

The critical role of polyketide synthase gene on the swainsonine biosynthesis in the fungus *Metarhizium anisopliae*

Lu Sun^{1#}, Enxia Huang^{1#}, Yu Zhang¹, Ziyu Guo¹, Kexin Wu¹, Yunhao Zhang¹, Chonghui Mo², Jinglong Wang³, Baoyu Zhao¹, Hao Lu^{1*}

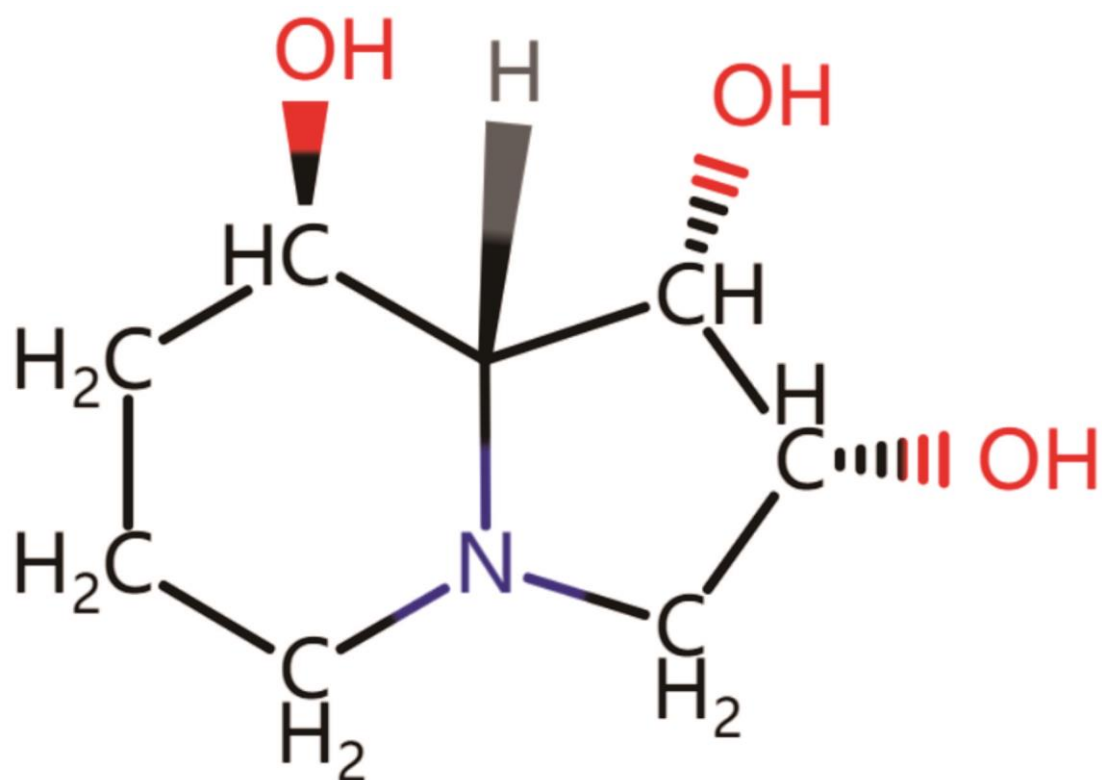
1 College of Veterinary Medicine, Northwest A&F University, Yangling, Shaanxi, 712100, China

