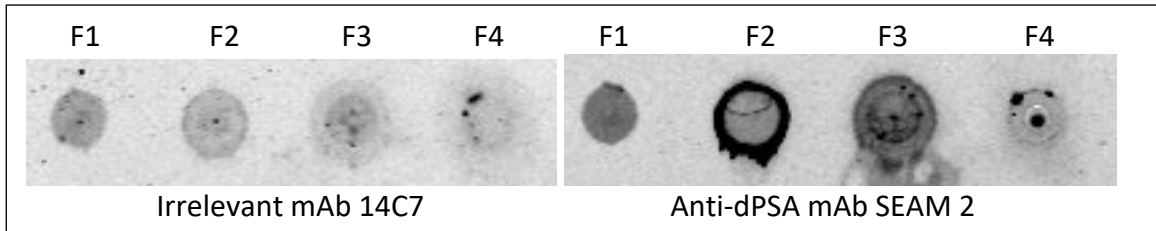
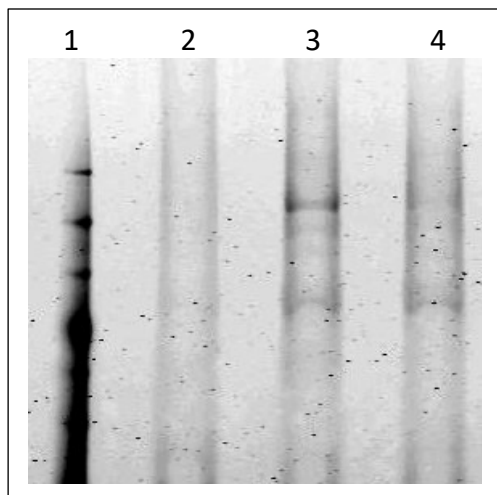


Additional file 1: Supplementary Figs. S1-S4



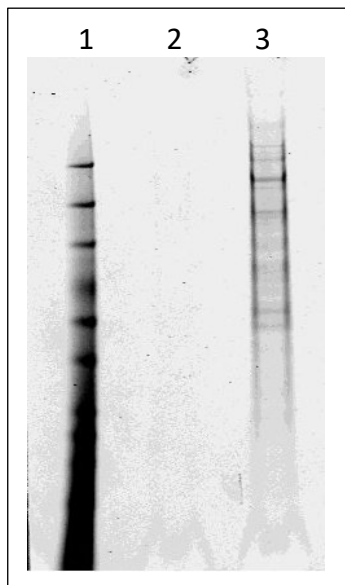
Supplementary Fig. S1. Immunodot blot of SK-MEL-28 cell differential detergent extraction fractions. Replicate portions of each detergent extract fraction from SK-MEL-28 cells prepared as described in Methods was spotted onto a PVDF membrane (Millipore) mounted on a dot blot apparatus (Topac). After blocking with 2% non-fat dry milk in DPBS blocking buffer, the section of the membrane was incubated with the indicated mAb for 1 hr at ambient temperature in the same blocking buffer and washed with DPBS containing 0.1% Tween 20 three times for 5 minutes. mAbs binding to the membrane were detected with IRDye®800CW Donkey anti-Mouse IgG (H+L) secondary antibody (LiCOR) using the same method as the primary mAb and images were recorded on an Odyssey® Fc Imaging System (LiCOR).



14

15 **Supplementary Fig. S2. Antigens immunoprecipitated from SK-MEL-28 melanoma cells**
 16 **differential detergent extraction fraction 2.** Immunoprecipitation was performed as described
 17 in the Methods using an irrelevant IgG3 mAb 14C7 (lane 2), anti-dPSA mAb SEAM 2 (lane 3)
 18 and anti-PSA mAb (SEAM 12). Proteins in the samples were resolved on 4–12% gradient SDS-
 19 PAGE gels (NuPAGE, Thermo Fisher Scientific). The gels were stained with SimplyBlue™
 20 Coomassie stain (Thermo Fisher Scientific). Individual bands were excised from the gel for
 21 LC/MS/MS analysis as described in the Methods. Protein molecular mass standards
 22 (Benchmark®, Invitrogen) are in lane 1.

23



24

25 **Supplementary Fig. S3. Antigens immunoprecipitated from Kelly neuroblastoma cells**

26 **differential detergent extraction fraction 2.** Immunoprecipitation was performed as described

27 in the Methods from Kelly neuroblastoma cell differential detergent fraction 2 using an irrelevant

28 IgG3 mAb 14C7 (lane 2), anti-dPSA mAb SEAM 2 (lane 3). Proteins in the samples were

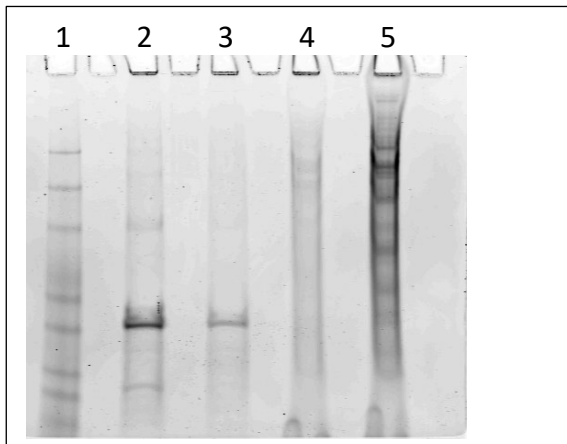
29 resolved on 4–12% gradient SDS-PAGE gels (NuPAGE, Thermo Fisher Scientific). The gels

30 were stained with SimplyBlue™ Coomassie stain (Thermo Fisher Scientific). Individual bands

31 were excised from the gel for LC/MS/MS analysis as described in the Methods. Protein

32 molecular mass standards (Benchmark®, Invitrogen) are in lane 1.

33



34

35 **Supplementary Fig. S4. Antigens immunoprecipitated from PBMCs and SNU-1 gastric**

36 **cancer cells differential detergent extraction fraction 2.** Immunoprecipitation was performed

37 as described in the Methods from PBMCs (lanes 2 and 4) or SNU-1 cells (lanes 4 and 5) using an

38 irrelevant protein, Neisserial Factor H binding protein (lanes 2 and 3), or anti-dPSA mAb SEAM

39 2 (lanes 4 and 5). Proteins in the samples were resolved on 4–12% gradient SDS-PAGE gels

40 (NuPAGE, Thermo Fisher Scientific). The gels were stained with SimplyBlue™ Coomassie

41 stain (Thermo Fisher Scientific). Individual bands were excised from the gel for LC/MS/MS

42 analysis as described in the Methods. Protein molecular mass standards (Benchmark®,

43 Invitrogen) are in lane 1.

44