

Supplementary Materials

Figure titles and legends

Figure S1: Impaired CCCP-induced TOM20 degradation in *PARK2* KO neurons

A) *PARK2* KO and control neurons were untreated or treated with vehicle (DMSO) or 10 μ M CCCP for 24, 48, 72, and 96 hrs and immunofluorescence stained for TOM20 (red) and Hoechst (blue) to visualize mitochondria and nuclei as indicated. Scale bar: 10 μ m. **B)** No significant differences in total cell numbers were observed upon CCCP or vehicle (0.1% DMSO) exposure although there was a clear trend towards fewer cells after 72 and 96 hrs. **C)** Quantification of TOM20+ mitochondrial area normalized to number of Hoechst+ nuclei showed that the mitochondrial area in control neurons was significantly reduced after 48, 72, and 96 hrs of CCCP treatment. In contrast, mitochondria were retained in the *PARK2* KO neurons. Vehicle (0.1% DMSO) did not affect area of TOM20 immunoreactivity. Data presented as mean $\pm$ SEM, n=9 technical replicates, data from 3 independent differentiations, Significant differences are indicated by **p < 0.01, ***p < 0.001 (control vs. *PARK2* KO), #p < 0.05 (untreated control vs. CCCP-treated control), one-way ANOVA followed by Dunnett's post hoc test for multiple comparisons

Figure S2: USP30 inhibition as a strategy to enhance mitophagy

A) USP30 opposes parkin-mediated ubiquitination. **B)** Western blotting and densitometric analysis demonstrating comparable levels of USP30 protein expression in *PARK2* KO and control neurons. Representative blots of two independent experiments.

Figure S3: Lactate dehydrogenase (LDH) release did not reveal cytotoxic effects of USP30 inhibitors (+/-CCCP)

Differentiated neurons were treated with 0 μ M (untreated), 0.75 μ M, 1.5 μ M, 3 μ M, 6 μ M of USP30i-37 and USP30i-3 inhibitors added 4 h prior to CCCP (10 μ M, 48 h). Necrotic cell death was assessed by LDH analysis of conditioned media from **A)** control neurons treated with USP30i-37, **B)** *PARK2* KO neurons treated with USP30i-37, **C)** control neurons treated with USP30i-3, and **D)** *PARK2* KO neurons treated with USP30i-3. No significant cytotoxicity was observed after treatment in either cell line, but there was a tendency to increase with increasing dose. Data presented as mean $\pm$ SEM, n=9-15 technical replicates, data from 3 independent differentiations.

Figure S4: Immunofluorescence staining showed that 3 μ M USP30 inhibitors did not affect the numbers of nuclei (DAPI), mature neurons (MAP2), or dopaminergic neurons (TH) (+/-CCCP)

Healthy control and *PARK2* KO neurons were treated with 0 μ M (untreated), 0.75 μ M, 1.5 μ M, 3 μ M, 6 μ M of USP30i-37 and USP30i-3. Cells were fixed and stained for DAPI (blue), MAP2 (red), and TH (green). **A)** Representative immunofluorescence pictures of DAPI+, MAP2+, and TH+ cells in healthy control and *PARK2* KO iPSC-derived neurons. Scale bars: 200 μ m. **B-E)** Quantification of total cell count in **B, C)** healthy control and **D, E)** *PARK2* KO neurons. The toxicity of USP30 compounds (decreased cell number and compromised cell morphology) increased in both cell lines at the highest concentrations. Data presented as mean $\pm$ SEM, n=9 technical replicates, data from 3 independent differentiations. Significant differences are indicated by *, #p < 0.05, **, ##p < 0.01, ***p < 0.001, one-way ANOVA followed by Dunnett's post hoc test for multiple comparisons.

Figure S5: Morphological assessment of nuclei in control and *PARK2* KO neurons after treatment with USP30i-37 and USP30i-3.

Differentiated neurons were treated with with 0 μ M (untreated), 0.75 μ M, 1.5 μ M, 3 μ M, 6 μ M of USP30i-37 and USP30i-3 inhibitors added 4 h prior to CCCP (10 μ M, 48 h). Nuclear morphology was assessed by quantification of **(A-B)** fragmented nuclei in control and *PARK2* KO neurons treated with **A)** USP30i-37 and **B)** USP30i-3, and **(C-D)** pycnotic nuclei in control and *PARK2* KO neurons treated with **C)** USP30i-37 and **D)** USP30i-3. Data presented as mean $\pm$ SEM, n=9 technical replicates, data from 3 independent differentiations. Significant differences are indicated by * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, ns: non significant, one-way ANOVA followed by Dunnett's post hoc test for multiple comparisons.

