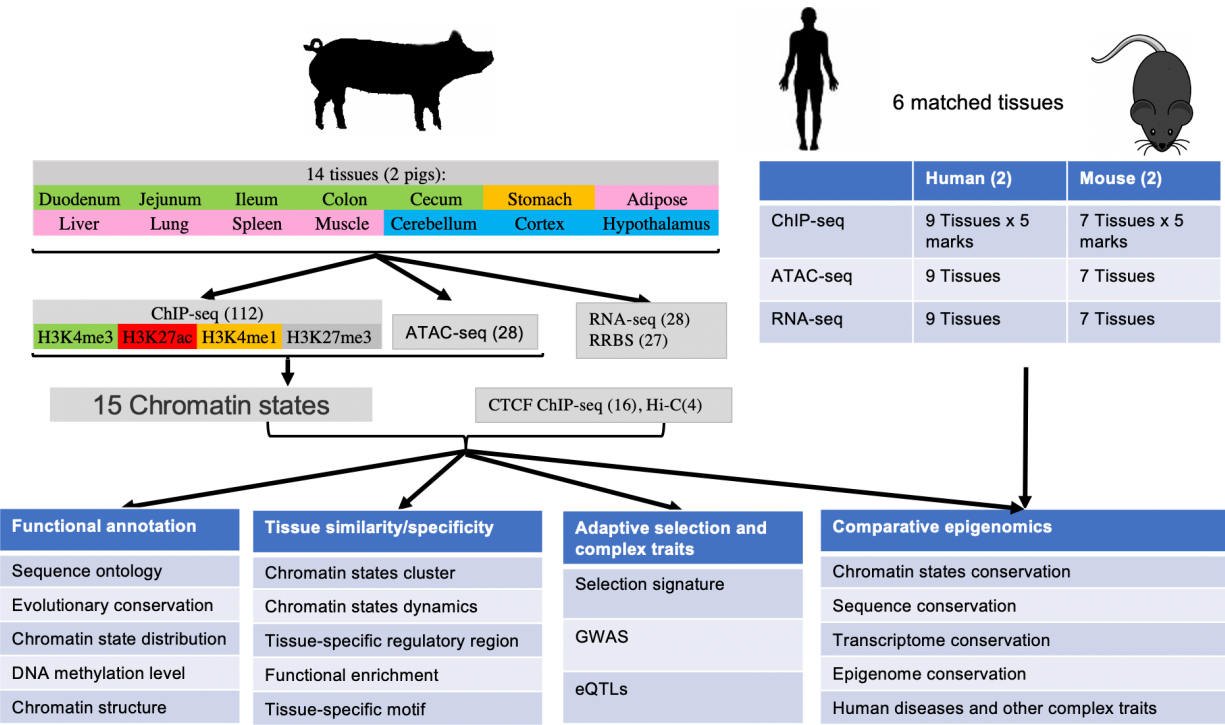
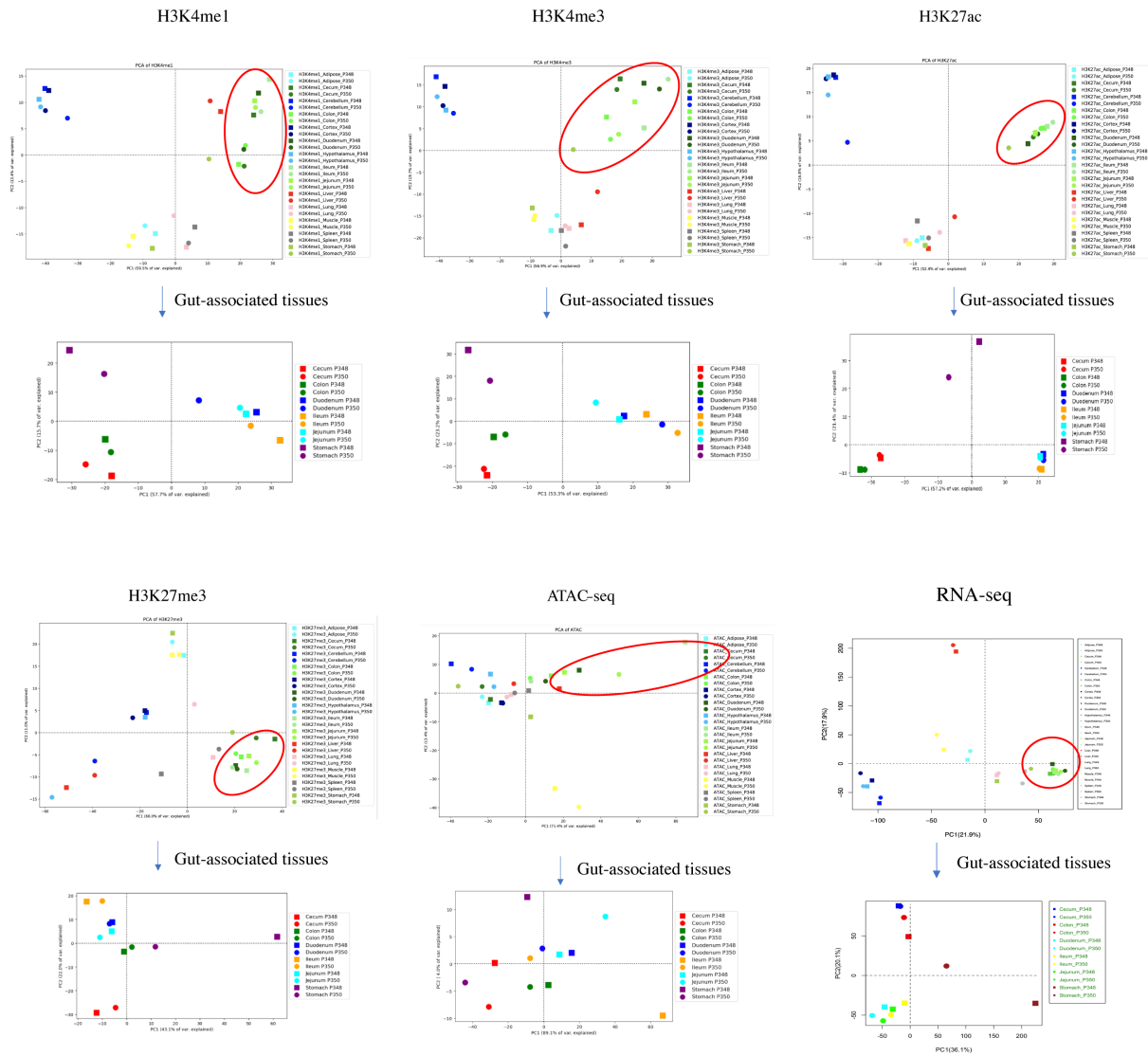
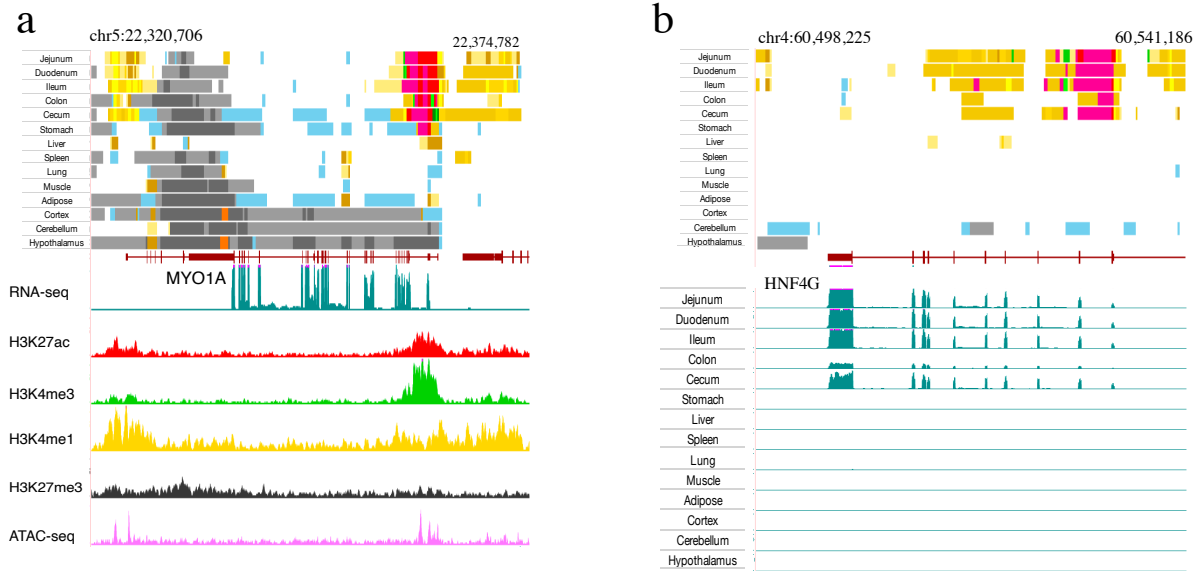




