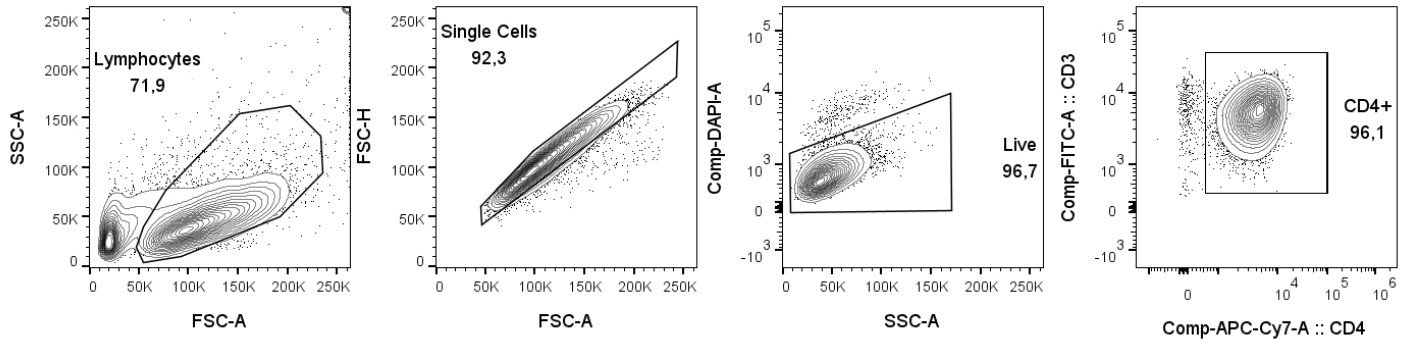
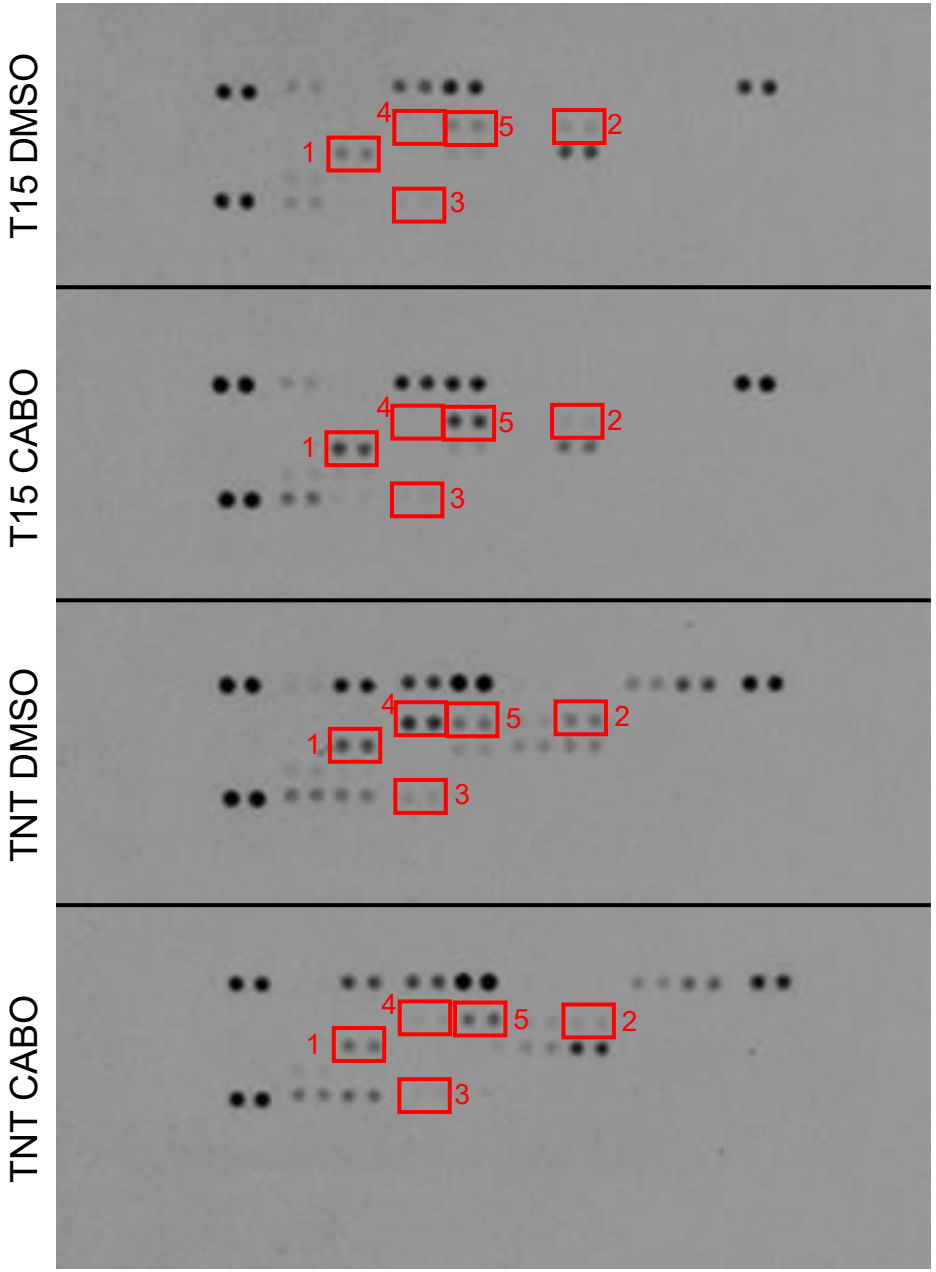
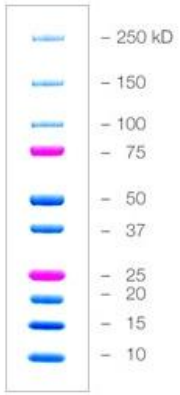
Gating Strategy