2 College of Agriculture and Animal Husbandry, Qinghai University, Xining, Qinghai, 810016, China

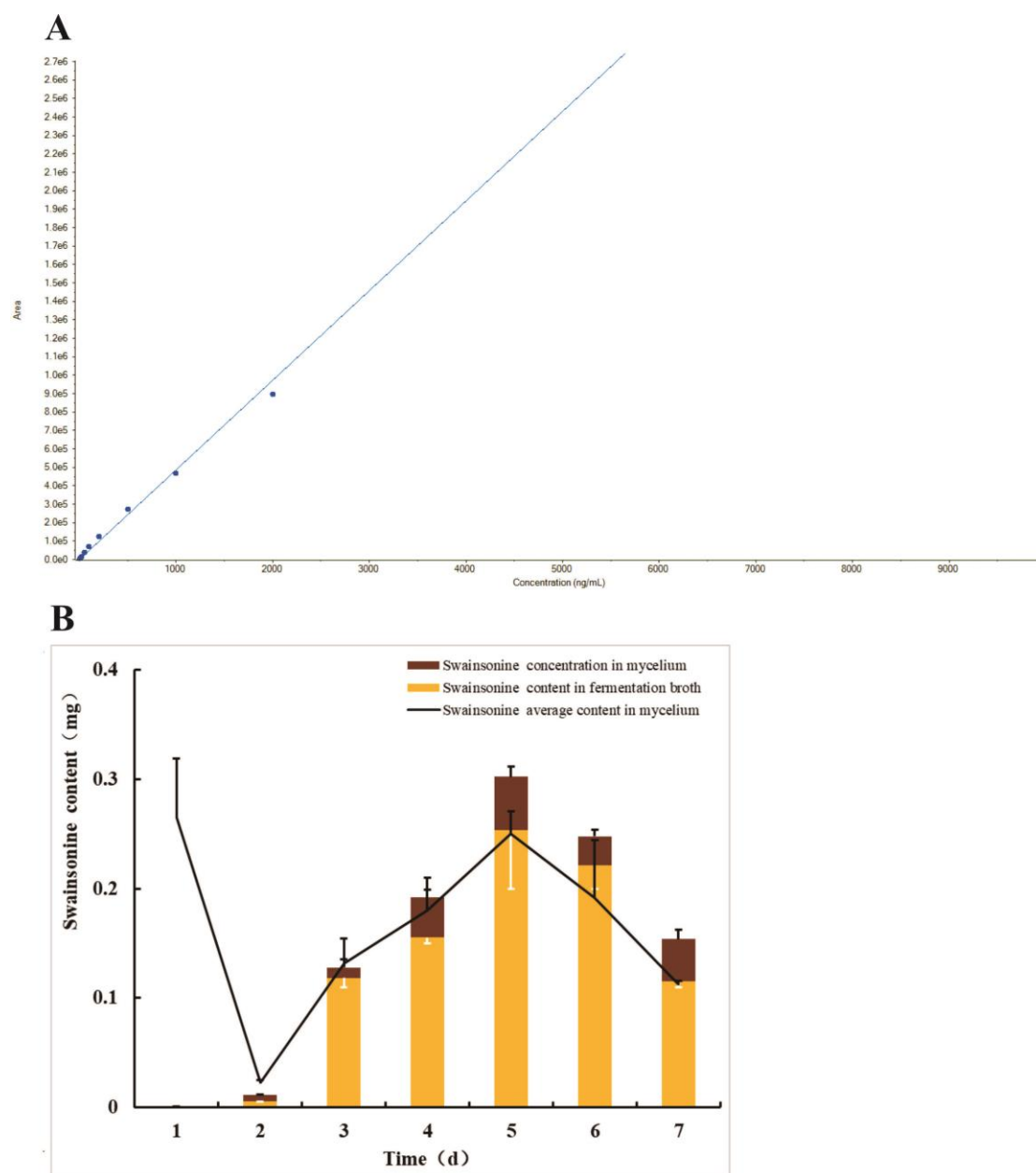
3 Tibet Academy of Agricultural and Animal Husbandry Sciences/State Key Laboratory of Barley and Yak Germplasm Resources and Genetic Improvement, Lhasa, Tibet, 850002, China

*Corresponding author. Lu H: E-mail: luhao@nwsuaf.edu.cn; Tel: +86-29-87092429; Fax: +86-29-87091032

