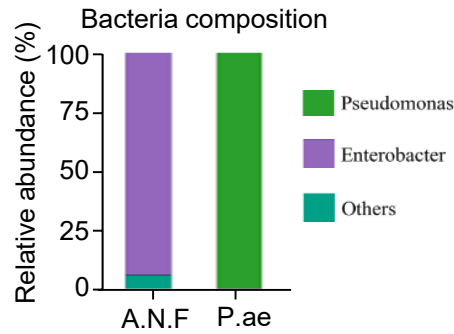
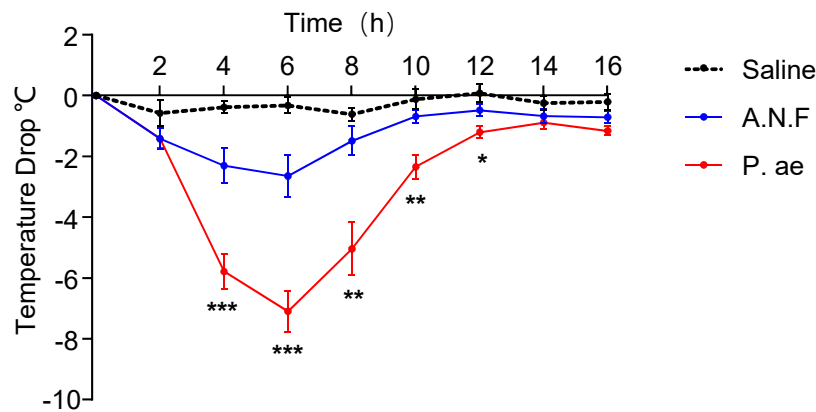


A



B



C

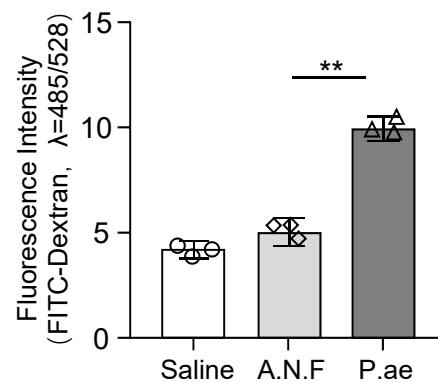


Figure S1. *P. ae* induced lung acute injury. (A) Bacteria composition in *P. ae* and A.N.F were detected by 16S rDNA sequencing. (B) C57BL/6 mice were intratracheally treated with *P. aeruginosa* (*P. ae*) or airway normal flora (A.N.F) (2×10^5 colony-forming units [CFUs] in a volume of 30 μ l per animal, $n = 5$) or same amount of sterile saline as control ($n = 3$) for 16 hours. Rectal temperatures at every two hours were detected. (C) C57BL/6 mice were infected with bacteria same as (B) for 16 hours, and then intratracheally treated with FITC-Dextran (10mg/kg body weight in 30 μ l PBS per animal) for 1 hours. Plasma samples were collected to detect fluorescence intensity ($\lambda = 485/528$, $n = 3$). Data in (B and C) are presented as means $\pm$ s.d. *** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$, as analyzed by two-tailed unpaired student's *t* test.