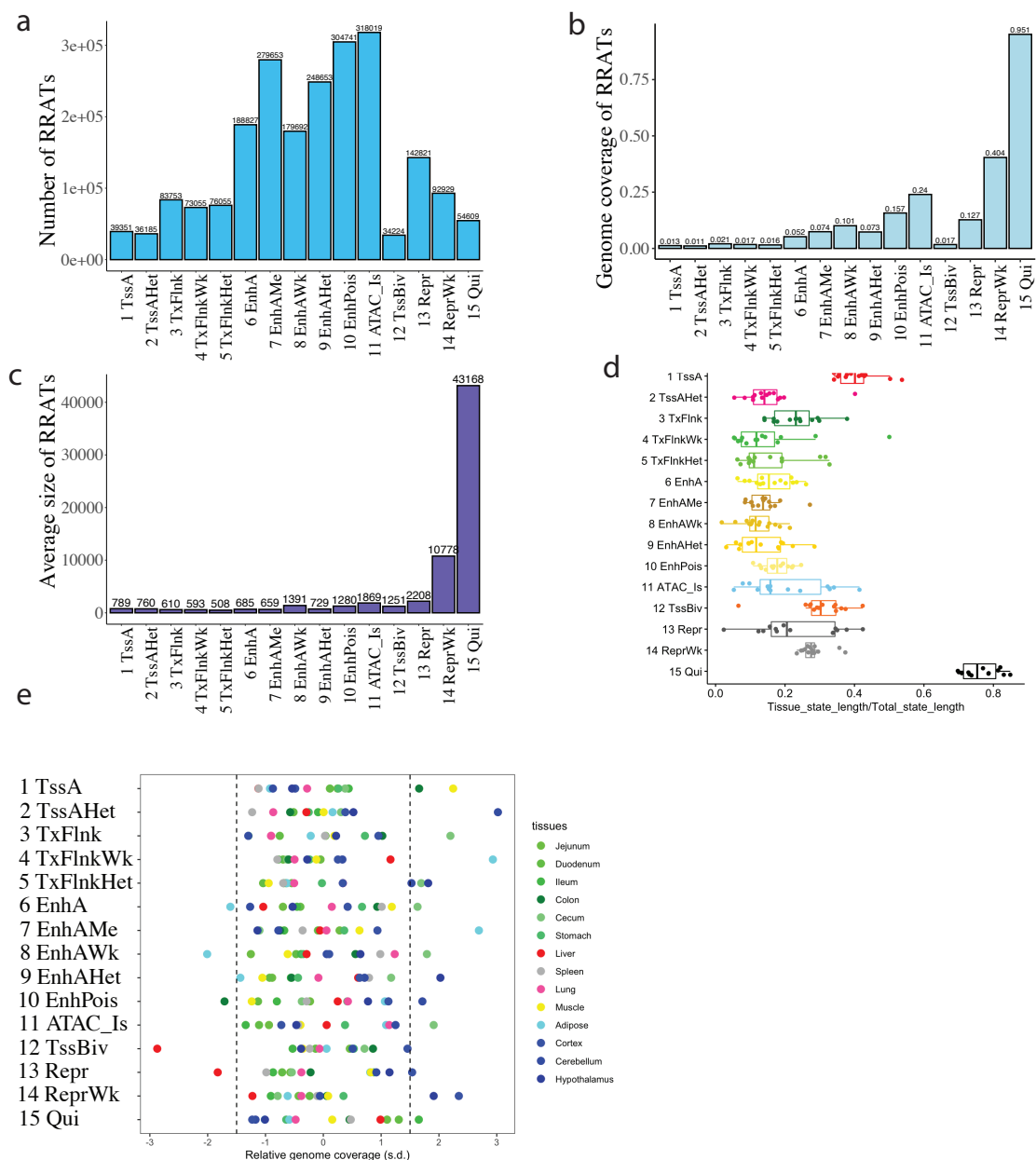
Supplementary Fig. 1 Study design of pig genome annotation and comparative epigenomic analysis. This study included 112 histone ChIP-seq, 28 ATAC-seq, 28 RNA-seq, 27 RRBS, 16 CTCF ChIP-seq, 4 Hi-C datasets cross 14 tissues from pig, and also integrated 126 human and 98 mouse datasets for comparative epigenomic analysis. The six matched tissues among pig, human, and mouse were small intestine, liver, spleen, lung, adipose, and brain cortex. The numbers in the brackets are the number of assays or samples included in the current study.



Supplementary Fig. 2 Principal component analysis (PCA) of five epi-marks and RNA-seq in 14 tissues in pig. The regions highlighted are gut-associated tissues.



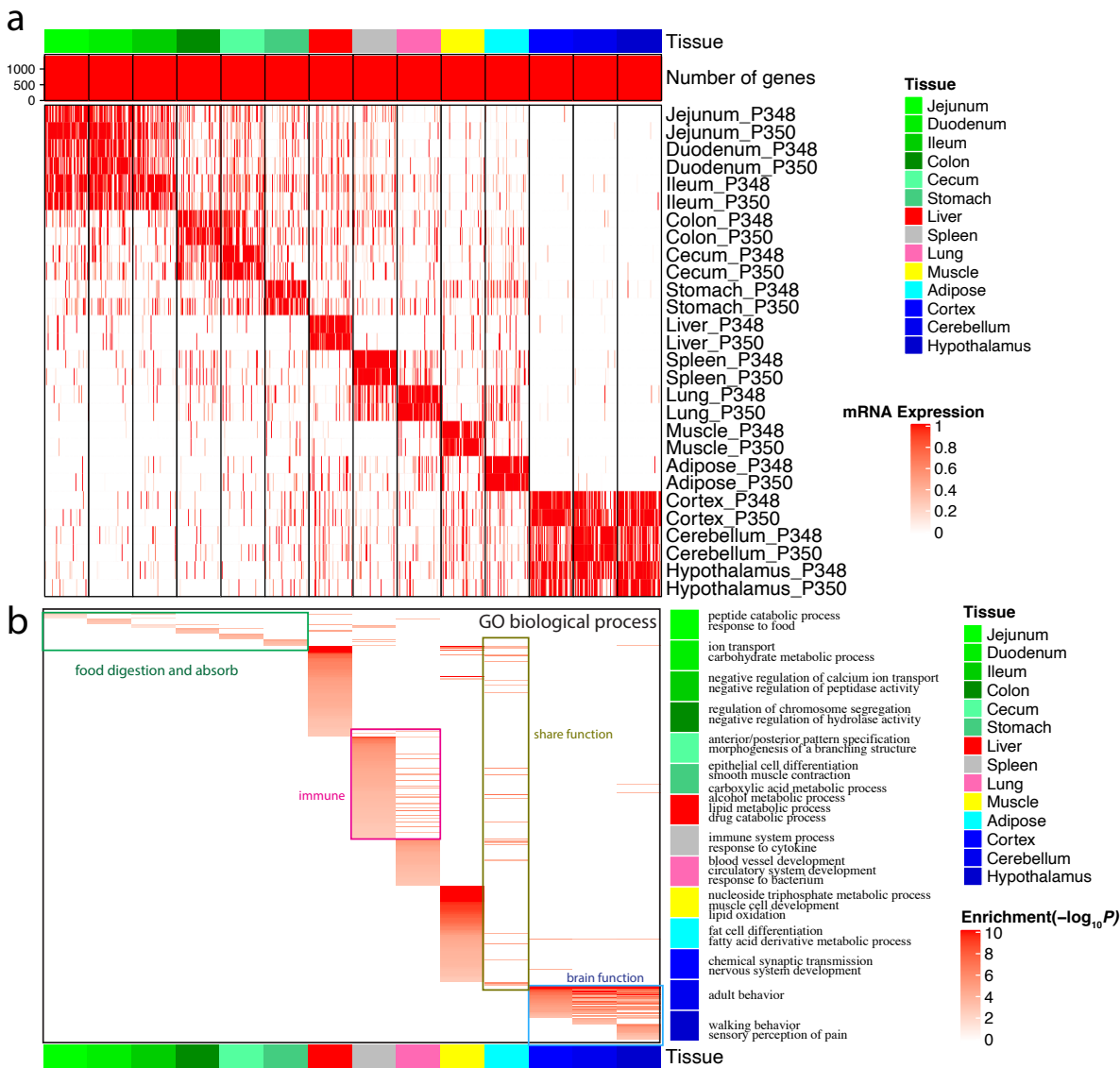
Supplementary Fig. 3 Example of regulation of gene expression by chromatin states. **a**, Chromatin state around *MYO1A* (chr5:22,320,706-22,374,782) in 14 tissues. **b**, Chromatin state around *HNF4G* (chr4:60,498,225-60,541,186) in 14 tissues. Vertical scale 0-200 for RNA-seq, 0-100 for H3K27ac and H3K4me3, and 0-50 for other marks and ATAC-seq.