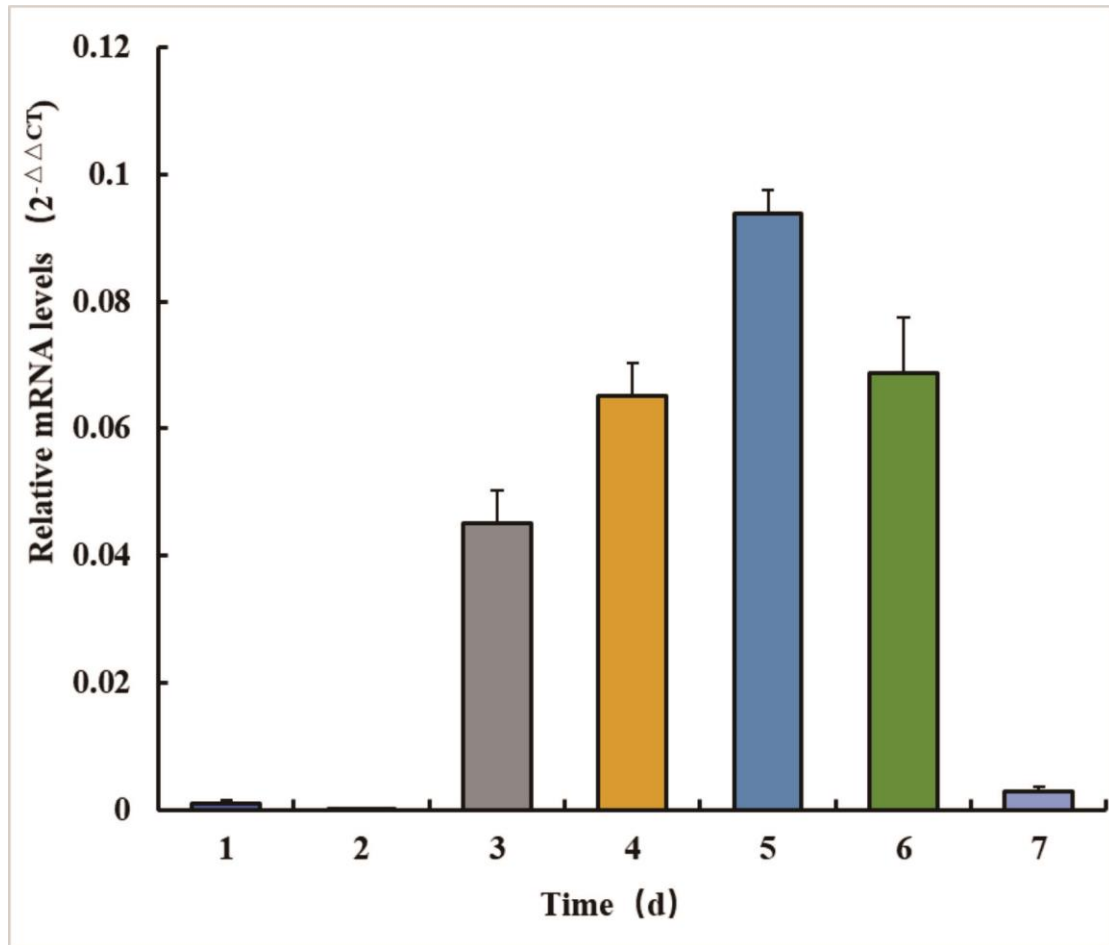
#These authors contributed equally to this work.