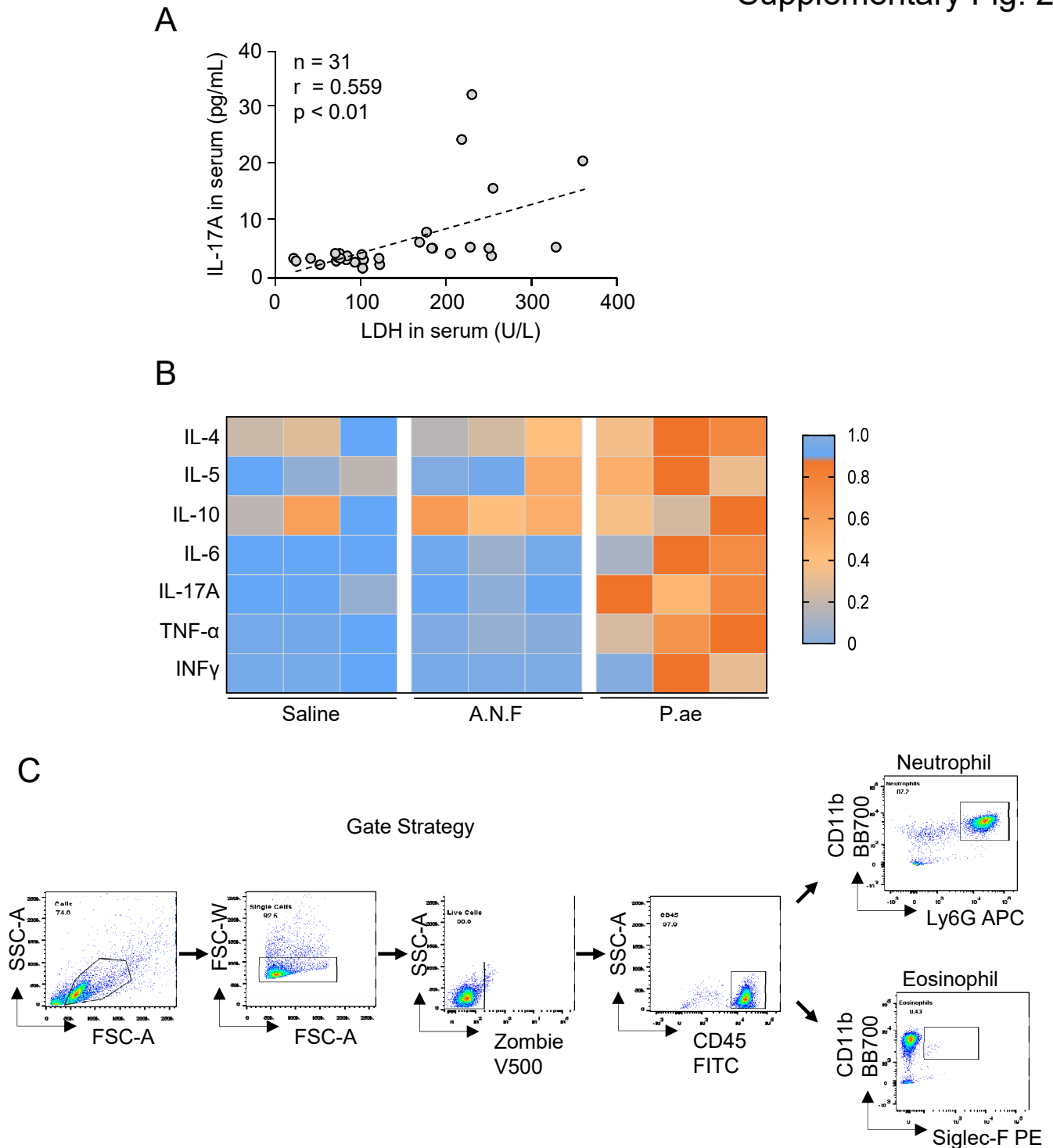


Figure S2. *P. ae* mediated lung inflammation. (A) The level of IL-17A and LDH in the serum of patients were detected by ELISA as mentioned in (Figure 1A and 2A). Linear correlation between IL-17A and LDH were analyzed by Pearson correlation coefficients test. (B and C) C57BL/6 mice were intratracheally treated with *P. ae*, A.N.F or sterile saline same as (Figure S1). Multiple cytokines (IL-4, IL-5, IL-10, IL-6, IL-17A, TNF- α , IFN γ) in BALF were detected through multi-analyte flow assay and presented in the Hot-map ($n = 3/\text{group}$) (B); Gate strategies of neutrophil and eosinophil through flow cytometry analysis (C).

