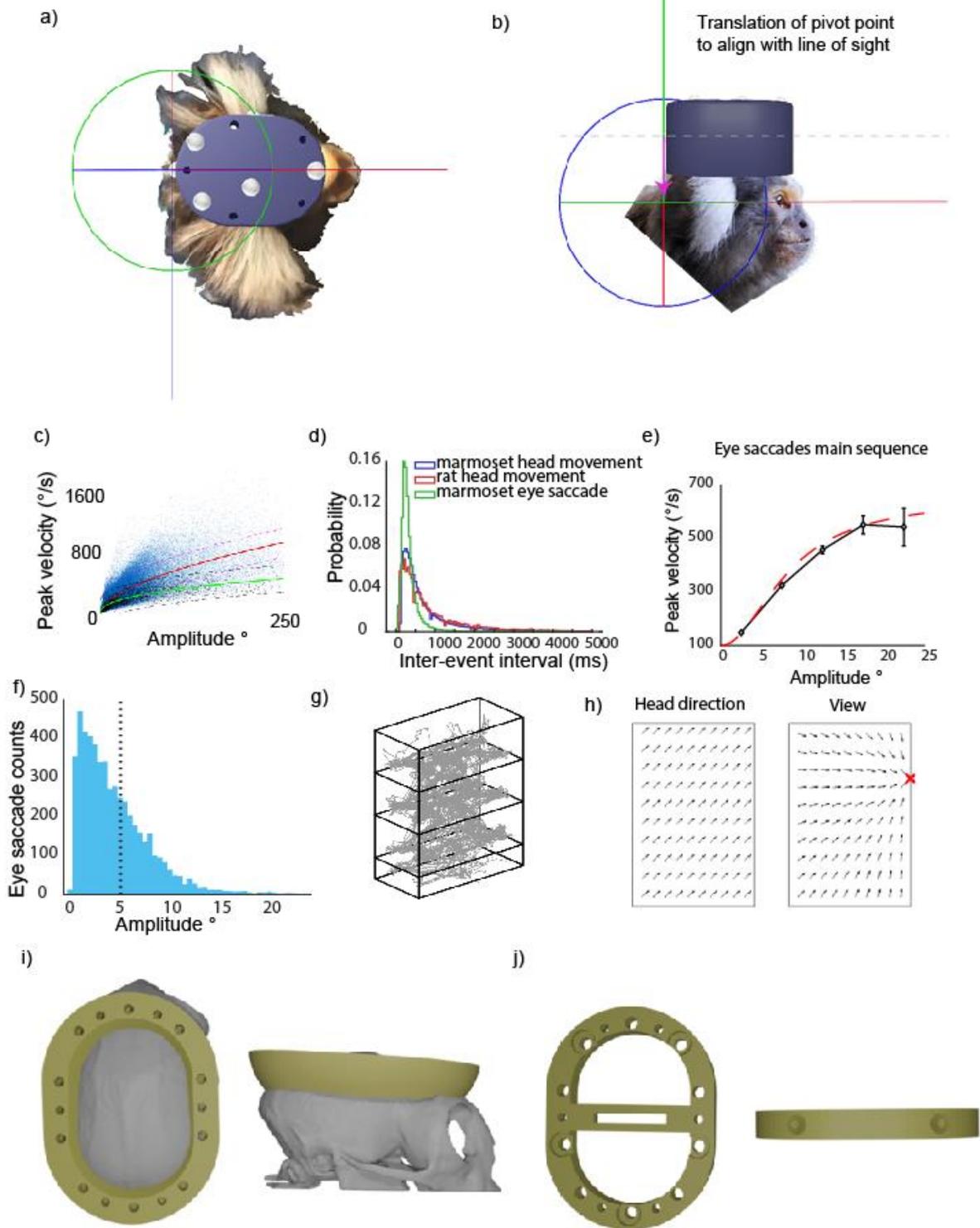
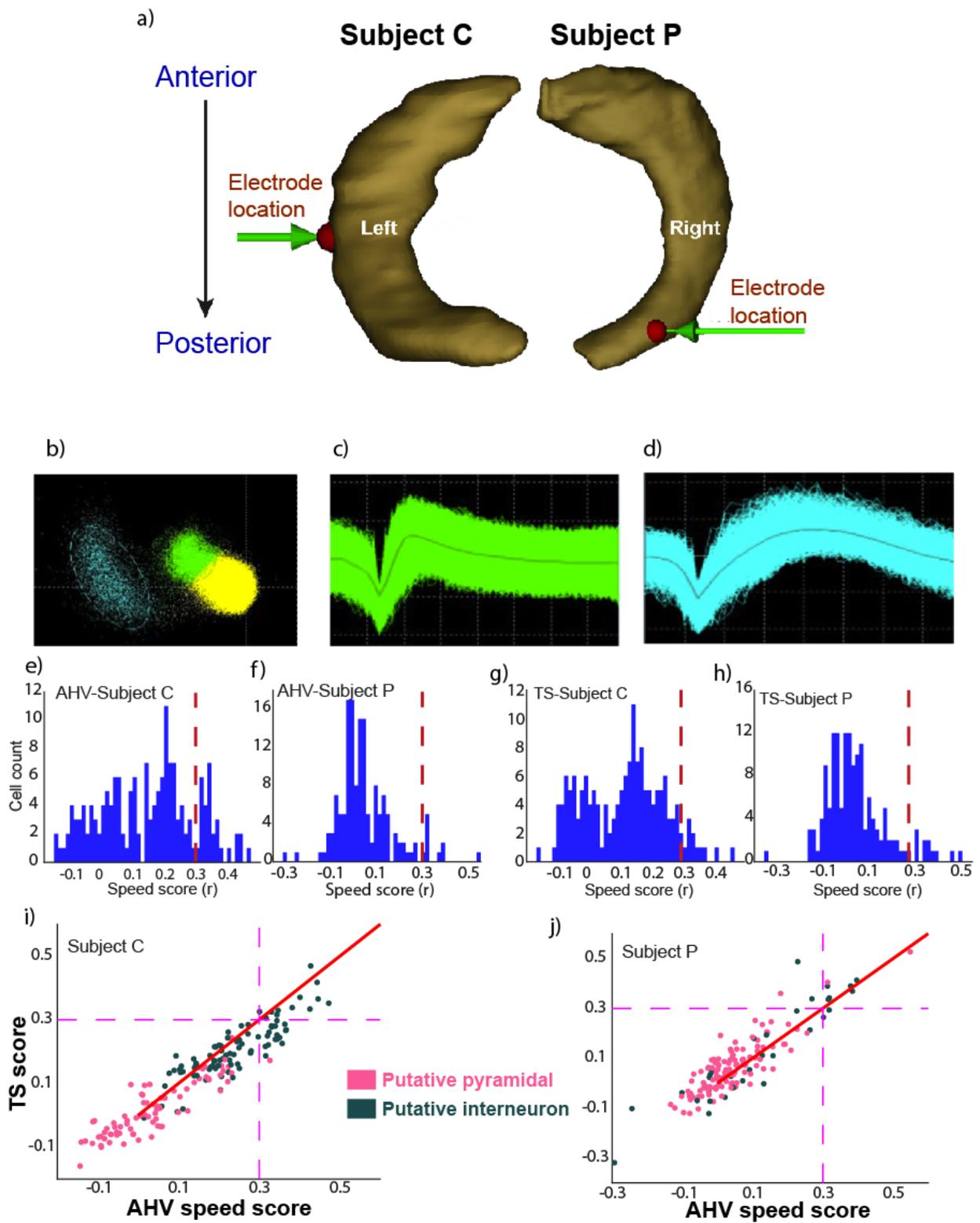


