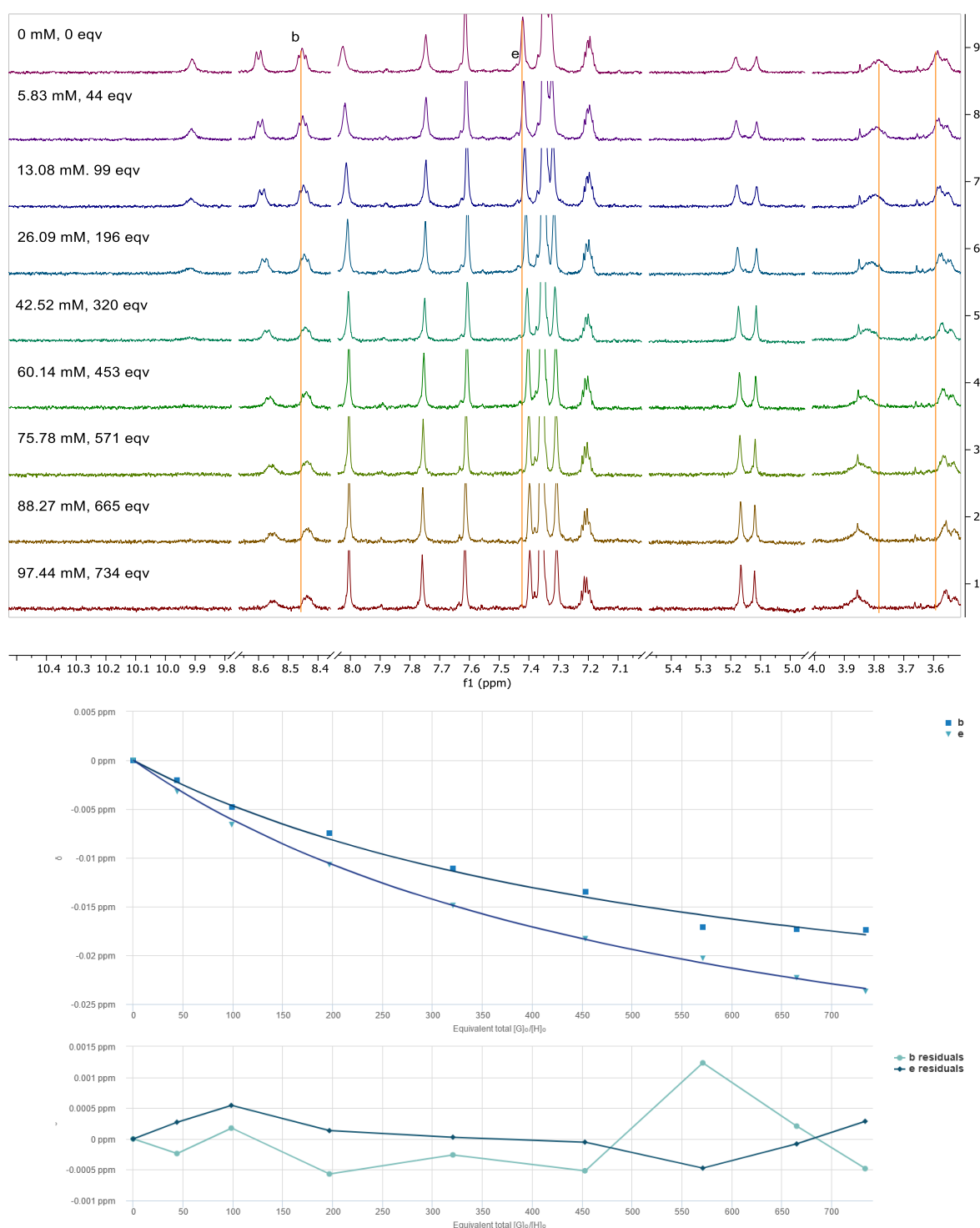
Supplementary Figure 47. Top) ^1H NMR spectra (600 MHz, DMSO- d_6) for receptor **9a** (524 μM) titrated with a combined solution of tetrabutylammonium chloride (20 mM) and receptor **9a** (524 μM). The equivalents and concentrations of guest added are listed in the graph. Bottom). Global fitting of the binding isotherms (protons a, c, b, d, e, i and o) from Bindfit to a 1:1 model $K_a = 405.2 \text{ M}^{-1}$ ($\pm 0.56 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/bdda8616-dcbf-46ca-8823-90cb8373d105>

Receptor 9b & acetate (H₂O/D₂O)



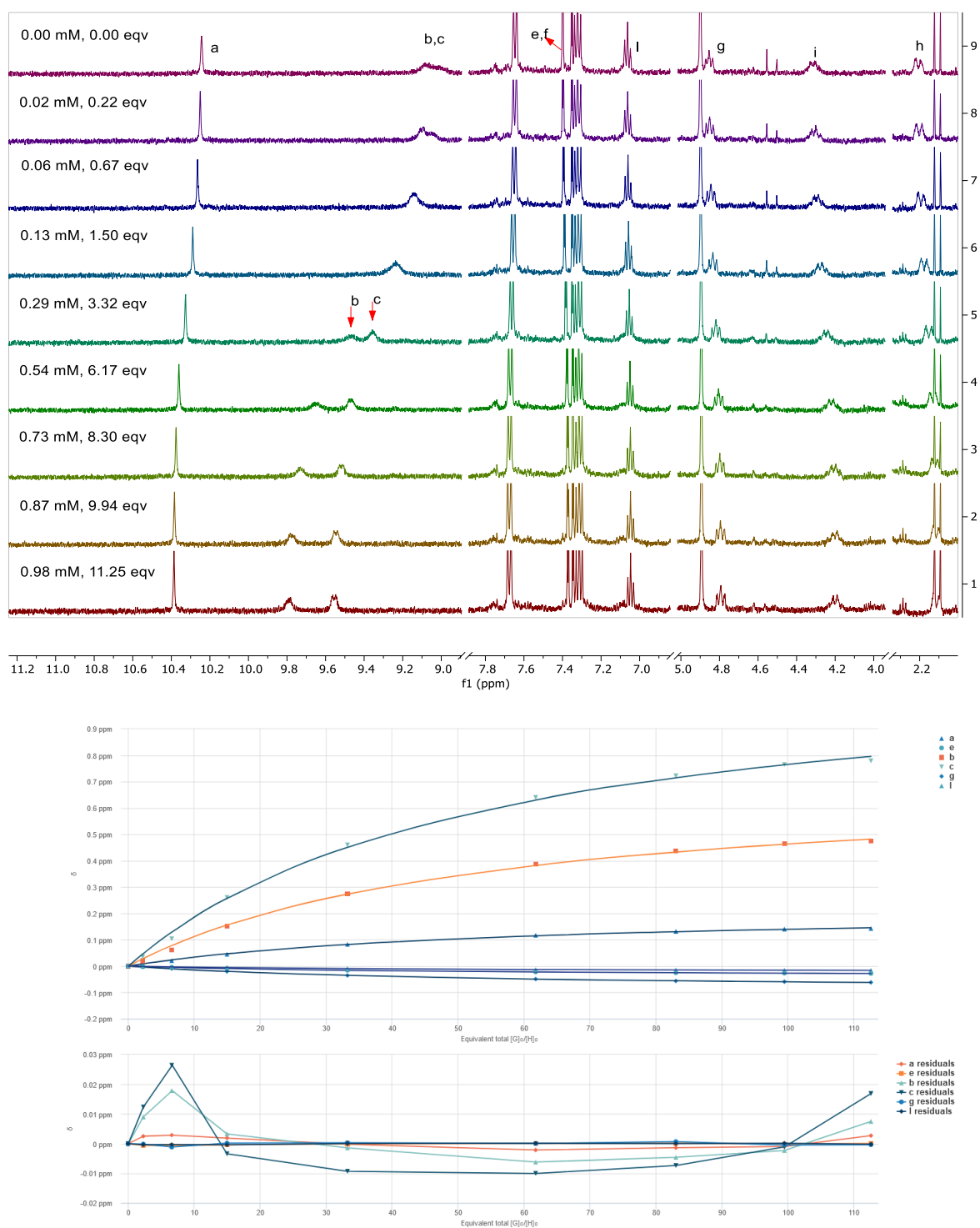
Supplementary Figure 48. Top) ¹H NMR (600 MHz, 9:1 H₂O/D₂O) spectra for receptor **9b** (133 μM) titrated with a combined solution of sodium acetate (100 mM) and receptor **9b** (133 μM). The concentration of guest is shown on each spectrum. pH = 7.53 at the end of titration. Bottom) Fitting of the binding isotherm (proton d) from Bindfit to a 1:1 model $K_a = 8.0 \text{ M}^{-1}$ ($\pm 7.6 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/62ac903b-f59f-4555-be81-15928ad94088>

Receptor 9b & chloride (H₂O/D₂O)



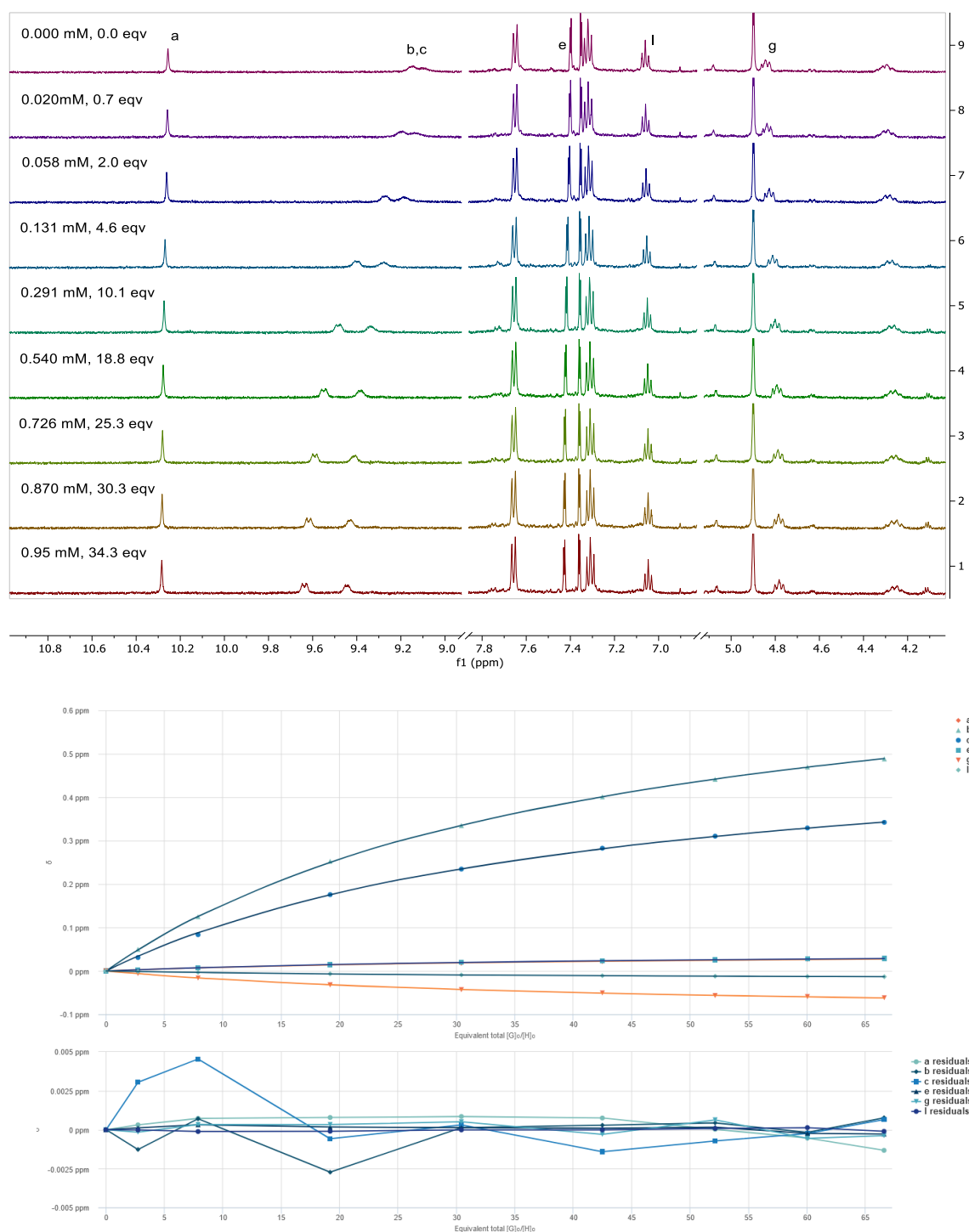
Supplementary Figure 49. Top). ¹H NMR spectra (600 MHz, 9:1 H₂O/D₂O) for receptor **9b** (133 μM) titrated with a combined solution of sodium chloride (200 mM) and receptor **9b** (133 μM). The equivalents and concentrations of guest added are listed in the graph. pH = 7.56 at the end of titration. Bottom). Global fitting of the binding isotherms (protons b, e) from Bindfit to a 1:1 model $K_a = 12.85 \text{ M}^{-1}$ ($\pm 3.26 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/0c39573d-f3c2-4f73-9260-8c1356858a77>

Receptor 10a & TBA acetate (DMSO- d_6)



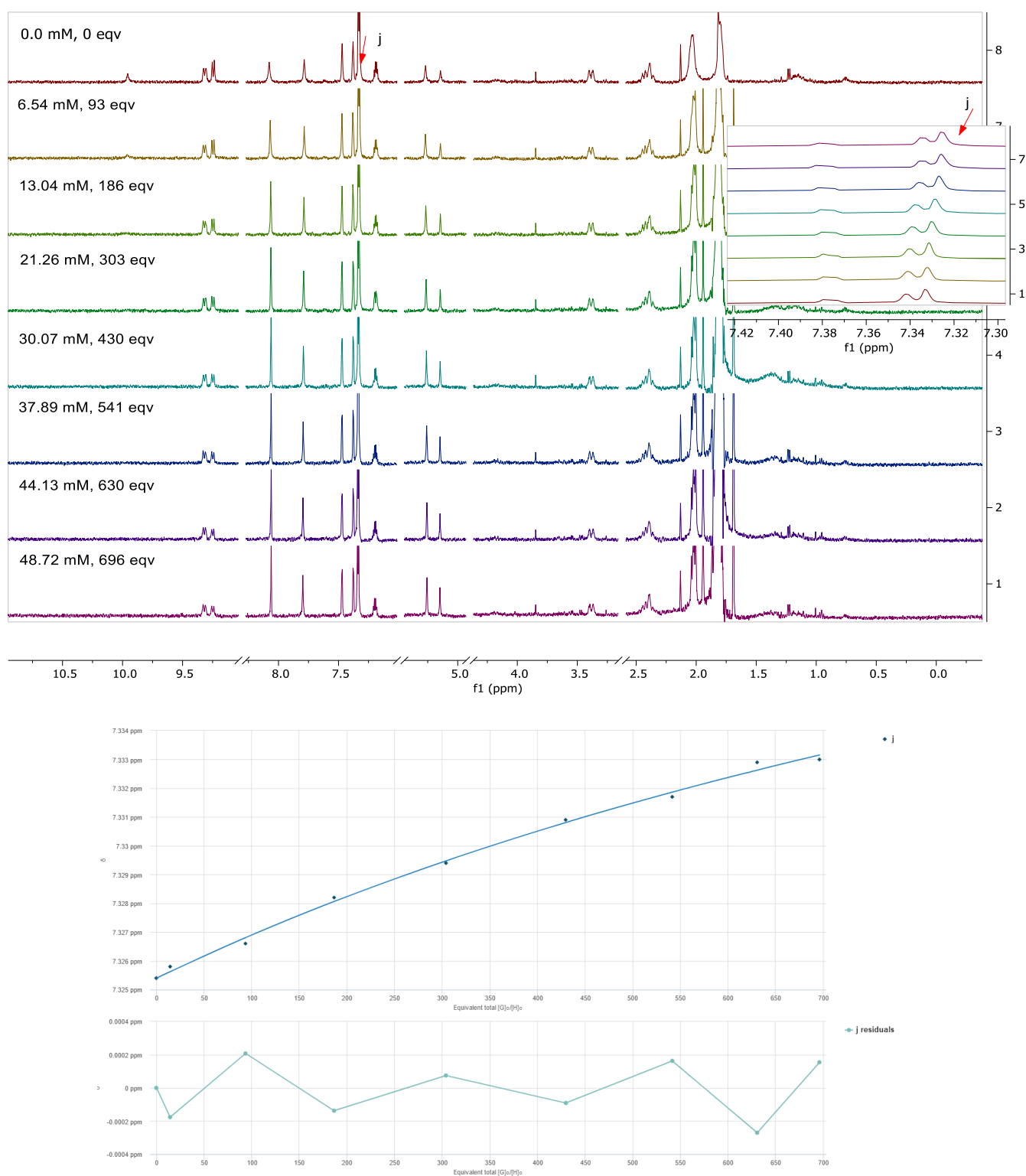
Supplementary Figure 50. Top) ^1H NMR spectra (500 MHz, DMSO- d_6) for receptor **10a** (87.5 μM) titrated with a combined solution of tetrabutylammonium acetate (2 mM) and receptor **10a** (87.5 μM). The equivalents and concentrations of guest added are listed in the graph. Bottom) Global fitting of the binding isotherms (protons a, b, c, e, g and i) from Bindfit to a 1:1 model $K_a = 2192 \text{ M}^{-1}$ ($\pm 1.7 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/c2a3e941-7f5e-4529-b500-e8893345559f>

Receptor 10a & TBA chloride (DMSO- d_6)



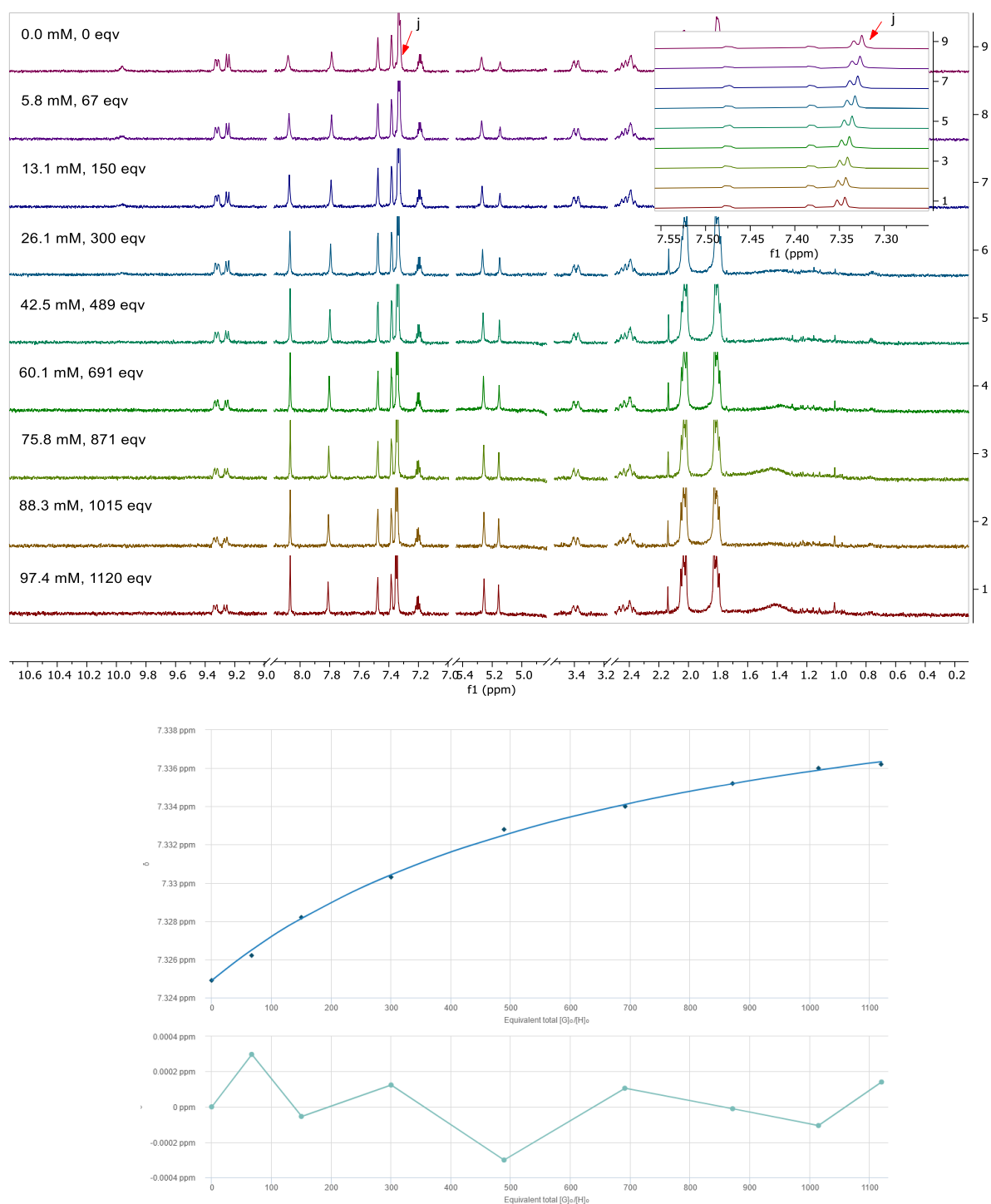
Supplementary Figure 51. Top) ^1H NMR spectra (500 MHz, DMSO- d_6) for receptor **10a** (143 μM) titrated with a combined solution of tetrabutylammonium chloride (20 mM) and receptor **10a** (143 μM). The equivalents and concentrations of guest added are listed in the graph. Bottom) Global fitting of the binding isotherms (protons a, c, b, e, g and l) from Bindfit to a 1:1 model $K_a = 169.7 \text{ M}^{-1}$ ($\pm 0.36 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/2ae8d51d-e58d-4510-a724-f75950f64405>

