

Supplementary Material

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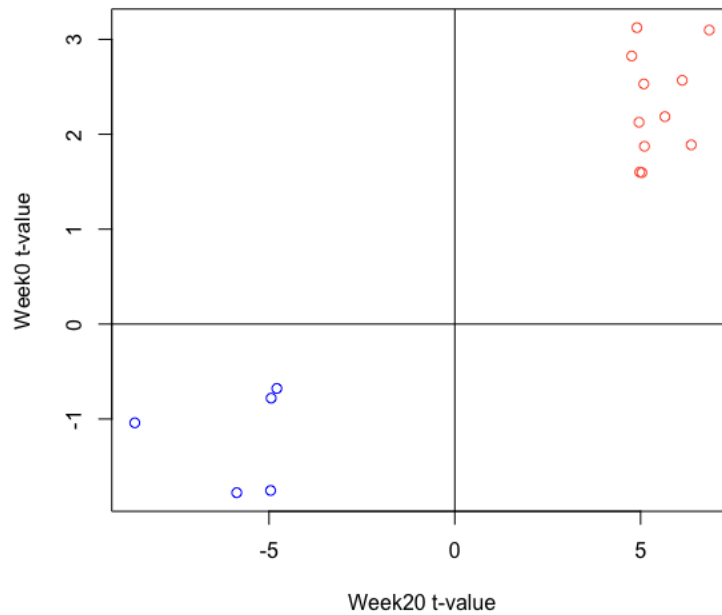
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Supplementary Table 1. Preservation of gene expression differences between baseline and response time point (week 20).

Symbol	Accession	Definition	FCh wk0	P wk0	FCh wk20	P wk20
Significant genes at baseline						
<i>PIK3CD</i>	NM_005026.2	phosphoinositide-3-kinase, catalytic, delta polypeptide (PIK3CD)	-1.56	7.10E-18	-1.22	0.3
<i>CX3CL1</i>	NM_002996.3	chemokine (C-X3-C motif) ligand 1 (CX3CL1)	1.94	7.40E-12	1.96	0.0019
<i>PLS3</i>	NM_005032.3	plastin 3 (T isoform) (PLS3)	1.71	2.00E-10	1.65	0.059
<i>DIXDC1</i>	NM_033425.1	DIX domain containing 1 (DIXDC1)	1.90	9.30E-10	1.69	0.01
<i>HCLS1</i>	NM_005335.3	hematopoietic cell-specific Lyn substrate 1 (HCLS1)	-1.63	4.40E-09	-1.68	0.076
<i>TMOD1</i>	NM_003275.1	tropomodulin 1 (TMOD1)	2.10	1.60E-08	1.76	0.029
<i>PCOLCE2</i>	NM_013363.2	procollagen C-endopeptidase enhancer 2 (PCOLCE2)	2.45	3.30E-07	2.39	0.061
<i>PPP1R3C</i>	NM_005398.3	protein phosphatase 1, regulatory (inhibitor) subunit 3C (PPP1R3C)	2.13	3.60E-07	2.09	0.011
<i>CRTAP</i>	NM_006371.3	cartilage associated protein (CRTAP)	1.58	4.80E-07	1.33	0.11
<i>PRELP</i>	NM_002725.3	proline/arginine-rich end leucine-rich repeat protein (PRELP), transcript variant 1	2.02	6.40E-07	1.79	0.11
<i>DKFZP686-A01247</i>	NM_014988.1	hypothetical protein (DKFZP686A01247)	2.30	7.10E-07	1.78	0.033
<i>ADA</i>	NM_000022.2	adenosine deaminase (ADA)	-1.73	7.10E-07	-1.45	0.08
<i>NT5DC2</i>	NM_022908.1	5'-nucleotidase domain containing 2 (NT5DC2)	-1.86	7.70E-07	-1.23	0.44
<i>UNQ689</i>	NM_212557.1	RSTI689 (UNQ689)	5.09	9.50E-07	5.53	0.0018