Figure S1

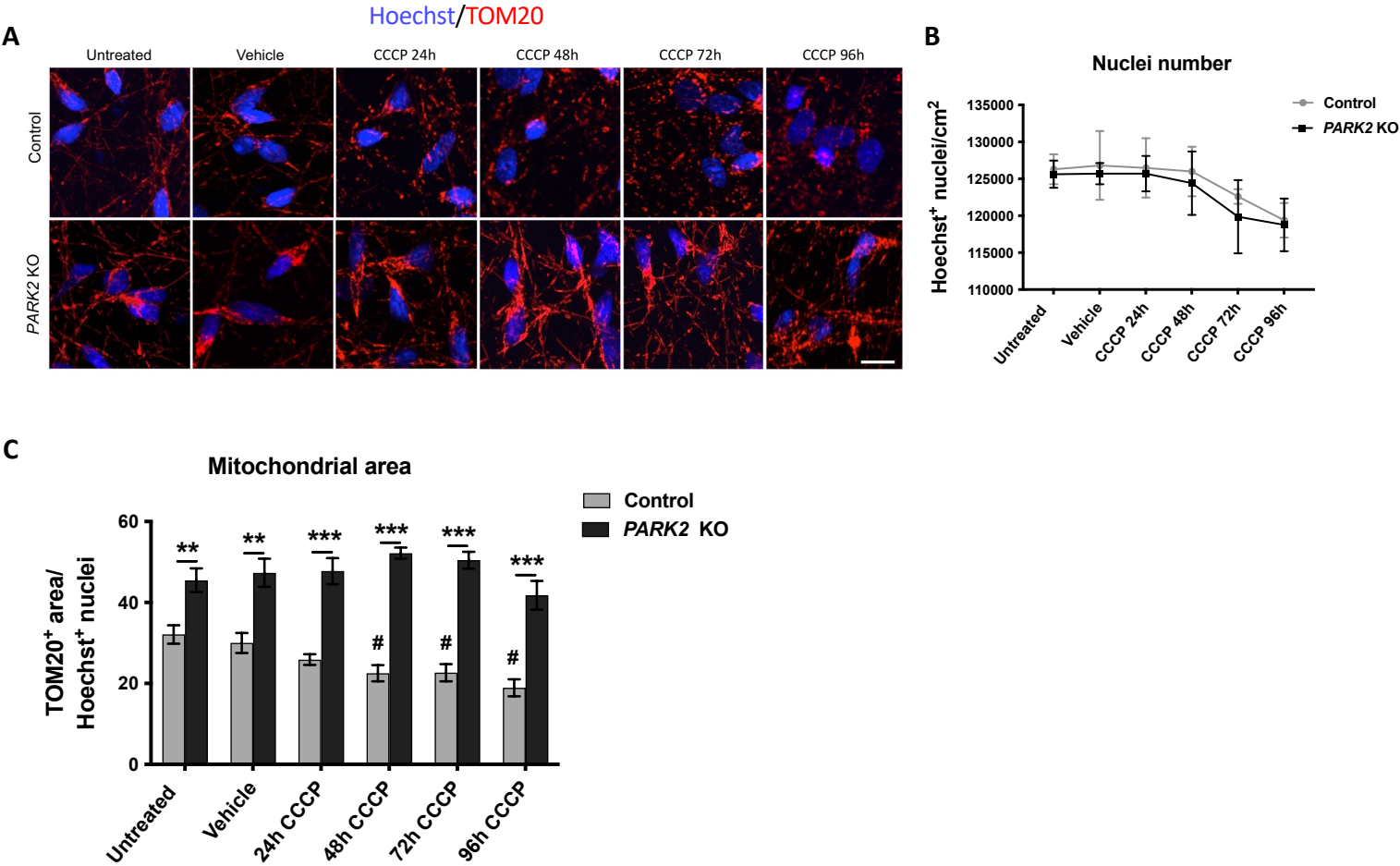
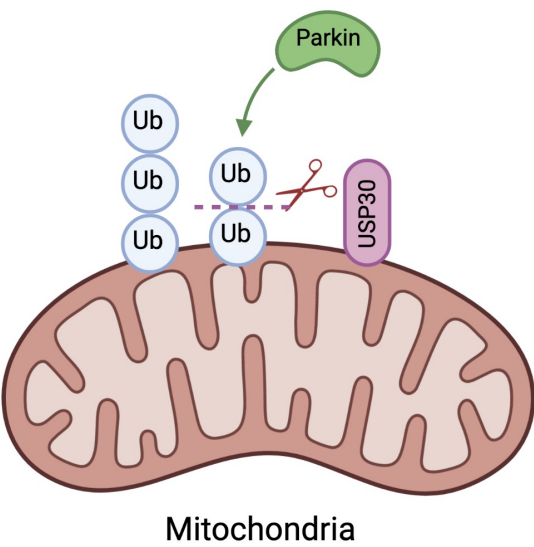


