

Supplementary Material

**Elovanoid-N34 is a homeostatic switch that modulates
TXNRD1 for cell survival**

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Running title: Elovanoid-N34 homeostatic switch modulates TXNRD1

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SUPPLEMENTARY FIGURES

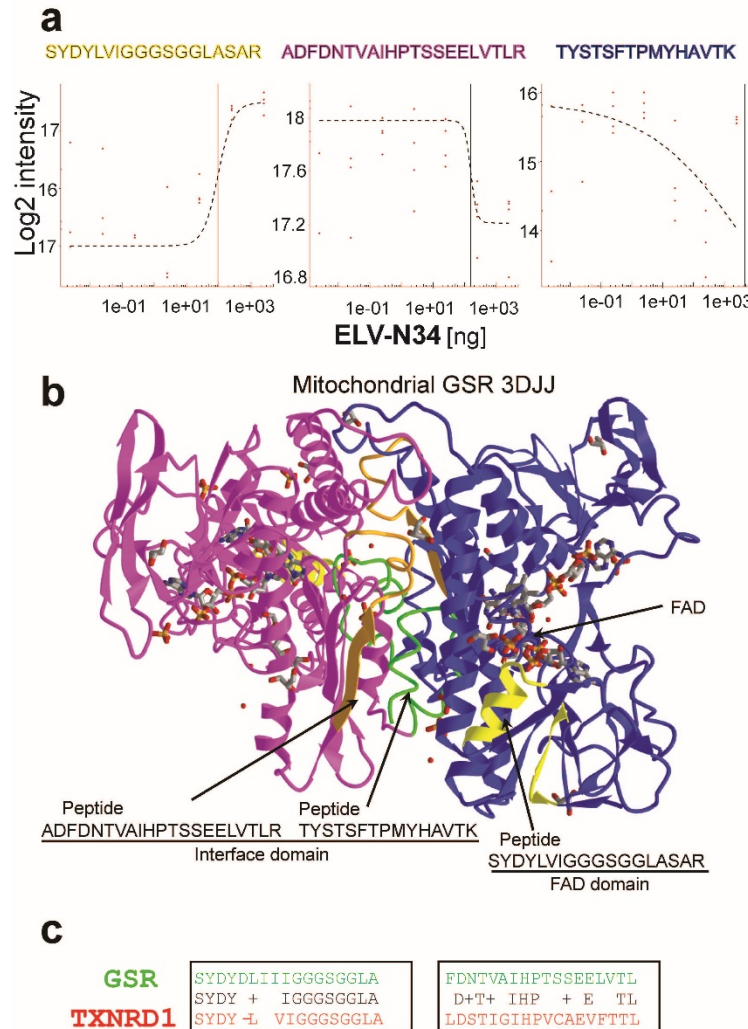


Fig. S1. Analysis of Target Candidate Protein Glutathione reductase, mitochondrial (GSR; P00390). The GSR displayed 3 peptides differentially proteolyzed in contact with ELV-N34. **a**, The plots show the peptide quantity vs. concentration of ELV-N34. The dotted line was determined by non-linear fit regression to obtain ED50 (vertical line). **b**, Structure shown is based on the X-ray chromatographic structure of human glutathione reductase (3DJJ), showing the 3D position of the peptides.

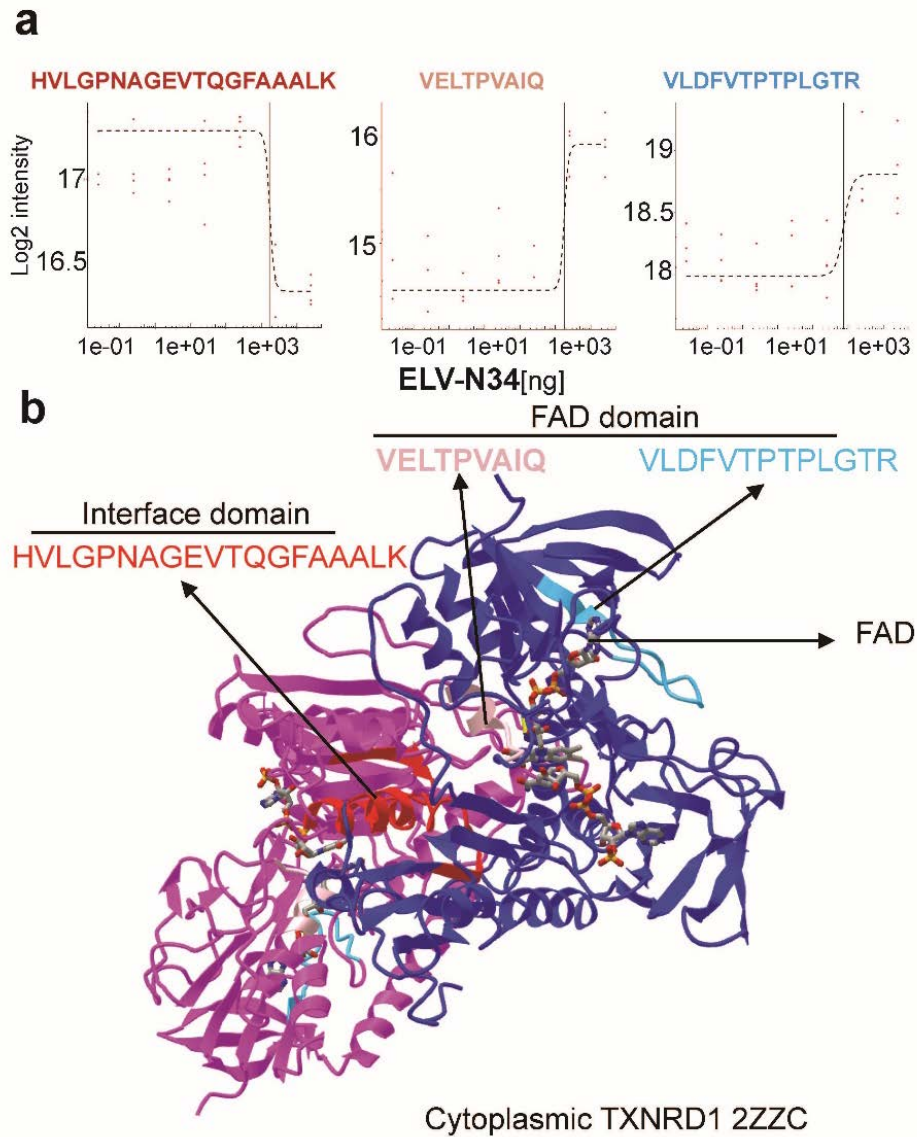


Fig. S2. Thioredoxin reductase, cytoplasmic (TXNRD1; Q16881). **a**, Plot shows the concentration curves for three of the 15 top quantified TXNRD1 peptides comparing DMSO and ELV-N34 treated samples. **b**, Highlighted peptides over the 3D ribbon model based on the X-ray crystallography (ID: 2ZZC).