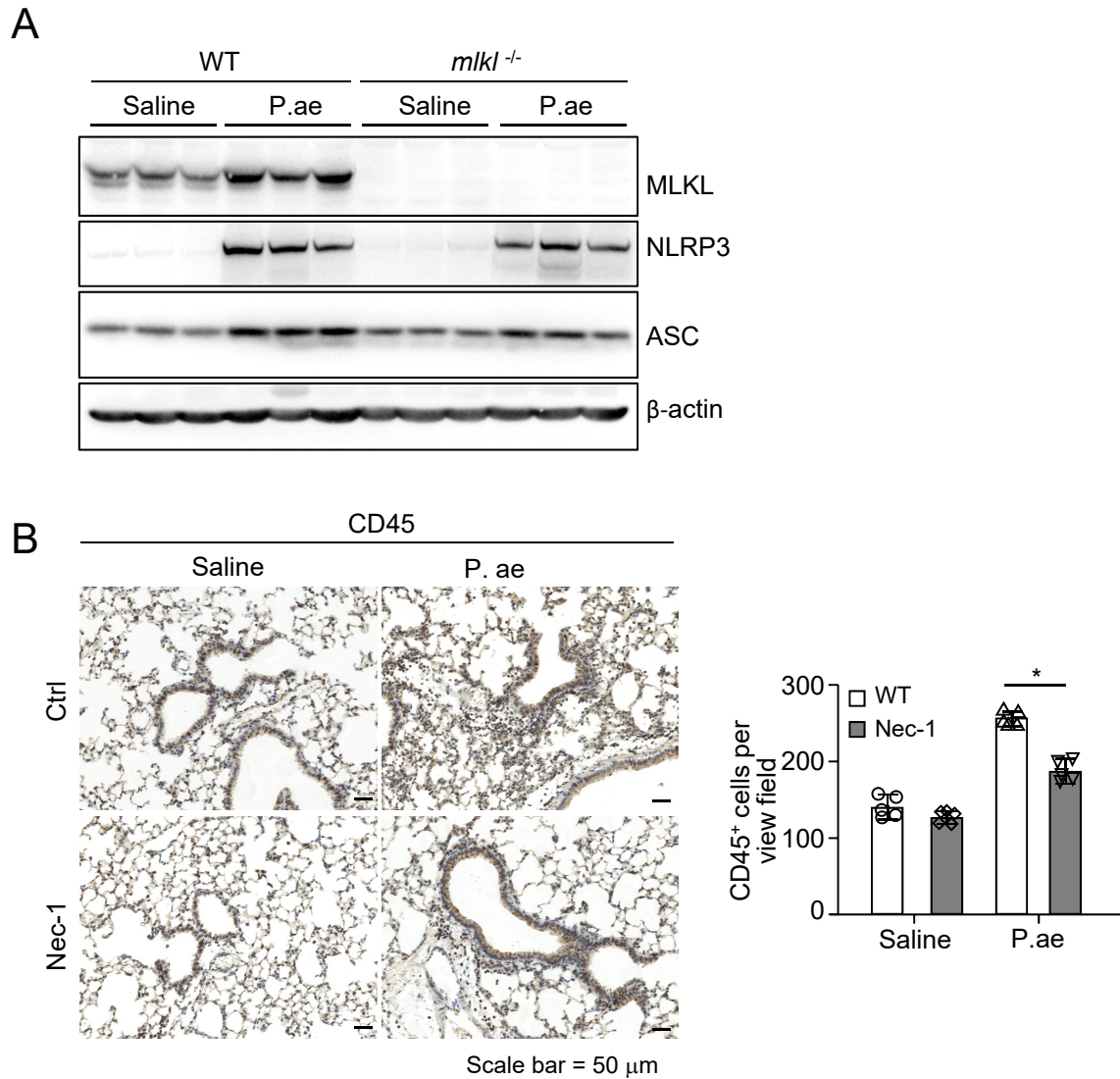


Figure S3. Pharmacological and genomic inhibition of necroptosis signaling ameliorates *P. aeruginosa* mediated acute lung injury and pulmonary inflammation. (A) *mlkl* deficient mice (*mlkl*^{-/-}) and WT littermates were intratracheally treated with *P. ae* (2×10^5 CFUs in 30 μ l per animal) or same amount of sterile saline as control for 16 hours. Immunoblot analysis of MLKL, NLRP3 and ASC level in the lungs as indicated, with β -actin as internal control. (B) C57BL/6 mice were intratracheally treated with Nec-1 (4 mg/kg body weight in 30 μ l 10 % DMSO solution per animal) or same amount of 10 % DMSO solution as control for 6 hours, and then intratracheally treated with *P. ae* (2×10^5 CFUs in 30 μ l per animal) or same amount of sterile saline as control for 16 hours (n = 5). IHC staining with anti-CD45 antibody in the representative lung sections as indicated; At least five random fields of each mouse were performed to analyzed the number of CD45⁺ cells through ImageJ program, and the scatter plot represents the average value of each sample. Data is presented as means $\pm$ s.d. * $p < 0.05$, as analyzed by two-tailed unpaired student's *t* test.