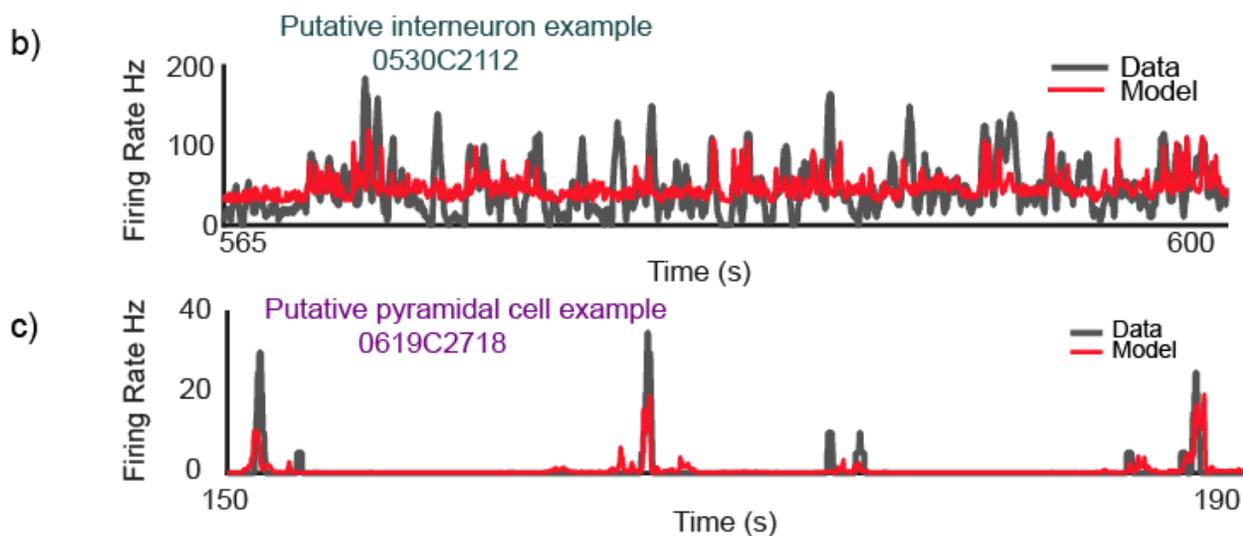
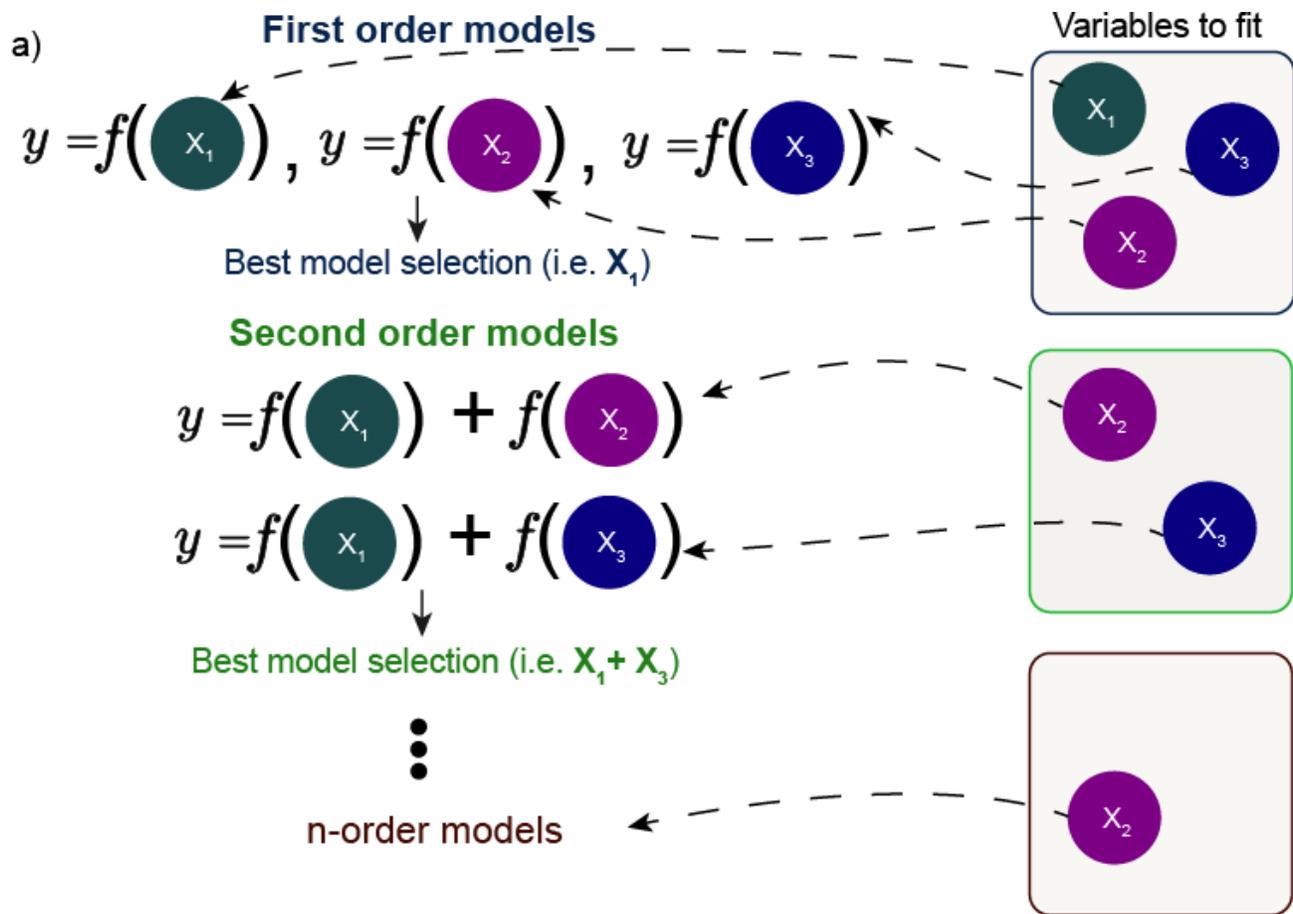
Supplementary Material:



Supplementary figure 1 (a) View from the top showing the alignment of the roll axis (red) with the visual axis, assuming a cyclopean eye. (b) Diagram illustrating the position of the pivot point (origin of rotation angles). The position of the pivot point (originally placed in the position of the most posterior retro-reflective marker) is linearly translated downwards, along the vertical axis to align with the subject's visual axis. (c) Main sequence (amplitude vs peak velocity) of head movements and Naka-Rushton function fit, for marmoset (blue markers, red line) and rat (black markers, green line); dashed lines correspond to 95% confidence intervals. (d) Distribution of inter-event interval for marmoset and rat head movements (blue and red distributions respectively) and marmoset eye saccades (green distribution). (e) Main sequence (amplitude vs peak velocity) of marmoset eye saccades, error bars convey 95% confidence intervals. The dashed line corresponds to a Naka-Rushton function fit. (f) Distribution of marmoset eye saccade amplitudes, dashed line placed at 5° . (g) Example trajectory traces (gray) of the 3D position during a recording session for subject C. (h) Illustration taken from a previous publication¹³⁵, displays the vector direction differences between head direction and view encoding. (i) 3D model of the recording chamber ring placed on the skull of the marmoset. (j) 3D model of the spacer that houses the electrode connector (omnientics 36pin), the spacer is placed on top of the chamber ring and it serves as a head-fixation base.

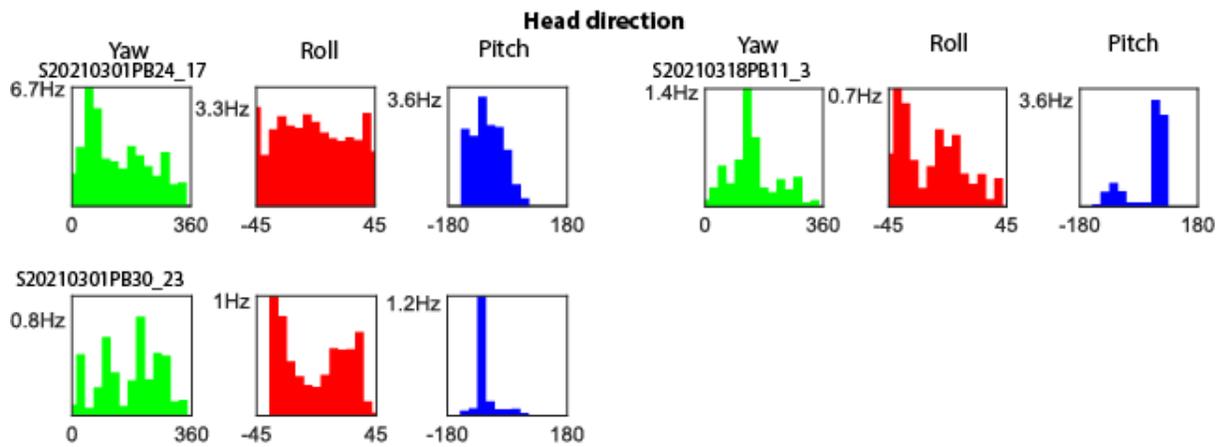
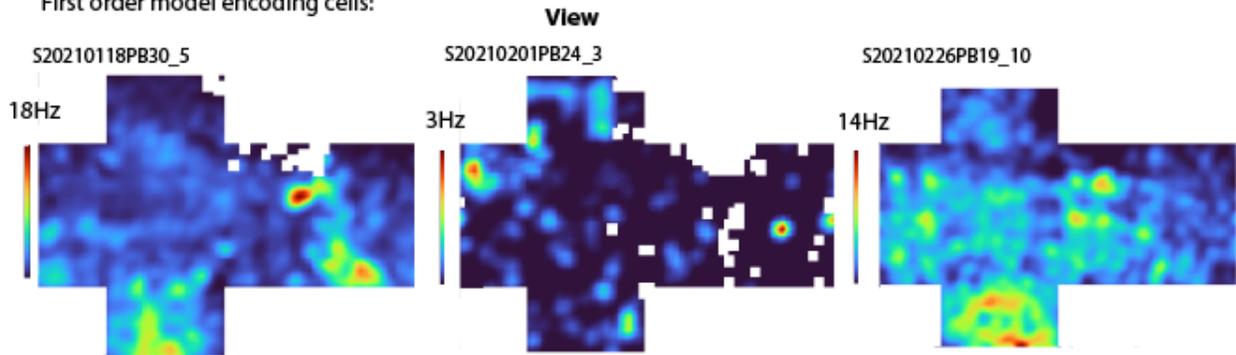


Supplementary figure 2 (a) MRI and micro-CT based 3D reconstruction of the electrode recording sites for subject C (left hippocampus) and subject P (right hippocampus). (b) Screenshot of a sorting session of one channel using Plexon software (Offline Sorter, Plexon Inc., Texas, USA). PCA view is displayed with labelled clusters, blue is a single unit, green is multi-unit activity, and yellow is the noise unit. (c) Multi-unit activity spike waveforms, aligned to the negative peak. (d) Single unit activity spike waveforms, aligned to the negative peak. (e,f) AHV speed scores for subject C and subject P respectively, the dashed line indicates a speed score of 0.3. (g,h) TS speed scores for subject C and subject P respectively, the dashed line indicates a speed score of 0.3. (i,j) Speed score distribution for both AHV and TS labelled according to cell type for subject C and subject P respectively, (green = putative interneuron, pink = putative pyramidal), the dashed line indicates a speed score of 0.3.