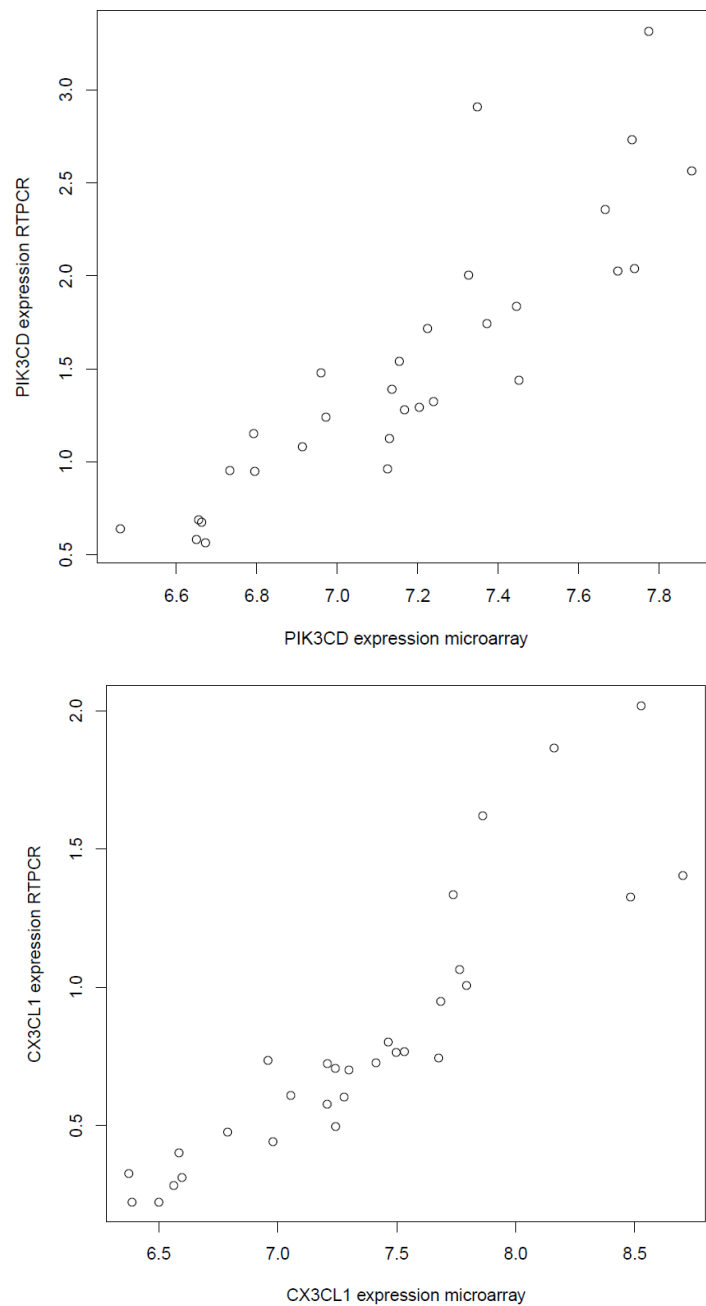
<i>SERPINH1</i>	NM_001235.2	serpin peptidase inhibitor, clade H (heat shock protein 47), member 1, (collagen binding protein 1) (SERPINH1)	-1.66	1.70E-06	-1.30	0.5
<i>OR2A9P</i>	NR_002157.1	olfactory receptor, family 2, subfamily A, member 9 pseudogene (OR2A9P) on chromosome 7.	2.01	1.90E-06	1.71	0.0047
Significant genes at week 20						
<i>RPS6KA2</i>	NM_001006932.1	ribosomal protein S6 kinase, 90kDa, polypeptide 2	-1.58	0.074	-1.53	3.10E-07
<i>LIPA</i>	NM_000235.2	lipase A, lysosomal acid, cholesterol esterase (Wolman disease)	-1.14	0.34	-1.50	1.50E-07
<i>ALPL</i>	NM_000478.2	alkaline phosphatase, liver/bone/kidney	-1.38	0.58	-2.77	7.90E-07
<i>PDE4B</i>	NM_002600.2	phosphodiesterase 4B, cAMP-specific	1.03	0.88	1.57	1.10E-07

List of genes that are significant after Bonferroni correction and with an (absolute) >1.5 fold-change (FC) between responders and non-responders to anti-TNF therapy at week 0 and week of response. In bold, the genes at week 20 that preserve the differential expression between the two groups at the nominal level ($P < 0.05$). Conversely, none of the differentially expressed genes at week 20 had a nominally significant difference at baseline.