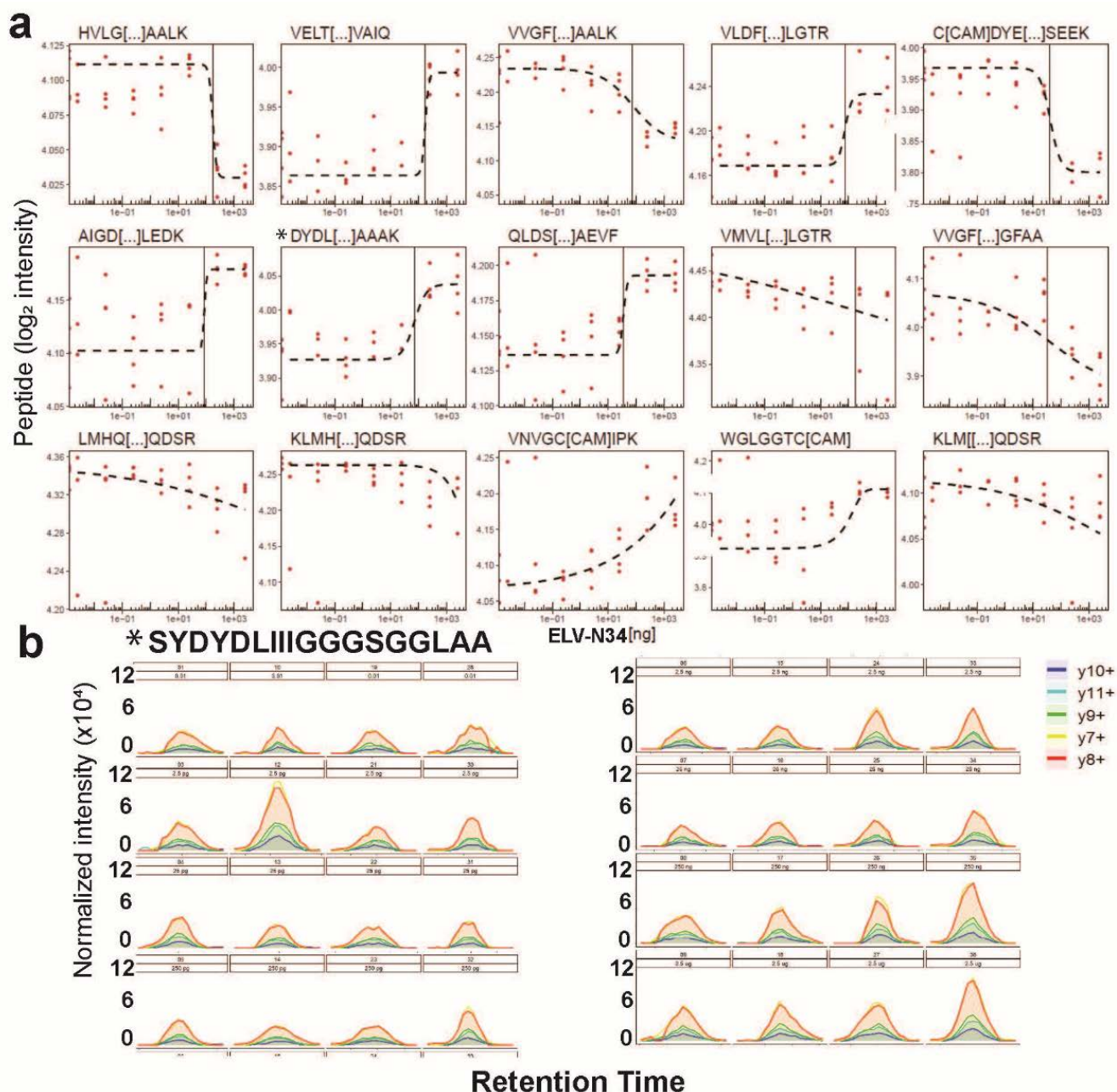


Fig. S3. Glutathione reductase (GSR; P00390). **a**, Plot shows changes of the top 15 quantified GSR peptides in DMSO and ELV-N34-treated samples. In the y-axis, the change is represented by $\log_2$ intensity, and the x-axis is the amount of ELV-N34 added in ng per reaction tested. Three peptides were significantly changed upon treatment with 250 nM ELV-N34 and used for

dose-response curve calculation. **b**, Extracted ion chromatogram plots show the response of peptide SYDYDLIIIGGSGGLAA (blue, green, and red lines show three). With an asterisk, we marked the peptide in **a** and **b**.

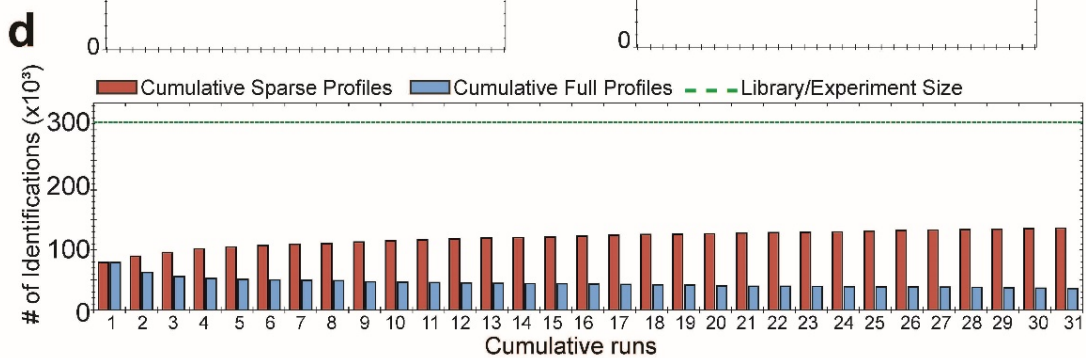
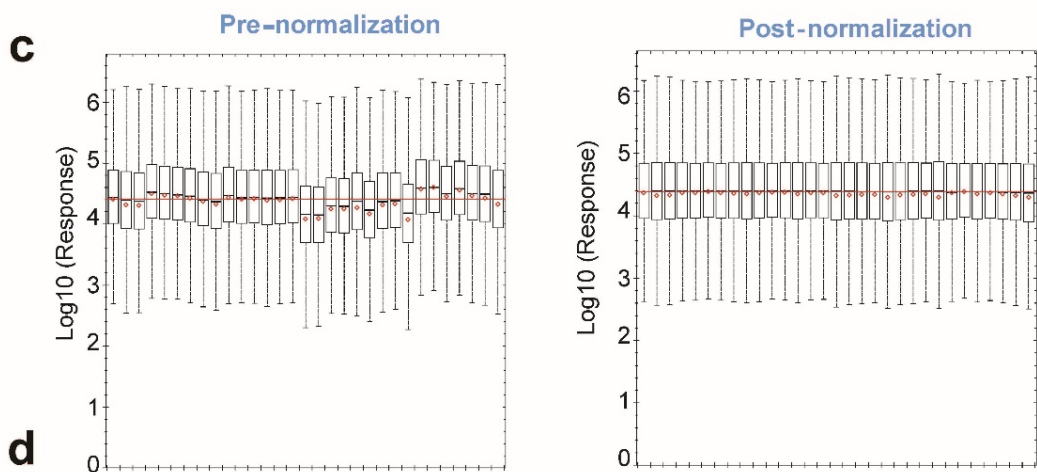
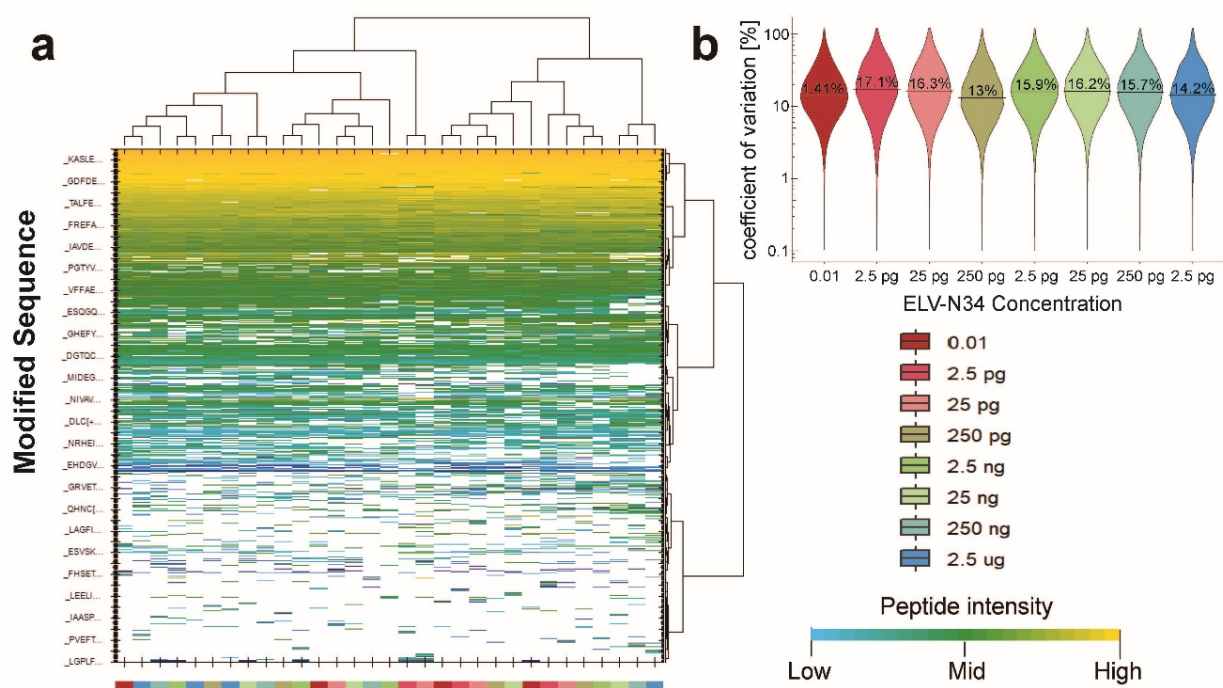


Fig. S4. Quality control of the first set of samples. **a**, Unsupervised clustering of 105'185 peptide intensities does not reconstruct sample groups. Hierarchical clustering analysis using the Manhattan distance measure using all peptide intensities across all samples was carried out. Clustered data is displayed as a heat map. Sample dendrogram (top) displays no separation according to sample group, indicating that no sample preparation or data acquisition artifact is confounding the analysis. **b**, Technical variation of target deconvolution experiment was assessed as coefficient of variance for peptide quantities between sample groups. Median technical variation was between 13.0 and 17.1 % within sample groups. Violin plot shows the distribution of coefficients of variation (CV) for peptide quantification, grouped by sample group. **c**, Minimal normalization was performed across all runs and was successful in reducing experimental variation. **d**, Number of precursors quantified across all runs. Data completeness is referred to as precursors quantified by HRM in the samples provided. Cumulative sparse profiles account for all precursors in the data set quantified at least once across the samples with high confidence (sparse data set). Cumulative full profiles account for all precursors in the data set quantified in each single sample with high confidence (complete data set). Size of the spectral library is shown as a dotted line.

