

Supplemental Information

***In vivo* hyperphosphorylation of tau is associated with synaptic loss and behavioral abnormalities in the absence of tau seeds**

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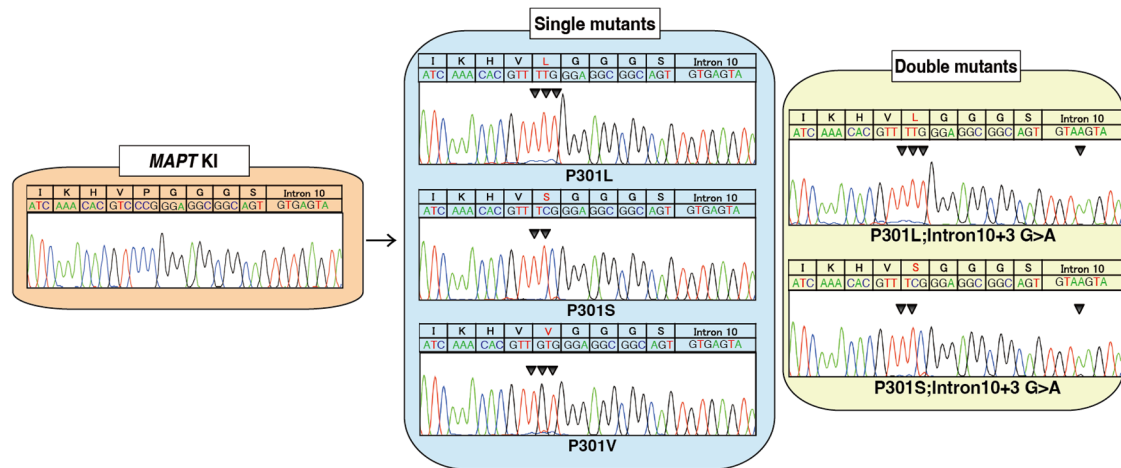
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Supplemental Figure 1. Sanger sequencing of other mutant *MAPT* KI mice. Sanger sequencing results determined *MAPT*^{P301L} KI, *MAPT*^{P301S} KI, *MAPT*^{P301V} KI, *MAPT*^{P301L; Int10+3} KI, and *MAPT*^{P301S; Int10+3} KI mice. Mutation loci are indicated by arrowheads in black and the substituted amino acids are highlighted in red.