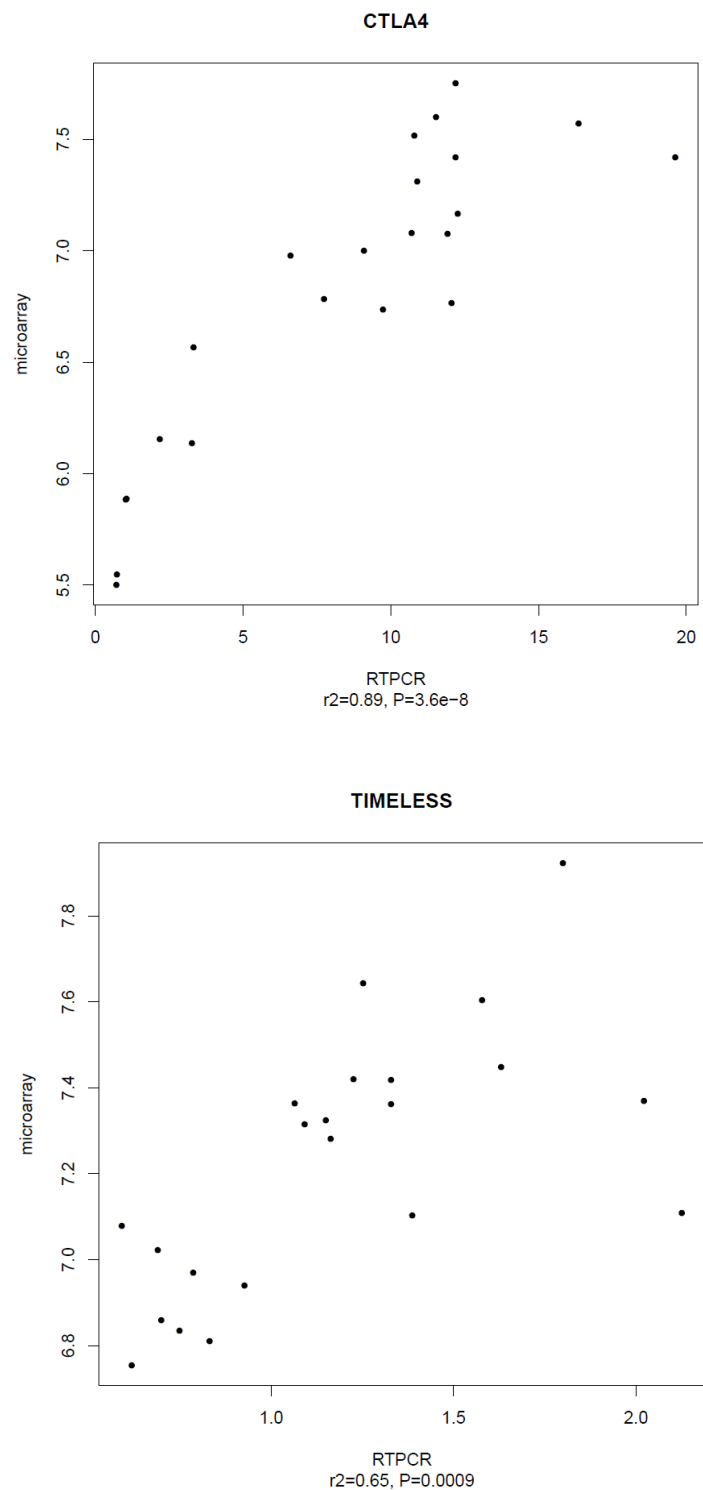
Supplementary Figure 1. Preservation of differential expression changes associated to anti-TNF response in the synovium from baseline. The t-values of the genes that are differential at week 0 between responders and non-responders (n=16) are plotted against the t-values of the same genes at week 0. While not all differences are significant at week 20, the direction of the change is clearly preserved in all overexpressed (red, n=11) and underexpressed (blue, n=5) genes.

Supplementary Figure 2. RTPCR replication of *CX3CL1* and *PIK3CD* genes.



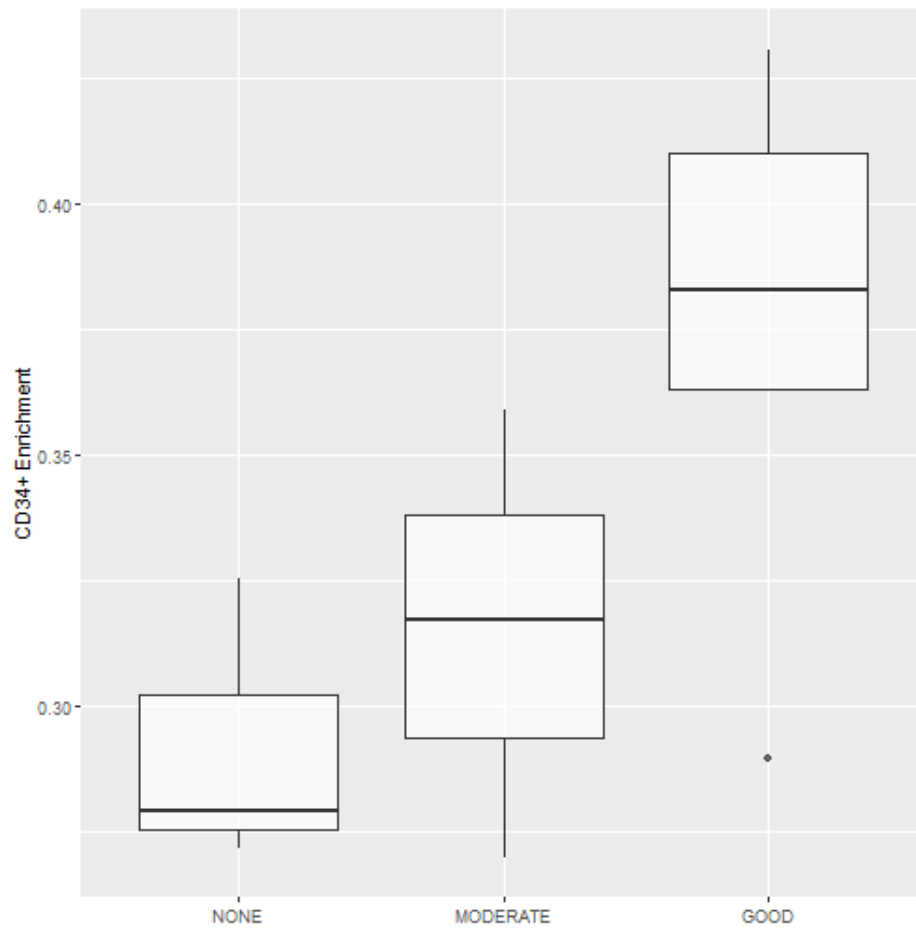
The gene expression of the two genes more differentially expressed at baseline between responders and non-responders was validated using Taqman QRT-PCR assays. The correlation between both platforms was highly significant ($r=0.86$ $P=2.42 \times 10^{-10}$ and $r=0.90$ $P=2.11 \times 10^{-11}$ for *PIK3CD* and *CX3CL1*, respectively).