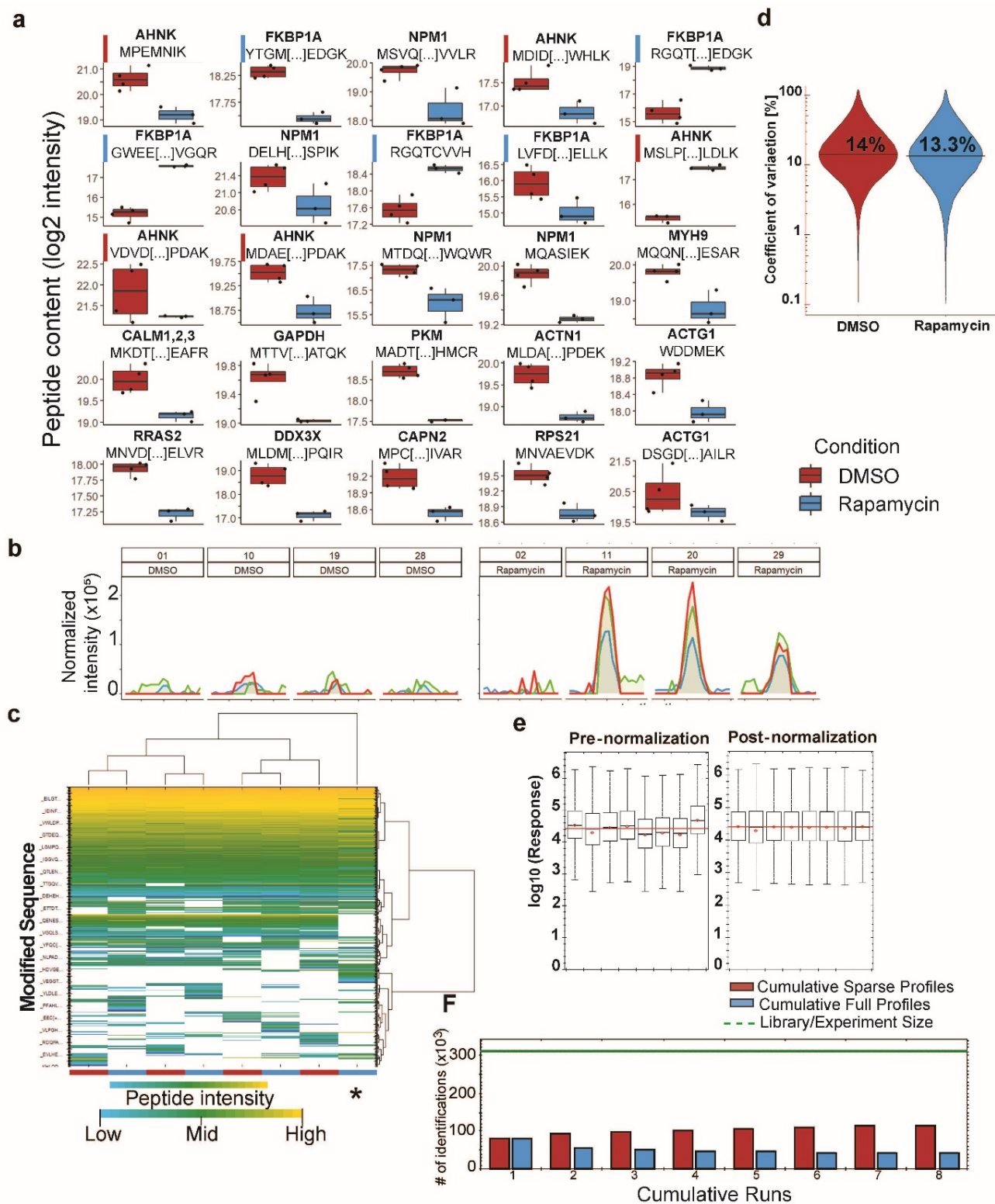


Fig. S5. Specificity control using Rapamycin instead of ELV-N34 for the first set of samples. The already known amount of 1'135 peptides was significantly changed between

samples treated with DMSO and Rapamycin. **a**, Box plot of the top 25 peptides and corresponding proteins significantly changed between DMSO and Rapamycin sample groups with 5 out of 25 high-ranking responder peptides. The candidate peptides shown were ranked by q-value. Data is grouped by sample groups and shown as box plots where the middle line depicts the average, and the limit of the boxes are the 1st and 3rd quartile. The bars show the standard deviation of the group. **b**, The known target of Rapamycin Peptidyl-prolyl cis-trans isomerase FKBP1A (FKBP1A; P62942). FKBP1A is identified, and extracted ion chromatogram plots show the response of peptide RGQTCVVHYTGMLEDGK²⁺. **c**, Hierarchical Clustering (Rapamycin Control), unsupervised clustering of all peptide intensities does not reconstruct sample groups; the Manhattan distance measure using all peptide intensities across all samples was carried out. Clustered data is displayed as heat map. Sample dendrogram (top) displays no separation according to sample group, indicating that no sample preparation or data acquisition artifact is confounding the analysis. One sample (*) was excluded from analysis due to excessive variance. **d**, Technical variation: Median technical variation was between 13.3 and 14.0 % within sample groups. Violin plot shows distribution of coefficients of variance (CV) for peptide quantification, grouped by sample group. **e**, Normalization: Minimal normalization was performed across all runs and was successful in reducing experimental variation. **f**, Number of precursors quantified across all runs. Data completeness is referred to as precursors quantified by HRM in the samples provided. Cumulative sparse profiles account for all precursors in the data set quantified at least once across the samples with high confidence (sparse data set). Cumulative full profiles account for all precursors in the data set quantified in each single sample with high confidence (complete data set). Size of the spectral library is shown as dotted line.

