

Fig. S1

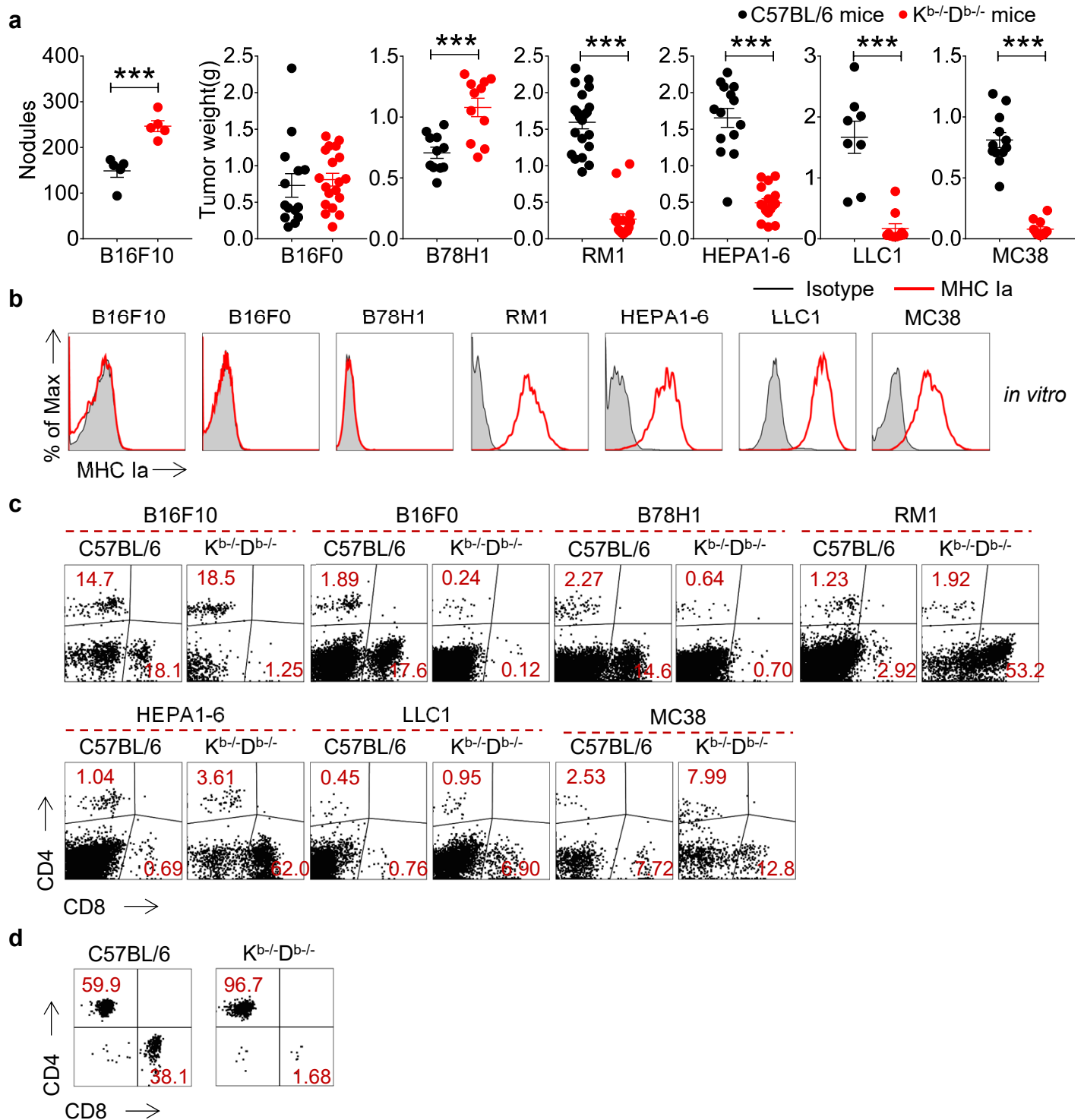
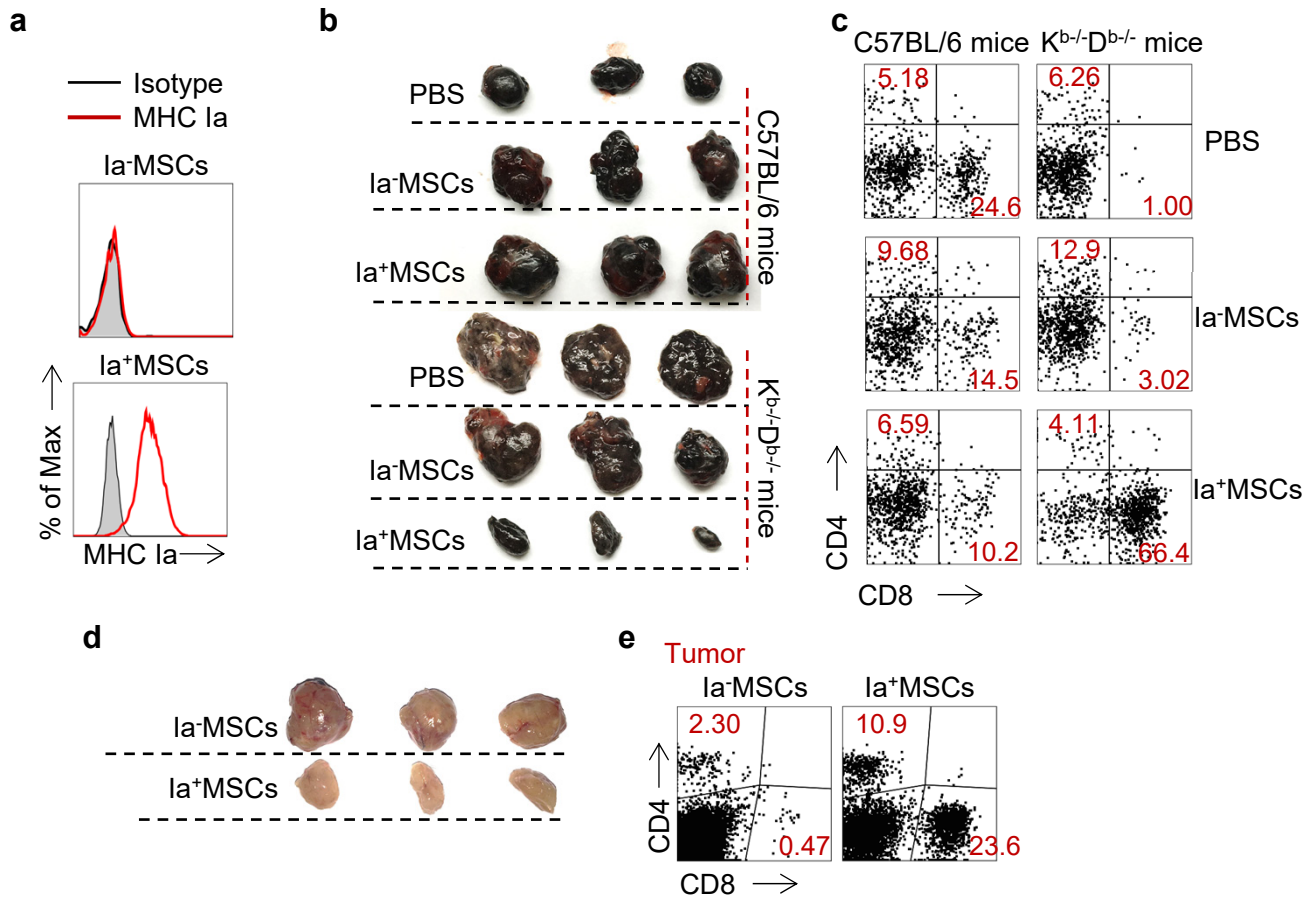


Fig. S1. The presence of MHC Ia inhibits tumor growth in K^{b-/-}D^{b-/-} mice, related to Fig. 1.