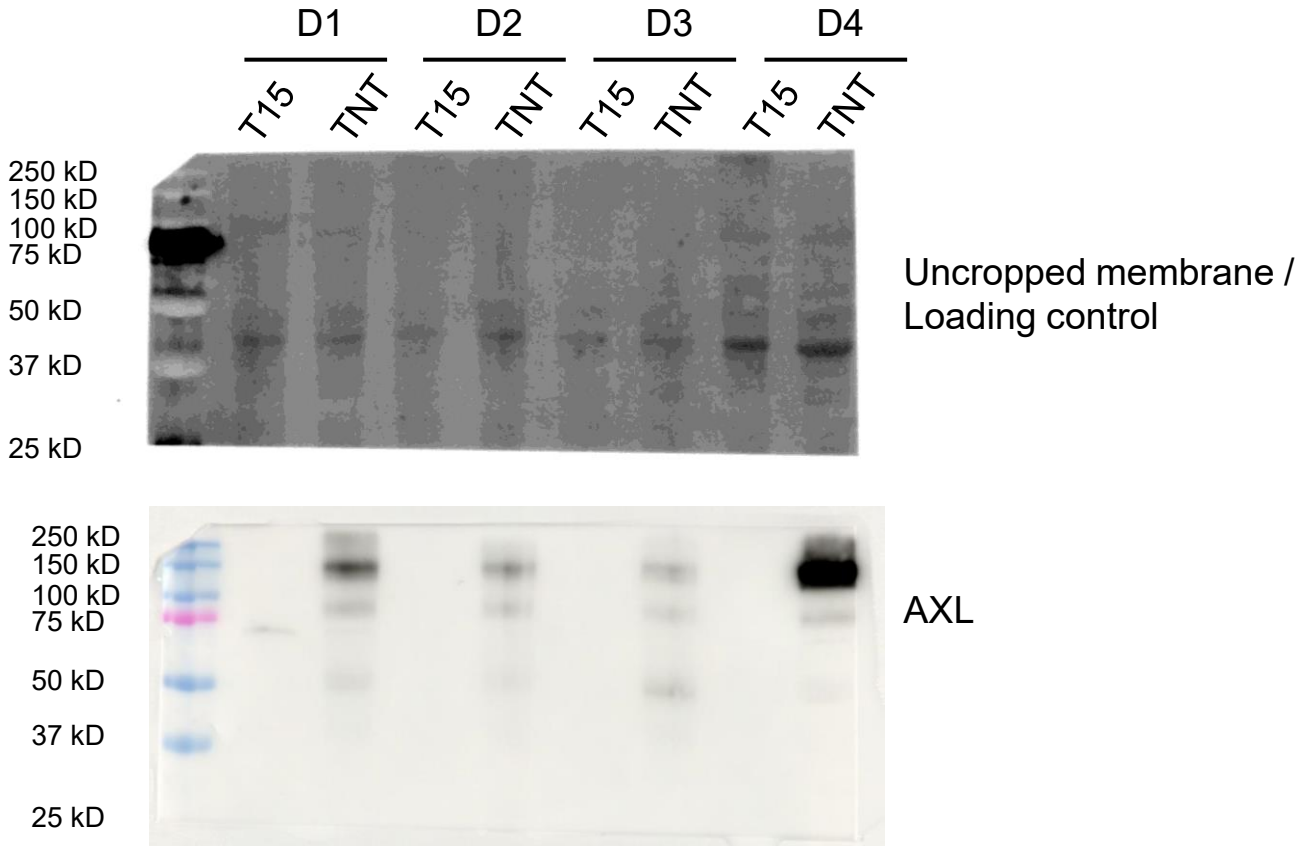
Supplementary Figure 1. Flow cytometry gating strategy. T-cell subsets were differentiated with FSC-A and SSC-A parameters, following single cells identification (FSC-A vs. FSC-H). Live subset was determined with Live/Dead Blue Staining Kit (Invitrogen), shown as DAPI parameter. Among live cells, CD3/CD4-positive cells were separated for further analysis.