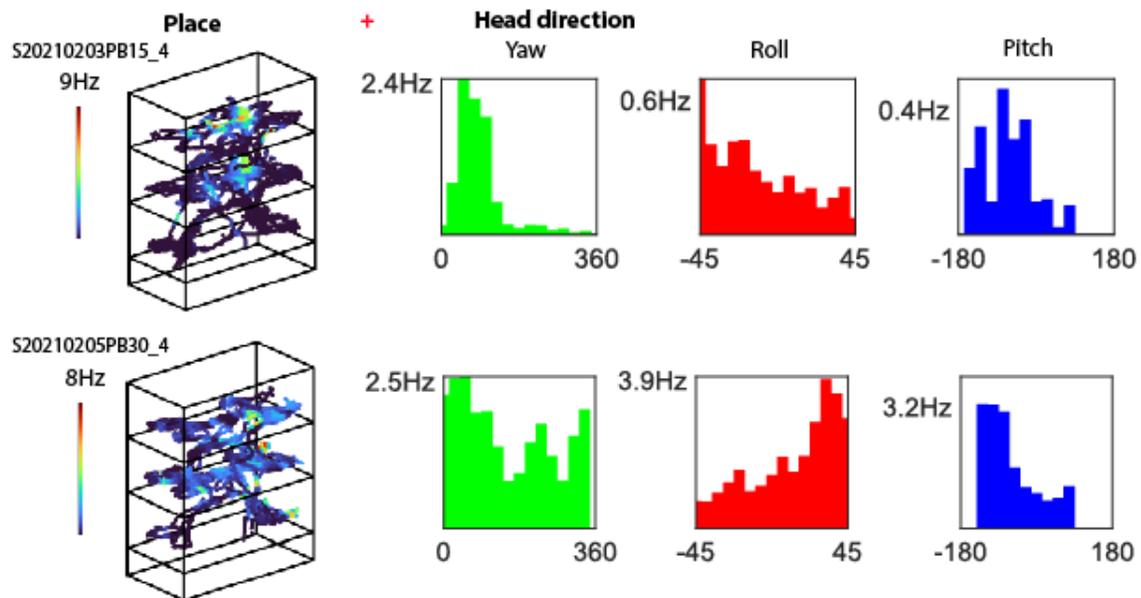


Supplementary figure 3 (a) Schematic describing the stepwise forward search GAM model selection process. (b) Real vs. predicted firing rate for a putative interneuron example cell (example cell from Fig.6c). (c) Real vs. predicted firing rate for a putative pyramidal example cell (example cell from Fig.6a).

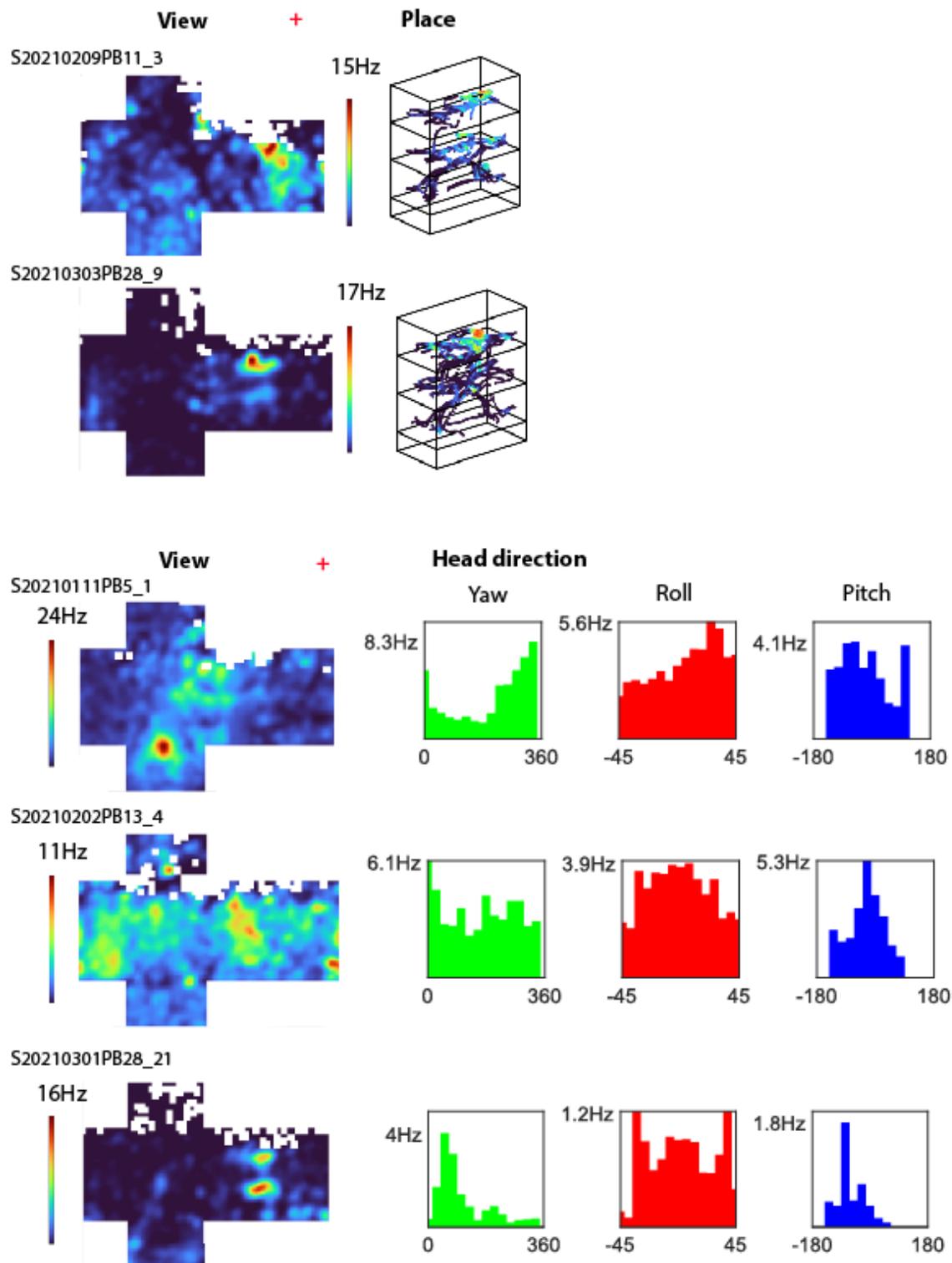
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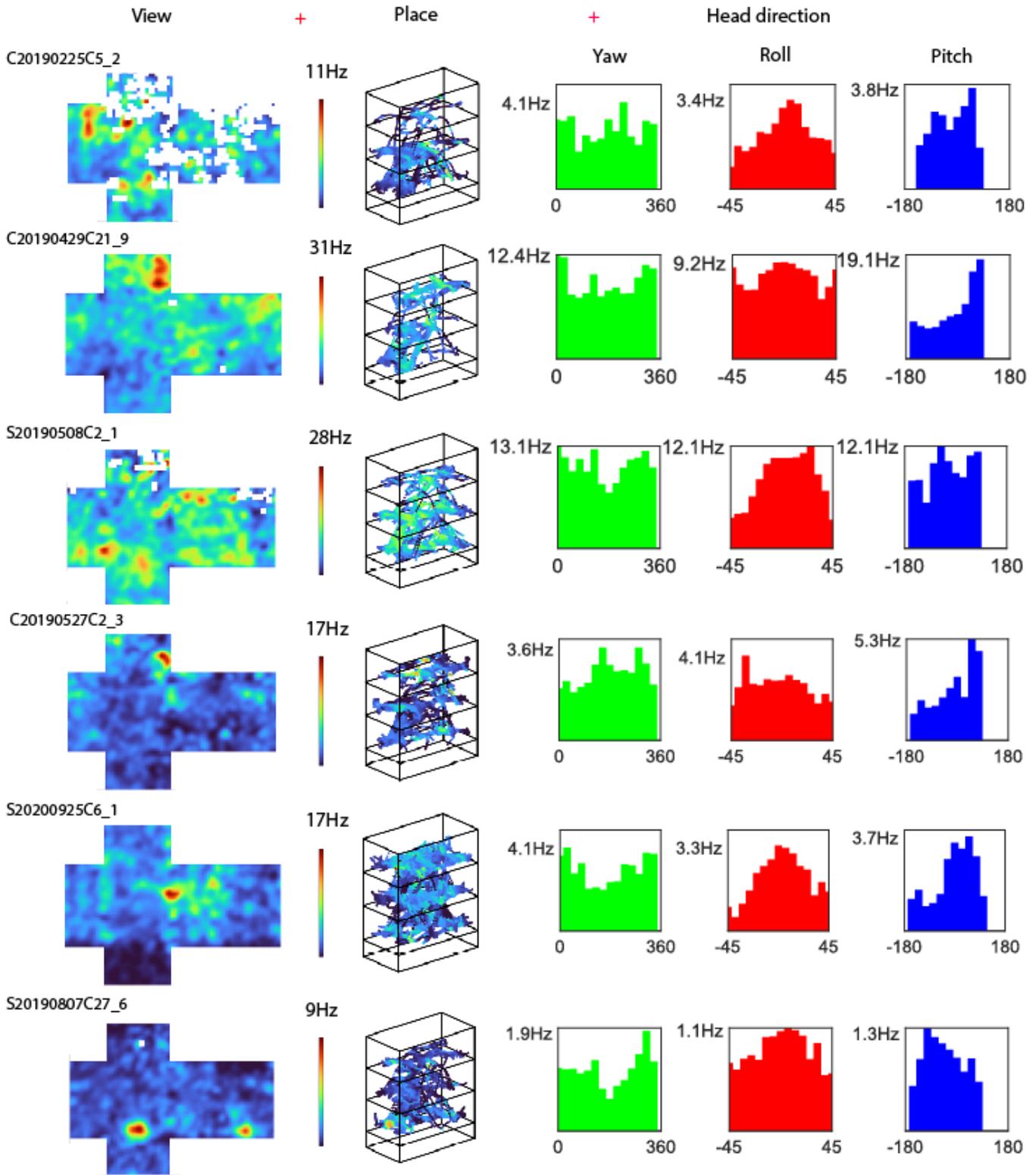
Second order model encoding cells:



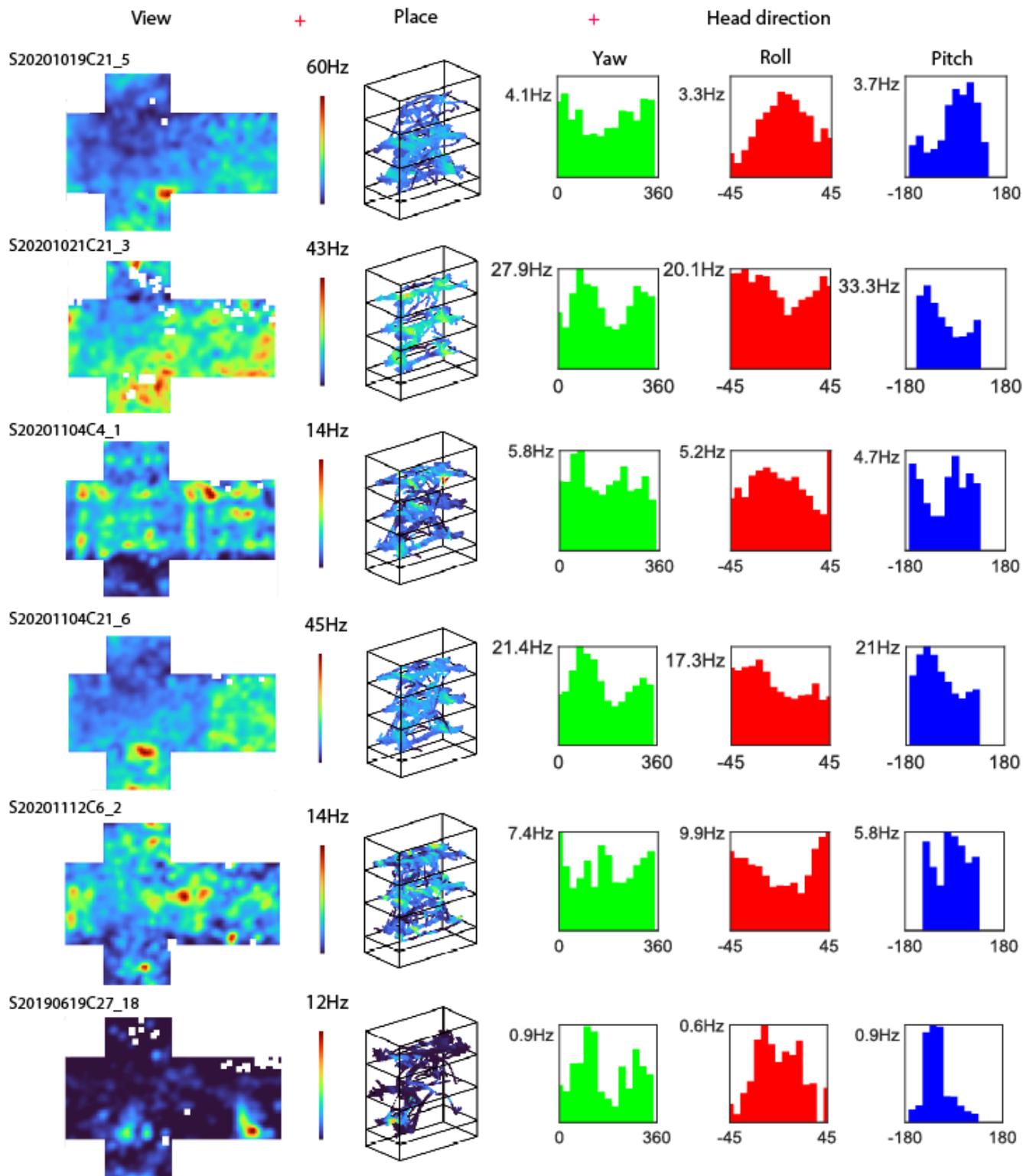
Second order model encoding cells:



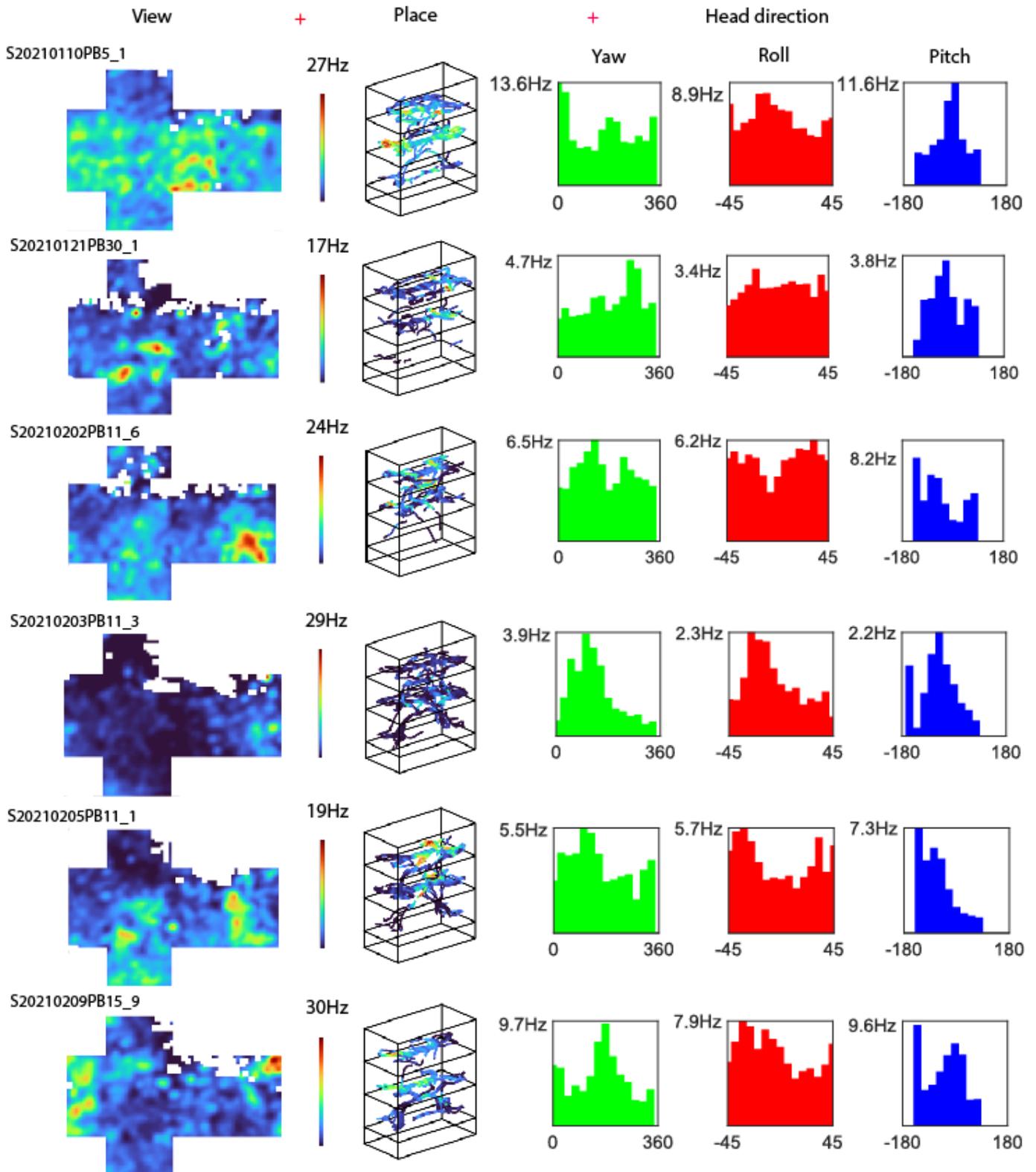
Third order model encoding cells:



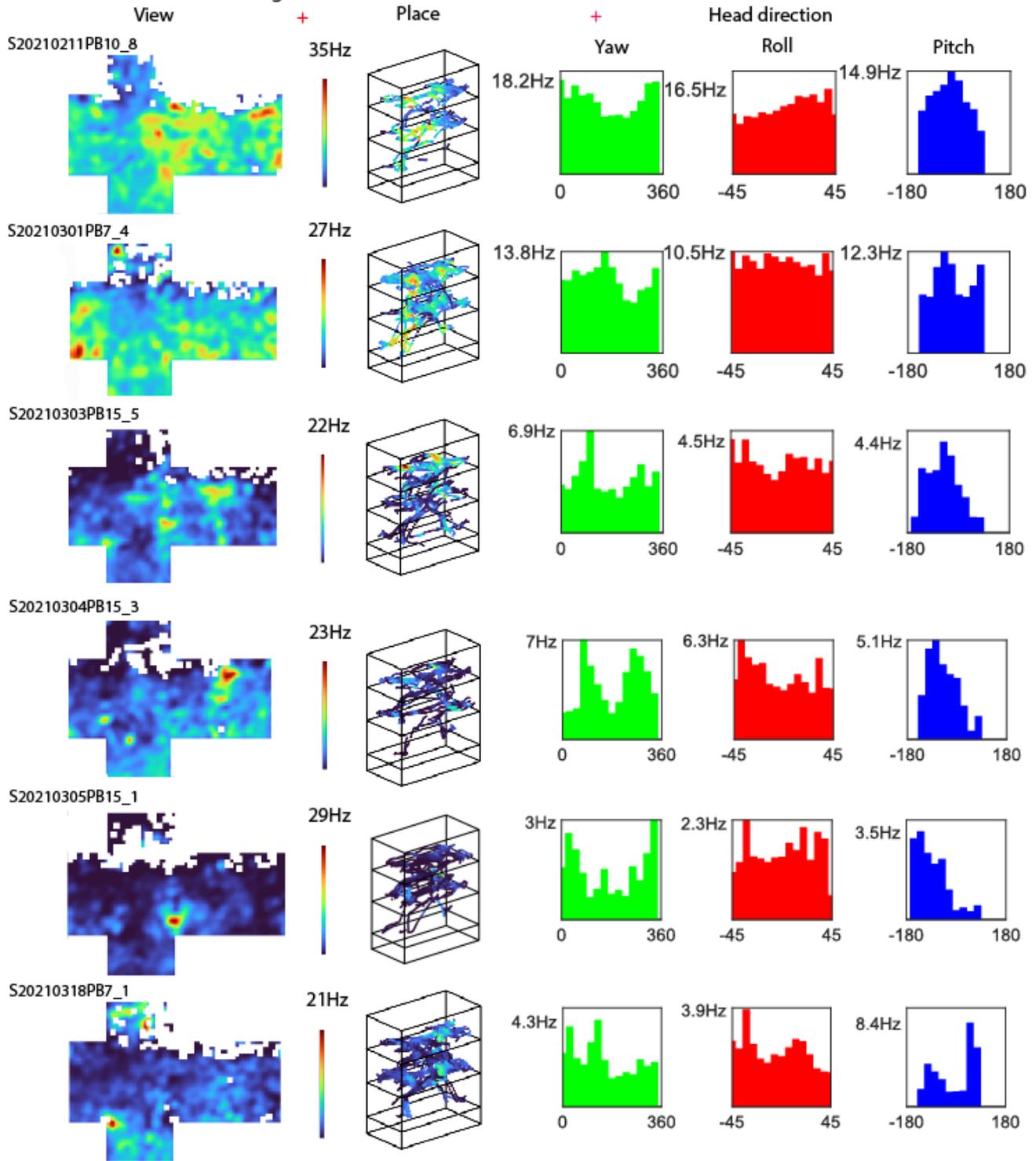
Third order model encoding cells:



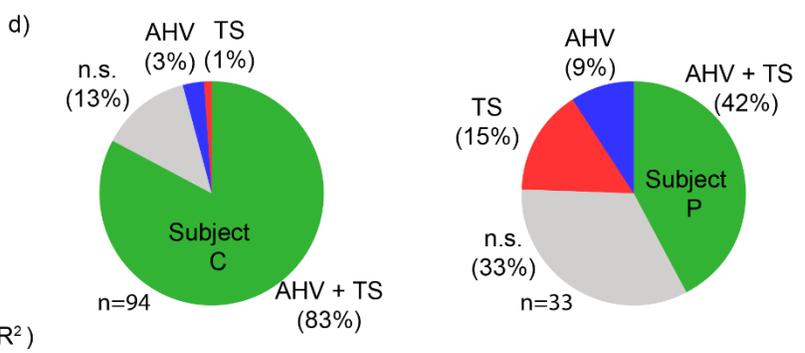
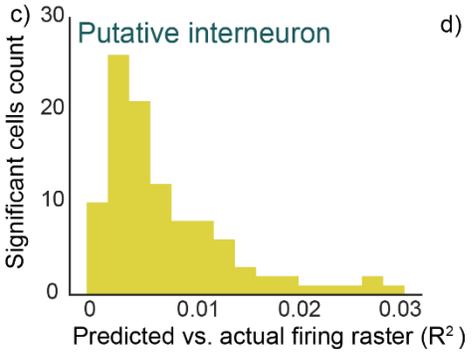
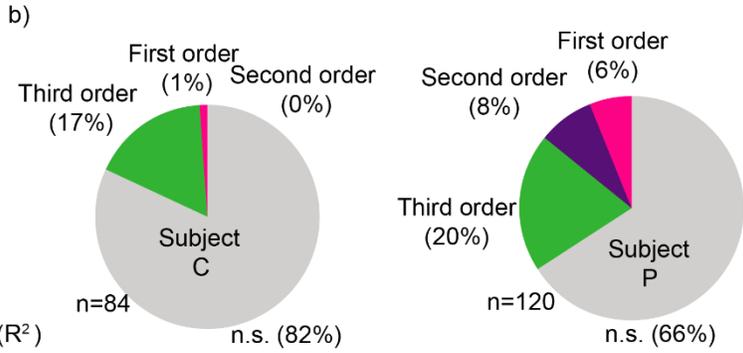
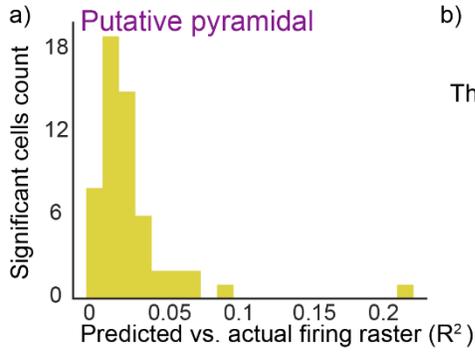
Third order model encoding cells:



Third order model encoding cells:



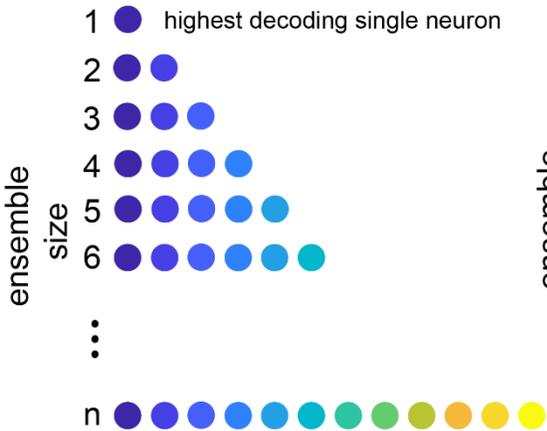
Supplementary figure 4 Additional single-cell examples are provided to illustrate the diverse types of encoding observed in putative pyramidal single neurons using the GAM model. Rate maps depicting view, place, and head direction are plotted to represent the corresponding behaviors encoded by individual neurons. In the case of cells encoding only one variable (first order models), a single rate map is observed. Conversely, in cells encoding multiple variables (second and third order models) multiple rate maps are displayed, indicating the specific behaviors being encoded. In head direction rate maps, firing rates corresponding to yaw angles (horizontal head direction) are color-coded as green, roll angles (lateral tilt) as red, and pitch angles (vertical axis) as blue. In the case of third order models, the view and place rate maps share the same color map, with the maximum firing rate indicated at the top of the color bar.