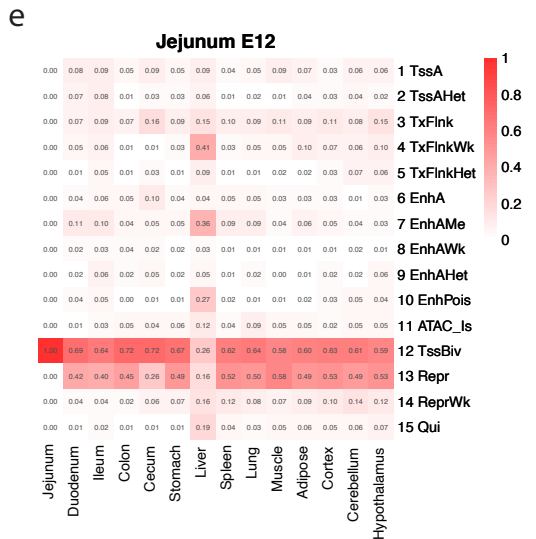
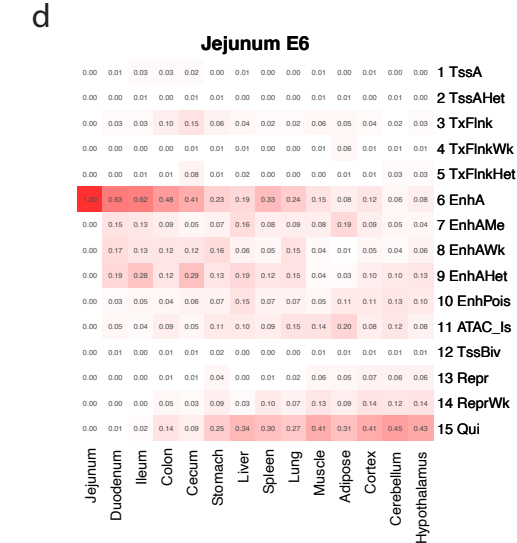
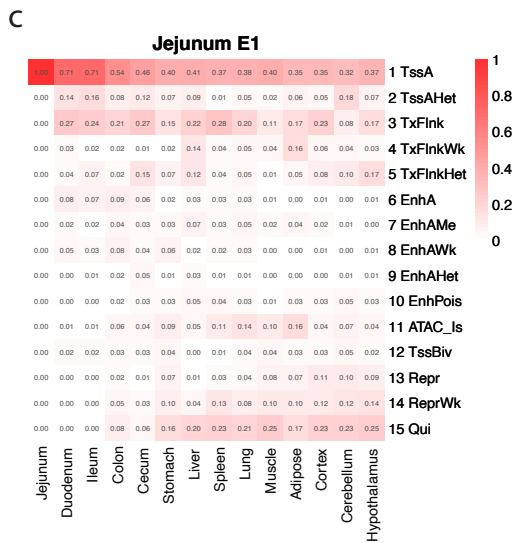
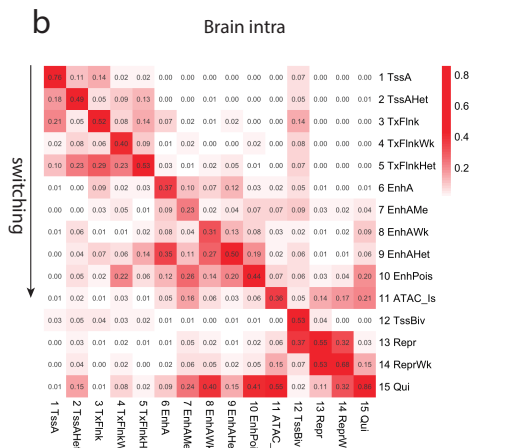
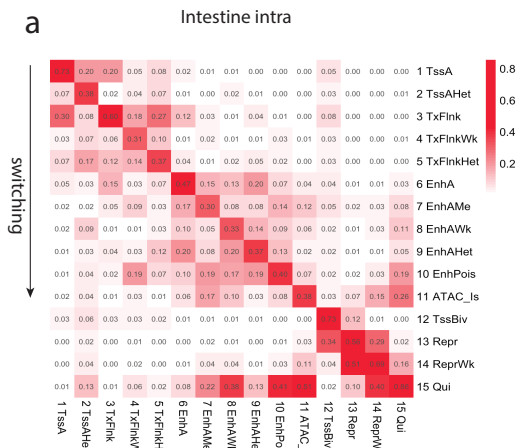
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37 **Supplementary Fig.4 Total number, genome coverage and characterizations of regulatory**
 38 **elements at each chromatin state across 14 tissues. a**, Total number of regulatory regions
 39 across 14 tissues (RRATs) for each chromatin state. Totally we identified 2,097,958 regulatory
 40 elements (exclude Qui) spanning 14 tissues including 39,351 active promoters (TssA), 188,827
 41 active strong enhancers (EnhA), and 142,821 repressors (Repr). **b**, Genome coverage of RRATs