Figure S2

A



B

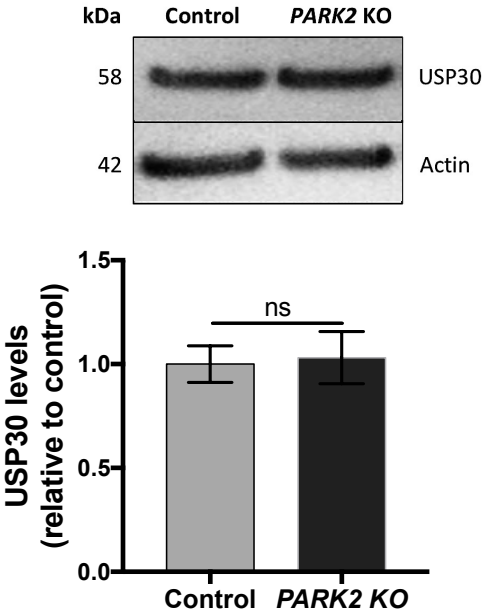
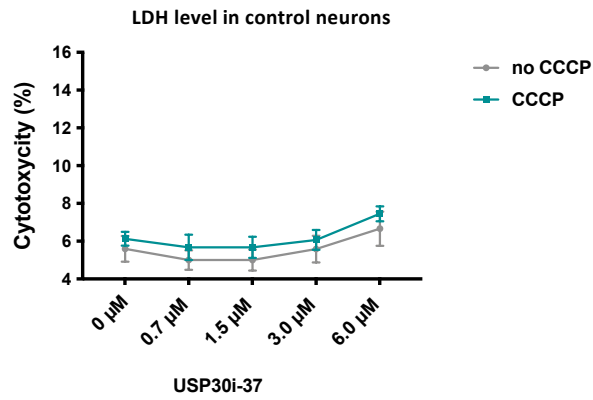
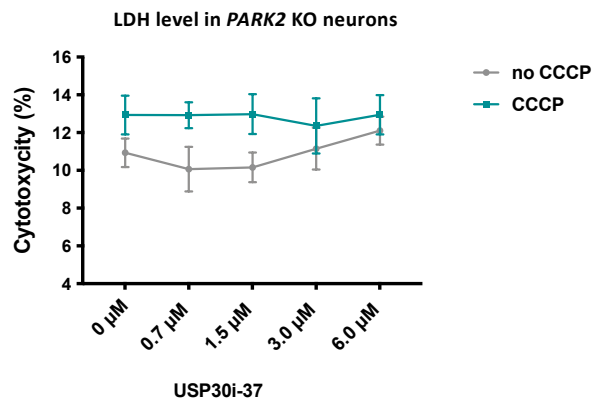