a

Svbp

Founder

MAPT KI

MAPT^{Int10+3} KI

MAPT^{S305N; Int10+3} KI

F4



b

Togaram1

Founder

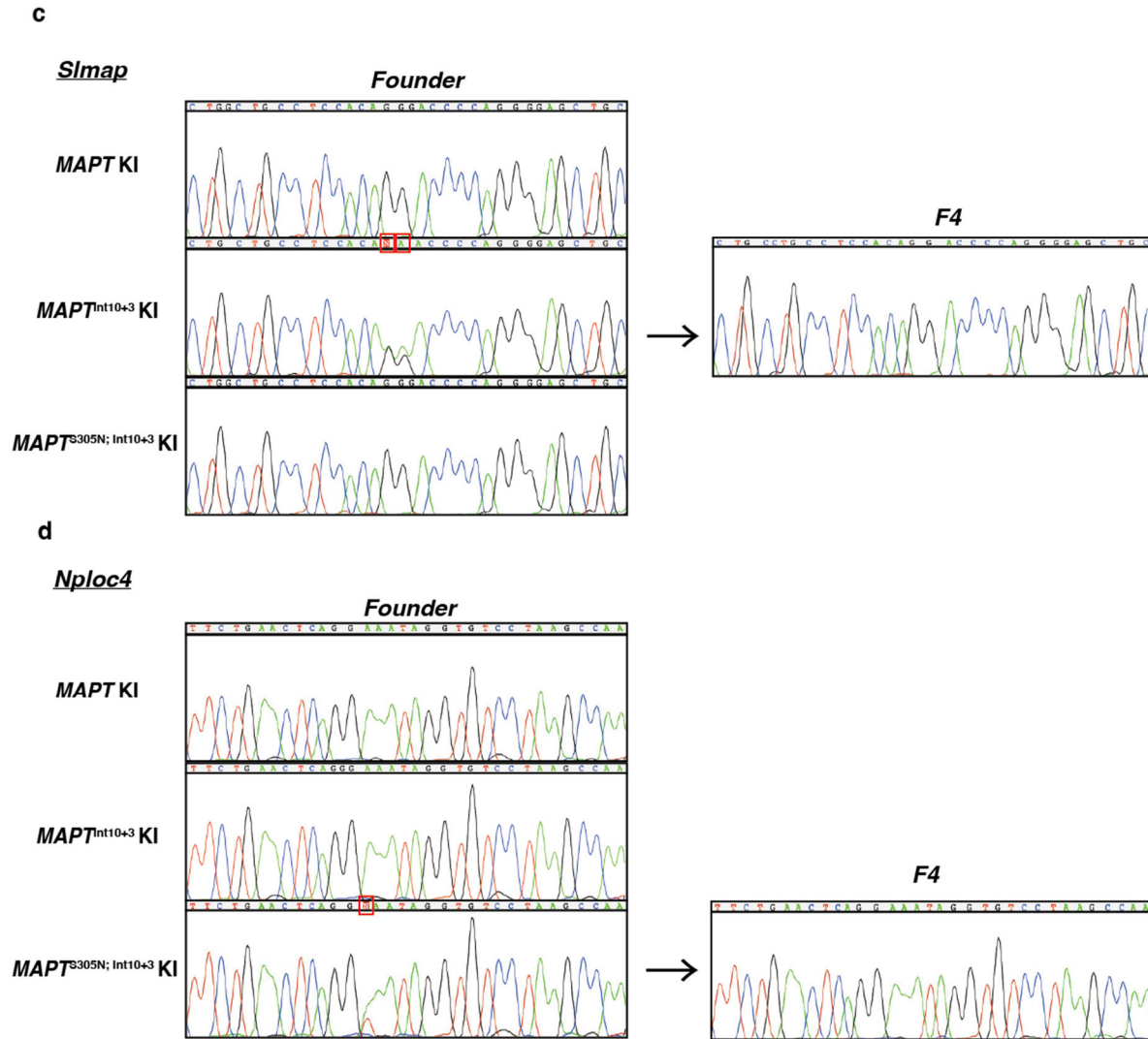
MAPT KI

MAPT^{Int10+3} KI

MAPT^{S305N; Int10+3} KI

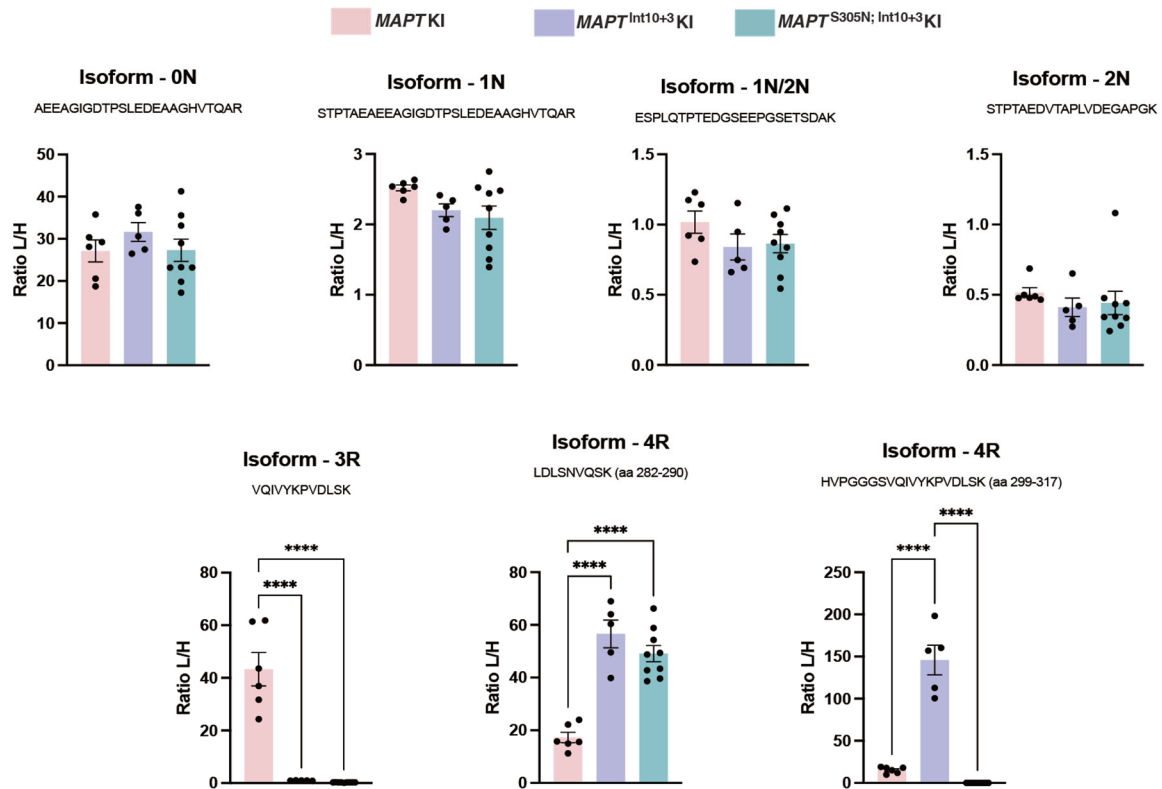
F4





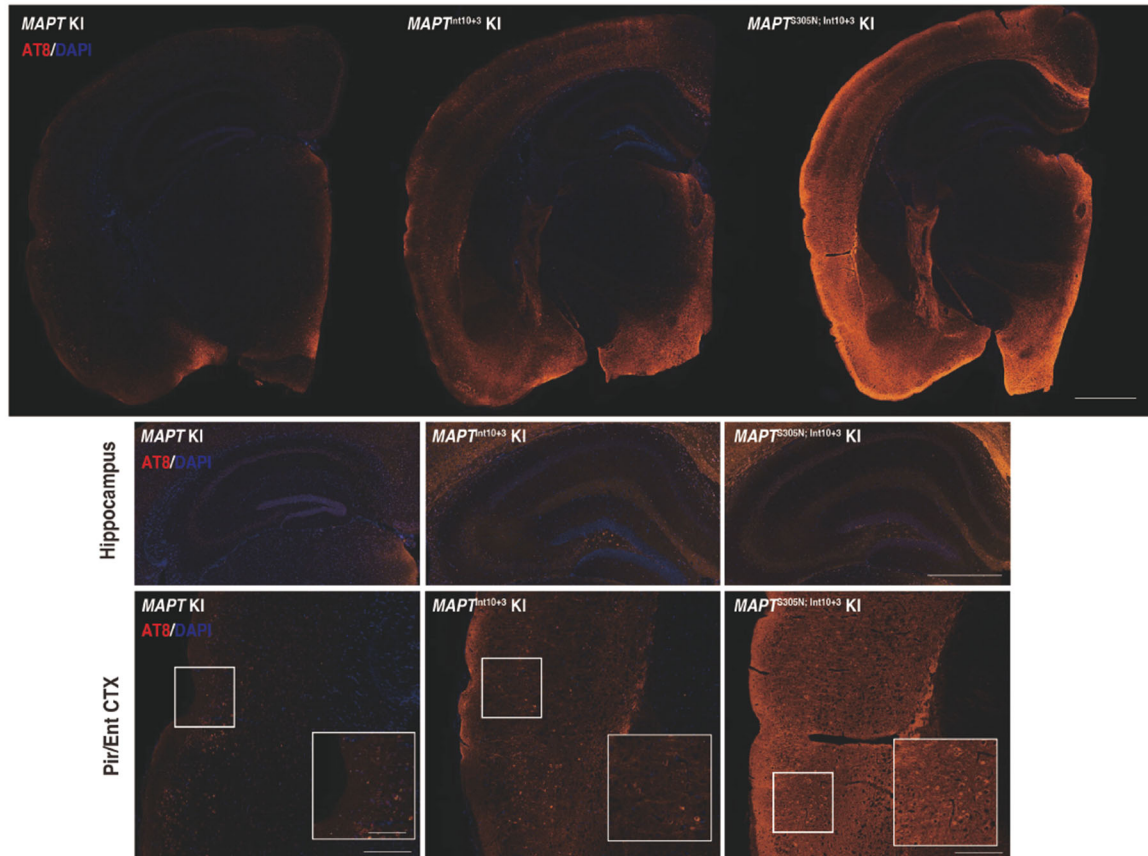
Supplemental Figure 2. Removal of off-target mutations in *MAPT*^{Int10+3} KI and *MAPT*^{S305N; Int10+3} KI mice by backcrossing with C57B6/J mice.

a-d, Off-target mutations identified in founders (highlighted in red boxes) were removed by backcrossing with C57B6/J mice more than four times.



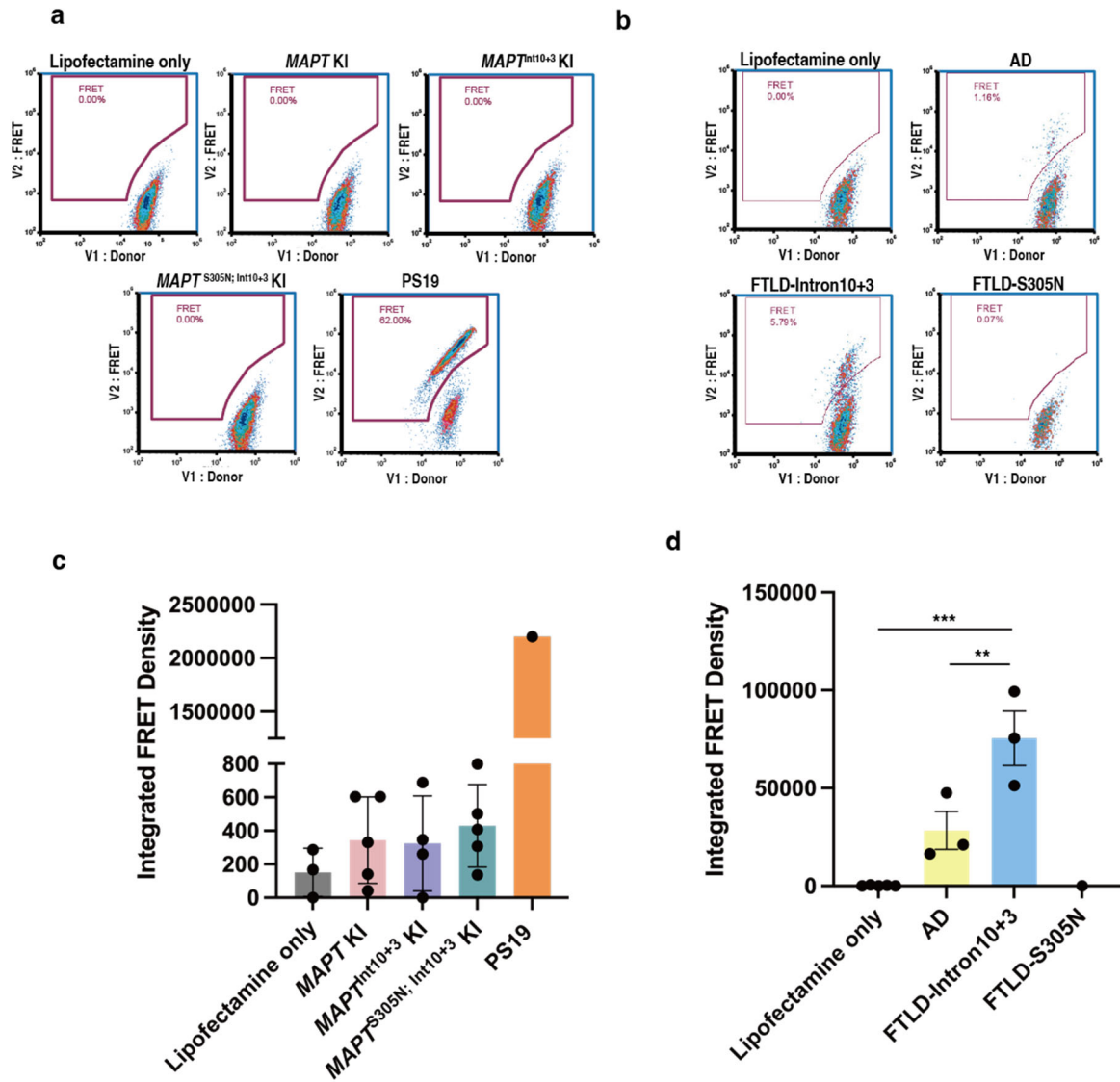
Supplemental Figure 3. LC-MS/MS analysis of tau in *MAPTKI*, *MAPT^{Int10+3} KI* and *MAPT^{S305N; Int10+3} KI* mice.

