

### **Supplementary information**

**Supplementary Figure 1** (supporting Figure 2). Subcellular localization, APC/C degradation assay and expression analysis of *S. pombe* Sor1. **a, b** Sor1 localizes to nucleus throughout the cell cycle. Cycling *S. pombe* cells expressing Sor1-GFP were fixed, stained with DAPI, and analyzed by fluorescence microscopy (**a**). Cycling *S. pombe* cells expressing Sor1-Flag were fixed and stained with antibodies against Flag and tubulin. Nuclei were visualized by DAPI staining (**b**). **c** *in vitro* assay shows no evidence that Sor1 is an APC/C substrate. **d** Mutating conserved Sor1 residues only slightly reduces Sor1 protein levels. Proteins extracted from cycling cells were analyzed by gel electrophoresis and Western blotting using anti-tubulin antibodies. The TAP epitope was detected using PAP antibodies (rabbit antiperoxidase antibody linked to peroxidase).

**Supplementary Figure 2** (supporting Figures 3 and 4). Meiotic and somatic *Atsororin* mutant phenotypes can be complemented with the wild type *AtSORORIN* gene. **a** Overall plant architecture and fertility are wild type-like in the complemented transgenic plant line, but not in the *Atsororin* mutant. **b** Seed counts demonstrate nearly wild type-like fertility of the complemented transgenic plant line, but sterility in the *Atsororin* mutant. Unpaired Mann-Whitney test has been applied (\*\*\*\* $p < 0.0001$ ). Somatic defects in *Atsororin* mutants are tissue-specific and WAPL-dependent. DNA was stained with DAPI (magenta) and fluorescence *in situ* hybridization (FISH) was performed to detect centromeric regions (green). **c** Spreads of cell nuclei from rosette leaf cells. Interphase stages were analyzed for wild type plants and *Atsororin*, *wapl1-1 wapl2* and *Atsororin wapl1-1 wapl2* mutants. The number of centromeric signals is indicated in the top left corner. Scale bar = 10  $\mu\text{m}$ . **d** Quantification of centromeric-FISH signals in interphase leaf nuclei. *Atsororin* mutants ( $n = 52$ ) have a significantly higher number of cells that have more than 10 signals, when compared to wild type ( $n = 84$ ), *wapl1-1 wapl2* ( $n = 68$ ) and *Atsororin wapl1-1 wapl2* ( $n = 82$ ). Chi-square test was performed (\*\*\*\* $p < 0.0001$ ).

**Supplementary Figure 3** (supporting Figure 7). Immunolocalization of the axis protein ASY1 and the meiosis-specific cohesin subunit REC8 in male meiocytes during late zygotene in wild type plants and *Atsororin*, *wapl1-1 wapl2* and *Atsororin wapl1-1 wapl2* mutants. Absence of AtSORORIN does not influence their time of deposition or their relative localisation on meiotic chromosomes. Scale bar = 10  $\mu\text{m}$ .

