

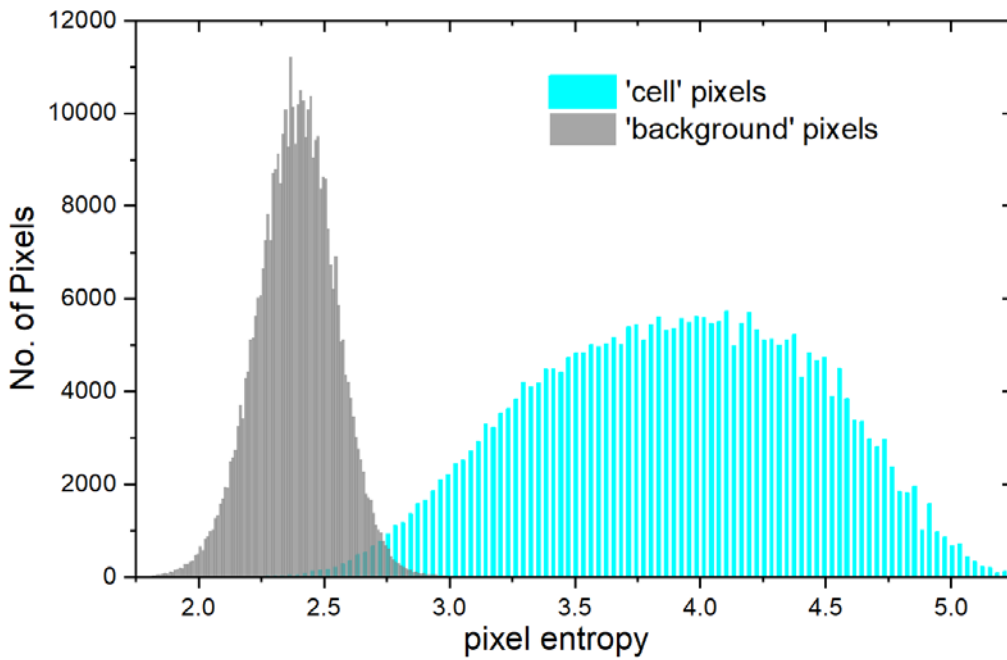
Supporting Information

A Self-Supervised Machine Learning Approach for Objective Live Cell Segmentation and Analysis

Michael C. Robitaille, Jeff M. Byers, Joseph A. Christodoulides, Marc P. Raphael

A. Static Feature Vectors

The self-supervised algorithm allows for automated training of as many static feature vectors as deemed helpful to robustly segmenting the cells from the background. In this work we utilized the `entropyfilt(I,nhood)` function in MATLAB to calculate the entropy of image *I* from a surrounding neighborhood of pixels, *nhood*, which was fixed to a 7 x 7 matrix. In a similar manner, we utilized the `imgradient(I,method)` function to assign a gradient to a given pixel based on 5 x 5 matrix neighborhood calculated with the 'intermediate' gradient operator method. An example of these feature vector histograms from *Dictyostelium* time course imagery are shown in Fig S1. Whether a pixel was labeled 'cell' or 'background' was determined by OF.



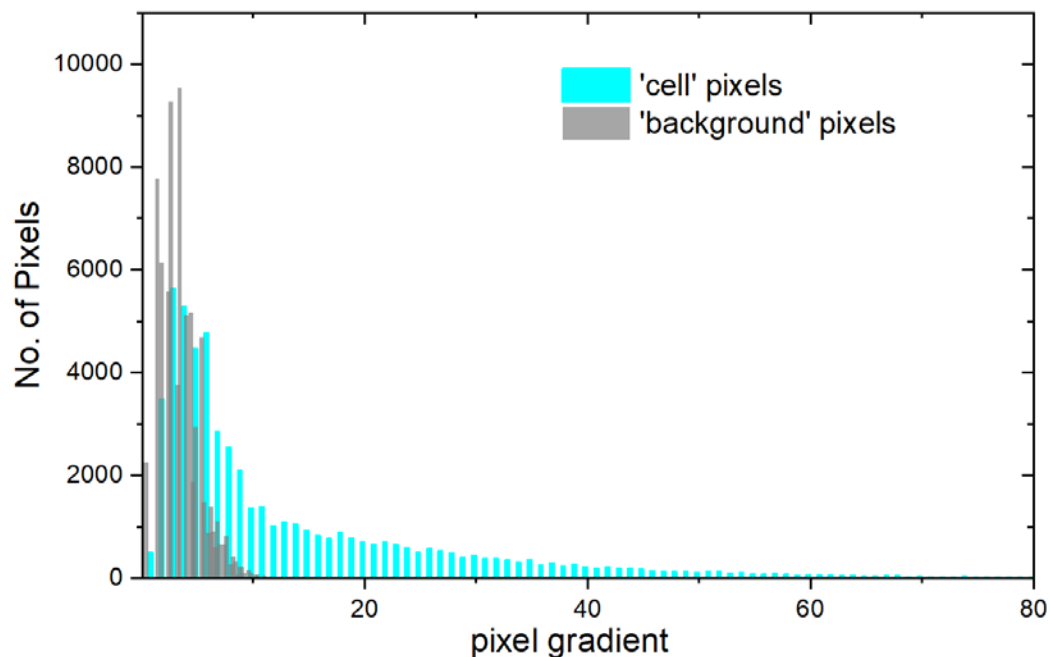


Fig S1. (top) Associated entropy feature vector histograms and (bottom) gradient feature vector histograms of the 'cell' and 'background' training pixels used for model training from transmitted light *Dictyostelium* images (10X objective).