Amount of tau peptides based on the isoform resulting from alternative splicing by LC-MS/MS analysis in the brains of 12-15-month-old *MAPTKI*, *MAPT^{Int10+3} KI*, and *MAPT^{S305N; Int10+3} KI* mice. 4R-tau (299-317 aa) in *MAPT^{S305N; Int10+3} KI* line was not detected because serine (S) at position 305 was converted to asparagine (N). Data represents the mean $\pm$ SEM. **** $P < 0.0001$ (One-way ANOVA followed by Holm-Sidak's multiple comparison test).



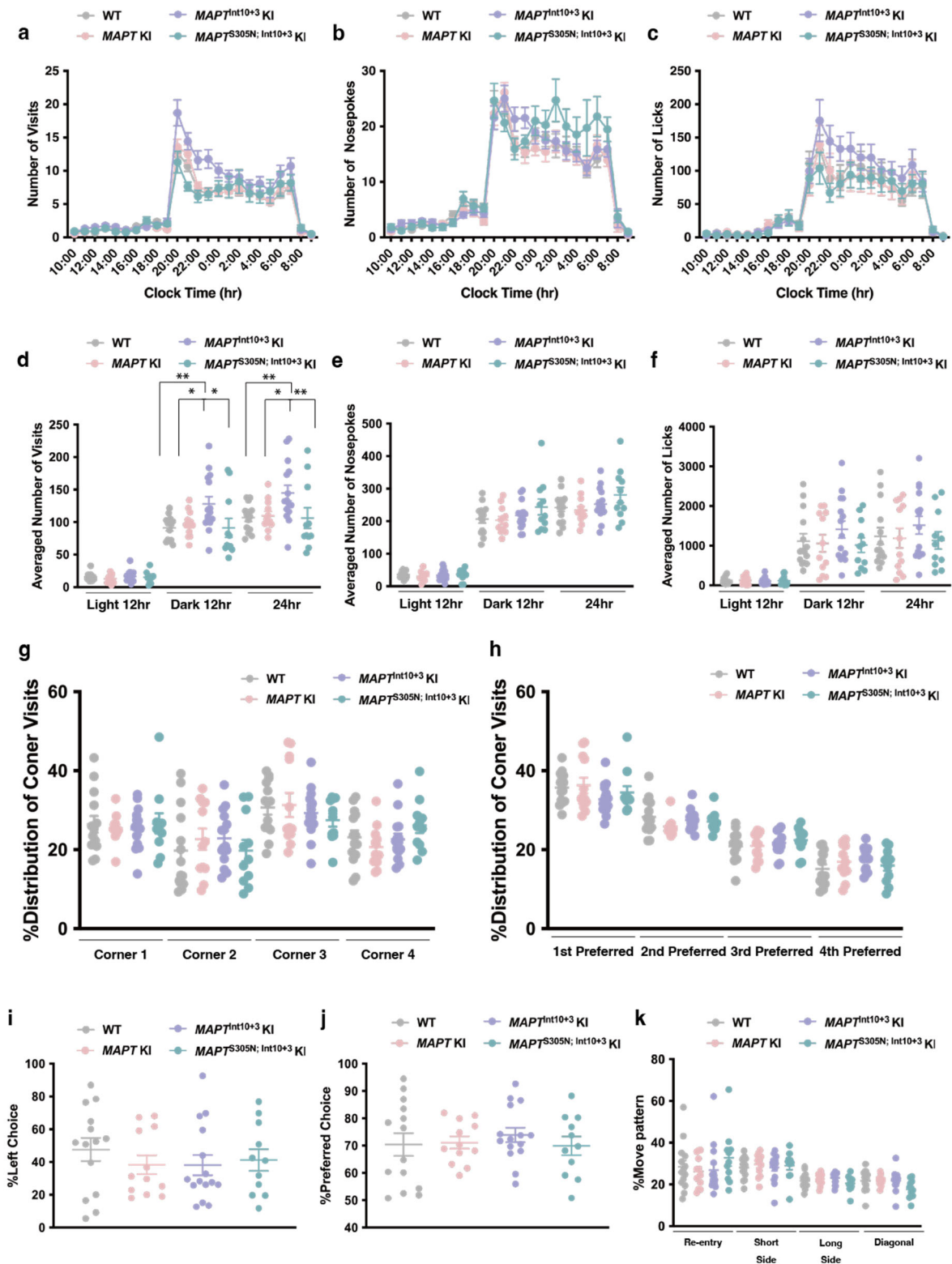
Supplemental Figure 4. Immunostaining of phospho-tau detected by AT8 antibody in *MAPT* KI, *MAPT*^{Int10+3} KI and *MAPT*^{S305N; Int10+3} KI mice.