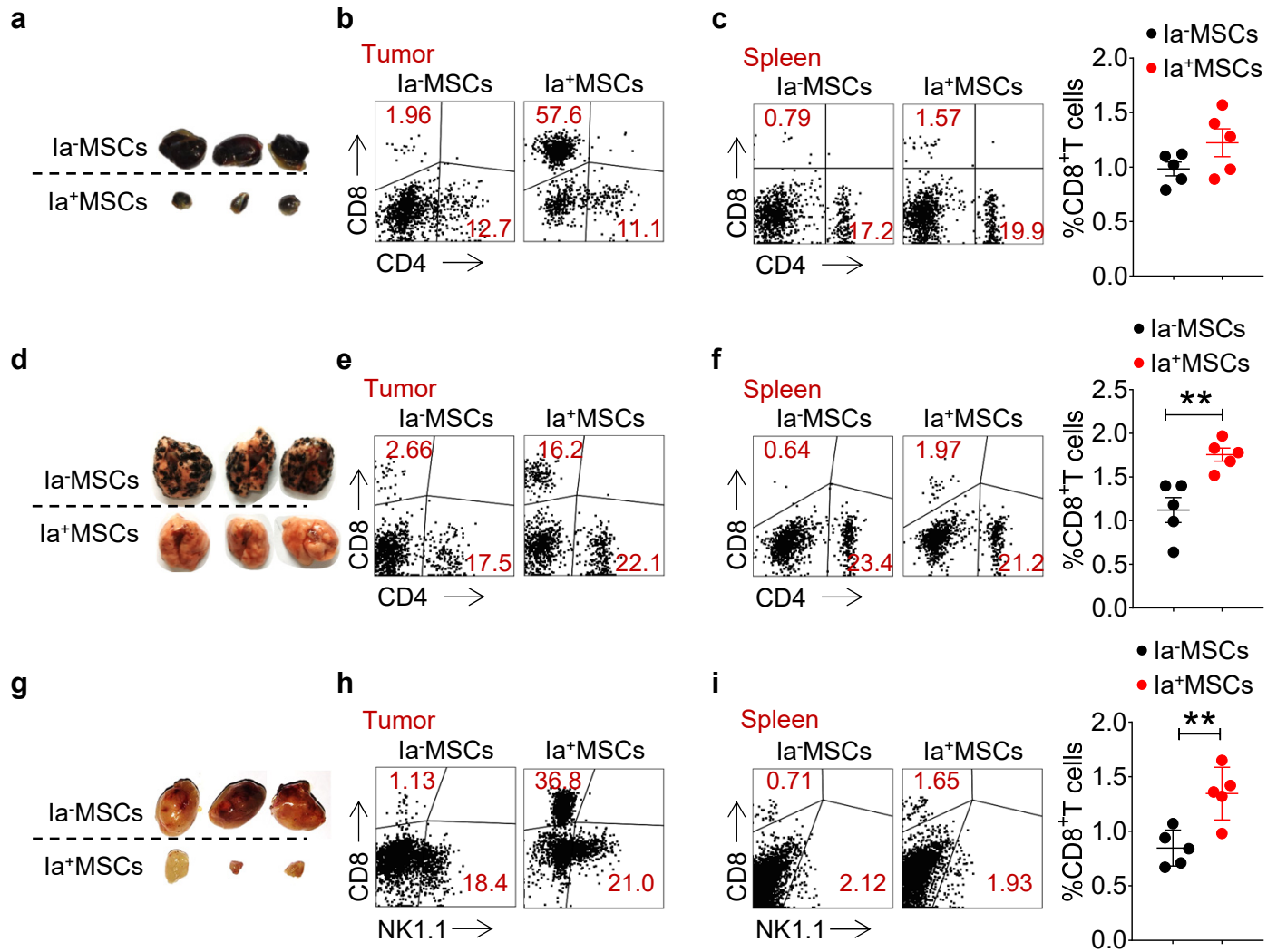
- (a) C57BL/6 and K^{b-/-}D^{b-/-} mice were inoculated with B16F10 cells (3×10^5 , i.v.) and other tumor cells (2×10^5 , i.m.) to evaluate tumor burden.
- (b) Analysis of MHC Ia expression levels on B16F10, B16F0, B78H1, RM1, HEPA1-6, LLC1, and MC38 by flow cytometry. Tumor cells were harvested from cell culture dishes.
- (c) C57BL/6 and K^{b-/-}D^{b-/-} mice were inoculated with B16F10 cells (3×10^5 , i.v.) and other tumor cells (2×10^5 , i.m.) and T cell infiltration were examined by flow cytometry.
- (d) The percentage of CD8⁺ T cells in splenic T cells of normal healthy C57BL/6 mice and K^{b-/-}D^{b-/-} mice.
- Data are presented as Mean $\pm$ SEM. ***p<0.001.

Fig. S2**Fig. S2. Co-injection of MHC Ia⁺ cells inhibits tumor growth in K^{b-/-}D^{d-/-} mice, related to Fig. 1.**

(a) The expression levels of MHC Ia on mesenchymal stromal cells isolated from K^{b-/-}D^{d-/-} mice (Ia⁻MSCs) or C57BL/6 mice (Ia⁺MSCs) were determined by flow cytometry.

(b-c) B16F0 cells (2×10^5) were i.m. administrated into C57BL/6 or K^{b-/-}D^{d-/-} mice alone or with mesenchymal stromal cells (2×10^5) derived from C57BL/6 mice (Ia⁺MSCs) or K^{b-/-}D^{d-/-} mice (Ia⁻MSCs). The mice were euthanized on day 13 post tumor inoculation to evaluate tumor burden (b) and CD8⁺ T cell infiltration at the tumor sites (c).

(d-e) B78H1 cells (2×10^5) were co-administrated with Ia⁻MSCs (2×10^5) or Ia⁺MSCs (2×10^5) to K^{b-/-}D^{d-/-} mice by i.m. injections. The mice were euthanized on day 13 post tumor inoculation to evaluate tumor burden (d) and CD8⁺ T cell infiltration at the tumor sites (e).

Fig. S3**Fig. S3. Administration of Ia⁺MSCs intraperitoneally inhibits tumor growth in $K^{b-/-}D^{b-/-}$ mice, related to Fig. 1.**

(a-c) Ia⁺MSCs or Ia⁻MSCs were i.p. administrated to $K^{b-/-}D^{b-/-}$ mice with i.m. inoculated B16F0. The size of tumor and the representative flow cytometry data of tumor infiltrated or splenic CD4⁺ T cells and CD8⁺ T cells were depicted.

(d-f) Ia⁺MSCs or Ia⁻MSCs were i.p. administrated to $K^{b-/-}D^{b-/-}$ mice with i.v. inoculated B16F10. The lungs and the representative flow cytometry data of tumor infiltrated or splenic CD4⁺ T cells and CD8⁺ T cells were depicted.