Supplementary Figure 3. QRTPCR replication of T_{PH} biomarker genes *CTLA4* and *TIMELESS*.

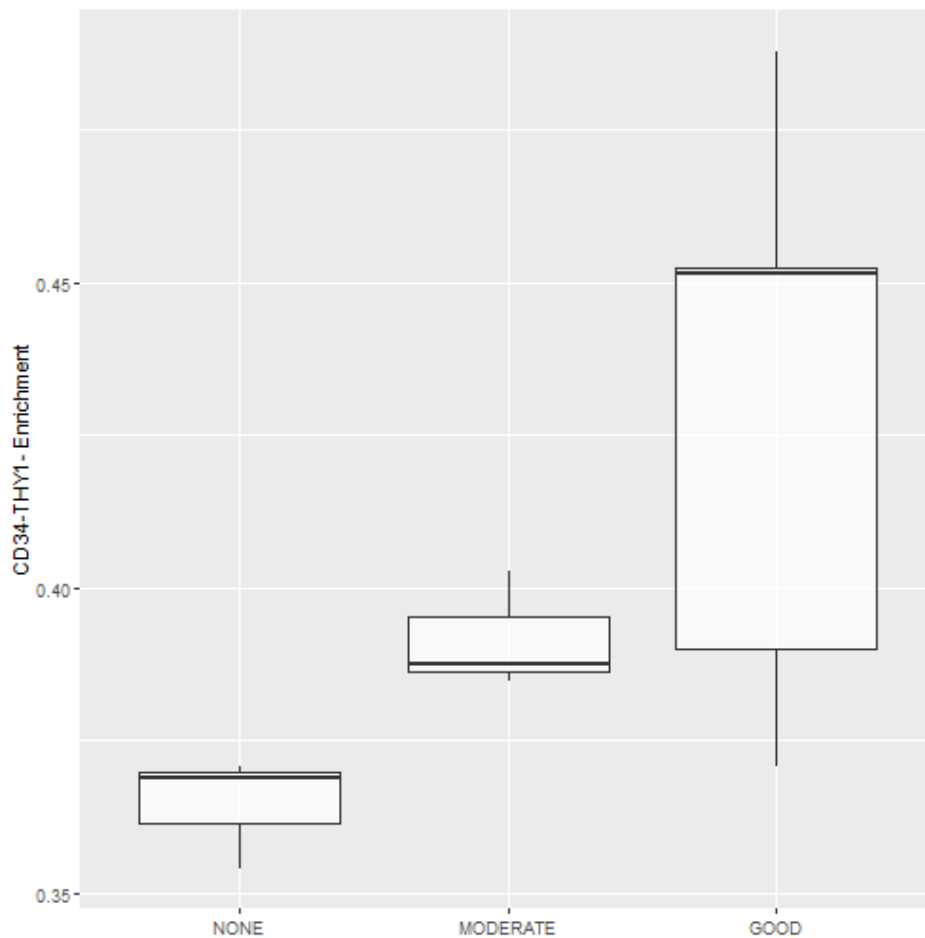


Plot of gene expression measures for *CTLA4* and *TIMELESS* as quantified using Illumina Beadchip microarrays and using Taqman quantitative RT-PCR. The high concordance between both platforms confirms the validity of the microarray gene expression quantification for accurate cell type deconvolution estimation.