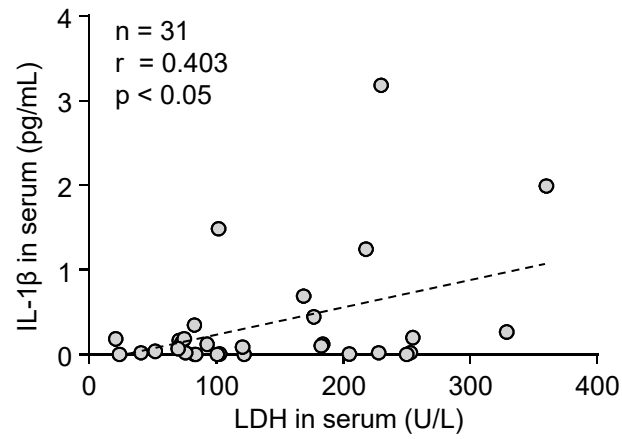


Figure S4. *P. ae* mediated NLRP3 inflammasome activation. The level of IL-1 β and LDH in the serum of patients were detected by ELISA as mentioned in (Figure 1A and 4A). Linear correlation between IL-1 β and LDH were analyzed by Pearson correlation coefficients test.

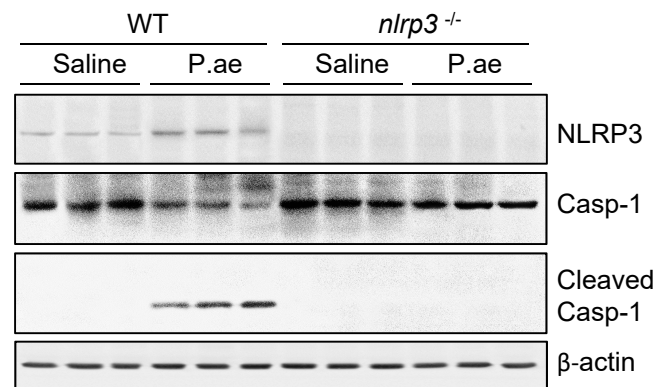


Figure S5. *nlrp3* ablation suppresses *P. aeruginosa* induced inflammasome activation. *nlrp3* deficient mice (*nlrp3*^{-/-}) and WT littermates were intratracheally treated with *P. ae* (2×10^5 CFUs in 30 μ l per animal) or same amount of sterile saline as control for 16 hours. Immunoblot analysis of NLRP3, Caspase-1 and cleaved caspase-1 level in the lungs as indicated, with β -actin as internal control.