(g-i) Ia⁺MSCs or Ia⁻MSCs were i.p. administrated to $K^{b-/-}D^{b-/-}$ mice with i.m. inoculated B78H1. The size of tumor and the representative flow cytometry data of tumor infiltrated or splenic NK1.1⁺ cells and CD8⁺ T cells were depicted.

Data are presented as Mean±SEM. **p<0.01.

Fig. S4

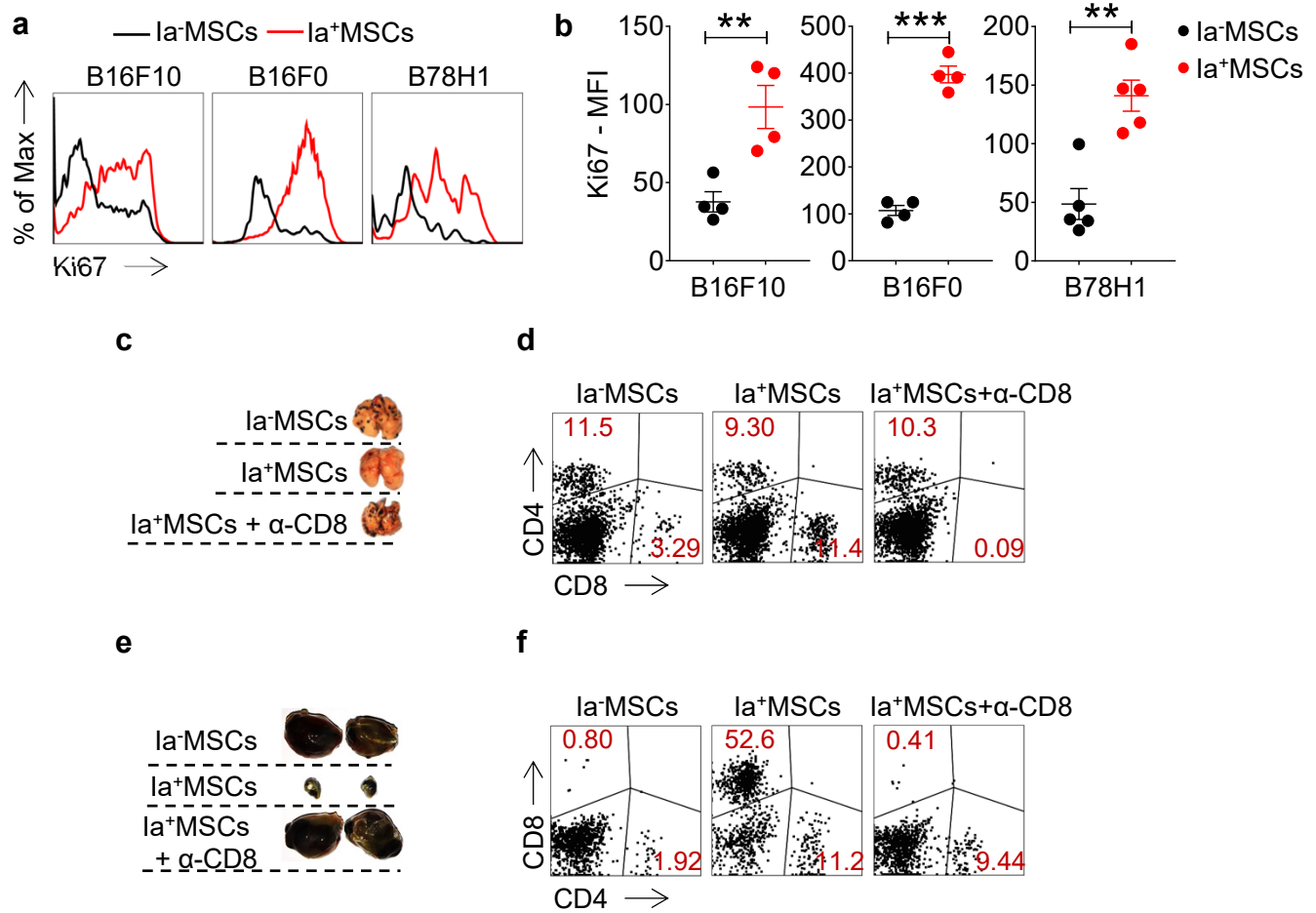


Fig. S4. Ib-CD8⁺ T cells mediate the tumor suppression effects of Ia⁺MSCs, related to Fig. 1.

(a-b) Ia⁺MSCs or Ia-MSCs were i.p administrated to K^{b-/-}D^{b-/-} mice with B16F0 (2×10^5 , i.m.), B16F10 (3×10^5 , i.v.), or B78H1 (2×10^5 , i.m.) inoculations. The Ki67 expression on tumor infiltrated CD8⁺ T cells was determined by flow cytometry.

(c-f) CD8 depletion antibodies (clone No: 2.43, 200 μg/mouse) were co-administrated together with Ia⁺MSCs to B16F0 (c-d) or B16F10 (e-f) tumor bearing mice. Tumor burden and the representative flow cytometry data of tumor infiltrated CD4⁺ T cells and CD8⁺ T cells were shown.

Data are presented as Mean ± SEM. **p<0.01, ***p<0.001.

Fig. S5

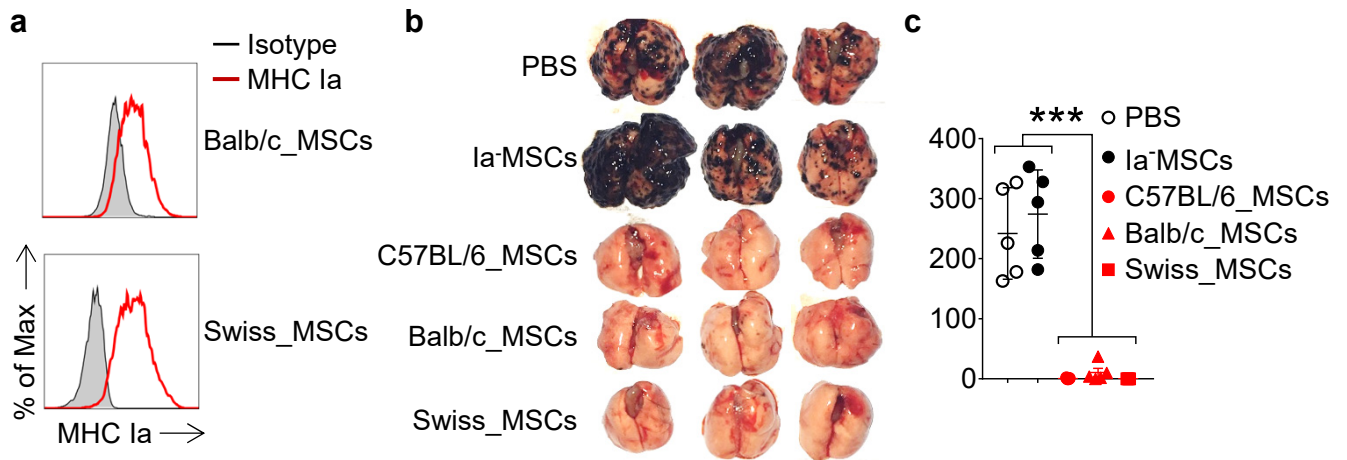


Fig. S5. Tumor suppression effects of Ia⁺MSCs in K^b^{-/-}D^b^{-/-} mice are not dependent on MHC haplotypes, related to Fig. 1.