Receptor 10b & acetate (H₂O/D₂O)



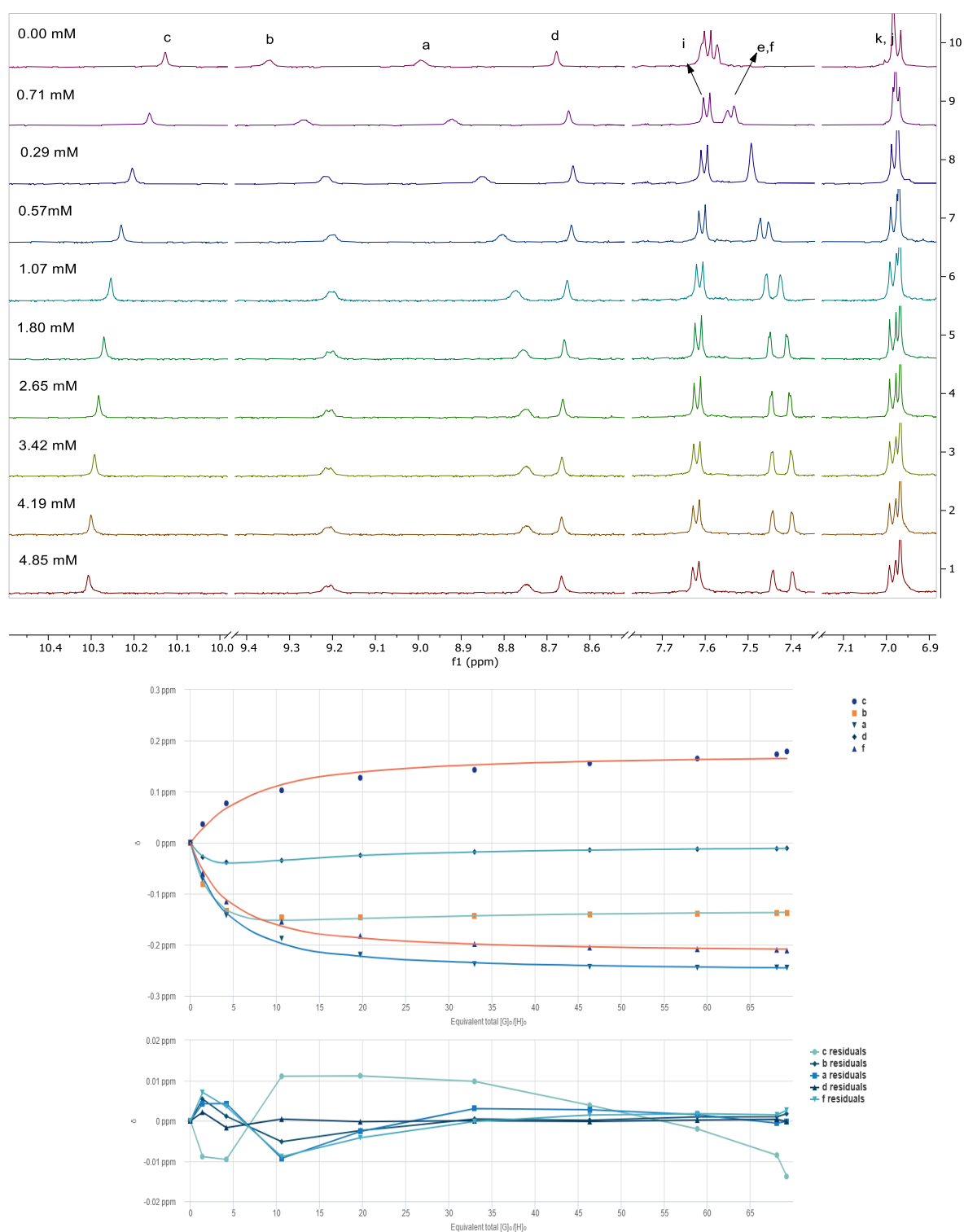
Supplementary Figure 52. Top) ¹H NMR (500 MHz, 9:1 H₂O/D₂O) spectra for receptor **10b** (66.7 μM) titrated with a combined solution of sodium acetate (100 mM) and receptor **10b** (66.7 μM). pH = 7.60 at the end of titration. Bottom). Fitting of the binding isotherm (proton j) from Bindfit to a 1:1 model $K_a = 8.84 \text{ M}^{-1}$ ($\pm 4.28 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/465327ae-6c72-4438-a821-538ef5ed5f58>

Receptor 10b & chloride (H₂O/D₂O)



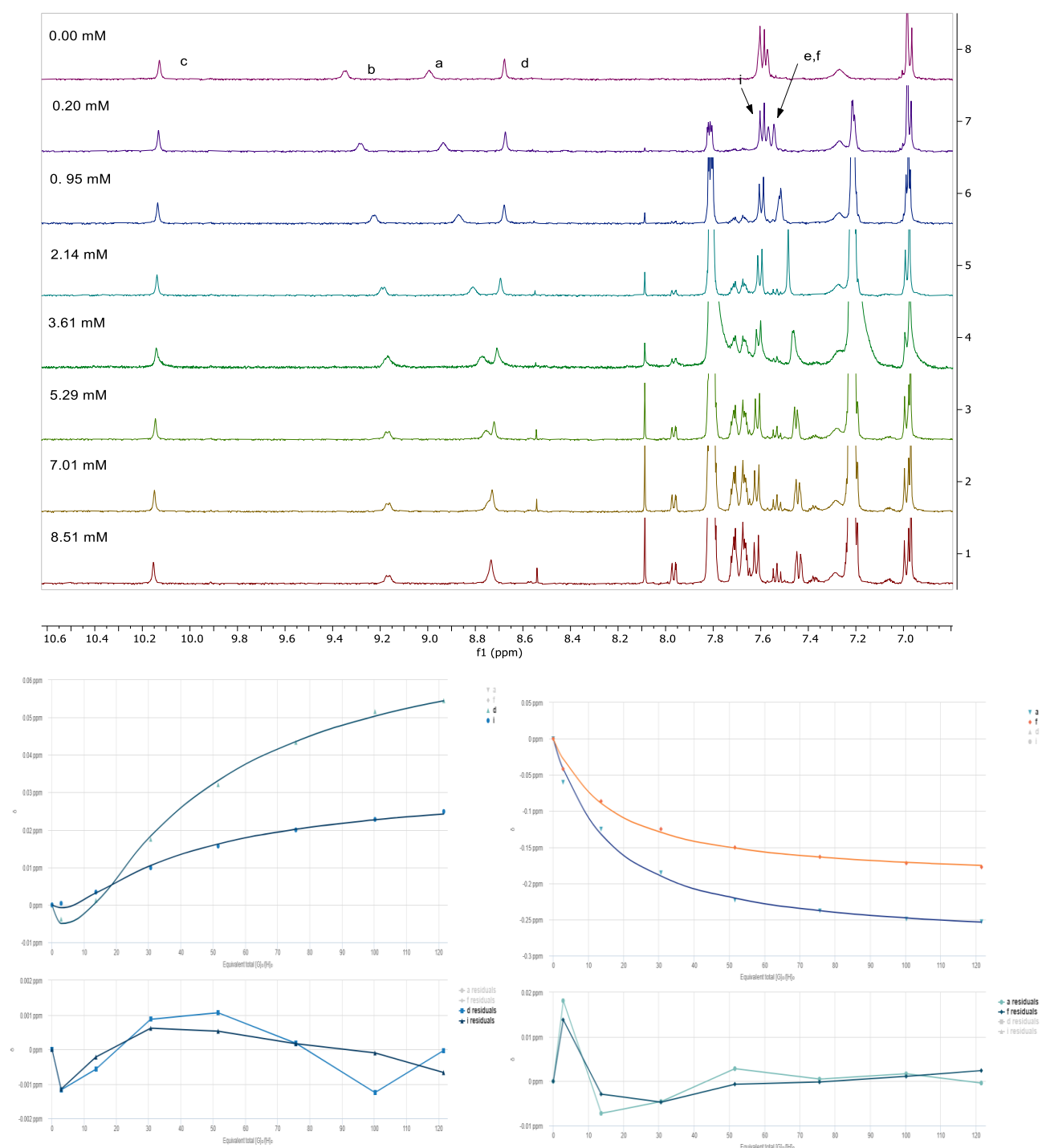
Supplementary Figure 53. Top) ¹H NMR (500 MHz, 9:1 H₂O/D₂O) spectra for receptor **10b** (66.7 μM) titrated with a combined solution of sodium chloride (100 mM) and receptor **10b** (66.7 μM). The concentration of guest is shown on each spectrum. pH = 7.47 at the end of titration. Bottom) Fitting of the binding isotherm (proton j) from Bindfit to a 1:1 model $K_a = 15.97 \text{ M}^{-1}$ ($\pm 4.16 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/1647bddf-5737-4dac-b08d-eea0da2c309b>

Receptor 20 & TBA acetate (DMSO- d_6)



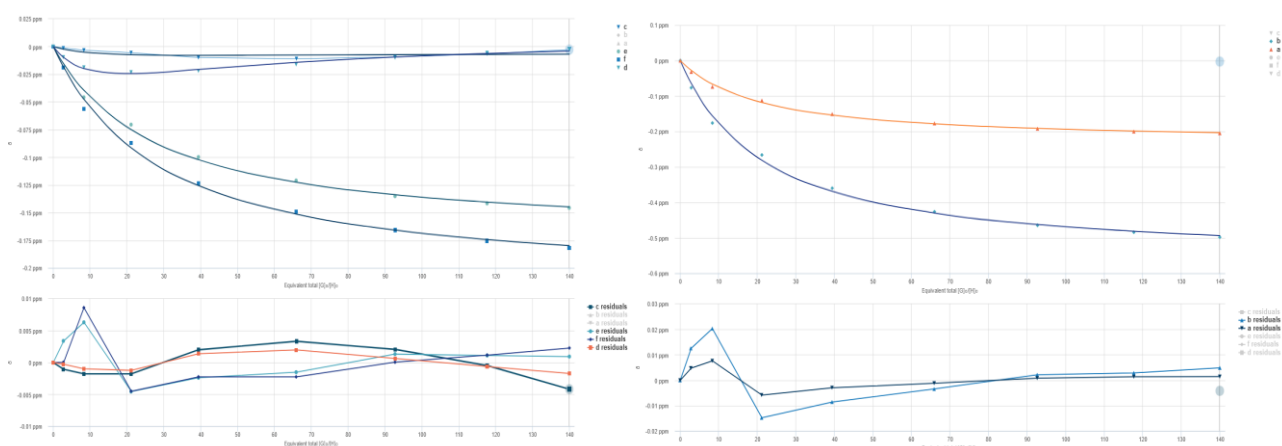
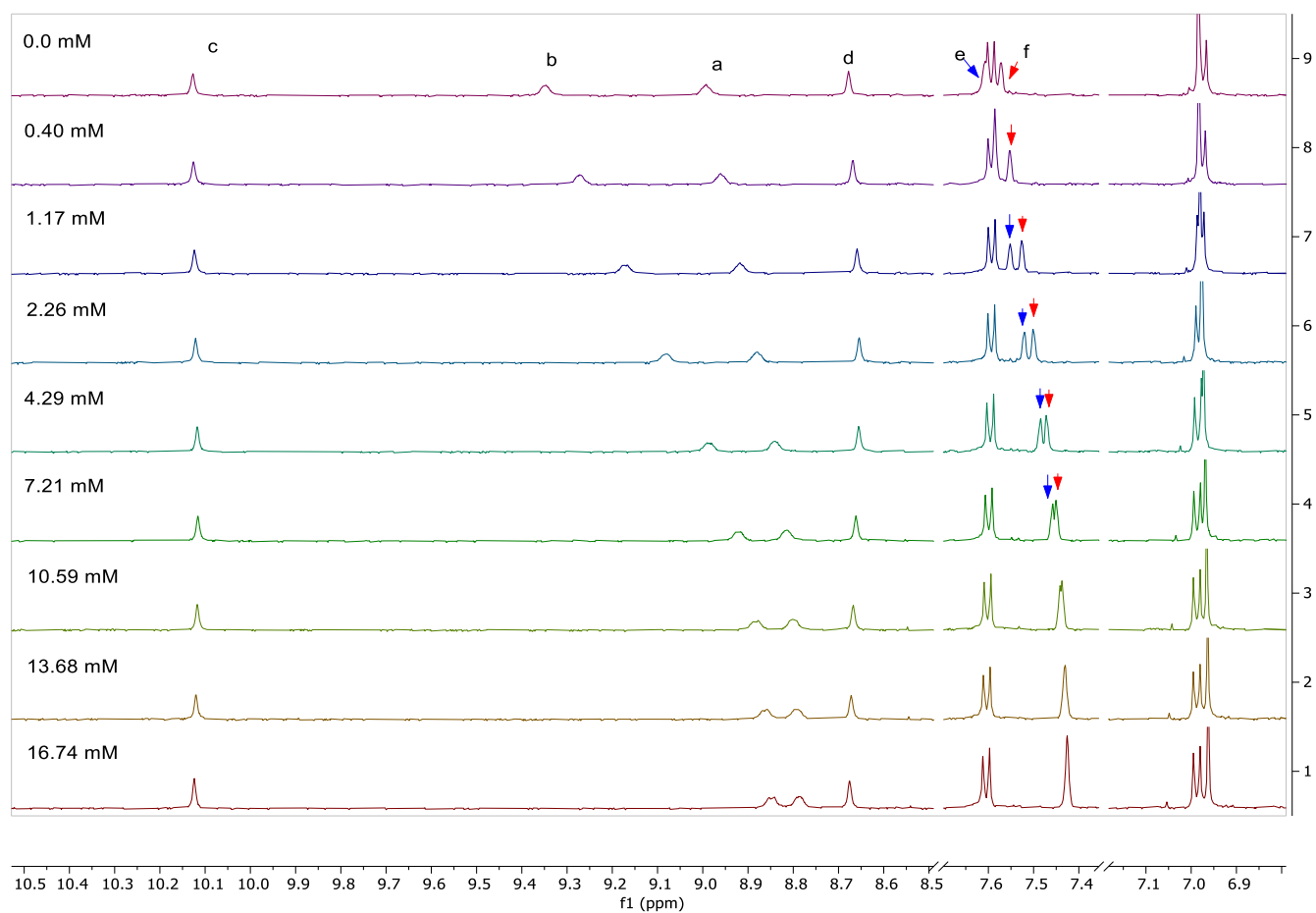
Supplementary Figure 54. Top) ^1H NMR (600 MHz, DMSO- d_6) spectra for receptor **20** (70 μM) titrated with a combined solution of tetrabutylammonium acetate (10 mM) and receptor **20** (70 μM). The concentration of guest is shown on each spectrum. Bottom) Global fitting of the binding isotherms (protons a, b, c, d, f) from Bindfit to a 1:2 non-cooperative model $K_a = 7564.4 \text{ M}^{-1} (\pm 7.4 \%)$. Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/90113f79-6294-4d3b-aad1-e8f96f95ef06>

Receptor 20 & TBA benzoate (DMSO- d_6)



Supplementary Figure 55. Top) ^1H NMR (500 MHz, DMSO- d_6) spectra for receptor **20** (70 μM) titrated with a combined solution of tetrabutylammonium benzoate (14.4 mM) and receptor **20** (70 μM). The concentration of guest is shown on each spectrum. Bottom) Global fitting of the binding isotherms (protons a, d, i and f) from Bindfit to a 1:2 non-cooperative model $K_a = 1650.2 \text{ M}^{-1}$ ($\pm 6.9 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/3bf198b5-89aa-4952-91b4-4ac1e54f20ef>

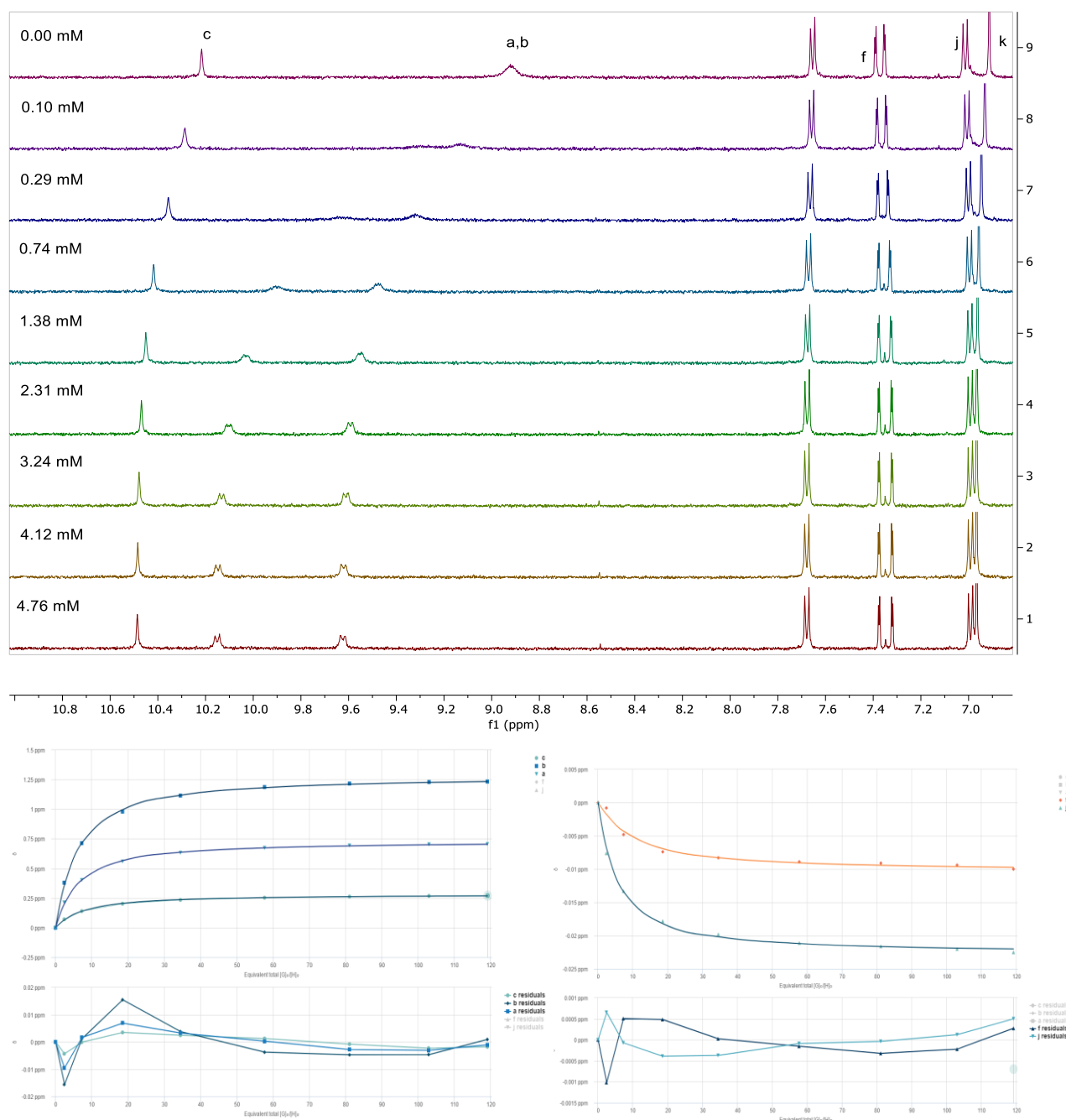
Receptor 20 & TBA chloride (DMSO- d_6)