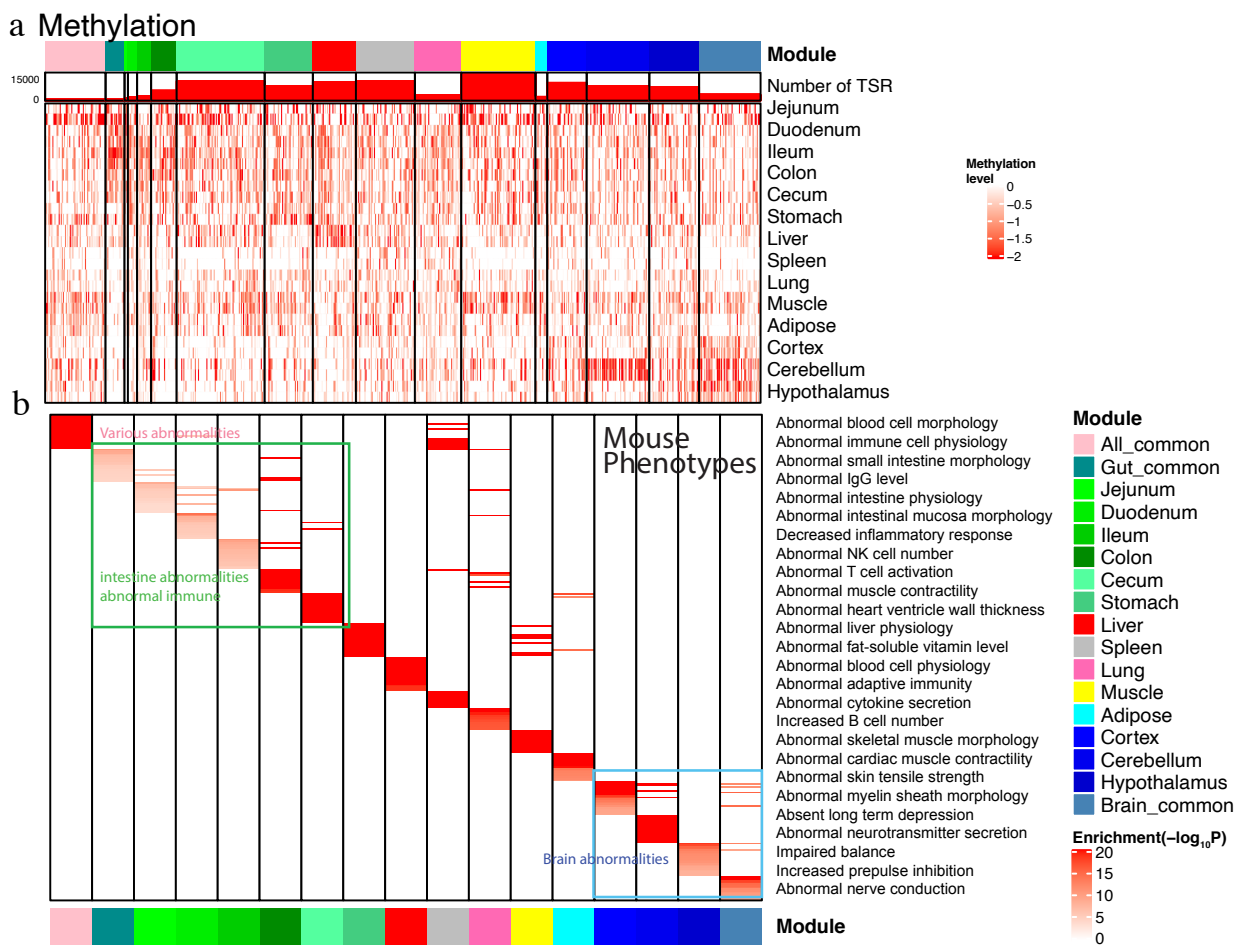
for each chromatin state. **c**, Average size of RRATs for each chromatin state. **d**, Average genome coverage for each chromatin state. **e**, Chromatin state relative genome coverage in different tissues.



Supplementary Fig. 5 The expression of tissue-specifically expressed genes (TSE) across 14 tissues (**a**) and their gene ontology (GO) functional enrichment (**b**).

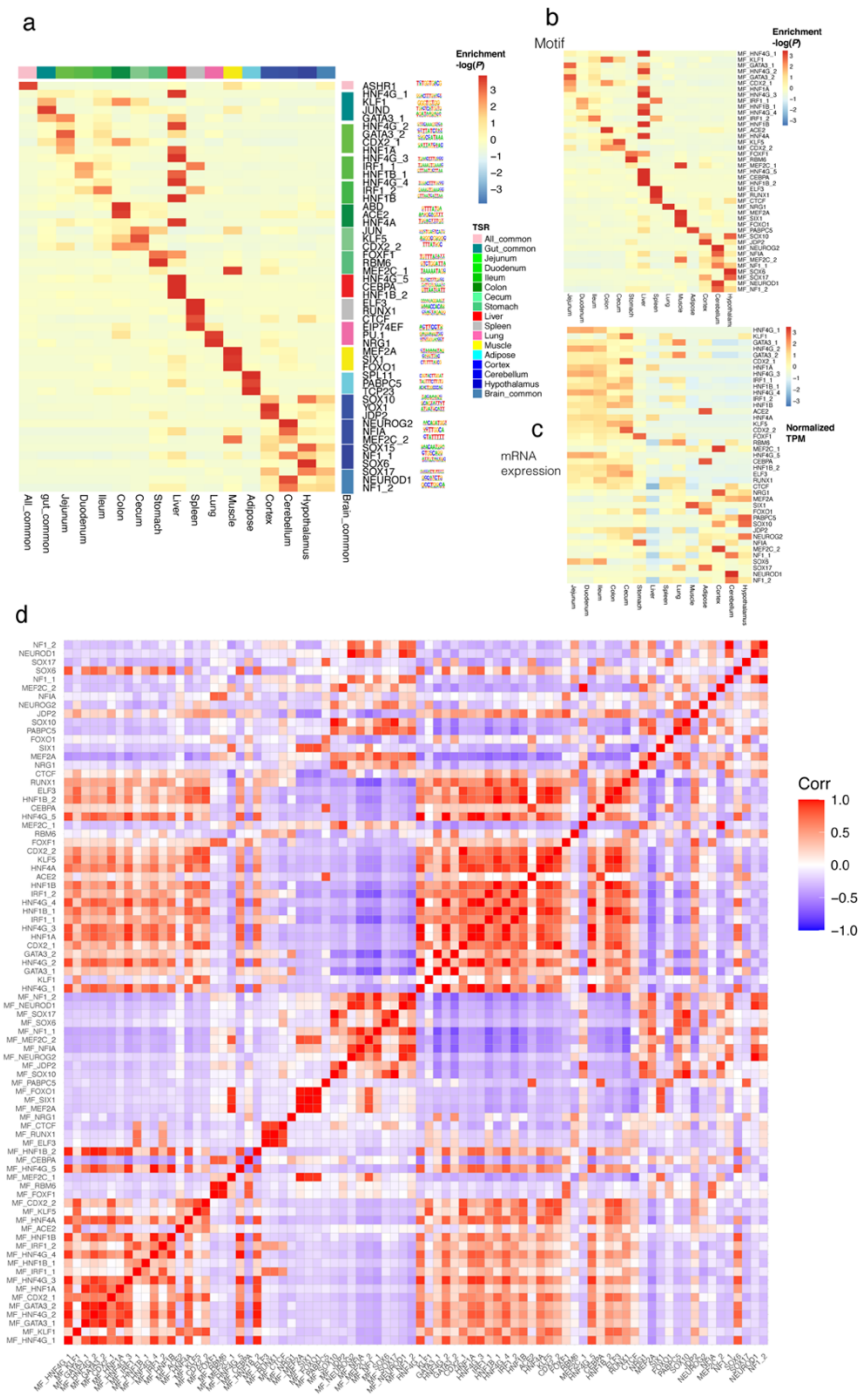


Supplementary Fig. 6 Chromatin states switching in tissues and TSE. **a,b** Chromatin states switching probabilities in between intestinal tissues, between brain tissues **c,d,e,f**. Chromatin state switching for proximal-promoters (E1), enhancers(E6), TssBivs (E12), repressors (E13) of tissue-specifically expressed genes (TSE) in jejunum. E1: 5,000bp around TSE gene's TSS, E6, E12, and E13: 25,000bp according to chromatin state density peak in Fig. 2g.