Immunostaining of phosphorylated tau detected by AT8 antibody in the brains of 15-month-old *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice. Scale bar represents 1 mm in upper panel (hemibrain), 500 μ m in the middle panel (hippocampus), 100 μ m in the lower panel (piriform/entorhinal (Pir/Ent) cortex) and 50 μ m in the insert.



Supplemental Figure 5. Seeding assays of tau with P301S biosensor cell line.

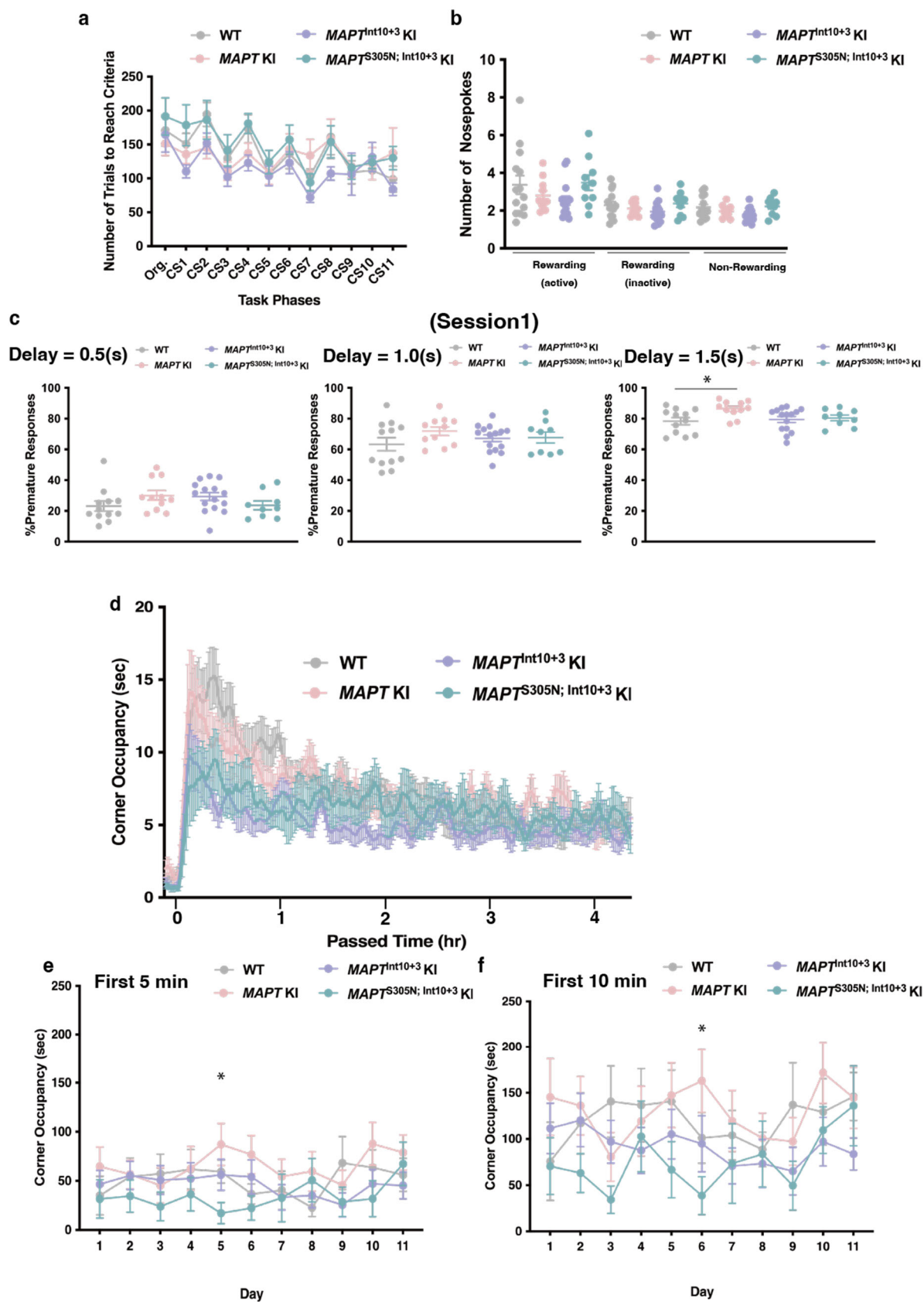
a, FRET *versus* donor (CFP) bivariate plots showing FRET-positive and FRET-negative P301S biosensor cells incubated with brain lysates from *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice at 15 months and from PS19 mice at 9 months. **b**, FRET *versus* donor (CFP) bivariate plots showing the FRET-positive and FRET-negative P301S biosensor cells incubated with extracts from FFPE-embedded AD and FTLD patients with Intron10+3 and S305N mutations in the *MAPT* gene. **c**, Integrated FRET density from brain lysates from *MAPT* KI, *MAPT*^{Int10+3} KI and *MAPT*^{S305N; Int10+3} KI mice at 15 months and PS19 mice at 9 months. **d**, Integrated FRET density from AD and FTLD patients with IVS10+3 and S305N mutations on the *MAPT* gene. Data represents the mean $\pm$ SEM. ** P <0.01, *** P <0.001 (one-way ANOVA with Tukey's multiple comparison test).