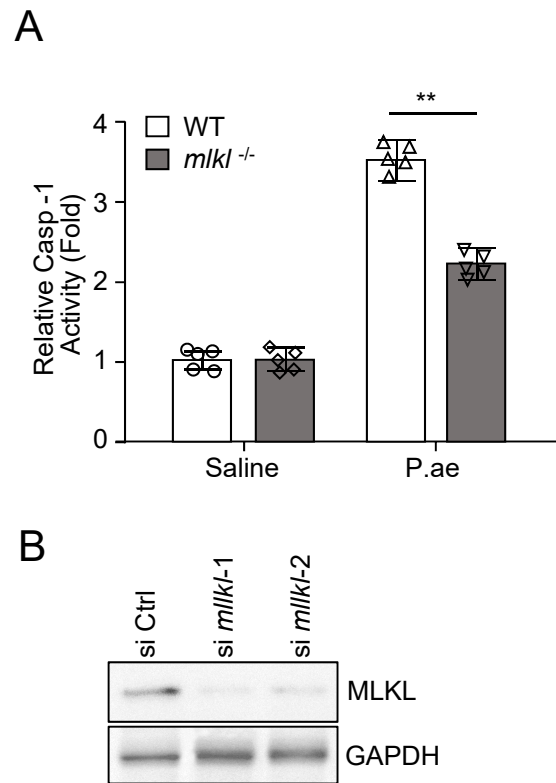
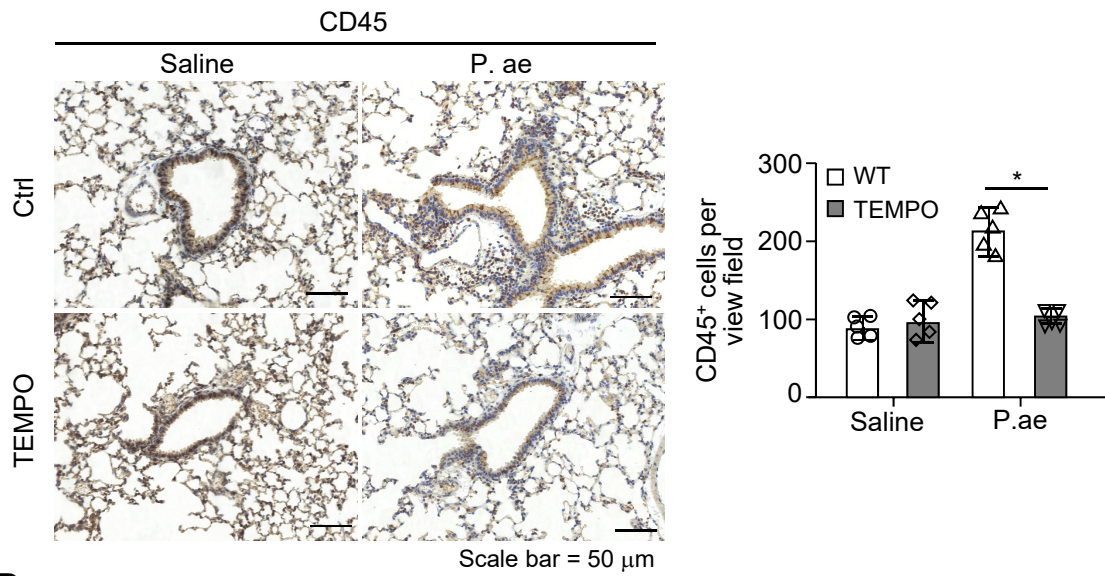
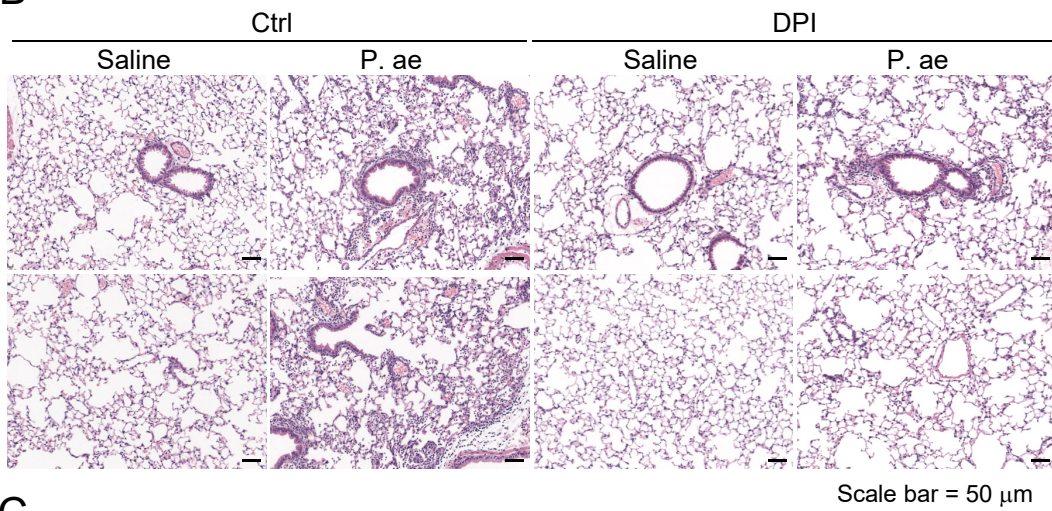