Figure S3

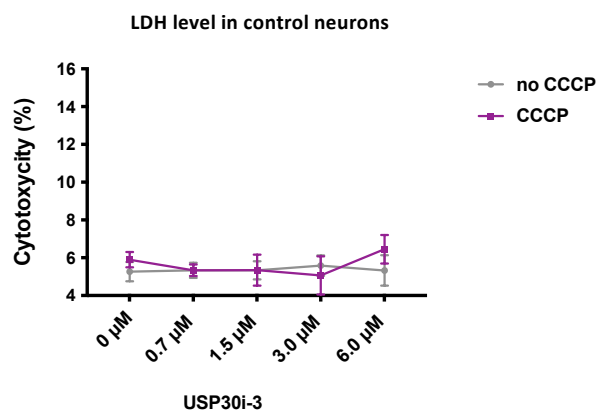
A



B



C



D

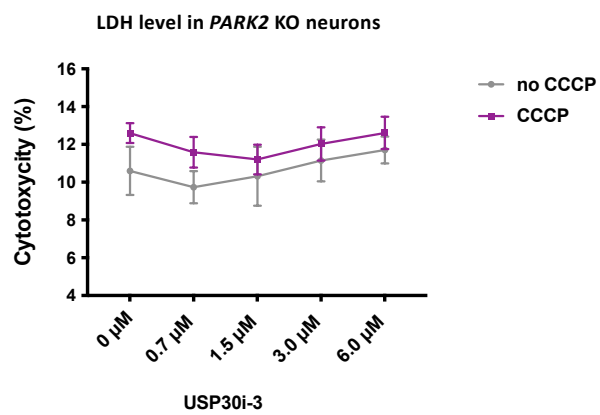
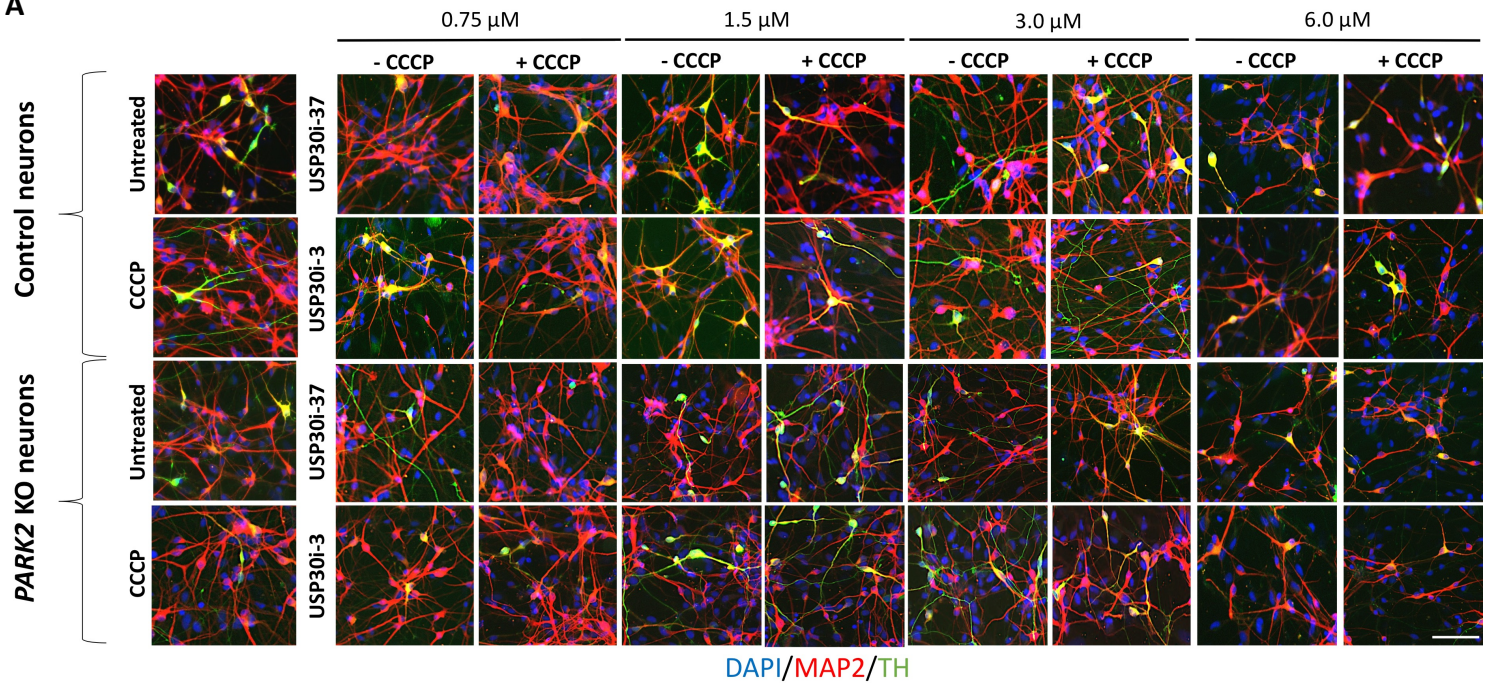
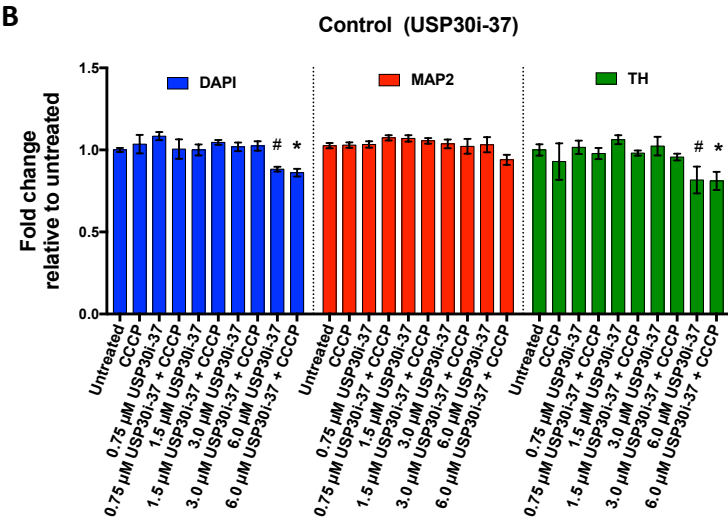