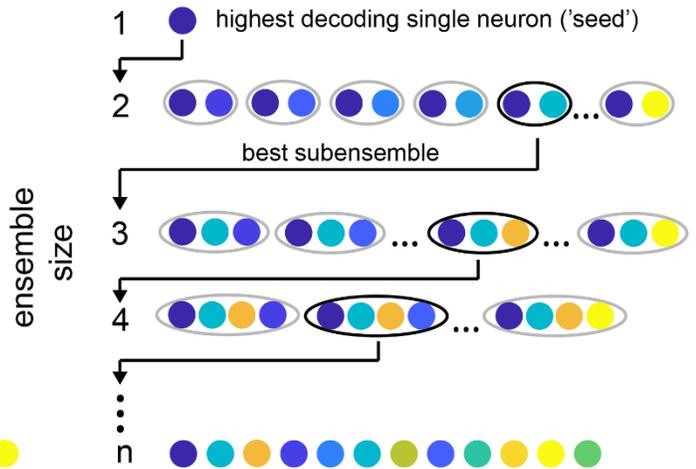
e) **Pseudopopulation pool of neurons**

most informative least informative

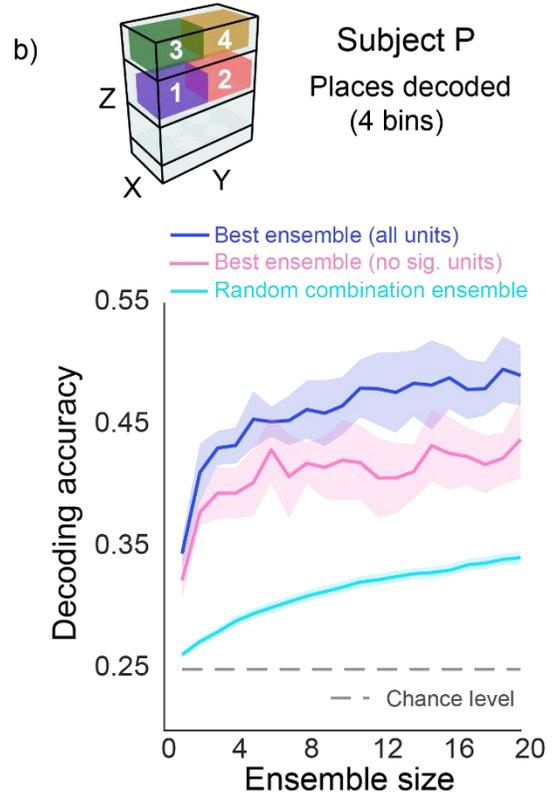
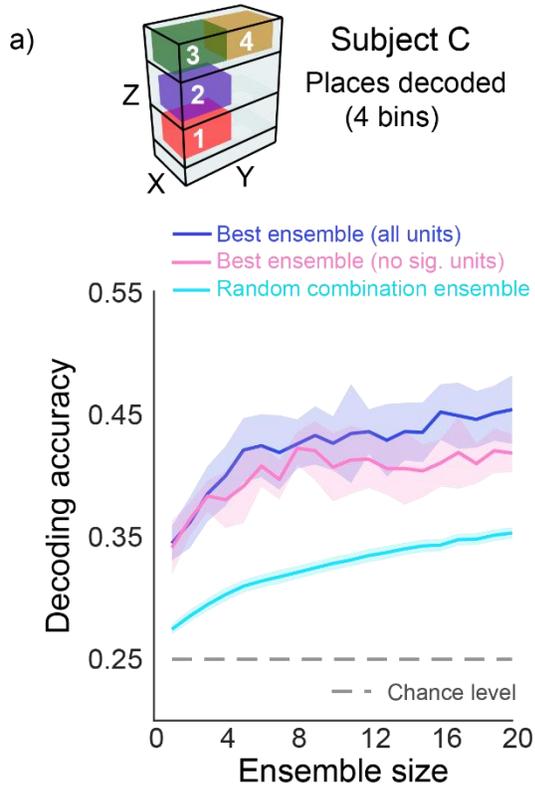
'Best individual neuron' procedure



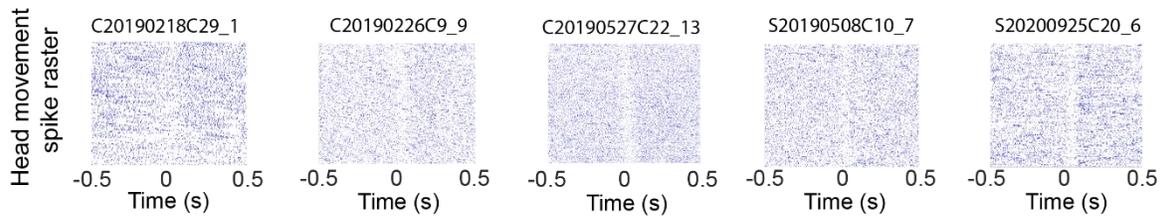
'Best ensemble' procedure



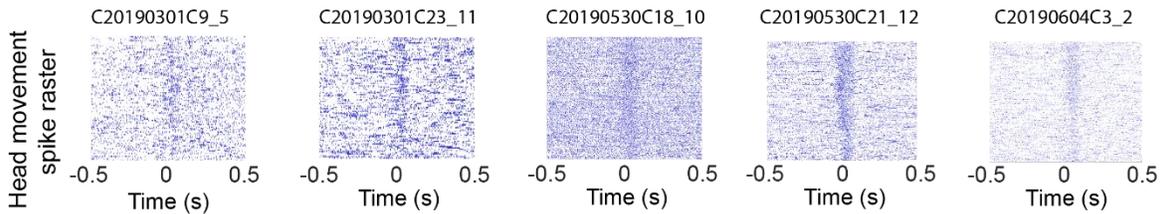
Supplementary figure 5 (a) Average R^2 between predicted and real spike rasters of all significantly encoding putative pyramidal cells. (b) Proportion of putative pyramidal encoding cells for subject C (left) and subject P (right). (c) Average R^2 between predicted and real spike rasters of all significantly encoding putative interneuron cells. (d) Proportion of putative interneuron encoding cells for subject C (left) and subject P (right). Since each recording session is divided into 5 equally-timed folds, each predicted raster is a continuous time series approximately 8-12 min. long. (e) Schematic describing the best ensemble building procedure. (left) The most informative neuron is selected, as assessed by the single-neuron with the highest SVM decoding accuracy of place. (right) The most informative neuron is used as a seed to iteratively add units (looped through the remaining units) to the subsequent subensembles until the 'best ensemble' n-combination of neurons is obtained.



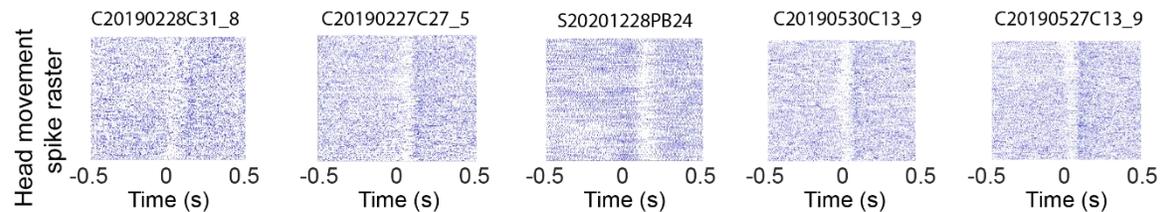
d) **Down-modulation**



e) **Up-modulation**



f) **Up/down-modulation**



Supplementary figure 6 (a,b) (top) 3D diagram of the binned place locations used to decode the subject's position, (bottom) blue and pink lines correspond to raw decoding accuracy (y axis) as a function ensemble size (number of neurons, x axis), shaded area corresponds to 95% confidence intervals. Blue solid lines correspond to the best ensemble constructed from a pool of all recorded putative pyramidal neurons. Pink solid lines correspond to the best ensemble constructed from a pool of non significantly selective cells (as per GAM encoding analysis). R^2 goodness of fit value is reported. The cyan lines correspond to decoding accuracy of a randomized combination of neurons (100 iterations), shaded area corresponds to 95% confidence intervals. The gray dashed lines correspond to chance decoding accuracy (1/4, 0.25). **(d,e,f)** Additional single-cell examples of down-modulated, up-modulated and up/down modulated cells respectively. Each raster plot corresponds to a single neuron and is aligned (time 0) with the peak velocity time of a head movement, where each row corresponds to one head movement event and each blue dot represents one neuronal spike, negative times indicate the neuron's firing before the peak velocity of a head movement, while positive times indicate the neuron's firing afterwards.

Supplementary movie 1: <https://youtu.be/Vz4lMer5g8o> (Recording session video example)