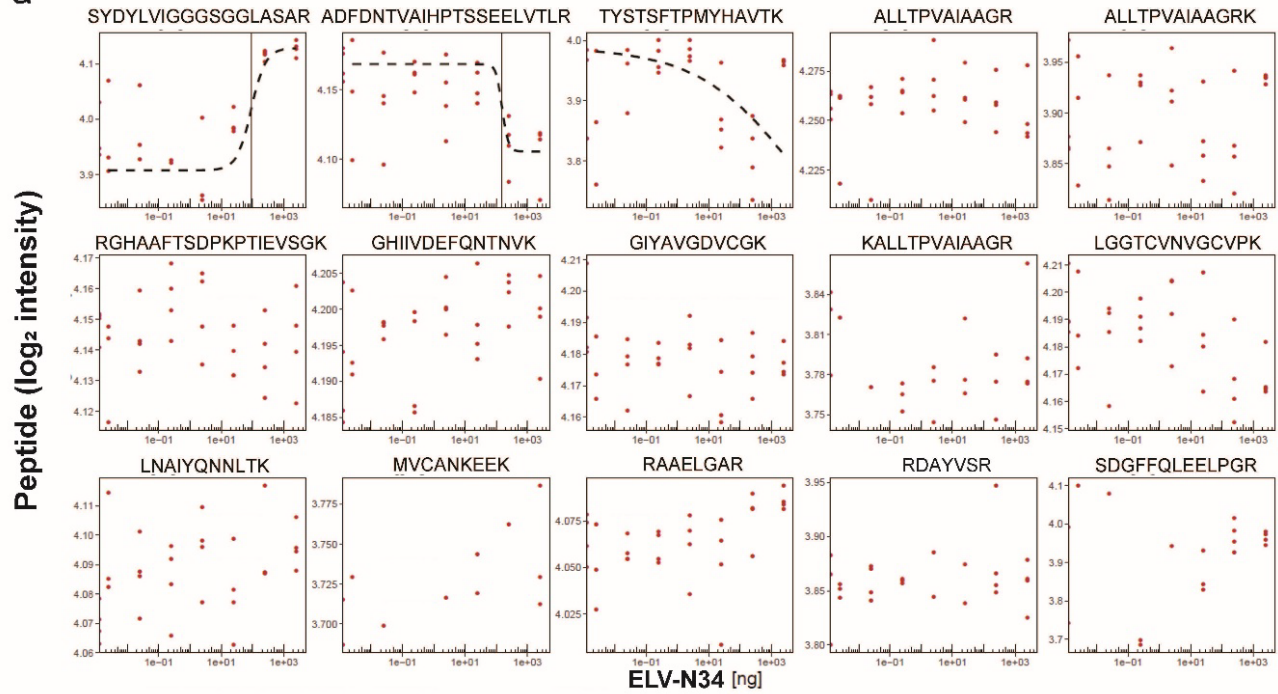
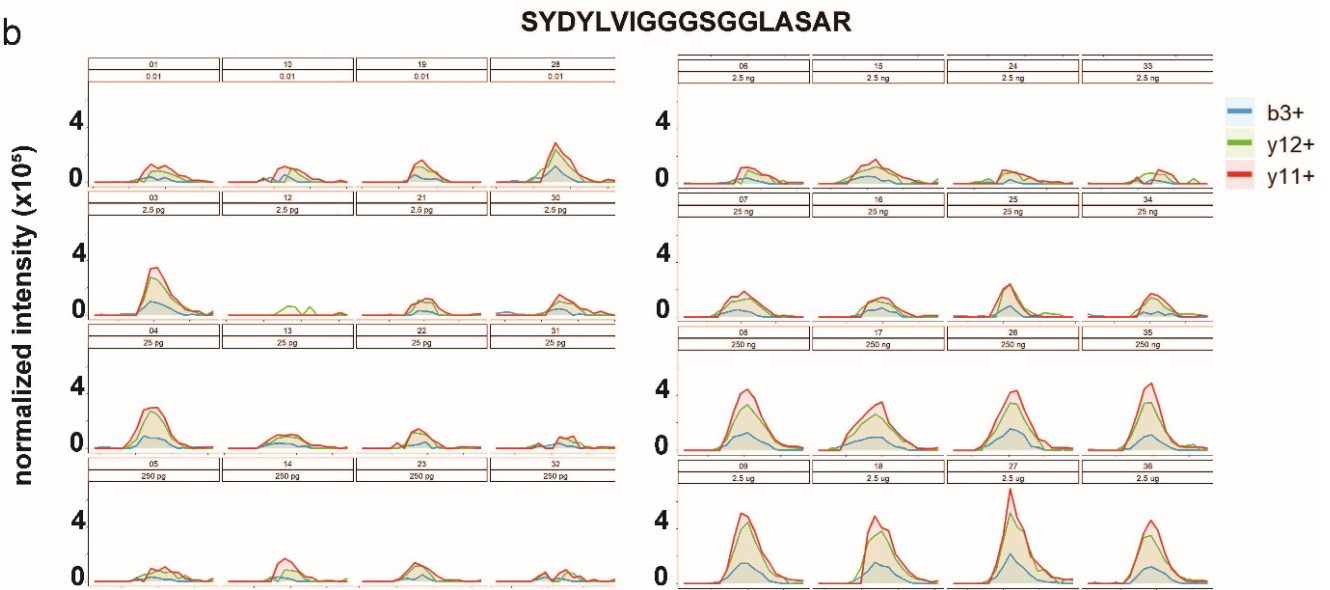
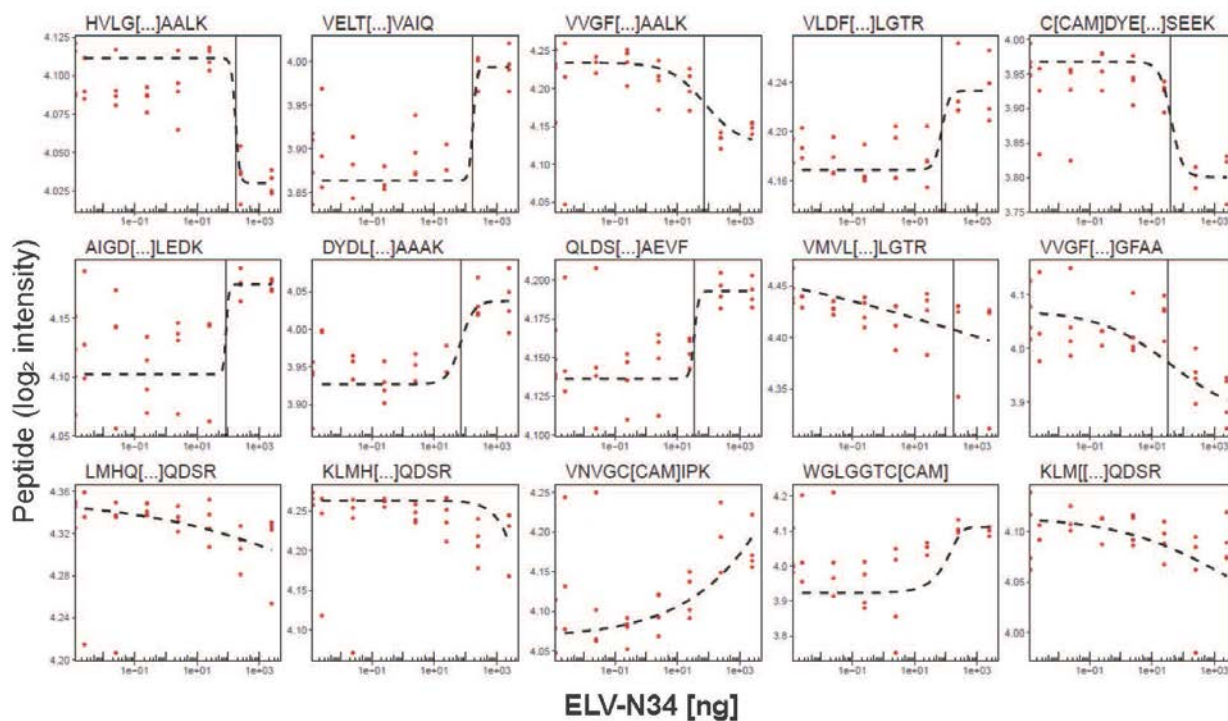
a**b**

Fig. S6. a, Glutathione reductase, mitochondrial (GSR; P00390) peptides that appeared in the UOS + ELV-N34 LiP comparison. From the top 15 quantified GSR peptides in the comparison DMSO and ELV-N34 treated samples, 3 peptides were significantly changed upon treatment

with 250 ng ELV-N34 and used for dose-response curve calculation. **b**, XIC plots show response of peptide SYDYLVIGGSGGLASAR.

a



b

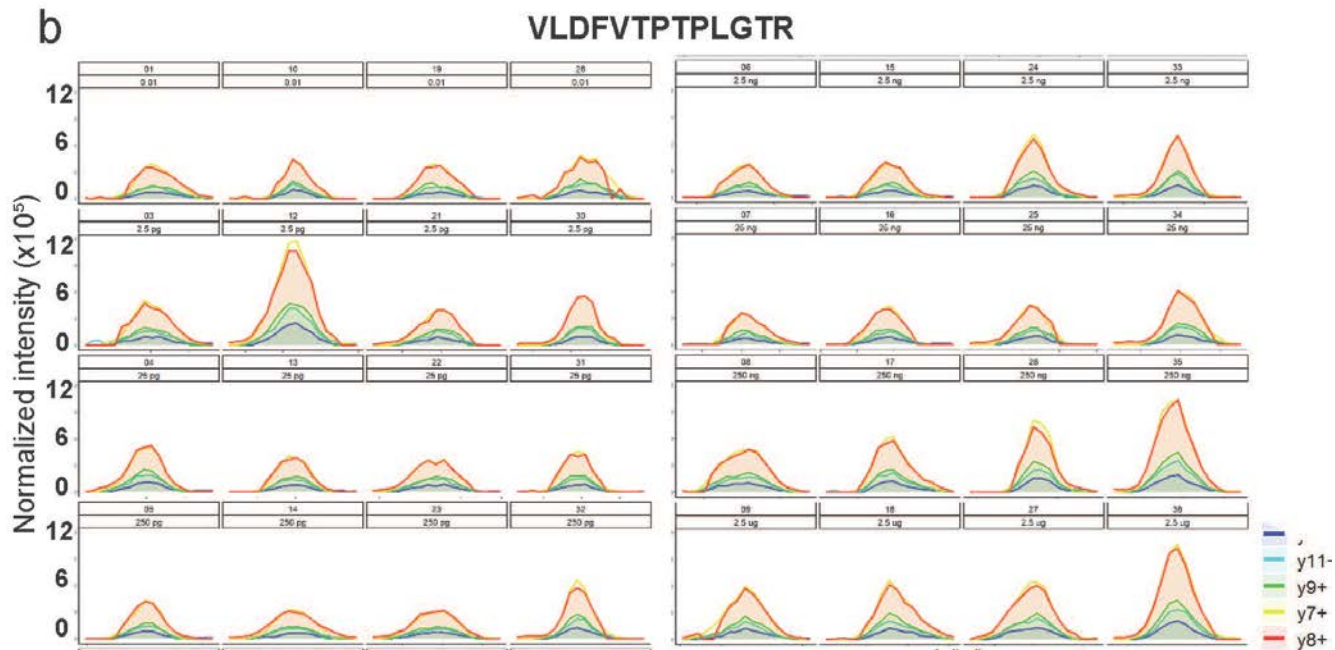


Fig. S7. Thioredoxin reductase 1, cytoplasmic (TXNRD1; Q16881). **a**, The plots show the fold change of the top 15 quantified TXNRD1 peptides in DMSO and ELV-N34 treated samples. More than 15 peptides were significantly changed upon treatment with 250 nM ELV-N34 and used for dose-response curve calculation. **b**, XIC plots show response of peptide VLDFVTPTPLGTR.