(a-c) Mesenchymal stromal cells isolated from C57BL/6 (K^bD^b) (C57BL/6_MSCs), Balb/c (K^dD^dL^d) (Balb/c_MSCs), and Swiss (K^aD^aL^a) (Swiss_MSCs) mice were administrated to B16F10 tumor bearing K^b^{-/-}D^b^{-/-} mice. The expression levels of MHC Ia on Balb/c_MSCs and Swiss_MSCs were analyzed (a). The tumor burdens in lungs were determined (b-c). Data are presented as Mean±SEM. ***p<0.001.

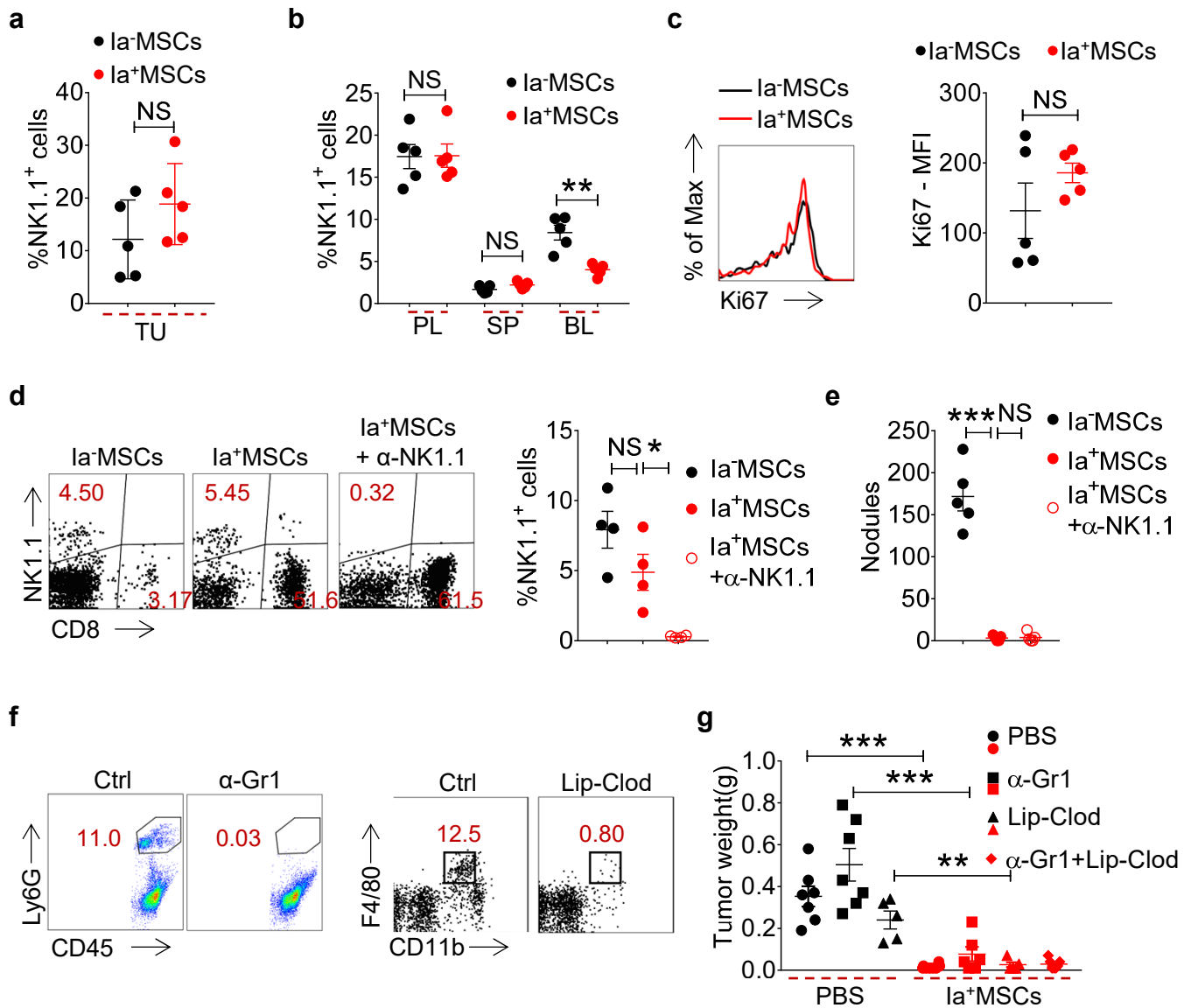
Fig. S6

Fig. S6. NK cells and myeloid cells are not involved in MHC Ia-mediated Ib-CD8⁺ T cell expansion and tumoricidal effects, related to Fig. 1.

(a-c) Ia⁺MSCs or Ia⁻MSCs were i.p. administrated to K^b-D^b- mice with i.v. injected B16F10. The percentage of NK1.1⁺ cells in tumor (TU), peritoneal lavage (PL), spleen (SP), and blood (BL) was determined (a-b). Ki67 staining was applied to measure the proliferation of tumor infiltrated NK1.1⁺ cells (c).

(d-e) NK1.1 depletion antibody (clone No: PK136, 200 μ g/mouse) was administrated to Ia⁺MSCs treated B16F10 tumor bearing K^b-D^b- mice. The efficiency of NK1.1⁺ cell deletion (d) and the impact of NK1.1⁺ cell deletion on tumor progression (e) were analyzed.

(f-g) Gr1 antibodies (clone No: RB6-8C5, 200 μ g/mouse) and clodronate liposomes were applied to deplete myeloid cells in Ia⁺MSCs treated B16F0 tumor bearing K^b-D^b- mice (f). The tumor weight after Gr1 antibodies and clodronate liposomes treatment was determined (g).

Data are presented as Mean $\pm$ SEM. * p <0.05, ** p <0.01, *** p <0.001, NS, no significance