Supplemental Figure 6. Basal activity parameters calculated from 2 weeks of data in IntelliCage.

a-c, Circadian variations in number of visits (**a**), nosepokes (**b**), and licks (**c**) in WT, *MAPT* KI, *MAPT*^{Int10+3} KI and *MAPT*^{S305N; Int10+3} KI mice. **d-f**, Average number of visits (**d**), nosepokes (**e**), and licks (**f**) in a day in WT, *MAPT* KI, *MAPT*^{Int10+3} KI and *MAPT*^{S305N; Int10+3} KI mice.

Int10+3 KI mice. **g**, Percentages of corner visit distribution for each corner displayed by WT, *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice. **h**, Percentages of corner visit distribution in sequence of preferred corners displayed by WT, *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice. **i**, Percentages of left bottle choice exhibited by WT, *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice. **j**, Percentages of preferred choice exhibited by WT, *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice. **k**, Percentages of movement pattern (Re-entry, Short side, Long side, Diagonal) displayed by WT, *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice. All experiments were performed with 14-16-month-old female mice (WT: n = 14, *MAPT* KI: n = 12, *MAPT*^{Int10+3} KI: n = 15, *MAPT*^{S305N; Int10+3} KI: n = 11). Data represent the mean $\pm$ SEM. **P*<0.05, ***P*<0.01 (two-way ANOVA with Tukey's multiple comparison test).



Supplemental Figure 7. Results from SP-FLEX (behavioral sequence learning and behavioral flexibility task), behavioral inhibition and competitive dominance tests

a, b, Number of trials to reach criteria and nosepokes of 14-16-month old WT, *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice in the CS (complete shift)-only session of Self-paced Learning and Behavioral Flexibility Test (SP-FLEX) (WT: n = 14, *MAPT* KI: n = 11, *MAPT*^{Int10+3} KI: n = 15, *MAPT*^{S305N; Int10+3} KI: n = 10). **c**, Percentage of premature responses displayed by 14-16-month-old WT, *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice in Session 1 of the behavioral inhibition test (WT: n = 12, *MAPT* KI: n = 11, *MAPT*^{Int10+3} KI: n = 15, *MAPT*^{S305N; Int10+3} KI: n = 9). **d**, Average corner occupancy time displayed by WT, *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice at 0-4h over 11 days. (WT: n = 10, *MAPT* KI: n = 9, *MAPT*^{Int10+3} KI: n = 14, *MAPT*^{S305N; Int10+3} KI: n = 9) **e**, Average corner occupancy time displayed by WT, *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice in the first 5 min over 11 days. **f**, Average corner occupancy time displayed by WT, *MAPT* KI, *MAPT*^{Int10+3} KI, and *MAPT*^{S305N; Int10+3} KI mice in the first 10 min over 11 days. In (**c**), (**e**), and (**f**), data represent the mean $\pm$ SEM. **P*<0.05 (two-way ANOVA with Tukey's multiple comparison test).

Supplemental Tables

Mutations	Substitution Efficiency (%)
No substitution	7/166 (4.2)
Indel	10/166 (6.0)
P301L	79/166 (47.6)
P301S	19/166 (11.4)
P301V	1/166 (0.6)
Int10+3 G>A	36/166 (21.7)
P301L; Int10+3 G>A	9/166 (5.4)
P301S; Int10+3 G>A	4/166 (2.4)
S305N; Int10+3 G>A	1/166 (0.6)

Supplemental Table 1. Summary of substitution efficiency and generation of tau mutants after BE microinjection targeting *MAPT*-P301 and *MAPT*-Intron10+3.

