

Supplementary Information

**Targeting PP2A-dependent autophagy enhances sensitivity to ruxolitinib in JAK2^{V617F}
myeloproliferative neoplasms**

Courdy et al.

Table of Contents :

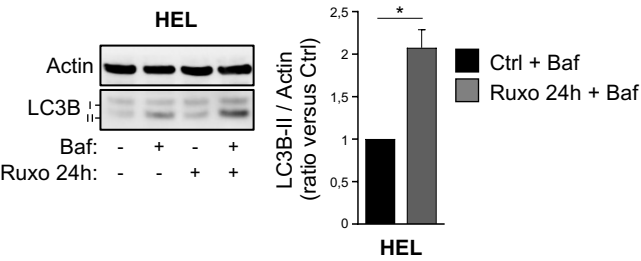
- Supplementary Table 1**
- Supplementary Figures 1- 4**

	ID study	From	JAK2 ^{V617F} allelic burden at diagnosis	Follow-up treatment
JAK2 ^{V617F} -positive MPN patients	12	Toulouse University Hospital	"Intermediate"	Phlebotomy
	13		45%	Phlebotomy
	16		60%	Phlebotomy
	17		36%	Phlebotomy
	18		23%	Phlebotomy
	19		45%	Phlebotomy
	20		13%	Phlebotomy
	22		45%	Phlebotomy
	23		87%	Phlebotomy
	29		85%	Phlebotomy + pegylated interferon
	31		43%	Phlebotomy
	32		<10%	Phlebotomy
	33		27%	Phlebotomy
	34		10%	Phlebotomy
	35		39%	Phlebotomy
	36		>50%	Phlebotomy
	37		21%	Phlebotomy
	38		23%	Phlebotomy
	39		9%	Phlebotomy
	40		<10%	Phlebotomy
	41		Not referred	Phlebotomy
	42		7%	Phlebotomy
JAK2 ^{WT} healthy donors	A	EFS, Toulouse	None	None
	B		None	None
	C		None	None
	D		None	None
	E		None	None
	F		None	None

Supplementary Table 1. Primary MPN patient and healthy donor characteristics.

Figure S1:

A



B

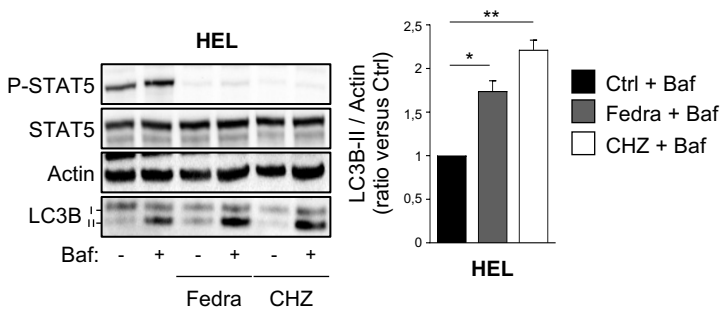


Figure S2:

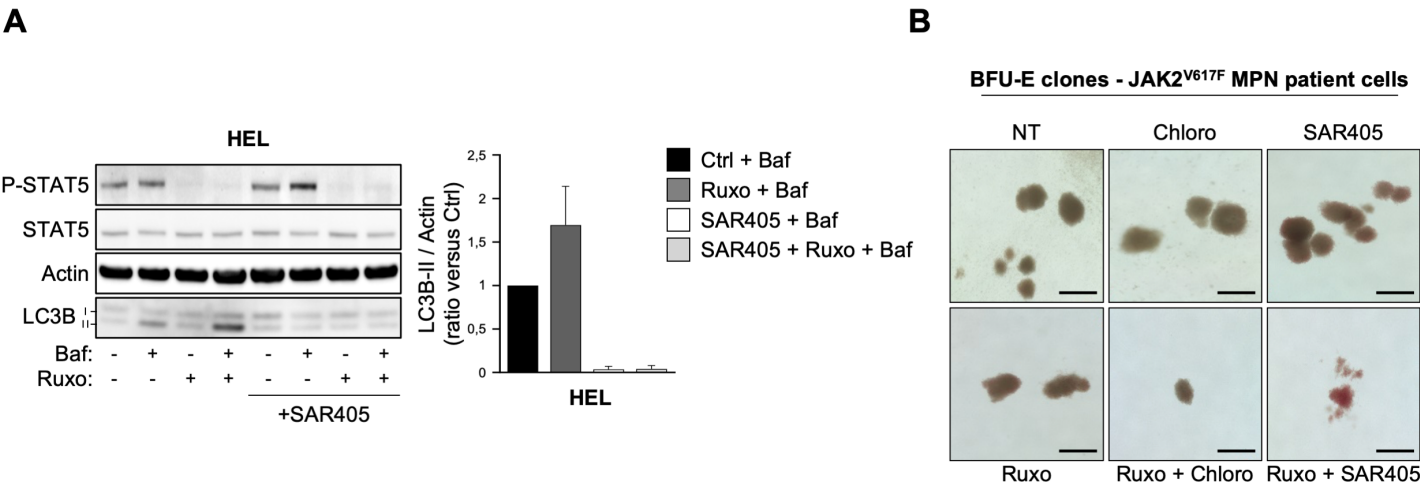


Figure S3:

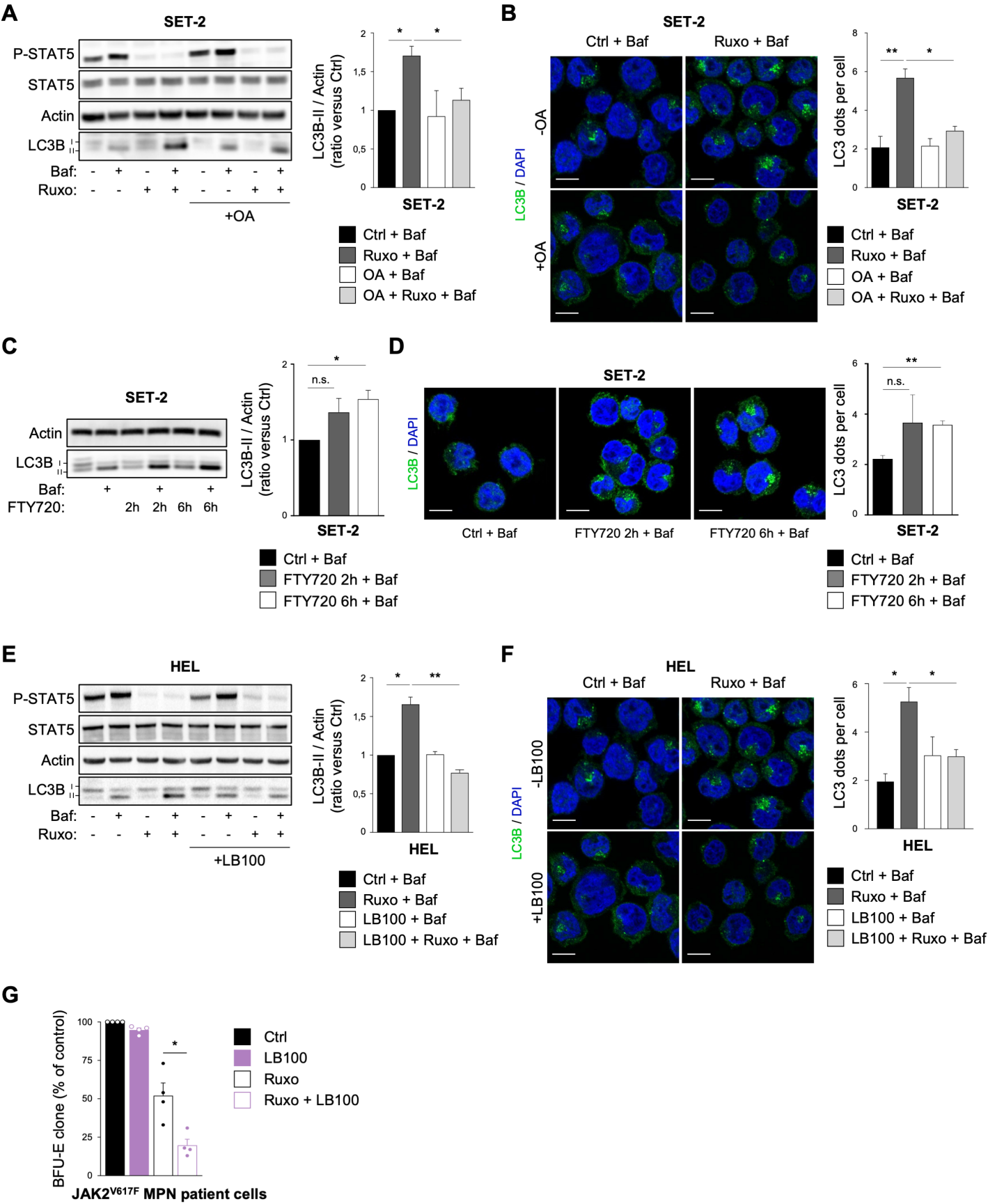


Figure S4:

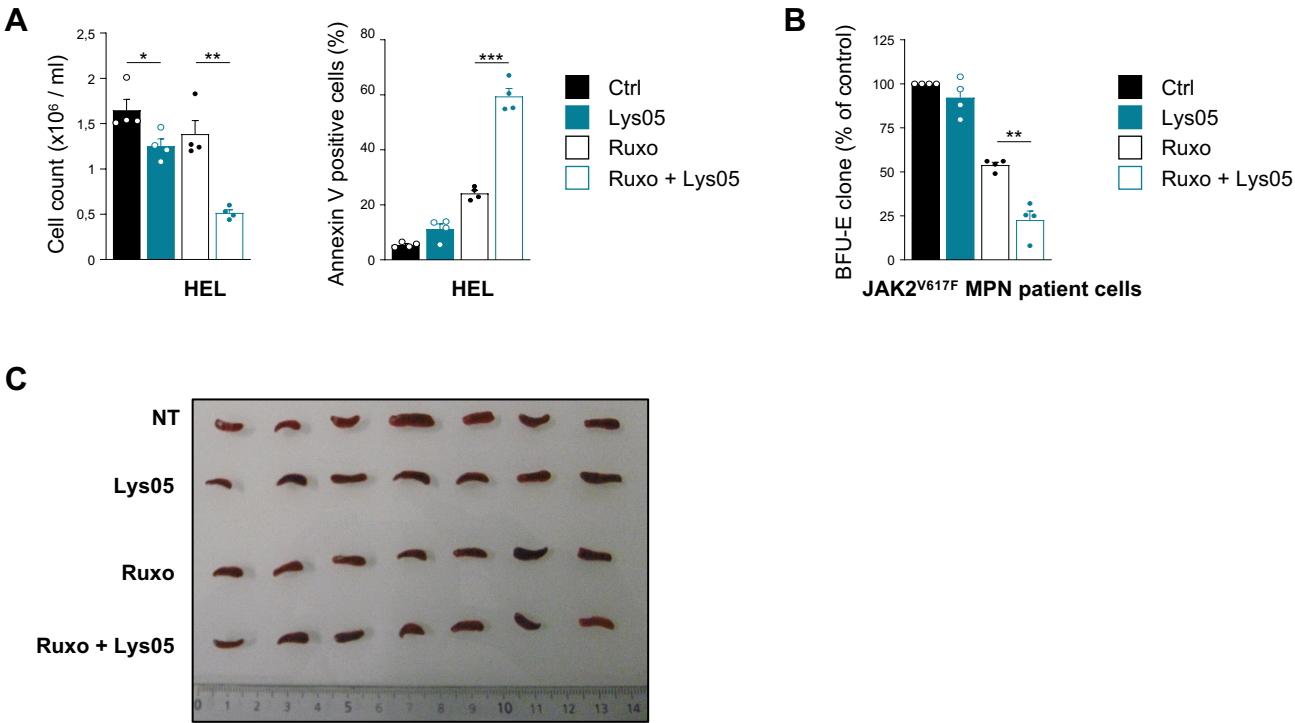


Figure S1. Ruxolitinib increases autophagy in JAK2^{V617F} cells. **A.** HEL cells were treated with ruxolitinib (1 μ M) for 24 hours in the presence or absence of bafilomycin (25nM) during 2 hours to monitor autophagy flux and processed for western blot analysis of LC3B. Actin was used as a loading control. Graphs represent the LC3B-II/actin ratios obtained by densitometric analysis ($n=3 \pm$ s.e.m.). **B.** HEL cells were treated with fedratinib (1 μ M) or CHZ868 (500nM) for 2 hours in the presence or absence of bafilomycin (25nM) to monitor autophagy flux and processed for western blot analysis of P-STAT5, STAT5 and LC3B. Actin was used as a loading control. Graphs represent the LC3B-II/actin ratios obtained by densitometric analysis ($n=3 \pm$ s.e.m.).