B. Microscopy Details

Main Text Figure	Cell Type	Optical Modality	Objective Magnification	Objective Numerical Aperture	Camera	Wait Time [t-1, t] (seconds)
Figure 2	MDA-MB-231 (human breast adenocarcinoma)	DIC	20X	0.8 (air)	Zeiss Axiocam 702 CMOS	300
Figure 3a	Hs27 (human foreskin, fibroblast)	Phase	10X	0.45 (air)	Hamamatsu ORCA R2 CCD	1200
Figure 3b	Dictyostelium Discoideum (amoeboid)	TL	10X	0.3 (air)	Zeiss Axiocam 702 CMOS	60
Figure 3c	MDA-MB-231 (human breast adenocarcinoma)	Phase	10X	0.3 (air)	Zeiss Axiocam 702 CMOS	600
Figure 3d	Hs27 (human foreskin, fibroblast)	IRM	40X	1.4 (oil)	Hamamatsu ORCA R2 CCD	600
Figure 3e	MDA-MB-231 (human breast adenocarcinoma)	DIC	20X	0.8 (air)	Zeiss Axiocam 702 CMOS	120
Figure 3f	A549 (human lung adenocarcinoma)	Fluorescence	100X	1.46 (oil)	iXon Ultra EMCCD	10
Figure 4	MDA-MB-231 (human breast adenocarcinoma)	Phase	10X	0.3 (air)	Zeiss Axiocam 702 CMOS	600

Table S1 Cell and optical details for imagery in the main text. DIC: Differential Interference Contrast; Phase: Phase Contrast; TL: Transmitted Light illumination; IRM: Interference Reflection Microscopy.

C. Segmented Cell Area as a function of Smoothing Disk Size

Hole filling was accomplished with the `imclose(I,SE)` function in Matlab in which `I` is the binary input image with 'cell' pixel values 1 and 'background' pixel values 0, and `SE` is structural element whose size and shape determines the extent of the blurring. We used a disk `SE` of radius r and by plotting the average segmented cell area in the field of view as a function of r we determined a range in which the area output remained relatively constant across all cell types and optical modalities. **Fig. S2a** shows the mean segmented area of Hs27 cells (phase contrast, 10X magnification) versus r . From $1 < r < 11$ the segmented area is stabilized, while above $r = 11$ discrete steps in area are observed as a result of cells being grouped together as shown in the cell segmentation images of **Fig. S2b** for $r = 5$ pixels versus **Fig. S2c** for $r = 20$ pixels. The $r = 5$ pixel value was found to reliably hole fill for all cells and optical modalities in this study.

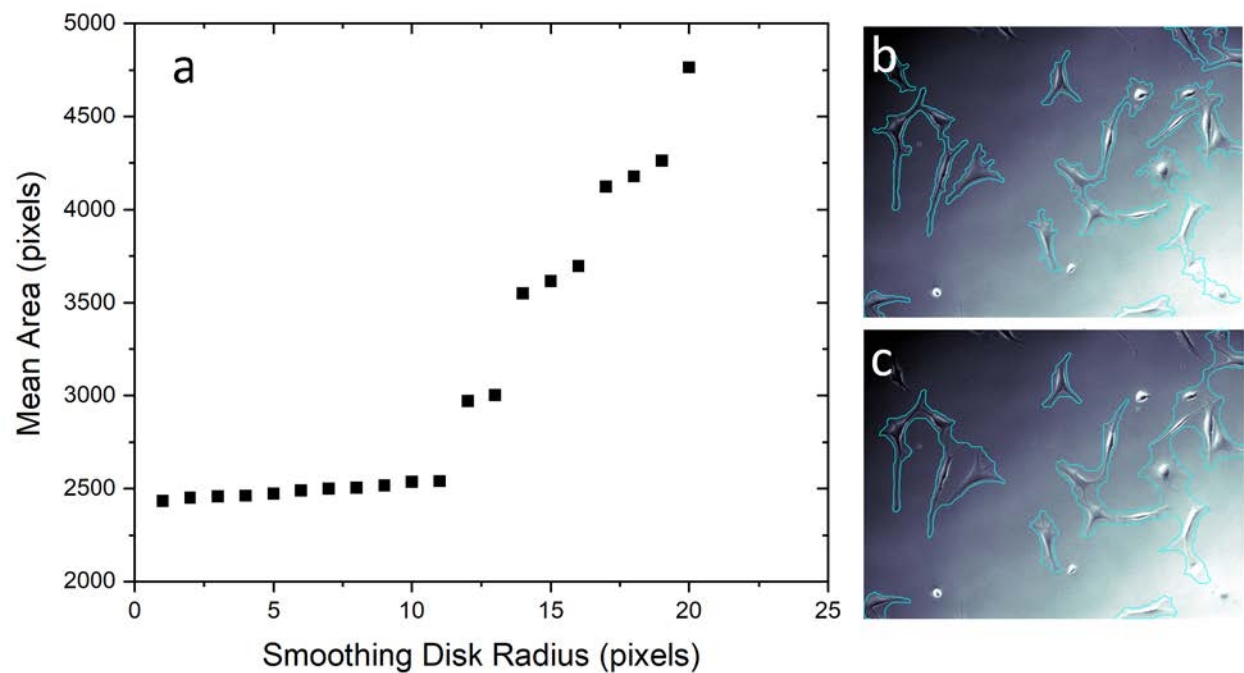


Fig S2. a. Mean area of the segmented cells in the field of view as a function of the smoothing disk radius. Segmented images of cells with **b.** $r = 5$ pixels and **c.** $r = 20$ pixels.