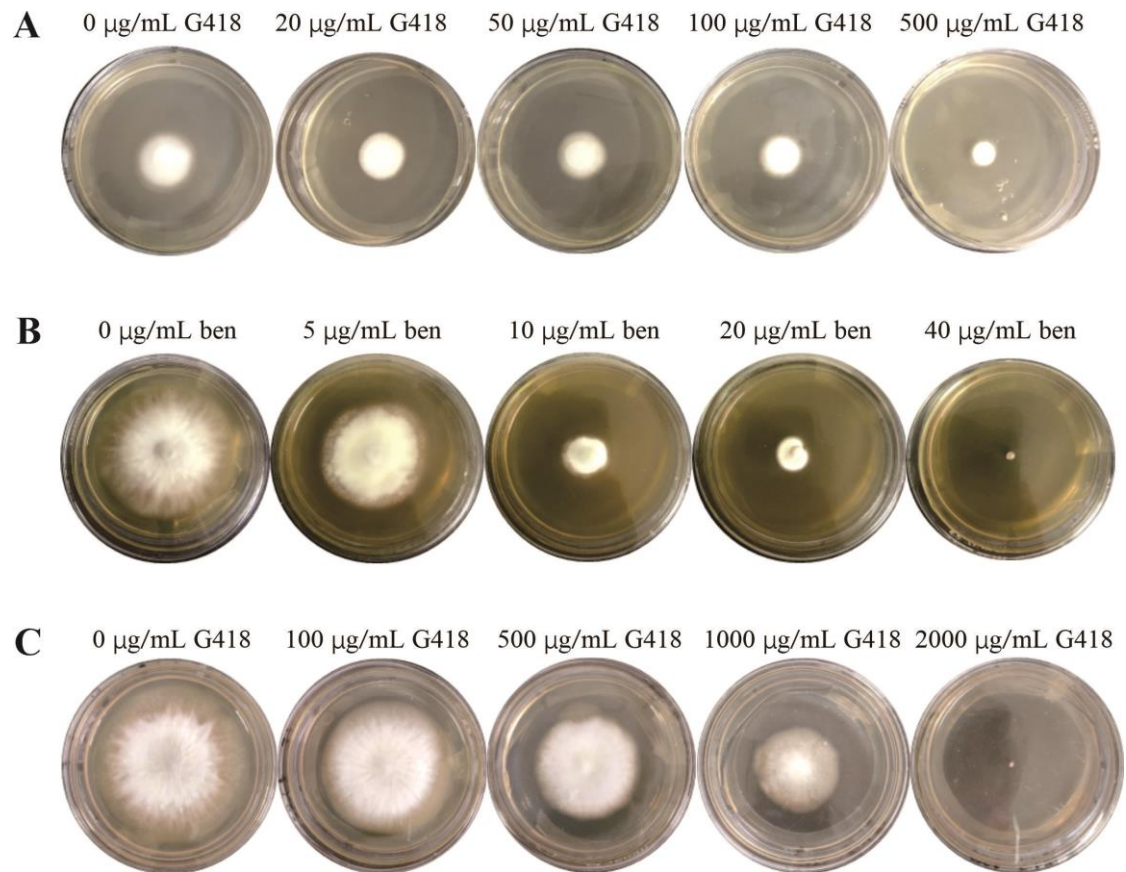
Supplementary Figure 1. Swainsonine chemical structure.



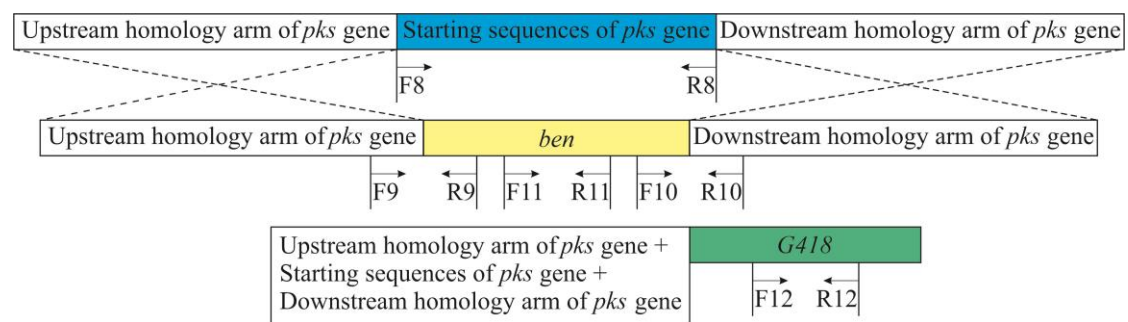
Supplementary Figure 2. Detection of SW content of *M. anisopliae* in different time. A: The standard curve was drawn according to the calculated regression equation: $Y=486.14665X+34.36851$ ($r=0.99049$). B: The content of swainsonine of *M. anisopliae* from 1st to 7th d. Error bars represent the standard error of the mean ($n = 3$).



Supplementary Figure 3. RT-qPCR detection of mRNA content of *M. anisopliae* in different time. Error bars represent the standard error of the mean (n = 3).