Fig. S7

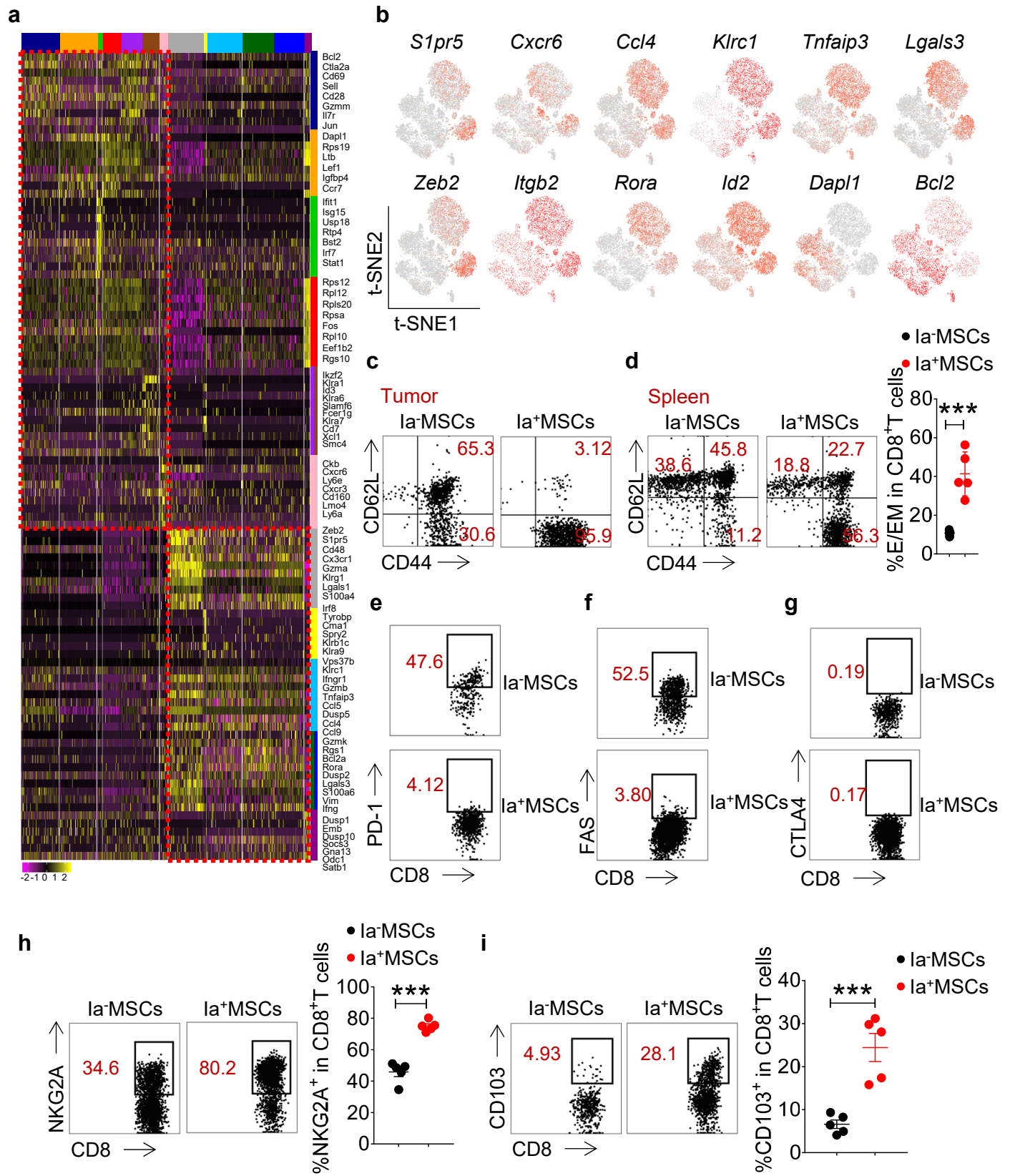


Fig. S7. Characterization of Ib-CD8⁺ T cells in tumor bearing mice treated with MSCs, related to Fig. 2 and Table S1.

(a) The unsupervised heatmap view of CD8⁺ T cell subsets and their marker genes in Fig. 2a.

(b) Relative expression marker genes of CD8⁺ subsets superimposed on the t-SNE projections shown in Fig. 2a.

(c-d) Ia⁻MSCs or Ia⁺MSCs were i.p. administrated to K^{b-/-}D^{b-/-} mice with B16F10 i.v. inoculation. The CD44 and CD62L expression in tumor infiltrated and splenic CD8⁺ T cells were examined. E/EM represented CD44⁺CD62L⁻ effector and effector memory CD8⁺ T cells.

(e-i) Ia⁻MSCs or Ia⁺MSCs were i.p. administrated to K^{b-/-}D^{b-/-} mice with B16F10 i.v. inoculation. The expression levels of PD-1 (e), FAS (f), CTLA4 (g), NKG2A (h), and CD103 (i) in tumor infiltrated CD8⁺ T cells were examined.

Data are presented as Mean±SEM. ***p<0.001.

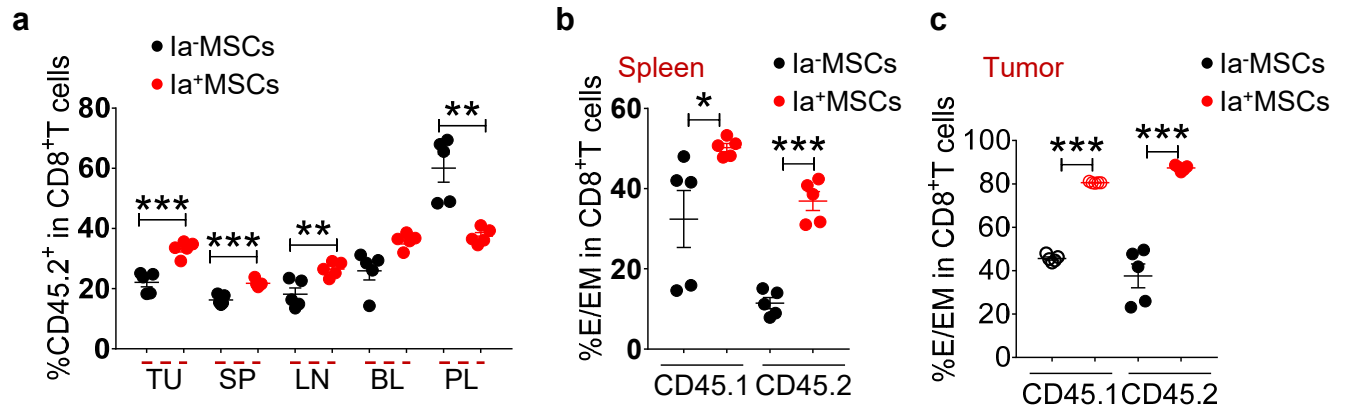
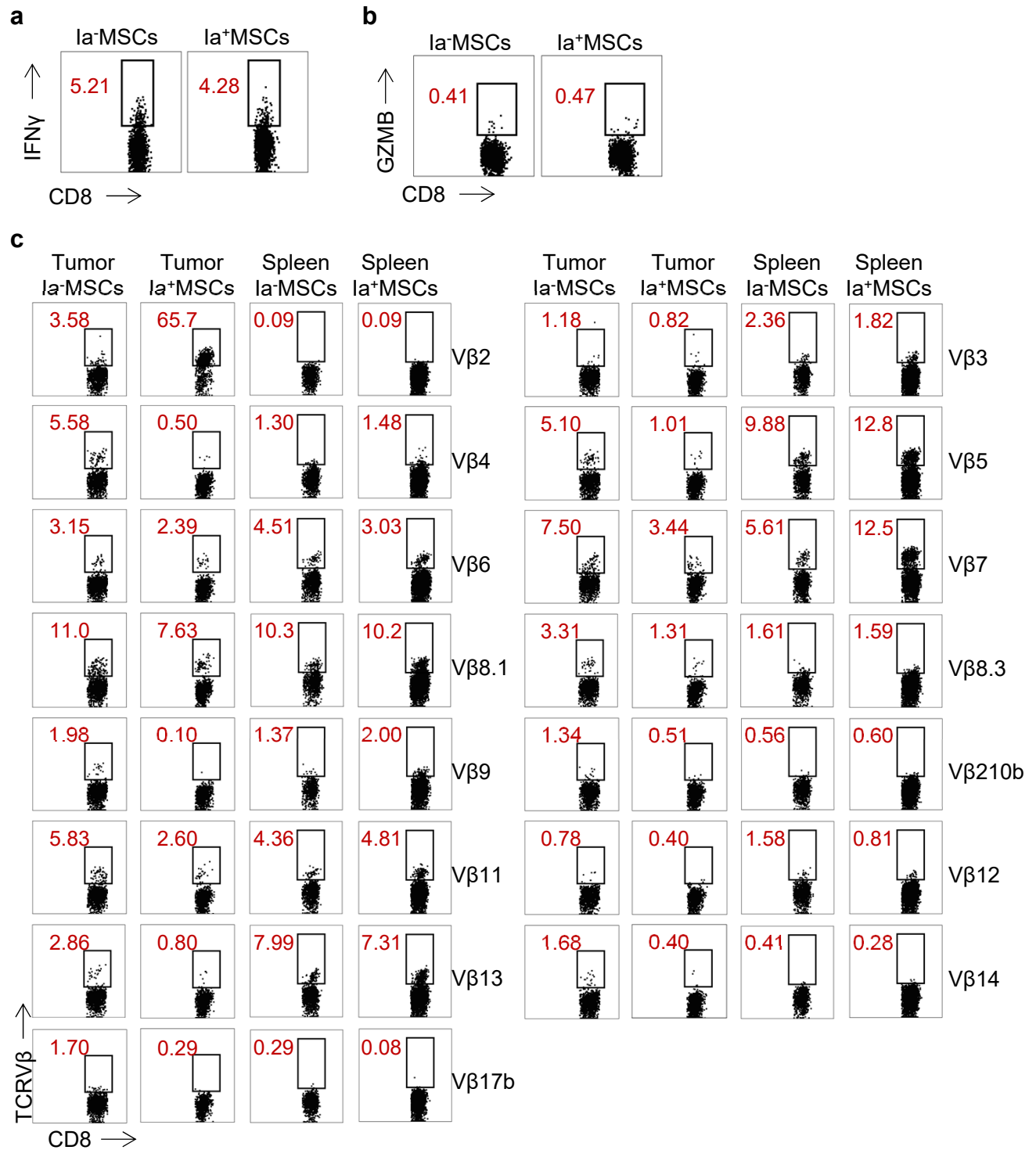
Fig. S8