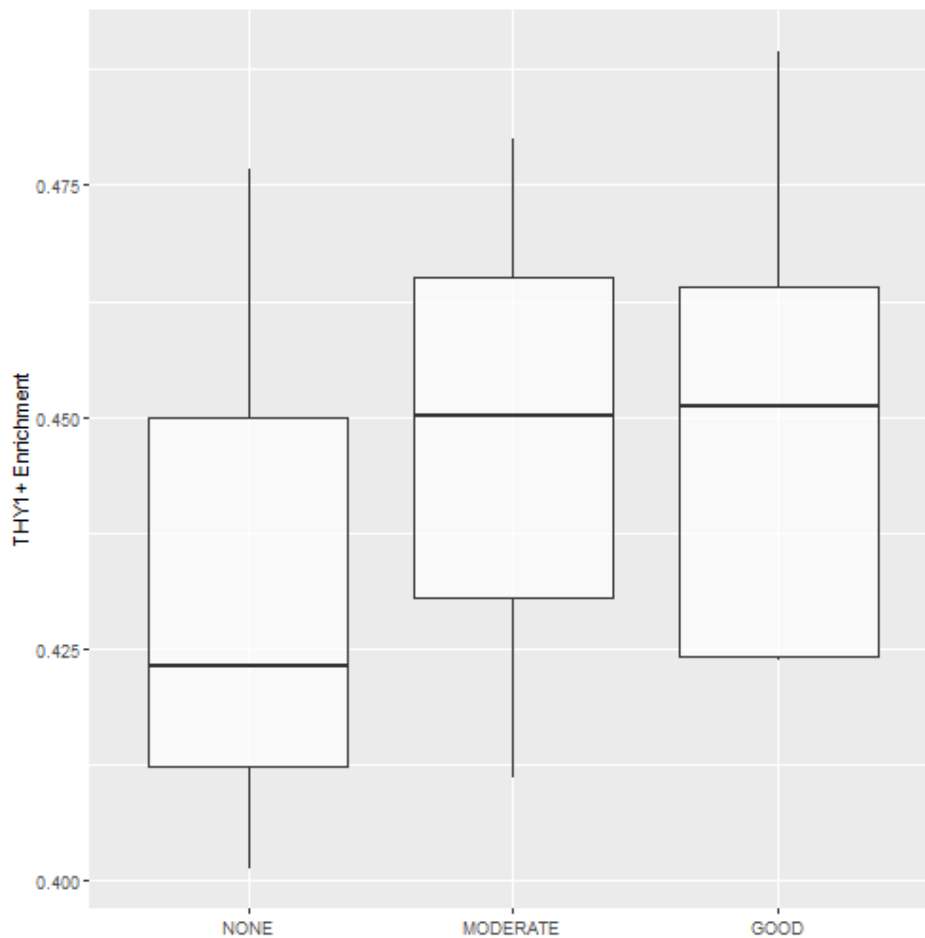
Supplementary Figure 4. Association of the three fibroblast subsets with anti-TNF response.



A. Boxplot of CD34+ fibroblast enrichment in the synovial membrane according to the response to anti-TNF therapy at week 20. Improvement in response increases linearly with the proportion of CD34+ fibroblast in the synovial membrane ($P=0.027$).

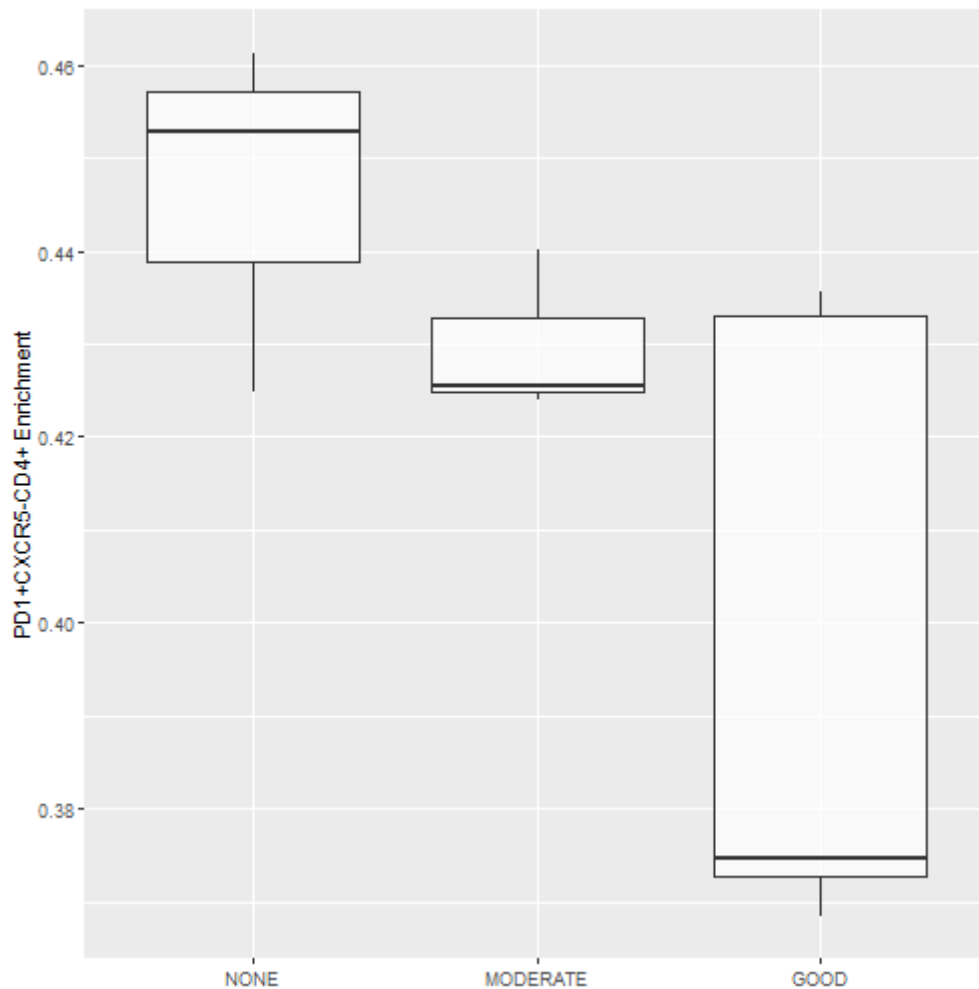


B. **Boxplot of CD34-THY1- fibroblast enrichment in the synovial membrane according to the response to anti-TNF therapy at week 20.** Improvement in response increases linearly with the proportion of CD34-THY1- fibroblasts in the synovial membrane ($P=0.021$).