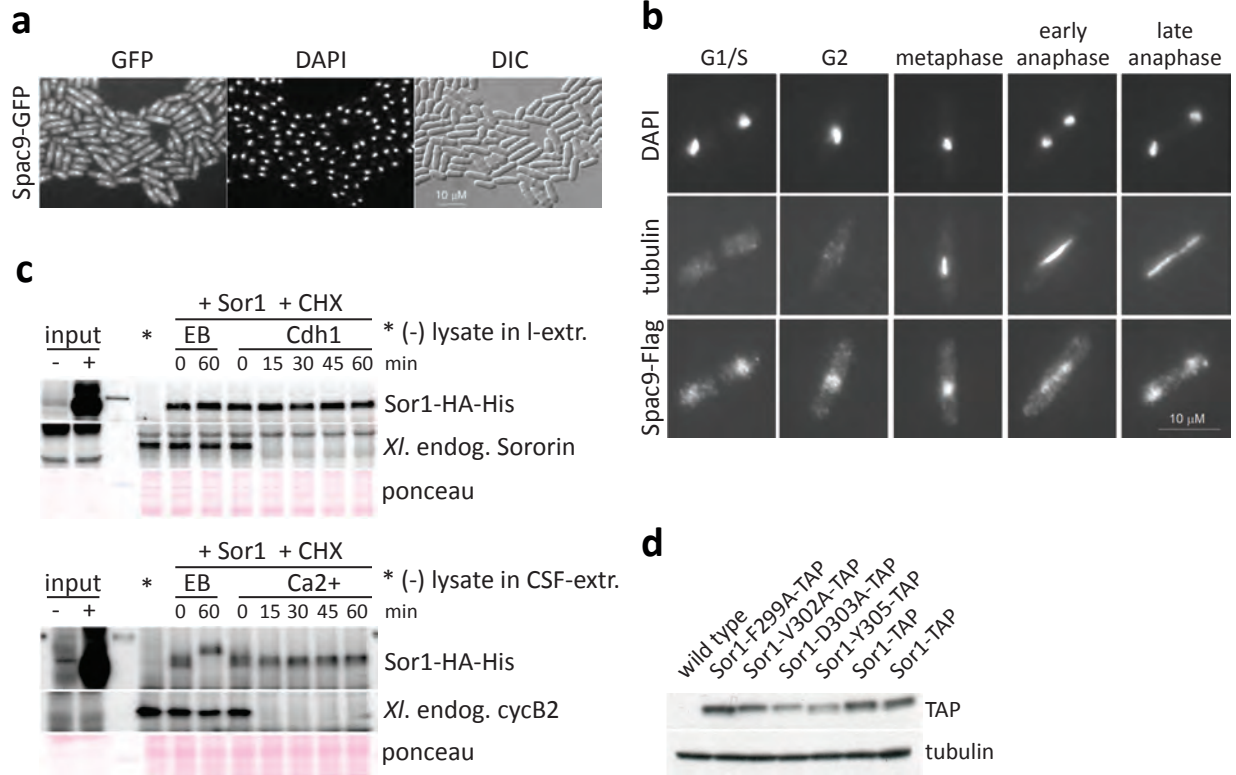
**Supplementary Movie 1.** Root tips from wild type plants display normal tissue organization and nucleus size.