Figure S6. *mlkl* ablation suppresses *P. aeruginosa* induced inflammasome activation. (A) *mlkl* deficient mice (*mlkl*^{-/-}) and WT littermates were intratracheally treated with *P. ae* (2×10^5 CFUs in 30 μ l per animal) or same amount of sterile saline as control for 16 hours. Representative lung lysis were subjected to caspase 1 activity analysis. Data is presented as means $\pm$ s.d. ** $p < 0.01$, as analyzed by two-tailed unpaired student's *t* test. (B) A549 cells were transfected with scramble siRNA (si Ctrl) or two different target sites of si *mlkl* for 48 h. Protein level of MLKL was detected by immunoblotting, with GAPDH as internal control.

A



B



C

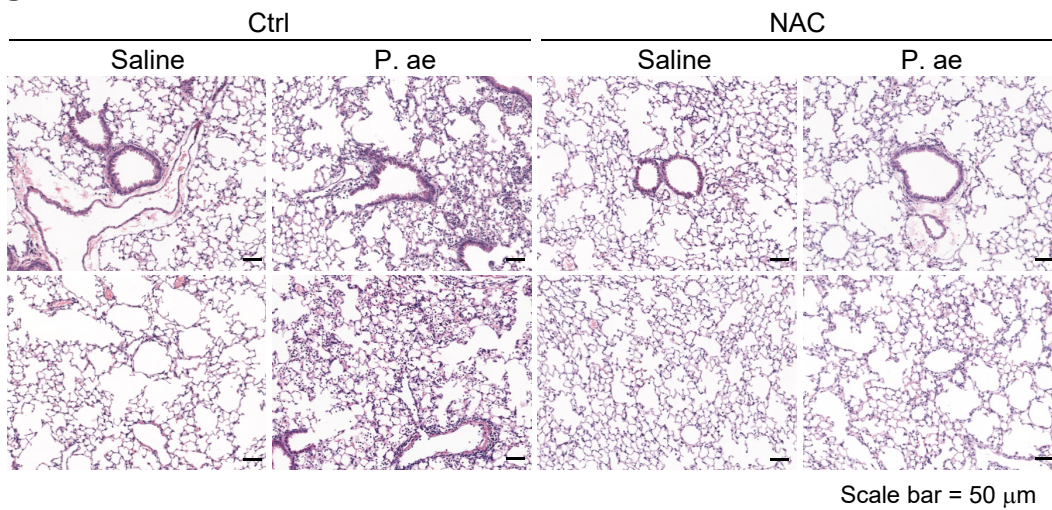


Figure S7. Antioxidants restrain *P. aeruginosa* induced acute lung injury. (A) C57BL/6 mice were intratracheally treated with Mito-TEMPO (20 mg/kg body weight in 30 μ l PBS per animal) or same amount of sterile saline as control for 6 hours, and then intratracheally treated with *P. ae* (2×10^5 CFUs in 30 μ l per animal) or same amount of sterile saline as control for 16 hours ($n = 5$). IHC staining with anti-CD45 antibody in the representative lung sections as indicated; At least five random fields of each mouse were performed to analyzed the number of CD45⁺ cells through ImageJ program, and the scatter plot represents the average value of each sample. Data is presented as means $\pm$ s.d. * $p < 0.05$, as analyzed by two-tailed unpaired student's *t* test. (B and C) C57BL/6 mice were intratracheally treated with DPI (1 mg/kg body weight in 10 % DMSO solution per mouse, B), NAC (125 mg/kg body weight in 30 μ l PBS per animal, C) or same amount of resolving buffer (10% DMSO or PBS) as control for 6 hours, followed with intratracheally treatment with *P. ae* (2×10^5 CFUs in 30 μ l per animal) or same amount of sterile saline as control for 16 hours ($n = 5$). H&E staining of the representative lung sections (Upper panel, bronchial area; Lower panel, alveolar area) from different treated mice as indicated.