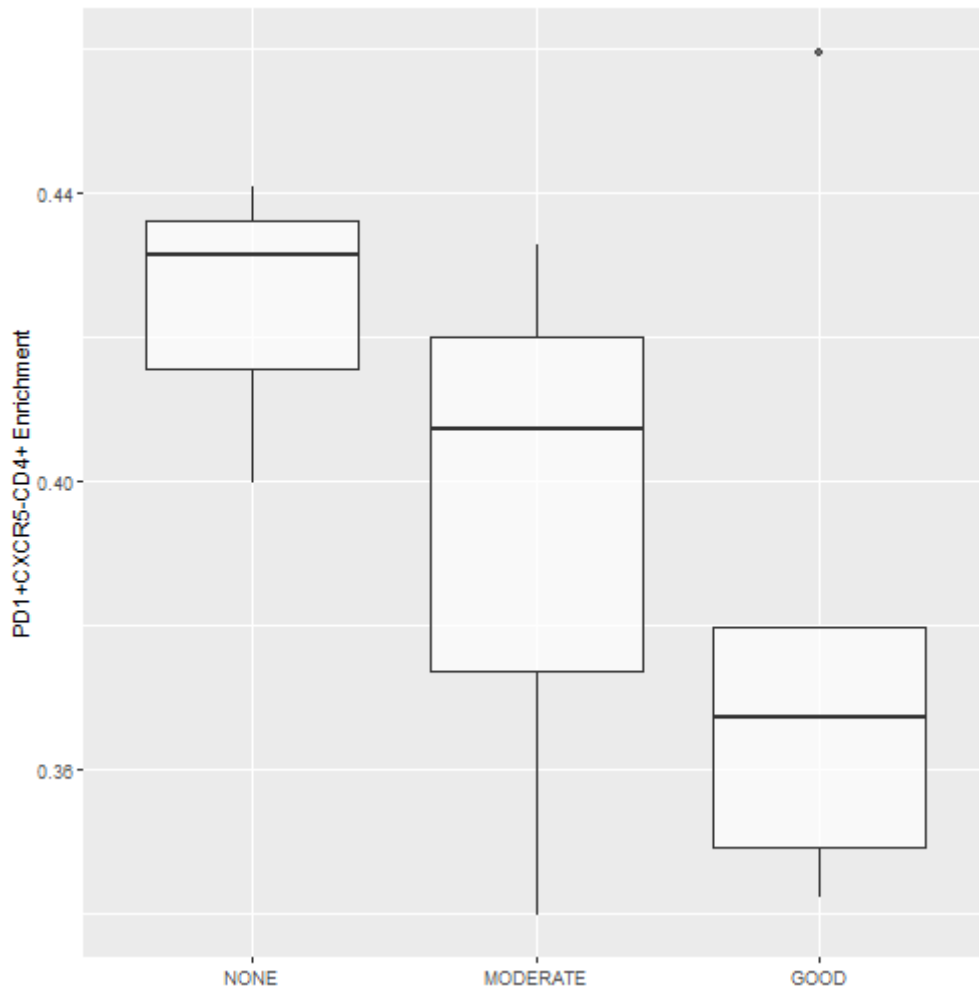
C. **Boxplot of THY1+ fibroblast enrichment in the synovial membrane according to the response to anti-TNF therapy at week 20.** The estimated proportion of THY1+ fibroblasts is not significantly associated with the response to anti-TNF therapy ($P=0.49$).

Supplementary Figure 5. Association of baseline PD1^{hi}CXCR5-CD4⁺ T cells with the response to anti-TNF therapy.