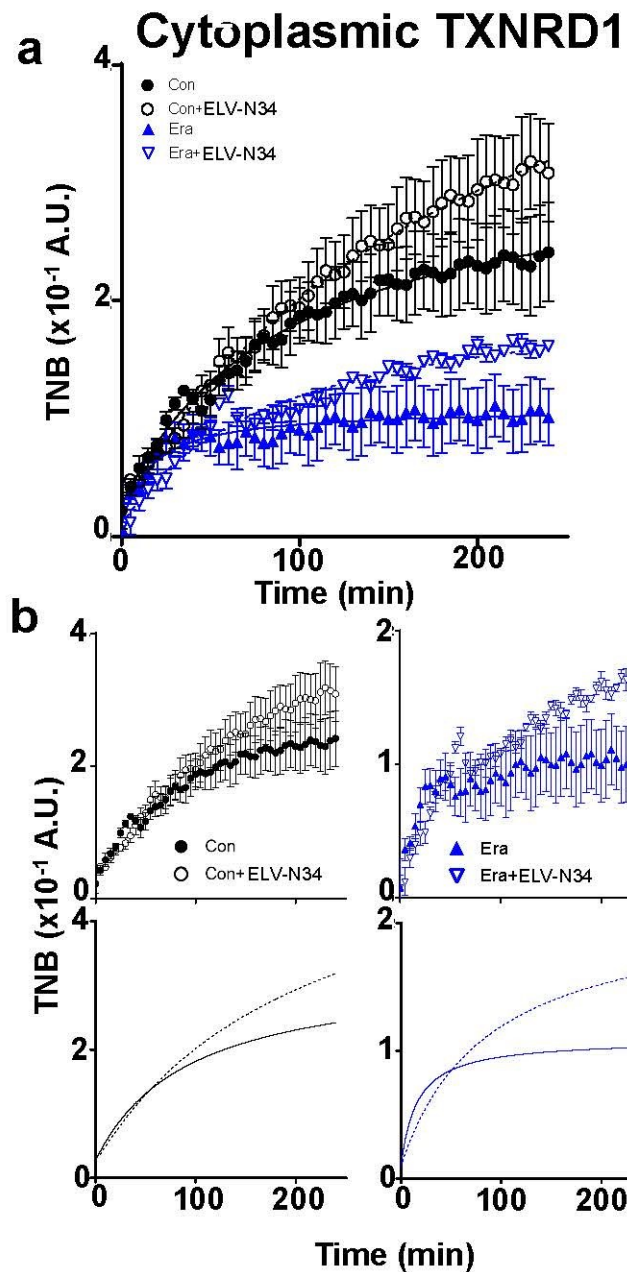


Fig. S8. Addition of ELV-N34 to the membrane and cytosolic fractions. Curves of Product vs. Time. **a-e**, 250 ng of ELV-N34 per 100 ng of cytoplasmic protein in 200 μ l reaction was added to cytosolic (**a, b**), and membrane (**c, d**) fractions of ABC cells exposed to Erastin or

naïve, the activity of TXNRD1 was recorded as Product (absorbance at 412 nm) vs. Time (min), and the subtraction resulting from the reaction and the reaction plus inhibitor was plotted.

Crude Extract TXNRD1

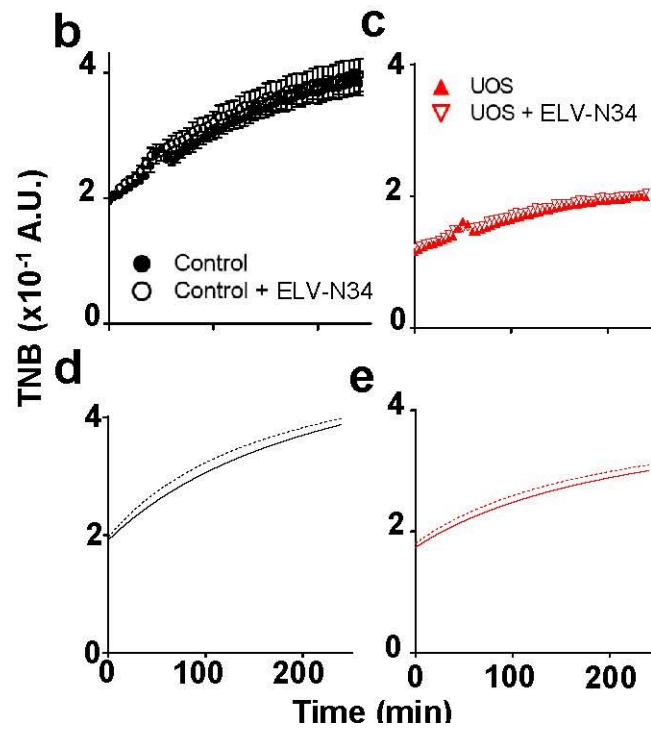
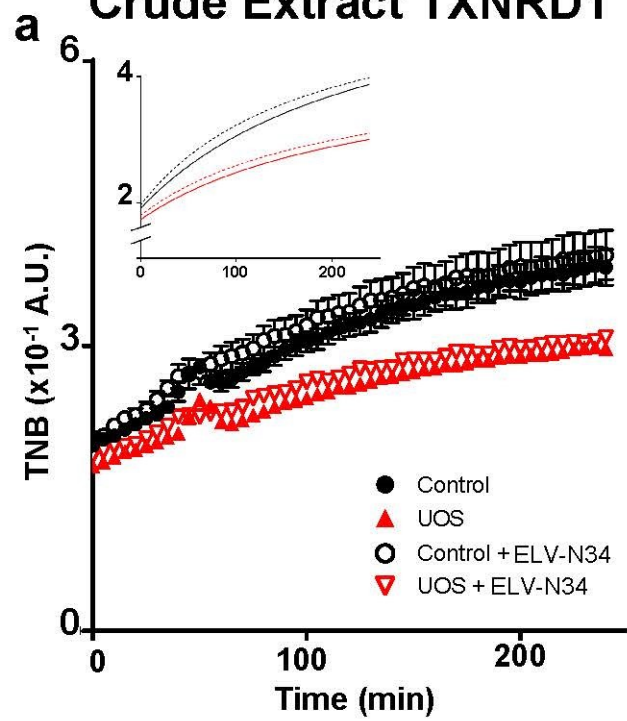


Fig. S9. Curves of Product vs. Time of the crude extract. a-e, 200 nM ELV-N34 was added to the ABC cells exposed to UOS or naïve. The activity of TXNRD1 was recorded on the total extract of the cells exposed to the mentioned treatments as Product (absorbance at 412 nm) vs. Time (min). Spureus reductase activity was ruled out using a specific inhibitor of TXNRD1. **a,** Merge curves of the four conditions. **b, c,** Mean of three observations and the bars represent the standard error of the mean for Control and Control + ELV-N34 (**b**) and UOS and UOS + ELV-N34 (**c**). **d, e,** The non-linear regression curves obtained for the data points in **b** and **c**.