Fig. S8. Peritoneal Ib-CD8⁺ T cells migrate to tumor and lymphoid organs after encountering Ia⁺MSCs, related to Fig. 2.

(a) B16F10 tumor bearing CD45.1⁺K^{b/-}D^{b/-} mice were i.p. injected with splenocytes derived from CD45.2⁺K^{b/-}D^{b/-} mice together with Ia⁻MSCs or Ia⁺MSCs. The percentage of transferred CD45.2⁺ Ib-CD8⁺ T cells in total CD8⁺ T cells in tumor (TU), spleen (SP), lymph node (LN), blood (BL), and peritoneal lavage (PL) were analyzed.

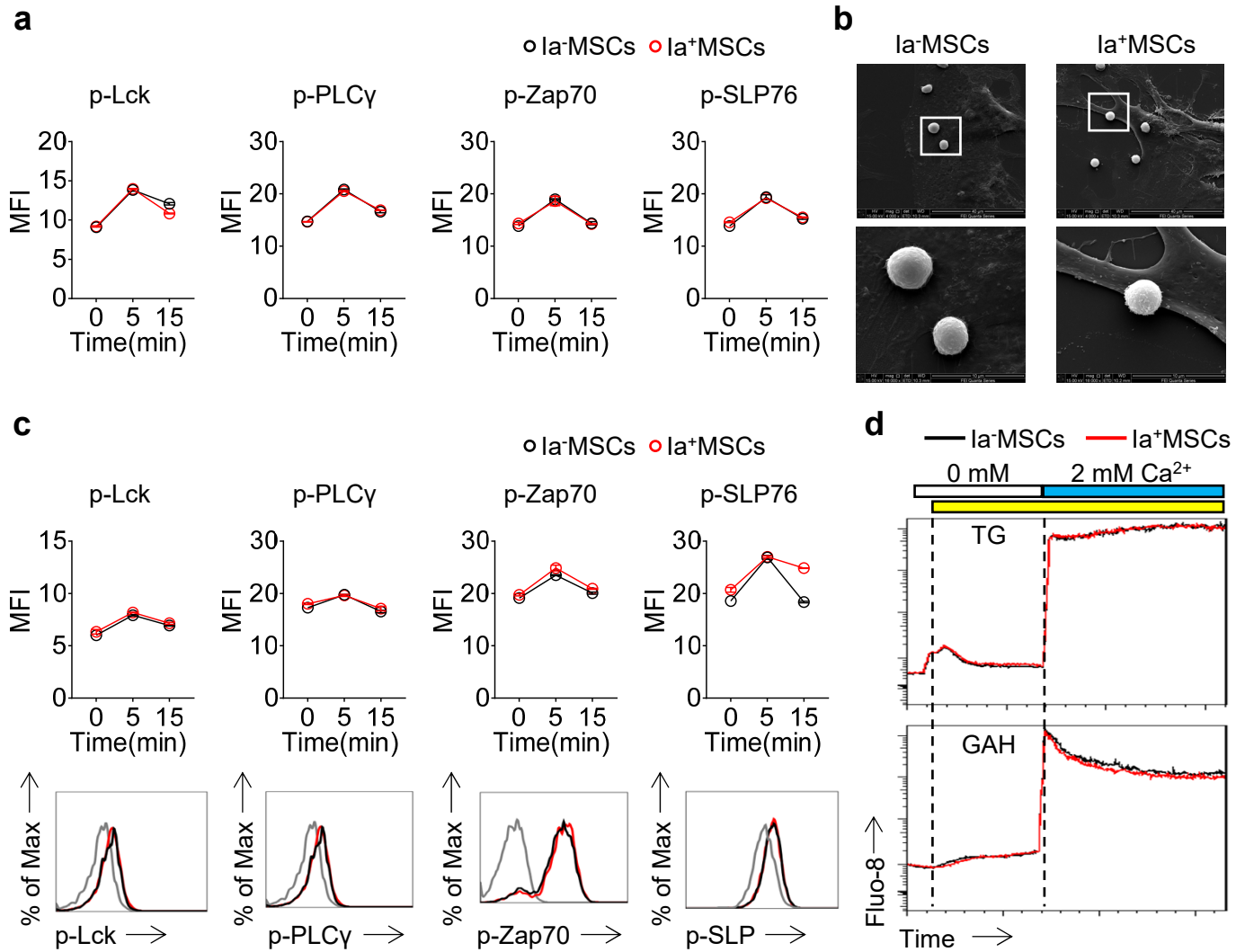
(b-c) The percentage of CD44⁺CD62L⁻ effector and effector memory (E/E/M) cells in CD45.1⁺ or CD45.2⁺ splenic or tumor infiltrated CD8⁺ T cells was determined by flow cytometry.

Data are presented as Mean±SEM. *p<0.05, **p<0.01, ***p<0.001.

Fig. S9**Fig. S9. TCRV β usage in CD8⁺ T cells of K^b^{-/-}D^b^{-/-} mice, related to Fig. 3.**

(a-b) Ia⁺MSCs were i.p. administrated to K^b^{-/-}D^b^{-/-} mice with B16F10 i.v. inoculation. The IFN γ (a) and GZMB (b) expression levels in splenic CD8⁺ T cells were examined.