Supplementary Figure 56. Top) ^1H NMR (600 MHz, $\text{DMSO}-d_6$) spectra for receptor **20** (70 μM) titrated with a combined solution of TBA chloride (40 mM) and receptor **20** (70 μM). The concentration of guest is shown on each spectrum. Bottom) Global fitting of the binding isotherms (protons a, b, c, d, e and f) from Bindfit to a 1:2 non-cooperative model $K_a = 1241.3 \text{ M}^{-1} (\pm 3.5 \%)$. Full fitted data is available online at:

<http://app.supramolecular.org/bindfit/view/aef14313-31d8-4dde-9c31-b34823ed8900>

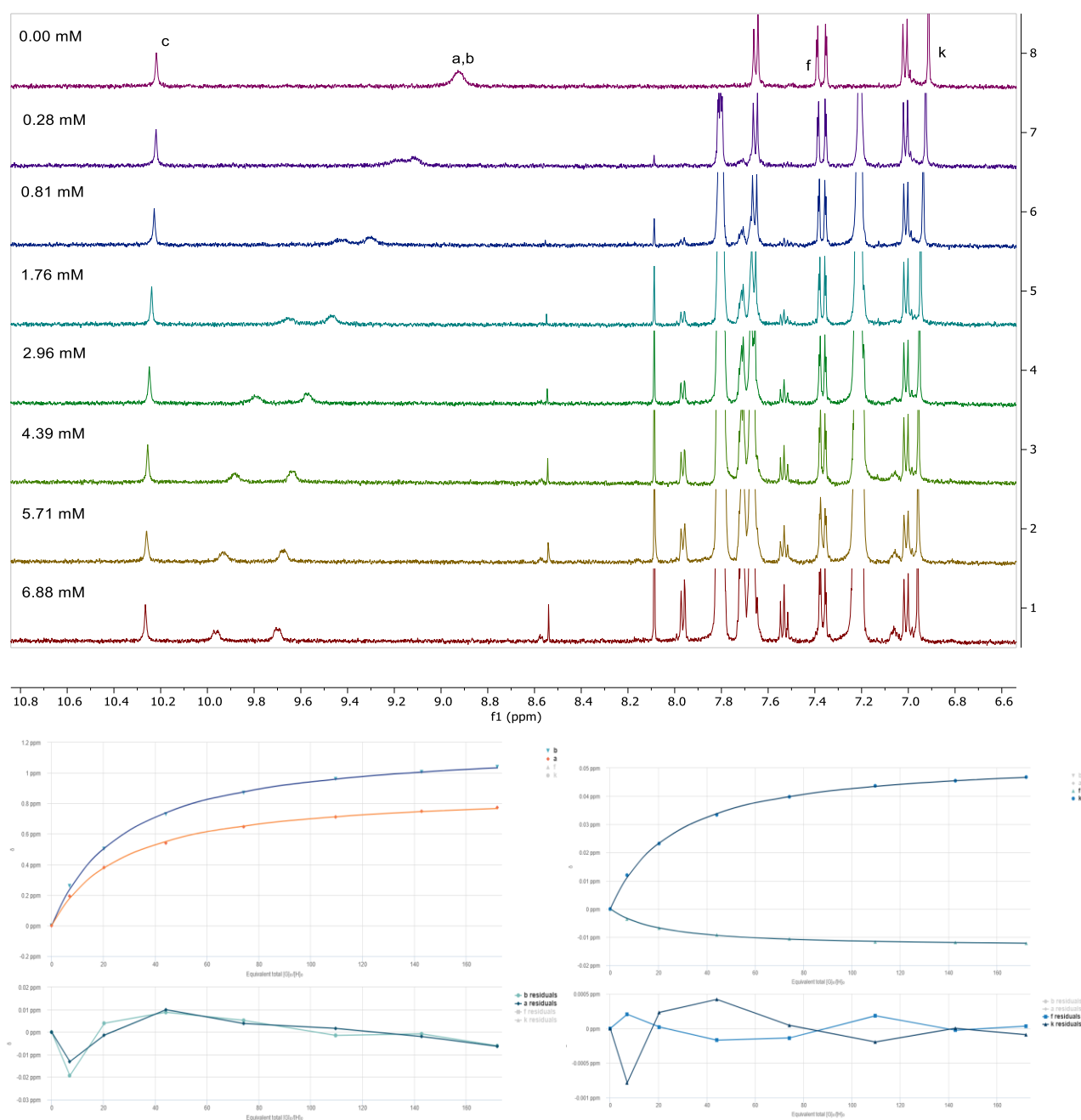
Receptor 21 & TBA acetate (DMSO- d_6)



Supplementary Figure 57. Top) ^1H NMR (500 MHz, DMSO- d_6) spectra for receptor **21** (40 μ M) titrated with a combined solution of TBA Acetate (10 mM) and receptor **21** (40 μ M). The concentration of guest is shown on each spectrum. Bottom) Global fitting of the binding isotherms (protons a, b, c, f, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 8354.8 \text{ M}^{-1}$ ($\pm 1.6 \%$). Full fitted data is available online at:

<http://app.supramolecular.org/bindfit/view/f85458bb-d149-47b4-ad6c-7beabe15d1fe>

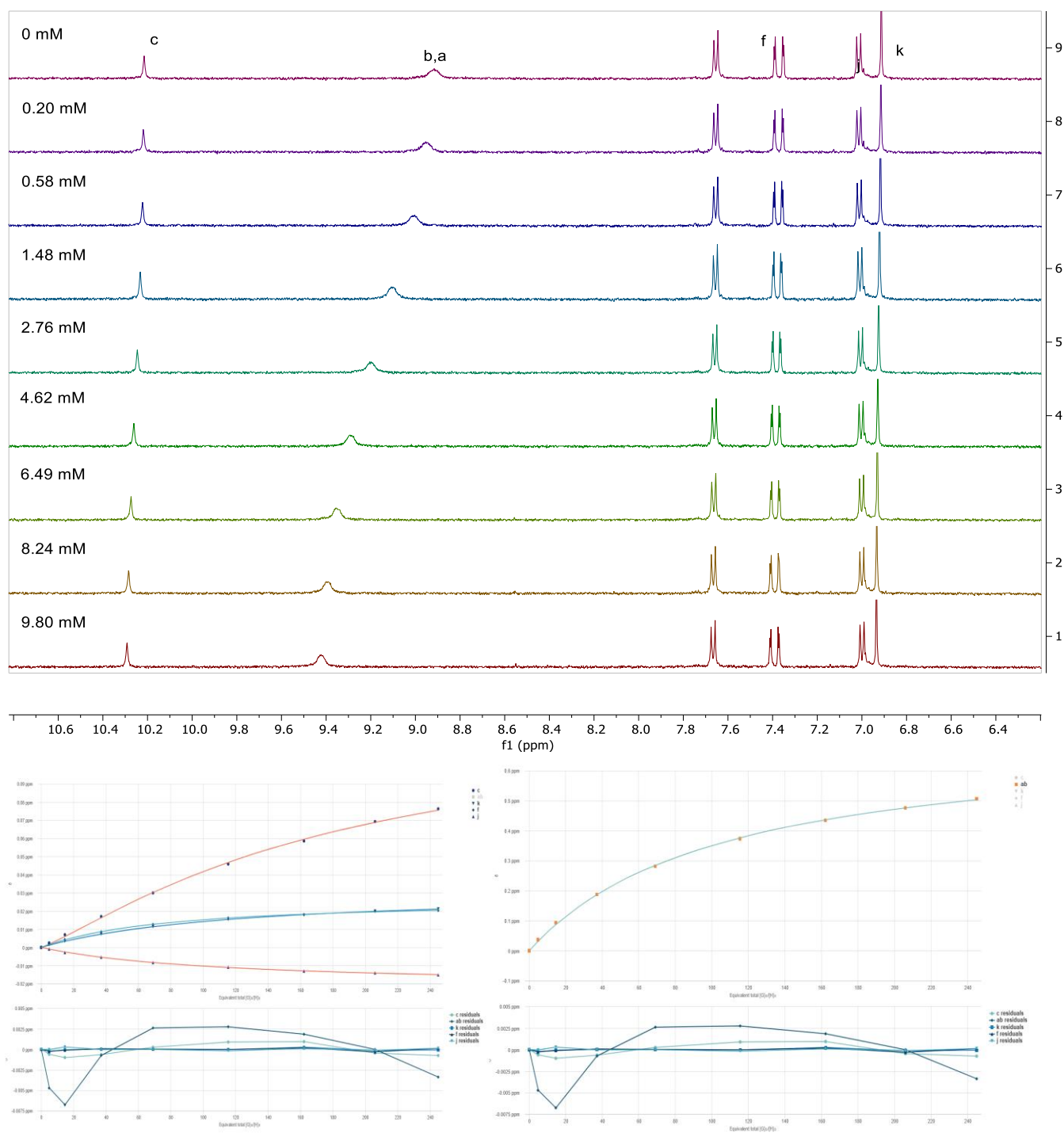
Receptor 21 & TBA benzoate (DMSO- d_6)



Supplementary Figure 58. Top) ^1H NMR (500 MHz, DMSO- d_6) spectra for receptor **21** (40 μM) titrated with a combined solution of TBA Benzoate (14.4 mM) and receptor **21** (40 μM). The concentration of guest is shown on each spectrum. Bottom) Global fitting of the binding isotherms (protons a, b, f and k) from Bindfit to a 1:2 non-cooperative model $K_a = 1820.6 \text{ M}^{-1}$ ($\pm 1.8 \%$). Full fitted data is available online at:

<http://app.supramolecular.org/bindfit/view/28aace97-2a48-4dec-993a-b5f988cecd3a>

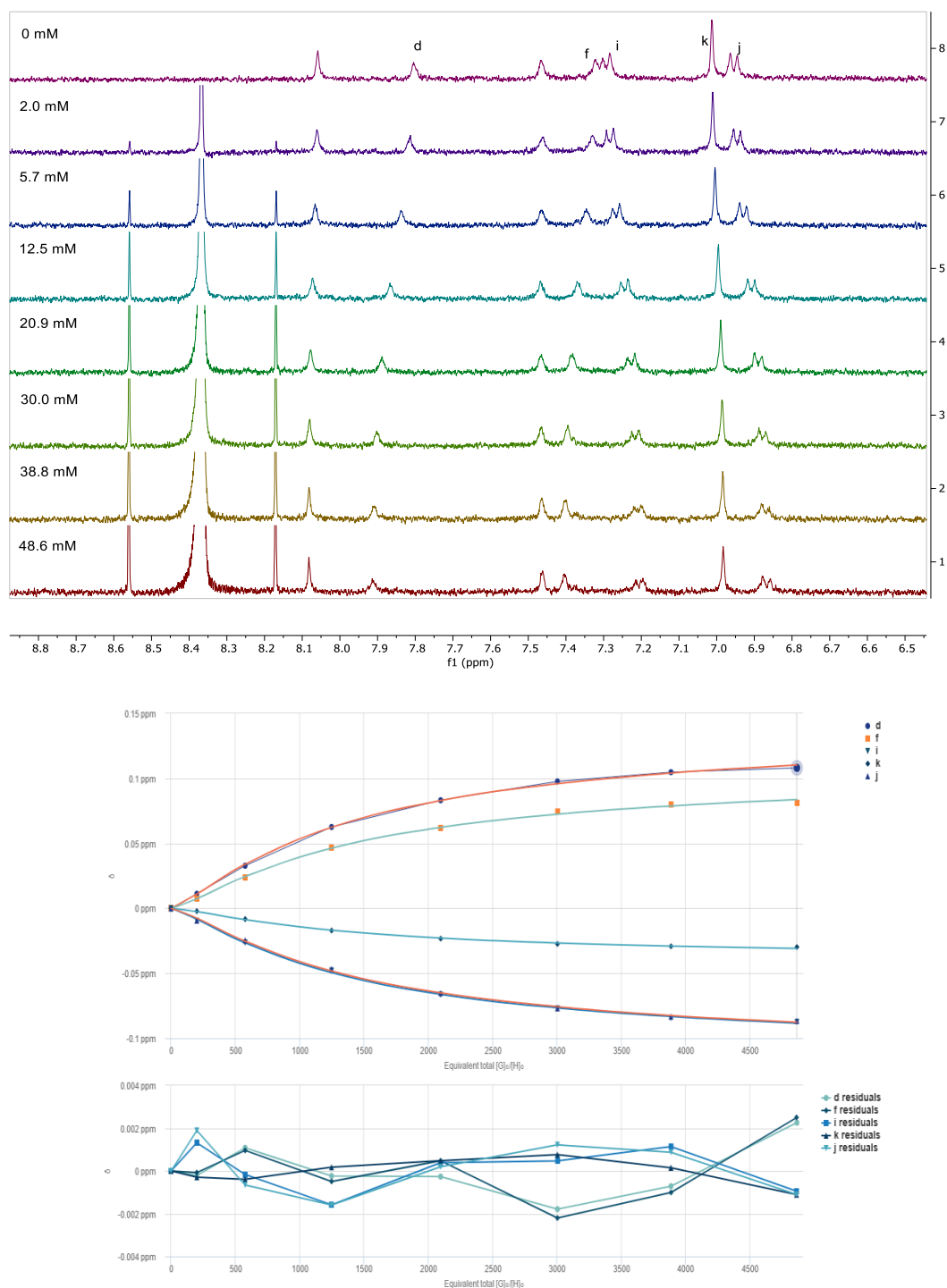
Receptor 21 & TBA chloride (DMSO- d_6)



Supplementary Figure 59. Top) ^1H NMR (500 MHz, DMSO- d_6) spectra for receptor **21** (40 μM) titrated with a combined solution of TBA chloride (20mM) and receptor **21** (40 μM). The concentration of guest is shown on each spectrum. Bottom) Global fitting of the binding isotherms (protons a, b, c, j, f and k) from Bindfit to a 1:2 non-cooperative model $K_a = 458.6 \text{ M}^{-1}$ ($\pm 0.87 \%$). Full fitted data is available online at:

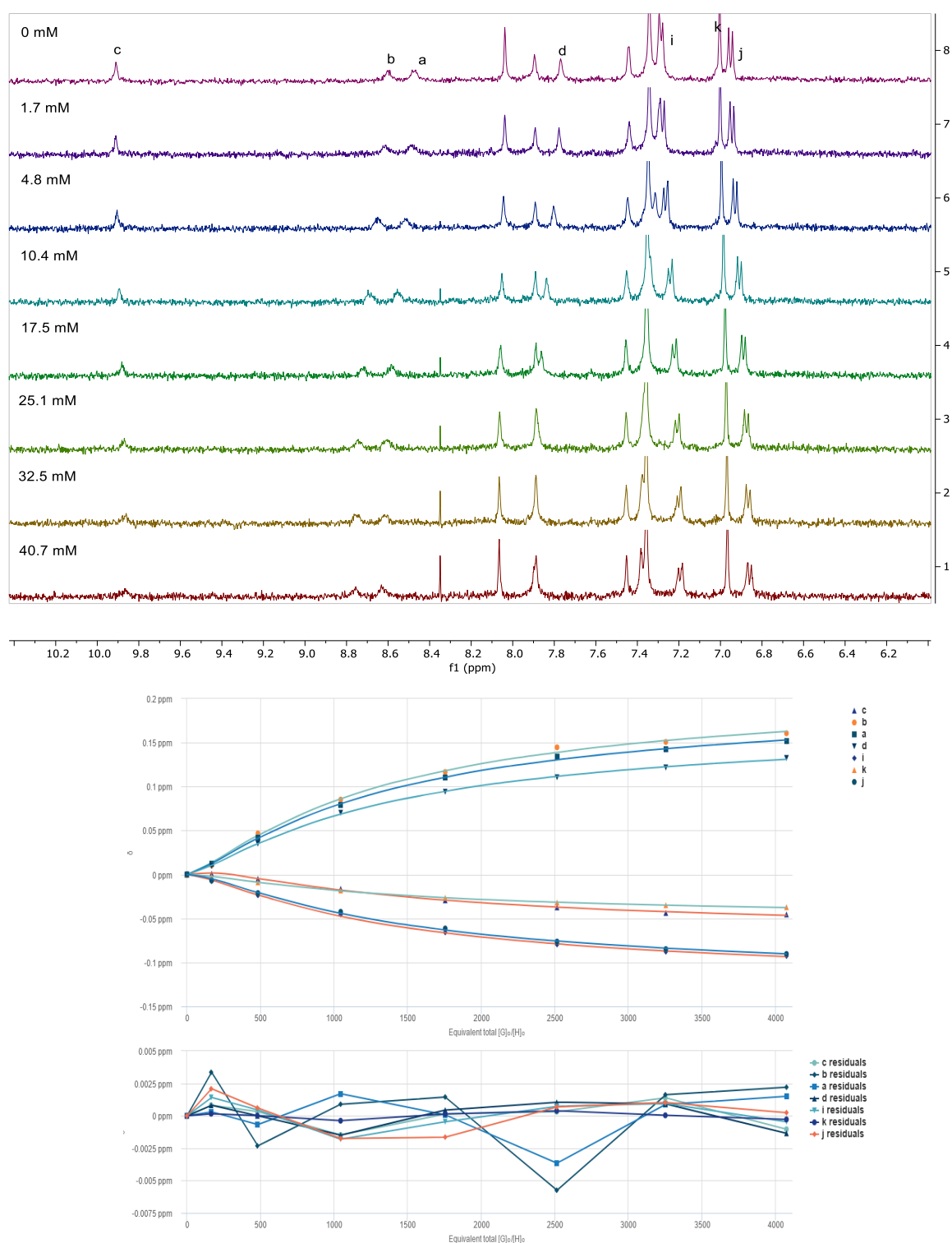
<http://app.supramolecular.org/bindfit/view/a33fa31c-141c-4383-8b9b-1e88ad02c2d1>

Receptor 5 & formate (D₂O)