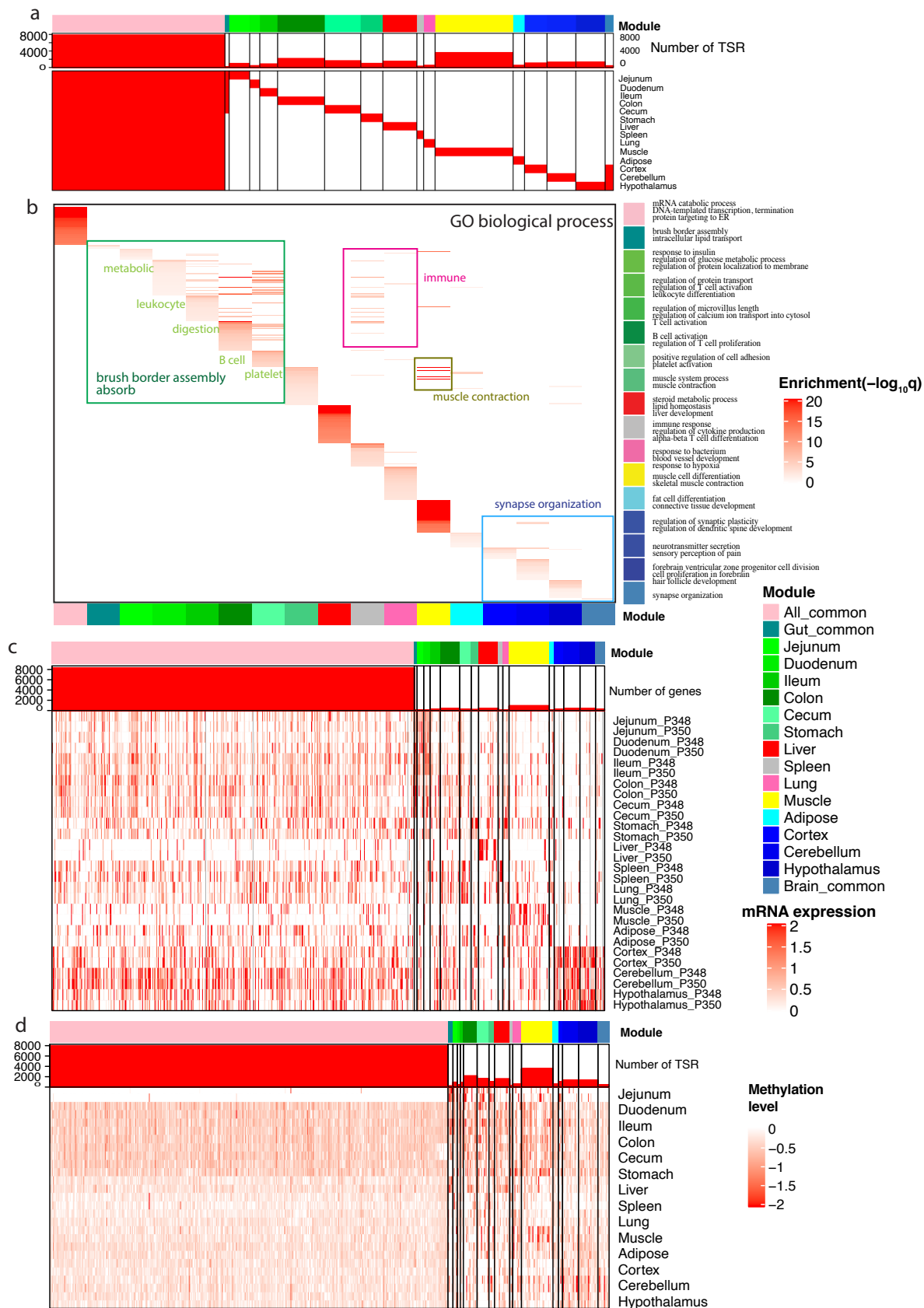


Supplementary Fig. 7 Methylation level and mouse phenotype enrichment of tissue-specific EnhAs. **a**, Methylation level of tissue-specific strong enhancers (EnhAs). The columns represent the genes in each module, the rows represent tissues. **b** Mouse phenotype enrichment of tissue-

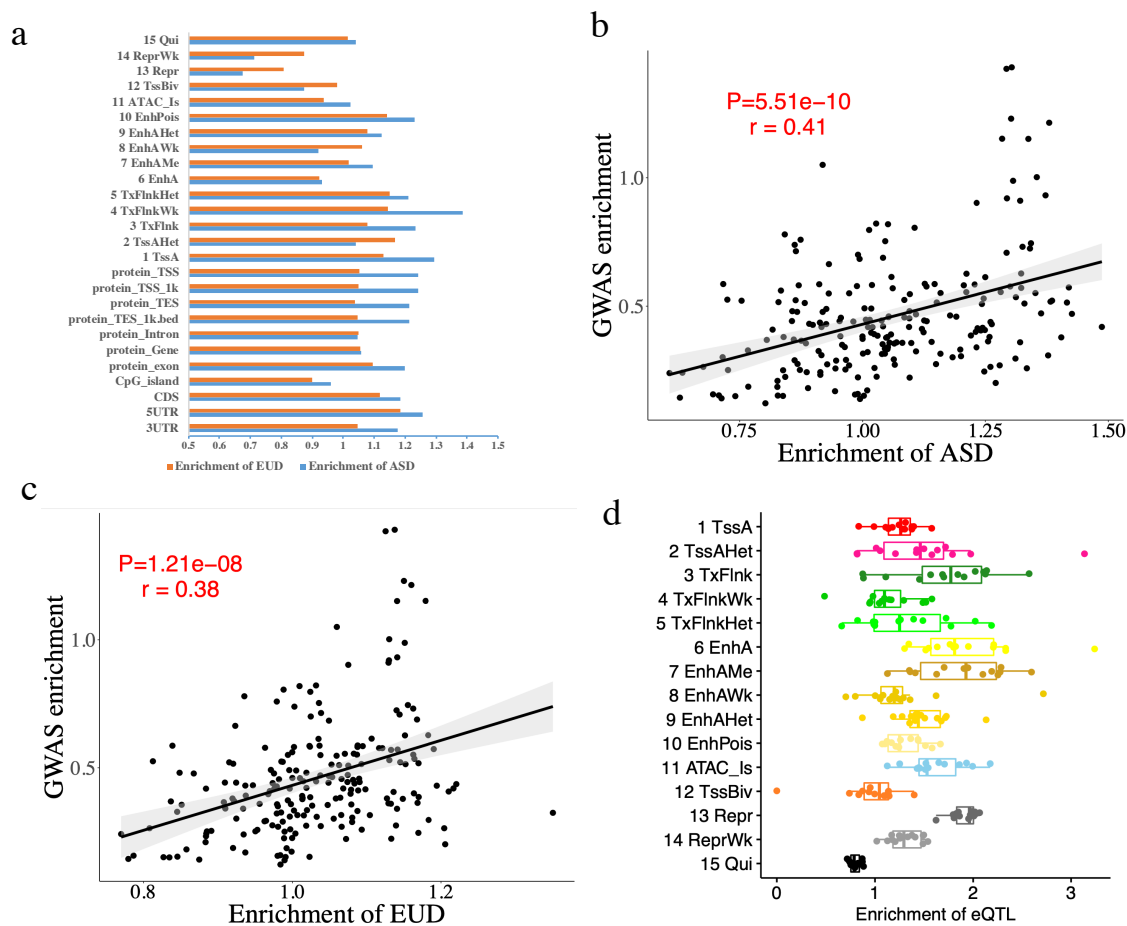
59 specific EnhAs. The columns represent 17 modules of strong enhancers. The rows represent
60 phenotypes in each module.



