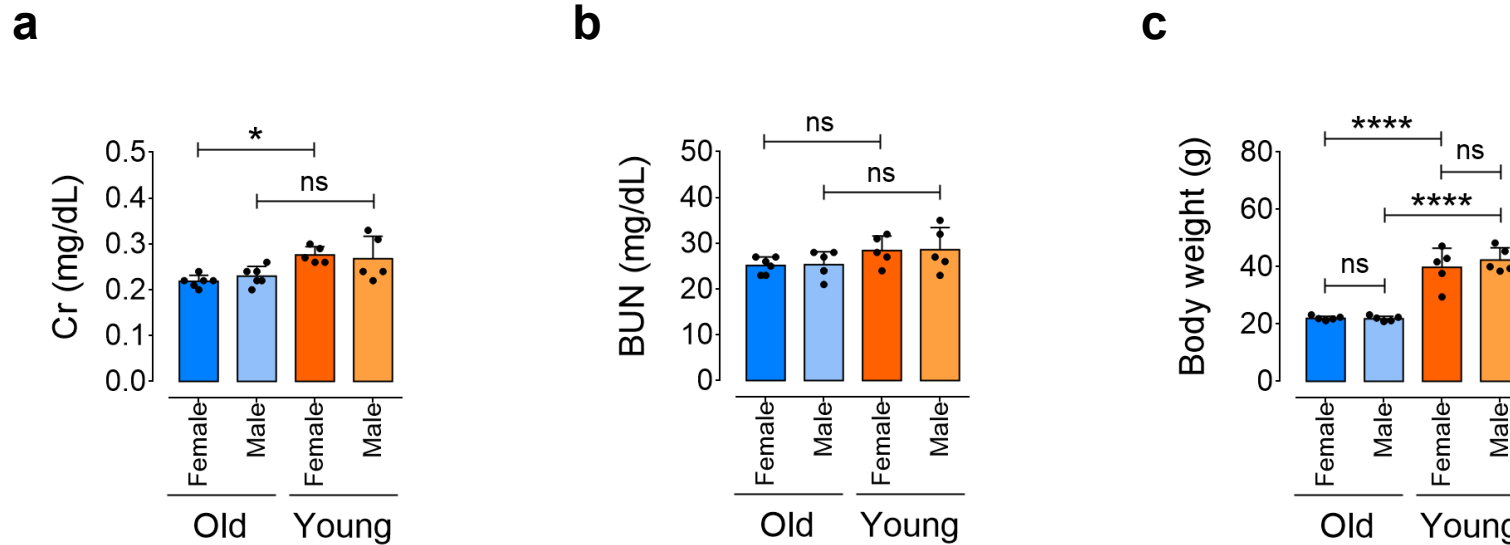
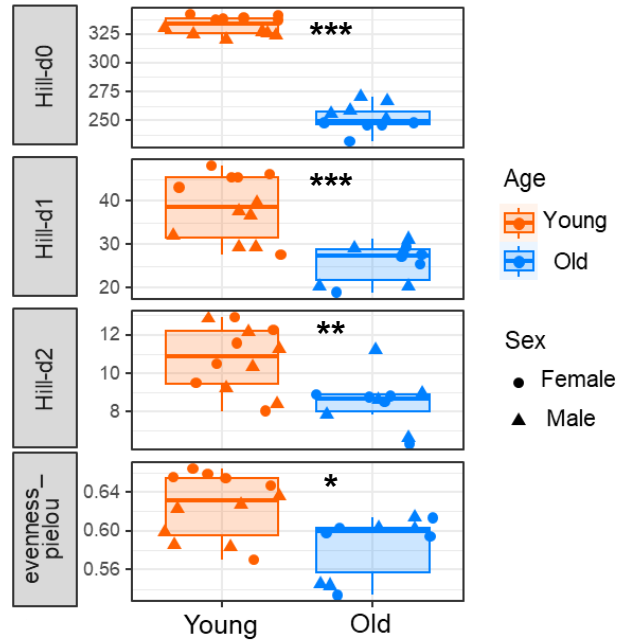