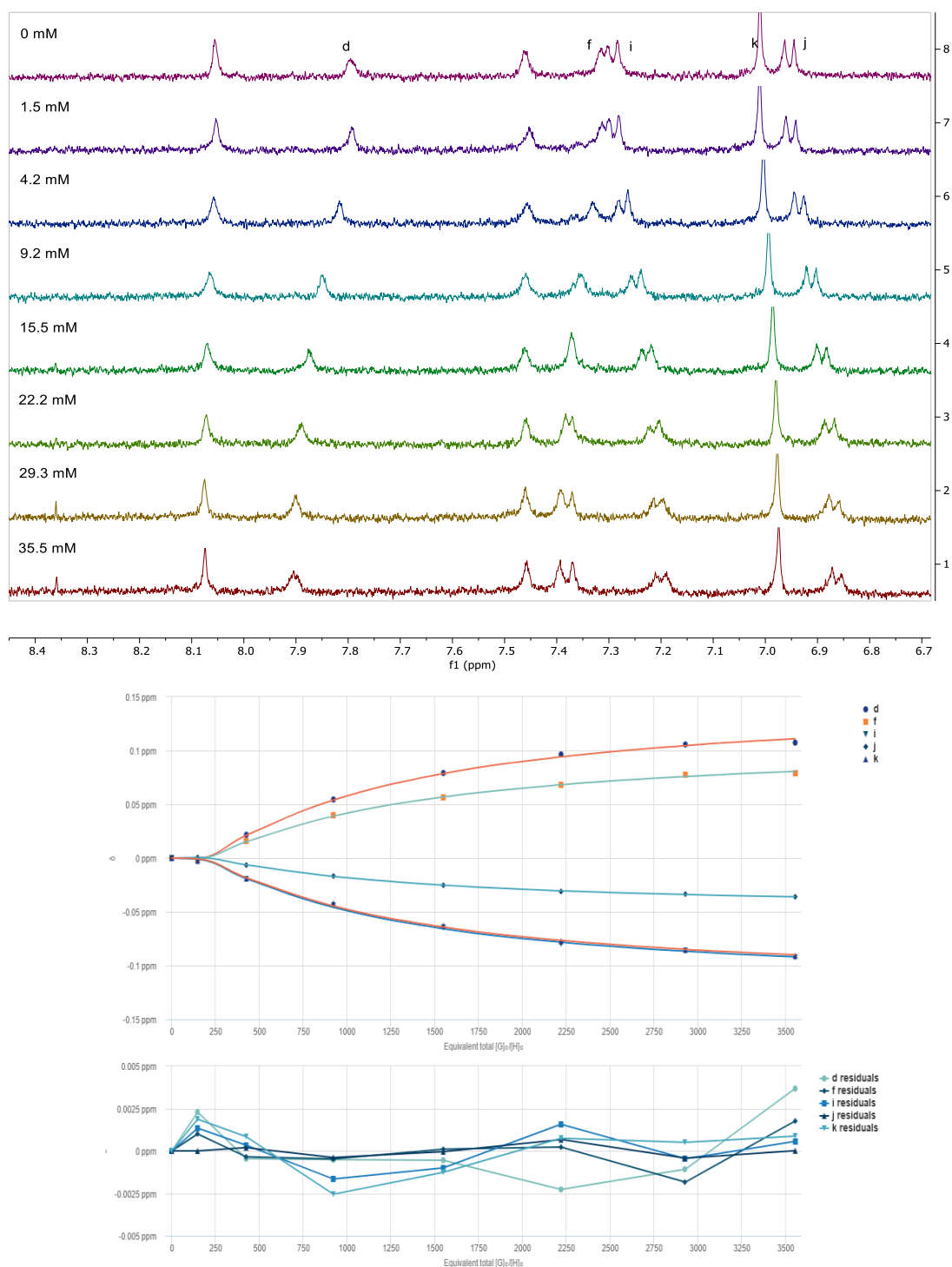
Supplementary Figure 60. Top) ¹H NMR (500 MHz, D₂O) spectra for receptor **5** (10 μM) titrated with a combined solution of sodium formate (102 mM) and receptor **5** (10 μM). The concentration of guest is shown on each spectrum. pH = 7.59 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons d, f, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 268.7 \text{ M}^{-1}$ ($\pm 1.8 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/c364696e-6e86-4596-875a-479a3a95a37e>

Receptor 5 & acetate (H₂O/D₂O)



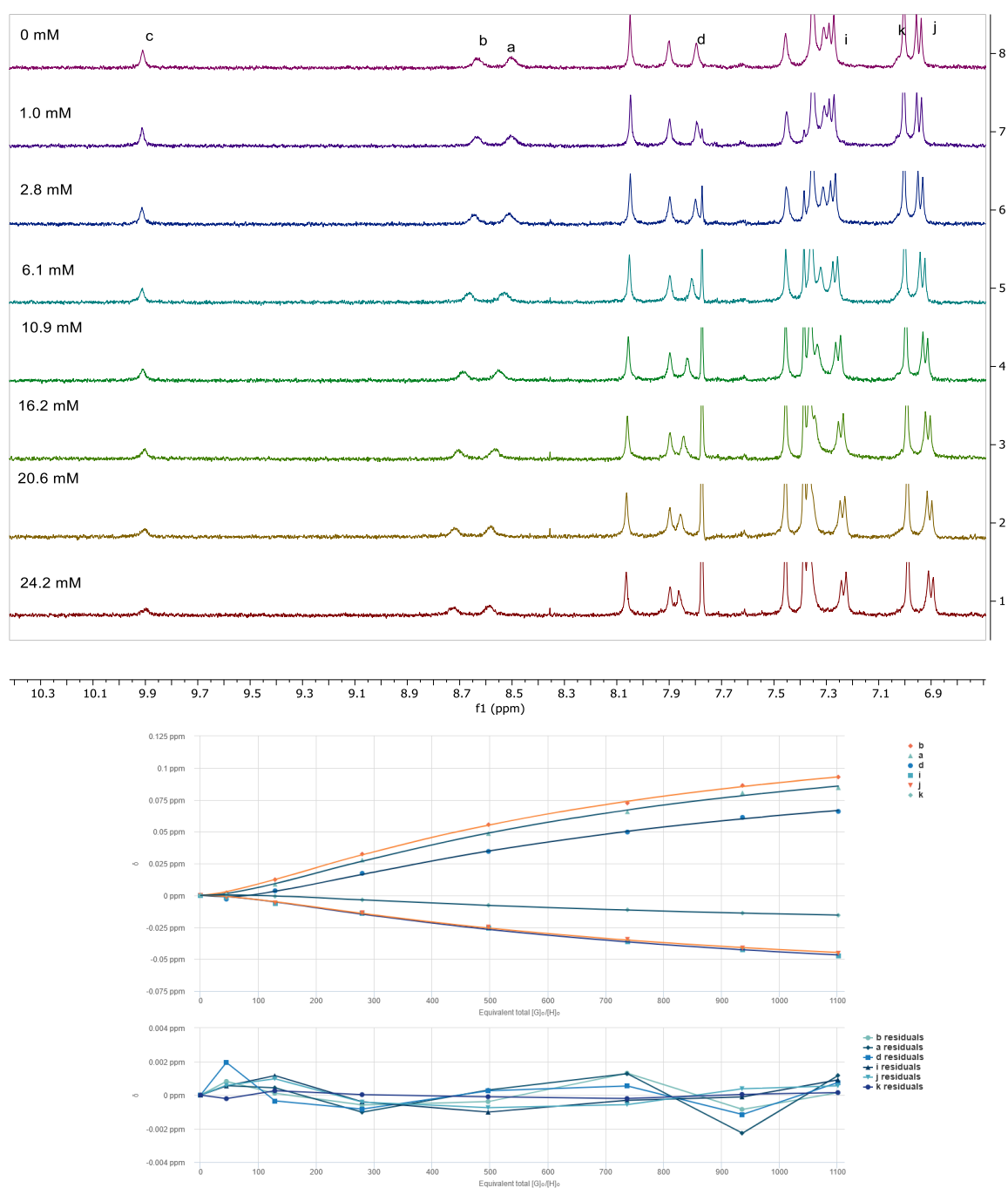
Supplementary Figure 61. Top) ¹H NMR (500 MHz, 9:1 H₂O/D₂O) spectra for receptor **5** (10 μM) titrated with a combined solution of sodium acetate (85 mM) and receptor **5** (10 μM). The concentration of guest is shown on each spectrum. pH = 7.41 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons a, b, c, d, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 279.2 \text{ M}^{-1}$ ($\pm 1.4 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/61d190f1-8139-465c-b285-8893d28f4643>

Receptor 5 & propionate (D₂O)



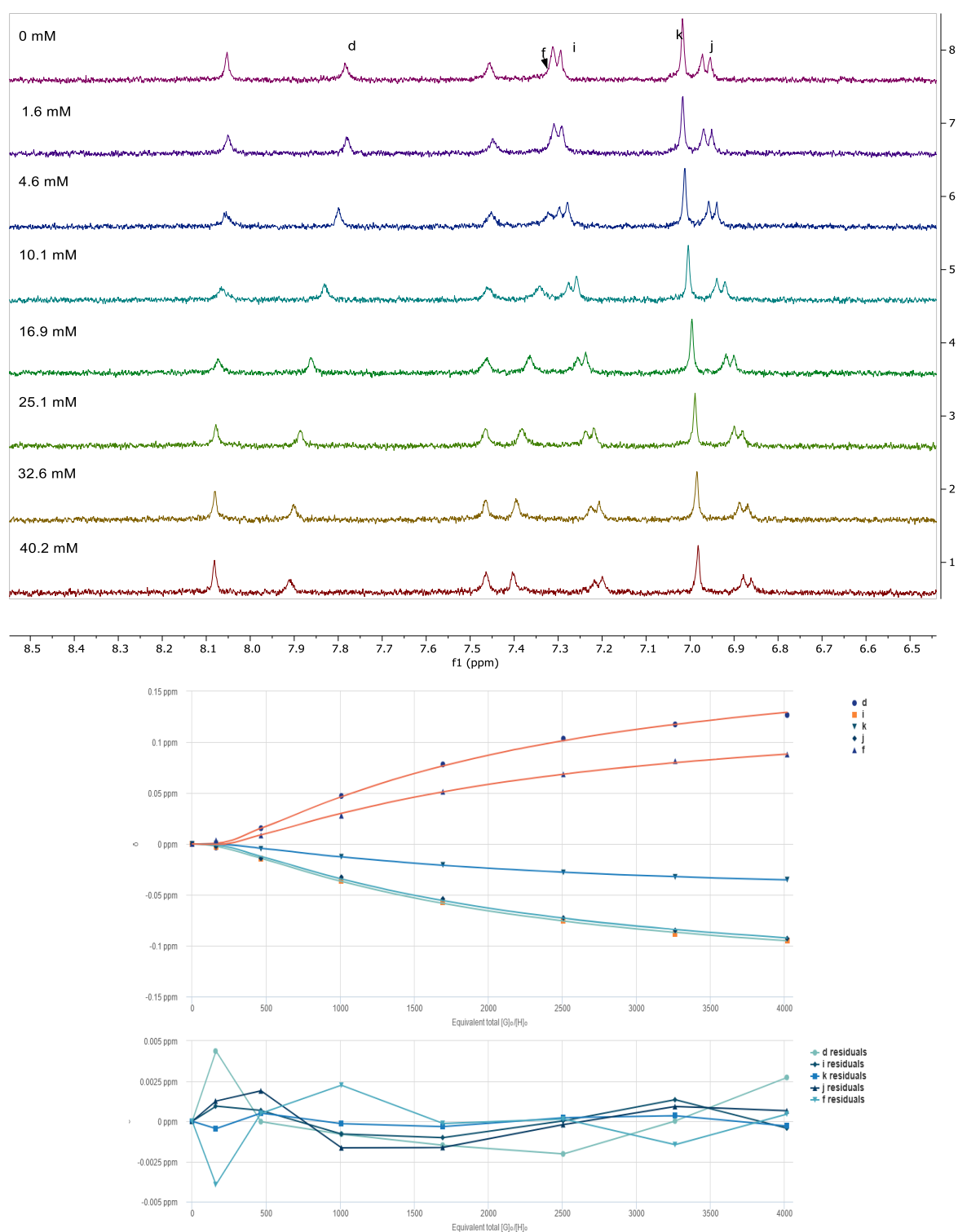
Supplementary Figure 62. Top) ¹H NMR (500 MHz, D₂O) spectra for receptor **5** (10 μM) titrated with a combined solution of sodium propionate (75 mM) and receptor **5** (10 μM). The concentration of guest is shown on each spectrum. pH = 7.52 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons d, f, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 415.0 \text{ M}^{-1} (\pm 1.8 \%)$. Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/a60d53a7-07de-4be7-bcf4-7442284016f5>

Receptor 5 & benzoate (H₂O/D₂O)

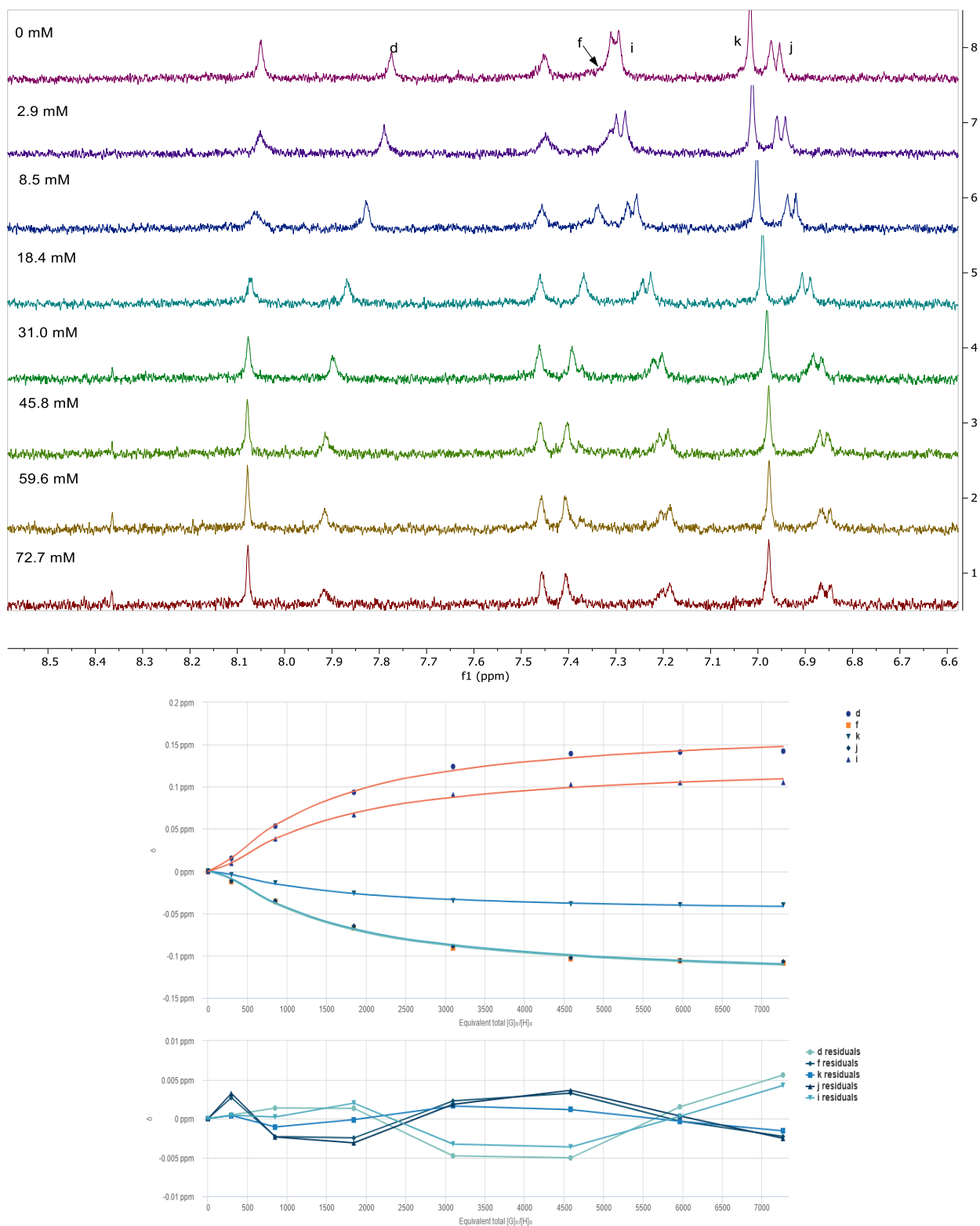


Supplementary Figure 63. Top) ¹H NMR (500 MHz, 9:1 H₂O/D₂O) spectra for receptor **5** (22 μM) titrated with a combined solution of sodium benzoate-*d*⁵ (50 mM) and receptor **5** (22 μM). The concentration of guest is shown on each spectrum. pH = 7.48 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons a, b, d, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 230.7 \text{ M}^{-1}$ ($\pm 1.0 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/83499776-e330-4edd-bbe7-8692016bf1e7>

Receptor 5 & *L* and *D*-Lactate (D₂O)

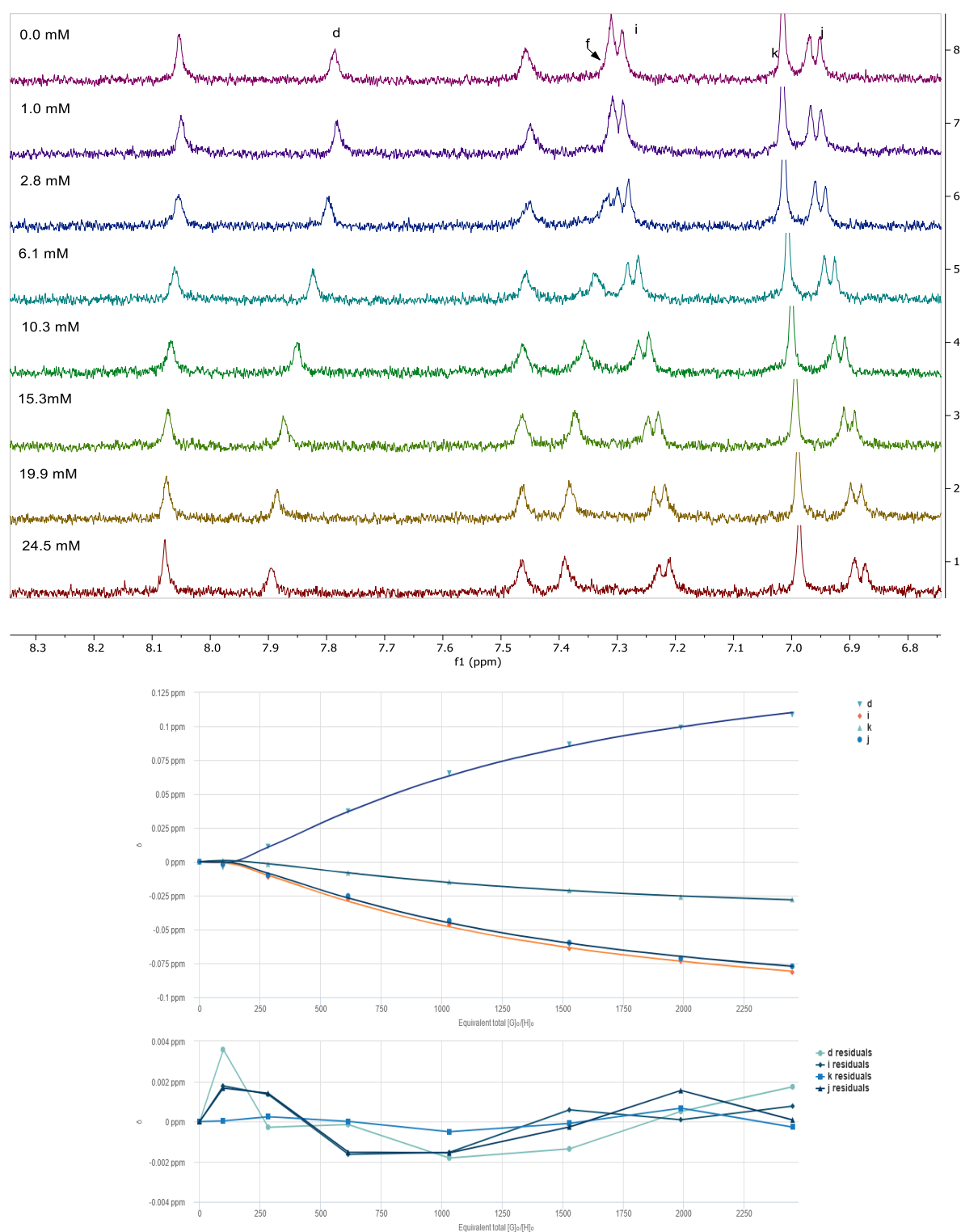


Supplementary Figure 64. Top) ¹H NMR (500 MHz, D₂O) spectra for receptor **5** (10 μM) titrated with a combined solution of sodium *L*-Lactate (82 mM) and receptor **5** (10 μM). The concentration of guest is shown on each spectrum. pH = 7.58 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons d, f, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 204.9 \text{ M}^{-1}$ ($\pm 1.6 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/75f1b5ed-b6dc-4254-ab2d-bf846df47fbd>