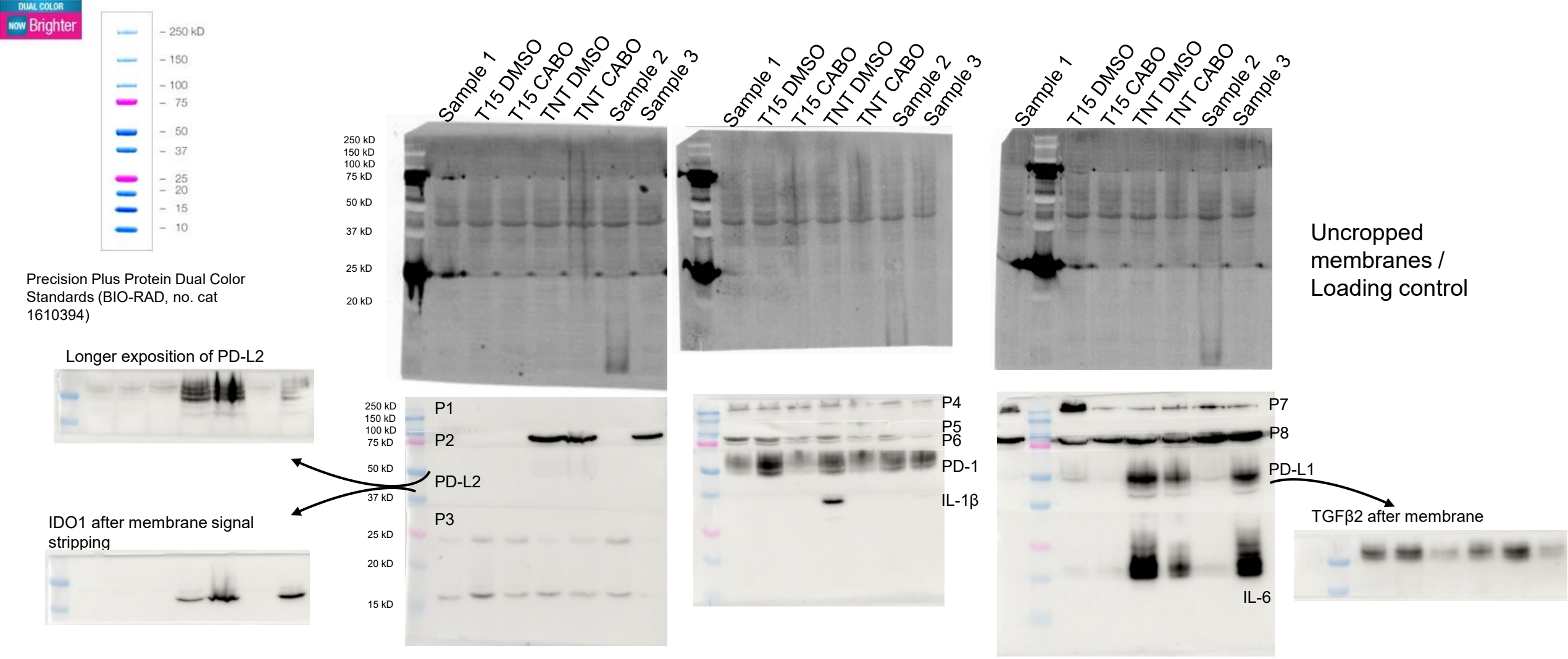
Supplementary Figure 2. Dot blots from cytokine array analysis executed on the supernatant collected from representative *in vitro* experiment. Cytokine array performed on the supernatant from four experimental groups with or without cabozantinib, which expression changed. 1 – IL-2, 2 – IFN-γ, 3 – TNF-α, 4 – G-CSF, 5 – GM-CSF.