## Supplementary Information

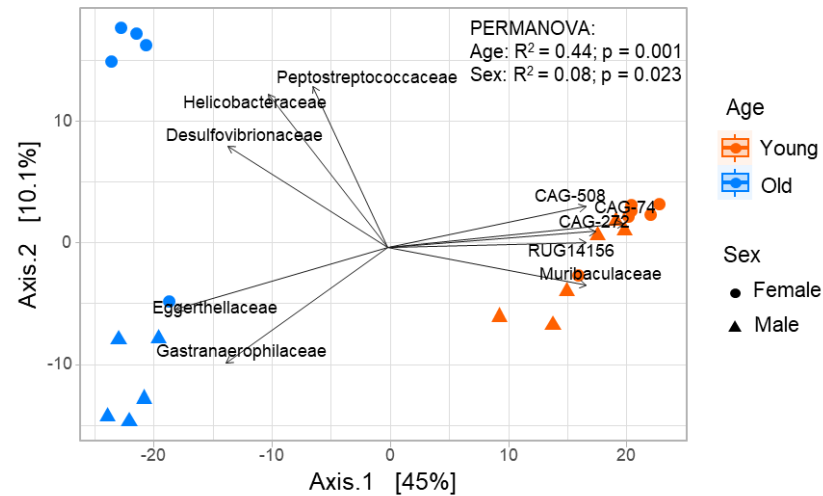


**Fig. 1. Kidney function profile in aging.** **a**, Analysis of body weight in female and male, old and young mice (n=5-6). **b,c**, Bar charts showing kidney function profiles, represented as concentrations of creatinine (Cr) (**b**) and BUN (**c**) in sera of female and male, old and young mice (n=5-6). Error bars represent SD (**a-c**). *P* values were calculated using two-way ANOVA followed by Tukey's post *hoc* test (**a-c**). (\**P*<0.05, \*\*\*\**P*<0.0001, ns, not significant). Source data are provided as a Source Data file.