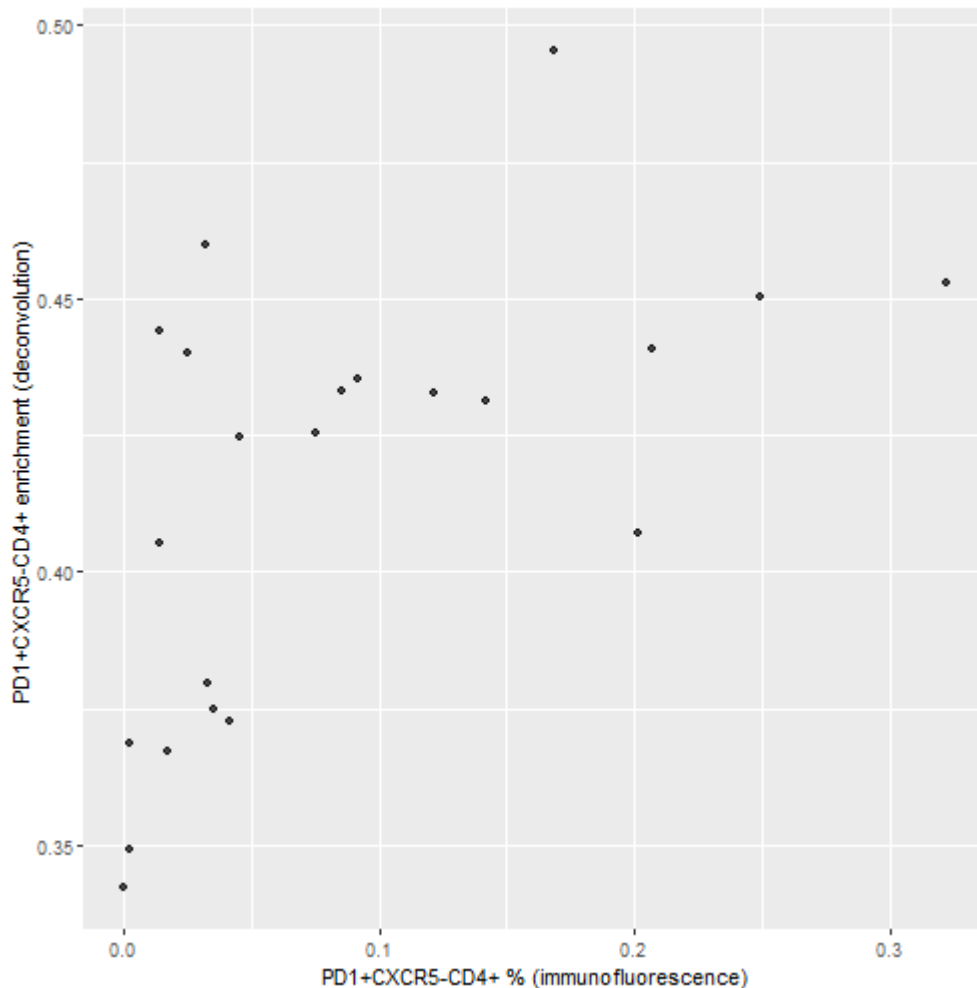
Boxplot of PD1^{hi}CXCR5-CD4⁺ T cell enrichment in the synovial membrane according to the response to anti-TNF therapy: week 0. A better response is associated with a lower proportion of peripheral helper T cells in the synovial membrane at baseline (P=0.022).

Supplementary Figure 6. Association of fibroblast types and T_{PH} cells with the response to anti-TNF therapy: week 20.



- A. **Boxplot of PD1^{hi}CXCR5-CD4⁺ T cell enrichment in the synovial membrane according to the response to anti-TNF therapy: week 20.** The trend between lower T_{PH} cells and a better response to anti-TNF therapy identified at baseline is maintained up to week 20 of therapy. Despite this, the association is no longer significant (P=0.29).

Supplementary Figure 6. Correlation between PD1^{hi}CXCR5-CD4⁺ T cell abundance estimation using the cell deconvolution approach with the percentage quantified using immunofluorescence.



The estimated proportion of T_{PH} cells in the synovial membrane using the cell-type deconvolution approach shows a high correlation with the percentage of cells directly quantified using immunofluorescence on the synovial membrane samples ($r^2=0.58$, $P=0.0051$). T_{PH} values for all $n=11$ for the two time points (baseline and wk20) are plotted. This result confirms the suitability of cell type deconvolution approach to estimate even low frequency cell types like T_{PH}.

Supplementary Table 4. Marker genes of the three synovial membrane fibroblast types.

CD34+ markers

<i>ADAM22</i>	<i>CACHD1</i>	<i>FZD4</i>	<i>PCDH18</i>
<i>ADAMTSL3</i>	<i>CAMK2D</i>	<i>G0S2</i>	<i>PKNOX2</i>
<i>AHR</i>	<i>CD34</i>	<i>GDA</i>	<i>RHOU</i>
<i>ANGPT1</i>	<i>CEP126</i>	<i>IQCC</i>	<i>SETBP1</i>
<i>ASPA</i>	<i>CLIP3</i>	<i>LPCAT2</i>	<i>SSPN</i>
<i>BICC1</i>	<i>CLLU1OS</i>	<i>MAB21L2</i>	<i>TBX5</i>
<i>C3</i>	<i>CP</i>	<i>MECOM</i>	<i>TMEM108</i>
<i>C6</i>	<i>CXCL12</i>	<i>NOVA1</i>	<i>TMEM133</i>
<i>C7</i>	<i>EGR1</i>	<i>OSR2</i>	<i>TRDMT1</i>