(c) K^b^{-/-}D^b^{-/-} mice were i.v. injected with B16F10 cells (3×10^5). The tumor bearing mice were i.p. injected with Ia⁻MSCs (1×10^6) or Ia⁺MSCs (1×10^6) every 3 days starting on day 0. After 16 days, splenic and tumor infiltrated CD8⁺ T cells were tested for their TCRV β usage.

Fig. S10**Fig. S10. MHC Ia could not prime Ia-CD8⁺ T cells, related to Fig. 6.**

- (a) Purified CD8⁺ T cells from C57BL/6 mice were activated in the presence of Ia⁺MSCs or Ia⁻MSCs. The phosphorylation of Lck, PLCγ, Zap70 and SLP76 was determined at 0, 5, and 15 min by flowcytometry.
- (b) CD8⁺ T cells were isolated from the spleen of C57BL/6 mice and co-cultured with Ia⁺MSCs or Ia⁻MSCs. The images by scanning electron microscopy were obtained after 24 hours co-culture.
- (c) CD8⁺ T cells from C57BL/6 mice were co-cultured with Ia⁺MSCs or Ia⁻MSCs for 24 hours. CD8⁺ T cells were then separated from co-culture system and stimulated with α-CD3/CD28. The phosphorylation of Lck, PLCγ, Zap70 and SLP76 in CD8⁺ T cells was determined. Histograms in the lower panel represents the signal intensities of phosphorylated protein after 5 min stimulation.
- (d) Store-operated Ca²⁺ influx in CD8⁺ T cells from C57BL/6 mice co-cultured with Ia⁺MSCs or Ia⁻MSCs in response to 1 μM thapsigargin (TG; upper panel) or goat-anti-hamster IgG (GAH; lower panel) mediated crosslinking of anti-CD3 and anti-CD28 antibodies.

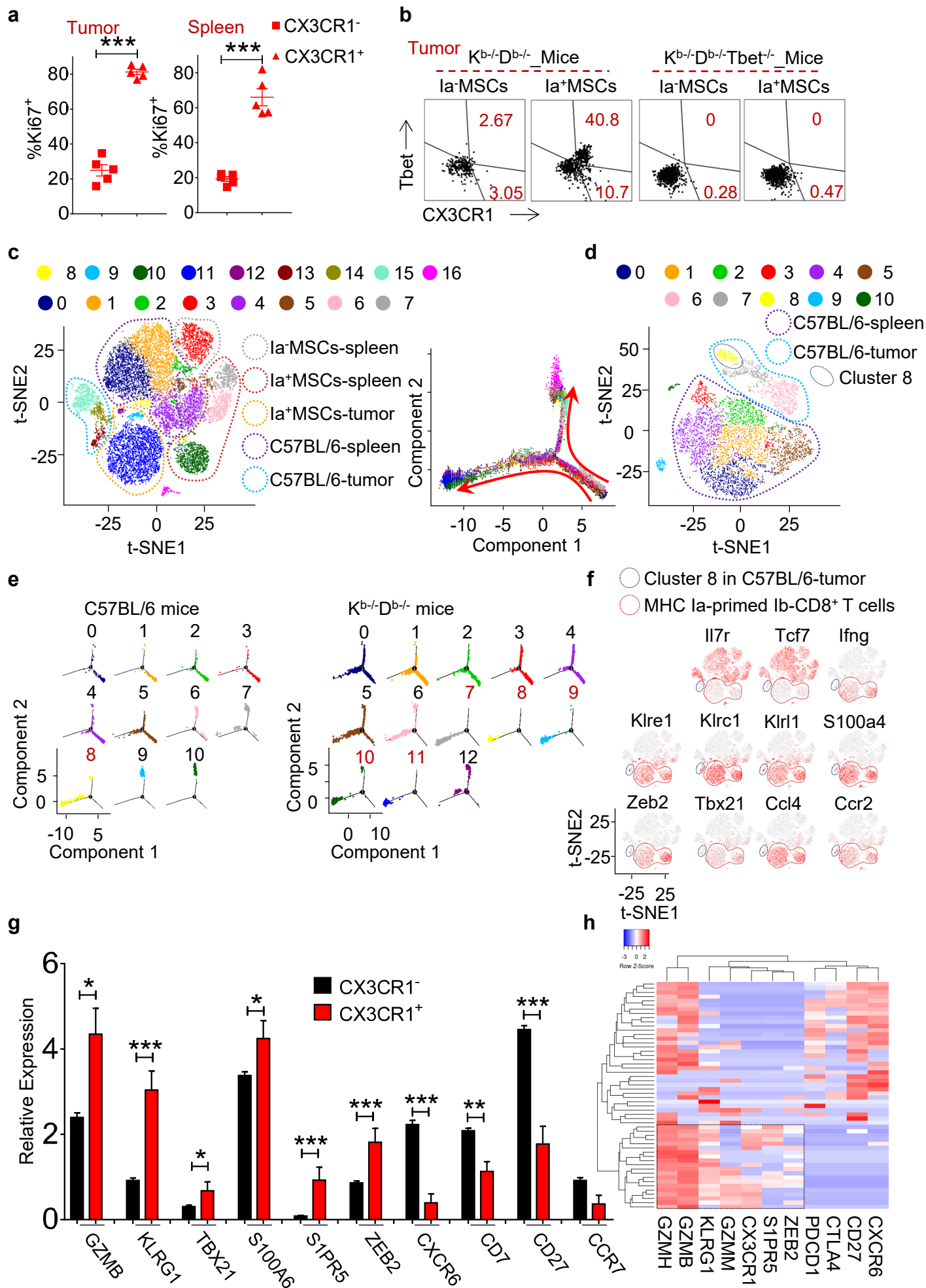
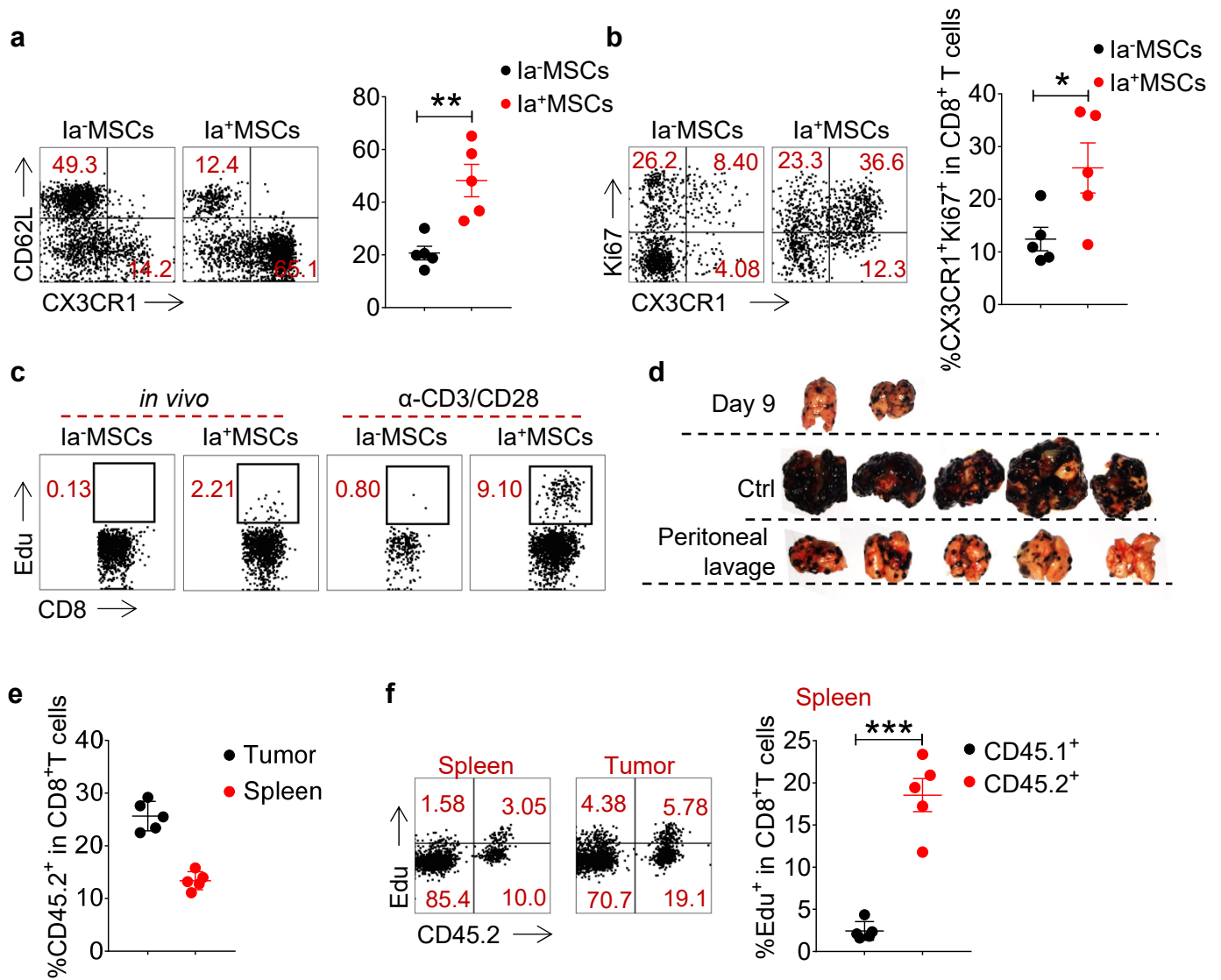
Fig. S11