a



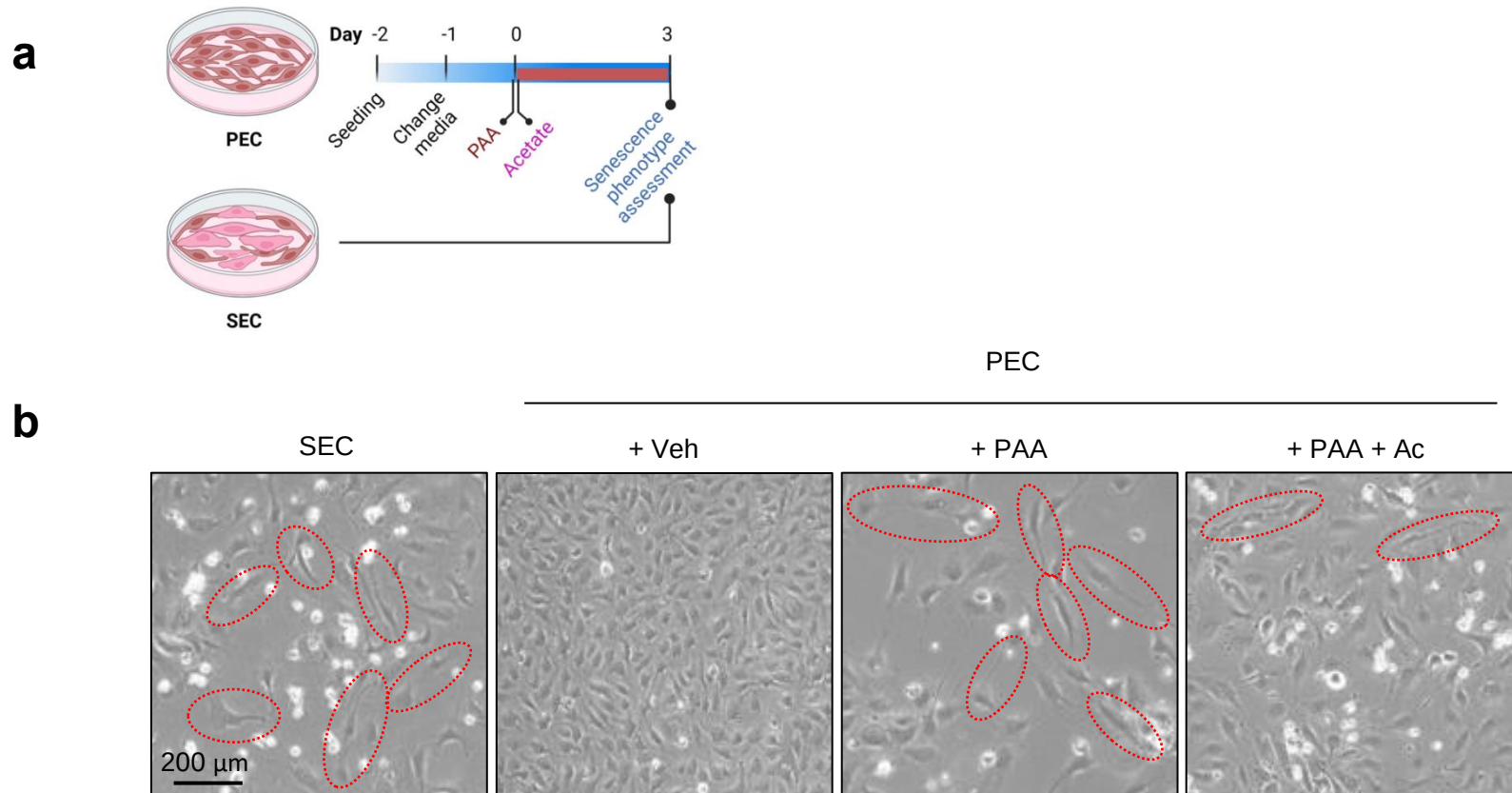
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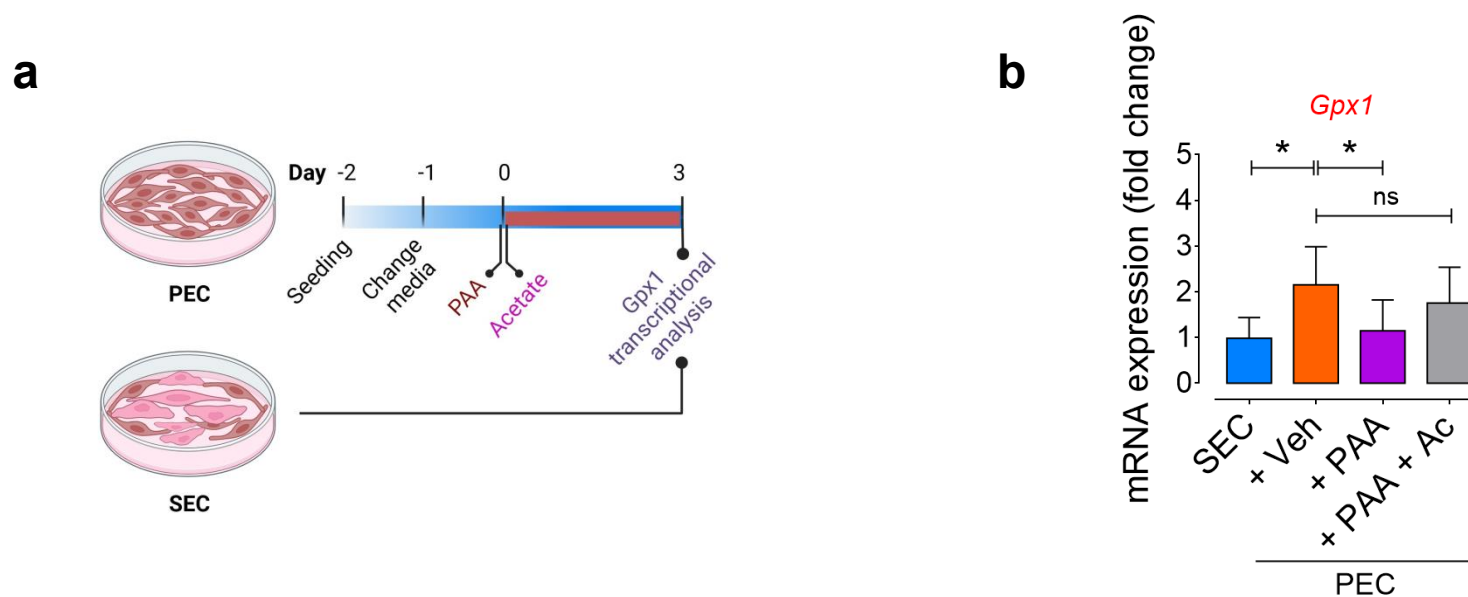
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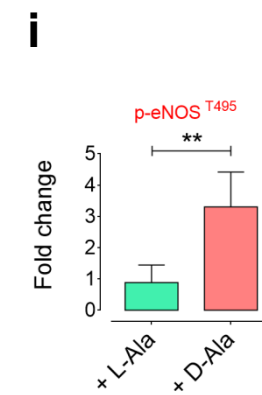
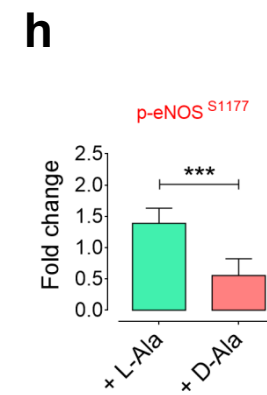
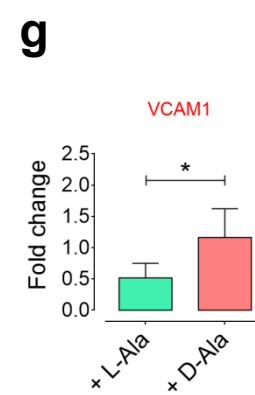
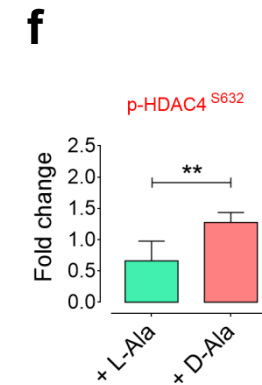
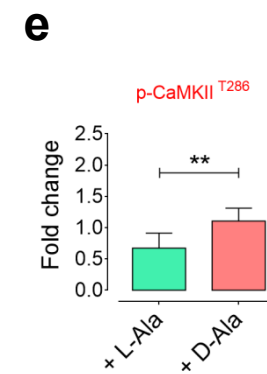
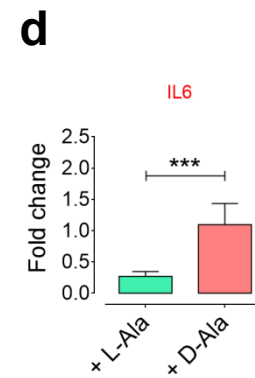
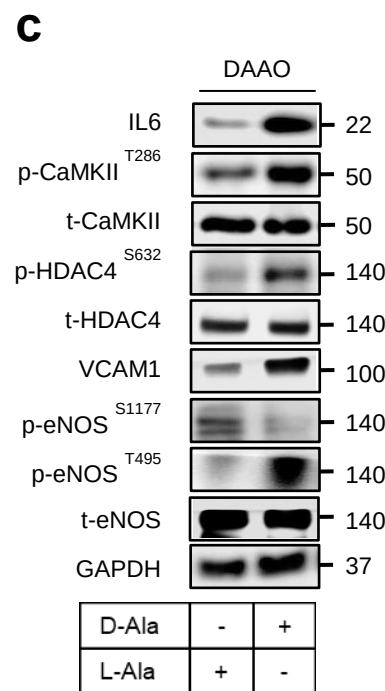
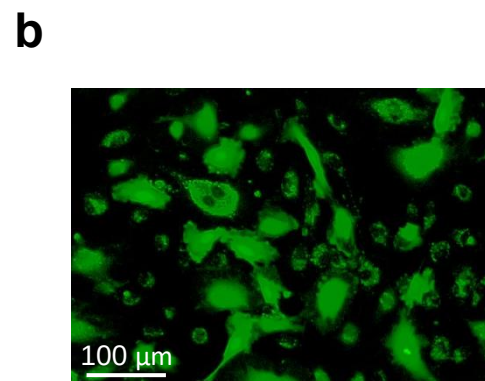
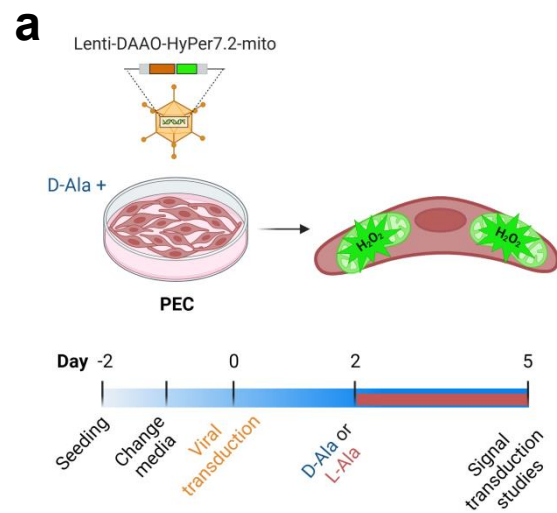
**Fig. 2. Age-associated gut microbiota alterations.** **a**, Box plots representing alpha diversity in feces of both female and male old and young mice measured by Hill-d0 to 2 and Pielou's evenness indices (n=5-6). **b**, Principal coordinate analysis (PCoA) plot showing beta diversity based on the community level changes in old and young mice (n=5-6). **c**, Heatmap depicting differentially abundant microbial profiles at the strain level (each color represents one bacterial taxa) in the fecal microbiota of female and male, old *versus* young mice (n=5-6). Error bars represent SEM (**a,b**). Data were analyzed using Kruskal-Wallis statistical test (**a**) and Atchinson distance and permutational multivariate analysis of variance (PERMANOVA) tests (**b**). (\* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ ). Source data are provided as a Source Data file.