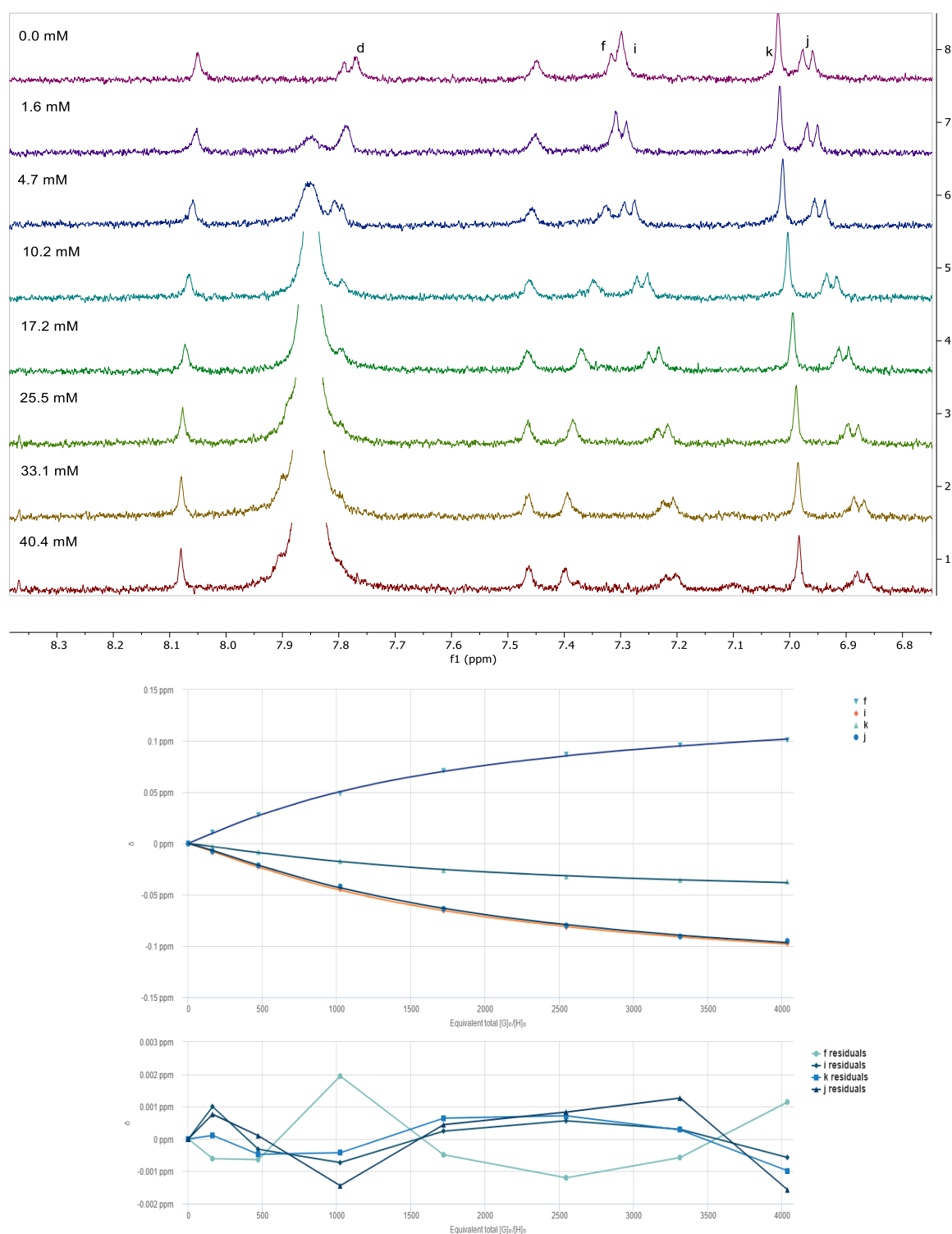
Supplementary Figure 65. Top) ^1H NMR (500 MHz, D_2O) spectra for receptor **5** (10 μM) titrated with a combined solution of sodium *D*-Lactate (150 mM) and receptor **5** (10 μM). The concentration of guest is shown on each spectrum. pH = 7.57 at the end of the titration Bottom) Global fitting of the binding isotherms (protons d, f, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 304.5 \text{ M}^{-1}$ ($\pm 3.5 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/10e4eb1e-063b-4243-9274-b83df416f8d0>

Receptor 5 & *O*-Ac-*L*-Lactate (D₂O)

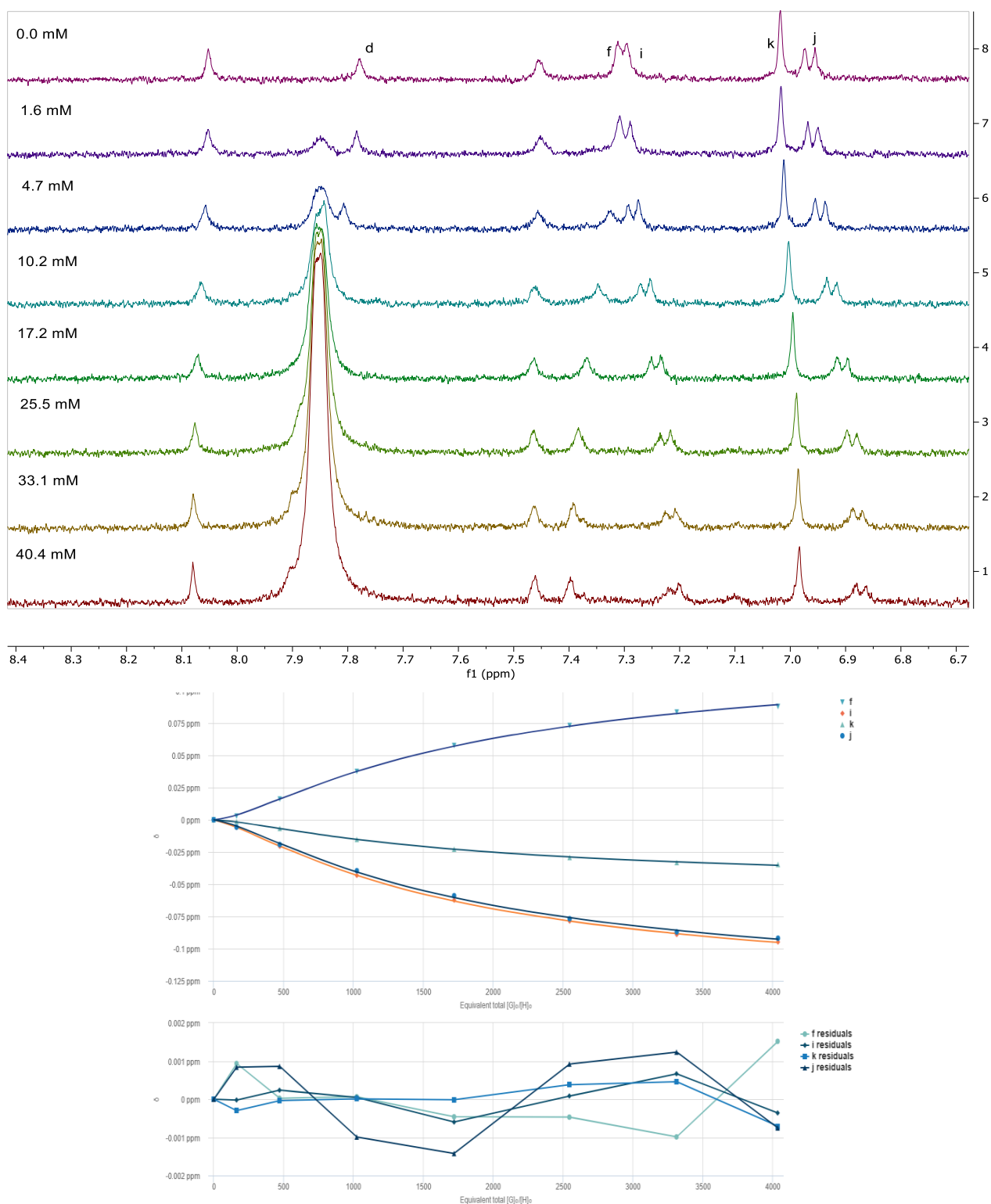


Supplementary Figure 66. Top) ¹H NMR (500 MHz, D₂O) spectra for receptor 5 (10 μM) titrated with a combined solution of sodium *O*-Ac-*L*-Lactate (50 mM) and receptor 5 (10 μM). The concentration of guest is shown on each spectrum. pH = 7.39 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons d, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 330.8 \text{ M}^{-1} (\pm 1.8 \%)$. Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/8a5adc39-1efc-409d-91f2-f3a868131b10>

Receptor 5 & *N*-Ac-*L* and *D*-Alanine (D₂O)

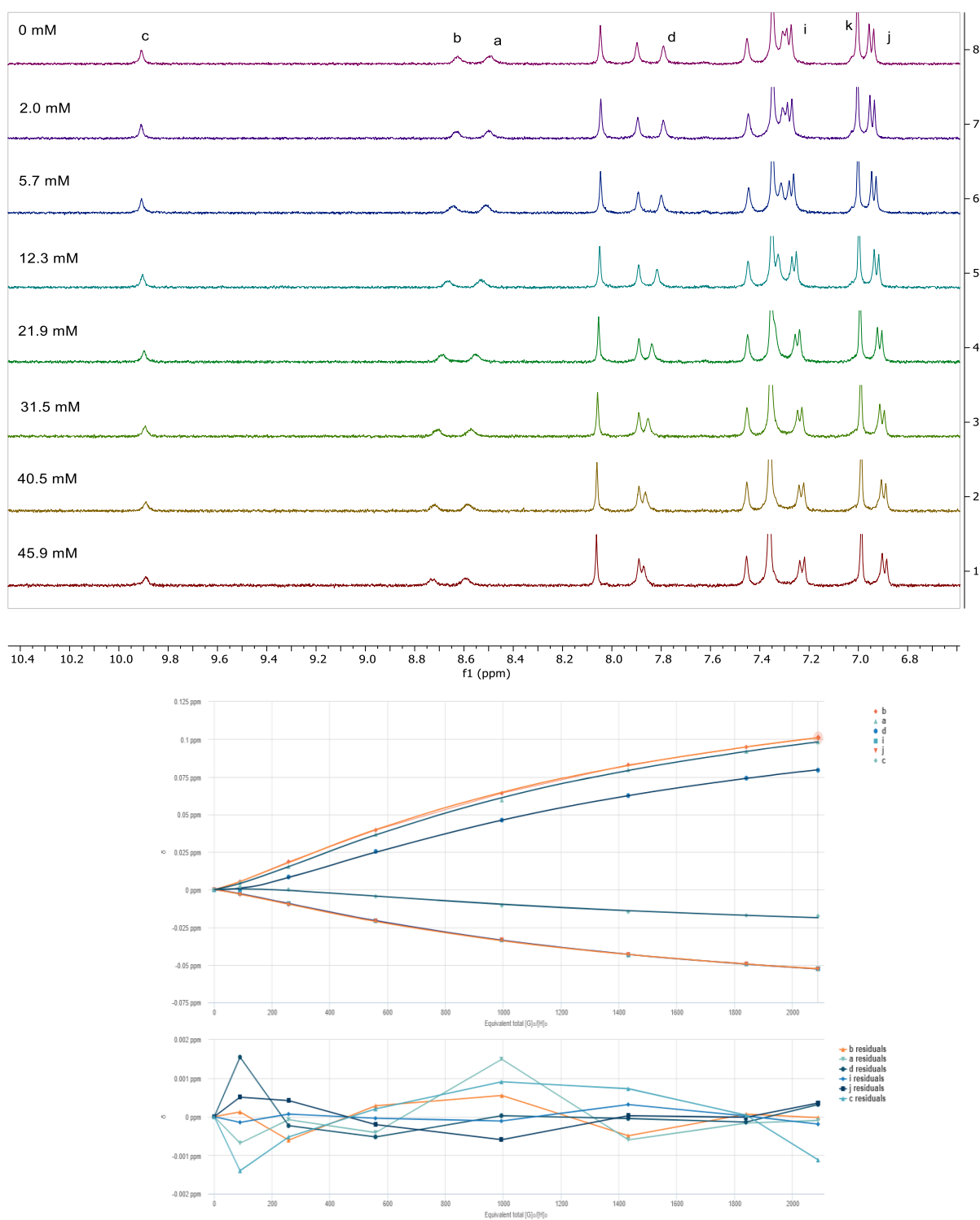


Supplementary Figure 67. Top) ¹H NMR (500 MHz, D₂O) spectra for receptor **5** (10 μM) titrated with a combined solution of sodium *N*-Ac-*L*-Alanine (83 mM) and receptor **5** (10 μM). The concentration of guest is shown on each spectrum. pH = 7.47 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons f, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 177.3 \text{ M}^{-1} (\pm 1.3 \%)$. Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/45c5ef57-73b9-4916-b5ed-2c96e6b45a23>



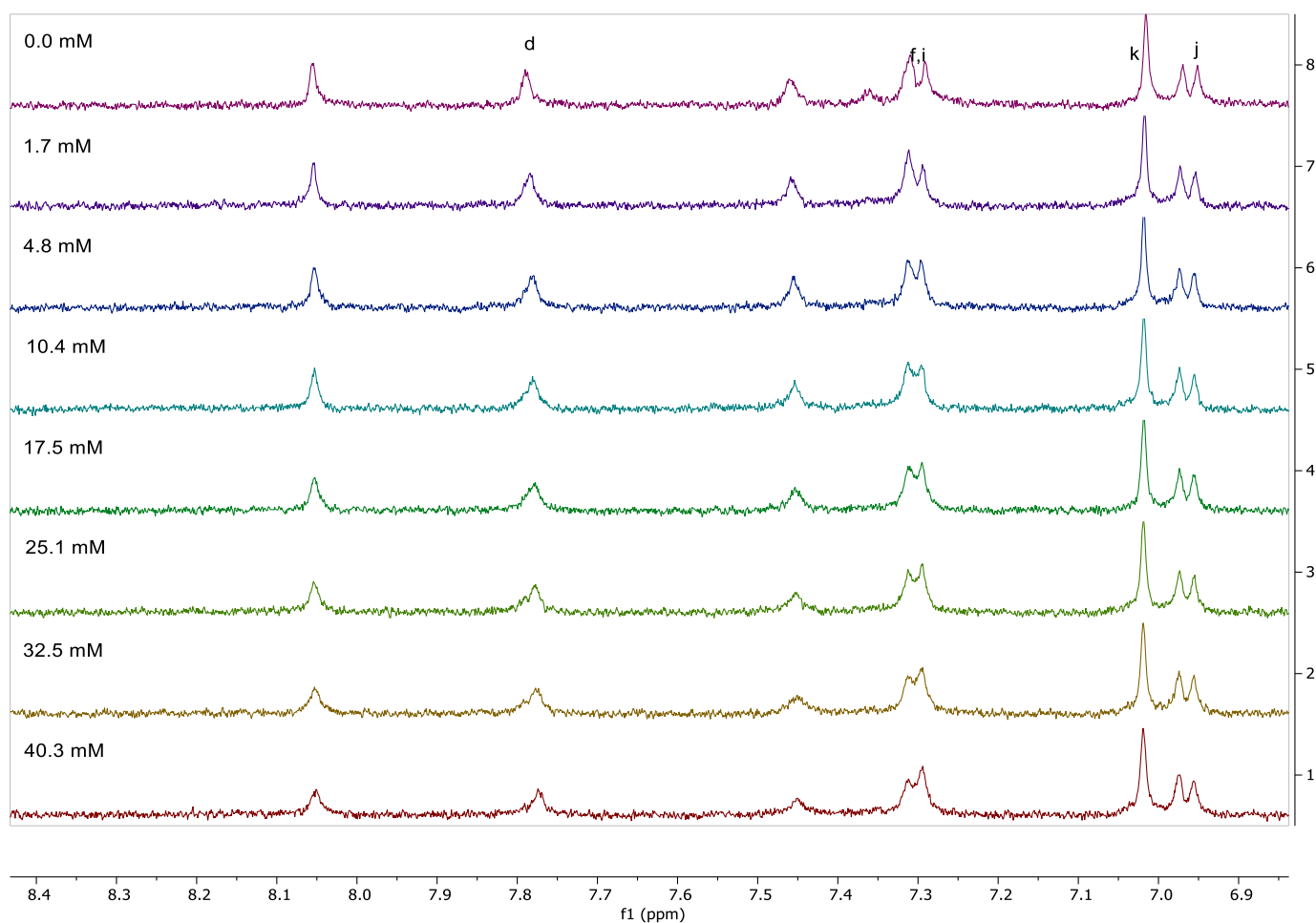
Supplementary Figure 68. Top) ¹H NMR (500 MHz, D₂O) spectra for receptor **5** (10 μM) titrated with a combined solution of sodium *N*-Ac-*D*-Alanine (83 mM) and receptor **5** (10 μM). The concentration of guest shown on each spectrum. pH = 7.42 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons f, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 205.5 \text{ M}^{-1}$ ($\pm 1.1 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/37f30eeb-a939-405f-a25b-c8da02febd96>

Receptor 5 & chloride (H₂O/D₂O)

