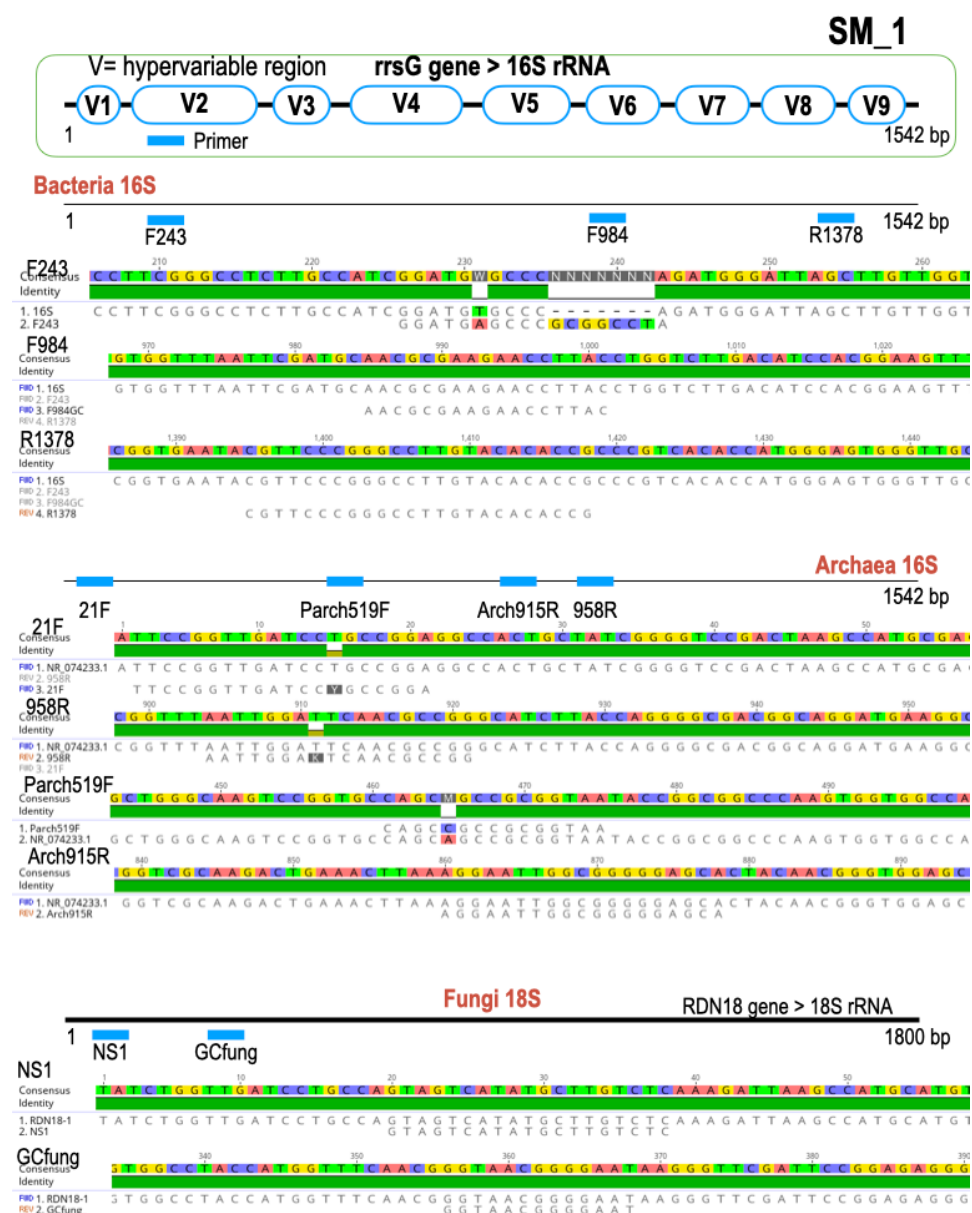


Online Resources 1

Archaea, the unseen kingdom in the gut microbiome of *Anastrepha obliqua*Amores GR¹, Zepeda-Ramos G¹, García-Fajardo LV¹, Hernandez Emilio², Guillén-Navarro K¹.¹ Laboratorio de Biotecnología Ambiental y Agroecológica, Grupo Académico de Biotecnología Ambiental, El Colegio de la Frontera Sur (ECOSUR), Tapachula, Chiapas, Mexico.² Programa Moscafrut DGSV-SENASICA-SAGARPA, Subdirección de Desarrollo de Métodos, Chiapas, Mexico.*Corresponding author: E-mail: kguillen@ecosur.mx

Hypervariable boundaries are illustrative regarding the gene

Online Resource 1. Primer annealing and PCR conditions.

The reaction mixture contained 0.20 pmol of each primer, 1x buffer, 0.20 mM dNTPs (dinucleotide triphosphate), 1.5 mM MgCl₂ (2 mM Fungi), 1 U *Taq* DNA polymerase (Invitrogen, USA), and template DNA for a total volume of 20 µl. Amplification protocol in bacteria consisted in a denaturation at 94 °C for 5 min, followed by 30 cycles of 1 min at 95 °C, 1 min at 58 °C and 40 sec at 72 °C, with a final extension at 72 °C for 5 min. The Gram positive (actinomycetes) nested amplification used the same protocol for second amplification; the first annealing temperature was modified at 53 °C (Das, Royer, and Leff, 2007). Archaea nested PCR protocol for the first reaction was: denaturation at 94 °C for 5 min, followed by 30 cycles of 40 sec at 94 °C, 40 sec at 53 °C, 1 min at 72 °C, and a final extension at 72 °C for 5 min. For the second reaction, the alignment temperature was changed to 57 °C. Fungi PCR conditions consisted of denaturation at 95 °C for 2 min, followed by 35 cycles of 30 sec at 95 °C, 30 sec at 50 °C, and 1 min at 72 °C, with a final extension at 72 °C for 5 min (Nikolcheva, Cockshutt, and Bärlocher, 2003). Controls were designed as follow, to confirm the absence of Gram positive (actinomycetes) in PCR negative samples, we determined the minimum amount of *Streptomyces* sp DNA that renders an amplicon using the corresponding primers. This PCR was done using serial dilutions (14 ng to 0.0014 ng) of DNA. Negative control included ultrapure water, whereas *Escherichia coli* DH5-α, *Streptomyces* sp., and *Trichoderma* sp., DNA were used as positive controls for bacteria, actinomycete, and fungi, respectively. Total DNA from an anaerobic plant sludge wastewater treatment was used as PCR positive control. For alignments, *rrsG* sequence was downloaded from <https://ecocyc.org/> for *E. coli* K12 MG1655; 16S rRNA sequence was downloaded at <https://www.ncbi.nlm.nih.gov/> using the accession number NR_074233.1 *Methanocaldococcus jannaschii*; and RDN18 gene sequence of *Saccharomyces cerevisiae* was downloaded at <https://www.yeastgenome.org/locus/S000006482>. Images were generated using free license of Geneious (Kearse, et al., 2012). Nucleotide mismatches at F243 is due to the 16S target employed for the alignment (i.e., *E. coli*) showing the region to identify Gram-positive bacteria.