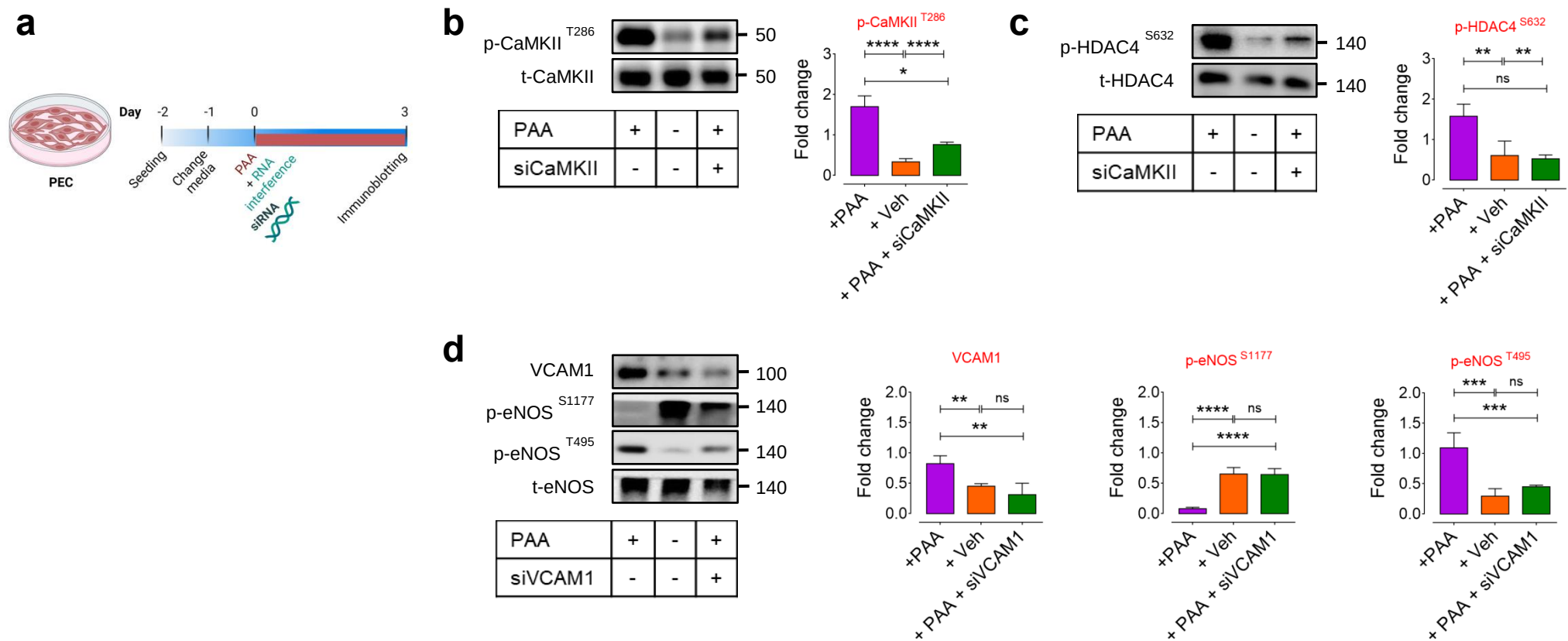
**Fig. 3. PAA induces senescence phenotype in endothelial cells.** **a**, Schematic diagram of the experimental setting: SECs, PECs, PAA-treated PECs (10  $\mu$ M, for 72 h), and PAA+Acetate-treated PECs (PAA: 10  $\mu$ M, sodium acetate: 3  $\mu$ M, for 72 h) were assessed for senescence phenotype. **b**, Representative bright-field images depicting cellular senescence-like phenotype, including enlarged, flattened, and multinucleated appearance, in PECs treated with PAA at the magnitude seen in replicative SECs. The images demonstrate that sodium acetate reduces the number of cells with morphologically senescence-like phenotype in PAA-exposed PECs (n=6). Scale bar, 200  $\mu$ m. Red circles indicate senescent-like cells. Experiments were triplicated independently.