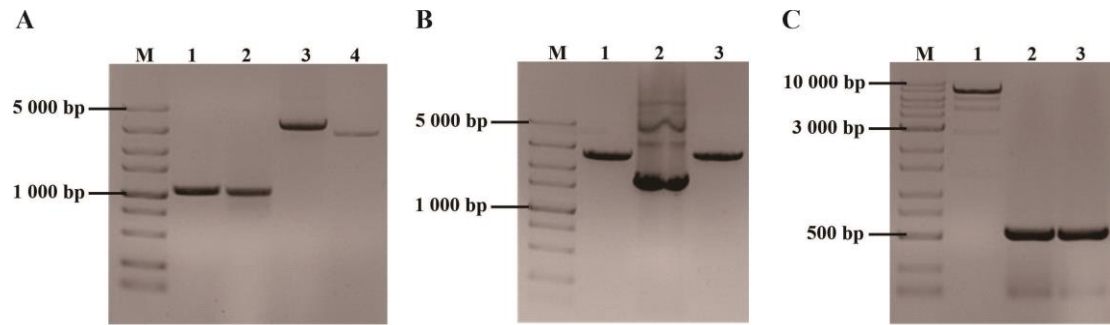


Supplementary Figure 4. Sensitivity Screening of *M. anisopliae*. A: The sensitivity of *M. anisopliae* to different concentrations of G418; B: The sensitivity of *M. anisopliae* to different concentrations of benomyl; C: Sensitivity of *pks* gene MT of *M. anisopliae* to different concentrations of G418.

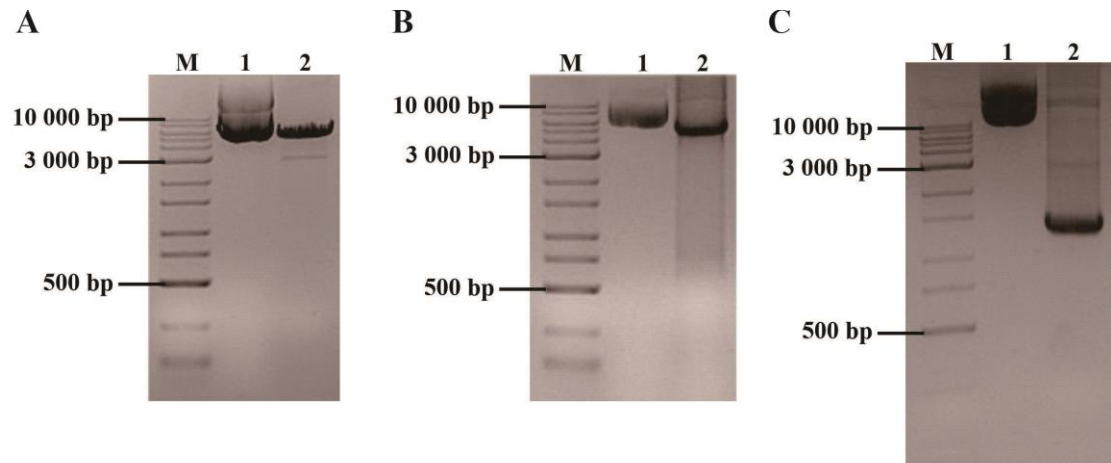


Supplementary Figure 5. Identification figure for *pks* gene MT, CT and RNAi strain.

the genomic DNA of *M. anisopliae*. The *G418* gene was amplified using primers L7/R7 from pSilent-Dual1. To produce a complementation vector, the *pks* gene and *G418* gene were inserted the MCS of pUC19 vector using In-Fusion cloning. C: The *PKS-1* (500 bp) and *PKS-2* (500 bp) fragment were amplified using primers L4/R4, L5/R5 respectively from the genomic DNA of *M. anisopliae*. To produce a RNAi vector, the *PKS-1* and *PKS-2* fragment were inserted the MCS1 and MCS2 of pSilent-1 vector respectively using In-Fusion cloning.



Supplementary Figure 7. Fragments of *pks* gene mutation vector, complementary vector and RNAi vector of *M. anisopliae*. A: Fragments of *pks* gene mutation vector, lane1: Upstream homology arm of *pks* gene (*PKS-I*: 998 bp), lane 2: Downstream homology arm of *pks* gene (*PKS-II*: 1,000 bp), lane 3: *Ben* gene (3,100 bp), 4: pUC19 plasmid digestion fragment; B: Fragments of *pks* gene complementary vector, lane 1: pUC19 plasmid digestion fragment, lane 2: *G418* gene (1,700 bp), lane 3: Upstream homology arm + *pks* starting sequence + downstream homology arm; C: Fragments of *pks* gene RNAi vector, lane 1: pSilent-1 plasmid digestion fragment, lane 2: *PKS-1* fragment (500 bp), lane 3: *PKS-2* fragment (500 bp).