Cytoplasmic TXNRD1

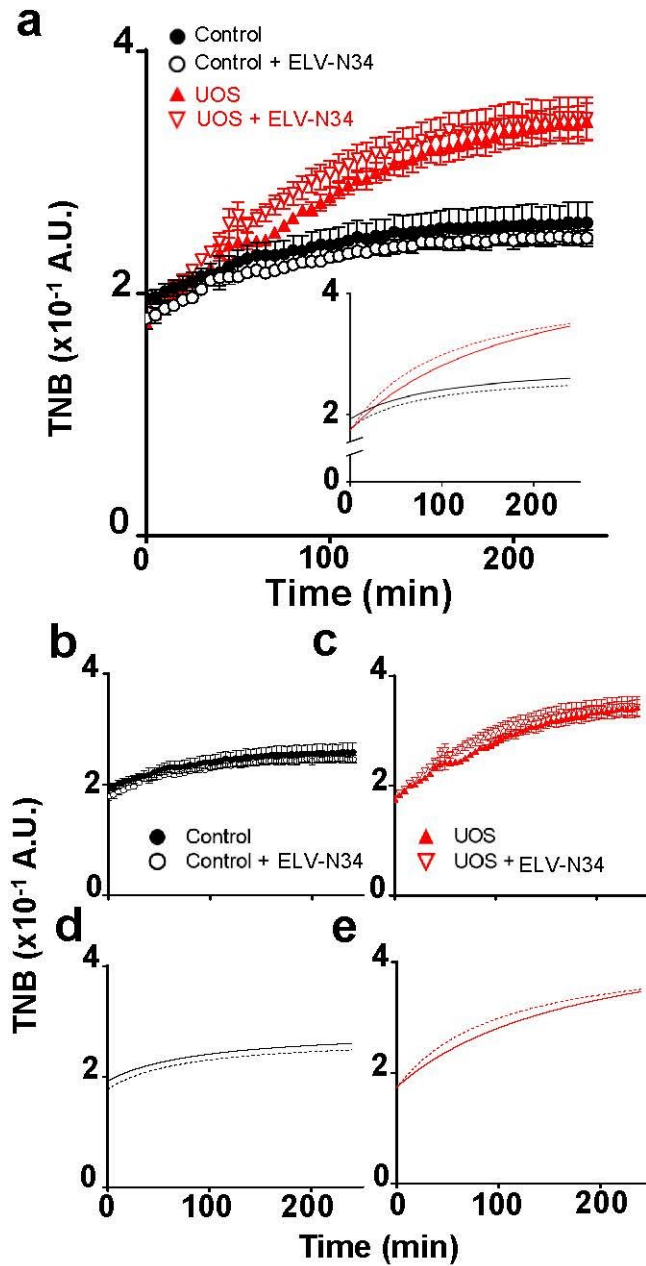


Fig. S10. Curves of Product vs. Time of the cytosolic TXNRD1. a-e, 200 nM ELV-N34 was added to the ABC cells exposed to UOS or naïve. The activity of TXNRD1 was recorded on the cytoplasmic fraction of the cells exposed to the mentioned treatments as product (absorbance at 412 nm) vs. Time (min). Spureus reductase activity was ruled out using a specific inhibitor of

TXNRD1. **a**, Merge curves of the four conditions. **b**, **c**, Mean of three observations and the bars represent the standard error of the mean for Control and Control + ELV-N34 (**b**) and UOS and UOS + ELV-N34 (**c**). **d**, **e**, The non-linear regression curves obtained for the data points in **b** and **c**.

Nuclear TXNRD1

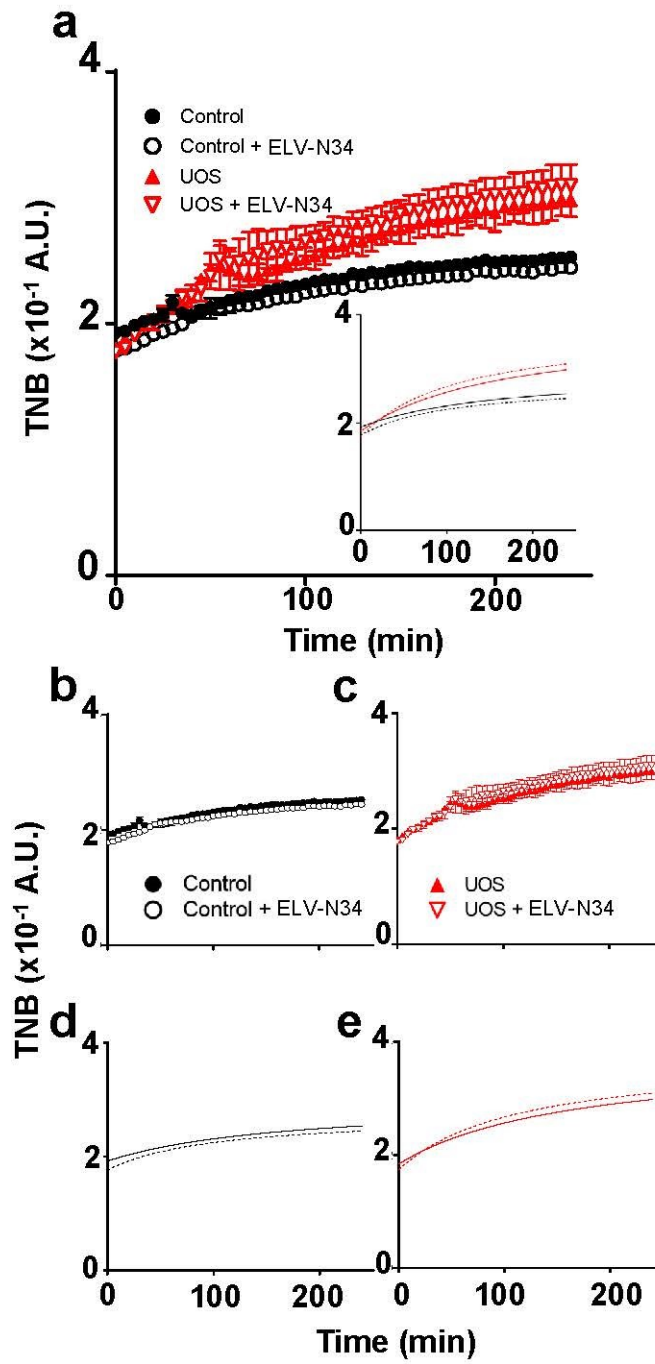


Fig. S11. Curves of Product vs. Time of the nuclear TXNRD1. a-e, 200 nM ELV-N34 was added to the ABC cells exposed to UOS or naïve. The activity of TXNRD1 was recorded on the

nuclear fraction of the cells exposed to the mentioned treatments as Product (absorbance at 412 nm) vs. Time (min). Spureus reductase activity was ruled out using a specific inhibitor of TXNRD1. **a**, Merge curves of the four conditions. **b**, **c**, Mean of three observations and the bars represent the standard error of the mean for Control and Control + ELV-N34 (**b**) and UOS and UOS + ELV-N34 (**c**). **d**, **e**, The non-linear regression curves obtained for the data points in **b** and **c**.

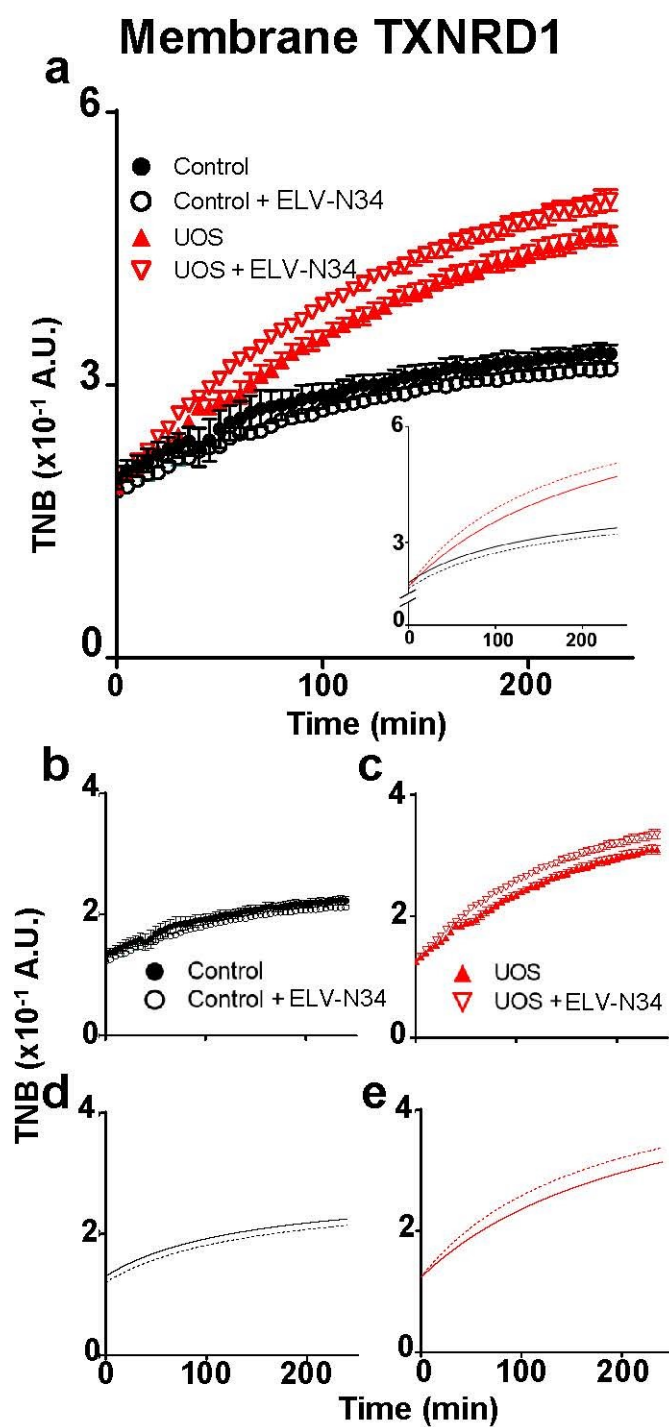


Fig. S12. Curves of Product vs. Time of the membrane TXNRD1. a-e, 200 nM ELV-N34 was added to the ABC cells exposed to UOS or naïve. The activity of TXNRD1 was recorded on the

membrane fraction of the cells exposed to the mentioned treatments as Product (absorbance at 412 nm) vs. Time (min). Spureus reductase activity was ruled out using a specific inhibitor of TXNRD1. **a**, Merge curves of the four conditions. **b**, **c**, Mean of three observations and the bars represent the standard error of the mean for Control and Control + ELV-N34 (**b**) and UOS and UOS + ELV-N34 (**c**). **d**, **e**, The non-linear regression curves obtained for the data points in **b** and **c**.