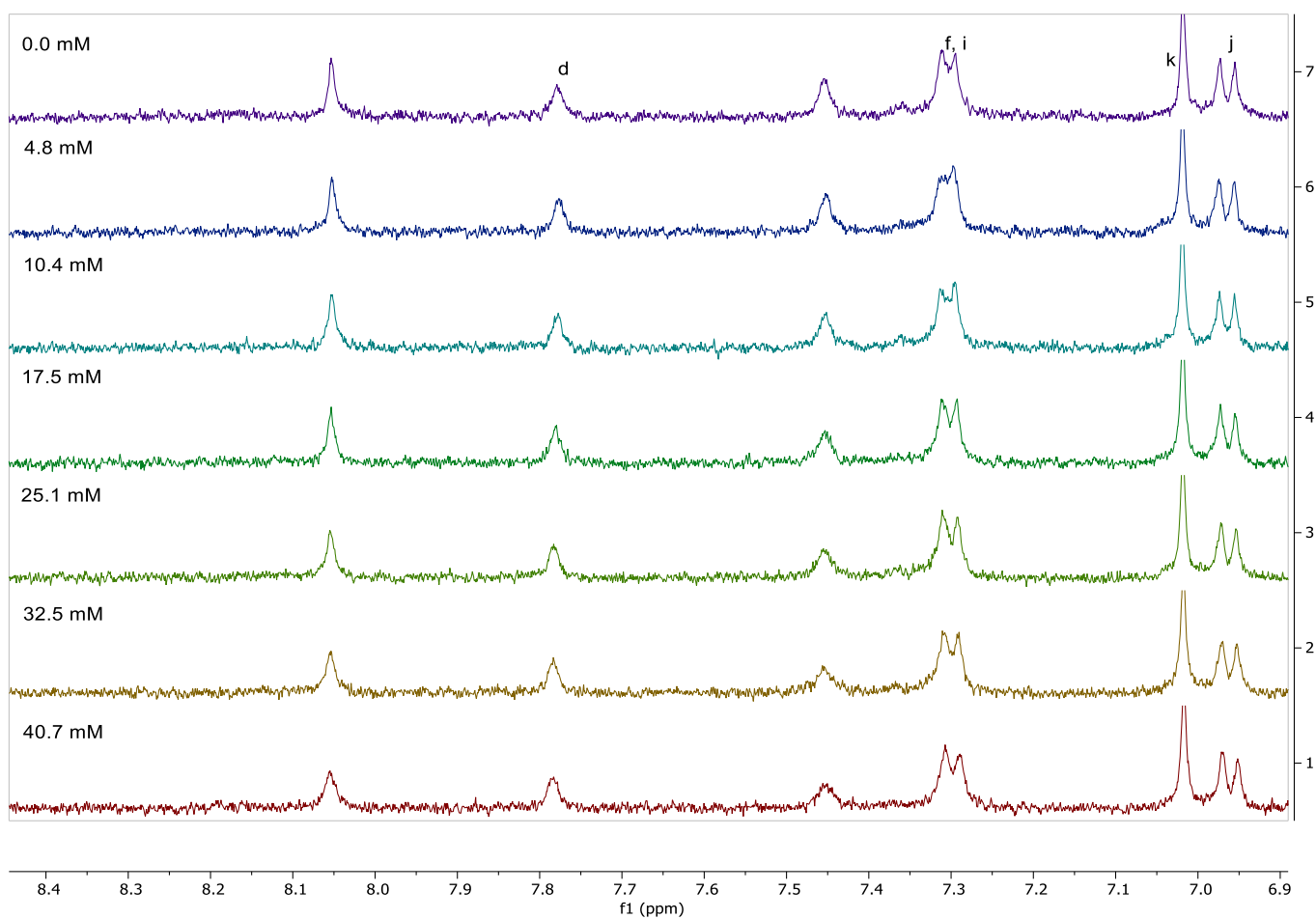


Supplementary Figure 69. Top) ¹H NMR (500 MHz, 9:1 H₂O/D₂O) spectra for receptor **5** (22 μM) titrated with a combined solution of sodium chloride (100 mM) and receptor **5** (22 μM). The concentration of guest is shown on each spectrum. pH = 7.43 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons a, b, c, d, i and j) from Bindfit to a 1:2 non-cooperative model $K_a = 114.9 \text{ M}^{-1}$ ($\pm 0.6 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/3cde4c30-29e0-4b7e-85f3-65e4c55a62f3>

Receptor 5 & *L/D*-Alanine (D₂O)

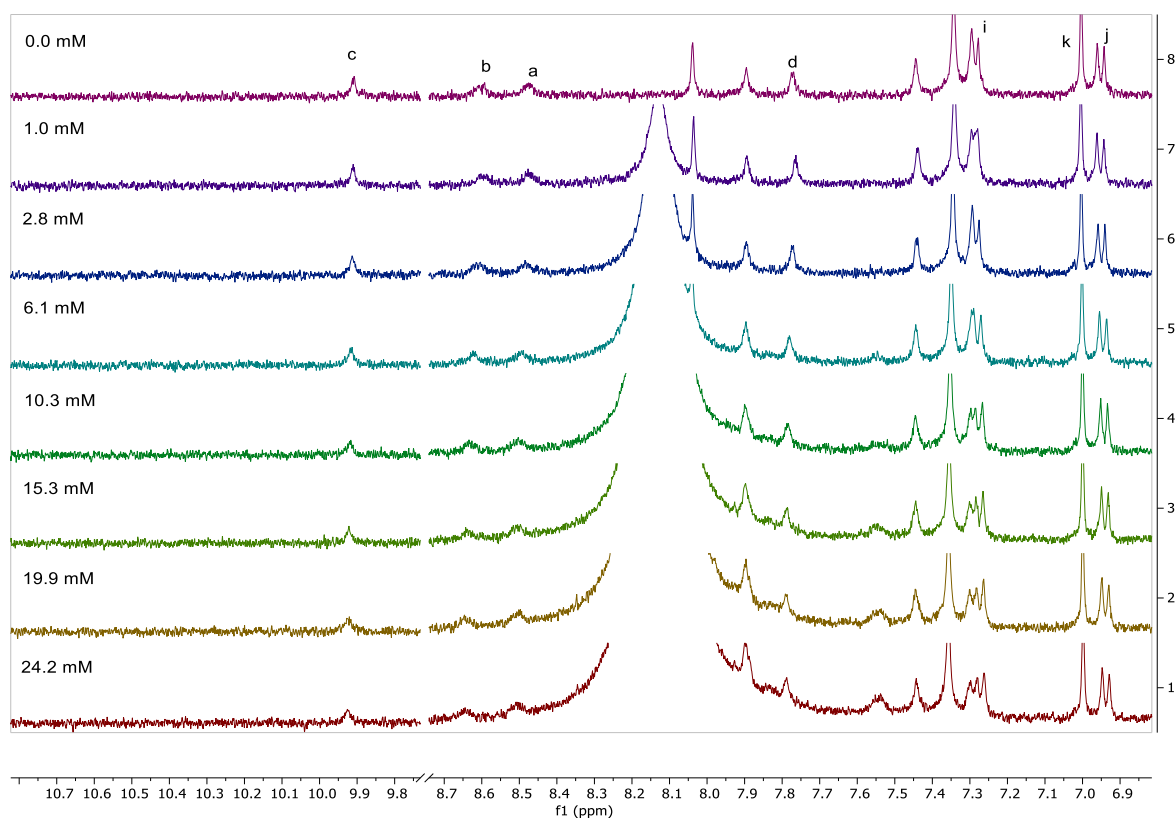


Supplementary Figure 70. ¹H NMR (500 MHz, D₂O) spectra for receptor **5** (10 μM) titrated with a combined solution of sodium *L*-Alanine (85 mM) and receptor **5** (10 μM). The concentration of guest is shown on each spectrum. pH = 7.50 at the end of the titration. No peak movements were observed.



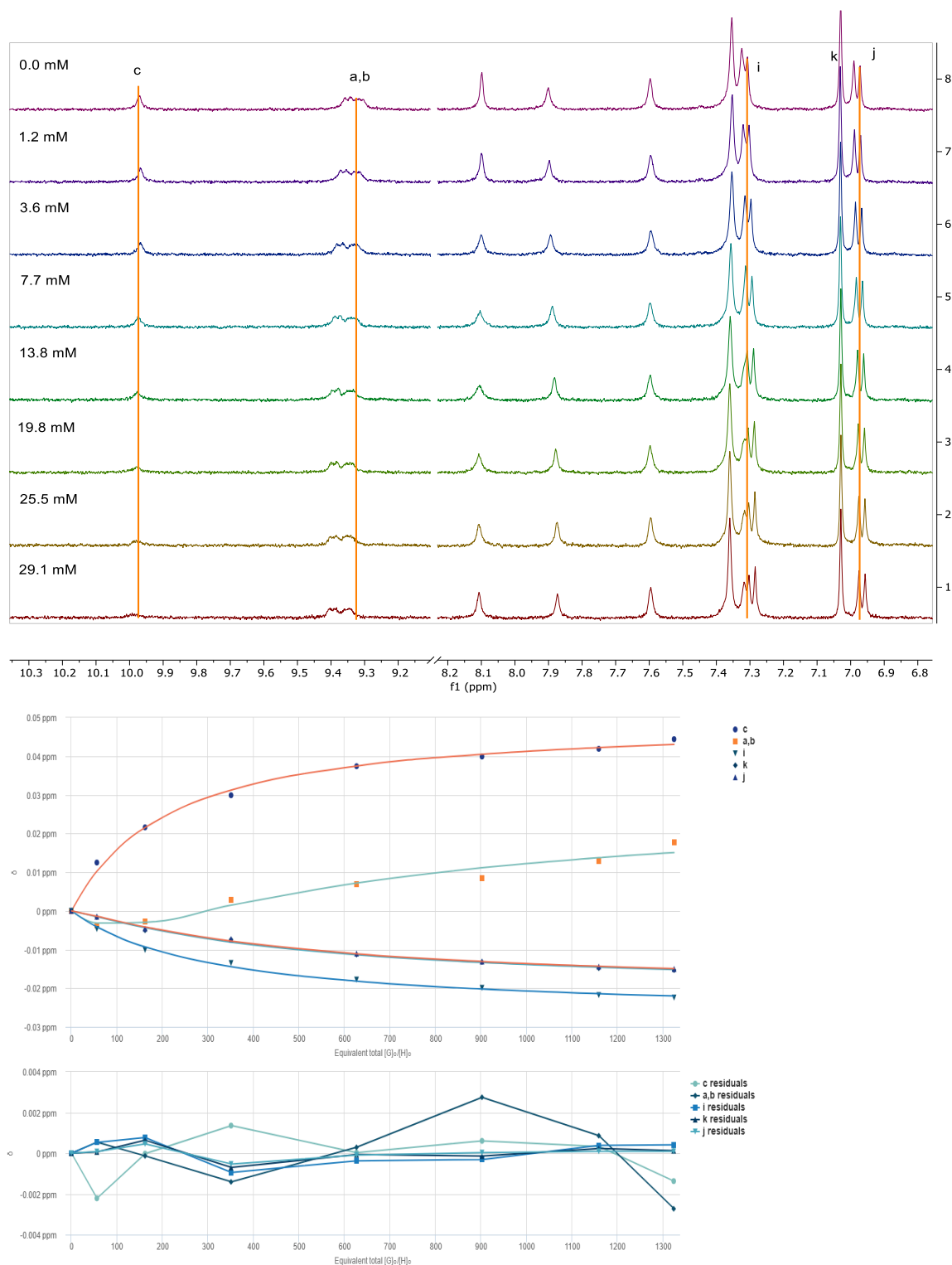
Supplementary Figure 71. ^1H NMR (500 MHz, D_2O) spectra for receptor **5** (10 μM) titrated with a combined solution of sodium *D*-Alanine (85 mM) and receptor **5** (10 μM). The concentration of guest is shown on each spectrum. pH = 7.55 at the end of the titration. No peak movements were observed.

Receptor 5 & *L*-Ala-*L*-Ala (D₂O)



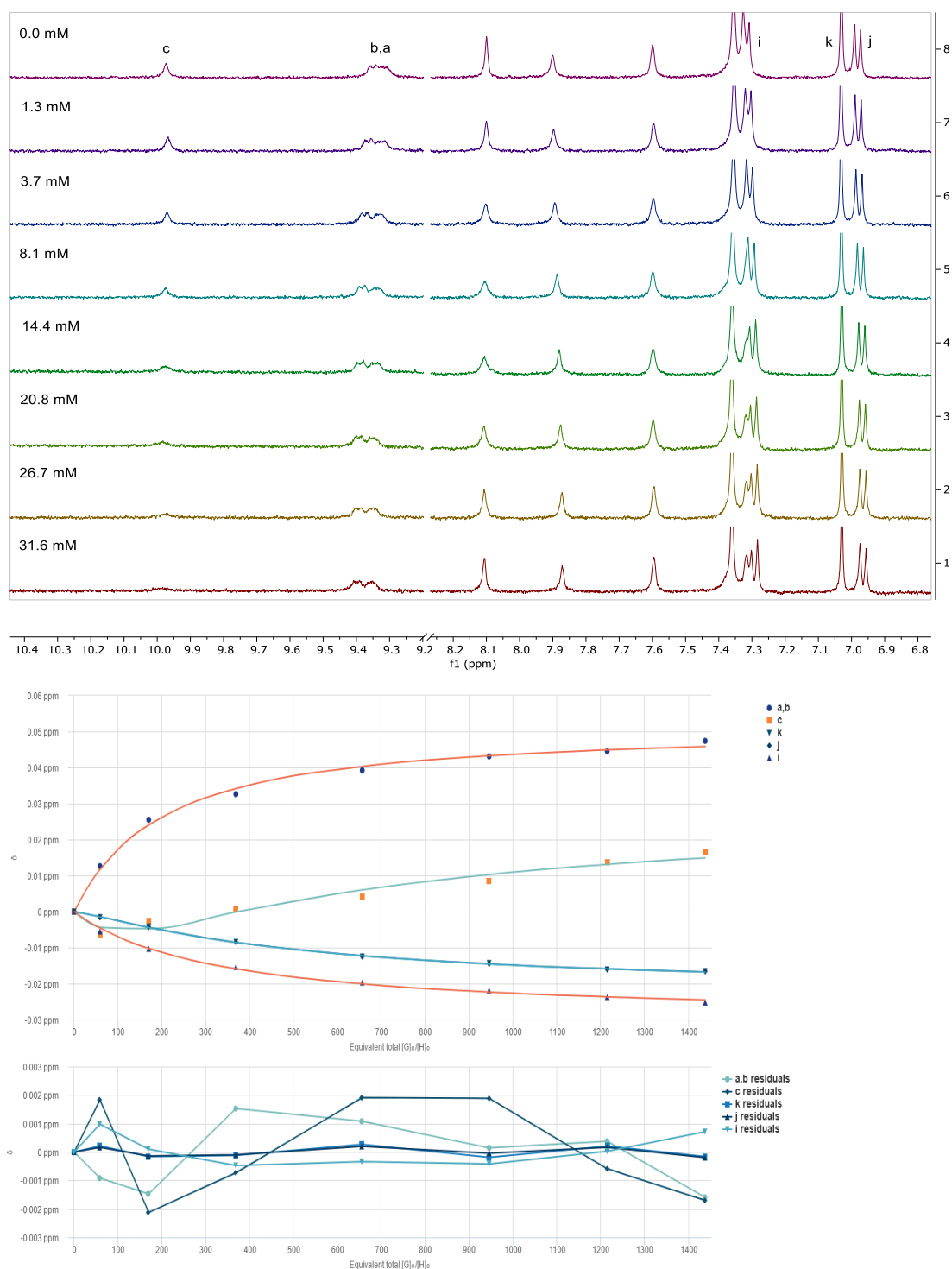
Supplementary Figure 72. ¹H NMR (500 MHz, D₂O) spectra for receptor **5** (10 μM) titrated with a combined solution of sodium *L*-Ala-*L*-Ala (50 mM) and receptor **5** (10 μM). The concentration of guest is shown on each spectrum. pH = 7.41 at the end of the titration. Very small peak movements were observed (< 0.02 ppm), compared to other titration spectra for receptor **5**, it is very likely that receptor **5** does not binding *L*-Ala-*L*-Ala.

Receptor 6 & acetate (H₂O/D₂O)



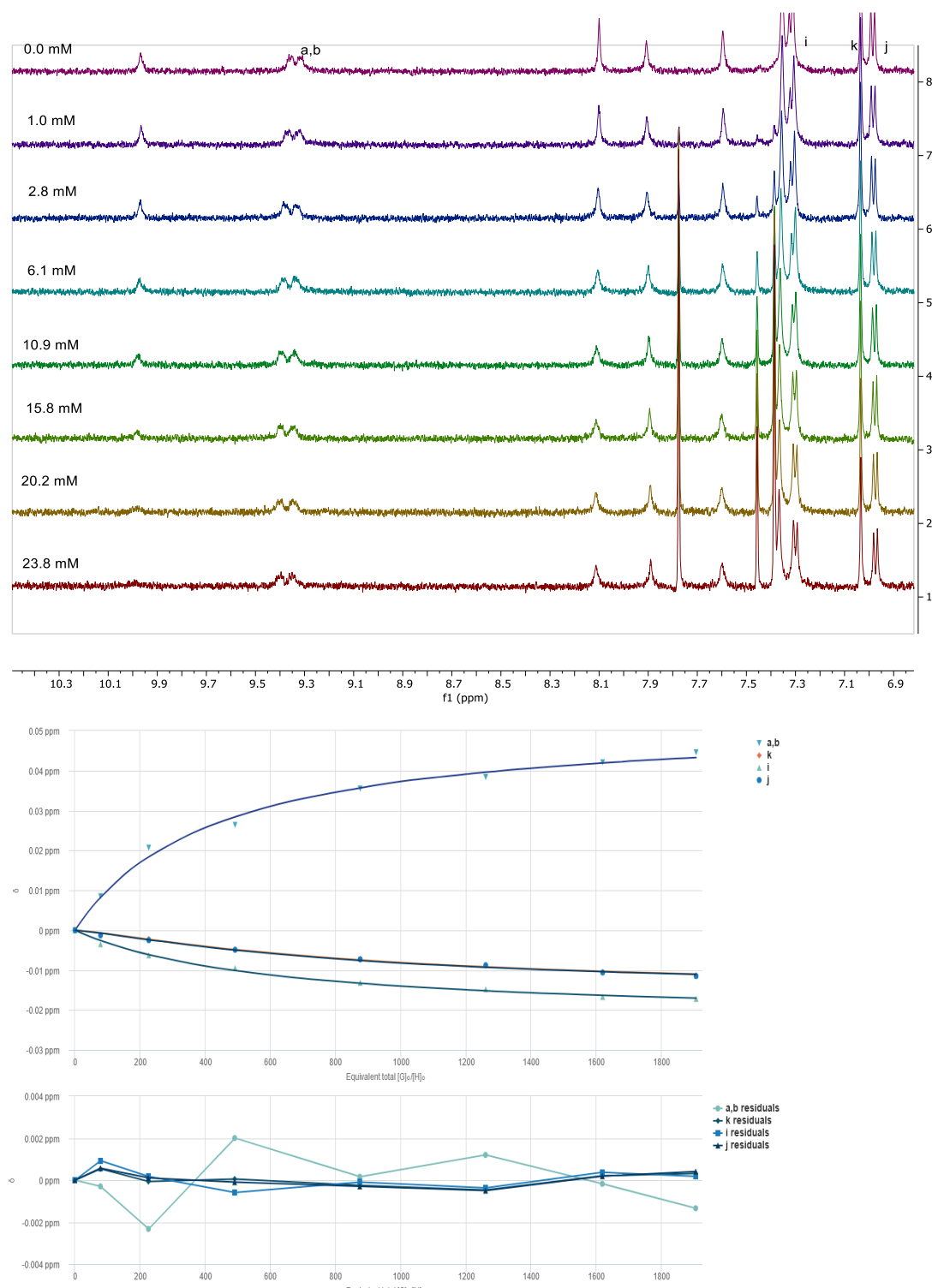
Supplementary Figure 73. Top) ¹H NMR (500 MHz, 9:1 H₂O/D₂O) spectra for receptor **6** (22 μM) titrated with a combined solution of sodium acetate (63 mM) and receptor **6** (22 μM). The concentration of guest is shown on each spectrum. pH = 7.48 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons a, b, c, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 315.2 \text{ M}^{-1} (\pm 5.4 \%)$. Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/9be4aa64-3c2b-4692-8953-9e09f626023e>

Receptor 6 & propionate (H₂O/D₂O)