Off-target candidate	Query type	Mismatch	Chr Position	Gene	Annotation	Off-target	
						<i>MAPT</i> KI ^{Intn10+3 G>A}	<i>MAPT</i> KI ^{S305N; Intn10+3 G>}
<i>MAPT</i> -P301-1	No indel	3	Chr11:70351826-70351849	<i>Alox15</i>	Intron	-	-
<i>MAPT</i> -P301-2	No indel	3	Chr4:119200276-119200299	<i>Svbp</i>	Intron	○	○
<i>MAPT</i> -P301-3	No indel	3	Chr9:64060473-64060496		Intergenic	-	-
<i>MAPT</i> -P301-4	No indel	3	Chr6:95725509-95725532		Intergenic	-	-
<i>MAPT</i> -P301-5	No indel	3	Chr12:64966002-64966025	<i>Togaram1</i>	Exon	○	○
<i>MAPT</i> -P301-6	No indel	3	Chr14:78141609-78141632		Intergenic	-	-
<i>MAPT</i> -P301-7	No indel	3	Chr2:126701111-126701134		Intergenic	-	-
<i>MAPT</i> -P301-8	No indel	3	Chr12:72964402-72964425		Intergenic	-	-
<i>MAPT</i> -P301-9	No indel	3	Chr1:180813179-180813202	<i>H3f3a</i>	Intron	-	-
<i>MAPT</i> -P301-10	No indel	3	Chr6:14688630-14688653	<i>Ppfbp1</i>	Non-coding	-	-
<i>MAPT</i> -P301-11	Del 8	2	Chr9:121021200-121021222	<i>Ulk4</i>	Intron	-	-
<i>MAPT</i> -P301-12	Del 3	2	ChrX:87776462-87776484	<i>Il1rapl1</i>	Intron	-	-
<i>MAPT</i> -P301-13	Del 2	2	Chr16:18687059-18687081		Intergenic	-	-
<i>MAPT</i> -P301-14	Del PAM 2	2	Chr2:167736542-167736564		Intergenic	-	-
<i>MAPT</i> -P301-15	Ins 17	2	Chr10:118675703-118675727		Intergenic	-	-
<i>MAPT</i> -P301-16	Ins 16	2	Chr2:133528883-133528907		Intergenic	-	-
<i>MAPT</i> -P301-17	Ins 13	2	Chr14:26429735-26429759	<i>Simap</i>	Intron	○	-
<i>MAPT</i> -P301-18	Ins 7	2	Chr6:146527101-146527125		Intergenic	-	-
<i>MAPT</i> -P301-19	Ins 5	2	Chr2:131262710-131262734	<i>Pank2</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-1	No indel	3	Chr14:31275639-31275661	<i>Dnah1</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-2	No indel	3	Chr10:67285234-67285256	<i>Nrbf2</i>	Non-coding	-	-
<i>MAPT</i> -intron10+3 G>A-3	No indel	3	Chr3:108148064-108148086		Intergenic	-	-
<i>MAPT</i> -intron10+3 G>A-4	No indel	3	Chr5:137062275-137062297	<i>Serpine1</i>	Non-coding	-	-
<i>MAPT</i> -intron10+3 G>A-5	No indel	3	Chr6:72577777-72577799	<i>Elmod3</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-6	No indel	3	Chr5:64508703-64508725		Intergenic	-	-
<i>MAPT</i> -intron10+3 G>A-7	Del 18	2	Chr6:59413774-59413795	<i>Gprn3</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-8	Del 18	2	Chr7:96207591-96207612	<i>Tenn4</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-9	Del 17	2	Chr2:168700314-168700335	<i>Atp9a</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-10	Del 16	2	Chr2:146621362-146621383		Intergenic	-	-
<i>MAPT</i> -intron10+3 G>A-11	Del 16	2	Chr8:69915135-69915156	<i>Gata2a</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-12	Del 16	2	Chr11:99056068-99056089		Intergenic	-	-
<i>MAPT</i> -intron10+3 G>A-13	Del 15	2	Chr2:167736538-167736559		Intergenic	-	-
<i>MAPT</i> -intron10+3 G>A-14	Del 14	2	Chr1:82288807-82288828	<i>Irs1</i>	Exon	-	-
<i>MAPT</i> -intron10+3 G>A-15	Del 14	2	Chr4:150626514-150626535		Intergenic	-	-
<i>MAPT</i> -intron10+3 G>A-16	Del 14	2	Chr17:30516288-30516309	<i>Btbd9</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-17	Del 14	2	Chr8:112093736-112093757		Intergenic	-	-
<i>MAPT</i> -intron10+3 G>A-18	Del 14	2	Chr12:84648519-84648540	<i>Vrtn</i>	Exon	-	-
<i>MAPT</i> -intron10+3 G>A-19	Del 14	2	Chr11:87664708-87664729	<i>Rnf43</i>	Exon	-	-
<i>MAPT</i> -intron10+3 G>A-20	Del 13	2	Chr4:139161305-139161326	<i>Gm33304</i>	Non-coding	-	-
<i>MAPT</i> -intron10+3 G>A-21	Del 12	2	Chr18:80778335-80778356	<i>Atp9b</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-22	Del 12	2	Chr2:119770882-119770903	<i>Rpap1</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-23	Del 12	2	Chr6:109857978-109857999		Intergenic	-	-
<i>MAPT</i> -intron10+3 G>A-24	Del 12	2	Chr5:142468117-142468138	<i>Ap5z1</i>	Exon	-	-
<i>MAPT</i> -intron10+3 G>A-25	Del 11	2	Chr17:25386575-25386596	<i>Cacna1h</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-26	Del 11	2	Chr9:119555524-119555545	<i>Scn5a</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-27	Del 9	2	Chr9:43536132-43536153		Intergenic	-	-
<i>MAPT</i> -intron10+3 G>A-28	Del 9	2	Chr7:125974764-125974785	<i>Gsg1l</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-29	Del 6	2	Chr12:91965237-91965258		Intergenic	-	-
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<i>MAPT</i> -intron10+3 G>A-31	Del 4, or Del 5	2	Chr4:43394834-43394855	<i>Rusc2</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-32	Del 4, or Del 5	2	Chr5:121818513-121818534	<i>Sh2b3</i>	Exon	-	-
<i>MAPT</i> -intron10+3 G>A-33	Del 3	2	Chr12:21295846-21295867	<i>Cpsf3</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-34	Del 3	2	Chr9:113870747-113870768	<i>Clasp2</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-35	Del 3	2	Chr6:125633288-125633309	<i>Vwf</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-36	Del 3	2	Chr2:130840193-130840214	<i>930402H24R</i>	Non-coding	-	-
<i>MAPT</i> -intron10+3 G>A-37	Del 1, or Del 2	2	Chr17:49049099-49049120	<i>Lrtn2</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-38	Ins 17	2	Chr16:22558979-22559002	<i>Dgkg</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-39	Ins 16	2	Chr15:74631843-74631866	<i>Mroh4</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-40	Ins 15	2	Chr17:54077434-54077457		Intergenic	-	-
<i>MAPT</i> -intron10+3 G>A-41	Ins 13	2	Chr7:139648287-139648310	<i>Clap46</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-42	Ins 13	2	Chr2:30194586-30194609	<i>Kyat1</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-43	Ins 6	2	Chr8:71290281-71290304	<i>Myo9b</i>	Non-coding	-	-
<i>MAPT</i> -intron10+3 G>A-44	Ins 6	2	Chr5:138859529-138859552		Intergenic	-	-
<i>MAPT</i> -intron10+3 G>A-45	Ins 6	2	Chr16:93903368-93903391	<i>Chaf1b</i>	Intron	-	-
<i>MAPT</i> -intron10+3 G>A-46	Ins 1	2	Chr13:63111073-63111096	<i>Aoepe</i>	Intron	-	-

Supplemental Table 2. Summary of off-target candidate sites predicted by COSMID for *MAPT*^{Intn10+3} KI and *MAPT*^{S305N; Intn10+3} KI mice.