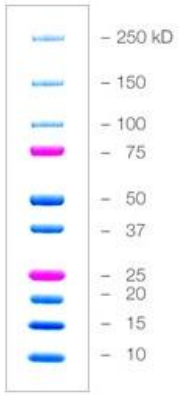
Precision Plus Protein Dual Color Standards (BIO-RAD, no. cat 1610394)



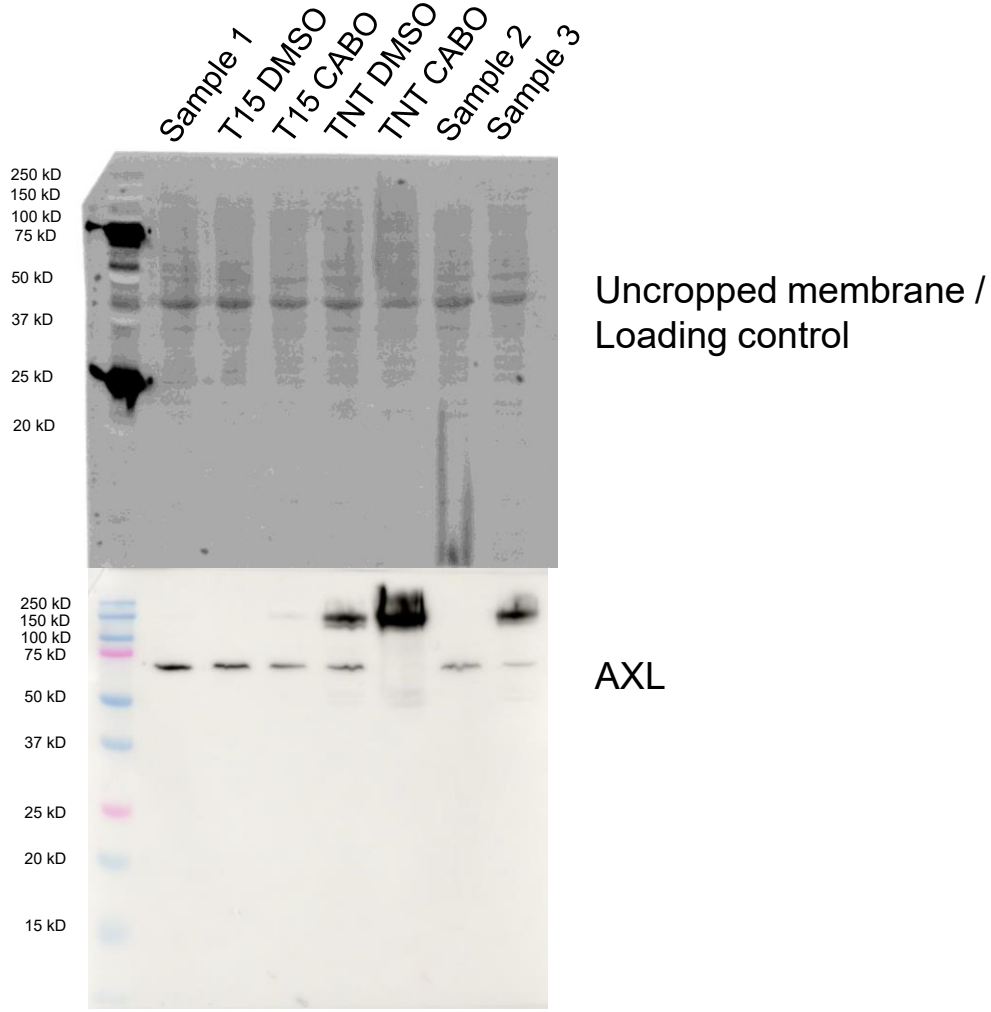
Supplementary Figure 3. Uncropped and unprocessed image of Figure 1A. The visualization of bands in the membrane was achieved using 0.5% TCE staining, which served as a protein loading control. Membranes were manually cut in a horizontal orientation using scissors to minimize the utilization of valuable samples. D1-D4 – donor number.



Supplementary Figure 4. Uncropped and unprocessed images of Figure 3B, Figure 5B and Figure 5F. The visualization of bands in the membrane was achieved using 0.5% TCE staining, which served as a protein loading control. Membranes were manually cut in a horizontal orientation using scissors to minimize the utilization of valuable samples. Chemiluminescence signal abrogation on the membranes were achieved using 30% H_2O_2 incubation for 20 min in 37°C. Samples 1-3 and proteins P1-P are irrelevant for this work.



Precision Plus Protein Dual Color Standards (BIO-RAD, no. cat 1610394)



Uncropped membrane / Loading control

AXL

Supplementary Figure 5. Uncropped and unprocessed image of Figure 4B. The visualization of bands in the membrane was achieved through the use of 0.5% TCE staining, which served as a protein loading control. Membranes were manually cut in a horizontal orientation using scissors to minimize the utilization of valuable samples. Samples 1-3 are irrelevant for this work.