**Fig. 4. PAA induces oxidative stress by downregulating antioxidant defense in PECs.** **a**, Schematic diagram of the experimental setting: SECs, PECs, PAA-treated PECs (10  $\mu$ M, for 72 h), and PAA+Acetate-treated PECs (PAA: 10  $\mu$ M, sodium acetate: 3  $\mu$ M, for 72 h) were subjected to glutathione peroxidase 1 (*Gpx1*) transcriptional analysis. **b**, qPCR analysis demonstrating the downregulation of *Gpx1* in PECs in response to PAA and restoration of the gene expression following the addition of sodium acetate (n=6). Error bars represent SD (**a**). Data represent triplicated biologically independent experiments. *P* values were calculated using one-way ANOVA followed by Tukey's post *hoc* test (**a-c**). (\**P*<0.05, ns, not significant). Source data are provided as a Source Data file.

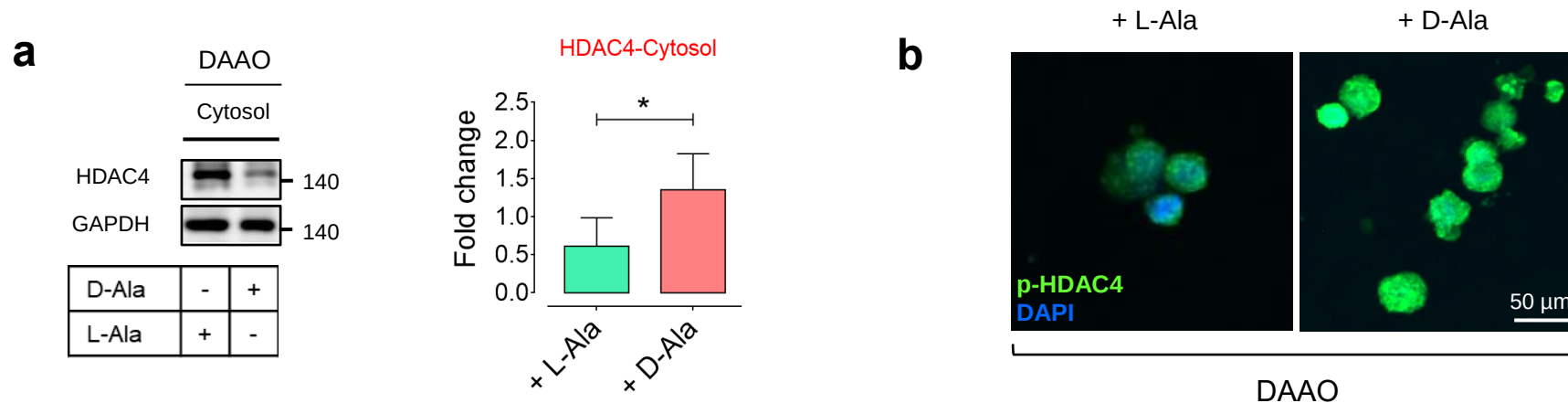


**Fig. 5. Chemogenetic mitochondrial H<sub>2</sub>O<sub>2</sub> regulates endothelial NO signaling through IL6-HDAC4 pathway.** **a**, Schematic diagram of the experimental setting: PECs were transduced with lentiviral vectors encoding D-aminoacid oxidase (DAAO)-HyPer7.2 targeted to the cell mitochondria for generation of mitochondrial H<sub>2</sub>O<sub>2</sub> in the presence D-alanine (10 mM) for 72 h. **b**, Representative widefield ratiometric HyPer7.2 images of PECs transduced with Lenti-DAAO-HyPer7.2-mito constructs. **c-i**, Immunoblots (**c**) and quantitative plots show that chemogenetic mitochondrial H<sub>2</sub>O<sub>2</sub>, produced by DAAO in response to D-alanine but not L-alanine, orchestrates IL6-mediated CaMKII-HDAC4 phosphorylation and the subsequent VCAM1-regulated eNOS phosphorylation at Ser1177 and Thr495 in PECs (n=6). Scale bar, 100  $\mu$ m. Error bars represent SD (**d-i**). Data represent triplicated biologically independent experiments. *P* values were calculated using a two-tailed unpaired Student's *t*-test (**d-i**). (\**P*<0.05, \*\**P*<0.01, \*\*\**P*<0.001, ns, not significant). Source data are provided as a Source Data file.

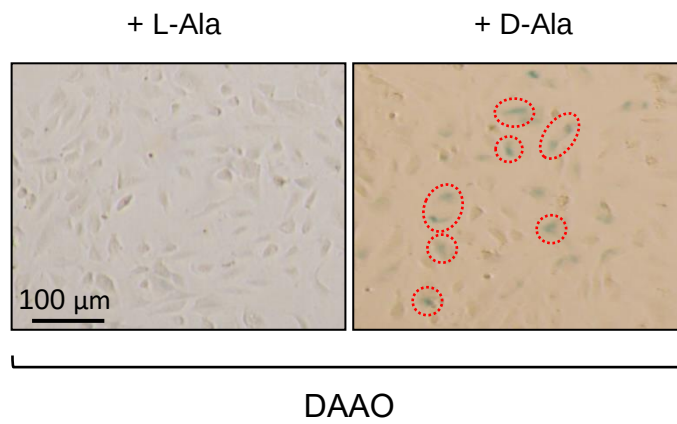
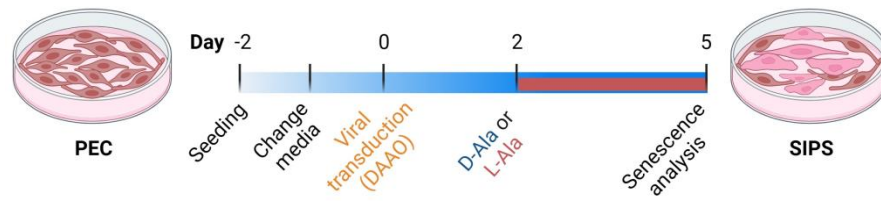
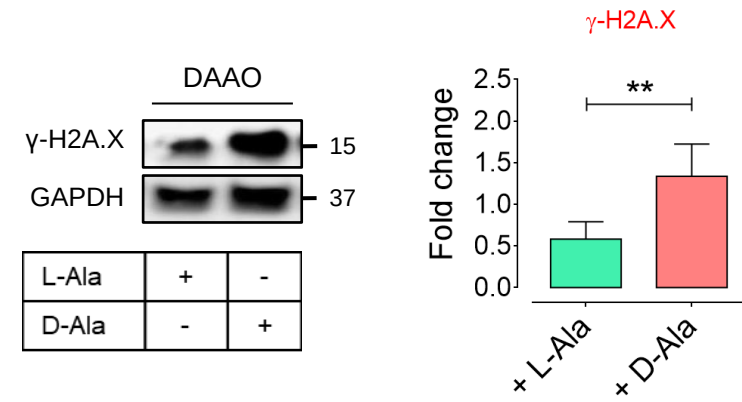
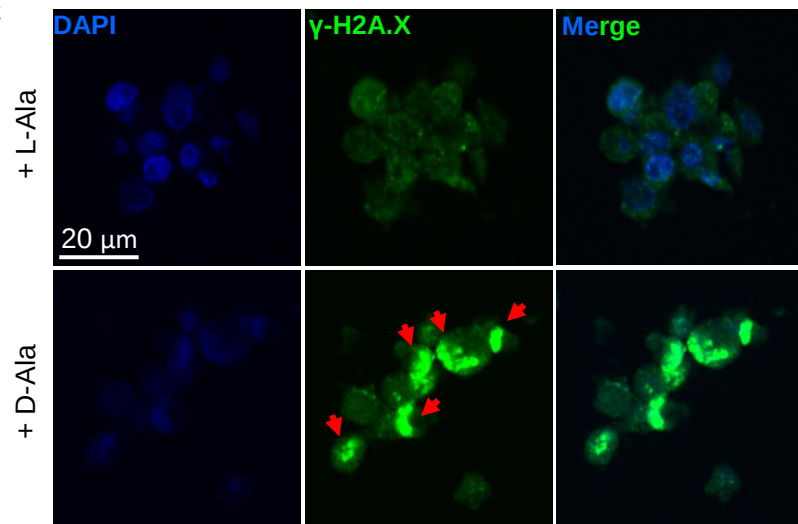


