nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🕱 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
×	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
×	A description of all covariates tested
×	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
x	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
x	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
X	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
×	Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated
	Our web collection on statistics for biologists contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection EPU 3.5.1, TEM User interface Titan 3.15.1, Digital Micrograph 3.22.1461.0

Data analysis

MotionCor2 1.4.5, cryoSPARC 4.4.1, Phenix 1.20.1-4487, ISOLDE 1.6.0, AlphaFold2, Coot 0.9.8.1, UCSF ChimeraX 1.6.1, Prism 6.07, GIMP 2.10.28, ImageJ 1.8.0_345, Tycho NT.6 software 1.3.2.880, AcquireMP (Refeyn) 2.3

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g., GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

The coordinates of the SLFN11 S753D structure have been deposited in the Protein Data Bank (PDB) under the accession code 9ERD and the cryo-EM reconstruction is available at the Electron Microscopy Data Bank (EMDB) under the EMBD accession code EMD-19912. The coordinates of the SLFN11 wt bound to tRNA-Leu and tRNA-Met structures have been deposited in the PDB under the accession codes 9ERE and 9ERF, respectively. The cryo-EM reconstructions are available at the EMDB under the EMBD accession codes EMD-19913 and EMD-19914, respectively.

Field-specific reporting					
Please select the or	ne below that is	the best fit for your research. If you are not sure, read the appropriate sections before making your selection.			
X Life sciences	□ Ве	chavioural & social sciences Ecological, evolutionary & environmental sciences			
For a reference copy of t	he document with a	Il sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>			
Life scier	ices stu	ıdy design			
All studies must dis	close on these (points even when the disclosure is negative.			
Sample size	No statistical methods were used to predetermine sample size. Sample sizes were chosen based on previous experience and the anticipated variance to obtain statistical significance and reproducibility. Size of cryo-EM dataset sample size was based on sufficient number of images and particles to obtain a high resolution reconstruction.				
Data exclusions	No data were excluded.				
Replication	Experiments that led to quantitative conclusions were performed in independent replicates as described in the figure legends.				
Randomization	No statistical calculation was involved that require randomization. For cryo-EM analyses, particles were randomly assigned to half-maps for resolution determination following the standard procedures in cryoSPARC.				
Blinding	No blinding was perfomed in order to avoid errors in sample naming.				
We require information	on from authors a	pecific materials, systems and methods bout some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material,			
•		your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response. Wethods			
✓ Antibodies ✓ ChIP-seq					
□ x Eukaryotic cell lines x □ Flow cytometry					
Palaeontology and archaeology MRI-based neuroimaging					
Animals and other organisms					
Human research participants					
Clinical data Dual use research of concern					
∡	search of concern				
Eukaryotic ce	ell lines				
Policy information about <u>cell lines</u>					
		Spodoptera frugiperda Sf21 insect cells (Thermo Fisher, 11497013) Trichoplusia ni High Five insect cells (Invitrogen, B85502)			

No methods were used for authentication.

Protein expression cell lines were not tested.

No commonly misidentified cell lines were used.

Authentication

(See <u>ICLAC</u> register)

Mycoplasma contamination

Commonly misidentified lines