Crude extract TXNRD1

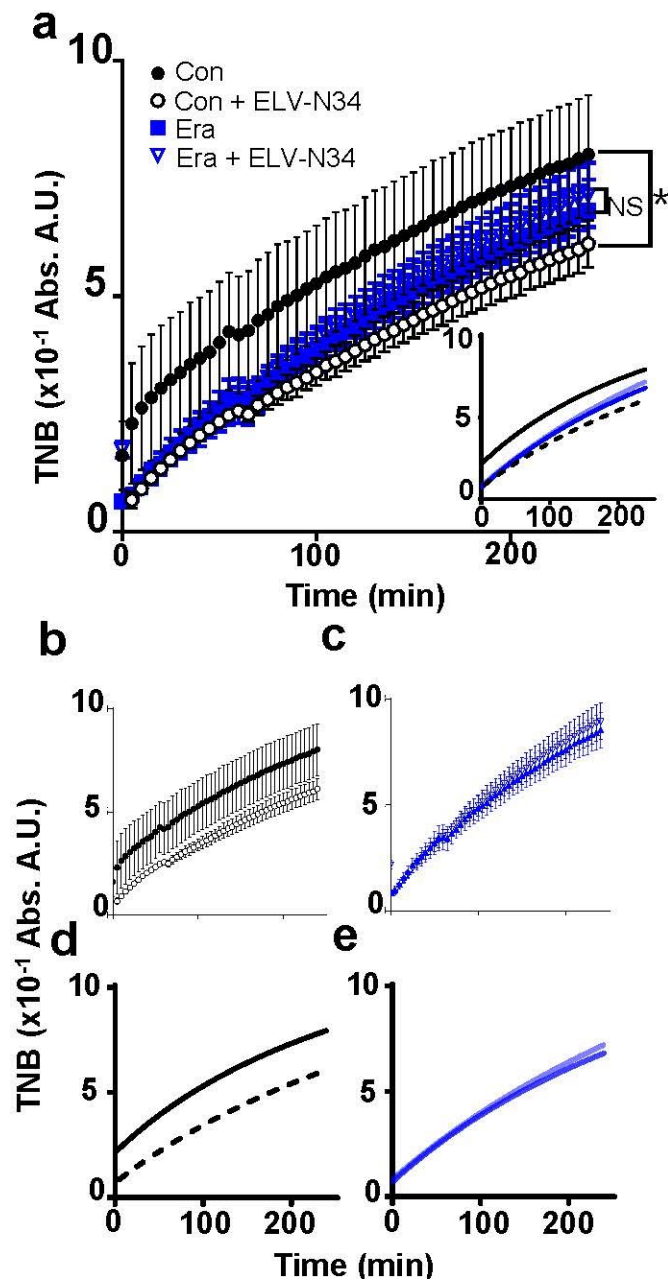


Fig. S13. Curves of Product vs. Time of the crude extract. a-e, 200 nM ELV-N34 was added to the ABC cells exposed to Erastin or naïve. The activity of TXNRD1 was recorded on the total extract of the cells exposed to the mentioned treatments as Product (absorbance at 412 nm) vs.

Time (min). Spureus reductase activity was ruled out using a specific inhibitor of TXNRD1. **a**, Merge curves of the four conditions. **b**, **c**, Mean of three observations and the bars represent the standard error of the mean for Control and Control + ELV-N34 (**b**) and UOS and UOS + ELV-N34 (**c**). **d**, **e**, The non-linear regression curves obtained for the data points in **b** and **c**.

Cytoplasmic TXNRD1

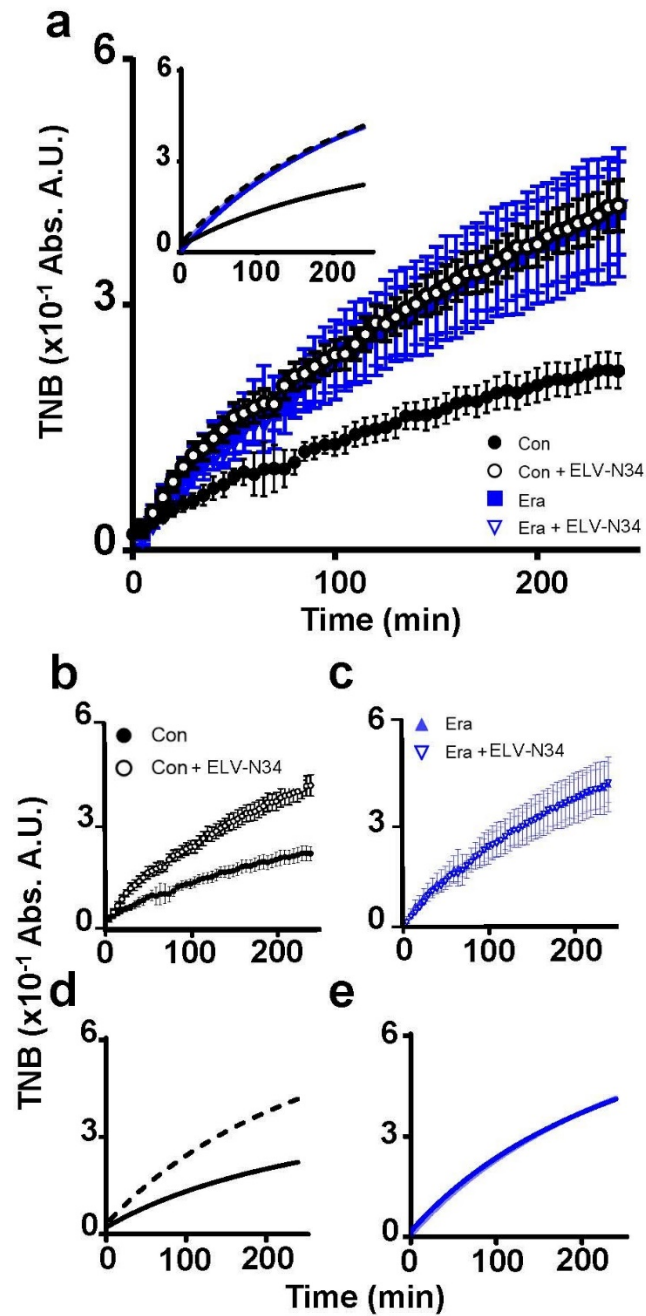


Fig. S14. Curves of Product vs. Time of the cytosolic TXNRD1. a-e, 200 nM ELV-N34 was added to the ABC cells exposed to Erastin or naïve. The activity of TXNRD1 was recorded on the cytoplasmic fraction of the cells exposed to the mentioned treatments as Product (absorbance

at 412 nm) vs. Time (min). Spurious reductase activity was ruled out using a specific inhibitor of TXNRD1. **a**, Merge curves of the four conditions. **b**, **c**, Mean of three observations and the bars represent the standard error of the mean for Control and Control + ELV-N34 (**b**) and UOS and UOS + ELV-N34 (**c**). **d**, **e**, The non-linear regression curves obtained for the data points in **b** and **c**.

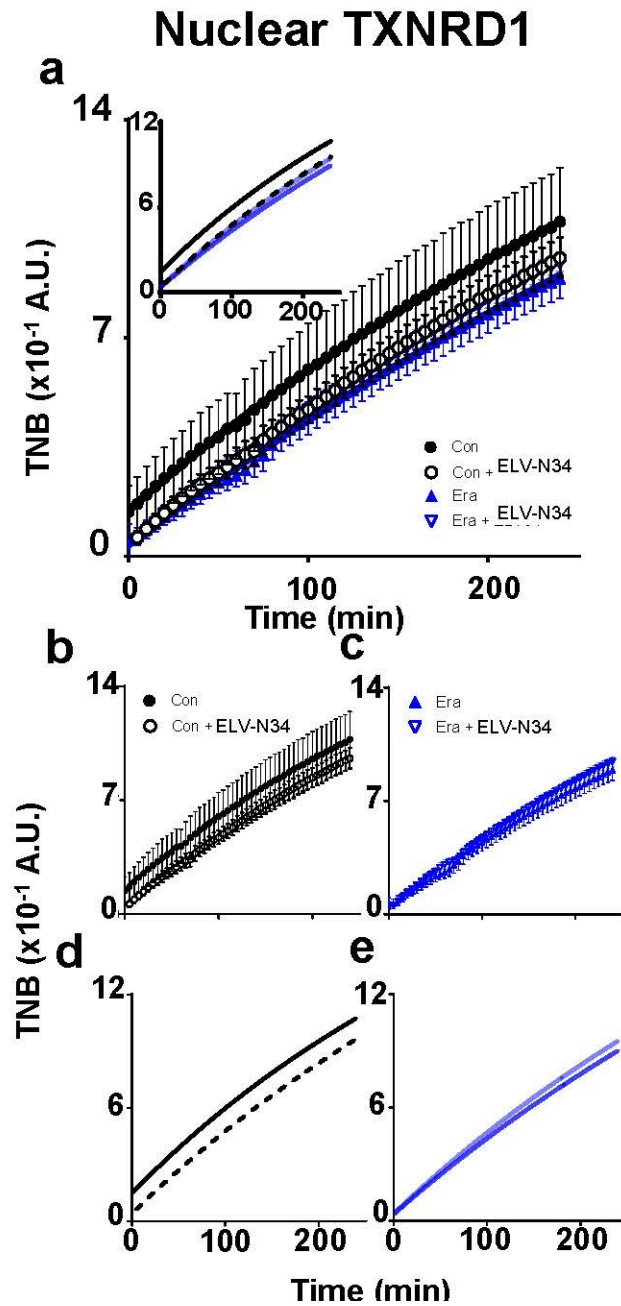


Fig. S15. Curves of Product vs. Time of the nuclear TXNRD1. a-e, 200 nM ELV-N34 was added to the ABC cells exposed to Erastin or naïve. The activity of TXNRD1 was recorded on the nuclear fraction of the cells exposed to the mentioned treatments as Product (absorbance at 412 nm) vs. Time (min). Spureus reductase activity was ruled out using a specific inhibitor of