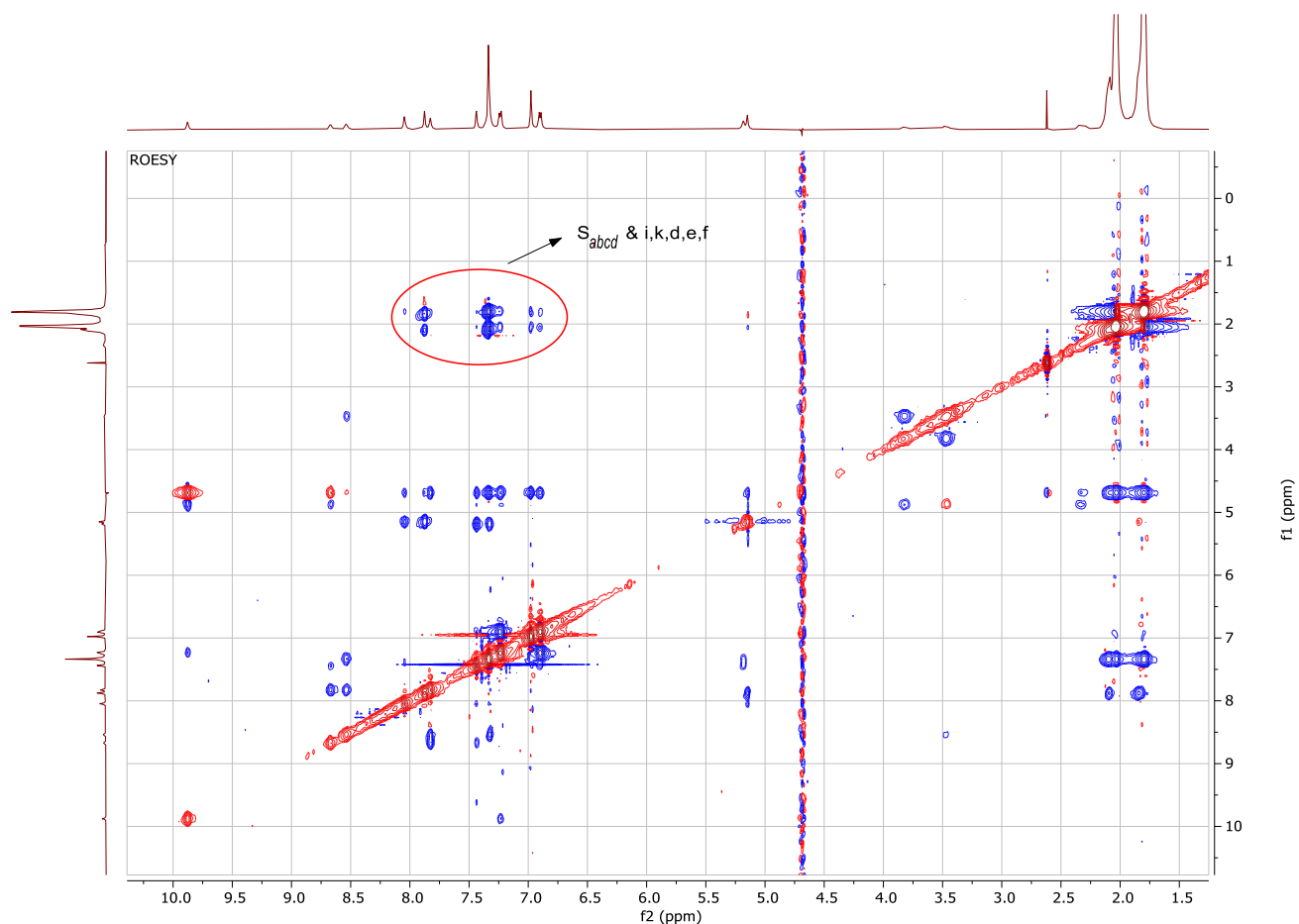
Supplementary Figure 74. Top) ¹H NMR (500 MHz, 9:1 H₂O/D₂O) spectra for receptor **6** (22 μM) titrated with a combined solution of sodium propionate (66 mM) and receptor **6** (22 μM). The concentration of guest is shown on each spectrum. pH = 7.52 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons a, b, c, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 311.9 \text{ M}^{-1}$ ($\pm 5.0 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/64882fad-c6c8-4de9-94ac-b9d16654a05b>

Receptor 6 & benzoate (H₂O/D₂O)

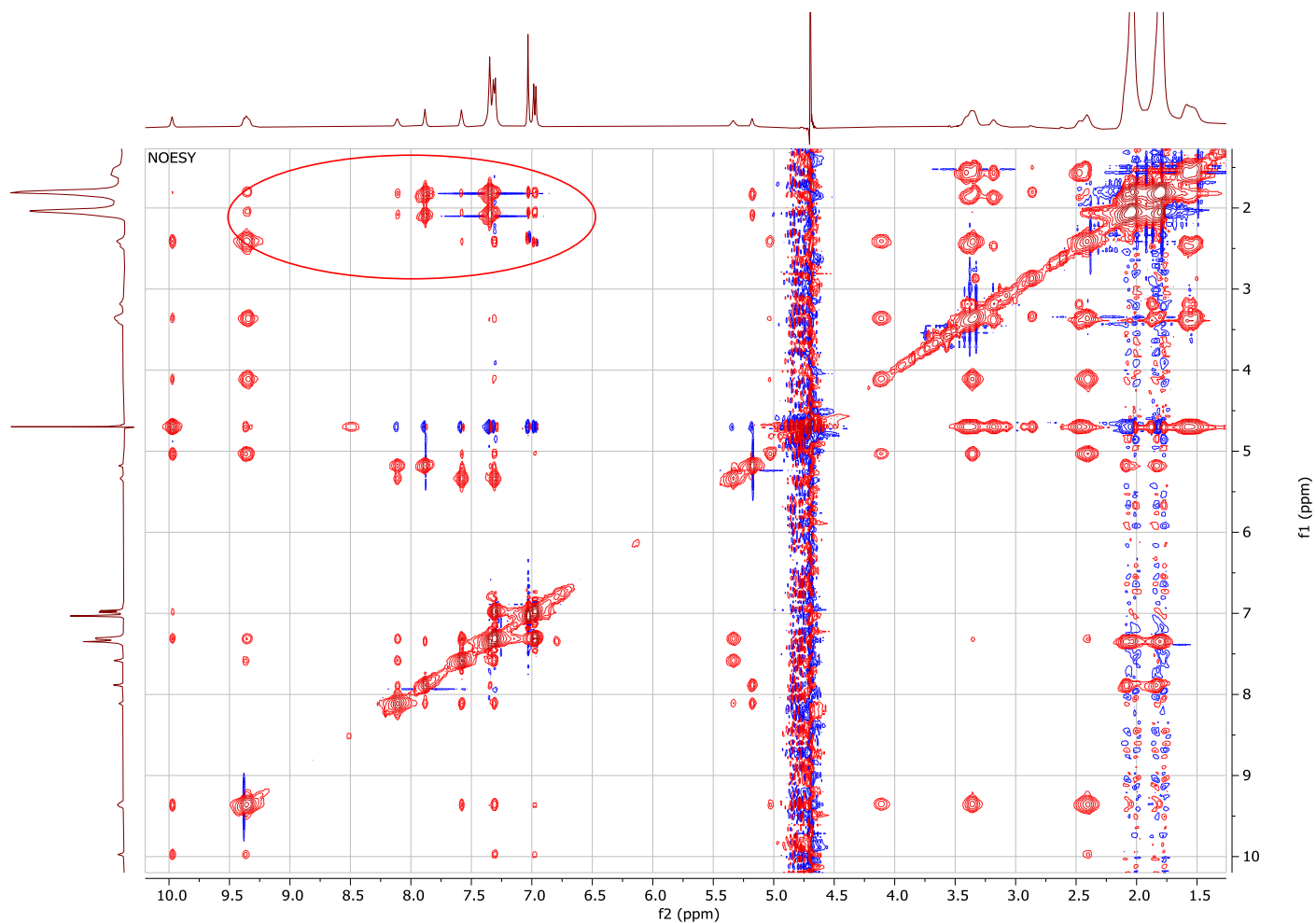


Supplementary Figure 75. Top) ¹H NMR (500 MHz, 9:1 H₂O/D₂O) spectra for receptor **6** (13 μM) titrated with a combined solution of sodium benzoate-*d*⁵ (50 mM) and receptor **6** (13 μM). The concentration of guest is shown on each spectrum. pH = 7.54 at the end of the titration. Bottom) Global fitting of the binding isotherms (protons a, b, i, j and k) from Bindfit to a 1:2 non-cooperative model $K_a = 317.7 \text{ M}^{-1}$ ($\pm 5.4 \%$). Full fitted data is available online at: <http://app.supramolecular.org/bindfit/view/5a94e312-c445-4639-8b6a-ac6773ec29a1>

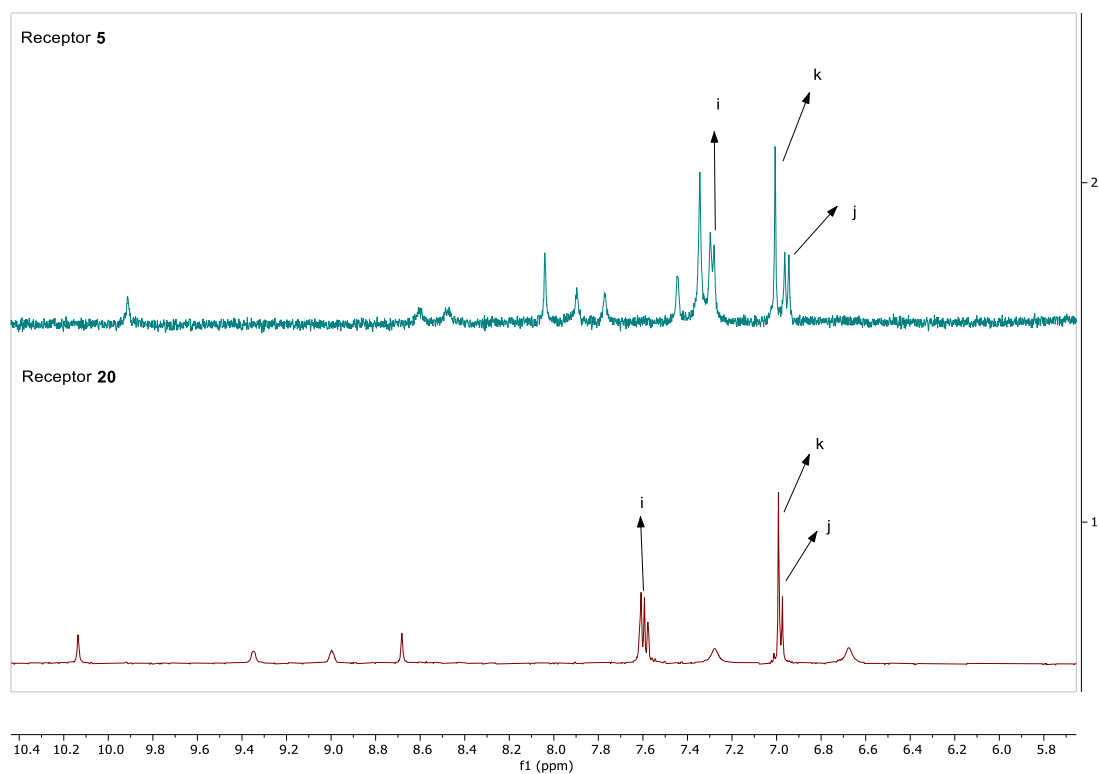
NMR Structural Studies of Hosts and Complexes



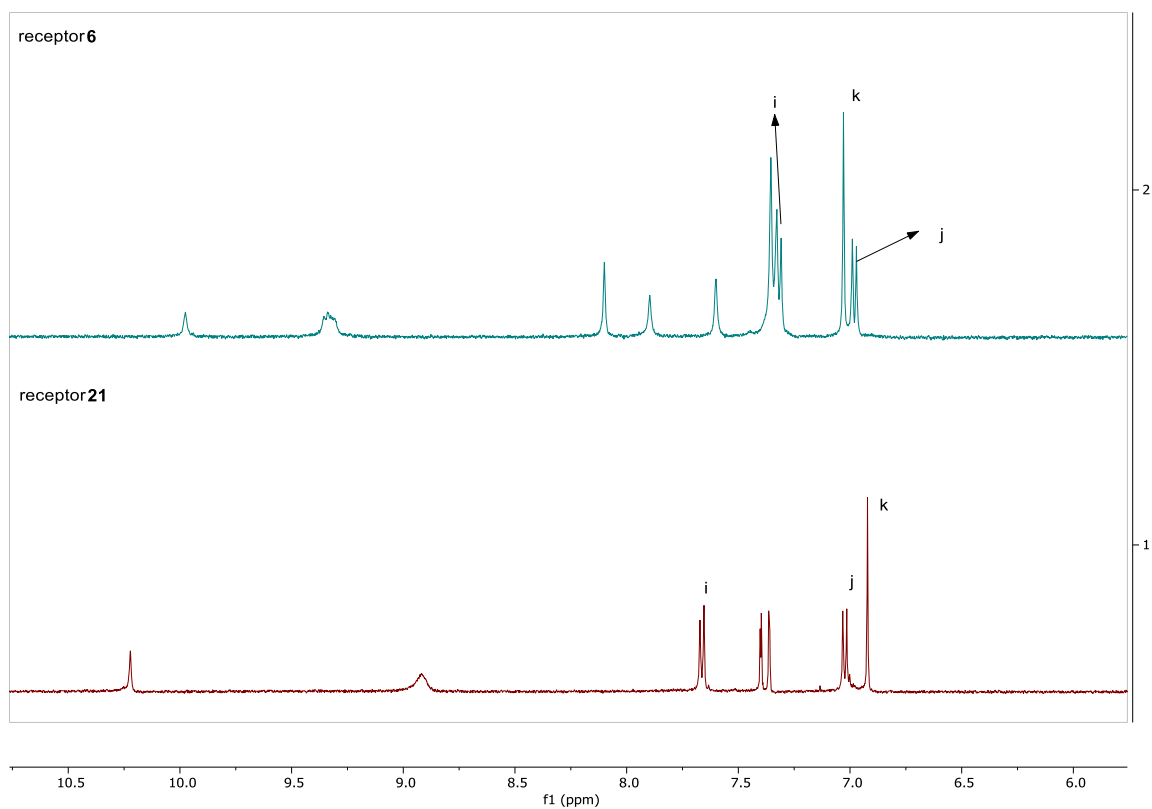
Supplementary Figure 76. 2D ROESY NMR spectrum (600 MHz) of receptor **5** (1.33 mM) in 9:1 $\text{H}_2\text{O}/\text{D}_2\text{O}$. Cross peaks in the red circle represent connections between solubilising groups and the receptor core aromatic protons, probably due to self-association at this relatively high concentration.



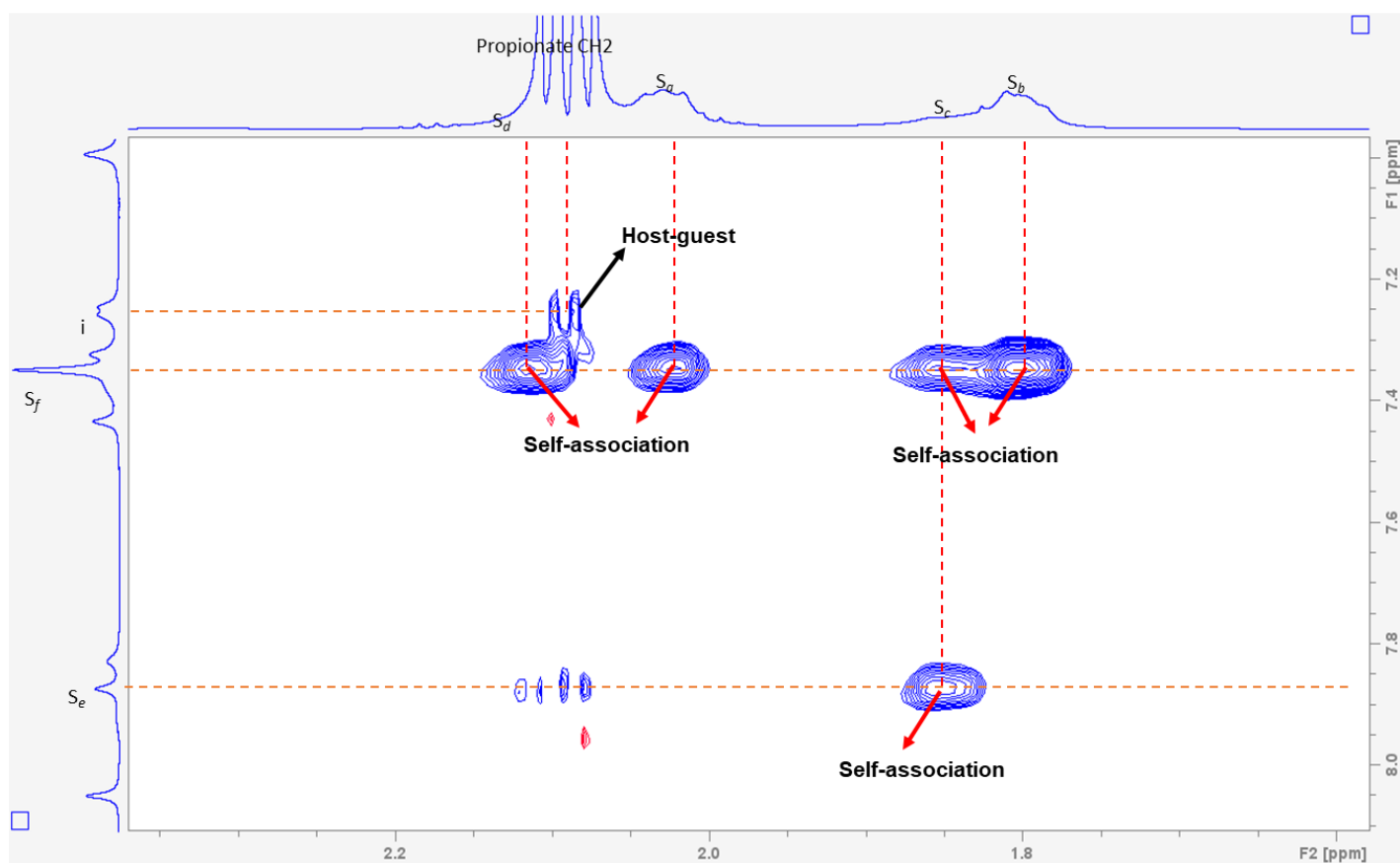
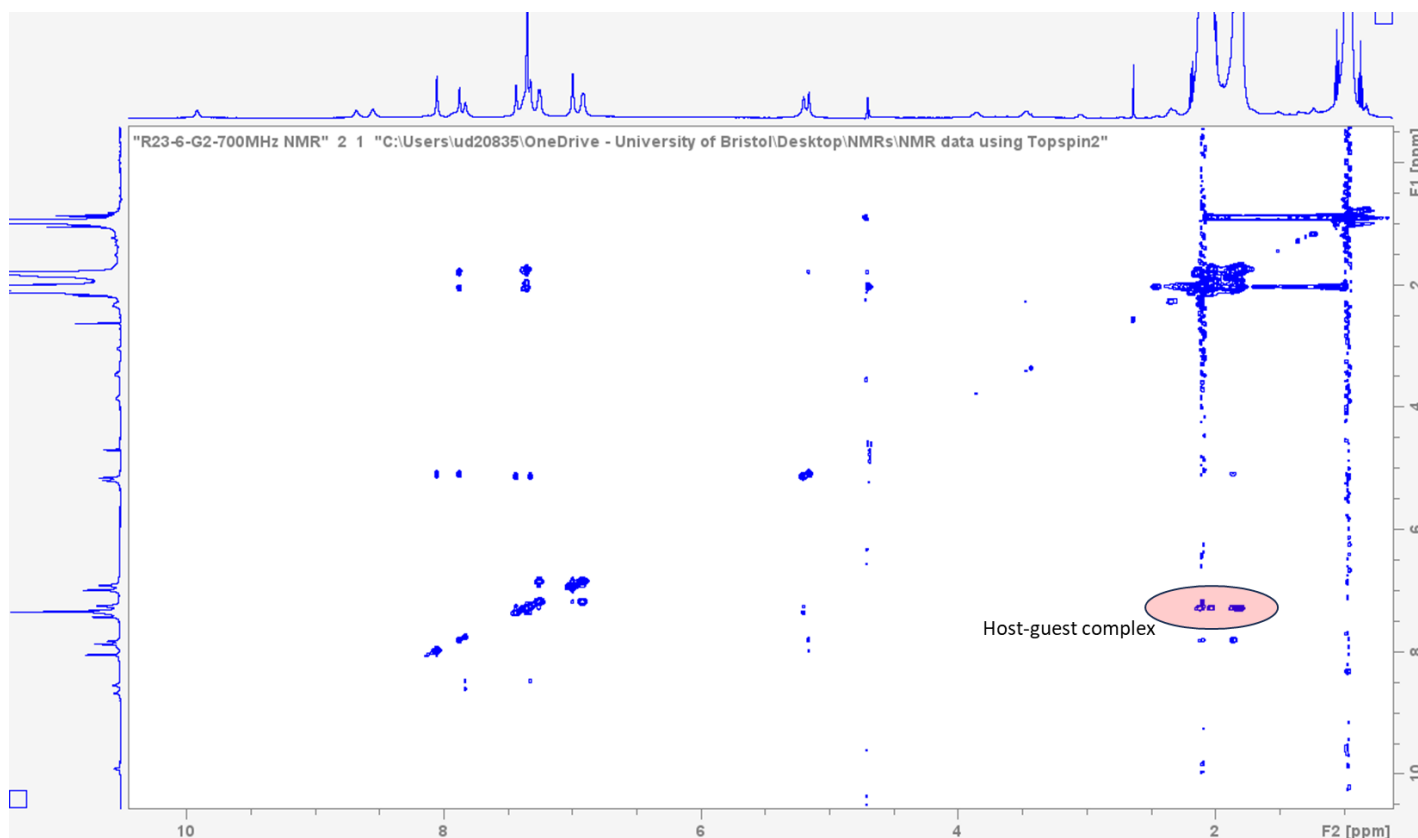
Supplementary Figure 77. 2D NOESY NMR spectrum (600 MHz, 350 ms mixing time) of receptor **6** (1.81 mM) in 9:1 H₂O/D₂O. Cross peaks in the red circle represent connections between solubilising groups and the receptor core aromatic protons, probably due to self-association at this relatively high concentration.