Supplementary Figure 8. *Pks* gene mutation vector, complementary vector and RNAi vector of *M. anisopliae* and its specific fragments. A: *Pks* gene mutation vector and its specific fragments, lane 1: *Pks* gene mutation vector, lane 2: Specific fragments (Upstream homology arm + *ben* gene +Downstream homology arm of *pks* gene) (5,100 bp); B: *Pks* gene complementary vector and its specific fragments, lane 1: *Pks* gene complementary vector; lane 2: Specific fragments (*G418* gene + *PKS-3*) (4,200 bp); C: *Pks* gene RNAi vector and its specific fragments, lane1: *Pks* gene RNAi vector; lane 2: Specific fragments (*PKS-1* in MCS1 and *PKS-2* in MCS2) (1,200 bp).

Supplementary Table 1. Identification primers for *PKS* gene MT, CT and RNAi strain

Primers	Products
F8: TGGATTTTCAGTTTCGAGGTT	500 bp
R8: ATGAGTGGCATCTCAGATC	
F9: GAATACCATCTCCTTCCAC	580 bp
R9: TGATCTGACCAGTTGCCTA	
F10: CGTCATGCATTGCAGATGA	439 bp
R10: GTGGCAATAATAGCTCAAGG	
F11: CACCTGTCTCCGTTTCC	278 bp
R11: TTTGGTCCTCAACCTCCTT	
F12: CACTGAAGCGGGAAGGGACT	492 bp
R12: GCGGCGATACCGTAAAGCAC	
ITS1: TCCGTAGGTGAACCTGCGG	534 bp
ITS4: TCCTCCGCTTATTGATATGC	

Supplementary Table 2. Primers is used to measure the expression of *PKS* gene

Genes	Primers	Products	GeneBank No.
<i>PKS</i>	F-Q: ACATTGCCCTCCTTTACC	245 bp	XM_007826620.1
	R-Q: TCGGCCAGTCACGAACA		
<i>its</i> (5.8s rDNA)	F-C: AGCGAAATGCGATAAGTAATG	178 bp	NR_132017.1
	R-C: GACGGCTGTGCTGGAAAA		

Supplementary Table 3. Primers for *PKS* gene of *M. anisopliae* to mutation vector, complementary vector and RNAi vector

Target sequence	Template	Primer sequence
<i>PKS</i>	DNA of <i>M. anisopliae</i>	F1: AAGTATCATGTTAGTTGCCG R1: GTGCATCCAGGTATAACTA
<i>ben</i> gene	pBARGPE1-EGFP-BenA	Ben-F: aactcgttgtaactcaagTCAAGAATTAATTCGGTCGA Ben-R: ttaagtgatctcggtagcCGGGACTTGGCATATG
<i>PKS-I</i>	DNA of <i>M. anisopliae</i>	F2: gcctgcaggctgactctagaGAGCTGAAACCATGGGA R2: tcgaccgaattaattctgaCTTGAGTTGTACAACGAGT
<i>PKS-II</i>	DNA of <i>M. anisopliae</i>	F3: ttaagtgatctcggtagcCGGGACTTGGCATATGTAA R3: aaacgacggccagtgaattcGGCAACGCATCCATTTGAA
<i>PKS-1</i>	DNA of <i>M. anisopliae</i>	F4: aacggcagatcttcgcatgcTGGATTTTCAGTTTCGAGGTT R4: taccacaggccttagcatgcATGAGTGGCATCTCAGATC
<i>PKS-2</i>	DNA of <i>M. anisopliae</i>	F5: tcgataccgtcgacctcgagATGAGTGGCATCTCAGATC R5: aagcttgtagctaccttcgagTGGATTTTCAGTTTCGAGGTT
<i>PKS-3</i>	DNA of <i>M. anisopliae</i>	F6: tccgtcaccagccctgggttGAGCTGAAACCATGGGAC R6: aaacgacggccagtgaattcGGCAACGCATCCATTTGAA
<i>G418</i> gene	pSilent-Dual1	F7: tccgtcaccagccctgggttGAGCTGAAACCATGGGAC R7: aaacgacggccagtgaattcGGCAACGCATCCATTTGAA