Figure S2. Autophagy inhibition enhances ruxolitinib efficacy in JAK2^{V617F} cells. **A.** HEL cells were treated with ruxolitinib (1 μ M) and SAR405 (3 μ M) for 2 hours in the presence or absence of bafilomycin (25nM) to monitor autophagy flux and processed for western blot analysis of P-STAT5, STAT5 and LC3B. Actin was used as a loading control. Graphs represent the LC3B-II/actin ratios obtained by densitometric analysis ($n=2 \pm$ s.e.m.). **B.** Representative picture of BFU-E clones after 14 days of CD34⁺ MPN patients' cells cultured in semi-solid medium supplemented or not with ruxolitinib (250nM) and chloroquine (3 μ M) or SAR405 (3 μ M). Scale bar: 300 μ m.

Figure S3. PP2A inhibition enhances ruxolitinib efficacy in JAK2^{V617F} cells. **A-B.** SET-2 cells were treated with ruxolitinib (1 μ M) and OA (5nM) for 2 hours in the presence or absence of bafilomycin (25nM) to monitor autophagy flux. Cells were processed for western blot analysis of P-STAT5, STAT5 and LC3B. Actin was used as a loading control. Graphs represent the LC3B-II/actin ratios obtained by densitometric analysis ($n=4 \pm$ s.e.m.), **(A)**. Cells were stained for LC3B and analyzed by confocal microscopy. Graphs represent the number of LC3B dots per cell ($n=4 \pm$ s.e.m.), **(B)**. Scale bar: 10 μ m. **C-D.** SET-2 cells were treated with FTY720 (2.5 μ M) for 2 or 6 hours in the presence or absence of bafilomycin (25nM) to monitor autophagy flux. Cells were processed for western blot analysis of LC3B. Actin was used as a

loading control. Graphs represent the LC3B-II/actin ratios obtained by densitometric analysis ($n=3 \pm \text{s.e.m.}$), (C). Cells were stained for LC3B and analyzed by confocal microscopy. Graphs represent the number of LC3B dots per cell ($n=3 \pm \text{s.e.m.}$), (D). Scale bar: 10 μm . E-F. HEL cells were treated with ruxolitinib (1 μM) and LB100 (2.5 μM) for 2 hours in the presence or absence of bafilomycin (25nM) to monitor autophagy flux. Cells were processed for western blot analysis of P-STAT5, STAT5 and LC3B. Actin was used as a loading control. Graphs represent the LC3B-II/actin ratios obtained by densitometric analysis ($n=3 \pm \text{s.e.m.}$), (E). Cells were stained for LC3B and analyzed by confocal microscopy. Graphs represent the number of LC3B dots per cell ($n=3 \pm \text{s.e.m.}$), (F). G. CD34⁺ cells from JAK2^{V617F} MPN patient samples were plated for colony forming assay in semi-solid medium supplemented or not with ruxolitinib (250nM) and LB100 (1 μM). After 14 days, clonogenic potential was assessed by counting the number of BFU-E clones per dish. Data are represented as percent of control ($n=4 \pm \text{s.e.m.}$).

Figure S4. Autophagy inhibitor Lys05 enhances ruxolitinib efficacy in JAK2^{V617F} cells *in vitro* and *in vivo*. A. HEL cells were treated or not with ruxolitinib (1 μM) in the presence or absence of Lys05 (5 μM). After 3 days, the number of cells was assessed by trypan blue exclusion counting (left panel; $n=4 \pm \text{s.e.m.}$) and the percentage of cell death was determined by Annexin-V labelling and flow cytometry analysis (right panel; $n=4 \pm \text{s.e.m.}$). B. CD34⁺ cells from JAK2^{V617F} MPN patient samples were plated for colony forming assay in semi-solid medium supplemented or not with ruxolitinib (250nM) and Lys05 (1 μM). After 14 days, clonogenic potential was assessed by counting the number of BFU-E clones per dish. Data are represented as percent of control ($n=4 \pm \text{s.e.m.}$). C. HEL cells were injected in NSG mice and, after 14 days of the indicated treatments their spleens were collected.