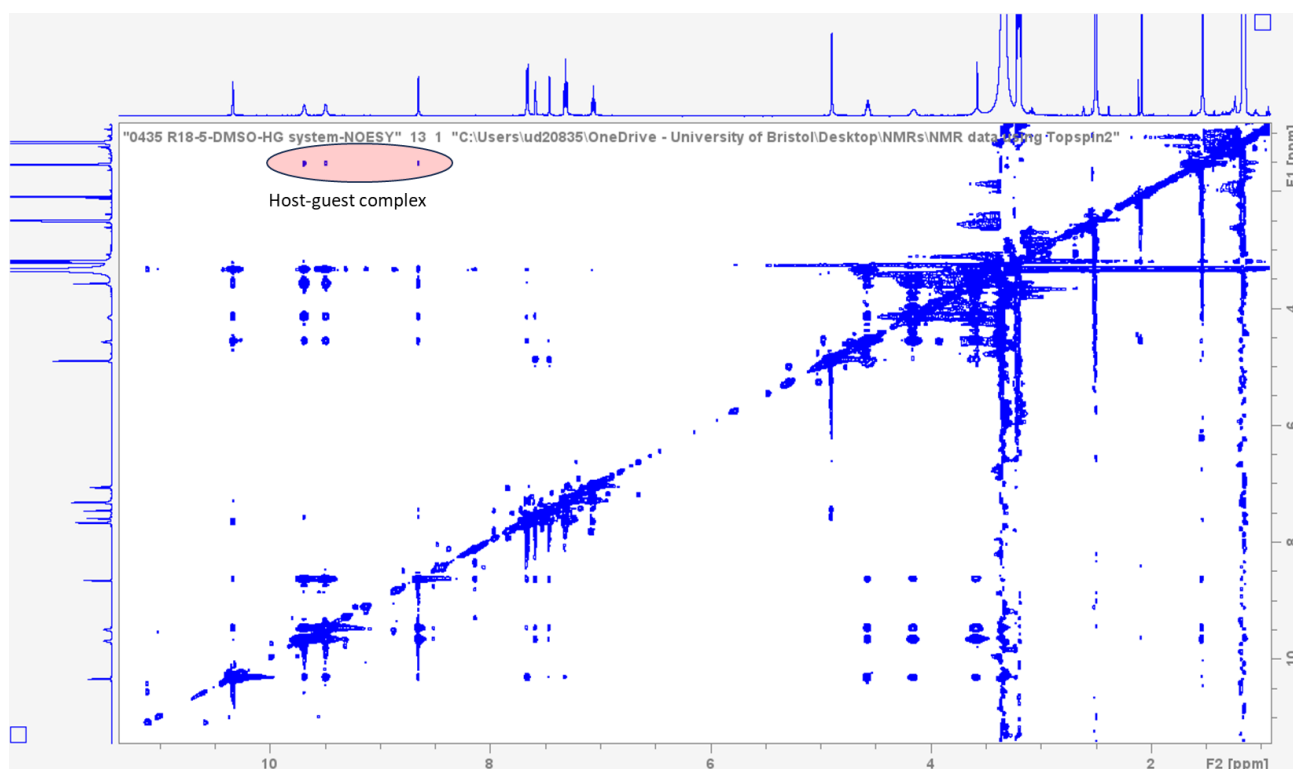
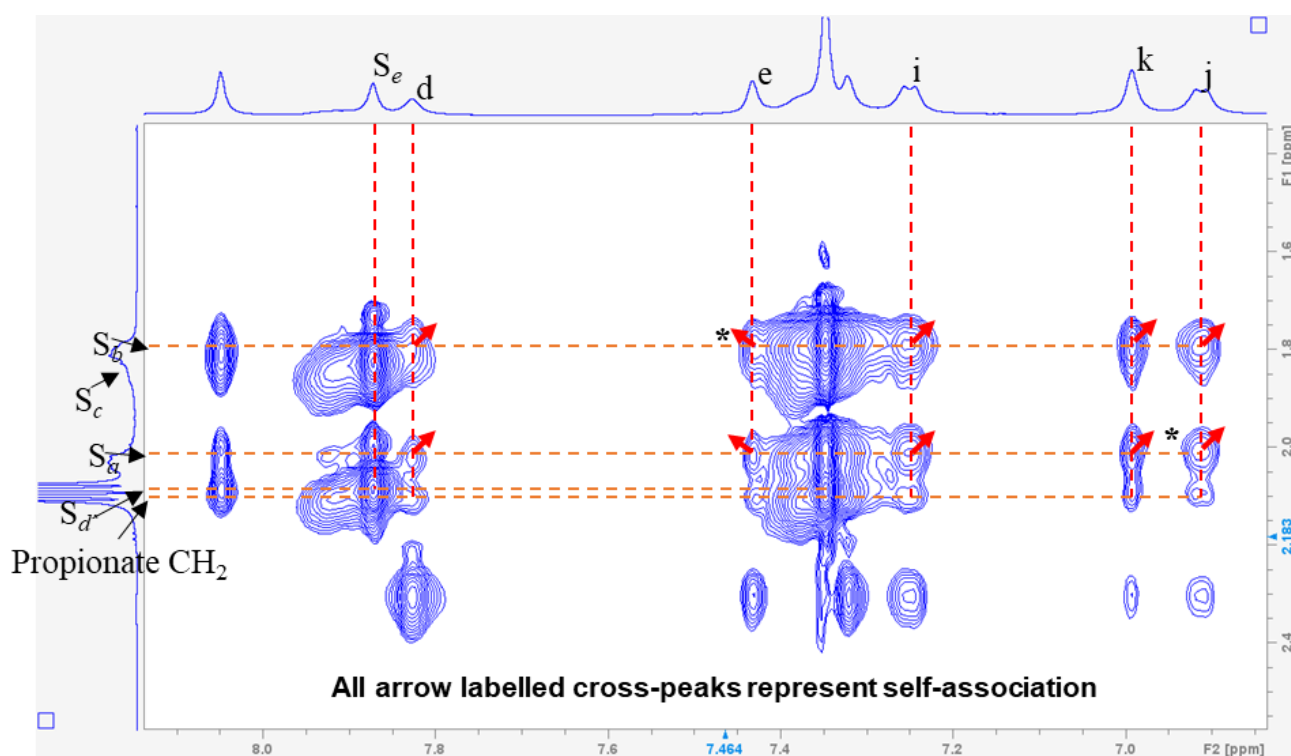


Supplementary Figure 78. Comparison between ¹H NMR spectrum (500 MHz) of receptor **5** (0.022 mM) in 9:1 H₂O/D₂O and ¹H NMR spectrum (600 MHz) of receptor **20** (0.17 mM) in DMSO-*d*₆.



Supplementary Figure 79. Comparison between ¹H NMR spectrum (500 MHz) of receptor **6** (0.056 mM) in 9:1 H₂O/D₂O and ¹H NMR spectrum (500 MHz) of receptor **21** (0.1 mM) in DMSO-*d*₆.

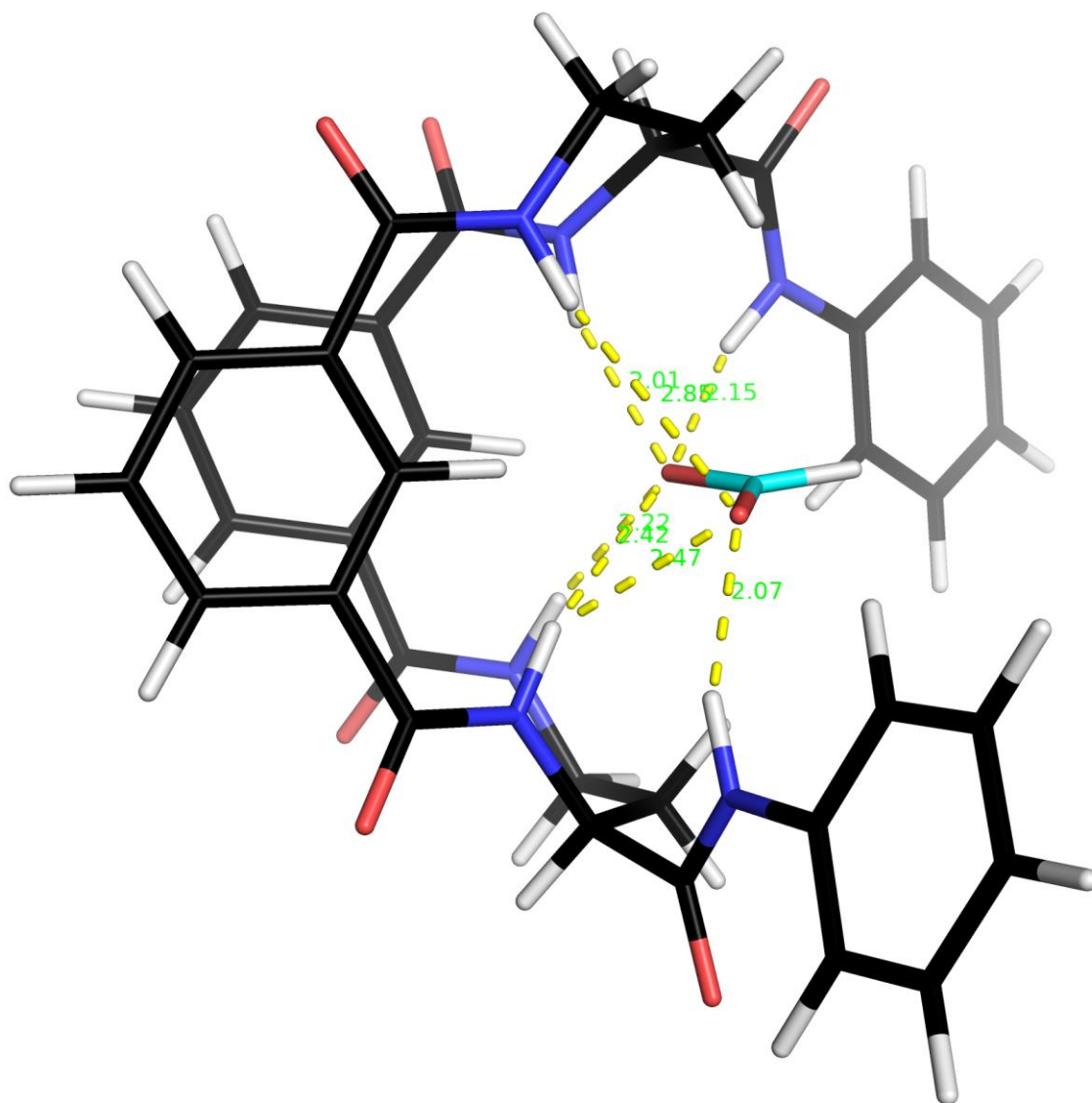




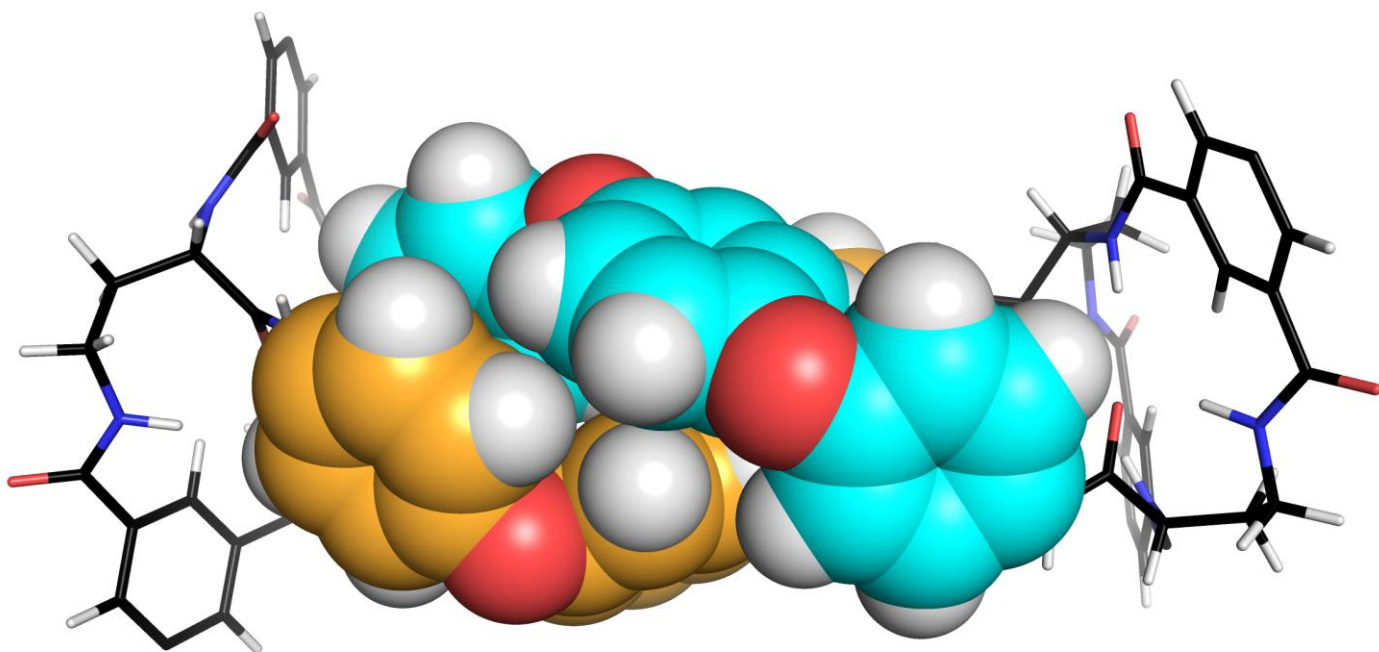
Supplementary Figure 80. NOESY spectra (700 MHz, 9:1 H₂O /D₂O, 600 ms) of receptor **5** (1.33 mM) + sodium propionate (250 mM).

3. Modelling studies

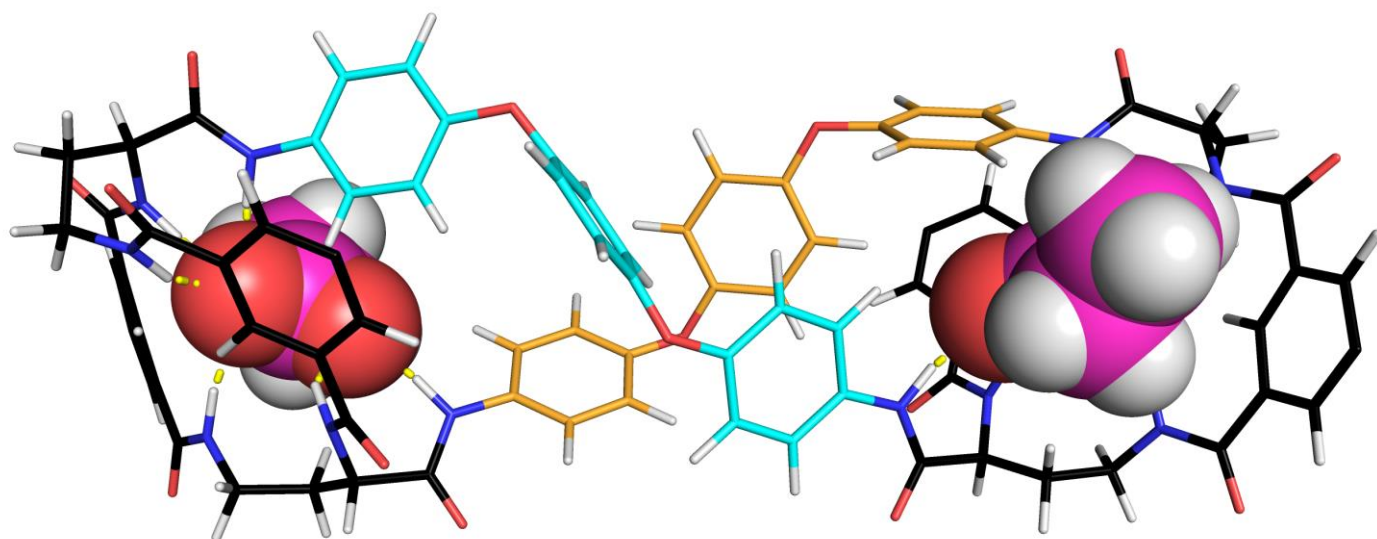
Modelling studies employed Maestro Version 13.8, with Batchmin V14.2 for energy minimisation. The calculations employed the OPLS4 force field, aqueous GB/SA solvation, and 4000 MCMM steps.



Supplementary Figure 81. . Lowest energy structure of macrocycle **9** (R = H) bound to formate anion from an MCMM simulation. Conformations were generated by opening of the macrocycle, bond rotations and ring closure. The ground state structure possesses seven short NH...O⁻ distances ranging from 2.01 – 2.85 Å (yellow broken lines).



Supplementary Figure 82. . Energy-minimised structure of receptor **5**, as also shown in Figure 5a, with space-filling representation for the bis-phenoxyphenyl bridges (cyan/gold carbons), and side chains omitted for clarity.



Supplementary Figure 83. . Energy-minimised structure of receptor **5** binding two propionate anions (magenta carbons) as also shown in Figure 5b. The perspective is chosen to highlight the offset between the bis-phenoxyphenyl bridges in the proposed binding conformation.

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- 1 Destecroix, H. *et al.* Affinity enhancement by dendritic side chains in synthetic carbohydrate receptors. *Angew Chem Int Ed.* **54**, 2057-2061 (2015).
 - 2 Shao, P. *et al.* Synthesis and Evaluation of a Tetra[6,7]quinoxalinoporphyrine-based Near Infrared Photosensitizer. *RSC Adv.* **7**, 50555-50561 (2017).
 - 3 Sugai, N., Heguri, H., Yamamoto, T. & Tezuka, Y. A Programmed Polymer Folding: Click and Clip Construction of Doubly Fused Tricyclic and Triply Fused Tetracyclic Polymer Topologies. *J. Am. Chem. Soc.* **133**, 19694-19697 (2011).
 - 4 Wang, F. *et al.* Metal coordination mediated reversible conversion between linear and cross-linked supramolecular polymers. *Angew Chem Int Ed.* **49**, 1090-1094 (2010).
 - 5 <http://supramolecular.org>