THY1+ markers

<i>ADCY7</i>	<i>CCNG1</i>	<i>FLRT2</i>	<i>PLXDC1</i>	<i>THY1</i>
<i>ADGRA2</i>	<i>COMP</i>	<i>FNDC1</i>	<i>PRKD1</i>	<i>TTC39C</i>
<i>ANKRD29</i>	<i>CRABP2</i>	<i>IGF1</i>	<i>PTGIS</i>	<i>VSTM4</i>
<i>ARL4C</i>	<i>CRISPLD2</i>	<i>LTBP2</i>	<i>SFRP4</i>	<i>WISP1</i>
<i>ARSB</i>	<i>CUTC</i>	<i>MBNL2</i>	<i>SMOC2</i>	
<i>ASPN</i>	<i>DPP4</i>	<i>MCUB</i>	<i>SRGAP1</i>	
<i>AXIN2</i>	<i>EDIL3</i>	<i>MYOF</i>	<i>STXBP5</i>	
<i>C12orf75</i>	<i>ELN</i>	<i>NLGN2</i>	<i>THBS1</i>	
<i>CADM1</i>	<i>FHL2</i>	<i>OAF</i>	<i>THBS2</i>	

CD34-THY1- markers

<i>ADAM10</i>	<i>ERRFI1</i>	<i>MET</i>	<i>PPIL6</i>	<i>TIMP3</i>
<i>ADGRA3</i>	<i>EZR</i>	<i>MFSD6</i>	<i>RBPM52</i>	<i>TMCO3</i>
<i>ADGRG2</i>	<i>FNIP2</i>	<i>MSN</i>	<i>RELL1</i>	<i>TMEM196</i>
<i>AGFG1</i>	<i>FOXO1</i>	<i>MT1G</i>	<i>RGS16</i>	<i>TSPAN15</i>
<i>BTC</i>	<i>GALNT12</i>	<i>MTHFD1L</i>	<i>RIOK3</i>	<i>TWISTNB</i>
<i>C10orf105</i>	<i>GFPT2</i>	<i>MTUS2</i>	<i>SEC11C</i>	<i>UBAP1</i>
<i>CLIP1</i>	<i>GPR18</i>	<i>NDP</i>	<i>SHTN1</i>	<i>UGP2</i>
<i>CNTNAP3</i>	<i>GPR183</i>	<i>NTN4</i>	<i>SIX3</i>	<i>USP24</i>
<i>CSN1S1</i>	<i>GPX3</i>	<i>OSTF1</i>	<i>SORBS2</i>	<i>UST</i>
<i>CUX1</i>	<i>GRAMD1B</i>	<i>PACSIN2</i>	<i>SOX5</i>	<i>VPS13A</i>
<i>CXCL8</i>	<i>HBEGF</i>	<i>PCSK6</i>	<i>STK38L</i>	<i>WARS</i>
<i>DAPK1</i>	<i>HMGA2</i>	<i>PDE8A</i>	<i>SUSD4</i>	<i>ZNF385B</i>
<i>DNASE1L3</i>	<i>HTRA4</i>	<i>PDLIM5</i>	<i>SV2B</i>	
<i>ENPP1</i>	<i>KIF1B</i>	<i>PGM3</i>	<i>SYT17</i>	
<i>EREG</i>	<i>LRRFIP2</i>	<i>PNP</i>	<i>TIAM2</i>	

Genes differentially overexpressed from each fibroblast subset compared to the other two were selected as markers for cell type deconvolution analysis.

Supplementary Table 5. Significantly overexpressed genes in PD-1^{hi} T cells according to Rao et al.

<i>PD-1</i>	<i>GZMK</i>	<i>F5</i>
<i>SCO2</i>	<i>DHFR</i>	<i>DUSP2</i>
<i>CDCA7</i>	<i>TIMELESS</i>	<i>MAF</i>
<i>TOX</i>	<i>DUSP4</i>	<i>PSMA4</i>
<i>TOX2</i>	<i>FABP5</i>	<i>SYT11</i>
<i>ENC1</i>	<i>FBXO41</i>	<i>UBE2A</i>
<i>HVCN1</i>	<i>MAP3K9</i>	<i>TBC1D4</i>
<i>CHN1</i>	<i>PMAIP1</i>	<i>UQCRC1</i>
<i>CCDC86</i>	<i>PRR5L</i>	<i>ANXA2</i>
<i>CCL5</i>	<i>MIS18BP1</i>	<i>SLAMF6</i>
<i>AKR1C3</i>	<i>EPSTI1</i>	<i>TUBB4B</i>
<i>CST7</i>	<i>BZRAP1</i>	<i>UBE2L6</i>
<i>CEP128</i>	<i>DPP3</i>	<i>ITM2A</i>
<i>MYL6B</i>	<i>TIGIT</i>	<i>RGS1</i>
<i>EZH2</i>	<i>FCRL3</i>	
<i>CXCR3</i>	<i>CTLA4</i>	
<i>ICA1</i>	<i>DDX54</i>	
<i>FANCI</i>	<i>RAB37</i>	
<i>FAM210A</i>	<i>SHMT2</i>	
<i>DIRC2</i>	<i>PFN1</i>	