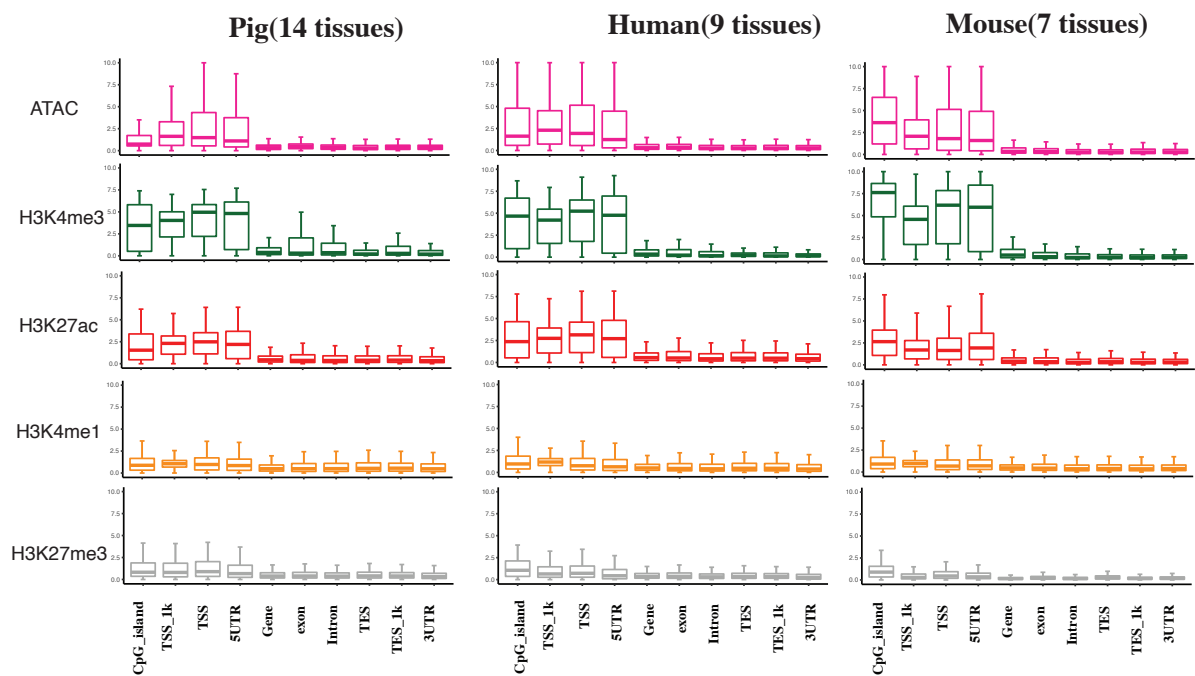
62 Supplementary Fig. 8 Motif enrichment of tissue-specific strong enhancers (EnhAs). **a**, Major
63 motif enrichment in 14 tissues. **b,c**, The corresponded motif enrichment and mRNA expression
64 in each tissues. (normalized and centered TPM). **d**, The correlation between motif enrichment
65 and the mRNA expression of corresponding transcription factors' motif.



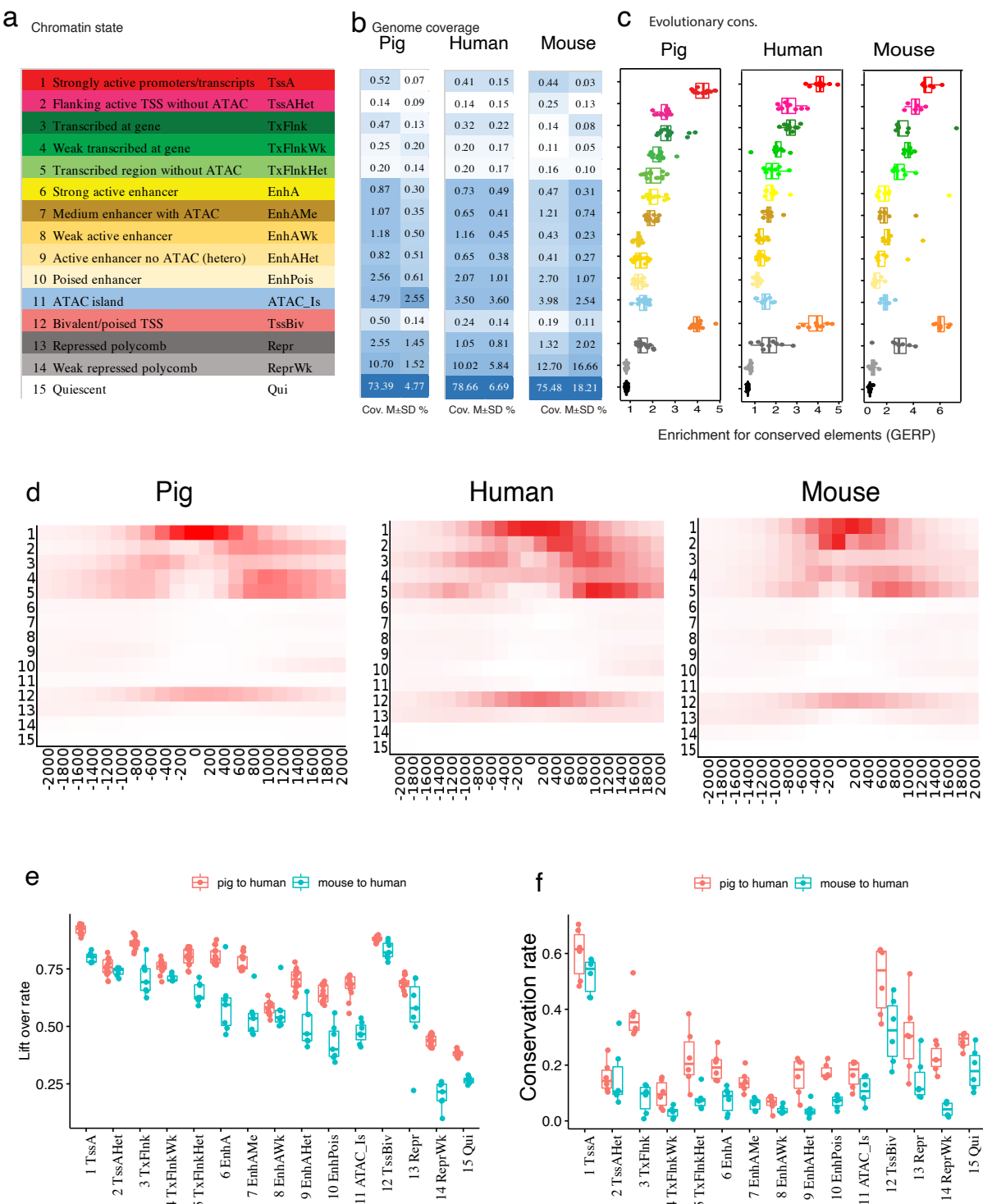
Supplementary Fig. 9 **Tissue-specific promoters (TssAs) and their potential functions in 14 tissues.** **a**, The number and enrichment distribution of 17 modules of promoters in tissues. TSR: tissue-specific regulatory elements. **b**, Functional enrichment of proximal (2000bp) genes for each module based on gene ontology (GO) biological processes. **c**, The mRNA expression (TPM) of promoters' target genes (2000bp around transcription start sites) in each module. **d**, Methylation level of tissue-specific promoters.