**Fig. 6. CaMKII-mediated SASP pathway underlies PAA-induced EC dysfunction.** **a**, Schematic diagram of the experimental setting: PECs were transfected with siCaMKII or siVCAM1 in the presence or absence of PAA (10  $\mu$ M). **b,c**, Immunoblots (left) and quantitative plots (right) characterizing the effects of PAA on the PAA-induced post-translational modifications of CaMKII (**b**) and its downstream epigenetic regulator HDAC4 (**c**) using CaMKII knockdown approach in PECs (n=6). **d**, Analysis of VCAM1 expression and eNOS phosphorylation at Ser1177 and Thr495 in siVCAM1-transfected PECs in the presence or absence of PAA (n=6). Error bars represent SD (**b-d**). Data represent triplicated biologically independent experiments. *P* values were calculated using one-way ANOVA followed by Tukey's post *hoc* test (**b-d**). (\**P*<0.05, \*\**P*<0.01, \*\*\**P*<0.001, \*\*\*\**P*<0.0001). Source data are provided as a Source Data file.

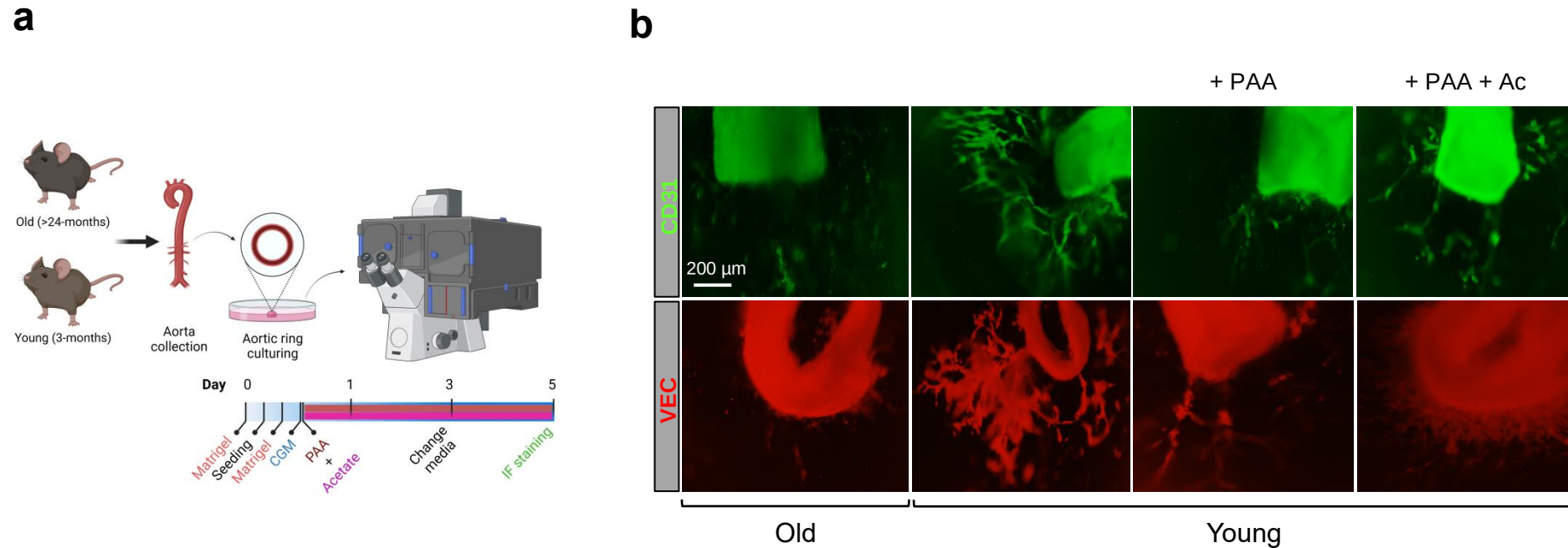




**Fig. 7. H<sub>2</sub>O<sub>2</sub> orchestrates HDAC4 phosphorylation and its nuclear export.** **a**, Immunoblots (left) and quantitative plot (right) reveal HDAC4 nuclear export, represented as an increased expression in the cytosol in response to intracellular H<sub>2</sub>O<sub>2</sub> generated by DAAO in the presence of D-alanine (10 mM, for 72 h) in PECs (n=6). **b**, p-HDAC4 immunostaining shows that intracellular H<sub>2</sub>O<sub>2</sub> markedly increases HDAC4 phosphorylation that facilitates its translocation towards the cell cytosol in PECs (n=6). Scale bar, 50  $\mu$ m. Error bars represent SD (**a**). Data represent triplicated biologically independent experiments. *P* values were calculated using a two-tailed unpaired Student's *t*-test (**a**). (\**P*<0.05, \*\**P*<0.01, \*\*\**P*<0.001, ns, not significant). Source data are provided as a Source Data file.

**a****b****c**

**Fig. 8. Chemogenetic H<sub>2</sub>O<sub>2</sub> promotes EC senescence.** **a**, Schematic diagram of the experimental setting: PECs were transduced with lentiviral vectors encoding D-aminoacid oxidase (DAAO) targeted to the cell mitochondria for generation of mitochondrial H<sub>2</sub>O<sub>2</sub> in the presence D-alanine (10 mM) for 72 h. Representative images of SA-β-gal staining demonstrates that DAAO-mediated mitochondrial H<sub>2</sub>O<sub>2</sub> induced cellular senescence in PECs, as shown by increased SA-β-gal-positive cells (%) (n=6). **b,c**, Immunoblots (**b**) and γ-H2A.X immunostaining images (**c**) reveal a marked increase in DDR following mitochondrial H<sub>2</sub>O<sub>2</sub> production in DAAO-transduced PECs incubated with D-alanine (n=6). Scale bar, 20 and 100 μm. Red circles indicate SA-β-gal-positive cells. Data represent triplicated biologically independent experiments. Source data are provided as a Source Data file.



**Fig. 9. PAA reduces aortic endothelial sprouting.** **a**, Schematic diagram of the experimental setting: aortic rings from old and young mice were subjected to PAA (10  $\mu$ M) for 72 h and tested for endothelial sprouting, followed by CD31 (green) and VEC (red) immunofluorescence staining. **b**, CD31 and VEC immunofluorescence staining in mouse aortas showing decreased endothelial sprouting after incubation of young aortic rings with PAA as opposed to vehicle. Immunofluorescence images reveal that co-treatment with sodium acetate markedly restores angiogenic capacity of aortic CD31, VEC-positive ECs (n=6). Scale bar, 200  $\mu$ m. Experiments were triplicated independently.

**Supplementary Table 1.** Gene-specific forward and reverse primers for RT-PCR amplification. *Gapdh*, Glyceraldehyde 3-phosphate dehydrogenase; *Gpx1*, Glutathione peroxidase 1; *IL1 $\alpha$* , Interleukin 1-alpha; *IL1 $\beta$* , Interleukin 1-beta; *IL6*, Interleukin 6; *p16*, Cdkn2a; *p19*, Cdkn2d; *p21*, Cdkn1a; *Tnfa*, tumor necrosis factor- $\alpha$ ; *Vcam1*, Vascular cell adhesion molecule 1.

| Gene         | Sequence                     |                              |
|--------------|------------------------------|------------------------------|
| Gapdh        | For: AACAGCAACTCCCACTCTTC    | Rev: CCTGTTGCTGTAGCCGTATT    |
| Gpx1         | For: CGACATCGAACCTGACATAGAA  | Rev: CAGAGTGCAGCCAGTAATCA    |
| IL1 $\alpha$ | For: GATACAGAGCACCCACAGAGAAC | Rev: GGCTAAGGAAACCCTAGGAAAG  |
| IL1 $\beta$  | For: TCCCAAAGTGCTGGGATTAC    | Rev: GAAAGCTCAGAGAGGAGGAAAG  |
| IL6          | For: GCCTGCATTAGGAGGTCTTT    | Rev: CCTGACACCAGCAAAGGATAA   |
| p16 (Cdkn2a) | For: GGTTCTCGCAGTACCATTGA    | Rev: CTGACTTCTGAGGTGGGTTTAG  |
| p19 (Cdkn2d) | For: GTCTCTCCGTTTCCCTTTCTTC  | Rev: GCTGGGCGCTTTATTTTCATTC  |
| p21 (Cdkn1a) | For: GGGTGCGGTGATGGATAAA     | Rev: ACTGCTGAGAACAGGAAGAAC   |
| Tnfa         | For: CCCTCGATGAAGCCCAATAAA   | Rev: CCCTCCCTCCATTCCTTAGATA  |
| Vcam1        | For: CCACTTGCCATCCTACTCTATTC | Rev: CAAGAGGGCTTAGGAGATTAAGG |