**Supplementary Movie 2.** Root cellular organization and nucleus size are highly affected in *Atsororin* mutant plants.

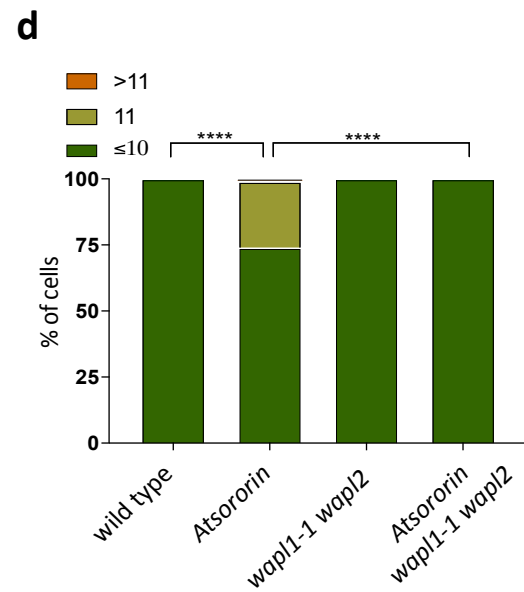
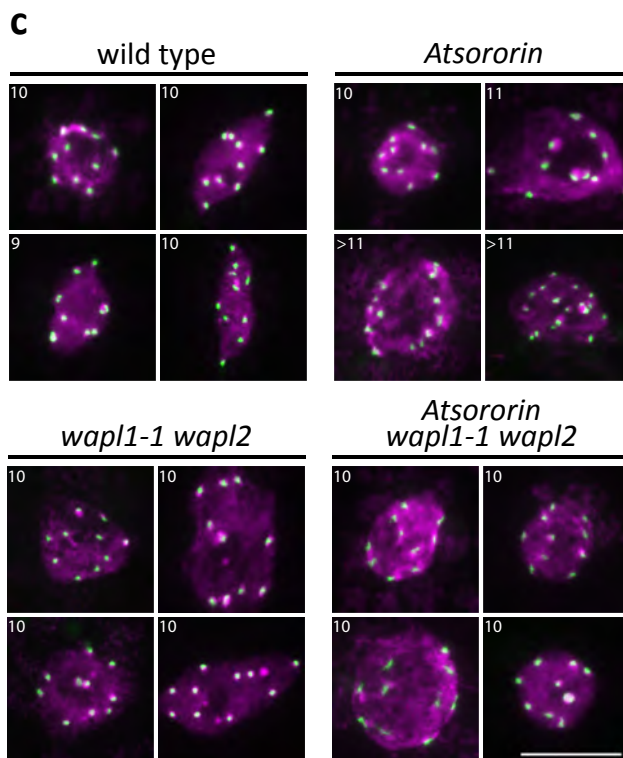
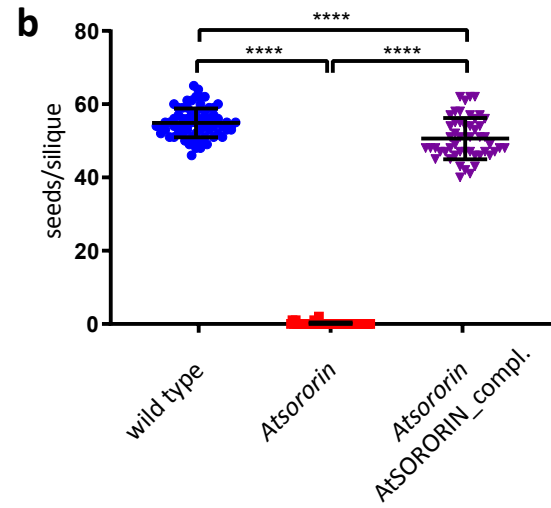
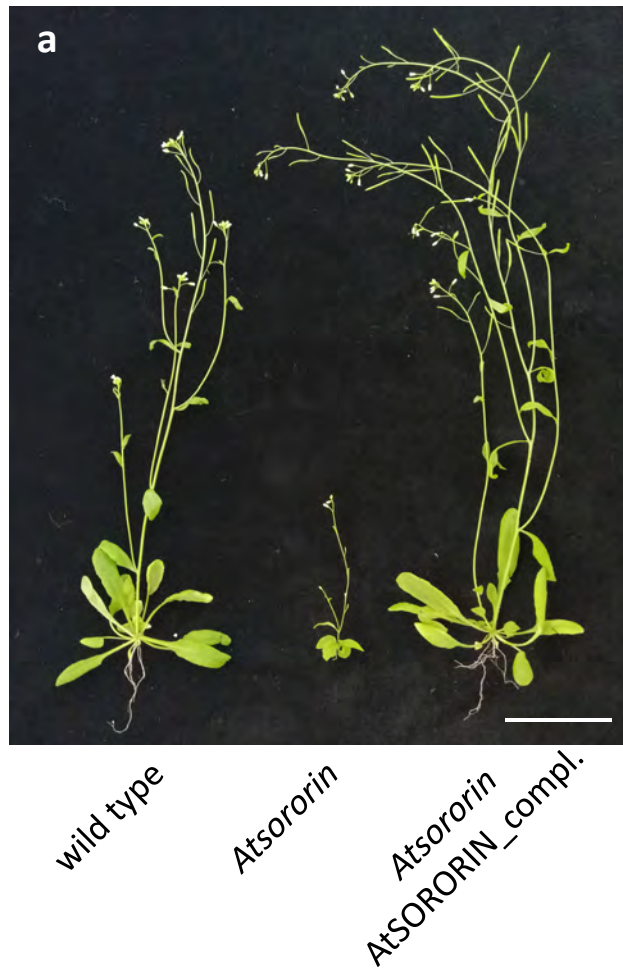
**Supplementary Movie 3.** Plants with mutations in both genes encoding WAPL (*wapl1-1 wapl2*) develop normal roots compared to wild type plants.

**Supplementary Movie 4.** *Atsororin wapl1-1 wapl2 triple* mutant plants develop normal roots, indicating that the *wapl1-1 wapl2* mutations suppress the effect of the *Atsororin* mutation with respect to root development.

# Supplementary Figure 1



# Supplementary Figure 2



## Supplementary Figure 3