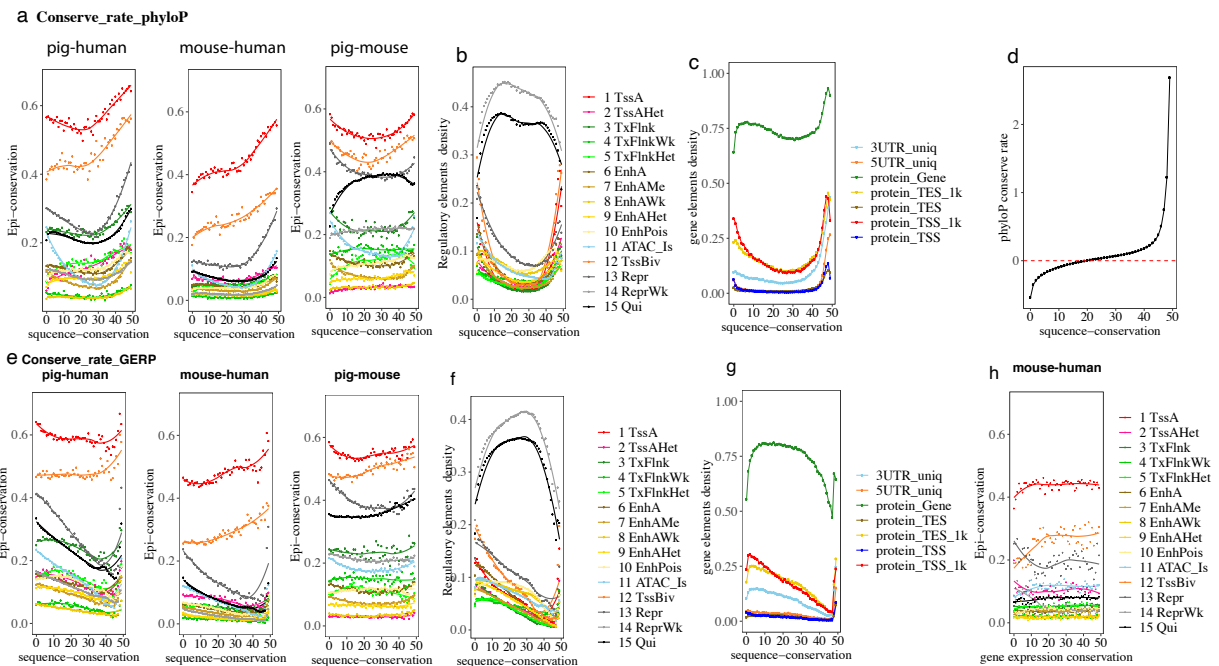
Supplementary Fig. 10 **Chromatin state enrichments of domestication selection signature and QTLs affecting complex traits in pigs.** **a**, Enrichment of selection signal of chromatin states compared with gene elements. ASD: Asian pig domestication; EUD: European pig domestication. **b,c**, Correlations among GWAS enrichment (44 traits) and domestication selection signature enrichment of chromatin states in tissues. **d**, Porcine muscle eQTL enrichment of chromatin states.



Supplementary Fig. 11 Traces of evolutionary conservation of epigenomes. Mark signal of gene elements among pig, human, and mouse.



Supplementary Fig. 12 Chromatin state conservation in pig, human, and mouse. **a**, 15 chromatin states in three species genomes defined by the ChromHMM. **b**, Genome coverage for each chromatin state in three species. **c**, Fold enrichments of chromatin states for mammalian conserved elements (GERP) in three species. **d**, Enrichment of chromatin state around genes in three species. **e**, The difference of lift over rate for each chromatin state between pig to human and mouse to human. **f**, The difference of conservation rate of each chromatin state between pig to human and mouse to human.



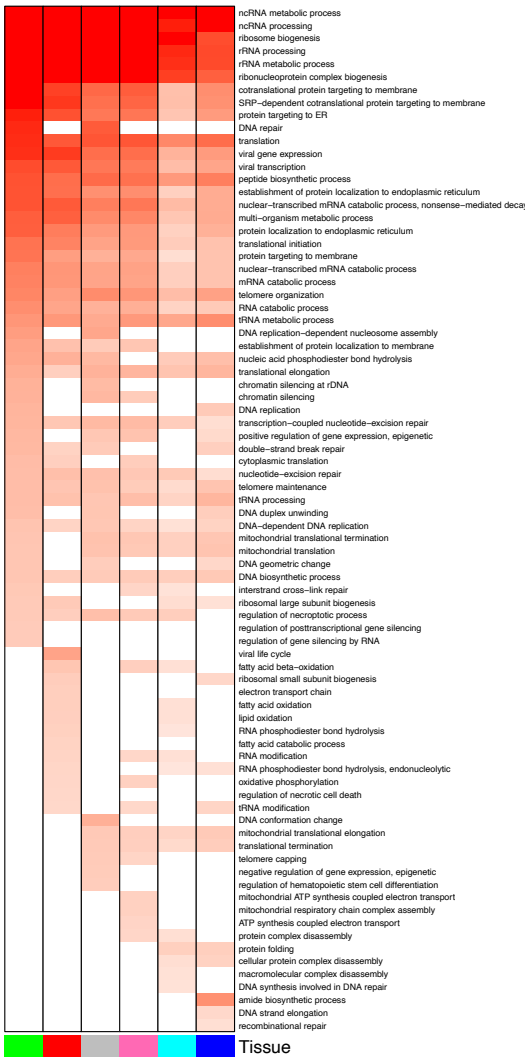
Supplementary Fig.13 Global comparison of genomic and epigenomic conservations. **a**, The relations between sequence conservation (phyloP conservation rate) and epigenomic conservation across 6 tissues. These segments were ordered from the fastest changing (0th), to slowest changing (49th). **b**, Regulatory elements density in different sequence conservation

102 segments. **c**, Gene elements densities in different sequence conservation segments. **d**, Average
103 phyloP conservation rate in each segment. **e**, The relations between sequence conservation
104 (GERP conservation rate) and epigenomic conservation. **f,g**, Regulatory and gene elements in
105 different GERP conservation segments. **h**, Relationship between expression conservation and
106 epigenomic conservation across 6 tissues in mouse-human pair. Expression conservation was
107 based on expression of 14,302 orthologous genes among 3 species. Regions were ordered from
108 the greatest difference (0th), to smallest difference (49th) in expression.

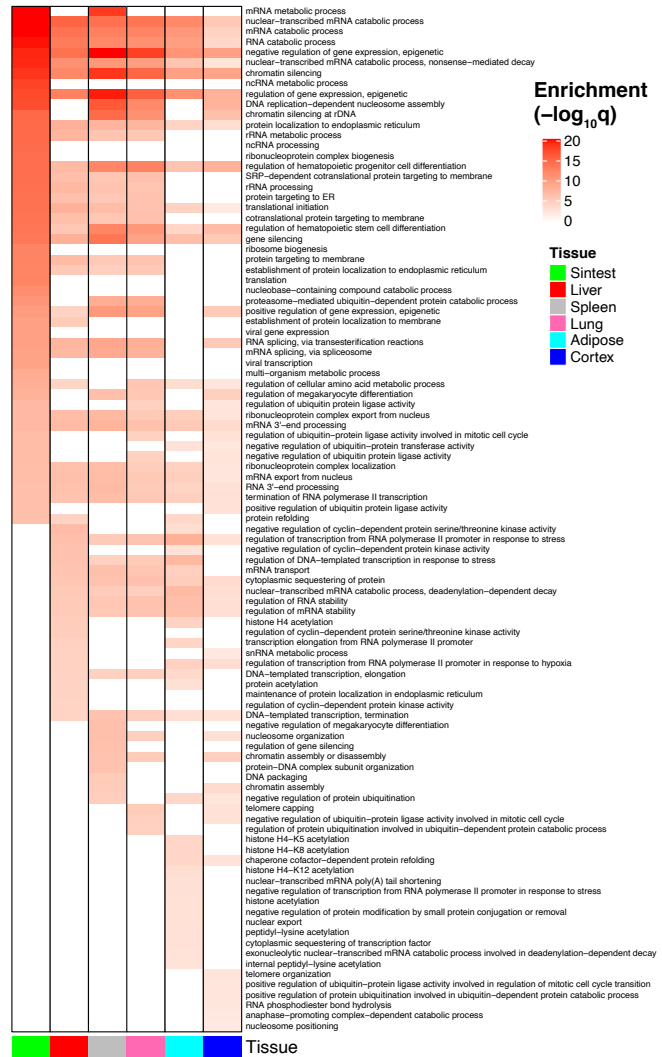
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110

Extremely variable TssA (human-pig share)