Fig. S11. Existence of tumoricidal CX3CR1⁺CD8⁺ T cells in wild type mice and melanoma patients, related to Fig. 6, Table S1, Table S4, Table S6, and Table S7.

- (a) In B16F10 tumor bearing K^{b/-}D^{b/-} mice receiving Ia⁺MSC administration, tumor infiltrated and splenic CD8⁺ T cells were classified into two groups based on CX3CR1 expression. The percentages of Ki67 positive cells in each group were determined by flow cytometry.
- (b) B16F10 tumor model was established in K^{b/-}D^{b/-} mice and K^{b/-}D^{b/-}Tbet^{-/-} mice by i.v injection of B16F10 cells. Ia⁻MSCs (1×10⁶) or Ia⁺MSCs (1×10⁶) were i.p. injected into tumor bearing mice every 3 days starting on day 0. The expression levels of CX3CR1 and Tbet in tumor infiltrated CD8⁺ T cells were determined by flow cytometry.
- (c) B16F10 melanoma were i.v. inoculated into C57BL/6 mice and K^{b/-}D^{b/-} mice. K^{b/-}D^{b/-} mice received i.p. administrated Ia⁺MSCs or Ia⁻MSCs every 3 days starting on day 0. On day 9, splenic CD8⁺ T cells from Ia⁻MSCs treated K^{b/-}D^{b/-} mice (Ia⁻MSCs-spleen), splenic or tumor infiltrated CD8⁺ T cells from Ia⁺MSCs treated K^{b/-}D^{b/-} mice (Ia⁺MSCs-spleen, Ia⁺MSCs-tumor), as well as splenic or tumor infiltrated CD8⁺ T cells from C57BL/6 mice (C57BL/6-spleen, C57BL/6-tumor) were isolated using magnet beads and subjected to single cell RNA sequencing. The t-SNE plot of the analyzed cells with their tissue origin was depicted (left panel). Pseudotime analysis of CD8⁺ T cells isolated from C57BL/6 mice and K^{b/-}D^{b/-} mice is shown (right panel).
- (d) The t-SNE plot view of splenic and tumor infiltrated CD8⁺ T cells isolated from C57BL/6 mice (C57BL/6-spleen, C57BL/6-tumor) was depicted.
- (e) The position of each CD8⁺ T cell subsets originated from either C57BL/6 mice (shown in Fig. S11d) or K^{b/-}D^{b/-} mice (shown in Fig. 2a) in the pseudotime trajectory of total CD8⁺ T cells (shown in Fig. S11c) was depicted.
- (f) Relative expression intensity of selected genes superimposed on the t-SNE projections shown in (c). Those MHC Ia primed Ib-CD8⁺ T cells (red dashed line) and CD8⁺ T cell population in C57BL/6 mice (blue dashed line) similar to MHC Ia primed Ib-CD8⁺ T cells are circled.
- (g-h) Re-analysis on tumor infiltrated CD8⁺ T cells derived from melanoma patients from published single cell transcriptome datasets. The average genes expression levels of CX3CR1⁺ and CX3CR1⁻ CD8⁺ T cells were determined (g) (Tirosh et al., 2016) and the heatmap view of selected gene expression of tumor infiltrated CD8⁺ T cells in melanoma patients (h) (Jerby-Arnon et al., 2018).

Data are presented as Mean±SEM. *p<0.05, **p<0.01, ***p<0.001.

Fig. S12**Fig. S12. MHC Ia primed CX3CR1⁺ Ib-CD8⁺ T cells are enriched in peritoneal lavage, related to Fig. 6.**

(a-b) The Ki67, CX3CR1, and CD62L expression levels on CD8⁺ T cells in the peritoneal lavages of K^b-D^b- B16F10 (3×10⁵, i.v.) tumor bearing mice receiving i.p. injected Ia⁺MSCs or Ia⁻MSCs were analyzed by flow cytometry.

(c) Edu incorporation assay was applied to measure the proliferation of peritoneal CD8⁺ T cells *in vivo*. The peritoneal CD8⁺ T cells were isolated using magnet beads and stimulated with α -CD3/CD28 *in vitro* to evaluate their responsiveness to TCR activation.

(d-f) B16F10 cells were i.v. injected into CD45.1⁺C57BL/6 mice and irradiated (5 Gy) on day 9. Then they received peritoneal lavage from Ia⁺MSCs pre-treated CD45.2⁺K^b-D^b- mice every 3 days. The mice were euthanized on day 22 post tumor inoculation. The photograph of the lungs of CD45.1⁺C57BL/6 mice with different treatment were shown (d). The tumor burden at the treatment time point on day 9 is also depicted (d). The reconstitution rates of CD45.2⁺Ib-CD8⁺ T cells in the spleen and tumor of wild type mice were determined (e). The ratio of proliferating cells among CD45.1⁺ host CD8⁺ T cells and CD45.2⁺ Ib-CD8⁺ T cells was measured by Edu incorporation assay (f).

Data are presented as Mean±SEM. *p<0.05, **p<0.01, ***p<0.001.