Figure S4

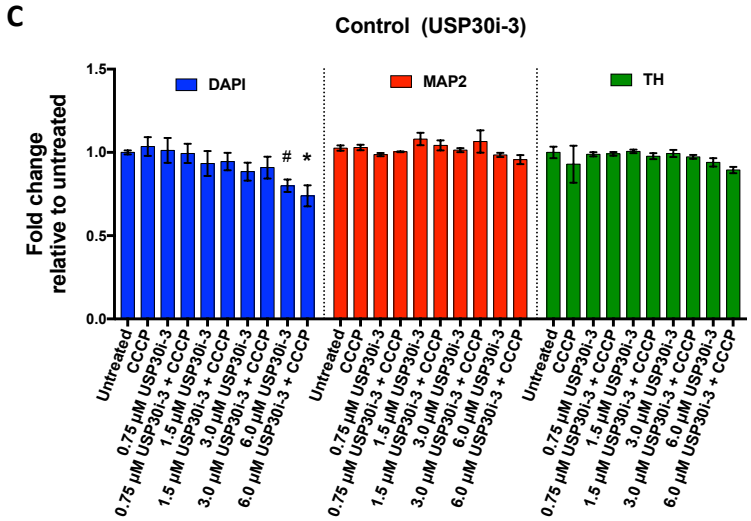
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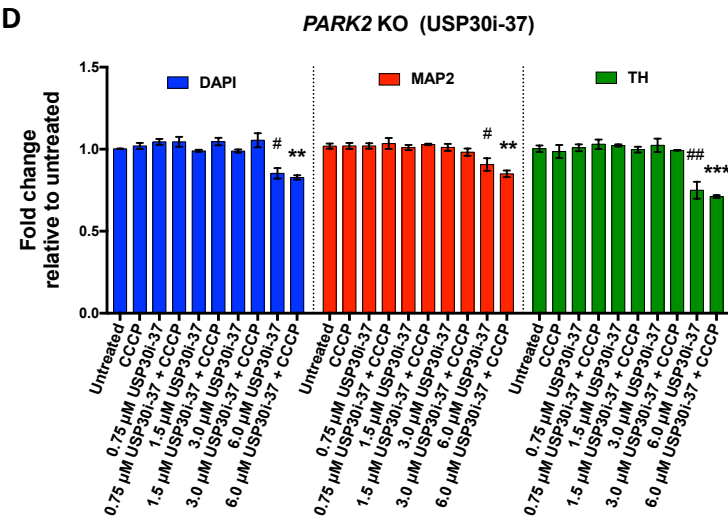
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C