Extremely conserved TssA (human-pig share)



Enrichment
($-\log_{10} q$)

20
15
10
5
0

Tissue

Sintest
Liver
Spleen
Lung
Adipose
Cortex

111

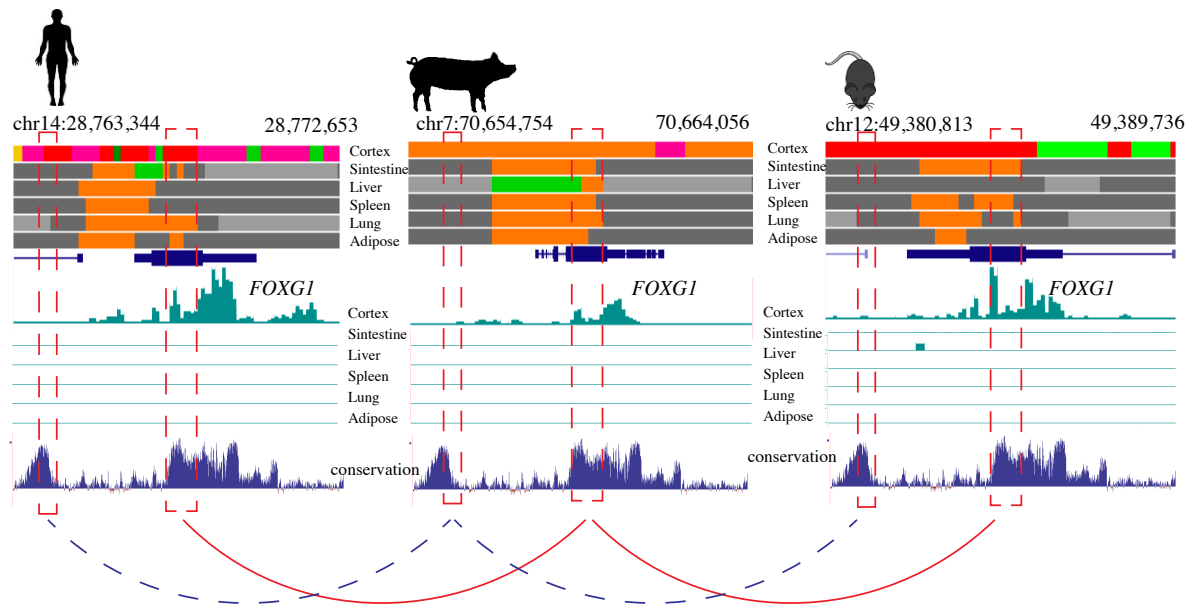
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Supplementary Fig. 14 GO functional enrichments of TssA regulatory elements for sequences

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extremely variable (0th) and extremely conserved regions (49th).

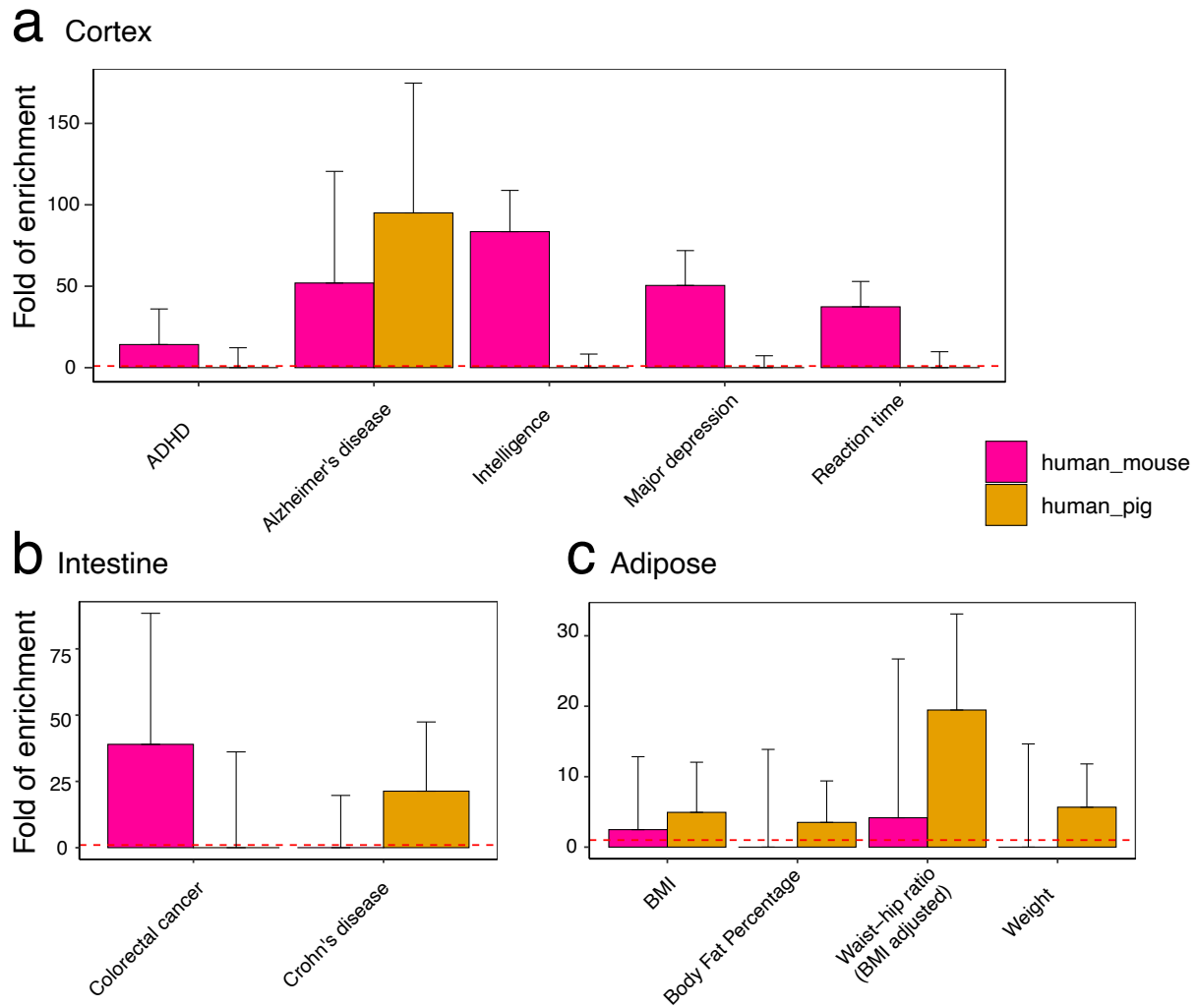
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116 Supplementary Fig. 15 Chromatin state, mRNA expression, sequence conservation around
 117 *FOXG1* locus in pig, human and mouse. In sequence extremely conserved region, human
 118 *FOXG1* has two brain cortex-specific TssAs (red dashed rectangle boxes) in which they are
 119 TssABiv in pigs. Aim Forkhead Box G1(*FOXG1*), is very important gene in cortical cell fate
 120 related with autism spectrum disorders and other brain disease^{1,2}, showed higher
 121 *FOXG1* expression in human than that in pig brain cortex. Vertical scale 0-110 for RNA-seq.

122



123

124 Supplementary Fig. 16 Different QTLs from human genome-wide association studies (GWAS)

125 (47 human diseases and complex traits) enrichment between human-pig and human-mouse

126 shared promoters (TssA) in brain cortex (**a**), small intestine (**b**), and adipose (**c**) tissues.

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129

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- 131 1. Mariani, J. *et al.* FOXP1-dependent dysregulation of GABA/glutamate neuron
132 differentiation in autism spectrum disorders. *Cell* **162**, 375-390 (2015).
133 2. Hanashima, C., Li, S.C., Shen, L., Lai, E. & Fishell, G. Foxg1 suppresses early cortical
134 cell fate. *Science* **303**, 56-59 (2004).

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