Sample \ antibody	AT8	CP13	PHF-1	AT180	AT270	TOC1	MC1
WT	-	-	-	-	-	-	-
<i>MAPT</i> KI	+	+	-	-	-	-	-
<i>MAPT</i> KI ^{Int10+3}	++	++	++	++	++	-	-
<i>MAPT</i> KI ^{S305N; Int10+3}	+++	+++	+++	++	++	-	-
FTLD-Int10+3	+++	+++	+++	+++	+	++	+
FTLD-S305N	+++	+++	+++	+++	+++	++	+

Supplemental Table 3. Summary of histological visual quantification analyses of tau phosphorylation detected by several phospho-tau antibodies.

The following antibodies were used at the dilutions described below.			
Antibody			Dilution
			WB IHC
CP-13	Kindly provided by Cristina D'Abramo		1:500 1:500
AT8	Inogenetics #90206 (anti-PHF-TAU) and ThermoFisher MN1020b		1:2000 1:500
PHF-1	Kindly provided by Cristina D'Abramo		1:2000 1:1000
AT180	Thermo #MN-1040		- 1:100
AT270	Thermo #1050		- 1:100
TOC1	Kindly provided by Lester I Binder		- 1:1500
MC1	Kindly provided by Peter Davis		- 1:500
Tau13	Santa Cruz #sc-21796		1:2000 -
Tau5	Thermo #AHB0042		1:2000 -
HT7	Thermo #MN-1000		1:2000 -
K9JA	Dako # A0024		1:10000 -
RD3	Merk Millipore #05-803		- 1:100
RD4	Merk Millipore #05-804		- 1:100
Synaptotagmin	Synaptic System 105-002		- 1:500
Homer	Synaptic System 160-011		- 1:500
VGLUT1	Synaptic System 135-303		- 1:500
β-actin	Sigma A5441		1:5000 -

Supplemental Table 4. Antibodies used for immunoblotting and immunohistochemical (IHC) analyses.

<i>In vitro</i> transcription for BE3	
T7-BE3-F	GCCGCTAATACGACTCACTATAG
T7-BE3-R	GCGGGTTAACTCAATGGT
<i>In vitro</i> transcription for sgRNAs	
sgRNA-MAPT-P301	taatacgactcactataGGTCCCGGAGGCGGCAGTGgttttagagctagaa
sgRNA-MAPT-Intron10+3 G>A	taatacgactcactataGGAATCAGCTGCGGCTCCgttttagagctagaa
CRISPRscan tail primer	AAAAGCACCGACTCGGTGCCACTTTTTCAAGTTGATAACGGACTAGCCTATTTTAACTTGCTATTCTA
RT-PCR for tau	
3R-Tau F	GTCCGTACTCCACCCAAGTC
3R-Tau R	TTTGTAAGTATTTGCACCTTC
4R-Tau F	GAAGCTGGATCTTAGCAACG
4R-Tau R	GACGTGTTTGATATTATCCT
Total Tau F	AGCCAAGACATCCACACGTT
Total Tau R	ATCAGAGGGTCTGAGCTACCA
G3PDH F	CCATGGCACCGTCAAGGCTGA
G3PDH R	GCCAGTAGAGGCAGGGATGAT
Sanger-sequencing for off-target sites	
Svbp F	GGGGGTCAGAGTGTTTTCAA
Svbp R	GGAGCTGGGGAGCTAAAAAG
Togaram1 F	AATGGTTCCCTCCAGCTCTC
Togaram1 R	TTTTTCTCGCCTCTCATGGT
Slmap F	CACTGGTTGTTGGAGCATCT
Slmap R	GCCAAACAAGACATTCAGCA
Nploc4 F	CAAAATGACCATCGGGAGAG
Nploc4 R	AGTCATCCTGCTGGCTCAAC

Supplemental Table 5. Primer lists for *in vitro* transcription of BEs and sgRNAs

Sample ID	Gender	Age	PMI	ApoE genotype	Braak Tau Stage	Region	Import source	Sample
P6/20	F	86	94.05	N/A	6	Frontal Cortex	from Q5BB	AD
P14/21	F	72	59.1	N/A	6	Frontal Cortex	from Q5BB	AD
P3/21	F	61	36.55	N/A	6	Frontal Cortex	from Q5BB	AD
HR09-5902	M	41.9	N/A	33	0	Frontal Cortex	from Mayo Clinic	FTLD-Intron10+3
HR10-25426	M	54.7	N/A	33	0	Frontal Cortex	from Mayo Clinic	FTLD-Intron10+3
HR10-64047	M	57	N/A	34	0	Frontal Cortex	from Mayo Clinic	FTLD-Intron10+3
H024	M	46	N/A	N/A	N/A	Frontal Cortex	from Iou National Hospital	FTLD-S305N

Supplemental Table 6. Human sample information