D



E

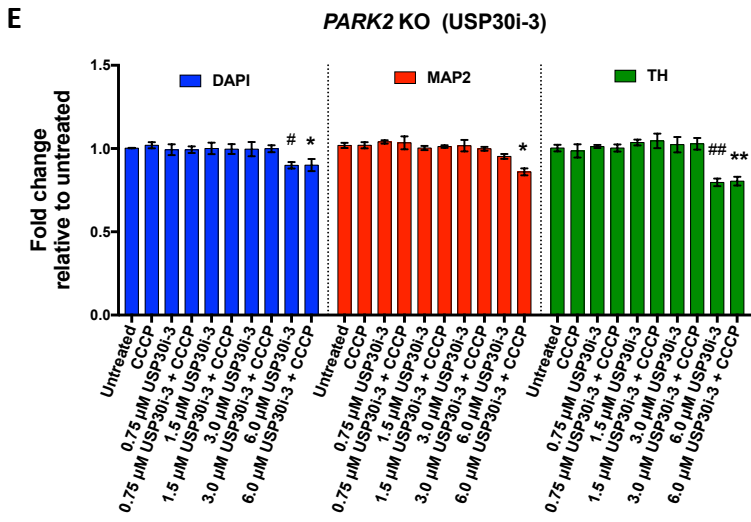
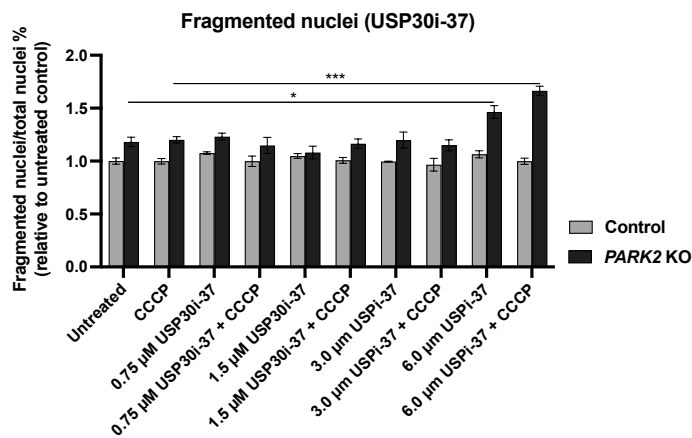
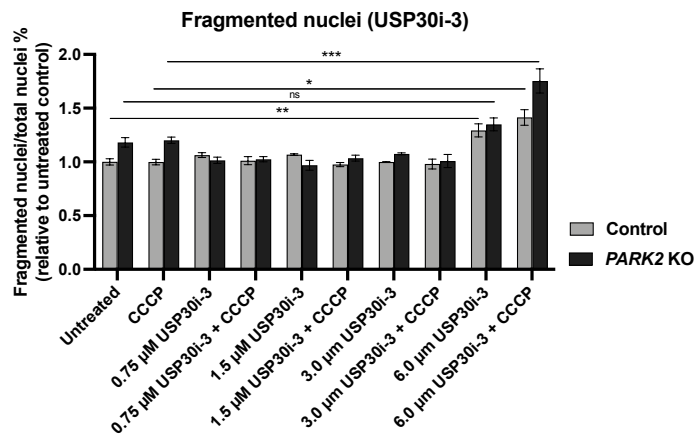


Figure S5

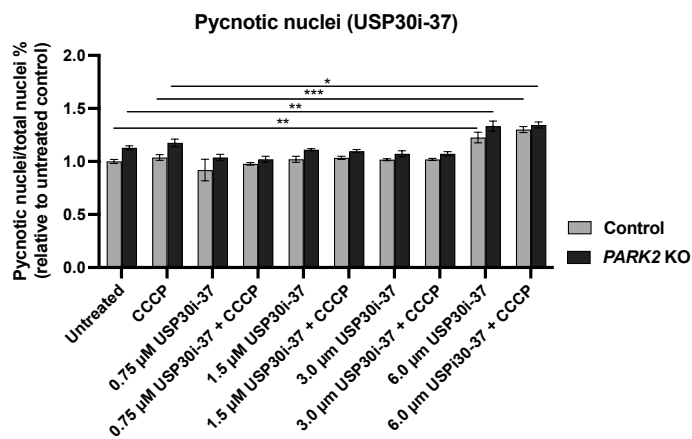
A



B



C



D