TXNRD1. **a**, Merge curves of the four conditions. **b**, **c**, Mean of three observations and the bars represent the standard error of the mean for Control and Control + ELV-N34 (**b**) and UOS and UOS + ELV-N34 (**c**). **d**, **e**, The non-linear regression curves obtained for the data points in **b** and **c**.

Membrane TXNRD1

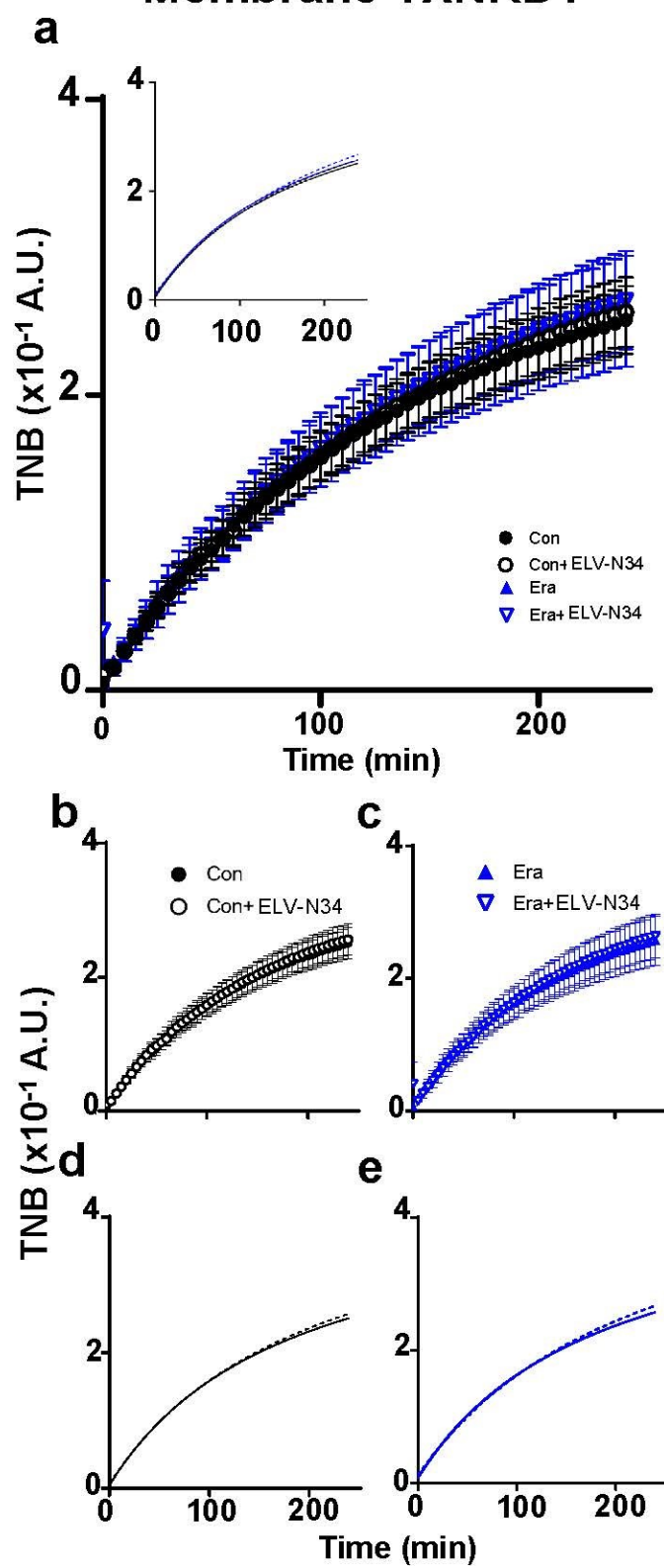


Fig. S16. Curves of Product vs. Time of the membrane TXNRD1. **a-e**, 200 nM ELV-N34 was added to the ABC cells exposed to Erastin or naïve. The activity of TXNRD1 was recorded on the membrane fraction of the cells exposed to the mentioned treatments as Product (absorbance at 412 nm) vs. Time (min). Spureus reductase activity was ruled out using a specific inhibitor of TXNRD1. **a**, Merge curves of the four conditions. **b, c**, Mean of three observations and the bars represent the standard error of the mean for Control and Control + ELV-N34 (**b**) and UOS and UOS + ELV-N34 (**c**). **d, e**, The non-linear regression curves obtained for the data points in b&c.

1 MGCAEGKAVA AAPTELQTK GKNGDGRRRS AKDHHPGKTL PENPAGFTST ATADSRALLQ

61 AYIDGHSVVI FSRSTCTRCT EVKKLFKSLC VPYFVLELDQ TEDGRALEGT LSELAETDL
AAETDL*
AAETDL*

121 PVVFEVKQRKI GGHGPTLKAY QEGRLQKLLK MNGPEDLPKS YDYDLIIIGG GSGGLAAAKE
DYDLIIIGG GSGGLAAK
1
PVVFEVK* S YDYDLIIIGG GSGGLAA
1
PVVFEVK* S YDYDLIIIGG GSGGLAA E
1

181 AAQYGKKVMV LDFVTPTPLG TRWGLGGTCV NVGCIPKKLM HQAALLGQAL QDSRNYGWKV
VMV LDFVTPTPLG TRWGLGGTCV NVGCIPKKLM HQAALLGQAL QDSR
2 3 4 5
KVMV LDFVTPTPLG TRWGLGGTCV NVGCIPK LM HQAALLGQAL QDSR
2 3 4
AAQYGKK AALLGQAL QDSR
2 3
KVMV LDFVTPTPLG TRWGLGGTCV NVGCIPK
1 2

241 EETVKHDWDR MIEAVQNHIG SLNWGYRVAL REKKVYENA YGQFIGPHRI KATNNKGKEK
A YGQFIGPHR
5

301 IYSAERFLIA TGERPRYLG PGDKEYCISS DDLFSLPYCP GKTIVVGASY VALECAGFLA
RFLIA TGERPR
4

361 GIGLDVTVMV RSILLRGFDQ DMANKIGEEM EEHGIKFIRQ FVPIKVEQIE AGTPGRLRVV
IGEEM EEHGIK
6
IGEEM EEHGIK
3

421 AQTSTNSEEII EGEYNIVMLA IGRDACTRKI GLETGVVKIN EKTGKIPVTD EEQTNVPYIY

481 AIGDILEDKV ELTPVAIQAG RLLAQRLYAG STVKCDYENV PTTVFTPLEY GACGLSEEKA
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6 7 8
AIGDILEDKV ELTPVAIQAG R CDYENV PTTVFTPLEY GACGLSEEK
7 8
AIGDILEDKV ELTPVAIQAG R RLYAG STVKCDYENV PTTVFTPLEY GACGLSEEK
5 6 7

541 VEKFGREENIE VYHSYFWPLE WTIPSRDNNK CYAKIICNTK DNERVVGFBV LGPNAGEVTQ
FGEENIE VYHSYFWPLE WTIPSR
8 9
VVGFBV LGPNAGEVTQ
9
HV LGPNAGEVTQ
4

601 GFAAALKCGL TKKQLDSTIG IHPVCAEVFT TLSVTKRSGA SILQAGCUGG
GFAAALK QLDSTIG IHPVCAEVF
9 10
KQLDSTIG IHPVCAEVF
9
GFAAALK
4

Fig. S17. Peptides obtained from the LiP study performed with ABC cells undergoing UOS (Fig. 1 and Table S2, green), Erastin (Fig. 2 and Table S4, blue), UOS (Fig. 2 and Table S3, red), and ELV-N34; UOS + NPD1 (purple).